

PHARMACEUTICAL COMPOSITION CONTAINING ELEMENE, PREPARATION METHOD THEREFOR, AND USE THEREOF

FIELD OF THE INVENTION

5 The present invention relates to the field of pharmaceutical formulations, in particular, to a pharmaceutical composition comprising elemene and preparation methods and use thereof.

BACKGROUND OF THE INVENTION

10 Elemene is a volatile oily compound extracted from plants such as *Curcuma Wenyujin* Y. H. Chen et C. ling and *Cymbopogon citratus* ((DC.)) Stapf. This compound appears as a pale yellow or yellow clear liquid with a pungent fennel smell. Currently, elemene is clinically used as a mixture containing α -elemene, β -elemene, δ -elemene and γ -elemene, which is widely used in malignant pleural effusion, lung cancer, gastrointestinal tumors and other superficial tumors. Since the elemene available in market and its formulations contain Cremophor EL and other
15 excipients are highly irritating, most patients will suffer from serious phlebitis, fever, local pain, allergic reaction or mild digestive tract reaction, as well as pyrexia and other adverse reactions after the use of the drugs available in market. In the actual clinical medication process, due to the high incidence of phlebitis and higher recurrence rate thereof, the speed of intravenous infusion should be strictly controlled, resulting in a long infusion time.

20 Elemene is almost insoluble in water and has strong lipid solubility, especially in organic solvents such as petroleum ether, diethyl ether and the like. Its molecular formula is $C_{15}H_{24}$ and only consists of hydrocarbons. Due to its unique chemical structure and physicochemical properties, there are great difficulties and obstacles in its preparation of pharmaceutical formulations. In order to increase the solubility of elemene, the Cremophor EL and other
25 excipients were added in drug prescription available in market during the preparation of elemene into injection, resulting in problems of serious hemolysis and great irritation. In addition, although the marketing of paclitaxel albumin nano-formulation provides a new form of pharmaceutical formulation for solving clinical side effects, due to the physicochemical properties of elemene, which is oily at room temperature, it is extremely challenging to formulate
30 elemene into albumin formulations. Therefore, there is currently no corresponding research or development.

SUMMARY OF THE INVENTION

35 The present application provides a pharmaceutical composition comprising elemene, an oil for injection and a protein carrier, which is able to effectively reduce one or more side effects of

pharmaceutical compositions administered into the human body.

In one or more embodiments of the present application, the protein carrier used in the pharmaceutical composition includes a protein, and any suitable protein may be used. Examples of suitable protein include, but are not limited to, albumin, immunoglobulin, including but not limited to IgA, lipoprotein, apolipoprotein B, alpha-acid glycoprotein, beta-2-macroglobulin, thyroglobulin, transferrin protein, fibronectin, factor VII, factor VIII, factor IX, factor X and analogues. The protein carrier may be natural or synthetic. In some embodiments, the protein carrier is a non-blood protein such as casein, alpha-lactalbumin, and beta-lactoglobulin. In some embodiments, the protein carrier includes albumin, such as human serum albumin (HSA), bovine serum albumin, and the like. Human serum albumin is a highly soluble globulin, Mr65K, composed of 585 amino acids. HSA is the most abundant protein in plasma and constitutes 70-80% of the colloidal osmotic pressure in human plasma.

In one or more embodiments of the present application, the “oil for injection” used in the pharmaceutical composition includes, but are not limited to, ethyl oleate, benzyl benzoate, medium chain triglyceride (MCT) and vegetable oil, among which the vegetable oil is selected from one or more of soybean oil, safflower oil, cottonseed oil, corn oil, sunflower oil, peanut oil, and olive oil. In one or more embodiments of the present application, the oil for injection is soybean oil.

In one or more embodiments of the present application, the pharmaceutical composition may be formulated into particles, wherein the particle size of the particles is less than 180 nm, for example, about 70-170 nm, preferably about 70-150 nm.

In one or more embodiments of the present application, the composition comprises nanoparticles of any shape (for instance, spherical or non-spherical shape), and in some embodiments, the average or intermediate diameter of the particles is no greater than about 180 nm. In some embodiments, the average or intermediate diameter of the particles is from about 50 nm to about 180 nm. In some embodiments, the average or intermediate diameter of the particles is from about 60 nm to about 180 nm. In some embodiments, the average or intermediate diameter of the particles is from about 70 nm to about 170 nm. In some embodiments, the average or intermediate diameter of the particles is from about 70 nm to about -170 nm. In some embodiments, the average or intermediate diameter of the particles is from about 70 to about 150 nm. In some embodiments, the average or intermediate diameter of the particles is about 60 nm, 70 nm, 80 nm, 90 nm, 100 nm, 110 nm, 120 nm, 130 nm, 140 nm, or 150 nm. In some implementations, the particles are sterile and filterable.

In one or more embodiments of the present application, the elemene in the pharmaceutical composition is selected from one or more of α -elemene, β -elemene, γ -elemene and δ -elemene.

In one or more embodiments of the present application, the elemene in the pharmaceutical composition is β -elemene.

In one or more embodiments of the present application, the ratio of the albumin carrier, the elemene and the oil for injection in the pharmaceutical composition is the weight ratio of the
5 feed.

In one or more embodiments of the present application, the weight ratio of the protein carrier to the elemene in the pharmaceutical composition is 0.520; for example, 0.5-10.

In one or more embodiments of the present application, the weight ratio of the oil for injection to the elemene in the pharmaceutical composition is 0.3510; for example, 0.5 to 10.

10 In one or more embodiments of the present application, the pharmaceutical composition further contains a lyoprotectant. The lyoprotectant is selected from one or more of glucose, sucrose, maltose, lactose, mannose, trehalose, glycine and dextran, preferably trehalose or sucrose. In one or more embodiments, wherein the ratio of the weight of the lyoprotectant to the volume of the solution of the pharmaceutical composition is 3:100-20:100 g/mL, preferably
15 5:100-10:100 g/mL.

In one or more embodiments of the present application, the pharmaceutical composition further contains one or more of isoosmotic adjusting agent, antioxidant, preservative, and pH regulator. The isoosmotic adjusting agent is one or more of glycerol, sorbitol, mannitol or glucose; the pH regulator is one or more of sodium hydroxide, sodium citrate, citric acid,
20 phosphoric acid, acetic acid or hydrochloric acid; the preservative is one or more of hydroxybenzene alkyl esters, benzoic acid, sodium benzoate, sorbic acid, chlorhexidine acetate, benzalkonium bromide; the antioxidant is one or more of sodium sulfite, sodium bisulfite, sodium metabisulfite, sodium thiosulfate, ascorbic acid, butylated hydroxyanisole, 2,6-di-tert-butylated hydroxytoluene, and vitamin E.

25 In one or more embodiments of the present application, there is also provided a pharmaceutical composition that no organic solvents is used during the preparation process thereof.

In one or more embodiments of the present application, the pharmaceutical composition may be formulated into particles, wherein the particle size of the particles is less than 180 nm,
30 for example, about 70-150 nm.

In one or more embodiments of the present application, wherein in the pharmaceutical composition, the weight ratio of the soybean oil to the elemene is about 0.35-3, and the weight ratio of the human serum albumin to the elemene is about 0.5-20; preferably, the weight ratio of the soybean oil to the elemene is about 0.5-3, the weight ratio of the human serum albumin to
35 the elemene is about 0.5-2.5; or in the pharmaceutical composition, the weight ratio of the

soybean oil to the elemene is about 0.35-1.5, and the weight ratio of the human serum albumin to the elemene is about 1.5-10.

In one or more embodiments of the present application, the weight ratio of soybean oil to elemene is about 0.5, and the weight ratio of human serum albumin to elemene is about 1.5.

5 In one or more embodiments of the present application, the weight ratio of soybean oil to elemene is about 0.75, and the weight ratio of human serum albumin to elemene is about 5.

In one or more embodiments of the present application, the weight ratio of soybean oil to elemene is about 1.5, and the weight ratio of human serum albumin to elemene is about 3-5.

In one or more embodiments of the present application, there is also provided a
10 pharmaceutical composition that contains an organic solvent during the preparation process. Wherein the organic solvent is selected from one or more of chloroform, dichloromethane, tertiary butanol, isopropanol, ethyl acetate, ethanol, tetrahydrofuran, dioxane, acetonitrile, acetone, dimethyl sulfoxide, dimethylformamide, and methylpyrrolidone. Wherein the liposoluble organic solvent is a mixed solvent of dichloromethane and ethanol.

15 Wherein the volume ratio of dichloromethane to ethanol is 1:1-8, preferably 1:4.

The pharmaceutical composition may be formulated into particles, and the particle size of the particles is about 70-150 nm.

In the pharmaceutical composition, the weight ratio of the soybean oil to the elemene is about 2.4-10, and the weight ratio of the human serum albumin to the elemene is about 0.5-10;
20 preferably, in the pharmaceutical composition, the weight ratio of the soybean oil to the elemene is about 2.4, the weight ratio of the human serum albumin to the elemene is about 3.

One or more embodiments of the present application provide a method for preparing the pharmaceutical composition of the present application, which comprises the following steps:

(1) mixing elemene and an oil for injection evenly, optionally adding an organic solvent, to
25 obtain a solution of the elemene and the oil for injection; dissolving a protein carrier in water to obtain a protein carrier solution;

(2) mixing the two solutions obtained in step (1) to form an emulsion;

(3) homogenizing the emulsion in step (2) under a high pressure, and optionally evaporating the emulsion under a reduced pressure to remove the organic solvent, to obtain a nanoparticle
30 solution.

One or more embodiments of the present application provide a method for preparing the pharmaceutical composition of the present application, which comprises the following steps:

(1) mixing elemene and an oil for injection evenly;

(2) taking an albumin and dissolving the albumin in water;

35 (3) mixing the both to form an emulsion;

(4) homogenizing the emulsion in step (3) under a high pressure to obtain a solution of albumin nanoparticles.

In one or more embodiments of the present application, in step (2), a shearing or ultrasonic method is used for mixing; and in step (3), a method of high-pressure homogenization or micro-jet homogenization is used for homogenization.

In one or more embodiments of the present application, step (3) may further comprise: flushing the pipeline of the homogenizer with an appropriate amount of trehalose aqueous solution to replace the homogeneous solution remaining in the homogenizer, and then flushing the pipeline of the homogenizer with an appropriate amount of ultrapure water to replace the residual trehalose aqueous solution, combining the trehalose aqueous solution with the homogeneous solution, mixing evenly, and then filtering.

In one or more embodiments of the present application, wherein the shearing may be performed by shearing with a mixer.

In one or more embodiments of the present application, wherein the speed of shearing with the mixer is 5000 rpm-10,000 rpm.

In one or more embodiments of the present application, wherein the shearing time of the mixer is 1 min-20 min, preferably for example 2 min-10 min or, more preferably 1 min-10 min.

In one or more embodiments of the present application, wherein the shearing is preferably performed multiple times with the mixer, preferably more than 2 times.

In one or more embodiments of the present application, preferably the first time is shearing at 5000 rpm for 1 min, and the second time is shearing at 10,000 rpm for 5 min.

In one or more embodiments of the present application, the pressure of the high-pressure homogenization is 200-1,600 bar, preferably for example 250-1,550 bar.

In one or more embodiments of the present application, wherein the time of the high-pressure homogenization is 1-10 min, for example, preferably 2-10 min, such as further preferably 5-8 min.

In one or more embodiments of the present application, wherein the high-pressure homogenization is preferably performed multiple times, preferably for example more than 2 times.

In one or more embodiments of the present application, the pressure of the micro-jet homogenization is 20,000 psi.

In one or more embodiments of the present application, wherein the time of the micro-jet homogenization is 30 min.

One or more embodiments of the present application further provide a method for preparing the pharmaceutical composition of the present application, which uses an organic solvent in the

preparation process.

In one or more embodiments of the present application, the method for preparing the pharmaceutical composition of the present application comprises the following steps:

(1) dissolving elemene and an oil for injection in a liposoluble organic solvent;

5 (2) taking an albumin and dissolving the albumin in water;

(3) mixing the two solutions obtained in step (1) and (2) to form an emulsion;

(4) homogenizing the emulsion in step (3);

(5) evaporating the emulsion in step (4) under a reduced pressure to remove the organic solvent, to obtain a solution of albumin nanoparticles.

10 In one or more embodiments of the present application, the organic solvent in step (1) is selected from one or more of chloroform, dichloromethane, tertiary butanol, isopropanol, ethyl acetate, ethanol, tetrahydrofuran, dioxane, acetonitrile, acetone, dimethyl sulfoxide, dimethylformamide, methylpyrrolidone; for example, a mixed solvent of dichloromethane and ethanol, and the volume ratio of ethanol to dichloromethane in the mixed solvent is 1:1-8, for
15 example 1:4.

In one or more embodiments of the present application, in the step (3), a shearing or ultrasonic method may be used for mixing; and in the step (4), a high-pressure or micro-jet method may be used for homogenization.

In one or more embodiments of the present application, wherein the shearing may be
20 performed by shearing with a mixer.

In one or more embodiments of the present application, wherein the speed of shearing with the mixer is 5,000-10,000 rpm.

In one or more embodiments of the present application, wherein the shearing time of the mixer is 1 min-20 min, for example 2-10 min.

25 In one or more embodiments of the present application, wherein the shearing is preferably performed multiple times with the mixer, preferably more than 2 times.

In one or more embodiments of the present application, preferably the first time is shearing at 5,000 rpm for 1 min, and the second time is shearing at 10,000 rpm for 5 min.

In one or more embodiments of the present application, wherein the pressure of the high-
30 pressure homogenization is 200-1,600 bar, for example 250-1,550 bar.

In one or more embodiments of the present application, wherein the time of high-pressure homogenization is 1-10 min, for example 2-10 min, such as further preferably 5-8 min.

In one or more embodiments of the present application, wherein the high-pressure homogenization is preferably performed multiple times, for example more than 2 times.

35 In one or more embodiments of the present application, the pressure of the micro-jet

homogenization is 20,000 psi.

In one or more embodiments of the present application, wherein the time of the micro-jet homogenization is 30 min.

5 In one or more embodiments of the present application, the method for preparing the pharmaceutical composition of the present application further comprises the step of lyophilizing the obtained solution of albumin nanoparticles.

One or more embodiments of the present application further provide use of the pharmaceutical composition of the present application in a medicament for the prevention or treatment of cancer.

10 In one or more embodiments of the present application, the cancer involved in the present application includes adrenocortical carcinoma, cryptogenic myeloid metaplasia, AIDS-related cancer, anal cancer, appendiceal cancer, astrocytoma, basal cell carcinoma, cholangiocarcinoma, bladder cancer, bone cancer, glioma, ependymoma, oligodendroglioma, meningioma, craniopharyngioma, hemangioblastoma, medulloblastoma, neuroectodermal tumor, visual
15 pathway and hypothalamic glioma and glioblastoma, bronchial adenoma, carcinoid tumor, central nervous system lymphoma, cervical cancer, colon cancer, colorectal cancer, chronic myeloproliferative disorder, endometrial cancer, ependymoma, Ewing's tumor family, eye cancer, gallbladder cancer, gastric cancer, gastrointestinal carcinoid tumor, gastrointestinal stromal tumor, germ cell carcinoma, gestational trophoblastic tumor, head and neck cancer, liver
20 cancer, laryngeal cancer, leukemia, lip and oral cancer, lung cancer, lymphoma, medulloblastoma, melanoma, mesothelioma, metastatic neck squamous cell carcinoma, multiple endocrine neoplasia syndrome, myelodysplastic syndrome, myelodysplastic/ myeloproliferative disease, carcinoma of nasal cavity and sinuses, neuroblastoma, neuroendocrine cancer, oropharyngeal cancer, nasopharyngeal cancer, brain tumor, bone metastasis cancer, intestinal
25 cancer, esophageal cancer, ovarian cancer, pancreatic cancer, skin cancer, parathyroid carcinoma, penile carcinoma, peritoneal cancer, pharyngeal carcinoma, pheochromocytoma, pineoblastoma and supratentorial primitive neuroectodermal tumor, pituitary tumor, pleuropulmonary blastoma, lymphoma, primary central nervous system lymphoma, pulmonary lymphangiomyomatosis, rectal cancer, kidney cancer, renal pelvis and ureter cancer, rhabdomyosarcoma, salivary gland
30 cancer, small intestine cancer, squamous epithelial cell carcinoma, testicular cancer, throat cancer, thymoma and thymic carcinoma, thyroid cancer, urethral cancer, vaginal cancer.

In one or more embodiments of the present application, the cancer involved in the present application includes lung cancer, liver cancer, esophageal cancer, nasopharyngeal cancer, brain tumor, bone metastasis cancer, gastric cancer, intestinal cancer, uterine cancer, cervical cancer,
35 germ cell carcinoma, endometrial cancer, gestational trophoblastic tumor, breast cancer, skin

cancer, lymphoma, leukemia, malignant melanoma.

In one or more embodiments of the present application, the brain tumor includes glioma, brain stem glioma, cerebellar or cerebral astrocytoma, malignant glioma, ependymoma, oligodendroglioma, meningioma, craniopharyngioma, hemangioblastoma, medulloblastoma, visual pathway and hypothalamic glioma or glioblastoma; the cerebellar or cerebral astrocytoma is fibrous cell astrocytoma or diffuse astrocytoma or anaplastic (malignant) astrocytoma.

In one or more embodiments of the present application, the pharmaceutical composition of the present application may be used in combination with radiotherapy and chemotherapy for lung cancer, liver cancer, esophageal cancer, nasopharyngeal cancer, brain tumor, bone metastasis and other malignant tumors to enhance the curative effect, reduce the toxic and side effects of radiotherapy and chemotherapy, and may also be used for interventional, intracavitary chemotherapy and the treatment of cancerous hydrothorax and ascite.

In one or more embodiments of the present application, the compounds of the present application may be used in combination with other drugs such as anticancer drugs, wherein the anticancer drugs include temozolomide, paclitaxel, docetaxel, taxane, gemcitabine, anti-VEGF drugs, PD-1 antibody drugs; wherein, anti-VEGF drugs include AVASTIN, ranibizumab, aflibercept or conbercept; wherein, PD-1 antibody drugs include nivolumab, pembrolizumab, toripalimab, sintilimab, cemiplimab, atezolizumab, avelumab, or durvalumab.

One or more embodiments of the present application provide a method for the treatment of human disease, comprising the administration of a therapeutically effective amount of the pharmaceutical composition of the present application. The pharmaceutical composition is administered by parenteral, by inhalation, intraperitoneal, intravesical, intramuscular, intravenous, intratracheal, subcutaneous, intraocular, intrathecal, transdermal, rectal or vaginal.

In one or more embodiments of the present application, the pharmaceutical composition of the present application may be formulated into a pharmaceutical formulation suitable for intravenous administration, a pharmaceutical formulation for parenteral administration, a pharmaceutical formulation for gastrointestinal administration, an aerosol, a pharmaceutical formulation for vaginal administration or other suitable pharmaceutical formulation, the pharmaceutical formulation is a solid formulation, a liquid formulation or a gas formulation.

In one or more embodiments of the present application, the pharmaceutical composition of the present application may be used as a liquid formulation for injection, which comprises elemene and a pharmaceutically acceptable carrier, wherein the pharmaceutically acceptable carrier includes an albumin carrier and an oil for injection; wherein the composition may be formulated into particles, wherein the particle size of the particles is 70-150 nm; and in the composition, the weight ratio of the albumin carrier to the elemene is about 0.5-20, and the

weight ratio of the oil for injection and the elemene is about 0.35-10.

In one or more embodiments of the present application, the liquid formulation of the present application may be a stable aqueous suspension reconstituted from a sterile lyophilized powder.

One or more embodiments of the present application provide a sealed container, which may
5 be a unit-dose container or a multi-dose container, comprising the pharmaceutical composition of the present application. The sealed container may be a prefilled syringe. The pharmaceutical composition is a liquid composition or a dry composition. The pharmaceutical composition is lyophilized and sterile.

In one or more embodiments of the present application, the nano-formulation or
10 nanoparticles described herein may be present in the form of a dry formulation (for instance, a lyophilized composition) or suspended in a biocompatible medium. The suitable biocompatible medium includes, but is not limited to, water, aqueous buffered medium, saline, buffered saline, optionally buffered amino acid solution, optionally buffered protein solution, optionally buffered sugar solution, optionally buffered vitamin solution, optionally buffered synthetic polymer
15 solution, lipid-containing emulsion, and the like.

Unless expressly stated otherwise, an “individual” as used herein is a mammal including, but not limited to, a primate, human, bovine, equine, feline, canine, or rodent.

As used herein, “treatment” is a method of obtaining beneficial or desired outcomes, including clinical outcomes. For the purposes of the present invention, beneficial or desired
20 clinical outcomes include, but are not limited to, any one or more of the following: reducing one or more symptoms resulting from the disease, reducing the severity of the disease, stabilizing the disease (for instance, preventing or delaying the exacerbation of the disease), preventing or delaying the spread (e.g. metastasis) of disease, preventing or delaying the occurrence or recurrence of disease, preventing or slowing the progression of the disease, improving the state
25 of the disease, providing remission (whether partial or total) of the disease, reducing the dose of one or more other drugs necessary to treat a disease, delaying the progression of the disease, increasing quality of life, and/or prolonging survival. In some embodiments, the composition reduces the severity of one or more symptoms associated with cancer, as compared to corresponding symptoms in the same subject prior to treatment or compared to corresponding
30 symptoms in other subjects who do not receiving the composition, by at least about any one of 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90%, 95%, or 100%. “Treatment” also includes alleviation of the pathological consequences of cancer. The methods of the present invention contemplate any one or more of these therapeutic aspects.

As used herein, “pharmaceutically acceptable” or “pharmacologically compatible” means
35 a substance that is not biological or otherwise undesirable may be incorporated into a

pharmaceutical composition administered to a patient, for instance, without causing any significant undesired biological effect or without interacting in a deleterious manner with any other ingredient contained in the composition. The pharmaceutically acceptable carrier or excipient preferably meets the standards required for toxicology and manufacturing testing.

Reference herein to “about” a value or parameter includes (and describes) embodiments that refer to the value or parameter itself. For example, description referring to “about X” includes description of “X”.

There are a variety of suitable formulations for the pharmaceutical composition of the present invention. The following formulations and methods are exemplary only and not limiting.

Formulations for oral administration may be made of (a) a liquid solution, for instance, an effective amount of the active ingredient dissolved in a diluent such as water, saline or orange juice, (b) capsules, sachets or tablets, each containing a predetermined amount of the active ingredient in solid or granular form, (c) a suspension in an appropriate liquid, and (d) a suitable emulsion. Tablet forms may include one or more of lactose, mannitol, corn starch, potato starch, microcrystalline cellulose, arabic gum, gelatin, colloidal silicon dioxide, croscarmellose sodium, talc, magnesium stearate, stearic acid and other excipients, colorants, diluents, buffers, wetting agents, preservatives, flavoring agents and pharmacologically compatible excipients. Lozenge forms may contain the active ingredient in spices, usually sucrose and arabic gum or tragacanth, and troches containing the active ingredient in an inert matrix, such as gelatin and glycerin, or sucrose and arabic gum, emulsions, gels, etc., in addition to the active ingredient, excipients known in the art are also contained.

Formulations for parenteral administration include an aqueous and nonaqueous, isotonic sterile injection solution, which may contain antioxidants, buffers, bacteriostatic agents, and solutes for rendering the formulation isotonic with the blood of the intended recipient, as well as an aqueous and non-aqueous sterile suspension which may include suspending agents, solubilizers, thickening agents, stabilizers and preservatives. The formulation may be presented in unit-dose or multi-dose sealed containers, such as ampoules and vials, and may be stored in a freeze-drying (lyophilizing) condition immediately before use, requiring only the addition of a sterile liquid excipient, such as water, for injection. Extemporaneous injection solutions and suspensions may be prepared from some kinds of the sterile powders, granules and tablets as described above.

Aerosols comprise the pharmaceutical composition of the present invention, the pharmaceutical composition includes an aqueous and nonaqueous, isotonic sterile solution which may contain antioxidants, buffers, bacteriostatic agents, and solutes, as well as an aqueous and non-aqueous sterile suspension which may include suspending agents, solubilizers,

thickening agents, stabilizers and preservatives; the pharmaceutical composition may be prepared as an aerosol formulation alone or in combination with other suitable components for administration by inhalation. These aerosol formulations may be placed in pressurized acceptable propellants such as difluoromethane, propane, nitrogen, and the like. They may also be formulated as a non-pressure formulation of drugs, for example in a nebulizer or atomizer.

Other suitable formulations are also possible, for example suppositories may be prepared by using various bases such as emulsified bases or water-soluble bases. Formulations for vaginal administration may be presented as pessaries, tampons, creams, gels, pastes, foams or spray formulations, containing suitable carriers known in the art in addition to the active ingredient.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 shows the hydratization diameter distribution of β -elemene albumin nanoparticles;

Figure 2 shows the potential diagram of β -elemene albumin nanoparticles;

Figure 3 shows the transmission electron microscope image of β -elemene albumin nanoparticles;

Figure 4 shows the stability investigation of particle size of different lyophilized samples after reconstitution;

Figure 5 shows the content stability investigation of different lyophilized samples after storage;

Figure 6 shows the survival of tumor-bearing mice in each group;

Figure 7 shows the results of the hemolytic experiment;

Figure 8 shows the tumor growth curve of the orthotopic tumor model in U87R1 mice;

Figure 9 shows the survival curve of the orthotopic tumor model in U87R1 mice.

DETAILED DESCRIPTION OF THE INVENTION

The following embodiments are further illustrations of the invention and are not a limitation of the scope of the invention. The present invention is further described in detail below with reference to the examples, but those skilled in the art should understand that the present invention is not limited to these examples and the preparation methods as used. Moreover, those skilled in the art can make equivalent substitutions, combinations, improvements or modifications to the present invention according to the description of the present invention, but these will all be included in the scope of the present invention.

The soybean oil involved in the present invention comes from soybean oil (SIO, injection grade, Lot19C4520/Lot18C4816); the human serum albumin (HSA) involved in the present invention comes from human serum albumin (20%, Baxter, LotA1U133A) or (20%, Grifols,

LotALAFD03042).

The commonly used detection methods of the present invention include particle size and particle size distribution, content determination, hemolysis experiment, etc., and the details are as follows:

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Particle size and particle size distribution

The particle size and the particle size distribution were determined with a particle size analyzer (MALVERN ZETASIZER NANO ZS90). 25 μ L of sample was taken, 1 mL of ultrapure water was added to dilute it for detection, detection angle: 90°; dispersion medium: water; 10 temperature: 25°C.

The particle size was the Z-average value (the average particle size of the sample) measured by the particle size analyzer, and the particle size distribution was the PDI value measured by the particle size analyzer.

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Content determination

It was determined by HPLC (General Rule 0512 of the Fourth Part of Chinese Pharmacopoeia 2015 Edition).

Chromatographic column was CHIRALPAK® IG (4.6×150 mm, 5 μ m), with acetonitrile-water (55:45) as mobile phase, at a flow rate of 1.0 mL/min. The injection volume was 20 μ L, 20 the column temperature was 30°C, and the detection wavelength was 210 nm.

Preparation of reference substance solution (0.2 mg/mL) (external standard method): about 10 mg of β -elemene reference substance was precisely weighed into a 50 mL volumetric flask with a small amount of acetonitrile added, acetonitrile was added to be dissolved and diluted to volume scale, the mixture was shaken well to obtain the reference substance solution.

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Preparation of test solution: the cap was removed from 1 bottle of this product and the product was precisely weighed (m_1), an appropriate amount of water was added (to make the ratio of API to water of 20 mg:1 mL), precisely weighed (m_2) and reconstituted by shaking. About 500 mg of the reconstituted solution (about equivalent to API 10 mg) (m_3) was precisely weighed and placed in a 50 mL volumetric flask, about 1.5 mL of water was added first, and then 30 acetonitrile was slowly added to 80% of the full range, sonicated for 10 min, cooled down, and acetonitrile was added again to dilute to volume scale, the mixture was shaken well, and filtered through a nylon filter with a pore size of 0.45 μ m, to obtain the test solution. After the reconstituted solution was used, the vial was washed with water and dried, and the empty vial was precisely weighed (m_4).

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20 μ L was precisely measured, injected into a liquid chromatograph, and the chromatogram

was recorded. The content of β -elemene ($C_{15}H_{24}$) in the test sample was calculated by the peak area according to the external standard method.

Calculation formula was:

$$\text{Content}(\%) = C_R \times \frac{A_X \times 50 \times (m_2 - m_4)}{A_R \times m_3 \times (m_1 - m_4)} \times 100\%$$

5 in the formula,

A_X : API peak area in the test solution

A_R : peak area of the reference substance

C_R : concentration of the reference substance

10 **Method for the determination of hemolysis value**

Preparation of blood cell suspension: a few milliliters of blood was taken from rabbits (male New Zealand rabbits, laboratory-suitable rearing for 1 week, body weight 2.5 kg-3 kg, blood was taken from the heart), and put in a beaker, the blood was stirred with a glass rod or bamboo stick to remove fibrinogen and get defibrinated blood. 0.9% sodium chloride solution was added
15 in an amount of about 10 times, mixed evenly, centrifuged at 1,500 rpm/min for 15 min, the supernatant was discarded, and the precipitated red blood cells were washed 2-3 times with 0.9% sodium chloride solution according to the above method, until the supernatant was not red. The obtained red blood cells were prepared into 2% blood cell suspension with 0.9% sodium chloride solution for later use.

20 Sample preparation: according to the sample content, a proper amount was taken and diluted successively with normal saline to obtain 1 mg/mL sample (actual content).

Reaction observation: each drug was added in the following manner, and after mixing, the mixture was immediately placed in a water bath at 37°C for incubation, and the hemolysis was observed after 3 hours.

25 Drug group: 2.5 mL of 2% blood cell suspension, 2.2 mL of normal saline, and 0.3 mL of drug were successively added to a test tube.

Positive group: 2.5 mL of 2% blood cell suspension and 2.5 mL of distilled water were successively added to a test tube.

Negative group: 2.5 mL of 2% blood cell suspension and 2.5 mL of normal saline were
30 successively added to a test tube.

Determination of OD value: 100 μ L of the supernatant was pipetted into a 96-well plate, and the OD value was measured at a wavelength of 540 nm.

The calculation method of hemolysis value: (OD value of the sample - OD value of the

negative group) / (OD value of the positive group - OD value of the negative group).

Test Example 1

The preparation of β -elemene albumin nanoparticles included the following steps: 1) 200 mg of elemene and 600 mg of soybean oil were weighted, and dissolved in 2 mL of dichloromethane to obtain an oil phase. 2) 650 mg of human serum albumin was dissolved in water for injection to obtain an aqueous solution of human serum albumin with a mass volume percentage of 3.6% (g/mL), that is, to obtain an aqueous phase. 3) Under the action of water bath ultrasound, the oil phase was added dropwise to the aqueous phase, and sonicated by a probe for 8 min (250 W) to produce a primary emulsion. 4) The obtained primary emulsion was transferred to a high-pressure homogenizer for high-pressure homogenization, the homogenization pressure was 1300 bar, and the homogenization was performed for 9 times. The organic solvent was removed from the homogenized system by rotary evaporation, and a 0.22 μ m microporous membrane was used for filtration and sterilization to obtain an aqueous solution of elemene albumin nanoparticles, and the metered volume was adjusted to 10 mg/mL with water for injection. 5) 3% sucrose was added to the obtained elemene albumin nanoparticles, and lyophilized. The lyophilization parameters were: pre-freezing at -45°C for 5h; starting the vacuum pump and keeping the vacuum degree at about 10 Pa, temperature programming, drying at -40°C for 4h, drying at -30°C for 10h, and then drying at -20°C, -10°C, 0°C, 5°C and 10°C for 4h respectively. The lyophilized samples prepared above were reconstituted in pure water, and the particle size, potential and microscopic morphology of the nanoparticles were investigated by the following methods.

The Malvern particle size analyzer was preheated for 30 min before measurement, 50 μ L of aqueous solution of the elemene albumin nanoparticles was taken, diluted to 1 mL with water, shaken slightly (to avoid bubbles), and placed in a Malvern Nano-ZS90 particle size dish or a potential dish to measure particle size and Zeta potential. The measurement temperature was 25°C, the equilibration time was 120 s, and the rest of the items were kept at the default values of the instrument, and the data was collected and processed by the Malvern V2.2 workstation. The particle size diagram and Zeta potential of the elemene albumin nanoparticles obtained were shown in Figure 1 and Figure 2. As shown in Figure 1 and 2, the particle size of the elemene albumin nanoparticles was about 110 nm, PDI was 0.127, and the potential was about -18 mV.

The aqueous solution of the elemene albumin nanoparticles was aspirated by a pipette and dropped on a copper mesh. After natural evaporation, 1% phosphotungstic acid solution was dropped on the copper mesh in the same way, and evaporated naturally, a few drops of pure water was aspirated to rinse the surface of the copper mesh carefully. After evaporation, the

morphology was observed by transmission electron microscope as shown in Figure 3, the nanoparticles were spherical, regular in shape, relatively uniform in particle size, with a particle diameter of about 100 nm.

5 **Test Example 2**

Elemene albumin nanoparticles was prepared according to the method of Test Experiment 1, wherein the oil for injection was employed with the same amount as those of soybean oil, corn oil, camellia oil, and medium chain triglyceride (MCT) respectively to prepare elemene albumin nanoparticles using different oils for injection as raw materials. The particle size of the prepared nanoparticles was determined by a laser particle size analyzer. After lyophilization, the drug content of each group of formulations before and after lyophilization was determined respectively, and the drug loss rate during lyophilization was calculated.

The preparation of elemene albumin nanoparticles without soybean oil included the following steps: 1) 200 mg of elemene was weighed and dissolved in 2 mL of dichloromethane to obtain an oil phase. 2) 650 mg of human serum albumin was dissolved in water for injection to obtain an aqueous solution of human serum albumin with a mass volume percentage of 3.6% (g/mL), that is, to obtain an aqueous phase. 3) Under the action of water bath ultrasound, the oil phase was added dropwise to the aqueous phase, and sonicated by a probe for 8 min (250 W) to produce a primary emulsion. 4) The obtained primary emulsion was transferred to a high-pressure homogenizer for high-pressure homogenization, the homogenization pressure was 1300 bar, and the homogenization was performed for 9 times. The organic solvent was removed from the homogenized system by rotary evaporation, and a 0.22 μ m microporous membrane was used for filtration and sterilization to obtain an aqueous solution of elemene albumin nanoparticles, and the metered volume was adjusted to 10 mg/mL with water for injection. 5) 3% sucrose was added to the obtained elemene albumin nanoparticles, and lyophilized according to the lyophilization parameters of Test Example 1. The drug content of a certain volume of the formulation before and after lyophilization was determined respectively, and the drug loss rate during lyophilization was calculated.

In addition, the groups of soybean oil, corn oil, camellia oil, and medium chain triglyceride only comprising no lyoprotectant of 3% sucrose were prepared according to the above methods, and the drug content of each certain volume of the formulations before and after lyophilization was determined respectively, and the drug loss rate during lyophilization was calculated. The results are shown in Table 1.

Table 1: Drug loss rate of samples prepared with different oils for injection after lyophilization

Lyophilization loss rate	Soybean oil	Corn oil	Camellia oil	MCT	None
3% sucrose contained	7.1%	10.4%	9.2%	11.8%	31.4%
Sucrose free	9.5%	13.3%	11.1%	13.0%	39.5%

Experiments show that the lyophilization loss rates of albumin nanoparticles prepared by mixing oil for injection with elemene were reduced in different degrees compared with nanoparticles prepared without oil for injection.

Test Example 3

According to the method of Test Example 1, the amount of soybean oil was adjusted, the elemene albumin nanoparticles were prepared by feeding according to the mass ratio of β -elemene/soybean oil of 5:1, 1:1, 1:3, 1:5, 1:10, 1:20 and without oils for injection.

The particle size of the prepared nanoparticles was determined by a laser particle size analyzer. After adding 3% sucrose and lyophilizing, the lyophilized samples were placed in a room temperature dryer. The samples were taken out periodically and reconstituted, and the changes of particle size and main drug content of the lyophilized samples within 30 days were determined. Specific detection parameters were as follows: instrument model: MALVERN ZETASIZER NANO ZS90, detection angle: 90°; detection mode: automatic; detection times: 3; detection type: nanoparticles; refractive index of the sample to be determined: 1.340; dispersion medium: water; temperature: 25°C; viscosity: 0.8872cp; refractive index of dispersion medium: 1.330; equilibration time: 120s; type of particle size dish : DTS0012.

The results were shown in Figure 4 and Figure 5. The particle size of the formulation without soybean oil during the preparation process was more than 200 nm after lyophilization. With the prolongation of storage time, the particle size of the sample after reconstitution showed a trend of increasing gradually, and the drug content showed a trend of decreasing. Gradually increasing the proportion of soybean oil in the formulation, the particle size of the lyophilized samples after reconstitution was all less than 200 nm, the stability of particle size was better, and the stability of the main drug content was also gradually enhanced. Wherein, when the mass ratio of soybean oil/elemene was in the range of 1-10, the prepared lyophilized sample of nanoparticles had no aggregation phenomenon within 30 days at room temperature, the particle size was 100-150 nm, the PDI was less than 0.3, and the content of the main drug of β -elemene changed little. When the amount of elemene was 1 part and the amount of soybean oil was less than 1 part, the prepared nanoparticles did not show a good enough stability.

Test Example 4

To investigate the therapeutic effect of the elemene albumin nanoparticles on mouse brain glioma, the experimental method was as follows.

5 Tumor modeling: the well-grown C6 cells in logarithmic phase were digested with trypsin, centrifuged, and the upper waste solution was discarded. The cells were washed with PBS, centrifuged, and the upper waste solution was discarded, and the cells were resuspended in PBS to make the final C6 cell density about 1×10^6 cells/5 μ L. Healthy male Kunming mice (28-30 g) were anesthetized by intraperitoneal injection of an appropriate amount of 4% chloral hydrate.

10 After shaving the hair on the top of the head, 75% ethanol was applied for disinfection, and a small incision was made along the sagittal midline of the head with sterile ophthalmic scissors, the tissue membrane was quickly removed with a cotton swab moistened with 10% hydrogen peroxide, the skull was separated, and the hydrogen peroxide was washed away with a cotton swab moistened with PBS. The head of the mouse was fixed on a brain stereotaxic device,

15 located at a position 1.8 mm to the right and 0.6 mm to the back of the bregma, a small hole was drilled at the locating point with the needle of a 2 mL syringe, which should not be too deep to avoid bleeding. 5 μ L of the above-mentioned C6 cells with a density of 1×10^6 cells/5 μ L was aspirated with a micro-injection needle. The needle was inserted by 4 mm and withdrawn by 1 mm along the vertical direction of the small hole to leave a space for cells, and the cell

20 suspension was slowly injected into the brain. The microneedle was taken out after remaining in the position of the small hole for 5 min. The wound was sutured with surgical thread, and finally the sutured surface was smeared with a cotton swab moistened with double antibody.

On the 10th day after tumor inoculation, the tumor-bearing mice were divided into normal saline group (NS), commercially available elemene liposomes (Lip, Dalian Jingang, approval

25 number: Guoyaozhunzi H10960114), elemene albumin nanoparticles (NPs) prepared in Test Example 1, with 12 mice in each group. The dosage of administration was 30 mg/kg, once every 2 days, for a total of 4 times. The survival time of each mouse was recorded, and the survival curve was made.

The investigation results of survival were shown in figure 6. The results of the experiment

30 showed that the C6 glioma-bearing mice administered with the elemene albumin nanoparticles exhibited longer survival. The average survival of the normal saline group was 21 days, the average survival of the elemene liposome group was 28 days, and the average survival of the elemene albumin nanoparticles group was 41 days. Therefore, the elemene albumin nanoparticles prepared in the experiment have an effect of prolonging the survival of C6-bearing

35 glioma mice.

Test Example 5

Hemolysis experiment:

New Zealand rabbits (male, laboratory-suitable housed for 1 week, body weight 2.5 kg-3 kg) were divided into two groups of commercially available elemene injection (Ailineng, CSPC Yuanda Pharmaceutical Co., Ltd., batch number 17050302) and elemene albumin nanoparticles (NPs) prepared in Test Example 1, with 2 mice in each group, the dosage of administration was 50 mg/kg. The two formulations were respectively prepared into medicinal solutions having a concentration of 6 mg/mL with glucose solution, and then the rabbits were administered by intravenous infusion.

Within 10-15 minutes after administration, about 1 mL of venous blood was drawn from the ear vein and placed in a centrifuge tube coated with heparin sodium, centrifuged at 3500 r/min for 15 minutes, and the supernatant was taken to observe the color.

The color of the obtained plasma was shown in figure 7. The result showed that the New Zealand rabbits administered with the elemene albumin nanoparticles did not show hemolysis, demonstrating little irritation and high safety.

Test Example 6

1.1 Preparation of experimental samples

Sample 1

3.33 g of β -elemene and 8 g of soybean oil were weighed and mixed to obtain an oil phase. 50 mL (10g) of 20% human serum albumin was measured, diluted evenly by adding an appropriate amount of ultrapure water, and the metered volume was adjusted to 333 mL to obtain an aqueous solution of human serum albumin with a mass volume percentage of 3% (g/mL), namely an aqueous phase. The aqueous phase was poured into the oil phase, and the mixer (FLUKO, FA30/30F) was used for shearing at 10,000 rpm for 10 min to form a primary emulsion. The obtained primary emulsion was transferred to a micro-jet homogenizer (Microfluidizer, M-110-EH) for homogenization, at a homogenization pressure of 20,000 psi for 30 min. The obtained emulsion was filtered by 0.45 μ m and 0.22 μ m PVDF plate filters, and the metered volume was adjusted, and 3% (g/mL) of sucrose was added as a lyoprotectant. The homogeneous solution added with the lyoprotectant was lyophilized, and after lyophilization, the lyophilized powder was reconstituted with sterilized ultrapure water. Particle size was 175.97 nm detected by a particle size analyzer, PDI was 0.188.

Sample 2

3.33 g of β -elemene and 8 g of soybean oil were weighed, dissolved in an organic solvent, and mixed to obtain an oil phase. The organic solvent was a mixed solvent of 26.67 mL dichloromethane and 6.67 mL anhydrous ethanol. 50 mL (10 g) of 20% human serum albumin was measured, diluted evenly by adding an appropriate amount of ultrapure water, and the metered volume was adjusted to 333 mL to obtain an aqueous solution of human serum albumin with a mass volume percentage of 3% (g/mL), namely an aqueous phase. The aqueous phase solution was poured into the oil phase, and the mixer (FLUKO, FA30/30F) was used for shearing at 10,000 rpm for 10 min to form a primary emulsion. The obtained primary emulsion was transferred to a micro-jet homogenizer (Microfluidizer, M-110-EH) for high-pressure homogenization, at a homogenization pressure of 20,000 psi for 30 min. After homogenization, the organic solvent was removed by a thin film evaporator (Shanghai DEA) from the organic solvent-containing emulsion, and the parameters were: temperature 30°C, vacuum degree 4000 pa, liquid feeding speed 100 rpm, scraper speed 50 rpm. The obtained emulsion was filtered by 0.45 μ m and 0.22 μ m PVDF plate filters, and the metered volume was adjusted, and 3% (g/mL) of sucrose was added as a lyoprotectant. The homogeneous solution added with the lyoprotectant was lyophilized, and after lyophilization, the lyophilized powder was reconstituted with sterilized ultrapure water. Particle size was 78.34 nm detected by a particle size analyzer, PDI was 0.207.

Sample 3 (Elemene injection (trade name: Ailineng)): CSPC Yuanda (Dalian) Pharmaceutical Co., Ltd., batch number: 19061502; particle size was 14.74 nm detected by a particle size analyzer, PDI: 0.035.

1.2 PK and mouse brain tissue distribution of elemene albumin nanoparticles

Kunming mice (male, SPF grade, body weight 20-25 g) were used for the experiment. The dosage of administration was 50 mg/kg, administered through the tail vein. Brain tissue samples were collected at different time periods and stored at -20°C in time for the detection of drug content in brain tissue. There were a total of 45 mice for three samples, 3 Kunming mice for each dosage form at each time point. The eyeballs were removed for blood collection at 5min, 15min, 30min, 1h, and 2h after administration, and brain tissues were collected to detect drug content in brain tissues. GC-MS was used to quantitatively analyze β -elemene in biological samples of plasma and brain tissue of the Kunming mice. The results of the study on the half-life in brain tissue and the brain tissue exposure are shown in Table 2.

Table 2 Results of studies on half-life and brain tissue exposure

	Sample 1	Sample 2	Sample 3
Particle size (nm)	175.97	78.34	14.7
Half-life in brain tissue (min)	41.8	57.5	33.6
Brain drug exposure (mg/L*min)	2808.7	3008.6	2812.7

Test Example 7**Preparation and stability investigation of sample 4**

5 1 g of β -elemene and 3 g of soybean oil were weighed and mixed to obtain an oil phase. 15 mL (3 g) of 20% human serum albumin was measured, diluted evenly by adding 85 mL of ultrapure water, to obtain an aqueous solution of human serum albumin with a mass volume percentage of 3% (g/mL), namely an aqueous phase. The aqueous phase solution was poured into the oil phase, and the mixer was used for shearing at 5000 rpm for 1 min, and at 10,000 rpm
10 for 5 min, to form a primary emulsion. The obtained primary emulsion was transferred to a high-pressure homogenizer for high-pressure homogenization at a homogenization pressure of 250-300 bar for 2 min, and at 1350-1400 bar for 9 min. After homogenization, the total volume of the homogeneous solution was measured with a measuring cylinder, which was about 50 mL, 5 g trehalose was added, shaken well to be dissolved, and filtered with a 0.45 μ m microporous
15 membrane to obtain a solution of elemene albumin nanoparticles. The particle size was 141.0 nm detected by a particle size analyzer, PDI was 0.162. The homogeneous solution added with trehalose was lyophilized, and after lyophilization, the lyophilized powder was reconstituted with sterilized ultrapure water. The particle size was 144.07 nm detected by a particle size analyzer, PDI was 0.123. The loss rate of elemene content before and after lyophilization was
20 1.27%.

Preparation and stability investigation of sample 5

4 g of β -elemene and 2 g of soybean oil were weighed and mixed to obtain an oil phase. 30 mL (6 g) of 20% human serum albumin was measured, diluted evenly by adding 170 mL of
25 ultrapure water, to obtain an aqueous solution of human serum albumin with a mass volume percentage of 3% (g/mL), namely an aqueous phase. The aqueous phase solution was poured into the oil phase, and the mixer was used for shearing at 5000 rpm for 1 min, and at 10,000 rpm for 10 min, to form a primary emulsion. The obtained primary emulsion was transferred to a high-pressure homogenizer for high-pressure homogenization at a homogenization pressure of

250-300 bar for 2 min, and at 1350-1400bar for 24 min. After homogenization, 150 mL of homogeneous solution was measured with a measuring cylinder, 15 g trehalose was added, shaken well to be dissolved, and filtered with a 0.45 μ m microporous membrane to obtain a solution of elemene albumin nanoparticles. The particle size was 132.4 nm detected by particle size analyzer, PDI was 0.133. The homogeneous solution added with trehalose was lyophilized, and after lyophilization, the lyophilized powder was reconstituted with sterilized ultrapure water. The particle size was 123.68 nm detected by a particle size analyzer, PDI was 0.144. The loss rate of elemene content after lyophilization was 1.58%.

Preparation and stability investigation of sample 6

1 g of β -elemene and 2 g of egg yolk lecithin were weighed, 50 mL of ultrapure water was added, and the mixer was used for shearing at 5000 rpm for 1 min, and shearing at 10,000 rpm for 5 min. Then the congealed fat adhered to the bottle wall was scraped into the solution, and sheared at 15,000rpm for 3 min again until no obvious undispersed phospholipid was found, and a primary emulsion was obtained. The primary emulsion was transferred to a high-pressure homogenizer for high-pressure homogenization at a homogenization pressure of 250-300 bar for 2 min, and at 1300-1350 bar for 12 min. After homogenization, the homogeneous solution was filtered with a 0.45 μ m filter membrane, and 50 mL of 2% HSA solution (containing 1 g of HSA) was added to the filtered homogeneous solution, and homogenized at 280 bar for 1 cycle; and the homogeneous solution was discharged, 40 mL was measured with a measuring cylinder, 4 g trehalose was added, shaken well to be dissolved, and filtered with a 0.45 μ m microporous membrane to obtain a solution of albumin nanoparticles. The particle size was 150.86 nm detected by a particle size analyzer, PDI was 0.114. The homogeneous solution added with trehalose was lyophilized, and after lyophilization, the lyophilized powder was reconstituted with sterilized ultrapure water. The particle size was 181.56 nm detected by a particle size analyzer, PDI was 0.251. The loss rate of elemene content after lyophilization was 16.47%.

Preparation and stability investigation of sample 7

1 g of β -elemene, 2 g of egg yolk lecithin and 0.5g of TPGS were weighed, 50 mL of ultrapure water was added, and the mixer was used for shearing at 5000 rpm for 1 min, and shearing at 10,000 rpm for 5 min. Then the congealed fat adhered to the bottle wall was scraped into the solution, and sheared at 15,000rpm for 3 min again until no obvious undispersed phospholipid was found, and a primary emulsion was obtained. The primary emulsion was transferred to a high-pressure homogenizer for high-pressure homogenization at a homogenization pressure of 250-300 bar for 2 min, and at 1300-1350 bar for 12 min. After

homogenization, the solution was filtered with a 0.22 μm filter membrane, and 50 mL of 2% HSA solution (containing 1 g of HSA) was added to the filtered homogeneous solution, homogenized at 140 bar for 1 cycle; and the homogeneous solution was discharged, 40 mL was measured with a measuring cylinder, 4 g of trehalose was added, shaken well to be dissolved, and filtered with a 0.45 μm microporous membrane to obtain a solution of albumin nanoparticles. The particle size was 93.11 nm detected by a particle size analyzer, PDI was 0.131. The homogeneous solution added with trehalose was lyophilized, and after lyophilization, the lyophilized powder was reconstituted with sterilized ultrapure water. The particle size was 811.70 nm detected by a particle size analyzer, PDI was 0.313. The loss rate of elemene content after lyophilization was 82.64%.

Example 1

200 mg of β -elemene and 600 mg of soybean oil were weighed, dissolved in 2 mL of an organic solvent, and mixed by vortex to obtain an oil phase. The organic solvent is dichloromethane, or ethyl acetate or a mixture of dichloromethane and ethanol. 650 mg of human serum albumin was taken and dissolved in 18 mL of water for injection to obtain an aqueous solution of human serum albumin with a mass volume percentage of 3.6% (g/mL), that is, to obtain an aqueous phase. Under the action of water bath ultrasound, the oil phase was added dropwise to the aqueous phase, and sonicated by a probe for 8 min (250 W) to produce a primary emulsion. The obtained primary emulsion was transferred to a high-pressure homogenizer for high-pressure homogenization, the homogenization pressure was 1000 bar, and the homogenization times was 16 times. The organic solvent was removed by rotary evaporation after homogenization, and a 0.22 μm microporous membrane was used for filtration and sterilization to obtain an aqueous solution of elemene albumin nanoparticles.

Table 3 Investigation results of single factor for organic solvents

Number	Organic solvent	Particle size (d.nm)	PDI
1	Without organic solvent	142.1	0.094
2	Ethyl acetate	122.0	0.164
3	Dichloromethane	108.8	0.129
4	Dichloromethane/ethanol (4:1)	98.5	0.130

The experimental results showed that, no matter whether β -elemene and soybean oil were dissolved in organic solvents or not during the preparation process, β -elemene albumin formulations with suitable particle size were prepared. Among various organic solvents, the nanoparticles prepared by dichloromethane or a mixed solvent of dichloromethane and ethanol had smaller particle size.

Example 2

200 mg of β -elemene and 600 mg of soybean oil were weighed, dissolved in a mixed solvent of dichloromethane and ethanol (dichloromethane:ethanol=4:1, 2 mL), and mixed by vortex to obtain an oil phase with a volume of 2 mL. 650 mg of human serum albumin was taken and dissolved in water for injection to obtain an aqueous solution of human serum albumin with a mass volume percentage of 3.6% (g/mL), and then 100 mg of polyethylene glycol-12-hydroxystearate was added, to obtain an aqueous phase. Under the action of water bath ultrasound, the oil phase was added dropwise to the aqueous phase, and sonicated by a probe for 8 min (250 W) to produce a primary emulsion. The obtained primary emulsion was transferred to a high-pressure homogenizer for high-pressure homogenization, the homogenization pressure was 1000 bar, and the homogenization times was 16 times. After homogenization, the organic solvent was removed by rotary evaporation, and a 0.22 μ m microporous membrane was used for filtration and sterilization to obtain an aqueous solution of elemene albumin nanoparticles. The measured particle size was 116.4 nm, PDI was 0.118.

Example 3

200 mg of β -elemene and 400 mg of medium and long chain fatty glyceride were weighed, dissolved in a mixed solvent of dichloromethane and ethanol (dichloromethane:ethanol=4:1, 2 mL), and mixed by vortex, to obtain an oil phase with a volume of 2 mL. 900 mg of human serum albumin was taken and dissolved in 18 mL of water for injection to obtain an aqueous solution of human serum albumin with 5.0% of mass volume percentage (g/mL), that is, to obtain an aqueous phase. Under the action of water bath ultrasound, the oil phase was added dropwise to the aqueous phase, and sonicated by a probe for 8 min (250 W) to produce a primary emulsion. The obtained primary emulsion was transferred to a high-pressure homogenizer for high-pressure homogenization, the homogenization pressure was 1000 bar, and the homogenization times was 16 times. After homogenization, the organic solvent was removed by rotary evaporation, and a 0.22 μ m microporous membrane was used for filtration and sterilization to obtain an aqueous solution of elemene albumin nanoparticles. The measured particle size was 105.3 nm, PDI was 0.116.

Example 4

800 mg of β -elemene and 2400 mg of sesame oil were weighed, dissolved in a mixed solvent of dichloromethane and ethanol (dichloromethane:ethanol=4:1, 10 mL), and mixed by vortex to obtain an oil phase. 4000 mg of human serum albumin was taken and dissolved in 100 mL of water for injection to obtain an aqueous solution of human serum albumin with 4% of mass volume percentage (g/mL), that is, to obtain an aqueous phase. The oil phase was added dropwise to the aqueous phase, and a tissue masher was used at 10,000 rpm for shearing for 10 min to prepare a primary emulsion. The obtained primary emulsion was transferred to a high-pressure homogenizer for high-pressure homogenization, the homogenization pressure was 1000 bar, and the homogenization times was 20 times. After homogenization, the organic solvent was removed by rotary evaporation, and a 0.22 μ m microporous membrane was used for filtration and sterilization to obtain an aqueous solution of elemene albumin nanoparticles. The measured particle size was 121.3 nm, PDI was 0.133.

Example 5

800 mg of β -elemene and 2400 mg of olive oil were weighed, successively added to a dichloromethane/ethanol mixed solvent (dichloromethane:ethanol=4:1, 5 mL), and mixed by vortex to obtain an oil phase. 4000 mg of human serum albumin was taken and dissolved in water for injection to obtain an aqueous solution of human serum albumin with a mass volume percentage of 4% (g/mL), to obtain an aqueous phase. The oil phase was added dropwise to the aqueous phase, and a tissue masher was used at 12000 rpm for shearing for 10 min to prepare a primary emulsion. The obtained primary emulsion was transferred to a high-pressure homogenizer for high-pressure homogenization, the homogenization pressure was 1400 bar, and the homogenization times was 20 times. The organic solvent was removed from the homogenized system by rotary evaporation. The rotary-evaporated medicinal solution was filtered and sterilized via a 0.22 μ m microporous membrane to obtain an aqueous solution of elemene albumin nanoparticles. 3% sucrose was added and lyophilized, capped after lyophilization, and stored at 4°C. The lyophilized samples were taken and reconstituted in water for injection, and the measured particle size was 112.6 nm, PDI was 0.101.

Example 6

200 mg of β -elemene and 600 mg of soybean oil were weighed, dissolved in a mixed solvent of dichloromethane and ethanol (dichloromethane:ethanol=4:1, 10 mL), and mixed by vortex to obtain an oil phase. 4000 mg of human serum albumin was dissolved in water for injection to

obtain an aqueous solution of human serum albumin with 10% of mass volume percentage (g/mL), that is, to obtain an aqueous phase. Under the action of water bath ultrasound, the oil phase was added dropwise to the aqueous phase, and sonicated by a probe for 8 min (250 W) to produce a primary emulsion. The obtained primary emulsion was transferred to a high-pressure homogenizer for high-pressure homogenization, the homogenization pressure was 1000 bar, and the homogenization times was 16 times. After homogenization, the organic solvent was removed by rotary evaporation, and a 0.22 μm microporous membrane was used for filtration and sterilization to obtain an aqueous solution of elemene albumin nanoparticles. The measured particle size was 140.3 nm, PDI was 0.164.

Example 7

200 mg of β -elemene and 1000 mg of soybean oil were weighed, dissolved in a mixed solvent of dichloromethane and ethanol (dichloromethane:ethanol=4:1, 5 mL), and mixed by vortex to obtain an oil phase. 650 mg of human serum albumin was taken and dissolved in water for injection to obtain an aqueous solution of human serum albumin with 3.6% of mass volume percentage (g/mL), that is, to obtain an aqueous phase. Under the action of water bath ultrasound, the oil phase was added dropwise to the aqueous phase, and sonicated by a probe for 8 min (250 W) to produce a primary emulsion. The obtained primary emulsion was transferred to a high-pressure homogenizer for high-pressure homogenization, the homogenization pressure was 1000 bar, and the homogenization times was 16 times. After homogenization, the organic solvent was removed by rotary evaporation, and a 0.22 μm microporous membrane was used for filtration and sterilization to obtain an aqueous solution of elemene albumin nanoparticles. The measured particle size was 125.3 nm, PDI was 0.148.

Example 8

800 mg of β -elemene and 2400 mg of soybean oil were weighed, dissolved in a mixed solvent of dichloromethane and ethanol (dichloromethane:ethanol=4:1, 10 mL), and mixed by vortex to obtain an oil phase. 4000 mg of human serum albumin was dissolved in water for injection to obtain an aqueous solution of human serum albumin with 4% of mass volume percentage (g/mL), that is, to obtain an aqueous phase. The oil phase was added dropwise to the aqueous phase, and a tissue masher was used at 10000 rpm for shearing for 10 min to prepare a primary emulsion. The obtained primary emulsion was transferred to a high-pressure homogenizer for high-pressure homogenization, the homogenization pressure was 1000 bar, and the homogenization times was 20 times. After homogenization, the organic solvent was removed by rotary evaporation, and a 0.22 μm microporous membrane was used for filtration and

sterilization to obtain an aqueous solution of elemene albumin nanoparticles. The measured particle size was 119.3 nm, PDI was 0.143. The prepared nanoparticle solution was taken and configured to a drug concentration of 10 mg/mL, an appropriate amount of lyoprotectant was added, and the amount of the lyoprotectant was 3% (g/mL), and lyophilized. After lyophilization, water for injection was added to reconstitute, and the particle size was determined. The results are shown in the following table.

Table 4 Particle size of elemene albumin nanoparticles with different lyoprotectants after lyophilization

Lyoprotectant	None	Glucose	Sucrose	Trehalose	Mannitol	Lactose	Maltose
Particle size (nm)	187.4	160.7	153.5	167.4	172.1	166.3	164.5
PDI	0.233	0.165	0.125	0.167	0.133	0.178	0.196

The experimental results showed that, during the lyophilization process of elemene albumin nanoparticles, various lyoprotectants were added to obtain lyophilized powders with loose structure and uniform texture, which could be reconstituted by adding water, and the particle size after reconstitution did not exceed 200 nm. Wherein, the nanoparticles obtained by reconstituting the sample by lyophilizing with sucrose as the lyoprotectant had the smallest particle size.

Example 9

500 mg of β -elemene and 1200 mg of medium and long chain fatty glyceride were weighed, dissolved in a mixed solvent of dichloromethane and ethanol (dichloromethane:ethanol=4:1, 5 mL), and mixed by vortex, to obtain an oil phase. 1500 mg of human serum albumin was taken and dissolved in 50 mL of water for injection to obtain an aqueous solution of human serum albumin with 3% of mass volume percentage (g/mL), to obtain an aqueous phase. The oil phase was added dropwise to the aqueous phase, and sonicated by a probe for 8 min (250 W) to produce a primary emulsion. The obtained primary emulsion was transferred to a high-pressure homogenizer for high-pressure homogenization, the homogenization pressure was 1400 bar, and the homogenization times was 10 times. After homogenization, the organic solvent was removed by rotary evaporation, and a 0.22 μ m microporous membrane was used for filtration and sterilization, the filtrate was diluted and configured to a drug concentration of 15 mg/mL, 3% sucrose was added, lyophilized, capped, and stored at 4°C. The lyophilized sample was taken

and reconstituted in water for injection, and the particle size was determined. The measured particle size was 103.5 nm, PDI was 0.123.

Example 10

800 mg of β -elemene and 2400 mg of sesame oil were weighed, successively added to a dichloromethane/isopropanol mixed solvent (dichloromethane:isopropanol=4:1, 5 mL), and mixed by vortex to obtain an oil phase. 4000 mg of human serum albumin was dissolved in water for injection to obtain an aqueous solution of human serum albumin with a mass volume percentage of 4% (g/mL), to obtain an aqueous phase. The oil phase was added dropwise to the aqueous phase, and a tissue masher was used at 10,000 rpm for shearing for 10 min to prepare a primary emulsion. The obtained primary emulsion was transferred to a high-pressure homogenizer for high-pressure homogenization, the homogenization pressure was 1000 bar, and the homogenization times was 20 times. The organic solvent was removed from the homogenized system by rotary evaporation. The rotary-evaporated medicinal solution was filtered and sterilized via a 0.22 μ m microporous membrane to obtain an aqueous solution of elemene albumin nanoparticles. The aqueous solution was diluted and configured to a drug concentration of 10 mg/mL, and 3% sucrose was added for lyophilization, capped after lyophilization, and stored at 4°C. The lyophilized sample was taken and reconstituted in water for injection, and the measured particle size was 115.2 nm, PDI was 0.102.

Example 11

800 mg of β -elemene and 2400 mg of olive oil were weighed, successively added to a dichloromethane/tertiary butanol mixed solvent (dichloromethane:tertiary butanol=4:1, 5 mL), and mixed by vortex to obtain an oil phase. 4000 mg of human serum albumin was dissolved in water for injection to obtain an aqueous solution of human serum albumin with a mass volume percentage of 4% (g/mL), to obtain an aqueous phase. The oil phase was added dropwise to the aqueous phase, and a tissue masher was used at 10,000 rpm for shearing for 10 min to prepare a primary emulsion. The obtained primary emulsion was transferred to a high-pressure homogenizer for high-pressure homogenization, the homogenization pressure was 1000 bar, and the homogenization times was 20 times. The organic solvent was removed from the homogenized system by rotary evaporation. The rotary-evaporated medicinal solution was filtered and sterilized via a 0.22 μ m microporous membrane to obtain an aqueous solution of elemene albumin nanoparticles. The aqueous solution was diluted and configured to a drug concentration of 10 mg/mL, and 3% sucrose was added for lyophilization, capped after lyophilization, and stored at 4°C. The lyophilized sample was taken and reconstituted in water

for injection, and the measured particle size was 109.6 nm, PDI was 0.113.

Example 12

800 mg of β -elemene and 2400 mg of sesame oil were weighed, successively added to a
5 dichloromethane/tetrahydrofuran mixed solvent (dichloromethane:tetrahydrofuran=4:1, 5 mL),
and mixed by vortex to obtain an oil phase with a volume of 5 mL. 4000 mg of human serum
albumin was taken and dissolved in water for injection to obtain an aqueous solution of human
serum albumin with a mass volume percentage of 4% (g/mL), to obtain an aqueous phase. The
oil phase was added dropwise to the aqueous phase, and a tissue masher was used at 10,000 rpm
10 for shearing for 10 min to prepare a primary emulsion. The obtained primary emulsion was
transferred to a high-pressure homogenizer for high-pressure homogenization, the
homogenization pressure was 1000 bar, and the homogenization times was 20 times. The organic
solvent was removed from the homogenized system by rotary evaporation. The rotary-
evaporated medicinal solution was filtered and sterilized via a 0.22 μ m microporous membrane
15 to obtain an aqueous solution of elemene albumin nanoparticles. 3% trehalose was added for
lyophilization, capped after lyophilization, and stored at 4°C. The lyophilized sample was taken
and reconstituted in water for injection, and the measured particle size was 112.6 nm, PDI was
0.101.

Example 13

500 mg of β -elemene and 1200 mg of soybean oil were weighed, successively added to
isopropanol (5 mL) and mixed by vortex to obtain an oil phase. 1500 mg of bovine serum
albumin was taken and dissolved in water for injection to obtain an aqueous solution of bovine
serum albumin with a mass volume percentage of 3% (g/mL), to obtain an aqueous phase. The
25 oil phase was added dropwise to the aqueous phase, and sonicated by a probe for 8 min (250 W)
to produce a primary emulsion. The obtained primary emulsion was transferred to a high-
pressure homogenizer for high-pressure homogenization, the homogenization pressure was 1400
bar, and the homogenization times was 10 times. The organic solvent was removed from the
homogenized system by rotary evaporation. The rotary-evaporated medicinal solution was
30 filtered and sterilized via a 0.22 μ m microporous membrane. The filtrate was diluted and
configured to a drug concentration of 10 mg/mL, and 3% sucrose was added for lyophilization,
capped after lyophilization, and stored at 4°C. The lyophilized samples were reconstituted in
water for injection, and the measured particle size was 128.1 nm, PDI was 0.132.

Example 14

500 mg of β -elemene and 1200 mg of soybean oil were weighed, successively added to tertiary butanol (5 mL) and mixed by vortex to obtain an oil phase. 1500 mg of bovine serum albumin was taken and dissolved in water for injection to obtain an aqueous solution of bovine serum albumin with a mass volume percentage of 3% (g/mL), to obtain an aqueous phase. The oil phase was added dropwise to the aqueous phase, and sonicated by a probe for 8 min (250 W) to produce a primary emulsion. The obtained primary emulsion was transferred to a high-pressure homogenizer for high-pressure homogenization, the homogenization pressure was 1400 bar, and the homogenization times was 10 times. After homogenization, the organic solvent was removed by rotary evaporation. The rotary-evaporated medicinal solution was filtered and sterilized via a 0.22 μ m microporous membrane. The filtrate was diluted and configured to a drug concentration of 5 mg/mL, and 3% sucrose was added for lyophilization, capped after lyophilization, and stored at 4°C. The lyophilized sample was taken and reconstituted in water for injection, and the measured particle size was 116.8 nm, PDI was 0.113.

Example 15

500 mg of β -elemene and 1200 mg of sesame oil were weighed, successively added to a dichloromethane/isopropanol mixed solvent (dichloromethane:isopropanol=4:1, 5 mL), and mixed by vortex to obtain an oil phase. 1500 mg of bovine serum albumin was dissolved in water for injection to obtain an aqueous solution of bovine serum albumin with a mass volume percentage of 3% (g/mL), to obtain an aqueous phase. The oil phase was added dropwise to the aqueous phase, and sonicated by a probe for 8 min (250 W) to produce a primary emulsion. The obtained primary emulsion was transferred to a high-pressure homogenizer for high-pressure homogenization, the homogenization pressure was 1400 bar, and the homogenization times was 10 times. After homogenization, the organic solvent was removed by rotary evaporation. The rotary-evaporated medicinal solution was filtered and sterilized via a 0.22 μ m microporous membrane. The filtrate was diluted and configured to a drug concentration of 1 mg/mL, and 3% sucrose was added for lyophilization, capped after lyophilization, and stored at 4°C. The lyophilized sample was taken and reconstituted in water for injection, and the measured particle size was 121.8 nm, PDI was 0.111.

Example 16

500 mg of β -elemene and 1200 mg of soybean oil were weighed, successively added to a dichloromethane/tertiary butanol mixed solvent (dichloromethane:tertiary butanol=4:1, 5 mL), and mixed by vortex to obtain an oil phase. 1500 mg of bovine serum albumin was dissolved in

water for injection to obtain an aqueous solution of bovine serum albumin with a mass volume percentage of 3% (g/mL), to obtain an aqueous phase. The oil phase was added dropwise to the aqueous phase, and sonicated by a probe for 8 min (250 W) to produce a primary emulsion. The obtained primary emulsion was transferred to a high-pressure homogenizer for high-pressure homogenization, the homogenization pressure was 1400 bar, and the homogenization times was 10 times. After homogenization, the organic solvent was removed by rotary evaporation. The rotary-evaporated medicinal solution was filtered and sterilized via a 0.22 μ m microporous membrane. The filtrate was diluted and configured to a drug concentration of 10 mg/mL, and 3% sucrose was added for lyophilization, capped after lyophilization, and stored at 4°C. The lyophilized sample was taken and reconstituted in water for injection, and the measured particle size was 121.1 nm, PDI was 0.109.

Example 17

500 mg of β -elemene and 1200 mg of olive oil were weighed, successively added to a dichloromethane/ethanol mixed solvent (dichloromethane:ethanol=4:1, 5 mL), and mixed by vortex to obtain an oil phase. 1500 mg of bovine serum albumin was dissolved in water for injection to obtain an aqueous solution of bovine serum albumin with 3% of mass volume percentage (g/mL), to obtain an aqueous phase. The oil phase was added dropwise to the aqueous phase, and sonicated by a probe for 8 min (250 W) to produce a primary emulsion. The obtained primary emulsion was transferred to a high-pressure homogenizer for high-pressure homogenization, the homogenization pressure was 1400 bar, and the homogenization times was 10 times. The organic solvent was removed from the homogenized system by rotary evaporation. The rotary-evaporated medicinal solution was filtered and sterilized via a 0.22 μ m microporous membrane. The filtrate was diluted and configured to a drug concentration of 12 mg/mL, and 3% sucrose was added for lyophilization, capped after lyophilization, and stored at 4°C. The lyophilized sample was taken and reconstituted in water for injection, and the measured particle size was 109.7 nm, PDI was 0.105.

Example 18

500 mg of β -elemene and 1200 mg of soybean oil were weighed, successively added to a dichloromethane/tetrahydrofuran mixed solvent (dichloromethane:tetrahydrofuran=4:1, 5 mL), and mixed by vortex, to obtain an oil phase. 1500 mg of bovine serum albumin was dissolved in water for injection to obtain an aqueous solution of bovine serum albumin with 3% of mass volume percentage (g/mL), to obtain an aqueous phase. The oil phase was added dropwise to the aqueous phase, and sonicated by a probe for 8 min (250 W) to produce a primary emulsion.

The obtained primary emulsion was transferred to a high-pressure homogenizer for high-pressure homogenization, the homogenization pressure was 1400 bar, and the homogenization times was 10 times. The organic solvent was removed from the homogenized system by rotary evaporation. The rotary-evaporated medicinal solution was filtered and sterilized via a 0.22 μm microporous membrane. The filtrate was diluted and configured to a drug concentration of 10 mg/mL, and 3% sucrose was added for lyophilization, capped after lyophilization, and stored at 4°C. The lyophilized sample was taken and reconstituted in water for injection, and the measured particle size was 112.7 nm, PDI was 0.112.

Example 19

200 mg of elemene and 600 mg of soybean oil were weighed, dissolved in a mixed solvent of dichloromethane and ethanol (dichloromethane:ethanol=4:1, 2 mL), and mixed by vortex, to obtain an oil phase. 650 mg of human serum albumin was dissolved in 18 mL of 0.5% sodium chloride solution to obtain a solution of human serum albumin with a mass volume percentage (g/mL) of 3.6%. Under the action of water bath ultrasound, the oil phase was added dropwise to the aqueous phase, and sonicated by a probe for 8 min (250 W) to produce a primary emulsion. The obtained primary emulsion was transferred to a high-pressure homogenizer for high-pressure homogenization, the homogenization pressure was 1000 bar, and the homogenization times was 16 times. The organic solvent was removed from the homogenized system by rotary evaporation, and a 0.22 μm microporous membrane was used for filtration and sterilization to obtain an aqueous solution of elemene albumin nanoparticles. The measured particle size was 126.1 nm, PDI was 0.122.

Example 20

500 mg of β -elemene and 1500 mg of soybean oil were weighed, dissolved in 5 mL of dichloromethane/ethanol (dichloromethane:ethanol=8:1) mixed solvent to obtain an oil phase; and 2500 mg of human serum albumin was weighed and dissolved in 45 mL of water to obtain an aqueous phase. Under the action of ultrasound, the two phases were mixed evenly, and the obtained solution was transferred to a high-pressure homogenizer for high-pressure homogenization, at 400-500 bar for 4-6 times, and at 1300-1400 bar for 8-10 times. After homogenization, then the organic solvent was removed by rotary evaporation, the microporous membrane was used for filtration and sterilization, the bubbles in the formulation were removed by standing still for a moment, and a solution of elemene albumin nanoparticles was obtained. The measured particle size of the nanoparticles was 99.4 nm, PDI was 0.096. The obtained solution of elemene albumin nanoparticles was adjusted to a concentration of 10 mg/mL, 3%

sucrose was added, lyophilized, and reconstituted by adding ultrapure water, the particle size was measured to be 101.6 nm, and the PDI was 0.113.

Example 21

5 The corresponding β -elemene, soybean oil, MCT (medium chain triglyceride) and surfactants (HS-15, namely macrogol 15-hydroxystearate; Tween 80; Poloxamer P188) were weighed according to the component amounts listed in Table 5 and mixed to obtain an oil phase. 20% human serum albumin (converted by mass as shown in Table 5) was taken, diluted by adding ultrapure water to a total volume of 50 mL and mixed evenly, to obtain an aqueous
10 solution of human serum albumin, namely an aqueous phase. An appropriate amount of surfactant was added to the aqueous phase solution, and the aqueous phase solution was poured into the oil phase, and sheared with a mixer (IKA, T25) at 5,000 rpm for 1 min and at 10,000 rpm for 5 min to form a primary emulsion. The obtained primary emulsion was transferred to a high-pressure homogenizer (ATS, AH-1500) for high-pressure homogenization at a
15 homogenization pressure of 250-300 bar for 2 min, and at 1450-1550 bar for 6 min. After homogenization, the total volume of the homogeneous solution was measured with a measuring cylinder, trehalose of 10% (g/mL) was added, shaken well to be dissolved, and filtered with a 0.45 μ m microporous membrane to obtain a solution of elemene albumin nanoparticles. The particle size and PDI were detected, and the hemolysis value was determined. The specific
20 results are shown in Table 5.

Table 5

Serial number	β -elemene /g	Soybean oil /g	MCT /g	HSA/g	Soybean oil/API (g/g)	HSA/API (g/g)	Surfactant	Hemolysis value	Particle size (nm)	PDI
1	0.5	0.6	0.6g	1.5	1.2	3	0.1g HS-15	31.6	182.7	0.156
2	0.5	0.6	0.6g	1.5	1.2	3	0.1g Tween 80	22.2	178.3	0.135
3	0.5	0.6	0.6g	1.5	1.2	3	0.1g Poloxamer P188	53.3	232.5	0.143
4	1	0.4	0.1g	1.5	0.4	1.5	/	6.4	128.3	0.167
5	1	0.6	0.15g	1.5	0.6	1.5	/	9.1	127.6	0.191
6	1	0.4	0.1g	5	0.4	5	/	13.8	103.4	0.162
7	1	0.6	0.15g	5	0.6	5	/	7	101.2	0.092

Example 22

β -elemene and soybean oil were weighed according to the component amounts listed in Table 6 and mixed evenly, to obtain an oil phase. Human serum albumin (converted by mass as shown in Table 6) was measured, diluted to a total volume of 50 mL by adding ultrapure water and mixed evenly, to obtain an aqueous solution of human serum albumin, namely an aqueous phase. The aqueous phase was poured into the oil phase, and sheared with a mixer (IKA, T25) at 5,000 rpm for 1 min and at 10,000 rpm for 5 min to form a primary emulsion. The obtained primary emulsion was transferred to a high-pressure homogenizer (ATS, AH-1500) for high-pressure homogenization at a homogenization pressure of 250-300 bar for 2 min, and at 1450-1550 bar for 6min. After homogenization, the total volume of the homogeneous solution was measured with a measuring cylinder, trehalose was added at a mass/volume ratio of 10% (g/mL), shaken well to be dissolved, and filtered with a 0.45 μ m microporous membrane to obtain a solution of elemene albumin nanoparticles. The particle size and PDI were detected, and the hemolysis value was determined. The specific results are shown in Table 6.

Table 6

Serial number	β -elemene /g	Soybean oil /g	HSA /g	Soybean oil/API (g/g)	HSA/API (g/g)	Hemolysis value	Particle size (nm)	PDI
1	1	0.5	1.5	0.5	1.5	7.8	130.1	0.161
2	1	0.75	1.5	0.75	1.5	5.2	132	0.16
3	1	0.5	5	0.5	5	8.3	111	0.084
4	1	0.75	5	0.75	5	3.8	111.8	0.071
5	1	0.5	10	0.5	10	13.7	94.99	0.107
6	0.5	1.2	1.5	2.4	3	15.1	130.3	0.079
7	0.5	1.2	5	2.4	10	13.1	131.1	0.08
8	0.5	1.2	7.5	2.4	15	11.7	132.9	0.085
9	0.5	1.2	10	2.4	20	13.5	127.3	0.067
10	0.5	1.2	1.5	2.4	3	11	143.2	0.161
11	1	0.35	1.5	0.35	1.5	9	115.43	0.132

Example 23

3.33 g of β -elemene and 8g of soybean oil were weighed and mixed to obtain an oil phase. 50 mL (10g) of 20% human serum albumin (HSA) was measured, diluted evenly by adding an appropriate amount of ultrapure water, to obtain an aqueous solution of human serum albumin with 3% of mass volume percentage (g/mL), namely an aqueous phase. The aqueous phase

solution was poured into the oil phase, and the mixer (FLUKO, FA30/30F) was used for shearing at 10,000 rpm for 10 min to form a primary emulsion. The obtained primary emulsion was transferred to a micro-jet homogenizer (Microfluidizer, M-110-EH) for homogenization, at a homogenization pressure of 20,000 psi for 30 min. The homogenized solution was filtered by 0.45 μm and 0.22 μm PVDF plate filters, and the metered volume was adjusted, and different lyoprotectants (g/mL) corresponding to Table 7 was added respectively. The homogeneous solution added with the lyoprotectant was lyophilized, and after lyophilization, the lyophilized powder was reconstituted with sterilized ultrapure water. The particle size, PDI and hemolysis value were detected. The specific results are shown in Table 7.

Table 7

Serial number	Prescription	Protectant	Hemolysis value	Particle size	PDI
1	3.33 g β -elemene 8g Soybean oil 10g HSA	3% Sucrose	13.4	175.97	0.188
2		10% Sucrose	13.4	138.76	0.119
3		3% Trehalose	11.3	170.91	0.192
4		10% Trehalose	14	138.39	0.158

Example 24

3.33 g of β -elemene and 8 g of soybean oil were weighed, dissolved in an organic solvent, and mixed to obtain an oil phase. The organic solvent was a mixed solvent of 26.67 mL dichloromethane and 6.67 mL anhydrous ethanol. 50 mL (10g) of 20% human serum albumin (HSA) was measured, diluted evenly by adding an appropriate amount of ultrapure water, to obtain an aqueous solution of human serum albumin with a mass volume percentage of 3% (g/mL), namely an aqueous phase. The aqueous phase solution was poured into the oil phase, and the mixer (FLUKO, FA30/30F) was used for shearing at 10,000 rpm for 10 min to form a primary emulsion. The obtained primary emulsion was transferred to a micro-jet homogenizer (Microfluidizer, M-110-EH) for high-pressure homogenization, at a homogenization pressure of 20,000 psi for 30 min. After homogenization, the organic solvent was removed by a thin film evaporator from the organic solvent-containing emulsion, and the parameters were: temperature 30°C, vacuum degree 4000 pa, liquid feeding speed 100 rpm, scraper speed 50 rpm. The obtained emulsion was filtered by 0.45 μm and 0.22 μm PVDF plate filters, and the metered volume was adjusted, and 10% (g/mL) of trehalose was added as a lyoprotectant. The homogeneous solution added with the lyoprotectant was lyophilized, and after lyophilization, the lyophilized powder

was reconstituted with sterilized ultrapure water. The detected particle size was 76.28, the PDI was 0.254 and the hemolysis value was 16.3.

Example 25

5 β -elemene and soybean oil were weighed according to the component amounts listed in Table 8, and dissolved in an organic solvent, the organic solvent was a mixed solvent of 1 mL of ethanol and 4 mL of dichloromethane, and mixed to obtain an oil phase. An appropriate amount of 20% human serum albumin (HSA) (converted by mass as shown in Table 8) was measured, diluted to a total volume of 50 mL by adding ultrapure water, namely an aqueous
10 phase. The aqueous phase solution was poured into the oil phase, and sheared with a mixer (IKA, T25) at 5,000 rpm for 1 min and 10,000 rpm for 5 min to form a primary emulsion. The obtained primary emulsion was transferred to a high-pressure homogenizer (ATS, AH-1500) for high-pressure homogenization at a homogenization pressure of 250-300 bar for 2 min, and at 1450-1550 bar for 6min. The organic solvent was removed with a rotary evaporator at 35°C, 150 rpm
15 and a vacuum of 30 mbar. After completion, the total volume of the homogeneous solution was measured with a measuring cylinder, trehalose was added at a mass/volume ratio of 10% (g/mL), shaken well to be dissolved, and filtered with a 0.45 μ m microporous membrane to obtain a solution of elemene albumin nanoparticles.

Table 8

Serial number	β -elemene /g	Soybean oil /g	HSA /g	Soybean oil/API (g/g)	HSA/API (g/g)
1	0.5	0.1	1.5	0.2	3
2	0.5	1.2	1.5	2.4	3
3	0.5	2.5	1.5	5	3
4	0.5	5	1.5	10	3
5	0.5	1.2	2.5	2.4	5
6	0.5	1.2	5	2.4	10
7	0.5	1.2	10	2.4	20

20

The prepared samples were placed in an oven at 30°C, and sampled at 0h, 24h, and 48h to determine particle size, PDI and β -elemene content. The measurement results are shown in Table 9.

Table 9

Serial number	0h		24h			48h		
	Particle size	PDI	Particle size	PDI	Content change (%)	Particle size	PDI	Content change (%)
1	326.9	0.07	348.5	0.177	-10.14	317.8	0.094	-18.84
2	79.11	0.14	80.22	0.229	-3.19	77.04	0.164	-5.32
3	99.59	0.174	101.6	0.203	-2.56	101.7	0.183	-3.85
4	121.8	0.194	125.4	0.211	0	119	0.216	-1.16
5	88.17	0.187	97.63	0.237	-2.35	99.19	0.219	-4.71
6	82.47	0.259	99.54	0.362	-3.70	102.6	0.271	-6.17
7	77.82	0.283	102.9	0.261	-2.0	104	0.22	-8.69

Example 26

5 β -elemene and soybean oil were weighed according to the component amounts listed in Table 10, to obtain an oil phase. An appropriate amount of 20% human serum albumin (HSA) (converted by mass as shown in Table 10) was measured, diluted to a total volume of 50 mL by adding ultrapure water, namely an aqueous phase. The aqueous phase solution was poured into the oil phase, and sheared with a mixer (IKA, T25) at 5,000 rpm for 1 min and at 10,000 rpm for 5 min to form a primary emulsion. The obtained primary emulsion was transferred to a high-
 10 pressure homogenizer (ATS, AH-1500) for high-pressure homogenization at a homogenization pressure of 250-300 bar for 2 min, and at 1350-1450 bar for 6 min. After completion, the total volume of the homogeneous solution was measured with a measuring cylinder, trehalose was added at the mass/volume ratio of (g/mL) shown in Table 10, shaken well to be dissolved, and filtered with a 0.45 μ m microporous membrane to obtain a solution of elemene albumin
 15 nanoparticles.

Table 10

Serial number	β -elemene/g	Soybean oil /g	HSA /g	Soybean oil/API (g/g)	HSA/API (g/g)	Protectant (g/mL)
1	1	1	1.5	1	1.5	10% Trehalose
2	1	1.5	1.5	1.5	1.5	10% Trehalose
3	1	2	1.5	2	1.5	10% Trehalose

4	1	2.5	1.5	2.5	1.5	10% Trehalose
5	1	3	1.5	3	1.5	10% Trehalose
6	1	0.5	0.5	0.5	0.5	10% Trehalose
7	1	0.5	1	0.5	1	10% Trehalose
8	1	0.5	1.5	0.5	1.5	10% Trehalose
9	1	0.5	2	0.5	2	10% Trehalose
10	1	0.5	2.5	0.5	2.5	10% Trehalose
11	1	0.5	1.5	0.5	1.5	20% Trehalose
12	1	0.5	1.5	0.5	1.5	15% Trehalose
13	1	0.5	1.5	0.5	1.5	5% Trehalose

The prepared samples were placed in an oven at 30°C, and sampled at 0h, 24h, and 48h to determine particle size, PDI and β -elemene content. The measurement results are shown in Table 11.

Table 11

Serial number	0h		24h			48h		
	Particle size	PDI	Particle size	PDI	Content change (%)	Particle size	PDI	Content change (%)
1	140.3	0.127	143.7	0.147	-3.60	135.5	0.165	-5.76
2	146.7	0.204	154.2	0.226	-3.64	149.5	0.208	-4.24
3	141.7	0.213	144.6	0.251	-1.83	141.4	0.265	-2.44
4	131.8	0.192	135.9	0.229	-3.05	132.3	0.222	-1.83
5	105.8	0.31	117.4	0.36	-1.24	106.8	0.285	-1.24
6	152.3	0.188	154.5	0.122	-3.59	151.3	0.13	-3.59
7	143.2	0.108	144.5	0.139	-3.85	142.2	0.129	-5.13
8	123.2	0.059	125.6	0.121	-4.19	121.6	0.089	-5.39
9	117.3	0.15	117.3	0.132	-4.79	116.8	0.126	-7.19
10	154.1	0.145	159.2	0.185	-4.17	163.5	0.167	-6.55
11	133.7	0.095	134.1	0.122	-4.61	131.5	0.118	-5.93
12	139.9	0.097	134	0.115	-5.63	128.4	0.194	-6.88
13	130.6	0.132	122.2	0.115	-4.14	128.7	0.159	-5.33

Example 27

2g of β -elemene and 1.5g of soybean oil were weighed and mixed to obtain an oil phase. 45 mL (9 g) of 20% human serum albumin was measured, namely an aqueous phase. The aqueous phase solution was poured into the oil phase, and sheared with a mixer (IKA, T25) at 5,000 rpm for 1 min and 10,000 rpm for 5 min to form a primary emulsion. The obtained primary emulsion was transferred to a high-pressure homogenizer (ATS, AH-1500) for high-pressure homogenization at a homogenization pressure of 250-300 bar for 2 min, and at 1450-1550 bar for 6 min, and the homogeneous solution was collected. The pipeline of the homogenizer was flushed with an aqueous solution of trehalose (25 mL of water + 10 g of trehalose) to replace the homogeneous solution remaining in the homogenizer, and then the pipeline of the homogenizer was flushed with 30 mL of ultrapure water to replace the residual aqueous solution of trehalose. The trehalose aqueous solution was combined with the homogeneous solution, mixed, and filtered through a 0.22 μ m sterile filter membrane. The particle size was 106.4 nm detected by a particle size analyzer, PDI was 0.08.

Example 28

Table 13

Serial number	Prescription (g)			Soybean/API (g/g)	HSA/API (g/g)	Protectant (g/mL)
	API	Soybean oil	HSA			
1	2	2	10	1	5	10% Trehalose
2	2	3	10	1.5	5	10% Trehalose
3	2	4	10	2	5	10% Trehalose
4	2	5	10	2.5	5	10% Trehalose
5	3	4.5	9	1.5	3	10% Trehalose

β -elemene and soybean oil were weighed as shown in Table 13 and mixed to obtain an oil phase. 20% human serum albumin (50 mL, 10 g) was measured, namely an aqueous phase. The aqueous phase solution was poured into the oil phase, and sheared with a mixer (IKA, T25) at 5,000 rpm for 1 min and at 10,000 rpm for 5 min to form a primary emulsion. The obtained solution was transferred to a high-pressure homogenizer (ATS, AH-1500) for high-pressure

homogenization at a homogenization pressure of 250-300 bar for 2 min, and at 1550-1550 bar for 6 min. 10 g of trehalose was weighed and dissolved to 50 mL by adding pure water (in which sample No. 5 was added with 15 g of trehalose and dissolved to 105 mL by adding pure water). The pipeline of the homogenizer was flushed with the aqueous solution of trehalose to replace the homogeneous solution remaining in the homogenizer, and the residual homogeneous solution was combined with the homogeneous solution, mixed, and filtered through a 0.45 μ m sterile filter membrane. Each sample was tested for hemolysis value. And the sample was divided into EP tubes, stored in a thermotank at 30°C, and the samples were taken at 0h, 24h, and 48h to determine particle size, PDI and β -elemene content. The results are shown in Table 14.

Table 14

Serial number	0h		24h			48h			Hemolysis value
	Particle size	PDI	Particle size	PDI	Content change %	Particle size	PDI	Content change %	
1	102.9	0.124	99.7	0.090	-4.22%	99.8	0.085	-5.42%	6.0
2	113.7	0.112	108.2	0.095	-1.81%	107.9	0.082	-4.82%	9.3
3	120.9	0.034	128.9	0.052	-2.33%	128.3	0.106	-4.07%	17.4
4	126.4	0.070	126.4	0.168	-2.33%	132.4	0.138	-2.91%	16.5
5	111.8	0.089	111.0	0.108	-5.52%	109.1	0.066	-6.08%	10.2

Example 29

2g of β -elemene and 1.5g of soybean oil were weighed and mixed to obtain an oil phase. 50 mL (10 g) of 20% human serum albumin was measured, namely an aqueous phase. The aqueous phase solution was poured into the oil phase, and sheared with a mixer (IKA, T25) at 5,000 rpm for 1 min and at 10,000 rpm for 5 min to form a primary emulsion. And 5 mL of water for injection was used for cleaning the shearing head, and combined with the primary emulsion. The obtained solution was transferred to a high-pressure homogenizer (ATS, AH-1500) for high-pressure homogenization at a homogenization pressure of 250-300 bar for 2 min, and at 1500-1550 bar for 6 min, and the homogeneous solution was collected. The pipeline of the homogenizer was flushed with 22.5 mL of an aqueous solution of trehalose to replace the homogeneous solution remaining in the homogenizer, and then the pipeline of the homogenizer was flushed with 22.5 mL of ultrapure water to replace the residual aqueous solution of trehalose.

The trehalose solution was combined with the homogeneous solution, mixed, and filtered via a 0.22 μm sterile filter membrane. The sample was divided into EP tubes, stored in refrigerator at 2-8°C, and the particle size and PDI were detected by sampling on the 0th day and second month, respectively. The results are shown in Table 12.

Table 12

Time	Particle size	PDI
Day 0	87.93	0.107
2nd month	89.39	0.119

Example 30 Drug Efficacy Study

Preparation of sample:

4.5 g of β -elemene and 10.8 g of soybean oil were weighed and mixed to obtain an oil phase. 4 mL of ethanol and 16 mL of dichloromethane were measured, and mixed evenly to obtain organic phase A. The oil phase was added to the organic phase and the metered volume was adjusted to 36 mL to obtain organic phase B.

13.5 g of human serum albumin was weighed and added to 405 mL of purified water, stirred to be dissolved. The organic phase B was added to the aqueous phase, and stirred with a high-speed shearing machine at gear A for 2 min, and then at gear B for 1 min to obtain primary emulsion A. The primary emulsion A was divided into 3 parts, and treated by a cell disrupter at 250w for 8 min to obtain primary emulsion B. The primary emulsion B was added into a high-pressure homogenizer, firstly at 200 bar for 3 cycles, and then at 1300 bar for 5 cycles to obtain albumin nanoparticles. The above nanoparticles were concentrated under reduced pressure at 37°C for 1 h, added with 12.30 g of 3% sucrose, filtered at 0.22 μm , and lyophilized. The particle size was 81.03 nm, PDI was 0.174.

Experimental Methods:

Establishment of a human-derived brain tumor xenograft mouse model: 50 nude mice were anesthetized with 2.5% isoflurane, a small hole (<0.5 mm) was drilled in the skull of the nude mice, and cells expressing the luminescent reporter gene (5×10^4 U87MGR1 Human glioma cells) were injected into the left forehead (2.5 mm lateral and 0.5 mm anterior bregma, 2.5 mm deep).

In vivo bioluminescence imaging assay method: nude mice received a single subcutaneous injection of D-luciferin solution (120 mg/kg). 10 min after injection of fluorescein, light images

of animals were taken with dark multicolor illumination during an exposure time of 1-5 min (for bioluminescence imaging of Gluc, a single intravenous injection of coelenterazine (20 mg/kg, at total volume of 150 μ L) was given). Intratumoral luciferase activity was obtained by recording photon counts with a CCD camera over 5 min without lighting. Image intensity was quantitatively analyzed using Perkin Elmer's Living image software 4.3.1.

One week after inoculation, in vivo bioluminescence imaging was performed to evaluate the tumor volume in nude mice. 32 nude mice with similar tumor volume were selected and randomly divided into 8 mice (half male and half female)/group, and the dose were administered by tail vein injection for 3 weeks, 3 times/week:

- 1) model group (control) (0.1% DMSO prepared by PBS), n=8;
- 2) 50 mg/kg temozolomide (TMZ), n=8;
- 3) 50 mg/kg Ailineng (ELE mic), n=8;
- 4) 50 mg/kg (Alb), n=8;

Tumor volume was measured weekly.

During the experiment, the animals were weighed weekly to monitor their healthy conditions.

Dosing was discontinued after two weeks of administration, followed by an observation for three weeks. The survival and tumor volume of mice in each group were recorded using Graphpad software to generate Kaplan-Meier plots. The tumor growth curve of the mouse orthotopic tumor model is shown in Figure 8 (the tumor-bearing nude mice were treated with Alb formulation, 3 times/week $\times$ 2 weeks (n=8, half male and half female); compared with temozolomide (TMZ), $p < 0.01$), the survival curve of the mouse orthotopic tumor model is shown in Figure 9 (the tumor-bearing nude mice were treated with Alb, 3 times/week $\times$ 2 weeks (n=8, half male and half female); compared with temozolomide (TMZ), $p < 0.01$).

Example 31 Safety Study

Preparation of sample:

Elemene injection (trade name: Ailineng): CSPC Yuanda (Dalian) Pharmaceutical Co., Ltd., batch number: 17050302)

β -elemene micelle formulation:

17.08 g of RH40 was weighed and heated at 60°C to be dissolved. 2 mL of β -elemene was added, heated and stirred at 60°C for 5 min. 14.40 g of propylene glycol and 38.72 g of purified water were weighed and mixed, then heated at 60°C for 10min. The aqueous solution of propylene glycol was added to the RH40 solution containing β -elemene, and stirred in a water bath at 60°C for 30 min to obtain β -elemene micelles. The above micelles were filtered at 0.22

μm, and divided into 5 mL/per formulation.

β-elemene albumin formulation:

2.5001 g of β-elemene and 6.001 g of soybean oil were weighed and mixed for later use. 2 mL of ethanol and 8 mL of dichloromethane were measured, and mixed evenly for later use. The volume of the oil phase was adjusted to 12 mL by the organic phase for later use.

4.5015 g of human serum albumin was weighed and added to 135 mL of purified water, stirred to be dissolved. The organic phase was added to the aqueous phase, and stirred with a high-speed shearing machine at gear A for 2 min, and then at gear B for 0.5 min to obtain primary emulsion A. The primary emulsion A was divided into 3 parts, and treated by a cell disrupter at 250W for 8 min to obtain primary emulsion B. The primary emulsion B was added into a high-pressure homogenizer, firstly at 200 bar for 3 cycles, and then at 1300-1400 bar for 5 cycles to obtain albumin nanoparticles. The above nanoparticles were concentrated under a reduced pressure at 37°C for 1 h, added with 1.20 g of sucrose, filtered at 0.22 μm, and lyophilized.

Solution Preparation:

The three formulations were prepared into 6 mg/mL solution using normal saline.

Ailineng: 7.5 mL of 20 mg/mL commercially available Ailineng was taken and diluted into 25 mL of 6 mg/mL sample solution with 17.5 mL of normal saline.

β-elemene micelle formulation: 5.91 mL of 25.41 mg/mL β-elemene micelle formulation was taken and diluted into 25 mL of 6 mg/mL sample solution with 19.09 mL of normal saline.

β-elemene albumin formulation: 12 β-elemene micelle formulations were taken, 2.01 mL of normal saline was added to each one, then mixed to obtain 24 mL of 6 mg/mL sample solution.

Test Methods:

Male KM mice, 20-25 g/mouse, were randomly divided into 13 groups according to body weight, with 7 mice in each group. The mice were grouped according to the prescribed dose by weight. The dosage of each sample was set as 100 mg/kg, 130 mg/kg, 170 mg/kg and 220 mg/kg, and another blank group was set. The mice were injected through tail vein, and the site of administration, reaction after administration and death conditions were observed after administration. The results of the study are shown in Table 15.

Table 15 Safety studies of different samples

Dosage Sample	100 mg/kg	130 mg/kg	170 mg/kg	220 mg/kg
Ailineng	reacted intensely; respiratory distress; motionless	reacted intensely; respiratory distress; lethargic, motionless; started to recover in about 5 min	3 died; reacted intensely; respiratory distress; lethargic; motionless, occasionally ataxia	6 died; respiratory distress, extremely low physical activity.
β -elemene micelle formulation	much less stresses than Ailineng group; a few with respiratory distress	much less stresses than Ailineng group; respiratory distress, activity decreasing, and started to recover in about 2 min	reacted intensely; respiratory distress; lethargic, motionless	3 died; respiratory distress; motionless
β -elemene albumin formulation	no obvious discomfort	no obvious discomfort, occasional mild respiratory distress	no obvious discomfort, occasional mild respiratory distress	no obvious discomfort, occasional respiratory distress

CLAIMS

1. A pharmaceutical composition comprising elemene, an oil for injection and a protein carrier.

5

2. The pharmaceutical composition according to claim 1, wherein the pharmaceutical composition is in the form of particles, and the particle size of the particles is less than 180 nm, preferably 50-180 nm.

10

3. The pharmaceutical composition according to claim 2, wherein the particle size of the particles is 70-170 nm.

4. The pharmaceutical composition according to claim 3, wherein the particle size of the particles is 70-150 nm.

15

5. The pharmaceutical composition according to claim 1, wherein the elemene is selected from one or more of α -elemene, β -elemene, γ -elemene and δ -elemene.

6. The pharmaceutical composition according to claim 5, wherein the elemene is β -elemene.

20

7. The pharmaceutical composition according to claim 1, wherein the protein carrier is albumin, and the albumin is selected from one or both of human serum albumin and bovine serum albumin.

25

8. The pharmaceutical composition according to claim 7, wherein the protein carrier is human serum albumin.

9. The pharmaceutical composition according to claim 1, wherein the weight ratio of the albumin carrier to the elemene is 0.5-20.

30

10. The pharmaceutical composition according to claim 9, wherein the weight ratio of the albumin carrier to the elemene is 0.5-10.

11. The pharmaceutical composition according to claim 1, wherein the weight ratio of the oil for injection and the elemene is 0.35-10.

12. The pharmaceutical composition according to claim 11, wherein the weight ratio of the
5 oil for injection and the elemene is 0.5-10.

13. The pharmaceutical composition according to any one of claims 1-12, wherein the oil for injection is one or more of soybean oil, safflower oil, cottonseed oil, corn oil, sunflower oil, peanut oil, olive oil, sesame oil, camellia oil, medium and long chain fatty glyceride or medium
10 chain triglyceride.

14. The pharmaceutical composition according to any one of claims 1-13, further comprising a lyoprotectant.

15 15. The pharmaceutical composition according to claim 14, wherein the lyoprotectant is selected from one or more of glucose, sucrose, maltose, lactose, mannose, trehalose, glycine and dextran.

16. The pharmaceutical composition according to claim 15, wherein the lyoprotectant is
20 trehalose or sucrose.

17. The pharmaceutical composition according to claims 14-16, wherein the ratio of the weight of the lyoprotectant to the volume of the solution of the pharmaceutical composition is 3:100-20:100 g/mL.
25

18. The pharmaceutical composition according to any one of claims 1-16, further comprising one or more of isoosmotic adjusting agent, antioxidant, preservative, and pH regulator.

30 19. The pharmaceutical composition according to claim 18, wherein the isoosmotic adjusting agent is one or more of glycerin, sorbitol, mannitol or glucose.

20. The pharmaceutical composition according to claim 18, wherein the pH regulator is one or more of sodium hydroxide, sodium citrate, citric acid, phosphoric acid, acetic acid or hydrochloric acid.

5 21. The pharmaceutical composition according to claim 18, wherein the preservative is selected from one or more of hydroxybenzene alkyl esters, benzoic acid, sodium benzoate, sorbic acid, chlorhexidine acetate, benzalkonium bromide.

10 22. The pharmaceutical composition according to claim 18, the antioxidant is selected from one or more of sodium sulfite, sodium bisulfite, sodium metabisulfite, sodium thiosulfate, ascorbic acid, butylated hydroxyanisole, 2,6-di-tert-butylated hydroxytoluene, and vitamin E.

23. The pharmaceutical composition according to any one of claims 1-22, wherein no organic solvent is included during preparation process thereof.

15

24. The pharmaceutical composition according to claim 23, wherein the pharmaceutical composition is prepared in the form of particles, and the particle size of the particles is less than 180 nm.

20 25. The pharmaceutical composition according to claim 24, wherein the particle size of the particles is 70-150 nm.

26. The pharmaceutical composition according to any one of claim 23-25, wherein in the pharmaceutical composition, the weight ratio of the soybean oil to the elemene is about 0.35-3; 25 the weight ratio of the human serum albumin to the elemene is about 0.5-20.

27. The pharmaceutical composition according to claim 26, wherein in the pharmaceutical composition, the weight ratio of the soybean oil to the elemene is about 0.5-3; the weight ratio of the human serum albumin to the elemene is about 0.5-2.5.

30

28. The pharmaceutical composition according to claim 26, wherein in the pharmaceutical composition, the weight ratio of the soybean oil to the elemene is about 0.35-1.5, preferably 0.35-0.75; the weight ratio of the human serum albumin to the elemene is about 1.5-10.

29. The pharmaceutical composition according to claim 28, wherein in the pharmaceutical composition, the weight ratio of the soybean oil to the elemene is about 0.5; the weight ratio of the human serum albumin to the elemene is about 1.5.

5 30. The pharmaceutical composition according to claim 28, wherein in the pharmaceutical composition, the weight ratio of the soybean oil to the elemene is about 0.75; the weight ratio of the human serum albumin to the elemene is about 5.

10 31. The pharmaceutical composition according to claim 28, wherein in the pharmaceutical composition, the weight ratio of the soybean oil to the elemene is about 1.5; the weight ratio of the human serum albumin to the elemene is about 3-5.

15 32. The pharmaceutical composition according to any one of claims 1-22, wherein organic solvent is included during preparation process thereof.

20 33. The pharmaceutical composition according to claim 32, wherein the organic solvent is selected from one or more of chloroform, dichloromethane, tertiary butanol, isopropanol, ethyl acetate, ethanol, tetrahydrofuran, dioxane, acetonitrile, acetone, dimethyl sulfoxide, dimethylformamide, and methylpyrrolidone.

34. The pharmaceutical composition according to claim 33, wherein the organic solvent is a mixed solvent of dichloromethane and ethanol.

25 35. The pharmaceutical composition according to claim 34, wherein the volume ratio of dichloromethane to ethanol is 1:1-8.

36. The pharmaceutical composition according to claim 35, wherein the volume ratio of dichloromethane to ethanol is 1:4.

30 37. The pharmaceutical composition according to any one of claims 33-36, wherein the particle size of the particles is about 70-150 nm.

38. The pharmaceutical composition according to claim 37, wherein in the pharmaceutical composition, the weight ratio of the soybean oil to the elemene is about 2.4-10; the weight ratio of the human serum albumin to the elemene is about 0.5-10.

5 39. The pharmaceutical composition according to claim 38, wherein in the pharmaceutical composition, the weight ratio of the soybean oil to the elemene is about 2.4; the weight ratio of the human serum albumin to the elemene is about 3.

10 40. A method for preparing the pharmaceutical composition of any one of claims 1-39, comprising the steps of:

(1) mixing elemene and an oil for injection evenly, optionally adding an organic solvent, to obtain a solution of the elemene and the oil for injection; dissolving a protein carrier in water to obtain a protein carrier solution;

(2) mixing the two solutions obtained in step (1) to form an emulsion;

15 (3) homogenizing the emulsion in step (2) under a high pressure, and optionally evaporating the emulsion under a reduced pressure to remove the organic solvent, to obtain a nanoparticle solution.

20 41. The method according to claim 40, wherein the organic solvent is used in step (1), preferably dissolving the elemene in the organic solvent and then mixing with the oil for injection.

25 42. The method according to claim 41, wherein the oil for injection is soybean oil, the albumin carrier is human serum albumin; the weight ratio of the soybean oil to the elemene is about 2.4-10; the weight ratio of the human serum albumin to the elemene is about 0.5-10.

43. The method according to claim 40, further comprising a step of lyophilizing the obtained solution of protein carrier nanoparticles.

30 44. The method according to claim 40, wherein, in step (2), a shearing or ultrasonic method is used for mixing; and in step (3), a high-pressure homogenization or micro-jet homogenization method is used for homogenization.

45. Use of the pharmaceutical composition of any one of claims 1-39 in the preparation of a medicament for the prevention or treatment of cancer.

46. The use according to claim 45, the cancer is adrenocortical carcinoma, cryptogenic
5 myeloid metaplasia, AIDS-related cancer, anal cancer, appendiceal cancer, astrocytoma, basal
cell carcinoma , cholangiocarcinoma, bladder cancer, bone cancer, glioma, ependymoma,
oligodendroglioma, meningioma, craniopharyngioma, hemangioblastoma, medulloblastoma,
neuroectodermal tumor, visual pathway and hypothalamic glioma and glioblastoma, breast
cancer, bronchial adenoma, carcinoid tumor, central nervous system lymphoma, cervical cancer,
10 colon cancer, colorectal cancer, chronic myeloproliferative disorder, endometrial cancer,
ependymoma, esophagus cancer, Ewing's tumor family, eye cancer, gallbladder cancer,
gastrointestinal carcinoid tumor, gastrointestinal stromal tumor, germ cell carcinoma, gestational
trophoblastic tumor, head and neck cancer, liver cancer, laryngeal cancer, leukemia, lip and oral
cancer, lung cancer, lymphoma, medulloblastoma, melanoma, mesothelioma, metastatic neck
15 squamous cell carcinoma, multiple endocrine neoplasia syndrome, myelodysplastic syndrome,
myelodysplastic/myeloproliferative disease, carcinoma of nasal cavity and sinuses,
nasopharyngeal cancer, neuroblastoma, neuroendocrine cancer, oropharyngeal cancer, brain
tumor, bone metastasis cancer, gastric cancer, intestinal cancer, esophageal cancer, ovarian
cancer, pancreatic cancer, breast cancer, skin cancer, parathyroid carcinoma, penile carcinoma,
20 peritoneal cancer, pharyngeal carcinoma, pheochromocytoma, pineoblastoma and supratentorial
primitive neuroectodermal tumor, pituitary tumor, pleuropulmonary blastoma, lymphoma,
primary central nervous system lymphoma, pulmonary lymphangiomyomatosis, rectal cancer,
kidney cancer, renal pelvis and ureter cancer, rhabdomyosarcoma, salivary gland cancer, skin
cancer, small intestine cancer, squamous epithelial cell carcinoma, testicular cancer, throat
25 cancer, thymoma and thymic carcinoma, thyroid cancer, urethral cancer, or vaginal cancer.

47. The use according to claim 46, wherein the cancer is lung cancer, liver cancer,
esophageal cancer, nasopharyngeal cancer, brain tumor, bone metastasis cancer, gastric cancer,
intestinal cancer, uterine cancer, cervical cancer, germ cell carcinoma, endometrial cancer,
30 gestational trophoblastic tumor, breast cancer, skin cancer, lymphoma, leukemia, or malignant
melanoma.

48. The use according to claim 47, wherein the cancer is brain tumor, preferably glioma, brain stem glioma, cerebellar or cerebral astrocytoma, malignant glioma, ependymoma, oligodendroglioma, meningioma, craniopharyngioma, hemangioblastoma, medulloblastoma, visual pathway and hypothalamic glioma or glioblastoma.

5

49. The use according to claim 48, wherein the cerebellar or cerebral astrocytoma is fibrous cell astrocytoma or diffuse astrocytoma or anaplastic (malignant) astrocytoma.

50. A method of treating a human disease comprising administering to a subject in need thereof a therapeutically effective amount of the pharmaceutical composition of any one of claims 1-37.

51. The method according to claim 50, wherein the pharmaceutical composition is administered by parenteral, by inhalation, intraperitoneal, intravesical, intramuscular, intravenous, intratracheal, subcutaneous, intraocular, intrathecal, transdermal, rectal or vaginal.

52. The method according to claim 51, wherein the administration is intravenous.

53. A pharmaceutical formulation, comprising the pharmaceutical composition of any one of claims 1-39.

54. The pharmaceutical formulation according to claim 53, wherein the pharmaceutical formulation is a pharmaceutical formulation for intravenous administration, a pharmaceutical formulation for parenteral administration, a pharmaceutical formulation for gastrointestinal administration, a pharmaceutical formulation for respiratory administration, a pharmaceutical formulation for vaginal administration or other suitable pharmaceutical formulation; the pharmaceutical formulation for respiratory administration is aerosol.

55. The pharmaceutical formulation according to claim 53, wherein the pharmaceutical formulation is a solid formulation, a liquid formulation or a gas formulation.

56. The pharmaceutical formulation according to claim 53, wherein the liquid formulation is a liquid formulation for injection, comprising elemene and a pharmaceutically acceptable carrier, wherein the pharmaceutically acceptable carrier comprises albumin and soybean oil.

57. The pharmaceutical formulation according to claim 55, the pharmaceutical formulation is a granule formulation, the particle size of the granule formulation is 70-150 nm, wherein in the composition, the weight ratio of the albumin to the elemene is 0.5-20, and the weight ratio of the oil for injection to the elemene is 0.35-10.

58. The pharmaceutical formulation according to claim 55, wherein the liquid formulation is a stable aqueous suspension reconstituted from a sterile lyophilized powder.

59. The pharmaceutical formulation according to claim 53, further comprising other drugs, including anticancer drugs.

60. The pharmaceutical formulation according to claim 59, wherein the anticancer drug comprises temozolomide, paclitaxel, docetaxel, taxane, gemcitabine, an anti-VEGF drug, or a PD-1 antibody drug.

61. The pharmaceutical formulation according to claim 60, wherein the anti-VEGF drug comprises AVASTIN, ranibizumab, aflibercept, or conbercept.

62. The pharmaceutical formulation according to claim 56, wherein the PD-1 antibody drug comprises nivolumab, pembrolizumab, toripalimab, sintilimab, cemiplimab, atezolizumab, avelumab, or durvalumab.

63. A sealed container comprising the pharmaceutical composition of any one of claims 1-39 or the pharmaceutical formulation of any one of claims 53-62.

64. The sealed container according to claim 63, wherein the sealed container is a unit-dose container or a multi-dose container.

65. The sealed container according to claim 63, wherein the pharmaceutical composition is a liquid composition or a dry solid composition.

66. The sealed container according to claim 63, wherein the pharmaceutical composition is lyophilized.

67. The sealed container according to claim 63, wherein the pharmaceutical composition is sterile.

5 68. The sealed container according to claim 63, wherein the sealed container is a prefilled syringe.