SOLUBLE HYALURONIDASE GLYCOPROTEIN (sHASEGP), PROCESS FOR PREPARING THE SAME, USES AND PHARMACEUTICAL COMPOSITIONS COMPRISING THEREOF

CROSS REFERENCE TO RELATED APPLICATION(S)

[0001] This application claims priority under 35 U.S.C.§119(e) to U.S. Serial Number 60/452,360, filed March 5, 2003, which is herein incorporated by reference in its entirety.

BACKGROUND OF THE INVENTION

FIELD OF THE INVENTION

[0002] The present invention is generally related to Neutral-Active, Soluble Hyaluronidase Glycoproteins (sHASEGP), portions thereof, particularly Hyaluronidase domains. More specifically, the invention is related to chemical modifications, pharmaceutical compositions, expression plasmids, methods for manufacture and therapeutic methods using the Hyaluronidase Glycoproteins and domains thereof and the encoding nucleic acid molecules for the therapeutic modification of glycosaminoglycans in the treatment of disease and for use to increase diffusion of other injected molecules less than 200 nanometers in diameter in an animal.

BACKGROUND INFORMATION

[0003] Glycosaminoglycans (GAGs) are complex linear polysaccharides of the extracellular matrix (ECM). GAG's are characterized by repeating disaccharide structures of an N-substituted hexosamine and an uronic acid, [hyaluronan (HA), chondroitin sulfate (CS), chondroitin (C), dermatan sulfate (DS), heparan sulfate (HS), heparin (H)], or a galactose, [keratan sulfate (KS)]. Except for HA, all exist covalently bound to core proteins. The GAGs with their core proteins are structurally referred to as proteoglycans (PGs).

[0004] Hyaluronan (HA) is found in mammals predominantly in connective tissues, skin, cartilage, and in synovial fluid. Hyaluronan is also the main constituent of the vitreous of the eye. In connective tissue, the water of hydration associated with hyaluronan creates spaces between tissues, thus creating an environment conducive to cell movement and proliferation. Hyaluronan plays a key role in biological phenomena associated with cell motility including rapid development, regeneration, repair, embryogenesis, embryological development, wound healing, angiogenesis, and tumorigenesis (Toole 1991 Cell Biol. Extracell. Matrix, Hay (ed), Plenum Press, New York, 1384-1386; Bertrand et al. 1992 Int. J. Cancer 52:1-6; Knudson et al, 1993 FASEB J. 7:1233-1241). In addition, hyaluronan levels correlate with tumor aggressiveness (Ozello et al. 1960 Cancer Res. 20:600-604; Takeuchi et al. 1976, Cancer Res. 36:2133-2139; Kimata et al. 1983 Cancer Res. 43:1347-1354).

[0005] HA is found in the extracellular matrix of many cells, especially in soft connective tissues. HA has been assigned various physiological functions, such as in water and plasma protein homeostasis (Laurent TC et al (1992) FASEB J 6: 2397-2404). HA production increases in proliferating cells and may play a role in mitosis. It has also been implicated in locomotion and cell migration. HA seems to play important roles in cell regulation, development, and differentiation (Laurent et al, supra).

[0006] HA has been used in clinical medicine. Its tissue protective and rheological properties have proved useful in ophthalmic surgery to protect the corneal endothelium during cataract surgery. Serum HA is diagnostic of liver disease and various inflammatory conditions, such as rheumatoid arthritis. Interstitial edema caused by accumulation of HA may cause disfunction in various organs (Laurent et al, supra).

[0007] Hyaluronan protein interactions also are involved in the structure of the extracellular matrix or "ground substance".

[0008] Hyaluronidases are a group of neutral- and acid-active enzymes found throughout the animal kingdom. Hyaluronidases vary with respect to substrate specificity, and mechanism of action.

[0009] There are three general classes of hyaluronidases:

[0010] 1. Mammalian-type hyaluronidases, (EC 3.2.1.35) which are endo-beta-N-acetylhexosaminidases with tetrasaccharides and hexasaccharides as the major end products. They have both hydrolytic and transglycosidase activities, and can degrade hyaluronan and chondroitin sulfates (CS), specifically C4-S and C6-S.

[0011] 2. Bacterial hyaluronidases (EC 4.2.99.1) degrade hyaluronan and, and to various extents, CS and DS. They are endo-beta-N-acetylhexosaminidases that operate by a beta elimination reaction that yields primarily disaccharide end products.

[0012] 3. Hyaluronidases (EC 3.2.1.36) from leeches, other parasites, and crustaceans are endo-beta-glucuronidases that generate tetrasaccharide and hexasaccharide end products through hydrolysis of the beta 1-3 linkage.

[0013] Mammalian hyaluronidases can be further divided into two groups: neutral active and acid active enzymes. There are six hyaluronidase-like genes in the human genome, HYAL1, HYAL2, HYAL3 HYAL4 HYALP1 and PH20/SPAM1. HYALP1 is a pseudogene, and HYAL3 has not been shown to possess enzyme activity toward any known substrates. HYAL4 is a chondroitinase and lacks activity towards hyaluronan. HYAL1 is the prototypical acid-active enzyme and PH20 is the prototypical neutral-active enzyme. Acid active hyaluronidases, such as HYAL1 and HYAL2 lack catalytic activity at neutral PH. For example, HYAL1 has no catalytic activity in vitro over pH 4.5 (Frost et al *Anal Biochemistry*, 1997). HYAL2 is an acid active enzyme with a very low specific activity *in vitro*.

[0014] The hyaluronidase-like enzymes can also be characterized by those which are locked to the plasma membrane via a glycosylphosphatidyl inositol anchor such as human HYAL2 and human PH20 (Danilkovitch-Miagkova, et al. *Proc Natl Acad Sci U S A*. 2003 Apr 15;100(8):4580-5, Phelps et al., *Science* 1988) and those which are soluble such as human HYAL1 (Frost et al, *Biochem Biophys Res Commun*. 1997 Jul 9;236(1):10-5). However, there are variations from species to species: bovine, PH20 for example is very loosely attached to the plasma membrane and is not anchored via a phospholipase sensitive anchor (Lalancette et al, *Biol Reprod*. 2001 Aug;65(2):628-36.). This unique feature of bovine hyaluronidase has permitted the use of the soluble bovine testes hyaluronidase enzyme as an extract for clinical use (Wydase®, Hyalase®). Other PH20 species are lipid anchored enzymes that are not insoluble without the use of detergents or

lipases. For example, human PH20 is anchored to the plasma membrane via a GPI anchor. Attempts to make human PH20 DNA constructs that would not introduce a lipid anchor into the polypeptide resulted in either a catalytically inactive enzyme, or an insoluble enzyme (Arming et al *Eur J Biochem.* 1997 Aug 1;247(3):810-4). Naturally occurring macaque sperm hyaluronidase is found in both a soluble and membrane bound form. While the 64kDa membrane bound form possesses enzyme activity at pH 7.0, the 54kDa form is only active at pH 4.0 (Cherr et al, Dev Biol. 1996 Apr 10;175(1):142-53.). Thus, soluble forms of PH20 are often lacking enzyme activity under neutral conditions.

[0015] Chondroitinases are enzymes found throughout the animal kingdom. These enzymes degrade glycosaminoglycans through an endoglycosidase reaction. Specific examples of known Chondroitinases include Chondroitinase ABC (derived from Proteus vulgaris; Japanese Patent Application Laid-open No 6-153947, T. Yamagata, H. Saito, O. Habuchi, and S. Suzuki, J. Biol. Chem., 243, 1523 (1968), S. Suzuki, H. Saito, T. Yamagata, K. Anno, N. Seno, Y. Kawai, and T. Furuhashi, J. Biol. Chem., 243, 1543 (1968)), Chondroitinase AC (derived from Flavobacterium heparinum; T. Yamagata, H. Saito, O. Habuchi, and S. Suzuki, J. Biol. Chem., 243, 1523 (1968)), Chondroitinase AC II (derived from Arthrobacter aurescens; K. Hiyama, and S. Okada, J. Biol. Chem., 250, 1824 (1975), K. Hiyama and S. Okada, J. Biochem. (Tokyo), 80, 1201 (1976)), Hyaluronidase ACIII (derived from Flavobacterium sp. Hp102; Hirofumi Miyazono, Hiroshi Kikuchi, Keiichi Yoshida, Kiyoshi Morikawa, and Kiyochika Tokuyasu, Seikagaku, 61, 1023 (1989)), Chondroitinase B (derived from Flavobacterium heparinum; Y. M. Michelacci and C. P. Dietrich, Biochem. Biophys. Res. Commun., 56, 973 (1974), Y. M. Michelacci and C. P. Dietrich, Biochem. J., 151, 121 (1975), Kenichi Maeyama, Akira Tawada, Akiko Ueno, and Keiichi Yoshida, Seikagaku, 57, 1189 (1985)), Chondroitinase C (derived from Flavobacterium sp. Hp102; Hirofumi Miyazono, Hiroshi Kikuchi, Kelichi Yoshida, Kiyoshi Morikawa, and Kiyochika Tokuyasu, Seikagaku, 61, 1023 (1939)), and the like.

[0016] Glycoproteins are composed of a polypeptide chain covalently bound to one or more carbohydrate moieties. There are two broad categories of glycoproteins that posses carbohydrates coupled though either N-glycosidic or O-glycosidic linkages to their constituent protein. The N- and O-linked glycans are attached to polypeptides through

asparagine-N-acetyl-D-glucosamine and serine (threonine)-N-acetyl-D-galactosamine linkages, respectively. Complex N-linked oligosaccharides do not contain terminal mannose residues. They contain only terminal N-acetylglucosamine, galactose, and/or sialic acid residues. Hybrid oligosaccharides contain terminal mannose residues as well as terminal N-acetylglucosamine, galactose, and/or sialic acid residues.

[0017] With N-linked glycoproteins, an oligosaccharide precursor is attached to the amino group of asparagine during peptide synthesis in the endoplasmic reticulum. The oligosaccharide moiety is then sequentially processed by a series of specific enzymes that delete and add sugar moieties. The processing occurs in the endoplasmic reticulum and continues with passage through the cis-, medial- and trans-Golgi apparatus.

SUMMARY OF THE INVENTION

[0018] Provided herein are members of the soluble, neutral active Hyaluronidase Glycoprotein family, particularly the human soluble PH-20 Hyaluronidase Glycoproteins (also referred to herein as sHASEGPs). The sHASEGP provided herein is a sHASEGP family member, designated herein as a sHASEGP. The soluble Hyaluronidase domain, and uses thereof are also provided.

[0019] The invention is based upon the discovery that a soluble, neutral-active hyaluronidase activity can be produced with high yield in a mammalian expression system by introducing nucleic acids that lack a narrow region encoding amino acids in the carboxy terminus of the human PH20 cDNA. Additional modifications of the sHASEGP to enhance secretion by use of non-native leader peptides are also provided. Further provided are methods to modify the sHASEGP to prolong its half life by way of masking the protein with polyethylene glycol and posttranslational modifications to native glycosylation. Previous attempts to generate secreted a neutral active human sHASEGP were unsuccessful. It was concluded that truncations of the human sHASEGP polypeptide resulted in both a loss of neutral enzymatic activity, and an inability of cells to secrete the recombinant protein in mammalian expression systems (Arming, et al Eur J Biochem 1997 Aug 1;247 (3):810-4). It is critical to generate neutral-acting secreted sHASEGP for commercial production and therapeutic utility as a hyaluronidase. The invention, disclosed herein, overcomes such challenges.

[0020] The invention further comprises a catalytically active human sHASEGP glycoprotein wherein the sHASEGP possesses at least one N-linked sugar moiety. The studies shown herein demonstrate that human PH20 requires N-linked glycans for catalytic activity, whereas bovine and bee venom hyaluronidases remain active without such N-linked glycans. A human hyaluronidase domain devoid of N-linked moieties is catalytically inactive. Thus classic recombinant DNA technology does not permit the production of a catalytically active human sHASEGP, unlike bee venom HASEGP, which can be produced in *e.* coli.

[0021] The invention includes methods and cells for generation of an N-linked sHASEGP glycoprotein polypeptide, by using of a cell capable of introducing said N-linked sugar moieties or by introduction of said N-linked moieties on a sHASEGP polypeptide. Methods of identifying properly glycosylated sHASEGP's are further disclosed.

[0022] Catalytically active Super-Sialated sHASEGP glycoproteins are also provided. Super-sialated sHASEGPs possess greater serum half-lives compared to naturally occurring non-sialated bovine and ovine testes sHASEGPs, and are thus preferable for both enzyme stability and use as intravenous drugs. The invention provides methods for the preparation of Super-Sialated sHASEGPs, compositions and uses thereof.

[0023] Proteins encoded by naturally GPI anchor deficient sHASEGP's splice variants are also provided.

[0024] Further provided are compositions of the sHASEGP comprising, a soluble sHASEGP glycoprotein with a metal ion, wherein the metal ion is Calcium, Magnesium or Sodium. sHASEGPs are optimally active in the presence of said metals. Formulations consisting of sHASEGP in the presence of said metal ions are also provided.

[0025] Modifications of sHASEGP to further prolong the half life are provided. Chemical modifications of a sHASEGP with polymers such as polyethylene glycol and dextran are provided. Such modifications shield sHASEGP's from removal from circulation and the immune system as well as glycosyaltion receptors for mannose and asialoglycoprotein. Further provide are methods to link to specific functional groups such as glycosylation sites, positively charged amino acids and cysteines.

[0026] Assays for identifying effectors, such as compounds, including small molecules, and conditions, such pH, temperature and ionic strength, that modulate the activation, expression or activity of sHASEGP are also provided herein. In exemplary assays, the effects of test compounds on the ability of a Hyaluronidase domain of sHASEGP to cleave a known substrate, typically a glycosaminoglycan or proteoglycan, are assessed. Agents, generally compounds, particularly small molecules, that modulate the activity of the Hyaluronidase domain are candidate compounds for modulating the activity of the sHASEGP. The Hyaluronidase domains can also be used to produce Hyaluronidase-specific antibodies with function perturbing activity. The Hyaluronidase domains provided herein include, but are not limited to, the N-terminal glycsoyl-hydrolase domain with C-terminal truncated portions thereof that exhibit catalytic activity in vitro.

[0027] Nucleic acid molecules encoding the proteins and Hyaluronidase domains are also provided. Nucleic acid molecules that encode a soluble Hyaluronidase domain or catalytically active portions thereof and also those that encode the full- length sHASEGP are provided. Nucleic acid encoding the Hyaluronidase domain and downstream nucleic acid is set forth in SEQ ID No. 6; and the Hyaluronidase domain of sHASEGP is set forth in SEQ ID No. 1 (amino acids 35-464). The protein sequence and encoding nucleic acid sequence of the full-length sHASEGP are set forth in SEQ ID Nos. 1 and 6.

[0028] Also provided are nucleic acid molecules that hybridize to such sHASEGP-encoding nucleic acid along their full-length or along at least about 70%, 80% or 90% of the full-length and encode the Hyaluronidase domain or portion thereof are provided. Hybridization is generally effected under conditions of at least low, generally at least moderate, and often high stringency.

[0029] The isolated nucleic acid fragment is DNA, including genomic or cDNA, or is RNA, or can include other components, such as protein nucleic acid or other nucleotide analogs. The isolated nucleic acid may include additional components, such as heterologous or native promoters, and other transcriptional and translational regulatory sequences, these genes may be linked to other genes, such as reporter genes or other indicator genes or genes that encode indicators.

[0030] Also provided is an isolated nucleic acid molecule that includes the sequence of molecules that is complementary to the nucleotide sequence encoding sHASEGP or the portion thereof.

[0031] Also provided are fragments thereof or oligonucleotides that can be used as probes or primers and that contain at least about 10,14,16 nucleotides, generally less than 1000 or less than or equal-to 100, set forth in SEQ ID NO. 6 (or the complement thereof); or contain at least about 30 nucleotides (or the complement thereof) or contain oligonucleotides that hybridize along their full-length (or at least about 70, 80 or 90% thereof) to any such fragments or oligonucleotides. The length of the fragments are a function of the purpose for which they are used and/or the complexity of the genome of interest. Generally probes and primers contain less than about 50,150 or 500 nucleotides.

[0032] Also provided are plasmids containing any of the nucleic acid molecules provided herein. Cells containing the plasmids are also provided. Such cells include, but are not limited to, bacterial cells, yeast cells, fungal cells, plant cells, insect cells and animal cells.

[0033] Also provided are enhanced mammalian expression systems using signal leaders capable of efficient secretion of sHASEGP. An example of such efficient secretory leader peptide amino acid sequence and fusion protein with sHASEGP is found in SEQ ID Nos. 43 and 46.

[0034] Also provided is a method of producing sHASEGP by growing the above-described cells under conditions whereby the sHASEGP is expressed by the cells, and recovering the expressed sHASEGP polypeptide or glycoprotein. Methods for isolating nucleic acid encoding other sHASEGPs are also provided.

[0035] Also provided are cells, generally eukaryotic cells, such as mammalian cells and yeast cells, in which the sHASEGP polypeptide is expressed on the surface of the cells. Such cells are used in drug screening assays to identify compounds that modulate the activity of the sHASEGP polypeptide. These assays, including in vitro binding assays, and transcription based assays in which signal transduction mediated directly or indirectly, such as via activation of pro-growth factors, by the sHASEGP is assessed.

[0036] Also provided are peptides encoded by such nucleic acid molecules. Included among those polypeptides is the sHASEGP Hyaluronidase domain or a polypeptide with

amino acid changes such that the specificity and/or Hyaluronidase activity remains substantially unchanged. In particular, a substantially purified mammalian sHASEGP glycoprotein is provided that includes a secreted neutral catalytically active

[0037] The invention also includes a Hyaluronidase catalytic domain and may additionally include other domains. The sHASEGP may form homodimers and can also form heterodimers with some other protein, such as a membrane-bound protein. Also provided is a substantially purified glycoprotein including a sequence of amino acids that has at least 60%, 70%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91 %, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% identity to the sHASEGP where the percentage identity is determined using standard algorithms and gap penalties that maximize the percentage identity.

[0038] Splice variants of the sHASEGP, particularly those with a catalytically active Hyaluronidase domain, are contemplated herein.

[0039] In other embodiments, substantially purified polypeptides that include a Hyaluronidase domain of a sHASEGP polypeptide or a catalytically active portion thereof, but that do not include the entire sequence of amino acids set forth in SEQ ID No. 1 are provided. Among these are polypeptides that include a sequence of amino acids that has at least 70%, 80%, 85%, 90%, 95% or 100% sequence identity to SEQ ID No. 1 or 3.

[0040] In a specific embodiment, a nucleic acid that encodes a eukaryotic Hyaluronidase glycoprotein, designated sHASEGP is provided. In particular, the nucleic acid includes the sequence of nucleotides set forth in SEQ ID No. 6, particularly set forth as nucleotides 106-1446 of SEQ ID NO. 6, or a portion there of that encodes a catalytically active polypeptide.

[0041] Also provided are nucleic acid molecules that hybridize under conditions of at least low stringency, generally moderate stringency, more typically high stringency to the SEQ ID NO. 6 or degenerates thereof.

[0042] In one embodiment, the isolated nucleic acid fragment hybridizes to a nucleic acid molecule containing the nucleotide sequence set forth in SEQ ID No: 6 (or degenerates thereof) under high stringency conditions. A full-length sHASEGP is set forth in SEQ ID No. 1 and is encoded by SEQ ID NO. 6 or degenerates thereof.

[0043] Also provided are muteins of the Hyaluronidase domain of sHASEGP, particularly muteins in which the Cys residue in the Hyaluronidase domain that is free i.e., does not form disulfide linkages with any other Cys residue in the Hyaluronidase domain) is substituted with another amino acid substitution, typically, although not necessarily, with a conservative amino acid substitution or a substitution that does not eliminate the activity, and muteins in which a specific glycosylation site (s) is eliminated.

[0044] sHASEGP polypeptides, including, but not limited to splice variants thereof, and nucleic acids encoding sHASEGPs, and domains, derivatives and analogs thereof are provided herein. Single chain secreted Hyaluronidase glycoproteins that have an N-terminus functionally equivalent to that generated by activation of a signal peptidase to form sHASEGP are also provided. There are seven potential N-linked glycosylation sites at N82, N166, N235, N254, N368, N393, N490 of sHASEGP as exemplified in SEQ ID NO: 1. Disulfide bonds form between the Cys residues C60-C351 and Cys residues C224 to C238 to form the core Hyaluronidase domain. However, additional cysteines are required in the carboxy terminus for neutral enzyme catalytic activity such that sHASEGP from amino acids 36 to Cys 464 in SEQ ID No.1 comprise the minimally active human sHASEGP hyaluronidase domain. Thus, N-linked glycosylation site N-490 is not required for proper sHASEGP activity.

[0045] N-linked glycosylation of the sHASEGP's are critical for their catalytic activity and stability. While altering the type of glycan modifying a glycoprotein can have dramatic affects on a protein's antigenicity, structural folding, solubility, and stability, most enzymes are not thought to require glycosylation for optimal enzyme activity. sHASEGPs are thus unique in this regard, that removal of N-linked glycosylation can result in near complete inactivation of the Hyaluronidase activity. The presence of N-linked glycans is critical for generating an active sHASEGP. Protein expression systems suitable for the introduction of critical N-linked glycosylation residues on sHASEGP are included. Additionally, the introduction of deglycosylated sHASEGP polypeptide in the presence of extracts capable of introducing N-linked glycans are included. In one aspect of the invention, complex glycosylation capped with sialation is described whereas others capped with free mannose residues are contemplated as well. Preferably, sialic acid residues are found in the terminal residues of N -linked glycosylation on sHASEGP.

[0046] N-linked oligosaccharides fall into several major types (oligomannose, complex, hybrid, sulfated), all of which have (Man) 3-GlcNAc-GlcNAc-cores attached via the amide nitrogen of Asn residues that fall within -Asn-Xaa-Thr/Ser- sequences (where Xaa is not Pro). Glycosylation at an -Asn-Xaa-Cys- site has been reported for coagulation protein C. N-linked sites are often indirectly assigned by the appearance of a "blank" cycle during sequencing. Positive identification can be made after release of the oligosaccharide by PNGase F, which converts the glycosylated Asn to Asp. After PNGase F release, N-linked oligosaccharides can be purified using Bio-Gel P-6 chromatography, with the oligosaccharide pool subjected to preparative high pH anion exchange chromatography (HPAEC) (Townsend et al., (1989) Anal. Biochem. 182, 1-8). Certain oligosaccharide isomers can be resolved using HPAEC. Fucose residues will shift elution positions earlier in the HPAEC chromatogram, while additional sialic acid residues will increase the retention time. Concurrent treatment of glycoproteins whose oligosaccharide structures are known (e.g., bovine fetuin, a-l acid glycoprotein, ovalbumin, RNAse B, transferrin) can facilitate assignment of the oligosaccharide peaks. The collected oligosaccharides can be characterized by a combination of compositional and methylation linkage analyses (Waegheet al., (1983) Carbohydr Res. 123, 281-304.), with anomeric configurations assigned by NMR spectroscopy (Van Halbeek (1993) in Methods Enzymol 230).

[0047] Formulations of sHASEGP's are also provided. sHASEGPs may be formulated in lyophilized forms and stabilized solutions. Formulations containing specific metal ions, such as calcium, magnesium, or sodium, are useful for optimal activity at neutral PH. In addition to stabilized solution formulations, slow release formulations are contemplated herein for extended removal of glycosaminoglycans. Also provided herein are kits providing for pre-packaged syringes of sHASEGP's for the administration of small volumes of sHASEGP for intraocular surgical procedures and other small volume procedures. Balanced salt formulations for *ex vivo* use in artificial reproductive technology procedures are also provided.

[0048] Methods for the use of sHASEGP's in the removal of glycosaminoglycans are also provided. sHASEGPs open channels in the interstitial space through degradation of glycosaminoglycans that permit the diffusion of molecules less than 500nm in size. These channels remain for a period of 24-48 hours depending on dose and formulation. Such

channels can be used to facilitate the diffusion of exogenously added molecules such as fluids, small molecules, proteins, nucleic acids and gene therapy vectors and other molecules less than 500nm in size.

[0049] sHASEGPs can also be used to remove excess glycosaminoglycans such as those that occur following ischemia reperfusion, inflammation, arteriosclerosis, edema, cancer, spinal cord injury and other forms of scarring. In some instances, sHASEGP's can be delivered systemically by intravenous infusion. This can be helpful when local access is not readily available such as the heart or brain or in the case of disseminated neoplasm wherein the disease is through the body. Super-Sialated sHASEGP's are preferable to increase serum half-life and distribution over native hyaluronidase enzymes that lack terminal sialic acids.

[0050] In some circumstances, such as spinal cord injury, glaucoma, and cosmetic treatments, sustained delivery is preferred.

[0051] In other indications, a single short acting dose is preferable. Temporary removal of glycosaminoglycans can be used to enhance the delivery of solutions and drugs into interstitial spaces. This can be useful for the diffusion of anesthesia and for the administration of therapeutic fluids, molecules and proteins. Subcutaneous and Intramuscular administration of molecules in the presence of sHASEGP's also facilitate their systemic distribution more rapidly. Such methods are very useful when intravenous access is not available or where more rapid systemic delivery of molecules is needed. Delivery of other large molecules such as Factor VIII, that are poorly bioavailable upon subcutaneous administration, may be injected with sHASEGP's to increase their availability.

[0052] Uses of sHASEGP's for enzymatic removal of the cumulus matrix surrounding oocytes are also provided. The removal of the cumulus matrix using a purified sHASEGP without the toxic contaminants of extract derived hyaluronidase permits more gentle recover of the oocyte with greater viabilities. Moreover, sHASEGP's can be manufactured without the use of cattle extracts or other organisms that carry viruses and other pathogens such as transmissbale spongiform ecephalopathies.

[0053] Injections of small volumes of sHASEGP for intraocular use may also be used for small spaces. SHASEGPs may be injected into the anterior chamber of the eye to remove excess viscoelastic substrates that are administered during surgery. Intraocular injection of sHASEGP's can also be used to reduce intraocular pressure in glaucoma, to dissolve vitreous aggregates, or "floaters", to clear vitreous hemorrhage, for the treatment of macular degeneration, to promote vitreo retinal detachment in diabetic retinopathy and to be mixed with other enzymes to promote reshaping of the cornea along with corrective lenses. It will be recognized that in some instances, the use of a long lasting sHASEGP such as a pegylated-sHASEGP will be desireable.

[0054] Co-formulations of sHASEGP with other substances may also be envisioned for injectable pens for small volume or rapid subcutaneous administration. Examples such as Epipen®, insulin, and other fluids can be formulated. The methods of the invention include administration of the sHASEGP polypeptide or pharmaceutical compositions containing sHASEGP prior to, simultaneously with or following administration of other therapeutic molecules. The sHASEGP may be administered at a site different from the site of administration of the therapeutic molecule or the sHASEGP may be administered at a site the same as the site of administration of the therapeutic molecule.

[0055] Hence, provided herein is a family of eukaryotic secreted neutral active hyaluronidase glycoproteins designated sHASEGP's, and functional domains, especially Hyaluronidase (or catalytic) domains thereof, muteins and other derivatives and analogs thereof. Also provided herein are nucleic acids encoding the sHASEGPs. Additionally provided are formulations and therapeutic uses of said sHASEGP's to treat disease and for use as tissue modifying enzymes.

BRIEF DESCRIPTION OF THE DRAWINGS

[0056] Figure 1 is a vector map of sHASEGP Vector HZ24.

DETAILED DESCRIPTION OF THE INVENTION

[0057] A. DEFINITIONS: Unless defined otherwise, all technical and scientific terms used herein have the same meaning as is commonly understood by one of skill in the art to which the invention (s) belong. All patents, patent applications, published applications and publications, Genbank sequences, websites and other published materials referred to throughout the entire disclosure herein, unless noted otherwise, are incorporated by reference in their entirety. In the event that there is a plurality of definitions for terms herein, those in this section prevail.

[0058] Where reference is made to a URL or other such identifier or address, it understood that such identifiers can change and particular information on the internet can come and go, but equivalent information can be found by searching the internet. Reference thereto evidences the availability and public dissemination of such information.

[0059] As used herein, the abbreviations for any protective groups, amino acids and other compounds, are, unless indicated otherwise, in accord with their common usage, recognized abbreviations, or the IUPAC-IUB Commission on Biochemical Nomenclature (see, (1972) Biochem. 11 : 942-944).

[0060] As used herein, eukaryotic Hyaluronidase refers to a diverse family of glycosaminoglycan endoglucosaminidases, wherein a glutamate residue in the Hyaluronidase hydrolyzes the beta 1,4 linkages of hyaluronan and chondroitin sulfates through an acid-base catalytic mechanism.

[0061] Of particular interest are sHASEGP's of mammalian, including human, origin. Those of skill in this art recognize that, in general, single amino acid substitutions in non-essential regions of a polypeptide do not substantially alter biological activity (see, e.g., Watson et al., (1987) Molecular Biology of the Gene, 4th Edition, The Benjamin/Cummings Pub. co., p. 224).

[0062] As used herein, membrane anchored sHASEGP, refers to a family of membrane anchored Hyaluronidases that share common structural features as described herein.

[0063] As used herein, soluble hyaluronidase refers to a polypeptide characterized by its solubility under physiologic conditions. Soluble HASEGP can be distinguished for

example by its partitioning into the aqueous phase of a Triton X-114 solution warmed to 37C (Bordier et al J Biol Chem. 1981 Feb 25;256(4):1604-7). Lipid anchored HASEGP on the other hand will partition into the detergent rich phase, but will partition into the detergent poor or aqueous phase following treatment with Phospholipase-C.

[0064] Thus, reference, for example, to "sHASEGP"encompasses all glycoproteins encoded by the sHASEGP gene family, including but not limited to: Human sHASEGP, mouse sHASEGP, or an equivalent molecule obtained from any other source or that has been prepared synthetically or that exhibits the same activity. Sequences of encoding nucleic acid molecules and the encoded amino acid sequences of exemplary sHASEGP's and/or domains thereof are set forth, for example in SEQ ID NO: 4. The term also encompasses sHASEGP with amino acid substitutions that do not substantially alter activity of each member and also encompasses splice variants thereof. Suitable substitutions, including, although not necessarily, conservative substitutions of amino acids, are known to those of skill in this art and can be made without eliminating the biological activity, such as the catalytic activity, of the resulting molecule.

[0065] As used herein, a sHASEGP, whenever referenced herein, includes at least one or all of or any combination of: a polypeptide encoded by the sequence of nucleotides set forth in SEQ ID NO. 6 or by a sequence of nucleotides that includes nucleotides that encode amino acids 1-509 of SEQ ID No. 1; a polypeptide encoded by a sequence of nucleotides that hybridizes under conditions of low, moderate or high stringency to the sequence of nucleotides set forth in SEQ ID NO. 6; a polypeptide that includes the sequence of amino acids set forth as amino acids 1-509 of SEQ ID No. 1; a polypeptide that includes a sequence of amino acids having at least about 60%, 70%, 75%, 80%, 81 %, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91 %, 92%, 93%, 94%, 95%, 96%, 97%, 98% or 99% sequence identity with the sequence of amino acids set forth in SEQ ID No. 1 or as amino acids 1-448 of SEQ ID No. 4 .

[0066] In particular, the sHASEGP polypeptide, with the Hyaluronidase domains as indicated in SEQ ID No. 4 is provided. The polypeptide is a single or two chain polypeptide. Smaller portions thereof that retain Hyaluronidase activity are also provided. The Hyaluronidase domains from sHASEGPs vary in size and constitution, including insertions and deletions in surface loops. Thus, for purposes herein, the catalytic domain

is a portion of a sHASEGP, as defined herein, and is homologous to a domain of other hyaluronidase like sequences, such as HYAL1, HYAL2, HYAL3, which have been previously identified; it was not recognized, however, that an isolated single chain form of the human Hyaluronidase domain could function in *in vitro* assays. The Aspartate and Glutamate residues necessary for activity are present in conserved motifs.

[0067] As used herein, a " neutral hyaluronidase domain of a soluble sHASEGP" refers to an beta 1,4 endoglucosaminidase domain of a sHASEGP that exhibits Hyaluronidase activity at neutral PH, is soluble under conditions as described and shares homology and structural features with the hyaluronidase glycosyl-hydrolase family domains but contains additional sequences in the carboxy terminus that are required for neutral activity. Hence it is at least the minimal portion of the domain that exhibits Hyaluronidase activity as assessed by standard in vitro assays and remains soluble. Contemplated herein are such Hyaluronidase domains and catalytically active portions thereof. Also provided are truncated forms of the Hyaluronidase domain that include the smallest fragment thereof that acts catalytically as a single chain form.

[0068] A Hyaluronidase domain of an sHASEGP, whenever referenced herein, includes at least one or all of or any combination of or a catalytically active portion of: an N-linked glycoprotein polypeptide that includes the sequence of amino acids set forth in SEQ ID No. 1; a polypeptide encoded by a sequence of nucleotides that hybridizes under conditions of low, moderate or high stringency to the sequence of nucleotides set forth in SEQ ID NO. 6; a polypeptide that includes the sequence of amino acids set forth in SEQ ID No. 1; a polypeptide that includes a sequence of amino acids having at least about 70%, 75%, 80%, 81 %, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91 %, 92%, 93%, 94%, 95%, 96%, 97%, 98% or 99% sequence identity with the sequence of amino acids set forth in SEQ ID No. 1; and/or a Hyaluronidase domain of a polypeptide encoded by a splice variant of the sHASEGP.

[0069] Thus, for purposes herein, the Hyaluronidase domain is a portion of a sHASEGP, as defined herein, and is homologous to a domain of other sHASEGP's. As with the larger class of enzymes of the hyaluronidase family, the sHASEGP catalytic domains share a high degree of amino acid sequence identity. The Asp and Glu residues necessary for activity are present in conserved motifs.

[0070] By active form is meant a form active in vivo and/or in vitro. As described herein, the Hyaluronidase domain also can exist as a soluble secreted glycoprotein. It is shown herein that, at least in vitro, the single chain forms of the sHASEGP's and the catalytic domains or enzymatically active portions thereof (typically C-terminal truncations) exhibit Hyaluronidase activity. Hence provided herein are isolated forms of the Hyaluronidase domains of sHASEGP's and their use in in vitro drug screening assays for identification of agents that modulate the activity thereof.

[0071] As used herein, the catalytically active domain of a sHASEGP refers to the neutral active endoglucosaminidase domain as defined by activity *in vitro* towards a glycosaminoglycan substrate.

[0072] sHASEGPs of interest include those that are active against chondroitin sulfates and chondroitin sulfate proteoglycans (CSPG's) in vivo and in vitro; and those that are active against hyaluronan. As used herein, a human sHASEGP is one encoded by nucleic acid, such as DNA, present in the genome of a human, including all allelic variants and conservative variations as long as they are not variants found in other mammals.

[0073] As used herein, nucleic acid encoding a Hyaluronidase domain or catalytically active portion of a sHASEGP" shall be construed as referring to a nucleic acid encoding only the recited single chain Hyaluronidase domain or active portion thereof, and not the other contiguous portions of the sHASEGP as a continuous sequence.

[0074] As used herein," disease" or "disorder" refers to a pathological condition in an organism resulting from, e.g., infection or genetic defect, and characterized by identifiable symptoms.

[0075] As used herein, a splice variant refers to a variant produced by differential processing of a primary transcript of genomic nucleic acid, such as DNA, that results in more than one type of mRNA. Splice variants of sHASEGPs are provided herein.

[0076] As used herein, the Hyaluronidase domain of a sHASEGP protein refers to the Hyaluronidase domain of a sHASEGP that exhibits neutral endoglucosaminidase activity. Hence it is at least the minimal portion of the protein that exhibits endoglucosaminidase activity as assessed by standard assays in vitro. Exemplary human Hyaluronidase domains

include at least a sufficient portion of sequences of amino acids set forth in SEQ ID No. 4 to exhibit endoglucosaminidase activity.

[0077] Also contemplated are nucleic acid molecules that encode a polypeptide that has endoglucosaminidase activity in an in vitro Hyaluronidase assay and that have at least 70%, 75%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91 %, 92%, 93%, 94%, 95%, 96%, 97%, 98% or 99% sequence identity with the full-length of a Hyaluronidase domain of an sHASEGP polypeptide, or that hybridize along their full-length or along at least about 70%, 80% or 90% of the full-length to a nucleic acids that encode a Hyaluronidase domain, particularly under conditions of moderate, generally high, stringency.

[0078] For the Hyaluronidase domains, residues at the in the N-terminal region can be critical yet not sufficient for activity. It is shown herein that the Hyaluronidase domain of the sHASEGP is catalytically active. Hence the Hyaluronidase domain generally requires the N-terminal amino acids thereof for activity; the C-terminus portion can be truncated until the last Cysteine residue yet requires additional amino acids to be optimally active. The amount that can be removed can be determined empirically by testing the polypeptide for Hyaluronidase activity in an in vitro assay that assesses catalytic cleavage.

[0079] Hence smaller portions of the Hyaluronidase domains, particularly the single chain domains, thereof that retain Hyaluronidase activity are contemplated. Such smaller versions generally are C-terminal truncated versions of the Hyaluronidase domains. The Hyaluronidase domains vary in size and constitution, including insertions and deletions in surface loops. Such domains exhibit conserved structure, including at least one structural feature, such as the proton donor, and/or other features of Hyaluronidase domains of endoglucosaminidases. Thus, for purposes herein, the Hyaluronidase domain is a single chain portion of a sHASEGP, as defined herein, but is homologous in its structural features and retention of sequence of similarity or homology the Hyaluronidase domain of other hyaluronidase-like sequences. The glycoprotein exhibits Hyaluronidase activity as a single chain.

[0080] As used herein, by homologous means about greater than 25% nucleic acid sequence identity, such as 25% 40%, 60%, 70%, 80%, 90% or 95%. If necessary the percentage homology will be specified. The terms"homology" and"identity"are often used

interchangeably. In general, sequences are aligned so that the highest order match is obtained (see, e.g.: Computational Molecular Biology, Lesk, A. M., ed., Oxford University Press, New York, 1988; Biocomputing : Informatics and Genome Projects, Smith, D. W., ed., Academic Press, New York, 1993; Computer Analysis of Sequence Data, Part/, Griffin, A. M., and Griffin, H. G., eds., Humana Press, New Jersey, 1994; Sequence Analysis in Molecular Biology, von Heinje, G., Academic Press, 1987; and Sequence Analysis Primer, Gribskov, M. and Devereux, J., eds., M Stockton Press, New York, 1991; Carillo et Al. (1988) et al. (1988) Slam J Applied Math 48] : 1073).

[0081] By sequence identity, the numbers of conserved amino acids is determined by standard alignment algorithms programs, and are used with default gap penalties established by each supplier. Substantially homologous nucleic acid molecules would hybridize typically at moderate stringency or at high stringency all along the length of the nucleic acid or along at least about 70%, 80% or 90% of the full-length nucleic acid molecule of interest. Also contemplated are nucleic acid molecules that contain degenerate codons in place of codons in the hybridizing nucleic acid molecule.

[0082] Whether any two nucleic acid molecules have nucleotide sequences that are at least, for example, 80%, 85%, 90%, 95%, 96%, 97%, 98% or 99% "identical" can be determined using known computer algorithms such as the "FASTA"program, using for example, the default parameters as in Pearson et al (1988) [Proc. Natl. Acad. Sci. USA 85]: 2444 (other programs include the GCG program package (Devereux, J., et al, Nucleic Acids Research 12: 387 (1984)), BLASTP, BLASTN, FASTA (Atschul, [S.] [F.,] [ET AL, J MOLEC BIOL 215]: 403 (1990); Guide to Huge Computers, Martin J. Bishop, [ED.,] Academic Press, San Diego, 1994, and [CARILLO ETA/.] (1988) SIAM J Applied Math 48: 1073). For example, the BLAST function of the National Center for Biotechnology Information database can be used to determine identity. Other commercially or publicly available programs include, DNASTAR"MEGALIGN"PROGRAM (Madison, WI) and the University of Wisconsin Genetics Computer Group (UWG)"Gap" program (Madison WI)). Percent homology or identity of proteins and/or nucleic acid molecules can be determined, for example, by comparing sequence information using a GAP computer program e.g. Needleman et al. (1970), J Mol Biol. 48: 443, as revised by Smith and Waterman Adv. Appl. Math (1981) 2:482). Briefly, the GAP program defines similarity

as the number of aligned symbols (i.e., nucleotides or amino acids) that are similar, divided by the total number of symbols in the shorter of the two sequences. Default parameters for the GAP program can include: (1) a unary comparison matrix (containing a value of 1 for identities and 0 for non-identities) and the weighted comparison matrix of Gribskov et al (1986) Nucl. Acids Res. 14: 6745, as described by Schwartz and Dayhoff, eds., Atlas Of Protein Sequence And Structure, National Biomedical Research Foundation, pp. 353-358 (1979); (2) a penalty of 3.0 for each gap and an additional 0.10 penalty for each symbol in each gap; and (3) no penalty for end gaps. Therefore, as used herein, the term"identity"represents a comparison between a test and a reference polypeptide or polynucleotide.

[0083] As used herein, the term at least"90% identical to" refers to percent identities from 90 to 99.99 relative to the reference polypeptides. Identity at a level of 90% or more is indicative of the fact that, assuming for exemplification purposes a test and reference polynucleotide length of 100 amino acids are compared. No more than 10% (i.e., 10 out of 100) amino acids in the test polypeptide differ from that of the reference polypeptides. Similar comparisons can be made between a test and reference polynucleotides. Such differences can be represented as point mutations randomly distributed over the entire length of an amino acid sequence or they can be clustered in one or more locations of varying length up to the maximum allowable, e.g. 10/100 amino acid difference (approximately 90% identity). Differences are defined as nucleic acid or amino acid substitutions, or deletions. At the level of homologies or identities above about 85-90%, the result should be independent of the program and gap parameters set; such high levels of identity can be assessed readily, often without relying on software.

[0084] As used herein, primer refers to an oligonucleotide containing two or more deoxyribonucleotides or ribonucleotides, typically more than three, from which synthesis of a primer extension product can be initiated. Experimental conditions conducive to synthesis include the presence of nucleoside triphosphates and an agent for polymerization and extension, such as DNA polymerase, and a suitable buffer, temperature and pH.

[0085] As used herein, animals include any animal, such as, but are not limited to, goats, cows, deer, sheep, rodents, pigs and humans. Non-human animals, exclude humans as the contemplated animal. The sHASEGPs provided herein are from any source, animal, plant,

prokaryotic and fungal. Most sHASEGP's are of animal origin, including mammalian origin.

[0086] As used herein, genetic therapy involves the transfer of heterologous nucleic acid, such as DNA, into certain cells, target cells, of a mammal, particularly a human, with a disorder or conditions for which such therapy is sought. The nucleic acid, such as DNA, is introduced into the selected target cells in a manner such that the heterologous nucleic acid, such as DNA, is expressed and a therapeutic product encoded thereby is produced.

[0087] Alternatively, the heterologous nucleic acid, such as DNA, can in some manner mediate expression of DNA that encodes the therapeutic product, or it can encode a product, such as a peptide or RNA that in some manner mediates, directly or indirectly, expression of a therapeutic product. Genetic therapy can also be used to deliver nucleic acid encoding a gene product that replaces a defective gene or supplements a gene product produced by the mammal or the cell in which it is introduced. The introduced nucleic acid can encode a therapeutic compound, such as a growth factor inhibitor thereof, or a tumor necrosis factor or inhibitor thereof, such as a receptor therefore, that is not normally produced in the mammalian host or that is not produced in therapeutically effective amounts or at a therapeutically useful time. The heterologous nucleic acid, such as DNA, encoding the therapeutic product can be modified prior to introduction into the cells of the afflicted host in order to enhance or otherwise alter the product or expression thereof. Genetic therapy can also involve delivery of an inhibitor or repressor or other modulator of gene expression.

[0088] As used herein, heterologous nucleic acid is nucleic acid that (if DNA encodes RNA) and proteins that are not normally produced in vivo by the cell in which it is expressed or that mediates or encodes mediators that alter expression of endogenous nucleic acid, such as DNA, by affecting transcription, translation, or other regulatable biochemical processes. Heterologous nucleic acid, such as DNA, can also be referred to as foreign nucleic acid, such as DNA. Any nucleic acid, such as DNA, that one of skill in the art would recognize or consider as heterologous or foreign to the cell in which is expressed is herein encompassed by heterologous nucleic acid; heterologous nucleic acid includes exogenously added nucleic acid that is also expressed endogenously. Examples of heterologous nucleic acid include, but are not limited to, nucleic acid that encodes

traceable marker proteins, such as a protein that confers drug resistance, nucleic acid that encodes therapeutically effective substances, such as anti-cancer agents, enzymes and hormones, and nucleic acid, such as DNA, that encodes other types of proteins, such as antibodies. Antibodies that are encoded by heterologous nucleic acid can be secreted or expressed on the surface of the cell in which the heterologous nucleic acid has been introduced.

[0089] Heterologous nucleic acid is generally not endogenous to the cell into which it is introduced, but has been obtained from another cell or prepared synthetically.

[0090] Generally, although not necessarily, such nucleic acid encodes RNA and proteins that are not normally produced by the cell in which it is expressed.

[0091] As used herein, a therapeutically effective product is a product that is encoded by heterologous nucleic acid, typically DNA, that, upon introduction of the nucleic acid into a host, a product is expressed that ameliorates or eliminates the symptoms, manifestations of an inherited or acquired disease or that cures the disease.

[0092] As used herein, recitation that a glycoprotein consists essentially of the Hyaluronidase domain means that the only sHASEGP portion of the polypeptide is a Hyaluronidase domain or a catalytically active portion thereof. The polypeptide can optionally, and generally will, include additional non-sHASEGP-derived sequences of amino acids.

[0093] As used herein, domain refers to a portion of a molecule, e.g., glycoproteins or the encoding nucleic acids that is structurally and/or functionally distinct from other portions of the molecule.

[0094] As used herein, Hyaluronidase refers to an enzyme catalyzing hydrolysis of glycosaminoglycans.

[0095] For clarity reference to Hyaluronidase refers to all forms, and particular forms will be specifically designated. For purposes herein, the Hyaluronidase domain includes the membrane bound and soluble forms of a sHASEGP protein.

[0096] As used herein, nucleic acids include DNA, RNA and analogs thereof, including protein nucleic acids (PNA) and mixture thereof. Nucleic acids can be single or double-

stranded. When referring to probes or primers, optionally labeled, with a detectable label, such as a fluorescent or radiolabel, single- stranded molecules are contemplated. Such molecules are typically of a length such that their target is statistically unique or of low copy number (typically less than 5, generally less than 3) for probing or priming a library. Generally a probe or primer contains at least 14,16 or 30 contiguous of sequence complementary to or identical a gene of interest. Probes and primers can be 10,20,30,50, 100 or more nucleic acids long.

[0097] As used herein, nucleic acid encoding a fragment or portion of a sHASEGP refers to a nucleic acid encoding only the recited fragment or portion of sHASEGP, and not the other contiguous portions of the sHASEGP.

[0098] As used herein, operative linkage of heterologous nucleic to regulatory and effector sequences of nucleotides, such as promoters, enhancers, transcriptional and translational stop sites, and other signal sequences refers to the relationship between such nucleic acid, such as DNA, and such sequences of nucleotides. For example, operative linkage of heterologous DNA to a promoter refers to the physical relationship between the DNA and the promoter such that the transcription of such DNA is initiated from the promoter by an RNA polymerase that specifically recognizes, binds to and transcribes the DNA in reading frame. Thus, operatively linked or operationally associated refers to the functional relationship of nucleic acid, such as DNA, with regulatory and effector sequences of nucleotides, such as promoters, enhancers, transcriptional and translational stop sites, and other signal sequences. For example, operative linkage of DNA to a promoter refers to the physical and functional relationship between the DNA and the promoter such that the transcription of such DNA is initiated from the promoter by an RNA polymerase that specifically recognizes, binds to and transcribes the DNA. In order to optimize expression and/or in vitro transcription, it can be necessary to remove, add or alter 5' Untranslated portions of the clones to eliminate extra, potential inappropriate alternative translation initiation i.e. start) codons or other sequences that can interfere with or reduce expression, either at the level of transcription or translation. Alternatively, consensus ribosome binding sites (see, e.g., Kozak J. Biol. Chem. 266: 19867- 19870 (1991) can be inserted immediately 5'of the start codon and can enhance expression. The desirability of (or need for) such modification can be empirically determined.

[0099] As used herein, a sequence complementary to at least a portion of an RNA, with reference to antisense oligonucleotides, means a sequence having sufficient complimentary to be able to hybridize with the RNA, generally under moderate or high stringency conditions, forming a stable duplex; in the case of double-stranded sHASEGP antisense nucleic acids, a single strand of the duplex DNA (or dsRNA) can thus be tested, or triplex formation can be assayed. The ability to hybridize depends on the degree of complimentarity and the length of the antisense nucleic acid. Generally, the longer the hybridizing nucleic acid, the more base mismatches with a sHASEGP encoding RNA it can contain and still form a stable duplex (or triplex, as the case can be). One skilled in the art can ascertain a tolerable degree of mismatch by use of standard procedures to determine the melting point of the hybridized complex.

[0100] For purposes herein, amino acid substitutions can be made in any of sHASEGPs and Hyaluronidase domains thereof provided that the resulting protein exhibits Hyaluronidase activity. Amino acid substitutions contemplated include conservative substitutions, such as those set forth in Table 1, which do not eliminate proteolytic activity. As described herein, substitutions that alter properties of the proteins, such as removal of cleavage sites and other such sites are also contemplated; such substitutions are generally non-conservative, but can be readily effected by those of skill in the art.

[0101] Suitable conservative substitutions of amino acids are known to those of skill in this art and can be made generally without altering the biological activity, for example enzymatic activity, of the resulting molecule. Those of skill in this art recognize that, in general, single amino acid substitutions in non-essential regions of a polypeptide do not substantially alter biological activity (see, e. g., Watson et al. Molecular Biology of the Gene, 4th Edition, 1987, The Benjamin/Cummings Pub. co., p.224). Also included within the definition, is the catalytically active fragment of a sHASEGP, particularly a single chain Hyaluronidase portion. Conservative amino acid substitutions are made, for example, in accordance with those set forth in TABLE 1 as follows:

[0102] TABLE 1 Original residue Conservative substitution Ala (A) Gly; Ser, Abu Arg (R) Lys, orn Asn (N) Gln ; His Cys (C) Ser Gin (Q) Asn Glu (E) ASP Gly (G) Ala ; Pro His (H) Asn; Gin Ile (I) Leu; Val ; Met; Nle ; Nva Leu (L); Val ; Met; Nle ; Nv Lys (K)

Arg; Gin ; Glu Met (M) Leu; Tyr; Ile ; NLe Val Ornitine Lys; Arg Phe (F) Met; Leu; Tyr Ser (S) Thr Thr (T) Ser Trp (W) Tyr Tyr (Y) Trp; Phe Val (V) ILE; Leu; Met; Nle ; Nv Other substitutions are also permissible and can be determined empirically or in accord with known conservative substitutions.

[0103] As used herein, Abu is 2-aminobutyric acid; Orn is ornithine. As used herein, the amino acids, which occur in the various amino acid sequences appearing herein, are identified according to their well-known, three- letter or one-letter abbreviations. The nucleotides, which occur in the various DNA fragments, are designated with the standard single-letter designations used routinely in the art.

[0104] As used herein, a probe or primer based on a nucleotide sequence disclosed herein, includes at least 10, 14, typically at least 16 contiguous sequence of nucleotides of SEQ ID NO. 6, and probes of at least 30, 50 or 100 contiguous sequence of nucleotides of SEQ ID NO. 6. The length of the probe or primer for unique hybridization is a function of the complexity of the genome of interest.

[0105] As used herein, amelioration of the symptoms of a particular disorder by administration of a particular pharmaceutical composition refers to any lessening, whether permanent or temporary, lasting or transient that can be attributed to or associated with administration of the composition.

[0106] As used herein, antisense polynucleotides refer to synthetic sequences of nucleotide bases complementary to mRNA or the sense strand of double- stranded DNA. Admixture of sense and antisense polynucleotides under appropriate conditions leads to the binding of the two molecules, or hybridization. When these polynucleotides bind to (hybridize with) mRNA, inhibition of protein synthesis (translation) occurs. When these polynucleotides bind to double-stranded DNA, inhibition of RNA synthesis (transcription) occurs.

[0107] The resulting inhibition of translation and/or transcription leads to an inhibition of the synthesis of the protein encoded by the sense strand. Antisense nucleic acid molecule typically contain a sufficient number of nucleotides to Specifically bind to a target nucleic acid, generally at least 5 contiguous nucleotides, often at least 14 or 16 or 30 contiguous

nucleotides or modified nucleotides complementary to the coding portion of a nucleic acid molecule that encodes a gene of interest, for example, nucleic acid encoding a single chain Hyaluronidase domain of an sHASEGP.

[0108] As used herein, an array refers to a collection of elements, such as antibodies, containing three or more members. An addressable array is one in which the members of the array are identifiable, typically by position on a solid phase support. Hence, in general the members of the array are immobilized on discrete identifiable loci on the surface of a solid phase.

[0109] As used herein, antibody refers to an immunoglobulin, whether natural or partially or wholly synthetically produced, including any derivative thereof that retains the specific binding ability the antibody. Hence antibody includes any protein having a binding domain that is homologous or substantially homologous to an immunoglobulin-binding domain. Antibodies include members of any immunoglobulin claims, including IgG, IgM, IgA, IgD and IgE.

[0110] As used herein, antibody fragment refers to any derivative of an antibody that is less then full-length, retaining at least a portion of the full-length antibody's specific binding ability. Examples of antibody fragments include, but are not limited to Fab, Fab', F(AB)2, single chain Fvs (scFV), FV, dsFV diabody and Fd fragments. The fragment can include multiple chains linked together, such as by disulfide bridges. An antibody fragment generally contains at least about 50 amino acids and typically at least 200 amino acids.

[0111] As used herein, an Fv antibody fragment is composed of one variable heavy domain (VH) and one variable light domain linked by noncovalent interactions.

[0112] As used herein, a dsFV refers to an Fv with an engineered intermolecular disulfide bond.

[0113] As used herein, an F (AB)2 fragment is an antibody fragment that results from digestion of an immunoglobulin with pepsin at pH 4.0-4.5; it can be recombinantly expressed to produce the equivalent fragment.

[0114] As used herein, Fab fragments are antibody fragments that result from digestion of an immunoglobulin with papain; they can be recombinantly expressed to produce the equivalent fragment.

[0115] As used herein, scFVs refer to antibody fragments that contain a variable light chain V, and variable heavy chain (VH) covalently connected by a polypeptide linker in any order. The linker is of a length such that the two variable domains are bridged without substantial interference. Included linkers are (Gly-Ser) n residues with some Glu or Lys residues dispersed throughout to increase solubility.

[0116] As used herein, humanized antibodies refer to antibodies that are modified to include human sequences of amino acids so that administration to a human does not provoke an immune response. Methods for preparation of such antibodies are known. For example, to produce such antibodies, the hybridoma or other prokaryotic or eukaryotic cell, such as an E. coli or a CHO cell, that expresses the monoclonal antibody are altered by recombinant DNA techniques to express an antibody in which the amino acid composition of the non-variable region is based on human antibodies. Computer programs have been designed to identify such regions.

[0117] As used herein, diabodies are dimeric scFV; diabodies typically have shorter peptide linkers than ScFVs, and they generally dimerize.

[0118] As used herein, production by recombinant means by using recombinant DNA methods means the use of the well known methods of molecular biology for expressing proteins encoded by cloned DNA.

[0119] As used herein the term assessing is intended to include quantitative and qualitative determination in the sense of obtaining an absolute value for the activity of an sHASEGP, or a domain thereof, present in the sample, and also of obtaining an index, ratio, percentage, visual or other value indicative of the level of the activity. Assessment can be direct or indirect and the chemical species actually detected need not of course be the proteolysis product itself but can for example be a derivative thereof or some further substance.

[0120] As used herein, biological activity refers to the in vivo activities of a compound or physiological responses that result upon in vivo administration of a compound, composition or other mixture. Biological activity, thus, encompasses therapeutic effects and pharmaceutical activity of such compounds, compositions and mixtures. Biological activities can be observed in in vitro systems designed to test or use such activities. Thus, for purposes herein the biological activity of a luciferase is its oxygenase activity whereby, upon oxidation of a substrate, light is produced.

[0121] As used herein, functional activity refers to a polypeptide or portion thereof that displays one or more activities associated with a full-length protein.

[0122] Functional activities include, but are not limited to, biological activity, catalytic or enzymatic activity, antigenicity (ability to bind to or compete with a polypeptide for binding to an anti-polypeptide antibody), immunogenicity, ability to form multimers, the ability to Specifically bind to a receptor or ligand for the polypeptide.

[0123] As used herein, a conjugate refers to the compounds provided herein that includes one or more sHASEGPs, including a sHASEGP, particularly single chain Hyaluronidase domains thereof, and one or more targeting agents. These conjugates include those produced by recombinant means as fusion proteins, those produced by chemical means, such as by chemical coupling, through, for example, coupling to sulfhydryl groups, and those produced by any other method whereby at least one sHASEGP, or a domain thereof, is linked, directly or indirectly via linker (s) to a targeting agent.

[0124] As used herein, a targeting agent is any moiety, such as a protein or effective portion thereof, that provides specific binding of the conjugate to a cell surface receptor, which, can internalize the conjugate or sHASEGP portion thereof. A targeting agent can also be one that promotes or facilitates, for example, affinity isolation or purification of the conjugate; attachment of the conjugate to a surface; or detection of the conjugate or complexes containing the conjugate.

[0125] As used herein, an antibody conjugate refers to a conjugate in which the targeting agent is an antibody.

[0126] As used herein, derivative or analog of a molecule refers to a portion derived from or a modified version of the molecule.

[0127] As used herein, an effective amount of a compound for treating a particular disease is an amount that is sufficient to ameliorate, or in some manner reduce the symptoms associated with the disease. Such amount can be administered as a single dosage or can be administered according to a regimen, whereby it is effective. The amount can cure the disease but, typically, is administered in order to ameliorate the symptoms of the disease. Repeated administration can be required to achieve the desired amelioration of symptoms.

[0128] As used herein equivalent, when referring to two sequences of nucleic acids means that the two sequences in question encode the same sequence of amino acids or equivalent proteins. When equivalent is used in referring to two proteins or peptides, it means that the two proteins or peptides have substantially the same amino acid sequence with only amino acid substitutions (such, as but not limited to, conservative changes such as those set forth in Table 1, above) that do not substantially alter the activity or function of the protein or peptide. When equivalent refers to a property, the property does not need to be present to the same extent (e. g., two peptides can exhibit different rates of the same type of enzymatic activity), but the activities are usually substantially the same. Complementary, when referring to two nucleotide sequences, means that the two sequences of nucleotides are capable of hybridizing, typically with less than 25%, 15%, 5% or 0% mismatches between opposed nucleotides. If necessary the percentage of complimentarity will be specified. Typically the two molecules are selected such that they will hybridize under conditions of high stringency.

[0129] As used herein, an agent that modulates the activity of a protein or expression of a gene or nucleic acid either decreases or increases or otherwise alters the activity of the protein or, in some manner up-or down-regulates or otherwise alters expression of the nucleic acid in a cell.

[0130] As used herein, inhibitor of the activity of an sHASEGP encompasses any substance that prohibits or decrease production, post-translational modification (s), maturation, or membrane localization of the sHASEGP or any substance that interferes

with or decreases the proteolytic efficacy of thereof, particularly of a single chain form in an in vitro screening assay.

[0131] As used herein, a method for treating or preventing neoplastic disease means that any of the symptoms, such as the tumor, metastasis thereof, the vascularization of the tumors or other parameters by which the disease is characterized are reduced, ameliorated, prevented, placed in a state of remission, or maintained in a state of remission. It also means that the hallmarks of neoplastic disease and metastasis can be eliminated, reduced or prevented by the treatment. Non-limiting examples of the hallmarks include uncontrolled degradation of the basement membrane and proximal extracellular matrix, migration, division, and organization of the endothelial cells into new functioning capillaries, and the persistence of such functioning capillaries.

[0132] As used herein, pharmaceutically acceptable salts, esters or other derivatives of the conjugates include any salts, esters or derivatives that can be readily prepared by those of skill in this art using known methods for such derivatization and that produce compounds that can be administered to animals or humans without substantial toxic effects and that either are pharmaceutical active or are prodrugs.

[0133] As used herein, a prodrug is a compound that, upon in vivo administration, is metabolized or otherwise converted to the biologically, pharmaceutically or therapeutically active form of the compound. To produce a prodrug, the pharmaceutical active compound is modified such that the active compound is regenerated by metabolic processes. The prodrug can be designed to alter the metabolic stability or the transport characteristics of a drug, to mask side effects or toxicity, to improve the flavor of a drug or to alter other characteristics or properties of a drug. By virtue of knowledge of pharmacodynamic processes and drug metabolism in vivo, those of skill in this art, once a pharmaceutically active compound is known, can design prodrugs of the compound (see, e. g., Nogrady (1985) Medicinal Chemistry A Biochemical Approach, Oxford University Press, New York, pages 388-392).

[0134] As used herein, a drug identified by the screening methods provided herein refers to any compound that is a candidate for use as a therapeutic or as a lead compound for the

design of a therapeutic. Such compounds can be small molecules, including small organic molecules, peptides, peptide mimetics, antisense molecules or dsRNA, such as RNAi, antibodies, fragments of antibodies, recombinant antibodies and other such compounds that can serve as drug candidates or lead compounds.

[0135] As used herein, a peptidomimetic is a compound that mimics the conformation and certain stereochemical features of the biologically active form of a particular peptide. In general, peptidomimetics are designed to mimic certain desirable properties of a compound, but not the undesirable properties, such as flexibility, that lead to a loss of a biologically active conformation and bond breakdown. Peptidomimetics may be prepared from biologically active compounds by replacing certain groups or bonds that contribute to the undesirable properties with bioisosteres. Bioisosteres are known to those of skill in the art. For example the methylene bioisostere CH2S has been used as an amide replacement in enkephalin analogs (see, e.g. Spatola (1983) pp. 267-357 in Chemistry and Biochemistry of Amino Acids, Peptides, and Proteins, Weistein, Ed. volume 7, Marcel Dekker, New York). Morphine, which can be administered orally, is a compound that is a peptidomimetic of the peptide endorphin. For purposes herein, cyclic peptides are included among pepidomimetics.

[0136] As used herein, a promoter region or promoter element refers to a segment of DNA or RNA that controls transcription of the DNA or RNA to which it is operatively linked. The promoter region includes specific sequences that are sufficient for RNA polymerase recognition, binding and transcription initiation.

[0137] This portion of the promoter region is referred to as the promoter. In addition, the promoter region includes sequences that modulate this recognition, binding and transcription initiation activity of RNA polymerase. These sequences can be cis acting or can be responsive to trans acting factors. Promoters, depending upon the nature of the regulation, can be constitutive or regulated. Exemplary promoters contemplated for use in prokaryotes include the bacteriophage T7 and T3 promoters.

[0138] As used herein, a receptor refers to a molecule that has an affinity for a given ligand. Receptors can be naturally occurring or synthetic molecules. Receptors can also be

referred to in the art as anti-ligands. As used herein, the receptor and anti-ligand are interchangeable. Receptors can be used in their unaltered state or as aggregates with other species. Receptors can be attached, covalently or noncovalently, or in physical contact with, to a binding member, either directly or indirectly via a specific binding substance or linker. Examples of receptors, include, but are not limited to: antibodies, cell membrane receptors surface receptors and internalizing receptors, monoclonal antibodies and antisera reactive with specific antigenic determinants such as on viruses, cells, or other materials], drugs, polynucleotides, nucleic acids, peptides, factors, lectins, sugars, polysaccharides, cells, cellular membranes, and organelles.

[0139] Examples of receptors and applications using such receptors, include but are not restricted to: a) enzymes: specific transport proteins or enzymes essential to survival of microorganisms, which could serve as targets for antibiotic [ligand] selection; b) antibodies: identification of a ligand-binding site on the antibody molecule that combines with the epitope of an antigen of interest can be investigated; determination of a sequence that mimics an antigenic epitope can lead to the development of vaccines of which the immunogen is based on one or more of such sequences or lead to the development of related diagnostic agents or compounds useful in therapeutic treatments such as for auto-immune diseases c) nucleic acids: identification of ligand, such as protein or RNA, binding sites; d) catalytic polypeptides : polymers, including polypeptides, that are capable of promoting a chemical reaction involving the conversion of one or more reactants to one or more products; such polypeptides generally include a binding site specific for at least one reactant or reaction intermediate and an active functionality proximate to the binding site, in which the functionality is capable of chemically modifying the bound reactant (see, e. g., U. S. Patent No. 5,215,899); e) hormone receptors: determination of the ligands that bind with high affinity to a receptor is useful in the development of hormone replacement therapies; for example, identification of ligands that bind to such receptors can lead to the development of drugs to control blood pressure; and f) opiate receptors: determination of ligands that bind to the opiate receptors in the brain is useful in the development of less-addictive replacements for morphine and related drugs.

[0140] As used herein, sample refers to anything that can contain an analyte for which an analyte assay is desired. The sample can be a biological sample, such as a biological fluid

or a biological tissue. Examples of biological fluids include urine, blood, plasma, serum, saliva, semen, stool, sputum, cerebral spinal fluid, tears, mucus, sperm, amniotic fluid or the like. Biological tissues are aggregate of cells, usually of a particular kind together with their intercellular substance that form one of the structural materials of a human, animal, plant, bacterial, fungal or viral structure, including connective, epithelium, muscle and nerve tissues. Examples of biological tissues also include organs, tumors, lymph nodes, arteries and individual cells.

[0141] As used herein: stringency of hybridization in determining percentage mismatch is as follows: 1) high stringency: 0.1 x SSPE, 0.1 % SDS, 65°C 2) medium stringency: 0.2 x SspE, 0.1 % SDS, 50°C 3 low stringency: 1.0 x SspE, 0.1 % SDS, 50°C. Those of skill in this art know that the washing step selects for stable hybrids and also know the ingredients of SspE (see, e. g., Sambrook, E. F. Fritsch, T. Maniatis, in: Molecular Cloning, A Laboratory Manual, Cold spring Harbor Laboratory Press 1989 Vol 3, p. B. 13, see, also, numerous catalogs that describe commonly used laboratory solutions). SspE is pH 7.4 phosphate- buffered 0.18 NaCl. Further, those of skill in the art recognize that the stability of hybrids is determined by TmT which is a function of the sodium ion concentration and temperature (Tm = 81.5° C-16.6 + 0.41 (% G+C)-600/L)) so that the only parameters in the wash conditions critical to hybrid stability are sodium ion concentration in the SspE (or SSC) and temperature.

[0142] It is understood that equivalent stringencies can be achieved using alternative buffers, salts and temperatures. By way of example and not limitation, procedures using conditions of low stringency are as follows (see also Shilo and Weinberg, Proc. Natl. Acad Sci USA 78: 6789-6792 (1981)): Filters containing DNA are pretreated for 6 hours at 40 C in a solution containing 35% formamide, 5X SSC, 50 mM Tris-HCl (pH 7.5), 5 mM EDTA, 0.1 % PVP, 0.1 % Ficoll 1% BSA, and 500ug/ml Denatured Salmon sperm DNA (10x) SSC is 1.5 M sodium chloride, and 0.15 M sodium citrate, adjusted to a pH of 7).

[0143] Hybridizations are carried out in the same solution with the following modifications: 0.02% PVP, 0.02% Ficoll 0.2% BSA, 100VG/M sperm DNA, 10% (wt/vol) dextran sulfate, and 5-20 X 106 cpm 32P-labled probe is used. Filters are incubated in hybridization mixture for 18-20 hours at 40C and then washed for 1.5 hours

at 55C in a solution containing 2X SSC, 25 mM Tris-HCI (pH 7.4), 5 mM EDTA, and 0.1 % SDS. The wash solution is replaced with fresh solution and incubated an additional 1.5 hours at 60C. Filters are blotted dry and exposed for autoradiography. If necessary, filters are washed for a third time at 65-68C and reexposed to film. Other conditions of low stringency which can be used are well known in the art e.g. as employed for cross-species hybridizations).

[0144] By way of example and not way of limitation, procedures using conditions of moderate stringency include, for example, but are not limited to, procedures using such conditions of moderate stringency are as follows: Filters containing DNA are pretreated for 6 hours at 55C in a solution containing 6X SSC, 5X Denhart's solution, 0.5% SDS and 100ug/ml denatured salmon sperm DNA. Hybridizations are carried out in the same solution and 5-20 X 106 32P labeled probe is used. Filters are incubated in hybridization mixture for 18-20 hours at 55C and then washed twice for 30 minutes at 60C in a solution containing 1X SSC and 0.1% SDS. Filters are blotted dry and exposed for autoradiography. Other conditions of moderate stringency that can be used are well known in the art. Washing of filters is done at 37C for 1 hour in a solution containing 2X SSC, 0.1 % SDS.

[0145] By way of example and not way of limitation, procedures using conditions of high stringency are as follows: Prehybridization of filters containing DNA is carried out for 8 hours to overnight at 65C in buffer composed of 6X SSC, 50 mM Tris-HCI (pH 7.5), 1 mM EDTA, 0.02% PVP, 0.02% Ficoll, 0.02% BSA, and 500ug/ml denatured salmon sperm DNA. Filters are hybridized for 48 hours at 65°C in prehybridization mixture containing 100ug/ml denatured salmon sperm DNA and 5-20 X 106 CPM 32P labeled probe. Washing of filters is done at 37C for 1 hour in a solution containing 2X SSC, 0.01 % PVP, 0.01 % Ficoll, and 0.01 % BSA. This is followed by a wash in 0.1X SSC at 50C for 45 minutes before autoradiography. Other conditions of high stringency that can be used are well known in the art.

[0146] The term substantially identical or substantially homologous or similar varies with the context as understood by those skilled in the relevant art and generally means at least

60% or 70%, preferably means at least 80%, 85% or more preferably at least 90%, and most preferably at least 95% identity.

[0147] As used herein, substantially identical to a product means sufficiently similar so that the property of interest is sufficiently unchanged so that the substantially identical product can be used in place of the product.

[0148] As used herein, substantially pure means sufficiently homogeneous to appear free of readily detectable impurities as determined by standard methods of analysis, such as thin layer chromatography (TLC), gel electrophoresis and high performance liquid chromatography (HPLC), used by those of skill in the art to assess such purity, or sufficiently pure such that further purification would not detectably alter the physical and chemical properties, such as enzymatic and biological activities, of the substance. Methods for purification of the compounds to produce substantially chemically pure compounds are known to those of skill in the art. A substantially chemically pure compound can, however, be a mixture of stereoisomers or isomers. In such instances, further purification might increase the specific activity of the compound.

[0149] As used herein, target cell refers to a cell that expresses a sHASEGP in vivo.

[0150] As used herein, test substance (or test compound) refers to a chemically defined compound (e.g., organic molecules, inorganic molecules, organic/inorganic molecules, proteins, peptides, nucleic acids, oligonucleotides, lipids, polysaccharides, saccharides, or hybrids among these molecules such as glycoproteins, etc.) or mixtures of compounds (e.g., a library of test compounds, natural extracts or culture supernatants, etc.) whose effect on an sHASEGP, particularly a single chain form that includes the Hyaluronidase domain or a sufficient portion thereof for activity, as determined by an in vitro method, such as the assays provided herein.

[0151] As used herein, the terms a therapeutic agent, therapeutic regimen, radioprotectant or chemotherapeutic mean conventional drugs and drug therapies, including vaccines, which are known to those skilled in the art. Radiotherapeutic agents are well known in the art.

[0152] As used herein, treatment means any manner in which the symptoms of a condition, disorder or disease are ameliorated or otherwise beneficially altered.

[0153] Treatment also encompasses any pharmaceutical use of the compositions herein.

[0154] As used herein, vector (or plasmid) refers to discrete elements that are used to introduce heterologous nucleic acid into cells for either expression or replication thereof. The vectors typically remain episomal, but can be designed to effect integration of a gene or portion thereof into a chromosome of the genome. Also contemplated are vectors that are artificial chromosomes, such as yeast artificial chromosomes and mammalian artificial chromosomes. Selection and use of such vehicles are well known to those of skill in the art. An expression vector includes vectors capable of expressing DNA that is operatively linked with regulatory sequences, such as promoter regions, that are capable of effecting expression of such DNA fragments. Thus, an expression vector refers to a recombinant DNA or RNA construct, such as a plasmid, a phage, recombinant virus or other vector that, upon introduction into an appropriate host cell, results in expression of the cloned DNA. Appropriate expression vectors are well known to those of skill in the art and include those that are repliable in eukaryotic cells and/or prokaryotic cells and those that remain episomal or those which integrate into the host cell genome.

[0155] As used herein, protein binding sequence refers to a protein or peptide sequence that is capable of specific binding to other protein or peptide sequences generally, to a set of protein or peptide sequences or to a particular protein or peptide sequence.

[0156] As used herein, epitope tag refers to a short stretch of amino acid residues corresponding to an epitope to facilitate subsequent biochemical and immunological analysis of the epitope tagged protein or peptide. Epitope tagging is achieved by including the sequence of the epitope tag to the protein-encoding sequence in an appropriate expression vector. Epitope tagged proteins can be affinity purified using highly specific antibodies raised against the tags.

[0157] As used herein, metal binding sequence refers to a protein or peptide sequence that is capable of specific binding to metal ions generally, to a set of metal ions or to a particular metal ion.

[0158] As used herein, a combination refers to any association between two or among more items.

[0159] As used herein, a composition refers to any mixture. It can be a solution, a suspension, liquid, powder, a paste, aqueous, non-aqueous or any combination thereof.

[0160] As used herein, fluid refers to any composition that can flow. Fluids thus encompass compositions that are in the form of semi-solids, pastes, solutions, aqueous mixtures, gels, lotions, creams and other such compositions.

[0161] As used herein, a cellular extract refers to a preparation or fraction which is made from a lysed or disrupted cell.

[0162] As used herein, an agent is said to be randomly selected when the agent is chosen randomly without considering the specific sequences involved in the association of a protein alone or with its associated substrates, binding partners, etc. An example of randomly selected agents is the use a chemical library or a peptide combinatorial library, or a growth broth of an organism or conditioned medium.

[0163] As used herein, an agent is said to be rationally selected or designed when the agent is chosen on a non-random basis that takes into account the sequence of the target site and/or its conformation in connection with the agent's action. As described in the Examples, there are proposed binding sites for Hyaluronidase and (catalytic) sites in the glycoprotein having SEQ ID NO: 1 or SEQ ID NO: 4. Agents can be rationally selected or rationally designed by utilizing the peptide sequences that make up these sites. For example, a rationally selected peptide agent can be a peptide whose amino acid sequence is identical to the ATP or calmodulin binding sites or domains.

[0164] Oligosaccharides are considered to have a reducing end and a non-reducing end, whether or not the saccharide at the reducing end is in fact a reducing sugar. In accordance with accepted nomenclature, oligosaccharides are depicted herein with the non-reducing end on the left and the reducing end on the right. All oligosaccharides described herein are described with the name or abbreviation for the non-reducing saccharide (e.g., Gal), followed by the configuration of the glycosidic bond (.alpha. or. beta.), the ring bond, the

ring position of the reducing saccharide involved in the bond, and then the name or abbreviation of the reducing saccharide (e.g., GlcNAc). The linkage between two sugars may be expressed, for example, as 2,3, 2.fw darw.3, or (2,3). Each saccharide is a pyranose.

[0165] As used herein, N-linked sugar moiety refers to an oligosaccharide attached to a sHASEGP via the amide nitrogen of Asn residues. N-linked oligosaccharides fall into several major types (oligomannose, complex, hybrid, sulfated), all of which have (Man) 3-GlcNAc-GlcNAc-cores attached via the amide nitrogen of Asn residues that fall within -Asn-Xaa-Thr/Ser- sequences (where Xaa is not Pro). N-linked sites are often indirectly assigned by the appearance of a "blank" cycle during sequencing. Positive identification can be made after release of the oligosaccharide by PNGase F, which converts the glycosylated Asn to Asp. After PNGase F release, N-linked oligosaccharides can be purified using Bio-Gel P-6 chromatography, with the oligosaccharide pool subjected to preparative high pH anion exchange chromatography (HPAEC) (Townsend et al., (1989) Anal. Biochem. 182, 1-8). Certain oligosaccharide isomers can be resolved using HPAEC. Fucose residues will shift elution positions earlier in the HPAEC chromatogram, while additional sialic acid residues will increase the retention time. Concurrent treatment of glycoproteins whose oligosaccharide structures are known (e.g., bovine fetuin, a-l acid glycoprotein, ovalbumin, RNAse B, transferrin) can facilitate assignment of the oligosaccharide peaks. The collected oligosaccharides can be characterized by a combination of compositional and methylation linkage analyses (Waegheet al., (1983) Carbohydr Res. 123, 281-304.), with anomeric configurations assigned by NMR spectroscopy (Van Halbeek (1993) in Methods Enzymol 230).

[0166] Alternatively, oligosaccharides can be identified by fluorescence assisted carbohydrate electrophoresis (FACE) Callewaert et al. (2001) Glycobiology 11, 275-281.

[0167] As used herein, the term "sialic acid" refers to any member of a family of nine-carbon carboxylated sugars. The most common member of the sialic acid family is N-acetylneuraminic acid (2-keto-5-acetamindo-3, 5-dideoxy-D-glycero-D-galactononulopyranos-1-onic acid (often abbreviated as Neu5Ac, NeuAc, or NANA). A second member of the family is N-glycolyl-neuraminic acid (Neu5Gc or NeuGc), in which

the N-acetyl group of NeuAc is hydroxylated. A third sialic acid family member is 2-keto-3-deoxy-nonulosonic acid (KDN) (Nadano et al. (1986) J. Biol. Chem. 261: 11550-11557; Kanamori et al. (1990) J. Biol. Chem. 265: 21811-21819. Also included are 9-substituted sialic acids such as a 9-O-C.sub.1 -C.sub.6 acyl-Neu5Ac like 9-O-lactyl-Neu5Ac or 9-O-acetyl-Neu5Ac, 9-deoxy-9-fluoro-Neu5Ac and 9-azido-9-deoxy-Neu5Ac. For review of the sialic acid family, see, e.g., Varki (1992) Glycobiology 2: 25-40; Sialic Acids: Chemistry, Metabolism and Function, R. Schauer, Ed. (Springer-Verlag, N.Y. (1992)). The synthesis and use of sialic acid compounds in a sialation procedure is disclosed in international application WO 92/16640, published Oct. 1, 1992.

[0168] As used herein, PNGase refers to an Asparagine Peptide specific N-glycosidase F such as the Flavobacterium maningoseptum peptide-N-glycosidase F. PNGASE enzymes are characterized by their specificity towards N-linked rather than O-linked oligosaccharides. Characterization of PNGASE efficacy can be defined by both SDS PAGE electrophoresis, or fluorescent assisted carbohydrate electrophoresis.

[0169] As used herein substantially terminated Sialation refers to N-linked oligosaccharides terminating with sialic acid residue as a terminal sugar. Terminal sialic acids can be identified by FACE analysis of released carbohydrates following treatment with neuraminidase.

[0170] The circulatory lifetime of glycoproteins in the blood is highly dependent on the composition and structure of its N-linked carbohydrate groups. This fact is of direct relevance for therapeutic glycoproteins that are intended to be administered parenterally. In general, maximal circulatory half-life of a glycoprotein requires that its N-linked carbohydrate groups terminate in the sequence NeuAc-Gal-GlcNAc. Without the terminal sialic acid (NeuAc), the glycoprotein is rapidly cleared from the blood by a mechanism involving the recognition of the underlying N-acetylgalactosamine (GalNAc) or galactose (Gal) residues (Goochee et al. (1991) Biol/Technology 9: 1347-1355). For this reason, ensuring the presence of terminal sialic acid on N-linked carbohydrate groups of therapeutic glycoproteins is an important consideration for their commercial development.

[0171] Circulating glycoproteins are exposed to sialidase(s) (or neuraminidase) which can remove terminal sialic acid residues. Typically the removal of the sialic acid exposes galactose residues, and these residues are recognized and bound by galactose-specific receptors in hepatocytes (reviewed in Ashwell and Harford (1982) Ann. Rev. Biochem. 51:531). Liver also contains other sugar-specific receptors which mediate removal of glycoproteins from circulation. Specificities of such receptors also include N-acetylglucosamine, mannose, fucose and phosphomannose. Glycoproteins cleared by the galactose receptors of hepatocytes undergo substantial degradation and then enter the bile; glycoproteins cleared by the mannose receptor of Kupffer cells enter the reticuloendothelial system (reviewed in Ashwell and Harford (1982) Ann. Rev. Biochem. 51:53).

[0172] As used herein Neutral Active refers to a sHASEGP glycoprotein with catalytic activity towards a glycosaminoglycan substrate in vitro at a PH between 5 and 8 under conditions of salt less than 150mM and buffering strength less than 50mM.

[0173] As used herein, a stabilized solution refers to a sHASEGP that retains greater than 60% of its initial activity after storage at room temperature for 30 days.

[0174] As used herein unless otherwise specified, a unit is expressed in turbidity reducing units (TRU). One TRU is defined as the amount of hyaluronidase activity required to reduce the turbidity of an acidified solution of hyaluronic acid and is equivalent to the U.S.P./National Formulary (NF XIII) units (NFU). The ELISA-like enzyme assay described herein can be related to the TRU, the NFU, and U.S.P. unit through a standard curve of a sample of hyaluronidase (e.g., USP or WHO standard) standardized through the U.S.P. Therefore, the enzyme activities determined by the ELISA-like enzyme assay are actually relative TRU, since enzyme activity is not actually measured using the turbidometric assay (Dorfman et al., 1948, J. Biol. Chem. 172:367).

[0175] As used herein, potency is defined by the amount of sHASEGP protein required to degrade substrate *in vitro* based upon a Turbidity Reducing Unit or Relative Turbidity Reducing Unit.

[0176] As used herein, specific activity refers to Units of activity per mg protein. The amount of sHASEGP protein is defined by the absorption of a solution of sHASEGP at 280 nm assuming a molar extinction coefficient of approximately 1.7, in units of M^{-1} cm^{-1}.

[0177] Polyethylene glycol (PEG) has been widely used in biomaterials, biotechnology and medicine primarily because PEG is a biocompatible, nontoxic, nonimmunogenic and water-soluble polymer (Zhao and Harris, ACS Symposium Series 680: 458-72, 1997). In the area of drug delivery, PEG derivatives have been widely used in covalent attachment (i.e., "PEGylation") to proteins to reduce immunogenicity, proteolysis and kidney clearance and to enhance solubility (Zalipsky, Adv. Drug Del. Rev. 16:157-82, 1995). Similarly, PEG has been attached to low molecular weight, relatively hydrophobic drugs to enhance solubility, reduce toxicity and alter biodistribution. Typically, PEGylated drugs are injected as solutions.

[0178] A closely related application is synthesis of crosslinked degradable PEG networks or formulations for use in drug delivery since much of the same chemistry used in design of degradable, soluble drug carriers can also be used in design of degradable gels (Sawhney et al., Macromolecules 26: 581-87, 1993). It is also known that intermacromolecular complexes can be formed by mixing solutions of two complementary polymers. Such complexes are generally stabilized by electrostatic interactions (polyanion-polycation) and/or hydrogen bonds (polyacid-polybase) between the polymers involved, and/or by hydrophobic interactions between the polymers in an aqueous surrounding (Krupers et al., Eur. Polym J. 32:785-790, 1996). For example, mixing solutions of polyacrylic acid (PAAc) and polyethylene oxide (PEO) under the proper conditions results in the formation of complexes based mostly on hydrogen bonding. Dissociation of these complexes at physiologic conditions has been used for delivery of free drugs (i.e., non-PEGylated). In addition, complexes of complementary polymers have been formed from both homopolymers and copolymers.

[0179] In one aspect, the polyethylene glycol has a molecular weight ranging from about 3 kD to about 50 kD, and preferably from about 5 kD to about 30 kD. Covalent attachment of the PEG to the drug (known as "PEGylation") may be accomplished by known chemical synthesis techniques. For example, in one aspect of the present invention, the

PEGylation of protein may be accomplished by reacting NHS-activated PEG with the protein under suitable reaction conditions.

[0180] While numerous reactions have been described for PEGylation, those that are most generally applicable confer directionality, utilise mild reaction conditions, and do not necessitate extensive downstream processing to remove toxic catalysts or bi-products. For instance, monomethoxyPEG (mPEG) has only one reactive terminal hydroxyl, and thus its use limits some of the heterogeneity of the resulting PEG-protein product mixture. Activation of the hydroxyl group at the end of the polymer opposite to the terminal methoxy group is generally necessary to accomplish efficient protein PEGylation, with the aim being to make the derivatised PEG more susceptible to nucleophilic attack. The attacking nucleophile is usually the epsilon-amino group of a lysyl residue, but other amines can also react (e.g. the N-terminal alpha-amine or the ring amines of histidine) if local conditions are favorable. A more directed attachment is possible in proteins containing a single lysine or cysteine. The latter residue can be targeted by PEG-maleimide for thiol-specific modification. Alternatively, PEG hydrazide can be reacted with periodate oxidized sHASEGP and reduced in the presence of $NaCNBH_3$. More specifically, PEGylated CMP sugars can be reacted with sHASEGP in the presence of appropriate glycosyl-transferases.One technique is the "PEGylation" technique where a number of polymeric molecules are coupled to the polypeptide in question. When using this technique the immune system has difficulties in recognizing the epitopes on the polypeptide's surface responsible for the formation of antibodies, thereby reducing the immune response. For polypeptides introduced directly into the circulatory system of the human body to give a particular physiological effect (i.e. pharmaceuticals) the typical potential immune response is an IgG and/or IgM response, while polypeptides which are inhaled through the respiratory system (i.e. industrial polypeptide) potentially may cause an IgE response (i.e. allergic response). One of the theories explaining the reduced immune response is that the polymeric molecule(s) shield(s) epitope(s) on the surface of the polypeptide responsible for the immune response leading to antibody formation. Another theory or at least a partial factor is that the heavier the conjugate is, the more reduced immune response is obtained.

[0181] The polymeric molecules coupled to the polypeptide may be any suitable polymeric molecule with a molecular weight as defined according to the invention, including natural and synthetic homopolymers, such as polyols (i.e. poly-OH), polyamines (i.e. poly-NH.sub.2) and polycarboxyl acids (i.e. poly-COOH), and further heteropolymers i.e. polymers comprising one or more different coupling groups e.g. a hydroxyl group and amine groups.

[0182] Examples of suitable polymeric molecules include polymeric molecules selected from the group comprising polyalkylene oxides (PAO), such as polyalkylene glycols (PAG), including polypropylene glycols (PEG), methoxypolyethylene glycols (mPEG) and polypropylene glycols, PEG-glycidyl ethers (Epox-PEG), PEG-oxycarbonylimidazole (CDI-PEG) branched polyethelene glycols (PEGs), polyvinyl alcohol)PVA), polycarboxylates, polyvinylpyrrolidone, poly-D,L-amino acids, polyethylene-co-maleic acid anhydride, polystyrene-co-malic acid arhydride, dextrans including carboxymethyl-dextrans, heparin, homologous albumin, celluloses, including methylcellulose, carboxymethylcellulose, ethylcellulosia, hydroxyethylcellulose carboxyethylcellulose and hydroxypropylcellulose, hydrolysates of chitosan, starches such as hydroxyethyl-starches and hydroxy propyl-starches, glycogen, agaroses and derivatives thereof, guar gum, pullulan, inulin, xanthan gum, carrageenan, pectin, alginic acid hydrolysates and bio-polymers.

[0183] Preferred polymeric molecules are non-toxic polymeric molecules such as (m)polyethylene glycol (mPEG) which further requires a relatively simple chemistry for its covalent coupling to attachment groups on the enzyme's surface.

[0184] Generally seen polyalkylene oxides (PAO), such as polyethylene oxides, such as PEG and especially mPEG, are the preferred polymeric molecules, as these polymeric molecules, in comparison to polysaccharides such as dextran, pullulan and the like, have few reactive groups capable of cross-linking, which is undesirable.

B. TISSUE EXPRESSION PROFILES sHASEGP.

[0185] While previously thought to be testis specific, human sHASEGP is expressed in multiple tissues in humans when using more sensitive techniques such as RT-PCR. The

sHASEGP transcript is found in medulla (brain), microvascular endothelium, prostate, breast, retina, pooled human melanocyte, fetal heart, and pregnant uterus. sHASEGP is also expressed in germ cell tumors. RT-PCR based detection of sHASEGP transcripts is generally required to detect levels in tissues other than testis.

C. ASSAYS FOR sHASEGP ENZYME ACTIVITY

[0186] TURBIDOMETRIC MICROTITER ASSAY FOR HYALURONIDASE ACTIVITY

[0187] Hyaluronidase activity can be detected by way of a modified turbidometric assay in acidified serum solution. The reagents required are as follows:

UV sterilized 2X-deionized water or sterile water for irrigation	Braun	R5000-01
Hylumed Medical – Sodium Hyaluronate, High Molecular Weight HA	Genzyme Advanced Biomaterials	4876
Hyaluronidase Reference Standard	USP	31200
Potassium Acetate, Granular, USP, ACS	JTBaker	2914-01
Acetic Acid, Glacial, 99+ %	Sigma	A-6283
Sodium Phosphate Monobasic Monohydrate, USP Granular	Mallinkrodt	7774
Sodium Phosphate Dibasic Anhydrous, USP	Mallinkrodt	7771
Sodium Chloride, Crystals, GR, ACS	EMScience	SX0420-5
Gelatin Hydrolysate Enzymatic	Sigma	G-0262
Horse Serum, Donor Herd, cell culture tested, Hybridoma culture tested, USA Origin	Sigma	H-1270
Human Serum Albumin 20 %	Griffols	
Hydrochloric Acid, ACS Reagent	Sigma	H-7020
Calcium Chloride, Dihydrate, Granular, USP, -FCC	JTBaker	1336-01

[0188] The following reagents are prepared: Acetate Buffer Solution- 14.0 g of potassium acetate and 25.0 mL of glacial acetic acid in water to make 1000 mL. Phosphate Buffer Solution- 2.5 g of sodium phosphate monobasic, 1.0 g of anhydrous sodium phosphate dibasic, and 8.2 g of sodium chloride in water to make 1000 mL. Enzyme Diluent Stock Solution- 500 mL of Phosphate Buffer Solution with 500 mL of water. Enzyme Diluent Working Solution- 33 mg of hydrolyzed gelatin in 50 mL of Enzyme Diluent Stock Solution-prepared within 2 hours of use. Sample Stabilization Buffer Solution ("SSB" Soln.)- 125 uL of a 20% Human Serum Albumin Solution and 50 uL of a 1 M Calcium Chloride solution in 50 mL of Enzyme Diluent Working Solution, and mix thoroughly. Serum Stock Solution-Dilute 1 volume of Horse Serum with 9 volumes of Acetate Buffer Solution. Adjust with 4 N hydrochloric acid to a pH of 3.1 and allow the solution to stand at room temperature for 18 to 24 hrs. Store the solution at 4 °C, and use within 30 days. Serum Working Solution- 10 mL of the Serum Stock Solution in 30 mL of the Acetate Buffer Solution, adjusted to room temperature. Hyaluronic Acid Stock Solution- Sodium Hyaluronic Acid to a concentration 5.0 mg/mL in water. Hyaluronic Acid Working Solution- 0.75 mL of the Hyaluronic Acid Stock Solution in 4.25 mL of the Phosphate Buffer Solution. Standard Stock Solution- One container of USP Reference Standard Hyaluronidase to a concentration 1000 Units/mL in water, aliquoted into 50 uL portions, and stored at -20 °C. Standard Working Solution- 40 uL of Standard Stock Solution in 960 uL of cold Enzyme Diluent Working Solution to obtain a solution having a known concentration of 40 Units/mL, prepared immediately before use in the assay.

[0189] All enzyme samples are diluted in a "Low Protein Binding" 96-well plate according to the following guidelines:

[0190] a) The range of maximum sensitivity of this assay is between 10-30 Units/mL. To minimize the number of times an assay must be repeated in order to get results that are within range, first determine the approximate number of total units/mL for the sample, and then choose a (whole number) dilution such that the final concentration is approximately 20 Units/ml.

[0191] b) Minimum Sample volumes needed to perform assay are as follows: FPLC Fractions=50 uL, Tissue Culture Supernatants=1 mL, Purified/Concentrated/Final Step Material=10 uL.

[0192] c) For samples with serial dilutions, 1:10 dilutions in the "Low Protein Binding" 96- well plate are made in triplicate by pipetting 360 uL of the "SSB" Solution and 40 uL of Sample into each well.

[0193] For preparation of USP Standard prepare the USP Standard Curve in the "Low Protein Binding" 96-well plate as follows:

USP Standard Curve:

Wells:	Standard:	Enzyme Diluent Soln.(in uL):	Standard Working Soln.(in uL):	Final Conc.(in Units/mL):
A1-A3	St01	0	100	40
B1-B3	St02	20	80	32
C1-C3	St03	40	60	24
D1-D3	St04	60	40	16
E1-E3	St05	80	20	8
F1-F3	St06	90	10	4
G1-G3	St07	100	0	0

[0194] For preparation of the Hyaluronic Acid Control in columns 1-3, prepare the H.A. Control in the "Flat Bottom" 96-well plate is prepared as follows:

[0195]

H.A. Controls:

Wells:	Control:	Hyaluronic Acid Working Soln.(in uL):	Enzyme Diluent Working Soln.(in uL):
H1-H3	Co01	0	60

[0196] The Reaction Plate: 30 uL per well of Hyaluronic Acid Working Solution is pippetted using a 50 uL 8- channel transfer pipette into a "Flat Bottom" 96-well microtiter plate leaving wells H1-H3 empty. 60 uL/well of Enzyme Diluent Working Solution is pipetted into wells H1-H3 of the same plate as the HA control.

[0197] Serum Working Solution: 40mL of Serum Working Solution is dispensed into a transfer basin and next to the Heat Block.

[0198] Pre-warming stage: Once both plates have been prepared, the Low Protein Binding 96-Well plate containing the diluted samples, standards, controls and the Flat Bottom 96-well plate containing the Hyaluronic Acid Working Solution are placed onto a heat block and allow them to warm for 5 min. at 37 °C.

[0199] The Reaction is initiated by the addition of Enzyme to Substrate: 30 uL from the enzyme plate into all of the wells in column #1 of the 96-Well flat bottom plate (containing the substrate) using a 5-50 uL 8-channel pipette. The Enzyme/Substrate reaction mixture is aspirated 5 times (drawing the solution up and down with the transfer during the first 15 seconds to ensure complete sample mixing. After mixing the enzyme and substrate, the tips are ejected and a new set of tips loaded on the transfer pipettor for the next column. A timer is restarted, and at time (t)=0:30, this process is repeated for column 2. At the next 30 second interval (t)=1:00, this is repeated process for column 3. This process is repeated moving from left to right across the plate, every 30 seconds until all of the wells contain both enzyme and substrate.

[0200] Stopping the reaction: When timer reaches 6 minutes (t)=6:00, 240 uL of the Serum Working Solution is pippetted into each well, using a 50-300 uL 8-channel transfer pipette, into column 1 of the 96-well flat bottom plate from the adjacent 50 mL Reagent Reservoir. The mixture is aspirated 3 times (drawing the solution up and down with the transfer Pipettor) during the first 10 seconds to ensure complete mixing. The process is repeated every 30 seconds, proceeding from column's 1 to 12 . Upon completion of the last column (column 12), the reaction plate is removed from the heat block and place the plate onto the read tray of the plate reader at 640 nM. A linear curve fit is generated from the standard curve that permits extrapolation of test samples.

[0201] ALTERNATIVE ASSAYS FOR HYALURONIDASE

[0202] BIOTINYLATED HYALURONAN MICROTITER ASSAY

[0203] The free carboxyl groups on glucuronic acid residues of Hyaluronan are biotinylated in a one step reaction using biotin-hydrazide (Pierce), Sulfo NHS (Pierce) and 1-Ethyl dimethylaminopropyl-carbodiimide (Sigma). This biotinylated HA substrate is

covalently coupled to a 96 well microtiter plate in a second reaction. At the completion of the enzyme reaction, residual substrate is detected with an avidin-peroxidase reaction that can be read in a standard ELISA plate reader. As the substrate is covalently bound to the microtiter plate, artifacts such as pH-dependent displacement of the biotinylated substrate does not occur. The sensitivity permits rapid measurement of Hyaluronidase activity from cultured cells and biological samples with an inter-assay variation of less than 10%.

[0204] The specific activity of hyaluronidase is expressed in turbidity reducing units (TRU). One TRU is defined as the amount of hyaluronidase activity required to reduce the turbidity of an acidified solution of hyaluronic acid and is equivalent to the U.S.P./National Formulary (NF XIII) units (NFU). The ELISA-like enzyme assay used for purification is related to the TRU, the NFU, and U.S.P. unit through a standard curve of a sample of hyaluronidase (e.g., USP) standardized through the U.S.P. Therefore, the enzyme activities determined by the ELISA-like enzyme assay are actually relative TRU, since enzyme activity is not actually measured using the turbidometric assay (Dorfman et al., 1948, J. Biol. Chem. 172:367).

[0205] Many Hyaluronidase assays have been based upon the measurement of the generation of new reducing N-acetylamino groups (Bonner and Cantey, *Clin. Chim. Acta* 13:746-752, 1966), or loss of viscosity (De Salegui et al., *Arch. Biochem. Biophys.* 121:548-554, 1967) or turbidity (Dorfman and Ott, *J. Biol. Chem.* 172:367, 1948). With purified substrates all of these methods suffice for determination of the presence or absence of endoglucosamidic activity.

[0206] Substantially purified glycosaminoglycan substrates can also be used for in a Gel Shift Assay. Glycosaminoglycans are mixed with recombinant sHASEGP to test for endoglucosidase activity that results in a shift in substrate mobility within the gel. Chondroitin-4 and 6 sulfate, dermatan sulfate, heparan-sulfate can be obtained from Sigma Chemical. Human umbilical cord Hyaluronan can be obtained from ICN. Each test substrate is diluted to 0.1mg/ml in a buffer range from pH 3.5-7.5. 10ul samples of purified sHASEGP or conditioned media from sHASEGP expressing cells as well as are mixed with 90ul of test substrate in desired buffer and incubated for 3 hours at 37C. Following incubation samples are neutralized with sample buffer (Tris EDTA PH 8.0,

Bromophenol Blue and glycerol) followed by electrophoresis. Glycosaminoglycans are detected by staining the gels in 0.5% Alcian Blue in 3% Glacial Acetic Acid overnight followed by destaining in 7% Glacial Acetic Acid. Degradation is determined by comparison substrate mobility in the presence and absence of enzyme.

[0207] Hyaluronidase activity can also be detected by substrate gel zymography (Guentenhoner et al., 1992, Matrix 388-396). In this assay a sample is applied to a SDS-PAGE gel containing hyaluronic acid and the proteins in the sample separated by electrophoresis. The gel is then incubated in an enzyme assay buffer and subsequently stained to detect the hyaluronic acid in the gel. Hyaluronidase activity is visualized as a cleared zone in the substrate gel.

[0208] D. IDENTIFICATION AND ISOLATION OF sHASEGP POLYPEPTIDE GENES.

[0209] The sHASEGP polypeptide gene and/or domains thereof, can be obtained by methods well known in the art for DNA isolation. Any method known to those of skill in the art for identification of nucleic acids that encode desired genes can be used. Any method available in the art can be used to obtain a full-length (i. e., encompassing the entire coding region) cDNA or genomic DNA clone encoding a sHASEGP polypeptide. For example, the polymerase chain reaction (PCR) can be used to amplify a sequence that is expressed in normal tissues, e.g., nucleic acids encoding a sHASEGP polypeptide (SEQ. Nos: 1 and 2), in a genomic or cDNA library. Oligonucleotide primers that hybridize to sequences at the 3'and 5' termini of the identified sequences can be used as primers to amplify by PCR sequences from a nucleic acid sample (RNA or DNA generally a cDNA library, from an appropriate source (e. testis, prostate, breast).

[0210] PCR can be carried out, e. g., by use of a Perkin-Elmer Cetus thermal cycler and Taq polymerase (Gene Amp). The DNA being amplified can include mRNA or cDNA or genomic DNA from any eukaryotic species. One can choose to synthesize several different degenerate primers, for use in the PCR reactions.

[0211] It is also possible to vary the stringency of hybridization conditions used in priming the PCR reactions, to amplify nucleic acid homologs (e. g., to obtain sHASEGP

polypeptide sequences from species other than humans or to obtain human sequences with homology to sHASEGP polypeptide) by allowing for greater or lesser degrees of nucleotide sequence similarity between the known nucleotide sequence and the nucleic acid homolog being isolated. For cross- species hybridization, low stringency to moderate stringency conditions are used. For same species hybridization, moderately stringent to highly stringent conditions are used. The conditions can be empirically determined.

[0212] After successful amplification of the nucleic acid containing all or a portion of the identified sHASEGP polypeptide sequence or of a nucleic acid encoding all or a portion of a sHASEGP polypeptide homolog, that segment can be molecularly cloned and sequenced, and used as a probe to isolate a complete cDNA or genomic clone. This, in turn, permits the determination of the gene's complete nucleotide sequence, the analysis of its expression, and the production of its protein product for functional analysis. Once the nucleotide sequence is determined, an open reading frame encoding the sHASEGP polypeptide gene protein product can be determined by any method well known in the art for determining open reading frames, for example, using publicly available computer programs for nucleotide sequence analysis. Once an open reading frame is defined, it is routine to determine the amino acid sequence of the protein encoded by the open reading frame. In this way, the nucleotide sequences of the entire sHASEGP polypeptide genes as well as the amino acid sequences of sHASEGP polypeptide proteins and analogs can be identified.

[0213] Any eukaryotic cell potentially can serve as the nucleic acid source for the molecular cloning of the sHASEGP polypeptide gene. The nucleic acids can be isolated from vertebrate, mammalian, human, porcine, bovine, feline, avian, equine, canine, as well as additional primate sources, insects, plants and other organisms. The DNA can be obtained by standard procedures known in the art from cloned DNA (e. g., a DNA"library"), by chemical synthesis, by cDNA cloning, or by the cloning of genomic DNA, or fragments thereof, purified from the desired cell (see, e. g., Sambrook et al. 1989, Molecular Cloning, A Laboratory Manual, 2d Ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York; Glover, D. M. Ed., 1985, DNA Cloning : A Practical Approach, MRL Press, Ltd., Oxford, U. K. Vol. 1 ,11. Clones derived from genomic DNA can contain regulatory and intron DNA regions in addition to coding

regions; clones derived from cDNA will contain only exon sequences. For any source, the gene is cloned into a suitable vector for propagation thereof.

[0214] In the molecular cloning of the gene from genomic DNA, DNA fragments are generated, some of which will encode the desired gene.

[0215] The DNA can be cleaved at specific sites using various restriction enzymes.

[0216] Alternatively, one can use DNAse in the presence of manganese to fragment the DNA, or the DNA can be physically sheared, for example, by sonication. The linear DNA fragments then can be separated according to size by standard techniques, including but not limited to, agarose and polyacrylamide gel electrophoresis and column chromatography.

[0217] Once the DNA fragments are generated, identification of the specific DNA fragment containing the desired gene can be accomplished in a number of ways.

[0218] For example, a portion of the sHASEGP polypeptide (of any species) gene (e. g., a PCR amplification product obtained as described above or an oligonucleotide having a sequence of a portion of the known nucleotide sequence) or its specific RNA, or a fragment thereof be purified and labeled, and the generated DNA fragments can be screened by nucleic acid hybridization to the labeled probe (Benton and Davis, Science 196: 180 (1977); Grunstein and Hogness, Proc. Natl. Acad. Sci. U. S. A. 72: 3961 (1975)). Those DNA fragments with substantial homology to the probe will hybridize. It is also possible to identify the appropriate fragment by restriction enzyme digestion (s) and comparison of fragment sizes with those expected according to a known restriction map if such is available or by DNA sequence analysis and comparison to the known nucleotide sequence of sHASEGP polypeptide. Further selection can be carried out on the basis of the properties of the gene. Alternatively, the presence of the gene can be detected by assays based on the physical, chemical, or- immunological properties of its expressed product. For example, cDNA clones, or DNA clones which hybrid-select the proper mRNA can be selected which produce a protein that, e. g., has similar or identical electrophoretic migration, isoelectric focusing behavior, proteolytic digestion maps, antigenic properties, Hyaluronidase activity. If an anti-sHASEGP polypeptide antibody is available, the protein

can be identified by binding of labeled antibody to the putatively sHASEGP polypeptide synthesizing clones, in an ELISA (enzyme-linked immunosorbent assay)-type procedure.

[0219] Alternatives to isolating the sHASEGP polypeptide genomic DNA include, but are not limited to, chemically synthesizing the gene sequence from a known sequence or making cDNA to the mRNA that encodes the sHASEGP polypeptide.

[0220] For example, RNA for cDNA cloning of the sHASEGP polypeptide gene can be isolated from cells expressing the protein. The identified and isolated nucleic acids then can be inserted into an appropriate cloning vector. A large number of vector-host systems known in the art can be used. Possible vectors include, but are not limited to, plasmids or modified viruses, but the vector system must be compatible with the host cell used. Such vectors include, but are not limited to, bacteriophages such as lambda derivatives, or plasmids such as pBR322 or pUC plasmid derivatives or the Bluescript vector (Stratagene, La Jolla, CA). The insertion into a cloning vector can, for example, be accomplished by ligating the DNA fragment into a cloning vector that has complementary cohesive termini.

[0221] If the complementary restriction sites used to fragment the DNA are not present in the cloning vector, the ends of the DNA molecules can be enzymatically modified. Alternatively, any site desired can be produced by ligating nucleotide sequences (linkers) onto the DNA termini; these ligated linkers can include specific chemically synthesized oligonucleotides encoding restriction endonuclease recognition sequences. In an alternative method, the cleaved vector and sHASEGP polypeptide gene can be modified by homopolymeric tailing.

[0222] Recombinant molecules can be introduced into host cells via transformation, transfection, infection, electroporation, calcium precipitation and other methods, so that many copies of the gene sequence are generated.

[0223] In specific embodiments, transformation of host cells with recombinant DNA molecules that incorporate the isolated sHASEGP polypeptide gene, cDNA, or synthesized DNA sequence enables generation of multiple copies of the gene.

[0224] Thus, the gene can be obtained in large quantities by growing transformants, isolating the recombinant DNA molecules from the transformants and, when necessary, retrieving the inserted gene from the isolated recombinant DNA.

[0225] E. VECTORS, PLASMIDS AND CELLS THAT CONTAIN NUCLEIC ACIDS ENCODING A SHASEGP POLYPEPTIDE OR HYALURONIDASE DOMAIN THEREOF AND EXPRESSION OF SHASEGP POLYPEPTIDES VECTORS AND CELLS.

[0226] For recombinant expression of one or more of the sHASEGP polypeptides, the nucleic acid containing all or a portion of the nucleotide sequence encoding the sHASEGP polypeptide can be inserted into an appropriate expression vector i.e., a vector that contains the necessary elements for the transcription and translation of the inserted protein coding sequence. The necessary transcriptional and translational signals can also be supplied by the native promoter for sHASEGP genes, and/or their flanking regions.

[0227] Also provided are vectors that contain nucleic acid encoding the sHASEGPs that can be introduced into an expression system capable of producing a soluble neutral active sHASEGP.

[0228] Cells containing the vectors are also provided. The cells include eukaryotic and prokaryotic cells, and the vectors suitable for use therein.

[0229] Eukaryotic cells, including dihydroflate reductase deficient Chinese Hamster Ovary Cells (DG44), containing the vectors are provided. Suitable cells include yeast cells, fungal cells, plant cells, insect cells and animal cells. The cells are used to produce a sHASEGP polypeptide or Hyaluronidase domain thereof by (a) growing the above-described cells under conditions whereby the encoded sHASEGP polypeptide or Hyaluronidase domain of the sHASEGP polypeptide is expressed by the cell, and then (b) recovering the expressed Hyaluronidase domain protein. In the exemplified embodiments, the Hyaluronidase domain is secreted into the medium.

[0230] In one embodiment, the vectors include a sequence of nucleotides that encodes a polypeptide that has Hyaluronidase activity and contains all or a portion of only the

Hyaluronidase domain, or multiple copies thereof, of a sHASEGP protein are provided. Also provided are vectors that comprise a sequence of nucleotides that encodes the Hyaluronidase domain and additional portions of a sHASEGP protein up to and including a full-length sHASEGP protein, as well as multiple copies thereof, are also provided. The vectors can be selected for expression of the sHASEGP protein or Hyaluronidase domain thereof in the cell or such that the sHASEGP protein is expressed as a secreted protein. Alternatively, the vectors can include signals necessary for secretion of encoded proteins. When the Hyaluronidase domain is expressed the nucleic acid is linked to nucleic acid encoding a secretion signal, such as the Saccharomyces cerevisiae a mating factor signal sequence or a portion thereof, or the native signal sequence.

[0231] In order to generate a soluble, neutral active sHASEGP, cells capable of introducing N-linked glycosylation are required. In the preferred embodiment, mammalian Chinese Hamster Ovary cells deficient in dihydrofolate reductase such as DG44, are electroporated with a plasmid encoding a strong mammalian promoter, such as CMV, nucleic acid encoding a sHASEGP followed by an internal ribosomal entry site, the mouse dihydrofolate reductase gene and the SV40 polyadenylation sequence as shown in **SEQ ID NO** 51. Such cells are then cultured in chemically defined medium in the absence of hypoxanthine and thymidine, followed by further gene amplification with increasing concentrations of methotrexate.

[0232] A variety of host-vector systems can be used to express the protein coding sequence. These include but are not limited to mammalian cell systems infected with virus e.g. vaccinia virus, adenovirus, etc.); insect cell systems infected with virus (e. g. baculovirus); microorganisms such as yeast containing yeast vectors; or bacteria transformed with bacteriophage, DNA, plasmid DNA, or cosmid DNA. The expression elements of vectors vary in their strengths and specificities. Depending on the host-vector system used, any one of a number of suitable transcription and translation elements can be used. Note that bacterial expression of sHASEGP DNA will not result in a catalytically active sHASEGP per se, but when combined with proper glycosylation machinery can be artificially glycosylated as such.

[0233] Any methods known to those of skill in the art for the insertion of nucleic acid fragments into a vector can be used to construct expression vectors containing a chimeric gene containing appropriate transcriptional /translational control signals and protein coding sequences. These methods can include in vitro recombinant DNA and synthetic techniques and in vivo recombinants (genetic recombination). Expression of nucleic acid sequences encoding sHASEGP polypeptide, or domains, derivatives, fragments or homologs thereof, can be regulated by a second nucleic acid sequence so that the genes or fragments thereof are expressed in a host transformed with the recombinant DNA molecule (s). For example, expression of the proteins can be controlled by any promoter/enhancer known in the art. In a specific embodiment, the promoter is not native to the genes for sHASEGP polypeptide. Promoters which can be used include but are not limited to the SV40 early promoter (Bernoist and Chambon, Nature 290: 304-310 (1981) the promoter contained in the 3'long terminal repeat of Rous sarcoma virus (Yamamoto et al., Ce//22: 787-797 (1980) the herpes thymidine kinase promoter (Wagner et al. Proc. Natl. Acad. Sci. USA 78: 1441-1445 (1981) the regulatory sequences of the metallothionein gene (Brinster et al., Nature 296: 39-42 (1982)); prokaryotic expression vectors such as the ß-Lactamase promoter (Villa-Kamaroff et al., Proc. Natl. Acad. Sci. USA 75 : 3727-3731 1978)) Or The TAC Promoter Deboer et al., Proc. Natl. Acad. Sci. USA 80: 21-25 (1983)); see also "Useful Proteins from Recombinant Bacteria" : in Scientific American 242: 79-94 (1980)); plant expression vectors containing the opaline synthetase promoter (Herrar-Estrella et al., Nature 303: 209-213 (1984)) or the cauliflower mosaic virus 35S RNA promoter (Garder et al., Nucleic Acids RES. 9 : 2871 (1981)), and the promoter of the photosynthetic enzyme ribulose bisphosphate carboxylase (Herrera-Estrella et AL., Nature 310: 115-120 (1984)) ; promoter elements from yeast and other fungi such as the Gal4 promoter, the alcohol dehydrogenase promoter, the phosphoglycerol kinase promoter, the alkaline phosphatase promoter, and the following animal transcriptional control regions that exhibit tissue specificity and have been used in transgenic animals : elastase I gene control region which is active in pancreatic acinar cells (Swift Et Al., Cell 38 : 639-646 (1984); Ornitz Et Al., Cold Spring Harbor Symp. Quant. Biol. 50: 399-409 (1986); Macdonald, Hepatology 7: 425-515 (1987)); insulin gene control region which is active in pancreatic beta cells (Hanahan et AL., Nature 315: 115-122 (1985)), immunoglobulin gene control region which is active in lymphoid cells

(Grosschedl et AL., Cell 38: 647-658 (1984); Adams et al., Nature 318: 533-538 (1985); Alexander et AL., Mol. Cell Biol. 7: 1436-1444 (1987)), mouse mammary tumor virus control region which is active in testicular, breast, lymphoid and mast cells (Leder et AL., CELL 45 : 485-495 (1986)), albumin gene control region which is active in liver (PINCKERT et AL., Genes and Devel. 1: 268- 276 (1987)), alpha-fetoprotein gene control region which is active in liver (Krumlauf et AL., Mol. Cell. Biol. 5: 1639-1648 (1985); Hammer et AL., Science 235: 53-58 1987)), alpha-1 antitrypsin gene control region which is active in liver (Kelsey et al., Genes And Devel. 1: 161-171 (1987)), beta globin gene control region which is active in myeloid cells (Mogram et al., Nature 315: 338-340 (1985); Kollias et AL., CE//46: 89-94 (1986)), myelin basic protein gene control region which is active in oligodendrocyte cells of the brain (Readhead et al., Cell 48: 703-712 (1987)), myosin light chain-2 gene control region which is active in skeletal muscle (Sani, Nature 314: 283-286 (1985)), and gonadotrophic releasing hormone gene control region which is active in gonadotrophs of the hypothalamus (Mason et al., Science 234: 1372-1378 (1986)).

[0234] In a specific embodiment, a vector is used that contains a promoter operably linked to nucleic acids encoding a sHASEGP polypeptide, or a domain, fragment, derivative or homolog, thereof, one or more origins of replication, and optionally, one or more selectable markers (e. G., an antibiotic resistance gene).

[0235] Specific initiation signals may also be required for efficient translation of a sHASEGP sequence. These signals include the ATG initiation codon and adjacent sequences. In cases where sHASEGP, its initiation codon and upstream sequences are inserted into the appropriate expression vector, no additional translational control signals may be needed. However, in cases where only coding sequence, or a portion thereof, is inserted, exogenous transcriptional control signals including the ATG initiation codon must be provided. Furthermore, the initiation codon must be in the correct reading frame to ensure transcription of the entire insert. Exogenous transcriptional elements and initiation codons can be of various origins, both natural and synthetic. The efficiency of expression may be enhanced by the inclusion of enhancers appropriate to the cell system in use (Scharf D et al (1994) Results Probl Cell Differ 20:125-62; Bittner et al (1987) Methods in Enzymol 153:516-544).

[0236] In addition, a host cell strain may be chosen for its ability to modulate the expression of the inserted sequences or to process the expressed protein in the desired fashion. Such modifications of the polypeptide include, but are not limited to, acetylation, carboxylation, glycosylation, phosphorylation, lipidation and acylation. Post-translational processing which cleaves a "prepro" form of the protein may also be important for correct insertion, folding and/or function. Different host cells such as CHO (DG44, DXB11 CHO-K1), HeLa, MDCK, 293, WI38, etc have specific cellular machinery and characteristic mechanisms for such post-translational activities and may be chosen to ensure the correct modification and processing of the introduced, foreign protein.

[0237] For long-term, high-yield production of recombinant proteins, stable expression is preferred. For example, cell lines that stably express sHASEGP may be transformed using expression vectors which contain viral origins of replication or endogenous expression elements and a selectable marker gene. Following the introduction of the vector, cells may be allowed to grow for 1-2 days in an enriched media before they are switched to selective media. The purpose of the selectable marker is to confer resistance to selection, and its presence allows growth and recovery of cells that successfully express the introduced sequences. Resistant clumps of stably transformed cells can be proliferated using tissue culture techniques appropriate to the cell type.

[0238] Any number of selection systems may be used to recover transformed cell lines. These include, but are not limited to, the herpes simplex virus thymidine kinase (Wigler M et al (1977) Cell 11:223-32) and adenine phosphoribosyltransferase (Lowy I et al (1980) Cell 22:817-23) genes which can be employed in TK- or APRT- cells, respectively. Also, antimetabolite, antibiotic or herbicide resistance can be used as the basis for selection; for example, DHFR which confers resistance to methotrexate (Wigler M et al (1980) Proc Natl Acad Sci 77:3567-70); npt, which confers resistance to the aminoglycosides neomycin and G-418 (Colbere-Garapin F et al (1981) J Mol Biol 150:1-14) and als or pat, which confer resistance to chlorsulfuron and phosphinotricin acetyltransferase, respectively (Murry, supra). Additional selectable genes have been described, for example, trpB, which allows cells to utilize indole in place of tryptophan, or hisD, which allows cells to utilize histinol in place of histidine (Hartman S C and R C Mulligan (1988) Proc Natl Acad Sci 85:8047-51). Recently, the use of visible markers has gained popularity

with such markers as anthocyanins, beta glucuronidase and its substrate, GUS, and luciferase and its substrate, luciferin, being widely used not only to identify transformants, but also to quantify the amount of transient or stable protein expression attributable to a specific vector system (Rhodes C A et al (1995) Methods Mol Biol 55:121-131).

[0239] IDENTIFICATION OF TRANSFORMANTS CONTAINING THE POLYNUCLEOTIDE SEQUENCE

[0240] Although the presence/absence of marker gene expression suggests that the gene of interest is also present, the presence and expression of an active sHASEGP should be confirmed. For example, if the sHASEGP is inserted within a marker gene sequence, recombinant cells containing sHASEGP can be identified by the absence of marker gene function. Alternatively, a marker gene can be placed in tandem with a sHASEGP sequence under the control of a single promoter. Expression of the marker gene in response to induction or selection usually indicates expression of the tandem sHASEGP as well. Detection of a properly glycosyalted neutral active sHASEGP can be determined by way of testing the conditioned media for sHASEGP enzyme activity under appropriate conditions.

[0241] PURIFICATION OF SHASEGP

[0242] Host cells transformed with a sHASEGP nucleotide sequence may be cultured under conditions suitable for the expression and recovery of the encoded protein from cell culture. The protein produced by a recombinant cell is preferably secreted but may be contained intracellularly depending on the sequence and/or the vector used. As will be understood by those of skill in the art, expression vectors containing sHASEGP can be designed with signal sequences that facilitate direct secretion of sHASEGP through a prokaryotic or eukaryotic cell membrane. Other recombinant constructions may join sHASEGP to nucleotide sequence encoding a polypeptide domain which will facilitate purification of soluble proteins (Kroll D J et al (1993) DNA Cell Biol 12:441-53; cf discussion of vectors infra containing fusion proteins).

[0243] sHASEGP may also be expressed as a recombinant protein with one or more additional polypeptide domains added to facilitate protein purification. Such purification

facilitating domains include, but are not limited to, metal chelating peptides such as histidine-tryptophan modules that allow purification on immobilized metals, protein A domains that allow purification on immobilized immunoglobulin, and the domain utilized in the FLAGS extension/affinity purification system (Immunex Corp, Seattle Wash.). The inclusion of a cleavable linker sequences such as Factor XA or enterokinase (Invitrogen, San Diego Calif.) between the purification domain and sHASEGP is useful to facilitate purification. One such expression vector provides for expression of a fusion protein compromising a sHASEGP and contains nucleic acid encoding 6 histidine residues followed by thioredoxin and an enterokinase cleavage site. The histidine residues facilitate purification on IMIAC (immobilized metal ion affinity chromatography as described in Porath et al (1992) Protein Expression and Purification 3: 263-281) while the enterokinase cleavage site provides a means for purifying the chemokine from the fusion protein.

[0244] In addition to recombinant production, fragments of sHASEGP may be produced by direct peptide synthesis using solid-phase techniques (cf Stewart et al (1969) Solid-Phase Peptide Synthesis, W H Freeman Co, San Francisco; Merrifield J (1963) J Am Chem Soc 85:2149-2154). In vitro protein synthesis may be performed using manual techniques or by automation. Automated synthesis may be achieved, for example, using Applied Biosystems 431A Peptide Synthesizer (Perkin Elmer, Foster City Calif.) in accordance with the instructions provided by the manufacturer. Various fragments of sHASEGP may be chemically synthesized separately and combined using chemical methods to produce the full-length molecule.

[0245] Expression vectors containing the coding sequences, or portions thereof, of a sHASEGP polypeptide, is made, for example, by subcloning the coding portions into the EcoRl restriction site of each of the three PGEX vectors (glutathione S- transferase expression vectors (Smith and Johnson, Gene 7: 31-40 (1988)). This allows for the expression of products in the correct reading frame. Exemplary vectors and systems for expression of the Hyaluronidase domains of the sHASEGP polypeptides include the well-known Pichia vectors (available, for example, from Invitrogen, San Diego, CA), particularly those designed for secretion of the encoded proteins. The protein can also be expressed cytoplasmically, such as in the inclusion bodies. One exemplary vector is described in the examples.

[0246] Plasmids for transformation of E. coli cells, include, for example, the pET expression vectors (see, U. S patent 4,952,496; available from Novagen, Madison, WI ; see, also literature published by Novagen describing the system).

[0247] Such plasmids include pET 11a, which contains the T71ac promoter, T7 terminator, the inducible E. coli lac operator, and the lac repressor gene; pET 12A-C, which contains the T7 promoter, T7 terminator, and the E. COLI OMPT secretion signal; and pET 15B and PET19B (Novagen, Madison, Wi), which contain a His-Tag leader sequence for use in purification with a His column and a thrombin cleavage site that permits cleavage following purification over the column; the T7-lac promoter region and the T7 terminator.

[0248] The vectors are introduced into host cells, such as Pichia cells and bacterial cells, such as E. coli, and the proteins expressed therein. Exemplary Pichia strains include, for example, GS115. Exemplary bacterial hosts contain chromosomal copies of DNA encoding T7 RNA polymerase operably linked to an inducible promoter, such as the LACUV promoter (see, U. S. Patent No. 4,952,496). Such hosts include, but are not limited to, the lysogenic E. coli strain BL21 (DE3).

[0249] The sHASEGP domains, derivatives and analogs can be produced by various methods known in the art. For example, once a recombinant cell expressing a sHASEGP polypeptide, or a domain, fragment or derivative thereof, is identified, the individual gene product can be isolated and analyzed. This is achieved by assays based on the physical and/or functional properties of the protein, including, but not limited to, radioactive labeling of the product followed by analysis by gel electrophoresis, immunoassay, cross-linking to marker-labeled product, and assays of proteolytic activity.

[0250] The sHASEGP polypeptides can be isolated and purified by standard methods known in the art (either from natural sources or recombinant host cells expressing the complexes or proteins), including but not restricted to column chromatography (E. g., ion exchange, affinity, gel exclusion, reversed-phase high pressure and fast protein liquid), differential centrifugation, differential solubility, or by any other standard technique used for the purification of proteins.

[0251] In one embodiment, a sHASEGP can be purified to homogeneity from the chemically defined conditioned media of HZ24 transfected and methotrexate amplified DG44 cells by 1) tangential flow diafiltration, 2) binding and elution from anion exchange chromatography, 3) flow through phenyl sepharose chromatography, 4) binding and elution from phenylboronate chromatography and 4) binding and elution with hydroxyapatite chromatography.

[0252] Functional properties can be evaluated using any suitable assay known in the art.

[0253] Alternatively, once a sHASEGP polypeptide or its domain or derivative is identified, the amino acid sequence of the protein can be deduced from the nucleotide sequence of the gene that encodes it. As a result, the protein or its domain or derivative can be synthesized by standard chemical methods known in the art (e. G. see Hunkapiller et al, Nature 310: 105-111 (1984)) followed by glycosylation *in vitro*.

[0254] Manipulations of sHASEGP polypeptide sequences can be made at the protein level. Also contemplated herein are sHASEGP polypeptide proteins, domains thereof, derivatives or analogs or fragments thereof, which are differentially modified during or after translation, e. g., by glycosylation, acetylation, phosphorylation, amidation, pegylation, derivatization by known protecting/blocking groups, proteolytic cleavage, linkage to an antibody molecule or other cellular ligand.

[0255] Any of numerous chemical modifications can be carried out by known techniques, including but not limited to specific chemical cleavage by cyanogen bromide, trypsin, chymotrypsin, papain, V8, NABH4, acetylation, formylation, oxidation, reduction, metabolic synthesis in the presence of tunicamycin and other such agents.

[0256] In addition, domains, analogs and derivatives of a sHASEGP polypeptide can be chemically synthesized. For example, a peptide corresponding to a portion of a sHASEGP polypeptide, which includes the desired domain or which mediates the desired activity in vitro can be synthesized by use of a peptide synthesizer.

[0257] Furthermore, if desired, nonclassical amino acids or chemical amino acid analogs can be introduced as a substitution or addition into the sHASEGP polypeptide sequence.

Non-classical amino acids include but are not limited to the D-isomers of the common amino acids, a-amino isobutyric acid, 4-aminobutyric acid, Abu, 2-aminobutyric acid, E-ABU, e-Ahx, 6-amino hexanoic acid, Aib, 2-amino isobutyric acid, 3-amino propionoic acid, ornithine, norleucine, norvaline, hydroxyproline, sarcosine, citrulline, cysteic acid, t-butylglycine, t-butylalanine, phenylglycine, cyclohexylalanine, ß-alanine, fluoro-amino acids, designer amino acids such as ß-methyl amino acids, ca-methyl amino acids, na-methyl amino acids, and amino acid analogs in general. Furthermore, the amino acid can be d (dextrorotary) or l (levorotary).

[0258] In cases where natural products are suspected of being mutant or are isolated from new species, the amino acid sequence of the sHASEGP polypeptide isolated from the natural source, as well as those expressed in vitro, or from synthesized expression vectors in vivo or in vitro, can be determined from analysis of the DNA sequence, or alternatively, by direct sequencing of the isolated protein. Such analysis can be performed by manual sequencing or through use of an automated amino acid sequenator.

[0259] Modifications- A variety of modifications of the sHASEGP polypeptides and domains are contemplated herein. A sHASEGP-encoding nucleic acid molecule can be modified by any of numerous strategies known in the art (Sambrook ET A/. (1990), Molecular Cloning, A Laboratory Manual, 2d ed., Cold Spring Harbor Laboratory, Cold Spring Harbor, New York). The sequences can be cleaved at appropriate sites with restriction endonuclease (s), followed by further enzymatic modification if desired, isolated, and ligated in vitro. In the production of the gene encoding a domain, derivative or analog of sHASEGP, care should be taken to ensure that the modified gene retains the original translational reading frame, uninterrupted by translational stop signals, in the gene region where the desired activity is encoded.

[0260] Additionally, the sHASEGP-encoding nucleic acid molecules can be mutated in vitro or in vivo, to create and/or destroy translation, initiation, and/or termination sequences, or to create variations in coding regions and/or form new restriction endonuclease sites or destroy pre-existing ones, to facilitate further in vitro modification. Also, as described herein muteins with primary sequence alterations, such as replacements of Cys residues and elimination or addition of glycosylation sites are contemplated; the

sHASEGP of SEQ ID No. 1 has seven potential glycosylation sites. Such mutations can be effected by any technique for mutagenesis known in the art, including, but not limited to, chemical mutagenesis and in vitro site-directed mutagenesis (Hutchinson et al., j. Biol. Chem. 253: 6551-6558 (1978)), use of TABE Linkers (Pharmacia). In one embodiment, for example, a sHASEGP polypeptide or domain thereof is modified to include a fluorescent label. In other specific embodiments, the sHASEGP polypeptide is modified to have a heterobifunctional reagent, such heterobifunctional reagents can be used to crosslink the members of the complex.

[0261] In addition, domains, analogs and derivatives of a sHASEGP can be chemically synthesized. For example, a peptide corresponding to a portion of a sHASEGP, which includes the desired domain or which mediates the desired activity in vitro can be synthesized by use of a peptide synthesizer. Furthermore, if desired, nonclassical amino acids or chemical amino acid analogs can be introduced as a substitution or addition into the sHASEGP sequence. Non-classical amino acids include but are not limited to the D-isomers of the common amino acids, a-amino isobutyric acid, 4-aminobutyric acid, Abu, 2-aminobutyric acid, S-ABU, e-Ahx, 6-amino hexanoic acid, Aib, 2-amino isobutyric acid, 3-amino propionoic acid, ornithine, norleucine, norvaline, hydroxyproline, sarcosine, citrulline, cysteic acid, t-butylglycine, t-butylalanine, phenylglycine, cyclohexylalanine, ß-alanine, fluoro-amino acids, designer amino acids such as ti- methyl amino acids, ca-methyl amino acids, na-methyl amino acids, and amino acid analogs in general. Furthermore, the amino acid can be d (dextrorotary) or l (levorotary).

[0262] F. GENERATION OF A FUNCTIONALLY ACTIVE GLYCOSYLATED SHASEGP WITH N-LINKED SUGAR MOIETIES.

[0263] Properly N-glycosylated human sHASEGP is required to generate a catalytically stable protein. N-linked glycosylation of sHASEGP can be achieved through various techniques. Glycosylation of sHASEGP can be achieved by introducing nucleic acids encoding sHASEGP into cells of eukaryotic origin capable of proper N-linked glycosylation or alternatively, by contacting sHASEGP polypeptide with cell free extracts or purified enzymes capable of introducing the desired N-linked sugar moieties.
SELECTION OF AN EXPRESSION SYSTEM

[0264] Eukaryotic cell expression systems vary in the extent and type of glycosylation they introduce into an ectopically expressed polypeptide. CHO cells are, for example, highly efficient at the introduction of N-linked glycosylation into an active sHASEGP polypeptide.

[0265] Additional eukaryotic expression systems that introduce N-linked glycosylation to generate a functional sHASEGP product can be tested by introducing a human sHASEGP expression plasmid into said cells and testing for neutral activity. Proper N-linked glycosylation can be determined by way of FACE analysis of PNGASE released oligosaccharides. Glycosylation profies of catalytically active sHASEGP's are further provided herein. Verification of glycosylation can also be made by treatment of sHASEGP from said cells with PNGASE-F or by growth of such cells in tunicamycin following introduction of sHASEGP encoding nucleic acids.

[0266] N-glycosylation of sHASEGP polypeptide *in vitro*. The sHASEGP polypeptide can be N-glycosylated by contact of sHASEGP polypeptide with cell-free extracts containing activity capable of transferring N-linked sugars to sHASEGP polypeptide such as canine microsomal membranes or through coupled transcription and translation as is commercially available (Promega Madison WI).

[0267] Oligosaccharides are considered to have a reducing end and a non-reducing end, whether or not the saccharide at the reducing end is in fact a reducing sugar. In accordance with accepted nomenclature, oligosaccharides are depicted herein with the non-reducing end on the left and the reducing end on the right. All oligosaccharides described herein are described with the name or abbreviation for the non-reducing saccharide (e.g., Gal), followed by the configuration of the glycosidic bond (.alpha. or. beta.), the ring bond, the ring position of the reducing saccharide involved in the bond, and then the name or abbreviation of the reducing saccharide (e.g., GlcNAc). The linkage between two sugars may be expressed, for example, as 2,3, 2.fw darw.3, or (2,3). Each saccharide is a pyranose.

[0268] As used herein, N-linked sugar moiety refers to an oligosaccharide attached to a sHASEGP via the amide nitrogen of Asn residues. N-linked oligosaccharides fall into

several major types (oligomannose, complex, hybrid, sulfated), all of which have (Man) 3-GlcNAc-GlcNAc-cores attached via the amide nitrogen of Asn residues that fall within -Asn-Xaa-Thr/Ser- sequences (where Xaa is not Pro). N-linked sites are often indirectly assigned by the appearance of a "blank" cycle during sequencing. Positive identification can be made after release of the oligosaccharide by PNGase F, which converts the glycosylated Asn to Asp. After PNGase F release, N-linked oligosaccharides can be purified using Bio-Gel P-6 chromatography, with the oligosaccharide pool subjected to preparative high pH anion exchange chromatography (HPAEC) (Townsend et al., (1989) Anal. Biochem. 182, 1-8). Certain oligosaccharide isomers can be resolved using HPAEC. Fucose residues will shift elution positions earlier in the HPAEC chromatogram, while additional sialic acid residues will increase the retention time. Concurrent treatment of glycoproteins whose oligosaccharide structures are known (e.g., bovine fetuin, a-l acid glycoprotein, ovalbumin, RNAse B, transferrin) can facilitate assignment of the oligosaccharide peaks. The collected oligosaccharides can be characterized by a combination of compositional and methylation linkage analyses (Waegheet al., (1983) Carbohydr Res. 123, 281-304.), with anomeric configurations assigned by NMR spectroscopy (Van Halbeek (1993) in Methods Enzymol 230).

[0269] Alternatively, oligosaccharides can be identified by fluorescence assisted carbohydrate electrophoresis (FACE) Callewaert et al. (2001) Glycobiology 11, 275-281.

[0270] G. DETECTION AND CHARACTERIZATION OF N-LINKED SUGAR MOIETIES ON sHASEGP

[0271] Determining whether a protein is in fact glycosylated is the initial step in glycoprotein glycan analysis. Polyacrylamide gel electrophoresis in the presence of sodium dodecyl sulfate (SDS-PAGE) has become the method of choice as the final step prior to protein sequencing. Glycosylated proteins often migrate as diffuse bands by SDS-PAGE. A marked decrease in bandwidth and change in migration position after treatment with peptide-N4- (N-acetyl-D-glucosaminyl) asparagine amidase (PNGase F) is considered diagnostic of N-linked glycosylation. If the other types of glycosylation are predominant other approaches must be used. Lectin blotting methods provide an approach that is independent of the class of glycosylation (N versus O). Lectins, carbohydrate-

binding proteins from various plant tissues, have both high affinity and narrow specificity for a wide range of defined sugar epitopes found on glycoprotein glycans (Cummings, R. D. (1994) Methods in Enzymol. 230, 66-86.). When conjugated with biotin or digoxigenin, they can be easily identified on membrane blots through a colorimetric reaction utilizing avidin or anti-digoxigenin antibodies conjugated with alkaline phosphatase (Haselbeck, et al. (1993) Methods in Mol. Biol. 14, 161-173.), analogous to secondary antibody-alkaline phosphatase reactions employed in Western blotting. Screening with a panel of lectins with well-defined specificity can provide considerable information about a glycoprotein's carbohydrate complement. Importantly, the color development amplification is sufficiently high that 10-50 ng of a glycoprotein can easily be seen on a membrane blot of an SDS-PAGE. Although lectins exhibit very high affinity for their cognate ligands, some do reveal significant avidity for structurally related epitopes. Thus, it is important to carefully note the possibility of cross-reactivity when choosing a panel of lectins, and apply those with the highest probability of individually distinguishing complex, hybrid and high mannose N-linked glycans from O-linked structures.

[0272] Monosaccharide analysis can also be used to determine whether sHASEGP is glycosylated and as in the case of lectin analysis provides additional information on structural features. Quantitative monosaccharide composition analysis i) identifies glycosylated proteins, ii) gives the molar ratio of individual sugars to protein, iii) suggests, in some cases, the presence of oligosaccharide classes, iv) is the first step in designing a structural elucidation strategy, and v) provides a measure of production consistency for recombinant glycoprotein therapeutics. In recent years high-pH anion-exchange chromatography with pulsed amperometric detection (HPAEC-PAD) has been extensively used to determine monosaccharide composition (Townsend, et al. (1995) in Carbohydrate Analysis: High-performance liquid chromatography and capillary electrophoresis (Z. El Rassi ed.). pp. 181-209.). More recently, fluorophore-based labeling methods have been introduced and many are available in kit form. A distinct advantage of fluorescent methods is an increase in sensitivity (50-fold). One potential disadvantage is that different monosaccharides may demonstrate different selectivity for the fluorophore during the coupling reaction, either in the hydrolysate or in the external standard mixture. However,

the increase in sensitivity and the ability to identify which monosaccharides are present from a small portion of the total amount of available glycoprotein, as well as the potential for greater sensitivity using laser induced fluorescence makes this approach attractive.

[0273] Monosaccharide composition analysis of small amounts of sHASEGP is best performed on PVDF (PSQ) membranes, after either electroblotting (Weitzhandleret al, (1993) J. Biol. Chem. 268, 5121-5130.) or if smaller aliquots are to be analyzed on dot blots. PVDF is an ideal matrix for carbohydrate analysis since neither mono- or oligosaccharides bind to the membrane, once released by either acid or enzymatic hydrolysis.

[0274] FACE analysis is an efficient means of detecting glycosylation profiles of sHASEGP's. FACE® N-Linked Oligosaccharide Profiling (Prozyme) with 30% oligosaccharide gels is one such mechanism. Oligosaccharides cleaved from 100 μg of glycoproteins by enzymatic digestion with N-Glycanase (a.k.a PNGase), labeled using the flourophore ANTS, and separated by electrophoresis can be used for detection of sHASEGP glycosylation profiles. The relative positions of the oligosaccharide bands are determined by running the sample and dilutions of the sample alongside an oligosaccharide standard ladder which designated the migration distance in Degree of Polymerization (DP) units.

[0275] H. SCREENING METHODS TO IDENTIFY COMPOUNDS THAT MODULATE sHASEGP ACTIVITY.

[0276] Several types of assays are exemplified and described herein. It is understood that the Hyaluronidase domains can be used in other assays. It is shown here, however, that the Hyaluronidase domains exhibit catalytic activity.

[0277] As such they are ideal for in vitro screening assays.

[0278] They can also be used in binding assays.

[0279] The sHASEGP full length zymogens, activated enzymes, and Hyaluronidase domains are contemplated for use in any screening assay known to those of skill in the art, including those provided herein. Hence the following description, if directed to

Hyaluronidase assays is intended to apply to use of a single chain Hyaluronidase domain or a catalytically active portion thereof of any Hyaluronidase, including a sHASEGP. Other assays, such as binding assays are provided herein, particularly for use with a sHASEGP, including any variants, such as splice variants thereof.

[0280] 1. Catalytic Assays for identification of agents that modulate the Hyaluronidase activity of a sHASEGP protein. Methods for identifying a modulator of the catalytic activity of a sHASEGP, particularly a single chain Hyaluronidase domain or catalytically active portion thereof, are provided herein. The methods can be practiced by: contacting the sHASEGP, a full-length zymogen or activated form, and particularly a single-chain domain thereof, with a substrate of the sHASEGP in the presence of a test substance, and detecting the proteolysis of the substrate, whereby the activity of the sHASEGP is assessed, and comparing the activity to a control. For example, a control can be the activity of the sHASEGP assessed by contacting a sHASEGP, including a full- length zymogen or activated form, and particularly a single-chain domain thereof, particularly a single-chain domain thereof, with a substrate of the sHASEGP, and detecting the proteolysis of the substrate, whereby the activity of the sHASEGP is assessed. The results in the presence and absence of the test compounds are compared. A difference in the activity indicates that the test substance modulates the activity of the sHASEGP. Activators of sHASEGP activation cleavage are also contemplated; such assays are discussed below.

[0281] In one embodiment a plurality of the test substances are screened simultaneously in the above screening method. In another embodiment, the sHASEGP is isolated from a target cell as a means for then identifying agents that are potentially specific for the target cell.

[0282] In another embodiment, a test substance is a therapeutic compound, and whereby a difference of the sHASEGP activity measured in the presence and in the absence of the test substance indicates that the target cell responds to the therapeutic compound.

[0283] One method includes the steps of (a) contacting the sHASEGP polypeptide or Hyaluronidase domain thereof with one or a plurality of test compounds under conditions

conducive to interaction between the ligand and the compounds; and (b) identifying one or more compounds in the plurality that specifically binds to the ligand.

[0284] Another method provided herein includes the steps of a) contacting a sHASEGP polypeptide or Hyaluronidase domain thereof with a substrate of the sHASEGP polypeptide, and detecting the degradation of substrate, whereby the activity of the sHASEGP polypeptide is assessed; b) contacting the sHASEGP polypeptide with a substrate of the sHASEGP polypeptide in the presence of a test substance, and detecting the degradation of the substrate, whereby the activity of the sHASEGP polypeptide is assessed; and c) comparing the activity of the sHASEGP polypeptide assessed in steps a) and b), whereby the activity measured in step a) differs from the activity measured in step b) indicates that the test substance modulates the activity of the sHASEGP polypeptide.

[0285] In another embodiment, a plurality of the test substances is screened simultaneously. In comparing the activity of a sHASEGP polypeptide in the presence and absence of a test substance to assess whether the test substance is a modulator of the sHASEGP polypeptide, it is unnecessary to assay the activity in parallel, although such parallel measurement is typical. It is possible to measure the activity of the sHASEGP polypeptide at one time point and compare the measured activity to a historical value of the activity of the sHASEGP polypeptide.

[0286] For instance, one can measure the activity of the sHASEGP polypeptide in the presence of a test substance and compare with historical value of the activity of the sHASEGP polypeptide measured previously in the absence of the test substance, and vice versa. This can be accomplished, for example, by providing the activity of the sHASEGP polypeptide on an insert or pamphlet provided with a kit for conducting the assay.

[0287] Methods for selecting substrates for a particular sHASEGP are described in the EXAMPLES, and particular Hyaluronidase assays are exemplified.

[0288] Combinations and kits containing the combinations optionally including instructions for performing the assays are provided. The combinations include a sHASEGP polypeptide and a substrate of the sHASEGP polypeptide to be assayed; and, optionally reagents for detecting proteolysis of the substrate. The substrates, which can be

chromogenic or fluorogenic molecules, including glycosaminoglycans, subject to proteolysis by a particular sHASEGP polypeptide, can be identified empirically by testing the ability of the sHASEGP polypeptide to cleave the test substrate. Substrates that are cleaved most effectively i.e. at the lowest concentrations and/or fastest rate or under desirable conditions), are identified.

[0289] Additionally provided herein is a kit containing the above-described combination. The kit optionally includes instructions for identifying a modulator of the activity of a sHASEGP polypeptide. Any sHASEGP polypeptide is contemplated as target for identifying modulators of the activity thereof.

[0290] 2. Binding assays. Also provided herein are methods for identification and isolation of agents, particularly compounds that bind to sHASEGPs. The assays are designed to identify agents that bind to the isolated Hyaluronidase domain (or a protein, other than a sHASEGP polypeptide, that contains the Hyaluronidase domain of a sHASEGP polypeptide), and to the activated form, including the activated form derived from the full-length zymogen or from an extended Hyaluronidase domain. The identified compounds are candidates or leads for identification of compounds for treatments of disorders and diseases involving aberrant Hyaluronidase activity. The sHASEGP polypeptides used in the methods include any sHASEGP polypeptide as defined herein, including the sHASEGP single chain Hyaluronidase domain or proteolytically active portion thereof.

[0291] A variety of methods are provided herein. These methods can be performed in solution or in solid phase reactions in which the sHASEGP polypeptide (s) or Hyaluronidase domain (s) thereof are linked, either directly or indirectly via a linker, to a solid support. Screening assays are described in the Examples, and these assays have been used to identify candidate compounds.

[0292] For purposes herein, all binding assays described above are provided for sHASEGP.

[0293] Methods for identifying an agent, such as a compound, that specifically binds to a sHASEGP single chain Hyaluronidase domain, a full-length activated sHASEGP or two

chain Hyaluronidase domain thereof are provided herein. The method can be practiced by (a) contacting the sHASEGP with one or a plurality of test agents under conditions conducive to binding between the sHASEGP and an agent; and (b) identifying one or more agents within the plurality that specifically binds to the sHASEGP.

[0294] For example, in practicing such methods the sHASEGP polypeptide is mixed with a potential binding partner or an extract or fraction of a cell under conditions that allow the association of potential binding partners with the polypeptide. After mixing, peptides, polypeptides, proteins or other molecules that have become associated with a sHASEGP are separated from the mixture. The binding partner that bound to the sHASEGP can then be removed and further analyzed. To identify and isolate a binding partner, the entire protein, for instance the entire disclosed protein of SEQ ID No. 1 can be used. Alternatively, a fragment of the protein can be used.

[0295] A variety of methods can be used to obtain cell extracts or body fluids, such as blood, serum, urine, sweat, synovial fluid, CSF and other such fluids.

[0296] For example, cells can be disrupted using either physical or chemical disruption methods. Examples of physical disruption methods include, but are not limited to, sonication and mechanical shearing. Examples of chemical lysis methods include, but are not limited to, detergent lysis and enzyme lysis. A skilled artisan can readily adapt methods for preparing cellular extracts in order to obtain extracts for use in the present methods.

[0297] Once an extract of a cell is prepared, the extract is mixed with the sHASEGP under conditions in which association of the protein with the binding partner can occur. A variety of conditions can be used, including conditions that resemble conditions found in the cytoplasm of a human cell or in a body fluid, such as blood. Features, such as osmolarity pH, temperature, and the concentration of cellular extract used, can be varied to optimize the association of the protein with the binding partner. Similarly, methods for isolation of molecules of interest from body fluids are known.

[0298] After mixing under appropriate conditions, the bound complex is separated from the mixture. A variety of techniques can be used to separate the mixture. For example,

antibodies specific to a sHASEGP can be used to immunoprecipitate the binding partner complex. Alternatively, standard chemical separation techniques such as chromatography and density/sediment centrifugation can be used.

[0299] After removing the non-associated cellular constituents in the extract, the binding partner can be dissociated from the complex using conventional methods. For example, dissociation can be accomplished by altering the salt concentration or pH of the mixture.

[0300] To aid in separating associated binding partner pairs from the mixed extract, the sHASEGP can be immobilized on a solid support. For example, the protein can be attached to a nitrocellulose matrix or acrylic beads. Attachment of the protein or a fragment thereof to a solid support aids in separating peptide/binding partner pairs from other constituents found in the extract. The identified binding partners can be either a single protein or a complex made up of two or more proteins.

[0301] Alternatively, the nucleic acid molecules encoding the single chain Hyaluronidases can be used in a yeast two-hybrid system. The yeast two-hybrid system has been used to identify other protein partner pairs and can readily be adapted to employ the nucleic acid molecules herein described.

[0302] Another in vitro binding assay, particularly for a sHASEGP, uses a mixture of a polypeptide that contains at least the catalytic domain of one of these proteins and one or more candidate binding targets or substrates. After incubating the mixture under appropriate conditions, the ability of the sHASEGP or a polypeptide fragment thereof containing the catalytic domain to bind to or interact with the candidate substrate is assessed. For cell-free binding assays, one of the components includes or is coupled to a detectable label. The label can provide for direct detection, such as radioactivity, luminescence, optical or electron density, etc., or indirect detection such as an epitope tag, an enzyme, etc. A variety of methods can be employed to detect the label depending on the nature of the label and other assay components. For example, the label can be detected bound to the solid substrate or a portion of the bound complex containing the label can be separated from the solid substrate, and the label thereafter detected.

[0303] 3. Detection of signal transduction sHASEGP, which is a membrane anchored protein, can be involved directly or indirectly in signal transduction directly as a cell surface receptor or indirectly by activating proteins, such as pro-growth factors that can initiate signal transduction.

[0304] In addition, secreted sHASEGP, such as the soluble domain of sHASEGP as described in SEQ ID NO. 4, can be involved in signal transduction either directly by binding to or interacting with a cell surface receptor or indirectly by activating proteins, such as pro-growth factors that can initiate signal transduction. Assays for assessing signal transduction are well known to those of skill in the art, and can be adapted for use with the sHASEGP polypeptide.

[0305] Assays for identifying agents that affect or alter signal transduction mediated directly or indirectly, such as via activation of a pro-growth factor, by a sHASEGP, particularly the full length or a sufficient portion to anchor the extracellular domain or a functional portion thereof of a sHASEGP on the surface of a cell are provided. Such assays, include, for example, transcription based assays in which modulation of a transduced signal is assessed by detecting an effect on an expression from a reporter gene (see, e. g., U. S. Patent No. 5,436,128).

[0306] 4. Methods for Identifying Agents that Modulate the Expression a Nucleic Acid encoding a sHASEGP. Another embodiment provides methods for identifying agents that modulate the expression of a nucleic acid encoding a sHASEGP. Such assays use any available means of monitoring for changes in the expression level of the nucleic acids encoding a sHASEGP.

[0307] Assay formats can be used to monitor the ability of the agent to modulate the expression of a nucleic acid encoding a sHASEGP. For instance, mRNA expression can be monitored directly by hybridization to the nucleic acids. Also enzyme assays as described can be used to detect agents that modulate the expression of sHASEGP.

[0308] Cell lines are exposed to the agent to be tested under appropriate conditions and time and total RNA or mRNA is isolated by standard procedures (see, e.g. Sambrook et al (1989) MOLECULAR CLONING : A LABORATORY MANUAL, 2nd Ed. Cold

Spring Harbor Laboratory Press). Probes to detect differences in RNA expression levels between cells exposed to the agent and control cells can be prepared from the nucleic acids. It is typical, but not necessary, to design probes which hybridize only with target nucleic acids under conditions of high stringency. Only highly complementary nucleic acid hybrids form under conditions of high stringency. Accordingly, the stringency of the assay conditions determines the amount of complementarity that should exist between two nucleic acid strands in order to form a hybrid. Stringency should be chosen to maximize the difference in stability between the probe: target hybrid and potential probe: non-target hybrids.

[0309] For example, N-and C-terminal fragments of the sHASEGP can be expressed in bacteria and used to search for proteins that bind to these fragments. Fusion proteins, such as His-tag or GST fusion to the N-or C- terminal regions of the sHASEGP can be prepared for use as a substrate. These fusion proteins can be coupled to, for example, Glutathione-Sepharose beads and then probed with cell lysates or body fluids. Prior to lysis, the cells or body fluids can be treated with a candidate agent that can modulate a sHASEGP or proteins that interact with domains thereon. Lysate proteins binding to the fusion proteins can be resolved by SDS-PAGE, isolated and identified by protein sequencing or mass spectroscopy, as is known in the art.

[0310] Antibody probes are prepared by immunizing suitable mammalian hosts in appropriate immunization protocols using the peptides, polypeptides or proteins if they are of sufficient length (e. g., 4,5,6,7,8,9,10,11,12,13,14,15,20, 25,30,35,40 or more consecutive amino acids the sHASEGP polypeptide or if required to enhance immunogenicity, conjugated to suitable carriers. Methods for preparing immunogenic conjugates with carriers, such as bovine serum albumin (BSA), keyhole limpet hemocyanin (KLH), or other carrier proteins are well known in the art. In some circumstances, direct conjugation using, for example, carbodiimide reagents can be effective; in other instances linking reagents such as those supplied by Pierce Chemical Co., Rockford, IL, can be desirable to provide accessibility to the hapten. Hapten peptides can be extended at either the amino or carboxy terminus with a Cys residue or interspersed with cysteine residues, for example, to facilitate linking to a carrier.

[0311] Administration of the immunogens is conducted generally by injection over a suitable time period and with use of suitable adjuvants, as is generally understood in the art. During the immunization schedule, titers of antibodies are taken to determine adequacy of antibody formation.

[0312] Anti-peptide antibodies can be generated using synthetic peptides corresponding to, for example, the carboxy terminal amino acids of the sHASEGP.

[0313] Synthetic peptides can be as small as 1-3 amino acids in length, generally at least 4 or more amino acid residues long. The peptides can be coupled to KLH using standard methods and can be immunized into animals, such as rabbits or ungulates. Polyclonal antibodies can then be purified, for example using Actigel beads containing the covalently bound peptide.

[0314] While the polyclonal antisera produced in this way can be satisfactory for some applications, for pharmaceutical compositions, use of monoclonal preparations are generally used. Immortalized cell lines which secrete the desired monoclonal antibodies can be prepared using the standard method of Kohler et al., (Nature 256: 495-7 (1975)) or modifications which effect immortalization of lymphocytes or spleen cells, as is generally known. The immortalized cell lines secreting the desired antibodies are screened by immunoassay in which the antigen is the peptide hapten, polypeptide or protein.

[0315] When the appropriate immortalized cell culture secreting the desired antibody is identified, the cells can be cultured either in vitro or by production in vivo via ascites fluid. Of particular interest, are monoclonal antibodies that recognize the catalytic domain or activation cleavage site (region) of a sHASEGP.

[0316] The antibodies or fragments can also be produced. Regions that bind specifically to the desired regions of receptor also can be produced in the context of chimeras with multiple species origin.

[0317] Agents that are assayed in the above method can be randomly selected or rationally selected or designed.

[0318] The agents can be, as examples, peptides, small molecules, and carbohydrates. A skilled artisan can readily recognize that there is no limit as to the structural nature of the agents.

[0319] The peptide agents can be prepared using standard solid phase (or solution phase) peptide synthesis methods, as is known in the art. In addition, the DNA encoding these peptides can be synthesized using commercially available oligonucleotide synthesis instrumentation and produced recombinantly using standard recombinant production systems. The production using solid phase peptide synthesis is necessitated if non-gene-encoded amino acids are to be included.

[0320] I. METHODS OF TREATMENT

[0321] sHASEGP's identified by the methods herein are used for treating or preventing abnormal accumulations of sHASEGP substrates in an animal, particularly a mammal, including a human. In one embodiment, the method includes administering to a mammal an effective amount of a sHASEGP glycoprotein, whereby the disease or disorder is treated or prevented.

[0322] In another embodiment, a sHASEGP inhibitor can be used in the treatment of an excess amount of neutral hyaluronidase activity. The mammal treated can be a human. The inhibitors provided herein are those identified by the screening assays. In addition, antibodies and antisense nucleic acids or double-stranded RNA (dsRNA), such as RNAi, are contemplated.

[0323] 1. Antisense treatment: In a specific embodiment, as described hereinabove, sHASEGP polypeptide function is reduced or inhibited by sHASEGP polypeptide antisense nucleic acids, to treat or prevent excessive chondroitinase activity. The therapeutic or prophylactic use of nucleic acids of at least six nucleotides, generally up to about 150 nucleotides, that are antisense to a gene or cDNA encoding sHASEGP polypeptide or a portion thereof is provided. A sHASEGP polypeptide"antisense"nucleic acid as used herein refers to a nucleic acid capable of hybridizing to a portion of a sHASEGP polypeptide RNA (generally mRNA) by virtue of some sequence complementarity, and generally under high stringency conditions. The antisense nucleic

acid can be complementary to a coding and/or noncoding region of a sHASEGP polypeptide mRNA. Such antisense nucleic acids have utility as therapeutics that reduce or inhibit sHASEGP polypeptide function, and can be used in the treatment or prevention of disorders as described supra.

[0324] The sHASEGP polypeptide antisense nucleic acids are of at least six nucleotides and are generally oligonucleotides (ranging from 6 to about 150 nucleotides including 6 to 50 nucleotides). The antisense molecule can be complementary to all or a portion of the Hyaluronidase domain. For example, the oligonucleotide is at least 10 nucleotides, at least 15 nucleotides, at least 100 nucleotides, or at least 125 nucleotides. The oligonucleotides can be DNA or RNA or chimeric mixtures or derivatives or modified versions thereof, single- stranded or double-stranded. The oligonucleotide can be modified at the base moiety, sugar moiety, or phosphate backbone. The oligonucleotide can include other appending groups such as peptides, or agents facilitating transport across the cell membrane (see e.g., Letsinger et al., Proc. Natl. Acad. Sci. USA 86: 6553-6556 (1989); Lemaitre et al., Proc. Natl. Acad. Sci. USA 84: 648-652 (1987); PCT Publication No. WO 88/09810, published December 15, 1988) or blood-brain barrier (see e.g., PCT Publication No. WO 89/10134, published April 25, 1988), hybridization-triggered cleavage agents (see e.g., Krol et al., BioTechniques 6: 958-976 (1988)) or intercalating agents (see e.g., Zon. Pharm. Res. 5: 539-549 (1988)).

[0325] The sHASEGP polypeptide antisense nucleic acid generally is an oligo- nucleotide, typically single-stranded DNA or RNA or an analog thereof or mixtures thereof. For example, the oligonucleotide includes a sequence antisense to a portion of a nucleic acid that encodes a human sHASEGP polypeptide. The oligonucleotide can be modified at any position on its structure with substituents generally known in the art.

[0326] The sHASEGP polypeptide antisense oligonucleotide can include at least one modified base moiety which is selected from the group including, but not limited to 5-fluorouracil, 5-bromouracil, 5-chlorouracil, 5-iodouracil, hypoxanthine, xanthine, 4-acetylcytosine, 5-(carboxyhydroxylmethyl) uracil, 5-carboxymethylaminomethyl-2-thiouridine, 5-carboxymethylaminomethyluracil, dihydrouracil, beta-D-galactosylqueosine, inosine, N6-isopentenyladenine, 1-methylguanine, 1-methylinosine,

2,2-dimethylguanine, 2-methyladenine, 2-methylguanine, 3-methylcytosine, 5-methylcytosine, N6-adenine, 7-methylguanine, 5-methylaminomethyluracil, 5-methoxyaminomethyl-2-thiouracil, beta-D-mannosylqueosine, 5-apos-methoxycarboxymethyluracil, 5-methoxyuracil, 2-methylthio-N6-isopentenyladenine, uracil-5-oxyacetic acid (v), wybutoxosine, pseudouracil, queosine, 2-thiocytosine, 5-methyl-2-thiouracil, 2-thiouracil, 4-thiouracil, 5-methyluracil, uracil-5-oxyacetic acid methylester, uracil-5-oxyacetic acid (v), 5-methyl-2-thiouracil, 3-(3-amino-3-n-2-carboxypropyl) uracil, (ACP3) w, and 2,6-diaminopurine.

[0327] In another embodiment, the oligonucleotide includes at least one modified sugar moiety selected from the group including but not limited to arabinose, 2-fluoroarabinose, xylulose, and hexose. The oligonucleotide can include at least one modified phosphate backbone selected from a phosphorothioate, a phosphorodithioate, a phosphoramidothioate, a phosphoramidate, a phosphordiamidate, a methylphosphonate, an alkyl phosphotriester, and a formacetal or analog thereof.

[0328] The oligonucleotide can be an a-anomeric oligonucleotide. An a-anomeric oligonucleotide forms specific double-stranded hybrids with complementary RNA in which the strands run parallel to each other (Gautier et al., Nucl. Acids Res. 15: 6625-6641 (1987)).

[0329] The oligonucleotide can be conjugated to another molecule, such as, but are not limited to, a peptide; hybridization triggered cross-linking agent, transport agent or a hybridization-triggered cleavage agent. The oligonucleotides can be synthesized by standard methods known in the art, e.g. by use of an automated DNA synthesizer (such as are commercially available from Biosearch, Applied Biosystems, etc.). As examples, phosphorothioate oligonucleotides can be synthesized by the method of Stein et al., Nucl. Acids Res. 16: 3209 (1988)), methylphosphonate oligonucleotides can be prepared by use of controlled pore glass polymer supports (Sarin et al., Proc. Natl. Acad. Sci. USA 85: 7448-7451 (1988)), etc. In a specific embodiment, the sHASEGP polypeptide antisense oligonucleotide includes catalytic RNA or a ribozyme (see, e.g., PCT International Publication WO 90/11364, published October 4,1990; Sarver et al., Science 247: 1222-1225 (1990)). In another embodiment, the oligonucleotide is a 2'-0- methylribonucleotide

(Inoue et al., Nucl. Acids Res. 15: 6131-6148 (1987)), or a chimeric RNA-DNA analogue Inoue et al., FEBS Lett. 215: 327-330 (1987)).

[0330] Alternatively, the oligonucleotide can be double-stranded RNA (dsRNA) such as RNAi.

[0331] In an alternative embodiment, the sHASEGP polypeptide antisense nucleic acid is produced intracellularly by transcription from an exogenous sequence.

[0332] For example, a vector can be introduced in vivo such that it is taken up by a cell, within which cell the vector or a portion thereof is transcribed, producing an antisense nucleic acid (RNA). Such a vector would contain a sequence encoding the sHASEGP polypeptide antisense nucleic acid. Such a vector can remain episomal or become chromosomally integrated, as long as it can be transcribed to produce the desired antisense RNA. Such vectors can be constructed by recombinant DNA technology methods standard in the art. Vectors can be plasmid, viral, or others known in the art, used for replication and expression in mammalian cells. Expression of the sequence encoding the sHASEGP polypeptide antisense RNA can be by any promoter known in the art to act in mammalian, including human, cells. Such promoters can be inducible or constitutive. Such promoters include but are not limited to: the SV40 early promoter region (Bernoist and Chambon, Nature 290: 304-310 (1981), the promoter contained in the 3' long terminal repeat of Rous sarcoma virus (Yamamoto et al., Ce//22: 787-797 (1980), the herpes thymidine kinase promoter (Wagner et al., Proc. Natl. Acad. Sci. USA 78: 1441-1445 (1981), the regulatory sequences of the metallothionein gene (Brinster et al., Nature 296: 39-42 (1982), etc.

[0333] The antisense nucleic acids include sequence complementary to at least a portion of an RNA transcript of a sHASEGP polypeptide gene, including a human sHASEGP polypeptide gene. Absolute complementarity is not required. The amount of sHASEGP polypeptide antisense nucleic acid that is effective in the treatment or prevention of neoplastic disease depends on the nature of the disease, and can be determined empirically by standard clinical techniques.

[0334] Where possible, it is desirable to determine the antisense cytotoxicity in cells in vitro, and then in useful animal model systems prior to testing and use in humans.

[0335] 2. RNA interference RNA interference (RNAi) (see, e.g. Chuang et al. (2000) Proc. Natl. Acad. Sci. USA 97: 4985) can be employed to inhibit the expression of a gene encoding a sHASEGP. Interfering RNA (RNAi) fragments, particularly double- stranded (ds) RNAi, can be used to generate loss-of-sHASEGP function. Methods relating to the use of RNAi to silence genes in organisms including, mammals, C. elegans, Drosophila and plants, and humans are known (see, e.g., Fire et al. (1998) Nature 391: 806-811; Fire (1999) Trends Genet. 15: 358-363; Sharp (2001) Genes Dev. 15: 485-490; Hammond et al. (2001) Nature Rev, Genet. 2: 110-119; Tuschl (2001) Chem. Biochem. 2: 239-245; Hamilton et al. (1999) Science 286: 950-952; Hammond et al. (2000) Nature 404: 293-296; Zamore et al. (2000) Cell 101: 25-33; Bernstein et al. (2001) Nature 409: 363-366; Elbashir et al. (2001) Genes Dev. 15: 188-200; Elbashir et al. (2001) Nature 411: 494-498; International PCT application No. WO 01/29058; International PCT application No. WO 99/32619).

[0336] Double-stranded RNA (dsRNA)-expressing constructs are introduced into a host, such as an animal or plant using, a repliable vector that remains episomal or integrates into the genome. By selecting appropriate sequences, expression of dsRNA can interfere with accumulation of endogenous mRNA encoding a sHASEGP. RNAi also can be used to inhibit expression in vitro.

[0337] Regions include at least about 21 (or 21) nucleotides that are selective (i.e. unique) for sHASEGP are used to prepare the RNAi. Smaller fragments of about 21 nucleotides can be transformed directly (i.e., in vitro or in vivo) into cells; larger RNAi dsRNA molecules are generally introduced using vectors that encode them. dsRNA molecules are at least about 21 bp long or longer, such as 50, 100, 150, 200 and longer. Methods, reagents and protocols for introducing nucleic acid molecules into cells in vitro and in vivo are known to those of skill in the art.

[0338] 3. Gene Therapy in an exemplary embodiment, nucleic acids that include a sequence of nucleotides encoding a sHASEGP polypeptide or functional domains or derivative thereof, are administered to promote sHASEGP polypeptide function, by way of gene therapy. Gene therapy refers to therapy performed by the administration of a nucleic acid to a subject. In this embodiment, the nucleic acid produces its encoded protein that

mediates a therapeutic effect by promoting sHASEGP polypeptide function. Any of the methods for gene therapy available in the art can be used (see, Goldspiel et al., Clinical Pharmacy 12: 488-505 (1993); Wu and Wu, Biotherapy 3: 87-95 (1991); Tolstoshev, An. Rev. Pharmacol. Toxicol. 32: 573-596 (1993); Mulligan, Science 260: 926-932 (1993); and Morgan and Anderson, An. Rev. Biochem. 62: 191-217 (1993); TIBTECH 11 5: 155-215 (1993). For example, one therapeutic composition for gene therapy includes a sHASEGP polypeptide-encoding nucleic acid that is part of an expression vector that expresses a sHASEGP polypeptide or domain, fragment or chimeric protein thereof in a suitable host. In particular, such a nucleic acid has a promoter operably linked to the sHASEGP polypeptide coding region, the promoter being inducible or constitutive, and, optionally, tissue-specific. In another particular embodiment, a nucleic acid molecule is used in which the sHASEGP polypeptide coding sequences and any other desired sequences are flanked by regions that promote homologous recombination at a desired site in the genome, thus providing for intrachromosomal expression of the sHASEGP protein nucleic acid (Koller and Smithies, Proc. Natl. Acad. Sci. USA 86: 8932-8935 (1989); Zijlstra et al., Nature 342: 435-438 (1989)).

[0339] Delivery of the nucleic acid into a patient can be either direct, in which case the patient is directly exposed to the nucleic acid or nucleic acid-carrying vector, or indirect, in which case, cells are first transformed with the nucleic acid in vitro, then transplanted into the patient. These two approaches are known, respectively, as in vivo or ex vivo gene therapy.

[0340] In a specific embodiment, the nucleic acid is directly administered in vivo, where it is expressed to produce the encoded product. This can be accomplished by any of numerous methods known in the art, e.g., by constructing it as part of an appropriate nucleic acid expression vector and administering it so that it becomes intracellular, e.g., by infection using a defective or attenuated retroviral or other viral vector (see U. S. Patent No. 4,980,286), or by direct injection of naked DNA, or by use of microparticle bombardment (e.g., a gene gun; Biolistic, Dupont), or coating with lipids or cell- surface receptors or transfecting agents, encapsulation in liposomes, microparticles, or microcapsules, or by administering it in linkage to a peptide which is known to enter the nucleus, by administering it in linkage to a ligand subject to receptor-mediated endocytosis

(see e.g., Wu and Wu, J. Biol. Chem, 262: 4429-4432 (1987)) (which can be used to target cell types specifically expressing the receptors), etc. In another embodiment, a nucleic acid-ligand complex can be formed in which the ligand is a fusogenic viral peptide to disrupt endosomes, allowing the nucleic acid to avoid lysosomal degradation. In yet another embodiment, the nucleic acid can be targeted in vivo for cell specific uptake and expression, by targeting a specific receptor (see, e.g., PCT Publications WO 92/06180 dated April 16,1992 (Wu et al.); WO 92/22635 dated December 23, 1992 (Wilson et al.); WO 92/20316 dated November 26, 1992 (Findeis et al.); WO 93/14188 dated July 22,1993 (Clarke et al.), WO 93/20221 dated October 14,1993 (Young)). Alternatively, the nucleic acid can be introduced intracellularly and incorporated within host cell DNA for expression, by homologous recombination (Koller and Smithies, Proc. Natl. Acad. Sci. USA 86: 8932-8935 (1989); Zijistra et al., Nature 342: 435-438 (1989)).

[0341] In a specific embodiment, a viral vector that contains the sHASEGP polypeptide nucleic acid is used. For example, a retroviral vector can be used (see Miller et al., Meth. Enzymol. 217: 581-599 (1993)). These retroviral vectors have been modified to delete retroviral sequences that are not necessary for packaging of the viral genome and integration into host cell DNA. The sHASEGP polypeptide nucleic acid to be used in gene therapy is cloned into the vector, which facilitates delivery of the gene into a patient. More detail about retroviral vectors can be found in Boesen et al., Biotherapy 6: 291-302 (1994), which describes the use of a retroviral vector to deliver the mdr1 gene to hematopoietic stem cells in order to make the stem cells more resistant to chemotherapy.

[0342] Other references illustrating the use of retroviral vectors in gene therapy are: Clowes et al., J. Clin. Invest. 93: 644-651 (1994); Kiem et al., Blood 83: 1467-1473 (1994); Salmons and Gunzberg, Human Gene Therapy 4: 129-141 (1993); and Grossman and Wilson, Curr. Opin. In Genetics And Devel. 3: 110-114 (1993).

[0343] Adenoviruses are other viral vectors that can be used in gene therapy. Adenoviruses are especially attractive vehicles for delivering genes to respiratory epithelia. Adenoviruses naturally infect respiratory epithelia where they cause a mild disease. Other targets for adenovirus-based delivery systems are liver, the central nervous system, endothelial cells, and muscle. Adenoviruses have the advantage of being capable

of infecting non-dividing cells. Kozarsky and Wilson, Current Opinion in Genetics and Development 3: 499-503 (1993) present a review of adenovirus-based gene therapy. Bout et al., Human Gene Therapy 5: 3-10 (1994) demonstrated the use of adenovirus vectors to transfer genes to the respiratory epithelia of rhesus monkeys. Other instances of the use of adenoviruses in gene therapy can be found in Rosenfeld et al., Science 252: 431-434 (1991); Rosenfeld et al., Cell 68: 143-155 (1992); and Mastrangeli et al., J. Clin. Invest. 91: 225-234 (1993).

[0344] Adeno-associated virus (AAV) has also been proposed for use in gene therapy (Walsh et al., Proc. Soc. Exp. Biol. Med. 204: 289-300 (1993).

[0345] Another approach to gene therapy involves transferring a gene to cells in tissue culture by such methods as electroporation, lipofection, calcium phosphate mediated transfection, or viral infection. Usually, the method of transfer includes the transfer of a selectable marker to the cells. The cells are then placed under selection to isolate those cells that have taken up and are expressing the transferred gene. Those cells are then delivered to a patient.

[0346] In this embodiment, the nucleic acid is introduced into a cell prior to administration in vivo of the resulting recombinant cell. Such introduction can be carried out by any method known in the art, including but not limited to transfection, electroporation, microinjection, infection with a viral or bacteriophage vector containing the nucleic acid sequences, cell fusion, chromosome-mediated gene transfer, microcell-mediated gene transfer, spheroplast fusion, etc. Numerous techniques are known in the art for the introduction of foreign genes into cells (see e.g., Loeffler and Behr, Meth. Enzymol. 217: 599-618 (1993); Cohen et al., Meth. Enzymol. 217: 618-644 (1993); Cline, Pharmac. Ther. 29: 69-92 (1985)) and can be used, provided that the necessary developmental and physiological functions of the recipient cells are not disrupted. The technique should provide for the stable transfer of the nucleic acid to the cell, so that the nucleic acid is expressible by the cell and generally heritable and expressible by its cell progeny.

[0347] The resulting recombinant cells can be delivered to a patient by various methods known in the art. In an embodiment, epithelial cells are injected, e.g., subcutaneously. In another embodiment, recombinant skin cells can be applied as a skin graft onto the patient. Recombinant blood cells (e.g., hematopoietic stem or progenitor cells) can be administered intravenously. The amount of cells envisioned for use depends on the desired effect, patient state, etc., and can be determined by one skilled in the art.

[0348] Cells into which a nucleic acid can be introduced for purposes of gene therapy encompass any desired, available cell type, and include but are not limited to epithelial cells, endothelial cells, keratinocytes, fibroblasts, muscle cells, hepatocytes; blood cells such as T lymphocytes, B lymphocytes, monocytes, macrophages, neutrophils, eosinophils, megakaryocytes, granulocytes; various stem or progenitor cells, in particular hematopoietic stem or progenitor cells, e.g., such as stem cells obtained from bone marrow, umbilical cord blood, peripheral blood, fetal liver, and other sources thereof.

[0349] For example, a cell used for gene therapy is autologous to the patient. In an embodiment in which recombinant cells are used in gene therapy, a sHASEGP polypeptide nucleic acid is introduced into the cells such that it is expressible by the cells or their progeny, and the recombinant cells are then administered in vivo for therapeutic effect. In a specific embodiment, stem or progenitor cells are used. Any stem and/or progenitor cells that can be isolated and maintained in vitro can potentially be used in accordance with this embodiment.

[0350] Such stem cells include but are not limited to hematopoietic stem cells (HSC), stem cells of epithelial tissues such as the skin and the lining of the gut, embryonic heart muscle cells, liver stem cells (PCT Publication WO 94/08598, dated April 28,1994), and neural stem cells (Stemple and Anderson, Cell 71: 973-985 (1992)).

[0351] Epithelial stem cells (ESC) or keratinocytes can be obtained from tissues such as the skin and the lining of the gut by known procedures (Rheinwald, Meth. Cell Bio. 21A: 229 (1980)). In stratified epithelial tissue such as the skin, renewal occurs by mitosis of stem cells within the germinal layer, the layer closest to the basal lamina. Stem cells within the lining of the gut provide for a rapid renewal rate of this tissue. ESC or

keratinocytes obtained from the skin or lining of the gut of a patient or donor can be grown in tissue culture (Rheinwald, Meth. Cell Bio. 21A: 229 (1980); Pittelkow and Scott, Cano. Clinic Proc. 61: 771 (1986)). If the ESC are provided by a donor, a method for suppression of host versus graft reactivity (e.g., irradiation, drug or antibody administration to promote moderate immunosuppression) also can be used.

[0352] With respect to hematopoietic stem cells (HSC), any technique which provides for the isolation, propagation, and maintenance in vitro of HSC can be used in this embodiment. Techniques by which this can be accomplished include (a) the isolation and establishment of HSC cultures from bone marrow cells isolated from the future host, or a donor, or (b) the use of previously established long-term HSC cultures, which can be allergenic or xenogenic.

[0353] Non-autologous HSC generally are used with a method of suppressing transplantation immune reactions of the future host/patient. In a particular embodiment, human bone marrow cells can be obtained from the posterior iliac crest by needle aspiration (see, e.g., Kodo et al., J. Clin. Invest. 73: 1377-1384 (1984)). For example, the HSC can be made highly enriched or in substantially pure form. This enrichment can be accomplished before, during, or after long- term culturing, and can be done by any techniques known in the art. Long-term cultures of bone marrow cells can be established and maintained by using, for example, modified Dexter cell culture techniques (Dexter et al., J. Cell Physiol. 91: 335 (1977)) or Witlock-Witte culture techniques (Witlock and Witte, Proc. Natl. Acad. Sci. USA 79: 3608-3612 (1982)).

[0354] In a specific embodiment, the nucleic acid to be introduced for purposes of gene therapy includes an inducible promoter operably linked to the coding region, such that expression of the nucleic acid is controllable by controlling the presence or absence of the appropriate inducer of transcription.

[0355] 3. Prodrugs- A method for treating tumors is provided. The method is practiced by administering a prodrug that is cleaved at a specific site by a HASEGP to release an active drug or precursor that can be converted to active drug in vivo. Upon contact with a cell that expresses sHASEGP activity, the prodrug is converted into an active drug. The

prodrug can be a conjugate that contains the active agent, such as an anti-tumor drug, such as a cytotoxic agent, or other therapeutic agent (TA), linked to a substrate for the targeted sHASEGP, such that the drug or agent is inactive or unable to enter a cell, in the conjugate, but is activated upon cleavage. The prodrug, for example, can contain an chondroitin sulfate molecule, typically a relatively short, less than about 20 disaccharide units, that is catalytically cleaved by the targeted sHASEGP. Cytotoxic agents, include, but are not limited to, alkylating agents, antiproliferative agents and tubulin binding agents. Others include, vinca drugs, mitomycins, bleomycins and taxanes.

[0356] J. PHARMACEUTICAL COMPOSITIONS AND MODES OF ADMINISTRATION

[0357] 1. Components of the compositions. Pharmaceutical compositions containing an active sHASEGP are provided herein. Also provided are combinations of compounds that modulate the activity of a sHASEGP polypeptide and another treatment or compound for treatment of a hyaluronidase disorder, such as an antibody compound.

[0358] The sHASEGP polypeptide and a second agent can be packaged as separate compositions for administration together or sequentially or intermittently. Alternatively, they can be provided as a single composition for administration or as two compositions for administration as a single composition. The combinations can be packaged as kits.

[0359] 2. Formulations and Route of Administration

[0360] The sHASEGP polypeptides and soluble human hyaluronidase domain thereof provided herein can be formulated as pharmaceutical compositions, typically for single dosage administration. The concentrations of the polypeptides in the formulations are effective for delivery of an amount, upon administration, that is effective for the intended treatment. Typically, the compositions are formulated for single dosage administration. To formulate a composition, the weight fraction of a sHASEGP polypeptide, soluble human hyaluronidase domains thereof or mixture thereof is dissolved, suspended, dispersed or otherwise mixed in a selected vehicle at an effective concentration such that the treated condition is relieved or ameliorated.

[0361] Pharmaceutical carriers or vehicles suitable for administration of the sHASEGP or soluble human hyaluronidase domains thereof provided herein include any such carriers known to those skilled in the art to be suitable for the particular mode of administration.

[0362] In addition, the polypeptides can be formulated as the sole pharmaceutically active ingredient in the composition or can be combined with other active ingredients. Liposomal suspensions, including tissue-targeted liposomes, can also be suitable as pharmaceutically acceptable carriers. These can be prepared according to methods known to those skilled in the art. For example, liposome formulations can be prepared as described in U. S. Patent No. 4,522,811.

[0363] The active sHASEGP or soluble human hyaluronidase domain thereof is included in the pharmaceutically acceptable carrier in an amount sufficient to exert a therapeutically useful effect in the absence of undesirable side effects on the patient treated. The therapeutically effective concentration can be determined empirically by testing the polypeptides in known *in vitro* and *in vivo* systems such as by using the assays provided herein or see, *e.g.*, Taliani *et al.* (1996) *Anal. Biochem. 240*: 60-67, Filocamo *et al.* (1997) *J. Virology 71*: 1417-1427, Sudo *et al.* (1996) *Antiviral Res. 32*: 9-18, Buffard *et al.* (1995) *Virology 209*: 52-59, Bianchi *et al.* (1996) *Anal. Biochem. 237*: 239-244, Hamatake *et al.* (1996) *Intervirology 39*:249-258, Steinkühler *et al.* (1998) *Biochem. 37*:8899-8905, D'Souza *et al.* (1995) *J. Gen. Virol. 76*:1729-1736, Takeshita *et al.* (1997) *Anal. Biochem. 247*: 242-246; see also, *e.g.*, Shimizu *et al.* (1994) *J. Virol. 68*: 8406-8408; Mizutani *et al.* (1996) *J. Virol. 70*: 7219-7223, Mizutani *et al.* (1996) *Biochem. Biophys. Res. Commun. 227*: 822-826, Lu *et al.* (1996) *Proc. Natl. Acad. Sci. (USA) 93*: 1412-1417, Hahm *et al.* (1996) *Virology 226*: 318-326, Ito *et al.* (1996) *J. Gen. Virol. 77*: 1043-1054, Mizutani *et al.* (1995) *Biochem. Biophys. Res. Commun. 212*: 906-911, Cho *et al.* (1997) *J. Virol. Meth. 65* :201-207 and then extrapolated therefrom for dosages for humans.

[0364] Typically a therapeutically effective dosage is contemplated. The amounts administered can be on the order of 0.001 to 1 mg/ml, including about 0.005-0.05 mg/ml and about 0.01 mg/ml, of blood volume. Pharmaceutical dosage unit forms are prepared to provide from about 1 mg to about 1000 mg, including from about 10 to about 500 mg, and

including about 25-75 mg of the essential active ingredient or a combination of essential ingredients per dosage unit form. The precise dosage can be empirically determined.

[0365] In some instances, a high Unit dose of human sHASEGP is preferable. For example, with intravenous administration of sHASEGP concentrations of sHASEGP from 500-100,000 Units per ml are preferable. Lyophilized formulations of sHASEGP are also ideal for storage of large Unit doses of sHASEGP. 200,000 Unit lyophilized vials of sHASEGP are contemplated for intravenous delivery.

[0366] High concentration doses are also contemplated for the delivery of small volumes of sHASEGP. Administration of 10-100ul of 5000 Units/ml sHASEGP is contemplated for injection in the anterior chamber to dissolve pre administered viscoelastic substances during cataract and phakic intraocular lens implantation surgeries. Small volume injections of 50-200U/ml doses are also contemplated for intravitreal procedures such as the treatment of vitreous hemorrhage or vitro-retinal detachment in diabetic retinopathy.

[0367] The active ingredient can be administered at once, or can be divided into a number of smaller doses to be administered at intervals of time. It is understood that the precise dosage and duration of treatment is a function of the disease being treated and can be determined empirically using known testing protocols or by extrapolation from in vivo or in vitro test data. It is to be noted that concentrations and dosage values can also vary with the severity of the condition to be alleviated. It is to be further understood that for any particular subject, specific dosage regimens should be adjusted over time according to the individual need and the professional judgment of the person administering or supervising the administration of the compositions, and that the concentration ranges set forth herein are exemplary only and are not intended to limit the scope or use of the claimed compositions and combinations containing them.

[0368] Pharmaceutically acceptable derivatives include acids, salts, esters, hydrates, solvates and prodrug forms. The derivative is typically selected such that its pharmacokinetic properties are superior to the corresponding neutral sHASEGP or soluble human hyaluronidase domain thereof.

[0369] Thus, effective concentrations or amounts of one or more of the polypeptides herein or pharmaceutically acceptable derivatives thereof are mixed with a suitable pharmaceutical carrier or vehicle for systemic, topical or local administration to form pharmaceutical compositions. sHASEGP polypeptides or soluble human hyaluronidase domains thereof are included in an amount effective for ameliorating or treating the disorder for which treatment is contemplated. The concentration of active polypeptide in the composition depends on absorption, inactivation, excretion rates of the active polypeptide, the dosage schedule, amount administered, particular formulation as well as other factors known to those of skill in the art.

[0370] The therapeutic agents for use in the methods can be administered by any route known to those of skill in the art, such as, but are not limited to, topically, intraarticularly, intracisternally, intraocularly, intraventricularly, intrathecally, intravenously, intramuscularly, intraperitoneally, intradermally, intratracheally, as well as by any combination of any two or more thereof. Dry powder pulmonary formulations can be envisioned as well.

[0371] The most suitable route for administration will vary depending upon the proposed use, such as, for example, use as a delivery agent to facilitate subcutaneous delivery of fluids, use to reduce intraocular pressure in the eyes of glaucoma patients receiving viscoelastics or use as a "spreading agent" to enhance the activity of chemotherapeutics, and the location of interest, such as a particular internal organ, a tumor growth, intraocular cavity and the epidermis. Modes of administration include, but are not limited to, topically, locally, intraarticularly, intracisternally, intraocularly, intraventricularly, intrathecally, intravenously, intramuscularly, intratracheally, intraperitoneally, intradermally, and by a combination of any two or more thereof. For example, for treatment of various cancers, such as squamous cell carcinoma, breast cancer, urinary bladder cancer and gastrointestinal cancer, local administration, including administration to the site of the tumor growth (*e.g.*, intrathecally, intraventricularly, or intracisternally) provides the advantage that the therapeutic agent can be administered in a high concentration without risk of the complications that can accompany systemic administration of a therapeutic agent.

[0372] Pharmaceutical and cosmetic carriers or vehicles suitable for administration of the sHASEGP polypeptides or soluble human hyaluronidase domain thereof provided herein include any such carriers known to those skilled in the art to be suitable for the particular mode of administration. In addition, the polypeptides can be formulated as the sole pharmaceutically active ingredient in the composition or can be combined with other active ingredients that do not impair the desired action, or with materials that supplement the desired action known to those of skill in the art. For example, the sHASEGP polypeptides provided herein can be used as a delivery or "spreading" agent in combination with a second active compound, such as a therapeutically effective agent, including, but not limited to a drug or a prodrug, to facilitate delivery of or to enhance the activity of the second active ingredient. In a particular embodiment, a sHASEGP polypeptide or a soluble human hyaluronidase domain thereof can be co-formulated with an anesthetic agent, such as Lignocaine, Bupivicaine or a mixture of the two, and, optionally, a hormonal agent, such as epinephrine, to decrease or stop blood uptake during ophthalmic surgery. A sHASEGP polypeptide or a soluble human hyaluronidase domain thereof can also be co-formulated with various chemotherapeutics, such as a toxin and a tumor necrosis factor, to enhance the activity of the chemotherapeutic and/or the accessibility of the target tumors to the chemotherapeutic. The active compound is included in the carrier in an amount sufficient to exert a therapeutically useful effect in the absence of serious toxic effects on the treated individual. The effective concentration can be determined empirically by testing the compounds using *in vitro* and *in vivo* systems, including the animal models described herein.

[0373] Solutions or suspensions used for parenteral, intradermal, subcutaneous, or topical application can include any of the following components: a sterile diluent, such as water for injection, saline solution, fixed oil, polyethylene glycol, glycerine, propylene glycol or other synthetic solvent; antimicrobial agents, such as benzyl alcohol and methyl parabens; antioxidants, such as ascorbic acid and sodium bisulfite; cheating agents, such as ethylenediaminetetraacetic acid (EDTA); buffers, such as acetates, citrates and phosphates; and agents for the adjustment of tonicity, including, but not limited to sodium chloride, calcium chloride, magnesium chloride, dextrose, glycerol or boric acid.

Parenteral preparations can be enclosed in ampoules, disposable syringes or single or multiple dose vials made of glass, plastic or other suitable material.

[0374] The sHASEGP polypeptides or soluble human hyaluronidase domains thereof can be suspended in micronized or other suitable form or can be derivatized to produce a more soluble active product or to produce a prodrug. The form of the resulting mixture depends upon a number of factors, including the intended mode of administration and the solubility of the polypeptide in the selected carrier or vehicle. The effective concentration is sufficient for ameliorating the targeted condition and can be empirically determined using methods known to those of skill in the art. To formulate a composition, the weight fraction of polypeptide is dissolved, suspended, dispersed, or otherwise mixed in a selected vehicle at an effective concentration such that the targeted condition is relieved or ameliorated.

[0375] In instances in which the sHASEGP polypeptides or soluble human hyaluronidase domain thereof exhibit insufficient solubility, methods for solubilizing polypeptides can be used. Such methods are known to those of skill in this art, and include, but are not limited to, using cosolvents, such as dimethylsulfoxide (DMSO), using surfactants, such as TWEEN® and Pluronic® F68; or dissolution in aqueous sodium bicarbonate. Derivatives of the polypeptides, such as prodrugs of the polypeptides can also be used in formulating effective pharmaceutical compositions. For ophthalmic indications, the compositions are formulated in an ophthalmically acceptable carrier. For the ophthalmic uses herein, local administration, either by topical administration or by injection is contemplated. Time-release formulations are also desirable. Typically, the compositions are formulated for single dosage administration, so that a single dose administers an effective amount.

[0376] Upon mixing or addition of the polypeptide with the vehicle, the resulting mixture can be a solution, suspension, emulsion or other composition and can be formulated as an aqueous mixtures, a creams, gels, ointments, emulsions, solutions, elixirs, lotions, suspensions, tinctures, pastes, foams, aerosols, irrigations, sprays, suppositories, bandages, or any other formulation suitable for systemic, topical or local administration.

[0377] The form of the resulting mixture depends upon a number of factors, including the intended mode of administration and the solubility of the compound in the selected carrier or vehicle. If necessary, pharmaceutically acceptable salts or other derivatives of the compounds are prepared. For local internal administration, such as, intramuscular, parenteral or intra-articular administration, the compounds are preferably formulated as a solution suspension in an aqueous-based medium, such as isotonically buffered saline or are combined with a biocompatible support or bioadhesive intended for internal administration.

[0378] The sHASEGP polypeptide or soluble human hyaluronidase domain thereof is included in the pharmaceutically acceptable carrier in an amount sufficient to exert a therapeutically useful effect in the absence of undesirable side effects on the patient treated. It is understood that number and degree of side effects depends upon the condition for which the compounds are administered. For example, certain toxic and undesirable side effects are tolerated when treating life-threatening illnesses that would not be tolerated when treating disorders of lesser consequence. Amounts effective for therapeutic use will, of course, depend on the severity of the disease and the weight and general state of the subject as well as the route of administration. Local administration of the therapeutic agent will typically require a smaller dosage than any mode of systemic administration, although the local concentration of the therapeutic agent can, in some cases, be higher following local administration than can be achieved with safety upon systemic administration.

[0379] Since individual subjects can present a wide variation in severity of symptoms and each therapeutic agent has its unique therapeutic characteristics, it is up to the practitioner to determine the response of a subject to treatment and vary the dosages accordingly. Dosages used *in vitro* can provide useful guidance in the amounts useful for *in situ* administration of the pharmaceutical composition, and animal models can in some cases be used to determine effective dosages for treatment of particular disorders. In general, however, for local administration, it is contemplated that an effective amount of the therapeutic agent will be an amount within the range from about 0.1 picograms (pg) up to about 1 ng per kg body weight. Various considerations in arriving at an effective amount are known to those of skill in the art and are described (see, *e.g.*, Goodman And Gilman's:

The Pharmacological Bases of Therapeutics, 8th ed., Pergamon Press, 1990; Remington's Pharmaceutical Sciences, 17th ed., Mack Publishing Co., Easton, Pa., 1990; and Mantyh *et al.*, (*Science, 278*: 275-79, 1997) involving the intrathecal injection of a neuronal specific ligand-toxin, each of which is herein incorporated by reference in its entirety).

[0380] The formulations of the sHASEGP polypeptides or soluble human hyaluronidase domains thereof for use herein include those suitable for oral, rectal, topical, inhalational, buccal (*e.g.*, sublingual), parenteral (*e.g.*, subcutaneous, intramuscular, intradermal, or intravenous), transdermal administration or any route. The most suitable route in any given case depends on the nature and severity of the condition being treated and on the nature of the particular active compound that is being used. The formulations are provided for administration to humans and animals in unit dosage forms, such as tablets, capsules, pills, powders, granules, sterile parenteral solutions or suspensions, and oral solutions or suspensions, and oil-water emulsions containing suitable quantities of the polypeptides and/or other agents or pharmaceutically acceptable derivatives thereof. The pharmaceutical therapeutically active polypeptides and/or other agents and derivatives thereof are typically formulated and administered in unit-dosage forms or multiple-dosage forms. A unit-dose form as used herein refers to physically discrete units suitable for human and animal subjects and packaged individually as is known in the art.

[0381] The pharmaceutical compositions are provided for administration to humans and animals in unit dosage forms, such as tablets, capsules, pills, powders, granules, sterile parenteral solutions or suspensions, and oral solutions or suspensions, and oil-water emulsions containing suitable quantities of the sHASEGP polypeptide or soluble human hyaluronidase domain thereof and, optionally, another agent or pharmaceutically acceptable derivatives thereof. The pharmaceutically therapeutically active compounds and derivatives thereof are typically formulated and administered in unit-dosage forms or multiple-dosage forms. A unit-dose form as used herein refers to physically discrete units suitable for human and animal subjects and packaged individually as is known in the art. Each unit-dose contains a predetermined quantity of the therapeutically active compound sufficient to produce the desired therapeutic effect, in association with the required pharmaceutical carrier, vehicle or diluent. Examples of unit-dose forms include, but are not limited to, ampoules, syringes and individually packaged tablets or capsules. For

example, a small volume formulation containing a stabilized solution with 1 to 5000 Units of sHASEGP in a small volume, such as 5 to 50 μl, can be prepackaged in a syringe for use, such as after viscoelastic injection. Unit-dose forms can be administered in fractions or multiples thereof. A multiple-dose form is a plurality of identical unit-dosage forms packaged in a single container to be administered in segregated unit-dose form. Examples of multiple-dose forms include vials, bottles of tablets or capsules or bottles of pints or gallons. Hence, multiple dose form is a multiple of unit-doses that are not segregated in packaging.

[0382] The composition can contain along with the active ingredient, such as a sHASEGP polypeptide: a diluent, such as lactose, sucrose, dicalcium phosphate, or carboxymethylcellulose; a lubricant, such as magnesium stearate, calcium stearate and talc; and a binder such as starch, natural gums, such as gum acaciagelatin, glucose, molasses, polvinylpyrrolidine, celluloses and derivatives thereof, povidone, crospovidones and other such binders known to those of skill in the art. Liquid pharmaceutically administrable compositions can, for example, be prepared by dissolving, dispersing, or otherwise mixing an active compound as defined above and optional pharmaceutical adjuvants in a carrier, such as, for example, water, saline, aqueous dextrose, glycerol, glycols, ethanol, and the like, to thereby form a solution or suspension. If desired, the pharmaceutical composition to be administered can also contain minor amounts of nontoxic auxiliary substances such as wetting agents, emulsifying agents, or solubilizing agents, pH buffering agents and the like, for example, acetate, sodium citrate, cyclodextrine derivatives, sorbitan monolaurate, triethanolamine sodium acetate, triethanolamine oleate, and other such agents. Methods of preparing such dosage forms are known, or will be apparent, to those skilled in this art (see *e.g.*, Remington's Pharmaceutical Sciences, Mack Publishing Company, Easton, Pa., 15th Edition, 1975). The composition or formulation to be administered contains a quantity of the active compound in an amount sufficient to alleviate the symptoms of the treated subject. For example, a standard stabilized formulation of sHASEGP or a soluble human hyaluronidase domain thereof as provided herein includes 150 Units/ml of the soluble glycoprotein formulated in EDTA, NaCl and $CaCl_2$. Additionally, an anti-bacterial or anti-fungal agent, including, but not limited to thiomersal, can be present in the formulation. Another

formulation provided herein is a stabilized solution or lyophilized form of sHASEGP or a soluble human hyaluronidase domain thereof in EDTA, NaCl and $CaCl_2$, containing an effective active amount of the soluble glycoprotein, such as 150 Unit/ml, with the addition of lactose, such as 13 mg/ml. Also provided herein is a formulation containing a stabilized solution or lyophilized form of sHASEGP or a soluble human hyaluronidase domain thereof in EDTA, NaCl and $CaCl_2$ containing an effective active amount of the soluble glycoprotein, such as 150 Unit/ml, with the addition of lactose, such as 13 mg/ml, and Albumin, Pluronic® F68, TWEEN® and/or other detergent. Another formulation provided herein, either lyophilized or as a stabilized solution, contains an effect amount of sHASEGP or a soluble human hyaluronidase domain thereof, such as 1 to 300 Units/ml, in EDTA, NaCl and $CaCl_2$.

[0383] Dosage forms or compositions containing active ingredient in the range of 0.005% to 100% with the balance made up from non-toxic carrier can be prepared. For oral administration, the pharmaceutical compositions can take the form of, for example, tablets or capsules prepared by conventional means with pharmaceutically acceptable excipients such as binding agents (*e.g.*, pregelatinized maize starch, polyvinyl pyrrolidone or hydroxypropyl methylcellulose); fillers (*e.g.*, lactose, microcrystalline cellulose or calcium hydrogen phosphate); lubricants (*e.g.*, magnesium stearate, talc or silica); disintegrants (*e.g.*, potato starch or sodium starch glycolate); or wetting agents (*e.g.*, sodium lauryl sulphate). The tablets can be coated by methods well known in the art.

[0384] The sHASEGPs or a soluble human hyaluronidase domain thereof or pharmaceutically acceptable derivatives can be prepared with carriers that protect the soluble glycoprotein against rapid elimination from the body, such as time release formulations or coatings. The compositions can include other pharmaceutically effective agents known in the general art to be of value in treating one or more of the diseases or medical conditions, including, but not limited to, a chemotherapeutic agent, an analgesic agent, an anti-inflammatory agent, an antimicrobial agent, an amoebicidal agent, a trichomonocidal agent, an anti-parkinson agent, an anti-malarial agent, an anticonvulsant agent, an anti-depressant agent, and antiarthritics agent, an anti-fungal agent, an antihypertensive agent, antipyretic agent, an anti-parasite agent, an antihistamine agent, an alpha-adrenargic agonist agent, an alpha blocker agent, an anesthetic agent, a bronchi

dilator agent, a biocide agent, a bactericide agent, a bacteriostat agent, a betadrenergic blocker agent, a calcium channel blocker agent, a cardiovascular drug agent, a contraceptive agent, a decongestant agent, a diuretic agent, a depressant agent, a diagnostic agent, a electrolyte agent, a hypnotic agent, a hormone agent, a hyperglycemic agent, a muscle relaxant agent, a muscle contractant agent, an ophthalmic agent, a parasympathomimetic agent, a psychic energizer agent, ophthalmic agent, a parasympathomimetic agent, a psychic energizer agent, a sedative agent, a sympathomimetic agent, a tranquilizer agent, an urinary agent, a vaginal agent, a viricide agent, a vitamin agent, a non-steroidal anti-inflammatory agent, an angiotensin converting enzyme inhibitor agent, a polypeptide, a protein, a nucleic acid, a drug, a prodrug, a organic molecule and a sleep inducer, to obtain desired combinations of properties. It is to be understood that such combination therapy constitutes a further aspect of the compositions and methods of treatment provided herein.

[0385] 1. COMPOSITIONS FOR ORAL ADMINISTRATION

[0386] Oral pharmaceutical dosage forms are solid, gel or liquid. The solid dosage forms are tablets, capsules, granules, and bulk powders. Types of oral tablets include compressed, chewable lozenges and tablets, which can be enteric-coated, sugarcoated or film-coated. Capsules can be hard or soft gelatin capsules, while granules and powders can be provided in non-effervescent or effervescent form with the combination of other ingredients known to those skilled in the art.

[0387] The pharmaceutical compositions containing a sHASEGP or a soluble human hyaluronidase domain thereof can be in liquid form, for example, solutions, syrups or suspensions, or can be presented as a drug product for reconstitution with water or other suitable vehicle before use. Such liquid preparations can be prepared by conventional means with pharmaceutically acceptable additives such as suspending agents (*e.g.*, sorbitol syrup, cellulose derivatives or hydrogenated edible fats); emulsifying agents (*e.g.*, lecithin or acacia); non-aqueous vehicles (*e.g.*, almond oil, oily esters, or fractionated vegetable oils); and preservatives (*e.g.*, methyl or propyl-p-hydroxybenzoates or sorbic acid).

[0388] In certain embodiments, the formulations are solid dosage forms, preferably capsules or tablets. The tablets, pills, capsules, troches and the like can contain any of the following ingredients, or compounds of a similar nature: a binder; a diluent; a disintegrating agent; a lubricant; a glidant; a sweetening agent; and a flavoring agent.

[0389] Examples of binders include microcrystalline cellulose, gum tragacanth, glucose solution, acacia mucilage, gelatin solution, sucrose and starch paste. Lubricants include talc, starch, magnesium or calcium stearate, lycopodium and stearic acid. Diluents include, for example, lactose, sucrose, starch, kaolin, salt, mannitol and dicalcium phosphate. Glidants include, but are not limited to, colloidal silicon dioxide. Disintegrating agents include crosscarmellose sodium, sodium starch glycolate, alginic acid, corn starch, potato starch, bentonite, methylcellulose, agar and carboxymethylcellulose. Coloring agents include, for example, any of the approved certified water soluble FD and C dyes, mixtures thereof; and water insoluble FD and C dyes suspended on alumina hydrate. Sweetening agents include sucrose, lactose, mannitol and artificial sweetening agents such as saccharin, and any number of spray dried flavors. Flavoring agents include natural flavors extracted from plants such as fruits and synthetic blends of compounds which produce a pleasant sensation, such as, but not limited to peppermint and methyl salicylate. Wetting agents include propylene glycol monostearate, sorbitan monooleate, diethylene glycol monolaurate and polyoxyethylene laural ether. Emetic-coatings include fatty acids, fats, waxes, shellac, ammoniated shellac and cellulose acetate phthalates. Film coatings include hydroxyethylcellulose, sodium carboxymethylcellulose, polyethylene glycol 4000 and cellulose acetate phthalate.

[0390] If oral administration is desired, the sHASEGP or a soluble human hyaluronidase domain thereof could be provided in a composition that protects it from the acidic environment of the stomach. For example, the composition can be formulated in an enteric coating that maintains its integrity in the stomach and releases the active compound in the intestine. The composition can also be formulated in combination with an antacid or other such ingredient.

[0391] When the dosage unit form is a capsule, it can contain, in addition to material of the above type, a liquid carrier such as a fatty oil. In addition, dosage unit forms can

contain various other materials which modify the physical form of the dosage unit, for example, coatings of sugar and other enteric agents. The compounds can also be administered as a component of an elixir, suspension, syrup, wafer, sprinkle, chewing gum or the like. A syrup can contain, in addition to the active compounds, sucrose as a sweetening agent and certain preservatives, dyes and colorings and flavors.

[0392] The sHASEGP or a soluble human hyaluronidase domain thereof can also be mixed with other active materials which do not impair the desired action, or with materials that supplement the desired action, such as antacids, H2 blockers, and diuretics. The active ingredient is a compound or pharmaceutically acceptable derivative thereof as described herein. Higher concentrations, up to about 98% by weight of the active ingredient can be included.

[0393] Pharmaceutically acceptable carriers included in tablets are binders, lubricants, diluents, disintegrating agents, coloring agents, flavoring agents, and wetting agents. Enteric-coated tablets, because of the enteric-coating, resist the action of stomach acid and dissolve or disintegrate in the neutral or alkaline intestines. Sugar-coated tablets are compressed tablets to which different layers of pharmaceutically acceptable substances are applied. Film-coated tablets are compressed tablets which have been coated with a polymer or other suitable coating. Multiple compressed tablets are compressed tablets made by more than one compression cycle utilizing the pharmaceutically acceptable substances previously mentioned. Coloring agents can also be used in the above dosage forms. Flavoring and sweetening agents are used in compressed tablets, sugar-coated, multiple compressed and chewable tablets. Flavoring and sweetening agents are especially useful in the formation of chewable tablets and lozenges.

[0394] Liquid oral dosage forms include aqueous solutions, emulsions, suspensions, solutions and/or suspensions reconstituted from non-effervescent granules and effervescent preparations reconstituted from effervescent granules. Aqueous solutions include, for example, elixirs and syrups. Emulsions are either oil-in-water or water-in-oil.

[0395] Elixirs are clear, sweetened, hydroalcoholic preparations. Pharmaceutically acceptable carriers used in elixirs include solvents. Syrups are concentrated aqueous

solutions of a sugar, for example, sucrose, and can contain a preservative. An emulsion is a two-phase system in which one liquid is dispersed in the form of small globules throughout another liquid. Pharmaceutically acceptable carriers used in emulsions are non-aqueous liquids, emulsifying agents and preservatives. Suspensions use pharmaceutically acceptable suspending agents and preservatives. Pharmaceutically acceptable substances used in non-effervescent granules, to be reconstituted into a liquid oral dosage form, include diluents, sweeteners and wetting agents. Pharmaceutically acceptable substances used in effervescent granules, to be reconstituted into a liquid oral dosage form, include organic acids and a source of carbon dioxide. Coloring and flavoring agents are used in all of the above dosage forms.

[0396] Solvents include glycerin, sorbitol, ethyl alcohol and syrup. Examples of preservatives include glycerin, methyl and propylparaben, benzoic add, sodium benzoate and alcohol. Examples of non-aqueous liquids utilized in emulsions include mineral oil and cottonseed oil. Examples of emulsifying agents include gelatin, acacia, tragacanth, bentonite, and surfactants such as polyoxyethylene sorbitan monooleate. Suspending agents include sodium carboxymethylcellulose, pectin, tragacanth, Veegum and acacia. Diluents include lactose and sucrose. Sweetening agents include sucrose, syrups, glycerin and artificial sweetening agents such as saccharin. Wetting agents include propylene glycol monostearate, sorbitan monooleate, diethylene glycol monolaurate and polyoxyethylene lauryl ether. Organic additives include citric and tartaric acid. Sources of carbon dioxide include sodium bicarbonate and sodium carbonate. Coloring agents include any of the approved certified water soluble FD and C dyes, and mixtures thereof. Flavoring agents include natural flavors extracted from plants such fruits, and synthetic blends of compounds that produce a pleasant taste sensation.

[0397] For a solid dosage form, the solution or suspension, in for example propylene carbonate, vegetable oils or triglycerides, is encapsulated in a gelatin capsule. Such solutions, and the preparation and encapsulation thereof, are disclosed in U.S. Patent Nos 4,328,245; 4,409,239; and 4,410,545. For a liquid dosage form, the solution, *e.g.*, for example, in a polyethylene glycol, can be diluted with a sufficient quantity of a pharmaceutically acceptable liquid carrier, *e.g.*, water, to be easily measured for administration.

[0398] Alternatively, liquid or semi-solid oral formulations can be prepared by dissolving or dispersing the sHASEGP or a soluble human hyaluronidase domain thereof in vegetable oils, glycols, triglycerides, propylene glycol esters (*e.g.*, propylene carbonate) and other such carriers, and encapsulating these solutions or suspensions in hard or soft gelatin capsule shells. Other useful formulations include those set forth in U.S. Patent Nos. Re 28,819 and 4,358,603.

[0399] Formulations suitable for buccal (sublingual) administration include, for example, lozenges containing the sHASEGP or a soluble human hyaluronidase domain thereof in a flavored base, usually sucrose and acacia or tragacanth; and pastilles containing the compound in an inert base such as gelatin and glycerin or sucrose and acacia.

[0400] In all embodiments, tablets and capsules formulations can be coated as known by those of skill in the art in order to modify or sustain dissolution of the active ingredient. Thus, for example, they can be coated with a conventional enterically digestible coating, such as phenylsalicylate, waxes and cellulose acetate phthalate.

[0401] 2. Injectables, Solutions and Emulsions

[0402] Parenteral administration of the sHASEGP or a soluble human hyaluronidase domain thereof, generally characterized by injection, either subcutaneously, intramuscularly or intravenously is also contemplated herein. Injectables can be prepared in conventional forms, either as liquid solutions or suspensions; solid forms suitable for solution or suspension in liquid prior to injection, or as emulsions. Suitable excipients are, for example, water, saline, dextrose, glycerol or ethanol. In addition, if desired, the pharmaceutical compositions to be administered can also contain minor amounts of non-toxic auxiliary substances such as wetting or emulsifying agents, pH buffering agents, stabilizers, solubility enhancers, and other such agents, such as, for example, sodium acetate, sorbitan monolaurate, triethanolamine oleate and cyclodextrins. Implantation of a slow-release or sustained-release system, such that a constant level of dosage is maintained (see, *e.g.*, U.S. Patent No. 3,710,795) is also contemplated herein. The percentage of the sHASEGP or a soluble human hyaluronidase domain thereof contained

in such parenteral compositions is dependent on the specific nature thereof, as well as the activity of the compound and the needs of the subject.

[0403] Parenteral administration of the compositions includes intravenous, subcutaneous and intramuscular administrations. Preparations for parenteral administration include sterile solutions ready for injection, sterile dry soluble products, such as lyophilized powders, ready to be combined with a solvent or sterile solution just prior to use, including hypodermic tablets, sterile suspensions ready for injection, sterile dry insoluble products ready to be combined with a vehicle just prior to use and sterile emulsions. The solutions can be either aqueous or nonaqueous.

[0404] If administered intravenously, suitable carriers include physiological saline or phosphate buffered saline (PBS), and solutions containing thickening and solubilizing agents, such as glucose, polyethylene glycol, and polypropylene glycol and mixtures thereof.

[0405] Pharmaceutically acceptable carriers used in parenteral preparations include aqueous vehicles, nonaqueous vehicles, antimicrobial agents, isotonic agents, buffers, antioxidants, local anesthetics, suspending and dispersing agents, emulsifying agents, sequestering or chelating agents and other pharmaceutically acceptable substances.

[0406] Examples of aqueous vehicles include Sodium Chloride Injection, Ringers Injection, Isotonic Dextrose Injection, Sterile Water Injection, Dextrose and Lactated Ringers Injection. Nonaqueous parenteral vehicles include fixed oils of vegetable origin, cottonseed oil, corn oil, sesame oil and peanut oil. Antimicrobial agents in bacteriostatic or fungistatic concentrations must be added to parenteral preparations packaged in multiple-dose containers which include phenols or cresols, mercurials, benzyl alcohol, chlorobutanol, methyl and propyl p-hydroxybenzoic acid esters, thiomersal, benzalkonium chloride and benzethonium chloride. Isotonic agents include sodium chloride and dextrose. Buffers include phosphate and citrate. Antioxidants include sodium bisulfate. Local anesthetics include procaine hydrochloride. Suspending and dispersing agents include sodium carboxymethylcelluose, hydroxypropyl methylcellulose and polyvinylpyrrolidone. Emulsifying agents include Polysorbate 80 (TWEEN® 80). A

sequestering or chelating agent of metal ions includes EDTA. Pharmaceutical carriers also include ethyl alcohol, polyethylene glycol and propylene glycol for water miscible vehicles and sodium hydroxide, hydrochloric acid, citric acid or lactic acid for pH adjustment.

[0407] The concentration of the pharmaceutically active compound is adjusted so that an injection provides an effective amount to produce the desired pharmacological effect. The exact dose depends on the age, weight and condition of the patient or animal as is known in the art.

[0408] The unit-dose parenteral preparations are packaged in an ampoule, a vial or a syringe with a needle. All preparations for parenteral administration must be sterile, as is known and practiced in the art.

[0409] Illustratively, intravenous or intraarterial infusion of a sterile aqueous solution containing an active compound is an effective mode of administration. Another embodiment is a sterile aqueous or oily solution or suspension containing an active material injected as necessary to produce the desired pharmacological effect.

[0410] Injectables are designed for local and systemic administration. Typically a therapeutically effective dosage is formulated to contain a concentration of at least about 0.1% w/w up to about 90% w/w or more, preferably more than 1% w/w of the active compound to the treated tissue(s). The active ingredient, such as a sHASEGP or a soluble human hyaluronidase domain thereof, can be administered at once, or can be divided into a number of smaller doses to be administered at intervals of time. It is understood that the precise dosage and duration of treatment is a function of the tissue being treated and can be determined empirically using known testing protocols or by extrapolation from *in vivo* or *in vitro* test data. It is to be noted that concentrations and dosage values can also vary with the age of the individual treated. It is to be further understood that for any particular subject, specific dosage regimens should be adjusted over time according to the individual need and the professional judgment of the person administering or supervising the administration of the formulations, and that the concentration ranges set forth herein are

exemplary only and are not intended to limit the scope or practice of the claimed formulations.

[0411] The compounds provided herein can be formulated for parenteral administration by injection, *e.g.*, by bolus injection or continuous infusion. Formulations for injection can be presented in unit dosage form, *e.g.*, in ampoules or in multi-dose containers, with an added preservative. The compositions can be suspensions, solutions or emulsions in oily or aqueous vehicles, and can contain formulatory agents such as suspending, stabilizing and/or dispersing agents. Alternatively, the active ingredient can be in powder form for reconstitution with a suitable vehicle, *e.g.*, sterile pyrogen-free water or other solvents, before use. For example, provided herein are parenteral formulations containing an effective amount of sHASEGP or a soluble human hyaluronidase domain thereof, such as 500 to 500,000 Units, in a stabilized solution or a lyophilized from.

[0412] The compound can be suspended in micronized or other suitable form or can be derivatized to produce a more soluble active product or to produce a prodrug. The form of the resulting mixture depends upon a number of factors, including the intended mode of administration and the solubility of the compound in the selected carrier or vehicle. The effective concentration is sufficient for ameliorating the symptoms of the condition and can be empirically determined.

[0413] 3. Lyophilized Powders

[0414] Also provided herein are lyophilized powders containing sHASEGP or a soluble human hyaluronidase domain thereof, which can be reconstituted for administration as solutions, emulsions and other mixtures. These formulations can also be reconstituted and formulated as solids or gels.

[0415] The sterile, lyophilized powder is prepared by dissolving a solid portion of or mixing an aliquot of a solution containing a sHASEGP or a soluble human hyaluronidase domain thereof in a suitable solvent. The solvent can contain an excipient that improves the stability or other pharmacological component of the powder or reconstituted solution, prepared from the powder. Excipients that can be used include, but are not limited to, dextrose, sorbitol, fructose, corn syrup, xylitol, glycerin, glucose, sucrose, lactose or other

suitable agent. The solvent can also contain a buffer, such as citrate, sodium or potassium phosphate or other such buffer known to those of skill in the art at, typically, about neutral pH. Subsequent sterile filtration of the solution followed by lyophilization under standard conditions known to those of skill in the art provides the lyophilized formulation. Generally, the solution resulting from the sterile filtration is apportioned into vials for lyophilization. Each vial can contain a single dosage, such as 10-1000 mg or 100-500 mg, or multiple dosages of the compound.

[0416] Briefly, the lyophilized powder is prepared by dissolving dextrose, sorbitol, fructose, corn syrup, xylitol, glycerin, glucose, sucrose, lactose or other suitable agent, about 1-20%, in a suitable buffer, such as citrate, sodium or potassium phosphate or other such buffer known to those of skill in the art at about neutral pH. Then, a selected salt, such as, for example, the sodium salt of the sHASEGP (about 1 gm of the salt per 10-100 gms of the buffer solution, typically about 1 gm/30 gms), is added to the resulting mixture above room temperature, such as at about 30-35°C, and stirred until it dissolves. The resulting mixture is diluted by adding more buffer, to decrease the resulting concentration of the salt by about 10-50%, typically about 15-25%). The resulting mixture is sterile filtered or treated to remove particulates and to insure sterility, and apportioned into vials for lyophilization. The lyophilized powder can be stored under appropriate conditions, such as at about 4 C to room temperature.

[0417] Reconstitution of this lyophilized powder with water for injection provides a formulation for use in parenteral administration. For reconstitution, a therapeutically effective amount of the lyophilized powder containing a sHASEGP or a soluble human hyaluronidase domain thereof is added per milliliter of sterile water or other suitable carrier. The precise amount depends upon the selected compound and can be empirically determined by methods known to those of skill in the art.

[0418] 4. Topical administration

[0419] Topical mixtures are prepared as described for the local and systemic administration. The resulting mixture can be a solution, suspension, emulsions or the like and are formulated as creams, gels, ointments, emulsions, solutions, elixirs, lotions,

suspensions, tinctures, pastes, foams, aerosols, irrigations, sprays, suppositories, bandages, dermal patches or any other formulations suitable for topical administration.

[0420] The compositions of sHASEGP or a soluble human hyaluronidase domain thereof or pharmaceutically acceptable derivatives thereof can be formulated as aerosols for topical application, such as by inhalation (see, *e.g.*, U.S. Patent Nos. 4,044,126, 4,414,209, and 4,364,923, which describe aerosols for delivery of a steroid useful for treatment inflammatory diseases, particularly asthma). These formulations for administration to the respiratory tract can be in the form of an aerosol or solution for a nebulizer, or as a microfine powder for insufflation, alone or in combination with an inert carrier such as lactose. In such a case, the particles of the formulation will typically have diameters of less than 50 microns, such as less than 10 microns.

[0421] For administration by inhalation, the compositions for use herein can be delivered in the form of an aerosol spray presentation from pressurized packs or a nebulizer, with the use of a suitable propellant, including, but not limited to, dichlorodifluoromethane, trichlorofluoromethane, dichlorotetrafluoroethane, carbon dioxide and other suitable gases. In the case of a pressurized aerosol, the dosage unit can be determined by providing a valve to deliver a metered amount. Capsules and cartridges of, *e.g.*, gelatin, for use in an inhaler or insufflator can be formulated containing a powder mix of the compound and a suitable powder base such as lactose or starch.

[0422] The compositions can be formulated for local or topical application, such as for topical application to the skin and mucous membranes, such as in the eye, in the form of gels, creams, and lotions and for application to the eye or for intracisternal or intraspinal application. Topical administration is contemplated for transdermal delivery and also for administration to the eyes or mucosa, or for inhalation therapies. Nasal solutions of the active compound alone or in combination with other pharmaceutically acceptable excipients can also be administered.

[0423] For example, formulations suitable for topical application to the skin or to the eye generally are formulated as an ointment, cream, lotion, paste, gel, spray, aerosol and oil. Carriers that can be used include vaseline, lanoline, polyethylene glycols, alcohols, and

combinations of two or more thereof. The topical formulations can further advantageously contain 0.05 to 15 percent by weight of thickeners, including, but not limited to, hydroxypropylmethylcellulose, methylcellulose, polyvinylpyrrolidone, polyvinyl alcohol, poly (alkylene glycols), poly/hydroxyalkyl, (meth) acrylates or poly (meth) acrylamides. A topical formulation is often applied by instillation or as an ointment into the conjunctival sac. It also can be used for irrigation or lubrication of the eye, facial sinuses, and external auditory meatus. The topical formulations in the liquid state can be also present in a hydrophilic three-dimensional polymer matrix in the form of a strip, contact lens, and the like from which the active components are released. It can also be injected into the anterior eye chamber and other places. For example, provided herein is a formulation for intraocular use after viscoelastic injecting containing a stabilized solution of an effective amount of a sHASEGP or a soluble human hyaluronidase domain thereof, such as 1 to 5000 Units of the soluble glycoprotein with 30 to 150,000 Units/mg of specific activity, in a small volume, such as 5 to 50 μl.

[0424] These solutions, particularly those intended for ophthalmic use, can be formulated as 0.01% - 10% isotonic solutions, pH about 5-7, with appropriate salts.

[0425] 5. Compositions for Other Routes of Administration

[0426] Other routes of administration, such as topical application, transdermal patches, and rectal administration are also contemplated herein.

[0427] For example, pharmaceutical dosage forms for rectal administration are rectal suppositories, capsules and tablets for systemic effect. Rectal suppositories are used herein mean solid bodies for insertion into the rectum that melt or soften at body temperature releasing one or more pharmacologically or therapeutically active ingredients. Pharmaceutically acceptable substances utilized in rectal suppositories are bases or vehicles and agents to raise the melting point. Examples of bases include cocoa butter (theobroma oil), glycerin-gelatin, carbowax (polyoxyethylene glycol) and appropriate mixtures of mono-, di- and triglycerides of fatty acids. Combinations of the various bases can be used. Agents to raise the melting point of suppositories include spermaceti and

wax. Rectal suppositories can be prepared either by the compressed method or by molding. The typical weight of a rectal suppository is about 2 to 3 gm.

[0428] Tablets and capsules for rectal administration are manufactured using the same pharmaceutically acceptable substance and by the same methods as for formulations for oral administration.

[0429] Formulations suitable for transdermal administration can be presented as discrete patches adapted to remain in intimate contact with the epidermis of the recipient for a prolonged period of time. Such patches suitably contain the active compound as an optionally buffered aqueous solution of, for example, 0.1 to 0.2 M concentration with respect to the active compound. Formulations suitable for transdermal administration can also be delivered by iontophoresis (see *e.g., Pharmaceutical Research 3 (6)*: 318 (1986)) and typically take the form of an optionally buffered aqueous solution of the active compound.

[0430] The pharmaceutical compositions can also be administered by controlled release means and/or delivery devices (see e.g., in U.S. Patent Nos. 3,536,809; 3,598,123; 3,630,200; 3,845,770; 3,847,770; 3,916,899; 4,008,719; 4,687,610; 4,769,027; 5,059,595; 5,073,543; 5,120,548; 5,354,566; 5,591,767; 5,639,476; 5,674,533 and 5,733,566). The active compounds or pharmaceutically acceptable derivatives can be prepared with carriers that protect the compound against rapid elimination from the body, such as time-release formulations or coatings.

[0431] In one embodiment of the compositions and methods provided herein, the therapeutic agent is administered locally in a slow release delivery vehicle, for example, encapsulated in a colloidal dispersion system or in polymer stabilized crystals. Useful colloidal dispersion systems include nanocapsules, microspheres, beads, and lipid-based systems, including oil-in-water emulsions, micelles, mixed micelles, and liposomes. For example, the colloidal dispersion system can be a liposome or microsphere. Liposomes are artificial membrane vesicles that are useful as slow release delivery vehicles when injected or implanted. Some examples of lipid-polymer conjugates and liposomes are disclosed in U.S. Patent No., 5,631,018, which are incorporated herein by reference in its

entirety. Other examples of slow release delivery vehicles are biodegradable hydrogel matrices (U.S. Patent No. 5,041, 292), dendritic polymer conjugates (U.S. Patent No. 5,714,166), and multivesicular liposomes (Depofoam®, Depotech, San Diego, CA) (U. S. Patent Nos. 5,723,147 and 5,766,627). One type of microspheres suitable for encapsulating therapeutic agents for local injection (*e.g.*, into subdermal tissue) is poly(D,L)lactide microspheres, as described in D. Fletcher, *Anesth. Analg. 84*:90-94, (1997). For example, a slow release formulation containing an effective amount of sHASEGP or a soluble human hyaluronidase domain thereof, such as 1 to 5000 Units/ml, can be employed for various uses or to treat various conditions, including, but not limited to, cosmetic formulations and treatment of spinal cord injuries.

[0432] Desirable blood levels can be maintained by a continuous infusion of the active agent as ascertained by plasma levels. It should be noted that the attending physician would know how to and when to terminate, interrupt or adjust therapy to lower dosage due to toxicity, or bone marrow, liver or kidney dysfunctions. Conversely, the attending physician would also know how to and when to adjust treatment to higher levels if the clinical response is not adequate (precluding toxic side effects).

[0433] The efficacy and/or toxicity of the sHASEGP polypeptide and/or its inhibitor (s), alone or in combination with other agents, such as therapeutically effective agents, also can be assessed by the methods known in the art (see, *e.g.*, O & Apos; Reilly, *Investigational New Drugs 15*: 5-13 (1997)).

[0434] 6. Articles of manufacture

[0435] The sHASEGP polypeptides or soluble human hyaluronidase domains thereof or compositions containing any of the preceding agents can be packaged as articles of manufacture containing packaging material, a compound or suitable derivative thereof provided herein, which is effective for treatment of a diseases or disorders contemplated herein, within the packaging material, and a label that indicates that the compound or a suitable derivative thereof is for treating the diseases or disorders contemplated herein. The label can optionally include the disorders for which the therapy is warranted.

[0436] The articles of manufacture provided herein contain packaging materials. Packaging materials for use in packaging pharmaceutical products are well known to those of skill in the art (see, *e.g.*, U.S. Patent Nos. 5,323,907, 5,052,558 and 5,033,352). Examples of pharmaceutical packaging materials include, but are not limited to, blister packs, bottles, tubes, inhalers, pumps, bags, vials, containers, syringes, bottles, and any packaging material suitable for a selected formulation and intended mode of administration and treatment. A wide array of formulations of the compounds and compositions provided herein are contemplated, as are variety treatments for any disorder in which HCV infection is implicated as a mediator or contributor to the symptoms or cause.

[0437] Kits containing the compositions and/or the combinations with instructions for administration thereof are also provided herein. The kit can further include a needle or syringe, typically packaged in sterile form, for injecting the complex, and/or a packaged alcohol pad. Instructions are optionally included for administration of the active agent by a clinician or by the patient. For example, provided herein is a kit containing a small volume syringe with an effective amount of sHASEGP or a soluble human hyaluronidase domain thereof, such as 1 to 5000 Units of the soluble glycoprotein, in a 5 to 50 μl volume, optionally containing a second syringe containing a viscoelastic. Also provided herein is a kit containing a small volume syringe containing an effective amount of sHASEGP or a soluble human hyaluronidase domain thereof, such as 1 to 500 Units of the soluble glycoprotein, and a therapeutic amount of a second active ingredient, such as a drug, a small molecule, a protein or a nucleic acid.

[0438] K. ANIMAL MODELS

[0439] Transgenic animal models and animals, such as rodents, including mice and rats, cows, chickens, pigs, goats, sheep, monkeys, including gorillas, and other primates, are provided herein. In particular, transgenic non-human animals that contain heterologous nucleic acid encoding a sHASEGP polypeptide or a transgenic animal in which expression of the polypeptide has been altered, such as by replacing or modifying the promoter region or other regulatory region of the endogenous gene are provided. Such an animal can by produced by promoting recombination between endogenous nucleic acid and an

exogenous sHASEGP gene that could be over-expressed or mis-expressed, such as by expression under a strong promoter, via homologous or other recombination event.

[0440] Transgenic animals can be produced by introducing the nucleic acid using any know method of delivery, including, but not limited to, microinjection, lipofection and other modes of gene delivery into a germline cell or somatic cells, such as an embryonic stem cell. Typically the nucleic acid is introduced into a cell, such as an embryonic stem cell (ES), followed by injecting the ES cells into a blastocyst, and implanting the blastocyst into a foster mother, which is followed by the birth of a transgenic animal. Generally introduction of a heterologous nucleic acid molecule into a chromosome of the animal occurs by a recombination between the heterologous sHASEGP-encoding nucleic acid and endogenous nucleic acid. The heterologous nucleic acid can be targeted to a specific chromosome. In some instances, knockout animals can be produced. Such an animal can be initially produced by promoting homologous recombination between a sHASEGP polypeptide gene in its chromosome and an exogenous sHASEGP polypeptide gene that has been rendered biologically inactive (typically by insertion of a heterologous sequence, e.g., an antibiotic resistance gene). In one embodiment, this homologous recombination is performed by transforming embryo-derived stem (ES) cells with a vector containing the insertionally inactivated sHASEGP polypeptide gene, such that homologous recombination occurs, followed by injecting the ES cells into a blastocyst, and implanting the blastocyst into a foster mother, followed by the birth of the chimeric animal ("knockout animal") in which a sHASEGP polypeptide gene has been inactivated (see Capecchi, Science 244: 1288-1292 (1989)). The chimeric animal can be bred to produce homozygous knockout animals, which can then be used to produce additional knockout animals. Knockout animals include, but are not limited to, mice, hamsters, sheep, pigs, cattle, and other non-human mammals. For example, a knockout mouse is produced. The resulting animals can serve as models of specific diseases, such as cancers, that exhibit under- expression of a sHASEGP polypeptide. Such knockout animals can be used as animal models of such diseases e.g., to screen for or test molecules for the ability to treat or prevent such diseases or disorders.

[0441] Other types of transgenic animals also can be produced, including those that over-express the sHASEGP polypeptide. Such animals include "knock-in" animals that are

animals in which the normal gene is replaced by a variant, such as a mutant, an over-expressed form, or other form. For example, one species', such as a rodent's endogenous gene can be replaced by the gene from another species, such as from a human. Animals also can be produced by non- homologous recombination into other sites in a chromosome; including animals that have a plurality of integration events.

[0442] After production of the first generation transgenic animal, a chimeric animal can be bred to produce additional animals with over-expressed or mis-expressed sHASEGP polypeptides. Such animals include, but are not limited to, mice, hamsters, sheep, pigs, cattle and other non-human mammals. The resulting animals can serve as models of specific diseases, such as cancers, that are exhibit over-expression or mis-expression of a sHASEGP polypeptide. Such animals can be used as animal models of such diseases e.g., to screen for or test molecules for the ability to treat or prevent such diseases or disorders. In a specific embodiment, a mouse with over-expressed or mis-expressed sHASEGP polypeptide is produced.

[0443] The following examples are included for illustrative purposes only and are not intended to limit the scope of the invention.

[0444] L. THERAPEUTIC USES OF sHASEGP

[0445] Naturally occurring hyaluronidase enzymes from slaughterhouses have been the principle source of clinical enzyme preparations for over forty years. Bovine and Ovine testicles are the principle source of this material. These clinical enzyme preparations however are very crude, sold in preparations ranging from 0.5-5% purity based upon known specific activities between 30-100,000 Units/mg. Thus their lack of purity combined with their slaughterhouse origin, leave them as both immunogenic to humans and as potential source of Jacob Creutzfeld disease and other bovine and ovine pathogens. Anaphylactic reactions to bovine and ovine hyaluronidase preparations are known to occur.

[0446] Cattle or bacterially derived hyaluronidase have been used in the treatment of diseases associated with excess hyaluronic acid and to enhance the circulation of physiological fluids and/or therapeutic agents. For example, bovine hyaluronidase can be

co injected with anesthesia in peribulbar, retrobulbar and sub-Tenon's blocks for ophthalmic surgical procedures. Moreover, increased surgical complications occur in its absence (Brown SM et al. *J Cataract Refract Surg*. 1999 Sep; 25(9): 1245-9.). Bovine hyaluronidase is also used as an antidote to local necrosis from paravenous injection of necrotic substances such as vinka alkaloids (Few, B.J. (1987) *Amer. J. Matern. Child Nurs*. **12**, 23-26). Bovine testes hyaluronidase is also useful for the treatment of ganglion cysts (Paul et al. *J Hand Surg* 1997 Apr; 22 (2): 219-21). Hyaluronidase can also be used to facilitate subcutaneous delivery of fluids in hypodermoclysis (Berger EY, *Am Geriatr Soc* 1984 Mar; 32(3):199-203). Hyaluronidase has also been utilized to reduce intraocular pressure in the eyes of glaucoma patients and cataract patients receiving viscoelastics (U.S. Pat. No. 4,820,516 issued Apr. 11, 1989).

[0447] Cattle or bacterially derived hyaluronidases have also been used as a "spreading agent" to enhance the activity of chemotherapeutics and/or the accessibility of tumors to chemotherapeutics (Schuller et al., 1991, Proc. Amer. Assoc. Cancer Res. 32:173, abstract no. 1034; Czejka et al., 1990, Pharmazie 45:H.9). Combination chemotherapy with hyaluronidase is effective in the treatment of a variety of cancers including urinary bladder cancer (Horn et al., 1985, J. Surg. Oncol. 28:304-307), squamous cell carcinoma (Kohno et al., 94, J. Cancer Res. Oncol. 120:293-297), breast cancer (Beckenlehner et al., 1992, J. Cancer Res. Oncol. 118:591-596), and gastrointestinal cancer (Scheithauer et al., 1988, Anticancer Res. 8:391-396). Hyaluronidase is effective as the sole therapeutic agent in the treatment of brain cancer (gliomas) (PCT published application no. WO88/02261, published Apr. 7, 1988). Administration of hyaluronidase also induces responsiveness of previously chemotherapy-resistant tumors of the pancreas, stomach, colon, ovaries, and breast (Baumgartner et al., 1988, Reg. Cancer Treat. 1:55-58; Zanker et al., 1986, Proc. Amer. Assoc. Cancer Res. 27:390). Unfortunately, the contaminants and non human nature of such hyaluronidases result in anaphylactic reactions.

[0448] In addition to its indirect anticancer effects, cattle derived hyaluronidase has direct anticarcinogenic effects. Hyaluronidase prevents growth of tumors transplanted into mice (De Maeyer et al., 1992, Int. J. Cancer 51:657-660) and inhibits tumor formation upon exposure to carcinogens (Pawlowski et al., 1979, Int. J. Cancer 23:105-109; Haberman et

al., 1981, Proceedings of the 17th Annual Meeting of the American Society of Clinical Oncology, Washington, D.C., 22:105, abstract no. 415).

[0449] Given the value of cattle-derived hyaluronidases as a therapeutic, particularly in chemotherapy in conjunction with conventional chemotherapeutics or as a chemotherapeutic in and of itself, there is a need in the field for substantially pure preparations of hyaluronidase of human origin. There is also a need for efficient, cost-effective methods of making hyaluronidase to provide commercially significant quantities of the enzyme. The present invention addresses these problems.

[0450] Hyaluronic acid is an essential component of the extracellular matrix. Hyaluronic acid is found in the connective tissue of mammals and is the main constituent of the vitreous of the eye. In connective tissue, the water of hydration associated with hyaluronic acid creates spaces between tissues, thus creating an environment conducive to cell movement and proliferation. Hyaluronic acid plays a key role in biological phenomena associated with cell motility including rapid development, regeneration, repair, embryogenesis, embryological development, wound healing, angiogenesis, and tumorigenesis (Toole, 1991, Cell Biol. Extracell. Matrix, Hay (ed), Plenum Press, New York, 1384-1386; Bertrand et al., 1992, Int. J. Cancer 52:1-6; Knudson et al., 1993, FASEB J. 7:1233-1241). In addition, hyaluronic acid levels correlate with tumor aggressiveness (Ozello et al., 1960, Cancer Res. 20:600-604; Takeuchi et al., 1976, Cancer Res. 36:2133-2139; Kimata et al., 1983, Cancer Res. 43:1347-1354).

[0451] Following spinal cord injury, glial scars are produced by astrocytes and contain chondroitin sulfate proteoglycans (CSPGs). CSPGs play a crucial role in the inhibition of axon growth (Levine, 1994; Powell et al., 1997). For example, during fetal development, CSPGs repel axons and inhibit neural cell adhesion. CSPG's also play an important role in boundary formation (Snow et al., 1990, 1992; Powell and Geller, 1999). In addition the expression of CSPG increases following injury of CNS (Mckeon et al., 1991; Davies et al., 1997).

[0452] Studies indicate that the inhibitory effects of CSPGs are principally due to the chondroitin sulfate (CS) glycosaminoglycan (GAG) sugar chain (Snow et al., 1990; Cole

and McCable, 1991; Geisert and Bidanset, 1993). This is supported by the finding that administration of bacterial chondroitinase in fact promote axon regeneration when administered intrathecally. Moreover, electrophysiological experiments determined that regenerated CST axons established functional connections (Bradbury, et al 2002). In addition to their direct inhibitory effects, CSPGs could also interact with cell adhesion molecules or neurotrophic factors to influence neurite outgrowth (Roberts et al., 1988; Ruoslahti and Yamaguchi, 1991; Milev et al., 1994). Recombinant mammalian Hyaluronidases are thus useful to reverse the inhibition of CSPG's in the glial scar and to promote axon regeneration following injury.

[0453] The amount of sHASEGP required to sufficiently degrade CSPG's in the glial scar will vary. In some cases repeated administration of 10-5000 Units of sHASEGP by intrathecal delivery will be required to remove the CSPG's in the scar. In other cases, sustained release of sHASEGP through use of a slow release formulation may be preferred. Alternatively, administration of gene therapy vectors encoding sHASEGP may be effective to enhance clearance of CSPG's.

[0454] sHASEGPs can also be utilized for the treatment of herniated disks in a process known as chemonucleolysis. Chondroitinase ABC, and enzyme cleaving similar substrates as sHASEGP can induce the reduction of intradiscal pressure in the lumbar spine. (Sasaki et al., 2001, Ishikawa et al., 1999). There are three types of disk injuries. A protruded disk is one that is intact but bulging. In an extruded disk, the fibrous wrapper has torn and the NP has oozed out, but is still connected to the disk. In a sequestered disk, a fragment of the NP has broken loose from the disk and is free in the spinal canal. Chemonucleolysis is effective on protruded and extruded disks, but not on sequestered disk injuries. In the United States, chemonucleolysis is approved only for use in the lumbar (lower) spine. In other countries, it has also been used successfully to treat cervical (upper spine) hernias. Chemonucleolysis is thus a conservative alternative to disk surgery when it is preferable to reduce disk pressure.

[0455] The precise composition and structure of the carbohydrate chain(s) on a glycoprotein can directly influence its serum lifetime, since cells in the liver and reticulo-endothelial system can bind and internalize circulating glycoproteins with specific

carbohydrates. Hepatocytes have receptors on their surfaces that recognize oligosaccharide chains with terminal (i.e., at the outermost end(s) of glycans relative to the polypeptide) Gal residues, macrophages contain receptors for terminal Man or GlcNAc residues, and hepatocytes and lymphocytes have receptors for exposed fucose residues. No sialic acid-specific receptors have been found, however. Although somewhat dependent on the spatial arrangement of the oligosaccharides, as a general rule, the greater the number of exposed sugar residues recognized by cell surface receptors in the liver and reticulo-endothelial system, the more rapidly a glycoprotein will be cleared from the serum. Because of the absence of sialic acid-specific receptors, however, oligosaccharides with all branches terminated, or "capped," with sialic acid will not promote the clearance of the protein to which they are attached.

[0456] The presence and nature of the oligosaccharide chain(s) on a glycoprotein can also affect important biochemical properties in addition to its recognition by sugar-specific receptors on liver and reticulo-endothelial cells. Removal of the carbohydrate from a glycoprotein will usually decrease its solubility, and it may also increase its susceptibility to proteolytic degradation by destabilizing the correct polypeptide folding pattern and/or unmasking protease-sensitive sites. For similar reasons, the glycosylation status of a protein can affect its recognition by the immune system.

[0457] sHASEGPs can be used to remove the cumulus cells surrounding an egg prior to cryopreservation and other In Vitro fertilization techniques such an intracytoplasmic sperm injection (ICSI). Hyaluronidase can be added to harvested oocytes between 10-200U/ml in buffered salt solutions. Oocytes are separated from the released cumulus cells through aspiration and washed through several washes with media lacking hyaluronidase. The eggs can then be processed for cryopreservation or IVF techniques.

[0458] sHASEGPs are also useful for the more effective penetration of chemotherapeutic agents into solid tumors. sHASEGPs can be injected intratumorally with anti-cancer agents or intravenously for disseminated cancers or hard to reach tumors. The anticancer agent can be a chemotherapeutic, an antibody, a peptide, or a gene therapy vector, virus or DNA. Additionally, sHASEGP's can be used to recruit tumor cells into the cycling pool for sensitization in previously chemorefractory tumors that have acquired multicultural

drug resistance St Croix et al Cancer Lett 1998 Sep 11; 131(1): 35-44). sHASEGPs are also useful to enhance delivery of biologics such as monoclonal antibodies, cytokines and other drugs to tumors that accumulate glycosaminoglycans. Many tumors delete genes involved with the catabolism of glycosaminoglycans such that localized accumulation can prevent antineoplastic agents and the immune system from reaching the tumor mass.

[0459] sHASEGP can also be used to increase the sensitivity of tumors that are resistant to conventional chemotherapy. In one embodiment, sHASEGP is administered to a patient having a tumor associated with a LuCa-1 defect in an amount effective to increase diffusion around the tumor site (e.g., to increase circulation of chemotherapeutic factors (e.g., to facilitate circulation and/or concentrations of chemotherapeutic agents in and around the tumor site), inhibit tumor cell motility (e.g., by HA degradation) and/or to lower the tumor cell(s) threshold of apoptosis (i.e., bring the tumor cell(s) to a state of anoikis), a state that renders the tumor cell(s) more susceptible to the action of chemotherapeutic agents or other agents that may facilitate cell death, preferably preferentially facilitate programmed cell death of cells in anoikis. Chemotherapeutic agents as used herein is meant to encompass all molecules, synthetic (e.g., cisplatin) as well as naturally occurring (e.g., tumor necrosis factor IF)), that facilitate inhibition of tumor cell growth, and preferably facilitate, more preferably preferentially facilitate tumor cell death.

[0460] Of particular interest is the use of sHASEGP for the treatment of metastatic and non-metastatic cancers, particularly metastatic cancers, having decreased to undetectable hyaluronidase activity relative to non-cancerous (normal) cells. sHASEGP can be used as a chemotherapeutic agent (alone or in combination with other chemotherapeutics) in the treatment of any of a variety of cancers, particularly invasive tumors. For example, sHASEGP can be used in the treatment of small lung cell carcinoma, squamous lung cell carcinoma, as well as cancers of the breast, ovaries, head and neck, or any other cancer associated with depressed levels of hyaluronidase or with a defective LuCa-1 (hpHAse) gene (e.g., a LuCa-1 gene that does not provide for expression of adequate hpHAse levels or encodes a defective hpHAse that does not provide for an adequate level of hyaluronidase activity) or other defect associated with decreased hyaluronan catabolism.

sHASEGP is preferable for the treatment of malignancies associated with deficient HA catabolism as it does not require cellular participation for degradation to occur.

[0461] The specific dosage appropriate for administration can be readily determined by one of ordinary skill in the art according to the factors discussed above (see, for example, Harrison's Principles of Internal Medicine, 11th Ed., 1987). In addition, the estimates for appropriate dosages in humans may be extrapolated from determinations of the level of enzymatic activity of sHASEGP in vitro and/or dosages effective in animal studies. For example, 70-300 TRU hyaluronidase is effective in reducing the tumor load in a scid mouse. Given this information, the corresponding dosages in the average 70 kg human would range from about 250,000-1,200,000 TRU hyaluronidase. The amount of sHASEGP administered to a human patient is generally in the range of 1 TRU to 5,000,000 TRU of enzymatic activity, preferably between about 1,000 TRU to 2,500,000 TRU, more preferably between about 100,000 TRU to 1,500,000 TRU, normally between about 250,000 TRU and 1,200,000 TRU, with about 725,000 TRU representing average prescribed doses.

[0462] In one embodiment, a sHASEGP is formulated in a 0.15 M saline solution containing sHASEGP at a concentration of about 150,000 TRU/cc. The formulation is then injected intravenously at 15,000 TRU/kg body weight of the patient. Alternatively, the enzyme formulation may also be injected subcutaneously to allow the hyaluronidase to perfuse around the tumor site. In a preferred embodiment, sHASEGP is injected peritumorally or into the tumor mass. In another preferred embodiment, sHASEGP is formulated as a liposome and is delivered by injection either intravenously or at or near the site of cancerous cells associated with a defect in the LuCa-1 (hpHAse) gene. Injection of sHASEGP intravenously results in sHASEGP in the tumor site. Moreover, Super Sialated sHASEGP is preferably for parenteral administration in that the terminal sialic acids on sHASEGP prevent the clearance of the enzyme from circulation by the reticuloendothelial system. Comparisons of super sialated sHASEGP to non -sialated bovine and ovine hyaluronidases reveal that substantially more favorable pharmacokinetics is achieved.

[0463] FACILITATION OF GENE THERAPY

[0464] The efficacy of most gene delivery vehicles *in vivo* does not correspond to the efficacy found *in vitro*. Glycosaminoglycans can hinder the transfer and diffusion of DNA and viral vectors into many cell types. The levels such extracellular matrix material can hinder the process considerably. Dubensky et al., (Proc Natl Acad Sci U S A 1984 Dec;81(23):7529-33) demonstrated that hyaluronidase when combined with collagenase could facilitate transduction of DNA *in vivo*. It has been demonstrated that adeno associated virus is also amenable to hyaluronidase mediated gene therapy Favre et al, (*Gene Ther* 2000 Aug;7(16):1417-20).

[0465] We have determined herein that channels of defined size in the extracellular matrix are opened with sHASEGP. These pores do not enhance the diffusion of substances greater than about 200-500nm in diameter. However, smaller molecules such as retroviruses, adenoviruses, adeno-associated viruses and DNA complexes are amenable to sHASEGP mediated diffusion.

[0466] Alternatively, viruses can be armed with the sHASEGP gene to facilitate their replication and spread within a target tissue for example. The target tissue can be a cancerous tissue whereby the virus is capable of selective replication within the tumor. The virus can also be a non-lytic virus wherein the virus selectively replicates under a tissue specific promoter. As the viruses replicate, the coexpression of sHASEGP with viral genes will facilitate the spread of the virus *in vivo*.

[0467] Alternatively the nucleic acid of interest and a sHASEGP, can be used simultaneously or consecutively or so as to be staggered over time. Simultaneously refers to a coadministration. In this case, these two essential components can be mixed to form a composition prior to being administered, or can be administered at the same time to the cell or the host organism. It is also possible to administer them consecutively, that is to say one after the other, irrespective of which component of the combination product according to the invention is administered first. Finally, it is possible to use a mode of administration which is staggered over time or is intermittent and which stops and restarts at intervals which may or may not be regular. It is pointed out that the routes and sites of

administration of the two components can be different. According to one particularly preferred embodiment, the sHASEGP is administered before the nucleic acid, with the route of administration of the two components preferably being similar. The time interval between the injections is not critical and can be defined by the skilled person. It is possible to recommend an interval of from 10 min to 72 h, advantageously of from 30 min to 48 h, preferably of from 1 to 24 h and, very preferably, of from 1 to 6 h.

[0468] In addition, the combination product according to the invention can also be combined with one or more molecule(s) which is/are intended to improve the nucleic acid administration. The molecules can be molecules which have a protective effect on the nucleic acid (protection with regard to degradation in the cell), which improve its penetration or its expression in the host cell (fusogenic peptide, nuclear localization signal, etc.), which enable one particular cell type to be targeted (ligand or antibody which recognizes a cell surface protein, etc.), or which prolong the therapeutic effect (immunosuppressive agent, etc.). The combination product can also be combined with agents that facilitate transfection (proteins, etc.).

[0469] The combination product according to the invention can be prepared with a view to local or parenteral administration or to administration by the digestive route. Routes which may in particular be mentioned are the intragastric, subcutaneous, intracardiac, intravenous, intraperitoneal, intrasynovial, intratumor, intrapulmonary, intranasal and intratracheal routes, and, very particularly, the intramuscular route. The administration can be effected by means of any technique of the art (injection, oral route, aerosol, instillation, etc.), as a single dose or as a dose that is repeated once or several times after a particular time interval. The route of administration can be adjusted to suit the gene of interest to be transferred and the disease to be treated. The formulation can include pharmaceutically acceptable vehicles (excipients, adjuvants, etc.). The substance leading to disorganization of the extracellular matrix and the nucleic acid of interest are preferably dissolved in a buffer which is suitable for pharmaceutical use and which can be hypertonic, hypotonic or isotonic. Various buffers can be envisaged. Those which may be mentioned by way of illustration are a physiological saline solution (0.9% NaCl), a nonphysiological saline solution (1.8% NaCl), a Hepes-Ringer solution, a Lactate-Ringer solution, a buffer which is based on Tris-HCl (10 mM Tris-HCl, pH 7.5 to 8, 1 mM EDTA; 10 mM Tris-HCl, pH

7.5 to 8, 1 mM MgCl.sub.2), a phosphate buffer (Krebs phosphate H.sub.2 O buffer), a sugar (glucose, sucrose, trehalose, etc.) solution, or simply water.

[0470] HYPODERMOCLYSIS

[0471] Hypodermoclysis, the subcutaneous infusion of fluids, is a useful and easy hydration technique suitable for mildly to moderately dehydrated adult patients, especially the elderly. The method is considered safe and does not pose any serious complications. The most frequent adverse effect is mild subcutaneous edema that can be treated by local massage or systemic diuretics. Approximately 3 L can be given in a 24-hour period at two separate sites. Common infusion sites are the chest, abdomen, thighs and upper arms. The preferred solution is normal saline, but other solutions, such as half-normal saline, glucose with saline or 5 percent glucose, can also be used. Potassium chloride can be added to the solution bag if needed. Additionally, other drugs can be delivered through similar routes. Human sHASEGP can be added to enhance fluid absorption and increase the overall rate of administration. Human sHASEGP is prefereable for repeated Hypodermoclysis over slaughter house-derived enzymes in that it not likely to be immunogenic as the bovine enzyme is known to be. It may be administered at home by family members or a nurse; the technique should be familiar to every family physician.

[0472] In ambulatory patients, hypodermoclysis sites include the abdomen, upper chest, above the breast, over an intercostal space and the scapular area. In bedridden patients, preferred sites are the thighs, the abdomen and the outer aspect of the upper arm. After one to four days, the needle and tubing should be changed, although infusion sets have been left in place for much longer periods without complications. Administration of 500-mL boluses over one or two hours three times a day can also be given, with 150 U of sHASEGP given at the subcutaneous site before the first morning infusion

[0473] FACILITATION OF THERAPEUTIC INJECTIONS.

[0474] Many molecules injected percutaneously reach circulation slowly or with very low efficiency. Several factors regulate the pharmacokinetics and pharmacodynamics of molecules injected subcutaneously (SC) or intramuscularly (IM). Generally, larger molecules reach circulation more slowly and less efficiently without active transport into

circulation. Subcutaneous bioavailability is determined by calculating the ratio of area under the curves for SC verses intravenous administration ($AUC_{SC}/AUC_{intravenous}$). A second factor is charge and affinity for matrix molecules that may play a role in sequestration of molecules subcutaneously. If these materials are degraded locally they may never reach their desired targets and thus demonstrate a decreased overall systemic bioavailability to the target organs.

[0475] Large proteins are normally given intravenously so the medicament directly available in the blood stream. It would however be advantageous if a medicament could be given subcutaneously, intramuscularly or intradermally as these administration forms are much easier to handle for the patient. Especially, if the medicament must be taken regularly during the whole life and treatment is to start early, already when the patient is a child. However, a medicament with a very large and labile molecule, such as coagulations factor VIII of 170 to 300 kDa, have normally a very low bioavailability if given subcutaneously, intramuscularly or intradermally, since the uptake is not enough and degradation is severe.

[0476] In addition to the need to increase bioavailability of many subcutaneously administered biologics, more rapid pharmacokinetics is also critically important in instances of emergency medicine. The time required to reach intravenous access in many patients can prevent an otherwise rapid acting drug when administered systemically from being utilized. In some cases failure to reach intravenous access is then followed by subcutaneous injection, which leads to additional delay in reaching the target organs. Thus, the more rapid availability of subcutaneous drugs would be of benefit as a first line of treatment rather than to risk the time required to achieve intravenous access. Examples of molecules that can be delivered subcutaneously as well as intravenously include epinephrine, atropine, narcan, lignocaine, and dextrose.

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[0480] An additional benefit of the invention lies in the ability to deliver equivalent or larger volumes of solutions SC or IM without the pain and morbidity associated with the pressure and volume of the solution at the site of injection.

[0481] VITREOUS HEMORHAGE

[0482] In an effort to minimize the potential for causing further detachment or tearing of the retina during performance of vitrectomy, it has previously been proposed in U.S. Pat. No. 5,292,509 (Hageman), to inject certain protease-free glycosaminoglycanase enzymes into the vitreous body, to cause the vitreous body to become uncoupled or "disinserted" from the retina, prior to removal of the vitreous body. Such disinsertion or uncoupling of the vitreous body is purported to minimize the likelihood that further tearing or detachment of the retina will occur as the vitreous body is removed. Examples of specific protease-free glycosaminoglycanase enzymes which may be used to bring about this vitreal disinsertion purportedly include; chondroitinase ABC, chondroitinase AC, chondroitinase B, chondroitin 4-sulfatase, chondroitin 6-sulfatase, hyaluronidase and beta-glucuronidase.

[0483] Although hyaluronidase enzyme has been known to be usable for various ophthalmic applications, including the vitrectomy adjunct application described in U.S. Pat. No. 5,292,509 (Hageman), published studies have indicated that the hyaluronidase enzyme may itself be toxic to the retina and/or other anatomical structures of the eye. See, The Safety of Intravitreal Hyaluronidase; Gottleib, J. L.; Antoszyk, A. N., Hatchell, D. L. and Soloupis, P., Invest Ophthalmol Vis Sci 31:11, 2345-52 (1990). Moreover, the used of impure slaughterhouse preparations of hyaluronidase can cause uveitis or inflammation of the eye. The use of human sHASEGP is thus preferable in both its increased potency, purity and lack of animal origin that can give rise to immunogenic reactions and antibody mediated neutralization following repeated administration. In another embodiment, a pegylated form of a sHASEGP can be injected into the eye. Such a pegylated sHASEGP is not cleared from the vitreous in such a rapid fashion and maintains its activity in the vitreous for a longer period of time.

[0484] The ophthalmic toxicity of some hyaluronidase preparations has been confirmed by other investigators, who have proposed that such hyaluronidase preparations be used as a toxic irritant for causing experimentally induced neovascularization of the eye, in animal toxicity models, (see An Experimental Model of Preretinal Neovascularization in the

Rabbit; Antoszyk, A. N., Gottleib, J. L., Casey, R. C., Hatchell, D. L. and Machemer, R., Invest Ophthalmol Vis Sci 32:1, 46-51 (1991). The use of a highly purified sHASEGP devoid of mercury-based and cattle or bacterially derived contaminants is preferable for intraocular procedures. Moreover, a recombinant human sHASEGP is preferable over slaughterhouse derived preparations in both purity lack of bovine pathogens and reduced risk of immunogenicity. Most preferably a pegylated sHASEGP is envisioned.

[0485] An enzymatic method using a human sHASEGP is thus provided for treating ophthalmic disorders of the mammalian eye. In one embodiment of the investion, said sHASEGP is PEGylated to prolong its residence within the vitreous and prevent localized uptake. Prevention of neovascularization, and the increased rate of clearance from the vitreous of materials toxic to retina, are accomplished by administering an amount of hyaluronidase effective to liquefy the vitreous humor of the treated eye without causing toxic damage to the eye. Liquefaction of the vitreous humor increases the rate of liquid exchange from the vitreal chamber. This increase in exchange removes those materials and conditions whose presence causes ophthalmologic and retinal damage.

[0486] COSMETIC USES OF sHASEGP

[0487] It is known that hyaluronidase has the effect of depolymerizing the long mucopolysaccharide chains of the fundamental substance, responsible for the retention of bound water and of the slowing, by capillary compression, of the diffusion of organic liquids, which eliminate metabolic wastes. Such retention of water and wastes associated with fat overloading of the lipocytes, constitutes classical "pigskin" edema or "orange peel" edema. This depolymerization will therefore cut the long chains of mucopolysaccharides into shorter chains, whence the elimination of the bound water, of wastes, restoration of the venous and lymphatic circulation and disappearance of local edema.

[0488] Use of sHASEGP by way of subcutaneous administration is thus preferred for the removal of glycosaminoglycans involved in the accumulation of so-called cellulite and to promote lymphatic flow. Human sHASEGP is preferred for the treatment of cellulite in that it is capable of removal of said glycosaminoglycans without the inflammatory

components of slaughter house derived proteins and is of high purity and is not likely to be immunogenic. The sHASEGP can be administered through repeated subcutaneous injections, through transdermal delivery in the form of ointments or creams or through the use of injectable slow release formulations to promote the continual degradation of glycosaminoglycans and prevent their return.

[0489] ORGAN TRANSPLANTATION

[0490] Hyaluronan has several biological effects, that are in part related to its molecular size (West, D.C., Kumar, S. Exp.Cell. Res. 183, 179-196, 1989). The content of hyaluronan in an organ increases in different conditions of inflammation of that organ. Thus, an increased concentration of hyaluronan has been shown in tissue from different organs characterized by inflammatory-immunological injury such as alveolitis (Nettelbladt 0 et al, Am Rev Resp Dis 1989; 139: 759-762) and myocardial infarction (Waldenstrom et al, J Clin Invest 1991; 88(5): 1622-1628). Other examples are allograft rejection after a renal (Ha'llgren et al, J Exp Med 1990a; 171: 2063-2076; Wells et al, Transplantation 1990; 50: 240-243), small bowel (Wallander et al, Transplant Int 1993; 6: 133-137) or cardiac (Hällgren et al, J Clin Invest 1990b;85:668-673) transplantation; or a myocardial inflammation of viral origin (Waldenstrdm et al, Eur J Clin Invest 1993; 23: 277-282).

[0491] The occurrence of interstitial edemas in connection with the grafting of an organ constitutes a severe problem in the field of transplantation surgery. As much as 25% of the grafts, will swell to such a degree that the function will temporarily be lost. Moreover, in 2-3% of the cases, the swelling causes disruption of the kidney, resulting in a massive haemorrhage.

[0492] SHASEGP may be used to degrade accumulated glycosaminoglycans in an organ transplant. Removal of such glycosaminoglycans promotes removal of water from the graft and thus organ function. Dose ranging from 500-10,000 Units/kg may be administered to reduce interstitial pressure as such.

[0493] Pathologic Accumulations of Glycosaminoglycans in the Brain

[0494] Hyaluronan levels are elevated in a number of cerebrospinal pathologic conditions. Levels of cerebrospinal hyaluronan are normally less than 200ug/L in adults (Laurent et al, Acta Neurol Scand 1996 Sep;94(3):194-206). These levels can elevate to over 8,000ug/L in diseases such as meningitis, spinal stenosis, head injury and cerebral infarction. Thus administration of sHASEGP by either intrathecal delivery or systemic injection of super sialated sHASEGP can be utilized to degrade critically elevated levels of substrate.

[0495] The lack of effective lymphatics in the brain can also lead to life threatening edema following head trauma. Hyaluronan accumulation is a result of increased synthesis by HA synthases, and decreased degradation. Accumulation of hyaluronan serves the purposed of increasing water content in the damaged tissue to facilitate leukocyte extravasation but can be lethal. Administration of human sHASEGP to a patient suffering from head trauma can thus removal tissue hyaluronan accumulation and the water associated with it. Human sHASEGP can be administered intrathecally through a shunt or alternatively, Super Sialated sHASEGP can be administered intravenously to reach the brain tissue.

[0496] Following and ischemic of the brain as occurs in stroke, the hyaluronan content increases dramatically due to increased expression of HA synthases and decreased catabolism. Failure of ion pumps and leakage of plasma into the interstitium results in fluid retention that if not properly cleared by the lymphatics, results in tissue necrosis. Some groups have attempted to prevent interstitial fluid accumulation following ischemia reperfusion by blocking vascular permeability. However, once the fluid has extravasated, preventing vascular permeability can prevent resolution of edema and exacerbate conditions.

[0497] Human sHASEGP can also be used in the treatment of edema associated with brain tumors, particularly that associated with glioblastoma multiform. The edema associated with brain tumors results from the accumulation of hyaluronan in the non-cancerous portions of the brain adjacent the tumor. Administration of hyaluronidase to the sites of hyaluronan accumulation (e.g., by intravenous injection or via a shunt) can relieve the edema associated with such malignancies by degrading the excess hyaluronan at these sites. Thus, hyaluronidase is successful in the treatment of brain tumors not only in the reduction of the tumor mass and inhibition of tumor growth and/or metastasis, but it also is

useful in relieving edema associated with the malignancy. Human sHASEGP can be administered for treatment of edema in a manner similar to that for administration of bovine testicular hyaluronidase to treat edema (see, e.g., Sa Earp Arq. Braz. Med. 44:217-20).

[0498] TREATMENT OF GLYCOSAMINOGLYCAN ACCUMULTATION IN CARDIOVASCULAR DISEASE

[0499] It has been shown that the administration of hyaluronidase in animal models following experimental myocardial infarct can reduce infarct size (Maclean, et. al Science 1976 Oct 8;194(4261):199-200). The proposed mechanism by which bovine hyaluronidase reduces infarct size in animals is by reducing hyaluronan accumulation that occurs following ischemia reperfusion. Reduction of infarct size is believed to occur from increased lymph drainage and increased tissue oxygenation and reduction of myocardial water content. While reduced infarct size could be obtained in animal models, the benefits were not realized in larger clinical studies in humans. Bovine testes hyaluronidase possesses a remarkably short serum half life of approximately 3 minutes in animals and man Wolf, et. al., J Pharmacol Exp Ther 1982 Aug;222(2):331-7. This short half-life is due to the terminal mannose residues that are readily recognized by the scavenger receptors of the reticuloendothelial system. While small animals may benefit from hyaluronidase due to a smaller vascular bed, an enzyme with increased half-life is needed. Super sialated sHASEGP possesses more favorable pharmacokinetics due to sialation for which there is no scavenger receptor. Super sialated sHASEGP in doses ranging from 100-200,000 Units /kg may be utilized to facilitate resolution of excess hyaluronan following ischemia reperfusion and to reduce infarct size.

[0500] Super sialated sHASEGP may also be used to limit coronary plaques from arteriosclerosis. Such plaques accumulate glycosaminoglycans and mediate macrophage and foam cell adhesion Kolodgie et al, Arterioscler Thromb Vasc Biol. 2002 Oct 1;22(10):1642-8. Administration of Super Sialated sHASEGP can be used to reduce plaque formation. As repeated administration of hyaluronidase is contemplated at doses from 100-100,000U/kg, the need to utilize a human recombinant protein with low risk of immunogenicity and increased half-life will result in superior reduction of plaques.

[0501] TREATMENT OF PERIPHERAL TISSUES NECROSIS

[0502] Tissue necrosis occurs in many diseases due to venous insufficiency. The lack of sufficient oxygenation is one of the main obstacles for regrowth of the tissue. It has been demonstrated that intra-arterial hyaluronidase treatment significantly improves the clinical picture in patients with peripheral arterial occlusive disease (Elder et. al, Lancet (1980) 648–649). sHASEGP can be injected intra-arterially 3-5 times a week at doses from 10-200,000 Units.

[0503] ENHANCEMENT OF ANESTHESIA

[0504] Slaughterhouse-derived hyaluronidase is commonly used for peribulbar block in local anesthesia prior ophthalmic surgery. The presence of the enzyme prevents the need for additional blocks and speeds the time to the onset of akinesia (loss of eye movement). Peribulbar and sub-Tenon's block are the most common applications of hyaluronidase for ophthalmic procedures. Since the discontinuation of Wydase®, reports of increased diplopia and ptosis have been reported with peribulbar block (Brown et al *J Cataract Refract Surg* 1999; 25:1245-9).

[0505] With Wyeth's discontinuance of Wydase®, bovine testes-derived hyaluronidase material is now supplied by compounding pharmacies. However, there are several concerns with using an extemporaneously compounded sterile product.http://www.ashp.org/shortage/hyaluronidase.cfm?cfid=11944667&CFToken=9426953 - ref#ref Compounded preparations are not FDA-approved products. As such, the FDA has no control over the quality or consistency of the manufacturing process.

[0506] SHASEGP from 10-500 Units can be mixed directly with 5ml 2% lidocaine (Xylocaine), 5 ml 0.5% bupivacaine (Marcaine) and optionally with epinephrine 1:200,000. sHASEGP can be used to increase the onset of akinesia and to remove the need for additional blocks. sHASEGP is also ideal for akinesia for cosmetic surgery in blepharoplasties and face lifts. sHASEGP can also be utilized following such surgical procedures to diffuse anti inflammatories and to reduce tissue swelling.

[0507] SHASEGP may also be mixed with a buffering solution such as bicarbonate to prevent discomfort during the injection procedure. SHASEGP can also be mixed with anesthesia for lacerations to both reduce the total volume of material required for injection and to reduce pain from swelling of tissue.

[0508] REDUCTION OF INTRAOCULAR PRESSURE

[0509] A common side effect occurring in postoperative cataract patients is a significant early, and occasionally prolonged, rise in intraocular pressure. Such a condition is sometimes serious, especially in patients with glaucomatous optic disc changes. Although the pressure increase tends to be more severe when visco-elastic agents such as hyaluronic acid are injected into the eye during surgery, the intraocular pressure can become elevated postoperatively even when such agents are not utilized. Furthermore, such a pressure increase can occur even when no additional medications are used during the surgical procedure. In some cases, it is advantageous to leave a viscoelastic agent in the eye, which often necessitates giving patients large doses of carbonic anhydrase inhibitors. These inhibitors lower the intraocular pressure by decreasing the formation of aqueous humor, a fluid that is normally secreted in the eye, by the ciliary body. Current methods for relieving postoperative pressure increases in the eye include various types of eye drops such as beta-adrenergic blocking agents, sympathomimetic agents, miotics, alpha II selective agents, carbonic anhydrase inhibitors and prostaglandin agents.

[0510] A preferred method of removing the viscoelastic such as hyaluronic acid is by injection of sHASEGP during or immediately following anterior segment or posterior segment surgical procedures, although other methods of administration known in the art are possible as well. It is preferred if the hyaluronic acid and the sHASEGP are administered by injection into the anterior chamber during anterior segment ocular surgical procedures to allow the hyaluronic acid to act as a spacer during the start of the surgical procedure. In some cases of corneal transplantation, the hyaluronic acid and sHASEGP combination may be placed on the surface of the intraocular structures prior to suturing the corneal transplant in place. This combination may also be used in posterior segment surgery, such as retina or vitreous surgery.

[0511] In some cases, it may be advisable to leave a visco-elastic agent such as Healon.TM., Viscoat.TM., or other space-occupying substances in the anterior chamber of the eye at the conclusion of surgery. This is especially true in positive pressure rise when the intraocular contents tend to come forward and press against the posterior surface of the cornea. If this occurs in an eye with a synthetic intraocular lens in place, pressure on the corneal endothelium can cause significant damage to the cells and subsequent corneal swelling and opacification can occur, which are associated with decreased vision. Typically, if a patient's intraocular pressure is significantly elevated at the conclusion of the operative procedure, it is necessary to give such a patient large doses of carbonic anhydrase inhibitors, as well as topical eye drops such as beta-blockers and alpha II agonists in order to decrease aqueous formation and/or to increase aqueous outflow. These agents all have significant side effects and, in some instances, are contraindicated in patients with various types of medical conditions such as breathing problems, heart disease or high blood pressure. However, the use of sHASEGP in these situations will eliminate the necessity of giving these patients large doses of such drugs.

[0512] Furthermore, there is a significant amount of hyaluronic acid in the trabecular meshwork. The sHASEGP will break this down and therefore improve the outflow of the aqueous through the trabecular meshwork. The patient's intraocular pressure will therefore decrease. The combination of sHASEGP with other anterior chamber agents, such as a methylcellulose (Ocucoat.RTM. for example, commercially available from Storz Instrument Co.), used as spacers and/or protective agents in cataract surgery, will also be efficacious in preventing significant pressure rises because it will in effect open the trabecular meshwork and allow more aqueous humor drainage by breaking down a significant amount of the hyaluronic acid present in the trabecular meshwork.

[0513] Removal of glycosaminoglycans from the trabecular meshwork is also useful for the reduction of intraocular pressure in individuals suffering form open angle glaucoma. Human sHASEGP can be administered by subconjunctuval injection or injection directly in the anterior chamber.

[0514] GANGLION CYSTS

[0515] The ganglion cyst (also known as a wrist cyst, Bible cyst, or dorsal tendon cyst) is the most common soft tissue mass of the hand. It is a fluid filled sac that can be felt below the skin. It is usually attached to a tendon sheath (lining which lubricates the tendon) in the hand or wrist or connected with an underlying joint; however, some have no obvious connection to any structures. These may also occur in the foot. It often occurs when there is a tear in the ligaments overlying the lining of tendons or joints and the lining herniates out of the ligamentous defect causing a bump under the skin. Because there is often inflammation associated, the inflamed tissue produces a jelly-like fluid that fills the protruding sac. They may be rock hard due to a high pressure of the mucous like fluid contained within the cyst, and are often mistaken for a bony prominence.

[0516] sHASEGP can be used to ameliorate ganglion cysts. Intralesional injection of sHASEGP from 5-1000 Units followed by fine needle aspiration will remove the cyst without the need for surgery. Corticosteroids may be optionally injected as well with the sHASEGP. Additional injection may be required for some patients.

[0517] MYXEDEMA

[0518] Glycosaminoglycan (GAG) infiltration of the skin is a feature of hyperthyroidism, hypothyroidism, pretibial myxedema, scleromyxedema, and scleredema. Hyaluronic acid is the main GAG in all the conditions and in normal skin. There is minimal histologic variability of GAG dermal distribution. The acquired cutaneous mucinoses exhibit similar skin GAG distribution and biochemical composition. The morphologic differences in fibroblastic activity suggest that the mucinoses of scleredema and scleromyxedema represent a local process, whereas the GAG infiltration of thyroid diseases may have a systemic origin. These disorders may be ameliorated with a sHASEGP from both a local and systemic route of administration. For chronic therapy, a PEGylated sHASEGP may be envisioned.

[0519] PULMONARY USES OF sHASEGP

[0520] Levels of Hyaluronan in broncheoalveolar lavages (BAL) from normal individuals are generally below 15ng/ml. However, BAL levels rise dramatically in conditions of respiratory distress (Bjermer *Br Med J* (Clin Res Ed) 1987 Oct 3;295(6602):803-6). In ARDS for example, hyaluronan levels can increase to 500ng/ml whereas in farmers lung, BAL levels can surpass 1000ng/ml (Hallgren et al *Am Rev Respir Dis*. 1989 Mar;139(3):682-7), (Larrson et al *Chest*. 1992 Jan;101(1):109-14). The increased hyaluronan in the lung can prevent oxygen diffusion and gas exchange as well as activating neutrophil and macrophage responses.

[0521] Bovine preparations of hyaluronidase are no preferable for the treatment of such conditions for a number of reasons. First, slaughterhouse testes-derived preparations of hyaluronidase are known to be contaminated with serine proteases such as acrosin. Secondly, the foreign nature of the bovine enzymes increase the probability of an anaphylactic reaction, which could result in death of the patient. Thus a highly purified preparation of recombinant human sHASEGP can be delivered by either pulmonary or intravenous delivery. Human sHASEGP can also be administered to patients suffering from other pulmonary complications that are associated with elevated glycosaminoglycans or to enhance the delivery of other co delivered molecules to the lung.

[0522] The invention will now be described in greater detail by reference to the following non-limiting examples

[0523] EXAMPLE 1

[0524] MICROTITER BASED HYALURONIDASE ASSAYS

[0525] The following example provides for a rapid assay for measurement of the hyaluronidase activity of sHASEGP. This assay can be related to the TRU, the IU or NFU through use of a W.H.O. standard preparation of hyaluronidase.

[0526] BIOTINYLATED HYALURONAN MICROTITER ASSAY

[0527] The free carboxyl groups on glucuronic acid residues of Hyaluronan are biotinylated in a one step reaction using biotin-hydrazide (Pierce), Sulfo NHS (Pierce) and 1-Ethyl dimethylaminopropyl-carbodiimide (Sigma). This biotinylated HA substrate is covalently coupled to a 96 well microtiter plate in a second reaction. At the completion of the enzyme reaction, residual substrate is detected with an avidin-peroxidase reaction that can be read in a standard ELISA plate reader. As the substrate is covalently bound to the microtiter plate, artifacts such as pH-dependent displacement of the biotinylated substrate does not occur. The sensitivity permits rapid measurement of Hyaluronidase activity from cultured cells and biological samples with an inter-assay variation of less than 10%.

[0528] a. Protocol

[0529] PREPARATION OF BIOTINYLATED HA SUBSTRATE

[0530] One hundred mg of HA (Sigma Chemicals) was dissolved in 0.1 M MES, pH 5.0, to a final concentration of 1 mg/ml and allowed to dissolve for at least 24 hr at 4°C prior to coupling of biotin. Sulfo-NHS (Pierce; Rockford IL) was added to the CS04 MES solution to a final concentration of 0.184 mg/ml. Biotin hydrazide (Pierce) was dissolved in DMSO as a stock solution of 100 mM and added to the CS04 solution to a final concentration of 1 mM. A stock solution of 1-ethyl -3-(3-dimethylaminopropyl) carbidodiimide (EDAC) was prepared as a 100 mM stock solution in distilled water and added to the HA biotin solution at a final concentration of 30 mM. This solution was left stirring overnight at 4°C. Unlinked biotin and EDAC were removed by dialysis against water with 3 changes of 1000x volume of water. The dialyzed, biotinylated HA (bHA) was aliquoted and stored at -20°C for up to several months.

[0531] Sulfo-NHS was diluted to 0.184 mg/ml in water with the bHA at a concentration of 0.2 mg/ml and pipetted into 96 well COVALINK-NH plates (NUNC; Placerville NJ) at 50 µl per well. EDAC was diluted to 0.123 mg/ml in water and pipetted into the COVALINK-NH plates with the bHA solution resulting in a final concentration of 10 µg/well bHA and 6.15 µg/well EDAC. The plates were incubated overnight at 4°C or

for 2 hr at 23°C, which gave comparable results. After covalent immobilization of bCS04 on the microtiter plates, the coupling solution was removed by shaking and the plates were washed 3 times in PBS containing 2M NaCl and 50 mM MgSO4 (Buffer A). The plates could be stored at 4°C for up to one week.

[0532] The COVALINK-NH plates with immobilized bHA were equilibrated with 100 µl/well assay buffer - either 0.1 M formate, pH 3.7, 0.1 M NaCl, 1% TRITON X-100 detergent, 5 mM saccharolactone for lysosomal Hyaluronidase; or 10mM Hepes PH 7.4 with 1mM CaCl2 and 1mg/ml Human Serum Albumin (ICN) for neutral-active enzymes. A set of standards for the calibration of enzyme activity against "relative Turbidity Reducing Units" (rTRU's) was generated by diluting bovine testicular hyaluronidase (Sigma Type VI-S) in neutral enzyme buffer from 1.0 to $1x10^{-6}$ rTRU/well and assaying 100 µl/well in triplicate. Samples of acid-active Hyaluronidase were diluted in lysosomal assay buffer from 1:10 to 1:130,000 were pipetted in triplicate at 100 µl/well. For most assays of tissue extracts and human plasma, a 30 min incubation at 37°C was sufficient. Positive and negative control wells (no enzyme or no ABC (see below), respectively) were included in triplicate.

[0533] The reaction was terminated by the addition of 200 µl/well of 6M Guanidine HCl followed by three washes of 300 µl/well with PBS, 2 M NaCl, 50 mM $MgSO_4$, 0.05% TWEEN 20 detergent (Buffer B). An avidin biotin complex (ABC) kit (Vector Labs; Burlingame CA) was prepared in 10 ml of PBS containing 0.1% TWEEN 20 detergent, which was preincubated for 30 min at room temperature during the incubation. The ABC solution was added (100 µl/well) and incubated for 30 min at room temperature. The plate was washed five times with Buffer B, then an o-phenylenediamine (OPD) substrate was added at 100 µl/well by dissolving one 10 mg tablet of OPD in 10 ml of 0.1 M citrate-PO_4 buffer, pH 5.3 and adding 7.5 µl of 30% H_20_2. The plate was incubated in the dark for 10-15 min, then read using a 492 nm filter in an ELISA plate reader (Titertek Multiskan PLUS; ICN) monitored by computer using the Delta Soft II plate reader software from Biometallics (Princeton NJ). A standard curve using the bovine testicular hyaluronidase was generated by a four parameter curve fit of the commercial hyaluronidase preparation and unknown samples were interpolated through their absorbance at 492 nm.

[0534] To analyze pH dependence of Hyaluronidases, purified recombinant sHASEGP and bovine testicular hyaluronidase are used. The pH dependence of enzyme activity is measured by diluting purified sHASEGP or partially purified bovine testicular hyaluronidase to 0.1 rTRU in the following buffers: 50 mM formate, pH 3-4.5; 50 mM acetate, pH 5-6; 50 mM MES, pH 6-7; or 50 mM HEPES, pH 7-8. Samples are assayed for 30 min at 37°C and activity was expressed as a percent of maximal activity. NaCl was not used in buffers, as it can alter the pH optima of testicular hyaluronidase preparations (Gold, *Biochem. J.* 205:69-74, 1982; Gacesa et al. *Biochem. Soc. Trans.* 7:1287-1289, 1979); physiological salt concentrations (0.15 M) decreased the apparent pH optimum, an effect that was more pronounced in purified preparations of the testicular enzyme than in the original crude sample.

[0535] b. Results

[0536] Hyaluronan was biotinylated in a one step reaction using biotin-hydrazide and EDAC. By limiting the EDAC, which couples the free carboxyl groups on HA with biotin hydrazide, only a small fraction of the total glucuronic acid residues on HA were labeled. This amount of EDAC (3×10^{-5} M) added to HA (2.8×10^{-3} M) results in a maximum of one molecule of biotin hydrazide coupled per 93 disaccharide units of HA.

[0537] A four-parameter curve fit of bovine testicular hyaluronidase standard reactions measured at pH 3.7, and diluted from 1.0 to 1×10^{-6} TRU/well, was prepared. Four parameter curve fits were established from the equation y=((A-D)/(1+ (conc/C)^B)) + D) where $\log_{it}$ y= ln (y'/1-y'), y'= (y-D)/(A-D), B=-b/ln 10 and C=EXP (a/B). The four parameters (A,B,C,D) were calculated with a software program that utilized the 2+2 algorithm with linear regression (Rodbard et al., *Clin. Chem.* 22:350, 1976). This curve fit incorporates the sigmoidal aspects the standard curve. Optimal accuracy for measurement of a sample typically occurs from 0.001 to 0.01 TRU/well for a 30min incubation. During a 60 min incubation, 1/1000th of a TRU is detectable. A standard logarithmic curve also can be utilized over a shorter range of values to establish a standard curve fit. Although the invention has been described in connection with specific preferred embodiments, it should be understood that the invention as claimed should not be unduly limited to such specific embodiments.

[0538] EXAMPLE 2

[0539] CLONING OF sHASEGP cDNA

[0540] Nucleic acid encoding Human sHASEGP may be obtained by one skilled in the art through a number of procedures including, but not limited to, artificial gene synthesis, RT-PCR, and cDNA library hybridization (for example see, Gmachl et al *FEBS* 336(3) 1993, Kimmel et al., *Proc. Natl. Acad. Sci. USA* 90 1993 10071-10075). Alternatively, clones encoding human sHASEGP may be obtained from IMAGE, or other suppliers of human gene sequences (Invitrogen Clone ID IOH10647).

[0541] The full length human PH20 cDNA was calculated to be 2009 nucleotides in length and contained an open reading frame of 1530 nucleotides. The 5' UTR is unusually large, which can indicate a retained intron and can inhibit translation by preventing the ribosome from binding to the correct initiating methionine codon due to 9 non coding start codons in the 5'UTR. The protein (Genbank Accession number NP_003108) is predicted to comprise 509 amino acids SEQ ID No. 1 with a calculated molecular mass of 58kDa.

[0542] For sequencing of clones, PCR amplified bands were excised, and eluted with the Gel Extraction Kit (Qiagen) and cloned into the appropriate vectors with compatible ends after restriction digestion. All sequencing reactions were performed on double stranded DNA with the *Taq* dye deoxy terminator cycle sequencing kit (Applied Biosystems) according to the manufacturer's instructions, and run on an ABI Prism™ automated sequencer (Applied Biosystems).

[0543] The human PH-20 open reading frame was obtained by amplifying a human testis cDNA library (Clontech, Palo Alto CA) by Polymerase Chain Reaction using primers SEQ ID NO 14 and SEQ ID NO 47. PCR products were digested with *Nhe*I and *Bam*HI and cloned into the *Nhe*I and *Bam*HI sites of the vector IRESpuro2 (Clontech).

[0544] EXAMPLE-4

[0545] ISOLATION OF SHASEGP FROM HUMAN PH20 cDNA

[0546] A catalytically active secreted recombinant human sHASEGP expression vector capable of effective glycosylation in mammalian cells was generated as described below. Other expression constructs with promoters and selection genes for different species such as yeast and insect cells that are also capable of generating sHASEGP are contemplated. Positive selection genes such as Glutamine Synthase or Dihydrofolate Reductase (DHFR) may also be used. The examples given below is not intended to restrict but is rather provided as an example of several plasmid expression systems that may be used.

[0547] In order to construct secreted forms of sHASEGP, truncation mutants that lack the hydrophobic C terminal end were constructed. Using a GPI cleavage prediction program the GPI anchor cleavage site was located around amino acid position N 483 in the full-length GPI-anchored protein. A set of seven nested 3' primers were used to construct a set of seven truncated deletion mutants lacking predicted GPI anchor starting at position Y 482 and deleted progressively by one amino acid. These primers were designed to have compatible Nhe1 (5') and BamH1 (3') sites to clone the truncation mutants in vector Irespuro2 either untagged with a stop codon in the 3' primer, or as a C terminus His tagged protein for ease of purification and detection. For example reverse primers SEQ ID No. 8, SEQ ID No. 9, and SEQ ID No. 10 were used to generate deletion mutants ending at position Y 482, F 481 and I 480 without a 6 His tag. Other mutant primers were generated with the same base design with the appropriate modifications to include and exclude the particular amino acids. For generating His-tagged variants the same set of primers are used as for non tagged variants except that primers lack the stop codon in the respective reverse primers, the forward primer remaining the same (for His tagged construction refer to primers with SEQ ID No 19, 20, 21, 22, 23,24 and 25 which are the reverse primers without stop codon corresponding to non tagged reverse primers for their respective constructs). Overlapping primers were used to construct a six amino acid spacer followed by hexahistidine within BamH1 and Not1 sites in Irespuro2 vector such that His-tagged mutants were generated by ligation of the PCR amplified and restriction

digested products within the Nhe1 and BamH1 sites in the his tag containing Irespuro2 vector.

[0548] To identify whether human sHASEGP could be modified at its carboxy terminus to generate a secreted and neutral active enzyme, a series of truncations were made from the GPI anchor attachment site to the predicted "catalytic domain" based upon homology with the bee venom enzyme.

[0549] DNA encoding the human sHASEGP full length GPI anchored clone in IRESPuro2 was used as a template to generate the various truncated deletion mutants. Software modeling programs gave several predicted cleavage sites for the full length polypeptide. One of such predicted sites was at amino acid position N 483 (SEQ ID No. 1). PCR primers were designed to successively truncate the protein from N483 to generate six deletion mutants starting at Y 482 (lacking N) and ending at E 477(lacking P).

[0550] a. Protocol

[0551] Generating truncation mutant lacking N483:

[0552] The full length GPI anchored sHASEGP clone between Nhe1 and BanH1 site in pIRESPuro2 was used as a template. This template was amplified with 5' primer containing NheI site that starts at starting Methionine of the native signal peptide at M 1(SEQ ID No. 14), and a 3' primer containing BamHI site that ends at Y 482 (SEQ ID No. 8). The PCR product was ran on a 1% agarose gel to resolve and confirm the correct sized amplified band, gel purified, restriction digested with NheI and BamHI and cloned into vector pIRESPuro2 (Clontech) between NheI and BamHI sites generating an expression vector for expressing this truncation mutant of SHASEGP ending at amino acid position N482 and lacking the GPI anchor with amino acid sequence (SEQ ID No. 5 for the sequence of the resulting polypeptide of sHASEGP up to Y 482) and nucleotide sequence (SEQ ID No. 48 – coding nucleotides for polypeptide in SEQ ID No. 5) as indicated.

[0553] Generation of the other truncation mutants lacking Y 482, F 481, I 480, Q 479, and P 478 respectively.

[0554] The same strategy was used with the only difference being using the appropriate 3' primer for each mutant. The respective 3' primers are as follows:

[0555] 3' primer for sHASEGP mutant that lacks the Y 482 -SEQ ID No. 9

[0556] 3' primer for mutant that lacks the F 481 –SEQ ID No. 10

[0557] 3' primer for mutant that lacks the I 480 –SEQ ID No. 11

[0558] 3' primer for mutant that lacks the Q 479 –SEQ ID No. 12

[0559] 3' primer for mutant that lacks the P 478 –SEQ ID No. 13

[0560] Generating further deletion mutants to determine the minimally active domain of sHASEGP:

[0561] Further deletions, in blocks of ten to twenty amino acids were made from the 3' end of innermost neutral pH active truncation mutant of sHASEGP, which is sHASEGP up to E 477. The NheI forward primer SEQ ID No. 14) was used with an appropriately positioned 3' primer to PCR amplify a deletion mutant of sHASEGP of the desired length from the carboxy terminal end. . For example PCR with primers described in SEQ ID No. 14 and SEQ ID No. 26 as the 5' and 3' primers respectively was used to generate the polypeptide in SEQ ID No. 49 when expressed from an expression construct in IresPuro2 vector. Similarly, PCR with reverse3' primers described in SEQ ID No 27,28,29,30,31 and 32 were used to generate deletion mutants ending at amino acid positions A 447, S 430, G 413, S 394, A 372, and S 347 respectively of the mature sHASEGP. The PCR products in each case were digested with NheI and BamHI enzymes and the digested product cloned into pIresPuro2 vector between NheI and BamHI sites. A few independent clones in the final expression construct from each group were tested for secreted neutral active sHASEGP activity by transient transfection in CHO cells in CD-CHO serum free media (Invitrogen, CA) and samples drawn at indicated time points for assay. Miniprep DNA prepared from overnight cultures was transfected with Genejuice (Novagen, CA)

transfection reagent following manufacturer recommended protocols. Hyaluronidase activity was measured by microtiter assay as described above.

[0562] b. Results

[0563] Hyaluronidase activity was measured in sHASEGP truncation mutants to identify the minimally active domain for secreted neutral active hyaluronidase activity.

AMINO ACID 1 TO :	U/ML/24HRS PH7.4
347	0.000
372	0.000
394	0.000
413	0.000
430	0.000
447	0.000
467	0.089
477	0.567
478	0.692
479	0.750
480	0.575
481	0.740
482	0.329
483	0.800
509	0.044

[0564] The results showed that all six one amino acid deletion mutants ending at indicated amino acids from Y 482 to E 477 gave higher secreted activity than GPI anchored sHASEGP.

[0565] The results also showed that deletions beyond A 467 eliminated any secreted activity. Secreted neutral activity from the A 467 clones decreased to approximately 10% of that found P478 or N 483 clones. It was therefore concluded that more of the carboxy

terminal domain of human sHASEGP was required to create the neutral active hyaluronidase domain than previously assumed from the bee venom enzyme. The cysteines in the carboxy terminal domain are thus necessary for neutral activity. A very narrow range spanning approximately 10 amino acids before the GPI cleavage site at N 483 thus defined the minimally active domain.

[0566] EXAMPLE-5 EFFECTS OF SIGNAL PEPTIDE MODIFICATION ON SHASEGP SECRETORY ACTIVITY.

[0567] Human sHASEGP possesses an unusually long predicted native leader peptide. Additionally, the existence of two adjacent cysteine residues in the leader peptide may lead to aggregation of polypeptide multimers within the endoplasmic reticulum during high level expression and therefore prevent high level expression of a sHASEGP. A series of more efficient secretory leader peptides were therefore tested to examine for their ability to enhance the targeting of sHASEGP for secretion.

[0568] a. Protocol

[0569] The Kappa leader peptide was constructed by overlapping primer annealing and extension PCR with primers corresponding to sequences in SEQ ID No 37, 38, 39 and 40. The resulting PCR amplified kappa sequence was amplified with flanking primers containing NheI site in the 5' end (as described in SEQ ID No. 41) and EcoR1 site at the 3' end (as described in SEQ ID No. 42). This allowed cloning the Kappa leader peptide (the polypeptide sequence is as described in SEQ ID No. 43) in the Litmus 39 (NEB) vector between NheI and EcoRI sites. sHASEGP has an internal EcoRI site; therefore this kappa construct between NheI site and EcoRI site was further amplified with a 5' SpeI primer (as described in SEQ ID No. 44) and a 3' MluI primer (as described in SEQ ID No. 45). sHASEGP without GPI anchor ending at P 478 was cut out from pIresPuro2 with NheI and BamHI and cloned into a Litmus 39(NEB) vector within the NheI and BamHI sites of theLitmus39 vector. This resulting sHASEGP-containing Litmus vector was digested with SpeI and MluI restriction enzymes and the kappa leader construct amplified with SpeI and MluI was cloned into it. Site directed mutagenesis was performed on this Litmus 39 vector containing both Kappa and sHASEGP sequences to generate the in

frame fusion of Kappa leader sequence to the mature polypeptide of sHASEGP. Primer pairs corresponding to SEQ ID No. 34 and 35 were used to generate the kappa leader with the native Asp as the terminal amino acid fused to the F 38 of sHASEGP (up to P 478) (as described in SEQ ID No. 46 for the polypeptide sequence of the fusion protein). Other primer pair combinations such as embodied by SEQ ID No. 33 with SEQ ID No. 35 were used to generate Kappa leader ending at the terminal Asp (D) fused to L 36 of SHASEGP, SEQ ID No. 33 with SEQ ID No. 36 were used to generate Kappa leader ending at the Gly (G) (before the terminal Asp (D)) fused to L 36 of SHASEGP, and SEQ ID No. 34 with SEQ ID No. 36 were used to generate Kappa ending at the Gly (G) (before the terminal Asp (D)) fused to F 38 of SHASEGP. The Kappa –sHASEGP fusions obtained by site directed mutagenesis were gel purified, digested with enzyme DpnI to digest any carryover parental DNA, and then digested with NheI and Bam HI and cloned in to the NheI/BamHI digested HisIresPuro2 backbone which has the his tag (six amino acid spacer followed by six histidines) cloned in between BamH1 and Not1 sites in pIRESPuro2 vector. Therefore upon ligation we obtain a construct that is NheI- kappa-SHASEGP-BamHI-His in pIresPuro2. Four sets of such construct were obtained that would correspond to the combinations of G or D at the Kappa leader end and L 36 or F 38 at the beginning of mature sHASEGP. A few independent clones from each type of construct were transfected into CHO cells in CD-CHO medium (Invitrogen, CA) to test whether the presence of kappa secretion leader would promote increased levels of secreted protein as compared to native secretion leader. Miniprep DNA prepared from overnight cultures were transfected with Genejuice (Novagen, CA) transfection reagent following manufacturer recommended protocols and samples were drawn for testing by microtiter assay at indicated time points. Hyaluronidase activity was measured by microtiter assay as described above.

[0570] Mouse IgG Kappa chain leader peptide sHASEGP fusion constructs were tested to test for higher levels of secreted neutral active sHASEGP activity.

[0571] b. Results

HUMAN sHASEGP GENE CONSTRUCT	U/ML/24HOURS PH 7.4
IgG Kappa Leader sHASEGP AA 38-478 HIS6	3.0257
Native Leader sHASEGP AA 1-478 HIS6	0.4857

[0572] The enzyme assay results indicated that the IgG Kappa leader was capable of enhancing secretion of sHASEGP approximately 7 to 8 fold higher than the native secretion leader when compared with clones P478, Y 482 or N 483 that lacked such a leader. Other kappa leader constructs with variations of the leader fusion site from the Asp or the Gly of the Kappa leader to L36 or F38 of sHASEGP yielded increased levels of secreted neutral active hyaluronidase activity as well. These examples are intended to expand rather than limit the scope of the invention, as other efficient secretory leader sequences may be utilized with the same technology.

[0573] EXAMPLE 6

[0574] GENERATION OF A HUMAN sHASEGP EXPRESSION VECTOR

[0575] A sHASEGP without an eptiope tag was generated by cloning into a bicistronic expression cassette, HZ24 (SEQ ID NO 47). The HZ24 plasmid vector for expression of sHASEGP comprises a pCI vector backbone (Promega), DNA sequence encoding amino acids 1-482 of human PH20 hyaluronidase, an internal ribosomal entry site (IRES) from the ECMV virus (Clontech), and the mouse dihydrofolate reductase (DHFR) gene. The pCI vector backbone also includes DNA encoding the Beta-lactamase resistance gene (AmpR), an f1 origin of replication, a Cytomegalovirus immediate-early enhancer/promoter region (CMV), a chimeric intron, and an SV40 late polyadenylation signal (SV40). The DNA encoding the sHASEGP construct contained a Kozak consensus sequence in the Methionine of the native signal leader and a stop codon at Tyrosine 482. The resultant construct pCI-PH20-IRES-DHFR-SV40pa (HZ-24) results in a single mRNA species driven by the CMV promoter that encodes amino acids 1-482 of PH20 and

amino acids 1-187 of the dihydrofolate reductase separated by the internal ribosomal entry site.

[0576] The human PH20 open reading frame was amplified from an Invitrogen ORF clone (IOH10647, Invitrogen, Carlsbad CA) with a 5'Primer that introduced an *Nhe*I site and Kozack consensus sequence before the Methionine of PH20 and a reverse primer that introduced a stop codon following Tyrosine 482 and introduced a *Bam*H1 restriction site. The resultant PCR product was ligated into the plasmid pIRESpuro2 (Clontech, Palo Alto, CA) following digest of the PH20 PCR fragment with *Nhe*I and *Bam*H1.

[0577] EXAMPLE-7

[0578] GENERATION OF A sHASEGP EXPRESSING CELL LINE

[0579] Non-transfected DG44 CHO cells growing in GIBCO Modified CD-CHO media for DHFR(-) cells, supplemented with 4mM Glutamine and 18 ml Plurionic F68/L (Gibco), were seeded at 0.5 x 10^6 cells/ml in a shaker flask in preparation for transfection. Cells were grown at 37°C in 5% CO_2 humidified incubator with 120 rpm for shaking. Exponentially growing non-transfected DG44 CHO cells were tested for viability prior to transfection.

[0580] 60,000,000 viable cells of the non-transfected DG44 CHO cell culture was pelleted and resuspended to a density of 20,000,000 cells in 0.7 mL of 2x transfection buffer (2X HeBS = 40 mM Hepes, pH 7.0, 274 mM NaCl, 10 mM KCl, 1.4 mM Na_2HPO_4, 12 mM dextrose). To each aliquot of resuspended cells, 0.09 mL of the linear HZ24 plasmid (250 ug) was added, and the cell/DNA solutions were transferred into 0.4 cm gap BTX (Gentronics) electroporation cuvettes at room temperature. A negative control electroporation was performed with no plasmid DNA mixed with the cells. The cell/plasmid mixes were electroporated with a capacitor discharge of 330 V and 960 uF or at 350 V and 960 uF.

[0581] The cells were removed from the cuvettes after electroporation and transferred into 5 mL of Modified CD-CHO media for DHFR(-) cells, supplemented with 4mM Glutamine

and 18 ml Plurionic F68/L (Gibco), and allowed to grow in a well of a 6-well tissue culture plate without selection for 2 days at 37°C in 5% CO_2 humidified incubator.

[0582] Two days post electroporation, 0.5 mL of tissue culture media was removed from each well and tested for presence of hyaluronidase activity.

[0583] Initial Hyaluronidase Activity of HZ24 Transfected DG44 CHO Cells At 40 Hours Post Transfection

	Dilution	Activity Units/ml
Transfection 1 330V	1 to 10	0.25
Transfection 2 350V	1 to 10	0.52
Negative Control	1 to 10	0.015

[0584] Cells from transfection 2 (350V), were collected from the tissue culture well, counted and diluted to 10,000 to 20,000 viable cells per mL. A 0.1 mL aliquot of the cell suspension was transferred to each well of five, 96 well round bottom tissue culture plates. 0.1 mL of CD-CHO media (GIBCO) containing 4 mM Glutamax-1, and without hypoxanthine and thymidine supplements were added to the wells containing cells (final volume 0.2 mL).

[0585] Ten clones were identified from the 5 plates grown without methotrexate.

Plate/Well ID	Relative Hyaluronidase Activity
1C3	261
2C2	261
3D3	261
3E5	243
3C6	174
2G8	103
1B9	304

2D9	273
4D10	302
1E11	242
A1 (+) control	333
H12 (-) control	0

[0586] Six HZ24 clones were expanded in culture and transferred into shaker flasks as single cell suspensions. Clones 3D3, 3E5, 2G8, 2D9, 1E11, and 4D10 were plated into 96-well round bottom tissue culture plates using a two-dimensional infinite dilution strategy. Diluted clones were grown in a background of 500 non-transfected DG44 CHO cells per well, to provide necessary growth factors for the initial days in culture. Ten plates were made per subclone.

[0587] Clone 3D3 produced 24 visual subclones. Significant hyaluronidase activity was measured in the supernatants from 8 of the 24 subclones (> 50 Units/mL), and these 8 subclones were expanded into T-25 tissue culture flasks in the presence of 50 nM methotrexate. Clone 3D3 50nM was further expanded in 500nM methotrexate giving rise to clones producing in excess of 1,000 Units/ml in shaker flasks (clone 3D3 5M).

[0588] EXAMPLE 8

[0589] PRODUCTION OF sHASEGP

[0590] A vial of 3D3 5M was thawed and expanded from T flasks through 1L spinner flasks in CHO CDM (Invitrogen, Carslbad CA) supplemented with 100nM Methotrexate and Glutamax (Invitrogen). Cells were transferred from spinner flasks to a 5L bioreactor (Braun) at an inoculation density of 4.0 x 10E5 viable cells per ml. Paramaters were temperature setpoint, 37C , pH 7.2 (starting Setpoint), with Dissolved Oxygen Setpoint 25% and an air overlay of 0-100 cc/min. At 168 hrs, 250ml of Feed #1 Medium (CD CHO + 50g/L Glucose) was added. At 216 hours, 250ml of Feed #2 Medium (CD CHO + 50g/L Glucose + 10mM Sodium Butyrate) was added, and at 264 hours 250ml of Feed #2 Medium was added. This process resulted in a final productivity of 1600 Units per ml with a maximal cell density of 6 million cells/ml. The addition of sodium butyrate was

found to dramatically enhance the production of sHASEGP in the final stages of production.

3D3-5M Growth & sHASEGP Production, 5L Bioreactor

Run Hours	Viable Cells x10E5	% Viable	Units/ml	Vol (mL)	[Glucose]	Feed
0	4.4	100	0	4500	547	
24	5.7	100	0	4500	536	
48	10.1	100	37	4500	501	
72	17.1	99	62	4500	421	
96	28.6	99	118	4500	325	
120	28.8	99	240	4500	274	
144	60.2	100	423	4500	161	
168	55	100	478	4500	92	250ml Feed #1
192	66.6	98	512	4750	370	
216	55.2	92	610	4750	573	250ml Feed#2
240	*53*	*88*	710	5000	*573*	
264	49.8	84	852	5000	474	250ml Feed #2
288	40	70	985	5250	770	
312	31	61	1467	5250	773	
336	25.4	52	1676	5250	690	

[0591] EXAMPLE 9

[0592] PURIFICATION OF sHASEGP

[0593] Conditioned media from the 3D3 clone was clarified by depth filtration and tangential flow diafiltration into 10mM Hepes pH 7.0. Soluble HASEGP was then purified by sequential chromatography on Q Sepharose (Pharmacia) ion exhange, Phenyl Sepharose (Pharmacia) hydrophobic interaction chromatography, phenyl boronate (Prometics) and Hydroxapatite Chromatography (Biorad, Richmond, CA).

[0594] SHASEGP bound to Q Sepharose and eluted at 400mM NaCl in the same buffer. The eluate was diluted with 2M Ammonium sulfate to a final concentration of 500mM ASO4 and passed through a Phenyl Sepharose (low sub) column, followed by binding under the same conditions to a phenyl boronate resin. The sHASEGP was eluted from the phenyl sepharose resin in Hepes pH 6.9 after washing at pH 9.0 in 50mM bicine without

AS04. The eluate was loaded onto a ceramic hydroxyapatite resin at pH 6.9 in 5mM PO4 1mM CaCl2 and eluted with 80mM PO4 pH 7.4 with 0.1mM CaCl2.

[0595] The resultant purified sHASEGP possessed a specific activity in excess of 65,000 USP Units/mg protein by way of the microturbidity assay using the USP reference standard. Purified sHASEGP eluted as a single peak from 24 to 26 minutes from a Pharmacia 5RPC styrene divinylbenzene column with a gradient between 0.1% TFA/H_2O and 0.1% TFA/90% acetonitrile/10% H_2O and resolved as a single broad 61 kDa band by SDS electrophoresis that reduced to a sharp 51kDa band upon treatment with PNGASE-F. N-terminal amino acid sequencing revealed that the leader peptide had been efficiently removed.

[0596] N-terminal Amino Acid Sequence biochemically purified sHASEGP.

Position	1	2	3	4	5	6	7	8	9	10	11
Theoretical	Leu	Asn	Phe	Arg	Ala	Pro	Pro	Val	Ile	Pro	Asn
Observed	-	Asn	Phe	Arg	Ala	Pro	Pro	Val	Ile	Pro	Asn

[0597] EXAMPLE 10

[0598] ANALYSIS OF DG44 CHO-DERIVED sHASEGP GLYCOSYLATION

[0599] Conflicting data exists as to whether sHASEGP's from different species require glycosylation for their catalytic activity. For example, it is reported that enzymatically active bee venom hyaluronidase can be synthesized in cells that lack glycosylation machinery, i.e. such as E. coli. Moreover, treatment of purified bovine testes hyaluronidase with PNGase did not inactivate enzyme activity (Yamagata et al 1997). Other studies report loss of activity following deglycosylation and that disulfide bonds are additionally required.

[0600] As all such previous tests were made using either crude or partially purified preparations however, it was not apparent whether the loss of activity was a result of exposure of deglycosylated enzyme to contaminating proteases in the crude preparations or a direct functional relationship between glycosylation and catalytic activity.

[0601] a. Protocol

[0602] To determine if functional N-linked glycosylation could be introduced into human sHASEGP using a CHO based expression system under protein free conditions, a cDNA encoding human sHASEGP-HIS6 was expressed in CHO cells using an IRESpuro bicistronic cassette in chemically defined media. Cells were grown for 72 hours in CHO CDM (Invitrogen/ Gibco) followed by concentration and tangential flow diafiltration on a Pellicon TFF unit (Millipore) with 30kDa cutoff membranes. The concentrate was exchanged with 10mM Hepes PH 7.4 50mM NaCl. The diafiltrate was then loaded on a DEAE streamline sepharose resin and eluted with a NaCl gradient from 0-1M NaCl on a Pharmacia FPLC resin. Human sHASEGP eluted between 10-30% NaCl. Levels of sHASEGP in column fractions determined that the majority of enzyme was recovered in the 10-30% NaCl gradient. The enzyme from the 10-30% NaCl gradient was then further purified through affinity chromatography on an IMAC resin charged with Ni. Human sHASEGP was eluted from the IMAC resin after washing with 10mM Imidizole with 50mM Acetate PH 5.0. The protein was concentrated and dialyzed against 10mM Hepes PH 7.4. The highly purified enzyme was determined to possess a specific activity of 97,000 Units/mg protein in the presence of 1mM Calcium and 1mg/ml HSA in the ELISA-based biotinylated substrate microtiter assay.

[0603] To detect changes in protein relative molecular mass, purified human sHASEGP was treated with PNGASE or Neuraminidase overnight followed by gel electrophoresis, electotransfer and western blot analysis with an HRP linked anti His6 monoclonal antibody (Qiagen) and ECL detection.

[0604] b. Results

[0605] Western blot analysis determined that the human sHASEGP produced in CHO cells was sensitive to PNGASE treatment. The relative molecule mass of human sHASEGP revealed that the protein was highly glycosylated. Upon complete overnight digestion with PNGASE, human sHASEGP reduced to a single species confirming that mild heterogeneity of the undigested band could be attributed to N-linked sugar residues. PNGaseF partial digestion showed a series of intermediates shifting from untreated and

progressive shift with longer treatment. Although bands were somewhat diffuse on a 7% Gel, at least 6 different intermediate isoforms could be visualized.

[0606] Treatment of sHASEGP with Neuraminidase revealed that CHO cells were in fact capable of synthesizing sialated human sHASEGP. Upon treatment with neuraminidase and Western Blot analysis of sHASEGP on 7% Gels, CHO derived Human recombinant sHASEGP revealed an approximately 1-3 kDa shift in mobility compared to untreated sHASEGP. This is thus the first report of the generation of a substantially sialated human sHASEGP. This is very valuable for both stability and to enhance serum half-life of a human sHASEGP as native sperm sHASEGP from many species lacks sialation and does not react with sialic acid specific lectins.

[0607] FACE analysis of sHASEGP

[0608] Analysis of active sHASEGP oligosaccharides by FACE analysis permits rapid determination of profiles of catalytically active sHASEGP's.

[0609] Protocol

[0610] Purified Hyaluronidase from the 3D3 5M clone was evaluated using FACE® N-Linked Oligosaccharide Profiling (Prozyme). Oligosaccharides were cleaved from 128.7 μg of glycoproteins by enzymatic digestion with N-Glycanase (a.k.a PNGase), labeled using the flourophore ANTS, and separated by electrophoresis. The relative positions of the oligosaccharide bands were determined by running the sample and dilutions of the sample alongside an oligosaccharide standard ladder which designated the migration distance in Degree of Polymerization (DP) units.

[0611] Results

[0612] The N-Profile for the Hyaluronidase sample consists of ten bands of which six (running concomitant with the oligosaccharide standard bands G5 – G12) had intensities greater than 9%. Furthermore, the band running alongside the G9 standard was the most intense with intensities of 35% – 46%.

[0613] sHAS

[0614] EGP oligosaccharide analysis

sHASEGP Oligosacharide	Degree of Polymerization	Percent of Total
1	15.64	1.2
2	13.68	3.4
3	11.61	10.0
4	10.04	10.4
5	8.37	35.4
6	7.32	9.7
7	6.14	9.0
8	5.57	12.4
9	3.84	2.3
10	3.26	0.5

[0615] EXAMPLE 11

[0616] DEPENDENCE OF SHASEGP N-LINKED GLYCOSYLATION FOR ENZYME ACTIVITY.

[0617] a. Protocol

[0618] Samples of purified HIS6 sHASEGP were mixed with buffer containing Neuraminidase and PNGASE with and without 50mm Octylglucoside overnight at 37C. Oligosaccharides were verified to have been removed by gel shift from Western Blot analysis.

[0619] b.

[0620] Results

SAMPLE	U/ML
No Rx	22.01
Neuraminidase O/N 50mM OG	23.57
PNGaseF w/ 50mM OG	0.0
PNGaseF without 50mM OG o/n	10.74

[0621] EXAMPLE-12

[0622] ACTIVITY OF SHASEGP TOWARDS SULFATED AND NON-SULFATED GLYCOSAMINOGLYCANS

[0623] In addition to the microtiter-based assay using HA, the substrate specificity of sHASEGP towards other glycosaminoglycans or proteoglycans can be tested using a gel shift assay with purified substrates to determine the activity of sHASEGP towards other glycosaminoglycans. Many Hyaluronidase assays have been based upon the measurement of the generation of new reducing N-acetylamino groups (Bonner and Cantey, *Clin. Chim. Acta* 13:746-752, 1966), or loss of viscosity (De Salegui et al., *Arch. Biochem. Biophys.* 121:548-554, 1967) or turbidity (Dorfman and Ott, *J. Biol. Chem.* 172:367, 1948). With purified substrates all of these methods suffice for determination of the presence or absence of endoglucosamidic activity.

[0624] a. Protocol

[0625] GEL SHIFT ASSAY- Purified substrates are mixed with recombinant sHASEGP to test for endoglucosidase activity that give rise to increased mobility in substrate within the gel. Chondroitin Sulfate A, Aggrecan and D were from Calbiochem. Hyaluronan (Human Umbilical Cord) Chondroitin Sulfate C, Dermatan sulfate, and Heparan-sulfate are obtained from Calbiochem. Human umbilical cord Hyaluronan was obtained from ICN. Each test substrate is diluted to 0.1mg/ml. 10ul samples of purified sHASEGP or conditioned media from sHASEGP expressing cells as well as are mixed with 90ul of test

substrate in desired buffer and incubated for 3 hours at 37C. Following incubation samples are neutralized with sample buffer (Tris EDTA PH 8.0, Bromophenol Blue and glycerol) followed by electrophoresis on 15% polyacrylamide gels. Glycosaminoglycans are detected by staining the gels in 0.5% Alcian Blue in 3% Glacial Acetic Acid overnight followed by destaining in 7% Glacial Acetic Acid. Degradation is determined by comparison substrate mobility in the presence and absence of enzyme.

[0626] b. Results

[0627] 100 Units of $sHASEGP_{HIS6}$ in 10ul was incubated with 90ul 10mM Hepes Buffer with 50ug/ml Human Serum Albumin for 2 hours at 37C containing 10ug of various glycosaminoglycans and proteoglycans. Electrophoretic analysis followed by Alcian blue staining revealed increased mobility shifts to a single species in Chondroitin Sulfate A, C and D, Aggrecan and Hyaluronan but not Heparan Sulfate nor Chondroitin Sulfate B. Whereas the undigested glycosaminoglycans ran as a smear in the middle of the gel, the digested products showed the majority of alcian blue stain running at the dye front with a small amount of material running as an incremental ladder.

[0628] EXAMPLE-13 EFECTS OF METAL IONS ON sHASEGP ACTIVATION

[0629] In addition to the requirement of glycosylation for optimal enzyme activity, human sHASEGP was found to be activated with cations for optimal enzyme activity. In the process of purification, sHASEGP was found to have a low specific acitivity following successive chromatography steps. The HIS6tagged sHASEGP was found to have a very low specific activity when purified to homogeneity from DEAE followed by successive Ni-IMAC purifications. As IMAC resins can chelate metal ions, various metals were added back to sHASEGP to determine the relative enzyme activity.

[0630] a. Protocol

[0631] Purified sHASEGP was tested following incubation with 0.1mM nickel (Ni), Cobalt (Co) Zinc (Zn) Calcium (Ca) and Magnesium (Mg) for 2 hours at room temperature followed by determination of hyaluronidase activity in microtiter based assay.

[0632] b. Results

Metal Salt Additive	Neutral Activity U/ml
NO ADDITIVES	11.909
100 uM Ni	6.0306
100 uM Co	8.972
100 uM Zn	3.7476
100 uM Ca	101.9892

[0633] A significant increase in hyaluronidase activity was found following incubation of sHASEGP with 0.1mM Calcium or 0.1mM Magnesium. No such activation was found following incubation with other metals. The addition of Calcium to sHASEGP increased the specific activity of the enzyme to approximately 97,000 units per mg protein based upon A280 measurement. A dose response curve of Calcium and Magnesium metals was then tested to determine the optimal concentration of metal ions to enzyme.

mM Divalent Metal	[Ca++]	[Mg++]
100	1	1.3
10	108	104
1	169	164
0.1	123	78
0.01	59	18
0.001	47	13
0.0001	39	13
0.00001	55	15

[0634] Activation of sHASEGP was found to occur in the micromolar range. Concentrations above 10mM were inhibitory for both Calcium and Magnesium. To rule out nonspecific activation of substrate rather than enzyme, Calcium Chloride in 10mM Hepes buffer was incubated with the immobilized biotinylated substrate on the microtiter

plate followed by washing. No activation was found when the enzyme was added to the Calcium preincubated plate that had been washed. The activation was also tested on phospholipase C released native sHASEGP which revealed a similar activation with Calcium ruling out an artifact of the carboxy terminus HIS6 epitope tag.

[0635] EXAMPLE 14

[0636] EFFECTS OF ALBUMIN ON THE ACTIVITY OF sHASEGP

[0637] It was found that the dilution of recombinant rHUPH20 and other preparations of slaughterhouse testes-derived hyaluronidases required albumin in addition to Calcium for optimal activity.

[0638] a. Protocol

[0639] Human Serum Albumin (ICN) was diluted into 10mM Hepes buffer with Calcium to determine the effects of albumin protein on enzyme activity. Enzyme assays with sHASEGP and commercial preparations were examined using both 1mM $CaCl_2$ and 1mg/ml Human Serum Albumin.

[0640] b. Results

[0641] Activation of hyaluronidase activity was found at high dilutions in the presence of albumin. It was not clear whether this activation was a result of preventing denaturation or if the albumin affected the availability of the substrate. A preferable formulation of human sHASEGP could therefore include Albumin and a metal salt consisting of either Calcium or Magnesium.

[0642] EXAMPLE-15 SPREADING ACTIVITY OF PURIFIED sHASEGP *IN VIVO*

[0643] a. Protocol

[0644] Purified sHASEGP in 10mM Hepes PH 7.4, 150mM NaCl 0.1% Pluronic was diluted to 0.5U/ul in pyrogen free water with 0.15M NaCl. A series of dilutions in 20ul final of Saline were made to give a total of 0.01, 0.05, 0.1 Units per injection. 20ul of Trypan Blue solution was added to a final volume of 40ul and injected subcutaneously

into the lateral skin on each side of balb $^{Nu/Nu}$ mice that had been previously anesthetized i.p. by ketamine/xylazine administration. Dye areas were measured in 2 dimensions with a microcaliper from t=0 to t=45 min. Area was represented as mm^2. As a control recombinant Human HYAL1 that lacks neutral activity but is secreted was included.

[0645] b. Results

TEST ARTICLE	DYE AREA @ 45MIN
A. Saline Control	51.5mm^2
B. sHASEGP 0.01U	76.8 mm^2
C. sHASEGP 0.05U	98.22mm^2
D. sHASEGP 0.10U	180.4mm^2
E. HYAL1 100U	67.48 mm^2

[0646] EXAMPLE-16 KINETICS OF sHASEGP DIFFUSION ACTIVITY

[0647] a. Protocol

[0648] Recombinant purified $sHASEGP_{His6}$ was separated into 2 aliquots. One was heated to 95C for 15minutes in a thermocycler with a heated lid. The other remained at room temperature. Thermal inactivation of enzyme activity was verified in the microtiter based enzyme assay. For kinetic analysis heat inactivated verses native material was tested. 4 Units of purified sHASEGP or equivalent heat inactivated material was injected subcutaneously with trypan blue dye. Areas were tested at various time points up to 15minutes.

[0649] b. Results

4 UNITS $t_{minute\ post\ injection}$	4 UNITS HEAT INACTIVATED $t_{minute\ post\ injection}$
t_0 = 52.38	t_0 = 50.58
t_3 = 116.51	t_3 =65.48
$t_{6.5}$= 181.93	$T_{6.5}$ =63.87
t_{10}= 216.96	T_{10} =65.80
t_{16}= 279.99	T_{16} =74.3

[0650] EXAMPLE-17 RESTORATION OF THE DERMAL BARRIER BROKEN DOWN BY sHASEGP

[0651] a. Protocol

[0652] To establish the regeneration time of the pores opened with sHASEGP following subcutaneous administration, 2Units of purified sHASEGP or saline control was injected into two opposing lateral sites subcutaneously in animals at t=0 followed by injection with trypan blue at the same site at 30min 60 min and 24 hours. Area of the dye diffusion at t=15 minutes post injection was recorded for each time point compared to the control.

[0653] b. Results

2 UNITS $T_{hour\ post\ injection\ sHASEGP}$	SALINE CONTROL $t_{hour\ post\ injection\ sHASEGP}$
$t_{0.5h}$= 183	$t_{0.5h}$= 54
t_{1hr}= 167	t_{1hr}= 50
t_{22hr}= 61	t_{22hr}= 48

[0654] The results demonstrate that the dermal barrier reconstitutes within 24 hours of administration of 2 Units of enzyme.

[0655] EXAMPLE-18 DETERMINATION OF THE SIZE OF CHANNELS OPENED BY sHASEGP

[0656] It was shown that human sHASEGP opened channels in the interstitial space sufficient to permit the diffusion of a small molecule, i.e. trypan blue dye. However, it was unknown what the upper limits were on the size of particles that could diffuse in the presence of sHASEGP.

[0657] a. Protocol

[0658] Florescent molecules of varying sizes were used to determine the size of the channels opened by human sHASEGP, Flouresceinated Dextrans of 4,400 and 2million Da Average Molecular Weight (Sigma) as well as flourescein labeled beads of defined diameters from 20nanometers to 500nanometers (Molecular Probes), were administered subcutaneously in a volume of 40ul with following injection of sHASEGP or saline control in the same sites. Area of the dye front was then measured in two dimensions at 15 minutes post injection.

[0659] b. Results

Diffusion Agent	Diffusion Test Particle Size	Area at 15min	Stand Dev
sHASEGP	4400 Da	**84.2**	25.7
Control	4400 Da	**38.0**	5.8
sHASEGP	2x10E6 Da	**141.2**	4.5
Control	2x10E6 Da	**51.7**	8.1
sHASEGP	20nm Diameter	**92.3**	20.6
Control	20nm Diameter	**51.6**	3.0
sHASEGP	100nm Diameter	**61.0**	5.7
Control	100nm Diameter	**40.0**	7.0
sHASEGP	200nm Diameter	**35.5**	1.6
Control	200nm Diameter	**27.9**	8.2
sHASEGP	500nm Diameter	**44.8**	13.6
Control	500nm Diameter	**41.2**	9.8

[0660] The results demonstrated that molecules from approximately 1kDa (Trypan Blue) to 50nm in diameter (Latex Beads) showed enhanced diffusion following administration of sHASEGP. While bovine serum albumin (66kDA) showed similar kinetics of diffusion to trypan blue, the 50nm latex beads required significantly more time to diffuse. 500nm beads showed no diffusion up to 480 minutes.

[0661] EXAMPLE-19 SERUM PHARMACOKINETICS PROFILES OF BIOTINYLATED ANTIBODIES FOLLOWING SUBCUTANEOUS CO-INJECTION OF HUMAN SHASEGP.

[0662] a. Protocol

[0663] Female Balb/c mice were anesthetized with a mixture of ketamine /xylazine. The mice were then injected subcutaneously with 20ul of 0.5mg/ml solution of biotinylated

mouse IgG mixed with 20ul of either saline or 20ul sHASEGP containing 4Units of activity.

[0664] b. Results

TIME POST INJECTION	CONTROL	sHASEGP (4U)
Serum IgG t=0 hrs	0 ng/ml	0 ng/ml
Serum IgG t=2hrs	0 ng/ml	360 ng/ml
Serum IgG t=51hrs	4152 ng/ml	4176 ng/ml

[0665] The results demonstrate that sHASEGP increases the kinetics of serum distribution of large molecules in circulation. Where no biotinylated IgG could be detected in the control group at 2 hours, 360ng/ml was apparent by 2 hours in the sHASEGP group.

[0666] EXAMPLE-20 SPREADING ACTIVITY OF SUBCUTANEOUSLY INJECTED MOLECULES FOLLOWING INTRAVENOUS INJECTION OF HUMAN sHASEGP

[0667] a. Protocol

[0668] Four sites for dye injection were utilized per dose of each Test Article and carrier control. Dye injection was 45 minutes after i.v. injection. Each dose of test or control article was injected i.v. into 2 animals. Measurement of the dye front area post 45 minute enzyme administration was calculated at 2.5, 5, 10 and 15 minutes for each dose or carrier control.

[0669] b. Results

[0670] Results demonstrated that highly purified sHASEGP was systemically available to distal tissues upon intravenous administration. The spreading activity of systemically administered sHASEGP was dose dependent, with a 10 unit injection being indistinguishable from carrier control.

Type	Dose IV	Time Minutes	Mean Area (mm^2	SD
PH20	1000	2.5	86.417	2.834193
PH20	1000	5	102.17	2.221146
PH20	1000	10	124.53	6.304944
PH20	1000	15	129.81	1.434319
PH20	300	2.5	59.137	7.218615
PH20	300	5	73.638	7.51197
PH20	300	10	87.092	8.686008
PH20	300	15	92.337	10.66466
PH20	100	2.5	56.308	7.741934
PH20	100	5	63.156	11.42052
PH20	100	10	76.519	16.18449
PH20	100	15	77.432	17.32264
PH20	30	2.5	50.534	10.64287
PH20	30	5	59.493	5.163971
PH20	30	10	68.102	11.00071
PH20	30	15	71.118	9.934212
PH20	10	2.5	36.4	3.807072
PH20	10	5	39.859	6.680932
PH20	10	10	45.649	4.44936
PH20	10	15	48.41	6.546835
Control	0	2.5	34.652	5.935037
Control	0	5	36.279	3.614544
Control	0	10	44.687	5.821216
Control	0	15	53.002	2.812439

CLAIMS

1. A substantially purified neutral active PH20 glycoprotein, wherein the glycoprotein consists of:

a) a polypeptide that consists of a sequence of amino acid residues that:

(i) is truncated at a C-terminal residue within 10 amino acids before or at amino acid 483 in the sequence of amino acids set forth in SEQ ID NO:1, or

(ii) consists of a sequence of amino acid residues having at least 98% sequence identity to the sequence of amino acids that consists of residues 36-483 of SEQ ID NO:1; and

b) at least one N-linked sugar moiety, wherein the N-linked sugar moiety is covalently attached to an asparagine residue of the polypeptide.

2. The glycoprotein of claim 1, wherein the polypeptide is selected from:

(a) a polypeptide that consists of a sequence of amino acids having at least 98% amino acid sequence identity with the sequence of amino acids that consists of residues 36-483 as set forth in SEQ ID NO. 1; and

(b) a polypeptide that consists of a sequence of amino acids encoded by the sequence of nucleotides 1-1449 as set forth in SEQ ID NO. 6.

3. The glycoprotein of claim 1, wherein the polypeptide consists of a sequence of amino acid residues having at least 98% sequence identity to the sequence of amino acids that consists of residues 36-483 of SEQ ID NO:1.

4. The glycoprotein of claim 1, wherein said sugar moiety is covalently attached to an asparagine residue selected from amino acids 82, 166, 235, 254, 368, or 393 as set forth in SEQ ID NO. 1.

5. A substantially purified neutral active PH20 glycoprotein, consisting of:

a) a polypeptide consisting of the sequence of amino acid residues set forth as 1-459 in SEQ ID NO:3; and

b) at least one N-linked sugar moiety, wherein the N-linked sugar moiety is covalently attached to an asparagine residue of the polypeptide, wherein

the glycoprotein is catalytically active; and

the glycoprotein is secreted.

6. The glycoprotein of claim 1, wherein said sugar moiety is covalently linked to said polypeptide through a PNGase sensitive bond.

7. The glycoprotein of claim 1, wherein said sugar moiety is a high mannose type.

8. The glycoprotein of claim 1, wherein said sugar moiety is a complex type.

9. The glycoprotein of claim 1, wherein said sugar moiety is a hybrid type.

10. The glycoprotein of claim 1, wherein said sugar moiety further comprises at least one terminated with sialic acid.

11. The glycoprotein of claim 10, wherein the polypeptide is super-sialated.

12. The glycoprotein of claim 1, wherein the polypeptide is truncated at an amino acid residue selected from among 477, 478, 479, 480, 481, 482 and 483.

13. A substantially purified neutral active PH20 glycoprotein that consists of:

a) a polypeptide that consists of:

(i) a sequence of amino acids residues set forth as 36-477, 36-478, 36-479, 36-480, 36-481, 36-482 or 36-483 of SEQ ID NO:1; or

(ii) a sequence of amino acids that contains amino acid substitutions in the sequence of amino acids set forth as 36-477, 36-478, 36-479, 36-480, 36-481, 36-482 or 36-483 of SEQ ID NO:1, whereby the amino-substituted polypeptide consists of a sequence of amino acids that has at least 91% amino acid sequence identity to a sequence of amino acids set forth as amino acid residues 1-477, 1-478, 1-479, 1-480, 1-481, 1-482 or 1-483 of SEQ ID NO:1; and

b) at least one N-linked sugar moiety, wherein the N-linked sugar moiety is covalently attached to an asparagine residue of the polypeptide.

14. The glycoprotein of claim 1 that consists of a sequence of amino acids set forth as amino acids 36-482 of SEQ ID NO:1.

15. The glycoprotein of claim 1, wherein the glycoprotein is a human glycoprotein.

16. The glycoprotein of claim 1, wherein the polypeptide is a polypeptide as set forth in SEQ ID NO. 4.

17. The glycoprotein of claim 1, with the proviso that the polypeptide does not comprise the complete sequence set forth in SEQ ID NO. 1 and includes at least amino acids 1-429 of SEQ ID NO. 4.

18. The glycoprotein of claim 1, wherein the polypeptide is encoded by a nucleic acid molecule that encodes amino acids 36-482 of SEQ ID NO:1 or amino acids 1-482 of SEQ ID NO:1.

19. The glycoprotein of claim 18, wherein the polypeptide is secreted in CHO cells.

20. The glycoprotein of claim 1, wherein the potency of the glycoprotein is greater than 40,000 USP Units/mg protein.

21. The glycoprotein of claim 20, wherein the glycoprotein is sialated.

22. The glycoprotein of claim 20, wherein the potency is greater than about 45,000, 55,000, 60,000, 80,000, 90,000, 95,000 or 100,000 USP Units/mg.

23. The glycoprotein of claim 20, wherein the potency is about 60,000 to 80,000 USP Units/mg.

24. The glycoprotein of claims 1 or 20, wherein the glycoprotein is modified with a polymer.

25. The glycoprotein of claim 24, wherein the polymer is PEG or dextran.

26. A composition comprising the glycoprotein of claim 20.

27. A nucleic acid molecule comprising a sequence of nucleotides that encodes a glycoprotein of claim 1 or claim 13, wherein the sequence of nucleotides includes a stop codon, whereby upon expression and secretion in a cell a polypeptide of claim 1 or 13 is produced.

28. The nucleic acid molecule of claim 27 that has a sequence of nucleotides set forth in SEQ ID NO:48 or that has a sequence of nucleotides set forth as nucleotides 106-1446 of SEQ ID NO:6.

29. A vector having a sequence as set forth in SEQ ID NO:51.

30. A vector comprising the nucleic acid molecule of claim 27 or claim 28.

31. The vector of claim 30 that is an expression vector.

32. The vector of claim 30 that is a eukaryotic vector.

33. The vector of claim 30 that includes a sequence of nucleotides that directs secretion of any polypeptide encoded by a sequence of nucleotides operatively linked thereto.

34. The vector of claim 30 that is a Pichia vector, an E. coli vector or a viral vector.

35. An isolated cell, comprising the vector of claim 30.

36. The cell of claim 35 that is a prokaryotic cell or a eukaryotic cell.

37. The cell of claim 35 selected from a bacterial cell, a yeast cell, a plant cell, an insect cell or an animal cell.

38. The cell of claim 35 that is a mammalian cell.

39. A method for producing a glycoprotein of claim 1, comprising:

introducing a nucleic acid that encodes the polypeptide of claim 1 operably linked to a suitable promoter into a cell capable of incorporating N-linked sugar moieties into the polypeptide;

culturing the cell under conditions whereby the encoded polypeptide is expressed by the cell and secreted into the cell culture medium; and

recovering the secreted polypeptide.

40. The method of claim 39, wherein the cell is a eukaryotic cell.

41. The method of claim 40, wherein the eukaryotic cell is selected from a mammalian cell, an insect cell, a yeast cell or a plant cell.

42. The method of claim 41, wherein the cell is a CHO cell.

43. A method for producing a glycoprotein, comprising:

culturing the cell of claim 35 under conditions whereby the encoded polypeptide is expressed by the cell; and

recovering the expressed polypeptide.

44. The use of the purified glycoprotein of claim 1 in the manufacture of a medicament.

45. A composition, comprising a substantially purified glycoprotein of claim 1 and a suitable pharmaceutical carrier.

46. The use of the glycoprotein of claim 1 in the manufacture of a medicament for treating an excess of glycosaminoglycan substrate in a subject, wherein the medicament removes said glycosaminoglycan substrate.

47. The use of claim 46, wherein said excess substrate is produced from a scar tissue.

48. The use of claim 47, wherein said scar tissue is a glial scar resulting from spinal cord injury.

49. The use of claim 47, wherein said scar tissue is a result of surgery.

50. The use of claim 47, wherein said scar tissue is a keloid scar.

51. The use of claim 50, wherein said substrate is associated with a herniated disk.

52. The use of the glycoprotein of claim 1 for the manufacture of a medicament, for increasing the diffusion of a therapeutic substance smaller than about 0.5 microns in diameter in a subject, wherein the medicament opens or forms channels smaller than about 0.5 microns in diameter in said subject.

53. The use of claim 52, wherein said glycoprotein is injected at a dose between 0.1 and 1500 USP Units.

54. The use of claim 52, wherein said glycoprotein polypeptide is mixed with the therapeutic substance prior to injection.

55. A kit, comprising:

(a) the glycoprotein of claim 1 at a dose between 0.1 and 1500 USP Units/ml in an acceptable carrier;

(b) at least one therapeutic substance in an acceptable carrier; and

(c) optionally, instructions for delivering therapeutic substances.

56. The kit of claim 55, wherein the glycoprotein and the therapeutic substance are provided as a mixture.

57. The use of claim 44, wherein the medicament removes excess glycosaminoglycans in a cardiovascular disorder.

58. The use of claim 57, wherein said cardiovascular disorder is selected from a myocardial infarct, congestive heart failure or arteriosclerosis.

59. The use of claim 44, wherein the medicament removes excess glycosaminoglycans in a tumor.

60. The use of the glycoprotein of claim 1 for the manufacture of a medicament for increasing delivery of a molecule to a tissue containing excess amounts of glycosaminoglycan, wherein the medicament degrades glycosaminoglycans sufficiently to open channels less than about 500 nm in diameter.

61. The use of claim 60, wherein the polypeptide is modified with a polymer.

62. The use of claim 61, wherein the polymer is PEG or dextran.

63. A pharmaceutical composition, comprising the substantially purified glycoprotein of claim 1 and a pharmaceutical carrier.

64. A pharmaceutical composition, comprising the substantially purified glycoprotein of claim 1.

65. The pharmaceutical composition of claim 64, wherein the glycoprotein is formulated in the composition with a pharmaceutically active agent.

66. The pharmaceutical composition of claim 65, wherein the pharmaceutically active agent is selected from the group of agents consisting of a chemotherapeutic agent, an analgesic agent, an anti-inflammatory agent, an antimicrobial agent, an amoebicidal agent, a trichomonacidal agent, an anti-parkinson agent, an anti-malarial agent, an anticonvulsant agent, an anti-depressant agent, an antiarthritic agent, an anti-fungal agent, an antihypertensive agent, an antipyretic agent, an anti-parasite agent, an antihistamine agent, an alpha-adrenergic agonist agent, an alpha blocker agent, an anesthetic agent, a bronchial dilator agent, a biocide agent, a bactericide agent, a bacteriostat agent, a beta-adrenergic blocker agent, a calcium channel blocker agent, a cardiovascular drug agent, a contraceptive agent, a decongestant agent, a diuretic agent, a depressant agent, a diagnostic agent, an electrolyte agent, a hypnotic agent, a hormone agent, a hyperglycemic agent, a muscle relaxant agent, a muscle contractant agent, an ophthalmic agent, a

parasympathomimetic agent, a psychic energizer agent, a sedative agent, a sympathomimetic agent, a tranquilizer agent, an urinary agent, a vaginal agent, a viricide agent, a vitamin agent, a non-steroidal anti-inflammatory agent, an angiotensin converting enzyme inhibitor agent, a polypeptide, a protein, a nucleic acid, a drug, an organic molecule, a sleep inducer, an insulin, a cytokine, an antibody and a monoclonal antibody.

67. The pharmaceutical composition of claim 66, wherein the chemotherapeutic agent is a toxin or a tumor necrosis factor.

68. The pharmaceutical composition of claim 66, wherein the anesthetic agent is lidocaine or bupivacaine.

69. The pharmaceutical composition of claim 66, further comprising a hormonal agent.

70. The pharmaceutical composition of claim 69, wherein the hormonal agent is epinephrine.

71. The pharmaceutical composition of claim 63, wherein the pharmaceutical carrier comprises a stabilizing solution.

72. The pharmaceutical composition of claim 71, wherein the stabilizing solution comprises, NaCl and a metal selected from $CaCl_2$ and $MgCl_2$.

73. The pharmaceutical composition of claim 71, wherein the stabilizing solution further comprises ethylenediamine tetra-acetic acid (EDTA).

74. The pharmaceutical composition of claim 71, wherein the stabilizing solution further comprises a carrier selected from the group comprising albumin, detergent or a surfactant.

75. The pharmaceutical composition of claim 71, wherein the stabilizing solution comprises phenol red, human serum albumin, HEPES, NaCl and a metal selected from $CaCl_2$ and $MgCl_2$.

76. The pharmaceutical composition of claim 75, wherein the concentration of the glycoprotein is about 80 USP Units/ml.

77. The pharmaceutical composition of claim 63, further comprising a surfactant.

78. The pharmaceutical composition of claim 77, wherein the surfactant is selected from sorbitan monolaurate; sorbitan monooleate; sorbitan palmitate; sorbitan monostearate; sorbitan_sesquioleate; sorbitan trioleate; polyoxyethylene oleic acid ester derivatives; polyoxyethylene lauryl amine derivatives; polyoxyethylene stearyl amine derivatives; polyoxyethylene oleyl amine derivatives; polyoxyethylene castor oil derivatives; polyoxyethylene hydrogenated castor oil derivatives; polyoxyethylene bis phenol ether derivatives; polyoxyethylene glycols; sorbitan fatty acid ester derivatives;

polyoxyethylene sorbitan fatty acid ester derivatives; polyoxyethylene-polyoxypropylene derivatives; polyethylene glycol hexadecyl ether (Brij® 56); polyethylene glycol octadecyl ether (Brij® 72); polyoxyethylene 10 oleyl ether (Brij® 97); t-Octylphenoxypolyethoxyethanol (TRITON® X 100); polyethylene glycol sorbitan monolaurate (TWEEN® 20); polyoxyethylene sorbitan monopalmitate (TWEEN® 40); polyethylene glycol sorbitan monostearate (TWEEN® 60); polyoxyethylene sorbitan Tristearate (TWEEN® 65); polyethylene glycol sorbitan monooleate (TWEEN® 80); polyoxyethylene sorbitan_Trioleate (TWEEN® 85); tris(hydroxymethyl) aminomethane lauryl sulfate (TRIZMA® dodecyl sulfate); block copolymer of polyethylene and polypropylene glycol (Pluronic® F68); or albumin.

79. The pharmaceutical composition of claim 63, wherein the concentration of the glycoprotein is between about 1 Unit/mg and 5000 USP Units/mg.

80. The pharmaceutical composition of claim 79, wherein the concentration of the glycoprotein is between about 1 Unit/mg and 300 USP Units/mg.

81. The pharmaceutical composition of claim 79, wherein the concentration of the glycoprotein is between about 1 Unit/mg and 150 USP Units/mg.

82. The pharmaceutical composition of claim 63, further comprising a metal ion selected from calcium and magnesium.

83. The pharmaceutical compositions of claim 63 that is formulated as a timed release or that is released upon contact with the site of application or targeted tissue.

84. The pharmaceutical composition of claim 63 that is formulated for topical, local, enteric, parenteral, intracystal, intracutaneous, intravitreal, subcutaneous, intramuscular, or intravenous administration.

85. The pharmaceutical composition of claim 63 that is formulated as spray, foam or aerosol, tablet or capsule.

86. The pharmaceutical composition of claim 85, that is a tablet or capsule and comprises an enteric coating.

87. The pharmaceutical composition of claim 86, wherein the coating is selected from cellulose acetate phthalate, polyethylene glycol, polyoxyethylene sorbitan, castor oil, ethyl cellulose pseudolatex, phenyl salicylate, n-butyl stearate, stearic acid and carnauba wax.

88. The pharmaceutical composition of claim 63 that is a lyophilized powder.

89. The pharmaceutical composition of claim 88, wherein the lyophilized powder is produced by a process, comprising:

(a) transferring the pharmaceutical composition into a buffer solution;

(b) sterile-filtering the resulting solution; and

(c) lyophilizing the filtered solution under standard conditions to produce a sterile powder.

90. The pharmaceutical composition of claim 89, wherein the buffer solution is selected from HEPES, Phosphate, Citrate and Bicarbonate.

91. The pharmaceutical composition of claim 90, wherein the buffer solution further comprises a sugar or carbohydrate.

92. The pharmaceutical composition of claim 91, wherein the sugar or carbohydrate is dextrose or lactose.

93. The pharmaceutical composition of claim 92, wherein the lactose is present at a concentration of between about 1 mg/ml to about 15 mg/ml.

94. The pharmaceutical composition of claim 89, wherein the buffer further comprises a surfactant.

95. The pharmaceutical composition of claim 94, wherein the surfactant is selected from sorbitan monolaurate; sorbitan monooleate; sorbitan palmitate; sorbitan monostearate; sorbitan_sesquioleate; sorbitan trioleate; polyoxyethylene oleic acid ester derivatives; polyoxyethylene lauryl amine derivatives; polyoxyethylene stearyl amine

derivatives; polyoxyethylene oleyl amine derivatives; polyoxyethylene castor oil derivatives; polyoxyethylene hydrogenated castor oil derivatives; polyoxyethylene bis phenol ether derivatives; polyoxyethylene glycols; sorbitan fatty acid ester derivatives; polyoxyethylene sorbitan fatty acid ester derivatives; polyoxyethylene-polyoxypropylene derivatives; polyethylene glycol hexadecyl ether (Brij® 56); polyethylene glycol octadecyl ether (Brij® 72); polyoxyethylene 10 oleyl ether (Brij® 97); t-Octylphenoxypolyethoxyethanol (TRITON® X 100); polyethylene glycol sorbitan monolaurate (TWEEN® 20); polyoxyethylene sorbitan_monopalmitate (TWEEN® 40); polyethylene glycol sorbitan monostearate (TWEEN® 60); polyoxyethylene sorbitan Tristearate (TWEEN® 65); polyethylene glycol sorbitan monooleate (TWEEN® 80); polyoxyethylene sorbitan_Trioleate (TWEEN® 85); tris(hydroxymethyl) aminomethane lauryl sulfate (TRIZMA® dodecyl sulfate); block copolymer of polyethylene and polypropylene glycol (Pluronic® F68); or albumin.

96. A combination comprising,

a sterile vial and the pharmaceutical composition of claim 63 wherein the composition is contained in the vial.

97. The combination of claim 96, wherein the sterile vial contains an amount of the pharmaceutical composition that is for single dose administration.

98. A combination comprising, a sterile vial containing the pharmaceutical composition of claim 84.

99. The combination of claim 96, wherein the sterile vial further contains an amount of sterile water for injection; and the final concentration of the glycoprotein is between about 1 and 5000 USP Units/ml.

100. A combination comprising, a sterile syringe containing the pharmaceutical composition of claim 63.

101. The combination of claim 100, wherein the sterile syringe has a volume of between about 5 to 50 μl.

102. The combination of claim 100, wherein the final concentration of the glycoprotein is between about 1 and about 5000 USP Units/ml.

103. A combination comprising, a sterile syringe containing the pharmaceutical composition of claim 84.

104. The combination of claim 103, wherein the final concentration of the glycoprotein is between about 1 and about 5000 USP Units/ml.

105. The combination of claim 103, wherein the pharmaceutical composition is dissolved in about 5 μl to about 50 μl of sterile water.

106. The combination of claim 103, further comprising a second syringe containing a pharmaceutically effective agent.

107. The combination of claim 106, wherein the pharmaceutically effective agent is selected from a chemotherapeutic agent, an analgesic agent, an anti-inflammatory agent, an antimicrobial agent, an amoebicidal agent, a trichomonacidal agent, an anti-parkinson agent, an anti-malarial agent, an anticonvulsant agent, an anti-depressant agent, and antiarthritic agent, an anti-fungal agent, an antihypertensive agent, an antipyretic agent, an anti-parasite agent, an antihistamine agent, an alpha-adrenergic agonist agent, an alpha blocker agent, an anesthetic agent, a bronchial dilator agent, a biocide agent, a bactericide agent, a bacteriostat agent, a beta-adrenergic blocker agent, a calcium channel blocker agent, a cardiovascular drug agent, a contraceptive agent, a decongestant agent, a diuretic agent, a depressant agent, a diagnostic agent, an electrolyte agent, a hypnotic agent, a hormone agent, a hyperglycemic agent, a muscle relaxant agent, a muscle contractant agent, an ophthalmic agent, a parasympathomimetic agent, a psychic energizer agent, a sedative agent, a sympathomimetic agent, a tranquilizer agent, an urinary agent, a vaginal agent, a viricide agent, a vitamin agent, a non-steroidal anti-inflammatory agent, an angiotensin converting enzyme inhibitor agent, a polypeptide, a protein, a nucleic acid, a drug, an organic molecule, a sleep inducer, an insulin, a cytokine, an antibody and a monoclonal antibody.

108. The combination of claim 106, wherein the pharmaceutically effective agent is a viscoelastic.

109. A kit comprising the combination of claim 103 and one or more of the following:

(a) packaging material; and

(b) instructions for using the kit for the use of the pharmaceutical composition.

110. The kit of claim 109, wherein the packaging material includes ice, dry ice, styrofoam, foam, plastic, cellophane, shrink wrap, bubble wrap, paper, cardboard, starch peanuts, twist ties, metal clips, metal cans, drierite, glass and rubber.

111. The use of claim 44 in the treatment of a pathologic accumulation of glycosaminoglycans, wherein said glycoprotein is free of bovine, ovine or bacterial enzymes, and ameliorates or reduces the accumulated glycosaminoglycan.

112. The use of claim 111, wherein the accumulation is selected from cardiovascular, cerebral, paraphimosis, myxedema, scleromyxedema and lymphedema.

113. The use of claim 44 in facilitating revascularization of necrotic tissue, wherein the glycoprotein induces the growth of neovasculature into said necrotic tissue.

114. The use of the glycoprotein of claim 1 for removal of glycosaminoglycans from the vitreous humor of a mammalian eye.

115. The glycoprotein of claim 1 wherein said N-linked glycan demonstrates a degree of polymerization of about 15.6, 13.7, 11.6, 10.0, 8.37, 7.32, 6.1, 5.6, 3.8 or 3.2.

116. The method of claim 39, wherein the cell is cultured in the presence of 0.1-1 mM Sodium Butyrate under conditions suitable for production of the polypeptide.

117. The method of claim 39, wherein the method of recovering the expressed polypeptide, comprises:

contacting the cell culture medium containing the expressed polypeptide under low ionic strength with an anion exchange resin at neutral pH:

eluting the polypeptide with about 400 mM of a salt;

contacting the polypeptide with hydrophobic interaction chromatography resin in the presence of about 0.5 M ammonium sulfate;

contacting the polypeptide in ammonium sulfate with phenyl boronate resin;

eluting the polypeptide in low salt at neutral pH;

contacting the polypeptide with Hydroxyapatite resin; and

eluting the polypeptide with about 100 mM Na Phosphate, thereby resulting in a polypeptide that is substantially purified.

118. The use of claim 44 for the manufacture of a medicament for treating a disorder of a mammalian eye by inducing liquefaction of the vitreous humor of the mammalian eye.

119. The use of claim 118, wherein the polypeptide is modified with a polymer.

120. The use of claim 119, wherein the polymer is PEG or dextran.

121. The use of a glycoprotein of claim 1 in the manufacture of a medicament, for reducing intraocular pressure in the eye of a subject, wherein the glycoprotein is substantially free of bovine, ovine or bacterial enzymes; and is greater than about 40,000 USP Units/mg protein.

122. The use of claim 121, wherein the glycoprotein is further characterized by being free of IgG.

123. The use of the glycoprotein of claim 1 in the manufacture of a medicament for treating cardiovascular disorder including myocardial infarct, congestive heart failure and arteriosclerosis, glycosaminoglycan accumulation, vitreous hemorrhage, herniated disc, cancer, intraocular pressure and myxedema, wherein the glycoprotein is present in an amount sufficient to remove excess glycosaminoglycans.