

Description

This invention relates generally to hormones and more particularly to glycoprotein hormones including the pituitary glycoproteins, and further provides methods for creating hormone analogs by site-directed mutagenesis.

Glycoprotein hormones, especially those synthesized and secreted by the anterior pituitary gland, play critically important roles in a variety of bodily functions including: metabolism, temperature regulation, growth, and reproduction. The pituitary glycoproteins, luteinizing hormone (LH), follicle stimulating hormone (FSH), and thyroid stimulating hormone (TSH) are similar in structure to the placental gonadotropin, human chorionic gonadotropin (hCG). Each of the molecules is actually a dimer consisting of two protein chains held together by non-covalent, ionic interactions. The alpha chain for each of the hormones is identical. The beta chain is the hormone-specific portion of the dimer.

Following secretion, the hormones travel in the blood stream to the target cells which contain membrane bound receptors. The hormone binds to the receptor and stimulates the cell. Typically, such stimulation involves an increase in activity of a specific intracellular regulatory enzyme which in turn catalyzes a biochemical reaction essential to the response of the cell. In the case of hCG, binding to the hCG receptor present upon the corpus luteum (an ovarian structure), stimulates the activity of the enzyme adenylate cyclase. This enzyme catalyzes the conversion of intracellular ATP to cyclic AMP (cAMP). cAMP stimulates the activity of other enzymes involved in the production of ovarian steroid hormones, especially progesterone. hCG-stimulated progesterone secretion is essential for the maintenance of pregnancy during the first trimester of gestation.

The exact mechanism by which a dimeric glycoprotein hormone, such as hCG, stimulates post-receptor events, such as activation of adenylate cyclase activity is unknown. By a variety of experimental manipulations, it has been shown, however, that the carbohydrate structures each attached to the hCG molecule by a linkage at respective asparagine residues (N-linked), play important roles in this regard. Treatment of glycoprotein hormones, such as LH, FSH, or hCG with hydrogen fluoride removes approximately 70 percent of the oligosaccharide side chains. The resultant partially deglycosylated molecules retain their receptor binding activity but are unable to stimulate any post-receptor events. Thus it is clear that the sugar portion of the glycoprotein hormone, while not directly involved with receptor binding, plays a critical role in post-receptor events, and therefore, biologic activity.

It is also known that in addition to a role in the *in vitro* bioactivity, oligosaccharides are important components of the molecule's survival time in circulation. Indeed, plasma half-life of a glycoprotein hormone is directly related to the amount of one particular sugar molecule, sialic acid, generally present upon the most distal portion of the oligosaccharide chain. The removal of the carbohydrate portions (by hydrogen fluoride treatment) would result in the production of hormones that bind to the receptor but fail to exert the expected biologic response. In addition, these molecules would have an extremely short plasma half-life since the lack of terminal sialic acid residues would increase the binding affinity to the hepatic asialoglycoprotein receptor thereby hastening clearance from the systemic circulation.

Many clinical endocrinopathies are the result of over production of stimulating hormones (e.g., excess TSH secretion resulting in hyperthyroidism). A conventional treatment for a pathologic state caused by a hormone excess would be the administration of a hormone antagonist. To be effective, an antagonist must bind with high affinity to the receptor but fail to activate post receptor events. From the earlier discussion, it can be anticipated that hydrogen fluoride treated hormones (that have had approximately 70 percent of carbohydrates removed) would be effective, competitive antagonists. Indeed, it has been shown that HF-treated hormones bind with somewhat greater affinity for the biologic receptor, compete effectively with native material, and diminish the expected biologic action of native hormone in a dose-dependent fashion.

At first glance, such a hormone preparation would appear to be a viable candidate for a competitive antagonist therapeutic agent. However, four problems are associated with large scale production of such preparations. First, all pituitary hormone preparations are generally contaminated with other hormones. Thus, while it may be possible to obtain a preparation of a hormone and partially deglycosylate it, the resultant preparation would also contain other deglycosylated hormones as contaminants which may disadvantageously produce unwanted and unacceptable side effects following administration. Secondly, preparations of partially purified hormones vary greatly in their potency, physicochemical characteristics and purity. Therefore, each batch produced would need to be analyzed carefully. The possibility of batch-to-batch variability would necessitate repeated execution of clinical trials to determine the effective dose. Thirdly, the method used to deglycosylate is incomplete and, therefore, somewhat uncontrollable. No information is available as to the variability associated with hydrogen fluoride treatment of successive batches of hormone. Potential variability in this process would also require extensive characterization of

each batch produced. Fourth, carbohydrate side chains also play an important role in dictating the plasma half-life of a molecule. Partially deglycosylated hormones have been shown to be rapidly cleared from the circulation following injection. Therefore, repeated injections of large doses of deglycosylated hormones would be required to achieve a desired effect. The need for such large doses of hormone to deglycosylate and administer creates yet another problem, namely availability. Large quantities of hormones are presently unavailable and could conceivably only be made available through recombinant DNA technology.

As mentioned above, deglycosylated hormones, while exhibiting the desired competitive antagonistic properties *in vitro* would be of little therapeutic value *in vivo* due to their extremely short plasma half-life. The only potential solution to this dilemma would be to preferentially remove carbohydrate residues that are responsible for imparting the molecules' biologic action and sparing those that provided the molecules' long plasma half-life. It is, however, impossible to obtain this result with conventional chemical or enzymatic means (i.e. hydrogen fluoride treatment or enzymatic digestion) because of the non-specific nature of the chemical treatment.

It is an object of the present invention to provide a method for preferentially removing carbohydrate residues that are responsible for imparting biologic action to molecules while sparing those associated with a long plasma half-life.

It is yet another object of the present invention to provide a method for obtaining molecules via non-chemical treatment which have the desired antagonistic nature coupled with a long half-life.

It is still another object of the present invention to provide a recombinant technique for obtaining therapeutically effective glycoprotein hormones having been partially deglycosylated by having at least one oligosaccharide chain entirely removed by having at least one glycosylation site, corresponding to the absent or removed oligosaccharide chain, absent or removed.

It is still yet another object of the present invention to provide novel hormonal competitive antagonists having therapeutic utility.

It is yet still another object of the present invention to provide competitive hormone antagonists having substantially batch-to-batch uniformity and consistent potency.

Therefore, according to a first aspect of the present invention there is provided a modified glycoprotein hormone retaining substantially all the receptor binding capacity exhibited by the hormone in unmodified form, the hormone being modified by having at least one oligosaccharide chain entirely absent or removed. In some cases not all the oligosaccharide chains may be absent or removed. This may be achieved when a glycosylation site corresponding to the absent or removed oligosaccharide chain is absent or removed the or by site - directed mutagenesis where the hormone is deglycosylated at a selected (glycosylation) site. The site may be on the "a" subunit of the hormone, such as the 52 and 78 amino acids positions. Preferably the hormone is substantially incapable of giving rise to post receptor biological activity.

Usually the hormone has at least one sialic acid residue, and it is advantageous that it has a plasma half life approximately equal to that of the natural unmodified hormone. Such hormones include luteinizing hormone, follicle stimulating hormone, thyroid stimulating hormone or human chorionic gonadotropin (hCG). The hormone will thus be capable of competing with the natural (unmodified) glycoprotein hormone, which will usually be of the same type, eg. luteinizing stimulating, thyroid stimulating or hCG.

The invention thus also extends to a glycoprotein hormone capable of competing with natural glycoprotein for binding to a receptor but substantially incapable of giving rise to post receptor biological activity where the hormone has at least one oligosaccharide chain entirely removed by having at least one glycosylation site corresponding to the absent or removed oligosaccharide chain, absent or removed,

A second aspect of the present invention relates to a modified glycoprotein hormone retaining substantially all the receptor binding capacity exhibited by the unmodified hormone, the hormone being modified by having at least one oligosaccharide chain entirely absent or removed by having at least one glycosylation site corresponding to the absent or removed oligosaccharide chain, absent or removed, for use in medicine.

A third aspect relates to a pharmaceutical composition comprising a modified glycoprotein hormone retaining substantially all the receptor binding capacity exhibited by the unmodified hormone, the hormone being modified by having at least one oligosaccharide chain entirely absent or removed by having at least one glycosylation site corresponding to the absent or removed oligosaccharide chain, absent or removed, and a pharmaceutically acceptable carrier.

A fourth aspect relates to the use of a modified glycoprotein hormone retaining substantially all the receptor binding capacity exhibited by the unmodified hormone the hormone being modified by having at least one oligosaccharide chain absent or removed by having at least one glycosylation site corresponding to the absent or removed oligosaccharide chain, absent or removed, in the preparation of a competitive unmodified hormone antagonist. The use aspect also covers the use of hCG modified in accordance with

the present invention in the preparation of an agent for effecting a first trimester abortion, and the use of the modified hormones of the present invention in the preparation of an agent for treating the overproduction of stimulating hormones.

A fifth aspect relates to a DNA sequence encoding for a modified glycoprotein hormone retaining all the receptor binding capacity exhibited by the hormone in unmodified form, the hormone being modified by having at least one oligosaccharide chain entirely absent or removed by having at least one glycosylation site corresponding to the absent or removed oligosaccharide absent or removed.

A sixth aspect relates to a vector comprising a DNA sequence of the fifth aspect.

A seventh aspect concerns a host comprising a vector of the sixth aspect.

An eighth aspect relates to a process for the preparation of a pharmaceutical composition comprising admixing a modified glycoprotein hormone retaining substantially all the receptor binding capacity exhibited by the unmodified hormone, the hormone being modified by having at least one oligosaccharide chain entirely absent or removed by having at least one glycosylation site corresponding to the absent or removed oligosaccharide absent or removed, with a pharmaceutically acceptable carrier.

Preferred features of the second to eighth aspects of the present invention are as the first aspect *mutatis mutandis*.

In accordance with the principles and objects of the present invention, there are provided methods for creating antagonistic molecules by removing specific glycosylation sites normally present on the molecule through a recombinant DNA technique called site-directed mutagenesis. Specifically, the method of the present invention employs the fact that N-linked oligosaccharides attach to the protein portion of hormones at asparagine residues only when such asparagine residues are present in the carbohydrate attachment signalling sequence: asparagine - any amino acid - threonine or serine. The method of the instant invention advantageously eliminates the glycosylation site by changing the nucleotide sequence of the gene that codes for the hormone. This is advantageously accomplished by mutating the codon for asparagine residues to some other amino acid, preferably one that preserves the overall charge of the protein or, alternatively, changing the threonine or serine codon to another amino acid thereby removing the glycosylation signal. The resultant expressed protein hormone will be completely lacking the sugar normally present at the now altered glycosylation site thereby rendering it a competitive antagonist.

Advantageously, the method allows for the continued specific existence of other carbohydrate attachment points for prolonging the plasma half-life of the molecule without disadvantageously affecting transduction of the post-receptor bioactivity. Accordingly, and unlike prior methods, the instant invention can provide a large quantity of uniform quality, competitive hormone antagonist with a long plasma half-life. The invention further comprises therapeutically effective formulations of these modified molecules including pharmaceutically acceptable salts thereof and also any combination of the foregoing with a carrier vehicle ideally selected to aid in the administration of the molecule to a mammal or other animal without deleteriously affecting either the molecule or the animal.

Certain specific hormones of the instant invention may also be advantageously employed as safe and effective first-trimester abortifacients. After a newly ovulated egg is fertilized within the oviducts, it travels to the uterus where it implants in the innermost layer, the endometrium. Shortly after implantation, the trophoectodermal layers differentiate into chorionic villi which are finger-like projections that "root" deeper within the endometrial layer. As the chorionic villi grow, one cell type within them begins to manufacture large amounts of hCG. Until these villi differentiate and proliferate into a placenta with steroidogenic capacity, the hCG that is secreted travels to the ovary to "rescue" the corpus luteum from its programmed demise by maintaining its steroidogenic capacity. Additionally, hCG stimulates the continued secretion of progesterone which prevents uterine contractility and sloughing of the endometrium (menstruation) which would otherwise terminate pregnancy. In contrast, failure to maintain adequate serum progesterone levels results in spontaneous abortion until after the first trimester of pregnancy when the placenta begins to supply adequate progesterone to maintain the conceptus. Thus treatment of pregnant women, during the first trimester, with an hCG competitive antagonist of the present invention will result in a failure of endogenous hCG to maintain the corpus luteum, thereby resulting in a fall in serum progesterone levels and abortion.

Site-directed mutagenesis provides a powerful technique for generating defined point mutants. The general procedure, is described by Zoller, M. J. and Smith, M. (1982) Nucl. Acids Res. 10, 6487-6500 and is summarized as follows: The DNA fragment of interest is cloned into an M13 derived vector and single-stranded DNA (ssDNA) is prepared. A synthetic oligonucleotide, usually 18-20 bases long and complementary to a region of the ssDNA except for a mismatch which will direct the mutagenesis, is used as a primer. The template and primer are annealed. Large fragment *E. coli* DNA Polymerase I (Klenow), deoxyribonucleotides and T4 DNA ligase are added and closed circular double-stranded DNA (ccDNA) is

synthesized and ligated. Enrichment for ccDNA is done by one of several methods. The enriched ccDNA is transfected with competent JM101 cells and a population of both mutant and wild-type molecules are obtained. Any of a variety of well known screening procedures may be advantageously used to differentiate between the mutant and wild-type molecules. The mutant molecule is then isolated and confirmed by sequencing.

There are several points to consider when employing this method. It is highly desirable to do a thorough computer search with the proposed oligonucleotide to ensure specific priming at the desired site. The oligonucleotide should be checked against the M13 vector and the DNA fragment to be mutagenized to be certain this is the case. Another potential difficulty preferably avoided is the increased background of wild-type molecules resulting from inefficient conversion of ssDNA to ds ccDNA. Zoller and Smith reference several methods available to overcome this potential problem. Zoller and Smith, themselves, employed alkaline sucrose gradients for their enrichment. Similarly, S1 nuclease treatment may be used.

In designing the mutagenic oligonucleotide, it is preferable either to create or destroy a restriction enzyme site where possible. By so doing the screening for mutant DNA is greatly facilitated by a simple restriction site-fragment size analysis.

If the creation or destruction of suitable restriction sites is not possible, screening may be accomplished by a hybridization assay technique using the mutagenic oligonucleotide as a probe. This latter method is advantageously independent of the mutation position and can be done using either phage or isolated DNA. The principle of the hybridization assay is that the mutagenic oligonucleotide will form a more stable duplex with the mutant DNA because they match perfectly; whereas, the wild-type has mismatching and thus a lower level of complementarity. Advantageously, the hybridization is initially carried out under conditions of low stringency so that both the mutant and wild-type hybridize to the ³²P-labeled oligonucleotide. The filters are then washed at increasingly higher temperatures until only the mutants hybridize. This method is very rapid and can be usually carried out in one day. While an isotopic label has been suggested for the probe, other labels may be employed although other labels are generally not as detectable as isotopic labels at very low levels.

The invention will now be described by way of Example only, and the following Examples serve to illustrate, rather than limit, the present invention.

Example 1 - Site-Selection:

There are two glycosylation sites in the human α subunit, occurring at amino acid positions 52 and 78. The present invention directs that the amino acid sequence at these positions, Asn-X-Thr be changed to Asn-X-Val, Asn-X-Met or Gln-X-Thr. For "a52", there are several possible mutations that fit the above scheme all involving 2 base mismatches. The codon used for Thr at position 54 is ACC. The codons for Val are GTT, GTC, GTA and GTG; for Met, ATG. So the possibilities are ACC\$GTC or ACC\$ATG. AAC codes for Asn at amino acid 52 and changing this to Gln (CAA or CAG) again involves a 2 base mismatch. Unfortunately, none of the above mutations creates a restriction enzyme site. Several sites are destroyed but these are very common and provide no useful screening information.

For "a78", there are two potential choices for mutation of Thr (ACG). To change this codon to Met, ATG, requires a one base substitution. For Val, GTG, a double mutation is necessary. The Asn\$Gln conversion requires the same changes as in a52. Again, no restriction sites are created nor are any destroyed at these positions. However, and as noted previously, a 2 base mismatch will be easier to screen for the mutant since there will be greater homology for its' oligonucleotide than for the wild-type.

Preferred embodiment for the conversion of the Asn\$Thr for a78 is given below.

	Asn	His	Met	Ala	Cys
mutant...	AG	AAC	CAC	ATG	GCG
					TGC C

5'

3'

a78

AG AAC CAC ACG GCG TGC C
Asn His Thr Ala Cys

oligo

TCTTGGTG TAC CGCACGG

3'

5'

Example 2 - Materials:

M13mp18 and mp19RF, IPTG, XGal and phenol were obtained from BRL (Bethesda Research Laboratories). T4 DNA ligase, T4 polynucleotide kinase, all restriction endonucleases and *E.coli* DNA polymerase I large fragment Klenow were purchased from New England Biolabs. ATP was obtained from Sigma. γ - 32 P-ATP and α - 32 P-ATP were purchased from DuPont/NEN Medical Products. PEG-6000 was obtained from Kodak. Acrylamide, bis-acrylamide and TEMED were purchased from BioRad. S1 nuclease, ribonuclease A and *E.coli* tRNA was obtained from Boehringer Mannheim. Deoxyribonucleotide triphosphates, pUC 18 and pUC 19 plasmids were purchased from PL-Pharmacia. Nitrocellulose filters were obtained from Schleicher and Schuell.

Buffers, Media and Stock Solutions

The following buffers were formulated:

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Buffers

10x kinase

500mM Tris-Cl(7.5)

100mM MgCl₂

10mM DTT

1mM spermidine

1mM EDTA

5	10x ligase	500mM Tris-Cl(7.5) 100mM MgCl ₂ 10mM DTT
10	10x ATP	10mM ATP, pH 7.0
15	10x dNTP's	2mM dATP, 2mM dCTP, 2 mM dGTP, 2mM DTTP pH 7.0
20	S1"stop"	50mM Tris base 20mM EDTA 1ug carrier tRNA
25	10x S1	300mM NaOAc (4.5) 500mM NaCl 45mM ZnSO ₄
30	1x TE	10mM Tris-Cl(8.0) 1mM EDTA
35	Solution A	200mM Tris-Cl(7.5) 100mM MgCl ₂ 500mM NaCl 10mM DTT
40	50x Denhardt's	1% Ficoll 1% Polyvinylpyrrolidone 1% BSA
45	1x SSC	150mM NaCl 15mM Na citrate

50

55

Media5x M9 Salts

10.5 g/L K_2HPO_4
 4.5 g/L KH_2PO_4
 1.0 g/L $(NH_4)_2 SO_4$
 0.5 g/L Na citrate

Autoclave and temper at 55°C.

Add, with sterile technique: 1 ml
 1M $MgSO_4 \cdot 7 H_2O$; 0.7 ml 0.7%
 thiamine, 10 ml 20% glucose.

Minimal Media Plates

Add 15g bactogar to 800 ml ddH₂O, autoclave. After tempering to 55°C, add 200 ml 5 X M9 salts and pour plates.

YT

8 g/L bactotryptone
 5 g/L yeast extract
 5 g/L NaCl
 Autoclave

Example 3 - Preparation of ssDNA:

The desired DNA fragment encoding for the alpha subunit was digested with the appropriate restriction enzyme and isolated from a polyacrylamide gel. The fragment was ligated into the appropriate site in the M13 vector of choice via conventional techniques. Approximately 300ng of fragment and 30ng digested M13 were mixed in 10x ligase buffer, 10x ATP and 400U T4 DNA ligase in a total volume of 50μl. The ligation reaction was incubated for four to six hours at room temperature or alternately it can be incubated for 20 hours at 16°C.

A loopful of JM101 cells was transferred from a M9 minimal plate to a flask containing the appropriate volume of YT and grown to a density of $OD_{660} = 0.3-0.4$ at 37°C with aeration. The cells were collected by centrifugation in a Sorvall SS-34 at 8,000rpm for ten minutes at 4°C, immediately resuspended in 0.5 volumes of ice-cold 50mM $CaCl_2$ and kept on ice for 20 minutes. The cells were centrifuged again and resuspended in 1/10 volume original growth volume with chilled 50mM $CaCl_2$. The competent cells are left on ice until ready to use.

Aliquots of the ligation reaction were added to the JM101 competent cells and transfected as described by Messing, J. (1984) Methods Enzym. 101, 20. ssDNA was prepared from individual plaques by growing 5 ml YT cultures with 50μl exponential JH101 for six to eight hours at 37°C. The cells were centrifuged at 3000rpm for ten minutes at 4°C and 4 ml supernatants were transferred to clean Corex tubes. PEG, NaCl and RNaseA were added to final concentrations of 4%, 500mM and 10μg/ml, respectively. The tubes were incubated at room temperature for 15 minutes. The solutions were then centrifuged at 8,000rpm for 10 to 15 minutes at 4°C in the SS-34 rotor. The supernatants were decanted and the pellets resuspended in 300μl 1xTE and transferred to microfuge Eppendorf tubes. A TE-saturated phenol, then ether extractions were done, followed by ethanol precipitation with 1/10 volume 3M NaOAc, pH 5.2. The dried pellet was resuspended in TE. A recombinant with the desired orientation was then chosen and a one liter culture of ssDNA was prepared by scaling-up the above method. This ssDNA served as template source for all

mutagenesis experiments.

Example 4 - 5' Phosphorylation of the Oligonucleotide:

For mutagenesis: 1000-3000 pmol of oligonucleotide was phosphorylated in a solution containing 2μl 10x kinase buffer, 2μl 10x ATP and 4U polynucleotide kinase in a total volume of 20μl. The reaction was incubated at 37°C for 30-60 minutes and terminated by incubation at 65°C for 10 minutes.

For hybridization: The oligonucleotide was phosphorylated as above in a volume of 50μl using 50-100μCi γ -³²P-ATP as the only source of ATP. To remove unincorporated nucleotide, the reaction was precipitated with a 1/2 volume 7.5M NH₄OAc and three volumes of cold ethanol. The pellet was washed several times with cold 70% ethanol, dried, and resuspended in 50μl TE.

Example 5 - Oligonucleotide - directed synthesis of ds ccDNA:

Annealing: Approximately 13 pmol of ssDNA, 260 pmol 5' phosphorylated oligonucleotide and 1μl of solution A were mixed in a total volume of 10μl. As a control, the above was done without the oligonucleotide. The reactions were incubated at 55°C for five minutes, then 22°C for five minutes.

Extension: 1.5μl 10x Klenow buffer, 1.5μl 10x dNTP'S and 5U Klenow were added to the annealing reactions in a final volume of 15μl. These reactions were incubated for five minutes at 22°C.

Ligation: To the above reactions, 1μl 10x ligase, 2μl 10x ATP and 400U T4 DNA ligase were added to a final volume of 20μl. The reactions were incubated for 20 hours at 16°C.

Example 6 - Enrichment for ccDNA:

To the ligation reaction, 3μl 10x S1 buffer and 0.9U S1 nuclease were added to a final volume of 30μl. A 15μl aliquot was immediately removed into a 1.5 ml Eppendorf tube containing 40μl S1 "stop" buffer. The S1 nuclease reaction was allowed to proceed for three minutes at room temperature. After three minutes, 40μl "stop" buffer was added to the original reaction eppendorf. The reactions were phenol:CHCl₃ extracted and ethanol precipitated. The pellets were then resuspended in TE.

Example 7 - Transfection of ccDNA:

Aliquots of the ligation reactions were used to transfect competent JM101 cells as previously described.

Example 8 - Screening:

A dry nitrocellulose filter was carefully placed on top of each transfection plate and placed at 4°C for 15-30 minutes. The filters were processed as described by Maniatis, L., Fritsch, E.F. and Sambrook, J. (1982) Molecular Cloning: A Laboratory Manual, Cold Spring Harbor. The baked filters were wetted with 2x SSC and prehybridized for two or more hours in a solution of 150mM Tris-Cl(8.0), 750mM NaCl, 10mM EDTA, 5x Denhardt's, 0.1% SDS and 0.1% sodium pyrophosphate in a sealed coaking bag at room temperature. The prehybridization solution was removed and replaced with fresh solution containing 1x 10⁶cpm/ml solution ³²P-labeled oligonucleotide. Hybridization was carried out at room temperature for 2-20 hours. The filters were then washed with 6X SCC, 0.1% SDS and 0.01M sodium phosphate (pH 7.2) for 20-30 minutes at increasing temperatures with autoradiography after each temperature change for one hour at -80°C with two intensifying screens. The final washing temperature was based on the equation: $T_m = 2^\circ\text{C} \times (\text{the number of A-T bases}) + 4^\circ\text{C} \times (\text{the number of G-C bases})$ pursuant to Hanahan, D. and Meselson, M. (1983) Methods Enzym. 100, 333-342. It was observed that the wild-type was usually off at $T_m - 10^\circ\text{C}$.

Example 9 - Isolation of ds mutant:

Single, well-isolated positives were removed into 5 ml YT with 50μl exponential JM101 as well as streaked into a YT plate with a JM101 lawn. The cultures were incubated at 37°C with aeration for six to eight hours and then centrifuged at 3,000rpm for ten minutes at 4°C. ds DNA was isolated as described in Maniatis, L., Fritsch, E.J. and Sambrook, J. (1982) Molecular Cloning: A Laboratory Manual, Cold Spring Harbor. If possible, mutations are confirmed by restriction digests. If not, the screening procedure is (ideally) repeated on the streaked plates. A large culture of the proposed mutant ds DNA was prepared pursuant to the method described in Maniatis, p.90.

Example 10 - DNA Sequencing:

The double-stranded mutant DNA was then digested with the appropriate restriction enzymes and isolated from polyacrylamide gels. These fragments were end-labeled and sequenced by the method described by Maxam, A.M. and Gilbert, W. (1980) *Methods Enzym.* 65, 499-560. Alternatively, the fragments can be subcloned into M13 or pUC vectors and sequenced by the dideoxy method described by Sanger, J., Nicklen, S. and Coulson, A. R. (1977) *Proc. Natl. Acad. Sci. U.S.A* 74, 5463-5467.

Transfection of mammalian cells and selection of a cell line produced by the above procedures can then be expressed by the following procedure: The mutant ds DNA can be ligated with Bam linkers and inserted into a bovine papilloma virus (BPV) expression cassette that includes the BPV genome, the mouse metallothionein promoter and SV40 poly (A) sequences as per Hsiung et al. (1984) *J. Mol. Applied Genetics* 2, 497. Mouse epithelioid cells (C127) may then be transfected using the calcium phosphate precipitation method. Transformed cells are ideally identified 10-14 days after transfection by their piled-up appearance. Foci of cells should then be subcloned to obtain a pure population and cultured in Dulbecco's Modified Eagle's Medium (DMEM) supplemented with 2% fetal bovine serum. Mutant hCG secreted into the culture medium will be purified by a three step process that included Tris Acryl Blue chromatography, ion exchange and gel filtration chromatography. Activity can be monitored through the purification process by a radioimmunoassay kit available for this purpose from Sero Laboratories.

The cell line that expresses the greatest amount of mutant hCG will stimulate the secretion of testosterone from suspension cultures of Leydig cells. Leydig cells can be obtained from adult male mice testes. Approximately 0.25×10^6 Leydig cells are ideally incubated in approximately 1 ml (DMEM) that contains one of several dose levels of native hCG added to the cells. A half-maximal dose of hCG may then be readily determined. A second set of suspension cultures should then be treated with a half-maximal dose of hCG and one of several increasing doses of the mutant recombinant hCG. A biologically inactive form of hCG will compete for binding to the receptor upon the Leydig cells and decrease the testosterone response of the native form.

Following the determination of the dose relationship between native and mutant hCG necessary to completely inhibit the native hCG-induced testosterone response, the mutant form can then be readily tested for its ability to induce abortion in a variety of species reported to secrete a chorionic gonadotropin, including the rhesus monkey and the optional antagonist hormone selected in accordance with the principles of the present invention.

Claims

Claims for the following Contracting States : AT, BE, CH, DE, FR, GB, IT, LI, LU, NL, SE

1. A modified glycoprotein hormone retaining substantially all the receptor binding capacity exhibited by the hormone in unmodified form, the hormone being modified by having at least one oligosaccharide chain entirely absent or removed by having at least one glycosylation site, corresponding to the absent or removed oligosaccharide chain, absent or removed.
2. A hormone as claimed in claim 1 which is substantially incapable of giving rise to post receptor biological activity.
3. A hormone as claimed in claim 1 or 2 having at least one sialic acid residue.
4. A hormone as claimed in any of claims 1 to 3 having a plasma half-life approximately equal to that of the natural unmodified hormone.
5. A hormone according to any of claims 1 to 4 wherein the glycosylation site is absent or removed from the a subunit.
6. A hormone as claimed in any of claims 1 to 5 which is luteinizing hormone (LH), follicle stimulating hormone (FSH), thyroid stimulating hormone (TSH) or human chorionic gonadotropin (hCG).
7. A modified glycoprotein hormone retaining substantially all the receptor binding capacity exhibited by the unmodified hormone, the hormone being modified by having at least one oligosaccharide chain entirely absent or removed by having at least one glycosylation site, corresponding to the absent or removed oligosaccharide chain, absent or removed, for use in medicine.

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The double-stranded mutant DNA was then digested with the appropriate restriction enzymes and isolated from polyacrylamide gels. These fragments were end-labeled and sequenced by the method described by Maxam, A.M. and Gilbert, W. (1980) *Methods Enzym.* 65, 499-560. Alternatively, the fragments can be subcloned into M13 or pUC vectors and sequenced by the dideoxy method described by Sanger, J., Nicklen, S. and Coulson, A. R. (1977) *Proc. Natl. Acad. Sci. U.S.A* 74, 5463-5467.

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Following the determination of the dose relationship between native and mutant hCG necessary to completely inhibit the native hCG-induced testosterone response, the mutant form can then be readily tested for its ability to induce abortion in a variety of species reported to secrete a chorionic gonadotropin, including the rhesus monkey and the optional antagonist hormone selected in accordance with the principles of the present invention.

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2. A hormone as claimed in claim 1 which is substantially incapable of giving rise to post receptor biological activity.
3. A hormone as claimed in claim 1 or 2 having at least one sialic acid residue.
4. A hormone as claimed in any of claims 1 to 3 having a plasma half-life approximately equal to that of the natural unmodified hormone.
5. A hormone according to any of claims 1 to 4 wherein the glycosylation site is absent or removed from the a subunit.
6. A hormone as claimed in any of claims 1 to 5 which is luteinizing hormone (LH), follicle stimulating hormone (FSH), thyroid stimulating hormone (TSH) or human chorionic gonadotropin (hCG).
7. A modified glycoprotein hormone retaining substantially all the receptor binding capacity exhibited by the unmodified hormone, the hormone being modified by having at least one oligosaccharide chain entirely absent or removed by having at least one glycosylation site, corresponding to the absent or removed oligosaccharide chain, absent or removed, for use in medicine.



8. A hormone as claimed in claim 7 which is luteinizing hormone, follicle stimulating hormone, thyroid stimulating hormone or human chorionic gonadotropin.
9. A pharmaceutical composition comprising a modified glycoprotein hormone retaining substantially all the receptor binding capacity exhibited by the unmodified hormone, the hormone being modified by having at least one oligosaccharide chain entirely absent or removed, by having at least one glycosylation site, corresponding to the absent or removed oligosaccharide chain, absent or removed, and a pharmaceutically acceptable carrier.
10. A composition as claimed in claim 10 wherein the hormone is luteinizing hormone, follicle stimulating hormone, thyroid stimulating hormone or human chorionic gonadotropin.
11. The use of a modified glycoprotein hormone retaining substantially all the receptor binding capacity exhibited by the unmodified hormone, the hormone being modified by having at least one oligosaccharide chain absent or removed, by having at least one glycosylation site, corresponding to the absent or removed oligosaccharide chain, absent or removed, in the preparation of a competitive unmodified hormone antagonist.
12. A DNA sequence encoding for a modified glycoprotein hormone retaining substantially all the receptor binding capacity exhibited by the hormone in unmodified form, the hormone being modified by having at least one oligosaccharide chain entirely absent or removed, by having at least one glycosylation site, corresponding to the absent or removed oligosaccharide chain, absent or removed.
13. A vector comprising a DNA sequence as claimed in claim 12.
14. A host comprising a vector as claimed in claim 13.

Claims for the following Contracting State : ES

1. A process for the preparation of a pharmaceutical composition comprising admixing a modified glycoprotein hormone retaining substantially all the receptor binding capacity exhibited by the hormone in unmodified form, the hormone being modified by having at least one oligosaccharide chain entirely absent or removed by having at least one glycosylation site, corresponding to the absent or removed oligosaccharide chain, absent or removed, with a pharmaceutically acceptable carrier.
2. A process as claimed in claim 1 wherein the hormone is substantially incapable of giving rise to post receptor biological activity.
3. A process as claimed in claim 1 or 2 wherein the hormone has at least one sialic acid residue.
4. A process as claimed in any of claims 1 to 3 wherein the hormone has a plasma half-life approximately equal to that of the natural unmodified hormone.
5. A process as claimed in any of claims 1 to 4 wherein the glycosylation site is absent or removed from the a subunit.
6. A process as claimed in any of claims 1 to 5 wherein the hormone is luteinizing hormone (LH), follicle stimulating hormone (FSH), thyroid stimulating hormone (TSH) or human chorionic gonadotropin (hCG).
7. The use of a modified glycoprotein hormone retaining substantially all the receptor binding capacity exhibited by the unmodified hormone, the hormone being modified by having at least one oligosaccharide chain entirely absent or removed by having at least one glycosylation site, corresponding to the absent or removed oligosaccharide chain, absent or removed, in the preparation of a competitive unmodified hormone antagonist.
8. The use according to claim 7 wherein the hormone is substantially incapable of giving rise to post-receptor biological activity.



9. The use as claimed in claim 7 or 8 wherein the hormone has at least one sialic acid residue.
10. The use according to any of claims 7 to 9 wherein the hormone has a plasma half-life approximately equal to that of the natural unmodified hormone.
11. The use according to any of claims 7 to 10 wherein the glycosylation site is absent or removed from the a subunit.
12. The use as claimed in any of claims 7 to 11 wherein the hormone is luteinizing hormone, follicle stimulating hormone, thyroid stimulating hormone or human chorionic gonadotropin.

Claims for the following Contracting State : GR

1. A process for the preparation of a pharmaceutical composition comprising admixing a modified glycoprotein hormone retaining substantially all the receptor binding capacity exhibited by the hormone in unmodified form, the hormone being modified by having at least one oligosaccharide chain entirely absent or removed by having at least one glycosylation site, corresponding to the absent or removed oligosaccharide chain, absent or removed, with a pharmaceutically acceptable carrier.
2. A process as claimed in claim 1 wherein the hormone is substantially incapable of giving rise to post receptor biological activity.
3. A process as claimed in claim 1 or 2 wherein the hormone has at least one sialic acid residue.
4. A process as claimed in any of claims 1 to 3 wherein the hormone has a plasma half-life approximately equal to that of the natural unmodified hormone.
5. A process as claimed in any of claims 1 to 4 wherein the glycosylation site is absent or removed from the a subunit.
6. A process as claimed in any of claims 1 to 5 wherein the hormone is luteinizing hormone (LH), follicle stimulating hormone (FSH), thyroid stimulating hormone (TSH) or human chorionic gonadotropin (hCG).
7. The use of a modified glycoprotein hormone retaining substantially all the receptor binding capacity exhibited by the unmodified hormone, the hormone being modified by having at least one oligosaccharide chain entirely absent or removed by having at least one glycosylation site, corresponding to the absent or removed oligosaccharide chain, absent or removed, in the preparation of a competitive unmodified hormone antagonist.
8. The use according to claim 7 wherein the glycosylation site is absent or removed from the a subunit.
9. The use as claimed in claim 7 or 8 wherein the hormone has at least one sialic acid residue.
10. The use as claimed in any of claims 7 to 9 wherein the hormone is luteinizing hormone, follicle stimulating hormone, thyroid stimulating hormone or human chorionic gonadotropin.
11. A DNA sequence encoding for a modified glycoprotein hormone retaining substantially all the receptor binding capacity exhibited by the hormone in unmodified form, the hormone being modified by having at least one oligosaccharide chain entirely absent or removed, by having at least one glycosylation site, corresponding to the absent or removed oligosaccharide chain, absent or removed.
12. A vector comprising a DNA sequence as claimed in claim 11.
13. A host comprising a vector as claimed in claim 12.



Patentansprüche**Patentansprüche für folgende Vertragsstaaten : AT, BE, CH, DE, FR, GB, IT, LI, LU, NL, SE**

- 5 1. Modifiziertes Glycoproteinormon, das im wesentlichen die gesamte Rezeptorbindekazapazität beibehält, die das Hormon in unmodifizierter Form zeigt, wobei das Hormon dadurch modifiziert ist, daß zumindest eine Oligosaccharidkette vollkommen fehlt oder entfernt wurde, indem zumindest eine Glycosylierungsstelle fehlt oder entfernt wurde, die der fehlenden oder entfernten Oligosaccharidkette entspricht.
- 10 2. Hormon nach Anspruch 1, das im wesentlichen unfähig ist, eine Erhöhung der nach der Bindung an den Rezeptor vorliegenden biologischen Aktivität hervorzurufen.
3. Hormon nach Anspruch 1 oder 2 mit zumindest einem Sialsäurerest.
- 15 4. Hormon nach einem der Ansprüche 1 bis 3 mit einer Plasmahalbwertszeit, die jener des natürlichen, unmodifizierten Hormons annähernd gleich ist.
5. Hormon nach einem der Ansprüche 1 bis 4, bei welchem die Glycosylierungsstelle fehlt oder von der "a"-Untereinheit entfernt wurde.
- 20 6. Hormon nach einem der Ansprüche 1 bis 5, welches das Luteinisierungshormon (LH), follikelstimulierendes Hormon (FSH), thyreotropes Hormon (TSH) oder Choriogonadotropin (hCG) ist.
- 25 7. Modifiziertes Glycoproteinormon, das im wesentlichen die gesamte Rezeptorbindekazapazität beibehält, die das unmodifizierte Hormon zeigt, wobei das Hormon dadurch modifiziert ist, daß zumindest eine Oligosaccharidkette vollkommen fehlt oder entfernt wurde, indem zumindest eine Glycosylierungsstelle fehlt oder entfernt wurde, die der fehlenden oder entfernten Oligosaccharidkette entspricht, zur Verwendung in der Medizin.
- 30 8. Hormon nach Anspruch 7, welches das Luteinisierungshormon, follikelstimulierendes Hormon, thyreotropes Hormon oder Choriogonadotropin ist.
- 35 9. Pharmazeutische Zusammensetzung, bestehend aus einem modifizierten Glycoproteinormon, das im wesentlichen die gesamte Rezeptorbindekazapazität beibehält, die das unmodifizierte Hormon zeigt, wobei das Hormon dadurch modifiziert ist, daß zumindest eine Oligosaccharidkette vollkommen fehlt oder entfernt wurde, indem zumindest eine Glycosylierungsstelle fehlt oder entfernt wurde, die der fehlenden oder entfernten Oligosaccharidkette entspricht, und einem pharmazeutisch annehmbaren Träger.
- 40 10. Zusammensetzung nach Anspruch 10, wobei das Hormon Luteinisierungshormon, follikelstimulierendes Hormon, thyreotropes Hormon oder Choriogonadotropin ist.
- 45 11. Verwendung eines modifizierten Glycoproteinormons, das im wesentlichen die gesamte Rezeptorbindekazapazität beibehält, die das unmodifizierte Hormon zeigt, in der Herstellung eines kompetitiven unmodifizierten Hormonantagonisten, wobei das Hormon dadurch modifiziert ist, daß zumindest eine Oligosaccharidkette vollkommen fehlt oder entfernt wurde, indem zumindest eine Glycosylierungsstelle fehlt oder entfernt wurde, die der fehlenden oder entfernten Oligosaccharidkette entspricht.
- 50 12. DNA-Sequenz, die für ein modifiziertes Glycoproteinormon codiert, das im wesentlichen die gesamte Rezeptorbindekazapazität beibehält, die das Hormon in unmodifizierter Form zeigt, wobei das Hormon dadurch modifiziert ist, daß zumindest eine Oligosaccharidkette vollkommen fehlt oder entfernt wurde, indem zumindest eine Glycosylierungsstelle fehlt oder entfernt wurde, die der fehlenden oder entfernten Oligosaccharidkette entspricht.
- 55 13. Vektor, der eine DNA-Sequenz nach Anspruch 12 umfaßt.
14. Wirt, der einen Vektor nach Anspruch 13 umfaßt.



Patentansprüche für folgenden Vertragsstaat : ES

1. Verfahren zur Herstellung einer pharmazeutischen Zusammensetzung, bestehend aus dem Vermischen eines modifizierten Glycoproteinhormons, das im wesentlichen die gesamte Rezeptorbindekazapazität beibehält, die das Hormon in unmodifizierter Form zeigt, wobei das Hormon dadurch modifiziert ist, daß zumindest eine Oligosaccharidkette vollkommen fehlt oder entfernt wurde, indem zumindest eine Glycosylierungsstelle fehlt oder entfernt wurde, die der fehlenden oder entfernten Oligosaccharidkette entspricht, mit einem pharmazeutisch verträglichen Träger.
2. Verfahren nach Anspruch 1, wobei das Hormon im wesentlichen unfähig ist, eine Erhöhung der nach der Bindung an den Rezeptor vorliegenden biologischen Aktivität hervorzurufen.
3. Verfahren nach Anspruch 1 oder 2, bei welchem das Hormon zumindest einen Sialsäurerest aufweist.
4. Verfahren nach einem der Ansprüche 1 bis 3, wobei das Hormon eine Plasmahalbwertszeit aufweist, die jener des natürlichen, unmodifizierten Hormons annähernd gleich ist.
5. Verfahren nach einem der Ansprüche 1 bis 4, wobei die Glycosylierungsstelle fehlt oder von der "a"-Untereinheit entfernt wurde.
6. Verfahren nach einem der Ansprüche 1 bis 5, wobei das Hormon Luteinisierungshormon (LH), follikelstimulierendes Hormon (FSH), thyreotropes Hormon (TSH) oder Choriongonadotropin (hCG) ist.
7. Verwendung eines modifizierten Glycoproteinhormons, das im wesentlichen die gesamte Rezeptorbindekazapazität beibehält, die das unmodifizierte Hormon zeigt, wobei das Hormon dadurch modifiziert ist, daß zumindest eine Oligosaccharidkette vollkommen fehlt oder entfernt wurde, indem zumindest eine Glycosylierungsstelle fehlt oder entfernt wurde, die der fehlenden oder entfernten Oligosaccharidkette entspricht, in der Herstellung eines kompetitiven, unmodifizierten Hormonantagonisten.
8. Verwendung nach Anspruch 7, wobei das Hormon im wesentlichen unfähig ist, eine Erhöhung der nach der Bindung an den Rezeptor vorliegenden biologischen Aktivität hervorzurufen.
9. Verwendung nach Anspruch 7 oder 8, wobei das Hormon zumindest einen Sialsäurerest aufweist.
10. Verwendung nach einem der Ansprüche 7 bis 9, wobei das Hormon eine Plasmahalbwertszeit aufweist, die jener des natürlichen, unmodifizierten Hormons annähernd gleich ist.
11. Verwendung nach einem der Ansprüche 7 bis 10, wobei die Glycosylierungsstelle fehlt oder von der "a"-Untereinheit entfernt wurde.
12. Verwendung nach einem der Ansprüche 7 bis 11, wobei das Hormon Luteinisierungshormon, follikelstimulierendes Hormon, thyreotropes Hormon oder Choriongonadotropin ist.

Patentansprüche für folgenden Vertragsstaat : GR

1. Verfahren zur Herstellung einer pharmazeutischen Zusammensetzung, bestehend aus dem Vermischen eines modifizierten Glycoproteinhormons, das im wesentlichen die gesamte Rezeptorbindekazapazität beibehält, die das Hormon in unmodifizierter Form zeigt, wobei das Hormon dadurch modifiziert ist, daß zumindest eine Oligosaccharidkette vollkommen fehlt oder entfernt wurde, indem zumindest eine Glycosylierungsstelle fehlt oder entfernt wurde, die der fehlenden oder entfernten Oligosaccharidkette entspricht, mit einem pharmazeutisch verträglichen Träger.
2. Verfahren nach Anspruch 1, wobei das Hormon im wesentlichen unfähig ist, eine Erhöhung der nach der Bindung an den Rezeptor vorliegenden biologischen Aktivität hervorzurufen.
3. Verfahren nach Anspruch 1 oder 2, bei welchem das Hormon zumindest einen Sialsäurerest aufweist.



4. Verfahren nach einem der Ansprüche 1 bis 3, wobei das Hormon eine Plasmahalbwertszeit aufweist, die jener des natürlichen, unmodifizierten Hormons annähernd gleich ist.
5. Verfahren nach einem der Ansprüche 1 bis 4, wobei die Glycosylierungsstelle fehlt oder von der "a"-Untereinheit entfernt wurde.
6. Verfahren nach einem der Ansprüche 1 bis 5, wobei das Hormon Luteinisierungshormon (LH), follikelstimulierendes Hormon (FSH), thyreotropes Hormon (TSH) oder Choriongonadotropin (hCG) ist.
7. Verwendung eines modifizierten Glycoproteinhormons, das im wesentlichen die gesamte Rezeptorbindekapazität beibehält, die das unmodifizierte Hormon zeigt, wobei das Hormon dadurch modifiziert ist, daß zumindest eine Oligosaccharidkette vollkommen fehlt oder entfernt wurde, indem zumindest eine Glycosylierungsstelle fehlt oder entfernt wurde, die der fehlenden oder entfernten Oligosaccharidkette entspricht, in der Herstellung eines kompetitiven, unmodifizierten Hormonantagonisten.
8. Verwendung nach Anspruch 7, wobei die Glycosylierungsstelle fehlt oder von der "a"-Untereinheit entfernt wurde.
9. Verwendung nach Anspruch 7 oder 8, wobei das Hormon zumindest einen Sialsäurerest aufweist.
10. Verwendung nach einem der Ansprüche 7 bis 9, wobei das Hormon Luteinisierungshormon, follikelstimulierendes Hormon, thyreotropes Hormon oder Choriongonadotropin ist.
11. DNA-Sequenz, die für ein modifiziertes Glycoproteinhormon codiert, das im wesentlichen die gesamte Rezeptorbindekapazität beibehält, die das Hormon in unmodifizierter Form zeigt, wobei das Hormon dadurch modifiziert ist, daß zumindest eine Oligosaccharidkette vollkommen fehlt oder entfernt wurde, indem zumindest eine Glycosylierungsstelle fehlt oder entfernt wurde, die der fehlenden oder entfernten Oligosaccharidkette entspricht.
12. Vektor, der eine DNA-Sequenz nach Anspruch 11 umfaßt.
13. Wirt, der einen Vektor nach Anspruch 12 umfaßt.

Revendications

35 **Revendications pour les Etats contractants suivants : AT, BE, CH, DE, FR, GB, IT, LI, LU, NL, SE**

1. Hormone glycoprotéique modifiée conservant substantiellement l'entière capacité de se fixer au récepteur que présente l'hormone sous sa forme non-modifiée, l'hormone étant modifiée par le fait qu'au moins une chaîne oligosaccharidique est totalement absente ou enlevée par le fait qu'au moins un site de glycosylation, correspondant à la chaîne oligosaccharidique absente ou enlevée, est absent ou enlevé.
2. Hormone suivant la revendication 1, qui est substantiellement incapable de déclencher une activité biologique après sa fixation au récepteur.
3. Hormone suivant la revendication 1 ou 2, présentant au moins un résidu acide sialique.
4. Hormone suivant l'une quelconque des revendications 1 à 3, ayant une demi-vie plasmatique à peu près égale à celle de l'hormone naturelle non-modifiée.
5. Hormone suivant l'une quelconque des revendications 1 à 4, dans laquelle le site de glycosylation est absent ou enlevé de la sous-unité a.
6. Hormone suivant l'une quelconque des revendications 1 à 5, qui est la lutéostimuline (LH), la folliculo-stimuline (FSH), la thyroïdo-stimuline (TSH) ou la gonadotrophine chorionique humaine (hCG).
7. Hormone glycoprotéique modifiée conservant substantiellement l'entière capacité de se fixer au récepteur que présente l'hormone non-modifiée, l'hormone étant modifiée par le fait qu'au moins une



chaîne oligosaccharidique est entièrement absente ou enlevée par le fait qu'au moins un site de glycosylation, correspondant à la chaîne oligosaccharidique absente ou enlevée, est absent ou enlevé, utilisée à des fins médicales.

- 5 8. Hormone suivant la revendication 7, qui est la lutéo-stimuline, la folliculo-stimuline, la thyroïdo-stimuline ou la gonadotrophine chorionique humaine.
- 10 9. Composition pharmaceutique comportant une hormone glycoprotéique modifiée conservant substantiellement l'entière capacité de se fixer au récepteur que présente l'hormone non-modifiée, l'hormone étant modifiée par le fait qu'au moins une chaîne oligosaccharidique est entièrement absente ou enlevée, par le fait qu'au moins un site de glycosylation, correspondant à la chaîne oligosaccharidique absente ou enlevée, est absent ou enlevé, et un porteur acceptable sur le plan pharmaceutique.
- 15 10. Composition suivant la revendication 10, dans laquelle l'hormone est la lutéo-stimuline, la folliculo-stimuline, la thyroïdo-stimuline ou la gonadotrophine chorionique humaine.
- 20 11. Utilisation d'une hormone glycoprotéique modifiée conservant substantiellement l'entière capacité de se fixer au récepteur que présente l'hormone non-modifiée, l'hormone étant modifiée par le fait qu'au moins une chaîne oligosaccharidique est absente ou enlevée, par le fait qu'au moins un site de glycosylation, correspondant à la chaîne oligosaccharidique absente ou enlevée, est absent ou enlevé, intervenant dans la préparation d'un antagoniste hormonal non-modifié et compétitif.
- 25 12. Séquence d'ADN codant pour une hormone glycoprotéique modifiée conservant substantiellement l'entière capacité de se fixer au récepteur que présente l'hormone sous sa forme non-modifiée, l'hormone étant modifiée par le fait qu'au moins une chaîne oligosaccharidique est entièrement absente ou enlevée, par le fait qu'au moins un site de glycosylation, correspondant à la chaîne oligosaccharidique absente ou enlevée, est absent ou enlevé.
- 30 13. Vecteur comportant une séquence d'ADN suivant la revendication 12.
14. Hôte comportant un vecteur suivant la revendication 13.

Revendications pour l'Etats contractants suivants : ES

- 35 1. Procédé de préparation d'une composition pharmaceutique comprenant le mélange d'une hormone glycoprotéique modifiée conservant l'entière capacité de se fixer au récepteur que présente l'hormone sous sa forme non-modifiée, avec un porteur acceptable sur le plan pharmaceutique, l'hormone étant modifiée par le fait qu'au moins une chaîne oligosaccharidique est entièrement absente ou enlevée par le fait qu'au moins un site de glycosylation, correspondant à la chaîne oligosaccharidique absente ou enlevée, est absent ou enlevé.
- 40 2. Procédé suivant la revendication 1, dans lequel l'hormone est substantiellement incapable de déclencher une activité biologique après sa fixation au récepteur.
- 45 3. Procédé suivant la revendication 1 ou 2, dans lequel l'hormone présente au moins un résidu acide sialique.
- 50 4. Procédé suivant l'une quelconque des revendications 1 à 3, dans lequel l'hormone a une demi-vie plasmatique à peu près égale à celle de l'hormone naturelle non-modifiée.
- 55 5. Procédé suivant l'une quelconque des revendications 1 à 4, dans lequel le site de glycosylation est absent ou enlevé de la sous-unité α .
6. Procédé suivant l'une quelconque des revendications 1 à 5, dans lequel l'hormone est la lutéo-stimuline (LH), la folliculo-stimuline (FSH) la thyroïdo-stimuline (TSH) ou la gonadotrophine chorionique humaine (hCG).



7. Utilisation d'une hormone glycoprotéique modifiée conservant substantiellement l'entière capacité de se fixer au récepteur que présente l'hormone non-modifiée, l'hormone étant modifiée par le fait qu'au moins une chaîne oligosaccharidique est entièrement absente ou enlevée par le fait qu'au moins un site de glycosylation correspondant à la chaîne oligosaccharidique absente ou enlevée, est absent ou enlevé, intervenant dans la préparation d'un antagoniste hormonal non-modifié et compétitif.
8. Utilisation suivant la revendication 7, dans laquelle l'hormone est substantiellement incapable de déclencher une activité biologique après sa fixation au récepteur.
9. Utilisation suivant la revendication 7 ou 8, dans laquelle l'hormone présente au moins un résidu acide sialique.
10. Utilisation suivant l'une quelconque des revendications 7 à 9, dans laquelle l'hormone a une demi-vie plasmatique à peu près égale à celle de l'hormone naturelle non-modifiée.
11. Utilisation suivant l'une quelconque des revendications 7 à 10, dans laquelle le site de glycosylation est absent ou enlevé de la sous-unité a.
12. Utilisation suivant l'une quelconque des revendications 7 à 11, dans laquelle l'hormone est la lutéostimuline, la folliculo-stimuline, la thyroïdo-stimuline ou la gonadotrophine chorionique humaine.

Revendications pour l'Etat contractant suivant : GR

1. Procédé de préparation d'une composition pharmaceutique comprenant le mélange d'une hormone glycoprotéique modifiée conservant l'entière capacité de se fixer au récepteur que présente l'hormone sous sa forme non-modifiée, avec un porteur acceptable sur le plan pharmaceutique, l'hormone étant modifiée par le fait qu'au moins une chaîne oligosaccharidique est entièrement absente ou enlevée par le fait qu'au moins un site de glycosylation, correspondant à la chaîne oligosaccharidique absente ou enlevée, est absent ou enlevé.
2. Procédé suivant la revendication 1, dans lequel l'hormone est substantiellement incapable de déclencher une activité biologique après sa fixation au récepteur.
3. Procédé suivant la revendication 1 ou 2, dans lequel l'hormone présente au moins un résidu acide sialique.
4. Procédé suivant l'une quelconque des revendications 1 à 3, dans lequel l'hormone a une demivie plasmatique à peu près égale à celle de l'hormone naturelle non-modifiée.
5. Procédé suivant l'une quelconque des revendications 1 à 4, dans lequel le site de glycosylation est absent ou enlevé de la sous-unité a.
6. Procédé suivant l'une quelconque des revendications 1 à 5, dans lequel l'hormone est la lutéostimuline (LH), la folliculo-stimuline (FSH) la thyroïdo-stimuline (TSH) ou la gonadotrophine chorionique humaine (hCG).
7. Utilisation d'une hormone glycoprotéique modifiée conservant substantiellement l'entière capacité de fixation des récepteurs que présente l'hormone non-modifiée, l'hormone étant modifiée par le fait qu'au moins une chaîne oligosaccharidique est entièrement absente ou enlevée par le fait qu'au moins un site de glycosylation correspondant à la chaîne oligosaccharidique absente ou enlevée, est absent ou enlevé, intervenant dans la préparation d'un antagoniste hormonal non-modifié et compétitif.
8. Utilisation suivant la revendication 7, dans laquelle le site de glycosylation est absent ou enlevé de la sous-unité a.
9. Utilisation suivant la revendication 7 ou 8, dans laquelle l'hormone présente au moins un résidu acide sialique.



10. Utilisation suivant l'une quelconque des revendications 7 à 9, dans laquelle l'hormone est la lutéostimuline, la folliculo-stimuline, la thyroïdo-stimuline ou la gonadotrophine chorionique humaine.

5 11. Séquence d'ADN codant pour une hormone glycoprotéique modifiée conservant substantiellement l'entière capacité de se fixer au récepteur que présente l'hormone sous sa forme non-modifiée, l'hormone étant modifiée par le fait qu'au moins une chaîne oligosaccharidique est entièrement absente ou enlevée, par le fait qu'au moins un site de glycosylation, correspondant à la chaîne oligosaccharidique absente ou enlevée, est absent ou enlevé.

10 12. Vecteur comportant une séquence d'ADN suivant la revendication 12.

13. Hôte comportant un vecteur suivant la revendication 13.

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