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METHOD FOR LYSING MYCOBACTERIA

FIELD OF THE INVENTION

The invention is in the field of molecular biology. In particular the invention is in the area of cell lysis. Most particularly the invention is a method for lysis of Mycobacteria.

BACKGROUND OF THE INVENTION

Mycobacteria are a large, diverse, and widely distributed family of aerobic, nonsporulating, nonmotile bacilli that have a high cell-wall lipid content and a slow growth rate. Members of the Mycobacterium genus vary tremendously in virulence. Some Mycobacteria are harmless while others like M. tuberculosis are significant pathogens. Mycobacterium species are differentiated by their growth rate, pigment production, animal virulence, and biochemical reactivity.

Many detection methods for determining the presence of pathogenic organisms such as those in the Mycobacteriaceae

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family rely on the lysis of those organisms. However, the lysis procedures for Mycobacteriaceae are laborious and time consuming. For example, chemical disruption of Mycobacteria is tedious and can take days. European Patent Application number 87 303641.2 discloses a method for releasing DNA or RNA from Mycobacteria which comprises the use of minute beads in combination with a sonicating bath. S. Hurley et al., J. Clin Microbiol. 25:2227 (1987) disclose a method for lysing mycobacteria that uses a mini-beadbeater cell disrupter and zirconium beads.

Recent advances in mycobacterial genetics and increased interest in opportunistic pathogens in patients like those suffering from acquired immunodeficiency syndrome have focused attention to the fact that a procedure for rapid lysis of Mycobacteriaceae is needed. It would be advantageous to have a process for lysing Mycobacteria that is simple, fast, and not disruptive to the material desired from the lysis.

SUMMARY OF THE INVENTION

The present invention provides a process for lysing Mycobacteria that is simple, fast, and not disruptive to the material desired from the lysis. In one embodiment is provided a process for lysing Mycobacteria which comprises exposing the bacteria to a lysis effective amount of the enzyme achromopeptidase.

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Further embodiments include isolating specific cellular components liberated from lysis of Mycobacteria using the method of the invention.

Specific embodiments also include the additional step of isolating nucleic acid from Mycobacteria and amplifying nucleic acid obtained from practicing the method of the present invention.

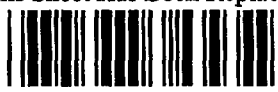
Other embodiments include the addition of a Mycobacteria identifying agent to the lysed Mycobacteria to identify the presence of Mycobacteria.

Embodiments also include kits comprising lysis effective amounts of achromopeptidase and a Mycobacteria identifying agent.

The method of the invention is particularly advantageous since only one step is involved, it is expedient compared to prior processes, and little instrumentation is necessary. By practicing the method of the invention it is possible to lyse Mycobacteria with minimal effort. In addition, practicing the invention results in liberating deoxyribonucleic acid (DNA) from Mycobacteria. Not only is DNA liberated, but the DNA is liberated in relative uniform size, rendering the DNA well suited for subsequent analysis by way of probe hybridization, restriction enzyme analysis, amplification, and the like.

As used in this document, "lysis effective amount of achromopeptidase" refers to that amount of achromopeptidase which liberates intracellular components such as DNA, RNA, and the like.

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DETAILED DESCRIPTION OF THE INVENTION

The present invention allows for the lysis and resultant liberation of DNA and cellular material from Mycobacteria. The enzyme employed in the process is achromopeptidase, also known as lysyl endopeptidase. The enzyme is readily available from a variety of sources, including commercial availability from vendors. The enzyme is also obtainable by isolation from the organism Achromobacter lyticus.

The exposure of Mycobacteria to the enzyme achromopeptidase is advantageous over known methods for lysis of Mycobacteria which involve the use of caustic chemicals, time consuming culturing, and mechanical methods which use the French press, the Hughes press, sonicating probes, bath sonicators, freeze-thawing, glass beads, the Ribi pressure cell, and the like.

Although numerous enzymes and procedures exist for lysing a variety of organisms, the application of the enzyme achromopeptidase to lyse Mycobacteria is unique. Mycobacteria are notorious for their inability to readily lyse. Those procedures that do result in lysis of Mycobacteria also generally destroy the contents of the cell that were desired. If the contents of the cell were not destroyed from the lysis procedure, it was generally the result of timely and laborious protocols. Mycobacteria are extremely resistant to physical

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stress and can be subjected to concentration and digestion procedures that kill ordinary bacteria. Thus, it is unexpected that the enzyme achromopeptidase, which can lyse less strenuous bacteria, can also lyse the extraordinarily lysis resistant Mycobacteria. It is also unexpected that achromopeptidase works so well in lysing Mycobacteria because other, more stringent conditions, do not work. However, the practice of the present invention results in Mycobacteria lysis and subsequent yield of useable pieces of DNA that are suitable for use for a variety of purposes such as detection methods and amplification, as well as liberating RNA and other cellular components.

Subsequent use of cellular components liberated from lysis include identification of Mycobacteria and amplification of nucleic acid by means such as polymerase chain reaction, ligase chain reaction, and the like. Identification can take place by means of Mycobacteria identifying agents. Identifying agents refers to those agents suitable for identifying Mycobacteria which include nucleic acid probes including deoxyribonucleic acid and ribonucleic acid, and the like.

The use of probes, for example, for identifying the presence of a particular Mycobacterium can be employed in a one step identification method. For example, once a sample is obtained, such as a sputum sample, the sputum is digested with a liquifying agent like N-Acetyl-L-Cysteine (NALC). After digestion and concentration a lysis effective amount of

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achromopeptidase is added to the sample, followed by the addition of an identifying agent. The presence of Mycobacteria can then be detected by a variety of means, depending on the marker (e.g., signal to be detected) chosen for use with the identifying agent. The means for identification of the presence of Mycobacteria is usually dictated by the identifying agent employed. For example, nucleic acid probes (e.g., specific for a Mycobacteria species) are typically labeled with ^{125}I, ^{32}P, fluorescent markers, and the like. The marker is then detected, which detection is an indication that the particular Mycobacteria is present. Other means for detection include Southern Blot analysis, electrophoretic gel visualization, and the like.

The Mycobacteria identifying agent and achromopeptidase can be conveniently provided in the form of a kit. Such a kit comprises at least one type of identifying agent and a lysis effective amount of achromopeptidase. Specific kits comprise identifying agents for any Mycobacteria or specific Mycobacterium. Specific kits also comprise particular Mycobacteria identifying means such as nucleic acid probes or antibodies. And, the means by which the identifying agent is detected can also be specific, for example, the agent can be designed for fluorescence, radioactive, and chemiluminescence detection and, if necessary, depending on sample requirements, liquification agents, isolation agents, and the like can be included in the kit.

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The process of the invention can be employed once the Mycobacteria have been obtained in the form of a sample such as sputum, or an isolated form. Mycobacteria are isolated from a variety of sources including feces, sputum, urine, serum, tissue, other body fluids or obtained from public or private culture collections, and the like. Mycobacteria obtained from the various sources are typically cultured, which is very time consuming, reaching three to six weeks culture time. However, by practicing the method of the invention, the need to culture can be eliminated. If culturing is not desired, the cells are generally first isolated from the source by conventional sample processing methods then usually pelleted by centrifugation and put into a cell suspension. The Mycobacteria in the cell suspension are then lysed. Lysis by the present invention comprises the addition of a lysis effective amount of achromopeptidase to the cells. The achromopeptidase does not have to be pure, and preferably is impure. This is advantageous in that purification steps are eliminated prior to use of the enzyme, and attention to contamination is eased. Preferably the enzyme is present in about 50 units to about 1000 units, most preferably the enzyme is present in about 100 units to about 300 units.

The process of the invention could be practiced without culturing. Unpurified biological samples from sputum, feces, tissue, blood, serum, and the like, can be lysed by practicing

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the invention and in the same sample could be identified with a Mycobacteria identifying agent. Thus the method comprises a simplified means for detecting Mycobacteria in a clinical, biological, food or environmental sample.

A typical protocol for lysing Mycobacteria with achromopeptidase comprises centrifugation of a sample of Mycobacteria for a brief amount of time (e.g., about five minutes) and discarding the resultant supernatant. The pellet of Mycobacteria can then be reconstituted in a buffered mixture of the achromopeptidase enzyme. Any suitable buffer will work. Suitable buffers include Trizma and NaCl, and Borate and NaCl. After a brief incubation (e.g., about thirty minutes) at about room temperature to about 50 degrees Centigrade lysis is complete and the liberated cellular contents can be conveniently isolated by conventional methods. Conventional methods for isolating DNA include phenol:chloroform extractions, glass binding with subsequent elution, and the like. Examples of conventional protocols for isolating DNA are found in references such as T. Maniatis et al., Molecular Cloning: A Laboratory Manual (Cold Spring Harbor Lab) (1982) and Boom et al., J. Clin. Micro 28:495 (1990).

The fact that the process of the present invention liberates DNA in utilizable size is significant. Although prior methods obtained lysis of Mycobacteria through laborious and time consuming methodology, the DNA liberated was generally not of useable size (it was degraded into pieces too small for

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subsequent use). It is important that liberated DNA be of sufficient size to allow for its subsequent use in detection procedures. Thus, despite the fact that a variety of lysis procedures obtain DNA, it is important to obtain DNA in useful quantity and size.

Important Mycobacteria that can be lysed by practicing the present invention include M. avium, M. intracellularae, M. gordonae, M. tuberculosis, M. kansasii, M. fortuitum, M.chelonae, M. bovis, M. scrofulaceum, M. paratuberculosis, M. marinum, M. simiae, M. szulgai, M. intracellulare, M. xenopi, M. ulcerans, M. leprae, M. lepraemurium, M. smegmatis, M. flavescens, M. terrae, M. nonchromogenicum, M. malmoense, M. asiaticum, M. vaccae, M. gastri, M. triviale, M. haemophilum, M. africanum, M. thermoresistable, and M. phlei. Several of the Mycobacteria are pathogenic. For example, M. tuberculosis, which already infects two billion people and infects an additional seven to nine million people each year, is an important Mycobacteria from an epidemiologic and clinical viewpoint In addition, M. averium, M. bovis, M. intracellularae, M. africanum, M. leprae, M. chelonae, M. paratuberculosis, and M. marinum, are also significant from an epidemiological and clinical viewpoint.

The practice of the present invention provides a rapid and simple lysis procedure for Mycobacteria that provides DNA, RNA and cellular components for subsequent use in a variety of detection procedures.

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The following examples illustrate the specific embodiments of the invention described in this document. As would be apparent to skilled artisans, various changes and modifications are possible and are contemplated within the scope of the invention described.

EXAMPLES

EXAMPLE 1

Purpose:

This experiment compares purified achromopeptidase enzyme to a crude product using equivalent amounts (by weight and by enzyme units) with Bactec cultures.

Materials:

- "Crude" achromopeptidase - (Sigma, St. Louis, MO, cat# A-7550 lot# 127F-68391)
- "Purified" achromopeptidase - (Sigma cat# A-3422 lot# 630H-0438)
- Pooled Bactec cultures of M. tuberculosis (strain H37RV American Type Culture Collection (ATCC)) and M. fortuitum
- Amresco (Solon, OH) TE (10mM TRIS-HCI, 1mM EDTA, pH 8.0) buffered phenol/chloroform (1:1)

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Procedure:

- Isolate 2ml pellets of M. tuberculosis and M. fortuitum (4 pellets for each species).
- Reconstitute pellets according to the following recipes:

"Crude" Achromopeptidase: 1 mg/ml 550 units/mg, 100 units = 182µL
"Purified" Achromopeptidase: 1 mg/ml 25,000 units/mg, 100 units = 4µL

#1 50ug Crude Achromopeptidase = 50µL + 450µL TRizma 9.0 (pre-pH adjusted mixes of TRIS-HCl + TRIS-hydroxide to give a set pH) (27 units)

#2 50ug Purified Achromopeptidase = 50µL + 450µL TRizma 9.0 (1250 units)

#3 100 units Crude Achromopeptidase = 182 µL + 318µL TRizma 9.0 (182 µg)

#4 100 units Purified Achromopeptidase = 4µL + 496µL TRizma 9.0 (4µg)

- Vortex all samples, then incubate at 50°C for 30 minutes.
- Perform 2 phenol/chloroform extractions, then ethanol. Precipitate and dry samples in speed-vac before reconstituting pellets in 10 µl TE buffer and running sample on 1% agarose gel.
- Samples were stained with 1ug/ml ethidium bromide and visualized under UV light.

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The results (band intensity) show that the crude enzyme works better than the partially purified enzyme. More DNA was released from M. tuberculosis and M. fortuitum with the crude enzyme.

EXAMPLE 2

Purpose:

This experiment examines 8 species of Mycobacteria with 100 units of achromopeptidase. The Mycobacteria are:

M. avium
M. scrofulaceum
M. intracellularae
M. gordonae
M. tuberculosis
M. kansasii
M. fortuitum
M. chelonae

Materials:

- Crude achromopeptidase
- 8 pooled Bactec cultures of Mycobacteria
- Amresco TE buffered phenol/chloroform

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Procedure:

- Isolate 2ml pellets of each species
- Reconstitute each pellet into 500µL TRizma 9.0
- Incubate for 30 minutes at 50°C with 100 units achromopeptidase (36µL or 5mg/ml stock = 2750 units/mg)
- Vortex while incubating (2 or 3 times)
- Perform 2 phenol/chloroform extractions on each
- Ethanol precipitate samples overnight at -20°C
- Spin 1/2 hour at 4°C, remove ethanol, dry pellets in virtis lyophilizer for 30 minutes. Resuspend in 17µL TE and 3µL loading dye
- Electrophorese in 1% agarose. Ethidium Bromide stain 10 minutes then visualize under UV light

Results indicate that DNA of same size is seen for all species (gel visualization).

EXAMPLE 3

Purpose:

To screen a series of enzymes for lysis efficiency of M. tuberculosis.

Materials:

- M. tuberculosis
- Lysozyme (Sigma cat# L-6876 lot# 39F-8210)
- Achromopeptidase (Sigma cat# A-3547 lot# 88F-0799)

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- Lipoxidase (Sigma cat# L-7395 lot# 118F-05421)
- Mixed Glycosidase A (From C. Lamdas, Sekagaku, Miles, Elkhart, Indiana, lot# 8L84803)
- Zymolase 20T (Sekagaku/Miles cat# 32-092-1 lot#2)
- Mixed Glycosidase B (From T. Cornelius, Sekagaku lot# ET84701)
- Lysing Enzymes A (From Tricoderma harzanium Sigma cat# L-2265 lot# 36F-080
- Lysing enzymes B (From Rhizoctonia Solani Sigma cat# L-8757 lot# 468-0273
- Phospholipase A (Sigma cat# D-6534 lot# 129F-8005)
- Lipase (Sigma cat# L-4384 lot# 88F-02081)
- Mutanolysin (Sigma cat# M-9901 lot# 98F-68211)
- Achromopeptidase/Mutanolysin cocktail (10mg/ml/1mg/ml)
- Gen-Probe (San Diego, CA) lysing reagent tube (lot# 92084 exp. 12/16/91)
- 1:1 Phenol/chloroform (TRIS saturated BRL 55090A lot# 71209/Baxter B&J cat# 67-66-3 lot# A W342
- Ethanol (Fisher (Pittsburgh, PA) E-575-500 lot# 887309) 70%
- 3M sodium acetate pH 5.5
- Lyophillizer (Savant speed vaccuum concentrator model
- TE buffer
- Other electrophoresis equipment

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Procedure:

1) Into 13 tubes, 2ml of pelleted M. tuberculosis was placed in each after being decanted.
2) To each portion was added 90µL H_2O (except the GEN probe tube, which received 100µL) and 10µL of the enzyme so that final concentration was 500mg/ml (50ug total).
3) All tubes were incubated at 37°C for 30 minutes (GEN probe run according to protocol).
4) 200µL of water was added to each tube to bring volume up to 300µL, then 2 Phenol/Chloroform extractions were performed (including GEN Probe).
5) 30µL of 3M sodium acetate and 600µL ethanol was added to the aqueous layer, and this mixture incubated at -20°C over the weekend.
6) Samples were spun at 4°C for thirty minutes, excess ethanol was removed, and pellets were dried in the Lyophillizer for 1 hour.
7) Samples were resuspended into 20µL TE + 3µL loading buffer and electrophoresed for 35 minutes at 150V (94VH)(1% agarose gel in 1x TAE buffer) (40mM TRIS-acetate/1mM EDTA).
8) Gel was ethidium bromide stained (1µg/1ml) and visualized under UV light.

Results show achromopeptidase released DNA of a distinct 23,000 base pair size. No other enzymes released much DNA.

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EXAMPLE 4

This experiment screens a second set of enzymes for the ability to lyse M. tuberculosis while preserving the DNA for possible detection procedures.

Materials

- M. tuberculosis
- Gen probe lysing tube

Enzymes

1. Achromopeptidase
2. O - Glycanase (Genzyme (Boston, MA) Code O-ASE Lot #39178)
3. Lyticase (from Arthrobacter lureus Sigma Cat #L-8137 Lot # 69F-6819)
4. Hyaluronidase (Type VI-S Sigma Cat #H-3631 Lot #88F-807
5. Thermolysin (Protease Type X from Bacillus Thermoproteclyticus rokko Sigma Cat #P-1512 Lot #978-0833
6. α-L-Fucosidase (Bohringer Mannheim, Indianapolis, Indiana Cat #104945 Lot #1109822-10)

Procedure

Samples were processed in same manner as those in Example

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3. Basically: 2 ml of M. tuberculosis used per sample. Samples reconstituted into 100 µl of enzyme in H_2O at 500µg/ml (50 µg total) concentration, all samples incubated at 37°C 30 minutes. Gen-probe was run as a control (which involved sonicating 15 minutes at 60°C). Two phenol/chloroform extractions were performed except one extra was done with the fucosidase sample, samples were precipitated in ethanol, lyophillized, resuspended in TE buffer and run on a 1% agarose gel, followed by ethidium bromide staining.

The visual results indicate the enzyme achromopeptidase liberates more DNA than other enzymes.

EXAMPLE 5

This experiment optimizes pH and buffer conditions for lysis of mycobacteria using achromopeptidase.

Materials:

- M. tuberculosis
- Bactec bottle #16
- Achromopeptidase (Sigma Cat# A-3547 Lot #88F-0799)
- Trizma 7.0 (E Cat #3503 Lot #89F-5615
- Trizma 8.0 (Sigma Cat #T-4753 Lot #28F-5628)
- Boric Acid (Aldrich, Milwaukee, WI Cat #26,646-2 Lot #3501TJ)
- Sodium Chloride (Sigma Cat # S-9625 Lot #27F-6086)

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- Phenol:chloroform 1:1 tris saturated
- 70% ethanol (Baxter, Charlotte, NC B+J brand Lot #E-K476
- 3M sodium acetate pH 5.2
- Freeze drying unit (Virtis model)
- Electrophoresis equipment

The following 4 buffers were prepared in 50 ml amounts:

1. 100 mM Trizma pH 7.0 + 10 mM NaCl
2. 100 mM Trizma pH 8.0 + 10 mM NaCl
3. 100 mM Trizma pH 9.0 + 10 mM NaCl
4. 60 mM Borate pH 9.0 + 10 mM NaCl

Four 2ml aliquots of M. tuberculosis were harvested and the pellets resuspended in 1 ml of each buffer.

Ten µl of a 5 mg/ml solution of achromopeptidase in H_2O was added to each 1 ml aliquot (50 µg total).

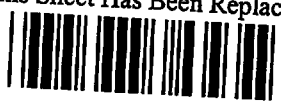
All samples were incubated at about 50°C 30 minutes, vortexing several times during incubation.

Each sample was subjected to two phenol/chloroform extractions, followed by an overnight incubation at -20°C with 2 volumes ethanol and 1/10 volume 3M sodium acetate.

DNA was precipitated and pellet was dried in a speed vaccuum, resuspended in TE tracing dye and run on agarose gel.

Results show that Trizma 9.0 + 10 mM NaCl is the best buffer. More DNA is seen using this buffer than the others.

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Although the invention has been described with respect to specific modifications, the details thereof are not to be construed as limitations, for it will be apparent that various equivalents, changes and modifications may be resorted to without departing from the spirit and scope thereof, and it is understood that such equivalent embodiments are to be included therein.

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WHAT IS CLAIMED IS:

1. A method for lysing Mycobacteria which comprises exposing the Mycobacteria to a lysis effective amount of the enzyme achromopeptidase.
2. The method of claim 1 in which the lysis effective amount of achromopeptidase is from about 50 to about 1000 units.
3. The method of claim 1 in which the Mycobacteria is selected from the group consisting of M. avium, M. intracellularae, M. gordonae, M. tuberculosis, M. kansasii, M. fortuitum, M. chelonae, M. bovis, M. scrofulaceum, M. paratuberculosis, M. phlei, M. marinum, M. simiae, M. scrofulaceum, M. szulgai,, M. intracellulare, M. leprae, M. xenopi, M. ulcerans, M. lepraemurium, M. flavescens, M. terrae, M. nonchromogenicum, M. malmoense, M. asiaticum, M. vaccae, M. gastri, M. triviale, M. haemophicum, M. africanum, M. thermoresistable, and M. smegmatis.
4. The method of claim 3 in which the Mycobacteria is M. tuberculosis.
5. The method of claim 1 which further comprises isolation of cellular components.
6. The method of claim 1 which further comprises amplification of Mycobacteria nucleic acid.
7. The method of claim 1 which further comprises the addition of a Mycobacteria identifying agent.
8. A kit comprising a Mycobacteria identifying agent and a lysis effective amount of achromopeptidase.

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9. The kit of claim 8 in which the Mycobacteria identifying agent is a nucleic acid probe.

10. The kit of claim 8 further comprising a liquification agent.

