EXPANSION OF DEFINITIVE ENDODERM CELLS

Related Applications

[0001] This application is a nonprovisional application of and claims priority to U.S. Provisional Patent Application No. 60/693,317, entitled EXPANSION OF ISOLATED DEFINITIVE ENDODERM CELLS, filed June 23, 2005 and a nonprovisional application of and claims priority to U.S. Provisional Patent Application No. 60/736,598, entitled MARKERS OF DEFINITIVE ENDODERM, filed November 14, 2005; this application is also a continuation-in-part of and claims priority to U.S. Patent Application Number 11/021,618, entitled DEFINITIVE ENDODERM, filed December 23, 2004, which claims priority under 35 U.S.C. § 119(e) as a nonprovisional application to U.S. Provisional Patent Application Number 60/587,942, entitled CHEMOKINE CELL SURFACE RECEPTOR FOR THE ISOLATION OF DEFINITIVE ENDODERM, filed July 14, 2004; U.S. Provisional Patent Application Number 60/586,566, entitled CHEMOKINE CELL SURFACE RECEPTOR FOR THE ISOLATION OF DEFINITIVE ENDODERM, filed July 9, 2004 and U.S. Provisional Patent Application Number 60/532,004, entitled DEFINITIVE ENDODERM, filed December 23, 2003.

Field of the Invention

[0002] The present invention relates to the fields of medicine and cell biology. In particular, the present invention relates to compositions of definitive endoderm cells which have been expanded either prior to or subsequent to enrichment, isolation and/or purification as well as methods of producing and using such cells.

Background

[0003] Human pluripotent stem cells, such as embryonic stem (ES) cells and embryonic germ (EG) cells, were first isolated in culture without fibroblast feeders in 1994 (Bongso et al., 1994) and with fibroblast feeders (Hogan, 1997). Later, Thomson, Reubinoff and Shamblott established continuous cultures of human ES and EG cells using mitotically inactivated mouse feeder layers (Reubinoff et al., 2000; Shamblott et al., 1998; Thomson et al., 1998).

[0004] Human ES and EG cells (hESCs) offer unique opportunities for investigating early stages of human development as well as for therapeutic intervention in several disease states, such as diabetes mellitus and Parkinson's disease. For example, the use of insulin-producing β-cells derived from hESCs would offer a vast improvement over current cell therapy procedures that utilize cells from donor pancreases for the treatment of diabetes. However, presently it is not known how to generate an insulin-producing β-cell from hESCs. As such, current cell therapy treatments for diabetes mellitus, which utilize islet cells from donor pancreases, are limited by the scarcity of high quality islet cells needed for transplant. Cell therapy for a single Type I diabetic patient requires a transplant of approximately 8×10^8 pancreatic islet cells. (Shapiro et al., 2000; Shapiro et al., 2001a; Shapiro et al., 2001b). As such, at least two healthy donor organs are required to obtain sufficient islet cells for a successful transplant. Human embryonic stem cells offer a source of starting material from which to develop substantial quantities of high quality differentiated cells for human cell therapies.

[0005] Two properties that make hESCs uniquely suited to cell therapy applications are pluripotence and the ability to maintain these cells in culture for prolonged periods. Pluripotency is defined by the ability of hESCs to differentiate to derivatives of all 3 primary germ layers (endoderm, mesoderm, ectoderm) which, in turn, form all somatic cell types of the mature organism in addition to extraembryonic tissues (e.g. placenta) and germ cells. Although pluripotency imparts extraordinary utility upon hESCs, this property also poses unique challenges for the study and manipulation of these cells and their derivatives. Owing to the large variety of cell types that may arise in differentiating hESC cultures, the vast majority of cell types are produced at very low efficiencies in mixed cell populations. In order to use hESCs as a starting material to generate cells that are useful in cell therapy applications, it would be advantageous to overcome the foregoing problems.

Summary of the Invention

[0006] Embodiments of the present invention relate to cell cultures comprising expanded definitive endoderm cells. Further embodiments described herein relate to methods for expanding enriched, isolated and/or purified definitive endoderm cells in culture.

[0006a] In a first aspect, the present invention refers to a method of expanding definitve endoderm cells in culture, said method comprising the steps of;

(a) obtaining a cell culture comprising definitive endoderm cells;

(b) isolating at least some of the definitive endoderm cells from at least some of the other cells in the cell culture, thereby producing a cell population enriched in definitive endoderm cells; and

(c) culturing said cell population enriched in definitive endoderm cells under conditions that permit the expansion of said definitive endoderm cells.

[0006b] In a second aspect, the present invention refers to an expanded definitive endoderm cell population produced by the method of the present invention.

[0006c] In a third aspect, the present invention refers to a method of expanding definitive endoderm cells in culture, said method comprising the

steps of: (a) obtaining a cell culture comprising definitive endoderm cells, and (b) passaging said definitive endoderm cells, thereby producing a plurality of cell cultures comprising definitive endoderm cells.

[0006d] In a fourth aspect, the present invention refers to an expanded definitive endoderm cell population produced by the method of the present invention.

[0007] Some of the methods described herein relate to the maintenance, growth, passage and/or expansion of definitive endoderm cells in cell culture. In such embodiments, cell cultures comprising definitive endoderm cells are obtained. The cells are then isolated so that at least some of the definitive endoderm cells are separated from at least some of the other cells in the cell culture, thereby producing a cell population that is enriched in definitive endoderm cells. In some embodiments, the enriched cell populations of definitive endoderm cells are cultured under conditions that permit the expansion of the definitive endoderm cells.

[0008] In other embodiments of the methods described herein, the definitive endoderm cells are multipotent cells that can differentiate into cells of the gut tube or organs derived therefrom. In preferred embodiments, the definitive endoderm cells are human definitive endoderm cells that are obtained by differentiating human embryonic stem cells (hESCs). In such embodiments, definitive endoderm cells can be derived from hESCs by contacting such cells with at least one growth factor from the TGFβ superfamily, such as activin A. In other embodiments, human and/or other definitive endoderm cells can be obtained from a pre-existing culture of definitive endoderm cells. In such embodiments, either a portion of or the entire culture may be used in the definitive endoderm expansion methods described herein.

[0009] In addition to obtaining cell cultures comprising definitive endoderm cells, some embodiments of the expansion methods described herein also comprise the step of producing enriched definitive endoderm cell populations. In some embodiments, such enriched definitive endoderm cell populations are produced by separating at least some of the definitive endoderm cells from at least some of the other cells in the cell cultures. As such, the at least some of the definitive endoderm cells are isolated from at least some of the other cells which remain in the cell culture. In some embodiments, the isolating step comprises providing the cells in the cell culture with a reagent which binds to a marker expressed in said definitive endoderm cells but which is not substantially expressed in said other cells present in the cell culture. The reagent-bound definitive endoderm cells are then separated from the non-reagent-bound cells, thereby producing an enriched definitive endoderm cell population. In some embodiments, the marker is CXCR4 and the reagent is an antibody with an affinity for CXCR4. In some embodiments, the definitive endoderm cells are separated by fluorescence activated cell sorting (FACS).

[0010] In still other embodiments, at least some of the definitive endoderm cells are separated from at least some of the other cells in the culture by specifically fluorescently labeling the definitive endoderm cells in culture and then separating the labeled cells from the unlabeled cells by FACS. In some embodiments, the fluorescence is produced by green fluorescent protein (GFP) or enhanced green fluorescent protein (EGFP). In some embodiments, the GFP and/or EGFP is expressed under the control of the SOX17 or the CXCR4 promoter.

[0011] In some embodiments, the enriched definitive endoderm cell populations that are produced as described above are substantially free of cells other than definitive endoderm cells. In other embodiments, the enriched definitive endoderm cell populations comprise from at least about 96% to at least about 100% definitive endoderm cells.

[0012] Additional embodiments of the methods described herein also include a culturing step that comprises plating the population enriched in definitive endoderm cells or a portion of the population. In some embodiments, the cells are plated on a surface coated with human fibronectin and/or poly-ornithine. In other embodiments, the culturing step comprises incubating the enriched definitive endoderm cell population or portion thereof in a medium comprising about 2% (v/v) serum. In some embodiments, the medium also comprises at least one growth factor. In certain embodiments, the at least one growth factor is a growth factor comprises a member of the TGFβ superfamily, such as activin A. Alternatively, the growth factor can be IGF1, bFGF, EGF or another growth factor. In such embodiments, the growth factor can be present in the medium at a concentration ranging from about 1 ng/ml to about 5000 ng/ml. In some embodiments, a combination of growth factors is present in the culture medium.

[0013] Additional embodiments described herein relate to methods of expanding definitive endoderm cells in culture by obtaining a cell culture comprising definitive endoderm cells and then passaging the definitive endoderm cells so as to produce a plurality of cell cultures comprising definitive endoderm cells. In some embodiments, the definitive endoderm cells obtained in cell culture are attached to a substrate, such as the surface of a cell culture flask or the surface of a microtiter plate. In some embodiments, the definitive endoderm cells are passaged using enzymatic methods. In other embodiments, the definitive endoderm cells are mechanically passaged. In yet

other embodiments, the definitive endoderm cells are passage using a cell dispersal buffer.

[0014] Still other embodiments described herein relate to expanded definitive endoderm cell cultures and/or populations produced by the methods described herein. In such embodiments, the definitive endoderm cells are multipotent cells that can differentiate into cells of the gut tube or organs derived therefrom.

[0015] In certain jurisdictions, there may not be any generally accepted definition of the term "comprising." As used herein, the term "comprising" is intended to represent "open" language which permits the inclusion of any additional elements. With this in mind, additional embodiments are described with reference to the numbered paragraphs below:

[0016] 1. A method of expanding definitive endoderm cells in culture, said method comprising the steps of: (a) obtaining a cell culture comprising definitive endoderm cells, (b) isolating at least some of the definitive endoderm cells from at least some of the other cells in the cell culture, thereby producing a cell population enriched in definitive endoderm cells; and (c) culturing said cell population enriched in definitive endoderm cells under conditions that permit the expansion of said definitive endoderm cells.

[0017] 2. The method of paragraph 1, wherein said definitive endoderm cells being multipotent cells that can differentiate into cells of the gut tube or organs derived therefrom.

[0018] 3. The method of paragraph 1, wherein said definitive endoderm cells are human definitive endoderm cells.

[0019] 4. The method of paragraph 3, wherein said definitive endoderm cells are derived from human embryonic stem cells (hESCs).

[0020] 5. The method of paragraph 4, wherein the obtaining step comprises contacting hESCs with at least one growth factor from the TGFβ superfamily so as to permit differentiation of at least some of said hESCs to definitive endoderm cells.

[0021] 6. The method of paragraph 5, wherein said at least one growth factor from the TGFβ superfamily comprises activin A.

[0022] 7. The method of paragraph 1, wherein the step of obtaining said cell culture comprising definitive endoderm further comprises obtaining a portion of an existing definitive endoderm culture.

[0023] 8. The method of paragraph 1, wherein said cell population enriched in definitive endoderm cells is substantially free of cells other than definitive endoderm cells.

[0024] 9. The method of paragraph 1, wherein said cell population enriched in definitive endoderm cells comprises at least about 96% definitive endoderm cells.

[0025] 10. The method of paragraph 1, wherein said cell population enriched in definitive endoderm cells comprises at least about 97% definitive endoderm cells.

[0026] 11. The method of paragraph 1, wherein said cell population enriched in definitive endoderm cells comprises at least about 98% definitive endoderm cells.

[0027] 12. The method of paragraph 1, wherein said cell population enriched in definitive endoderm cells comprises at least about 99% definitive endoderm cells.

[0028] 13. The method of paragraph 1, wherein said cell population enriched in definitive endoderm cells comprises about 100% definitive endoderm cells.

[0029] 14. The method of paragraph 1, wherein said isolating step comprises providing said cell culture with a reagent which binds to a marker expressed in said definitive endoderm cells but which is not substantially expressed in said other cells present in the cell culture, and separating said definitive endoderm cells bound to said reagent from said other cells present in the cell culture, thereby producing a cell population enriched in definitive endoderm cells.

[0030] 15. The method of paragraph 14, wherein said marker is CXCR4.

[0031] 16. The method of paragraph 14, wherein said reagent is an antibody.

[0032] 17. The method of paragraph 16, wherein said antibody has affinity for CXCR4.

[0033] 18. The method of paragraph 14, wherein said definitive endoderm cells bound to said reagent are separated from said other cells present in the cell culture by fluorescence activated cell sorting (FACS).

[0034] 19. The method of paragraph 1, wherein said isolating step comprises separating fluorescently-labeled definitive endoderm cells from unlabeled cells.

[0035] 20. The method of paragraph 19, wherein said fluorescently-labeled definitive endoderm cells are labeled as a result of the expression of enhanced green fluorescent protein (EGFP).

[0036] 21. The method of paragraph 20, wherein the expression of EGFP is under control of the SOX17 promoter.

[0037] 22. The method of paragraph 20, wherein the expression of EGFP is under control of the CXCR4 promoter.

[0038] 23. The method of paragraph 19, wherein said fluorescently-labeled definitive endoderm cells are separated from unlabeled cells by FACS.

[0039] 24. The method of paragraph 1, wherein said culturing step comprises plating said population enriched in definitive endoderm cells or a portion thereof.

[0040] 25. The method of paragraph 24, wherein said population enriched in definitive endoderm cells or a portion thereof is plated on a surface coated with human fibronectin.

[0041] 26. The method of paragraph 25, wherein said surface is coated with poly-ornithine.

[0042] 27. The method of paragraph 1, wherein said culturing step comprises incubating said population enriched in definitive endoderm cells or a portion thereof in a medium comprising about 2% (v/v) serum.

[0043] 28. The method of paragraph 1, wherein said culturing step comprises incubating said population enriched in definitive endoderm cells or a portion thereof in a medium comprising greater than about 2% (v/v) serum.

[0044] 29. The method of paragraph 1, wherein said culturing step comprises incubating said population enriched in definitive endoderm cells or a portion thereof in a medium comprising less than about 2% (v/v) serum.

[0045] 30. The method of paragraph 1, wherein said culturing step comprises incubating said population enriched in definitive endoderm cells or a portion thereof in a medium comprising at least one growth factor.

[0046] 31. The method of paragraph 30, wherein said at least one growth factor is a growth factor from the TGFβ superfamily of growth factors.

[0047] 32. The method of paragraph 31, wherein said at least one growth factor from the TGFβ superfamily of growth factors comprises activin A.

[0048] 33. The method of paragraph 32, wherein said activin A is present in said medium at a concentration of about 100 ng/ml.

[0049] 34. The method of paragraph 30, wherein said at least one growth factor comprises IGF1.

[0050] 35. The method of paragraph 34, wherein said IGF1 is present in said medium at a concentration of about 100 ng/ml.

[0051] 36. The method of paragraph 30, wherein said at least one growth factor comprises a combination of activin A and IGF1.

[0052] 37. The method of paragraph 30, wherein said at least one growth factor comprises bFGF.

[0053] 38. The method of paragraph 37, wherein said bFGF is present in said medium at a concentration of about 12 ng/ml.

[0054] 39. The method of paragraph 30, wherein said at least one growth factor comprises EGF.

[0055] 40. The method of paragraph 39, wherein said EGF is present in said medium at a concentration of about 10 ng/ml.

[0056] 41. The method of paragraph 30, wherein said at least one growth factor comprises a combination of activin A, bFGF and EGF.

[0057] 42. An expanded definitive endoderm cell population produced by the method of paragraph 1.

[0058] 43. A method of expanding definitive endoderm cells in culture, said method comprising the steps of: (a) obtaining a cell culture comprising definitive endoderm cells, and (b) passaging said definitive endoderm cells, thereby producing a plurality of cell cultures comprising definitive endoderm cells.

[0059] 44. The method of paragraph 43, wherein the step of passaging said definitive endoderm cells comprises providing at least one enzyme to said cell culture.

[0060] 45. The method of paragraph 44, wherein said at least one enzyme comprises at least one protease.

[0061] 46. The method of paragraph 45, wherein said at least one protease comprises trypsin.

[0062] 47. The method of paragraph 43, wherein the step of passaging said definitive endoderm cells comprises mechanically disrupting contacts between said definitive endoderm cells.

[0063] 48. The method of paragraph 43, wherein the step of passaging said definitive endoderm cells comprises incubating said definitive endoderm cells in a cell dispersal buffer.

[0064] 49. The method of paragraph 43, wherein said definitive endoderm cells are attached to a substrate.

[0065] 50. The method of paragraph 49, wherein the step of passaging said definitive endoderm cells comprises detaching said definitive endoderm cells from said substrate.

[0066] 51. The method of paragraph 50, wherein said substrate is a surface of a tissue culture flask.

[0067] 52. The method of paragraph 50, wherein said substrate is a surface of a microtiter plate.

[0068] 53. An expanded definitive endoderm cell population produced by the method of paragraph 43.

[0069] It will be appreciated that the methods and compositions described above relate to cells cultured *in vitro*. However, the above-described *in vitro* differentiated cell compositions may be used for *in vivo* applications.

[0070] Additional embodiments may also be found in United States Provisional Patent Application No. 60/532,004, entitled DEFINITIVE ENDODERM, filed December 23, 2003; U.S. Provisional Patent Application No. 60/566,293, entitled PDX1 EXPRESSING ENDODERM, filed April 27, 2004; U.S. Provisional Patent Application No. 60/586,566, entitled CHEMOKINE CELL SURFACE RECEPTOR FOR THE ISOLATION OF DEFINITIVE ENDODERM, filed July 9, 2004; U.S. Provisional Patent Application No. 60/587,942, entitled CHEMOKINE CELL SURFACE RECEPTOR FOR THE ISOLATION OF DEFINITIVE ENDODERM, filed July 14, 2004; U.S. Patent Application No. 11/021,618, entitled DEFINITIVE ENDODERM, filed December 23, 2004; U.S. Patent Application No. 11/115,868, entitled PDX1 EXPRESSING ENDODERM, filed April 26, 2005; U.S. Patent Application No. 11/165,305, entitled METHODS FOR IDENTIFYING FACTORS FOR

DIFFERENTIATING DEFINITIVE ENDODERM, filed June 23, 2005; U.S. Provisional Patent Application No. 60/693364, entitled PREPRIMITIVE STREAK AND MESENDODERM CELLS, filed June 23, 2005; U.S. Provisional Patent Application No. 60/693,317, entitled EXPANSION OF ISOLATED DEFINITIVE ENDODERM CELLS, filed June 23, 2005; and U.S. Provisional Patent Application No. 60/736,598, entitled MARKERS OF DEFINITIVE ENDODERM, filed November 14, 2005, the disclosures of which are incorporated herein by reference in their entireties.

Brief Description of the Drawings

[0071] Figure 1 is a schematic of a proposed differentiation pathway for the production of beta-cells from hESCs. The first step in the pathway commits the ES cell to the definitive endoderm lineage and represents one of the earliest known steps in the further differentiation of ES cells to pancreatic endoderm, endocrine endoderm, or islet/beta-cell. Some factors useful for mediating this transition are members of the TGFβ family which include, but are not limited to, activins and nodals. Exemplary markers for defining the definitive endoderm target cell are SOX17, GATA4, HNF3b, MIX1 and CXCR4.

[0072] Figure 2 is a diagram of the human SOX17 cDNA which displays the positions of conserved motifs and highlights the region used for the immunization procedure by GENOVAC.

[0073] Figure 3 is a relational dendrogram illustrating that SOX17 is most closely related to SOX7 and somewhat less to SOX18. The SOX17 proteins are more closely related among species homologs than to other members of the SOX group F subfamily within the same species.

[0074] Figure 4 is a Western blot probed with the rat anti-SOX17 antibody. This blot demonstrates the specificity of this antibody for human SOX17 protein over-expressed in fibroblasts (lane 1) and a lack of immunoreactivity with EGFP (lane 2) or the most closely related SOX family member, SOX7 (lane 3).

[0075] Figures 5A-B are micrographs showing a cluster of $SOX17^+$ cells that display a significant number of AFP^+ co-labeled cells (A). This is in striking contrast to other $SOX17^+$ clusters (B) where little or no AFP^+ cells are observed.

[0076] Figures 6A-C are micrographs showing parietal endoderm and SOX17. Panel A shows immunocytochemistry for human Thrombomodulin (TM) protein located

on the cell surface of parietal endoderm cells in randomly differentiated cultures of hES cells. Panel B is the identical field shown in A double-labeled for TM and SOX17. Panel C is the phase contrast image of the same field with DAPI labeled nuclei. Note the complete correlation of DAPI labeled nuclei and SOX17 labeling.

[0077] Figures 7A-B are bar charts showing SOX17 gene expression by quantitative PCR (Q-PCR) and anti-SOX17 positive cells by SOX17-specific antibody. Panel A shows that activin A increases SOX17 gene expression while retinoic acid (RA) strongly suppresses SOX17 expression relative to the undifferentiated control media (SR20). Panel B shows the identical pattern as well as a similar magnitude of these changes is reflected in $SOX17^+$ cell number, indicating that Q-PCR measurement of SOX17 gene expression is very reflective of changes at the single cell level.

[0078] Figure 8A is a bar chart which shows that a culture of differentiating hESCs in the presence of activin A maintains a low level of AFP gene expression while cells allowed to randomly differentiate in 10% fetal bovine serum (FBS) exhibit a strong upregulation of AFP. The difference in expression levels is approximately 7-fold.

[0079] Figures 8B-C are images of two micrographs showing that the suppression of AFP expression by activin A is also evident at the single cell level as indicated by the very rare and small clusters of AFP^+ cells observed in activin A treatment conditions (bottom) relative to 10% FBS alone (top).

[0080] Figures 9A-B are comparative images showing the quantitation of the AFP^+ cell number using flow cytometry. This figure demonstrates that the magnitude of change in AFP gene expression (Figure 8A) in the presence (right panel) and absence (left panel) of activin A exactly corresponds to the number of AFP^+ cells, further supporting the utility of Q-PCR analyses to indicate changes occurring at the individual cell level.

[0081] Figures 10A-F are micrographs which show that exposure of hESCs to nodal, activin A and activin B (NAA) yields a striking increase in the number of $SOX17^+$ cells over the period of 5 days (A-C). By comparing to the relative abundance of $SOX17^+$ cells to the total number of cells present in each field, as indicated by DAPI stained nuclei (D-F), it can be seen that approximately 30-50% of all cells are immunoreactive for SOX17 after five days treatment with NAA.

[0082] Figure 11 is a bar chart which demonstrates that activin A (0, 10, 30 or 100 ng/ml) dose-dependently increases SOX17 gene expression in differentiating hESCs. Increased expression is already robust after 3 days of treatment on adherent cultures and continues through subsequent 1, 3 and 5 days of suspension culture as well.

[0083] Figures 12A-C are bar charts which demonstrate the effect of activin A on the expression of MIXL1 (panel A), GATA4 (panel B) and HNF3b (panel C). Activin A dose-dependent increases are also observed for three other markers of definitive endoderm; MIXL1, GATA4 and HNF3b. The magnitudes of increased expression in response to activin dose are strikingly similar to those observed for SOX17, strongly indicating that activin A is specifying a population of cells that co-express all four genes ($SOX17^+$, $MIXL1^+$, $GATA4^+$ and $HNF3b^+$).

[0084] Figures 13A-C are bar charts which demonstrate the effect of activin A on the expression of AFP (panel A), SOX7 (panel B) and SPARC (panel C). There is an activin A dose-dependent decrease in expression of the visceral endoderm marker AFP. Markers of primitive endoderm (SOX7) and parietal endoderm (SPARC) remain either unchanged or exhibit suppression at some time points indicating that activin A does not act to specify these extra-embryonic endoderm cell types. This further supports the fact that the increased expression of SOX17, MIXL1, GATA4, and HNF3b are due to an increase in the number of definitive endoderm cells in response to activin A.

[0085] Figures 14A-B are bar charts showing the effect of activin A on ZIC1 (panel A) and Brachyury expression (panel B) Consistent expression of the neural marker ZIC1 demonstrates that there is not a dose-dependent effect of activin A on neural differentiation. There is a notable suppression of mesoderm differentiation mediated by 100 ng/ml of activin A treatment as indicated by the decreased expression of brachyury. This is likely the result of the increased specification of definitive endoderm from the mesendoderm precursors. Lower levels of activin A treatment (10 and 30 ng/ml) maintain the expression of brachyury at later time points of differentiation relative to untreated control cultures.

[0086] Figures 15A-B are micrographs showing decreased parietal endoderm differentiation in response to treatment with activins. Regions of TM^{hi} parietal endoderm are found through the culture (A) when differentiated in serum alone, while differentiation to TM^+ cells is scarce when activins are included (B) and overall intensity of TM immunoreactivity is lower.

[0087] Figures 16A-D are micrographs which show marker expression in response to treatment with activin A and activin B. hESCs were treated for four consecutive days with activin A and activin B and triple labeled with SOX17, AFP and TM antibodies. Panel A - SOX17; Panel B - AFP; Panel C - TM; and Panel D -

Phase/DAPI. Notice the numerous SOX17 positive cells (A) associated with the complete absence of AFP (B) and TM (C) immunoreactivity.

[0088] Figure 17 is a micrograph showing the appearance of definitive endoderm and visceral endoderm in vitro from hESCs. The regions of visceral endoderm are identified by $AFP^{hi}/SOX17^{lo/-}$ while definitive endoderm displays the complete opposite profile, $SOX17^{hi}/AFP^{lo/-}$. This field was selectively chosen due to the proximity of these two regions to each other. However, there are numerous times when $SOX17^{hi}/AFP^{lo/-}$ regions are observed in absolute isolation from any regions of AFP^{hi} cells, suggesting the separate origination of the definitive endoderm cells from visceral endoderm cells.

[0089] Figure 18 is a diagram depicting the TGFβ family of ligands and receptors. Factors activating AR Smads and BR Smads are useful in the production of definitive endoderm from human embryonic stem cells (see, *J Cell Physiol.*187:265-76).

[0090] Figure 19 is a bar chart showing the induction of SOX17 expression over time as a result of treatment with individual and combinations of TGFβ factors.

[0091] Figure 20 is a bar chart showing the increase in $SOX17^{+}$ cell number with time as a result of treatment with combinations of TGFβ factors.

[0092] Figure 21 is a bar chart showing induction of SOX17 expression over time as a result of treatment with combinations of TGFβ factors.

[0093] Figure 22 is a bar chart showing that activin A induces a dose-dependent increase in $SOX17^{+}$ cell number.

[0094] Figure 23 is a bar chart showing that addition of Wnt3a to activin A and activin B treated cultures increases SOX17 expression above the levels induced by activin A and activin B alone.

[0095] Figures 24A-C are bar charts showing differentiation to definitive endoderm is enhanced in low FBS conditions. Treatment of hESCs with activins A and B in media containing 2% FBS (2AA) yields a 2-3 times greater level of SOX17 expression as compared to the same treatment in 10% FBS media (10AA) (panel A). Induction of the definitive endoderm marker MIXL1 (panel B) is also affected in the same way and the suppression of AFP (visceral endoderm) (panel C) is greater in 2% FBS than in 10% FBS conditions.

[0096] Figures 25A-D are micrographs which show $SOX17^{+}$ cells are dividing in culture. SOX17 immunoreactive cells are present at the differentiating edge of an

hESC colony (C, D) and are labeled with proliferating cell nuclear antigen (PCNA) (panel B) yet are not co-labeled with OCT4 (panel C). In addition, clear mitotic figures can be seen by DAPI labeling of nuclei in both $SOX17^+$ cells (arrows) as well as $OCT4^+$, undifferentiated hESCs (arrowheads) (D).

[0097] Figure 26 is a bar chart showing the relative expression level of CXCR4 in differentiating hESCs under various media conditions.

[0098] Figures 27A-D are bar charts that show how a panel of definitive endoderm markers share a very similar pattern of expression to CXCR4 across the same differentiation treatments displayed in Figure 26.

[0099] Figures 28A-E are bar charts showing how markers for mesoderm (BRACHYURY, MOX1), ectoderm (SOX1, ZIC1) and visceral endoderm (SOX7) exhibit an inverse relationship to CXCR4 expression across the same treatments displayed in Figure 26.

[0100] Figures 29A-F are micrographs that show the relative difference in SOX17 immunoreactive cells across three of the media conditions displayed in Figures 26-28.

[0101] Figures 30A-C are flow cytometry dot plots that demonstrate the increase in $CXCR4^+$ cell number with increasing concentration of activin A added to the differentiation media.

[0102] Figures 31A-D are bar charts that show the $CXCR4^+$ cells isolated from the high dose activin A treatment (A100-CX+) are even further enriched for definitive endoderm markers than the parent population (A100).

[0103] Figure 32 is a bar chart showing gene expression from $CXCR4^+$ and $CXCR4^-$ cells isolated using fluorescence-activated cell sorting (FACS) as well as gene expression in the parent populations. This demonstrates that the $CXCR4^+$ cells contain essentially all the CXCR4 gene expression present in each parent population and the $CXCR4^-$ populations contain very little or no CXCR4 gene expression.

[0104] Figures 33A-D are bar charts that demonstrate the depletion of mesoderm (BRACHYURY, MOX1), ectoderm (ZIC1) and visceral endoderm (SOX7) gene expression in the CXCR4+ cells isolated from the high dose activin A treatment which is already suppressed in expression of these non-definitive endoderm markers.

[0105] Figures 34A-M are bar charts showing the expression patterns of marker genes that can be used to identify definitive endoderm cells. The expression analysis of definitive endoderm markers, FGF17, VWF, CALCR, FOXQ1, CMKOR1 and

CRIP1 is shown in panels G-L, respectively. The expression analysis of previously described lineage marking genes, SOX17, SOX7, SOX17/SOX7, TM, ZIC1, and MOX1 is shown in panels A-F, respectively. Panel M shows the expression analysis of CXCR4. With respect to each of panels A-M, the column labeled hESC indicates gene expression from purified human embryonic stem cells; 2NF indicates cells treated with 2% FBS, no activin addition; 0.1A100 indicates cells treated with 0.1% FBS, 100 ng/ml activin A; 1A100 indicates cells treated with 1% FBS, 100 ng/ml activin A; and 2A100 indicates cells treated with 2% FBS, 100 ng/ml activin A.

[0106] Figures 35A D are bar charts showing the expression patterns of definitive endoderm marker genes in cell cultures maintained for 36 days under various growth conditions. The expression analysis of definitive endoderm markers SOX17, GSC, MIXL1 and CXCR4 is shown in panels A-D, respectively. EB and EV are used to designate to separate cell populations each from the hCyT25 hESC line. The abbreviation NF indicates cells grown in the absence of activin A, whereas A100 indicates cells grown in the presence of 100 ng/ml this factor. EGF indicates 50 ng/ml epidermal growth factor.

[0107] Figure 36 is a diagram showing the cell differentiation, isolation and expansion procedure for definitive endoderm cells. The abbreviations are as follows: hESC refers to human embryonic stem cells; d5 hESC-DE refers to unpurified definitive endoderm cells; d6 FACS-DE refers to CXCR4 antibody/FACS purified definitive endoderm cells; and p1 d10 FACS-DE refers to purified definitive endoderm cells that have been passaged once and grown for 10 additional days subsequent to passage. RNA samples are taken and analyzed for marker expression at each of the indicated days.

[0108] Figures 37A-F are bar charts showing the expression patterns of various embryonic cell lineage marker genes in cell cultures that have been differentiated to definitive endoderm then subsequently purified using the CXCR4 antibody in conjunction with fluorescence activated cell sorting (FACS). The abbreviations are as follows: p96 hESC refers to mRNA from the 96th passage of CyT25 human embryonic stem cells; d5 DE refers to mRNA from unpurified definitive endoderm cells on the fifth day of differentiation from p96 hESCs; NF d6-FACS refers to mRNA taken 11 days post differentiation from CXCR4 antibody/FACS purified definitive endoderm cells incubated in the absence of activin A; A refers to mRNA taken 11 days post differentiation from CXCR4 antibody/FACS purified definitive endoderm cells incubated in the presence of 100 ng/ml activin A; AI refers to mRNA taken 11 days post differentiation from CXCR4

antibody/FACS purified definitive endoderm cells incubated in the presence of 100 ng/ml activin A and 100 ng/ml IGF1; AFE refers to mRNA taken 11 days post differentiation from CXCR4 antibody/FACS purified definitive endoderm cells incubated in the presence of 100 ng/ml activin A, 12 ng/ml bFGF and 10 ng/ml EGF; NF p1-d10-FACS refers to mRNA taken 10 days post passage of CXCR4 antibody/FACS purified definitive endoderm cells incubated in the absence of activin A; A refers to mRNA taken 10 days post passage from CXCR4 antibody/FACS purified definitive endoderm cells incubated in the presence of 100 ng/ml activin A; AI refers to mRNA taken 10 days post passage from CXCR4 antibody/FACS purified definitive endoderm cells incubated in the presence of 100 ng/ml activin A and 100 ng/ml IGF1; and AFE refers to mRNA taken 10 days post passage from CXCR4 antibody/FACS purified definitive endoderm cells incubated in the presence of 100 ng/ml activin A, 12 ng/ml bFGF and 10 ng/ml EGF. Panel A – SOX17; B – GSC; C – OCT4; D – Brachyury; E – ZIC1; F – SOX1..

Detailed Description

[0109] A crucial stage in early human development termed gastrulation occurs 2-3 weeks after fertilization. Gastrulation is extremely significant because it is at this time that the three primary germ layers are first specified and organized (Lu et al., 2001; Schoenwolf and Smith, 2000). The ectoderm is responsible for the eventual formation of the outer coverings of the body and the entire nervous system whereas the heart, blood, bone, skeletal muscle and other connective tissues are derived from the mesoderm. Definitive endoderm is defined as the germ layer that is responsible for formation of the entire gut tube which includes the esophagus, stomach and small and large intestines, and the organs which derive from the gut tube such as the lungs, liver, thymus, parathyroid and thyroid glands, gall bladder and pancreas (Grapin-Botton and Melton, 2000; Kimelman and Griffin, 2000; Tremblay et al., 2000; Wells and Melton, 1999; Wells and Melton, 2000). A very important distinction should be made between the definitive endoderm and the completely separate lineage of cells termed primitive endoderm. The primitive endoderm is primarily responsible for formation of extra-embryonic tissues, mainly the parietal and visceral endoderm portions of the placental yolk sac and the extracellular matrix material of Reichert's membrane.

[0110] During gastrulation, the process of definitive endoderm formation begins with a cellular migration event in which mesendoderm cells (cells competent to form mesoderm or endoderm) migrate through a structure called the primitive streak.

Definitive endoderm is derived from cells, which migrate through the anterior portion of the streak and through the node (a specialized structure at the anterior-most region of the streak). As migration occurs, definitive endoderm populates first the most anterior gut tube and culminates with the formation of the posterior end of the gut tube.

[0111] Definitive endoderm and endoderm cells derived therefrom represent important multipotent starting points for the derivation of cells which make up terminally differentiated tissues and/or organs derived from the definitive endoderm lineage. Such cells, tissues and/or organs are extremely useful in cell therapies. Because large numbers of cells are usually necessary for successful cell therapy applications, it is beneficial to begin differentiation procedures with large numbers of cells of a single cell type. As embryonic stem cells in culture differentiate to definitive endoderm, not every embryonic stem cell is converted to the definitive endoderm cell type. To overcome this problem, definitive endoderm cells growing in mixed cell cultures can be enriched, isolated and/or purified using the methodology described herein. After such enrichment, isolation and/or purification, the resulting definitive endoderm cells may not be easy to grow in culture. Methods described herein improve the ability of enriched, isolated and/or purified definitive endoderm cells to grow and expand in cell culture. Because definitive endoderm cells can now be expanded in culture subsequent to enrichment, isolation and/or purification, cells, tissues and/or organs derived from definitive endoderm cells can be produced in greater numbers.

[0112] Some embodiments of the present invention relate to methods of expanding definitive endoderm cells in cell culture. In some embodiments, definitive endoderm cells are enriched by separating these cells from other cells in a mixed cell culture. The enriched definitive endoderm cells are then cultured under conditions which permit their expansion.

Definitions

[0113] Certain terms and phrases as used throughout this application have the meanings provided as follows:

[0114] As used herein, "embryonic" refers to a range of developmental stages of an organism beginning with a single zygote and ending with a multicellular structure that no longer comprises pluripotent or totipotent cells other than developed gametic cells. In addition to embryos derived by gamete fusion, the term "embryonic" refers to embryos derived by somatic cell nuclear transfer.

[0115] As used herein, "multipotent" or "multipotent cell" refers to a cell type that can give rise to a limited number of other particular cell types.

[0116] As used herein, "expression" refers to the production of a material or substance as well as the level or amount of production of a material or substance. Thus, determining the expression of a specific marker refers to detecting either the relative or absolute amount of the marker that is expressed or simply detecting the presence or absence of the marker.

[0117] As used herein, "marker" refers to any molecule that can be observed or detected. For example, a marker can include, but is not limited to, a nucleic acid, such as a transcript of a specific gene, a polypeptide product of a gene, a non-gene product polypeptide, a glycoprotein, a carbohydrate, a glycolipd, a lipid, a lipoprotein or a small molecule (for example, molecules having a molecular weight of less than 10,000 amu)

[0118] When used in connection with cell cultures and/or cell populations, the term "portion" means any non-zero amount of the cell culture or cell population, which ranges from a single cell to the entirety of the cell culture or cells population.

[0119] With respect to cells in cell cultures or in cell populations, the phrase "substantially free of" means that the specified cell type of which the cell culture or cell population is free, is present in an amount of less than about 5% of the total number of cells present in the cell culture or cell population.

[0120] With respect to cell culture medium, as used herein, "low serum RPMI" refers to a low serum containing medium, wherein the serum concentration is gradually increased over a defined time period. For example, in one embodiment, low serum RPMI comprises a concentration of about 0.2% fetal bovine serum (FBS) on the first day of cell growth, about 0.5% FBS on the second day of cell growth and about 2% FBS on the third through fifth day of cell growth. In another embodiment, low serum RPMI comprises a concentration of about 0% on day one, about 0.2% on day two and about 2% on the third and subsequent days.

[0121] As used herein, the terms "bFGF" and "FGF2" are used interchangeably.

Definitive Endoderm Cells and Processes Related Thereto

[0122] Embodiments described herein relate to novel, defined processes for the production of definitive endoderm cells in culture by differentiating pluripotent cells, such as stem cells into multipotent definitive endoderm cells. As described above,

definitive endoderm cells do not differentiate into tissues produced from ectoderm or mesoderm, but rather, differentiate into the gut tube as well as organs that are derived from the gut tube. In certain preferred embodiments, the definitive endoderm cells are derived from hESCs. Such processes can provide the basis for efficient production of human endodermal derived tissues such as pancreas, liver, lung, stomach, intestine, thyroid and thymus. For example, production of definitive endoderm may be the first step in differentiation of a stem cell to a functional insulin-producing β-cell. To obtain useful quantities of insulin-producing β-cells, high efficiency of differentiation is desirable for each of the differentiation steps that occur prior to reaching the pancreatic islet/β-cell fate. Since differentiation of stem cells to definitive endoderm cells represents perhaps the earliest step towards the production of functional pancreatic islet/β-cells (as shown in Figure 1), high efficiency of differentiation at this step is particularly desirable.

[0123] In view of the desirability of efficient differentiation of pluripotent cells to definitive endoderm cells, some aspects of the differentiation processes described herein relate to *in vitro* methodology that results in approximately 50-80% conversion of pluripotent cells to definitive endoderm cells. Typically, such methods encompass the application of culture and growth factor conditions in a defined and temporally specified fashion. Further enrichment of the cell population for definitive endoderm cells can be achieved by isolation and/or purification of the definitive endoderm cells from other cells in the population by using a reagent that specifically binds to definitive endoderm cells. As such, some embodiments described herein relate to definitive endoderm cells as well as methods for producing and isolating and/or purifying such cells.

[0124] In order to determine the amount of definitive endoderm cells in a cell culture or cell population, a method of distinguishing this cell type from the other cells in the culture or in the population is desirable. Accordingly, certain embodiments described herein relate to cell markers whose presence, absence and/or relative expression levels are specific for definitive endoderm and methods for detecting and determining the expression of such markers.

[0125] In some embodiments described herein, the presence, absence and/or level of expression of a marker is determined by quantitative PCR (Q-PCR). For example, the amount of transcript produced by certain genetic markers, such as SOX17, CXCR4, OCT4, AFP, TM, SPARC, SOX7, MIXL1, GATA4, HNF3b, GSC, FGF17, VWF, CALCR, FOXQ1, CMKOR1, CRIP1 and other markers described herein is

determined by quantitative Q-PCR. In other embodiments, immunohistochemistry is used to detect the proteins expressed by the above-mentioned genes. In still other embodiments, Q-PCR and immunohistochemical techniques are both used to identify and determine the amount or relative proportions of such markers.

[0126] By using methods, such as those described above, to determine the expression of one or more appropriate markers, it is possible to identify definitive endoderm cells, as well as determine the proportion of definitive endoderm cells in a cell culture or cell population. For example, in some embodiments of the present invention, the definitive endoderm cells or cell populations that are produced express the SOX17 and/or the CXCR4 gene at a level of about 2 orders of magnitude greater than non-definitive endoderm cell types or cell populations. In other embodiments, the definitive endoderm cells or cell populations that are produced express the SOX17 and/or the CXCR4 gene at a level of more than 2 orders of magnitude greater than non-definitive endoderm cell types or cell populations. In still other embodiments, the definitive endoderm cells or cell populations that are produced express one or more of the markers selected from the group consisting of SOX17, CXCR4, GSC, FGF17, VWF, CALCR, FOXQ1, CMKOR1 and CRIP1 at a level of about 2 or more than 2 orders of magnitude greater than non-definitive endoderm cell types or cell populations. In some embodiments described herein, definitive endoderm cells do not substantially express PDX1.

[0127] Embodiments described herein also relate to definitive endoderm compositions. For example, some embodiments relate to cell cultures comprising definitive endoderm, whereas others relate to cell populations enriched in definitive endoderm cells. Some preferred embodiments relate to cell cultures which comprise definitive endoderm cells, wherein at least about 50-80% of the cells in culture are definitive endoderm cells. An especially preferred embodiment relates to cells cultures comprising human cells, wherein at least about 50-80% of the human cells in culture are definitive endoderm cells. Because the efficiency of the differentiation procedure can be adjusted by modifying certain parameters, which include but are not limited to, cell growth conditions, growth factor concentrations and the timing of culture steps, the differentiation procedures described herein can result in about 5%, about 10%, about 15%, about 20%, about 25%, about 30%, about 35%, about 40%, about 45%, about 50%, about 55%, about 60%, about 65%, about 70%, about 75%, about 80%, about 85%, about 90%, about 95%, or greater than about 95% conversion of pluripotent cells to definitive

endoderm. In other preferred embodiments, conversion of a pluripotent cell population, such as a stem cell population, to substantially pure definitive endoderm cell population is contemplated.

[0128] The compositions and methods described herein have several useful features. For example, the cell cultures and cell populations comprising definitive endoderm as well as the methods for producing such cell cultures and cell populations are useful for modeling the early stages of human development. Furthermore, the compositions and methods described herein can also serve for therapeutic intervention in disease states, such as diabetes mellitus. For example, since definitive endoderm serves as the source for only a limited number of tissues, it can be used in the development of pure tissue or cell types.

Production of Definitive Endoderm from Pluripotent Cells

[0129] Processes for differentiating pluripotent cells to produce cell cultures and enriched cell populations comprising definitive endoderm is described below and in US Patent No. 11/021,618, entitled DEFINITIVE ENDODERM, filed December 23, 2004, the disclosure of which is incorporated herein by reference in its entirety. In some of these processes, the pluripotent cells used as starting material are stem cells. In certain processes, definitive endoderm cell cultures and enriched cell populations comprising definitive endoderm cells are produced from embryonic stem cells. A preferred method for deriving definitive endoderm cells utilizes human embryonic stem cells as the starting material for definitive endoderm production. Such pluripotent cells can be cells that originate from the morula, embryonic inner cell mass or those obtained from embryonic gonadal ridges. Human embryonic stem cells can be maintained in culture in a pluripotent state without substantial differentiation using methods that are known in the art. Such methods are described, for example, in US Patent Nos. 5,453,357, 5,670,372, 5,690,926 5,843,780, 6,200,806 and 6,251,671 the disclosures of which are incorporated herein by reference in their entireties.

[0130] In some processes for producing definitive endoderm cells, hESCs are maintained on a feeder layer. In such processes, any feeder layer which allows hESCs to be maintained in a pluripotent state can be used. One commonly used feeder layer for the cultivation of human embryonic stem cells is a layer of mouse fibroblasts. More recently, human fibroblast feeder layers have been developed for use in the cultivation of hESCs (see US Patent Application No. 2002/0072117, the disclosure of which is incorporated

herein by reference in its entirety). Alternative processes for producing definitive endoderm permit the maintenance of pluripotent hESC without the use of a feeder layer. Methods of maintaining pluripotent hESCs under feeder-free conditions have been described in US Patent Application No. 2003/0175956, the disclosure of which is incorporated herein by reference in its entirety.

[0131] The human embryonic stem cells used herein can be maintained in culture either with or without serum. In some embryonic stem cell maintenance procedures, serum replacement is used. In others, serum free culture techniques, such as those described in US Patent Application No. 2003/0190748, the disclosure of which is incorporated herein by reference in its entirety, are used.

[0132] Stem cells are maintained in culture in a pluripotent state by routine passage until it is desired that they be differentiated into definitive endoderm. In some processes, differentiation to definitive endoderm is achieved by providing to the stem cell culture a growth factor of the TGFβ superfamily in an amount sufficient to promote differentiation to definitive endoderm. Growth factors of the TGFβ superfamily which are useful for the production of definitive endoderm are selected from the Nodal/Activin or BMP subgroups. In some preferred differentiation processes, the growth factor is selected from the group consisting of Nodal, activin A, activin B and BMP4. Additionally, the growth factor Wnt3a and other Wnt family members are useful for the production of definitive endoderm cells. In certain differentiation processes, combinations of any of the above-mentioned growth factors can be used.

[0133] With respect to some of the processes for the differentiation of pluripotent stem cells to definitive endoderm cells, the above-mentioned growth factors are provided to the cells so that the growth factors are present in the cultures at concentrations sufficient to promote differentiation of at least a portion of the stem cells to definitive endoderm cells. In some processes, the above-mentioned growth factors are present in the cell culture at a concentration of at least about 5 ng/ml, at least about 10 ng/ml, at least about 25 ng/ml, at least about 50 ng/ml, at least about 75 ng/ml, at least about 100 ng/ml, at least about 200 ng/ml, at least about 300 ng/ml, at least about 400 ng/ml, at least about 500 ng/ml, at least about 1000 ng/ml, at least about 2000 ng/ml, at least about 3000 ng/ml, at least about 4000 ng/ml, at least about 5000 ng/ml or more than about 5000 ng/ml.

[0134] In certain processes for the differentiation of pluripotent stem cells to definitive endoderm cells, the above-mentioned growth factors are removed from the cell culture subsequent to their addition. For example, the growth factors can be removed within about one day, about two days, about three days, about four days, about five days, about six days, about seven days, about eight days, about nine days or about ten days after their addition. In a preferred processes, the growth factors are removed about four days after their addition.

[0135] Cultures of definitive endoderm cells can be grown in medium containing reduced serum or no serum. Under certain culture conditions, serum concentrations can range from about 0.05% v/v to about 20% v/v. For example, in some differentiation processes, the serum concentration of the medium can be less than about 0.05% (v/v), less than about 0.1% (v/v), less than about 0.2% (v/v), less than about 0.3% (v/v), less than about 0.4% (v/v), less than about 0.5% (v/v), less than about 0.6% (v/v), less than about 0.7% (v/v), less than about 0.8% (v/v), less than about 0.9% (v/v), less than about 1% (v/v), less than about 2% (v/v), less than about 3% (v/v), less than about 4% (v/v), less than about 5% (v/v), less than about 6% (v/v), less than about 7% (v/v), less than about 8% (v/v), less than about 9% (v/v), less than about 10% (v/v), less than about 15% (v/v) or less than about 20% (v/v). In some processes, definitive endoderm cells are grown without serum or with serum replacement. In still other processes, definitive endoderm cells are grown in the presence of B27. In such processes, the concentration of B27 supplement can range from about 0.1% v/v to about 20% v/v.

Monitoring the Differentiation of Pluripotent Cells to Definitive Endoderm

[0136] The progression of the hESC culture to definitive endoderm can be monitored by determining the expression of markers characteristic of definitive endoderm. In some processes, the expression of certain markers is determined by detecting the presence or absence of the marker. Alternatively, the expression of certain markers can be determined by measuring the level at which the marker is present in the cells of the cell culture or cell population. In such processes, the measurement of marker expression can be qualitative or quantitative. One method of quantitating the expression of markers that are produced by marker genes is through the use of quantitative PCR (Q-PCR). Methods of performing Q-PCR are well known in the art. Other methods which are known in the art can also be used to quantitate marker gene expression. For example, the expression of a marker gene product can be detected by using antibodies specific for

the marker gene product of interest. In certain processes, the expression of marker genes characteristic of definitive endoderm as well as the lack of significant expression of marker genes characteristic of hESCs and other cell types is determined.

[0137] As described further in the Examples below, a reliable marker of definitive endoderm is the SOX17 gene. As such, the definitive endoderm cells produced by the processes described herein express the SOX17 marker gene, thereby producing the SOX17 gene product. Other markers of definitive endoderm are MIXL1, GATA4, HNF3b, GSC, FGF17, VWF, CALCR, FOXQ1, CMKOR1 and CRIP1. Since definitive endoderm cells express the SOX17 marker gene at a level higher than that of the SOX7 marker gene, which is characteristic of primitive and visceral endoderm (see Table 1), in some processes, the expression of both SOX17 and SOX7 is monitored. In other processes, expression of the both the SOX17 marker gene and the OCT4 marker gene, which is characteristic of hESCs, is monitored. Additionally, because definitive endoderm cells express the SOX17 marker gene at a level higher than that of the AFP, SPARC or Thrombomodulin (TM) marker genes, the expression of these genes can also be monitored.

[0138] Another marker of definitive endoderm is the CXCR4 gene. The CXCR4 gene encodes a cell surface chemokine receptor whose ligand is the chemoattractant SDF-1. The principal roles of the CXCR4 receptor-bearing cells in the adult are believed to be the migration of hematopoetic cells to the bone marrow, lymphocyte trafficking and the differentiation of various B cell and macrophage blood cell lineages [Kim, C., and Broxmeyer, H. J. Leukocyte Biol. 65, 6-15 (1999)]. The CXCR4 receptor also functions as a coreceptor for the entry of HIV-1 into T-cells [Feng, Y., et al. Science, 272, 872-877 (1996)]. In an extensive series of studies carried out by [McGrath, K.E. et al. Dev. Biology 213, 442-456 (1999)], the expression of the chemokine receptor CXCR4 and its unique ligand, SDF-1 [Kim, C., and Broxmyer, H., J. Leukocyte Biol. 65, 6-15 (1999)], were delineated during early development and adult life in the mouse. The CXCR4/SDF1 interaction in development became apparent when it was demonstrated that if either gene was disrupted in transgenic mice [Nagasawa et al. Nature, 382, 635-638 (1996)], Ma, Q., et al Immunity, 10, 463-471 (1999)] it resulted in late embryonic lethality. McGrath et al. demonstrated that CXCR4 is the most abundant chemokine receptor messenger RNA detected during early gastrulating embryos (E7.5) using a combination of RNase protection and *in situ* hybridization methodologies. In the gastrulating embryo, CXCR4/SDF-1 signaling appears to be mainly involved in inducing

migration of primitive-streak germlayer cells and is expressed on definitive endoderm, mesoderm and extraembryonic mesoderm present at this time. In E7.2-7.8 mouse embryos, CXCR4 and alpha-fetoprotein are mutually exclusive indicating a lack of expression in visceral endoderm [McGrath, K.E. et al. Dev. Biology 213, 442-456 (1999)].

[0139] Since definitive endoderm cells produced by differentiating pluripotent cells express the CXCR4 marker gene, expression of CXCR4 can be monitored in order to track the production of definitive endoderm cells. Additionally, definitive endoderm cells produced by the methods described herein express other markers of definitive endoderm including, but not limited to, SOX17, MIXL1, GATA4, HNF3b, GSC, FGF17, VWF, CALCR, FOXQ1, CMKOR1 and CRIP1. Since definitive endoderm cells express the CXCR4 marker gene at a level higher than that of the SOX7 marker gene, the expression of both CXCR4 and SOX7 can be monitored. In other processes, expression of both the CXCR4 marker gene and the OCT4 marker gene, is monitored. Additionally, because definitive endoderm cells express the CXCR4 marker gene at a level higher than that of the AFP, SPARC or Thrombomodulin (TM) marker genes, the expression of these genes can also be monitored.

[0140] It will be appreciated that expression of CXCR4 in endodermal cells does not preclude the expression of SOX17. As such, definitive endoderm cells produced by the processes described herein will substantially express SOX17 and CXCR4 but will not substantially express AFP, TM, SPARC or PDX1.

[0141] It will be appreciated that SOX17 and/or CXCR4 marker expression is induced over a range of different levels in definitive endoderm cells depending on the differentiation conditions. As such, in some embodiments described herein, the expression of the SOX17 marker and/or the CXCR4 marker in definitive endoderm cells or cell populations is at least about 2-fold higher to at least about 10,000-fold higher than the expression of the SOX17 marker and/or the CXCR4 marker in non-definitive endoderm cells or cell populations, for example pluripotent stem cells. In other embodiments, the expression of the SOX17 marker and/or the CXCR4 marker in definitive endoderm cells or cell populations is at least about 4-fold higher, at least about 6-fold higher, at least about 8-fold higher, at least about 10-fold higher, at least about 15-fold higher, at least about 20-fold higher, at least about 40-fold higher, at least about 80-fold higher, at least about 100-fold higher, at least about 150-fold higher, at least about 200-fold higher, at least about 500-fold higher, at least about 750-fold higher, at least

about 1000-fold higher, at least about 2500-fold higher, at least about 5000-fold higher, at least about 7500-fold higher or at least about 10,000-fold higher than the expression of the SOX17 marker and/or the CXCR4 marker in non-definitive endoderm cells or cell populations, for example pluripotent stem cells. In some embodiments, the expression of the SOX17 marker and/or CXCR4 marker in definitive endoderm cells or cell populations is infinitely higher than the expression of the SOX17 marker and/or the CXCR4 marker in non-definitive endoderm cells or cell populations, for example pluripotent stem cells.

[0142] It will also be appreciated that in some embodiments described herein, the expression of markers selected from the group consisting of GATA4, MIXL1, HNF3b, GSC, FGF17, VWF, CALCR, FOXQ1, CMKOR1 and CRIP1 in definitive endoderm cells or cell populations is increased as compared to the expression of GATA4, MIXL1, HNF3b, GSC, FGF17, VWF, CALCR, FOXQ1, CMKOR1 and CRIP1 in non-definitive endoderm cells or cell populations.

[0143] Additionally, it will be appreciated that there is a range of differences between the expression level of the SOX17 marker and the expression levels of the OCT4, SPARC, AFP, TM and/or SOX7 markers in definitive endoderm cells. Similarly, there exists a range of differences between the expression level of the CXCR4 marker and the expression levels of the OCT4, SPARC, AFP, TM and/or SOX7 markers in definitive endoderm cells. As such, in some embodiments described herein, the expression of the SOX17 marker or the CXCR4 marker is at least about 2-fold higher to at least about 10,000-fold higher than the expression of OCT4, SPARC, AFP, TM and/or SOX7 markers. In other embodiments, the expression of the SOX17 marker or the CXCR4 marker is at least about 4-fold higher, at least about 6-fold higher, at least about 8-fold higher, at least about 10-fold higher, at least about 15-fold higher, at least about 20-fold higher, at least about 40-fold higher, at least about 80-fold higher, at least about 100-fold higher, at least about 150-fold higher, at least about 200-fold higher, at least about 500-fold higher, at least about 750-fold higher, at least about 1000-fold higher, at least about 2500-fold higher, at least about 5000-fold higher, at least about 7500-fold higher or at least about 10,000-fold higher than the expression of OCT4, SPARC, AFP, TM and/or SOX7 markers. In some embodiments, OCT4, SPARC, AFP, TM and/or SOX7 markers are not significantly expressed in definitive endoderm cells.

[0144] It will also be appreciated that in some embodiments described herein, the expression of markers selected from the group consisting of GATA4, MIXL1, HNF3b, GSC, FGF17, VWF, CALCR, FOXQ1, CMKOR1 and CRIP1 in definitive

endoderm cells is increased as compared to the expression of OCT4, SPARC, AFP, TM and/or SOX7 in definitive endoderm cells.

Enrichment, Isolation and/or Purification of Definitive Endoderm

[0145] Definitive endoderm cells produced by any of the above-described processes can be enriched, isolated and/or purified by using an affinity tag that is specific for such cells. Examples of affinity tags specific for definitive endoderm cells are antibodies, ligands or other binding agents that are specific to a marker molecule, such as a polypeptide, that is present on the cell surface of definitive endoderm cells but which is not substantially present on other cell types that would be found in a cell culture produced by the methods described herein. In some processes, an antibody which binds to CXCR4 is used as an affinity tag for the enrichment, isolation or purification of definitive endoderm cells. In other processes, the chemokine SDF-1 or other molecules based on SDF-1 can also be used as affinity tags. Such molecules include, but not limited to, SDF-1 fragments, SDF-1 fusions or SDF-1 mimetics.

[0146] Methods for making antibodies and using them for cell isolation are known in the art and such methods can be implemented for use with the antibodies and definitive endoderm cells described herein. In one process, an antibody which binds to CXCR4 is attached to a magnetic bead and then allowed to bind to definitive endoderm cells in a cell culture which has been enzymatically treated to reduce intercellular and substrate adhesion. The cell/antibody/bead complexes are then exposed to a movable magnetic field which is used to separate bead-bound definitive endoderm cells from unbound cells. Once the definitive endoderm cells are physically separated from other cells in culture, the antibody binding is disrupted and the cells are replated in appropriate tissue culture medium.

[0147] Additional methods for obtaining enriched, isolated or purified definitive endoderm cell cultures or populations can also be used. For example, in some embodiments, the CXCR4 antibody is incubated with a definitive endoderm-containing cell culture that has been treated to reduce intercellular and substrate adhesion. The cells are then washed, centrifuged and resuspended. The cell suspension is then incubated with a secondary antibody, such as an FITC-conjugated antibody that is capable of binding to the primary antibody. The cells are then washed, centrifuged and resuspended in buffer. The cell suspension is then analyzed and sorted using a fluorescence activated cell sorter (FACS). CXCR4-positive cells are collected separately from CXCR4-negative cells,

thereby resulting in the isolation of such cell types. If desired, the isolated cell compositions can be further purified by using an alternate affinity-based method or by additional rounds of sorting using the same or different markers that are specific for definitive endoderm.

[0148] In still other processes, definitive endoderm cells are enriched, isolated and/or purified using a ligand or other molecule that binds to CXCR4. In some processes, the molecule is SDF-1 or a fragment, fusion or mimetic thereof.

[0149] In some embodiments of the processes described herein, definitive endoderm cells are fluorescently labeled then isolated from non-labeled cells by using a fluorescence activated cell sorter (FACS). In such embodiments, a nucleic acid encoding green fluorescent protein (GFP) or another nucleic acid encoding an expressible fluorescent marker gene is used to label PDX1-positive cells. For example, in some embodiments, at least one copy of a nucleic acid encoding GFP or a biologically active fragment thereof is introduced into a pluripotent cell, preferably a human embryonic stem cell, downstream of the SOX17 or CXCR4 promoter such that the expression of the GFP gene product or biologically active fragment thereof is under control of the SOX17 or CXCR4 promoter. In some embodiments, the entire coding region of the nucleic acid, which encodes SOX17 or CXCR4, is replaced by a nucleic acid encoding GFP or a biologically active fragment thereof. In other embodiments, the nucleic acid encoding GFP or a biologically active fragment thereof is fused in frame with at least a portion of the nucleic acid encoding SOX17 or CXCR4, thereby generating a fusion protein. In such embodiments, the fusion protein retains a fluorescent activity similar to GFP.

[0150] Fluorescently marked cells, such as the above-described pluripotent cells, are differentiated to definitive endoderm as described previously above. Because definitive endoderm cells express the fluorescent marker gene, whereas other cell types do not, definitive endoderm cells can be separated from the other cell types. In some embodiments, cell suspensions comprising a mixture of fluorescently-labeled definitive endoderm cells and unlabeled non-definitive endoderm cells are sorted using a FACS. Definitive endoderm cells are collected separately from non-fluorescing cells, thereby resulting in the isolation of definitive endoderm. If desired, the isolated cell compositions can be further purified by additional rounds of sorting using the same or different markers that are specific for definitive endoderm.

[0151] In preferred processes, definitive endoderm cells are enriched, isolated and/or purified from other non-definitive endoderm cells after the stem cell cultures are

induced to differentiate towards the definitive endoderm lineage. It will be appreciated that the above-described enrichment, isolation and purification procedures can be used with such cultures at any stage of differentiation.

[0152] In addition to the procedures just described, definitive endoderm cells may also be isolated by other techniques for cell isolation. Additionally, definitive endoderm cells may also be enriched or isolated by methods of serial subculture in growth conditions which promote the selective survival or selective expansion of the definitive endoderm cells.

[0153] Using the methods described herein, enriched, isolated and/or purified populations of definitive endoderm cells and or tissues can be produced *in vitro* from pluripotent cell cultures or cell populations, such as stem cell cultures or populations, which have undergone at least some differentiation. In some methods, the cells undergo random differentiation. In a preferred method, however, the cells are directed to differentiate primarily into definitive endoderm. Some preferred enrichment, isolation and/or purification methods relate to the *in vitro* production of definitive endoderm from human embryonic stem cells.

[0154] Using the methods described herein, cell populations or cell cultures can be enriched in definitive endoderm content by at least about 2- to about 1000-fold as compared to untreated cell populations or cell cultures. In some embodiments, definitive endoderm cells can be enriched by at least about 5- to about 500-fold as compared to untreated cell populations or cell cultures. In other embodiments, definitive endoderm cells can be enriched from at least about 10- to about 200-fold as compared to untreated cell populations or cell cultures. In still other embodiments, definitive endoderm cells can be enriched from at least about 20- to about 100-fold as compared to untreated cell populations or cell cultures. In yet other embodiments, definitive endoderm cells can be enriched from at least about 40- to about 80-fold as compared to untreated cell populations or cell cultures. In certain embodiments, definitive endoderm cells can be enriched from at least about 2- to about 20-fold as compared to untreated cell populations or cell cultures.

Compositions Comprising Definitive Endoderm

[0155] Cell compositions produced by the above-described methods include cell cultures comprising definitive endoderm and cell populations enriched in definitive endoderm. For example, cell cultures which comprise definitive endoderm cells, wherein

at least about 50-80% of the cells in culture are definitive endoderm cells, can be produced. Because the efficiency of the differentiation process can be adjusted by modifying certain parameters, which include but are not limited to, cell growth conditions, growth factor concentrations and the timing of culture steps, the differentiation procedures described herein can result in about 5%, about 10%, about 15%, about 20%, about 25%, about 30%, about 35%, about 40%, about 45%, about 50%, about 55%, about 60%, about 65%, about 70%, about 75%, about 80%, about 85%, about 90%, about 95%, or greater than about 95% conversion of pluripotent cells to definitive endoderm. In processes in which isolation of definitive endoderm cells is employed, for example, by using an affinity reagent that binds to the CXCR4 receptor, a substantially pure definitive endoderm cell population can be recovered.

[0156] Some embodiments described herein relate to compositions, such as cell populations and cell cultures, that comprise both pluripotent cells, such as stem cells, and definitive endoderm cells. For example, using the methods described herein, compositions comprising mixtures of hESCs and definitive endoderm cells can be produced. In some embodiments, compositions comprising at least about 5 definitive endoderm cells for about every 95 pluripotent cells are produced. In other embodiments, compositions comprising at least about 95 definitive endoderm cells for about every 5 pluripotent cells are produced. Additionally, compositions comprising other ratios of definitive endoderm cells to pluripotent cells are contemplated. For example, compositions comprising at least about 1 definitive endoderm cell for about every 1,000,000 pluripotent cells, at least about 1 definitive endoderm cell for about every 100,000 pluripotent cells, at least about 1 definitive endoderm cell for about every 10,000 pluripotent cells, at least about 1 definitive endoderm cell for about every 1000 pluripotent cells, at least about 1 definitive endoderm cell for about every 500 pluripotent cells, at least about 1 definitive endoderm cell for about every 100 pluripotent cells, at least about 1 definitive endoderm cell for about every 10 pluripotent cells, at least about 1 definitive endoderm cell for about every 5 pluripotent cells, at least about 1 definitive endoderm cell for about every 2 pluripotent cells, at least about 2 definitive endoderm cells for about every 1 pluripotent cell, at least about 5 definitive endoderm cells for about every 1 pluripotent cell, at least about 10 definitive endoderm cells for about every 1 pluripotent cell, at least about 20 definitive endoderm cells for about every 1 pluripotent cell, at least about 50 definitive endoderm cells for about every 1 pluripotent cell, at least about 100 definitive endoderm cells for about every 1 pluripotent cell, at least about 1000

definitive endoderm cells for about every 1 pluripotent cell, at least about 10,000 definitive endoderm cells for about every 1 pluripotent cell, at least about 100,000 definitive endoderm cells for about every 1 pluripotent cell and at least about 1,000,000 definitive endoderm cells for about every 1 pluripotent cell are contemplated. In some embodiments, the pluripotent cells are human pluripotent stem cells. In certain embodiments the stem cells are derived from a morula, the inner cell mass of an embryo or the gonadal ridges of an embryo. In certain other embodiments, the pluripotent cells are derived from the gondal or germ tissues of a multicellular structure that has developed past the embryonic stage.

[0157] Some embodiments described herein relate to cell cultures or cell populations comprising from at least about 5% definitive endoderm cells to at least about 95% definitive endoderm cells. In some embodiments the cell cultures or cell populations comprise mammalian cells. In preferred embodiments, the cell cultures or cell populations comprise human cells. For example, certain specific embodiments relate to cell cultures comprising human cells, wherein from at least about 5% to at least about 95% of the human cells are definitive endoderm cells. Other embodiments relate to cell cultures comprising human cells, wherein at least about 5%, at least about 10%, at least about 15%, at least about 20%, at least about 25%, at least about 30%, at least about 35%, at least about 40%, at least about 45%, at least about 50%, at least about 55%, at least about 60%, at least about 65%, at least about 70%, at least about 75%, at least about 80%, at least about 85%, at least about 90% or greater than 90% of the human cells are definitive endoderm cells. In embodiments where the cell cultures or cell populations comprise human feeder cells, the above percentages are calculated without respect to the human feeder cells in the cell cultures or cell populations.

[0158] Further embodiments described herein relate to compositions, such as cell cultures or cell populations, comprising human cells, such as human definitive endoderm cells, wherein the expression of either the SOX17 or the CXCR4 marker is greater than the expression of the OCT4, SPARC, alpha-fetoprotein (AFP), Thrombomodulin (TM) and/or SOX7 marker in at least about 5% of the human cells. In other embodiments, the expression of either the SOX17 or the CXCR4 marker is greater than the expression of the OCT4, SPARC, AFP, TM and/or SOX7 marker in at least about 10% of the human cells, in at least about 15% of the human cells, in at least about 20% of the human cells, in at least about 25% of the human cells, in at least about 30% of the human cells, in at least about 35% of the human cells, in at least about 40% of the

human cells, in at least about 45% of the human cells, in at least about 50% of the human cells, in at least about 55% of the human cells, in at least about 60% of the human cells, in at least about 65% of the human cells, in at least about 70% of the human cells, in at least about 75% of the human cells, in at least about 80% of the human cells, in at least about 85% of the human cells, in at least about 90% of the human cells, in at least about 95% of the human cells or in greater than 95% of the human cells. In embodiments where the cell cultures or cell populations comprise human feeder cells, the above percentages are calculated without respect to the human feeder cells in the cell cultures or cell populations.

[0159] It will be appreciated that some embodiments described herein relate to compositions, such as cell cultures or cell populations, comprising human cells, such as human definitive endoderm cells, wherein the expression of one or more markers selected from the group consisting of GATA4, MIXL1, HNF3b, GSC, FGF17, VWF, CALCR, FOXQ1, CMKOR1 and CRIP1 is greater than the expression of the OCT4, SPARC, AFP, TM and/or SOX7 markers in from at least about 5% to greater than at least about 95% of the human cells. In embodiments where the cell cultures or cell populations comprise human feeder cells, the above percentages are calculated without respect to the human feeder cells in the cell cultures or cell populations.

[0160] Still other embodiments described herein relate to compositions, such as cell cultures or cell populations, comprising human cells, such as human definitive endoderm cells, wherein the expression both the SOX17 and the CXCR4 marker is greater than the expression of the OCT4, SPARC, AFP, TM and/or SOX7 marker in at least about 5% of the human cells. In other embodiments, the expression of both the SOX17 and the CXCR4 marker is greater than the expression of the OCT4, SPARC, AFP, TM and/or SOX7 marker in at least about 10% of the human cells, in at least about 15% of the human cells, in at least about 20% of the human cells, in at least about 25% of the human cells, in at least about 30% of the human cells, in at least about 35% of the human cells, in at least about 40% of the human cells, in at least about 45% of the human cells, in at least about 50% of the human cells, in at least about 55% of the human cells, in at least about 60% of the human cells, in at least about 65% of the human cells, in at least about 70% of the human cells, in at least about 75% of the human cells, in at least about 80% of the human cells, in at least about 85% of the human cells, in at least about 90% of the human cells, in at least about 95% of the human cells or in greater than 95% of the human cells. In embodiments where the cell cultures or cell populations comprise human

feeder cells, the above percentages are calculated without respect to the human feeder cells in the cell cultures or cell populations.

[0161] It will be appreciated that some embodiments described herein relate to compositions, such as cell cultures or cell populations, comprising human cells, such as human definitive endoderm cells, wherein the expression of the GATA4, MIXL1, HNF3b, GSC, FGF17, VWF, CALCR, FOXQ1, CMKOR1 and CRIP1 markers is greater than the expression of the OCT4, SPARC, AFP, TM and/or SOX7 markers in from at least about 5% to greater than at least about 95% of the human cells. In embodiments where the cell cultures or cell populations comprise human feeder cells, the above percentages are calculated without respect to the human feeder cells in the cell cultures or cell populations.

[0162] Additional embodiments described herein relate to compositions, such as cell cultures or cell populations, comprising mammalian endodermal cells, such as human endoderm cells, wherein the expression of either the SOX17 or the CXCR4 marker is greater than the expression of the OCT4, SPARC, AFP, TM and/or SOX7 marker in at least about 5% of the endodermal cells. In other embodiments, the expression of either the SOX17 or the CXCR4 marker is greater than the expression of the OCT4, SPARC, AFP, TM and/or SOX7 marker in at least about 10% of the endodermal cells, in at least about 15% of the endodermal cells, in at least about 20% of the endodermal cells, in at least about 25% of the endodermal cells, in at least about 30% of the endodermal cells, in at least about 35% of the endodermal cells, in at least about 40% of the endodermal cells, in at least about 45% of the endodermal cells, in at least about 50% of the endodermal cells, in at least about 55% of the endodermal cells, in at least about 60% of the endodermal cells, in at least about 65% of the endodermal cells, in at least about 70% of the endodermal cells, in at least about 75% of the endodermal cells, in at least about 80% of the endodermal cells, in at least about 85% of the endodermal cells, in at least about 90% of the endodermal cells, in at least about 95% of the endodermal cells or in greater than 95% of the endodermal cells.

[0163] It will be appreciated that some embodiments described herein relate to compositions, such as cell cultures or cell populations comprising mammalian endodermal cells, wherein the expression of one or more markers selected from the group consisting of GATA4, MIXL1, HNF3b, GSC, FGF17, VWF, CALCR, FOXQ1, CMKOR1 and CRIP1 is greater than the expression of the OCT4, SPARC, AFP, TM

and/or SOX7 markers in from at least about 5% to greater than at least about 95% of the endodermal cells.

[0164] Still other embodiments described herein relate to compositions, such as cell cultures or cell populations, comprising mammalian endodermal cells, such as human endodermal cells, wherein the expression of both the SOX17 and the CXCR4 marker is greater than the expression of the OCT4, SPARC, AFP, TM and/or SOX7 marker in at least about 5% of the endodermal cells. In other embodiments, the expression of both the SOX17 and the CXCR4 marker is greater than the expression of the OCT4, SPARC, AFP, TM and/or SOX7 marker in at least about 10% of the endodermal cells, in at least about 15% of the endodermal cells, in at least about 20% of the endodermal cells, in at least about 25% of the endodermal cells, in at least about 30% of the endodermal cells, in at least about 35% of the endodermal cells, in at least about 40% of the endodermal cells, in at least about 45% of the endodermal cells, in at least about 50% of the endodermal cells, in at least about 55% of the endodermal cells, in at least about 60% of the endodermal cells, in at least about 65% of the endodermal cells, in at least about 70% of the endodermal cells, in at least about 75% of the endodermal cells, in at least about 80% of the endodermal cells, in at least about 85% of the endodermal cells, in at least about 90% of the endodermal cells, in at least about 95% of the endodermal cells or in greater than 95% of the endodermal cells.

[0165] It will be appreciated that some embodiments described herein relate to compositions, such as cell cultures or cell populations comprising mammalian endodermal cells, wherein the expression of the GATA4, MIXL1, HNF3b, GSC, FGF17, VWF, CALCR, FOXQ1, CMKOR1 and CRIP1 markers is greater than the expression of the OCT4, SPARC, AFP, TM and/or SOX7 markers in from at least about 5% to greater than at least about 95% of the endodermal cells.

[0166] Using the methods described herein, compositions comprising definitive endoderm cells substantially free of other cell types can be produced. In some embodiments described herein, the definitive endoderm cell populations or cell cultures produced by the methods described herein are substantially free of cells that significantly express the OCT4, SOX7, AFP, SPARC, TM, ZIC1 or BRACH marker genes.

[0167] In one embodiment, a description of a definitive endoderm cell based on the expression of marker genes is, SOX17 high, MIXL1 high, AFP low, SPARC low, Thrombomodulin low, SOX7 low, CXCR4 high.

Expansion of Definitive Endoderm Cells

[0168] According to some of the *in vitro* methods described herein, definitive endoderm cells are maintained, grown, passaged and/or expanded while in cell culture. In some embodiments the definitive endoderm cells are maintained, grown, passaged and/or expanded without any significant differentiation. In other words, in such embodiments, the definitive endoderm cells maintain the definitive endoderm phenotype while being maintained, grown, passaged and/or expanded in cell culture.

[0169] In some embodiments, definitive endoderm cells used in the expansion methods described herein, are multipotent cells that can differentiate into cells of the gut tube or organs derived therefrom. Such cells include, but are not limited to, cells of the pancreas, liver, lungs, stomach, intestine, thyroid, thymus, pharynx, gallbladder and urinary bladder as well as precursors of such cells. Additionally, these cells can further develop into higher order structures such as tissues and/or organs. In some embodiments, the definitive endoderm cells are human definitive endoderm cells.

[0170] Some embodiments of the methods described herein comprise a step of obtaining a cell culture comprising definitive endoderm cells. The cell culture can be a pure culture of definitive endoderm cells or a mixed cell culture that comprises definitive endoderm cells as well as cells of other types. For example, the cell culture can be a culture comprising both definitive endoderm cells and human embryonic stem cells (hESCs). In some embodiments, the definitive endoderm cell culture is obtained by differentiating *in vitro* cell cultures of hESCs. In certain embodiments, the hESCs are derived from a morula, the inner cell mass of an embryo or the gonadal ridges of an embryo. In certain other embodiments, the pluripotent cells are derived from the gonadal or germ tissues of a multicellular structure that has developed past the embryonic stage.

[0171] Methods of differentiating hESCs so as to produce cell cultures comprising human definitive endoderm cells have been described throughout this application and in U.S. Patent Application No. 11/021,618, entitled DEFINITIVE ENDODERM, filed December 23, 2004, the disclosure of which is incorporated herein by reference in its entirety. However, it will be appreciated that any known methods for producing human definitive endoderm cells from hESCs or from other human cell types can be used. In some embodiments described herein, the cultures of definitive endoderm cells produced by differentiating hESCs can be mixed definitive endoderm cultures, which comprise definitive endoderm cells and one or more types of other cells, enriched definitive endoderm cell cultures and/or purified definitive endoderm cell cultures. Some

methods for obtaining definitive endoderm cells from hESCs comprise contacting or otherwise providing the hESCs with at least one growth factor from the TGFβ superfamily. Such growth factors can include, but are not limited to, Nodal, activin A and activin B. In some embodiments, the growth factor is provided to the hESCs at a concentration ranging from 5 ng/ml to 5000 ng/ml. In certain embodiments the growth factor is provided to the hESCs in culture at a concentration of at least about 5 ng/ml, at least about 10 ng/ml, at least about 25 ng/ml, at least about 50 ng/ml, at least about 75 ng/ml, at least about 100 ng/ml, at least about 200 ng/ml, at least about 300 ng/ml, at least about 400 ng/ml, at least about 500 ng/ml, at least about 1000 ng/ml, at least about 2000 ng/ml, at least about 3000 ng/ml, at least about 4000 ng/ml, at least about 5000 ng/ml or more than about 5000 ng/ml.

[0172] In other embodiments of the methods described herein, definitive endoderm cells can be obtained from a pre-existing culture of definitive endoderm cells. In such embodiments, either a portion of or the entire culture may be used in the definitive endoderm expansion methods described herein.

[0173] In addition to obtaining cell cultures comprising definitive endoderm cells, some embodiments of the expansion methods described herein also comprise the step of isolating at least some of the definitive endoderm cells from the cell culture. In such embodiments, at least some of the definitive endoderm cells are separated from at least some of the other cells in the cell culture, thereby producing a cell population enriched in definitive endoderm cells. In some embodiments, at least some of the definitive endoderm cell are removed from the cell culture while at least some of the other cells remain in the cell culture. Other cells that are present in the cell culture can include, but are not limited to, hESCs, primitive endoderm, trophectoderm, mesoderm and ectoderm.

[0174] In other embodiments described herein, the isolating step comprises providing the cells in the cell culture with a reagent which binds to a marker expressed in said definitive endoderm cells but which is not substantially expressed in said other cells present in the cell culture. As described previously herein, in some embodiments, the marker can be any cell surface marker that is specific to definitive endoderm cells. One such marker that is described throughout this application (see especially the Examples below) is the CXCR4 marker. As described previously herein, the reagent-bound definitive endoderm cells can be separated from the non-reagent-bound cells by numerous

methods. For example, an antibody against the CXCR4 receptor that is selectively present on the surface of definitive endoderm cells, can be provided to definitive endoderm cells in a cell culture. Antibody-bound definitive endoderm cells can then be separated from other cells in the culture by, for example, fluorescent activated cell sorting (FACS), binding the antibody to a solid support or isolating appropriately tagged antibody in a magnetic field. In some embodiments, the antibody is released from the definitive endoderm cells after the separation process.

[0175] As an alternative means of separation, at least some of the definitive endoderm cells are separated from at least some of the other cells in the culture by specifically fluorescently labeling the definitive endoderm cells in culture and then separating the labeled cells from the unlabeled cells by FACS. As described previously above and in the Examples, in such embodiments, hESCs are transfected with a vector comprising a fluorescent reporter gene under the control of the promoter of a marker gene that is highly expressed in definitive endoderm cells but not significantly expressed in other cell types. In some embodiments, the fluorescent reporter gene is the gene encoding green fluorescent protein (GFP) or enhanced green fluorescent protein (EGFP). In some embodiments, the GFP and/or EGFP is expressed under the control of the SOX17 or the CXCR4 promoter. Transfected hESCs are then grown in culture in the presence of a differentiation factor that specifically induces the production of definitive endoderm. In preferred embodiments, the differentiation factor is activin A. In other preferred embodiments, activin A is added to the cell culture at a concentration of 100 ng/ml.

[0176] In some embodiments described herein, the enriched definitive endoderm cell populations that are produced as a result of the isolating step are substantially free of cells other than definitive endoderm cells. In other embodiments, the enriched definitive endoderm cell populations comprise from at least about 96% to at least about 100% definitive endoderm cells. In still other embodiments, the enriched definitive endoderm cell populations comprise from at least about 96%, at least about 97%, at least about 98%, at least about 99% and at least about 100% definitive endoderm cells.

[0177] According to further embodiments of the expansion methods described herein a cell culture step is contemplated. For example, some embodiments include a culturing step that comprises plating the population enriched in definitive endoderm cells or a portion of the population. In some embodiments, the cells are plated on a surface coated with human fibronectin. In other embodiments the plates are coated with poly-

ornithine. In still other embodiments, the plates are coated with poly-ornithine and human fibronectin. In preferred embodiments, the plates are IVF plates coated with both poly-ornithine and human fibronectin. It will be appreciated that although human fibronectin is a preferred coating for the plates described herein, fibronectin from other sources is sufficient for coating plates.

[0178] In other embodiments, the culturing step comprises incubating the enriched definitive endoderm cell population or portion thereof in an expansion medium comprising about 2% (v/v) serum. In some embodiments, the serum concentration can range from about 0% (v/v) to about 20% (v/v). For example, in some methods described herein, the serum concentration of the medium can be about 0.05% (v/v), about 0.1% (v/v), about 0.2% (v/v), about 0.3% (v/v), about 0.4% (v/v), about 0.5% (v/v), about 0.6% (v/v), about 0.7% (v/v), about 0.8% (v/v), about 0.9% (v/v), about 1% (v/v), about 2% (v/v), about 3% (v/v), about 4% (v/v), about 5% (v/v), about 6% (v/v), about 7% (v/v), about 8% (v/v), about 9% (v/v), about 10% (v/v), about 15% (v/v) or about 20% (v/v). In some embodiments, serum replacement is included in the medium.

[0179] In still other embodiments of the expansion methods described herein, the expansion medium also comprises at least one growth factor. In certain embodiments, the at least one growth factor is a growth factor comprises a member of the TGFβ superfamily. In such embodiments, the at least one growth factor of the TGFβ superfamily includes, but is not limited to Nodal, activin A, activin B and combinations of these growth factors. Alternatively, in some embodiments, the at least one growth factor can be IGF1 or a combination of IGF and a growth factor of the TGFβ superfamily. In other embodiments, the at least one growth factor can be bFGF, EGF or another growth factor. In yet other embodiments, the at least one growth factor can be a combination of bFGF, EGF and a growth factor of the TGFβ superfamily. In each of the above embodiments, one or more of the growth factors can be present at a concentration ranging from about 1 ng/ml to about 5000 ng/ml. In such embodiments, the concentration of growth factor in the medium is at least about 5 ng/ml, at least about 10 ng/ml, at least about 25 ng/ml, at least about 50 ng/ml, at least about 75 ng/ml, at least about 100 ng/ml, at least about 200 ng/ml, at least about 300 ng/ml, at least about 400 ng/ml, at least about 500 ng/ml, at least about 1000 ng/ml, at least about 2000 ng/ml, at least about 3000 ng/ml, at least about 4000 ng/ml, at least about 5000 ng/ml or more than about 5000 ng/ml. In certain embodiments, a combination of growth factors is present in the culture

medium. In such embodiments, each growth factor is present in the medium at a concentration of at a concentration of at least about 5 ng/ml, at least about 10 ng/ml, at least about 25 ng/ml, at least about 50 ng/ml, at least about 75 ng/ml, at least about 100 ng/ml, at least about 200 ng/ml, at least about 300 ng/ml, at least about 400 ng/ml, at least about 500 ng/ml, at least about 1000 ng/ml, at least about 2000 ng/ml, at least about 3000 ng/ml, at least about 4000 ng/ml, at least about 5000 ng/ml or more than about 5000 ng/ml.

[0180] In addition to the above-described expansion methods, in some embodiments definitive endoderm cells are expanded by first obtaining a cell culture comprising definitive endoderm cells and then passaging the definitive endoderm cells so as to produce a plurality of cell cultures comprising definitive endoderm cells. These methods of expanding definitive endoderm cells by passaging the cells can be performed using any definitive endoderm culture regardless of how such culture is obtained. For example, these methods can be performed as part of the culturing step that follows the cell isolation step in the above-described expansion methods, or alternatively, this methods can be performed using definitive endoderm cells that have been freshly differentiated from hESCs.

[0181] In accordance with certain aspects of the expansion methods described herein, the step of passaging definitive endoderm cells comprises providing at least one enzyme to a cell culture comprising definitive endoderm cells. For example, the at least one enzyme can be one or more enzymes selected from the group consisting of papain, pronase, type I collagenase, type II collagenase, type III collagenase, type IV collagenase, trypsin, hyaluronidase, elastase, DNase I, and dispase. In some embodiments, the at least one enzyme comprises at least one protease. In preferred embodiments, the at least one protease comprises trypsin. For example, in certain embodiments, definitive endoderm cells growing in a culture vessel are passaged with trypsin by first removing the culture medium from the cells. Next, a sterile trypsin solution is provided to the definitive endoderm cells for several minutes at room temperature. The trypsin solution is then gently removed so as not to disturb the cells. After the trypsin solution has been removed, the definitive endoderm cells are provided with a culture medium, such as RPMI with 2% (v/v) serum, and the cell culture vessel is then agitated so as to disrupt cell adhesions and generate a cell suspension. In some embodiments, the cell culture medium comprises trypsin inhibitor to inactivate residual trypsin.

[0182] It will be appreciated that trypsin can be provided in a variety of sterile solutions, for example, trypsin can be provided to the definitive endoderm cells in a balanced salt solution, such as Hanks balanced salt solution. Alternatively, trypsin can be provided to the definitive endoderm in a medium with or without serum, for example in low serum RPMI.

[0183] In accordance with other aspects of the expansion methods described herein, the step of passaging definitive endoderm cells comprises mechanically disrupting contacts between said definitive endoderm cells. Such mechanical disruption techniques should be sufficient to substantially disrupt cell contacts and the substrate, however, these techniques should not be so harsh as to affect cell viability. Mechanical cell disruption techniques, such as trituration, are known to those of ordinary skill in the art.

[0184] In accordance with yet other aspects of the expansion methods described herein, the step of passaging definitive endoderm cells comprises incubating said definitive endoderm cells in a cell dispersal buffer. The cell dispersal buffer can be any dispersal buffer known in the art, for example, commercially available chemical dissociation buffers.

[0185] In some embodiments of the expansion methods described herein, the definitive endoderm cells are grown in a cell culture vessel. Cell culture vessels can include, but are not limited to, tissue culture flasks and cell culture plate, such as microtiter plates. In some embodiments, the definitive endoderm cells in culture are attached to a substrate. In certain embodiments, the step of passaging said definitive endoderm cells comprises detaching said definitive endoderm cells from the substrate. In preferred embodiments, the substrate is a surface of a tissue culture flask. In other preferred embodiments, the substrate is a surface of a microtiter plate.

EXAMPLES

[0187] Many of the examples below describe the use of pluripotent human cells. Methods of producing pluripotent human cells are well known in the art and have been described numerous scientific publications, including U.S. Patent Nos. 5,453,357, 5,670,372, 5,690,926, 6,090,622, 6,200,806 and 6,251,671 as well as U.S. Patent Application Publication No. 2004/0229350, the disclosures of which are incorporated herein by reference in their entireties.

EXAMPLE 1

Human ES cells

[0188] For our studies of endoderm development we employed human embryonic stem cells, which are pluripotent and can divide seemingly indefinitely in culture while maintaining a normal karyotype. ES cells were derived from the 5-day-old embryo inner cell mass using either immunological or mechanical methods for isolation. In particular, the human embryonic stem cell line hESCyt-25 was derived from a supernumerary frozen embryo from an *in vitro* fertilization cycle following informed consent by the patient. Upon thawing the hatched blastocyst was plated on mouse embryonic fibroblasts (MEF), in ES medium (DMEM, 20% FBS, non essential amino acids, beta-mercaptoethanol, and FGF2). The embryo adhered to the culture dish and after approximately two weeks, regions of undifferentiated hESCs were transferred to new dishes with MEFs. Transfer was accomplished with mechanical cutting and a brief digestion with dispase, followed by mechanical removal of the cell clusters, washing and re-plating. Since derivation, hESCyt-25 has been serially passaged over 100 times. We employed the hESCyt-25 human embryonic stem cell line as our starting material for the production of definitive endoderm.

[0189] It will be appreciated by those of skill in the art that stem cells or other pluripotent cells can also be used as starting material for the differentiation procedures described herein. For example, cells obtained from embryonic gonadal ridges, which can be isolated by methods known in the art, can be used as pluripotent cellular starting material.

EXAMPLE 2

hESCyt-25 Characterization

[0190] The human embryonic stem cell line, hESCyt-25 has maintained a normal morphology, karyotype, growth and self-renewal properties over 18 months in culture. This cell line displays strong immunoreactivity for the OCT4, SSEA-4 and TRA-1-60 antigens, all of which, are characteristic of undifferentiated hESCs and displays alkaline phosphatase activity as well as a morphology identical to other established hESC lines. Furthermore, the human stem cell line, hESCyt-25, also readily forms embryoid bodies (EBs) when cultured in suspension. As a demonstration of its pluripotent nature, hESCyT-25 differentiates into various cell types that represent the three principal germ layers. Ectoderm production was demonstrated by Q-PCR for ZIC1 as well as

immunocytochemistry (ICC) for nestin and more mature neuronal markers. Immunocytochemical staining for β-III tubulin was observed in clusters of elongated cells, characteristic of early neurons. Previously, we treated EBs in suspension with retinoic acid, to induce differentiation of pluripotent stem cells to visceral endoderm (VE), an extra-embryonic lineage. Treated cells expressed high levels of α-fetoprotein (AFP) and SOX7, two markers of VE, by 54 hours of treatment. Cells differentiated in monolayer expressed AFP in sporadic patches as demonstrated by immunocytochemical staining. As will be described below, the hESCyT-25 cell line was also capable of forming definitive endoderm, as validated by real-time quantitative polymerase chain reaction (Q-PCR) and immunocytochemistry for SOX17, in the absence of AFP expression. To demonstrate differentiation to mesoderm, differentiating EBs were analyzed for Brachyury gene expression at several time points. Brachyury expression increased progressively over the course of the experiment. In view of the foregoing, the hESCyT-25 line is pluripotent as shown by the ability to form cells representing the three germ layers.

EXAMPLE 3

Production of SOX17 Antibody

[0191] A primary obstacle to the identification of definitive endoderm in hESC cultures is the lack of appropriate tools. We therefore undertook the production of an antibody raised against human SOX17 protein.

[0192] The marker SOX17 is expressed throughout the definitive endoderm as it forms during gastrulation and its expression is maintained in the gut tube (although levels of expression vary along the A-P axis) until around the onset of organogenesis. SOX17 is also expressed in a subset of extra-embryonic endoderm cells. No expression of this protein has been observed in mesoderm or ectoderm. It has now been discovered that SOX17 is an appropriate marker for the definitive endoderm lineage when used in conjunction with markers to exclude extra-embryonic lineages.

[0193] As described in detail herein, the SOX17 antibody was utilized to specifically examine effects of various treatments and differentiation procedures aimed at the production of SOX17 positive definitive endoderm cells. Other antibodies reactive to AFP, SPARC and Thrombomodulin were also employed to rule out the production of visceral and parietal endoderm (extra-embryonic endoderm).

[0194] In order to produce an antibody against SOX17, a portion of the human SOX17 cDNA (SEQ ID NO: 1) corresponding to amino acids 172-414 (SEQ ID NO: 2) in the carboxyterminal end of the SOX17 protein (Figure 2) was used for genetic immunization in rats at the antibody production company, GENOVAC (Freiberg, Germany), according to procedures developed there. Procedures for genetic immunization can be found in US Patent Nos. 5,830,876, 5,817,637, 6,165,993 and 6,261,281 as well as International Patent Application Publication Nos. WO00/29442 and WO99/13915, the disclosures of which are incorporated herein by reference in their entireties.

[0195] Other suitable methods for genetic immunization are also described in the non-patent literature. For example, Barry et al. describe the production of monoclonal antibodies by genetic immunization in *Biotechniques* **16**: 616-620, 1994, the disclosure of which is incorporated herein by reference in its entirety. Specific examples of genetic immunization methods to produce antibodies against specific proteins can be found, for example, in Costaglia *et al.*, (1998) Genetic immunization against the human thyrotropin receptor causes thyroiditis and allows production of monoclonal antibodies recognizing the native receptor, *J. Immunol.* 160: 1458-1465; Kilpatrick *et al* (1998) Gene gun delivered DNA-based immunizations mediate rapid production of murine monoclonal antibodies to the Flt-3 receptor, *Hybridoma* 17: 569-576; Schmolke *et al.*, (1998) Identification of hepatitis G virus particles in human serum by E2-specific monoclonal antibodies generated by DNA immunization, *J. Virol.* 72: 4541-4545; Krasemann et al., (1999) Generation of monoclonal antibodies against proteins with an unconventional nucleic acid-based immunization strategy, *J. Biotechnol.* 73: 119-129; and Ulivieri *et al.*, (1996) Generation of a monoclonal antibody to a defined portion of the *Heliobacter pylori* vacuolating cytotoxin by DNA immunization, *J. Biotechnol.* 51: 191-194, the disclosures of which are incorporated herein by reference in their entireties.

[0196] SOX7 and SOX18 are the closest Sox family relatives to SOX17 as depicted in the relational dendrogram shown in Figure 3. We employed the human SOX7 polypeptide as a negative control to demonstrate that the SOX17 antibody produced by genetic immunization is specific for SOX17 and does not react with its closest family member. In particular, SOX7 and other proteins were expressed in human fibroblasts, and then, analyzed for cross reactivity with the SOX17 antibody by Western blot and ICC. For example, the following methods were utilized for the production of the SOX17, SOX7 and EGFP expression vectors, their transfection into human fibroblasts and

analysis by Western blot. Expression vectors employed for the production of SOX17, SOX7, and EGFP were pCMV6 (OriGene Technologies, Inc., Rockville, MD), pCMV-SPORT6 (Invitrogen, Carlsbad, CA) and pEGFP-N1 (Clonetech, Palo Alto, CA), respectively. For protein production, telomerase immortalized MDX human fibroblasts were transiently transfected with supercoiled DNA in the presence of Lipofectamine 2000 (Invitrogen, Carlsbad, CA). Total cellular lysates were collected 36 hours post-transfection in 50 mM TRIS-HCl (pH 8), 150 mM NaCl, 0.1% SDS, 0.5% deoxycholate, containing a cocktail of protease inhibitors (Roche Diagnostics Corporation, Indianapolis, IN). Western blot analysis of 100 µg of cellular proteins, separated by SDS-PAGE on NuPAGE (4-12 % gradient polyacrylamide, Invitrogen, Carlsbad, CA), and transferred by electro-blotting onto PDVF membranes (Hercules, CA), were probed with a 1/1000 dilution of the rat SOX17 anti-serum in 10 mM TRIS-HCl (pH 8), 150 mM NaCl, 10% BSA, 0.05 % Tween-20 (Sigma, St. Louis, MO), followed by Alkaline Phosphatase conjugated anti-rat IgG (Jackson ImmunoResearch Laboratories, West Grove, PA), and revealed through Vector Black Alkaline Phosphatase staining (Vector Laboratories, Burlingame, CA). The proteins size standard used was wide range color markers (Sigma, St. Louis, MO).

[0197] In Figure 4, protein extracts made from human fibroblast cells that were transiently transfected with SOX17, SOX7 or EGFP cDNA's were probed on Western blots with the SOX17 antibody. Only the protein extract from hSOX17 transfected cells produced a band of ~51Kda which closely matched the predicted 46 Kda molecular weight of the human SOX17 protein. There was no reactivity of the SOX17 antibody to extracts made from either human SOX7 or EGFP transfected cells. Furthermore, the SOX17 antibody clearly labeled the nuclei of human fibroblast cells transfected with the hSOX17 expression construct but did not label cells transfected with EGFP alone. As such, the SOX17 antibody exhibits specificity by ICC.

EXAMPLE 4

Validation of SOX17 Antibody as a Marker of Definitive Endoderm

[0198] Partially differentiated hESCs were co-labeled with SOX17 and AFP antibodies to demonstrate that the SOX17 antibody is specific for human SOX17 protein and furthermore marks definitive endoderm. It has been demonstrated that SOX17, SOX7 (which is a closely related member of the SOX gene family subgroup F (Figure 3)) and AFP are each expressed in visceral endoderm. However, AFP and SOX7 are not

expressed in definitive endoderm cells at levels detectable by ICC, and thus, they can be employed as negative markers for bonifide definitive endoderm cells. It was shown that SOX17 antibody labels populations of cells that exist as discrete groupings of cells or are intermingled with AFP positive cells. In particular, Figure 5A shows that small numbers of SOX17 cells were co-labeled with AFP; however, regions were also found where there were little or no AFP^{+} cells in the field of $SOX17^{+}$ cells (Figure 5B). Similarly, since parietal endoderm has been reported to express SOX17, antibody co-labeling with SOX17 together with the parietal markers SPARC and/or Thrombomodulin (TM) can be used to identify the $SOX17^{+}$ cells that are parietal endoderm. As shown in Figures 6A-C, Thrombomodulin and SOX17 co-labeled parietal endoderm cells were produced by random differentiation of hES cells.

[0199] In view of the above cell labeling experiments, the identity of a definitive endoderm cell can be established by the marker profile $SOX17^{hi}/AFP^{lo}/[TM^{lo}$ or $SPARC^{lo}]$. In other words, the expression of the SOX17 marker is greater than the expression of the AFP marker, which is characteristic of visceral endoderm, and the TM or SPARC markers, which are characteristic of parietal endoderm. Accordingly, those cells positive for SOX17 but negative for AFP and negative for TM or SPARC are definitive endoderm.

[0200] As a further evidence of the specificity of the $SOX17^{hi}/AFP^{lo}/TM^{lo}/SPARC^{lo}$ marker profile as predictive of definitive endoderm, SOX17 and AFP gene expression was quantitatively compared to the relative number of antibody labeled cells. As shown in Figure 7A, hESCs treated with retinoic acid (visceral endoderm inducer), or activin A (definitive endoderm inducer), resulted in a 10-fold difference in the level of SOX17 mRNA expression. This result mirrored the 10-fold difference in SOX17 antibody-labeled cell number (Figure 7B). Furthermore, as shown in Figure 8A, activin A treatment of hESCs suppressed AFP gene expression by 6.8-fold in comparison to no treatment. This was visually reflected by a dramatic decrease in the number of AFP labeled cells in these cultures as shown in Figures 8B-C. To quantify this further, it was demonstrated that this approximately 7-fold decrease in AFP gene expression was the result of a similar 7-fold decrease in AFP antibody-labeled cell number as measured by flow cytometry (Figures 9A-B). This result is extremely significant in that it indicates that quantitative changes in gene expression as seen by Q-PCR mirror changes in cell type specification as observed by antibody staining.

[0201] Incubation of hESCs in the presence of Nodal family members (Nodal, activin A and activin B - NAA) resulted in a significant increase in SOX17 antibody-labeled cells over time. By 5 days of continuous activin treatment greater than 50% of the cells were labeled with SOX17 (Figures 10A-F). There were few or no cells labeled with AFP after 5 days of activin treatment.

[0202] In summary, the antibody produced against the carboxy-terminal 242 amino acids of the human SOX17 protein identified human SOX17 protein on Western blots but did not recognize SOX7, it's closest Sox family relative. The SOX17 antibody recognized a subset of cells in differentiating hESC cultures that were primarily $SOX17^{+}/AFP^{lo/-}$ (greater than 95% of labeled cells) as well as a small percentage (< 5%) of cells that co-label for SOX17 and AFP (visceral endoderm). Treatment of hESC cultures with activins resulted in a marked elevation of SOX17 gene expression as well as SOX17 labeled cells and dramatically suppressed the expression of AFP mRNA and the number of cells labeled with AFP antibody.

EXAMPLE 5

Q-PCR Gene Expression Assay

[0203] In the following experiments, real-time quantitative RT-PCR (Q-PCR) was the primary assay used for screening the effects of various treatments on hESC differentiation. In particular, real-time measurements of gene expression were analyzed for multiple marker genes at multiple time points by Q-PCR. Marker genes characteristic of the desired as well as undesired cell types were evaluated to gain a better understanding of the overall dynamics of the cellular populations. The strength of Q-PCR analysis includes its extreme sensitivity and relative ease of developing the necessary markers, as the genome sequence is readily available. Furthermore, the extremely high sensitivity of Q-PCR permits detection of gene expression from a relatively small number of cells within a much larger population. In addition, the ability to detect very low levels of gene expression provides indications for "differentiation bias" within the population. The bias towards a particular differentiation pathway, prior to the overt differentiation of those cellular phenotypes, is unrecognizable using immunocytochemical techniques. For this reason, Q-PCR provides a method of analysis that is at least complementary and potentially much superior to immunocytochemical techniques for screening the success of differentiation treatments. Additionally, Q-PCR provides a mechanism by which to

evaluate the success of a differentiation protocol in a quantitative format at semi-high throughput scales of analysis.

[0204] The approach taken here was to perform relative quantitation using SYBR Green chemistry on a Rotor Gene 3000 instrument (Corbett Research) and a two-step RT-PCR format. Such an approach allowed for the banking of cDNA samples for analysis of additional marker genes in the future, thus avoiding variability in the reverse transcription efficiency between samples.

[0205] Primers were designed to lie over exon-exon boundaries or span introns of at least 800 bp when possible, as this has been empirically determined to eliminate amplification from contaminating genomic DNA. When marker genes were employed that do not contain introns or they possess pseudogenes, DNase I treatment of RNA samples was performed.

[0206] We routinely used Q-PCR to measure the gene expression of multiple markers of target and non-target cell types in order to provide a broad profile description of gene expression in cell samples. The markers relevant for the early phases of hESC differentiation (specifically ectoderm, mesoderm, definitive endoderm and extra-embryonic endoderm) and for which validated primer sets are available are provided below in Table 1. The human specificity of these primer sets has also been demonstrated. This is an important fact since the hESCs were often grown on mouse feeder layers. Most typically, triplicate samples were taken for each condition and independently analyzed in duplicate to assess the biological variability associated with each quantitative determination.

[0207] To generate PCR template, total RNA was isolated using RNeasy (Qiagen) and quantitated using RiboGreen (Molecular Probes). Reverse transcription from 350-500 ng of total RNA was carried out using the iScript reverse transcriptase kit (BioRad), which contains a mix of oligo-dT and random primers. Each 20 μL reaction was subsequently diluted up to 100 μL total volume and 3 μL was used in each 10 μL Q-PCR reaction containing 400 nM forward and reverse primers and 5 μL 2X SYBR Green master mix (Qiagen). Two step cycling parameters were used employing a 5 second denature at 85-94°C (specifically selected according to the melting temp of the amplicon for each primer set) followed by a 45 second anneal/extend at 60°C. Fluorescence data was collected during the last 15 seconds of each extension phase. A three point, 10-fold dilution series was used to generate the standard curve for each run and cycle thresholds

(Ct's) were converted to quantitative values based on this standard curve. The quantitated values for each sample were normalized to housekeeping gene performance and then average and standard deviations were calculated for triplicate samples. At the conclusion of PCR cycling, a melt curve analysis was performed to ascertain the specificity of the reaction. A single specific product was indicated by a single peak at the T_m appropriate for that PCR amplicon. In addition, reactions performed without reverse transcriptase served as the negative control and do not amplify.

[0208] A first step in establishing the Q-PCR methodology was validation of appropriate housekeeping genes (HGs) in the experimental system. Since the HG was used to normalize across samples for the RNA input, RNA integrity and RT efficiency, it was of value that the HG exhibited a constant level of expression over time in all sample types in order for the normalization to be meaningful. We measured the expression levels of *Cyclophilin G, hypoxanthine phosphoribosyltransferase 1 (HPRT), beta-2-microglobulin, hydroxymethylbiane synthase (HMBS), TATA-binding protein (TBP), and glucoronidase beta (GUS)* in differentiating hESCs. Our results indicated that *beta-2-microglobulin* expression levels increased over the course of differentiation and therefore we excluded the use of this gene for normalization. The other genes exhibited consistent expression levels over time as well as across treatments. We routinely used both Cyclophilin G and GUS to calculate a normalization factor for all samples. The use of multiple HGs simultaneously reduces the variability inherent to the normalization process and increases the reliability of the relative gene expression values.

[0209] After obtaining genes for use in normalization, Q-PCR was then utilized to determine the relative gene expression levels of many marker genes across samples receiving different experimental treatments. The marker genes employed have been chosen because they exhibit enrichment in specific populations representative of the early germ layers and in particular have focused on sets of genes that are differentially expressed in definitive endoderm and extra-embryonic endoderm. These genes as well as their relative enrichment profiles are highlighted in Table 1.

TABLE 1

Germ Layer	Gene	Expression Domains
Endoderm	SOX17	definitive, visceral and parietal endoderm
	MIXL1	endoderm and mesoderm
	GATA4	definitive and primitive endoderm
	HNF3b	definitive endoderm and primitive endoderm, mesoderm, neural plate
	GSC	endoderm and mesoderm
Extra-embryonic	SOX7	visceral endoderm
	AFP	visceral endoderm. liver
	SPARC	parietal endoderm
	TM	parietal endoderm/trophectoderm
Ectoderm	ZIC1	neural tube, neural progenitors
Mesoderm	BRACH	nascent mesoderm

[0210] Since many genes are expressed in more than one germ layer it is useful to quantitatively compare expression levels of many genes within the same experiment. SOX17 is expressed in definitive endoderm and to a smaller extent in visceral and parietal endoderm. SOX7 and AFP are expressed in visceral endoderm at this early developmental time point. SPARC and TM are expressed in parietal endoderm and Brachyury is expressed in early mesoderm.

[0211] Definitive endoderm cells were predicted to express high levels of SOX17 mRNA and low levels of AFP and SOX7 (visceral endoderm), SPARC (parietal endoderm) and Brachyury (mesoderm). In addition, ZIC1 was used here to further rule out induction of early ectoderm. Finally, GATA4 and HNF3b were expressed in both definitive and extra-embryonic endoderm, and thus, correlate with SOX17 expression in definitive endoderm (Table 1). A representative experiment is shown in Figures 11-14 which demonstrates how the marker genes described in Table 1 correlate with each other among the various samples, thus highlighting specific patterns of differentiation to definitive endoderm and extra-embryonic endoderm as well as to mesodermal and neural cell types.

[0212] In view of the above data it is clear that increasing doses of activin resulted in increasing SOX17 gene expression. Further this SOX17 expression predominantly represented definitive endoderm as opposed to extra-embryonic endoderm. This conclusion stems from the observation that SOX17 gene expression was inversely correlated with AFP, SOX7, and SPARC gene expression.

EXAMPLE 6

Directed Differentiation of Human ES Cells to Definitive Endoderm

[0213] Human ES cell cultures randomly differentiate if cultured under conditions that do not actively maintain their undifferentiated state. This heterogeneous differentiation results in production of extra-embryonic endoderm cells comprised of both parietal and visceral endoderm (AFP, SPARC and SOX7 expression) as well as early ectodermal and mesodermal derivatives as marked by ZIC1 and Nestin (ectoderm) and Brachyury (mesoderm) expression. Definitive endoderm cell appearance has not been examined or specified for lack of specific antibody markers in ES cell cultures. As such, and by default, early definitive endoderm production in ES cell cultures has not been well studied. Since satisfactory antibody reagents for definitive endoderm cells have been unavailable, most of the characterization has focused on ectoderm and extra-embryonic endoderm. Overall, there are significantly greater numbers of extra-embryonic and neurectodermal cell types in comparison to $SOX17^{hi}$ definitive endoderm cells in randomly differentiated ES cell cultures.

[0214] As undifferentiated hESC colonies expand on a bed of fibroblast feeders, the cells at the edges of the colony take on alternative morphologies that are distinct from those cells residing within the interior of the colony. Many of these outer edge cells can be distinguished by their less uniform, larger cell body morphology and by the expression of higher levels of OCT4. It has been described that as ES cells begin to differentiate they alter the levels of OCT4 expression up or down relative to undifferentiated ES cells. Alteration of OCT4 levels above or below the undifferentiated threshold may signify the initial stages of differentiation away from the pluripotent state.

[0215] When undifferentiated colonies were examined by SOX17 immunocytochemistry, occasionally small 10-15-cell clusters of SOX17-positive cells were detected at random locations on the periphery and at the junctions between undifferentiated hESC colonies. As noted above, these scattered pockets of outer colony edges appeared to be some of the first cells to differentiate away from the classical ES cell morphology as the colony expanded in size and became more crowded. Younger, smaller fully undifferentiated colonies (< 1mm; 4-5 days old) showed no SOX17 positive cells within or at the edges of the colonies while older, larger colonies (1-2 mm diameter, > 5days old) had sporadic isolated patches of SOX17 positive, AFP negative cells at the periphery of some colonies or in regions interior to the edge that did not display the

classical hESC morphology described previously. Given that this was the first development of an effective SOX17 antibody, definitive endoderm cells generated in such early "undifferentiated" ES cell cultures have never been previously demonstrated.

[0216] Based on negative correlations of SOX17 and SPARC gene expression levels by Q-PCR, the vast majority of these SOX17 positive, AFP negative cells will be negative for parietal endoderm markers by antibody co-labeling. This was specifically demonstrated for TM-expressing parietal endoderm cells as shown in Figures 15A-B. Exposure to Nodal factors activin A and B resulted in a dramatic decrease in the intensity of TM expression and the number of TM positive cells. By triple labeling using SOX17, AFP and TM antibodies on an activin treated culture, clusters of SOX17 positive cells that were also negative for AFP and TM were observed (Figures 16A-D). These are the first cellular demonstrations of SOX17 positive definitive endoderm cells in differentiating hESC cultures (Figures 16A-D and 17).

[0217] With the SOX17 antibody and Q-PCR tools described above we have explored a number of procedures capable of efficiently programming hESCs to become $SOX17^{hi}/AFP^{lo}$ / $SPARC/TM^{lo}$ definitive endoderm cells. We applied a variety of differentiation protocols aimed at increasing the number and proliferative capacity of these cells as measured at the population level by Q-PCR for SOX17 gene expression and at the level of individual cells by antibody labeling of SOX17 protein.

[0218] We were the first to analyze and describe the effect of TGFβ family growth factors, such as Nodal/activin/BMP, for use in creating definitive endoderm cells from embryonic stem cells in *in vitro* cell cultures. In typical experiments, activin A, activin B, BMP or combinations of these growth factors were added to cultures of undifferentiated human stem cell line hESCyt-25 to begin the differentiation process.

[0219] As shown in Figure 19, addition of activin A at 100 ng/ml resulted in a 19-fold induction of SOX17 gene expression vs. undifferentiated hESCs by day 4 of differentiation. Adding activin B, a second member of the activin family, together with activin A, resulted in a 37-fold induction over undifferentiated hESCs by day 4 of combined activin treatment. Finally, adding a third member of the TGFβ family from the Nodal/Activin and BMP subgroups, BMP4, together with activin A and activin B, increased the fold induction to 57 times that of undifferentiated hESCs (Figure 19). When SOX17 induction with activins and BMP was compared to no factor medium controls 5-, 10-, and 15-fold inductions resulted at the 4-day time point. By five days of

triple treatment with activins A, B and BMP, SOX17 was induced more than 70 times higher than hESCs. These data indicate that higher doses and longer treatment times of the Nodal/activin TGFβ family members results in increased expression of SOX17.

[0220] Nodal and related molecules activin A, B and BMP facilitate the expression of SOX17 and definitive endoderm formation *in vivo* or *in vitro*. Furthermore, addition of BMP results in an improved SOX17 induction possibly through the further induction of Cripto, the Nodal co-receptor.

[0221] We have demonstrated that the combination of activins A and B together with BMP4 result in additive increases in SOX17 induction and hence definitive endoderm formation. BMP4 addition for prolonged periods (>4 days), in combination with activin A and B may induce SOX17 in parietal and visceral endoderm as well as definitive endoderm. In some embodiments of the present invention, it is therefore valuable to remove BMP4 from the treatment within 4 days of addition.

[0222] To determine the effect of TGFβ factor treatment at the individual cell level, a time course of TGFβ factor addition was examined using SOX17 antibody labeling. As previously shown in Figures 10A-F, there was a dramatic increase in the relative number of SOX17 labeled cells over time. The relative quantification (Figure 20) shows more than a 20-fold increase in SOX17-labeled cells. This result indicates that both the numbers of cells as well SOX17 gene expression level are increasing with time of TGFβ factor exposure. As shown in Figure 21, after four days of exposure to Nodal, activin A, activin B and BMP4, the level of SOX17 induction reached 168-fold over undifferentiated hESCs. Figure 22 shows that the relative number of SOX17-positive cells was also dose responsive. activin A doses of 100 ng/ml or more were capable of potently inducing SOX17 gene expression and cell number.

[0223] In addition to the TGFβ family members, the Wnt family of molecules may play a role in specification and/or maintenance of definitive endoderm. The use of Wnt molecules was also beneficial for the differentiation of hESCs to definitive endoderm as indicated by the increased SOX17 gene expression in samples that were treated with activins plus Wnt3a over that of activins alone (Figure 23).

[0224] All of the experiments described above were performed using a tissue culture medium containing 10% serum with added factors. Surprisingly, we discovered that the concentration of serum had an effect on the level of SOX17 expression in the

presence of added activins as shown in Figures 24A-C. When serum levels were reduced from 10% to 2%, SOX17 expression tripled in the presence of activins A and B.

[0225] Finally, we demonstrated that activin induced $SOX17^+$ cells divide in culture as depicted in Figures 25A-D. The arrows show cells labeled with SOX17/PCNA/DAPI that are in mitosis as evidenced by the PCNA/DAPI-labeled mitotic plate pattern and the phase contrast mitotic profile.

EXAMPLE 7

Chemokine receptor 4 (CXCR4) expression correlates with markers for definitive endoderm and not markers for mesoderm, ectoderm or visceral endoderm

[0226] As described above, hESCs can be induced to differentiate to the definitive endoderm germ layer by the application of cytokines of the TGFβ family and more specifically of the activin/nodal subfamily. Additionally, we have shown that the proportion of fetal bovine serum (FBS) in the differentiation culture medium effects the efficiency of definitive endoderm differentiation from hESCs. This effect is such that at a given concentration of activin A in the medium, higher levels of FBS will inhibit maximal differentiation to definitive endoderm. In the absence of exogenous activin A, differentiation of hESCs to the definitive endoderm lineage is very inefficient and the FBS concentration has much milder effects on the differentiation process of hESCs.

[0227] In these experiments, hESCs were differentiated by growing in RPMI medium (Invitrogen, Carlsbad, CA; cat# 61870-036) supplemented with 0.5%, 2.0% or 10% FBS and either with or without 100 ng/ml activin A for 6 days. In addition, a gradient of FBS ranging from 0.5% to 2.0% over the first three days of differentiation was also used in conjunction with 100 ng/ml of activin A. After the 6 days, replicate samples were collected from each culture condition and analyzed for relative gene expression by real-time quantitative PCR. The remaining cells were fixed for immunofluorescent detection of SOX17 protein.

[0228] The expression levels of CXCR4 varied dramatically across the 7 culture conditions used (Figure 26). In general, CXCR4 expression was high in activin A treated cultures (A100) and low in those which did not receive exogenous activin A (NF). In addition, among the A100 treated cultures, CXCR4 expression was highest when FBS concentration was lowest. There was a remarkable decrease in CXCR4 level in the 10%

FBS condition such that the relative expression was more in line with the conditions that did not receive activin A (NF).

[0229] As described above, expression of the SOX17, GSC, MIXL1, and HNF3β genes is consistent with the characterization of a cell as definitive endoderm. The relative expression of these four genes across the 7 differentiation conditions mirrors that of CXCR4 (Figures 27A-D). This demonstrates that CXCR4 is also a marker of definitive endoderm.

[0230] Ectoderm and mesoderm lineages can be distinguished from definitive endoderm by their expression of various markers. Early mesoderm expresses the genes Brachyury and MOX1 while nascent neuro-ectoderm expresses SOX1 and ZIC1. Figures 28A-D demonstrate that the cultures which did not receive exogenous activin A were preferentially enriched for mesoderm and ectoderm gene expression and that among the activin A treated cultures, the 10% FBS condition also had increased levels of mesoderm and ectoderm marker expression. These patterns of expression were inverse to that of CXCR4 and indicated that CXCR4 was not highly expressed in mesoderm or ectoderm derived from hESCs at this developmental time period.

[0231] Early during mammalian development, differentiation to extra-embryonic lineages also occurs. Of particular relevance here is the differentiation of visceral endoderm that shares the expression of many genes in common with definitive endoderm, including SOX17. To distinguish definitive endoderm from extra-embryonic visceral endoderm one should examine a marker that is distinct between these two. SOX7 represents a marker that is expressed in the visceral endoderm but not in the definitive endoderm lineage. Thus, culture conditions that exhibit robust SOX17 gene expression in the absence of SOX7 expression are likely to contain definitive and not visceral endoderm. It is shown in Figure 28E that SOX7 was highly expressed in cultures that did not receive activin A, SOX7 also exhibited increased expression even in the presence of activin A when FBS was included at 10%. This pattern is the inverse of the CXCR4 expression pattern and suggests that CXCR4 is not highly expressed in visceral endoderm.

[0232] The relative number of SOX17 immunoreactive ($SOX17^+$) cells present in each of the differentiation conditions mentioned above was also determined. When hESCs were differentiated in the presence of high dose activin A and low FBS concentration (0.5% - 2.0%) $SOX17^+$ cells were ubiquitously distributed throughout the

culture. When high dose activin A was used but FBS was included at 10% (v/v), the $SOX17^+$ cells appeared at much lower frequency and always appeared in isolated clusters rather than evenly distributed throughout the culture (Figures 29A and C as well as B and E). A further decrease in $SOX17^+$ cells was seen when no exogenous activin A was used. Under these conditions the $SOX17^+$ cells also appeared in clusters and these clusters were smaller and much more rare than those found in the high activin A, low FBS treatment (Figure 29 C and F). These results demonstrate that the CXCR4 expression patterns not only correspond to definitive endoderm gene expression but also to the number of definitive endoderm cells in each condition.

EXAMPLE 8

Differentiation conditions that enrich for definitive endoderm increase the proportion of CXCR4 positive cells

[0233] The dose of activin A also effects the efficiency at which definitive endoderm can be derived from hESCs. This example demonstrates that increasing the dose of activin A increases the proportion of $CXCR4^+$ cells in the culture.

[0234] hESCs were differentiated in RPMI media supplemented with 0.5%-2% FBS (increased from 0.5% to 1.0% to 2.0% over the first 3 days of differentiation) and either 0, 10, or 100 ng/ml of activin A. After 7 days of differentiation the cells were dissociated in PBS without Ca^{2+}/Mg^{2+} containing 2% FBS and 2 mM (EDTA) for 5 minutes at room temperature. The cells were filtered through 35 μm nylon filters, counted and pelleted. Pellets were resuspended in a small volume of 50% human serum/50% normal donkey serum and incubated for 2 minutes on ice to block non-specific antibody binding sites. To this, 1 μl of mouse anti-CXCR4 antibody (Abcam, cat# ab10403-100) was added per 50 μl (containing approximately 10^5 cells) and labeling proceeded for 45 minutes on ice. Cells were washed by adding 5 ml of PBS containing 2% human serum (buffer) and pelleted. A second wash with 5 ml of buffer was completed then cells were resuspended in 50 μl buffer per 10^5 cells. Secondary antibody (FITC conjugated donkey anti-mouse; Jackson ImmunoResearch, cat# 715-096-151) was added at 5 μg/ml final concentration and allowed to label for 30 minutes followed by two washes in buffer as above. Cells were resuspended at $5x10^6$ cells/ml in buffer and analyzed and sorted using a FACS Vantage (Beckton Dickenson) by the staff at the flow cytometry core facility (The Scripps Research Institute). Cells were collected directly

into RLT lysis buffer (Qiagen) for subsequent isolation of total RNA for gene expression analysis by real-time quantitative PCR.

[0235] The number of $CXCR4^+$ cells as determined by flow cytometry were observed to increase dramatically as the dose of activin A was increased in the differentiation culture media (Figures 30A-C). The $CXCR4^+$ cells were those falling within the R4 gate and this gate was set using a secondary antibody-only control for which 0.2% of events were located in the R4 gate. The dramatically increased numbers of $CXCR4^+$ cells correlates with a robust increase in definitive endoderm gene expression as activin A dose is increased (Figures 31A-D).

EXAMPLE 9

Isolation of CXCR4 positive cells enriches for definitive endoderm gene expression and depletes cells expressing markers of mesoderm, ectoderm and visceral endoderm

[0236] The $CXCR4^+$ and $CXCR4^-$ cells identified in Example 8 above were collected and analyzed for relative gene expression and the gene expression of the parent populations was determined simultaneously.

[0237] The relative levels of CXCR4 gene expression was dramatically increased with increasing dose of activin A (Figure 32). This correlated very well with the activin A dose-dependent increase of $CXCR4^+$ cells (Figures 30A-C). It is also clear that isolation of the $CXCR4^+$ cells from each population accounted for nearly all of the CXCR4 gene expression in that population. This demonstrates the efficiency of the FACS method for collecting these cells.

[0238] Gene expression analysis revealed that the $CXCR4^+$ cells contain not only the majority of the CXCR4 gene expression, but they also contained gene expression for other markers of definitive endoderm. As shown in Figures 31A-D, the $CXCR4^+$ cells were further enriched over the parent A100 population for SOX17, GSC, HNF3B, and MIXL1. In addition, the $CXCR4^-$ fraction contained very little gene expression for these definitive endoderm markers. Moreover, the $CXCR4^+$ and $CXCR4^-$ populations displayed the inverse pattern of gene expression for markers of mesoderm, ectoderm and extra-embryonic endoderm. Figures 33A-D shows that the $CXCR4^+$ cells were depleted for gene expression of Brachyury, MOX1, ZIC1, and SOX7 relative to the A100 parent population. This A100 parent population was already low in expression of these markers relative to the low dose or no activin A conditions. These results show that the isolation

of $CXCR4^+$ cells from hESCs differentiated in the presence of high activin A yields a population that is highly enriched for and substantially pure definitive endoderm.

EXAMPLE 10

Quantitation of Definitive Endoderm Cells in a Cell Population Using CXCR4

[0239] To confirm the quantitation of the proportion of definitive endoderm cells present in a cell culture or cell population as determined previously herein and as determined in United States Provisional Patent Application No. 60/532,004, entitled DEFINITIVE ENDODERM, filed December 23, 2003, the disclosure of which is incorporated herein by reference in its entirety, cells expressing CXCR4 and other markers of definitive endoderm were analyzed by FACS.

[0240] Using the methods such as those described in the above Examples, hESCs were differentiated to produce definitive endoderm. In particular, to increase the yield and purity in differentiating cell cultures, the serum concentration of the medium was controlled as follows: 0.2% FBS on day1, 1.0% FBS on day 2 and 2.0% FBS on days 3-6. Differentiated cultures were sorted by FACS using three cell surface epitopes, E-Cadherin, CXCR4, and Thrombomodulin. Sorted cell populations were then analyzed by Q-PCR to determine relative expression levels of markers for definitive and extraembryonic-endoderm as well as other cell types. CXCR4 sorted cells taken from optimally differentiated cultures resulted in the isolation of definitive endoderm cells that were >98% pure.

[0241] Table 2 shows the results of a marker analysis for a definitive endoderm culture that was differentiated from hESCs using the methods described herein.

Table 2

Composition of Definitive Endoderm Cultures

Marker(s)	Percent of culture	Percent Definitive Endoderm	Percent Extraembryonic endoderm	Percent hES cells
SOX17	70-80	100		
Thrombomodulin	<2	0	75	
AFP	<1	0	25	
CXCR4	70-80	100	0	
ECAD	10	0		100
other (ECAD neg.)	10-20			
Total	100	100	100	100

[0242] In particular, Table 2 indicates that CXCR4 and SOX17 positive cells (endoderm) comprised from 70%-80% of the cells in the cell culture. Of these SOX17-expressing cells, less than 2% expressed TM (parietal endoderm) and less than 1% expressed AFP (visceral endoderm). After subtracting the proportion of TM-positive and AFP-positive cells (combined parietal and visceral endoderm; 3% total) from the proportion of SOX17/CXCR4 positive cells, it can be seen that about 67% to about 77% of the cell culture was definitive endoderm. Approximately 10% of the cells were positive for E-Cadherin (ECAD), which is a marker for hESCs, and about 10-20% of the cells were of other cell types.

[0243] We have discovered that the purity of definitive endoderm in the differentiating cell cultures that are obtained prior to FACS separation can be improved as compared to the above-described low serum procedure by maintaining the FBS concentration at $\leq$0.5% throughout the 5-6 day differentiation procedure. However, maintaining the cell culture at $\leq$0.5% throughout the 5-6 day differentiation procedure also results in a reduced number of total definitive endoderm cells that are produced.

[0244] Definitive endoderm cells produced by methods described herein have been maintained and expanded in culture in the presence of activin for greater than 50 days without appreciable differentiation. In such cases, SOX17, CXCR4, MIXL1, GATA4, HNF3β expression is maintained over the culture period. Additionally, TM, SPARC, OCT4, AFP, SOX7, ZIC1 and BRACH were not detected in these cultures. It is likely that such cells can be maintained and expanded in culture for substantially longer than 50 days without appreciable differentiation.

EXAMPLE 11

Additional Marker of Definitive Endoderm Cells

[0245] In the following experiment, RNA was isolated from purified definitive endoderm and human embryonic stem cell populations. Gene expression was then analyzed by gene chip analysis of the RNA from each purified population. Q-PCR was also performed to further investigate the potential of genes expressed in definitive endoderm, but not in embryonic stem cells, as a marker for definitive endoderm.

[0246] Human embryonic stem cells (hESCs) were maintained in DMEM/F12 media supplemented with 20% KnockOut Serum Replacement, 4 ng/ml recombinant human basic fibroblast growth factor (bFGF), 0.1 mM 2-mercaptoethanol, L-glutamine,

non-essential amino acids and penicillin/streptomycin. hESCs were differentiated to definitive endoderm by culturing for 5 days in RPMI media supplemented with 100 ng/ml of recombinant human activin A, fetal bovine serum (FBS), and penicillin/streptomycin. The concentration of FBS was varied each day as follows: 0.1% (first day), 0.2% (second day), 2% (days 3-5).

[0247] Cells were isolated by fluorescence activated cell sorting (FACS) in order to obtain purified populations of hESCs and definitive endoderm for gene expression analysis. Immuno-purification was achieved for hESCs using SSEA4 antigen (R&D Systems, cat# FAB1435P) and for definitive endoderm using CXCR4 (R&D Systems, cat# FAB170P). Cells were dissociated using trypsin/EDTA (Invitrogen, cat# 25300-054), washed in phosphate buffered saline (PBS) containing 2% human serum and resuspended in 100% human serum on ice for 10 minutes to block non-specific binding. Staining was carried out for 30 minutes on ice by adding 200 µl of phycoerythrin-conjugated antibody to 5 x 10^6 cells in 800 µl human serum. Cells were washed twice with 8 ml of PBS buffer and resuspended in 1 ml of the same. FACS isolation was carried out by the core facility of The Scripps Research Institute using a FACS Vantage (BD Biosciences). Cells were collected directly into RLT lysis buffer and RNA was isolated by RNeasy according to the manufacturers instructions (Qiagen).

[0248] Purified RNA was submitted in duplicate to Expression Analysis (Durham, NC) for generation of the expression profile data using the Affymetrix platform and U133 Plus 2.0 high-density oligonucleotide arrays. Data presented is a group comparison that identifies genes differentially expressed between the two populations, hESCs and definitive endoderm. Genes that exhibited a robust upward change in expression level over that found in hESCs were selected as new candidate markers that are highly characteristic of definitive endoderm. Select genes were assayed by Q-PCR, as described above, to verify the gene expression changes found on the gene chip and also to investigate the expression pattern of these genes during a time course of hESC differentiation.

[0249] Figures 34A-M show the gene expression results for certain markers. Results are displayed for cell cultures analyzed 1, 3 and 5 days after the addition of 100 ng/ml activin A, CXCR4-expressing definitive endoderm cells purified at the end of the five day differentiation procedure (CXDE), and in purified hESCs. A comparison of Figures 34C and G-M demonstrates that the six marker genes, FGF17, VWF, CALCR, FOXQ1, CMKOR1 and CRIP1, exhibit an expression pattern that is almost identical to

each other and which is also identical to the pattern of expression of CXCR4 and the ratio of SOX17/SOX7. As described previously, SOX17 is expressed in both the definitive endoderm as well as in the SOX7-expressing extra-embryonic endoderm. Since SOX7 is not expressed in the definitive endoderm, the ratio of SOX17/SOX7 provides a reliable estimate of definitive endoderm contribution to the SOX17 expression witnessed in the population as a whole. The similarity of panels G-L and M to panel C indicates that FGF17, VWF, CALCR, FOXQ1, CMKOR1 and CRIP1 are likely markers of definitive endoderm and that they are not significantly expressed in extra-embryonic endoderm cells.

[0250] It will be appreciated that the Q-PCR results described herein can be further confirmed by ICC.

EXAMPLE 12

Generation of SOX17 Promoter-EGFP Transgenic hESC Lines and CXCR4 Promoter-EGFP Transgenic hESC Lines

[0251] As an alternative to purification of definitive endoderm using the CXCR4-specific antibody, EGFP fusions to either the SOX17 or the CXCR4 promoters can be used. In particular, this Example describes the construction of a vector comprising a reporter cassette which comprises a reporter gene under the control of the SOX17 regulatory region. Additionally, the construction of a vector comprising a reporter cassette which comprises a reporter gene under the control of the CXCR4 regulatory region is described. This Example also describes the preparation of a cell, such as a human embryonic stem cell, transfected with one or more of these vectors as well as a cell having this one or both of these reporter cassettes integrated into its genome.

[0252] SOX17-expressing definitive endoderm cell lines and CXRC4-expressing definitive endoderm cell lines genetically tagged with a reporter gene are constructed by placing a GFP reporter gene under the control of the regulatory region (promoter) of the SOX17 gene or the CXCR4 gene, respectively. First, a plasmid construct in which EGFP expression is driven by the human SOX17 or CXCR4 gene promoter is generated by replacing the CMV promoter of vector pEGFP-N1 (Clontech) with the human SOX17 or CXCR4 control region. These control regions contain the characterized regulatory elements of either the SOX17 or the CXCR4 gene, and they is sufficient to confer the normal expression pattern of these genes in transgenic mice. In

the resulting vector, expression of EFGP is driven by either the SOX17 promoter or the CXCR4 promoter. In some experiments, this vector can be transfected into hESCs.

[0253] The SOX17 promoter/EGFP cassette or the CXCR4 promoter/EGFP cassette is excised from the above vector, and then subcloned into a selection vector containing the neomycin phosphotransferase gene under control of the phosphoglycerate kinase-1 promoter. The selection cassette is flanked by flp recombinase recognition sites to allow removal of the cassette. This selection vector is linearized, and then introduced into hESCs using standard lipofection methods. Following 10-14 days of selection in G418, undifferentiated transgenic hESC clones is isolated and expanded.

[0254] It will be appreciated that reporter genes other than GFP or EGFP can be used in any of the above-described constructs provided that the reporter allows for cell separation by FACS

EXAMPLE 13

Alternative Isolation of Definitive Endoderm

[0255] The following Example demonstrates that hESCs comprising a SOX17 or CXCR4 promoter/EGFP cassette can be differentiated into definitive endoderm cells and then subsequently isolated by fluorescence-activated cell sorting (FACS).

[0256] SOX17 or CXCR4 promoter/EGFP transgenic hESCs are differentiated for approximately 6, 12 and 18 hours in growth medium containing 100 ng/ml activin A and no serum. The differentiated cells are then harvested by trypsin digestion and sorted on a Becton Dickinson FACS Diva directly into RNA lysis buffer or PBS. A sample of single live cells is taken without gating for EGFP and single live cells are gated into EGFP positive and GFP negative populations. In a separate experiment, the EGFP positive fraction is separated into two equally sized populations according to fluorescence intensity (Hi and Lo).

[0257] Following sorting, cell populations are analyzed by both Q-PCR and immunocytochemistry. For Q-PCR analysis, RNA is prepared using Qiagen RNeasy columns and then converted to cDNA. Q-PCR is conducted as described previously. For immunocytochemistry analysis, cells are sorted into PBS, fixed for 10 minutes in 4% paraformaldehyde, and adhered to glass slides using a Cytospin centrifuge. The primary antibody SOX17 or CXCR4. An appropriate secondary antibody conjugated to FITC (green) or Rhodamine (Red) is used to detect binding of the primary antibody.

[0258] Sorted cells are further subjected to Q-PCR analysis. Differentiated cells show a correlation of EGFP fluorescence with endogenous SOX17 or CXCR4 expression gene expression. Compared to non-fluorescing cells, the EGFP positive cells show a greater than 2-fold increase in SOX17 or CXCR4 expression levels. The separation of high and low EGFP intensity cells indicates that EGFP expression level correlates with SOX17 or CXCR4 expression level. In addition to SOX17 or CXCR4 mRNA analysis, sorted cells are subjected to immunocytochemistry analysis of SOX17 or CXCR4 polypeptide (in embodiments where CXCR4/EGFP fusions are used, SOX17 polypeptide expression is analyzed and in cases where SOX17/EGFP fusions are used, CXCR4 polypeptide expression is analyzed). Substantial expression of either the SOX17 or CXCR4 polypeptides can be seen in the enriched in the EGFP positive fraction. In contrast, little expression of either the SOX17 or CXCR4 polypeptides is seen in the EGFP negative fraction.

[0259] Given these results, at least about 5% of the cells present in the differentiated cell cultures prior to sorting are SOX17/CXCR4-positive definitive endoderm cells. At least about 90% of the cells in the sorted cell populations are SOX17/CXCR4-positive definitive endoderm cells.

EXAMPLE 14

Passage of Definitive Endoderm Cells in Culture

[0260] This Example demonstrates that the definitive endoderm cells described herein can be maintained in cell culture and passaged without further differentiation.

[0261] Definitive endoderm cells were differentiated from two related passages, designated EB and EV, of the CyT25 hESC line in the presence of 100 ng/ml activin A in low serum RMPI. The low serum RPMI contained 0%(v/v) fetal bovine serum (FBS) on day 1, 0.2% (v/v) FBS on day two and 2% serum on each day thereafter. After four days of differentiation, the cells maintained in culture in either the presence or absence of 100 ng/ml activin A for a total of 36 days as measured from induction of differentiation. During the 36 day culture period, the definitive endoderm cells were passaged twice. Furthermore, on days 29-36 the cells of the group designated EV were additionally contacted with 50 mg/ml EGF. On days 4, 9, 23, 29 and 36 of culture, Q-PCR was used to measure the expression of marker genes indicative of definitive endoderm .

[0262] Figures 35A-D show that, in cell cultures provided with 100 ng/ml activin A, expression of the definitive endoderm markers SOX17, GSC, MIXL1 and CXCR4 was maintained during the 32 day culture period subsequent to the derivation of the definitive endoderm cells from hESCs (days 4 to 36). Little expression of these markers was observed in the cell cultures grown in the absence of activin A. Addition of 50 ng/ml EGF did not appear to significantly increase the expression of any of the definitive endoderm markers.

EXAMPLE 15

Expansion of Purified Definitive Endoderm Cells

[0263] This Example demonstrates that the definitive endoderm cells described herein can be differentiated from hESCs, purified and then regrown and expanded in cell culture.

[0264] Figure 36 shows the design of a definitive endoderm purification/expansion experiment. In particular, definitive endoderm cells were differentiated from the 96th passage of hESC line CyT25 in the presence of 100 ng/ml activin A in low serum RMPI. The low serum RPMI contained 0%(v/v) fetal bovine serum (FBS) on day 1, 0.2% (v/v) FBS on day two and 2% serum on each day thereafter. After five days of differentiation, the cells were subjected to FACS purification using antibody against CXCR4 as described in previous Examples. The purified cell population was then cultured on IVF dishes coated with poly-ornithine and 10 µg/ml human fibronectin in RPMI containing 2% FBS under one of the following four growth factor conditions: no added factor (NF); 100 ng/ml activin A (A); 100 ng/ml activin A and 100 ng/ml IGF1 (AI); or 100 ng/ml activin A, 12 ng/ml bFGF and 10 ng/ml EGF (AFE). On day 11, the expanded definitive endoderm cells were passaged using the standard trypsinization method. Each of the cell cultures were then grown for an additional 10 days after passage (a total of 21 days subsequent to the first contact with activin A). Samples of mRNA were obtained at days 0, 5, 11 and 21 as indicated in Figure 36.

[0265] Figures 37A-E show the expression of marker genes for various embryonic cell types at each of the sample time points for each of the culture conditions. As shown in Figures 37A-B, the definitive endoderm markers SOX17 and GSC were highly expressed five day old unpurified definitive endoderm cultures but were not expressed in hESCs. This expression is in contrast to that of the hESC marker, OCT4 (Figure 37C). Six days after the purification of definitive endoderm cells (day 11) the

expression of SOX17 and GSC expression remained high in each of the cell cultures treated with growth factor(s) but not in cell cultures grown in the absence of activin A (Figure 37A-B). A similar pattern of expression was observed for these markers 10 days after passage (day 21) (Figures 37A-B). No expression of mRNA for markers of hESCs (OCT4), mesoderm (brachyury), or ectoderm (ZIC1 and SOX1) was observed in any of the cell cultures subsequent to purification. This result indicates that purified definitive endoderm cells do not form hESCs or cells of the other two embryonic cell lineages even in the absence of activin A (Figures 37C-F).

[0266] The methods, compositions, and devices described herein are presently representative of preferred embodiments and are exemplary and are not intended as limitations on the scope of the invention. Changes therein and other uses will occur to those skilled in the art which are encompassed within the spirit of the invention and are defined by the scope of the disclosure. Accordingly, it will be apparent to one skilled in the art that varying substitutions and modifications may be made to the invention disclosed herein without departing from the scope and spirit of the invention.

[0267] As used in the claims below and throughout this disclosure, by the phrase "consisting essentially of" is meant including any elements listed after the phrase, and limited to other elements that do not interfere with or contribute to the activity or action specified in the disclosure for the listed elements. Thus, the phrase "consisting essentially of" indicates that the listed elements are required or mandatory, but that other elements are optional and may or may not be present depending upon whether or not they affect the activity or action of the listed elements.

References

[0268] Numerous literature and patent references have been cited in the present patent application. Each and every reference that is cited in this patent application is incorporated by reference herein in its entirety.

[0269] For some references, the complete citation is in the body of the text. For other references the citation in the body of the text is by author and year, the complete citation being as follows:

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WHAT IS CLAIMED IS:

1. A method of expanding definitive endoderm cells in culture, said method comprising the steps of;

   (a) obtaining a cell culture comprising definitive endoderm cells;

   (b) isolating at least some of the definitive endoderm cells from at least some of the other cells in the cell culture, thereby producing a cell population enriched in definitive endoderm cells; and

   (c) culturing said cell population enriched in definitive endoderm cells under conditions that permit the expansion of said definitive endoderm cells.

2. The method of claim 1, wherein said definitive endoderm cells being multipotent cells that can differentiate into cells of the gut tube or organs derived therefrom.

3. The method of claim 1 or 2, wherein said definitive endoderm cells are human definitive endoderm cells.

4. The method of claim 3, wherein said definitive endoderm cells are derived from human embryonic stem cells (hESCs).

5. The method of claim 4, wherein the obtaining step comprises contacting hESCs with at least one growth factor from the TGFβ superfamily so as to permit differentiation of at least some of said hESCs to definitive endoderm cells.

6. The method of claim 5, wherein said at least one growth factor from the TGFβ superfamily comprises activin A.

7. The method of any one of claims 1 to 6, wherein the step of obtaining said cell culture comprising definitive endoderm further comprises obtaining a portion of an existing definitive endoderm culture.

8. The method of any one of claims 1 to 7, wherein said cell population enriched in definitive endoderm cells is substantially free of cells other than definitive endoderm cells.

9. The method of any one of claims 1 to 7, wherein said cell population enriched in definitive endoderm cells comprises at least about 96% definitive endoderm cells.

10. The method of any one of claims 1 to 9, wherein said isolating step comprises providing said cell culture with a reagent which binds to a marker expressed in said definitive endoderm cells but which is not substantially expressed in said other cells present in the cell culture, and separating said definitive endoderm cells bound to said reagent from said other cells present in the cell culture, thereby producing a cell population enriched in definitive endoderm cells.

11. The method of claim 10, wherein said marker is CXCR4.

12. The method of any one of claims 1 to 9, wherein said isolating step comprises separating fluorescently-labeled definitive endoderm cells from unlabeled cells.

13. The method of claim 12, wherein said fluorescently-labeled definitive endoderm cells are labeled as a result of the expression of enhanced green fluorescent protein (EGFP).

14. The method of claim 13, wherein the expression of EGFP is under control of the SOX17 promoter.

15. The method of claim 13, wherein the expression of EGFP is under control of the CXCR4 promoter.

16. The method of any one of claims 1 to 15, wherein said culturing step comprises plating said population enriched in definitive endoderm cells or a portion thereof.

17. The method of claim 16, wherein said population enriched in definitive endoderm cells or a portion thereof is plated on a surface coated with human fibronectin.

18. The method of any one of claims 1 to 15, wherein said culturing step comprises incubating said population enriched in definitive endoderm cells or a portion thereof in a medium comprising less than about 2% (v/v) serum.

19. The method of any one of claims 1 to 15, wherein said culturing step comprises incubating said population enriched in definitive endoderm cells or a portion thereof in a medium comprising at least one growth factor.

20. The method of claim 19, wherein said at least one growth factor is a growth factor from the TGFβ superfamily of growth factors.

21. The method of claim 20, wherein said at least one growth factor from the TGFβ superfamily of growth factors comprises activin A.

22. The method of claim 21, wherein said activin A is present in said medium at a concentration of about 100 ng/ml.

23. The method of claim 19, wherein said at least one growth factor is selected from the group consisting of IGF1, activin A, bFGF and EGF, or a combination thereof.

24. An expanded definitive endoderm cell population produced by the method of any one of claims 1 to 23.

25. A method of expanding definitive endoderm cells in culture, said method comprising the steps of: (a) obtaining a cell culture comprising definitive endoderm cells, and (b) passaging said definitive endoderm cells, thereby producing a plurality of cell cultures comprising definitive endoderm cells.

26. The method of claim 25, wherein the step of passaging said definitive endoderm cells comprises mechanically disrupting contacts between said definitive endoderm cells.

27. The method of claim 25, wherein the step of passaging said definitive endoderm cells comprises incubating said definitive endoderm cells in a cell dispersal buffer.

28. The method of any one of claims 25 to 27, wherein said definitive endoderm cells are attached to a substrate.

29. The method of claim 28, wherein the step of passaging said definitive endoderm cells comprises detaching said definitive endoderm cells from said substrate.

30. An expanded definitive endoderm cell population produced by the method of any one of claims 25 to 29.

EXPANSION OF DEFINITIVE ENDODERM CELLS

Abstract

Disclosed herein are cell cultures comprising expanded definitive endoderm cells as well as methods for expanding definitive endoderm cells in culture.

Figure 36.

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Baetge, Emmanuel, E.
Carpenter, Melissa

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