BINDING PROTEINS SPECIFIC FOR INSULIN-LIKE GROWTH FACTORS AND USES THEREOF

CROSS-REFERENCE TO RELATED APPLICATIONS

[0001] This application claims priority under 35 U.S.C. § 119 to U.S. Provisional Application Serial No. 60/750,085, filed December 13, 2005; U.S. Provisional Application Serial No. 60/750,772, filed December 14, 2005; U.S. Provisional Application Serial No. 60/774,747, filed February 17, 2005; and U.S. Provisional Application Serial No. 60/808,183, filed May 24, 2006, each of which is incorporated herein by reference in its entirety.

BACKGROUND OF THE INVENTION

Field of the Invention

[0002] The invention relates to binding proteins that bind to insulin-like growth factor-2 (IGF-II) with cross-reactivity to insulin-like growth factor-1 (IGF-I) and uses of such binding proteins. More specifically, the invention relates to monoclonal antibodies directed to IGF-II with cross-reactivity to IGF-I and uses of these antibodies. Aspects of the invention also relate to hybridomas or other cell lines expressing such antibodies.

Description of the Related Art

[0003] Insulin-like growth factor IGF-I and IGF-II are small polypeptides involved in regulating cell proliferation, survival, differentiation and transformation. IGFs exert their various actions by primarily interacting with a specific cell surface receptor, the IGF-I receptor (IGF-IR) and activating various intracellular signaling cascades. IGFs circulate in serum mostly bound to IGF-binding proteins (IGFBP-1 to 6). The interaction of IGFs with the IGF-IR is regulated by the IGFBPs, and IGFs can only bind to the IGF-IR once released from the IGFBPs (mostly by proteolysis of the IGFBPs). IGF-I can also bind to a hybrid receptor comprised of IGF-IR and insulin receptor (IR) subunits. IGF-II has been shown to bind to the "A" isoform of the insulin receptor.

[0004] Malignant transformation involves the imbalance of diverse processes such as cell growth, differentiation, apoptosis, and transformation. IGF-I and IGF-II have

been implicated in the pathophysiology of a wide range of conditions, and are thought to play a role in tumorigenesis due to the mitogenic and antiapoptotic properties mediated by the receptor IGF-IR. LeRoith and Roberts, *Cancer Lett.* **195**:127-137 (2003).

[0005] IGF-I was discovered as a growth factor produced by the liver under the regulatory control of pituitary growth hormone and was originally designated somatomedin-C. Salmon *et al., J. Lab. Clin. Med.* **49**:825-826 (1957). Both IGF-I and IGF-II are expressed ubiquitously and act as endocrine, paracrine, and autocrine growth factors, through their interaction with the IGF-IR, a trans-membrane tyrosine kinase that is structurally and functionally related to the insulin receptor (IR). IGF-I functions primarily by activating the IGF-IR, whereas IGF-II can act through either the IGF-IR or through the IR-A isoform. LeRoith and Roberts, Cancer Lett. **195**:127-137 (2003). Additionally, the interaction of both IGF-I and IGF-II with the IGF-binding proteins may affect the half-life and bioavailability of the IGFs, as well as their direct interaction with receptors in some cases. Rajaram *et al., Endocr. Rev.* **18**:801-831 (1997).

[0006] IGF-I has a long-term impact on cell proliferation, differentiation, and apoptosis. Experiments in cultured osteosarcoma and breast cancer cells suggested that IGF-I is a potent mitogen and exerts its mitogenic action by increasing DNA synthesis and by stimulating the expression of cyclin D1, which accelerates progression of the cell cycle from G_1 to S phase. Furlanetto *et al., Mol. Endocrinol.* **8**:510-517 (1994); Dufourny *et al., J. Biol. Chem.* **272**:311663-31171 (1997). Suppression of cyclin D1 expression in pancreatic cancer cells abolished the mitogenic effect of IGF-I. Kornmann *et al., J. Clin. Invest.* **101**:344-352 (1998). In addition to stimulating cell cycle progression, IGF-I also inhibits apoptosis. IGF-I was shown to stimulate the expression of Bcl proteins and to suppress expression of Bax, which results in an increase in the relative amount of the Bcl/Bax heterodimer, thereby blocking initiation of the apoptotic pathway. Minshall *et al., J. Immunol.* **159**:1225-1232 (1997); Parrizas *et al., Endocrinology* **138**:1355-1358 (1997); Wang *et al., Endocrinology* **139**:1354-1360 (1998).

[0007] Like IGF-I, IGF-II also has mitogenic and antiapoptotic actions and regulates cell proliferation and differentiation. Compared with IGF-I, high concentrations of IGF-II circulate in serum. High serum IGF-II concentrations have been found in patients with colorectal cancer, with a trend towards higher concentrations in advanced disease. Renehan *et al., Br. J. Cancer* **83**:1344-1350. Additionally, most primary tumors and transformed cell lines overexpress IGF-II mRNA and protein. Werner and LeRoith *Adv. Cancer Res.* **68**:183-223 (1996). Overexpression of IGF-II in colon cancer is

associated with an aggressive phenotype, and the loss of imprinting (loss of allele-specific expression) of the IGF-II gene may be important in colorectal carcinogenesis. Michell *et al.*, *Br. J. Cancer* **76**:60-66 (1997); Takano et al., *Oncology* **59**:210-216 (2000). Cancer cells with a strong tendency to metastasize have four-fold higher levels of IGF-II expression than those cells with a low ability to metastasize. Guerra *et al.*, *Int. J. Cancer* **65**:812-820 (1996).

[0008] Research and clinical studies have highlighted the role of the IGF family members in the development, maintenance and progression of cancer. Many cancer cells have been shown to overexpress the IGF-IR and/or the IGF ligands. For example, IGF-I and IGF-II are strong mitogens for a wide variety of cancer cell lines, including sarcoma, leukemia, and cancers of the prostate, breast, lung, colon, stomach, esophagus, liver, pancreas, kidney, thyroid, brain, ovary, and uterus. Macaulay *et al.*, *Br. J. Cancer* **65**:311-320 (1992); Oku *et al.*, *Anticancer Res.* **11**:1591-1595 (1991); LeRoith *et al.*, *Ann. Intern. Med.* 122:54-59 (1995); Yaginuma *et al.*, *Oncology* **54**:502-507 (1997); Singh *et al.*, *Endocrinology* **137**:1764-1774 (1996); Frostad *et al.*, *Eur. J. Haematol* **62**:191-198 (1999). When IGF-I was administered to malignant colon cancer cells, they became resistant to cytokine-induced apoptosis. Remacle-Bonnet *et al.*, *Cancer Res.* **60**:2007-2017 (2000).

[0009] The role of IGFs in cancer is also supported by epidemiologic studies, which showed that high levels of circulating IGF-I and low levels of IGFBP-3 are associated with an increased risk for development of several common cancers (prostate, breast, colorectal and lung). Mantzoros *et al.*, *Br. J. Cancer* **76**:1115-1118 (1997); Hankinson *et al.*, *Lancet* **351**:1393-1396 (1998); Ma *et al.*, *J. Natl.Cancer Inst.* **91**:620-625 (1999); Karasik *et al.*, *J. Clin. Endocrinol Metab.* **78**:271-276 (1994). These results suggest that IGF-I and IGF-II act as powerful mitogenic and anti-apoptotic signals, and that their overexpression correlates with poor prognosis in patients with several types of cancer.

[0010] Using knockout mouse models, several studies have further established the role of IGFs in tumor growth. With the development of the technology for tissue specific, conditional gene deletion, a mouse model of liver IGF-I deficiency (LID) was developed. Liver-specific deletion of the *igf1* gene abrogated expression of IGF-I mRNA and caused a dramatic reduction in circulating IGF-I levels. Yakar *et al.*, *Proc. Natl. Acad. Sci. USA* **96**:7324-7329 (1999). When mammary tumors were induced in the LID mouse, reduced circulating IGF-1 levels resulted in significant reductions in cancer

development, growth, and metastases, whereas increased circulating IGF-1 levels were associated with enhanced tumor growth. Wu *et al.*, *Cancer Res.* **63**:4384-4388 (2003).

[0011] Several papers have reported that inhibition of IGF-IR expression and/or signaling leads to inhibition of tumor growth, both *in vitro* and *in vivo*. Inhibition of IGF signaling has also been shown to increase the susceptibility of tumor cells to chemotherapeutic agents. A variety of strategies (antisense oligonucleotides, soluble receptor, inhibitory peptides, dominant negative receptor mutants, small molecules inhibiting the kinase activity and anti-hIGF-IR antibodies) have been developed to inhibit the IGF-IR signaling pathway in tumor cells. One approach has been to target the kinase activity of IGF-IR with small molecule inhibitors. Two compounds were recently identified as small molecule kinase inhibitors capable of selectively inhibiting the IGF-IR. Garcia-Echeverria *et al.*, *Cancer Cell* **5**:231-239 (2004); Mitsiades *et al.*, *Cancer Cell* 5:221-230 (2004). Inhibition of IGF-IR kinase activity abrogated IGF-I-mediated survival and colony formation in soft agar of MCF-7 human breast cancer cells. Garcia-Echeverria *et al.*, *Cancer Cell* **5**:231-239 (2004). When an IGF-IR kinase inhibitor was administered to mice bearing tumor xenografts, IGF-IR signaling in tumor xenografts was inhibited and the growth of IGF-IR-driven fibrosarcomas was significantly reduced. Garcia-Echeverria *et al.*, *Cancer Cell* 5:231-239 (2004). A similar effect was observed on hematologic malignancies, especially multiple myeloma. In multiple myeloma cells, a small molecule IGF-IR kinase inhibitor demonstrated a >16-fold greater potency against the IGF-1R, as compared to the insulin receptor, and was similarly effective in inhibiting cell growth and survival. Mitsiades *et al.*, *Cancer Cell* **5**:221-230 (2004). The same compound was injected intraperitoneally into mice and inhibited multiple myeloma cell growth and enhanced survival of the mice. Mitsiades *et al.*, *Cancer Cell* **5**:221-230 (2004). When combined with other chemotherapeutics at subtherapeutic doses, inhibition of IGF-IR kinase activity synergistically reduced tumor burden. Mitsiades *et al.*, *Cancer Cell* **5**:221-230 (2004).

[0012] Another approach to inhibit IGF signaling has been the development of neutralizing antibodies directed against the receptor IGF-IR. Various groups have developed antibodies to IGF-IR that inhibit receptor IGF-I-stimulated autophosphorylation, induce receptor internalization and degradation, and reduce proliferation and survival of diverse human cancer cell lines. Hailey *et al.*, *Mol Cancer Ther.* **1**:1349-1353 (2002); Maloney *et al.*, *Cancer Res.* **63**:5073-5083 (2003); Benini *et al.*, *Clin. Cancer Res.* 7:1790-1797 (2001); Burtrum *et al.*, *Cancer Res.* **63**:8912-8921

(2003). Additionally, in xenograft tumor models, IGF-IR blockade resulted in significant growth inhibition of breast, renal and pancreatic tumors *in vivo*. Burtrum *et al.*, *Cancer Res.* **63**:8912-8921 (2003); Maloney *et al.*, *Cancer Res.* **63**:5073-5083 (2003). Experiments utilizing chimeric humanized IGF-IR antibodies yielded similar results, inhibiting growth of breast cancer cells *in vitro* and in tumor xenografts. Sachdev et al., Cancer Res. 63:627-635 (2003). Other humanized IGF-IR antibodies blocked IGF-I-induced tyrosine phosphorylation and growth inhibition in breast and non small cell lung tumors, as well as *in vivo*. Cohen *et al.*, *Clin. Cancer Res.* **11**:2063-2073 (2005); Goetsch *et al.*, *Int. J. Cancer* **113**:316-328 (2005).

[0013] Increased IGF-I levels have also been associated with several non-cancerous pathological conditions, including acromegaly and gigantism (Barkan, Cleveland Clin. J. Med. 65: 343, 347-349, 1998), while abnormal IGF-I/IGF-II receptor function has been implicated in psoriasis (Wraight et al., Nat. Biotech. 18: 521-526, 2000), atherosclerosis and smooth muscle restenosis of blood vessels following angioplasty (Bayes-Genis et al., Circ. Res. 86: 125-130, 2000). Increased IGF-I levels have been implicated in diabetes or in complications associated with diabetes, such as microvascular proliferation (Smith et al., Nat. Med. 5: 1390-1395, 1999).

[0014] Antibodies to IGF-I and IGF-II have been disclosed in the art. See, for example, Goya *et al.*, *Cancer Res.* **64**:6252-6258 (2004); Miyamoto *et al.*, *Clin. Cancer Res.* **11**:3494-3502 (2005). Additionally, see WO 05/18671, WO 05/28515 and WO 03/93317.

SUMMARY

[0015] Embodiments of the invention relate to binding proteins that specifically bind to insulin-like growth factors and reduce tumor growth. In one embodiment, the binding proteins are fully human monoclonal antibodies, or binding fragments thereof that specifically bind to insulin-like growth factors and reduce tumor growth. Mechanisms by which this can be achieved can include and are not limited to either inhibition of binding of IGF-I/II to its receptor IGF-IR, inhibition of IGF-I/II-induced IGF-IR signaling, or increased clearance of IGF-I/II, therein reducing the effective concentration of IGF-I/II.

[0016] Thus, some embodiments provide a fully human isolated specific binding protein that preferentially binds to insulin-like growth factor-II (IGF-II) with cross-reactivity to insulin-like growth factor I (IGF-I) and neutralizes IGF-I and IGF-II

activity. In certain aspects, the binding protein binds to IGF-II with at least 2.5 times greater affinity than to IGF-I. In other aspects, the binding protein binds to IGF-II with at least 3, at least 4, at least 5, at least 7, at least 10, at least 50, at least 60, at least 100 or at least 150 times greater affinity than to IGF-I.

[0017] In some embodiments, the specific binding protein has an EC_{50} of no more than 15 nM for inhibiting IGF-I-dependent IGF-I receptor phosphorylation in NIH3T3 cells expressing IGF-1R ectopically. In some aspects, the specific binding protein has an EC_{50} of no more than 15 nM, no more than 10 nM, or no more than 8 nM for inhibiting IGF-I-dependent IGF-I receptor phosphorylation in NIH3T3 cells expressing IGF-1R ectopically.

[0018] In some embodiments, the specific binding protein has an EC_{50} of no more than 5 nM, no more than 4 nM, or no more than 3 nM for inhibiting IGF-II-dependent IGF-I receptor phosphorylation in NIH3T3 cells expressing IGF-1R ectopically.

[0019] In other embodiments, the specific binding protein inhibits greater than 70% of IGF-II dependent proliferation of NIH3T3 cells that express recombinant hIGF-IR with an EC_{50} of no more than 25 nM, no more than 20 nM, no more than 15 nM, or no more than 10 nM.

[0020] In other embodiments, the specific binding protein inhibits greater than 70% of IGF-I dependent proliferation of NIH3T3 cells that express recombinant hIGF-IR with an EC_{50} of no more than 40 nM, no more than 30 nM, or no more than 25 nM.

[0021] In certain embodiments, the specific binding protein competes for binding with a monoclonal antibody comprising a variable heavy chain sequence selected from the group consisting of SEQ ID NO.: 2, SEQ ID NO.: 6, SEQ ID NO.: 10, SEQ ID NO.: 14 and SEQ ID NO.: 18, and comprising a variable light chain sequence selected from the group consisting of SEQ ID NO.: 4, SEQ ID NO.: 8, SEQ ID NO.: 12 and SEQ ID NO.: 16.

[0022] One embodiment of the invention is a fully human antibody that binds to IGF-I with a Kd less than 500 picomolar (pM). More preferably, the antibody binds with a Kd less than 450 picomolar (pM). More preferably, the antibody binds with a Kd less than 410 picomolar (pM). More preferably, the antibody binds with a K_d of less than 350 pM. Even more preferably, the antibody binds with a K_d of less than 300 pM. Affinity and/or avidity measurements can be measured by BIACORE®, as described herein.

[0023] Yet another embodiment of the invention is a fully human monoclonal antibody that binds to IGF-II with a K_d of less than 175 picomolar (pM). More preferably, the antibody binds with a Kd less than 100 picomolar (pM). More preferably, the antibody binds with a Kd less than 50 picomolar (pM). More preferably, the antibody binds with a Kd less than 5 picomolar (pM). Even more preferably, the antibody binds with a K_d of less than 2 pM.

[0024] In certain embodiments, the specific binding protein is a fully human monoclonal antibody or a binding fragment of a fully human monoclonal antibody. The binding fragments can include fragments such as Fab, Fab' or $F(ab')_2$ and Fv.

[0025] One embodiment of the invention comprises fully human monoclonal antibodies 7.251.3 (ATCC Accession Number PTA-7422), 7.34.1 (ATCC Accession Number PTA-7423) and 7.159.2 (ATCC Accession Number PTA-7424) which specifically bind to IGF-I/II, as discussed in more detail below.

[0026] In some embodiments the specific binding protein that binds to insulin-like growth factor-II (IGF-II) with cross-reactivity to insulin-like growth factor-I (IGF-I), or binding fragment thereof can include a heavy chain polypeptide having the sequence of SEQ ID NO.: 6, and a light chain polypeptide having the sequence of SEQ ID NO.: 8.

[0027] The specific binding protein can include a heavy chain polypeptide having the sequence of SEQ ID NO.: 10, and a light chain polypeptide having the sequence of SEQ ID NO.: 12.

[0028] The specific binding protein of the invention can include heavy chain polypeptide having the sequence of SEQ ID NO.: 14 and a light chain polypeptide having the sequence of SEQ ID NO.: 16.

[0029] In certain embodiments, the specific binding protein can be in a mixture with a pharmaceutically acceptable carrier.

[0030] Another embodiment includes isolated nucleic acid molecules encoding any of the specific binding proteins described herein, vectors having isolated nucleic acid molecules encoding the specific binding proteins, or a host cell transformed with any of such nucleic acid molecules and vectors.

[0031] In certain embodiments the specific binding protein that binds to insulin-like growth factor-II (IGF-II) with cross-reactivity to insulin-like growth factor-I (IGF-I), or binding fragment thereof does not bind specifically to IGF-II or IGF-I proteins when said proteins are bound to Insulin Growth Factor Binding Proteins.

[0032] Further embodiments include methods of determining the level of insulin-like growth factor-II (IGF-II) and insulin-like growth factor I (IGF-I) in a patient sample. These methods can include providing a patient sample; contacting the sample with a specific binding protein that binds to insulin-like growth factor-II (IGF-II) with cross-reactivity to insulin-like growth factor-I (IGF-I), or binding fragment thereof; and determining the level of IGF-I and IGF-II in said sample. In some aspects, the patient sample is blood.

[0033] Additional embodiments include methods of treating a malignant tumor in a mammal. These methods can include selecting a mammal in need of treatment for a malignant tumor; and administering to the mammal a therapeutically effective dose of a specific binding protein that binds to insulin-like growth factor-II (IGF-II) with cross-reactivity to insulin-like growth factor-I (IGF-I), or binding fragment thereof. In some aspects the animal is human. In some aspects the binding protein is a fully human monoclonal antibody, and is selected from the group consisting of mAb 7.251.3 (ATCC Accession Number PTA-7422), mAb 7.34.1 (ATCC Accession Number PTA-7423), and mAb 7.159.2 (ATCC Accession Number PTA-7424).

[0034] Treatable diseases can include melanoma, non-small cell lung cancer, glioma, hepatocellular (liver) carcinoma, thyroid tumor, gastric (stomach) cancer, prostrate cancer, breast cancer, ovarian cancer, bladder cancer, lung cancer, glioblastoma, endometrial cancer, kidney cancer, colon cancer, pancreatic cancer, and epidermoid carcinoma.

[0035] Additional embodiments include methods of treating a growth factor-dependent disease in a mammal. These methods include selecting a mammal in need of treatment for a growth factor-dependent disease; and administering to said mammal a therapeutically effective dose of a specific binding protein that binds to insulin-like growth factor-II (IGF-II) with cross-reactivity to insulin-like growth factor-I (IGF-I), or binding fragment thereof. In some aspects, the mammal can be human. In some aspects the binding protein is a fully human monoclonal antibody, and is selected from the group consisting of mAb 7.251.3 (ATCC Accession Number PTA-7422), mAb 7.34.1 (ATCC Accession Number PTA-7423), and mAb 7.159.2 (ATCC Accession Number PTA-7424).

[0036] Treatable growth factor-dependent diseases can include osteoporosis, diabetes, and cardiovascular diseases. Other treatable disease conditions include acromegaly and gigantism, psoriasis, atherosclerosis and smooth muscle restenosis of blood vessels, as well as diabetes.

[0037] Additional embodiments include a conjugate comprising a fully human monoclonal antibody that binds to insulin-like growth factor-II (IGF-II) with cross-reactivity to insulin-like growth factor-I (IGF-I), or a binding fragment thereof and a therapeutic agent. In some aspects the therapeutic agent can be a toxin, a radioisotope, or a pharmaceutical composition.

[0038] In other embodiments, the invention provides fully human monoclonal antibodies, or binding fragment thereof, that bind to insulin-like growth factor-II (IGF-II) with cross-reactivity to insulin-like growth factor-I (IGF-I), and comprise a heavy chain complementarity determining region 1 (CDR1) having the amino acid sequence of "Ser Tyr Tyr Trp Ser" (SEQ ID NO: 21); a heavy chain complementarity determining region 2 (CDR2) having the amino acid sequence of "Tyr Phe Phe Tyr Ser Gly Tyr Thr Asn Tyr Asn Pro Ser Leu Lys Ser" (SEQ ID NO: 22); and a heavy chain complementarity determining region 3 (CDR3) having the amino acid sequence of "Ile Thr Gly Thr Thr Lys Gly Gly Met Asp Val" (SEQ ID NO: 23).

[0039] Further embodiments include fully human monoclonal antibodies, or binding fragment thereof, having a light chain complementarity determining region 1 (CDR1) having the amino acid sequence of "Thr Gly Ser Ser Ser Asn Ile Gly Ala Gly Tyr Asp Val His" (SEQ ID NO: 24). Antibodies herein can also include a light chain complementarity determining region 2 (CDR2) having the amino acid sequence of "Gly Asn Asn Asn Arg Pro Ser" (SEQ ID NO: 25); and a light chain complementarity determining region 3 (CDR3) having the amino acid sequence of "Gln Ser Phe Asp Ser Ser Leu Ser Gly Ser Val" (SEQ ID NO: 26).

[0040] In other embodiments, the invention provides fully human monoclonal antibodies, or binding fragment thereof, that bind to insulin-like growth factor-II (IGF-II) with cross-reactivity to insulin-like growth factor-I (IGF-I), and comprise a heavy chain complementarity determining region 1 (CDR1) having the amino acid sequence of "Ser Tyr Tyr Trp Ser" (SEQ ID NO: 27); a heavy chain complementarity determining region 2 (CDR2) having the amino acid sequence of "Tyr Phe Phe Tyr Ser Gly Tyr Thr Asn Tyr Asn Pro Ser Leu Lys Ser" (SEQ ID NO: 28); and a heavy chain complementarity determining region 3 (CDR3) having the amino acid sequence of "Ile Thr Gly Thr Thr Lys Gly Gly Met Asp Val" (SEQ ID NO: 29).

[0041] Further embodiments include fully human monoclonal antibodies, or binding fragment thereof, having a light chain complementarity determining region 1 (CDR1) having the amino acid sequence of "Thr Gly Arg Ser Ser Asn Ile Gly Ala Gly

Tyr Asp Val His" (SEQ ID NO: 30); a light chain complementarity determining region 2 (CDR2) having the amino acid sequence of "Gly Asn Ser Asn Arg Pro Ser" (SEQ ID NO: 31); and a light chain complementarity determining region 3 (CDR3) having the amino acid sequence of "Gln Ser Tyr Asp Ser Ser Leu Ser Gly Ser Val" (SEQ ID NO: 32).

[0042] In other embodiments, the invention provides fully human monoclonal antibodies, or binding fragment thereof, that bind to insulin-like growth factor-II (IGF-II) with cross-reactivity to insulin-like growth factor-I (IGF-I), and comprise a heavy chain complementarity determining region 1 (CDR1) having the amino acid sequence of "Ser Tyr Asp Ile Asn" (SEQ ID NO: 33); a heavy chain complementarity determining region 2 (CDR2) having the amino acid sequence of "Trp Met Asn Pro Asn Ser Gly Asn Thr Gly Tyr Ala Gln Lys Phe Gln Gly" (SEQ ID NO: 34); and a heavy chain complementarity determining region 3 (CDR3) having the amino acid sequence of "Asp Pro Tyr Tyr Tyr Tyr Tyr Gly Met Asp Val" (SEQ ID NO: 35).

[0043] Further embodiments include fully human monoclonal antibodies, or binding fragment thereof, having a light chain complementarity determining region 1 (CDR1) having the amino acid sequence of "Ser Gly Ser Ser Ser Asn Ile Glu Asn Asn His Val Ser" (SEQ ID NO: 36); a light chain complementarity determining region 2 (CDR2) having the amino acid sequence of "Asp Asn Asn Lys Arg Pro Ser" (SEQ ID NO: 37); and a light chain complementarity determining region 3 (CDR3) having the amino acid sequence of "Glu Thr Trp Asp Thr Ser Leu Ser Ala Gly Arg Val" (SEQ ID NO: 38).

[0044] In other embodiments, the invention provides fully human monoclonal antibodies, or binding fragment thereof, that bind to insulin-like growth factor-II (IGF-II) with cross-reactivity to insulin-like growth factor-I (IGF-I), and comprise a heavy chain complementarity determining region 1 (CDR1) having the amino acid sequence of "Ser Ser Ser Tyr Tyr Trp Gly" (SEQ ID NO: 81); a heavy chain complementarity determining region 2 (CDR2) having the amino acid sequence of "Gly Ile Tyr Tyr Ser Gly Ser Thr Tyr Tyr Asn Pro Ser Leu Lys Ser" (SEQ ID NO: 82); and a heavy chain complementarity determining region 3 (CDR3) having the amino acid sequence of "Gln Arg Gly His Ser Ser Gly Trp Trp Tyr Phe Asp Leu" (SEQ ID NO: 83).

[0045] Further embodiments include fully human monoclonal antibodies, or binding fragment thereof, having a light chain complementarity determining region 1 (CDR1) having the amino acid sequence of "Arg Ala Ser Gln Gly Ile Ser Ser Tyr Leu Ala" (SEQ ID NO: 84); a light chain complementarity determining region 2 (CDR2)

having the amino acid sequence of "Ala Ala Ser Ser Leu Gln Ser" (SEQ ID NO: 85); and a light chain complementarity determining region 3 (CDR3) having the amino acid sequence of "Gln Gln Ala Asn Asn Phe Pro Phe Thr" (SEQ ID NO: 86).

[0046] In other embodiments, the invention provides fully human monoclonal antibodies, or binding fragment thereof, that bind to insulin-like growth factor-II (IGF-II) with cross-reactivity to insulin-like growth factor-I (IGF-I), and comprise a heavy chain complementarity determining region 1 (CDR1) having the amino acid sequence of "Ser Ser Ser Asn Tyr Trp Gly" (SEQ ID NO: 87); a heavy chain complementarity determining region 2 (CDR2) having the amino acid sequence of "Gly Ile Tyr Tyr Ser Gly Ser Thr Tyr Tyr Asn Pro Ser Leu Arg Ser" (SEQ ID NO: 88); and a heavy chain complementarity determining region 3 (CDR3) having the amino acid sequence of "Gln Arg Gly His Ser Ser Gly Trp Trp Tyr Phe Asp Leu" (SEQ ID NO: 89).

[0047] Further embodiments include fully human monoclonal antibodies, or binding fragment thereof, having a light chain complementarity determining region 1 (CDR1) having the amino acid sequence of "Arg Ala Ser Arg Gly Ile Ser Ser Trp Leu Ala" (SEQ ID NO: 90); a light chain complementarity determining region 2 (CDR2) having the amino acid sequence of "Thr Ala Ser Ser Leu Gln Ser" (SEQ ID NO: 91); and a light chain complementarity determining region 3 (CDR3) having the amino acid sequence of "Gln Gln Ala Asn Ser Phe Pro Phe Thr" (SEQ ID NO: 92).

[0048] Some embodiments provide the use of the specific binding proteins described herein in the preparation of a medicament for the treatment of a malignant tumor. In some aspects, the specific binding protein can be a fully human monoclonal antibody. In certain aspects, the binding protein is mAb 7.251.3 (ATCC Accession Number PTA-7422) or mAb 7.34.1 (ATCC Accession Number PTA-7423) or mAb 7.159.2 (ATCC Accession Number PTA-7424). In some aspects, the medicament is for use in combination with a second anti-neoplastic agent selected from the group consisting of an antibody, a chemotherapeutic agent, and a radioactive drug. In some aspects, the medicament is for use in conjunction with or following a conventional surgery, a bone marrow stem cell transplantation or a peripheral stem cell transplantation.

[0049] The malignant tumor can be melanoma, non-small cell lung cancer, glioma, hepatocellular (liver) carcinoma, thyroid tumor, gastric (stomach) cancer, prostrate cancer, breast cancer, ovarian cancer, bladder cancer, lung cancer, glioblastoma, endometrial cancer, kidney cancer, colon cancer, pancreatic cancer, and epidermoid carcinoma, for example.

[0050] Other embodiments provide the use of the specific binding proteins described herein in the preparation of a medicament for the treatment of a growth factor-dependent disease. In some aspects, the specific binding protein is a fully human monoclonal antibody and can be selected from the group consisting of mAb 7.251.3 (ATCC Accession Number PTA-7422), mAb 7.34.1 (ATCC Accession Number PTA-7423), and mAb 7.159.2 (ATCC Accession Number PTA-7424).

[0051] The growth factor-dependent disease can be osteoporosis, diabetes, and cardiovascular diseases, for example.

[0052] Preferably, the antibody comprises a heavy chain amino acid sequence having a complementarity determining region (CDR) with one or more of the sequences shown in Table 11. For example, the antibody can comprise a heavy chain amino acid sequence having the CDR1, CDR2, or CDR3 of one or more of the sequences shown in Table 11, or a combination thereof. It is noted that those of ordinary skill in the art can readily accomplish CDR determinations. See for example, Kabat *et al.*, *Sequences of Proteins of Immunological Interest,* Fifth Edition, NIH Publication 91-3242, Bethesda MD (1991), vols. 1-3.

[0053] Embodiments of the invention described herein relate to monoclonal antibodies that bind IGF-I/II and affect IGF-I/II function. Other embodiments relate to fully human anti-IGF-I/II antibodies and anti-IGF-I/II antibody preparations with desirable properties from a therapeutic perspective, including high binding affinity for IGF-I/II, the ability to neutralize IGF-I/II *in vitro* and *in vivo*, and the ability to inhibit IGF-I/II induced cell proliferation.

BRIEF DESCRIPTION OF THE DRAWINGS

[0054] Figure 1 is a graph showing inhibition of xenograft tumor growth in nude mice of NIH3T3 cells expressing IGF-II and IGF-IR (Clone 32 cells) with mAbs 7.159.2, 7.34.1, 7.251.3 compared to IgG2 and PBS controls. Mean tumor volume is shown on the y-axis and time after implantation is shown on the x-axis.

[0055] Figure 2 is a graph showing body weight in Clone 32 xenograft mice treated with mAbs 7.159.2, 7.34.1, 7.251.3 compared to IgG2 and PBS controls. Mean body weight is shown on the y-axis and time after implantation is shown on the x-axis.

[0056] Figure 3 is a graph showing inhibition of xenograft tumor growth in nude mice of NIH3T3 cells expressing IGF-I and IGF-IR (P12 cells) with mAb 7.159.2

compared to PBS control. Mean tumor volume is shown on the y-axis and time after implantation (indicated by date) is shown on the x-axis.

DETAILED DESCRIPTION

[0057] Embodiments of the invention described herein relate to binding proteins that specifically bind to IGF-II with cross reactivity to IGF-I (referred to herein as "IGFI/II"). In some embodiments, the binding proteins are antibodies, or binding fragments thereof, and bind to IGF-II with cross-reactivity to IGF-I and inhibit the binding of these proteins to their receptor, IGF-IR. Other embodiments of the invention include fully human neutralizing anti-IGF-I/II antibodies, and antibody preparations that are therapeutically useful and bind both insulin-like growth factors. Such anti-IGF-I/II antibody preparations preferably have desirable therapeutic properties, including strong binding affinity for IGF-I/II, the ability to neutralize IGF-I/II *in vitro*, and the ability to inhibit IGF-I/II-induced cell proliferation *in vivo*.

[0058] Embodiments of the invention also include isolated binding fragments of anti-IGF-I/II antibodies. Preferably, the binding fragments are derived from fully human anti-IGF-I/II antibodies. Exemplary fragments include Fv, Fab' or other well know antibody fragments, as described in more detail below. Embodiments of the invention also include cells that express fully human antibodies against IGF-I/II. Examples of cells include hybridomas, or recombinantly created cells, such as Chinese hamster ovary (CHO) cells that produce antibodies against IGF-I/II.

[0059] In addition, embodiments of the invention include methods of using these antibodies for treating diseases. Anti-IGF-I/II antibodies are useful for preventing IGF-I/II mediated IGF-I/II signal transduction, thereby inhibiting cell proliferation. The mechanism of action of this inhibition may include inhibition of IGF-I/II from binding to its receptor, IGF-IR, inhibition of IGF-I/II induced IGF-IR signaling, or enhanced clearance of IGF-I/II therein lowering the effective concentration of IGF-I/II for binding to IGF-IR. Diseases that are treatable through this inhibition mechanism include, but are not limited to, neoplastic diseases, such as, melanoma, non-small cell lung cancer, glioma, hepatocellular (liver) carcinoma, gynecologic tumors, head and neck cancer, esophageal cancer, glioblastoma, and cancers and tumors of the thyroid, stomach, prostrate, breast, ovary, bladder, lung, uterus, kidney, colon, and pancreas, salivary gland, and colorectum.

[0060] Other embodiments of the invention include diagnostic assays for specifically determining the quantity of IGF-I/II in a biological sample. The assay kit can include anti-IGF-I/II antibodies along with the necessary labels for detecting such antibodies. These diagnostic assays are useful to screen for growth factor-related diseases including, but not limited to, neoplastic diseases, such as, melanoma, non-small cell lung cancer, glioma, hepatocellular (liver) carcinoma, gynecologic tumors, head and neck cancer, esophageal cancer, glioblastoma, and carcinoma of the thyroid, stomach, prostrate, breast, ovary, bladder, lung, uterus, kidney, colon, and pancreas, salivary gland, and colorectum. Other non-neoplastic disease conditions may include acromegaly and gigantism, psoriasis, osteoporosis, atherosclerosis and smooth muscle restenosis of blood vessels, as well as diabetes.

[0061] Further embodiments, features, and the like regarding anti-IGF-I/II antibodies are provided in additional detail below.

Sequence Listing

[0062] Embodiments of the invention include the specific anti-IGF-I/II antibodies listed below in Table 1. This table reports the identification number of each anti-IGF-I/II antibody, along with the SEQ ID number of the corresponding heavy chain and light chain genes. Further, the germline sequences from which each heavy chain and light chain derive are also provided below in Table 1.

[0063] Each antibody has been given an identification number that includes either two or three numbers separated by one or two decimal points. In some cases, several clones of one antibody were prepared. Although the clones have the identical nucleic acid and amino acid sequences as the parent sequence, they may also be listed separately, with the clone number indicated by the number to the right of a second decimal point. Thus, for example, the nucleic acid and amino acid sequences of antibody 7.159.2 are identical to the sequences of antibody 7.159.1.

[0064] As can be seen by comparing the sequences in the sequence listing, SEQ ID NOs.: 1-20 differ from SEQ ID NOs.: 39-58 because SEQ ID NOs.: 39-58 include the untranslated, signal peptide, and constant domain regions for each sequenced heavy or light chain.

TABLE 1

mAb ID No.:	Sequence	SEQ ID NO:
7.158.1	Nucleotide sequence encoding the variable region of the heavy chain	1
	Amino acid sequence encoding the variable region of the heavy chain	2
	Nucleotide sequence encoding the variable region of the light chain	3
	Amino acid sequence encoding the variable region of the light chain	4
7.159.2	Nucleotide sequence encoding the variable region of the heavy chain	5
	Amino acid sequence encoding the variable region of the heavy chain	6
	Nucleotide sequence encoding the variable region of the light chain	7
	Amino acid sequence encoding the variable region of the light chain	8
7.34.1	Nucleotide sequence encoding the variable region of the heavy chain	9
	Amino acid sequence encoding the variable region of the heavy chain	10
	Nucleotide sequence encoding the variable region of the light chain	11
	Amino acid sequence encoding the variable region of the light chain	12
7.251.3	Nucleotide sequence encoding the variable region of the heavy chain	13
	Amino acid sequence encoding the variable region of the heavy chain	14
	Nucleotide sequence encoding the variable region of the light chain	15
	Amino acid sequence encoding the variable region of the light chain	16
7.234.1	Nucleotide sequence encoding the variable region of the heavy chain	17
	Amino acid sequence encoding the variable region of the heavy chain	18
	Nucleotide sequence encoding the variable region of the light chain	19
	Amino acid sequence encoding the variable region of the light chain	20
7.158.1	Nucleotide sequence encoding the variable region of the heavy chain	39
	Amino acid sequence encoding the variable region of the heavy chain	40
	Nucleotide sequence encoding the variable region of the light chain	41
	Amino acid sequence encoding the variable region of the light chain	42
7.159.2	Nucleotide sequence encoding the variable region of the heavy chain	43
	Amino acid sequence encoding the variable region of the heavy chain	44
	Nucleotide sequence encoding the variable region of the light chain	45
	Amino acid sequence encoding the variable region of the light chain	46
7.34.1	Nucleotide sequence encoding the variable region of the heavy chain	47
	Amino acid sequence encoding the variable region of the heavy chain	48
	Nucleotide sequence encoding the variable region of the light chain	49
	Amino acid sequence encoding the variable region of the light chain	50
7.251.3	Nucleotide sequence encoding the variable region of the heavy chain	51
	Amino acid sequence encoding the variable region of the heavy chain	52
	Nucleotide sequence encoding the variable region of the light chain	53
	Amino acid sequence encoding the variable region of the light chain	54
7.234.1	Nucleotide sequence encoding the variable region of the heavy chain	55
	Amino acid sequence encoding the variable region of the heavy chain	56
	Nucleotide sequence encoding the variable region of the light chain	57
	Amino acid sequence encoding the variable region of the light chain	58
Germline (7.158.1)	Nucleotide sequence encoding the variable region of the heavy chain	59
	Amino acid sequence encoding the variable region of the heavy chain	60
	Nucleotide sequence encoding the variable region of the light chain	61

	Amino acid sequence encoding the variable region of the light chain	62
Germline (7.159.1)	Nucleotide sequence encoding the variable region of the heavy chain	63
	Amino acid sequence encoding the variable region of the heavy chain	64
	Nucleotide sequence encoding the variable region of the light chain	65
	Amino acid sequence encoding the variable region of the light chain	66
Germline (7.34.1)	Nucleotide sequence encoding the variable region of the heavy chain	67
	Amino acid sequence encoding the variable region of the heavy chain	68
	Nucleotide sequence encoding the variable region of the light chain	69
	Amino acid sequence encoding the variable region of the light chain	70
Germline (7.251.3)	Nucleotide sequence encoding the variable region of the heavy chain	71
	Amino acid sequence encoding the variable region of the heavy chain	72
	Nucleotide sequence encoding the variable region of the light chain	73
	Amino acid sequence encoding the variable region of the light chain	74

Definitions

[0065] Unless otherwise defined, scientific and technical terms used herein shall have the meanings that are commonly understood by those of ordinary skill in the art. Further, unless otherwise required by context, singular terms shall include pluralities and plural terms shall include the singular. Generally, nomenclatures utilized in connection with, and techniques of, cell and tissue culture, molecular biology, and protein and oligo- or polynucleotide chemistry and hybridization described herein are those well known and commonly used in the art.

[0066] Standard techniques are used for recombinant DNA, oligonucleotide synthesis, and tissue culture and transformation (e.g., electroporation, lipofection). Enzymatic reactions and purification techniques are performed according to manufacturer's specifications or as commonly accomplished in the art or as described herein. The foregoing techniques and procedures are generally performed according to conventional methods well known in the art and as described in various general and more specific references that are cited and discussed throughout the present specification. *See e.g.,* Sambrook *et al. Molecular Cloning: A Laboratory Manual* (3rd ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. (2001)), which is incorporated herein by reference. The nomenclatures utilized in connection with, and the laboratory procedures and techniques of, analytical chemistry, synthetic organic chemistry, and medicinal and pharmaceutical chemistry described herein are those well known and commonly used in the art. Standard techniques are used for chemical syntheses, chemical analyses, pharmaceutical preparation, formulation, and delivery, and treatment of patients.

[0067] As utilized in accordance with the present disclosure, the following terms, unless otherwise indicated, shall be understood to have the following meanings:

[0068] The term "IGF-I" refers to the molecule Insulin-like growth factor-I, and the term "IGF-II" refers to the molecule Insulin-like growth factor-II. The term "IGF-I/II" refers to both molecules Insulin-like growth factors-I and -II, and relates to the preferential binding to IGF-II with cross-reactivity to IGF-I. Thus, an antibody that binds to IGF-I/II will preferentially bind to IGF-II, but would cross-react with IGF-I, binding to IGF-II with higher affinity than to IGF-I. For example, the antibody can bind to IGF-II with 2.5 times greater affinity than to IGF-I. In certain embodiments, the antibody can bind to IGF-II with at least 5, at least 10, at least 25, at least 50 or at least 150 times greater affinity than to IGF-I.

[0069] The term "neutralizing" when referring to an antibody relates to the ability of an antibody to eliminate, or significantly reduce, the activity of a target antigen. Accordingly, a "neutralizing" anti-IGF-I/II antibody is capable of eliminating or significantly reducing the activity of IGF-I/II. A neutralizing IGF-I/II antibody may, for example, act by blocking the binding of IGF-I/II to its receptor IGF-IR. By blocking this binding, the IGF-IR mediated signal transduction is significantly, or completely, eliminated. Ideally, a neutralizing antibody against IGF-I/II inhibits cell proliferation.

[0070] The term "isolated polynucleotide" as used herein shall mean a polynucleotide that has been isolated from its naturally occurring environment. Such polynucleotides may be genomic, cDNA, or synthetic. Isolated polynucleotides preferably are not associated with all or a portion of the polynucleotides they associate with in nature. The isolated polynucleotides may be operably linked to another polynucleotide that it is not linked to in nature. In addition, isolated polynucleotides preferably do not occur in nature as part of a larger sequence.

[0071] The term "isolated protein" referred to herein means a protein that has been isolated from its naturally occurring environment. Such proteins may be derived from genomic DNA, cDNA, recombinant DNA, recombinant RNA, or synthetic origin or some combination thereof, which by virtue of its origin, or source of derivation, the "isolated protein" (1) is not associated with proteins found in nature, (2) is free of other proteins from the same source, e.g. free of murine proteins, (3) is expressed by a cell from a different species, or (4) does not occur in nature.

[0072] The term "polypeptide" is used herein as a generic term to refer to native protein, fragments, or analogs of a polypeptide sequence. Hence, native protein,

fragments, and analogs are species of the polypeptide genus. Preferred polypeptides in accordance with the invention comprise the human heavy chain immunoglobulin molecules and the human kappa light chain immunoglobulin molecules, as well as antibody molecules formed by combinations comprising the heavy chain immunoglobulin molecules with light chain immunoglobulin molecules, such as the kappa or lambda light chain immunoglobulin molecules, and vice versa, as well as fragments and analogs thereof. Preferred polypeptides in accordance with the invention may also comprise solely the human heavy chain immunoglobulin molecules or fragments thereof.

[0073] The term "naturally-occurring" as used herein as applied to an object refers to the fact that an object can be found in nature. For example, a polypeptide or polynucleotide sequence that is present in an organism (including viruses) that can be isolated from a source in nature and which has not been intentionally modified by man in the laboratory or otherwise is naturally-occurring.

[0074] The term "operably linked" as used herein refers to positions of components so described that are in a relationship permitting them to function in their intended manner. For example, a control sequence "operably linked" to a coding sequence is connected in such a way that expression of the coding sequence is achieved under conditions compatible with the control sequences.

[0075] The term "polynucleotide" as referred to herein means a polymeric form of nucleotides of at least 10 bases in length, either ribonucleotides or deoxynucleotides or a modified form of either type of nucleotide, or RNA-DNA hetero-duplexes. The term includes single and double stranded forms of DNA.

[0076] The term "oligonucleotide" referred to herein includes naturally occurring, and modified nucleotides linked together by naturally occurring, and non-naturally occurring linkages. Oligonucleotides are a polynucleotide subset generally comprising a length of 200 bases or fewer. Preferably, oligonucleotides are 10 to 60 bases in length and most preferably 12, 13, 14, 15, 16, 17, 18, 19, or 20 to 40 bases in length. Oligonucleotides are usually single stranded, e.g. for probes; although oligonucleotides may be double stranded, e.g. for use in the construction of a gene mutant. Oligonucleotides can be either sense or antisense oligonucleotides.

[0077] The term "naturally occurring nucleotides" referred to herein includes deoxyribonucleotides and ribonucleotides. The term "modified nucleotides" referred to herein includes nucleotides with modified or substituted sugar groups and the like. The term "oligonucleotide linkages" referred to herein includes oligonucleotides linkages such

as phosphorothioate, phosphorodithioate, phosphoroselenoate, phosphorodiselenoate, phosphoroanilothioate, phosphoraniladate, phosphoroamidate, and the like. *See e.g.,* LaPlanche *et al. Nucl. Acids Res.* 14:9081 (1986); Stec *et al. J. Am. Chem. Soc.* 106:6077 (1984); Stein *et al. Nucl. Acids Res.* 16:3209 (1988); Zon *et al. Anti-Cancer Drug Design* 6:539 (1991); Zon *et al. Oligonucleotides and Analogues: A Practical Approach*, pp. 87-108 (F. Eckstein, Ed., Oxford University Press, Oxford England (1991)); Stec *et al.* U.S. Patent No. 5,151,510; Uhlmann and Peyman *Chemical Reviews* 90:543 (1990), the disclosures of which are hereby incorporated by reference. An oligonucleotide can include a label for detection, if desired.

[0078] The term "selectively hybridize" referred to herein means to detectably and specifically bind. Polynucleotides, oligonucleotides and fragments thereof selectively hybridize to nucleic acid strands under hybridization and wash conditions that minimize appreciable amounts of detectable binding to nonspecific nucleic acids. High stringency conditions can be used to achieve selective hybridization conditions as known in the art and discussed herein. Generally, the nucleic acid sequence homology between the polynucleotides, oligonucleotides, or antibody fragments and a nucleic acid sequence of interest will be at least 80%, and more typically with preferably increasing homologies of at least 85%, 90%, 95%, 99%, and 100%.

[0079] Two amino acid sequences are "homologous" if there is a partial or complete identity between their sequences. For example, 85% homology means that 85% of the amino acids are identical when the two sequences are aligned for maximum matching. Gaps (in either of the two sequences being matched) are allowed in maximizing matching; gap lengths of 5 or less are preferred with 2 or less being more preferred. Alternatively and preferably, two protein sequences (or polypeptide sequences derived from them of at least about 30 amino acids in length) are homologous, as this term is used herein, if they have an alignment score of at more than 5 (in standard deviation units) using the program ALIGN with the mutation data matrix and a gap penalty of 6 or greater. *See* Dayhoff, M.O., in *Atlas of Protein Sequence and Structure*, pp. 101-110 (Volume 5, National Biomedical Research Foundation (1972)) and Supplement 2 to this volume, pp. 1-10. The two sequences or parts thereof are more preferably homologous if their amino acids are greater than or equal to 50% identical when optimally aligned using the ALIGN program. It should be appreciated that there can be differing regions of homology within two orthologous sequences. For example,

the functional sites of mouse and human orthologues may have a higher degree of homology than non-functional regions.

[0080] The term "corresponds to" is used herein to mean that a polynucleotide sequence is homologous (i.e., is identical, not strictly evolutionarily related) to all or a portion of a reference polynucleotide sequence, or that a polypeptide sequence is identical to a reference polypeptide sequence.

[0081] In contradistinction, the term "complementary to" is used herein to mean that the complementary sequence is homologous to all or a portion of a reference polynucleotide sequence. For illustration, the nucleotide sequence "TATAC" corresponds to a reference sequence "TATAC" and is complementary to a reference sequence "GTATA".

[0082] The following terms are used to describe the sequence relationships between two or more polynucleotide or amino acid sequences: "reference sequence", "comparison window", "sequence identity", "percentage of sequence identity", and "substantial identity". A "reference sequence" is a defined sequence used as a basis for a sequence comparison. A reference sequence may be a subset of a larger sequence, for example, as a segment of a full-length cDNA or gene sequence given in a sequence listing or may comprise a complete cDNA or gene sequence. Generally, a reference sequence is at least 18 nucleotides or 6 amino acids in length, frequently at least 24 nucleotides or 8 amino acids in length, and often at least 48 nucleotides or 16 amino acids in length. Since two polynucleotides or amino acid sequences may each (1) comprise a sequence (i.e., a portion of the complete polynucleotide or amino acid sequence) that is similar between the two molecules, and (2) may further comprise a sequence that is divergent between the two polynucleotides or amino acid sequences, sequence comparisons between two (or more) molecules are typically performed by comparing sequences of the two molecules over a "comparison window" to identify and compare local regions of sequence similarity. A "comparison window", as used herein, refers to a conceptual segment of at least about 18 contiguous nucleotide positions or about 6 amino acids wherein the polynucleotide sequence or amino acid sequence is compared to a reference sequence of at least 18 contiguous nucleotides or 6 amino acid sequences and wherein the portion of the polynucleotide sequence in the comparison window may include additions, deletions, substitutions, and the like (i.e., gaps) of 20 percent or less as compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. Optimal alignment of sequences for aligning a comparison window may

be conducted by the local homology algorithm of Smith and Waterman *Adv. Appl. Math.* 2:482 (1981), by the homology alignment algorithm of Needleman and Wunsch *J. Mol. Biol.* 48:443 (1970), by the search for similarity method of Pearson and Lipman *Proc. Natl. Acad. Sci. (U.S.A.)* 85:2444 (1988), by computerized implementations of these algorithms (GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package Release 7.0, (Genetics Computer Group, 575 Science Dr., Madison, Wis.), GENEWORKS™, or MACVECTOR® software packages), or by inspection, and the best alignment (i.e., resulting in the highest percentage of homology over the comparison window) generated by the various methods is selected.

[0083] The term "sequence identity" means that two polynucleotide or amino acid sequences are identical (i.e., on a nucleotide-by-nucleotide or residue-by-residue basis) over the comparison window. The term "percentage of sequence identity" is calculated by comparing two optimally aligned sequences over the window of comparison, determining the number of positions at which the identical nucleic acid base (e.g., A, T, C, G, U, or I) or amino acid residue occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the comparison window (i.e., the window size), and multiplying the result by 100 to yield the percentage of sequence identity. The terms "substantial identity" as used herein denotes a characteristic of a polynucleotide or amino acid sequence, wherein the polynucleotide or amino acid comprises a sequence that has at least 85 percent sequence identity, preferably at least 90 to 95 percent sequence identity, more preferably at least 99 percent sequence identity, as compared to a reference sequence over a comparison window of at least 18 nucleotide (6 amino acid) positions, frequently over a window of at least 24-48 nucleotide (8-16 amino acid) positions, wherein the percentage of sequence identity is calculated by comparing the reference sequence to the sequence which may include deletions or additions which total 20 percent or less of the reference sequence over the comparison window. The reference sequence may be a subset of a larger sequence.

[0084] As used herein, the twenty conventional amino acids and their abbreviations follow conventional usage. *See Immunology - A Synthesis* (2nd Edition, E.S. Golub and D.R. Gren, Eds., Sinauer Associates, Sunderland, Mass. (1991)), which is incorporated herein by reference. Stereoisomers (e.g., D-amino acids) of the twenty conventional amino acids, unnatural amino acids such as α-, α-disubstituted amino acids,

N-alkyl amino acids, lactic acid, and other unconventional amino acids may also be suitable components for polypeptides of the present invention. Examples of unconventional amino acids include: 4-hydroxyproline, γ-carboxyglutamate, ε-N,N,N-trimethyllysine, ε-N-acetyllysine, O-phosphoserine, N-acetylserine, N-formylmethionine, 3-methylhistidine, 5-hydroxylysine, σ-N-methylarginine, and other similar amino acids and imino acids (e.g., 4-hydroxyproline). In the polypeptide notation used herein, the left-hand direction is the amino terminal direction and the right-hand direction is the carboxy-terminal direction, in accordance with standard usage and convention.

[0085] Similarly, unless specified otherwise, the left-hand end of single-stranded polynucleotide sequences is the 5' end; the left-hand direction of double-stranded polynucleotide sequences is referred to as the 5' direction. The direction of 5' to 3' addition of nascent RNA transcripts is referred to as the transcription direction; sequence regions on the DNA strand having the same sequence as the RNA and which are 5' to the 5' end of the RNA transcript are referred to as "upstream sequences"; sequence regions on the DNA strand having the same sequence as the RNA and which are 3' to the 3' end of the RNA transcript are referred to as "downstream sequences".

[0086] As applied to polypeptides, the term "substantial identity" means that two peptide sequences, when optimally aligned, such as by the programs GAP or BESTFIT using default gap weights, share at least 80 percent sequence identity, preferably at least 90 percent sequence identity, more preferably at least 95 percent sequence identity, and most preferably at least 99 percent sequence identity. Preferably, residue positions that are not identical differ by conservative amino acid substitutions. Conservative amino acid substitutions refer to the interchangeability of residues having similar side chains. For example, a group of amino acids having aliphatic side chains is glycine, alanine, valine, leucine, and isoleucine; a group of amino acids having aliphatic-hydroxyl side chains is serine and threonine; a group of amino acids having amide-containing side chains is asparagine and glutamine; a group of amino acids having aromatic side chains is phenylalanine, tyrosine, and tryptophan; a group of amino acids having basic side chains is lysine, arginine, and histidine; and a group of amino acids having sulfur-containing side chains is cysteine and methionine. Preferred conservative amino acids substitution groups are: valine-leucine-isoleucine, phenylalanine-tyrosine, lysine-arginine, alanine-valine, glutamic-aspartic, and asparagine-glutamine.

[0087] As discussed herein, minor variations in the amino acid sequences of antibodies or immunoglobulin molecules are contemplated as being encompassed by the present invention, providing that the variations in the amino acid sequence maintain at least 75%, more preferably at least 80%, 90%, 95%, and most preferably 99% sequence identity to the antibodies or immunoglobulin molecules described herein. In particular, conservative amino acid replacements are contemplated. Conservative replacements are those that take place within a family of amino acids that have related side chains. Genetically encoded amino acids are generally divided into families: (1) acidic=aspartate, glutamate; (2) basic=lysine, arginine, histidine; (3) non-polar=alanine, valine, leucine, isoleucine, proline, phenylalanine, methionine, tryptophan; and (4) uncharged polar=glycine, asparagine, glutamine, cysteine, serine, threonine, tyrosine. More preferred families are: serine and threonine are an aliphatic-hydroxy family; asparagine and glutamine are an amide-containing family; alanine, valine, leucine and isoleucine are an aliphatic family; and phenylalanine, tryptophan, and tyrosine are an aromatic family. For example, it is reasonable to expect that an isolated replacement of a leucine with an isoleucine or valine, an aspartate with a glutamate, a threonine with a serine, or a similar replacement of an amino acid with a structurally related amino acid will not have a major effect on the binding function or properties of the resulting molecule, especially if the replacement does not involve an amino acid within a framework site. Whether an amino acid change results in a functional peptide can readily be determined by assaying the specific activity of the polypeptide derivative. Assays are described in detail herein. Fragments or analogs of antibodies or immunoglobulin molecules can be readily prepared by those of ordinary skill in the art. Preferred amino- and carboxy-termini of fragments or analogs occur near boundaries of functional domains. Structural and functional domains can be identified by comparison of the nucleotide and/or amino acid sequence data to public or proprietary sequence databases. Preferably, computerized comparison methods are used to identify sequence motifs or predicted protein conformation domains that occur in other proteins of known structure and/or function. Methods to identify protein sequences that fold into a known three-dimensional structure are known. Bowie *et al. Science* 253:164 (1991). Thus, the foregoing examples demonstrate that those of skill in the art can recognize sequence motifs and structural conformations that may be used to define structural and functional domains in accordance with the antibodies described herein.

[0088] Preferred amino acid substitutions are those which: (1) reduce susceptibility to proteolysis, (2) reduce susceptibility to oxidation, (3) alter binding affinity for forming protein complexes, (4) alter binding affinities, and (5) confer or modify other physicochemical or functional properties of such analogs. Analogs can include various muteins of a sequence other than the naturally-occurring peptide sequence. For example, single or multiple amino acid substitutions (preferably conservative amino acid substitutions) may be made in the naturally-occurring sequence (preferably in the portion of the polypeptide outside the domain(s) forming intermolecular contacts. A conservative amino acid substitution should not substantially change the structural characteristics of the parent sequence (e.g., a replacement amino acid should not tend to break a helix that occurs in the parent sequence, or disrupt other types of secondary structure that characterizes the parent sequence). Examples of art-recognized polypeptide secondary and tertiary structures are described in *Proteins, Structures and Molecular Principles* (Creighton, Ed., W. H. Freeman and Company, New York (1984)); *Introduction to Protein Structure* (C. Branden and J. Tooze, eds., Garland Publishing, New York, N.Y. (1991)); and Thornton et at. *Nature* 354:105 (1991), which are each incorporated herein by reference.

[0089] The term "polypeptide fragment" as used herein refers to a polypeptide that has an amino-terminal and/or carboxy-terminal deletion, but where the remaining amino acid sequence is identical to the corresponding positions in the naturally-occurring sequence deduced, for example, from a full-length cDNA sequence. Fragments typically are at least 5, 6, 8 or 10 amino acids long, preferably at least 14 amino acids long, more preferably at least 20 amino acids long, usually at least 50 amino acids long, and even more preferably at least 70 amino acids long. The term "analog" as used herein refers to polypeptides which are comprised of a segment of at least 25 amino acids that has substantial identity to a portion of a deduced amino acid sequence and which has at least one of the following properties: (1) specific binding to IGF-I/II, under suitable binding conditions, (2) ability to block appropriate IGF-I/II binding, or (3) ability to inhibit IGF-I/II activity. Typically, polypeptide analogs comprise a conservative amino acid substitution (or addition or deletion) with respect to the naturally-occurring sequence. Analogs typically are at least 20 amino acids long, preferably at least 50 amino acids long or longer, and can often be as long as a full-length naturally-occurring polypeptide.

[0090] Peptide analogs are commonly used in the pharmaceutical industry as non-peptide drugs with properties analogous to those of the template peptide. These

types of non-peptide compound are termed "peptide mimetics" or "peptidomimetics". Fauchere, *J. Adv. Drug Res.* 15:29 (1986); Veber and Freidinger *TINS* p.392 (1985); and Evans *et al. J. Med. Chem.* 30:1229 (1987), which are incorporated herein by reference. Such compounds are often developed with the aid of computerized molecular modeling. Peptide mimetics that are structurally similar to therapeutically useful peptides may be used to produce an equivalent therapeutic or prophylactic effect. Generally, peptidomimetics are structurally similar to a paradigm polypeptide (i.e., a polypeptide that has a biochemical property or pharmacological activity), such as human antibody, but have one or more peptide linkages optionally replaced by a linkage selected from the group consisting of: --CH_2NH--, --CH_2S--, --CH_2-CH_2--, --CH=CH--(cis and trans), --$COCH_2$--, --$CH(OH)CH_2$--, and –CH_2SO--, by methods well known in the art. Systematic substitution of one or more amino acids of a consensus sequence with a D-amino acid of the same type (e.g., D-lysine in place of L-lysine) may be used to generate more stable peptides. In addition, constrained peptides comprising a consensus sequence or a substantially identical consensus sequence variation may be generated by methods known in the art (Rizo and Gierasch *Ann. Rev. Biochem.* 61:387 (1992), incorporated herein by reference); for example, by adding internal cysteine residues capable of forming intramolecular disulfide bridges which cyclize the peptide.

[0091] As used herein, the term "antibody" refers to a polypeptide or group of polypeptides that are comprised of at least one binding domain that is formed from the folding of polypeptide chains having three-dimensional binding spaces with internal surface shapes and charge distributions complementary to the features of an antigenic determinant of an antigen. An antibody typically has a tetrameric form, comprising two identical pairs of polypeptide chains, each pair having one "light" and one "heavy" chain. The variable regions of each light/heavy chain pair form an antibody binding site.

[0092] "Binding fragments" of an antibody are produced by recombinant DNA techniques, or by enzymatic or chemical cleavage of intact antibodies. Binding fragments include Fab, Fab', $F(ab')_2$, Fv, and single-chain antibodies. An antibody other than a "bispecific" or "bifunctional" antibody is understood to have each of its binding sites identical. An antibody substantially inhibits adhesion of a receptor to a counterreceptor when an excess of antibody reduces the quantity of receptor bound to counterreceptor by at least about 20%, 40%, 60% or 80%, and more usually greater than about 85% (as measured in an *in vitro* competitive binding assay).

[0093] As used herein, a "binding protein" or a "specific binding protein" are proteins that specifically bind to a target molecule. Antibodies, and binding fragments of antibodies, are binding proteins.

[0094] The term "epitope" includes any protein determinant capable of specific binding to an immunoglobulin or T-cell receptor. Epitopic determinants usually consist of chemically active surface groupings of molecules such as amino acids or sugar side chains and may, but not always, have specific three-dimensional structural characteristics, as well as specific charge characteristics. An antibody is said to specifically bind an antigen when the dissociation constant is ≤ 1 μM, preferably ≤ 100 nM and most preferably ≤ 10 nM.

[0095] The term "agent" is used herein to denote a chemical compound, a mixture of chemical compounds, a biological macromolecule, or an extract made from biological materials.

[0096] "Active" or "activity" in regard to an IGF-I/II polypeptide refers to a portion of an IGF-I/II polypeptide that has a biological or an immunological activity of a native IGF-I/II polypeptide. "Biological" when used herein refers to a biological function that results from the activity of the native IGF-I/II polypeptide. A preferred IGF-I/II biological activity includes, for example, IGF-I/II induced cell proliferation.

[0097] "Mammal" when used herein refers to any animal that is considered a mammal. Preferably, the mammal is human.

[0098] Digestion of antibodies with the enzyme, papain, results in two identical antigen-binding fragments, known also as "Fab" fragments, and a "Fc" fragment, having no antigen-binding activity but having the ability to crystallize. Digestion of antibodies with the enzyme, pepsin, results in the a $F(ab')_2$ fragment in which the two arms of the antibody molecule remain linked and comprise two-antigen binding sites. The $F(ab')_2$ fragment has the ability to crosslink antigen.

[0099] "Fv" when used herein refers to the minimum fragment of an antibody that retains both antigen-recognition and antigen-binding sites.

[0100] "Fab" when used herein refers to a fragment of an antibody that comprises the constant domain of the light chain and the CH1 domain of the heavy chain.

[0101] The term "mAb" refers to monoclonal antibody.

[0102] "Liposome" when used herein refers to a small vesicle that may be useful for delivery of drugs that may include the IGF-I/II polypeptide of the invention or antibodies to such an IGF-I/II polypeptide to a mammal.

[0103] "Label" or "labeled" as used herein refers to the addition of a detectable moiety to a polypeptide, for example, a radiolabel, fluorescent label, enzymatic label chemiluminescent labeled or a biotinyl group. Radioisotopes or radionuclides may include ^{3}H, ^{14}C, ^{15}N, ^{35}S, ^{90}Y, ^{99}Tc, ^{111}In, ^{125}I, ^{131}I, fluorescent labels may include rhodamine, lanthanide phosphors or FITC and enzymatic labels may include horseradish peroxidase, β-galactosidase, luciferase, alkaline phosphatase.

[0104] The term "pharmaceutical agent or drug" as used herein refers to a chemical compound or composition capable of inducing a desired therapeutic effect when properly administered to a patient. Other chemistry terms herein are used according to conventional usage in the art, as exemplified by *The McGraw-Hill Dictionary of Chemical Terms* (Parker, S., Ed., McGraw-Hill, San Francisco (1985)), (incorporated herein by reference).

[0105] As used herein, "substantially pure" means an object species is the predominant species present (i.e., on a molar basis it is more abundant than any other individual species in the composition), and preferably a substantially purified fraction is a composition wherein the object species comprises at least about 50 percent (on a molar basis) of all macromolecular species present. Generally, a substantially pure composition will comprise more than about 80 percent of all macromolecular species present in the composition, more preferably more than about 85%, 90%, 95%, and 99%. Most preferably, the object species is purified to essential homogeneity (contaminant species cannot be detected in the composition by conventional detection methods) wherein the composition consists essentially of a single macromolecular species.

[0106] The term "patient" includes human and veterinary subjects.

Human Antibodies and Humanization of Antibodies

[0107] Human antibodies avoid some of the problems associated with antibodies that possess murine or rat variable and/or constant regions. The presence of such murine or rat derived proteins can lead to the rapid clearance of the antibodies or can lead to the generation of an immune response against the antibody by a patient. In order to avoid the utilization of murine or rat derived antibodies, fully human antibodies can be generated through the introduction of functional human antibody loci into a rodent, other

mammal or animal so that the rodent, other mammal or animal produces fully human antibodies.

[0108] One method for generating fully human antibodies is through the use of XenoMouse® strains of mice that have been engineered to contain up to but less than 1000 kb-sized germline configured fragments of the human heavy chain locus and kappa light chain locus. *See* Mendez *et al. Nature Genetics* 15:146-156 (1997) and Green and Jakobovits *J. Exp. Med.* 188:483-495 (1998). The XenoMouse® strains are available from Abgenix, Inc. (Fremont, CA).

[0109] The production of the XenoMouse® strains of mice is further discussed and delineated in U.S. Patent Application Serial Nos. 07/466,008, filed January 12, 1990, 07/610,515, filed November 8, 1990, 07/919,297, filed July 24, 1992, 07/922,649, filed July 30, 1992, 08/031,801, filed March 15, 1993, 08/112,848, filed August 27, 1993, 08/234,145, filed April 28, 1994, 08/376,279, filed January 20, 1995, 08/430, 938, filed April 27, 1995, 08/464,584, filed June 5, 1995, 08/464,582, filed June 5, 1995, 08/463,191, filed June 5, 1995, 08/462,837, filed June 5, 1995, 08/486,853, filed June 5, 1995, 08/486,857, filed June 5, 1995, 08/486,859, filed June 5, 1995, 08/462,513, filed June 5, 1995, 08/724,752, filed October 2, 1996, 08/759,620, filed December 3, 1996, U.S. Publication 2003/0093820, filed November 30, 2001 and U.S. Patent Nos. 6,162,963, 6,150,584, 6,114,598, 6,075,181, and 5,939,598 and Japanese Patent Nos. 3 068 180 B2, 3 068 506 B2, and 3 068 507 B2. *See also* European Patent No., EP 0 463 151 B1, grant published June 12, 1996, International Patent Application No., WO 94/02602, published February 3, 1994, International Patent Application No., WO 96/34096, published October 31, 1996, WO 98/24893, published June 11, 1998, WO 00/76310, published December 21, 2000. The disclosures of each of the above-cited patents, applications, and references are hereby incorporated by reference in their entirety.

[0110] In an alternative approach, others, including GenPharm International, Inc., have utilized a "minilocus" approach. In the minilocus approach, an exogenous Ig locus is mimicked through the inclusion of pieces (individual genes) from the Ig locus. Thus, one or more V_H genes, one or more D_H genes, one or more J_H genes, a mu constant region, and usually a second constant region (preferably a gamma constant region) are formed into a construct for insertion into an animal. This approach is described in U.S. Patent No. 5,545,807 to Surani *et al.* and U.S. Patent Nos. 5,545,806, 5,625,825, 5,625,126, 5,633,425, 5,661,016, 5,770,429, 5,789,650, 5,814,318, 5,877,397, 5,874,299,

and 6,255,458 each to Lonberg and Kay, U.S. Patent No. 5,591,669 and 6,023.010 to Krimpenfort and Berns, U.S. Patent Nos. 5,612,205, 5,721,367, and 5,789,215 to Berns *et al.*, and U.S. Patent No. 5,643,763 to Choi and Dunn, and GenPharm International U.S. Patent Application Serial Nos. 07/574,748, filed August 29, 1990, 07/575,962, filed August 31, 1990, 07/810,279, filed December 17, 1991, 07/853,408, filed March 18, 1992, 07/904,068, filed June 23, 1992, 07/990,860, filed December 16, 1992, 08/053,131, filed April 26, 1993, 08/096,762, filed July 22, 1993, 08/155,301, filed November 18, 1993, 08/161,739, filed December 3, 1993, 08/165,699, filed December 10, 1993, 08/209,741, filed March 9, 1994, the disclosures of which are hereby incorporated by reference. *See also* European Patent No. 0 546 073 B1, International Patent Application Nos. WO 92/03918, WO 92/22645, WO 92/22647, WO 92/22670, WO 93/12227, WO 94/00569, WO 94/25585, WO 96/14436, WO 97/13852, and WO 98/24884 and U.S. Patent No. 5,981,175, the disclosures of which are hereby incorporated by reference in their entirety. *See further* Taylor *et al.*, 1992, Chen *et al.*, 1993, Tuaillon *et al.*, 1993, Choi *et al.*, 1993, Lonberg *et al.*, (1994), Taylor *et al.*, (1994), and Tuaillon *et al.*, (1995), Fishwild *et al.*, (1996), the disclosures of which are hereby incorporated by reference in their entirety.

[0111] Kirin has also demonstrated the generation of human antibodies from mice in which, through microcell fusion, large pieces of chromosomes, or entire chromosomes, have been introduced. *See* European Patent Application Nos. 773 288 and 843 961, the disclosures of which are hereby incorporated by reference. Additionally, KM™– mice, which are the result of cross-breeding of Kirin's Tc mice with Medarex's minilocus (Humab) mice have been generated. These mice possess the human IgH transchromosome of the Kirin mice and the kappa chain transgene of the Genpharm mice (Ishida *et al.*, Cloning Stem Cells, (2002) 4:91-102).

[0112] Human antibodies can also be derived by in vitro methods. Suitable examples include but are not limited to phage display (CAT, Morphosys, Dyax, Biosite/Medarex, Xoma, Symphogen, Alexion (formerly Proliferon), Affimed) ribosome display (CAT), yeast display, and the like.

Preparation of Antibodies

[0113] Antibodies, as described herein, were prepared through the utilization of the XenoMouse® technology, as described below. Such mice, then, are capable of

producing human immunoglobulin molecules and antibodies and are deficient in the production of murine immunoglobulin molecules and antibodies. Technologies utilized for achieving the same are disclosed in the patents, applications, and references disclosed in the background section herein. In particular, however, a preferred embodiment of transgenic production of mice and antibodies therefrom is disclosed in U.S. Patent Application Serial No. 08/759,620, filed December 3, 1996 and International Patent Application Nos. WO 98/24893, published June 11, 1998 and WO 00/76310, published December 21, 2000, the disclosures of which are hereby incorporated by reference. *See also* Mendez *et al. Nature Genetics* 15:146-156 (1997), the disclosure of which is hereby incorporated by reference.

[0114] Through the use of such technology, fully human monoclonal antibodies to a variety of antigens have been produced. Essentially, XenoMouse® lines of mice are immunized with an antigen of interest (e.g. IGF-I/II), lymphatic cells (such as B-cells) are recovered from the hyper-immunized mice, and the recovered lymphocytes are fused with a myeloid-type cell line to prepare immortal hybridoma cell lines. These hybridoma cell lines are screened and selected to identify hybridoma cell lines that produced antibodies specific to the antigen of interest. Provided herein are methods for the production of multiple hybridoma cell lines that produce antibodies specific to IGF-I/II. Further, provided herein are characterization of the antibodies produced by such cell lines, including nucleotide and amino acid sequence analyses of the heavy and light chains of such antibodies.

[0115] Alternatively, instead of being fused to myeloma cells to generate hybridomas, B cells can be directly assayed. For example, CD19+ B cells can be isolated from hyperimmune XenoMouse® mice and allowed to proliferate and differentiate into antibody-secreting plasma cells. Antibodies from the cell supernatants are then screened by ELISA for reactivity against the IGF-I/II immunogen. The supernatants might also be screened for immunoreactivity against fragments of IGF-I/II to further map the different antibodies for binding to domains of functional interest on IGF-I/II. The antibodies may also be screened against other related human chemokines and against the rat, the mouse, and non-human primate, such as cynomolgus monkey, orthologues of IGF-I/II, the last to determine species cross-reactivity. B cells from wells containing antibodies of interest may be immortalized by various methods including fusion to make hybridomas either from individual or from pooled wells, or by infection with EBV or transfection by known

immortalizing genes and then plating in suitable medium. Alternatively, single plasma cells secreting antibodies with the desired specificities are then isolated using an IGF-I/II-specific hemolytic plaque assay (Babcook *et al.*, *Proc. Natl. Acad. Sci. USA* 93:7843-48 (1996)). Cells targeted for lysis are preferably sheep red blood cells (SRBCs) coated with the IGF-I/II antigen.

[0116] In the presence of a B-cell culture containing plasma cells secreting the immunoglobulin of interest and complement, the formation of a plaque indicates specific IGF-I/II-mediated lysis of the sheep red blood cells surrounding the plasma cell of interest. The single antigen-specific plasma cell in the center of the plaque can be isolated and the genetic information that encodes the specificity of the antibody is isolated from the single plasma cell. Using reverse-transcription followed by PCR (RT-PCR), the DNA encoding the heavy and light chain variable regions of the antibody can be cloned. Such cloned DNA can then be further inserted into a suitable expression vector, preferably a vector cassette such as a pcDNA, more preferably such a pcDNA vector containing the constant domains of immunglobulin heavy and light chain. The generated vector can then be transfected into host cells, e.g., HEK293 cells, CHO cells, and cultured in conventional nutrient media modified as appropriate for inducing transcription, selecting transformants, or amplifying the genes encoding the desired sequences.

[0117] In general, antibodies produced by the fused hybridomas were human IgG2 heavy chains with fully human kappa or lambda light chains. Antibodies described herein possess human IgG4 heavy chains as well as IgG2 heavy chains. Antibodies can also be of other human isotypes, including IgG1. The antibodies possessed high affinities, typically possessing a Kd of from about 10^{-6} through about 10^{-12} M or below, when measured by solid phase and solution phase techniques. Antibodies possessing a KD of at least 10^{-11} M are preferred to inhibit the activity of IGF-I/II.

[0118] As will be appreciated, anti-IGF-I/II antibodies can be expressed in cell lines other than hybridoma cell lines. Sequences encoding particular antibodies can be used to transform a suitable mammalian host cell. Transformation can be by any known method for introducing polynucleotides into a host cell, including, for example packaging the polynucleotide in a virus (or into a viral vector) and transducing a host cell with the virus (or vector) or by transfection procedures known in the art, as exemplified by U.S. Patent Nos. 4,399,216, 4,912,040, 4,740,461, and 4,959,455 (which patents are hereby incorporated herein by reference). The transformation procedure used depends upon the host to be transformed. Methods for introducing heterologous polynucleotides

into mammalian cells are well known in the art and include dextran-mediated transfection, calcium phosphate precipitation, polybrene mediated transfection, protoplast fusion, electroporation, encapsulation of the polynucleotide(s) in liposomes, and direct microinjection of the DNA into nuclei.

[0119] Mammalian cell lines available as hosts for expression are well known in the art and include many immortalized cell lines available from the American Type Culture Collection (ATCC), including but not limited to Chinese hamster ovary (CHO) cells, HeLa cells, baby hamster kidney (BHK) cells, monkey kidney cells (COS), human hepatocellular carcinoma cells (e.g., Hep G2), human epithelial kidney 293 cells, and a number of other cell lines. Cell lines of particular preference are selected through determining which cell lines have high expression levels and produce antibodies with constitutive IGF-I/II binding properties.

[0120] Anti-IGF-I/II antibodies are useful in the detection of IGF-I/II in patient samples and accordingly are useful as diagnostics for disease states as described herein. In addition, based on their ability to significantly neutralize IGF-I/II activity (as demonstrated in the Examples below), anti-IGF-I/II antibodies have therapeutic effects in treating symptoms and conditions resulting from IGF-I/II expression. In specific embodiments, the antibodies and methods herein relate to the treatment of symptoms resulting from IGF-I/II induced cell proliferation. Further embodiments involve using the antibodies and methods described herein to treat diseases including neoplastic diseases, such as, melanoma, non-small cell lung cancer, glioma, hepatocellular (liver) carcinoma, thyroid tumor, gastric (stomach) cancer, prostrate cancer, breast cancer, ovarian cancer, bladder cancer, lung cancer, glioblastoma, endometrial cancer, kidney cancer, colon cancer, gynecologic tumors, head and neck cancer, esophageal cancer, and pancreatic cancer. Other non-neoplastic disease conditions may include acromegaly and gigantism, psoriasis, osteoporosis, atherosclerosis and smooth muscle restenosis of blood vessels, as well as diabetes.

Therapeutic Administration and Formulations

[0121] Embodiments of the invention include sterile pharmaceutical formulations of anti-IGF-I/II antibodies that are useful as treatments for diseases. Such formulations would inhibit the binding of IGF-I/II to its receptor IGF-IR, thereby effectively treating pathological conditions where, for example, serum or tissue IGF-I/II

is abnormally elevated. Anti-IGF-I/II antibodies preferably possess adequate affinity to potently neutralize IGF-I/II, and preferably have an adequate duration of action to allow for infrequent dosing in humans. A prolonged duration of action will allow for less frequent and more convenient dosing schedules by alternate parenteral routes such as subcutaneous or intramuscular injection.

[0122] Sterile formulations can be created, for example, by filtration through sterile filtration membranes, prior to or following lyophilization and reconstitution of the antibody. The antibody ordinarily will be stored in lyophilized form or in solution. Therapeutic antibody compositions generally are placed into a container having a sterile access port, for example, an intravenous solution bag or vial having an adapter that allows retrieval of the formulation, such as a stopper pierceable by a hypodermic injection needle.

[0123] The route of antibody administration is in accord with known methods, e.g., injection or infusion by intravenous, intraperitoneal, intracerebral, intramuscular, intraocular, intraarterial, intrathecal, inhalation or intralesional routes, or by sustained release systems as noted below. The antibody is preferably administered continuously by infusion or by bolus injection.

[0124] An effective amount of antibody to be employed therapeutically will depend, for example, upon the therapeutic objectives, the route of administration, and the condition of the patient. Accordingly, it is preferred that the therapist titer the dosage and modify the route of administration as required to obtain the optimal therapeutic effect. Typically, the clinician will administer antibody until a dosage is reached that achieves the desired effect. The progress of this therapy is easily monitored by conventional assays or by the assays described herein.

[0125] Antibodies, as described herein, can be prepared in a mixture with a pharmaceutically acceptable carrier. This therapeutic composition can be administered intravenously or through the nose or lung, preferably as a liquid or powder aerosol (lyophilized). The composition may also be administered parenterally or subcutaneously as desired. When administered systemically, the therapeutic composition should be sterile, pyrogen-free and in a parenterally acceptable solution having due regard for pH, isotonicity, and stability. These conditions are known to those skilled in the art. Briefly, dosage formulations of the compounds described herein are prepared for storage or administration by mixing the compound having the desired degree of purity with physiologically acceptable carriers, excipients, or stabilizers. Such materials are non-

toxic to the recipients at the dosages and concentrations employed, and include buffers such as TRIS HCl, phosphate, citrate, acetate and other organic acid salts; antioxidants such as ascorbic acid; low molecular weight (less than about ten residues) peptides such as polyarginine, proteins, such as serum albumin, gelatin, or immunoglobulins; hydrophilic polymers such as polyvinylpyrrolidinone; amino acids such as glycine, glutamic acid, aspartic acid, or arginine; monosaccharides, disaccharides, and other carbohydrates including cellulose or its derivatives, glucose, mannose, or dextrins; chelating agents such as EDTA; sugar alcohols such as mannitol or sorbitol; counterions such as sodium and/or nonionic surfactants such as TWEEN, PLURONICS or polyethyleneglycol.

[0126] Sterile compositions for injection can be formulated according to conventional pharmaceutical practice as described in *Remington: The Science and Practice of Pharmacy* (20th ed, Lippincott Williams & Wilkens Publishers (2003)). For example, dissolution or suspension of the active compound in a vehicle such as water or naturally occurring vegetable oil like sesame, peanut, or cottonseed oil or a synthetic fatty vehicle like ethyl oleate or the like may be desired. Buffers, preservatives, antioxidants and the like can be incorporated according to accepted pharmaceutical practice.

[0127] Suitable examples of sustained-release preparations include semipermeable matrices of solid hydrophobic polymers containing the polypeptide, which matrices are in the form of shaped articles, films or microcapsules. Examples of sustained-release matrices include polyesters, hydrogels (e.g., poly(2-hydroxyethyl-methacrylate) as described by Langer *et al.*, *J. Biomed Mater. Res.*, (1981) 15:167-277 and Langer, *Chem. Tech.*, (1982) 12:98-105, or poly(vinylalcohol)), polylactides (U.S. Pat. No. 3,773,919, EP 58,481), copolymers of L-glutamic acid and gamma ethyl-L-glutamate (Sidman *et al.*, *Biopolymers*, (1983) 22:547-556), non-degradable ethylene-vinyl acetate (Langer *et al.*, *supra*), degradable lactic acid-glycolic acid copolymers such as the LUPRON Depot™ (injectable microspheres composed of lactic acid-glycolic acid copolymer and leuprolide acetate), and poly-D-(-)-3-hydroxybutyric acid (EP 133,988).

[0128] While polymers such as ethylene-vinyl acetate and lactic acid-glycolic acid enable release of molecules for over 100 days, certain hydrogels release proteins for shorter time periods. When encapsulated proteins remain in the body for a long time, they may denature or aggregate as a result of exposure to moisture at 37°C, resulting in a loss of biological activity and possible changes in immunogenicity. Rational strategies

can be devised for protein stabilization depending on the mechanism involved. For example, if the aggregation mechanism is discovered to be intermolecular S-S bond formation through disulfide interchange, stabilization may be achieved by modifying sulfhydryl residues, lyophilizing from acidic solutions, controlling moisture content, using appropriate additives, and developing specific polymer matrix compositions.

[0129] Sustained-released compositions also include preparations of crystals of the antibody suspended in suitable formulations capable of maintaining crystals in suspension. These preparations when injected subcutaneously or intraperitonealy can produce a sustained release effect. Other compositions also include liposomally entrapped antibodies. Liposomes containing such antibodies are prepared by methods known per se: U.S. Pat. No. DE 3,218,121; Epstein *et al., Proc. Natl. Acad. Sci. USA*, (1985) 82:3688-3692; Hwang *et al., Proc. Natl. Acad. Sci. USA*, (1980) 77:4030-4034; EP 52,322; EP 36,676; EP 88,046; EP 143,949; 142,641; Japanese patent application 83-118008; U.S. Pat. Nos. 4,485,045 and 4,544,545; and EP 102,324.

[0130] The dosage of the antibody formulation for a given patient will be determined by the attending physician taking into consideration various factors known to modify the action of drugs including severity and type of disease, body weight, sex, diet, time and route of administration, other medications and other relevant clinical factors. Therapeutically effective dosages may be determined by either *in vitro* or *in vivo* methods.

[0131] An effective amount of the antibodies, described herein, to be employed therapeutically will depend, for example, upon the therapeutic objectives, the route of administration, and the condition of the patient. Accordingly, it is preferred for the therapist to titer the dosage and modify the route of administration as required to obtain the optimal therapeutic effect. A typical daily dosage might range from about 0.001mg/kg to up to 100mg/kg or more, depending on the factors mentioned above. Typically, the clinician will administer the therapeutic antibody until a dosage is reached that achieves the desired effect. The progress of this therapy is easily monitored by conventional assays or as described herein.

[0132] It will be appreciated that administration of therapeutic entities in accordance with the compositions and methods herein will be administered with suitable carriers, excipients, and other agents that are incorporated into formulations to provide improved transfer, delivery, tolerance, and the like. These formulations include, for example, powders, pastes, ointments, jellies, waxes, oils, lipids, lipid (cationic or anionic)

containing vesicles (such as Lipofectin™), DNA conjugates, anhydrous absorption pastes, oil-in-water and water-in-oil emulsions, emulsions carbowax (polyethylene glycols of various molecular weights), semi-solid gels, and semi-solid mixtures containing carbowax. Any of the foregoing mixtures may be appropriate in treatments and therapies in accordance with the present invention, provided that the active ingredient in the formulation is not inactivated by the formulation and the formulation is physiologically compatible and tolerable with the route of administration. *See also* Baldrick P. "Pharmaceutical excipient development: the need for preclinical guidance." *Regul. Toxicol. Pharmacol.* 32(2):210-8 (2000), Wang W. "Lyophilization and development of solid protein pharmaceuticals." *Int. J. Pharm.* 203(1-2):1-60 (2000), Charman WN "Lipids, lipophilic drugs, and oral drug delivery-some emerging concepts." *J Pharm Sci* .89(8):967-78 (2000), Powell *et al.* "Compendium of excipients for parenteral formulations" *PDA J Pharm Sci Technol.* 52:238-311 (1998) and the citations therein for additional information related to formulations, excipients and carriers well known to pharmaceutical chemists.

Design and Generation of Other Therapeutics

[0133] In accordance with the present invention and based on the activity of the antibodies that are produced and characterized herein with respect to IGF-I/II, the design of other therapeutic modalities is facilitated and disclosed to one of skill in the art. Such modalities include, without limitation, advanced antibody therapeutics, such as bispecific antibodies, immunotoxins, radiolabeled therapeutics, and single antibody V domains, antibody-like binding agent based on other than V region scaffolds, generation of peptide therapeutics, gene therapies, particularly intrabodies, antisense therapeutics, and small molecules.

[0134] In connection with the generation of advanced antibody therapeutics, where complement fixation is a desirable attribute, it can be possible to sidestep the dependence on complement for cell killing through the use of bispecifics, immunotoxins, or radiolabels, for example.

[0135] For example, bispecific antibodies can be generated that comprise (i) two antibodies, one with a specificity to IGF-I/II and another to a second molecule, that are conjugated together, (ii) a single antibody that has one chain specific to IGF-I/II and a second chain specific to a second molecule, or (iii) a single chain antibody that has specificity to both IGF-I/II and the other molecule. Such bispecific antibodies can be

generated using techniques that are well known; for example, in connection with (i) and (ii) *see e.g.,* Fanger et al. *Immunol Methods* 4:72-81 (1994) and Wright and Harris, *supra.* and in connection with (iii) *see e.g.,* Traunecker et al. *Int. J. Cancer (Suppl.)* 7:51-52 (1992). In each case, the second specificity can be made as desired. For example, the second specificity can be made to the heavy chain activation receptors, including, without limitation, CD16 or CD64 (*see e.g.,* Deo et al. 18:127 (1997)) or CD89 (*see e.g.,* Valerius et al. *Blood* 90:4485-4492 (1997)).

[0136] Antibodies can also be modified to act as immunotoxins utilizing techniques that are well known in the art. *See e.g.,* Vitetta *Immunol Today* 14:252 (1993). *See also* U.S. Patent No. 5,194,594. In connection with the preparation of radiolabeled antibodies, such modified antibodies can also be readily prepared utilizing techniques that are well known in the art. *See e.g.,* Junghans et al. in *Cancer Chemotherapy and Biotherapy* 655-686 (2d edition, Chafner and Longo, eds., Lippincott Raven (1996)). *See also* U.S. Patent Nos. 4,681,581, 4,735,210, 5,101,827, 5,102,990 (RE 35,500), 5,648,471, and 5,697,902. Each of immunotoxins and radiolabeled molecules would be likely to kill cells expressing the desired multimeric enzyme subunit oligomerization domain. In some embodiments, a pharmaceutical composition comprising an effective amount of the antibody in association with a pharmaceutically acceptable carrier or diluent is provided.

[0137] In some embodiments, an anti-IGF-I/II antibody is linked to an agent (*e.g.,* radioisotope, pharmaceutical composition, or a toxin). Preferably, such antibodies can be used for the treatment of diseases, such diseases can relate to cells expressing IGF-I/II or cells overexpressing IGF-I/II. For example, it is contemplated that the drug possesses the pharmaceutical property selected from the group of antimitotic, alkylating, antimetabolite, antiangiogenic, apoptotic, alkaloid, COX-2, and antibiotic agents and combinations thereof. The drug can be selected from the group of nitrogen mustards, ethylenimine derivatives, alkyl sulfonates, nitrosoureas, triazenes, folic acid analogs, anthracyclines, taxanes, COX-2 inhibitors, pyrimidine analogs, purine analogs, antimetabolites, antibiotics, enzymes, epipodophyllotoxins, platinum coordination complexes, vinca alkaloids, substituted ureas, methyl hydrazine derivatives, adrenocortical suppressants, antagonists, endostatin, taxols, camptothecins, oxaliplatin, doxorubicins and their analogs, and a combination thereof.

[0138] Examples of toxins further include gelonin, Pseudomonas exotoxin (PE), PE40, PE38, diphtheria toxin, ricin, ricin, abrin, alpha toxin, saporin, ribonuclease

(RNase), DNase I, Staphylococcal enterotoxin-A, pokeweed antiviral protein, gelonin, Pseudomonas endotoxin, as well as derivatives, combinations and modifications thereof.

[0139] Examples of radioisotopes include gamma-emitters, positron-emitters, and x-ray emitters that can be used for localization and/or therapy, and beta-emitters and alpha-emitters that can be used for therapy. The radioisotopes described previously as useful for diagnostics, prognostics and staging are also useful for therapeutics. Non-limiting examples of anti-cancer or anti-leukemia agents include anthracyclines such as doxorubicin (adriamycin), daunorubicin (daunomycin), idarubicin, detorubicin, carminomycin, epirubicin, esorubicin, and morpholino and substituted derivatives, combinations and modifications thereof. Exemplary pharmaceutical agents include cis-platinum, taxol, calicheamicin, vincristine, cytarabine (Ara-C), cyclophosphamide, prednisone, daunorubicin, idarubicin, fludarabine, chlorambucil, interferon alpha, hydroxyurea, temozolomide, thalidomide, and bleomycin, and derivatives, combinations and modifications thereof. Preferably, the anti-cancer or anti-leukemia is doxorubicin, morpholinodoxorubicin, or morpholinodaunorubicin.

[0140] As will be appreciated by one of skill in the art, in the above embodiments, while affinity values can be important, other factors can be as important or more so, depending upon the particular function of the antibody. For example, for an immunotoxin (toxin associated with an antibody), the act of binding of the antibody to the target can be useful; however, in some embodiments, it is the internalization of the toxin into the cell that is the desired end result. As such, antibodies with a high percent internalization can be desirable in these situations. Thus, in one embodiment, antibodies with a high efficiency in internalization are contemplated. A high efficiency of internalization can be measured as a percent internalized antibody, and can be from a low value to 100%. For example, in varying embodiments, 0.1-5, 5-10, 10-20, 20-30, 30-40, 40-45, 45-50, 50-60, 60-70, 70-80, 80-90, 90-99, and 99-100% can be a high efficiency. As will be appreciated by one of skill in the art, the desirable efficiency can be different in different embodiments, depending upon, for example, the associated agent, the amount of antibody that can be administered to an area, the side effects of the antibody-agent complex, the type (*e.g.*, cancer type) and severity of the problem to be treated.

[0141] In other embodiments, the antibodies disclosed herein provide an assay kit for the detection of IGF-I/II expression in mammalian tissues or cells in order to screen for a disease or disorder associated with changes in expression of IGF-I/II. The kit

comprises an antibody that binds IGF-I/II and means for indicating the reaction of the antibody with the antigen, if present.

[0142] In some embodiments, an article of manufacture is provided comprising a container, comprising a composition containing an anti-IGF-I/II antibody, and a package insert or label indicating that the composition can be used to treat disease mediated by IGF-I/II expression. Preferably a mammal, and more preferably, a human, receives the anti-IGF-I/II antibody.

Combinations

[0143] The anti-IGF-I/II antibodies defined hereinbefore may be applied as a sole therapy or may involve, in addition to the compound of the invention, conventional surgery or radiotherapy or chemotherapy. Such chemotherapy may include one or more of the following categories of anti-tumour agents :-

(i) antiproliferative/antineoplastic drugs and combinations thereof, as used in medical oncology, such as alkylating agents (for example cis-platin, carboplatin, cyclophosphamide, nitrogen mustard, melphalan, chlorambucil, busulphan and nitrosoureas); antimetabolites (for example antifolates such as fluoropyrimidines like 5-fluorouracil and tegafur, raltitrexed, methotrexate, cytosine arabinoside and hydroxyurea; antitumour antibiotics (for example anthracyclines like adriamycin, bleomycin, doxorubicin, daunomycin, epirubicin, idarubicin, mitomycin-C, dactinomycin and mithramycin); antimitotic agents (for example vinca alkaloids like vincristine, vinblastine, vindesine and vinorelbine and taxoids like taxol and taxotere); and topoisomerase inhibitors (for example epipodophyllotoxins like etoposide and teniposide, amsacrine, topotecan and camptothecin);

(ii) cytostatic agents such as antioestrogens (for example tamoxifen, toremifene, raloxifene, droloxifene and iodoxyfene), oestrogen receptor down regulators (for example fulvestrant), antiandrogens (for example bicalutamide, flutamide, nilutamide and cyproterone acetate), LHRH antagonists or LHRH agonists (for example goserelin, leuprorelin and buserelin), progestogens (for example megestrol acetate), aromatase inhibitors (for example as anastrozole, letrozole, vorazole and exemestane) and inhibitors of 5α-reductase such as finasteride;

(iii) agents which inhibit cancer cell invasion (for example metalloproteinase inhibitors like marimastat and inhibitors of urokinase plasminogen activator receptor function);

(iv) inhibitors of growth factor function, for example such inhibitors include growth factor antibodies, growth factor receptor antibodies (for example the anti-erbb2 antibody trastuzumab [Herceptin™] and the anti-erbb1 antibody cetuximab [C225]) , farnesyl transferase inhibitors, MEK inhibitors, tyrosine kinase inhibitors and serine/threonine kinase inhibitors, for example inhibitors of the epidermal growth factor family (for example EGFR family tyrosine kinase inhibitors such as *N*-(3-chloro-4-fluorophenyl)-7-methoxy-6-(3-morpholinopropoxy)quinazolin-4-amine (gefitinib, AZD1839), *N*-(3-ethynylphenyl)-6,7-bis(2-methoxyethoxy)quinazolin-4-amine (erlotinib, OSI-774) and 6-acrylamido-*N*-(3-chloro-4-fluorophenyl)-7-(3-morpholinopropoxy)quinazolin-4-amine (CI 1033)), for example inhibitors of the platelet-derived growth factor family and for example inhibitors of the hepatocyte growth factor family;

(v) antiangiogenic agents such as those which inhibit the effects of vascular endothelial growth factor, (for example the anti-vascular endothelial cell growth factor antibody bevacizumab [Avastin™], compounds such as those disclosed in International Patent Applications WO 97/22596, WO 97/30035, WO 97/32856 and WO 98/13354) and compounds that work by other mechanisms (for example linomide, inhibitors of integrin αvβ3 function, angiostatin and inhibitors of the action of angiopoietins e.g angiopoietin 1 and angiopoietin 2);

(vi) vascular damaging agents such as Combretastatin A4 and compounds disclosed in International Patent Applications WO 99/02166, WO00/40529, WO 00/41669, WO01/92224, WO02/04434 and WO02/08213;

(vii) antisense therapies, for example those which are directed to the targets listed above, such as ISIS 2503, an anti-ras antisense;

(viii) gene therapy approaches, including for example approaches to replace aberrant genes such as aberrant p53 or aberrant BRCA1 or BRCA2, GDEPT (gene-directed enzyme pro-drug therapy) approaches such as those using cytosine deaminase, thymidine kinase or a bacterial nitroreductase enzyme and approaches to increase patient tolerance to chemotherapy or radiotherapy such as multi-drug resistance gene therapy;

(ix) immunotherapy approaches, including for example ex-vivo and in-vivo approaches to increase the immunogenicity of patient tumour cells, such as transfection with cytokines such as interleukin 2, interleukin 4 or granulocyte-macrophage colony stimulating factor, approaches to decrease T-cell anergy, approaches using transfected immune cells such as cytokine-transfected dendritic cells, approaches using cytokine-transfected tumour cell lines and approaches using anti-idiotypic antibodies;

(x) cell cycle inhibitors including for example CDK inhibitiors (eg flavopiridol) and other inhibitors of cell cycle checkpoints (eg checkpoint kinase); inhibitors of aurora kinase and other kinases involved in mitosis and cytokinesis regulation (eg mitotic kinesins); and histone deacetylase inhibitors;

(xi) endothelin antagonists, including endothelin A antagonists, endothelin B antagonists and endothelin A and B antagonists; for example ZD4054 and ZD1611 (WO 96 40681), atrasentan and YM598; and

(xii) biotherapeutic therapeutic approaches for example those which use peptides or proteins (such as antibodies or soluble external receptor domain constructions) which either sequest receptor ligands, block ligand binding to receptor or decrease receptor signalling (e.g. due to enhanced receptor degradation or lowered expression levels)

[0144] Such conjoint treatment may be achieved by way of the simultaneous, sequential or separate dosing of the individual components of the treatment. Such combination products employ the compounds of this invention within the dosage range described hereinbefore and the other pharmaceutically-active agent within its approved dosage range.

EXAMPLES

[0145] The following examples, including the experiments conducted and results achieved are provided for illustrative purposes only and are not to be construed as limiting upon the teachings herein.

EXAMPLE 1

Immunization and TITERING

Immunization

[0146] Recombinant human IGF-I and IGF-II obtained from R&D Systems, Inc. (Minneapolis, MN Cat. No. 291-G1 and 292-G2 respectively) were used as antigens. Monoclonal antibodies against IGF-I/II were developed by sequentially immunizing XenoMouse® mice (XenoMouse strains XMG2 and XMG4 (3C-1 strain), Abgenix, Inc. Fremont, CA). XenoMouse animals were immunized via footpad route for all injections. The total volume of each injection was 50 μl per mouse, 25 μl per footpad. A total of ten (10) mice were immunized in each group. Each injection was with 10 μg per mouse of IGF-I or IGF-II alone or conjugated to Keyhole Limpet Hemocyanin (KLH) antigen as a carrier, as detailed in Table 2. The first injection was made up in Dulbecco's PBS (DPBS) and admixed 1:1 v/v with Titermax Gold Adjuvant (SIGMA Cat. #T2684, lot #K1599). A total of 8 to 11 additional boosts were then administered over a period of 27 to 38 days, admixed with 25 μg of Adju-Phos (aluminum phosphate gel, Catalog # 1452-250, batch #8937, HCI Biosector) and 10 μg CpG (15 μl of ImmunEasy Mouse Adjuvant, catalog # 303101; lot #11553042; Qiagen) per mouse, followed by a final boost of 10 μg of antigen in pyrogen-free DPBS, without adjuvant. For combined immunization (animals immunized with both IGF-I and IGF-II), the second antigen was given in the last two (2) boosts.

TABLE 2. IMMUNIZATION SUMMARY

Immunization Group	Initial Immunogen	Final Immunogen	KLH Conjugated	Isotype of Mice	Fusion Group
1	IGF-1	IGF-1	–	IgG2-κλ	1
3	IGF-1	IGF-1	–	IgG4-κλ	1
5	IGF-1	IGF-1	+	IgG2-κλ	1
7	IGF-1	IGF-1	+	IgG4-κλ	1
2	IGF-2	IGF-2	–	IgG2-κλ	2
4	IGF-2	IGF-2	–	IgG4-κλ	2
6	IGF-2	IGF-2	+	IgG2-κλ	2
8	IGF-2	IGF-2	+	IgG4-κλ	2
9	IGF-1	IGF-2	–	IgG2-κλ	3
11	IGF-1	IGF-2	–	IgG4-κλ	3
13	IGF-1	IGF-2	+	IgG2-κλ	3
15	IGF-1	IGF-2	+	IgG4-κλ	3
10	IGF-2	IGF-1	–	IgG2-κλ	4
12	IGF-2	IGF-1	–	IgG4-κλ	4

14	IGF-2	IGF-1	+	IgG2-κλ	4
16	IGF-2	IGF-1	+	IgG4-κλ	4

EXAMPLE 2

RECOVERY OF LYMPHOCYTES, B-CELL ISOLATIONS, FUSIONS AND GENERATION OF HYBRIDOMAS

[0147] Immunized mice were sacrificed by cervical dislocation, and the draining lymph nodes harvested and pooled from each cohort. The lymphoid cells were dissociated by grinding in DMEM to release the cells from the tissues and the cells were suspended in DMEM. The cells were counted, and 0.9 ml DMEM per 100 million lymphocytes added to the cell pellet to resuspend the cells gently but completely. Using 100 µl of CD90+ magnetic beads per 100 million cells, the cells were labeled by incubating the cells with the magnetic beads at 4°C for 15 minutes. The magnetically labeled cell suspension containing up to 10^8 positive cells (or up to $2x10^9$ total cells) was loaded onto a LS+ column and the column washed with DMEM. The total effluent was collected as the CD90-negative fraction (most of these cells were expected to be B cells).

[0148] The fusion was performed by mixing washed enriched B cells from above and nonsecretory myeloma P3X63Ag8.653 cells purchased from ATCC, cat.# CRL 1580 (Kearney et al, J. Immunol. 123, 1979, 1548-1550) at a ratio of 1:1. The cell mixture was gently pelleted by centrifugation at 800 x g. After complete removal of the supernatant, the cells were treated with 2-4 mL of Pronase solution (CalBiochem, cat. # 53702; 0.5 mg/ml in PBS) for no more than 2 minutes. Then 3–5 ml of FBS was added to stop the enzyme activity and the suspension was adjusted to 40 ml total volume using electro cell fusion solution, (ECFS, 0.3M Sucrose, Sigma, Cat# S7903, 0.1mM Magnesium Acetate, Sigma, Cat# M2545, 0.1mM Calcium Acetate, Sigma, Cat# C4705). The supernatant was removed after centrifugation and the cells were resuspended in 40 ml ECFS. This wash step was repeated and the cells again were resuspended in ECFS to a concentration of $2x10^6$ cells/ml.

[0149] Electro-cell fusion was performed using a fusion generator (model ECM2001, Genetronic, Inc., San Diego, CA). The fusion chamber size used was 2.0 ml, using the following instrument settings:

[0150] Alignment condition: voltage: 50 V, time: 50 sec.

[0151] Membrane breaking at: voltage: 3000 V, time: 30 µsec

[0152] Post-fusion holding time: 3 sec

[0153] After ECF, the cell suspensions were carefully removed from the fusion chamber under sterile conditions and transferred into a sterile tube containing the same volume of Hybridoma Culture Medium (DMEM, JRH Biosciences), 15 % FBS (Hyclone), supplemented with L-glutamine, pen/strep, OPI (oxaloacetate, pyruvate, bovine insulin) (all from Sigma) and IL-6 (Boehringer Mannheim). The cells were incubated for 15-30 minutes at 37°C, and then centrifuged at 400 x g (1000 rpm) for five minutes. The cells were gently resuspended in a small volume of Hybridoma Selection Medium (Hybridoma Culture Medium supplemented with 0.5x HA (Sigma, cat. # A9666)), and the volume adjusted appropriately with more Hybridoma Selection Medium, based on a final plating of $5x10^6$ B cells total per 96-well plate and 200 µl per well. The cells were mixed gently and pipetted into 96-well plates and allowed to grow. On day 7 or 10, one-half the medium was removed, and the cells re-fed with Hybridoma Selection Medium.

EXAMPLE 3

SELECTION OF CANDIDATE ANTIBODIES BY ELISA

[0154] After 14 days of culture, hybridoma supernatants were screened for IGF-I/II-specific monoclonal antibodies. The ELISA plates (Fisher, Cat. No. 12-565-136) were coated with 50 µl/well of human IGF-I or IGF-II (2 µg/ml) in Coating Buffer (0.1 M Carbonate Buffer, pH 9.6, $NaHCO_3$ 8.4 g/L), then incubated at 4°C overnight. After incubation, the plates were washed with Washing Buffer (0.05% Tween 20 in PBS) 3 times. 200 µl/well Blocking Buffer (0.5% BSA, 0.1% Tween 20, 0.01% Thimerosal in 1x PBS) were added and the plates incubated at room temperature for 1 hour. After incubation, the plates were washed with Washing Buffer three times. 50 µl/well of hybridoma supernatants, and positive and negative controls were added and the plates incubated at room temperature for 2 hours.

[0155] After incubation, the plates were washed three times with Washing Buffer. 100 µl/well of detection antibody goat anti-huIgGFc-HRP (Caltag, Cat. No. H10507), was added and the plates incubated at room temperature for 1 hour. In a secondary screen, the positives in first screening were screened in two sets, one for human IgG (heavy chain) detection and the other for human Ig kappa light chain detection (goat anti-hIg kappa-HRP (Southern Biotechnology, Cat. No. 2060-05) in order to demonstrate fully human composition for both IgG and Ig kappa. After incubation, the

plates were washed three times with Washing Buffer. 100 µl/well of TMB (BioFX Lab. Cat. No. TMSK-0100-01) were added and the plates allowed to develop for about 10 minutes (until negative control wells barely started to show color). 50 µl/well stop solution (TMB Stop Solution, (BioFX Lab. Cat. No. STPR-0100-01) was then added and the plates read on an ELISA plate reader at 450nm. As indicated in Table 3, there were a total of 1,233 wells containing antibodies against IGF-I and -II.

[0156] All antibodies that bound in the ELISA assay were counter screened for binding to insulin by ELISA in order to exclude those that cross-reacted with insulin. The ELISA plates (Fisher, Cat. No. 12-565-136) were coated with 50 µl/well of recombinant insulin (concentration: 1µg/ml; Sigma, catalog # I2643) in Coating Buffer (0.1 M Carbonate Buffer, pH 9.6, $NaHCO_3$ 8.4 g/L), then incubated at 4°C overnight. As detailed in Table 3, a total of 1,122 antibodies from the original 1233 antibodies did cross react with insulin.

TABLE 3. SCREENING SUMMARY

			Confirmation Screen			
Fn #	**Mouse strain**	**Immunogen**	**Target with hG detection**	**Target with hK+hL detection**	**IGF-I(+) & IGF-II(+)**	**IGF-I(+) & IGF-II(+) and hu-Insulin (-)**
1	G2 KL	IGF-I-KLH	IGF-II	IGF-I	36	28
2	G4 KL	IGF-I-KLH	IGF-II	IGF-I	65	55
3	G2 KL	IGF-II or IGF-II-KLH	IGF-I	IGF-II	168	150
4	G4 KL	IGF-II or IGF-II-KLH	IGF-I	IGF-II	197	194
5	G2 KL	IGF-I/-II-KLH	IGF-II	IGF-II	54	50
6	G4 KL	IGF-I/-II-KLH	IGF-II	IGF-II	101	86
7	G2 KL	IGF-II/-I-KLH	IGF-II	IGF-II	294	271
8	G4 KL	IGF-II/-I or IGF-II/-I-KLH	IGF-II	IGF-II	318	288
		Total F1 to F4			466	427
		Total F5 to F8			767	695
		Total			**1,233**	**1,122**

[0157] Finally, the antibodies that were selected in the counter-screen were then tested by ELISA to confirm binding to mouse IGF-I and IGF-II proteins. A total of 683 hybridoma lines .were identified that have cross-reactivity with mouse IGF-I/II. Accordingly, these hybridoma lines expressed antibodies that bound to human IGF-I, human IGF-II, mouse IGF-I and mouse IGF-II, but did not bind to human insulin.

EXAMPLE 4

INHIBITION OF IGF-I AND IGF-II BINDING TO IGF-IR

[0158] The purpose of this study was to screen the 683 anti-IGF-I/II human IgG2 and IgG4 antibodies at the hybridoma supernatant stage for neutralizing activity, as determined by inhibition of IGF-I and IGF-II binding to the IGF-IR receptor. Thus, a receptor/ligand binding assay was performed with NIH3T3 cells that overexpress the human IGF-IR receptor, as described below.

[0159] Briefly, multi-screen filter plates (MultiScreen 0.65 μM 96-well PVDF, Millipore, Cat. No. MADV N0B 10) were blocked with blocking buffer (PBS containing 10%BSA with 0.02%NaN_3) at 200μL/well overnight at 4°C. [^{125}I]-labeled IGF (Amersham Life Sciences Cat No. IM172 (IGF-I) or IM238 (IGF-II)) at 100μCi/ml and 50nM was diluted to the appropriate concentration (70pM final for IGF-I and 200pM final for IGF-II) in binding buffer (PBS containing 2%BSA with 0.02% NaN_3). The blocking buffer-coated filter plate was washed once with 200μL PBS, and 50μL anti-IGF-I/II Ab supernatants (diluted in binding buffer to 25% final volume) were preincubated with 25μL of [^{125}I]-IGF in the MultiScreen plate for 30-60 minutes on ice. Subconfluent NIH3T3 mouse fibroblasts stably expressing hIGF-IR (obtained from AstraZeneca) were harvested with trypsin and resuspended in cold binding buffer at $6x10^6$/ml, and 25μL of cells were added to the plate for a two-hour incubation on ice. The plate was washed four times with 200μL cold PBS and dried overnight. Twenty-five μL/well of scintillant (SuperMix cocktail, Wallac/Perkin Elmer Cat No. 1200-439) was added and the plates were read using a Microbeta Trilux reader (Wallac).

[0160] The following controls were used per screening plate: no antibody (total IGF bound), control neutralizing anti-IGF-I (#05-172, Upstate) or anti-IGF-II (#MAB292, R&D Systems) mAbs at 50μg/ml (non-specific background) and 0.075 to 0.5μg/ml (approximate EC50 values of the neutralizing antibodies), and isotype-matched control human IgG2 (PK16.3.1, Abgenix, lot #360-154) or IgG4 (108.2.1, Abgenix,

lot#718-53A) mAbs at a concentration of 0.5μg/ml (approximate EC50 value of neutralizing antibodies). An additional titration of control neutralizing antibodies and isotype-matched control human antibodies was added to one plate per screening assay (1/10 serial dilution from 50ug/ml (333.3nM)). All controls with or without antibodies were prepared in binding buffer supplemented with anti-KLH human IgG2 or IgG4 exhaust supernatant at 25% final volume.

[0161] The percentage of inhibition was determined as follows:

% Inhibition= ([(Mean CPM Total ^{125}I-IGF bound)- (Mean CPM ^{125}I-IGF bound in the presence of antibody)] / [(Mean CPM Total ^{125}I-IGF bound)-(Mean CPM ^{125}I-IGF bound in the presence of an excess of control neutralizing antibody*)]) x100

[0162] * the non-specific background was determined as CPM of cells with an excess of control neutralizing anti-IGF Ab (50ug/ml, 333.3nM), which was found to be equivalent to an excess of cold IGF (less than or equal to 10% of total CPM)

[0163] The anti-IGF-I/II supernatant screening was split by isotype because of radiolabeled ligand availability issues. As shown in Table 4, supernatants from the anti-IGF-I/II antibodies with an IgG2 isotype (293 total) were first screened against radiolabeled IGF-I. A cut-off at 40% inhibition was initially applied to this screening (i.e. hybridoma lines inhibiting at 40% and above were selected), and 111 hits were selected for subsequent screening against IGF-II. Of the 111 hits, a total of 91 lines were found to inhibit IGF-II binding to its receptor with a 50% cut-off. A total of 71 final hits were selected by taking supernatants that neutralized 50% of both IGF-I and IGF-II activity.

[0164] All the supernatants expressing IgG4 isotypes (390 total) were initially screened against radiolabeled IGF-II, and 232 hits with a cut-off at 50% inhibition were subsequently screened against IGF-I. A total of 90 lines were able to inhibit IGF-I binding to its receptor with a 50% cut-off. After combining the hits for IgG2 (71) and IgG4 (90), a total of 161 lines were obtained which inhibited IGF-I and IGF-II by 50% or more.

[0165] In conclusion, from the 683 original supernatants, 343 (111 IgG2 and 232 IgG4, 50.2%) were selected from the first screening with either IGF-I or IGF-II. A total of 161 final hits were obtained (23.6% of original lines), which are able to block

both IGF-I and IGF-II binding to IGF-IR with an overall cut-off criteria of 50% inhibition.

TABLE 4. ANTI-IGF-I/II EXHAUST SUPERNATANT SCREENING SUMMARY (50% CUT-OFF)

			ESN Activity								
Fusion #	**Mouse strain**	**Immunogen**	**hIGF-II with hG^+/hK^+ or hG^+/hL^+**	**R/L Binding Assay IGF-I**	**IGF-I+ % Total**	**R/L Binding Assay IGF-II on IGF-I+**	**IGF-I/II+ % Total**	**R/L Binding Assay IGF-II**	**IGF-II+ % Total**	**R/L Binding Assay IGF-I on IGF-II+**	**IGF-I/II+ % Total**
1	G2 KL	IGF-I-KLH	14	12	85.7	1	7.1				
2	G4 KL	IGF-I-KLH	20					4	20.0	4	20.0
3	G2 KL	IGF-II or IGF-II-KLH	49	13	26.5	4	8.2				
4	G4 KL	IGF-II or IGF-II-KLH	114					86	75.4	36	31.6
5	G2 KL	IGF-I/-II-KLH	32	11	34.4	7	21.9				
6	G4 KL	IGF-I/-II-KLH	47					21	44.7	8	17.0
7	G2 KL	IGF-II/-I-KLH	198	75	37.9	59	29.8				
8	G4 KL	IGF-II/-I or IGF-II/-I-KLH	209					121	57.9	42	20.1
			683	111	16.3	**71**	10.4	232	34.0	**90**	13.2
				Cut-off 40%	**IgG2**				**IgG4**		

EXAMPLE 5

HIGH ANTIGEN AND LIMITED ANTIGEN ELISAS

[0166] In order to determine the relative affinities among the 161 hybridoma lines selected in Example 4, as well as the concentration of antibody in the supernatants of each line, high antigen (HA) and limited antigen (LA) ELISA assays were carried out. In the HA quantitation assay, the high antigen concentration and overnight incubation limit the effect of antibody affinity, allowing for quantitation of the relative amount of antigen-specific antibody present in each sample. The low antigen concentration in the LA assay limits the effect of antibody concentration and results in a ranking of antibodies based on their relative affinity.

High Antigen Quantitation Assay

[0167] ELISA plates were coated with relatively large amounts of either IGF-I or IGF-II antigen (R&D Systems, Inc., Minneapolis, MN Cat. No. 291-G1 and 292-G2 respectively) at 500ng/ml (67nM). Antibody-containing hybridoma supernatants were titrated over a dilution range of 1:50 to 1:12200. A control of a known IGF-specific antibody (R&D Systems, Inc., Minneapolis, MN Cat. No. MAB291 and MAB292 respectively) was used to define the linear range of the assay. Data within the linear range were then used to derive the relative concentration of the IGF-specific antibody in each titrated sample.

Limited Antigen Assay

[0168] Microtiter plates were coated with low concentrations of antigen. Fifty microliters (50 µL) of IGF-I or IGF-II at 64, 32, 16, 8, 4, and 2 ng/ml (covering a range of 8.5 nM to 0.26 nM) in 1% skim milk / 1X PBS pH 7.4 / 0.5% azide was added to each well. The plate was incubated for 30 minutes.

[0169] Plates were washed four times (4X) with water, and 50µL of hybridoma supernatant containing test antibodies diluted 1:25 in 1% skim milk / 1X PBS pH 7.4 / 0.5% azide were added to the wells. Plates were wrapped tightly with plastic wrap or paraffin film, and incubated overnight with shaking at room temperature.

[0170] On the following day, all plates were washed five times (5X) and 50 µL goat anti-Human IgG Fc HRP polyclonal antibody at a concentration of 0.5 ug/ml in 1%

milk, 1X PBS pH 7.4 was added to each well. The plates were incubated for 1 hour at room temperature.

[0171] Plates were washed at least five times (5X with tap water). Fifty microliters (50) µL of HPR substrate TMB was added to each well, and the plate were incubated for 30 minutes. The HRP-TMB reaction was stopped by adding 50 µL of 1M phosphoric acid to each well. Optical density (absorbance) at 450 nm was measured for each well of the plate.

Data Analysis

[0172] OD values of test antibodies were averaged and the range was calculated. Antibodies with the highest signal and acceptably low standard deviation were selected as antibodies having a higher affinity for the antigen than did a reference antibody.

[0173] An analysis was then made to select top antibodies based on either neutralization (Example 4), potency (low antibody concentration as determined by HA ELISA and high inhibition of ligand binding), affinity (LA ELISA), or all three criteria. From this analysis, a list of 25 antibodies was generated. A separate analysis based on average % inhibition of IGF-I and -II binding and affinity for both IGF-I and IGF-II generated a second list of 25 antibodies. Sixteen antibodies were common to both lists, resulting in a final list of 40 antibodies. The LA and HA results for these 40 antibodies are summarized in Table 5. These 40 lines were selected for cloning, of which 33 were successfully cloned.

TABLE 5. RESULTS OF HIGH AND LIMITED ANTIGEN ELISA FOR TOP 40 ANTIBODIES

	IGF1 HA		IGF2 HA		IGF1 LA	IGF2 LA	IGF1 LA	IGF2 LA
Line ID	Avg (ug/ml) HA1	Std. Dev HA1	Avg (ug/ml) HA2	Std. Dev HA2	IGF1 2ng/ml	IGF2 2ng/ml	IGF1 4ng/ml	IGF2 4ng/ml
4.121	1.16	0.16	3.56	1.34	0.56	0.53	1.14	0.90
4.141	1.99	0.22	1.99	0.21	0.57	0.79	1.25	1.52
4.142	4.70	0.21	3.65	0.31	0.78	1.00	1.76	1.78
4.143	1.74	0.17	2.03	0.41	0.60	0.99	1.20	1.91
4.25	1.26	0.23	1.48	0.36	0.71	0.81	1.31	1.54

	IGF1 HA		IGF2 HA		IGF1 LA	IGF2 LA	IGF1 LA	IGF2 LA
Line ID	Avg (ug/ml) HA1	Std. Dev HA1	Avg (ug/ml) HA2	Std. Dev HA2	IGF1 2ng/ml	IGF2 2ng/ml	IGF1 4ng/ml	IGF2 4ng/ml
4.69	7.17	1.16	6.50	0.53	0.80	0.80	1.50	1.54
4.90	1.15	0.12	3.68	0.77	0.58	0.48	1.04	0.89
7.118	10.34	1.26	10.32	1.90	0.81	0.85	1.56	1.44
7.123	13.4	3.2	12.0	2.8	1.0	1.7	1.66	2.58
7.127	7.28	0.44	6.59	1.55	1.19	1.37	2.29	2.40
7.130	4.32	0.32	3.51	1.34	0.64	1.17	1.36	1.98
7.146	12.04	0.98	9.63	0.6	0.2	0.86	0.29	1.58
7.158	9.29	0.49	7.1	0.56	1.71	1.42	3.00	2.46
7.159	16.53	1.83	41.1		0.56	1.65	0.98	2.47
7.160	4.9	0.5	5.1	0.2	1.7	2.2	3.14	3.30
7.175	8.46	0.49	6.21	1.22	0.13	0.34	0.14	0.62
7.202	11.94	1.98	15.24	1.72	1.11	1.68	2.28	2.69
7.212	11.30	1.90	10.86	1.26	0.97	0.93	2.22	1.54
7.215	10.11	2.05	10.94	1.39	1.01	1.09	2.25	1.93
7.23	4.30	0.26	3.99	0.29	0.55	1.22	1.40	2.17
7.234	4.7	1.4	3.1	0.4	0.7	1.4	1.79	2.44
7.251	3	0.41	1.93	0.17	1.09	1.02	1.31	1.53
7.252	8.25	0.51	5.55	1.68	1.22	1.23	2.53	2.09
7.268	7.58	0.42	5.07	0.92	1.47	1.37	3.06	2.42
7.29	12.5	1.24	23.53	4.7	0.18	0.39	0.27	0.49
7.3	13.18	2.12	8.83	0.58	0.81	1.28	2.07	1.96
7.34	12.54	1.99	14.67	3.05	0.21	1.07	0.44	1.84
7.41	3.69	0.19	4.97	0.8	1.02	1.53	2.21	2.32
7.56	14.6	2.0	21.7	3.7	1.3	1.4	2.38	2.46
7.58	17.52	0.01	27.54	6.22	0.21	1.15	0.47	1.82
7.66	6.02	0.81	6.18	0.71	0.49	0.97	1.42	1.53
7.77	8.42	0.18	7.25	1.12	0.64	0.46	1.50	1.00
7.85	22.67	0.68	23.63	0.93	0.1	0.33	0.16	0.51
7.99	7.9	0.2	5.9	1.6	0.9	1.0	1.87	1.65
8.119	1.26	0.00	0.77	0.16	2.37	0.79	3.77	1.36
8.141	5.96	0.50	4.12	0.61	1.80	0.61	3.02	1.08
8.146	4.03	0.45	2.55	0.74	0.97	1.06	2.13	1.98
8.287	4.8	0.1	2.8	0.8	2.2	1.6	3.80	2.91
8.8	2.00	0.17	1.45	0.25	1.72	0.77	3.13	1.46
8.86	3.15	0.19	2.36	0.24	0.92	1.35	1.76	2.18

EXAMPLE 6

BINDING OF ANTIBODIES TO IGF-I AND IGF-II BOUND TO IGFBP-3

[0174] IGF-I and –II circulate in serum mostly bound to IGF-binding proteins (IGFBPs). One aim was to identify antibodies that do not recognize IGFs in complex with IGFBPs, in order to avoid *in vivo* depletion of anti-IGF antibodies. The following assay format was developed for the characterization of antibodies that recognize IGF-I or IGF-II when these growth factors are complexed with IGFBP-3. Specifically, this assay tested the ability of IGF in IGF/anti-IGF antibody complexes to bind IGFBP-3.

Antibody-Mediated Block of Capture of IGF by IGFBP-3

[0175] An assay was developed wherein complexes were pre-formed between IGF-I or IGF-II and IGF-specific antibodies from the aforementioned examples. The ability of these complexes to bind to IGFBP-3 was tested using AlphaScreen assay technology (PerkinElmer). In a 384-well plate, 10 µL 1:20 diluted hybridoma supernatants were mixed with 10 µL of 3 nM biotinylated IGF-I or IGF-II and incubated at room temperature for 2 hours. Streptavidin-coated AlphaScreen donor beads and IGFBP-3-coupled AlphaScreen acceptor beads (10 uL of a mixture, for a 1/60 final dilution of the hybridoma supernatants) were added, and the incubation was continued for another hour. Samples were then read in a Packard Fusion plate reader.

[0176] Three commercially available anti-IGF monoclonal antibodies M23 (Cell Sciences), 05-172 (Upstate) and MAB291 (R&D Systems) showed different abilities to inhibit IGF binding to IGFBP with IC50 values ranging from low ng/mL to 100 ng/mL. No inhibition of IGF-I binding to the IGFBP-3 was observed with irrelevant mouse IgG and human IgG up to 10 µg/mL, suggesting that the anti-IGF-I effect is specific. Commercially available monoclonal antibodies 05-166 (Upstate) and MAB292 (R&D) showed a significant difference in affinity for inhibition of IGF-II / IGFBP-3 interactions. These experiments show that anti-IGF mAbs can block the binding of IGF to IGFBP-3, giving an assay that could be used for screening purified antibodies from hybridoma lines. The next step was to evaluate the effects of exhausted hybridoma medium on the assay signal.

[0177] Serial dilutions of the hybridoma medium and anti-KLH hybridoma exhaust supernatants were tested in the assay system. When hybridoma supernatants were diluted 1:10 in preparation for preincubation with IGFI/II (final dilution in the assay was

1:60), there was almost no effect of the medium on the assay results. Based on these data, hybridoma supernatants were diluted for preincubation with IGF, providing the preferred 1/60 dilution final dilution in the assay.

[0178] Six hundred eighty-three exhaust supernatants positive for IGF-I and IGF-II binding were examined for their ability to inhibit binding of IGF to IGFBP-3. Inhibition above 50% for IGF-I and above 60% for IGF-II were used as cut-off criteria. The summary results of the screen using these cut-offs are shown in Table 6.

TABLE 6. NUMBERS OF POSITIVE HITS IDENTIFIED IN THE SCREEN

		IGF-I	IGF-II	IGF-I/II
Samples	Inhibition->	>50%	>60%	
376	(plates 1-4)	48	51	19
307	(plates 5-8)	39	78	32
683	Total	87	129	51

[0179] The IGFBP competition assay using the AlphaScreen assay identified 87 samples inhibiting IGF-I binding to IGFBP-3 and 129 samples inhibiting IGF-II binding to IGFBP-3 among 683 tested supernatants. Fifty-one samples demonstrated dual competition of IGF-I and IGF-II. However, in order to more carefully reproduce the function or behavior of the antibodies *in vivo*, where the IGF and the IGFBP complex would be largely preformed, additional assays, as described in example 8 were performed.

EXAMPLE 7

DETERMINATION OF ANTI-IGF-I AND IGF-II ANTIBODY AFFINITY USING BIACORE ANALYSIS (LOW RESOLUTION SCREEN)

Low Resolution Screen of 34 Purified Monoclonal Antibodies

[0180] The label-free surface plasmon resonance (SPR), or Biacore, was utilized to measure the antibody affinity to the antigen. For this purpose, a high-density goat anti-human antibody surface over a CM5 Biacore chip was prepared using routine amine coupling. All the mAbs were diluted to approximately 20 μg/ml in HBS-P running buffer containing 100 μg/ml BSA. Each mAb was captured on a separate surface using a 30-second contact time at 10 μL/min., and a 5-minute wash for stabilization of the mAb baseline.

[0181] IGF-I was injected at 335.3 nM over all surfaces at 23°C for 120 seconds, followed by a 5-minute dissociation, using a flow rate of 100 μL/min. The samples were

prepared in the HBS-P running buffer described above. The surfaces were regenerated after every capture/injection cycle with one 15-second pulse of 146mM phosphoric acid (pH 1.5). The same capture/injection cycles were repeated for each antibody with 114.7 nM IGF-II. Drift-corrected binding data for the 34 mAbs was prepared by subtracting the signal from a control flow cell and subtracting the baseline drift of a buffer injected just prior to each antigen injection. Data were fit globally to a 1:1 interaction model using CLAMP to determine the binding kinetics (David G. Myszka and Thomas Morton (1998) "CLAMP©: a biosensor kinetic data analysis program," TIBS 23, 149-150). A mass transport coefficient was used in fitting the data. The kinetic analysis results of IGF-I and IGF-II binding at 25°C are listed in Table 7 below. The mAbs are ranked from highest to lowest affinity.

TABLE 7. IGF-I AND IGF-II LOW RESOLUTION BIACORE SCREEN OF 34 MONOCLONAL ANTIBODIES

	IGF-II			IGF-I		
Sample	k_a ($M^{-1}s^{-1}$)	k_d (s^{-1})	K_D (pM)	k_a ($M^{-1}s^{-1}$)	k_d (s^{-1})	K_D (pM)
7.159.2	3.5×10^6	1.0×10^{-5}*	2.9	4.3×10^6	9.3×10^{-4}	216.0
8.86.1	6.1×10^6	2.4×10^{-4}	39.3	3.4×10^6	1.3×10^{-2}	3823.0
4.25.1	9.3×10^6	4.1×10^{-4}	44.1	5.1×10^6	6.9×10^{-3}	1353.0
7.234.2	6.4×10^6	2.9×10^{-4}	45.3	6.7×10^6	2.2×10^{-3}	328.0
7.160.2	4.6×10^6	2.5×10^{-4}	54.3	5.6×10^6	3.3×10^{-3}	589.0
7.146.3	3.2×10^6	1.8×10^{-4}	56.2	3.8×10^6	8.7×10^{-3}	2289.0
7.34.1	3.0×10^6	1.8×10^{-4}	60.0	5.2×10^6	3.2×10^{-3}	615.0
7.123.1	4.0×10^6	3.4×10^{-4}	85.0	4.3×10^6	9.0×10^{-3}	2093.0
7.202.3	1.8×10^6	1.7×10^{-4}	94.4	1.2×10^6	5.4×10^{-3}	4500.0
4.141.1	4.9×10^6	4.7×10^{-4}	95.9	*	*	*
7.215.2	3.2×10^6	3.3×10^{-4}	103.0	3.6×10^6	2.6×10^{-2}	7222.0
8.287.2	2.4×10^6	2.5×10^{-4}	104.0	7.7×10^5	1.3×10^{-3}	1688.0
8.146.2	7.7×10^6	8.0×10^{-4}	104.0	2.4×10^6	1.3×10^{-2}	5417.0
4.143.2	1.1×10^7	1.2×10^{-3}	109.0	6.4×10^6	1.9×10^{-2}	2969.0
7.251.3	3.5×10^6	4.3×10^{-4}	123.0	4.6×10^6	4.3×10^{-3}	935.0
7.99.1	6.9×10^6	9.9×10^{-4}	143.0	6.0×10^6	4.8×10^{-3}	800.0

	IGF-II			IGF-I		
Sample	k_a ($M^{-1}s^{-1}$)	k_d (s^{-1})	K_D (pM)	k_a ($M^{-1}s^{-1}$)	k_d (s^{-1})	K_D (pM)
4.142.2	5.3×10^6	8.5×10^{-4}	160.0	8.5×10^6	1.9×10^{-2}	2235.0
7.41.3	3.2×10^6	5.5×10^{-4}	172.0	5.2×10^6	2.9×10^{-3}	558.0
7.56.3	3.3×10^6	6.0×10^{-4}	182.0	4.8×10^6	3.1×10^{-3}	646.0
7.127.1	4.1×10^6	7.6×10^{-4}	185.0	4.9×10^6	3.5×10^{-3}	714.0
8.8.3	4.0×10^6	7.8×10^{-4}	195.0	3.1×10^6	8.2×10^{-4}	264.0
7.158.2	3.4×10^6	6.7×10^{-4}	197.0	4.4×10^6	2.2×10^{-3}	500.0
7.23.3	3.3×10^6	6.5×10^{-4}	197.0	4.1×10^6	5.8×10^{-3}	1415.0
7.252.1	3.2×10^6	6.5×10^{-4}	203.0	2.3×10^6	2.1×10^{-3}	913.0
7.66.1	3.6×10^6	7.7×10^{-4}	214.0	2.0×10^6	2.5×10^{-3}	1250.0
7.130.1	4.2×10^6	1.0×10^{-3}	238.0	4.6×10^6	1.8×10^{-2}	3913.0
4.90.2	4.9×10^6	1.2×10^{-3}	245.0	*	*	*
7.3.3	3.2×10^6	8.8×10^{-4}	275.0	4.1×10^6	3.9×10^{-3}	951.0
7.118.1	4.6×10^6	1.5×10^{-3}	326.0	5.6×10^6	5.1×10^{-3}	911.0
7.212.1	4.2×10^6	1.6×10^{-3}	381.0	3.1×10^6	6.0×10^{-3}	1935.0
7.175.2	1.3×10^6	7.4×10^{-4}	569.0	1.8×10^6	2.0×10^{-2}	11111.0
4.121.1	5.5×10^6*	3.2×10^{-3}*	582.0*	1.5×10^6	2.1×10^{-3}	1400.0
7.85.2	1.9×10^6	1.3×10^{-3}	684.0	2.2×10^6	2.9×10^{-2}	13182.0
7.58.3	8.9×10^6	7.9×10^{-3}	888.0	7.2×10^6	3.2×10^{-2}	4444.0

IGF-I Binding Data

[0182] Most mAbs fit a 1:1 model reasonably well. MAbs 4.90.2 and 4.141.1 were characterized by extremely complex data. These mAbs were listed with an asterisk in Table 7 because no meaningful kinetic constants could be estimated from the 1:1 model fit. The latter off-rate phase appears to be very slow for both of these mAbs (at least 1×10^{-5} sec^{-1}), which might make these two mAbs useful as therapeutic compounds.

IGF-II Binding Data

[0183] Most mAbs fit a 1:1 model reasonably well. The off-rate for mAb 7.159.2 was held constant at 1 X 10^{-5} sec^{-1} because there was not enough decay data to adequately estimate k_d.

[0184] The low-resolution Biacore studies in this example are designed as a semi-quantitative ranking approach. In order to acquire more accurate information regarding the characteristic rate constants and affinities of individual mAbs, high-resolution Biacore studies were carried out as described in Example 8.

EXAMPLE 8

DETERMINATION OF ANTI-IGF-I AND IGF-II ANTIBODY AFFINITY USING BIACORE ANALYSIS (HIGH RESOLUTION SCREEN)

[0185] A high resolution Biacore analysis was performed to further measure the antibody affinity to the antigen. mAbs 7.159.2, 7.234.2, 7.34.1, 7.251.3, and 7.160.2 were each captured and the IGF-I and IGF-II antigens were each injected over a range of concentrations. The resulting binding constants are listed in Table 8.

TABLE 8. ANTI-IGF ANTIBODY AFFINITY DETERMINED BY LOW-AND HIGH-RESOLUTION BIACORE ANALYSIS

mAb	Low resolution K_D (pM)		High Resolution K_D (pM)	
	IGF-I	IGF-II	IGF-I	IGF-II
7.159.2	216.0	2.9	294.0	1.9
7.234.2	328.0	45.3	3760.0	295.0
7.34.1	615.0	60.0	436.0 421.0	164.0 162.0
7.251.3	935.0	123.0	452.0	47.4
7.160.2	589.0	54.3	2800.0	237.0

[0186] Thus, embodiments of the invention can include an antibody that will preferentially bind to IGF-II, but that will cross-react with IGF-I, binding to IGF-II with higher affinity than to IGF-I. For example, the antibody can bind to IGF-II with 2.5 times greater affinity than to IGF-I. In certain embodiments, the antibody can bind to IGF-II with at least 5, at least 10, at least 25, at least 50 or at least 150 times greater affinity than to IGF-I.

Screening of Preformed IGF-I/GFBP-3 Complexes

[0187] The IGFBP competition assay described in Example 6 identified 87 samples inhibiting IGF-I binding to IGFBP-3 and 129 samples inhibiting IGF-II binding to IGFBP-3 among 683 tested supernatants. Fifty-one samples demonstrated dual competition of IGF-I and IGF-II. However, in order to more carefully reproduce the function or behavior of the antibodies *in vivo*, where the IGF and the IGFBP complex would be largely preformed, the following Biacore assays were performed on selected antibodies.

[0188] Six selected antibodies were screened to determine whether they bind IGF-I or IGF-II in complex with IGFBP. All six of the selected mAbs (7.159.2, 7.146.3, 7.34.1, 7.251.3, 7.58.3, and unrelated control antibody ABX-MA1) were covalently immobilized to a high surface capacity (5,400-12,800 RUs) on two CM5 Biacore chips using routine amine coupling with a Biacore 2000 instrument. One flow cell on each CM5 chip was activated and blocked (no mAb immobilized) for use as a control surface.

[0189] Next, IGF-I and IGFBP-3 were mixed together in Hepes buffered saline, pH 7.4, 0.005% P-20, 100 μg/ml BSA (HBS-P), to make a final solution of 193 nM and 454 nM, respectively. IGF-II and IGFBP-3 were mixed together to make a final solution of 192 nM and 455 nM, respectively. Under these conditions, IGF-I and IGF-II were 99.97% complexed by IGFBP-3. Equilibrium was reached within minutes under these conditions. Solutions of complexed IGF-I/IGFBP-3 and IGF-II/IGFBP-3 were flowed across the various mAb surfaces at 40 μL/min and 23 °C, for 180 seconds and dissociation was followed for 120 seconds. Uncomplexed IGF-I and IGF-II were then flowed across each surface at 193 nM and 192 nM, respectively, and IGFBP-3 was flowed across each surface at 454 nM. The surfaces were regenerated with a 20 second pulse of 10 mM glycine, pH 2.0.

[0190] The sensorgrams were processed using the program Scrubber by subtracting the bulk refractive index change and any nonspecific binding signal of the analyte to the blank surface from the binding signal from surfaces with mAb immobilized. After blank correction subtraction, the sensorgrams were referenced a second time by subtracting an average sensorgram for buffer injections over a specific flow cell. This "double reference" corrected the mAb binding sensorgrams for any systematic errors present on a particular flow cell.

[0191] Complexed and uncomplexed IGF-I/IGFBP-3 and IGF-II/IGFBP-3 bound fairly weakly to the bound antibodies, with a rough estimate of the nonspecific binding interaction being a K_D>1 μM for all six mAbs, including negative control ABX-MA1 (See Table 9). However, with ABX-MA1 the IGF-I/II binding was weak and indicated nonspecific binding interactions occurred with all these three analytes. Apparently, the IGF/IGFBP-3 complexes bind slightly stronger to all these mAbs than IGFBP-3 does alone. However, because both IGF-I, IGF-II and IGBP-3 appear to bind nonspecifically to these mAbs themselves, when they are both bound together, this results in an even "stickier" nonspecific binding protein complex, which explains the greater binding signal for the complex. The IGF-I/II/IGFBP-3 complexes and IGFBP-3 bound to the control surface significantly also indicating the nonspecificity of these two proteins. However, in the sensorgrams below this background binding is subtracted out in the first reference during data processing, as described above.

[0192] This experiment suggests that although 51 of the samples were previously shown to inhibit binding of IGF-I/II to IGFBP3 (Example 6), the antibodies may also bind to the IGF/IGFBP complex *in vitro*.

TABLE 9. BINDING SUMMARY FOR IGF-I/IGFBP-3 AND IGF-II/IGFBP-3 BINDING TO SIX MABS.

mAb	IGF-I/IGFBP-3 complex	IGF-II/IGFBP-3 complex	IGFBP-3	IGF-I (or II)
7.159.2	+	+	+	+++
7.146.3	+	++	+	+++
7.34.1	+	++	+	+++
7.251.3	++	++	++	+++
7.58.3	++	++	++	+++
ABX-MA1	+	+	+	+

+, slight binding relative to IGF-I or IGF-II to the mAb
++, medium binding relative to IGF-I or IGF-II to the mAb
+++, strong binding relative to IGF-I or IGF-II binding to the mAb
*These ratings DO NOT indicate the K_D for these interactions.

EXAMPLE 9

DETERMINATION OF ANTI-INSULIN ANTIBODY AFFINITY USING BIACORE ANALYSIS (LOW RESOLUTION SCREEN)

[0193] The cross-reactivity of antibodies to IGF-I/II was further investigated by measuring the affinity of the mAbs to human insulin. IGF-I/II antibodies were immobilized to the CM5 Biacore chips, and insulin in solution was injected for the determination of the on-rate and off-rate. Five mAbs, including 7.234.2, 7.34.1, 7.159.2, 7.160.2, and 7.251.3, were tested in this experiment. Insulin diluted to 502 nM in the running buffer was injected over all capture surfaces.

[0194] No insulin binding to any of the mAbs was observed at 502 nM insulin. These results suggest that there is no apparent cross-reactivity of the IGF-I/II mAbs with insulin.

EXAMPLE 10

BINNING OF ANTIBODIES

[0195] Epitope binning was performed to determine which of the anti-IGF-I/II antibodies would cross compete with one another, and thus were likely to bind to the same epitope on IGF-I/II. The binning process is described in U.S. Patent Application

20030175760, also described in Jia et al., J. Immunol. Methods, (2004) 288:91-98, both of which are incorporated by reference in entirety. Briefly, Luminex beads were coupled with mouse anti-huIgG (Pharmingen #555784) following the protein coupling protocol provided on the Luminex website. Pre-coupled beads were prepared for coupling to primary unknown antibody using the following procedure, protecting the beads from light. Individual tubes were used for each unknown supernatant. The volume of supernatant needed was calculated using the following formula: (nX2+10) x 50 µl (where n = total number of samples). A concentration of 0.1 µg/ml was used in this assay. The bead stock was gently vortexed, and diluted in supernatant to a concentration of 2500 of each bead in 50 µl per well or $0.5X10^5$ beads/ml.

[0196] Samples were incubated on a shaker in the dark at room temperature overnight.

[0197] The filter plate was pre-wetted by adding 200 µl wash buffer per well, which was then aspirated. 50 µl of each bead was added to each well of the filter plate. Samples were washed once by adding 100 µl/well wash buffer and aspirating. Antigen and controls were added to the filter plate at 50 µl/well. The plate was covered, incubated in the dark for 1 hour on a shaker, and then samples were washed 3 times. A secondary unknown antibody was then added at 50 µl/well. A concentration of 0.1 µg/ml was used for the primary antibody. The plate was then incubated in the dark for 2 hours at room temperature on a shaker, and then samples were washed 3 times. 50 µl/well of biotinylated mouse anti-human IgG (Pharmingen #555785) diluted at 1:500 was added, and samples were incubated in the dark for 1 hour with shaking at room temperature.

[0198] Samples were washed 3 times. 50 µl/well Streptavidin-PE at a 1:1000 dilution was added, and samples were incubated in the dark for 15 minutes with shaking at room temperature. After running two wash cycles on the Luminex100, samples were washed 3 times. Contents in each well were resuspended in 80 µl blocking buffer. Samples were carefully mixed with pipetting several times to resuspend the beads. Samples were then analyzed on the Luminex100. Results are presented below in Table 10.

TABLE 10. BINS FOR TOP 34 IGF-I/II ANTIBODIES POSITIVE IN FUNCTIONAL ASSAY

IGF-I			
Bin 1	**Bin 2**	**Bin 3**	**No Bin**
7.3.3	7.58.3	7.175.2	7.215.2
7.23.3	8.287.2	7.85.2	
7.66.1		4.90.2	
7.56.3		4.141.1	
7.160.2		7.146.3	
7.41.3		7.34.1	
4.121.1		7.159.2	
8.146.2		7.251.3	
7.252.1			
7.123.1			
7.212.1			
7.234.2			
7.99.1			
7.127.1			
4.25.1			
8.8.3			
7.158.2			
7.202.3			
7.130.1			
8.86.1			
4.142.2			
7.118.1			
4.143.2			

IGF-II			
Bin 1	**Bin 2**	**Bin 3**	**No Bin**
7.3.3	7.158.2	7.175.2	7.215.2
7.127.1	8.146.2	4.90.2	
7.99.1	7.252.1	4.141.1	
7.123.1	8.86.1	7.85.2	
7.212.1		7.251.3	
7.234.2		7.159.2	
7.130.1		7.146.3	
7.118.1		7.34.1	
8.287.2			
7.58.3			
7.66.1			
7.41.3			
7.56.3			
7.160.2			
7.202.3			
8.8.3			
4.25.1			
7.23.3			
4.142.2			
4.143.2			
4.121.1			

EXAMPLE 11

STRUCTURAL ANALYSIS OF ANTI-IGF-I/II ANTIBODIES

[0199] The variable heavy chains and the variable light chains of several antibodies were sequenced to determine their DNA sequences. The complete sequence information for the anti-IGF-I/II antibodies is provided in the sequence listing with nucleotide and amino acid sequences for each gamma and kappa chain combination. The variable heavy sequences were analyzed to determine the VH family, the D-region sequence and the J-region sequence. The sequences were then translated to determine the primary

amino acid sequence and compared to the germline VH, D and J-region sequences to assess somatic hypermutations.

[0200] The alignment of the sequences of these antibodies to their germline genes are shown in the following tables. Table 11 is a table comparing the antibody heavy chain regions to their cognate germ line heavy chain region. Table 12 is a table comparing the antibody kappa light chain regions to their cognate germ line light chain region. Mutations away from germline are shown as the new amino acid.

[0201] The variable (V) regions of immunoglobulin chains are encoded by multiple germ line DNA segments, which are joined into functional variable regions (V_HDJ_H or V_KJ_K) during B-cell ontogeny. The molecular and genetic diversity of the antibody response to IGF-I/II was studied in detail. These assays revealed several points specific to anti-IGF-I/II antibodies.

[0202] Analysis of five individual antibodies specific to IGF-I/II resulted in the determination that the antibodies were derived from three different germline VH genes, four of them from the VH4 family, with 2 antibodies being derived from the VH4-39 gene segment. Tables 11 and 12 show the results of this analysis.

[0203] It should be appreciated that amino acid sequences among the sister clones collected from each hybridoma are identical. For example, the heavy chain and light chain sequences for mAb 7.159.2 are identical to the sequences shown in Tables 11 and 12 for mAb 7.159.1.

[0204] The heavy chain CDR1s of the antibodies of the invention have a sequence as disclosed in Table 11. The CDR1s disclosed in Table 11 are of the Kabat definition. Alternatively, the CDR1s can be defined using an alternative definition so as to include the last five residues of the FR1 sequence. For example, for antibody 7.159.1 the FR1 sequence is QVQLVQSGAEVKKPGASVKVSCKAS (SEQ ID NO.: 93) and the CDR1 sequence is GYTFTSYDIN (SEQ ID NO.: 94); for antibody 7.158.1 the FR1 sequence is QLQLQESGPGLVKPSETLSLTCTVS (SEQ ID NO.: 95) and the CDR1 sequence is GGSIRSSSYYWG (SEQ ID NO.: 96); for antibody 7.234.1 the FR1 sequence is QLQLQESGPGLVKPSETLSLTCTVS (SEQ ID NO.: 97) and the CDR1 sequence is GGSINSSSNYWG (SEQ ID NO.: 98); for antibody 7.34.1 the FR1 sequence is QVQLQESGPGLVKPSETLSLTCTVS (SEQ ID NO.: 99) and the CDR1 sequence is

GGSISSYYWS (SEQ ID NO.: 100); and for antibody 7.251.3 the FR1 sequence is QVQLQESGPGLVKPSETLSLTCTVS (SEQ ID NO.: 101) and the CDR1 sequence is GGSISSYYWS (SEQ ID NO.: 102).

[0205] It should also be appreciated that where a particular antibody differs from its respective germline sequence at the amino acid level, the antibody sequence can be mutated back to the germline sequence. Such corrective mutations can occur at one, two, three or more positions, or a combination of any of the mutated positions, using standard molecular biological techniques. By way of non-limiting example, Table 12 shows that the light chain sequence of mAb 7.34.1 (SEQ ID NO.: 12) differs from the corresponding germline sequence (SEQ ID NO.:80) through a Pro to Ala mutation (mutation 1) in the FR1 region, and via a Phe to Leu mutation (mutation 2) in the FR2 region. Thus, the amino acid or nucleotide sequence encoding the light chain of mAb 7.34.1 can be modified to change mutation 1 to yield the germline sequence at the site of mutation 1. Further, the amino acid or nucleotide sequence encoding the light chain of mAb 7.34.1 can be modified to change mutation 2 to yield the germline sequence at the site of mutation 2. Still further, the amino acid or nucleotide sequence encoding the light chain of mAb 7.34.1 can be modified to change both mutation 1 and mutation 2 to yield the germline sequence at the sites of both mutations 1 and 2.

TABLE 11. HEAVY CHAIN ANALYSIS

Chain Name	SEQ ID NO.	V	D	J	FR1	CDR1	FR2	CDR2	FR3	CDR3	FR4
Germ-line**	75	VH1-8	N.A.	JH6B	QVQLVQSGAEVKKPG ASVKVSCKASGYTFT	SYDI N	WVRQATG QGLEWMG	WMNPNSGNT GYAQKFQG	RVTMTRNTSISTAYMELS SLRSEDTAVYYCAR	##YYYYY GMDV	WGQGTT VTVSSA
7_159_1	6	"	"	"	QVQLVQSGAEVKKPG ASVKVSCKASGYTFT	SYDI N	WVRQATG QGLEWMG	WMNPNSGNT GYAQKFQG	RVTMTRNTSISTAYMELS SLRSEDTAVYYCAR	DPYYYYY GMDV	WGQGTT VTVSSA
Germline	77	VH4-39	D6-19	JH2	QLQLQESGPGLVKPS ETLSLTCTVSGGSIS	SSSY YWG	WIRQPPG KGLEWIG	SIYYSGSTY YNPSLKS	RVTISVDTSKNQFSLKLS SVTAADTAVYYCAR	####SS# #WYFDL	WGRGTL VTVSSA
7_158_1	2	"	"	"	QLQLQESGPGLVKPS ETLSLTCTVSGGSIR	SSSY YWG	WIRQPPG KGLEWIG	GIYYSGSTY YNPSLKS	RVTMSVDTSKNQFSLKLS SVTAADTAVYYCAR	QRGHSSG WWYFDL	WGRGTL VTVSSA
7_234_1	18	"	"	"	QLQLQESGPGLVKPS ETLSLTCTVSGGSIN	SSSN YWG	WIRQPPG KGLAWIG	GIYYSGSTY YNPSLRS	RVTMSVDTSKNQFSLKLS SVTAADTAVYYCAR	QRGHSSG WWYFDL	WGRGTL VTVSSA
Germline	79	VH4-59	D1-20	JH6B	QVQLQESGPGLVKPS ETLSLTCTVSGGSIS	SYYW S	WIRQPPG KGLEWIG	YIYYSGSTN YNPSLKS	RVTISVDTSKNQFSLKLS SVTAADTAVYYCA#R	ITGT### GMDV	WGQGTT VTVSSA
7_34_1	10	"	"	"	QVQLQESGPGLVKPS ETLSLTCTVSGGSIS	SYYW S	WIRQPPG RGLEWIG	YFFYSGYTN YNPSLKS	RVTMSVDTSKNQFSLKLS SVTAADTAVYYCAC	ITGTTKG GMDV	WGQGAT VTVSSA
7_251_3	14	"	"	"	QVQLQESGPGLVKPS ETLSLTCTVSGGSIS	SYYW S	WIRQPPG KGLEWIG	YFFYSGYTN YNPSLKS	RVTISVDTSKNQFSLKLS SVTAADTAVYYCAC	ITGTTKG GMDV	WGQGTT VTVSSA

* The hatch designation (#) indicates a space in the germline and is used to show a proper alignment with the antibody sequences shown in the table.

** The germline sequences shown in the above table are for alignment purposes, and it should be realized that each individual antibody region exists in its own location within the variable regions of immunoglobulin germline DNA segments *in vivo*.

TABLE 12. LIGHT CHAIN ANALYSIS

Chain Name	SEQ ID NO.	V	J	FR1	CDR1	FR2	CDR2	FR3	CDR3	FR4
Germline	76	V1-19	JL2	QSVLTQPPSVSA APGQKVTISC	SGSSSNI GNNYVS	WYQQLPGT APKLLIY	DNNKRPS	GIPDRFSGSKSGTSAT LGITGLQTGDEADYYC	GTWDSSL SA##V	FGGGTK LTVLG
7_159_1	8	"	"	QSVLTQPPSVSA APGQKVTISC	SGSSSNI ENNHVS	WYQQLPGT APKLLIY	DNNKRPS	GIPDRFSGSKSGTSAT LGITGLQTGDEADYYC	ETWDTSL SAGRV	FGGGTK LTVLG
Germline	78	L5	JK3	DIQMTQSPSSVS ASVGDRVTITC	RASQGIS SWLA	WYQQKPGK APKLLIY	AASSLQS	GVPSRFSGSGSGTDFT LTISSLQPEDFATYYC	QQANSFP FT	FGPGTK VDIKR
7_158_1	4	"	"	DIQMTQSPSSVS ASVGDSVTITC	RASQGIS SYLA	WYQQKPGK APKLLIY	AASSLQS	GVPSRFSGNGSGTDFT LTISSLQPEDFATYYC	QQANNFP FT	FGPGTK VDIKR
7_234_1	20	"	"	DIQMTQSPSSVS ASVGDRVTITC	RASRGIS SWLA	WYQQRPGK APKLLIY	TASSLQS	GVPSRFSGSGSGTDFT LTISSLQPEDFATYYC	QQANSFP FT	FGPGTK VDIKR
Germline	80	V1-13	JL2.	QSVLTQPPSVSG APGQRVTISC	TGSSSNI GAGYDVH	WYQQLPGT APKLLIY	GNSNRPS	GVPDRFSGSKSGTSAS LAITGLQAEDEADYYC	QSYDSSL SGSV	FGGGTK LTVLG
7_34_1	12	"	"	QSVLTQAPSVSG APGQRVTISC	TGRSSNI GAGYDVH	WYQQFPGT APKLLIY	GNSNRPS	GVPDRFSGSKSGTSAS LAITGLQAEDEADYYC	QSYDSSL SGSV	FGGGTK LTVLG
7_251_3	16	"	"	QSVLTQPPSVSG APGQRVTISC	TGSSSNI GAGYDVH	WYQQLPGT APKLLIY	GNNNRPS	GVPDRFSGSKSGTSAS LAITGLQADDEADYYC	QSFDSSL SGSV	FGGGTK LTVLG

* The hatch designation (#) indicates a space in the germline and is used to show a proper alignment with the antibody sequences shown in the table.

** The germline sequences shown in the above table are for alignment purposes, and it should be realized that each individual antibody region exists in its own location within the variable regions of immunoglobulin germline DNA segments *in vivo*.

EXAMPLE 12

INHIBITION OF IGF-I AND IGF-II-INDUCED PHOSPHORYLATION OF hIGF-IR ECTOPICALLY EXPRESSED IN NIH3T3 CELLS

[0206] IGF ligands exert their proliferation and anti-apoptosis functions by activating receptor tyrosine kinase activity in the IGF-IR receptor. In order to evaluate the anti IGF-I/II antibodies for their ability to inhibit IGF-induced phosphorylation of IGF-IR, NIH3T3 cells ectopically expressing hIGF-IR, were used in the following assay.

[0207] NIH3T3 cells ectopically expressing the human IGF-IR were seeded in a 96-well plate at a density of 10,000 cells per well and incubated overnight in starvation media (1% charcoal stripped FBS). The following day, the growth medium was discarded, the wells were gently washed twice with PBS, and 100μL of serum-free medium (0% FBS) was added to starve the cells. After 1-2 hours, 100ul of serum-free medium with 0.05% BSA containing either IGF-I (10nM) or IGF-II (10nM) that was pre-incubated for 60 minutes at 37°C with various antibody concentrations, was added to the cells in triplicate. The stimulation was allowed to occur for 10 minutes at 37°C, after stimulation, media removed and 100uL 3.7%fromaldehyde in PBS/3%BSA added to each well and incubated at RT for 20 min. The cells were then washed 2X with PBS and 100uL permeabilization buffer (0.1% Triton-X in 3%BSA/PBS) was added to each well. This was allowed to incubate at RT for 10 min, discarded and 100ul of 0.6% hydrogen peroxide in PBS/3% BSA was added to inactivate any endogenous peroxidase activity. After a 20min RT incubation, the cells were washed 3X with PBS/0.1% Tween-20 and blocked by adding 100uL 10%FBS in PBS/0.1% Tween-20 at RT for 1hr. The Blocking Buffer was then removed and 50uL anti-phospho IGFIR antibody at 1ug/ml (cat#44-804, BioSource) was added to each well in 10%FBS/PBS-T. After a 2hr RT incubation cells were washed 3X with PBST soaking for 5 minutes between each wash. After the washes 50ul/well of a Goat anti Rabbit IgGFc-HRP secondary antibody diluted 1:250 in Blocking Buffer was added to each of the well. After a 1 hour RT incubation the cells were washed 3X for 5 minutes with PBST as before and tapped dry. 50ul of ECL reagent (DuoLux) was then added and RLUs was read immediately.

[0208] Thirty-two (32) antibody lines were screened, and two independent assays were performed for each antigen. The results for the top ten antibodies are summarized in Table 13 below.

TABLE 13. SUMMARY OF INHIBITION OF IGF-DEPENDENT IGF-IR PHOSPHORYLATION IN NIH3T3 CELLS

	IGF-I pTYR Results (n=2)				IGF-II pTYR Results (n=2)			
	% MAX Activation		EC50 (nM)*		% MAX Activation		EC50 (nM)*	
mAb ID	Mean	SD	Mean	SD	Mean	SD	Mean	SD
7.159.2	16.5%	6.6%	7.4	0.8	28.6%	5.4%	3.1	0.2
7.34.1	14.2%	1.8%	9.4	0.8	21.5%	3.9%	2.5	0.1
7.146.3	19.5%	3.9%	19.0	5.7	23.6%	2.1%	3.6	0.2
7.251.3	16.9%	5.4%	14.5	0.7	15.9%	1.5%	3.0	0.9
7.234.2	21.1%	3.0%	24.3	1.0	21.1%	0.1%	7.7	0.1
7.160.2	33.6%	6.4%	22.9	0.1	21.5%	0.6%	4.7	0.2
7.158.2	22.7%	0.9%	28.3	0.5	33.7%	2.4%	11.3	3.2
7.56.3	31.3%	2.2%	25.1	4.3	21.2%	0.2%	6.3	0.5
7.118.1	24.1%	5.2%	40.8	2.6	21.8%	3.2%	13.9	5.3
7.41.3	33.1%	6.1%	47.1	7.0	29.5%	4.4%	3.5	4.0

*These assays may have been run under antigen limiting conditions given the mAb KD for IGF-I and IGF-II.

EXAMPLE 13

INHIBITION OF IGF-I AND IGF-II-INDUCED PROLIFERATION OF NIH3T3 CELLS TRANSFECTED WITH HIGF-IR

[0209] As discussed above, one of the criteria for neutralizing IGF-I/II antibodies is the ability to inhibit IGF-induced proliferation. In order to evaluate the antibodies for their ability to inhibit IGF-induced proliferation, NIH3T3 cells ectopically expressing hIGF-IR, were used in the following assay.

[0210] NIH3T3 cells ectopically expressing hIGF-IR were seeded in a 96-well plate at a density of 5000 cells per well and cultured overnight in starvation medium (1% charcoal stripped FBS). The following day, the growth medium was discarded, the wells were gently washed twice in medium without serum, and 100μL of serum-free medium was added to starve the cells. 100μL of starvation media containing 15ng/ml IGFI or 50ng/ml IGFII pre-incubated for 30 min at 37°C with various antibody concentrations was added to the cells in duplicate or triplicate. Following a 20hr incubation cells are

pulsed with BrdU for 2hrs and the degree of incorporation (proliferation) was quantitated using the Cell Proliferation ELISA kit from Roche (Roche, Cat#1 647 229).

[0211] A total of 32 antibody lines were screened, and two or three independent assays were performed for each antigen. The results for the top 10 antibodies are summarized in Table 14 below.

TABLE 14. SUMMARY OF INHIBITION OF IGF-DEPENDENT PROLIFERATION OF NIH3T3/hIGF-IR CELLS

	IGF-I Proliferation Assay				IGF-II Proliferation Assay			
	% Inhibition		EC50 (nM)*		% Inhibition		EC50 (nM)*	
mAb ID	Mean (n=3)	SD	Mean (n=3)	SD	Mean (n=2)	SD	Mean (n=2)	SD
7.159.2	77.0%	9.6%	24.1	5.9	99.3%	0.6%	7.6	2.5
7.34.1	72.6%	5.6%	23.4	8.1	73.6%	11.8%	16.3	0.4
7.146.3	65.3%	5.5%	37.2	4.5	82.0%	6.9%	15.9	3.4
7.251.3	72.4%	15.3%	38.9	4.3	79.2%	7.8%	22.0	3.7
7.234.2	67.3%	6.9%	40.6	4.6	62.1%	17.2%	24.3	2.4
7.160.2	62.8%	5.7%	47.6	10.7	45.9%	0.8%	24.7	2.8
7.158.2	57.4%	19.5%	42.8	1.7	54.6%	6.6%	36.0	4.2
7.56.3	50.2%	7.8%	65.7	31.9	48.0%	10.9%	38.3	7.5
7.118.1	59.4%	14.5%	1626.6	2714.5	68.3%	0.8%	49.9	3.8
7.41.3	29.5%	14.7%	76.3	35.9	51.9%	13.7%	61.9	23.7

*These assays may have been run under antigen limiting conditions given the mAb KD for IGF-I and IGF-II.

EXAMPLE 14

INHIBITION OF IGF-I AND IGF-II-INDUCED PHOSPHORYLATION OF hIGF-IR EXPRESSED IN BxPC3 HUMAN PANCREATIC TUMOR CELLS

[0212] IGF-I/II exert their proliferation and anti-apoptosis functions by activating receptor tyrosine kinase activity in the IGF-IR receptor. In order to evaluate the antibodies for their ability to inhibit IGF-induced phosphorylation of IGF-IR, BxPC3 human pancreatic tumor cells, which express endogenous hIGF-IR, were used in the following assay.

[0213] BxPC3 cells were seeded in a 96-well plate at a density of 55,000 cells per well and incubated overnight in regular growth medium. The following day, the growth medium was discarded, the wells were gently washed twice in medium without serum, and 100μL of serum-free medium was added to starve the cells. After 24 hours,

the medium was discarded, and the cells were gently washed once in medium without serum. Serum-free medium with 0.05% BSA containing either IGF-I (20ng/ml) or IGF-II (75ng/ml) was pre-incubated for 30 minutes at 37°C with various antibody concentrations, and 100µL was then added to the cells in triplicate. The plates were incubated for 15 minutes at 37°C, and were subsequently rinsed with cold PBS. 100µL of lysis buffer was added to the wells and the plates were incubated for 30 minutes at 4°C. The lysates were spun down at 2000 rpm for 10 minutes at 4°C, and the supernatant was collected. IGF-IR phosphorylation was quantitated using the Duoset human phosphor-IGF-IR ELISA kit (R&D Systems, Cat. No. DYC1770).

[0214] Ten antibody lines were screened, and two independent assays were performed for each antigen. The results are summarized in Table 15 below.

TABLE 15. SUMMARY OF INHIBITION OF IGF-DEPENDENT IGF-IR PHOSPHORYLATION

	IGF-I pTYR Results (n=2)				**IGF-II pTYR Results (n=2)**			
	n=1		**n=2**		**n=1**		**n=2**	
mAb ID	**Max % Inhibition (333.3nM)**	**EC50 (nM)**	**Max % Inhibition (333.3nM)**	**EC50 (nM)**	**Max % Inhibition (333.3nM)**	**EC50 (nM)**	***Max % Inhibition (133.3nM)**	**EC50 (nM)**
7.159.2	100.0	3.3	100.0	1.6	100.0	1.6	91.2	1.7
7.34.1	100.0	5.9	98.5	3.8	100.0	2.0	89.7	1.9
7.146.3	96.4	16.1	94.2	10.7	100.0	2.0	87.9	2.0
7.251.3	95.7	7.5	95.2	5.3	100.0	N.D.	91.3	2.6
7.234.2	97.3	5.1	91.5	2.9	98.5	1.9	77.3	2.3
7.160.2	93.4	5.3	89.2	3.1	88.6	1.7	73.2	2.5
7.158.2	92.9	4.5	89.4	3.6	92.4	N.D.	74.0	4.2
7.56.3	84.9	N.D.	88.7	6.5	91.4	10.1	66.2	5.1
7.118.1	90.5	13.1	90.6	11.8	95.7	17.9	78.0	13.1
7.41.3	88.6	6.5	86.5	6.5	88.6	4.5	70.6	3.1

*333nM for the last 3 antibodies.

N.D.: Not Determined

EXAMPLE 15

INHIBITION OF IGF-I AND IGF-II-INDUCED PROLIFERATION OF BxPC3 HUMAN PANCREATIC TUMOR CELLS

[0215] As discussed above, one of the criteria for neutralizing IGF antibodies is the ability to inhibit IGF-induced proliferation. In order to evaluate the antibodies for

their ability to inhibit IGF-induced proliferation, BxPC3 human pancreatic tumor cells, which express endogenous hIGF-IR, were used in the following assay.

[0216] BxPC3 cells were seeded in a 96-well plate at a density of 2000 cells per well and cultured overnight in regular growth medium. The following day, the growth medium was discarded, the wells were gently washed twice in medium without serum, and 100μL of serum-free medium with 10μg/ml transferrin and 0.1% BSA (assay medium) was added to starve the cells. After 24 hours, the medium was discarded, the cells were gently washed once in medium without serum, and 100μL of assay medium containing 20ng/ml IGF preincubated for 30 min at 37°C with various antibody concentrations was added to the cells in duplicate or triplicate. The plates were incubated for 3 days, and proliferation was quantitated using the CellTiter-Glo reagent (Promega).

[0217] Ten antibody lines were screened, and two or three independent assays were performed for each antigen. The results are summarized in Table 16 below. Based on the functional data below and the data from the Example 14, the four best antibodies were selected. IGF-I-induced proliferation assay data was excluded from the selection criteria because of the high assay variability observed.

TABLE 16. SUMMARY OF INHIBITION OF IGF-DEPENDENT PROLIFERATION OF BxPC3 HUMAN PANCREATIC TUMOR CELLS

	IGF-II Proliferation Results (n=2)			
	n=1		**n=2**	
MAb ID	**Max % Inhibition (333.3nM)**	**EC50 (nM)**	***Max % Inhibition (133.3nM)**	**EC50 (nM)**
7.159.2	120.0	0.8	118.3	0.9
7.34.1	117.0	0.5	109.0	6.7
7.146.3	128.7	3.0	119.5	7.3
7.251.3	128.5	0.5	105.3	4.4
7.234.2	111.7	N.D.	200.7	2.6
7.160.2	79.7	1.1	155.7	N.D.
7.158.2	86.3	0.0013	148.3	N.D.
7.56.3	87.0	N.D.	112.3	102.0
7.118.1	114.0	34.0	137.0	54.7
7.41.3	102.0	N.D.	73.0	N.D.

*333nM for the last 3 antibodies

N.D.: Not Determined

EXAMPLE 16

DETERMINATION OF CROSS-REACTIVITY WITH MOUSE IGF-I, IGF-II AND INSULIN

[0218] One objective was to develop antibodies that were specific to IGF-I and IGF-II but that have no cross-reactivity with insulin. In order to perform later experiments in animals, the antibodies should also cross-react with murine IGF-I/II but not murine insulin. Accordingly, ELISA assays were performed to determine whether selected antibodies were able to cross-react with murine IGFs or insulin.

[0219] As shown in Table 17, five of the top ten antibodies were tested for cross-reactivity with mouse or rat insulin by ELISA. The ELISAs showed that these antibodies had no cross-reactivity with mouse or rat insulin, compared to negative control antibody PK16.3.1 and in contrast to positive control anti-rat insulin antibody.

TABLE 17. CROSS-REACTIVITY WITH MOUSE INSULIN

	OD 450 with different Ag		
Antibodies	**mouse Insulin**	**Rat Insulin**	**No Ag**
7.159.2	0.52	0.52	0.56
7.160.2	0.60	0.57	0.62
7.34.1	0.48	0.47	0.55
7.251.3	0.55	0.53	0.56
7.234.2	0.51	0.49	0.66
Serum	1.28	1.23	1.34
anti Rat Insulin	2.52	3.06	0.10
PK16.3.1	0.58	0.58	0.62

EXAMPLE 17

INHIBITION OF MOUSE IGF-I AND IGF-II-INDUCED PHOSPHORYLATION OF HUMAN IGF-IR ECTOPICALLY EXPRESSED IN NIH3T3 CELLS

[0220] The monoclonal antibodies with cross-reactivity with mouse IGF-I and IGF-II were further tested in order to determine the extent they inhibit IGF-induced phosphorylation of the IGF-IR. This assay was performed as previously described using NIH3T3 cells ectopically expressing the hIGF-IR receptor. The results of this assay are summarized in Table 18.

[0221] NIH3T3 cells ectopically expressing the human IGF-IR were seeded in a 96-well plate at a density of 10,000 cells per well and incubated overnight in starvation media (1% charcoal stripped FBS). The following day, the growth medium was

discarded, the wells were gently washed twice with PBS, and 100μL of serum-free medium (0% FBS) was added to starve the cells. After 1-2 hours, 100ul of serum-free medium with 0.05% BSA containing either mouse IGF-I (10nM) or IGF-II (10nM) (R&D Systems, Inc., Minneapolis, MN Cat. No. 791-MG and 792-MG respectively) that was pre-incubated for 60 minutes at 37°C with various antibody concentrations, was added to the cells in triplicate. The stimulation was allowed to occur for 10 minutes at 37°C, after stimulation, media removed and 100uL 3.7% formaldehyde in PBS/3%BSA added to each well and incubated at RT for 20 min. The cells were then washed 2X with PBS and 100uL permeabilization buffer (0.1% Triton-X in 3%BSA/PBS) was added to each well. This was allowed to incubate at RT for 10 min, discarded and 100ul of 0.6% hydrogen peroxide in PBS/3% BSA was added to inactivate any endogenous peroxidase activity. After a 20min RT incubation, the cells were washed 3X with PBS/0.1% Tween-20 and blocked by adding 100uL 10%FBS in PBS/0.1% Tween-20 at RT for 1hr. The Blocking Buffer was then removed and 50uL anti-phospho IGFIR antibody at 1ug/ml (cat#44-804, BioSource) was added to each well in 10%FBS/PBS-T. After a 2hr RT incubation cells were washed 3X with PBST soaking for 5 minutes between each wash. After the washes 50ul/well of a Goat anti Rabbit IgGFc-HRP secondary antibody diluted 1:250 in Blocking Buffer was added to each of the well. After a 1 hour RT incubation the cells were washed 3X for 5 minutes with PBST as before and tapped dry. 50ul of ECL reagent (DuoLux) was then added and RLUs was read immediately.

TABLE 18. INHIBITION OF MOUSE IGF-INDUCED PHOSPHORYLATION OF hIGF-IR

	Mouse IGF-I EC50 (nM)		**Mouse IGF-II EC50 (nM)**	
mAb ID	**n=1**	**n=2**	**n=1**	**n=2**
7.159.2	2.8	5.7	3.1	5.0
7.34.1	6.0	10.2	4.0	9.7
7.251.3	6.7	10.6	5.4	8.7
7.234.2	46.0		36.1	
7.160.2	49.5		225.2	

EXAMPLE 18

INHIBITION OF THE GROWTH OF NIH3T3 CELLS EXPRESSING IGF-II AND IGF-1R *IN VIVO* IN NUDE MICE

[0222] In order to evaluate the antibodies for their ability to inhibit IGF-II-induced proliferation *in vivo*, the following experiments were performed.

[0223] Female nude mice 6-8 weeks of age (supplied by Charles River Laboratories, Wilmington, MA, USA) were implanted subcutaneously with $5x10^6$ Clone 32 cells (NIH3T3 cells ectopically overexpressing human IGF-II and human IGF-1R). The cells were suspended in PBS in a total inoculum volume of 330 µl. The tumors were allowed to grow to 100-200mm^3 prior to treatment with monoclonal antibodies 7.159.2, 7.34.1 and 7.251.3. Antibodies or IgG2 isotype control antibody suspended in PBS were administered intraperitoneally to randomized groups of 9 or 12 mice weekly for 4 weeks at 5 or 50 mg/kg from Day 22. PBS was administered as a vehicle control to a further group of 11 mice weekly for 4 weeks from Day 22. Tumor size and body weight was measured 2-3 times per week. The results are summarized in Figures 1 and 2.

[0224] Significant tumor growth inhibition was observed (see Figure 1) with no significant weight loss occurring in any group (see Figure 2). Antibodies 7.159.2, 7.34.1 and 7.251.3 significantly inhibited the growth of Clone 32 tumors at 5 and 50 mg/kg/week (see Figure 1).

EXAMPLE 19

INHIBITION OF THE GROWTH OF NIH3T3 CELLS EXPRESSING IGF-I AND IGF-1R IN VIVO IN NUDE MICE

[0225] In the previous example, the antibodies were shown to inhibit IGF-II-induced proliferation *in vivo*. In order to evaluate the antibodies for their ability to inhibit IGF-I-induced proliferation *in vivo*, the following experiments were performed.

[0226] Female nude mice (Alderley Park strain derived from Swiss nu/nu mice strain, supplied by AstraZeneca) were implanted with $5x10^6$ viable P12 cells [NIH3T3 cells ectopically overexpressing human IGF-I and human IGF-1R (Pietrzkowski et al, Cell Growth & Differentiation, 3, 199-205, 1992)] subcutaneously in the left flank. The cells were suspended in PBS in a total inoculum volume of 0.1 ml. Two groups of animals (each n=10) were dosed twice weekly from day of NIH3T3 cell implant with either mAb 7.159.2 at 1.0 mg per mouse or with an equivalent volume of PBS vehicle (0.3 ml) for the same schedule. All doses were given intraperitoneally via the (i.p.) route. Animal body weights were measured daily and, once established, tumor measurements

were taken twice weekly using calipers. The volume for all measurable tumors was calculated from the caliper measurements assuming an ovoid shape. The results are summarized in Figure 3.

[0227] As shown in Figure 3, significant tumor growth inhibition was observed with mAb 7.159.2 following twice weekly i.p. administration of 1.0mg antibody/mouse. No significant weight loss was observed in any of the groups of animals.

EXAMPLE 20

INHIBITION OF TUMOR CELL GROWTH IN HUMAN PATIENTS

[0228] A group of human cancer patients diagnosed with pancreatic cancer is randomized into treatment groups. Each patient group is treated with weekly intravenous injections of mAb 7.159.2, 7.34.1 or 7.251.3 described herein. Each patient is dosed with an effective amount of the antibody ranging from 50 mg/kg to 2,250 mg/kg for 4-8 weeks. A control group is given only the standard chemotherapeutic regimen.

[0229] At periodic times during and after the treatment regimen, tumor burden is assessed by magnetic resonance imaging (MRI). It is found that the patients who have received weekly antibody treatment with mAb 7.159.2, 7.34.1 or 7.251.3 show significant reductions in tumor size, compared to patients that do not receive antibody treatment. In some treated patients, the tumors are no longer detectable. In contrast, tumor size increases or remains substantially the same in the control group.

EXAMPLE 21

INHIBITION OF TUMOR CELL GROWTH IN A HUMAN PATIENT

[0230] A human patient is diagnosed with a malignant tumor. The patient is treated with weekly intravenous injections of mAb 7.159.2 for 8 weeks. At periodic times during and after the treatment regimen, tumor burden is assessed by magnetic resonance imaging (MRI). Significant reductions in tumor size are found.

EXAMPLE 22

TREATMENT OF ACROMEGALY IN A HUMAN PATIENT

[0231] An adult male is diagnosed with acromegaly. The patient is treated with bi-weekly intravenous injections of mAb 7.34.1 over a period of 2 years. As a result, the patient experiences a significant reduction in the symptoms of acromegaly.

EXAMPLE 23

TREATMENT OF PSORIASIS IN A HUMAN PATIENT

[0232] An adult female is diagnosed with severe psoriasis. The patient is treated with bi-weekly intravenous injections of mAb 7.251.3 over a period of 3 weeks. As a result, the patient experiences a significant reduction in the symptoms of psoriasis.

EXAMPLE 24

TREATMENT OF OSTEOPOROSIS IN A HUMAN PATIENT

[0233] An adult female is diagnosed with osteoporosis. The patient is treated with bi-weekly intravenous injections of mAb 7.159.2 over a period of a year. As a result, there is a significant reduction in loss of bone density.

EXAMPLE 25

TREATMENT OF ATHEROSCLEROSIS IN A HUMAN PATIENT

[0234] An adult male is diagnosed with atherosclerosis. The patient is treated with bi-weekly intravenous injections of mAb 7.34.1 over a period of a year. As a result, the patient experiences a reduction in the symptoms of atherosclerosis, such as angina pectoris.

EXAMPLE 26

TREATMENT OF RESTENOSIS IN A HUMAN PATIENT

[0235] An adult male receives angioplasty to relieve a blocked artery. Following the angioplasty procedure, the patient is treated with bi-weekly intravenous injections of mAb 7.251.3 over a period of a year. As a result, the patient does not experience restenosis of the treated artery.

EXAMPLE 27

TREATMENT OF DIABETES IN A HUMAN PATIENT

[0236] An adult female is diagnosed with diabetes. The patient is treated with bi-weekly intravenous injections of mAb 7.159.2 over a period of a year. As a result, the symptoms of diabetes are reduced.

SEQUENCES

[0237] The sub-cloned hybridomas were sequenced to determine their primary structure at both the nucleotide and amino acid level for both the variable heavy and the variable light chain genes. The nucleotide and polypeptide sequences of the variable regions of the monoclonal antibodies against IGF-I and IGF-II, as listed in Table 1, are provided in the Sequence Listing.

INCORPORATION BY REFERENCE

[0238] All references cited herein, including patents, patent applications, papers, text books, and the like, and the references cited therein, to the extent that they are not already, are hereby incorporated herein by reference in their entirety.

EQUIVALENTS

[0239] The foregoing written specification is considered to be sufficient to enable one skilled in the art to practice the invention. The foregoing description and Examples detail certain preferred embodiments of the invention and describes the best mode contemplated by the inventors. It will be appreciated, however, that no matter how detailed the foregoing may appear in text, the invention may be practiced in many ways and the invention should be construed in accordance with the appended claims and any equivalents thereof.

Claims

1. A fully human isolated specific binding protein that preferentially binds to insulin-like growth factor-II (IGF-II) with cross-reactivity to insulin-like growth factor I (IGF-I) and neutralizes IGF-I and IGF-II activity, wherein said binding protein, or binding fragment thereof, comprises:

   a heavy chain complementarity determining region 1 (CDR1) having the amino acid sequence of "Ser Tyr Asp Ile Asn" (SEQ ID NO: 33);
   a heavy chain complementarity determining region 2 (CDR2) having the amino acid sequence of "Trp Met Asn Pro Asn Ser Gly Asn Thr Gly Tyr Ala Gln Lys Phe Gln Gly" (SEQ ID NO: 34);
   a heavy chain complementarity determining region 3 (CDR3) having the amino acid sequence of "Asp Pro Tyr Tyr Tyr Tyr Tyr Gly Met Asp Val" (SEQ ID NO: 35);
   a light chain complementarity determining region 1 (CDR1) having the amino acid sequence of "Ser Gly Ser Ser Ser Asn Ile Glu Asn Asn His Val Ser" (SEQ ID NO: 36);
   a light chain complementarity determining region 2 (CDR2) having the amino acid sequence of "Asp Asn Asn Lys Arg Pro Ser" (SEQ ID NO: 37); and
   a light chain complementarity determining region 3 (CDR3) having the amino acid sequence of "Glu Thr Trp Asp Thr Ser Leu Ser Ala Gly Arg Val" (SEQ ID NO: 38).

2. The specific binding protein of claim 1, wherein said binding protein is monoclonal antibody 7.159.2 (ATCC Accession Number PTA-7424).

3. The specific binding protein of claim 1, wherein said binding protein comprises a heavy chain polypeptide having the sequence of SEQ ID NO.: 6;
   or wherein said binding protein comprises a light chain polypeptide having the sequence of SEQ ID NO.: 8.

4. The specific binding protein of any of Claims 1 or 3, wherein said binding protein comprises a heavy chain polypeptide having the sequence of SEQ ID NO.: 6; and wherein said binding protein comprises a light chain polypeptide having the sequence of SEQ ID NO.: 8.

5. The specific binding protein of any one of the preceding claims, wherein said binding protein is a fully human monoclonal antibody; or a binding fragment of a fully human monoclonal antibody.

6. The specific binding protein of Claim 5, wherein said binding fragment is selected from the group consisting of Fab, Fab' or $F(ab')_2$ and Fv.

7. The specific binding protein of any of Claims 1 to 6 in a mixture with a pharmaceutically acceptable carrier.

8. A nucleic acid molecule encoding the specific binding protein of any one of claims 1 to 6.

9. A vector comprising the nucleic acid molecule of Claim 8.

10. A host cell comprising the vector of Claim 9.

11. The specific binding protein of any one of Claims 1 to 6, wherein said specific binding protein does not bind specifically to IGF-II or IGF-I proteins when said proteins are bound to Insulin Growth Factor Binding Proteins.

12. A method of determining the level of insulin-like growth factor-II (IGF-II) and insulin-like growth factor I (IGF-I) in a patient sample comprising:

    contacting a sample from a patient with the specific binding protein of any one of claims 1 to 6; and
    determining the level of IGF-I and IGF-II in said sample.

13. Use of the specific binding protein of any one of Claims 1 to 6 in the preparation of a medicament for the treatment of a malignant tumor.

14. The use of Claim 13, wherein said malignant tumor is selected from the group consisting of: melanoma, non-small cell lung cancer, glioma, hepatocellular (liver) carcinoma, thyroid tumor, gastric (stomach) cancer, prostrate cancer, breast cancer, ovarian cancer, bladder cancer, lung cancer, glioblastoma, endometrial cancer, kidney cancer, colon cancer, pancreatic cancer, and epidermoid carcinoma.

15. The use of Claim 13 or 14, wherein said medicament is for use in combination with a second anti-neoplastic agent selected from the group consisting of an antibody, a chemotherapeutic agent, and a radioactive drug.

16. The use of Claim 13 or 14, wherein said medicament is for use in conjunction with or following a conventional surgery, a bone marrow stem cell transplantation or a peripheral stem cell transplantation.

17. A conjugate comprising the antibody of Claim 5 or a binding fragment thereof and a therapeutic agent.

18. The conjugate of Claim 17, wherein the therapeutic agent is a toxin; a radioisotope; or a pharmaceutical composition.