

enzyme and an inhibitor to the reaction, together with reagents for detecting interference of the oligonucleotide probe with the inhibitor, as an indication of the presence of a particular target nucleic acid sequence. The kit may further contain the reagents for performing the methods of this invention.

[0015] Accordingly, it is an object of the invention to not encompass within the invention any previously known product, process of making the product, or method of using the product such that Applicants reserve the right and hereby disclose a disclaimer of any previously known product, process, or method. It is further noted that the invention does not intend to encompass within the scope of the invention any product, process, or making of the product or method of using the product, which does not meet the written description and enablement requirements of the USPTO (35 U.S.C. §112, first paragraph) or the EPO (Article 83 of the EPC), such that Applicants reserve the right and hereby disclose a disclaimer of any previously described product, process of making the product, or method of using the product.

[0016] It is noted that in this disclosure and particularly in the claims and/or paragraphs, terms such as “comprises”, “comprised”, “comprising” and the like can have the meaning attributed to it in U.S. Patent law; e.g., they can mean “includes”, “included”, “including”, and the like; and that terms such as “consisting essentially of” and “consists essentially of” have the meaning ascribed to them in U.S. Patent law, e.g., they allow for elements not explicitly recited, but exclude elements that are found in the prior art or that affect a basic or novel characteristic of the invention.

[0017] These and other embodiments are disclosed or are obvious from and encompassed by, the following Detailed Description.

BRIEF DESCRIPTION OF THE DRAWINGS

[0018] The following detailed description, given by way of example, ~~but not intended to limit the invention solely to the specific embodiments described,~~ may best be understood in conjunction with the accompanying drawings.

[0019] FIGS. 1A and 1B are schematics showing the reaction in use to detect a target nucleic acid, wherein the detector is conjugated to a fluorophore and the inhibitor is conjugated to a quencher. FIG. 1A depicts an oligonucleotide probe that matches the target nucleic acid acting as a primer for DNA polymerization. The resulting double stranded DNA blocks the binding site

human or animal and includes, ~~for example but is not limited to~~, genomic DNA, PCR amplicon, cDNA, and others. The target nucleic acid may be double stranded or single stranded. In one example, the single stranded target nucleic acid is DNA. In one embodiment, a double stranded target nucleic acid is first converted to a single stranded target nucleic acid. In yet another embodiment, PCR is performed on the target nucleic acid prior to detection. In one aspect of this embodiment, the PCR product is subsequently converted to single stranded form. Methods for converting a double stranded nucleic acid into a single stranded nucleic acid are known in the art and are described, for example, by Mitsis et al., "Characterization of the interaction of lambda exonuclease with the ends of DNA", *Nucleic Acids Res* (1999) vol. 27, no. 15, pp. 3057-3063; Sanchez, et al., "Linear-After-The-Exponential (LATE)-PCR: An advanced method of asymmetric PCR and its uses in quantitative real-time analysis", *PNAS* (2004) vol. 101, no. 7, pp. 1933-1938; Chen, et al., "Asynchronous PCR", In: D. Park, ed. 2011. *PCR Protocols* (Methods in Molecular Biology, vol. 687). New Jersey: Humana Press Inc., pp. 231-243.

[0031] The methods of the invention may be used to detect the presence or absence of a nucleic acid sequence within a target nucleic acid. In one embodiment, a single nucleotide within the target nucleic acid sequence is detected. In one aspect of this embodiment, a particular locus may be queried to detect the presence or absence of a particular nucleic acid sequence variance. A "variance" is a difference in the nucleotide sequence among related polynucleotides. The difference may be the deletion of one or more nucleotides from the sequence of one polynucleotide compared to the sequence of a related polynucleotide, the addition of one or more nucleotides or the substitution of one nucleotide for another. The terms "mutation," "polymorphism" and "variance" are used interchangeably herein. As used herein, the term "variance" in the singular is to be construed to include multiple variances, i.e., two or more nucleotide additions, deletions and/or substitutions in the same polynucleotide. A "point mutation" refers to a single substitution of one nucleotide for another.

[0032] For example, a particular locus may be queried to detect the presence or absence of a single nucleotide polymorphism. A "single nucleotide polymorphism" or "SNP" refers to a variation in the nucleotide sequence of a polynucleotide that differs from another polynucleotide by a single nucleotide difference. A SNP included, for example ~~and without limitation~~, exchanging one A for one C, G or T, or one C for one G, T or C and so on, in the entire sequence of polynucleotide. Additionally, it is possible to have more than one SNP in a particular nucleic

Hybridization Probes”, In: V. Didenko, ed. 2006. *Fluorescent Energy Transfer Nucleic Acid Probes: Designs and Protocols* (Methods in Molecular Biology, vol. 335). New Jersey: Humana Press Inc., pp.3-16. Examples of fluorophores that can be employed in the present invention include, ~~but are not limited to,~~ those described by Marras 2006 and further described herein below. The particular location of the fluorophore in relation to the detector is not critical to the present invention. The fluorophore can be attached anywhere along the detector, including the 5' end, the 3' end or anywhere internally along the detector.

[0036] The methods of the invention may further comprise the use of an inhibitor. The inhibitor is an oligonucleotide that is analogous to, and hybridizes with, the detector. The inhibitor functions to allow a signal to be detected only if an oligonucleotide probe matches the target nucleic acid. Hybridization of the inhibitor to the detector takes place in standard reaction buffer, for example, in a DNA polymerase reaction buffer whereby the detector and the inhibitor are mixed in the buffer at the appropriate temperature. In one example, the reaction may be heated to 95°C for a period of 30 seconds and then chilled to 5°C below the annealing temperature of the inhibitor.

[0037] In one embodiment, the inhibitor is conjugated to a quencher. The quencher is a molecule that functions to decrease, i.e., quench the intensity of the fluorescence by transferring energy from a first fluorophore to a second fluorophore or to a non-fluorescent molecule. The particular quencher employed is not critical to the present invention. Quenchers are known in the art and are described, for example by, Marras 2006. Examples of quenchers that can be employed in the present invention include, ~~but are not limited to,~~ those described by Marras 2006 and further described herein below. The particular location of the quencher in relation to the inhibitor is not critical to the present invention. The quencher can be attached anywhere along the inhibitor, including the 5' end, the 3' end or anywhere internally along the inhibitor.

[0038] In an alternative embodiment, the detector is conjugated to a quencher. The particular quencher employed is not critical to the present invention. Examples of quenchers that can be employed in the present invention include, ~~but are not limited to,~~ those described by Marras 2006 and further described herein below. The particular location of the quencher in relation to the detector is not critical to the present invention. The quencher can be attached anywhere along the detector, including the 5' end, the 3' end or anywhere internally along the detector.

[0039] In an alternative embodiment, the inhibitor is conjugated to a fluorophore. The particular fluorophore employed is not critical to the present invention. Examples of fluorophores that can be employed in the present invention include, ~~but are not limited to,~~ those described by Marras 2006 and further described herein below. The location of the fluorophore in relation to the inhibitor is not critical to the present invention. The fluorophore can be attached anywhere along the inhibitor, including the 5' end, the 3' end or anywhere internally along the inhibitor.

[0040] In another embodiment of the methods of this invention, the inhibitor is conjugated to biotin ("inhibitor-biotin conjugate"). Inhibitor-biotin conjugates can be obtained commercially from various vendors (e.g., Integrated DNA Technologies, Inc., Eurofins MWG Operon, Eurogentec, TriLink BioTechnologies, Inc.). Examples of commercially available purification kits include Agencourt®-AMPure® XP (Beckman Coulter, Inc.), and QIAquick 96 PCR Purification kit (Qiagen). The particular location of the biotin in relation to the inhibitor is not critical to the present invention. The biotin can be attached anywhere along the inhibitor, including the 5' end, the 3' end or anywhere internally along the inhibitor.

[0041] In another embodiment of the methods of this invention, the detector or the inhibitor is conjugated to a molecule that will facilitate isolation of the detector or the inhibitor, respectively, from the reaction. The molecule that will facilitate this isolation may be, for example, an epitope, which is anything capable of reacting with an antibody. The epitope may be, for example, an antigen, peptide or protein. In one example, the epitope is biotin.

[0042] In another embodiment of the methods of this invention, the detector or the inhibitor is conjugated to a molecule that will facilitate detection of the detector or the inhibitor, respectively, within the reaction. The molecule that will facilitate this detection may be, for example, an epitope. The epitope may be, for example, an antigen, peptide or protein. In one example, the epitope is biotin. In another example, the epitope is a peptide that can be detected using a chemiluminescent assay, e.g., by using secondary antibodies conjugated to the HRP enzyme.

[0043] In the above embodiments wherein the detector or the inhibitor is conjugated to biotin, the conjugation can be achieved by PCR amplification of the detector or the inhibitor with a biotin conjugated primer (commercially available from Integrated DNA Technologies, Inc., Operon, Eurogentec, and TriLink BioTechnologies, Inc.). In the above embodiments wherein

the epitope is detected using antibodies conjugated to the HRP enzymes, conjugation kits may be obtained commercially from Solulink or the antibody-HRP enzyme conjugates may be ordered directly from commercial vendors, e.g., Eurogentec).

[0044] In one embodiment of the methods of this invention, when the detector is conjugated to a fluorophore, the inhibitor is conjugated to a quencher. In another embodiment of the methods of this invention, when the detector is conjugated to a quencher, the inhibitor is conjugated to a fluorophore. The efficiency of energy transfer between the detector and the inhibitor or the inhibitor and the detector, respectively, is specific to the fluorophore-quencher pair chosen and should be optimized accordingly, as described by Marras 2006 and product literature provided by known commercial vendors listed herein below.

[0045] In another embodiment of the methods of this invention, the detector is conjugated to a first fluorophore and the inhibitor is conjugated to a second fluorophore, such that emission from the first fluorophore will excite the second fluorophore by energy transfer, and wherein the transferred energy is emitted as fluorescence characteristic of the second fluorophore. This quenching phenomenon is known in the art and described, for example, by Marras 2006. Fluorophore-fluorophore pairs are also known in the art and are described, for example by, Marras 2006. Examples of fluorophore-fluorophore pairs that can be employed in the present invention include, ~~but are not limited to,~~ those described by Marras 2006 and further described herein below. The particular location of the first fluorophore in relation to the detector and the second fluorophore in relation to the inhibitor is not critical to the present invention. The first fluorophore can be attached anywhere along the detector, including the 5' end, the 3' end or anywhere internally along the detector. The second fluorophore can be attached anywhere along the inhibitor, including the 5' end, the 3' end or anywhere internally along the inhibitor. The efficiency of energy transfer between the detector and inhibitor is specific to the fluorophore-fluorophore pairs chosen and should be optimized accordingly.

[0046] The fluorophore and quencher of a fluorophore-quencher pair or the first fluorophore and second fluorophore of a fluorophore-fluorophore pair (referred to individually and collectively as "FRET pair(s)") may be placed anywhere in relation to each other as long as the distance ("FRET distance") does not fall beyond the efficient energy transfer distance of the particular FRET pair. The importance of FRET distance is known in the art and is described, for

example, by Marras 2006. The efficiency of energy transfer between the detector and the inhibitor is specific to the FRET pair chosen and should be optimized accordingly.

[0047] As illustrated in FIG. 3, the fluorophore can be located anywhere on the detector and the quencher can be located anywhere on the inhibitor, as long as they are within FRET distance of each other. Alternatively, as illustrated in FIG. 4, the quencher can be located anywhere on the detector and the fluorophore can be located anywhere on the inhibitor as long as they are within FRET distance of each other. Thus, the fluorophore and the quencher must be within an acceptable FRET distance to each other that allows the reaction to occur. This FRET distance is particular to the fluorophore-quencher pair chosen and each fluorophore and quencher can be on adjacent nucleotides, tens of nucleotides apart or even hundreds of nucleotides apart, depending on the particular FRET pair chosen.

[0048] In one example, the fluorophore is on the 5' end of the detector and the quencher is on the 3' end of the inhibitor. In another example, the quencher is on the 5' end of the detector and the fluorophore is on the 3' end of the inhibitor.

[0049] As illustrated in FIG. 5, the first fluorophore can be located anywhere on the detector and the second fluorophore can be located anywhere on the inhibitor, as long as they are within FRET distance of each other. Thus, the first fluorophore and the second fluorophore must be within an acceptable distance to each other that allows the reaction to occur. This FRET distance is particular to the fluorophore-fluorophore pair and each fluorophore can be on adjacent nucleotides, tens of nucleotides apart or even hundreds of nucleotides apart, depending on the particular FRET pair chosen.

[0050] In one example, the detector is conjugated to a FAM fluorophore on its 5' end and the inhibitor is conjugated to a TAMRA fluorophore positioned approximately 15nt away from the FAM fluorophore.

[0051] The selection of a particular fluorophore-quencher or fluorophore-fluorophore pair is not critical. Examples of the categories of fluorophores that can be employed in the present invention include, ~~but are not limited to~~ adjacent probes (e.g., LightCycler® hybridization probes, available from Sigma-Aldrich®), 5'-nuclease probes (or TaqMan® probes, available from PREMIER Biosoft®), minor groove binder (Taqman® MGB probes, available from Applied Biosystems®) probes, molecular beacon probes, Scorpions® primers (available from PREMIER Biosoft and Biosearch Technologies), and strand-displacement probes (or Yin-Yang probes).

[0052] Examples of the specific fluorophores that may be employed in the present invention include, ~~but are not limited to~~ fluorescein and derivatives thereof (e.g., fluorescein isothianate (FITC), carboxyfluorescein (FAM), tetrachlorofluorescein (TET), 2',7'-difluorofluorescein (Oregon Green® 488), Oregon Green® 514 carboxylic acid, and a fluorescein with chloro and methoxy substituents (JOE and 6-JOE)); rhodamine derivatives (e.g., tetramethyl rhodamine (TAMRA), tetramethyl rhodamine iso-thiocyanate (TRITC), tetramethylrhodamine (TMR), carboxy-X-rhodamine (ROX), Texas Red (a mixture of isomeric sulfonyl chlorides and sulforhodamine; Invitrogen™) and Texas Red-X (Texas Red succinimidyl ester, which contains an additional seven-atom aminohexanoyl spacer ("X") between the fluorophore and its reactive group; Invitrogen™), and Rhodamine X); cyanine (Cy) dyes (e.g., Cy3, Cy5 and Cy5.5) and cyanine derivatives (e.g., indocarbocyanine (Quasar® 570, Quasar® 670 and Quasar® 705), Oregon Green® isothiocyanate, and eosin isothiocyanate (EITC)); N-hydroxysuccinimidyl 1-pyrenebutyrate (PYB); N-hydroxysuccinimidyl 1-pyrenesulfonate (PYS); (5-(2'-aminoethyl)aminonaphthalene (EDANS); CAL Fluor® Gold 540, CAL Fluor® Orange 560, Fluor® Red 590, CAL Fluor® Red 610, and CAL Fluor® Red 635 (proprietary fluorophores available from Biosearch Technologies, Inc.); VIC®; HEX® (a 6-isomer phosphoramidite); and NED®.

[0053] Examples of the specific quenchers that may be employed in the present invention include, ~~but are not limited to~~ Black Hole Quencher® dyes (BHQ®-1, BHQ®-2, BHQ®-3); p-(dimethyl aminophenylazo)benzoic acid (DABCYL); Deep Dark Quencher DDQ-I (Eurogentec); Eosin (2',4',5',7'-tetrabromofluorescein); Eclipse® Dark Quencher (Eurogentec); Iowa Black® Quenchers, e.g., Iowa Black® FQ and Iowa Black® RQ (Integrated DNA Technologies, Inc.); QSY-7, QSY-9 and QSY-21 (Molecular Probes®).

[0054] Examples of specific fluorophore-quencher pairs that may be employed in the present invention include, ~~but are not limited to~~, fluorescein/DABCYL, EDANS/DABCYL, CAL Fluor® Gold 540/BHQ®-1, Cy3/BHQ-1, FAM/BHQ®-1, TET/BHQ®-1, JOE/BHQ®-1, HEX/BHQ®-1, Oregon Green®/BHQ-1, Cy3/BHQ®-2, Cy5/BHQ-2, ROX/BHQ®-2, TAMRA/BHQ-2, Cy5/BHQ®-3, and Cy5.5/BHQ®-3.

[0055] Examples of specific fluorophore-fluorophore pairs that may be employed in the present invention include, ~~but are not limited to~~, FAM/TAMRA, FITC/TAMRA, FITC/Rhodamine X, PYS/FITC, FITC/EITC, FITC/PYB, FITC/Texas Red, and FITC/TRITC.

[0056] The methods of the invention may further comprise the use of an oligonucleotide probe that functions as a query molecule looking for a match to a sequence (the “query sequence”) in the target nucleic acid. If the oligonucleotide probe matches the target nucleic acid, it hybridizes to it and acts as a primer for nucleic acid polymerase chain extension. As the extension proceeds, it will displace the inhibitor from the detector. If the oligonucleotide probe does not match the target nucleic acid, it will not hybridize to the target nucleic acid and no chain extension will occur, allowing the inhibitor to remain attached to the detector.

[0057] The methods of the invention may further comprise the use of a polymerase enzyme. This may be any enzyme with strand-displacement capacity. Examples of commercially available polymerase enzymes include, ~~but are not limited to:~~ Klenow fragment (New England Biolabs® Inc.), *Taq* DNA polymerase (QIAGEN), 9°N™ DNA polymerase (New England Biolabs® Inc.), Deep Vent™ DNA polymerase (New England Biolabs® Inc.), Manta DNA polymerase (Enzymatics®), *Bst* DNA polymerase (New England Biolabs® Inc.), and phi29 DNA polymerase (New England Biolabs® Inc.).

[0058] In one embodiment of the methods of this invention, the detector is conjugated to a fluorophore and the inhibitor is conjugated to a quencher. In this example, the detector is incorporated into the target nucleic acid, followed by addition of an oligonucleotide probe, a polymerase and an inhibitor to the reaction. The fluorophore emits fluorescence until the inhibitor binds to the detector and quenches the fluorescent signal by way of the quencher. If the oligonucleotide probe finds a match in the target nucleic acid (as illustrated in FIG. 1A), the polymerase will extend the probe until it reaches the inhibitor and displaces the inhibitor from the detector. As a result, the quencher no longer quenches the fluorescence emitted by the fluorophore and a relatively strong fluorescent signal is emitted. If the oligonucleotide probe does not find a match in the target nucleic acid (as illustrated in FIG. 1B), the inhibitor will remain in place and the quencher will continue to quench the fluorescence emitted by the fluorophore, resulting in a continued relatively low fluorescent signal.

[0059] In another embodiment of the methods of this invention, the detector is conjugated to a quencher and the inhibitor is conjugated to a fluorophore. In this example, the detector is incorporated into the target nucleic acid, followed by addition of an oligonucleotide probe, a polymerase and an inhibitor to the reaction. The inhibitor binds to the detector and the fluorescence emitted by the fluorophore is quenched by the quencher of the detector. If the

[0063] FIG. 6 illustrates an example where the inhibitor is conjugated to an epitope wherein the inhibitor-epitope conjugate may comprise biotin as the epitope. In this example, the reaction proceeds as above and, after purification, the target nucleic acid is tested for the presence of inhibitor bound to the target nucleic acid by quantifying the amount of biotin remaining in the assay. For example, the remaining biotin may be detected by using an anti-biotin, HRP-linked antibody in a chemiluminescent assay, as described previously.

[0064] The results of the detection methods of this invention, referred to herein as "data", associated with a particular target nucleic acid sequence may then be kept in an accessible database, and may or may not be associated with other data from that particular human or animal associated with the target nucleic acid sequence or with data from other humans or animals. Data obtained may be stored in a database that can be integrated or associated with and/or crossmatched to other databases.

[0065] The methods and kits of this invention may further be associated with a network interface. The term "network interface" is defined herein to include any person or computer system capable of accessing data, depositing data, combining data, analyzing data, searching data, transmitting data or storing data. The term is broadly defined to be a person analyzing the data, the electronic hardware and software systems used in the analysis, the databases storing the data analysis, and any storage media capable of storing the data. ~~Non-limiting e~~Examples of network interfaces include people, automated laboratory equipment, computers and computer networks, data storage devices such as, ~~but not limited to,~~ disks, hard drives or memory chips.

[0066] The methods and kits of this invention may further provide for detecting a target nucleic acid sequence in a nucleic acid sample within an emulsion. An "emulsion", as used herein, is a stable mixture of at least two immiscible or partially immiscible liquids. In general, immiscible liquids tend to separate into two distinct phases. Accordingly, a surfactant may be added to stabilize the emulsion by reducing surface tension between the at least two immiscible or partially immiscible liquids and/or to stabilize the interface. For example, an emulsion according to the methods and kits of this invention may comprise a plurality of aqueous droplets in an immiscible oil, such as fluorocarbon oil, silicon oil or hydrocarbon oil where the droplet size ranges from about 0.5 to 5000 microns in diameter. A "droplet", as used herein, means an isolated aqueous or lipophilic phase within a continuous phase having any shape, for example

~~but not limited to~~, cylindrical, spherical and ellipsoidal, as well as flattened, stretched or irregular shapes and so on.

[0067] Although the present invention and its advantages have been described in detail, it should be understood that various changes, substitutions and alterations can be made herein without departing from the spirit and scope of the invention as defined in the appended claims.

[0068] The present invention will be further illustrated in the following Examples which are given for illustration purposes only ~~and are not intended to limit the invention in any way.~~

Examples

Example 1

[0069] In this example, an assay was performed whereby a detector conjugated with a FAM fluorophore and incorporated into a target nucleic acid, was incubated at 34C for 10 minutes in the presence of an inhibitor conjugated with an Iowa Black FQ quencher (Integrated DNA Technologies, Inc.), Manta DNA polymerase (Enzymatics®), dNTPs, the appropriate buffer (supplied with the DNA polymerase from Enzymatics®) and a short oligonucleotide probe having either an exact match to the target nucleic acid or a single position mismatch at one of 6 different positions along the 8 nucleotide long probe as illustrated in FIG. 8.

[0070] Fluorescence was measured using a microplate reader for the 7 reactions. As illustrated in FIG. 9, fluorescence in the sample containing a matched probe was 2 to 5 fold higher than in samples containing mismatched probes.

Example 2

[0071] In this example, an assay was performed wherein a set of oligonucleotide probes was used to distinguish between 2 different alleles in patient samples. The TNNT2 gene was first PCR amplified (exons 4-5, 477bp) using genomic DNA from one patient who was a heterozygote for an A/G SNP and another patient who was a homozygote (G). One of the PCR primers (forward) was labeled with a FAM fluorophore (Integrated DNA Technologies, Inc.). The PCR the samples were converted to single stranded DNA and mixed with an inhibitor conjugated to the Iowa Black® FQ quencher (Integrated DNA Technologies, Inc.), Manta DNA polymerase (Enzymatics®), dNTPs, the appropriate buffer (supplied with the enzyme from Enzymatics®), and a short probe that either matched the SNP allele (allele 1), the "WT" allele (allele 2) or did not match the target (i.e., it was a randomly chosen from a library of hexamer

probes). The fluorescence of each sample was measured after incubation for 30 minutes at 34°C. As illustrated in FIG. 10, relatively high fluorescence is seen with probes matching both alleles 1 and 2 for the heterozygous patient but only allele 2 for the homozygous patient, and relatively low fluorescence is also seen for all mismatched probes.

Example 3

[0072] In this example, a 191bp single stranded DNA amplicon containing a detector conjugated to a FAM fluorophore (Integrated DNA Technologies, Inc.) was incubated for 30 minutes at 34°C and then combined with an inhibitor conjugated to the Iowa Black® FQ quencher (Integrated DNA Technologies, Inc.), Manta DNA polymerase (Enzymatics®), dNTPs, the appropriate buffer (supplied with the enzyme from Enzymatics®), and one of 24 short probes (6 to 8 nucleotide long) that either exactly matched the target (18 probes) or was a mismatch (6 probes). Each one of the 24 mixtures was emulsified into picoliter sized droplets, after which fluorescence was detected for thousands of emulsified drops (1700 drops were measured for each probe on average). The average fluorescence of each probe type and the standard deviations are shown in FIG. 11. Mismatched probes (negatives) all have low fluorescence and all matched probes (positives) have high fluorescence (bars=standard deviation).

* * *

~~[0073] — Having thus described in detail preferred embodiments of the present invention, it is to be understood that the invention defined by the above paragraphs is not to be limited to particular details set forth in the above description as many apparent variations thereof are possible without departing from the spirit or scope of the present invention. —~~

WHAT IS CLAIMED IS:

1. A method for detecting a target nucleic acid comprising:
 - a) incorporating a detector oligonucleotide into a target nucleic acid to form a reaction;
 - b) adding an oligonucleotide probe to the reaction;
 - c) adding an inhibitor to the reaction; and
 - d) detecting interference of the oligonucleotide probe with the inhibitor as an indication of the presence of the target nucleic acid; wherein the inhibitor is an inhibitor oligonucleotide and the detecting comprises performing a strand displacement reaction with a polymerase that extends the oligonucleotide probe and displaces the inhibitor oligonucleotide if the oligonucleotide probe hybridizes to the target nucleic acid.
2. A method according to claim 1, wherein the target nucleic acid is single stranded.
3. A method according to claim 1, wherein a single nucleotide within the target nucleic acid is detected.
4. A method according to claim 1, wherein a short sequence of nucleotides within the target nucleic acid is detected.
5. A method according to claim 1, wherein the entire target nucleic acid is detected.
6. A method according to claim 1, wherein a variance within the target nucleic acid is detected.
7. A method according to claim 1, wherein a mutation within the target nucleic acid is detected.
8. A method according to claim 1, wherein a polymorphism within the target nucleic acid is detected.
9. A method according to claim 1, wherein the detector oligonucleotide is conjugated to a fluorophore.
10. A method according to claim 1, wherein the inhibitor is conjugated to a quencher.
11. A method according to claim 1, wherein the detector oligonucleotide is conjugated to a quencher.
12. A method according to claim 1, wherein the inhibitor is conjugated to a fluorophore.
13. A method according to claim 1, wherein the detector oligonucleotide is conjugated to an epitope.
14. A method according to claim 1, wherein the inhibitor is conjugated to an epitope.
15. A method according to claim 1, wherein more than one target nucleic acid is detected.