

HTLV-I AND HTLV-II GP21 DERIVED PEPTIDES FOR USE IN DIAGNOSTICS.

This application is a continuation-in-part of co-
 pending U.S. patent application for "HTLV-I and HTLV-II
 Peptide Antigens and Methods", Serial No. 07/653,091,
 filed February 08, 1991, which in turn is a
 continuation-in-part of U.S. patent application "HTLV-I
 Peptide Antigen and Methods", Serial No. 366,313, filed
 June 13, 1989, now U.S. Patent No. 5,066,579, issued
 November 19, 1991, which in turn is a continuation of
 U. S. patent application for "HTLV-I Peptide Antigen
 and Methods," Serial No. 948,270, filed December 31,
 1986, now abandoned.

1. Field of the Invention

The present invention relates to an HTLV-specific
 peptide and to methods of preparing and using the anti-
 gen.

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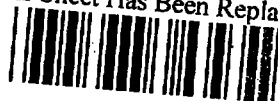


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15 3. Background of the Invention

The human T-cell leukemia viruses (HTLV) represent a family of T-cell retroviruses with three known members. HTLV type I (HTLV-I) has transforming activity *in vitro* and is etiologically linked to adult T-cell
20 leukemia, which is known to be endemic in several parts of the world. HTLV-II is another retrovirus having transforming capacity *in vitro*, and has been isolated from a patient with a T-cell variant of hairy cell leukemia (for a review of HTLV-I and II see Cann and
25 Chen). HTLV-III, which has also been called lymphadenopathy-associated virus and is now known as the human immunodeficiency virus (HIV), is lytic for certain kinds of T cells and has been linked to the etiology of acquired immunodeficiency syndrome (AIDS).
30 Unlike the HTLV-I and -II viruses, HTLV-III is not known to have *in vitro* transforming activity.

The diagnosis of HTLV-I infection is usually based on serum antibody response to HTLV-I peptide antigens. This usually involves an initial screening assay to
35 identify HTLV-I antibodies, based on an enzyme immunoassay (EIA) with HTLV-I virion peptides. The assays presently used for blood screening detect about 0.5 to

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0.05% HTLV-I and HTLV-II positives in blood donors in the United States; of these about 4 out of 5 are false positives. Therefore, positive sera must be further tested in a confirmatory assay, using Western blotted HTLV-I viral lysate. Current blood testing procedures require that individuals possess antibodies to both the HTLV-I p24 gag protein and at least one of the envelope proteins gp46, and gp68 (public health service working group). However, it has proven to be technically difficult to detect gp46 or gp68 proteins using a Western blot assay. Therefore a second round of confirmatory radioimmunoprecipitation assays must often be performed to detect antibody reaction to the HTLV-I envelope proteins.

A partial solution to this problem was provided by the molecular cloning of a 134 amino acid portion of the transmembrane glycoprotein gp21 (Samuel et al.). The recombinant protein, referred to as p21E protein, is reactive with sera from both HTLV-I and HTLV-II infected individuals, and has been successfully incorporated into Western blot assays for confirmation of HTLV infection (Lillehoj et al., Lipka et al. 1991). However, the p21E protein was also found to be reactive with 0.6% of HTLV negative blood donors (Lal, et al.). In addition, much higher rates of reactivity to p21E (approximately 5% in U.S. blood donors) are observed in individuals who are reactive in HTLV screening EIA tests, but who do not Possess antibodies to both HTLV-I gag and env gene products when tested by HTLV confirmatory assays and thus do not meet established criteria for being HTLV infected (Lal, et al.; Lipka, et al., 1991). Additionally in the study by Lipka et al. (Lipka et al. 1991) all of the p21E reactive-HTLV indeterminate individuals were negative for the presence of HTLV-I and HTLV-II nucleic acids when tested by PCR using HTLV-I and HTLV-II specific primers and probes. Therefore some individuals who are not

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infected with HTLV-I or HTLV-II possess antibodies which react with the p21E antigen. This fact has limited the use of the p21E recombinant protein, particularly in HTLV screening assays in which a high rate of false positives with HTLV-negative sera would result in the needless disposal of donated blood.

It would therefore be desirable to provide an improved method for detecting HTLV-I and HTLV-II positive sera. In particular, the improved test should be capable of detecting all HTLV-I and HTLV-II positive sera, with a minimum number of false positives, and also be able to distinguish HTLV-I from HTLV-II infected sera.

4. Summary of the Invention

The invention includes, in one aspect, a peptide having an HTLV-specific antigenic region consisting essentially of the amino acid sequence identified by SEQ ID NO: 1. The peptide is characterized by (i) immunoreactivity to sera from human subjects infected with HTLV-I or HTLV-II, and (ii) non-immunoreactivity with sera that is (1) obtained from humans who are not infected with HTLV-I or HTLV-II, but (2) immunoreactive with HTLV-1 p21E antigen.

Also disclosed is a kit for detecting the presence of HTLV-I or HTLV-II infection in a human serum. The kit includes (a) a solid support, (b) the above HTLV-specific peptide attached to the support, and (c) a reporter reagent for detecting the presence of human antibodies bound to said support.

In one embodiment, for use in detecting HTLV-I or HTLV-II infection, the solid support includes two reaction zones, one coated with the above HTLV-specific peptide, and a second zone coated with a HTLV-specific antigenic region consisting essentially of the amino acid sequence identified by SEQ ID NO: 2. This peptide is characterized by immunoreactivity with sera that is

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(1) obtained from humans who are not infected with HTLV-I or HTLV-II, but (2) immunoreactive with HTLV-1 p21E antigen.

5 The kit may also contain one or more reaction
zones designed to allow discrimination between HTLV-I
and HTLV-II infection. One preferred peptide for use
in this kit is a peptide having an HTLV-I specific
antigenic region consisting essentially of the sequence
identified by SEQ NO ID: 3. Alternatively, or in
10 addition, the kit may include a peptide having an HTLV-
II specific antigenic region consisting essentially of
the sequence identified by SEQ NO ID: 4.

15 In another aspect, the invention includes a method
of positively identifying HTLV-I or HTLV-II infection
in a human subject. The test includes reacting serum
from the subject with a peptide containing an HTLV-
specific antigenic region consisting essentially of the
amino acid sequence identified by SEQ ID NO: 1, to form
an immune complex the peptide and antibodies in serum
20 from a subject having an HTLV-I or HTLV-II infection.
The peptide is then examined for the presence of the
immune complex.

25 Also disclosed is a vaccine composition for
immunizing an individual against HTLV-I and HTLV-II
infection. The composition includes a peptide having
an HTLV-specific antigenic region consisting
essentially of the amino acid sequence identified by
SEQ ID NO: 1. The peptide is coupled to a carrier
protein.

30 In still another aspect, the invention includes a
passive vaccine from HTLV prophylaxis and treatment,
and method of passive vaccination employing a human
monoclonal antibody or human recombinant antibody
specific against the 2B3A peptide.

35 These and other objects and features of the pre-
sent invention will become more fully apparent when the
following detailed description of the invention is read

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in conjunction with the accompanying drawings.

Brief Description of the Drawings

Fig 1 shows, at the top, the HTLV-I genome, in the middle, an expanded portion of the genome containing the coding regions for gp46 and gp21 envelope proteins, and in the lower portion of the figure, the coding regions corresponding to recombinant HTLV-I peptides p21E, 2A2B, 2B3A, and 3A3B within the gp21 coding region, and the HTLV-I MTA-1, MTA-4, and MTA-5 peptides in the gp46 coding region.

Figs. 2A and 2B show the polynucleotide coding sequence (SEQ ID NO: 5) and the corresponding amino acid sequence (SEQ ID NO: 6) for the p21E env protein of HTLV-I (Fig. 2A), and the polynucleotide coding sequence (SEQ ID NO: 7) and the corresponding amino acid sequence (SEQ ID NO: 8) for the p21E region of the gp21 env protein of HTLV-II (Fig. 2B). The 5' ends of the HTLV-I specific sequences of the oligonucleotide primers used in the construction of the HTLV-I recombinant peptides are indicated.

Figs. 3A-3C show the polynucleotide sequences of forward primers (Fig. 3A) identified herein as 1A (SEQ ID NO: 9), 2A (SEQ ID NO: 10), MF1 (SEQ ID NO: 11), MF2 (SEQ ID NO: 12) and 3A (SEQ ID NO: 13), and the reverse primers (Fig. 3B) identified herein as 1B (SEQ ID NO: 14), 2B (SEQ ID NO: 15), MR1 (SEQ ID NO: 16), MR2 (SEQ ID NO: 17), and 3B (SEQ ID NO: 18), where the restriction enzyme recognition sequences located in the primers are underlined. The restriction enzyme sites of the modified pGEX plasmid pGEX-GLI are also presented (SEQ ID NO: 44) (Fig. 3C).

Fig. 4 shows the amino acid sequences of the HTLV-I recombinant peptides identified as p21E (SEQ ID NO: 6), 1A1B (SEQ ID NO: 19), 2A2B (SEQ ID NO: 20), 2A3B (SEQ ID NO: 21), 2B3A (SEQ ID NO: 22), 3A3B (SEQ ID NO: 30), MF1R2 (SEQ ID NO: 24), and MF2R1 (SEQ ID NO: 25).

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Fig. 5 shows the amino acid sequences of the peptide 2B3A which was actually constructed (SEQ ID NO: 22) and the corresponding sequence from three different HTLV-I strains (SEQ ID NOS: 26, 27, and 28), a
 5 corresponding HTLV-II 2B3A peptide (SEQ ID NO: 29), and the consensus sequence for the 2B3A peptide (SEQ ID NO: 1).

Fig. 6 shows the amino acid sequences of the peptide 3A3B which was actually constructed (SEQ ID NO: 23) and the corresponding sequence from three different
 10 HTLV-I strains (SEQ ID NOS: 31, 32, and 2), a corresponding HTLV-II 3A3B peptide (SEQ ID NO: 34), and the consensus sequence for the 3A3B peptide (SEQ ID NO: 2).

Fig. 7 shows a comparison of the polynucleotide coding sequences for the HTLV-I peptide 2B3A (SEQ ID NO: 35) and the homologous HTLV-II peptide 2B3A(II) (SEQ ID NO: 36).

Fig. 8 shows a comparison of the polynucleotide coding sequences for the HTLV-I peptide 3A3B (SEQ ID NO: 37) and the homologous HTLV-II peptide 3A3B(II) (SEQ ID NO: 38).

Fig. 9 shows the amino acid sequences of homologous regions of HTLV-I and HTLV-II gp46 protein
 25 in the region of the gp46 peptides identified in Fig. 1 as MTA-1 (SEQ ID NO: 3), MTA-4 (SEQ ID NO: 39), and MTA-5 (SEQ ID NO: 40), additional HTLV-I peptide K163 (SEQ ID NO: 41), and analogous HTLV-II peptides identified herein as K15 (SEQ ID NO: 42) and 4 (SEQ ID
 30 NO: 4).

Figs. 10A-10D illustrate a 4-zone solid phase assay plate for use in an assay method for detecting HTLV-I or HTLV-II infection in human sera (10A),
 35 showing a typical assay result for an HTLV-I positive individual (Fig. 10B), for an HTLV-II positive individual (Fig. 10C), and for a false positive (Fig. 10D).



Detailed Description of the InventionI. Definitions

Unless indicated otherwise, the terms given below have the following meanings:

5 "HTLV-composite env sequence" is formed by aligning homologous amino acid sequences of regions of the env transmembrane protein of different strains of HTLV-I and HTLV-II, and selecting, at each position (i) the consensus amino acid at that position, or (ii) if
10 the amino acids at that position are different among the HTLV-I and HTLV-II sequences, all of the amino acid variations at that position. For example, the 2B3A HTLV composite sequence shown in Fig. 5 contains amino acid variations within the amino acid sequences of the
15 2B3A peptide for three different HTLV-I strains (SEQ ID NOS: 26, 27, and 28), a corresponding HTLV-II 2B3A peptide (SEQ ID NO: 29), and the consensus sequence for the 2B3A peptide (SEQ ID NO: 1).

20 "HTLV-specific" peptide means a peptide having an HTLV composite sequence, i.e., either an HTLV-I amino acid sequence, and HTLV-II amino acid sequence, or one of the HTLV composite sequences.

25 "Non-infected individuals" refers to humans whose serum may contain antibodies which are crossreactive with some HTLV-I or HTLV-II proteins, e.g., p21E protein, but whose serum shows no evidence of HTLV-I or HTLV-II infection, as judged by 1.> Failure to detect antibodies to both HTLV gag and env proteins in a confirmatory assay and/or 2.> the failure to detect
30 HTLV-specific sequences by sequence amplification with HTLV-specific primers using the polymerase chain reaction (PCR) method (Kwok et al., Lipka et al. 1991).

35 II. HTLV-I and HTLV-II Env Peptides

Fig. 1, upper frame, represents a portion of an HTLV-I genome which is expressed in the env

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transmembrane glycoproteins gp46 and gp21. The coding region extends from basepair 5180 to 6116 for gp46, and between 6117 and 6644 for the gp21 protein (expanded portion of the genome in Fig. 1).

5 The coding region between bases 6096 and 6497 encodes a recombinant protein, designated p21E, which is known to react with sera from HTLV-I or HTLV-II infected individuals, but which is also reactive with a small percentage of individuals who are not infected with either HTLV virus (Lal et al., Lipka et al., 10 1991). Contained within the p21E coding region, and shown at the lower right in the figure, are coding regions for three HTLV-I peptides identified herein as 2A2B, 2B3A, and 3A3B. The present invention includes the 2B3A peptide, and a diagnostic assay kit and method 15 which includes the 2B3A peptide. The kit and method may also include the 3A3B peptide. The properties of the 2B3A and 3A3B peptides which are related to the assay kit and method for detecting to the HTLV-I or HTLV-II infected human sera are discussed below. 20

 The coding region 5565 to 5895, shown at the lower left in the figure contains coding regions for three MTA peptides described in co-pending U.S. patent application for "HTLV-I and HTLV-II Peptide Antigens and Methods", Serial No. 07/653,091, filed February 08, 25 1991, and identified in the co-pending application and herein as MTA-1, MTA-4, and MTA-5. These peptides may also be used, in combination with the 2B3A peptide, or the 2B3A peptide plus the 3A3B peptide, in an assay kit and method for assaying for HTLV-I or HTLV-II infected 30 human sera. The properties of the MTA peptides which are pertinent to the use in such a kit and method are given below.

35 A. 2B3A Peptide

 Fig. 4 shows the amino acid sequences of a 2B3A peptide from HTLV-I (SEQ ID NO: 22). As will be seen

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below, the peptide was expressed by PCR amplification of an HTLV-I, strain MT2 virus, using primers (which contribute 7 codons on each end of the amplified coding region) designed from the nucleotide coding sequence of the HTLV-I variant ATK isolated by Seiki et al. Thus, the seven amino acids at either end of the peptide correspond in sequence to the 2B3A peptide from the HTLV-I strain ATK and the remaining interior amino acids correspond in sequence to the 2B3A peptide from HTLV-I, strain MT2. The single-letter and triple-letter amino acid codes in the figure conform to standard convention (e.g., Maniatis).

According to an important aspect of the invention, and as detailed below, the peptide is (i) immunoreactive with sera from human subjects infected with HTLV-I or HTLV-II, but (ii) not immunoreactive with sera that is immunoreactive with HTLV-1 p21E antigen, but which shows no evidence of HTLV-I or HTLV-II infection, for example, by PCR analysis of viral sequences in the serum sample, as discussed below.

Fig. 5 shows the amino acid sequences of peptides containing the 2B3A for three different HTLV-I strains, identified herein as ATK, MT2, and B41281, and one HTLV-II peptide, identified herein as strain MO. These nucleotide and amino acid sequences of these HTLV variants were obtained from the Genbank sequence data base. The locus names of the strains which can be used to obtain the sequences from the Genbank data base are as follows; HTLV1A = env region of the HTLV-I variant ATK, HTVENVAA = env region of the HTLV-I variant MT2, B41281 = another more divergent HTLV-I isolate, and HL3V2CG = MoT strain of HTLV-II. The figure also shows the amino acid sequence matching between the upper 2B3A sequence and the corresponding regions of the three HTLV-I strains and the one HTLV-II strain. The sequence homology between the uppermost 2B3A peptide and the corresponding portion of the individual-strain

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gp21 protein is shown direct above the amino acid sequence of each strain. The degree of homology is indicated by ":" for identical sequences and a blank for non-matching amino acids residues.

5 The 2B3A sequences (i.e., the portions of the gp21 amino acid sequences corresponding to the HTLV-I peptide antigen 2B3A) are identified in the figure as follows; SEQ ID NO: 26 = ATK variant of HTLV-I; SEQ ID NO: 28 = MT2 variant of HTLV-I, and SEQ ID NO: 28 = the
10 HTLV variant identified by the locus name B41281. The corresponding HTLV-II 2B3A peptide from strain Mo is identified herein by SEQ ID NO: 29. The strains displayed in figure 5 and figure 6 below are representative of the different amino acid sequences
15 obtained from approximately 30 separate variants of HTLV-I, STLV, and HTLV-II (the amino acid and nucleotide sequences of all of the variants can be obtained from the Genbank database).

 The consensus sequence of the five 2B3A sequences, i.e., the HTLV composite sequence for this region, is
20 shown at the bottom of the figure, and is identified herein by SEQ ID NO: 1. This sequence is constructed from the consensus amino acids, where there is complete consensus among the five peptides, and by the known
25 variations in amino acid, at the six positions (X_1 - X_6) where amino acid variations occur. In this sequence X_1 is K or Q, X_2 is L or I, X_3 is K or R, X_4 is I or V, and X_5 is R or C, and X_6 is P or L. SEQ ID NO: 1 thus includes the five sequences identified above as SEQ ID
30 NO: 22, SEQ ID NOS: 26, SEQ ID NO: 28, SEQ ID NO: 28, and SEQ ID NO: 29. It is to be expected that amino acid substitutions other than those specifically included in SEQ ID NO: 1 may also be allowed, so long as they do not substantially affect the
35 immunoreactivity of the 2B3A peptide as described below. More generally, the 2B3A peptide of the invention includes an HTLV-specific peptide consisting

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essentially of the amino acid sequence identified by SEQ ID NO: 1, where the peptide is characterized by:

- (i) immunoreactivity to sera from human subjects infected with HTLV-I or HTLV-II, and
- (ii) non-immunoreactivity with sera that is (1) immunoreactive with HTLV-1 p21E antigen, but (2) obtained from humans who are not infected with HTLV-I or HTLV-II.

10 B. 3A3B Peptide

 The 3A3B peptide from HTLV-I has the upper amino acid sequence shown in Fig. 4, and is identified by SEQ ID NO: 30. The peptide is intended, in combination with the 2B3A peptide, for use in an assay kit and method for screening human sera for HTLV-I or HTLV-II infection.

 As noted above, sera obtained from patients infected with HTLV-I or HTLV-II contain antibodies which are immunoreactive with the HTLV-I gp21 protein and the p21E recombinant protein derived from gp21. Accordingly, these peptides have been employed routinely in immunoassay methods and kits for detecting HTLV (I or II) infection in humans. The peptides, however, also are crossreactive with antibodies contained in the serum of a certain percentage non-infected individuals, giving false positives in the assay. The frequency of crossreactivity appears to be about 1% of HTLV-negative blood donors, at least in particular locals in the U.S. where statistics are available.

 The 2B3A peptide described above is immunoreactive with sera from HTLV-infected individuals, but is not immunoreactive with non-infected individuals, as discussed above. This peptide provides a useful antigen for detecting HTLV infection without the false positives generated by the p21E protein. It would be further useful, in an HTLV assay, to provide a protein

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which detects non-infected p21E positive individuals, but not HTLV-infected individuals, as a confirmation that the serum tested is non-infected, even though it is cross-reactive with the p21E protein.

5 The 3A3B peptide described herein has the desired properties, in that the peptide is characterized by immunoreactivity with sera that is (1) obtained from humans who are not infected with HTLV-I or HTLV-II, but (2) immunoreactive with HTLV-1 p21E antigen.

10 This peptide was expressed by a coding region sequence by PCR amplification of an HTLV-I, strain MT2 virus, using primers (which contribute 7 codons on each end of the amplified coding region) from the HTLV-I variant ATK. Thus, the seven amino acids at either end of the peptide correspond in sequence to the 3A3B peptide from the HTLV-I variant ATK, and the remaining interior amino acids correspond in sequence to the 3A3B peptide from HTLV-I, strain MT2. The corresponding peptide from HTLV-II, designated herein as SEQ ID NO: 20 34 has amino acid sequence shown at the top in Fig. 6, and is identified herein as SEQ ID NO: 34.

Fig. 6 shows the amino acid sequences of peptides containing the 3A3B for three different HTLV-I strains. The 3A3B sequences are identified in the figure as 25 follows; SEQ ID NOS: 31 = ATK variant of HTLV-I; SEQ ID NO: 32 = MT2 variant of HTLV-I, and SEQ ID NO: 33 = the HTLV variant identified by the locus name HTVENVCH. The corresponding HTLV-II 3A3B peptide from strain Mo is identified herein by SEQ ID NO: 34. These nucleotide and amino acid sequences of the strains were obtained 30 from the Genbank database as described above.

The figure also shows the amino acid sequence matching between the upper 3A3B sequence and the corresponding regions of the three HTLV-I strains and 35 the one HTLV-II strain. The sequence homology between the uppermost 3A3B peptide and the corresponding portion of the individual-strain gp21 protein is shown

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directly above the amino acid sequence of each strain. The degree of homology is indicated by ":" for identical sequences and a blank for non-matching amino acids residues.

5 The consensus sequence of the five 3A3B sequences, i.e., the HTLV composite sequence for this region, is shown at the bottom of the figure, and is identified herein by SEQ ID NO: 2. As above, this sequence is constructed from the consensus amino acids, where there is complete consensus among the five peptides, and by the known variations in amino acid, at the eight positions (X_1 - X_8) where amino acid variations occur. In this sequence X_1 is R or C, X_2 is P or L, X_3 is T or S, X_4 is S or T, X_5 is S or P, X_6 is I, M, or V, X_7 is N or K, and X_8 is L or I. SEQ ID NO: 2 thus includes the five sequences identified above as SEQ ID NO: 30, SEQ ID NOS: 31, SEQ ID NO: 32, SEQ ID NO: 33, and SEQ ID NO: 34. It is anticipated that amino acid substitutions other than those included in SEQ ID NO: 2 may also be allowed, so long as they do not substantially affect the immunoreactivity of the 3A3B peptide as described below.

20 More generally, the 3A3B peptide includes an HTLV-specific peptide consisting essentially of the amino acid sequence identified by SEQ ID NO: 2, where the peptide is characterized by immunoreactivity with sera that is (1) immunoreactive with HTLV-1 p21E antigen, but (2) obtained from humans who are not infected with HTLV-I or HTLV-II.

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C. HTLV-I and HTLV-II Specific Peptides

35 The 2B3A peptide, and the combination of the 2B3A and 3A3B peptides described above are designed for use in an immunoassay method and kit for detecting HTLV-I or HTLV-II infected human sera. The HTLV-I MTA peptides and corresponding gp46 peptides from HTLV-II described in this section are useful in such an assay

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and kit for distinguishing between infection by HTLV-I and infection by HTLV-II infection.

The preparation and properties of the MTA peptides and the corresponding HTLV-II peptides have been described in co-pending U.S. patent application for "HTLV-I and HTLV-II Peptide Antigens and Methods", Serial No. 07/653,091, filed February 08, 1991, and corresponding PCT patent application pct/us92/00823, published August 20, 1992, both of which are incorporated herein by reference. The coding regions of MTA peptides identified as MTA-1, MTA-4, and MTA-5 are given in Fig. 1, and encompass the region between bases 5565 and 5895.

Fig. 9 shows the amino acid sequences of MTA-1 (SEQ ID NO: 3), MTA-4 (SEQ ID NO: 39), and MTA-5 (SEQ ID NO: 40). All three peptides are immunoreactive specifically with HTLV-I infected sera, i.e., are not immunoreactive with HTLV-II infected sera. One smaller HTLV-I peptide, identified in Fig. 10 as K163 (SEQ ID NO: 41) is also immunoreactive specifically with HTLV-I infected sera.

The lower portion of Fig. 9 shows corresponding HTLV-II sequences which are specifically immunoreactive with HTLV-II sera, i.e., don't show immunoreactivity with sera from patients infected with HTLV-I only. These include K15 (SEQ ID NO: 42), K34 (SEQ ID NO: 43), and 4 (SEQ ID NO: 4). The smaller peptide antigen, K34 has been shown to have essentially the same immunoreactivity to HTLV-II infected sera as do the larger peptide antigens K15 and K55.

The HTLV-I and HTLV-II specific peptides described in above may be prepared by the recombinant methods described in the above-cited PCT application PCT/US92/00823.

II. Identification of 2B3A and 3A3B Peptides

The p21E polynucleotide coding sequence,

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identified herein as SEQ ID NO: 5, is shown in Fig. 2, and extending from base 6096 to 6497 was derived from a p21E cloning vector (Samuel et al.). A number of peptides within this region were tested for their immunoreactivity with HTLV-I and HTLV-II infected sera, and with non-infected sera which is immunoreactive with p21E peptide.

A. Preparation of Bacterial Lysates

The peptides which were constructed are shown in Fig. 4. These include recombinant peptides identified as p21E (SEQ ID NO: 6), 1A1B (SEQ ID NO: 19), 2A2B (SEQ ID NO: 20), 2A3B (SEQ ID NO: 21), 2B3A (SEQ ID NO: 22), 3A3B (SEQ ID NO: 30), MF1R2 (SEQ ID NO: 24), AND MF2R1 (SEQ ID NO: 25)

The peptides were prepared by (i) PCR amplification of the selected coding region from the HTLV-I strain MT2, insertion of the amplified coding sequence in a pGEX-GLI expression vector, and transformation of competent *E. coli* host cells with the expression vector, as detailed in Example 1A.

Figs. 3A and 3B show the sequences of the four forward and four reverse primers, respectively, used in construction the peptide coding regions. These include forward primers identified as FP-1A (SEQ ID NO: 9), FP-2A (SEQ ID NO: 10), FP-MF1 (SEQ ID NO: 11), and FP-3A (SEQ ID NO: 13), and the reverse primers identified herein as RP-1B (SEQ ID NO: 14), RP-2B (SEQ ID NO: 15), RP-MR1 (SEQ ID NO: 16), and RP-3B (SEQ ID NO: 18). The 5' ends of the primers are equipped with recognition sequences for one or more of the following restriction enzymes Nco I, BamH I, and EcoR I. The amplified DNAs can then be cut with the appropriate restriction enzymes for ligation into a similarly digested pGEX-GLI expression vector. The cloning site of pGEX-GLI is also presented in Fig 3C.

Each selected peptide coding region is constructed

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by PCR amplification with a selected forward and reverse primer. For example, to construct the coding region of the 1A1B peptide, forward primer FP-1A and reverse primer RP-1B are used to amplify the p21 coding sequence. To construct the coding region of the 2B3A peptide, forward primer MF1 and reverse primer MR1 are employed in PCR amplification of the sequence.

B. Screening of Bacterial Lysates

The recombinant peptides from above were screened in a Western blot assay format described in Example 1C. Briefly, whole cell bacterial lysate transformed with a selected expression vector was fractionated by SDS/PAGE (Laemmli), electroblotted onto nitrocellulose and examined for immunoreactivity with (i) anti-HTLV-I antibodies obtained from EBV-activated lymphocytes from HTLV-I infected individuals (Example 1B), and (ii) HTLV-infected and control sera. Details are given in Example 1C below.

The results are shown in Table 1 below. In the table "ND" means "not done"; "I" indicates sera from an HTLV-I infected individual; "II" indicates sera from an HTLV-I infected individual, where the diagnosis of HTLV-I and HTLV-II was confirmed by PCR; "UnInf" indicates sera from an uninfected individual. "Sup" indicates tissue culture supernatant (diluted 1:2) from a culture of EBV activated peripheral B-cells from an HTLV-I positive donor; and "Ind" indicates sera that are reactive with the recombinant protein p21E, but that are negative for the presence of HTLV-I or HTLV-II infection by PCR using HTLV-I and HTLV-II specific primers and probes.

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Table 1

Sera	Type	p21E	1A1B	2A3B	2A2B	2B3A	3A3B
J254	I	+	+	+	-	+	-
J253	I	+	-	+	-	+	-
J183	I	+	ND	+	-	+	-
J313	I	+	-	+	-	+	-
J103	I	+	-	+	-	+	+
J332	II	+	-	+	-	+	-
J317	II	+	-	+	-	+	+
J309	II	+	-	+	-	+	-
GE9	Sup	+	-	+	-	+	-
5E4	Sup	+	-	+	-	+	-
J376	Uninf	-	-	-	-	-	-
JCO1	Ind	+	ND	+	-	-	+
JCO2	Ind	+	ND	+	-	-	+
JCO3	Ind	+	ND	+	-	-	+
JCO4	Ind	+	ND	+	-	-	+
JCO5	Ind	+	ND	+	-	-	+
JCO6	Ind	+	ND	+	-	-	+
JCO7	Ind	+	ND	+	-	-	+

Overall sera from 10 of the 10 HTLV-I/II infected individuals tested reacted strongly with the relatively large p21E recombinant protein 2A3B. However, only 2 out of the 10 HTLV-I/II antisera tested reacted with either the 2A2B or the 3A3B recombinant proteins. Similarly, when the anti-p21E EBV activated B cell line tissue culture supernatants were tested they both reacted with the recombinant protein 2A3B but not either 2A2B or 3A3B. This suggests that the central portion of 2A3B contains the immunodominant epitope of HTLV-I gp21.

This was confirmed when the recombinant protein 2B3A, which contained 44 amino acids from the central portion of the 2A3B was tested. Ten of the 10 HTLV-I/II sera tested reacted with the 2B3A recombinant protein. In addition, both of the anti-p21E EBV

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I/II sera tested reacted with the 2B3A recombinant protein. In addition, both of the anti-p21E EBV activated B cells tested produced antibodies which recognized the 2B3A.

5 The location of the epitope recognized by sera that react with the p21E recombinant protein, but are negative for HTLV-I or HTLV-II nucleic acids by PCR was also determined. Seven of 7 p21E indeterminate sera reacted with the 2A3B and 3A3B recombinant proteins.
10 None of the 7 p21E indeterminate sera reacted with the 2B3A recombinant protein. Thus the epitopes recognized by sera from HTLV-I infected individuals and individuals who are not infected, but possess an antibody recognizing p21E are differentiatable.

15 With reference again to Fig. 4, it is seen that the 2B3A peptide is divided roughly in half by the MF1R1 and MF2R2 peptides. Preliminary studies conducted in support of the invention indicated that neither peptide is immunoreactive with 2 HTLV-I and 2
20 HTLV-II sera that reacted strongly with the 2B3A peptide. This indicates that the majority of the sequences of the 2B3A peptide are required for immunoreactivity with HTLV positive sera.

25 C. Preparation of Purified Proteins

Recombinant 2A2B, 2B3A, and 3A3B peptides were prepared from bacterial lysates of transformed bacteria, as detailed in Example 1D. Briefly, the cells were grown under conditions which induced the
30 expression of recombinant protein, pelleted by centrifugation, and lysed by several cycles of freezing and thawing. After lysis, proteins were solubilized by addition of Triton-X100™ and insoluble cellular debris was pelleted by centrifugation.

35 The supernatant fraction was passed over a glutathione agarose column, and bound proteins were eluted with 5 mM glutathione. Protein containing

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analysis of the protein fractions.

It will be appreciated that the 2B3A and 3A3B peptides may be prepared directly, by solid-phase peptide synthesis methods, e.g., as described by Mitchell et al., or by alternative recombinant systems.

D. Immunoreactivity of Purified Proteins

The purified 2A2B, 2B3A, and 3A3B peptides from above were examined in a Western blot format for immunoreactivity with 56 sera from HTLV-positive individuals, 7 sera from HTLV negative individuals, and 18 sera from p21E reactive, HTLV indeterminate individuals, as identified in Table 2 below. As seen, the 2B3A peptide was immunoreactive with all of the HTLV-I and HTLV-II positive sera, but was not crossreactive with non-infected sera. By contrast, the 3A3B peptide was reactive with some of the HTLV-positive sera, but reacted with all 18 of the non-infected sera (which were also immunoreactive with the p21E peptide).

Table 2

Sera	p21E Recombinant Proteins		
	2A2B	2B3A	3A3B
HTLV-I Infected PCR +	1/8	26/26	14/26
HTLV-II Infected PCR +	0/8	28/28	19/28
p21E Immunoreactive HTLV PCR (-)	4/16	0/18	18/18
HTLV Negative/p21E Negative	0/1	0/7	1/7

IV. HTLV-I and HTLV-II Diagnostic Method and Kit

Four basic types of diagnostic applications of the 2B3A peptide (defined to include the composite 2B3A peptide) of the invention will be described.

The First general assay type is an enzyme-immuno-assay for screening human sera for HTLV-I or HTLV-II infection. In this assay format, a solid phase reagent

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The First general assay type is an enzyme-immuno-assay for screening human sera for HTLV-I or HTLV-II infection. In this assay format, a solid phase reagent having surface-bound 2B3A peptide is reacted with
5 analyte serum, under conditions which allow antibody binding to the peptide on the reagent. The assay may also utilize additional HTLV peptides, either recombinant or derived from HTLV viral lysate, bound to the solid phase. After washing the reagent to remove
10 unbound serum components, the reagent is reacted with an reporter-labeled anti-human antibody, to bind reporter to the reagent in proportion to the amount of anti-HTLV-I or HTLV-II specific antibody bound to the 2B3A peptide and any other HTLV peptide(s) also bound
15 to the solid support. The reagent is again washed, to remove unbound antibody, and the amount of reporter associated with the reagent is determined. The reporter-labeled antibody, and additional reagents which may be required for reporter detection, are also
20 referred to herein as reporter means for detecting the presence of human antibody bound to the peptide antigen on the solid support.

The second assay type is an enzyme immunoassay for both detecting antibodies to HTLV-I and HTLV-II and
25 differentiating HTLV-I and HTLV-II infected individuals. In this assay format, recombinant peptide antigens capable of specifically detecting antibodies against HTLV-I, or HTLV-II, or both HTLV-I and HTLV-II are attached to a solid phase reagent at different
30 locations. Duplicate samples of the serum to be tested are added to the appropriate regions of the solid phase reagent, after which the assay is preformed essentially as described above.

Fig. 10A illustrates a specific embodiment of a
35 solid phase reagent of the this type. The reagent includes a solid support 10 which forms part of a kit for use in detecting HTLV-I or HTLV-II infection in

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human serum samples. The support has first, second, third, and fourth reaction zones, indicated at 12, 14, 16, and 18, respectively. Reaction zone 12 has surface attached 2B3A peptide molecules. This zone is

5 immunoreactive with antibodies from HTLV-positive serum samples (HTLV-I or HTLV-II infection), but not with "false positives" which are immunoreactive with the p21E peptide but show no evidence of HTLV-I or HTLV-II infection.

10 Reaction zone 14 has surface attached 3A3B peptide molecules. This zone is immunoreactive with antibodies which are crossreactive with the HTLV-I p21E peptide, but in which the serum itself shows no evidence of HTLV-I or HTLV-II infection, e.g., by PCR detection of
15 HTLV-specific sequences.

Reaction zone 16 has surface attached peptide molecules which are immunoreactive specifically with serum antibodies from HTLV-I infected individuals. One preferred HTLV-I peptide is the MTA-1 peptide
20 identified above (SEQ ID NO: 3). A variety of related gp46 peptides, including MTA-4, MTA-5, and K163 (all shown in Fig. 11) may also be employed.

Reaction zone 18 has surface attached peptide molecules which are immunoreactive specifically with
25 serum antibodies from HTLV-II infected individuals. One preferred HTLV-I peptide is the K55 peptide identified above (SEQ ID NO: 4). Other preferred HTLV-II peptides are the K15 and K34 peptides identified above (SEQ ID NO: 4 and SEQ ID NO: 43).

30 The solid surface reagent in the above assay is prepared by known techniques for attaching protein material to solid support material, such as a polymer support or the like. The support may be provided with reactive surface groups, such as amine aldehyde,
35 carboxyl, alcohol, or sulfhydryl groups. The peptide attachment methods generally include non-specific adsorption of the protein to the support, or covalent

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attachment of the protein, typically through a free amine group, to a chemically reactive group on the solid support, such as an activated carboxyl, hydroxyl, or aldehyde group. Methods for attaching peptides to solid support surface, either by non-specific adsorption, or chemical derivatization, are well known.

In a typical assay method, a suitable dilution of a serum sample is placed in contact with each of the four reaction zones in the solid-phase reagent. Generally, the serum sample is placed on each zone in an amount sufficient to cover the zone, e.g., 50-200 μ l serum sample. The serum is incubated with the reagent under conditions sufficient to allow immunoreaction of the serum antibodies with the support-bound peptides. Typically reaction conditions are at 37° C for 30-60 minutes.

Following incubation, the reagent is washed with physiological buffer of the like to remove non-bound and non-specifically bound serum material. To each of the washed zones is then added a drop of detection reagent(s), such as enzyme-labeled anti-human antibody, to bind enzyme to the reagent in proportion to the amount of bound anti-HTLV-I antibody on the solid support. The reagent is again washed, to remove unbound antibody, and the amount of enzyme associated with the reagent is determined.

Fig. 10B illustrates the reaction-zone pattern which may be observed in a serum sample from an HTLV-I infected individual, where the cross-hatching indicates a detectable immunoreaction. In the present case, no reaction with the second zone has occurred, although some HTLV-I samples may be immunoreactive with the 3A3B peptide in the second zone (Table 2 above). The reaction with the third, but not the fourth zone, indicates infection by HTLV-I only.

Fig. 10C illustrates the reaction-zone pattern which may be observed in a serum sample from an HTLV-II

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infected individual. In this case, the serum antibodies are immunoreactive with the 3A3B peptide as well as the 2B3A peptide (Table 2 above). The reaction with the fourth, but not the third zone, indicates infection by HTLV-II only.

Finally, Fig. 10D illustrates the reaction-zone pattern which may be observed in a serum sample from a non-infected, but cross-reactive serum. The absence of reaction with the first zone indicates no infection by HTLV-I or HTLV-II, even though the serum contains a cross-reactive antibody which recognizes the 3A3B peptide. The absence of reaction with either the third or fourth zones further confirms the absence of HTLV-I or HTLV-II infection in the test serum.

A third general assay type is a Western blot assay for use in confirming HTLV-I or HTLV-II antisera. This assay format includes, in addition to one of the gp21 peptide antigens described in this invention, one or more gp46 recombinant peptides, described in U.S. Patent No. _____ and PCT patent application PCT/US92/00823 that are effective at detecting and differentiating serum antibodies to HTLV-I and HTLV-II. In one preferred format, the confirmatory peptides include the p24 gag protein from HTLV-I viral lysate and the p21E recombinant envelope protein containing a large portion of the HTLV-I gp21 envelope protein. The HTLV-I viral lysate detects almost all sera containing antibodies to HTLV-I and/or HTLV-II gag proteins. Antibodies to the HTLV-I and HTLV-II env region are detected by the p21E recombinant protein, however, some sera from uninfected individuals will also react with the p21E protein (Lal, et al.; Lipka, et al., 1991). The sera is diagnosed as being infected with HTLV-I or HTLV-II by the reactivity it displays towards the HTLV-I gp46 peptide antigen MTA1 and the HTLV-II gp46 peptide antigen K55. Thus, if a particular infected serum reacts with MTA1 and not K55 the individual is

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infected with HTLV-I. If the reverse is true the individual can be diagnosed as being infected with HTLV-II. Evaluations of this Western blot confirmatory assay have been reported by the applicants and co-workers (Roberts, et al.; Lipka, et al., 1992b). In these studies the described assay had a specificity of 99% and a sensitivity of 99% for detecting HTLV-I and HTLV-II infected individuals. Details of the blot procedure are given in Example 2B and in the cited publications.

In another embodiment of the Western blot assay the p21E recombinant protein is replaced by the gp21 peptide 2B3A described in this invention. In this format either the HTLV-I or HTLV-II version of the 2B3A peptide detects antibodies to either the HTLV-I or HTLV-II gp21 protein. Due to the lack of reactivity of the 2B3A peptide with sera from uninfected individuals who crossreact with the p21E recombinant protein, this assay would be expected to have a much greater specificity for HTLV infected individuals with essentially the same sensitivity currently exhibited by tests using the p21E recombinant protein.

A final embodiment might include both the 2B3A and 3A3B peptides. Truly HTLV infected individuals would react with 2B3A and possibly the 3A3B peptides. Individuals who are not infected with HTLV-I or HTLV-II but who possess antibodies which cross react with the p21E recombinant protein, and so may be positive on HTLV screening assays, will react only with the 3A3B protein. This embodiment would have the advantage of providing an antigen which will give a positive signal for both the truly HTLV infected individuals and the uninfected, HTLV indeterminate individuals. This allows for a more precise determination of a given individual's seroreactivity.

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V. HTLV-Peptide Vaccine Compositions

Also included in the invention is a vaccine composition containing the 2P3A peptide immunogenic peptide carrier to which the peptide is bound. More specifically, the vaccine contains a peptide having an HTLV-specific antigenic region consisting essentially of the amino acid sequence identified by SEQ ID NO: 22, in combination with a an immunogenic peptide carrier.

Particularly useful immunogenic carriers for the peptide(s) include keyhole limpet hemocyanin (KLH), tetanus toxoid, poly-L-(Lys:Glu), peanut agglutinin, poly-D-lysine, diphtheria toxoid, ovalbumin, soybean agglutinin, bovine serum albumin (BSA), human serum albumin, and the like.

The 2B3A peptide may be conjugated to the carrier by a variety of known methods, including chemical derivatization and by genetic engineering techniques. Such latter technique is disclosed in more detail by Gerald Quinnan, "Proceedings of a Workshop," November 13-14, 1984. Vaccines and inocula of the present invention may be administered by injection, usually intramuscularly or subcutaneously, orally by means of an enteric capsule or tablet, as a suppository, as a nasal spray, and by other suitable routes of administration. For a human patient, a suitable dose of the polypeptide depends, in part, upon the chosen route of administration and a number of other factors. Included among those factors are the body weight of the mammal to be immunized, the carrier when used, the adjuvant when used, and the number of inoculations desired to be used.

Individual inoculations for a human patient typically contain unit doses of about 10 micrograms to about 100 milligrams of polypeptide, exclusive of any carrier to which the polypeptide may be linked. If desired, a series of doses may be administered over a period of time for optimum immunity. Unit dosage forms

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of the vaccine can also be provided, if desired, containing the aforementioned amounts of the polypeptide.

5 In any event, the immunogen contained in a vaccine or an inoculum is present in an "effective amount," which amount depends upon a variety of factors as is well known in the immunological arts, e.g., the body weight of the mammal to be immunized, the carrier moiety used, the adjuvant used, the duration of protection
10 sought, and the desired immunization protocol.

VI. Anti-2B3A Peptide Antibodies

This section describes the preparation of human monoclonal antibodies (Mabs) specific against the 2B3A
15 peptide, the preparation of human recombinant antibodies (Rabs) specific against the 2B3A peptide, and uses of the antibodies as a passive vaccine against HTLV-I viral infection.

20 A. Preparation of Human Anti-2B3A Mabs

Hybridomas which produce anti-2B3A antibodies can be generated by fusing an anti-2B3A antibody-producing lymphocyte isolated from B lymphocytes from an HTLV-I or HTLV-II infected human, with a fusion partner
25 myeloma cell, using hybridoma production techniques known in the art (Harlow).

In one exemplary method of hybridoma production, B-lymphocytes isolated from the peripheral blood of an asymptomatic HTLV-I infected individual are activated
30 with Epstein-Barr virus (EBV), and subsequently selected for antibodies which are immunoreactive with the 2B3A peptides, as described in Example 1B.

Culture of cells showing positive anti-2B3A activity are expanded and fused with a suitable human
35 or mouse-human myeloma fusion partner, such as the GLI-H7 myeloma partner cells described in the Materials section below, using polyethylene glycol (PEG) as a

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fusogen. Hybridomas are then selected by growth in a medium containing hypoxanthine, aminopterin, thymidine and ouabain, according to well-established criteria (Mitchell).

5 Supernatants of hybridoma cultures are tested for presence of immunoglobulins which are immunoreactive with the 2B3A peptide, for example using methods described in Example 2.

10 Positive hybridoma cultures identified by the above steps are subcloned by limiting dilution and re-tested for immunoreactivity with the 2B3A peptide. Positive subclones are expanded and are further tested for immunoglobulin isotype and for 2B3A peptide immunoreactivity.

15

B. Preparation of Human Recombinant Anti-2B3A Antibodies

Cultures of hybridomas which produce anti-2B3A
20 Mabs are prepared as above. Messenger RNA (mRNA) is isolated from the cells, and the mRNA is used to produce corresponding cDNA's according to well-known methods (Maniatis).

25 The coding sequences for the light-chain and heavy-chain variable regions of the immunoglobulin genes are amplified by PCR methods employing known PCR primers for heavy and light chain IgG variable regions (Larrick, 1989, 1991, 1992). The amplified coding sequence fragment for the light-chain variable region is purified, cut between appropriate restriction
30 enzymes, and inserted into a suitable expression vector for expression of IgG light chain, following published procedures (Larrick, 1989, 1991, 1992). The construction of one suitable expression vector, identified herein as pSXR.D.kappa-IgG is given in
35 Example 3.

The amplified coding sequence fragment for the heavy-chain variable region is similarly purified, cut appropriate restriction enzymes, and inserted into a

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suitable expression vector for expression of IgG heavy chain, following published procedures (Larrick, 1989, 1991). One suitable expression vector for expressing the heavy-chain variable region is the pcDNA1/neo.IgG1 vector. The vector can be constructed by modifying a pcDNA1 vector (Invitrogen, San Diego, CA) by addition of a IgG heavy-chain constant coding region (Larrick, 1992). The two plasmids are then cotransfected into either CHO or GLI-H7 cells using lipofection or electroporation. Selection for the heavy plasmid was carried out using antibiotic GENETICIN™ (G418, BRL #860-1811I). Cells are assayed for IgG production and subcloned. The subclones are assayed for binding to the 2B3A peptide. Positive clones were subcloned several times to guarantee purity.

Alternatively, a clone selection method, such as has been described (Huse, McCafferty) may be used to generate recombinant anti-2B3A antibodies, employing as a source of B lymphocyte variable-region coding regions, cDNAs prepared from B lymphocytes isolated from a human infected with HTLV-I or HTLV-II.

The cDNA's are inserted into phage vectors, the vectors are expressed in a suitable *E. coli* bacterial host, and the released phage are selected by affinity binding to a solid support containing the antigen. The captured phage are then used to reinfect a bacterial host.

C. Passive Vaccine Compositions

The human anti-2B3A Mabs or Rabs prepared as above are employed in a vaccine composition for use in the treatment and/or prophylaxis of HTLV-I and/or HTLV-II infection. In this approach Mabs or Rabs which recognize the 2B3A peptide are administered to individuals who may have been exposed to and/or infected with HTLV-I or HTLV-II. Binding of these antibodies to HTLV-I infected cells then provides a

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humoral immune response in which macrophages destroy B lymphocytes having bound anti HTLV-I antibodies.

The anti-HTLV-I antibodies are formulated in a suitable solution for injection, typically by
5 parenteral route, to form the vaccine composition. The composition is administered in an amount effective to block HTLV infection, in an uninfected individual, or to inhibit viral replication in an infected patient. Preferred antibody dosage is in the range between about
10 0.5 to 5 mg antibody/kg body weight. The composition may be administered at spaced intervals, preferably about 1 to 4 week intervals, for treatment of an infected individual. Vaccination may be prior to an expected infection, or in treating existing HTLV-I
15 infection.

In one general application, infants whose mothers are diagnosed as having HTLV-I are injected with the antibody composition, to prevent development of the viral infection, particularly when the infant is breast
20 fed over an extended period. The antibodies may be administered parenterally, e.g., intramuscularly, subcutaneously, or intravenously, or in the case of infants, also by oral administration, in a method for treating or preventing HTLV-I by immune prophylaxis.

25

VII. Identification of 3A3B Crossreactive Protein(s)

The identification of HTLV epitope 3A3B which is specifically crossreactive with HTLV-noninfected humans can be used, according to procedures described in this
30 section, for identifying the protein(s) which are responsible for inducing the cross-reactive antibody.

A. Antibody Identification of Protein

Polyclonal or monoclonal antibodies specific
35 against the 3A3B peptide may be employed to isolate and identify cross-reactive peptides present in the serum of cross-reactive individuals, i.e., individuals who

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are negative for the presence of HTLV virus, but whose serum is cross-reactive with the HTLV p21E antigen.

5 The anti-3A3B antibody may be a polyclonal or monoclonal antibody made according to conventional techniques, using the 3A3B peptide as antigen (Harlow). The peptide antigen is preferably conjugated to a suitable carrier protein, such as keyhole limpet hemocyanin, and injected into a suitable animal, such as a rabbit, for generating polyclonal antibodies, or a
10 mouse, or producing Mabs.

Alternatively, the antibody may be purified by affinity purification by attaching the 3A3B peptide to a solid support, and capturing antibodies from cross-reactive individuals, employing known affinity
15 purification methods (Harlow).

To isolate the cross-reactive protein from a cross-reactive individual, anti-3A3B antibody from above is attached to a solid support, by standard derivatization methods, and a sample of plasma or whole
20 cell lysate isolated from the 3A3B crossreactive individual is passed over the support bed, to entrap cross-reactive proteins on the support. The trapped proteins can be released from the column, candidate proteins can be purified by preparative SDS-PAGE or
25 Chromatography, and the purified proteins can be sequenced using standard methods.

Once a partial protein sequence is obtained, it is possible to identify the coding sequence responsible for the peptide using methods well known to the art
30 (Maniatis et al., SISPA patent). One such method would involve isolating nucleic acids from a serum and/or PBMC sample from a 3A3B crossreactive individual followed by construction of a λ gt10 library. The resulting library would be probed with labeled
35 degenerate primers to identify clones containing hybridizing sequences in the serum which may encode the isolated 3A3B crossreactive protein. Other

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methodologies, which are beyond the scope of this application may also be employed.

5 The following examples illustrate various aspects of the invention, but are in no way intended to limit the scope thereof.

Materials

10 The materials used in the following examples were as follows:

Enzymes: DNAase I and alkaline phosphatase were obtained by Boehringer Mannheim Biochemicals (BMB, Indianapolis, IN); EcoRI, EcoRI methylase, DNA ligase, and Polymerase I, from New England Biolabs (NEB, Beverly, MA); and RNase was obtained from Sigma (St. Louis, MO).

15 Other reagents: EcoRI linkers were obtained from NEB; and nitro blue tetrazolium (NBT), 5-bromo-4-chloro-3-indolyl phosphate (BCIP), 5-bromo-4-chloro-3-indolyl- β -D-galactopyranoside (X-gal) and isopropyl β -D-thiogalactopyranoside (IPTG) were obtained from Sigma.

20 The GLI-H7 cell line is a human-mouse heteromyeloma fusion partner cell line, constructed from fusion of a mouse NS-1 myeloma with a human activated B lymphocyte, as described by Carroll. The HEp-2 cell line is an epidermoid carcinoma (from human larynx) cell line, obtained from the American Type Culture Collection (ATCC), Rockville, MD (ATCC-CCL-23).
30 Chinese Hamster Ovary cells (DUX-B11) were obtained from L.A. Chasin (Urlaub).

Cell culture media (PFHM, RPMI, IMDM) were obtained from Gibco Laboratories.

35

Example 1

Preparation of 2B3A and 3A3B Recombinant Peptides

A. Constructing Expression Vectors

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A plasmid containing a full-copy DNA insert derived from the HTLV-I genome was obtained from Drs. R.C. Gallo and F. Wong-Staal of the Laboratory of Tumor Cell Biology, National Institutes of Health (Bethesda, MD).

The gp21 coding region (Fig. 2) was derived from the gp21 coding region present in the HTLV-I clone sp65 MT-2 obtained from Dr. Wong-Stahl. Oligonucleotide primers designed to amplify selected portions of HTLV-I gp21 (Figure 3) were synthesized on an automated synthesizer (Applied Biosystems, Foster City, CA), following manufacturers instructions. All of the primers contained either a BamHI, NcoI and/or EcoR I site located at their 5' ends to facilitate cloning of the amplified DNAs as in-frame insertion into the pGEX-GLI expression system.

PCR was performed according the manufacturer's instructions (Perkin-Elmer/Cetus, Norwalk, CT) and all PCR reactions contained 2 ng of the HTLV-I clone sp65 MT-2 (generously provided by Dr. F. Wong-Staal) as template and 1.0 μ M of the appropriate oligonucleotide primers. PCR amplification was carried out for 25 cycles of template denaturation (1 minute at 94°C), primer annealing (2 minutes at 50°C), and primer extension (2 minutes at 72°C). Amplified DNAs were purified, digested for 2 hours with the appropriate restriction enzymes and ligated into the expression vector pGEX-GLI, which is a modified version of the commercially available vector pGEX-2 (Pharmacia, Piscataway, NJ), which had previously digested with the same restriction enzymes.

B. Preparing Antibodies for Screening

Peripheral B cells were isolated from an asymptomatic HTLV-I infected individual and activated with Epstein-Barr virus (EBV) at 10^4 cells per well in a 96 well microtiter plate as previously described

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(Perkins; Fong). After 2 days in culture specific anti-HTLV-I IgG activity was assessed using an HTLV-I viral lysate-based enzyme immunoassay (Diagnostic Biotechnology, Singapore). Anti HTLV-I activity was detected, and the EMB immortalized B cells were then grown in culture for about 1 month during which time the spent supernatants from the cultures were saved. Subsequent Western blot analysis using a HTLV-I confirmatory assay (Diagnostic Biotechnology) determined that 3 of the EBV activated B cell lines, designated 3E9, 5G4, and 6E9, reacted strongly with the recombinant env protein p21E. The supernatants were then diluted 1/2 in BLOTTO and used to screen the isolated gp21 recombinant proteins. Upon subsequent fusion to mouse-human heteromyeloma cells, none of these 3 EBV activated B cell lines were successfully fused and the antibody production of the EBV activated B cells eventually ceased. However, the activated B cell supernatants did provide a highly specific antibody preparation which was useful in the screening procedure described below. In addition, the isolation of antibodies to the 2B3A from 3 separate EBV activated B cells confirms that antibodies to the 2B3A peptide antigen are a major component of the immune response of infected individuals to HTLV-I or HTLV-II.

C. Screening Immunogenic Peptides

Plasmid containing bacteria were screened for protein production by Western blot analysis of crude lysates prepared from 2 ml cultures of the transformed *E. coli*. One-tenth volume of the whole cell lysate was loaded per lane and was electrophoresed on a 12.0% polyacrylamide SDS gel (Laemmli). The resulting gel was electroblotted onto nitrocellulose filter paper (Schleicher and Schuell, Keene, NH) and the HTLV-I Western blots were incubated overnight at room temperature with EBV activated B cell tissue culture

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supernatants diluted 1/2 (Example 1B), or HTLV-infected or control antisera diluted 1/100. All sera and supernatants were diluted into BLOTTO (10 mM TRIS-HCl, pH 7.4, 5% nonfat dry milk, 2.5% normal goat sera, and 0.5% Tween-20). The Western blots were washed 3 times with TTBS wash buffer (10 mM TRIS-HCl pH 7.4, 150 mM NaCl, 0.05% Tween-20) for 5 minutes each.

Bound human IgG was detected by a 1 hour incubation with goat-anti-human IgG conjugated to alkaline phosphatase (Promega, Madison, WI) for 1 hour. This was followed by 4,5 minute rinses with TTBS. Bound second antibody was detected by incubating the strips in a substrate solution containing 5-bromo-4-chloro 3-indolylphosphate (BCIP) and Nitroblue tetrazolium (NBT) in 100 mM TRIS-HCl, pH 9.5, and 50 mM MgCl₂. The resulting Western blots were screened with sera from HTLV-I infected or uninfected individuals. The results of these analyses are presented in Table 1.

The recombinant clones 2A3B, 2A2B, 2B3A, and 3A3B were all DNA sequenced by the dideoxy termination procedure (Maniatis et al.). The sequence of the DNA inserts obtained was consistent with the DNA sequences of the primers and templates used in their construction, and would allow for the production of the desired recombinant proteins.

D. Purification of Recombinant Antigens

Purification of recombinant fusion protein was performed essentially as described (20). Briefly, a 10 ml overnight culture of bacteria containing the recombinant plasmid of interest was diluted 1/100 into flasks containing 500 mls NZYDT media (Maniatis, et al.) with 100 µg/ml ampicillin. Expression of fusion protein was induced by the addition of IPTG (final concentration 0.2 mM) to log-phase cultures. The cultures were grown for an addition 3 to 4 hours at

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37°C at which point the bacteria were pelleted by centrifugation at 5000 $\times$ g for 10 minutes. The cells were resuspended in 20 mls cold MTBS and were lysed by several cycles of freezing and thawing. After lysis
5 proteins were solubilized by the addition of Triton-X100 (Sigma, St. Louis, MO) to 1.0%, DNase I to 1 μ g/ml, and Aprotinin to 1.0%. After incubation for 5 minutes at 25°C, insoluble cellular debris was pelleted by centrifugation 2 times at 10,000 $\times$ g for 10 minutes
10 and the supernatants were reserved. Aliquots from both the pellet and supernatant fraction were analyzed by SDS-PAGE (Laemmli) to determine if the recombinant proteins were solubilized by the above procedure.

The 2A3B, 2A2B, and 3A3B recombinant proteins were
15 all present in the soluble fraction. For all 3 of the recombinant proteins a 1 litre culture resulted in the purification of 1-2 mgs of fusion protein at a purity of approximately 50%. The supernatants were then passed through a column containing 0.8 ml of
20 glutathione agarose (Pharmacia, Piscataway, NJ), which was pre-treated as recommended by the manufacturer. The column was washed with 10 mls of MTBS plus 1% Triton and 1% Apoprotinin followed by a 5 ml wash with MTBS alone. Bound proteins were eluted with buffer
25 containing 5 mM glutathione in 50 mM Tris pH 8.0 and 10, 1 ml fractions were collected. The location of the peak of eluted protein was determined by measuring the absorbance at 280 nm of the fractions and by SDS-PAGE analysis of aliquots of the fractions. Fractions
30 containing significant amounts of protein were pooled and aliquots of this pool were frozen at -70°C for subsequent analysis.

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Example 2Serological Paneling of the Purified p21E Recombinants
vs. HTLV-I, HTLV-II and p21E Indeterminate SeraA. Antisera

5 The antisera used in these analyses included a
well-characterized panel of sera from 26 HTLV-I, and 28
HTLV-II infected individuals (Lipka et al. 1991;
Hadlock et al. 1992). All of the HTLV-I and HTLV-II
sera had antibody profiles meeting standard criteria
10 for HTLV infection (antibodies to p24 gag and gp46
and/or gp68 env proteins). In addition, the sera were
typed as being HTLV-I infected both by virtue of their
positive reactivity towards the recombinant HTLV-I
antigen MTA1 and/or through PCR using HTLV-I specific
15 oligonucleotide primers and probes (Lipka, et al.
1992b). Similarly, the 32 HTLV-II infected individuals
were identified by their reactivity towards the
recombinant HTLV-II antigen K55 and/or by PCR using
HTLV-II specific primers and probes (Lipka et al.
20 1992b).

 The gp21 peptide antigens were also tested for
their immunoreactivity with sera from 7 HTLV-I negative
individuals and sera from 18 individuals whose sera
were reactive with the p21E recombinant protein but who
25 were negative for the presence of HTLV nucleic acids
when tested by PCR using HTLV-I and HTLV-I specific
primers and probes. In addition these 18 p21E reactive
sera did not meet serological criteria for being HTLV
infected. The HTLV-I and HTLV-I infected individuals
30 were all from the Northern California area, with the
exception of antisera J103 which was from a HTLV-I
infected Japanese blood donor. The uninfected sera
were all derived from blood donors to the Stanford
University Blood Bank. The 18 p21E reactive-HTLV
35 negative sera were either from blood donors from the
Northern California area or were provided by the Center
for Disease Control, Atlanta, Georgia.

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B. Seroreactivity of Purified Recombinant Peptides

The recombinant proteins 2A2B, 2B3A, and 3A3B were prepared as in Example 1. Aliquots of the purified proteins were separated under reducing conditions on a 11.5% polyacrylamide gel (Laemmli). The resolved proteins were electroblotted onto a nitrocellulose membrane, blocked with BLOTTO, air dried, and cut into 3 mm wide strips.

In the assay, the test strips from above were first rehydrated in TTBS buffer, and the strips were incubated overnight with human test sera, diluted 1:50 in BLOTTO. The strips were washed several times with wash buffer, then incubated for one hour with goat anti-human IgG conjugated to alkaline phosphatase (Bio-Rad, Hercules, CA). After washing, color development was achieved by incubating the strips in a substrate solution containing NBT and BCIP in 100 mM Tris-HCl buffer, pH 9.5, 50 mM MgCl₂. Color development was continued until a uniform background developed on the strip and was halted by rinsing the strips two times with de-ionized water. The recombinant proteins 2A2B, 2B3A, and 3A3B were tested against the panels of HTLV infected and negative sera described in example 2A. The results of the immunoassays are given in Table 2 above.

Example 3

Construction of pSXR.D.kappa-Ig

Construction of the mammalian expression vector pSXR.D.kappa-Ig was performed in several steps. The base vector is pUC18. Into the BamHI site of pUC18 was inserted the 585 bp BamHI-BGLII fragment from the HBV surface antigen gene comprising the polyadenylation signal (Larrick, et al., 1992). The orientation was such that the BamHI site of the HBV insert was nearest to the EcoRI site of the vector. The region between

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the HindIII and HincII sites of the polylinker were removed by digesting the plasmid with those two enzymes and then blunting the HindIII site using the Klenow fragment of DNA polymerase. After ligation, the
5 resultant plasmid was designated pUCHBV3'. A unique SalI site was inserted into the BamHI site using a synthetic oligonucleotide primer.

Between the unique EcoRI site and the SalI site were inserted the following fragments in a series of
10 ligations: (a) An SV40 Early promoter, bounded by an EcoRI site (created by the addition of a synthetic oligonucleotide into the PvuII site immediately preceding the SV40 promoter and the HindIII site immediately preceding the T antigen initiation codon,
15 (b) a stuffer fragment derived from an irrelevant cDNA bounded by a HindIII site and ending in an XbaI site, (c) an XbaI-BglII fragment from pcDNA1/neo containing the SV40 polyadenylation signal and the RSV promoter. The XbaI site is provided in the vector and the BglII
20 site was inserted using PCR cloning to be situated at the junction between the RSV promoter and the Neo selectable marker, and (d) the murine DHRF cDNA, bounded by a BglII site at the 3' end. The restriction sites were inserted using PCR.

25

While the invention has been described with reference to particular embodiments, methods of construction, and uses, it will be clear to those in the art that various other uses, formulations, and
30 methods of practice are within the contemplation of the present invention.

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SEQUENCE LISTING

(1) GENERAL INFORMATION:

- 5 (i) APPLICANT: Hadlock, Kenneth G.
Goh, Chin-Joo
Foung, Steven K.H.
- (ii) TITLE OF INVENTION: Method and Assay for HTLV
- 10 (iii) NUMBER OF SEQUENCES: 46
- (iv) CORRESPONDENCE ADDRESS:
(A) ADDRESSEE: Law Offices of Peter Dehlinger
(B) STREET: 350 Cambridge Avenue, Suite 300
15 (C) CITY: Palo Alto
(D) STATE: CA
(E) COUNTRY: USA
(F) ZIP: 94306
- 20 (v) COMPUTER READABLE FORM:
(A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version #1.25
- 25 (vi) CURRENT APPLICATION DATA:
(A) APPLICATION NUMBER: US 08/014,153
(B) FILING DATE: 05-FEB-1993
(C) CLASSIFICATION:
- 30 (vii) PRIOR APPLICATION DATA:
(A) APPLICATION NUMBER: US 07/653,091
(B) FILING DATE: 08-FEB-1991
- 35 (vii) PRIOR APPLICATION DATA:
(A) APPLICATION NUMBER: US 07/366,313
(B) FILING DATE: 13-JUN-1989
- 40 (vii) PRIOR APPLICATION DATA:
(A) APPLICATION NUMBER: US 06/948,270
(B) FILING DATE: 31-DEC-1986
- 45 (viii) ATTORNEY/AGENT INFORMATION:
(A) NAME: Fabian, Gary R.
(B) REGISTRATION NUMBER: 33,875

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(C) REFERENCE/DOCKET NUMBER: 4600-0106

(ix) TELECOMMUNICATION INFORMATION:

(A) TELEPHONE: (415) 324-0880

(B) TELEFAX: (415) 324-0960

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 44 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

(C) INDIVIDUAL ISOLATE: 2B3AC, Fig. 5

(ix) FEATURE:

(A) NAME/KEY: Modified-site

(B) LOCATION: 6

(D) OTHER INFORMATION: /note= "where Xaa is K or Q"

(ix) FEATURE:

(A) NAME/KEY: Modified-site

(B) LOCATION: 8

(D) OTHER INFORMATION: /note= "where Xaa is L or I"

(ix) FEATURE:

(A) NAME/KEY: Modified-site

(B) LOCATION: 10

(D) OTHER INFORMATION: /note= "where Xaa is K or R"

(ix) FEATURE:

(A) NAME/KEY: Modified-site

(B) LOCATION: 11

(D) OTHER INFORMATION: /note= "where Xaa is I or V"

(ix) FEATURE:

(A) NAME/KEY: Modified-site

(B) LOCATION: 36

(D) OTHER INFORMATION: /note= "where Xaa is L or I"

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44

(ix) FEATURE:

- (A) NAME/KEY: Modified-site
 (B) LOCATION: 7
 (D) OTHER INFORMATION: /note= "where Xaa is P or L"

5

(ix) FEATURE:

- (A) NAME/KEY: Modified-site
 (B) LOCATION: 10
 (D) OTHER INFORMATION: /note= "where Xaa is T or S"

10

(ix) FEATURE:

- (A) NAME/KEY: Modified-site
 (B) LOCATION: 12
 (D) OTHER INFORMATION: /note= "where Xaa is S or T"

15

(ix) FEATURE:

- (A) NAME/KEY: Modified-site
 (B) LOCATION: 15
 (D) OTHER INFORMATION: /note= "where Xaa is S or P"

20

(ix) FEATURE:

- (A) NAME/KEY: Modified-site
 (B) LOCATION: 16
 (D) OTHER INFORMATION: /note= "where Xaa is I, M or V"

25

(ix) FEATURE:

- (A) NAME/KEY: Modified-site
 (B) LOCATION: 25
 (D) OTHER INFORMATION: /note= "where Xaa is N or K"

30

(ix) FEATURE:

- (A) NAME/KEY: Modified-site
 (B) LOCATION: 28
 (D) OTHER INFORMATION: /note= "where Xaa is L or I"

35

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Gln Glu Gln Cys Xaa Phe Xaa Asn Ile Xaa Asn Xaa His Val Xaa

40

Xaa

1 5 10 15

Leu Arg Pro Pro Leu Glu Xaa Arg Val Xaa Thr Gly Trp

45

Gly

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45

20

25

30

Leu Asn

5

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

10

(A) LENGTH: 48 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

15

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

20

(C) INDIVIDUAL ISOLATE: MTA-1, Fig. 9

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

25

Asn

Ser Leu Leu Val Asp Ala Pro Gly Tyr Asp Pro Ile Trp Phe Leu

1

5

10

15

30

His

Thr Glu Pro Ser Gln Leu Pro Pro Thr Ala Pro Pro Leu Leu Pro

20

25

30

35

Lys

Ser Asn Leu Asp His Ile Leu Glu Pro Ser Ile Pro Trp Lys Ser

35

40

45

40

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

45

(A) LENGTH: 44 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

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(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

5 (vi) ORIGINAL SOURCE:

(C) INDIVIDUAL ISOLATE: K55 or "4," Fig. 9

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

10

Asp Ala Pro Gly Tyr Asp Pro Leu Trp Phe Ile Thr Ser Glu Pro
Thr
1 5 10 15

15

Gln Pro Pro Pro Thr Ser Pro Pro Leu Val His Asp Ser Asp Leu
Glu
20 25 30

20

His Val Leu Thr Pro Ser Thr Ser Trp Thr Thr Lys
35 40

(2) INFORMATION FOR SEQ ID NO:5:

25

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 427 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

30

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

35

(vi) ORIGINAL SOURCE:

(C) INDIVIDUAL ISOLATE: p21E(I)CS, Fig. 2A

40

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

TCCTTGTCAC CTGTTCCAC CCTAGGATCC CGCTCCCGCC GAGCGGTACC GGTGGCGGTC
60

45

TGGCTTGCT CCGCCCTGGC CATGGGAGCC GGAGTGGCTG GCGGGATTAC CGGCTCCATG

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120

TCCCTCGCCT CAGGAAAGAG CCTCCTACAT GAGGTGGACA AAGATATTTT CCAGTTAACT

180

5

CAAGCAATAG TCAAAAACCA CAAAATCTA CTCAAATTG CGCAGTATGC TGCCAGAAC

240

AGACGAGGCC TTGATCTCCT GTTCTGGGAG CAAGGAGGAT TATGCAAAGC ATTACAAGAA

10

300

CAGTGCCGTT TTCCGAATAT TACCAATTCC CATGTCCCAA TACTACAAGA AAGACCCCCC

360

15

CTTGAGAATC GAGTCCTGAC TGGCTGGGGC CTTRACTGGG ACCTTGGCCT CTCACAGTGG

420

GCTCGAG

427

20

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:

25

(A) LENGTH: 142 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

30

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

35

(C) INDIVIDUAL ISOLATE: p21E(I), Fig. 2A

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

40

Val Ser Leu Ser Pro Val Pro Thr Leu Gly Ser Arg Ser Arg Arg Ala

1

5

10

15

45

Val Pro Val Ala Val Trp Leu Val Ser Ala Leu Ala Met Gly Ala Gly

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20 25 30

5 Leu Ala Gly Gly Ile Thr Gly Ser Met Ser Leu Ala Ser Gly Lys Ser

35 40 45

10 Val Leu His Glu Val Asp Lys Asp Ile Ser Gln Leu Thr Gln Ala Ile

50 55 60

15 Asn Lys Asn His Lys Asn Leu Leu Lys Ile Ala Gln Tyr Ala Ala Gln

65 70 75

80

20 Lys Arg Arg Gly Leu Asp Leu Leu Phe Trp Glu Gln Gly Gly Leu Cys

85 90 95

25 Val Ala Leu Gln Glu Gln Cys Arg Phe Pro Asn Ile Thr Asn Ser His

100 105 110

30 Gly Pro Ile Leu Gln Glu Arg Pro Pro Leu Glu Asn Arg Val Leu Thr

115 120 125

35 Trp Gly Leu Asn Trp Asp Leu Gly Leu Ser Gln Trp Ala Arg

130 135 140

(2) INFORMATION FOR SEQ ID NO:7:

- 40 (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 480 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear
- 45

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(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

5

(vi) ORIGINAL SOURCE:

(C) INDIVIDUAL ISOLATE: p21E(II)CS, Fig. 2B

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

10

TTCCCTCGCT CCCGTACCTC CTCGGCGGAC AAGACGCCGC CGTGCCGTTT CAATAGCAGT
60

15

GTGGCTTGTC TCCGCCCTAG CGGCGGGAAC AGGTATCGCT GGTGGAGTAA CAGGCTCCCT
120ATCTCTGGCT TCCAGTAAAA GCCTTCTCCT CGAGGTGAC AAAGACATCT CCCACCTTAC
180

20

CCAGGCCATA GTCAAAAATC ATCAAAACAT CCTCCGGGTT GCACAGTATG CAGCCCCAAA
240TAGACGAGGA TTAGACCTCC TATTCTGGGA ACAAGGGGGT TTGTGCAAGG CCATACAGGA
300

25

GCAATGTTGC TTCCTCAACA TCAGTAACAC TCATGTATCC GTCCTCCAGG AACGGCCCCC
360

30

TCTTGAAAAA CGTGTCATCA CCGGCTGGGG ACTAAACTGG GATCTTGGAC TGTCCCAATG
420GGCAGGAGAA GCCCTCCAGA CAGGCATAAC CATTCTCGCT CTACTCCTCC TCGTCATATT
480

35

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:

40

(A) LENGTH: 160 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

45

(iii) HYPOTHETICAL: NO

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50

(vi) ORIGINAL SOURCE:

(C) INDIVIDUAL ISOLATE: p21E(II), Fig. 2B

5

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

	Ser	Leu	Ala	Pro	Val	Pro	Pro	Pro	Ala	Thr	Arg	Arg	Arg	Arg	Ala
Val															
10	1				5					10					15
	Pro	Ile	Ala	Val	Trp	Leu	Val	Ser	Ala	Leu	Ala	Ala	Gly	Thr	Gly
Ile															
15					20					25					30
	Ala	Gly	Gly	Val	Thr	Gly	Ser	Leu	Ser	Leu	Ala	Ser	Ser	Lys	Ser
Leu															
20					35					40					45
	Leu	Leu	Glu	Val	Asp	Lys	Asp	Ile	Ser	His	Leu	Thr	Gln	Ala	Ile
Val															
25					50					55					60
	Lys	Asn	His	Gln	Asn	Ile	Leu	Arg	Val	Ala	Gln	Tyr	Ala	Ala	Gln
Asn															
30	65							70					75		
80															
	Arg	Arg	Gly	Leu	Asp	Leu	Leu	Phe	Trp	Glu	Gln	Gly	Gly	Leu	Cys
Lys															
35								85				90			95
	Ala	Ile	Gln	Glu	Gln	Cys	Cys	Phe	Leu	Asn	Ile	Ser	Asn	Thr	His
Val															
40								100				105			110
	Ser	Val	Leu	Gln	Glu	Arg	Pro	Pro	Leu	Glu	Lys	Arg	Val	Ile	Thr
Gly															
45								115				120			125

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Trp Gly Leu Asn Trp Asp Leu Gly Leu Ser Gln Trp Ala Arg Glu
Ala

130

135

140

5

Leu Gln Thr Gly Ile Thr Ile Leu Ala Leu Leu Leu Val Ile
Leu

145

150

155

160

10

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:

15

(A) LENGTH: 39 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

20

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

25

(C) INDIVIDUAL ISOLATE: FP-1A, Fig. 3A

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

30

TCCGAATTCT CCATGGGTTTCT CTTGTCACCT GTTCCCACC

39

(2) INFORMATION FOR SEQ ID NO:10:

35

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 36 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

40

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

45

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(vi) ORIGINAL SOURCE:

(C) INDIVIDUAL ISOLATE: FP-2A, Fig. 3A

5

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

TCCGAATTCG GATCCTGGCT TGTCTCCGCC CTGGCC
36

10

(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:

15

- (A) LENGTH: 36 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

20

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

25

(C) INDIVIDUAL ISOLATE: FP-MF1, Fig. 3A

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

30

GCGAATTCGG ATCCATAGTC AAAAACCACA AAAATC
36

(2) INFORMATION FOR SEQ ID NO:12:

35

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 34 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

40

(ii) MOLECULE TYPE: DNA (genomic)

(i: HYPOTHETICAL: NO

45

(vi) ORIGINAL SOURCE:

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53

(C) INDIVIDUAL ISOLATE: MF2, Fig. 3A

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

5

CCGAATTCGG ATCCCTCCTG TTCTGGGAGC AAGG

34

10

(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 42 base pairs

(B) TYPE: nucleic acid

15

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

20

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

(C) INDIVIDUAL ISOLATE: FP-3A, Fig. 3A

25

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

TCCGAATTCA CTAGTGGATC CCAAGAACAG TGCCGTTTTTC CG

42

30

(2) INFORMATION FOR SEQ ID NO:14:

(i) SEQUENCE CHARACTERISTICS:

35

(A) LENGTH: 48 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

40

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

45

(C) INDIVIDUAL ISOLATE: RP-1B, Fig. 3B

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

ACCACTAGTA CCACCACCAC CGAATTCCAC CGGTACCGCT CGGCGGGA

5

48

(2) INFORMATION FOR SEQ ID NO:15:

10

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 30 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

15

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

20

(vi) ORIGINAL SOURCE:

(C) INDIVIDUAL ISOLATE: RP-2B, Fig. 3B

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

25

TGGGAATTCG TGGTTTTTGA CTATTGCTTG

30

30

(2) INFORMATION FOR SEQ ID NO:16:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 26 base pairs

(B) TYPE: nucleic acid

35

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

40

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

(C) INDIVIDUAL ISOLATE: RP-MR1, Fig. 3B

45

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55

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

CGCGAATTCG GAAAACGGCA CTGTTC

25

5

(2) INFORMATION FOR SEQ ID NO:17:

(i) SEQUENCE CHARACTERISTICS:

10

(A) LENGTH: 31 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

15

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

20

(C) INDIVIDUAL ISOLATE: MR2, Fig. 3B

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

25

CGCGAATTC AGGAGATCAA GGCCTCGTCT G

31

(2) INFORMATION FOR SEQ ID NO:18:

30

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 30 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

35

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

40

(vi) ORIGINAL SOURCE:

(C) INDIVIDUAL ISOLATE: RP-3B, Fig. 3B

45

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

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TGGGAATTCG TTAAGGCCCC AGCCAGTCAG
30

5 (2) INFORMATION FOR SEQ ID NO:19:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 18 amino acids

(B) TYPE: amino acid

10 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

15 (iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

(C) INDIVIDUAL ISOLATE: 1A1B, Fig. 4

20 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

Ser Leu Ser Pro Val Pro Thr Leu Gly Ser Arg Ser Arg Arg Ala
Val

1 5 10 15

25

Pro Val

30 (2) INFORMATION FOR SEQ ID NO:20:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 47 amino acids

(B) TYPE: amino acid

35 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

40

(vi) ORIGINAL SOURCE:

(C) INDIVIDUAL ISOLATE: 2A2B, Fig. 4

45 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

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Trp Leu Val Ser Ala Leu Ala Met Gly Ala Gly Val Ala Gly Gly
Ile
1 5 10 15

5

Thr Gly Ser Met Ser Leu Ala Ser Gly Lys Ser Leu Leu His Glu
Val
20 25 30

10

Asp Lys Asp Ile Ser Gln Leu Thr Gln Ala Ile Val Lys Asn His
35 40 45

15

(2) INFORMATION FOR SEQ ID NO:21:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 86 amino acids

(B) TYPE: amino acid

20

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

25

(vi) ORIGINAL SOURCE:

(C) INDIVIDUAL ISOLATE: 2A3B, Fig. 4

30

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

Trp Leu Val Ser Ala Leu Ala Met Gly Ala Gly Val Ala Gly Gly
Ile
1 5 10 15

35

Thr Gly Ser Met Ser Leu Ala Ser Gly Lys Ser Leu Leu His Glu
Val
20 25 30

40

Asp Lys Asp Ile Ser Gln Leu Thr Gln Ala Ile Val Lys Asn His
Lys
35 40 45

45

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Asn Leu Leu Lys Ile Ala Gln Tyr Ala Ala Gln Asn Arg Arg Gly
Leu

50 55 60

5

Asp Leu Leu Phe Trp Glu Gln Gly Gly Leu Cys Lys Ala Leu Gln
Glu

65 70 75

80

10

Gln Cys Cys Phe Leu Asn

85

15

(2) INFORMATION FOR SEQ ID NO:22:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 44 amino acids

(B) TYPE: amino acid

20

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

25

(vi) ORIGINAL SOURCE:

(C) INDIVIDUAL ISOLATE: 2B3A, Fig. 5

30

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

Ile Val Lys Asn His Lys Asn Leu Leu Lys Ile Ala Gln Tyr Ala
Ala

1 5 10 15

35

Gln Asn Arg Arg Gly Leu Asp Leu Leu Phe Trp Glu Gln Gly Gly
Leu

20 25 30

40

Cys Lys Ala Leu Gln Glu Gln Cys Arg Phe Pro Asn

35

40

45

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(2) INFORMATION FOR SEQ ID NO:23:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 34 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

(C) INDIVIDUAL ISOLATE: 3A3B, Fig. 6

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

Gln Glu Gln Cys Arg Phe Pro Asn Ile Thr Asn Ser His Val Ser
Ile

20 1 5 10 15

Leu Gln Glu Arg Pro Pro Leu Glu Asn Arg Val Leu Thr Gly Trp
Gly

25 20 25 30

Leu Asn

30

(2) INFORMATION FOR SEQ ID NO:24:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 25 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

(C) INDIVIDUAL ISOLATE: MF1R2, Fig. 4

45

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60

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

Ile Val Lys Asn His Lys Asn Leu Leu Lys Ile Ala Gln Tyr Ala
 Ala
 5 1 5 10 15

Gln Asn Arg Arg Gly Leu Asp Leu Leu
 10 20 25

(2) INFORMATION FOR SEQ ID NO:25:

- (i) SEQUENCE CHARACTERISTICS:
 15 (A) LENGTH: 21 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
 20
- (iii) HYPOTHETICAL: NO
- (vi) ORIGINAL SOURCE:
 25 (C) INDIVIDUAL ISOLATE: MF2R1, Fig. 4

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

Leu Leu Phe Trp Glu Gln Gly Gly Leu Cys Lys Ala Leu Gln Glu
 30 Gln
 1 5 10 15

Cys Arg Phe Pro Asn
 35 20

(2) INFORMATION FOR SEQ ID NO:26:

- (i) SEQUENCE CHARACTERISTICS:
 40 (A) LENGTH: 63 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
 45

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(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

(C) INDIVIDUAL ISOLATE: 2B3AS, Fig. 5

5

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

[illegible]

(2) INFORMATION FOR SEQ ID NO:27:

30 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 63 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

35 (ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

40 (C) INDIVIDUAL ISOLATE: 2B3AM, Fig. 5

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

45 His Glu Val Asp Lys Asp Ile Ser Gln Leu Thr Gln Ala Ile Val

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Lys
 1 5 10 15
 5 Asn His Lys Asn Leu Leu Lys Ile Ala Gln Tyr Ala Ala Gln Asn
 Arg
 20 25 30
 10 Arg Gly Leu Asp Leu Leu Phe Trp Glu Gln Gly Gly Leu Cys Lys
 Ala
 35 40 45
 15 Leu Gln Glu Gln Cys Cys Phe Leu Asn Ile Thr Asn Ser His Val
 50 55 60

(2) INFORMATION FOR SEQ ID NO:28:

20

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 63 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

25

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

30

(vi) ORIGINAL SOURCE:

(C) INDIVIDUAL ISOLATE: 2B3AB, Fig. 5

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

35

His Glu Val Asp Lys Asp Ile Ser Gln Leu Thr Gln Ala Ile Val
 Lys
 1 5 10 15

40

Asn His Lys Asn Leu Leu Lys Val Ala Gln Tyr Ala Ala Gln Asn
 Arg
 20 25 30

45

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Arg Gly Leu Asp Leu Leu Phe Trp Glu Gln Gly Gly Leu Cys Lys
Ala

35

40

45

5

Leu Gln Glu Gln Cys Cys Phe Leu Asn Ile Thr Asn Ser His Val
50 55 60

10

(2) INFORMATION FOR SEQ ID NO:29:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 63 amino acids

(B) TYPE: amino acid

15

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

20

(vi) ORIGINAL SOURCE:

(C) INDIVIDUAL ISOLATE: 2B3AMO, Fig. 5

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

25

Lys Asp Ile Ser His Leu Thr Gln Ala Ile Val Lys Asn His Gln
Asn
1 5 10 15

30

Ile Leu Arg Val Ala Gln Tyr Ala Ala Gln Asn Arg Arg Gly Leu
Asp
20 25 30

35

Leu Leu Phe Trp Glu Gln Gly Gly Leu Cys Lys Ala Ile Gln Glu
Gln
35 40 45

40

Cys Cys Phe Leu Asn Ile Ser Asn Thr His Val Ser Val Leu Gln
50 55 60

45

(2) INFORMATION FOR SEQ ID NO:30:

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- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 8 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

5

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

10

(vi) ORIGINAL SOURCE:

(C) INDIVIDUAL ISOLATE: 3A3B, Fig. 4

15

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

Gln Glu Gln Cys Arg Phe Pro Asn
 1 5

20

(2) INFORMATION FOR SEQ ID NO:31:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 53 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

25

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

30

(vi) ORIGINAL SOURCE:

(C) INDIVIDUAL ISOLATE: 3A3BS, Fig. 6

35

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:

Glu Gln Gly Gly Leu Cys Lys Ala Leu Gln Glu Gln Cys Arg Phe
 Pro
 1 5 10 15

40

Asn Ile Thr Asn Ser His Val Pro Ile Leu Gln Glu Arg Pro Pro
 Leu

45

20

25

30

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65

Glu Asn Arg Val Leu Thr Gly Trp Gly Leu Asn Trp Asp Leu Gly
 Leu
 35 40 45

5

Ser Gln Trp Ala Arg
 50

10

(2) INFORMATION FOR SEQ ID NO:32:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 53 amino acids

(B) TYPE: amino acid

15

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

20

(vi) ORIGINAL SOURCE:

(C) INDIVIDUAL ISOLATE: 3A3BM, Fig. 6

25

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:

Glu Gln Gly Gly Leu Cys Lys Ala Leu Gln Glu Gln Cys Cys Phe
 Leu
 1 5 10 15

30

Asn Ile Thr Asn Ser His Val Ser Ile Leu Gln Glu Arg Pro Pro
 Leu
 20 25 30

35

Glu Asn Arg Val Leu Thr Gly Trp Gly Leu Asn Trp Asp Leu Gly
 Leu
 35 40 45

40

Ser Gln Trp Ala Arg
 50

45

(2) INFORMATION FOR SEQ ID NO:33:

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(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 53 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

5.

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

10

(vi) ORIGINAL SOURCE:
(C) INDIVIDUAL ISOLATE: 3A3BCH, Fig. 6

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

15

Glu Gln Gly Gly Leu Cys Lys Ala Leu Gln Glu Gln Cys Cys Phe
Leu
1 5 10 15

20

Asn Ile Thr Asn Ser His Val Ser Met Leu Gln Glu Arg Pro Pro
Leu
20 25 30

25

Glu Asn Arg Val Leu Thr Gly Trp Gly Leu Asn Trp Asp Leu Gly
Leu
35 40 45

30

Ser Gln Trp Ala Arg
50

35

(2) INFORMATION FOR SEQ ID NO:34:

(i) **SEQUENCE CHARACTERISTICS:**

- (A) **LENGTH:** 53 amino acids
- (B) **TYPE:** amino acid
- (D) **TOPOLOGY:** linear

40

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

45

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(vi) ORIGINAL SOURCE:

(C) INDIVIDUAL ISOLATE: 3A3BMO, Fig. 6

5 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:

Leu Cys Lys Ala Ile Gln Glu Gln Cys Cys Phe Leu Asn Ile Ser
Asn

10 1 5 10 15

Thr His Val Ser Val Leu Gln Glu Arg Pro Pro Leu Glu Lys Arg
Val

15 20 25 30

Ile Thr Gly Trp Gly Leu Asn Trp Asp Leu Gly Leu Ser Gln Trp
Ala

20 35 40 45

Arg Glu Ala Leu Gln
50

25

(2) INFORMATION FOR SEQ ID NO:35:

(i) SEQUENCE CHARACTERISTICS:

30 (A) LENGTH: 133 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

35

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

40 (C) INDIVIDUAL ISOLATE: 2B3A(I), Fig. 7

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:

45 CCATAGTCAA AAACCACAAA AATCTACTCA AAATTGCGCA GTATGCTGCC CAGAACAGAC
60

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GAC TTTGA TCTCCTGTTT TGGGAGCAAG GAGGATTATG CAAAGCATTG CAAGAACAGT
1.

5 GCCGTTTTTC GAA
133

(2) INFORMATION FOR SEQ ID NO:36:

10 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 133 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

15 (ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

20 (vi) ORIGINAL SOURCE:
(C) INDIVIDUAL ISOLATE: 2B3A(II), Fig. 7

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:

25 CCATAGTCAA AAATCATCAA AACATCTCTC GGGTTGCACA GTATGCAGCC CAAATAGAC
60

30 GAGGATTAGA CTCCTATTC TGGGAACAAG GGGGTTTGTG CAAGGCCATA CAGGAGCAAT
120

GTTCCTTCCT CAA
133

35

(2) INFORMATION FOR SEQ ID NO:37:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 102 base pairs
40 (B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)
45

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(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

(C) INDIVIDUAL ISOLATE: 3A3B(I), Fig. 8

5

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:

CAAGAACAGT GCCGTTTTCC GAATATCACT AATTCCTATG TCTCAATACT ACAGGAAAGA
60

10

CCCCCCTTG AGAATCGAGT CCTGACTGGC TGGGGCCTTA AC
102

15

(2) INFORMATION FOR SEQ ID NO:38:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 102 base pairs

20

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

25

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

(C) INDIVIDUAL ISOLATE: 3A3B(II), Fig. 8

30

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:

CAGGAGCAAT GTTGCTTCCT CAACATCAGT AACACTCATG TATCCGTCCT CCAGGAACGG
60

35

CCCCCTCTTG AAAAACGTGT CATCACCAGC TGGGGACTAA AC
102

40

(2) INFORMATION FOR SEQ ID NO:39:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 47 amino acids

45

(B) TYPE: amino acid

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(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

5 (iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

(C) INDIVIDUAL ISOLATE: MTA-4, Fig. 9

10

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:39:

Cys Gly Phe Pro Ser Ser Leu Leu Val Asp Ala Pro Gly Tyr Asp
 Pro
 15 1 5 10 15

Ile Trp Phe Leu Asn Thr Glu Pro Ser Gln Leu Pro Pro Thr Ala
 Pro
 20 20 25 30

Pro Leu Leu Pro His Ser Asn Leu Asp His Ile Leu Glu Pro Ser
 35 40 45

25

(2) INFORMATION FOR SEQ ID NO:40:

(i) SEQUENCE CHARACTERISTICS:

30 (A) LENGTH: 77 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

35

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

(C) INDIVIDUAL ISOLATE: MTA-5, Fig. 9

40

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:

45 Ser Leu Leu Val Asp Ala Pro Gly Tyr Asp Pro Ile Trp Phe Leu
 Asn

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		1	5	10	15
5	His	Thr Glu Pro Ser Gln Leu Pro Pro Thr Ala Pro Pro Leu Leu Pro			
		20	25	30	
10	Lys	Ser Asn Leu Asp His Ile Leu Glu Pro Ser Ile Pro Trp Lys Ser			
		35	40	45	
15	Ile	Leu Leu Thr Leu Val Gln Leu Thr Leu Gln Ser Thr Tyr Tyr Cys			
		50	55	60	
20		Val Cys Ile Asp Arg Ala Ser Leu Ser Thr Trp His Val			
		65	70	75	

(2) INFORMATION FOR SEQ ID NO:41:

- 25 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 18 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear
- 30 (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (vi) ORIGINAL SOURCE:
 35 (C) INDIVIDUAL ISOLATE: K163, Fig. 9

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:

40	Glu	Thr Ala Pro Pro Leu Leu Pro His Ser Asn Leu Asp His Ile Leu			
		1	5	10	15
45		Pro Ser			

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(2) INFORMATION FOR SEQ ID NO:42:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 49 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

(C) INDIVIDUAL ISOLATE: K15, Fig. 9

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:42:

Met Thr Leu Leu Val Asp Ala Pro Gly Tyr Asp Pro Leu Trp Phe
Ile
20 1 5 10 15

Thr Ser Glu Pro Thr Gln Pro Pro Pro Thr Ser Pro Pro Leu Val
His
25 20 25 30

Asp Ser Asp Leu Glu His Val Leu Thr Pro Ser Thr Ser Trp Thr
Thr
30 35 40 45

Lys

35

(2) INFORMATION FOR SEQ ID NO:43:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 17 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

45

(iii) HYPOTHETICAL: NO

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(vi) ORIGINAL SOURCE:

(C) INDIVIDUAL ISOLATE: K34, Fig. 9

1 Val Asp Lys

45

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:43:

Ser Pro Pro Leu Val His Asp Ser Asp Leu Glu His Val Leu Thr

1 Lys Asn Leu

Pro

1 5 10 15

Ser

1 Leu Asp Leu

(2) INFORMATION FOR SEQ ID NO:44:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 68 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

1 Glu Gln Cys

95

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

(C) INDIVIDUAL ISOLATE: MODIFIED pGEX, FIG. 3C

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 3..68

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:44:

AT CCT CCA AAA TCG GAT CTG GTT CCG CGT GGT TCC ATG GGT GGA TCC.

47

Pro Pro Lys Ser Asp Leu Val Pro Arg Gly Ser Met Gly Gly Ser

1 5 10 15

GAA TTC ATC GTG ACT GAC TGA

68

Glu Phe Ile Val Thr Asp

20

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IT IS CLAIMED:

1. A peptide having an HTLV-specific antigenic region consisting essentially of the amino acid sequence identified by SEQ ID NO: 1, said peptide being characterized by:

(i) immunoreactivity to sera from human subjects infected with HTLV-I or HTLV-II, and

(ii) non-immunoreactivity with sera that are (1) immunoreactive with HTLV-1 p21E antigen, but (2) obtained from humans who are not infected with HTLV-I or HTLV-II.

Ser Glu
15

2. The peptide of claim 1, wherein the peptide sequence is identified by SEQ ID NO: 22 contained within SEQ ID NO: 1.

3. The peptide of claim 1, wherein the peptide sequence is identified by SEQ ID NO: 29 contained within SEQ ID NO: 1.

4. The peptide of claim 1, for use in a solid-phase assay for detecting HTLV-I or HTLV-II infection in a human serum sample, which is carried on a solid-support.

5. A kit for detecting the presence of HTLV-I or HTLV-II infection in a human serum, comprising

(a) a solid support;

(b) attached to the support, in a first reaction zone, a peptide having an HTLV-specific antigenic region consisting essentially of the amino acid sequence identified by SEQ ID NO: 1, said peptide being characterized by:

1 Trp Leu

15

(i) immunoreactivity to sera from human subjects infected with HTLV-I or HTLV-II, and

e Thr Gly

(ii) non-immunoreactivity with sera that are (1)

30

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immunoreactive with HTLV-1 p21E antigen, but (2) obtained from humans who are not infected with HTLV-I or HTLV-II; and

5 (c) reporter means for detecting the presence of human antibodies bound to said support.

10 6. The kit of claim 5, wherein the peptide sequence is identified by SEQ ID NO: 22 contained with SEQ ID NO: 1.

7. The kit antigen of claim 5, wherein the peptide sequence is identified by SEQ ID NO: 29 contained within SEQ ID NO: 1.

15 8. The kit of claim 5, which further includes a second reaction zone in the solid support, and attached to this second zone, a second HTLV-specific antigenic region consisting essentially of the amino acid sequence identified by SEQ ID NO: 2, and being
20 characterized by immunoreactivity with sera that are (1) obtained from humans who are not infected with HTLV-I or HTLV-II, but (2) immunoreactive with HTLV-1 p21E antigen.

25 9. The kit of claim 5, which further includes a third reaction zone in the solid support, and attached to this third zone, a third HTLV-specific antigen capable of discriminating between serum antibodies specific against HTLV-I and HTLV-II.

30 10. The kit of claim 9, wherein the second peptide contains an HTLV-I specific antigenic region consisting essentially of the sequence identified by SEQ NO ID: 3.

35 11. The kit of claim 9, wherein the second peptide contains an HTLV-II specific antigenic region

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consisting essentially of the sequence identified by
SEQ NO ID: 4.

12. A kit for detecting the presence of HTLV-I or
5 HTLV-II infection in a human serum, comprising

(a) a solid support having first and second, and
third reaction zones,

(b) immobilized at said first reaction zone, a
first peptide having an HTLV-specific antigenic region
10 consisting essentially of the amino acid sequence
identified by SEQ ID NO: 1, said peptide being
characterized by:

(i) immunoreactivity to sera from human subjects
infected with HTLV-I or HTLV-II, and

15 (ii) non-immunoreactivity with sera that are (1)
immunoreactive with HTLV-1 p21E antigen, but (2)
obtained from humans who are not infected with HTLV-I
or HTLV-II;

(c) immobilized at said second zone, a second
20 peptide having an HTLV-specific antigenic region
consisting essentially of the amino acid sequence
identified by SEQ ID NO: 2, and being characterized by
immunoreactivity with sera that are (1) immunoreactive
with HTLV-1 p21E antigen, but (2) obtained from humans
25 who are not infected with HTLV-I or HTLV-II;

(d) immobilized at said third zone, a third HTLV-
specific antigen capable of discriminating between
serum antibodies specific against HTLV-I and HTLV-II;
and

30 (e) reporter means for detecting the presence of
human antibodies bound to said support.

13. The kit of claim 12, wherein said first
peptide sequence is identified by SEQ ID NO: 22
35 contained within SEQ ID NO: 1, and said second peptide
sequence is identified by SEQ ID NO: 30 contained
within SEQ ID NO: 2.

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14. The kit of claim 12, wherein said third peptide has an HTLV-I-specific antigenic region consisting essentially of the amino acid sequence identified by SEQ ID NO: 3, said peptide being characterized by:

(A) immunoreactivity to sera from human subjects infected with HTLV-I, and

(ii) non-immunoreactivity to sera from human subjects infected with HTLV-II only.

15. The kit of claim 12, wherein said third peptide has an HTLV-II-specific antigenic region consisting essentially of the amino acid sequence identified by SEQ ID NO: 4, said peptide being characterized by:

(A) immunoreactivity to sera from human subjects infected with HTLV-II, and

(ii) non-reactivity to sera from human subjects with infected with HTLV-I only.

16. A method of positively identifying HTLV-I or HTLV-II infection in a human subject, comprising reacting serum from the subject with a peptide containing an HTLV-specific antigenic region consisting essentially of the amino acid sequence identified by SEQ ID NO: 1,

by said reacting, forming an immune complex between said peptide and antibodies in serum from a subject having an HTLV-I or HTLV-II infection, and

examining the peptide for the presence of immunocomplex, evidencing the presence of HTLV-I or HTLV-II antibodies in the serum.

17. The method of claim 16, wherein the peptide sequence is identified by SEQ ID NO: 30 contained within SEQ ID NO: 2.

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