

KITS AND DEVICES

This invention relates to kits and devices suitable for use in the testing for the presence of a large number of diagnostic markers, including those used in the identification of cancer, infectious
5 disease and transplant organ rejection. They are also useful for companion diagnostic testing in which a panel of markers must be identified reliably and at low cost.

The polymerase chain reaction (PCR) is a well-known and powerful technique for amplifying DNA or RNA present in laboratory and diagnostic samples to a point where they can be reliably
10 detected and/or quantified. However, when applied for the purposes of investigating analyte samples containing low-levels of such molecules, it suffers from a number of limitations. First, whilst the technique can detect as little as a single target molecule, it is prone to generating false positive results due to unwanted amplification of other nucleic acid sequences present in the sample. This makes the choice of oligonucleotide primers used to initiate the reaction key; which
15 in turn makes designing primers with the required level of specificity relatively complex. As a consequence, many PCR-based tests available on the market today have limited specificity.

A second drawback is that multiplexing of PCR-based methods is in practice limited to at most tens of target sequences (frequently no more than 10) with the avoidance of primer-primer interactions resulting in the need for relatively narrow operational windows.

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Another issue is that, because the PCR reaction cycles in an exponential fashion, quantification of the target is difficult; small variations in the efficiency of the reaction having a huge impact on the amount of detectable material generated. Even with appropriate controls and calibrations in place, quantification is thus typically limited to an accuracy within a factor of
25 around 3.

Finally, mutations in the region targeted for investigation by PCR amplification methods can have unwanted side effects. For example, there have been instances where FDA-approved tests have had to be withdrawn because the target organism underwent mutation in the genetic region
30 targeted by the test primers resulting in large numbers of false negatives. Conversely, if a specific single nucleotide polymorphism (SNP) is targeted for amplification the PCR method will often give a false positive when the wild-type variant is present. Avoiding this requires very careful primer design and further limits the efficacy of multiplexing. This is particularly relevant when searching

for panels of SNPs as is a common requirement in cancer testing/screening or companion diagnostics.

US2006/110765 A1 (Wang et al) teaches enzymatic cleavage at a mismatch, which is typically an inefficient and not a highly specific reaction. Moreover, off target hybridisation of the probes as disclosed in Wang et al to similar sequences in a sample would result in false positive results because of the use of cleavage at mismatches. It would also not be possible to distinguish between two different genetic variants in the same or near-neighbouring positions as they would all result in cleavage and amplification of the same probe. The teachings of Wang et al would therefore result in low sensitivity and specificity of the reaction scheme. In contrast, the technical effect of the method as disclosed by the present invention provides a fast, efficient method with high specificity to dsDNA, which can be effectively blocked by a mismatch. In addition, the method of the present invention is extremely specific to the targeted genetic variant, enabling discrimination between different variants in the same or near-neighbouring positions.

US2009/239283 A1 (Liu et al) teaches the use of non-extendable 3' ends which are removed by pyrophosphorolysis, necessitating the genetic engineering of custom polymerases capable of removing the 3' blocking modification. In contrast the present invention utilises the natural pyrophosphorolysis activity inherent in existing polymerases and does not use 3' blocking modifications. The method as disclosed in Liu et al also relies on the removal of only the terminal base from a fraction of the probe to enable subsequent amplification, and is limited to this embodiment by the use of 3' blocking modification. In contrast, the methods disclosed in the present invention enables embodiments, in which progressive removal of multiple bases from the probe is required to set off the reaction, making it substantially more robust to transient off-target annealing either to the background DNA or to other probes, which can result in the unwanted removal of the terminal base.

Summary of Invention

We have now developed kits and devices for use with a new method which builds on our experience using the pyrophosphorolysis method employed in our earlier patent (see PCT/GB2019/052017) to overcome many of these limitations. In doing so, it harnesses the double-strand specificity of pyrophosphorolysis; a reaction which will not proceed efficiently with single-stranded oligonucleotide substrates or double-stranded substrates which include blocking

groups or nucleotide mismatches. Thus, according to the present invention, there is provided a kit comprising:

- 5 (a) a single-stranded probe oligonucleotide A_0 , capable of forming a first intermediate product with a target polynucleotide sequence, said intermediate product being at least partially double-stranded;
- (b) a ligase;
- (c) a pyrophosphorolysing enzyme capable of digesting the first intermediate product in the 3'-5' direction from the end of A_0 to create a partially digested strand A_1 ;
- 10 (d) at least one single-stranded primer oligonucleotide that is substantially complementary to a portion of A_0 ;
- (e) an amplification enzyme;
- (f) suitable buffers.

And a device comprising:

- 15 at least a fluid pathway between a first region, a second region and a third region, wherein the first region comprises one or more wells, wherein each well comprises:
 - dNTPs;
 - at least one single-stranded primer oligonucleotide;
 - an amplification enzyme for the initial amplification of DNA present in a sample;
- 20 and
- wherein the second region comprises one or more wells, wherein each well comprises:
 - a single-stranded probe oligonucleotide A_0 , capable of forming a first intermediate product with a target polynucleotide sequence, said intermediate product being at least partially double-stranded;
 - 25 a pyrophosphorolysing enzyme capable of digesting the first intermediate product in the 3'-5' direction from the end of A_0 to create a partially digested strand A_1 ;
 - at least one source of pyrophosphate ion;
 - a ligase;
 - and
- 30
- wherein the third region comprises one or more wells, wherein each well comprises:
 - dNTPs;
 - buffers;

an amplification enzyme;
a means for detecting a signal derived from A_1 or a portion thereof, or multiple copies of A_1 or multiple copies of a portion thereof; and

5 wherein the wells of the second region or the wells of the third region further comprise at least one single-stranded primer oligonucleotide that is substantially complementary to a portion of A_0 .

The analytes to which the method of the invention can be applied are those nucleic acids,
10 such as naturally-occurring or synthetic DNA or RNA molecules, which include the target polynucleotide sequence(s) being sought. In one embodiment, the analyte will typically be present in an aqueous solution containing it and other biological material and in one embodiment the analyte will be present along with other background nucleic acid molecules which are not of interest for the purposes of the test. In some embodiments, the analyte will be present in low
15 amounts relative to these other nucleic acid components. Preferably, for example where the analyte is derived from a biological specimen containing cellular material, prior to performing step (b) of the method some or all of these other nucleic acids and extraneous biological material will have been removed using sample-preparation techniques such as filtration, centrifuging, chromatography or electrophoresis. Suitably, the analyte is derived from a biological sample
20 taken from a mammalian subject (especially a human patient) such as blood, plasma, sputum, urine, skin or a biopsy. In one embodiment, the biological sample will be subjected to lysis in order that the analyte is released by disrupting any cells present. In other embodiments, the analyte may already be present in free form within the sample itself; for example cell-free DNA circulating in blood or plasma.

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Brief Description of Drawings

Fig. 1: Protocols for simplified polynucleotide sequence detection methods.

Fig. 2: A graph comparing the level of fluorescence detected (representing the presence of a particular target analyte sequence) when the 5'-3' exonuclease digestion step happens during the pre-amplification step and when it is moved to the pyrophosphorolysis/ligation step of the protocol (as in protocols 3-5). In this example, the 5'-3' exonuclease is Lambda.

Fig. 3: The inventors have tested the method of Protocol 3 of the current invention using a range of different PPL enzymes. Fig. 3 (A) shows detection of 1% MAF T790M using Mako, Klenow and Bsu. Fig. 3(B) shows the detection of 0.5% MAF T790M using Bst LF at a range of different Ppi concentrations. All four enzymes performed very well even without extended optimisation.

Fig. 4: Results for the detection of 1% MAF, T790M, using the methods of Protocol 4 of the current invention using four different pyrophosphorolysing (PPL) enzymes: Mako, Klenow, Bsu and Bst LF.

Fig. 5: Graphs showing the level of fluorescence detected (representing the presence of particular target analyte sequence) of 0.5%, 0.10% and 0.05% MAF Exon19 del_6223 detected according to Protocol 1 and Protocol 4.

Fig. 6: The inventors have detected EGFR exon 20 T790M at 0.10%, 0.50% and 1% MAF according to Protocol 4.

Fig. 7: shows detection of EGFR exon 20 T790M at 1% MAF with and without the presence of an exonuclease in the RCA step.

Fig. 8: The inventors have investigated what effect the PPL:RCA mix ratio has on the intensity of signal detected for 0.5% MAF EGFR exon 20 T790M, the results of which are shown in Fig. 8. As can be seen a ratio of 1:2 PPL:RCA mix results in the lowest signal intensity but at the earliest time point. This is followed closely in time by 1:4 PPL:RCA mix which has a greater signal intensity. The largest signal intensity is seen for 1:8 PPL:RCA mix at the latest time point in the reaction.

Fig. 9: shows the results of comparison experiments performed according to Protocol 4 using SybrGreenI (50°C and 60°C) and Syto82 (50°C and 60°C).

Fig. 10: The inventors have investigated the use of two different enzymes, BST L.F and BST 2.0 WS,
5 for RCA according to Protocol 4.

Fig. 11: The inventors have investigated the effect of different PPL enzymes on the RCA reaction at different PPL:RCA reaction mixture ratios. The results of which can be seen in Fig. 11(A) 1:4 PPL:RCA and Fig. 11(B) 1:8 PPL:RCA. All PPL enzymes impact the RCA reaction at 1:4 PPL:RCA ratio
10 other than BST L.F. At 1:8 PPL:RCA ratio, all enzymes apart from BST L.F and Klenow impact the RCA reaction.

Fig. 12: Fluorescence measurement results for Example 11 showing that when oligonucleotide 3 and 4 are both present, the fluorescent signal appears faster in the reaction, showing that
15 pyrophosphorolysis and ligation of oligonucleotide 3 has occurred in the first reaction mixture.

Fig. 13: Detection of T790M and C797S_2389 mutations at 1% allele fraction in the same reaction.

Fig. 14: Detection of three mutations at the same time in one well at 0.5% allele fraction: G719X_6239, G719X_6252, G719X_6253.

Fig.15: A schematic representation of the circularisation of A_1 to form A_2 against the analyte target sequence. A_0 is progressively digested against the target in the 3'-5' direction from the 3' end of A_0 to form partially digested strand A_1 , this is shown as steps (A) and (B). This progressive digestion reveals the region of the target that is complementary to the 5' end of A_0/A_1 and the 5' end of A_1 then hybridises to this region, this is shown in step (C). A_1 is then ligated together to
20 form circularised A_2 , step (D).
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Fig. 16: Fluorescence measurement results for Example 14 showing results from an embodiment wherein pyrophosphorolysis of A_0 to form A_1 occurs followed by circularisation of A_1 to form A_2 against a target sequence.
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Fig. 17: A single-stranded probe oligonucleotide A_0 anneals to a target polynucleotide sequence to create a first intermediate product which is at least partially double-stranded and in which the 3' end of A_0 forms a double-stranded complex with the target polynucleotide sequence. In this simplified embodiment of the invention there are two molecules of A_0 present and one target polynucleotide sequence, in order to illustrate how A_0 that has not annealed to a target does not take part in further steps of the method. In this illustrative example, the 3' end of A_0 anneals to
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the target polynucleotide sequence whilst the 5' end of A_0 does not. The 5' end of A_0 comprises a 5' chemical blocking group, a common priming sequence and a barcode region.

5 The partially double-stranded first intermediate product undergoes pyrophosphorolysis in the presence of a pyrophosphorolysing enzyme in the 3'-5' direction from the 3' end of A_0 to create a partially digested strand A_1 , the analyte and the undigested A_0 molecule which did not anneal to a target.

10 Fig. 18: A_1 is annealed to a single-stranded trigger oligonucleotide B and the A_1 strand is extended in the 5'-3' direction against B to create an oligonucleotide A_2 . In this illustrative example, trigger oligonucleotide B has a 5' chemical block. Any undigested A_0 anneals to the trigger oligonucleotide B, however it is unable to be extended in the 5'-3' direction against B to generate sequences that are the targets for later parts of the method. In this example, A_2 is primed with at least one single-stranded primer oligonucleotide and multiple copies of A_2 , or a region of A_2 are created.

15 Fig. 19: A_1 is annealed to a splint oligonucleotide D, and then circularised by ligation of its 3' and 5' ends. The now circularised A_2 is primed with at least one single-stranded primer oligonucleotide and multiple copies of A_2 , or a region of A_2 are created. In this illustrative example, the splint oligonucleotide D is unable to extend against A_1 by virtue of either a 3'-modification (chemical in this illustration) or through a nucleotide mismatch between the 3' end of D and the corresponding region of A_2 .

20 Fig. 20: The 3' region of a splint oligonucleotide D anneals to the 3' region of A_1 whilst the 5' region of the splint oligonucleotide D anneals to the 5' region of a ligation probe C. Thus, a second intermediate product A_2 is formed comprised of A_1 , C and optionally an intermediate region formed by extension of A_1 in the 5'-3' direction to meet the 5' end of C. In this illustrative example, the ligation probe C has a 3' chemical blocking group so that a 3'-5' exonuclease can be used to digest any non-ligated A_1 .

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30 A_2 is primed with at least one single-stranded primer oligonucleotide and multiple copies of A_2 , or a region of A_2 are created.

Description of Embodiments.

In an aspect of the present invention, there is provided a method of detecting a target polynucleotide sequence in a given nucleic acid analyte present in a sample, the method comprising the steps of:

- 5 (a) introducing one or more nucleic acid analytes to a first reaction mixture comprising:
- i. a single-stranded probe oligonucleotide A_0 ;
 - ii. a pyrophosphorolysing enzyme; and
 - iii. a ligase
- 10 wherein A_0 is pyrophosphorolysed in the 3'-5' direction from the 3' end to create at least a partially digested strand A_1 and A_1 undergoes ligation to form A_2 ;
- (b) detecting a signal derived from the products of the previous step, wherein the products are A_2 or a portion thereof, or multiple copies of A_2 or multiple copies of a portion thereof, and inferring therefrom the presence or absence of the
- 15 polynucleotide target sequence in the analyte.

In some embodiments, the first reaction mixture further comprises a source of pyrophosphate ions.

- 20 In some embodiments, the first reaction mixture further comprises at least one single-stranded primer oligonucleotide that is substantially complementary to a portion of A_0 and deoxyribonucleotide triphosphates (dNTPs).

In some embodiments, the dNTPs are optional.

In some embodiments, the first reaction mixture further comprises an amplification enzyme.

- 25 In some embodiments, the products of step (a) are introduced to a second reaction mixture prior to step (b), said second reaction mixture comprising at least one single-stranded primer oligonucleotide and dNTPs.

In some embodiments, the second reaction mixture further comprises an amplification enzyme.

In some embodiments, one or more nucleic acid analytes may be introduced to a first and second reaction mixture concurrently.

- 30 In some embodiments, one or more nucleic acid analytes may be introduced to a first and second reaction mixture consecutively.

In some embodiments, the dNTPs are hot start dNTPs.

Hot start dNTPs are dNTPs which are modified with a thermolabile protecting group at the 3' terminus. The presence of this modification blocks DNA polymerase nucleotide incorporation until the nucleotide protecting group is removed using a heat activation step.

5 In some embodiments, during step (a) the analyte anneals to the single-stranded probe oligonucleotide A_0 to create a first intermediate product which is at least partially double-stranded and in which the 3' end of A_0 forms a double-stranded complex with the analyte target sequence.

10 In some embodiments, during step (a) the first intermediate product is pyrophosphorolysed in the 3'-5' direction from the 3' end of A_0 to create partially digested strand A_1 and the analyte. In some embodiments, the first reaction mixture further comprises a ligation probe oligonucleotide C and the partially digested strand A_1 is ligated at the 3' end to the 5' end of C to create an oligonucleotide A_2 .

15 In some embodiments, the partially digested strand A_1 is circularised through ligation of its 3' and 5' ends.

In some embodiments, the ligation of A_1 occurs:

20 during step (a); or
during step (b); or
inbetween steps (a) and (b).

In one embodiment, A_1 is circularised against the analyte target sequence. In this embodiment, the region of the target that is revealed by progressive digestion of A_0 , in the 3'-5' direction from the 3' end of A_0 to form A_1 , is complementary to the 5' end of A_0/A_1 . In this embodiment, a ligase may be used to ligate the 3' and 5' ends of A_1 to form a circularised oligonucleotide A_2 . This is shown, for example in Figure 15. In one embodiment, the 5' end of A_0/A_1 is complementary to the target across a region that is 5-50 nucleotides in length. In one embodiment, it is 5-25 nucleotides in length. In one embodiment, it is 5-20 nucleotides in length. In one embodiment, it is 5-15
30 nucleotides in length. In one embodiment, it is 5-12 nucleotides in length. In one embodiment, it is 5-10 nucleotides in length.

In some embodiments, the first reaction mixture further comprises a 5'-3' exonuclease and the 5' end of A₀ is rendered resistant to 5'-3' exonuclease digestion.

In some embodiments, after amplification of the given nucleic acid analyte and prior to addition of the first reaction mixture (step (a)), the sample is further treated with a proteinase.

- 5 In some embodiments, the first reaction mixture further comprises a phosphatase or phosphohydrolase.

In some embodiments, prior to or during step (b) the products of the previous step are treated with a pyrophosphatase.

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In some embodiments, prior to or during step (b) the products of the previous step are treated with an exonuclease.

- 15 In some embodiments, the oligonucleotide C further comprises a 3' or internal modification protecting it from 3'-5' exonuclease digestion.

In some embodiments, the oligonucleotide C further comprises a 5' modification protecting it from 5'-3' exonuclease digestion.

In some embodiments, the first or second reaction mixture further comprises a splint oligonucleotide D.

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In some embodiments, D comprises an oligonucleotide region complementary to the 3' end of A₁ and a region complementary to either the 5' end of oligonucleotide C or to the 5' end of A₁.

In some embodiments, D is unable to undergo extension against A₁ by virtue of either a 3' modification or through a mismatch between the 3' end of D and the corresponding region of A₁.

- 25 In some embodiments, the enzyme which performs pyrophosphorolysis of A₀ to form partially digested strand A₁ also amplifies A₂.

In some embodiments, detection is achieved using one or more oligonucleotide fluorescent binding dyes or molecular probes.

- 30 In some embodiments, an increase in signal over time resulting from the generation of amplicons of A₂ is used to infer the concentration of the target sequence in the analyte.

In some embodiments, multiple probes A₀ are employed, each selective for a different target sequence and each including an identification region, and further characterised in that the

amplicons derived from A_2 include this identification region and therefore the target sequences present in the analyte are inferred through the detection of the identification region(s).

In some embodiments, detection of the identification regions(s) is carried out using molecular probes or through sequencing.

In some embodiments, the final step of the method further comprises the steps of:

- i. labelling the products of step (b) using one or more oligonucleotide fluorescent binding dyes or molecular probes;
- ii. measuring the fluorescent signal of the products;
- 10 iii. exposing the products to a set of denaturing conditions; and

identifying the polynucleotide target sequence in the analyte by monitoring changes in the fluorescent signal of the products during exposure to the denaturing conditions.

In some embodiments, the one or more nucleic acid analytes are split into multiple reaction volumes, each volume having a one or more probe oligonucleotide A_0 , introduced to detect different target sequences.

In some embodiments, the different probes A_0 comprise common priming sites, allowing a single primer or single set of primers to be used for amplification of a region of A_2 .

In some embodiments, there is provided a method of detecting a target polynucleotide sequence in a given nucleic acid analyte present in a sample, the method comprising the steps of:

- (a) amplifying a given nucleic acid analyte present in the sample;
- (b) introducing the products of step (a) to a first reaction mixture comprising:
 - i. a single-stranded probe oligonucleotide A_0 ;
 - ii. a pyrophosphorolysing enzyme; and
 - 25 iii. a ligase

wherein A_0 is pyrophosphorolysed in the 3'-5' direction from the 3' end to create at least a partially digested strand A_1 and A_1 undergoes ligation to form A_2 ;

- (c) detecting a signal derived from the products of the previous step, wherein the products are A_2 or a portion thereof, or multiple copies of A_2 or multiple copies of a portion thereof, and inferring therefrom the presence or absence of the polynucleotide target sequence in the analyte.

According to the present invention, there are provided methods of detecting a target polynucleotide sequence in a given nucleic acid analyte. The analytes to which the various methods of the invention can be applied may be prepared from the biological sample mentioned above by a series of preliminary steps designed to amplify the analyte and separate it from the background genomic DNA which is typically present in significant excess. This method is generally applicable to the production of single-stranded target analytes and is therefore useful in situations other than when it is integrated with or further comprises part of the method of the first aspect of the invention. Accordingly, there is provided a method for preparing at least one single-stranded analyte of a nucleic acid comprised of a target polynucleotide region characterised by the steps of (1) producing amplicons of the analyte(s) by subjecting a biological sample comprised of the analyte(s) and optionally background genomic DNA to cycles of amplification.

In some preferred embodiments amplification is carried out using the polymerase chain reaction (PCR) in the presence of a polymerase, nucleoside triphosphates and at least one corresponding primer pair wherein one of the primers includes a 5'-3' exonuclease blocking group and (2) optionally digesting the product of step (1) with an exonuclease having 5'-3' exonucleolytic activity. In one embodiment, the method may further comprise (3) reacting the product of step (2) with a proteinase to destroy the polymerase and thereafter (4) deactivating the proteinase by heating the product of step (3) to a temperature in excess of 50°C.

In some preferred embodiments steps (1) to (4) are carried out prior to step (a) of the method of the first aspect of the invention to produce an integrated method of detecting target sequences derived from a biological sample. In another embodiment, the biological sample has undergone cell lysis before step (1) is carried out.

In some embodiments of step (1) the nucleoside triphosphates are a mixture of the four deoxynucleoside triphosphates characteristic of naturally occurring DNA. In a preferred embodiment the mixture of deoxynucleoside triphosphates comprise deoxyuridine triphosphate (dUTP) instead of deoxythymidine triphosphate (dTTP) and step (1) is further carried out in the presence of the enzyme dUTP-DNA glycolase (UDG) to remove any contaminating amplicons from previous assays. In yet another embodiment, a high fidelity polymerase is used in step (1) for

example one of those sold under the trade name Phusion® or Q5. In yet another embodiment, the polymerase may be KAPA HiFi Uracil+ DNA Polymerase.

High-fidelity DNA polymerases have several safeguards to protect against both making and propagating mistakes while copying DNA. Such enzymes have a significant binding preference for the correct versus the incorrect nucleoside triphosphate during polymerization. If an incorrect nucleotide does bind in the polymerase active site, incorporation is slowed due to the sub-optimal architecture of the active site complex. This lag time increases the opportunity for the incorrect nucleotide to dissociate before polymerase progression, thereby allowing the process to start again, with a correct nucleoside triphosphate. If an incorrect nucleotide is inserted, proofreading DNA polymerases have an extra line of defense. The perturbation caused by the mispaired bases is detected, and the polymerase moves the 3' end of the growing DNA chain into a proofreading 3'→5' exonuclease domain. There, the incorrect nucleotide is removed by the 3'→5' exonuclease activity, whereupon the chain is moved back into the polymerase domain, where polymerization can continue.

In some embodiments, the nucleoside triphosphates are a mixture of synthetic or modified deoxynucleoside triphosphates.

In some embodiments, the nucleoside triphosphates are a mixture of the four deoxynucleoside triphosphates and synthetic or modified deoxynucleotide triphosphates.

In some embodiments, step (1) is carried out using a limited amount of primer and an excess of amplification cycles. By this means a fixed amount of amplicons is produced regardless of the initial amount of analyte. Thus the need for analyte quantification prior to subsequent steps is avoided. In another embodiment of step (1), which has the advantage of obviating the need for step (2), amplification is carried out in the presence of a primer pair where one of the two primers is present in excess of the other, resulting in generation of single-stranded amplicons once one primer is fully utilised.

In some preferred embodiments of step (2), the 5' primer is blocked with an exonuclease blocking group selected from phosphorothioate linkages, inverted bases, DNA spacers and other oligonucleotide modifications commonly known in the art. In another embodiment the other primer in the pair has a phosphate group at its 5' end.

In some embodiments, in step (3) the proteinase employed is proteinase K and step (4) is carried out by heating to a temperature of 80 to 100°C for up to 30 minutes. In one embodiment, in step (3) the proteinase employed is proteinase K, step (3) is carried out by heating to a temperature of 55°C for 5 minutes and step (4) is carried out by heating to a temperature of 95°C for 10 minutes.

- 5 In another embodiment at some point after step (2) the reaction medium is treated with a phosphatase or phosphohydrolase to remove any residual nucleoside triphosphates which may be present.

- 10 In some embodiments, the target polynucleotide sequence in the analyte will be a gene or chromosomal region within the DNA or RNA of a cancerous tumour cell and will be characterised by the presence of one or more mutations; for example in the form of one or more single nucleotide polymorphisms (SNPs). Thus the invention will be useful in the monitoring of and/or treatment for disease recurrence. Patients who have been declared free of disease following treatment may be monitored over time to detect the recurrence of disease. This needs to be done
- 15 non-invasively and requires sensitive detection of target sequences from blood samples. Similarly, for some cancers there are residual cancer cells that remain in a patient after treatment. Monitoring of the levels of these cells (or cell free DNA) present in the patient's blood, using the current invention, allows detection of recurrence of disease or failure of current therapy and the need to switch to an alternative.

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- In some embodiments, detection of the target polynucleotide sequence will allow repeated testing of patient samples during treatment of disease to allow early detection of developed resistance to therapy. For example, epidermal growth factor receptor (EGFR) inhibitors, such as gefitinib, erlotinib, are commonly used as first line treatments for non-small cell lung cancer
- 25 (NSCLC). During treatment the tumour will often develop mutations in the EGFR gene (e.g T790M, C797S) which confer resistance to the treatment. Early detection of these mutations allows transfer of the patient onto alternative therapies.

- 30 In some embodiments, the target polynucleotide sequence in the analyte will be a gene or chromosomal region within the DNA or RNA of fetal origin and will be characterised by the presence of one or more mutations; for example in the form of one or more single nucleotide polymorphisms (SNPs). Thus, the invention may be used to detect mutations at very low allele fractions, at an earlier stage of pregnancy than other available testing techniques.

In another embodiment, the target polynucleotide sequence may be a gene or genomic region derived from an otherwise healthy individual but the genetic information obtained may assist in generating valuable companion diagnostic information allowing medical or therapeutic conclusions to be drawn across one or more defined groups within the human population.

5 In yet another embodiment, the target polynucleotide sequence may be characteristic of an infectious disease, or of resistance of an infectious disease to treatment with certain therapies; for example a polynucleotide sequence characteristic of a gene or chromosomal region of a bacterium or a virus, or a mutation therein conferring resistance to therapy.

10 In some embodiments, the target polynucleotide sequence may be characteristic of donor DNA. When a transplanted organ is rejected by the patient, the DNA from this organ is shed into the patient's bloodstream. Early detection of this DNA would allow early detection of rejection. This could be achieved using custom panels of donor-specific markers, or by using panels of variants known to be common in the population, some of which will be present in the donor and some in
15 the recipient. Routine monitoring of organ recipients over time is thus enabled by the claimed method.

In yet another embodiment, various versions of the method using different combinations of probes (see below) are employed in parallel so that the analyte can be simultaneously screened
20 for multiple target sequences; for example sources of cancer, cancer indicators or multiple sources of infection. In this approach, the amplified products obtained in by parallel application of the method are contacted with a detection panel comprised of one or more oligonucleotide binding dyes or sequence specific molecular probes such as a molecular beacon, hairpin probe or the like. Thus, in another aspect of the invention there is provided the use of at least one probe
25 and optionally one ligation oligonucleotide in combination with one or more chemical and biological probes selective for the target polynucleotide sequences or with the use of sequencing to identify the amplified probe regions.

In some embodiments, the single-stranded probe oligonucleotide A_0 comprises a priming region
30 and a 3' end which is complementary to the target polynucleotide sequence to be detected. By this means, a first intermediate product is created which is at least partially double-stranded. In one embodiment, this step is carried out in the presence of excess A_0 and in an aqueous medium containing the analyte and any other nucleic acid molecules.

During step (a), the double-stranded region of the first intermediate product is pyrophosphorolysed in the 3'-5' direction from the 3' end of its A_0 strand. As a consequence, the A_0 strand is progressively digested to create a partially digested strand; hereinafter referred to as A_1 . Where the probe oligonucleotide erroneously hybridises with a non-target sequence, the pyrophosphorolysis reaction will stop at any mismatches, preventing subsequent steps of the method from proceeding. In another embodiment, this digestion continues until A_1 lacks sufficient complementarity with the analyte or a target region therein to form a stable duplex. At this point, the various strands then separate by melting, thereby producing A_1 in single-stranded form.

Under typical pyrophosphorolysis conditions, this separation occurs when there are between 6 and 20 complementary nucleotides between the analyte and A_0 .

In another embodiment, the digestion continues until A_1 lacks sufficient complementarity with the analyte or target region therein for the pyrophosphorolysing enzyme to bind or for the pyrophosphorolysing reaction to continue. This typically occurs when there are between 6 and 20 complementary nucleotides remaining between the analyte and probe. In some embodiments, this occurs when there are between 6 and 40 complementary nucleotides remaining.

In another embodiment, in which a splint oligonucleotide D having complementarity to the 5' and 3' ends of A_1 is employed (see below), the digestion continues until the length of complementarity between A_1 and the target is reduced to the point at which it is energetically favourable for oligo D to displace the analyte molecule from A_1 . This typically occurs when the region of complementarity between A_1 and the analyte molecule is of similar or shorter length than the region of complementarity between oligo D and the 3' end of A_1 , but may also occur when the complementarity between A_1 and the analyte molecule is longer than this due to the favourability of intra-molecular hybridisation of the oligo D, which may already be hybridised to the 5' end of A_1 .

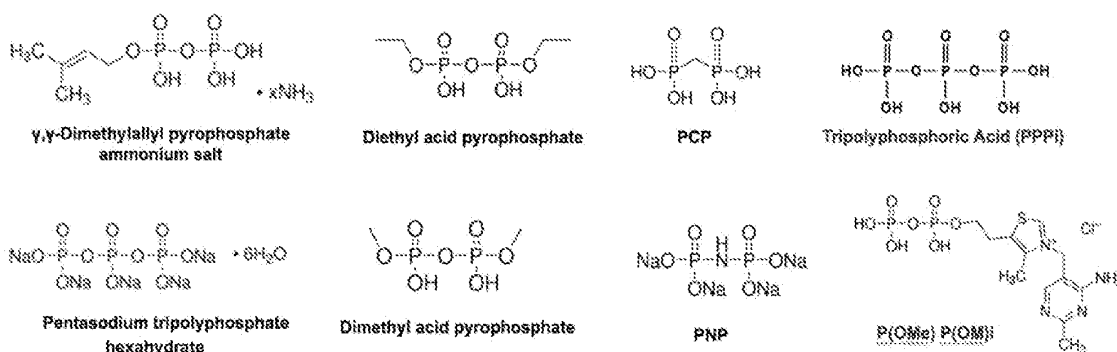
In another embodiment, in which the ligation of A_1 is performed using the analyte molecule as a splint (see Figure 16), the digestion continues until the 5' end of A_1 is able to hybridise to the analyte molecule such that the 3' and 5' ends of A_1 are neighbouring and are separated only by a nick, at which point they are ligated together by the ligase and digestion is no longer able to proceed.

Suitably, pyrophosphorolysis is carried out in the reaction medium at a temperature in the range 20 to 90°C in the presence of at least a polymerase exhibiting pyrophosphorolysis activity and a source of pyrophosphate ion. Further information about the pyrophosphorolysis reaction as applied to the digestion of polynucleotides can be found for example in J. Biol. Chem. 244 (1969) pp. 3019-3028 or our earlier patent application.

In some embodiments, the pyrophosphorolysis step is driven by the presence of a source of excess polypyrophosphate, suitable sources including those compounds containing 3 or more phosphorous atoms.

In some embodiments, the first reaction mixture comprises a source of excess polypyrophosphate.

In some embodiments, the pyrophosphorolysis step is driven by the presence of a source of excess modified pyrophosphate. Suitable modified pyrophosphates include those with other atoms or groups substituted in place of the bridging oxygen, or pyrophosphate (or polypyrophosphate) with substitutions or modifying groups on the other oxygens. The person skilled in the art will understand that there are many such examples of modified pyrophosphate which would be suitable for use in the current invention, a non-limiting selection of which are:



In some embodiments, the first reaction mixture comprises a source of excess modified polypyrophosphate.

In some preferred embodiments, the source of pyrophosphate ion is PNP, PCP or Triphosphoric Acid (PPPi).

Further, but not limiting, examples of sources of pyrophosphate ion for use in the pyrophosphorolysis step (b) may be found in WO2014/165210 and WO00/49180.

- 5 In some embodiments, the source of excess modified pyrophosphate can be represented as Y-H wherein Y corresponds to the general formula $(X-O)_2P(=B)-(Z-P(=B)(O-X))_n$ - wherein n is an integer from 1 to 4; each Z- is selected independently from -O-, -NH- or -CH₂-; each B is independently either O or S; the X groups are independently selected from -H, -Na, -K, alkyl, alkenyl, or a heterocyclic group with the proviso that when both Z and B correspond to -O- and when n is 1 at least one X group is not H.

- In some embodiments, Y corresponds to the general formula $(X-O)_2P(=B)-(Z-P(=B)(O-X))_n$ - wherein n is 1, 2, 3 or 4. In another embodiment, the Y group corresponds to the general formula $(X-O)_2P(=O)-Z-P(=O)(O-H)$ - wherein one of the X groups is -H. In yet another preferred embodiment, Y corresponds to the general formula $(X-O)_2P(=O)-Z-P(=O)(O-X)-$ wherein at least one of the X groups is selected from methyl, ethyl, allyl or dimethylallyl.

- In an alternative embodiment, Y corresponds to either of the general formulae $(H-O)_2P(=O)-Z-P(=O)(O-H)$ - wherein Z is either -NH- or -CH₂- or $(X-O)_2P(=O)-Z-P(=O)(O-X)-$ wherein the X groups are all either -Na or -K and Z is either -NH- or -CH₂-.

20

In other embodiments, Y corresponds to the general formula $(H-O)_2P(=B)-O-P(=B)(O-H)$ - wherein each B group is independently either O or S, with at least one being S.

- Specific examples of preferred embodiments of Y include those of the formula $(X1-O)(HO)P(=O)-Z-P(=O)(O-X2)$ wherein Z is O, NH or CH₂ and (a) X1 is γ,γ-dimethylallyl, and X2 is -H; or (b) X1 and X2 are both methyl; or (c) X1 and X2 are both ethyl; or (d) X1 is methyl and X2 is ethyl or vice versa.

25

- In some embodiments, where molecular probes are to be used for detection, the probe oligonucleotide A₀ is configured to include an oligonucleotide identification region on the 5' side of the region complementary to the target sequence, and the molecular probes employed are designed to anneal to this identification region. In one embodiment, only the 3' region of A₀ is able to anneal to the target; i.e. any other regions lack sufficient complementarity with the analyte for a stable duplex to exist at the temperature at which the pyrophosphorolysis step is carried out. Here and throughout, by the term 'sufficient complementarity' is meant that, to the

30

extent that a given region has complementarity with a given region on the analyte, the region of complementarity is more than 10 nucleotides long.

In a further aspect of the methods of the invention there is provided alternate embodiments in which the phosphorolysis step of any previous embodiment is replaced with an exonuclease

5 digestion step using a double-strand specific exonuclease. The person skilled in the art will understand that double-strand specific exonucleases include those that read in the 3'-5' direction, such as ExoIII, and those that read in the 5'-3' direction, such as Lambda Exo, amongst many others.

10 In some embodiments of the invention wherein the exonuclease digestion step utilises a double strand-specific 5'-3' exonuclease, it is the 5' end of A_0 that is complementary to the target analyte and the common priming sequence and blocking group are located on the 3' side of the region complementary to the target. In a further embodiment, where molecular probes are to be used for detection the probe oligonucleotide A_0 is configured to include an oligonucleotide
15 identification region on the 3' side of the region complementary to the target sequence, and the molecular probes employed are designed to anneal to this identification region.

In embodiments of the invention wherein the exonuclease digestion step utilises a double strand-specific 5'-3' exonuclease, an exonuclease having 3' to 5' exonucleolytic activity can optionally be
20 added to the first reaction mixture for the purpose of digesting any other nucleic acid molecules present whilst leaving A_0 and any material comprising partially digested strand A_1 intact. Suitably, this resistance to exonucleolysis is achieved as described elsewhere in this application.

In one preferable embodiment of the invention, the 5' end of A_0 or an internal site on the 5' side
25 of the priming region is rendered resistant to exonucleolysis. By this means and after, or simultaneously with, the pyrophosphorolysis step, an exonuclease having 5'-3' exonucleolytic activity can optionally be added to the reaction medium for the purpose of digesting any other nucleic acid molecules present whilst leaving A_0 and any material comprising the partially digested strand A_1 intact. Suitably, this resistance to exonucleolysis is achieved by introducing one or more
30 blocking groups into the oligonucleotide A_0 at the required point. In one embodiment, these blocking groups may be selected from phosphorothioate linkages and other backbone modifications commonly used in the art, C3 spacers, phosphate groups, modified bases and the like.

In some embodiments, the identification region will comprise or have embedded within a barcoding region which has a unique sequence and is adapted to be indirectly identified using a sequence-specific molecular probe applied to the amplified components A_2 or directly by the sequencing of these components. Examples of molecular probes which may be used include, but are not limited to, molecular beacons, TaqMan® probes, Scorpion® probes and the like.

In all embodiments the A_2 strand or a desired region thereof is caused to undergo amplification so that multiple, typically many millions, of copies are made. This is achieved by priming a region of A_2 and subsequently any amplicons derived therefrom with single-stranded primer oligonucleotides, provided for example in the form of a forward/reverse or sense/antisense pair, which can anneal to a complementary region thereon. The primed strand then becomes the point of origin for amplification. Amplification methods include, but are not limited to, thermal cycling and isothermal methods such as the polymerase chain reaction, recombinase polymerase amplification and rolling circle amplification; the last of these being applicable when A_2 is circularised. By any of these means, many amplicon copies of a region of A_2 and in some instances its sequence complement can be rapidly created. The exact methodologies for performing any of these amplification methods will be well-known to one of ordinary skill and the exact conditions and temperature regimes employed are readily available in the general literature to which the reader is directed. Specifically, in the case of the polymerase chain reaction (PCR), the methodology generally comprises extending the primer oligonucleotide against the A_2 strand in the 5'-3' direction using a polymerase and a source of the various single nucleoside triphosphates until a complementary strand is produced; dehybridising the double-stranded product produced to regenerate the A_2 strand and the complementary strand; re-priming the A_2 strand and any of its amplicons and thereafter repeating these extension/dehybridisation/repriming steps multiple times to build-up a concentration of A_2 amplicons to a level where they can be reliably detected.

In some embodiments, the first reaction mixture further comprises a ligation probe oligonucleotide C, and the partially digested strand A_1 is ligated at the 3' end to the 5' end of C, while in another embodiment, A_1 is circularised through ligation of its 3' and 5' ends; in each case to create an oligonucleotide A_2 .

In one embodiment, the ligation of A_1 occurs:

during step (a); or

during step (b); or
inbetween steps (a) and (b).

In one embodiment, A₁ is optionally extended in 5'-3' direction prior to ligation.

- 5 In some embodiments, this optional extension and the ligation are performed against the target oligonucleotide, while in another embodiment they are performed through addition of a further splint oligonucleotide D to which A₁ anneals prior to extension and/or ligation. In one embodiment, D comprises an oligonucleotide region complementary to the 3' end of A₁ and a region complementary to either the 5' end of oligonucleotide C or to the 5' end of A₁. In another
10 embodiment, D is unable to extend against A₁ by virtue of either a 3'-end modification or through a nucleotide mismatch between the 3' end of D and the corresponding region of A₁.

- In some embodiments, the ligation probe C has a 5' region complementary to at least part of a 5' end region of a splint oligonucleotide D or to the target oligonucleotide. By such means, a second
15 intermediate product is formed in which the A₂ strand is comprised of A₁, C and optionally an intermediate region formed by extension of A₁ in the 5'-3' direction to meet the 5' end of C. In such an embodiment, the primers employed in step (c) (see below) are chosen to amplify at least a region of A₂ including the site at which ligation of the A₁ to C has occurred. In this embodiment, we have found that it is advantageous to include a 3' blocking group on C so that a 3'-5'
20 exonuclease can be used to digest any non-ligated A₁ prior to amplification. Suitable polymerases which may be used for the extension of A₁ prior to ligation include but are not limited to Hemo KlenTaq, Mako and Stoffel Fragment.

- In some embodiments, the first reaction mixture further comprises a phosphatase or
25 phosphohydrolase to remove by hydrolysis the nucleoside triphosphates produced by the pyrophosphorolysis reaction thereby ensuring that the pyrophosphorolysis reaction can continue and does not become out-competed by the forward polymerisation reaction.

- In some embodiments, prior to or during step (b) the products of the previous step are treated
30 with a pyrophosphatase to hydrolyse the pyrophosphate ion, preventing further pyrophosphorolysis from occurring and favouring the forward polymerisation reaction.

In some embodiments, prior to or during step (b) the products of the previous step are treated with an exonuclease.

In some embodiments, the enzyme which performs pyrophosphorolysis of A_0 to form partially digested strand A_1 also amplifies A_2 . The person skilled in the art will appreciate there exists many such enzymes.

5

The amplicons are detected and the information obtained are used to infer whether the polynucleotide target sequence is present or absent in the original analyte and/or a property associated therewith. For example, by this means a target sequence characteristic of a cancerous tumour cell may be detected with reference to specific SNPs being looked for. In another
10 embodiment, a target sequence characteristic of the genome of a virus or bacterium (including new mutations thereof) may be detected. Many methods of detecting the amplicons or identification regions can be employed including for example an oligonucleotide binding dye, a sequence-specific molecular probe such as fluorescently-labelled molecular beacon or hairpin probe. Alternatively, direct sequencing of the A_2 amplicons can be performed using one of the
15 direct sequencing methods employed or reported in the art. Where oligonucleotide binding dyes, fluorescently labelled beacons or probes are employed it is convenient to detect the amplicons using an arrangement comprising a source of stimulating electromagnetic radiation (laser, LED, lamp etc.) and a photodetector arranged to detect emitted fluorescent light and to generate therefrom a signal comprising a data stream which can be analysed by a microprocessor or a
20 computer using specifically-designed algorithms.

In some embodiments, detection is achieved using one or more oligonucleotide fluorescent binding dyes or molecular probes. In such embodiments, an increase in signal over time resulting from the generation of amplicons of A_2 is used to infer the concentration of the target sequence
25 in the analyte. In one embodiment that the final step of the method further comprises the steps of:

- i. labelling the products of step (b) using one or more oligonucleotide fluorescent binding dyes or molecular probes;
 - ii. measuring the fluorescent signal of the products;
 - 30 iii. exposing the products to a set of denaturing conditions; and
- identifying the polynucleotide target sequence in the analyte by monitoring changes in the fluorescent signal of the products during exposure to the denaturing conditions.

In some embodiments, multiple probes A_0 are employed, each selective for a different target sequence and each including an identification region, and further characterised in that the amplicons of A_2 include this identification region and therefore the target sequences present in the analyte are inferred through the detection of the identification region(s). In such
5 embodiments detection of the identification regions(s) is carried out using molecular probes or through sequencing.

In some embodiments, is the one or more nucleic acid analytes are split into multiple reaction volumes, each volume having a one or more probe oligonucleotide A_0 , introduced to detect
10 different target sequences.

In some embodiments, wherein different probes A_0 are used the different probes A_0 comprise one or more common priming sites, allowing a single primer or single set of primers to be used for amplification.
15

In another aspect of the invention, there is provided a method of identifying a target polynucleotide sequence in a given nucleic acid analyte characterised by the steps of any previous embodiment of the invention wherein the multiple copies of A_2 , or a region of A_2 , are labelled using one or more oligonucleotide fluorescent binding dyes or molecular probes. The fluorescent
20 signal of these multiple copies is measured and the multiple copies are exposed to a set of denaturing conditions. The target polynucleotide sequence is the identified by monitoring a change in the fluorescent signal of the multiple copies during exposure to the denaturing conditions.

In some embodiments, the denaturing conditions may be provided by varying the temperature e.g. increasing the temperature to a point where the double strand begins to dissociate. Additionally or alternatively, the denaturing conditions may also be provided by varying the pH such that the conditions are acidic or alkaline, or by adding in additives or agents such as a strong acid or base, a concentrated inorganic salt or organic solvent e.g. alcohol.
25

In another aspect of the invention, there is provided the use of the methods described above to screen mammalian subjects, especially human patients, for the presence of infectious diseases, cancer or for the purpose of generating companion diagnostic information.
30

- In a further aspect of the invention, there is provided control probes for use in the methods as described above. Embodiments of the current invention include those wherein the presence of a specific target sequence, or sequences, is elucidated by the generation of a fluorescent signal. In such embodiments, there may inevitably be a level of signal generated from non-target DNA present in the sample. For a given sample, this background signal has a later onset than the 'true' signal, but this onset may vary between samples. Accurate detection of the presence of low concentrations of target sequence, or sequences, thus relies on knowledge of what signal is expected in its absence. For contrived samples references are available, but for true 'blind' samples from patients this is not the case. The control probes (E_0) are utilised to determine the expected background signal profile for each assay probe. The control probe targets a sequence not expected to be present in the sample and the signal generated from this probe can then be used to infer the expected rate of signal generation from the sample in the absence of target sequence.
- 15 Thus there is provided a method of detecting a target polynucleotide sequence in a given nucleic acid analyte according to any of the previously described methods characterised by the steps of:
- a. either subsequently or concurrently repeating the steps of the methods using a second single-stranded probe oligonucleotide E_0 having a 3' end region at least partially mismatched to the target sequence, either using a separate aliquot of the sample or in the same aliquot and using a second detection channel;
 - b. inferring the background signal expected to be generated from A_0 in the absence of any target analyte in the sample; and
 - c. through comparison of the expected background signal inferred in (a) with the actual signal observed in the presence of the target analyte inferring the presence or absence of the polynucleotide target sequence in the analyte.

- In some embodiments, the control probe (E_0) and A_0 are added to separate portions of the sample while in another embodiment the E_0 and A_0 are added to the same portion of the sample and different detection channels (e.g. different colour dyes) used to measure their respective signals.
- 30 The signal generated by E_0 may then be utilised to infer and correct for the background signal expected to be generated by A_0 in the absence of the polynucleotide target sequence in the sample. For example, a correction of the background signal may involve the subtraction of the signal observed from E_0 from that observed from A_0 , or through the calibration of the signal

observed from A_0 using a calibration curve of the relative signals generated by A_0 and E_0 under varying conditions.

5 In some embodiments, a single E_0 can be used to calibrate all of the assay probes which may be produced.

10 In some embodiments, a separate E_0 may be used to calibrate each amplicon of the sample DNA generated in an initial amplification step. Each amplicon may contain multiple mutations/target sequences of interest, but a single E_0 will be sufficient to calibrate all of the assay probes against a single amplicon.

15 In a further embodiment, a separate E_0 may be used for each target sequence. For example, if a C>T mutation is being targeted, an E_0 could be designed that targets a C>G mutation in the same site that is not known to occur in patients. The signal profile generated by E_0 under various conditions can be assessed in calibration reactions and these data used to infer the signal expected from the assay probe targeting the C>T variant when this variant is not present.

Some embodiments, of the method of the invention can be seen in Figures 17 to 20.

20 In Figure 17, a single-stranded probe oligonucleotide A_0 anneals to a target polynucleotide sequence to create a first intermediate product which is at least partially double-stranded and in which the 3' end of A_0 forms a double-stranded complex with the target polynucleotide sequence. In this simplified embodiment of the invention there are two molecules of A_0 present and one target polynucleotide sequence, in order to illustrate how A_0 that has not annealed to a target does not take part in further steps of the method. In this illustrative example, the 3' end of A_0 anneals to the target polynucleotide sequence whilst the 5' end of A_0 does not. The 5' end of A_0 comprises a 5' chemical blocking group, a common priming sequence and a barcode region.

25 The partially double-stranded first intermediate product undergoes pyrophosphorolysis in the presence of a pyrophosphorolysing enzyme in the 3'-5' direction from the 3' end of A_0 to create a partially digested strand A_1 , the analyte and the undigested A_0 molecule which did not anneal to a target.

In Figure 18, A_1 is annealed to a single-stranded trigger oligonucleotide B and the A_1 strand is extended in the 5'-3' direction against B to create an oligonucleotide A_2 . In this illustrative example, trigger oligonucleotide B has a 5' chemical block. Any undigested A_0 anneals to the trigger oligonucleotide B, however it is unable to be extended in the 5'-3' direction against B to generate sequences that are the targets for later parts of the method. In this example, A_2 is primed with at least one single-stranded primer oligonucleotide and multiple copies of A_2 , or a region of A_2 are created.

In Figure 19, A_1 is annealed to a splint oligonucleotide D, and then circularised by ligation of its 3' and 5' ends. The now circularised A_2 is primed with at least one single-stranded primer oligonucleotide and multiple copies of A_2 , or a region of A_2 are created. In this illustrative example, the splint oligonucleotide D is unable to extend against A_1 by virtue of either a 3'-modification (chemical in this illustration) or through a nucleotide mismatch between the 3' end of D and the corresponding region of A_2 .

In Figure 20, the 3' region of a splint oligonucleotide D anneals to the 3' region of A_1 whilst the 5' region of the splint oligonucleotide D anneals to the 5' region of a ligation probe C. Thus, a second intermediate product A_2 is formed comprised of A_1 , C and optionally an intermediate region formed by extension of A_1 in the 5'-3' direction to meet the 5' end of C. In this illustrative example, the ligation probe C has a 3' chemical blocking group so that a 3'-5' exonuclease can be used to digest any non-ligated A_1 .

A_2 is primed with at least one single-stranded primer oligonucleotide and multiple copies of A_2 , or a region of A_2 are created.

The specificity of the methods of the current invention may be improved by the introduction of blocking oligonucleotides. For example a blocking oligo nucleotide can be introduced so as to hybridise to at least a portion of wild-type DNA, promoting annealing of A_0 only to the target polynucleotide sequences and not the wildtype. Alternatively or additionally, blocking oligonucleotides can be used to improve the specificity of the polymerase chain reaction (PCR) to prevent amplification of any wild type sequence present. The general technique used is to design an oligonucleotide that anneals between the PCR primers and is not able to be displaced or digested by the PCR polymerase. The oligonucleotide is designed to anneal to the non-target (usually healthy) sequence, while being mismatched (often by a single base) to the target

(mutant) sequence. This mismatch results in a different melting temperature against the two sequences, the oligonucleotide being designed to remain annealed to the non-target sequence at the PCR extension temperature while dissociating from the target sequence.

- 5 The blocking oligonucleotides may often have modifications to prevent its digestion by the exonuclease activity of the PCR polymerase, or to enhance the melting temperature differential between the target and non-target sequence.

The incorporation of a locked nucleic acid (LNA) or other melting temperature altering modification into a blocking oligonucleotide can significantly increase the differential in melting
10 temperature of the oligonucleotide against target and non-target sequences.

Thus there is provided an embodiment of the invention wherein blocking oligonucleotides are used. The blocking oligonucleotides must be resistant to the pyrophosphorolysing (PPL) reaction to ensure they are not digested or displaced. This can be achieved in a number of different ways, for example via mismatches at their 3' ends or through modifications such as phosphorothioate
15 bonds or spacers.

In such embodiments or an aspect of the present invention where blocking oligonucleotides are used, the method of detecting a target polynucleotide sequence in a given nucleic acid analyte is characterised by annealing single-stranded blocking oligonucleotides to at least a subset of non-
20 target polynucleotide sequences before, or during, the same step wherein the analyte target sequence is annealed to a single-stranded probe oligonucleotide A_0 to create a first intermediate product which is at least partially double-stranded and in which the 3' end of A_0 forms a double-stranded complex with the analyte target sequence.

25 In some embodiments, the blocking oligonucleotides are made to be resistant to the pyrophosphorolysing reaction via mismatches at their 3' ends. In another embodiment, the blocking oligonucleotides are made to be resistant via the presence of a 3'-blocking group. In another embodiment the blocking oligonucleotides are made to be resistant via the presence of spacers or other internal modifications. In a further embodiment the blocking oligonucleotides
30 include both a melting temperature increasing modification or modified nucleotide base and are rendered resistant to pyrophosphorolysis.

References herein to 'phosphatase enzymes' refer to any enzymes, or functional fragments thereof, with the ability to remove by hydrolysis the nucleoside triphosphates produced by the methods of the current invention. This includes any enzymes, or functional fragments thereof, with the ability to cleave a phosphoric acid monoester into a phosphate ion and an alcohol.

- 5 References herein to 'pyrophosphatase enzymes' refer to any enzymes, or functional fragments thereof, with the ability to catalyse the conversion of one ion of pyrophosphate to two phosphate ions.

10 This also includes inorganic pyrophosphatases and inorganic diphosphatases. A non-limiting example is thermostable inorganic pyrophosphate (TIPP).

In some embodiments there is provided a modified version of any previously described embodiment wherein the use of a pyrophosphatase is optional.

In some embodiments of the invention there is provided a kit for use in a method of detecting a target polynucleotide sequence in a given nucleic acid analyte present in a sample, comprising:

- 15 (a) a single-stranded probe oligonucleotide A_0 , capable of forming a first intermediate product with a target polynucleotide sequence, said intermediate product being at least partially double-stranded;
- (b) a ligase;
- (c) a pyrophosphorolysing enzyme capable of digesting the first intermediate
- 20 product in the 3'-5' direction from the end of A_0 to create a partially digested strand A_1 ;
- (d) at least one single-stranded primer oligonucleotide that is substantially complementary to a portion of A_0 ;
- (e) an amplification enzyme; and
- 25 (f) suitable buffers.

In one embodiment, the 3' end of A_0 is perfectly complementary to the target polynucleotide sequence.

- 30 In one embodiment, the ligase is substantially lacking in single-strand ligation activity.

In some embodiments, the kit comprises a single-stranded probe oligonucleotide A_0 , capable of forming a first intermediate product with a target polynucleotide sequence, said intermediate product being at least partially double-stranded;

- (a) a ligase;
- 5 (b) a pyrophosphorolysing enzyme capable of digesting the first intermediate product in the 3'-5' direction from the end of A_0 to create a partially digested strand A_1 ;
- (c) suitable buffers.

In some embodiments, the kit may alternatively further comprise:

- 10 - two or more Ligation Chain Reaction (LCR) probe oligonucleotides that are complementary to adjacent sequences on A_1 wherein when the probes are successfully annealed the 5' phosphate of one LCR probe is directly adjacent to the 3'OH of the other LCR probe; and
- one or more ligases.

15

In some embodiments, in the presence of A_2 the two LCR probes will successfully anneal to A_2 and be ligated together to form one oligonucleotide molecule which subsequently acts as a new target for second-round covalent ligation, leading to geometric amplification of the target of interest, in this case A_2 . The ligated products, or amplicons, are complementary to A_2 and function as targets

20 in the next cycle of amplification. Thus, exponential amplification of the specific target DNA sequences is achieved through repeated cycles of denaturation, hybridization, and ligation in the presence of excess LCR probes. From this, the presence of A_2 and hence the target polynucleotide sequence is inferred.

- 25 In some embodiments, in the presence of A_2 the two PCR probes will successfully anneal to A_2 and be ligated together to form one oligonucleotide molecule which then acts as a new target for second-round covalent ligation, leading to geometric amplification of the target of interest, in this case A_2 , which is then detected.

30 In some embodiments, the kit may alternatively further comprise:

- A ligation probe oligonucleotide C;
- A splint oligonucleotide D;

wherein C has a 5' phosphate, the 3' end of a splint oligonucleotide D is complementary to the 5' end of C and the 5' end of D is complementary to the 3' end of A₁ such that A₁ and C are capable of being ligated together to form A₂.

- 5 In some embodiments, the kit may further comprise:
- A hairpin oligonucleotide 1 (HO1) comprising a fluorophore-quencher pair, wherein HO1 is complementary to A₂ and when annealed to A₂ the hairpin structure of HO1 opens and the fluorophore-quencher pair separate; and
 - A hairpin oligonucleotide 2 (HO2) comprising a fluorophore-quencher pair, wherein
10 HO2 is complementary to the open HO1 and when annealed to HO1 the hairpin structure of HO2 opens and the fluorophore-quencher pair separate.

In some embodiments, the kit may further comprise a plurality of HO1 and HO2.

- 15 In some embodiments, the kit may alternatively further comprise an oligonucleotide A comprising a substrate arm, a partial catalytic core and a sensor arm;
- an oligonucleotide B comprising a substrate arm, a partial catalytic core and a sensor arm; and
 - a substrate comprising a fluorophore quencher pair;
- 20 wherein the sensor arms of oligonucleotides A and B are complementary to flanking regions of A₂ such that in the presence of A₂, oligonucleotides A and B are combined to form a catalytically, multicomponent nucleic acid enzyme (MNAzyme).

- In some embodiments, the kit may alternatively further comprise a partially double-stranded
25 nucleic acid construct wherein:

- one strand comprises at least one RNA base, at least one fluorophore and wherein a region of this strand is complementary to a region of A₂ and wherein this strand may be referred to as
30 the 'substrate' strand; and
- the other strand comprises at least one quencher and wherein a region of this strand is complementary to a region of A₂ adjacent to that which the substrate strand is complementary to, such that in the presence of A₂ the partially double stranded nucleic acid
35 construct becomes substantially more double-stranded.

In other words, the partially double stranded nucleic acid construct, in the presence of A_2 , has a double-stranded portion which is greater in size.

- 5 In some embodiments, the kit may further comprise an enzyme for removal of the at least one RNA base.

In some embodiments, the enzyme is Uracil-DNA Glycosylase (UDG) and the RNA base is uracil.

- 10 In some embodiments, the kit may alternatively further comprise:
- an oligonucleotide complementary to a region of A_2 including the site of ligation, comprising one or multiple fluorophores arranged such that their fluorescence is quenched either by their proximity to each other or to one or more fluorescence quenchers;
 - 15 - a double strand specific DNA digestion enzyme;

wherein, in the presence of A_2 , the labelled oligonucleotide is digested such that the fluorophores are separated from each other or from their corresponding quenchers, and a fluorescent signal, and hence the presence of A_2 , is detectable.

- 20 In some embodiments, the double strand specific DNA digestion enzyme is an exonuclease.

In some embodiments, the double strand specific DNA digestion enzyme is a polymerase with proofreading activity.

- 25 In some embodiments, the fluorophore of the kit may be selected from dyes of the fluorescein family, the carboxyrhodamine family, the cyanine family, the rhodamine family, polyhalofluorescein-family dyes, hexachlorofluorescein-family dyes, coumarin-family dyes, oxazine-family dyes, thiazine-family dyes, squaraine-family dyes, and chelated lanthanide-family dyes.
- 30 In some embodiments, the fluorophore of the kit may be selected from any of the commercially available dyes.

In some embodiments, the quencher of the kit may be selected from those available those available under the trade designations Black Hole™, Eclipse™. Dark, Qx1J, and Iowa Black™.

In some embodiments, the quencher of the kit may be selected from any of the commercially available quenchers.

5

In some embodiments, the kit may further comprise one or more partially double stranded DNA constructs wherein each construct contains one or more fluorophores and one or more quenchers. In some embodiments, when the construct is partially double-stranded the one or more fluorophores and one or more quenchers are located in close enough proximity to each other such that sufficient quenching of the one or more fluorophores occurs.

10

In some embodiments, the construct is one strand of DNA with a self-complementary region that is looped back on itself.

In some embodiments, the construct comprises one primer of a primer pair.

In some embodiments, the kit may further comprise the other primer of a primer pair.

15

In some embodiments, a portion of the single stranded section of the construct hybridises to A_2 and is extended against it by a DNA polymerase. In some embodiments, the other primer of the primer pair then hybridises to the extended construct. This primer is then extended against the construct, displacing the self-complementary region. Thus, the one or more fluorophores and one or more dyes are separated sufficiently for a fluorescent signal to be detected, indicating the presence of A_2 .

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In such an embodiment the construct may be known as a Sunrise Primer.

In some embodiments, the construct comprises two separate DNA strands.

In some embodiments, a portion of the single stranded section of the construct hybridises to A_2 and is extended against it by a DNA polymerase. In some embodiments, the other primer of the primer pair then hybridises to the extended construct. This primer is then extended against the construct, in the direction of the double stranded section, displacing the shorter of the DNA strands and thus the one or more fluorophores and one or more dyes are separated sufficiently for a fluorescent signal to be detected, indicating the presence of A_2 .

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In such an embodiment the construct may be known as a Molecular Zipper.

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The person skilled in the art will appreciate that for both the Sunrise Primer and Molecular Zipper it is possible for the one or more fluorophores and the one or more quencher pairs to be located at various positions within each respective construct. The key feature is that each pair is located in sufficient proximity to one another that in the absence of A_2 , i.e. when no extension and strand displacement has occurred, no fluorescent signal is emitted.

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In one embodiment the kit further comprises a source of pyrophosphate ion.

Suitable source(s) of pyrophosphate ion are as described previously.

In some embodiments, the kit further comprises suitable positive and negative controls.

- 5 In some embodiments, the kit may further comprise one or more control probes (E_0) which are as previously described.

In some embodiments, the kit may further comprise one or more blocking oligonucleotides which are as previously described.

- 10 In some embodiments, the kit may further comprise one or more control probes (E_0) and one or more blocking oligonucleotides.

In some embodiments, the 5' end of A_0 may be rendered resistant to 5'-3' exonuclease digestion and the kit may further comprise a 5'-3' exonuclease.

In some embodiments a kit may further comprise a ligation probe oligonucleotide C.

In some embodiments a kit may further comprise a splint oligonucleotide D.

- 15 In some embodiments, a kit may comprise both C and D.

The ligation probe C may comprise a 3' or internal modification protecting it from 3'-5' exonuclease digestion.

D may comprise an oligonucleotide region complementary to the 3' end of A_1 and a region complementary to either the 5' end of oligonucleotide C or the 5' end of A_1 .

- 20 In some embodiments, D may be unable to undergo extension against A_1 by virtue of either a 3' modification or through a mismatch between the 3' end of D and the corresponding region of A_1 or C.

In some embodiments, the kit may further comprise dNTPs, a polymerase and suitable buffers for the initial amplification of a target polynucleotide sequence present in a sample.

- 25 In some embodiments, the kit may further comprise a dUTP incorporating high fidelity polymerase, dUTPs and uracil-DNA N-glycosylase (UDG).

In some embodiments, the kit may further comprise a phosphatase or a phosphohydrolase.

In some embodiments, the kit may further comprise a pyrophosphatase. The pyrophosphatase may be hot start.

- 30 In some embodiments, the kit may further comprise a proteinase.

In some embodiments, the kit may further comprise one or more oligonucleotide binding dyes or molecular probes.

In some embodiments, the kit may further comprise multiple A_0 , each selective for a different target sequence and each including an identification region.

5 In some embodiments, the kit may further comprise an enzyme for the formation of DNA from an RNA template.

In some embodiments, the enzyme is a reverse transcriptase.

10 In some embodiments, the one or more enzymes of the kit may be hot start.

In some embodiments, the one or more enzymes of the kit may be thermostable.

15 In some embodiments, the kit may further comprise suitable washing and buffer reagents.

In some embodiments, the amplification enzyme, of (e), and the pyrophosphorolysing enzyme are the same.

20 In some embodiments, the amplification enzyme and the pyrophosphorolysis enzyme are the same.

25 The kit may further comprise purification devices and reagents for isolating and/or purifying a portion of polynucleotides, following treatment as described herein. Suitable reagents are well known in the art and include gel filtration columns and washing buffers.

In some embodiments, there is provided a kit comprising:

- (a) a single-stranded probe oligonucleotide A_0 , capable of forming a first intermediate product with a target polynucleotide sequence, said intermediate product being at least partially double-stranded;
- 30 (b) a ligase;
- (c) a pyrophosphorolysing enzyme capable of digesting the first intermediate product in the 3'-5' direction from the end of A_0 to create a partially digested strand A_1 ;

(d) suitable buffers.

In some embodiments, the kit may further comprise a source of pyrophosphate ion.

5 In some embodiments, the kit may further comprise suitable positive and negative controls.

In some embodiments of the kit, the 5' end of A_0 is rendered resistant to 5'-3' exonuclease digestion and the kit may further comprise a 5'-3' exonuclease.

10 In some embodiments, the kit may further comprise dNTPs, a polymerase and suitable buffers for the initial amplification of a target polynucleotide sequence present in a sample.

In some embodiments, the kit may further comprise a dUTP incorporating high fidelity polymerase, dUTPs and uracil-DNA N-glycosylase (UDG).

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In some embodiments, the kit may further comprise a proteinase.

In some embodiments, the kit may further comprise a ligation probe oligonucleotide C.

20 In some embodiments, the kit may further comprise a splint oligonucleotide D.

In some embodiments, the kit may further comprise a ligation probe oligonucleotide C and a splint oligonucleotide D.

25 In some embodiments of the kit, oligonucleotide C comprises a 3' or internal modification protecting it from 3'-5' exonuclease digestion.

In some embodiments of the kit, D comprises an oligonucleotide region complementary to the 3' end of A_1 and a region complementary to either the 5' end of oligonucleotide C or to the 5' end of

30 A_1 .

In some embodiments of the kit, D is unable to undergo extension against A_1 by virtue of either a 3' modification or through a mismatch between the 3' end of D and the corresponding region of A_1 or C.

- 5 In some embodiments, the kit further comprises at least one single-stranded primer oligonucleotide that is substantially complementary to a portion of A_0 , an amplification enzyme and dNTPs.

- 10 In some embodiments, the kit further comprises one or more oligonucleotide binding dyes or molecular probes.

In some embodiments, the kit further comprises multiple A_0 , each selective for a different target sequence and each including an identification region.

- 15 In some embodiments, the kit further comprises:
- two or more Ligation Chain Reaction (LCR) probe oligonucleotides that are complementary to adjacent sequences on A_1 wherein when the probes are successfully annealed the 5' phosphate of one LCR probe is directly adjacent to the 3'OH of the other LCR probe; and
 - 20 - one or more ligases.

In some embodiments, the kit further comprises one or more polymerases.

- 25 In some embodiments of the kit, the one or more polymerases are the same as the pyrophosphorolysis enzyme.

In some embodiments, the kit further comprises:

- 30 - A ligation probe oligonucleotide C;
- A splint oligonucleotide D;

wherein E has a 5' phosphate, the 3' end of a splint oligonucleotide D is complementary to the 5' end of E and the 5' end of D is complementary to the 3' end of A_1 such that A_1 and E are capable of being ligated together to form A_2 .

In some embodiments, the kit further comprises:

- A hairpin oligonucleotide 1 (HO1) comprising a fluorophore-quencher pair, wherein HO1 is complementary to A_2 and when annealed to A_2 the hairpin structure of HO1 opens and the fluorophore-quencher pair separate; and
- A hairpin oligonucleotide 2 (HO2) comprising a fluorophore-quencher pair, wherein HO2 is complementary to the open HO1 and when annealed to HO1 the hairpin structure of HO2 opens and the fluorophore-quencher pair separate.

In some embodiments, the kit further comprises a plurality of HO1 and HO2.

In some embodiments, the kit further comprises the kit further comprises:

- an oligonucleotide A comprising a substrate arm, a partial catalytic core and a sensor arm;
- an oligonucleotide B comprising a substrate arm, a partial catalytic core and a sensor arm; and
- a substrate comprising a fluorophore quencher pair;

wherein the sensor arms of oligonucleotides A and B are complementary to flanking regions of A_2 such that in the presence of A_2 , oligonucleotides A and B are combined to form a catalytically, multicomponent nucleic acid enzyme (MNAzyme).

In some embodiments, the kit further comprises a partially double-stranded nucleic acid construct wherein:

- one strand comprises at least one RNA base, at least one fluorophore and wherein a region of this strand is complementary to a region of A_2 and wherein this strand may be referred to as the 'substrate' strand; and
- the other strand comprises at least one quencher and wherein a region of this strand is complementary to a region of A_2 adjacent to that which the substrate strand is complementary to, such that in the presence of A_2 the partially stranded nucleic acid construct becomes substantially more double-stranded.

In some embodiments, the kit further comprises an enzyme for removal of the at least one RNA base.

5 In some embodiments of the kit, the enzyme is Uracil-DNA Glycosylase (UDG) and the RNA base is uracil.

In some embodiments, the kit further comprises:

- 10 - an oligonucleotide complementary to a region of A₂ including the site of ligation, comprising one or multiple fluorophores arranged such that their fluorescence is quenched either by their proximity to each other or to one or more fluorescence quenchers;
- a double strand specific DNA digestion enzyme;

wherein, in the presence of A₂, the labelled oligonucleotide is digested such that the fluorophores are separated from each other or from their corresponding quenchers, and a fluorescent signal,
15 and hence the presence of A₂, is detectable.

In some embodiments of the kit, the double strand specific DNA digestion enzyme is an exonuclease.

20 In some embodiments of the kit, the double strand specific DNA digestion enzyme is a polymerase with proofreading activity.

In some embodiments of the kit, the fluorophore is selected from dyes of the fluorescein family, the carboxyrhodamine family, the cyanine family, the rhodamine family, polyhalofluorescein-
25 family dyes, hexachlorofluorescein-family dyes, coumarin-family dyes, oxazine-family dyes, thiazine-family dyes, squaraine-family dyes, and chelated lanthanide-family dyes.

In some embodiments of the kit, the quencher is selected from those available under the trade designations Black Hole™, Eclipse™, Dark, Qx1J, and Iowa Black™.

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In some embodiments, the kit further comprises a phosphatase or a phosphohydrolase.

In some embodiments, the kit further comprises a pyrophosphatase.

In some embodiments, the kit further comprises an enzyme for the formation of DNA from an RNA template.

5 In some embodiments of the kit, the enzyme is a reverse transcriptase.

In some embodiments of the kit, one or more enzymes are hot start.

In some embodiments of the kit, one or more enzymes are thermostable.

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In some embodiments, the kit may further comprise suitable washing and buffer reagents.

In one embodiment of the invention there is provided a device comprising:

15 at least a fluid pathway between a first region, a second region and a third region,
wherein the first region comprises one or more wells, wherein each well comprises:
dNTPs;
at least one single-stranded primer oligonucleotide;
an amplification enzyme for the initial amplification of DNA present in a sample; and
wherein the second region comprises one or more wells, wherein each well comprises:
20 a single-stranded probe oligonucleotide A_0 , capable of forming a first intermediate
product with a target polynucleotide sequence, said intermediate product being at least
partially double-stranded;
a pyrophosphorolysing enzyme capable of digesting the first intermediate product in the
3'-5' direction from the end of A_0 to create a partially digested strand A_1 ; and
25 wherein the third region comprises one or more wells, wherein each well comprises:
dNTPs;
buffers;
an amplification enzyme;
a means for detecting a signal derived from A_2 or a portion thereof, or multiple copies of
30 A_2 or multiple copies of a portion thereof; and
wherein the wells of the second region or the wells of the third region further comprise at least
one single-stranded primer oligonucleotide that is substantially complementary to a portion of A_0 .

In some embodiments, a means for detecting a signal is located within one or more wells of the third region.

In some embodiments, a means for detecting a signal is located within the third region of the device.

- 5 In some embodiments, a means for detecting a signal is located within an adjacent region of the device.

In some embodiments, the dNTPs of each well of the first region may be dUTP, dGTP, dATP and dCTP and each well may further comprise a dUTP incorporating high fidelity polymerase and
10 uracil-DNA N-glycosylase (UDG).

In some embodiments, the dNTPs of each well of the third region may be dUTP, dGTP, dATP and dCTP and each well may further comprise a dUTP incorporating high fidelity polymerase and uracil-DNA N-glycosylase (UDG).

15

In some embodiments, each well of the second region may further comprise a source of pyrophosphate ion.

In some embodiments, the 5' end of A_0 may be rendered resistant to 5'-3' exonuclease digestion
20 and the wells of the second region may further comprises a 5'-3' exonuclease.

In some embodiments, each well of the second or third regions may further comprise a ligase and a ligation probe oligonucleotide C or a splint oligonucleotide D.

- 25 The ligation probe C may comprise a 3' or internal modification protecting it from 3'-5' exonuclease digestion.

The splint oligonucleotide D may comprise an oligonucleotide region complementary to the 3' end of A_1 and a region complementary to either the 5' end of oligonucleotide C or to the 5' end of A_1 .

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D may be unable to undergo extension against A_1 by virtue of either a 3' modification or through a mismatch between the 3' end of D and the corresponding region of A_1 or C.

In some embodiments, the dNTPs may be hot start and each well of the second region may further comprise a phosphatase or a phosphohydrolase.

In some embodiments, each well of the second region may further comprise a pyrophosphatase.

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In some embodiments, the pyrophosphatase may be a hot start.

In some embodiments, each well of the third region may further comprise one or more oligonucleotide binding dyes or molecular probes.

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In some embodiments, each well of the second region may comprise at least one or more different A_0 that is selective for a target sequence including an identification region.

In some embodiments, the amplification enzyme and the pyrophosphorolysing enzyme in the second region may be the same.

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In some embodiments, there may be a fourth region comprising one or more wells, wherein each well may comprise a proteinase and wherein said fourth region may be located between the first and second regions.

In some embodiments, the second and third regions of the device may be combined such that the wells of the second region further comprise:

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dNTPs;

buffers;

an amplification enzyme; and

a means for detecting a signal derived from A_1 or a portion thereof, or multiple copies of A_1 or

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multiple copies of a portion thereof.

In some embodiments, a means for detecting a signal is located within one or more wells of the second region.

In some embodiments, a means for detecting a signal is located within the second region of the device.

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In some embodiments, a means for detecting a signal is located within an adjacent region of the device.

In some embodiments, there is provided a device comprising:

a fluid pathway between a first region and second region, wherein the first region comprises one or more wells, wherein one or more well comprises:

5 a single-stranded probe oligonucleotide A_0 , which is capable of forming a first intermediate product with a target polynucleotide sequence, said intermediate product being at least partially double-stranded;

a pyrophosphorolysing enzyme capable of digesting the first intermediate product in the 3'-5' direction from the end of A_0 to create a partially digested strand A_1 ;

10 and

one or more ligases capable of ligating A_1 to create an oligonucleotide A_2 .

wherein the second region comprises one or more wells.

In some embodiments, one or more well of the first region may further comprise a source of ions to drive the pyrophosphorolysis reaction forward.

15 In some embodiments, the ions are pyrophosphate ions.

In some embodiments, the 5' end of A_0 is resistant to 5'-3' exonuclease digestion and wherein the wells of the first region further comprise a 5'-3' exonuclease.

20 In some embodiments, the device may further comprise a third region comprising one or more wells which is joined to the first region by a fluid pathway and wherein one or more wells of the third region comprises:

dNTPs;

25 a single-stranded primer oligonucleotide; and

an amplification enzyme.

In some embodiments, the dNTPs of the third region may be dUTP, dGTP, dCTP and dATP; the amplification enzyme may be a dUTP incorporating high fidelity polymerase; and

30 one or more wells of the third region may further comprise uracil-DNA N-glycosylase

in some embodiments, the device may further comprise a fourth region, located between the first and third regions, comprising one or more wells, wherein one or more wells may comprise a proteinase.

35 In some embodiments, the one or more wells of the first or second regions may further comprise a ligase and a ligation probe oligonucleotide C which is complementary to a region of A_0 .

In some embodiments, the one or more wells of the first or second regions may further comprise a ligase and a splint oligonucleotide D which is complementary to a region of A_0 .

5 In some embodiments, the one or more wells of the first or second region may further comprise a ligase, a splint oligonucleotide D and a ligation probe oligonucleotide C.

In some embodiments, the ligation probe oligonucleotide C may comprise a 3' or internal modification protecting it from 3'-5' exonuclease digestion.

10 In some embodiments, D may comprise an oligonucleotide region complementary to the 3' end of A_1 and a region complementary to either the 5' end of oligonucleotide C or to the 5' end of A_1 .

15 In some embodiments, D may be unable to undergo extension against A_1 by virtue of either a 3' modification or through a mismatch between the 3' end of D and the corresponding region of A_1 or C.

In some embodiments, one or more wells of the first region may comprise at least one or more different A_0 each selective for a different target sequence and each including an identification region.

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In some embodiments, the wells of the second region may comprise:

dNTPs;

buffers;

an amplification enzyme;

25 a means for detecting a signal derived from A_1 or a portion thereof, or multiple copies of A_1 or multiple copies of a portion thereof.

In some embodiments, a means for detecting a signal is located within one or more wells of the second region.

30 In some embodiments, a means for detecting a signal is located within the second region of the device.

In some embodiments, a means for detecting a signal is located within an adjacent region of the device.

In some embodiments, one or more wells of the second region may further comprise one or more oligonucleotide binding dyes or molecular probes.

5 In some embodiments, the amplification enzyme and the pyrophosphorolysing enzyme of the device are the same.

In some embodiments, the wells of the second region further comprise:

- 10 - two or more Ligation Chain Reaction (LCR) probe oligonucleotides that are complementary to adjacent sequences on A_1 wherein when the probes are successfully annealed the 5' phosphate of one LCR probe is directly adjacent to the 3'OH of the other LCR probe; and
- one or more ligases.

15 In some embodiments, the wells of the second region may comprise:

- A ligation probe oligonucleotide C;
- A splint oligonucleotide D;

20 wherein C has a 5' phosphate, the 3' end of a splint oligonucleotide D is complementary to the 5' end of C and the 5' end of D is complementary to the 3' end of A_1 such that A_1 and C are capable of being ligated together to form an oligonucleotide A_2 .

In some embodiments, the wells of the second region may further comprise:

- 25 - A hairpin oligonucleotide 1 (HO1) comprising a fluorophore-quencher pair, wherein HO1 is complementary to A_2 and when annealed to A_2 the hairpin structure of HO1 opens and the fluorophore-quencher pair separate; and
- A hairpin oligonucleotide 2 (HO2) comprising a fluorophore-quencher pair, wherein HO2 is complementary to the open HO1 and when annealed to HO1 the hairpin structure of HO2 opens and the fluorophore-quencher pair separate.

30 In some embodiments, the wells of the second region may further comprise a plurality of HO1 and HO2.

In some embodiments, the wells of the second region may further comprise:

- 35 - an oligonucleotide A comprising a substrate arm, a partial catalytic core and a sensor arm;

- an oligonucleotide B comprising a substrate arm, a partial catalytic core and a sensor arm; and

- a substrate comprising a fluorophore quencher pair;

wherein the sensor arms of oligonucleotides A and B are complementary to flanking regions of A_2

5 such that in the presence of A_2 , oligonucleotides A and B are combined to form a catalytically, multicomponent nucleic acid enzyme (MNAzyme).

In some embodiments, the wells of the second region may comprise a partially double-stranded nucleic acid construct wherein:

10 - one strand comprises at least one RNA base, at least one fluorophore and wherein a region of this strand is complementary to a region of A_2 and wherein this strand may be referred to as the 'substrate' strand; and

- the other strand comprises at least one quencher and wherein a region of this strand is complementary to a region of A_2 adjacent to that which the substrate strand is complementary to,
15 such that in the presence of A_2 the partially stranded nucleic acid construct becomes substantially more double-stranded.

In some embodiments, the wells of the second region may further comprise an enzyme for the removal of the at least one RNA base.

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In some embodiments, the enzyme is Uracil-DNA Glycosylase (UDG) and the RNA base is uracil.

In some embodiments, one or more wells of the second region may further comprise:

25 an oligonucleotide complementary to a region of A_2 including the site of ligation, comprising one or multiple fluorophores arranged such that their fluorescence is quenched either by their proximity to each other or to one or more fluorescence quenchers;

a double strand specific DNA digestion enzyme;

30 wherein, in the presence of A_2 , the labelled oligonucleotide is digested such that the fluorophores are separated from each other or from their corresponding quenchers, and a fluorescent signal, and hence the presence of A_2 , is detectable.

In some embodiments, the double strand specific DNA digestion enzyme is an exonuclease.

In some embodiments, the double-strand specific DNA digestion enzyme is a polymerase with proofreading activity.

- 5 In some embodiments, the fluorophore is selected from dyes of the fluorescein family, the carboxyrhodamine family, the cyanine family, the rhodamine family, polyhalofluorescein-family dyes, hexachlorofluorescein-family dyes, coumarin-family dyes, oxazine-family dyes, thiazine-family dyes, squaraine-family dyes, and chelated lanthanide-family dyes.

- 10 In some embodiments, the fluorophore of the device may be selected from any of the commercially available dyes.

In some embodiments, the quencher of the device is selected from those available under the trade designations Black Hole™, Eclipse™ Dark, Qx1J, Iowa Black™, ZEN and/or TAO.

- 15 In some embodiments, the quencher of the device may be selected from any of the commercially available quencher.

- 20 In some embodiments, one or more wells of the second region may further comprise one or more partially double stranded DNA constructs wherein each construct contains one or more fluorophores and one or more quenchers. In some embodiments, when the construct is partially double-stranded the one or more fluorophores and one or more quenchers are located in close enough proximity to each other such that sufficient quenching of the one or more fluorophores occurs.

- 25 In some embodiments, the construct is one strand of DNA with a self-complementary region that is looped back on itself.

In some embodiments, the construct comprises one primer of a primer pair.

- 30 In some embodiments, one or more wells of the second region may further comprise the other primer of a primer pair.

In some embodiments, a portion of the single stranded section of the construct hybridises to A₂ and is extended against it by a DNA polymerase. In some embodiments, the other primer of the

primer pair then hybridises to the extended construct, displaying A_2 . This primer is then extended against the construct, displacing the self-complementary region. Thus, the one or more fluorophores and one or more dyes are separated sufficiently for a fluorescent signal to be detected, indicating the presence of A_2 .

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In such an embodiment the construct may be known as a Sunrise Primer.

In some embodiments, the construct comprises two separate DNA strands.

10 In some embodiments, a portion of the single stranded section of the construct hybridises to A_2 and is extended against it by a DNA polymerase. In some embodiments, the other primer of the primer pair then hybridises to the extended construct, displaying A_2 . This primer is then extended against the construct, in the direction of the double stranded section, displacing the shorter of the DNA strands and thus the one or more fluorophores and one or more dyes are separated
15 sufficiently for a fluorescent signal to be detected, indicating the presence of A_2 .

In such an embodiment the construct may be known as a Molecular Zipper.

The person skilled in the art will appreciate that for both the Sunrise Primer and Molecular Zipper
20 it is possible for the one or more fluorophores and the one or more quencher pairs to be located at various positions within each respective construct. The key feature is that each pair is located in sufficient proximity to one another that in the absence of A_2 , i.e. when no extension and strand displacement has occurred, no fluorescent signal is emitted.

25 In some embodiments, one or more wells of one or more regions may further comprise a pyrophosphatase.

In some embodiments, one or more wells of one or more regions of the device may further comprise a phosphatase or a phosphohydrolase.

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In some embodiments, one or more wells of the first region of the device may further comprise an enzyme for the formation of DNA from an RNA template.

In some embodiments, the enzyme is a reverse transcriptase.

In some embodiments, one or more enzymes present in the device are hot start.

- 5 In some embodiments, one or more enzymes present in the device are thermostable.

In some embodiments, the first and second regions of the device are combined.

- 10 In some embodiments, there is located, between one or more wells of a region/and or between one or more regions of the device, one or more fluidic pathways.

In some embodiments, the first region may be fluidically connected to a sample container via a fluidic interface.

- 15 In some embodiments, heating and/or cooling elements may be present at one or more regions of the device.

In some embodiments, heating and/or cooling may be applied to one or more regions of the device.

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In some embodiments, each region of the device may independently comprise at least 100 or 200 wells.

- 25 In some embodiments, each region of the device may independently comprise between about 100 and 300, 400, 500, 600, 700, 800, 900, 1000, 1100, 1200, 1300, 1400, 1500 or more wells. The wells may be of any shape and their locations may be arranged in any format or pattern on a substrate.

- 30 In some embodiments, the well-substrate can be constructed from a metal (e.g. gold, platinum, or nickel alloy as non-limiting examples), ceramic, glass, or other PCR compatible polymer material, or a composite material. The well-substrate includes a plurality of wells.

In some embodiments, the wells may be formed in a well-substrate as blind-holes or through-holes. The wells may be created within a well-substrate, for example, by laser drilling (e.g. excimer or solid-state laser), ultrasonic embossing, hot embossing lithography, electroforming a nickel mold, injection molding, and injection compression molding.

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In some embodiments, individual well volume may range from 0.1 to 1500nl. In one embodiment, 0.5 to 50nL. Each well may have a volume of about 0.1, 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, 0.9, 1, 1.5, 2, 2.5, 3, 3.5, 4, 4.5, 5, 5.5, 6, 6.5, 7, 7.5, 8, 8.5, 9, 9.5, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 25, 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, 110, 120, 130, 140, 150, 160, 170, 180,

10 190, 200, 225, 250, 275, 300, 325, 350, 375, 400, 425, 450, 475, or 500 nL.

In some embodiments, well dimensions may have any shape, for example, circular, elliptical, square, rectangular, ovoid, hexagonal, octagonal, conical, and other shapes well known to persons of skill in the art.

15 In some embodiments, well shapes may have cross-sectional areas that vary along an axis. For example, a square hole may taper from a first size to a second size that is a fraction of the first size.

In some embodiments, well dimensions may be square with diameters and depths being approximately equal.

20 In some embodiments, walls that define the wells may be non-parallel.

In some embodiments, walls that define the wells may converge to a point. Well dimensions can be derived from the total volume capacity of the well-substrate.

In some embodiments, well depths may range from 25 μm to 1000 μm .

25 In one embodiment, wells may have a depth of 25, 50, 75, 100, 150, 200, 250, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950 or 1000 μm .

In some embodiments, well diameter may range from about 25 μm to about 500 μm .

In some embodiments, wells may have a width of 25, 50, 75, 100, 125, 150, 175, 200, 225, 250, 275, 300, 325, 350, 375, 400, 425, 450, 475 or 500 μm .

30 In some embodiments, portions of one or more regions of the device may be modified to encourage or discourage fluid adhered. Surfaces defining the wells may be coated with a hydrophilic material (or modified to be hydrophilic), and thus encourage retention of fluid.

In some embodiments, portions of one or more regions of the device, may be coated with a hydrophobic material (or modified to be hydrophobic) and thus discourage retention of fluid thereon. The person skilled in the art will understand that other surface treatments may be performed such that fluid is preferably held within the wells, but not on upper surfaces so as to encourage draining of excess fluid.

In some embodiments, the wells of the well-substrate may be patterned to have a simple geometric pattern of aligned rows and columns, or patterns arranged diagonally or hexagonally. In one embodiment, the wells of the well-substrate may be patterned to have complex geometric patterns, such as chaotic patterns or isogeometric design patterns.

In some embodiments, the wells may be geometrically separated from one another and/or feature large depth to width ratios to help prevent cross-contamination of reagents. In some embodiments, the device may comprise one or more axillary regions which are usable to provide process fluids, such as oil or other chemical solutions to one or more of the regions of the device. Such auxiliary regions may be fluidically connected to one or more of the regions of the device via one or more membranes, valves and/or pressure severable substrates (i.e. materials that break when subjected to a pre-determined amount of pressure from fluid within an auxiliary region or adjacent portion of the fluid pathway) such as metal foil or thin film.

In some embodiments, the fluid pathway of the device may include extensive torturous portions. A torturous path between the inlet passage of the fluid pathway and one or more of the regions of the device can be helpful for control and handling of fluid processes. A torturous path can help reduce formation of gas bubbles that can interfere with flowing oil through the fluid pathway.

In some embodiments, the device may further comprises a gas permeable membrane which enables gas to be evacuated from the wells of one or more regions of the device, while not allowing fluid to pass through. The gas permeable membrane may be adhered to the well-substrate of the device by a gas permeable adhesive. In one embodiment, the membrane may be constructed from polydimethylsiloxane (PDMS), and has a thickness ranging from 20-1000 μm . In some embodiments the membrane may have a thickness ranging from 100-200 μm .

In some embodiments, all or portions of the well-substrate may contain conductive metal portions (e.g., gold) to enable heat transfer from the metal to the wells. In one embodiment, the interior surfaces of wells may be coated with a metal to enable heat transfer.

- 5 In some embodiments, after appropriate reagents have filled the wells of one or more regions of the device, an isolation oil or thermally conductive liquid may be applied to the device to prevent cross-talk.

- 10 In some embodiments, the wells of one or more regions of the device may be shaped to taper from a large diameter to a smaller diameter, similar to a cone. Cone-shaped wells with sloped walls enables the use of a non-contact deposition method for reagents (e.g., ink jet). The conical shape also aids in drying and has been found to prevent bubbles and leaks when a gas permeable membrane is present.

- 15 In some embodiments, the wells of one or more regions of the device may be filled by advancing a sample fluid (e.g. via pressure) along the fluid pathway of the device. As the fluid passes over the wells of one or more regions of the device, each well becomes filled with fluid, which is primarily retained within the wells via surface tension. As previously described, portions of the well-substrate of the device may be coated with a hydrophilic/hydrophobic substance as desired
20 to encourage complete and uniform filling of the wells as the sample fluid passes over.

- In some embodiments, the wells of one or more regions of the device may be 'capped' with oil following filling. This can then aid in reducing evaporation when the well-substrate is subjected to heat cycling. In one embodiment, following oil capping, an aqueous solution can fill one or more regions of the device to improve thermal conductivity.

25

In some embodiments, the stationary aqueous solution may be pressurised within one or more regions of the device to halt the movement of fluid and any bubbles.

- In some embodiments, oil such as mineral oil may be used for the isolation of the wells of one or more regions of the device and to provide thermal conductivity. However, any thermal conductive
30 liquid, such as fluorinated liquids (e.g., 3M FC-40) can be used. References to oil in this disclosure should be understood to include such alternatives as the skilled person in the art will appreciate are applicable.

In some embodiments, the device may further comprise one or more sensor assemblies.

In some embodiments, the one or more sensor assemblies may comprise a charge coupled device (CCD)/complementary metal-oxide-semiconductor (CMOS) detector coupled to a fiber optic face plate (FOPF). A filter may be layered on top of the FOPF, and placed against or adjacent to the well-substrate. In one embodiment, the filter can be layered (bonded) directly on top of the CCD with the FOPF placed on top.

In some embodiments, a hydration fluid, such as distilled water, may be heated within the first region or one of the auxiliary regions such that one or more regions of the device has up to 100% humidity, or at least sufficient humidity to prevent over evaporation during thermal cycling.

In some embodiments, after filing of the device is complete, the well-substrate may be heated by an external device that is in thermal contact with the device to perform thermal cycling for PCR.

In some embodiments, non-contact methods of heating may be employed, such as RFID, Curie point, inductive or microwave heating. These and other non-contact methods of heating will be well known to the person skilled in the art. During thermal cycling, the device may be monitored for chemical reactions via the sensor arrangements previously described.

In some embodiments, reagents that are deposited in one or more of the wells of one or more of the regions of the device are deposited in a pre-determined arrangement.

20

In some embodiments there is provided a method comprising:

providing a sample fluid to a fluid pathway of a device wherein the device comprises at least a fluid pathway between a first region, a second region and a third region, wherein the first, second and third regions independently comprise one or more wells;

25 filling the second region with the amplified fluid from the first region such that one or more wells of the second region is coated with the amplified fluid;

 evacuating the amplified fluid from the second region such that one or more wells remain wetted with at least some of the amplified fluid;

 filling the third region with the fluid evacuated from the second region such that one or more wells of the third region is coated with this fluid; and

30 evacuating the fluid from the third chamber such that the one or more wells remains wetted with at least some of this fluid.

In some embodiments of the method, the fluid pathway may be valveless.

In some embodiments of the method, the evacuated second region may be filled with a hydrophobic substance.

5 In some embodiments of the method, the evacuated third region may be filled with a hydrophobic substance.

In some embodiments of the method, the hydrophobic substance may be supplied from an oil chamber that is in fluid communication with the second and third regions.

In some embodiments of the method, the sample fluid may be routed along the fluid pathway in a serpentine manner.

10 In some embodiments, the method may further comprise applying heating and cooling cycles to the one or more of the first, second or third regions.

Various further aspects and embodiments of the present invention will be apparent to those skilled in the art in view of the present disclosure.

15 “and/or” where used herein is to be taken as specific disclosure of each of the two specified features or components with or without the other. For example “A and/or B” is to be taken as specific disclosure of each of (i) A, (ii) B and (iii) A and B, just as if each is set out individually herein.

20 Unless context dictates otherwise, the descriptions and definitions of the features set out above are not limited to any particular aspect or embodiment of the invention and apply equally to all aspects and embodiments which are described.

25 It will further be appreciated by those skilled in the art that although the invention has been described by way of example with reference to several embodiments, it is not limited to the disclosed embodiments and that alternative embodiments could be constructed without departing from the scope of the invention as defined in the appended claims.

Example 1 – Simplified Protocols

For the purpose of this, and following sections, embodiments of the invention are exemplified and referred to as Protocol 1-5 respectively.

Fig. 1 provides an overview of the different Protocols.

5

The table below shows an overview of the time taken to perform each Protocol:

Protocol Step	Time [min]				
	1	2	3	4	5
PCR	60-100				
Exo 5-3'	30		-		
Proteinase K	70		15		-
PPL	30	30	30	15	
Ligation	20	20			
TIPP	10		10	50*	
Exo	10	10			
RCA	50				
Total time	280-320	270-310	165-205	140-180	125-165

10 In one embodiment, TIPP is absent from any one of the protocols, methods, kits and or devices of the invention.

In one embodiment, a 5'-3' exonuclease is absent from any one of the protocols, methods, kits and or devices of the invention.

15

5

The tables below show an overview of the enzymes which may be used in each Protocol:

Protocol Step	Enzyme				
	1	2	3	4	5
PCR	High Fidelity Polymerase, Uracil-DNA Glycosylase				
Exo	5-->3' Exonuclease		-		
Proteinase	Proteinase				-
PPL	Pplase, diphosphatase		Pplase, Ligase, 5-->3' exo, diphosphatase		
Ligation	Ligase	Ligase, Pyrophosphatase			
TIPP	Pyrophosphatase	Pyrophosphatase	Exonuclease 3-->5' or/and 5-->3', Pyrophosphatase	Pyrophosphatase, DNA polymerase	
Exo	Exonuclease 3-->5' or/and 5-->3'				
Amplification	DNA polymerase				
Number of enzymes	12	12	10	9	8

In one embodiment, the presence of pyrophosphatase is optional.

In one embodiment, the presence of a 5'-3' exonuclease is optional.

- 5 In one embodiment, the presence of UDG is optional.

Protocol Step	Enzyme				
	1	2	3	4	5
PCR	PhusionU/Q5, UDG				
Exo 5-3'	Lambda, PNK		-		
Proteinase K	Proteinase K				
PPL	Mako/Klenow/BST L.F./Bsu, Apyrase		Mako/Klenow/BST L.F./Bsu, E.coli Ligase/T4 Ligase/T3 Ligase/HiFi Ligase/ 9oN Ligase, Lambda exo, Apyrase		
Ligation	E.coli Ligase/T4 Ligase/T3 Ligase/HiFi Ligase/ 9oN Ligase	E.coli Ligase/T4 Ligase/T3 Ligase/HiFi Ligase/ 9oN Ligase, TIPP			
TIPP	TIPP		ExoIII, TIPP	TIPP, BST L.F/BST 2.0 WS/Klenow/phi29/	

Exo	T5, Exo III			AmpliTaq/TaqPolymerase/Q5/Phusion Flex	
Amplificati on	BST L.F./BST 2.0 WS/Klenow/phi29				
Number of enzymes	12	12	10	9	8

As can be seen, the inventors have reduced total number of enzymes needed thus reducing the cost and complexity of the method. Surprisingly, the inventors discovered that moving the 5'-3' exonuclease addition from the pre-amplification step to the pyrophosphorolysis/ligation step of the protocol (as in protocols 3-5) results in a higher fluorescent signal (representing detection of particular target analyte sequence) as shown in Fig. 2.

Example 2: Pyrophosphorolysing (PPL) Enzymes

The inventors have tested the method of Protocol 3 of the current invention using a range of different PPL enzymes, the results of which can be seen in Fig. 3. Fig. 3 (A) shows detection of 1% MAF T790M using Mako, Klenow and Bsu. Fig. 3(B) shows the detection of 0.5% MAF T790M using Bst LF at a range of different PPI concentrations.

The inventors have tested the method of Protocol 4 of the current invention using a range of different PPL enzymes, the results of which can be seen in Fig. 4.

Example 3: Protocol 1 vs Protocol 4

The inventors have detected Exon19 del_6223 at 0.5%, 0.10% and 0.05% MAF, which can be seen in Fig. 5, using both Protocol 1 and Protocol 4. As can be seen, the fluorescent peaks are greater when using Protocol 4.

Example 4: Protocol 4 – Sensitivity

The inventors have detected EGFR exon 20 T790M mutations at 0.10%, 0.50% and 1% MAF as shown in Fig. 6 according to Protocol 4.

Example 5: Protocol 4 – Is an exonuclease digestion step needed during RCA?

The inventors have demonstrated that an exonuclease digestion step during RCA is not essential. However, a detectable signal is detected later in the RCA if the exonuclease digestion step is omitted. Fig. 7 shows detection of EGFR exon 20 T790M at 1% MAF with and without the presence of an exonuclease in the RCA step.

Example 6: Protocol 4 – PPL:RCA mix ratio

The inventors have investigated what effect the PPL:RCA mix ratio has on the intensity of signal detected for 0.5% MAF EGFR exon 20 T790M, the results of which are shown in Fig. 8. As can be seen a ratio of 1:2 PPL:RCA mix results in the lowest signal intensity but at the earliest time point. This is followed closely in time by 1:4 PPL:RCA mix which has a greater signal intensity. The largest signal intensity is seen for 1:8 PPL:RCA mix at the latest time point in the reaction.

Example 7: Protocol 4 – Dye choice

The inventors have investigated whether the dye used during RCA can be optimised. Fig. 9 shows the results of comparison experiments performed according to Protocol 4 using SybrGreenI (50°C and 60°C) and Syto82 (50°C and 60°C). The Syto82 dye allows the RCA to be run at a lower temperature of 50°C, whereas SybrGreenI requires the higher 60°C temperature. A lower RCA temperature is needed for Protocol 5 which removes the addition of Proteinase K to the reaction mixture. The amplification enzyme used for preparing at least one single-stranded analyte of a nucleic acid comprised of a target polynucleotide region, for detection using the methods of the current invention, requires a temperature of greater than 50°C to work. The use of SybrGreenI necessitates a reaction temperature of 60°C and thus Proteinase K must be added at some point during the method to deactivate the amplification enzyme prior to RCA.

A lower RCA temperature may allow the methods of the invention to be carried out in a plate reader instead of qPCR.

The reaction utilising Syto82 is faster, as can be seen in Fig. 9, and although the total amount of fluorescence is lower for Syto82 – this can be alleviated by the use of a higher concentration of

Syto82 dye.

Example 8: Protocol 4 – BST L.F. vs BST 2.0 WS

The inventors have investigated the use of two different enzymes, BST L.F and BST 2.0 WS, for
5 RCA according to Protocol 4 to detect 0.5% MAF EGFR exon 20 T790M mutation. The results of
this are shown in Fig. 10 where it can be seen that the reaction is fastest with BST 2.0 WS. BST 2.0
WS is designed to incorporate dUTP, which helps with the speed of the reaction. There is a
negligible difference in total signal intensity achieved between BST L.F. and BST 2.0 WS. According
to its description, provided by New England Biolabs (NEB), BST 2.0 WS should be more stable and
10 active only above 45°C.

Example 9: Effect of PPL Enzymes on Signal Detection

The inventors have investigated the effect of different PPL enzymes on the RCA reaction at
different PPL:RCA reaction mixture ratios. The results of which can be seen in Fig. 11(A) 1:4
15 PPL:RCA and Fig. 11(B) 1:8 PPL:RCA. All PPL enzymes, excepting BST, impact the RCA reaction at
1:4 PPL:RCA ratio. At 1:8 PPL:RCA ratio, BST and Klenow have no impact on the RCA reaction.

Example 10: Pyrophosphorolysis, ligation specificity against single nucleotide mismatches

20 A single-stranded first oligonucleotide 1 (SEQ ID NO: 1) was prepared, having the following
nucleotide sequence:

5'-

/5Phos/A*T*G*TTTCGATGAGCTTTGACAATACTTGAAGCTCGCAGATATAGGATGTTGCGATAGT
CCAGGAGGCTGC-3'

25

A single-stranded ligating oligonucleotide 2 (SEQ ID NO: 2) was prepared, having the
following nucleotide sequence:

SEQ ID NO: 2: 5'- TGTCAAAGCTCATCGAACATCCTGGACTATGTCTCC-3'

wherein A, C, G, and T represent nucleotides bearing the relevant characteristic nucleobase of
30 DNA,

/5Phos/ represent 5' end phosphate

* represent phosphorothioate bond

A set of single-stranded oligonucleotides 3-4 (SEQ ID NOs 3-4) was also prepared, having the following nucleotide sequences in the 5' to 3' direction:

SEQ ID NO: 3:

5 TGCTGGGCATCTGCCTCACCTCCACCGTGCAGCTCATCACGCAGCTCATGCCCTTCGGCAGCCTCC
TGGACTATG

SEQ ID NO: 4:

TGCTGGGCATCTGCCTCACCTCCACCGTGCAGCTCATCACGCAGCTCATGCCCTTCGGCTGCCTCC
TGGACTATG

10 wherein oligonucleotide 3 includes a 17 base region complementary to the 17 bases at the 3' end of oligonucleotide 1 and oligonucleotide 4 include the same region with single nucleotide mismatches at positions 3. SEQ IDs 3 and 4 are part of the human EGFR gene with/without the C797S mutation respectively.

15 A first reaction mixture was then prepared, having a composition corresponding to that derived from the following formulation:

0.5uL 20x buffer pH 7.0
0.25 uL 5x buffer pH 8.0
0.25 uL 5x HF buffer
20 0.2uL oligonucleotide 1, 1000 nM
0.3 uL oligonucleotide 2, 1000 nM
1uL oligonucleotide 2 (500 nM) or mixture of oligo 2 and 3 (500 and 0.5 nM respectively),
0.3U Klenow Fragment exo- (NEB)
0.01uL inorganic pyrophosphate, 10mM
25 0.0132 U Apyrase (ex. NEB)
1 U E. coli DNA Ligase (ex. NEB)
Water to 10uL

wherein the 20x buffer comprised the following mixture:

200uL Tris Acetate, 1M, pH 7.0
30 342.5uL aqueous Magnesium Acetate, 1M
120uL aqueous Potassium Acetate, 5M
50uL Triton X-100 surfactant (10%)
Water to 1mL

wherein the 5x buffer comprised the following mixture:

50uL Trizma Acetate, 1M, pH 8.0
 25uL aqueous Magnesium Acetate, 1M
 25uL aqueous Potassium Acetate, 5M
 50uL Triton X-100 surfactant (10%)
 5 Water to 1mL

Pyrophosphorolysis, followed by circularisation, via ligation of, oligonucleotide 1 was then carried out by incubating the mixture at 45°C for 15 minutes and the resulting product mixture was used in the amplification reaction (Example 11).

10

Example 11: Amplification of circularised probe

A pair of single stranded oligonucleotide primers 1 (SEQ ID NO 5) and 2 (SEQ ID NO 6) were prepared, having the following nucleotide sequences:

(SEQ ID NO: 5)1: TCGCAACATCCTATATCTGC

15 (SEQ ID NO: 6) 2: TGAGCTTTGACAATACTTGA

wherein A, C, G, and T represent nucleotides bearing the relevant characteristic nucleobase of DNA.

20 A second reaction mixture was then prepared, having a composition corresponding to that derived from the following formulation:

3 uL 10x Thermopol buffer
 3.2 U BST 2.0 WS
 0.32 uL oligonucleotide 1, 10uM
 0.32 uL oligonucleotide 2, 10uM
 25 1.125 uL Syto82, 30 uM
 0.165 U Inorganic Pyrophosphatase
 1.2 uL dNTPs mix, 10 mM
 1.25 uL reaction mixture in Example 10
 Water to 11.25 uL

30

wherein the 10x Thermopol buffer comprised the following mixture:

200 uL Tris-HCl pH=8.8, 1M
 100 uL (NH₄)₂SO₄, 1M

100 μ L mM KCl, 1M
20 mM MgSO₄, 1M
10 μ L Triton® X-100, 10%
Water to 1mL

5

The reaction mix was then incubated at 50°C for 40 minute and the resulting reaction product was then analysed by real-time fluorescence. The results of which are shown in Fig. 12. From this analysis it can be seen that when oligonucleotide 3 and 4 are both present, the fluorescent signal appears faster in the reaction, showing that pyrophosphorolysis and ligation of

10

oligonucleotide 3 has occurred in the first reaction mixture.

Example 12: Multi-colour detection using Sunrise Primers**1. Target oligo dilution**

WT oligo dilution is made of following ingredients

0.5x A7 buffer

5 0.5x Phusion U buffer

200 nM WT oligonucleotide (SEQ ID NO: 7)

Total volume: 5 uL

T790M and C797S 1% AF mutant oligo mix:

10 0.5x A7 buffer

0.5x Phusion U buffer

100 nM WT oligonucleotide (SEQ ID NO: 7)

2 nM T790M oligonucleotide (SEQ ID NO: 8)

2 nM C797S_2389 oligo (SEQ ID NO: 9)

15 Total volume: 5 uL

WT oligonucleotide (SEQ ID NO: 7):

5'-

20 CATCTGCCTCACCTCCACCGTGCAGCTCATCACGCAGCTCATGCCCTTCGGCTGCCTCCTGGACTATGTCCG
GGAACACAAAGACAATAT-3'

T790M oligonucleotide (SEQ ID NO: 8):

5'-

25 CATCTGCCTCACCTCCACCGTGCAGCTCATCATGCAGCTCATGCCCTTCGGCTGCCTCCTGGACTATGTCCG
GGAACACAAAGACAATAT-3'

C797S_2389 oligonucleotide (SEQ ID NO: 9):

5'-

30 CATCTGCCTCACCTCCACCGTGCAGCTCATCACGCAGCTCATGCCCTTCGGCAGCCTCCTGGACTATGTCCG
GGAACACAAAGACAATAT-3'

1x A7 composition

Tris Acetate pH=8.0 10mM

Potassium Acetate 25mM

35 Magnesium Acetate 5mM

Triton-X 0.01%

PhusionU buffer

The PhusionU buffer composition is not publicly available.

40

2. PPL

A mixture was prepared corresponding to:

1xBFF1

37.5 U/mL Mako DNA Polymerase (3' → 5' exo-)

45 100 U/mL E.coli Ligase

1.2 U/mL apyrase

0.6 mM PPi

20 nM T790M probe

20 nM C797S_2389 probe

30 nM T790M splint oligonucleotide
30 nM C797S_2389 splint oligonucleotide
5 uL of WT or 1%AF mutant dilution point 1.
Total volume 10 uL

- 5 This mixture was then incubated at 41°C for 30 min.

1xBFF1 composition

- Tris Acetate pH=7.0 10mM
Potassium Acetate 30mM
10 Magnesium Acetate 17.125mM
Triton-X 0.01%

T790M probe (SEQ ID NO : 10):

5'-

- 15 /5Phos/A*T*G*TTTCGATGAGCTTTGACAATACTTGAGCACGGCAGATATAGGATGTTGCGAAGGGCATGA
GCTGCATGATGAGCTG'-3'

C797S_2389 probe (SEQ ID NO : 11):

5'-

- 20 /5Phos/A*T*G*TTTCGATGAGCTTTGACAATACTTGAAGCTCGCAGATATAGGATGTTGCGATAGTCCAGG
AGGCTGC-3'

where * represents a phosphorothioate bond

3. TIPP

- 25 A mixture was prepared corresponding to:

1xA7

66.6 U/mL TIPP

10 uL mixture from point 2.

Total volume: 20 uL

- 30 This mixture was then incubated at 25°C for 5 min, 95°C for 5 min.

4. Ligation

A mixture was prepared corresponding to:

- 35 1xA7

100 U/mL E. coli ligase

20 uL mixture from point 3.

10 nM T790M splint oligonucleotide

10 nM C797S_2389 splint oligonucleotide

- 40 Total volume: 30 uL

This mixture was then incubated at 37°C for 10 min, 95°C for 10 min.

T790M splint oligonucleotide (SEQ ID NO: 12):

5'- TGTCAAAGCTCATCGAACATGCCCTTCGCAACATCT-3'

- 45

C797S_2389 splint oligonucleotide (SEQ ID NO: 13):

5'- TGTCAAAGCTCATCGAACATTCCTGGACTATCGCAT-3'

5. Exonuclease treatment

A mixture was prepared corresponding to:

1xA7

100 U/mL E. coli ligase

5 30 uL mixture from point 4.

625 U/mL of Exonuclease III

62.5 U/mL T5 Exonuclease

Total volume: 40 uL

This mixture was then incubated at 30°C for 5 min, 95°C for 5 min.

10

6. RCA

A mixture corresponding to the following was prepared:

1x Thermopol buffer (53.2 mM Tris-HCl, 26.6 mM (NH₄)₂SO₄, 26.6 mM KCl, 5.32 mM MgSO₄, 0.266% Triton® X-100, pH 8.8)

15 0.2 uM Primer mix 1

0.4 uM Reverse primer

533.3 U/mL BST L.F.

0.4 mM dNTPs

10 uL of reaction mixture from point 5.

20 Total volume 15 uL

Primer mix 1:

Cy5 primer (SEQ ID NO: 14):

5'-

25 /Qusar670/ACGCCTGGTTACCGAGCCAGGTTTCGCACATGTAGGCTCGGTAACCAGGCG/BHQ2/ACATCCT
ATATCTGCCGTGC-3'

TexasRed primer (SEQ ID NO: 15):

5'-

30 /TexasRed/ACGCCTGGTTACAGGTTTCGCACATGTAGTAACCAGGCG/BHQ2/CAACATCCTATATCTGCGA
G-3'

where /BHQ2/ represents Black Hole Quencher;

Reverse primer (SEQ ID NO: 16):

35 5'-ATGTTGATGAGCTTTGACA-3'

The mixture was then incubated at 60°C for 90 min. Fluorescent measurements were taken every 1 minute. Cq was obtained based on auto-threshold given by Bio-rad machine. The results of this can be seen in Fig 13.

40

Example 13: Multi-colour detection using Molecular Zippers

1. Target oligo dilution

WT oligo dilution is made of following ingredients

45 0.5x A7 buffer

0.5x Q5U buffer

100 nM WT oligonucleotide (SEQ ID NO: 17)

Total volume: 1.25 uL

G719X_6239, G719X_6252, G719X_6253 0.5% AF mutant oligonucleotide mix:

0.5x A7 buffer

0.5x Q5U buffer

100 nM WT oligonucleotide (SEQ ID NO: 17)

5 0.5 nM G719X_6239 oligonucleotide (SEQ ID NO: 18)

0.5 nM G719X_6252 oligonucleotide (SEQ ID NO: 19)

0.5 nM G719X_6253 oligonucleotide (SEQ ID NO: 20)

Total volume: 1.25 uL

10 WT oligonucleotide (SEQ ID NO: 17):

5'-

CCCAACCAAGCTCTCTTGAGGATCTTGAAGGAACTGAATTCAAAAAGATCAAAGTGCTGGGCTCCGGTGC
GTTCGGCACGGTGTATAAGGTAAGGTCCC-3'

15 G719X_6239 oligonucleotide (SEQ ID NO: 18):

5'-

CCCAACCAAGCTCTCTTGAGGATCTTGAAGGAACTGAATTCAAAAAGATCAAAGTGCTGGCCTCCGGTGC
GTTCGGCACGGTGTATAAGGTAAGGTCCC-3'

20 G719X_6252 oligonucleotide (SEQ ID NO: 19):

5'-

CCCAACCAAGCTCTCTTGAGGATCTTGAAGGAACTGAATTCAAAAAGATCAAAGTGCTGAGCTCCGGTGC
GTTCGGCACGGTGTATAAGGTAAGGTCCC-3'

25 G719X_6253 oligonucleotide (SEQ ID NO: 20):

5'-

CCCAACCAAGCTCTCTTGAGGATCTTGAAGGAACTGAATTCAAAAAGATCAAAGTGCTGTGCTCCGGTGC
GTTCGGCACGGTGTATAAGGTAAGGTCCC-3'

30

1x A7 composition

Tris Acetate pH=8.0 10mM

Potassium Acetate 25mM

Magnesium Acetate 5mM

35 Triton-X 0.01%

Q5U buffer

The Q5U buffer composition is not publicly available.

40

2. Pyrophosphorolysis (PPL) and ligation

A mixture was prepared corresponding to:

1xBFF1

10 U/mL Klenow (exo-)

45 100 U/mL E.coli Ligase

1.2 U/mL apyrase

100 U/mL Lambda exo

0.25 mM PPi

6.6 nM G719X_6239 probe oligonucleotide (SEQ ID NO: 21)

6.6 nM G719X_6252 probe oligonucleotide (SEQ ID NO: 22)

6.6 nM G719X_6253 probe oligonucleotide (SEQ ID NO: 23)

30 nM splint oligonucleotide (SEQ ID NO: 24)

1.25 uL of mixture from point 2.

5 Total volume 10 uL

This mixture was then incubated at 45°C for 15 min.

1xBFF1 composition

Tris Acetate pH=7.0 10mM

10 Potassium Acetate 30mM

Magnesium Acetate 17.125mM

Triton-X 0.01%

G719X_6239 probe oligonucleotide (SEQ ID NO: 21):

15 5'-

/5Phos/A*T*G*TTGATGAGCTTTGACAATACTTGACATGCGCAGATATAGGATGTTGCGAAACGCACCG
GAGGCCAGCACTTTG-3'

G719X_6252 probe oligonucleotide (SEQ ID NO: 22):

20 5'-

/5Phos/A*T*G*TTGATGAGCTTTGACAATACTTGACATGCCGAGTAATGAGAGTTTCGCAAACGCACCG
GAGCTCAGCACTTTG-3'

G719X_6253 probe oligonucleotide (SEQ ID NO: 23):

25 5'-

/5Phos/A*T*G*TTGATGAGCTTTGACAATACTTGACATGCGAGCAATTAGGTAGTGTCGTAACGCACCG
GAGCACAGCACTTTG-3'

Splint oligonucleotide (SEQ ID NO: 24):

30 5'-TGTCAAAGCTCATCGAACATCCGGTTCGTCGCAA-5'

where * represents a phosphorothioate bond

3. Detection – RCA

35 A mixture corresponding to the following was prepared:

2.66x Thermopol buffer (53.2 mM Tris-HCl, 26.6 mM (NH₄)₂SO₄, 26.6 mM KCl, 5.32 mM MgSO₄,
0.266% Triton® X-100, pH 8.8)

0.28 uM Dye primer mix 1

0.56 uM Quencher primer 1

40 0.28 uM Quencher primer 2

0.84 uM Reverse primer

568.8 U/mL BST 2.0 WarmStart

14.67 U/mL TIPP

1.06 mM dNTPs

45 1.25 uL of reaction mixture from point 2.

Total volume 11.25 uL

Dye primer mix 1 consists of:

Dye primer 1 (SEQ ID NO: 25):

5'-/5Cy5/A*CTGACCAGCTCCATGACAATCGCTGTCGCCATGATCGATCGCAACATCCTATATCTGC-3'

Dye primer 2 (SEQ ID NO: 26):

5'-/5TEX615/A*CTGACCAGCTCCATGACAATCGCTGTCGCCATGATCGATGCGAACTCTCATTACTCG-3'

5

Dye primer 3 (SEQ ID NO: 27):

5'-/5HEX/T*ACGACCGACTCACTCCTTACAGCAGTCCGAGTATGCTACGACACTACCTAATTGCTC-3'

where * represents a phosphorothioate bond, /5Cy5/ represent Cy5 dye on 5' end, /5TEX615 represent TEX dye on 5' end, /5HEX/ represents Hex dye on 5' end

10

Quencher primer 1 (SEQ ID NO: 28):

5'-TCGATCATGGCGACAGCGATTGTCATGGAGCTGGTCAGT/3IAbRQSp/-3'

where /3IAbRQSp/ represents 3' Iowa Black® RQ quencher

15

Quencher primer 2 (SEQ ID NO: 29):

5'-AGCATACTGCGGACTGCTGTAAGGAGTGAGTCGGTCGTA/3IAbkFQ/-3'

where /3IAbkFQ/ represents 3' Iowa Black® FQ quencher

Reverse primer (SEQ ID NO: 30):

20

5'-T*G*AGCTTTGACAATACTTGA-3'

where * represents a phosphorothioate bond

The mixture was then incubated at 58°C for 150 min. Fluorescent measurements were taken every 1 minute. The results of this can be seen in Fig 14.

25

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Example 14: Pyrophosphorolysis and ligation against target

1. Oligonucleotide dilution preparation
- 5 Dilution of oligonucleotides were prepared in 0.5x A7 and 0.5x Q5 buffer:
- WT oligonucleotide 200nM
- +/-Mutant oligonucleotide 500pM
- Total volume 1.25 uL
- 10 WT oligonucleotide (SEQ ID NO 31):
- 5'-
CTGCTGGGCATCTGCCTCACCTCCACCGTGCAGCTCATCACGCAGCTCATGCCCTTCGGCTGCCTCCTGGAC
TATGTCCGG-3'
- 15 Mutant oligonucleotide (SEQ ID NO 32):
- 5'-
CTGCTGGGCATCTGCCTCACCTCCACCGTGCAGCTCATCATGCAGCTCATGCCCTTCGGCTGCCTCCTGGAC
TATGTCCGG-3'
2. Pyrophosphorolysis and ligation
- 20 A PPL mixture was prepared consisting of:
- 1xBFF1
- 10 U/mL Klenow (exo-)
- 100 U/mL E.coli Ligase
- 25 1.2 U/mL apyrase
- 100 U/mL Lambda exo
- 0.25 mM PPi
- 20 nM Probe A₀
- 1.25 uL Oligos from point 1
- 30 Total volume 10 uL

Probe A₀ (SEQ ID NO 33):

5'-
/5Phos/A*G*C*TGCATCTGAGCTTTGACAATACTTGAGCACGGCAGATATAGGATGTTGCGAAG
5 GGCATGAGCTGCATGATG-3'

where * Phosphorothioate bonds

1xBFF1 composition

Tris Acetate pH=7.0 10mM

10 Potassium Acetate 30mM

Magnesium Acetate 17.125mM

Triton-X 0.01%

1xA7 composition

15 Tris Acetate pH=8.0 10mM

Potassium Acetate 25mM

Magnesium Acetate 5mM

Triton-X 0.01%

20 Q5 buffer

The Q5 buffer composition is not publicly available.

The resulting mixture was incubated at 45°C for 15 minutes.

25 3. Detection – RCA

A RCA mixture was prepared consisting of:

2.66x Thermopol buffer (53.2 mM Tris-HCl, 26.6 mM (NH₄)₂SO₄, 26.6 mM KCl, 5.32 mM MgSO₄,
0.266% Triton-X, pH 8.8)

0.28 uM Primer mix

30 284.4 U/mL BST 2.0 WarmStart

14.67 U/mL TIPP

1.06 mM dNTPs

Syto82 dye 3uM

1.25 uL of reaction from point 2

Total volume 11.25uL

5

Primer mix:

Fwd (SEQ ID NO 34): 5'- T*C*GCAACATCCTATATCTGC-3'

Rev (SEQ ID NO 35): 5'- ATGTTGCGAAGGGCATATGT-3'

10 The resulting mixture was incubated at 50°C for 70 minutes.

Fluorescent readings were taken every 1 minute. The results can be seen in Fig.16.

Example 15: Further selected applications of the invention and embodiments**KRAS detection**

5 The KRAS gene controls cell proliferation, when it is mutated this negative signalling is disrupted and cells are able to continuously proliferate, often developing into cancer. A single amino acid substitution, and in particular a single nucleotide substitution, is responsible for an activating mutation implicated in various cancers: lung adenocarcinoma, mucinous adenoma, ductal carcinoma of the pancreas and colorectal cancer. KRAS mutations have been used as prognostic biomarkers, for example, in lung cancer.

10 Driver mutations in KRAS are associated with up to 20% of human cancers and there are targeted therapies in development against this mutation and its associated disease(s), a non-limiting list of some such therapies can be seen in the table below:

Name	Target	Manufacturer	Status	Brand Name(s)
AMG 510	KRAS ^{G12C}	Amgen	Trials	N/A
MRTX-849	KRAS ^{G12C}	Mirati Therapeutics	Trials	N/A
ARS-3248	KRAS ^{G12C}	Wellspring Biosciences, Inc/Janssen Biotech, Inc.	Trials	N/A

15 The presence of KRAS mutation has been found to reflect a very poor response to the EGFR inhibitors panitumumab (Vectibix) and cetuximab (Erbix). Activating mutations in the gene that encodes KRAS occurs in 30%-50% of colorectal cancers and studies show that patients whose tumours express this mutated version of the KRAS gene will not respond to panitumumab and cetuximab. The presence of the wild-type KRAS gene is not a guarantee that a patient will respond to these drugs, however, studies have shown that cetuximab has significant efficacy in metastatic colorectal cancer patients with wild-type KRAS tumours. Lung cancer patients who are positive for KRAS mutation (wild-type EGFR) have a response rate estimated at 5% or less for the EGFR antagonists erlotinib or gefitinib compared with a 60% response rate in patients who do not possess a KRAS mutation.

25 The early detection of the emergence of KRAS mutations (activating or over-expression), a frequent driver of acquired resistance to cetuximab therapy (anti-EGFR therapy) in colorectal cancers, allows the modification of treatment (for example the early initiation of a mitogen-activated protein kinase kinase [MEK] inhibitor) to delay or reverse resistance and thus it is advantageous that the methods of the current invention allow the quick and cheap detection of the KRAS status of patients.

30 A non-limiting list of mutations is: G12D, G12A, G12C, G13D, G12V, G12S, G12R, A59T/E/G, Q61H, Q61K, Q61R/L, K117N and A146P/T/V.

A further non-limiting list of mutations is shown in the table below:

Exon	Mutation name	COSM Number	Mutation sequence
2	G12A	COSM522	c.35G>C
2	G12C	COSM516	c.34G>T
2	G12D	COSM521	c.35G>A
2	G12F	COSM512	c.34_35delinsTT
2	G12R	COSM518	c.34G>C
2	G12S	COSM517	c.34G>A
2	G12V	COSM520	c.35G>T
2	G12V	COSM515	c.35_36delinsTC
2	G13A	COSM533	c.38G>C
2	G13C	COSM527	c.37G>T
2	G13D	COSM532	c.38G>A
2	G13R	COSM529	c.37G>C
2	G13S	COSM528	c.37G>A
3	Q61E	COSM550	c.181C>G
3	Q61H	COSM1146992	c.183A>T
3	Q61H	COSM554/COSM1135364	c.183A>C
3	Q61K	COSM549/COSM1159597	c.181C>A
3	Q61L	COSM553	c.182A>T
3	Q61R	COSM552	c.182A>G

BRAF detection

BRAF is a human gene that encodes for a protein called B-Raf which is involved in sending signals inside cells which are involved in directing cell growth. It has been shown to be mutated in some human cancers. B-Raf is a member of the Raf kinase family of growth signal transduction protein kinases and plays a role in regulating the MAP Kinase/ERKs signalling pathway, which affects, amongst other things, cell division.

Certain other inherited BRAF mutations cause birth defects.

More than 30 mutations of the BRAF gene have been identified that are associated with human cancers. In 90% of cases, thymine is substituted with adenine at nucleotide 1799. This leads to valine (V) being substituted for by glutamate (E) at codon 600 (now referred to as V600E) in the activation segment found in human cancers. This mutation has been widely observed in:

- Colorectal cancer
- Melanoma
- Papillary thyroid carcinoma
- Non-small cell lung cancer
- Ameloblastoma

A non-limiting list of other mutations which have been found are: R461I, I462S, G463E, G463V, G465A, G465E, G465V, G468A, G468E, N580S, E585K, D593V, F594L, G595R, L596V, T598I, V599D, V599E, V599K, V599R, V600K and A727V.

Drugs that treat cancers driven by BRAF mutations have been developed; vemurafenib and dabrafenib are approved by the FDA for the treatment of late-stage melanoma. The response rate to treatment with vemurafenib was 53% for metastatic melanoma, compared to 7-12% for the former best chemotherapeutic dacarbazine.

ERBB2/HER2 detection

Human epidermal growth factor receptor 2 (HER2), also known as CD340 (cluster of differentiation 340), proto-oncogene Neu, Erbb2 (rodent) or ERBB2 (human) is a protein encoded by the ERBB2 gene. Amplification or over-expression of this oncogene plays an important role in the progression of aggressive types of breast cancer. Over-expression of the ERBB2 gene is also known to occur in ovarian, stomach, adenocarcinoma of the lung, and aggressive forms of uterine cancer and 30% of salivary duct carcinomas. Structural alterations have also been identified that cause ligand-independent firing of the receptor in the absence of over-expression.

There are numerous targeted therapies approved and in development against this mutation and its associated disease(s), a non-limiting list of some such therapies can be seen in the table below:

Name	Target	Manufacturer	Status	Brand Name(s)
Trastuzumab	HER2 overexpression	Genentech	Approved	Herceptin

Pertuzumab	Dimerisation of HER2 and HER3 receptors	Genentech	Approved	Perjeta
Margetuximab	HER2 overexpression	Raven biotechnologies/MacroGenics	Trials	N/A
NeuVax	Vaccine	Galena Biopharma	Trials	N/A

HER2 testing is routinely performed in breast cancer patients to assess prognosis, monitor response to treatment and to determine suitability for targeted therapy (trastuzumab etc.). As trastuzumab is expensive and associated with serious side effects (cardiotoxicity) it is important that only HER2+ patients are selected to receive it and thus it is advantageous that the methods of the current invention allow the quick and cheap detection of the HER2 status of patients.

In one embodiment, the presence or absence of the ERBB2 Exon 20 insertion mutations is detected using the methods of the current invention.

A further non-limiting list of ERBB2 mutations is shown in the table below:

Exon	Mutation name	COSM Number	Mutation sequence
20	A775_G776insYVMA	COSM12558	c.2324_2325ins>TTACGTGATGGC
	G778_P780dup	COSM12556	c.2332_2340dup>GGCTCCCCA
	G776delinsVC	COSM1651739	c.2326_2327ins>TGT
	P780_Y781insGSP	COSM21607	c.2339_2340ins>TGGCTCCCC
	A775_G776insYVMA/Y772_A775dup	COSM20959	c.2313_2324dup>ATACGTGATGGC
17	V659E_AA	COSM3724566	c.1976_1977delinsTT>AA
	V659E_AG	COSM6503262	? c.1976_1977delinsTT>AG

EML4-ALK detection

EML4-ALK is an abnormal gene fusion of echinoderm microtubule-associated protein-like 4 (EML4) gene and anaplastic lymphoma kinase (ALK) gene. This gene fusion leads to the production of the protein EML4-ALK, which appears to promote and maintain the malignant behaviour of cancer cells. EML4-ALK positive lung cancer is a primary malignant lung tumour whose cells contain this mutation.

There are numerous targeted therapies approved and in development against this mutation and its associated disease(s), a non-limiting list of some such therapies can be seen in the table below:

Name	Generation	Target	Manufacturer	Status	Brand Name(s)
Crizotinib	1	EML4/ALK	Pfizer	FDA approved 2011	Xalkori, Crizalk
Certinib	2	ALK	Novartis	FDA approved 2014	Zykadia
Alectinib	2	ALK	Genentec	Japan 2014, FDA approved 2015	Alecensa
Brigatinib	2	ALK/ EGFR	Takeda	FDA approved 2017	Alunbrig, Briganix
Ensartinib	2	ALK	XCovery	Trial(s)	N/A
Lorlatinib	3	ROS1/ALK	Pfizer	Trial(s)	N/A

EML4-ALK gene fusions are responsible for approximately 5% of non-small cell lung cancers (NSCLC), with about 9,000 new cases in the US per year and about 45,000 worldwide.

- 5 There are numerous variants of EML4-ALK with all variants having the essential coiled-coil domain in the EML4 N-terminal portion and in the kinase domain of ALK exon 20 that are needed for transforming activity. Fusion of exon 13 of EML4 with exon 20 of ALK (variant 1: V1) (detection of which can be seen in Fig. 20), exon 20 of EML4 with exon 20 of ALK (V2), and exon 6 of EML4 with exon 20 of ALK (V3) are some of more common variants. The clinical significance of these different variants has only recently become clearer.
- 10 V3 has emerged as a marker suitable for the selection of patients who are likely to have shorter progression-free survival (PFS) after non-tyrosine kinase inhibitor (TKI) treatment such as chemotherapy and radiotherapy. There is further evidence that V3 is associated with shorter PFS of those patients who receive first- and second- generation treatment lines and worse overall survival (OS) compared to V1 and V2 of EML4-ALK.
- 15 It has also been found that V3-positive patients develop resistance to first and second treatment lines though the development of resistance mutations and possibly facilitated by incomplete tumour cell suppression due to a higher IC50 of wild-type V3. Detection of the unfavourable V3 could be used to select patients requiring more aggressive surveillance and treatment strategies. It appears that administration of the third generation Lorlatinib to patients with V3 may confer
- 20 longer PFS over those with V1 and thus it is advantageous that the methods of the current invention allow the quick and cheap detection of which variant a patient may have.

The methods of the current invention further allow the detection of resistance mutations such as, but not limited to: G1202R, G1269A, E1210K, D1203, S1206C, L1196M, F1174C, I1171T, I1171N/S, V1180L, T1151K and C1156Y.

- 25 G1202R, for example, is a solvent-front mutation which causes interference with drug binding and confers a high level of resistance to first- and second-generation ALK inhibitors. Thus it is advantageous that the methods of the current invention allow identification of those patients

who may possess this mutation and benefit from treatment initiation on a third generation treatment rather than a first or second.

A further non-limiting list of EML4-ALK mutations is shown in the table below:

Exon	Mutation	COSM Number	Mutation sequence
Rearrangements (always exon 20)	EML4-ALK	COSF408	E13_A20
		COSF474	E6ins33_A20
		COSF409	E20_A20
		COSF411	E6_A20

5 EGFR Detection

The identification of the epidermal growth factor receptor (EGFR) as an oncogene led to the development of targeted therapies such as gefitinib, erlotinib, afatinib, brigatinib and icotinib for lung cancer, and cetuximab for colon cancer. However, many people develop resistance to these therapies. Two primary sources of resistance are the T790M mutation and the MET oncogene.

- 10 EGFR mutations occur in EGFR exons 18–21 and mutations in exons 18, 19 and 21 and indicate suitability for treatment with EGFR-TKIs (tyrosine kinase inhibitors). Mutations in exon 20 (with the exception of a few mutations) show the tumours are EGFR-TKI resistant and not suitable for treatment with EGFR-TKIs.

- 15 The two most common EGFR mutations are short in-frame deletions of exon 19 and a point mutation (CTG to CGG) in exon 21 at nucleotide 2573, which results in substitution of leucine by arginine at codon 858 (L858R). Together, these two mutations account for ~90% of all *EGFR* mutations in non-small cell lung cancer (NSCLC). Screening for these mutations in patients with NSCLC can be used to predict which patients will respond to TKIs.

- 20 Thus it is advantageous that the methods of the current invention allow identification of those patients who may possess these mutations and benefit from treatment initiation on TKIs. The person skilled in the art will appreciate that the methods of the current invention allow identification of a range of EGFR mutations, a non-exhaustive list of such mutations is: G719X, Ex19Del, S768I, Ex20Ins and L861Q.

A further non-limiting list of mutations is shown in the table below:

Exon	Mutation name	COSM Number	Mutation sequence
Exon 18	G719A	6239	c.2156G>C
	G719S	6252	c.2155G>A
	G719C	6253	c.2155G>T
Exon 19	del_6210	6210	2240_2251del12
	del_6218	6218	2239_2247del9
	del_6220	6220	2238_2255del18

	del_6223	6223	2235_2249del15
	del_6225	6225	2236_2250del15
	del_6255	6255	2239_2256del18
	del_12367	12367	2237_2254del18
	del_12369	12369	2240_2254del15
	del_12370	12370	2240_2257del18
	del_12382	12382	2239_2248TTAAGAGAAG>C
	del_12383	12383	2239_2251>C
	del_12384	12384	2237_2255>T
	del_12385	12385	2235_2255>AAT
	del_12386	12386	2237_2252>T
	del_12387	12387	2239_2258>CA
	del_12403	12403	2239_2256>CAA
	del_12416	12416	2237_2253>TTGCT
	del_12419	12419	2238_2252>GCA
	del_12422	12422	2238_2248>GC
	del_12678	12678	2237_2251del15
	del_12728	12728	2236_2253del18
	del_13550	13550	2235_2248>AATTC
	del_13551	13551	2235_2252>AAT
	del_13552	13552	2235_2251>AATTC
	del_13556	13556	2253_2276del24
	del_18427	18427	2237_2257>TCT
	del_26038	26038	2233_2247del15
	del_6256	6256	? c.2254_2277del24
	del_28517	28517	? c.2235_2246del12
	del_1190791	1190791	? c.2234_2248del15
	del_26718	26718	? c.2250_2264del15
	del_133189	133189	? c.2236_2256del21
	del_24970	24970	? c.2239_2262del24
	I740_K745dup	26443	c.2217_2234dup
	K745_E746insVPVAIK	26444	c.2219_2236dup
Exon 20	T790M	6240	2369C>T
	S768I	6241	2303G>T
	p.A767_V769dup	12376	2307_2308ins9GCCAGCGTG
	p.H773dup	12377	2319_2320insCAC
	p.D770_N771insG	12378	2310_2311insGGT
	p.S768_D770dup	13428	2311_2312ins9GCGTGACACA

	p.A767_V769dup	13558	2309_2310AC>CCAGCGTGGAT
	A763_Y764insFQEA	26720	2284-5_2290dupTCCAGGAAGCCT
	C797S_2389	6493937	2389T>A
	C797S_2390	5945664	2390G>C
Exon 21	L858R	6224	c.2573T>G
	L858R	12429	2573_2574TG>GT
	L861Q	6213	c.2582T>A

ROS1

ROS1 is a receptor tyrosine kinase (encoded by the gene ROS1) with structural similarity to the anaplastic lymphoma kinase (ALK) protein; it is encoded by the c-ros oncogene.

- 5 A non-limiting list of ROS1 mutations is shown in the table below:

Exon	Mutation name	COSM Number	Mutation sequence
Rearrangements	CD74-ROS1	COSF1202	C6_R32
		COSF1200	C6_R34
		COSF1478	C6_R35
	SLC34A2-ROS1	COSF1196	S4_R32
		COSF1198	S4_R34
		COSF1259	S13del2046_R32
		COSF126	S13del2046_R34
	SDC4-ROS1	COSF1265	S2_R32
		COSF1278	S4_R32
		COSF1671	S2_R34
		COSF1280	S4_R34
	EZR-ROS1	COSF1267	E10_R34
	GOPC-ROS1	-	G4_R35

		COSF1188	G4_R36
		COSF1139	G8_R35
	LRIG3-ROS1	COSF1269	L16_R35
	TPM3-ROS1	COSF1273	T8_R35

RET proto-oncogene

The RET proto-oncogene encodes a receptor tyrosine kinase for members of the glial cell line-derived neurotrophic factor (GDNF) family of extracellular signalling molecules.

- 5 A non-limiting list of RET mutations is shown in the table below:

Exon	Mutation name	COSM Number	Mutation sequence
Rearrangements	KIF5B-RET	COSF1232	K15-R12
		COSF1230	K16-R12
		COSF1253	K22-R12
		COSF1234	K23-R12
		COSF1255 (actually 15_11del107)	K15-R11
		COSF1262	K24-R11
		COSF1242	K24-R8
			K24-R7
	CCDC6-RET	COSF1271	C1-R12
	NCOA4-RET	COSF1341	N6-R12
	TRIM33-RET		T14-R12

MET Exon 14

MET exon 14 skipping occurs with an approximately 5% frequency in NSCLC and is seen in both squamous and adenocarcinoma histology.

A non-limiting list of MET mutations is shown in the table below:

Exon	Mutation name	COSM Number	Mutation sequence
Skipping 14	MET-MET	COSM29312	M13_M15

5

NTRK proto-oncogenes

NTRK gene fusions lead to abnormal proteins called TRK fusion proteins, which may cause cancer cells to grow. NTRK gene fusions may be found in some types of cancer, including cancers of the brain, head and neck, thyroid, soft tissue, lung, and colon. Also called neurotrophic tyrosine receptor kinase gene fusion.

10

A non-limiting list of NTRK mutations is shown in the table below:

Exon	Mutation name	COSM Number	Mutation sequence
NTRK1 Rearrangements	CD74-NTRK1		C7-N10
	MPRIIP-NTRK1		M22-N12
	TFG-NTRK1	COSF1323	T5-N10
	TPM3-NTRK1	COSF1329	T8-N10
NTRK3 Rearrangements	ETV6-NTRK3	COSF571	E5-N15
	ETV6-NTRK3	COSF1534	E4-N14
	ETV6-NTRK3	COSF823	E4-N15

15

Panels

In one embodiment of the invention, there is provided a panel comprising a plurality of probe molecules (A_0) wherein each A_0 is complementary to a target mutation. The mutation may be selected from any mutation previously, or subsequently, described or known. The person skilled in the art will thus appreciate that within the scope of the invention are included panels which may be useful in the detection of one or more mutations to the any of the proto-oncogenes or oncogenes previously, or subsequently, described or known.

In one embodiment, the panel comprises 5-500 individual probe molecules, each complementary to a specific target mutation. In one embodiment, the panel comprises 5-400 individual probe molecules, each complementary to a specific target mutation. In one embodiment, the panel comprises 5-300 individual probe molecules, each complementary to a specific target mutation. In one embodiment, the panel comprises 5-200 individual probe molecules, each complementary to a specific target mutation. In one embodiment, the panel comprises 5-100 individual probe molecules, each complementary to a specific target mutation. In one embodiment, the panel comprises 5-50 individual probe molecules, each complementary to a specific target mutation.

In one embodiment, there may be a plurality of probe molecules specific to the same mutation. In one embodiment, there may be only one probe molecule specific to each mutation of the panel.

In one embodiment, there is provided a panel, wherein the panel comprises a plurality of probe molecules wherein one or more probes are complementary to an EGFR mutation, one or more probes are complementary to a KRAS mutation, one or more probes are complementary to a ERBB2/HER2 mutation, one or more probes are complementary to a EML4-ALK mutation, one or more probes are complementary to a ROS1 mutation, one or more probes are complementary to a RET mutation and one or more probes are complementary to a MET mutation.

In one embodiment, there is provided a panel, wherein the panel comprises a plurality of probe molecules wherein one or more probes may be complementary to an EGFR mutation, one or more probes may be complementary to a KRAS mutation, one or more probes may be complementary to a ERBB2/HER2 mutation, one or more probes may be complementary to a EML4-ALK mutation, one or more probes may be complementary to a ROS1 mutation, one or more probes may be complementary to a RET mutation and one or more probes may be complementary to a MET mutation.

In one embodiment, there is provided a panel of probes selective for one or more EGFR, KRAS, BRAF, ERBB2/HER2, EML4-ALK, ROS1, RET, MET mutations.

In one embodiment, there is provided a panel of probe molecules selective for EGFR mutations.

In one embodiment, there is provided a panel of probe molecules selective for KRAS mutations.

In one embodiment, there is provided a panel of probe molecules selective for BRAF mutations.

In one embodiment, there is provided a panel of probe molecules selective for ERBB2/HER2 mutations.

In one embodiment, there is provided a panel of probe molecules selective for EML4-ALK mutations.

- 5 In one embodiment, there is provided a panel of probe molecules selective for ROS1 mutations.

In one embodiment, there is provided a panel of probe molecules selective for RET mutations.

In one embodiment, there is provided a panel of probe molecules selective for NTRK mutations.

In one embodiment, there is provided a panel of probe molecules selective for ROS1 mutations.

- 10 In one embodiment, there is provided a panel of probe molecules selective for MET exon 14 mutations.

In one embodiment, there is provided a panel comprising a plurality of probe molecules selective for one or more coding sequences (CDSs).

In one embodiment, there is provided a method of detecting one or more mutations using one or more of the previously described panels.

- 15 In one embodiment, there is provided a method of detecting the presence or absence of one or more mutations using one or more of the previously described panels.

In one embodiment, there is provided a kit comprising a panel, which may be as previously or subsequently described, in combination with one or more reagents, which may be as previously or subsequently described.

- 20 The person skilled in the art will appreciate that embodiments of kits that disclose A₀, include within their scope embodiments wherein there is a panel comprising a plurality of A₀.

In one embodiment, there is provided a device wherein one or more regions of the device comprise one or more panels, which may be as previously, or subsequently, described.

- 25 **Companion diagnostics**

The methods of the present invention can be used to detect specific genetic markers in a sample which may be used to help guide the selection of appropriate therapy. These markers may be tumour-specific mutations, or may be wild-type genomic sequences, and may be detected using tissue, blood or any other patient sample type.

30

Resistance monitoring

Repeated testing of patient samples during treatment of disease may allow early detection of developed resistance to therapy. As an example of this application is in non-small cell lung carcinoma (NSCLC), in which epidermal growth factor receptor (EGFR) inhibitors (e.g. gefitinib, erlotinib) are commonly used as first line treatments. During treatment the tumour can often develop mutations in the EGFR gene (e.g. T790M, C797S) which confer resistance to the drug.

35

Early detection of these mutations may allow for transfer of the patient onto alternative therapies (e.g. Tagrisso).

5 Typically patients being monitored for resistance onset can be too sick for repeated tissue biopsy to be carried out. Repeated tissue biopsy may also be expensive, invasive and carries associated risks. It is preferable to test from blood, but there may be very low copy numbers of the mutations of interest in a reasonable blood drawn sample. Monitoring therefore requires sensitive testing from blood samples using a method of the present invention in which the method is simple and cost effective to carry out such that it can be regularly performed.

10

Recurrence monitoring

In this application example, patients who have been declared free of disease following treatment may be monitored over time to detect the recurrence of disease. This needs to be done non-invasively and requires sensitive detection of target sequences from blood samples. By using the method of the present invention, it provides a simple and low-cost method that can be regularly performed. The sequences targeted may be generic mutations known to be common in the disease of interest, or can be custom panels of targets designed for a specific patient based on detection of variants in the tumour tissue prior to remission.

15

Minimal Residual Disease (MRD) monitoring

For some cancers there are residual cancer cells that remain in a patient after treatment, it is a major cause of relapse in cancer and leukaemia. MRD monitoring and testing has several important roles: determining whether treatment has eradicated the cancer or whether traces remain, comparing the efficacy of different treatments, monitoring patient remission status as well as detecting recurrence of leukaemia, and choosing the treatment that will best meet those needs.

20

Screening

Population screening for early detection of disease has been a long-held goal, particularly in cancer diagnostics. The challenge is two-fold: the identification of panels of markers which allow confident detection of disease without too many false negatives, and the development of a method with sufficient sensitivity and low enough cost. The methods of the present invention could be used to address larger panels of mutations than PCR-based tests but with a much simpler workflow and lower cost than sequencing-based diagnostics.

25

30

35

Organ transplant rejection

When a transplanted organ is rejected by the recipient, the DNA from this organ is shed into the recipient's bloodstream. Early detection of this DNA would allow early detection of rejection. This could be achieved using custom panels of donor-specific markers, or by using panels of variants known to be common in the population, some of which will be present in the donor and some in the recipient. Routine monitoring of organ recipients over time could be enabled by the low cost and simple workflow of the present invention disclosed herein.

40

Non-invasive prenatal testing (NIPT)

45 It has long been known that fetal DNA is present in maternal blood, and the NIPT market is now quite saturated with companies using sequencing to identify mutations and count copy numbers of specific chromosomes to enable detection of fetal abnormalities. The methods of the present invention as disclosed herein have the ability to detect mutations at very low allele fractions, potentially allowing earlier detection of fetal DNA. Identification of common mutations in a given

population would allow assays to be developed that target mutations that may be present in either the maternal or fetal DNA or to allow detection of abnormalities at an earlier stage of pregnancy.

- 5 Various further aspects and embodiments of the present invention will be apparent to those skilled in the art in view of the present disclosure.

“and/or” where used herein is to be taken as specific disclosure of each of the two specified features or components with or without the other. For example “A and/or B” is to be taken as
10 specific disclosure of each of (i) A, (ii) B and (iii) A and B, just as if each is set out individually herein.

Unless context dictates otherwise, the descriptions and definitions of the features set out above are not limited to any particular aspect or embodiment of the invention and apply equally to all
15 aspects and embodiments which are described.

It will further be appreciated by those skilled in the art that although the invention has been described by way of example with reference to several embodiments, it is not limited to the disclosed embodiments and that alternative embodiments could be constructed without
20 departing from the scope of the invention as defined in the appended claims.

The person skilled in the art will understand that references to ‘partially digested strand A_1 ’ may refer to the single-stranded oligonucleotide formed by progressive digestion of A_0 when hybridised to a target analyte sequence, in the 3’-5’ direction until the strands dissociate due to
25 lack of complementarity.

The person skilled in the art will understand that references to ‘partially double-stranded’ nucleic acids may refer to nucleic acids wherein one or more portions are double-stranded and one or more portions are single-stranded.

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The person skilled in the art will understand that references to ‘substantially double-stranded’ nucleic acids may refer to nucleic acids wherein one or more portions are double-stranded and one or more smaller portions are single-stranded.

Claims:

1. A kit comprising:
 - (a) a single-stranded probe oligonucleotide A_0 , capable of forming a first intermediate product with a target polynucleotide sequence, said intermediate product being at least partially double-stranded;
 - (b) a ligase;
 - (c) a pyrophosphorolysing enzyme capable of digesting the first intermediate product in the 3'-5' direction from the end of A_0 to create a partially digested strand A_1 ;
 - (d) a source of ions to drive the pyrophosphorolysis reaction, wherein optionally the ions are pyrophosphate ions; and
 - (e) suitable buffers.
2. A kit as claimed in claim 1, further comprising positive and negative controls.
3. A kit as claimed in any one preceding claim, wherein the 5' end of A_0 is rendered resistant to 5'-3' exonuclease digestion and wherein the kit further comprises a 5'-3' exonuclease.
4. A kit as claimed in any one of claims 1 to 3, wherein the kit further comprises deoxynucleotide triphosphates (dNTPs), a polymerase and buffers for the initial amplification of a target polynucleotide sequence present in a sample.
5. A kit as claimed in claim 4, wherein the kit further comprises a dUTP incorporating high fidelity polymerase, dUTPs and uracil-DNA N-glycosylase (UDG).
6. A kit as claimed in any one of claims 1 to 5, wherein the kit further comprises a proteinase.
7. A kit as claimed in any one of claims 1 to 6, further comprising a ligation probe oligonucleotide C.
8. A kit as claimed in any one of claims 1 to 6, further comprising a splint oligonucleotide D.
9. A kit as claimed any one of claims 1 to 6, further comprising a ligation probe oligonucleotide C and a splint oligonucleotide D.
10. A kit as claimed in claim 7 or 9, wherein the ligation probe oligonucleotide C comprises a 3' or internal modification protecting it from 3'-5' exonuclease digestion and the kit further comprises a 3'-5' exonuclease.

11. A kit as claimed in claim 8, 9 or 10, wherein D comprises an oligonucleotide region complementary to the 3' end of A₁ and a region complementary to either the 5' end of oligonucleotide C or to the 5' end of A₁.
12. A kit as claimed in claim 11, wherein D is unable to undergo extension against A₁ by virtue of either a 3' modification or through a mismatch between the 3' end of D and the corresponding region of A₁ or C.
13. A kit as claimed in any one preceding claim, wherein the kit further comprises at least one single-stranded primer oligonucleotide that is substantially complementary to a portion of A₀, an amplification enzyme, dNTPs and one or more oligonucleotide binding dyes or molecular probes.
14. A kit as claimed in any one preceding claim, wherein the kit further comprises multiple A₀, each selective for a different target sequence and each including an identification region.
15. A kit as claimed in any one of claims 1 to 12, wherein the kit further comprises:
- two or more Ligation Chain Reaction (LCR) probe oligonucleotides that are complementary to adjacent sequences on A₁ wherein when the probes are successfully annealed the 5' phosphate of one LCR probe is directly adjacent to the 3'OH of the other LCR probe; and
 - one or more ligases;
16. A kit as claimed in claim 13, wherein the amplification enzyme is the same as the pyrophosphorolysis enzyme.
17. A kit as claimed in any one of claims 1 to 12, wherein the kit further comprises
- A ligation probe oligonucleotide C;
 - A splint oligonucleotide D;
- wherein C has a 5' phosphate, the 3' end of a splint oligonucleotide D is complementary to the 5' end of C and the 5' end of D is complementary to the 3' end of A₁ such that A₁ and C are capable of being ligated together to form A₂.
18. A kit as claimed in claim 17, wherein the kit further comprises:
- A hairpin oligonucleotide 1 (HO1) comprising a fluorophore-quencher pair, wherein HO1 is complementary to A₂ and when annealed to A₂ the hairpin structure of HO1 opens and the fluorophore-quencher pair separate; and
 - A hairpin oligonucleotide 2 (HO2) comprising a fluorophore-quencher pair, wherein HO2 is complementary to the open HO1 and when annealed to HO1 the hairpin structure of HO2 opens and the fluorophore-quencher pair separate.

19. A kit as claimed in claim 18, wherein the kit further comprises a plurality of HO1 and HO2.
20. A kit as claimed in any one of claims 1 to 12, wherein the kit further comprises:
- an oligonucleotide A comprising a substrate arm, a partial catalytic core and a sensor arm;
 - an oligonucleotide B comprising a substrate arm, a partial catalytic core and a sensor arm; and
 - a substrate comprising a fluorophore quencher pair;
- wherein the sensor arms of oligonucleotides A and B are complementary to flanking regions of A₂ such that in the presence of A₂, oligonucleotides A and B are combined to form a catalytically, multicomponent nucleic acid enzyme (MNAzyme).
21. A kit as claimed in any one of claims 1 to 12, wherein the kit further comprises a partially double-stranded nucleic acid construct wherein:
- one strand comprises at least one RNA base, at least one fluorophore and wherein a region of this strand is complementary to a region of A₂ and wherein this strand is the 'substrate' strand; and
 - the other strand comprises at least one quencher and wherein a region of this strand is complementary to a region of A₂ adjacent to that which the substrate strand is complementary to, such that in the presence of A₂ the partially double stranded nucleic acid construct has a greater double-stranded portion
22. A kit as claimed in claim 21, wherein the kit further comprises an enzyme for removal of the at least one RNA base.
23. A kit as claimed in any one of claims 1 to 12, wherein the kit further comprises:
- an oligonucleotide complementary to a region of A₂ including the site of ligation, comprising one or multiple fluorophores arranged such that their fluorescence is quenched either by their proximity to each other or to one or more fluorescence quenchers;
 - a double strand specific DNA digestion enzyme;
- wherein, in the presence of A₂, the labelled oligonucleotide is digested such that the fluorophores are separated from each other or from their corresponding quenchers, and a fluorescent signal, and hence the presence of A₂, is detectable.
24. A kit as claimed in any one of claims 1 to 23, wherein the kit further comprises a phosphatase or a phosphohydrolase.
25. A kit as claimed in any one of claims 1 to 24, wherein the kit further comprises a pyrophosphatase.

26. A kit as claimed in any one of claims 1 to 25, wherein the kit further comprises an enzyme for the formation of DNA from an RNA template.

27. A device comprising:

- 5 a fluid pathway between a first region and second region, wherein the first region comprises one or more wells, wherein one or more well comprises:
- a single-stranded probe oligonucleotide A_0 , which is capable of forming a first intermediate product with a target polynucleotide sequence, said intermediate product being at least partially double-stranded;
- 10 a pyrophosphorolysing enzyme capable of digesting the first intermediate product in the 3'-5' direction from the end of A_0 to create a partially digested strand A_1 ; a source of ions to drive the pyrophosphorolysis reaction forward, wherein optionally the ions are pyrophosphate ions;
- and
- one or more ligases capable of ligating A_1 to create an oligonucleotide A_2 ;
- 15 wherein the second region comprises one or more wells.

28. The device of claim 27, wherein the 5' end of A_0 is resistant to 5'-3' exonuclease digestion and wherein the wells of the first region further comprise a 5'-3' exonuclease.

- 20 29. The device of any one of claims 27 or 28, wherein the device further comprises a third region comprising one or more wells which is joined to the first region by a fluid pathway and wherein one or more wells of the third region comprises:
- dNTPs;
- at least one single-stranded primer oligonucleotide; and
- 25 an amplification enzyme.

30. The device of claim 29 wherein:

- the dNTPs of the third region are dUTP, dGTP, dCTP and dATP;
- the amplification enzyme is a dUTP incorporating high fidelity polymerase; and
- 30 one or more wells of the third region further comprises uracil-DNA N-glycosylase.

31. The device of claim 29 or claim 30 wherein the device further comprises a fourth region, located between the first and third regions, comprising one or more wells, wherein one or more wells comprises a proteinase.
- 5 32. The device of any one of claims 27 to 31, wherein one or more wells of the first or second regions further comprises a ligase and a ligation probe oligonucleotide C.
33. The device of any one of claims 27 to 31, wherein one or more wells of the first or second regions further comprises a ligase and a splint oligonucleotide D which is
10 complementary to a region of A_0 .
34. The device of any one of claims 27 to 31, wherein one or more wells of the first or second region further comprises a ligase, a splint oligonucleotide D and a ligation probe oligonucleotide C.
15
35. The device of claim 32 or claim 34, wherein the ligation probe oligonucleotide C comprises a 3' or internal modification protecting it from 3'-5' exonuclease digestion.
36. The device of any one of claims 33, 34 or 35, wherein D comprises an oligonucleotide
20 region complementary to the 3' end of A_1 and a region complementary to either the 5' end of oligonucleotide C or to the 5' end of A_1 .
37. The device of any one of claims 27 to 36, wherein one or more wells of the first region comprises at least one or more different A_0 , each selective for a different target
25 sequence.
38. The device of claim 37, wherein the wells of the second region comprise:
dNTPs;
buffers;
an amplification enzyme;
30 one or more oligonucleotide binding dyes or molecular probes; and
a means for detecting a signal derived from A_1 or a portion thereof, or multiple copies of A_1 or multiple copies of a portion thereof.

39. The device of any one of claims 27 to 36, wherein the wells of the second region further comprise:

- two or more Ligation Chain Reaction (LCR) probe oligonucleotides that are complementary to adjacent sequences on A_1 wherein when the probes are successfully annealed the 5' phosphate of one LCR probe is directly adjacent to the 3'OH of the other LCR probe; and
- one or more ligases.

40. The device of any one of claims 27 to 36, wherein the wells of the second region further comprise:

- A ligation probe oligonucleotide C;
- A splint oligonucleotide D;

wherein C has a 5' phosphate, the 3' end of a splint oligonucleotide D is complementary to the 5' end of C and the 5' end of D is complementary to the 3' end of A_1 such that A_1 and C are capable of being ligated together to form an oligonucleotide A_2 .

41. The device of claim 40, wherein the wells of the second region further comprise:

- A hairpin oligonucleotide 1 (HO1) comprising a fluorophore-quencher pair, wherein HO1 is complementary to A_2 and when annealed to A_2 the hairpin structure of HO1 opens and the fluorophore-quencher pair separate; and
- A hairpin oligonucleotide 2 (HO2) comprising a fluorophore-quencher pair, wherein HO2 is complementary to the open HO1 and when annealed to HO1 the hairpin structure of HO2 opens and the fluorophore-quencher pair separate.

42. The device of any one of claims 27 to 36, wherein the wells of the second region further comprise:

- an oligonucleotide A comprising a substrate arm, a partial catalytic core and a sensor arm;
- an oligonucleotide B comprising a substrate arm, a partial catalytic core and a sensor arm; and
- a substrate comprising a fluorophore quencher pair;

wherein the sensor arms of oligonucleotides A and B are complementary to flanking regions of A_2 such that in the presence of A_2 , oligonucleotides A and B are combined to form a catalytically, multicomponent nucleic acid enzyme (MNAzyme).

5 43. The device of any one of claims 27 to 36, wherein the wells of the second region further comprise a partially double-stranded nucleic acid construct wherein:

- 10 - one strand comprises at least one RNA base, at least one fluorophore and wherein a region of this strand is complementary to a region of A_2 and wherein this strand may be referred to as the 'substrate' strand;
- 15 - the other strand comprises at least one quencher and wherein a region of this strand is complementary to a region of A_2 adjacent to that which the substrate strand is complementary to, such that in the presence of A_2 the partially stranded nucleic acid construct becomes substantially more double-stranded; and

wherein the wells of the second region further comprise an enzyme for removal of the at least one RNA base.

20 44. The device of any one of claims 27 to 36 wherein one or more wells of the second region further comprises:

an oligonucleotide complementary to a region of A_2 including the site of ligation, comprising one or multiple fluorophores arranged such that their fluorescence is quenched either by their proximity to each other or to one or more fluorescence quenchers;
a double strand specific DNA digestion enzyme;

25 wherein, in the presence of A_2 , the labelled oligonucleotide is digested such that the fluorophores are separated from each other or from their corresponding quenchers, and a fluorescent signal, and hence the presence of A_2 , is detectable.

30 45. A device as claimed in any one of claims 27 to 44 wherein one or more wells of one or more regions further comprises a pyrophosphatase.

46. A device as claimed in any one of claims 27 to 45 wherein one or more wells of one or more regions further comprises a phosphatase or a phosphohydrolase.

47. A device as claimed in any one of claims 27 to 46 wherein one or more wells of the first region further comprises an enzyme for the formation of DNA from an RNA template.

48. A device as claimed in any one of claims 27 to 47 wherein the first and second regions are combined.