IMMUNOGENIC PEPTIDE COMPOSITION FOR THE PREVENTION AND TREATMENT OF ALZHEIMER'S DISEASE

FIELD OF THE INVENTION

[0001] The present invention relates to a composition comprsing a peptide immunogen useful for the prevention and treatment of Alzheimer's Disease. More particularly, the peptide immunogen comprises a main functional/regulatory site, an N-terminal fragment of Amyloid β (Aβ) peptide linked to a helper T cell epitope (Th) having multiple class II MHC binding motifs. The peptide immunogen elicit a site-directed immune response against the main functional/regulatory site of the Aβ peptide and generate antibodies, which are highly cross-reactive to the soluble $A\beta_{1-42}$ peptide and the amyloid plaques formed in the brain of Alzheimer's Disease patients. The antibodies elicited being cross reactive to the soluble $A\beta_{1-42}$ peptide, promote fibril disaggregation and inhibit fibrillar aggregation leading to immunoneutralization of the "soluble Aβ-derived toxins"; and being cross-reactive to the amyloid plaques, accelerate the clearance of these plaques from the brain. Thus, the composition of the invention comprising the peptide immunogen is useful for the prevention and treatment of Alzheimer's Disease.

BACKGROUND OF THE INVENTION

[0002] Alzheimer's Disease (AD) is a chronic, neurodegenerative disorder characterized by a loss of cognitive ability and severe behavoral abnormalities in a patient leading to the eventual death of the patient. There are currently 2.5 to 4.0 million AD patients in the U.S. and 17 to 25 million worldwide. It is the fourth leading cause of death in Western cultures, preceded only by heart disease, cancer, and stroke. ARICEPT®, an acetylcholinesterase inhibitor has been approved by the FDA for decelerating the rate of decline of Alzheimer patients. However, it is effective only for a limited period of time and in some patients. Up to the present there is no definitive treatment or cure for this devastating disease.

[0003] Two microscopic deposits, i.e., neurofibrillary tangles (NFT) and senile amyloid plaques, were identified by Alois Alzheimer as the pathologic hallmarks of the disease. The neurofibrillary tangles consist of two 10 nm wide filaments twisted around each other, referred to as paired helical filaments (PHFs), a major component of which is phosphorylated tau. The phosphorylation of serine at amino acid 262 of the tau protein represents a crucial step leading to physiological dysfunction of tau. PHFs are intracellular and are found in many of the abnormal dendritic and axonal processes, or neurites that make up the periphery of senile amyloid plaques. The senile amyloid plaques consist of disorganized neurophil filaments in an area of up to 150 μm in cross section with an extra-cellular core of amyloid deposit. The cerebral amyloid plaques are ultrastructurally distinct from PHFs and consist of 4-8 nm wide filaments that are not wound together in pairs. The plaque core consists of aggregates of a peptide, initially referred to as A4, with a relative molecular mass (*M*) of about 4,000 (Masters *et a1., Proc Natl Acad Sci USA*, 1985, 82:4245-4249).

[0004] A partial amino acid sequence of A4, now renamed amyloid β peptide (or $A\beta_{1-42}$), shows that it is similar to the amyloid β protein isolated from cerebral blood vessels of patients with Alzheimer's disease or Down's syndrome (Glenner and Wong, *Biochem Biophys Res Comm*, 1984; 120:885-890; 122:1131-1135).

[0005] $A\beta_{1-42}$ has been hypothesized to be related to AD for a number of reasons. Firstly, in peripheral amyloidoses, e.g., primary light chain disease or secondary AA amyloidosis, large amyloid burdens strongly correlate with tissue and organ dysfunction. Secondly, amyloid plaque density positively correlates with premortem dementia scores in AD. Thirdly, $A\beta_{1-42}$ deposition is the earliest neuropathological marker in AD and related disorders such as Down's syndrome, where it can precede NFT formation by 2-3 decades. Fourthly, β-amyloidosis is relatively specific to AD and related disorders. Fifthly, $A\beta_{1-42}$ is toxic to neurons (Yankner et al., *Science*, 1990; 250:279-282). Lastly, missense mutations in the

structural amyloid precursor protein (APP) gene cause early onset of familial AD (Goate *et al.*, *Nature*, 1991; 349:704-706; Mullan *et al.*, *Nature Genetics*, 1992; 1:345-347). Notably, one such mutation causes dramatic $A\beta_{1-42}$ overproduction (Citron *et al.*, *Nature*, 1992; 360:672-674).

[0006] In 1987, Kang *et al.* (*Nature*, 1987; 325:733-737) and three other groups (see 1987 status reports by Anderton, *Nature*, 1987; 325:658-659 and Barnes, *Science*, 1987; 235:846-847) independently cloned the gene from which $A\beta_{1-42}$ is derived. This gene, now known as the amyloid precursor protein (APP), encodes a protein of 695 amino-acid residues with a MW of about 79,000 that is expressed in virtually all tissues. There are at least five splicing variants of APP, four of which contain the β-amyloid peptide sequence.

[0007] Four genes have been implicated in familial forms of AD. Three of the genes, βAPP, *presenilin I*, and *presenilin* 2, when mutated, cause autosomal dominant early forms of AD. The fourth gene, Apolipoprotein E, has a naturally occurring polymorphic form, ApoE4, that represents a major genetic risk factor for the development of the disease. The concept that alterations in several distinct genes can lead to a chronic imbalance between $A\beta_{1-42}$ production and its clearance, with the resulting aggregation of first the 42-residue and then the 40-residue peptide into cytotoxic plaques, is supported by available evidence. The evidence strongly suggests that defects in each of these four genes predispose the AD phenotype by (1) enhancing the production and/or the deposition of $A\beta_{1-42}$ peptides or (2) by decreasing the clearance of ApoE4 from tissue (Selkoe, *J Biol Chem*, 1996; 271:18295-18298).

[0008] From available data, it appears that aggregated but not monomeric $A\beta_{1-42}$ peptides can induce cell dysfunction and death *in vitro* by a range of presumably interrelated mechanisms. These include oxidative injury (Thomas et al., *Nature*, 1996; 380:168-171; Behl *et al.*, *Cell*, 1994; 77:817-827), alterations in intracellular calcium homeostasis (Arispe et al., *Proc Natl Acad Sci USA*, 1993; 90:567-571), and cytoskeletal reorganization (Busciglio et al., *Neuron*, 1995;

14:879-888). Sufficient knowledge of some of the principal steps in the amyloid-induced cascade has emerged, even though the cascade hypothesis is hotly contested.

[0009] Pharmalogical approaches of identifying small molecules which could inhibit one or another step of the amyloid induced cascade is now well under way. Of particular interest are two approaches: attempts to interfere with the aggregation of $A\beta_{1-42}$ peptides by decreasing the secretion of $A\beta_{1-42}$ peptides from neuronal and glial cells or inhibit the toxicity that these extracellular aggregates produce on neurons and glial cells and their processes. A third approach attempts to control the specialized inflammatory response that appears to be triggered by aggregated $A\beta_{1-42}$ (including microglial stimulation, activation of the classical complement cascade, cytokine release, and reactive astrocytosis) may prove to be of benefit to Alzheimer patients.

[0010] Aside from the above-mentioned pharmacological approaches for AD intervention, immunological interventions have also been attempted. Soloman *et al.* (*Proc Natl Acad. Sci,* 1996; 93:452-455; *Proc Natl Aca. Sci*, 1997; 94:4109-4112) showed that three specific monoclonal antibodies, directed toward a site in the N-terminal region of the human $A\beta_{1-42}$ peptide, bind in varying degrees to preformed fibrils leading to their disaggregation and inhibition of their neurotoxic effect. The antibodies were also found to prevent the formation of fibrillar $A\beta_{1-42}$. Solomon et al. (WO 01/18169) also attempted to prepare a phage display of an epitope of the $A\beta_{1-42}$ peptide and administering the phage displayed epitope or peptide containing the epotope intraperitonially to mice to elicit antibodies to the $A\beta_{1-42}$ peptide. *In vitro* testing with rat phenochromocytoma showed that a 1:5 dilution of the anitsera prevented the neurotoxicity of $A\beta_{1-42}$. The antiserum at a dilution of 1:5 and 1:20 was also shown to disrupt the fibril structure of Aβ *in vitro* with extensive deterioration of fibril morphology. However, the adjuvant used was for the first injection was Complete Freund's Adjuvant with the incomplete Freund's Adjuvant for the second injection. The adjuvants used are entirely

unsuitable for use in humans. Moreover, the levels of antibodies generated were too low to be effective despite the use of these harsh adjuvants.

[0011] Subsequently, Schenk *et al.* (*Nature*, 1999; 400:173-177) showed that immunization with $A\beta_{1-42}$ peptide inhibits the formation of amyloid plaques and the associated dystrophic neurites in a mouse model of AD. However, due to the low immunogenicity of the $A\beta_{1-42}$ peptide, the method employed required repeated administrations of the antigen with a harsh lesion-forming adjuvant to obtain the higher levels of anti- $A\beta_{1-42}$ plaque antibodies necessary to affect plaque formation. Moreover, it was cautioned that immunization with $A\beta_{1-42}$ might induce more accumulation of the toxic amyloid itself (Araujo, DM & Cotman, CW, *Brain Res*, 1992; 569, 141-145).

[0012] Despite these criticisms, additional studies in transgenic AD mice models through similar active immunization have lent credence to the immunoprophylaxis and immunotherapeutic approaches for AD. Janus et al. (*Nature*, 2000; 408:979-982) described $A\beta_{1-42}$ peptide immunization in a mouse model for AD that reduced behavior impairment and plaques. Morgan et al. (*Nature*, 2000; 408:982-985) described $A\beta_{1-42}$ peptide vaccination to prevent memory loss in the mouse model.

[0013] Direct support for the effectiveness of immune therapy came from the observation that peripheral administration of antibodies, monoclonal or polyclonal, against Aβ-peptide reduced amyloid burden (WO 99/27944; Bard *et al.*, *Nature Medicine*, 2000; 6:916-919). Despite relatively modest serum levels, these passively administered antibodies, monoclonal 3D6 (anti-$A\beta_{1-5}$) and 10D5 (anti-$A\beta_{1-12}$) or polyclonal anti- $A\beta_{1-42}$, were able to enter the central nervous system. There, the antibodies bound to plaques and induced clearance of pre-existing amyloid plaques. Bard *et al.*, reported that when examined in an *ex vivo* assay with brain sections of PDAPP mice (i.e., mice transgenic for an APP mini-gene driven by a platelet-derived growth factor promoter) or AD patient brain

tissue, antibodies against Aβ-peptide triggered microglial cells to clear plaques through Fc receptor-mediated phagocytosis and subsequent peptide degradation. This study demonstrated that passively administered antibodies against $A\beta_{1-42}$ peptide and the $A\beta_{1-42}$ N-terminus region reduced the extent of plaque deposition in a mouse model of AD; and that monoclonal antibodies or polyclonal antibodies elicited by site-directed vaccines are able to enter the CNS at therapeutically relevant levels.

[0014] Despite the promising findings of immunological intervention in mice model for AD, a vaccine against AD suitable for humans remains a long way off (Chapman, *Nature*, 2000; 408:915-916). The principal hurdles reside in the extensive work necessary to design and formulate an immunogenic composition that is uselful in humans before a practicable vaccine for AD can be achieved. Some of the issues that rely on experimental data for guidance are: (1) What is the specific target site for antibody recognition within the Aβ? (2) In what form should the immunogen be presented? (3) What other sites need to be included before an immunogen is achieved that will elicit a therapeutic level of antibody? (4) What is an effective vaccine delivery system employing a clinically acceptable adjuvant for humans?.

[0015] A major gap exists between what has been disclosed in the literature and what remains to be done. What is the suitable specific target site (i.e., the polymerized $A\beta_{1-42}$ plaque or the monomeric soluble $A\beta_{1-42}$ peptide) and how the specific site is to be engineered for immunological intervention. In spite of some 5,000 publications on $A\beta_{1-42}$ over the past decade, the amyloid cascade hypothesis is hotly debated and the issue: the form in which $A\beta_{1-42}$ should be used for intervention remains contentious. At the heart of the problem, argued by Terry and colleagues, is the weak correlation between fibrillar amyloid load and measures of neurological dysfunction (The Neuropathology of Alzheimer Disease and the Structure Basis of its Alterations, Ed. by Terry et al., Alzheimer Disease, p187-206, Lippincott Williams and Wilkins, 1999).

[0016] In AD patients, amyloid deposits often form at a distance from the site of neuron damage. The best correlation with pathological dementia is loss of synaptic terminals. However, the loss of synaptic terminals correlates poorly with amyloid load. If the manifestations of disease correlate weakly with amyloid load, then what is the role of Aβ? The article by Klein et al, titled "Targeting small $A\beta_{1-42}$ oligomers: the solution to an Alzheimer's disease conumdrum?"(*Trends in Neurosciences*, 2001; 24:219-224) suggests that fibrils are not the only toxic form of Aβ, and perhaps not the neurotoxin that is most relevant to AD. Small oligomers and protofibrils, also termed as $A\beta_{1-42}$ derived diffusible ligands (ADDLs), may also have potent toxic neurological activity.

[0017] An AD vaccine for successful immunological intervention will require an immunogen designed to elicit site-directed high affinity antibodies that bind to the senile plaques in the brain tissue to accelerate the clearance of the plaque by the Glial cells, and immunoneutralize the soluble Aβ-derived toxins.

[0018] The problem of raising high affinity site-directed antibodies against poorly immunogenic site-specific peptides have been known for decades. Immunologists and vaccinologists often resort to the classical hapten [peptide]-carrier protein conjugate approach as demonstrated in WO 99/27944. For the development of a site-directed vaccine against AD, Frenkel et al. attempted immunization against $A\beta_{1-42}$ plaques through "EFRH"-phage administration (*Proc Natl Acad. Sci* 2000; 97:11455-11459, WO 01/18169) as mentioned above.

[0019] The approaches: using $A\beta_{1-42}$ peptide aggregate or $A\beta_{1-42}$ peptide fragment-carrier protein conjugates (WO99/27944) and using filamentous phage displaying "EFRH peptide" as the agents to induce immune responses against an amyloid deposit in a patient, are cumbersome and ineffective. For example, after the fourth immunization of 10^{11} phages displaying the EFRH epitope, >95% of the antibodies in the guinea pig immune sera are against the phages. Only a small population (<5%) of the antibodies is against the soluble $A\beta_{1-42}$ peptide (Frenkel et al., Vaccine 2001, 19:2615-2619; WO 01/18169).

[0020] Less cumbersome methods were described in EP 526,511 and WO 99/27944, which disclosed the administration of $A\beta_{1-42}$ peptide to treat patients with pre-established AD and the administration of $A\beta_{1-42}$ or other immunogens to a patient under conditions that generate a "beneficial" immune response in the AD patient. However, a review of WO99/27944 show that there are major deficiencies in the vaccine design disclosed therein.

[0021] In particular, the problem lies in the lack of a pharmaceutically acceptable and effective vaccine delivery system. WO99/27944 disclosed $A\beta_{1-42}$ or active fragments of $A\beta_{1-42}$ conjugated to a carrier molecule such as cholera toxin as the active vaccine component. See page 4 of WO 99/27944. Although page 5 taught that a pharmaceutical composition comprising the immunogen should be free of Complete Freund's Adjuvant [CFA], the only examples showing the efficacy of the $A\beta_{1-42}$ vaccine for the treatment of AD in transgenic mice employed large doses of aggregated $A\beta_{42}$ peptide in CFA. Despite repetitive recital of preferred adjuvants that are to be used with the disclosed immunogenic agents to enhance the immune response, experimental data showed that only the formulations employing CFA/ICFA provided a sufficiently high titer of anitbodies. See, page 25 of WO 99/27944. In example 1, the prophylactic efficacy of $A\beta_{1-42}$ against AD was demonstrated in PDAPP mice. However, the formulations administered consist a dose of 100ug per mouse of aggregated $A\beta_{42}$ emulsified in Complete Freund's Adjuvant [CFA] (p34 of WO 99/27944) followed by multiple booster doses of the same $A\beta_{1-42}$ peptide emulsified in Incomplete Freund's Adjuvant. In Example IX, the immune responses in mice to different adjuvants were studied. When the adjuvants: MPL, Alum, QS21, and CFA/ICFA were used with the purportedly potent immunogen AN1792 (i.e., aggregated human $A\beta_{42}$), the level of antibodies to $A\beta_{1-4}$ were reduced at a statistically significant level in comparison to mice that received the CFA/ICFA vaccines. See, Table 9, and pages 59-64 of WO 99/27944.

[0022] In the case where $A\beta_{1-42}$ peptide fragments were used (human $A\beta_{1-42}$ peptides of amino acids 1-5, 1-12, 13-28, and 33-42), each fragment was conjugated to sheep anti-mouse IgG as the protein carrier. In a later disclosure, the efficacy of antibodies to Aβ peptide fragments could only be shown by passive immunization with monoclonal antibodies (Bard *et al.*, *Nature Medicine* 2000; 6:916-919). The efficacy of these fragments conjugated to sheep anti-mouse IgG was not shown. Therefore, the only immunogen shown to be effective was the aggregated $A\beta_{1-42}$ peptide in CFA/ICFA.

[0023] Up to the present, all of the vaccine formulations shown to be effective employed CFA/IFA as the adjuvant. Peptide immunogens targeting $A\beta_{1-42}$ have thus far been prepared by conjugation of the various $A\beta_{1-42}$ fragments to sheep anti-mouse immunoglobulin, conjugation of synthetic $A\beta_{13-28}$ via *m*-maleimidobenzoyl-N-hydroxysuccinimide ester to anti-CD3 antibody, or aggregated $A\beta_{1-42}$ peptide alone. These immunogens, i.e., $A\beta_{42}$ peptide alone or $A\beta_{1-42}$ peptide-carrier protein conjugates, were emulsified with complete Freund's adjuvant for the first immunization, followed by subsequent boosts in incomplete Freund's adjuvant (Johnson-Wood et al., *Proc Natl Acad Sci USA*, 1997; 94:1550-1555; Seubert et al., *Nature*, 1992; 359:325-327; Schenk et al., *Nature*, 1999; 400: 173-177; Janus et al., *Nature* 2000; 408:979-982; and Morgan et al., *Nature*, 2000; 408:982-985). The formulations disclosed in WO 99/27944 or others using CFA and ICFA as adjuvants causes lesions and are too harsh for use in humans. Thus, none of the vaccine compositions for AD described in the prior art are suitable for use in humans.

[0024] In summary, despite statements suggesting the potential of $A\beta_{1-42}$ peptide for the treatment of AD in view of the previous disclosures of Kline (EP 526,511), no problem solving vaccine formulations were really offered in WO99/27944 to address this key problem.

[0025] Another disadvantage with the peptide-carrier protein conjugates and $A\beta_{1-42}$ aggregates is that these molecules are highly complex and are difficult to characterize and it is difficult to develop effective quality control procedures for the manufacturing process. A further disadvantage is that, $A\beta_{1-42}$ peptide or its fragments are self molecules when administered to humans. Therefore, they are less immunogenic or non-immunogenic in humans. It is, thus, necessary to develop clinically acceptable vaccine formulations for administration in humans.

[0026] It is known that promiscuous Th epitopes may be employed to evoke efficient T cell help and may be combined with poorly immunogenic B cell epitopes to provide potent immunogens. Well-designed promiscuous Th/B cell epitopes chimeric peptides have been shown to be useful in eliciting Th responses and resultant antibody responses in most members of a genetically diverse population expressing diverse MHC haplotypes. Promiscuous Th from a number of pathogens, such as measles virus F protein and hepatitis B virus surface antigen, are known. Tables 1 and 2 lists many of the known promiscuous Th that have been shown to be effective in potentiating a short poorly immunogenic peptide, the decapeptide hormone LHRH (US 5,759,551, and 6,025,468).

[0027] Potent Th epitopes range in size from approximately 15-40 amino acid residues in length, often share common structural features, and may contain specific landmark sequences. For example, a common feature of a Th is that it contains amphipathic helices, alpha-helical structures with hydrophobic amino acid residues dominating one face of the helix and with charged and polar residues dominating the surrounding faces (Cease et al., *Proc Natl Acad Sci USA*, 1987; 84: 4249-4253). Th epitopes frequently contain additional primary amino acid patterns such as a Gly or charged residue followed by two to three hydrophobic residues, followed in turn by a charged or polar residue. This pattern defines what are called Rothbard sequences. Th epitopes often obey the 1, 4, 5, 8 rule, where a positively charged residue is followed by hydrophobic residues at the fourth, fifth and eighth positions after the charged residue. Since all of these structures are composed of common hydrophobic, charged and polar amino

acids, each structure can exist simultaneously within a single Th epitope (Partidos et al., *J Gen Virol*, 1991; 72:1293). Most, if not all, of the promiscuous T cell epitopes fit at least one of the periodicities described above. These features may be incorporated into the designs of idealized artificial Th sites, including combinatorial Th epitopes. With respect to the design of combinatorial Th sites, lists of variable positions and preferred amino acids are available for MHC-binding motifs (Meister et al., *Vaccine*, 1995; 13:581-591). Furthermore, a method for producing combinatorial Th has been disclosed for combinatorial library peptides termed structured synthetic antigen library (Wang et al., WO 95/11998). Thus, the 1, 4, 5, 8 rule can be applied together with known combinatorial MHC-binding motifs to assign invariant and degenerate positions in a combinatorial Th site, and to select residues for the degenerate sites to vastly enlarge the range of immune responsiveness of an artificial Th. See, Table 2, WO 99/66957, and WO 95/11998.

[0028] Wang *et al.* (US 5,759,551) suggested the use of immunostimulatory elements to render the self protein Amylin immunogenic. Wang *et al.* suggested the administration of immunogenic synthetic amylin peptides as vaccines for the treatment of non-insulin dependent diabetes mellitus (NIDDM), an amyloidogenic disease caused by overproduction of Amylin (column 19, lines 9-39, US 5,759,551). Amylin is a 37 amino acid residues peptide hormone produced by the β cells in the islets of Langerhans. Overproduction of Amylin will result in the depositon of insoluble amyloid leading to amyloidogenic disease in the pancreas. Similar to the overproduction of Amylin, overproduction of the $A\beta_{1-42}$ peptide will lead to the deposition of insoluble amyloid in the brain of AD patients. However, there is limited sequence homology between Amylin and the $A\beta_{1-42}$ peptide. Only a short stretch of amino acids residues, VGSN, of $Amylin_{32-35}$ corresponds to $A\beta_{24-27}$. Antibodies produced against the Amylin peptide is not expected to be cross reactive to soluble $A\beta_{1-42}$ peptides nor accelerate the clearance of amyloid plaques in the brain in view of the studies by Soloman *et al.* and Schenk *et al.*, which showed that the sequence EFRH is critical.

[0029] It is the object of the invention to develop an immunogen that will enable the generation of high levels of high affinity antibodies against the N-terminal functional site of the $A\beta_{1-42}$ peptide with high cross-reactivity to the senile plaques in the brain of AD patients. The anitbodies generated by binding to the $A\beta_{1-42}$ peptide and the senile plaques is expected to accelerate the clearance of these plaques from the brain, promote fibril disaggregation, inhibit fibrillar aggregation, and immunoneutralization of the "soluble Aβ-derived toxins" [also termed as Aβ-derived diffusible ligands or ADDLs].

[0030] It is a further objective of the present invention to develop a vaccine delivery vehicle that is suitable for human or veterinary use for the prophylaxis and treatment of Alzheimer's Disease.

BRIEF DESCRIPTION OF THE DRAWINGS

[0031] Figures 1a, 1b, 1c, 1d, 1e and 1f are photographs showing Immunoperoxidase staining of serial sections from 2 AD brains, using Avidin-Biotinylated Antibody Complex (ABC) method with immune and preimmune sera at 1:100 dilution under 10X magnification. Figures 1a, and 1d show significant binding of antibodies to both senile plaques and Aβ plaques (both labelled as "P") on thioflavine S positive blood vessels (labelled as "BV"). The antibodies were generated in guinea pigs using $A\beta_{1-28}$-εK-MVF Th1-16 (SEQ ID NO:74) prepared in ISA51 water-in-oil emulsion. Figures 1b and 1e show the cross reactivity of antibodies raised against the same peptide immunogen in CFA/ICFA. Figures 1c and 1f show brain sections using preimmune sera.

[0032] Figures 2a, 2b, 2c, 2d, and 2e are photographs showing Immunoperoxidase staining of serial sections of AD brain with immune and preimmune sera at 1:100 dilution and under 40X magnification. Figures 2a and 2d showed that the antibodies in guinea pigs immunized with $A\beta_{1-28}$-εK-MVF Th1-16 (SEQ ID NO:74) prepared in ISA51 water-in-oil emulsion strongly stained the plaques (P) forming a pattern of cores. Figure 2b is a photograph of the staining

pattern of AD pig brain sections using the same immunogen in CFA/ICFA formulation. The anti-sera reacted predominantly with plaques on the blood vessels (BV). Figure 2c is a photograph of a guinea pig brain section with preimmune serum and showed no staining. Figure 2e shows the brain section with hyperimmune sera generated by immunization with $A\beta_{1-28}$ peptide alone in CFA/ICFA showing a surprisingly weak staining pattern despite the strong reactivity with $A\beta_{1-28}$ by ELISA.

BRIEF DESCRIPTION OF THE INVENTION

A first aspect of the present invention provides a peptide immunogen of about 20 to 100 amino acids long comprising: (i) a helper T cell (Th) epitope selected from the group consisting of SEQ ID Nos: 1 to 64; (ii) an N-terminal fragment of $A\beta_{1-42}$ peptide, SEQ ID NO:65, consisting of from 10 to 28 amino acid residues wherein each fragment comprises amino acid residue 1 of the $A\beta_{1-42}$ peptide or an immunologically functional analog of the N-terminal fragment of $A\beta_{1-42}$ peptide; and (iii) optionally a spacer consisting of at least an amino acid to separate the immunogenic domains.

A second aspect of the present invention provides a peptide immunogen selected from the group consisting of SEQ ID Nos: 70, 71, 72, 73, and 74.

A third aspect of the present invention provides the peptide immunogen represented by one of the following formulae: $(A)_n$(N-terminal fragment of $A\beta_{1-42}$ peptide)-$(B)_o$-$(Th)_m$-X; or $(A)_n$-$(Th)_m$-$(B)_o$-(N-terminal fragment of $A\beta_{1-42}$ peptide)-X; wherein each A is independently an amino acid; each B is a linking group selected from the group consisting of an amino acid, gly-gly, (α, ε-N)-Lys, and Pro-Pro-Xaa-Pro-Xaa-Pro (SEQ ID NO:73); Th comprise an amino acid sequence that constitutes a helper T cell epitope, selected from the group consisting of SEQ ID Nos: 1-64 and an immune enhancing analog thereof; (N-terminal fragment of $A\beta_{1-42}$ peptide) is 10 to about 28 amino acid residues and

wherein each fragment comprises EFRH of the $A\beta_{1-42}$ peptide and immunologically functional analog thereof; X is an α-COOH or α-$CONH_2$ of an amino acid; n is from 0 to about 10; m is from 1 to about 4; and o is from 0 to about 10.

A fourth aspect of the present invention provides a composition comprising a pepetide immunogen of the first aspect and a pharmaceutically acceptable adjuvant and/or carrier selected from the group consisting of alum, liposyn, saponin, squalene, L121, emulsigen monophosphyryl lipid A (MPL), polysorbate 80, QS21, Montanide ISA51, ISA35, ISA206 and ISA 720.

A fifth aspect of the present invention provides the use of a composition of the fourth aspect, for the manufacture of a medicament for preventing or treating Alzheimer's disease.

A sixth aspect of the present invention provides the use of a composition of the fourth aspect, for the manufacture of a medicament for producing antibodies to $A\beta_{1-42}$ peptide that is cross reactive to soluble Aβ peptides and brain tissue plaques formed therefrom.

[0031] An embodiment provides an immunogenic composition comprising synthetic peptides capable of inducing antibodies against the main functional/regulatory site of the Aβ peptide with high cross-reactivity both to the soluble $A\beta_{1-42}$ peptide and the plaques in the brain of Alzheimer's Disease (AD) patients. The immunogenic composition when administered to an AD patient or a person predisposed to AD is expected to accelerate the clearance of amyloid plaques and immunoneutralization of the soluble Aβ derived toxins in the brain to prevent and treat AD. In particular, a peptide immunogen of disclosed comprise a Th epitope selected from the group consisting of SEQ ID NOS: 1-64 and the immunologically functional analogs thereof linked to a short N-terminal $A\beta_{1-42}$ peptide fragment selected from the group consisting of 10 to 28 amino acid residues comprising

EFRH of the $A\beta_{1-42}$ peptide, SEQ ID NO: 65, or an immunologically functional analog of the $A\beta_{1-42}$ peptide fragment. Preferably the $A\beta_{1-42}$ peptide fragment is selected from the group SEQ ID NOS: 66-69 or a immunologically functional analogs thereof.

[0032] Another embodiment provides an immunogenic composition comprising an immunologically effective amount of a peptide composition in a pharmaceutically acceptable vaccine formulation comprising an adjuvant or emulsifier selected from the group consisting of liposyn, saponin, squalene, L121, emulsigen monophosphyryl lipid A (MPL), polysorbate 80, QS21, Montanide ISA51, ISA35, ISA206 and ISA 720 as well as the other efficacious adjuvants and emulsifiers.

[0033] A further embodiment provides a method for the induction of accelerated clearance of amyloid plaques and immunoneutralization of the soluble Aβ- derived toxins in the brain to prevent and treat Alzheimer Disease in a mammal by administering one or more of the immunogenic peptides to the mammal for a time and under conditions sufficient to induce antibodies directed against the functional/regulatory site of the $A\beta_{1-42}$ peptide. A typical example of a vaccine disclosed is a peptide composition comprising 5-1000 µg of the peptide immunogen in a vaccine formulated as a water in oil emulsion in a pharmaceutical acceptable adjuvant and/or carrier. A typical method of administering the vaccine is to inject intramuscularly the vaccine formulation at 0.5-2mL per dose on an immunization schedule of 0, 4, and 8 weeks intervals.

[0034] Yet a further embodiment provides an immunogenic synthetic peptide of about 30 to about 60 amino acids consisting of a helper T cell (Th) epitope, linked to an N-terminal $A\beta_{1-42}$ peptide fragment selected from the group consisting of 10 to 28 amino acids with each fragment comprising amino acid residue 1 of the $A\beta_{1-42}$

peptide. See SEQ ID NO: 65 wherein D, Aspartic acid, is designated as amino acid residue 1. Preferably the N terminal $A\beta_{1-42}$ peptide fragment is selected from the group SEQ ID NOs: 66-69 or a peptide analog of N-terminal fragmentof $A\beta_{1-42}$ peptide. Optionally, amino acid spacers to separate the immunogenic domains may be included. The immunogenic domain elements separated by spacers can be covalently joined in any order provided that either the immunoreactivity of the peptide hapten is substantially preserved or that immunoreactivity to the N-terminal Aβ peptide fragment, soluble $A\beta_{1-42}$ peptide, and the plaques is generated.

[0035] An important factor affecting immunogenicity of a synthetic peptide for an N-terminal $A\beta_{1-42}$ fragment immunogen is its presentation to the immune

system by T helper cell epitopes (Th). Such Th is most reliably supplied to the peptide immunogen by foreign Th epitopes placed on a separate Th peptide domain element that are extrinsic to the target Aβ peptide. Such peptide immunogens may be produced as hybrid polypeptides by recombinant DNA expression. They may also be more simply and less expensively supplied as a synthetic peptide immunogen comprising the target hapten B cell site from Aβ peptide and T-helper epitopes (Th) appropriate for the host. Such peptides react with helper T-cell receptors and the class II MHC molecules, in addition to antibody binding sites (Babbitt et al., *Nature*, 1985; 317:359) and thus stimulate a tightly site-specific antibody response to the target antibody binding site. Previously such Th was supplied for workable $A\beta_{1-42}$ peptide immunogens by Th intrinsic to aggregated full length Aβ peptide (WO 99/66957; WO 1999/27944; Janus et al., 2000, Morgan et al., 2000) and can be supplied by carrier protein. A wholly synthetic peptide immunogen enjoys the following advantages over $A\beta_{1-42}$ peptide aggregates, carrier conjugates and recombinant polypeptides in that the product is chemically defined for easy quality control. The synthetic peptide immunogen is stable. No elaborate downstream processing nor an elaborate manufacturing facility is needed. The immune response is site-specific and focused on the Aβ target and not the carrier. Thus, undesirable responses such as epitopic suppression are avoided.

[0036] Immunogenicity of synthetic N-terminal functional-site directed Aβ peptide immunogens can be optimized by (1) combining N-terminal $A\beta_{1-42}$ peptide fragment with selected foreign promiscuous Th sites to which the majority of a population are responsive; and (2) combining Aβ peptide fragment with Th whose repertoire is enlarged through combinatorial chemistry, and thereby accommodate to the variable immune responsiveness of a genetically diverse population.

[0037] It has been found that peptides composition of the present invention are effective in stimulating the production of antibodies against the main

functional/regulatory site of the Aβ peptide, with cross-reactivities to the soluble $A\beta_{1-42}$ and the plaques in the brains of AD patients. Based on the immunogenicity data obtained in guinea pigs and baboons, and the data obtained from the immunoperoxidase staining of the amyloid plaques present in human AD brain sections by the spicific immune sera obtained, it is expected that the peptide immunogens of the present invention formulated appropriately are effective in humans. It is to be noted that the data obtained in baboons are particularly significant in that this is a species whose immune response closely resemble those of humans.

DETAILED DESCRIPTION OF THE INVENTION

[0038] This invention is directed to a novel peptide composition for the generation of high titer polyclonal antibodies with specificity for the main functional/regulatory site of the Aβ peptide, with cross-reactivities to the soluble $A\beta_{1-42}$ and the plaques in the brain of Alzheimer Disease (AD) patients. The antibodies generated by the peptide composition are highly site-specific and bind to the Aβ peptides and to amyloids plaques in the brain. Thus, the present invention provides an effective method for accelerating the clearance of amyloid plaques and immunoneutralization of soluble Aβ derived toxins in the brains for the prevention and treatment of AD.

[0039] N-terminal $A\beta_{1-42}$ peptide fragments selected from the group consisting of 10 to 28 amino acids wherein each fragment comprises EFRH of the $A\beta_{1-42}$ peptide (SEQ ID NO:65), are short linear peptides which, by themselves are non-immunogenic. The short $A\beta_{1-42}$ peptide fragments can be immuno-potentiated by chemical coupling to a carrier protein, for example, keyhole limpet hemocyanin (KLH) or by fusion to a carrier polypeptide through recombinant DNA expression, for example, hepatitis B surface antigen. The deficiency of such "Aβ peptide(s)-carrier protein" vaccines is that a major portion of antibodies generated are non-functional antibodies directed against the carrier protein.

[0040] The immunogens of the present invention are wholly synthetic peptide immunogens comprising N-terminal fragment of $A\beta_{1-42}$ peptide of 10 to 28 amino acids with each fragment comprising EFRH of the $A\beta_{1-42}$ peptide covalently linked to promiscuous Th epitopes selected from the group consisting of SEQ ID NOs: 1 to 64. The immunogens of the invention elicit the production of site-specific antibodies which bind to the $A\beta_{1-42}$ peptide and its aggregates and are cross reactive with amyloid plaques in the brain to provide for accelerated clearance of amyloid plaques and immunoneutralization of the soluble Aβ-derived toxins in the brain. Thus, the immunogen of the present invention is useful in preventing and treating AD.

[0041] The helper T cell epitopes (Th) useful in the invention comprise multiple class II MHC binding motifs. Specific examples of Th covalently linked to an N-terminal $A\beta_{1-42}$ peptide fragment are provided. The results of anti-sera from animals immunized with the immunogen peptides of the present invention demonstrate that potent site-directed Aβ peptide reactive antibodies are generated, in a genetically diverse host population.

[0042] Generally, the synthetic immunogenic peptide of the present invention are approximately 20 to 100 amino acids long and comprise:

(i) a helper T cell (Th) epitope selected from the group consisting of SEQ ID Nos: 1 to 64;

(ii) an N-terminal fragment of $A\beta_{1-42}$ peptide from about 10 to about 28 amino acid residues wherein each fragment comprises EFRH of the $A\beta_{1-42}$ peptide; and

(iii) optionally a spacer consisting of at least an amino acid to separate the immunogenic domains.

[0043] Preferably, the N terminal fragment of the $A\beta_{1-42}$ peptide is selected from the group consisting of SEQ ID NOS: 66-69 and an immunologically effective

analog thereof. The Th peptide is covalently attached to either the N- or C-terminus of the target N-terminal fragment of $A\beta_{1-42}$ peptide optionally with a spacer (e.g., Gly-Gly, ε-N Lys).

The peptide immunogen of this invention is represented by one of the following formula:

$(A)_n$-(N-terminal fragment of $A\beta_{1-42}$ peptide)-$(B)_o$-$(Th)_m$-X;or
$(A)_n$-$(Th)_m$-$(B)_o$-(N-terminal fragment of $A\beta_{1-42}$ peptide)-X;

wherein

each A is independently an amino acid;

each B is a linking group selected from the group consisting of an amino acid, gly-gly, (α, ε-N)lys, Pro-Pro-Xaa-Pro-Xaa-Pro (SEQ ID NO:73);

Each Th comprise an amino acid sequence that constitutes a helper T cell epitope, or an immune enhancing analog or segment thereof;

(N-terminal fragment of $A\beta_{1-42}$ peptide) is a synthetic peptide B cell target site antigen and is a fragment of about 10 to about 28 amino acid residues wherein each fragment comprises EFRH of the $A\beta_{1-42}$ peptide or an immunologically functional analog thereof;

X is an α-COOH or α-$CONH_2$ of an amino acid ;

n is from 0 to about 10;
m is from 1 to about 4; and
o is from 0 to about 10.

[0044] The peptide immunogen of the present invention comprises from about 20 to about 100 amino acid residues, preferably from about 25 to about 60 amino acid residues. Preferably, the (N-terminal fragment of $A\beta_{1-42}$ peptide) is

selected from the group consisting of SEQ ID Nos: 66-69 and preferably the Th epitope is selected from the group consisting of SEQ ID NOs: 1, 3, 4, 5, 6, 7, 8, 9, 20, 38-40, 47-51 and 52-54. Preferably, m=1, n=1, and o=1 or 2.

[0045] When A is an amino acid, it is a non-naturally occurring or naturally occurring amino acid. Non-naturally occurring amino acids include, but are not limited to, ε-N lysine, ß-alanine, ornithine, norleucine, norvaline, hydroxyproline, thyroxine, γ-amino butyric acid, homoserine, citrulline and the like. Naturally-occurring amino acids include alanine, arginine, asparagine, aspartic acid, cysteine, glutamic acid, glutamine, glycine, histidine, isoleucine, leucine, lysine, methionine, phenylalanine, proline, serine, threonine, tryptophan, tyrosine and valine. when m is greater than one, and two or more of A are amino acids, then each amino acid may independently be the same or different. $(A)_n$ may include a spacer, e.g., Gly-Gly, ε-N Lys.

[0046] B is a spacer and is an amino acid which can be naturally occurring or the non-naturally occurring amino acids as described above. Each B is independently the same or different. The amino acids of B can also provide a spacer, e.g., Gly-Gly, ε-Lys, or lysine between the promiscuous Th epitope and the N-terminal fragment of $A\beta_{1-42}$ peptide (e.g., SEQ ID NOs:66-69) or an immunologically functional analog thereof. In addition by physically separating the Th epitope from the B cell epitope, i.e., the N-terminal fragments of $A\beta_{1-42}$ peptide or its immunologically functional analog, the Gly-Gly or ε-Lys spacer can disrupt any artifactual secondary structures created by the joining of the Th epitope with an N-terminal fragment of $A\beta_{1-42}$ peptide or its immunologically functional analog and thereby eliminate interference between the Th and/or B cell responses. The amino acids of B can also form a spacer which acts as a flexible hinge that enhances separation of the Th and the N-terminal fragments of $A\beta_{1-42}$ peptide. Examples of sequences encoding flexible hinges are found in the immunoglobulin heavy chain hinge region. Flexible hinge sequences are often proline rich. One particularly useful flexible hinge is provided by the sequence Pro-Pro-Xaa-Pro-

Xaa-Pro (SEQ ID NO:77), where Xaa is any amino acid, and preferably aspartic acid. The conformational separation provided by the amino acids of B permits more efficient interactions between the presented peptide immunogen and the appropriate Th cells and B cells to enhances the immune responses to the Th epitope and the antibody-eliciting epitope or their immunologically functional analogs.

[0047] Th is a sequence of amino acids (natural or non-natural amino acids) that comprises a Th epitope. A Th epitope may be a continuous or discontinuous epitope. In a discontinuous Th epitope, not every amino acid of Th is necessary. A Th epitope, or an analog or fragment thereof, is capable of enhancing or stimulating an immune response to the N-terminal fragment of $A\beta_{1-42}$ peptide. Th epitopes that are immunodominant and promiscuous are highly and broadly reactive across animal and human populations with widely divergent MHC types (Partidos *et al.*, 1991; US 5,759,551). The Th epitope of the subject peptides is about 10 to about 50 amino acids, preferably from about 10 to about 30 amino acids. When multiple Th epitopes are present (i.e., $m \geq 2$), each Th epitope may be the same or different. A Th segment comprises a contiguous portion of a Th epitope that is sufficient to enhance or stimulate an immune response to the N-terminal fragment of $A\beta_{1-42}$ peptide.

[0048] Th epitopes of the present invention include those derived from foreign pathogens including but not limited to those exemplified in Table 1 (SEQ ID Nos:1-21). Further, Th epitopes include idealized artificial Th and artificial idealized combinatorial Th disclosed in WO 99/66957 and listed here in Table 2 as SEQ ID Nos 22-64. Peptides comprising combinatorial Th are produced simultaneously in a single solid-phase peptide synthesis in tandem with the N-terminal fragment of $A\beta_{1-42}$ peptide, A and B. The Th epitopes also include immunologically functional analogs thereof, having conservative substitutions, additions, deletions and insertions therein of from one to about 10 amino acid residues as long as the Th-stimulating function has not been essentially modified.

[0049] In the synthetic peptides of this invention, the Th epitope is covalently attached through a spacer B to either the N terminus or C terminus of the N-terminal fragment of $A\beta_{1-42}$ peptide or an immunologically functional analog thereof. An immunologically functional analog of the N-terminal fragment of $A\beta_{1-42}$ peptide may comprise conservative substitutions, additions, deletions, or insertions of from one to about four amino acid residues as long as immune responses that are crossreactive with the $A\beta_{1-42}$ peptides are elicited. The conservative substitutions, additions, and insertions can be accomplished with natural or non-natural amino acids as defined above.

[0050] The preferred peptide immunogens of this invention are those comprising the N-terminal fragment of the $A\beta_{1-42}$ peptide fragments selected from the group consisting of SEQ ID NOs: 66-69 or an immunologically functional analog thereof; a spacer (e.g., Gly-Gly, ε-Lys); a Th epitope selected from the group consisting of an HB_s Th (SEQ ID NO:1); HB_c Th (SEQ ID NO:20); MVF Th (SEQ ID NOS:8, 9); PT Th (SEQ ID NOs:4, 5, 7), TT Th (SEQ ID NOs:3, 4, 6); CT Th (SEQ ID NOs:12, 21); DT Th (SEQ ID NO:13, 14), MVF Th derived artificial Th selected from the group consisting of SEQ ID Nos:38-40, 47-51); HBV Th derived artificial Th selected from the group consisting of SEQ ID NOS: 52-54. See Tables 1 and 2.

[0051] Peptide compositions which contain a cocktail of the subject peptide immunogens with two or more Th epitopes may enhance immunoefficacy in a broader population and thus provide an improved immune response to the $A\beta_{1-42}$ peptides and their fragments.

[0052] The peptide immunogens of this invention can be made by chemical synthesis methods which are well known to the ordinarily skilled artisan. See, for example, Fields et al., Chapter 3 in *Synthetic Peptides: A User's Guide*, ed. Grant, W. H. Freeman & Co., New York, NY, 1992, p. 77. Hence, peptides can be synthesized using the automated Merrifield techniques of solid phase synthesis with the α-NH_2 protected by either t-Boc or F-moc chemistry using side chain

protected amino acids on, for example, an Applied Biosystems Peptide Synthesizer Model 430A or 431. Preparation of peptide constructs comprising combinatorial library peptides for Th epitopes can be accomplished by providing a mixture of alternative amino acids for coupling at a given variable position. After complete assembly of the desired peptide immunogen, the resin is treated according to standard procedures to cleave the peptide from the resin and deblock the functional groups on the amino acid side chains. The free peptide is purified by HPLC and characterized biochemically, for example, by amino acid analysis or by sequencing. Purification and characterization methods for peptides are well known to one of ordinary skill in the art.

[0053] The immunogen of the present invention may also be prepared as a branched polymer by synthesis of the desired peptide construct directly onto a branched poly-lysyl core resin (Wang, et al., *Science*, 1991; 254:285-288).

[0054] Alternatively, the longer synthetic peptide immunogens can be synthesized by well known recombinant DNA techniques. Such techniques are provided in well-known standard manuals with detailed protocols. To construct a gene encoding a peptide of this invention, the amino acid sequence is reverse translated to obtain a nucleic acid sequence encoding the amino acid sequence, preferably with codons that are optimum for the organism in which the gene is to be expressed. Next, a synthetic gene is made, typically by synthesizing oligonucleotides which encode the peptide and any regulatory elements, if necessary. The synthetic gene is inserted in a suitable cloning vector and transfected into a host cell. The peptide is then expressed under suitable conditions appropriate for the selected expression system and host. The peptide is purified and characterized by standard methods.

[0055] The efficacy of the peptide composition of the present invention can be established by injecting an animal, for example, guinea pigs, with an immunogenic composition comprising peptides of the invention. See, Table 4, SEQ ID NOS:70-75. The humoral immune response to the N-terminal fragment of

$A\beta_{1-42}$ peptide and the soluble $A\beta_{1-42}$ peptide are monitored. A detailed description of the procedures used is provided in the Examples hereinbelow.

[0056] Another aspect of this invention provides a peptide composition comprising an immunologically effective amount of one or more of the peptide immunogens of this invention in a pharmaceutically acceptable delivery system. Accordingly, the subject peptide composition can be formulated as a vaccine using pharmaceutically acceptable adjuvants, carriers or other ingredients routinely employed in the formulation of vaccines. Among the ingredients that can be used in this invention are adjuvants or emulsifiers including alum, liposyn, saponin, squalene, L121, emulsigen monophosphyryl lipid A (MPL), polysorbate 80, QS21, Montanide ISA51, ISA35, ISA206 and ISA 720 as well as the other efficacious adjuvants and emulsifiers. The composition may be formulated for immediate release or sustained release. The composition may also be formulated for induction of systemic immunity, e.g., by entrapment in or coadministration with microparticles. Such formulations are readily available to one of ordinary skill in the art.

[0057] The immunogens of the present invention can be administered via any conventional route, such as subcutaneous, oral, intramuscular, parenteral or enteral route. The immunogens can be administered in a single dose or in multiple doses. A suitable immunization schedule is readily determined and available to one of ordinary skill in the art.

[0058] The peptide composition of the present invention comprises an effective amount of one or more of the peptide immunogens of the present invention and a pharmaceutically acceptable carrier. Such a composition in a suitable dosage unit form generally contains about 0.25 μg to about 500 μg of the immunogen per kg body weight. When delivered in multiple doses, the effective amount may be conveniently divided per dosage unit. For example, an initial dose, e.g. 0.0025-0.5 mg per kg body weight; preferably 1-50 μg per kg of body weight of the peptide immunogen is to be administered by injection, preferably

intramuscularly, followed by repeat (booster) doses of a similar amount. Dosage will depend on the age, weight and general health of the subject as is well known in the vaccine and therapeutic arts.

[0059] The immune response of the synthetic $A\beta_{1-42}$ peptide immunogens can be improved by delivery through entrapment in or on biodegradable microparticles of the type described by O'Hagan *et al.* (*Vaccine*, 1991; 9: 768-771). The immunogens can be encapsulated with or without an adjuvant in biodegradable microparticles, to potentiate immune responses, and to provide time-controlled release for sustained or periodic responses, and for oral administration, (O'Hagan *et al.*, 1991; and, Eldridge *et al.*, 1991; 28: 287-294).

[0060] The following examples are provided to illustrate the invention. The scope of the invention is not to be limited to the specific peptide immunogens and compositions provided. The examples demonstrate that the peptide immunogens of the present invention are useful for eliciting site-directed antibodies to both $A\beta_{1-10}$ and $A\beta_{1-14}$ fragments as well as cross-reactive antibodies to soluble $A\beta_{1-42}$ peptides as early as 4 weeks after the initial immunization.

EXAMPLE 1

TYPICAL METHODS TO SYNTHESIZE Aβ PEPTIDE IMMUNOGENS OF THE PRESENT INVENTION

[0061] Peptide immunogens listed in Table 4 (SEQ ID NOS:70-76) were synthesized individually by the Merrifield solid-phase synthesis technique on Applied Biosystems automated peptide synthesizers (Models 430, 431 and 433A) using Fmoc chemistry. Preparation of peptide immunogens comprising a combinatorial library Th, i.e., idealized artificial Th site such as MvF derived Th1-8 (SEQ ID NOs:38-40), can be accomplished by providing a mixture of the desired amino acids for chemical coupling at a given position as specified in the design. After complete assembly of the desired peptide, the resin was treated according to standard procedure using trifluoroacetic acid to cleave the peptide from the resin and deblock the protecting groups on the amino acid side chains. The cleaved,

extracted and washed peptides were purified by HPLC and characterized by mass spectrometry and reverse phase HPLC.

EXAMPLE 2

EVALUATION OF THE IMMUNOGENICITY OF THE Aβ PEPTIDE IMMUNOGENS OF THE PRESENT INVENTION

[0062] Aβ-derived peptide immunogens were evaluated on groups of guinea pigs as specified by the experimental immunization protocol outlined below and by serological assays for determination of immunogenicity.

Standard Experimental Design:

Immunogens: (1) individual peptide immunogen; or

(2) a mixture of equal molar peptide immunogens as specified in each example.

Dose: 100 μg in 0.5 mL per immunization unless otherwise specified

Route: intramuscular unless otherwise specified

Adjuvants: Complete Freund's Adjuvant (CFA)/ Incomplete Adjuvant (IFA); or water in oil emulsions unless otherwise specified. CFA/IFA groups received CFA week 0, IFA in subsequent weeks.

Dose Schedule: 0, 3, and 6 weeks or otherwise specified.

Bleed Schedule: weeks 0, 5, 8 or otherwise specified

Species: Duncan-Hartley guinea pigs or otherwise specified

Assay: Specific ELISAs for each immune serum's anti-peptide activity. The Solid phase substrate was the Aβ peptide fragment e.g. $A\beta_{1-14}$ or full length $A\beta_{1-42}$ (SEQ ID NOs: 67 and 65). Blood was

collected and processed into serum, and stored prior to ELISA with the target peptides.

[0063] The immunoreactivities of the antibodies elicited against Aβ peptides and against the soluble $A\beta_{1-42}$ peptides were determined by ELISAs (enzyme-linked immunosorbent assays) using 96-well flat bottom microtiter plates which were coated with the $A\beta_{1-42}$ peptide fragments, SEQ ID NOs: 67 or 65 as the immunosorbent. Aliquots (100 μL) of the peptide immunogen solution at a concentration of 5 μg/mL were incubated for 1 hour at 37°C. The plates were blocked by another incubation at 37°C for 1 hour with a 3% gelatin/PBS solution. The blocked plates were then dried and used for the assay. Aliquots (100 μL) of the test immune sera, starting with a 1:100 dilution in a sample dilution buffer and ten-fold serial dilutions thereafter, were added to the peptide coated plates. The plates were incubated for 1 hour at 37°C.

[0064] The plates were washed six times with 0.05% PBS/Tween® buffer. 100 μL of horseradish peroxidase labeled goat-anti-species specific antibody was added at appropriate dilutions in conjugate dilution buffer (Phosphate buffer containing 0.5M NaCl, and normal goat serum). The plates were incubated for 1 hour at 37°C before being washed as above. Aliquots (100 μL) of o-phenylenediamine substrate solution were then added. The color was allowed to develop for 5-15 minutes before the enzymatic color reaction was stopped by the addition of 50 μL 2N H_2SO_4. The A_{492nm} of the contents of each well was read in a plate reader. ELISA titers were calculated based on linear regression analysis of the absorbances, with cutoff A_{492nm} set at 0.5. The cutoff value chosen was rigorous with the values for diluted normal control samples being less than 0.15.

EXAMPLE 3

CHARACTERIZATION OF THE RELATIVE IMMUNOGENICITIES OF $A\beta_{1-42}$ AND ITS N-TERMINAL FRAGMENTS FOR OPTIMIZATION OF DESIGN FOR SITE-DIRECTED Aβ PEPTIDE-BASED SYNTHETIC VACCINE

[0065] To design a total synthetic vaccine that generates a high level of high affinity antibodies against the Aβ peptides with high cross-reactivity to the soluble $A\beta_{1-42}$ peptides and the plaques in the brain of AD patients, the relative immunogenicities of $A\beta_{1-42}$ and its N-terminal fragments were characterized initially. In order to determine the relative immunological properties of the various regions within $A\beta_{1-42}$ peptide, a mild adjuvant suitable for human use, alum was employed in the first study. The relative immunogenicities of $A\beta_{1-42}$ peptide and an N-terminal fragment thereof, $A\beta_{1-28}$ were compared. The immunogenicity evaluation was conducted according to procedures described in Example 2. Unexpectedly, $A\beta_{1-28}$ was found to be more immunogenic than the $A\beta_{1-42}$ peptide, indicating that there is immunosuppression within C-terminal fragment $A\beta_{29-42}$ (Table 5).

[0066] Subsequently, the immunogenicities of $A\beta_{1-28}$ was compared to $A\beta_{1-14}$, a shorter N-terminal fragment of $A\beta_{1-42}$. A more potent adjuvant suitable for human use (Montanide ISA51, Seppic, Paris, FR) was employed for the preparation of a water-in-oil emulsion for formulating the vaccine. Based on the data obtained as shown in Table 6, the relative immunogenicities for the three Aβ peptides (i.e. $A\beta_{1-14}$, $A\beta_{1-28}$ and $A\beta_{1-42}$) were ranked $A\beta_{1-28} > A\beta_{1-42} > A\beta_{1-14}$. Surprisingly, the loss of the C-terminal 14mer from $A\beta_{1-42}$ improved rather than reduced the immunogenicity. The antibody response against Aβ is primarily directed to the N-terminal region, particularly the $A\beta_{1-14}$ N-terminal fragment as shown by ELISA data (Table 6). However, a further shortening of the $A\beta_{1-28}$ fragment from the C-terminal to form the $A\beta_{1-14}$ fragment resulted in a loss in immunogenicity.

[0067] The short $A\beta_{1-14}$ fragment contains the main functional/regulatory site, EFRH, located at positions 3-6 of the $A\beta_{1-42}$ peptide as reported by Solomon *et al.* The blocking of this epitope by antibodies modulates the dynamics of aggregation as well resolubilization of already formed aggregates (Soloman *et al.*, *Proc Natl Acad. Sci,* 1996; 93:452-455; *Proc Natl Aca. Sci*, 1997; 94:4109-4112). Most of the anti- $A\beta_{1-28}$ and $A\beta_{1-42}$ antibodies are directed against the N-terinal fragment of the $A\beta_{1-42}$ peptide containing this epitope (Table 6). However, the $A\beta_{1-14}$ fragment by itself was poorly immunogenic. The results of this experiment suggest the presence of an intrinsic Th epitope within the $A\beta_{15-28}$ segment. This intrinsic Th epitope accounts for the modest immunogenicities of $A\beta_{1-28}$ and $A\beta_{1-42}$ peptides in guinea pigs.

[0068] The presence of a Th epitope in the $A\beta_{15-28}$ fragment is desirable. However, it is desirable to be able to engineer a more potent immunogen for a successful human vaccine when faced with the limitation of a restricted human MHC molecule, the number of appropriate doses and the type of adjuvants permitted for human use. Therefore, we attempted the linkage of a foreign or extrinsic Th such as that derived from HBV Th (SEQ ID NO: 1) to the C-terminal of the $A\beta_{1-28}$ peptide (SEQ ID NO:66). The extrinsic Th epitope significantly enhanced the immunogenicity of the $A\beta_{1-28}$ fragment as shown in Table 6. The antibody response to the engineered immunogen with the $A\beta_{1-28}$ fragment remained directed to the functional N-terminal fragment of peptide immunogen (SEQ ID NO: 70) making this construct a better immunogen than the $A\beta_{1-28}$ fragment or $A\beta_{1-42}$ fragment alone. This peptide immunogen (SEQ ID NO: 70) represents a peptide immunogen with the formula:

$$(A)_n\,(\text{N-terminal fragment of A}\beta\text{ peptide})\,\text{-}(B)_o\,\text{-}(Th)_m$$

wherein:

A is αNH_2, with $A\beta_{1-28}$ being an N-terminal fragment of $A\beta_{1-42}$;

B is glycine;

Th is a helper T cell epitope derived from a foreign pathogen, HBsAg Th (SEQ ID NO: 1), and wherein n is 1, m is 1 and o is 2.

EXAMPLE 4

LOWER LIMIT OF N-TERMINAL FRAGMENT OF Aβ FOR THE DEVELOPMENT OF Aβ BASED SYNTHETIC VACCINE FOR AD

[0069] Since the main functional/regulatory site comprising the EFRH residues is located at positions 3-6 of the $A\beta_{1-42}$ peptide (Soloman *et al. Proc Natl Acad. Sci,* 1996; 93:452-455; *Proc Natl Aca. Sci*, 1997; 94:4109-4112), it was useful to explore the shortest N-terminal fragment of $A\beta_{1-42}$ peptide as an optimal B cell target site on Aβ for incorporation into the synthetic immunogen of the present invention.

[0070] Each of several short non-immunogenic N-terminal fragments of Aβ, $A\beta_{1-10}$, $A\beta_{1-12}$, $A\beta_{1-14}$ along with $A\beta_{1-28}$ was incorporated into immunogens designed with a representative idealized artificial Th (SEQ ID NO:51). Linkage was through an εN-Lys spacer. The engineered constructs were formulated with strong adjuvants due to the expected low immunogenicity of the short Aβ fragments. The three synthetic constructs were formulated in complete and incomplete Freund's adjuvant and tested for their immunogenicities based on procedures as described in Example 2. As shown in Table 7, all four peptide immunogens were highly immunogenic with Log_{10} ELISA titers in the range from 4.3 to 5.6 [i.e. $10^{4.3}$ to $10^{5.6}$] with very high crossreactivities to the full length $A\beta_{1-42}$ peptide after only four weeks from the initial immunization. More importantly, fragments as small as $A\beta_{1-10}$, $A\beta_{1-12}$ and $A\beta_{1-14}$ each linked to the idealized artificial Th (SEQ ID NO:51) were found to be highly immunogenic after linkage to a disclosed artificial Th epitope (Table 7). These peptide immunogens were designed in accordance with the formula:

$(A)_n$-(N-terminal fragment of Aβ peptide)-$(B)_o$-$(Th)_m$

wherein:

A is αNH_2, wherein the N-terminal fragment is $A\beta_{1-10}$, $A\beta_{1-12}$, $A\beta_{1-14}$ or $A\beta_{1-28}$;

B is ε-N Lysine, a spacer linked through its epsilon amino group to the next amino acid;

Th is a helper T cell epitope derived from an idealized artificial Th, MVF Th1-16(SEQ ID NO:51), wherein n is 1, m is 1 and o is 1.

[0071] It was found that further reduction in the length of the N-terminal fragment of Aβ to less than a 10mer would result in more limited, thus undesirable, immunogenicity. It appears that peptides smaller than 10 amino acids are problematic for receptor recognition by class II MHC molecules (Immunology, Fifth edition, ed. Roitt *et al.*, 1998, Mosby International Ltd., London, pp88-89).

[0072] Based on this study of Aβ, the useful B cell site derived from $A\beta_{1-42}$ should be in the size range of about 10 to about 28 residues.

EXAMPLE 5

SITE-DIRECTED IMMUNOREACTIVITY TARGETED BY THE SYNTHETIC PEPTIDE IMMUNOGEN LINKED TO ARTIFICIAL Th EPITOPE

[0073] The non-immunogenic N-terminal fragment such as $A\beta_{1-14}$ of Aβ peptide was linked either through an εN-lysine spacer to an artificial Th peptide designated as MVF Th 1-16 (SEQ ID NO:51), or through a standard chemical coupling procedure to a conventional carrier protein KLH. The two immunogenic constructs were evaluated in guinea pigs for their relative "site-directed" immunogenicities to Aβ peptide and the resultant respective reactivity of the antibodies towards their respective carriers, the artificial Th epitope or the KLH carrier protein, according to the procedures described in Example 2. The short $A\beta_{1-14}$ peptide alone as a control immunogen, and the two immunogenic

constructs were formulated in a water-in-oil emulsion containing the adjuvant ISA51, a formulation that is suitable for human use. As shown in Table 8, the N-terminal
$A\beta_{1-14}$ fragment by itself is non-immunogenic as expected. The synthetic immunogen comprising $A\beta_{1-14}$ fragment and artificial Th (SEQ ID NO: 73) was found to be highly immunogenic in eliciting site-directed antibodies to $A\beta_{1-14}$. The antibodies were also found to be highly cross-reactive to soluble $A\beta_{1-42}$ peptide as early as 4 weeks after the initial immunization (Log_{10} titers of 4.094 and 4.126 for 4 and 6 weeks post initial immunization respectively). When these Aβ-reactive high titer immune sera were tested by ELISA on the MVF Th1-16 peptide (SEQ ID NO 51) coated plate, they were found to be negative (Log_{10} titer of 0.038 and 0.064 for 4 and 6 weeks post initial immunization respectively)showing that irrelevant antibodies were not produced. The data obtained as shown in Table 8 clearly demonstrated the highly specific site-directed characteristic of the peptide immunogen of the present invention.

[0074] The immunogens with the carrier protein KLH was found to be highly immunoreactive with the conventional peptide-carrier protein conjugate (e.g. Log_{10} titers of 4.903 and 5.018 for 4 and 6 weeks post initial immunization respectively). However, the antibodies elicited were only moderately crossreactive with the soluble $A\beta_{1-42}$ peptide (e.g. with Log_{10} titers of 3.342 and 2.736 for 4 and 6 weeks post initial immunization respectively). This is approximately 10X to 100X less than SEQ ID NO:73. Unexpectedly, the peptide immunogens of the present invention were highly site-directed and focused. Only functionally important antibodies towards the anti-aggregation and disaggregation sites on the N-terminal fragment of the Aβ peptide were generated rather than towards irrelevant carrier sites.

EXAMPLE 6

EVALUATION OF Aβ PEPTIDE IMMUNOGEN BY CROSS-REACTIVITIES TO SENILE PLAQUES

[0075] Brains of AD patients with plaques and tangles and thioflavine S positive blood vessels (TSBV) containing amyloid plaques were used for evaluation of cross-reactivities to polymeric senile plaques of the immune sera raised in guinea pigs and baboons against Aβ peptide immunogens. Plaques and TSBV reactivities were detected by immunoperoxidase staining using Avidin-Biotinylated antibody Complex (ABC) method or by immunofluorescence staining using rhodamine conjugated Fab fragment of species specific anti-IgG. All guinea pig sera were tested at a dilution of 1:100 with end point titers determined for some of the samples. All baboon sera were tested at a dilution of 1:50. The evaluation of the immune and preimmune sera were kindly performed under code by Dr. Gaskin as described (Gaskin *et al., J. Exp Med.* 165:245, 1987).

[0076] In Figure 1, serial cross sections of brains from 2 AD patients were initially examined at 10X magnification. Sections (a), (b) and (c) are from AD Brain 1 and (d), (e) and (f) are from AD brain 2. Preimmune normal serum and immune sera from guinea pigs collected at 6 weeks post-initial immunization were tested by immunoperoxidase staining on cryostat sections from AD temporal cortex rich in plaques and neurofilament tangles (NFT). The immune sera used in the first study shown on slides Figures 1a and 1d were obtained from animals immunized with $A\beta_{1-28}$ -εK-MvF Th1-16 (SEQ ID NO:74) prepared in ISA51 water-in-oil emulsion. The results show significant binding to both senile plaques and amyloid plaques on the thioflavine S positive blood vessels (TSBV). The cross-reactivities of the immune sera raised against the equivalent immunogen prepared in CFA/ICFA are shown in slides Figures 1b and 1d. Unexpectedly, in contrast to the results obtained with the vaccine formulated with ISA51, preferential binding to the $A\beta_{1-28}$ plaques on the blood vessels (TSBV) were observed for the sera raised against the CFA/ICFA vaccine. This means that the antibodies elicited by the vaccine formulated with ISA51 is distinguishable from the antibodies raised by the vaccine formulated in CFA/ICFA. Moreover, the

antibodies generated by the vaccines formulated according to the present invention provided antibodies that have the desired higher cross reactivity to senile plaques in the brain tissue. Preimmune serum gave no staining in corresponding serial sections shown in slides Figures 1c and 1f.

[0077] Further Immunoperoxidase staining of serial cross sections of AD brain 1 with preimmune and immune sera at 1:100 dilution are shown in Figures 2a to 2e at 40X magnification. The sera obtained from animals immunized with $A\beta_{1-28}$-εK-MVF Th 1-16 (Seq ID NO:74) prepared in ISA 51 water-in-oil emulsion strongly stained the plaques forming a pattern of cores as shown in slides Figures 2a and 2d. Again, surprisingly, staining with immune sera prepared against the corresponding CFA/ICFA formulation gave a different staining pattern in that reactivities with plaques were predominantly on the blood vessels as shown in Figure 2b rather than with the plaques in the brain tissue. Preimmune serum did not stain the sections as shown in Figure 2c. The hyperimmune sera generated by immunization with $A\beta_{1-42}$ peptide alone in CFA/ICFA, despite its strong reactivities with $A\beta_{1-28}$ by ELISA, gave a surprisingly weak staining pattern in the section shown in Figure 2e.

[0078] Similar immunostaining of AD brain tissue was performed with 11 pooled immune and preimmune sera obtained from guinea pigs immunized with the various vaccine formulations described in Examples 3, 4 and 5. These sera were also evaluated for their antibody reactivities with the functional-site by $A\beta_{1-14}$ ELISA, and with the soluble $A\beta_{1-42}$ by $A\beta_{1-42}$ ELISA (Table 9). In general, parallel trends were found with sera tested in all three assays. As shown in Table 9, the anti-peptide reactivities of the pre-immune serum and the sera raised against the short peptide $A\beta_{1-14}$ alone formulated in ISA51 water-in-oil emulsion by ELISA were low and the cross-reactivities to plaques were negligible. Modest reactivities were found with sera from animals vaccinated with $A\beta_{1-28}$ peptide alone formulated in Alum and in ISA51, and $A\beta_{1-14}$ conjugated to KLH and formulated in ISA51. Whereas, significant site-directed reactivities to the functional $A\beta_{1-14}$ site, to soluble Aβ, and to the plaques and TSBV in AD patient brain tissue sections were found with sera from animals

immunized with synthetic Aβ /Th immunogens of the present invention. The results obtained from these studies, therefore, demonstrate excellent and useful immunogenicity of the peptide immunogens comprising the N-terminal fragment of $A\beta_{1-42}$ having amino acids from 1-28 to about 1-10, linked to foreign Th epitopes. Moreover, the results showed that the presence of a foreign Th epitope improves the immunogenicity of the peptide immunogens of the present invention to a surprising extent. The peptide immunogens of the present invention in clinically acceptable vaccine formulations acceptable to use in humans generated antibodies having the desired cross-reactivity to senile plaques in the brain tissues of AD patients.

EXAMPLE 7

THE IMMUNOGENICITY OF REPRESENTATIVE Aβ PEPTIDE VACCINES IN BABOONS AS PREDICTOR OF IMMUNOTHERAPEUTIC EFFICACY FOR AD

[0079] A representative synthetic immunogen, $A\beta_{1-28}$-ε-K-MvF Th1-16 (SEQ ID NO:74), formulated in ISA51 water-in-oil emulsion at dose levels of 25ug/0.5mL,100ug/0.5mL and 400ug/0.5mL were given to three baboons Y299, X398, X1198 at 0, 3 and 6 weeks schedule from initial immunization. Pre-immune sera and sera at weeks 5 and 8 weeks post initial immunization (wpi) were collected. For comparison, a fourth baboon X798 was given 100ug/0.5mL doses of an equimolar mixture of free peptides $A\beta_{1-28}$ and $A\beta_{1-42}$ formulated in alum, the standard adjuvant approved for human use. Preimmune sera were used as the negative control.

[0080] Sera from all four immunized animals were collected and evaluated for their antibody reactivities with the functional site by $A\beta_{1-14}$ ELISA, and for reactivities with soluble $A\beta_{1-42}$ by $A\beta_{1-42}$ ELISA (for sera collected at 0, 5 and 8 wpi). The cross-reactivities of the anti-sera (8wpi only) with the senile plaques and the plaques in thioflavine S positive blood vessels were evaluated by immunostaining as described in Example 6. Instead of using anti-baboon Ig, the

antibody detector used is an Fab fragment from anti-human IgG that recognizes all human isotypes and is cross-reactive with baboon IgG.

[0081] Parallel trends again were found with sera tested in all three assays. As shown in Table 10, pre-immune sera were negative. Modest ELISA reactivities were found with serum from animal X798 vaccinated with $A\beta_{1-28}$ and $A\beta_{1-42}$ formulated in Alum. However, the reactivity of this serum was weak for the recognition of senile plaques. In contrast, significant site-directed reactivities to the functional-site of $A\beta_{1-14}$, to soluble $A\beta_{1-42}$, and to the plaques and TSBV in AD patient brain sections were found with sera collected at 8 weeks post initial immunization from animals immunized with the representative composition of the invention (SEQ ID NO:74) at both the 100ug/0.5mL and 400ug/0.5mL doses formulated with ISA51. The results obtained from this baboon study, therefore, demonstrated the usefulness of the immunogen of the present invention in a vaccine formulation appropriate for humans. The improvement in immunogenicity (10 to 100X increase in specific antibody titers to the functional-site of Aβ) is very significant in comparison to the peptide vaccine of the prior art with the immune responsiveness in baboons closely resembling that of humans.

[0082] Similarly, a mixture containing two to three synthetic immunogens of the present invention can be used for formulation into vaccines at from about 25 to 1000 ug per dose to elicit functional anti-$A\beta_{1-14}$ antibodies in genetically diverse human populations for the prevention and treatment of AD. Broad immunogenicity in humans is expected due to the presence of a promiscuous Th epitope in the peptide immunogen of the invention that provides for achieving broad MHC recognition.

[0083]

Table 1

Pathogen-derived Promiscuous T Helper Cell Epitopes (Th)

Description of Th	Amino Acid Sequence	SEQ ID NO
HBs Th[a]	FFLLTRILTIPQSLD	1
PT_1 Th[a]	KKLRRLLYMIYMSGLAVRVHVSKEEQYYDY	2
TT_1 Th[a]	KKQYIKANSKFIGITEL	3
TT_2 Th[a]	KKFNNFTVSFWLRVPKVSASHL	4
PT_{1A} Th[a]	YMSGLAVRVHVSKEE	5
TT_3 Th[a]	YDPNYLRTDSDKDRFLQTMVKLFNRIK	6
PT_2 Th[a]	GAYARCPNGTRALTVAELRGNAEL	7
MVF_1 Th[a]	LSEIKGVIVHRLEGV	8
MVF_2 Th[a]	GILESRGI KARITHVDTESY	9
TT_4 Th[a]	WVRDIIDDFTNESSQKT	10
TT_5 Th[a]	DVSTIVPYIGPALNHV	11
CT Th[a]	ALNIWDRFDVFCTLGATTGYLKGNS	12
DT1 Th[a]	DSETADNLEKTVAALSILPGHGC	13
DT2 Tha	EEIVAQSIALSSLMVAQAIPLVGELVDIGFAATNFVESC	14
PF Th[a]	DHEKKHAKMEKASSVFNVVNS	15
SM Th[a]	KWFKTNAPNGVDEKHRH	16
$TraT_1$ Th[a]	GLQGKHADAVKAKG	17
TraT2 Th[a]	GLAAGLVGMAADAMVEDVN	18
$TraT_3$ Th[a]	STETGNQHHYQTRVVSNANK	19
HB_{c50-69}[b]	SDFFPSVRDLLDTASALYRE	20
CTP_{11} Th[c]	TINKPKGYVGKE	21

[a] US 5,759,551

[b] Ferrari et al., J Clin Invest, 1991; 88:214

[c] Stagg et al., Immunology, 1993; 71:1

[0084]

Table 2

Artificial Idealized Th and Combinatorial Library Idealized Artificial Th

a. MVF Th and Th epitopes derived therefrom

Th Identifier	Amino Acid Sequence	SEQ ID NO
MVF Th1	LSEIKGVIVHRLEGV	22
SSAL1 Th1	DLSDLKGLLLHKLDGL	23
	EI EIR III RIE I	24
	V V VVV V V	25
	F F FFF F F	26
MVF Th1-1	ISEIKGVIVHKIEGI	27
	MT RT TRM TM	28
	L L V	29
MVF Th1-2	ISEIKGVIVHKIEGI	30
	T RT TR T	31
MVF Th1-3	MSEIKGVIVHKLEGM	32
	LT MRT TRM TV	33
MVF Th1-4	ISEIKGVIVHKIEGI	34
MVF Th1-5	ITEIRTVIVTRIETI	35
MVF Th1-6	MSEMKGVIVHKMEGM	36
MVF Th1-7	LTEIRTVIVTRLETV	37
MVF Th1-8	ISISEIKGVIVHKIEGILF	38
	MT RT TRM TM	39
	L L V	40
MVF Th1-9	ISISEIKGVIVHKIEGILF	41
	T RT TR T	42
MVF Th1-10	ISLSEIKGVIVHKLEGMLF	43
	MT MRT TRM TV	44
MVF Th1-11	ISLTEIRTVIVTRLETVLF	45
	I I I	46
MVF Th1-12	ISISEIKGVIVHKIEGILF	47
MVF Th1-13	ISITEIRTVIVTRIETILF	48
MVF Th1-14	ISMSEMKGVIVHKMEGMLF	49
MVF Th1-15	ISLTEIRTVIVTRLETVLF	50
MVF Th1-16	ISITEIKGVIVHRIETILF	51

b. HBsAg Th, Prototype and Derivatives

Th Identifier	Amino Acid Sequence	SEQ ID NO
HbsAg-Th1	FFLLTRILTIPQSLD	52
HbsAg-Th1-1	KKKFFLLTRILTIPQSLD	53
HbsAg-Th1-2	FFLLTRILTIPQSL	54
SSAL2 Th2	KKKLFLLTKLLTLPQSLD	55
	RRRIKII RII I L IR	56
	VRVV VV V I V	57
	F FF FF F V F	58
	F	59
HbsAg Th1-3	KKKIITITRIITIITTID	60
HbsAg Th1-4	KKKIITITRIITIITTI	61
HbsAg Th1-5	KKKMMTMTRMITMITTID	62
HbsAg Th1-6	FITMDTKFLLASTHIL	63
HbsAg Th1-7	KKKFITMDTKFLLASTHIL	64

[0085]

Table 3

Amino Acid Sequences of $A\beta_{1-42}$ Peptides and its N-terminus Fragments

SEQ ID NO		Amino Acid Sequence
SEQ ID NO:65	$A\beta_{1-42}$	DAEFRHDSGYEVHHQKLVFFAEDVGSNKGAIIGLMVGGVVIA
SEQ ID NO:66	$A\beta_{1-28}$	DAEFRHDSGYEVHHQKLVFFAEDVGSNK
SEQ ID NO:67	$A\beta_{1-14}$	DAEFRHDSGYEVHH
SEQ ID NO:68	$A\beta_{1-12}$	DAEFRHDSGYEV
SEQ ID NO:69	$A\beta_{1-10}$	DAEFRHDSGY

[0086]

Table 4

Immunogen	Amino Acid Sequence	SEQ ID NO
$A\beta_{1-28}$-GG-HBV Th	DAEFRHDSGYEVHHQKLVFFAEDVGSNK-GG-FFLLTRILTIPQSLD	70
$A\beta_{1-10}$-εK-IS-MVF Th1-16	DAEFRHDSGY-εK-ISITEIKGVIVHRIETILF	71
$A\beta_{1-12}$-εK-IS-MVF Th1-16	DAEFRHDSGYEV-εK-ISITEIKGVIVHRIETILF	72
$A\beta_{1-14}$-εK-IS-MVF Th1-16	DAEFRHDSGYEVHH-εK-ISITEIKGVIVHRIETILF	73
$A\beta_{1-28}$-εK-IS-MVF Th1-16	DAEFRHDSGYEVHHQKLVFFAEDVGSNK-εK-ISITEIKGVIVHRIETILF	74
$A\beta_{1-14}$-εK-MVF Th1-9	DAEFRHDSGYEVHH-εK-ISISEIKGVIVHKIEGILF T RT TR T	75 76

[0087]

Table 5

Immunogen	Adjuvant	GP ID #	ELISA Titer (Log_{10})							
			4 WPI				6 WPI			
			$A\beta_{1-14}$	Avg.	$A\beta_{1-42}$	Avg.	$A\beta_{1-14}$	Avg.	$A\beta_{1-42}$	Avg.
$A\beta_{1-28}$ (SEQ ID NO:66)	Alum	1630	1.244	2.326	0.878	2.401	0.888	1.966	1.202	2.405
		1631	3.408		3.924		3.044		3.608	
$A\beta_{1-42}$ (SEQ ID NO:65)	Alum	1634	0.773	1.124	0.680	1.461	1.062	1.784	1.203	1.807
		1635	1.474		2.242		2.505		2.510	

[0088]

Table 6

Immunogen	Adjuvant	GP ID #	ELISA Titer (Log_{10})							
			4 WPI				6 WPI			
			$A\beta_{1-14}$	Avg.	$A\beta_{1-42}$	Avg.	$A\beta_{1-14}$	Avg.	$A\beta_{1-42}$	Avg.
$A\beta_{1-14}$ (SEQ ID NO: 67)	ISA 51	1658	1.168	1.129	1.229	0.975	1.100	1.271	1.285	1.080
		1659	1.090		0.720		1.441		0.874	
$A\beta_{1-28}$ (SEQ ID NO: 66)	ISA51	1632	2.341	2.291	3.656	3.382	2.276	2.715	3.359	3.455
		1633	2.241		3.107		3.153		3.550	
$A\beta_{1-28}$- GG-HBVTh (SEQ ID NO: 70)	ISA51	1642	4.792	4.612	4.526	4.582	4.548	4.498	4.441	4.261
		1643	4.432		4.637		4.447		4.081	
$A\beta_{1-42}$ (SEQ ID NO: 65)	ISA51	1636	2.724	1.864	3.603	2.402	2.286	1.997	3.250	2.873
		1637	1.004		1.201		1.707		2.495	

[0089]

Table 7

Immunogen	Adjuvant	GP ID #	ELISA Titer (Log_{10})							
			4 WPI				6 WPI			
			$A\beta_{1-14}$	Avg.	$A\beta_{1-42}$	Avg.	$A\beta_{1-14}$	Avg.	$A\beta_{1-42}$	Avg.
$A\beta_{1-10}$-εK-MVF Th1-16 (SEQ ID NO: 71)	CFA/IFA	1666	4.293	4.495	4.924	5.087	4.414	4.320	5.180	5.265
		1667	4.696		5.250		4.225		5.350	
$A\beta_{1-12}$-εK-MVF Th1-16 (SEQ ID NO: 72)	CFA/IFA	1664	4.577	4.495	5.100	4.891	5.320	4.545	6.000	5.278
		1665	4.322		4.682		3.700		4.555	
$A\beta_{1-14}$-εK-MVF Th1-16 (SEQ ID NO: 73)	CFA/IFA	1660	3.700	3.285	4.677	5.060	4.544	4.683	5.250	5.625
		1661	4.764		5.443		4.822		6.000	
$A\beta_{1-28}$-εK-MVF Th1-16 (SEQ ID NO: 74)	CFA/IFA	1584	3.355	3.201	4.610	4.328	2.743	3.592	4.487	4.901
		1585	3.707		4.688		3.731		5.155	
		1586	2.545		3.685		4.304		5.061	

[0090]

Table 8

Immunogen	Adjuvant	GP ID #	ELISA Titer (Log_{10})							
			4 WPI				6 WPI			
			$A\beta_{1-42}$	Avg.	Th peptide or KLH	Avg	$A\beta_{1-42}$	Avg.	Th peptide or KLH	Avg
$A\beta_{1-14}$ (SEQ ID NO: 67)	ISA 51	1658	1.229	0.975	NA	NA	1.285	1.080	NA	NA
		1659	0.720		NA		0.874		NA	
$A\beta_{1-14}$-εK-MVF Th1-16 (SEQ ID NO: 73)	ISA 51	1662	4.388	4.094	0.006	0.038	4.559	4.126	0.065	0.064
		1663	3.800		0.070		3.693		0.063	
KLH-(C) $A\beta_{1-14}$ (SEQ ID NO: 67)	ISA 51	1670	3.181	3.342	4.672	4.903	2.625	2.736	4.876	5.018
		1671	3.502		5.133		2.846		5.160	

[0091]

Table 9

Vaccine Formulation	GP ID#	ELISA Titer (Log_{10})				Immunostaining[a] of serial frozen sections of AD's brain tissue	
		$A\beta_{1-42}$		$A\beta_{1-14}$			
			Avg		Avg	Plaque	TSBV
$A\beta_{1-28}$ in Alum	1630	0.878	2.401	1.244	2.326	+1	+4
	1631	3.924		3.408			
$A\beta_{1-28}$ in ISA51	1632	3.686	3.397	2.341	2.291	+3	+5
	1633	3.107		2.241			
$A\beta_{1-28}$-εK-MVF Th1-16 in CFA/IFA (SEQ ID NO: 74)	1584	4.610	4.328	3.355	3.201	+4	+6
	1585	4.688		3.707			
	1586	3.685		2.540			
$A\beta_{1-28}$-εK-MVF Th1-16 in ISA51 (SEQ ID NO: 74)	1642	3.603	4.582	2.724	3.510	+4	+6
	1643	1.201		1.004			
$A\beta_{1-14}$ in ISA51	1658	1.229	0.975	1.168	1.129	Neg	Neg
	1659	0.720		1.090			
$A\beta_{1-14}$-εK-MVF Th1-16 in CFA/IFA (SEQ ID NO: 73)	1660	4.677	5.060	3.700	4.232	+4	+6
	1661	5.443		4.764			
$A\beta_{1-14}$-εK-MVF Th1-16 in ISA51 (SEQ ID NO: 73)	1662	4.388	4.094	3.551	3.285	+4	+6
	1663	3.800		3.018			
$A\beta_{1-12}$-εK-MVF Th1-16 in CFA/IFA (SEQ ID NO: 72)	1664	5.100	4.891	4.577	4.450	+4	+6
	1665	4.682		4.322			
$A\beta_{1-10}$-εK-MVF Th1-16 in CFA/IFA (SEQ ID NO: 71)	1666	4.924	5.087	4.293	4.455	+4	+5
	1667	5.250		4.696			
KLH-(C) $A\beta_{1-14}$ in ISA51	1670	3.181	3.342	3.280	3.102	+2	+4
	1635	3.502		2.924			
Negative Control (preimmune serum)		<0.5		<0.5		Neg	Neg

a: Serial dilution @ 1:100

[0092]

Table 10

Group #	Vaccine Formulation	Dose	ELISA Titer (Log_{10})						Immunostaining of frozen sections of AD brain (8 wpi)	
			$A\beta_{1-42}$			$A\beta_{1-14}$				
			0 WPI	5 WPI	8 WPI	0 WPI	5 WPI	8 WPI	Plaques	TSBV
1	$A\beta_{1-28}$-εKV-MVF Th1-16 in ISA51	25 µg	0.894	2.962	2.736	0.665	1.745	2.706	+2	+
2		100 µg	0.610	2.987	3.640	0.794	2.816	4.800	+4	+6
3		400 µg	0.696	2.696	4.050	0.539	4.250	3.799	+4	+6
4	$A\beta_{1-28}$+ $A\beta_{1-42}$ in Alum	100 µg	0.897	1.963	2.485	0.798	0.727	2.850	+	+
5	Negative control	-	-	-	-	-	-	-	Neg	Neg

I claim:

1. A peptide immunogen of about 20 to 100 amino acids long comprising:

(i) a helper T cell (Th) epitope selected from the group consisting of SEQ ID Nos: 1 to 64;

(ii) an N-terminal fragment of $A\beta_{1-42}$ peptide, SEQ ID NO:65, consisting of from 10 to 28 amino acid residues wherein each fragment comprises amino acid residue 1 of the $A\beta_{1-42}$ peptide or an immunologically functional analog of the N-teminal fragment of $A\beta_{1-42}$ peptide; and

(iii) optionally a spacer consisting of at least an amino acid to separate the immunogenic domains.

2. A peptide immunogen of claim 1, wherein the spacer is selected from the group consisting of an amino acid, Gly-Gly, (α, ε-N)-Lys, and Pro-Pro-Xaa-Pro-Xaa-Pro (SEQ ID NO:73).

3. A peptide immunogen of claim 2, wherein the spacer is Gly-Gly.

4. A peptide immunogen of claim 2, wherein the spacer is ε-N-Lys.

5. A peptide immunogen of claim 1, wherein the N-terminal fragment of $A\beta_{1-42}$ peptide is selected from the group consisting of SEQ ID NOs: 66-69 and an immunologically functional analog thereof.

6. A peptide immunogen of any one of claims 2, 3, or 4, wherein the N-terminal fragment of $A\beta_{1-42}$ peptide is selected from the group consisting of SEQ ID NOs: 66-69 and an immunologically functional analog thereof.

7. A peptide immunogen of claim 1, wherein Th is selected from the group consisting of SEQ ID NOs: 1, 3, 4, 5, 6, 7, 8, 9, 20, 38-40, 47-51 and 52-54.

8. A peptide immunogen of any one of claims 2, 3, or 4, wherein Th is selected from the group consisting of SEQ ID NOs: 1, 3, 4, 5, 6, 7, 8, 9, 20, 38-40, 47-51 and 52-54..

9. A peptide immunogen selected from the group consisting of SEQ ID NOs: 70, 71, 72, 73, and 74.

10. A peptide immunogen of claim 9 consisting of SEQ ID NO: 73.

11. A peptide immunogen of claim 9 consisting of SEQ ID NO: 74.

12. The peptide immunogen represented by one of the following formulae:

$(A)_n$-(N-terminal fragment of $A\beta_{1-42}$ peptide)-$(B)_o$-$(Th)_m$-X;or

$(A)_n$-$(Th)_m$-$(B)_o$-(N-terminal fragment of $A\beta_{1-42}$ peptide)-X;

wherein

each A is independently an amino acid;

each B is a linking group selected from the group consisting of an amino acid, gly-gly, (α, ε-N)-Lys, and Pro-Pro-Xaa-Pro-Xaa-Pro (SEQ ID NO:73);

Th comprise an amino acid sequence that constitutes a helper T cell epitope, selected from the group consisting of SEQ ID NOs: 1-64 and an immune enhancing analog thereof;

(N-terminal fragment of $A\beta_{1-42}$ peptide) is 10 to about 28 amino acid residues and wherein each fragment comprises EFRH of the $A\beta_{1-42}$ peptide and immunologically functional analog thereof;

X is an α-COOH or α-$CONH_2$ of an amino acid ;

n is from 0 to about 10;
m is from 1 to about 4; and
o is from 0 to about 10.

13. A peptide immunogen of claim 12, wherein the spacer is Gly-Gly.

14. A peptide immunogen of claim 12, wherein the spacer is ε-N-Lys.

15. A peptide immunogen of claim 12, wherein the N-terminal fragment of $A\beta_{1-42}$ peptide is selected from the group consisting of SEQ ID NOs: 66-69 and an immunologically effective analog thereof.

16. A peptide immunogen of any one of claims 13, or 14 , wherein the N-terminal fragment of $A\beta_{1-42}$ peptide is selected from the group consisting of SEQ ID NOs: 66-69 and an immunologically functional analog thereof

17. A peptide immunogen of claim 12, wherein Th is selected from the group consisting of SEQ ID NOs: 1, 3, 4, 5, 6, 7, 8, 9, 20, 38-40, 47-51 and 52-54..

18. A peptide immunogen of any one of claims 13, or 14 wherein Th is selected from the group consisting of SEQ ID NOs: 1, 3, 4, 5, 6, 7, 8, 9, 20, 38-40, 47-51 and 52-54.

19. A peptide immunogen of claim 15 wherein Th is selected from the group consisting of SEQ ID NOs: 1, 3, 4, 5, 6, 7, 8, 9, 20, 38-40, 47-51 and 52-54.

20. A peptide immunogen of claim 16 wherein Th is selected from the group consisting of SEQ ID NOs: 1, 3, 4, 5, 6, 7, 8, 9, 20, 38-40, 47-51 and 52-54.

21. A composition comprising a pepetide immunogen of claim 1 and a pharmaceutically acceptable adjuvant and/or carrier selected from the group consisting of alum, liposyn, saponin, squalene, L121, emulsigen monophosphyryl lipid A (MPL), polysorbate 80, QS21, Montanide ISA51, ISA35, ISA206 and ISA 720.

22. A composition comprising a pepetide immunogen of claim 2 and a pharmaceutically acceptable adjuvant and/or carrier selected from the group consisting of alum, liposyn, saponin, squalene, L121, emulsigen monophosphyryl lipid A (MPL), polysorbate 80, QS21, Montanide ISA51, ISA35, ISA206 and ISA 720.

23. A composition comprising a pepetide immunogen of claim 3 and a pharmaceutically acceptable adjuvant and/or carrier selected from the group consisting of alum, liposyn, saponin, squalene, L121, emulsigen monophosphyryl lipid A (MPL), polysorbate 80, QS21, Montanide ISA51, ISA35, ISA206 and ISA 720.

24. A composition comprising a pepetide immunogen of claim 4 and a pharmaceutically acceptable adjuvant and/or carrier selected from the group consisting of alum, liposyn, saponin, squalene, L121, emulsigen monophosphyryl lipid A (MPL), polysorbate 80, QS21, Montanide ISA51, ISA35, ISA206 and ISA 720.

25. A composition comprising a pepetide immunogen of claim 5 and a pharmaceutically acceptable adjuvant and/or carrier selected from the group consisting of alum, liposyn, saponin, squalene, L121, emulsigen monophosphyryl lipid A (MPL), polysorbate 80, QS21, Montanide ISA51, ISA35, ISA206 and ISA 720.

26. A composition comprising a pepetide immunogen of claim 6 and a pharmaceutically acceptable adjuvant and/or carrier selected from the group consisting of alum, liposyn, saponin, squalene, L121, emulsigen monophosphyryl lipid A (MPL), polysorbate 80, QS21, Montanide ISA51, ISA35, ISA206 and ISA 720.

27. A composition comprising a pepetide immunogen of claim 7 and a pharmaceutically acceptable adjuvant and/or carrier selected from the group consisting of alum, liposyn, saponin, squalene, L121, emulsigen monophosphyryl lipid A (MPL), polysorbate 80, QS21, Montanide ISA51, ISA35, ISA206 and ISA 720.

28. A composition comprising a pepetide immunogen of claim 8 and a pharmaceutically acceptable adjuvant and/or carrier selected from the group consisting of alum, liposyn, saponin, squalene, L121, emulsigen monophosphyryl lipid A (MPL), polysorbate 80, QS21, Montanide ISA51, ISA35, ISA206 and ISA 720.

29. A composition comprising a pepetide immunogen of claim 9 and a pharmaceutically acceptable adjuvant and/or carrier selected from the group consisting of alum, liposyn, saponin, squalene, L121, emulsigen monophosphyryl lipid A (MPL), polysorbate 80, QS21, Montanide ISA51, ISA35, ISA206 and ISA 720.

30. A composition comprising a pepetide immunogen of claim 10 and a pharmaceutically acceptable adjuvant and/or carrier selected from the group consisting of alum, liposyn, saponin, squalene, L121, emulsigen monophosphyryl lipid A (MPL), polysorbate 80, QS21, Montanide ISA51, ISA35, ISA206 and ISA 720.

31. A composition comprising a pepetide immunogen of claim 11 and a pharmaceutically acceptable adjuvant and/or carrier selected from the group consisting of alum, liposyn, saponin, squalene, L121, emulsigen monophosphyryl lipid A (MPL), polysorbate 80, QS21, Montanide ISA51, ISA35, ISA206 and ISA 720.

32. A composition comprising a pepetide immunogen of claim 12 and a pharmaceutically acceptable adjuvant and/or carrier selected from the group consisting of alum, liposyn, saponin, squalene, L121, emulsigen monophosphyryl lipid A (MPL), polysorbate 80, QS21, Montanide ISA51, ISA35, ISA206 and ISA 720

33. A composition comprising a pepetide immunogen of claim 13 and a pharmaceutically acceptable adjuvant and/or carrier selected from the group consisting of alum, liposyn, saponin, squalene, L121, emulsigen monophosphyryl lipid A (MPL), polysorbate 80, QS21, Montanide ISA51, ISA35, ISA206 and ISA 720.

34. A composition comprising a pepetide immunogen of claim 14 and a pharmaceutically acceptable adjuvant and/or carrier selected from the group consisting of alum, liposyn, saponin, squalene, L121, emulsigen monophosphyryl lipid A (MPL), polysorbate 80, QS21, Montanide ISA51, ISA35, ISA206 and ISA 720.

35. A composition comprising a pepetide immunogen of claim 15 and a pharmaceutically acceptable adjuvant and/or carrier selected from the group consisting of alum, liposyn, saponin, squalene, L121, emulsigen monophosphyryl lipid A (MPL), polysorbate 80, QS21, Montanide ISA51, ISA35, ISA206 and ISA 720.

36. A composition comprising a pepetide immunogen of claim 16 and a pharmaceutically acceptable adjuvant and/or carrier selected from the group consisting of alum, liposyn, saponin, squalene, L121, emulsigen monophosphyryl lipid A (MPL), polysorbate 80, QS21, Montanide ISA51, ISA35, ISA206 and ISA 720.

37. A composition comprising a pepetide immunogen of claim 17 and a pharmaceutically acceptable adjuvant and/or carrier selected from the group consisting of alum, liposyn, saponin, squalene, L121, emulsigen monophosphyryl lipid A (MPL), polysorbate 80, QS21, Montanide ISA51, ISA35, ISA206 and ISA 720.

38. A composition comprising a pepetide immunogen of claim 18 and a pharmaceutically acceptable adjuvant and/or carrier selected from the group consisting of alum, liposyn, saponin, squalene, L121, emulsigen monophosphyryl lipid A (MPL), polysorbate 80, QS21, Montanide ISA51, ISA35, ISA206 and ISA 720.

39. A composition comprising a pepetide immunogen of claim 19 and a pharmaceutically acceptable adjuvant and/or carrier selected from the group consisting of alum, liposyn, saponin, squalene, L121, emulsigen monophosphyryl lipid A (MPL), polysorbate 80, QS21, Montanide ISA51, ISA35, ISA206 and ISA 720.

40. A composition comprising a pepetide immunogen of claim 20 and a pharmaceutically acceptable adjuvant and/or carrier selected from the group consisting of alum, liposyn, saponin, squalene, L121, emulsigen monophosphyryl lipid A (MPL), polysorbate 80, QS21, Montanide ISA51, ISA35, ISA206 and ISA 720.

41. Use of a composition of claim 21, for the manufacture of a medicament for preventing or treating Alzheimer's disease.

42. Use of a composition of claim 22, for the manufacture of a medicament for preventing or treating Alzheimer's disease.

43. Use of a composition of claim 23, for the manufacture of a medicament for preventing or treating Alzheimer's disease.

44. Use of a composition of claim 24, for the manufacture of a medicament for preventing or treating Alzheimer's disease.

45. Use of a composition of claim 25, for the manufacture of a medicament for preventing or treating Alzheimer's disease.

46. Use of a composition of claim 26, for the manufacture of a medicament for preventing or treating Alzheimer's disease.

47. Use of a composition of claim 27, for the manufacture of a medicament for preventing or treating Alzheimer's disease.

48. Use of a composition of claim 28, for the manufacture of a medicament for preventing or treating Alzheimer's disease.

49. Use of a composition of claim 29, for the manufacture of a medicament for preventing or treating Alzheimer's disease.

50. Use of a composition of claim 30, for the manufacture of a medicament for preventing or treating Alzheimer's disease.

51. Use of a composition of claim 31, for the manufacture of a medicament for preventing or treating Alzheimer's disease.

52. Use a composition of claim 32, for the manufacture of a medicament for preventing or treating Alzheimer's disease.

53. Use of a composition of claim 33, for the manufacture of a medicament for preventing or treating Alzheimer's disease.

54. Use of a composition of claim 34, for the manufacture of a medicament for preventing Alzheimer's disease.

55. Use of a composition of claim 35, for the manufacture of a medicament for preventing or treating Alzheimer's disease.

56. Use of a composition of claim 36, for the manufacture of a medicament for preventing or treating Alzheimer's disease.

57. Use of a composition of claim 37, for the manufacture of a medicament for preventing or treating Alzheimer's disease.

58. Use of a composition of claim 38, for the manufacture of a medicament for preventing or treating Alzheimer's disease.

59. Use of a composition of claim 39, for the manufacture of a medicament for preventing or treating Alzheimer's disease.

60. Use of a composition of claim 40, for the manufacture of a medicament for preventing or treating Alzheimer's disease.

61. Use of a composition of claim 21, for the manufacture of a medicament for producing antibodies to $A\beta_{1-42}$ peptide that is cross reactive to soluble Aβ peptides and brain tissue plaques formed therefrom.

62. Use of a composition of claim 22, for the manufacture of a medicament for producing antibodies to $A\beta_{1-42}$ peptide that is cross reactive to soluble Aβ peptides and brain tissue plaques formed therefrom.

63. Use of a composition of claim 23, for the manufacture of a medicament for producing antibodies to $A\beta_{1-42}$ peptide that is cross reactive to soluble Aβ peptides and brain tissue plaques formed therefrom.

64. Use of a composition of claim 24, for the manufacture of a medicament for producing antibodies to $A\beta_{1-42}$ peptide that is cross reactive to soluble Aβ peptides and brain tissue plaques formed therefrom.

65. Use of a composition of claim 25, for the manufacture of a medicament for producing antibodies to $A\beta_{1-42}$ peptide that is cross reactive to soluble Aβ peptides and brain tissue plaques formed therefrom.

66. Use of a composition of claim 26, for the manufacture of a medicament for producing antibodies to $A\beta_{1-42}$ peptide that is cross reactive to soluble Aβ peptides and brain tissue plaques formed therefrom.

67. Use of a composition of claim 27, for the manufacture of a medicament for producing antibodies to $A\beta_{1-42}$ peptide that is cross reactive to soluble Aβ peptides and brain tissue plaques formed therefrom.

68. Use of a composition of claim 28, for the manufacture of a medicament for producing antibodies to $A\beta_{1-42}$ peptide that is cross reactive to soluble Aβ peptides and brain tissue plaques formed therefrom.

69. Use of a composition of claim 29, for the manufacture of a medicament for producing antibodies to $A\beta_{1-42}$ peptide that is cross reactive to soluble Aβ peptides and brain tissue plaques formed therefrom.

70. Use of a composition of claim 30, for the manufacture of a medicament for producing antibodies to $A\beta_{1-42}$ peptide that is cross reactive to soluble Aβ peptides and brain tissue plaques formed therefrom.

71. Use of a composition of claim 31, for the manufacture of a medicament for producing antibodies to $A\beta_{1-42}$ peptide that is cross reactive to soluble Aβ peptides and brain tissue plaques formed therefrom.

72. Use of a composition of claim 32, for the manufacture of a medicament for producing antibodies to $A\beta_{1-42}$ peptide that is cross reactive to soluble Aβ peptides and brain tissue plaques formed therefrom.

73. Use of a composition of claim 33, for the manufacture of a medicament for producing antibodies to $A\beta_{1-42}$ peptide that is cross reactive to soluble Aβ peptides and brain tissue plaques formed therefrom.

74. Use of a composition of claim 34, for the manufacture of a medicament for producing antibodies to $A\beta_{1-42}$ peptide that is cross reactive to soluble Aβ peptides and brain tissue plaques formed therefrom.

75. Use of a composition of claim 35, for the manufacture of a medicament for producing antibodies to $A\beta_{1-42}$ peptide that is cross reactive to soluble Aβ peptides and brain tissue plaques formed therefrom.

76. Use of a composition of claim 36, for the manufacture of a medicament for producing antibodies to $A\beta_{1-42}$ peptide that is cross reactive to soluble Aβ peptides and brain tissue plaques formed therefrom.

77. Use of a composition of claim 37, for the manufacture of a medicament for producing antibodies to $A\beta_{1-42}$ peptide that is cross reactive to soluble Aβ peptides and brain tissue plaques formed therefrom.

78. Use of a composition of claim 38, for the manufacture of a medicament for producing antibodies to $A\beta_{1-42}$ peptide that is cross reactive to soluble Aβ peptides and brain tissue plaques formed therefrom.

79. Use of a composition of claim 39, for the manufacture of a medicament for producing antibodies to $A\beta_{1-42}$ peptide that is cross reactive to soluble Aβ peptides and brain tissue plaques formed therefrom.

80. Use of a composition of claim 40, for the manufacture of a medicament for producing antibodies to $A\beta_{1-42}$ peptide that is cross reactive to soluble Aβ peptides and brain tissue plaques formed therefrom.

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<140> TBA
<141> 2001-5-25

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Leu Phe

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Leu Phe

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20 25 30

Leu Phe