

METHOD FOR SEPARATING EIGHTEEN COMPONENTS IN TRADITIONAL CHINESE MEDICINE COMPOSITION

TECHNICAL FIELD

5 The invention belongs to the field of quality analysis and control of traditional Chinese medicine, and specifically relates to a method for separating eighteen components in traditional Chinese medicine composition.

BACKGROUND OF ART

10 The traditional Chinese medicine compound is the main form of traditional Chinese medicine. After thousands of years of clinical use, the compound can obtain a stronger therapeutic effect than a single medicine, which has fully proved the scientific nature of the compound composition.

A pharmaceutical composition consisting of 13 traditional Chinese medicines, such as *Fructus Forsythiae*, *Flos Lonicerae* and *Herba Ephedrae*, has the function of clearing distemper and detoxifying, ventilating lung and releasing heat. Clinical studies have confirmed that the pharmaceutical composition has a definite and significant effect on treating influenza and acute upper respiratory infections.

In order to clarify the pharmacological action mechanism of this compound and the scientific
20 connotation of the compatibility regulation of compounded drugs, it is necessary to conduct a systematic study on its fundamentals of substances.

SUMMARY OF THE PRESENT INVENTION

The objective of the present invention is to provide a method for separating eighteen components
25 in a traditional Chinese medicine composition, on which the problem to be solved is that it is difficult to achieve rapid separation of high polar components because of the diversity of chemical components and the complex chemical structure of traditional Chinese medicine compounds. The composite reverse rapid separation technology can effectively separate the large polar components of the traditional Chinese medicine composition in a short time, and obtain a
30 series of representative compounds.

In order to achieve the above objective, a method for separating eighteen components in a traditional Chinese medicine composition is provided, the traditional Chinese medicine composition is made from crude drugs with the following part by weight: *Fructus Forsythiae* 200-300, *Herba Ephedrae* 60-100, *Radix et Rhizoma Rhei* 40-60, *Herba Houttuyniae* 200-300,

Flos Lonicerae 200-300, *Radix Isatidis* 200-300, *Herba Pogostemonis* 60-100, *Rhizoma Dryopteris Crassirhizomae* 200-300, *Radix Rhodiolae* 60-100, menthol 5-9, *Semen Armeniacae Amarum* 60-100, *Radix Glycyrrhizae* 60-100, *Gypsum Fibrosum* 200-300, wherein the method for separating includes the following steps:

5 (1) the total extract of the traditional Chinese medicine composition is separated by AB-8 macroporous resin, and then eluted with water, 10% ethanol, and 30% ethanol in sequence, and then the 30% ethanol eluate is collected and the solvent is recovered to obtain a 30% ethanol extract;

10 (2) the 30% ethanol extract obtained in step (1) is loaded to the reversed-phase silica gel ODS-AQ-HG, S-50 μ m, and mixed; after a mixed ODS is naturally dried, the mixed ODS is loaded to a sample column and the medium pressure preparation liquid phase is loaded for separation, the separation column packing is ODS-AQ-HG, S-50 μ m; successively 10% methanol is used, 5 fractions are obtained in order of elution, and numbers thereof are 10%-1, 10%-2, 10%-3, 10%-4, 10%-5; 20% methanol is used for eluting, 6 fractions are obtained in order of
15 elution, and numbers thereof are 20%-1, 20%-2, 20%-3, 20%-4, 20%-5, and 20%-6; the eluate is collected and the solvent is recovered respectively, and eluting dry extracts which are numbered 10%-1, 10%-2, 10%-3, 10%-4, 10%-5 and eluting dry extracts which are numbered 20%-1, 20%-2, 20%-3, 20%-4, 20%-5 and 20%-6 are obtained;

20 (3) the 10%-1 eluting dry extract obtained in step (2) is dissolved with 30% methanol, and the solution is filtered through a 0.45 μ m microporous membrane and preliminarily separated by high-performance liquid chromatography; the mobile phase is methanol-water 22:78, the flow rate is 1ml/min, the detection wavelength is 210nm; chromatographic peaks with retention time of 3-9min, 9-11min, 19-22min, 22-26min, 26-30min, 37-41min and 44-48min are collected, and the solvent is recovered under reduced pressure, and the following separations are performed
25 respectively:

3-9min chromatographic peak: being further purified by high performance liquid chromatography, mobile phase: methanol-water, 5:95, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 $\times$ 20 mm, S-10 μ m; under these conditions, the chromatographic peak with retention time of 25-27 min is collected, and the
30 solvent is recovered under reduced pressure to obtain compound 14: Cornoside;

9-11min chromatographic peak: being further purified by high performance liquid chromatography, mobile phase: methanol-water, 12:88, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 $\times$ 20 mm, S-10

µm; under these conditions, the chromatographic peak with retention time of 32-35min is collected, and the solvent is recovered under reduced pressure to obtain compound 10: Ferruginoside B;

19-22min chromatographic peak: being further purified by high performance liquid chromatography, mobile phase: methanol-water, 18:82, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10 µm; under these conditions, the chromatographic peak with a retention time of 31-35 min is collected, and the solvent is recovered under reduced pressure to obtain compound 9: Forsythoside E;

22-26min chromatographic peak: being further purified by high performance liquid chromatography, mobile phase: methanol-water, 16:84, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10 µm; under these conditions, the chromatographic peak with a retention time of 32-37 min is collected, and the solvent is recovered under reduced pressure to obtain compound 18: 3,4-dihydroxybenzaldehyde;

26-30min chromatographic peak: being further purified by high performance liquid chromatography, mobile phase: methanol-water, 18:82, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10 µm; under these conditions, the chromatographic peak with a retention time of 38-42 min is collected, and the solvent is recovered under reduced pressure to obtain a mixture of compound 11: D-Amygdalin and compound 12: L-Amygdalin;

37-41min chromatographic peak: being further purified by high performance liquid chromatography, mobile phase: methanol-water, 18:82, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10 µm; under these conditions, the chromatographic peak with a retention time of 52-56 min is collected, and the solvent is recovered under reduced pressure to obtain compound 13: Sambunigrin;

44-48min chromatographic peak: being further purified by high performance liquid chromatography, mobile phase: methanol-water, 22:78, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10 µm; under these conditions, the chromatographic peak with a retention time of 41-44 min is collected, and the solvent is recovered under reduced pressure to obtain compound 15: 4-Hydroxy-4-methylenecarbomethoxy-cyclohexa-2,5-dienone;

(4) the 20%-2 eluting dry extract obtained in step (2) is dissolved in 30% methanol and filtered through a 0.45 μm microporous filter membrane, and high-performance liquid chromatography is used for preliminary separation; mobile phase is methanol-water, 22:78, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 $\times$ 20 mm, S-10 μm ; chromatographic peaks with retention time of 14-17min, 17-19min, 22-24min, 29-34min, and 35-40min are collected, and the solvent is recovered under reduced pressure, and the following separations are performed respectively:

14-17min chromatographic peak: being further purified by high performance liquid chromatography, mobile phase: acetonitrile-water, 15:85, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 $\times$ 20 mm, S-10 μm ; under these conditions, the chromatographic peaks with retention time of 38-40min and 45-47min are collected, the solvent in 45-47min chromatographic peaks is recovered under reduced pressure to obtain compound 7: Lianqiaoxingan C; the solvent in 38-40 min chromatographic peaks is recovered under reduced pressure and then purified by high performance liquid chromatography, the mobile phase: acetonitrile-water, 13:87, the flow rate:10 ml/min, and the detection wavelength: 210 nm, chromatographic column: 250 $\times$ 20 mm, S-10 μm , under this condition, the chromatographic peak with retention time of 52-56min is collected, and the solvent is recovered under reduced pressure to obtain compound 6: Ferruginoside A;

17-19min chromatographic peak: being further purified by high performance liquid chromatography, mobile phase: acetonitrile-water, 15:85, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 $\times$ 20 mm, S-10 μm ; under these conditions, chromatographic peaks with retention time of 28-30 min and 37-44 min are collected, and the solvent is recovered under reduced pressure to obtain compound 16: Liriodendrin and compound 2: Forsythoside I respectively;

22-24min chromatographic peak: being further purified by high performance liquid chromatography, mobile phase: acetonitrile-water, 17:83, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 $\times$ 20 mm, S-10 μm ; under these conditions, the chromatographic peak with a retention time of 22-25 min is collected, and the solvent is recovered under reduced pressure to obtain compound 8: Calceolarioside C;

29-34min chromatographic peak: after recovering the solvent under reduced pressure, white solids are precipitated after the solvent being evaporated during the standing process, and centrifuged at 5000rpm to obtain compound 17: glycyrrhizin-7-O- β -D-glucoside; supernatant is further purified by high performance liquid chromatography, mobile phase: acetonitrile-water,

16:84, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10 μm; under these conditions, the chromatographic peak with retention time of 40-45min is collected, and the solvent is recovered under reduced pressure to obtain compound 3: Forsythoside H;

5 35-40min chromatographic peak: being further purified by high performance liquid chromatography, mobile phase: acetonitrile-water, 16:84, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10 μm; under these conditions, the chromatographic peak with retention time of 34-45 min is collected, and the solvent is recovered under reduced pressure to obtain compound 1:
10 Forsythoside A;

(5) the fraction 20%-4 and fraction 20%-5 obtained in step (2) are mixed, and is dissolved with 30% methanol and filtered through a 0.45μm microporous membrane, and high-performance liquid chromatography is used for preliminary separation; mobile phase: acetonitrile-water, 15:85, flow rate: 15ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D
15 ODS-A, 250 × 20 mm, S-10 μm; chromatographic peaks with retention time of 15-17 and 35-40 min are collected, and the solvent is recovered under reduced pressure; wherein,

15-17 min chromatographic peaks: being further purified by high performance liquid chromatography, mobile phase: acetonitrile-water, 15:85, flow rate: 12ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10
20 μm; under these conditions, chromatographic peak with retention time of 15-16 min is collected, and the solvent is recovered under reduced pressure to obtain compound 5: Isolugrandoside;

35-40min chromatographic peak: being further purified by high performance liquid chromatography, mobile phase: methanol-water, 25:75, flow rate: 12ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10
25 μm; under these conditions, the chromatographic peak with retention time of 26-29 min is collected, and the solvent is recovered under reduced pressure to obtain compound 4: Lugrandoside.

In the present invention, (1) the concentration of methanol and ethanol used in each separation step are both volume concentration (vol%); (2) the amount ratio of the methanol-water, the
30 amount ratio of the acetonitrile-water used as the mobile phase for preliminary separation by high-performance liquid chromatography are all volume ratios; (3) the chromatographic peak of each retention time collected after preliminary separation by high-performance liquid chromatography is further purified and separated by high-performance liquid chromatography, means that the samples obtained from the fractions corresponding to each chromatographic peak

with retention time after the solvent recovery are further purified.

According to the method for separating eighteen components provided by the present invention, preferably, in the step (1), the amounts of the eluent water, 10% ethanol, and 30% ethanol are 40ml eluent water, 17.6ml 10% ethanol, 45ml 30% ethanol for 1g the total extract of Chinese medicine composition.

According to the method for separating eighteen components provided by the present invention, preferably, the amount of the reversed-phase silica gel ODS-AQ-HG in step (2) is: 8g of the reversed-phase silica gel ODS-AQ-HG for 1 g of 30% ethanol extract obtained in step (1), wherein 3g is used for mixing with 1 g of 30% ethanol extract obtained in step (1), and 5g is used as the packing for the medium pressure separation column.

Preferably, in step (2), successively 10% methanol is used; 5 fractions are obtained in order of elution, and the amount of eluent for each fraction is: 66.7ml of 10% methanol for 1g of 30% ethanol extract obtained in step (1); 20% methanol is used for eluting, 6 fractions are obtained in order of elution, and the amount of eluent for each fraction is: 44.4-66.7ml of 20% methanol for 1g of 30% ethanol extract obtained in step (1).

According to the method for separating eighteen components provided by the present invention, preferably, the traditional Chinese medicine composition is made from crude drugs with the following part by weight:

Fructus Forsythiae 200, *Flos Lonicerae* 300, *Radix Isatidis* 200, *Radix et Rhizoma Rhei* 40, *Herba Pogostemonis* 60, *Rhizoma Dryopteris Crassirhizomae* 300, *Radix Rhodiolae* 100, menthol 9, *Herba Ephedrae* 60, *Semen Armeniacae Amarum* 100, *Herba Houttuyniae* 200, *Radix Glycyrrhizae* 100, *Gypsum Fibrosum* 200.

Preferably, the traditional Chinese medicine composition is made from crude drugs with the following part by weight:

Fructus Forsythiae 300, *Flos Lonicerae* 200, *Radix Isatidis* 300, *Radix et Rhizoma Rhei* 60, *Herba Pogostemonis* 100, *Rhizoma Dryopteris Crassirhizomae* 200, *Radix Rhodiolae* 60, menthol 5, *Herba Ephedrae* 100, *Semen Armeniacae Amarum* 60, *Herba Houttuyniae* 300, *Radix Glycyrrhizae* 60, *Gypsum Fibrosum* 300.

Preferably, the traditional Chinese medicine composition is made from crude drugs with the following part by weight:

Fructus Forsythiae 278, *Flos Lonicerae* 294, *Radix Isatidis* 285, *Radix et Rhizoma Rhei* 55, *Herba Pogostemonis* 95, *Rhizoma Dryopteris Crassirhizomae* 290, *Radix Rhodiolae* 87, menthol

8.5, *Herba Ephedrae* 88, *Semen Armeniacae Amarum* 80, *Herba Houltuyniae* 284, *Radix Glycyrrhizae* 95, *Plaster* 277.

According to the method for separating eighteen components provided by the present invention, preferably, the total extract of the traditional Chinese medicine composition is made by the following steps:

(1) the crude drugs are weighed according to the weight ratio, chosen for the clean ones and ground;

(2) the *Herba Pogostemonis* is ground, 10 times amount of water is added to extract the volatile oil, and extraction time of the volatile oil is 8 hours, and then the volatile oil is collected and is reserved for use; after the extract is filtered, the residue is discarded and the filtrate is reserved for use;

(3) *Fructus Forsythiae*, *Herba Ephedrae*, *Herba Houltuyniae*, and *Radix et Rhizoma Rhei* are extracted with 12 times amount of 70% ethanol for 3 times, 2.5 hours each time; extracts are mixed and filtrated, the ethanol is recovered, and filtrate is reserved for use;

(4) *Flos Lonicerae*, *Gypsum Fibrosum*, *Radix Isatidis*, *Rhizoma Dryopteris Crassirhizomae*, *Radix Glycyrrhizae* and *Radix Rhodiolae* are boiled in 12 times amount of water, then the *Semen Armeniacae Amarum* is added and boiled twice, 1 hour each time; extracts are mixed and filtered, and filtrate obtained is combined with the filtrate obtained after extracting *Herba Pogostemonis* in step (2), and condensed into a clear extract having a relative density of 1.10-1.15 at 60°C, and then ethanol is added to adjust alcohol concentration to be 70%, refrigerated (for example, 4°C), filtered, and the ethanol is recovered until no alcohol taste, the clear extract is obtained and reserved for use;

(5) the clear extract obtained in step (4) is combined with the alcohol extract obtained in step (3), and then condensed into a clear extract having relative density of 1.15-1.20 at 60°C; dried to obtain total extract and reserved for use.

The pharmaceutical composition of the present invention is made from 13 kinds of traditional Chinese medicines, such as *Fructus Forsythiae*, *Flos Lonicerae* and *Herba Ephedrae*, which has the functions of clearing distemper and detoxifying, ventilating lung and releasing heat, and is used for treating influenza. It has a definite and significant effect on treating influenza and acute upper respiratory infections. In order to elucidate the pharmacological action mechanism of this compound and the scientific connotation of the compatibility regulation of compounded drugs, it is necessary to conduct a systematic study on its fundamentals of substances.

In view of the above, the chemical ingredients of the pharmaceutical composition according to

the present invention was studied deeply (see Patent No. CN1483463 A for details), and 18 components are separated and identified in the 30% ethanol portion from the macroporous adsorption resin column of its total extract. 18 compounds are Forsythoside A (1), Forsythoside I (2), Forsythoside H (3), Lugrandoside (4), Isolugrandoside (5), Ferruginoside A (6),
5 Lianqiaoxingan C (7), Calceolarioside C (8), Forsythoside E (9), Ferruginoside B (10), D-Amygdalin (11), L-Amygdalin (12), Sambunigrin (13), Cornoside (14), 4-Hydroxy-4-methylenecarbomethoxy-cyclohexa-2,5-dienone (15), Liriodendrin (16), glycyrrhizin-7-O- β -D-glucoside (17), 3,4-diHydroxybenzaldehyde (18), respectively.

10 Among them, compound 2 to compound 8, compound 10, compound 13 to compound 18 are separated from the total extract described in the present invention for the first time; there is no report on the separation of compound 4 to compound 6, compound 10, compound 15 and compound 16 from any single drug in the compound drugs, and the nuclear magnetic data of compound 8 in DMSO-d₆ solvent is assigned for the first time.

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The beneficial effects of the technical solution in the present invention lie in:

(1) macroporous resin adsorption column chromatography, reversed-phase low-pressure column chromatography, and high-pressure preparative liquid chromatography technology are combined to achieve effective separation of large polar components in the traditional Chinese medicine
20 composition;

(2) the technical solution in the present invention has high separation efficiency of the chemical components of the traditional Chinese medicine composition, and a representative monomer compound in the compound drugs can be obtained in a short time;

(3) compared with the prior technology using a single silica gel column chromatography, the
25 technical solution in the present invention has less consumption of organic reagents and less environmental pollution during the separation process.

EMBODIMENT

In order to understand the technical features and contents of the present invention in detail,
30 preferred embodiments of the present invention will be described in more detail below. Although the preferred embodiments of the present invention are described in the examples, it should be understood that the present invention can be implemented in various forms and should not be limited by the embodiments set forth herein.

In an embodiment of the present invention, the method for separating eighteen components in a traditional Chinese medicine composition is provided, the traditional Chinese medicine composition is made from crude drugs with the following part by weight: *Fructus Forsythiae* 200-300, *Herba Ephedrae* 60-100, *Radix et Rhizoma Rhei* 40-60, *Herba Houttuyniae* 200-300, 5 *Flos Lonicerae* 200-300, *Radix Isatidis* 200-300, *Herba Pogostemonis* 60-100, *Rhizoma Dryopteris Crassirhizomae* 200-300, *Radix Rhodiolae* 60-100, menthol 5-9, *Semen Armeniacae Amarum* 60-100, *Radix Glycyrrhizae* 60-100, *Gypsum Fibrosum* 200-300, wherein the method for separating includes the following steps:

(1) the traditional Chinese medicine composition is prepared into total extract, and then the total 10 extract of the traditional Chinese medicine composition is separated by AB-8 macroporous resin, and then eluted with water, 1 vol.% ethanol, and 30 vol.% ethanol in sequence, and then the 30 vol.% ethanol eluate is collected and the solvent is recovered to obtain a 30 % ethanol extract;

(2) the 30% ethanol extract obtained in step (1) is loaded to the reversed-phase silica gel ODS-AQ-HG, S-50 μ m and mixed ODS is obtained by mixing; after a mixed ODS is naturally 15 dried, the mixed ODS is loaded to a sample column (for example, its length is 8cm and its diameter is 4cm) and the medium pressure preparation liquid phase is used for separation, the intermediate pressure separation column (for example, 40cm in length and 5cm in diameter) is packed with ODS-AQ-HG, S-50 μ m; first, 10 vol.% methanol is used, 5 fractions are obtained in order of elution, and numbers thereof are 10%-1, 10%-2, 10%-3, 10%-4, 10%-5; then, 20 vol.% 20 methanol is used for eluting, 6 fractions are obtained in order of elution, and numbers thereof are 20%-1, 20%-2, 20%-3, 20%-4, 20%-5, and 20%-6; 10% methanol eluent and 20% methanol eluent are collected respectively and the solvent is recovered respectively, and eluting dry extracts which are numbered 10%-1, 10%-2, 10%-3, 10%-4, 10%-5 and eluting dry extracts which are numbered 20%-1, 20%-2, 20%-3, 20%-4, 20%-5 and 20%-6 are obtained;

(3) the 10%-1 eluting dry extract obtained in step (2) is dissolved with 30 vol.% methanol, and the solution is filtered through a 0.45 μ m microporous membrane and preliminarily separated by high-performance liquid chromatography; the mobile phase is methanol-water 22:78, the flow rate is 1ml/min, the detection wavelength is 210nm; chromatographic peaks with retention time of 3-9min, 9-11min, 19-22min, 22-26min, 26-30min, 37-41min and 44-48min are collected, and 30 the solvent is recovered under reduced pressure, and the following separations are performed respectively (the objects of separation here are the samples from the fractions corresponding to each chromatographic peak with retention time after solvent recovery; 3-9min chromatographic peak, 9-11min chromatographic peak, 19-22min chromatographic peak, 22-26min chromatographic peak, 26-30min chromatographic peak, 37-41min chromatographic peak, and

44-48min chromatographic peak appearing below respectively correspond to the fractions of the numbers in turn: fraction 3-9, fraction 9-11, fraction 19-22, fraction 22-26, fraction 26-30, fraction 37-41, fraction 44-48):

5 the sample which is from the fraction corresponding to 3-9min chromatographic peak after solvent recovery, is further purified by high performance liquid chromatography, mobile phase: methanol-water, 5:95, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10 μm; under these conditions, the fraction corresponding to the chromatographic peak with retention time of 25-27 min is collected, and the solvent is recovered under reduced pressure to obtain compound 14: Cornoside;

10 the sample which is from the fraction corresponding to 9-11min chromatographic peak after solvent recovery, is further purified by high performance liquid chromatography, mobile phase: methanol-water, 12:88, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10 μm; under these conditions, the fraction corresponding to the chromatographic peak with retention time of 32-35min is collected, and the
15 solvent is recovered under reduced pressure to obtain compound 10: Ferruginoside B;

the sample which is from the fraction corresponding to 19-22min chromatographic peak after solvent recovery, is further purified by high performance liquid chromatography, mobile phase: methanol-water, 18:82, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10 μm; under these conditions, the fraction
20 corresponding to the chromatographic peak with a retention time of 31-35 min is collected, and the solvent is recovered under reduced pressure to obtain compound 9: Forsythoside E;

the sample which is from the fraction corresponding to 22-26min chromatographic peak after solvent recovery, is further purified by high performance liquid chromatography, mobile phase: methanol-water, 16:84, flow rate: 10ml/min, detection wavelength 210nm, chromatographic
25 column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10 μm; under these conditions, the fraction corresponding to the chromatographic peak with a retention time of 32-37 min is collected, and the solvent is recovered under reduced pressure to obtain compound 18: 3,4-dihydroxybenzaldehyde;

the sample which is from the fraction corresponding to 26-30min chromatographic peak after
30 solvent recovery, is further purified by high performance liquid chromatography, mobile phase: methanol-water, 18:82, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10 μm; under these conditions, the fraction corresponding to the chromatographic peak with a retention time of 38-42 min is collected, and

the solvent is recovered under reduced pressure to obtain a mixture of compound 11: D-Amygdalin and compound 12: L-Amygdalin;

the sample which is from the fraction corresponding to 37-41min chromatographic peak after solvent recovery, is further purified by high performance liquid chromatography, mobile phase:
5 methanol-water, 18:82, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10 μm; under these conditions, the fraction corresponding to the chromatographic peak with a retention time of 52-56 min is collected, and the solvent is recovered under reduced pressure to obtain compound 13: Sambunigrin;

the sample which is from the fraction corresponding to 44-48min chromatographic peak after
10 solvent recovery, is further purified by high performance liquid chromatography, mobile phase: methanol-water, 22:78, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10 μm; under these conditions, the fraction corresponding to the chromatographic peak with a retention time of 41-44 min is collected, and the solvent is recovered under reduced pressure to obtain compound 15:
15 4-Hydroxy-4-methylenecarbomethoxy-cyclohexa-2,5-dienone;

(4) the 20%-2 eluting dry extract obtained in step (2) is dissolved in 30 vol.% methanol and filtered through a 0.45 μm microporous filter membrane, and the high-performance liquid chromatography is used for preliminary separation; the mobile phase is methanol-water, 22:78, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D
20 ODS-A, 250 × 20 mm, S-10μm; the fractions corresponding to chromatographic peaks with retention time of 14-17min, 17-19min, 22-24min, 29-34min, and 35-40min are collected respectively, and the solvent is recovered under reduced pressure, and the following separations are performed respectively (the object of separation here are the sample which is from the fraction corresponding to each chromatographic peak with retention time after solvent recovery;
25 14-17min chromatographic peak, 17-19min chromatographic peak, 22-24min chromatographic peak, 29-34min chromatographic peaks, and 35-40min chromatographic peaks appearing below respectively correspond to the fractions of the numbers in turn: fraction 14-17, fraction 17-19, fraction 22-24, fraction 29-34, fraction 35-40):

the sample which is from the fraction corresponding to 14-17min chromatographic peak after
30 solvent recovery, is further purified by high performance liquid chromatography, mobile phase: acetonitrile-water, 15:85, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10 μm; under these conditions, the fractions correspond to the chromatographic peaks with retention time of 38-40min and 45-47min are collected, the solvent in the fraction correspond to 45-47min chromatographic peaks is recovered

under reduced pressure to obtain compound 7: Lianqiaoxingan C; the solvent in the fraction correspond to 38-40 min chromatographic peaks is recovered under reduced pressure and then purified by high performance liquid chromatography, the mobile phase: acetonitrile-water, 13:87, the flow rate:10 ml/min, and the detection wavelength: 210 nm. 250 × 20 mm, S-10 μm, under
5 this condition, the fraction correspond to the chromatographic peak with retention time of 52-56min is collected,, and the solvent is recovered under reduced pressure to obtain compound 6: Ferruginoside A;

the sample which is from the fraction corresponding to 17-19min chromatographic peak after solvent recovery, is further purified by high performance liquid chromatography, mobile phase:
10 acetonitrile-water, 15:85, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10 μm; under these conditions, the fractions correspond to chromatographic peaks with retention time of 28-30 min and 37-44 min are collected, and the solvent is recovered under reduced pressure to obtain compound 16: Liriodendrin and compound 2: Forsythoside I respectively;

15 the sample which is from the fraction corresponding to 22-24min chromatographic peak after solvent recovery, is further purified by high performance liquid chromatography, mobile phase: acetonitrile-water, 17:83, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10 μm; under these conditions, the fraction correspond to the chromatographic peak with a retention time of 22-25 min is collected, and the
20 solvent is recovered under reduced pressure to obtain compound 8: Calceolarioside C;

the sample which is from the fraction corresponding to 29-34min chromatographic peak after solvent recovery under reduced pressure, white solids are precipitated after the solvent being evaporated during the standing process, and centrifuged at 5000rpm to obtain compound 17: glycyrrhizin-7-O-β-D-glucoside; the supernatant obtained by centrifugation is further purified by
25 high performance liquid chromatography, mobile phase: acetonitrile-water, 16:84, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10 μm; under these conditions, the fraction correspond to the chromatographic peak with retention time of 40-45min is collected, and the solvent is recovered under reduced pressure to obtain compound 3: Forsythoside H;

30 the sample which is from the fraction corresponding to 35-40min chromatographic peak after solvent recovery, is further purified by high performance liquid chromatography, mobile phase: acetonitrile-water, 16:84, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10 μm; under these conditions, the fraction correspond to the chromatographic peak with retention time of 34-45 min is collected, and the

solvent is recovered under reduced pressure to obtain compound 1: Forsythoside A;

(5) the fraction 20%-4 and fraction 20%-5 obtained in step (2) are mixed, and is dissolved with 30 vol.% methanol and filtered through a 0.45 μ m microporous membrane, and the high-performance liquid chromatography is used for preliminary separation; mobile phase: acetonitrile-water, 15:85, flow rate: 15ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 $\times$ 20 mm, S-10 μ m; the fractions correspond to chromatographic peaks with retention time of 15-17 and 35-40 min are collected, and the solvent is recovered under reduced pressure; wherein,

the sample which is from the fraction corresponding to 15-17 min chromatographic peak after solvent recovery, is further purified by high performance liquid chromatography, mobile phase: acetonitrile-water, 15:85, flow rate: 12ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 $\times$ 20 mm, S-10 μ m; under these conditions, the fraction correspond to the chromatographic peak with retention time of 15-16 min is collected, and the solvent is recovered under reduced pressure to obtain compound 5: Isolugrandoside;

the sample which is from the fraction corresponding to 35-40min chromatographic peak after solvent recovery, is further purified by high performance liquid chromatography, mobile phase: methanol-water, 25:75, flow rate: 12ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 $\times$ 20 mm, S-10 μ m; under these conditions, the fraction correspond to the chromatographic peak with retention time of 26-29 min is collected, and the solvent is recovered under reduced pressure to obtain compound 4: Lugrandoside.

In some preferred embodiments, in the step (1), the amounts of the water, 10% ethanol, and 30% ethanol used as the eluent are 40ml water for 1g of the total extract of Chinese medicine composition, 17.6ml 10% ethanol for 1g of the total extract of Chinese medicine composition, 45ml 30% ethanol for 1g of the total extract of Chinese medicine composition.

In some preferred embodiments, the amount of the reversed-phase silica gel ODS-AQ-HG in step (2) is: 8g of the reversed-phase silica gel ODS-AQ-HG for 1 g of 30% ethanol extract obtained in step (1), wherein 3g of the reversed-phase silica gel ODS-AQ-HG is used for mixing with 1 g of 30% ethanol extract obtained in step (1), and the remaining 5g is used as the packing for the medium pressure separation column.

Preferably, in step (2), successively 10% methanol and 20% methanol are used to elute; when eluting with 10% methanol, 5 fractions are obtained in order of elution, and the amount of eluent for each fraction is: 66.7ml of 10% methanol for 1g of 30% ethanol extract obtained in step (1); when eluting with 20% methanol, 6 fractions are obtained in order of elution, and the amount of

eluent for each fraction is: 44.4-66.7ml of 20% methanol for 1g of 30% ethanol extract obtained in step (1).

In some embodiments, the traditional Chinese medicine composition is made from crude drugs with the following part by weight:

- 5 *Fructus Forsythiae* 200, *Flos Lonicerae* 300, *Radix Isatidis* 200, *Radix et Rhizoma Rhei* 40, *Herba Pogostemonis* 60, *Rhizoma Dryopteris Crassirhizomae* 300, *Radix Rhodiolae* 100, menthol 9, *Herba Ephedrae* 60, *Semen Armeniacae Amarum* 100, *Herba Houttuyniae* 200, *Radix Glycyrrhizae* 100, *Gypsum Fibrosum* 200.

- 10 In some embodiments, the traditional Chinese medicine composition is made from crude drugs with the following part by weight:

Fructus Forsythiae 300, *Flos Lonicerae* 200, *Radix Isatidis* 300, *Radix et Rhizoma Rhei* 60, *Herba Pogostemonis* 100, *Rhizoma Dryopteris Crassirhizomae* 200, *Radix Rhodiolae* 60, menthol 5, *Herba Ephedrae* 100, *Semen Armeniacae Amarum* 60, *Herba Houttuyniae* 300, *Radix Glycyrrhizae* 60, *Gypsum Fibrosum* 300.

- 15 In some examples, the traditional Chinese medicine composition is made from crude drugs with the following part by weight:

- Fructus Forsythiae* 278, *Flos Lonicerae* 294, *Radix Isatidis* 285, *Radix et Rhizoma Rhei* 55, *Herba Pogostemonis* 95, *Rhizoma Dryopteris Crassirhizomae* 290, *Radix Rhodiolae* 87, menthol 8.5, *Herba Ephedrae* 88, *Semen Armeniacae Amarum* 80, *Herba Houttuyniae* 284, *Radix Glycyrrhizae* 95, *Gypsum Fibrosum* 277.

The total extract of the traditional Chinese medicine composition as described above is made by the following steps:

- (1) the crude drugs are weighed according to the weight ratio, chosen for the clean ones and ground;
- 25 (2) the *Herba Pogostemonis* is further ground, and the volatile oil is extracted by adding water, which is 10 times the weight of the *Herba Pogostemonis*; the extraction time of volatile oil is 8 hours, and then the volatile oil is collected and is reserved for use; after the extract is filtered, the residue is discarded and the filtrate is reserved for use;
- 30 (3) *Fructus Forsythiae*, *Herba Ephedrae*, *Herba Houttuyniae*, and *Radix et Rhizoma Rhei* are extracted with 70wt% ethanol for 3 times, 2.5 hours each time; wherein the weight of 70% ethanol is 12 times the total weight of the *Fructus Forsythiae*, *Herba Ephedrae*, *Herba Houttuyniae*, and *Radix et Rhizoma Rhei*; then the extracts are mixed and filtrated, the ethanol is recovered, and the filtrate is reserved for use;

(4) *Flos Lonicerae*, *Gypsum Fibrosum*, *Radix Isatidis*, *Rhizoma Dryopteris Crassirhizomae*, *Radix Glycyrrhizae* and *Radix Rhodiolae* are boiled in water, wherein the weight of water is 12 times the total weight of the *Flos Lonicerae*, *Gypsum Fibrosum*, *Radix Isatidis*, *Rhizoma Dryopteris Crassirhizomae*, *Radix Glycyrrhizae* and *Radix Rhodiolae*; then the *Semen Armeniacae Amarum* is added and boiled twice, 1 hour each time; the extracts are mixed and filtered, and the filtrate obtained is combined with the filtrate obtained after extracting *Herba Pogostemonis* in step (2), and condensed into a clear extract having a relative density of 1.10-1.15 at 60°C, and then ethanol is added to adjust alcohol concentration to be 70%, refrigerated (for example, 4°C) , filtered, and the ethanol is recovered until no alcohol taste, the clear extract is obtained and reserved for use;

(5) the clear extract obtained in step (4) is combined with the alcohol extract obtained in step (3), and then condensed into a clear extract having relative density of 1.15-1.20 at 60°C; dried to obtain total extract and reserved for use.

EXAMPLES

Example 1

The traditional Chinese medicine composition is made from crude drugs with the following part by weight: 20kg of *Fructus Forsythiae*, 30kg of *Flos Lonicerae*, 20kg of *Radix Isatidis*, 4kg of *Radix et Rhizoma Rhei*, 6kg of *Herba Pogostemonis*, 30kg of *Rhizoma Dryopteris Crassirhizomae*, 10kg of *Radix Rhodiolae*, 0.9kg of menthol, 6kg of *Herba Ephedrae*, 10kg of *Semen Armeniacae Amarum*, 20kg of *Herba Houttuyniae*, 10kg of *Radix Glycyrrhizae*, 20kg of *Gypsum Fibrosum*; **the total extract of the traditional Chinese medicine composition is made by the following steps:**

- (1) the crude drugs were weighed according to the weight ratio, chosen for the clean ones and ground;
- (2) the *Herba Pogostemonis* was further ground, and the volatile oil was extracted by adding water, which was 10 times the weight of the *Herba Pogostemonis*; the extraction time of volatile oil was 8 hours, and then the volatile oil was collected and was reserved for use; after the extract was filtered, the residue was discarded and the filtrate was reserved for use;
- (3) *Fructus Forsythiae*, *Herba Ephedrae*, *Herba Houttuyniae*, and *Radix et Rhizoma Rhei* were extracted with 70wt% ethanol for 3 times, 2.5 hours each time; wherein the weight of 70wt% ethanol was 12 times the total weight of the *Fructus Forsythiae*, *Herba Ephedrae*, *Herba Houttuyniae*, and *Radix et Rhizoma Rhei*; then the extracts were mixed and filtrated, the ethanol was recovered, and the filtrate was reserved for use;

(4) *Flos Lonicerae*, *Gypsum Fibrosum*, *Radix Isatidis*, *Rhizoma Dryopteris Crassirhizomae*, *Radix Glycyrrhizae* and *Radix Rhodiolae* were boiled in water, wherein the weight of water was 12 times the total weight of the *Flos Lonicerae*, *Gypsum Fibrosum*, *Radix Isatidis*, *Rhizoma Dryopteris Crassirhizomae*, *Radix Glycyrrhizae* and *Radix Rhodiolae*; then the *Semen Armeniacae Amarum* was added and boiled twice, 1 hour each time; the extracts were mixed and filtered, and the filtrate obtained was combined with the filtrate obtained after extracting *Herba Pogostemonis* in step (2), and condensed into a clear extract having a relative density of 1.15 at 60°C, and then ethanol was added to adjust alcohol concentration to be 70wt%, refrigerated under 4°C, filtered, and the ethanol was recovered until no alcohol taste, the clear extract was obtained and reserved for use;

(5) the clear extract obtained in step (4) was combined with the alcohol extract obtained in step (3), and then condensed into a clear extract having relative density of 1.20 at 60°C; dried to obtain total extract (batch number: B1509001) of the traditional Chinese medicine composition and reserved for use.

The steps of the separation method were as follows:

1. Instruments and materials

Bruker AVIIIHD 600 NMR spectrometer (Bruker, Switzerland);
Agilent NMR VNMRS 600 Nuclear magnetic resonance spectrometer (Agilent, USA);
Synapt G2-S Mass Mass spectrometer (Waters, USA);
Combi Flash Rf Medium-low pressure preparative liquid chromatograph (Teledvne ISCO, USA);
Waters 2489-1525 Preparative liquid chromatograph (Waters, USA);
NU3000 Preparative liquid chromatograph (Jiangsu Hanbang Technology Co., Ltd.);
Milli-Q Pure water purifier (US Millipore company);
AL204 Analysis electronic balance (Mettler Toledo, USA);
AB135-S Analytical electronic balance (Mettler Toledo, USA);
TGL-16G Centrifuge (Shanghai Anting Scientific Instrument Factory);
YMC ODS-A-HG 50 µm Reversed phase silica gel (YMC Japan);
YMC-Pack R & D ODS-A (length 250 × diameter 20 mm, filler particle size S-10 µm, Japan YMC Corporation);

AB-8 Macroporous resin (Tianjin Xijinna Environmental Protection Material Technology Co., Ltd.);

The total extract of the pharmaceutical composition in the present invention (Shijiazhuang Yiling Pharmaceutical Co., Ltd., batch number: B1509001);

5 Chromatographically pure acetonitrile and methanol (Shanghai Adamas Reagent Company);

Unless otherwise specified, the chemical reagents used in the present invention are analytical reagents (Beijing Chemical Plant).

Unless otherwise specified, the selection of the above several liquid chromatographs in the high performance liquid chromatography of the examples is related to the use conditions of the
10 chromatography in the technical solution.

Extraction and separation

(1) The total extract of the pharmaceutical composition in the present invention was 5 kg (batch number: B1509001), which was adsorbed by the AB-8 macroporous resin, and then eluted with
15 200 liters of water, 88 liters of 10% ethanol, 225 liters of 30% ethanol, 250 liters of 50 % ethanol, 150 liters of 70% ethanol and 162 liters of 95% ethanol successively, and then the solvent was recovered to obtain the corresponding extracts, wherein 625.0 g was the 30% ethanol extract.

(2) 90.0 g of the 30% ethanol extract was taken and 270.0 g of reversed-phase silica gel YMC ODS-AQ-HG (S-50 μ m) was added to be mixed; after a mixed ODS was naturally dried, the
20 mixed ODS was loaded to the sample column, a Combi Flash Rf medium-low pressure preparative liquid chromatograph was used for separation (separation column size: 49 $\times$ 460mm, packing: 450g ODS-AQ-HG (S-50 μ m)), and 30 liters of 10% methanol, 32 liters of 20% methanol, 24 liters of 30% methanol, 22 liters of 35% methanol, 18 liters of 40% methanol, 18 liters of 45% methanol, 12 liters of 50% methanol, 10 liters of 70% methanol, and 6 liters of
25 anhydrous methanol were used for eluting, respectively; the solvent was recovered under reduced pressure in each elution part; 5 fractions (the solvent amount of each fraction was 6 liters) were obtained in order of elution in the 10% methanol elution part: 4.5g eluting dry extract (No. 10%-1), 4.0 g eluting dry extract (No. 10%-2), 3.5 g eluting dry extract (No. 10%-3), 2.0 g eluting dry extract (No.10%-4), and 0.9 g eluting dry extract (No. 10%-5), respectively; six
30 fractions (the solvent amount of each fraction was 6 liters) were obtained in order of elution in the 20% methanol elution part: 2.4 g eluting dry extract (No. 20%-1), 8.6 g eluting dry extract (No. 20%-2), 3.8 g eluting dry extract (No. 20%-3), 3.1 g eluting dry extract (No. 20%-4), 4.0 g eluting dry extract (No. 20%-5), and 1.7 g eluting dry extract (No. 20%-6), respectively.

(3) 2.5 g eluting dry extract with fraction number 10%-1 was taken, and was dissolved with 30% methanol, and then the solution was filtered through a 0.45 μm microporous filter membrane and preliminarily separated by high performance liquid chromatography (mobile phase: methanol-water, the volume ratio of methanol and water was 22:78, the flow rate was 10ml/min, the detection wavelength was 210nm, and the column was YMC-Pack R & D ODS-A (250 $\times$ 20 mm, S-10 μm)); the fractions corresponding to chromatographic peaks with retention time of 3-9min, 9-11min, 19-22min, 22-26min, 26-30min, 37-41min, and 44-48min were collected respectively, and the solvent was recovered under reduced pressure; the above fractions were purified by high efficient liquid chromatography, and multiple monomer compounds were obtained; the specific separation conditions were as follows:

Separation of compound 14: the sample which was from the fraction corresponding to 3-9 min chromatographic peak after solvent recovery, was further purified by high performance liquid chromatography (mobile phase: methanol-water, methanol to water volume ratio of 5:95, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A (250 $\times$ 20 mm, S-10 μm)); under these conditions, the fraction corresponding to the chromatographic peak with a retention time of 25-27min was collected, and the solvent was recovered under reduced pressure to obtain compound 14 (22 mg).

Separation of compound 10: the sample which was from the fraction corresponding to 9-11min chromatographic peak after solvent recovery, was further purified by high performance liquid chromatography (mobile phase: methanol-water, methanol: water volume ratio 12:88, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A (250 $\times$ 20 mm, S-10 μm)); under these conditions, the fraction corresponding to the chromatographic peak with a retention time of 32-35min was collected, and the solvent was recovered under reduced pressure to obtain compound 10 (15 mg).

Separation of compound 9: the sample which was from the fraction corresponding to 19-22min chromatographic peak after solvent recovery, was further purified by high-performance liquid chromatography (mobile phase: methanol-water, methanol to water volume ratio of 18:82, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A (250 $\times$ 20 mm, S-10 μm)); under these conditions, the fraction corresponding to the chromatographic peak with a retention time of 31-35min was collected, and the solvent was recovered under reduced pressure to obtain compound 9 (79 mg).

Separation of compound 18: the sample which was from the fraction corresponding to 22-26min chromatographic peak after solvent recovery, was further purified by high performance liquid chromatography (mobile phase: methanol-water, methanol to water volume ratio of 16:84, flow

rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A (250 × 20 mm, S-10 μm)); under these conditions, the fraction corresponding to the chromatographic peak with a retention time of 32-37min was collected, and the solvent was recovered under reduced pressure to obtain compound 18 (8 mg).

- 5 Separation of compound 11 and compound 12: the sample which was from the fraction corresponding to 26-30min chromatographic peak after solvent recovery, was further purified by high-performance liquid chromatography (mobile phase: methanol-water, the volume ratio of methanol to water was 18:82, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A (250 × 20 mm, S-10 μm)); under these
10 conditions, the fraction corresponding to the chromatographic peak with a retention time of 38-42min was collected, and the solvent was recovered under reduced pressure to obtain a mixture of compounds 11 and 12 (159 mg).

- Separation of compound 13: the sample which was from the fraction corresponding to 37-41min chromatographic peak after solvent recovery, was recovered by high-performance liquid
15 chromatography (mobile phase: methanol-water, methanol: water volume ratio 18:82, flow rate: 10ml)/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A (250 × 20 mm, S-10 μm)); under these conditions, the fraction corresponding to the chromatographic peak with a retention time of 52-56min was collected, and the solvent was recovered under reduced pressure to obtain compound 13 (8 mg).

- 20 Separation of compound 15: the sample which was from the fraction corresponding to 44-48min chromatographic peak after solvent recovery, was further purified by high-performance liquid chromatography (mobile phase: methanol-water, methanol: water volume ratio 22:78, flow rate: 10ml)/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A (250 × 20 mm, S-10 μm)); under these conditions, the fraction corresponding to the
25 chromatographic peak with a retention time of 41-44min was collected, and the solvent was recovered under reduced pressure to obtain compound 15 (10 mg).

- (4) 2.9 g of eluting dry extract with fraction number 20%-2 was taken, and was dissolved with 30% methanol, and then the solution was filtered through a 0.45 μm microporous filter membrane and preliminarily separated by high-performance liquid chromatography (mobile phase:
30 methanol-water, the volume ratio of methanol and water was 22:78, the flow rate was 10ml/min, the detection wavelength was 210nm, and the column was YMC-Pack R & D ODS-A (250 × 20 mm, S-10 μm)); the fractions corresponding to chromatographic peaks with retention time of 14-17min, 17-19min, 22-24min, 29-34min, and 35-40min were collected respectively, and the solvent was recovered under reduced pressure; the above fractions were purified by high efficient

liquid chromatography, and multiple monomer compounds were obtained; the specific separation conditions were as follows:

Separation of compound 6 and compound 7: the sample which was from the fraction corresponding to 14-17min chromatographic peak after solvent recovery was further purified by high-performance liquid chromatography (mobile phase: acetonitrile-water, the volume ratio of acetonitrile to water was 15:85, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A (250 × 20 mm, S-10 μm)); under these conditions, the fractions corresponding to chromatographic peaks with retention time of 38-40min and 45-47min were collected, and the solvent was recovered under reduced pressure to obtain the fractions corresponding to a chromatographic peak of 38-40 min and compound 7 (4 mg); the fraction corresponding to the chromatographic peak of 38-40 min was purified by high performance liquid chromatography (mobile phase: acetonitrile-water, the volume ratio of acetonitrile to water was 13:87, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A (250 × 20 mm, S-10 μm)); under these conditions, the fraction corresponding to the chromatographic peak at 52-56 min was collected, and the solvent was recovered under reduced pressure to obtain compound 6 (4 mg).

Separation of compound 16 and compound 2: the sample which was from the fraction corresponding to 17-19min chromatographic peak after solvent recovery was further purified by high performance liquid chromatography (mobile phase: acetonitrile-water, the volume ratio of acetonitrile to water was 15:85, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A (250 × 20 mm, S-10 μm)); under these conditions, the fractions corresponding to chromatographic peaks with retention time of 28-30min and 37-44min were collected, and the solvents were recovered under reduced pressure to obtain compound 16 (3 mg) and compound 2 (117 mg).

Separation of compound 8: the sample which was from the fraction corresponding to 22-24min chromatographic peak after solvent recovery was further purified by high performance liquid chromatography (mobile phase: acetonitrile-water, acetonitrile to water volume ratio 17:83, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A (250 × 20 mm, S-10 μm)); under these conditions, the fraction corresponding to the chromatographic peak with a retention time of 22-25min was collected, and the solvent was recovered under reduced pressure to obtain compound 8 (36 mg).

Separation of compound 17 and compound 3: the sample which was from the fraction corresponding to 29-34min chromatographic peak after solvent recovery under reduced pressure was left at room temperature; after the solvent evaporated, a white solid was precipitated and

centrifuged at 5000 rpm to obtain compound 17 (10mg). The supernatant was further purified by high-performance liquid phase (mobile phase: acetonitrile-water, acetonitrile to water volume ratio of 16:84, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A (250 × 20 mm, S-10 μm)); under these conditions, the fraction
5 corresponding to the chromatographic peak with a retention time of 40-45min was collected, and the solvent was recovered under reduced pressure to obtain compound 3 (19 mg).

Separation of Compound 1: the sample which was from the fraction corresponding to 35-40min chromatographic peak was further purified by high performance liquid chromatography (mobile phase: acetonitrile-water, acetonitrile to water volume ratio of 16:84, flow rate: 10ml/min,
10 detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A (250 × 20 mm, S-10 μm)); under these conditions, the fraction corresponding to the chromatographic peak with a retention time of 34-45min was collected, and the solvent was recovered under reduced pressure to obtain Compound 1 (161 mg).

(5) Separation of compound 4 and compound 5: the fraction numbered 20%-4 and fraction
15 numbered 20%-5 were combined, and 2.0g of the mixed fraction was dissolved with 30% methanol, and then was filtered through a 0.45 μm microporous filter membrane and preliminarily separated by high performance liquid chromatography (mobile phase: acetonitrile-water, volume ratio of acetonitrile to water: 15:85, flow rate: 15ml/min, detection wavelength: 210nm, chromatographic column: YMC-pack R&D ODS-A (250 × 20 mm, S-10
20 μm)); under these conditions, the fractions corresponding to chromatographic peaks with retention time of 13-15 min and 15-17 min were collected, respectively, and the solvent was recovered under reduced pressure.

The sample which was from the fraction corresponding to 35-40min chromatographic peak after solvent recovery was further purified by high performance liquid chromatography (mobile phase:
25 methanol-water, methanol to water volume ratio of 25:75, flow rate: 12ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A (250 × 20 mm, S-10 μm)); under these conditions, the fraction corresponding to the chromatographic peak with a retention time of 26-29min was collected, and the solvent was recovered under reduced pressure to obtain compound 4 (32 mg).

30 The sample which was from the fraction corresponding to 15-17 min chromatographic peak after solvent recovery was further purified by high performance liquid chromatography (mobile phase: acetonitrile-water, acetonitrile to water volume ratio of 15:85, flow rate: 12ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A (250 × 20 mm, S-10 μm)); under these conditions, the fraction corresponding to the chromatographic peak with a

retention time of 15-16min was collected, and the solvent was recovered under reduced pressure to obtain compound 5 (4 mg).

Structure Identification

5 Compound **1**: pale green glassy solid (methanol), ESI-MS m/z : 623.2 $[M-H]^-$, and the molecular formula of the compound was determined to be $C_{29}H_{36}O_{15}$ based on NMR data.

1H -NMR (DMSO- d_6 , 600 MHz) δ_H : 6.63 (1H, d, J = 1.9 Hz, H-2), 6.64 (1H, d, J = 8.1 Hz, H-5), 6.50 (1H, dd, J = 8.1, 1.9 Hz, H-6), 2.69 (2H, m, H-7), 4.31 (1H, d, J = 7.8 Hz, H-1'), 4.51 (1H, br s, H-1''), 1.05 (1H, d, J = 6.2 Hz, H-6''), 7.05 (1H, br s, H-2'''), 6.77 (1H, d, J = 8.1 Hz, H-5'''), 7.00 (1H, br s, H-6'''), 7.50 (1H, d, J = 15.8 Hz, H-7'''), 6.26 (1H, d, J = 15.8 Hz, H-8''').

^{13}C -NMR (DMSO- d_6 , 150 MHz) δ_C : 129.3 (C-1), 115.6 (C-2), 145.1 (C-3), 143.6 (C-4), 116.4 (C-5), 119.6 (C-6), 35.2 (C-7), 70.4 (C-8), 103.0 (C-1'), 73.1 (C-2'), 73.6 (C-3'), 71.0 (C-4'), 74.0 (C-5'), 66.2 (C-6'), 100.6 (C-1''), 70.7 (C-2''), 70.4 (C-3''), 71.9 (C-4''), 68.5 (C-5''), 17.9 (C-6''), 125.6 (C-1'''), 115.0 (C-2'''), 145.7 (C-3'''), 148.6 (C-4'''), 115.9 (C-5'''), 121.5 (C-6'''), 145.7 (C-7'''), 113.9 (C-8'''), 166.0 (C-9''').

The above hydrogen spectrum characteristics and carbon spectrum data were basically consistent with the report in reference [1], and the compound was identified as Forsythoside A.

Compound **2**: pale green glassy solid (methanol), ESI-MS m/z : 623.2 $[M-H]^-$, and the molecular formula of the compound was determined to be $C_{29}H_{36}O_{15}$ based on NMR data.

20 1H -NMR (DMSO- d_6 , 600 MHz) δ_H : 6.62 (1H, d, J = 1.8 Hz, H-2), 6.64 (1H, d, J = 8.1 Hz, H-5), 6.50 (1H, dd, J = 8.1, 1.8 Hz, H-6), 2.69 (2H, m, H-7), 4.35 (1H, d, J = 7.7 Hz, H-1'), 4.60 (1H, br s, H-1''), 1.14 (1H, d, J = 6.2 Hz, H-6''), 7.05 (1H, br s, H-2'''), 6.77 (1H, d, J = 8.0 Hz, H-5'''), 7.01 (1H, dd, J = 8.0, 1.7 Hz, H-6'''), 7.47 (1H, d, J = 15.8 Hz, H-7'''), 6.28 (1H, d, J = 15.8 Hz, H-8''').

25 ^{13}C -NMR (DMSO- d_6 , 150 MHz) δ_C : 129.3 (C-1), 115.6 (C-2), 144.9 (C-3), 143.6 (C-4), 116.4 (C-5), 119.7 (C-6), 35.2 (C-7), 70.3 (C-8), 102.8 (C-1'), 71.5 (C-2'), 77.6 (C-3'), 68.3 (C-4'), 75.2 (C-5'), 66.6 (C-6'), 100.8 (C-1''), 70.8 (C-2''), 70.6 (C-3''), 72.0 (C-4''), 68.5 (C-5''), 18.1 (C-6''), 125.8 (C-1'''), 114.9 (C-2'''), 145.7 (C-3'''), 148.4 (C-4'''), 115.9 (C-5'''), 121.3 (C-6'''), 145.1 (C-7'''), 114.8 (C-8'''), 166.2 (C-9''').

30 The above hydrogen spectrum characteristics and carbon spectrum data were basically consistent with those reported in reference [1], and the compound was identified as Forsythoside I.

Compound **3**: transparent glassy solid (methanol), ESI-MS m/z : 623.2 $[M-H]^-$, and the molecular formula of the compound was determined to be $C_{29}H_{36}O_{15}$ based on NMR data.

1H -NMR (DMSO- d_6 , 600 MHz) δ_H : 6.54 (1H, br s, H-2), 6.55 (1H, d, J = 8.2 Hz, H-5), 6.41

(1H, dd, $J = 8.2, 2.0$ Hz, H-6), 2.56 (2H, m, H-7), 4.49 (1H, d, $J = 8.1$ Hz, H-1'), 4.60 (1H, br s, H-1''), 1.14 (1H, d, $J = 6.8$ Hz, H-6''), 7.06 (1H, d, $J = 1.9$ Hz, H-2'''), 6.77 (1H, d, $J = 8.2$ Hz, H-5'''), 7.02 (1H, dd, $J = 8.2, 1.9$ Hz, H-6'''), 7.49 (1H, d, $J = 15.8$ Hz, H-7'''), 6.28 (1H, d, $J = 15.8$ Hz, H-8''').

5 ^{13}C -NMR (DMSO- d_6 , 150 MHz) δ_{C} : 129.1 (C-1), 115.4 (C-2), 144.8 (C-3), 143.4 (C-4), 116.2 (C-5), 119.5 (C-6), 35.0 (C-7), 69.8 (C-8), 100.2 (C-1'), 73.4 (C-2'), 74.1 (C-3'), 70.6 (C-4'), 75.4 (C-5'), 66.6 (C-6'), 100.7 (C-1''), 70.4 (C-2''), 70.3 (C-3''), 71.9 (C-4''), 68.4 (C-5''), 17.9 (C-6''), 125.6 (C-1'''), 114.9 (C-2'''), 145.5 (C-3'''), 148.3 (C-4'''), 115.8 (C-5'''), 121.2 (C-6'''), 144.8 (C-7'''), 114.2 (C-8'''), 165.6 (C-9''').

10 The above hydrogen spectrum characteristics and carbon spectrum data were basically consistent with those reported in reference [2], and the compound was identified as Forsythoside H.

Compound **4**: transparent glassy solid (methanol), ESI-MS m/z : 639.2 $[\text{M-H}]^-$, and the molecular formula of the compound was determined to be $\text{C}_{29}\text{H}_{36}\text{O}_{16}$ based on NMR data.

^1H -NMR (DMSO- d_6 , 600 MHz) δ_{H} : 6.64 (1H, o, H-2), 6.64 (1H, o, H-5), 6.51 (1H, br d, $J = 7.3$ Hz, H-6), 2.69 (2H, m, H-7), 4.31 (1H, d, $J = 7.5$ Hz, H-1'), 4.19 (1H, d, $J = 7.7$ Hz, H-1''), 7.06 (1H, br s, H-2'''), 6.77 (1H, o, H-5'''), 7.01 (1H, d, $J = 7.1$ Hz, H-6'''), 7.49 (1H, d, $J = 15.8$ Hz, H-7'''), 6.26 (1H, d, $J = 15.8$ Hz, H-8''').

^{13}C -NMR (DMSO- d_6 , 150 MHz) δ_{C} : 129.7 (C-1), 116.8 (C-2), 145.4 (C-3), 140.0 (C-4), 116.3 (C-5), 120.0 (C-6), 35.5 (C-7), 71.7 (C-8), 103.1 (C-1'), 73.9 (C-2'), 74.4 (C-3'), 70.7 (C-4'), 20 73.7 (C-5'), 68.6 (C-6'), 103.7 (C-1''), 74.0 (C-2''), 77.0 (C-3''), 70.4 (C-4''), 77.3 (C-5''), 61.4 (C-6''), 125.9 (C-1'''), 115.4 (C-2'''), 146.1 (C-3'''), 149.1 (C-4'''), 116.0 (C-5'''), 121.9 (C-6'''), 145.4 (C-7'''), 114.2 (C-8'''), 166.7 (C-9''').

The above carbon spectrum data were basically consistent with the report in reference [3], and the compound was identified as Lugrandoside.

25 Compound **5**: transparent glassy solid (methanol), ESI-MS m/z : 639.2 $[\text{M-H}]^-$, and the molecular formula of the compound was determined to be $\text{C}_{29}\text{H}_{36}\text{O}_{16}$ based on NMR data.

^1H -NMR (DMSO- d_6 , 600 MHz) δ_{H} : 6.64 (2H, br s, H-2, 5), 6.52 (1H, dd, $J = 8.0, 1.7$ Hz, H-6), 2.69 (2H, m, H-7), 4.36 (1H, d, $J = 7.7$ Hz, H-1'), 4.25 (1H, d, $J = 7.7$ Hz, H-1''), 7.06 (1H, br s, H-2'''), 6.78 (1H, d, $J = 7.9$ Hz, H-5'''), 7.01 (1H, d, $J = 7.9$ Hz, H-6'''), 7.48 (1H, d, $J = 15.9$ Hz, H-7'''), 6.30 (1H, d, $J = 15.9$ Hz, H-8''').

^{13}C -NMR (DMSO- d_6 , 150 MHz) δ_{C} : 129.8 (C-1), 116.8 (C-2), 145.3 (C-3), 143.9 (C-4), 116.3 (C-5), 120.1 (C-6), 35.5 (C-7), 70.6 (C-8), 103.0 (C-1'), 71.9 (C-2'), 78.0 (C-3'), 68.3 (C-4'), 75.8 (C-5'), 68.5 (C-6'), 103.8 (C-1''), 74.0 (C-2''), 77.3 (C-3''), 70.5 (C-4''), 77.1 (C-5''), 61.5 (C-6''), 126.1 (C-1'''), 116.0 (C-2'''), 145.4 (C-3'''), 148.7 (C-4'''), 116.3 (C-5'''), 121.7 (C-6'''),

146.0 (C-7'''), 115.2 (C-8'''), 166.6 (C-9''').

The above hydrogen spectrum characteristics and carbon spectrum data were basically consistent with those reported in reference [4], and the compound was identified as Isolugrandoside.

Compound 6: light brown glassy solid (methanol), ESI-MS m/z : 639.2 $[M-H]^-$, and the molecular formula of the compound was determined to be $C_{29}H_{36}O_{16}$ based on NMR data.

1H -NMR (DMSO- d_6 , 600 MHz) δ_H : 6.55 (1H, br s, H-2), 6.54 (1H, d, $J = 8.0$ Hz, H-5), 6.42 (1H, dd, $J = 8.0, 1.9$ Hz, H-6), 2.56 (2H, m, H-7), 4.49 (1H, d, $J = 8.1$ Hz, H-1'), 4.24 (1H, d, $J = 7.8$ Hz, H-1''), 7.06 (1H, br s, H-2'''), 6.77 (1H, d, $J = 8.1$ Hz, H-5'''), 7.02 (1H, dd, $J = 8.1, 1.9$ Hz, H-6'''), 7.49 (1H, d, $J = 15.8$ Hz, H-7'''), 6.28 (1H, d, $J = 15.8$ Hz, H-8''').

^{13}C -NMR (DMSO- d_6 , 150 MHz) δ_C : 129.3 (C-1), 115.9 (C-2), 145.0 (C-3), 143.5 (C-4), 116.3 (C-5), 119.7 (C-6), 35.1 (C-7), 69.9 (C-8), 100.2 (C-1'), 74.3 (C-2'), 75.8 (C-3'), 70.2 (C-4'), 73.5 (C-5'), 68.4 (C-6'), 103.5 (C-1''), 73.6 (C-2''), 77.0 (C-3''), 70.1 (C-4''), 76.8 (C-5''), 61.2 (C-6''), 125.7 (C-1'''), 115.5 (C-2'''), 145.7 (C-3'''), 148.4 (C-4'''), 115.5 (C-5'''), 121.4 (C-6'''), 145.2 (C-7'''), 115.0 (C-8'''), 165.8 (C-9''').

The above hydrogen spectrum characteristics and carbon spectrum data were basically consistent with those reported in reference [4].

Compound 7: light brown glassy solid (methanol), ESI-MS m/z : 609.2 $[M-H]^-$, and the molecular formula of the compound was determined to be $C_{28}H_{34}O_{15}$ based on NMR data.

1H -NMR (DMSO- d_6 , 600 MHz) δ_H : 6.63 (1H, brs, H-2), 6.62 (1H, d, $J = 8.0$ Hz, H-5), 6.50 (1H, dd, $J = 8.0, 2.0$ Hz, H-6), 2.69 (2H, m, H-7), 4.33 (1H, d, $J = 7.7$ Hz, H-1'), 4.89 (1H, t, $J = 9.4$ Hz, H-3'), 4.19 (1H, d, $J = 7.6$ Hz, H-1''), 7.05 (1H, br s, H-2'''), 6.77 (1H, d, $J = 8.0$ Hz, H-5'''), 7.01 (1H, dd, $J = 8.0, 1.9$ Hz, H-6'''), 7.47 (1H, d, $J = 15.7$ Hz, H-7'''), 6.29 (1H, d, $J = 15.7$ Hz, H-8''').

^{13}C -NMR (DMSO- d_6 , 150 MHz) δ_C : 129.3 (C-1), 116.5 (C-2), 145.1 (C-3), 143.6 (C-4), 115.9 (C-5), 119.7 (C-6), 35.2 (C-7), 70.3 (C-8), 102.7 (C-1'), 71.5 (C-2'), 77.6 (C-3'), 67.9 (C-4'), 75.4 (C-5'), 68.1 (C-6'), 104.2 (C-1''), 73.4 (C-2''), 76.7 (C-3''), 69.7 (C-4''), 65.8 (C-5''), 125.8 (C-1'''), 114.8 (C-2'''), 145.7 (C-3'''), 148.4 (C-4'''), 115.6 (C-5'''), 121.3 (C-6'''), 145.1 (C-7'''), 114.9 (C-8'''), 166.3 (C-9''').

The characteristics of the above hydrogen spectrum were basically consistent with those reported in reference [5]. The structure of the compound was further confirmed by the two-dimensional spectrum, and the carbon signal was assigned to identify the compound as Lianqiaoxingan C.

Compound 8: light brown glassy solid (methanol), ESI-MS m/z : 609.2 $[M-H]^-$, and the molecular formula of the compound was determined to be $C_{28}H_{34}O_{15}$ based on NMR data.

1H -NMR (DMSO- d_6 , 600 MHz) δ_H : 6.64 (1H, brs, H-2), 6.63 (1H, d, $J = 8.0$ Hz, H-5), 6.51

(1H, dd, $J = 8.0, 1.8$ Hz, H-6), 2.70 (2H, m, H-7), 3.62, 3.89 (2H, m, H-8), 4.30 (1H, d, $J = 8.1$ Hz, H-1'), 4.64 (1H, t, $J = 9.8$ Hz, H-4'), 4.15 (1H, d, $J = 7.6$ Hz, H-1''), 7.06 (1H, br s, H-2'''), 6.77 (1H, d, $J = 7.9$ Hz, H-5'''), 7.02 (1H, dd, $J = 7.9, 1.4$ Hz, H-6'''), 7.49 (1H, d, $J = 15.8$ Hz, H-7'''), 6.27 (1H, d, $J = 15.8$ Hz, H-8''').

5 ^{13}C -NMR (DMSO- d_6 , 150 MHz) δ_{C} : 129.3 (C-1), 115.6 (C-2), 145.7 (C-3), 143.6 (C-4), 116.4 (C-5), 119.6 (C-6), 35.2 (C-7), 70.3 (C-8), 102.8 (C-1'), 74.0 (C-2'), 73.6 (C-3'), 71.3 (C-4'), 73.3 (C-5'), 68.2 (C-6'), 103.9 (C-1''), 73.4 (C-2''), 76.5 (C-3''), 69.6 (C-4''), 65.8 (C-5''), 125.6 (C-1'''), 115.0 (C-2'''), 145.0 (C-3'''), 148.6 (C-4'''), 115.9 (C-5'''), 121.6 (C-6'''), 145.8 (C-7'''), 113.9 (C-8'''), 166.3 (C-9''').

10 The above hydrogen spectrum characteristics and carbon spectrum data were basically consistent with those reported in reference [6]. The carbon and hydrocarbon signals were assigned by two-dimensional nuclear magnetic data, and the compound was identified as Calceolarioside C.

Compound **9**: transparent glassy solid (methanol), ESI-MS m/z : 461.2 $[\text{M-H}]^-$, and the molecular formula of the compound was determined to be $\text{C}_{20}\text{H}_{30}\text{O}_{12}$ based on NMR data.

15 ^1H -NMR (DMSO- d_6 , 400 MHz) δ_{H} : 6.59 (1H, d, $J = 3.2$ Hz, H-2), 6.61 (1H, d, $J = 8.0$ Hz, H-5), 6.47 (1H, dd, $J = 8.0, 2.0$ Hz, H-6), 2.64 (2H, t, $J = 7.6$ Hz, H-7), 1.11 (3H, d, $J = 6.4$ Hz, H-6'').

^{13}C -NMR (DMSO- d_6 , 100 MHz) δ_{C} : 129.3 (C-1), 116.3 (C-2), 144.9 (C-3), 143.5 (C-4), 115.5 (C-5), 119.5 (C-6), 35.2 (C-7), 70.1 (C-8), 103.0 (C-1'), 73.4 (C-2'), 76.7 (C-3'), 70.2 (C-4'), 20 75.4 (C-5'), 67.0 (C-6'), 100.8 (C-1''), 70.7 (C-2''), 70.5 (C-3''), 72.0 (C-4''), 68.4 (C-5''), 18.0 (C-6'').

The above hydrogen spectrum characteristics and carbon spectrum data were basically consistent with those reported in reference [7], and the compound was identified as Forsythoside E.

25 Compound **10**: transparent glassy solid (methanol), ESI-MS m/z : 477.2 $[\text{M-H}]^-$, and the molecular formula of the compound was determined to be $\text{C}_{20}\text{H}_{30}\text{O}_{13}$ based on NMR data.

^1H -NMR (DMSO- d_6 , 600 MHz) δ_{H} : 6.60 (1H, d, $J = 1.8$ Hz, H-2), 6.61 (1H, d, $J = 8.2$ Hz, H-5), 6.47 (1H, dd, $J = 8.2, 1.8$ Hz, H-6), 4.15 (1H, d, $J = 7.8$ Hz, H-1'), 4.22 (1H, d, $J = 7.8$ Hz, H-1').

30 ^{13}C -NMR (DMSO- d_6 , 150 MHz) δ_{C} : 129.3 (C-1), 116.3 (C-2), 144.9 (C-3), 143.5 (C-4), 115.5 (C-5), 119.5 (C-6), 35.1 (C-7), 70.0 (C-8), 102.8 (C-1'), 73.4 (C-2'), 76.7 (C-3'), 69.9 (C-4'), 75.7 (C-5'), 68.4 (C-6'), 103.3 (C-1''), 73.4 (C-2''), 76.9 (C-3''), 70.0 (C-4''), 76.7 (C-5''), 61.0 (C-6'').

The above hydrogen spectrum characteristics and carbon spectrum data were basically consistent with those reported in reference [8], and the compound was identified as Ferruginoside B.

Compound **11** and Compound **12** were white powdery solids (methanol), and the nuclear magnetic resonance spectrum showed that the component to be measured was a mixture of a pair of epimers.

Comparing the NMR data with those reported in reference [9], it can be confirmed that this
5 component was a mixture of D-Amygdalin and L-Amygdalin, and the molecular weight of the mixture was ESI-MS m/z : 456.2 $[M-H]^-$.

The NMR data of compound **11** D-Amygdalin was assigned as follows: 1H -NMR (DMSO- d_6 , 400 MHz) δ_H : 7.55 (2H, o, H-4, 8), 7.48 (3H, o, H-5, 6, 7), 5.99 (1H, s, H-2), 4.20-4.60 (2H, o, H-1', 1''); ^{13}C -NMR (DMSO- d_6 , 100 MHz) δ_C : 118.9 (C-1), 68.5 (C-2), 133.9 (C-3), 129.6 (C-6),
10 127.3 (C-5,7), 129.0 (C-4,8), 103.7 (C-1'), 73.1 (C-2'), 76.8 (C-3'), 70.1 (C-4'), 76.6 (C-5'), 66.8 (C-6'), 101.6 (C-1''), 73.8 (C-2''), 76.8 (C-3''), 70.1 (C-4''), 76.5 (C-5''), 61.1 (C-6').

The NMR data of compound **12** L-Amygdalin was assigned as follows: 1H -NMR (DMSO- d_6 , 400 MHz) δ_H : 7.55 (2H, o, H-4, 8), 7.48 (3H, o, H-5,6,7), 5.99 (1H, s, H-2), 4.20-4.60 (2H, o, H-1', 1''); ^{13}C -NMR (DMSO- d_6 , 100 MHz) δ_C : 118.2 (C-1), 69.0 (C-2), 133.8 (C-3), 128.9 (C-4,8), 127.5
15 (C-5,7), 129.6 (C-6), 104.0 (C-1'), 72.9 (C-2'), 76.8 (C-3'), 70.3 (C-4'), 76.5 (C-5'), 67.1 (C-6'), 101.0 (C-1''), 73.6 (C-2''), 76.8 (C-3''), 70.0 (C-4''), 76.2 (C-5''), 61.1 (C-6').

Compound **13**: amorphous solid (methanol) ESI-MS m/z : 340.1 $[M + HCOO]^-$. The molecular formula of the compound was determined to be $C_{14}H_{17}NO_6$ based on NMR data.

1H -NMR (DMSO- d_6 , 600 MHz) δ_H : 7.56 (2H, d, $J = 7.7$ Hz, H-2,6), 7.48 (2H, o, H-3,5),
20 7.48 (1H, o, H-4), 6.03 (1H, s, H-7), 4.19 (1H, d, $J = 6.6$ Hz, H-1'), 3.70 (1H, br d, $J = 11.6$ Hz, H-6'a), 3.51 (1H, br d, $J = 11.0$ Hz, H-6'b), 3.09 (4H, o, H-2', H-3', H-4', H-5'); ^{13}C -NMR (DMSO- d_6 , 150 MHz) δ_C : 134.4 (C-1), 128.1 (C-2,6), 129.7 (C-3,5), 130.3 (C-4), 67.3 (C-7), 119.5 (C-8), 101.8 (C-1'), 73.9 (C-2'), 77.2 (C-3'), 70.6 (C-4'), 78.0 (C-5'), 61.8 (C-6').

The above hydrogen spectrum characteristics and carbon spectrum data were basically consistent
25 with those reported in the reference [10], and the compound was identified as Sambunigrin.

Compound **14**: amorphous solid (methanol), ESI-MS m/z : 315.1 $[M-H]^-$, and the molecular formula of the compound was determined to be $C_{14}H_{20}O_8$ based on NMR data.

1H -NMR (DMSO- d_6 , 600 MHz) δ_H : 6.04 (2H, d, $J = 10.1$ Hz, H-2, 6), 6.97 (2H, dd, $J = 10.1$ Hz, 2.6 Hz H-3, 5), 4.07 (1H, d, $J = 7.8$ Hz, H-1'), 1.89 (2H, m, H-1'').
30 ^{13}C -NMR (DMSO- d_6 , 150 MHz) δ_C : 185.3 (C-1), 126.4 (C-2, 6), 153.4 (C-3 or C-5), 153.3 (C-3 or C-5), 67.3 (C-4), 39.7 (C-1'), 63.9 (C-2'), 102.9 (C-1''), 73.4 (C-2''), 76.9 (C-3''), 70.0 (C-4''), 76.7 (C-5''), 61.0 (C-6'').

The above hydrogen spectrum characteristics and carbon spectrum data were basically consistent with those reported in reference [11], and the compound was identified as Cornoside.

Compound **15**: amorphous solid (methanol), ESI-MS m/z : 195.1 $[M-H]^-$, and the molecular formula of the compound was determined to be $C_{10}H_{12}O_4$ based on NMR data.

1H -NMR (DMSO- d_6 , 600 MHz) δ_H : 7.02 (2H, br d, $J = 10.0$ Hz H-2, 6), 6.08 (2H, br d, $J = 10.0$ Hz, H-3, 5), 2.69 (2H, s, H-1'), 4.01 (2H, q, $J = 14.2$ Hz, 7.1 Hz, H-3'), 1.13 (3H, td, $J = 7.1$ Hz, 1.4 Hz, H-4').

^{13}C -NMR (DMSO- d_6 , 150 MHz) δ_C : 185.2 (C-1), 126.8 (C-2, C-6), 151.7 (C-3, C-5), 66.6 (C-4), 44.9 (C-1'), 168.6 (C-2'), 60.2 (C-3'), 14.1 (C-4').

The above hydrogen spectrum characteristics and carbon spectrum data were basically consistent with those reported in reference [12], and the compound was identified as 4-Hydroxy-4-methylenecarbomethoxy-cyclohexa-2,5-dienone.

Compound **16**: white crystal (methanol), ESI-MS m/z : 787.2 $[M + HCOO]^-$, 765.3 $[M + Na]^+$, and the molecular formula of the compound was determined to be $C_{34}H_{46}O_{18}$ by NMR data.

1H -NMR (DMSO- d_6 , 600 MHz) δ_H : 3.76 (12H, s, OCH_3 -2, 2', 6, 6'), 6.66 (4H, s, H-3, 3', 5, 5'), 4.67 (2H, d, $J = 3.8$ Hz, H-7, 7'), 4.20 (2H, m, H-9 α , 9' α), 3.83 (2H, dd, $J = 9.2, 3.4$ Hz, H-9 β , 9' β).

^{13}C -NMR (DMSO- d_6 , 150 MHz) δ_C : 137.3 (C-1, 1'), 152.8 (C-2, 2', 6, 6'), 104.3 (C-3, 3', 5, 5'), 133.8 (C-4, 4'), 85.2 (C-7, 7'), 53.7 (C-8, 8'), 71.5 (C-9, 9'), 56.6 (OCH_3 -2, 2', 6, 6'), 102.8 (C-1'', 1'''), 74.3 (C-2'', 2'''), 77.3 (C-3'', 3'''), 70.0 (C-4'', 4'''), 76.6 (C-5'', 5'''), 61.0 (C-6'', 6''').

The above hydrogen spectrum characteristics and carbon spectrum data were basically consistent with those reported in reference [13,14], and the compound was identified as Liriodendrin.

Compound **17**: white powder (methanol), ESI-MS m/z : 417.1 $[M-H]^-$, and the molecular formula of the compound was determined to be $C_{21}H_{22}O_9$ based on NMR data.

1H -NMR (DMSO- d_6 , 600 MHz) δ_H : 5.53 (1H, dd, $J = 12.7, 2.8$ Hz, H-2), 7.65 (1H, d, $J = 8.6$ Hz, H-5), 6.35 (1H, d, $J = 2.2$ Hz, H-8), 6.51 (1H, dd, $J = 8.6, 2.2$ Hz, H-6), 7.45 (2H, d, $J = 8.7$ Hz, H-2', 6'), 7.07 (2H, d, $J = 8.8$ Hz, H-3', 5'), 3.15 (1H, o, H-3a) 2.68 (1H, dd, $J = 16.7, 2.9$ Hz, H-3b), 3.2-3.5 (4H, m, , 2'', 3'', 4'', 5''), 4.89 (1H, d, $J = 7.5$ Hz, H-6''a), 3.70 (1H, d, $J = 12.0$ Hz, H-6''b). ^{13}C -NMR (DMSO- d_6 , 150 MHz) δ_C : 78.8 (C-2), 43.3 (C-3), 190.0 (C-4), 132.5 (C-5), 110.7 (C-6), 164.8 (C-7), 113.6 (C-8), 163.2 (C-9), 102.7 (C-10), 128.5 (C-1'), 128.1 (C-2', 6'), 116.3 (C-3', 5'), 157.6 (C-4'), 100.4 (C-1''), 73.3 (C-2''), 76.7 (C-3''), 69.8 (C-4''), 77.2 (C-5''), 60.8 (C-6'').

The above hydrogen spectrum characteristics and carbon spectrum data were basically consistent with the report in reference [15], and the compound was identified as glycyrrhizin-7- O - β -D-glucoside.

Compound **18**: light yellow amorphous solid (methanol), ESI-MS m/z : 137.0 $[M-H]^-$, and the

molecular formula of the compound was determined to be C₇H₆O₃ based on the NMR data.

¹H-NMR (DMSO-*d*₆, 600 MHz) δ_H : 9.68 (CHO), 7.22 (1H, br s, H-2), 6.89 (1H, d, *J* = 8.4 Hz, H-3), 7.25 (1H, br d, *J* = 7.8 Hz, H-4).

¹³C-NMR (DMSO-*d*₆, 150 MHz) δ_C : 191.1 (CHO), 128.8 (C-1), 115.5 (C-2), 145.9 (C-3),
5 152.2 (C-4), 114.3 (C-5), 124.5 (C-6).

The above hydrogen spectrum characteristics and carbon spectrum data were basically consistent with the reports in reference [16,17], and the compound was identified as 3,4-dihydroxybenzaldehyde.

10 Example 2

The traditional Chinese medicine composition is made from crude drugs with the following part by weight: *Fructus Forsythiae* 30kg, *Flos Lonicerae* 20kg, *Radix Isatidis* 30kg, *Radix et Rhizoma Rhei* 6kg, *Herba Pogostemonis* 10kg, *Rhizoma Dryopteris Crassirhizomae* 20kg, *Radix Rhodiolae* 6kg, menthol 0.5kg, *Herba Ephedrae* 10kg, *Semen Armeniacae Amarum* 6kg, *Herba*
15 *Houttuyniae* 30kg, *Radix Glycyrrhizae* 6kg, *Gypsum Fibrosum* 30kg; **the total extract of the traditional Chinese medicine composition was made by the following steps:**

(1) the crude drugs were weighed according to the weight ratio, chosen for the clean ones and ground;

(2) the *Herba Pogostemonis* was further ground, and the volatile oil was extracted by adding
20 water, which was 10 times the weight of the *Herba Pogostemonis*; the extraction time of volatile oil was 8 hours, and then the volatile oil was collected and was reserved for use; after the extract was filtered, the residue was discarded and the filtrate was reserved for use;

(3) *Fructus Forsythiae*, *Herba Ephedrae*, *Herba Houttuyniae*, and *Radix et Rhizoma Rhei* were extracted with 70wt% ethanol for 3 times, 2.5 hours each time; wherein the weight of 70wt%
25 ethanol was 12 times the total weight of the *Fructus Forsythiae*, *Herba Ephedrae*, *Herba Houttuyniae*, and *Radix et Rhizoma Rhei*; then the extracts were mixed and filtrated, the ethanol was recovered, and the filtrate was reserved for use;

(4) *Flos Lonicerae*, *Gypsum Fibrosum*, *Radix Isatidis*, *Rhizoma Dryopteris Crassirhizomae*, *Radix Glycyrrhizae* and *Radix Rhodiolae* were boiled in water, wherein the weight of water was
30 12 times of the total weight of the *Flos Lonicerae*, *Gypsum Fibrosum*, *Radix Isatidis*, *Rhizoma Dryopteris Crassirhizomae*, *Radix Glycyrrhizae* and *Radix Rhodiolae*; then the *Semen Armeniacae Amarum* was added and boiled twice, 1 hour each time; the extracts were mixed and filtered, and the filtrate obtained was combined with the filtrate obtained after extracting *Herba*

Pogostemonis in step (2), and condensed into a clear extract having a relative density of 1.10 at 60°C, and then ethanol was added to adjust alcohol concentration to be 70wt%, refrigerated under 4°C, filtered, and the ethanol was recovered until no alcohol taste, the clear extract was obtained and reserved for use;

- 5 (5) the clear extract obtained in step (4) was combined with the alcohol extract obtained in step (3), and then condensed into a clear extract having relative density of 1.15 measured at 60°C; and dried to obtain the total extract and reserved for use.

The separation method steps were the same as those in Example 1. As a result, the eighteen compounds described in Example 1 were separated.

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Example 3

The traditional Chinese medicine composition is made from crude drugs with the following part by weight: 27.8 kg of *Fructus Forsythiae*, 29.4 kg of *Flos Lonicerae*, 28.5 kg of *Radix Isatidis*, 5.5 kg of *Radix et Rhizoma Rhei*, 9.5 kg of *Herba Pogostemonis*, 29 kg of *Rhizoma Dryopteris*
15 *Crassirhizomae*, 8.7 kg of *Radix Rhodiolae*, 0.85kg of menthol, 8.8kg of *Herba Ephedrae*, 8kg of *Semen Armeniacae Amarum*, 28.4kg of *Herba Houttuyniae*, 9.5kg of *Radix Glycyrrhizae*, 27.7kg of *Gypsum Fibrosum*; **the total extract of the traditional Chinese medicine composition was made by the following steps:**

- (1) the crude drugs were weighed according to the weight ratio, chosen for the clean ones and
20 ground;

(2) the *Herba Pogostemonis* was further ground, and the volatile oil was extracted by adding water, which was 10 times the weight of the *Herba Pogostemonis*; the extraction time of volatile oil was 8 hours, and then the volatile oil was collected and was reserved for use; after the extract was filtered, the residue was discarded and the filtrate was reserved for use;

- 25 (3) *Fructus Forsythiae*, *Herba Ephedrae*, *Herba Houttuyniae*, and *Radix et Rhizoma Rhei* were extracted with 70wt% ethanol for 3 times, 2.5 hours each time; wherein the weight of 70wt% ethanol was 12 times the total weight of the *Fructus Forsythiae*, *Herba Ephedrae*, *Herba Houttuyniae*, and *Radix et Rhizoma Rhei*; then the extracts were mixed and filtrated, the ethanol was recovered, and the filtrate was reserved for use;

- 30 (4) *Flos Lonicerae*, *Gypsum Fibrosum*, *Radix Isatidis*, *Rhizoma Dryopteris Crassirhizomae*, *Radix Glycyrrhizae* and *Radix Rhodiolae* were boiled in water, wherein the weight of water was 12 times of the total weight of the *Flos Lonicerae*, *Gypsum Fibrosum*, *Radix Isatidis*, *Rhizoma Dryopteris Crassirhizomae*, *Radix Glycyrrhizae* and *Radix Rhodiolae*; then the *Semen*

Armeniacae Amarum was added and boiled twice, 1 hour each time; the extracts were mixed and filtered, and the filtrate obtained was combined with the filtrate obtained after extracting *Herba Pogostemonis* in step (2), and condensed into a clear extract having a relative density of 1.13 at 60°C, and then ethanol was added to adjust alcohol concentration to be 70wt%, refrigerated under 4°C, filtered, and the ethanol was recovered until no alcohol taste, the clear extract was obtained and reserved for use;

(5) the clear extract obtained in step (4) was combined with the alcohol extract obtained in step (3), and then condensed into a clear extract having relative density of 1.18 at 60°C; dried to obtain the total extract and reserved for use.

The separation method steps were the same as those in Example 1. As a result, the eighteen compounds described in Example 1 were separated.

The references mentioned in the examples are as follows:

- [1] Fan Yi, Chen Ling, Zhu Jie, *et al.*, Chemical compositions of Forsythia leaf [J]. Chinese Journal of Experimental Traditional Medical Formulae, 2015, 12 (24): 22-25.
- [2] Wang F N, Ma Z Q, Liu Y, *et al.*, New Phenylethanoid Glycosides from the Fruits of *Forsythia Suspense* (Thunb.) Vahl [J]. *Molecules*, 2009, 14 (3): 1324-1331.
- [3] Zheng Xiaoke, Li Jun, Feng Sanitary, *et al.*, Studies on the phenylethanol glycosides of Lithocholic grass [J]. Chinese Traditional and Herbal Drugs, 2002, 33 (10): 19-21.
- [4] Lin Sheng, Liu Mingtao, Wang Sujuan, *et al.*, Powder glycosides and phenylethanol glycosides of the *Ligustrum sinense* [J]. China Journal of Traditional Chinese medicine, 2010, 35 (8): 992-996.
- [5] Xia Y G , Yang B Y , Liang J, *et al.*, Caffeoyl Phenylethanoid Glycosides from Unripe Fruits of *Forsythia Suspensai* [J]. *CHEM NAT COMPD+*, 2015, 51 (4): 656-659.
- [6] Nicoletti M, Galeffi C, Messana I, *et al.*, Phenylpropanoid glycosides from *Calceolaria hypericina* [J]. *Phytochemistry*, 1988, 27 (2): 639-641.
- [7] Endo K, Hikino H. Structures of rengyol, rengyoxide and rengyolone, new cyclohexylethane derivatives from *Forsythia suspensa* fruits [J]. *Can. J. Chem.*, 1984, 62 (9): 2011-2014.
- [8] Calis I, Tasdemir D, Sticher O, *et al.*, Phenylethanoid Glycosides from *Digitalis ferruginea* subsp. *farruginea* (=D.aurea LINDLEY)(Scrophulariaceae) [J]. *Chem. Pharm. Bull.*, 1999, 47 (9): 1305-1307.
- [9] Xiao Wei, Xie Xue, Song Yaling, *et al.*, Preparation method of Amygdalin: China, CN105461765A [P], 2016-04-06.
- [10] Liu Hua, Shen Juanjuan, Zhang Dongming, *et al.*, Studies on the polar components of

Elsholtzia haichowensis Sun [J]. China Journal of Experimental Traditional Medical Formulae, 2010, 16 (8): 84-86.

[11] Hong M L, Jin K K, Jai M J, *et al.*, Analysis of the inhibitory activity of *Abeliophyllum distichum* leaf constituents against aldose reductase by using high-speed counter current chromatography[J]. *Arch. Pharm. Res.*, 2013, 36(9): 1104–1112.

[12] Muhammad P, Ahmad S, Nawaz H R , *et al.*, New acetylated quinols from *Ajuga parviflora* [J]. *Fitoterapia*, 1999, 70 (3): 229-232.

[13] Lami N, Kadota S, Kikuchi T, *et al.*, Constituents of the roots of *Boerhaavia diffusa* L. III. Identification of Ca²⁺ channel antagonistic compound from the methanol extract [J]. *Chem. Pharm. Bull.*, 1991, 39 (6): 1551-1555.

[14] He Jun, Zhang Xunjie, Ma Bingzhi, *et al.*, Studies on chemical constituents of *Solanum lyratum thunb* [J]. Chinese Pharmaceutical Journal, 2015, 50 (23): 2035-2038.

[15] Jin Q H, Lee C, Lee J W, *et al.*, Chemical Constituents from the Fruits of *Prunus mume* [J]. *Nat. Prod. Sci.*, 2012, 18 (3): 200-203.

[16] Kang H S, Choi J H, Cho W K, *et al.*, A Sphingolipid and Tyrosinase Inhibitors from the Fruiting Body of *Phellinus linteus* [J]. *Arch. Pharm. Res.*, 2004, 27 (7): 742-750.

[17] Trimbak B M, Tushar M K, Ramakrishna G B. Oxidative decarboxylation of arylacetic acids in water: One-pot transition-metal-free synthesis of aldehydes and ketones[J]. *Tetrahedron Letters*, 2017, 58(29): 2822-2825.

The examples of the present invention have been described above, the above description is exemplary, not exhaustive, and is not limited to the disclosed examples. Many modifications and variations will be apparent to those skilled in the art without departing from the scope and spirit of the various examples described.

Claims

1. A method for separating eighteen components in a traditional Chinese medicine composition, the traditional Chinese medicine composition is made from crude drugs with the following part by weight: *Fructus Forsythiae* 200-300, *Herba Ephedrae* 60-100, *Radix et Rhizoma Rhei* 40-60, *Herba Houttuyniae* 200-300, *Flos Lonicerae* 200-300, *Radix Isatidis* 200-300, *Herba Pogostemonis* 60-100, *Rhizoma Dryopteris Crassirhizomae* 200-300, *Radix Rhodiolae* 60-100, menthol 5-9, *Semen Armeniacae Amarum* 60-100, *Radix Glycyrrhizae* 60-100, *Gypsum Fibrosum* 200-300, wherein the method for separating includes the following steps:
- (1) the total extract of the traditional Chinese medicine composition is separated by AB-8 macroporous resin, and then eluted with water, 10% ethanol, and 30% ethanol in sequence, and then the 30% ethanol eluate is collected and the solvent is recovered to obtain a 30% ethanol extract;
- (2) the 30% ethanol extract obtained in step (1) is loaded to the reversed-phase silica gel ODS-AQ-HG, S-50 μ m, and mixed; after a mixed ODS is naturally dried, the mixed ODS is loaded to a sample column, and the medium pressure preparation liquid phase is loaded for separation, the separation column packing is ODS-AQ-HG, S-50 μ m; successively 10% methanol is used, 5 fractions are obtained in order of elution, and numbers thereof are 10%-1, 10%-2, 10%-3, 10%-4, 10%-5; 20% methanol is used for eluting, 6 fractions are obtained in order of elution, and numbers thereof are 20%-1, 20%-2, 20%-3, 20%-4, 20%-5, and 20%-6; the eluate is collected and the solvent is recovered respectively, and eluting dry extracts which are numbered 10%-1, 10%-2, 10%-3, 10%-4, 10%-5 and eluting dry extracts which are numbered 20%-1, 20%-2, 20%-3, 20%-4, 20%-5 and 20%-6 are obtained;
- (3) the 10%-1 eluting dry extract obtained in step (2) is dissolved with 30% methanol, and the solution is filtered through a 0.45 μ m microporous membrane and preliminarily separated by high-performance liquid chromatography; the mobile phase is methanol-water 22:78, the flow rate is 1ml/min, the detection wavelength is 210nm; chromatographic peaks with retention time of 3-9min, 9-11min, 19-22min, 22-26min, 26-30min, 37-41min and 44-48min are collected, and the solvent is recovered under reduced pressure; and the following separations are performed respectively:
- 3-9min chromatographic peak: being further purified by high performance liquid chromatography, mobile phase: methanol-water, 5:95, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 $\times$ 20 mm, S-10 μ m; under these conditions, the chromatographic peak with retention time of 25-27 min is collected, and the solvent is recovered under reduced pressure to obtain compound 14: Cornoside;

9-11min chromatographic peak: being further purified by high performance liquid chromatography, mobile phase: methanol-water, 12:88, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10 μm; under these conditions, the chromatographic peak with retention time of 32-35min is collected, and the solvent is recovered under reduced pressure to obtain compound 10: Ferruginoside B;

19-22min chromatographic peak: being further purified by high performance liquid chromatography, mobile phase: methanol-water, 18:82, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10 μm; under these conditions, the chromatographic peak with a retention time of 31-35 min is collected, and the solvent is recovered under reduced pressure to obtain compound 9: Forsythoside E;

22-26min chromatographic peak: being further purified by high performance liquid chromatography, mobile phase: methanol-water, 16:84, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10 μm; under these conditions, the chromatographic peak with a retention time of 32-37 min is collected, and the solvent is recovered under reduced pressure to obtain compound 18: 3,4-dihydroxybenzaldehyde;

26-30min chromatographic peak: being further purified by high performance liquid chromatography, mobile phase: methanol-water, 18:82, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10 μm; under these conditions, the chromatographic peak with a retention time of 38-42 min is collected, and the solvent is recovered under reduced pressure to obtain a mixture of compound 11: D-Amygdalin and compound 12: L-Amygdalin;

37-41min chromatographic peak: being further purified by high performance liquid chromatography, mobile phase: methanol-water, 18:82, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10 μm; under these conditions, the chromatographic peak with a retention time of 52-56 min is collected, and the solvent is recovered under reduced pressure to obtain compound 13: Sambunigrin;

44-48min chromatographic peak: being further purified by high performance liquid chromatography, mobile phase: methanol-water, 22:78, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10

μm; under these conditions, the chromatographic peak with a retention time of 41-44 min is collected, and the solvent is recovered under reduced pressure to obtain compound 15: 4-Hydroxy-4-methylenecarbomethoxy-cyclohexa-2,5-dienone;

(4) the 20%-2 eluting dry extract obtained in step (2) is dissolved in 30% methanol and filtered
5 through a 0.45 μm microporous filter membrane, and high-performance liquid chromatography is used for preliminary separation; mobile phase is methanol-water, 22:78, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10μm; chromatographic peaks with retention time of 14-17min, 17-19min, 22-24min, 29-34min, and 35-40min are collected, and the solvent is recovered under reduced pressure, and
10 the following separations are performed respectively:

14-17min chromatographic peak: being further purified by high performance liquid chromatography, mobile phase: acetonitrile-water, 15:85, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10 μm; under these conditions, the chromatographic peaks with retention time of 38-40min and
15 45-47min are collected, the solvent in 45-47min chromatographic peaks is recovered under reduced pressure to obtain compound 7: Lianqiaoxingan C; the solvent in 38-40 min chromatographic peaks is recovered under reduced pressure and then purified by high performance liquid chromatography, the mobile phase: acetonitrile-water, 13:87, the flow rate:10 ml/min, and the detection wavelength: 210 nm, chromatographic column: 250 × 20 mm, S-10 μm,
20 under this condition, the chromatographic peak with retention time of 52-56min is collected, and the solvent is recovered under reduced pressure to obtain compound 6: Ferruginoside A;

17-19min chromatographic peak: being further purified by high performance liquid chromatography, mobile phase: acetonitrile-water, 15:85, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10
25 μm; under these conditions, chromatographic peaks with retention time of 28-30 min and 37-44 min are collected, and the solvent is recovered under reduced pressure to obtain compound 16: Liriodendrin and compound 2: Forsythoside I respectively;

22-24min chromatographic peak: being further purified by high performance liquid chromatography, mobile phase: acetonitrile-water, 17:83, flow rate: 10ml/min, detection
30 wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10 μm; under these conditions, the chromatographic peak with a retention time of 22-25 min is collected, and the solvent is recovered under reduced pressure to obtain compound 8: Calceolarioside C;

29-34min chromatographic peak: after recovering the solvent under reduced pressure, white solids are precipitated during standing process, and centrifuged at 5000rpm to obtain compound 17: glycyrrhizin-7-O-β-D-glucoside; supernatant is further purified by high performance liquid chromatography, mobile phase: acetonitrile-water, 16:84, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10 μm; under these conditions, the chromatographic peak with retention time of 40-45min is collected, and the solvent is recovered under reduced pressure to obtain compound 3: Forsythoside H;

35-40min chromatographic peak: being further purified by high performance liquid chromatography, mobile phase: acetonitrile-water, 16:84, flow rate: 10ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10 μm; under these conditions, the chromatographic peak with retention time of 34-45 min is collected, and the solvent is recovered under reduced pressure to obtain compound 1: Forsythoside A;

(5) the fraction 20%-4 and fraction 20%-5 obtained in step (2) are mixed, and is dissolved with 30% methanol and filtered through a 0.45μm microporous membrane, and high-performance liquid chromatography is used for preliminary separation; mobile phase: acetonitrile-water, 15:85, flow rate: 15ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10 μm; chromatographic peaks with retention time of 15-17 and 35-40 min are collected, and the solvent is recovered under reduced pressure; wherein,

15-17 min chromatographic peak: being further purified by high performance liquid chromatography, mobile phase: acetonitrile-water, 15:85, flow rate: 12ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10 μm; under these conditions, chromatographic peak with retention time of 15-16 min is collected, and the solvent is recovered under reduced pressure to obtain compound 5: Isolugrandoside;

35-40min chromatographic peak: being further purified by high performance liquid chromatography, mobile phase: methanol-water, 25:75, flow rate: 12ml/min, detection wavelength 210nm, chromatographic column: YMC-Pack R & D ODS-A, 250 × 20 mm, S-10 μm; under these conditions, the chromatographic peak with retention time of 26-29 min is collected, and the solvent is recovered under reduced pressure to obtain compound 4: Lugrandoside.

2. The method for separating eighteen components according to claim 1, wherein in the step (1), the amounts of the eluent water, 10% ethanol, and 30% ethanol are 40ml eluent water, 17.6ml 10%

ethanol, 45ml 30% ethanol for 1g the total extract of Chinese medicine composition.

3. The method for separating eighteen components according to claim 1, wherein the amount of the reversed-phase silica gel ODS-AQ-HG in step (2) is: 8g of the reversed-phase silica gel ODS-AQ-HG for 1 g of 30% ethanol extract obtained in step (1), wherein 3g is used for mixing with 1 g of 30% ethanol extract obtained in step (1), and 5g is used as the packing for the medium pressure separation column.

4. The method for separating eighteen components according to claim 1, wherein in step (2), successively 10% methanol is used; 5 fractions are obtained in order of elution, and the amount of eluent for each fraction is: 66.7ml of 10% methanol for 1g of 30% ethanol extract obtained in step (1); 20% methanol is used for eluting, 6 fractions are obtained in order of elution, and the amount of eluent for each fraction is: 44.4-66.7ml of 20% methanol for 1g of 30% ethanol extract obtained in step (1).

5. The method for separating eighteen components according to any one of claims 1-4, wherein the traditional Chinese medicine composition is made from crude drugs with the following part by weight:

Fructus Forsythiae 200, *Flos Lonicerae* 300, *Radix Isatidis* 200, *Radix et Rhizoma Rhei* 40, *Herba Pogostemonis* 60, *Rhizoma Dryopteris Crassirhizomae* 300, *Radix Rhodiolae* 100, menthol 9, *Herba Ephedrae* 60, *Semen Armeniacae Amarum* 100, *Herba Houttuyniae* 200, *Radix Glycyrrhizae* 100, *Gypsum Fibrosum* 200.

6. The method for separating eighteen components according to any one of claims 1-4, wherein the traditional Chinese medicine composition is made from crude drugs with the following part by weight:

Fructus Forsythiae 300, *Flos Lonicerae* 200, *Radix Isatidis* 300, *Radix et Rhizoma Rhei* 60, *Herba Pogostemonis* 100, *Rhizoma Dryopteris Crassirhizomae* 200, *Radix Rhodiolae* 60, menthol 5, *Herba Ephedrae* 100, *Semen Armeniacae Amarum* 60, *Herba Houttuyniae* 300, *Radix Glycyrrhizae* 60, *Gypsum Fibrosum* 300.

7. The method for separating eighteen components according to any one of claims 1-4, wherein the traditional Chinese medicine composition is made from crude drugs with the following part by weight:

5 *Fructus Forsythiae* 278, *Flos Lonicerae* 294, *Radix Isatidis* 285, *Radix et Rhizoma Rhei* 55, *Herba Pogostemonis* 95, *Rhizoma Dryopteris Crassirhizomae* 290, *Radix Rhodiolae* 87, menthol 8.5, *Herba Ephedrae* 88, *Semen Armeniacae Amarum* 80, *Herba Houttuyniae* 284, *Radix Glycyrrhizae* 95, Plaster 277.

10 8. The method for separating eighteen components according to any one of claims 1-4, wherein the total extract of the traditional Chinese medicine composition is made by the following steps:

(1) the crude drugs are weighed according to the weight ratio, chosen for the clean ones and ground;

(2) the *Herba Pogostemonis* is ground, 10 times amount of water is added to extract the volatile oil, and extraction time of the volatile oil is 8 hours, and then the volatile oil is collected and is reserved for use; after extract is filtered, the residue is discarded and the filtrate is reserved for use;

(3) *Fructus Forsythiae*, *Herba Ephedrae*, *Herba Houttuyniae*, and *Radix et Rhizoma Rhei* are extracted with 12 times amount of 70% ethanol for 3 times, 2.5 hours each time; extracts are mixed and filtrated, the ethanol is recovered, and filtrate is reserved for use;

20 (4) *Flos Lonicerae*, *Gypsum Fibrosum*, *Radix Isatidis*, *Rhizoma Dryopteris Crassirhizomae*, *Radix Glycyrrhizae* and *Radix Rhodiolae* are boiled in 12 times amount of water, then the *Semen Armeniacae Amarum* is added and boiled twice, 1 hour each time; extracts are mixed and filtered, and filtrate obtained is combined with the filtrate obtained after extracting *Herba Pogostemonis* in step (2), and condensed into a clear extract having a relative density of 1.10-1.15 at 60°C, and then ethanol is added to adjust alcohol concentration to be 70%, refrigerated, filtered, and the ethanol is recovered until no alcohol taste, the clear extract is obtained and reserved for use;

25 (5) the clear extract obtained in step (4) is combined with the alcohol extract obtained in step (3), and then condensed into a clear extract having relative density of 1.15-1.20 at 60°C; dried to obtain total extract and reserved for use.

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