

Caspase imaging probes

The present invention relates to molecular probes (substrates) that allow the observation of the catalytic activity of individual proteolytic enzymes or groups of proteolytic enzymes in *in vitro* assays, in cells or in multicellular organisms. The invention furthermore relates to methods for the synthesis and the design of such probes (substrates).

Background of the Invention

Proteolytic enzymes (proteases) cleave or degrade other enzymes or peptides in- and outside of the living cell. Proteases are involved in a multitude of vital processes, many of which are critical in cellular signalling and tissue homeostasis. Aberrant or enhanced activity of proteases is associated with a variety of diseases including cancer, osteoarthritis, artherosclerosis, inflammation and many others. Since proteolytic activity has to remain under stringent control in living systems many proteases are expressed as inactive precursor proteins (zymogens) which are activated by controlled proteolytic cleavage. Additional control of proteolytic activity results from endogenous inhibitors that bind to and thereby inactivate catalytically active form of the enzyme. In view of this stringent regulation the investigation of protease function in cellular or physiological events requires the monitoring of protease activity rather than the monitoring of protease expression alone. Consequently, a variety of activity based chemical probes have been proposed in the literature. Commonly applied protease probes generate a detectable signal either (i) through enzymatic cleavage of a peptide bond leading to a change of the spectroscopic properties of a reporter system or (ii) by covalent attachment of a mechanism based inhibitor to the protease of interest. The localization and quantitative investigation of the activity and inhibition of a specific protease or a group of proteases (e.g. in cell-based assays or whole-animal imaging experiments) require the development of imaging probes that (i) reach the physiologically relevant *locus* of protease action (e.g. the cytosol of a cell or a specific organ in whole animal imaging) and (ii) are selective for the

desired protease or a group of proteases. The generation of protease selective probes has imposed a considerable challenge for the field. The present invention relates (i) to novel selective probes for cysteine proteases preferably from the caspases subfamily, (ii) to the application of these probes for in-vitro assays, in
5 cells or in multicellular organisms (e.g. by the means of molecular imaging) and (iii) to methods for the synthesis and the design of such probes.

Within recent years several molecular imaging technologies (optical and non-optical) have become more and more important for the non-invasive visualisation
10 of specific molecular targets and pathways *in vivo*. Since the information content of any image signal is primarily a function of internal contrast, the development of internally quenched imaging probes that are activable upon enzymatic reaction (e.g. cleavage of a peptide bond) has been commonly applied to image and localize catalytically active proteases. The generation of probes that are selective
15 for individual proteases and exhibit the ability to reach the *locus* of protease action *in vivo* has rarely been achieved with conventional approaches. Medicinal chemists in the pharmaceutical industry face related challenges in the development of drugs with appropriate pharmacokinetic properties and appropriate specificity for a given target. In our invention we have devised a new
20 route towards selective activity based probes for cysteine proteases and have applied this approach to proteases from the caspases subfamily.

Cysteine proteases are characterized by a cysteine residue in the active site which serves as a nucleophile during catalysis. The catalytic cysteine is
25 commonly hydrogen bonded with appropriate neighboring residues, so that a thiolate ion can be formed. When a substrate is recognized by the protease, the scissile peptide bond is placed in proximity to the catalytic cysteine, which attacks the carbonyl carbon forming an oxoanion intermediate. The amide bond is then cleaved liberating the C-terminal peptide as an amine. The N-terminal portion of
30 the scissile peptide remains in the covalent acyl-enzyme intermediate, which is subsequently cleaved by water, resulting in regeneration of the enzyme. The N-terminal cleavage product of the substrate is liberated as a carboxylic acid.

Caspases are a family of cysteinyl aspartate-specific proteases. The human genome encodes 11 caspases. Eight of them (caspase-2,3,6,7,8,9,10 and 14) function in apoptosis or programmed cell death. They process through a highly regulated signalling cascade. In a hierarchical order, some initiator caspases (caspase-2,8,9 and 10) cleave and activate effector caspases (caspase-3,6 and 7). These caspases are involved in cancers, autoimmune diseases, degenerative disorders and strokes. Three other Caspases (caspase-1, 4 and 5) serve a distinct function: inflammation mediated by activation of a subset of inflammatory cytokines.

Caspase-1 or interleukin-1 β -converting enzyme (ICE) is primarily found in monocytic cells. This protease is responsible for the production of the pro-inflammatory cytokines interleukin-1-beta and interleukine-18. Inhibition of caspase-1 has been shown to be beneficial in models of human inflammation disease, including rheumatoid arthritis, osteoarthritis, inflammatory bowel disease and asthma.

Caspase-3 is responsible for proteolytic cleavage of a variety of fundamental proteins including cytoskeletal proteins, kinases and DNA-repair enzymes. It is a critical mediator of apoptotsis in neurons. Inhibition of caspase-3 have shown efficacy in models such as stroke, traumatic brain spinal cord injury, hypoxic brain damage, cardiac ischemia and reperfusion injury.

Caspase-8 is an apoptosis initiator caspase, downstream of TNF super-family death receptors. Its substrates include apoptosis-related effector caspases and pro-apoptotic Bcl-2 family members. Resistance to apoptosis in cancer has been linked to low expression levels of caspase-8 and inhibition of caspase-8 increases resistance to apoptosis-inducing stressors such as chemotherapy and radiation. Thus caspase-8 is an attractive target for therapy of tumours and metastatic lesions. Knockout studies reveal as well several other potential roles for caspases-8 which are independent of apoptosis. For example, caspase-8

knockouts exhibit deficiencies in leukocyte differentiation, proliferation and immune response.

For proteolytic enzymes, it is their activity, rather than mere expression level, that dictates their functional role in cell physiology and pathology. Accordingly, molecules that inhibit the activity of caspases are useful as therapeutic agents in the treatment of diseases and in the development of specific imaging biomarkers that visualize the proteolytic activity as well as their inhibition through drug candidates may accelerate target validation, drug development and even clinical trials (H. Pien *et al.* Drug Discovery Today, 2005, 10, 259-266). Using activity based imaging reagents, a specific protein or protein family can be readily monitored in complex protein mixtures, intact cells, and even *in vivo*. Furthermore, enzyme class specific probes can be used to develop screens for small molecule inhibitors that can be used for functional studies (D.A. Jeffery, M. Bogoy Curr. Opp. Biotech. 2003, 14, 87-95).

So far, activity based imaging probes incorporating a peptide substrate have been developed to monitor and label in cell based assays caspase-1 (W.Nishii *et al.*, FEBS Letters 2002, 518, 149-153) or caspase-3 (S. Mizukami *et al.*, FEBS Letters 1999, 453, 356-360). Furthermore a near-infrared fluorescent probe has been reported to detect caspase-1 activity in living animals (S. Messerli *et al.*, Neoplasia 2004, 6, 95-105).

The enzymatic mechanism used by the caspases has been well studied and is highly conserved. From the investigation and screening data of cleavable peptides, electrophilic substrate analogs have been developed that only react in the context of this conserved active site. The electrophilic center in such probes is usually part of a so called "warhead", a molecular entity that is optimized in its electrophilic character and its geometric placement to fit perfectly into the active site of caspases, where it reacts with the catalytic cysteine residue. A wide variety of such electrophilic substrates have been described as mechanism based cysteine protease inhibitors including for example but not exclusively: diazomethyl ketones,

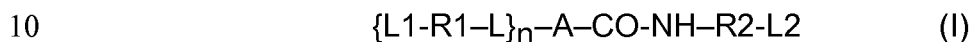
fluoromethyl ketones, acyloxymethyl ketones, O-acylhydroxylamines, vinyl sulfones and epoxysuccinic derivatives (S. Verhelst, M. Bogoy QSAR Comb. Sci. 2005, 24, 261-269).

- 5 Another tool to monitor protease activity consists in bioluminescent assay. This method make use of amino-modified beetles pro-luciferine (caged luciferine) or carboxy-terminal derivatives thereof linked to a protease substrate. A first proteolytic cleavage releases luciferine which is subsequently converted by luciferase, detectable as a luminescent signal. This secondary assay has a similar
10 application spectra than fluorescent probes and present the additionally advantage of a high signal to noise ratio.

To be effective as biological tools, protease inhibitors must be not only very potent but also highly selective in binding to a particular protease. The development of
15 small molecule inhibitors for specific proteases has often started from peptide substrates. Although peptides display a diverse range of biological properties, their use as drugs can be compromised by their instability and their low oral bioavailability. To be effective drugs, protease inhibitors with reduced peptide-like character, high stability against non selective proteolytic degradation, high
20 selectivity for a given protease, and good bioavailability to the *locus* of protease action are desirable. These requirements led to the development of caspases inhibitors A-B where A is a chemical scaffolds covalently linked to an electrophilic warhead B. In presence of caspase, B reacts covalently with the catalytic cysteine (mechanism based inhibitor). In many cases, the selectivity and pharmacokinetic
25 properties of such inhibitors were successfully optimized in the context of biomedical research. To enable the effective nucleophilic attack of the catalytic cysteine, the electrophilic center of such inhibitors must be oriented precisely within the active site of the enzyme. The special arrangement of catalytic cysteine to the electrophilic carbon atom of the warhead corresponds well to the spatial
30 arrangement of the catalytic cysteine and the peptide carbonyl of a scissile peptide substrate. This comparison guided us to the idea that a "redesign" of optimized covalent inhibitors (with a chemical scaffold A and an electrophilic

warhead B) into a cleavable substrate should be possible. Since the chemical scaffold A can be considered as the determinant of inhibitor selectivity, our approach would allow for the transfer of the selectivity or parts of the selectivity of an optimized inhibitor into an activity based chemical probe. We refer to this process as “reversed design” of selective activity based probes from selective caspase inhibitors.

The invention relates to molecular probes for cysteine proteases of the formula (I)



wherein

A is a group recognizable by a caspase;

15

R1 is a linker;

R2 is a bond or a linker;

L is a bond or a group allowing for a facile conjugation of the group L1;

L1 and L2 are, independent of each other, at least one label optionally bound to a solid support; and

20

n is 1;

or

25

R2 is a bond;

L2 is a substrate, suitable for a coupled bioluminescent assay; and

n is 0.

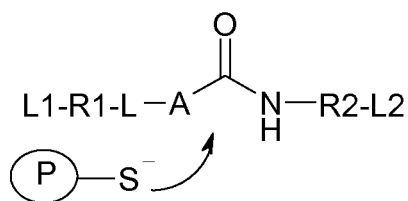
The compounds of the formula (I) are activity based probes (substrates) for cysteine proteases, preferably from the caspase family.

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In their most basic form, the chemical probe consists of four functional elements,
 a) an amide group –CO-NH- as a reactive group, that can be cleaved by the
 action of a protease, b) a scaffold A which defines the selectivity for a given
 protease target, c) linker moieties R1 and R2 to connect subunits to each other
 5 and d) set of label L1 and L2 for detection.

Group A is preferably the main determinant for specificity towards a given
 caspase or a group of caspases, preferably for caspase-1,3 and 8, e.g. as shown
 in compounds 1-43 in Table 1,2 and 3. Activity-based probes of the present
 10 invention show selectivity for a given caspase of the factor 1000 to 1, preferably a
 factor 10 to 1, wherein selectivity is defined by the relative turnover number
 (turnover number with enzyme 1 versus turnover number with enzyme 2) at a
 preferred substrate concentration. The relative turnover number is determined for
 each enzyme pair by dividing the turnover number of the enzyme of interest
 15 (enzyme 1) by the turnover number of another enzyme against which selectivity is
 desired (enzyme 2). For *in vivo* applications high selectivity is desired at low (e.g.
 micromolar or submicromolar) substrate concentrations.

Scheme 1 shows the reaction of a protease P with a substrate wherein A
 20 represents the specificity determinant, and P represents the protease with its
 reactive cysteine comprising the thiol group S⁻:



(Scheme 1)

25 The reaction rate is dependent on the structure of the substrate.

The linker group R1 or R2 is preferably a flexible linker connected to a label L1 or
 L2, respectively, or a plurality of same or different label L2 or L1. The linker group

is chosen in the context of the envisioned application, i.e. in context of an activity based imaging probe for a specific protease. The linker may also increase the solubility of the substrate in the appropriate solvent. The linkers used are chemically stable under the conditions of the actual application. The linker does not interfere with the reaction of a selected protease target nor with the detection of the label L1 and/or L2, but may be constructed such as to be cleaved at some point in time. More specifically, the linker group R1 or R2 is a straight or branched chain alkylene group with 1 to 300 carbon atoms, wherein optionally

(a) one or more carbon atoms are replaced by oxygen, in particular wherein every third carbon atom is replaced by oxygen, e.g. a polyethyleneoxy group with 1 to 100 ethyleneoxy units; and/or

(b) one or more carbon atoms are replaced by nitrogen carrying a hydrogen atom, and the adjacent carbon atoms are substituted by oxo, representing an amide function -NH-CO- ; and/or

(c) one or more carbon atoms are replaced by an ester function -O-CO- ;

(d) the bond between two adjacent carbon atoms is a double or a triple bond; and/or

(e) two adjacent carbon atoms are replaced by a disulfide linkage.

The label L1 and L2 of the substrate can be chosen by those skilled in the art dependent on the application for which the probe is intended.

The label L1 and L2 is independently of each other a spectroscopic probe such as a fluorophore; a quencher or a chromophore; a magnetic probe; a contrast reagent; a molecule which is one part of a specific binding pair which is capable of specifically binding to a partner; a molecule which is a substrate for an enzyme, a molecule covalently attached to a polymeric support, a dendrimer, a glass slide, a microtiter plate known to those proficient in the art; or a molecule possessing a combination of any of the properties listed above.

A preferred embodiment of the present invention is the use of a modified aminoluciferin or a carboxy-terminal protected derivative thereof as a reporter

group, which upon cleavage from the central scaffold A can generate a luminescent signal through its conversion by a luciferase. Therefore, label L2 may alternatively be a substrate, suitable for a coupled bioluminescent assay, characterized in a modified aminoluciferin or a carboxy-terminal protected derivative thereof as a reporter group.

US7148030 discloses examples of bioluminescent protease assays comprising peptides as caspase substrates which are linked to modified aminoluciferines.

- 10 Preferred is a probe which consists of intramolecularly quenched fluorescent probes comprising a polymeric backbone and a plurality of fluorochromes covalently linked *via* scaffold A to the backbone at a density which leads to fluorescent quenching.
- 15 Another preferred embodiment of the present invention is the use of a dendritic macromolecule onto which two or more fluorophores are covalently linked *via* scaffold A at a density which leads to fluorescent quenching. The use of a polymeric probe has the advantage of localized probe delivery (targeting) and a prolonged circulation time in the blood stream of an animal or humans. Polymer
- 20 conjugation alters the biodistribution of low-molecular-weight substances, enabling tumour-specific targeting (by the enhanced permeability and retention effect (EPR effect)) with reduced access to sites of toxicity and the combination of polymer conjugates with low-molecular-weight imaging probes is a most preferred embodiment of the present invention for imaging of multicellular organisms
- 25 including mammals such as mice, rats etc.. The polymeric backbone can consist of any biocompatible polymer and may comprise a polypeptide, a polysaccharide, a nucleic acid or a synthetic polymer. A comprehensive summary of polymers useful in the context of the present invention can be found in M.J. Vincent *et al.* Trends Biotech. 2006, 24, 39-47 and R. Duncan, Nature Reviews Cancer, 2006,
- 30 688-701. A further description of polymers useful in the context of the present invention is disclosed in WO99/58161. The polymeric or dendrimeric probe can comprise protective chains covalently linked to the backbone or the dendritic

molecule. Protective chains include polyethylene glycol, methoxypolyethyleneglycol and further copolymers of ethyleneglycol.

The probe of the present invention can additionally comprise a targeting moiety
5 such as an antibody, an antibody fragment, a receptor-binding ligand, a peptide fragment or a synthetic protein inhibitor.

Label L1 and L2 can further be positively charged linear or branched polymers. Said polymers are known to those skilled in the art to facilitate the transfer of
10 attached molecules over the plasma membrane of living cells. This is especially preferred for substances which otherwise have a low cell membrane permeability or are in effect impermeable for the cell membrane of living cells. A non cell permeable chemical probe will become cell membrane permeable upon conjugation to such a group L1 or L2. Such cell membrane transport enhancer
15 groups L1 and L2 comprise, for example, a linear poly(arginine) of D- and/or L-arginine with 6 – 15 arginine residues, linear polymers of 6 – 15 subunits each of which carry a guanidinium group, an oligomer or a short-length polymer of from 6 to up to 50 subunits, a portion of which have attached guanidinium groups, and/or parts of the sequence of the HIV-tat protein, for example the subunit Tat49-Tat57
20 (RKKRRQRRR in the one letter amino acid code). A linear poly(arginine) of D- and/or L-arginine with 6 – 15 arginine residues is preferably utilized as polymeric label in case L1 is one member and L2 is the other member of two interacting spectroscopic probes L1 / L2, such as in a FRET pair.

25 Most preferred as label L1 and/or L2 are spectroscopic probes. Most preferred as label L2 are molecules representing one part of a spectroscopic interaction pair with L1, furthermore a label which is capable of specifically binding to a partner and molecules covalently attached to a solid support.

30 Particularly preferred are label such that L1 is one member and L2 is the other member of two interacting spectroscopic probes L1 / L2, wherein energy can be transferred non-radiatively between the donor and acceptor (quencher) through

either dynamic or static quenching. Such said pair of label L1 / L2 changes its spectroscopic properties upon reaction/cleavage through the corresponding caspase protease. An example of such a pair of label L1 / L2 is a FRET (Förster resonance energy transfer) pair, e.g. a pro-fluorescent probe covalently labelled at one end (e.g. L1) with a donor (reporter), and the another position (L2) with an acceptor (quencher), or *vice versa*.

In particular, L1 is a donor (reporter) and L2 is an acceptor (quencher), or L1 is a quencher and L2 is a reporter. In using this probe, the reaction of the cysteine protease with the probe will lead to a change in fluorescence. The reporter-quencher distance within the double labelled substrate is changed upon reaction with the protease leading to a spatial separation of reporter and quencher which causes the appearance of fluorescence or change of the emission wavelength. A broad selection of reporter groups may be used as label L1 or L2, respectively, including e.g. near infra-red emitting fluorophores. The substrate containing reporter and quencher remains dark until it reacts with the protease, whereupon the reaction mixture is "lit up" switching on the fluorophore emission, since the reporter label and the quencher label are now spatially separated. Fluorescence quenching and energy transfer can be measured by the emission of only one of the two labels, the quenched or energy donor label. When energy transfer occurs and the energy accepting label is also fluorescent, the acceptor label fluorescence can also be measured. A donor label of these two interacting label can be chosen from chemoluminescent donor probes which eliminates the need of an excitation lamp and reduces acceptor background fluorescence. The mentioned particular method using such double-labelled substrates is useful to determine reaction kinetics based on fluorescence time measurements, and may be applied *in vivo* as well as *in vitro*.

Alternatively, the label L2 may be a solid support or be additionally attached to solid support or attached or attachable to a polymer/ solid support. Linear poly(arginine) of D- and/or L-arginine with 6 – 15 arginine residues is preferably utilized as polymeric label for a L1 / L2 FRET pair.

Particular preferred combinations are two different affinity label, especially a pair of spectroscopic interacting label L1 / L2, e.g. a FRET pair. An affinity label is defined as a molecule which is one part of a specific binding pair which is capable of specifically binding to a partner. A specific binding pair considered is e.g. biotin and avidin or streptavidin furthermore methotrexate, which is a tight-binding inhibitor of the enzyme dihydrofolate reductase (DHFR).

Appropriate pairs of reporters and quenchers can be chosen by those skilled in the art. Typically reporter and quencher are fluorescent dyes with large spectral overlap as, for example, fluorescein as a reporter and rhodamine as a quencher. Other quenchers are gold clusters, and metal cryptates.

A second class of quenchers used in this invention are "dark quenchers", i.e. dyes without native fluorescence having absorption spectra that overlap with the emission spectra of common reporter dyes leading to maximal FRET quenching. Furthermore pairs of dyes can be chosen such that their absorption bands overlap in order to promote a resonance dipole-dipole interaction mechanism within a ground state complex (static quenching).

Particular fluorophores and quenchers considered are: Alexa dyes, including Alexa 350, Alexa 488, Alexa 532, Alexa 546, Alexa 555, Alexa 635 and Alexa 647 (US5696157, US6130101, US6716979); dimethylaminocoumarin (e.g. 7-dimethylaminocoumarin-4-acetic acid succinimidyl ester supplied as product D374 by Invitrogen, CA 92008, USA); quenchers QSY 35, QSY 9 and QSY 21 (Invitrogen, CA 92008, USA); Cyanine-3 (Cy 3), Cyanine 5 (Cy 5) and Cyanine 5.5 (Cy 5.5) (Amersham – GE Healthcare, Solingen, Germany); BHQ-1, BHQ-2 and BHQ-3 (Black Hole Quencher™ of Biosearch Technologies, Inc., Novato, CA 94949, USA); fluorophores ATTO 488, ATTO 532, ATTO 600 and ATTO 655 and quenchers ATTO 540Q and ATTO 612Q (Atto-Tec, D57076 Siegen, Germany); fluorophores DY-505, DY-547, DY-632 and DY-647 (Dyomics, Jena, Germany); 5/6-carboxyfluorescein, tetramethylrhodamine, 4-

dimethylaminoazobenzene-4'-sulfonyl derivatives (Dabsyl) and 4-dimethylaminoazobenzene-4'-carbonyl derivatives (Dabcyl). These can be advantageously combined in the following combinations:

Fluorophore

- Alexa 350, dimethylaminocoumarin, 5/6-carboxyfluorescein, Alexa 488, ATTO 488, DY-505
- 5/6-carboxyfluorescein, Alexa 488, Alexa 532, Alexa 546, Alexa 555, ATTO 488, ATTO 532, tetramethylrhodamine, Cy 3, DY-505, DY-547,
- Alexa 635, Alexa 647, ATTO 600, ATTO 655, DY-632, Cy 5, DY-647 Cy 5.5

Quencher

- Dabsyl, Dabcyl, BHQ 1, QSY 35
- BHQ 2, QSY 9, ATTO 540Q
- BHQ 3, ATTO 612Q, QSY 21

5

Bioluminescent assays that are linked to an enzymatic event yield light coupled to the instantaneous rate of catalysis. The method comprises an amino-modified beetle amino-luciferin or a carboxy-terminal protected derivative thereof where the amino-group of aminoluciferin is linked via an amide bond to the central scaffold A, resulting in a substrate that is recognized and subsequently cleaved by a caspase. The enzymatic activity of a caspase leads to the cleavage of the peptide bond which links the aminoluciferin to the scaffold A liberating the aminoluciferin a substrate for a luciferase. The following reaction of the luciferase with its substrate yields a detectable signal (luminescence). The method thus relates caspase activity with a second enzymatic reaction, generating luminescence as a read-out signal. This type of assay requires the development of a "pro-luciferin" ("caged luciferin"), which is recognized by a luciferase as a substrate only when converted to luciferine by a precedent enzymatic event e.g. proteolytic cleavage. In this way, the luminescent signal is directly dependent on the previous enzymatic event. It is therefore a further embodiment of the present invention to provide a probe for detecting proteolytic activity of caspases by means of luminescence.

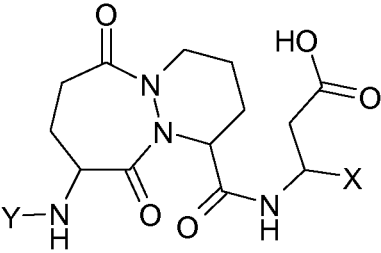
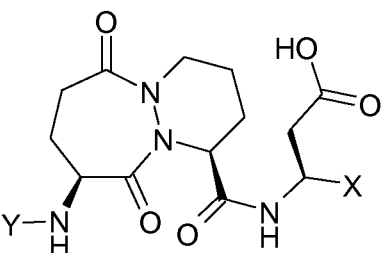
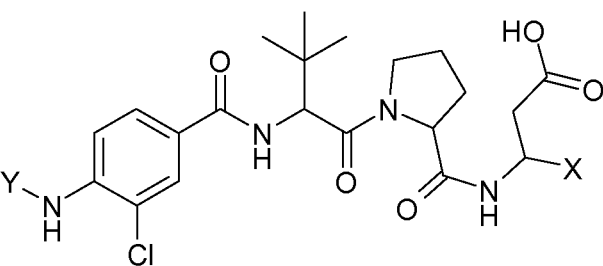
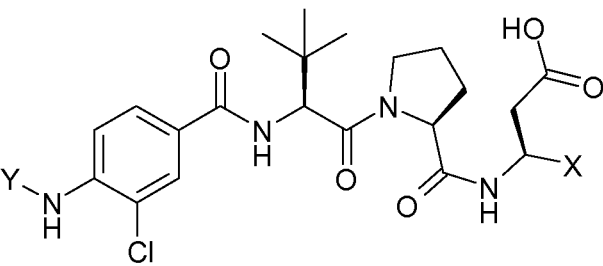
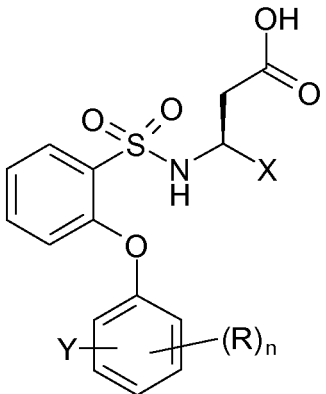
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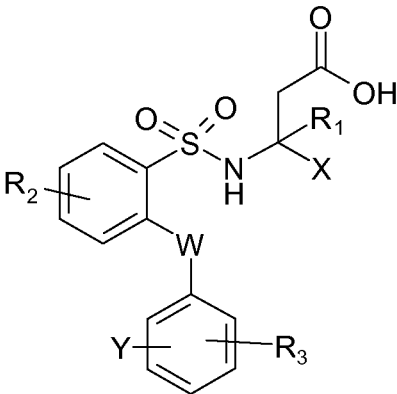
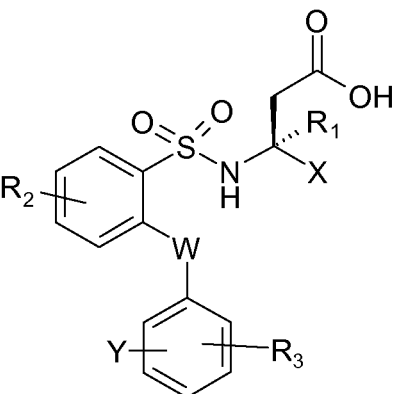
In a particular embodiment, the method involves a substrate wherein L2 is a solid support or attached to a solid support further carrying one member of the reporter / quencher pair, or wherein L2 is a combination of a solid support and one member of the reporter / quencher pair, and L1 is the other member of this pair. In this way, the dark solid support becomes fluorescent upon reaction with the appropriate protease.

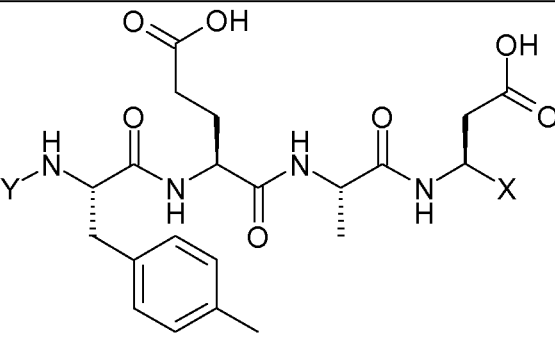
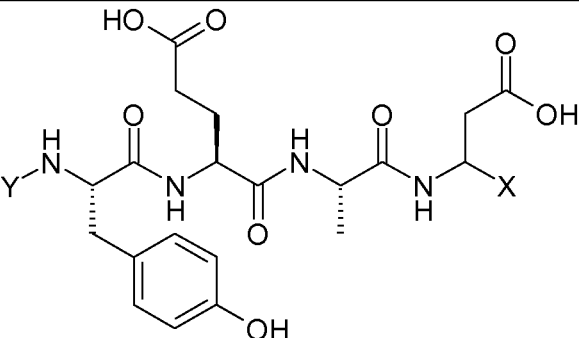
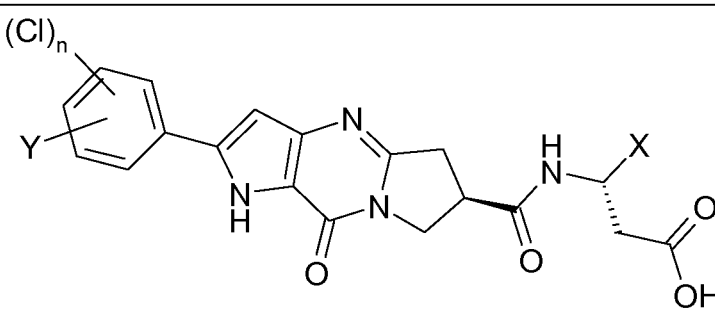
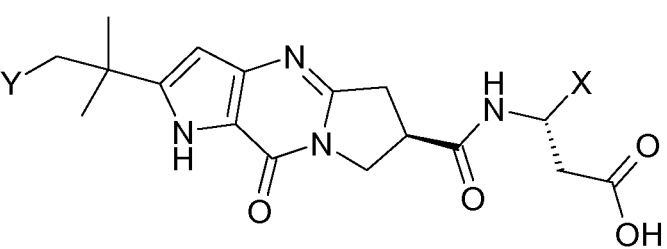
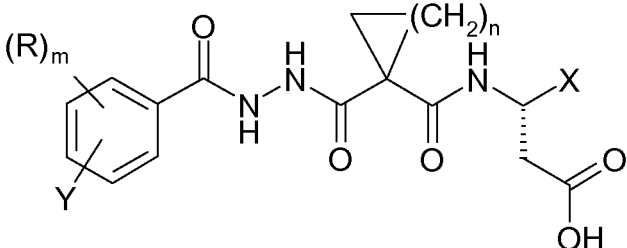
A solid support, may be a glass slide, a microtiter plate or any polymer known to those proficient in the art, e.g. a functionalized polymers (preferably in the form of beads), chemically modified oxidic surfaces, e.g. silicon dioxide, tantalum pentoxide or titanium dioxide, or also chemically modified metal surfaces, e.g. noble metal surfaces such as gold or silver surfaces. A solid support may also be a suitable sensor element.

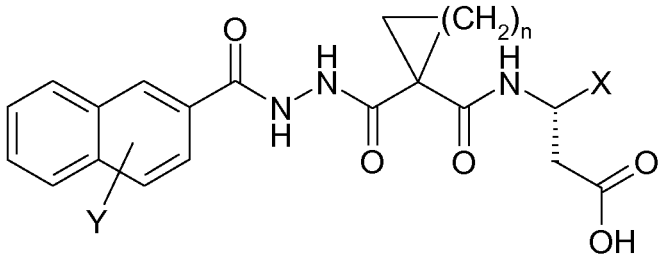
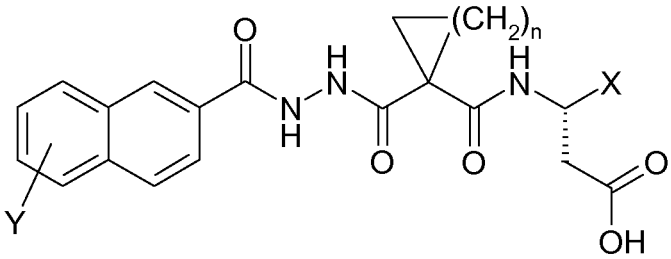
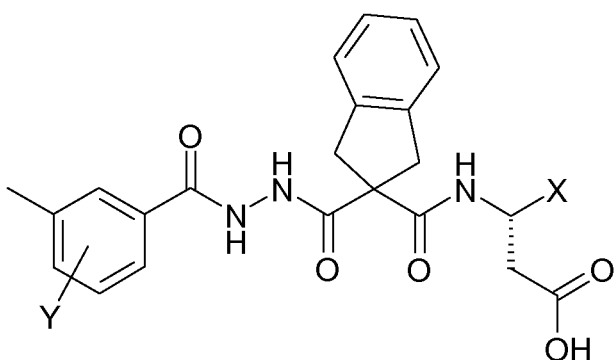
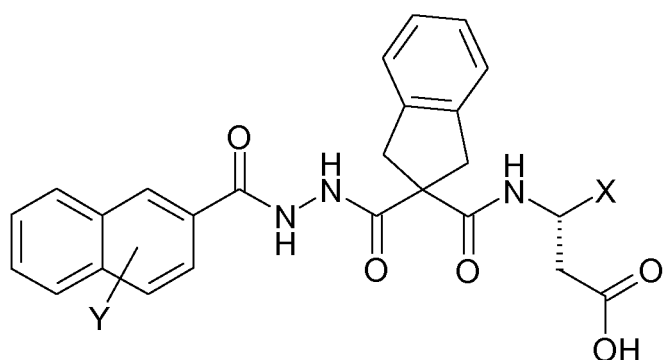
Preferably, the compound of the formula (I) comprises a group A being an inhibitor of caspase-1. The preparation of scaffolds A having caspase-1 inhibitory activity is for example described in US5670494; WO9526958; WO9722619; WO9816504; WO0190063; WO03106460; WO03104231; WO03103677; W. G. Harter, *Bioorg. Med. Chem. Lett.* 2004, 14, 809-812; Shahripour et al., *Bioorg. Med. Chem. Lett.* 2001, 11, 2779-2782; Shahripour et al., *Bioorg. Med. Chem.* 2002, 10, 31-40; M. C. Laufersweiler et al., *Bioorg. Med. Chem. Lett.* 2005, 15, 4322-4326; K. T. Chapman, *Bioorg. Med. Chem. Lett.* 1992, 2, 613-618; Dolle et al., *J. Med. Chem.* 1997, 40, 1941-1946; D. L. Soper et al., *Bioorg. Med. Chem. Lett.* 2006, 16, 4233-4236; D. L. Soper et al., *Bioorg. Med. Chem.* 2006, 14, 7880-7892; D. J. Lauffer et al., *Bioorg. Med. Chem. Lett.* 2002, 12, 1225-1227; and C. D. Ellis et al., *Bioorg. Med. Chem. Lett.* 2006, 16, 4728-4732. More preferred, the compound of the formula (I) is a probe for caspase-1 characterized by a compound comprising the following preferred scaffolds A (Table 1):

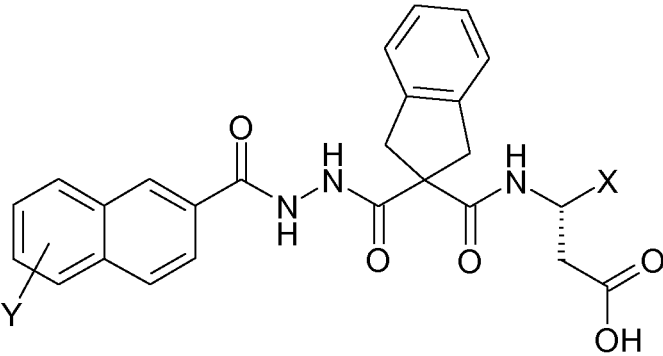
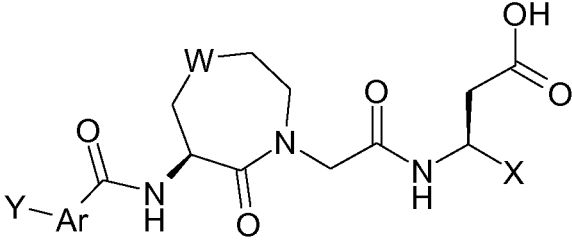
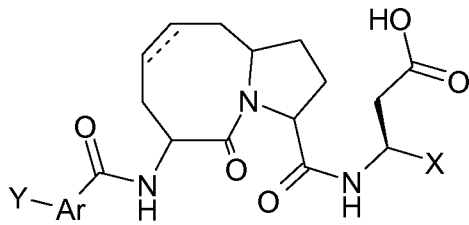
Table 1 – Examples of selective probes (I) for caspase-1

1.	 <p>preferably in the configuration:</p> 
2.	 <p>preferably in the configuration:</p> 
3.	

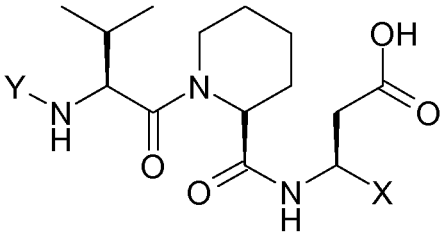
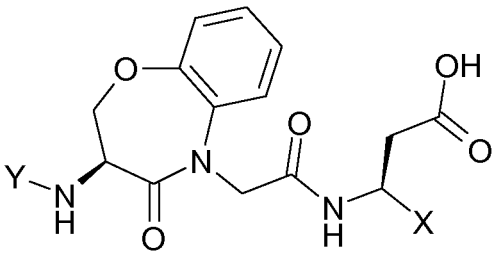
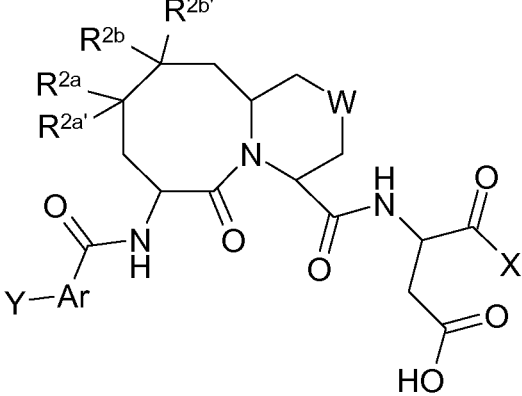
	wherein R is (C ₁ -C ₅)alkyl, phenyl, (C ₅ -C ₆)cycloalkyl; and n is 1-3.
4.	 <p>preferably in the configuration:</p>  <p>wherein R₁ is hydrogen, (C₁-C₆)alkyl, aryl or -CH₂-aryl, R₂ and R₃ are independently of each other hydrogen, or an aryl, substituted aryl, heteroaryl, substituted heteroaryl, cycloalkyl, substituted cycloalkyl, heterocycle, or substituted heterocycle group that is fused to the phenyl group that contains a group R₂ as a substituent; W is a bond, NR₅, CO, S, O, SO₂, O(CHR₅)_n, CHR₅, NR₅CO, CONR₅, OCHR₅, CHR₅O, SCHR₅, CHR₅S, SO₂NR₅, (C₁-C₆)alkyl, NR₅SO₂, CH₂CHR₅, CHR₅CH₂, COCH₂ or CH₂CO; wherein R₅ is independently hydrogen, (C₁-C₆)alkyl, aryl, (CH₂)_n-aryl, or (CH₂)_n-cycloalkyl; and each n is independently 0 to 5.</p>

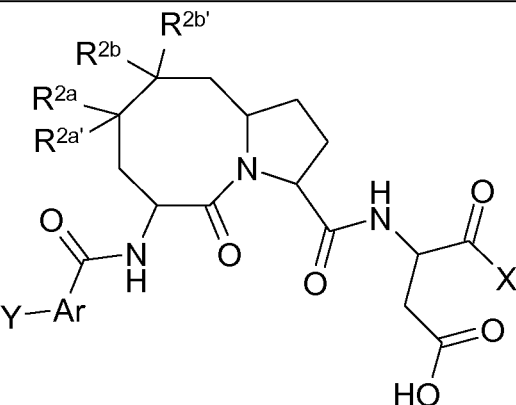
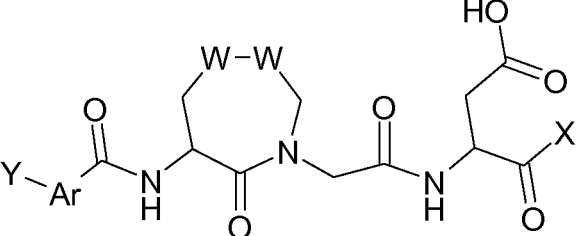
6.	
7.	
8.	 <p>wherein n is 0 or 1;</p>
9.	
10.	

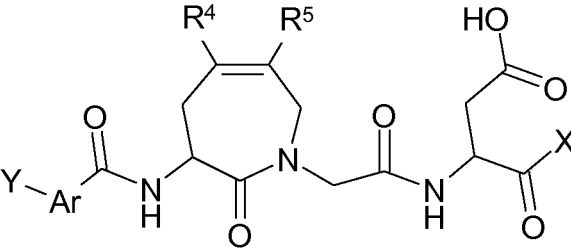
	wherein n is 1-4 m is 1 or 2, and R is methyl or methoxy.
11.	 wherein n is 1-4.
12.	 wherein n is 1-4.
13.	
14.	

15.	
16.	 <p>wherein W is S or S(O)₂, and Ar is aryl or heteroaryl, preferably phenyl, naphthyl, benzothiophene or isoquinolyl.</p>
17.	 <p>wherein Ar is an aryl or heteroaryl group selected from phenyl, benzothiophene, isoquinolyl, cinnamyl, naphthyl, which is optionally once or independently twice substituted by methoxy, chloro, methyl, CF₃, and wherein</p> <p></p> <p>means either a single or a double bond.</p>

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25.	 <p>preferably in the all-(S) configuration, wherein Ar is aryl or heteroaryl; W is CH₂, O or NR⁹ wherein R⁹ is hydrogen or (C₁-C₆)alkyl, aryl, heteroaryl, heterocyclyl; R^{2a}, R^{2a'}, R^{2b} and R^{2b'} are each independently hydrogen, hydroxyl, N(R⁶)₂, halogen, (C₁-C₄)alkyl, (C₁-C₄)alkoxy, and mixtures thereof wherein R⁶ is hydrogen, (C₁-C₆)alkyl, cycloalkyl, (C₆-C₁₀)aryl; or R^{2a} and R^{2b} can taken together to form a double bond.</p>

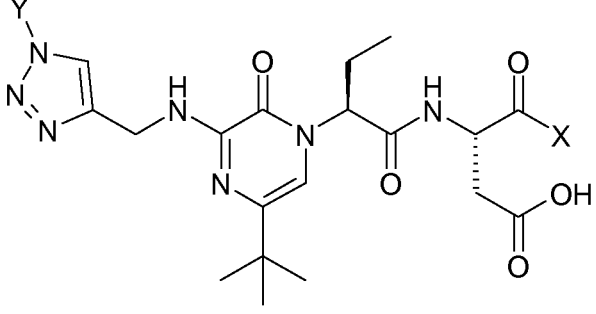
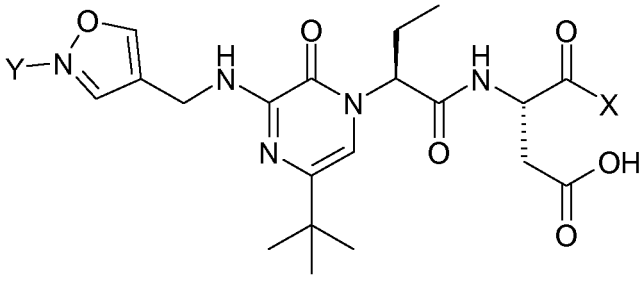
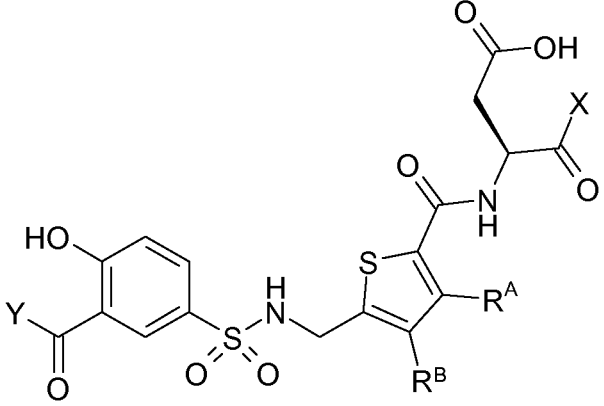
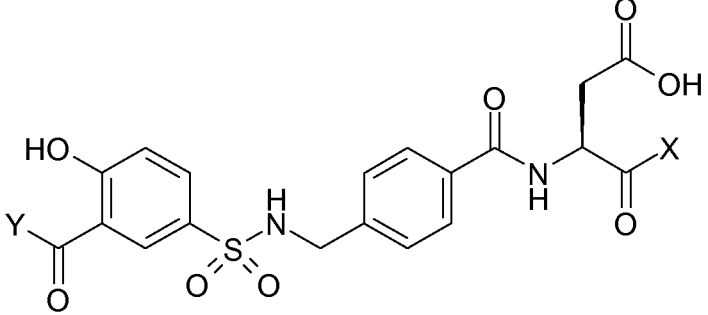
26.	 <p>preferably in the all-(S) configuration, wherein Ar is aryl or heteroaryl; R^{2a}, $R^{2a'}$, R^{2b} and $R^{2b'}$ are each independently hydrogen, hydroxyl, $N(R^6)_2$, halogen, (C_1-C_4)alkyl, (C_1-C_4)alkoxy, and mixtures thereof wherein R^6 is hydrogen, (C_1-C_6)alkyl, cycloalkyl, (C_6-C_{10})aryl; or R^{2a} and R^{2b} can taken together to form a double bond.</p>
27.	 <p>preferred in the all-(S) configuration wherein Ar is aryl or heteroaryl; and W is independently selected from: $C(R^1)_2$; $C(O)$; NR^2; S; $S(O)$; $S(O)_2$; wherein R^1 and R^2 are independently hydrogen, $[C(R^3)_2]_p(CH=CH)_qR^3$, $C(=Z)R^3$, $C(=Z)[C(R^3)_2]_p(CH=CH)_qR^3$, $C(=Z)N(R^3)_2$, $C(=Z)NR^3N(R^3)_2$, CN, CF_3, $N(R^3)_2$, NR^3CN, $NR^3C(=Z)R^3$, $NRC(=Z)N(R^3)_2$, $NHN(R^3)_2$, $NHOR^3$, NO_2, OR^3, OCF_3, F, Cl, Br, I, SO_3H, OSO_3H, $SO_2N(R^3)_2$, SO_2R^3, $P(O)(OR^3)R^3$, $P(O)(OR^3)_2$; wherein p is 0 to 12; wherein q is 0 to 12; wherein Z is O,</p>

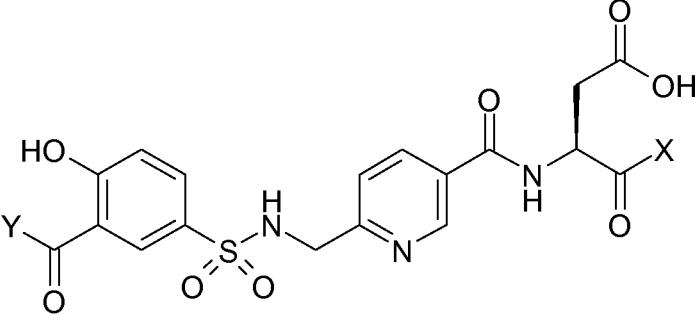
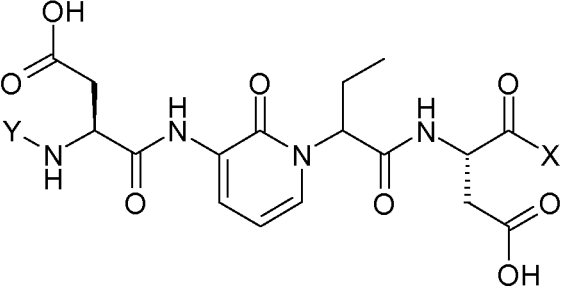
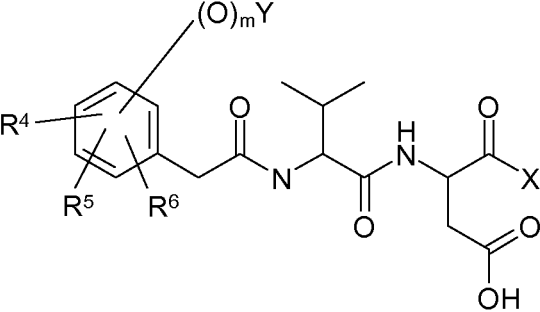
	S, NR ³ ; wherein R ³ is independently hydrogen, alkyl, cycloalkyl, aryl, heteroaryl, heterocyl.
28.	 <p>preferred in the all-(S) configuration</p> <p>wherein Ar is aryl or heteroaryl; and R⁴ and R⁵ are independently selected from: C(R¹)₂; C(O); NR²; S; S(O); S(O)₂; wherein R¹ and R² are independently hydrogen, [C(R³)₂]_p(CH=CH)_qR³, C(=Z)R³, C(=Z)[C(R³)₂]_p(CH=CH)_qR³, C(=Z)N(R³)₂, C(=Z)NR³N(R³)₂, CN, CF₃, N(R³)₂, NR³CN, NR³C(=Z)R³, NRC(=Z)N(R³)₂, NHN(R³)₂, NHOR³, NO₂, OR³, OCF₃, F, Cl, Br, I, SO₃H, OSO₃H, SO₂N(R³)₂, SO₂R³, P(O)(OR³)R³, P(O)(OR³)₂; wherein p is 0 to 12; wherein q is 0 to 12; wherein Z is O, S, NR³; wherein R³ is independently hydrogen, alkyl, cycloalkyl, aryl, heteroaryl, heterocyl.</p>

wherein the variables in the groups 1. to 28. are defined as indicated in the definition next to the respective compound; X is -CONH-R²-L²; Y is -L-R¹-L¹; and R¹, R², L, L¹ and L² are as described above.

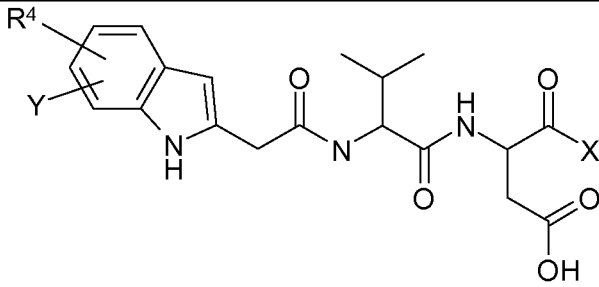
- 5 Further preferably, the compound of the formula (I) comprises a group A being an inhibitor of caspase-3. The preparation of scaffolds A having caspase-3 inhibitory activity is for example described in WO0032620; WO0055127; WO0105772; WO03024955; WO 2008/008264; P. Tawa et al., Cell Death and Differentiation 2004, 11, 439-447; Micale et al., J. Med. Chem. 2004, 47, 6455-6458; and Berger
- 10 et al., Molecular Cell, 2006, 23, 509-521. More preferred, the compound of the formula (I) is a probe for caspase-3 characterized by a compound comprising the following preferred scaffolds A (Table 2):

Table 2 – Examples of selective probes (I) for casapse-3

29.	
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31.	 <p>wherein R^A and R^B are independently hydrogen, (C_1-C_6)alkyl, hydroxyl, (C_1-C_6)alkoxy or halogen.</p>
32.	

33.	
34.	
35.	 <p>preferably in the all-(S) configuration, wherein m is 0 or 1; and R⁴, R⁵ and R⁶ are independently selected from the group consisting of:</p> <ol style="list-style-type: none"> 1) H, 2) halogen, 3) (C₁-C₄)alkoxy optionally substituted with 1-3 halogen atoms, 4) NO₂, 5) OH, 6) benzyloxy, the benzyl portion of which is optionally substituted with 1-2 members selected from the group consisting of: halogen, CN, (C₁-C₄)alkyl and (C₁-C₄)alkoxy, said alkyl and alkoxy being optionally substituted with 1-3 halogen groups,

	<p>7) NH(C₁-C₄)acyl,</p> <p>8) (C₁-C₄)acyl,</p> <p>9) O-(C₁-C₄)alkyl-CO₂H, optionally esterified with a (C₁-C₆)alkyl or a (C₅-C₇)cycloalkyl group,</p> <p>10) CH=CH-CO₂H,</p> <p>11) CO₂H,</p> <p>12) (C₁-C₅)alkyl-CO₂H,</p> <p>13) C(O)NH₂, optionally substituted on the nitrogen atom by 1-2 (C₁-C₄)alkyl groups;</p> <p>14) (C₁-C₅)alkyl-C(O)NH₂, optionally substituted on the nitrogen atom by 1-2 (C₁-C₄)alkyl groups;</p> <p>15) S(O)₀₋₂-(C₁-C₄)alkyl;</p> <p>16) (C₁-C₂)alkyl-S(O)₀₋₂-(C₁-C₄)alkyl;</p> <p>17) S(O)₀₋₂-(C₁-C₆)alkyl or S(O)₀₋₂-phenyl, said alkyl and phenyl portions thereof being optionally substituted with 1-3 members selected from the group consisting of: halogen, CN, (C₁-C₄)alkyl and (C₁-C₄)alkoxy, said alkyl and alkoxy being optionally substituted by 1-3 halogen groups,</p> <p>18) benzoyl optionally substituted by 1-2 members selected from the group consisting of: halogen, CN, (C₁-C₄)alkyl and (C₁-C₄)alkoxy, said alkyl and alkoxy groups being optionally substituted by 1-3 halogen groups,</p> <p>19) phenyl or naphthyl, optionally substituted with 1-2 members selected from the group consisting of: halogen, CN, (C₁-C₄)alkyl and (C₁-C₄)alkoxy, said alkyl and alkoxy being optionally substituted with 1-3 halogen groups,</p> <p>20) CN,</p> <p>21) (C₁-C₄)alkylene-HET₂, wherein HET₂ represents a 5-7 membered</p>
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	<p>aromatic or non-aromatic ring containing 1-4 heteroatoms selected from O, S, NH and N(C₁-C₄) and optionally containing 1-2 oxo groups, and optionally substituted with 1-3 (C₁-C₄)alkyl, OH, halogen or (C₁-C₄)acyl groups;</p> <p>22) O-(C₁-C₄)alkyl-HET3, wherein HET3 is a 5 or 6 membered aromatic or non-aromatic ring containing from 1 to 3 heteroatoms selected from O, S and N, and optionally substituted with one or two groups selected from halogen and (C₁-C₄)alkyl, and optionally containing 1-2 oxo groups, and</p> <p>23) HET4, wherein HET4 is a 5 or 6 membered aromatic or non-aromatic ring, and the benzofused analogs thereof, containing from 1 to 4 heteroatoms selected from O, S and N, and is optionally substituted by one or two groups selected from halogen, (C₁-C₄)alkyl and (C₁-C₄)acyl; and wherein halogen includes F, Cl, Br and I.</p>
36.	 <p>preferably in the all-(S) configuration,</p> <p>wherein R⁴ is selected from the group consisting of:</p> <ol style="list-style-type: none"> 1) H, 2) halogen, 3) (C₁-C₄)alkoxy optionally substituted with 1-3 halogen atoms, 4) NO₂, 5) OH, 6) benzyloxy, the benzyl portion of which is optionally substituted with 1-2 members selected from the group consisting of: halogen, CN, (C₁-C₄)alkyl and (C₁-C₄)alkoxy, said alkyl and alkoxy being optionally

	<p>substituted with 1-3 halogen groups,</p> <p>7) $\text{NH}(\text{C}_1\text{-C}_4)\text{acyl}$,</p> <p>8) $(\text{C}_1\text{-C}_4)\text{acyl}$,</p> <p>9) $\text{O}-(\text{C}_1\text{-C}_4)\text{alkyl-CO}_2\text{H}$, optionally esterified with a $(\text{C}_1\text{-C}_6)\text{alkyl}$ or a $(\text{C}_5\text{-C}_7)\text{cycloalkyl}$ group,</p> <p>10) $\text{CH}=\text{CH-CO}_2\text{H}$,</p> <p>11) CO_2H,</p> <p>12) $(\text{C}_1\text{-C}_5)\text{alkyl-CO}_2\text{H}$,</p> <p>13) $\text{C}(\text{O})\text{NH}_2$, optionally substituted on the nitrogen atom by 1-2 $(\text{C}_1\text{-C}_4)\text{alkyl}$ groups;</p> <p>14) $(\text{C}_1\text{-C}_5)\text{alkyl-C}(\text{O})\text{NH}_2$, optionally substituted on the nitrogen atom by 1-2 $(\text{C}_1\text{-C}_4)\text{alkyl}$ groups;</p> <p>15) $\text{S}(\text{O})_{0-2}-(\text{C}_1\text{-C}_4)\text{alkyl}$;</p> <p>16) $(\text{C}_1\text{-C}_2)\text{alkyl-S}(\text{O})_{0-2}-(\text{C}_1\text{-C}_4)\text{alkyl}$;</p> <p>17) $\text{S}(\text{O})_{0-2}-(\text{C}_1\text{-C}_6)\text{alkyl}$ or $\text{S}(\text{O})_{0-2}\text{-phenyl}$, said alkyl and phenyl portions thereof being optionally substituted with 1-3 members selected from the group consisting of: halogen, CN, $(\text{C}_1\text{-C}_4)\text{alkyl}$ and $(\text{C}_1\text{-C}_4)\text{alkoxy}$, said alkyl and alkoxy being optionally substituted by 1-3 halogen groups,</p> <p>18) benzoyl optionally substituted by 1-2 members selected from the group consisting of: halogen, CN, $(\text{C}_1\text{-C}_4)\text{alkyl}$ and $(\text{C}_1\text{-C}_4)\text{alkoxy}$, said alkyl and alkoxy groups being optionally substituted by 1-3 halogen groups,</p> <p>19) phenyl or naphthyl, optionally substituted with 1-2 members selected from the group consisting of: halogen, CN, $(\text{C}_1\text{-C}_4)\text{alkyl}$ and $(\text{C}_1\text{-C}_4)\text{alkoxy}$, said alkyl and alkoxy being optionally substituted with 1-3 halogen groups,</p> <p>20) CN,</p>
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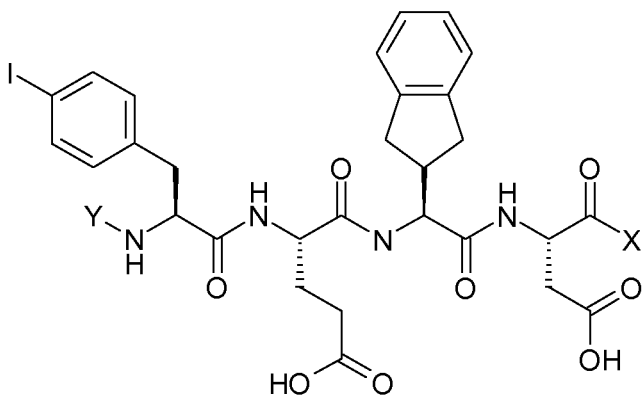
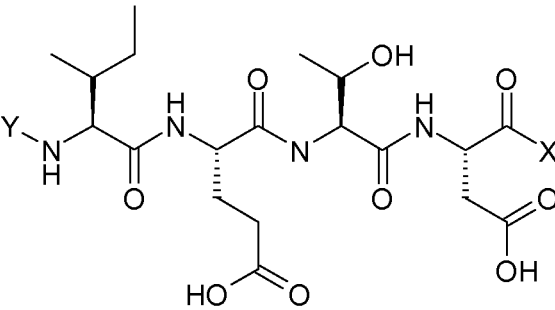
	<p>21) (C₁-C₄)alkylene-HET2, wherein HET2 represents a 5-7 membered aromatic or non-aromatic ring containing 1-4 heteroatoms selected from O, S, NH and N(C₁-C₄) and optionally containing 1-2 oxo groups, and optionally substituted with 1-3 (C₁-C₄)alkyl, OH, halogen or (C₁-C₄)acyl groups;</p> <p>22) O-(C₁-C₄)alkyl-HET3, wherein HET3 is a 5 or 6 membered aromatic or non-aromatic ring containing from 1 to 3 heteroatoms selected from O, S and N, and optionally substituted with one or two groups selected from halogen and (C₁-C₄)alkyl, and optionally containing 1-2 oxo groups, and</p> <p>23) HET4, wherein HET4 is a 5 or 6 membered aromatic or non-aromatic ring, and the benzofused analogs thereof, containing from 1 to 4 heteroatoms selected from O, S and N, and is optionally substituted by one or two groups selected from halogen, (C₁-C₄)alkyl and (C₁-C₄)acyl; and wherein halogen includes F, Cl, Br and I.</p>
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41.	
42.	

wherein the variables in the groups 29. to 42. are defined as indicated in the definition next to the respective compound; X is -CONH-R₂-L₂; Y is -L-R₁-L₁; and R₁, R₂, L, L₁ and L₂ are as described above.

Further preferably, the compound of the formula (I) comprises a group A being an inhibitor of caspase-8. The preparation of scaffolds A having caspase-8 inhibitory activity is for example described in Berger et al., Molecular Cell, 2006, 23, 509-521; and Garcia-Calvo, J. Biol. Chem. 1998, 273 (49), 32608-32613. More preferred, the compound of the formula (I) is a probe for caspase-8 characterized by a compound comprising the following preferred scaffolds A (Table 3):

Table 3 – Examples of selective probes (I) for caspase-8

43.	
44.	

wherein X is -CONH-R2-L2; Y is -L-R1-L1; and R1, R2, L, L1 and L2 are as described above.

The following definitions apply if not otherwise stated:

Alkyl means a straight or branched chain hydrocarbon having 1 to 6 carbon atoms.

Examples of (C₁-C₆)alkyl groups are methyl, ethyl, propyl, isopropyl, isobutyl, butyl, tert-butyl, sec-butyl, pentyl, and hexyl.

Acyl is defined as a group -C(=O)alkyl.

Aryl is defined as an aromatic hydrocarbon having 6 to 10 carbon atoms.

Examples of aryl groups include phenyl and naphthyl.

- 5 Heteroaryl is defined as an aryl group wherein one or more carbon atom of the aromatic hydrocarbon has been replaced with a heteroatom wherein the term "heteroatom" includes oxygen, nitrogen, sulfur, and phosphorus. Examples of heteroaryl groups include furan, thiophene, benzothiophene, pyrrole, thiazole, pyridine, pyrimidine, pyrazine, benzofuran, indole, coumarin, quinoline,
10 isoquinoline, and naphthyridine.

Cycloalkyl means a cyclic alkyl group having 3 to 10 carbon atoms. Examples of cycloalkyl groups include cyclopropane, cyclobutane, cyclopentane, and cyclohexane.

- 15 Heterocycle or heterocyclyl means a cycloalkyl group on which one or more carbon atom has been replaced with a heteroatom. Examples of heterocycles include piperazine, morpholine, and piperidine.

- 20 The aryl, heteroaryl, or cycloalkyl groups may be substituted with one or more substituents, which can be the same or different. Examples of suitable substituents include alkyl, alkoxy, thioalkoxy, hydroxy, halogen, trifluoromethyl, amino, alkylamino, dialkylamino, NO₂, CN, CO₂H, CO₂alkyl, SO₃H, CHO, C(=O)alkyl, CONH₂, CONH-alkyl, CONHR_q, C(=O)N(alkyl)₂, (CH₂)_nNH₂, OH,
25 CF₃, O(C₁-C₆)alkyl, (CH₂)_nNH-alkyl, NHR_q, NHCOR_q, phenyl, where n is 1 to 5 and R_q is hydrogen or (C₁-C₆)alkyl.

- The activity based probes of the present invention may be synthesized by using
30 appropriate protecting group chemistry known in the art to build up the central scaffold A and to attach either linker and label L1 or L2 to this unit *via* a group L and a group -C(O)-NH-. Appropriate building blocks as well as FRET-pairs such

as the cyanine-dyes (e.g. Cy3 B, Cy 5.5, Cy 7) are commercially available (e.g. Sigma-Aldrich, GE-Healthcare). For a subset of probe, described in this invention, the solid-phase synthesis method is particularly useful (B. J. Merrifield, Methods in Enzymology 1997, 289, 3-13). Depending on the synthetic requirements,
5 attachment linker, quencher or fluorophore may be done on the solid support or by solution phase chemistry.

In general, reactive side chain residues on the central scaffold A and optionally the group L will be protected and liberated sequentially for further modification
10 with the subunits L1R1 and L2R2 respectively. Conjugation of these subunits can be accomplished by known methods of chemical synthesis. Particular useful is the reaction between a dye active ester and a primary amine group of the scaffold A to connect both units *via* an amide bond. Intermediates as well as final probe molecules may be purified by high performance liquid chromatography (HPLC)
15 and characterized by mass spectrometry and analytical HPLC before they are used in labelling and imaging experiments.

The present invention is illustrated in the following paragraph by several but non-limiting examples:

20 In a preferred embodiment, the probe of the formula (I) comprises a scaffold A which is derived from a tetrapeptide caspase-1 inhibitor (Table 1, compound 2) bearing a chromophore at the C-terminal side and at the N-terminal side. Appropriate chromophores are chosen in a way that their spectral properties are
25 suitable for fluorescence resonance energy transfer (FRET). Chromophores can be fluorescent or non fluorescent. In principle, a broad variety of chromophores may be used in the present invention, as long as the central requirement that is a spectral change after proteolytic cleavage of a peptide bond is met. The attachment of such interacting chromophores and the central scaffold is made
30 optionally *via* linker units.

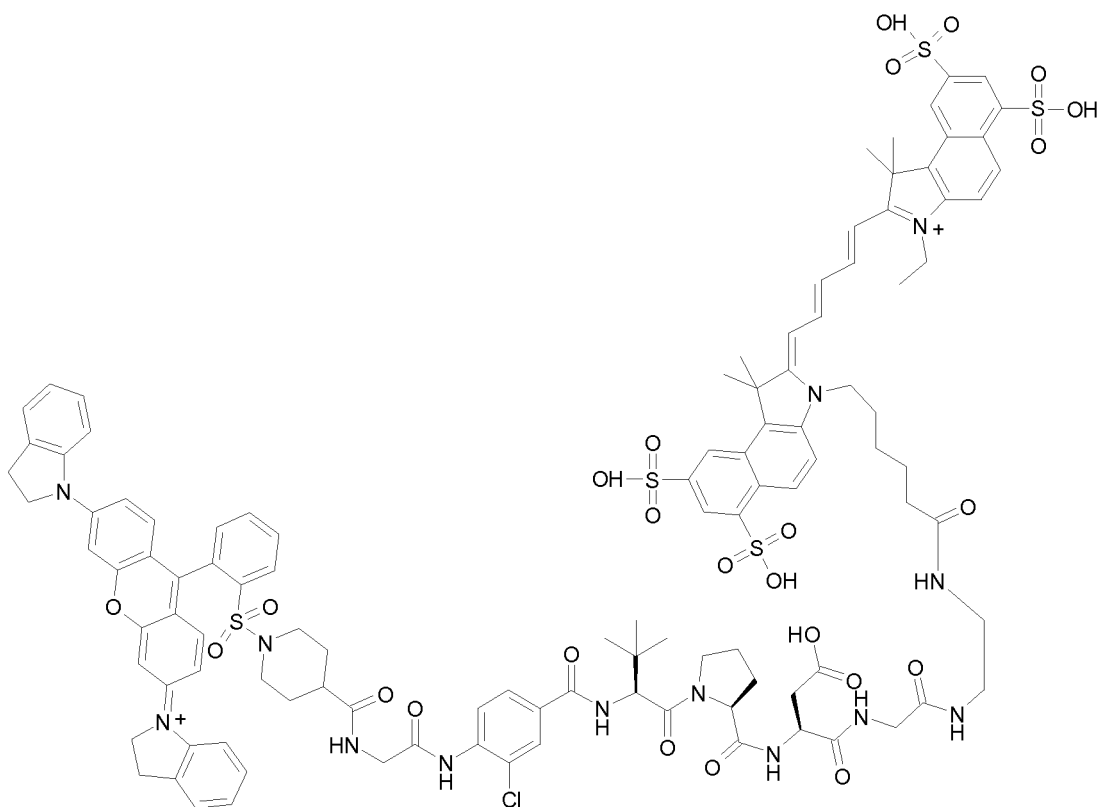
Preferably, the fluorophore are chosen from the group of xanthene- or cyanine dyes. More preferred are cyanine dyes from the group of carbacyanines, thiacyanines, oxacyanines and azacyanines. Cyanine dyes suitable to be used in the context of the present invention are disclosed in US5268468 and US5627027.

5 They include the dyes with the trademark (Amersham, GE Healthcare) Cy 3, Cy 3B, Cy 3.5, Cy 5, Cy 5.5, Cy 7 and Cy 7.5.

Preferably, the quencher unit is a non-fluorescent chromophore which include 2,4-dinitrophenyl, 4-(4-dimethylaminophenyl)azobenzoic acid (DABCYL), 7-methoxy-
10 coumarin-4-yl)acetyl and non fluorescent cyanine-dyes as described in WO9964519.

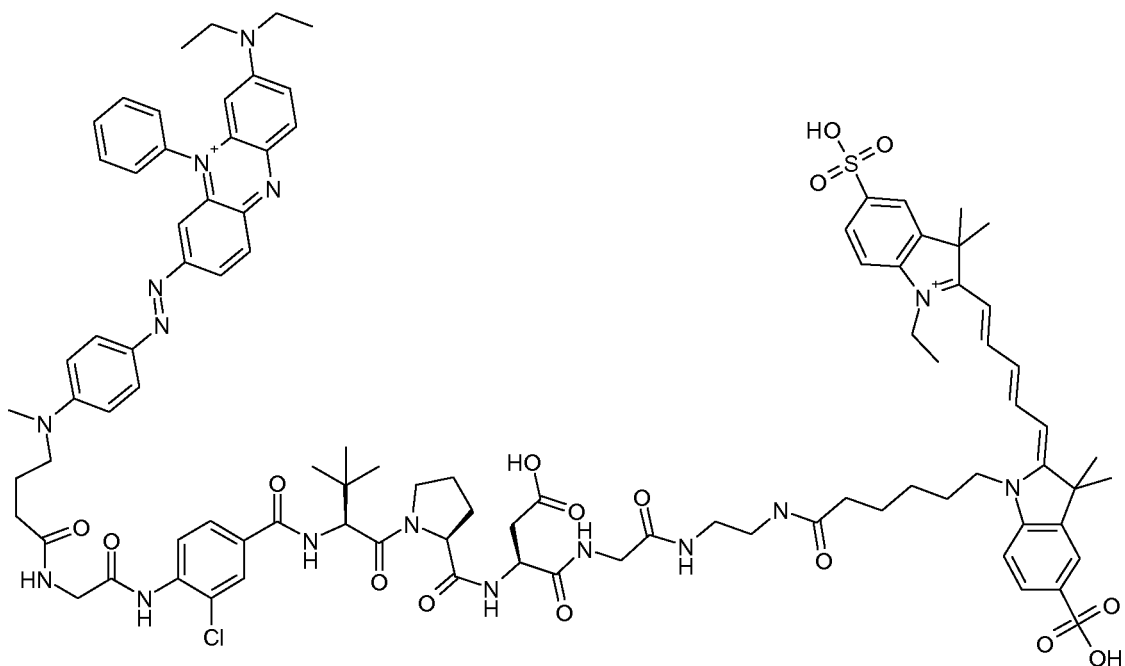
In a preferred embodiment, the quencher does not show a significant emission and more preferably is a non-fluorescent chromophore. In this embodiment, the
15 imaging reagent comprises a fluorophore and a non-fluorescent (dark) acceptor chromophore.

More preferred is a probe of the formula (I) based on a tetrapeptide scaffold (Table 1, compound 2) bearing a QSY 21-Quencher at the N-terminal side and a
20 CY 5.5 fluorophore at the C-terminal side (Scheme 2):



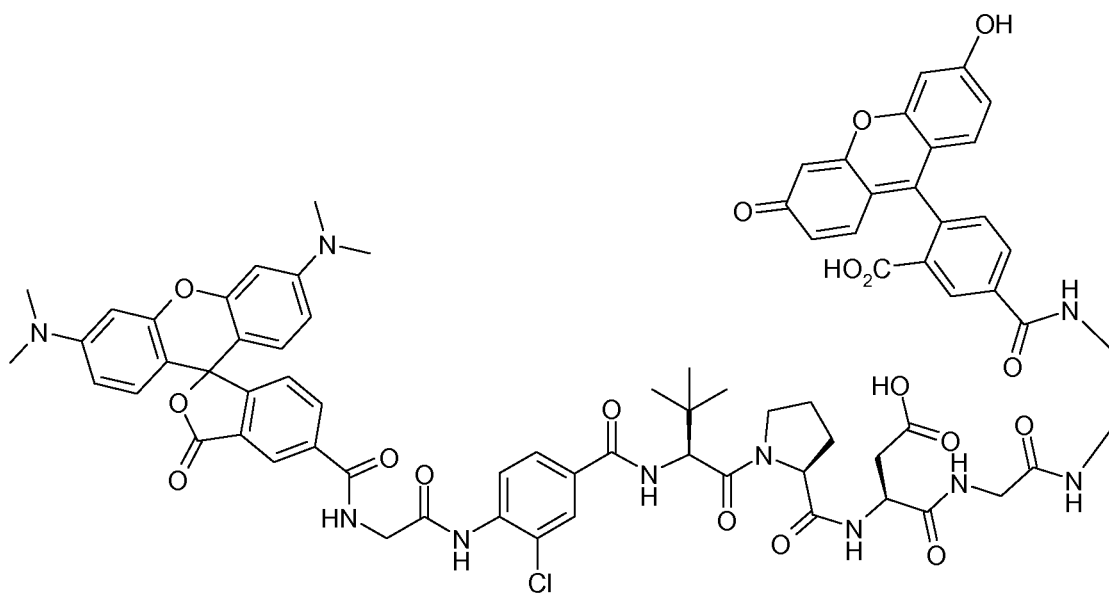
(Scheme 2).

A further preferred embodiment includes the same scaffold bearing the dark
5 quencher BHQ 3 at the N-terminal side and a Cy 7 fluorophore at the C-terminal
side (Scheme 3):



(Scheme 3).

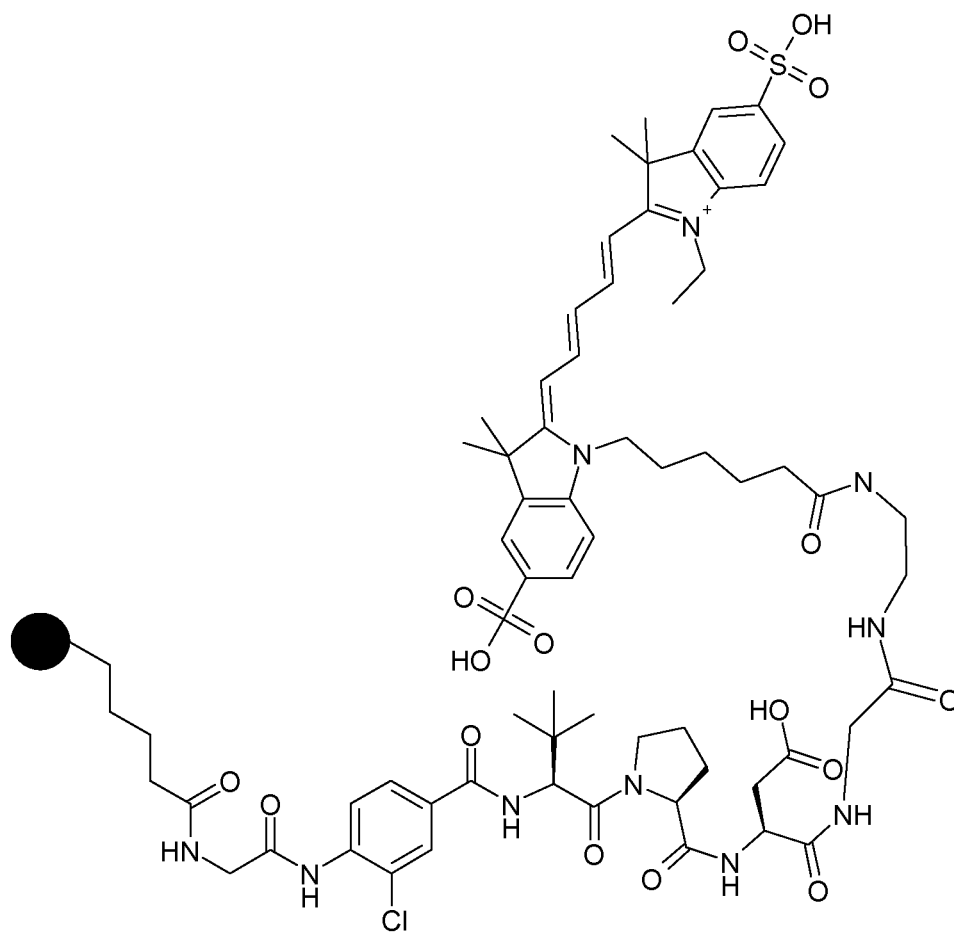
In a preferred embodiment, fluorescein and tetramethylrhodamine are chosen as an interacting FRET pair and the tetramethylrhodamine is placed at the N-terminal side of the scaffold whereas the fluorescein is linked at the C-terminal side as shown in (Scheme 4):



(Scheme 4).

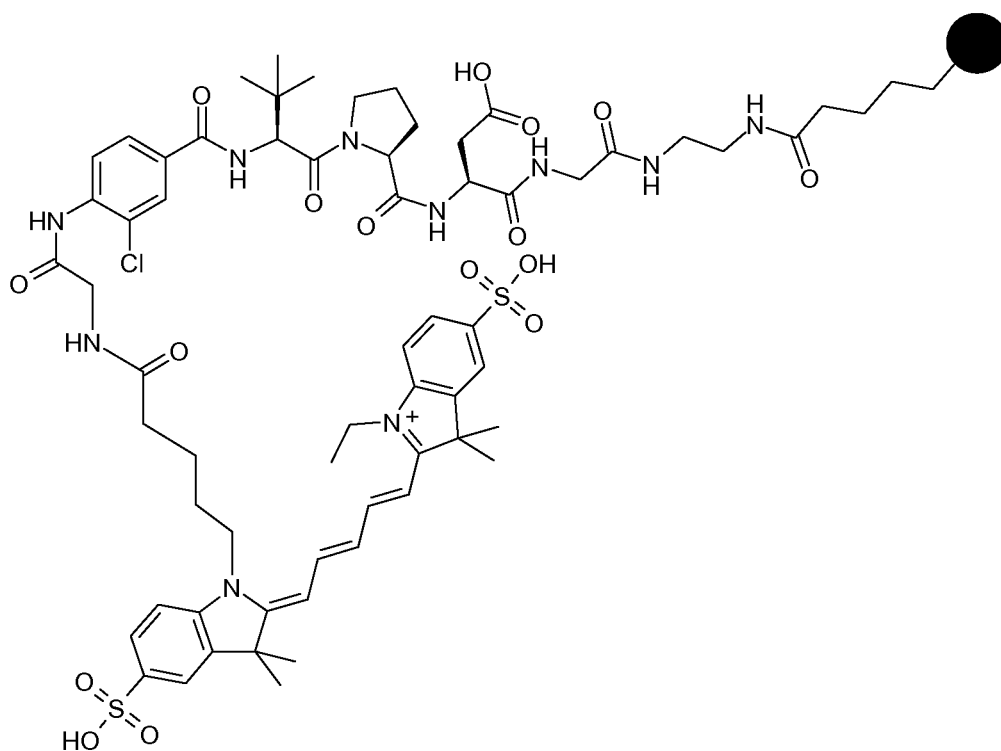
In a further preferred embodiment, one interaction partner of the FRET pair comprises a nanoparticle. More preferred in the context of the present inventions are CdSe nanoparticles (e.g. Quantum-dots), lanthanide-ion doped oxide nanoparticles (e.g. $\text{Y}_{0.6}\text{Eu}_{0.4}\text{VO}_4$) and iron-oxide nanoparticles (e.g. AminoSPARK 680 and AminoSPARK 750 supplied by VisEn Medical, Inc., MA 01801, USA). If such nanoparticles are used as a donor in a FRET pair, they can be excited at a wavelength much shorter than the acceptor absorption thus minimizing direct acceptor excitation. In addition, the narrow donor emission does not overlap with acceptor emission. Furthermore, such nanoparticles proved to be much more photostable than organic dyes which undergo fast photobleaching. Activated quantum dots for chemical conjugation are commercially available (Invitrogen, CA 92008, USA) and their emission wavelength can be chosen from a variety of products.

Schemes 5 and 6 show quantum dot based probes of the formula (I) that are specific for caspase-1. Thus, in a further preferred probe of the formula (I) the quantum dot (e.g. QD605 supplied by Invitrogen, CA 92008, USA) might be positioned *via* an appropriate linker either at the N-terminal side of the caspase-1 probe (Scheme 5) or at the C-terminal side of the caspase-1 probe (Scheme 6).



(Scheme 5);

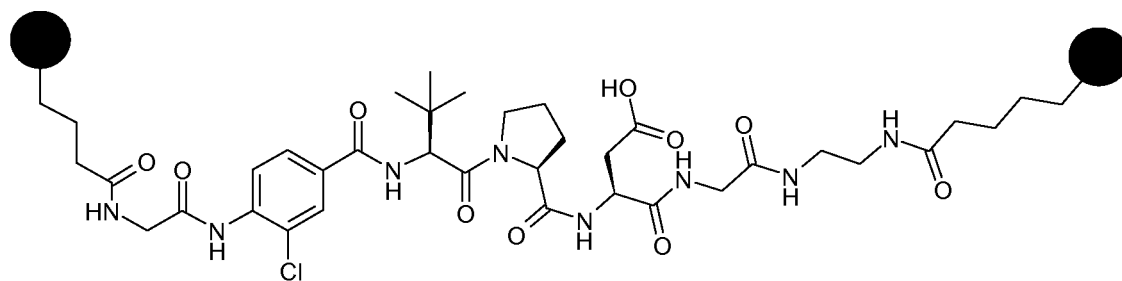
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(Scheme 6).

The quantum dot is represented as a black circle and an appropriate acceptor molecule is represented by the cyanine-dye CY 7.

In a further preferred embodiment, the quantum dots in the probe of the formula (I) are connected to gold-nanoparticles *via* a proteolytic cleavable subunit (Scheme 7):

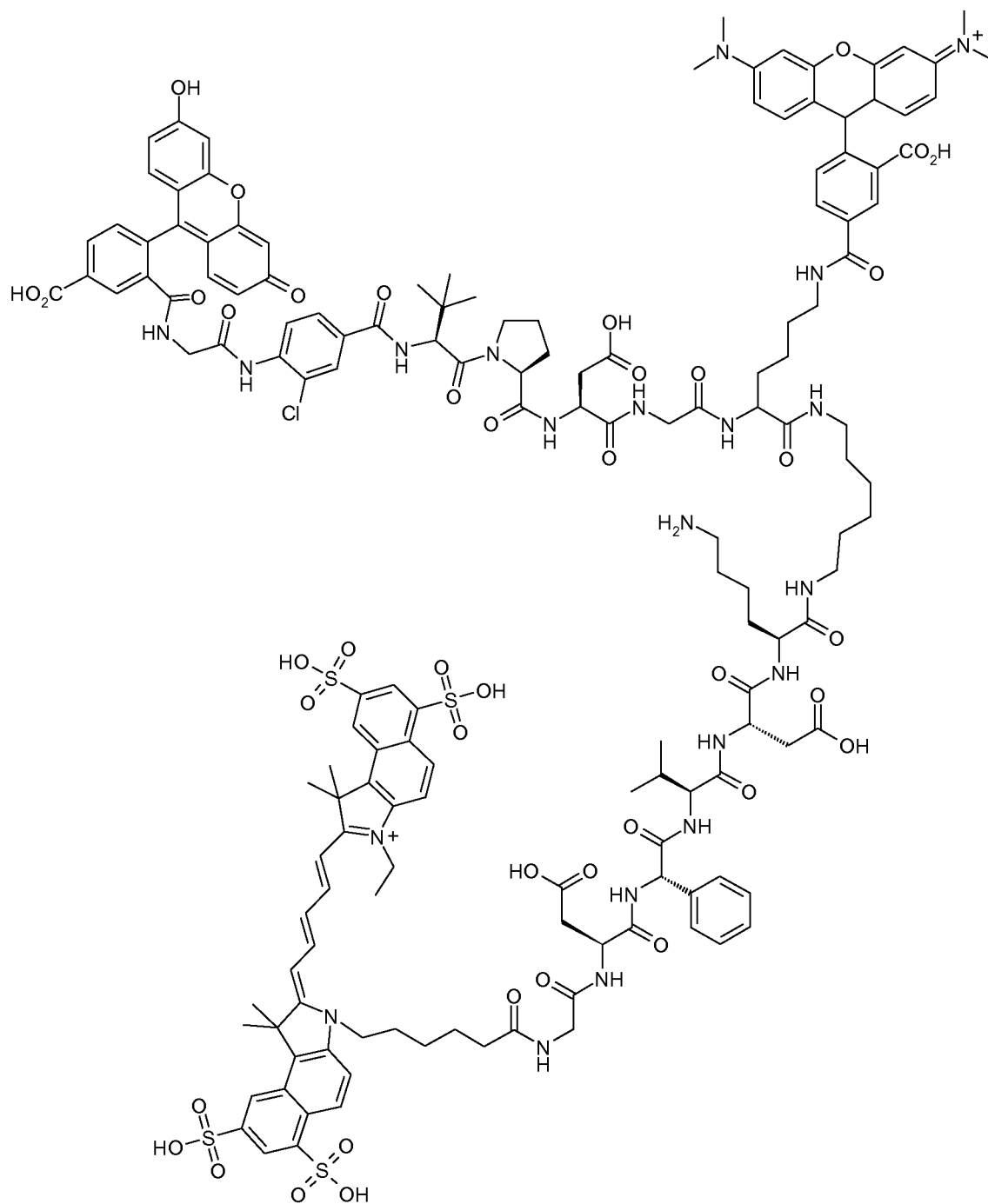


(Scheme 7).

The quantum dot and the gold-nanoparticle are represented as a black circle.

Gold nanoparticles (AuNPs) have been shown as effective quenchers for organic fluorescent dyes as well as for quantum dots. The application of quantum dots in combination with AuNPs is e.g. disclosed in WO2006126570.

- 5 In a further preferred embodiment, the probe of the formula (I) consists of a multi-FRET system wherein two specific protease probes are covalently linked together (Scheme 8):

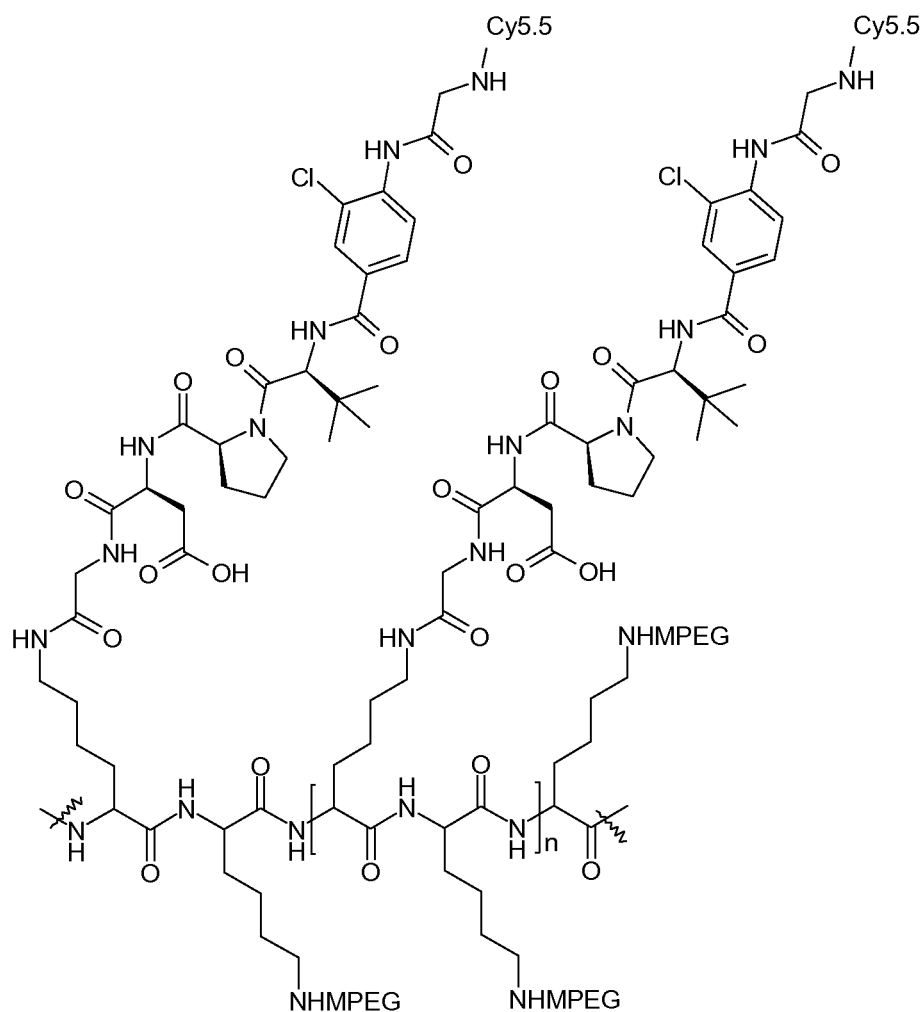


(Scheme 8).

In this configuration it is possible to excite at a single wavelength and use the
5 different emission ratios as unique FRET signatures (K.E Sapsford et al., Angew.
Chem. Int. Ed. 2006, 45, 4562- 4588). This probe combines two specificities in
one molecule that is a scaffold for caspase-1 and a scaffold for caspase-3.

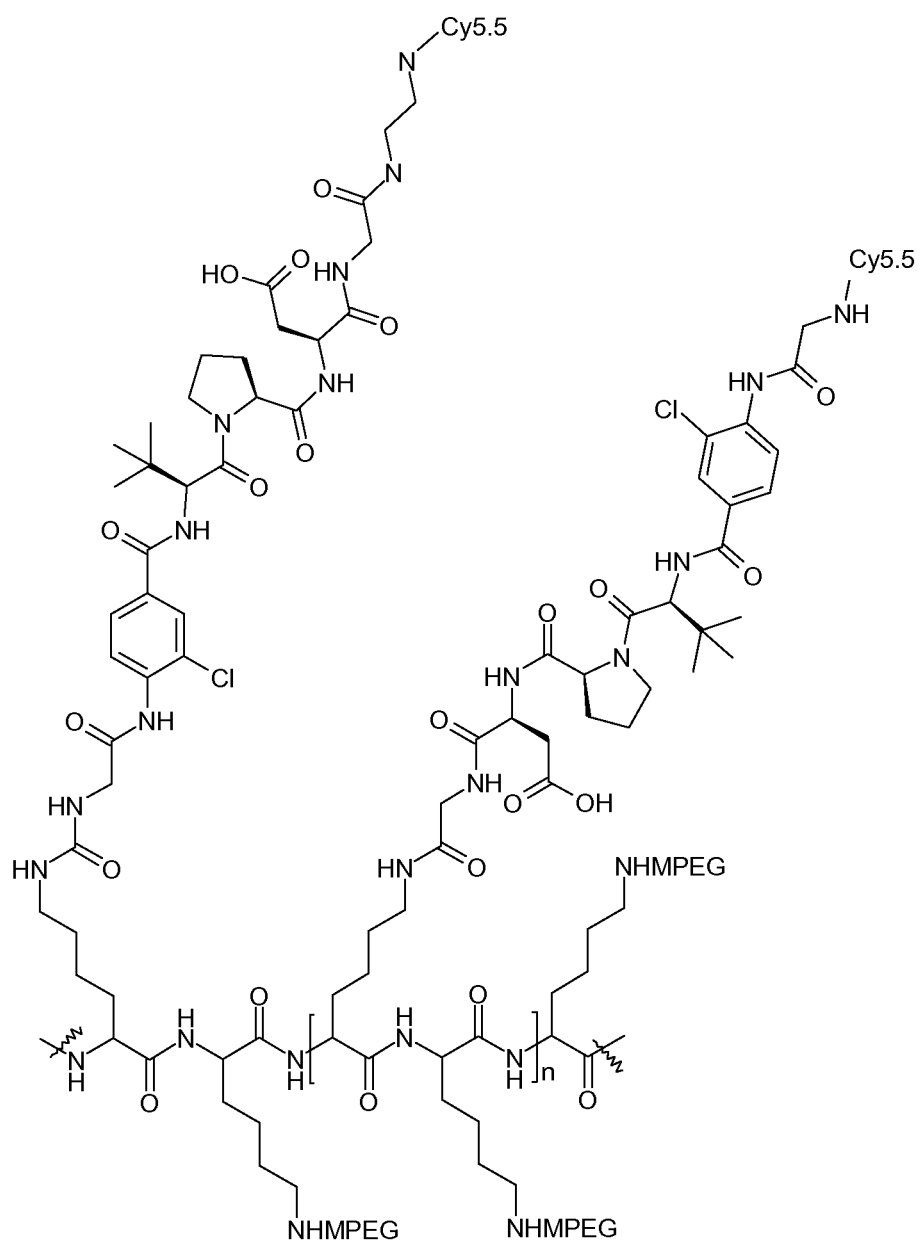
In a further preferred embodiment, the probes of the formula (I) are designed to have a long circulation time, have high tumoral accumulation and contain quenched fluorescent markers which become fluorescent in the near-infrared spectrum after enzyme activation. These probes are based on synthetic graft copolymer [partially methoxy poly(ethylene glycol) modified poly-L-lysine] onto which multiple NIR fluorochromes were attached to free poly-lysine residues. The fluorescence of these macromolecules is highly reduced, due to internal quenching by the high density and close proximity of the NIR-chromophores.

As an example, Scheme 9 shows a polymer-based caspase-1 probe where the connection of A to the poly-lysine backbone of D- and/or L- lysine is achieved *via* a linker at the C-terminal side whereas the NIR-chromophore Cy 5.5 is attached *via* a linker at the N-terminal side:



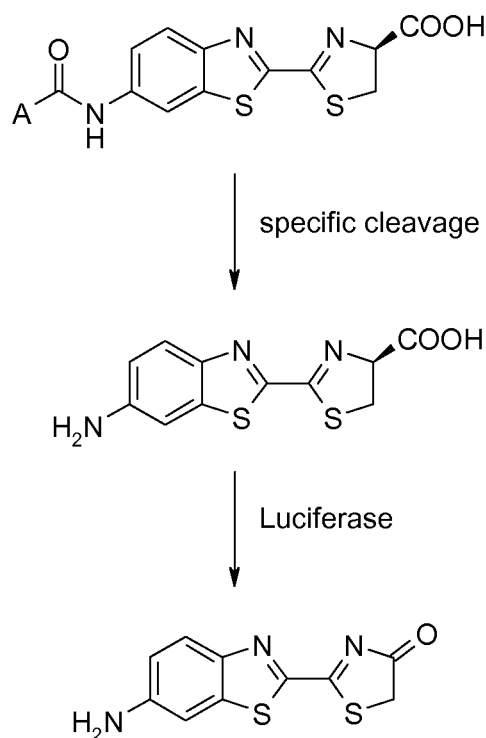
(Scheme 9).

The inverse situation is shown in Scheme 10, where the connection of A to the
 5 poly-lysine backbone of D- and/or L- lysine is achieved *via* a linker at the N-
 terminal side whereas the NIR-chromophore Cy 5.5 is attached *via* a linker at the
 C-terminal side:

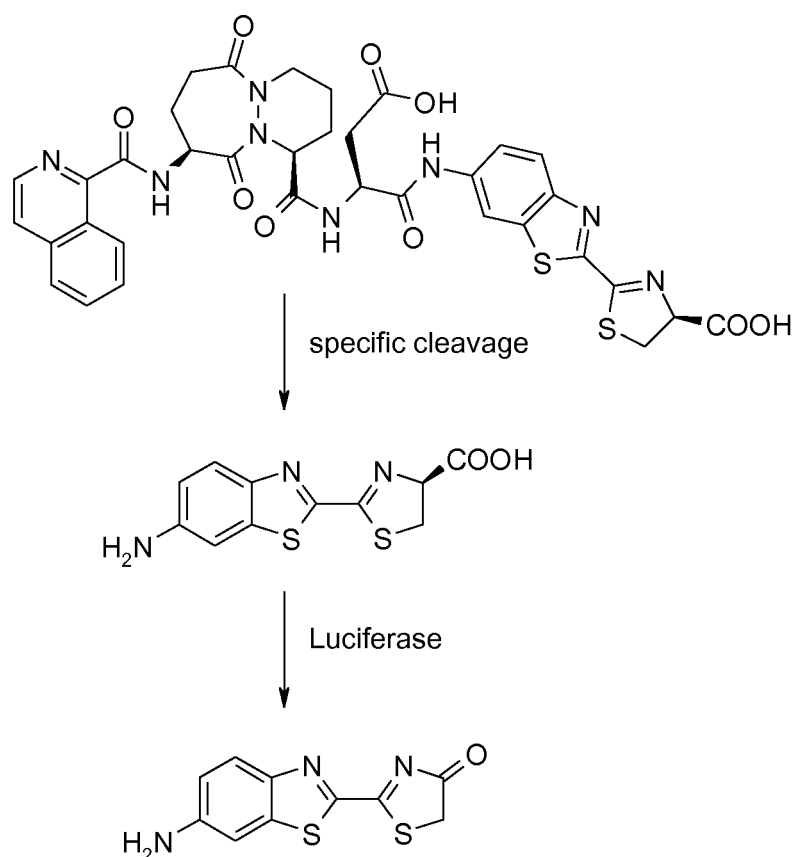


(Scheme 10).

In a further preferred embodiment, the probes of the formula (I) are designed to be used in an homogeneous enzyme linked luminescence assay. The following scheme shows the above-mentioned mechanism of action generically. The luciferine is a substrate for luciferase and a luminescent signal will be generated by a second enzymatic reaction:



The following scheme shows the above-mentioned mechanism of action, where luciferine is masked with a pyridazinodiazepine-derivative and liberated through the proteolytic activity of said caspase-1:



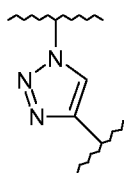
The invention further relates to a method for the design of a molecular probe for the observation of the catalytic activity of one individual proteolytic enzyme or groups of proteolytic enzymes, such as e.g. one caspase or several caspases, in *in vitro* assays, in cells or in multicellular organisms, characterized in transforming an inhibitor for an individual proteolytic enzyme or a group of proteolytic enzymes into a selective imaging probe for these individual proteolytic enzyme or group of proteolytic enzymes, preferably caspase enzymes. To achieve this we replace the electrophilic groups of certain known caspase inhibitors with a scissile amide bond. Preferred compounds are synthesized in a way that a detectable signal is generated by the enzymatic (e.g. proteolytic) activity of a specific target. Particularly, preferred probes comprise internally quenched fluorophores (e.g. appropriate FRET-pairs) linked to (i) the specificity determinant A at the N-terminal portion of the scissile bond and (ii) at the C-terminal portion of the scissile bond. The invention allows for the transfer elements of desirable and previously optimized properties of known inhibitors into novel activity based probes.

Caspase inhibitors described in the prior art utilize an electrophilic warhead in P1 position. The activity based probes of the present invention make use of said known scaffolds and introduce two modifications, firstly the conversion of the electrophilic warhead into a scissile amide group and secondly the positioning of
5 interacting labelling pairs or property modulators on both sides of the scissile amide bound.

In vitro, the reaction of the protease with the substrate of the invention can
10 generally be either performed in cell extracts or with purified or enriched forms of the protease. For *in vivo* application, the reporters are preferably emitters in the near infra red (NIR) region because that region is absent of interfering biofluorescence. Known cyanine NIR dyes matching these requirements are preferably incorporated in the substrates of the present invention.

15 The molecular architecture of compounds of the formula (I) consists of a central scaffold A bearing an amide functional group and two subunits L1R1 and L2R2 respectively. L2R2 is, as shown in formula (I), always connected to scaffold A *via* an amide group since the amide group can be cleaved by the caspase enzyme.
20 Appropriate functional groups for the attachment of subunits L1R1 to scaffold A can be chosen by those skilled in the art, and examples are given below. The specific functional groups L' in the precursor compound can be placed on the scaffold A for the attachment of suitable L1R1 subunits to yield the group L within the compound of the formula (I) are limited only by the requirement of the
25 synthesis strategy and the final use of such substrate as an activity based imaging reagent. Thus their selection will depend upon the specific reagents chosen to build the desired substrates. Examples of functional groups L' which can be provided on scaffold A to connect A with the subunit L1R1 include fluoro, chloro, bromo, cyano, nitro, amino, azido, alkylcarbonylamino, carboxy, carbamoyl,
30 alkoxycarbonyl, aryloxycarbonyl, carbaldehyde, hydroxy, alkoxy, aryloxy, alkylcarbonyloxy, arylcarbonyloxy, a carbon-carbon double bond, a carbon-carbon triple bond, and the like. Most preferable examples include amino, azido, hydroxy,

cyano, carboxy, carbamoyl, carbaldehyde, or a carbon-carbon double or a carbon-carbon triple bond. Thus, L is preferably a direct bond or a group selected from



- , $-(NR_x)-$, $-O-$, $-C=N-$, $-C(=O)-$, $-C(=O)-NH-$, $-NH-C(=O)-$, $-C(=O)H$, $-CR_x=CR_y-$, $-C\equiv C-$ and phenyl, wherein R_x and R_y are independently H or $(C_1-C_6)alkyl$.

In particular, the preferred synthesis of a compound of formula (I) makes use of orthogonally protected functional groups. Such a choice of protective groups allows for a separate deprotection so that each released functionality in turn can be further chemically manipulated towards the attachment of the corresponding subunits to scaffold A. Appropriate protecting groups for the envisioned functionalities can be chosen by those skilled in the art, and are e.g. summarized in T.W. Greene and P.G.M. Wuts in "Protective Groups in Organic Synthesis", John Wiley & Sons, New York 1991.

Compounds of the formula $L'-A-CO-OH$ (scaffolds) can be prepared by standard methods known in the art, e.g. as described in international patent applications US5670494; WO9526958; WO9722619; WO9816504; WO0032620; WO0055127; WO0105772; WO0190063; WO03024955; WO03106460; WO03104231; WO03103677; W. G. Harter, Bioorg. Med. Chem. Lett. 2004, 14, 809-812; Shahripour et al., Bioorg. Med. Chem. Lett. 2001, 11, 2779-2782; Shahripour et al., Bioorg. Med. Chem. 2002, 10, 31-40; M. C. Lauferweiler et al., Bioorg. Med. Chem. Lett. 2005, 15, 4322-4326; K. T. Chapman, Bioorg. Med. Chem. Lett. 1992, 2, 613-618; Dolle et al., J. Med. Chem. 1997, 40, 1941-1946; D. L. Soper et al., Bioorg. Med. Chem. Lett. 2006, 16, 4233-4236; D. L. Soper et al., Bioorg. Med. Chem. 2006, 14, 7880-7892; D. J. Lauffer et al., Bioorg. Med. Chem. Lett. 2002, 12, 1225-1227; C. D. Ellis et al., Bioorg. Med. Chem. Lett. 2006, 16, 4728-4732; P. Tawa et al., Cell Death and Differentiation 2004, 11, 439-447;

Micale et al., J. Med. Chem. 2004, 47, 6455-6458; Berger et al., Molecular Cell, 2006, 23, 509-521; and Garcia-Calvo, J. Biol. Chem. 1998, 273 (49), 32608-32613.

- 5 The present invention also relates to a method for the preparation of a compound of the formula (I) characterized in, if n is 1:

(a) a compound of the formula (II)



wherein A is as defined above in its generic and preferred meanings and L' is fluoro, chloro, bromo, cyano, nitro, amino, azido, alkylcarbonylamino, carboxy, carbamoyl, alkoxy, carbonyl, aryloxy, carbonyl, hydroxy, alkoxy, 15 aryloxy, alkylcarbonyloxy, arylcarbonyloxy, a carbon-carbon double bond, a carbon-carbon triple bond, preferably amino, azido, hydroxy, cyano, carboxy, carbamoyl, carbonyl, or a carbon-carbon double or a carbon-carbon triple bond, more preferred amino,

- 20 is reacted under conditions known to a skilled person with a compound of the formula L1-R1-H wherein L1 is as defined above in its generic and preferred meanings to a compound of the formula (III)



(b) the compound (III) is reacted with a compound H₂N-R2-L2 to a compound of the formula (I).

- Optionally, the synthesis of the compound of the formula (I) makes use of 30 orthogonally protected functional groups. Such a choice of protective groups allows for a separate deprotection so that each released functionality in turn can be further manipulated chemically either to attach a label to it or for the

introduction of further extension of the linker R1 and/or R2. Appropriate protecting groups for the envisioned functionalities can be chosen by those skilled in the art, and are e.g. summarized in T.W. Greene and P.G.M. Wuts in "Protective Groups in Organic

5 Synthesis", John Wiley & Sons, New York 1991.

A further method for the preparation of the probe of the formula (I) wherein n is 1 comprises

10 (a1) the reaction of a compound of the formula (II) with a compound of the formula (IV)



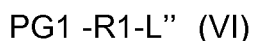
15 to a compound of the formula (V)



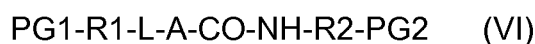
under conditions known to the skilled person,

20

(b) subsequently reacting the compound (V) with a compound (VI)



25 to a compound



under conditions known to the skilled person for the respective groups,

30

wherein PG1 and PG2 are independent of each other protecting groups, preferably orthogonally protecting groups,

L'' is the respective connecting group for L' to be selected by the person skilled in the art, or bond,

(c1) the group PG2 of the compound (VI) is cleaved and the resulting compound is reacted with a label L2, and subsequently the protecting group PG1 is cleaved and the resulting compound is reacted with a label L1 to a compound of the formula (I), or

(c2) the group PG1 of the compound (VI) is cleaved and the resulting compound is reacted with a label L1, and subsequently the protecting group PG2 is cleaved and the resulting compound is reacted with a label L2 to a compound of the formula (I).

In step (b), preferred combinations of L' and L'' and reaction types (in brackets) are as follows:

When L' is fluoro, chloro, bromo, iodo, L'' is amino (R-NH₂), hydroxy (R-OH), triple-bond (Sonogashira Reaction), a double bond (Heck reaction), an alkyl borane (Suzuki-reaction);

when L' is cyano, L'' is amino (R-NH₂), hydroxy (R-OH), thiol (R-SH);

when L' is amino, L'' is an activated carboxylic acid (NHS-ester,...), an carbaldehyde, fluoro, chloro, bromo, iodo;

when L' is azido, L'' is a triple bond, a phosphine moiety (Staudinger ligation);

when L' is carboxy, L'' is amino, hydroxyl, hydrazide;

when L' is alkoxycarbonyl, L'' is amino, hydroxyl, hydrazide;

when L' is aryloxy carbonyl, L'' is amino, hydroxyl, hydrazide;

when L' is hydroxy, L'' is fluoro, chloro, bromo, iodo, hydroxy (Mitsunobu-reaction), carboxy;

when L' is carbaldehyde, L'' is amino, hydrazine;

when L' is carbon-carbon double bond, bromo, chloro, iodo (Heck reaction), an alkyl borane (Suzuki-reaction);

when L' is a carbon-carbon triple bond, L'' is bromo, chloro, iodo (Sonogashira Reaction), azido.

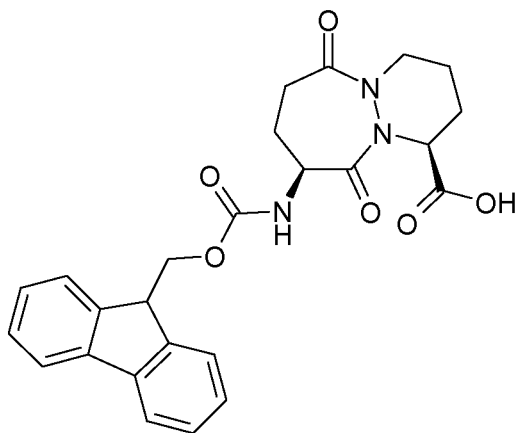
Compounds of the formula (I) wherein n is 0 can be prepared by reacting a compound of the formula A-CO-OH (IV) with a compound H₂N-R₂-L₂ to the probe of the formula (I).

5

Preferably cysteine protease substrates functionalized with different label are synthesized on the solid support.

For the synthesis of caspase probes of the formula (I) with a peptidomimetic structure non-peptidic building blocks may be utilized for the solid-phase synthesis. Building block syntheses are further described in Examples 8.

Building block (VII) is preferably used for the synthesis of caspase-1 probes, e.g. the compounds of Examples 1 and 2,



15

(VII).

The probes of the present inventions are preferably probes for caspase-1, caspase-3 or caspase-8.

The probes of the present invention are used in the context of molecular imaging *in vitro*, in cell-culture experiments, *ex-vivo* experiments or in a living organism (*in vivo*), including screening and whole animal imaging. Mostly preferred are imaging modalities such as optical imaging and magnetic resonance imaging (MRI).

20

The probes of the present invention are intended to be used for diagnostic imaging of protease activity. Most preferred are applications which provide methods of monitoring the effect of a drug or drug-like substance towards the targeted proteases. Administration of such a drug or drug like substance should
5 have a measurable effect to the signal from the probe of the present invention.

A further most preferred aspect of the probes of the present invention is their use as imaging reagents in surgical guidance and to monitor the effect of medical treatment. Surgical guidance includes the detection of tumour margin and
10 detection of progression of tumour metastasis.

Therefore, a further aspect of the present invention is method of imaging a living organism, comprising:

- (a) administering to said organism a probe of the formula (I),
- 15 (b) exposing said organism to electromagnetic radiation which excites non-quenched fluorophore to produce a detectible signal, and
- (c) detecting said signal and creating an image thereby.

Alternatively, the method of imaging a living organism comprises:

- 20 a) administering to said organism a probe of the formula (I),
- (b) exposing said organism to electromagnetic radiation which excites fluorophore to produce a detectible signal; and
- (c) detecting said signal and creating an image thereby.

25 A "living organism" may be any live cell or whole organism comprising the cysteine protease to-be-detected, preferably the living organism is a mammal, e.g. a mouse or a rat.

The probes of the present invention are highly selective, whereby a risk of false
30 positives can be avoided.

Abbreviations:

DMF = dimethylformamide

DMSO = dimethylsulfoxide

DCM = dichloromethane

5 equiv. = equivalents

sat. = saturated

THF = tetrahydrofuran

DIPEA = diisopropylethyl amine

HOAt = 1-Hydroxy-7-azabenzotriazole

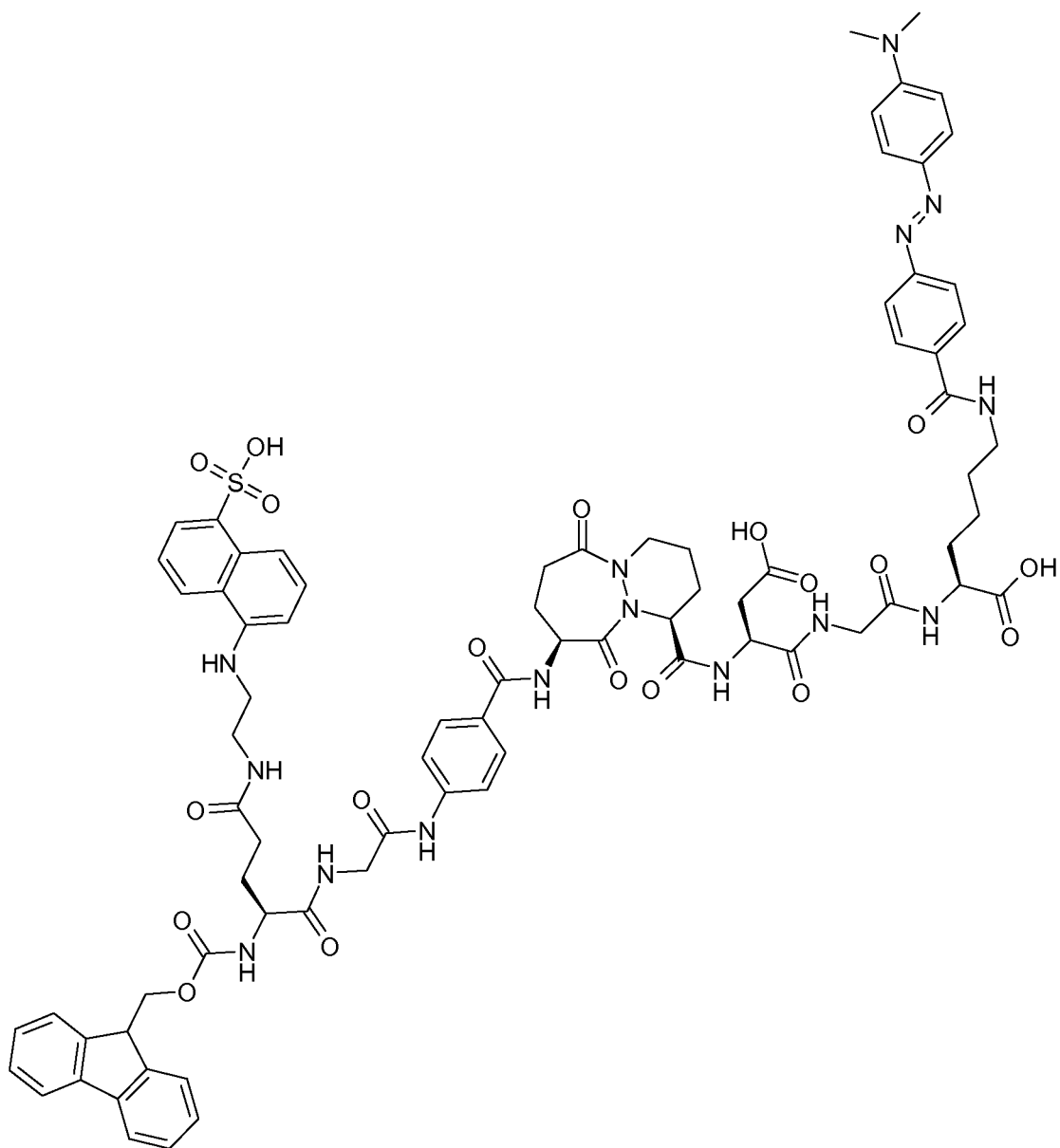
10 HATU = *O*-(7-Azabenzotriazol-1-yl)-*N,N,N',N'*-tetramethyluronium
hexafluorophosphate,

NHS = N-hydroxysuccinimidyl ester

General Procedure for Solid Phase Peptide Synthesis:

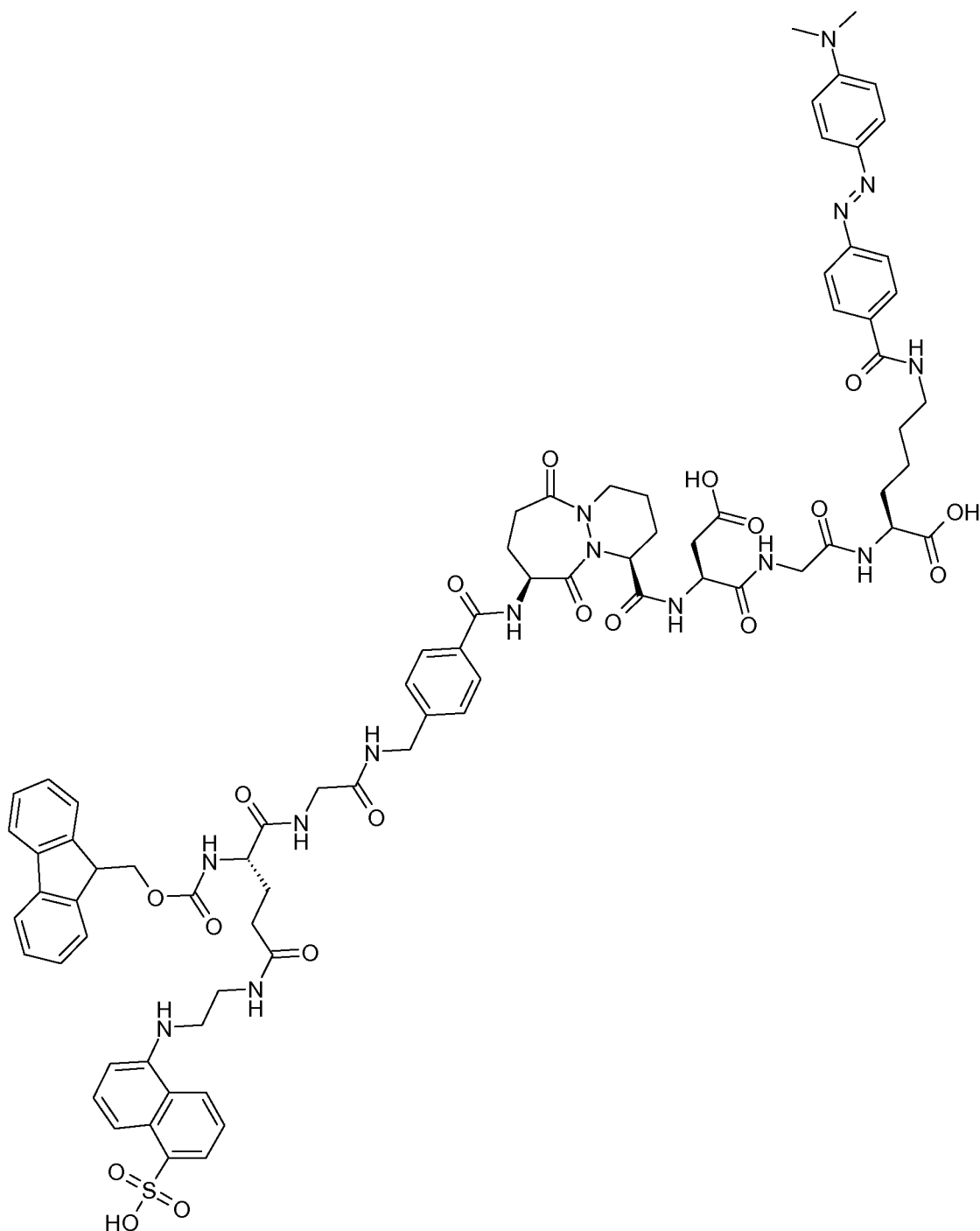
15

The following probes were synthesized using standard solid phase peptide synthesis. The 2-chlorotrityl-resin was used as solid support. For the loading of the resin, 2 equiv. of Fmoc-protected amino acid and 3 equiv. of DIPEA were solved in DCM and the reaction mixture was added to the resin (loading: 1.4 mmol/g). The reaction mixture was shaken at room temperature over night. The resin was washed with DCM and DMF. For Fmoc-deprotection the resin was treated two times for 15 minutes with 30% piperidine/DMF solution. For solid phase peptide synthesis a standard protocol was used: 4 equiv. of Fmoc-protected amino acid, 4 equiv. of HATU, 4 equiv. of HOAt and 8 equiv. of DIPEA were solved in a mixture of DCM/DMF (1/1). The reaction mixture was stirred at room temperature for 20 minutes and then added to the resin. The reaction mixture was shaken for 2 hours or longer if the Fmoc-protected amino acid were sterically hindered. For cleavage from the solid phase, the resin was treated with 5% TFA in DCM two times for 15 minutes. The solvent was coevaporated with toluene under reduced pressure and the final product was purified by preparative HPLC (Gradient: H₂O+0.05% TFA; 5 to 95% CH₃CN).

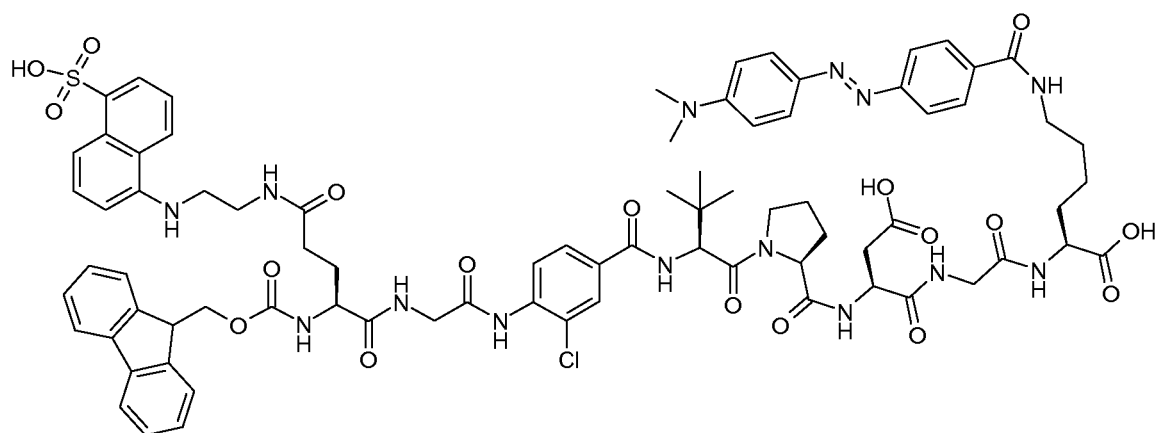
Example 1: Caspase-1 probe

The compound was prepared on solid-support according to the general procedure and purified by HPLC (H₂O+0.05% TFA; 4-95% CH₃CN). Calculated: [M+H]⁺ =

5 1569,70, found: [M+H]⁺ = 1569,45. Yield: 54%.

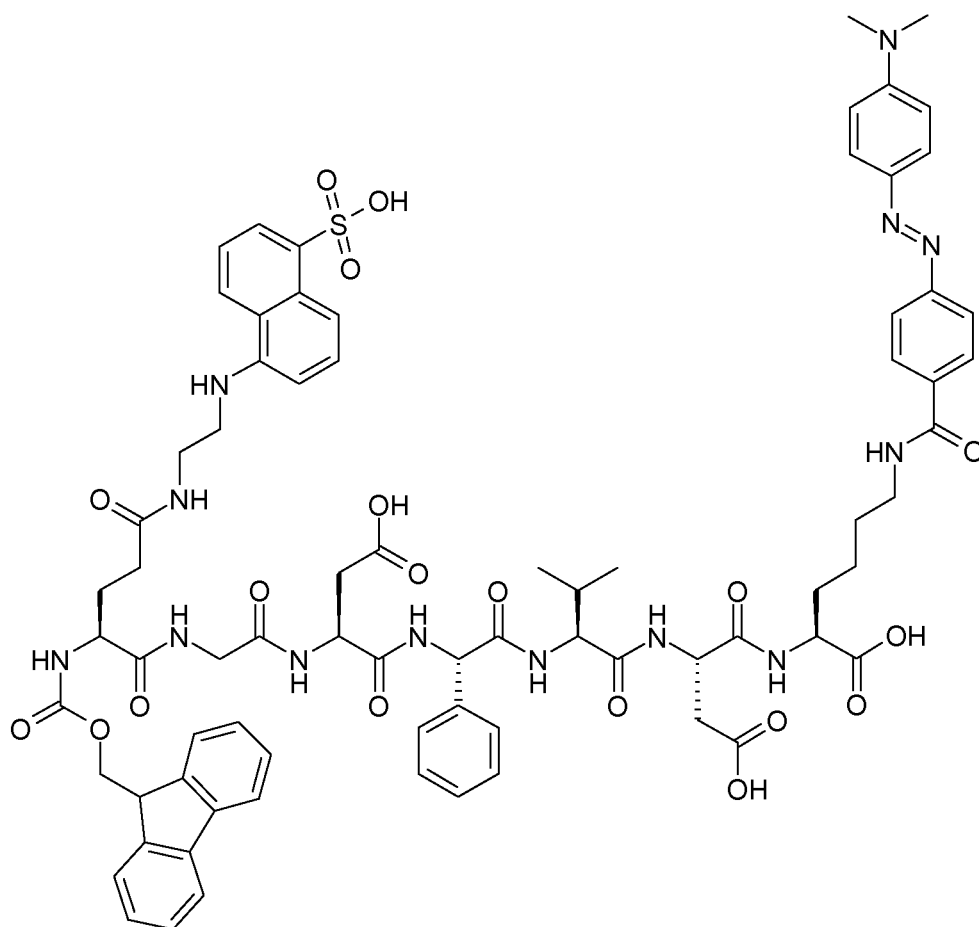
Example 2: Caspase-1 probe

The compound was prepared on solid-support according to the general procedure
5 and purified by HPLC (H₂O+0.05% TFA; 4-95% CH₃CN). Calculated: [M+H]⁺ =
1583,73 found: [M+H]⁺ = 1583,2 . Yield: 72%.

Example 3: Caspase-1 probe

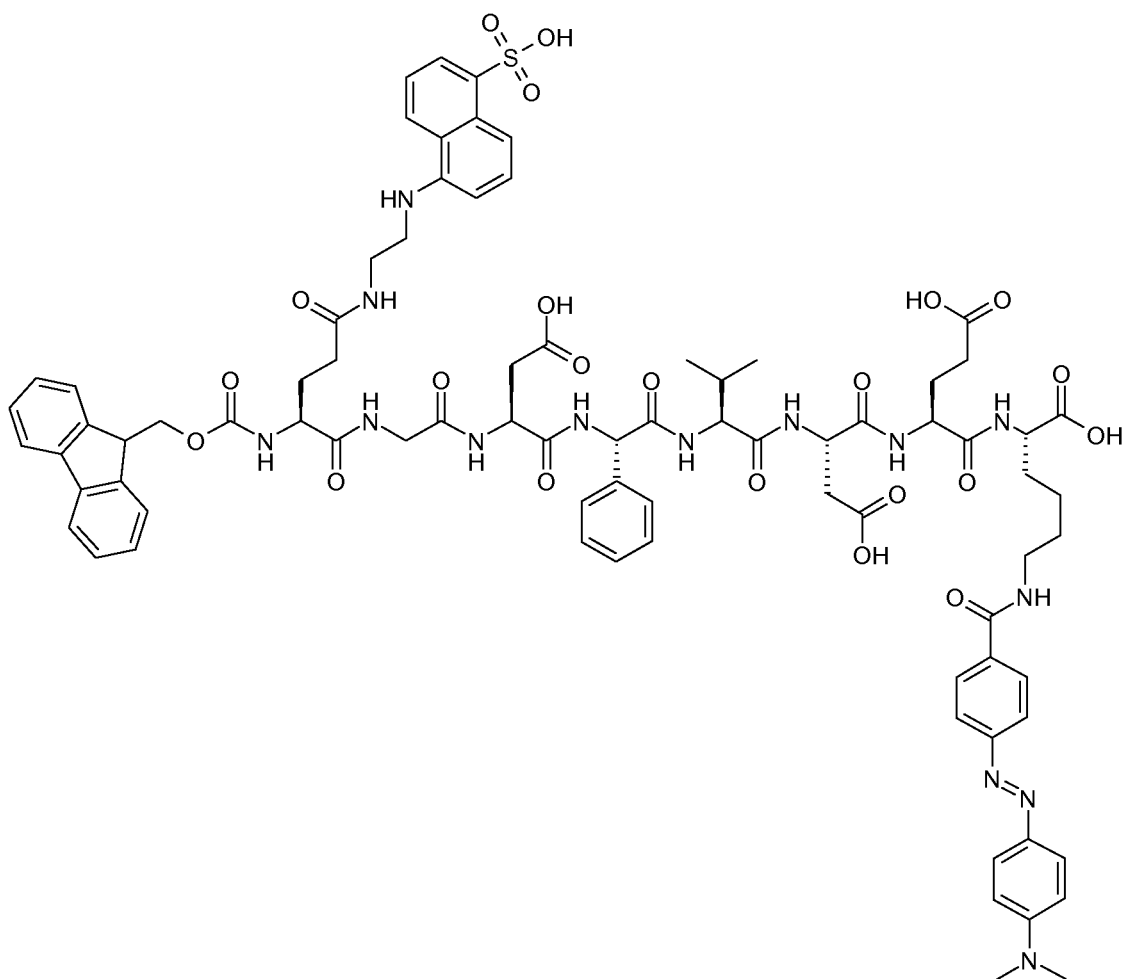
The compound was prepared on solid-support according to the general procedure and purified by HPLC (H₂O+0.05% TFA; 4-95% CH₃CN). Calculated: [M+H]⁺ =

5 1591,19 found: [M+H]⁺ = 1591,50. Yield: 66%.

Example 4: Caspase-3 probe

The compound was prepared on solid-support according to the general procedure and purified by HPLC (H₂O+0.05% TFA; 4-95% CH₃CN). Calculated: [M+H]⁺ = 1517,66 found: [M+H]⁺ = 1517,55. Yield: 59%.

5 Example 5: Caspase-3 probe

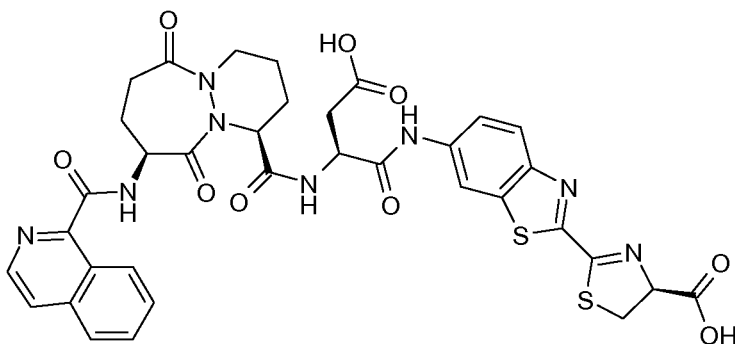


The compound was prepared on solid-support according to the general procedure and purified by HPLC (H₂O+0.05% TFA; 4-95% CH₃CN). Calculated: [M+H]⁺ = 1546,79 found: [M+H]⁺ = 1546,35. Yield: 61%.

Building block (VII) has been prepared according to the procedure described in WO9722619.

Example 8: Caspase-1 bioluminescent probe

5

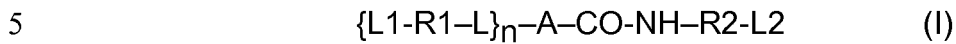


The compound was prepared on solid-support, starting from 6-Fmoc-Amino-D-Luciferin, according to the general procedure and purified by HPLC (H₂O+0.05% TFA; 4-95% CH₃CN). Calculated: [M+H]⁺ = 772.82, found: [M+H]⁺ = 773.15. Yield:

10 13%.

Claims:

1. Molecular probe for cysteine proteases of the formula (I)



wherein

A is a group recognizable by a caspase;

10

R1 is a linker;

R2 is a bond or a linker;

L is a bond or a group allowing for a facile conjugation of the group L1;

L1 and L2 are, independent of each other, at least one label optionally bound to a

15 solid support; and

n is 1,

or

20 R2 is a bond;

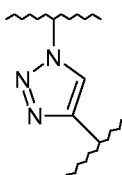
L2 is a substrate, suitable for a coupled bioluminescent assay; and

n is 0.

2. Probe according to claim 1, wherein the caspase is caspase-1, caspase-3

25 or caspase-8.

3. Probe according to any one of claims 1 to 2, wherein L is a direct bond or a group selected from



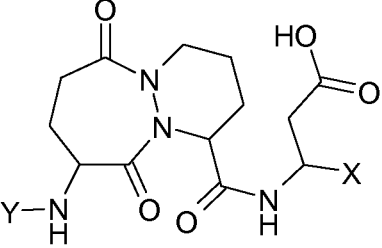
, $-(NR_x)-$, $-O-$, $-C=N-$, $-C(=O)-$, $-C(=O)-NH-$, $-NH-C(=O)-$, $-C(=O)H$, $-CR_x=CR_y-$, $-C\equiv C-$ and phenyl, wherein R_x and R_y are independently H or (C_1-C_6) alkyl.

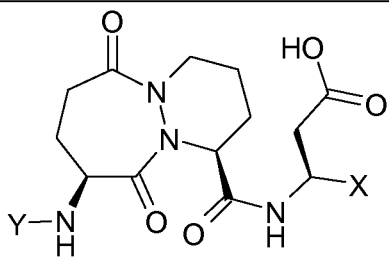
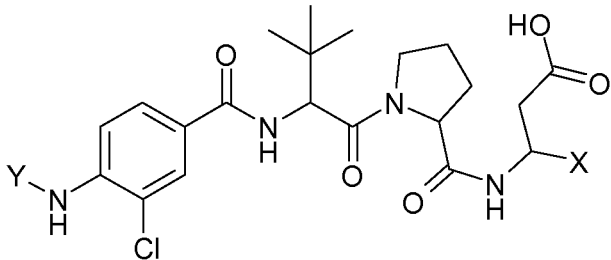
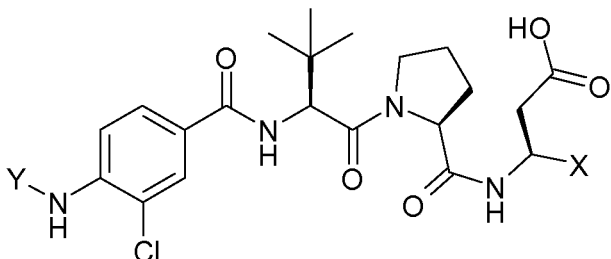
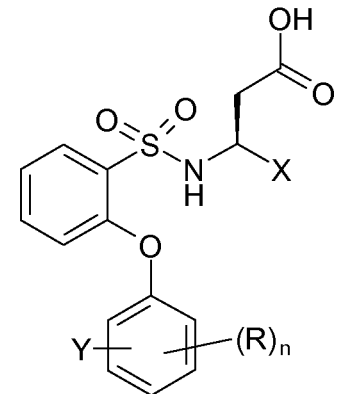
- 5 4. Probe according to any of claims 1 to 3, wherein R_1 or R_2 is a straight or branched chain alkylene group with 1 to 300 carbon atoms, wherein optionally
 - (a) one or more carbon atoms are replaced by oxygen, in particular wherein every third carbon atom is replaced by oxygen, e.g. a polyethyleneoxy group with 1 to 100 ethyleneoxy units; and/or
 - 10 (b) one or more carbon atoms are replaced by nitrogen carrying a hydrogen atom, and the adjacent carbon atoms are substituted by oxo, representing an amide function $-NH-CO-$; and/or
 - (c) one or more carbon atoms are replaced by an ester function $-O-CO-$;
 - (d) the bond between two adjacent carbon atoms is a double or a triple bond;
 - 15 and/or
 - (e) two adjacent carbon atoms are replaced by a disulfide linkage.

- 5 5. Probe according to any one of claims 1 to 4, wherein label L_1 and L_2 are independently of each other a spectroscopic probe such as a fluorophore; a
 - 20 quencher or a chromophore; a magnetic probe; a contrast reagent; a molecule which is one part of a specific binding pair which is capable of specifically binding to a partner; a molecule covalently attached to a solid support, where the support may be a glass slide, a microtiter plate or any polymer known to those proficient in the art; a biomolecule with desirable enzymatic, chemical or physical properties;
 - 25 or a molecule possessing a combination of any of the properties listed above; or a positively charged linear or branched polymer.

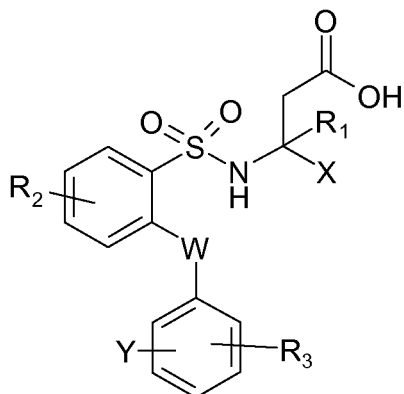
6. Probe according to claim 5, wherein label L_1 and L_2 are independently of each other bound to a positively charged linear or branched polymer.

7. Probe according to claim 6, wherein one label L1 and L2 is a linear poly(arginine) of D- and/or L-arginine with 6 – 15 arginine residues.
- 5 8. Probe according to any one of claims 5 to 7, wherein L1 is one member and L2 is the other member of two interacting spectroscopic probes L1 / L2.
9. Probe according to claim 8, wherein L1 / L2 is a FRET pair.
- 10 10. Probe according to claim 9, wherein one L1 / L2 is a fluorophor selected from Alexa 350, dimethylaminocoumarin, 5/6-carboxyfluorescein, Alexa 488, ATTO 488, DY-505, 5/6-carboxyfluorescein, Alexa 488, Alexa 532, Alexa 546, Alexa 555, ATTO 488, ATTO 532, tetramethylrhodamine, Cy 3, DY-505, DY-547, Alexa 635, Alexa 647, ATTO 600, ATTO 655, DY-632, Cy 5, DY-647 Cy 5.5, and
15 the other label L1 / L2 is a quencher selected from Dabsyl, Dabcyl, BHQ 1, QSY 35, BHQ 2, QSY 9, ATTO 540Q, BHQ 3, ATTO 612Q, QSY 21.
11. Probe according to any one of claims 1 to 4, wherein n is 0, R2 is a bond and L2 is a substrate, suitable for a coupled bioluminescent assay characterized
20 in a modified aminoluciferin or a carboxy-terminal protected derivative thereof as a reporter group, which upon cleavage from the central scaffold A can generate a luminescent signal through its conversion by a luciferase.
12. Selective caspase-1 probe according to any one of claims 1 to 11
25 characterized in a compound 1. – 28. of the table below:

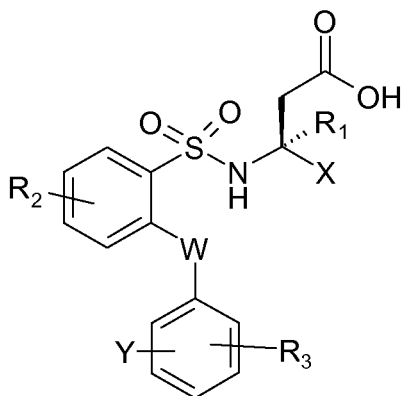
1.	 <p>preferably in the configuration:</p>
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2.	 <p>preferably in the configuration:</p> 
3.	 <p>wherein R is (C₁-C₅)alkyl, phenyl, (C₅-C₆)cycloalkyl; and n is 1-3;</p>

4.



preferably in the configuration:

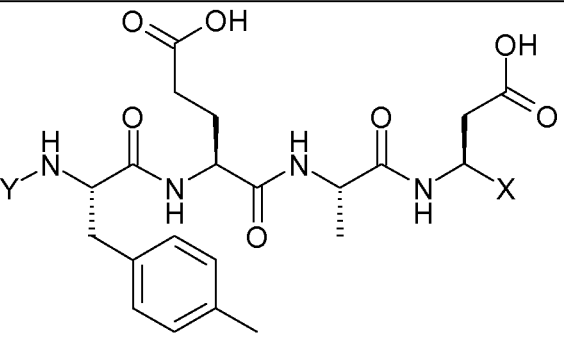
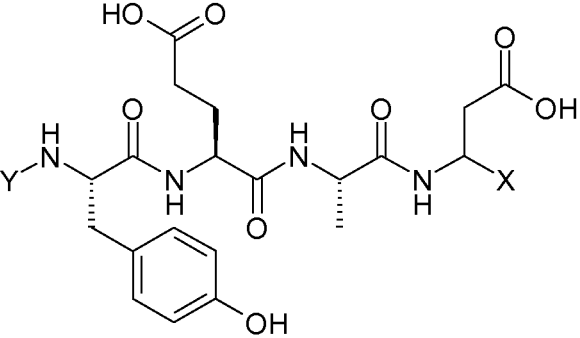
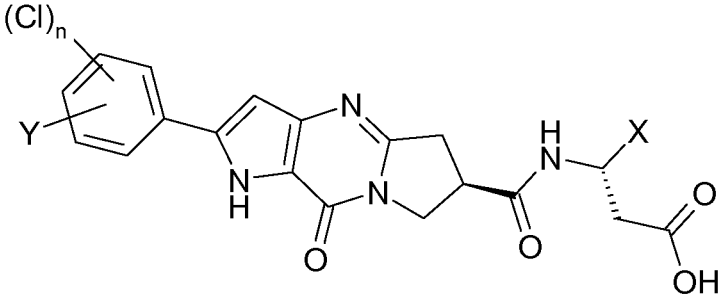
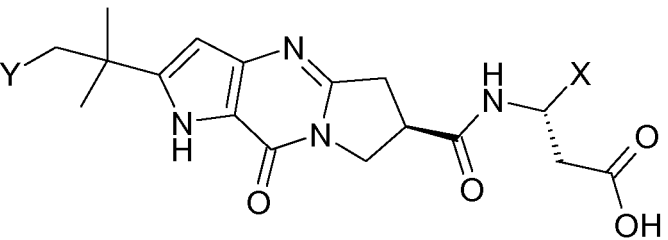
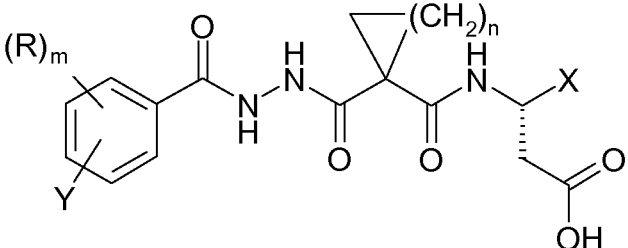


wherein R_1 is hydrogen, (C_1-C_6) alkyl, aryl or $-CH_2$ -aryl,
 R_2 and R_3 are independently of each other hydrogen, or an aryl,
 substituted aryl, heteroaryl, substituted heteroaryl, cycloalkyl,
 substituted cycloalkyl, heterocycle, or substituted heterocycle group
 that is fused to the phenyl group that contains a group R_2 as a
 substituent;

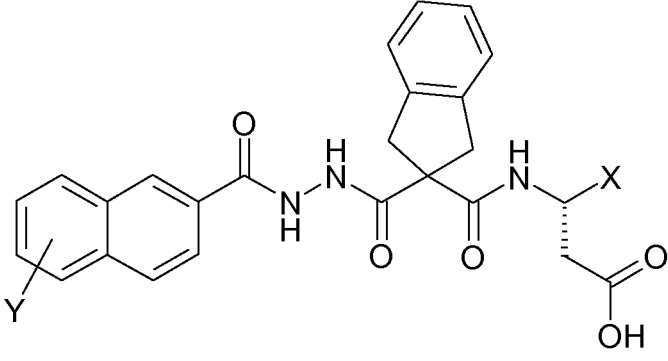
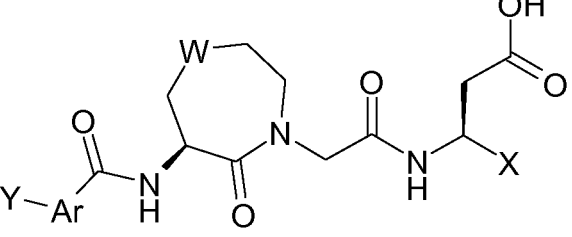
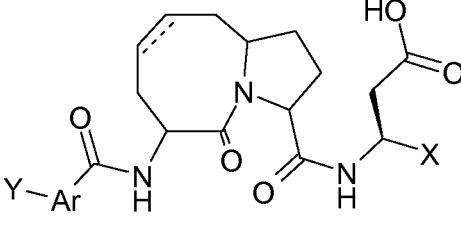
W is a bond, NR_5 , CO , S , O , SO_2 , $O(CHR_5)_n$, CHR_5 ,

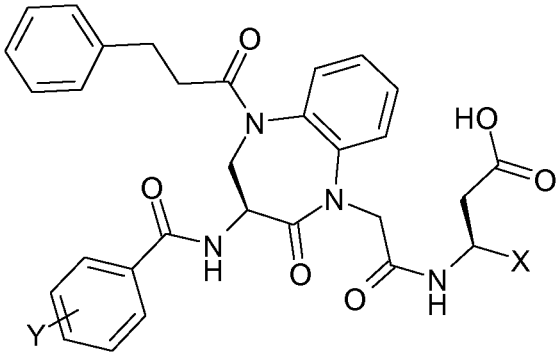
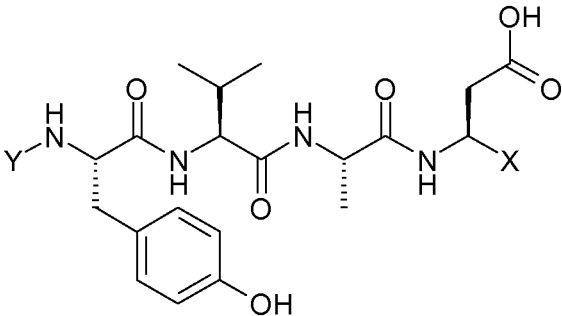
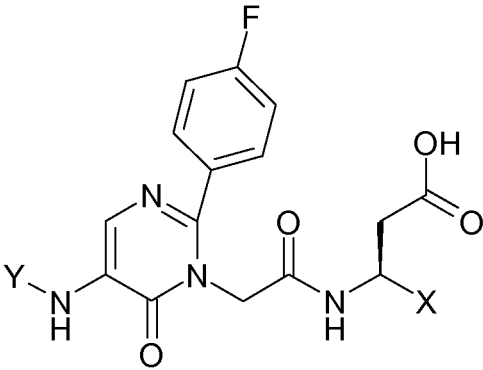
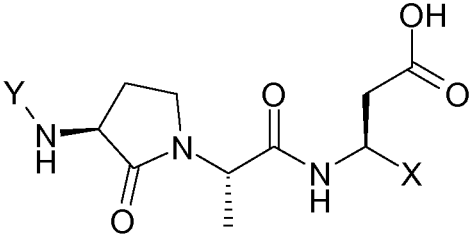
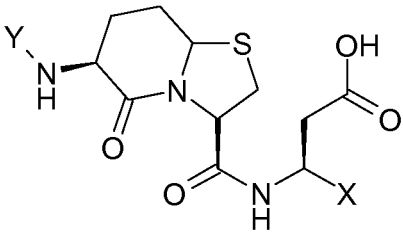
NR_5CO , $CONR_5$, $OCHR_5$, CHR_5O , $SCHR_5$, CHR_5S , SO_2NR_5 , (C_1-C_6) alkyl, NR_5SO_2 , CH_2CHR_5 , CHR_5CH_2 , $COCH_2$ or CH_2CO ;

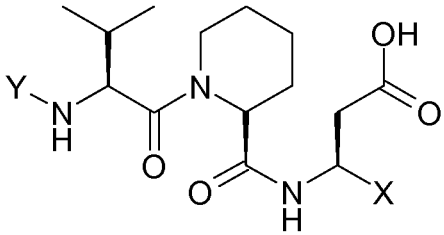
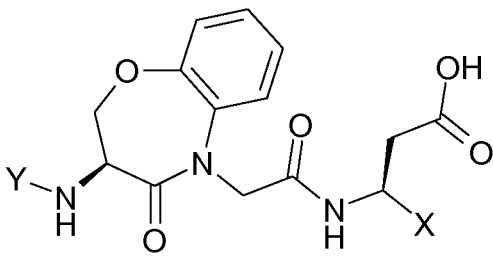
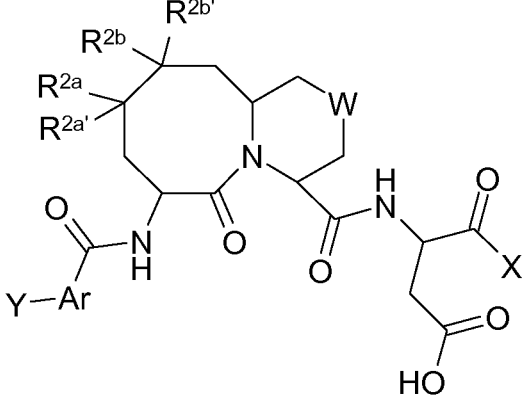
wherein R_5 is independently hydrogen, (C_1-C_6) alkyl, aryl, $(CH_2)_n$ -aryl, or $(CH_2)_n$ -cycloalkyl; and each n is independently 0 to 5;

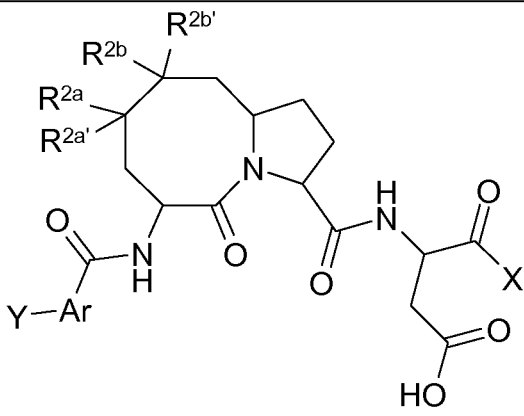
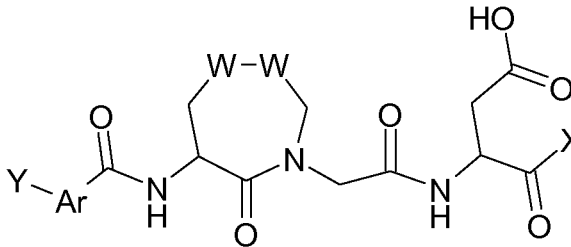
6.	
7.	
8.	 <p>wherein n is 0 or 1;</p>
9.	
10.	

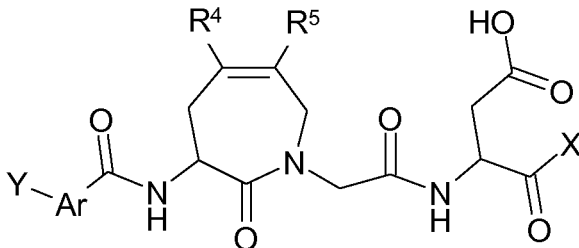
	wherein n is 1-4 m is 1 or 2, and R is methyl or methoxy;
11.	<p>wherein n is 1-4;</p>
12.	<p>wherein n is 1-4;</p>
13.	<p></p>
14.	<p></p>

15.	
16.	 <p>wherein W is S or S(O)₂, and Ar is aryl or heteroaryl, preferably phenyl, naphthyl, benzothiophene or isoquinolyl;</p>
17.	 <p>wherein Ar is an aryl or heteroaryl group selected from phenyl, benzothiophene, isoquinolyl, cinnamyl, naphthyl, which is optionally once or independently twice substituted by methoxy, chloro, methyl, CF₃, and wherein</p> <p></p> <p>means either a single or a double bond;</p>

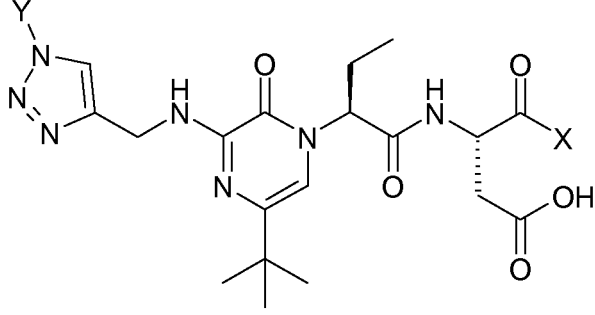
18.	
19.	
20.	
21.	
22.	

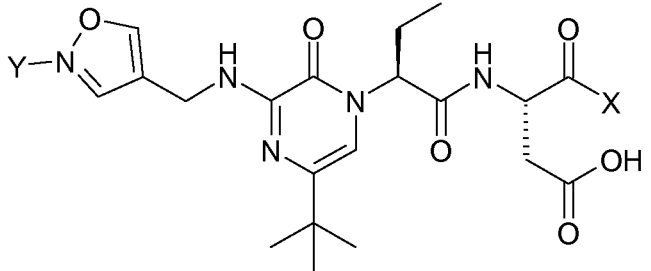
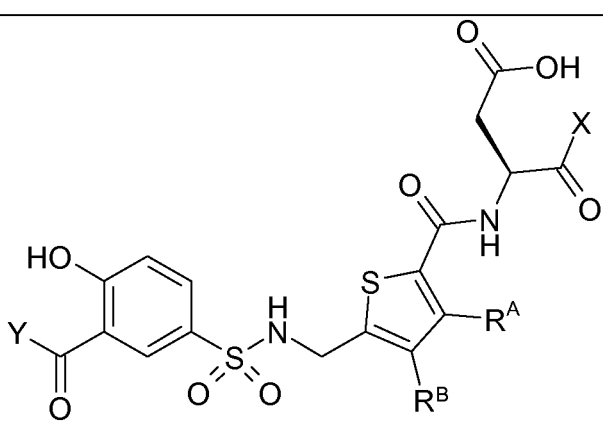
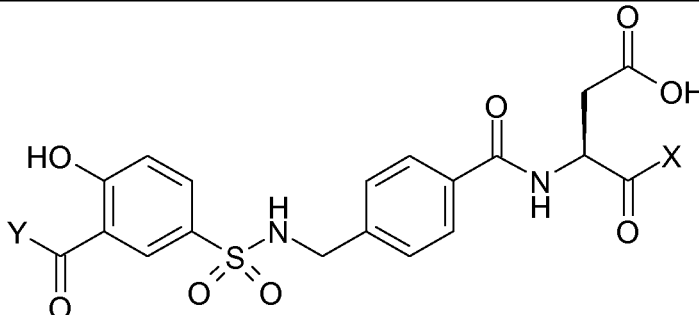
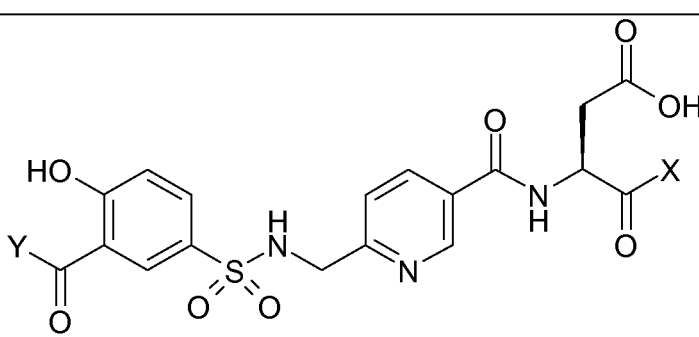
23.	
24.	
25.	 <p>preferably in the all-(S) configuration, wherein Ar is aryl or heteroaryl; W is CH₂, O or NR₉ wherein R₉ is hydrogen or (C₁-C₆)alkyl, aryl, heteroaryl, heterocyclyl; R^{2a}, R^{2a'}, R^{2b} and R^{2b'} are each independently hydrogen, hydroxyl, N(R⁶)₂, halogen, (C₁-C₄)alkyl, (C₁-C₄)alkoxy, and mixtures thereof wherein R⁶ is hydrogen, (C₁-C₆)alkyl, cycloalkyl, (C₆-C₁₀)aryl; or R^{2a} and R^{2b} can taken together to form a double bond;</p>

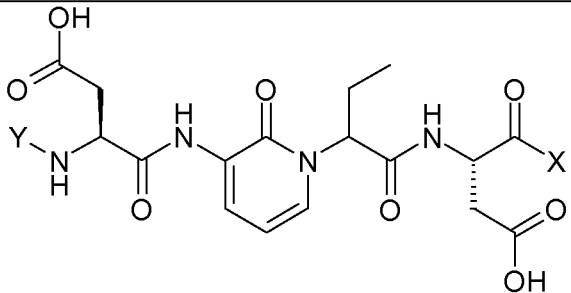
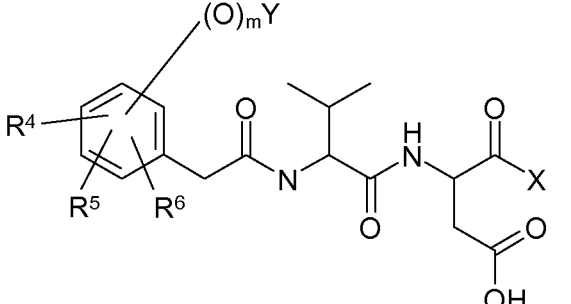
26.	 <p>preferably in the all-(S) configuration, wherein Ar is aryl or heteroaryl; R^{2a}, R^{2a'}, R^{2b} and R^{2b'} are each independently hydrogen, hydroxyl, N(R⁶)₂, halogen, (C₁-C₄)alkyl, (C₁-C₄)alkoxy, and mixtures thereof wherein R⁶ is hydrogen, (C₁-C₆)alkyl, cycloalkyl, (C₆-C₁₀)aryl; or R^{2a} and R^{2b} can taken together to form a double bond;</p>
27.	 <p>preferred in the all-(S) configuration wherein Ar is aryl or heteroaryl; and W is independently selected from: C(R¹)₂; C(O); NR²; S; S(O); S(O)₂; wherein R¹ and R² are independently hydrogen, [C(R³)₂]_p(CH=CH)_qR³, C(=Z)R³, C(=Z)[C(R³)₂]_p(CH=CH)_qR³, C(=Z)N(R³)₂, C(=Z)NR³N(R³)₂, CN, CF₃, N(R³)₂, NR³CN, NR³C(=Z)R³, NRC(=Z)N(R³)₂, NHN(R³)₂, NHOR³, NO₂, OR³, OCF₃, F, Cl, Br, I, SO₃H, OSO₃H, SO₂N(R³)₂, SO₂R³,</p>

	$P(O)(OR^3)R^3$, $P(O)(OR^3)_2$; wherein p is 0 to 12; wherein q is 0 to 12; wherein Z is O, S, NR^3 ; wherein R^3 is independently hydrogen, alkyl, cycloalkyl, aryl, heteroaryl, heterocyllyl.
28.	 <p>preferred in the all-(S) configuration</p> <p>wherein Ar is aryl or heteroaryl; and R^4 and R^5 are independently selected from: $C(R^1)_2$; $C(O)$; NR^2; S; $S(O)$; $S(O)_2$; wherein R^1 and R^2 are independently hydrogen, $[C(R^3)_2]_p(CH=CH)_qR^3$, $C(=Z)R^3$, $C(=Z)[C(R^3)_2]_p(CH=CH)_qR^3$, $C(=Z)N(R^3)_2$, $C(=Z)NR^3N(R^3)_2$, CN, CF_3, $N(R^3)_2$, NR^3CN, $NR^3C(=Z)R^3$, $NRC(=Z)N(R^3)_2$, $NHN(R^3)_2$, $NHOR^3$, NO_2, OR^3, OCF_3, F, Cl, Br, I, SO_3H, OSO_3H, $SO_2N(R^3)_2$, SO_2R^3, $P(O)(OR^3)R^3$, $P(O)(OR^3)_2$; wherein p is 0 to 12; wherein q is 0 to 12; wherein Z is O, S, NR^3; wherein R^3 is independently hydrogen, alkyl, cycloalkyl, aryl, heteroaryl, heterocyllyl.</p>

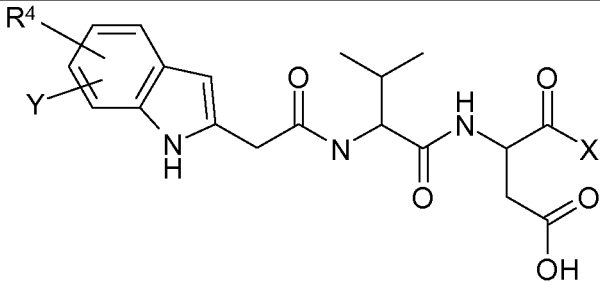
13. Selective caspase-3 probe according to any one of claims 1 to 11 characterized in a compound 29. – 42. of the table below :

29.	
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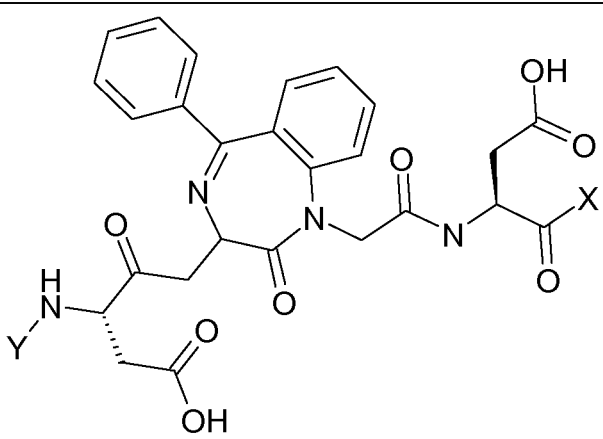
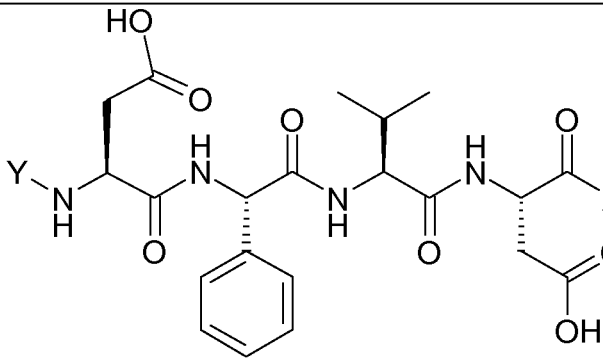
30.	
31.	 <p>wherein R^A and R^B are independently hydrogen, (C₁-C₆)alkyl, hydroxyl, (C₁-C₆)alkoxy or halogen.</p>
32.	
33.	

34.	
35.	 <p>preferably in the all-(S) configuration, wherein m is 0 or 1; and R⁴, R⁵ and R⁶ are independently selected from the group consisting of:</p> <ol style="list-style-type: none"> 1) H, 2) halogen, 3) (C₁-C₄)alkoxy optionally substituted with 1-3 halogen atoms, 4) NO₂, 5) OH, 6) benzyloxy, the benzyl portion of which is optionally substituted with 1-2 members selected from the group consisting of: halogen, CN, (C₁-C₄)alkyl and (C₁-C₄)alkoxy, said alkyl and alkoxy being optionally substituted with 1-3 halogen groups, 7) NH(C₁-C₄)acyl, 8) (C₁-C₄)acyl, 9) O-(C₁-C₄)alkyl-CO₂H, optionally esterified with a (C₁-C₆)alkyl or a (C₅-C₇)cycloalkyl group, 10) CH=CH-CO₂H,

- | | |
|--|--|
| | <p>11) CO₂H,</p> <p>12) (C₁-C₅)alkyl-CO₂H,</p> <p>13) C(O)NH₂, optionally substituted on the nitrogen atom by 1-2 (C₁-C₄)alkyl groups;</p> <p>14) (C₁-C₅)alkyl-C(O)NH₂, optionally substituted on the nitrogen atom by 1-2 (C₁-C₄)alkyl groups;</p> <p>15) S(O)₀₋₂-(C₁-C₄)alkyl;</p> <p>16) (C₁-C₂)alkyl-S(O)₀₋₂-(C₁-C₄)alkyl;</p> <p>17) S(O)₀₋₂-(C₁-C₆)alkyl or S(O)₀₋₂-phenyl, said alkyl and phenyl portions thereof being optionally substituted with 1-3 members selected from the group consisting of: halogen, CN, (C₁-C₄)alkyl and (C₁-C₄)alkoxy, said alkyl and alkoxy being optionally substituted by 1-3 halogen groups,</p> <p>18) benzoyl optionally substituted by 1-2 members selected from the group consisting of: halogen, CN, (C₁-C₄)alkyl and (C₁-C₄)alkoxy, said alkyl and alkoxy groups being optionally substituted by 1-3 halogen groups,</p> <p>19) phenyl or naphthyl, optionally substituted with 1-2 members selected from the group consisting of: halogen, CN, (C₁-C₄)alkyl and (C₁-C₄)alkoxy, said alkyl and alkoxy being optionally substituted with 1-3 halogen groups,</p> <p>20) CN,</p> <p>21) (C₁-C₄)alkylene-HET2, wherein HET2 represents a 5-7 membered aromatic or non-aromatic ring containing 1-4 heteroatoms selected from O, S, NH and N(C₁-C₄) and optionally containing 1-2 oxo groups, and optionally substituted with 1-3 (C₁-C₄)alkyl, OH, halogen or (C₁-C₄)acyl groups;</p> <p>22) O-(C₁-C₄)alkyl-HET3, wherein HET3 is a 5 or 6 membered</p> |
|--|--|

	<p>aromatic or non-aromatic ring containing from 1 to 3 heteroatoms selected from O, S and N, and optionally substituted with one or two groups selected from halogen and (C₁-C₄)alkyl, and optionally containing 1-2 oxo groups, and</p> <p>23) HET4, wherein HET4 is a 5 or 6 membered aromatic or non-aromatic ring, and the benzofused analogs thereof, containing from 1 to 4 heteroatoms selected from O, S and N, and is optionally substituted by one or two groups selected from halogen, (C₁-C₄)alkyl and (C₁-C₄)acyl; and wherein halogen includes F, Cl, Br and I.</p>
36.	 <p>preferably in the all-(S) configuration,</p> <p>wherein R⁴ is selected from the group consisting of:</p> <ol style="list-style-type: none"> 1) H, 2) halogen, 3) (C₁-C₄)alkoxy optionally substituted with 1-3 halogen atoms, 4) NO₂, 5) OH, 6) benzyloxy, the benzyl portion of which is optionally substituted with 1-2 members selected from the group consisting of: halogen, CN, (C₁-C₄)alkyl and (C₁-C₄)alkoxy, said alkyl and alkoxy being optionally substituted with 1-3 halogen groups, 7) NH(C₁-C₄)acyl, 8) (C₁-C₄)acyl, 9) O-(C₁-C₄)alkyl-CO₂H, optionally esterified with a (C₁-C₆)alkyl or a (C₅-C₇)cycloalkyl group,

- | |
|--|
| <p>10) CH=CH-CO₂H,</p> <p>11) CO₂H,</p> <p>12) (C₁-C₅)alkyl-CO₂H,</p> <p>13) C(O)NH₂, optionally substituted on the nitrogen atom by 1-2 (C₁-C₄)alkyl groups;</p> <p>14) (C₁-C₅)alkyl-C(O)NH₂, optionally substituted on the nitrogen atom by 1-2 (C₁-C₄)alkyl groups;</p> <p>15) S(O)₀₋₂-(C₁-C₄)alkyl;</p> <p>16) (C₁-C₂)alkyl-S(O)₀₋₂-(C₁-C₄)alkyl;</p> <p>17) S(O)₀₋₂-(C₁-C₆)alkyl or S(O)₀₋₂-phenyl, said alkyl and phenyl portions thereof being optionally substituted with 1-3 members selected from the group consisting of: halogen, CN, (C₁-C₄)alkyl and (C₁-C₄)alkoxy, said alkyl and alkoxy being optionally substituted by 1-3 halogen groups,</p> <p>18) benzoyl optionally substituted by 1-2 members selected from the group consisting of: halogen, CN, (C₁-C₄)alkyl and (C₁-C₄)alkoxy, said alkyl and alkoxy groups being optionally substituted by 1-3 halogen groups,</p> <p>19) phenyl or naphthyl, optionally substituted with 1-2 members selected from the group consisting of: halogen, CN, (C₁-C₄)alkyl and (C₁-C₄)alkoxy, said alkyl and alkoxy being optionally substituted with 1-3 halogen groups,</p> <p>20) CN,</p> <p>21) (C₁-C₄)alkylene-HET₂, wherein HET₂ represents a 5-7 membered aromatic or non-aromatic ring containing 1-4 heteroatoms selected from O, S, NH and N(C₁-C₄) and optionally containing 1-2 oxo groups, and optionally substituted with 1-3 (C₁-C₄)alkyl, OH, halogen or (C₁-C₄)acyl groups;</p> |
|--|

	<p>22) O-(C₁-C₄)alkyl-HET3, wherein HET3 is a 5 or 6 membered aromatic or non-aromatic ring containing from 1 to 3 heteroatoms selected from O, S and N, and optionally substituted with one or two groups selected from halogen and (C₁-C₄)alkyl, and optionally containing 1-2 oxo groups, and</p> <p>23) HET4, wherein HET4 is a 5 or 6 membered aromatic or non-aromatic ring, and the benzofused analogs thereof, containing from 1 to 4 heteroatoms selected from O, S and N, and is optionally substituted by one or two groups selected from halogen, (C₁-C₄)alkyl and (C₁-C₄)acyl; and wherein halogen includes F, Cl, Br and I.</p>
37.	
38.	

39.	
40.	
41.	
42.	

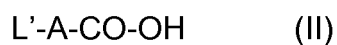
14. Selective caspase-8 probe according to any one of claims 1 to 11 characterized in a compound 43. – 44. of the table below:

43.	
44.	

15. Preparation of a probe of the formula (I) according to claim 1 to 14 characterized in

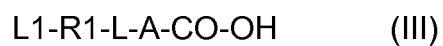
5 if n is 1:

(a) reacting a compound of the formula (II)



10

with a compound of the formula L1-R1-H to a compound of the formula (III)



15 (b) reacting the compound (III) with a compound H₂N-R2-L2 to the probe of the formula (I),

wherein L' is fluoro, chloro, bromo, cyano, nitro, amino, azido, alkylcarbonylamino, carboxy, carbamoyl, alkoxycarbonyl, aryloxy, carbonyl, carbaldehyde, hydroxy, alkoxy, aryloxy, alkylcarbonyloxy, arylcarbonyloxy, a carbon-carbon double bond, a carbon-carbon triple bond, preferably amino, azido, hydroxy, cyano, carboxy, carbamoyl, carbaldehyde, or a carbon-carbon double or a carbon-carbon triple bond, more preferred amino, and

R1 and/or R2 may be protected by suitable orthogonally protecting groups and sequentially cleaved in the course of the preparation of the compound (I); and

if n is 0:

reacting a compound of the formula A-CO-OH (IV) with a compound H₂N-R₂-L₂ to the probe of the formula (I).

16. Use of a probe of the formula (I) according to claims 1 to 14 for molecular imaging in vitro, in cell-culture experiments, ex-vivo experiments or in a living organism.

17. Use of a probe of the formula (I) according to claims 1 to 14 for imaging a living organism, comprising:

- (a) administering to said organism a probe of the formula (I),
- (b) exposing said organism to electromagnetic radiation which excites non-quenched fluorophore to produce a detectible signal, and
- (c) detecting said signal and creating an image thereby.

18. Use of a probe of the formula (I) according to claims 1 to 14 for imaging a living organism, comprising:

- (a) administering to said organism a probe of the formula (I),
- (b) exposing said organism to electromagnetic radiation which excites fluorophore to produce a detectible signal; and
- (c) detecting said signal and creating an image thereby.