METHODS OF REDUCING VIRUCIDAL ACTIVITY IN PCV-2 COMPOSITIONS AND PCV-2 COMPOSITIONS WITH AN IMPROVED IMMUNOGENICITY

This application relates to and claims priority to U.S. Provisional Patent Application No. 61/309,408, which was filed March 1, 2010, and U.S. Provisional Patent Application No. 61/239,192, which was filed September 2, 2009. All of which are incorporated herein by reference in their entirety. All applications are commonly owned.

SEQUENCE LISTING

The present application includes a sequence listing in accordance with 37 C.F.R. 1.821 – 1.825. The sequence listing accompanying this application is hereby incorporated by reference in its entirety.

BACKGROUND

Field of Invention

The present invention relates to methods and compositions, for reducing the virucidal activity of compositions that would normally exhibit some degree of virucidal activity. By using the methods of the present invention, the virucidal activity of such compositions can be reduced in comparison to the virucidal activity of a composition that does not include the steps of the present invention. More specifically, the present invention relates to methods for producing antigenic Porcine Circovirus Type II (PCV-2) compositions such that they show relatively little or no virucidal activity as compared to the compositions known in the art using current detection methods, and in particular, as compared to compositions not produced by a method according the present invention. The present invention further relates to a novel immunogenic composition, preferably a PCV-2 containing composition produced according to the method provided by the present patent application, preferably characterized by reduced or no virucidal activity relative to comparable compositions described in the art. According to a further aspect, the present invention also provides immunogenic compositions comprising purified PCV-2 antigen, preferably purified PCV-2 antigen with an improved immunogenicity.

Description of the Prior Art

Porcine circovirus type 2 (PCV-2) is a small (17 -22 nm in diameter), icosahedral, non-enveloped DNA virus, which contains a single-stranded circular genome. PCV-2 shares

approximately 80% sequence identity with porcine circovirus type 1 (PCV-1). However, in contrast with PCV-1, which is generally non-virulent, swine infected with PCV-2 exhibit a syndrome commonly referred to as Post-weaning Multisystemic Wasting Syndrome (PMWS). PMWS is clinically characterized by wasting, paleness of the skin, unthriftiness, respiratory distress, diarrhea, icterus, and jaundice. In some affected swine, a combination of all symptoms will be apparent while other swine will only have one or two of these symptoms. During necropsy, microscopic and macroscopic lesions also appear on multiple tissues and organs, with lymphoid organs being the most common site for lesions. A strong correlation has been observed between the amount of PCV-2 nucleic acid or antigen and the severity of microscopic lymphoid lesions. Mortality rates for swine infected with PCV-2 can approach 80%. In addition to PMWS, PCV-2 has been associated with several other infections including pseudorabies, porcine reproductive and respiratory syndrome (PRRS), Glasser's disease, streptococcal meningitis, salmonellosis, postweaning colibacillosis, dietetic hepatosis, and suppurative bronchopneumonia.

Several vaccine are available to reduced the impact of PCV-2 infections in pigs. U.S. Patent No. 6,703,023 provides a DNA based vaccine for the prophylaxis of pigs against PMWS. In WO 03/049703 production of a live chimeric vaccine is described, comprising the non-pathogenic PCV1 virus in which, however, the ORF2 protein is replaced by the ORF2 protein of the pathogenic PCV-2. WO 99/18214 and WO 99/29717 have provided several PCV-2 strains and procedures for the preparation of a killed PVC2 vaccine. Preparation of subunit vaccines have also been described in WO 99/18214 and WO 99/29717. An effective ORF2 based subunit vaccine has been reported in WO 06/072065. A further ORF-2 based subunit vaccine is described also in WO 07/28823. However, none of the vaccine described in the prior art includes a non-virucidal and/or purified PCV-2 antigen, preferably a highly purified PCV-2 ORF2 antigen.

Immunogenic compositions against PCV-2 and various immunogenic compositions against other pathogens often have a virucidal effect on other antigens. Current regulatory standards (9 CFR 113.35) permit some virucidal activity in multivalent compositions, but this virucidal activity cannot result in a loss of more than 0.7 logs/ml of a live virus or less than 0.7 logs/ml CFU of live bacteria when combined with the other components of the immunogenic composition. Compositions that have more virucidal activity than permitted cannot be combined with other antigens to create a multivalent vaccine.

Open reading frame 2 (ORF2) protein of PCV-2, having an approximate molecular weight of 30 kDa when run on SDS-PAGE gel, has been utilized in the past as an antigenic

component in vaccines and immunogenic compositions for PCV-2. Typical methods of obtaining ORF2 for use in such vaccines and compositions generally consist of amplifying the PCV-2 DNA coding for ORF2, expressing the ORF2 protein within a host cell, and extracting the ORF2 protein from the host cell via cell lysis. The recovered ORF2 cell lysate is then used as the antigenic portion of an immunogenic composition or vaccine. In some cases the ORF2 containing cell lysate is separated from the cell debris.

What is needed is a method for reducing the virucidal activity of PCV-2-containing immunogenic compositions and antigens therein such that regulatory requirements can be met and efficacious multivalent compositions can be administered. What is further needed are methods for decreasing or reducing the virucidal activity and effect of PCV-2-containing compositions on Porcine Reproductive and Respiratory Syndrome Virus (PRRSV). What is still further needed are immunogenic compositions that have undergone the methods of the present invention such that their virucidal activity has been reduced to acceptable standards and can be combined with other antigens to form multivalent immunogenic compositions.

SUMMARY

The practice of the present disclosure will employ, unless otherwise indicated, conventional techniques of molecular biology, microbiology, recombinant DNA technology, protein chemistry and immunology, which are within the skill of the art. Such techniques are explained fully in the literature. See, e.g., Sambrook, Fritsch & Maniatis, Molecular Cloning: A Laboratory Manual, Vols. I, II and III, Second Edition (1989); DNA Cloning, Vols. I and II (D. N. Glover ed. 1985); Oligonucleotide Synthesis (M. J. Gait ed. 1984); Nucleic Acid Hybridization (B. D. Hames & S. J. Higgins eds. 1984); Animal Cell Culture (R. K. Freshney ed. 1986); Immobilized Cells and Enzymes (IRL press, 1986); Perbal, B., A Practical Guide to Molecular Cloning (1984); the series, Methods In Enzymology (S. Colowick and N. Kaplan eds., Academic Press, Inc.); Protein purification methods – a practical approach (E.L.V. Harris and S. Angal, eds., IRL Press at Oxford University Press); and Handbook of Experimental Immunology, Vols. I-IV (D. M. Weir and C. C. Blackwell eds., 1986, Blackwell Scientific Publications).

Before describing the present disclosure in detail, it is to be understood that this disclosure is not limited to particular DNA, polypeptide sequences or process parameters as such may, of course, vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments of the disclosure only, and is not intended to be limiting. It must be noted that, as used in this specification and the appended claims, the

singular forms "a", "an" and "the" include plural referents unless the content clearly dictates otherwise. Thus, for example, reference to "an antigen" includes a mixture of two or more antigens, reference to "an excipient" includes mixtures of two or more excipients, and the like

The present disclosure solves the problems inherent in the prior art and provides a distinct advance in the state of the art. Generally, the present disclosure provides a method of producing a PCV-2 antigenic composition comprising the steps of i) obtaining a first liquid containing a PCV-2 antigen, and ii) removing at least a portion of the first liquid from the PCV-2 antigen. Preferably the PCV-2 antigen is used as or in the PCV-2 antigenic composition.

In a first aspect, there is provided a method of producing a PCV-2 antigenic composition comprising the steps of: i) obtaining a first liquid containing therein PCV-2 antigen; ii) removing at least a portion of the first liquid from the PCV-2 antigen by an exchange of the portion of the first liquid against a second liquid, wherein the second liquid is different from the first liquid; and iii) admixing the PCV-2 antigen remaining after step ii) with a further component selected from the group consisting of pharmaceutically acceptable carriers, adjuvants, diluents, excipients, and combinations thereof.

In a second aspect, there is provided a PCV-2 antigenic composition obtainable by a method as defined above.

In a third aspect, there is provided an immunogenic composition comprising a PCV-2 antigenic composition obtainable by a method as defined above.

In a fourth aspect, there is provided an immunogenic composition comprising the PCV-2 antigenic composition as defined above.

In a fifth aspect, there is provided the immunogenic composition as defined above for reducing one or more clinical symptoms of a PCV-2 infection in an animal as compared to an animal not receiving the immunogenic composition.

For purposes of the present disclosure, a "first liquid" refers to liquid, aqueous, or fluid media typically used in combination with cells, antigens, immunogenic compositions, vaccines, and the like. Preferably, the first liquid comprises media from an antigenic composition, more preferably, the first liquid comprises or preferably consists of cell culture media used for the production of recombinant proteins in cultivated host cells. The cultivated host cells can be bacteria, yeasts, insect cells, animal cells, and mammalian cells, with insect and mammalian cells being particularly preferred. Thus the first fluid may comprise or consist of media for the cultivation of bacteria, yeast, insect cells, animal cells, or mammalian cells. Preferably, the cell media is serum free cell media, and most preferably the culture media is EX-CELL® 420 serum free media, when insect cells are used. EX-CELL® 420 is a complete medium that is protein-free and contains L-glutamine, and was developed and optimized for the serum-free growth of Sf9 and Sf21 insect cell lines.

A "second liquid", for purposes of the present disclosure, refers to any liquid normally used in combination with cells, antigen, immunogenic compositions, vaccines, and the like, which is different from the first liquid. Preferably, the second liquid is an aqueous solution, even more preferably a pharmaceutically acceptable solution, and even more preferably a buffer, such as a saline or phosphate buffer and the like. Most preferably, the second fluid is characterized by not being virucidal to any live virus or any live bacteria (herein, unless explicitly stated or apparent from the context the term "virucidal" is inclusive of bactericidal activity), when the live virus or live bacteria is cultivated in or stored in such a fluid.

"Portion", for purposes of the present disclosure, refers to any amount which does not encompass the entire amount. For example, a portion of liquid would be anything less than 100% of the volume of the liquid, such as 90% of the liquid, 80% of the liquid, 70% of the liquid, and all amounts between more than 0% and less than 100%.

A "PCV-2 antigen" refers to any composition of matter that comprises at least one antigen that can induce, stimulate or enhance the immune response against PCV-2 infection, when administered to an animal, preferably to a pig. Preferably, the PCV-2 antigen is the whole PCV-2 virus, preferably in an inactivated form, a live modified or attenuated PCV-2 virus, a chimeric virus that comprises at least an immunogenic amino acid sequence of PCV-2, or any other polypeptide or component that comprises at least an immunogenic amino acid sequence of PCV-2, preferably ORF2. The terms "immunogenic protein", "immunogenic polypeptide" or "immunogenic amino acid sequence" as used herein refer to any amino acid sequence of PCV-2, which elicits an immune response in a host against PCV-2. Preferably, such immunogenic protein, immunogenic polypeptide or immunogenic amino acid of PCV-2 is any one of those disclosed or provided in the international patent application WO2006/072065 (the contents and teachings of which are hereby incorporated by reference), or is any other PCV-2 polypeptide known in the art. For instance, a representative sequence of PCV-2 ORF2 DNA comprises the nucleotide sequence Genbank Accession No. AF086834 (SEQ ID NO: 3) and SEQ ID NO: 4.

However, it is understood by those of skilled in the art that this sequence could vary by as much as 1-10% in sequence homology and still retain the antigenic characteristics that render it useful in immunogenic compositions. The antigenic characteristics of an immunological composition can be, for example, estimated by the challenge experiment as provided by Example 4 of WO06/072065. Moreover, the antigenic characteristic of a modified antigen is still retained, when the modified antigen confers at least 70%, preferably 80%, more preferably 90% or more of the protective immunity as compared to the PCV-2 ORF2 protein, encoded by the polynucleotide sequence of SEQ ID NO:3 or SEQ ID NO:4 as provided in WO06/072065. Further preferred PCV-2 ORF2 antigens are as follow:

i) a polypeptide comprising the sequence of SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID NO: 9, SEQ ID NO: 10 or SEQ ID NO: 11 of WO06/072065;

ii) any polypeptide that is at least 80% homologous and/or identical to the polypeptide of i),

iii) any immunogenic portion of the polypeptides of i) and/or ii)

iv) the immunogenic portion of iii), comprising at least 5, preferably 8, more preferably 10 contiguous amino acids of any of the sequences of SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID NO: 9, SEQ ID NO: 10 or SEQ ID NO: 11 of WO06/072065,

v) a polypeptide that is encoded by a DNA comprising the sequence of SEQ ID NO: 3 or SEQ ID NO: 4 of WO06/072065.

vi) any polypeptide that is encoded by a polynucleotide that is at least 80% homologous and/or identical to the polynucleotide of v),

vii) any immunogenic portion of the polypeptides encoded by the polynucleotide of v) and/or vi),

viii) the immunogenic portion of vii), wherein the polynucleotide coding for the immunogenic portion comprises at least 30 contiguous nucleotides included in the sequences of SEQ ID NO: 3 or SEQ ID NO: 4 of WO06/072065.

The sequence listing of WO06/072065 is identical with the sequence listing attached to this application.

Preferably any of the immunogenic portions described above having the antigenic characteristics of PCV-2 ORF2 antigen that is encoded by the sequence of SEQ ID NO: 3 or SEQ ID NO: 4 of WO06/072065.

"Sequence Identity" as it is known in the art refers to a relationship between two or more polypeptide sequences or two or more polynucleotide sequences, namely a reference sequence and a given sequence to be compared with the reference sequence. Sequence identity is determined by comparing the given sequence to the reference sequence after the sequences have been optimally aligned to produce the highest degree of sequence similarity, as determined by the match between strings of such sequences. Upon such alignment, sequence identity is ascertained on a position-by-position basis, e.g., the sequences are "identical" at a particular position if at that position, the nucleotides or amino acid residues are identical. The total number of such position identities is then divided by the total number of nucleotides or residues in the reference sequence to give % sequence identity. As an illustration, by a polynucleotide having a nucleotide sequence having at least, for example, 85%, preferably 90%, even more preferably 95% "sequence identity" to a reference nucleotide sequence, it is intended that the nucleotide sequence of the given polynucleotide is identical to the reference sequence except that the given polynucleotide sequence may include up to 15, preferably up to 10, even more preferably up to 5 point mutations per each 100 nucleotides of the reference nucleotide sequence. In other words, in a polynucleotide having a nucleotide sequence having at least 85%, preferably 90%, even more preferably 95% identity relative to the reference nucleotide sequence, up to 15%, preferably 10%, even more preferably 5% of the nucleotides in the reference sequence may be deleted or substituted with another nucleotide, or a number of nucleotides up to 15%, preferably 10%, even more preferably 5% of the total nucleotides in the reference sequence may be inserted into the

reference sequence. These mutations of the reference sequence may occur at the 5' or 3' terminal positions of the reference nucleotide sequence or anywhere between those terminal positions, interspersed either individually among nucleotides in the reference sequence or in one or more contiguous groups within the reference sequence. Analogously, by a polypeptide having a given amino acid sequence having at least, for example, 85%, preferably 90%, even more preferably 95% sequence identity to a reference amino acid sequence, it is intended that the given amino acid sequence of the polypeptide is identical to the reference sequence except that the given polypeptide sequence may include up to 15, preferably up to 10, even more preferably up to 5 amino acid alterations per each 100 amino acids of the reference amino acid sequence. In other words, to obtain a given polypeptide sequence having at least 85%, preferably 90%, even more preferably 95% sequence identity with a reference amino acid sequence, up to 15%, preferably up to 10%, even more preferably up to 5% of the amino acid residues in the reference sequence may be deleted or substituted with another amino acid, or a number of amino acids up to 15%, preferably up to 10%, even more preferably up to 5% of the total number of amino acid residues in the reference sequence may be inserted into the reference sequence. These alterations of the reference sequence may occur at the amino or the carboxy terminal positions of the reference amino acid sequence or anywhere between those terminal positions, interspersed either individually among residues in the reference sequence or in the one or more contiguous groups within the reference sequence. Preferably, residue positions that are not identical differ by conservative amino acid substitutions. However, conservative substitutions are not included as a match when determining sequence identity.

"Live" virus or bacterium, for purposes of the present invention, refers to a virus or bacterium that is capable of replicating in a host. A preferred live virus and a preferred live bacterium of the present invention are the PRRS virus and the *Mycoplasma hyopneumonia* bacterium, respectively. However, the term live virus or live bacterium is not limited to PRRS virus and *Mycoplasma hypneumoniae*, respectively.

The portion of the first liquid can be removed from the PCV-2 antigen by an exchange of the portion of the first liquid against a second liquid, wherein the second liquid is different from the first liquid (see definition of second fluid). Thus according to a further aspect, the present application provides a method of producing a PCV-2 antigenic composition comprising the steps of i) obtaining a first liquid containing a PCV-2 antigen, ii) removing at least a portion of the first liquid from the PCV-2 antigen, wherein the portion of the first liquid is removed from the PCV-2 antigen by an exchange of the portion of the first liquid

against a second liquid, and wherein the second liquid is different from the first liquid. Preferably the exchange of the portion of the first liquid with the second liquid comprises the steps of a) adding the second liquid to the first liquid which contains the PCV-2 antigen and b) concentrating the PCV-2 antigen by removing a portion of the first and second liquids from the PCV-2 antigen. Thus according to a further aspect, the present application provides a method of producing a PCV-2 antigenic composition comprising the steps of i) obtaining a first liquid containing a PCV-2 antigen, ii) removing at least a portion of the first liquid from the PCV-2 antigen by an exchange of at least a portion of the first liquid against a second liquid comprising the steps a) adding the second liquid to the first liquid which contains the PCV-2 antigen and b) concentrating the PCV-2 antigen by removing a portion of the first and second liquids from the PCV-2 antigen.

The portion of the first liquid can be removed from the PCV-2 antigen by a filtration step utilizing a filter. However, any other method known to a person skilled in the art can be used to remove the portion of any fluids, including the first and, whenever applicable, a portion of the second fluid from the PCV-2 antigen. Such method, for instance includes but is not limited to centrifugation and/or chromatography. However, filtration is most preferred. A preferred filtration method to remove the portion of the first fluid, or any other fluid, whenever applicable, comprises ultra- and/or dia-filtration. Ultra- and dia-filtration are standard methods known to a person skilled in the art, described for example in detail in *Protein Purification Methods - A Practical Approach – editors: E.L.V. Harris and S. Angel, Oxford University Press 1995* (the contents and teachings of which are hereby incorporated by reference). In particular, in Chapter 3 of that textbook, several methods and types of equipment are described, all of which can be used by an ordinary person skilled in the art in an exemplary manner for the purpose of the present invention. Thus according to a further aspect, the present application provides a method of producing a PCV-2 antigenic composition comprising the steps of i) obtaining a first liquid containing a PCV-2 antigen, ii) removing at least a portion of the first liquid from the PCV-2 antigen, wherein the portion of the first liquid is removed from the PCV-2 antigen by filtration, preferably by dia- or ultra-filtration. Preferably, the portion of the first liquid is removed from the PCV-2 antigen by an exchange of at least a portion of the first liquid against a second liquid comprising the steps a) adding the second liquid to the first liquid which contains the PCV-2 antigen and b) concentrating the PCV-2 antigen by removing a portion of the first and second liquids from the PCV-2 antigen.

As defined above, a preferred second liquid to be used in any of the methods described is a buffer, preferably a physiologically acceptable buffer with saline being particularly preferred. Thus according to a further aspect, the present application provides a method of producing a PCV-2 antigenic composition comprising the steps of i) obtaining a first liquid containing a PCV-2 antigen, ii) removing at least a portion of the first liquid from the PCV-2 antigen, by an exchange against a buffer, preferably a physiologically acceptable buffer such as saline or phosphate buffer or the like. Preferably the portion of the first liquid is removed from the PCV-2 antigen by filtration, preferably by dia- and/or ultra-filtration. More preferably, the portion the exchange of at least a portion of the first liquid against the buffer, preferably the physiologically acceptable buffer, such as saline or phosphate buffer or the like, comprising the steps a) adding the buffer, preferably the physiologically acceptable buffer, such as saline or phosphate buffer or the like, to the first liquid which contains the PCV-2 antigen and b) concentrating the PCV-2 antigen by removing a portion of the first and the fluid which is a buffer, preferably a physiologically acceptable buffer, such as saline or phosphate buffer or the like, from the PCV-2 antigen, preferably by filtration, even more preferably by dia- and/or ultra-filtration.

The concentrating step and the liquid addition step of the method as described herein can be performed substantially simultaneously or alternatively, the concentrating step and the liquid addition step are performed sequentially. Thus according to a further aspect, the present application provides a method of producing a PCV-2 antigenic composition comprising the steps of i) obtaining a first liquid containing a PCV-2 antigen, ii) removing at least a portion of the first liquid from the PCV-2 antigen by an exchange of a portion of the first liquid against a second liquid comprising the steps of a) adding the second liquid to the first liquid which contains the PCV-2 antigen and b) concentrating the PCV-2 antigen by removing a portion of the first and second liquids from the PCV-2 antigen, wherein the liquid addition step is performed substantially simultaneously or sequentially. Preferably the portion of the first liquid and in the case of the addition of the second liquid, the mixture of the first and the second fluid is removed from the PCV-2 antigen by filtration, preferably by dia- and/or ultra-filtration.

When the concentrating step and liquid addition step are performed sequentially, the order of the steps does not matter. For example, in a further aspect, the liquid addition step occurs prior to the concentrating step and in an alternative aspect, the concentrating step occurs prior to the liquid addition step. The liquid addition step and the concentrating step, regardless of the order in which they are performed, can be performed multiple times. For

example, each of these respective steps can be performed at least two, at least three, at least four, at least five, at least 10, up to as many times as desired. In one aspect, the concentrating step and the liquid addition step are each performed at least two times. In another aspect, the concentrating step and the liquid addition step are each performed at least three times. Thus, according to a further aspect of the present application, a method of producing a PCV-2 antigenic composition is provided wherein the method generally comprises the steps of i) obtaining a first liquid containing a PCV-2 antigen, ii) removing at least a portion of the first liquid from the PCV-2 antigen by an exchange of the portion of the first liquid against a second liquid, wherein the exchange is performed multiple times. Preferably the exchange of the portion of the first fluid against a portion of the second fluid comprises the steps of a) adding the second liquid to the first liquid which contains the PCV-2 antigen and b) concentrating the PCV-2 antigen by removing a portion of the first and second liquids from the PCV-2 antigen, wherein the liquid addition step and concentration step are performed multiple times, for instance, two times, three times, 5 times, 10 times, etc. Preferably, the liquid addition step and concentration step are performed two times, most preferably three times. As described above, filtration is the preferred method to remove a portion of the first liquid, or in case of multiple removing steps as described above, to remove a portion of the mixture of the first and the second fluid, from the PCV-2 antigen.

The filter can be any conventional filter in the art. Preferably, the filter includes a semi-permeable membrane. In a further preferred form, the semi-permeable membrane has an average pore size that is smaller than the PCV-2 antigen to thereby prevent passage of at least 90% of the PCV-2 antigen through the semi-permeable membrane pores and withhold the PCV-2 antigen by the filter. In a further aspect, the filter has an average pore size which prevents passage of at least 90% of proteins of 50 kDa to 500 kDa in size, more preferably, the filter has an average pore size which prevents passage of at least 90% of proteins of 75 kDa to 400 kDa in size, and most preferably, the filter has an average pore size which prevents passage of at least 90% of proteins of 100 kDa to 300 kDa in size. This pore size is preferred, when the PCV-2 antigen is produced as whole virus or as virus like particles. In a still further aspect, the semi-permeable membrane includes a material selected from the group consisting of polysulfone, polyethersulfone, and regenerated cellulose. However, any other material that allows removing of a portion of the first fluid, and in case of a multiple process step, removing of a mixture of the first and the second fluid from the PCV-2 antigen can be used. The filter can be selected from the group consisting of a hollow fiber membrane ultra filtration cartridge, flat sheets, or a cassette, with a hollow fiber membrane ultra filtration

cartridge being particularly preferred. Thus, according to a further aspect of the present application, a method of producing a PCV-2 antigenic composition is provided as described above. The method generally comprises the steps of i) obtaining a first liquid containing a PCV-2 antigen, ii) removing at least a portion of the first liquid from the PCV-2 antigen by a filtration step, wherein the filter preferably is or comprises a semi-permeable membrane. Preferably, the semi-permeable membrane has an average pore size that is smaller than the PCV-2 antigen and prevent passage of at least 90% of the PCV-2 antigen through the semi-permeable membrane pores. Preferably the average pore size of the semi-permeable membrane prevents passage of at least 90% of proteins of 50 kDa to 500 kDa in size, more preferably, at least 90% of proteins of 75 kDa to 400 kDa in size, and most preferably at least 90% of proteins of 100 kDa to 300 kDa in size. This pore size is preferred, when the PCV-2 antigen is produced as whole virus or as virus like particles. As described above, the removing step in general includes the exchange of the portion of the first fluid against a portion of the second fluid comprising the steps of a) adding the second liquid to the first liquid which contains the PCV-2 antigen and b) concentrating the PCV-2 antigen by removing a portion of the first and second liquids from the PCV-2 antigen, wherein the liquid addition step and concentration step are performed multiple times, for instance, two times, three times, 5 times, 10 times, etc. Preferably, the liquid addition step and the concentration step are performed two times, most preferably three times.

The concentration step of the method provided herein is performed such that the PCV-2 antigen is concentrated from 3X to 50X in comparison to the volume of the first liquid. More preferably, the concentrating step is done such that the PCV-2 antigen is concentrated 4X to 20X in comparison to the volume of the first liquid. Most preferably, concentration step is done such that the PCV-2 antigen is concentrated from 7X to 10X in comparison to the volume of the first liquid. Thus according to a further aspect, the present application provides a method of producing a PCV-2 antigenic composition comprising the steps of i) obtaining a first liquid containing a PCV-2 antigen, ii) removing at least a portion of the first liquid from the PCV-2 antigen, wherein the portion of the first liquid is removed from the PCV-2 antigen, and wherein the PCV-2 antigen is concentrated from 3X to 50X, preferably from 4X to 20X, and even more preferably from 7X to 10X in comparison to the volume of the first liquid. Preferably, the portion of the first fluid is removed from the PCV-2 antigen by an exchange of the portion of the first liquid against a second liquid comprising the steps of a) adding the second liquid to the first liquid which contains the PCV-2 antigen and b) concentrating the PCV-2 antigen from 3X to 50X, preferably from 4X to 20X, and even more preferably from

7X to 10X in comparison to the volume of the first liquid by removing a portion of the first and second liquids from the PCV-2 antigen. Preferably, the liquid addition step and concentration step are performed multiple times, preferably two times, even more preferably three times. In such case, not only the first liquid is removed, but also a mixture of the first and second liquid. Preferably each liquid addition step is performed substantially simultaneously or sequentially. When the concentrating step and liquid addition step are performed sequentially, the order of the steps does not matter. Moreover, the concentration step is preferably done by filtration - preferably dia- and/or ultra-filtration, utilizing a filter, which preferably contains a semi-permeable membrane. The semi-permeable membrane preferably has an average pore size that is smaller than the PCV-2 antigen and prevent passage of at least 90% of the PCV-2 antigen through the semi-permeable membrane pores. Preferably the average pore size of the semi-permeable membrane is prevents passage of at least 90% of proteins of 50 kDa to 500 kDa in size, more preferably, at least 90% of proteins of 75 kDa to 400 kDa in size, and most preferably at least 90% of proteins of 100 kDa to 300 kDa in size. This pore size is preferred, when the PCV-2 antigen is produced as whole virus or as virus like particles.

In a further aspect, the virucidal activity of the PCV-2 antigenic composition produced by the methods herein is reduced by at least 10% as compared to the liquid that has not undergone the method. More preferably, the virucidal activity of the PCV-2 antigenic composition is reduced by at least 50% as compared to the first liquid that has not undergone the method. Still more preferably, the virucidal activity of the PCV-2 antigenic composition is reduced by at least 70% as compared to the first liquid that has not undergone the method.

For the purpose of the current invention the term "virucidal activity" means, that a fluid, solution or composition inactivates or kills a live virus or live bacteria to a certain extent, when the fluid, solution or composition is mixed with such live virus or live bacteria. Thus, a reduction of the virucidal activity of a fluid, solution or composition by at least 10% means, that the survival rate of a live virus or live bacteria is 90% higher in a fluid, solution or composition that has undergone any of the methods described herein, as compared to a fluid, solution or composition, that has not undergone any of the method described herein. According to the present invention, the PRRS virus, preferably PRRS virus having the ATCC accession number VR 2332, is the reference virus for the determination of virucidal activity. To determine the virucidal activity with regard to a bacterium, it is proposed to use the *Mycoplasma hyopneumonia* bacterium, preferably the J-strain of *Mycoplasma hyopneumonia.*

Thus according to a further aspect, the present application provides a method of producing a PCV-2 antigenic composition comprising the steps of i) obtaining a first liquid containing a PCV-2 antigen, ii) removing at least a portion of the first liquid from the PCV-2 antigen, wherein the virucidal activity - preferably in respect to PRRS virus - of the PCV-2 antigenic composition obtained after step ii) is reduced by at least 10%, preferably at least 50%, more preferably at least 70%, even more preferably at least 90% as compared to that of the first liquid. Preferably, the portion of the first liquid having virucidal activity is removed from the PCV-2 antigen by an exchange of a portion of the first liquid against a second liquid. The exchange is preferably done in such a manner that it comprises the steps of a) adding the second liquid to the first liquid which contains the PCV-2 antigen and b) concentrating the PCV-2 antigen, preferably from 3X to 50X, even more preferably from 4X to 20X, and even more preferably from 7X to 10X in comparison to the volume of the first liquid by removing a portion of the first and second liquids from the PCV-2 antigen. Preferably, the liquid addition step and concentration step are performed multiple times, preferably two times, and even more preferably three times. In such case, not only the first liquid is removed, but also a mixture of the first and second liquid. Preferably each liquid addition step is performed substantially simultaneously or sequentially as described above. When the concentrating step and liquid addition step are performed sequentially, the order of the steps does not matter. Moreover, the concentration step is preferably done by filtration - preferably by dia- and/or ultra-filtration, utilizing a filter, which preferably contains a semi-permeable membrane. The semi-permeable membrane preferably has an average pore size that is smaller than the PCV-2 antigen and prevents passage of at least 90% of the PCV-2 antigen through the semi-permeable membrane pores. Preferably the average pore size of the semi-permeable membrane or of any other filter that is used herein, prevents passage of at least 90% of proteins of 50 kDa to 500 kDa in size, more preferably, at least 90% of proteins of 75 kDa to 400 kDa in size, and most preferably at least 90% of proteins of 100 kDa to 300 kDa in size. This pore size is preferred, when the PCV-2 antigen is produced as whole virus or as virus like particles.

In a further aspect, the method further comprises the step of harvesting the PCV-2 antigen obtained after at least a portion of the first liquid is removed from the PCV-2 antigen.

As used herein, "harvesting" or "harvest" refers to the collecting or recovering of the PCV-2 antigen. Any conventional method known in the art can be used to recover the PCV-2 antigen either when an antigen is being produced for use with the methods and compositions of the present application, or when the PCV-2 antigen is undergoing the methods described

herein. In a particularly preferred manner of harvesting, the portion of the first liquid is removed from the PCV-2 antigen via a filtration step and the PCV-2 antigen is recovered or harvested from the filter retard. In a more preferred form, the PCV-2 antigen is harvested or collected, or recovered from the retard of a semi-permeable membrane having the pore size described herein. Thus, according to a further aspect, the present application provides a method of producing a PCV-2 antigenic composition comprising the steps of i) obtaining a first liquid containing a PCV-2 antigen, ii) removing at least a portion of the first liquid from the PCV-2 antigen, wherein the PCV-2 antigen obtained after the step ii) is harvested. Preferably, the portion of the first liquid is removed from the PCV-2 antigen by an exchange of a portion of the first liquid against a second liquid. The exchange is preferably done such that it comprises the steps of a) adding a second liquid to the first liquid which contains the PCV-2 antigen and b) concentrating the PCV-2 antigen, preferably from 3X to 50X, even more preferably from 4X to 20X, and even more preferably from 7X to 10X in comparison to the volume of the first liquid by removing a portion of the first and second liquids from the PCV-2 antigen. Preferably, the liquid addition step and concentration step are performed multiple times, preferably two times, even more preferably three times. In such cases, not only the first liquid is removed, but also a mixture of the first and second liquid. Preferably each liquid addition step is performed substantially simultaneously or sequentially as described above. When the concentrating step and liquid addition step are performed sequentially, the order of the steps does not matter. Moreover, the concentration step is preferably done by filtration - preferably by dia- and/or ultra-filtration, utilizing a filter, which preferably contains a semi-permeable membrane. The semi-permeable membrane preferably has an average pore size that is smaller than the PCV-2 antigen and prevent passage of at least 90% of the PCV-2 antigen through the semi-permeable membrane pores and withholds the PCV-2 antigen within the filter for harvesting or recovery. Preferably the average pore size of the semi-permeable membrane or of any other filter that is used herein, prevents passage of at least 90% of proteins of 50 kDa to 500 kDa in size, more preferably, at least 90% of proteins of 75 kDa to 400 kDa in size, and most preferably at least 90% of proteins of 100 kDa to 300 kDa in size. This pore size is preferred, when the PCV-2 antigen is produced as whole virus or as virus like particles.

The PCV-2 antigen remaining after undergoing the methods provided herein, preferably after being harvested from the filter retard, is admixed with a further component selected from the group consisting of pharmaceutically acceptable carriers, adjuvants, diluents, excipients, and combinations thereof. Preferably, the further component is an

adjuvant, even more preferably wherein the adjuvant is a polymer of acrylic or methacrylic acid, and still more preferably wherein the adjuvant is Carbomer (the generic name for synthetic high molecular weight polymers of acrylic acid).

As used herein, "a pharmaceutical-acceptable carrier" and a "veterinary acceptable carrier" includes any and all solvents, dispersion media, coatings, stabilizing agents, diluents, preservatives, antibacterial and antifungal agents, isotonic agents, adsorption delaying agents, and the like.

"Adjuvants" as used herein, can include aluminum hydroxide and aluminum phosphate, saponins e.g., Quil A, QS-21 (Cambridge Biotech Inc., Cambridge MA), GPI-0100 (Galenica Pharmaceuticals, Inc., Birmingham, AL), water-in-oil emulsion, oil-in-water emulsion, water-in-oil-in-water emulsion. The emulsion can be based in particular on light liquid paraffin oil (European Pharmacopea type); isoprenoid oil such as squalane or squalene ; oil resulting from theoligomerization of alkenes, in particular of isobutene or decene; esters of acids or of alcohols containing a linear alkyl group, more particularly plant oils, ethyl oleate, propylene glycol di-(caprylate/caprate), glyceryl tri-(caprylate/caprate) or propylene glycol dioleate; esters of branched fatty acids or alcohols, in particular isostearic acid esters. The oil is used in combination with emulsifiers to form the emulsion. The emulsifiers are preferably nonionic surfactants, in particular esters of sorbitan, of mannide (e.g. anhydromannitol oleate), of glycol, of polyglycerol, of propylene glycol and of oleic, isostearic, ricinoleic or hydroxystearic acid, which are optionally ethoxylated, and polyoxypropylene-polyoxyethylene copolymer blocks, in particular the Pluronic products, especially L121. See Hunter et al., The Theory and Practical Application of Adjuvants (Ed.Stewart-Tull, D. E. S.). JohnWiley and Sons, NY, pp51-94 (1995) and Todd et al., Vaccine 15:564-570 (1997). For example, it is possible to use the SPT emulsion described on page 147 of "Vaccine Design, The Subunit and Adjuvant Approach" edited by M. Powell and M. Newman, Plenum Press, 1995, and the emulsion MF59 described on page 183 of this same book. Further suitable adjuvants include, but are not limited to, the RIBI adjuvant system (Ribi Inc.), Block co-polymer (CytRx, Atlanta GA), SAF-M (Chiron, Emeryville CA), monophosphoryl lipid A, Avridine lipid-amine adjuvant, heat-labile enterotoxin from E. coli (recombinant or otherwise), cholera toxin, IMS 1314 or muramyl dipeptide among many others. Among the copolymers of maleic anhydride and alkenyl derivative, the copolymers EMA (Monsanto), which are copolymers of maleic anhydride and ethylene, are included. The dissolution of these polymers in water leads to an acid solution that will be neutralized, preferably to

physiological pH, in order to give the adjuvant solution into which the immunogenic, immunological or vaccine composition itself will be incorporated.

A further instance of an adjuvant is a compound chosen from the polymers of acrylic or methacrylic acid and the copolymers of maleic anhydride and alkenyl derivative. Advantageous adjuvant compounds are the polymers of acrylic or methacrylic acid which are cross-linked, especially with polyalkenyl ethers of sugars or polyalcohols. These compounds are known by the term carbomer (Phameuropa Vol. 8, No. 2, June 1996). Persons skilled in the art can also refer to U. S. Patent No. 2,909,462 which describes such acrylic polymers cross-linked with a polyhydroxylated compound having at least 3 hydroxyl groups, preferably not more than 8, the hydrogen atoms of at least three hydroxyls being replaced by unsaturated aliphatic radicals having at least 2 carbon atoms. The preferred radicals are those containing from 2 to 4 carbon atoms, e.g. vinyls, allyls and other ethylenically unsaturated groups. The unsaturated radicals may themselves contain other substituents, such as methyl. The products sold under the name CARBOPOL®; (BF Goodrich, Ohio, USA) are particularly appropriate. They are polymers of acrylic acid cross-linked with polyalkenyl ethers or divinyl glycol or cross-linked with an allyl sucrose or with allyl pentaerythritol. Among them, there may be mentioned CARBOPOL® 974P, 934P and 971P. Most preferred is the use of CARBOPOL® 971P.

Preferably, the adjuvant is added in an amount of about 100 µg to about 10 mg per dose. Even more preferably the adjuvant is added in an amount of about 100 µg to about 10 mg per dose. Still more preferably the adjuvant is added in an amount of about 500 µg to about 5 mg per dose. Still more preferably the adjuvant is added in an amount of about 750 µg to about 2.5 mg per dose. Most preferably the adjuvant is added in an amount of about 1 mg per dose.

"Diluents" can include water, saline, dextrose, ethanol, glycerol, and the like. Isotonic agents can include sodium chloride, dextrose, mannitol, sorbitol, and lactose, among others. Stabilizers include albumin and alkali salts of ethylendiamintetracetic acid, among others.

A "preservative" as used herein refers to an anti-microbiological active agent, such as for example Gentamycin, Merthiolate, and the like. In particular adding of a preservative is most preferred for the preparation of a multi-dose composition. Those anti-microbiological active agents are added in concentrations effective to prevent the composition of interest for any microbiological contamination or for inhibition of any microbiological growth within the composition of interest.

Thus, according to a further aspect, the present application provides a method of producing a PCV-2 antigenic composition comprising the steps of i) obtaining a first liquid containing a PCV-2 antigen, ii) removing at least a portion of the first liquid from the PCV-2 antigen, further comprising the step of admixing the PCV-2 antigen remaining after step ii) with a further component selected from the group consisting of pharmaceutically acceptable carriers, adjuvants, diluents, excipients, and combinations thereof. Preferably wherein the further component is an adjuvant, even more preferably wherein the adjuvant is a polymer of acrylic or methacrylic acid, and still more preferably wherein the adjuvant is Carbomer. Preferably, the portion of the first liquid is removed from the PCV-2 antigen by an exchange of a portion of the first liquid against a second liquid. The exchange is preferably done such that it comprises the steps of a) adding the second liquid to the first liquid which contains the PCV-2 antigen and b) concentrating the PCV-2 antigen, preferably from 3X to 50X, even more preferably from 4X to 20X, and even more preferably from 7X to 10X in comparison to the volume of the first liquid by removing a portion of the first and second liquids from the PCV-2 antigen. Preferably, the liquid addition step and the concentration step are performed multiple times, preferably two times, and even more preferably three times. In such cases, not only the first liquid is removed, but also a mixture of the first and second liquid. Preferably each liquid addition step is performed substantially simultaneously or sequentially as described above. When the concentrating step and liquid addition step are performed sequentially, the order of the steps does not matter. Moreover, the concentration step is preferably done by filtration - preferably by dia- and/or ultrafiltration, utilizing a filter, which preferably contains a semi-permeable membrane. The semi-permeable membrane preferably has an average pore size that is smaller than the PCV-2 antigen and prevents passage of at least 90% of the PCV-2 antigen through the semi-permeable membrane pores and withholds the PCV-2 antigen within the filter for harvesting or recovery. Preferably the average pore size of the semi-permeable membrane or of any other filter that is used herein, prevents passage of at least 90% of proteins of 50 kDa to 500 kDa in size, more preferably, at least 90% of proteins of 75 kDa to 400 kDa in size, and most preferably at least 90% of proteins of 100 kDa to 300 kDa in size. This pore size is preferred, when the PCV-2 antigen is produced as whole virus or as virus like particles.

The PCV-2 antigen used in the methods described above can be any PCV-2 antigen as defined herein. Preferably the PCV-2 antigen comprises the ORF-2 protein of PCV-2, more preferably recombinant ORF-2 protein of PCV-2, and still more preferably virus like particles of ORF-2 protein, and even more preferably the antigen included in INGELVAC

CIRCOFLEX®. Thus, according to a further aspect of the present application, the present application provides a method of producing a PCV-2 antigenic composition comprising the steps of i) obtaining a first liquid containing a PCV-2 antigen, ii) removing at least a portion of the first liquid from the PCV-2 antigen, wherein the PCV-2 antigen comprises the ORF-2 protein of PCV-2, more preferably recombinant ORF-2 protein of PCV-2, and still more preferably virus like particles of ORF-2 protein. Preferably, the portion of the first liquid is removed from the PCV-2 antigen by an exchange of a portion of the first liquid against a second liquid. The exchange is preferably done such that it comprises the steps of a) adding the second liquid to the first liquid which contains the PCV-2 antigen and b) concentrating the PCV-2 antigen, preferably from 3X to 50X, even more preferably from 4X to 20X, and even more preferably from 7X to 10X in comparison to the volume of the first liquid by removing a portion of the first and second liquids from the PCV-2 antigen.

Preferably, the liquid addition step and concentration step are performed multiple times, preferably two times, and even more preferably three times. In such cases, not only the first liquid is removed, but also a mixture of the first and second liquid. Preferably each liquid addition step is performed substantially simultaneously or sequentially as described above. When the concentrating step and liquid addition step are performed sequentially, the order of the steps does not matter. Moreover, the concentration step is preferably done by filtration - preferably by dia- and/or ultrafiltration, utilizing a filter, which preferably contains a semi-permeable membrane. The semi-permeable membrane preferably has an average pore size that is smaller than the PCV-2 antigen and prevents passage of at least 90% of the PCV-2 antigen through the semi-permeable membrane pores and withholds the PCV-2 antigen within the filter for harvesting or recovery. Preferably the average pore size of the semi-permeable membrane or of any other filter that is used herein, prevents passage of at least 90% of proteins of 50 kDa to 500 kDa in size, more preferably, at least 90% of proteins of 75 kDa to 400 kDa in size, and most preferably at least 90% of proteins of 100 kDa to 300 kDa in size. This pore size is preferred, when the PCV-2 antigen is produced as whole virus or as virus like particles.

The first liquid containing the PCV-2 antigen used can be obtained by any method known in the art. Preferably, the first liquid containing the PCV-2 antigen as well as PCV-2 antigen can be obtained by any of the methods described in the international patent application WO2006/072065 (the contents and teachings of which are hereby incorporated by reference). In particular, the PCV-2 antigen, when expressed recombinantly *in vitro* in host

cells, can be obtained via a viral vector, preferably a recombinant baculovirus viral vector, containing and expressing the PCV-2 antigen, preferably, PCV-2 ORF-2.

Vectors and methods for making and/or using vectors (or recombinants) for expression of the PCV-2 antigen, preferably the PCV-2 ORF2 antigen can be by or analogous to the methods disclosed in: U.S. Patent Nos. 4,603,112, 4,769,330, 5,174,993, 5,505,941, 5,338,683, 5,494,807, 4,722,848, 5,942,235, 5,364,773, 5,762,938, 5,770,212, 5,942,235, 382,425, PCT publications WO 94/16716, WO 96/39491, WO 95/30018, Paoletti, "Applications of pox virus vectors to vaccination: An update, "PNAS USA 93: 11349-11353, October 1996, Moss, "Genetically engineered poxviruses for recombinant gene expression, vaccination. and safety," PNAS USA 93: 11341-11348, October 1996, Smith et al., U. S. Patent No. 4,745,051, (recombinant baculovirus), Richardson, C.D. (Editor), Methods in Molecular Biology 39, "Baculovirus Expression Protocols" (1995 Humana Press Inc.), Smith et al., "Production of Human Beta Interferon in Insect Cells Infected with a Baculovirus Expression Vector", Molecular and Cellular Biology, Dec., 1983, Vol. 3, No. 12, p. 2156-2165; Pennock et al., "Strong and Regulated Expression of Escherichia coli B-Galactosidase in Infect Cells with a Baculovirus vector, "Molecular and Cellular Biology Mar. 1984, Vol. 4, No. 3, p. 399-406; EPA0 370 573, U. S. application No. 920,197, filed October 16,1986, EP Patent publication No. 265785, U. S. Patent No. 4,769,331 (recombinant herpesvirus), Roizman, "The function of herpes simplex virus genes: A primer for genetic engineering of novel vectors," PNAS USA 93:11307-11312, October 1996, Andreansky et al., "The application of genetically engineered herpes simplex viruses to the treatment of experimental brain tumors," PNAS USA 93: 11313-11318, October 1996, Robertson et al. "Epstein-Barr virus vectors for gene delivery to B lymphocytes", PNAS USA 93: 11334-11340, October 1996, Frolov et al., "Alphavirus-based expression vectors: Strategies and applications," PNAS USA 93: 11371-11377, October 1996, Kitson et al., J. Virol. 65,3068-3075,1991; U. S. Patent Nos. 5,591,439, 5,552,143, WO 98/00166, allowed U. S. applications Serial Nos. 08/675,556, and 08/675,566 both filed July 3,1996 (recombinant adenovirus), Grunhaus et al., 1992,"Adenovirus as cloning vectors," Seminars in Virology (Vol. 3) p. 237-52, 1993, Ballay et al. EMBO Journal, vol. 4, p. 3861-65,Graham, Tibtech 8,85-87, April, 1990, Prevec et al., J. Gen Virol. 70,42434, PCT WO 91/11525, Felgner et al. (1994), J. Biol. Chem. 269,2550-2561, Science, 259: 1745-49,1993 and McClements et al., "Immunization with DNA vaccines encoding glycoprotein D or glycoprotein B, alone or in combination, induces protective immunity in animal models of herpes simplex virus-2 disease", PNAS USA 93: 11414-11420, October 1996, and U. S. Patent Nos. 5,591,639, 5,589,466, and 5,580,859, as well as WO

90/11092, WO93/19183, WO94/21797, WO95/11307, WO95/20660, Tang et al., Nature and Furth et al. Analytical Biochemistry, relating to DNA expression vectors, inter alia. See also WO 98/33510; Ju et al., Diabetologia, 41: 736-739,1998 (lentiviral expression system); Sanford et al., U. S. Patent No. 4,945,050; Fischbachet al. (Intracel), WO 90/01543; Robinson et al., seminars in Immunology vol. 9, pp. 271-283 (1997), (DNA vector systems); Szoka et al., U. S. Patent No. (method of inserting DNA into living cells); McCormick et al., U. S. Patent No. 5,677,178 (use of cytopathic viruses); and U. S. Patent No. 5,928,913 (vectors for gene delivery), as well as other documents cited herein. The expression of PCV-2 ORF2 antigen in insect cells is described, for instance, in WO 06/072065. The purified PCV-2 ORF2 antigen according to the invention can be obtained by several methods known in the art. Preferred methods are those described herein. The PCV-2 ORF2 antigen can be produced recombinantly *in vitro* by the method comprising the steps **i)** permitting infection of susceptible cells in culture with a recombinant viral vector containing PCV-2 ORF2 coding sequence, wherein the PCV-2 ORF2 protein is expressed by the recombinant viral vector, and **ii)** thereafter recovering the PCV-2 ORF2 antigen from cell culture. The PCV-2 ORF2 antigen is recovered by harvesting the whole (i.e. intact) SF+ cells expressing the PCV-2 ORF2 antigen.

Thus, according to a further aspect of the present application, the present application provides a method of producing a PCV-2 antigenic composition comprising the steps of i) obtaining a first liquid containing a PCV-2 antigen, ii) removing at least a portion of the first liquid from the PCV-2 antigen, wherein the PCV-2 antigen is obtained via a viral vector, preferably a recombinant baculovirus viral vector, containing and expressing the PCV-2 antigen, preferably, PCV-2 ORF-2, and wherein the PCV-2 antigen comprises the ORF-2 protein of PCV-2, more preferably recombinant ORF-2 protein of PCV-2, and still more preferably virus like particles of ORF-2 protein. When a viral vector, in particular a recombinant baculovirus containing and expressing the PCV-2 antigen is used to produce/obtain the PCV-2 antigen, the method described above further comprises the step of inactivating the viral vector, preferably the recombinant baculovirus viral vector with a DNA inactivating agent, preferably in the presence of about 1 to about 20 mM of binary ethylenimine. Preferably, the inactivating step is performed after at least a portion of the first liquid is removed from the PCV-2 antigen, more preferably after the PCV-2 antigen is harvested. Even more preferably, the inactivating step is performed after the portion of the first liquid is removed from the PCV-2 antigen by an exchange of a portion of the first liquid against a second liquid. When the exchange of a portion of the first liquid against a second

liquid is done such that it comprises the steps of a) adding the second liquid to the first liquid which contains the PCV-2 antigen and b) concentrating the PCV-2 antigen, preferably from 3X to 50X, even more preferably from 4X to 20X, and even more preferably from 7X to 10X in comparison to the volume of the first liquid by removing a portion of the first and second liquids from the PCV-2 antigen, the inactivating step is done after the concentration step. When the liquid addition step and concentration step are performed multiple times, preferably two times, even more preferably three times, the inactivation step is performed after the last liquid addition step and concentration step. When the concentration step is done by filtration - preferably by dia- and/or ultra-filtration, utilizing a filter, preferably containing a semi-permeable membrane, the inactivation step is performed after the filtration step described above, preferably utilizing a semi-permeable membrane. The semi-permeable membrane preferably has an average pore size that is smaller than the PCV-2 antigen and prevent passage of at least 90% of the PCV-2 antigen through the semi-permeable membrane pores and withhold the PCV-2 antigen within the filter for harvesting or recovery. Preferably the average pore size of the semi-permeable membrane or of any other filter that is used herein, prevents passage of at least 90% of proteins of 50 kDa to 500 kDa in size, more preferably, at least 90% of proteins of 75 kDa to 400 kDa in size, and most preferably at least 90% of proteins of 100 kDa to 300 kDa in size. This pore size is preferred, when the PCV-2 antigen is produced as whole virus or as virus like particles.

"DNA inactivating agent", for purposes of the present invention, refers to any chemical agent which deactivates the DNA, preferably, DNA of a pathogen, such that the pathogen cannot cause active infection or be infective or replicate, but is still capable of inducing an immune response in a subject. Preferably, the DNA inactivating agent is formalin.

Thus, according to a further aspect, the present application provides a method of producing a PCV-2 antigenic composition comprising the steps of i) obtaining a first liquid containing a PCV-2 antigen, ii) removing at least a portion of the first liquid from the PCV-2 antigen, wherein the PCV-2 antigen is obtained via a viral vector, preferably a recombinant baculovirus viral vector, containing and expressing the PCV-2 antigen, preferably, PCV-2 ORF-2, wherein the method further comprises the step of inactivating the viral vector, preferably the recombinant baculovirus viral vector with a DNA inactivating agent, preferably in the presence of about 1 to about 20 mM of binary ethylenimine, and wherein the PCV-2 antigen comprises the ORF-2 protein of PCV-2, more preferably recombinant ORF-2 protein of PCV-2, and still more preferably virus like particles of ORF-2 protein. Preferably, the

inactivating step is performed after at least a portion of the first liquid is removed from the PCV-2 antigen, more preferably after the PCV-2 antigen is harvested. Even more preferably, the inactivating step is performed after the portion of the first liquid is removed from the PCV-2 antigen by an exchange of a portion of the first liquid against a second liquid. When the exchange of a portion of the first liquid against a second liquid is done such that it comprises the steps of a) adding the second liquid to the first liquid which contains the PCV-2 antigen and b) concentrating the PCV-2 antigen, preferably from 3X to 50X, even more preferably from 4X to 20X, even more preferably from 7X to 10X in comparison to the volume of the first liquid by removing a portion of the first and second liquids from the PCV-2 antigen, the inactivating step is done after the concentration step. When the liquid addition step and concentration step are performed multiple times, preferably two times, even more preferably three times, such inactivation step is performed after the last liquid addition step and concentration step. When the concentration step is done by filtration - preferably by dia- and/or ultra-filtration, utilizing a filter, preferably containing a semi-permeable membrane, the inactivation step is performed after the filtration step described above, preferably utilizing a semi-permeable membrane. The semi-permeable membrane preferably has an average pore size that is smaller than the PCV-2 antigen and prevents passage of at least 90% of the PCV-2 antigen through the semi-permeable membrane pores and withholds the PCV-2 antigen within the filter for harvesting or recovery. Preferably the average pore size of the semi-permeable membrane or of any other filter that is used herein, prevents passage of at least 90% of proteins of 50 kDa to 500 kDa in size, more preferably, at least 90% of proteins of 75 kDa to 400 kDa in size, and most preferably at least 90% of proteins of 100 kDa to 300 kDa in size. This pore size is preferred, when the PCV-2 antigen is produced as whole virus or as virus like particles.

In the case that a DNA inactivating agent is used in the method according to the invention the method further comprises the step of adding an amount of an agent that neutralizes the DNA inactivating agent, the amount being equivalent to the amount of the DNA inactivating agent wherein the agent that neutralizes the DNA inactivating agent comprises a sodium thiosulfate solution concentrated to a final concentration of about 1 to about 20 mM and wherein the DNA inactivating agent is BEI. Preferably, the inactivating step is performed after at least a portion of the first liquid is removed from the PCV-2 antigen.

"Agent that neutralizes the inactivating agent" or "neutralizing agent", as used herein, refers to any agent capable of neutralizing the inactivating agents listed above such that the

inactivating agent is no longer capable of inactivating DNA. The agent that neutralizes the inactivating agent is preferably sodium thiosulfate.

Thus, according to a further aspect, the present application provides a method of producing a PCV-2 antigenic composition comprising the steps of i) obtaining a PCV-2 antigen in a first liquid wherein the PCV-2 antigen is obtained via a viral vector, preferably a recombinant baculovirus viral vector, containing and expressing the PCV-2 antigen, preferably, PCV-2 ORF-2, and wherein the PCV-2 antigen comprises the ORF-2 protein of PCV-2, more preferably recombinant ORF-2 protein of PCV-2, and still more preferably virus like particles of ORF-2 protein; ii) removing at least a portion of the first liquid from the PCV-2 antigen; iii) inactivating the recombinant baculovirus viral vector with a DNA inactivating agent, preferably in the presence of about 1 to about 20 mM of binary ethylenimine; iv) adding an amount of a neutralizing agent that neutralizes the inactivating agent, the amount of neutralizing agent being equivalent to the amount of the inactivating agent, wherein the neutralizing agent preferably comprises a sodium thiosulfate solution preferably concentrated to a final concentration of about 1 to about 20 mM and wherein the inactivating agent preferably comprises BEI. Preferably, the inactivating and neutralization step is performed after at least a portion of the first liquid is removed from the PCV-2 antigen, more preferably after the PCV-2 antigen is harvested. Even more preferably, the inactivating and neutralization step is performed after the portion of the first liquid is removed from the PCV-2 antigen by an exchange of a portion of the first liquid against a second liquid. When the exchange of a portion of the first liquid against a second liquid is done such that it comprises the steps of a) adding the second liquid to the first liquid which contains the PCV-2 antigen and b) concentrating the PCV-2 antigen, preferably from 3X to 50X, even more preferably from 4X to 20X, and even more preferably from 7X to 10X in comparison to the volume of the first liquid by removing a portion of the first and second liquids from the PCV-2 antigen, the inactivating and neutralization step is done after the concentration step. When the liquid addition step and concentration step are performed multiple times, preferably two times, even more preferably three times, the inactivation and neutralization step is performed after the last liquid addition step and concentration step. When the concentration step is done by filtration - preferably by dia- and/or ultrafiltration, utilizing a filter, preferably containing a semi-permeable membrane, the inactivation and neutralization step is performed after the filtration step described above, preferably utilizing a semi-permeable membrane. The semi-permeable membrane preferably has an average pore size that is smaller than the PCV-2 antigen and prevents passage of at least 90% of the PCV-2 antigen through the semi-

permeable membrane pores and withholds the PCV-2 antigen within the filter for harvesting or recovery. Preferably the average pore size of the semi-permeable membrane or of any other filter that is used herein, prevents passage of at least 90% of proteins of 50 kDa to 500 kDa in size, more preferably, at least 90% of proteins of 75 kDa to 400 kDa in size, and most preferably at least 90% of proteins of 100 kDa to 300 kDa in size. This pore size is preferred, when the PCV-2 antigen is produced as whole virus or as virus like particles.

In a further aspect of the present application, the method described above further comprises the steps admixing the PCV-2 antigen obtained after the inactivating and neutralizing steps with a further component selected from the group consisting of pharmaceutically acceptable carriers, adjuvants, diluents, excipients, and combinations thereof. Thus, according to a further aspect, the present application provides a method of producing a PCV-2 antigenic composition comprising the steps of i) obtaining a PCV-2 antigen in a first liquid, wherein the PCV-2 antigen is obtained via a viral vector, preferably a recombinant baculovirus viral vector, containing and expressing the PCV-2 antigen, preferably, PCV-2 ORF-2, and wherein the PCV-2 antigen comprises the ORF-2 protein of PCV-2, more preferably recombinant ORF-2 protein of PCV-2, and still more preferably virus like particles of ORF-2 protein; ii) removing at least a portion of the first liquid from the PCV-2 antigen; iii) inactivating the recombinant baculovirus viral vector with a DNA inactivating agent, preferably in the presence of about 1 to about 20 mM of binary ethylenimine; iv) adding an amount of a neutralizing agent that neutralizes the inactivating agent, the amount of neutralizing agent preferably being equivalent to the amount of the inactivating agent, wherein the neutralizing agent preferably comprises a sodium thiosulfate solution preferably concentrated to a final concentration of about 1 to about 20 mM and wherein the inactivating agent preferably comprises BEI; and v) admixing the PCV-2 antigen obtained in step iv) with a further component selected from the group consisting of pharmaceutically acceptable carriers, adjuvants, diluents, excipients, and combinations thereof. Preferably, the PCV-2 antigen comprises the ORF-2 protein of PCV-2, more preferably recombinant ORF-2 protein of PCV-2, and still more preferably virus like particles of ORF-2 protein. Preferably, in step ii), the portion of the first liquid is removed from the PCV-2 antigen by an exchange of a portion of the first liquid against a second liquid. The exchange is preferably done such that it comprises the steps of a) adding the second liquid to the first liquid which contains the PCV-2 antigen and b) concentrating the PCV-2 antigen, preferably from 3X to 50X, even more preferably from 4X to 20X, and even more preferably from 7X to 10X in comparison to the volume of the first liquid by removing a portion of the

first and second liquids from the PCV-2 antigen. Preferably, the liquid addition step and concentration step are performed multiple times, preferably two times, even more preferably three times. In such case, not only the first liquid is removed, but also a mixture of the first and second liquid. Preferably each liquid addition step is performed substantially simultaneously or sequentially as described above. When the concentrating step and liquid addition step are performed sequentially, the order of the steps does not matter. Moreover, the concentration step is preferably done by filtration - preferably by dia- and/or ultrafiltration, utilizing a filter, which preferably contains a semi-permeable membrane. The semi-permeable membrane preferably has an average pore size that is smaller than the PCV-2 antigen and prevents passage of at least 90% of the PCV-2 antigen through the semi-permeable membrane pores and withholds the PCV-2 antigen within the filter for harvesting or recovery. Preferably the average pore size of the semi-permeable membrane or of any other filter that is used herein, prevents passage of at least 90% of proteins of 50 kDa to 500 kDa in size, more preferably, at least 90% of proteins of 75 kDa to 400 kDa in size, and most preferably at least 90% of proteins of 100 kDa to 300 kDa in size. This pore size is preferred, when the PCV-2 antigen is produced as whole virus or as virus like particles.

According to a further aspect, any of the method described above to obtain a PCV-2 antigen with reduced virucidal activity can include further purification steps to obtain a purified PCV-2 antigen. It was surprisingly found that an antigenic or immunogenic composition comprising a purified PCV-2 antigen, preferably in combination with an adjuvant, not only shows a reduced virucidal activity as described herein, but also shows an increased immunogenicity as compared to an immunogenic composition, which does not comprise a purified PCV-2 antigen, means which comprises a non-purified or crude PCV-2 antigen.

The term "purified PCV-2 antigen" means, that the PCV-2 antigen is purified in a preparation to an extent of more than 50% (w/w), preferably of more than 60% (w/w), preferably of more than 70% (w/w), preferably of more than 80% (w/w), preferably of more than 85% (w/w), more preferably of more than 90% (w/w), even more preferable of more than 95% (w/w) with reference to the total amount of protein included in the immunogenic composition. In other words, if a preparation comprises a PCV-2 antigen with purity grade of 80% (w/w), such preparation comprise not more than 20% (w/w) of non PCV-2 proteins with reference to the total amount of protein included in the immunogenic composition. Preferably, the grade of purity is measured in the preparation, i.e. in the immunogenic composition before admixing with adjuvant or any other excipients or inactivating agent. However, if the

adjuvant used in the final immunogenic composition is a non-protein based adjuvant, the addition of the adjuvant does not have any effect of the purity value. The purity grade of the PCV-2 antigen can be estimated by standard methods known to a person skilled in the art, for instance by Imperial Protein Stain (Pierce) after SDS-PAGE separation, gas chromatography, HPLC analyses, etc. The preferred method according to this invention to estimate the purity or purity grade of a PCV-2 antigen in a preparation i.e. an immunogenic composition is the Imperial Protein Stain (Pierce) staining, which is done as follows: The preparation comprising the PCV-2 antigen are separated via NuPAGE 10% Bis-Tris gels (Invitrogen) using the NuPAGE MOPS buffer system (Invitrogen). Gels were run under denaturing (all buffers have SDS in them) and reducing conditions (the loading buffer has 2-mercaptoethanol). After loading the gels with samples, the gels were run for 55 min at 200 Volts constant. Once the run was completed, the gels were stained using Imperial Protein Stain (Pierce) and destained according the manufacturer's instructions.

In contrast, the term "non-purified" or "crude" PCV-2 antigen refers to a crude preparation comprising PCV-2 antigen. PCV-2 antigen is normally produced *in vitro* in cell culture. Thus, a crude PCV-2 antigen refers to a mixture of PCV-2 antigen and the cell culture or cell culture material used for the production of the PCV-2 antigen. Moreover, a non-purified PCV-2 antigen also means a partial purified PCV-2 antigen, preferably having a purity grade of less than 50% (w/w), more preferred of less than 40% (w/w), even more preferred of less than 30% (w/w), even more preferred of less than 20% (w/w) with reference to the total amount of protein included in the immunogenic composition.

In addition, the terms "increased immunogenicity or improved immunogenicity" as used herein, mean that the immune response caused by an immunogenic composition comprising an antigen of interest is increased as compared to a reference immunogenic composition comprising a different antigen or different purity grade of the antigen, whether this immune response is a cellular mediated and/or antibody mediated immune response. According to a preferred embodiment, the term increased immunogenicity or improved immunogenicity means, that the antibody mediated immune response elicited by an immunogenic composition comprising the antigen of interest is increased as compared to a reference immunogenic composition comprising a different antigen or a different purity grade of the antigen. In this regard antibody mediated immune response means, that the production of antibodies, which are specific to the antigen of interest is increased as compared to the antibody production elicited by a reference immunogenic composition comprising a different antigen or a different purity grade of the antigen.

The term "increased" means, that the cellular and/or antibody mediated immune response is increased by at least 10%, preferably by at least 20%, more preferably by at least 30%, even more preferably by at least 40%, even more preferably by at least 50%, even more preferably by at least 75%, most preferably by at least 100% as compared to the cellular and/or antibody mediated immune response elicited by a reference immunogenic composition comprising a different antigen or a different purity grade of the antigen.

It is in the general knowledge of a person skilled in the art how to measure the cellular and/or antibody mediated immune response. In particular, it is clear to such person skilled in the art either to compare the cellular mediated immune response of the immunogenic composition of interest with cellular mediated immune response of the reference, or the antibody mediated immune response of the immunogenic composition of interest with that of the reference composition, but neither the cellular mediated immune response of a immunogenic composition of interest with the antibody mediated immune response of the reference or *vice versa*. Moreover, the cellular mediated immune response can be measured, for instance, by measuring the activation of cytotoxic T-cells by an immunogenic composition/antigen of interest. The antibody mediated immune response can be measured, for instance, by measuring the amount of antigen specific antibodies, generated in cause of the administration of the immunogenic composition comprising such antigen to an animal. The cellular and/or antibody mediated immune response can be measured, for instance, by using a mouse model. According to the current invention, the mouse model is used as the reference method.

The term "immunogenic composition" means, but is not limited to, a composition of matter that comprises at least one antigen which elicits a cellular and/ or antibody-mediated immune response in a host against the antigen of interest. Usually, an "immune response" includes but is not limited to one or more of the following effects: the production or activation of antibodies, B cells, helper T cells, suppressor T cells, and/or cytotoxic T cells and/or gamma-delta T cells, directed specifically to an antigen or antigens included in the composition or vaccine of interest. Preferably, the host will display either a therapeutic or protective immune response such that resistance to new infection will be enhanced and/or the clinical severity of the disease reduced. In such a case the immunogenic composition is a "vaccine". Such protection will be demonstrated by either a reduction or lack of symptoms normally displayed by an infected host, a quicker recovery time and/or a lowered viral titer in the infected host.

Further purification of the PCV-2 antigen can be achieved with chromatography procedures, preferably a two-step chromatography procedure. If the PCV-2 antigen is assembled to virus like particles (VLP), one step, preferably the first step, is preferably a size exclusion (gel filtration) chromatography, which can be done, for instance, by using a Sephacryl S300 matrix. In lab scale use of HiPrep 26/60 Sephacryl S300HR columns are most preferred. However, any other size exclusion chromatography matrices known to a person skilled in the art can be used, which allow the separation of the PCV-2 ORF2 VLPs from the culture filtrate or supernatant. Suitable matrices are described, for instance, in E.L.V. Harris and S. Angel (eds.), Protein purification methods – a practical approach, IRL Press Oxford 1995). The gel filtration chromatography can be conducted, for instance, by loading the column with the crude preparation comprising the PCV-2 antigen with a flow-rate of 1.0 ml/min and eluting the column with 1.5 column volume of a buffer comprising 20 mM Tris, pH 6.5, 5 mM DTT. However, the PCV-2 ORF2 antigen can also be purified by using affinity chromatography, for instance, via selective binding to an immobilized PCV-2 ORF2 specific antibody, or any other method known to a person skilled in the art.

Thus according to a preferred embodiment the present invention provides a method of producing a PCV-2 antigenic composition comprising the steps of i) obtaining a first liquid containing a PCV-2 antigen, ii) removing at least a portion of the first liquid from the PCV-2 antigen and iii) purifying the harvest of step ii) comprising the PCV-2 antigen, preferably the PCV-2 ORF2 antigen by chromatographic procedure. Preferably size exclusion chromatography is performed as described herein, preferably as described in Example 3. Preferably, the size exclusion results in an immunogenic composition having purity grade of more than 80% (w/w), preferably more than 90% (w/w) with reference to the total amount of protein included in the immunogenic composition prior to the mixture with the adjuvant. The purity grade can be estimated by Imperial Protein Stain (Pierce) staining after SDS PAGE via NuPAGE 10% Bis-Tris gels (Invitrogen) using the NuPAGE MOPS buffer system (Invitrogen).

Thus according to a preferred embodiment the present invention provides a method of producing a PCV-2 antigenic composition comprising the steps of i) obtaining a first liquid containing a PCV-2 antigen, ii) removing at least a portion of the first liquid from the PCV-2 antigen and iii) purifying the harvest of step ii) comprising the PCV-2 antigen by size exclusion chromatography (gel filtration).

In order to obtain a higher purity grade a second chromatography step can be done,

which however is different from the first one. For instance if the first purification step / chromatography step is size exclusion (gel filtration) the second should different from that e.g. an affinity chromatography, ion exchange chromatography, etc. Preferably, if the first step to purify PCV-2 antigen, preferably to purify PCV-2 ORF2 antigen is a size exclusion (gel filtration) chromatography, the second step can be ion-exchange chromatography, preferably anion-exchange chromatography (AIEX). A preferred anion-exchange chromatography matrix for the purification of PCV-2 antigen, preferably the PCV-2 ORF2 antigen is Q Sepharose. In a small scale of about 50 ml, use of 5 ml HiTrap Q Sepharose HP columns are most preferred. The anion exchange chromatography can be conducted, for instance, as described in Example 3. Briefly, about 50 ml of the void volume fraction pool from the size exclusion chromatography step can be loaded onto the AIEX column at a flow rate of 3.0 ml/min. Following a washing step using, for instance, 20 mM Tris, pH 6.5, 5 mM DTT to remove unbound material, protein can be eluted with a single step of 8 column volumes of the following buffer (20 mM Tris, pH 6.5, 5 mM DTT, 1.0 M NaCl) The flow-through from the AIEX run can be loaded back onto the Q Sepharose column and eluted as described above to increase the yield. This two step technique (size exclusion followed by anion-exchange chromatography) effectively separates PCV-2 ORF2 antigen from most of the other protein components of the culture harvest.

Thus according to a preferred embodiment the present invention provides a method of producing a PCV-2 antigenic composition comprising the steps of i) obtaining a first liquid containing a PCV-2 antigen, ii) removing at least a portion of the first liquid from the PCV-2 antigen and iii) purifying the harvest of step ii) comprising the PCV-2 antigen, by a two-step chromatography. Preferably the first chromatography step is different from the second step. If the first step is a size exclusion (gel filtration) chromatography, the second step can be ion-exchange chromatography, preferably anion-exchange chromatography (AIEX). Preferably, in any of the methods described above, which include one or more further purification steps to obtain a purified PCV-2 antigen, preferably a PCV-2 ORF-2 protein, the portion of the first liquid is removed from the PCV-2 antigen by an exchange of a portion of the first liquid against a second liquid. The exchange is preferably done such that it comprises the steps of a) adding the second liquid to the first liquid which contains the PCV-2 antigen and b) concentrating the PCV-2 antigen, preferably from 3X to 50X, even more preferably from 4X to 20X, and even more preferably from 7X to 10X in comparison to the volume of the first liquid by removing a portion of the first and second liquids from the PCV-2 antigen. Preferably, the liquid addition step and concentration step are performed multiple times,

preferably two times, and even more preferably three times. In such cases, not only the first liquid is removed, but also a mixture of the first and second liquid. Preferably each liquid addition step is performed substantially simultaneously or sequentially as described above. When the concentrating step and liquid addition step are performed sequentially, the order of the steps does not matter. Moreover, the concentration step is preferably done by filtration - preferably by dia- or ultrafiltration, utilizing a filter, which preferably contains a semi-permeable membrane. The semi-permeable membrane preferably has an average pore size that is smaller than the PCV-2 antigen and prevents passage of at least 90% of the PCV-2 antigen through the semi-permeable membrane pores and withholds the PCV-2 antigen within the filter for harvesting or recovery. Preferably the average pore size of the semi-permeable membrane or of any other filter that is used herein, prevents passage of at least 90% of proteins of 50 kDa to 500 kDa in size, more preferably, at least 90% of proteins of 75 kDa to 400 kDa in size, and most preferably at least 90% of proteins of 100 kDa to 300 kDa in size. This pore size is preferred, when the PCV-2 antigen is produced as whole virus or as virus like particles. In preferred forms, the method of producing a PCV-2 antigenic composition described above further comprises the steps of i) obtaining a PCV-2 antigen in a first liquid wherein the PCV-2 antigen is obtained via a viral vector, preferably a recombinant baculovirus viral vector, containing and expressing the PCV-2 antigen, preferably, PCV-2 ORF-2, and wherein the PCV-2 antigen comprises the ORF-2 protein of PCV-2, more preferably recombinant ORF-2 protein of PCV-2, and still more preferably virus like particles of ORF-2 protein; ii) removing at least a portion of the first liquid from the PCV-2 antigen; iii) inactivating the recombinant baculovirus viral vector with a DNA inactivating agent, preferably in the presence of about 1 to about 20 mM of binary ethylenimine; iv) adding an amount of a neutralizing agent that neutralizes the inactivating agent, the amount of neutralizing agent being equivalent to the amount of the inactivating agent, wherein the neutralizing agent preferably comprises a sodium thiosulfate solution preferably concentrated to a final concentration of about 1 to about 20 mM and wherein the inactivating agent preferably comprises BEI; and v) admixing the PCV-2 antigen obtained in step iv) with a further component selected from the group consisting of pharmaceutically acceptable carriers, adjuvants, diluents, excipients, and combinations thereof. The further purification, preferably, the two step purification strategy including the pre-filtration step results in an immunogenic composition having purity grade of more than 80% (w/w), preferably of more than 85% (w/w), even more preferred of more than 90% (w/w), most preferred of more than 95% (w/w)

with reference to the total amount of protein included in the immunogenic composition prior to the mixture with any adjuvant.

The PCV-2 antigenic composition produced by the method described herein causes a loss of less than 1 log $TCID_{50}$ of a live virus or less than 1 log CFU per ml of a live bacterium, when the live virus or live bacterium is mixed with the PCV-2 antigenic composition and incubated for 2 or more hours, preferably for more than 4 hours, even more preferably for more than 12 hours, even more preferably for more than 24 hours, even more preferably for more than 2 days, even more preferably for more than 4 days, even more preferably for more than 7 days, even more preferably for more than 2 weeks, even more preferably for more than 4 weeks, even more preferably for more than 2 months, even more preferably for more than 3 months, even more preferably for more than 4 months, even more preferably for more than 6 months, even more preferably for more than 9 months, even more preferably for more than 12 months, even more preferably for more than 18 months, most preferably for more than 2 years. More preferably, the PCV-2 antigenic composition produced by the method described herein causes a loss of less than 0.9 log $TCID_{50}$ per ml of a live virus or less than 0.9 log CFU per ml of a live bacterium, when the live virus or live bacterium is mixed and incubated with the PCV-2 antigenic composition for 2 or more hours, preferably for more than 4 hours, even more preferably for more than 12 hours, even more preferably for more than 24 hours, even more preferably for more than 2 days, even more preferably for more than 4 days, even more preferably for more than 7 days, even more preferably for more than 2 weeks, even more preferably for more than 4 weeks, even more preferably for more than 2 months, even more preferably for more than 3 months, even more preferably for more than 4 months, even more preferably for more than 6 months, even more preferably for more than 9 months, even more preferably for more than 12 months, even more preferably for more than 18 months, most preferably for more than 2 years. Even more preferably, the PCV-2 antigenic composition produced by the method described herein causes a loss of less than 0.7 log $TCID_{50}$ per ml of a live virus or less than 0.7 log CFU per ml of a live bacterium, when the live virus or live bacterium is mixed and incubated with the PCV-2 antigenic composition for 2 or more hours, preferably for more than 4 hours, even more preferably for more than 12 hours, even more preferably for more than 24 hours, even more preferably for more than 2 days, even more preferably for more than 4 days, even more preferably for more than 7 days, even more preferably for more than 2 weeks, even more preferably for more than 4 weeks, even more preferably for more than 2 months, even more preferably for more than 3 months, even more preferably for more than 4 months, even more

preferably for more than 6 months, even more preferably for more than 9 months, even more preferably for more than 12 months, even more preferably for more than 18 months, most preferably for more than 2 years. Still more preferably, the PCV-2 antigenic composition produced by steps by the method described herein causes a loss of less than 0.5 log $TCID_{50}$ per ml of a live virus or less than 0.5 log CFU per ml of a live bacterium, when the live virus or live bacterium is mixed and incubated with the PCV-2 antigenic composition for 2 or more hours , preferably for more than 4 hours, even more preferably for more than 12 hours, even more preferably for more than 24 hours, even more preferably for more than 2 days, even more preferably for more than 4 days, even more preferably for more than 7 days, even more preferably for more than 2 weeks, even more preferably for more than 4 weeks, even more preferably for more than 2 months, even more preferably for more than 3 months, even more preferably for more than 4 months, even more preferably for more than 6 months, even more preferably for more than 9 months, even more preferably for more than 12 months, even more preferably for more than 18 months, most preferably for more than 2 years. Even more preferably, the PCV-2 antigenic composition produced by the method described herein causes a loss of less than 0.3 log $TCID_{50}$ per ml of a live virus or less than 0.3 log CFU per ml of a live bacterium, when the live virus or live bacterium is mixed and incubated with the PCV-2 antigenic composition for 2 or more hours, preferably for more than 4 hours, even more preferably for more than 12 hours, even more preferably for more than 24 hours, even more preferably for more than 2 days, even more preferably for more than 4 days, even more preferably for more than 7 days, even more preferably for more than 2 weeks, even more preferably for more than 4 weeks, even more preferably for more than 2 months, even more preferably for more than 3 months, even more preferably for more than 4 months, even more preferably for more than 6 months, even more preferably for more than 9 months, even more preferably for more than 12 months, even more preferably for more than 18 months, most preferably for more than 2 years. The live virus can be any live virus, but preferably the live virus is the PRRS virus, preferably the PRRS virus having the ATCC accession number VR 2332. The live bacterium can be any bacterium, but is preferably the *Mycoplasma hyopneumonia* bacterium, preferably the J-strain of *Mycoplasma hyopneumonia*. The $TCID_{50}$ per ml can be estimated by a standard in vitro titration assay which allow the estimation of the amount of a live virus. The CFU per ml can be determined also by a standard in vitro titration assay which allows the estimation of the amount of a live bacterium. The term "per ml" preferably refers to 1 ml of a fluid. Such purified PCV-2 antigen, does not only show reduced virucidal activity, as defined herein, it also shows an increased immunogenicity as compared

to a non-purified PCV-2 antigen as defined herein, preferably such purified PCV-2 antigen increases the cellular and/or antibody mediated immune response by at least 10%, preferably by at least 20%, more preferably by at least 30%, even more preferably by at least 40%, even more preferably by at least 50%, even more preferably by at least 75%, most preferably by at least 100% as compared to the cellular and/or antibody mediated immune response elicited by a reference immunogenic composition comprising a non-purified PCV-2 antigen.

Thus according to a further aspect, the present application provides a method of producing a PCV-2 antigenic composition comprising the steps of i) obtaining a first liquid containing a PCV-2 antigen, ii) removing at least a portion of the first liquid from the PCV-2 antigen, wherein the PCV-2 antigenic composition obtained after step ii) causes a loss of less than 1 log $TCID_{50}$ – preferably per ml - , preferably less than 0.9 log $TCID_{50,}$ - preferably per ml -, even more preferably less than 0.7 log $TCID_{50}$ - preferably per ml -, even more preferably less than 0.5 log $TCID_{50}$ - preferably per ml -, most preferably less than 0.3 log $TCID_{50}$ - preferably per ml - of a live virus, preferably of a live PRRSV or less than 1 log CFU - preferably per ml -, preferably less than 0.9 log CFU - preferably per ml -, even more preferably less than 0.7 log CFU - preferably per ml -, even more preferably less than 0.5 log CFU - preferably per ml -, most preferably less than 0.3 log CFU - preferably per ml - of a live bacterium, preferably of *Mycoplasma hyopneumoniae*, when the live virus, preferably PRRSV or live bacterium, preferably *Mycoplasma hyopneumoniae* is mixed and incubated with the PCV-2 antigenic composition for 2 or more hours, preferably for more than 4 hours, even more preferably for more than 12 hours, even more preferably for more than 24 hours, even more preferably for more than 2 days, even more preferably for more than 4 days, even more preferably for more than 7 days, even more preferably for more than 2 weeks, even more preferably for more than 4 weeks, even more preferably for more than 2 months, even more preferably for more than 3 months, even more preferably for more than 4 months, even more preferably for more than 6 months, even more preferably for more than 9 months, even more preferably for more than 12 months, even more preferably for more than 18 months, most preferably for more than 2 years. Preferably, the portion of the first liquid is removed from the PCV-2 antigen by an exchange of a portion of the first liquid against a second liquid. The exchange is preferably done in such that it comprises the steps of a) adding the second liquid to the first liquid which contains the PCV-2 antigen and b) concentrating the PCV-2 antigen, preferably from 3X to 50X, even more preferably from 4X to 20X, even more preferably from 7X to 10X in comparison to the volume of the first liquid by removing a portion of the first and second liquids from the PCV-2 antigen. Preferably, the liquid addition

step and concentration step are performed multiple times, preferably two times, even more preferably three times. In such case, not only the first liquid is removed, but also a mixture of the first and second liquid. Preferably each liquid addition step is performed substantially simultaneously or sequentially as described above. When the concentrating step and liquid addition step are performed sequentially, the order of the steps does not matter. Moreover, the concentration step is preferably done by filtration - preferably by dia- and/or ultra-filtration, utilizing a filter, which preferably contains a semi-permeable membrane. The semi-permeable membrane preferably has an average pore size that is smaller than the PCV-2 antigen and prevents passage of at least 90% of the PCV-2 antigen through the semi-permeable membrane pores and withholds the PCV-2 antigen within the filter for harvesting or recovery. Preferably the average pore size of the semi-permeable membrane or of any other filter that is used herein, prevents passage of at least 90% of proteins of 50 kDa to 500 kDa in size, more preferably, at least 90% of proteins of 75 kDa to 400 kDa in size, and most preferably at least 90% of proteins of 100 kDa to 300 kDa in size. This pore size is preferred, when the PCV-2 antigen is produced as whole virus or as virus like particles. When the PCV-2 antigen is obtained via a viral vector, preferably a recombinant baculovirus viral vector, containing and expressing the PCV-2 antigen, preferably, PCV-2 ORF-2, the process further comprises iii) inactivating the recombinant baculovirus viral vector with a DNA inactivating agent, preferably in the presence of about 1 to about 20 mM of binary ethylenimine; iv) adding an amount of a neutralizing agent that neutralizes the inactivating agent, the amount of neutralizing agent being equivalent to the amount of the inactivating agent, wherein the neutralizing agent preferably comprises a sodium thiosulfate solution preferably concentrated to a final concentration of about 1 to about 20 mM and wherein the inactivating agent preferably comprises BEI. Preferably, the inactivating and neutralization steps are performed after at least a portion of the first liquid is removed from the PCV-2 antigen, more preferably after the PCV-2 antigen is harvested. Even more preferably, the inactivating and neutralization steps are performed after the portion of the first liquid is removed from the PCV-2 antigen by an exchange of a portion of the first liquid against a second liquid. When the exchange of a portion of the first liquid against a second liquid is done such that it comprises the steps of a) adding the second liquid to the first liquid which contains the PCV-2 antigen and b) concentrating the PCV-2 antigen, preferably from 3X to 50X, even more preferably from 4X to 20X, even more preferably from 7X to 10X in comparison to the volume of the first liquid by removing a portion of the first and second liquids from the PCV-2 antigen, the inactivating and neutralization steps are done after the concentration step.

When the liquid addition step and concentration step are performed multiple times, preferably two times, and even more preferably three times, such inactivation and neutralization steps are performed after the last liquid addition step and concentration step. When the concentration step is done by filtration - preferably by dia- and/or ultrafiltration, utilizing a filter, preferably containing a semi-permeable membrane, the inactivation and neutralization steps are performed after the filtration step described above, preferably utilizing a semi-permeable membrane. The semi-permeable membrane preferably has an average pore size that is smaller than the PCV-2 antigen and prevents passage of at least 90% of the PCV-2 antigen through the semi-permeable membrane pores and withholds the PCV-2 antigen within the filter for harvesting or recovery. Preferably the average pore size of the semi-permeable membrane or of any other filter that is used herein, prevents passage of at least 90% of proteins of 50 kDa to 500 kDa in size, more preferably, at least 90% of proteins of 75 kDa to 400 kDa in size, and most preferably at least 90% of proteins of 100 kDa to 300 kDa in size. This pore size is preferred, when the PCV-2 antigen is produced as whole virus or as virus like particles. Preferably, further purification to obtain a purified PCV-2 antigen as defined herein, can be achieved by performing further purification step comprising iii) purifying the harvest of step ii) comprising the PCV-2 antigen, which is obtained after the removal of a portion of the first liquid, by a chromatography a step. In order to obtain a higher purity grade a second chromatography step can be done, which however is different from the first one. For instance if the first purification step / chromatography step is size exclusion (gel filtration) the second should different from that e.g. an affinity chromatography, ion exchange chromatography, etc. Preferably, if the first step to purify PCV-2 antigen, preferably to purify PCV-2 ORF2 antigen is a size exclusion (gel filtration) chromatography, the second step can be ion-exchange chromatography, preferably anion-exchange chromatography (AIEX). A preferred anion-exchange chromatography matrix for the purification of PCV-2 antigen, preferably the PCV-2 ORF2 antigen is Q Sepharose. In a small scale of about 50 ml, use of 5 ml HiTrap Q Sepharose HP columns are most preferred. The anion exchange chromatography can be conducted, for instance, as described in Example 3. Briefly, about 50 ml of the void volume fraction pool from the size exclusion chromatography step can be loaded onto the AIEX column at a flow rate of 3.0 ml/min. Following a washing step using, for instance, 20 mM Tris, pH 6.5, 5 mM DTT to remove unbound material, protein can be eluted with a single step of 8 column volumes of the following buffer (20 mM Tris, pH 6.5, 5 mM DTT, 1.0 M NaCl) The flow-through from the AIEX run can be loaded back onto the Q Sepharose column and eluted as described above to increase the yield. This two step technique (size exclusion

followed by anion-exchange chromatography) effectively separates PCV-2 ORF2 antigen from most of the other protein components of the culture harvest.

The PCV-2 antigenic composition obtained according to the method described above, or the PCV-2 antigen used in step i) of the method described above, can be combined with at least one additional antigen, preferably a viral or bacterial antigen, and even more preferably, a viral or bacterial antigen from at least one other disease-causing organism in swine. The additional antigen can be any one of those disclosed in the international patent application WO2007/094893 (the contents and teachings of which are hereby incorporated by reference). Briefly, the additional antigens can be antigens of any other disease-causing organisms of swine. Preferably the "another disease-causing organisms" of swine are selected from the group consisting of: Actinobacillus pleuropneumonia (1); Adenovirus (2); Alphavirus such as Eastern equine encephalomyelitis viruses (3); Bordetella bronchiseptica (4); Brachyspira spp. (5), preferably B. hyodyentheriae (6); B. piosicoli (7), Brucella suis, preferably biovars 1, 2, and 3 (8); Clasical swine fever virus (9); Clostridium spp. (10), preferably Cl. difficile (11), Cl. perfringens types A, B, and C (12), Cl. novyi (13), Cl.septicum (14), Cl. tetani (15); Coronavirus (16), preferably Porcine Respiratory Corona virus (17); Eperythrozoonosis suis (18); Erysipelothrix rhsiopathiae (19) Escherichia coli (20); Haemophilus parasuis, preferably subtypes 1, 7 and 14 (21) Hemagglutinating encephalomyelitis virus (22); Japanese Encephalitis Virus (23); Lawsonia intracellularis(24) Leptospira spp. (25), preferably Leptospira australis (26); Leptospira canicola (27); Leptospira grippotyphosa (28); Leptospira icterohaemorrhagicae (29); and Leptospira interrogans (30); Leptospira pomona (31); Leptospira tarassovi (32); Mycobacterium spp. (33) preferably M. avium (34), M. intracellulare (35) and M.bovis (36); Mycoplasma hyopneumoniae (37); Pasteurella multocida (38); Porcine cytomegalovirus (39); Porcine Parvovirus (40); Porcine Reproductive and Respiratory Syndrome Virus (41); Pseudorabies virus (42); Rotavirus (43); Salmonella spp. (44), preferably S. thyhimurium (45) and S. choleraesuis (46); Staph. hyicus (47); Staphylococcus spp. (48) preferably Streptococcus spp. (49), preferably Strep. suis (50); Swine herpes virus (51); Swine Influenza Virus (52); Swine pox virus (53); Swine pox virus (54); Vesicular stomatitis virus (55); Virus of vesicular exanthema of swine (56); Leptospira Hardjo (57); and/or Mycoplasma hyosynoviae (58).

Thus, according to a further aspect of the present invention, the present invention provides a method of producing a PCV-2 antigenic composition comprising the steps of i) obtaining a PCV-2 antigen in a first liquid; ii) removing at least a portion of the first liquid from the PCV-2 antigen; and combining the PCV-2 antigen with at least one additional

antigen, preferably a viral or bacterial antigen, and more preferably a viral or bacterial antigen from at least one other disease-causing organism in swine. Preferably, the PCV-2 antigen comprises the ORF-2 protein of PCV-2, more preferably recombinant ORF-2 protein of PCV-2, and still more preferably virus like particles of ORF-2 protein. Preferably, the portion of the first liquid is removed from the PCV-2 antigen by an exchange of a portion of the first liquid against a second liquid. The exchange is preferably done such that it comprises the steps of a) adding the second liquid to the first liquid which contains the PCV-2 antigen and b) concentrating the PCV-2 antigen, preferably from 3X to 50X, even more preferably from 4X to 20X, and even more preferably from 7X to 10X in comparison to the volume of the first liquid by removing a portion of the first and second liquids from the PCV-2 antigen. Preferably, the liquid addition step and concentration step are performed multiple times, preferably two times, and even more preferably three times. In such cases, not only the first liquid is removed, but also a mixture of the first and second liquid. Preferably each liquid addition step is performed substantially simultaneously or sequentially as described above. When the concentrating step and liquid addition step are performed sequentially, the order of the steps does not matter. Moreover, the concentration step is preferably done by filtration - preferably by dia- or ultrafiltration, utilizing a filter, which preferably contains a semi-permeable membrane. The semi-permeable membrane preferably has an average pore size that is smaller than the PCV-2 antigen and prevents passage of at least 90% of the PCV-2 antigen through the semi-permeable membrane pores and withholds the PCV-2 antigen within the filter for harvesting or recovery. Preferably the average pore size of the semi-permeable membrane or of any other filter that is used herein, prevents passage of at least 90% of proteins of 50 kDa to 500 kDa in size, more preferably, at least 90% of proteins of 75 kDa to 400 kDa in size, and most preferably at least 90% of proteins of 100 kDa to 300 kDa in size. This pore size is preferred, when the PCV-2 antigen is produced as whole virus or as virus like particles. Further purification to obtain a purified PCV-2 antigen can be done as described above.

In preferred forms, the method of producing a PCV-2 antigenic composition described above further comprises the steps of i) obtaining a PCV-2 antigen in a first liquid wherein the PCV-2 antigen is obtained via a viral vector, preferably a recombinant baculovirus viral vector, containing and expressing the PCV-2 antigen, preferably, PCV-2 ORF-2, and wherein the PCV-2 antigen comprises the ORF-2 protein of PCV-2, more preferably recombinant ORF-2 protein of PCV-2, and still more preferably virus like particles of ORF-2 protein; ii) removing at least a portion of the first liquid from the PCV-2 antigen; iii) inactivating the

recombinant baculovirus viral vector with a DNA inactivating agent, preferably in the presence of about 1 to about 20 mM of binary ethylenimine; iv) adding an amount of a neutralizing agent that neutralizes the inactivating agent, the amount of neutralizing agent being equivalent to the amount of the inactivating agent, wherein the neutralizing agent preferably comprises a sodium thiosulfate solution preferably concentrated to a final concentration of about 1 to about 20 mM and wherein the inactivating agent preferably comprises BEI; and v) admixing the PCV-2 antigen obtained in step iv) with a further component selected from the group consisting of pharmaceutically acceptable carriers, adjuvants, diluents, excipients, and combinations thereof.

In a further aspect of the method, the at least one additional antigen is a viral antigen, preferably an antigen from Porcine Reproductive and Respiratory Syndrome Virus. Even more preferably, the Porcine Reproductive and Respiratory Syndrome Virus antigen comprises a live virus, and still more preferably a modified live virus, even more preferably a modified live attenuated virus. Still more preferably, the modified live Porcine Reproductive and Respiratory Syndrome Virus antigen comprises a modified live virus strain of ATCC Accession Number VR 2332, and still more preferably comprises INGELVAC® PRRS MLV. Thus, according to a further aspect, the present application provides a method of producing a PCV-2 antigenic composition comprising the steps of i) obtaining a first liquid containing a PCV-2 antigen, ii) removing at least a portion of the first liquid from the PCV-2 antigen, and combining the PCV-2 antigen with an antigen from Porcine Reproductive and Respiratory Syndrome Virus. Preferably, the Porcine Reproductive and Respiratory Syndrome Virus antigen comprises a live virus, still more preferably a modified live virus, and even more preferably a modified live attenuated virus. Still more preferably, the modified live Porcine Reproductive and Respiratory Syndrome Virus antigen comprises a modified live virus strain of ATCC Accession Number VR 2332, and still more preferably comprises INGELVAC® PRRS MLV. Preferably the PCV-2 antigen comprises the ORF-2 protein of PCV-2, more preferably recombinant ORF-2 protein of PCV-2, and still more preferably virus like particles of ORF-2 protein. Preferably, the portion of the first liquid is removed from the PCV-2 antigen by an exchange of a portion of the first liquid against a second liquid. The exchange is preferably done such that it comprises the steps of a) adding the second liquid to the first liquid which contains the PCV-2 antigen and b) concentrating the PCV-2 antigen, preferably from 3X to 50X, even more preferably from 4X to 20X, and even more preferably from 7X to 10X in comparison to the volume of the first liquid by removing a portion of the first and second liquids from the PCV-2 antigen. Preferably, the liquid addition step and concentration

step are performed multiple times, preferably two times, even more preferably three times. In such case, not only the first liquid is removed, but also a mixture of the first and second liquid. Preferably each liquid addition step is performed substantially simultaneously or sequentially as described above. When the concentrating step and liquid addition step are performed sequentially, the order of the steps does not matter. Moreover, the concentration step is preferably done by filtration - preferably by dia- and/or ultrafiltration, utilizing a filter, which preferably contains a semi-permeable membrane. The semi-permeable membrane preferably has an average pore size that is smaller than the PCV-2 antigen and prevents passage of at least 90% of the PCV-2 antigen through the semi-permeable membrane pores and withholds the PCV-2 antigen within the filter for harvesting or recovery. Preferably the average pore size of the semi-permeable membrane or of any other filter that is used herein, prevents passage of at least 90% of proteins of 50 kDa to 500 kDa in size, more preferably, at least 90% of proteins of 75 kDa to 400 kDa in size, and most preferably at least 90% of proteins of 100 kDa to 300 kDa in size. This pore size is preferred, when the PCV-2 antigen is produced as whole virus or as virus like particles. Further purification to obtain a purified PCV-2 antigen can be done as described above.

In a further aspect of the present application, the at least one additional antigen is a bacterial antigen, preferably *Mycoplasma hyopneumoniae*. Preferably the *Mycoplasma hyopneumoniae* antigen is a bacterin, and more preferably, the *Mycoplasma hyopneumoniae* bacterin is INGELVAC® MYCOFLEX. Thus, according to a further aspect, the present application provides a method of producing a PCV-2 antigenic composition comprising the steps of i) obtaining a first liquid containing a PCV-2 antigen, ii) removing at least a portion of the first liquid from the PCV-2 antigen, and combining the PCV-2 antigen with a bacterial antigen, preferably *Mycoplasma hyopneumoniae*. Preferably the *Mycoplasma hyopneumoniae* antigen is a bacterin, and more preferably, the *Mycoplasma hyopneumoniae* bacterin is INGELVAC® MYCOFLEX. Preferably the PCV-2 antigen comprises the ORF-2 protein of PCV-2, more preferably recombinant ORF-2 protein of PCV-2, and still more preferably virus like particles of ORF-2 protein. Preferably, the portion of the first liquid is removed from the PCV-2 antigen by an exchange of a portion of the first liquid against a second liquid. The exchange is preferably done such that it comprises the steps of a) adding the second liquid to the first liquid which contains the PCV-2 antigen and b) concentrating the PCV-2 antigen, preferably from 3X to 50X, even more preferably from 4X to 20X, and even more preferably from 7X to 10X in comparison to the volume of the first liquid by removing a portion of the first and second liquids from the PCV-2 antigen. Preferably, the liquid

addition step and concentration step are performed multiple times, preferably two times, and even more preferably three times. In such cases, not only the first liquid is removed, but also a mixture of the first and second liquid. Preferably each liquid addition step is performed substantially simultaneously or sequentially as described above. When the concentrating step and liquid addition step are performed sequentially, the order of the steps does not matter. Moreover, the concentration step is preferably done by filtration - preferably by dia- or ultrafiltration, utilizing a filter, which preferably contains a semi-permeable membrane. The semi-permeable membrane preferably has an average pore size that is smaller than the PCV-2 antigen and prevents passage of at least 90% of the PCV-2 antigen through the semi-permeable membrane pores and withholds the PCV-2 antigen within the filter for harvesting or recovery. Preferably the average pore size of the semi-permeable membrane or of any other filter that is used herein, prevents passage of at least 90% of proteins of 50 kDa to 500 kDa in size, more preferably, at least 90% of proteins of 75 kDa to 400 kDa in size, and most preferably at least 90% of proteins of 100 kDa to 300 kDa in size. This pore size is preferred, when the PCV-2 antigen is produced as whole virus or as virus like particles. Further purification to obtain a purified PCV-2 antigen can be done as described above.

In a further aspect of the present application, the at least one additional antigen includes a viral antigen, preferably a Porcine Reproductive and Respiratory Syndrome Virus antigen, as described above, and a bacterial antigen, preferably a *Mycoplasma hyopneumoniae* antigen, as described above. Preferably, the Porcine Reproductive and Respiratory Syndrome Virus antigen comprises a live virus, more preferably a modified live virus, and still more preferably, comprises a modified live virus strain of ATCC Accession Number VR 2332, and still more preferably comprises INGELVAC® PRRS MLV. Preferably, the *Mycoplasma hyopneumoniae* antigen is a bacterin, and more preferably, the *Mycoplasma hyopneumoniae* bacterin is INGELVAC® MYCOFLEX Thus, according to a further aspect, the present application provides a method of producing a PCV-2 antigenic composition comprising the steps of i) obtaining a first liquid containing a PCV-2 antigen, ii) removing at least a portion of the first liquid from the PCV-2 antigen, and combining the PCV-2 antigen with a viral antigen, preferably a Porcine Reproductive and Respiratory Syndrome Virus antigen, as described above, and a bacterial antigen, preferably a *Mycoplasma hyopneumoniae* antigen, as described above. Preferably, the Porcine Reproductive and Respiratory Syndrome Virus antigen comprises a live virus, more preferably a modified live virus, and still more preferably, comprises a modified live virus strain of ATCC Accession Number VR 2332, and still more preferably comprises INGELVAC® PRRS MLV. Preferably, the *Mycoplasma*

hyopneumoniae antigen is a bacterin, and more preferably, the *Mycoplasma hyopneumoniae* bacterin is INGELVAC® MYCOFLEX. Preferably the PCV-2 antigen comprises the ORF-2 protein of PCV-2, more preferably recombinant ORF-2 protein of PCV-2, and still more preferably virus like particles of ORF-2 protein. Preferably, the portion of the first liquid is removed from the PCV-2 antigen by an exchange of a portion of the first liquid against a second liquid. The exchange is preferably done such that it comprises the steps of a) adding the second liquid to the first liquid which contains the PCV-2 antigen and b) concentrating the PCV-2 antigen, preferably from 3X to 50X, even more preferably from 4X to 20X, and even more preferably from 7X to 10X in comparison to the volume of the first liquid by removing a portion of the first and second liquids from the PCV-2 antigen. Preferably, the liquid addition step and concentration step are performed multiple times, preferably two times, and even more preferably three times. In such cases, not only the first liquid is removed, but also a mixture of the first and second liquid. Preferably the liquid addition step is performed substantially simultaneously or sequentially as described above. When the concentrating step and liquid addition step are performed sequentially, the order of the steps does not matter. Moreover, the concentration step is preferably done by filtration - preferably by dia- and/or ultrafiltration, utilizing a filter, which preferably contains a semi-permeable membrane. The semi-permeable membrane preferably has an average pore size that is smaller than the PCV-2 antigen and prevents passage of at least 90% of the PCV-2 antigen through the semi-permeable membrane pores and withholds the PCV-2 antigen within the filter for harvesting or recovery. Preferably the average pore size of the semi-permeable membrane or of any other filter that is used herein, prevents passage of at least 90% of proteins of 50 kDa to 500 kDa in size, more preferably, at least 90% of proteins of 75 kDa to 400 kDa in size, and most preferably at least 90% of proteins of 100 kDa to 300 kDa in size. This pore size is preferred, when the PCV-2 antigen is produced as whole virus or as virus like particles. Further purification to obtain a purified PCV-2 antigen can be done as described above.

The present application does not only provide methods of producing PCV-2 antigenic compositions, it also relates to a PCV-2 antigenic composition. Thus, according to a further aspect the present patent application further provides a PCV-2 antigenic composition characterized in such that the PCV-2 antigenic composition causes a loss of less than 1 log $TCID_{50}$ of a live virus or less than 1 log CFU per ml of a live bacterium, when the live virus or live bacterium is mixed with the PCV-2 antigenic composition and incubated for 2 or more hours, preferably for more than 4 hours, even more preferably for more than 12 hours, even more preferably for more than 24 hours, even more preferably for more than 2 days, even

more preferably for more than 4 days, even more preferably for more than 7 days, even more preferably for more than 2 weeks, even more preferably for more than 4 weeks, even more preferably for more than 2 months, even more preferably for more than 3 months, even more preferably for more than 4 months, even more preferably for more than 6 months, even more preferably for more than 9 months, even more preferably for more than 12 months, even more preferably for more than 18 months, and most preferably for more than 2 years. More preferably, the PCV-2 antigenic composition produced by the method described herein causes a loss of a live virus or less than 0.9 log CFU per ml of a live bacterium, when the live virus or live bacterium is mixed and incubated with the PCV-2 antigenic composition for 2 or more hours, preferably for more than 4 hours, even more preferably for more than 12 hours, even more preferably for more than 24 hours, even more preferably for more than 2 days, even more preferably for more than 4 days, even more preferably for more than 7 days, even more preferably for more than 2 weeks, even more preferably for more than 4 weeks, even more preferably for more than 2 months, even more preferably for more than 3 months, even more preferably for more than 4 months, even more preferably for more than 6 months, even more preferably for more than 9 months, even more preferably for more than 12 months, even more preferably for more than 18 months, and most preferably for more than 2 years. Even more preferably, the PCV-2 antigenic composition causes a loss of less than 0.7 log $TCID_{50}$ per ml of a live virus or less than 0.7 log CFU per ml of a live bacterium, when the live virus or live bacterium is mixed and incubated with the PCV-2 antigenic composition for 2 or more hours, preferably for more than 4 hours, even more preferably for more than 12 hours, even more preferably for more than 24 hours, even more preferably for more than 2 days, even more preferably for more than 4 days, even more preferably for more than 7 days, even more preferably for more than 2 weeks, even more preferably for more than 4 weeks, even more preferably for more than 2 months, even more preferably for more than 3 months, even more preferably for more than 4 months, even more preferably for more than 6 months, even more preferably for more than 9 months, even more preferably for more than 12 months, even more preferably for more than 18 months, and most preferably for more than 2 years.. Still more preferably, the PCV-2 antigenic composition causes a loss of less than 0.5 log $TCID_{50}$ per ml of a live virus or less than 0.5 log CFU per ml of a live bacterium, when the live virus or live bacterium is mixed and incubated with the PCV-2 antigenic composition for 2 or more hours , preferably for more than 4 hours, even more preferably for more than 12 hours, even more preferably for more than 24 hours, even more preferably for more than 2 days, even more preferably for more than 4 days, even more preferably for more than 7 days, even more

preferably for more than 2 weeks, even more preferably for more than 4 weeks, even more preferably for more than 2 months, even more preferably for more than 3 months, even more preferably for more than 4 months, even more preferably for more than 6 months, even more preferably for more than 9 months, even more preferably for more than 12 months, even more preferably for more than 18 months, and most preferably for more than 2 years. Even more preferably, the PCV-2 antigenic composition causes a loss of less than 0.3 log $TCID_{50}$ per ml of a live virus or less than 0.3 log CFU per ml of a live bacterium, when the live virus or live bacterium is mixed and incubated with the PCV-2 antigenic composition for 2 or more hours, preferably for more than 4 hours, even more preferably for more than 12 hours, even more preferably for more than 24 hours, even more preferably for more than 2 days, even more preferably for more than 4 days, even more preferably for more than 7 days, even more preferably for more than 2 weeks, even more preferably for more than 4 weeks, even more preferably for more than 2 months, even more preferably for more than 3 months, even more preferably for more than 4 months, even more preferably for more than 6 months, even more preferably for more than 9 months, even more preferably for more than 12 months, even more preferably for more than 18 months, and most preferably for more than 2 years. The live virus can be any live virus, but preferably the live virus is the PRRS virus, preferably the PRRS virus having the ATCC accession number VR 2332. The live bacterium can be any bacterium, but is preferably the *Mycoplasma hyopneumonia* bacterium, preferably the J-strain of *Mycoplasma hyopneumonia*. The $TCID_{50}$ per ml can be estimated by a standard in vitro titration assay which allow the estimation of the amount of a live virus. The CFU per ml can be determined also by a standard in vitro titration assay which allows the estimation of the amount of a live bacterium. The term "per ml" preferably refers to 1 ml of a fluid.

In a further aspect, the PCV-2 antigenic composition described above comprises a further component selected from the group consisting of pharmaceutically acceptable carriers, adjuvants, diluents, excipients, and combinations thereof. Preferably, the further component is an adjuvant, even more preferably wherein the adjuvant is a polymer of acrylic or methacrylic acid, and still more preferably wherein the adjuvant is Carbomer. Preferably, the adjuvant is added in an amount of about 100 µg to about 10 mg per dose. Even more preferably the adjuvant is added in an amount of about 100 µg to about 10 mg per dose. Still more preferably the adjuvant is added in an amount of about 500 µg to about 5 mg per dose. Still more preferably the adjuvant is added in an amount of about 750 µg to about 2.5 mg per dose. Most preferably the adjuvant is added in an amount of about 1 mg per dose.

The present application does not only provide methods of producing PCV-2 antigenic compositions and/or the PCV-2 antigenic compositions as defined above, it also relates to a PCV-2 antigenic composition that is obtainable by any of the methods described herein. Thus, in a further aspect the present application relates to a PCV-2 antigenic composition that is obtained by a method comprising the steps of i) obtaining a first liquid containing a PCV-2 antigen, ii) removing at least a portion of the first liquid from the PCV-2 antigen. Preferably the PCV-2 antigen is used as or in the PCV-2 antigenic composition. The term "a PCV-2 antigenic composition obtained by a method provided herein" also means that the PCV-2 antigenic composition is obtainable by a method provided herein. According to a further aspect, the present application also relates to the PCV-2 antigenic composition that is obtained by removing the portion of the first liquid from the PCV-2 antigen by an exchange of the portion of the first liquid against a second liquid, wherein the second liquid is different from the first liquid. Thus according to a further aspect, the present application relates to a PCV-2 antigenic composition obtained by a method comprising the steps of i) obtaining a first liquid containing a PCV-2 antigen, ii) removing at least a portion of the first liquid from the PCV-2 antigen, wherein the portion of the first liquid is removed from the PCV-2 antigen by an exchange of the portion of the first liquid against a second liquid, wherein the second liquid is different from the first liquid. Preferably the exchange of the portion of the first liquid with the second liquid comprises the steps of a) adding the second liquid to the first liquid which contains the PCV-2 antigen and b) concentrating the PCV-2 antigen by removing a portion of the first and second liquids from the PCV-2 antigen.

According to a further aspect, the PCV-2 antigenic composition is preferably obtained by a method wherein the portion of the first liquid is removed from the PCV-2 antigen by a filtration step utilizing a filter. However, any other methods known to a person skilled in the art can be used to remove the portion of the first and second fluid from the PCV-2 antigen, for instance centrifugation and/or chromatography. However, filtration is most preferred. Preferred filtration methods to remove the portion of the first fluid comprise ultra- and/or dia-filtration. The concentrating step and the liquid addition step of the method as described herein can be performed substantially simultaneously or alternatively, the concentrating step and the liquid addition step are performed sequentially. Thus according to a further aspect, the present application relates to a PCV-2 antigenic composition obtained by a method comprising the steps of i) obtaining a first liquid containing a PCV-2 antigen, ii) removing at least a portion of the first liquid from the PCV-2 antigen, wherein the portion of the first liquid is removed from the PCV-2 antigen by an exchange of the portion of the first liquid

against a second liquid, wherein the second liquid is different from the first liquid. Preferably the exchange of the portion of the first liquid with the second liquid comprises the steps of a) adding the second liquid to the first liquid which contains the PCV-2 antigen and b) concentrating the PCV-2 antigen by removing a portion of the first and second liquids from the PCV-2 antigen, wherein the liquid addition step is performed substantially simultaneously or sequentially. When the concentrating step and liquid addition step are performed sequentially, the order of the steps does not matter. For example, in a further aspect, the liquid addition step occurs prior to the concentrating step and in an alternative aspect, the concentrating step occurs prior to the liquid addition step.

In a further aspect, the present application relates to a PCV-2 antigenic composition that can be obtained using a method described herein, wherein the liquid addition step and the concentrating step, regardless of the order in which they are performed, can be performed multiple times. For example, each of these respective steps can be performed at least two, at least three, at least four, at least five, at least 10, up to as many times as desired. In one aspect, the concentrating step and the liquid addition step are each performed at least two times. In another aspect, the concentrating step and the liquid addition step are each performed at least three times.

In a further aspect of the present application, the PCV-2 antigenic composition of the present invention is obtained as described above, wherein filtration is the preferred method to remove a portion of the first liquid, or in case of multiple removing steps as described above, a portion of the mixture of the first and the second fluid from the PCV-2 antigen. The filter can be any conventional filter in the art. Preferably, the filter includes a semi-permeable membrane. In a further preferred form, the semi-permeable membrane has an average pore size that is smaller than the PCV-2 antigen to thereby prevent passage of at least 90% of the PCV-2 antigen through the semi-permeable membrane pores and withhold the PCV-2 antigen by the filter. In a further aspect, the filter has an average pore size which prevents passage of at least 90% of proteins of 50 kDa to 500 kDa in size, more preferably, the filter has an average pore size which prevents passage of at least 90% of proteins of 75 kDa to 400 kDa in size, and most preferably, the filter has an average pore size which prevents passage of at least 90% of proteins of 100 kDa to 300 kDa in size. This pore size is preferred, when the PCV-2 antigen is produced as whole virus or as virus like particles. In a still further aspect, the semi-permeable membrane includes a material selected from the group consisting of polysulfone, polyethersulfone, and regenerated cellulose. However, any other material can be used, which allows removing of a portion of the first fluid, and in case of a multiple process step,

removing of a mixture of the first and the second fluid from the PCV-2 antigen. In a further aspect, the filter is selected from the group consisting of a hollow fiber membrane ultra filtration cartridge, flat sheets, or a cassette, with a hollow fiber membrane ultra filtration cartridge being particularly preferred.

Thus, according to a further aspect, the present application relates to a PCV-2 antigenic composition that is obtained using the methods as described above, wherein the filter preferably is or comprises a semi-permeable membrane. Preferably, the semi-permeable membrane has an average pore size that is smaller than the PCV-2 antigen and prevent passage of at least 90% of the PCV-2 antigen through the semi-permeable membrane pores. Preferably the average pore size of the semi-permeable membrane prevents passage of at least 90% of proteins of 50 kDa to 500 kDa in size, more preferably, at least 90% of proteins of 75 kDa to 400 kDa in size, and most preferably at least 90% of proteins of 100 kDa to 300 kDa in size. This pore size is preferred, when the PCV-2 antigen is produced as whole virus or as virus like particles. As described above, the removing step in general includes the exchange of the portion of the first fluid against a portion of the second fluid comprises the steps of a) adding the second liquid to the first liquid which contains the PCV-2 antigen and b) concentrating the PCV-2 antigen by removing a portion of the first and second liquids from the PCV-2 antigen, wherein the liquid addition step and concentration step are performed multiple times, for instance, two times, three times, 5 times, 10 times, etc. Preferably, the liquid addition step and concentration step are performed two times, most preferably three times.

The concentration step of the method provided herein to obtain the PCV-2 antigenic composition is performed such that the PCV-2 antigen is concentrated from 3X to 50X in comparison to the volume of the first liquid. More preferably, the concentrating step is done in such that the PCV-2 antigen is concentrated 4X to 20X in comparison to the volume of the first liquid. Most preferably, concentration step is done in such that the PCV-2 antigen is concentrated from 7X to 10X in comparison to the volume of the first liquid. Thus according to a further aspect, the present application relates to a PCV-2 antigenic composition obtained by a method described above, wherein the PCV-2 antigen is concentrated from 3X to 50X, preferably from 4X to 20X, and even more preferably from 7X to 10X in comparison to the volume of the first liquid. Preferably, the portion of the first fluid is removed from the PCV-2 antigen by an exchange of the portion of the first liquid against a second liquid comprising the steps of a) adding the second liquid to the first liquid which contains the PCV-2 antigen and b) concentrating the PCV-2 antigen from 3X to 50X, preferably from 4X to 20X, and even

more preferably from 7X to 10X in comparison to the volume of the first liquid by removing a portion of the first and second liquids from the PCV-2 antigen. Preferably the liquid addition step is performed substantially simultaneously or sequentially with the concentrating step. When the concentrating step and liquid addition step are performed sequentially, the order of the steps does not matter. Moreover, the concentration step is preferably done by filtration - preferably dia- and/or ultrafiltration, utilizing a filter, which preferably contains a semi-permeable membrane. The semi-permeable membrane preferably has an average pore size that is smaller than the PCV-2 antigen and prevents passage of at least 90% of the PCV-2 antigen through the semi-permeable membrane pores. Preferably the average pore size of the semi-permeable membrane prevents passage of at least 90% of proteins of 50 kDa to 500 kDa in size, more preferably, at least 90% of proteins of 75 kDa to 400 kDa in size, and most preferably at least 90% of proteins of 100 kDa to 300 kDa in size. This pore size is preferred, when the PCV-2 antigen is produced as whole virus or as virus like particles.

Preferably, further purification to obtain PCV-2 antigenic composition comprising a purified PCV-2 antigen as defined herein, can be achieved by performing further purification step comprising iii) purifying the harvest of step ii) comprising the PCV-2 antigen (of any methods described herein), which is obtained after the removal of a portion of the first liquid, by a chromatography a step. In order to obtain a higher purity grade a second chromatography step can be done, which however is different from the first one. For instance if the first purification step / chromatography step is size exclusion (gel filtration) the second should different from that e.g. an affinity chromatography, ion exchange chromatography, etc. Preferably, if the first step to purify PCV-2 antigen, preferably to purify PCV-2 ORF2 antigen is a size exclusion (gel filtration) chromatography, the second step can be ion-exchange chromatography, preferably anion-exchange chromatography (AIEX). A preferred anion-exchange chromatography matrix for the purification of PCV-2 antigen, preferably the PCV-2 ORF2 antigen is Q Sepharose. In a small scale of about 50 ml, use of 5 ml HiTrap Q Sepharose HP columns are most preferred. The anion exchange chromatography can be conducted, for instance, as described in Example 3. Briefly, about 50 ml of the void volume fraction pool from the size exclusion chromatography step can be loaded onto the AIEX column at a flow rate of 3.0 ml/min. Following a washing step using, for instance, 20 mM Tris, pH 6.5, 5 mM DTT to remove unbound material, protein can be eluted with a single step of 8 column volumes of the following buffer (20 mM Tris, pH 6.5, 5 mM DTT, 1.0 M NaCl) The flow-through from the AIEX run can be loaded back onto the Q Sepharose column and eluted as described above to increase the yield. This two step technique (size exclusion

followed by anion-exchange chromatography) effectively separates PCV-2 ORF2 antigen from most of the other protein components of the culture harvest.

In a further aspect, the virucidal activity of the PCV-2 antigenic composition produced by the methods described herein is reduced by at least 10% as compared to the liquid that has not undergone the method. More preferably, the virucidal activity of the PCV-2 antigenic composition is reduced by at least 50% as compared to the first liquid that has not undergone the method. Still more preferably, the virucidal activity of the PCV-2 antigenic composition is reduced by at least 70% as compared to the first liquid that has not undergone the method.

Thus according to a further aspect, the present application relates to PCV-2 antigenic composition obtained by a method comprising the steps of i) obtaining a first liquid containing a PCV-2 antigen, ii) removing at least a portion of the first liquid from the PCV-2 antigen, wherein the virucidal activity - preferably in respect to PRRS virus - of the PCV-2 antigenic composition obtained after step ii) is reduced by at least 10%, preferably at least 50%, more preferably at least 70%, even more preferably at least 90% as compared to that of the first liquid. Preferably, the portion of the first liquid having virucidal activity is removed from the PCV-2 antigen by an exchange of a portion of the first liquid against a second liquid. The exchange is preferably done in such that it comprises the steps of a) adding the second liquid to the first liquid which contains the PCV-2 antigen and b) concentrating the PCV-2 antigen, preferably from 3X to 50X, even more preferably from 4X to 20X, and even more preferably from 7X to 10X in comparison to the volume of the first liquid by removing a portion of the first and second liquids from the PCV-2 antigen. Preferably the liquid addition step is performed substantially simultaneously or sequentially with the concentrating step as described above. When the concentrating step and liquid addition step are performed sequentially, the order of the steps does not matter. Moreover, the concentration step is preferably done by filtration - preferably by dia- and/or ultra-filtration, utilizing a filter, which preferably contains a semi-permeable membrane. The semi-permeable membrane preferably has an average pore size that is smaller than the PCV-2 antigen and prevent passage of at least 90% of the PCV-2 antigen through the semi-permeable membrane pores. Preferably the average pore size of the semi-permeable membrane or of any other filter that is used herein, prevents passage of at least 90% of proteins of 50 kDa to 500 kDa in size, more preferably, at least 90% of proteins of 75 kDa to 400 kDa in size, and most preferably at least 90% of proteins of 100 kDa to 300 kDa in size. This pore size is preferred, when the PCV-2 antigen is produced as whole virus or as virus like particles. Further purification to obtain a purified PCV-2 antigen can be done as described above.

According to a further aspect, the present application relates to a PCV-2 antigenic composition obtained by a method described herein, wherein the PCV-2 antigenic composition causes a loss of less than 1 log $TCID_{50}$ – preferably per ml - , preferably less than 0.9 log $TCID_{50,}$ - preferably per ml -, even more preferably less than 0.7 log $TCID_{50}$ - preferably per ml -, even more preferably less than 0.5 log $TCID_{50}$ - preferably per ml -, most preferably less than 0.3 log $TCID_{50}$ - preferably per ml - of a live virus, preferably of a live PRRSV or less than 1 log CFU - preferably per ml -, preferably less than 0.9 log CFU - preferably per ml -, even more preferably less than 0.7 log CFU - preferably per ml -, even more preferably less than 0.5 log CFU - preferably per ml -, most preferably less than 0.3 log CFU - preferably per ml - of a live bacterium, preferably of *Mycoplasma hyopneumoniae*, when the live virus, preferably PRRSV or live bacterium, preferably *Mycoplasma hyopneumoniae* is mixed and incubated with the PCV-2 antigenic composition for 2 or more hours, preferably for more than 4 hours, even more preferably for more than 12 hours, even more preferably for more than 24 hours, even more preferably for more than 2 days, even more preferably for more than 4 days, even more preferably for more than 7 days, even more preferably for more than 2 weeks, even more preferably for more than 4 weeks, even more preferably for more than 2 months, even more preferably for more than 3 months, even more preferably for more than 4 months, even more preferably for more than 6 months, even more preferably for more than 9 months, even more preferably for more than 12 months, even more preferably for more than 18 months, and most preferably for more than 2 years. The live virus can be any live virus, but preferably the live virus is the PRRS virus, preferably the PRRS virus having the ATCC accession number VR 2332. The live bacterium can be any bacterium, but is preferably the *Mycoplasma hyopneumoniae* bacterium, preferably the J-strain of *Mycoplasma hyopneumoniae*. The $TCID_{50}$ per ml can be estimated by a standard in vitro titration assay which allow the estimation of the amount of a live virus. The CFU per ml can be determined also by a standard in vitro titration assay which allows the estimation of the amount of a live bacterium. The term "per ml" preferably refers to 1 ml of a fluid.

In a further aspect, the present patent application relates to a PCV-2 antigenic composition that is obtained by a method described above, further comprising the step of harvesting the PCV-2 antigen remaining after step ii). This harvesting can be done in any conventional manner. In a particularly preferred manner of harvesting, the portion of the first liquid is removed from the PCV-2 antigen via a filtration step and the PCV-2 antigen is recovered or harvested from the filter retard.

In a further aspect, the PCV-2 antigenic composition obtained by any of the methods described herein is admixed with a further component selected from the group consisting of pharmaceutically acceptable carriers, adjuvants, diluents, excipients, and combinations thereof. Preferably, the further component is an adjuvant, even more preferably wherein the adjuvant is a polymer of acrylic or methacrylic acid, and still more preferably wherein the adjuvant is Carbomer.

Thus, according to a further aspect, the present application provides a PCV-2 antigenic composition obtained by a method described above, further comprising the step of admixing the PCV-2 antigen obtained by the method described herein with a further component selected from the group consisting of pharmaceutically acceptable carriers, adjuvants, diluents, excipients, and combinations thereof. Preferably the further component is an adjuvant, even more preferably wherein the adjuvant is a polymer of acrylic or methacrylic acid, and still more preferably wherein the adjuvant is Carbomer. Preferably, the adjuvant is added in an amount of about 100 μg to about 10 mg per dose. Even more preferably the adjuvant is added in an amount of about 100 μg to about 10 mg per dose. Still more preferably the adjuvant is added in an amount of about 500 μg to about 5 mg per dose. Still more preferably the adjuvant is added in an amount of about 750 μg to about 2.5 mg per dose. Most preferably the adjuvant is added in an amount of about 1 mg per dose.

In a further aspect, the PCV-2 antigenic composition described above comprises the ORF-2 protein of PCV-2, more preferably recombinant ORF-2 protein of PCV-2, and still more preferably virus like particles of ORF-2 protein. Thus, according to a further aspect of the present application, the present application provides a PCV-2 antigenic composition obtained by a method described above, wherein the PCV-2 antigen comprises the ORF-2 protein of PCV-2, more preferably recombinant ORF-2 protein of PCV-2, and still more preferably virus like particles of ORF-2 protein.

As mentioned above, the PCV-2 antigen used in the method described herein can be obtained by any method known in the art. Preferably, the PCV-2 antigen is obtained via a viral vector, preferably a recombinant baculovirus viral vector, containing and expressing the PCV-2 antigen, preferably, PCV-2 ORF-2. In preferred forms, the PCV-2 antigen is obtained following the procedures described in WO2006/072065 (the teachings and content of which were previously incorporated by reference). Thus, according to a further aspect of the present application, the present application provides a PCV-2 antigenic composition obtained by a method described above, wherein the PCV-2 antigen is obtained via a viral vector, preferably a recombinant baculovirus viral vector, containing and expressing the PCV-2 antigen,

preferably, PCV-2 ORF-2, and wherein the PCV-2 antigen comprises the ORF-2 protein of PCV-2, more preferably recombinant ORF-2 protein of PCV-2, and still more preferably virus like particles of ORF-2 protein.

In a further aspect of the present application, the PCV-2 antigenic composition is obtained by the method described above and further comprises the step of inactivating the recombinant baculovirus viral vector with a DNA inactivating agent, preferably in the presence of about 1 to about 20 mM of binary ethylenimine. In preferred forms, the method further comprises the step of adding an amount of an agent that neutralizes the DNA inactivating agent, the amount being equivalent to the amount of the DNA inactivating agent wherein the agent that neutralizes the DNA inactivating agent comprises a sodium thiosulfate solution concentrated to a final concentration of about 1 to about 20 mM and wherein the DNA inactivating agent is BEI. Preferably, the inactivating step is performed after at least a portion of the first liquid is removed from the PCV-2 antigen.

In a further aspect of the present application, the PCV-2 antigenic composition is obtained by the method described above further comprising the steps of admixing the PCV-2 antigen obtained after the inactivating and neutralizing steps. Thus, according to a further aspect, the present application provides a PCV-2 antigenic composition obtained by a method described above comprising the steps of i) obtaining a PCV-2 antigen in a first liquid; ii) removing at least a portion of the first liquid from the PCV-2 antigen; iii) inactivating the recombinant baculovirus viral vector with a DNA inactivating agent, preferably in the presence of about 1 to about 20 mM of binary ethylenimine; iv) adding an amount of a neutralizing agent that neutralizes the inactivating agent, the amount of neutralizing agent being equivalent to the amount of the inactivating agent, wherein the neutralizing agent preferably comprises a sodium thiosulfate solution preferably concentrated to a final concentration of about 1 to about 20 mM and wherein the inactivating agent preferably comprises BEI; and, preferably step v), comprising admixing the PCV-2 antigen obtained in step iv) with a further component selected from the group consisting of pharmaceutically acceptable carriers, adjuvants, diluents, excipients, and combinations thereof.

In a further aspect of the present application, the PCV-2 antigenic composition described above, preferably obtained by the methods described above, further comprises at least one additional antigen, preferably a viral or bacterial antigen, and more preferably a viral or bacterial antigen from at least one other disease-causing organism in swine. In a further aspect the at least one additional antigen is Porcine Reproductive and Respiratory Syndrome Virus. Even more preferably, the Porcine Reproductive and Respiratory Syndrome Virus

antigen comprises a live virus, and still more preferably a modified live virus. Still more preferably, the modified live Porcine Reproductive and Respiratory Syndrome Virus antigen comprises a modified live virus strain of ATCC Accession Number VR 2332, and still more preferably comprises INGELVAC® PRRS MLV. In a further aspect of the present application, the at least one additional antigen is *Mycoplasma hyopneumoniae*. Preferably the *Mycoplasma hyopneumoniae* antigen is a bacterin, and more preferably, the *Mycoplasma hyopneumoniae* bacterin is INGELVAC® MYCOFLEX. In a further aspect of the present application, the PCV-2 antigenic composition described above, preferably obtained by the methods described above further comprises Porcine Reproductive and Respiratory Syndrome Virus antigen, preferably a modified live Porcine Reproductive and Respiratory Syndrome Virus, still more preferably, the Porcine Reproductive and Respiratory Syndrome Virus having the ATCC Accession Number VR 2332, or the Porcine Reproductive and Respiratory Syndrome Virus included in INGELVAC® PRRS MLV or INGELVAC® PRRS ATP. In a further aspect of the present application, the PCV-2 antigenic composition described above, preferably obtained by the methods described above further comprises *Mycoplasma hyopneumoniae,* preferably *Mycoplasma hyopneumoniae* bacterin, and more preferably INGELVAC® MYCOFLEX or the *Mycoplasma hyopneumoniae* bacterin included in INGELVAC® MYCOFLEX. In a further aspect, the PCV-2 antigenic composition described herein, comprises a Porcine Reproductive and Respiratory Syndrome Virus, preferably any one of those described above and a *Mycoplasma hyopneumoniae,* preferably any one of those described above.

When the PCV-2 antigenic composition comprising the at least one additional antigen from at least one other disease-causing organism in swine as described above, preferably Porcine Reproductive and Respiratory Syndrome Virus and/or *Mycoplasma hyopneumoniae* antigen is obtained by a method described herein, the method comprises the steps of i) obtaining a PCV-2 antigen in a first liquid; ii) removing at least a portion of the first liquid from the PCV-2 antigen; and combining the PCV-2 antigen with at least one additional antigen, preferably a viral or bacterial antigen, and more preferably a viral or bacterial antigen from at least one other disease-causing organism in swine. Preferably, the PCV-2 antigen comprises the ORF-2 protein of PCV-2, more preferably recombinant ORF-2 protein of PCV-2, and still more preferably virus like particles of ORF-2 protein. Preferably, the portion of the first liquid is removed from the PCV-2 antigen by an exchange of a portion of the first liquid against a second liquid. The exchange is preferably done such that it comprises the steps of a) adding the second liquid to the first liquid which contains the PCV-2 antigen and

b) concentrating the PCV-2 antigen, preferably from 3X to 50X, even more preferably from 4X to 20X, and even more preferably from 7X to 10X in comparison to the volume of the first liquid by removing a portion of the first and second liquids from the PCV-2 antigen. Preferably, the liquid addition step and concentration step are performed multiple times, preferably two times, even more preferably three times. In such cases, not only the first liquid is removed, but also a mixture of the first and second liquid. Preferably each liquid addition step is performed substantially simultaneously or sequentially as described above. When the concentrating step and liquid addition step are performed sequentially, the order of the steps does not matter. Moreover, the concentration step is preferably done by filtration - preferably by dia- or ultrafiltration, utilizing a filter, which preferably contains a semi-permeable membrane. The semi-permeable membrane preferably has an average pore size that is smaller than the PCV-2 antigen and prevents passage of at least 90% of the PCV-2 antigen through the semi-permeable membrane pores and withholds the PCV-2 antigen within the filter for harvesting or recovery. Preferably the average pore size of the semi-permeable membrane or of any other filter that is used herein, prevents passage of at least 90% of proteins of 50 kDa to 500 kDa in size, more preferably, at least 90% of proteins of 75 kDa to 400 kDa in size, and most preferably at least 90% of proteins of 100 kDa to 300 kDa in size. This pore size is preferred, when the PCV-2 antigen is produced as whole virus or as virus like particles.

The present invention as defined above, provides new methods of producing a PCV-2 antigen and immunogenic compositions comprising a PCV-2 antigen, wherein the PCV-2 antigen shows a reduced virucidal activity and/or increased immunogenicity (each as defined herein), wherein the method comprises the steps of i) obtaining a first liquid containing a PCV-2 antigen, ii) removing at least a portion of the first liquid from the PCV-2 antigen. Moreover, the present invention also provides a PCV-2 antigen as well as immunogenic compositions comprising such PCV-2 antigen showing a reduced virucidal activity and/or increased immunogenicity (each as defined herein). According to a further aspect, the PCV-2 antigen as well as the immunogenic compositions comprising a purified PCV-2 antigen showing a reduced virucidal activity and/or increased immunogenicity can alternatively be obtained by the following method (II). The purified PCV-2 antigen according to the invention, preferably the purified PCV-2 ORF2 antigen, can be obtained by the purification of a PCV-2 virus preparation, in particular by the purification of the whole virus. Whole virus preparations are described for instance in WO 99/18214 or WO 03/049703. Moreover, purified PCV-2 antigen can also be obtained by the purification of a recombinant expressed

PCV-2 antigen, preferably by the purification of a recombinant PCV-2 ORF2 antigen. Expression systems for the production of recombinant PCV-2 antigen, preferably for the production of recombinant PCV-2 ORF2 antigens are well known in the art and include, but not limited to, bacterial expression systems, yeast expression systems, insect cell or mammalian expression systems. Vectors and methods for making and/ or using vectors (or recombinants) for the expression of the PCV-2 antigens are described in the application elswhere.

Preferred cells are those susceptible for infection with an appropriate recombinant viral vector, containing a PCV-2 ORF2 DNA and expressing the PCV-2 ORF2 protein. Preferably the cells are insect cells, and more preferably, they include the insect cells sold under the trademark SF+ insect cells (Protein Sciences Corporation, Meriden, CT). Preferred cell cultures have a cell count between about 0.3 - 2.0 x 10^6 cells/mL, more preferably from about 0.35 - 1.9 x 10^6 cells/mL, still more preferably from about 0.4 - 1.8 x 10^6 cells/mL, even more preferably from about 0.45 - 1.7 x 10^6 cells/mL, and most preferably from about 0.5 - 1.5 x 10^6 cells/mL.

Preferred viral vectors include baculovirus such as BaculoGold (BD Biosciences Pharmingen, San Diego, CA), in particular provided that the production cells are insect cells. Although the baculovirus expression system is preferred, it is understood by those of skill in the art that other expression systems, including those described above will work for purposes of the present invention, namely the expression of PCV-2 ORF2 antigen.

Appropriate growth media will also be determinable by those of skill in the art with a preferred growth media being serum-free insect cell media such as Excell 420 (JRH Biosciences, Inc., Lenexa, KS) and the like.

The recombinant viral vector containing the PCV-2 ORF2 DNA sequences has a preferred multiplicity of infection (MOI) of between about 0.03 - 1.5, more preferably from about 0.05 - 1.3, still more preferably from about 0.09 - 1.1, and most preferably from about 0.1 - 1.0, when used for the infection of the susceptible cells. Preferably the MOIs mentioned above relates to one mL of cell culture fluid. Preferably, the method described herein comprises the infection of 0.35 - 1.9 x 10^6 cells/mL, still more preferably of about 0.4 - 1.8 x 10^6 cells/mL, even more preferably of about 0.45 - 1.7 x 10^6 cells/mL, and most preferably of about 0.5 - 1.5 x 10^6 cells/mL with a recombinant viral vector containing a PCV-2 ORF2 DNA and expressing the PCV-2 ORF2 antigen protein having a MOI (multiplicity of infection) of between about 0.03 - 1.5, more preferably from about 0.05 - 1.3, still more preferably from about 0.09 - 1.1, and most preferably from about 0.1 - 1.0.

The infected cells are then incubated over a period of up to ten days, more preferably from about two days to about ten days, still more preferably from about four days to about nine days, and most preferably from about five days to about eight days. Preferred incubation conditions include a temperature between about 22 - 32°C, more preferably from about 24 - 30°C, still more preferably from about 25 - 29°C, even more preferably from about 26 - 28°C, and most preferably about 27°C. Preferably, the SF+ cells are observed following inoculation for characteristic baculovirus-induced changes. Such observation may include monitoring cell density trends and the decrease in viability during the post-infection period. It was found that peak viral titer is observed 3 - 5 days after infection and peak PCV-2 ORF2 antigen production in cells is obtained between days 5 and 8 post infection and/or when cell viability decreases to less than 10%.

The PCV-2 ORF2 antigen can be purified from the harvest by standard methods known to a person skilled in the art, for example by those described in Protein purification methods – a practical approach (E.L.V. Harris and S. Angal, eds., IRL Press at Oxford University Press). Those methods include, but are not limited to, separation by centrifugation and/or filtration, precipitation, size exclusion (gel filtration) chromatography, affinity chromatography, metal chelate chromatography, ion-exchange chromatography covalent chromatography, hydrophobic interaction chromatography, etc.

The recovery process of the PCV-2 antigen, preferably the PCV-2 ORF2 antigen, preferably begins with the separation of cell debris from the expressed PCV-2 ORF2 antigen via a separation step. Preferred separation steps include filtration, centrifugation at speeds up to about 20,000xg, continuous flow centrifugation, chromatographic separation using ion exchange or gel filtration, and conventional immunoaffinity methods. Those methods are known to persons skilled in the art for example by (E.L.V. Harris and S. Angel (eds.), Protein purification methods – a practical approach, IRL Press Oxford 1995). The most preferred separation methods include centrifugation at speeds up to about 20,000xg and filtration. Preferred filtration methods include dead-end microfiltration and tangential flow (or cross flow) filtration including hollow fiber filtration dead-end micro filtration. Of these, dead-end microfiltration is preferred. Preferred pore sizes for dead-end microfiltration are between about 0.30 - 1.35 μm, more preferably between about 0.35 - 1.25 μm, still more preferably between about 0.40 - 1.1 μm, and most preferably between about 0.45 - 1.0 μm. It is believed that any conventional filtration membrane will work for purposes of the present invention and polyethersulfone membranes are preferred. Any low weight nucleic acid species are removed during the filtration step.

Further purification of PCV-2 antigen, preferably of the PCV-2 ORF2 antigen can be achieved with chromatography procedures, preferably a two-step chromatography procedure. However it is also possible to start with the chromatography procedure in the event, the loading material does not comprise cell debris.

If the PCV-2 antigen is assembled to virus like particles (VLP), the first step is preferably a size exclusion (gel filtration) chromatography, which can be done, for instance, by using a Sephacryl S300 matrix. In lab scale use of HiPrep 26/60 Sephacryl S300HR columns are most preferred. However, any other size exclusion chromatography matrices known to a person skilled in the art can be used, which allow the separation of the PCV-2 ORF2 VLPs from the culture filtrate or supernatant. Suitable matrices are described, for instance, in E.L.V. Harris and S. Angel (eds.), Protein purification methods – a practical approach, IRL Press Oxford 1995). The gel filtration chromatography can be conducted, for instance, by loading the column with the crude preparation comprising the PCV-2 antigen with a flow-rate of 1.0 ml/min and eluting the column with 1.5 column volume of a buffer comprising 20 mM Tris, pH 6.5, 5 mM DTT. However, the PCV-2 ORF2 antigen can also be purified by using affinity chromatography, for instance, via selective binding to an immobilized PCV-2 ORF2 specific antibody, or any other method known to a person skilled in the art.

Thus according to a preferred embodiment, the immunogenic composition comprising a purified PCV-2 antigen, preferably a purified PCV-2 ORF2 antigen, and the adjuvant, is obtainable by a process comprising the steps

a) Expressing the PCV-2 antigen, prefer the PCV-2 ORF2 antigen in a host cell;
b) Harvesting the cell culture obtaining PCV-2 antigen, preferably the PCV-2 ORF2 antigen;
c) Purifying the harvest comprising the PCV-2 antigen, preferably the PCV-2 ORF2 antigen by size exclusion chromatography (gel filtration);
d) Admixing the purified PCV-2 antigen, preferably the PCV-2 ORF2 antigen with an adjuvant.

According to a preferred embodiment, the size exclusion chromatography is performed as described herein, preferably as described in Example 3. Preferably, the size exclusion results in an immunogenic composition having purity grade of more than 80% (w/w), preferably more than 90% (w/w) with reference to the total amount of protein included in the immunogenic composition prior to the mixture with the adjuvant. The purity grade can

be estimated by Imperial Protein Stain (Pierce) staining after SDS PAGE via NuPAGE 10% Bis-Tris gels (Invitrogen) using the NuPAGE MOPS buffer system (Invitrogen).

In order to obtain a higher purity grade a second chromatography step can be done, which however is different from the first one. For instance if the first purification step / chromatography step is size exclusion (gel filtration) the second should different from that e.g. an affinity chromatography, ion exchange chromatography, etc.

Preferably, if the first step to purify PCV-2 antigen, preferably to purify PCV-2 ORF2 antigen is a size exclusion (gel filtration) chromatography, the second step can be ion-exchange chromatography, preferably anion-exchange chromatography (AIEX). A preferred anion-exchange chromatography matrix for the purification of PCV-2 antigen, preferably the PCV-2 ORF2 antigen is Q Sepharose. In a small scale of about 50 ml, use of 5 ml HiTrap Q Sepharose HP columns are most preferred. The anion exchange chromatography can be conducted, for instance, as described in Example 3. Briefly, about 50 ml of the void volume fraction pool from the size exclusion chromatography step can be loaded onto the AIEX column at a flow rate of 3.0 ml/min. Following a washing step using, for instance, 20 mM Tris, pH 6.5, 5 mM DTT to remove unbound material, protein can be eluted with a single step of 8 column volumes of the following buffer (20 mM Tris, pH 6.5, 5 mM DTT, 1.0 M NaCl) The flow-through from the AIEX run can be loaded back onto the Q Sepharose column and eluted as described above to increase the yield. This two step technique (size exclusion followed by anion-exchange chromatography) effectively separates PCV-2 ORF2 antigen from most of the other protein components of the culture harvest.

Thus according to a preferred embodiment, the immunogenic composition comprising a purified PCV-2 antigen, preferably the PCV-2 ORF2 antigen, and the adjuvant, is obtainable by a process comprising the steps

a) Expressing the PCV-2 antigen, prefer the PCV-2 ORF2 antigen in a host cell;
b) Harvesting the cell culture obtaining PCV-2 antigen, preferably the PCV-2 ORF2 antigen;
c) Purifying the harvest comprising the PCV-2 antigen, preferably the PCV-2 ORF2 antigen by size exclusion chromatography (gel filtration) followed by anion exchange chromatography; and
d) Admixing the purified PCV-2 antigen, preferably the PCV-2 ORF2 antigen with an adjuvant.

According to a preferred embodiment, the size exclusion chromatography and the

anion exchange chromatography are performed as described herein, preferably as described in Example 3. Preferably, the two step purification strategy results in an immunogenic composition having purity grade of more than 90% (w/w), preferably more than 95% (w/w) with reference to the total amount of protein included in the immunogenic composition prior to the mixture with the adjuvant. The purity grade can be estimated by Imperial Protein Stain (Pierce) staining after SDS PAGE via NuPAGE 10% Bis-Tris gels (Invitrogen) using the NuPAGE MOPS buffer system (Invitrogen).

As described above, the recovery process of the PCV-2 antigen, preferably the PCV" ORF2 antigen begins with the separation of cell debris from the expressed PCV-2 ORF2 antigen via a separation step. A preferred separation step includes a micro filtration through a filter having a pore size of about 0.6 μm to about 2 μm, preferably having a pore size of about 0.8 mm to about 1.2 μm.

Thus the immunogenic composition comprising a purified PCV-2 antigen, preferably the PCV-2 ORF2 antigen, and the adjuvant, is obtainable by a process comprising the steps

a) Expressing the PCV-2 antigen, prefer the PCV-2 ORF2 antigen in a host cell;
b) Harvesting the cell culture obtaining PCV-2 antigen, preferably the PCV-2 ORF2 antigen;
c) Filtering the harvest obtained under step b) through a filter having a pore size of 0.6 to 2.0 μm.
d) Purifying the filtrate comprising the PCV-2 antigen, preferably the PCV-2 ORF2 antigen and obtained under step c)by size exclusion chromatography (gel filtration) optionally followed by anion exchange chromatography; and
e) Admixing the purified PCV-2 antigen, prefer the PCV-2 ORF2 antigen with an adjuvant.

According to a preferred embodiment, the micro-filtration, size exclusion chromatography and the anion exchange chromatography are performed as described herein, preferably as described in Example 3. Preferably, the two step purification strategy including the pre-filtration step results in an immunogenic composition having purity grade of more than 90% (w/w), preferably more than 95% (w/w) with reference to the total amount of protein included in the immunogenic composition prior to the mixture with the adjuvant. The purity grade can be estimated by Imperial Protein Stain (Pierce) staining after SDS PAGE via NuPAGE 10% Bis-Tris gels (Invitrogen) using the NuPAGE MOPS buffer system (Invitrogen).

The immunogenic compositions comprising the purified PCV-2 antigen, preferably the purified PCV-2 ORF2 antigen described herein, preferably those obtainable by the methods described herein are characterized by an increased immunogenicity as compared to an immunogenic composition not comprising such a purified PCV-2 antigen or purified PCV-2 ORF2 antigen.

In the event, viral vectors such as a recombinant poxvirus, adenovirus or baculovirus is used to produce the PCV-2 antigen, preferably the PCV-2 ORF2 antigen, it is recommended to inactivate the viral nucleic acid by an appropriate inactivation treatment. Such inactivation may occur anytime during the purification of the PCV-2 antigen, preferably the PCV-2 ORF2 antigen. Thus, inactivation may occur immediately after the harvest of the cell culture fluid comprising the PCV-2 antigen, preferably the PCV-2 ORF2 antigen, or after the micro-filtration of the of PCV-2 antigen, preferably of the PCV-2 ORF2 antigen, if micro-filtration is done, prior or after the purification step, for instance, prior to or after the gel filtration, and prior to or after the anion exchange chromatography, if this is done.

Any conventional inactivation method can be used for purposes of the present invention. Thus, inactivation can be performed by chemical and/or physical treatments. In preferred forms, the volume of harvest fluids is determined and the temperature is brought to between about 32°C - 42°C, more preferably between about 34°C - 40°C, and most preferably between about 35°C - 39°C. Preferred inactivation methods include the addition cyclized binary ethylenimine (BEI), preferably in a concentration of about 1 to about 20 mM, preferably of about 2 to about 10 mM, still more preferably of about 2 to about 8 mM, still more preferably of about 3 to about 7 mM, most preferably of about 5 mM. For example the inactivation includes the addition of a solution of 2-bromoethyleneamine hydrobromide (BEA), preferably of about 0.4M, which has been cyclized to 0.2M binary ethylenimine (BEI) in 0.3N NaOH, to the fluids to give a final concentration of about 5mM BEI. Preferably, the fluids are then stirred continuously for 2 - 96 hours and the inactivated harvest fluids can be stored frozen at - 40°C or below or between about 1°C - 7°C. After inactivation is completed a sodium thiosulfate solution, preferably at 1.0M is added to neutralize any residual BEI. Preferably, the sodium thiosulfate is added in equivalent amount as compared to the BEI added prior to for inactivation. For example, in the event BEI is added to a final concentration of 5mM, a 1.0M sodium thiosulfate solution is added to give a final minimum concentration of 5 mM to neutralize any residual BEI.

Prior to the mixing of the purified PCV-2 antigen, preferably of the PCV-2 ORF2

antigen with an adjuvant, it is also recommended to dialyze the purified PCV-2 antigen, preferably the PCV-2 ORF2 antigen against phosphate buffered saline, pH 7.4 or any other physiological buffer.

The methods described above result in an PCV-2 antigen with reduced virucidal activity as defined herein as well as in an improved immunogenicity, if the PCV-2 antigen has a purity grade of more than 50% (w/w), preferably of more than 70% (w/w), even more preferred of more than 80% (w/w), even more preferred of more than 85% (w/w), even more preferred of more than 90% (w/w), most preferred of more than 95% (w/w) with reference to the total amount of protein included in the immunogenic composition prior to the mixture with any adjuvant. However, the purified PCV-2 antigen obtainable according to this method II can also be mixed and used together with an adjuvant, preferably with any of the adjuvants described herein. The preferred adjuvant is a Carbopol, preferably in a concentration of about 0.1 to 10 mg/ml, more preferred in a concentration of 0.5 to 5 mg/ml, most preferably of about 1 mg/ml of the final immunogenic composition.

Again, the present invention does not only provides any of the methods described herein, including the alternative method II, it also provides a PCV-2 antigen, preferably a purified PCV-2 antigen, most preferably a purified PCV-2 ORF-2 protein obtainable by any of the methods described herein, including the alternative method II. Moreover, the present invention also provides PCV-2 antigenic compositions comprising a PCV-2 antigen, preferably a purified PCV-2 antigen, most preferably a purified PCV-2 ORF-2 protein obtainable by any of the methods described herein, including the alternative method II. The amount of the PCV-2 antigen, in particular of the purified PCV-2 ORF2 antigen in the final immunogenic composition should be in a range from about 0.25 to about 400 µg per dose with reference to the final immunogenic composition. Preferably the finally immunogenic composition should include an amount of PCV-2 antigen, preferably of PCV-2 ORF2 antigen in a range from about 2 to about 200 µg/dose, even more preferably from about 3 to about 150 µg/dose, still more preferably from about 4 to about 100 µg/dose, still more preferably from about 5 to about 80 µg/dose, still more preferably from about 6 to about 60 µg/dose, even more preferably from about 7 to about 50 µg/dose, even more preferably from about 8 to about 40 µg/dose, still more preferably from about 8 to about 32 µg/dose, even more preferably from about 8 to about 24 µg/dose, and most preferred from about 8 to about 16 µg/dose.

The immunogenic compositions provided herewith, including those obtainable by the method II comprises one or more additional antigens of another disease-causing organism.

Those "another disease-causing organisms" are defined above. Preferably the additional antigen is Porcine Reproductive and Respiratory Syndrome Virus. Even more preferably, the Porcine Reproductive and Respiratory Syndrome Virus antigen comprises a live virus, and still more preferably a modified live virus. Still more preferably, the modified live Porcine Reproductive and Respiratory Syndrome Virus antigen comprises a modified live virus strain of ATCC Accession Number VR 2332, and still more preferably comprises INGELVAC® PRRS MLV. In a further aspect of the present application, the additional antigen is *Mycoplasma hyopneumoniae*. Preferably the *Mycoplasma hyopneumoniae* antigen is a bacterin, and more preferably, the *Mycoplasma hyopneumoniae* bacterin is INGELVAC® MYCOFLEX. Most preferred are combinations with, both antigen of Porcine Reproductive and Respiratory Syndrome Virus and *Mycoplasma hyopneumoniae*.

Due to the increased immunogenicity of the immunogenic composition including the purified PCV-2 antigen, preferably the purified PCV-2 ORF2 antigen provided herewith, the immunogenic compositions can be used for reducing the incidence or reducing the severity of clinical signs caused by or being associated with PCV-2 infections as compared to an animal not receiving that immunogenic composition.

The term "reduction in the incidence of or severity of clinical signs" shall mean that any of such signs are reduced in incidence or severity in animals receiving an administration of the vaccine in comparison with a "control group" of animals when both have been infected with or challenged by the pathogen from which the immunological active component(s) in the vaccine are derived and wherein the control group has not received an administration of the vaccine or immunogenic composition. In this context, the term "decrease" or "reduction" means a reduction of at least 10%, preferably 25%, even more preferably 50%, most preferably of more than 100% in the vaccinated group as compared to the control group not vaccinated.

As used herein, "clinical symptoms" or "clinical signs" shall refer to signs of infection from a pathogen that are directly observable from a live animal such as symptoms. Representative examples will depend on the pathogen selected but can include things such as nasal discharge, lethargy, coughing, elevated fever, weight gain or loss, dehydration, diarrhea, swelling, lameness, and the like. PCV-2 clinical signs can include wasting, paleness of the skin, unthriftiness, respiratory distress, diarrhea, icterus, and jaundice.

Reducing in the incidence of or the severity of clinical signs caused by or being associated with PCV-2 infections in an animal can be reached by the administration of only a single dose of such immunogenic composition to an animal in need of such treatment.

However, the immunogenic composition provided herewith can also be administered in two doses or more doses, with an interval of 2 to 4 weeks between the administration of the first dose and the any subsequent dose. Thus, according to a further embodiment the immunogenic composition provided herewith including the purified PCV-2 antigen, preferably the purified PCV-2 ORF2 antigen can be administered in one, two or more doses to an animal in need thereof.

In particular, in a further aspect of the present application, an immunogenic composition comprising a PCV-2 antigenic composition as described above is provided wherein the immunogenic composition, when administered to an animal, reduces lymphoid depletion and inflammation by at least 80% in an animal as compared to an animal not receiving the immunogenic composition. Thus, in a further aspect of the present application, an immunogenic composition is provided comprising a PCV-2 antigenic composition as described above and the immunogenic composition reduces lymphoid depletion and inflammation by at least 80% in an animal that has received an administration of the immunogenic composition as compared to an animal not receiving the immunogenic composition.

In a further aspect of the present application, an immunogenic composition comprising a PCV-2 antigenic composition as described above is provided, wherein the immunogenic composition, when administered to an animal, reduces lung lesions by at least 80% in an animal as compared to an animal not receiving the immunogenic composition. Thus, in a further aspect of the present application, an immunogenic composition comprising a PCV-2 antigenic composition is provided as described above and the immunogenic composition reduces lung lesions by at least 80% in an animal that has received an administration of the immunogenic composition as compared to an animal not receiving the immunogenic composition.

In a further aspect of the present invention, an immunogenic composition comprising a PCV-2 antigenic composition, as described above, is provided wherein the immunogenic composition induces a protective immune response against PCV-2 after the administration of one dose of the immunogenic composition. The immunogenic composition comprising a PCV-2 antigenic composition can be of any volume including 1 ml, 2 ml, 3 ml, 4 ml, 5 ml and higher. In preferred forms, 2 ml of the immunogenic composition comprises one dose of the PCV-2 antigen. Thus, in a further aspect of the present invention, an immunogenic composition as described above is provided wherein the immunogenic composition comprising a PCV-2 antigenic composition induces a protective immune response against

PCV-2 after the administration of one dose of the immunogenic composition. In a further aspect, 2 ml of the immunogenic composition comprises one dose of the PCV-2 antigen.

As used herein, a "protective immune response" refers to a reduced incidence of or reduced severity of clinical, pathological, or histopathological signs or symptoms of infection from a pathogen of interest up to and including the complete prevention of such signs or symptoms.

The term "Pathological" signs shall refer to signs of infection that are observable at the microscopic or molecular level, through biochemical testing, or with the naked eye upon necropsy. For PCV-2, pathological signs will include microscopic and macroscopic lesions on multiple tissues and organs, with lymphoid organs being the most common site for lesions.

The term "Histopathological" signs shall refer to signs of tissue changes resulting from infection.

The terms, "clinical symptoms" or "clinical signs" are defined above.

In a further aspect of the present invention, an immunogenic composition comprising a PCV-2 antigenic composition and a PRRRS antigen, preferably any one of the PRRS antigens described herein, as described above, is provided wherein the immunogenic composition induces a protective immune response against PRRS virus after the administration of one dose of the immunogenic composition. Again, any dosage volume can be produced, but in preferred forms, 2 ml of the immunogenic composition comprises one dose of the PRRS antigen and one dose of the PCV-2 antigen. Thus, in a further aspect of the present invention, an immunogenic composition as described above comprising a PRRSV and a PCV-2 antigenic composition as described herein, is provided wherein the immunogenic composition induces a protective immune response against PRRS after the administration of one dose of the immunogenic composition. In a further aspect, 2 ml of the immunogenic composition comprises one dose of the PRRS antigen and one dose of the PCV-2 antigen.

In a further aspect of the present invention, an immunogenic composition comprising a PCV-2 antigenic composition as described herein and *Mycoplasma hyopneumoniae* antigen as described above, is provided wherein the immunogenic composition induces a protective immune response against *Mycoplasma hyopneumoniae* after the administration of one dose of the immunogenic composition. Again, any dosage volume can be produced, but in preferred forms, 2 ml of the immunogenic composition comprises one dose of the *Mycoplasma hyopneumoniae* antigen and one dose of a PCV-2 antigen. Thus, in a further aspect of the present invention, an immunogenic composition as described above is provided wherein the immunogenic composition induces a protective immune response against *Mycoplasma*

hyopneumoniae after the administration of one dose of the immunogenic composition comprising a PCV-2 antigenic composition as described herein and *Mycoplasma hyopneumoniae* antigen. In a further aspect, 2 ml of the immunogenic composition comprises one dose of the *Mycoplasma hyopneumoniae* antigen.

In a further aspect of the present application, an immunogenic composition, as described above, is prepared for the administration of 2 ml per dose.

In a further aspect of the present application, a method of reducing one or more clinical symptoms of a PCV-2 infection in an animal as compared to an animal not receiving the immunogenic composition is provided. In general, the method comprises the step of administering to an animal any of the immunogenic compositions comprising a PCV-2 antigenic or composition as described above. Preferably, one or more clinical symptoms of a PCV-2 infection are reduced after the administration of a single dose of the or immunogenic composition. Thus, according to a further aspect of the present application, a method of reducing one or more clinical symptoms of a PCV-2 infection in an animal as compared to an animal not receiving the immunogenic composition comprising a PCV-2 antigenic composition as described herein is provided. In general, the method comprises the step of administering to an animal any of the immunogenic compositions comprising a PCV-2 antigenic composition described above, wherein one or more clinical symptoms of a PCV-2 infection are reduced, preferably after the administration of a single dose of the immunogenic composition comprising a PCV-2 antigenic composition as described herein.

In a further aspect of the present application, a method of reducing one or more clinical symptoms of a PRRS infection in an animal as compared to an animal not receiving the immunogenic composition is provided. In general, the method comprises the step of administering to an animal any of the immunogenic compositions described above comprising a PCV-2 antigenic composition as described herein and a PRRS Virus as described herein. Preferably, one or more clinical symptoms of a PRRS infection are reduced after the administration of a single dose of the immunogenic composition comprising a PCV-2 antigenic composition as described herein and a PRRS Virus as described herein. Thus, according to a further aspect of the present application, a method of reducing one or more clinical symptoms of a PRRS infection in an animal as compared to an animal not receiving the immunogenic composition comprising a PCV-2 antigenic composition as described herein and a PRRS Virus as described herein, is provided. Clinical signs of Porcine Reproductive and Respiratory Syndrome Virus (PRRSV) include, but are not limited to inappetance, fever, abortion, transient discoloration, prolonged anoestrus, coughing, respiratory signs, mastitis,

agalactia, lethargy, mummified piglets, stillbirths, weak piglets at birth, reduction in farrowing rate, early farrowing, diarrhea, wasting, sneezing, eye discharge, pale skin, mortality, and combinations thereof.

In a further aspect of the present application, a method of reducing one or more clinical symptoms of a *Mycoplasma hyopneumoniae* infection in an animal as compared to an animal not receiving the immunogenic composition comprises a PCV-2 antigenic composition as described herein and a *Mycoplasma hyopneumoniae* antigen as described herein, is provided. In general, the method comprises the step of administering to an animal any of the immunogenic compositions described above. Preferably, one or more clinical symptoms of a *Mycoplasma hyopneumoniae* infection are reduced after the administration of a single dose of the immunogenic composition comprising a PCV-2 antigenic composition as described herein and a *Mycoplasma hyopneumoniae* antigen as described herein. Thus, according to a further aspect of the present application, a method of reducing one or more clinical symptoms of a *Mycoplasma hyopneumoniae* infection in an animal as compared to an animal not receiving the immunogenic composition comprising a PCV-2 antigenic composition as described herein and a *Mycoplasma hyopneumoniae* antigen as described herein is provided. Clinical signs of *Mycoplasma hyopneumoniae* (*M. hyo*) infection include, but are not limited to a dry cough, impaired performance, and lung lesions.

The immunogenic composition comprising the purified PCV-2 antigen, preferably the PCV-2 ORF2 antigen as provided herein, has improved immunogenicity. Therefore, the immunogenic composition provided herewith is suitable to improve the immune response in an animal receiving such immunogenic composition. Thus, according to a further embodiment, the present invention provides a method for improving the immune response in an animal against PCV-2 comprising the step: adminstistering a the immunogenic composition as described herein and having a purified PCV-2 antigen, preferably a purified PCV-2 ORF-2 protein as provided herewith, to an animal in need thereof. According to a preferred aspect, the PCV-2 antigen, preferably the PCV-2 ORF2 antigen used in such method is purified to an extent of more than 60% (w/w), preferably more than 60% (w/w), even more preferred to more than 70% (w/w), even more preferred to more than 80% (w/w), even more preferred to more than 90% (w/w), most preferred to more than 95% (w/w) with reference to the total amount of protein included in the immunogenic composition. The purity grade can be estimated by Imperial Protein Stain (Pierce) staining after SDS PAGE via NuPAGE 10% Bis-Tris gels (Invitrogen) using the NuPAGE MOPS buffer system (Invitrogen). The PCV-2, and preferably the PCV-2 ORF2 can be purified using conventional

methods well known to a person skilled in the art.

DESCRIPTION OF THE DRAWINGS

The following drawings form part of the present specification and are included to further demonstrate certain aspects of the present invention. The invention may be better understood by reference to one or more of these drawings in combination with the detailed description of specific embodiments presented herein.

FIG. 1 shows the results of ultrafiltration configuration pilot scale using a10% Bis-Tris/MOPS gel that demonstrates the presence of the ORF 2 after the filtration process. Lanes were loaded as follows: (1) marker; (2) n/a; (3) n/a; (4) 24 - 180/181 Pre conc – 20 µl; (5) 25 - 180/181 1x antigens – 20 µl; (6) 26 - 180/181 filter wash – 20 µl; (7) 27 - PCV 504 Preconc – 8 µl; (8) 28 - PCV 504 Perm – 20 µl; (9) 29 - PCV 504 1X – 20 µl; (10) 092704PD – 20 µl; (11) marker.

DETAILED DESCRIPTION

The following examples set forth preferred materials and procedures in accordance with the present invention. It is to be understood, however, that these examples are provided by way of illustration only, and nothing therein should be deemed a limitation upon the overall scope of the invention.

EXAMPLE 1

This example describes a laboratory scale and pilot scale process for manufacturing concentrated PCV-2 ORF2 antigen that will have a reduced virucidal activity in comparison to manufacturing processes that do not include the steps of the present invention. Specifically, the effects that the present invention has on the virucidal activity of the PCV-2 ORF2 antigen on PRRS virus will be determined.

Materials and Methods

Production Antigen:
Laboratory Scale:
PCV SUB037 H1-F, 18.94 kg
PCV 1025, 20.6 kg
PCV 180/181, 20.0 kg
PCV SUB 504PD, 40kg

Pilot Scale:
PCV SUB 506PD, 362 kg
PCV SUB 507PD, 384 kg

PCV SUB512PD, 430 kg
PCV SUB 513PD, 405 kg

Ultra Filtration Cartridges: GE Healthcare, Steam-In-Place (SIP), hollow fiber membrane cartridges

UFP-100-E-55-STM: 100,000 NMWC, 1mm diameter tubule; used in UF-002 in X109, laboratory scale.

UFP-300-E-55-STM: 300,000 NMWC, 1mm diameter tubule; used in UF-002 in X109, laboratory scale.

UFP-100-E-65-MSM: 100,000 NMWC, 1mm diameter tubule; used in UF-B2614, in APU-1, pilot scale.

UFP-300-E55-SMO: 300,000 NMWC, 1mm diameter tubule; used in UF-2713in VP-1, pilot scale.

The following Ultrafiltration equipment was used in the feasibility evaluation and initial process development:

Table 1. Equipment

Process Step	Procedure	Equipment	Identification
Ultrafiltration	Antigen Concentration	Flex-Stand Concentrator	UF-002 in X109, laboratory scale
		UF Skid	UF-B2614 in APU-1, pilot scale
		UF Skid	UF-2713 in VP-1, pilot scale

Manufacturing Process:

Ultrafiltration (UF) Configuration: Laboratory Scale

50 liter carboys containing filtered, inactivated, neutralized, PCV-2 ORF2 material generated in building P were used in the concentration process with the GE Healthcare (Amersham) Flex Stand 30L size UF skid #002.

The initial concentration processes used a "batch" diafiltration scheme, whereby approximately 20kg of antigen material was transferred to the UF skid and concentrated through a 100,000 NMWC hollow fiber cartridge (UFP-100-E-55-STM). The 100,000

NMWC concentration processes used PCV-2 ORF2 lots SUB037PD and PCV1025 material from PD and Manufacturing, respectively.

These two initial runs were concentrated to approximately 4x of the original volume and were Q.S.'d (quantity substantiated) in the feed tank back to the original transfer volume. The concentrated material was treated in this manner for a total of 2 concentrations per lot number, with the third and final concentration harvested as a concentrate and a portion Q.S.'d to 1X of the original volume. Samples were drawn pre-concentration, at each concentration step and at each Q.S. step. Permeate samples were drawn during each concentration step.

The next two consecutive runs concentrated the PCV-2 ORF2 antigen without a saline wash. The concentrated material was sampled at approximately 4x and then at final concentration. Approximately 20kg of antigen material was transferred to the UF skid holding tank and concentrated through a 300,000 NMWC hollow fiber cartridge (UFP-300-E-55-STM). A second 20L volume was added to the holding tank with the concentrate from the first 20L. This was concentrated to final volume. The 300,000 NMWC concentration processes used PCV-2 ORF2 lots PCV 180/181 pool and SUB504PD generated by Manufacturing and PD, respectively. Samples were drawn pre-concentration and at each concentration step. Permeate samples were drawn during each concentration step.

Ultrafiltration Configuration: Pilot Scale

Pilot scale processes utilized SUB lots 506PD, 512PD and513PD. Antigen pre-concentration volumes ranged from approximately 350L to 430L. Lots SUB506PD and SUB513PD were transferred to DSP (down-stream processing) 2602 and concentrated with UF-B2614 in APU-1 using a 100,000 NMWC filter (UFP-100-65-E-MSM), with 4.2 m^2 surface area. SUB512PD was transferred to DSP 2701 and concentrated with UF-2713 using a 300,000 NMWC filter (UFP-300-E-SMO) with 2.1 m^2 surface area. Final concentrated material was harvested for each lot and stored at 4°C for analysis.

Results and Conclusions

Ultrafiltration (UF) Configuration: Laboratory Scale

Filtration with 100,000 NMWC (100kDa) versus 300,000 NMWC (300kDa) filters was comparable in concentration times and was feasible when considering a full scale process. Filtration times for the 100kDa filter, concentrating 4X, with approximately 18L to 26L, ranged from 14 minutes to 32 minutes, with the shorter times resulting from the saline

wash steps (Table 2). Filtration times for the 300kDa filter, concentrating 3.2X, 7X for PCV180/181and 21.5X for SUB504PD with approximately 40L of material concentrated in two consecutive 20L volumes yielded 3.2X at 25 minutes, 7X at 23 minutes and 21.5X at 32 minutes. Some time variation is expected due to the time taken to get a concentration process to a target trans-membrane pressure (TMP) of 10.25 psi.

Process flux values ranged from 27.43 lmh to 32.00 lmh for the PCV180/181 lot, with the 32.00 lmh value resulting from a spike toward the end of the concentration process. Flux values for SUB 504PD material were 28.57 lmh during the first-20L concentration and 35.71 lmh during the 2nd-20L concentration. (Tables 3 and 4)

Table 2. Process Data

Lot #	Starting Concentration Volume	X Conc.	Time to Concentrate (min)
PCV 037 QS-0 (100kDa)	18.94	4.17	24
PCV 037 QS-1 (100kDa)	18.76	4.75	16
PCV 037 QS-2 (100kDa)	19.05	4.70	14
PCV 1025 QS-0 (100kDa)	20.59	4.39	28
PCV 1025 QS-1 (100kDa)	20.24	4.65	29
PCV 1025 QS-2 (100kDa)	18.71	3.6	17
PCV 180/181 conc-1 (300kDa)	20	3.50	25
PCV 180/181 conc-2 (300kDa)	26.01	7.00	23
SUB 504PD conc-2 (300kDa)	24.72	21.50	32

Table 3. Process Data

PCV 180/181: 300,000 NMWC Filter				
	TIME	TMP	PERM FLOW (ml/min)	FLUX (lmh)
Conc-1	9:55	9	N/A	N/A
Conc-2	10:23	12.5	N/A	N/A
Conc-2	10:30	12.5	960	27.43
Conc-2	10:33	14.5	960	27.43
Conc-2	10:36	10.5	1120	32.00
Conc-2	10:39	11	810	23.14

Table 4. Process Data

SUB504PD: 300,000 NMWC Filter				
	TIME	TMP	PERM FLOW (ml/min)	FLUX (lmh)
Conc-1	15:29	9.5	1000	28.57
Conc-2	16:00	10.25	1250	35.71

Change in potency post filtration was found unchanged when the concentrated material was Q.S.'d back to 1X volume, as with SUB 037 reconstituted material and PCV 1025 reconstituted material. Concentrate antigen content values pushed the limits of the assay beyond the validated approximate 64 µg limit, as is seen in the values in tables 5 through 8 where antigen content amounts are compared to the expected calculated amounts. Permeate values from the concentrations performed using SUB 037, PCV 180/181 and SUB 504PD antigens showed no significant loss of material due to filtration. All permeate antigen content amounts fell into the undetectable range of the assay. PCV 1025 antigen permeate antigen content amounts were not collected.

Table 5. SUB 037 Change in PCV-2 Antigen Content (in µg)

Lot Number /vol	PreConc. Antigen Content	PostConc. Antigen Content	Concentration Factor	Calc Antigen Content	Change from Calculated Antigen Content	Gain/Loss from Calc RP
SUB 037 (18.94kg)--4.7x—100kDa--PDX	56	137.6	4.7	263.2	-125.6	loss
Concentrated and Reconstituted with Saline: QS-1	56	61.6	1	56	5.6	gain
Concentrated and Reconstituted with Saline: QS-2	56	62.7	1	56	6.7	gain
Concentrated and Reconstituted with Saline: QS-3	56	55.8	1	56	-0.2	loss
SUB 037 permeates 1,2,3	--	0			No loss	

Table 6. PCV 1025 Change in PCV-2 Antigen Content (in µg)

Lot Number /vol	PreConc. Antigen Content	PostConc. Antigen Content	Concentration Factor	Calc Antigen Content	Change from Calculated Antigen Content	Gain/Loss from Calc RP
1025 (20.46kg)--4.5X--100kDa--PDX	70.88	288.64	4.5	318.96	-30.32	loss
Concentrated and Reconstituted with Saline-1	$N/A_{(NO\ SAMPLE)}$	N/A	1	N/A	N/A	no sample
Concentrated and Reconstituted with Saline-2	70.88	66.24	1	70.88	-4.64	loss
Concentrated and Reconstituted with Saline-3	70.88	76.00	1	70.88	5.12	gain
1025 permeate	--	N/A			N/A	

Table 7. Change in PCV-2 Antigen Content (in µg)

Lot Number /vol	PreConc. Antigen Content	PostConc. Antigen Content	Concentration Factor	Calc Antigen Content	Change from Calculated Antigen Content	Gain/Loss from Calc RP
180/181 (40.3kg)--3.5X--300kDa--PDX	43.36	90.8	3.5	151.76	-60.96	loss
180/181 **7.2X**	43.36	247.04	7.2	312.19	-65.15	loss
180/181 permeate	--	0			No loss	N/A

Table 8. Change in PCV-2 Antigen Content (in µg)

Lot Number /vol	PreConc. Antigen Content	PostConc. Antigen Content	Concentration Factor	Calc Antigen Content	Change from Calculated Antigen Content	Gain/Loss from Calc RP
SUB504 (40.43kg) 4.3X—300kDa--PDX	22.24	68.16	4.3	95.63	-27.47	loss
SUB504 **20X**	22.24	448.48	20	444.8	3.68	gain
SUB504 permeate	--	0			No loss	

SDS-PAGE gels were run with material from PCV 180/181 and SUB 504PD in R&D. The ORF2 band residing at approximately 27 kDa in Figure 1 was consistent with the banding pattern of the reference in lane 10. This band size has previously been determined to be ORF2. The permeate material from SUB 504PD, 300kDa filtration concentration, exhibited an absence of banding at the 27kDa site. No ORF2 protein was apparently lost with this pore size filter. The gel was run under reducing conditions.

Virucidal activity of the pre-concentrate antigen, concentrated antigen, reconstituted antigen and filtration permeate was tested against the PRRS virus vaccine. Initial results from Quality Control (QC) for SUB 037 were unsatisfactory for the pre-concentrated and concentrated (100kDa filter) material, which had been reconstituted back to 1X with saline. However, when the concentrated material was formulated into vaccines, concentrations up to 80% inclusion in the vaccine reduced this virucidal activity to a satisfactory level well below the acceptance limit of 0.7 log/ml loss of PRRS virus titer. Permeate material from SUB 037 showed borderline passing to unsatisfactory levels of virucidal activity. See Table 9.

PCV 1025 pre-concentrated material was found unsatisfactory for virucidal activity to PRRS with loss in PRRS titer at 1.5 log/ml loss. The three saline reconstituted concentrate (100kDa filter) materials were passing at 0.5 log/ml loss and 0.6 log/ml loss with one of the reconstituted concentrates satisfactory with "no change" in PRRS titer. Permeate samples were not tested for this lot. See Table 10.

PCV 180/181 pre-concentrated vaccine material was unsatisfactory for virucidal activity to PRRS for 2 of the 3 vaccines prepared. Percent antigen inclusion levels ranged from 37.0% to 55.5%. The highest % inclusion pre-concentrate vaccine was found satisfactory.

Vaccines prepared from 1X (concentrate reconstituted to 1X with saline) material were found satisfactory for virucidal activity to PRRS virus. Percent antigen inclusion levels ranged from 44% to 66%. See Table 11.

SUB 504PD vaccines prepared from pre-concentrated antigen with 79.5% vaccine inclusion were satisfactory for virucidal activity to PRRS virus. Vaccines prepared from 4.3X concentrated antigen with 23.5-35% vaccine inclusion and from 21.5X concentrated antigen with 3.5-5.5% vaccine inclusion were also satisfactory. Last, the filter wash antigen prepared with 72% inclusion level was found satisfactory for virucidal activity to PRRS virus.

Table 9. SUB 037 Virucidal Activity

Sample ID	Change in Potency log/ml	Sat/Unsat
SUB 037 PreConc (18.94kg)-- 4.7x--100kDa--PDX Anitgen Content 56 pre / 137.6 post	1.4	unsat
Concentrated and Reconstituted with Saline-1	0.8	unsat
Concentrated and Reconstituted with Saline-2	1.3	unsat
Concentrated and Reconstituted with Saline-3	1.3	unsat
SUB 037 permeate-1	0.6	sat
SUB 037 permeate-2	1.0	unsat
SUB 037 permeate-3	0.5	sat
Vaccine: 20% inclusion Ames**	-0.2	sat
Vaccine: 40% inclusion Ames**	-0.2	sat
Vaccine: 60% inclusion Ames**	0.2	sat
Vaccine: 80% inclusion Ames**	0.1	sat

Table 10. PCV 1025 Virucidal Activity

Sample ID	Change in Potency log/ml	Sat/Unsat
PCV1025 PreConc (20.46kg)--4.5x--100kDa--PDX Anitgen Content 70.8 pre / 288.64 post	1.5	unsat
Concentrated and Reconstituted with Saline-1	0.6	sat
Concentrated and Reconstituted with Saline-2	no change	sat
Concentrated and Reconstituted with Saline-3	0.5	sat
1025 permeate	No submission	n/a

Table 11. PCV 180/181 Virucidal Activity

Sample ID	% Vaccine Inclusion	Change in Potency log/ml	Sat/Unsat
PCV180/181 (49.3kg)--3.5X--300kDa--PDX Antigen Content = 43.36 µg pre/Antigen Content (1) 90.88 µg/Antigen Content (2) 247.04 µg	n/a	n/a	n/a
Preconcentrate vaccine Antigen Content = 16 µg/8.8 µg Actual	37.0	1.2 log/ml loss	unsat
Preconcentrate vaccine Antigen Content = 19.2 µg/8.8 µg Actual	44.5	1.2 log/ml loss	unsat
Preconcentrate vaccine Antigen Content = 24 µg/15.36 µg Actual	55.5	0.8 log/ml gain	sat
7X reconstituted to 1x vaccine Antigen Content = 16 µg/8.48 µg Actual	44.0	0.6 log/ml gain	sat
7X reconstituted to 1x vaccine Antigen Content = 19.24 µg/12.32 µg Actual	53.0	no change	sat
7X reconstituted to 1x vaccine Antigen Content = 24 µg/14.08 µg Actual	66.0	0.3 log/ml gain	sat

Table 12. SUB 504PD Virucidal Activity

Sample ID	% Vaccine Inclusion	Change in Potency log/ml	Sat/Unsat
SUB504PD (~40kg) 4.3X--300kDa--PDX Antigen Content = 22.24 µg pre/Antigen Content (1) = 68.16 µg Antigen Content (2) = 448.48 µg	n/a	n/a	n/a
Preconcentrate vaccine Antigen Content = 16 µg/10.24 µg Actual	79.5	0.3 log/ml loss	sat
4.3X vaccine Antigen Content = 16 µg/18.56 µg Actual	23.5	0.1 log/ml gain	sat
4.3X vaccine Antigen Content = 19.2 µg/3.04 µg Actual	28.0	0.7 log/ml gain	sat
4.3X vaccine Antigen Content = 24 µg/5.28 µg Actual	35.0	0.1 log/ml gain	sat
21.5X vaccine Antigen Content = 16 µg/10.08 µg Actual	3.5	0.1 log/ml loss	sat
21.5X vaccine Antigen Content = 19.2 µg/15.2 µg Actual	4.5	0.1 log/ml gain	sat
21.5X vaccine Antigen Content = 24 µg/3.36 µg Actual	5.5	0.3 log/ml gain	sat
Filter Wash Antigen Content = 16 µg/12.96 µg Actual	72.0	0.7 gain	sat

Table 13. Process Data

SUB 506: 100,000 NMWC FILTER			
TIME	TMP (psi)	PERM FLOW (ml/min)	FLUX (lmh)
13:25	12	800	11.43
13:59	11.5	3300	47.14
15:09	12.5	3800	54.29
15:29	12	2100	30.00

Table 14. Process Data

SUB 513: 100,000 NMWC FILTER			
TIME	TMP (psi)	PERM FLOW (ml/min)	FLUX (lmh)
6:49	11.5	2600	37.14
8:06	11.5	2700	38.57

Table 15. Process Data

SUB 512: 300,000 NMWC FILTER			
TIME	TMP (psi)	PERM FLOW (ml/min)	FLUX (lmh)
15:25	16	5000	142.86
18:00	13	800	22.86
20:03	17.5	3500	100.00
21:05	17.5	3000	85.71
21:47	18	2500	71.43
21:59	12.5	4000	114.29

Discussion

Porcine Circovirus Vaccine, Type 2, Killed Baculovirus Vector is a global product manufactured by Boehringer Ingelheim Vetmedica, Inc., in St. Joseph, Missouri and used in the INGELVAC CIRCOFLEX® product. At harvest, virus fluids are aseptically filtered through one or more 2-15μm pre-filters, and then a 0.8-1.0μm filter for final filtration. BEI (binary ethylenimine) stock solution is added to the harvest fluids to a final concentration of 5mM BEI. The fluids are stirred continuously for a minimum of 72 hours and a maximum of 96 hours and may be stored frozen at ≤ 40°C or at 4°C ± 3°C. A 1.0M sodium thiosulfate solution is added to a final concentration of 5mM to neutralize any residual BEI.

The neutralized antigen is blended with 0.5% Carbopol solution to 20% v/v with the PCV-2 ORF2 protein content in the final product adjusted by the addition of saline to meet the minimum release requirements of a relative potency greater than, or equal to, 1.0. After bulking, the serial may be stored at 4°C or filled.

PCV-2 ORF2 material was concentrated post-neutralization by hollow fiber cartridge ultrafiltration. The concentrated material was further processed with two diafiltration volumes of saline solution. Preferred ultrafiltration nominal molecular weight cut-off (NMWC) pore size were determined to include 100,000 NMWC and 300,000 NMWC, each with a 1.0mm tubule lumen diameter. Both pore sizes were included to provide flexibility in manufacturing in the event of interrupted supply of filter cartridges by the manufacturer. Sodium dodecyl sulfate – polyacrylamide gel electrophoresis (SDS PAGE) gels and potency data indicated no difference in the antigen protein or the potency between the two filter pore sizes.

EXAMPLE 2

This example compares the relative yields of ORF2 using methods of the present invention with methods that are known in the prior art. It is understood that this example represents one of many possible methods for obtaining PCV-2 ORF2 for use with the present methods and compositions.

Materials and Methods

Four 1000mL spinner flasks were each seeded with approximately $1.0x10^6$ Sf+ cells/ml in 300mL of insect serum free media, Excell 420 (JRH Biosciences, Inc., Lenexa, KS). The master cell culture is identified as SF+ (*Spodoptera frugiperda*) Master Cell Stock, passage 19, Lot#N112-095W. The cells used to generate the SF+ Master Cell Stock were obtained from Protein Sciences Corporation, Inc., Meriden, CT. The SF+ cell line for this example was confined between passages 19 and 59. Other passages will work for purposes of the present invention, but in order to scale the process up for large scale production, at least 19 passages will probably be necessary and passages beyond 59 may have an effect on expression, although this was not investigated. In more detail, the initial SF+ cell cultures from liquid nitrogen storage were grown in Excell 420 media in suspension in sterile spinner flasks with constant agitation. The cultures were grown in 100 mL to 250mL spinner flasks with 25 to 150 mL of Excell 420 serum-free media. When the cells had multiplied to a cell density of 1.0 - 8.0 x 10^6 cells/mL, they were split to new vessels with a planting density of 0.5 - 1.5 x 10^6 cells/mL. Subsequent expansion cultures were grown in spinner flasks up to 36 liters in size or in stainless steel bioreactors of up to 300 liters for a period of 2-7 days at 25 - 29°C.

After seeding, the flasks were incubated at 27°C for four hours. Subsequently, each flask was seeded with a recombinant baculovirus containing the PCV-2 ORF2 gene (SEQ ID

NO: 4). The recombinant baculovirus containing the PCV-2 ORF2 gene was generated as follows: the PCV-2 ORF2 gene from a North American strain of PCV-2 was PCR amplified to contain a 5' Kozak's sequence (SEQ ID NO: 1) and a 3' EcoR1 site (SEQ ID NO: 2), cloned into the pGEM-T-Easy vector (Promega, Madison, WI). Then, it was subsequently excised and subcloned into the transfer vector pVL1392 (BD Biosciences Pharmingen, San Diego, CA). The subcloned portion is represented herein as SEQ ID NO: 7. The pVL1392 plasmid containing the PCV-2 ORF2 gene was designated N47-064Y and then co-transfected with BaculoGold® (BD Biosciences Pharmingen) baculovirus DNA into Sf+ insect cells (Protein Sciences, Meriden, CT) to generate the recombinant baculovirus containing the PCV-2 ORF2 gene. The new construct is provided herein as SEQ ID NO: 8. The recombinant baculovirus containing the PCV-2 ORF2 gene was plaque-purified and Master Seed Virus (MSV) was propagated on the SF+ cell line, aliquotted, and stored at -70°C. The MSV was positively identified as PCV-2 ORF2 baculovirus by PCR-RFLP using baculovirus specific primers. Insect cells infected with PCV-2 ORF2 baculovirus to generate MSV or Working Seed Virus express PCV-2 ORF2 antigen as detected by polyclonal serum or monoclonal antibodies in an indirect fluorescent antibody assay. Additionally, the identity of the PCV-2 ORF2 baculovirus was confirmed by N-terminal amino acid sequencing. The PCV-2 ORF2 baculovirus MSV was also tested for purity in accordance with 9 C.F.R. 113.27 (c), 113.28, and 113.55. Each recombinant baculovirus seeded into the spinner flasks had varying multiplicities of infection (MOIs). Flask 1 was seeded with 7.52mL of .088 MOI seed; flask 2 was seeded with 3.01mL of 0.36MOI seed; flask 3 was seeded with 1.5mL of 0.18MOI seed; and flask 4 was seeded with 0.75mL of 0.09MOI seed.

After being seeded with the baculovirus, the flasks were then incubated at 27 ± 2°C for 7 days and were also agitated at 100rpm during that time. The flasks used ventilated caps to allow for air flow. Samples from each flask were taken every 24 hours for the next 7 days. After extraction, each sample was centrifuged, and both the pellet and the supernatant were separated and then microfiltered through a 0.45-1.0 μm pore size membrane.

Results and Conclusions

The resulting samples then had the amount of ORF2 present within them quantified via an ELISA assay. The ELISA assay was conducted with capture antibody Swine anti-PCV-2 Pab IgG Prot. G purified (diluted 1:250 in PBS) diluted to 1:6000 in 0.05M Carbonate buffer (pH 9.6). 100 μL of the antibody was then placed in the wells of the mictrotiter plate, sealed, and incubated overnight at 37°C. The plate was then washed three times with a wash solution which comprised 0.5mL of Tween 20 (Sigma, St. Louis, MO), 100mL of 10X D-

PBS (Gibco Invitrogen, Carlsbad, CA) and 899.5mL of distilled water. Subsequently, 250 µL of a blocking solution (5g Carnation Non-fat dry milk (Nestle, Glendale, CA) in 10mL of D-PBS QS to 100mL with distilled water) was added to each of the wells. The next step was to wash the test plate and then add pre-diluted antigen. The pre-diluted antigen was produced by adding 200 µL of diluent solution (0.5mL Tween 20 in 999.5mL D-PBS) to each of the wells on a dilution plate. The sample was then diluted at a 1:240 ratio and a 1:480 ratio, and 100 µL of each of these diluted samples was then added to one of the top wells on the dilution plate (i.e. one top well received 100 µL of the 1:240 dilution and the other received 100 µL of the 1:480 dilution). Serial dilutions were then done for the remainder of the plate by removing 100 µL form each successive well and transferring it to the next well on the plate. Each well was mixed prior to doing the next transfer. The test plate washing included washing the plate three times with the wash buffer. The plate was then sealed and incubated for an hour at 37°C before being washed three more times with the wash buffer. The detection antibody used was monoclonal antibody to PCV ORF2. It was diluted to 1:300 in diluent solution, and 100 µL of the diluted detection antibody was then added to the wells. The plate was then sealed and incubated for an hour at 37°C before being washed three times with the wash buffer. Conjugate diluent was then prepared by adding normal rabbit serum (Jackson Immunoresearch, West Grove, PA) to the diluent solution to 1% concentration. Conjugate antibody Goat anti-mouse (H+l)-HRP (Jackson Immunoresearch) was diluted in the conjugate diluent to 1:10,000. 100 µL of the diluted conjugate antibody was then added to each of the wells. The plate was then sealed and incubated for 45 minutes at 37°C before being washed three times with the wash buffer. 100 µL of substrate (TMB Peroxidase Substrate, Kirkgaard and Perry Laboratories (KPL), Gaithersberg, MD), mixed with an equal volume of Peroxidase Substrate B (KPL) was added to each of the wells. The plate was incubated at room temperature for 15 minutes. 100 µL of 1N HCL solution was then added to all of the wells to stop the reaction. The plate was then run through an ELISA reader.

The results of this assay are provided in Table 17 below:

Table 17

Day	Flask	ORF2 in pellet (µg)	ORF2 in supernatant (µg)
3	1	47.53	12
3	2	57.46	22
3	3	53.44	14
3	4	58.64	12

Day	Flask	ORF2 in pellet (µg)	ORF2 in supernatant (µg)
4	1	43.01	44
4	2	65.61	62
4	3	70.56	32
4	4	64.97	24
5	1	31.74	100
5	2	34.93	142
5	3	47.84	90
5	4	55.14	86
6	1	14.7	158
6	2	18.13	182
6	3	34.78	140
6	4	36.88	146
7	1	6.54	176
7	2	12.09	190
7	3	15.84	158
7	4	15.19	152

These results indicate that when the incubation time is extended, expression of ORF2 into the supernatant of the centrifuged cells and media is greater than expression in the pellet of the centrifuged cells and media. Accordingly, allowing the ORF2 expression to proceed for at least 5 days and recovering it in the supernate rather than allowing expression to proceed for less than 5 days and recovering ORF2 from the cells, provides a great increase in ORF2 yields, and a significant improvement over prior methods.

EXAMPLE 3

Purification of ORF2 was achieved by microfiltration process followed by a two-step chromatography scheme. The harvest obtained in Example 1 was filtered trough a micro filter membrane having a pore size of 1.2 µm. The micorfiltrate was then purified by size exclusion (gel filtration) using a HiPrep 26/60 Sephacryl S300HR column. A starting sample of 20 ml of the filtrate comprising the PCV-2 ORF2 was loaded on the HiPrep 26/60 Sephacryl S300HR column at a flow rate of 1.0 ml/min and was eluted with 1.5 column volumes of Buffer A (20 mM Tris, pH 6.5, 5 mM DTT). Eight milliliter fractions were

collected during the elution step. Fractions No. 10 - 16 (milititers 10 to 16 of the eluate) from the size exclusion chromatography were pooled and utilized as the starting sample for anion exchange (AIEX) chromatography. These fractions represent the void volume of the sizing column, which is where the PCV-2 ORF2 elutes due to the large molecular weight of PCV-2 ORF2 protein. This technique effectively separates ORF2 from most of the other protein components of an antigen sample.

AIEX was performed using a 5 ml HiTrap Q Sepharose HP column. Approximately 48 ml of the void volume fraction pool from the size exclusion experiment was loaded onto the AIEX HiTrap Q Sepharose HP column at a flow rate of 3.0 ml/min. Following a washing step with loading Buffer A (20 mM Tris, pH 6.5, 5 mM DTT) to remove unbound material, protein was eluted with a single step of 8 column volumes of Buffer B (20 mM Tris, pH 6.5, 5 mM DTT, 1.0 M NaCl) and 5 ml fractions were collected. Peak fractions No. 8 and 9 were collected and pooled. The flow-through from the AIEX run was loaded back onto the Q Sepharose column and eluted as described above. From the second run, fractions No 7, 8, and 9 were pooled with the fractions from the first run. A third run of the flow-through material did not result in a significant peak fraction in the eluate, so no fractions were saved from that run.

The fraction pool of approximately 25 ml was dialyzed overnight at 4°C against 2 liter of phosphate buffered saline, pH 7.4 (Gibco). After dialysis, ORF2 was >95% pure based on SDS-PAGE analysis.

EXAMPLE 4

Solutions of 2-bromoethylamine hydrobromide (BEA), sodium hydroxide (NaOH), and sodium thiosulfate ($Na_2S_2O_3$) were prepared. BEA solution was made by weighing out 1.63 g of BEA (Sigma, B65705, lot 05316EE) and dissolving in 20 ml of purified water (dH_2O, aqua dest., here: 'water'). The final concentration of this solution was 0.4 M BEA. The NaOH solution was made by weighing 0.33 g of NaOH (JTBaker, 3722-01, lot E01470) and dissolving in 20 ml of water. The final concentration of this solution was 0.41 M NaOH. Sodium thiosulfate ($Na_2S_2O_3$) solution was prepared by weighing 25 g of $Na_2S_2O_3$ (Sigma S7026, lot 106K0178) and dissolving in 100 ml of water. Once dissolved, the solution was filtered through a 0.2 µm bottle-top filter to sterilize. The final concentration of this solution was 1.0 M $Na_2S_2O_3$.

To prepare binary ethylenimine (BEI), 20 ml of 0.4 M BEA solution was mixed with 20 ml of 0.41 M NaOH and the initial pH was determined to be ~12.5 – 14.0. The mixture

was incubated at 37°C for one hour and the pH was checked again. The pH after incubation was ~7.0 – 7.5, and this indicated a successful cyclization reaction of BEA into BEI. The final concentration of BEI was calculated to be ~0.2 M (20 ml of 0.4 M BEA cyclized with an excess of base (0.41 M) in a volume of 40 ml.

The inactivation reactions were as follows (per 100 ml material to be inactivated): The materials to be inactivated were mixed with 2.5 ml freshly prepared BEI. The inactivation reactions were incubated for 72 hr at 37°C with stirring to continually mix the solutions. After 72 hr, the reactions were neutralized by addition 0.5 ml of 1.0 M sodium thiosulfate. After allowing the thiosulfate to completely mix into the solutions (~15 min of mixing), the inactivated and neutralized materials were stored at 4°C prior to formulation with adjuvant.

EXAMPLE 5

Preparation of the test samples:

In order to estimate the immunogenicity of highly purified ORF2 antigen (purity grade of higher than 90%) as compared to non- or less purified ORF2 antigen, 5 ml batches of several test samples were prepared:

Table 18: Test samples

Test sample No	Description
#1	Highly purified ORF2 antigen, inactivated with BEI and mixed with 1mg/ml Carbopol
#2	Highly purified ORF2 antigen mixed with insect cell debris inactivated with BEI and mixed with 1mg/ml Carbopol
#3	Insect cell debris (mock control)
#4	PCV-2 ORF2 antigen, non filtered, non purified mixed with 1mg/ml Carbopol
#5	PCV-2 ORF2 antigen, non filtered, non purified, inactivated with BEI and mixed with 1mg/ml Carbopol
#6	PCV-2 ORF2 antigen, non purified, inactivated with BEI and mixed with 1mg/ml Carbopol

Test sample #1 was produced as follows: PCV-2 ORF2 antigen was produced as described in Example 1 and highly purified as described in Example 3. The highly purified PCV-2 ORF 2 antigen was inactivated with BEI as described in Example 4. After BEI inactivation, the PCV-2 ORF2 antigen was content was adjusted to an amount of about 32 to 32,5 µg per ml test sample and mixed with 1 mg Carbopol 971P (BF Goodrich, Ohio, USA) per ml test sample.

Test sample #2 was produced as follows: PCV-2 ORF2 antigen was produced as described in Example 1 and highly purified as described in Example 3. The highly purified

PCV-2 ORF 2 antigen was inactivated with BEI as described in Example 4. After BEI inactivation, PCV-2 ORF2 antigen was mixed with insect cell debris and Carbopol. The final test sample included about 2.06 x 10^6 insect cells, about 32 to 32.5 µg and 1 mg Carbopol 971P per ml test sample.

Test sample #3 was prepared by mixing about 2.06 x 10^6 insect cells with 1 mg Carbopol 971P per ml test sample. Before mixing, insect cells were inactivated by BEI as described in Example 3.

Test sample #4 was produced as follows: the PCV-2 ORF2 antigen was produced as described in Example 1. The PCV-2 ORF2 antigen content in the supernatant was adjusted to an amount of about 32 to 32.5 µg per ml test sample and mixed with 1 mg Carbopol 971P per ml test sample.

Test sample #5 was produced as follows: the PCV-2 ORF2 antigen was produced as described in Example 1. The supernatant was then used for BEI inactivation as described in Example 3. After BEI inactivation, PCV-2 ORF2 antigen was mixed with insect cell debris and Carbopol. The final test sample included about 2.06 x 10^6 insect cells, about 32 to 32.5 µg and 1 mg Carbopol 971P per ml test.

Test sample #6 was produced as follows: PCV-2 ORF2 antigen was produced as described in Example 1. The supernatant of Example 1 then was filtered through a 1.2 µm laboratory scale filter. This filter size was determined previously to be sufficient to filter intact and broken insect cells while allowing the PCV-2 ORF2 antigen to pass through the filter. Afterwards, the filtrate was BEI inactivated as described in Example 3. After BEI inactivation, the PCV-2 ORF2 antigen was content was adjusted to an amount of about 32 to 32.5 µg per ml test sample and mixed with 1 mg Carbopol 971P.

Testing of the Immunogenicity of each of the test sample

Clinical Phase:

One-hundred and fifty female Balb/C mice were sourced from Jackson Laboratories (United States) and acclimated for seven days. One mouse from each cage was randomly selected for blood collection on Day 0 for a total of twenty-six samples. A total of ten mice were each inoculated by the subcutaneous route with 0.1 – 0.2mL of Dulbecco's Phosphate Buffer.

A total of twenty mice were each inoculated by the subcutaneous route with 0.1 – 0.2mL of each test sample (test samples #1 to #6). Each cage contained five mice and all mice in each cage were in the same treatment group. On day twenty-one, all mice were

terminally bled. Each blood sample was allowed to clot and the serum was collected by centrifugation. All samples were kept in separate tubes and stored at - 80C±10°C until testing. Mice were disposed of by incineration.

Sample Testing:

The immunogenicity of each test sample was estimated by measuring the PCV-2 specific antibody response of each test sample by using an in-house PCV-2 specific ELISA. The value of immunogenicity of each test sample is given as a Relative Immunogenicity (RI) value in table 2. This Relative Immunogenicity value is a measure for the ORF2 specific antibody titer obtained in an immunized animal per standardized amount of ORF2 antigen used for immunization.

Instead of using the in-house ELISA, the amount of PCV-2 specific antibodies can also be measured by using the ELISA assay described by Nawagitgul, P., et al. in *Modified indirect porcine circovirus (PCV) type 2-based and recombinant capsid protein (ORF2)-based ELISA for the detection of antibodies to PCV* Clin. Diagn. Lab. Immunol. **9**:33-40 (2002), which teaching and content hereby is incorporated by reference. The value measured in such assay can also be used to calculate the Relative Immunogenicity value (*see below*).

An aliquot of serum from each mouse was pooled with cage mates for a total of twenty-six samples for day 21. An aliquot of all day 0 serum samples were pooled into one sample. The reference was diluted two-fold starting at 1:2 and added in triplicate to each corresponding well. The positive and negative controls were added in triplicate. Each sample was serially diluted two-fold and added to the plate starting at 1:200 in triplicate. The final absorbance at 450nm was read using a monthly calibrated SoftMax™ Plate reader and all raw OD values were captured electronically and analyzed with Statlia (Brendan Scientific) to calculate the Relative Immunogenicity (RI) values.

Results:

The calculated RI value for the amount of produced antibodies after immunization showed that the purified PCV-2 ORF2 formulation evoked the highest serological (antibody) response to highly purified PCV-2 ORF2 antigen. The formulation of purified PCV-2 ORF2 together with insect cell debris resulted in a decrease in Relative Immunogenicity (*i.e.* Immunogenicity) of ORF2 compared to highly purified PCV-2 ORF2 alone. Insect cells alone did not generate an antibody response against PCV-2 ORF2 antigen at all. Test samples 4 to 6, which also do not contain highly purified PCV-2 ORF2 antigen showed also a decreased Relative Immunogenicity compared to highly purified PCV-2 ORF2 alone.

What is claimed is:

1. A method of producing a PCV-2 antigenic composition comprising the steps of:

i) obtaining a first liquid containing therein PCV-2 antigen;

ii) removing at least a portion of the first liquid from the PCV-2 antigen by an exchange of the portion of the first liquid against a second liquid, wherein the second liquid is different from the first liquid; and

iii) admixing the PCV-2 antigen remaining after step ii) with a further component selected from the group consisting of pharmaceutically acceptable carriers, adjuvants, diluents, excipients, and combinations thereof.

2. The method according to claim 1, wherein the exchange of the portion of the first liquid with the second liquid comprises the steps of:

a) liquid addition comprising adding the second liquid to the first liquid which contains the PCV-2 antigen; and

b) concentrating the PCV-2 antigen by removing a portion of the first and second liquids.

3. The method according to claim 1 or 2, wherein the portion of the first liquid is removed from the PCV-2 antigen by a filtration step utilizing a filter.

4. The method according to claim 2 or 3, wherein the concentrating step and the liquid addition step are performed simultaneously.

5. The method according to any one of claims 2 to 4, wherein the concentrating step and the liquid addition step are performed at least two times.

6. The method according to any one of claims 3 to 5, wherein the filter includes a semi-permeable membrane.

7. The method according to claim 6, wherein the semi-permeable membrane has an average pore size that is smaller than the PCV-2 antigen to thereby prevent passage of at least 90% of the PCV-2 antigen through the semi-permeable membrane pores and hold the PCV-2 antigen within the filter.

8. The method according to any one of claims 3 to 7, wherein the filter has an average pore size which prevents passage of at least 90% of proteins of 50 kDa to 500 kDa in size.

9. The method according to any one of claims 2 to 8, wherein the concentrating step concentrates the antigen from 3X to 50X in comparison to the volume of the first liquid.

10. The method according to any one of claims 1 to 9, wherein the virucidal activity of the PCV-2 antigenic composition is reduced by at least 10% as compared to the liquid that has not undergone the method.

11. The method according to any one of claims 1 to 10, wherein the PCV-2 antigenic composition produced by steps i) to ii) causes a loss of less than 1 log $TCID_{50}$ per ml of a live virus or less than 1 log CFU per ml of a live bacterium, when the live virus or live bacterium is mixed with the PCV-2 antigenic composition for 2 or more hours.

12. The method according to any one of claims 1 to 11, wherein the PCV-2 antigenic composition produced by steps i) to ii) causes a loss of less than 0.7 log $TCID_{50}$ per ml of a live virus or less than 0.7 log CFU per ml of a live bacterium, when the live virus or live bacterium is mixed with the PCV-2 antigenic composition for 2 or more hours.

13. The method according to any one of claims 1 to 12, wherein the method further comprises the step of harvesting the PCV-2 antigen remaining after step ii).

14. The method according to claim 13, wherein method further comprising the step of purifying the harvest of step ii) comprising the PCV-2 antigen, by chromatographic procedure.

15. The method according to claim 14, wherein the PCV-2 antigen is purified to a purity grade of the PCV-2 antigen of more than 50% (w/w) with reference to the total amount of protein.

16. The method according to any one of claims 1 to 15, wherein the further component is an adjuvant.

17. The method according to claim 16, wherein the adjuvant is Carbomer.

18. The method according to any one claims 1 to 17, wherein the PCV-2 antigen comprises the ORF-2 protein of PCV-2.

19. The method according to any one of claims 1 to 18, wherein the PCV-2 antigen comprises virus like particles of ORF-2 protein.

20. The method according to any one of claims 1 to 19, wherein the method further comprises the step of combining the PCV-2 antigenic composition with at least one additional antigen.

21. The method according to claim 20, wherein the at least one additional antigen includes Porcine Reproductive and Respiratory Syndrome Virus antigen and/or *Mycoplasma hyopneumoniae* antigen.

22. A PCV-2 antigenic composition obtainable by a method according to any one of claims 1 to 21.

23. The PCV-2 antigenic composition according to claim 22, wherein the PCV-2 antigenic composition causes a loss of less than 1 log $TCID_{50}$ per ml of a live virus or less than 1 log CFU per ml live bacterium, when the live virus or live bacterium is mixed with the PCV-2 antigenic composition for at least 2 hours.

24. The PCV-2 antigenic composition according to claim 23, wherein the PCV-2 antigenic composition causes a loss of less than 0.7 log $TCID_{50}$ per ml of a live virus or less than 0.7 log CFU per ml of a live bacterium, when the live virus or live bacterium is mixed with the PCV-2 antigenic composition for at least 2 hours.

25. The PCV-2 antigenic composition according to claim 23 or 24, wherein the live virus is Porcine Reproductive and Respiratory Syndrome (PRRS) virus.

26. An immunogenic composition comprising a PCV-2 antigenic composition obtainable by a method according to any one of claims 1 to 21.

27. An immunogenic composition comprising the PCV-2 antigenic composition according to any one of claims 22 to 24.

28. The immunogenic composition according to claim 27, wherein the immunogenic composition further comprises an attenuated live virus or an attenuated live bacterium from at least one other disease-causing organism in swine.

29. The immunogenic composition according to claim 28, wherein the attenuated live virus is Porcine Reproductive and Respiratory Syndrome (PRRS) virus.

30. The immunogenic composition according to any of claims 27 to 29, wherein the immunogenic composition further comprises a *Mycoplasma hyopneumoniae* bacterin.

31. The immunogenic composition according to any one of claims 27 to 30, wherein the immunogenic composition further comprises an adjuvant.

32. The immunogenic composition according to claim 31, wherein the adjuvant is Carbomer.

33. The immunogenic composition according to claims 26 to 32, wherein the immunogenic composition induces a protective immune response against PCV-2 after the administration of one dose of the immunogenic composition.

34. The immunogenic composition according to claim 26 to 32, wherein administration of the immunogenic composition reduces lymphoid depletion and inflammation by at least 80% in an animal as compared to an animal not receiving the immunogenic composition.

35. The immunogenic composition according to claims 29 to 32, wherein the immunogenic composition induces a protective immune response against PRRS virus after the administration of one dose of the immunogenic composition.

36. The immunogenic composition according to claims 30 to 32, wherein the immunogenic composition induces a protective immune response against *Mycoplasma hyopneumoniae* after administration of one dose of the immunogenic composition.

37. The immunogenic composition according to claims 26 to 32, wherein 2 ml of the immunogenic composition comprises 1 dose of the PCV-2 antigen.

38. The immunogenic composition according to any one of claims 26 to 32 for reducing one or more clinical symptoms of a PCV-2 infection in an animal as compared to an animal not receiving the immunogenic composition.