as described above by adding to NIH/3T3 cells (cell line for mouse fibroblast). That is, NIH/3T3 cells were cultured in DMEM medium (10% (v/v)FBS) at 37°C under conditions of 5% CO_2, 1×10^3 cells were plated on each well of a 96-well plate with DMEM medium (5% (v/v)FBS), and cultured at 37°C under conditions of 5% CO_2 for 24 hours. Subsequently, replacement was carried out with a medium formed by mixing purified human FGF2 derived from the SP56 strain and the SP67 strain or recombinant hFGF2 (R&D systems) with DMEM medium (5% (v/v)FBS) so that the FGF2 concentration was 0.25 ng/mL to 10 ng/mL. As a negative control, replacement with a medium that was mixed with PBS(-) instead of the FGF2 solution was also carried out in the same manner. These plates were cultured at 37°C under conditions of 5% CO_2 for 4 days.

100 μL of Cell Counting kit-8 (Dojindo) was added to the plate, incubation was then carried out at 37°C under conditions of 5% CO_2 for a further 2 hours, and the activity in promoting cell proliferation for the NIH/3T3 cells was measured by measuring the absorbance at wavelengths of 450 nm and 630 nm (reference wavelength).

[0102]

(3) Results

The results are shown in FIG. 6.

Human FGF2 secreted by the SP56 and SP67 strains had concentration-dependent activity in promoting cell proliferation.

[0103]

Example 6: comparison of stability of amount of FGF2 secreted

(1) Method

10 mL of MRS (containing 75 μg/mL spectinomycin and 1% vitamin C-containing liquid) medium was inoculated at 1% with a glycerol stock of *Bifidobacterium longum* 105A/pFGF12a strain described in WO2013/008881, the FGF110 strain, and the FGF111 strain, and anaerobically cultured at 37°C for 24 hours, thus giving an activated culture broth. Subsequently, the same