COMPOSITIONS AND METHODS RELATING TO OSTEOARTHRITIS

Field of the Invention

The invention relates to the profiling of differential gene expression in specific human tissue types through the construction and use of cDNA libraries and microarrays.

1. Background of the Invention

Osteoarthritis (OA) is a chronic disease in which the articular cartilage that lies on the ends of bones that forms the articulating surface of the joints gradually degenerates over time. There are many factors that are believed to predispose a patient to osteoarthritis including genetic susceptibility, obesity, accidental or athletic trauma, surgery, drugs and heavy physical demands. Osteoarthritis is initiated by damage to the cartilage of joints. The two most common injuries to joints are sports-related injuries and long term "repetitive use" joint injuries. Joints most commonly affected by osteoarthritis are the knees, hips and hands. In most cases, due to the essential weight-bearing function of the knees and hips, osteoarthritis in these joints causes much more disability than osteoarthritis of the hands. As cartilage degeneration progresses, secondary changes occur in other tissues in and around joints including bone, muscle, ligaments, menisci and synovium. The net effect of the primary failure of cartilage tissue and secondary damage to other tissues is that the patient experiences pain, swelling, weakness and loss of functional ability in the afflicted joint(s). These symptoms frequently progress to the point that they have a significant impact in terms of lost productivity and or quality of life consequences for the patient.

Articular cartilage is predominantly composed of chondrocytes, type II collagen, proteoglycans and water. Articular cartilage has no blood or nerve supply and chondrocytes are the only type of cell in this tissue. Chondrocytes are responsible for manufacturing the type II collagen and proteoglycans that form the cartilage matrix. This matrix in turn has physical-chemical properties that allow for saturation of the matrix with water. The net effect of this structural-functional relationship is that articular cartilage has exceptional wear characteristics and allows for almost frictionless movement between the articulating cartilage surfaces. In the absence of osteoarthritis, articular cartilage often provides a lifetime of pain-free weight bearing and unrestricted joint motion even under demanding physical conditions.

During fetal development, articular cartilage is initially derived from the interzone of mesenchymal condensations. The mesenchymal cells cluster together and synthesize matrix proteins. The tissue is recognized as cartilage when the accumulation of matrix separates the cells, which are spherical in shape and are now called chondrocytes. During cartilage formation and growth, chondrocytes proliferate rapidly and synthesize large volumes of matrix. Prior to skeletal maturity, chondrocytes are at their highest level of metabolic activity. As skeletal maturation is reached, the rate of chondrocyte metabolic activity and cell division declines. After completion of skeletal growth, most chondrocytes do not divide but do continue to synthesize matrix proteins such as collagens, proteoglycans and other noncollagenous proteins (1, 2).

Like all living tissues, articular cartilage is continually undergoing a process of renewal in which "old" cells and matrix components are being removed (catabolic activity) and "new" cells and molecules are being produced (anabolic activity). Relative to most tissues, the rate of anabolic/catabolic turnover in articular cartilage is low. Long-term maintenance of the structural integrity of mature cartilage relies on the proper balance between matrix synthesis and degradation. Chondrocytes maintain matrix equilibrium by responding to chemical and mechanical stimuli from their environment. Appropriate and effective chondrocyte responses to these stimuli are essential for cartilage homeostasis. Disruption of homeostasis through either inadequate anabolic activity or excessive catabolic activity can result in cartilage degradation and osteoarthritis (3). Most tissues that are damaged and have increased catabolic activity are able to mount an increased anabolic response that allows for tissue healing. Unfortunately, chondrocytes have very limited ability to up-regulate their anabolic activity and increase the synthesis of proteoglycan and type II collagen in response to damage or loss of cartilage matrix. This fundamental limitation of chondrocytes is the core problem that has precluded the development of therapies that can prevent and cure osteoarthritis. Additionally, there is a need for a definitive diagnostic test for detecting early osteoarthritis, and a prognostic test that effectively monitors a patient's response to therapy.

Joint pain is the most common manifestation of early osteoarthritis. The pain tends to be episodic lasting days to weeks and remitting spontaneously. Although redness and swelling of joints is uncommon, joints become tender during a flare-up of osteoarthritis.

"Mild" or "early stage osteoarthritis" is difficult to diagnose. The physician relies primarily on the patient's history and physical exam to make the diagnosis of mild osteoarthritis. X-rays do not show the underlying early changes in articular cartilage. There are no recognized biochemical markers used to confirm the diagnosis of early stage osteoarthritis.

X-ray changes confirm the diagnosis of moderate osteoarthritis. X-rays of normal joints reveal well preserved symmetrical joint spaces. Changes seen on the x-rays of patients with osteoarthritis include new bone formation (osteophytes), joint space narrowing and sclerosis (bone thickening). There are no recognized biochemical markers used to confirm the diagnosis of "moderate osteoarthritis" at this stage.

The clinical exam of a joint with severe osteoarthritis reveals tenderness, joint deformity and a loss of mobility. Passive joint movement during examination may elicit crepitus or the grinding of bone-on-bone as the joint moves. X-ray changes are often profound: the joint space may be obliterated and misalignment of the joint can be seen. New bone formation (osteophytes) is prominent. Again, there are no recognized biochemical markers used to confirm the diagnosis of "severe osteoarthritis".

"Osteoarthritis" is the most common chronic joint disease. It is characterized by progressive degeneration and eventual loss of cartilage. Currently, there is a need for an effective therapy that will alter the course of osteoarthritis. Further advances in preventing, modifying or curing the osteoarthritic disease process critically depends, at least in part, on a thorough understanding of the molecular mechanisms underlying anabolic and catabolic processes in cartilage. Since cellular functions are substantially determined by the genes that the cells express, elucidating the genes expressed in articular cartilage at different developmental and disease stages will inevitably provide new insights into the molecules and mechanisms involved in cartilage formation, injury, disease and repair.

cDNA libraries from putatively normal and severely osteoarthritic human cartilage tissue have been constructed (Kumar et al., 46th Annual Meeting, Orthopaedic Res. Soc., Abstract, p. 1031). However, this work does not adequately address the differentiation of chondrocyte gene expression from differing severities of osteoarthritic human cartilage (mild, moderate, marked and severe). In addition, the "normal cartilage" samples were obtained from deceased donors more than

24 hours after death. Thus, this cDNA library does not truly reflect normal chondrocyte gene expression due to the rapid degeneration of RNA that occurs after cessation of perfusion to the sampled joint, as demonstrated by baboon studies, presented herein below.

Summary of the Invention

The invention relates to one or more profiles of gene expression for human fetal articular cartilage, and the cartilage of normal, mildly, moderately, markedly and severely osteoarthritic individuals, and thus to a method for identifying genes that play critical roles in cartilage injury, repair and disease progression. Given the inherently low anabolic activity in adult chondrocytes, identification of key replicative and/or anabolic genes expressed by fetal but not adult chondrocytes, has important implications for developing novel disease modifying therapies for adult cartilage injury and osteoarthritis.

One aspect of the invention is to isolate chondrocyte enriched or chondrocyte-specific polynucleotide sequences.

In one embodiment, one or more polynucleotide sequences selected from the group consisting of the sequences identified in Figure 6A which correspond to genes 1-5807 identified in Figure 6 and sequences identified in Figure 13 are isolated.

In another embodiment, a vector comprising one or more polynucleotide sequences selected from the sequences identified in Figure 6A corresponding to genes 1-5807 of Figure 6 and sequences identified in Figure 13 are constructed.

In yet another embodiment, a host cell comprising said vector is constructed.

Another aspect of the invention is to provide a composition comprising one or more chondrocyte enriched or chondrocyte-specific polynucleotide sequences.

Another aspect of the invention is to provide a composition comprising one or more chondrocyte enriched or chondrocyte-specific polynucleotide sequences isolated from one or more of (a) a fetus, (b) normal, (c) mildly osteoarthritic, (d) moderately osteoarthritic, (e) markedly osteoarthritic, or (f) severely osteoarthritic cartilage samples.

Another embodiment of the invention provides a composition comprising one or more polynucleotide sequences selected from the group of sequences identified in Figure 6B which are isolated from a fetal cDNA library as disclosed herein.

Another embodiment of the invention provides a composition comprising one or more polynucleotide sequences selected from the group of sequences identified in Figure 6C which are isolated from a normal cDNA library as disclosed herein.

Another embodiment of the invention provides a composition comprising one or more polynucleotide sequences selected from the group of sequences identified in Figure 6D which are isolated from a mild osteoarthritic chondrocyte library as disclosed herein.

Another embodiment of the invention provides a composition comprising one or more polynucleotide sequences selected from the group of sequences identified in Figure 6E which are isolated from a severe osteoarthritic chondrocyte library as disclosed herein.

Another embodiment of the invention provides a composition comprising one or more polynucleotide sequences selected from the group of sequences identified in Figures 6B, 6C, 6D and 6E.

Another embodiment of the invention provides a composition comprising one or more polynucleotide sequences where at least one of said polynucleotide sequences is differentially expressed in cartilage from a patient diagnosed with mild osteoarthritis relative to cartilage from a normal individual, wherein cartilage isolated from the normal individual is isolated from cartilage tissue obtained less than 14 hours post-mortem.

Another embodiment of the invention provides a composition comprising one or more polynucleotide sequences where at least one of the polynucleotide sequences is differentially expressed in cartilage from a patient diagnosed with severe osteoarthritis relative to cartilage derived from a normal individual, wherein cartilage isolated from the normal individual is isolated from cartilage tissue obtained less than 14 hours post-mortem.

Another embodiment of the invention provides a composition comprising one or more polynucleotide sequences wherein at least one of the polynucleotide sequences is differentially

expressed in cartilage from a patient diagnosed with moderate osteoarthritis relative to cartilage from a normal individual, wherein cartilage isolated from the normal individual is isolated from cartilage tissue obtained less than 14 hours post-mortem.

Another embodiment of the invention provides a composition comprising one or more polynucleotide sequences wherein at least one of the polynucleotide sequences is differentially expressed in cartilage from a patient diagnosed with marked osteoarthritis relative to cartilage derived from a normal individual, wherein cartilage isolated from the normal individual is isolated from cartilage tissue obtained less than 14 hours post-mortem.

Another embodiment of the invention provides a composition comprising one or more polynucleotide sequences wherein at least one of the polynucleotide sequences is differentially expressed in cartilage isolated from any two or more of the following sources (a) fetus, or (b) patient with mild osteoarthritis, (c) patient with moderate osteoarthritis, (d) patient with marked osteoarthritis, (e) patient with severe osteoarthritis or (f) cartilage isolated from cartilage tissue obtained from a normal individual.

Another embodiment of the invention provides a composition comprising one or more polynucleotide sequences identified in Figure 9 and/or sequences identified in Figure 6A which correspond to the genes disclosed in Figure 9.

Another embodiment of the invention provides a composition comprising one or more polynucleotide sequences identified in Figure 11 and/or those sequences identified in Figure 6A which correspond to the genes disclosed in Figure 11.

Another embodiment of the invention provides a composition comprising one or more polynucleotide sequences identified in Figure 6A which correspond to the genes disclosed in Figures 15 and Figures 16.

Another embodiment of the invention provides a composition comprising one or more polynucleotide sequences identified in Figure 6A which correspond to the genes disclosed in Figure 6.

Another embodiment of the invention provides a composition comprising one or more polynucleotide sequences selected from sequences identified Figure 13.

A further aspect of the invention relates to nucleic acid arrays comprising a plurality of chondrocyte enriched or chondrocyte-specific nucleic acid member sequences.

In one embodiment, the invention provides an array comprising a plurality of nucleic acid members, wherein at least one member is differentially expressed in cartilage from a patient diagnosed with mild osteoarthritis, as compared to cartilage from a normal individual, and a solid substrate, wherein each nucleic acid member has a unique position on the array and is stably associated with the solid substrate.

In another embodiment, the invention provides an array comprising a plurality of nucleic acid members, wherein at least one member is differentially expressed in cartilage isolated from a patient diagnosed with severe osteoarthritis, as compared to cartilage from a normal individual, and a solid substrate, wherein each nucleic acid member has a unique position on the array and is stably associated with the solid substrate.

In another embodiment, the invention provides an array comprising a plurality of nucleic acid members, wherein at least one member is differentially expressed in cartilage isolated from a patient diagnosed with moderate osteoarthritis, as compared to cartilage from a normal individual, and a solid substrate, wherein each nucleic acid member has a unique position on the array and is stably associated with the solid substrate.

In another embodiment, the invention provides an array comprising a plurality of nucleic acid members, wherein at least one member is differentially expressed in cartilage isolated from a patient diagnosed with marked osteoarthritis, as compared to cartilage from a normal individual, and a solid substrate, wherein each nucleic acid member has a unique position on the array and is stably associated with the solid substrate.

In another embodiment, the invention provides an array comprising a plurality of nucleic acid members, wherein at least one member is differentially expressed in cartilage isolated from a

fetus, as compared to cartilage from a normal individual, and a solid substrate, wherein each nucleic acid member has a unique position on the array and is stably associated with the solid substrate.

In a preferred embodiment, cartilage is isolated from a living normal individual.

In another preferred embodiment, the cartilage is isolated from the normal individual in less than 14 hours post-mortem.

In another embodiment, the invention provides an array comprising a plurality of nucleic acid members and a solid substrate, where at least one member is differentially expressed in cartilage isolated from any two or more of the following sources: (a) a fetus, (b) patient with mild osteoarthritis, (c) patient with moderate osteoarthritis, (d) patient with marked osteoarthritis, (e) patient with severe osteoarthritis or (f) cartilage isolated from a normal individual and where each nucleic acid member has a unique position on the array and is stably associated with the solid substrate.

In one embodiment, each nucleic acid member on an array according to the invention, is at least 50 nucleotides.

In another embodiment, an array according to the invention comprises from 10 to 20,000 positions.

In yet another embodiment, an array according to the invention further includes negative and positive control sequences and RNA quality control sequences. Control sequences can be selected from the group consisting of cDNA sequences of housekeeping genes, plant gene sequences (and/or their cDNA sequences), bacterial sequences, PCR products, vector sequences, and combinations thereof.

Another aspect of the invention relates to novel methods for diagnosing osteoarthritis.

In one embodiment, a method for diagnosing mild osteoarthritis in a patient comprises hybridizing a nucleic acid sample corresponding to RNA (e.g., a sample comprising RNA or cDNA or amplified products of RNA or cDNA) to an array comprising a solid substrate and a plurality of nucleic acid members, wherein at least one member is differentially expressed in cartilage isolated

from a patient diagnosed with mild osteoarthritis, as compared to cartilage isolated from a normal individual and wherein each nucleic acid member has a unique position and is stably associated with the solid substrate. The cartilage isolated from the normal individual is isolated from cartilage tissue less than 14 hours post-mortem. Hybridization of the nucleic acid sample to one or more of the nucleic acid members is indicative of mild osteoarthritis.

In another embodiment, a method of diagnosing moderate osteoarthritis in a patient comprises hybridizing a nucleic acid sample corresponding to RNA to an array comprising a solid substrate and a plurality of nucleic acid members, wherein at least one member is differentially expressed in cartilage isolated from a patient diagnosed with moderate osteoarthritis, as compared to cartilage isolated from a normal individual and wherein each nucleic acid member has a unique position and is stably associated with the solid substrate. Cartilage isolated from the normal individual is isolated from cartilage tissue less than 14 hours post-mortem. Hybridization of the nucleic acid sample to one or more of the nucleic acid members on the array is indicative of moderate osteoarthritis.

In yet another embodiment, a method of diagnosing marked osteoarthritis in a patient comprises hybridizing a nucleic acid sample corresponding to RNA to an array comprising a solid substrate and a plurality of nucleic acid members, wherein at least one member is differentially expressed in cartilage isolated from a patient diagnosed with marked osteoarthritis, as compared to cartilage isolated from a normal individual and each nucleic acid member has a unique position and is stably associated with the solid substrate. Like the above arrays, cartilage isolated from the normal individual is isolated from cartilage tissue less than 14 hours post-mortem. Hybridization of the nucleic acid sample to one or more of the nucleic acid members is indicative of marked osteoarthritis.

In a further embodiment, a method of diagnosing severe osteoarthritis in a patient comprises hybridizing a nucleic acid sample corresponding to RNA to an array comprising a solid substrate and a plurality of nucleic acid members wherein at least one member is differentially expressed in cartilage isolated from a patient diagnosed with severe osteoarthritis, as compared to cartilage isolated from a normal individual and each nucleic acid member has a unique position and is stably associated with the solid substrate. Like the above arrays, cartilage from the normal individual is

isolated from cartilage tissue less than 14 hours post-mortem. Hybridization of the nucleic acid sample to one or more of the nucleic acid members is indicative of severe osteoarthritis.

In a preferred embodiment, the method of diagnosis comprises isolating a cartilage sample from a patient at a specific stage of osteoarthritis (e.g., mild, moderate, marked, or severe).

In another preferred embodiment, the method of diagnosis further comprises the step of preparing an RNA sample from a cartilage sample.

In another preferred embodiment, the method of diagnosis further comprises the step of preparing an RNA sample from blood.

In another preferred embodiment, the method of diagnosis further comprises the step of preparing an RNA sample from synovial fluid.

Another aspect of the invention relates to a method of identifying an agent that increases or decreases the expression of one or more polynucleotide sequences that are differentially expressed in a chondrocyte derived from a fetus or from patient(s) with a chondrocyte disease selected from the group consisting of: mild osteoarthritis, moderate osteoarthritis, marked osteoarthritis and severe osteoarthritis. The method comprises incubating a chondrocyte isolated from a cartilage sample obtained from a normal individual less than 14 hours post-mortem with a candidate agent. RNA is isolated from the chondrocyte and a probe is hybridized to the RNA which corresponds to a polynucleotide sequence which is differentially expressed in a chondrocyte from any two or more of the following developmental or disease stages: a fetus, normal, mild osteoarthritic, moderate osteoarthritic, marked osteoarthritic and severe osteoarthritic. Differential hybridization of the probe to the RNA from normal individual(s) relative to hybridization of the probe to RNA from any one or more of: fetus(es), patient(s) with mild osteoarthritis, patient(s) with moderate osteoarthritis, patient(s) with marked osteoarthritis and patient(s) with severe osteoarthritis identifies the RNA which specifically hybridizes to the probe as a differentially expressed chondrocyte-specific polynucleotide sequence and identifies the candidate agent as one which increases or decreases the expression of the chondrocyte-specific polynucleotide sequence.

The method also can be performed by evaluating cDNA corresponding to RNAs obtained from chondrocytes.

This method also can be performed by evaluating cDNA corresponding to RNAs obtained from blood.

This method also can be performed by evaluating cDNA corresponding to RNAs obtained from synovial fluid.

The invention further relates to methods of preparing chondrocyte cDNA libraries.

In one embodiment, a method of preparing a chondrocyte cDNA library comprises: a) isolating chondrocytes from a cartilage sample from a normal individual, wherein the cartilage sample is obtained less than 14 hours post-mortem, b) isolating total RNA from the chondrocytes, c) synthesizing cDNA from the mRNA in the total RNA, and d) ligating the cDNA into a vector.

In another embodiment, a method of preparing a chondrocyte cDNA library comprises: a) isolating chondrocytes from a cartilage sample from a normal individual, wherein the normal individual is living, b) isolating total RNA from the chondrocytes, c) synthesizing cDNA from mRNA in the total RNA, and d) ligating the cDNA into a vector.

In another embodiment, a method of preparing a chondrocyte cDNA library comprises: a) isolating chondrocytes from a cartilage sample from a fetus, b) isolating total RNA from the chondrocytes, c) synthesizing cDNA from mRNA in the total RNA, and d) ligating the cDNA into a vector.

In another embodiment, a method of preparing a chondrocyte cDNA library is provided comprising, a) isolating chondrocytes from a cartilage sample from a patient diagnosed with mild, moderate, marked or severe osteoarthritis, b) isolating total RNA from the chondrocytes, c) synthesizing cDNA from mRNA in the total RNA, and d) ligating the cDNA into a vector.

The invention also relates to a method of making an array which comprises a plurality of nucleic acid members comprising nucleic acid sequences selected from the group consisting of sequences of Figure 14 on a solid support comprising a surface with a plurality of pre-selected

unique regions. The method comprises: spotting each nucleic acid member individually onto a unique pre-selected region, and stably associating each nucleic acid member with the solid support at the pre-selected region.

In a preferred embodiment, at least one nucleic acid member is differentially expressed in cartilage isolated from a patient diagnosed with mild, moderate, marked, or severe osteoarthritis, as compared to cartilage isolated from a normal individual, wherein cartilage isolated from the normal individual is isolated from cartilage tissue less than 14 hours post-mortem.

In another preferred embodiment, at least one nucleic acid member is differentially expressed in cartilage isolated from a patient diagnosed with mild, moderate, marked, or severe osteoarthritis, as compared to cartilage isolated from a fetus.

In another preferred embodiment, at least one nucleic acid member is differentially expressed in cartilage isolated from a fetus, as compared to a cartilage isolated from a normal individual, wherein cartilage isolated from the normal individual is isolated from cartilage tissue less than 14 hours post-mortem.

In another preferred embodiment, at least one nucleic acid member is differentially expressed in cartilage isolated from any two of the following sources: (a) fetus, (b) a normal individual wherein cartilage isolated from the normal individual is isolated from cartilage tissue less than 14 hours post-mortem, and (c) a patient diagnosed with mild osteoarthritis, (d) a patient diagnosed with moderate osteoarthritis, (e) a patient diagnosed with marked osteoarthritis, or (f) a patient diagnosed with severe osteoarthritis.

The invention also provides kits comprising one or more of the compositions and/or arrays described above and packaging means therefore.

Brief Description of the Drawings

The objects and features of the invention can be better understood with reference to the following detailed description and drawings.

Figure 1, is a graph according to one embodiment of the invention showing the relative EST frequency level of selected extracellular matrix (ECM) proteins among the fetal, normal, mildly osteoarthritic and severe osteoarthritic cartilage cDNA libraries. The percentages were calculated by dividing the number of ESTs matched to a certain type of ECM protein by the total number of ECM ESTs per library. Legend: COL=collagen, PGL=proteoglycan, CMP=cartilage matrix proteins, OSN=osteonectin, FN=fibronectin, CRTL 1=cartilage link protein.

Figure 2, is a graph according to one embodiment of the invention showing the relative EST frequency level of collagens among the fetal, normal, mildly osteoarthritic and severely osteoarthritic cartilage cDNA libraries. The percentages were calculated by dividing the total number of collagen ESTs in a particular library by the total number of collagen ESTs from the four cartilage libraries.

Figure 3, is a graph according to one embodiment of the invention showing the relative EST levels of specific collagen types among the fetal, normal, mildly osteoarthritic and severely osteoarthritic cartilage cDNA libraries. Percentages were calculated by dividing the total number of ESTs for each type of collagen in a particular library by the total number of collagen ESTs from each library.

Figure 4, is a graph according to one embodiment of the invention showing the relative EST frequency level of selected chondrocyte genes among the fetal, normal, mild osteoarthritic and severe osteoarthritic cDNA libraries. Percentages were calculated by dividing the number of ESTs for each gene by the total number of unique genes in each library. Legend: DCN=decorin/chondroitin dermatan sulfate proteoglycan (PG40), HSP90=heatshock protein 90/alpha gene sequence, MSF=megakaryocyte stimulating factor/proteoglycan 4/superficial zone protein, B2M=beta 2 microglobulin, MGP=matrix Gla protein, LUM=lumican, TB4=thymosin beta 4, OSF-2=mRNA for osteoblast specific factor 2, CHI=chitinase, Vim=vimentin.

Figure 5, is a table according to one embodiment of the invention showing the total number of ESTs in each of the four cDNA libraries and the breakdown of what the ESTs represent, including the number of novel sequences (ie. ESTs with no significant match) in each library.

Figure 6, is a table according to one embodiment of the invention listing the unique known genes (5,807) identified in the four cDNA libraries to date.

Figure 6A, is a table according to one embodiment of the invention listing the names of the EST sequences identified in the four cDNA libraries that represent each of the unique known genes identified in Figure 6.

Figure 6B, is a table according to one embodiment of the invention listing the names of all of the EST sequences identified from the cDNA library constructed from fetal cartilage tissue.

Figure 6C, is a table according to one embodiment of the invention listing the names of all of the EST sequences identified from the cDNA library constructed from normal cartilage tissue where such tissue is obtained less than 14 hours post-mortem.

Figure 6D, is a table according to one embodiment of the invention listing the names of all of the EST sequences identified from the cDNA library constructed from cartilage of patients with mild osteoarthritis.

Figure 6E, is a table according to one embodiment of the invention listing the names of all of the EST sequences identified from the cDNA library constructed from cartilage of patients with severe osteoarthritis.

Figure 7, is a table according to one embodiment of the invention showing the characterization of the total number of ESTs from the four cDNA libraries (57,422) based on the functional classification of unique known genes represented by the ESTs.

Figure 8, is a list of known and novel EST clones from the mild and severe cDNA libraries comprising a microarray according to one embodiment of the invention.

Figure 9, is a table showing candidate upregulated genes detected in the mild osteoarthritis cDNA library based on the microarray analysis according to one embodiment of the invention.

Figure 10, is a table showing candidate down-regulated genes detected in the mild osteoarthritis cDNA library based on the microarray analysis according to one embodiment of the invention.

Figure 11, is a table showing candidate up-regulated genes detected in the severe osteoarthritis cDNA library based on the microarray analysis according to one embodiment of the invention.

Figure 12, is a table showing candidate down-regulated genes detected in the severe osteoarthritic cDNA library based on the microarray analysis according to one embodiment of the invention.

Figure 13, is a table listing the EST sequence names representing novel sequences identified in each of the four cDNA libraries to date according to one embodiment of the invention.

Figure 14, is a CD ROM, attached hereto, containing all of the EST sequences identified from the four human cartilage cDNA libraries according to one embodiment of the invention. The names of all of the EST sequences on the CD-ROM are listed in Figures 6B, 6C, 6D and 6E.

Figure 15, contains a list of genes that have been identified through EST frequency analysis as being differentially expressed between fetal and normal cDNA libraries according to one embodiment of the invention.

Figure 16, contains a list of genes that have been identified through EST frequency analysis as being differentially expressed between mild and severe osteoarthritis cDNA libraries according to one embodiment of the invention.

Figure 17, is a bar graph showing the level of beta-2 microglobulin (B2M) in synovial fluid from normal individuals and patients with different stages of osteoarthritis according to one embodiment of the invention. Legend: nor=normal individual, mioa=patient with mild osteoarthritis, mooa=patient with moderate osteoarthritis, maoa=patient with marked osteoarthritis, seoa=patient with severe osteoarthritis.

Figure 18, is a bar graph showing the level of beta 2 microglobulin (B2M) in medium cultured from cartilage from patients with severe osteoarthritis at varying time periods during culturing according to one embodiment of the invention.

Figure 19, is a black and white representation of a two-color fluorescent scan, according to one embodiment of the invention, showing genes preferentially expressed in non-B2M-treated chondrocytes (which would appear as green spots) and genes preferentially expressed in B2M-treated chondrocytes (which would appear as reddish spots). Genes expressed at approximately equal levels would appear as yellow spots. B2M=beta 2 microglobulin.

Detailed Description of the Invention

The invention relates to methods of profiling gene sequences expressed in human chondrocytes to identify differential gene expression in chondrocytes at different stages of development and disease. Differentially expressed genes and their products (e.g., mRNAs and proteins) can be used in methods for diagnosis, prognosis, screening, or treatment of osteoarthritis.

Definitions

The following definitions are provided for specific terms which are used in the following written description.

As used herein, "osteoarthritis" refers to a chronic disease in which the articular cartilage that lies on the ends of bones that form the articulating surface of the joints gradually degenerates over time. Cartilage degeneration can be caused by an imbalanced catabolic activity (removal of "old" cells and matrix components) and anabolic activity (production of "new" cells and molecules) (Westacott et al., 1996, *Semin Arthritis Rheum*, 25:254-72).

As used herein, "cartilage" or "articular cartilage" refers to elastic, translucent connective tissue in mammals, including human and other species. Cartilage is composed predominantly of chondrocytes, type II collagen, small amounts of other collagen types, other noncollagenous proteins, proteoglycans and water, and is usually surrounded by a perichondrium, made up of fibroblasts, in a matrix of type I and type II collagen as well as other proteoglycans. Although most cartilage becomes bone upon maturation, some cartilage remains in its original form in locations such as the nose, ears, knees, and other joints. The cartilage has no blood or nerve supply and chondrocytes are the only type of cell in this tissue.

As used herein, "chondrocyte" refers to cartilage cells.

As used herein, "synovial fluid" refers to fluid secreted from the "synovial sac" which surrounds each joint. Synovial fluid serves to protect the joint, lubricate the joint and provide nourishment to the articular cartilage. Synovial fluid useful according to the invention contains cells from which RNA can be isolated according to methods well known in the art as described herein.

As used herein, the term "osteoarthritis (OA) staging" or "osteoarthritis (OA) grading" refers to determining the degree of advancement or progression of the disease in the cartilage. In order to classify cartilage into different disease stages, a scoring system is used according to known methods in the art. Preferably the scoring system described in Marshall (Marshall W., 1996, *The Journal of Rheumatology*, 23:582-584, incorporated by reference) is used. According to this

method, each of the 6 articular surfaces (patella, femoral trochlea, medial femoral condyle, medial tibial plateau, lateral femoral condyle and lateral tibial plateau) is assigned a cartilage grade based on the worst lesion present on that specific surface. A scoring system is then applied in which each articular surface receives an OA severity number value that reflects the cartilage severity grade for that surface. For example, if the medial femoral condyle has a grade I lesion as its most severe cartilage damage a value of 1 is assigned. A total score for the patient is then derived from the sum of the scores on the 6 articular surfaces. Based on the total score, each patient is placed into one of 4 OA groups: mild (early) (1-6), moderate (7-12), marked (13-18) and severe (>18).

As used herein, "diagnosis" refers to a process of determining if an individual is afflicted with a disease or ailment. "Diagnosis of OA" or "OA diagnosis", according to the invention, means determining if an individual is afflicted with OA, or, once a patient is diagnosed, determining the OA stage or grade as used herein based on the medical history and physical examination of the patient using methods known in the art (i.e., joint X ray). Preferably, OA stages are measured using the scoring system described by Marshall, *supra*. "Prognosis of OA" refers to a prediction of the probable occurrence and/or progression of OA in a patient, as well as the likelihood of recovery from OA, or the likelihood of ameliorating symptoms of OA or the likelihood of reversing the effects of OA.

As used herein, "patient" refers to a mammal who is diagnosed with a mild, moderate, marked, or severe form of OA.

As used herein, "normal" refers to an individual who has not shown any OA symptoms or has not been diagnosed with cartilage injury or OA. "Normal", according to the invention, also refers to a sample taken from a normal individual within 14 hours post-mortem. A normal cartilage tissue sample, for example, refers to the whole or a piece of cartilage isolated from cartilage tissue within 14 hours post-mortem from an individual who was not diagnosed with OA and whose corpse does not show any symptoms of OA at the time of tissue removal. In alternative embodiments of the invention, the "normal" cartilage tissue sample is isolated from cartilage tissue less than 14 hours post-mortem, e.g., within 13 hours, 12 hours, 11 hours, 10 hours, 9 hours, 8 hours, 7 hours, 6 hours, 5 hours, 4 hours, 3 hours, 2 hours, or 1 hour post-mortem. In one embodiment of the

invention, the "normal" cartilage sample is isolated at 14 hours post-mortem and the integrity of mRNA samples extracted is confirmed.

As used herein, "mRNA integrity" refers to the quality of mRNA extracts from cartilage samples. mRNA extracts with good integrity do not appear to be degraded when examined by methods well known in the art, for example, by RNA agarose gel electrophoresis (e.g., Ausubel et al., John Weley & Sons, Inc., 1997, *Current Protocols in Molecular Biology*). Preferably, the mRNA samples have good integrity (e.g., less than 10%, preferably, less than 5%, and more preferably, less than 1% of the mRNA is degraded) to truly represent the gene expression levels of the cartilage samples from which they are extracted.

As used herein, "fetal" cartilage samples refer to samples taken from a fetus. The chondrocytes of fetal cartilage have a higher level of metabolic activity and cell division rates as compared to chondrocytes from cartilage derived from either a normal adult or from an adult diagnosed with any stage of OA (mild, moderate, marked and severe).

As used herein, "polynucleotide(s)", which includes "nucleic acid(s)" "nucleic acid sequences", "sequences" and "Express Sequence Tags"(EST(s)), generally refers to any polyribonucleotide or poly-deoxyribonucleotide, which may be unmodified RNA or DNA or modified RNA or DNA. "Polynucleotides" include, without limitation, single-and double-stranded nucleic acids. As used herein, the term "polynucleotide(s)" also includes DNAs or RNAs as described above, that contain one or more modified bases. Thus, DNAs or RNAs with backbones modified for stability or for other reasons are "polynucleotides". The term "polynucleotides" as it is used herein embraces such chemically, enzymatically or metabolically modified forms of polynucleotides, as well as the chemical forms of DNA and RNA characteristic of viruses and cells, including for example, simple and complex cells.

As used herein, "isolated" or "purified" when used in reference to a nucleic acid means that a naturally occurring sequence has been removed from its normal cellular (e.g., chromosomal) environment or is synthesized in a non-natural environment (e.g., artificially synthesized). Thus, an "isolated" or "purified" sequence may be in a cell-free solution or placed in a different cellular environment. The term "purified" does not imply that the sequence is the only nucleotide present,

but that it is essentially free (about 90-95% pure) of non-nucleotide material naturally associated with it, and thus is distinguished from isolated chromosomes.

As used herein, the term "probe" refers to an oligonucleotide which forms a duplex structure with a sequence in the target nucleic acid, due to complementarity of at least one sequence in the probe with a sequence in the target region.

As defined herein, a "nucleic acid array" refers a plurality of unique nucleic acids (or "nucleic acid members") attached to one surface of a solid support at a density exceeding 20 different nucleic acids/cm^2 wherein each of the nucleic acid members is attached to the surface of the solid support in a non-identical pre-selected region. In one embodiment, the nucleic acid member attached to the surface of the solid support is DNA. In a preferred embodiment, the nucleic acid member attached to the surface of the solid support is cDNA. In another preferred embodiment, the nucleic acid member attached to the surface of the solid support is cDNA synthesized by polymerase chain reaction (PCR). Preferably, a nucleic acid member of the array according to the invention is at least 50 nucleotides in length. Preferably, a nucleic acid member of the array is less than 6,000 nucleotides in length. More preferably, a nucleic acid member of the array comprises an array less than 500 nucleotides in length. In one embodiment, the array comprises at least 500 different nucleic acid members attached to one surface of the solid support. In another embodiment, the array comprises at least 10 different nucleic acid members attached to one surface of the solid support. In yet another embodiment, the array comprises at least 10,000 different nucleic acid members attached to one surface of the solid support. In yet another embodiment, the array comprises at least 15,000 different nucleic acid members attached to one surface of the solid support. The term "nucleic acid", as used herein, is interchangeable with the term "polynucleotide".

As used herein, "a plurality of" or "a set of" refers to more than two, for example, 3 or more, 100 or more, or 1000 or more, or 10,000 or more.

As used herein, "attaching" or "spotting" refers to a process of depositing a nucleic acid onto a solid substrate to form a nucleic acid array such that the nucleic acid is irreversibly bound to the solid substrate via covalent bonds, hydrogen bonds or ionic interactions.

As used herein, "stably associated" refers to a nucleic acid that is irreversibly bound to a solid substrate to form an array via covalent bonds, hydrogen bonds or ionic interactions such that the nucleic acid retains its unique pre-selected position relative to all other nucleic acids that are stably associated with an array, or to all other pre-selected regions on the solid substrate under conditions in which an array is typically analyzed (i.e., during one or more steps of hybridization, washes, and/or scanning, etc.).

As used herein, "solid substrate" or "solid support" refers to a material having a rigid or semi-rigid surface. The terms "substrate" and "support" are used interchangeably herein with the terms "solid substrate" and "solid support". The solid support may be biological, non-biological, organic, inorganic, or a combination of any of these, existing as particles, strands, precipitates, gels, sheets, tubing, spheres, beads, containers, capillaries, pads, slices, films, plates, slides, chips, etc. Often, the substrate is a silicon or glass surface, (poly)tetrafluoroethylene, (poly)vinylidendifluoride, polystyrene, polycarbonate, a charged membrane, such as nylon 66 or nitrocellulose, or combinations thereof. In a preferred embodiment, the solid support is glass. Preferably, at least one surface of the substrate will be substantially flat. Preferably, the surface of the solid support will contain reactive groups, including, but not limited to, carboxyl, amino, hydroxyl, thiol, and the like. In one embodiment, the surface is optically transparent.

As used herein, "pre-selected region", "predefined region", or "unique position" refers to a localized area on a substrate which is, was, or is intended to be used for the deposit of a nucleic acid and is otherwise referred to herein in the alternative as a "selected region" or simply a "region." The pre-selected region may have any convenient shape, e.g., circular, rectangular, elliptical, wedge-shaped, etc. In some embodiments, a pre-selected region is smaller than about 1 cm^2, more preferably less than 1 mm^2, still more preferably less than 0.5 mm^2, and in some embodiments less than 0.1 mm^2. A nucleic acid member at a "pre-selected region", "predefined region", or "unique position" is one whose identity (e.g., sequence) can be determined by virtue of its position at the region or unique position.

As used herein, a " nucleic acid target" or "a target nucleic acid" is defined as a nucleic acid capable of binding to a nucleic acid member of complementary sequence through one or more types of chemical bonds, usually through complementary base pairing, i.e., through hydrogen bond

formation. As used herein, a nucleic acid target may include natural (i. e., A, G, C, or T) or modified bases (7-deazaguanosine, inosine, etc.). In addition, the bases in nucleic acid probe may be joined by a linkage other than a phosphodiester bond, so long as it does not interfere with hybridization (i.e., the probe still specifically binds to its complementary sequence under standard stringent or selective hybridization conditions). Thus, nucleic acid targets may be peptide nucleic acids in which the constituent bases are joined by peptide bonds rather than phosphodiester linkages. Preferably, the nucleic acid targets are derived from human cartilage, blood or synovial fluid extracts. More preferably, the nucleic acid targets are single- or double-stranded DNA, RNA, or DNA-RNA hybrids, from human cartilage, blood or synovial fluid RNA extracts, and preferably from mRNA extracts.

As used herein, a "cartilage nucleic acid sample", refers to nucleic acids derived from cartilage. Preferably, a cartilage nucleic acid sample is RNA or is a nucleic acid corresponding to RNA, for example, cDNA.

As used herein, the term "hybridizing to" or "hybridization" refers to the hydrogen binding with a complementary nucleic acid, via an interaction between for example, a target nucleic acid sequence and a nucleic acid member in an array.

As used herein, "specific hybridization" or "selective hybridization" refers to hybridization which occurs when two nucleic acid sequences are substantially complementary (at least about 65% complementary over a stretch of at least 14 to 25 nucleotides, preferably at least about 75%, more preferably at least about 90% complementary). See Kanehisa, M., 1984, *Nucleic acids Res.*, 12:203, incorporated herein by reference. As a result, it is expected that a certain degree of mismatch is tolerated. Such mismatch may be small, such as a mono-, di- or tri-nucleotide. Alternatively, a region of mismatch can encompass loops, which are defined as regions in which there exists a mismatch in an uninterrupted series of four or more nucleotides. Numerous factors influence the efficiency and selectivity of hybridization of two nucleic acids, for example, a nucleic acid member on a array, to a target nucleic acid sequence. These factors include nucleic acid member length, nucleotide sequence and/or composition, hybridization temperature, buffer composition and potential for steric hindrance in the region to which the nucleic acid member is required to hybridize. A positive correlation exists between the nucleic acid member length and

both the efficiency and accuracy with which a nucleic acid member will anneal to a target sequence. In particular, longer sequences have a higher melting temperature (T_M) than do shorter ones, and are less likely to be repeated within a given target sequence, thereby minimizing promiscuous hybridization. Hybridization temperature varies inversely with nucleic acid member annealing efficiency, as does the concentration of organic solvents, e.g., formamide, that might be included in a hybridization mixture, while increases in salt concentration facilitate binding. Under stringent annealing conditions, longer nucleic acids, hybridize more efficiently than do shorter ones, which are sufficient under more permissive conditions.

As used herein, the term "differential hybridization" refers to a probe that can hybridize to a same polynucleotide sequence obtained from two or more samples at different levels. A "differential hybridization" means that the ratio of the level of hybridization of the probe to the polynucleotide sequence isolated from one sample as compared to the polynucleotide sequence isolated from another sample is not equal to 1.0. For example, the ratio of the level of hybridization of the probe to the polynucleotide sequence isolated from one sample as compared to the polynucleotide sequence isolated from another sample is greater than or less than 1.0 and includes greater than 1.5 and less than 0.7 greater than 2 and less than 0.5. A differential hybridization also exists if the hybridization is detectable in one sample but not another sample.

As herein used, the term "standard stringent conditions" means hybridization will occur only if there is at least 95% and preferably, at least 97% identity between the sequences, wherein the region of identity comprises at least 10 nucleotides. In one embodiment, the sequences hybridize under stringent conditions following incubation of the sequences overnight at 42°C, followed by stringent washes (0.2X SSC at 65° C). As several factors affect the stringency of hybridization, the combination of parameters is more important than the absolute measure of a single factor.

As used herein, the term "level of expression" refers to the measurable expression level of a given nucleic acid. The level of expression of a nucleic acid is determined by methods well known in the art. The term "differentially expressed" or "changes in the level of expression" refers to an increase or decrease in the measurable expression level of a given nucleic acid. As used herein, "differentially expressed" when referring to microarray analysis means the ratio of the level of

expression of a given polynucleotide in one sample and the expression level of the given polynucleotide in another sample is not equal to 1.0. "Differentially expressed" when referring to microarray analysis according to the invention also means the ratio of the expression level of a given polynucleotide in one sample and the expression level of the given polynucleotide in another sample where the ratio is greater than or less than 1.0 and includes greater than 1.5 and less than 0.7, as well as greater than 2.0 and less than 0.5. A nucleic acid also is said to be differentially expressed in two samples if one of the two samples contains no detectable expression of the nucleic acid. Absolute quantification of the level of expression of a nucleic acid can be accomplished by including known concentration(s) of one or more control nucleic acid species, generating a standard curve based on the amount of the control nucleic acid and extrapolating the expression level of the "unknown" nucleic acid species from the hybridization intensities of the unknown with respect to the standard curve. The level of expression is measured by hybridization analysis using labeled target nucleic acids according to methods well known in the art. The label on the target nucleic acid can be a luminescent label, an enzymatic label, a radioactive label, a chemical label or a physical label. Preferably, target nucleic acids are labeled with a fluorescent molecule. Preferred fluorescent labels include, but are not limited to: fluorescein, amino coumarin acetic acid, tetramethylrhodamine isothiocyanate (TRITC), Texas Red, Cy3 and Cy5.

As used herein "differentially expressed" when referring to EST analysis refers to the relative expression level of a gene based on the frequency of ESTs representing the gene derived from a cDNA library as compared to the frequency of ESTs representing the same gene derived from another cDNA library. As described herein, the "relative EST frequency" of an EST is calculated by dividing the number of ESTsrepresenting each specific gene by the total number of ESTs analyzed. Differences in "relative EST frequency" may be used as an indication of differential gene expression.

As used herein, the term "significant match", when referring to nucleic acid sequences, means that two nucleic acid sequences exhibit at least 65% identity, at least 70%, at least 75%, at least 80%, at least 85%, and preferably, at least 90% identity, using comparison methods well known in the art (i.e., Altschul, S.F. et al., 1997, *Nucl. Acids Res.*, 25:3389-3402; Schäffer, A.A. et al., 1999, *Bioinformatics* 15:1000-1011). As used herein, "significant match" encompasses non-contiguous or scattered identical nucleotides so long as the sequences exhibit at least 65%, and

preferably, at least 70%, at least 75%, at least 80%, at least 85%, and preferably, at least 90% identity, when maximally aligned using alignment methods routine in the art.

As used herein, a "novel sequence" or "novel expressed sequence tag (EST)" refers to a nucleic acid sequence which has no significant match to any existing sequence in the "nt", "nr", "est", "gss" and "htg" databases available through NCBI at the time each novel sequence was compared. "No significant match" preferably refers to a less than 65% match between a novel sequence being queried against other sequences in the database, and preferably, a less than 50% match, a less than 40% match, or a less than 30% match, after maximally aligning sequences using methods routine in the art.

As used herein, a "known sequence" refers to a nucleic acid sequence which has significant match to at least one existing sequence in the "nt", "nr", "est", "gss" and "htg" databases available through NCBI. "Known sequence with a function" refers to a nucleic acid with significant match to an existing sequence which encodes a polypeptide with a known function. "Known sequence with no function" refers to a nucleic acid that exhibits a significant match to an existing sequence which encodes a polypeptide of unknown function.

As used herein, a "chondrocyte-specific nucleic acid" is a nucleic acid sequence which is expressed at a detectable level in a chondrocyte and is not expressed at a detectable level in any other cell types as indicated by having no significant match to any sequence in any of the available databases comprising sequences from other cell types.

As used herein, a "chondrocyte enriched nucleic acid" or "chondrocyte enriched sequence" refers to a sequence which is differentially expressed in chondrocytes as compared to non-chondrocytes.

As used herein, "indicative of disease" refers to an expression pattern which is diagnostic of disease such that the expression pattern is found significantly more often in patients with a disease than in patients without the disease (as determined using routine statistical methods setting confidence levels at 95%). Preferably, an expression pattern which is indicative of disease is found in at least 70% of patients who have the disease and is found in less than 10% of patients who do not have the disease. More preferably, an expression pattern which is indicative of disease is found

in at least 75%, at least 80%, at least 85%, at least 90%, at least 95% or more in patients who have the disease and is found in less than 10%, less than 8%, less than 5%, less than 2.5%, or less than 1% of patients who do not have the disease.

As used herein, a "gene expression pattern" or "gene expression profile" comprises the pattern of expression of one or more of a set of nucleic acid sequences where one or more members of the set are differentially expressed.

As used herein, "a nucleic acid array expression profile" is generated from the hybridization of nucleic acids derived from a sample to one or more nucleic acid members comprising an array according to the invention.

As used herein, a "therapeutic agent" or "agent" refers to a compound that increases or decreases the expression of a polynucleotide sequence that is differentially expressed in a chondrocyte from any two of the following developmental or osteoarthritis disease stages: (a) fetal, (b) mild, (c) moderate, (d) marked and (e) severe, or (f) chondrocyte from a normal individual, as defined herein. A therapeutic agent according to the invention also refers to a compound that increases or decreases the anabolic activity of a chondrocyte. The invention provides for a "therapeutic agent" that 1) prevents the onset of osteoarthritis; 2) reduces, delays, or eliminates osteoarthritis symptoms such as pain, swelling, weakness and loss of functional ability in the afflicted joints; 3) reduces, delays, or eliminates cartilage degeneration, and/or enhances chondrocyte metabolic activity and cell division rates; and/or 4) restores one or more expression profiles of one or more disease-indicative nucleic acids of a patient to a profile more similar to that of a normal individual when administered to a patient.

Identifying Chondrocyte Enriched and Chondrocyte-Specific Polynucleotide Sequences

cDNA libraries were constructed from human fetal, normal, mild osteoarthritic and severe osteoarthritic cartilage samples. The known and novel clones derived from these libraries were then used to construct human chondrocyte-specific microarrays to generate differential gene expression profiles useful as a diagnostic tool for detection of mild (early stage) osteoarthritis. Arrays of the invention are useful as a gold standard for osteoarthritis diagnosis and for use to identify and monitor therapeutic efficacy of new drug targets.

One effective and rapid way of characterizing gene expression patterns in a given tissue is through large-scale partial sequencing of a cDNA library produced from such a tissue to generate expressed sequence tags (ESTs). This approach has provided both quantitative and qualitative information on gene expression in a variety of tissues and cells (4-7). Since cDNA libraries represent gene transcription in the cells of the tissue used to construct the library, gene expression profiles generated by random sampling and sequencing is used for detailed genetic-level comparison between developmental, normal and pathological states of the tissue examined.

Many human genes are expressed at different levels in cartilage of different developmental (fetal vs. mature) or disease states. In some cases, a gene is not expressed at all in some developmental or disease states, and at high levels in others (see Figure 6, 15 and 16 for examples). According to the invention, differential analysis of chondrocyte gene expression during different stages of cartilage developmental and in different disease states using an EST-based approach has identified genes that play important roles in osteoarthritis pathogenesis and cartilage repair. The advantage of this method is that it provides gene expression information on a larger scale than other methods. The cDNA clones generated by this approach are also useful for functional studies of certain genes. This type of genomic-based approach has provided important novel insights into our understanding of the osteoarthritis disease process and provides for novel diagnostic, prognostic and therapeutic approaches.

Samples

Cartilage

In one aspect, cartilage is obtained from a fetus using methods known in the art. The chondrocytes of fetal cartilage have a higher level of metabolic activity and cell division rates as compared to chondrocytes from cartilage from either a normal adult or from an individual diagnosed with any stage of osteoarthritis (mild, moderate, marked and severe).

In another aspect, cartilage is obtained from a normal individual who is alive or is obtained from cartilage tissue less than 14 hours post mortem, according to methods known in the art and described below. Normal articular cartilage from human adults are obtained using any known method. However, truly normal cartilage cannot generally be sampled from live donors due to

ethical considerations. Preferably, normal cartilage samples are obtained from deceased donors, within a fourteen-hour post-mortem window after cessation of perfusion to the sampled joint, to minimize the degradation of RNA observed beyond the window. In other embodiments, the "normal" tissue is obtained less than 14 hours post-mortem, such as 13, 12, 11, 10, 9, 8, 6, 4, 2,or 1 hour post-mortem. A baboon study was conducted to confirm this approach and is described herein below in Example 11. Preferably the normal cartilage is obtained less than 14 hours post-mortem. More prefably, the normal cartilage is obtained less than 12 hours post-mortem.

Preferably, cartilage also is isolated from the following disease stages of osteoarthritis: mild, marked, moderate and severe. Human cartilage samples from osteoarthritic individuals are obtained using any known method. Preferably the cartilage is obtained from individuals undergoing arthroscopy or total knee replacements and samples are stored in liquid nitrogen until needed. In a preferred embodiment, a minimum of 0.05 g of cartilage sample is isolated to obtain 2 μg total RNA extract for the construction of a cDNA library. In another preferred embodiment, a minimum of 0.025 g cartilage sample is isolated to obtain 1 μg total RNA extract to use as a target sample for a microarray. A cartilage sample that is useful according to the invention is in an amount that is sufficient for the detection of one or more polynucleotide sequences according to the invention.

Blood and Synovial Fluid

Samples useful according to the invention also include blood and synovial fluid samples.

In one aspect, blood is obtained from a normal patient or from an individual diagnosed with, or suspected of having, osteoarthritis according to methods of phlebotomy well known in the art. A blood sample useful according to the invention is in an amount ranging from 1 μl to 100ml, preferably 10 μl to 50 ml, more preferably 10 μl to 25ml and most preferably 10 μl to 1 ml. A blood sample that is useful according to the invention is in an amount that is sufficient for the detection of one or more polynucleotide sequences according to the invention. In one embodiment, polynucleotides contained within the blood sample are amplified, for example, by polymerase chain reaction (PCR) or by RT-PCR. Other amplification methods known in the art are also encompassed within the scope of the invention (e.g., ligase chain reaction, NASBA, 3SR, and the like).

A synovial fluid sample is obtained from an individual diagnosed with, or suspected of having osteoarthritis according to methods well known in the art. Preferably, synovial fluid is collected from a human knee joint by aspiration at arthroscopy. A synovial fluid sample useful according to the invention is in an amount ranging from 0.1 ml to 20 ml and preferably 0.5 ml to 10 ml. A synovial fluid sample that is useful according to the invention is in an amount that is sufficient for the detection of one or more polynucleotide sequences according to the invention.

Developmental and Disease Stages of Articular Cartilage

Chondrocytes are preferably obtained from any of the following developmental and disease stages: fetal, normal, mild osteoarthritic, moderate osteoarthritic, marked osteoarthritic or severe osteoarthritic.

Cartilage isolated from a human fetus (e.g., during fetal development) is characterized above, and is useful according to the invention for analysis of fetal chondrocytes.

Cartilage isolated from a "normal" individual, defined herein, also is useful according to the invention for isolation and analysis of "normal" chondrocytes.

Cartilage isolated from a patient diagnosed with any one of: mild, moderate, marked and severe osteoarthritis also is useful in the present invention.

In order to classify cartilage according to disease state, a scoring system is used, whereby subjective decisions by the arthroscopist are minimized. The scoring system which defines disease states described herein is that of Marshall, *supra*, incorporated herein by reference. According to this method, each of the 6 articular surfaces (patella, femoral trochlea, medial femoral condyle, medial tibial plateau, lateral femoral condyle and lateral tibial plateau) is assigned a cartilage grade based on the worst lesion present on that specific surface. A scoring system is then applied in which each articular surface receives an osteoarthritis severity number value that reflects the cartilage severity grade for that surface, as described in Table 1.

Table 1. Articular Cartilage Grading System

Grade	Articular Cartilage	Points

0	Normal	0
I	Surface intact-softening, edema	1
II	Surface-disrupted-partial thickness lesions (no extension to bone)	2
III	Full thickness lesions-extensions to intact bone	3
IV	Bone erosion or eburnation	4

For example, if the medial femoral condyle has a grade I lesion as its most severe cartilage damage, a value of 1 is assigned. A total score for the patient is then derived from the sum of the scores of the 6 articular surfaces. Based on the total score, each patient is placed into one of 4 osteoarthritis groups: mild (1-6), moderate (7-12), marked (13-18) and severe (>18).

RNA Preparation

In one aspect, RNA is isolated from cartilage samples from various disease or developmental stages as described herein. Samples can be from single patients or can be pooled from multiple patients.

In another aspect, RNA is isolated directly from synovial fluid of persons with various disease or developmental stages of osteoarthritis as described herein. Samples can be from single patients or can be pooled from multiple patients.

In another aspect, RNA is isolated directly from blood samples of persons with various disease or developmental stages of osteoarthritis as described herein. Samples can be from single patients or can be pooled from multiple patients.

Total RNA is extracted from the cartilage samples according to methods well known in the art. In one embodiment, RNA is purified from cartilage tissue according to the following method. Following removal of a tissue of interest from an individual or patient, the tissue is quick frozen in liquid nitrogen, to prevent degradation of RNA. Upon the addition of a volume of tissue guanidinium solution, tissue samples are ground in a tissuemizer with two or three 10-second bursts. To prepare tissue guanidinium solution (1 L) 590.8 g guanidinium isothiocyanate is dissolved in approximately 400 ml DEPC-treated H_2O. 25 ml of 2 M Tris-Cl, pH 7.5 (0.05 M

final) and 20 ml Na_2EDTA (0.01 M final) is added, the solution is stirred overnight, the volume is adjusted to 950 ml, and 50 ml 2-ME is added.

Homogenized tissue samples are subjected to centrifugation for 10 min at 12,000 x g at 12°C. The resulting supernatant is incubated for 2 min at 65°C in the presence of 0.1 volume of 20% Sarkosyl, layered over 9 ml of a 5.7M CsCl solution (0.1g CsCl/ml), and separated by centrifugation overnight at 113,000 x g at 22°C. After careful removal of the supernatant, the tube is inverted and drained. The bottom of the tube (containing the RNA pellet) is placed in a 50 ml plastic tube and incubated overnight (or longer) at 4°C in the presence of 3 ml tissue resuspension buffer (5 mM EDTA, 0.5% (v/v) Sarkosyl, 5% (v/v) 2-ME) to allow complete resuspension of the RNA pellet. The resulting RNA solution is extracted sequentially with 25:24:1 phenol/chloroform/isoamyl alcohol, followed by 24:1 chloroform/isoamyl alcohol, precipitated by the addition of 3 M sodium acetate, pH 5.2, and 2.5 volumes of 100% ethanol, and resuspended in DEPC water (Chirgwin et al., 1979, *Biochemistry*, 18:5294).

Alternatively, RNA is isolated from cartilage tissue according to the following single step protocol. The tissue of interest is prepared by homogenization in a glass teflon homogenizer in 1 ml denaturing solution (4M guanidinium thiosulfate, 25 mM sodium citrate, pH 7.0, 0.1M 2-ME, 0.5% (w/v) N-laurylsarkosine) per 100mg tissue. Following transfer of the homogenate to a 5-ml polypropylene tube, 0.1 ml of 2 M sodium acetate, pH 4, 1 ml water-saturated phenol, and 0.2 ml of 49:1 chloroform/isoamyl alcohol are added sequentially. The sample is mixed after the addition of each component, and incubated for 15 min at 0-4°C after all components have been added. The sample is separated by centrifugation for 20 min at 10,000 x g, 4°C, precipitated by the addition of 1 ml of 100% isopropanol, incubated for 30 minutes at -20°C and pelleted by centrifugation for 10 minutes at 10,000 x g, 4°C. The resulting RNA pellet is dissolved in 0.3 ml denaturing solution, transferred to a microfuge tube, precipitated by the addition of 0.3 ml of 100% isopropanol for 30 minutes at -20°C, and centrifuged for 10 minutes at 10,000 x g at 4°C. The RNA pellet is washed in 70% ethanol, dried, and resuspended in 100-200µl DEPC-treated water or DEPC-treated 0.5% SDS (Chomczynski and Sacchi, 1987, *Anal. Biochem.*, 162:156).

Preferably, the cartilage samples are finely powdered under liquid nitrogen and total RNA is extracted using TRIzol® reagent (GIBCO/BRL).

Alternatively, RNA is isolated from blood by the following protocol. Lysis Buffer is added to blood sample in a ratio of 3 parts Lysis Buffer to 1 part blood (Lysis Buffer (1L) 0.6g EDTA; 1.0g $KHCO_2$, 8.2g NH_4Cl adjusted to pH 7.4 (using NaOH)). Sample is mixed and placed on ice for 5-10 minutes until transparent. Lysed sample is centrifuged at 1000 rpm for 10 minutes at 4°C, and supernatant is aspirated. Pellet is resuspended in 5ml Lysis Buffer, and centrifuged again at 1000 rpm for 10 minutes at 4°C. Pelleted cells are homogenized using TRIzol® (GIBCO/BRL) in a ratio of approximately 6ml of TRIzol® for every 10ml of the original blood sample and vortexed well. Samples are left for 5 minutes at room temperature. RNA is extracted using 1.2 ml of chloroform per 1 ml of TRIzol®. Sample is centrifuged at 12,000 x g for 5 minutes at 4°C and upper layer is collected. To upper layer, isopropanol is added in ratio of 0.5 ml per 1 ml of TRIzol®. Sample is left overnight at -20°C or for one hour at -20°C. RNA is pelleted in accordance with known methods, RNA pellet air dried, and pellet resuspended in DEPC treated ddH_2O. RNA samples can also be stored in 75% ethanol wherein said samples are stable at room temperature for transportation.

Alternatively, RNA is isolated from synovial fluid using TRIzol® reagent (GIBCO/BRL).

Purity and integrity of RNA is assessed by absorbance at 260/280nm and agarose gel electrophoresis followed by inspection under ultraviolet light.

Construction of cDNA libraries

cDNA libraries are constructed according to methods well known in the art (see for example Ausubel, *supra*, and Sambrook, *supra*, incorporated herein by reference).

In one aspect, cDNA samples, i.e., DNA that is complementary to RNA such as mRNA are prepared. The preparation of cDNA is well-known and well-documented in the prior art.

cDNA may be prepared according to the following method. Total cellular RNA is isolated (as described) and passed through a column of oligo(dT)-cellulose to isolate polyA RNA. The bound polyA mRNAs are eluted from the column with a low ionic strength buffer. To produce cDNA molecules, short deoxythymidine oligonucleotides (12-20 nucleotides) are hybridized to the polyA tails to be used as primers for reverse transcriptase, an enzyme that uses RNA as a template

for DNA synthesis. Alternatively, or additionally, mRNA species are primed from many positions by using short oligonucleotide fragments comprising numerous sequences complementary to the mRNA of interest as primers for cDNA synthesis. The resultant RNA-DNA hybrid is converted to a double stranded DNA molecule by a variety of enzymatic steps well-known in the art (Watson et al., 1992, *Recombinant DNA*, 2nd edition, Scientific American Books, New York).

To construct a cDNA library, the poly $(A)^+$ RNA fraction may be isolated by oligo-dT cellulose chromatography (Pharmacia), and 3-5 ug poly $(A)^+$ RNA is used to construct a cDNA library in the λ ZAP Express vector (Stratagene). Alternatively, cDNA libraries may be constructed into λTriplEx2 vector through a PCR-based method, using SMART (Switching Mechanism At 5' end of RNA Transcript) cDNA Library Construction Kit (Clontech). First-strand cDNA is synthesized with an Xho I-oligo (dT) adapter-primer in the presence of 5'-methyl dCTP. After second-strand synthesis and ligation of EcoRI adapters, the cDNAs are digested with Xho I, resulting in cDNA flanked by EcoRI sites at the 5'-ends and Xho I sites at the 3'-ends. Digested cDNAs are size-fractionated in Sephacryl S-500 spin columns (Stratagene), then ligated into the λ ZAP Express vector predigested with EcoRI and Xho I. The resulting DNA/cDNA concatomers are packaged using Gigapack Gold packaging extracts. After titration, aliquots of primary packaging mix are stored in 7% DMSO at -80°C as primary library stocks, and the rest are amplified to establish stable library stocks.

From the amplified library, phage plaques are plated onto an appropriate medium. Preferably, phage plaques are plated at a density of 200-500 pfu/150 mm plate onto an *Escherichia coli* XL1-blue MRF' lawn with IPTG/X-gal for color selection. The plaques are then randomly picked and positive inserts are identified by polymerase chain reaction (PCR), according to methods well known in the art and described hereinbelow. Preferably, plaques are picked into 75 ul suspension media buffer (100 mM NaCl, 10 mM $MgSO_4$, 1 mM Tris, pH7.5, 0.02% gelatin). Phage elutes (5 ul) may be used for PCR reactions (50 ul total volume) with 125 umol/L of each dNTP (Pharmacia), 10 pmol each of modified T3 (5'-GCCAAGCTCGAAATTAACCCTCACTAAAGGG-3') and T7 (5'-CCAGTGAATTGTAATACGACTCACTATAGGGCG-3') primers, and 2 U of Taq DNA polymerase (Pharmacia). Reactions are cycled in a DNA Thermal Cycler (Perkin-Elmer) [denaturation at 95°C for 5 minutes, followed by 30 cycles of amplification (94°C, 45 seconds; 55°

C, 30 seconds; 72°C, 3 minutes) and a terminal isothermal extension (72°C, 3 minutes)]. Agarose gel electrophoresis is used to assess the presence and purity of inserts.

The PCR product is then subjected to DNA sequencing using known methods (see Ausubel et al., *supra* and Sambrook et al., *supra*). Methods of sequencing employ such enzymes as the Klenow fragment of DNA polymerase I, Sequenase® (US Biochemical Corp, Cleveland, OH), Taq polymerase (Perkin Elmer, Norwalk, CT), thermostable T7 polymerase (Amersham, Chicago, IL), or combinations of recombinant polymerases and proofreading exonucleases such as the ELONGASE Amplification System (Gibco BRL, Gaithersburg, MD). Preferably, the process is automated with machines such as the Hamilton Micro Lab 2200 (Hamilton, Reno NV), Peltier Thermal Cycler (PTC200; MJ Research, Watertown, MA), the ABI 377 DNA sequencers (Perkin Elmer), and the PE Biosystems ABI Prism 3700 DNA Analyzer..

PCR products are first subjected to DNA sequencing reactions using specific primers, BigDye™ Terminator Cycle Sequencing v2.0 Ready Reaction (PE Biosystems), Tris MgCl buffer and water in a thermocycler. Sequencing reactions were incubated at 94°C for 2 minutes, followed by 25 cycles of 94°C, 30 seconds; 55°C, 20 seconds; and 72°C, 1 minute; and 15 cycles of 94°C, 30 seconds; and 72°C for 1 minute; and 72°C for 5 minutes. Reactions were then put on hold at 4°C until purified using methods well known in the prior art (i.e. alcohol precipitation or ethanol precipitation). Automated sequencing is preferably carried out with a PE Biosystems ABI Prism 3700 DNA Analyzer.

PCR

In one aspect, polynucleotide sequences of the invention are amplified by the polymerase chain reaction (PCR). PCR methods are well-known to those skilled in the art.

PCR provides a method for rapidly amplifying a particular polynucleotide sequence by using multiple cycles of DNA replication catalyzed by a thermostable, DNA-dependent DNA polymerase to amplify the target sequence of interest. PCR requires the presence of a nucleic acid to be amplified, two single-stranded oligonucleotide primers flanking the sequence to be amplified, a DNA polymerase, deoxyribonucleoside triphosphates, a buffer and salts.

The method of PCR is well known in the art. PCR, is performed as described in Mullis and Faloona, 1987, *Methods Enzymol.*, 155: 335, herein incorporated by reference.

PCR is performed using template DNA (at least 1fg; more usefully, 1-1000 ng) and at least 25 pmol of oligonucleotide primers. A typical reaction mixture includes: 2µl of DNA, 25 pmol of oligonucleotide primer, 2.5 µl of 10H PCR buffer 1 (Perkin-Elmer, Foster City, CA), 0.4 µl of 1.25 µM dNTP, 0.15 µl (or 2.5 units) of Taq DNA polymerase (Perkin Elmer, Foster City, CA) and deionized water to a total volume of 25 µl. Mineral oil is overlaid and the PCR is performed using a programmable thermal cycler.

The length and temperature of each step of a PCR cycle, as well as the number of cycles, are adjusted according to the stringency requirements in effect. Annealing temperature and timing are determined both by the efficiency with which a primer is expected to anneal to a template and the degree of mismatch that is to be tolerated. The ability to optimize the stringency of primer annealing conditions is well within the knowledge of one of moderate skill in the art. An annealing temperature of between 30°C and 72°C is used. Initial denaturation of the template molecules normally occurs at between 92°C and 99°C for 4 minutes, followed by 20-40 cycles consisting of denaturation (94-99°C for 15 seconds to 1 minute), annealing (temperature determined as discussed above; 1-2 minutes), and extension (72°C for 1 minute). The final extension step is generally carried out for 4 minutes at 72°C, and may be followed by an indefinite (0-24 hour) step at 4°C.

Several techniques for detecting PCR products quantitatively without electrophoresis may be useful according to the invention . One of these techniques, for which there are commercially available kits such as Taqman™ (Perkin Elmer, Foster City, CA), is performed with a transcript-specific antisense probe. This probe is specific for the PCR product (e.g. a nucleic acid fragment derived from a gene) and is prepared with a quencher and fluorescent reporter probe complexed to the 5' end of the oligonucleotide. Different fluorescent markers are attached to different reporters, allowing for measurement of two products in one reaction. When Taq DNA polymerase is activated, it cleaves off the fluorescent reporters of the probe bound to the template by virtue of its 5'-to-3' exonuclease activity. In the absence of the quenchers, the reporters now fluoresce. The color change in the reporters is proportional to the

amount of each specific product and is measured by a fluorometer; therefore, the amount of each color is measured and the PCR product is quantified. The PCR reactions are performed in 96 well plates so that samples derived from many individuals are processed and measured simultaneously. The Taqman™ system has the additional advantage of not requiring gel electrophoresis and allows for quantification when used with a standard curve.

Polynucleotide Sequences Useful According to the Invention

The invention provides for isolated polynucleotide sequences including ESTs which can be used as probes, arrayed on microarrays, and/or used for the development of therapies to treat osteoarthritis.

In one aspect, cartilage gene expression profiles at different developmental stages are identified. Another aspect of the invention is to monitor cartilage gene expression profiles of osteoarthritis patients diagnosed with different stages of osteoarthritis. A third aspect of the invention is to screen for potential therapeutic agents which alter the gene expression profile of diseased cartilage cells. The invention therefore provides for polynucleotide sequences that are present at each of the following developmental and disease stages: normal, fetal, mild osteoarthritic, moderate osteoarthritic, marked osteoarthritic and severe osteoarthritic. The invention also provides for polynucleotide sequences that are differentially expressed in any two of the following developmental and disease stages: normal, fetal, mild osteoarthritic, moderate osteoarthritic, marked osteoarthritic and severe osteoarthritic.

Polynucleotides useful according to the invention are prepared by isolating cartilage tissue samples from a developmental or disease stage (normal, fetal, mild osteoarthritic, moderate osteoarthritic, marked osteoarthritic and severe osteoarthritic), preparing a cDNA library (as described above), and performing large-scale partial sequencing (described herein) of the cDNA library to generate Expressed Sequence Tags (ESTs). An EST useful according to the invention is preferably in the range of 50-1000 nucleotides and most preferably 50-500 nucleotides in length.

The invention provides for polynucleotide sequences or ESTs that are categorized as "novel" or "known", including "known sequences with a function" and "known sequences without a known function", all defined herein.

Nucleic Acid Members and Probes

In one aspect, the invention provides nucleic acid members and probes that bind specifically to a target nucleic acid sequence (e.g., present in a cartilage nucleic acid sample).

Nucleic acid members are stably associated with a solid support to comprise an array according to the invention. The length of a nucleic acid member can range from 50 to 6000 nucleotides, 100 to 500 nucleotides, and in other embodiments, from 500 to 1500 nucleotides. The nucleic acid members may be single or double stranded, and/or may be PCR fragments amplified from cDNA.

The invention also provides for polynucleotide sequences comprising a probe. In a certain embodiment, a probe is labeled, according to methods known in the art. A probe according to the invention is 50 to 5000 nucleotides, more preferably 100-500 nucleotides and most preferably 50 to 250 nucleotides in length. The probe may be single or double stranded, and may be a PCR fragment amplified from cDNA.

The nucleic acid members and probes according to the invention can be used to detect target sequences such as chondrocyte enriched or chondrocyte-specific sequences, and preferably sequences whose presence in a sample are indicative ,or diagnostic or prognostic, of a stage of osteoarthritis.

The target nucleic acid sequences to be analyzed are preferably from human cartilage, blood or synovial fluid and preferably comprise RNA or nucleic acid corresponding to RNA, (i.e., cDNA or amplified products of RNA or cDNAs).

Data Acquisition and Analysis of EST Sequences

The invention provides for EST sequences including "novel sequences", "novel expressed sequence tags (ESTs)" and "known sequences" including "known sequences with a function" and "known sequences with no known function".

The generated EST sequences are searched against available databases, including the "nt", "nr", "est", "gss" and "htg" databases available through NCBI to determine putative identities for ESTs matching to known genes or other ESTs. Relative EST frequency level can then be

calculated using known methods. Functional characterization of ESTs with known gene matches are made according to any known method. Preferably, generated EST sequences are compared to the non-redundant Genbank/EMBL/DDBJ and dbEST databases using the BLAST algorithm (8). A minimum value of $P = 10^{-10}$ and nucleotide sequence identity >95%, wherein the sequence identity is non-contiguous or scattered, are required for assignments of putative identities for ESTs matching to known genes or to other ESTs. Construction of a non-redundant list of genes represented in the EST set is done with the help of Unigene, Entrez and PubMed at the National Center for Biotechnology Information (NCBI) site (http://www.ncbi.nlm.nih.gov/). Relative gene expression frequency is calculated by dividing the number of EST copies for each gene by the total number of ESTs analyzed.

Genes are identified from ESTs according to known methods. To identify novel genes from an EST sequence, the EST should preferably be at least 100 nucleotides in length, and more preferably 150 nucleotides in length, for annotation. Preferably, the EST exhibits open reading frame characteristics (i.e., can encode a putative polypeptide).

Because of the completion of the Human Genome Project, a specific EST which matches with a genomic sequence can be mapped onto a specific chromosome based on the chromosomal location of the genomic sequence. However, no function may be known for the protein encoded by the sequence and the EST would then be considered "novel" in a functional sense. In one aspect, the invention is used to identify a novel EST which is part of a larger known sequence for which no function is known is used to determine the function of a gene comprising the EST (e.g., such as the role of expression products produced by the gene in chondrogenesis and/or in a pathology affecting chondrocytes). Alternatively, or additionally, the EST can be used to identify an mRNA or polypeptide encoded by the larger sequence as a diagnostic or prognostic marker of chondrogenesis and/or of a pathology affecting chondrocytes.

Having identified an EST corresponding to a larger sequence as chondrocyte enriched or chondrocyte-specific, other portions of the larger sequence which comprises the EST can be used in assays to elucidate gene function, e.g., to isolate polypeptides encoded by the gene, to generate antibodies specifically reactive with these polypeptides, to identify binding partners of the polypeptides (receptors, ligands, agonists, antagonists and the like) and/or to detect the expression of the gene (or lack thereof) in chondrocytes in fetal, adult, normal, and/or diseased individuals.

In another aspect, the invention provides for polynucleotide sequences that do not demonstrate a "significant match" to any of the publicly known sequences in sequence databases at the time a query is done. Longer genomic segments comprising these types of novel EST sequences can be identified by probing genomic libraries, while longer expressed sequences can be identified in cDNA libraries and/or by performing polymerase extension reactions (e.g., RACE) using EST sequences to derive primer sequences as is known in the art. Longer fragments can be mapped to particular chromosomes by FISH and other techniques and their sequences compared to known sequences in genomic and/or expressed sequence databases and further functional analysis can be performed as described above.

Using the methods according to the invention, out of a total of 57,422 ESTs from the four cDNA libraries, no significant match was found for 618 sequences. The remaining sequences were characterized as shown in Figure 5.

Identified genes can be catalogued according to their putative function. Functional characterization of ESTs with known gene matches is preferably made according to the categories described by Hwang et al (5). The distribution of genes in each of the subcellular categories is indicative of the dynamic state of the tissue and will provide important insights into the osteoarthritis disease process. The results of this analysis are provided in Figure 7 where the total number of ESTs identified by the method in different human cartilage libraries are characterized based on the functional classification of known genes identified in each library.

Alternative methods for analyzing ESTs are also available. For example, the ESTs from each library may be assembled into contigs with sequence alignment, editing, and assembly programs such as PHRED and PHRAP (Ewing, et al., 1998, *Genome Res.* 3:175, incorporated herein; http://bozeman.genome.washington.edu/). Contig redundancy is reduced by clustering nonoverlapping sequence contigs using the EST clone identification number, which is common for the nonoverlapping 5' and 3' sequence reads for a single EST cDNA clone. In one aspect, the consensus sequence from each cluster is compared to the non-redundant Genbank/EMBL/DDBJ and dbEST databases using the BLAST algorithm with the help of unigene, Entrez and PubMed at the NCBI site.

Known Polynucleotide Sequences or ESTs and Novel Polynucleotide Sequences or ESTs

An EST that exhibits a significant match (> 65%, and preferably 90% or greater, identity) to at least one existing sequence in an existing polynucleotide sequence database is characterized as a "known" sequence according to the invention. Within this category, some known ESTs match to existing sequences which encode polypeptides with known function(s) and are referred to as a "known sequence with a function". Other "known" ESTs exhibit significant match to existing sequences which encode polypeptides of unknown function(s) and are referred to as a "known sequence with no known function".

In one aspect, the invention also provides for known polynucleotide sequences that are chondrocyte enriched or chondrocyte-specific.

EST sequences which have no significant match (less than 65% identity) to any existing sequence in the above cited available databases are categorized as novel ESTs. These novel ESTs are considered chondrocyte-specific since they are not matched to any other genes or ESTs derived from any other tissue. To identify a novel gene from an EST sequence, the EST is preferably at least 150 nucleotides in length. More preferably, the EST encodes at least part of an open reading frame, that is, a polynucleotide sequence between a translation initiation codon and a termination codon, which is potentially translated into a polypeptide sequence.

The invention provides for known and novel polynucleotide sequences that are uniquely expressed in normal, fetal, mild osteoarthritic, moderate osteoarthritic, marked osteoarthritic and severe osteoarthritic cartilage. Figures 6 and 13, show unique known genes and names of the novel sequences identified to date in the fetal, normal, mild osteoarthritic and severe osteoarthritic cDNA libraries using the methods according to the invention.

The invention also provides for known and novel polynucleotide sequences that are upregulated and downregulated in normal, fetal, mild osteoarthritic, moderate osteoarthritic, marked osteoarthritic and severe osteoarthritic cartilage. In one aspect, polynucleotide sequences are enriched in chondrocytes compared to cells which are non-chondrocytes, or in chondrocytes from individuals with osteoarthritis compared to normal individuals, or in chondrocytes from particular stages of development or disease compared to particular other stages of development or disease.

The invention also provides for polynucleotide sequences that are differentially expressed in cartilage from any two of the following developmental and disease stages: fetal, normal, mild osteoarthritic, moderate osteoarthritic, marked osteoarthritic and severe osteoarthritic.

Relative EST frequency is calculated by dividing the number of EST copies for each gene by the total number of ESTs analyzed. The chondrocyte-specific expression of a number of novel ESTs has been confirmed by methods known in the art. Useful methods for measuring gene expression in a tissue include RT PCR, Northern blot, etc.

Novel Nucleic Acid Molecules

Many of the novel nucleic acid molecules of the present invention are differentially expressed between the mild and severe osteoarthritis disease states and are thus useful as potential drug targets or markers for the osteoarthritis disease process. The invention also provides one or more nucleic acid molecules that are differentially expressed in two or more of the following developmental and disease stages: fetal, normal, mild osteoarthritic, moderate osteoarthritic, marked osteoarthritic and severe osteoarthritic. The invention further provides for one or more novel clones that are differentially expressed in two or more of the following developmental and disease stages: fetal, normal, mild osteoarthritic, moderate osteoarthritic, marked osteoarthritic and severe osteoarthritic.

Microarrays

Polynucleotide Microarrays

Any combination of the polynucleotide sequences generated from any of the chondrocyte cDNA libraries are used for the construction of a microarray. In one embodiment, the microarray is chondrocyte-specific and is anticipated to encompass the entire spectrum of genes that are important in the osteoarthritis disease process. A microarray according to the invention preferably comprises between 10 and 20,000 nucleic acid members, and more preferably comprises at least 5000 nucleic acid members. The nucleic acid members are known or novel polynucleotide sequences described herein, or any combination thereof. A microarray according to the invention is used to confirm differential gene expression profiles of genes that are specifically expressed at different cartilage development and osteoarthritis disease stages.

The invention also provides for a microarray comprising genes that are differentially expressed between normal and mild osteoarthritis patients to allow for the identification of early risk factors for osteoarthritis development. The invention also provides for a microarray for osteoarthritis diagnosis comprising one or more polynucleotide sequences that are differentially expressed between a normal individual and a patient diagnosed with mild, moderate, marked or severe osteoarthritis. Such arrays also may be used for prognostic methods to monitor a patient's response to therapy. Preferably, an array for osteoarthritis diagnosis comprises 10-20,000 nucleic acid members and more preferably 50-15,000 nucleic acid members. In one embodiment, the above microarrays are used to identify a therapeutic agent that modulates the anabolic activity of a chondrocyte or changes (e.g., increases or decreases) the level of expression of at least one polynucleotide sequence that is differentially expressed in a chondrocyte derived from any of the following chondrocyte disease or developmental stages: fetal, normal, mild osteoarthritic, moderate osteoarthritic, marked osteoarthritic and severe osteoarthritic.

The target nucleic acid samples that are hybridized to and analyzed with a microarray of the invention are preferably from human cartilage, blood or synovial fluid. A limitation for this procedure lies in the amount of RNA available for use as a target nucleic acid sample. Preferably, at least 1 microgram of total RNA is obtained for use according to this invention. This is advantageous because the amount of RNA in synovial fluid and in many cartilage biopsy samples is very minimal.

Construction of a Microarray

In one aspect, cDNAs generated from human cartilage cDNA libraries are arrayed on a microarray. Preferably, a microarray according to the invention comprises chondrocyte enriched or chondrocyte-specific genes and includes the whole spectrum of genes that are important in the osteoarthritis disease process.

The EST frequency analysis in Figure 6 (and portions thereof shown in Figures 15 and 16) shows the differential gene expression profiles for known genes. Microarrays according to the invention may be used to confirm these profiles and may also be used to show differential expression profiles between different developmental stages and osteoarthritis disease states for novel EST sequences. These novel EST sequences may be further characterized by cluster and

alignment analyses to determine how many unique genes are represented by the novel EST sequences. The novel unique genes identified may provide a basis for identifying key markers in osteoarthritis disease progression and treatment.

In the subject methods, an array of nucleic acid members stably associated with the surface of a substantially solid support is contacted with a sample comprising target polynucleotides under hybridization conditions sufficient to produce a hybridization pattern of complementary nucleic acid members/target complexes in which one or more complementary nucleic acid members at unique positions on the array specifically hybridize to target nucleic acids. The identity of target nucleic acids which hybridize can be determined with reference to location of nucleic acid members on the array.

The nucleic acid members may be produced using established techniques such as polymerase chain reaction (PCR) and reverse transcription (RT). These methods are similar to those currently known in the art (see e.g., *PCR Strategies*, Michael A. Innis (Editor), et al. (1995) and *PCR: Introduction to Biotechniques Series*, C. R. Newton, A. Graham (1997)). Amplified polynucleotides are purified by methods well known in the art (e.g., column purification or alcohol precipitation). A polynucleotide is considered pure when it has been isolated so as to be substantially free of primers and incomplete products produced during the synthesis of the desired polynucleotide. Preferably, a purified polynucleotide will also be substantially free of contaminants which may hinder or otherwise mask the specific binding activity of the molecule.

A microarray according to the invention comprises a plurality of unique polynucleotides attached to one surface of a solid support at a density exceeding 20 different polynucleotides/cm^2, wherein each of the polynucleotides is attached to the surface of the solid support in a non-identical pre-selected region. Each associated sample on the array comprises a polynucleotide composition, of known identity, usually of known sequence, as described in greater detail below. Any conceivable substrate may be employed in the invention.

In one embodiment, the polynucleotide attached to the surface of the solid support is DNA. In a preferred embodiment, the polynucleotide attached to the surface of the solid support is cDNA or RNA. In another preferred embodiment, the polynucleotide attached to the surface of the solid support is cDNA synthesized by polymerase chain reaction (PCR). Preferably, a nucleic acid

member in the array, according to the invention, is at least 50 nucleotides in length. In one embodiment, a nucleic acid member is at least 150 nucleotides in length. Preferably, a nucleic acid member is less than 1000 nucleotides in length. More preferably, a nucleic acid member is less than 500 nucleotides in length. In one embodiment, an array comprises at least 10 different polynucleotides attached to one surface of the solid support. In another embodiment, the array comprises at least 100 different polynucleotides attached to one surface of the solid support. In yet another embodiment, the array comprises at least 10,000 different polynucleotides attached to one surface of the solid support. In yet another embodiment, the array comprises at least 15,000 different polynucleotides attached to one surface of the solid support.

In the arrays of the invention, the polynucleotide compositions are stably associated with the surface of a solid support, wherein the support may be a flexible or rigid solid support. By "stably associated" is meant that each nucleic acid member maintains a unique position relative to the solid support under hybridization and washing conditions. As such, the samples are non-covalently or covalently stably associated with the support surface. Examples of non-covalent association include non-specific adsorption, binding based on electrostatic interactions (e.g., ion pair interactions), hydrophobic interactions, hydrogen bonding interactions, specific binding through a specific binding pair member covalently attached to the support surface, and the like. Examples of covalent binding include covalent bonds formed between the polynucleotides and a functional group present on the surface of the rigid support (e.g., --OH), where the functional group may be naturally occurring or present as a member of an introduced linking group, as described in greater detail below

The amount of polynucleotide present in each composition will be sufficient to provide for adequate hybridization and detection of target polynucleotide sequences during the assay in which the array is employed. Generally, the amount of each nucleic acid member stably associated with the solid support of the array is at least about 0.001 ng, preferably at least about 0.02 ng and more preferably at least about 0.05 ng, where the amount may be as high as 1000 ng or higher, but will usually not exceed about 20 ng. Where the nucleic acid member is "spotted" onto the solid support in a spot comprising an overall circular dimension, the diameter of the "spot" will generally range from about 10 to 5,000 μm, usually from about 20 to 2,000 μm and more usually from about 100 to 200 μm.

Control nucleic acid members may be present on the array including nucleic acid members comprising oligonucleotides or polynucleotides corresponding to genomic DNA, housekeeping genes, vector sequences, plant nucleic acid sequence, negative and positive control genes, and the like. Control nucleic acid members are calibrating or control genes whose function is not to tell whether a particular "key" gene of interest is expressed, but rather to provide other useful information, such as background or basal level of expression.

Other control polynucleotides are spotted on the array and used as target expression control polynucleotides and mismatch control nucleotides to monitor non-specific binding or cross-hybridization to a polynucleotide in the sample other than the target to which the probe is directed. Mismatch probes thus indicate whether a hybridization is specific or not. For example, if the target is present, the perfectly matched probes should be consistently brighter than the mismatched probes. In addition, if all control mismatches are present, the mismatch probes are used to detect a mutation.

Solid Substrate

An array according to the invention comprises either a flexible or rigid substrate. A flexible substrate is capable of being bent, folded or similarly manipulated without breakage. Examples of solid materials which are flexible solid supports with respect to the present invention include membranes, e.g., nylon, flexible plastic films, and the like. By "rigid" is meant that the support is solid and does not readily bend, i.e., the support is not flexible. As such, the rigid substrates of the subject arrays are sufficient to provide physical support and structure to the associated polynucleotides present thereon under the assay conditions in which the array is employed, particularly under high throughput handling conditions.

The substrate may be biological, non-biological, organic, inorganic, or a combination of any of these, existing as particles, strands, precipitates, gels, sheets, tubing, spheres, beads, containers, capillaries, pads, slices, films, plates, slides, chips, etc. The substrate may have any convenient shape, such as a disc, square, sphere, circle, etc. The substrate is preferably flat or planar but may take on a variety of alternative surface configurations. The substrate may be a polymerized Langmuir Blodgett film, functionalized glass, Si, Ge, GaAs, GaP, SiO_2, SIN_4, modified silicon, or any one of a wide variety of gels or polymers such as (poly)tetrafluoroethylene,

(poly)vinylidenedifluoride, polystyrene, polycarbonate, or combinations thereof. Other substrate materials will be readily apparent to those of skill in the art upon review of this disclosure.

In a preferred embodiment the substrate is flat glass or single-crystal silicon. According to some embodiments, the surface of the substrate is etched using well-known techniques to provide for desired surface features. For example, by way of formation of trenches, v-grooves, mesa structures, or the like, the synthesis regions may be more closely placed within the focus point of impinging light, be provided with reflective "mirror" structures for maximization of light collection from fluorescent sources, etc.

Surfaces on the solid substrate will usually, though not always, be composed of the same material as the substrate. Alternatively, the surface may be composed of any of a wide variety of materials, for example, polymers, plastics, resins, polysaccharides, silica or silica-based materials, carbon, metals, inorganic glasses, membranes, or any of the above-listed substrate materials. In some embodiments the surface may provide for the use of caged binding members which are attached firmly to the surface of the substrate. Preferably, the surface will contain reactive groups, which are carboxyl, amino, hydroxyl, or the like. Most preferably, the surface will be optically transparent and will have surface Si--OH functionalities, such as are found on silica surfaces.

The surface of the substrate is preferably provided with a layer of linker molecules, although it will be understood that the linker molecules are not required elements of the invention. The linker molecules are preferably of sufficient length to permit polynucleotides of the invention and on a substrate to hybridize to other polynucleotide molecules and to interact freely with molecules exposed to the substrate.

Often, the substrate is a silicon or glass surface, (poly)tetrafluoroethylene, (poly)vinylidendifluoride, polystyrene, polycarbonate, a charged membrane, such as nylon 66 or nitrocellulose, or combinations thereof. In a preferred embodiment, the solid support is glass. Preferably, at least one surface of the substrate will be substantially flat. Preferably, the surface of the solid support will contain reactive groups, including, but not limited to, carboxyl, amino, hydroxyl, thiol, or the like. In one embodiment, the surface is optically transparent. In a preferred embodiment, the substrate is a poly-lysine coated slide or Gamma amino propyl silane-coated Corning Microarray Technology-GAPS or CMT-GAP2 coated slides.

Any solid support to which a nucleic acid member may be attached may be used in the invention. Examples of suitable solid support materials include, but are not limited to, silicates such as glass and silica gel, cellulose and nitrocellulose papers, nylon, polystyrene, polymethacrylate, latex, rubber, and fluorocarbon resins such as TEFLON™.

The solid support material may be used in a wide variety of shapes including, but not limited to slides and beads. Slides provide several functional advantages and thus are a preferred form of solid support. Due to their flat surface, probe and hybridization reagents are minimized using glass slides. Slides also enable the targeted application of reagents, are easy to keep at a constant temperature, are easy to wash and facilitate the direct visualization of RNA and/or DNA immobilized on the solid support. Removal of RNA and/or DNA immobilized on the solid support is also facilitated using slides.

The particular material selected as the solid support is not essential to the invention, as long as it provides the described function. Normally, those who make or use the invention will select the best commercially available material based upon the economics of cost and availability, the expected application requirements of the final product, and the demands of the overall manufacturing process.

Spotting Method

In one aspect, The invention provides for arrays wherein each nucleic acid member comprising the array is spotted onto a solid support.

Preferably, spotting is carried out as follows. PCR products (~40 ul) of cDNA clones from osteoarthritis, fetal or normal cartilage cDNA libraries, in the same 96-well tubes used for amplification, are precipitated with 4 ul (1/10 volume) of 3M sodium acetate (pH 5.2) and 100 ul (2.5 volumes) of ethanol and stored overnight at –20°C. They are then centrifuged at 3,300 rpm at 4°C for 1 hour. The obtained pellets are washed with 50 ul ice-cold 70% ethanol and centrifuged again for 30 minutes. The pellets are then air-dried and resuspended well in 20ul 3X SSC or in 50% dimethylsulfoxide (DMSO) overnight. The samples are then spotted, either singly or in duplicate, onto polylysine-coated slides (Sigma Cat. No. P0425) using a robotic GMS 417 or 427 arrayer (Affymetrix, Ca).

The boundaries of the spots on the microarray may be marked with a diamond scriber (as the spots become invisible after post-processing). The arrays are rehydrated by suspending the slides over a dish of warm particle free ddH_20 for approximately one minute (the spots will swell slightly but will not run into each other) and snap-dried on a 70-80°C inverted heating block for 3 seconds. Nucleic acid is then UV crosslinked to the slide (Stratagene, Stratalinker, 65 mJ – set display to "650" which is 650 x 100 uJ) or the array is baked at 80C for two to four hours prior to hybridization. The arrays are placed in a slide rack. An empty slide chamber is prepared and filled with the following solution: 3.0 grams of succinic anhydride (Aldrich) was dissolved in 189 ml of 1-methyl-2-pyrrolidinone (rapid addition of reagent is crucial); immediately after the last flake of succinic anhydride is dissolved, -21.0 ml of 0.2 M sodium borate is mixed in and the solution is poured into the slide chamber. The slide rack is plunged rapidly and evenly in the slide chamber and vigorously shaken up and down for a few seconds, making sure the slides never leave the solution, and then mixed on an orbital shaker for 15-20 minutes. The slide rack is then gently plunged in 95°C ddH_20 for 2 minutes, followed by plunging five times in 95% ethanol. The slides are then air dried by allowing excess ethanol to drip onto paper towels. The arrays are stored in the slide box at room temperature until use.

Numerous methods may be used for attachment of the nucleic acid members of the invention to the substrate (a process referred to as "spotting"). For example, polynucleotides are attached using the techniques of, for example U.S. Pat. No. 5,807,522, which is incorporated herein by reference, for teaching methods of polymer attachment.

Alternatively, spotting may be carried out using contact printing technology as is known in the art.

Kits

The invention provides for kits for performing expression assays using the arrays of the present invention. Such kits according to the subject invention will at least comprise the arrays of the invention having associated nucleic acid members and packaging means therefore. The kits may further comprise one or more additional reagents employed in the various methods, such as: 1) primers for generating test polynucleotides; 2) dNTPs and/or rNTPs (either premixed or separate), optionally with one or more uniquely labeled dNTPs and/or rNTPs (e.g., biotinylated or Cy3 or Cy5

tagged dNTPs); 3) post synthesis labeling reagents, such as chemically active derivatives of fluorescent dyes; 4) enzymes, such as reverse transcriptases, DNA polymerases, and the like; 5) various buffer mediums, e.g., hybridization and washing buffers; 6) labeled probe purification reagents and components, like spin columns, etc.; and 7) signal generation and detection reagents, e.g., streptavidin-alkaline phosphatase conjugate, chemifluorescent or chemiluminescent substrate, and the like.

Use of a Microarray

Polynucleotide arrays according to the invention can be used in high throughput techniques that can assay a large number of polynucleotides in a sample comprising one or more target nucleic acid sequences. The arrays of the subject invention find use in a variety of applications, including gene expression analysis, diagnosis of osteoarthritis and prognosis of osteoarthritis, monitoring a patient's response to therapy, drug screening, and the like.

In one aspect, the arrays of the invention are used in, among other applications, differential gene expression assays. For example, arrays are useful in the differential expression analysis of: (a) diseased osteoarthritis and normal tissue; (b) tissues representing different stages of osteoarthritis; (c) developing cartilage (e.g., fetal cartilage); (d) chondrocyte responses to external or internal stimuli; (e) cartilage/chondrocyte response to treatment; (f) cartilage tissue engineering; (g) pharmacogenomics; and the like. The arrays are also useful in broad scale expression screening for drug discovery and research, such as the effect of a particular active agent on the expression pattern of genes in a particular cell, where such information is used to reveal drug efficacy and toxicity, environmental monitoring, disease research and the like. For example, high expression of a particular polynucleotide sequence in an osteoarthritis sample (mild, moderate, marked, or severe), which is not observed in a corresponding normal cell, can indicate an osteoarthritis-specific gene product.

Target Preparation

The targets for the microarrays according to the invention are preferably derived from human cartilage, blood or synovial fluid.

A target polynucleotide is capable of binding to a polynucleotide probe or nucleic acid member of complementary sequence through one or more types of chemical bonds, usually through complementary base pairing, usually through hydrogen bond formation.

As used herein, a "polynucleotide derived from an mRNA transcript: or a "polynucleotide corresponding to an mRNA" refers to a polynucleotide for which synthesis of the mRNA transcript or a sub-sequence thereof has ultimately served as a template. Thus, a cDNA reverse transcribed from an mRNA, an RNA transcribed from that cDNA, a DNA amplified from the cDNA, an RNA transcribed from the amplified DNA, etc., are all derived from or correspond to the mRNA transcript and detection of such derived or corresponding products is indicative of or proportional to the presence and/or abundance of the original transcript in a sample. Thus, suitable target nucleic acid samples include, but are not limited to, mRNA transcripts of a gene or genes, cDNA reverse transcribed from the mRNA, cRNA transcribed from the cDNA, DNA amplified from a gene or genes, RNA transcribed from amplified DNA, and the like. The polynucleotide targets used herein are preferably derived from human cartilage, blood or synovial fluid. Preferably, the targets are polynucleotides derived from human cartilage, blood or synovial fluid extracts. Polynucleotides can be single- or double-stranded DNA, RNA, or DNA-RNA hybrids synthesized from human cartilage, blood or synovial fluid mRNA extracts using methods known in the art, for example, reverse transcription or PCR.

In the simplest embodiment, such a polynucleotide target comprises total mRNA or a nucleic acid sample corresponding to mRNA (e.g., cDNA) isolated from cartilage, blood, or synovial fluid samples. In another embodiment, total mRNA is isolated from a given sample using, for example, an acid guanidinium-phenol-chloroform extraction method and polyA+ mRNA is isolated by oligo dT column chromatography or by using (dT)n magnetic beads (see, e.g., Sambrook et al., *Molecular Cloning: A Laboratory Manual* (2nd ed.), Vols. 1-3, Cold Spring Harbor Laboratory, (1989), or Current Protocols in Molecular Biology, F. Ausubel et al., ed. Greene Publishing and Wiley-Interscience, New York (1987). In a preferred embodiment, total RNA is extracted using TRIzol® reagent (GIBCO/BRL, Invitrogen Life Technologies, Cat. No. 15596). Purity and integrity of RNA is assessed by absorbance at 260/280nm and agarose gel electrophoresis followed by inspection under ultraviolet light.

In some embodiments, it is desirable to amplify the target nucleic acid sample prior to hybridization, for example, when synovial fluid is used. One of skill in the art will appreciate that whatever amplification method is used, if a quantitative result is desired, care must be taken to use a method that maintains or controls for the relative frequencies of the amplified polynucleotides. Methods of "quantitative" amplification are well known to those of skill in the art. For example, quantitative PCR involves simultaneously co-amplifying a known quantity of a control sequence using the same primers. This provides an internal standard that may be used to calibrate the PCR reaction. The high density array may then include probes specific to the internal standard for quantification of the amplified polynucleotide. Detailed protocols for quantitative PCR are provided in *PCR Protocols, A Guide to Methods and Applications*, Innis et al., Academic Press, Inc. N.Y., (1990).

Other suitable amplification methods include, but are not limited to polymerase chain reaction (PCR) (Innis, et al., *PCR Protocols. A Guide to Methods and Application.* Academic Press, Inc. San Diego, (1990)), ligase chain reaction (LCR) (see Wu and Wallace, 1989, *Genomics*, 4:560; Landegren, et al., 1988, *Science*, 241:1077 and Barringer, et al., 1990, *Gene*, 89:117, transcription amplification (Kwoh, et al., 1989, *Proc. Natl. Acad. Sci. USA*, 86: 1173), and self-sustained sequence replication (Guatelli, et al., 1990, *Proc. Nat. Acad. Sci. USA,* 87: 1874).

In a particularly preferred embodiment, the target nucleic acid sample mRNA is reverse transcribed with a reverse transcriptase and a primer consisting of oligo dT and a sequence encoding the phage T7 promoter to provide single-stranded DNA template. The second DNA strand is polymerized using a DNA polymerase. After synthesis of double-stranded cDNA, T7 RNA polymerase is added and RNA is transcribed from the cDNA template. Successive rounds of transcription from each single cDNA template results in amplified RNA. Methods of *in vitro* transcription are well known to those of skill in the art (see, e.g., Sambrook, *supra.*) and this particular method is described in detail by Van Gelder, et al., 1990, *Proc. Natl. Acad. Sci. USA*, 87: 1663-1667 who demonstrate that *in vitro* amplification according to this method preserves the relative frequencies of the various RNA transcripts. Moreover, Eberwine et al. *Proc. Natl. Acad. Sci. USA*, 89: 3010-3014 provide a protocol that uses two rounds of amplification via *in vitro* transcription to achieve greater than 10^6 fold amplification of the original starting material thereby permitting expression monitoring even where biological samples are limited.

Labeling of Target or Nucleic Acid Probe

Either the target or the probe can be labeled.

Any analytically detectable marker that is attached to or incorporated into a molecule may be used in the invention. An analytically detectable marker refers to any molecule, moiety or atom which is analytically detected and quantified.

Detectable labels suitable for use in the present invention include any composition detectable by spectroscopic, photochemical, biochemical, immunochemical, electrical, optical or chemical means. Useful labels in the present invention include biotin for staining with labeled streptavidin conjugate, magnetic beads (e.g., Dynabeads™), fluorescent dyes (e.g., fluorescein, texas red, rhodamine, green fluorescent protein, and the like), radiolabels (e.g., ^{3}H, ^{125}I, 35S, ^{14}C, or ^{32}P), enzymes (e.g., horse radish peroxidase, alkaline phosphatase and others commonly used in an ELISA), and colorimetric labels such as colloidal gold or colored glass or plastic (e.g., polystyrene, polypropylene, latex, etc.) beads. Patents teaching the use of such labels include U.S. Pat. Nos. 3,817,837; 3,850,752; 3,939,350; 3,996,345; 4,277,437; 4,275,149; and 4,366,241, the entireties of which are incorporated by reference herein.

Means of detecting such labels are well known to those of skill in the art. Thus, for example, radiolabels may be detected using photographic film or scintillation counters, fluorescent markers may be detected using a photodetector to detect emitted light. Enzymatic labels are typically detected by providing the enzyme with a substrate and detecting the reaction product produced by the action of the enzyme on the substrate, and colorimetric labels are detected by simply visualizing the colored label.

The labels may be incorporated by any of a number of means well known to those of skill in the art. However, in a preferred embodiment, the label is simultaneously incorporated during the amplification step in the preparation of the sample polynucleotides. Thus, for example, polymerase chain reaction (PCR) with labeled primers or labeled nucleotides will provide a labeled amplification product. In a preferred embodiment, transcription amplification, as described above, using a labeled nucleotide (e.g. fluorescein-labeled UTP and/or CTP) incorporates a label into the transcribed polynucleotides.

Alternatively, a label may be added directly to the original polynucleotide sample (e.g., mRNA, polyA mRNA, cDNA, etc.) or to the amplification product after the amplification is completed. Means of attaching labels to polynucleotides are well known to those of skill in the art and include, for example, nick translation or end-labeling (e.g. with a labeled RNA) by kinasing of the polynucleotide and subsequent attachment (ligation) of a polynucleotide linker joining the sample polynucleotide to a label (e.g., a fluorophore).

In a preferred embodiment, the fluorescent modifications are by cyanine dyes e.g. Cy-3/Cy-5 dUTP, Cy-3/Cy-5 dCTP (Amersham Pharmacia) or alexa dyes (Khan,et al., 1998, *Cancer Res.* 58:5009-5013).

In a preferred embodiment, the two target samples used for comparison are labeled with different fluorescent dyes which produce distinguishable detection signals, for example, targets made from normal cartilage are labeled with Cy5 and targets made from mild osteoarthritis cartilage are labeled with Cy3. The differently labeled target samples are hybridized to the same microarray simultaneously. In a preferred embodiment, the labeled targets are purified using methods known in the art, e.g., by ethanol purification or column purification.

In a preferred embodiment, the target will include one or more control molecules which hybridize to control probes on the microarray to normalize signals generated from the microarray. Preferably, labeled normalization targets are polynucleotide sequences that are perfectly complementary to control oligonucleotides that are spotted onto the microarray as described above. The signals obtained from the normalization controls after hybridization provide a control for variations in hybridization conditions, label intensity, "reading" efficiency and other factors that may cause the signal of a perfect hybridization to vary between arrays. In a preferred embodiment, signals (e.g., fluorescence intensity) read from all other probes in the array are divided by the signal (e.g., fluorescence intensity) from the control probes, thereby normalizing the measurements.

Preferred normalization targets are selected to reflect the average length of the other targets present in the sample, however, they are selected to cover a range of lengths. The normalization control(s) also can be selected to reflect the (average) base composition of the other probes in the array, however, in a preferred embodiment, only one or a few normalization probes are used and

they are selected such that they hybridize well (i.e., have no secondary structure and do not self hybridize) and do not match any target molecules.

Normalization probes are localized at any position in the array or at multiple positions throughout the array to control for spatial variation in hybridization efficiency. In a preferred embodiment, normalization controls are located at the corners or edges of the array as well as in the middle.

Hybridization Conditions

Polynucleotide hybridization involves providing a denatured probe or target nucleic acid member and target polynucleotide under conditions where the probe or target nucleic acid member and its complementary target can form stable hybrid duplexes through complementary base pairing. The polynucleotides that do not form hybrid duplexes are then washed away leaving the hybridized polynucleotides to be detected, typically through detection of an attached detectable label. It is generally recognized that polynucleotides are denatured by increasing the temperature or decreasing the salt concentration of the buffer containing the polynucleotides. Under low stringency conditions (e.g., low temperature and/or high salt) hybrid duplexes (e.g., DNA:DNA, RNA:RNA, or RNA:DNA) will form even where the annealed sequences are not perfectly complementary. Thus specificity of hybridization is reduced at lower stringency. Conversely, at higher stringency (e.g., higher temperature or lower salt) successful hybridization requires fewer mismatches.

The invention provides for hybridization conditions comprising the Dig hybridization mix (Boehringer); or formamide-based hybridization solutions, for example as described in Ausubel et al., *supra* and Sambrook et al. *supra.*

Methods of optimizing hybridization conditions are well known to those of skill in the art (see, e.g., *Laboratory Techniques in Biochemistry and Molecular Biology*, Vol. 24: *Hybridization With Polynucleotide Probes*, P. Tijssen, ed. Elsevier, N.Y., (1993)).

Following hybridization, non-hybridized labeled or unlabeled polynucleotide is removed from the support surface, conveniently by washing, thereby generating a pattern of hybridized target polynucleotide on the substrate surface. A variety of wash solutions are known to those of skill in the art and may be used. The resultant hybridization patterns of labeled, hybridized

oligonucleotides and/or polynucleotides may be visualized or detected in a variety of ways, with the particular manner of detection being chosen based on the particular label of the test polynucleotide, where representative detection means include scintillation counting, autoradiography, fluorescence measurement, calorimetric measurement, light emission measurement and the like.

Image Acquisition and Data Analysis

Following hybridization and any washing step(s) and/or subsequent treatments, as described above, the resultant hybridization pattern is detected. In detecting or visualizing the hybridization pattern, the intensity or signal value of the label will be not only be detected but quantified, by which is meant that the signal from each spot of the hybridization will be measured and compared to a unit value corresponding to the signal emitted by a known number of end labeled target polynucleotides to obtain a count or absolute value of the copy number of each end-labeled target that is hybridized to a particular spot on the array in the hybridization pattern.

Methods for analyzing the data collected from hybridization to arrays are well known in the art. For example, where detection of hybridization involves a fluorescent label, data analysis can include the steps of determining fluorescent intensity as a function of substrate position from the data collected, removing outliers, i.e., data deviating from a predetermined statistical distribution, and calculating the relative binding affinity of the test polynucleotides from the remaining data. The resulting data is displayed as an image with the intensity in each region varying according to the binding affinity between associated oligonucleotides and/or polynucleotides and the test polynucleotides.

The following detection protocol is used for the simultaneous analysis of two cartilage samples to be compared, wherein each sample is labeled with a different fluorescent dye.

Each element of the microarray is scanned for the first fluorescent color. The intensity of the fluorescence at each array element is proportional to the expression level of that gene in the sample.

The scanning operation is repeated for the second fluorescent label. The ratio of the two fluorescent intensities provides a highly accurate and quantitative measurement of the relative gene expression level in the two tissue samples.

In a preferred embodiment, fluorescence intensities of immobilized target nucleic acid sequences were determined from images taken with a custom confocal microscope equipped with laser excitation sources and interference filters appropriate for the Cy3 and Cy5 fluors. Separate scans were taken for each fluor at a resolution of 225 μm^2 per pixel and 65,536 gray levels. Image segmentation to identify areas of hybridization, normalization of the intensities between the two fluor images, and calculation of the normalized mean fluorescent values at each target are as described (Khan, et al., 1998, *Cancer Res.* 58:5009-5013. Chen, et al., 1997, *Biomed. Optics* 2:364-374). Normalization between the images is used to adjust for the different efficiencies in labeling and detection with the two different fluors. This is achieved by equilibrating to a value of one the signal intensity ratio of a set of internal control genes spotted on the array.

In another preferred embodiment, the array is scanned in the Cy 3 and Cy5 channels and stored as separate 16-bit TIFF images. The images are incorporated and analysed using software which includes a gridding process to capture the hybridization intensity data from each spot on the array. The fluorescence intensity and background-subtracted hybridization intensity of each spot is collected and a ratio of measured mean intensities of Cy5 to Cy3 is calculated. A liner regression approach is used for normalization and assumes that a scatter plot of the measured Cy5 versus Cy3 intensities should have a scope of one. The average of the ratios is calculated and used to rescale the data and adjust the slope to one. A post-normalization cutoff of greater than 1.0 fold up- or down-regulation is used to identify differentially expressed genes.

Following detection or visualization, the hybridization pattern is used to determine quantitative information about the genetic profile of the labeled target polynucleotide sample that was contacted with the array to generate the hybridization pattern, as well as the physiological source from which the labeled target polynucleotide sample was derived. By "genetic profile" is meant information regarding the types of polynucleotides present in the sample, e.g., such as the types of genes to which they are complementary, and/or the copy number of each particular polynucleotide in the sample. From this data, one can also derive information about the physiological source from which the target polynucleotide sample was derived, such as the types of genes expressed in the tissue or cell which is the physiological source of the target, as well as the levels of expression of each gene, particularly in quantitative terms.

Where one uses the subject methods to compare target polynucleotides from two or more physiological sources, the hybridization patterns may be compared to identify differences between the patterns. Where arrays in which each of the different nucleic acid members corresponds to a known gene are employed, any discrepancies are related to a differential expression of a particular gene in the physiological sources being compared. Thus, the subject methods find use in differential gene expression assays, where one may use the subject methods in the differential expression analysis of: (a) diseased vs. normal tissue, e.g., osteoarthritic and normal tissue, (b) tissue derived from different stages of osteoarthritis; and the like.

In a particularly preferred embodiment, where it is desired to quantify the transcription level (and thereby expression) of one or more polynucleotide sequences in a sample, the target nucleic acid sample is one in which the concentration of the mRNA transcript(s) of the gene or genes, or the concentration of the polynucleotides derived from the mRNA transcript(s), is proportional to the transcription level (and therefore expression level) of that gene. Similarly, it is preferred that the hybridization signal intensity be proportional to the amount of hybridized polynucleotide. While it is preferred that the proportionality be relatively strict (e.g., a doubling in transcription rate results in a doubling in mRNA transcript in the sample polynucleotide pool and a doubling in hybridization signal), one of skill will appreciate that the proportionality can be more relaxed and even non-linear and still provide meaningful results. Thus, for example, an assay where a 5 fold difference in concentration of the target mRNA results in a 3- to 6-fold difference in hybridization intensity is sufficient for most purposes. Where more precise quantification is required, appropriate controls are run to correct for variations introduced in sample preparation and hybridization as described herein. In addition, serial dilutions of "standard" target mRNAs are used to prepare calibration curves according to methods well known to those of skill in the art. Of course, where simple detection of the presence or absence of a transcript is desired, no elaborate control or calibration is required.

For example, if a microarray nucleic acid member is not labeled after hybridization, this indicates that the gene comprising that nucleic acid member is not expressed in either sample. If a nucleic acid member is labeled with a single color, it indicates that a labeled gene was expressed only in one sample. The labeling of a nucleic acid member comprising an array with both colors indicates that the gene was expressed in both samples. Even genes expressed once per cell are

detected (1 part in 100,000 sensitivity). A difference in expression intensity in the two samples being compared is indicative of differential expression, the ratio of the intensity in the two samples being not equal to 1.0, preferably less than 0.7 or greater than 1.2, more preferably less than 0.5 or greater than 1.5.

Many human genes are expressed at different levels in cartilage of different developmental (fetal vs. mature) or disease states. In some cases, a gene is not expressed at all in some developmental or disease states, and at high levels in others. Differential analysis of chondrocyte gene expression in differing cartilage states using an EST-based approach is used to identify genes that may play important roles in osteoarthritis pathogenesis and cartilage repair. The advantage of this method is that it can provide gene expression information on a larger scale than other methods. The cDNA clones generated by this approach is useful for future functional studies of certain genes. This type of genomic-based approach can provide important novel insights into our understanding of the osteoarthritis disease process and provide for novel diagnostic, prognostic and therapeutic approaches.

Diagnostic or Prognostic Tests

The invention also provides for diagnostic tests for detecting osteoarthritis. The invention also provides for prognostic tests for monitoring a patient's response to therapy.

According to the method of the invention, mild, moderate, marked or severe osteoarthritis is detected by obtaining a cartilage sample from a patient. In alternative embodiments, a blood or synovial fluid sample is obtained from a patient . A sample comprising nucleic acid corresponding to RNA (i.e., RNA or cDNA) is prepared from the patient cartilage (or blood or synovial fluid) sample. The sample comprising nucleic acid corresponding to RNA is hybridized to an array comprising a solid substrate and a plurality of nucleic acid members, wherein at least one member is differentially expressed in cartilage isolated from a patient diagnosed with mild, moderate, marked or severe osteoarthritis, as compared to a "normal individual", according to the invention. According to this diagnostic test, hybridization of the sample comprising nucleic acid corresponding to RNA to one or more nucleic acid members on the array is indicative of disease.

A patient response to therapy is monitored by using a prognostic test according to the invention. In one aspect, a prognostic test according to the invention comprises obtaining a

cartilage sample from a patient prior to treatment, during the course of treatment and after treatment. Preferably, the patient is treated for at least 12 hours before a sample is taken. In alternative embodiments, blood or synovial fluid samples are obtained from a patient prior to treatment, during the course of treatment and after treatment . A sample comprising nucleic acid corresponding to RNA (i.e., RNA or cDNA) is prepared from the patient cartilage (or blood or synovial fluid) samples. The samples comprising nucleic acid corresponding to RNA are hybridized to an array comprising a solid substrate and a plurality of nucleic acid members, wherein at least one member is differentially expressed in cartilage isolated from a patient diagnosed with mild, moderate, marked or severe osteoarthritis, as compared to a normal individual, according to the invention. Arrays are selected in accordance with the diagnostic state of the patient whose treatment is being monitored. According to this prognostic test, differential hybridization of the samples comprising nucleic acid corresponding to RNA isolated prior to and after treatment to one or more nucleic acid members on the array is indicative of an effective treatment. Preferably, gene expression profiles in patients being treated changes to resemble more closely gene expression profiles in patients with less severe forms of the disease or more preferably more closely resembles gene expression profiles in normal patients. The extent of change in a gene expression profile can be further correlated with various therapeutic endpoints such as a decrease in the severity and/or occurrence of one or more symptoms associated with the disease.

Therapeutic Agents

A useful therapeutic agent according to the invention can increase or decrease the anabolic and/or the catabolic activity of a chondrocyte. Preferably, a therapeutic agent can increase or decrease the anabolic and/or catabolic activity of a chondrocyte by greater than 1.0-fold, more preferably, 1.5-5-fold, and most preferably, 5-100-fold, as compared to an untreated chondrocyte.

In one embodiment, a therapeutic agent changes (e.g., increases or decreases) the level of expression of at least one polynucleotide sequence that is differentially expressed in a chondrocyte derived from any of the following chondrocyte disease or developmental stages: fetal, normal, mild osteoarthritic, moderate osteoarthritic, marked osteoarthritic and severe osteoarthritic. Preferably, a therapeutic agent causes a change in the level of expression of a polynucleotide sequence or increase or decrease in the expression of a polynucleotide sequence that is differentially expressed in a chondrocyte derived from any of the following chondrocyte disease or developmental stages:

fetal, normal, mild osteoarthritic, moderate osteoarthritic, marked osteoarthritic and severe osteoarthritic, wherein the change is greater than 1.0-fold, more preferably 1.5-5-fold, and most preferably 5-100-fold, more or less than the level of expression in the absence of a candidate therapeutic agent.

In another embodiment, a therapeutic agent according to the invention can ameliorate at least one of the symptoms and/or changes associated with osteoarthritis including cartilage degeneration, or pain, swelling, weakness and/or loss of functional ability in the afflicted joints, associated with cartilage degeneration.

The candidate therapeutic agent may be a synthetic compound, or a mixture of compounds, or may be a natural product (*e.g.* a plant extract or culture supernatant).

Candidate therapeutic agents or compounds from large libraries of synthetic or natural compounds can be screened. Numerous means are currently used for random and directed synthesis of saccharide, peptide, and nucleic acid-based compounds. Synthetic compound libraries are commercially available from a number of companies including Maybridge Chemical Co. (Trevillet, Cornwall, UK), Comgenex (Princeton, NJ), Brandon Associates (Merrimack, NH), and Microsource (New Milford, CT). A rare chemical library is available from Aldrich (Milwaukee, WI). Combinatorial libraries are available and are prepared. Alternatively, libraries of natural compounds in the form of bacterial, fungal, plant and animal extracts are available from e.g., Pan Laboratories (Bothell, WA) or MycoSearch (NC), or are readily produceable by methods well known in the art. Additionally, natural and synthetically produced libraries and compounds are readily modified through conventional chemical, physical, and biochemical means.

Useful compounds may be found within numerous chemical classes. Useful compounds may be organic compounds, or small organic compounds. Small organic compounds have a molecular weight of more than 50 yet less than about 2,500 daltons, preferably less than about 750, more preferably less than about 350 daltons. Exemplary classes include heterocycles, peptides, saccharides, steroids, and the like. The compounds may be modified to enhance efficacy, stability, pharmaceutical compatibility, and the like. Structural identification of an agent may be used to identify, generate, or screen additional agents. For example, where peptide agents are identified, they may be modified in a variety of ways to enhance their stability, such as using an unnatural

amino acid, such as a D-amino acid, particularly D-alanine, by functionalizing the amino or carboxylic terminus, e.g. for the amino group, acylation or alkylation, and for the carboxyl group, esterification or amidification, or the like.

A therapeutic agent, according to the invention, can be a gene corresponding to an EST sequence identified from any of the cDNA libraries constructed from cartilage of different development and disease stages.

Each cDNA library revealed a number of EST sequences specific to the particular stage. The ESTs are first characterized according to their putative function (Tables 2-6) and their expression is confirmed by using microarrays, as described herein. Since osteoarthritis is a chronic disease caused by an imbalance between catabolic activity and anabolic activity, that is, an increase in catabolic activity and/or a decrease in anabolic activity, normal- or fetal-specific ESTs may be important in maintaining the normal metabolic function of cartilage so as to maintain a balance between the catabolic activity and the anabolic activity. Therefore, an increased expression of a full length gene sequence corresponding to one or more of these ESTs may restore the anabolic activity in disease cartilage. Therapy involving altered gene expression (e.g., gene therapy, gene disruption, antisense therapy, and the like) is useful according to the invention.

A full-length gene sequence corresponding to one of the normal- or fetal-specific genes is cloned by methods known in the art (e.g., Ausubel et al., John Weley & Sons, Inc., 1997, *Current Protocols in Molecular Biology*). A cloned sequence is transfected into disease chondrocytes isolated from any stage of osteoarthritis (e.g., mild, moderate, marked, and severe). The ability of normal- or fetal-specific genes to complement the anabolic defect in the disease chondrocytes is accessed.

In one embodiment, this is achieved by examining the expression profile of disease chondrocytes transfected with a normal- or fetal-specific gene. A normal- or fetal-specific gene which is capable of restoring the expression profile of disease chondrocytes to more closely resemble that of normal or fetal chondrocytes is a useful candidate for treatment of osteoarthritis.

In another embodiment, the anabolic activity of disease chondrocytes transfected with a normal- or fetal- specific gene is measured as described by Westacott et al. (1996, *Semin Arthritis*

Rheum, 25:254-72). A normal- or fetal-specific gene which increases the anabolic activity is useful for treatment of osteoarthritis.

Once a therapeutic gene is defined, the gene sequence is subcloned into a vector suitable for the purpose of gene therapy. Murine leukemia virus (MLV)-based retroviral vectors are one of the most widely used gene delivery vehicles in gene therapy clinical trials and have been employed in almost 70% of approved protocols (Ali, M. et al., 1994, *Gene Ther.*, 1:367-384; Marshall, 1995, *Science*, 269:1050-1055, 1995). Other useful vectors are also known in the art (e.g., Carter and Samulski, 2000, *Int. J. Mol. Med.* 6:17-27; Lever et al., 1999, *Biochem. Soc. Trans.* 27: 841-7). Methods for gene therapy of human diseases are described in U.S. Patent Nos. 6,190,907; 6,187,305; 6,140,087; and 6,129,705, for example, the entireties of which are incorporated by reference herein.

Dosage and Administration

Therapeutic agents of the invention are administered to a patient, preferably in a biologically compatible solution or a pharmaceutically acceptable delivery vehicle, by ingestion, injection, inhalation or any number of other methods routine in the art. The dosages administered will vary from patient to patient. A "therapeutically effective dose" is determined, for example, by the level of enhancement of function (e.g., increased or decreased chondrocyte anabolic activity, or an increase or decrease in the expression of at least one polynucleotide sequence that is differentially expressed in a chondrocyte derived from any of the following chondrocyte disease or developmental stages: fetal, normal, mild osteoarthritic, moderate osteoarthritic, marked osteoarthritic or severe osteoarthritic).

A therapeutic agent according to the invention is administered in a single dose. This dosage may be repeated daily, weekly, monthly, yearly, or as considered appropriate by the treating physician.

Pharmaceutical Compositions

The invention provides for compositions comprising a therapeutic agent according to the invention admixed with a physiologically compatible carrier. As used herein, "physiologically compatible carrier" refers to a physiologically acceptable diluent such as water, phosphate buffered saline, or saline, and further may include an adjuvant. Adjuvants such as incomplete Freund's adjuvant, aluminum phosphate, aluminum hydroxide, or alum are materials well known in the art.

The invention also provides for pharmaceutical compositions. In addition to the active ingredients, these pharmaceutical compositions may contain suitable pharmaceutically acceptable carrier preparations which is used pharmaceutically.

Pharmaceutical compositions for oral administration are formulated using pharmaceutically acceptable carriers well known in the art in dosages suitable for oral administration. Such carriers enable the pharmaceutical compositions to be formulated as tablets, pills, dragees, capsules, liquids, gels, syrups, slurries, suspensions and the like, for ingestion by the patient.

Pharmaceutical preparations for oral use are obtained through a combination of active compounds with solid excipient, optionally grinding a resulting mixture, and processing the mixture of granules, after adding suitable auxiliaries, if desired, to obtain tablets or dragee cores. Suitable excipients are carbohydrate or protein fillers such as sugars, including lactose, sucrose, mannitol, or sorbitol; starch from corn, wheat, rice, potato, or other plants; cellulose such as methyl cellulose, hydroxypropylmethyl-cellulose, or sodium carboxymethyl cellulose; and gums including arabic and tragacanth; and proteins such as gelatin and collagen. If desired, disintegrating or solubilizing agents may be added, such as the cross-linked polyvinyl pyrrolidone, agar, alginic acid, or a salt thereof, such as sodium alginate.

Dragee cores are provided with suitable coatings such as concentrated sugar solutions, which may also contain gum arabic, talc, polyvinylpyrrolidone, carbopol gel, polyethylene glycol, and/or titanium dioxide, lacquer solutions, and suitable organic solvents or solvent mixtures. Dyestuffs or pigments may be added to the tablets or dragee coatings for product identification or to characterize the quantity of active compound, i.e., dosage.

Pharmaceutical preparations which are used orally include push-fit capsules made of gelatin, as well as soft, sealed capsules made of gelatin and a coating such as glycerol or sorbitol. Push-fit capsules can contain active ingredients mixed with a filler or binders such as lactose or starches, lubricants such as talc or magnesium stearate, and, optionally, stabilizers. In soft capsules, the active compounds may be dissolved or suspended in suitable liquids, such as fatty oils, liquid paraffin, or liquid polyethylene glycol with or without stabilizers.

Pharmaceutical formulations for parenteral administration include aqueous solutions of active compounds. For injection, the pharmaceutical compositions of the invention may be formulated in aqueous solutions, preferably in physiologically compatible buffers such as Hank's solution, Ringer' solution, or physiologically buffered saline. Aqueous injection suspensions may contain substances which increase the viscosity of the suspension, such as sodium carboxymethyl cellulose, sorbitol, or dextran. Additionally, suspensions of the active solvents or vehicles include fatty oils such as sesame oil, or synthetic fatty acid esters, such as ethyl oleate or triglycerides, or liposomes. Optionally, the suspension may also contain suitable stabilizers or agents which increase the solubility of the compounds to allow for the preparation of highly concentrated solutions.

For nasal administration, penetrants appropriate to the particular barrier to be permeated are used in the formulation. Such penetrants are generally known in the art.

The pharmaceutical compositions of the present invention may be manufactured in a manner known in the art, e.g. by means of conventional mixing, dissolving, granulating, dragee-making, levitating, emulsifying, encapsulating, entrapping or lyophilizing processes.

The pharmaceutical composition may be provided as a salt and are formed with many acids, including but not limited to hydrochloric, sulfuric, acetic, lactic, tartaric, malic, succinic, etc. Salts tend to be more soluble in aqueous or other protonic solvents that are the corresponding free base forms. In other cases, the preferred preparation may be a lyophilized powder in 1mM-50 mM histidine, 0.1%-2% sucrose, 2%-7% mannitol at a pH range of 4.5 to 5.5 that is combined with buffer prior to use.

After pharmaceutical compositions comprising a therapeutic agent of the invention formulated in a acceptable carrier have been prepared, they are placed in an appropriate container

and labeled for treatment of an indicated condition with information including amount, frequency and method of administration.

Efficacy of Osteoarthritis Therapy Using Defined Therapeutic Agents

The efficacy of the therapy using any of the therapeutic agents according to the invention is determined by a medical practitioner. This determination may be related to alleviating osteoarthritis symptoms such as pain, swelling, weakness and loss of functional ability in the afflicted joint(s), and/or criteria for osteoarthritis diagnosis and staging described in Marshall (1996, supra).

The above disclosure generally describes the present invention. A more complete understanding can be obtained by reference to the following specific examples, which are provided herein for purposes of illustration only and are not intended to limit the scope of the invention.

Examples

The examples below are non-limiting and are merely representative of various aspects and features of the present invention

Example 1: RNA Extraction And Fetal cDNA Library Construction

A cDNA library was prepared from fetal cartilage. ESTs were obtained from the cDNA library and evaluated to create one or more gene expression profiles for fetal chondrocytes.

Human fetal femoral cartilage RNA was extracted from pooled specimens of aborted fetuses (8-12 weeks). Samples were finely powdered under liquid nitrogen, and total RNA was extracted using TRIzol® reagent (GIBCO/BRL). Purity and integrity of RNA was assessed by absorbance at 260/280nm and agarose gel electrophoresis. The poly $(A)^+$ RNA fraction was isolated by oligo-dT cellulose chromatography (Pharmacia), and 3-5 ug poly $(A)^+$ RNA was used to construct a cDNA library in the λ ZAP Express vector (Stratagene). First-strand cDNA was synthesized with an Xho I-oligo (dT) adapter-primer in the presence of 5'-methyl dCTP. After second-strand synthesis and ligation of EcoRI adapters, the cDNA was digested with Xho I, resulting in cDNA flanked by EcoRI sites at the 5'-ends and Xho I sites at the 3'-ends. Digested cDNAs were size-fractionated in Sephacryl S-500 spin columns (Stratagene), then ligated into the λ ZAP Express vector predigested

with EcoRI and Xho I. The resulting DNA/cDNA concatomers were packaged using Gigapack Gold packaging extracts. After titration, aliquots of primary packaging mix were stored in 7% DMSO at -80°C as primary library stocks, and the rest were amplified to establish stable library stocks.

Large-scale sequencing of cDNA inserts

From the amplified λ ZAP Express library, phage plaques were plated at a density of 200-500 pfu/150 mm plate onto *Escherichia coli* XL1-blue MRF' lawn with IPTG/X-gal for color selection. Plaques were picked into 75 ul suspension media buffer (100 mM NaCl, 10 mM $MgSO_4$, 1 mM Tris, pH7.5, 0.02% gelatin). Phage elutes (5 ul) were used for PCR reactions (50 ul total volume) with 125 umol/L of each dNTP (Pharmacia), 10 pmol each of modified T3 (5'-GCCAAGCTCGAAATTAACCCTCACTAAAG GG-3') and T7 (5'-CCAGTGAATTGTAATACGACTCACTATAGGGCG-3') primers, and 2 U of Taq DNA polymerase (Pharmacia). Reactions were cycled in a DNA Thermal Cycler (Perkin-Elmer) [denaturation at 95°C for 5 minutes, followed by 30 cycles of amplification (94°C, 45 seconds; 55° C, 30 seconds; 72°C, 3 minutes) and a terminal isothermal extension (72°C, 3 minutes)]. Agarose gel electrophoresis was used to assess the presence and purity of inserts. PCR products are subjected to DNA sequencing reactions using specific primers, BigDye™ Terminator Cycle Sequencing v2.0 Ready Reaction (PE Biosystems), Tris MgCl buffer and water in a thermocycler. Sequencing reactions were incubated at 94°C for 2 minutes, followed by 25 cycles of 94°C, 30 seconds; 55°C, 20 seconds; and 72°C, 1 minute; and 15 cycles of 94°C, 30 seconds; and 72°C for 1 minute; and 72°C for 5minues. Reactions were then put on hold at 4°C until purified through methods well known in the prior art (i.e. column purification or alcohol precipitation). Automated sequencing was carried out with a PE Biosystems ABI Prism 3700 DNA Analyzer.

Sequences were manually edited or edited using Sequencher software (GeneCodes). All edited EST sequences were compared to the non-redundant Genbank/EMBL/DDBJ and dbEST databases using the BLAST algorithm (8). A minimum value of $P = 10^{-10}$ and nucleotide sequence identity >95% were required for assignments of putative identities for ESTs matching to known genes or to other ESTs. Construction of a non-redundant list of genes represented in the EST set was done with the help of Unigene, Entrez and PubMed at the National Center for Biotechnology Information (NCBI) site (http://www.ncbi.nlm.nih.gov/). Relative gene expression frequency was

calculated by dividing the number of EST copies for each gene by the total number of ESTs analyzed. Functional characterization of ESTs with known gene matches was made according to the categories described by Hwang et al., "A Genome-Based Resource for Molecular Cardiovascular Medicine: Toward a Compendium of Cardiovascular Genes." *Circulation* 1997;96:4146-203).

A total of 13,398 ESTs were obtained from the human fetal cartilage cDNA library. Of these, 5,747 ESTs (41.8%) matched to known gene sequences, 1,855 ESTs (13.4%) matched to other ESTs, and 3,053 (22.0%) matched to mitochondrial, ribosomal, vector and cDNA/hypothetical protein sequences. The 209 ESTs (4.7%) that did not match to any known sequences were designated as novel. The remainder matched to genomic DNA sequences (1,948 ESTs, 13.8%) and repetitive sequences (586 ESTs, 4.3%).

The 13,398 EST sequences in the fetal library were characterized based on the functional classification of the 2,579 unique known genes they represented. The following table sets out the results of this analysis.

Table 2. Fetal Cartilage Library

Putative Function	Percent (%) of Representation in the Library	Number of genes
Cell division	7.06	182
Cell signaling/communication	15.01	387
Cell structure/motility	10.90	281
Cell/organism defense	7.60	196
Gene/protein expression	22.22	573
Metabolism	14.89	384
Unclassified	22.33	576
Total known/unique genes analyzed	100.00	2,579

Example 2: RNA Extraction And Normal Adult cDNA Library Construction

A cDNA library was prepared from normal adult cartilage. ESTs were obtained from the cDNA library and characterized to create one or more gene expression profiles for normal adult chondrocytes.

Large-scale sequencing of cDNA inserts

cDNA libraries were constructed into λTripleEx2 vector through a PCR-based method, using SMART (Switching Mechanism At 5' end of RNA Transcript) cDNA Library Construction Kit (Clontech). Phage plaques were randomly picked and positive inserts were identified by PCR. Agarose gel electrophoresis was used to assess the presence and purity of inserts. PCR product was then subjected to automated DNA sequencing with a 5' vector-specific forward primer and sequenced by ABI PRISM 377 DNA sequencer (Perkin Elmer) and ABI PRISM 3700 DNA Analyzer (Applied Biosystems). All generated EST sequences were searched against the nonredundant Genebank/EMBL/DDBL, dbEST and GSS databases. A minimum value of $p=10^{-10}$ and nucleotide sequence identity >90% were required for assignments of putative identities for EST-matching to known genes or other ESTs. Relative EST frequency level was calculated by dividing the EST number matched to that gene into the total number of ESTs obtained from the library.

Sequences were manually edited or edited using Sequencher software (GeneCodes). All edited EST sequences were compared to the non-redundant Genbank/EMBL/DDBJ and dbEST databases using the BLAST algorithm (8). A minimum value of $P=10^{-10}$ and nucleotide sequence identity >95% were required for assignments of putative identities for ESTs matching to known genes or to other ESTs. Construction of a non-redundant list of genes represented in the EST set was done with the help of Unigene, Entrez and PubMed at the National Center for Biotechnology Information (NCBI) site (http://www.ncbi.nlm.nih.gov/). Relative gene expression frequency was calculated by dividing the number of EST copies for each gene by the total number of ESTs analyzed. Functional characterization of ESTs with known gene matches was made according to the categories described by Hwang et al (Hwang DM, Dempsey AA, Wang RX, Rezvani M, Barrans JD, Dai KS, et al. A Genome-Based Resource for Molecular Cardiovascular Medicine: Toward a Compendium of Cardiovascular Genes. Circulation 1997;96:4146-203).

A total of 17,151 ESTs were obtained from the normal cartilage cDNA library. Of these, 6,755 ESTs (44.2%) matched to 2,518 known genes. 1.4% (132 ESTs) showed no significant match and were thus designated as novel. Characterization of the 17,151 EST sequences based on functional classification of known/unique genes resulted in the following table:

Table 3. Normal Adult Cartilage Library		
Putative function	Percent (%) of representation in the library	Number of genes
Cell division	6.13	160
Cell signaling/communication	13.52	353
Cell structure/motility	9.00	235
Cell/organism defense	7.51	196
Gene/protein expression	20.08	524
Metabolism	13.14	343
Unclassified	27.09	707
Total known/unique genes analyzed	100.00	2518

Example 3: RNA Extraction and cDNA Library Construction From Mild Osteoarthritic Chondrocytes and Severe Osteoarthritic Chondrocytes

A cDNA library was prepared from mild osteoarthritic cartilage and severe osteoarthritic cartilage. ESTs were obtained from the cDNA libraries and characterized to create one or more gene expression profiles for mild osteoarthritic chondrocytes and severe osteoarthritic chondrocytes.

Articular cartilage was obtained during either arthroscopic knee surgery or total knee replacement. The cartilage samples were obtained from either areas of very early cartilage degeneration (mild) or from sites of end stage disease (severe). cDNA libraries were constructed as described for normal adult samples (Example 2).

Large-scale sequencing of cDNA inserts

cDNA libraries were constructed into λTripleEx2 vector through a PCR-based method, using SMART (Switching Mechanism At 5' end of RNA Transcript) cDNA Library Construction Kit (Clontech). Phage plaques were randomly picked and positive inserts were identified by PCR. Agarose gel electrophoresis was used to assess the presence and purity of inserts. PCR product was then subjected to automated DNA sequencing with a 5' vector-specific forward primer and sequenced by ABI PRISM 377 DNA sequencer (Perkin Elmer) and ABI PRISM 3700 DNA Analyzer (Applied Biosystems). All generated EST sequences were searched against the

nonredundant Genebank/EMBL/DDBL, dbEST and GSS databases. A minimum value of $p=10^{-10}$ and nucleotide sequence identity >90% were required for assignments of putative identities for EST-matching to known genes or other ESTs. Relative EST frequency level was calculated by dividing the EST number matched to that gene into the total number of ESTs obtained from the library.

Sequences were manually edited or edited using Sequencher software (GeneCodes). All edited EST sequences were compared to the non-redundant Genbank/EMBL/DDBJ and dbEST databases using the BLAST algorithm (8). A minimum value of $P=10^{-10}$ and nucleotide sequence identity >95% were required for assignments of putative identities for ESTs matching to known genes or to other ESTs.

Construction of a non-redundant list of genes represented in the EST set was done with the help of Unigene, Entrez and PubMed at the National Center for Biotechnology Information (NCBI) site (http://www.ncbi.nlm.nih.gov/). Relative gene expression frequency was calculated by dividing the number of EST copies for each gene by the total number of ESTs analyzed.

Functional characterization of ESTs with known gene matches was made according to the categories described by Hwang et al (Hwang DM, Dempsey AA, Wang RX, Rezvani M, Barrans JD, Dai KS, et al. A Genome-Based Resource for Molecular Cardiovascular Medicine: Toward a Compendium of Cardiovascular Genes. Circulation 1997;96:4146-203).

A total of 12,651 ESTs and 14,222 ESTs were obtained from the mild and severe OA cDNA libraries (Table 5 and Table 6), respectively. About 43% of mild and 51% of severe OA ESTs matched to known genes in the database. Approximately 2.6% and 1.5% of the ESTs, respectively, resulted in no significant match and were thus designated as novel. The sequences for each of the ESTs is disclosed in the CD ROM described as Figure 14 wherein each sequence is prescribed a SEQ ID No. The following table provides a correlation between the SEQ ID NO in Figure 14 and the clone ID name for these sequences as disclosed in other Figures herein.

SEQ IDNo.	Clone ID	SEQ IDNo.	Clone ID	SEQ IDNo.	Clone ID	SEQ IDNo.	Clone ID
1	BFCN0001	55	BFCN0109	109	BFCN0207	163	BFCS0003
2	BFCN0002	56	BFCN0112	110	BFCN0208	164	BFCS0005
3	BFCN0003	57	BFCN0113	111	BFCN0209	165	BFCS0006
4	BFCN0005	58	BFCN0114	112	BFCN0210	166	BFCS0007
5	BFCN0006	59	BFCN0115	113	BFCN0211	167	BFCS0008
6	BFCN0007	60	BFCN0116	114	BFCN0213	168	BFCS0009
7	BFCN0008	61	BFCN0117	115	BFCN0214	169	BFCS0014
8	BFCN0009	62	BFCN0118	116	BFCN0215	170	BFCS0021
9	BFCN0010	63	BFCN0119	117	BFCN0216	171	BFCS0022
10	BFCN0012	64	BFCN0120	118	BFCN0217	172	BFCS0024
11	BFCN0013	65	BFCN0124	119	BFCN0219	173	BFCS0027
12	BFCN0018	66	BFCN0127	120	BFCN0220	174	BFCS0034
13	BFCN0019	67	BFCN0128	121	BFCN0222	175	BFCS0035
14	BFCN0021	68	BFCN0130	122	BFCN0224	176	BFCS0037
15	BFCN0024	69	BFCN0133	123	BFCN0225	177	BFCS0038
16	BFCN0027	70	BFCN0134	124	BFCN0226	178	BFCS0039
17	BFCN0029	71	BFCN0135	125	BFCN0227	179	BFCS0041
18	BFCN0031	72	BFCN0136	126	BFCN0228	180	BFCS0042
19	BFCN0034	73	BFCN0138	127	BFCN0229	181	BFCS0043
20	BFCN0038	74	BFCN0139	128	BFCN0232	182	BFCS0045
21	BFCN0039	75	BFCN0140	129	BFCN0233	183	BFCS0047
22	BFCN0040	76	BFCN0142	130	BFCN0235	184	BFCS0048
23	BFCN0042	77	BFCN0156	131	BFCN0236	185	BFCS0049
24	BFCN0045	78	BFCN0164	132	BFCN0238	186	BFCS0050
25	BFCN0047	79	BFCN0168	133	BFCN0239	187	BFCS0054
26	BFCN0048	80	BFCN0171	134	BFCN0245	188	BFCS0055
27	BFCN0049	81	BFCN0172	135	BFCN0246	189	BFCS0057
28	BFCN0050	82	BFCN0173	136	BFCN0247	190	BFCS0058
29	BFCN0051	83	BFCN0177	137	BFCN0248	191	BFCS0074
30	BFCN0053	84	BFCN0178	138	BFCN0249	192	BFCS0077
31	BFCN0055	85	BFCN0179	139	BFCN0250	193	BFCS0079
32	BFCN0056	86	BFCN0180	140	BFCN0251	194	BFCS0081
33	BFCN0059	87	BFCN0181	141	BFCN0252	195	BFCS0082
34	BFCN0060	88	BFCN0182	142	BFCN0253	196	BFCS0083
35	BFCN0062	89	BFCN0185	143	BFCN0254	197	BFCS0088
36	BFCN0065	90	BFCN0186	144	BFCN0255	198	BFCS0089
37	BFCN0067	91	BFCN0190	145	BFCN0256	199	BFCS0092
38	BFCN0072	92	BFCN0192	146	BFCN0259	200	BFCS0093
39	BFCN0073	93	BFCN0194	147	BFCN0261	201	BFCS0094
40	BFCN0075	94	BFCN0195	148	BFCN0265	202	Bfcs0037n
41	BFCN0079	95	BFCN0196	149	BFCN0266	203	Bfcs0039n
42	BFCN0081	96	BFCN0197	150	BFCN0267	204	Bfcs0047n
43	BFCN0082	97	BFCN0198	151	BFCN0268	205	Bfcs0048n
44	BFCN0083	98	BFCN0199	152	BFCN0270	206	BFCS0195
45	BFCN0085	99	Bfcn0168n	153	BFCN0271	207	BFCS0196
46	BFCN0090	100	Bfcn0182n	154	BFCN0272	208	BFCS0198
47	BFCN0092	101	Bfcn0185n	155	BFCN0273	209	BFCS0199
48	BFCN0093	102	Bfcn0190n	156	BFCN0274	210	Bfcs0195n
49	BFCN0094	103	BFCN0200	157	Bfcn0202n	211	BFCS0202
50	BFCN0096	104	BFCN0202	158	Bfcn0206n	212	BFCS0203
51	BFCN0097	105	BFCN0203	159	Bfcn0215n	213	BFCS0205
52	BFCN0098	106	BFCN0204	160	Bfcn0224n	214	BFCS0206
53	Bfcn0056n	107	BFCN0205	161	Bfcn0198n	215	BFCS0207
54	BFCN0105	108	BFCN0206	162	BFCS0001	216	BFCS0208

217	BFCS0212	271	BFCS0320	325	BFCS0491	379	Bfcw0253n
218	BFCS0214	272	BFCS0321	326	BFCS0492	380	Bfcw0064n
219	BFCS0216	273	BFCS0322	327	BFCS0493	381	Bfcw0060n
220	BFCS0219	274	BFCS0324	328	BFCS0494	382	Bfcw0572n
221	BFCS0220	275	BFCS0326	329	BFCS0495	383	Bfcw0183n
222	BFCS0223	276	BFCS0330	330	BFCS0496	384	Bfcw0482n
223	BFCS0228	277	BFCS0331	331	BFCS0498	385	Bfcw0292n
224	BFCS0229	278	BFCS0332	332	Bfcs0421n	386	Bfcw0191n
225	BFCS0231	279	BFCS0335	333	Bfcs0468n	387	Bfcw0340n
226	BFCS0232	280	BFCS0336	334	Bfcs0469n	388	BFCW0008
227	BFCS0233	281	BFCS0337	335	BFCS0500	389	BFCW0009
228	BFCS0238	282	BFCS0338	336	BFCS0501	390	BFCW0010
229	BFCS0239	283	BFCS0342	337	BFCS0502	391	BFCW0014
230	BFCS0241	284	BFCS0343	338	BFCS0503	392	BFCW0019
231	BFCS0244	285	BFCS0345	339	BFCS0504	393	BFCW0020
232	BFCS0246	286	BFCS0346	340	BFCS0508	394	BFCW0023
233	BFCS0257	287	BFCS0347	341	BFCS0509	395	BFCW0024
234	BFCS0259	288	BFCS0368	342	BFCS0513	396	BFCW0026
235	BFCS0260	289	BFCS0369	343	BFCS0516	397	BFCW0035
236	BFCS0261	290	BFCS0371	344	BFCS0518	398	BFCW0036
237	BFCS0264	291	BFCS0377	345	BFCS0519	399	BFCW0038
238	BFCS0265	292	BFCS0379	346	BFCS0520	400	BFCW0054
239	BFCS0266	293	BFCS0384	347	BFCS0522	401	BFCW0055
240	BFCS0269	294	BFCS0389	348	BFCS0523	402	BFCW0056
241	BFCS0270	295	BFCS0390	349	BFCS0524	403	BFCW0060
242	BFCS0276	296	BFCS0391	350	BFCS0526	404	BFCW0062
243	BFCS0277	297	BFCS0392	351	BFCS0527	405	BFCW0064
244	BFCS0280	298	BFCS0393	352	BFCS0531	406	BFCW0065
245	BFCS0281	299	BFCS0396	353	BFCS0532	407	BFCW0067
246	BFCS0283	300	BFCS0398	354	BFCS0533	408	BFCW0069
247	BFCS0284	301	BFCS0399	355	BFCS0535	409	BFCW0071
248	BFCS0285	302	Bfcs0309n	356	BFCS0538	410	BFCW0072
249	BFCS0286	303	Bfcs0312n	357	BFCS0539	411	BFCW0073
250	BFCS0289	304	Bfcs0315n	358	BFCS0541	412	BFCW0074
251	BFCS0292	305	Bfcs0346n	359	BFCS0544	413	BFCW0076
252	BFCS0296	306	BFCS0404	360	BFCS0545	414	BFCW0078
253	BFCS0297	307	BFCS0407	361	BFCS0547	415	BFCW0079
254	BFCS0299	308	BFCS0408	362	BFCS0548	416	BFCW0081
255	Bfcs0206n	309	BFCS0417	363	BFCS0549	417	BFCW0083
256	Bfcs0207n	310	BFCS0420	364	BFCS0552	418	BFCW0085
257	Bfcs0208n	311	BFCS0421	365	BFCS0553	419	BFCW0088
258	Bfcs0239n	312	BFCS0457	366	BFCS0557	420	BFCW0090
259	BFCS0300	313	BFCS0462	367	BFCS0559	421	BFCW0092
260	BFCS0302	314	BFCS0463	368	BFCS0560	422	BFCW0093
261	BFCS0303	315	BFCS0468	369	BFCS0563	423	BFCW0094
262	BFCS0309	316	BFCS0469	370	Bfcs0518n	424	Bfcw0019n
263	BFCS0311	317	BFCS0478	371	Bfcs0520n	425	Bfcw0026n
264	BFCS0312	318	BFCS0479	372	Bfcs0545n	426	Bfcw0036n
265	BFCS0313	319	BFCS0481	373	Bfcs0553n	427	bfcw0043n
266	BFCS0314	320	BFCS0483	374	Bfcs0057n	428	BFCW0100
267	BFCS0315	321	BFCS0484	375	Bfcs0269n	429	BFCW0102
268	BFCS0316	322	BFCS0485	376	Bfcs0088n	430	BFCW0103
269	BFCS0317	323	BFCS0487	377	Bfcs0347n	431	BFCW0108
270	BFCS0319	324	BFCS0489	378	Bfcw0254n	432	BFCW0109

433	BFCW0111	487	BFCW0218	541	BFCW0312	595	BFCW0401
434	BFCW0112	488	BFCW0219	542	BFCW0313	596	BFCW0402
435	BFCW0114	489	BFCW0220	543	BFCW0314	597	BFCW0403
436	BFCW0115	490	BFCW0223	544	BFCW0316	598	BFCW0404
437	BFCW0118	491	BFCW0224	545	BFCW0317	599	BFCW0406
438	BFCW0132	492	BFCW0225	546	BFCW0318	600	BFCW0407
439	BFCW0133	493	BFCW0226	547	BFCW0319	601	BFCW0408
440	BFCW0134	494	BFCW0228	548	BFCW0320	602	BFCW0409
441	BFCW0137	495	BFCW0230	549	BFCW0323	603	BFCW0412
442	BFCW0139	496	BFCW0231	550	BFCW0325	604	BFCW0413
443	BFCW0140	497	BFCW0235	551	BFCW0326	605	BFCW0414
444	BFCW0144	498	BFCW0236	552	BFCW0327	606	BFCW0415
445	BFCW0145	499	BFCW0238	553	BFCW0329	607	BFCW0416
446	BFCW0146	500	BFCW0239	554	BFCW0330	608	BFCW0420
447	BFCW0147	501	BFCW0240	555	BFCW0331	609	BFCW0421
448	BFCW0148	502	BFCW0241	556	BFCW0332	610	BFCW0422
449	BFCW0150	503	BFCW0244	557	BFCW0333	611	BFCW0423
450	BFCW0151	504	BFCW0245	558	BFCW0334	612	BFCW0424
451	BFCW0154	505	BFCW0246	559	BFCW0335	613	BFCW0425
452	BFCW0159	506	BFCW0248	560	BFCW0336	614	BFCW0426
453	BFCW0160	507	BFCW0250	561	BFCW0337	615	BFCW0429
454	BFCW0162	508	BFCW0251	562	BFCW0339	616	BFCW0430
455	BFCW0166	509	BFCW0252	563	BFCW0340	617	BFCW0431
456	BFCW0169	510	BFCW0253	564	BFCW0341	618	BFCW0432
457	BFCW0170	511	BFCW0254	565	BFCW0345	619	BFCW0433
458	BFCW0172	512	BFCW0255	566	BFCW0348	620	BFCW0435
459	BFCW0176	513	BFCW0256	567	BFCW0352	621	BFCW0436
460	BFCW0177	514	BFCW0258	568	BFCW0369	622	BFCW0438
461	BFCW0179	515	BFCW0261	569	BFCW0370	623	BFCW0440
462	BFCW0180	516	BFCW0266	570	BFCW0371	624	BFCW0445
463	BFCW0183	517	BFCW0268	571	BFCW0372	625	BFCW0457
464	BFCW0184	518	BFCW0275	572	BFCW0373	626	BFCW0458
465	BFCW0186	519	BFCW0276	573	BFCW0375	627	BFCW0459
466	BFCW0188	520	BFCW0277	574	BFCW0378	628	BFCW0460
467	BFCW0189	521	BFCW0280	575	BFCW0379	629	BFCW0461
468	BFCW0191	522	BFCW0281	576	BFCW0380	630	BFCW0462
469	BFCW0192	523	BFCW0282	577	BFCW0382	631	BFCW0464
470	BFCW0194	524	BFCW0286	578	BFCW0384	632	BFCW0467
471	BFCW0197	525	BFCW0287	579	BFCW0386	633	BFCW0469
472	BFCW0198	526	BFCW0288	580	BFCW0388	634	BFCW0472
473	Bfcw0100n	527	BFCW0289	581	BFCW0389	635	BFCW0476
474	Bfcw0102n	528	BFCW0291	582	BFCW0390	636	BFCW0478
475	Bfcw0109n	529	BFCW0292	583	BFCW0391	637	BFCW0479
476	Bfcw0139n	530	BFCW0293	584	BFCW0393	638	BFCW0480
477	BFCW0200	531	BFCW0294	585	BFCW0394	639	BFCW0481
478	BFCW0202	532	BFCW0296	586	BFCW0395	640	BFCW0482
479	BFCW0206	533	Bfcw0202n	587	BFCW0396	641	BFCW0483
480	BFCW0207	534	Bfcw0206n	588	BFCW0397	642	BFCW0485
481	BFCW0209	535	Bfcw0207n	589	BFCW0398	643	BFCW0487
482	BFCW0210	536	Bfcw0209n	590	Bfcw0314n	644	BFCW0488
483	BFCW0212	537	BFCW0304	591	Bfcw0330n	645	BFCW0489
484	BFCW0215	538	BFCW0307	592	Bfcw0334n	646	BFCW0490
485	BFCW0216	539	BFCW0310	593	Bfcw0335n	647	BFCW0492
486	BFCW0217	540	BFCW0311	594	BFCW0400	648	BFCW0493

649	Bfcw0407n	703	BFCW0598	757	CR0027	811	CR0118
650	Bfcw0413n	704	Bfcw0599	758	CR0027X	812	CR0119
651	Bfcw0430n	705	Bfcw0540n	759	CR0028	813	CR0120
652	Bfcw0458n	706	Bfcw0542n	760	CR0029	814	CR0121
653	BFCW0500	707	Bfcw0551n	761	CR0030	815	CR0124
654	BFCW0506	708	Bfcw0567n	762	CR0033	816	CR0125
655	BFCW0510	709	Bfcw0601	763	CR0038	817	CR0128
656	BFCW0511	710	Bfcw0604	764	CR0039	818	CR0131
657	BFCW0513	711	Bfcw0605	765	CR0039X	819	CR0133
658	BFCW0515	712	Bfcw0607	766	CR0040	820	CR0135
659	BFCW0516	713	Bfcw0608	767	CR0040X	821	CR0136
660	BFCW0517	714	Bfcw0609	768	CR0042	822	CR0138
661	BFCW0518	715	Bfcw0610	769	CR0042X	823	CR0140
662	BFCW0519	716	Bfcw0056n	770	CR0043	824	CR0143
663	BFCW0520	717	Bfcw0569n	771	CR0044	825	CR0144
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2620	FCR2018	2674	FCR2097	2728	FCR2160	2782	FCR2240
2621	FCR2019	2675	FCR2097N	2729	FCR2161	2783	FCR2241
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2628	FCR2032	2682	FCR2108	2736	FCR2174	2790	FCR2249N
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9294	hfcr2064	9348	hfcr2251	9402	hfcr2389	9456	hfcr2555
9295	hfcr2065	9349	hfcr2252	9403	hfcr2390	9457	hfcr2556
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9300	hfcr2071	9354	hfcr2263	9408	hfcr2499	9462	hfcr2563
9301	hfcr2073	9355	hfcr2264	9409	hfcr2501	9463	hfcr2565
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9305	hfcr2077	9359	hfcr2275	9413	hfcr2505	9467	hfcr2570
9306	hfcr2078	9360	hfcr2282	9414	hfcr2506	9468	hfcr2572
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9308	hfcr2080	9362	hfcr2287	9416	hfcr2509	9470	hfcr2574
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9310	hfcr2082	9364	hfcr2294	9418	hfcr2511	9472	hfcr2576
9311	hfcr2084	9365	hfcr2295	9419	hfcr2512	9473	hfcr2578
9312	hfcr2114	9366	hfcr2296	9420	hfcr2513	9474	hfcr2580
9313	hfcr2128	9367	hfcr2297	9421	hfcr2514	9475	hfcr2581
9314	hfcr2129	9368	hfcr2299	9422	hfcr2515	9476	hfcr2582
9315	hfcr2131	9369	hfcr2301	9423	hfcr2516	9477	hfcr2583
9316	hfcr2138	9370	hfcr2306	9424	hfcr2517	9478	hfcr2584
9317	hfcr2140	9371	hfcr2310	9425	hfcr2519	9479	hfcr2586
9318	hfcr2141	9372	hfcr2312	9426	hfcr2520	9480	hfcr2587
9319	hfcr2148	9373	hfcr2313	9427	hfcr2521	9481	hfcr2588
9320	hfcr2150	9374	hfcr2314	9428	hfcr2522	9482	hfcr2589
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9548	hfcr2682	9602	hfcr2754	9656	hfcr2825	9710	hfcr2899
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9552	hfcr2687	9606	hfcr2758	9660	hfcr2832	9714	hfcr2907
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9555	hfcr2690	9609	hfcr2761	9663	hfcr2836	9717	hfcr2910
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9557	hfcr2695	9611	hfcr2766	9665	hfcr2838	9719	hfcr2912
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10155	hfcr3572	10209	hfcr3653	10263	hfcr3734	10317	hfcr3799
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10185	hfcr3614	10239	hfcr3700	10293	hfcr3766	10347	hfcr3847
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10198	hfcr3634	10252	hfcr3721	10306	hfcr3781	10360	hfcr3867
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10374	hfcr3882	10428	hfcr3944	10482	hfcr4028	10536	hfcr4115
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10383	hfcr3892	10437	hfcr3960	10491	hfcr4046	10545	hfcr4124
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31796	ncr7763	31850	ncr7840	31904	ncr7922	31958	ncr7999
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31802	ncr7770	31856	ncr7848	31910	ncr7929	31964	ncr8012
31803	ncr7771	31857	ncr7849	31911	ncr7931	31965	ncr8013
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31805	ncr7773	31859	ncr7852	31913	ncr7933	31967	ncr8017
31806	ncr7774	31860	ncr7853	31914	ncr7934	31968	ncr8018

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31971	ncr8024	32025	ncr8100	32079	ncr8174	32133	ncr8250
31972	ncr8025	32026	ncr8101	32080	ncr8175	32134	ncr8251
31973	ncr8026	32027	ncr8103	32081	ncr8176	32135	ncr8252
31974	ncr8027	32028	ncr8107	32082	ncr8177	32136	ncr8253
31975	ncr8030	32029	ncr8108	32083	ncr8180	32137	ncr8254
31976	ncr8031	32030	ncr8109	32084	ncr8181	32138	ncr8256
31977	ncr8032	32031	ncr8110	32085	ncr8182	32139	ncr8259
31978	ncr8033	32032	ncr8111	32086	ncr8183	32140	ncr8260
31979	ncr8034	32033	ncr8112	32087	ncr8184	32141	ncr8261
31980	ncr8035	32034	ncr8113	32088	ncr8186	32142	ncr8263
31981	ncr8036	32035	ncr8114	32089	ncr8187	32143	ncr8267
31982	ncr8038	32036	ncr8115	32090	ncr8188	32144	ncr8268
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32014	ncr8083	32068	ncr8156	32122	ncr8236	32176	ncr8316
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32017	ncr8089	32071	ncr8160	32125	ncr8241	32179	ncr8320
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32216	ncr8381	32270	ncr8468	32324	ncr8544	32378	ncr8637
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58155	seob7741	58209	seob7901	58263	seob7969	58317	seob8034
58156	seob7742	58210	seob7902	58264	seob7970	58318	seob8035
58157	seob7743	58211	seob7903	58265	seob7972	58319	seob8037
58158	seob7744	58212	seob7905	58266	seob7973	58320	seob8039
58159	seob7745	58213	seob7906	58267	seob7974	58321	seob8040
58160	seob7746	58214	seob7907	58268	seob7975	58322	seob8041
58161	seob7747	58215	seob7908	58269	seob7977	58323	seob8042
58162	seob7748	58216	seob7909	58270	seob7978	58324	seob8044
58163	seob7749	58217	seob7910	58271	seob7979	58325	seob8045
58164	seob7750	58218	seob7911	58272	seob7980	58326	seob8046
58165	seob7751	58219	seob7912	58273	seob7981	58327	seob8047
58166	seob7752	58220	seob7915	58274	seob7982	58328	seob8048
58167	seob7753	58221	seob7916	58275	seob7983	58329	seob8051
58168	seob7754	58222	seob7917	58276	seob7984	58330	seob8052
58169	seob7755	58223	seob7918	58277	seob7986	58331	seob8053
58170	seob7756	58224	seob7919	58278	seob7987	58332	seob8054
58171	seob7757	58225	seob7920	58279	seob7989	58333	seob8055
58172	seob7758	58226	seob7921	58280	seob7990	58334	seob8060
58173	seob7759	58227	seob7923	58281	seob7992	58335	seob8063
58174	seob7760	58228	seob7924	58282	seob7993	58336	seob8065
58175	seob7763	58229	seob7926	58283	seob7994	58337	seob8066
58176	seob7764	58230	seob7928	58284	seob7995	58338	seob8067
58177	seob7765	58231	seob7929	58285	seob7996	58339	seob8068
58178	seob7766	58232	seob7930	58286	seob7998	58340	seob8069
58179	seob7769	58233	seob7931	58287	seob7999	58341	seob8071
58180	seob7866	58234	seob7933	58288	seob8000	58342	seob8072
58181	seob7868	58235	seob7934	58289	seob8001	58343	seob8073
58182	seob7869	58236	seob7935	58290	seob8002	58344	seob8076
58183	seob7870	58237	seob7936	58291	seob8004	58345	seob8077
58184	seob7871	58238	seob7937	58292	seob8006	58346	seob8078
58185	seob7872	58239	seob7938	58293	seob8007	58347	seob8079
58186	seob7873	58240	seob7939	58294	seob8008	58348	seob8080
58187	seob7874	58241	seob7940	58295	seob8009	58349	seob8081
58188	seob7875	58242	seob7941	58296	seob8010	58350	seob8082
58189	seob7876	58243	seob7942	58297	seob8011	58351	seob8083
58190	seob7877	58244	seob7944	58298	seob8012	58352	seob8084
58191	seob7878	58245	seob7945	58299	seob8013	58353	seob8085
58192	seob7879	58246	seob7946	58300	seob8015	58354	seob8086
58193	seob7880	58247	seob7947	58301	seob8016	58355	seob8087

58356	seob8088	58410	seob8188	58464	seob8256	58518	seob8323
58357	seob8089	58411	seob8189	58465	seob8257	58519	seob8324
58358	seob8090	58412	seob8190	58466	seob8258	58520	seob8326
58359	seob8092	58413	seob8191	58467	seob8260	58521	seob8328
58360	seob8093	58414	seob8192	58468	seob8261	58522	seob8329
58361	seob8094	58415	seob8193	58469	seob8262	58523	seob8330
58362	seob8095	58416	seob8194	58470	seob8263	58524	seob8332
58363	seob8096	58417	seob8196	58471	seob8264	58525	seob8333
58364	seob8097	58418	seob8198	58472	seob8265	58526	seob8334
58365	seob8099	58419	seob8200	58473	seob8266	58527	seob8335
58366	seob8100	58420	seob8202	58474	seob8267	58528	seob8336
58367	seob8101	58421	seob8204	58475	seob8268	58529	seob8337
58368	seob8102	58422	seob8205	58476	seob8269	58530	seob8338
58369	seob8104	58423	seob8207	58477	seob8271	58531	seob8339
58370	seob8106	58424	seob8208	58478	seob8275	58532	seob8341
58371	seob8107	58425	seob8209	58479	seob8276	58533	seob8343
58372	seob8108	58426	seob8210	58480	seob8277	58534	seob8344
58373	seob8110	58427	seob8211	58481	seob8278	58535	seob8345
58374	seob8129	58428	seob8212	58482	seob8279	58536	SOA0001
58375	seob8130	58429	seob8214	58483	seob8280	58537	soa0001n
58376	seob8132	58430	seob8215	58484	seob8281	58538	SOA0002
58377	seob8135	58431	seob8216	58485	seob8282	58539	SOA0004
58378	seob8138	58432	seob8217	58486	seob8284	58540	soa0004n
58379	seob8140	58433	seob8219	58487	seob8285	58541	SOA0005
58380	seob8141	58434	seob8220	58488	seob8286	58542	soa0005n
58381	seob8154	58435	seob8221	58489	seob8287	58543	SOA0006
58382	seob8155	58436	seob8223	58490	seob8288	58544	soa0006n
58383	seob8157	58437	seob8224	58491	seob8289	58545	SOA0007
58384	seob8158	58438	seob8225	58492	seob8291	58546	soa0007n
58385	seob8159	58439	seob8226	58493	seob8292	58547	SOA0008
58386	seob8160	58440	seob8227	58494	seob8293	58548	SOA0012
58387	seob8161	58441	seob8229	58495	seob8294	58549	soa0012n
58388	seob8162	58442	seob8231	58496	seob8296	58550	SOA0017
58389	seob8163	58443	seob8232	58497	seob8297	58551	SOA0021
58390	seob8164	58444	seob8233	58498	seob8298	58552	SOA0022
58391	seob8166	58445	seob8235	58499	seob8299	58553	soa0022n
58392	seob8167	58446	seob8236	58500	seob8300	58554	SOA0024
58393	seob8168	58447	seob8237	58501	seob8301	58555	SOA0026
58394	seob8169	58448	seob8238	58502	seob8303	58556	SOA0027
58395	seob8171	58449	seob8239	58503	seob8305	58557	SOA0028
58396	seob8172	58450	seob8240	58504	seob8306	58558	soa0028n
58397	seob8173	58451	seob8241	58505	seob8308	58559	SOA0031
58398	seob8174	58452	seob8242	58506	seob8309	58560	SOA0033
58399	seob8176	58453	seob8243	58507	seob8310	58561	SOA0035
58400	seob8177	58454	seob8244	58508	seob8311	58562	SOA0038
58401	seob8178	58455	seob8245	58509	seob8312	58563	soa0038n
58402	seob8179	58456	seob8246	58510	seob8313	58564	SOA0039
58403	seob8180	58457	seob8247	58511	seob8314	58565	soa0039n
58404	seob8181	58458	seob8248	58512	seob8315	58566	SOA0040
58405	seob8182	58459	seob8249	58513	seob8317	58567	soa0040n
58406	seob8184	58460	seob8250	58514	seob8319	58568	SOA0042
58407	seob8185	58461	seob8252	58515	seob8320	58569	soa0042n
58408	seob8186	58462	seob8254	58516	seob8321	58570	SOA0043
58409	seob8187	58463	seob8255	58517	seob8322	58571	soa0043n

58572	SOA0044	58626	SOA0143	58680	SOA0241	58734	SOA0327
58573	SOA0046	58627	SOA0145	58681	SOA0242	58735	SOA0328
58574	SOA0047	58628	SOA0146	58682	SOA0245	58736	SOA0329
58575	SOA0049	58629	soa0146n	58683	soa0245n	58737	soa0329n
58576	soa0049n	58630	SOA0147	58684	SOA0248	58738	SOA0330
58577	SOA0050	58631	SOA0148	58685	SOA0249	58739	SOA0331
58578	SOA0053	58632	SOA0149	58686	SOA0251	58740	SOA0332
58579	soa0053n	58633	SOA0154	58687	SOA0253	58741	SOA0334
58580	SOA0055	58634	SOA0156	58688	SOA0256	58742	SOA0335
58581	SOA0056	58635	SOA0158	58689	SOA0257	58743	SOA0337
58582	SOA0058	58636	SOA0161	58690	SOA0262	58744	SOA0338
58583	SOA0059	58637	SOA0163	58691	SOA0263	58745	SOA0340
58584	SOA0060	58638	SOA0165	58692	SOA0264	58746	SOA0341
58585	SOA0064	58639	SOA0195	58693	SOA0267	58747	SOA0342
58586	SOA0065	58640	SOA0196	58694	SOA0269	58748	SOA0343
58587	SOA0068	58641	soa0196n	58695	SOA0271	58749	soa0343n
58588	SOA0070	58642	SOA0197	58696	soa0271n	58750	SOA0345
58589	SOA0071	58643	soa0197n	58697	SOA0274	58751	soa0345n
58590	SOA0076	58644	SOA0198	58698	SOA0275	58752	SOA0346
58591	SOA0077	58645	soa0198n	58699	SOA0277	58753	soa0346n
58592	SOA0078	58646	SOA0201	58700	soa0277n	58754	SOA0347
58593	soa0078n	58647	soa0201n	58701	SOA0278	58755	SOA0348
58594	SOA0079	58648	SOA0204	58702	SOA0281	58756	SOA0349
58595	SOA0082	58649	soa0204n	58703	SOA0282	58757	SOA0351
58596	SOA0083	58650	SOA0207	58704	SOA0283	58758	SOA0353
58597	SOA0085	58651	SOA0208	58705	SOA0284	58759	SOA0354
58598	SOA0089	58652	SOA0209	58706	SOA0285	58760	SOA0356
58599	SOA0092	58653	SOA0210	58707	SOA0286	58761	SOA0357
58600	SOA0093	58654	SOA0212	58708	SOA0288	58762	SOA0360
58601	soa0093n	58655	SOA0213	58709	SOA0289	58763	soa0360n
58602	SOA0095	58656	SOA0214	58710	SOA0291	58764	SOA0362
58603	SOA0096	58657	SOA0215	58711	soa0291n	58765	SOA0363
58604	SOA0100	58658	SOA0216	58712	SOA0292	58766	soa0363n
58605	SOA0101	58659	SOA0217	58713	soa0292n	58767	SOA0365
58606	SOA0103	58660	SOA0219	58714	SOA0294	58768	SOA0368
58607	SOA0105	58661	SOA0220	58715	soa0294n	58769	SOA0369
58608	SOA0107	58662	SOA0221	58716	SOA0298	58770	SOA0370
58609	SOA0109	58663	SOA0222	58717	soa0298n	58771	SOA0372
58610	SOA0111	58664	SOA0223	58718	SOA0300	58772	SOA0373
58611	soa0111n	58665	SOA0224	58719	soa0300n	58773	soa0373n
58612	SOA0116	58666	SOA0225	58720	SOA0301	58774	SOA0375
58613	SOA0117	58667	SOA0228	58721	soa0301n	58775	SOA0376
58614	SOA0121	58668	SOA0229	58722	SOA0303	58776	SOA0377
58615	SOA0122	58669	SOA0230	58723	SOA0304	58777	SOA0379
58616	SOA0125	58670	soa0230n	58724	SOA0306	58778	SOA0381
58617	SOA0131	58671	SOA0231	58725	soa0306n	58779	SOA0382
58618	SOA0132	58672	SOA0233	58726	SOA0307	58780	soa0382n
58619	SOA0133	58673	SOA0234	58727	SOA0308	58781	SOA0384
58620	SOA0134	58674	SOA0236	58728	SOA0310	58782	SOA0387
58621	SOA0138	58675	SOA0237	58729	SOA0315	58783	SOA0388
58622	SOA0140	58676	soa0237n	58730	SOA0317	58784	soa0388n
58623	soa0140n	58677	SOA0239	58731	SOA0319	58785	SOA0389
58624	SOA0141	58678	SOA0240	58732	SOA0322	58786	SOA0391
58625	SOA0142	58679	soa0240n	58733	SOA0323	58787	SOA0393

58788	SOA0397	58840	SOA0481	58892	SOA0558	58944	SOA0636
58789	SOA0399	58841	SOA0482	58893	SOA0559	58945	soa0636n
58790	SOA0401	58842	SOA0483	58894	SOA0560	58946	SOA0637
58791	SOA0403	58843	SOA0484	58895	SOA0561	58947	soa0637n
58792	SOA0405	58844	SOA0485	58896	SOA0563	58948	SOA0639
58793	soa0405n	58845	SOA0486	58897	SOA0564	58949	SOA0640
58794	SOA0406	58846	soa0486n	58898	soa0564n	58950	SOA0641
58795	SOA0409	58847	SOA0487	58899	SOA0565	58951	SOA0642
58796	SOA0410	58848	SOA0488	58900	SOA0567	58952	SOA0643
58797	SOA0411	58849	SOA0489	58901	SOA0568	58953	SOA0646
58798	SOA0412	58850	soa0489n	58902	soa0568n	58954	SOA0647
58799	SOA0413	58851	SOA0490	58903	SOA0569	58955	SOA0648
58800	SOA0415	58852	SOA0491	58904	SOA0570	58956	SOA0650
58801	SOA0416	58853	SOA0493	58905	SOA0571	58957	SOA0651
58802	SOA0417	58854	SOA0495	58906	SOA0575	58958	SOA0652
58803	SOA0419	58855	SOA0496	58907	SOA0579	58959	SOA0654
58804	SOA0420	58856	SOA0498	58908	SOA0580	58960	soa0659
58805	SOA0421	58857	SOA0501	58909	SOA0583	58961	soa0660
58806	SOA0426	58858	SOA0503	58910	SOA0585	58962	soa0661
58807	SOA0427	58859	SOA0505	58911	soa0585n	58963	soa0662
58808	SOA0428	58860	SOA0506	58912	SOA0589	58964	soa0667
58809	SOA0429	58861	SOA0514	58913	SOA0591	58965	soa0670
58810	soa0435	58862	SOA0516	58914	SOA0593	58966	SOA0673
58811	SOA0435N	58863	SOA0518	58915	SOA0594	58967	SOA0674
58812	soa0436	58864	SOA0520	58916	SOA0598	58968	soa0674n
58813	soa0437	58865	SOA0521	58917	SOA0600	58969	SOA0675
58814	soa0439	58866	soa0521n	58918	SOA0601	58970	SOA0677
58815	soa0440	58867	SOA0523	58919	SOA0602	58971	soa0677n
58816	soa0442	58868	SOA0525	58920	SOA0604	58972	SOA0678
58817	SOA0442N	58869	SOA0526	58921	SOA0605	58973	SOA0679
58818	SOA0444	58870	SOA0527	58922	SOA0606	58974	SOA0684
58819	SOA0445	58871	SOA0529	58923	SOA0608	58975	SOA0685
58820	SOA0448	58872	soa0529n	58924	SOA0609	58976	SOA0688
58821	SOA0449	58873	SOA0532	58925	soa0609n	58977	SOA0690
58822	SOA0450	58874	SOA0533	58926	SOA0611	58978	SOA0692
58823	SOA0453	58875	soa0533n	58927	SOA0612	58979	SOA0693
58824	SOA0461	58876	SOA0535	58928	SOA0613	58980	SOA0694
58825	soa0461n	58877	SOA0536	58929	soa0613n	58981	SOA0698
58826	SOA0463	58878	SOA0537	58930	SOA0614	58982	SOA0701
58827	soa0463n	58879	SOA0539	58931	SOA0615	58983	SOA0704
58828	SOA0464	58880	soa0539n	58932	SOA0616	58984	SOA0705
58829	SOA0466	58881	SOA0540	58933	SOA0619	58985	soa0705n
58830	soa0466n	58882	soa0540n	58934	SOA0620	58986	SOA0706
58831	SOA0467	58883	SOA0541	58935	SOA0621	58987	SOA0707
58832	SOA0468	58884	SOA0542	58936	SOA0622	58988	soa0712
58833	SOA0470	58885	SOA0544	58937	SOA0623	58989	soa0713
58834	SOA0471	58886	SOA0545	58938	SOA0630	58990	soa0715
58835	SOA0473	58887	SOA0546	58939	SOA0631	58991	soa0716
58836	SOA0476	58888	SOA0549	58940	SOA0632	58992	soa0718
58837	SOA0477	58889	SOA0550	58941	SOA0633		
58838	soa0477n	58890	SOA0552	58942	soa0633n		
58839	SOA0478	58891	SOA0554	58943	SOA0634		

Characterization of the 12,651 EST sequences from the mild OA library and of the 14,222 EST sequences from the severe OA library based on functional characterization of the known genes represented resulted in the following tables:

Table 4. Mild OA Cartilage Library		
Putative Function	Percent (%) of Representation in the Library	Number of Genes
Cell division	6.39	127
Cell signaling/communication	15.31	304
Cell structure/motility	9.16	182
Cell/organism defense	8.41	167
Gene/protein expression	21.60	429
Metabolism	13.95	277
Unclassified	22.76	452
Total known/unique genes analyzed	100.00	1938

Table 5. Severe OA Cartilage Library		
Putative Function	Percent (%) of Representation in the Library	Number of genes
Cell division	6.81	157
Cell signaling/communication	14.14	326
Cell structure/motility	8.50	196
Cell/organism defense	7.98	184
Gene/protein expression	22.94	529
Metabolism	13.53	312
Unclassified	23.94	552
Total known/unique genes analyzed	100.00	2256

Example 4: Identification Of Differentially Expressed Genes In Fetal, Normal Mild Osteoarthritic, And Severe Osteoarthritic Cartilage

Genes that are differentially expressed as defined herein between normal, mild, severe and fetal cartilage were identified through relative EST frequency analysis (see Figure 6). Of the 5,807 known unique genes identified in Figure 6, 405 genes were found to be expressed in all four tissue types. Examples of the possible subanalyses are shown in Figures 15 and 16. Some of these genes with particularly marked differential expression are shown in Figure 4. The relative frequency of ESTs representing collagens (Figures 2 and 3) and selected extracellular matrix proteins (see Figure 1) were also analyzed.

Example 5: Microarray Construction

A microarray according to the invention was constructed as follows.

PCR products (~40 ul) of cDNA clones from OA cartilage cDNA libraries, in the same 96-well tubes used for amplification, are precipitated with 4 ul (1/10 volume) of 3M sodium acetate (pH 5.2) and 100 ul (2.5 volumes) of ethanol and stored overnight at –20°C. They are then centrifuged at 3,300 rpm at 4°C for 1 hour. The obtained pellets were washed with 50 ul ice-cold 70% ethanol and centrifuged again for 30 minutes. The pellets are then air-dried and resuspended well in 50% dimethylsulfoxide (DMSO) or 20ul 3X SSC overnight. The samples are then deposited either singly or in duplicate onto Gamma Amino Propyl Silane (Corning CMT-GAPS or CMT-GAP2, Catalog No. 40003, 40004) or polylysine-coated slides (Sigma Cat. No. P0425) using a robotic GMS 417 or 427 arrayer (Affymetrix, CA). The boundaries of the DNA spots on the microarray are marked with a diamond scriber. The invention provides for arrays wherein 10-20,000 PCR products are spotted onto a solid support to prepare an array.

The arrays are rehydrated by suspending the slides over a dish of warm particle free ddH_20 for approximately one minute (the spots will swell slightly but not run into each other)

and snap-dried on a 70-80°C inverted heating block for 3 seconds. DNA is then UV crosslinked to the slide (Stratagene, Stratalinker, 65 mJ – set display to "650" which is 650 x 100 uJ) or baked at 80C for two to four hours. The arrays are placed in a slide rack. An empty slide chamber is prepared and filled with the following solution: 3.0 grams of succinic anhydride (Aldrich) is dissolved in 189 ml of 1-methyl-2-pyrrolidinone (rapid addition of reagent is crucial); immediately after the last flake of succinic anhydride dissolved, 21.0 ml of 0.2 M sodium borate is mixed in and the solution is poured into the slide chamber. The slide rack is plunged rapidly and evenly in the slide chamber and vigorously shaken up and down for a few seconds, making sure the slides never leave the solution, and then mixed on an orbital shaker for 15-20 minutes. The slide rack is then gently plunged in 95°C ddH_20 for 2 minutes, followed by plunging five times in 95% ethanol. The slides are then air dried by allowing excess ethanol to drip onto paper towels. The arrays are then stored in the slide box at room temperature until use.

Example 6: Target Nucleic acid Preparation and Hybridization

Preparation of Fluorescent DNA Probe from mRNA

Fluorescently labeled target nucleic acid samples are prepared for analysis with an array of the invention.

2 μg Oligo-dT primers are annealed to 2 ug of mRNA isolated from a cartilage sample from patient diagnosed with osteoarthritis or suspected of having osteoarthritis in a total volume of 15 ul, by heating to 70°C for 10 min, and cooled on ice. The mRNA is reverse transcribed by incubating the sample at 42°C for 1.5-2 hours in a 100 μl volume containing a final concentration of 50 mM Tris-HCl (pH 8.3), 75 mM KCl, 3 mM MgCl2, 25 mM DTT, 25 mM unlabeled dNTPs, 400 units of Superscript II (200 U/uL, Gibco BRL), and 15 mM of Cy3 or Cy5 (Amersham). RNA is then degraded by addition of 15μl of 0.1N NaOH, and incubation at 70°C for 10 min. The reaction mixture is neutralized by addition of 15μl of

0.1N HCL, and the volume is brought to 500µl with TE (10mM Tris, 1mM EDTA), and 20 µg of Cot1 human DNA (Gibco-BRL) is added.

The labeled target nucleic acid sample is purified by centrifugation in a Centricon-30 micro-concentrator (Amicon). If two different target nucleic acid samples (e.g., two samples derived from different patients) are being analyzed and compared by hybridization to the same array, each target nucleic acid sample is labeled with a different fluorescent label (e.g., Cy3 and Cy5) and separately concentrated. The separately concentrated target nucleic acid samples (Cy3 and Cy5 labeled) are combined into a fresh centricon, washed with 500µl TE, and concentrated again to a volume of less than 7µl. 1µL of 10µ g/µl polyA RNA (Sigma, #P9403) and 1 µl of 10µg/ul tRNA (Gibco-BRL, #15401-011) is added and the volume is adjusted to 9.5 µl with distilled water. For final target nucleic acid preparation 2.1µl 20XSSC (1.5M NaCl, 150mM NaCitrate (pH8.0)) and 0.35µl 10%SDS is added.

Hybridization

Labeled nucleic acid is denatured by heating for 2 min at 100°C, and incubated at 37°C for 20-30 min before being placed on a nucleic acid array under a 22mm x 22mm glass cover slip. Hybridization is carried out at 65°C for 14 to 18 hours in a custom slide chamber with humidity maintained by a small reservoir of 3XSSC. The array is washed by submersion and agitation for 2-5 min in 2X SSC with 0.1%SDS, followed by 1X SSC, and 0.1X SSC. Finally, the array is dried by centrifugation for 2 min in a slide rack in a Beckman GS-6 tabletop centrifuge in Microplus carriers at 650 RPM for 2 min.

Example 7: Signal Detection And Data Generation

Following hybridization of an array with one or more labeled target nucleic acid samples, arrays are scanned immediately using a GMS Scanner 418 and Scanalyzer software (Michael Eisen, Stanford University), followed by GeneSpring software (Silicon Genetics,

CA) analysis. Alternatively, a GMS Scanner 428 and Jaguar software may be used followed by GeneSpring software analysis

If one target nucleic acid sample is analyzed, the sample is labeled with one fluorescent dye (e.g., Cy3 or Cy5).

After hybridization to a microarray as described in Example 6, fluorescence intensities at the associated nucleic acid members on the microarray are determined from images taken with a custom confocal microscope equipped with laser excitation sources and interference filters appropriate for the Cy3 or Cy5 fluors.

The presence of Cy3 or Cy5 fluorescent dye on the microarray indicates hybridization of a target nucleic acid and a specific nucleic acid member on the microarray. The intensity of Cy3 or Cy5 fluorescence represents the amount of target nucleic acid which is hybridized to the nucleic acid member on the microarray, and is indicative of the expression level of the specific nucleic acid member sequence in the target sample.

When two target nucleic acid samples are being analyzed and compared (e.g., mild osteoarthritic vs severe osteoarthritic), one target nucleic acid sample (for example, mild osteoarthritic) is labeled with fluorescent dye Cy3, the other target nucleic acid sample (for example, severe osteoarthritis) is labeled with fluorescent dye Cy5.

After hybridization as described in Example 6, fluorescence intensities at the associated nucleic acid members on the microarray are determined from images taken with a custom confocal microscope equipped with laser excitation sources and interference filters appropriate for the Cy3 and Cy5 fluors. Separate scans are taken for each fluor at a resolution of 225 μm^2 per pixel and 65,536 gray levels. Normalization between the images is used to adjust for the different efficiencies in labeling and detection with the two different fluors. This is achieved by manual matching of the detection sensitivities to bring a set of internal

control genes to nearly equal intensity followed by computational calculation of the residual scalar required for optimal intensity matching for this set of genes.

The presence of Cy3 or Cy5 fluorescent dye on the microarray indicates hybridization of a target nucleic acid and a specific nucleic acid member on the microarray. The intensities of Cy3 or Cy5 fluorescence represent the amount of target nucleic acid which is hybridized to the nucleic acid member on the microarray, and is indicative of the expression level of the specific nucleic acid member sequence in the target sample. If a nucleic acid member on the array shows no color, it indicates that the gene in that element is not expressed in either sample. If a nucleic acid member on the array shows a single color, it indicates that a labeled gene is expressed only in that cell sample. The appearance of both colors indicates that the gene is expressed in both tissue samples. The differences in Cy3 and Cy5 fluorescence intensities, after normalization, are indicative of differences of expression levels of the associated nucleic acid member sequence in the two samples for comparison. Differences in expression intensity between the two samples greater than 1.0 foldare used as an indication of differential gene expression.

The array is scanned in the Cy 3 and Cy5 channels and stored as separate 16-bit TIFF images. The images are incorporated and analysed using Scanalyzer software which includes a gridding process to capture the hybridization intensity data from each spot on the array. The fluorescence intensity and background-subtracted hybridization intensity of each spot is collected and a ratio of measured mean intensities of Cy5 to Cy3 is calculated. A liner regression approach is used for normalization and assumes that a scatter plot of the measured Cy5 versus Cy3 intensities should have a scope of one. The average of the ratios is calculated and used to rescale the data and adjust the slope to one. A post-normalization cutoff of greater than 1.0 fold up- or down-regulation is used to identify differentially expressed genes.

Analysis of a microarray comprising some of the sequences in Figure 14, resulted in 36 candidate upregulated genes in the mild OA library that showed a greater than 2-fold

median ratio and 47 candidate downregulated genes that showed a less than 0.2-fold median ratio (Figures 9 and 10, respectively,). A total of 38 candidate upregulated genes were also identified in the severe OA library that showed a greater than 2-fold median ratio and 51 candidate downregulated genes that showed a less than 0.2-fold median ratio (Figures 11 and 12, respectively,). According to this embodiment, the microarray was hybridized with a target nucleic acid sample derived from an individual diagnosed with mild osteoarthritis and a target nucleic acid sample derived from an individual diagnosed with severe osteoarthritis. As would be clear to a person skilled in the art, similar analysis can be performed for any of the sequences identified in Figure 13, or the sequences identified in Figure 6A which correspond to the genes disclosed in Figure 6 using the methods disclosed herein.

Example 8: Chondrocyte-Specific Gene Microarray And Diagnosis Microarray Construction

A collection of nucleic acid members are spotted on a glass slide as described in Example 5 for the construction of a mild OA diagnosis microarray. A collection of nucleic acid members are spotted on a glass slide as described in Example 5 for the construction of a severe OA diagnosis microarray. A collection of chondrocyte specific nucleic acid members are spotted on a glass slide as described in Example 5 for the construction of a chondrocyte-specific gene microarray. The nucleic acid members spotted onto the microarrays described are selected from those named in Figures 6B, 6C, 6D and 6E.

Example 9: Diagnosis

Target nucleic acid samples are prepared from cartilage RNA extracts of an individual (as described in Example 6) and hybridized to a microarray comprising a collection of nucleic acid members wherein at least one member is differentially expressed in cartilage isolated from a patient diagnosed with mild, moderate, marked or severe osteoarthritis, as compared to cartilage isolated from a normal individual as defined herein (as described in Example 6). A hybridization pattern is generated and analyzed as in Example 7. For example, the

hybridization of target nucleic acid samples to one or more nucleic acid members on the microarray comprising a collection of nucleic acid members wherein at least one member is differentially expressed in mild osteoarthritis cartilage as compared to a normal individual is indicative of a mild osteoarthritis of the individual from whom the target nucleic acid sample is derived. The hybridization of target nucleic acid samples to one or more nucleic acid members on the microarray comprising a collection of nucleic acid members differentially expressed in severe osteoarthritis cartilage as compared to the normal individual is indicative of severe osteoarthritis of the individual from whom the target nucleic acid sample is derived.

Example 10: Therapeutic Agent Screening

A candidate therapeutic agent that increases or decreases the expression of one or more polynucleotide sequences that are differentially expressed in a chondrocyte derived from any of the following chondrocyte disease or developmental stages: fetal, mild osteoarthritic, moderate osteoarthritic, marked osteoarthritic and severe osteoarthritic, is screened according to the following method.

Chondrocytes are isolated from a "normal" individual and are incubated in the presence and absence of a candidate agent for varying amounts of time (i.e., 30 min, 1 hr, 5 hr, 24 hr, 48 hr and 96 hrs). When screening for therapeutic genes, a clone of a full gene sequence corresponding to an EST in Figure 6A or Figure 13 is used to transfect chondrocytes. The transfected chondrocytes are cultured for varying amounts of time (i.e., 1, 2, 3, 5, 7, 10, or 14 days). Following incubation, target nucleic acid samples are prepared from the chondrocytes and hybridized to a nucleic acid probe corresponding to a polynucleotide sequence which is differentially expressed in a chondrocyte derived from at least any two of the following of: fetal, normal, mild osteoarthritic, moderate osteoarthritic and severe osteoarthritic. The nucleic acid probe is labeled, for example with a radioactive label, according to methods well-known in the art and described herein. Hybridization is carried out by northern blot, for example as described in Ausubel et al., *supra* or Sambrook et al., *supra*). The differential hybridization, as defined herein, of the probe to the target nucleic acid samples from normal relative to RNA from any one of fetal, mild osteoarthritic, moderate osteoarthritic, marked osteoarthritic and severe osteoarthritic is indicative of the level of expression of RNA corresponding to a differentially expressed chondrocyte specific polynucleotide sequence. A change in the level of expression of the probe sequence as a result of the incubation step in the presence of the candidate agent, is indicative of an agent that increases or decreases the expression of the corresponding chondrocyte specific polynucleotide sequence.

Example 11: Assessing The Integrity Of Cartilage RNA Isolated Post-Mortem

The following Baboon cartilage study was preformed to evaluate the quality of freshly isolated RNA and RNA isolated at various times post-mortem.

Nine vials of baboon cartilage were obtained, and stored in liquid nitrogen till use.

Baboon cartilage from each vial was weighed and finely powdered under liquid nitrogen. The sample was then homogenized in TRIzol® reagent (0.1g/ml TRIzol®) and total RNA was extracted. The quantity of RNA was calculated according to the OD_{260} value. The appearance of two sharp bands on the RNA gel indicated that the RNA was of good quality.

RT-PCR was performed for the gene expression of collagen type II (COL2A1), B-actin and GAPDH, using 0.1ug total RNA from each sample.

The RNA gel pattern clearly shows that the RNA was not degraded up to 12 hours post-mortem (Table 7). Therefore stable RNA should be expected from the biopsy sample within 12 hours after death.

Table 7. Integrity Of Cartilage RNA Isolated Post-Mortem

Sample No.	Time Taken	Weight (g)	Total RNA (ug) - Based on OD260	RNA Gel (non Dil)	Col2A1	b-actin	GAPDH
1	Fresh	0.175	8	OK	++	++	++
2	1hr pm	0.29	9	OK	++	++	++
3	2hr	0.29	11.36	OK	++	+/-	+/-
4	3hr	0.25	2.8	OK	++	+/-	+/-
5	6hr	0.53	8.0	OK	++	+	+/-
6	8hr	0.18	5.26	OK	++	+	-

7	10hr	0.38	9.35	OK	++	+	+/-
8	12hr	0.20	6.7	OK	++	+/-	-
9	24hr	0.41	9.35	SMEAR	+/-	-	-

Collagen type II is abundant and specific to normal articular cartilage. Its mRNA level was comparable among all the samples except #9 (24 hours post-mortem). It should be noted that samples taken earlier will better reflect the natural *in vivo* state.

Example 12. Expressed Sequence Tags (ESTs) Analysis of Human Chondrocyte Gene Expression in Mild and Severe Osteoarthritic Cartilage

Large-scale partial sequencing of cDNA libraries obtained from human fetal cartilage was performed to identify expressed sequence tags (ESTs) corresponding to genes that might play critical roles in OA progression. Large scale sequencing of cDNA libraries from human normal, mild and severe OA cartilage was also performed and a total of over 44,000 ESTs from the three cDNA libraries were analyzed.

Normal cartilage was obtained from the donor program of Department of Orthopaedics and Rehabilitation, University of Miami. OA cartilage samples were obtained from either areas of very early cartilage degeneration (mild) or from sites of end stage disease (severe) during either arthroscopic knee surgery or total knee replacement. Total RNA from cartilage was extracted using TRIzol® reagent (GIBCO). cDNA libraries were constructed into λTriplEx2 vector through a PCR-based method, using SMART (Switching Mechanism At 5' end of RNA Transcript) cDNA Library Construction Kit (Clontech) as described above. Phage plaques were randomly picked and positive inserts were identified by PCR. Agarose gel electrophoresis was used to assess the presence and purity of inserts. PCR product was then subjected to automated DNA sequencing with a 5'vector-specific forward primer and sequenced by ABI PRISM 377 DNA sequencer (Perkin Elmer) and ABI PRISM 3700 DNA Analyzer (Applied Biosystems). All generated EST sequences were searched against the

nonredundant Genebank/EMBL/DDBL, dbEST and GSS databases. A minimum value of $p=10^{-10}$ and nucleotide sequence identity >90% were required for assignments of putative identities for EST-matching to known genes or other ESTs. Relative EST frequency level was calculated by dividing the EST number matched to that gene into the total number of ESTs obtained from the library.

A total of 17,151 ESTs, 12,651 ESTs and 14,222 ESTs were obtained from normal, mild and severe OA cDNA libraries respectively and used for gene expression profiling. About 44% of the total ESTs from these three cDNA libraries matched to known genes in the database, and about 0.9% of the ESTs (409) resulted in no significant match to known sequences and were thus designated as novel. Nonredundancy analysis of the known gene matches resulted in the identification of 2,518 unique genes in normal, 1,938 in mild and 2,256 in severe OA cartilage. Differentially expressed known genes amongst fetal (22), normal, mild, and severe OA cartilage (23) were identified by examining relative EST frequency levels as shown in Figure 6.

Some of the genes with particularly marked differential expression are shown in Figure 4 provided herewith. Heat shock protein 90 (HSP90) was the gene with the most abundant ESTs matches in mild OA. Its transcript level was low in fetal cartilage. Beta-2 microglobulin (B2M) level was higher in diseased cartilage than normal cartilage, and significantly higher in diseased cartilage than in fetal cartilage. Its EST levels in mild and severe OA were similar. Osteoblast specific factor 2 (OSF-2p1) was highly expressed in severe OA compared to fetal, mild and normal cartilage. Another differentially expressed gene was megakaryocyte stimulating factor (MSF, also known as superficial zone protein, or proteoglycan 4). It had a significantly higher expression in mild OA than in severe OA.

The relative frequency of ESTs representing the collagens was also analyzed as shown in Figure 3.

Noncollagenous matrix protein profiles showed higher EST levels of decorin (DCN), fibronectin (FN), lumican (LUM) and matrix G1a protein (MGP) in both mild and severe OA cartilage as shown in Figures 1 and 4 provided herewith.

Example 13. Microarray Analysis Of Beta-2 Microglobulin (B2M) Expression In Human Osteoarthritis

As discussed above, Beta-2 microglobulin (B2M) had a high EST expression level in mild and severe OA cartilage. B2M is a nonglycosylated polypeptide that is elevated in inflammatory and malignant diseases. It has been shown to induce stromelysin and cyclooxygenase-2 synthesis in human synovial fibroblasts (24, 25).

B2M expression during different stages of osteoarthritis was evaluated. Human OA synovial fluid (SF) was collected from human knee joint by aspiration at arthroscopy or total knee replacement. Normal samples were collected from volunteers with no history of knee injury or arthritis. Organ culture was performed as follows: human severe OA cartilage slices were cultured at one slice/well in a 24-well plate in DMEM (Dulbecco's modified Eagle medium), with 10% FCS, 100 units/ml penicillin and 100 mg/ml streptomycin (DMEM++) at 37°C in a humidified atmosphere of 5% CO_2. Cultured medium (20 ul) was then collected at different time points for B2M testing. B2M levels in synovial fluid and cartilage organ cultured medium were measured using a B2M enzyme immunoassay test kit (ALPCO). Statistical significance was assessed by Student's t-test with P values less than 0.05 being considered significant. Cell culture of chondrocytes from patients with severe OA was performed as follows. Chondrocytes were derived from cartilage from patients with severe OA through collagenase type II digestion. Cells were then seeded at 6.5×10^4/well (3.2×10^4/ml) in a 6-well plate and treated with or without 10 ug/ml B2M (Sigma) for 72 hr.

Microarrays containing 5184 chondrocyte-specific cDNA clones were used for gene expression profiling.

The average B2M levels detected in normal (nor), mild (mioa), moderate (mooa), marked (maoa) and severe OA (seoa) synovial fluid are shown in Figure 17. B2M in osteoarthritis synovial fluid is significantly higher than that in normal. However, no significant difference was found in B2M levels among different osteoarthritis stages.

To assess if chondrocytes contribute B2M secretion, medium from cultured severe OA cartilage was collected and tested for B2M. Figure 18 shows the release of B2M is detectable after 24 hour culture and continues to increase during the 72 hour study period. At 72 hours, the accumulation of B2M was about 2.1 ug/g cartilage. Similar results were obtained across three experimental runs, each using cartilage from a different donor.

Genes regulated by B2M were detected through microarray technology as described above. Figure 19 shows a black and white representation of a two-color fluorescent scan. Cy3 labeling (which would appear as green spots) correspond to genes preferentially expressed in non-B2M treated chondrocytes, while Cy5 labeling (which would appear as reddish spots) represent genes preferentially expressed in B2M treated chondrocytes. Genes expressed at approximately equal levels would appear as yellow spots. The identity of genes was determined by the location of nucleic acid members on the array. Some of the genes that were up or down-regulated at least two-fold by B2M are listed in Table 8.

Table 8. Genes Regulated by B2M

Up-Regulated	Down-Regulated
Adrenomedullin	hypothetical protein (KIAA0102)
chitinase precursor=YKL-39	intersectin short form
collagen type III, alpha 1	KARP-1 binding protein 2 (KAB2)
manganese superoxide dismutase (SOD-2)	peripheral myelin protein 22 (PMP22)
syntaxin 7	putative GTP binding protein

Variations, modifications, and other implementations of what is described herein will occur to those of ordinary skill in the art without departing from the spirit and scope of the invention. The references provided below are incorporated herein by reference in their entireties.

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What is claimed is:

Claims

1. A composition for use in diagnosing or prognosing osteoarthritis, said composition comprising two or more isolated polynucleotides, each said polynucleotide capable of selectively hybridizing to a unique gene identified as differentially expressed in cartilage derived from a patient diagnosed with osteoarthritis relative to cartilage derived from a normal individual.

2. The composition of claim 1 wherein each said unique genes are chosen from the genes disclosed in Figure 6.

3. The composition of claim 1 or 2 wherein two or more of said polynucleotides are chosen from the ESTs identified in Figure 6A whose sequences are disclosed in Figure 14.

4. The use of a composition of claim 1, 2, or 3 for diagnosis or prognosis of osteoarthritis.

5. A composition for use in diagnosing or prognosing mild osteoarthritis, said composition comprising two or more isolated polynucleotides, each said polynucleotide capable of selectively hybridizing to a unique gene identified as differentially expressed in cartilage derived from a patient diagnosed with mild osteoarthritis relative to cartilage derived from a normal individual.

6. A composition for use in diagnosing or prognosing mild osteoarthritis, said composition comprising two or more isolated polynucleotides, each said polynucleotide capable of selectively hybridizing to a unique gene identified as differentially expressed in cartilage derived from a patient diagnosed with mild osteoarthritis relative to cartilage derived from a patient diagnosed with severe osteoarthritis.

7. The composition of claim 5, or 6 wherein each said unique genes are chosen from the genes disclosed in Figure 6.

8. The composition of claim 5 or 6 wherein each said unique genes are chosen from the genes disclosed in Figure 9, 10 or 16.

9. The composition of claim 5, 6, 7 or 8 wherein two or more of said polynucleotides are chosen from the ESTs identified in Figure 6A whose sequences are disclosed in Figure 14.

10. The use of a composition of claim 5, 6, 7, 8, or 9 for diagnosing or prognosing mild osteoarthritis.

11. A composition for use in diagnosing or prognosing severe osteoarthritis, said composition comprising two or more isolated polynucleotides, each said polynucleotide capable of selectively hybridizing to a unique gene identified as differentially expressed in cartilage derived from a patient diagnosed with severe osteoarthritis relative to cartilage derived from a normal individual.

12. A composition for use in diagnosing or prognosing severe osteoarthritis, said composition comprising two or more isolated polynucleotides, each said polynucleotide capable of selectively hybridizing to a unique gene identified as differentially expressed in cartilage derived from a patient diagnosed with severe osteoarthritis relative to cartilage derived from a patient diagnosed with mild osteoarthritis.

13. The composition of claim 11 or 12 wherein each said unique genes are chosen from the genes disclosed in Figure 6.

14. The composition of claim 11 or 12 wherein each said unique genes are chosen from the genes disclosed in Figure 11, 12 or 16.

15. The composition of claim 11, 12 13 or 14 wherein two or more of said polynucleotides are chosen from the ESTs identified in Figure 6A whose sequences are disclosed in Figure 14.

16. The use of a composition of claim 11, 12, 13, 14 or 15 for diagnosing or prognosing severe osteoarthritis.

17. An array comprising the composition of claim 1, 2, or 3 and a substrate; wherein each polynucleotide has a unique position on said array and is stably associated with said substrate.

18. The use of an array of claim 17 for diagnosing or prognosing osteoarthritis.

19. An array comprising the composition of claim 5, 6, 7, 8 or 9 and a substrate; wherein each polynucleotide has a unique position on said array and is stably associated with said substrate.

20. The use of an array of claim 19 for diagnosing or prognosing mild osteoarthritis.

21. An array comprising the composition of claim 11, 12, 13, 14 or 15 and a substrate; wherein each polynucleotide has a unique position on said array and is stably associated with said substrate.

22. The use of an array of claim 21 for diagnosing or prognosing severe osteoarthritis.

23. An array of claim 17, 19 or 21 wherein each polynucleotide is at least 50 nucleotides in length.

24. An array of claim 17, 19 or 21 wherein each polynucleotide is at least 100 nucleotides in length.

25. A method of diagnosing or prognosing osteoarthritis in a patient, comprising: hybridizing an isolated sample from said patient with an array of claim 17; generating a hybridization pattern for RNA, cDNA or EST of said isolated sample and comparing said hybridization pattern with a hybridization pattern generated for an isolated sample from a normal individual; and comparing the two hybridization patterns to determine whether there is differential expression of polynucleotides isolated from said patient as compared to said normal individual, wherein said differential expression is indicative of osteoarthritis.

26. A method of diagnosing or prognosing mild osteoarthritis in a patient, comprising: hybridizing an isolated sample from said patient with an array of claim 19 generating a hybridization pattern for RNA, cDNA or EST of said isolated sample and comparing said hybridization pattern with a hybridization pattern generated for an isolated sample from a normal individual; and comparing the two hybridization patterns to determine whether there is differential expression of polynucleotides isolated from said patient as compared to said normal individual, wherein said differential expression is indicative of mild osteoarthritis

27. A method of diagnosing or prognosing severe osteoarthritis in a patient, comprising: hybridizing an isolated sample from said patient sample with an array of claim 21 generating a hybridization pattern for RNA, cDNA or EST of said isolated sample and comparing said hybridization pattern with a hybridization pattern generated for an isolated sample from a normal individual; and comparing the two hybridization patterns to determine whether there is differential expression of polynucleotides isolated from said patient as compared to said normal individual, wherein said differential expression is indicative of severe osteoarthritis.

28. The method of claim 25, 26, or 27 further comprising the step of isolating said RNA from said isolated sample.

29. The method of claim 25, 26, or 27 wherein said isolated sample is a cartilage sample.

30. The method of claim 25, 26 or 27 wherein said isolated sample is a blood sample.

31. The method of claim 25, 26 or 27 wherein said isolated sample is a synovial fluid sample.

32. A kit comprising an array of claim 17, 19 or 21 and packaging means therefor.

33. A kit of claim 28 further comprising instructions wherein said instructions recite the method for using said kit according to claim 23, 24, 25, 26, 27 or 28.