DNA MARKER PROFILE DATA ANALYSIS

BACKGROUND OF THE INVENTION

Field of the Invention

The present invention relates generally to DNA marker analysis and more particularly to methods for processing raw DNA marker profile data into a format that facilitates analysis of the raw data.

Background and Related Art

DNA markers are molecular genetic markers identified by studying genomic DNA samples. The genome of higher organisms is contained in rod-like structures of tightly coiled DNA (deoxyribonucleic acid) found in the cell nucleus of plants and animals, called chromosomes. Every strand of DNA has pieces or sequences of nucleotides or bases (consisting of adenine, guanine, cytosine and thymine) that contain genetic information which contribute to the function of a gene (exons) and sequences that apparently supply no relevant genetic information at all (introns, minisatellites or microsatellites). Intron, minisatellite or microsatellite sequences are repeated within the gene and in other genes of a DNA sample. Each organism has a unique pattern of these minisatellites (the only exception being multiple individuals from a single zygote, i.e., identical twins), which can be analyzed.

DNA markers are used for mapping and tagging physical traits of interest, and as indicators of genetic diversity. DNA polymorphism can be used for investigating the organization of genomes and for the construction of genetic maps, which can provide detailed blueprints for strategies of gene isolation by map-based cloning, marker-assisted selection, and introgression and dissection of complex traits. Genetic mapping offers modern breeders and scientists a powerful array of tools for analyzing the inheritance of important physiological traits in animals and plants.

The investigation of taxonomic units at the species level and the determination of the uniqueness of a species are essential for conservational, systemic, ecological and evolutionary studies. Knowledge of genetic relationships among genotypes in plant breeding programs permits the organization of germplasm, including elite lines, and provides for more efficient parental selection. Exotic germplasms are important sources of genes exerting highly desirable qualitative effects on traits, such as biotic and abiotic stress resistance. Transfer of such genes in breeding programs can be enhanced through marker-assisted selection with backcrossing. DNA markers also can help determine, regarding diversity questions, how populations of given species are distributed, how genetically distinct different populations are with respect to each other, and how much genetic variation is present in and among populations.

Many different techniques for obtaining DNA markers have been developed in recent years, which include:

RFLP (Restriction Fragment Length Polymorphism):

The RFLP technique is based on the fact that base substitution in a restriction endonuclease site or base insertions or deletions between sites can result in detectable differences in the length of DNA fragments when a DNA sample is digested with restriction enzymes. DNA from an organism is isolated and digested with suitable restriction enzymes and size fractionated by gel electrophoresis. The DNA fragments are then transferred to a membrane before being hybridized to radioisotope labeled probes. The RFLP method was the first and most widely used method in obtaining genetic markers and provides very reproducible results. In many species, however, such polymorphism occurs at a low frequency, and the RFLP method requires large amounts of DNA, which in turn requires that a large amount of the sample be collected.

RAPD (Random Amplified Polymorphic DNA):

PCR (Polymerase Chain Reaction) amplifies genomic DNA with a single random primer of about ten bases, and the amplified products are separated by agarose gel electrophoresis. It has the advantage of being cheap, simple and fast to perform, and does not require prior sequence information. However, this method can result in non-reproducible bands, faint bands, or spurious bands that cannot be scored confidently.

DAF (DNA Amplification Fingerprinting):

This technique is similar to RAPD, except that very short (e.g., five to eight nucleotide) random primers are used for the amplification of genomic DNA, and the PCR fragments are imaged by polyacrylamide gel electrophoresis followed by silver staining. This method provides more

variation, making it possible to distinguish between closely related cultivars. However, this method also suffers from the reproducibility and non-genetic origin problems of RAPD.

SSR (Simple Sequence Repeats):

In animal and plant genomes, microsatellite regions are rich in tandem repeats of two or five nucleotides. Such sequences are named simple sequence repeats. Such repeats are associated with high levels of polymorphism, and are well suited for generating genetic markers. Primers are used to amplify the SSR regions and the resulting PCR fragments are resolved by polyacrylamide gel electrophoresis. This method provides the high polymorphism of genetic origin, requires very little DNA, and give reproducible and repetitive results. However, prior sequence information is needed on multiple microsatellite loci for a particular organism in order to utilize this method of obtaining DNA markers.

VNTR (Variable Number Tandem Repeats):

Repetitive sequences of 10-100 base pairs occurring in tandem arrays of up to 1,000 units distributed throughout the genome are known as minisatellites. Variations in the number of tandem repeats of minisatellite DNA regions have been used as molecular markers to detect high levels of polymorphism, even between closely related individuals among the population of a single species. Mutation rates at minisatellite loci have been estimated to be as high as $2x10^3$ per meiosis. VNTR is widely used in forensic investigations.

AFLP (Amplified Fragment Length Polymorphism):

AFLP is based on the selective PCR amplification of primer-ligated restriction fragments from a total

digestion of genomic DNA. It requires only a small amount of genomic DNA, does not require prior sequence information, and has the ability to amplify sequences from a large number of restriction fragments. It provides high polymorphism of genetic origin and gives reproducible results.

In any of the above methods, after hybridization or PCR reactions and electrophoresis, data is obtained by a variety of methods including autoradiography, ethidium bromide staining, silver staining, and fluorescence detection. Fragment sizes are calculated or estimated from DNA size standards run either alongside the samples or mixed with the samples. A typical result comprises the detection or imaging of a pattern of DNA fragments of different sizes at different intensities or peaks, called a DNA "fingerprint." The uniqueness of any DNA fingerprint is dependent on many factors, mainly the source DNA and probe or primer combination, but also on reaction conditions. When reaction conditions are standardized, the DNA fingerprint becomes specific to the source DNA and probe or primer combination.

Improvements in technology have greatly increased the speed at which raw DNA marker data can be generated. In comparison data collection and analysis is less developed and is the main bottleneck in restricting the full exploitation of the potential associated with DNA fingerprinting methods. Manual interpretation of the raw data is tedious, time-consuming, and subjective, and the results from different batches even within the same laboratory are difficult to compare. Consequently, it is almost impossible to exchange and compare quantitative results from different laboratories. There thus exists in

the art a need for a simple, standardized format for expressing DNA fingerprint data that will greatly facilitate the development of the entire field.

SUMMARY OF THE INVENTION

The present invention provides a solution to the shortcomings of the prior art as discussed above.

In particular, the present invention provides a method for obtaining DNA fingerprint profile data, including the steps of measuring peak intensity and size of each DNA fragment in a sample of genomic DNA, classifying the peak intensities of said fragments according to a predetermined discrete intensity level scale, aligning the sizes of said fragments into corresponding ones of discrete size bins, entering the classified peak intensities of said fragments into a data record in a sequence determined by said aligned sizes, and storing the record.

According to another aspect of the invention, a computer program product is provided, including a computer-readable medium having computer-executable code recorded thereon for obtaining DNA fingerprint profile data, said computer-executable code including code modules for measuring peak intensity and size of each DNA fragment in a sample of genomic DNA; for classifying the peak intensities of said fragments according to a predetermined discrete intensity level scale; for aligning the sizes of said fragments into corresponding ones of discrete size bins; for entering the classified peak intensities of said fragments into a data record in a sequence determined by said aligned sizes; and storing the record in a computer-readable storage medium.

BRIEF DESCRIPTION OF THE DRAWINGS

The invention will be described in detail with reference to the following drawings in which:

Fig. 1 is a flow diagram of a method of processing DNA marker data according to one preferred embodiment of the present invention;

Fig. 2 is a table showing a format for expressing peak intensity of detected DNA fragments according to one embodiment of the invention;

Figs. 3A and 3B are diagrams for explaining a method of binning DNA fragments according to the invention;

Fig. 4 is a flow diagram of a method of binning DNA fragments according to the invention;

Fig. 5 is an example of an Amplified-type signature record of a DNA fingerprint according to the present invention;

Figs. 6A and 6B are tables providing values for carrying out scoring of DNA fingerprints according to the signature records obtained by the present invention; and

Fig. 7 is a flow diagram of the scoring process according to one preferred embodiment of the present invention.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

Referring to Fig. 1, one preferred method for DNA marker data analysis according to the present invention is illustrated. The analysis according to the invention preferably is carried out on a computer (such as a PC, minicomputer, mainframe, work station, server, etc.), using a software package performing the functions of the invention described in detail below.

At step 100, the labeled DNA fragments of a DNA sample that has been processed to obtain DNA markers according to any one of the abovementioned methods, are resolved, such as by running the sample through a gel electrophoresis. For example, using the AFLP technique, many small-sized PCR-generated fragments (typically in the range of 50-500 base pairs) are obtained, which comprise a DNA fingerprint. Depending upon whether radioisotope labeled primers or fluorescent labeled primers are used, different resolving processes are carried out. For radioisotope labeled primers, the samples are electrophoresed on a gel and the gel is exposed to X-ray film for several days. The exposed film is then manually interpreted.

For fluorescent labeled primers, the samples are run on an electrophoresis gel and fluorescent emission signals are detected in real time by a fluorescence sensor (such as a CCD camera or the like). The signals are digitized and inputted to a host computer for processing. Results are provided in tabular form. The use of fluorescent labeled primers is thus much faster than radioisotope primers and provides more reproducible results. An example of an automatic sequencer suitable for use in obtaining DNA marker data is the commercially available ABI 377 sequencer. Suitable DNA sequencers are available from a number of different manufacturers.

At step 102, the peak intensities of each fragment and the size of the fragment (given in bps or base pair length) are measured (such as by the fluorescence sensor coupled to a host processor or computer). The obtained measurements are stored in a memory storage medium (such

as a solid state memory, hard drive, magnetic tape drive, etc.) at step 104.

Raw peak data can be obtained by inputting appropriate commands to the software running the DNA sequencer. For example, an "Export Lane-to-Raw" command in the software interfacing with the ABI 377 exports to memory a raw lane text file containing summarized peak data in columnar form, as illustrated below.

G,1	35.29	51.49	151	544	683
G,2	35.65	53.22	296	2046	690
G,3	36.12	55.44	299	1834	699
G,4	36.32	56.42	378	1529	703
G,5	36.73	58.38	765	6140	711
G,6	37.35	61.31	156	963	723
G,7	37.72	63.01	171	979	730
G,8	38.03	64.46	514	2982	736

The six columns represent, respectively, peak, time, size, height (intensity), area and point. The present invention makes use of only the size (i.e., number of base pairs in the fragment) and the height (i.e., the intensity).

At step 106, the peak intensities of the stored fragments are normalized. In this regard, the intensities may be normalized by average amplitude, wherein unity would represent the average height of the peaks over the entire size range of the waveform.

At step 108, the normalized peak intensities are classified or transformed into one of five discrete peak levels, as shown in Fig. 2. The empirical ranges shown in Fig. 2 were chosen after experimental adjustments, and

were found by the inventors to work very well at producing consistent, reproducible signatures. According to one preferred embodiment of the invention, peaks are classified based on five discrete levels. This would allow adaptation to existing nucleotide sequence analysis software. However, peak intensities may be classified into any number of suitable levels without departing from the spirit of the invention.

The character set 'ABCD.' was chosen to represent the peak levels. The one-to-one correspondence between this character set and the universally used 'ATCGN' characters for representing DNA sequences enables adaptation of numerous sequence comparison software packages presently available on the market to the DNA fingerprint signature data of the present invention. In particular, such adaptation would involve simple adjustment of a software program's weight matrix to properly analyze the sequence as DNA marker peak data instead of DNA sequence data.

Next, at step 110, the stored DNA marker data are classified into discrete size (i.e., length) bins. Theoretically, an AFLP gel run should register peaks only at integral sizes (i.e., base pair lengths), because DNA fragments consist simply of multiple connected base pairs. However, because different base types are slightly different in size, peaks frequently are detected at fractional length sizes. In order to form the Amplified-type DNA marker signatures according to the invention, the detected peaks must be correlated to base pair positions, thus it is necessary to fit the experimental data into discrete sizes or "bins" each representing a base pair position or location starting from the beginning of a DNA fragment.

A typical method of binning is to form a ladder of the entire gel run, by lining up the gel lanes from the same run side-by-side so as to average out individual lane errors and to obtain a set of clear consensus rungs on the ladder into which the peaks are binned. However, this approach requires data from all lanes in the same gel run to be collected before the binning can be performed. This collective dependency exhibits an unacceptable deviation from the consistency and reproducibility requirements of the present invention, and thus a different method is used according to the invention, which allows an accurate and unbiased binning of peaks from a single lane, without the need to reference all other lanes in the run. Consequently, raw lane files of 2-3 Kb can be stored together with the generated DNA marker signature files according to the invention, which allows the signatures to be recalculated using later-developed algorithms.

A binning algorithm based on the borrowed concepts from physics of "spring" and "rubberband" energies is used according to the invention, since simple numeric rounding was found not to produce satisfactory results in binning a sequence of real-valued peaks. For example, as shown in Fig. 3A, for an experimentally obtained fragment sequence of sizes 67.3, 68.5, 69.4, 70.2 and 71.9, simple rounding would bin the sequence into 67, 69, 69, 70 and 72, obviously problematic since two peaks (68.5 and 69.4) would be binned into the same position although they are almost one whole position apart.

In order to arrive at a good fit, the relative distance between the peaks as well as their real positions is considered according to the invention. These two considerations can be combined mathematically into a

single potential energy function, based on the physical models of a spring and a rubberband.

If one end of a compressible spring is fixed and the other end displaced by a distance x, the spring exerts a force acting in the direction against the displacement, and is expressed as $F = -kx$, where k is the spring constant, and the minus sign indicates that the force is opposite to the direction of the displacement x. Integrating the force function gives the potential energy of the spring, which is expressed as $\frac{1}{2} kx^2$, and which indicates that the potential energy is proportional to the square of the displacement.

The main difference between a spring and a rubberband is that a rubberband can only be pulled (expanded), and not pushed back (compressed) as can a spring. Functionally, this means that a rubberband is equivalent to a spring of zero length.

The Spring-and-Rubberband model is applied to the peak binning problem by assuming, as shown in Fig. 3A, that a spring maintains the distance between two adjacent peaks, and a rubberband pulls the real-valued peaks toward integral size points. A corollary of this model is that the binning should be localized within a close cluster of consecutive peaks, so as to allow sequence fragments to move together as a whole (and thus maintain their shape as much as possible), and to reflect the fact that relative distances between peaks become less significant as they become farther apart. Thus, according to a preferred embodiment of the binning process according to the invention, as shown in Fig. 4 at step 1101, fragment sequences are grouped into clusters of smaller fragment sequences consisting of adjacent peaks no farther than 2.5

positions apart. Optimal formulae for representing the energy of the springs and rubberbands for purposes of binning were experimentally determined as shown in step 1102 of Fig. 4, where E_r is the rubberband energy, E_s is the spring energy, and E is the combined potential energy of the forces of the springs and rubberbands each acting on the peaks. In the equations, x is the displacement of the peaks and k_r and k_s are the proportionality constants. Since the system works best when $k_s/k_r = 3$, k_r can be set to 1 and k_s can be set to 3, such that $E = x_r^3 + 3x_s^2$.

At step 1103, the displacements x_s and x_r are varied in a methodical fashion to achieve the lowest combined potential energy E, and thus the best fit to the discrete bin sizes. Once the lowest or minimum potential energy E is obtained, the peaks are binned according to the indicated displacements at step 1104. For example, as shown in Fig. 3B, the first "spring" is compressed from 1.2 units to one unit, while the second "spring" is stretched from 0.9 units to one unit (in contrast with the result of rounding as shown in Fig. 3A, where the first spring is stretched from 1.2 to 2 units, and the second spring is compressed from 0.9 to zero units). Similarly, the second and third "rubberbands" have respective displacements of 0.5 units and 0.4 units, as in Fig. 3A.

Once the normalized peaks are classified and the fragments binned into discrete sizes, at step 112 (Fig. 1) the DNA marker signature data is formatted into a standardized data entry record as shown in Fig. 5. The record has three distinct sections or fields: the first field is a unique accession code that functions as an identifier for a particular sample. The accession code contains information pertaining to the source DNA, the

type of DNA marker generation technique, and the probe ID or primer combination. For example, as shown in Fig. 5, 501 designates a two-letter organism type (OR) and four-character alphanumeric organism ID, (D011) in this case an orchid cultivar. Reference numeral 502 designates a 1-character DNA marker technique, such as 'A' for AFLP, 'R' for RFLP, 'D' for RAPD, 'S' for SSR, and 'V' for VNTR (in the example, AFLP was used to generate the markers). Numeral 503 is a 1 or 2 primer pair code or probe code in the form of an alphabetic letter followed by a digit. In the case of primer combinations, an alphabet code indicates a primer in the 3' direction, and a digit code indicates a primer in the 5' direction. In the example, 'B1' indicates a predetermined primer combination for AFLP selective amplification.

The second field of the fingerprint signature according to the invention consists of other information, such as unit size (spacing), starting and ending positions, and a description of the sequence. Hence, numeral 504 indicates that the sequence spacing is 1 bp, numeral 505 indicates that the starting size for the fingerprint is 50, numeral 506 indicates that the ending size is 101, and 507 and 508 indicates an AFLP profile for D. *sonia*. The first line of the signature, containing the first and second fields, begins with a symbol such as a right brace (}); all subsequent lines not starting with '}' are interpreted as the signature sequence. Thus, signature sequence 509 in Fig. 5 gives the peak values for each fragment from 50 bp to 101 bp, in terms of the discrete levels A,B,C,D, and '.'. The data record so formed according to the invention is then stored at step 114 into a suitable data storage medium, such as a solid-

state memory, magnetic or optical recording medium, floppy disk, hard disk, etc.

Among the many advantages of this new format are the immediately informative nature of the format, providing information on a peak's location and its relative intensity; results for different samples are aligned and thus easily compared; exchange of results among different laboratories is simple and straightforward (allowing simple text typing); fingerprints can be easily put into and retrieved from databases; and large-scale data analysis becomes possible. Thousands of fingerprints or more can be compared and analyzed, and markers linked to certain phenotype can be found through the use of computers, eliminating many man-hours of manual analysis.

To take full advantage of the novel data marker format, the signature must allow some degree of comparison to differentiate between similar and unrelated species. A scoring system is used which adapts the reward-penalty concept used in BLAST sequence comparisons, wherein a positive score is awarded for each matching character, and a negative score is given as a penalty for each mismatch.

Fig. 6A illustrates the peak reward scale, and Fig. 6B illustrates the peak penalty scale according to the invention. The score awarded for matching peaks is determined by their intensity. If both have positive peaks at a location, two sequences are said to match at that position. If the intensities are different (i.e., an 'A' matched with a 'C'), the lower of the peaks is used to obtain the score value. Alternatively, for every peak that exists on one sequence but is absent (i.e., zero) on the other, a penalty is subtracted from the total score, according to the value of the missing peak's intensity.

Thus, scoring AB..C.D against AB..C.D gives a total score of 22 (10+6+4+2), while scoring CB..A.D against CB....D, a -4 penalty is incurred for the missing 'A', giving a total score of 8 (4+6-4+2).

Fig. 7 illustrates a scoring method according to one embodiment of the invention. First, at step 701, two signature strings are compared with each other by searching for exact matches. Scoring is performed within a predetermined window, i.e., within a predetermined size range (such as from position 50 to position 350, for example). In the event that either of the sequences falls outside the boundaries of the window, the window is shortened to include only the range where both sequences have values.

As exactly matching peaks are found, at step 702 the matching characters are replaced in the string with a dummy symbol, to prevent peaks from being matched multiple times when additional searching is performed using modified search parameters (as described below). After all exactly matching peaks are found, at step 703 a search is performed on the remaining peaks to find peaks of the same intensity in a position immediately adjacent to the position of the peak under consideration. This step compensates for possibly misaligned sequences, where strings have matching peak intensities but are deviated from each other by one position, e.g., 'AB.CD.' with '.AB.CD'. In the event that matching peaks are found deviating by one position, the amount of the reward score is one half the points shown in Fig. 6A. Thus, scoring 'AB.CD.' against '.AB.CD' considering the one position deviation would result in a score of 11 (5+3+2+1), while interpreting the strings as having differing positive

intensities at positions 2 and 5, with unmatched peaks at positions 1 and 6 and a C peak at position 4 deviating to a B peak at position 3, the resulting score would be reduced to 5 (-4+6+2+2-1). Matching peaks are again replaced with dummy characters at step 702, and processing advances to step 704, where a search is performed for peaks of differing intensity at the same position, and then step 705 where a search is performed for peaks of differing intensities at adjacent positions. Since all matched pairs of peaks have been progressively removed from the two strings being compared, the peaks remaining after step 705 represent mismatched peaks which will contribute to scoring penalties. At step 706 the award points are added for all previous matches, and at step 707 the (negative) penalty points are added for all mismatches, to arrive at a final score.

Negative scores are taken into account because sequences with mismatched peaks are considered even more different than sequences with no peaks. However, the use of negative scores makes the final score somewhat dependent upon the width of the scoring window, as the longer the sequences, the higher the probability of generating a higher overall score.

Because of this, a second scoring result is introduced which is expressed as a percentage value with a maximum of 100%. Within a designated scoring window, the percentage score of sequence B against sequence A is defined as the ratio of the score of B against A, to the highest possible score against A for the designated range (which would be attained if two identical A sequences were compared against each other).

While the score is a symmetric function in that score (A,B) = score (B,A), the percentage is an asymmetric function in that percent (A,B) = 100% x score (A,B)/score (A,A). The additional percentage metric thus is useful to indicate how good a match two sequences are within a given window. For example, considering a score between 'AB.CC.' and 'AB.CD.' (score = 22) compared with the same score for sequences in a wider window, such as 'ACD.A.CAB.CDC' and '.AB.C..BACD.C' (score = 22), the first pair of sequences is clearly a closer match than the second, which fact is reflected in the percentage scores when the second sequence of each pair is scored against the first of each pair (percentage = 93% versus 39%).

The invention having been thus described, it will be apparent to those skilled in the art that the same may be varied in many ways without departing from the spirit and scope of the inventions. All such modifications are intended to be encompassed by the following claims.

WHAT IS CLAIMED IS:

1. A method for obtaining DNA fingerprint profile data, comprising the steps of:

measuring peak intensity and size of each DNA fragment in a sample of genomic DNA;

classifying the peak intensities of said fragments according to a predetermined discrete intensity level scale;

aligning the sizes of said fragments into corresponding ones of discrete size bins;

entering the classified peak intensities of said fragments into a data record in a sequence determined by said aligned sizes; and

storing the record.

2. The method of claim 1, wherein said sample is obtained by ALFP.

3. The method of claim 1, wherein said sample is obtained by RLFP.

4. The method of claim 1, wherein said sample is obtained by SSR PCR.

5. The method of claim 1, wherein said sample is obtained by VNTR PCR.

6. The method of claim 1, further comprising the step of normalizing the measured peak intensities of said fragments before classifying said peak intensities according to said discrete intensity level scale.

7. The method of claim 6, wherein said discrete intensity level scale comprises at least five discrete peak levels.

8. The method of claim 1, wherein the step of aligning comprises the steps of:

grouping fragment sequences of said fragments into clusters, wherein adjacent peaks of each cluster are less

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than or equal to a predetermined number of discrete positions apart;

assigning a potential energy value to each cluster, said potential energy value being proportional to the spacing between adjacent peaks of the cluster and to the amount of displacement required to bin the peaks into discrete size bins;

varying the displacement of said peaks so that said potential energy value is minimized; and

aligning said peaks into discrete size bins according to the displacement values that result in a minimized potential energy value.

9. The method of claim 1, wherein said step of entering comprises the step of creating a data record having:
an information field including

an identification of the source of the genomic DNA,

the method of producing DNA fragments from said source,

the spacing between successive fragments,

the starting size of said fragments, and

the ending size of said fragments; and

a sequence field containing a sequence of said classified peak intensities.

10. The method of claim 9, further comprising the step of scoring a comparison between two data records, including the steps of assigning reward points to matches of peak intensities and penalty points to mismatches of peak intensities, based on the relative magnitude of said peak intensities, comparing said two data records for identical matches at corresponding size positions, comparing said two data records for identical matches at adjacent size positions, comparing said two data records for non-

identical matches at corresponding size positions, and comparing said two data records for non-identical matches at adjacent size positions, and totaling said reward points and penalty points according to found matches and remaining mismatches to obtain a score.

11. The method of claim 10, further comprising the step of obtaining a percentage metric for two compared data records, by obtaining a ratio of said score to a score obtained by matching one of said two data records to itself.

12. A computer program product, comprising:

a computer-readable medium having computer-executable code recorded thereon for obtaining DNA fingerprint profile data, said computer-executable code comprising:

means for measuring peak intensity and size of each DNA fragment in a sample of genomic DNA;

means for classifying the peak intensities of said fragments according to a predetermined discrete intensity level scale;

means for aligning the sizes of said fragments into corresponding ones of discrete size bins;

means for entering the classified peak intensities of said fragments into a data record in a sequence determined by said aligned sizes; and

means storing the record in a computer-readable storage medium.

13. The computer program product of claim 12, further comprising:

means for normalizing the measured peak intensities of said fragments before classifying said peak intensities according to said discrete intensity level scale.

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14. The computer program product of claim 13, wherein said discrete intensity level scale comprises at least five discrete peak levels.

15. The computer program product of claim 12, wherein said means for aligning comprises:

means for grouping fragment sequences of said fragments into clusters, wherein adjacent peaks of each cluster are less than or equal to a predetermined number of discrete positions apart;

means for assigning a potential energy value to each cluster, said potential energy value being proportional to the spacing between adjacent peaks of the cluster and to the amount of displacement required to bin the peaks into discrete size bins;

means for varying the displacement of said peaks so that said potential energy value is minimized; and

means for aligning said peaks into discrete size bins according to the displacement values that result in a minimized potential energy value.

16. The computer program product of claim 12, wherein said means for entering comprises means for creating a data record having:

an information field including

an identification of the source of the genomic DNA,

the method of producing DNA fragments from said source,

the spacing between successive fragments,

the starting size of said fragments, and

the ending size of said fragments; and

a sequence field containing a sequence of said classified peak intensities.

17. A DNA fingerprint data record stored in a computer-readable storage medium, said DNA fingerprint data including size and peak intensity of DNA fragments produced from a DNA sample obtained from a source of genomic DNA, said data record comprising:

an information field including

an identification of said source of genomic DNA,

the method of producing DNA fragments from said source,

the spacing between successive fragments,

the starting size of said fragments, and

the ending size of said fragments; and

a sequence field containing a sequence of classified peak intensities.