WHAT IS CLAIMED IS:

1. A method for making mammalian cells that produce recombinant adeno-associated viral (rAAV) vector, comprising the steps:

(a) providing a plasmid/polyethylenimine (PEI) mixture of components (i), (ii) and (iii):

(i) one or more plasmids comprising nucleic acids encoding AAV packaging proteins and nucleic acids encoding helper functions;

(ii) a plasmid comprising a transgene that encodes a protein or is transcribed into a transcript of interest; and

(iii) a polyethylenimine (PEI) solution,

(b) contacting cells with the plasmid/PEI mixture of step (a) to produce a plasmid/PEI cell culture;

(c) adding valproic acid, a salt thereof or a derivative comprising an amino acid linked or conjugated thereto to the plasmid/PEI cell culture to produce a second mixture; and

(d) incubating said second mixture of step (c) thereby making transfected cells that produce recombinant rAAV vector.

2. The method of claim 1, further comprising step (e) harvesting said transfected cells produced in step (d) and/or culture medium from the transfected cells produced in step (d) to produce a cell and/or culture medium harvest.

3. The method of claim 1, further comprising step (e) culturing, expanding, isolating or selecting for cells that have been transfected with the plasmids comprising nucleic acids encoding AAV packaging proteins and nucleic acids encoding helper functions and the plasmid comprising the transgene.

4. The method of claim 2, further comprising isolating and/or purifying recombinant AAV vector from the transfected cells produced in step (d) and/or culture medium from the transfected cells produced in step (d).

5. The method of claim 2, further comprising step (f) isolating and/or purifying recombinant AAV vector from the transfected cells and/or culture medium harvest produced in step (e).

6. The method of claim 1, wherein the transgene encodes a protein.

7. The method of any of claims 1 – 6, wherein the transgene encodes a wild-type, or functional variant blood clotting factor, apoE2, TPP1, argininosuccinate synthase, copper transporting ATPase 2, acid alpha-glucosidase, β-Glucocerebrosidase, α-galactosidase or C1 inhibitor serine protease inhibitor.

8. The method of claim 7, wherein the wild-type, or functional variant blood clotting factor is Factor VII, Factor VIII, or Factor IX.

9. The method of any of claims 1 – 8, wherein step (c) comprises adding valproic acid.

10. The method of any of claims 1 – 9, wherein the valproic acid, salt or derivative thereof is added at the time of step (b) or immediately after step (b) or up to 3 hours after step (b).

11. The method of any of claims 1 – 9, wherein the valproic acid, salt or derivative thereof is added two or more times after step (b).

12. The method of claim 10, wherein the valproic acid, salt or derivative thereof is first added at the time of or immediately after step (b) or up to 3 hours after step (b) and added again 12-72 hours after step (b), or added again 12-48 hours after step (b), or added again 12-24 hours after step (b).

13. The method of any of claims 1 – 12, wherein said plasmids (i) and (ii) are in a molar ratio range of 1:0.01 to 1:100, and wherein the mixture of components (i), (ii) and (iii) has been incubated for a period of time from 10 seconds to 4 hours prior to step (b).

14. The method of any of claims 1 – 13, wherein said plasmids are in a PEI:plasmid weight ratio in the range of 0.1:1 to 5:1, or in a PEI:plasmid weight ratio in the range of 5:1 to 0.1:1.

15. The method of any of claims 1 – 14, further comprising adding free PEI to the cells after step (b), or prior to or at the time of or after step (c); or further comprising adding free PEI to the cells at the time of or after step (b), or at the time of or after step (c).

16. The method of claim 15, wherein said free PEI is added so that the PEI:plasmid weight ratio is in the range of 0.1:1 to 5:1, or is in the range of 5:1 to 0.1:1.

17. The method of claim 15 or claim 16, wherein the PEI in the plasmid/PEI mixture comprises linear polyethylenimine; or a hydrolyzed linear polyethylenimine; or a hydrolyzed linear polyethylenimine with a molecular weight in the range of 4,000 to 160,000 and/or in the range of 2,500 to 250,000 molecular weight in free base form.

18. The method of any of claims 15 – 17, wherein the molar ratio of nitrogen (N) in the free PEI and the PEI in the plasmid/PEI mixture to phosphate (P) in the plasmid/PEI mixture is in the range of 1:1 to 50:1 (N:P).

19. The method of any of claims 15 – 18, wherein the amount of free PEI is 10% to 90% of a sum of PEI present in the plasmid/PEI mixture and the free PEI; or wherein the amount of free PEI is 0.1 μg/mL to 10 μg/mL.

20. The method of any of claims 15 – 16 or 19, wherein the PEI solution and/or free PEI comprises a solution having a pH from 7.0 to 8.0.

21. The method of any of claims 15 – 20, wherein said plasmids and PEI have been incubated from 10 seconds to 4 hours with each other prior to step (a).

22. The method of any of claims 15 – 20, wherein said plasmids and PEI have been incubated from 1 minute to 30 minutes with each other prior to step (a).

23. The method of any of claims 15 – 22, wherein the mixture of components (i), (ii) and (iii) are incubated together for 10 seconds to 4 hours prior to step (b); or wherein the incubating of step (d) is for at least 4 hours.

24. The method of any of claims 15 – 23, wherein the cells are human embryonic kidney (HEK) or Chinese hamster ovary (CHO) cells; or wherein the cells comprise Human Embryonic Kidney (HEK) 293 cells; or wherein said cells are HEK 293E, HEK 293F or HEK 293T cells.

25. The method of any of claims 15 – 24, wherein the cells are stably or transiently transfected; or wherein the cells are in suspension culture; or wherein the cells are adherent; or wherein said cells are grown or maintained in a serum-free culture medium.

26. The method of any of claims 15 – 25, wherein said cells are at a density in the range of 1×10^5 cells/mL to 1×10^8 cells/mL when contacted with said plasmid/PEI mixture and/or when contacted with said free PEI.

27. The method of any of claims 15 – 26, wherein viability of said cells when contacted with said plasmid/PEI mixture or with said free PEI is 60% or greater than 60%, or wherein said cells are in log phase growth when contacted with said plasmid/PEI mixture.

28. The method of any of claims 1 – 27, wherein the total amount of plasmids is in the range of 0.1 µg to 15 µg per mL of cells.

29. The method of any of claims 1 – 28, wherein the molar ratio of the plasmid comprising the transgene to the one or more plasmids comprising nucleic acids encoding AAV packaging proteins and/or nucleic acids encoding helper functions is from 1:5 to 1:1, or is from 1:1 to 5:1.

30. The method of any of claims 1 – 29, wherein said one or more plasmids comprises a first plasmid comprising the nucleic acids encoding AAV packaging proteins comprising AAV rep and AAV cap, and a second plasmid comprising the nucleic acids encoding helper functions comprising E2 and E4 proteins.

31. The method of claim 30, wherein the molar ratio of the plasmid comprising the transgene to the first plasmid comprising the nucleic acids encoding AAV packaging proteins to the second plasmid comprising the nucleic acids encoding helper functions is in the range of 1-5:1:1, or 1:1-5:1, or 1:1:1-5.

32. The method of any of claims 1 – 31, wherein the encoded AAV packaging proteins comprise AAV rep and AAV cap.

33. The method of any of claims 1 – 32, wherein the encoded helper functions comprise adenovirus E2 and E4 proteins, VA RNA and helper proteins not encoded by AAV.

34. The method of any of claims 1 – 33, wherein the recombinant AAV vector comprises any of AAV serotypes 1-12, and an AAV VP1, VP2 and VP3 capsid protein.

35. The method of any of claims 1 – 34, wherein the adeno-associated viral (AAV) vector comprises a capsid VP1, VP2 and VP3 protein sequence or inverted terminal repeat sequence having 70% or more sequence identity to a capsid protein sequence or to an inverted terminal repeat (ITR) sequence of any capsid protein sequence or ITR selected from AAV1, AAV2, AAV3, AAV4, AAV5, AAV6, AAV7, AAV8, AAV9, and AAV10 serotypes; or wherein the adeno-associated viral (AAV) vector comprises a capsid VP1, VP2 and/or VP3 protein sequence having 70% or more sequence identity to a capsid protein sequence selected from SEQ ID NO:1 and SEQ ID NO:2.

36. The method of any of claims 1 – 35, wherein the AAV vector comprises an AAV serotype or an AAV pseudotype, wherein said AAV pseudotype comprises an AAV

capsid serotype different from the ITR serotype; or wherein the AAV vector comprises a capsid VP1, VP2 and VP3 protein or inverted terminal repeat of any serotype selected from AAV1, AAV2, AAV3, AAV4, AAV5, AAV6, AAV7, AAV8, AAV9, AAV10, SEQ ID NO:1 and SEQ ID NO:2; or wherein the AAV vector further comprises an intron, an expression control element, one or more adeno-associated virus (AAV) inverted terminal repeats (ITRs) and/or a filler polynucleotide sequence.

37. The method of claim 36, wherein the intron is within or flanks the nucleic acid that encodes a protein or is transcribed into a transcript of interest, or wherein the expression control element is operably linked to the nucleic acid that encodes a protein or is transcribed into a transcript of interest, or wherein the AAV ITR(s) flanks the 5' or 3' terminus of the nucleic acid that encodes a protein or is transcribed into a transcript of interest, or wherein the filler polynucleotide sequence flanks the 5' or 3' terminus of the nucleic acid that encodes a protein or is transcribed into a transcript of interest.

38. The method of any of claims 1 – 37, wherein the cells are subcultured to a reduced cell density prior to contact with said plasmid/PEI mixture.

39. The method of any of claims 1 – 38, wherein the cells are contacted with said plasmid/PEI mixture between a period of 2 days to 5 days after subculture.

40. The method of any of claims 15 – 27, wherein the amount of plasmids introduced into said transfected cells is at least 50% greater with the step of adding free PEI to the cell culture compared to without adding free PEI to the cell culture; or wherein the amount of plasmids introduced into said transfected cells is at least 50% greater with the step of adding the valproic acid, salt or derivative thereof compared to without adding the valproic acid, salt or derivative thereof.

41. The method of any of claims 15 – 16, 19 or 40, wherein the amount of recombinant AAV vector produced is at least 50% or greater with the step of adding free PEI to the plasmid/PEI cell culture compared to without adding free PEI to the plasmid/PEI cell

culture; or wherein the amount of recombinant AAV vector produced is 1-5, 5-8, 8-10 or 10-20 fold greater with the step of adding free PEI to the plasmid/PEI cell culture compared to without adding free PEI to the plasmid/PEI cell culture; or wherein the amount of recombinant AAV vector produced is 1-5, 5-8, 8-10 or 10-15 fold greater with the step of adding the valproic acid, salt or derivative thereof compared to without adding the valproic acid, salt or derivative thereof to the plasmid/PEI cell culture.

42. The method of any of claims 1 – 41, wherein after step (c) the valproic acid, salt or derivative thereof is at a concentration from 0.1mM to 25mM.

43. The method of claim 15 or 16, wherein the free PEI comprises linear polyethylenimine; or a hydrolyzed linear polyethylenimine; or a hydrolyzed linear polyethylenimine with a molecular weight in the range of 4,000 to 160,000 and/or in the range of 2,500 to 250,000 molecular weight in free base form.