

ARGININE DEIMINASE WITH REDUCED CROSS-REACTIVITY TOWARD ADI – PEG 20
ANTIBODIES FOR CANCER TREATMENT

CROSS-REFERENCE TO RELATED APPLICATIONS

5 This application claims priority under 35 U.S.C. § 119(e) to U.S. Application No. 62/051,182, filed September 16, 2014, which is incorporated by reference in its entirety.

STATEMENT REGARDING SEQUENCE LISTING

10 The Sequence Listing associated with this application is provided in text format in lieu of a paper copy, and is hereby incorporated by reference into the specification. The name of the text file containing the Sequence Listing is POLA_005_01WO_SeqList_ST25.txt. The text file is about 97 KB, was created on September 15, 2015, and is being submitted electronically via EFS-Web.

15 **BACKGROUND**

Technical Field

 The present invention relates generally to arginine deiminase (ADI) proteins, including ADI proteins having reduced cross-reactivity with ADI-PEG 20 antibodies. Such ADI proteins are useful for treating arginine-dependent or related diseases such as cancer.

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Description of the Related Art

 Amino acid deprivation therapy can be an effective treatment of some forms of cancer. To date, there is one known clinical example relevant to this approach which utilizes asparaginase to lower circulating levels of asparagine and inhibit protein synthesis. This treatment is particularly effective for acute lymphoblastic leukemia (Avramis 2005, Viera
25 Pinheiro 2004). Acute lymphoblastic leukemia cells require the amino acid asparagine for growth and proliferation. In contrast, most normal human cells are capable of synthesizing asparagine and are unaffected by asparagine depletion. Therefore, decreasing serum asparagine with asparaginase can selectively kill the cancer cells without harming the
30 normal cells, tissues, and host. An *E. coli* derived form of asparaginase has been approved for human use. However, asparaginase is found only in microbes; which makes it highly immunogenic in humans and also has a short serum half-life following injection (Avramis 2005). To make asparaginase a more effective drug, these drawbacks were minimized by formulating the *E. coli* derived asparaginase with polyethylene glycol (PEG) to reduce the
35 immunogenicity of this enzyme and the associated allergic reactions. In addition, PEG greatly prolongs the circulating half-life of asparaginase, which reduces both the frequency

of treatment and the total cost of the therapy. PEG formulated asparaginase is approved for use and is marketed under the trade name Oncaspar® (Oncaspar® 2011, Avramis 2005, Viera Pinheiro 2004, Fu 2007, Zeidan 2008).

Arginine is another non-essential amino acid for humans and mice (for review see Rogers 1994). In humans, arginine can be synthesized from citrulline in two steps via the Krebs (urea) cycle enzymes argininosuccinate synthetase (ASS, L-citrulline:L-aspartate ligase [AMP-forming], EC 6.3.4.5) and argininosuccinate lyase (ASL, L-argininosuccinate arginine-lyase, EC 4.3.2.) (Haines 2011, Wu 2009, Morris 2006, Husson 2003, Tapiero 2002, Rogers 1994). ASS catalyzes the conversion of citrulline and aspartic acid to argininosuccinate, which is then converted to arginine and fumaric acid by ASL. An arginine deficient diet in humans does not evoke hyperammonemia, orotic aciduria, nor alter the rate of whole body nitric oxide (NO) synthesis in adult humans (Tapiero 2002, Castillo 1995, Rogers 1994, Carey 1987, Barbul 1986, Snyderman 1959, Rose 1949). Although preterm infants appear to require arginine (Wu 2004), arginine levels do not correlate with age among infants, children and young adults (Lücke 2007). In 1992, Takaku and Sugimura separately reported that human melanomas and hepatocellular carcinoma (HCC) cell lines appear to require arginine for growth. Other studies showed that pegylated ADI was effective for the treatment of melanomas and hepatomas with few adverse effects.

ADI-PEG 20 treatment requires multiple doses over a period of time. After a number of treatments, anti-ADI-PEG 20 antibodies can develop that may limit its continued effectiveness. Therefore, there is a need in the art for ADI that has reduced cross-reactivity to anti-ADI-PEG20 antibodies for use in treatment in order to improve and extend the efficacy of arginine depletion therapy. The present invention provides this and other advantages for the treatment of cancers.

References: Avramis VI, Panosyan EH. 2005. Clin Pharmacokinet 44:367-393; Barbul A. 1986. J Parenteral Enteral Nutr 10:227-238; Carey GP, *et al.* 1987. J Nutr 117:1734-1739; Castillo L, *et al.* 1995. Am J Physiol 268 (Endocrinol Metab 31):E360-367; Fu CH, Sakamoto KM. 2007. Expert Opin Pharmacother 8:1977-1984; Haines RJ, *et al.* 2011. Int J Biochem Mol Biol 2:8-23; Husson A, *et al.* 2003. Eur J Biochem 270:1887-1899; Lücke T, *et al.* 2007. Clin Chem Lab Med 45:1525-1530; Morris SM Jr. 2006. Am J Clin Nutr 83(Suppl):598S-512S; Rogers QR. 1994. In Proceedings from a Symposium Honoring Willard J. Visek - from Ammonia to Cancer and Gene Expression. Special Publication 86 - April, 1994, Agriculture Experiment Station, University of Illinois, 211 Mumford Hall, Urbana, IL 61801, pp. 9-21; Tapiero H, *et al.* 2002. Biomed Pharmacother 56:439- 445, 2002; Viera Pinheiro JP, Boos J. 2004. Br J Haematol 125: 117-127; Wu G, *et al.* 2009. Amino Acids 37:153-168; Wu G, *et al.* 2004. J Nutr Biochem 15:442-451; Zeidan A, *et al.* 2008. Expert Opin Biol Ther 9:111-119).

BRIEF SUMMARY

In one aspect of the present disclosure, there is provided a therapeutic composition comprising an isolated arginine deiminase, or a fragment thereof having arginine deiminase (ADI) activity, and a pharmaceutically-acceptable carrier, wherein the isolated arginine deiminase has reduced cross-reactivity with patient anti-ADI-PEG 20 antibodies and comprises the amino acid sequence set forth in SEQ ID NO: 9 or SEQ ID NO: 22, or a variant thereof that displays at least 90% sequence identity to SEQ ID NO: 9 or SEQ ID NO: 22.

In another aspect of the present disclosure, there is provided a use of a therapeutic composition disclosed herein in the manufacture of a medicament for treating, ameliorating the symptoms of, or inhibiting the progression of a cancer in a patient.

In a further aspect of the present disclosure, there is provided a use of a ADI-PEG 20 and a therapeutic composition disclosed herein in the manufacture of a medicament for treating, ameliorating the symptoms of, or inhibiting the progression of a cancer in a patient, wherein said ADI-PEG 20 is to be administered to the patient, and after a period of time, said therapeutic composition is to be administered to the patient.

In another aspect of the present disclosure, there is provided a therapeutic composition disclosed herein for use in treating, ameliorating the symptoms of, or inhibiting the progression of a cancer.

Certain examples relate to an isolated arginine deiminase, wherein the isolated arginine deiminase has reduced cross-reactivity with patient anti-ADI-PEG 20 antibodies. Also included are therapeutic or pharmaceutical compositions comprising an isolated arginine deiminase or a fragment thereof having ADI activity, and a pharmaceutically-acceptable carrier. In certain examples, the composition is sterile and/or substantially free of pyrogens such as endotoxins. In some examples, the isolated arginine deiminase having reduced cross-reactivity with patient anti-ADI-PEG 20 antibodies is not from *M. hominis*. In some examples, the isolated arginine deiminase having reduced cross-reactivity with patient anti-ADI-PEG 20 antibodies is from an organism listed in Table 1.

In certain examples the isolated arginine deiminase having reduced cross-reactivity with patient anti-ADI-PEG 20 antibodies has one or more properties comparable to or better than those of ADI-PEG 20. In this regard, the one or more properties includes, but is not limited to, K_{cat}, K_m, pH optimum, stability, *in vivo* proteolytic stability, or no requirement for ions or cofactors that are not already present in blood, or any combination thereof. In some examples, the isolated arginine deiminase having reduced cross-reactivity with patient anti-ADI-PEG 20 antibodies, has at least 5, 10, 15, or 20 surface residue changes as compared to *M. hominis* arginine deiminase. In certain examples,

the isolated arginine deiminase having reduced cross-reactivity with patient anti-ADI-PEG 20 antibodies has between about 20 and 135 surface residue changes, between about 40 and 100 surface residue changes, between about 30 and 60 surface residue changes, between about 80 and 100 surface residues changes, or between about 100 and 120 surface residues changes, as compared to *M. hominis* arginine deiminase.

In certain examples, the isolated arginine deiminase having reduced cross-reactivity with patient anti-ADI-PEG 20 antibodies is from *Mycoplasma salivarium*, *Mycoplasma spumans*, *Mycoplasma canadense*, *Mycoplasma auris*, *Mycoplasma hyosynoviae*, *Mycoplasma cloacale*, *Mycoplasma anseris*, *Mycoplasma alkalescens*, *Mycoplasma orale*, *Mycoplasma iners*, *Mycoplasma meleagridis*, *Mycoplasma alvi*, *Mycoplasma penetrans*, *Mycoplasma gallinarum*, *Mycoplasma pirum*, *Mycoplasma primatum*, *Mycoplasma fermentans*, *Mycoplasma lipofaciens*, *Mycoplasma felifaucium*, *Mycoplasma imitans*, *Mycoplasma opalescens*, *Mycoplasma moatsii*, *Mycoplasma elephantis*, *Mycoplasma pneumoniae*, *Mycoplasma testudinis*, *Mycoplasma* sp. CAG:877, or *Mycoplasma* sp. CAG:472. Illustrative arginine deiminase having reduced cross-reactivity with patient anti-ADI-PEG 20 antibodies comprise any one or more of the amino acid sequences set forth in SEQ ID NOS:2-28.

In some examples, the isolated arginine deiminase having reduced cross-reactivity with patient anti-ADI-PEG 20 antibodies has been modified to remove at least one pegylation site. In certain examples of the arginine deiminase having reduced cross-reactivity with patient anti-ADI-PEG 20 antibodies, at least one lysine residue has been modified by an amino acid substitution. In this regard, in certain examples, at least about 5 lysine residues, at least about 10 lysine residues, or at least about 20 lysine residues have been modified by an amino acid substitution.

In some examples, the arginine deiminase having reduced cross-reactivity with patient anti-ADI-PEG 20 antibodies is covalently bonded via a linker to a PEG molecule. In this regard, the arginine deiminase having reduced cross-reactivity with patient anti-ADI-PEG 20 antibodies may be covalently bonded to one or more PEG molecule, such as to about 1 to about 10 or about 2 to about 8 PEG molecules. The PEG molecules may be straight chain or branch chain PEG molecules and may have a total weight average molecular weight of from about 1,000 to about 40,000, or a total weight average molecular weight of from about 10,000 to about 30,000. In some examples, where the PEG is covalently bonded to an ADI_r described herein via a linker, the linker may comprise a succinyl group, an amide group, an imide group, a carbamate group, an ester group, an epoxy group, a carboxyl group, a hydroxyl group, a carbohydrate, a tyrosine group, a cysteine group, a histidine group, a methylene group, or any combinations thereof. In specific examples, the source of the succinyl group is succinimidyl succinate.

Also included are polynucleotides encoding an isolated arginine deiminase described herein, vectors comprising the polynucleotide, and isolated host cells comprising the vectors.

Certain examples relate to compositions comprising an isolated arginine deiminase having reduced cross-reactivity with patient anti-ADI-PEG 20 antibodies as described herein, and a physiologically acceptable carrier. In certain examples, the compositions further comprise a chemotherapeutic agent. Exemplary chemotherapeutic agents include, but are not limited to, docetaxel, carboplatin, cyclophosphamide, gemcitabine, cisplatin, sorafenib, sunitinib, and everolimus.

Also included are methods of treating, ameliorating the symptoms of, or inhibiting the progression of a cancer comprising administering to a patient in need thereof a therapeutically effective amount of a composition comprising the isolated arginine deiminase having reduced cross-reactivity with patient anti-ADI-PEG 20 antibodies as described herein and a physiologically acceptable carrier, thereby treating, ameliorating the symptoms of, or inhibiting the progression of the cancer. In certain examples, the patient in need thereof has been determined to have anti-ADI-PEG 20 antibodies. In some examples, the cancer is selected from the group consisting of hepatocellular carcinoma, melanoma including metastatic melanoma, pancreatic cancer, prostate cancer, small cell lung cancer,

mesothelioma, lymphocytic leukemia, chronic myelogenous leukemia, lymphoma, hepatoma, sarcoma, leukemia, acute myeloid leukemia, relapsed acute myeloid leukemia, breast cancer, ovarian cancer, colorectal cancer, gastric cancer, glioma, glioblastoma multiforme, non-small cell lung cancer (NSCLC), kidney cancer, bladder cancer, uterine cancer, esophageal cancer, brain cancer, head and neck cancers, cervical cancer, testicular cancer, and stomach cancer.

Some examples include methods of treating, ameliorating the symptoms of, or inhibiting the progression of a cancer comprising administering to a patient in need thereof a therapeutically effective amount of a composition comprising ADI-PEG 20, and after a period of time, administering to the patient a composition comprising the isolated arginine deiminase having reduced cross-reactivity with patient anti-ADI-PEG 20 antibodies as described herein and a physiologically acceptable carrier, thereby treating, ameliorating the symptoms of, or inhibiting the progression of the cancer. The period of time may be determined, for example, by detecting a predetermined level of anti-ADI-PEG 20 antibodies in the patient and/or measuring or otherwise observing ADI activity in the patient, wherein the composition comprising the isolated arginine deiminase having reduced cross-reactivity with patient anti-ADI-PEG 20 antibodies is administered following detection of the predetermined level of said anti-ADI-PEG 20 antibodies and/or measurement or observation of a predetermined level of ADI activity in the patient.

Also included are isolated arginine deiminase proteins described herein for use in the preparation or manufacture of a medicament for treating, ameliorating the symptoms of, or inhibiting the progression of a cancer.

BRIEF DESCRIPTION OF THE DRAWINGS

Figures 1A-1D illustrate a variety of cysteine-reactive PEG molecules that can be conjugated to the ADI enzymes described herein.

DETAILED DESCRIPTION

Embodiments of the present invention relate to selected ADI enzymes, which in some embodiments are conjugated with PEG through a linker, e.g., a stable linker. In some embodiments, the ADI enzymes are engineered or selected to have a small or otherwise reduced number of surface lysine residues, for example, relative to a wild-type sequence or a reference sequence (see, e.g., Table 1). The selected ADI enzymes are chosen from a large number of ADI enzymes, from different organisms, based on their beneficial properties. These properties include the ability of the enzyme to establish and maintain low arginine concentrations in human blood through ADI conversion of arginine to citrulline and ammonia. In some embodiments, the selected ADI molecules have reduced cross-reactivity toward

anti-ADI-PEG 20 antibodies as compared to ADI-PEG 20, such antibodies possibly resulting from a patient's previous treatment with ADI-PEG 20.

In certain embodiments, the ADI enzymes are pegylated to provide protection against renal clearance and proteolysis, as well as reduced immunogenicity or antigenicity. To increase the effectiveness of the pegylation, modifications to the enzymes may be engineered to reduce the number of surface lysine residues and therefore limit the number of available PEG attachment sites. In some instances, reducing the number of lysine residues provides more complete and uniform pegylation at the remaining lysine attachment residues.

In some embodiments, the PEG linker selected to attach methoxy-PEG to ADI provides a chemically-stable linkage. It is expected that a stable linker will increase the molecule's bioactive lifetime. A chemically stable linker will also eliminate hydrolysis and reduce an immune response that might occur to a de-pegylated linker attached to the enzyme surface.

These cumulative specifications result in one or more molecules that effectively remove arginine from a patient's blood and are not neutralized or cleared by anti-ADI-PEG 20 antibodies from previous arginine depletion therapy. The molecules are pegylated so as to delay neutralization and clearance due to their own immunogenicity. These factors will permit their use instead of ADI-PEG 20 or in addition to ADI-PEG 20 (e.g., as a follow-on drug) to extend arginine depletion therapy and therefore increase effectiveness of arginine depletion treatment as an anti-cancer therapeutic.

Normal cells do not require arginine for growth, since they can synthesize arginine from citrulline in a two-step process catalyzed by ASS and ASL. In contrast, certain cancers do not express ASS. Certain cancers do not express ASL, and other cancers may have diminished expression of, or may not express ASS and/or ASL. Therefore, these cancers are auxotrophic for arginine. This metabolic difference may be capitalized upon to develop a safe and effective therapy to treat these forms of cancer. ADI catalyzes the conversion of arginine to citrulline via the arginine dihydrolase pathway, and may thus be used to eliminate arginine.

The practice of the present invention will employ, unless indicated specifically to the contrary, conventional methods of virology, immunology, microbiology, molecular biology and recombinant DNA techniques within the skill of the art, many of which are described below for the purpose of illustration. Such techniques are explained fully in the literature. See, e.g., *Current Protocols in Protein Science*, *Current Protocols in Molecular Biology* or *Current Protocols in Immunology*, John Wiley & Sons, New York, N.Y. (2009); Ausubel *et al.*, *Short Protocols in Molecular Biology*, 3rd ed., Wiley & Sons, 1995; Sambrook and Russell, *Molecular Cloning: A Laboratory Manual* (3rd Edition, 2001); Maniatis *et al.* *Molecular*

Cloning: A Laboratory Manual (1982); *DNA Cloning: A Practical Approach*, vol. I & II (D. Glover, ed.); *Oligonucleotide Synthesis* (N. Gait, ed., 1984); *Nucleic Acid Hybridization* (B. Hames & S. Higgins, eds., 1985); *Transcription and Translation* (B. Hames & S. Higgins, eds., 1984); *Animal Cell Culture* (R. Freshney, ed., 1986); Perbal, *A Practical Guide to Molecular Cloning* (1984) and other like references.

Standard techniques may be used for recombinant DNA, oligonucleotide synthesis, and tissue culture and transformation (e.g., electroporation, lipofection). Enzymatic reactions and purification techniques may be performed according to manufacturer's specifications or as commonly accomplished in the art or as described herein. These and related techniques and procedures may be generally performed according to conventional methods well known in the art and as described in various general and more specific references that are cited and discussed throughout the present specification. Unless specific definitions are provided, the nomenclature utilized in connection with, and the laboratory procedures and techniques of, molecular biology, analytical chemistry, synthetic organic chemistry, and medicinal and pharmaceutical chemistry described herein are those well-known and commonly used in the art. Standard techniques may be used for recombinant technology, molecular biological, microbiological, chemical syntheses, chemical analyses, pharmaceutical preparation, formulation, and delivery, and treatment of patients.

As used in this specification and the appended claims, the singular forms "a," "an" and "the" include plural references unless the content clearly dictates otherwise.

By "about" is meant a quantity, level, value, number, frequency, percentage, dimension, size, amount, weight or length that varies by as much as 30, 25, 20, 15, 10, 9, 8, 7, 6, 5, 4, 3, 2 or 1% to a reference quantity, level, value, number, frequency, percentage, dimension, size, amount, weight or length.

As used herein, the term "amino acid" includes both naturally-occurring and non-natural amino acids as well as amino acid analogs and mimetics. Naturally-occurring amino acids include the 20 (L)-amino acids utilized during protein biosynthesis as well as others such as 4-hydroxyproline, hydroxylysine, desmosine, isodesmosine, homocysteine, citrulline and ornithine, for example. Non-natural amino acids include, for example, norleucine, norvaline, p-fluorophenylalanine, ethionine, and the like, which are known to a person skilled in the art. Amino acid analogs include modified forms of natural and non-natural amino acids. Such modifications can include, for example, substitution or replacement of chemical groups and moieties on the amino acid or by derivatization of the amino acid. Amino acid mimetics include, for example, organic structures which exhibit functionally similar properties such as charge and charge spacing characteristic of the reference amino acid. For example, an organic structure which mimics arginine (Arg or R) would have a positive charge moiety located in similar molecular space and having the same degree of mobility as the e-amino

group of the side chain of the naturally occurring Arg amino acid. Mimetics also include constrained structures so as to maintain optimal spacing and charge interactions of the amino acid or of the amino acid functional groups. Those skilled in the art know or can determine what structures constitute functionally equivalent amino acid analogs and amino acid mimetics.

Throughout this specification, unless the context requires otherwise, the word “comprise”, or variations such as “comprises” or “comprising”, will be understood to imply the inclusion of a stated element or integer or group of elements or integers but not the exclusion of any other element or integer or group of elements or integers.

“Biocompatible” refers to materials or compounds which are generally not injurious to biological functions and which will not result in any degree of unacceptable toxicity, including allergenic and disease states.

The terms “endotoxin free” and “substantially endotoxin free” relate generally to compositions, solvents, and/or vessels that contain at most trace amounts (e.g., amounts having no clinically adverse physiological effects to a subject) of endotoxin, and preferably undetectable amounts of endotoxin. Endotoxins are toxins associated with certain bacteria, typically gram-negative bacteria, although endotoxins may be found in gram-positive bacteria, such as *Listeria monocytogenes*. The most prevalent endotoxins are lipopolysaccharides (LPS) or lipo-oligo-saccharides (LOS) found in the outer membrane of various Gram-negative bacteria, and which represent a central pathogenic feature in the ability of these bacteria to cause disease. Small amounts of endotoxin in humans may produce fever, a lowering of the blood pressure, and activation of inflammation and coagulation, among other adverse physiological effects.

Therefore, in pharmaceutical production, it is often desirable to remove most or all traces of endotoxin from drug products and/or drug containers, because even small amounts may cause adverse effects in humans. A depyrogenation oven may be used for this purpose, as temperatures in excess of 300°C are typically required to break down most endotoxins. For instance, based on primary packaging material such as syringes or vials, the combination of a glass temperature of 250°C and a holding time of 30 minutes is often sufficient to achieve a 3 log reduction in endotoxin levels. Other methods of removing endotoxins are contemplated, including, for example, chromatography and filtration methods, as described herein and known in the art. Also included are methods of producing polypeptides in and isolating them from eukaryotic cells such as mammalian cells to reduce, if not eliminate, the risk of endotoxins being present in a composition of the invention.

Included are methods of producing polypeptides in and isolating them from serum free cells.

Endotoxins can be detected using routine techniques known in the art. For example, the Limulus Amoebocyte Lysate assay, which utilizes blood from the horseshoe crab, is a

highly sensitive assay for detecting presence of endotoxin. In this test, very low levels of LPS can cause detectable coagulation of the limulus lysate due a powerful enzymatic cascade that amplifies this reaction. Endotoxins can also be quantitated by enzyme-linked immunosorbent assay (ELISA). To be substantially endotoxin free, endotoxin levels may be less than about 0.001, 0.005, 0.01, 0.02, 0.03, 0.04, 0.05, 0.06, 0.08, 0.09, 0.1, 0.5, 1.0, 1.5, 2, 2.5, 3, 4, 5, 6, 7, 8, 9, or 10 EU/ml. Typically, 1 ng lipopolysaccharide (LPS) corresponds to about 1-10 EU.

The "half-life" of a polypeptide can refer to the time it takes for the polypeptide to lose half of its pharmacologic, physiologic, or other activity, relative to such activity at the time of administration into the serum or tissue of an organism, or relative to any other defined time-point. "Half-life" can also refer to the time it takes for the amount or concentration of a polypeptide to be reduced by half of a starting amount administered into the serum or tissue of an organism, relative to such amount or concentration at the time of administration into the serum or tissue of an organism, or relative to any other defined time-point. The half-life can be measured in serum and/or any one or more selected tissues.

"Homology" refers to the percentage number of amino acids that are identical or constitute conservative substitutions. Homology may be determined using sequence comparison programs such as GAP (Deveraux et al., *Nucleic Acids Research*. 12, 387-395, 1984), which is incorporated herein by reference. In this way sequences of a similar or substantially different length to those cited herein could be compared by insertion of gaps into the alignment, such gaps being determined, for example, by the comparison algorithm used by GAP.

The terms "modulating" and "altering" include "increasing," "enhancing" or "stimulating," as well as "decreasing" or "reducing," typically in a statistically significant or a physiologically significant amount or degree relative to a control. An "increased," "stimulated" or "enhanced" amount is typically a "statistically significant" amount, and may include an increase that is 1.1, 1.2, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 30 or more times (e.g., 500, 1000 times) (including all integers and ranges in between e.g., 1.5, 1.6, 1.7, 1.8, etc.) the amount produced by no composition (e.g., the absence of agent) or a control composition. A "decreased" or "reduced" amount is typically a "statistically significant" amount, and may include a 1%, 2%, 3%, 4%, 5%, 6%, 7%, 8%, 9%, 10%, 11%, 12%, 13%, 14%, 15%, 16%, 17%, 18%, 19%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, or 100% decrease (including all integers and ranges in between) in the amount produced by no composition (e.g., the absence of an agent) or a control composition. Examples of comparisons and "statistically significant" amounts are described herein.

“Patient” or “subject” refers to an animal, in certain embodiments a mammal, and in specific embodiments, a human.

In certain embodiments, the “purity” of any given agent (e.g., ADlr, ADlr-PEG) in a composition may be specifically defined. For instance, certain compositions may comprise an agent that is at least 80, 85, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, or 100% pure, including all decimals and ranges in between, as measured, for example and by no means limiting, by high performance liquid chromatography (HPLC), a well-known form of column chromatography used frequently in biochemistry and analytical chemistry to separate, identify, and quantify compounds.

The term “reference sequence” refers generally to a nucleic acid coding sequence, or amino acid sequence, to which another sequence is being compared. All polypeptide and polynucleotide sequences described herein are included as references sequences, including those described by name and those described in the Tables and the Sequence Listing.

The terms “sequence identity” or, for example, comprising a “sequence 50% identical to,” as used herein, refer to the extent that sequences are identical on a nucleotide-by-nucleotide basis or an amino acid-by-amino acid basis over a window of comparison. Thus, a “percentage of sequence identity” may be calculated by comparing two optimally aligned sequences over the window of comparison, determining the number of positions at which the identical nucleic acid base (e.g., A, T, C, G, I) or the identical amino acid residue (e.g., Ala, Pro, Ser, Thr, Gly, Val, Leu, Ile, Phe, Tyr, Trp, Lys, Arg, His, Asp, Glu, Asn, Gln, Cys and Met) occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the window of comparison (i.e., the window size), and multiplying the result by 100 to yield the percentage of sequence identity.

Terms used to describe sequence relationships between two or more polypeptides include “reference sequence,” “comparison window,” “sequence identity,” “percentage of sequence identity” and “substantial identity.” A “reference sequence” can be at least 12 but frequently 15 to 18 and often at least 25 monomer units, inclusive of nucleotides and amino acid residues, in length. Because two polypeptides may each comprise (1) a sequence (i.e., only a portion of the complete polypeptides sequence) that is similar between the two polypeptides, and (2) a sequence that is divergent between the two polypeptides, sequence comparisons between two (or more) polypeptides are typically performed by comparing sequences of the two polypeptides over a “comparison window” to identify and compare local regions of sequence similarity. A “comparison window” refers to a conceptual segment of at least 6 contiguous positions, usually about 50 to about 100, more usually about 100 to about 150 in which a sequence is compared to a reference sequence of the same number of contiguous positions after the two sequences are optimally aligned. The comparison window

may comprise additions or deletions (i.e., gaps) of about 20% or less as compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. Optimal alignment of sequences for aligning a comparison window may be conducted by computerized implementations of algorithms (GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package Release 7.0, Genetics Computer Group, 575 Science Drive Madison, WI, USA) or by inspection and the best alignment (i.e., resulting in the highest percentage homology over the comparison window) generated by any of the various methods selected. Reference also may be made to the BLAST family of programs as for example disclosed by Altschul et al. (Nucl. Acids Res. 25:3389, 1997). A detailed discussion of sequence analysis can be found in Unit 19.3 of Ausubel et al. ("Current Protocols in Molecular Biology," John Wiley & Sons Inc, 1994-1998, Chapter 15).

Calculations of sequence similarity or sequence identity between sequences (the terms are used interchangeably herein) can be performed as follows. To determine the percent identity of two amino acid sequences, or of two nucleic acid sequences, the sequences can be aligned for optimal comparison purposes (e.g., gaps can be introduced in one or both of a first and a second amino acid or nucleic acid sequence for optimal alignment and non-homologous sequences can be disregarded for comparison purposes). In certain embodiments, the length of a reference sequence aligned for comparison purposes is at least 30%, preferably at least 40%, more preferably at least 50%, 60%, and even more preferably at least 70%, 80%, 90%, or 100% of the length of the reference sequence. The amino acid residues or nucleotides at corresponding amino acid positions or nucleotide positions are then compared. When a position in the first sequence is occupied by the same amino acid residue or nucleotide as the corresponding position in the second sequence, then the molecules are identical at that position.

The percent identity between the two sequences is a function of the number of identical positions shared by the sequences, taking into account the number of gaps, and the length of each gap, which need to be introduced for optimal alignment of the two sequences.

The comparison of sequences and determination of percent identity between two sequences can be accomplished using a mathematical algorithm. In some embodiments, the percent identity between two amino acid sequences is determined using the Needleman and Wunsch algorithm (J. Mol. Biol. 48: 444-453, 1970) which has been incorporated into the GAP program in the GCG software package, using either a Blossum 62 matrix or a PAM250 matrix, and a gap weight of 16, 14, 12, 10, 8, 6, or 4 and a length weight of 1, 2, 3, 4, 5, or 6. In some embodiments, the percent identity between two nucleotide sequences is determined using the GAP program in the GCG software package, using a NWSgapdna.CMP matrix and a gap weight of 40, 50, 60, 70, or 80 and a length weight of 1, 2, 3, 4, 5, or 6. Another exemplary set of parameters (and the one that should be used unless otherwise specified)

includes a Blossum 62 scoring matrix with a gap penalty of 12, a gap extend penalty of 4, and a frameshift gap penalty of 5. The percent identity between two amino acid or nucleotide sequences can also be determined using the algorithm of E. Meyers and W. Miller (Cabios. 4:11-17, 1989) which has been incorporated into the ALIGN program (version 2.0), using a PAM120 weight residue table, a gap length penalty of 12 and a gap penalty of 4.

The term "solubility" refers to the property of ADIr enzyme provided herein to dissolve in a liquid solvent and form a homogeneous solution. Solubility is typically expressed as a concentration, either by mass of solute per unit volume of solvent (g of solute per kg of solvent, g per dL (100 mL), mg/ml, etc.), molarity, molality, mole fraction or other similar descriptions of concentration. The maximum equilibrium amount of solute that can dissolve per amount of solvent is the solubility of that solute in that solvent under the specified conditions, including temperature, pressure, pH, and the nature of the solvent. In certain embodiments, solubility is measured at physiological pH, or other pH, for example, at pH 5.0, pH 6.0, pH 7.0, pH 7.2, pH 7.4, pH 7.6, pH 7.8, or pH 8.0. In certain embodiments, solubility is measured in water or a physiological buffer such as PBS or NaCl (with or without NaP) or other buffer/composition described herein. In specific embodiments, solubility is measured at relatively lower pH (e.g., pH 6.0) and relatively higher salt (e.g., 500mM NaCl and 10mM NaP). In certain embodiments, solubility is measured in a biological fluid (solvent) such as blood or serum. In certain embodiments, the temperature can be about room temperature (e.g., about 20, 21, 22, 23, 24, 25°C) or about body temperature (37°C). In certain embodiments, an ADIr enzyme has a solubility of at least about 0.1, 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, 0.9, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 25, or 30 mg/ml at room temperature or at 37°C.

"Substantially" or "essentially" means nearly totally or completely, for instance, 95%, 96%, 97%, 98%, 99% or greater of some given quantity.

By "statistically significant," it is meant that the result was unlikely to have occurred by chance. Statistical significance can be determined by any method known in the art. Commonly used measures of significance include the p-value, which is the frequency or probability with which the observed event would occur, if the null hypothesis were true. If the obtained p-value is smaller than the significance level, then the null hypothesis is rejected. In simple cases, the significance level is defined at a p-value of 0.05 or less.

Each embodiment in this specification is to be applied *mutatis mutandis* to every other embodiment unless expressly stated otherwise.

Throughout the present disclosure, the following abbreviations may be used: PEG, polyethylene glycol; ADI, arginine deiminase; SS, succinimidyl succinate; SSA, succinimidyl succinimide; SPA, succinimidyl propionate; NHS, N-hydroxy-succinimide; ASS1 or ASS, argininosuccinate synthetase; ASL, argininosuccinate lyase.

A polynucleotide encoding an ADI enzyme may be derived, cloned, isolated, synthesized or produced from any source, including, for example, microorganisms, recombinant biotechnology or any combination thereof. For example, arginine deiminase may be cloned from microorganisms of the genera *Mycoplasma*. In certain embodiments, arginine deiminase is cloned from *Mycoplasma salivarium*, *Mycoplasma spumans*, *Mycoplasma canadense*, *Mycoplasma auris*, *Mycoplasma hyosynoviae*, *Mycoplasma cloacale*, *Mycoplasma anseris*, *Mycoplasma alkalescens*, *Mycoplasma orale*, *Mycoplasma iners*, *Mycoplasma meleagridis*, *Mycoplasma alvi*, *Mycoplasma penetrans*, *Mycoplasma gallinarum*, *Mycoplasma pirum*, *Mycoplasma primum*, *Mycoplasma fermentans*, *Mycoplasma lipofaciens*, *Mycoplasma felifaucium*, *Mycoplasma imitans*, *Mycoplasma opalescens*, *Mycoplasma moatsii*, *Mycoplasma elephantis*, *Mycoplasma pneumoniae*, *Mycoplasma testudinis*, *Mycoplasma sp. CAG:877*, or *Mycoplasma sp. CAG:472*, or any combination thereof. In some embodiments, the arginine deiminase is cloned from a species listed in Table 1. In particular embodiments, the ADI comprises the amino acid sequence of any one of SEQ ID NOs: 2-28, or a variant or fragment or extension thereof having ADI activity (e.g., able to metabolize arginine into citrulline and ammonia). Such ADI enzymes can be prepared or synthesized using known techniques.

In certain embodiments, the ADI enzymes as described herein are compared to the benchmark ADI-PEG 20 molecule derived from *M. hominis*. As used herein, “ADI-PEG 20” refers to the ADI molecule known in the art and described for example in U.S. Patent Nos. 6,183,738 and 6,635,462; see also Ascierto et al., 2005. Pegylated arginine deiminase treatment of patients with metastatic melanoma: results from phase I and II studies. J Clin Oncol 23(30): 7660– 7668; Izzo F, et al. (2004) Pegylated arginine deiminase treatment of patients with unresectable hepatocellular carcinoma: results from phase I/II studies. J Clin Oncol 22(10): 1815–1822; Holtsberg FW, et al. (2002), Poly(ethylene glycol) (PEG) conjugated arginine deiminase: effects of PEG formulations on its pharmacological properties. J Control Release 80(1– 3): 259–271; Kelly et al., (2012) British Journal of Cancer 106, 324 – 332. As would be recognized by the skilled artisan, this molecule is a pegylated (PEG 20,000) ADI enzyme derived from *M. hominis*, and has two substitutions (K112E; P210S) relative to the wild type *M. hominis* ADI enzyme.

The arginine deiminase enzymes as described herein are screened from a large number of ADI enzymes and are believed to have reduced level of reactivity with anti-ADI-PEG 20 antibodies from patients and/or other beneficial properties. Anti-ADI-PEG 20 antibodies can appear in subjects treated with ADI-PEG 20 and can be measured using known methodologies. Reactivity to anti-ADI-PEG 20 antibodies can be determined for example using ELISA or other similar assays known to the skilled artisan.

In this regard, ADI-PEG 20 can be used as a comparison to assess cross-reactivity level to patient anti-ADI-PEG 20 antibodies. A cross-reactivity level that is statistically significantly lower than that of ADI-PEG 20 to patient anti-ADI-PEG 20 antibodies may be useful herein. In certain embodiments, the arginine deiminase enzymes as described herein have low or no cross-reactivity to anti-ADI-PEG 20 antibodies. In certain embodiments, any reduction in reactivity to anti-ADI-PEG 20 antibodies as compared to reactivity with ADI-PEG 20 can be beneficial as such an ADI enzyme would improve treatment options for patients in need of arginine depletion therapy. Thus, in some embodiments, the arginine deiminase enzymes as described herein have reduced cross-reactivity to patient anti-ADI-PEG 20 antibodies as compared to ADI-PEG 20 reactivity to such antibodies.

“ADIr” is used herein to refer to an ADI enzyme of the present invention having reduced cross-reactivity to anti-ADI-PEG 20 antibodies as compared to ADI-PEG 20 reactivity to such antibodies. “ADIr” nomenclature is used to distinguish the molecules identified herein from ADI and ADI-PEG 20 as known in the art. Examples of ADIr enzymes include SEQ ID NOS:2-28, and variants thereof that differ in amino acid sequence from SEQ ID NO:1 or ADI-PEG 20.

In some embodiments, the ADIr enzymes of the invention have characteristics or properties comparable to or better than those of ADI-PEG 20, in order to reduce and maintain low blood arginine levels for effective cancer treatment. Examples of such properties include Kcat, Km, pH optimum, stability, *in vivo* proteolytic stability and lack of requirement for ions or cofactors not already present in the blood, or any combination thereof. In certain embodiments, an ADIr as described herein has properties that are about or at least 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or higher, than comparable properties of ADI-PEG 20. In some embodiments an ADIr described herein has properties that are about or at least about 100%, 105%, 110%, 120%, 140%, 150%, 160%, 180%, 200%, 220%, 240%, 250%, 260%, 280%, 300%, 320%, 340, 350%, 360%, 400%, 420%, 450%, 460%, 500%, 520%, 550% or higher than the specific property of ADI-PEG 20 being compared.

Thus, in certain embodiments, an ADIr has a Kcat that is about or at least about 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% of the Kcat of ADI-PEG 20, or better. In certain embodiments, an ADIr has a Kcat that is about or at least about 100%, 105%, 110%, 120%, 125%, 140%, 150%, 160%, 180%, 200%, 220%, 240%, 250%, 260%, 280%, 300%, 320%, 340, 350%, 360%, 400%, 420%, 450%, 460%, 500%, 520%, 550% or higher, times that of the ADI-PEG 20 Kcat. In certain embodiments, the Kcat of the ADIr enzymes described herein, or compositions comprising

same, is about 0.5 sec^{-1} to about 15 sec^{-1} , about 1 sec^{-1} to about 12 sec^{-1} , about 1 sec^{-1} to about 10 sec^{-1} , about 1.5 sec^{-1} to about 9 sec^{-1} , about 2 sec^{-1} to about 8 sec^{-1} or about 2.5 sec^{-1} to about 7 sec^{-1} . In certain embodiments, the ADI_r or ADI_r-PEG in a composition has a K_{cat} of about 2.5 sec^{-1} to about 7.5 sec^{-1} . In some embodiments, the ADI_r or ADI_r-PEG in a composition has a K_{cat} of about 2.5 sec^{-1} , about 3 sec^{-1} , about 3.5 sec^{-1} , about 4 sec^{-1} , about 4.5 sec^{-1} , about 5 sec^{-1} , about 5.5 sec^{-1} , about 6 sec^{-1} , about 6.5 sec^{-1} , about 7 sec^{-1} , about 7.2 sec^{-1} , about 7.5 sec^{-1} , about 8 sec^{-1} , about 10 sec^{-1} , about 15 sec^{-1} , about 20 sec^{-1} , about 25 sec^{-1} , about 30 sec^{-1} , about 35 sec^{-1} , about 40 sec^{-1} , about 45 sec^{-1} , about 50 sec^{-1} , about 55 sec^{-1} , about 60 sec^{-1} , about 65 sec^{-1} , about 70 sec^{-1} , about 75 sec^{-1} , about 80 sec^{-1} , about 85 sec^{-1} , about 90 sec^{-1} , about 95 sec^{-1} , or about 100 sec^{-1} .

In certain embodiments, an ADI_r has a K_m that is about or at least about 5%, 6%, 7%, 8%, 9%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% of the K_m of ADI-PEG 20, or better. In certain embodiments, an ADI_r has a K_m that is about or at least about 100%, 105%, 110%, 120%, 130%, 140%, 150%, 160%, 180%, 200%, 220%, 240%, or 250% that of the K_m of ADI-PEG 20. In particular embodiments, an ADI_r, or a pegylated formulation thereof, has a K_m of from about $0.5 \mu\text{M}$ to about $50 \mu\text{M}$, or about $1.6 \mu\text{M}$ to about $48 \mu\text{M}$, or about $0.5 \mu\text{M}$ to about $15 \mu\text{M}$, from about $1 \mu\text{M}$ to about $12 \mu\text{M}$, about $1 \mu\text{M}$ to about $10 \mu\text{M}$, about $1.5 \mu\text{M}$ to about $9 \mu\text{M}$, about $1.5 \mu\text{M}$ to about $8 \mu\text{M}$, or about $1.5 \mu\text{M}$ to about $7 \mu\text{M}$. In certain embodiments, the ADI_r or ADI_r-PEG in a composition has a K_m of about $1.5 \mu\text{M}$ to about $6.5 \mu\text{M}$. In some embodiments, the ADI_r or pegylated formulation thereof has a K_m of about $1.5 \mu\text{M}$, about $1.6 \mu\text{M}$, about $2 \mu\text{M}$, about $2.5 \mu\text{M}$, about $3 \mu\text{M}$, about $3.5 \mu\text{M}$, about $4 \mu\text{M}$, about $4.5 \mu\text{M}$, about $5 \mu\text{M}$, about $5.5 \mu\text{M}$, about $6 \mu\text{M}$, about $6.5 \mu\text{M}$, about $7 \mu\text{M}$, about $8 \mu\text{M}$, about $9 \mu\text{M}$, about $10 \mu\text{M}$, about $12 \mu\text{M}$, about $14 \mu\text{M}$, about $15 \mu\text{M}$, about $16 \mu\text{M}$, about $18 \mu\text{M}$, about $20 \mu\text{M}$, about $22 \mu\text{M}$, about $24 \mu\text{M}$, about $25 \mu\text{M}$, about $26 \mu\text{M}$, about $28 \mu\text{M}$, about $30 \mu\text{M}$, about $32 \mu\text{M}$, about $34 \mu\text{M}$, about $35 \mu\text{M}$, about $36 \mu\text{M}$, about $38 \mu\text{M}$, about $40 \mu\text{M}$, about $42 \mu\text{M}$, about $44 \mu\text{M}$, about $45 \mu\text{M}$, about $46 \mu\text{M}$, about $48 \mu\text{M}$, or about $50 \mu\text{M}$.

In certain embodiments, an ADI_r functions at a pH close to the physiological pH of human blood. Thus, in some embodiments, an ADI_r functions at a pH of about 4 to about 10.8, or about 6 to about 8, or about 6.5 to about 7.5. In certain embodiments, an ADI_r has good enzyme activity at about pH 7.4.

In certain embodiments, an ADI_r has stability during long term storage and temperature and proteolytic stability during treatment in the human body. In some embodiments, an ADI_r does not require ions or cofactors for activity that are not already present in blood.

In certain embodiments, an ADI_r described herein generally has an amino acid sequence sufficiently different from *M. hominis* so that there are surface residue changes which will reduce or eliminate antigenic sites for anti-ADI-PEG 20 antibodies. In some embodiments, there will be no cross reactivity (e.g., no statistically significant cross-reactivity) between the selected ADI_r molecule and existing anti-ADI-PEG 20 antibodies in a subject, and a completely new immune response will be generated in a subject rather than a maturation of the existing response to *M. hominis* ADI. Thus, in some embodiments, an ADI_r as described herein has from 20% - 85% sequence identity to *M. hominis* ADI as set forth in SEQ ID NO:1. In certain embodiments, an ADI_r as described herein has even lower percent sequence identity to *M. hominis* ADI, such as 10% or 15% identity. In certain embodiments, an ADI_r as described herein has 20%, 21%, 22%, 23%, 24%, 25%, 26%, 27%, 28%, 29%, 30%, 31%, 32%, 33%, 34%, 35%, 36%, 37%, 38%, 39%, 40%, 41%, 42%, 43%, 44%, 45%, 46%, 47%, 48%, 49%, 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82% or even 83% identity to *M. hominis* ADI, and still has reduced cross-reactivity toward anti-ADI-PEG 20 antibodies.

In certain embodiments, an ADI_r as described herein has from about 10-140, 15-140, or 25-140 surface residue changes as compared to *M. hominis* ADI. Surface residues can be identified from the crystal structure of *M. hominis* ADI and surface residues for ADI from other organisms can be determined by sequence homology. An ADI_r as described herein may have about 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100, 101, 102, 103, 104, 105, 106, 107, 108, 109, 110, 111, 112, 113, 114, 115, 116, 117, 118, 119, 120, 121, 122, 123, 124, 125, 126, 127, 128, 129, 130, 131, 132, 133, 134, 135, 136, 137, 138, 139, or about 140 surface residue changes as compared to *M. hominis* ADI (see SEQ ID NO:1).

In some embodiments, an ADI_r as described herein has from about 10-140, 15-140, or 25-140 residue changes as compared to *M. hominis* ADI. Such residue changes need not only be of surface amino acid residues. Such residue changes (or additions or deletions) can be at either end of the molecule or may be at any residue of the ADI, such that the modified ADI has the desired ADI activity as described herein. Residues to be changed can be identified from the crystal structure of *M. hominis* ADI and residues for ADI from other organisms can be determined by sequence homology. An ADI_r as described herein may have about 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100, 101, 102, 103, 104, 105, 106, 107, 108, 109, 110, 111, 112, 113, 114, 115, 116, 117, 118, 119, 120, 121, 122, 123, 124, 125, 126, 127, 128, 129, 130, 131, 132, 133, 134, 135, 136, 137, 138, 139, or about 140 amino acid residue changes as compared to *M. hominis* ADI (see SEQ ID NO:1).

From a large number of ADI enzymes, Table 1 lists 27 ADIr enzymes and their percent sequence identity relative to *M. hominis* ADI.

In certain embodiments, the ADIr enzymes identified herein from a number of selected species, have a defined number of surface lysine residues (in certain embodiments, e.g., up to 30 or more). Certain of the ADIr enzymes identified herein that have reduced cross-reactivity with anti-ADI-PEG 20 antibodies have about, at least about, or no more than about 0, 1, 2, 3, 4, 5, 10, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, or 60 surface lysine residues, including all ranges in between.

The terms “polypeptide,” “protein” and “peptide” are used interchangeably and mean a polymer of amino acids not limited to any particular length. The term “enzyme” includes polypeptide or protein catalysts, and with respect to ADIr is used interchangeably with protein, polypeptide, or peptide. The terms include modifications such as myristoylation, sulfation, glycosylation, phosphorylation and addition or deletion of signal sequences. The terms “polypeptide” or “protein” means one or more chains of amino acids, wherein each chain comprises amino acids covalently linked by peptide bonds, and wherein said polypeptide or protein can comprise a plurality of chains non-covalently and/or covalently linked together by peptide bonds, having the sequence of native proteins, that is, proteins produced by naturally-occurring and specifically non-recombinant cells, or genetically-engineered or recombinant cells, and comprise molecules having the amino acid sequence of the native protein, or molecules having deletions from, additions to, and/or substitutions of one or more amino acids of the native sequence. The terms “polypeptide” and “protein” specifically encompass the ADIr enzymes/proteins described herein, or sequences that have deletions from, additions to, and/or substitutions of one or more amino acid of the ADIr proteins. In certain embodiments, the polypeptide is a “recombinant” polypeptide, produced by recombinant cell that comprises one or more recombinant DNA molecules, which are typically made of heterologous polynucleotide sequences or combinations of polynucleotide sequences that would not otherwise be found in the cell.

The term “isolated protein” referred to herein means that a subject protein (1) is free of at least some other proteins with which it would typically be found in nature, (2) is essentially free of other proteins from the same source, e.g., from the same species, (3) is expressed by a cell from a different species, (4) has been separated from at least about 50 percent of polynucleotides, lipids, carbohydrates, or other materials with which it is associated in nature, (5) is not associated (by covalent or non-covalent interaction) with portions of a protein with which the “isolated protein” is associated in nature, (6) is operably associated (by covalent or non-covalent interaction) with a polypeptide with which it is not associated in nature, or (7) does not occur in nature. Such an isolated protein can be

encoded by genomic DNA, cDNA, mRNA or other RNA, of may be of synthetic origin, or any combination thereof. In certain embodiments, the isolated protein is substantially free from proteins or polypeptides or other contaminants that are found in its natural environment that would interfere with its use (therapeutic, diagnostic, prophylactic, research or otherwise).

5 The term "variant" includes a polypeptide that differs from a reference polypeptide specifically disclosed herein (e.g., SEQ ID NOS:1-28) by one or more substitutions, deletions, additions and/or insertions. Variant polypeptides are biologically active, that is, they continue to possess the enzymatic or binding activity of a reference polypeptide. Such variants may result from, for example, genetic polymorphism and/or from human
10 manipulation.

 In many instances, a biologically active variant will contain one or more conservative substitutions. A "conservative substitution" is one in which an amino acid is substituted for another amino acid that has similar properties, such that one skilled in the art of peptide chemistry would expect the secondary structure and hydrophobic nature of the polypeptide to
15 be substantially unchanged. As described above, modifications may be made in the structure of the polynucleotides and polypeptides described herein and still obtain a functional molecule that encodes a variant or derivative polypeptide with desirable characteristics.

 For example, certain amino acids may be substituted for other amino acids in a
20 protein structure without appreciable loss of interactive binding capacity with structures such as, for example, antigen-binding regions of antibodies or binding sites on substrate molecules. Since it is the interactive capacity and nature of a protein that defines that protein's biological functional activity, certain amino acid sequence substitutions can be made in a protein sequence, and, of course, its underlying DNA coding sequence, and
25 nevertheless obtain a protein with like properties. It is thus contemplated that various changes may be made in the peptide sequences of the disclosed compositions, or corresponding DNA sequences which encode said peptides without appreciable loss of their utility.

 In making such changes, the hydrophobic index of amino acids may be considered.
30 The importance of the hydrophobic amino acid index in conferring interactive biologic function on a protein is generally understood in the art (Kyte & Doolittle, 1982, incorporated herein by reference). It is accepted that the relative hydrophobic character of the amino acid contributes to the secondary structure of the resultant protein, which in turn defines the interaction of the protein with other molecules, for example, enzymes, substrates, receptors,
35 DNA, antibodies, antigens, and the like. Each amino acid has been assigned a hydrophobic index on the basis of its hydrophobicity and charge characteristics (Kyte & Doolittle, 1982). These values are: isoleucine (+4.5); valine (+4.2); leucine (+3.8); phenylalanine (+2.8);

cysteine (+2.5); methionine (+1.9); alanine (+1.8); glycine (−0.4); threonine (−0.7); serine (−0.8); tryptophan (−0.9); tyrosine (−1.3); proline (−1.6); histidine (−3.2); glutamate (−3.5); glutamine (−3.5); aspartate (−3.5); asparagine (−3.5); lysine (−3.9); and arginine (−4.5). It is known in the art that certain amino acids may be substituted by other amino acids having a similar hydropathic index or score and still result in a protein with similar biological activity, *i.e.*, still obtain a biological functionally equivalent protein. In making such changes, the substitution of amino acids whose hydropathic indices are within ± 2 is preferred, those within ± 1 are particularly preferred, and those within ± 0.5 are even more particularly preferred.

It is also understood in the art that the substitution of like amino acids can be made effectively on the basis of hydrophilicity. U.S. Patent 4,554,101 (specifically incorporated herein by reference in its entirety), states that the greatest local average hydrophilicity of a protein, as governed by the hydrophilicity of its adjacent amino acids, correlates with a biological property of the protein. As detailed in U. S. Patent 4,554,101, the following hydrophilicity values have been assigned to amino acid residues: arginine (+3.0); lysine (+3.0); aspartate (+3.0 $\pm$ 1); glutamate (+3.0 $\pm$ 1); serine (+0.3); asparagine (+0.2); glutamine (+0.2); glycine (0); threonine (−0.4); proline (−0.5 $\pm$ 1); alanine (−0.5); histidine (−0.5); cysteine (−1.0); methionine (−1.3); valine (−1.5); leucine (−1.8); isoleucine (−1.8); tyrosine (−2.3); phenylalanine (−2.5); tryptophan (−3.4). It is understood that an amino acid can be substituted for another having a similar hydrophilicity value and still obtain a biologically equivalent, and in particular, an immunologically equivalent protein. In such changes, the substitution of amino acids whose hydrophilicity values are within ± 2 is preferred, those within ± 1 are particularly preferred, and those within ± 0.5 are even more particularly preferred.

As outlined above, amino acid substitutions are generally therefore based on the relative similarity of the amino acid side-chain substituents, for example, their hydrophobicity, hydrophilicity, charge, size, and the like. Exemplary substitutions that take various of the foregoing characteristics into consideration are well known to those of skill in the art and include: arginine and lysine; glutamate and aspartate; serine and threonine; glutamine and asparagine; and valine, leucine and isoleucine.

Amino acid substitutions may further be made on the basis of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity and/or the amphipathic nature of the residues. For example, negatively charged amino acids include aspartic acid and glutamic acid; positively charged amino acids include lysine and arginine; and amino acids with uncharged polar head groups having similar hydrophilicity values include leucine, isoleucine and valine; glycine and alanine; asparagine and glutamine; and serine, threonine, phenylalanine and tyrosine. Other groups of amino acids that may represent conservative

changes include: (1) ala, pro, gly, glu, asp, gln, asn, ser, thr; (2) cys, ser, tyr, thr; (3) val, ile, leu, met, ala, phe; (4) lys, arg, his; and (5) phe, tyr, trp, his.

A variant may also, or alternatively, contain non-conservative changes. In a preferred embodiment, variant polypeptides differ from a native sequence by substitution, deletion or addition of fewer than about 10, 9, 8, 7, 6, 5, 4, 3, 2 amino acids, or even 1 amino acid. Variants may also (or alternatively) be modified by, for example, the deletion or addition of amino acids that have minimal influence on the immunogenicity, secondary structure, enzymatic activity, and/or hydropathic nature of the polypeptide.

In general, variants will display about or at least about 30%, 40%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% similarity or sequence identity or sequence homology to a reference polypeptide sequence (e.g., SEQ ID NOS:1-28). Moreover, sequences differing from the native or parent sequences by the addition (e.g., C-terminal addition, N-terminal addition, both), deletion, truncation, insertion, or substitution of about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 30, 40, 50, 60, 70, 80, 90, 100 or more amino acids but which retain the properties or activities of a parent or reference polypeptide sequence are contemplated.

In some embodiments, variant polypeptides differ from reference sequence by at least one but by less than 50, 40, 30, 20, 15, 10, 8, 6, 5, 4, 3 or 2 amino acid residue(s). In some embodiments, variant polypeptides differ from a reference sequence by about or at least 0.5% or 1% but less than 20%, 15%, 10% or 5% of the residues. (If this comparison requires alignment, the sequences should be aligned for maximum similarity. "Looped" out sequences from deletions or insertions, or mismatches, are considered differences.).

In some embodiments, an ADI_r will be about 300 to about 500 amino acids in length, including all integers and ranges in between. In specific embodiments, the ADI_r will be about 300, 305, 310, 315, 320, 325, 330, 335, 340, 345, 350, 355, 360, 365, 370, 375, 380, 385, 390, 395, 400, 405, 410, 415, 420, 425, 430, 435, 440, 445, 450, 455, 460, 465, 470, 475, 480, 485, 490, 495, or 500 amino acids in length, including all integers and ranges in between.

The term "polypeptide fragment" refers to a polypeptide that has an amino-terminal deletion, a carboxyl-terminal deletion, and/or an internal deletion or substitution of a naturally-occurring or recombinantly-produced polypeptide. In certain embodiments, a polypeptide fragment can comprise an amino acid chain at least 5 to about 400 amino acids long. It will be appreciated that in certain embodiments, fragments are at least 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, 110, 150, 200, 250, 300, 350, or 400 amino acids long. Particularly useful polypeptide fragments include functional domains, including the catalytic ADI domains of the

ADlr described herein. In the case of an ADlr, useful fragments include, but are not limited to, the catalytic domain and the α -helical domain.

Many activated PEGs used for conjugation to ADI covalently bond to lysine residues. There are usually many fewer PEG molecules attached to ADI than there are lysine residues. Both the number and distribution of attachments can be heterogeneous from molecule to molecule. Any particular lysine residue will be modified in only a small fraction of the ADI molecules. This site modification heterogeneity and low PEG occupancy can result in problems with both drug characterization and the effectiveness of PEG shielding at antigenic sites. Therefore, in certain embodiments, the selected ADlr enzymes as described herein, are modified by lysine replacement with other residue types to reduce the number of lysine residues. This produces a more uniformly pegylated protein and increases the PEG occupancy at the remaining lysine residues. Specific lysine residues chosen to be changed to other residues will be selected in order to preserve enzyme activity. This more uniform pegylation is expected to provide increased protection against proteolysis in blood and increased shielding of antigenic sites from patient antibodies.

In certain embodiments, the ADlr enzyme is modified as described in U.S. Patent No. 6,635,462. In particular, modifications of one or more of the naturally-occurring amino acid residues of an ADlr can provide for an enzyme that is more easily renatured and formulated thereby improving the manufacture of ADlr and therapeutic compositions comprising the same. In some embodiments, the ADlr enzyme is modified to remove one or more lysine residues (*e.g.*, the lysine can be substituted with another amino acid or analogues thereof, or a non-natural amino acid). In particular, in some embodiments, the ADlr enzyme is modified to be free of the lysine at a position equivalent to 112, 374, 405 or 408 of SEQ ID NO:1 (*M. hominis* ADI), or a combination of one or more of these positions. In some embodiments, the ADlr enzyme is modified to be free of one or more lysine residues, for example, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, or more lysine residues, should they be present, can be substituted with another amino acid or analogues thereof, or a non-natural amino acid. In specific embodiments, an ADlr enzyme has 5 lysine residues substituted, for example, at an equivalent position to position 7, 88, 137, 209, and 380 of SEQ ID NO: 1. In some embodiments, an ADlr enzyme has 10 lysine residues substituted, for example, at positions equivalent to positions 7, 9, 59, 88, 115, 116, 137, 178, 209, and 380 of SEQ ID NO: 1. In certain embodiments, an ADlr enzyme has 15 lysine residues substituted, for example, at positions equivalent to positions 7, 9, 59, 66, 88, 91, 93, 115, 116, 137, 141, 178, 209, 279, and at position 380 of SEQ ID NO: 1. In some embodiments, an ADlr enzyme has 21 lysine residues substituted, for example, at positions equivalent to positions 7, 9, 56, 59, 66, 88, 91, 93, 96, 115, 116, 137, 141, 178, 209, 254, 279, 325, 326, 380, and 406 of SEQ ID NO: 1.

In some instances, a native ADI_r may be found in microorganisms and is thus immunogenic and rapidly cleared from circulation in a patient. These problems may be overcome by modifying an ADI_r to create a “modified ADI_r” enzyme. Thus, certain embodiments include an ADI_r enzyme that comprises a “modifying agent,” examples of which included but are not limited to macromolecule polymers, proteins, peptides, polysaccharides, and other compounds. The ADI_r enzyme and the modifying agent may be linked by either covalent bonds or non-covalent interaction to form a stable conjugate or a stable composition to achieve a desired effect. In certain embodiments, the modified ADI_r retains the biological activity of a corresponding unmodified ADI_r (e.g., of the same or similar sequence) and has a longer half-life *in vivo* and lower antigenicity than the corresponding unmodified ADI_r. In certain embodiments, the modified ADI_r retains at least 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99% or more of the biological activity of the corresponding unmodified ADI_r. Generally, the modified ADI_r retains biological activity sufficient for therapeutic use.

In some embodiments, a modifying agent can be a polymer or a protein or a fragment thereof that is biocompatible and can increase the half-life of ADI_r in blood. The modifying agent can be either chemically coupled to ADI_r or where applicable, linked to the ADI_r via fusion protein expression.

Macromolecule polymers may include a non-peptide macromolecule polymer, which in certain embodiments, may have its own bioactivity. Suitable polymers include, but are not limited to, polyenol compounds, polyether compounds, polyvinylpyrrolidone, poly amino acids, copolymer of divinyl ether and maleic anhydride, N-(2-hydroxypropyl)-methacrylamide, polysaccharide, polyoxyethylated polyol, heparin or its fragment, poly-alkyl-ethylene glycol and its derivatives, copolymers of poly-alkyl-ethylene glycol and its derivatives, poly(vinyl ethyl ether), a,P-Poly[(2-hydroxyethyl)-DL-aspartamide], polycarboxylates, poly oxyethylene-oxymethylenes, polyacryloyl morpholines, copolymer of amino compounds and oxyolefin, poly hyaluronic acid, polyoxiranes, copolymer of ethanedioic acid and malonic acid, poly (1,3-dioxolane), ethylene and maleic hydrazide copolymer, poly sialic acid, cyclodextrin, etc. In certain embodiments, the polymer is polyethylene glycol.

The polyenol compounds as used herein include, but are not limited to, polyethylene glycol (including monomethoxy polyethylene glycol, monohydroxyl polyethylene glycol), polyvinyl alcohol, polyallyl alcohol, polybutenol and the like, and their derivatives, such as lipids.

The polyether compounds include, but are not limited to poly alkylene glycol (HO((CH₂)_xO)_nH), polypropylene glycol, polyoxyrethylene (HO((CH₂)₂O)_nH), polyvinyl alcohol ((CH₂CHOH)_n).

Poly amino acids include, but are not limited to, polymers of one type of amino acid or copolymers of two or more types of amino acids, for example, polyalanine or polylysine, or block co-polymers thereof.

Polysaccharides include but are not limited to, glucosan and its derivatives, for example dextran sulfate, cellulose and its derivatives (including methyl cellulose and carboxymethyl cellulose), starch and its derivatives, polysucrose, etc.

In specific embodiments, the ADIr is modified by coupling with protein(s) or peptide(s), wherein one or more proteins or peptides are directly or indirectly linked to ADIr. The proteins can either be naturally-existing proteins or their fragments, including but not limited to naturally existing human serum proteins or their fragments, such as thyroxine-binding protein, transthyretin, α 1-acid glycoprotein, transferrin, fibrinogen, immunoglobulin, Ig Fc regions, albumin, and fragments thereof. By "fragment" is meant any portion of a protein that is smaller than the whole protein but which retains the desired function of the protein. The ADIr as described herein may be directly or indirectly linked to a protein via a covalent bond. Direct linking means that one amino acid of ADIr is directly linked to one amino acid of the modifying protein, via a peptide bond or a disulfide bridge. Indirect linking refers to the linkages between a ADIr and a modifying protein, via originally existing chemical groups there between or specific chemical groups added through biological or chemical means, or the combination of the above-mentioned linkages.

In particular embodiments, the ADIr is modified by covalent attachment to one or more PEG molecules. ADIr covalently modified with PEG (with or without a linker) may be hereinafter referred to as "ADIr-PEG." When compared to unmodified ADIr, ADIr-PEG retains most of its enzymatic activity, is far less immunogenic or antigenic, has a greatly extended circulating half-life, and is more efficacious in the treatment of tumors.

"Polyethylene glycol" or "PEG" refers to mixtures of condensation polymers of ethylene oxide and water, in a branched or straight chain, represented by the general formula $H(OCH_2CH_2)_nOH$, wherein n is at least 4. "Polyethylene glycol" or "PEG" is used in combination with a numeric suffix to indicate the approximate weight average molecular weight thereof. For example, PEG5,000 refers to PEG having a total weight average molecular weight of about 5,000; PEG12,000 refers to PEG having a total weight average molecular weight of about 12,000; and PEG20,000 refers to PEG having a total weight average molecular weight of about 20,000.

In some embodiments, the PEG has a total weight average molecular weight of about 1,000 to about 50,000; about 3,000 to about 40,000; about 5,000 to about 30,000; about 8,000 to about 30,000; about 11,000 to about 30,000; about 12,000 to about 28,000; about 16,000 to about 24,000; about 18,000 to about 22,000; or about 19,000 to about 21,000. In some embodiments, the PEG has a total weight average molecular weight of

about 1,000 to about 50,000; about 3,000 to about 30,000; about 3,000 to about 20,000; about 4,000 to about 12,000; about 4,000 to about 10,000; about 4,000 to about 8,000; about 4,000 to about 6,000; or about 5,000. In specific embodiments, the PEG has a total weight average molecular weight of about 20,000. Generally, PEG with a molecular weight of 30,000 or more is difficult to dissolve, and yields of the formulated product may be reduced. The PEG may be a branched or straight chain. Generally, increasing the molecular weight of the PEG decreases the immunogenicity of the ADIr. The PEG may be a branched or straight chain, and in certain embodiments is a straight chain. The PEG having a molecular weight described herein may be used in conjunction with ADIr, and optionally, a biocompatible linker, to treat graft versus host disease (GVHD) or cancer, including, for example, hepatocellular carcinoma, acute myeloid leukemia, such as relapsed acute myeloid leukemia, breast cancer, ovarian cancer, colorectal cancer, gastric cancer, glioma, glioblastoma multiforme, non-small cell lung cancer (NSCLC), kidney cancer, bladder cancer, uterine cancer, esophageal cancer, brain cancer, head and neck cancers, cervical cancer, testicular cancer, stomach cancer, or esophageal cancer, among other cancers described herein.

Certain embodiments employ thiol, sulfhydryl, or cysteine-reactive PEG(s). In some embodiments, the thiol, sulfhydryl, or cysteine-reactive PEG(s) are attached to one or more naturally-occurring cysteine residues, one or more introduced cysteine residues (e.g., substitution of one or more wild-type residues with cysteine residue(s)), insertion of one or more cysteine residues), or any combination thereof (see, e.g., Doherty et al., *Bioconjug Chem.* 16:1291-98, 2005). In certain embodiments, certain of the wild-type ADI cysteines residues may be first substituted with another amino acid to prevent attachment of the PEG polymer to wild-type cysteines, for example, to prevent the PEG(s) from disrupting an otherwise desirable biological activity. Some embodiments employ one or more non-natural cysteine derivatives (e.g., homocysteine) instead of cysteine.

Non-limiting examples of thiol, sulfhydryl, or cysteine-reactive PEGs include Methoxy PEG Maleimides (M-PEG-MAL) (e.g., MW 2000, MW 5000, MW 10000, MW 20000, MW 30000, MW 40000). M-PEG-MALs react with the thiol groups on cysteine side chains in proteins and peptides to generate a stable 3-thiosuccinimidyl ether linkage. This reaction is highly selective and can take place under mild conditions at about pH 5.0-6.5 in the presence of other functional groups. Particular examples of commercially-available thiol, sulfhydryl, or cysteine-reactive PEG molecules are illustrated in Figures 1A-1D. Thus, in certain embodiments, an ADIr enzyme is conjugated to any one or more of the thiol, sulfhydryl, or cysteine-reactive PEG molecules described herein.

ADIr may be covalently bonded to a modifying agent, such as PEG, with or without a linker, although a preferred embodiment utilizes a linker. ADIr may be covalently bonded to

PEG via a biocompatible linker using methods known in the art, as described, for example, by Park *et al*, Anticancer Res., 1:373-376 (1981); and Zaplipsky and Lee, Polyethylene Glycol Chemistry: Biotechnical and Biomedical Applications, J. M. Harris, ed., Plenum Press, NY, Chapter 21 (1992), the disclosures of which are hereby incorporated by reference herein
5 in their entirety. In some instances, ADI_r may be coupled directly (i.e., without a linker) to a modifying agent such as PEG, for example, through an amino group, a sulfhydryl group, a hydroxyl group, a carboxyl group, or other group.

The linker used to covalently attach ADI_r to a modifying agent (e.g. PEG) can be any biocompatible linker. As discussed above, "biocompatible" indicates that the compound or
10 group is non-toxic and may be utilized *in vitro* or *in vivo* without causing injury, sickness, disease, or death. A modifying agent such as PEG can be bonded to the linker, for example, via an ether bond, a thiol bond, an amide bond, or other bond.

In some embodiments, suitable linkers can have an overall chain length of about 1-100 atoms, 1-80 atoms, 1-60 atoms, 1-40 atoms, 1-30 atoms, 1-20 atoms, or 1, 2, 3, 4, 5, 6,
15 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, or 20 atoms, for example, wherein the atoms in the chain comprise C, S, N, P, and/or O. In certain embodiments, a linker is optional, e.g., a PEG conjugated ADI_r enzyme does not comprise a linker. In some instances, a linker group includes, for example, a succinyl group, an amide group, an imide group, a carbamate group, an ester group, an epoxy group, a carboxyl group, a hydroxyl group, a carbohydrate,
20 a tyrosine group, a cysteine group, a histidine group, a methylene group, and combinations thereof. Particular examples of stable linkers includesuccinimide, propionic acid, carboxymethylate linkages, ethers, carbamates, amides, amines, carbamides, imides, aliphatic C-C bonds, and thio ethers. In certain embodiments, the biocompatible linker is a succinimidyl succinate (SS) group.

Other suitable linkers include an oxycarbonylimidazole group (including, for example, carbonylimidazole (CDI)), a nitro phenyl group (including, for example, nitrophenyl carbonate (NCP) or trichlorophenyl carbonate (TCP)), a trysylate group, an aldehyde group, an isocyanate group, a vinylsulfone group, or a primary amine. In certain embodiments, the linker is derived from SS, SPA, SCM, or NHS; in certain embodiments, SS, SPA, or NHS are
30 used, and in some embodiments, SS or SPA are used. Thus, in certain embodiments, potential linkers can be formed from methoxy-PEG succinimidyl succinate(SS), methoxy-PEG succinimidyl glutarate(SG), methoxy-PEG succinimidyl carbonate (SC), methoxy-PEG succinimidyl carboxymethyl ester (SCM), methoxy-PEG₂ N-hydroxy succinimide (NHS), methoxy-PEG succinimidyl butanoate (SBA), methoxy-PEG succinimidyl propionate (SPA),
35 methoxy-PEG succinimidyl glutaramide, and/or methoxy-PEG succinimidyl succinimide.

Additional examples of linkers include, but are not limited to, one or more of the following: —O—, —NH—, —S—, —C(O)—, C(O)—NH, NH—C(O)—NH, O—C(O)—NH, —

C(S)—, —CH₂—, —CH₂—CH₂—, —CH₂—CH₂—CH₂—, —CH₂—CH₂—CH₂—CH₂—, —
 O—CH₂—, —CH₂—O—, —O—CH₂—CH₂—, —CH₂—O—CH₂—, —CH₂—CH₂—O—, —
 O—CH₂—CH₂—CH₂—, —CH₂—O—CH₂—CH₂—, —CH₂—CH₂—O—CH₂—, —CH₂—
 CH₂—CH₂—O—, —O—CH₂—CH₂—CH₂—CH₂—, —CH₂—O—CH₂—CH₂—CH₂—, —
 5 CH₂—CH₂—O—CH₂—CH₂—, —CH₂—CH₂—CH₂—O—CH₂—, —CH₂—CH₂—CH₂—
 CH₂—O—, —C(O)—NH—CH₂—, —C(O)—NH—CH₂—CH₂—, —CH₂—C(O)—NH—
 CH₂—, —CH₂—CH₂—C(O)—NH—, —C(O)—NH—CH₂—CH₂—CH₂—, —CH₂—C(O)—
 NH—CH₂—CH₂—, —CH₂—CH₂—C(O)—NH—CH₂—, —CH₂—CH₂—CH₂—C(O)—NH—
 , —C(O)—NH—CH₂—CH₂—CH₂—CH₂—, —CH₂—C(O)—NH—CH₂—CH₂—CH₂—, —
 10 CH₂—CH₂—C(O)—NH—CH₂—CH₂—, —CH₂—CH₂—CH₂—C(O)—NH—CH₂—, —
 CH₂—CH₂—CH₂—C(O)—NH—CH₂—CH₂—, —CH₂—CH₂—CH₂—CH₂—C(O)—NH—, —
 —NH—C(O)—CH₂—, —CH₂—NH—C(O)—CH₂—, —CH₂—CH₂—NH—C(O)—CH₂—, —
 NH—C(O)—CH₂—CH₂—, —CH₂—NH—C(O)—CH₂—CH₂—, —CH₂—CH₂—NH—C(O)—
 CH₂—CH₂—, —C(O)—NH—CH₂—, —C(O)—NH—CH₂—CH₂—, —O—C(O)—NH—CH₂—, —
 15 —O—C(O)—NH—CH₂—CH₂—, —NH—CH₂—, —NH—CH₂—CH₂—, —CH₂—NH—
 CH₂—, —CH₂—CH₂—NH—CH₂—, —C(O)—CH₂—, —C(O)—CH₂—CH₂—, —CH₂—
 C(O)—CH₂—, —CH₂—CH₂—C(O)—CH₂—, —CH₂—CH₂—C(O)—CH₂—CH₂—, —
 CH₂—CH₂—C(O)—, —CH₂—CH₂—CH₂—C(O)—NH—CH₂—CH₂—NH—, —CH₂—
 CH₂—CH₂—C(O)—NH—CH₂—CH₂—NH—C(O)—, —CH₂—CH₂—CH₂—C(O)—NH—
 20 CH₂—CH₂—NH—C(O)—CH₂—, bivalent cycloalkyl group, —N(R₆)—, R₆ is H or an
 organic radical selected from the group consisting of alkyl, substituted alkyl, alkenyl,
 substituted alkenyl, alkynyl, substituted alkynyl, aryl and substituted aryl. Additionally, any of
 the linker moieties described herein may further include an ethylene oxide oligomer chain
 comprising 1 to 20 ethylene oxide monomer units [i.e., -(CH₂CH₂O)₁₋₂₀]. That is, the ethylene
 25 oxide oligomer chain can occur before or after the linker, and optionally in between any two
 atoms of a linker moiety comprised of two or more atoms. Also, the oligomer chain would not
 be considered part of the linker moiety if the oligomer is adjacent to a polymer segment and
 merely represent an extension of the polymer segment.

Specific exemplary PEG molecules and linkers are described in Table A below.

Table A. PEG and Linkers		
PEG	Linker	Comments
Methoxy-PEG succinimidyl hexanoate	amide	pH 7-8, lower reactivity
Methoxy-PEG succinimidyl butanoate (SBA)	amide	pH 7-8, longer hydrolysis time than SPA (~23 min)

Methoxy-PEG succinimidyl propionate (SPA)	amide	Tan, 1998, Metase; Basu, IFN; Games, Phe Am. Lyase; better than SCM (~16 min)
Methoxy-PEG succinimidyl carboxymethyl ester (SCM)	amide	pH 7-8, RT, 1 hr rxn time, extremely reactive, 0.75 min at pH 8, 25°C, arginase
Methoxy-PEG succinimidyl glutaramide	amide	pH 7-8, RT, 90% complete
Methoxy-PEG succinimidyl succinamide	amide	pH 7-8, RT, 95% complete
MethoxyPEG2 NHS		Gamez, Phe Am. Lyase; Basu, IFNa2a40K, Nulasta (G-CSF), pegfilgrastim (G-CSF)
Methoxy-PEG succinimidyl carbonate (SC)	urethane	Hydrolysis ½ longer than SCM, Wang, 2006, M. art. ADI, Basu, IFNa2b, slow release
Methoxy-PEG succinimidyl glutarate (SG)	ester	Yang, 2004, Metase
Methoxy-PEG succinimidyl succinate (SS)	ester	Adenosine deaminase, asparaginase, ADI-PEG 20
PEG-maleimide		
PEG-vinylsulfone		
PEG-iodoacetamide		
orthopyridyl disulfide-PEG		

In certain embodiments, the ADIr enzyme comprises one or more PEG molecules and/or linkers as described herein (e.g., in Table A).

5 The attachment of PEG to ADIr increases the circulating half-life of ADIr. Generally, PEG is attached to a primary amine of ADIr. Selection of the attachment site of PEG, or other modifying agent, on the ADIr is determined by the role of each of the sites within the active domain of the protein, as would be known to the skilled artisan. PEG may be attached to the primary amines of ADIr without substantial loss of enzymatic activity. For example, the lysine residues present in ADIr are all possible points at which ADIr as described herein can be attached to PEG via a biocompatible linker, such as SS, SPA, SCM, SSA and/or NHS.

10 PEG may also be attached to other sites on ADIr, as would be apparent to one skilled in the art in view of the present disclosure.

From 1 to about 30 PEG molecules may be covalently bonded to ADIr. In certain embodiments, ADIr is modified with (i.e., comprises) one PEG molecule. In some

15 embodiments, the ADIr is modified with more than one PEG molecule. In particular embodiments, the ADIr is modified with about 1 to about 10, or from about 7 to about 15

PEG molecules, or from about 2 to about 8 or about 9 to about 12 PEG molecules. In some embodiments, the ADIr is modified with about 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, or 15 PEG molecules. In specific embodiments, ADIr is modified with 4.5 – 5.5 PEG molecules per ADIr. In some embodiment, ADIr is modified with 5 ± 1.5 PEG molecules.

5 In certain embodiments, about 15% to about 70% of the primary amino groups in ADIr are modified with PEG, in some embodiments about 20% to about 65%, about 25% to about 60%, or in certain embodiments about 30% to about 55%, or 45% to about 50%, or in some embodiments about 50% of the primary amino groups in arginine deiminase are modified with PEG. When PEG is covalently bonded to the end terminus of ADIr, it may be
10 desirable to have only 1 PEG molecule utilized. Increasing the number of PEG units on ADIr increases the circulating half-life of the enzyme. However, increasing the number of PEG units on ADIr decreases the specific activity of the enzyme. Thus, a balance needs to be achieved between the two, as would be apparent to one skilled in the art in view of the present disclosure.

15 In some embodiments, a common feature of biocompatible linkers is that they attach to a primary amine of arginine deiminase via a succinimide group. Once coupled with ADIr, SS-PEG has an ester linkage next to the PEG, which may render this site sensitive to serum esterase, which may release PEG from ADIr in the body. SPA-PEG and PEG2-NHS do not have an ester linkage, so they are not sensitive to serum esterase.

20 PEG which is attached to the protein may be either a straight chain, as with SS-PEG, SPA-PEG and SC-PEG, or a branched chain of PEG may be used, as with PEG2-NHS.

In certain embodiments, pegylation sites associated with ADIr located at or adjacent to the catalytic region of the enzyme are modified. In certain embodiments, the phrase “pegylation site” is defined as any site or position of ADI or a ADIr that may be covalently
25 modified with polyethylene glycol. A “pegylation site” can be considered located at or adjacent the catalytic region of the enzyme where pegylation of the site results in a significant reduction in catalytic activity of the enzyme. The pegylation of such sites has traditionally resulted in the inactivation of the enzyme. For example, ADI from *Mycoplasma hominis* has a lysine at the 112 position which can be considered to be at or adjacent the
30 catalytic region of the enzyme. The attachment of PEG to this lysine at the 112 position can inactivate the enzyme. In addition, ADI from *Mycoplasma hominis* has a cysteine at the 397 position which can be considered to be at or adjacent the catalytic region of the enzyme. The amino acid substitutions for cysteine at the 397 position can inactivate the enzyme. In particular, substituting alanine, histidine, arginine, serine, lysine or tyrosine for cysteine at
35 the 397 position can result in a loss of all detectable enzyme activity. ADI from *Mycoplasma hominis* also has three lysine residues located near this conserved cysteine, in particular

Lys374, Lys405 and Lys408. The attachment of PEG to Lys374, Lys405, Lys408 or combinations thereof can inactivate the enzyme.

It is to be understood that ADIr derived from other organisms may also have pegylation sites corresponding to 112 position of ADI from *Mycoplasma hominis*. In addition, ADI from some organisms may have lysine residues corresponding to the same general location as the 112 position of ADI from *Mycoplasma hominis*. The location of lysine in ADI from such organisms are known to the skilled person and are described in U.S. Patent No. 6,635,462.

Thus, some embodiments provide for certain amino acid substitutions in the polypeptide chain of ADIr. These amino acid substitutions provide for modified ADIr that loses less activity when modified by a modifying agent, e.g., upon pegylation. By eliminating pegylation sites, or other known modification sites, at or adjacent to the catalytic region of enzyme, optimal modification, e.g., pegylation, can be achieved without the loss of activity.

In some embodiments, for example, as noted above, the amino acid substitutions employ non-natural amino acids for conjugation to PEG or other modifying agent (see, e.g., de Graaf et al., Bioconjug Chem. 20:1281-95, 2009). Certain embodiments thus include an ADIr enzyme that is conjugated to one or more PEGs via one or more non-natural amino acids. In some embodiments the non-natural amino acid comprises a side chain having a functional group selected from the group consisting of: an alkyl, aryl, aryl halide, vinyl halide, alkyl halide, acetyl, ketone, aziridine, nitrile, nitro, halide, acyl, keto, azido, hydroxyl, hydrazine, cyano, halo, hydrazide, alkenyl, alkynyl, ether, thio ether, epoxide, sulfone, boronic acid, boronate ester, borane, phenylboronic acid, thiol, seleno, sulfonyl, borate, boronate, phospho, phosphono, phosphine, heterocyclic-, pyridyl, naphthyl, benzophenone, a constrained ring such as a cyclooctyne, thioester, enone, imine, aldehyde, ester, thioacid, hydroxylamine, amino, carboxylic acid, alpha-keto carboxylic acid, alpha or beta unsaturated acids and amides, glyoxyl amide, and an organosilane group. In some embodiments, the non-natural amino acid is selected from the group consisting of: p-acetyl-L-phenylalanine, O-methyl-L-tyrosine, L-3-(2-naphthyl)alanine, 3-methyl-phenylalanine, O-4-allyl-L-tyrosine, homocysteine, 4-propyl-L-tyrosine, tri-O-acetyl-GlcNAc β -serine, β -O-GlcNAc-L-serine, tri-O-acetyl-GalNAc- α -threonine, α -GalNAc-L-threonine, L-Dopa, a fluorinated phenylalanine, isopropyl-L-phenylalanine, p-azido-L-phenylalanine, p-acyl-L-phenylalanine, p-benzoyl-L-phenylalanine, L-phosphoserine, phosphoserine, phosphotyrosine, p-iodo-phenylalanine, p-bromophenylalanine, p-amino-L-phenylalanine, and isopropyl-L-phenylalanine.

While ADIr-PEG is the illustrative modified ADIr described herein, as would be recognized by the skilled person ADIr may be modified with other polymers or appropriate

molecules for the desired effect, in particular reducing antigenicity and increasing serum half-life.

It is to be understood that some embodiments are based on the understanding that certain structural characteristics of arginine deiminase may prevent or interfere with the proper and rapid renaturation when produced via recombinant technology. In particular, these structural characteristics hinder or prevent the enzyme from assuming an active conformation during recombinant production. In some embodiments, the term “active conformation” is defined as a three-dimensional structure that allows for enzymatic activity by unmodified or modified arginine deiminase. The active conformation may, in particular, be necessary for catalyzing the conversion of arginine into citrulline. The term “structural characteristic” may be defined as any trait, quality or property of the polypeptide chain resulting from a particular amino acid or combination of amino acids. For instance, arginine deiminase may contain an amino acid that results in a bend or kink in the normal peptide chain and thus hinders the enzyme from assuming an active conformation during renaturation of the enzyme. In particular, arginine deiminase from *Mycoplasma hominis* has a proline at the 210 position that may result in a bend or kink in the peptide chain, making it more difficult to renature the enzyme during recombinant production. It is to be understood that arginine deiminase derived from other organisms may also have sites corresponding to the 210 position of arginine deiminase from *Mycoplasma hominis*.

Some embodiments thus provide for certain amino acid substitutions in the polypeptide chain of wild type arginine deiminases. Examples include substitutions that eliminate the problematic structural characteristics in the peptide chain of arginine deiminase. Also included are substitutions that provide for improved renaturation of the modified arginine deiminase. These amino acid substitutions make possible rapid renaturation of modified arginine deiminases using reduced amounts of buffer. These amino acid substitutions may also provide for increased yields of renatured modified arginine deiminase. In some embodiments, the modified arginine deiminase has an amino acid substitution at P210 or the equivalent residue. As mentioned above, arginine deiminase derived from *Mycoplasma hominis* has the amino acid proline located at the 210 position. While not limiting the present invention, it is believed that the presence of the amino acid proline at position 210 results in a bend or kink in the normal polypeptide chain that increases the difficulty of renaturing (i.e., refolding) certain arginine deiminase enzymes. Substitutions for proline at position 210 make possible the rapid renaturation of modified arginine deiminase using reduced amounts of buffer. Substitutions for proline at position 210 (or the equivalent residue) may also provide for increased yields of renatured modified arginine deiminase. In some embodiments, the proline at position 210 (or the equivalent residue) is substituted with serine. Non-limiting examples of other substitutions include

Pro210 to Thr210, Pro210 to Arg210, Pro210 to Asn210, Pro210 to Gln210 or Pro210 to Met210. By eliminating those structural characteristics associated with the amino acid of position 210 of the wild-type arginine deiminase, optimal refolding of the enzyme can be achieved.

5 The methods provided herein can involve either *in vitro* or *in vivo* applications. In the case of *in vitro* applications, including cell culture applications, the compounds described herein can be added to the cells in cultures and then incubated. The compounds described herein may also be used to facilitate the production of monoclonal and/or polyclonal antibodies, using antibody production techniques well known in the art. The monoclonal and/or polyclonal antibodies can then be used in a wide variety of diagnostic applications, as would be apparent to one skilled in the art.

10 The *in vivo* means of administration of the compounds or compositions described herein will vary depending upon the intended application. Administration of the ADIr compositions described herein, in pure form or in an appropriate pharmaceutical composition, can be carried out via any of the accepted modes of administration of agents for serving similar utilities. The pharmaceutical compositions can be prepared by combining ADIr, e.g., ADIr-PEG, ADIr-PEG 20, with an appropriate physiologically acceptable carrier, diluent or excipient, and may be formulated into preparations in solid, semi-solid, liquid or gaseous forms, such as tablets, capsules, powders, granules, ointments, solutions, suppositories, injections, inhalants, gels, microspheres, and aerosols. In addition, other pharmaceutically active ingredients (including other anti-cancer agents as described elsewhere herein) and/or suitable excipients such as salts, buffers and stabilizers may, but need not, be present within the composition.

15 Administration may be achieved by a variety of different routes, including oral, parenteral, nasal, intravenous, intradermal, subcutaneous or topical. Modes of administration depend upon the nature of the condition to be treated or prevented. Thus, an ADIr enzyme (e.g., ADIr-PEG, ADIr-PEG 20) can be administered orally, intranasally, intraperitoneally, parenterally, intravenously, intralymphatically, intratumorally, intramuscularly, interstitially, intra-arterially, subcutaneously, intraocularly, intrasynovial, transepithelial, and transdermally. An amount that, following administration, reduces, inhibits, prevents or delays the progression and/or metastasis of a cancer is considered effective. In certain embodiments, the ADIr compositions herein increase median survival time of patients by a statistically significant amount. In some embodiments, the ADIr treatments described herein increase median survival time of a patient by 4 weeks, 5 weeks, 6 weeks, 7 weeks, 8 weeks, 9 weeks, 10 weeks, 15 weeks, 20 weeks, 25 weeks, 30 weeks, 40 weeks, or longer. In certain embodiments, ADIr treatments increase median survival time of a patient by 1 year, 2 years, 3 years, or longer. In some embodiments, the ADIr treatments described herein

increase progression-free survival by 2 weeks, 3 weeks, 4 weeks, 5 weeks, 6 weeks, 7 weeks, 8 weeks, 9 weeks, 10 weeks or longer. In certain embodiments, the ADIr treatments described herein increase progression-free survival by 1 year, 2 years, 3 years, or longer.

In certain embodiments, the amount administered is sufficient to result in tumor regression, as indicated by a statistically significant decrease in the amount of viable tumor, for example, at least a 10%, 20%, 30%, 40%, 50% or greater decrease in tumor mass, or by altered (e.g., decreased with statistical significance) scan dimensions. In certain embodiments, the amount administered is sufficient to result in stable disease. In certain embodiments, the amount administered is sufficient to result in clinically relevant reduction in symptoms of a particular disease indication known to the skilled clinician.

In certain embodiments the amount administered is sufficient to inhibit NO synthesis, inhibit angiogenesis, and or is sufficient to induce apoptosis in tumor cells or any combination thereof. NO synthesis, angiogenesis and apoptosis may be measured using methods known in the art, see, e.g., *Current Protocols in Immunology* or *Current Protocols in Molecular Biology*, John Wiley & Sons, New York, N.Y.(2009 and updates thereto); Ausubel *et al.*, *Short Protocols in Molecular Biology*, 3rd ed., Wiley & Sons, 1995; and other like references. In some embodiments the amount administered inhibits NO synthesis and inhibits the growth of melanoma and complements, adds to, or synergizes with other chemotherapies as described herein, such as cisplatin. Accordingly, some embodiments provide a method of treating melanoma by administering ADIr-PEG 20 in combination with cisplatin, wherein the treatment depletes endogenous nitric oxide (NO).

The precise dosage and duration of treatment is a function of the disease being treated and may be determined empirically using known testing protocols or by testing the compositions in model systems known in the art and extrapolating therefrom. Controlled clinical trials may also be performed. Dosages may also vary with the severity of the condition to be alleviated. A pharmaceutical composition is generally formulated and administered to exert a therapeutically useful effect while minimizing undesirable side effects. The composition may be administered one time, or may be divided into a number of smaller doses to be administered at intervals of time. For any particular subject, specific dosage regimens may be adjusted over time according to the individual need.

The ADIr (e.g., ADIr-PEG) compositions may be administered alone or in combination with other known cancer treatments, such as radiation therapy, chemotherapy, transplantation, immunotherapy, hormone therapy, photodynamic therapy, *etc.* The compositions may also be administered in combination with antibiotics.

The ADIr compositions may also be administered alone or in combination with ADI-PEG 20 therapy. In certain embodiments, the ADIr as described herein are used in patients who have been treated with ADI-PEG 20 and who have developed anti-ADI-PEG 20

antibodies. Such patients no longer benefit from ADI-PEG 20 treatment as the enzyme is neutralized by the antibodies. Thus, in certain embodiments, the invention provides a method of treating, ameliorating the symptoms of, or inhibiting the progression of a cancer comprising administering to a patient in need thereof a therapeutically effective amount of a composition comprising ADI-PEG 20, and after a period of time, administering to the patient a composition comprising an ADIr as described herein, thereby treating, ameliorating the symptoms of, or inhibiting the progression of the cancer.

In some embodiments of the method, the period of time is determined by detecting a predetermined level of anti-ADI-PEG 20 antibodies in the patient, wherein the composition comprising an ADIr is administered following detection of the predetermined level of said anti-ADI-PEG 20 antibodies. In certain embodiments, threshold level(s) or predetermined levels of anti-ADI-PEG 20 antibodies in patients to be treated with ADI-PEG 20 and an ADIr enzyme described herein can be established. A “predetermined threshold level” (also referred to as “predetermined level” or “predetermined cut-off value”), or sometimes referred to as a predetermined cut off, of anti-ADI-PEG 20 antibodies may be established using methods known in the art, for example, using Receiver Operator Characteristic curves or “ROC” curves. In some embodiments, even very low levels of anti-ADI-PEG 20 antibodies is deemed sufficient to warrant switching treatment from ADI-PEG 20 to an ADIr-PEG described herein. In certain embodiments, an appropriate level of anti-ADI-PEG 20 that will determine when to terminate ADI-PEG 20 treatment and begin treatment with an ADIr composition described herein can be determined by the skilled clinician.

In some embodiments, the period of time is determined by detecting or otherwise observing ADI activity in the patient, wherein the composition is administered following detection or observation of a predetermined level of ADI activity. In particular embodiments, the composition is administered following detection or observation of a reduced level of ADI activity in the patient. ADI activity can be measured directly, for example, by assaying a biological sample for at least one indicator of ADI activity, or indirectly, for example, by observing the desired or intended effect of the ADI-PEG 20 treatment. In certain embodiments, an appropriate level of ADI activity that will determine when to terminate ADI-PEG 20 treatment and begin treatment with an ADIr-PEG described herein can be determined by the skilled clinician.

Typical routes of administering these and related pharmaceutical compositions thus include, without limitation, oral, topical, transdermal, inhalation, parenteral, sublingual, buccal, rectal, vaginal, and intranasal. The term parenteral as used herein includes subcutaneous injections, intravenous, intramuscular, intrasternal injection or infusion techniques.

Certain pharmaceutical compositions are formulated so as to allow the active ingredients contained therein to be bioavailable upon administration of the composition to a patient. Compositions that will be administered to a subject or patient may take the form of one or more dosage units, where for example, a tablet may be a single dosage unit, and a container of a herein described ADIr composition in aerosol form may hold a plurality of dosage units. Actual methods of preparing such dosage forms are known, or will be apparent, to those skilled in this art; for example, see *Remington: The Science and Practice of Pharmacy*, 20th Edition (Philadelphia College of Pharmacy and Science, 2000). The composition to be administered will, in any event, contain a therapeutically effective amount of an ADIr-PEG described herein, such as ADIr-PEG 20, for treatment of a disease or condition of interest in accordance with teachings herein. In certain embodiments, the pharmaceutical or therapeutic compositions are sterile and/or pyrogen-free.

A pharmaceutical composition may be in the form of a solid or liquid. In some embodiments, the carrier(s) are particulate, so that the compositions are, for example, in tablet or powder form. The carrier(s) may be liquid, with the compositions being, for example, an oral oil, injectable liquid or an aerosol, which is useful in, for example, inhalatory administration. When intended for oral administration, the pharmaceutical composition is generally either solid or liquid form, where semi-solid, semi-liquid, suspension and gel forms are included within the forms considered herein as either solid or liquid.

As a solid composition for oral administration, the pharmaceutical composition may be formulated into a powder, granule, compressed tablet, pill, capsule, chewing gum, wafer or the like. Such a solid composition will typically contain one or more inert diluents or edible carriers. In addition, one or more of the following may be present: binders such as carboxymethylcellulose, ethyl cellulose, microcrystalline cellulose, gum tragacanth or gelatin; excipients such as starch, lactose or dextrins, disintegrating agents such as alginic acid, sodium alginate, Primogel, corn starch and the like; lubricants such as magnesium stearate or Sterotex; glidants such as colloidal silicon dioxide; sweetening agents such as sucrose or saccharin; a flavoring agent such as peppermint, methyl salicylate or orange flavoring; and a coloring agent. When the pharmaceutical composition is in the form of a capsule, for example, a gelatin capsule, it may contain, in addition to materials of the above type, a liquid carrier such as polyethylene glycol or oil.

The pharmaceutical composition may be in the form of a liquid, for example, an elixir, syrup, solution, emulsion or suspension. The liquid may be for oral administration or for delivery by injection, as two examples. When intended for oral administration, preferred composition contain, in addition to the present compounds, one or more of a sweetening agent, preservatives, dye/colorant and flavor enhancer. In a composition intended to be

administered by injection, one or more of a surfactant, preservative, wetting agent, dispersing agent, suspending agent, buffer, stabilizer and isotonic agent may be included.

The liquid pharmaceutical compositions, whether they be solutions, suspensions or other like form, may include one or more of the following adjuvants: sterile diluents such as water for injection, saline solution, in certain embodiments, physiological saline, Ringer's solution, isotonic sodium chloride, fixed oils such as synthetic mono or diglycerides which may serve as the solvent or suspending medium, polyethylene glycols, glycerin, propylene glycol or other solvents; antibacterial agents such as benzyl alcohol or methyl paraben; antioxidants such as ascorbic acid or sodium bisulfite; chelating agents such as ethylenediaminetetraacetic acid; buffers such as acetates, citrates or phosphates and agents for the adjustment of tonicity such as sodium chloride or dextrose. The parenteral preparation can be enclosed in ampoules, disposable syringes or multiple dose vials made of glass or plastic. Physiological saline is a preferred adjuvant. An injectable pharmaceutical composition is preferably sterile.

A liquid pharmaceutical composition intended for either parenteral or oral administration should contain an amount of ADI_r as herein disclosed (e.g., ADI_r-PEG, ADI_r-PEG 20) such that a suitable dosage will be obtained. Typically, this amount is at least 0.01% of ADI_r in the composition. When intended for oral administration, this amount may be varied to be between 0.1 and about 70% of the weight of the composition. Certain oral pharmaceutical compositions contain between about 4% and about 75% of ADI_r. In certain embodiments, pharmaceutical compositions and preparations are prepared so that a parenteral dosage unit contains between 0.01 to 10% by weight of ADI_r prior to dilution.

The pharmaceutical composition may be intended for topical administration, in which case the carrier may suitably comprise a solution, emulsion, ointment or gel base. The base, for example, may comprise one or more of the following: petrolatum, lanolin, polyethylene glycols, bee wax, mineral oil, diluents such as water and alcohol, and emulsifiers and stabilizers. Thickening agents may be present in a pharmaceutical composition for topical administration. If intended for transdermal administration, the composition may include a transdermal patch or iontophoresis device. The pharmaceutical composition may be intended for rectal administration, in the form, for example, of a suppository, which will melt in the rectum and release the drug. The composition for rectal administration may contain an oleaginous base as a suitable nonirritating excipient. Such bases include, without limitation, lanolin, cocoa butter and polyethylene glycol.

The pharmaceutical composition may include various materials, which modify the physical form of a solid or liquid dosage unit. For example, the composition may include materials that form a coating shell around the active ingredients. The materials that form the coating shell are typically inert, and may be selected from, for example, sugar, shellac, and

other enteric coating agents. Alternatively, the active ingredients may be encased in a gelatin capsule. The pharmaceutical composition in solid or liquid form may include an agent that binds to ADIr (e.g., ADIr-PEG) and thereby assists in the delivery of the compound. Suitable agents that may act in this capacity include monoclonal or polyclonal antibodies, one or more proteins or a liposome. The pharmaceutical composition may consist essentially of dosage units that can be administered as an aerosol. The term aerosol is used to denote a variety of systems ranging from those of colloidal nature to systems consisting of pressurized packages. Delivery may be by a liquefied or compressed gas or by a suitable pump system that dispenses the active ingredients. Aerosols may be delivered in single phase, bi-phasic, or tri-phasic systems in order to deliver the active ingredient(s). Delivery of the aerosol includes the necessary container, activators, valves, sub-containers, and the like, which together may form a kit. One of ordinary skill in the art, without undue experimentation may determine preferred aerosols.

The pharmaceutical compositions may be prepared by methodology well known in the pharmaceutical art. For example, a pharmaceutical composition intended to be administered by injection can be prepared by combining a composition that comprises ADIr (e.g., ADIr-PEG) as described herein and optionally, one or more of salts, buffers and/or stabilizers, with sterile, distilled water so as to form a solution. A surfactant may be added to facilitate the formation of a homogeneous solution or suspension. Surfactants are compounds that non-covalently interact with the ADIr (e.g., ADIr-PEG) composition so as to facilitate dissolution or homogeneous suspension of the ADIr (e.g., ADIr-PEG) in the aqueous delivery system.

The compositions may be administered in a therapeutically effective amount, which will vary depending upon a variety of factors including the activity of the specific compound (e.g., ADIr-PEG) employed; the metabolic stability and length of action of the compound; the age, body weight, general health, sex, and diet of the patient; the mode and time of administration; the rate of excretion; the drug combination; the severity of the particular disorder or condition; and the subject undergoing therapy.

A therapeutically effective amount of a compound described herein is an amount that is effective to inhibit tumor growth. Generally, treatment is initiated with small dosages which can be increased by small increments until the optimum effect under the circumstances is achieved. Generally, a therapeutic dosage of compounds may be from about 1 to about 200 mg/kg twice a week to about once every two weeks. For example, the dosage may be about 1 mg/kg once a week as a 2 ml intravenous injection to about 20 mg/kg once every 3 days. In some embodiments, the dose may be from about 50 IU/m² to about 700 IU/m², administered about once every 3 days, about once a week, about twice a week, or about once every 2 weeks. In certain embodiments, the dose may be about 50 IU/m², 60 IU/m², 70

IU/m², 80 IU/m², 90 IU/m², 100 IU/m², 110 IU/m², 120 IU/m², 130 IU/m², 140 IU/m², 150 IU/m², 160 IU/m², 170 IU/m², 180 IU/m², 190 IU/m², 200 IU/m², 210 IU/m², 220 IU/m², 230 IU/m², 240 IU/m², 250 IU/m², 260 IU/m², 270 IU/m², 280 IU/m², 290 IU/m², 300 IU/m², 310 IU/m², about 320 IU/m², about 330 IU/m², 340 IU/m² about 350 IU/m², 360 IU/m², 370 IU/m², 380 IU/m², 390 IU/m², 400 IU/m², 410 IU/m², 420 IU/m², 430 IU/m², 440 IU/m², 450 IU/m², 500 IU/m², 550 IU/m², 600 IU/m², 620 IU/m², 630 IU/m², 640 IU/m², 650 IU/m², 660 IU/m², 670 IU/m², 680 IU/m², 690 IU/m², or about 700 IU/m² administered about once every 3 days, about once a week, about twice a week, or about once every 2 weeks. In certain embodiments, the dose may be modified as desired by the skilled clinician.

In some instances, the optimum dosage with ADIr-SS-PEG5,000 may be about twice a week, while the optimum dosage with ADIr-SS-PEG20,000 may be from about once a week to about once every two weeks. In certain embodiments, the optimum dosage with ADIr-SS-PEG20,000 may be about twice a week.

ADIr (e.g., ADIr-PEG) may be mixed with a phosphate buffered saline solution, or any other appropriate solution known to those skilled in the art, prior to injection. In some embodiments, a liquid composition comprising ADIr-PEG comprises about 10 to about 12 mg of ADIr; about 20 to about 40 mg of polyethylene glycol; about 1.27 mg $\pm$ 5% monobasic sodium phosphate, USP; about 3 mg $\pm$ 5% dibasic sodium phosphate, USP; about 7.6 mg $\pm$ 5% sodium chloride, USP; at a pH of about 6.6 to about 7; in an appropriate amount of water for injection (e.g., about 1 ml or about 2 ml).

In some embodiments, a liquid composition comprising an ADIr-PEG comprises histidine – HCl, and in certain embodiments, the composition buffer is from about 0.0035 M Histidine-HCl to about 0.35 M Histidine-HCl. In one particular embodiment, the composition is formulated in a buffer comprising 0.035 M Histidine-HCl at pH 6.8 with 0.13 M sodium chloride. In certain embodiments, the composition is formulated in a buffer comprising 0.02 M sodium phosphate buffer at pH 6.8 with 0.13 M sodium chloride. In some embodiments, a liquid composition comprising ADIr-PEG comprises about 10 to about 12 mg of ADIr; about 20 to about 40 mg of polyethylene glycol; about 5.4 mg $\pm$ 5% Histidine, USP; about 7.6 mg $\pm$ 5% sodium chloride, USP; at a pH of about 6.6 to about 7; in an appropriate amount of water for injection (e.g., about 1 ml or about 2 ml).

In some embodiments, a composition comprising ADIr (e.g., ADIr-PEG) has a pH of about 5 to about 9, about 6 to about 8, or about 6.5 to about 7.5. In some embodiments, the composition comprising ADIr has a pH of about 6.8 ± 1.0 .

In some embodiments, free PEG in a composition comprising ADIr (e.g., ADIr-PEG) is between 1-10%. In some embodiments, it is less than 7%, less than 6%, less than 5%, less than 4%, less than 3%, less than 2% or less than 1% of the total PEG. In certain embodiments, the unmodified ADIr in a composition comprising ADIr (e.g., ADIr-PEG) is less

than about 1%, 0.9%, 0.8%, 0.7%, 0.6%, 0.5%, 0.4%, 0.3%, 0.2% or less than 0.1%.

Generally, compositions comprising ADIr-PEG have total impurities less than or equal to about 4%, 3%, 2%, 1.5%, 1% or 0.5%. In some embodiments, the endotoxin limit meets the requirements stated in USP, i.e., ≤ 50 EU/mL.

5 In some embodiments, the free sulfhydryl in a composition comprising ADIr (e.g., ADIr-PEG) is greater than about 90%. In some embodiments, the free sulfhydryl in a composition comprising ADIr or ADIr-PEG is about 91%, about 92%, about 93%, about 94% or about 95%, about 96% about 97%, about 98% about 99% or more.

10 In some embodiments, the ADIr (e.g., ADIr-PEG) in a composition has a K_m of from about 0.1 μM or 0.5 μM to about 15 μM , or is from about 1 μM to about 12 μM , about 1 μM to about 10 μM , about 1.5 μM to about 9 μM , about 1.5 μM to about 8 μM or about 1.5 μM to about 7 μM . In certain embodiments, the ADIr (e.g., ADIr-PEG) in a composition has a K_m of about 1.0 μM to about 10 μM or about 1.5 μM to about 6.5 μM . In some embodiments, the ADIr or ADIr-PEG in a composition has a K_m of about, at least about, or less than about 0.1
15 μM , about 0.5 μM , about 1.0 μM , about 1.5 μM , about 2 μM , about 2.5 μM , about 3 μM , about 3.5 μM , about 4 μM , about 4.5 μM , about 5 μM , about 5.5 μM , about 6 μM , about 6.5 μM , or about 7 μM , or about 8 μM , or about 9 μM , or about 10 μM .

 In some embodiments, the ADIr (e.g., ADIr-PEG) in a composition has a K_{cat} of from about 0.5 sec^{-1} to about 80 sec^{-1} , or about 0.5 sec^{-1} to about 70 sec^{-1} , or about 0.5 sec^{-1} to about 60 sec^{-1} , or about 0.5 sec^{-1} to about 50 sec^{-1} , or about 0.5 sec^{-1} to about 40 sec^{-1} , or about 0.5 sec^{-1} to about 30 sec^{-1} , or about 0.5 sec^{-1} to about 20 sec^{-1} , or about 0.5 sec^{-1} to about 15 sec^{-1} , or is from about 0.5 sec^{-1} to about 80 sec^{-1} , or about 1 sec^{-1} to about 80 sec^{-1} , or about 5 sec^{-1} to about 80 sec^{-1} , or about 10 sec^{-1} to about 80 sec^{-1} , or about 20 sec^{-1} to about 80 sec^{-1} , or about 30 sec^{-1} to about 80 sec^{-1} , or about 40 sec^{-1} to about 80 sec^{-1} , or about 50 sec^{-1} to about 80 sec^{-1} , or about 60 sec^{-1} to about 80 sec^{-1} , or about 70 sec^{-1} to about 80 sec^{-1} , or about 1 sec^{-1} to about 12 sec^{-1} , about 1 sec^{-1} to about 10 sec^{-1} , about 1.5 sec^{-1} to about 9 sec^{-1} , about 2 sec^{-1} to about 8 sec^{-1} or about 2.5 sec^{-1} to about 7 sec^{-1} . In certain embodiments, the ADIr (e.g., ADIr-PEG) in a composition has a K_{cat} of about 2.5 sec^{-1} to about 7.5 sec^{-1} . In some embodiments, the ADIr or ADIr-PEG in a composition has a
25 K_{cat} of about or at least about 2.5 sec^{-1} , about 3 sec^{-1} , about 3.5 sec^{-1} , about 4 sec^{-1} , about 4.5 sec^{-1} , about 5 sec^{-1} , about 5.5 sec^{-1} , about 6 sec^{-1} , about 6.5 sec^{-1} , about 7 sec^{-1} , about 7.5 sec^{-1} or about 8 sec^{-1} , about 10 sec^{-1} , about 15 sec^{-1} , about 20 sec^{-1} , about 25 sec^{-1} , about 30 sec^{-1} , about 35 sec^{-1} , about 40 sec^{-1} , about 45 sec^{-1} , about 50 sec^{-1} , about 55 sec^{-1} , about 60 sec^{-1} , about 65 sec^{-1} , about 70 sec^{-1} , about 75 sec^{-1} , about 80 sec^{-1} , about 85 sec^{-1} ,
30 about 90 sec^{-1} , about 95 sec^{-1} , or about 100 sec^{-1} .

 In some embodiments, the ADIr (e.g., ADIr-PEG) in a composition has a conductivity (also referred to in the art as specific conductance) of about 5 mS/cm to about 20 mS/cm, or

about 5 mS/cm to about 15 mS/cm, about 7 mS/cm to about 15 mS/cm, about 9 mS/cm to about 15 mS/cm or about 10 mS/cm to about 15 mS/cm. In some embodiments, the ADIr (e.g., ADIr-PEG) in a composition has a conductivity of about 9 mS/cm, about 10 mS/cm, about 11 mS/cm, about 12 mS/cm or about 13 mS/cm, about 14 mS/cm or about 15 mS/cm.

5 In certain embodiments, the ADIr (e.g., ADIr-PEG) in a composition has a conductivity of about 13 mS/cm $\pm$ 1.0 mS/cm.

In some embodiments, the ADIr (e.g., ADIr-PEG) in a composition has an osmolality of about 50 mOsm/kg to about 500 mOsm/kg, about 100 mOsm/kg to about 400 mOsm/kg, about 150 mOsm/kg to about 350 mOsm/kg, about 200 mOsm/kg to about 350 mOsm/kg or
10 about 250 mOsm/kg to about 350 mOsm/kg. In certain embodiments, the ADIr (e.g., ADIr-PEG) in a composition has an osmolality of about 300 $\pm$ 30 mOsm/kg.

In some embodiments, the protein concentration is about 11.0 $\pm$ 1.0 mg/mL. In certain embodiments, the protein concentration is between about 8 and about 15 mg/mL. In certain embodiments, the protein concentration is about 8, 9, 10, 10.5, 11, 11.5, 12, 12.5, 13,
15 13.5, 14, or 15 mg/mL.

In some embodiments, the specific enzyme activity is between about 5.0 and 90 IU/mg or between about 5 and 55 IU/mg, where 1 IU is defined as the amount of enzyme that converts one μ mol of arginine into one μ mol of citrulline and 1 μ mol of ammonia in one minute at 37° C and the potency is 100 $\pm$ 20 IU/mL. In certain embodiments, the specific
20 enzyme activity is about 5.5, 6, 6.5, 7, 7.5, 8, 8.5, 9.0, 9.5, 10, 10.5, 11, 11.5, 12, 12.5, 13, 13.5, 14, 14.5, 15, 15.5, 16, 16.5, 17, 17.5, 18, 18.5, 19, 19.5, 20, 20.5, 21, 21.5, 22, 22.5, 23, 23.5, 24, 24.5, 25, 25.5, 26, 26.5, 27, 27.5, 28, 28.5, 29, 29.5, 30, 30.5, 35, 40, 45, 50, 55, 55, 60, 65, 70, 75, 80, 85, 90, 95, or 100 $\pm$ 2.0 IU/mg. In one particular embodiment, the specific enzyme activity is 9 $\pm$ 2.0 IU/mg.

25 Compositions comprising an ADIr (e.g., ADIr-PEG) described herein can be administered simultaneously with, prior to, or after administration of one or more other therapeutic agents, including ADI-PEG 20. Such combination therapy may include administration of a single pharmaceutical dosage formulation which contains a compound of the invention and one or more additional active agents, as well as administration of
30 compositions comprising an ADIr described herein (e.g., ADIr-PEG) and each active agent in its own separate pharmaceutical dosage formulation. For example, the ADIr (e.g., ADIr-PEG) and the other active agent can be administered to the patient together in a single oral dosage composition such as a tablet or capsule, or each agent administered in separate oral dosage formulations. Similarly, the ADIr (e.g., ADIr-PEG) and the other active agent can be
35 administered to the patient together in a single parenteral dosage composition such as in a saline solution or other physiologically acceptable solution, or each agent administered in separate parenteral dosage formulations, by the same or different routes (e.g., one by

injection, one by oral). Where separate dosage formulations are used, the compositions comprising the ADIr (e.g., ADIr-PEG) and one or more additional active agents can be administered at essentially the same time, *i.e.*, concurrently, or at separately staggered times, *i.e.*, sequentially and in any order; combination therapy is understood to include all these regimens.

Thus, in certain embodiments, also contemplated is the administration of the ADIr compositions of this disclosure in combination with one or more other therapeutic agents. Such therapeutic agents may be accepted in the art as a standard treatment for a particular disease state as described herein, such as a particular cancer or GVHD. Exemplary therapeutic agents contemplated include cytokines, growth factors, steroids, NSAIDs, DMARDs, anti-inflammatories, chemotherapeutics, radiotherapeutics, autophagy inhibitors, or other active and ancillary agents.

In certain embodiments, the ADIr (e.g., ADIr-PEG) compositions disclosed herein may be administered in conjunction with any number of chemotherapeutic agents. Examples of chemotherapeutic agents include alkylating agents such as thiotepa and cyclophosphamide (CYTOXANTM); alkyl sulfonates such as busulfan, improsulfan and piposulfan; aziridines such as benzodopa, carboquone, meturedopa, and uredopa; ethylenimines and methylamelamines including altretamine, triethylenemelamine, triethylenephosphoramidate, triethylenethiophosphoramidate and trimethylolomelamine; nitrogen mustards such as chlorambucil, chlornaphazine, cholophosphamide, estramustine, ifosfamide, mechlorethamine, mechlorethamine oxide hydrochloride, melphalan, novembichin, phenesterine, prednimustine, trofosfamide, uracil mustard; nitrosureas such as carmustine, chlorozotocin, fotemustine, lomustine, nimustine, ranimustine; antibiotics such as aclacinomysins, actinomycin, authramycin, azaserine, bleomycins, cactinomycin, calicheamicin, carabycin, carminomycin, carzinophilin, chromomycins, dactinomycin, daunorubicin, detorubicin, 6-diazo-5-oxo-L-norleucine, doxorubicin, epirubicin, esorubicin, idarubicin, marcellomycin, mitomycins, mycophenolic acid, nogalamycin, olivomycins, peplomycin, potfiromycin, puromycin, quelamycin, rodorubicin, streptonigrin, streptozocin, tubercidin, ubenimex, zinostatin, zorubicin; anti-metabolites such as methotrexate and 5-fluorouracil (5-FU); folic acid analogues such as denopterin, methotrexate, pteropterin, trimetrexate; purine analogs such as fludarabine, 6-mercaptopurine, thiamiprine, thioguanine; pyrimidine analogs such as ancitabine, azacitidine, 6-azauridine, carmofur, cytarabine, dideoxyuridine, doxifluridine, enocitabine, floxuridine, 5-FU; androgens such as calusterone, dromostanolone propionate, epitioestanol, mepitioestane, testolactone; anti-adrenals such as aminoglutethimide, mitotane, trilostane; folic acid replenisher such as frolinic acid; aceglatone; aldophosphamide glycoside; aminolevulinic acid; amsacrine; bestrabucil; bisantrene; edatraxate; defofamine; demecolcine; diaziquone; elformithine;

elliptinium acetate; etoglucid; gallium nitrate; hydroxyurea; lentinan; lonidamine; mitoguazone; mitoxantrone; mopidamol; nitracrine; pentostatin; phenamet; pirarubicin; podophyllinic acid; 2-ethylhydrazide; procarbazine; PSK; razoxane; sizofiran; spirogermanium; tenuazonic acid; triaziquone; 2, 2',2"-trichlorotriethylamine; urethan;

5 vindesine; dacarbazine; mannomustine; mitobronitol; mitolactol; pipobroman; gacytosine; arabinoside ("Ara-C"); cyclophosphamide; thiotepa; taxoids, e.g. paclitaxel (TAXOL®, Bristol-Myers Squibb Oncology, Princeton, N.J.) and docetaxel (TAXOTERE®, Rhne-Poulenc Rorer, Antony, France); chlorambucil; gemcitabine; 6-thioguanine; mercaptopurine; methotrexate; platinum analogs such as cisplatin and carboplatin; vinblastine; platinum;

10 etoposide (VP-16); ifosfamide; mitomycin C; mitoxantrone; vincristine; vinorelbine; navelbine; novantrone; teniposide; daunomycin; aminopterin; xeloda; ibandronate; CPT-11; topoisomerase inhibitor RFS 2000; difluoromethylomithine (DMFO); retinoic acid derivatives such as Targretin™ (bexarotene), Panretin™ (alitretinoin) ; ONTAK™ (denileukin diftitox) ; esperamicins; capecitabine; and pharmaceutically acceptable salts, acids or derivatives of

15 any of the above. Also included in this definition are anti-hormonal agents that act to regulate or inhibit hormone action on tumors such as anti-estrogens including for example tamoxifen, raloxifene, aromatase inhibiting 4(5)-imidazoles, 4-hydroxytamoxifen, trioxifene, keoxifene, LY117018, onapristone, and toremifene (Fareston); and anti-androgens such as flutamide, nilutamide, bicalutamide, leuprolide, and goserelin. Further chemotherapeutic agents include

20 sorafenib and other protein kinase inhibitors such as afatinib, axitinib, bevacizumab, cetuximab, crizotinib, dasatinib, erlotinib, fostamatinib, gefitinib, imatinib, lapatinib, lenvatinib, mubritinib, nilotinib, panitumumab, pazopanib, pegaptanib, ranibizumab, ruxolitinib, trastuzumab, vandetanib, vemurafenib, and sunitinib; sirolimus (rapamycin), everolimus and other mTOR inhibitors. Pharmaceutically acceptable salts, acids or derivatives of any of the

25 above are also contemplated for use herein.

In certain embodiments, the ADI_r (e.g., ADI_r-PEG) compositions disclosed herein may be administered in conjunction with any number of autophagy inhibitors. In some preferred embodiments, the autophagy inhibitor is selected from the group consisting of: chloroquine, 3-methyladenine, hydroxychloroquine (Plaquenil.TM.), bafilomycin A1, 5-amino-

30 4-imidazole carboxamide riboside (AICAR), okadaic acid, autophagy-suppressive algal toxins which inhibit protein phosphatases of type 2A or type 1, analogues of cAMP, and drugs which elevate cAMP levels, adenosine, N6-mercaptopurine riboside, wortmannin, and vinblastine. In addition, antisense or siRNA that inhibits expression of proteins essential for autophagy, such as for example ATG5, may also be used.

35 In some embodiments, the combination of ADI_r (e.g., ADI_r-PEG) with one or more therapeutic agents acts complementary, additively, or synergistically. In this regard, complementary or synergizing agents are described herein, which include a therapeutic

agent (e.g., chemotherapeutic agent, autophagy inhibitor, mTOR inhibitor, or any other therapeutic agent used for the treatment of cancer, GVHD, or inflammatory bowel disease as described herein) that is capable of acting complementary or synergistically with ADIr-PEG as provided herein, where such complementarity or synergy manifests as a detectable effect that is greater (i.e., in a statistically significant manner relative to an appropriate control condition) in magnitude than the effect that can be detected when the chemotherapeutic agent is present but the ADIr (e.g., ADIr-PEG) composition is absent, and/or when the ADIr (e.g., ADIr-PEG) is present but the chemotherapeutic agent is absent. Methods for measuring synergy and complementarity are known in the art (see e.g., Cancer Res January 15, 2010 70; 440).

The compositions comprising ADIr (e.g., ADIr-PEG) and optionally other therapeutic agents, as described herein, may be used in therapeutic methods for treating cancer and methods for preventing metastasis of a cancer. Thus, some embodiments include methods for treating, ameliorating the symptoms of, or inhibiting the progression of or prevention of a variety of different cancers. Some embodiments include methods for treating, ameliorating the symptoms of, or inhibiting the progression of GVHD. Particular embodiments include methods for treating, ameliorating the symptoms of, or inhibiting the progression of a cancer or GVHD in a patient comprising administering to the patient a therapeutically effective amount of ADIr composition as described herein, optionally, following treatment with ADIr-PEG 20, particularly where a patient develops anti-ADIr-PEG 20 antibodies, thereby treating, ameliorating the symptoms of, or inhibiting the progression of the cancer or GVHD. Thus, the ADIr compositions described herein may be administered to an individual afflicted with inflammatory bowel disease (e.g., Crohn's disease; ulcerative colitis), GVHD or a cancer, including, but not limited to hepatocellular carcinoma, leukemia (e.g. acute myeloid leukemia and relapsed acute myeloid leukemia), melanoma including metastatic melanoma, sarcomas (including, but not limited to, metastatic sarcomas, uterine leiomyosarcoma), pancreatic cancer, prostate cancer (such as, but not limited to, hormone refractory prostate cancer), mesothelioma, lymphatic leukemia, chronic myelogenous leukemia, lymphoma, small cell lung cancer, breast cancer, ovarian cancer, colorectal cancer, gastric cancer (including, but not limited to, gastric adenocarcinoma), glioma, glioblastoma multi-form, retinoblastoma, neuroblastoma, non-small cell lung cancer (NSCLC), kidney cancer (including but not limited to renal cell carcinoma), bladder cancer, uterine cancer, esophageal cancer, brain cancer, head and neck cancers (including, but not limited to, squamous cell carcinoma of the head and neck; cancer of the tongue), cervical cancer, testicular cancer, gallbladder, cholangiocarcinoma, and stomach cancer.

Also included are methods of treating, ameliorating the symptoms of, or inhibiting the progression of a cancer in a patient comprising administering to the patient a composition

comprising ADIr (e.g., ADIr-PEG), and optionally one or more other therapeutic agents, as described herein, wherein the cancer is deficient in ASS, ASL, or both. In this regard, ASS or ASL deficiency may be a reduction in expression as measured by mRNA expression or protein expression, or may be a reduction in protein activity, and generally comprises a statistically significant reduction in expression or activity as determined by the skilled person. Reduced ASS or ASL expression or activity may be a reduction in expression or activity of about 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, or more, as compared to expression or activity in an appropriate control sample known to be cancer free. In certain embodiments, ASS or ASL expression or activity is reduced by at least twofold as compared to expression or activity in a non-cancer control sample.

In certain embodiments, the reduced expression or activity of ASS or ASL results from methylation of the ASS or ASL promoter or inhibition of the ASS or ASL promoter. In some embodiments the reduction in expression or activity of ASS or ASL results from a DNA mutation (e.g., one or more point mutations, small deletions, insertions, and the like) or a chromosomal abnormality resulting in deletion of the gene. In some embodiments, the cancer is ASS or ASL negative, meaning no expression or activity is observed.

Reduction in ASS or ASL expression or activity may be measured using any methods known in the art, such as but not limited to, quantitative PCR, immunohistochemistry, enzyme activity assays (e.g., assay to measure conversion of citrulline into argininosuccinate or conversion of argininosuccinate into arginine and fumarate), and the like.

Thus, certain embodiments include methods for treating, ameliorating the symptoms of, or inhibiting the progression of a cancer in a patient comprising administering to the patient a composition comprising an ADIr as described herein (e.g., ADIr-PEG), wherein the cancer exhibits reduced expression or activity of ASS or ASL, or both, wherein the cancer includes, but is not limited to hepatocellular carcinoma, leukemia (e.g. acute myeloid leukemia and relapsed acute myeloid leukemia), melanoma including metastatic melanoma, sarcomas (including, but not limited to, metastatic sarcomas, uterine leiomyosarcoma), pancreatic cancer, prostate cancer (such as, but not limited to, hormone refractory prostate cancer), mesothelioma, lymphatic leukemia, chronic myelogenous leukemia, lymphoma, small cell lung cancer, breast cancer, ovarian cancer, colorectal cancer, gastric cancer (including, but not limited to, gastric adenocarcinoma), glioma, glioblastoma multi-form, retinoblastoma, neuroblastoma, non-small cell lung cancer (NSCLC), kidney cancer (including but not limited to renal cell carcinoma), bladder cancer, uterine cancer, esophageal cancer, brain cancer, head and neck cancers (including, but not limited to, squamous cell carcinoma of the head and neck; cancer of the tongue), cervical cancer, testicular cancer, gallbladder, cholangiocarcinoma, and stomach cancer.

Various studies in the literature have shown that ASS is deficient in the following tumors: acute myelogenous leukemia (AML), bladder, breast, colorectal, gastric, glioblastoma, HCC, lymphoma, melanoma, mesothelioma, non-small cell lung, ovarian, pancreatic, prostate, renal, sarcoma, and small cell lung. Accordingly, treatment of these
5 ASS-deficient cancers is specifically contemplated herein, with ADIr-PEG alone or in combination with other treatments, including treatment first with ADI-PEG 20.

Also included are methods for treating, ameliorating the symptoms of, or inhibiting the progression of cancer in a patient comprising administering to the patient a composition comprising ADIr as described herein (e.g., ADIr-PEG, ADIr-PEG 20), in combination with an
10 autophagy inhibitor. Some embodiments include methods for treating cancer in a patient comprising administering to the patient a therapeutically effective amount of a composition comprising ADIr as described herein in combination with autophagy inhibitor wherein the cancer is pancreatic cancer or small cell lung cancer.

Some embodiments include methods of treatment where administration of the
15 compositions comprising ADIr described herein depletes arginine in the plasma for at least one month, 2 months, 3 months, 4 months, 5 months, 6 months or longer. Certain embodiments include methods of treatment where administration of the compositions comprising ADIr described herein depletes arginine in the plasma for at least one month, 2 months, 3 months, 4 months, 5 months, 6 months or longer after terminating treatment with
20 ADI-PEG 20 following detection of anti-ADI-PEG 20 antibodies.

EXAMPLES

EXAMPLE 1

SCREENING AND SELECTION OF ADI ENZYMES THAT HAVE LOW CROSS-REACTIVITY WITH PATIENT ANTI-ADI-PEG 20 ANTIBODIES

This example describes the screening and selection of ADI enzymes that have low cross-reactivity with patient anti-ADI-PEG 20 antibodies.

From a large number of ADI enzymes, Table 1 lists 27 ADI enzymes selected for their sequence percent identity relative to *M. hominis* ADI.

Table 1: Selected ADI Sequences with Varying Degrees of Similarity to <i>M. hominis</i> ADI			
ORGANISM	PERCENT IDENTITY	SEQUENCE ACCESSION NUMBER	SEQ ID NO:
<i>Mycoplasma hominis</i>	100.0	gi 728876	1
<i>Mycoplasma salivarium</i>	81.95	gi 639205998	2
<i>Mycoplasma spumans</i>	81.51	gi 652844751	3
<i>Mycoplasma canadense</i>	81.22	gi 780004832	20
<i>Mycoplasma auris</i>	78.54	gi 490558584	4
<i>Mycoplasma hyosynoviae</i>	78.78	gi 738491959	5
<i>Mycoplasma cloacale</i>	78.97	gi 657216914	6
<i>Mycoplasma anseris</i>	78.73	gi 697347301	21
<i>Mycoplasma alkalescens</i>	77.67	gi 488970417	7
<i>Mycoplasma orale</i>	79.27	gi 551316378	8
<i>Mycoplasma iners</i>	60.20	gi 657715567	9
<i>Mycoplasma meleagridis</i>	59.1	gi 803632330	22
<i>Mycoplasma alvi</i>	58.46	gi 697091625	23
<i>Mycoplasma penetrans</i>	58.1	gi 768722109	24
<i>Mycoplasma gallinarum</i>	58.06	gi 653082428	10
<i>Mycoplasma pirum</i>	56.05	gi 652846075	11
<i>Mycoplasma primatum</i>	56.86	gi 657716867	12
<i>Mycoplasma fermentans</i>	56.11	gi 753838319	25
<i>Mycoplasma lipofaciens</i>	55.64	gi 652840093	13
<i>Mycoplasma felifaucium</i>	55.85	gi 653084388	14
<i>Mycoplasma imitans</i>	53.85	gi 652843200	15
<i>Mycoplasma opalescens</i>	54.25	gi 659858924	16
<i>Mycoplasma moatsii</i>	50.24	gi 551314758	17
<i>Mycoplasma elephantis</i>	52.63	gi 653084064	18
<i>Mycoplasma pneumoniae</i>	51.97	gi 12643471	26
<i>Mycoplasma testudinis</i>	49.50	gi 652838532	19
<i>Mycoplasma sp. CAG:877</i>	39.28	gi 524101565	27
<i>Mycoplasma sp. CAG:472</i>	38.07	gi 524735679	28

A number of ADI enzymes from a variety of organisms were characterized to determine which enzymes would be expected to remove and maintain low concentrations of arginine in patient blood, even in the presence of anti-ADI-PEG 20 antibodies. As detailed below, these studies show that ADI from a number of species that are closely related to *M. hominis*, based on sequence identity, have sufficiently good enzyme catalytic properties,

measured at various temperatures and pH, and also show reduced cross-reactivity with anti-ADI-PEG 20 antibodies.

ADI Preparation. Recombinant ADI enzymes were cloned, expressed, and purified for testing according to standard protocols, as described, for example, in Gallego et al., PLOS One, 7(10):e47886, 2012; Monstadt and Holldorf, Biochem. J. 273:739-745, 1990; Joo Noh et al., Molecules and Cells. 13:137-143, 2002; and Sugimura et al., Infection and Immunity. 58:2510-2515, 1990.

Human Anti-ADI-PEG20 Antibody Purification. Anti-ADI-PEG20 antibody was purified from plasma samples of patients who had received ADI-PEG20 during a clinical study. For example, a total of 60 ml of plasma was pooled from 8 different patients that had reached high titer (titer ≥ 4) against ADI-PEG20 as determined by an ELISA assay. A two-step purification was used, a Protein "A" chromatography (GE Healthcare) followed by an ADI affinity chromatography. About 20 mg of purified antibody was obtained and stored at -80°C in aliquots until needed.

ADI Enzyme Assays. Arginine deiminase (ADI) catalyzes the conversion of L-arginine to L-citrulline and ammonia. The amount of L-citrulline can be detected by a colorimetric endpoint assay (see, for example, Knipp and Vasak, Analytical Biochem. 286:257-264, 2000) and compared to a standard curve of known amounts of L-citrulline in order to calculate the specific activity of ADI expressed as IU/mg of protein. One IU of enzyme activity was defined as the amount of enzyme that produces 1 μmol of citrulline per minute at the pH and temperature being tested. Standard assay conditions were performed at 37°C in Physiological HEPES Buffer (PHB) 50 mM HEPES, 160 mM NaCl pH 7.4 (Lang and Zander, Clin Chem Lab Med. 37:563-571, 1999) plus 0.1% BSA. All samples and standards were run in duplicate or triplicate where conditions permitted.

K_m and K_{cat} values were determined by using a variation of the activity assay described above. As with the activity assay (or unless otherwise indicated), all reactions were run at 37°C in PHB plus 0.1% BSA. Enzyme concentration, reaction time, and substrate concentration range were adjusted for each of the ADI or ADI_r constructs to account for their differences in activity. In general, 2 nM enzyme, 5 minute reaction time, and a 0 – 160 μM arginine were used as starting conditions. When optimizing the conditions, particular attention was paid towards the amount of substrate consumed as a percentage of total substrate added to the reaction. Typically, the lower limit of detection was about 1 μM of citrulline with the lower limit of quantitation being about 2 μM . A citrulline standard curve was run on every plate and used to quantify the citrulline produced by the enzymatic reaction.

Activity assays were also performed to assess enzymatic activity in the presence of anti-ADI-PEG20 (antibody neutralization profiles). These assays were performed as

described above and in the presence of 640 nM, 320 nM, 160 nM, 80 nM, 40 nM, 20 nM, 10 nM, and 0 nM of anti-ADI-PEG20 antibodies.

5 Calculations. The citrulline concentration (μM) produced in each reaction well was calculated and averaged using the citrulline standard curve. The velocity of each reaction was then calculated in $\mu\text{M}/\text{min}/50 \text{ nM ADI}$. Specific activity (IU/mg or $\mu\text{moles product}/\text{min}/\text{mg ADI}$) was calculated by multiplying this value by the “IU” factor (IU factor was calculated from the molecular weight of the ADI and the reaction volume).

The data are shown in Tables 2-7 below.

Table 2. Selected Native ADI Sequences and Properties			
Organism	Specific Activity (IU/mg)	Kcat (s-1)	Km (μM)
<i>Mycoplasma hominis</i>	+	+	+
<i>Mycoplasma salivarium</i>	++	++	+
<i>Mycoplasma spumans</i>	+++	+++	+
<i>Mycoplasma canadense</i>	+	+	+
<i>Mycoplasma cloacale</i>	++	+	+
<i>Mycoplasma hyosynoviae</i>	-	-	+
<i>Mycoplasma anseris</i>	+	-	+
<i>Mycoplasma auris</i>	+	-	+
<i>Mycoplasma alkalescens</i>	++	++	-
<i>Mycoplasma iners</i>	++++	++++	+
<i>Mycoplasma meleagridis</i>	+++	++	+
<i>Mycoplasma penetrans</i>	++	++	-
<i>Mycoplasma gallinarum</i>	++++	+++	+
<i>Mycoplasma primatum</i>	+++	+++	+
<i>Mycoplasma fermentans</i>	-	-	ND
<i>Mycoplasma lipofaciens</i>	-	-	ND
<i>Mycoplasma opalescens</i>	++	++	-
<i>Mycoplasma imitans</i>	-	ND	ND
<i>Mycoplasma sp. CAG:877</i>	-	ND	ND
<i>Mycoplasma sp. CAG:472</i>	-	ND	ND

Table 3. Selected PEG-ADI Sequences and Properties			
Organism	Specific Activity (IU/mg)	Kcat (s-1)	Km (μM)
<i>Mycoplasma hominis</i>	+	+	+
<i>Mycoplasma salivarium</i>	++	++	+
<i>Mycoplasma spumans</i>	+++	+++	+
<i>Mycoplasma canadense</i>	++	++	-

<i>Mycoplasma cloacale</i>	++	++	+
<i>Mycoplasma hyosynoviae</i>	-	+	ND
<i>Mycoplasma anseris</i>	++	++	-
<i>Mycoplasma auris</i>	++	++	-
<i>Mycoplasma alkalescens</i>	-	+	ND
<i>Mycoplasma iners</i>	++++	++++	+
<i>Mycoplasma meleagridis</i>	+++	++	+
<i>Mycoplasma penetrans</i>	+	+	-
<i>Mycoplasma gallinarum</i>	++++	++++	+
<i>Mycoplasma primum</i>	+++	+++	+
<i>Mycoplasma opalescens</i>	-	ND	ND

Table 4. Selected Native ADI Sequences and Properties

Organism	Specific Activity at Concentration of α -ADI Ab Indicated (nM)*							
	640	320	160	80	40	20	10	0
<i>Mycoplasma hominis</i>	ND	33	39	59	76	86	94	100
<i>Mycoplasma salivarium</i>	25	49	69	84	92	96	96	100
<i>Mycoplasma spumans</i>	42	59	76	87	95	96	98	100
<i>Mycoplasma canadense</i>	27	56	89	104	121	122	113	100
<i>Mycoplasma cloacale</i>	24	50	72	85	95	98	98	100
<i>Mycoplasma hyosynoviae</i>	22	47	72	83	101	100	91	100
<i>Mycoplasma anseris</i>	39	60	90	99	109	106	106	100
<i>Mycoplasma auris</i>	45	64	81	90	95	99	100	100
<i>Mycoplasma alkalescens</i>	27	48	71	83	92	94	95	100
<i>Mycoplasma iners</i>	63	80	89	93	96	98	99	100
<i>Mycoplasma meleagridis</i>	64	90	104	101	110	101	112	100
<i>Mycoplasma penetrans</i>	67	86	91	90	89	106	100	100
<i>Mycoplasma gallinarum</i>	67	82	89	94	96	96	97	100
<i>Mycoplasma primum</i>	64	79	89	94	98	99	99	100
<i>Mycoplasma opalescens</i>	66	91	95	102	106	106	104	100
*Indicated as percentage (%) specific activity (IU/mg) relative to activity with no antibody (0 nM)								

Table 5. Selected PEG-ADI Sequences and Properties

Organism	Specific Activity at Concentration of α -ADI Ab Indicated (nM)*							
	640	320	160	80	40	20	10	0

<i>Mycoplasma hominis</i>	ND	53	64	77	89	93	94	100
<i>Mycoplasma salivarium</i>	43	59	77	85	91	94	95	100
<i>Mycoplasma spumans</i>	58	72	83	91	97	97	99	100
<i>Mycoplasma canadense</i>	42	62	79	94	91	92	95	100
<i>Mycoplasma cloacale</i>	31	52	72	82	93	95	95	100
<i>Mycoplasma anseris</i>	41	59	71	91	93	93	99	100
<i>Mycoplasma auris</i>	56	70	82	88	93	95	96	100
<i>Mycoplasma alkalescens</i>	42	59	78	87	92	97	98	100
<i>Mycoplasma iners</i>	65	84	92	94	98	99	94	100
<i>Mycoplasma meleagridis</i>	56	79	93	101	106	105	107	100
<i>Mycoplasma penetrans</i>	69	85	93	101	104	104	105	100
<i>Mycoplasma gallinarum</i>	62	77	87	91	94	99	100	100
<i>Mycoplasma primatum</i>	65	80	86	91	93	96	98	100
*Indicated as percentage (%) specific activity (IU/mg) relative to activity with no antibody (0 nM)								

Table 6. Selected Native-ADI Sequences and Properties								
Organism	Specific Activity* at Temperature Indicated							
	20°C	24°C	28°C	32°C	37°C	40°C	44°C	48°C
<i>Mycoplasma hominis</i>	37	50	66	85	100	105	103	80
<i>Mycoplasma salivarium</i>	20	32	46	69	100	120	147	168
<i>Mycoplasma spumans</i>	25	38	53	75	100	118	135	135
<i>Mycoplasma canadense</i>	21	32	43	74	100	127	160	185
<i>Mycoplasma cloacale</i>	17	28	44	65	100	124	147	180
<i>Mycoplasma hyosynoviae</i>	16	26	41	64	100	104	126	124
<i>Mycoplasma anseris</i>	17	23	44	60	100	128	159	179
<i>Mycoplasma auris</i>	18	29	44	67	100	122	157	174
<i>Mycoplasma alkalescens</i>	20	31	45	67	100	119	146	160
<i>Mycoplasma iners</i>	36	52	63	78	100	125	142	191
<i>Mycoplasma meleagridis</i>	40	49	60	75	100	95	58	20
<i>Mycoplasma penetrans</i>	53	52	67	74	100	108	118	114
<i>Mycoplasma gallinarum</i>	41	51	67	81	100	119	145	171
<i>Mycoplasma primatum</i>	31	46	60	79	100	116	124	128
<i>Mycoplasma opalescens</i>	29	35	47	66	100	106	135	132
* Indicated as percentage (%) specific activity (IU/mg) relative to activity at 37°C								

Table 7. Selected Native-ADI Sequences and Properties												
Organism	Specific Activity* at pH Indicated											
	2.4	3.2	4	4.8	5.6	6.4	7	7.6	8.4	9.2	10	10.4
<i>Mycoplasma hominis</i>	5	6	17	66	94	71	88	58	17	26	18	5
<i>Mycoplasma salivarium</i>	1	1	1	55	124	132	114	82	46	59	40	10

<i>Mycoplasma spumans</i>	1	1	1	58	117	119	108	84	44	62	45	10
<i>Mycoplasma canadense</i>	0	1	1	67	148	149	144	96	47	67	49	20
<i>Mycoplasma cloacale</i>	0	0	0	43	126	130	113	86	46	64	46	13
<i>Mycoplasma hyosynoviae</i>	0	1	1	22	124	132	117	85	41	39	26	6
<i>Mycoplasma anseris</i>	0	0	1	51	159	171	135	108	55	82	59	22
<i>Mycoplasma auris</i>	0	1	1	1	87	92	98	87	50	69	48	4
<i>Mycoplasma alkalescens</i>	0	0	0	38	113	123	106	78	38	53	35	7
<i>Mycoplasma iners</i>	0	1	15	32	76	83	84	78	45	66	55	26
<i>Mycoplasma meleagridis</i>	0	3	3	6	84	90	103	98	46	55	40	5
<i>Mycoplasma penetrans</i>	0	0	7	41	123	130	129	100	42	64	51	14
<i>Mycoplasma gallinarum</i>	0	1	18	36	91	90	104	92	50	69	56	26
<i>Mycoplasma primatum</i>	0	1	0	27	81	94	99	92	50	76	60	17
<i>Mycoplasma opalescens</i>	0	1	1	2	83	97	115	93	51	84	65	23
* Indicated as percentage (%) specific activity (IU/mg) relative to activity at pH 7.4												

These data show, inter alia, that native and PEGylated ADI enzymes that are highly homologous to *M. hominis* ADI (about 50-100 percent identity) maintained excellent catalytic activity (Tables 2-4), including at various pH (Table 6) and temperature-defined (Table 7) conditions. They also showed reduced affinity toward patient anti-ADI-PEG 20 antibodies, as measured by (increased) enzyme activity in the presence of anti-ADI-PEG 20 antibodies (Tables 4-5), for example, relative to that of *Mycoplasma hominis*. Accordingly, these ADI enzymes may have therapeutic utility for use in therapy for the treatment of cancer, either alone or following ADI-PEG 20 treatment, to extend and/or increase the effectiveness of arginine depletion therapy.

The various embodiments described above can be combined to provide further embodiments. All of the U.S. patents, U.S. patent application publications, U.S. patent application, foreign patents, foreign patent application and non-patent publications referred to in this specification and/or listed in the Application Data Sheet are incorporated herein by reference, in their entirety. Aspects of the embodiments can be modified, if necessary to employ concepts of the various patents, application and publications to provide yet further embodiments.

These and other changes can be made to the embodiments in light of the above-detailed description. In general, in the following claims, the terms used should not be construed to limit the claims to the specific embodiments disclosed in the specification and

the claims, but should be construed to include all possible embodiments along with the full scope of equivalents to which such claims are entitled. Accordingly, the claims are not limited by the disclosure.

Claims

1. A therapeutic composition comprising an isolated arginine deiminase, or a fragment thereof having arginine deiminase (ADI) activity, and a pharmaceutically-acceptable carrier, wherein the isolated arginine deiminase has reduced cross-reactivity with patient anti-ADI-PEG 20 antibodies and comprises the amino acid sequence set forth in SEQ ID NO: 9 or SEQ ID NO: 22, or a variant thereof that displays at least 90% sequence identity to SEQ ID NO: 9 or SEQ ID NO: 22.
2. The therapeutic composition of claim 1 wherein the isolated arginine deiminase has one or more properties comparable to or better than those of ADI-PEG 20.
3. The therapeutic composition of claim 2 wherein the one or more properties is K_{cat}, K_m, pH optimum, stability, *in vivo* proteolytic stability, or no requirement for ions or cofactors that are not already present in blood, or any combination thereof.
4. The therapeutic composition of any one of claims 1-3 wherein the isolated arginine deiminase has been modified to remove at least one pegylation site.
5. The therapeutic composition of any one of claims 1-4 wherein at least one lysine residue has been modified by an amino acid substitution.
6. The therapeutic composition of any one of claims 1 to 5 wherein at least 5 lysine residues have been modified by an amino acid substitution.
7. The therapeutic composition of any one of claims 1 to 6 wherein at least 10 lysine residues have been modified by an amino acid substitution.
8. The therapeutic composition of any one of claims 1 to 7 wherein at least 15 lysine residues have been modified by an amino acid substitution.
9. The therapeutic composition of any one of claims 1 to 8 wherein at least 20 lysine residues have been modified by an amino acid substitution.
10. The therapeutic composition of any one of claims 1-9 wherein the arginine deiminase is covalently bonded via a linker to a PEG molecule.
11. The therapeutic composition of claim 10 wherein the arginine deiminase is covalently bonded to more than one PEG molecule.
12. The therapeutic composition of claim 10 or 11 wherein the arginine deiminase is covalently bonded to 1 to 10 PEG molecules, or 2 to 8 PEG molecules.
13. The therapeutic composition of any one of claims 10 to 12 wherein the PEG molecules are straight chain or branch chain PEG molecules.

14. The therapeutic composition of any one of claims 10 to 13 wherein the PEG has a total weight average molecular weight of from 1,000 to 40,000, or from 10,000 to 30,000.

15. The therapeutic composition of any one of claims 10 to 14 wherein the linker is a succinyl group, an amide group, an imide group, a carbamate group, an ester group, an epoxy group, a carboxyl group, a hydroxyl group, a carbohydrate, a tyrosine group, a cysteine group, a histidine group, a methylene group, or any combinations thereof.

16. The therapeutic composition of claim 15 wherein the source of the succinyl group is succinimidyl succinate.

17. The therapeutic composition of one any of claims 1 to 16 further comprising a chemotherapeutic agent.

18. The therapeutic composition of claim 17 wherein the chemotherapeutic agent is selected from the group consisting of docetaxel, carboplatin, cyclophosphamide, gemcitabine, cisplatin, sorafenib, sunitinib and everolimus.

19. Use of a therapeutic composition of any one of claims 1 to 18 in the manufacture of a medicament for treating, ameliorating the symptoms of, or inhibiting the progression of a cancer in a patient.

20. The use of claim 19 wherein the patient in need thereof has been determined to have anti-ADI-PEG 20 antibodies.

21. The use of claim 19 or 20 wherein the cancer is selected from the group consisting of hepatocellular carcinoma, melanoma, metastatic melanoma, pancreatic cancer, prostate cancer, small cell lung cancer, mesothelioma, lymphocytic leukemia, chronic myelogenous leukemia, lymphoma, hepatoma, sarcoma, leukemia, acute myeloid leukemia, relapsed acute myeloid leukemia, breast cancer, ovarian cancer, colorectal cancer, gastric cancer, glioma, glioblastoma multiforme, non-small cell lung cancer (NSCLC), kidney cancer, bladder cancer, uterine cancer, esophageal cancer, brain cancer, head and neck cancers, cervical cancer, testicular cancer, and stomach cancer.

22. Use of a ADI-PEG 20 and a therapeutic composition of any one of claims 1 to 18 in the manufacture of a medicament for treating, ameliorating the symptoms of, or inhibiting the progression of a cancer in a patient, wherein said ADI-PEG 20 is to be administered to the patient, and after a period of time, said therapeutic composition is to be administered to the patient.

23. The use of claim 22 wherein the period of time is determined by detecting a predetermined level of anti-ADI-PEG 20 antibodies in the patient, wherein the therapeutic composition is to be administered following detection of the predetermined level of said anti-ADI-PEG 20 antibodies.

24. The use of claim 22 wherein the period of time is determined by detecting ADI activity in the patient, wherein the therapeutic composition is to be administered following detection of a predetermined or reduced level of ADI activity.

25. The therapeutic composition of any one of claims 1 to 18 for use in treating, ameliorating the symptoms of, or inhibiting the progression of a cancer.

ARGININE DEIMINASE WITH REDUCED CROSS-REACTIVITY TOWARD ADI – PEG 20
ANTIBODIES FOR CANCER TREATMENT

ABSTRACT

The present invention relates generally to isolated to arginine deiminase (ADI) proteins that have reduced cross-reactivity with anti-ADI-PEG 20 antibodies as compared to ADI-PEG 20, but which can have functional characteristics comparable to or better than ADI-PEG 20, compositions comprising the ADI proteins, and related methods of treating arginine-dependent diseases or related diseases such as cancer.

[Fig. 1C]