1. A method of producing aggregates of non-genetically modified non-human-animal-derived pluripotent stem cells (PSCs) comprising the steps of:

    a. seeding at least one PSC in an expansion medium, the expansion medium is a serum-free liquid medium comprising a combination of the growth factor bFGF, at least one additional growth factor of the transforming growth factor beta (TGF-β, TGFB) superfamily and at least one small molecule selected from the group consisting of an inhibitor of the Wnt-β-catenin signaling pathway selected from the group consisting of IWR1, JW67, NSC668036, KY02111, niclosamide, DKK1 or Beta-Catenin siRNA, porcupine inhibitor, and IWP-2; PD 0325901 (C16H14F3IN2O4); and A 83-01 (C25H19N5S)-1; to form a seeding suspension culture; and

    b. growing the suspension culture under conditions enabling aggregate formation and aggregate expansion, thereby forming homogenous aggregates of said PSCs.

2. The method of claim 1, wherein the seeding and growing is within a vessel having walls of a material having essentially no cell adherence capacity.

3. The method of claim 1, wherein the at least one PSCs is a fresh cell or a cell obtained from a frozen stock.

4. The method of any one of claims 1-3, wherein step (b) of growing the suspension culture comprises the steps of disaggregating the formed homogenous aggregates into smaller aggregates and/or single cells and re-aggregating the smaller aggregates and/or single cells to re-form homogenous aggregates.

5. The method of claim 4, wherein disaggregating the homogenous aggregates comprises exposing said aggregates to a dissociation reagent and/or to a dissociation force.

6. The method of any one of claims 4-5, wherein re-aggregating comprises seeding the smaller aggregates and/or single cells in the expansion medium.

7. The method of any one of claims 4-6, wherein the steps of disaggregating the formed homogenous aggregates into smaller aggregates and/or single cells and re-aggregating the smaller aggregates and/or single cells are repeated at least once.

8. The method of claim 7, wherein the steps of disaggregating the formed homogenous

aggregates into smaller aggregates and/or single cells and re-aggregating the smaller aggregates and/or single cells are repeated until a cell concentration of from 10^6 cell/ml to 10^9 cells/ml is reached.

9. The method of any one of claims 1-8, wherein the steps of seeding and re-aggregating further comprise adding an inhibitor of Rho-associated protein kinase (Rock) to the expansion medium.

10. The method of any one of claims 1-9, said method is performed in a closed system.

11. The method of any one of claims 1-10, wherein the inhibitor of the Wnt-β-catenin signaling pathway is selected from the group consisting of IWR1, JW67, NSC668036, KY02111, niclosamide, DKK1 or si-Beta-Catenin, porcupine inhibitor, and IWP-2.

12. The method of claim 11, wherein the inhibitor of the Wnt-β-catenin signaling pathway is IWR1.

13. The method of any one of claims 1-12, wherein the expansion medium comprises a combination comprising bFGF and at least one small molecule.

14. The method of claims 13, wherein the expansion medium comprises a combination consisting of the growth factor bFGF and the small molecule IWR1.

15. The method of any one of claims 1-12, wherein the at least one additional growth factor is a protein of the transforming growth factor beta (TGF-β) superfamily.

16. The method of claim 15, wherein the TGF-β is selected from the group consisting of TGF-β-1, TGFβ-3, Activin-A and any combination thereof.

17. The method of claim 16, wherein the expansion medium comprises a combination comprising bFGF and at least one of TGF-β-1, TGFβ-3, and Activin-A.

18. The method of claim 16, wherein the expansion medium comprises a combination consisting of the growth factors bFGF, at least one of TGF-β-1, TGFβ-3, and Activin-A, and the small molecule is IWR1.

19. The method of any one of claims 1-18, wherein step (a) of seeding the PSCs further comprises encapsulating the at least one PSCs within a particle and seeding a plurality of the particles within the expansion medium.

20. The method of claim 19, wherein the particle comprises an inner core comprising the at least one PSC surrounded by at least one outer shell layer.

21. The method of claim 20, wherein the at least one outer shell layer is a fluid permeable food grade, dissolvable material.

22. The method of claim 21, wherein the food grade, dissolvable material is selected from group consisting of a polymer, a polymer precursor, a copolymer, and any combination thereof.

23. The method of any one of claims 19-22, wherein the inner core further comprises at least one compound supporting proliferation of the at least one non-genetically modified non-human-animal-derived PSCs.

24. The method of claim 23, wherein the at least one compound supporting proliferation is formulated in a slow-release and/or suspended release formulation.

25. The method of any one of claims 19-24, wherein the encapsulated PSCs are proliferating within the inner core to form PSC aggregates within the particle.

26. The method of any one of claims 1-25, wherein the suspension culture is incubated under dynamic rotation, static conditions or a combination thereof.

27. The method of any one of claims 1-26, wherein the suspension culture is grown at a temperature which is the body temperature of the non-human-animal species from which the cells are derived.

28. The method of any one of claims 1-27, wherein the volume of the suspension culture is from 50 ml to 15,000 liters per vessel.

29. The method of any one of claims 1-28, wherein the PSC aggregates formed have an average diameter of from 50µm to 500µm.

30. The method of any one of claims 1-29, wherein the non-human-animal is selected from the group consisting of ungulate, poultry, aquatic animals, invertebrate and reptiles.

31. The method of claim 30, wherein the ungulate is selected from the group consisting of a bovine, an ovine, an equine, a pig, a giraffe, a camel, a deer, a hippopotamus, or a rhinoceros.

32. The method of any one of claims 30-31, wherein the non-human-animal PSCs are selected from the group consisting of non-genetically modified induced PSCs (iPSCs) and embryonic stem cells (ESCs)).

33. The method of claim 32, wherein the non-human-animal is bovine, and wherein the

bovine PSCs are embryonic stem cells produced by a method comprising the steps of:

i. obtaining at least one bovine pre-embryo;

ii. culturing the at least one pre-embryo to reach blastocyst or enhanced blastocyst stage;

iii. obtaining at least one cell from the blastocyst; and

iv. culturing the at least one cell in a culture medium comprising a combination of the growth factor bFGF, at least one additional growth factor of the transforming growth factor beta (TGF-β, TGFB) superfamily and at least one small molecule selected from the group consisting of an inhibitor of the Wnt-β-catenin signaling pathway selected from the group consisting of IWR1, JW67, NSC668036, KY02111, niclosamide, DKK1 or Beta-Catenin siRNA, porcupine inhibitor, and IWP-2; PD 0325901 (C16H14F3IN2O4); and A 83-01 (C25H19N5S)-1; to obtain a plurality of bovine-derived embryonic stem cells.

34. The method of claim 33, wherein the least one bovine pre-embryo is obtained by embryo flushing procedure.

35. The method of claim 32, wherein the iPSCs are produced by a method comprising introducing into at least one non-human-animal-derived cell a combination of:

a. at least one reprogramming mRNA encoding reprogramming factor; and

b. at least one double-stranded microRNA;

wherein the at least one reprogramming mRNA is not integrated into the genome of the cell, thereby producing at least one iPSC.

36. The method of claim 35, said method further comprises introducing into the at least one non-human-animal-derived cell at least one immune evasion mRNA.

37. The method of any one of claims 35-36, wherein the double-stranded microRNA is selected from the group consisting of miR-302a, miR-302b, miR-302c, miR-302d, miR-367, miR-218, miR-449b and any combination thereof.

38. The method of claim 32, wherein the iPSCs are produced by a method comprising introducing into at least one non-human-animal-derived cell a combination of:

a. at least one reprogramming mRNA encoding reprogramming factor; and

b. at least one inhibitor of at least one microRNA endogenous to the non-human-animal -derived cell;

thereby producing at least one iPSC.

39. The method of claim 38, said method further comprises introducing into the at least one non-human-animal -derived cell at least one immune evasion mRNA.

40. The method of any one of claims 38-39, wherein the at least one inhibitor is directed to miR-145.

41. The method of any one of claims 38-40, wherein the at least one inhibitor is an RNA inhibitory (RNAi) molecule.

42. The method of any one of claims 36-41, wherein introducing the at least one reprogramming mRNA, and/or the at least one immune evasion mRNA and/or the at least one double-stranded microRNA and/or the at least one miRNA inhibitor is performed in a serum-free liquid expansion medium, and wherein said at least one mRNA is not integrated into the genome of the cell.

43. The method of any one of claims 36-42, wherein the reprogramming mRNA introduced into the non-human-animal-derived cell or cells is selected from the group consisting of OCT4, SOX2, KLF4, cMYC, NANOG, LIN28, KLF5 and any combination thereof.

44. The method of claim 36 or claim 39, wherein the immune evasion mRNA introduced into the non-human-animal -derived cell or cells is selected from the group consisting of E3, K3, B18R [EKB] from vaccinia virus and any combination thereof.

45. The method of any one of claims 35-44, wherein the non-human-animal is bovine, and wherein the bovine-derived cells are obtained from bovine umbilical cord, bovine nasopharyngeal mucosa, bovine blood and bovine milk.

46. The method of any one of claims 1-45, wherein the homogenous aggregates comprise at least 70% of cells expressing at least one pluripotency marker.

47. The method of claim 46, wherein the at least one pluripotency marker is selected from the group consisting of stage-specific embryonic antigen-4 (SSEA4), octamer-binding transcription factor 4 (Oct4), Sall4 transcription factor, Nanog homeobox transcription factor, Lin28A translation factor, DNA-methyltransferase Dnmt3b, and a combination thereof.

48. A plurality of homogenous aggregates comprising at least 70% viable non-genetically modified non-human-animal-derived PSCs.

49. A suspension comprising serum-free liquid medium and a plurality of homogenous aggregates, wherein the aggregates comprise at least 70% viable, non-genetically modified non-human-animal-derived PSCs.

50. The suspension of claim 49, wherein the non-genetically modified, non-human-animal derived PSCs are dividing every 16-32 hours.

51. The suspension of any one of claims 49-50, wherein the non-genetically modified non-human-animal-derived PSCs express at least one pluripotency marker.

52. The suspension of claim 51, wherein the pluripotency marker is selected from the group consisting of stage-specific embryonic antigen-4 (SSEA4), octamer-binding transcription factor 4 (Oct4), Sall4 transcription factor, Nanog homeobox transcription factor, Lin28A translation factor, DNA-methyltransferase Dnmt3b, and a combination thereof.

53. The suspension of any one of claims 49-52 wherein the PSCs express at least one surface protein contributing to cell-cell adhesion.

54. The suspension of any one of claims 49-53, wherein the aggregates have an average size of from 200µm to 500µm.

55. The suspension of any one of claims 49-54, said suspension comprises from 10^6 to 10^9 non-genetically modified, non-human-animal derived PSCs.

56. The suspension of any one of claims 49-55 wherein the non-human-animal is selected from the group consisting of ungulate, poultry, aquatic animals, invertebrate and reptiles.

57. The suspension of claim 56, wherein the ungulate is selected from the group consisting of a bovine, an ovine, an equine, a pig, a giraffe, a camel, a deer, a hippopotamus, and a rhinoceros.

58. The suspension of claim 57, wherein the ungulate is bovine.

59. A cell grown meat comprising the plurality of homogenous aggregates of non-genetically modified non-human animal-derived PSCs or a suspension comprising same according to any one of claims 48-58.