

EPITOPES OF ANTI-SERINE PROTEASE INHIBITOR KAZAL (SPIK) ANTIBODIES

CROSS-REFERENCE TO RELATED APPLICATIONS

[0001] This application claims priority benefit of the filing date of US Provisional Patent Application Serial Number 62/899,024, filed on September 11, 2019, the disclosure of which application is herein incorporated by reference in its entirety.

GOVERNMENT RIGHTS

[0002] This invention was made with government support under grant number 2R44CA165314-02A1 and FAIN number R44CA165314 awarded by the National Institutes of Health (NIH) under the Small Business Innovation Research (SBIR) program. The government has certain rights in the invention.

FIELD OF THE INVENTION

[0003] The present invention relates to two classes of anti-AS-SPIK antibodies that specifically bind to one of two different conformational epitopes, along with methods of making such antibodies, compositions, including pharmaceutical compositions, comprising such antibodies, and their use to diagnose and/or treat disorders characterized by the expression of AS-SPIK (e.g., liver cancer). Diagnostic methods and kits comprising the anti-AS-SPIK antibodies are also disclosed.

BACKGROUND

[0004] The liver is one of the largest organs in the body. The liver has many functions, including the production of enzymes and bile required for the digestion of food, regulation of glycogen storage, plasma protein synthesis, hormone production, and detoxification of various metabolites. Liver disorders include liver cancers such as Hepatocellular Carcinoma (HCC) and intrahepatic Cholangiocarcinoma (ICC), viral infections, cirrhosis, and other inflammatory disorders of the liver affect millions of people worldwide. For example, over 5 million individuals in the U.S. and over 450 million individuals worldwide suffer from hepatitis B virus (HBV) and hepatitis C virus (HCV) infections, and over 30% of these infected individuals are at a high risk of developing liver cancer. (1-4) Despite advances in diagnosis and treatment, liver cancer remains an important cause of both morbidity and mortality. Primary liver cancer, or cancer that originates in the liver, has a five-year survival rate of less than 10%. However, if liver cancer is detected early and during its most treatable stages, the survival rate increases to almost 40%. Patients with early-stage liver cancer may have few or no symptoms. Current detection methods, such as serological methods, ultrasound, computed tomography (CT) scans, magnetic resonance imaging (MRI), and angiography, can be unreliable

due to low sensitivity and the potential for operator error. Imaging techniques, which are costly, may be less accurate for the detection of smaller, early stage tumors (1, 2). Liver biopsy, which is still considered the most reliable method for distinguishing benign from malignant tumors, is invasive and requires surgery (3). There is a continuing need for new methods of diagnosing and treating liver cancer, especially for those affected by liver cirrhosis, viral infections, and inflammatory disorders of the liver

[0005] Serine protease Inhibitor Kazal (SPIK/SPINK1) is a small secreted protein with 79 amino acids (4). It was first discovered in the pancreas as an inhibitor of the autoactivation of trypsinogen (5, 6). Recent studies have suggested that expression of SPIK is elevated in liver cancer such as hepatocellular carcinoma (HCC) and intrahepatic cholangiocarcinoma (ICC), but has limited or no activity in normal tissues including liver, but outside of pancreas (7-9). The use of SPIK as a cancer biomarker has been impeded by the fact that levels of serum SPIK are also elevated in the presence of other diseases, especially pancreatitis (10-12). We have found that for SPIK secreted by liver cancer cells, an additional, at least 9-residue long fragment in the N-terminus, is retained, while it is removed in SPIK secreted from normal cell such as pancreatic cell (13). We call this kind SPIK as NS-SPIK (Normal Secreted SPIK). However, we found that the SPIK secreted by liver cancer cell is larger than NS-SPIK, an additional, at least 9-residue long fragment of this 23 amino acid, is retained during secretion (11). We named SPIK secreted by liver cancer cell as AS-SPIK or LC-SPIK (Abnormal Secreted SPIK or Liver Cancer secreted SPIK). Herein the AS-SPIK and LC-SPIK have same meaning. Our further study found that in fact an additional 23 amino acids, not only 9 amino acid at the N-terminus of SPIK (SEQ ID NO: 6), is retained in AS-SPIK, while it is removed from the NS-SPIK. The reason why the cancer cells secrete un-cut SPIK is unknown. Our working hypothesis is that because SPIK is a serine protease inhibitor, over-expression of SPIK in cancer cells suppresses the activity of signal peptide peptidase, one kind of serine protease, resulting in un-attenuated, full-length protein being secreted from cancer cells. While we believe we understand certain events that occur during the expression of AS-SPIK, the compositions and methods of the present invention are not limited to those that work by affecting any particular cellular mechanism. Therefore, it should be possible to differentiate between SPIK produced from cancerous liver cells and SPIK generated by other non-cancerous diseases, by using an antibody which can recognize and bind specifically to this domain. For this purpose, we have developed a series of monoclonal antibodies, which solely recognize cancerous SPIK (AS-SPIK, Abnormal Secreted SPIK), but not normal SPIK (NS-SPIK, normal secreted SPIK), with high sensitivity and specificity. Using this antibody, we further developed an ELISA test kit (SERAVUE®), and assessed the performance of AS-SPIK in distinguishing HCC from other liver disease such as HBV and HCV infection, liver cirrhosis, and non-liver disease such as pancreatitis and as well as healthy subject. The clinical study results suggest that the subject antibodies and related methods can be used to selectively and reliably detect

HCC, even in its very early stages, without interference from other liver or non-liver diseases, which will tremendously improve the diagnosis and treatment of HCC.

SUMMARY OF THE INVENTION

[0006] Aspects of the invention include isolated antibodies that specifically bind to a conformational epitope of an AS-SPIK protein, and do not specifically bind to a NS-SPIK protein, wherein the conformational epitope of the AS-SPIK protein comprises: one or more amino acids selected from the group consisting of: L14, L15, S16, L17, D24 and S25 of SEQ ID NO: 2; and one or more amino acids selected from the group consisting of: C58, V59, L60, C61, and F62 of SEQ ID NO: 2.

[0007] In some embodiments, the conformational epitope comprises amino acids L14, L15, S16, and L17 of SEQ ID NO: 2. In some embodiments, the conformational epitope comprises amino acids L60 and C61 of SEQ ID NO: 2. In some embodiments, the conformational epitope comprises amino acids L14, L15, S16, L17, L60, and C61 of SEQ ID NO: 2. In some embodiments, the conformational epitope further comprises amino acids D24 and S25 of SEQ ID NO: 2. In some embodiments, the conformational epitope further comprises amino acids C58, V59 and F62 of SEQ ID NO: 2. In some embodiments, the conformational epitope comprises amino acids L14, L15, S16, L17, D24, S25, C58, V59, L60, C61 and F62 of SEQ ID NO: 2.

[0008] In some embodiments, an isolated antibody comprises: a CDRH1 sequence comprising S6; and/or a CDRH2 sequence comprising I2, G5, G6, Y10 and K16; and/or a CDRH3 sequence comprising G4 and Y7; and/or a CDRL1 sequence comprising Q4 and S9; and/or a CDRL2 sequence comprising A2, S3, T4 and S7; and/or a CDRL3 sequence comprising Q1, Q2, Y4 and S5.

[0009] In some embodiments, an isolated antibody comprises: a CDRH1 sequence comprising S6; a CDRH2 sequence comprising I2, G5, G6, Y10 and K16; a CDRH3 sequence comprising G4 and Y7; a CDRL1 sequence comprising Q4 and S9; a CDRL2 sequence comprising A2, S3, T4 and S7; and a CDRL3 sequence comprising Q1, Q2, Y4 and S5.

[0010] Aspects of the invention include isolated antibodies that specifically bind to a conformational epitope of an AS-SPIK protein, and do not specifically bind to a NS-SPIK protein, wherein the conformational epitope of the AS-SPIK protein comprises: one or more amino acids selected from the group consisting of: L36, N37, I42 and Y43 of SEQ ID NO: 2; and one or more amino acids selected from the group consisting of: R67, Q68, I71 and L72 of SEQ ID NO: 2.

[0011] In some embodiments, the conformational epitope comprises amino acids L36 and N37 of SEQ ID NO: 2. In some embodiments, the conformational epitope comprises amino acids I42 and Y43 of SEQ ID NO: 2. In some embodiments, the conformational epitope comprises amino acids L36, N37, I42 and Y43

of SEQ ID NO: 2. In some embodiments, the conformational epitope comprises amino acids R67, Q68, I71 and L72 of SEQ ID NO: 2. In some embodiments, the conformational epitope comprises amino acids L36, N37, I42, Y43, R67, Q68, I71 and L72 of SEQ ID NO: 2.

[0012] In some embodiments, an antibody comprises: a CDRH1 sequence comprising Y3, S7 and W9; and/or a CDRH2 sequence comprising A1, I2, G4, G6 and Y10; and/or a CDRH3 sequence comprising R1 and D7; and/or a CDRL1 sequence comprising A2, S3, Q4, I6, Y9, L10 and S11; and/or a CDRL2 sequence comprising A2, S3, L5 and S7; and/or a CDRL3 sequence comprising Q1, Q2, and T5.

[0013] In some embodiments, an antibody comprises: a CDRH1 sequence comprising Y3, S7 and W9; a CDRH2 sequence comprising A1, I2, G4, G6 and Y10; a CDRH3 sequence comprising R1 and D7; a CDRL1 sequence comprising A2, S3, Q4, I6, Y9, L10 and S11; a CDRL2 sequence comprising A2, S3, L5 and S7; and a CDRL3 sequence comprising Q1, Q2, and T5.

[0014] In some embodiments, an antibody is multi-specific. In some embodiments, an antibody is bispecific. In some embodiments, an antibody has binding affinity to an effector cell. In some embodiments, an antibody has binding affinity to a T-cell antigen. In some embodiments, the T-cell antigen comprises a CD3 protein. In some embodiments, an antibody is a monoclonal antibody. In some embodiments, an antibody is in a CAR-T format.

[0015] Aspects of the invention include immunoconjugates comprising an antibody as described herein, covalently attached to a cytotoxic agent. In some embodiments, the cytotoxic agent is selected from the group consisting of: a toxin, a chemotherapeutic agent, a drug moiety, an antibiotic, a radioactive isotope and a nucleolytic enzyme.

[0016] In some embodiments, an immunoconjugate has the formula Ab-(L-D)_p, wherein: Ab is an antibody as described herein; L is a linker; D is a drug moiety; and p is an integer that ranges from 1 to 8. In some embodiments, D is selected from the group consisting of: a maytansinoid, an auristatin and dolostatin. In some embodiments, L comprises one or more linkers selected from the group consisting of 6-maleimidocaproyl (MC), maleimidopropanoyl (MP), valine-citrulline (val-cit), alanine-phenylalanine (ala-phe), p-aminobenzyloxycarbonyl (PAB), N-Succinimidyl 4-(2-pyridylthio)pentanoate (SPP), N-succinimidyl 4-(N-maleimidomethyl)cyclohexane-1 carboxylate (SMCC), 4-(2-Pyridyldithio)butyric acid-N-hydroxysuccinimide ester (SPDB), and N-Succinimidyl (4-iodo-acetyl)aminobenzoate (SIAB).

[0017] Aspects of the invention include pharmaceutical compositions comprising an antibody or immunoconjugate as described herein.

[0018] Aspects of the invention include methods for the treatment of a disorder characterized by expression of AS-SPIK, comprising administering to a subject with said disorder an antibody or immunoconjugate as described herein, or a pharmaceutical composition as described herein.

[0019] Aspects of the invention include use of an antibody or immunoconjugate as described herein, in the preparation of a medicament for the treatment of a disorder characterized by expression of AS-SPIK.

[0020] Aspects of the invention include an antibody or immunoconjugate as described herein for use in the treatment of a disorder characterized by expression of AS-SPIK.

[0021] In some embodiments, the disorder is a liver disorder. In some embodiments, the liver disorder is hepatocellular carcinoma. In some embodiments, the liver disorder is intrahepatic cholangiocarcinoma. In some embodiments, the liver disorder is a viral infection. In some embodiments, the liver disorder is an inflammatory liver disorder. In some embodiments, the inflammatory liver disorder is cirrhosis of the liver.

[0022] Aspects of the invention include a polynucleotide encoding an antibody as described herein. Aspects of the invention include a vector comprising a polynucleotide as described herein. Aspects of the invention include a host cell comprising a vector as described herein.

[0023] Aspects of the invention include methods of producing an antibody or immunoconjugate as described herein, comprising growing a host cell as described herein under conditions permissive for expression of the antibody, and isolating the antibody from the cell.

[0024] Aspects of the invention include a diagnostic method for determining whether a subject has or is at risk of developing a disorder characterized by expression of AS-SPIK, the method comprising: contacting a biological test sample from the subject with an AS-SPIK antibody as described herein to generate an AS-SPIK-antibody complex; detecting a concentration of the AS-SPIK-antibody complex in the biological test sample; and comparing the concentration of the AS-SPIK-antibody complex to a reference value to determine whether the subject has or is at risk of developing the disorder.

[0025] Aspects of the invention include a diagnostic method for determining whether a subject has or is at risk of developing a disorder characterized by expression of AS-SPIK, the method comprising: contacting a biological test sample from the subject with a first antibody or antigen-binding fragment that specifically binds to SPIK to form a SPIK-antibody complex; contacting the SPIK-antibody complex with an AS-SPIK antibody or antigen-binding fragment as described herein to generate an AS-SPIK-antibody complex; detecting a concentration of the AS-SPIK-antibody complex in the biological test sample; and comparing the concentration of the AS-SPIK-antibody complex to a reference value to determine whether the subject has or is at risk of developing the disorder.

[0026] In some embodiments, the antibody or antigen-binding fragment comprises a detectable label. In some embodiments, the disorder is a liver disorder. In some embodiments, the liver disorder is selected from the group consisting of: hepatocellular carcinoma, intrahepatic cholangiocarcinoma, viral infection of the liver, inflammatory disorder of the liver, and cirrhosis of the liver.

[0027] Aspects of the invention include kits comprising an antibody or immunoconjugate as described herein. In some embodiments, a kit further comprises an antibody that specifically binds to SPIK.

BRIEF DESCRIPTION OF THE DRAWINGS

- [0028] FIG. 1 shows the size of AS-SPIK and NS-SPIK using gel electrophoresis.
- [0029] FIG. 2 shows the N-terminal sequence of AS-SPIK by Edman N-terminal analysis.
- [0030] FIG. 3 shows a comparison of the amino acid sequences of AS-SPIK and NS-SPIK.
- [0031] FIG. 4 shows a comparison of the 3D structure of AS-SPIK with that of NS-SPIK.
- [0032] FIG. 5 shows the test results of binding activity of antibodies that bind to AS-SPIK and NS-SPIK.
- [0033] FIG. 6 shows the results of binding tests of anti-AS-SPIK antibodies to synthesized peptides.
- [0034] FIG. 7 the shows the 3D structure (crystal model) of AS-SPIK and its epitopes.
- [0035] FIG. 8 shows the result of inhibition tests for locating the binding site of Class I anti-AS-SPIK antibodies.
- [0036] FIG. 9 shows the consensus amino acids of CDRs in Class I anti-AS-SPIK antibodies.
- [0037] FIG. 10 shows the consensus amino acids of CDRs in Class II anti-AS-SPIK antibodies.
- [0038] FIG. 11 provides an illustration of the mechanism for the AS-SPIK detection kit.
- [0039] FIG. 12, panels A, is a graph showing that IM-CA22 binds specifically to AS-SPIK but not NS-SPIK, while antibody IM-BA2 binds to both AS-SPIK and NS-SPIK. Panel B is a graph showing that AS-SPIK is elevated in HCC, but not in pancreatitis or healthy patients. Panel C is a graph showing that NS-SPIK is elevated in pancreatitis patients but does not interfere with the performance of the AS-SPIK-based detection kit.
- [0040] FIG. 13 is a table (Table 1) that shows the binding characteristics of Class I and Class II anti-AS-SPIK antibodies.
- [0041] FIG. 14 is a table (Table 2) that shows the regions for anti-AS-SPIK binding predicted by a CLIPS analysis study, and the amino acid residues that constitute the epitopes therein.
- [0042] FIG. 15 is a table (Table 3) that shows the amino acid sequences of four example Class I antibodies that bind to Epitope I, as described further herein.
- [0043] FIG. 16 is a table (Table 4) that shows the amino acid sequences of four example Class II antibodies that bind to Epitope II, as described further herein.
- [0044] FIG. 17 is a table (Table 5) that shows serum AS-SPIK levels in a clinical study with 512 samples.
- [0045] FIG. 18 is a table (Table 6) that provides a performance summary of AS-SPIK v. AFP in the detection of HCC.
- [0046] FIG. 19 is a table (Table 7) that shows AS-SPIK levels in early v. late stage HCC.

[0047] FIG. 20 is a table (Table 8) that shows AS-SPIK levels in HCC, by BCLC stage.

DETAILED DESCRIPTION

[0048] The practice of the present invention will employ, unless otherwise indicated, conventional techniques of molecular biology (including recombinant techniques), microbiology, cell biology, biochemistry, and immunology, which are within the skill of the art. Such techniques are explained fully in the literature, such as, “Molecular Cloning: A Laboratory Manual”, second edition (Sambrook et al., 1989); “Oligonucleotide Synthesis” (M. J. Gait, ed., 1984); “Animal Cell Culture” (R. I. Freshney, ed., 1987); “Methods in Enzymology” (Academic Press, Inc.); “Current Protocols in Molecular Biology” (F. M. Ausubel et al., eds., 1987, and periodic updates); “PCR: The Polymerase Chain Reaction”, (Mullis et al., ed., 1994); “A Practical Guide to Molecular Cloning” (Perbal Bernard V., 1988); “Phage Display: A Laboratory Manual” (Barbas et al., 2001); Harlow, Lane and Harlow, Using Antibodies: A Laboratory Manual: Portable Protocol No. I, Cold Spring Harbor Laboratory (1998); and Harlow and Lane, Antibodies: A Laboratory Manual, Cold Spring Harbor Laboratory; (1988).

[0049] Where a range of values is provided, it is understood that each intervening value, to the tenth of the unit of the lower limit unless the context clearly dictates otherwise, between the upper and lower limit of that range and any other stated or intervening value in that stated range is encompassed within the invention. The upper and lower limits of these smaller ranges may independently be included in the smaller ranges is also encompassed within the invention, subject to any specifically excluded limit in the stated range. Where the stated range includes one or both of the limits, ranges excluding either or both of those included limits are also included in the invention.

[0050] Unless indicated otherwise, antibody residues herein are numbered according to the Kabat numbering system (e.g., Kabat et al., Sequences of Immunological Interest. 5th Ed. Public Health Service, National Institutes of Health, Bethesda, Md. (1991)).

[0051] In the following description, numerous specific details are set forth to provide a more thorough understanding of the present invention. However, it will be apparent to one of skill in the art that the present invention may be practiced without one or more of these specific details. In other instances, well-known features and procedures well known to those skilled in the art have not been described in order to avoid obscuring the invention.

[0052] All references cited throughout the disclosure, including patent applications and publications, are incorporated by reference herein in their entirety.

Definitions

[0053] For purposes of interpreting this specification, the following definitions will apply and whenever appropriate, terms used in the singular will also include the plural and vice versa. In the event that any definition set forth conflicts with any document incorporated herein by reference, the definition set forth below shall control.

[0054] An “epitope” is the site on the surface of an antigen molecule to which a single antibody molecule binds. Generally, an antigen has several or many different epitopes and reacts with many different antibodies. The term specifically includes linear epitopes and conformational epitopes. The term includes any molecular determinant capable of specific binding to an antibody. In certain embodiments, an epitope determinant includes chemically active surface groupings of molecules such as amino acids, sugar side chains, phosphoryl, or sulfonyl, and, in certain embodiments, may have specific three dimensional structural characteristics, and/or specific charge characteristics. An epitope is a region of an antigen that is bound by an antibody. A “binding region” is a region on a binding target bound by a binding molecule.

[0055] “Epitope mapping” is the process of identifying the binding sites, or epitopes, of antibodies on their target antigens. Antibody epitopes may be linear epitopes or conformational epitopes. Linear epitopes are formed by a continuous sequence of amino acids in a protein. Conformational epitopes are formed of amino acids that are discontinuous in the protein sequence, but which are brought together upon folding of the protein into its three-dimensional structure.

[0056] “Epitope binning”, as defined herein, is the process of grouping antibodies based on the epitopes they recognize. More particularly, epitope binning comprises methods and systems for discriminating the epitope recognition properties of different antibodies, combined with computational processes for clustering antibodies based on their epitope recognition properties and identifying antibodies having distinct binding specificities.

[0057] An antibody binds “essentially the same epitope” as a reference antibody when the two antibodies recognize identical or sterically overlapping epitopes. The most widely used and rapid methods for determining whether two epitopes bind to identical or sterically overlapping epitopes are competition assays, which can be configured in any number of different formats, using either labeled antigen or labeled antibody. Usually, the antigen is immobilized on a 96-well plate, and the ability of unlabeled antibodies to block the binding of labeled antibodies is measured using radioactive or enzyme labels.

[0058] A “modification” of an amino acid residue/position, as used herein, refers to a change of a primary amino acid sequence as compared to a starting amino acid sequence, wherein the change results from a sequence alteration involving said amino acid residue/positions. For example, typical modifications include substitution of the residue (or at said position) with another amino acid (e.g., a conservative or non-

conservative substitution), insertion of one or more (generally fewer than 5 or 3) amino acids adjacent to said residue/position, and deletion of said residue/position. An “amino acid substitution” or variation thereof, refers to the replacement of an existing amino acid residue in a predetermined (starting) amino acid sequence with a different amino acid residue. Generally and preferably, a modification results in an alteration in at least one physical or biochemical activity of the variant polypeptide compared to a polypeptide comprising the starting (or “wild type”) amino acid sequence. For example, in the case of an antibody, a physical or biochemical activity that is altered can be binding affinity, binding capability and/or binding effect upon a target molecule.

[0059] The term “antibody” includes monoclonal antibodies (including full length antibodies which have an immunoglobulin Fc region), single-chain molecules, as well as antibody fragments (e.g., Fab, F(ab')₂, and Fv). The term “immunoglobulin” (Ig) is used interchangeably with “antibody” herein. The basic 4-chain antibody unit is a heterotetrameric glycoprotein composed of two identical light (L) chains and two identical heavy (H) chains. Unless noted otherwise, the term “antibody” is used herein in the broadest sense and specifically includes all isotypes, sub-classes and forms of antibodies, including IgG, IgM, IgA, IgD, and IgE antibodies and their fragments, preferably antigen-binding fragments.

[0060] Unless stated otherwise, the term “antibody” specifically includes native human and non-human IgG1, IgG2 (IgG2a, IgG2b), IgG3, IgG4, IgE, IgA, IgD and IgM antibodies, including naturally occurring variants.

[0061] The term “monoclonal antibody” as used herein refers to an antibody obtained from a population of substantially homogeneous antibodies, i.e., the individual antibodies comprising the population are identical except for possible naturally occurring mutations that may be present in minor amounts. Monoclonal antibodies are highly specific, being directed against a single antigenic site. Furthermore, in contrast to conventional (polyclonal) antibody preparations which typically include different antibodies directed against different determinants (epitopes), each monoclonal antibody is directed against a single determinant on the antigen. The modifier “monoclonal” indicates the character of the antibody as being obtained from a substantially homogeneous population of antibodies, and is not to be construed as requiring production of the antibody by any particular method. For example, the monoclonal antibodies to be used in accordance with the present invention may be made by the hybridoma method first described by Kohler et al. (1975) Nature 256:495, or may be made by recombinant DNA methods (see, e.g., U.S. Patent No. 4,816,567). The “monoclonal antibodies” may also be isolated from phage antibody libraries using the techniques described in Clackson et al. (1991) Nature 352:624-628 and Marks et al. (1991) J. Mol. Biol. 222:581-597, for example.

[0062] The monoclonal antibodies herein specifically include “chimeric” antibodies (immunoglobulins) in which a portion of the heavy and/or light chain is identical with or homologous to corresponding sequences in antibodies derived from a particular species, while the remainder of the chain(s) is identical with or homologous to corresponding sequences in antibodies derived from another species, as well as fragments of such antibodies, so long as they exhibit the desired biological activity (U.S. Patent No. 4,816,567; and Morrison et al. (1984) *Proc. Natl. Acad. Sci. USA* 81:6851-6855).

[0063] “Humanized” forms of non-human (e.g., murine) antibodies are antibodies which contain minimal sequence derived from non-human immunoglobulin. For the most part, humanized antibodies are human immunoglobulins (recipient antibody) in which residues from a hypervariable region of the recipient are replaced by residues from a hypervariable region of a non-human species (donor antibody) such as mouse, rat, rabbit or nonhuman primate having the desired specificity, affinity, and capacity. In some instances, Fv framework region (FR) residues of the human immunoglobulin are also replaced by corresponding non-human residues. Furthermore, humanized antibodies may comprise residues which are not found in the recipient antibody or in the donor antibody. These modifications are made to further refine antibody performance. In general, the humanized antibody will comprise substantially all of at least one, and typically two, variable domains, in which all or substantially all of the hypervariable loops correspond to those of a non-human immunoglobulin and all or substantially all of the FR regions are those of a human immunoglobulin sequence. The humanized antibody optionally also will comprise at least a portion of an immunoglobulin constant region (Fc), typically that of a human immunoglobulin. For further details, see Jones et al. (1986) *Nature* 321:522-525; Riechmann et al. (1988) *Nature* 332:323-329; and Presta (1992) *Curr. Op. Struct. Biol.* 2:593-596.

[0064] “Percent (%) amino acid sequence identity” with respect to a reference polypeptide sequence is defined as the percentage of amino acid residues in a candidate sequence that are identical with the amino acid residues in the reference polypeptide sequence, after aligning the sequences and introducing gaps, if necessary, to achieve the maximum percent sequence identity, and not considering any conservative substitutions as part of the sequence identity. Alignment for purposes of determining percent amino acid sequence identity can be achieved in various ways that are within the skill in the art, for instance, using publicly available computer software such as BLAST, BLAST-2, ALIGN or Megalign (DNASTAR) software. Those skilled in the art can determine appropriate parameters for aligning sequences, including any algorithms needed to achieve maximal alignment over the full length of the sequences being compared. For purposes herein, however, % amino acid sequence identity values are generated using the sequence comparison computer program ALIGN-2.

[0065] As used herein, the term “percent sequence homology” refers to the degree of homology between any given query sequence and a subject sequence. For example, a naturally occurring AS-SPIK polypeptide or NS-SPIK polypeptide can be the query sequence and a fragment of an AS-SPIK polypeptide or an NS-SPIK polypeptide can be the subject sequence. Similarly, a fragment of an AS-SPIK polypeptide or an NS-SPIK polypeptide can be the query sequence and a biologically active variant thereof can be the subject sequence.

[0066] The term “consensus sequence” as used herein means a sequence of amino acid or nucleotide residues that represent the most frequent residues found at each position in a sequence alignment, after aligning the sequences and introducing gaps, if necessary, to achieve the maximum sequence match, and not considering any conservative substitutions as part of the sequence identity.

[0067] An “isolated” antibody herein is one which has been identified and separated and/or recovered from a component of its natural environment in a recombinant host cell. Contaminant components of its natural environment are materials which would interfere with diagnostic or therapeutic uses for the antibody, and may include enzymes, hormones, and other proteinaceous or non-proteinaceous solutes, as well as undesired byproducts of the production. In a preferred embodiment, an isolated antibody herein will be purified (1) to greater than 95% by weight, or greater than 98% by weight, or greater than 99% by weight, as determined by SDS-PAGE or SEC- HPLC methods, (2) to a degree sufficient to obtain at least 15 residues of N-terminal or internal amino acid sequence by use of an amino acid sequencer, or (3) to homogeneity by SDS-PAGE under reducing or non-reducing conditions using Coomassie blue or, preferably, silver stain. Ordinarily, an isolated antibody will be prepared by at least one purification step.

[0068] In the case of IgGs, the 4-chain unit is generally about 150,000 Daltons. Each L chain is linked to an H chain by one covalent disulfide bond, while the two H chains are linked to each other by one or more disulfide bonds depending on the H chain isotype. Each H and L chain also has regularly spaced intra-chain disulfide bridges. Each H chain has at the N-terminus, a variable domain (VH) followed by three constant domains (CH) for each of the α and γ chains and four CH domains for μ and ϵ isotypes. Each L chain has at the N-terminus, a variable domain (VL) followed by a constant domain at its other end. The VL is aligned with the VH and the CL is aligned with the first constant domain of the heavy chain (CH1). Particular amino acid residues are believed to form an interface between the light chain and heavy chain variable domains. The pairing of a VH and VL together forms a single antigen-binding site.

[0069] The term “polypeptide” is used herein in the broadest sense and includes peptide sequences. The term “peptide” generally describes linear molecular chains of amino acids containing up to about 60, preferably up to about 30 amino acids covalently linked by peptide bonds.

[0070] The term “specific binding” or “specifically binds to” or is “specific for” refers to the binding of an antibody to a target antigen, e.g., an epitope on a particular polypeptide, peptide, or other target (e.g., a glycoprotein target), and means binding that is measurably different from a non-specific interaction (e.g., a non-specific interaction may be binding to bovine serum albumin or casein). Specific binding can be measured, for example, by determining binding of an antibody to a target molecule compared to binding to a control molecule. For example, specific binding can be determined by competition with a control molecule that is similar to the target, for example, an excess of non-labeled target. In this case, specific binding is indicated if the binding of the labeled target to a probe is competitively inhibited by excess unlabeled target. The term “specific binding” or “specifically binds to” or is “specific for” a particular polypeptide or an epitope on a particular polypeptide target as used herein can be exhibited, for example, by a molecule having a K_d for the target of at least about 200 nM, alternatively at least about 150 nM, alternatively at least about 100nM, alternatively at least about 60 nM, alternatively at least about 50 nM, alternatively at least about 40 nM, alternatively at least about 30 nM, alternatively at least about 20 nM, alternatively at least about 10 nM, alternatively at least about 8 nM, alternatively at least about 6 nM, alternatively at least about 4 nM, alternatively at least about 2 nM, alternatively at least about 1 nM, or greater. In certain instances, the term “specific binding” refers to binding where a molecule binds to a particular polypeptide or epitope on a particular polypeptide without substantially binding to any other polypeptide or polypeptide epitope.

[0071] “Binding affinity” refers to the strength of the sum total of noncovalent interactions between a single binding site of a molecule (e.g., an antibody) and its binding partner (e.g., an antigen). Unless indicated otherwise, as used herein, “binding affinity” refers to intrinsic binding affinity which reflects a 1:1 interaction between members of a binding pair (e.g., antibody and antigen). The affinity of a molecule X for its partner Y can generally be represented by the dissociation constant (K_d). For example, the K_d can be about 200 nM, 150 nM, 100 nM, 60 nM, 50 nM, 40 nM, 30 nM, 20 nM, 10 nM, 8 nM, 6 nM, 4 nM, 2 nM, 1 nM, or stronger. Affinity can be measured by common methods known in the art, including those described herein. Low-affinity antibodies generally bind antigen slowly and tend to dissociate readily, whereas high-affinity antibodies generally bind antigen faster and tend to remain bound longer. A variety of methods of measuring binding affinity are known in the art.

[0072] As used herein, the “ K_d ” or “ K_d value” refers to a dissociation constant measured by a technique appropriate for the antibody and target pair, for example using surface plasmon resonance assays, for example, using a BIAcore™-2000 or a BIAcore™-3000 (BIAcore, Inc., Piscataway, N.J.) at 25°C with immobilized antigen CM5 chips at about 10 response units (RU).

[0073] The term “valent” as used herein denotes the presence of a specified number of binding sites in an antibody. As such, the term “bivalent” denotes the presence of two binding sites.

[0074] “Polyepitopic specificity” refers to the ability to specifically bind to two or more different epitopes on the same or different target(s). “Monospecific” refers to the ability to bind only one epitope. In some embodiments, an antibody binds to each epitope with an affinity of at least 10^{-7} M, or 10^{-8} M or better.

[0075] The term “target” or “binding target” is used in the broadest sense and specifically includes, without limitation, polypeptides, nucleic acids, carbohydrates, lipids, cells, and other molecules with or without biological function as they exist in nature.

[0076] The term “antigen” refers to an entity or fragment thereof, which can bind to an antibody or trigger a cellular immune response. An immunogen refers to an antigen, which can elicit an immune response in an organism, particularly an animal, more particularly a mammal including a human. The term antigen includes regions known as antigenic determinants or epitopes, as defined above.

[0077] As used herein, the term “immunogenic” refers to substances that elicit the production of antibodies, and/or activate T-cells and/or other reactive immune cells directed against an antigen of the immunogen.

[0078] An “antigen-binding site” or “antigen-binding region” of an antibody of the present invention typically contains six hypervariable regions (HVRs) which contribute in varying degrees to the affinity of the binding site for antigen. The term “complementarity determining region” or “CDR” is used interchangeably herein with the term “hypervariable region” or “HVR”. There are three heavy chain variable domain HVRs (HVR-H1, HVR-H2 and HVR-H3) and three light chain variable domain HVRs (HVR-L1, HVR-L2 and HVR-L3). The extent of HVR and framework regions (FRs) is determined by comparison to a compiled database of amino acid sequences in which those regions have been defined according to variability among the sequences and/or structural information from antibody/antigen complexes. Also included within the scope of the invention are functional antigen binding sites comprised of fewer HVRs (i.e., where binding specificity is determined by three, four or five HVRs). Less than a complete set of 6 HVRs may be sufficient for binding to some binding targets. Thus, in some instances, the HVRs of a VH or a VL domain alone will be sufficient. Furthermore, certain antibodies might have non-HVR-associated binding sites for an antigen. Such binding sites are specifically included within the present definition.

[0079] A “naked antibody” for the purposes herein is an antibody that is not conjugated to a cytotoxic moiety or radiolabel.

[0080] An “antibody-drug conjugate” (ADC) or immunoconjugate means an antibody, or antigen-binding fragment thereof, conjugated to a cytotoxic agent, such as a chemotherapeutic agent, a drug, a growth inhibitory agent, a toxin (e.g., an enzymatically active toxin of bacterial, fungal, plant, or animal origin, or fragments thereof), or a radioactive isotope (i.e., a radioconjugate).

[0081] The term “host cell” as used herein denotes any kind of cellular system which can be engineered to generate the antibodies according to the current invention. In one embodiment, Chinese hamster ovary (CHO) cells are used as host cells.

[0082] As used herein, the expressions “cell,” “cell line,” and “cell culture” are used interchangeably and all such designations include progeny. Thus, the words “transformants” and “transformed cells” include the primary subject cell and cultures derived therefrom without regard for the number of transfers. It is also understood that all progeny may not be precisely identical in DNA content, due to deliberate or inadvertent mutations. Variant progeny that have the same function or biological activity as screened for in the originally transformed cell are included.

[0083] A nucleic acid is “operably linked” when it is placed in a functional relationship with another nucleic acid sequence. For example, DNA for a pre-sequence or secretory leader is operably linked to DNA for a polypeptide if it is expressed as a pre-protein that participates in the secretion of the polypeptide; a promoter or enhancer is operably linked to a coding sequence if it affects the transcription of the sequence; or a ribosome binding site is operably linked to a coding sequence if it is positioned so as to facilitate translation. Generally, “operably linked” means that the DNA sequences being linked are contiguous, and, in the case of a secretory leader, contiguous and in reading frame. However, enhancers do not have to be contiguous. Linking is accomplished by ligation at convenient restriction sites. If such sites do not exist, the synthetic oligonucleotide adaptors or linkers are used in accordance with conventional practice.

[0084] The terms “anti-AS-SPIK antibody”, “AS-SPIK antibody”, or “an antibody that binds to AS-SPIK” all refer to an antibody that is capable of binding AS-SPIK with sufficient affinity such that the antibody is useful as a diagnostic and/or therapeutic agent in targeting AS-SPIK.

[0085] The term “variable” refers to the fact that certain segments of the variable domains differ extensively in sequence among antibodies. The “variable” or “V” domain mediates antigen binding and defines specificity of a particular antibody for its particular antigen. However, the variability is not evenly distributed across the 110-amino acid span of the variable domains. Instead, the V regions consist of relatively invariant stretches called framework regions (FRs) of 15-30 amino acids separated by shorter regions of extreme variability called “hypervariable regions” that are each 9-12 amino acids long. The variable domains of native heavy and light chains each comprise four FRs, largely adopting a β -sheet configuration, connected by three hypervariable regions, which form loops connecting, and in some cases forming part of, the β -sheet structure. The hypervariable regions in each chain are held together in close proximity by the FRs and, with the hypervariable regions from the other chain, contribute to the formation of the antigen-binding site of antibodies (see Kabat et al., *Sequences of Proteins of Immunological Interest*, 5th Ed. Public Health Service, National Institutes of Health, Bethesda, MD. (1991)).

[0086] An “intact” antibody is one which comprises an antigen-binding site as well as a light chain constant domain (CL) and at least heavy chain constant domains of the particular antibody class. For example, an intact IgG antibody comprises an antigen-binding site, a light chain constant domain CL, and at least heavy chain constant domains CH1 (C γ 1), CH2 (C γ 2) and CH3 (C γ 3). An intact IgM antibody comprises an antigen-binding site, a light chain constant domain CL, and at least heavy chain constant domains CM1 (C μ 1), CM2 (C μ 2), CM3 (C μ 3) and CM4 (C μ 4). An intact IgA antibody comprises an antigen-binding site, a light chain constant domain CL, and at least heavy chain constant domains CA1 (C α 1), CA2 (C α 2) and CA3 (C α 3). An intact IgD antibody comprises an antigen-binding site, a light chain constant domain CL, and at least heavy chain constant domains CD1 (C δ 1), CD2 (C δ 2) and CD3 (C δ 3). An intact IgE antibody comprises an antigen-binding site, a light chain constant domain CL, and at least heavy chain constant domains CE1 (C ϵ 1), CE2 (C ϵ 2), CE3 (C ϵ 3) and CE4 (C ϵ 4). The constant domains can be native sequence constant domains (e.g., human native sequence constant domains) or amino acid sequence variants thereof. Preferably, an intact antibody has one or more effector functions.

[0087] “Antibody fragments” or “antigen-binding fragments” of antibodies comprise a portion of an intact antibody, preferably the antigen binding or variable region, of the intact antibody. Non-limiting examples of antibody fragments include Fab, Fab', F(ab')₂, and Fv fragments; diabodies; linear antibodies (see U.S. Patent No. 5,641,870, Example 2; Zapata et al., Protein Eng. 8(10): 1057-1062 (1995)); single-chain antibody molecules; and multi-specific antibodies formed from antibody fragments. In one embodiment, an antibody fragment comprises an antigen binding site of an intact antibody and thus retains the ability to bind antigen. Those of ordinary skill in the art will understand that an antibody fragment can be generated from any intact antibody, e.g., from an IgG, IgM, IgA, IgD, or IgE antibody, by separating at least an antigen-binding portion of the antibody from the remainder of its light and heavy chains to create an antigen-binding fragment. In certain embodiments, an antibody fragment can comprise an antigen-binding region of an antibody, as well as one or more additional domains of a light and/or heavy chain of the antibody. For example, in some embodiments, an antibody fragment can comprise an antigen-binding region comprising a VH and a VL domain, a light chain constant domain CL, and one or more heavy chain constant domains, e.g., a CH1 (C γ 1) domain, a CM1 (C μ 1) domain, a CA1 (C α 1) domain, a CD1 (C δ 1) domain, or a CE1 (C ϵ 1) domain.

[0088] In the case of IgG antibody fragments, papain digestion produces two identical antigen-binding fragments, called “Fab” fragments, and a residual “Fc” fragment, a designation reflecting the ability to crystallize readily. The Fab fragment consists of an entire L chain along with the variable region domain of the H chain (VH), and the first constant domain of one heavy chain (CH1). Each Fab fragment is monovalent with respect to antigen binding, i.e., it has a single antigen-binding site. Pepsin treatment of an

IgG antibody yields a single large F(ab')₂ fragment which roughly corresponds to two disulfide linked Fab fragments having divalent antigen-binding activity and is still capable of cross-linking antigen. Fab' fragments differ from Fab fragments by having additional few residues at the carboxy terminus of the CH1 domain including one or more cysteines from the antibody hinge region. Fab'-SH is the designation herein for Fab' in which the cysteine residue(s) of the constant domains bear a free thiol group. F(ab')₂ antibody fragments originally were produced as pairs of Fab' fragments which have hinge cysteines between them. Other chemical couplings of antibody fragments are also known.

[0089] The Fc fragment of an IgG antibody comprises the carboxy-terminal portions of both H chains held together by disulfides. The effector functions of antibodies are determined by sequences in the Fc region, which region is also the part recognized by Fc receptors (FcR) found on certain types of cells.

[0090] A "native-sequence Fc region" comprises an amino acid sequence identical to the amino acid sequence of an Fc region found in nature. Native-sequence human Fc regions include, for example, a native-sequence human IgG1 Fc region (non-A and A allotypes); native-sequence human IgG2 Fc region; native-sequence human IgG3 Fc region; and native-sequence human IgG4 Fc region, as well as naturally occurring variants thereof.

[0091] A "variant Fc region" comprises an amino acid sequence that differs from that of a native-sequence Fc region by virtue of at least one amino acid modification, preferably one or more amino acid substitution(s). Preferably, a variant Fc region has at least one amino acid substitution compared to a native-sequence Fc region or to the Fc region of a parent polypeptide, e.g., from about one to about ten amino acid substitutions, and preferably from about one to about five amino acid substitutions in a native-sequence Fc region or in the Fc region of the parent polypeptide. A variant Fc region herein will preferably possess at least about 80% homology with a native-sequence Fc region and/or with an Fc region of a parent polypeptide, and most preferably at least about 90% homology therewith, more preferably at least about 95% homology therewith.

[0092] The human IgG1 amino acid sequence is provided by UniProtKB No. P01857, which is incorporated by reference herein in its entirety. The human IgG2 amino acid sequence is provided by UniProtKB No. P01859, which is incorporated by reference herein in its entirety. The human IgG3 amino acid sequence is provided by UniProtKB No. P01860, which is incorporated by reference herein in its entirety. The human IgG4 amino acid sequence is provided by UniProtKB No. P01861, which is incorporated by reference herein in its entirety.

[0093] "Fv" is the minimum antibody fragment which contains a complete antigen-recognition and binding site. This fragment consists of a dimer of one heavy- and one light-chain variable region domain in tight, non-covalent association. In a single-chain Fv (scFv) species, one heavy- and one light-chain variable

domain can be covalently linked by a flexible peptide linker such that the light and heavy chains can associate in a “dimeric” structure analogous to that in a two-chain Fv species. From the folding of these two domains emanate six hypervariable loops (3 loops each from the H and L chain) that contribute the amino acid residues for antigen binding and confer antigen binding specificity to the antibody. However, even a single variable domain (or half of an Fv comprising only three HVRs specific for an antigen) has the ability to recognize and bind antigen, although at a lower affinity than the entire binding site.

[0094] “Single-chain Fv” also abbreviated as “sFv” or “scFv” are antibody fragments that comprise the VH and VL antibody domains connected into a single polypeptide chain. Preferably, the sFv polypeptide further comprises a polypeptide linker between the VH and VL domains which enables the sFv to form the desired structure for antigen binding. For a review of sFv, see Pluckthun in *The Pharmacology of Monoclonal Antibodies*, vol. 113, Rosenberg and Moore eds., Springer-Verlag, New York, pp. 269-315 (1994); Borrebaeck 1995, *infra*.

[0095] The term “chimeric antigen receptor” or “CAR” is used herein in the broadest sense to refer to an engineered receptor, which grafts a desired binding specificity (e.g., the antigen-binding region of a monoclonal antibody or other ligand) to membrane-spanning and intracellular-signaling domains. Typically, the receptor is used to graft the specificity of a monoclonal antibody onto a T cell to create a chimeric antigen receptor (CAR). (Dai et al., *J Natl Cancer Inst*, 2016; 108(7):djv439; and Jackson et al., *Nature Reviews Clinical Oncology*, 2016; 13:370–383.).

[0096] As used herein, the term “effector cell” refers to an immune cell which is involved in the effector phase of an immune response, as opposed to the cognitive and activation phases of an immune response. Some effector cells express specific Fc receptors and carry out specific immune functions. In some embodiments, an effector cell such as a natural killer cell is capable of inducing antibody-dependent cellular cytotoxicity (ADCC). For example, monocytes and macrophages, which express FcR, are involved in specific killing of target cells and presenting antigens to other components of the immune system, or binding to cells that present antigens. In some embodiments, an effector cell may phagocytose a target antigen or target cell.

[0097] “Human effector cells” are leukocytes which express receptors such as T cell receptors or FcRs and perform effector functions. Preferably, the cells express at least FcγRIII and perform ADCC effector function. Examples of human leukocytes which mediate ADCC include natural killer (NK) cells, monocytes, cytotoxic T cells and neutrophils; with NK cells being preferred. The effector cells may be isolated from a native source thereof, e.g., from blood or PBMCs as described herein.

[0098] The term “immune cell” is used herein in the broadest sense, including, without limitation, cells of myeloid or lymphoid origin, for instance lymphocytes (such as B cells and T cells including cytolytic T

cells (CTLs)), killer cells, natural killer (NK) cells, macrophages, monocytes, eosinophils, polymorphonuclear cells, such as neutrophils, granulocytes, mast cells, and basophils.

[0099] Antibody “effector functions” refer to those biological activities attributable to the Fc region (a native sequence Fc region or amino acid sequence variant Fc region) of an antibody. Examples of antibody effector functions include C1q binding; complement dependent cytotoxicity; Fc receptor binding; antibody-dependent cell-mediated cytotoxicity (ADCC); phagocytosis; down regulation of cell surface receptors (e.g., B cell receptor; BCR), etc.

[0100] “Antibody-dependent cell-mediated cytotoxicity” and “ADCC” refer to a cell-mediated reaction in which nonspecific cytotoxic cells that express Fc receptors (FcRs) (e.g., Natural Killer (NK) cells, neutrophils, and macrophages) recognize bound antibody on a target cell and subsequently cause lysis of the target cell. The primary cells for mediating ADCC, NK cells, express Fc γ RIII only, whereas monocytes express Fc γ RI, Fc γ RII and Fc γ RIII. FcR expression on hematopoietic cells is summarized in Table 3 on page 464 of Ravetch and Kinet, *Annu. Rev. Immunol* 9:457-92 (1991). To assess ADCC activity of a molecule of interest, an in vitro ADCC assay, such as that described in US Patent No. 5,500,362 or 5,821,337 may be performed. Useful effector cells for such assays include peripheral blood mononuclear cells (PBMC) and Natural Killer (NK) cells. Alternatively, or additionally, ADCC activity of the molecule of interest may be assessed in vivo, e.g., in an animal model such as that disclosed in Clynes et al. *PNAS (USA)* 95:652-656 (1998).

[0101] “Complement dependent cytotoxicity” or “CDC” refers to the ability of a molecule to lyse a target in the presence of complement. The complement activation pathway is initiated by the binding of the first component of the complement system (C1q) to a molecule (e.g., an antibody) complexed with a cognate antigen. To assess complement activation, a CDC assay, e.g., as described in Gazzano-Santoro et al., *J. Immunol. Methods* 202:163 (1996), may be performed.

[0102] A “blocking” antibody or an “antagonist” or “antagonistic” antibody is one which inhibits or reduces a biological activity of an antigen to which it binds. Preferred blocking antibodies or antagonist antibodies are capable of substantially or completely inhibiting a biological activity of an antigen.

[0103] An antibody “which binds” an antigen of interest, e.g., an AS-SPIK or NS-SPIK polypeptide, is one that binds the antigen with sufficient affinity such that the antibody is useful as a therapeutic agent in targeting a cell or tissue expressing the antigen, and does not significantly cross-react with other proteins. With regard to the binding of an antibody to a target molecule, the term “specific binding” or “specifically binds to” or is “specific for” a particular polypeptide or an epitope on a particular polypeptide target means binding that is measurably different from a non-specific interaction. Specific binding can be measured, for example, by determining binding of a molecule compared to binding of a control molecule, which generally

is a molecule of similar structure that does not have binding activity. For example, specific binding can be determined by competition with a control molecule that is similar to the target, for example, an excess of non-labeled target. In this case, specific binding is indicated if the binding of the labeled target to a probe is competitively inhibited by excess unlabeled target. In one embodiment, the term “specific binding” refers to binding where a molecule binds to a particular polypeptide or epitope on a particular polypeptide without substantially binding to any other polypeptide or polypeptide epitope.

[0104] The terms “cancer” and “cancerous” refer to or describe the physiological condition in mammals that is typically characterized by unregulated cell growth. A “tumor” comprises one or more cancerous cells. Examples of cancer include, but are not limited to, carcinoma, lymphoma, blastoma, sarcoma, and leukemia or lymphoid malignancies. More particular examples of such cancers include squamous cell cancer (e.g., epithelial squamous cell cancer), skin cancer, melanoma, lung cancer, including small-cell lung cancer, non-small cell lung cancer (“NSCLC”), adenocarcinoma of the lung and squamous carcinoma of the lung, cancer of the peritoneum, gastric or stomach cancer including gastrointestinal cancer, pancreatic cancer (e.g., pancreatic ductal adenocarcinoma), glioblastoma, cervical cancer, ovarian cancer (e.g., high grade serous ovarian carcinoma), liver cancer (e.g., hepatocellular carcinoma (HCC), intrahepatic cholangiocarcinoma (ICC)), bladder cancer (e.g., urothelial bladder cancer), testicular (germ cell tumor) cancer, hepatoma, breast cancer, brain cancer (e.g., astrocytoma), colon cancer, rectal cancer, colorectal cancer, endometrial or uterine carcinoma, salivary gland carcinoma, kidney or renal cancer (e.g., renal cell carcinoma, nephroblastoma or Wilms’ tumour), prostate cancer, vulval cancer, thyroid cancer, hepatic carcinoma, anal carcinoma, penile carcinoma, as well as head and neck cancer. Additional examples of cancer include, without limitation, retinoblastoma, thecomas, arrhenoblastomas, hepatoma, hematologic malignancies including non-Hodgkin’s lymphoma (NHL), multiple myeloma and acute hematologic malignancies, endometrial or uterine carcinoma, endometriosis, fibrosarcomas, choriocarcinoma, salivary gland carcinoma, vulval cancer, thyroid cancer, esophageal carcinomas, hepatic carcinoma, anal carcinoma, penile carcinoma, nasopharyngeal carcinoma, laryngeal carcinomas, Kaposi's sarcoma, melanoma, skin carcinomas, Schwannoma, oligodendroglioma, neuroblastomas, rhabdomyosarcoma, osteogenic sarcoma, leiomyosarcomas, and urinary tract carcinomas.

[0105] The term “metastatic cancer” means the state of cancer where the cancer cells of a tissue of origin are transmitted from the original site to one or more sites elsewhere in the body, by the blood vessels or lymphatics, to form one or more secondary tumors in one or more organs besides the tissue of origin.

[0106] As used herein, an “AS-SPIK-associated disorder” of a “disorder that is characterized by expression of AS-SPIK” is a disorder that is associated with expression or over-expression of an AS-SPIK gene or gene product (an AS-SPIK polypeptide), which can be any disorder that is characterized by cells that

express normal or elevated levels of AS-SPIK, relative to suitable control cells. Suitable control cells can be cells from an individual who is not affected with an AS- SPIK-expressing or over-expressing cancer, or they may be non-cancerous cells from either the subject in need, or they may be non-cancerous cells from another individual who is affected with an AS-SPIK-expressing or over-expressing cancer. One prominent example of an AS-SPIK-associated disorder is liver cancer.

[0107] The terms “cell proliferative disorder” and “proliferative disorder” refer to disorders that are associated with some degree of abnormal cell proliferation. In one embodiment, the cell proliferative disorder is cancer.

[0108] “Tumor”, as used herein, refers to all neoplastic cell growth and proliferation, whether malignant or benign, and all pre-cancerous and cancerous cells and tissues.

[0109] The terms “predictive” and “prognostic” as used herein are also interchangeable, in the sense of meaning that the methods for prediction or prognostication are to allow the person practicing the method to select patients that are deemed (usually in advance of treatment, but not necessarily) more likely to respond to treatment with an anti-cancer agent, including an anti-AS- SPIK antibody.

[0110] The terms “treat”, “treatment” or “treating” as used herein refer to both therapeutic treatment and prophylactic or preventative measures, wherein the object is to prevent or slow down (lessen) a targeted pathological condition or disorder. A subject in need of treatment includes those already having a particular condition or disorder, as well as those prone to having the disorder or those in whom the disorder is to be prevented.

Detailed Description of the Invention

[0111] As demonstrated herein, AS-SPIK has the complete amino acid sequence of genetic SPIK, but NS-SPIK is shorter than AS-SPIK due to the removal of 23 amino acids in its N-terminus during secretion. This difference in size was confirmed by gel electrophoresis and Edman Degradation protein sequencing.

[0112] Additionally, the data herein demonstrate that the 3D conformations of AS-SPIK and NS-SPIK are different. Retaining the additional 23 amino acids in its N-terminus significantly alters the conformation of AS-SPIK relative to NS-SPIK. Three structural/conformational changes are visualized through the comparison of the 3D structures (crystal) of AS-SPIK and NS-SPIK. The first change is in the N-terminus of protein. The extra 23-residue fragment in AS-SPIK projects outwards and extends past the main body of the protein; in contrast, the N-terminus of NS-SPIK does not have an additional fragment that protrudes from the main body of the protein. Second, due to the longer N-terminus of AS-SPIK, the first loop in AS-SPIK is flatter and angled differently compared to the corresponding loop in NS-SPIK. This difference leads to more space between the first loop and alpha helix in AS-SPIK, which exposes amino acids that are

on the interior and inaccessible in NS-SPIK. Finally, the longer N-terminus of AS-SPIK also changes the relative position and distance between the N-terminus and alpha-helix of the protein, increasing the space in this local region (FIG. 4).

[0113] Due to these differences in size and conformation, it is possible to generate many different antibodies that specifically bind to AS-SPIK, but do not bind to NS-SPIK. This was confirmed by developing approximately 20 different monoclonal antibodies in mice and rabbits, which specifically bind to AS-SPIK, and do not bind to NS-SPIK. FIG. 5 shows that 8 of the developed monoclonal antibodies (IM-A1, IM-B10, IM-C6, IM-E2, IM-CA22, IM-CA18, IM-CA46, and IMCA77), whether mouse or rabbit, strongly bind to AS-SPIK while having negligible to no binding activity with NS-SPIK (similar to the negative control). Poly S, a polyclonal antibody developed in sheep, strongly binds to both AS-SPIK and NS-SPIK, as it contains multiple antibodies that bind to various epitopes.

[0114] Through additional analyses of the aforementioned 8 antibodies, we determined that all of them bind to conformational epitopes. This is supported by testing each antibody with various synthesized peptides in order to evaluate their potential to bind to various combinations of linear epitope regions of AS-SPIK and NS-SPIK. Four peptides, each containing sequences from different regions of AS-SPIK, were synthesized by BioMatik (Wilmington DE), including Peptide A, which contains the entire sequence of AS-SPIK but without any disulfide bonds due to the inactivation of cysteine side groups. Peptide B contains the AS-SPIK fragment M₁-G₅₀, Peptide C contains the AS-SPIK fragment D₂₃-G₅₀, and Peptide D contains the AS-SPIK fragment N₅₁-C₇₉. The test results show that none of these 8 antibodies binds to any of the synthetic peptides, and supports the hypothesis that the anti-AS-SPIK antibodies we developed bind to conformational epitopes (FIG. 6).

[0115] The data provided herein demonstrate that the anti-AS-SPIK antibodies described herein can be divided into two classes: Class I and Class II. Class I antibodies function similarly to IM-CA22, and include IM-A1, IM-B10, IM-CA18, IM-D2, IM-D3, IM-D5 and IM-G2. Class II antibodies function similarly to IM-E2, and include IM-C6, IM-CA46, IM-CA77, IM-A6, IM-B3, IM-F5 and IM-G6. Most importantly, we demonstrate that any Class I antibody can work with any Class II antibody as a pair in a sandwich ELISA test, and vice versa. This is due to the fact that Class I antibodies bind to a conformational epitope that is different from the conformational epitope to which Class II antibodies bind. Additionally, when used as a pair in a sandwich ELISA, antibodies in Class I entirely inhibit binding with any other Class I antibody, and antibodies in Class II entirely inhibit binding with any other Class II antibody (FIG. 13; Table 1).

[0116] In addition to the competition test results described herein, we performed Precision Epitope Mapping with CLIPS (Chemically Linked Peptides on Scaffolds) Peptide Arrays to more conclusively show that the binding epitope of Class I antibodies is distinct from that of Class II antibodies. The CLIPS epitope

mapping suggests that Class I antibodies binds to Epitope I, which is a discontinuous, conformational epitope composed of at least two separate regions within AS-SPIK. Similarly, Class II antibodies bind to Epitope II, which is also a discontinuous, conformational epitope composed of another two separate regions within AS-SPIK (FIG. 14; Table 2).

[0117] The CLIPS study identified γ FLLSALALLSLSGNTGADSLGREAA₂₉ (SEQ ID NO: 7) and δ CVLCFENRKRQ₆₈ (SEQ ID NO: 8) as the essential binding sites for all Class I antibodies. Within these binding sites of Epitope I, the critical residues are δ L₁₇ (SEQ ID NO: 12), δ S₂₅ (SEQ ID NO: 13), and δ CVLCF₆₂ (SEQ ID NO: 14) (FIG. 14; Table 2).

[0118] The CLIPS study also identified δ LNGCTKIYD₄₄ (SEQ ID NO: 9) and δ NRKRQTSILIQ₇₄ (SEQ ID NO: 10) as the essential binding sites for all Class II antibodies. Within these binding sites of Epitope II, the critical residues are δ LN₃₇ (SEQ ID NO: 15), δ IY₄₃ (SEQ ID NO: 16), δ RQ₆₈ (SEQ ID NO: 17), and δ IL₇₂ (SEQ ID NO: 18) (FIG. 14; Table 2).

[0119] In addition to defining the epitopes for both classes of antibodies, also provided herein are structural similarities among each class of antibodies, which are defining characteristics of their respective antibody genera. Class I antibodies have remarkable similarity in their CDRs and the following residues are conserved to a high degree: a) S6 in CDRH1, b) I2, G5, G6, Y10 and K16 in CDRH2, c) G4 and Y7 in CDRH3, d) Q4 and S9 in CDRL1, e) A2, S3, T4 and S7 in CDRL2, and f) Q1, Q2, Y4 and S5 in CDRL3 (FIG. 9). Class II antibodies also have remarkable similarity in their CDRs and the following residues are conserved to a high degree: a) Y3, S7 and W9 in CDRH1, b) A1, I2, G4, G6 and Y10 in CDRH2, c) R1 and D7 in CDRH3, d) A2, S3, Q4, I6, Y9, L10 and S11 in CDRL1, e) A2, S3, L5 and S7 in CDRL2, and f) Q1, Q2, and T5 in CDRL3 (FIG. 10). In some embodiments, anti-AS-SPIK antibodies can comprise any suitable combination of CDR sequences that comprise the conserved amino acid residues listed above. For example, in some embodiments, a Class I anti-AS-SPIK antibody can comprise 1, 2, 3, 4, 5, or all six of the of the following CDR sequences, or any combination thereof: a) S6 in CDRH1, b) I2, G5, G6, Y10 and K16 in CDRH2, c) G4 and Y7 in CDRH3, d) Q4 and S9 in CDRL1, e) A2, S3, T4 and S7 in CDRL2, and f) Q1, Q2, Y4 and S5 in CDRL3 (FIG. 9). In some embodiments, a Class II anti-AS-SPIK antibody can comprise 1, 2, 3, 4, 5, or all six of the of the following CDR sequences, or any combination thereof: a) Y3, S7 and W9 in CDRH1, b) A1, I2, G4, G6 and Y10 in CDRH2, c) R1 and D7 in CDRH3, d) A2, S3, Q4, I6, Y9, L10 and S11 in CDRL1, e) A2, S3, L5 and S7 in CDRL2, and f) Q1, Q2, and T5 in CDRL3 (FIG. 10).

[0120] The amino acid sequences of various examples of Class I antibodies that bind to Epitope I are provided in Table 3 (FIG. 15). The amino acid sequences of various examples of Class II antibodies that bind to Epitope II are provided in Table 4 (FIG. 16).

[0121] Aspects of the present invention are based, at least in part, on the discovery that certain disorders are characterized by expression of a unique form of serine protease inhibitor Kazal (SPIK). One prominent example is liver cancer, which includes, without limitation, hepatocellular carcinoma (HCC) and intrahepatic cholangiocarcinoma (ICC). More specifically, the inventors have found that certain cancers, such as liver cancer, express a form of SPIK that includes an additional 23 amino acids at the N-terminus of the secreted SPIK polypeptide. This 23 amino acid segment (SEQ ID NO: 6) is not found in the SPIK polypeptide secreted from normal cells, such as pancreatic cells. This is consistent with our previous report that the first 9 amino acids of this 23 amino acid segment may exist in unprocessed SPIK secreted by a liver cancer cell line. Lu et al., Immunology 2011; 134(4):398-408. We may refer to the longer form of SPIK as AS-SPIK or Abnormal Secreted SPIK. We may also refer to AS-SPIK produced by liver cancer cells as LC-SPIK or Liver Cancer Secreted SPIK. The terms AS-SPIK and LC-SPIK are used interchangeably herein. An exemplary AS-SPIK polypeptide can have the amino acid sequence of SEQ ID NO: 2. We may refer to the form of SPIK secreted by normal cells, such as pancreas cells, as NS-SPIK or Normal Secreted SPIK. An exemplary NS-SPIK polypeptide can have the amino acid sequence of SEQ ID NO: 4.

[0122] The inventors have demonstrated herein that AS-SPIK is different from NS-SPIK in terms of both its size and conformation (three-dimensional structure). Antibodies which selectively bind to AS-SPIK, but not to NS-SPIK, can be divided into two classes (Class I and Class II), according to the epitope to which they bind. Class I anti-AS-SPIK antibodies bind to Epitope I and Class II anti-AS-SPIK antibodies bind to Epitope II, both of which epitopes are described herein. Epitope mapping and analysis of the 3D structure of AS-SPIK shows that each epitope is conformational and discontinuous, consisting of at least two separate regions, and the specific epitope sequences also are identified. In addition, further analysis of antibodies of each class has identified consensus sequences for Class I and Class II antibodies that are critical to their functionality. Methods of using such antibodies to diagnose and/or treat disorders characterized by the expression of AS-SPIK (e.g., liver cancer) are also described herein.

[0123] Accordingly, aspects of the invention include compositions, such as antibodies, that specifically or preferentially bind to AS-SPIK, and that do not bind to NS-SPIK. Also provided are AS-SPIK complexes. AS-SPIK complexes in accordance with embodiments of the invention comprise an antibody that specifically or preferentially binds to AS-SPIK, and an AS-SPIK polypeptide, or fragment thereof.

[0124] Aspects of the invention include antibody-drug conjugates (ADCs) that comprise an antibody as described herein (Ab), a linker (L), and a drug moiety (D). In some embodiments, an ADC has the formula Ab-(L-D)_p, where p is an integer that ranges from 1 to 8.

[0125] Aspects of the invention also include methods of using the subject antibodies for the detection of a disorder characterized by expression of AS-SPIK, e.g., a liver disorder, such as a liver cancer, for example, HCC or ICC.

[0126] While we believe we understand certain events that occur during the expression of AS- SPIK, the compositions and methods of the present invention are not limited to those that work by affecting any particular cellular mechanism. Without being held to theory, the inventors hypothesize that because SPIK is a protease inhibitor, over-expression of SPIK in cancer cells suppresses the activity of signal peptidase, one kind of protease, resulting in un-attenuated, full-length protein being secreted from cancer cells.

Compositions

[0127] The compositions provided herein include antibodies that specifically or preferentially bind to AS-SPIK and that do not bind to NS-SPIK.

[0128] Serine protease inhibitor Kazal (SPIK), also known as SPINK1, PSTI, and TATI, is a small protein that has been shown to broadly regulate the activity of many cellular proteases, such as trypsin-like proteases and chymotrypsin-like proteases. Greene, LJ, J Surg Oncol. 1975;7(2):151- 154; Horii et al., Biochemical and biophysical research communications 1987;149(2):635-641; Stenman, UH, Clin Chem. 2002;48(8):1206-1209. SPIK may also play a role in inhibition of apoptosis. Lu et al., Immunology 2011;134(4):398-408. Exemplary human SPIK amino acid sequences include GenBank Accession Number: M11949, GI Number: 190687; GenBank Accession Number: NM003122, GI: 657940887; and GeneBank Accession Number: BC025790, GI: 19343607.

Antibodies

[0129] The antibodies provided herein can include an antibody that specifically or preferentially binds to a conformational epitope on an AS-SPIK protein. In some embodiments, an antibody specifically or preferentially binds to a discontinuous, conformational epitope, as described above. Antibodies in accordance with embodiments of the invention may be polyclonal or monoclonal, particularly monoclonal, and may be produced by human, mouse, rabbit, sheep or goat cells, or by hybridomas derived from these cells. In some embodiments, an antibody can be humanized, or chimeric.

[0130] Antibodies in accordance with embodiments of the invention can assume various configurations and encompass proteins consisting of one or more polypeptides substantially encoded by immunoglobulin genes. Any one of a variety of antibody structures can be used, including the intact antibody, antibody multimers, or antibody fragments or other variants thereof that include functional, antigen-binding regions of the antibody. The term “immunoglobulin” may be used synonymously with “antibody.” The antibodies

may be monoclonal or polyclonal in origin. Regardless of the source of the antibody, suitable antibodies include intact antibodies, for example, IgG tetramers having two heavy (H) chains and two light (L) chains, single chain antibodies, chimeric antibodies, humanized antibodies, complementary determining region (CDR)-grafted antibodies as well as antibody fragments, e.g., Fab, Fab', F(ab')₂, scFv, Fv, and recombinant antibodies derived from such fragments, e.g., camelbodies, microantibodies, diabodies and bispecific antibodies.

[0131] An intact antibody is one that comprises an antigen-binding variable region (VH and VL) as well as a light chain constant domain (CL) and heavy chain constant domains, CH1, CH2 and CH3. The constant domains may be native sequence constant domains (e.g. human native sequence constant domains) or amino acid sequence variants thereof. As is well known in the art, the VH and VL regions are further subdivided into regions of hypervariability, termed “complementarity determining regions” (CDRs), interspersed with the more conserved framework regions (FRs). The CDR of an antibody typically includes amino acid sequences that together define the binding affinity and specificity of the natural Fv region of a native immunoglobulin binding site.

[0132] An anti-AS-SPIK antibody can be from any class of immunoglobulin, for example, IgA, IgG, IgE, IgD, IgM (as well as subtypes thereof (e.g., IgG1, IgG2, IgG3, and IgG4)), and the light chains of the immunoglobulin may be of types kappa or lambda. The recognized human immunoglobulin genes include the kappa, lambda, alpha (IgA1 and IgA2), gamma (IgG1, IgG2, IgG3, IgG4), delta, epsilon, and mu constant region genes, as well as the myriad immunoglobulin variable region genes.

[0133] The term “antigen-binding portion” of an immunoglobulin or antibody refers generally to a portion of an immunoglobulin that specifically or preferentially binds to a target, in this case, a conformational epitope of an AS-SPIK protein. An antigen-binding portion of an immunoglobulin is therefore a molecule in which one or more immunoglobulin chains are not full length, but which specifically or preferentially binds to a target. Examples of antigen-binding portions or fragments include: (i) an Fab fragment, a monovalent fragment consisting of the VLC, VHC, CL and CH1 domains; (ii) a F(ab')₂ fragment, a bivalent fragment comprising two Fab fragments linked by a disulfide bridge at the hinge region; (iii) a Fv fragment consisting of the VLC and VHC domains of a single arm of an antibody, and (v) an isolated CDR having sufficient framework to specifically or preferentially bind, e.g., an antigen binding portion of a variable region. An antigen-binding portion of a light chain variable region and an antigen binding portion of a heavy chain variable region, e.g., the two domains of the Fv fragment, VLC and VHC, can be joined, using recombinant methods, by a synthetic linker that enables them to be made as a single protein chain in which the VLC and VHC regions pair to form monovalent molecules (known as single chain Fv (scFv)). Such scFvs are encompassed by the term “antigen-binding portion” of an antibody.

[0134] An “Fv” fragment is the minimum antibody fragment that contains a complete antigen- recognition and binding site. This region consists of a dimer of one heavy chain and one light chain variable domain in tight, con-covalent association. It is in this configuration that three hypervariable regions of each variable domain interact to define an antigen-binding site on the surface of the VH-VL dimer. While six hypervariable regions confer antigen-binding specificity, even a single variable domain (or half of an Fv comprising only three hypervariable regions specific for an antigen) has the ability to recognize and bind antigen, although at a lower affinity than the entire binding site. To improve stability, the VH-VL domains may be connected by a flexible peptide linker such as (Gly4Ser)₃ to form a single chain Fv or scFV antibody fragment or may be engineered to form a disulfide bond by introducing two cysteine residues in the framework regions to yield a disulfide stabilized Fv (dsFv).

[0135] Fragments of antibodies are suitable for use in the methods provided so long as they retain the desired epitope specificity of the full-length antibody and/or sufficient specificity to bind AS-SPIK and not NS-SPIK.

[0136] The antigen-binding domains of the antibodies described herein can be utilized in the production of T-cell engager molecules (e.g., bispecific T-cell engagers, aka BiTE molecules) as well as CAR-T structures. T-cell engager molecules are described, for example, in Huehls et al., Bispecific T cell engagers for cancer immunotherapy, *Immunol Cell Biol.* 2015 Mar; 93(3):290-296. CAR-T structures comprising single-domain antibodies as a binding (targeting) domain are described, for example, in Iri-Sofla et al., 2011, *Experimental Cell Research* 317:2630-2641 and Jamnani et al., 2014, *Biochim Biophys Acta*, 1840:378-386.

[0137] Methods for preparing antibody fragments encompass both biochemical methods (e.g. proteolytic digestion of intact antibodies which may be followed by chemical cross-linking) and recombinant DNA-based methods in which immunoglobulin sequences are genetically engineered to direct the synthesis of the desired fragments. Antibody fragments can be obtained by proteolysis of the whole immunoglobulin by the non-specific thiolprotease, papain. Papain digestion yields two identical antigen-binding fragments, termed “Fab fragments,” each with a single antigen-binding site, and a residual “Fc fragment.” The various fractions can be separated by protein A-Sepharose or ion exchange chromatography. The usual procedure for preparation of F(ab')₂ fragments from IgG of rabbit and human origin is limited proteolysis by the enzyme pepsin. Pepsin treatment of intact antibodies yields an F(ab')₂ fragment that has two antigen-combining sites and is still capable of cross-linking antigen. A Fab fragment contains the constant domain of the light chain and the first constant domain (CH1) of the heavy chain. Fab' fragments differ from Fab fragments by the addition of a few residues at the carboxyl terminus of the heavy chain CH1 domain

including one or more cysteine(s) from the antibody hinge region. F(ab')₂ antibody fragments were originally produced as pairs of Fab' fragments that have hinge cysteines between them.

[0138] Also within the scope of the present invention are methods of making an anti-AS-SPIK antibody. For example, variable regions can be constructed using PCR mutagenesis methods to alter DNA sequences encoding an immunoglobulin chain (e.g., using methods employed to generate humanized immunoglobulins).

[0139] Monoclonal antibodies are homogeneous antibodies of identical antigenic specificity produced by a single clone of antibody-producing cells, and polyclonal antibodies generally recognize different epitopes on the same antigen and are produced by more than one clone of antibody producing cells. Each monoclonal antibody is directed against a single determinant on the antigen. The modifier, monoclonal, indicates the character of the antibody as being obtained from a substantially homogeneous population of antibodies, and is not to be construed as requiring production of the antibody by any particular method.

[0140] The monoclonal antibodies herein can include chimeric antibodies, i.e., antibodies that typically have a portion of the heavy and/or light chain identical with or homologous to corresponding sequences in antibodies derived from a particular species or belonging to a particular antibody class or subclass, while the remainder of the chain(s) is identical with or homologous to corresponding sequences in antibodies derived from another species or belonging to another antibody class or subclass, as well as fragments of such antibodies, so long as they exhibit the desired biological activity. Chimeric antibodies of interest include primatized antibodies comprising variable domain antigen-binding sequences derived from a non-human primate (e.g. apes, Old World monkeys, New World monkeys, prosimians) and human constant region sequences.

[0141] Murine and rabbit monoclonal antibodies were generated through the immunization of a mouse or a rabbit with specifically designed recombinant proteins, that have the extra 23 amino acid sequence found in AS-SPIK (SEQ ID NO: 6) that is not found in NS-SPIK, in addition to the common region (SEQ ID NO: 4), which is the amino acid sequence found both in NS-SPIK and AS-SPIK. In some embodiments, the recombinant proteins may not need to have the entire 23 amino acid sequence (SEQ ID NO: 6) to generate an antibody being effective at binding only to AS-SPIK but not to NS-SPIK.

[0142] Methods for producing monoclonal antibodies can include purification steps. For example, the antibodies can generally be further purified, for example, using filtration, centrifugation and various chromatographic methods, such as HPLC or affinity chromatography, all of which are techniques well known to one of ordinary skill in the art. These purification techniques each involve fractionation to separate the desired antibody from other components of a mixture. Analytical methods particularly suited to the

preparation of antibodies include, for example, protein A-Sepharose and/or protein G-Sepharose chromatography.

[0143] The anti-AS-SPIK antibodies of the invention may include CDRs from a human or non-human source. "Humanized" antibodies are generally chimeric or mutant monoclonal antibodies from mouse, rat, hamster, rabbit or other species, bearing human constant and/or variable region domains or specific changes. The framework of the immunoglobulin can be human, humanized, or non-human (e.g., a murine framework modified to decrease antigenicity in humans), or a synthetic framework (e.g., a consensus sequence). Humanized immunoglobulins are those in which the framework residues correspond to human germline sequences and the CDRs result from V(D)J recombination and somatic mutations. However, humanized immunoglobulins may also comprise amino acid residues not encoded in human germline immunoglobulin nucleic acid sequences (e.g., mutations introduced by random or site-specific mutagenesis *ex vivo*). An antibody variable domain gene based on germline sequence but possessing framework mutations introduced by, for example, an *in vivo* somatic mutational process is termed "human."

[0144] Humanized antibodies may be engineered by a variety of methods known in the art including, for example: (1) grafting the non-human complementarity determining regions (CDRs) onto a human framework and constant region (a process referred to in the art as humanizing), or, alternatively, (2) transplanting the entire non-human variable domains, but providing them with a human-like surface by replacement of surface residues (a process referred to in the art as veneering). Humanized antibodies can include both humanized and veneered antibodies. Similarly, human antibodies can be made by introducing human immunoglobulin loci into transgenic animals, e.g., mice in which the endogenous immunoglobulin genes have been partially or completely inactivated. Upon challenge, human antibody production is observed, which closely resembles that seen in humans in all respects, including gene rearrangement, assembly, and antibody repertoire.

[0145] In addition to chimeric and humanized antibodies, fully human antibodies can be derived from transgenic mice having human immunoglobulin genes. In some embodiments, antibodies may be produced and identified by scFv-phage display libraries.

[0146] The anti-AS-SPIK antibodies may be modified to modulate their antigen binding affinity, their effector functions, or their pharmacokinetics. In particular, random mutations can be made in the CDRs and products screened to identify antibodies with higher affinities and/or higher specificities. Typically, the CDRs may differ in 1 or 2 amino acids.

[0147] CDR shuffling and implantation technologies can be used with the antibodies provided herein, for example. CDR shuffling inserts CDR sequences into a specific framework region. CDR implantation techniques permit random combination of CDR sequences into a single master framework. Using such

techniques, CDR sequences of the anti-AS-SPIK antibody, for example, can be mutagenized to create a plurality of different sequences, which can be incorporated into a scaffold sequence and the resultant antibody variants screened for desired characteristics, e.g., higher affinity.

[0148] Our study of the function of SPIK shows that SPIK can bind Granzyme A (GzmA) and inhibit it from inducing apoptosis. Lu et al., *Immunology* 2011;134(4):398-408. GzmA is a cytotoxic serine protease secreted by activated CTLs and NK cells to kill target cells during immune surveillance. The role of GzmA-induced apoptosis in removal of malignant cells, such as tumor precursor/tumor germ cells, has been confirmed. Pardo et al., *Eur J Immunol* 2002;32(10):2881-2887. Therefore, it is possible that over-expression of AS-SPIK in liver cancer cells will cause the cell to be resistant to the apoptosis induced by GzmA during immune-clearance. This results in the escape of these cancer cells from killing by the immune pathways. Lu et al., *Immunology* 2011;134(4):398-408. Based on this hypothesis, and without being held to theory, we conclude that suppression of the over-expression of AS-SPIK, or the inhibition of the activity of over-expressed AS-SPIK, may restore the immuno-killing of cancer cells induced by GzmA during the human body's immune clearance.

[0149] Anti-AS-SPIK antibodies in accordance with embodiments of the invention can inhibit the activity of AS-SPIK, as demonstrated by the disclosure of PCT Application No. PCT/US19/20999, the disclosure of which is incorporated herein by reference in its entirety. Therefore, it is possible to use an anti-AS-SPIK antibody to block the binding of AS-SPIK with GzmA, free the GzmA, and restore the apoptotic killing of these cancer cells via immune-clearance. For this purpose, an anti-SPIK antibody may be used in the treatment of disorders characterized by the expression of AS-SPIK, including, but not limited to, cancer, viral infection, and inflammation.

[0150] One therapeutic use of antibodies is through humanization. Therapy with humanized monoclonal antibodies is an area that is being developed rapidly and their specificity and efficiency are well studied. Rothemberg, ME, *Cell* 2016;165(3):509. The subject anti-AS-SPIK monoclonal antibodies, including but not limited to IM-CA22, IM-A1, IM-B10, IM-CA18, IM-D2, IM-D3, IM-D5, IM-G2, IM-E2, IM-C6, IM-CA46, IM-CA77, IM-A6, IM-B3, IM-F5 and IM-G6, and other antibodies of the invention, such as antibodies that bind to Epitope I or Epitope II, as described herein, and which are able to inhibit the activity of SPIK, can also be humanized and used for treatment of disease.

[0151] Recombinant technology using, for example phagemid technology, allows for preparation of antibodies having a desired specificity from recombinant genes encoding a range of antibodies. Certain recombinant techniques involve isolation of antibody genes by immunological screening of combinatorial immunoglobulin phage expression libraries prepared from RNA isolated from spleen of an immunized animal. For such methods, combinatorial immunoglobulin phagemid libraries can be prepared from RNA

isolated from spleen of an immunized animal, and phagemids expressing appropriate antibodies can be selected by panning using cells expressing antigen and control cells.

[0152] In addition to the combinatorial immunoglobulin phage expression libraries disclosed above, one molecular cloning approach is to prepare antibodies from transgenic mice containing human antibody libraries. Such transgenic animals can be employed to produce human antibodies of a single isotype, more specifically an isotype that is essential for B cell maturation, such as IgM and possibly IgD.

[0153] The anti-AS-SPIK immunoglobulins may be modified to reduce or abolish glycosylation. An immunoglobulin that lacks glycosylation may be an immunoglobulin that is not glycosylated at all; that is not fully glycosylated; or that is atypically glycosylated (i.e., the glycosylation pattern for the mutant differs from the glycosylation pattern of the corresponding wild type immunoglobulin). The IgG polypeptides include one or more (e.g., 1, 2, or 3 or more) mutations that attenuate glycosylation, i.e., mutations that result in an IgG CH2 domain that lacks glycosylation, or is not fully glycosylated or is atypically glycosylated. The oligosaccharide structure can also be modified, for example, by eliminating the fucose moiety from the N-linked glycan.

[0154] Antibodies can also be modified to increase their stability and or solubility in vivo by conjugation to non-protein polymers, e.g., polyethylene glycol. Any PEGylation method can be used as long as the anti-AS-SPIK antibody retains the ability to selectively bind AS-SPIK and not NS-SPIK.

[0155] A wide variety of antibody/immunoglobulin frameworks or scaffolds can be employed so long as the resulting polypeptide includes at least one binding region that is specific for the target, i.e., AS-SPIK. Such frameworks or scaffolds include the five main idiotypes of human immunoglobulins, or fragments thereof (such as those disclosed elsewhere herein), and include immunoglobulins of other animal species, preferably having humanized aspects. Single heavy-chain antibodies such as those identified in camelids are of particular interest in this regard.

[0156] The anti-AS-SPIK antibodies of the invention specifically or preferentially bind to an epitope on AS-SPIK and do not bind to an epitope on NS-SPIK. An epitope refers to an antigenic determinant on a target that is specifically bound by the paratope, i.e., the binding site of an antibody. Epitopic determinants usually consist of chemically active surface groupings of molecules such as amino acids or sugar side chains, and typically have specific three-dimensional structural characteristics, as well as specific charge characteristics. Epitopes generally have between about 4 to about 10, preferably 4 to 8, contiguous amino acids (a linear or continuous epitope), or alternatively can be a set of noncontiguous amino acids that define a particular structure (e.g., a conformational epitope). Thus, an epitope can consist of at least 4, at least 6, at least 8, at least 10, and at least 12 such amino acids. Methods of determining the spatial conformation of amino acids are known in the art, and include, for example, x-ray crystallography, 2-dimensional nuclear

magnetic resonance, and Precision Epitope Mapping with CLIPS (Chemically Linked Peptides on Scaffolds) Peptide Array (Timmerman, Puijk et al., *J Mol Recognit*, 20(5), 283-299, (2007).

[0157] Methods of predicting other potential epitopes to which an antibody can bind can include but are not limited to, Kyte-Doolittle Analysis (Kyte and Doolittle, *J. Mol. Biol.* 157:105-132 (1982)), Hopp and Woods Analysis (Hopp and Woods, *Proc. Natl. Acad. Sci. USA* 78:3824-3828 (1981); Hopp and Woods, *Mol. Immunol.* 20:483-489 (1983); Hopp, *J. Immunol. Methods* 88:1-18 (1986)), Jameson-Wolf Analysis (Jameson and Wolf, *Comput. Appl. Biosci.* 4:181-186 (1988)), and Emini Analysis (Emini et al., *Virology* 140:13-20 (1985)), Chou and Fasman analysis (Ponomarenko & Regenmortel, *Structural Bioinformatics*, 2009), Karplus and Schulz Analysis (Kolaskar and Tongaonkar Analysis Kolaskar & Tongaonkar, *FEBS Letters*, 172-174 (1990)), and Parker analysis. In some embodiments potential epitopes are determined through correlations with known antigenic sites from other studies and these predictive techniques can be combined with structural data, such as X-ray crystallographic data. Epitope prediction may also include techniques that predict both continuous and discontinuous epitopes. Methods of predicting discontinuous epitopes includes but are not limited to the following: DiscoTope, BEpro, ElliPro, SEPPA, EPITOPIA, EPCES, Bpredictor, and EPMeta (Yao et al., *PLOS ONE*, (2013)). In some embodiments, potential epitopes are identified by determining theoretical extracellular domains. Analysis algorithms such as TMPred (see Hofmann and Stoffel, *Biol. Chem.* 374:166 (1993)) or TMHMM (Krogh et al., *J. Mol. Biol.*, 305(3):567-580 (2001)) can be used to make such predictions. Other algorithms, such as SignalP 3.0 (Bednsten et al., *J. Mol. Biol.* 340(4):783-795 (2004)) can be used to predict the presence of signal peptides and to predict where those peptides would be cleaved from the full-length protein. The portions of the proteins on the outside of the cell can serve as targets for antibody interaction.

[0158] The compositions of the present invention include antibodies described herein that (1) exhibit a threshold level of binding activity; (2) do not significantly cross-react with known related polypeptide molecules; (3) bind to AS-SPIK and (4) do not bind to NS-SPIK. The binding affinity of an antibody can be readily determined by one of ordinary skill in the art, for example, by Scatchard analysis (Scatchard, *Ann. NY Acad. Sci.* 51:660-672 (1949)).

[0159] In some embodiments, the anti-AS-SPIK antibodies can bind to their target epitopes or mimetic decoys at least 1.5-fold, 2-fold, 5-fold, 10-fold, 100-fold, 103-fold, 104-fold, 105-fold, 106-fold or greater for the target AS-SPIK than to other proteins predicted to have some homology to AS-SPIK, for example, NS-SPIK.

[0160] In some embodiments, the anti-AS-SPIK antibodies bind with high affinity of 10^{-4} M or less, 10^{-7} M or less, 10^{-9} M or less or with subnanomolar affinity (0.9, 0.8, 0.7, 0.6, 0.5, 0.4, 0.3, 0.2, 0.1 nM or even less). In some embodiments, the binding affinity of the anti-AS-SPIK antibodies for their respective targets

is at least 1×10^6 Ka. In some embodiments the binding affinity of the anti-AS-SPIK antibodies for AS-SPIK is at least 5×10^6 Ka, at least 1×10^7 Ka, at least 2×10^7 Ka, at least 1×10^8 Ka, or greater. Antibodies may also be described or specified in terms of their binding affinity to AS-SPIK. In some embodiments binding affinities include those with a Kd less than 5×10^{-2} M, 10^{-2} M, 5×10^{-3} M, 10^{-3} M, 5×10^{-3} M, 10^{-4} M, 5×10^{-5} M, 10^{-5} M, 5×10^{-6} M, 10^{-6} M, 5×10^{-7} M, 10^{-7} M, 5×10^{-8} M, 10^{-8} M, 5×10^{-9} M, 10^{-9} M, 5×10^{-10} M, 10^{-10} M, 5×10^{-11} M, 10^{-11} M, 5×10^{-12} M, 10^{-12} M, 5×10^{-13} M, 10^{-13} M, 5×10^{-14} M, 10^{-14} M, 5×10^{-15} M, or 10^{-15} M, or less.

[0161] The antibodies of the invention may bind with an affinity of 10^{-4} M or less, 10^{-7} M or less, 10^{-9} M or less or with sub-nanomolar affinity (0.9, 0.8, 0.7, 0.6, 0.5, 0.4, 0.3, 0.2, 0.1 nM or even less). In some embodiments, the binding affinity of the anti-AS-SPIK antibodies for their respective targets is at least 1×10^6 Ka. In some embodiments, the binding affinity of the anti-AS-SPIK antibodies for AS-SPIK is at least 5×10^6 Ka, at least 1×10^7 Ka, at least 2×10^7 Ka, at least 1×10^8 Ka, or greater. In some embodiments, the binding affinities include those with a Kd less than 5×10^{-2} M, 10^{-2} M, 5×10^{-3} M, 10^{-3} M, 5×10^{-3} M, 10^{-4} M, 5×10^{-5} M, 10^{-5} M, 5×10^{-6} M, 10^{-6} M, 5×10^{-7} M, 10^{-7} M, 5×10^{-8} M, 10^{-8} M, 5×10^{-9} M, 5×10^{-10} M, 10^{-10} M, 5×10^{-11} M, 10^{-11} M, 5×10^{-12} M, 10^{-12} M, 5×10^{-13} M, 10^{-13} M, 5×10^{-14} M, 10^{-14} M, 5×10^{-15} M, or 10^{-15} M, or less. In contrast thereto, the term “non-specifically binding”, e.g. to NS-SPIK, as used herein refers to a binding affinity that is by a factor of at least 1.5, 2, 5, 10, 100, 10^3 , 10^4 , 10^5 , 10^6 or greater less than that determined for the “specific binding”, e.g. to AS-SPIK. Affinities, such as Kd, may be measured by a radio-labeled antigen-binding assay (radioimmuno assay, RIA) performed with a Fab-version of an antibody of interest and its antigen. According to another embodiment, Kd may be measured using surface plasmon resonance assays with immobilized antigen. In a preferred embodiment, the antibody of the invention specifically or preferentially binds to AS-SPIK and does not specifically bind to NS-SPIK, wherein the affinity of the antibody to AS-SPIK is at least 1.5-fold, 2-fold, 5-fold, 10-fold, 100-fold, 10^3 -fold, 10^4 -fold, 10^5 -fold or 10^6 -fold greater than to NS-SPIK.

[0162] In some embodiments, the antibodies do not bind to known related polypeptide molecules; for example, they bind AS-SPIK but not known related polypeptides, for example, NS-SPIK. Antibodies may be screened against known related polypeptides to isolate an antibody population that specifically or preferentially binds AS-SPIK. For example, antibodies specific for AS-SPIK will flow through a column comprising NS-SPIK adhered to insoluble matrix under appropriate buffer conditions. Such screening allows isolation of polyclonal and monoclonal antibodies non- cross-reactive to closely related polypeptides. Other methods of screening and isolation of specific antibodies include, without limitation, for example, concurrent immunoelectrophoresis, radioimmunoassay (RIA), radioimmunoprecipitation,

enzyme-linked immunosorbent assay (ELISA), dot blot or Western blot assay, inhibition or competition assay, and sandwich assay.

[0163] Antibodies in accordance with embodiments of the invention can include a detectable label, which may also be referred to as a reporter (e.g., a detectable reporter). In some embodiments, a detectable label can be any molecule that is covalently linked to an antibody (e.g., an anti-AS-SPIK antibody) or a biologically-active fragment thereof that allows for qualitative and/or quantitative assessment of the expression or activity of the tagged peptide. The activity can include a biological activity, a physico-chemical activity, or a combination thereof. Both the form and position of the detectable label can vary, as long as the labeled antibody retains biological activity. Many different labels can be used, and the choice of a particular label will depend upon the desired application. Labeled anti-AS-SPIK antibodies can be used, for example, for assessing the levels of AS-SPIK in a biological sample, e.g., urine, saliva, cerebrospinal fluid, blood or a biopsy sample.

[0164] Detectable labels can include enzymes, photo-affinity ligands, radioisotopes, and fluorescent or chemiluminescent compounds. Exemplary enzymatic labels can include horseradish peroxidase, alkaline phosphatase, β -galactosidase, and urease. The covalent linkage of an anti-AS-SPIK antibody to an enzyme may be performed by different methods, for example, the coupling with glutaraldehyde via free amino groups. Alternatively, anti-AS-SPIK antibody can be coupled to the enzyme via sugar residues. Other enzyme containing carbohydrates can also be coupled to the antibody in this manner. Enzyme coupling may also be performed by interlinking the amino groups of the antibody with free thiol groups of an enzyme, such as β -galactosidase, using a heterobifunctional linker, such as succinimidyl 6-(N-maleimido) hexanoate. The horseradish-peroxidase detection system can be used, for example, with the chromogenic substrate tetramethylbenzidine (TMB), which yields a soluble product in the presence of hydrogen peroxide that is detectable at 450 nm. The alkaline phosphatase detection system can be used with the chromogenic substrate p-nitrophenyl phosphate, for example, which yields a soluble product readily detectable at 405 nm. Similarly, the β -galactosidase detection system can be used with the chromogenic substrate o-nitrophenyl-P-D-galactopyranoside (ONPG), which yields a soluble product detectable at 410 nm. A urease detection system can be used with a substrate, such as urea-bromocresol purple.

[0165] A detectable label can be a fluorescent label, including, but not limited to, fluorescein isothiocyanate, rhodamine, phycoerytherin, phycocyanin, allophycocyanin, o-phthaldehyde and fluorescamine; a chemiluminescent compound selected from the group consisting of luminol, isoluminol, an aromatic acridinium ester, an imidazole, an acridinium salt and an oxalate ester; a liposome or dextran; or a bioluminescent compound such as luciferin, luciferase and aequorin. Alternatively or in addition, detectable labels include, but are not limited to, a radiopaque or contrast agent such as barium, diatrizoate,

ethiodized oil, gallium citrate, iocarmic acid, iocetamic acid, iodamide, iodipamide, iodoxamic acid, iogulamide, iohexol, iopamidol, iopanoic acid, ioprocemic acid, iosefamic acid, ioseric acid, iosulamide meglumine, iosememic acid, iotasul, iotetric acid, iothalamic acid, iotroxic acid, ioxaglic acid, ioxotrizoic acid, ipodate, meglumine, metrizamide, metrizoate, propyl iodone, and thallous chloride.

[0166] Labels can be added during synthesis or post-synthetically. Recombinant anti-AS-SPIK antibodies or biologically active variants thereof can also be labeled by the addition of labeled precursors (e.g., radiolabeled amino acids) to the culture medium in which the transformed cells are grown. In some embodiments, analogues or variants of peptides can be used in order to facilitate incorporation of detectable markers. For example, any N-terminal phenylalanine residue can be replaced with a closely related aromatic amino acid, such as tyrosine, that can be easily labeled with ^{125}I . In some embodiments, additional functional groups that support effective labeling can be added to the fragments of an anti-AS-SPIK antibody or biologically active variants thereof. For example, a 3-tributyltinbenzoyl group can be added to the N-terminus of the native structure; subsequent displacement of the tributyltin group with ^{125}I will generate a radiolabeled iodobenzoyl group.

Antibody Drug Conjugates (ADCs)

[0167] Aspects of the invention include immunoconjugates, or antibody-drug conjugates (ADC), comprising an antibody conjugated to a cytotoxic agent such as a chemotherapeutic agent, a drug, a growth inhibitory agent, a toxin (e.g., an enzymatically active toxin of bacterial, fungal, plant, or animal origin, or fragments thereof), or a radioactive isotope (i.e., a radioconjugate). In another aspect, the invention further provides methods of using the immunoconjugates. In one aspect, an immunoconjugate comprises any of the above anti-AS-SPIK antibodies covalently attached to a cytotoxic agent or a detectable agent. ADCs are described, for example, in US Patent No. 8,362,213, the disclosure of which is incorporated by reference herein in its entirety.

[0168] The use of ADCs for the local delivery of cytotoxic or cytostatic agents, i.e., drugs to kill or inhibit tumor cells in the treatment of cancer (Lambert, J. (2005) *Curr. Opinion in Pharmacology* 5:543-549; Wu et al (2005) *Nature Biotechnology* 23(9):1137-1146; Payne, G. (2003) *Cancer Cell* 3:207-212; Syrigos and Epenetos (1999) *Anticancer Research* 19:605-614; Niculescu-Duvaz and Springer (1997) *Adv. Drug Del. Rev.* 26:151-172; U.S. Pat. No. 4,975,278) allows targeted delivery of the drug moiety to tumors, and intracellular accumulation therein, where systemic administration of these unconjugated drug agents may result in unacceptable levels of toxicity to normal cells as well as the tumor cells sought to be eliminated (Baldwin et al (1986) *Lancet* pp. (Mar. 15, 1986):603-05; Thorpe, (1985) "Antibody Carriers Of Cytotoxic Agents In Cancer Therapy: A Review," in *Monoclonal Antibodies '84: Biological And Clinical*

Applications, A. Pinchera et al (ed.s), pp. 475-506). Efforts to improve the therapeutic index, i.e., maximal efficacy and minimal toxicity of ADC have focused on the selectivity of polyclonal (Rowland et al (1986) *Cancer Immunol. Immunother.*, 21:183-87) and monoclonal antibodies (mAbs) as well as drug-linking and drug-releasing properties (Lambert, J. (2005) *Curr. Opinion in Pharmacology* 5:543-549). Drug moieties used in ADCs include bacterial protein toxins such as diphtheria toxin, plant protein toxins such as ricin, small molecules such as auristatins, geldanamycin (Mandler et al (2000) *J. of the Nat. Cancer Inst.* 92(19):1573-1581; Mandler et al (2000) *Bioorganic & Med. Chem. Letters* 10:1025-1028; Mandler et al (2002) *Bioconjugate Chem.* 13:786-791), maytansinoids (EP 1391213; Liu et al (1996) *Proc. Natl. Acad. Sci. USA* 93:8618-8623), calicheamicin (Lode et al (1998) *Cancer Res.* 58:2928; Hinman et al (1993) *Cancer Res.* 53:3336-3342), daunomycin, doxorubicin, methotrexate, and vindesine (Rowland et al (1986) *supra*). The drug moieties may affect cytotoxic and cytostatic mechanisms including tubulin binding, DNA binding, or topoisomerase inhibition. Some cytotoxic drugs tend to be inactive or less active when conjugated to large antibodies or protein receptor ligands.

[0169] The auristatin peptides, auristatin E (AE) and monomethylauristatin (MMAE), synthetic analogs of dolastatin (WO 02/088172), have been conjugated as drug moieties to: (i) chimeric monoclonal antibodies cBR96 (specific to Lewis Y on carcinomas); (ii) cAC10 which is specific to CD30 on hematological malignancies (Klussman, et al (2004), *Bioconjugate Chemistry* 15(4):765-773; Doronina et al (2003) *Nature Biotechnology* 21(7):778-784; Francisco et al (2003) *Blood* 102(4):1458-1465; US 2004/0018194; (iii) anti-CD20 antibodies such as rituxan (WO 04/032828) for the treatment of CD20-expressing cancers and immune disorders; (iv) anti-EphB2R antibody 2H9 for treatment of colorectal cancer (Mao et al (2004) *Cancer Research* 64(3):781-788); (v) E-selectin antibody (Bhaskar et al (2003) *Cancer Res.* 63:6387-6394); (vi) trastuzumab (HERCEPTIN®, US 2005/0238649), and (vi) anti-CD30 antibodies (WO 03/043583). Variants of auristatin E are disclosed in U.S. Pat. No. 5,767,237 and U.S. Pat. No. 6,124,431. Monomethyl auristatin E conjugated to monoclonal antibodies are disclosed in Senter et al, *Proceedings of the American Association for Cancer Research*, Volume 45, Abstract Number 623, presented Mar. 28, 2004. Auristatin analogs MMAE and MMAF have been conjugated to various antibodies (US 2005/0238649).

[0170] Conventional means of attaching, i.e., linking through covalent bonds, a drug moiety to an antibody generally leads to a heterogeneous mixture of molecules where the drug moieties are attached at a number of sites on the antibody. For example, cytotoxic drugs have typically been conjugated to antibodies through the often-numerous lysine residues of an antibody, generating a heterogeneous antibody-drug conjugate mixture. Depending on reaction conditions, the heterogeneous mixture typically contains a distribution of antibodies with from 0 to about 8, or more, attached drug moieties. In addition, within each subgroup of

conjugates with a particular integer ratio of drug moieties to antibody, is a potentially heterogeneous mixture where the drug moiety is attached at various sites on the antibody. Analytical and preparative methods may be inadequate to separate and characterize the antibody-drug conjugate species molecules within the heterogeneous mixture resulting from a conjugation reaction. Antibodies are large, complex and structurally diverse biomolecules, often with many reactive functional groups. Their reactivities with linker reagents and drug-linker intermediates are dependent on factors such as pH, concentration, salt concentration, and co-solvents. Furthermore, the multistep conjugation process may be non-reproducible due to difficulties in controlling the reaction conditions and characterizing reactants and intermediates.

[0171] Cysteine thiols are reactive at neutral pH, unlike most amines which are protonated and less nucleophilic near pH 7. Since free thiol (RSH, sulfhydryl) groups are relatively reactive, proteins with cysteine residues often exist in their oxidized form as disulfide-linked oligomers or have internally bridged disulfide groups. Extracellular proteins generally do not have free thiols (Garman, 1997, *Non-Radioactive Labelling: A Practical Approach*, Academic Press, London, at page 55). Antibody cysteine thiol groups are generally more reactive, i.e. more nucleophilic, towards electrophilic conjugation reagents than antibody amine or hydroxyl groups. Cysteine residues have been introduced into proteins by genetic engineering techniques to form covalent attachments to ligands or to form new intramolecular disulfide bonds (Better et al (1994) *J. Biol. Chem.* 13:9644-9650; Bernhard et al (1994) *Bioconjugate Chem.* 5:126-132; Greenwood et al (1994) *Therapeutic Immunology* 1:247-255; Tu et al (1999) *Proc. Natl. Acad. Sci. USA* 96:4862-4867; Kanno et al (2000) *J. of Biotechnology*, 76:207-214; Chmura et al (2001) *Proc. Nat. Acad. Sci. USA* 98(15):8480-8484; U.S. Pat. No. 6,248,564). However, engineering in cysteine thiol groups by the mutation of various amino acid residues of a protein to cysteine amino acids is potentially problematic, particularly in the case of unpaired (free Cys) residues or those which are relatively accessible for reaction or oxidation. In concentrated solutions of the protein, whether in the periplasm of *E. coli*, culture supernatants, or partially or completely purified protein, unpaired Cys residues on the surface of the protein can pair and oxidize to form intermolecular disulfides, and hence protein dimers or multimers. Disulfide dimer formation renders the new Cys unreactive for conjugation to a drug, ligand, or other label. Furthermore, if the protein oxidatively forms an intramolecular disulfide bond between the newly engineered Cys and an existing Cys residue, both Cys thiol groups are unavailable for active site participation and interactions. Furthermore, the protein may be rendered inactive or non-specific, by misfolding or loss of tertiary structure (Zhang et al (2002) *Anal. Biochem.* 311:1-9).

[0172] Cysteine-engineered antibodies have been designed as FAB antibody fragments (thioFab) and expressed as full-length, IgG monoclonal (thioMab) antibodies (Junutula, J. R. et al. (2008) *J Immunol Methods* 332:41-52; US 2007/0092940, the contents of which are incorporated by reference). ThioFab and

ThioMab antibodies have been conjugated through linkers at the newly introduced cysteine thiols with thiol-reactive linker reagents and drug-linker reagents to prepare antibody drug conjugates (Thio ADC).

[0173] All references cited herein, including patent applications and publications, are incorporated by reference in their entirety.

Polypeptides

[0174] Aspects of the invention include compositions that comprise a SPIK polypeptide, for example, an AS-SPIK polypeptide encoded by the nucleic acid sequence of SEQ ID NO: 1. The terms “peptide,” “polypeptide,” and “protein” are used interchangeably herein, although typically they refer to peptide sequences of varying sizes. We may refer to the amino acid-based compositions of the invention as “polypeptides” to convey that they are linear polymers of amino acid residues, and to help distinguish them from full-length proteins. A polypeptide in accordance with embodiments of the invention can “constitute” or “include” a fragment of an AS-SPIK polypeptide or an NS-SPIK polypeptide, and the invention encompasses polypeptides that constitute or include biologically active variants of an AS-SPIK polypeptide or an NS-SPIK polypeptide. It will be understood that the polypeptides can therefore include only a fragment of an AS-SPIK polypeptide or an NS-SPIK polypeptide (or a biologically active variant thereof) but may include additional residues as well. Biologically active variants will retain sufficient activity to inhibit proteases.

[0175] The bonds between the amino acid residues can be conventional peptide bonds or another covalent bond (such as an ester or ether bond), and the polypeptides can be modified by amidation, phosphorylation or glycosylation. A modification can affect the polypeptide backbone and/or one or more side chains. Chemical modifications can be naturally occurring modifications made in vivo following translation of an mRNA encoding the polypeptide (e.g., glycosylation in a bacterial host) or synthetic modifications made in vitro. A biologically active variant of an AS-SPIK polypeptide or an NS-SPIK polypeptide can include one or more structural modifications resulting from any combination of naturally occurring (i.e., made naturally in vivo) and with synthetic modifications (i.e., naturally occurring or non-naturally occurring modifications made in vitro). Examples of modifications include, but are not limited to, amidation (e.g., replacement of the free carboxyl group at the C-terminus by an amino group); biotinylation (e.g., acylation of lysine or other reactive amino acid residues with a biotin molecule); glycosylation (e.g., addition of a glycosyl group to either asparagines, hydroxylysine, serine or threonine residues to generate a glycoprotein or glycopeptide); acetylation (e.g., the addition of an acetyl group, typically at the N-terminus of a polypeptide); alkylation (e.g., the addition of an alkyl group); isoprenylation (e.g., the addition of an isoprenoid group); lipoylation (e.g. attachment of a lipoate

moiety); and phosphorylation (e.g., addition of a phosphate group to serine, tyrosine, threonine or histidine).

[0176] One or more of the amino acid residues in a biologically active variant may be a non-naturally occurring amino acid residue. Naturally occurring amino acid residues include those naturally encoded by the genetic code as well as non-standard amino acids (e.g., amino acids having the D-configuration instead of the L-configuration). The present peptides can also include amino acid residues that are modified versions of standard residues (e.g. pyrrolysine can be used in place of lysine and selenocysteine can be used in place of cysteine). Non-naturally occurring amino acid residues are those that have not been found in nature, but that conform to the basic formula of an amino acid and can be incorporated into a peptide. These include D-alloisoleucine(2R,3S)-2-amino-3-methylpentanoic acid and L-cyclopentyl glycine (S)-2-amino-2-cyclopentyl acetic acid. For other examples, one can consult textbooks or the worldwide web (a site is currently maintained by the California Institute of Technology and displays structures of non-natural amino acids that have been successfully incorporated into functional proteins).

[0177] Alternatively, or in addition, one or more of the amino acid residues in a biologically active variant can be a naturally occurring residue that differs from the naturally occurring residue found in the corresponding position in a wildtype sequence. In other words, biologically active variants can include one or more, particularly one or two, amino acid substitutions. We may refer to a substitution, addition, or deletion of amino acid residues as a mutation of the wildtype sequence. As noted, the substitution can replace a naturally occurring amino acid residue with a non-naturally occurring residue or just a different naturally occurring residue. Further the substitution can constitute a conservative or non-conservative substitution. Conservative amino acid substitutions typically include substitutions within the following groups: glycine and alanine; valine, isoleucine, and leucine; aspartic acid and glutamic acid; asparagine, glutamine, serine and threonine; lysine, histidine and arginine; and phenylalanine and tyrosine.

[0178] The polypeptides that are biologically active variants of AS-SPIK can be characterized in terms of the extent to which their sequence is similar to or homologous to the corresponding wild-type polypeptide. For example, the sequence of a biologically active variant can be at least or about 80% homologous to (or identical to) corresponding residues in the wild-type polypeptide. For example, a biologically active variant of an AS-SPIK polypeptide or an NS-SPIK polypeptide can have an amino acid sequence with at least or about 80% sequence homology (e.g., at least or about 85%, 90%, 95%, 96%, 97%, 98%, or 99% sequence homology) (or the recited percentage identity) to an AS-SPIK or NS-SPIK polypeptide (SEQ ID NOs: 2, 4) or to a homolog or ortholog thereof.

[0179] A biologically active variant of an AS-SPIK polypeptide or an NS-SPIK polypeptide will retain sufficient biological activity to be useful in the present methods. The biologically active variants will retain

sufficient activity to function as an inhibitor of protease activity. The biological activity can be assessed in ways known to one of ordinary skill in the art and includes, without limitation, in vitro cleavage assays or functional assays.

[0180] Polypeptides can be generated by a variety of methods including, for example, recombinant techniques or chemical synthesis. Once generated, polypeptides can be isolated and purified to any desired extent. For example, one can use lyophilization following, for example, reversed phase (preferably) or normal phase HPLC, or size exclusion or partition chromatography on polysaccharide gel media such as Sephadex G-25. The composition of the final polypeptide may be confirmed by amino acid analysis after degradation of the peptide by standard means, by amino acid sequencing, or by FAB-MS techniques. Salts, including acid salts, esters, amides, and N-acyl derivatives of an amino group of a polypeptide may be prepared using methods known in the art, and such peptides are useful in the context of the present invention.

[0181] Also provided are AS-SPIK complexes. AS-SPIK complexes in accordance with embodiments of the invention comprise an antibody of the invention, as described herein, which specifically or preferentially binds to AS-SPIK, and an AS-SPIK polypeptide or fragment thereof. The antibody can be any of the anti-AS-SPIK antibodies described herein. The AS-SPIK polypeptide or fragment thereof can be AS-SPIK polypeptides or fragments thereof described herein. In some embodiments, the antibody is an anti-AS-SPIK monoclonal antibody that binds to Epitope I or Epitope II, as described herein. In some embodiments, the AS-SPIK polypeptide is a polypeptide with an amino acid sequence having at least 98% homology to (or identity to) the amino acid sequence of SEQ ID NO: 2. In some embodiments, the AS-SPIK polypeptide is a polypeptide having the amino acid sequence of SEQ ID NO: 2.

[0182] The specific binding of an anti-AS-SPIK antibody such as antibody that binds to Epitope I or Epitope II, as described herein, can form an immune-complex with AS-SPIK or AS-SPIK peptide under certain conditions. The complex can be precipitated from solution for further analysis, for example, with a sandwich ELISA test. Using a 96-well plate immobilized with a second anti-SPIK antibody as a carrier, the immune complex can be caught by plate. The amount of AS-SPIK immune-complex formed can then be determined, if the antibodies in the complex are labeled with a reporter such as horseradish peroxidase (HPR). The AS-SPIK immune-complex also can be caught by agarose beads linking with protein A or G for western blot analysis.

Nucleic acids

[0183] The terms “nucleic acid” and “polynucleotide” are used interchangeably herein to refer to both RNA and DNA, including cDNA, genomic DNA, synthetic DNA, and DNA (or RNA) containing nucleic

acid analogs, any of which may encode a polypeptide of the invention and all of which are encompassed by the invention. Polynucleotides can have essentially any three-dimensional structure. A nucleic acid can be double-stranded or single-stranded (*i.e.*, a sense strand or an antisense strand). Non-limiting examples of polynucleotides include genes, gene fragments, exons, introns, messenger RNA (mRNA) and portions thereof, transfer RNA, ribosomal RNA, siRNA, micro-RNA, ribozymes, cDNA, recombinant polynucleotides, branched polynucleotides, plasmids, vectors, isolated DNA of any sequence, isolated RNA of any sequence, nucleic acid probes, and primers, as well as nucleic acid analogs. In the context of the present invention, nucleic acids can encode a fragment of a naturally occurring AS-SPIK polypeptide or NS-SPIK polypeptide or a biologically active variant thereof. Non-limiting examples of nucleic acid sequences in accordance with embodiments of the invention include SEQ ID NOs: 1 and 3, or a biologically active fragment or variant thereof.

[0184] An “isolated” nucleic acid can be, for example, a naturally-occurring DNA molecule or a fragment thereof, provided that at least one of the nucleic acid sequences normally found immediately flanking that DNA molecule in a naturally-occurring genome is removed or absent. Thus, an isolated nucleic acid includes, but is not limited to, a DNA molecule that exists as a separate molecule, independent of other sequences (e.g., a chemically synthesized nucleic acid, or a cDNA or genomic DNA fragment produced by the polymerase chain reaction (PCR) or restriction endonuclease treatment). An isolated nucleic acid also refers to a DNA molecule that is incorporated into a vector, an autonomously replicating plasmid, a virus, or into the genomic DNA of a prokaryote or eukaryote. In addition, an isolated nucleic acid can include an engineered nucleic acid such as a DNA molecule that is part of a hybrid or fusion nucleic acid. A nucleic acid existing among many (e.g., dozens, or hundreds to millions) of other nucleic acids within, for example, cDNA libraries or genomic libraries, or gel slices containing a genomic DNA restriction digest, is not an isolated nucleic acid.

[0185] Isolated nucleic acid molecules can be produced, for example, by polymerase chain reaction (PCR) techniques, which can be used to obtain an isolated nucleic acid containing a nucleotide sequence described herein, including nucleotide sequences encoding a polypeptide described herein. PCR can be used to amplify specific sequences from DNA as well as RNA, including sequences from total genomic DNA or total cellular RNA. Generally, sequence information from the ends of the region of interest or beyond is employed to design oligonucleotide primers that are identical or similar in sequence to opposite strands of the template to be amplified. Various PCR strategies also are available by which site-specific nucleotide sequence modifications can be introduced into a template nucleic acid.

[0186] Isolated nucleic acids also can be chemically synthesized, either as a single nucleic acid molecule (e.g., using automated DNA synthesis in the 3' to 5' direction using phosphoramidite technology) or as

a series of oligonucleotides. For example, one or more pairs of long oligonucleotides (*e.g.*, >50-100 nucleotides) can be synthesized that contain the desired sequence, with each pair containing a short segment of complementarity (*e.g.*, about 15 nucleotides) such that a duplex is formed when the oligonucleotide pair is annealed. DNA polymerase is used to extend the oligonucleotides, resulting in a single, double-stranded nucleic acid molecule per oligonucleotide pair, which then can be ligated into a vector. Isolated nucleic acids of the invention also can be obtained by mutagenesis of, *e.g.*, a naturally occurring portion of an AS-SPIK- or NS- SPIK-encoding DNA (in accordance with, for example, the formula above).

[0187] Two nucleic acids or the polypeptides they encode may be described as having a certain degree of homology or identity to one another. For example, AS-SPIK polypeptide or an NS-SPIK polypeptide and a biologically active variant thereof may be described as exhibiting a certain degree of homology or identity. Alignments may be assembled by locating short AS-SPIK polypeptide or an NS-SPIK polypeptide sequences in the Protein Information Research (PIR) site (<http://pir.georgetown.edu>), followed by analysis with the “short nearly identical sequences” Basic Local Alignment Search Tool (BLAST) algorithm on the NCBI website (<http://www.ncbi.nlm.nih.gov/blast>).

[0188] To determine sequence homology or identity, a query nucleic acid or amino acid sequence can be aligned to one or more subject nucleic acid or amino acid sequences, respectively, using a computer program, such as, for example, BioEdit (version 4.8.5, North Carolina State University), which allows alignments of nucleic acid or protein sequences to be carried out across their entire length (global alignment), or ALIGN-2, as described above.

[0189] BioEdit calculates the best match between a query and one or more subject sequences and aligns them so that identities, similarities and differences can be determined. Gaps of one or more residues can be inserted into a query sequence, a subject sequence, or both, to maximize sequence alignments. For fast pair wise alignment of nucleic acid sequences, the following default parameters are used: word size: 2; window size: 4; scoring method: percentage; number of top diagonals: 4; and gap penalty: 5. For multiple alignments of nucleic acid sequences, the following parameters are used: gap opening penalty: 10.0; gap extension penalty: 5.0; and weight transitions: yes. For fast pair wise alignment of protein sequences, the following parameters are used: word size: 1; window size: 5; scoring method: percentage; number of top diagonals: 5; gap penalty: 3. For multiple alignment of protein sequences, the following parameters are used: weight matrix: blosum; gap opening penalty: 10.0; gap extension penalty: 0.05; hydrophilic gaps: on; hydrophilic residues: Gly, Pro, Ser, Asn, Asp, Gln, Glu, Arg, and Lys; residue-specific gap penalties: on. The output is a sequence alignment that reflects the relationship between sequences.

[0190] To determine a percent homology between a query sequence and a subject sequence, BioEdit divides the number of identities in the best alignment by the number of residues compared (gap positions are excluded), and multiplies the result by 100. The output is the percent homology of the subject sequence with respect to the query sequence. It is noted that the percent homology value can be rounded to the nearest tenth. For example, 78.11, 78.12, 78.13, and 78.14 are rounded down to 78.1, while 78.15, 78.16, 78.17, 78.18, and 78.19 are rounded up to 78.2.

[0191] The nucleic acids and polypeptides described herein may be referred to as “exogenous.” The term “exogenous” indicates that the nucleic acid or polypeptide is part of, or encoded by, a recombinant nucleic acid construct, or is not in its natural environment. For example, an exogenous nucleic acid can be a sequence from one species introduced into another species, i.e., a heterologous nucleic acid. Typically, such an exogenous nucleic acid is introduced into the other species via a recombinant nucleic acid construct. An exogenous nucleic acid can also be a sequence that is native to an organism and that has been reintroduced into cells of that organism. An exogenous nucleic acid that includes a native sequence can often be distinguished from the naturally occurring sequence by the presence of non-natural sequences linked to the exogenous nucleic acid, e.g., non-native regulatory sequences flanking a native sequence in a recombinant nucleic acid construct. In addition, stably transformed exogenous nucleic acids typically are integrated at positions other than the position where the native sequence is found.

[0192] Recombinant constructs are also provided herein and can be used to transform cells in order to express AS-SPIK. A recombinant nucleic acid construct comprises a nucleic acid encoding an AS-SPIK or NS-SPIK sequence operably linked to a regulatory region suitable for expressing the AS-SPIK or NS-SPIK in the particular cell. It will be appreciated that a number of nucleic acids can encode a polypeptide having a particular amino acid sequence. The degeneracy of the genetic code is well known in the art. For many amino acids, there is more than one nucleotide triplet that serves as the codon for the amino acid. For example, codons in the coding sequence for AS-SPIK or NS-SPIK can be modified such that optimal expression in a particular organism is obtained, using appropriate codon bias tables for that organism.

[0193] Vectors containing nucleic acids such as those described herein also are provided. A “vector” is a replicon, such as a plasmid, phage, or cosmid, into which another DNA segment may be inserted so as to bring about the replication of the inserted segment. Generally, a vector is capable of replication when associated with the proper control elements. Suitable vector backbones include, for example, those routinely used in the art such as plasmids, viruses, artificial chromosomes, BACs, YACs, or PACs. The term “vector” includes cloning and expression vectors, as well as viral vectors and integrating vectors. An “expression vector” is a vector that includes a regulatory region. A wide variety of host/expression vector combinations

may be used to express the nucleic acid sequences described herein. Suitable expression vectors include, but are not limited to, plasmids and viral vectors derived from, for example, bacteriophage, baculoviruses, and retroviruses.

[0194] The vectors provided herein also can include, for example, origins of replication, scaffold attachment regions (SARs), and/or markers. A marker gene can confer a selectable phenotype on a host cell. For example, a marker can confer biocide resistance, such as resistance to an antibiotic (e.g., kanamycin, G418, bleomycin, or hygromycin). As noted above, an expression vector can include a tag sequence designed to facilitate manipulation or detection (e.g., purification or localization) of the expressed polypeptide. Tag sequences, such as green fluorescent protein (GFP), glutathione S-transferase (GST), polyhistidine, c-myc, hemagglutinin, or Flag™ tag (Kodak, New Haven, CT) sequences typically are expressed as a fusion with the encoded polypeptide. Such tags can be inserted anywhere within the polypeptide, including at either the carboxyl or amino terminus.

[0195] Additional expression vectors also can include, for example, segments of chromosomal, non-chromosomal and synthetic DNA sequences. Suitable vectors include derivatives of SV40 and known bacterial plasmids, e.g., E. coli plasmids col E1, pCR1, pBR322, pMal-C2, pET, pGEX, pMB9 and their derivatives, plasmids such as RP4; phage DNAs, e.g., the numerous derivatives of phage 1, e.g., NM989, and other phage DNA, e.g., M13 and filamentous single stranded phage DNA; yeast plasmids such as the 2μ plasmid or derivatives thereof, vectors useful in eukaryotic cells, such as vectors useful in insect or mammalian cells; vectors derived from combinations of plasmids and phage DNAs, such as plasmids that have been modified to employ phage DNA or other expression control sequences.

[0196] The vector can also include a regulatory region. The term “regulatory region” refers to nucleotide sequences that influence transcription or translation initiation and rate, and stability and/or mobility of a transcription or translation product. Regulatory regions include, but are not limited to, promoter sequences, enhancer sequences, response elements, protein recognition sites, inducible elements, protein binding sequences, 5' and 3' untranslated regions (UTRs), transcriptional start sites, termination sequences, polyadenylation sequences, nuclear localization signals, and introns.

[0197] As used herein, the term “operably linked” refers to positioning of a regulatory region and a sequence to be transcribed in a nucleic acid so as to influence transcription or translation of such a sequence. For example, to bring a coding sequence under the control of a promoter, the translation initiation site of the translational reading frame of the polypeptide is typically positioned between one and about fifty nucleotides downstream of the promoter. A promoter can, however, be positioned as much as about 5,000 nucleotides upstream of the translation initiation site or about 2,000 nucleotides upstream of the transcription start site. A promoter typically comprises at least a core (basal) promoter. A promoter also

may include at least one control element, such as an enhancer sequence, an upstream element or an upstream activation region (UAR). The choice of promoters to be included depends upon several factors, including, but not limited to, efficiency, selectability, inducibility, desired expression level, and cell- or tissue-preferential expression. It is a routine matter for one of skill in the art to modulate the expression of a coding sequence by appropriately selecting and positioning promoters and other regulatory regions relative to the coding sequence.

[0198] A vector comprising an AS-SPIK or NS-SPIK nucleic acid sequence can be formulated in such a way as to promote uptake by a cell, i.e., a prokaryotic or eukaryotic cell, for example, a mammalian cell. Useful vector systems and formulations are described above. In some embodiments the vector can deliver the compositions to a specific cell type. The invention is not so limited however, and other methods of DNA delivery such as chemical transfection, using, for example calcium phosphate, DEAE dextran, liposomes, lipoplexes, surfactants, and perfluoro chemical liquids are also contemplated, as are physical delivery methods, such as electroporation, micro injection, ballistic particles, and “gene gun” systems. In some embodiments, the polynucleotides of the invention may also be used with a microdelivery vehicle such as cationic liposomes, other lipid-containing complexes, and other macromolecular complexes capable of mediating delivery of a polynucleotide to a host cell. Another delivery method is to use single stranded DNA producing vectors which can produce the expressed products intracellularly.

[0199] It is another aspect of the present invention to provide methods of producing the antibodies described herein. Although antibodies can be prepared by chemical synthesis, they are typically produced by methods of recombinant DNA technology, such as co-expression of all the chains making up the protein in a single recombinant host cell, or co-expression of a heavy chain polypeptide and an antibody, e.g., a human antibody. In addition, the antibody heavy and light chains can also be expressed using a single polycistronic expression vector. Purification of individual polypeptides is achieved using standard protein purification technologies such as affinity (protein A) chromatography, size exclusion chromatography and/or hydrophobic interaction chromatography.

[0200] It is another aspect of the present invention to provide pharmaceutical compositions comprising one or more proteins of the present invention in admixture with a suitable pharmaceutically acceptable carrier. Pharmaceutically acceptable carriers as used herein are exemplified, but not limited to, adjuvants, solid carriers, water, buffers, or other carriers used in the art to hold therapeutic components, or combinations thereof.

[0201] Therapeutic formulations of the proteins (e.g., antibodies) used in accordance with the present invention are prepared for storage by mixing proteins having the desired degree of purity with optional pharmaceutically acceptable carriers, excipients or stabilizers (see, e.g. Remington's Pharmaceutical

Sciences 16th edition, Osol, A. Ed. (1980)), such as in the form of lyophilized formulations or aqueous solutions. Acceptable carriers, excipients, or stabilizers are nontoxic to recipients at the dosages and concentrations employed, and include buffers such as phosphate, citrate, and other organic acids; antioxidants including ascorbic acid and methionine; preservatives (such as octadecyldimethylbenzyl ammonium chloride; hexamethonium chloride; benzalkonium chloride, benzethonium chloride; phenol, butyl or benzyl alcohol; alkyl parabens such as methyl or propyl paraben; catechol; resorcinol; cyclohexanol; 3-pentanol; and m-cresol); low molecular weight (less than about 10 residues) polypeptides; proteins, such as serum albumin, gelatin, or immunoglobulins; hydrophilic polymers such as polyvinylpyrrolidone; amino acids such as glycine, glutamine, asparagine, histidine, arginine, or lysine; monosaccharides, disaccharides, and other carbohydrates including glucose, mannose, or dextrans; chelating agents such as EDTA; sugars such as sucrose, mannitol, trehalose or sorbitol; salt-forming counter-ions such as sodium; metal complexes (e.g. Zn- protein complexes); and/or non-ionic surfactants such as TWEEN™, PLURONICS™ or polyethylene glycol (PEG).

Methods of Use

[0202] The compositions disclosed herein are generally and variously useful for the diagnosis and/or treatment of disorders that are characterized by the expression of AS-SPIK. Such disorders include, but are not limited to, cancers, viral infections, and inflammatory disorders. One prominent example is liver cancer. Other non-limiting examples include those cancers described herein in connection with the definition of the term “cancer”. Accordingly, aspects of the invention involve methods for diagnosing and/or treating a cancer (e.g., a liver cancer) in a subject having a said cancer, or who is at risk for developing said cancer. The terms “subject”, “patient”, and “individual” are used interchangeably herein.

[0203] In some embodiments, the methods involve contacting a biological test sample from a subject with an AS-SPIK antibody or antigen-binding fragment to generate an AS-SPIK-antibody complex; detecting a concentration of the AS-SPIK-antibody complex in the biological test sample; and comparing the concentration of the AS-SPIK-antibody complex to a reference value to determine whether the subject has or is at risk of developing the disorder. In certain embodiments, the methods comprise contacting a biological test sample with a first antibody or antigen-binding fragment that binds to SPIK to generate a SPIK-antibody complex; contacting the SPIK-antibody complex with an AS-SPIK antibody or antigen-binding fragment to generate an AS-SPIK-antibody complex in the biological test sample; and comparing the concentration of the AS-SPIK-antibody complex to a reference value to determine whether the subject has or is at risk of developing the disorder. Several non-limiting examples of antibodies that can be utilized in such methods are described herein.

[0204] In some embodiments, the methods involve administering a therapeutically effective amount of an antibody, or antigen-binding fragment thereof, or antibody-drug conjugate, as described herein, to a patient suffering from a disease or disorder characterized by the expression of AS-SPIK.

Liver cancer

[0205] One prominent example of a disorder that is characterized by expression of AS-SPIK is liver cancer. Liver cancer encompasses a wide range of conditions that result in damage to the liver or impaired liver function. Liver cancer can result, for example, from infectious agents, disease, trauma, or genetic conditions or a combination of infectious agents, disease, trauma, and genetic conditions.

[0206] Liver cancer can include diseases which involve abnormal cell growth, such as primary liver cancer, for example, hepatocellular carcinoma, cholangiocarcinoma, angiosarcoma, and hepatoblastoma. Such cancers can include cancers at any stage of disease progression, such as HCC from very early stages (Barcelona Clinic Liver Cancer system (BCLC) stages 0 and tumor size < 2 cm), early stages (BCLC stage A, tumor size between 2cm and 5 cm), middle stages (BCLC stage B, intermediate tumor size > 5cm), late stages (BCLC stage C and D, advanced stage), or metastatic stages, (Pons et al., *HPB* 2005;7(1):35-41 and ICC from ICC early stages (Stage I, II and IIIa, tumor size < 2cm), middle stages (Stage IIIb and IIIc, tumor size $\geq$ 2cm), and late stage (stage IV) (Farges et al., *Cancer* 2011;117(10):2170-2177).

[0207] Liver cancer can also be induced by infectious diseases caused by viruses, such as Hepatitis B, Hepatitis C, and Hepatitis D. Regardless of the specific hepatitis virus, such infections can be either acute or chronic.

[0208] Liver cancer can also arise from liver damage, for example, liver cirrhosis. Cirrhosis, a late stage scarring or fibrosis of the liver, can be caused by many forms of liver diseases and conditions. Cirrhosis can occur as the result of genetic conditions, for example hemochromatosis, cystic fibrosis, Wilson's disease, and autoimmune disorders. Cirrhosis can also arise from hepatitis viral infections and alcohol consumption.

[0209] Liver cancer also can be caused by other diseases including, but are not limited to alcoholic liver disease, disorders related to abnormal fat content in the liver such as fatty liver, non-alcoholic fatty liver disease, non-alcoholic steatosis, and liver fibrosis.

Biological samples

[0210] A "biological sample", "test sample" or "sample" refers to a sample obtained or derived from a patient. The sample can be, for example, a body fluid sample. Exemplary body fluid samples include

blood, serum, plasma, urine, saliva, semen, stool, sputum, cerebral spinal fluid, tears, mucus, amniotic fluid, or any combination thereof. In some embodiments, a biological sample can be a tissue sample. Exemplary tissue samples include a biopsy specimen, such as a liver biopsy specimen, or a primary cell culture specimen prepared from a patient's cells, or supernatant from the primary culture.

Immunoassays

[0211] Aspects of the invention include diagnostic assay methods, e.g., diagnostic immunoassays, which can be used to detect the presence or absence of AS-SPIK in a test sample. The immunoassay format used for the detection of AS-SPIK can be configured in a variety of ways. The immunoassays can include both homogeneous and heterogeneous assays, competitive and non-competitive assays, direct and indirect assays, and "sandwich" assays. Useful formats include, but are not limited to, enzyme immunoassays, for example, enzyme linked immunosorbent assays (ELISA), chemiluminescent immune-assays (CLIA), electrochemiluminescent assays, radioimmunoassay, immunofluorescence, fluorescence anisotropy, immunoprecipitation, equilibrium dialysis, immunodiffusion, immunoblotting, agglutination, luminescent proximity assays, and nephelometry.

[0212] Regardless of the format, the biological sample is contacted with an anti-AS-SPIK antibody of the present invention. In some embodiments, the biological sample can be immobilized on a solid support. In some embodiments, the biological sample is contacted with an anti-SPIK antibody of the invention that has been immobilized on a solid support. The solid support can be, for example, a plastic surface, a glass surface, a paper or fibrous surface, or the surface of a particle. More specifically, the support can include a microplate, a bead, a polyvinylidene difluoride (PVDF) membrane, nitrocellulose membrane, nylon membrane, porous membranes, non-porous membranes. The composition of the substrate can be varied. For example, substrates or support can comprise glass, cellulose-based materials, thermoplastic polymers, such as polyethylene, polypropylene, or polyester, sintered structures composed of particulate materials (e.g., glass or various thermoplastic polymers), or cast membrane film composed of nitrocellulose, nylon, or polysulfone. In general embodiments, the substrate may be any surface or support upon which an antibody or a polypeptide can be immobilized, including one or more of a solid support (e.g., glass such as a glass slide or a coated plate, silica, plastic or derivatized plastic, paramagnetic or non-magnetic metal), a semi-solid support (e.g., a polymeric material, a gel, agarose, or other matrix), and/or a porous support (e.g., a filter, a nylon or nitrocellulose membrane or other membrane). In some embodiments, synthetic polymers can be used as a substrate, including, e.g., polystyrene, polypropylene, polyglycidylmethacrylate, aminated or carboxylated polystyrenes, polyacrylamides, polyamides, and polyvinylchlorides.

[0213] In some embodiments, the immunoassay format can be a two antibody “sandwich” assay. The biological sample is contacted with an anti-SPIK antibody of the invention that has been immobilized on a solid support, for example, microtiter plate. The sample and the first antibody are incubated under conditions that favor specific binding and the formation of a SPIK-antibody complex. Following the contacting step, unbound constituents of the biological sample are removed. Then, the complex is contacted with a second anti-SPIK antibody. The second antibody binds to a different SPIK epitope than the epitope bound by the first antibody. Thus, the first and second antibodies do not competitively inhibit one another for binding to SPIK. In some embodiments, the first antibody can recognize an epitope, i.e., an antigenic determinant, present on both AS-SPIK and NS-SPIK. We may refer to such an antibody as a “pan-SPIK” antibody. Alternatively, the first antibody can recognize an epitope present only on AS-SPIK. In some embodiments, the second antibody can recognize an epitope, i.e., an antigenic determinant, present on both AS-SPIK and NS-SPIK. Alternatively, the second antibody can recognize an epitope present only on AS-SPIK or NS-SPIK. Thus, the sandwich assay can be configured such that the first antibody is a pan-SPIK antibody and the second antibody specifically or preferentially binds to AS-SPIK and does not specifically bind to NS-SPIK. Alternatively, the sandwich assay can be configured such that both the first and second antibodies specifically or preferentially bind to AS-SPIK and do not specifically bind to NS-SPIK.

[0214] Antibody binding can be measured in a variety of ways. The signal, for example, generated by a detectable label, can be analyzed and, if applicable, quantified using an optical scanner or other image acquisition device and software that permits the measurement of the signal, for example a fluorescent signal a luminescent signal, or a phosphorescent signal, or a radioactive signal, associated with complex formation. Exemplary instrumentation for measuring a detectable signal can include, but is not limited to microplate readers, fluorimeters, spectrophotometers, and gamma counters.

Reference samples

[0215] The level of AS-SPIK in a biological sample can be compared with that of a reference sample. Standard reference levels typically represent the average AS-SPIK levels derived from a population of individuals. The reference population may include individuals of similar age, body size, ethnic background or general health as the individual in question. Thus, the AS-SPIK levels in a patient's sample can be compared to values derived from: 1) individuals who are known to have a liver cancer and who express AS-SPIK and whose bodily fluids contain AS-SPIK; 2) individuals who do not have a liver cancer and whose bodily fluids contain low levels of AS-SPIK.

[0216] In general, an elevated level of AS-SPIK can be any level of AS-SPIK that is greater, preferably at least 1, 2, 3, 4 or 5% greater, more preferably at least 5% greater, than either the level of AS-SPIK found in a control sample or greater than the average level of AS-SPIK found in samples from a population of normal healthy individuals who do not have a liver cancer (reference value). A reduced level of AS-SPIK can be any level of AS-SPIK that is less than either the level of AS-SPIK found in a control sample or less than the average level of AS-SPIK found in samples from a population of individuals having a liver cancer. Any population size can be used to determine the average level of AS-SPIK found in samples from a population of normal healthy individuals. For example, a population of between 2 and 250, e.g., 2, 3, 4, 5, 10, 15, 20, 25, 30, 40, 50, 100, 150, 200, 250 or more individuals can be used to determine the average level of AS-SPIK in samples from a population of normal healthy individuals, with greater accuracy in the measurement coming from larger sample populations.

[0217] In some embodiments, a reference chart can be used to determine whether or not a particular level of AS-SPIK in a sample is elevated relative to a control sample or a larger population. For example, a reference chart can contain the normal range of AS-SPIK found in healthy individuals of the same age, ethnic background or general health as the individual in question. Using this reference chart, any level of AS-SPIK measured in a sample can be classified as being low, normal, or elevated relative to a control sample or relative to an average value derived from a larger population. The term “elevated level” is defined as a level, which is higher, preferably at least 2% higher, more preferably at least 5% higher, than a reference level.

[0218] Alternatively, or in addition, the level of AS-SPIK in a biological sample can be “normalized” against the level of one or more additional biological markers, for example another marker whose expression is independent of AS-SPIK expression. That is, the levels of the additional marker can be evaluated in parallel with those of AS-SPIK, either at the same time or on a separate occasion. The additional marker can serve as an internal control for sample preparation, handling and storage as well as day-to-day assay variability. The values for the level AS-SPIK and the additional marker may be expressed as a ratio and the ratio may be compared to similar ratio obtained for a reference sample or population. A useful second marker can be alpha-fetoprotein.

[0219] In some embodiments, the methods can include the use of a standard reference set. The reference set can include one or more samples of a purified SPIK polypeptide or fragment thereof. When multiple samples are used, these can be of different concentrations. In one embodiment, the reference set can include 6 samples of recombinant AS-SPIK at concentrations of 50ng/ml, 30ng/ml, 8ng/ml, 3ng/ml, 1ng/ml and 0 ng/ml of AS-SPIK. The recombinant AS-SPIK can be purified with affinity chromatography (HPLC) using either anti-AS-SPIK antibody such as IM-CA22 or anti-tag antibodies. The reference value

in blood or other body fluids can vary. However, the skilled person is in a position to determine the average level of AS-SPIK in the different body fluids of the respective populations and to determine a respective reference value, which assures that the level of AS-SPIK in patients having the liver cancer to be determined is well above the reference value, whereas the level of patients not suffering from a liver cancer to be detected or of healthy individuals is well below the respective reference value. In a preferred embodiment, the reference value is about 5%, more preferably about 7%, even more preferably about 10%, higher than the average level of AS-SPIK found in samples from a population of normal healthy individuals. It is noted that the levels of AS-SPIK in the biological sample and in the control sample are to be determined via the same method, so that comparability is given. The absolute values of e.g. AS-SPIK levels can be determined via calibration curves using recombinant AS-SPIK as described above.

Control Samples

[0220] In some embodiments, a positive control can include a sample of AS-SPIK produced by a eukaryotic cell or cell line. For example, a useful control can be medium containing 100 ng/ml of AS-SPIK from a stable cell line S2-3. This was created by the inventors by inserting the DNA sequence of AS-SPIK into the chromosomes of the HCC cells under the control of an artificial promotor which over-expressed AS-SPIK.

[0221] Methods disclosed herein are useful in the detection of a liver cancer in a patient suspected of having or at risk for a liver cancer. The methods can also be used in the analysis of samples from a patient who has been treated for a liver cancer, for example, hepatocellular carcinoma, in order to determine whether the patient is at risk for experiencing a remission of hepatocellular carcinoma. The methods can also be used for monitoring the course of the treatment, for example treatment with a therapeutic agent such as a small molecule drug or therapeutic antibody, chemotherapy, radiation therapy or surgery, to determine efficacy of the treatment and to allow to managing clinician to alter the treatment if needed. The methods may also be used in the detection, monitoring, or analysis of a patient suffering from or at risk for any disorder that is associated with a modulation, for example an increase, in the level of AS-SPIK in a biological sample, for example, a blood or serum sample, obtained from the patient.

[0222] The methods disclosed herein can be used in conjunction with other standard diagnostic methods, for example serological analyses of liver enzymes or alpha-fetoprotein, ultrasound (sonography), computed tomography (CT scan), magnetic resonance imaging (MRI), angiography, laparoscopy, or biopsy.

Articles of manufacture

[0223] The compositions described herein can be packaged in suitable containers labeled, for example, for use in the detection, identification, and quantification of AS-SPIK in a biological sample. The articles of manufacture, also referred to as “kits”, may include antibodies, antigen-binding fragments of antibodies, and/or antibody-drug conjugates of the present invention, media, purified samples of antigen for use as positive controls, or any combination thereof. The containers included in the kits can include a composition comprising an antibody of the present invention that specifically or preferentially binds to AS-SPIK but not to NS-SPIK. A kit can also include an antibody that binds to both AS-SPIK and NS-SPIK. Suitable buffers for diluting or reconstituting test samples and antibodies may also be provided. Some of the components may be provided in dry form, and may require reconstitution. The anti-SPIK antibody can be pre-bound to an assay device, for example, a microplate. Thus, in one embodiment, a kit for the detection, identification and quantification of AS-SPIK comprises an anti-AS-SPIK antibody and a pan-SPIK antibody. The kit may optionally comprise a detectable label.

[0224] Accordingly, packaged products (e.g., sterile containers containing one or more of the compositions described herein and packaged for storage, shipment, or sale at concentrated or ready-to-use concentrations) and kits, including at least one composition of the invention, e.g., an anti-AS-SPIK antibody, are also within the scope of the invention. A product can include a container (e.g., a vial, jar, bottle, bag, or the like) containing one or more compositions of the invention. In addition, an article of manufacture further may include, for example, packaging materials, instructions for use, syringes, delivery devices, buffers or other control reagents for treating or monitoring the condition for which diagnosis or treatment is required.

[0225] Reagents for particular types of assays can also be provided in kits of the invention. Thus, the kits can include a population of beads (e.g., suitable for an agglutination assay or a lateral flow assay), or a plate (e.g., a plate suitable for an ELISA assay). In other embodiments, the kits comprise a device, such as a lateral flow immunoassay device, an analytical rotor, or an electrochemical, optical, or optoelectronic sensor. The population of beads, the plate, and the devices are useful for performing an immunoassay. For example, they can be useful for detecting formation of a first agent-analyte-second agent complex.

[0226] In addition, the kits can include various diluents and buffers, labeled conjugates or other agents for the detection of specifically bound antigens or antibodies, and other signal-generating reagents, such as enzyme substrates, cofactors and chromogens. The kits can include one or more reference samples of varying concentrations, for example, purified recombinant AS-SPIK. The kits can also include a positive control, for example a cell supernatant from a cell line that over expresses AS-SPIK. Other components

of a kit can include coating reagents, polyclonal or monoclonal capture antibodies specific for an antigen or analyte to be tested, or a cocktail of two or more of the antibodies, purified or semi-purified extracts of these antigens as standards, monoclonal antibody detector antibodies, an anti-mouse, anti-dog, anti-chicken, or anti-human antibody with indicator molecule conjugated thereto, indicator charts for colorimetric comparisons, disposable gloves, decontamination instructions, applicator sticks or containers, a sample preparatory cup, etc. In one embodiment, a kit comprises buffers or other reagents appropriate for constituting a reaction medium allowing the formation of a peptide-antibody complex.

[0227] Such kits provide a convenient, efficient way for a clinician to determine whether subject has or is at risk for a liver cancer. Thus, in certain embodiments, the kits further comprise instructions for use. The product may also include a legend (e.g., a printed label or insert or other medium describing the product's use (e.g., an audio- or videotape)). The legend can be associated with the container (e.g., affixed to the container) and can describe the manner in which the assay should be performed, indications therefor, and other uses.

[0228] The following examples are offered for illustrative purposes only, and are not intended to limit the scope of the present invention in any way. While several embodiments have been provided in the present disclosure, it should be understood that the disclosed compositions and methods might be embodied in many other specific forms without departing from the spirit or scope of the present disclosure. The present examples are to be considered as illustrative and not restrictive, and the intention is not to be limited to the details given herein. Various examples of changes, substitutions, and alterations are ascertainable by one skilled in the art and could be made without departing from the spirit and scope disclosed herein.

EXAMPLES

Example 1. AS-SPIK is larger than NS-SPIK

[0229] Serine protease Inhibitor Kazal (SPIK/SPINK1) is a small secreted protein with 79 amino acids (4). Normally, the secreted SPIK protein is shorter than genetic SPIK due to the removal of 23 amino acids in its N-terminus, which is believed to act as a signal peptide, resulting in a secreted protein that contains only 56 residues (14, 15). However, in liver cancer cells, AS-SPIK retains this additional fragment and is therefore larger than NS-SPIK. This compositional difference between AS-SPIK and NS-SPIK is confirmed by size analysis (gel electrophoresis) and Edman Degradation-based protein sequencing. Briefly, AS-SPIK was purified from the media of S2-3 cells, a cell line we constructed that expresses high levels of AS-SPIK (16), and NS-SPIK was purified from the media of pancreatic cells using HPLC. 1µg of each

protein was run on a 5-15% gradient SDS-PAGE gel (Invitrogen, Carlsbad, CA). After transfer to a PVDF membrane, proteins were visualized by Coomassie Blue staining. FIG. 1, which captures the results from this testing, shows that the size of NS-SPIK produced by pancreatic cells was around 6.5KD, which was consistent with the published sequence data (SEQ ID NO: 4), confirming that the first 23 amino acids in NS-SPIK are removed during secretion (14, 17). In contrast, the size of AS-SPIK was approximately 10 KD and larger than NS-SPIK, which is again consistent with the size of the full-length SPIK protein (SEQ ID NO: 2).

[0230] To determine the exact sequence of AS-SPIK, the AS-SPIK bands from the above gel electrophoresis were cut from the membrane. Alphalyse Inc. (Palo Alto, CA) performed the Edman degradation sequence analysis, with the results shown in FIG. 2. Here, the sequence predicted by Edman degradation in the N-terminus of S2-3 cells-secreted-SPIK is shown in red. The Edman degradation data shows that the N-terminus of AS-SPIK matched the sequence of residues 2-6 of SPIK (Excluding the first Methionine of the start codon) (see FIG. 2), suggesting that the full 23 amino acid sequence in the N-terminus of SPIK (starting with (M)K), which is removed in NS-SPIK, was retained in AS-SPIK after secretion. The full amino acid sequences of both NS-SPIK and AS-SPIK are shown in FIG. 3, highlighting this difference.

Example 2. The conformation of AS-SPIK is different from that of NS-SPIK

[0231] The inclusion of these additional 23 residues in the N-terminus of AS-SPIK in turn changes the conformation of the protein. This change is visualized by comparing the 3D structure of AS-SPIK and NS-SPIK. The crystal structure of AS-SPIK was determined by CLISP protein epitope mapping study (Pepscan, Lelystad, Netherlands) and the crystal structure of NS-SPIK was taken from a peer-reviewed publication by Hecht et al (18). Both structures are shown side by side in FIG 4. Through a visual comparison, three conformational differences between AS-SPIK and NS-SPIK can be identified, which are outlined in red. Box I shows the N-terminus of both NS-SPIK and AS-SPIK. The extra 23-residue fragment in AS-SPIK projects outwards and extends past the main body of the protein; in contrast, the N-terminus of NS-SPIK is does not have an additional fragment which protrudes from the main body of the protein. This exposed fragment greatly increases the likelihood of other proteins, such as antibodies, of selectively interacting with AS-SPIK and not NS-SPIK. Box II shows that, due to the longer N-terminus of AS-SPIK, the first loop in AS-SPIK is flatter and angled differently compared to the corresponding loop in NS-SPIK. This difference leads to more space between the first loop and alpha helix (FIG. 4, box II) in AS-SPIK, which exposes amino acids that are on the interior and inaccessible in NS-SPIK. Finally, the longer N-terminus of AS-SPIK also changes the relative position and distance between the N-terminus and alpha-helix of the

protein. The low position of the N-terminus relative to the alpha helix in AS-SPIK is highlighted in crystal structure in box III. These conformational changes and differences in tertiary structure suggest that different conformational antibodies could be generated to specifically target either form of SPIK.

Example 3. Development of antibodies that selectively bind to AS-SPIK, but not NS-SPIK

[0232] Due to the differences between AS-SPIK and NS-SPIK described in above, it is possible to develop antibodies that solely recognize AS-SPIK, but not NS-SPIK. As proof of this, we have developed approximately 20 monoclonal antibodies which only bind AS-SPIK, but not NS-SPIK, in both mice and rabbits. At the same time, we have developed a polyclonal antibody from sheep, which binds to both AS-SPIK and NS-SPIK. Briefly, mice and rabbits were immunized with a series of recombinant proteins consisted of 1) a Tag 2) a linker sequence, 3) a sequence of varying length that is a subset of SEQ ID NO: 6, and 4) the common region of AS-SPIK and NS-SPIK (SEQ ID NO: 4). To develop the monoclonal antibodies, the animals were injected with the recombinant proteins with the sequence described above. The blood was tested after 4 injections, and single clones were established for samples that tested positive. In order to identify antibodies that only bind to AS-SPIK, but not NS-SPIK, clones were screened by ELISA test using plates coated with partially purified AS-SPIK and NS-SPIK. Those that tested positive to AS-SPIK but negative to NS-SPIK were then selected. The cell lines with the highest binding affinity to AS-SPIK were expanded and hybridomas were established. The resulting antibodies were then purified from the cell culture medium using a protein G affinity column. To generate polyclonal antibodies in sheep, the sheep was immunized with one of the recombinant proteins described above. After four injections, the serum was collected and the polyclonal antibody was purified from serum.

[0233] FIG. 5 shows the binding test results of 8 example antibodies from this group of 20 monoclonal antibodies: IM-A1, IM-B10, IM-C6, IM-E2, IM-CA22, IM-CA18, IM-CA46, IM-CA77 and IM-Poly S (polyclonal antibody from sheep) (the sequences of these 8 antibodies are provided Tables 3 and 4 (FIGS. 15 and 16), and are also provided in US Provisional Patent Application Serial No. 62/639,850, PCT Patent Application No. PCT/US19/20999 and US Provisional Patent Application Serial No. 62/871565, the disclosures of which applications are incorporated by reference herein in their entireties. FIG. 5 shows that all 8 of these example monoclonal antibodies, whether they were developed in mice or rabbits, strongly bind to AS-SPIK. In contrast, binding activity to NS-SPIK is at the background level, similar to negative control (FIG. 5, Neg control). As expected, the polyclonal sheep antibody strongly binds to both AS-SPIK and NS-SPIK, as polyclonal antibodies contain multiple different antibodies that bind to a variety of epitopes, including those that may be common across both NS-SPIK and AS-SPIK (FIG. 5, Poly S). This

data confirms that antibodies which selectively bind to AS-SPIK and not NS-SPIK can be developed in multiple animal models.

[0234] As a control, we simultaneously developed a series of monoclonal antibodies in mice, using a peptide containing the sequence ₃₆ELNGCTKIYDPV₄₇, which is a common sequence in both AS-SPIK and NS-SPIK (SEQ ID NO: 4) and is colored green in FIG. 4. This led to the production of antibody IM-BA2, which binds to both AS-SPIK and NS-SPIK (data not shown). However, we were unable to generate an antibody that binds specifically to AS-SPIK but not NS-SPIK using this method, suggesting that epitopes specific to AS-SPIK are conformational and not linear.

Example 4. The anti-AS-SPIK antibodies are conformation-dependent

[0235] In order to further support the observation that epitopes specific to AS-SPIK and not NS-SPIK are conformation-dependent, additional tests to evaluate antibody binding to various synthesized peptides was completed. Four peptides, each containing sequences from different regions of AS-SPIK were synthesized by BioMatik (Wilmington DE). Peptide A contains the entire sequence of AS-SPIK, but without the formation of any disulfide bonds via the inactivation of cysteine side-groups. Peptide B contains the sequence of AS-SPIK fragment from M₁-G₅₀, Peptide C contains the sequence of AS-SPIK fragment from D₂₃-G₅₀ and Peptide D contains the sequence of AS-SPIK fragment from N₅₁-C₇₉. The binding activity of our monoclonal antibodies to AS-SPIK and the other peptides was tested by ELISA and results for IM-CA22 are shown in FIG. 6. Similar results were obtained from all other monoclonal anti-AS-SPIK antibodies we have developed, including IM-CA18, IM-CA46, IM-CA77, IM-A1, IM-A6, IM-B10, IM-C6, IM-D3, IM-D5, IM-E2, IM-F5, IM-G6 and IM-G7. Briefly, a 96-well plate was coated with Native AS-SPIK and different peptides, the plate then reacted with anti-AS-SPIK antibody (IM-CA22 herein) labeled with HRP. The OD_{450nm} was determined after incubating the plate with the substrate, TMB. FIG. 6 shows that antibody IM-CA22 only recognizes native protein, but does not recognize any of the synthesized peptides, including Peptide A, which contains the entire native AS-SPIK sequence. This result further supports the observation that anti-AS-SPIK antibodies are conformation-dependent antibodies. In contrast, our polyclonal antibody Poly S recognizes all synthesized peptides as well as native AS-SPIK (FIG. 6, Poly S), suggesting there are both linear epitope antibodies as well as conformation-dependent epitope antibodies in Poly S. However, the differences in conformation that we identified above, in conjunction with these test results, strongly demonstrate that the monoclonal antibodies described herein have conformation-dependent epitopes.

Example 5. Class I and Class II anti-AS-SPIK antibodies

[0236] In order to identify possible binding epitopes of anti-AS-SPIK antibodies, we established an ELISA test system to examine if all of the anti-AS-SPIK antibodies that we developed bind to the same epitope. Briefly, we coated a plate with one antibody, and then incubated the plate with native AS-SPIK from S2-3 cell. After the AS-SPIK is captured on the plate, the second anti-AS-SPIK labelled with HRP was added, followed by a substrate. The OD value was determined after the color was developed. If a negative result was obtained, this suggests that the capture and signal antibodies bind to the same epitope or to epitopes close enough that binding to one inhibits binding to the other. In contrast, if a positive result was obtained, it suggests these two antibodies bind to different epitopes, which are far enough apart so that both can be accessed by different antibodies simultaneously. Poly S was used as a control. Our data demonstrate that all of the anti-AS-SPIK antibodies we have developed can be divided into two classes: Class I antibodies that include IM-CA22, IM-A1, IM-B10, IM-CA18, IM-D2, IM-D3, IM-D5 and IM-G2, and Class II antibodies that include IM-E2, IM-C6, IM-CA46, IM-CA77, IM-A6, IM-B3, IM-F5 and IM-G6. The defining feature of these antibody classes is that any antibody in Class I can work with any antibody in class II, and vice versa. This means that they give positive results in the ELISA test described above, and can function well as an antibody pair in a sandwich ELISA. Our data also demonstrate that all of these antibodies bind to one or two different conformational epitopes. Table 1 (FIG. 13) shows the results from these competition ELISA tests, using 4 Class I antibodies: IM-A1, IM-B10, IM-CA22 and IM-CA18, and 4 Class II antibodies: IM-C6, IM-E2, IM-CA46 and IM-CA77. Briefly, the plate was coated with each antibody from Class II at 100ng/ml and then reacted with AS-SPIK. After washing, a signal antibody (labeled with HRP) from Class I was added, and the binding activity was determined. The results of this are shown in Table 1A (FIG. 13). These tests were then repeated using antibodies within the same class for both the capture antibody and the signal antibody, with the results of Class I-Class I pairs shown in Table 1B (FIG. 13) and the results of Class II-Class II pairs shown in Table 1C (FIG. 13). For all cells, the listed concentrations represent the lowest concentration of signal antibody that still gives a positive result, with a lower concentration suggesting a stronger binding affinity. Based on these results, it is clear that all antibodies in Class I can work with any antibodies in Class II, while antibodies in the same class almost completely inhibit target binding, even when the signal antibody is present in very high concentrations (500ng/ml). Finally, and unsurprisingly, all antibodies listed here can work with our polyclonal antibody Poly S, suggesting that the test system is very efficient.

Example 6. Epitope I and epitope II of AS-SPIK

[0237] With the understanding that antibodies within the same class likely bind to very similar epitopes, the next step is to determine the exact binding sites for Class I and Class II anti-AS-SPIK antibodies. To achieve this, we performed epitope mapping using 8 monoclonal anti-AS-SPIK antibodies, which includes 4 Class I antibodies (IM-CA22, IM-CA18, IM-A1 and IM-B10) and 4 Class II antibodies (IM-CA46, IM-CA77, IM-C6 and IM-E2). IM-CA22, IM-CA18, IMCA46 and IMCA77 were generated in mice and IM-A1, IM-B10, IM-C6 and IM-E2 were generated in rabbits. The epitope mapping was also performed on Poly S, a polyclonal antibody from sheep. The Precision Epitope Mapping with CLIPS (Chemically Linked Peptides on Scaffolds) Peptide Arrays (19) was done by Pepscan (Lelystad, Netherlands).

[0238] CLIPS technology structurally fixes peptides into defined 3D structures. The CLIPS reaction takes place between bromo groups of the CLIPS scaffold and thiol sidechains of cysteines introduced into peptide constructs. The reaction is ultra-fast, very specific and is performed under mild conditions. Using this elegant chemistry, native protein sequences are transformed into CLIPS constructs with a range of structures. CLIPS technology is now routinely used to shape peptide libraries into single, double or triple looped structures as well as sheet- and helix-like folds, which allows mimicking conformational and discontinuous binding sites. The sequence dependent, conformational dependent and discontinuous conformational epitope thus can be determined.

[0239] An array of more than 2,400 independent peptides was synthesized and the binding of monoclonal antibodies IM-CA18, IM-CA22, IM-CA46, IM-CA77, IM-A1, IM-B10, IM-C6, IM-E2 and polyclonal antibody Poly S was tested and analyzed. Results of this epitope mapping are visualized using bar plots, heatmaps, tables, and on the structure modeled in Swiss-model using PDB entry 1HPT.pdb. Possible binding regions within AS-SPIK for each antibody are predicted, including linear, conformation-dependent, and discontinuous conformation-dependent epitopes. The 3D structure (crystal model) of these AS-SPIK binding regions is also visualized.

[0240] Because AS-SPIK is a small protein with 79 amino acids, constructing the conformational dependent epitopes as determined by CLIPS requires only 4 antigenic regions. These regions are highlighted in FIG. 7. The first region is G₅-A₂₉, which includes two parts: part I (G₅-A₂₃), which is shown in light blue, only exists in AS-SPIK. Part II (D₂₄-A₂₉), which is shown in yellow, exists in both AS-SPIK and NS-SPIK (SEQ ID NO 4); the second region is K₃₁-C₄₇, shown in green; the third region is I₄₂-N₅₆, shown in blue; the fourth region is D₅₀-C₇₉, shown in grey. However, the results from CLIPS suggest that only regions 1, 2, and 4 are directly involved in the binding of AS-SPIK and anti-AS-SPIK antibodies. Table 2 (FIG. 14) shows the amino acids which comprise the possible binding regions for anti-AS-SPIK antibodies as predicted by CLIPS. The CLIPS study also shows that each antibody binds to at least two

separate regions, implying that all anti-AS-SPIK antibodies described herein bind to discontinuous, conformational epitopes. This is consistent with our previous observation that anti-AS-SPIK antibodies are exclusively conformational dependent antibodies.

Example 7. The critical residues of epitope I and epitope II

[0241] In addition to identifying the potential binding regions of AS-SPIK for both classes of antibodies, the CLIPS study also evaluates the relative importance of amino acids within each region to its binding affinity. Region 1 (7FLLSALALLSLSGNTGADSLGRE₂₉, SEQ ID NO: 7) and region 4 (58CVLCFENRKRQ₆₈, SEQ ID NO: 8) comprise the essential binding sites for all Class I antibodies. The most critical amino acids for binding functionality in region 1 are 14LLSL₁₇ (SEQ ID NO: 12), and to a lesser degree 24DS₂₅ (SEQ ID NO: 13). In region 4, the critical residues are 58CVLCF₂₆ (SEQ ID NO: 14). Together, these amino acids comprise a discontinuous conformational epitope for all Class I antibodies, called Epitope I (Table 2 (FIG. 14) and FIG. 7). The discontinuous nature of this epitope is further supported by an inhibition test we performed to confirm these findings (shown in FIG. 8). Here, we synthesized a short peptide with 9 amino acids containing 14LLSL₁₇, which can inhibit the binding between native AS-SPIK and Class I antibody in this region. However, the inhibition of binding is incomplete, suggesting that there is a distinct second region of AS-SPIK that is responsible for the binding. This is consistent with our hypothesis that the anti-AS-SPIK binds a discontinuous conformational dependent epitope which should involve at least two separate regions in AS-SPIK. The CLIPS study also shows that the amino acids in region 2 (36LNGCTKIYD₄₄, SEQ ID NO: 9) and region 4 (64NRKRQTSILIQ₇₅, SEQ ID NO: 10) comprise the essential binding sites for all Class II antibodies. The most critical amino acids for binding functionality in region 2 are 36LN₃₇ (SEQ ID NO: 15) and 42IY₄₃ (SEQ ID NO: 16). In region 4, the critical residues are 67RQ₆₈ (SEQ ID NO: 17) and 71IL₇₂ (SEQ ID NO: 18). Together, these amino acids comprise a discontinuous conformational epitope for all Class II antibodies, called Epitope II (Table 2 (FIG. 14) and FIG. 7). Results from the CLIPS study on binding epitopes for Poly S suggests that the dominant antibody in poly S can be categorized as a Class II anti-AS-SPIK antibody, because it binds to Epitope II, similar to other Class II antibodies.

Example 8. Consensus sequences within Class I and Class II antibody CDRs

[0242] Now that we have defined Epitopes I and II and identified examples of antibodies that bind to them, the next step is to identify structural similarities between antibodies within each class. The CDRs in 4 Class I antibodies (IM-A (rabbit), IM-B10 (rabbit), IM-CA22 (mouse) and IM-CA18 (mouse)) were compared and analyzed. Consensus sequences of their CDRs were determined using the software BioEdit (North

Carolina State University), and are presented in FIG. 9. Interestingly, although 2 antibodies here are from mice and another 2 antibodies are from rabbits, we found that at least one amino acid in their CDRs is conserved in all of the antibodies. The most conserved CDR in these 4 Class I antibodies is CDRL2. 4 of 7 (57%) amino acids in this CDR are identical. The conserved residues within each of these CDRs is a defining characteristic of this genus of antibodies (Class I). A similar study of Class II antibodies IM-C6 (rabbit), IM-E2 (rabbit), IM-CA46 (mouse) and IMCA77 (mouse) leads to equally important findings. FIG. 10 lists the CDR consensus sequences for these 4 Class II antibodies. The number of conserved amino acids in CDRs of Class II antibodies is higher than in Class I. We found that at least 2 amino acids in each of their CDRs are conserved. The most conserved CDR in these 4 Class II antibodies is CDRL1, with 7 of 11 (64%) amino acids in this CDR being identical. The conserved residues within each of these CDRs is a defining characteristic of this genus of antibodies (Class II).

Example 9. Clinical Evaluation

[0243] AS-SPIK test kits based on a sandwich ELISA (Enzyme-Linked Immunosorbent Assay) and utilizing a monoclonal antibody (IM-CA22), which solely binds to AS-SPIK, as the capture antibody and an HRP-conjugated polyclonal anti-AS-SPIK antibody (Poly S) as the signal antibody, were used for clinical evaluation. The mechanism of this kit is illustrated in FIG. 11.

Study design and sample population:

[0244] Serum samples were collected from a total of 512 unique study subjects in a prospective, blinded study, and were sourced through various research institutions under a study protocol approved by each site's Institutional Review Board (IRB). Informed patient consent was obtained for all study participants. Of the 512 samples, 164 were from HCC patients, who were positively diagnosed using biopsy, CT, and/or MRI. This also included 81 patients with early stage HCC specifically (BCLC Stage 0 and A). The remaining 348 subjects were part of various control groups, including cirrhosis, non-cirrhotic chronic HBV/HCV, pancreatitis, and healthy subjects.

[0245] Serum levels of AS-SPIK were quantified using an ELISA-based test kit, which utilizes the monoclonal antibody IM-CA22, whose amino acid sequence information is provided herein (SEQ ID NO: 75 and SEQ ID NO: 76). AFP, which is the most commonly used biomarker for HCC, was quantitatively determined for all patients by each research institution, using FDA-approved AFP tests in their certified clinical laboratories. Receiver operating curves (ROCs) were constructed for both AS-SPIK and AFP, and their sensitivity, specificity, and AUC (area under the curve) were compared. For this analysis, only the intended use population, those with liver disease, was considered (HCC,

cirrhosis, and HBV/HCV). Pancreatitis patients were evaluated separately to confirm that normal pancreatic SPIK (NS-SPIK) does not interfere with the AS-SPIK test, while healthy patients were used as a baseline negative control.

Results:

A). Serum AS-SPIK is significantly elevated in HCC patients, including early-stage HCC

[0246] The results demonstrated that the mean serum level of AS-SPIK in all HCC (45.2 ng/mL, 95% CI: 40.5 to 49.9) was significantly higher than in all control groups ($p < 0.001$) (FIG. 17; Table 5). For the 81 patients with early stage HCC (BCLC stage 0 and A), the mean concentration of serum AS-SPIK was 38.1 ng/mL (95% CI: 32.1 to 44.2), and was significantly different from the serum AS-SPIK levels in all control groups ($P < 0.001$) (FIG. 17; Table 5). These results are consistent with previous studies described in Examples 12 and 13, as well as FIGS. 15 and 16 of WO2019/173503, the disclosure of which is incorporated by reference herein in its entirety.

B). The performance of AS-SPIK and AFP in distinguishing HCC from other liver disease.

[0247] Overall, AS-SPIK demonstrated significantly better sensitivity and specificity than AFP and is shown in Table 6 (FIG. 18). The AUC for AS-SPIK in detecting HCC using liver disease patients as controls was 0.87 (95% CI: 0.83 to 0.91), compared to 0.70 (95% CI: 0.64 to 0.76) for AFP. Using 21.5 ng/mL as a cutoff value for serum AS-SPIK, the sensitivity and specificity of AS-SPIK were 80% and 90%, respectively. Comparatively, using the standard 20.0 ng/mL as a cutoff value of serum AFP, the sensitivity and specificity were only 52% and 86%, respectively, which is significantly lower than that of AS-SPIK ($P < 0.05$). For early stage HCC, the AUC for AS-SPIK was 0.84 (95% CI: 0.79 to 0.89), compared to only 0.61 (95% CI: 0.53 to 0.70) for AFP. Using 21.5 ng/mL as a cutoff value, AS-SPIK's sensitivity in detecting HCC in its early stages was 72% with 90% specificity, which is significantly higher than the 42% sensitivity and 86% specificity for AFP (FIG. 18; Table 6).

C). The test may be used for the prediction of HCC stage and monitoring/prognosis.

[0248] In this study, a correlation between AS-SPIK levels and the stage of cancer progression was observed (FIG. 19; Table 7). The mean level of AS-SPIK for patients in the early stage group was 38.1 ng/mL (95% CI: 32.1 - 44.1), while the mean level of AS-SPIK in the late stage group was 52.2 ng/mL (95% CI: 45.3 - 59.1). Comparing these values, we see a statistically significant difference between the

two groups ($P < 0.05$), showing that there is a correlation between progression of HCC by stage and increased levels of AS-SPIK (FIG. 19; Table 7).

[0249] This is further supported by the observation that the mean values of AS-SPIK were consistently higher for later and more advanced stages when divided by BCLC stage. Patients in the very early stage group (BCLC stage 0) had the lowest mean AS-SPIK value of only 33.7 ng/mL, while terminal Stage D patients had the highest mean AS-SPIK value of 65.6 ng/mL. Analysis based strictly on BCLC classification, however, led to over-stratification and uneven sample sizes, especially for hard to recruit groups such as the BCLC stage 0 (very early) and stage D (terminal) groups. Thus, the results were not statistically significant ($P > 0.05$) (FIG. 20; Table 8). These results suggest that AS-SPIK may have potential as a tool to monitor HCC progression.

D). AS-SPIK Detection Kit solely detected AS-SPIK, but not NS-SPIK in patients' serum

[0250] Because AS-SPIK is a liver-cancer specific isoform of SPIK, which contains an additional fragment in its N-terminus, we evaluated whether NS-SPIK (pancreatic SPIK) interferes with the AS-SPIK test described herein. FIG. 12, panel A, shows that two monoclonal antibodies, IM-CA22 (recognizes only AS-SPIK) and IM-BA2 (recognizes both AS-SPIK and NS-SPIK), were used to coat plates, and then reacted with either AS-SPIK or NS-SPIK. IM-CA22 and the AS-SPIK test kit only recognize AS-SPIK, while IM-BA2 detects and confirms the presence of both AS-and NS-SPIK. FIG. 12, panel B shows serum levels of AS-SPIK in the 24 pancreatitis patients from the clinical study, who are expected to have elevated levels of pancreatic SPIK. The data demonstrate that AS-SPIK levels in these patients (7.4 ng/mL) were similar to those of healthy patients (7.4 ng/mL, $P > 0.99$), and significantly lower than in HCC patients of the clinical study (45.2 ng/mL, $P < 0.001$). FIG. 12, panel C confirms the elevated serum levels of NS-SPIK in pancreatitis patients and the observation that it does not interfere with the AS-SPIK detection kit described herein.

Sequence Information:

Name:	Sequence:	SEQ ID NO:
DNA sequence of full-length AS-SPIK, 237 NT	ATGAAGGTAACAGGCATCTTTCTTCTCAGTGCCTTG GCCCTGTTGAGTCTATCTGGTAACACTGGAGCTGAC TCCCTGGGAAGAGAGGCCAAATGTTACAATGAACT TAATGGATGCACCAAGATATATGACCCTGTCTGTG GGACTGATGGAAATACTTATCCCAATGAATGCGTG TTATGTTTTGAAAATCGGAAACGCCAGACTTCTAT CCTCATTCAAAAATCTGGGCCTTGC	1

Name:	Sequence:	SEQ ID NO:
Protein sequence of full-length AS-SPIK, 79AA	MKVTGIFLLSALALLSLSGNTGADSLGREAKCYNELN GCTKIYDPVCGTDGNTYPNECVLCFENRKRQTSILIQKSGPC	2
DNA sequence of full-length NS-SPIK, 168 NT	GACTCCCTGGGAAGAGAGGCCAAATGTTACA ATGAACTTAATGGATGCACCAAGATATATGA CCCTGTCTGTGGGACTGATGGAAATACTTATC CCAATGAATGCGTGTTATGTTTTGAAAATCGG AAACGCCAGACTTCTATCCTCATTCAAAAATCT GGGCCTTGC	3
Protein sequence of full-length NS-SPIK, 56AA	DSLGREAKCYNELNGCTKIYDPVCGTDGNTYPN ECVLCFENRKRQTSILIQKSGPC	4
DNA sequence only existing in AS-SPIK, 69NT	ATGAAGGTAACAGGCATCTTTCTTCTCAGTGC CTTGGCCCTGTTGAGTCTATCTGGTAACACTG GAGCT	5
Protein sequence only existing in AS-SPIK, 23AA	MKVTGIFLLSALALLSLSGNTGA	6
First region of AS-SPIK composes epitope I	FLLSALALLSLSGNTGADSLGREA	7
Second region of AS-SPIK composes epitope I	CVLCFENRKRQ	8
First region of AS-SPIK composes epitope II	LNGCTKIYD	9
Second region of AS-SPIK composes epitope II	NRKRQTSILIQ	10

Name:	Sequence:	SEQ ID NO:
The sequence only existing in AS-SPIK but not in NS-SPIK in epitope I	FLLSALALLSLSGNTGA	11
The critical residues of epitope I	LLSL	12
The critical residues of epitope I	DS	13
The critical residues of epitope I	CVLCF	14
The critical residues of epitope II	LN	15
The critical residues of epitope II	IY	16
The critical residues of epitope II	RQ	17
The critical residues of epitope II	IL	18
CDRH1 A1	SSNAIS	19
CDRH1 B10	SSYGVS	20
CDRH1 CA22	GYTFTDYYIN	21
CDRH1 CA18	GFTFSRYAMS	22
CDRH2 A1	AIGSSGSTYYASWAKS	23
CDRH2 B10	SIWSGGTTDYASWAKS	24
CDRH2 CA22	WIYPGSGNPIYNENFKD	25
CDRH2 CA18	SISIGGTYTYYPDSVKD	26
CDRH3 A1	RWENIGYTNVRLDL	27
CDRH3 B10	RGGYDYGYSNI	28
CDRH3 CA22	EWGCAMDS	29
CDRH3 CA18	EDYGFDY	30
CDRL1 A1	QASQSISTALA	31
CDRL1 B10	QASESISSYLS	32

Name:	Sequence:	SEQ ID NO:
CDRL1 CA22	KSSQSLN-SGNQKNYLA	33
CDRL1 CA18	KASQDVSTAVA	34
CDRL2 A1	GASTLAS	35
CDRL2 B10	RASTLAS	36
CDRL2 CA22	GASTRES	37
CDRL2 CA18	WASTRHT	38
CDRL3 A1	QQGYSTSDVDNA	39
CDRL3 B10	QQGYSVSNVDNI	40
CDRL3 CA22	QSDYSHPYT	41
CDRL3 CA18	HQHYSTYT	42
CDRH1 C6	SYAISW	43
CDRH1 E2	SAYAISW	44
CDRH1 CA46	GYTFTSYWMQ	45
CDRH1 CA77	GYTFSSNWIE	46
CDRH2 C6	AINTYGGTYIASWAKS	47
CDRH2 E2	AINSGGSAYIANWAKS	48
CDRH2 CA46	AIYPGDGDTRYTQKFED	49
CDRH2 CA77	QIFPGRDTTNYNEKFKG	50
CDRH3 C6	RDFDS-DAYTSASGGMDP	51
CDRH3 E2	REDIY-DYGGAFDP	52
CDRH3 CA46	GANYANIRFAY	53
CDRH3 CA77	RQEESDYDGSSHLNYGMDY	54
CDRL1 C6	QASQSINNYLS	55
CDRL1 E2	QASQGI SSYLS	56
CDRL1 CA46	RASQDITNYLN	57
CDRL1 CA77	RASQEISGHLN	58
CDRL2 C6	RASTLAS	59
CDRL2 E2	AATTLVS	60
CDRL2 CA46	YTSRLHS	61
CDRL2 CA77	AASI-LDS	62
CDRL3 C6	QQGYTSNVDNV	63

Name:	Sequence:	SEQ ID NO:
CDRL3 E2	QQDYTTSNVDNT	64
CDRL3 CA46	QQGNTVPWT	65
CDRL3 CA77	LQYTDYPWT	66
CA-46 variable region heavy chain	QGHLLQSGAELARPGTSVKLSCKASGYTFTSYWMQWVK QRPQGQLEWIGAIYPGDGDTRYTQKFEDKATLTADKSSS TAYMQLSNLASEDSAYYYCARGANYANIRFAYWGQGT LTVSA	67
CA-46 variable region light chain	DIQMTQTTSSLSASLGDRVSISCRASQDITNYLNWYQQKP DGTVKLLIFYTSRLHSGVPSRFSGSGSGTNFSLTISNLEQ DIATYFCQQGNTVPWTFGGGKLEIK	68
CA-77 variable region heavy chain	QVQLQQSGAELMKPGASVKISCKATGYTFSSNWIEWIK QRPGHGLEWIGQIFPGRDITNYNEKFKGKATFTADTSS NTAYMQLSSLTSEDSAVYYCARRQEEFSDYYGSSHL NYGMDYWGQGTSTVSS	69
CA-77 variable region light chain	DIQMTQSPSSLSASLGERVSLTCRASQEISGHLSWLQK PDGTIKRLIYAASILDGVPKRFSGSRSGSDYSLTISNLE SEDFADYYCLQYTDYPWTFGGGKVEIK	70
C6 variable region heavy chain	LVAVLKGVQCQSVKESEGLFKPTDALTLTCTVS GFSLSSYAIWVRQAPGSGLEWIGAINTYGGTYA SWAKSRSTITRNTNENTVTLKMTSLTAADTATYFC ARDFDSDAYTSASGGMDWGPGLVTVSSGQPKAP SFFPLAPCCGDTPR	71
C6 variable region light chain	WLPGARCAYDMTQTPASVEVAVGGTVTIKCQASQ SINNYLSWYQQIPGQPPKLLIYRASTLASGVSSRFK SGSGTQFTLTISGVQCADAATYYCQQGYTSNVDNV FGGGTEVVVKGDPVAPTFLIFPPSAD	72
E2 variable region heavy chain	LVAVLKGVQCQSVKESEGLFKPTDTLTLTCTV SGFSLSAIYAIWVRQAPGNLEWIGAINSGGSA YYANWAKSRSTITRNTNLNTVTLKMTSLTAAD TATYFCAREDIYDYGGAFDPWGPGLVTVSTGQ PKLHH	73
E2 variable region light chain	WLPGARCAYDMTQTPASVEVTVGGTVTIKCQAS QGISSYLSWYQQKPGQPPKLLIYAATTLVSGVSSR FKGSGSGTQFTLTISGVECADEATYYCQQDYTTS NVDNTFGGGTEVVVKGDPVAPTFLIFPPSAD	74
CA-22 variable region heavy chain	QIQLQQSGPELVKPGTSVKLSCKASGYTFTDYYINWVK QRPQGQLEWIGWIYPGSGNPIYNENFKDKATLTVDTSS TTAYLQLSSLTSEDSAVYFCAREWGCAMDSWGQGT SVTVSSAKTTAP SVYPLAP	75
CA-22 variable region light chain	DIVMTQSPSSLSVSTGEKVTMSCKSSQSLNLSGNQKNY LAWYQQKPGQSPKLLIYGASTRESGVPDRFTGSGSGTE FTLTISVQAEDLAVYYCQSDYSHPYTFGGGKLEIK	76

Name:	Sequence:	SEQ ID NO:
CA-18 variable region heavy chain	MNFVLSLIFLALILKGVQCEVQLVESGGGLVKPG RSLKLSCAASGFTFSRYAMSWVRQTPEKRLEGV ASISIGGTYYTYPDSVKDRFTISRDNANTLYLQ MNSLRSEDAMYYCVREDYGFQDYWGQGTLLVTVSS	77
CA-18 variable region light chain	DIVMTQSHKFMSTSVGDRVSITCKASQDVSTAVA WYQQKPGQSPKLLIYWASTRHTGVPDRFTGSGSG TDYTLTISSVQAEDLALYYCHQHYSTYTFGGGTKLEIK	78
A1 variable region heavy chain	LVAVLKGVQCQSVKESEGGLFKPTDTLTLTC TVSGFSLSSNAISWVRQAPNGLEWIGAIGSS GSTYYASWAKSRSTVTRNTNLNTVTLKMTSL TAADTATYFCARWENIGYTNVRLDLWGQGT LVTVSSGQPKAPSVFPLAPCCGDTTPSS	79
A1 variable region light chain	WLPGARCAIDMTQTPASVEVAVGGTVTIKCQ ASQSISTALAWYQQKPGQPPKLLIYGASTLASG VSSRFKSGSGTQFTLTISGVECADEATYYCQ GYSTSDVDNAFGGGTEG	80
B10 variable region heavy chain	LVAVLKGVQCQSVKESEGGLFKPTDTLT LTCTVSGFSLSSYGVSWSVRQAPGKGLEW IGSIWSGGTTDYASWAKSRSTITRNTNEN TVTLKVTSLTAADTATYFCARGGYDYG ASNIWGPGLTVTVSSGQPKAPS	81
B10 variable region light chain	LWLPGARCAIDMTQTPASVEVAVGGTVTIK CQASESISYLSWYQQKPGQPPKLLIYRASTL ASGVPSRFSGSGSGTEFTLTISDGQCDDAATY YCQQGYSVSNVDNIFGGGTEVVVKGDPVAPT VLIFPPSAD	82

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