

MYCELIATED COFFEE PRODUCTS AND METHODS FOR MAKING MYCELIATED COFFEE PRODUCTS

CROSS-REFERENCE TO RELATED APPLICATIONS

[0001] U.S. Provisional Application No. 61/802,256, filed March 15, 2013 and entitled “Myceliated Agricultural Substrates for Production of Nutraceutical Formulations and Functional Foods”; U.S. Patent Application No. 13/844,685, filed March 15, 2013 and entitled “A Method for Myceliating Coffee (amended)”; U.S. Patent Application No. 13/859,719, filed April 9, 2013 and entitled “Method of Myceliation of Agricultural Substrates for Producing Functional Foods and Nutraceuticals”; U.S. Patent Application No. 13/874,832, filed May 1, 2013 and entitled “Extract of a Myceliated Agricultural Substrate and it's Use as a Nutraceutical Composition”; U.S. Provisional Application No. 61/844,498, filed July 10, 2013 and entitled “Method for Manufacturing Coffee by Fermentation”; U.S. Provisional Application No. 61/396,863, filed July 23, 2013 and entitled “Coffee Decaffeination Process Including Fungal Components”; U.S. Provisional Application No. 61/866,371, filed August 15, 2013 and entitled “Cocoa Beans Including Metaboloites of 2-Methoxy-3-Isopropylpyrazine and Method for Metabolizing 2-Methoxy-3-Isopropylpyrazine in Cocoa Beans”; U.S. Provisional Application No. 61/867,501, filed August 19, 2013 and entitled “Cocoa Beans including Methyl Pyrazines Produced by Fermentation, and a Method of Increasing Methyl Pyrazines in Cocoa”; U.S. Patent Application No. 14/020,512, filed September 6, 2013 and entitled “Improved Method for Myceliating Raw Coffee Beans Including Removal of Chlorogenic Acids”; U.S. Patent Application No. 14/020,781, filed September 6, 2013 and entitled “Method for Myceliating Raw Coffee Beans Including Removal of Chlorogenic Acids”; U.S. Provisional Application No. 61/878,037, filed September 15, 2013 and entitled “Myceliating Green Coffee Beans to Lessen the Tooth-Staining Impact of Coffee Melanoidins”; and U.S. Provisional Application No. 61/896,097, filed October 27, 2013 and entitled “Cocoa Myceliation in an Aqueous Solution”; all of which are incorporated herein by reference in their entirety.

TECHNICAL FIELD

[0002] The methods and products relate to use of fungal strains to improve flavor in agricultural substrates, in particular, coffee.

BACKGROUND

[0003] Coffee has been mixed or infused with fungal components such as dried mushroom powders, or extracts. These mixtures have the benefit of providing additional nutritional

value derived from fungal polysaccharides. However, this method of mixing fungal components does little to improve the flavor of the coffee.

[0004] U.S. Patent Publication 20100239711 A1 to Pei-Jung Let et al. describes a method of manufacturing coffee by solid state fermentation using fungal mycelium. It is a natural process. Untreated green coffee is deposited into a dust-free container, the coffee is inoculated with a fungal strain, carried out to perform a sterile operation for implanting fungi into the coffee beans, and a fungal fermentation process is initiated. *Antrodia Camphorate*, a fungal strain native to Taiwan is utilized. The process takes between 15 to 60 days. U.S. The entire disclosure of Patent Publication 20100239711 is incorporated herein by reference in its entirety.

[0005] Many strains of Fungi can co-exist with chlorogenic acids, but the presence of chlorogenic acids reduces the ability of these fungal strains to optimally metabolize various substrates, including green coffee beans.

[0006] A portion of the coffee market is represented by decaffeinated coffee. Current caffeine extraction techniques include pre-wetting the beans to a moisture content of 40% or more then extraction with a water immiscible organic solvent, and, after decaffeination, the residual solvent is removed from the beans (de-solventizing), usually by steam stripping.

[0007] A disadvantage to these processes of caffeine removal is that there is some removal of coffee flavor and aroma compounds or precursors, especially water-insoluble waxes, during green bean decaffeination.

[0008] A need remains in the art for improved coffee products having reduced levels of undesirable taste components and/or increased levels of flavor and/or health promoting components relative to green coffee beans, and for methods of obtaining such products.

BRIEF DESCRIPTION OF THE DRAWINGS

[0009] FIG. 1 is a flow chart of a method of creating a bio-available mycelium extract in accordance with the present invention.

[0010] FIG. 2 is a flow chart of a method of creating an extract of myceliated agricultural product for human consumption to effectuate neuro-regeneration and neuro-protection in humans in accordance with the present invention.

[0011] FIG. 3 is a flow chart of a method of removing caffeine from green coffee beans according to the instant invention.

[0012] FIG. 4 is a flow chart of a method of creating a myceliated coffee product.

[0013] FIG. 5 is a flow chart of a method of creating a myceliated coffee product.

[0014] FIG. 6 is a conveyer used in accordance with the present invention.

[0015] FIG. 7 is a conveyer used in accordance with the present invention to deliver containers to an autoclave.

[0016] FIG. 8(A), 8(B), 8(C) show autoclavable bags having a pebbled surface, a stack of autoclavable bags, and a rolled autoclavable material.

SUMMARY OF THE INVENTION

[0017] In one embodiment, the present invention provides a method for the preparation of a myceliated coffee product. This method includes the step of providing prepared green coffee beans, which includes providing green coffee beans and sterilizing the green coffee beans to provide prepared green coffee beans. The method also includes the step of providing a prepared fungal component. The method also comprises inoculating the prepared green coffee beans with the prepared fungal component and culturing the inoculum to prepare the myceliated coffee product.

[0018] In one embodiment, the method includes reducing the amount of undesirable taste components. Reducing the amount of undesirable taste components optionally includes at least one, or two aqueous extractions of the green coffee beans. An undesirable taste component includes chlorogenic acid.

[0019] In one embodiment, the method includes a step of hydrating the green coffee beans, optionally to about 60%. The methods of the instant invention also include where the prepared fungal component is *G. lucidum*, *C. sinensis*, and/or *T. melanosporum*, optionally, *G. lucidum* strain 806. The methods of the invention include screening a number of strains of fungi and selecting a strain having an enhanced ability to grow on, metabolize or utilize green coffee beans, and/or selecting a strain that is capable of enhanced removal of one or more undesirable taste components from the green coffee beans, and/or enhanced removal of caffeine from the green coffee beans.

[0020] In another embodiment, the prepared fungal component is maintained on an undefined media comprising an aqueous green coffee bean extract and an energy source. The maintenance of the strain of fungi causes an adaptation of the fungi resulting in enhancement of the fungi's ability to grow on, metabolize or utilize green coffee beans.

[0021] The inventive methods also include where the culturing step is a fermentation step carried out under semi-anaerobic conditions, optionally for about 7 days.

[0022] The methods of the instant invention result in myceliated coffee products having reduced levels of undesirable taste components, such as chlorogenic acids, and increased

levels of myceliation products, such as beta glucans and polysaccharides, relative to the green coffee beans.

[0023] The green coffee beans may be from *Coffea Arabica* or *Coffea robusta*.

[0024] The present invention includes a myceliated coffee product and green coffee beans prepared by the methods of the invention, and a myceliated coffee product having reduced levels of undesirable taste components and increased levels of myceliation products relative to green coffee beans.

[0025] Methods also include a method for the reduction of the levels of undesirable taste components in green coffee beans and a method for the increase in levels of myceliation products in green coffee beans as herein described.

DETAILED DESCRIPTION OF THE INVENTION

[0026] In one embodiment, the present invention provides a method for the preparation of a myceliated coffee product. This method includes the step of providing prepared green coffee beans, which includes providing green coffee beans and sterilizing the green coffee beans to provide prepared green coffee beans. The method also includes the step of providing a prepared fungal component. The method also comprises inoculating the prepared green coffee beans with the prepared fungal component and culturing the prepared green coffee beans and prepared fungal component to allow myceliation to produce the myceliated coffee product. These steps may be performed in any order.

[0027] In one embodiment, prepared green coffee beans are provided, which includes the step of providing green coffee beans. Coffee refers to genus *Coffea* which is a genus of flowering plants whose seeds, called coffee beans, are used to make coffee. It is a member of the Rubiaceae family. Coffee beans may be selected from one of several coffee varieties which are diverse cultivars derived through selective breeding or natural selection of coffee plants. Coffee beans of the same variety grown in different locations may have distinctive characteristics such as flavor (flavor criteria include terms such as "citrus-like" or "earthy"), caffeine content, body or mouthfeel, and acidity. Green coffee beans useful for the present invention may be any species of coffee, including *Coffea arabica* and *Coffea robusta* (also known as *Coffea conephora*); additional species of coffee useful for the present invention include *Coffea benghalensis*, or Bengal coffee; *Coffea congensis*, or Congo coffee; *Coffea liberica*, or Liberian coffee; *Coffea stenophylla*, or Sierra Leonian coffee; *Coffea excelsia*, another Liberian coffee; *Coffea bonnierii*; *Coffea gallienii*; and *Coffea mogenetii*. The invention includes any varietals or strains of the species listed above. For example, many Arabica varietals are named after the country or region in which they are predominantly

found, or in which they originated. Some of the exemplary varieties of Arabica coffee include Typica, Bourbon, Caturra, Catuai, Mundo Nova, and Blue Mountain. Also included in the invention are and derivative species of coffee including any genetically-modified (GMO) strains or cultivars and also any heirloom variety (non-GMO) strains or cultivars of coffee.

[0028] A green coffee bean refers to a raw, unroasted coffee bean. Generally, the raw fruit of the coffee plant is referred to as a coffee cherry. To prepare the green coffee bean, generally, the coffee cherry has the fruit or pulp removed and the seed, or fruit, is then dried. Fruit and pulp may be removed from the green coffee bean by various methods known in the art and include a wet process where the fruit/pulp is removed from the coffee bean prior to drying, and a dry process where the whole cherries are dried prior to mechanical hulling, sorting, grading, and bagging takes place. Dried coffee beans useful for the instant process may include dried coffee beans that are fresh, or may be subject to an aging process. Dried coffee beans may be stored in burlap bags, lined burlap bags, or in vacuum sealed containers prior to entry into the instant process.

[0029] In some embodiments, the green coffee bean is not dried prior to being used in the processes of the instant invention. In this embodiment, after the green coffee bean is harvested, the green coffee bean has the pulp removed by any processes known in the art, and then can be used in the present invention without further green coffee bean treatment, such as drying. In this embodiment, the hydration and/or washing step as described below is not necessary or is obviated by the use of the undried green coffee bean.

[0030] HYDRATION AND WASHING STEP

[0031] In one embodiment, the green coffee beans are prepared for use in the instant methods of the invention, resulting in a prepared green coffee bean.

[0032] In some embodiments, the green coffee beans are prepared by a step of hydrating the green coffee beans. Hydration is particularly useful where the green coffee beans have been dried. Hydration ensures that the green coffee beans have optimal moisture content for the culturing (myceliation) process. Hydration may be accomplished by a number of methods known in the art.

[0033] The hydration may be accomplished by an aqueous medium. The aqueous medium includes water and optionally, additional excipients. Water may be distilled or mineralized. Other excipients can be added to the water, such as buffers to maintain a certain pH, sodium chloride, citric acid and/or ascorbic acid. The pH may be neutral or adjusted. The

temperature of the aqueous medium may be room temperature, or elevated in temperature to accelerate the hydration process.

[0034] Hydration may be accomplished by allowing the green coffee beans to soak in the aqueous medium for any appropriate length of time, ranging from a few seconds or less to overnight. The soaking step for the hydration and/or aqueous extraction step may be less than a second, at least five seconds, at least ten seconds, at least thirty seconds, at least a minute, at least five minutes, at least ten minutes, at least twenty minutes, at least thirty minutes, at least forty minutes, at least fifty minutes, at least an hour, at least an hour and a half, at least two hours, at least two and a half hours, at least three hours, at least four hours, at least five hours, at least six hours, at least seven hours, at least eight hours, at least ten hours, at least twelve hours, or at least fifteen hours, at least eighteen hours, at least twenty four hours, at least thirty six hours, or at least forty-eight hours. However, the time for the hydration step should be selected in view of the fact that the green coffee beans are not sterile and soaking for too long of a time may encourage the growth of undesirable organisms.

[0035] The green coffee beans may be hydrated at any temperature that allows for effective hydration; in one embodiment, the temperature of the aqueous component temperature is room temperature. Hydration temperature should be selected in view of the fact that at high temperatures, desirable flavor components may be altered.

[0036] Hydration may be performed under normal atmospheric pressure or may be performed under increased pressures to accelerate the hydration process and/or aqueous extraction process, such as between 1 atmospheric pressure and 2 atmospheric pressures, for example, at 1.5 atmospheres.

[0037] Moisture content of the hydrated green coffee beans is optionally between about 20% and about 80% moisture content, between a 40% and 70% moisture content. In one embodiment, the moisture content is at least about 30%, at least about 50%, or at least about 60%.

[0038] The hydration step may occur in a container. In one embodiment, the container is a drum, such as a 55 gallon drum. In this embodiment, volumetric 1:1 (equal volumes) ratio green coffee beans:aqueous component are allowed to stand for 5 – 24 hours. For example, the 55 gallon drum is filled half-way to the top with green coffee beans and aqueous medium is added to completely fill the drum. In this embodiment, the green coffee beans may absorb all of the aqueous component. In another embodiment, the cacao beans or other agricultural substrate, held in a container, may be filled with water to completely submerge the beans, in one embodiment, without significant excess.

[0039] In another embodiment, green coffee beans that have been demucilaged but not yet dried, which on average have a moisture content of 60 %, may be used. This method avoids the hydration step.

[0040] In one embodiment, the step of providing prepared coffee beans optionally includes a step of removing undesirable taste components by washing or rinsing the green coffee beans. The wash or rinse may be the aqueous medium as described above. In one embodiment, the green coffee beans are optionally washed or rinsed prior to, during, or after the optional hydration step. Washing, draining and/or rinsing the green coffee beans can be performed by any method known in the art. The green coffee beans may be washed one time, at least two times, at least three times, at least four times, at least five times, at least ten times, at least fifteen times, at least twenty times, at least fifty times or more. In one embodiment the wash step is performed two times. The wash or rinse step may include optional soaking times as described herein.

[0041] In one embodiment, the green coffee beans are washed by a method of filling a container holding the green coffee beans with water, allowing the water to soak for 10 seconds to 4 hours, draining the water off and repeating the steps as many times as desired, or to raise the beans to the desired moisture level. The washing or rinsing step may also be carried out until the green coffee beans have had a determined amount of undesirable taste component removed.

[0042] The green coffee beans may be washed at any temperature that allows for the efficient extraction of undesirable taste components; in one embodiment, the temperature of the aqueous medium temperature is room temperature. Wash temperature should be selected in mind of the fact that at high temperatures, desirable flavor components may be altered, destroyed and/or extracted.

[0043] In another embodiment, the excess aqueous medium or component is removed and/or separated and/or drained from the hydrated coffee beans after the hydration step. This step may also be referred to as an aqueous extraction step. This step may be done to remove undesirable taste components.

[0044] The major components of coffee include caffeine, minerals, tannic acid, cellulose, water, fat, protein and fibers. Coffee contains methylxanthine such as caffeine, theophylline, and theobromine, flavonoids, phenols, phenolic acids, volatile alkaloids, non-volatile alkaloids. Coffee contains some undesirable taste components as well. These components will contribute to a perception of a harsh and/or bitter taste to the coffee. These tastes are commonly mitigated by addition of sugar or cream to mask the bitter components. Reduced

bitterness and/or harshness is noted in more premium, expensive coffee varieties such as Arabica coffee.

[0045] The hydration step, aqueous extraction step, wash and/or rinse step can optionally reduce and/or remove undesirable taste components from the green coffee beans and may be carried out as described herein until the desired amount of undesirable taste component has been removed from the green coffee beans.

[0046] Green coffee beans (and roasted conventional coffee beans) are known to contain a number of components which contribute to a harsh or bitter flavor. These flavor components are considered undesirable in coffee. One such component is chlorogenic acid which is an ester of caffeic acid and quinic acid and may be described as an acrylic acid derivative. During roasting the smoke from the coffee is toxic to the respiratory tract. Chlorogenic acids contribute to a harsh and/or bitter taste in coffee and coffee products, and may be toxic to both humans, mammals, and various strains of fungi.

[0047] Chlorogenic acid has a green color and its presence in the green coffee beans contributes to the green color of the beans. Optionally, even where steps are performed to remove chlorogenic acid while creating the prepared green coffee beans, at least some amount of chlorogenic acid is left in the prepared green coffee beans, as it contributes to the characteristic taste of coffee. In some embodiments at least some chlorogenic acid in the green coffee beans is retained. Chlorogenic acid has been reported to carry certain health benefits, particularly in its ability to mediate insulin levels and thus aid in adipose metabolism. It has also been identified as a powerful antioxidant. Chlorogenic acid is converted to a lactone upon roasting as a result of dehydration at the quinnic acid moiety. The lactone may be incorporated into macromolecules known as melanoidins, molecules that mass upwards of 25,000 kD and are the result of numerous and subsequent Maillard reactions (any of the carbonyls or hydroxyl groups on the lactone could serve as nucleophiles in this reaction), making them less bioavailable than in pure form. Moreover, extensive roasting (i.e. dark roast) will degrade the lactone into hydroxylated phenylindanes. Therefore the roasted myceliation product may not contain chlorogenic acid, rather it may contain the lactone. The chlorogenic acid lactone bio-functionalities are much less understood. Chlorogenic acid has also been implicated as an important tastant in coffee taste profile.

[0048] If quantitative removal of chlorogenic acid from green coffee beans is desired, green coffee beans may be autoclaved in excess water; for example 1 lb green coffee beans may be mixed in 3 L of water and autoclaved on a liquid cycle for 30 – 80 minutes, resulting in nearly white beans. However, quantitative removal of chlorogenic acid is not preferred.

[0049] Without being bound by theory, the inventors believe that green coffee beans' chlorogenic acid can reduce the growth of at least some of the fungal strains of the present invention and may further interfere with some or all of the processes of the present invention by interfering or reducing growth of the fungi and/or ability of the fungi to metabolize, or myceliate, the green coffee beans. When chlorogenic acids are not removed from the green coffee beans, higher moisture contents (for example, 60%) of the green coffee beans are preferred. Removal of at least some of the chlorogenic acid allows for the culturing or myceliation step to occur at lower moisture content such as 30% and greater. The inventors attribute this to coffee's 20% cellulose content.

[0050] In some embodiments, as optionally measured by the intensity and/or presence of green color of the green coffee beans, the aqueous extraction, wash and/or rinse step is carried out until about 5% of chlorogenic acids are removed; in other embodiments, up to 10%, up to 15%, up to 20%, up to 25%, up to 30%, up to 35%, up to 40%, up to 45%, up to 50%, up to 55%, up to 60%, up to 65%, up to 70%, up to 75%, up to 80%, up to 85%, up to 90%, or up to 95% of chlorogenic acids are removed in the processes of the instant invention. In some embodiments, about 25% to about 80% of the chlorogenic acids are removed. In one embodiment, about 45-50% of the chlorogenic acids are removed.

[0051] Other undesirable taste components contributing to a bitter taste in coffee include quinic acid, 5-hydroxymethylfurfural, 2-methylfuran, furfuryl alcohol, trigonelline, caffeic acid, citric acid, malic acid, lactic acid, pyruvic acid, acetic acid, pyrazine, thiazole, quinolone, phenylpyridine, caffeine, among others. Robusta coffee contains higher levels of both caffeine and chlorogenic acids, leading to increased bitterness and astringency in Robusta coffee. Additional undesirable taste components can include one or more of theophylline, theobromine, paraxanthine, liberine, methyl-liberine, trigonelline (N-methylnicotinate); hydrophobic amino acids such as isoleucine, leucine, valine, tyrosine, phenylalanine, gamma-aminobutyric acid; diketopiperazines such as cyclo(proline-proline), cyclo(proline-leucine), and cyclo(proline-isoleucine); acetic acid, propionic acid, butanoic acid, pentanoic acid, hexanoic acid, heptanoic acid, octanoic acid; nonanoic acid; decanoic acid and derivatives of such fatty acids; 3-methyl-valeric acid, acetaldehyde, propanal, butanal, pentanal; carboxylic acid-5-hydroxytryptamides with an amide bond to fatty acids (unsaturated C6 to C24); the triglycerides linoleic acid, palmitic acid and related esters; diterpenes including cafestol, kahweol, 16-O-methyl-kafeestol, cafestal and kahweal; chlorogenic acids; polyphenols; and chlorogenic acids such as ferulic acid and 3,4-dimethoxycinnamic acid, which are connected by an ester bond to the hydroxyl groups of

quinic acid. Other harsh and/or bitter flavor components include chlorogenic acid lactones and breakdown products of the lactones such as phenylindanes. One or more of the above-named compounds may be reduced and/or removed by the methods of the invention.

[0052] In some embodiments, determination of the extent of the removal of at least one undesirable taste component is determined by the appearance, taste and/or chemical composition (by methods known in the art) of the myceliated coffee product. Alternatively, the green coffee bean's appearance or chemical composition may be determined by known methods. This determination may be quantitative, e.g., the chemical composition of the myceliated coffee product may be measured by assay methods, or determined qualitatively by taste testing by skilled persons.

[0053] In one embodiment, up to 5% of one or more of the undesirable taste components are removed; in other embodiments, up to 10%, up to 15%, up to 20%, up to 25%, up to 30%, up to 35%, up to 40%, up to 45%, up to 50%, up to 55%, up to 60%, up to 65%, up to 70%, up to 75%, up to 80%, up to 85%, up to 90%, or up to 95% of one or more of the undesirable flavor components are removed in the processes of the instant invention. In one embodiment, one or more of the undesirable flavor components are quantitatively removed.

[0054] Removal of undesirable taste components may allow for increasing the value of poorer quality coffee and/or rendering it more drinkable. Myceliated coffee products produced by this method may be used to blend with less expensive coffee beans leading to a lower cost product having improved taste properties. The amount of sugar, milk and substitutes thereof to be added to the coffee may be reduced. The instant methods lead to enhanced flavor profile of the myceliated coffee products due to a perception that the myceliated coffee products provide a richer, smoother, and/or sweeter coffee with less bitter, harsh, and/or acidic tastes. Green coffee beans and myceliated coffee beans that have been subjected to the hydration and/or aqueous extraction steps as described above will have reduced amounts of chlorogenic acid. Myceliated coffee products of the present invention will demonstrate improved flavor. In one embodiment, the improved flavor results from removal and/or reduction of bitter-tasting compounds such as chlorogenic acid in the myceliated coffee beans.

[0055] In one embodiment, reduction of the desirable flavor components such as volatile oils is minimized by the processes of the present invention. In processing green coffee beans from Robusta coffee, the art teaches to steam treat, steam extract, or steam strip the beans prior to roasting, which can remove many desirable volatile oils from the Robusta coffee beans. The processes of the instant invention avoid the steam roasting step for Robusta

coffee beans, thereby helping to preserve the desirable volatile oils that contribute to coffee flavor.

[0056] In an optional step, coffee beans can be decaffeinated by conventional processes prior to, subsequent to, or in addition to the methods of the instant invention.

[0057] HEAT TREATMENT

[0058] The methods of the present invention further optionally comprise a method of heat treatment such as pasteurizing and/or sterilizing the green coffee beans. In one embodiment, the green coffee beans are sterilized to provide prepared green coffee beans. This step may be accomplished by any method known in the art. For example, this step may be performed under atmospheric pressure or under increased pressure. This step may also be referred to as "pre-processing." This step is performed to reduce or remove undesirable microbial or fungal organism contaminants on the green coffee beans.

[0059] Methods for pasteurization and/or sterilization may be carried out as known in the art. As an example of pasteurization, green coffee beans may be subjected to dry heat treatment at atmospheric pressure at 145 °F to 190 °F for 30 to 90 minutes, alternatively at 140 °F to 210 °F for 20-100 minutes.

[0060] Sterilization of the green coffee beans may be performed as is known in the art. For example, green coffee beans may be sterilized by heating under pressure at 15 lb/in² at 121-122 °C for 20 to 100 minutes, such as 90 minutes. Other examples of suitable sterilization conditions include sterilization to 108-109 °C at 5 lb/in², 115-116 °C at 10 lb/in², for 20 to 90 minutes. In another embodiment, the steam is superheated to 250-255 °F. The pressures may vary from 5-25 lbs per square inch. Green coffee beans may be sterilized in a container. The container may optionally be the same container as the container used for the aqueous extraction and/or hydration step. The container may be optionally sealed and the green coffee beans may be sterilized by the application of heat to the exterior of the container. In one embodiment, the heat is provided by applying steam to the exterior of the container for a sufficient period of time to allow for sterilization of the contents.

[0061] The sealed container of some embodiments can provide some advantages. For example, sealing the container minimizes outflow of flavor components and aromatic components from the green coffee beans, which can be noticed by the lack of coffee aroma from steam from the pressure cooker or autoclave during the sterilization process. Sealing also prevents water-soluble flavor and aromatic components from escaping the green coffee beans directly into steam, hot air, or heated water.

[0062] Suitable containers include containers known in the art for mushroom cultivation. Optionally the containers have a section for exchanging air or gases but do not allow passage of any other component. Such sections are known in the art and include filter strips. In one embodiment, the container is a drum, for example, a 55 gallon drum.

[0063] In some embodiments, the containers of the instant invention can be glass, stainless steel, temperature-resistant high density polyethylene or polypropylene bags. Fermentors and bioreactors can also be used as containers of the instant invention. In some embodiments, the containers have a means for gas exchange that precludes passage of contaminants, such as filter zones or valves.

[0064] In one embodiment the container is a bag, for example, an autoclavable, polypropylene bag with filter strips, an autoclavable, high density polyethylene bag with filter zones, and a gamma-irradiated polyethylene bag with filter zones.

[0065] A further advantage of the bags described above is that when sealed, they conform to shape of the green coffee beans when pressurized during the sterilization step. Conforming the bags to the shape of the green coffee beans inhibits movement of the green coffee beans relative to each other, preventing or minimizing degradation of the surface of the beans. This conforming of the bag to the shape of the green coffee beans also improves heat transfer, as the lack of air prevents air insulation of the green coffee beans from heat. The bags can be of any dimension. In one embodiment, bags are elongated or flattened to hasten the heating process, for example, the length may be three times the diameter of the bag. This dimension may also facilitate the advantageous stacking of bags or positioning of bags for sterilization.

[0066] The size of the bags to be used can be chosen according to the volume or amount of green coffee beans to treat by the methods of the present invention. Exemplary amounts of green coffee beans to use per bag include 1 lb of green coffee beans to 150 lbs of green coffee beans, although larger and smaller amounts of green coffee beans are contemplated. For example, amounts of 1 gram or 10,000 lb of green coffee beans can be treated by the method.

[0067] In another embodiment, the bags are flattened, having a thickness of 1/10th or less than the sum of the peripheral edges of each bag. The bags can be round in shape, having a circumference that defines the peripheral edges of each bag. Alternatively, the bags can be rectangular so that the sum of the sides defines the peripheral edges of each bag. The bags can be conjoined so that a series of rectangular bags can be easily handled in a production environment. All bags have breathable patches (filter strips) that provide for the approximation of an anaerobic environment. In yet another embodiment, the bags are

flattened to hold a layer of beans being less than three beans thick. Accordingly, heat quickly penetrates the flattened bags to the beans to effectuate sterilization or pasteurization. In this embodiment, due to the pressurization, the bag will conform to the shape of the green coffee beans, and this will yield a pebbled surface on the outer surface of each bag when pressure is applied. The pebbled bag surface forms interstitial spaces that allow heat to penetrate between bags that are stacked to accelerate the sterilization or pasteurization process. The pebbled surface of the bags also induces turbulent fluid flow along the bag surface to improve heat transfer to the green coffee beans.

[0068] In another embodiment, the green coffee beans are vacuum packed in the bags to eliminate air that could draw volatile flavor or aromatic components from the bags.

[0069] In another embodiment, the bags are replaced by sheets of autoclavable material, such as BPA-free plastic. One base sheet is continuously dispensed along the top of a conveyor, green coffee beans are then laid on the dispensed base sheet. A second top sheet is overlaid upon the green coffee beans and sealed to the base sheet. A vacuum is applied between the top and bottom sheet to evacuate air, then the sheets are sealed at predetermined distances to form sections. Each section holds a pre-determined volume of green coffee beans. The sections are conveyed through an autoclave, or oven, to effectuate the pasteurization or sterilization process. Heat may be applied in a pressurized or non-pressurized environment in the form of steam, hot water under pressure, hot air in turbulent or laminar flow over the sheets, or other heated fluid. In a variation of this embodiment, the sections containing the green coffee beans are rolled and placed in an autoclave for pressurization or sterilization. One roll can contain many sections.

[0070] Since the green coffee beans cause a pebbled surface on the exterior of the sheets, interstitial space exists on the outside surface of the sheets to hasten the pasteurization or sterilization process by allowing heated fluid to readily penetrate between sheets. The pebbled sheet surface also induces turbulent fluid flow that further improves heat transfer to the green coffee beans. The pebbled surface inhibits relative movement between the beans to assure that the green coffee beans do not crack, break or rub.

[0071] FUNGAL COMPONENT

[0072] The fungal component to use with the present invention can be a fungi from phylum Basidiomycotina of Eumycota, including any Fungi belonging to Polyporaceae and Hericiaceae, wherein Fungi selected from Basidiomycotina of Eumycota include Eumycota, including at least one selected from Basidiomycotina and Ascomycotina, including the strains: *Hericium erinaceus*, *Pleurotus ostreatus*, *Pleurotus eryngii*, *Pleurotus citrinopileatus*,

Pleurotus djamor, *Tramete versicolor*, *Lentinula edodes*, *Armillariella mellea*, *Tricholoma matsutake*, *Flammulina velutipes*, *Vovariella volvacea*, *Agaricus campestris*, *Agaricus blazei*, *Grifola frondosa*, *Pholiota nameko*, *Agrocybe cylindracea*, *Boletus ornatipes*, *Ganoderma lucidum*, *Ganoderma applanatum*, *Hypsizygus marmoreus*, *Morchella hortensis*, *Phellinus linteus*, *Auricularia auricula*, *Tremella fuciformis*, *Inonotus obliquus*, *Fomes fometarius*, *Laetiporus sulfureus*, *Bridgeoporus nobilissimus*, *Cordyceps sinensis*, *Cordyceps militaris*, *Xylaria nigripes*, *Tuber melanosporum*, *Polyporus umbellatus*. The Fungi used in this invention for clean myceliation of green coffee beans may also include Ascomycotina of Eumycota, including Clavicipitaceae, wherein the Fungi selected from Ascomycotina of Eumycota include Clavicipitaceae such as *Cordyceps sinensis* and *Cordyceps militaris*; and Xylariaceae, such as *Xylaria nigripes*. Combinations of the above-identified strains are also contemplated. In some embodiments, the present invention utilizes *Ganoderma lucidum* or *Cordyceps sinensis*.

[0073] Generally, the invention does not contemplate use of the following fungi: *Rhizopus chinensis*, *R. oligosporus*, *Aspergillus flavusoryzae*, *A. tamari*, *A. niger*, *A. nidulans*, *A. sojae*, *Fusarium venenatum*, *F. graminearum*, *Saccharomyces cerevisiae*, *S. exiguus*, *S. pombe*, *Saccharomycopsis (Candida) lipolytica*, *Candida utilis*, *C. krusei* or *C. tropicalis*, *Pichia saitoi*, *Kluyveromyces fragilis*, *Endomycopsis fibuliger*, the Ascomycete *Chaetomium*, *Zygosaccharomyces rouxii*, *Mucor racemosus*, *Geotrichum candidum*, *Penicillium camemberti*, *P. notatum*, *P. griseofulvum*, *P. grisea*, *P. chrysogenum*, *P. roqueforti*, *P. nalgiovense*, *Neurospora intermedia*, *Amylomyces rouxii*, *Endomycopsis burtonii*, *Psycilocibin*, *Monascus purpureus*, *Debaryomyces hansenii*, *Ashbya gossypii*, *Blakeslea trispora*, *Tolypocladium niveum*, *T. inflatum*, *Streptomyces*, *Neocosmospora*, *Stachybotrys*, *Beauveria*, *Cephalosporium acremonium*, *C. acremonium*, *Gibberella fujikuroi*, *Fusidium coccineum*, *Monascus ruber*, *Claviceps fusiformis*, *C. paspali*, *C. purpurea*, *Aminita muscaria*, or *A. phalloides*.

[0074] Fungal components useful in the present invention may be prepared by methods as described herein. For example, in one embodiment, a pure strain of fungus is used. In some embodiments, the pure strain of fungus is able to effectively grow on and/or myceliate the prepared coffee beans to prepare the myceliated coffee products. Any strain of fungus identified herein which is capable of effectively growing on and/or myceliating prepared coffee beans can be used for the methods of the present invention.

[0075] It was surprisingly found by the inventors of the instant invention that some fungal strains have enhanced and/or increased ability to grow on, metabolize or otherwise utilize

and/or modify green coffee beans and/or remove one or more undesirable taste components from the green coffee beans and/or better tolerate the presence of green coffee beans (or extract) in media. In one embodiment, the undesirable taste component is chlorogenic acid. In another embodiment, the fungal component reduces or removes caffeine from green coffee beans.

[0076] Therefore, the methods of the invention have as an optional additional step, a method of selecting a fungal component having an enhanced and/or increased ability to grow on, metabolize or otherwise utilize and/or modify green coffee beans and/or remove one or more undesirable taste components from the green coffee beans, and/or remove caffeine and/or better tolerate the presence of green coffee beans (or extract) in media. This method comprises screening a number of strains of a desired fungal species to select for a suitable fungal component (strain) which exhibits the enhanced and/or increased ability to grow on, metabolize or otherwise utilize and/or modify green coffee beans and/or remove one or more undesirable taste components and/or caffeine from the green coffee beans, and/or is better able to tolerate the presence of green coffee beans, and using this selected strain(s) in the methods of the invention.

[0077] In one embodiment, a suitable strain will generate a distinctively dark ring of coffee matter around the perimeter of the colony, showing that the strain rejects and actively excludes certain aspects of coffee as a food source. The dark color of the ring, without being bound by theory, shows that chlorogenic acid is rejected as a food source and removed/volatilized away from the strain.

[0078] In one embodiment, a pure strain of any commercially available *Ganoderma lucidum* is used as the fungal component. While all strains of *Ganoderma lucidum* are effective for the present invention, it was surprisingly found that some selected strains have the enhanced abilities useful for the present invention as described herein. One such strain useful for the fungal component of the present invention is *Ganoderma lucidum* strain 806, (Alice Chen; Buffalo, NY; 4/94) commercially available from Pennsylvania State University (The Pennsylvania State University Mushroom Culture Collection, available from the College of Agriculture Sciences, Department of Plant Pathology and Environmental Microbiology, 117 Buckhout Laboratory, The Pennsylvania State University, University Park, Pennsylvania, USA 16802.)

[0079] This strain was surprisingly determined by the present inventors to more efficiently grow on, metabolize or otherwise utilize and/or modify green coffee beans and/or tolerate green coffee beans and/or remove one or more undesirable taste components from the green

coffee beans, including chlorogenic acid. In another embodiment, this strain can remove and/or reduce the amount of caffeine in green coffee beans. Therefore, in one embodiment, the fungal component is *Ganoderma lucidum* strain 806 Alice Chen; Buffalo, NY; 4/94. These selected strain(s) were deposited with ATCC as described hereinbelow.

[0080] In one embodiment, a pure strain of any commercially available *Cordyceps sinensis* is used as the fungal component. While all strains of *Cordyceps sinensis* are effective for the present invention, it was surprisingly found that some selected strains have the enhanced abilities useful for the present invention as described herein. One such strain useful for the fungal component of the present invention is *Cordyceps sinensis* (Strain 1009 Caterpillar Fungus; Colorado Corp, 1/2014), commercially available from Pennsylvania State University (The Pennsylvania State University Mushroom Culture Collection, available from the College of Agriculture Sciences, Department of Plant Pathology and Environmental Microbiology, 117 Buckhout Laboratory, The Pennsylvania State University, University Park, Pennsylvania, USA 16802.) These selected strain(s) were deposited with ATCC as described hereinbelow.

[0081] This strain was surprisingly determined by the present inventors to more efficiently grow on, metabolize or otherwise utilize and/or modify green coffee beans and/or remove one or more undesirable taste components from the green coffee beans, including chlorogenic acid and/or better tolerate the presence of green coffee beans (or extract) in media. In another embodiment, this strain can remove and/or reduce the amount of caffeine in green coffee beans. Therefore, in one embodiment, the fungal component is *Cordyceps sinensis* (Strain 1009 Caterpillar Fungus; Colorado Corp, 1/2014). These selected strain(s) were deposited with ATCC as described hereinbelow.

[0082] Similarly selected strains for *H. erinaceus*, *T. versicolor*, *L. edodes*, *T. matsutake*, *F. velutipes*, *A. blazei*, *G. frondosa*, *P. nameko*, *L. officinalis*, *M. hortensis*, *M. angusticeps*, *A. auricula*, *T. fuciformis*, *I. obliquus*, *F. fomentarius*, *L. sulfureus*, for example, (or for any species of fungi mentioned herein) were obtained by screening a number of strains of each species to select for a suitable fungal component (strain) which exhibits the enhanced and/or increased ability to grow on, metabolize or otherwise utilize and/or modify green coffee beans and/or remove one or more undesirable taste components and/or caffeine from the green coffee beans, and/or is better able to tolerate the presence of green coffee beans, and using this selected strain(s) in the methods of the invention. Therefore, in some embodiments, the selected strain(s) of *H. erinaceus*, *T. versicolor*, *L. edodes*, *T. matsutake*, *F. velutipes*, *A. blazei*, *G. frondosa*, *P. nameko*, *L. officinalis*, *M. hortensis*, *M. angusticeps*, *A.*

auricula, *T. fuciformis*, *I. obliquus*, *F. fomentarius*, *L. sulfureus* are used in the processes of the instant invention. These selected strain(s) were deposited with ATCC as described hereinbelow.

[0083] In one embodiment, a pure strain of *Tuber melanosporum* is commercially obtained. In another embodiment, at least one strain is generated from wild truffle mushroom(s) and used in the instant invention as the fungal component. Although antibiotics may be used in the culture, more vigorous growing strains were found where culture medium lacked antibiotics. For example, a pure strain comprising a strain maternal to the original truffle mushroom can be generated by culturing on undefined agar comprising oak leaves and oak twigs. A pure strain comprising a strain paternal to the original truffle mushroom can be generated by culturing on undefined agar comprising oak leaves and oak twigs. A pure strain comprising a strain comprising a cultivar derived from non-sexual genetic recombination or anastomosis of both the maternal and the paternal mating types can be generated by culturing on undefined agar comprising oak leaves and oak twigs. In some embodiments, appropriate isolates (strains) to use include isolates that result in a pleasant flavorful aroma/taste in the myceliated coffee, reminiscent of flowers (in some embodiments, achieved by the maternal strains to a truffle mushroom as described herein), or isolates (strains) that result in a pleasant flavorful aroma/taste, reminiscent of truffles, (in some embodiments, achieved by a cultivar derived from non-sexual genetic recombination or anastomosis of both the maternal and the paternal mating types, as described herein). These selected strain(s) were deposited with ATCC as described herein.

[0084] In one embodiment, the methods of the invention include screening a number of strains of *T. melanosporum*, created as described above, to select for a suitable fungal component which exhibits the enhanced and/or increased ability to grow on, metabolize or otherwise utilize and/or modify green coffee beans and/or remove one or more undesirable taste components and/or caffeine from the green coffee beans, and/or is better able to tolerate the presence of green coffee beans, and using this selected strain(s) in the methods of the invention. These selected strain(s) were deposited with ATCC as described herein.

[0085] All strains referenced herein are deposited with the ATCC at 10801 University Boulevard, Manassas, Virginia 20110-2209 USA under the Budapest Treaty provisions. The deposit will irrevocably and without restriction or condition be available to the public upon issuance of a patent and will be maintained under the terms of the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purposes of Patent Procedure. These deposits were made merely as a convenience for those of skill in the art and

are not an admission that a deposit is required under 35 U.S.C. §112. However, it should be understood that the availability of a deposit does not constitute a license to practice the subject invention in derogation of patent rights granted by government action. The deposit will be maintained without restriction in the ATCC Depository, which is a public depository, for a period of 30 years, or 5 years after the most recent request, or for the enforceable life of the patent, whichever is longer, and will be replaced if it ever becomes nonviable during that period.

[0086] MAINTENANCE AND ADAPTATION OF FUNGAL COMPONENT

[0087] Fungal components useful in the present invention may be prepared by methods as described herein. For example, in one embodiment, the fungal component is optionally grown, maintained and/or propagated in an undefined medium comprising green coffee bean extract prior to use for inoculation of the green coffee beans. In one embodiment, the fungal component is indefinitely maintained in the undefined medium comprising green coffee bean extract in the solid-state, floating, and submerged morphologies. Without being bound by theory, the inventors believe that maintenance of the fungal component on undefined media comprising green coffee beans plays an important role in the long-term viability and health of the fungal component. It is believed that the perpetual and subtle changes made from batch to batch of agar and liquid media when using undefined medium comprising green coffee beans effectively avoids the phenomenon of undesirable genetic drift that will occur over time to the fungal component when are maintained on identical (defined) media.

[0088] The undefined medium comprising green coffee bean extract may be made by a number of methods. In one embodiment, the undefined medium comprises aqueous green coffee extract. Optionally, additional energy sources can be added. Materials are optionally organic and water at least RO filtered. It has been surprisingly found by the inventors that the medium may comprise aqueous green coffee extract without any additional added excipients, such as an additional energy source for growing fungi of the present invention.

[0089] Solid media comprising undefined medium comprising green coffee bean extract. In one embodiment, undefined medium comprising green coffee bean extract agar media to grow fungi for the eventual purpose of myceliating sterilized green coffee beans can comprise filtrate of green coffee beans. In one embodiment, 0.1 - 100 lbs. of green coffee beans are in 0.1 - 100 L of water, respectively, for 1 minute to 12 hours. The filtrate was collected through 1 - 3 filtrations of the mixture, and 14-17 g/L of agar was added. This base solution was then mixed with the filtrate of undefined vegetable (of any type but ideally organic) aqueous extract, such as malt extract, yeast extract, potato. In one embodiment the

vegetable is potato. Aqueous potato mixture can be prepared by softening 1 – 300 g of potato mass in boiling or pressurized water, mashed, and the filtrate was collected through 1-3 filtrations. Optionally, fruit juice with no added sugars can also be added to the base green coffee agar media. In one embodiment, the medium comprises 0.1-10 % by weight of malt extract, 0.1-10 % by weight undefined vegetable extract with essence of green coffee bean, 0.1-10 % by weight of yeast extract, 0.1-10 % by weight of peptone, 0.1-10% by weight of glucose, 20-80 % by weight of water, and 1-90% by weight whole green coffee beans or green coffee bean extract.

[0090] As a non-limiting example of the media, for example, 2 lbs green coffee beans, either pulverized or whole can be mixed with ¼ gallon water at room temperature. The mixture may be blended. The mixture is then allowed to extract for 20 minutes with shaking, then filtered three times through fine mesh. Separately, about 5 organic potatoes are placed in 10 L of water and autoclaved 20 minutes to soften the potatoes. The potatoes are then pulverized with a potato masher, and then filtered through fine mesh three times. 1 L of commercial unsweetened fruit juice can be added. These solutions are combined and autoclaved.

[0091] Once prepared, the media can be sterilized by any method known in the art. Once the media cooled, it was poured into Petri plates and fungal cultures were propagated from plate to plate in sterile operation, as known in the art. Slants for test tubes and flasks may be prepared by this method. Petri plates can be inoculated with floating and submerged liquid tissue culture, and with myceliated substrate.

[0092] Liquid media undefined medium comprising green coffee bean extract. Green coffee bean extract and undefined vegetable extract were prepared as described for solid media, except that no agar is added. If preparing to make a floating culture, 1 – 10 tablespoons of flour was added to the mixture, in one embodiment, about 1 tablespoon per 10-15 L of culture. The media can be sterilized by methods known in the art. Once cooled, the vessel can be inoculated in sterile operation with a colonized section Petri plate, from other liquid tissue cultures, or from samples of myceliated substrate.

[0093] In one embodiment, the fungal component for inoculation into green coffee beans can be prepared by submerged liquid tissue culture using the undefined medium comprising green coffee bean extract liquid media as defined herein and agitated on a shaker table. In one embodiment, the agitation rate is 50-240 rpm, or 85-95 RPM, and incubated for 4-90 days. In one embodiment, the temperature of incubation is 87-89 °F.

[0094] In one embodiment, the fungal component is trained and/or adapted and/or maintained in its ability to efficiently grow on, metabolize or otherwise utilize and/or modify green coffee beans. In one embodiment, the fungal component is selected and/or trained and/or adapted and/or maintained in its ability to remove or reduce one or more undesirable taste components from the green coffee beans or to remove or reduce the amount of caffeine. Methods to determine whether an undesirable taste component and/or caffeine has been reduced or removed has been disclosed herein and also be found in the art.

[0095] In one embodiment, the trained and/or adapted and/or maintained fungal component is prepared from disinfected wild and healthy fungi. Such fungi with changed, improved, and adapted properties as described herein, relative to the starting strains, either selected or unselected, were developed by these methods. These adapted strains were deposited with the ATCC as described elsewhere herein. In one embodiment, the trained and/or adapted and/or maintained fungal component is prepared from *Ganoderma lucidum*. In one embodiment, the trained and/or adapted and/or maintained fungal component is prepared from *Ganoderma lucidum* strain 806 Alice Chen; Buffalo, NY; 4/94. In another embodiment, the trained and/or adapted and/or maintained fungal component is prepared from *Cordyceps sinensis* (Strain 1009 Caterpillar Fungus; Colorado Corp, 1/2014). In one embodiment, the trained and/or adapted and/or maintained fungal component is prepared from *H. erinaceus*, *T. versicolor*, *L. edodes*, *T. matsutake*, *F. velutipes*, *A. blazei*, *G. frondosa*, *P. nameko*, *L. officinalis*, *M. hortensis*, *M. angusticeps*, *A. auricula*, *T. fuciformis*, *I. obliquus*, *F. fomentarius*, *L. sulfureus*. In one embodiment, the trained and/or adapted and/or maintained fungal component is prepared from a pure strain of *Tuber melanosporum*, obtained by culturing a truffle mushroom by the methods described herein. These fungi having changed, improved, and adapted properties as described herein, relative to the starting strains, were deposited with ATCC as described herein.

[0096] The training and/or adaption and/or maintenance step as described herein can be optionally conducted on undefined medium comprising green coffee bean extract liquid media or solid media as defined herein. In one embodiment, the fungi may be cultivated for 4 – 90 days at any temperature known in the art for cultivating fungi, for example, 87 – 89 °F. Re-inoculation of the cultivated fungal component into fresh media as described herein can be performed at an appropriate time as determined by one of skill in the art depending on the growth rate, growth cycle, and appearance of the fungal component. The cycle of growth and re-inoculation of the fungal component into fresh media, in some embodiments, is performed more than one time, more than two times, more than three times, more than four times, more

than five times, more than ten times, more than fifteen times, more than twenty times, more than twenty five times, more than thirty times, more than forty times, more than fifty times, more than seventy five times, or one hundred times or more. The fungal component by these methods can, for example, better recognize green coffee beans or any particular component of green coffee beans as an energy source, better tolerate the presence of green coffee bean extract in media (as measured by an enhanced growth rate, for example), better remove undesirable taste components, or better remove caffeine. In one embodiment, the undesirable taste component to be removed and/or reduced is chlorogenic acid.

[0097] Therefore, the methods of the invention have as an optional additional step, a method of preparing a trained and/or adapted and/or maintained fungal component comprising a fungal component having an enhanced and/or increased ability to grow on, metabolize or otherwise utilize and/or modify green coffee beans and/or remove one or more undesirable taste components from the green coffee beans, and/or remove caffeine, and use of the trained and/or adapted and/or maintained fungal component in the present invention. The methods of the invention further comprise use of any of the trained, adapted, and/or maintained fungal component(s) as described herein, in the methods of the instant invention.

[0098] PREPARATION OF FUNGAL COMPONENT FOR INOCULATION OF GREEN COFFEE BEANS

[0099] In one embodiment, methods for preparing the fungal component to inoculate the prepared green coffee beans include scaling up a fungal component as defined herein in liquid culture. Such fungal component which is readied for inoculation of the prepared coffee beans is called a "prepared fungal component."

[00100] In one embodiment, the prepared fungal component is in solid culture. In another embodiment, the prepared fungal component is in liquid culture. In another embodiment the prepared fungal component is a mixture of solid and liquid culture. Liquid culture can be accomplished by any means known in the art and includes use of a bioreactor. For example, when using a bioreactor to prepare the fungal component, the bioreactor can be prepared by diluting undefined medium comprising green coffee bean extract liquid media up to 1000x with filtered/RO water. The jacket of the bioreactor may be steamed in one embodiment to sterilize the media, or alternatively, the media can be sterilized by way of injecting steam into the vessel.

[00101] The media to use in preparation of a fungal component for use to inoculate the prepared green coffee beans, may be any suitable media known in the art, or may be made by the methods disclosed herein. The media may further comprise trace elements and organic

substances such as water, nucleic acids, and minerals. The media may be diluted up to 1000x with filtered/RO water. Dilution can be 1x, about 2x, about 3x, about 4x, about 5x, about 6x, about 7x, about 8x, about 9x, about 10x, about 15x, about 20x, about 25x, about 30x, about 35x, about 40x, about 45x, about 50x, about 55x, about 60x, about 65x, about 70x, about 80x, about 90x, about 100x, about 150x, about 200x, about 250x, about 300x, about 350x, about 400x, about 450x, about 500x, about 550x, about 600x, about 650x, about 700x, about 750x, about 800x, about 850x, about 900x, about 950x or about 1000x. In some embodiments, the dilution is about 5x to about 100 x. For a 100 L bioreactor, media can be diluted about 10x, for example.

[00102] In one embodiment, to inoculate the reactor, media may be pumped from another reactor through a sterilized line with an inline pump, or creating positive pressure by sparging air into the reactor with an air compressor that runs the air through inline 0.2/0.5 micron capsule filters then through a check valve with a specific cracking pressure, for example, 2-3 psi.

[00103] Methods to inoculate the bioreactor to prepare the fungal component include inoculating the bioreactor with an excised colonized section of Petri plate and/or a sample of myceliated agriculture using sterile procedure, or by pouring a floating or submerged liquid tissue culture into the bioreactor through the nozzle.

[00104] Optionally, the fungal component may be agitated during culturing by methods known in the art. For example, in a bioreactor, the agitation may be accomplished by a combination of sparged air and a motorized paddle which allows both a turbulent environment and shear mechanical force. The inventors, without limitation, have found that the combination is superior to running either method individually, as sparged air creates the most turbulence at the top half of the culture, while affecting the bottom less, which can be kept agitated by a motorized paddle, while the paddle does not have to run at such a high RPM as normally used in the art. The combination creates the proper small hyphal sphere sizes without damaging the mycelia.

[00105] Liquid state fermentation agitation and swirling techniques are known in the art and include mechanical shearing using magnetic stir bars, stainless steel impellers, injection of sterile high pressure ambient air, injection at high-pressure of sterile media, and or the use of shaker tables. Higher agitation and swirling rates, in conjunction with air and media injections, produce small mycelial spheres.

[00106] The fungal component can be grown until ready for inoculation of the prepared green coffee beans as determined by one of skill in the art. In some embodiments, the fungal

component can be grown for 48 hours prior to use in inoculating the green coffee beans. Determination of whether the fungal cultures comprising the fungal component are suitable for inoculation of the prepared green coffee beans can be determined by one of skill in the art. For example, in one embodiment, the fungal culture, when in liquid media, is suitable for inoculation while in exponential growth phase, either early or late. Senescent cultures and cultures in earlier growth phases with lower amounts of mycelia/mL can be used, but are not preferred. The prepared fungal component optionally appears well grown through in the media, with visible mycelia growing through every mL visible by microscope and unassisted vision.

[00107] In order to effect the most efficient myceliation of the green coffee, the fungal component have defined hyphal sphere sizes which enables hyphae growth in three dimensions around the spherical conglomeration of the culture of the fungal strain. In one embodiment the hyphal sphere size is less than 10 mm in diameter, less than 2 mm in diameter, less than 1 mm in diameter, less than 100 microns in diameter, less 10 microns in diameter, less than 5 microns in diameter, less than 2 microns in diameter, or less than 1 micron in diameter. In another embodiment, the hyphal spherical conglomeration has a size range of 5 microns to 1 millimeter in diameter, or, a size range of 10-50 microns in diameter.

[00108] These methods result in a prepared fungal component for inoculation of prepared green coffee beans.

[00109] INOCULATION AND MYCELIATION OF THE PREPARED GREEN COFFEE BEANS

[00110] The prepared green coffee beans are inoculated with the prepared fungal component. The prepared fungal component to be used can be any fungal component as defined in the instant invention. The inoculation of the prepared fungal component onto the prepared green coffee beans can be carried out by any method known in the art. This step may be variously referred to as the culturing step, the fermentation step, and/or the myceliation step.

[00111] The myceliation may take place in a container as described herein. In one embodiment, the myceliation takes place in a 55 gallon drum as described herein. In this embodiment, the 55 gallon drums has a lid containing two ports, and one port can be used as an inoculation port, while the other can be used to sparge in filtered air to the bottom of the culture, and simultaneously serve as a vent. In some embodiments, the inoculation port is a quick disconnect socket, which is attached to a quick disconnect plug at the end of a harvesting line during inoculation. Optionally, prepared green coffee beans in a plurality of

drums can be myceliated in one cycle via a sterilized manifold connected to bioreactor harvesting line, with the included infrastructure to target any drum in singular or all together at once. In one embodiment, a system for volumetrically consistent dispensing of inoculant per culture is used.

[00112] In one embodiment, the culture may be pneumatically injected into a container comprising the prepared green coffee beans. Moisture may optionally be injected into bags to optimize mycelial growth. In another embodiment the prepared green coffee beans are inoculated by pouring the culture into the container holding the sterilized coffee beans either manually or through a valve built into the fermentor or bioreactor, from any variety of liquid tissue culture.

[00113] In one embodiment, the prepared green coffee beans are cooled to a temperature of between 80-100 ° F prior to inoculation with the prepared fungal component. Cooling may be accomplished by refrigeration or at room temperature. The step of myceliating the prepared green coffee beans can take place for between 4 and 90 days, for between about seven and twenty one days, and in one embodiment, for about seven days, and at any temperature, for example, at 87 – 89 °F. Multiplication of the fungal mycelia by fermentation is carried out by efficiently controlling environmental light, such as by a control model of 40% lighting and 60% dark, and also by controlling sterile airflow and temperature at 86-88 or 87 – 89 ° F, or between 12 and 35 °C, or between 24 °C to 32 °C.

[00114] Relative humidity of this culturing, myceliation and/or fermentation step is between 20% and 80%, in some embodiments, about 60%.

[00115] The step of myceliating the prepared green coffee beans is preferably accomplished in an anaerobic or semi-anaerobic environment. Methods known in the art can be used to induce and/or maintain facultative anaerobic metabolic activity of the prepared fungal component as described by the Pasteur Effect. In an alternate embodiment, the prepared green coffee beans are removed from the sheets and deposited in large stainless steel vats in a sterile environment. The vats regulate oxygen levels and temperature, and enable the facultative anaerobic activity and mycelial growth on the prepared green coffee beans. Facultative anaerobic activity metabolizes more cellulose of per unit of time, meaning that the coffee substrate is consumed at a more rapid rate than in an aerobic environment. In some cases mycelial growth is nine times faster than in an aerobic environment (that is, nine times more cellulose molecules are metabolized to ATP). Another benefit is that the anaerobic environment inhibits fruiting body growth. An anaerobic environment also assures a reduction in unwanted bacterial growth, and other unwanted microbial growth.

[00116] Expansion of the fungus mycelia is monitored by microscopy, and schedules of growth documented by photography.

[00117] The longer the incubation period, the greater the production of the mycelium dry weight and the greater the flavor enhancement of the prepared green coffee. Some strains will form primordial tissue and fruit bodies by 30 days (*Hericium erinaceus* is particularly prone to fruiting while in culture, as is *Ganoderma lucidum* and *Flamulina velutipes*). In some embodiments, harvesting the myceliated coffee beans is performed before the culture fruits body tissue. However, a prolonged period of incubation of a specific fungus does not guarantee a high production of metabolite or the accumulation of active material.

[00118] Determination of when to harvest the myceliated coffee product may be determined by a number of methods. Harvesting is generally performed with a timing to optimize the taste profile of the myceliated coffee product according to the taste profile desired. For example, the scent profile of the myceliation culture can be used by the trained person to determine when the culture is ready. Determination of the appearance of the culture may also be done by the trained person. In some embodiments, harvesting can be done when the amount of the mycelia in the culture are in the approximate amount of 2-3 fully grown (standard size) petri plates (for *G. lucidum*), or when the amount of the mycelia are in the approximate amounts of 10-12 fully grown standard petri plates (for *C. sinensis*), per 8 lbs of coffee. Analytical methods of analysis including high performance liquid chromatography (HPLC) may be employed to carry out measurement of total biomolecules in order to determine the optimum composition and cultivation conditions and the appropriate time(s) for harvesting the fungi.

[00119] In a non-limiting example of the present invention, about 8 lbs of prepared green coffee beans according to the processes of the invention, in a sealed container as described herein, was inoculated with about 25 ml of prepared fungal component as described herein. A lowered oxygen environment was obtained. The myceliation or culturing was allowed to proceed for seven days at 87 – 89 °F, although as little as four days or as long as sixty or more days is also suitable. Harvesting was performed when observers determined that an appropriate taste profile for the myceliated coffee product had been obtained.

[00120] REDUCTION OF CAFFEINE AND/OR UNDESIRABLE TASTE COMPONENTS DURING MYCELIATION

[00121] The step of culturing or myceliation may also cause reduction and/or removal of undesirable taste components as described herein and/or caffeine. In some embodiments, determination of the extent of the removal of at least one undesirable taste component is

determined by the appearance, taste and/or chemical composition of the myceliated coffee product as is known in the art. This determination may be quantitative, e.g., the chemical composition of the myceliated coffee product may be measured by assay methods for one or more of the undesirable taste components by assay methods as known in the art, or determined qualitatively by taste testing by skilled persons.

[00122] In one embodiment, up to 5% of one or more of the undesirable taste components are removed; in other embodiments, up to 10%, up to 15%, up to 20%, up to 25%, up to 30%, up to 35%, up to 40%, up to 45%, up to 50%, up to 55%, up to 60%, up to 65%, up to 70%, up to 75%, up to 80%, up to 85%, up to 90%, or up to 95% of one or more of the undesirable flavor components are removed in the processes of the instant invention. In one embodiment, one or more of the undesirable flavor components are quantitatively removed. The invention also relates to myceliated coffee products having reduced levels of undesirable taste components as described herein.

[00123] In one embodiment, the undesirable taste component is chlorogenic acid and up to 5% of chlorogenic acids are removed; in other embodiments, up to 10%, up to 15%, up to 20%, up to 25%, up to 30%, up to 35%, up to 40%, up to 45%, up to 50%, up to 55%, up to 60%, up to 65%, up to 70%, up to 75%, up to 80%, up to 85%, up to 90%, or up to 95% of chlorogenic acids are removed in the processes of the instant invention. The invention also relates to myceliated coffee products having reduced levels of chlorogenic acid as described herein.

[00124] In one embodiment, caffeine is removed from the prepared coffee beans during the culturing or myceliation step. In one embodiment, up to 5% of caffeine is removed; in other embodiments, up to 10%, up to 15%, up to 20%, up to 25%, up to 30%, up to 35%, up to 40%, up to 45%, up to 50%, up to 55%, up to 60%, up to 65%, up to 70%, up to 75%, up to 80%, up to 85%, up to 90%, or up to 95% of caffeine is removed in the processes of the instant invention. The invention also relates to myceliated coffee products having reduced levels of caffeine as described herein.

[00125] Removal of undesirable taste components may allow for increasing the value of poorer quality coffee and/or rendering it more drinkable. Myceliated coffee products produced by this method may be used to blend with less expensive coffee beans leading to a lower cost product having improved taste properties. The amount of sugar, milk and substitutes thereof to be added to the coffee may be reduced. The instant methods lead to enhanced flavor profile of the myceliated coffee products due to a perception that the

myceliated coffee products provide a richer, smoother, and/or sweeter coffee with less bitter, harsh, and/or acidic tastes.

[00126] ADDITION OF FLAVOR AND/OR HEALTH PROMOTING COMPONENTS

[00127] The culturing or myceliation processes of the instant invention, in some embodiments, provide a myceliated coffee product with added flavor and/or health promoting components. For example, the myceliated coffee products may contain exogenously added anti-tumor and immunomodulatory health promoting components.

[00128] Fungi are metabolically similar to animals but structurally similar to plants in that they possess a rigid cell wall formed largely of long sugar molecule chains joined by somewhat difficult to digest beta (b-) linkages and to a smaller extent more easily digestible alpha (a-) linkages in conjunction with membrane-bound proteins. In contrast, plant cell walls (such as green coffee beans) are made of cellulose polysaccharides whose 1->4 glycosidic glucose linkages are likewise difficult for humans to digest but are digestible by fungi. Fungi cell walls are primarily composed of 1->3 glycosidic linkages, with 1->6 linked side chains, and therefore they may be broken down by minimal processing using water, heat and mechanical treatment into smaller, more easily digestible, immunologically-active polysaccharide molecules of variable microparticulate size called beta glucans, and related glycoprotein compounds. The immune response to beta glucan is dependent upon a- or b-glucan structure, which has primary, secondary, and chiral tertiary structures, explaining the differences in immune response to each fungi's unique a- and or b- glucan profile. Thus, myceliated coffee products have added health promoting components including the molecules described above. Other health promoting components present in the myceliated coffee products may be components that have various properties such as immunomodulating, anti-aging, aphrodisiac, anti-tumour, anti-viral, anti-bacterial, and or anti-fungal properties and include compounds such as a- and b-glucans, glycoproteins, proteins, ergosterols, sterols, triterpenes, and fatty acids, glucomannan, riboglucan, sterpuric acid, mannitol, ribitol, guanosine and adenosine.

[00129] Methyl pyrazines, which develop naturally in cocoa, are among the most important desirable flavor compounds found in cocoa. In the presence of heat, such as roasting, they are produced through Maillard reactions i.e. between sugars and amino acids or peptides. Without being bound by theory, it is believed that methyl pyrazines are also produced through fermentation and/or myceliation, which leads to increased and/or altered methyl

pyrazines in myceliated coffee products and myceliated green coffee beans and other agricultural products.

[00130] *Agaricus blazei* may be used for addition of unique α- and β-linked glucans called glucomannan and riboglucan, which are antiviral, into the myceliated coffee product. Other *A. blazei* polysaccharide extracts may have anti-cancer effects and may be co-therapeutic with other mycelial extract of Fungi listed herein. Methods to optimize biomass and extracellular polysaccharide production have been reported. Therefore, myceliation with *A. blazei* and myceliated coffee products containing flavor and/or health promoting components derived from *A. blazei* as described herein are also included in the instant invention.

[00131] *Cordyceps sinensis* (*C. sinensis*) produces cordycepic acid, adenosine, D-mannitol, and cordycepinadenosine which are immunomodulating and anti-viral. *C. Sinensis* extracts have been shown to be anti-aging and aphrodisiacal. Mycelial sterols isolated from *C. sinensis* have been shown to inhibit the proliferation of numerous cancer cell lines. *C. sinensis* mycelial polysaccharide extracts have been shown to induce hypoglycemia. Therefore, myceliation with *C. sinensis* and myceliated coffee products containing flavor and/or health promoting components derived from *C. sinensis* as described herein are also included in the instant invention.

[00132] *Flammulina velutipes* mycelium has been shown to have a polysaccharide profile that is immunomodulating. *F. velutipes* mycelium composes a unique ergosterol and amino acid profile, sterpuric acid, mannitol, ribitol, and the nucleosides guanosine and adenosine, Enokipodins A-D extracted from *F. velutipes* mycelium are broad spectrum antimicrobial terpenes. The proteins flammulin and velutin exhibit anti-HIV and anti-HPV activity. Therefore, myceliation with *F. velutipes* and myceliated coffee products containing flavor and/or health promoting components derived from *F. velutipes* as described herein are also included in the instant invention.

[00133] *Ganoderma lucidum*'s polysaccharide profile has been shown to be immunomodulating in human cell lines and also in clinical studies. *G. lucidum* mycelial extracts have anti-peroxidative, anti-inflammatory, and anti-mutagenic properties. *G. lucidum* extracts have been shown to be anti-aging and aphrodisiacal. The triterpenoid profile of *G. lucidum* has been determined and shown to be anti-hepatotoxic and hepatoprotective, anti-tumor, anti-angiogenic, anti-hypertensive, hypocholesterolemic, anti-histaminic, and anti-HIV. *G. lucidum*, in addition to producing polysaccharides and glycoproteins, likewise

produce triterpenes, such as ganoderic and lucidenic acids, phenolic compounds, and sterols which also have high biological activity and therapeutic properties and are in themselves anti-oxidant, anti-tumor, anti-bacterial, anti-cancer, anti-inflammatory, anti-histamine, hypotensive, sedative, and meditative after oral consumption. Therefore, myceliation with *G. lucidum* and myceliated coffee products containing flavor and/or health promoting components derived from *G. lucidum* as described herein are also included in the instant invention.

[00134] *Grifola frondosa*'s polysaccharide profile has been shown to be immunomodulating and anti-oxidative, *G. frondosa* produces ergosterols and an anti-oxidative profile of fatty acids. The anti-tumor effects of *G. frondosa* extracts on in vitro cancer cell lines have been investigated, and shows promise for diabetes patients as being hypoglycemic. Therefore, myceliation with *G. frondosa* and myceliated coffee products containing flavor and/or health promoting components derived from *G. frondosa* as described herein are also included in the instant invention.

[00135] *Hericium erinaceus* mycelial and fruiting body extracts have been shown to be anti-mutagenic and immunomodulatory across various cell lines. *H. erinaceus* uniquely produces hericenones in fruit bodies and erinacines in mycelium, structurally determined compounds that can pass the blood-brain barrier and promote secretion of Nerve Growth Factor (NGF) in certain regions of the brain. Erinacenes have been shown to be greater potentiators of NGF expression than hericenones. Therefore, myceliation with *H. erinaceus* and myceliated coffee products containing flavor and/or health promoting components derived from *H. erinaceus* as described herein are also included in the instant invention.

[00136] Aspects of *Lentinula edodes*' polysaccharide profile has been determined and shown to be immunomodulating and antiviral. Lentinan and other metabolites have been studied for their numerous health care benefits. In some countries, lentinan is classified as an "antineoplastic polysaccharide" and is available for clinical use. Addition of lentinan to standard cancer therapies has been shown to result in increased tumor necrosis and with hepatocellular carcinoma and improved quality of life in patients with esophageal carcinoma. Therefore, myceliation with *L. edodes* and myceliated coffee products containing flavor and/or health promoting components derived from *L. edodes* as described herein are also included in the instant invention.

[00137] *Phellinus linteus* extracts have been shown to exhibit antitumor activity. *Polyporus umbellatus* polysaccharide extracts have been studied and shown to be anti-cancer,

immunomodulating, anti-malarial, and hepatoprotective. *Inonotus obliquus* mycelial polysaccharide extract has demonstrated anti-tumor, hypoglycemic, and anti-oxidative properties. *Pleurotus ostreatus* mycelium and fruit body composition have been shown to be very similar, differing only in amino acid content. The mycelial polysaccharide profile consists primarily of laminarin, the extract of which has been shown to be immunomodulating. Lovastatin, isolated from the mycelial broth of *P. ostreatus*, exhibits anti-carcinoma activity, inhibits growth of bacteria and Fungi, and lowers cholesterol. *Trametes versicolor* produces heteroglucans with a-(1-4)- and b-(1-3) glycosidic linkages with fucose in PSK (Krestin) and rhamnose and arabinose in PSP, have been shown to be anti-tumor and immunomodulatory. PSK, an approved drug, is a mycelial extract which exhibits immunomodulating, anti-viral, and cholesterol regulating properties. Mycelial polysachharide extracts of *Tremella fuciformis* have been shown to be therapeutic for various circulatory disorders, to be neurologically healthy, anti-carcinoma, anti-tumor, and anti-aging.

[00138] Therefore, myceliation with *Phellenis linteus*, *Polyporus umbellatus*, *Inonotus obliquus*, *Pleurotus ostreatus*, *Trametes versicolo*, and/or *Tremella fuciformis* (and any other fungal species described herein) and myceliated coffee products containing flavor and/or health promoting components derived from *Phellenis linteus*, *Polyporus umbellatus*, *Inonotus obliquus*, *Pleurotus ostreatus*, *Trametes versicolo*, and/or *Tremella fuciformis* (or any other fungal species described herein) are also included in the instant invention.

[00139] The amounts of flavor components or health promoting components added by the fungal component as described herein can be estimated by one of knowledge in the art, and includes up to 1 ng of the component per unit myceliated coffee product, or up to 5 ng, up to 10 ng, up to 50 ng, up to 100 ng, up to 500 ng, up to 1 µg, up to 5 µg, up to 10 µg, up to 50 µg, up to 100 µg, up to 500 ug, up to 1 mg, up to 2 mg, up to 5 mg, up to 10 mg, up to 20 mg, up to 50 mg, up to 100 mg, or up to 500 mg per unit myceliated coffee product. A unit of myceliated coffee product can be variously defined as 1 g, 1 lb, 1 kg, and the like.

[00140] FURTHER PROCESSING OF MYCELIATED COFFEE PRODUCT

[00141] In some embodiments, once fully myceliated, the myceliated coffee product is optionally rinsed after myceliation. Rinsing may be performed to remove some or all parts of the mycelia and/or other non-green coffee bean matter.

[00142] In some embodiments, one fully myceliated, the myceliated coffee product is optionally dried. Drying can be accomplished by means known in the art for drying green

coffee beans. For example, myceliated coffee product may be spread on a dry surface to dry. In one embodiment, the myceliated coffee product is dried down to about a 11 to 13 % moisture content.

[00143] Optionally, the dried or undried myceliated coffee product can be roasted and/or toasted by conventional methods known in the art. The optional roasting step provides for deactivating the fungus, which may be desirable to reduce the risk of mycoses.

[00144] The myceliated coffee product may optionally be extracted to prepare an extract for use in food and/or drink products. For example, 1000 g of myceliated roasted coffee beans may be fully extracted, with agitation; using 10 to 1000 ml of 121-122°C pressurized water as a buffer, containing 0.01 % to 10 % citric acid and 0.01 to 10 % ascorbic acid. The resulting aqueous extract may be further purified and concentrated using conventional methods.

Myceliated coffee product extracts may be given an extended shelf life by methods known in the art such as formulation in 18 % to 24 % alcohol, 45 % to 60 % glycerol, or addition of 2.5 X volumes of honey or similar sugar such as maple syrup or evaporated cane sugar.

[00145] Removal of undesirable taste components and/or caffeine, and/or addition of myceliation products which increase flavor and/or promote health, will result in myceliated coffee products, and, and may increase the value of poorer quality coffee and/or render it more drinkable. Myceliated coffee products produced by this method may be used to blend with less expensive coffee beans leading to a lower cost product having improved taste properties such as a richer, smoother, and/or sweeter coffee with less bitter, harsh, and/or acidic tastes.

[00146] EMBODIMENTS

[00147] FIG. 1 shows a flow chart of one embodiment of a method of creating an extract of myceliated agricultural product (such as a myceliated coffee product) for human consumption. The step 10 provides an agricultural substrate, the step 12 inoculates the substrate by liquid media comprising an aliquot of culture derived from liquid state fermentation, the step 14 enables mycelium growth and the step 16 grows mycelium on the substrate, the step 18 includes boiling the substrate in water and separating the water-substrate mixture into aqueous and non-aqueous components after mycelium growth on the substrate reaches a desired stage, the step 20 adds a mixture of small molecule components to the aqueous components whereby the aqueous components mix in bio-available combination with the small molecule components to facilitate water solubility of the extract.

[00148] In one embodiment, the step 24 utilizes a culture of Basidiomycota fungi to enable step 12. In an alternate embodiment, the step 26 utilizes and Ascomycota fungi to enable step 12.

[00149] FIG. 2 shows a method of creating an extract of myceliated agricultural product for human consumption to effectuate neuroregeneration and neuro-protection in humans in accordance with the present invention.

[00150] The method includes the step 30 of providing an agricultural substrate, the step 32 of inoculating the substrate by liquid media comprising an aliquot of culture derived from liquid state fermentation, the step 34 of enabling mycelium growth and growing mycelium on the substrate, the step 36 of boiling the substrate in water and separating the water-substrate mixture into an aqueous and nonaqueous components. Step 36 occurs after mycelium growth on the substrate reaches a desired stage. The step 38 adds a mixture of small molecule components to the aqueous components to increase water solubility of the extract to enhance passive absorption and uptake in humans.

[00151] The step 32 of inoculating includes the step 40 of inoculating with a fungal component of the present invention.

[00152] The step 34 of enabling and growing mycelium on the substrate includes regulating temperature and humidity in a sterile environment. The mycelium then grows automatically. The mycelium is grown in a container that allows only a small amount of sterile air to enter the container. Volumes of sterile air are regulated by capping the container. This modulates ambient oxygen in the container.

[00153] The step 34 grows the mycelium to a desired stage, for example, where fruiting bodies begin to appear on the surface of the substrate.

[00154] The step 36 boils the substrate in water to separate the polysaccharides in the mycelium and other beneficial components to suspend in the water, separating aqueous component from the residual substrate solids. The non-aqueous component includes the substrate solids. Separating the aqueous component from the non-aqueous component can be accomplished by filtering or siphoning. It can be appreciated that other methods known in the art can be utilized.

[00155] The step 38 adds a mixture of small molecule components to the aqueous component. This enables the aqueous components to be more bio-available in vivo.

[00156] FIG. 3 shows a method of the present invention 10. The method 10 includes the step 12 of providing a fungal component in a liquid tissue culture medium, the step 14 of adding a

mixture including caffeine, such as the undefined medium containing green coffee bean extract, to the liquid tissue culture medium to train the fungal component to consume caffeine, the step 16 of pasteurizing or sterilizing the green coffee beans, the step 18 of hydrating the green coffee beans to 10-80% moisture content, the step 20 of inoculating the green coffee beans with the fungal component, the step 22 of providing optimal temperature, oxygen content and humidity to enable fungal growth, and the step 24 of drying and roasting the green coffee beans.

[00157] FIG. 4 shows a method 48 of manufacturing coffee in accordance with the present invention. The method 48 includes the step 50 providing coffee beans in container. Preferably, the container is an autoclavable bag.

[00158] The step 22 includes reducing oxygen levels to facilitate facultative anaerobic mycelium growth on the green coffee beans to maximize the consumption rates of caffeine by the mycelium of the fungal component. In the step 50 the container is sealed, allowing only a minimal leakage.

[00159] The step 52 hydrates the green coffee beans to optimal moisture content. In one embodiment the step 52 of hydrating the green coffee beans includes soaking the green coffee beans in water for two hours, and draining the green coffee beans for two hours before sterilization or pasteurization. This is an important step because hydration of the green coffee beans accelerates mycelial growth. The step of hydrating the green coffee beans also removes unwanted residue, and can remove some undesirable water-soluble components from the green coffee beans. In an alternate embodiment, a fluid other than water is used. For example, water infused with an enzyme can eliminate undesirable components from the green coffee beans prior to the step 54 of sterilization. The pH of the water can be regulated to optimize hydration step 52. Distilled water, or mineralized water, can be used for the step 52 of hydration 52.

[00160] The step 54 sterilizes the green coffee beans with steam and the step 56 pressurizes the container. The step 56 of pressurization the container is typically included with the sterilization step 54. Accordingly, the sterilization step 54 is accomplished with pressure ranging from 5-25 lbs per square inch. Preferably the pressure is 10-20 lbs per square inch. In one embodiment, the pressure is 15 lbs per square inch.

[00161] Sterilization under the step 54 may be accomplished under the conditions of 121-122°C at 15 lb/in²; 115-116°C at 10/in²; and 108-109°C at 5 lb/in² for a time period of 20 to 100 minutes, or pasteurization under the conditions of dry heat treatment at 140°C - 210°C for a period of 20 to 100 minutes. These parameters can be varied.

[00162] The step 58 inoculates the bags with a fungal component. In one embodiment, the step 58 is accomplished with a liquid tissue culture having fungal hyphael sphere sizes of 5-100 microns in diameter to assure consistent inoculation. In another embodiment, solid state tissue culture is added to the green coffee beans and mixed with the green coffee beans in a consistent manner. In yet another embodiment both liquid tissue culture and solid state tissue cultures are both added to the green coffee beans in the container to reduce the time required to propagate fungal mycelium on the sterilized coffee beans as described in step 60.

[00163] In one embodiment the step 58 is accomplished by cooling the container to 80-100 °F after the steps 54 and 56 and injecting fungal liquid tissue culture directly into the container. The step 60 propagates the fungal mycelium within the same container. The container may be agitated or spun to assure consistent distribution of the fungal liquid tissue culture. In an alternate embodiment, the green coffee beans are transferred from the containers into a vat for the step 58 of inoculation.

[00164] In another embodiment, the step 58 is accomplished with a liquid tissue culture derived from liquid state fermentation. The liquid tissue culture yields a pure culture of a fungal strain comprising spherical conglomeration with size less than 2 millimeters in diameter, which enables hyphae growth in three dimensions around the spherical conglomeration of the culture of the fungal strain. In a variation of this embodiment, liquid state fermentation yields a pure culture of a fungal strain comprising spherical conglomeration with size ranging from 5 microns to 1 millimeter in diameter due to shearing of hyphae structures during a process of agitation or stirring of liquid media. Agitation and/or swirling processes can include, but not limited to, for example, mechanical shearing using magnetic stir bars, stainless steel impellers, injection of sterile high-pressure ambient air, injection at high-pressure of sterile media, and/or use of shaker tables. Higher agitation and swirling rates, in conjunction with air and media injections, produce small mycelial spheres, aliquots of which are used to inoculate solid-state agricultural substrate(s) for subsequent semi-anaerobic fermentation. Optimally the mycelial spherical conglomerations are less than 100 microns in diameter, and more preferably, within the range of 10-50 microns in diameter.

[00165] The step 58 of inoculating the bags includes using a floating or submerged liquid-state culture, myceliated substrate such as grain or coffee, or petri plate culture inoculate a solid-state cultivation medium comprised of sterilized or pasteurized green coffee beans. Special attention is paid to sub-strains that exhibit enhanced ability to ferment coffee. These strains can be selected and propagated, being cultured on petri-plate and liquid media composed of a certain concentration of green coffee bean extract (as in, four to five green

coffee beans are added to the liquid tissue culture media prep), in a submerged or floating liquid state, and then used to inoculate sterilized green coffee beans at a faster rate than strains and sub-strains that are not conditioned to ferment coffee.

[00166] The step 60 of propagating fungal mycelium on the sterilized coffee beans is preferably accomplished in an anaerobic environment. Accordingly, once substrate (coffee beans) has been sterilized or pasteurized, the subsequent facultative anaerobic metabolic activity of the fungi as described by the Pasteur Effect can be induced after the step 58 of inoculation.

[00167] FIG. 5 shows a method 10 of the present invention. The method 10 includes the step 12 of providing green coffee beans in a container, the step 14 of preparing the green coffee beans for fungal myceliation by removing chlorogenic acids. In one embodiment the step 14 includes rinsing the beans in an aqueous solution.

[00168] The method 10 further includes the step 16 of pasteurizing or sterilizing the green coffee beans, the step 18 hydrates the green coffee beans to 10%-80% moisture content, the step 20 inoculates the green coffee beans with a fungal component, the step 22 regulates temperature, oxygen and humidity in the container to myceliate the green coffee beans. This step 22 optimizes the rate of myceliation. The step 24 rinses the myceliated coffee beans to eliminate surface build-up of mycelial metabolites and other undesired material caused by the myceliation step 22.

[00169] FIG. 6 shows a conveyer 10, a first sheet 12 of autoclavable material, a second sheet 14 of autoclavable material, and a hopper 16 dispensing coffee beans between the sheets 12 and 14. The first sheet 12 extends from a roll 24 of autoclavable material and is laid on the conveyer 10. The hopper 16 releases coffee beans on the first sheet 12. The second sheet 14 is initially rolled in a roll 26 and then laid on the coffee beans and the first sheet 12. The first sheet 12 has edges 22a and 22b. The second sheet 14 has edges 20a and 20b, which seal against the edges 22a and 22b of the first sheet 12 to form a large autoclavable bag, which can be rolled as shown in FIG. 8c and designed to hold over 150 lbs of coffee beans.

[00170] A vacuum may be optionally applied to extract air from between the sheets 24a and 24b, which optimizes heat transfer to the coffee beans 18 when the coffee beans 18 are delivered to a sterilization chamber. The vacuum also and inhibits relative movement between the coffee beans 18. The conveyer 10 delivers the coffee beans 18 into a sterilization chamber.

[00171] FIG. 7 shows the conveyer 10. The hopper 16 delivers coffee beans to an autoclavable bag 30. The autoclavable bag 30 is then sealed and laid on the conveyer 10. The

conveyer 10 Transports numerous autoclavable bags 30, 32, 34, 36, 38 and 40, to an autoclave 42. The autoclave 42 in one embodiment applies heated fluid such as steam, and pressure, to the autoclavable bags 30, 32, 34, 36, 38 and 40 to effectuate sterilization or pasteurization of the coffee beans 18.

[00172] FIG. 8a shows an autoclavable bag 30 filled with coffee beans. The autoclavable bag 30 is under pressure causing a pebbled surface 44. Alternatively, the autoclavable bag 30 is vacuum sealed to cause the pebbled surface 44.

[00173] FIG. 8b shows autoclavable bags 30, 32, 34, 36 and 38 stacked for sterilization or pasteurization in an autoclave. The pebbled surface 44 improves heat transfer to the green coffee beans enclosed in the bag 30. The autoclavable bag 30 includes at least one breather patch 45 adhered to the pebbled surface of the bag 30. Preferably, the breather patch 45 includes vents having a 0.1 micron diameter to enable a minimal amount of air flow through the bag 30 and to assure that mycelium propagated within the bag 30 propagate anaerobically. It can be appreciated that the breather patches can have vents of between 0.05-100 microns in diameter. Preferably the breather patch 45 vents having a 5 micron or less vent diameter. More than one breather patch 45 can be adhered to the pebbled surface of the bag 30, and can be used on containers other than bags in accordance with the present invention.

[00174] FIG. 8c shows a continuous bag 46 of autoclavable material enclosing coffee beans. The continuous bag 46 is sealed to enable the bag 46 to be sterilized or pasteurized in an autoclave. In an alternate embodiment, the bag 46 is sealed in sections so that the continuous bag 46 has multiple sections, which are isolated from each other to inhibit movement of air and coffee between sections. Turbulent airflow during sterilization is shown with arrows passing through the bag 46 and around its surfaces.

[00175] The following examples are provided for illustrative purposes only and are not intended to limit the scope of the invention.

[00176] EXAMPLE 1

[00177] Specific and pure strains of Fungi obtained from referenced collections were manipulated in sterile environments in 1 gal to 10 gal plastic bags, 1 qt to 1 gal glass jar, or on 10 cm to 15 cm petri plates, using undefined, organic fruit and vegetable-based media including green coffee bean extract with 1.5% agar (w/v), in order to monitor and ensure the general vigor and health of strains.

[00178] Mycelial samples were grown in a gentle, ambient sterile airflow for 2 to 4 weeks, then excised from petri plates and subsequently used for inoculation into liquid-state

fermentation employing a similar undefined fruit and vegetable-based media (but with no agar), using ambient air, in 1 qt to 1 gal glass jars. Some samples were grown in agitated and some were grown in unagitated cultures in ambient air in stainless steel tanks designed for commercial beer brewing and/or fermentation.

[00179] The unagitated liquid state fermentation formed a floating mass of hyphae which exhibited continuous growth at interface of liquid and air. The mycelium of agitated and/or swirling cultures grew very quickly as hyphael spheres, which being hydrated, remained submerged, and had the appearance of gelatinous beads in small diameter. Hydrated hyphael spheres collapsed upon desiccation, wherein they were used for inoculating petri-plates for strain propagation and quality control.

[00180] Sphere diameter in liquid-state fermentation was found to be inversely proportional to agitation intensity and volume. Hyphael shear became more efficient at higher agitation and swirling intensity, and once sheared, hyphae formed new spheres of smallest possible diameter, growing in size until they sheared again. When employed in continuous liquid-state fermentation, there existed a constant ratio of sphere diameters, and therefore a constant supply of spheres on the order of microns was produced.

[00181] Thus, this example demonstrated that mycelia sphere diameter was manipulated for more efficient inoculation with inoculation efficiency being inversely proportional to sphere diameter.

[00182] EXAMPLE 2

[00183] Mycelial cultures from unagitated liquid state fermentation (growth period of 2 to 4 weeks) formed a floating mass of hyphae, which were gently blended with a sharp, sterile cutting device prior to being used for inoculation. Gentle blending was achieved by mixing or low homogenization in a commercial blender in short bursts at slow speeds. Aliquots of blended liquid-state culture were used to inoculate sterilized unprocessed fruits and or vegetables, cereal grains, and/or culinary seed, or pasteurized culinary spice, medicinal herbs, natural flavorings, tea mixes, green vanilla beans, green cocoa beans, and green coffee beans.

[00184] EXAMPLE 3

[00185] Substrates for myceliation (containing both substrate and inoculated mycelial culture) in jars or bags were gently mixed every few days until they commanded the substrate and became somewhat resistant to mixing or shaking, usually 2 to 4 weeks depending upon strain. The products were then in a tempeh form. The myceliated green vanilla beans were cooked or baked; the myceliated green cocoa beans were baked or toasted; and the

myceliated green coffee beans were toasted or roasted. Myceliated grain presented in tempeh form, or as an ingredient in food(s) including soups, stir fries, breads, and meat-substitutes, was made safe to eat, and bio-available, by cooking on low to medium heat, 145 ° F to 165 ° F, for 10 min to 60 min, at some point prior to consumption. Other cultures in jars or bags, such as herbs and spices were dried at 100 ° F to 145 ° F for 1 h to 24 h, packaged and used conventionally.

[00186] Myceliated honey formulations were stirred for 10 min to 90 min at 100 ° F to 125 ° F, then poured into small glass bottles. Moreover, myceliated agricultural products were reformulated into value added products such as egg noodles, meat substitutes, specialty flavorings, cooking sauces, soup ingredients and the like.

[00187] EXAMPLE 4

[00188] For a large batch liquid-state and solid-state operation, pure cultures were grown aerobically and inoculated into large industrial liquid-state and large solid-state commercial processors operated continuously and semi-anaerobically for large-scale fermentation of food products. After cultures of media turned completely white or a representative color thereof for a particular species, and had completely overgrown and commanded the medium and were resistant to gentle mixing, the contents were harvested, removed to plastic bags and refrigerated for quick use at either 40 °F, or frozen for long-term storage, and subsequent utilization, at -20 °F. Fermented media were prepared into gourmet human foods including: “tempeh style” meat substitutes, egg-noodles, specialty flavorings, breads, extracts and cooking-sauces, or used directly as a fresh ingredient in soup and/or stir fried recipes, or packaged.

[00189] EXAMPLE 5

[00190] Agricultural substrates completely myceliated by inoculating with pure cultures of fungal strains selected from *A. blazei*, *C. sinensis*, *G. lucidum*, *H. erinaceus*, *G. frondosa*, *P. eryngii*, *P. ostreatus*, *P. citrinopileatus*, *P. djamor*, *T. versicolor*, *L. edodes*, *F. velutipes*, *V. volvacea*, *H. marmoreus*, *P. nameko*, *T. melanosporum*, *M. hortensis*, *P. umbellatus*, and *T. fuciformis* were subjected to heat treatment 1 hour to 24 hours prior to harvest for 1 min to 2 hours at 145 °F to 195 °F followed by recovery at room temperature for 45 min to 48 hours. This process showed remarkable decrease in RNA levels and were formulated into different nutraceutical compositions.

[00191] EXAMPLE 6

[00192] Small Batch Work

[00193] 48 lbs. of coffee was divided into 48 equal portions in clean quart ball jars with lids constructed to enable gaseous diffusion past a collar. These 48, 1 lbs. masses of coffee were soaked with $\frac{3}{4}$ quart of water for two hours. The water in the mixtures was filtered off. The jars of coffee were then subjected to 90 minutes of sterilization temperatures at 15 psi, and placed in a sterile laminar air flow to cool for 8 hours. Once cool, the prepared green coffee beans were inoculated with half to whole colonies of fungus selected from one of the following: *Ganoderma lucidum*, *Cordyceps sinensis*, *Tuber melanosporum*, *Hericium erinaceus*, *Agaricus blazei*, *Grifola frondosa*, *Pleurotus ostreatus*, *Trametes versicolor*, *Laetiporus sulphureus*, *Flammulina velutipes*, *Lentinula edodes*, *Morchella angusticeps*, *Morchella crassipes*, *Morchella hesculenta*, *Tremella fuciformis*, and *Inonotus obliquus*, doing three of each, growing on an undefined vegetable and fruit juice agar media containing green coffee extract as described in Example 8, with sterile tools and in sterile operation inside the laminar flow hood. The cultures myceliated for 7 to 21 days, with samples of each being pulled out for drying and roasting at the 7th, 14th, and 21st days. The smell of the culture and taste of the myceliated green coffee beans at the 7th day indicated that the cultures were complete, though longer myceliation periods yielded greater cell mass.

[00194] Large Batch Work

[00195] 528 lbs. of green coffee beans were soaked in two different procedures. In the first procedure, the beans were soaked three times, for 20 minutes each soak, in the second procedure, the beans were soaked for 20 minutes through a constant stream of filtered water. The beans were then packed into polypropylene bags with 0.2 micron breather patches, with the tops of the bags folded over with rubber bands wrapped around the sides of the bags, such that steam and gas diffusion could occur through breather patch and through the folded sides of the bags. The bags were sterilized under a liquid cycle at 22 psi for 80 minutes, and then allowed to cool for 8 hours. The bags were inoculated with fungi from the following species: *Ganoderma lucidum*, *Cordyceps sinensis*, *Tuber melanosporum*, and *Morchella angusticeps*. The *Ganoderma lucidum* culture was grown in a bioreactor, with 10 L of organic potato extract, 2 L of green coffee extract, and 1 L organic mango juice diluted to 100 liters with RO water. The bioreactor was sparged with compressed air filtered through two inline 0.2 micron hydrophobic capsule filters, and the reactor was kept under 2-3 psi through the use of check valves on the air supply and venting lines with 2-3 psi cracking pressure ratings. The inoculant was readily grown in 48 hours, and was harvested through a diaphragm valve located at the bottom of the reactor, which led to a harvesting line that had teed and valved off access to a steam line and steam trap, with an inline check valve, through six feet of

flexible stainless steel hosing, to a solenoid valve connected to a timer and foot switch, followed by a flow metering valve to an elbowed sanitary fitting. While being steamed, the elbowed sanitary fitting was connected to a ball valve that connected to the steam exhaust manifold. The ball valve was closed after steaming the line, and the ball valve was detached from the harvesting line once entered into a laminar flow hood, so as to keep the whole line sterile. The *Cordyceps sinensis*, *Tuber melanosporum*, and *Morchella angusticeps* cultures were grown in 4 L flasks, in 1.5 L of the same media used in the bioreactor pre-dilution. These cultures were grown for six days, and were used to inoculate the bags of sterilized green coffee beans. The beans were myceliated for 7 days, where their smell conferred the desired taste profile of the beverage made from the roasted myceliated beans, whereupon they were dried on the 8th day to a 13% moisture content.

[00196] EXAMPLE 7

[00197] A suitable fungi for use in the methods of the present invention was prepared by the following methods. The following *G. lucidum* strains were purchased commercially from the Pennsylvania State University mushroom culture collection: 496 Ling ZHI; Singapore commercial line; 7/85; 502 IFO #8436; IFO-Japan; 7/30/85; 510 Red oak, State College, PA; D.J. Royse; 9/85; 549 Y.H. Park, ASI-Korea; 12/5/85; 550 Y.H. Park, ASI-Korea; 12/5/85; 551 Y.H. Park, ASI-Korea; 12/5/85; 580 Y.H. Park, ASI-Korea; 2/10/85; 607 Y.H. Park, ASI-Korea; 2/19/85; 617 Y.H. Park, ASI-Korea; 2/25/85; 618 Y.H. Park, ASI-Korea; 2/25/85; 619 Y.H. Park, ASI-Korea; 2/25/85; 620 Y.H. Park, ASI-Korea; 2/25/85; 621 Y.H. Park, ASI-Korea; 2/25/85; 622 Y.H. Park, ASI-Korea; 2/25/85; 623 Y.H. Park, ASI-Korea; 2/25/85; 624 Y.H. Park, ASI-Korea; 2/25/85; 625 Y.H. Park, ASI-Korea; 2/25/85; 626 Y.H. Park, ASI-Korea; 2/25/85; 627 Y.H. Park, ASI-Korea; 2/25/85; 665 Quimio; Philippines; 3/6/86; 669 Y.H. Park, ASI-Korea; 3/25/86; 686 B. W. Yoo; 4/28/86; 724 T. Mitchel, Lawn PSU Forestry Bldg. 9/16/90; 806 Alice Chen; Buffalo, NY; 4/94; 807 Alice Chen; North Carolina; 4/94; 841 White Oak; PSU Campus; J. Peplinski; 8/99. The above strains were cultured using the media described herein comprising green coffee bean extract (see Example (9)). Many strains were unable to grow and/or died on the media. Surprisingly, the inventors found that *G. lucidum* strain 806 Alice Chen; Buffalo, NY was able to grow on the media comprising green coffee bean extract and was selected for further use in accordance with the instant invention.

[00198] EXAMPLE 8

[00199] Fungi (including *G. lucidum* strain 806, *C. sinensis*, and *T. melanosporum* as described herein, also *H. erinaceus*, *T. versicolor*, *L. edodes*, *T. matsutake*, *F. velutipes*, *A.*

blazei, *G. frondosa*, *P. nameko*, *L. officinalis*, *M. hortensis*, *M. angusticeps*, *A. auricula*, *T. fuciformis*, *I. obliquus*, *F. fomentarius*, *L. sulfureus*) were maintained on a culture comprising an undefined media including extract of green coffee beans. Experiments showed that use of the media including extract of green coffee beans to culture the maintained the fungi's ability to tolerate, grow on, metabolize, remove or reduce caffeine or undesirable flavor components. It was also found that successive propagations of fungi as defined above caused enhancement and/or improvement of the fungi's ability to tolerate, grow on, metabolize, remove or reduce caffeine or decrease undesirable flavor components, resulting in training or adapting the fungi to undefined media including extract of green coffee beans. Such fungi with changed, improved, and adapted properties as described herein, relative to the starting strains, either selected or unselected, were developed. These adapted strains were deposited with the ATCC as described elsewhere herein.

[00200] The undefined media including extract of green coffee beans was made as follows: 2 lbs green coffee beans, pulverized was mixed with ¼ gallon water at room temperature. The mixture was allowed to extract for 20 minutes with shaking, then filtered three times through fine mesh. Separately, about 5 organic potatoes were placed in 10 L of water and autoclaved 20 minutes to soften the potatoes. The potatoes were then pulverized with a potato masher, and then filtered through fine mesh three times. 1 L of commercial unsweetened fruit juice was be added. These solutions are combined and autoclaved. This recipe was also scaled up or down as required.

[00201] The washed green coffee beans were soaked in water and the moisture content was raised to about 30%. At other times the moisture content was raised to about 60%. At this point, the bean as well quenched of chlorogenic acid, as evidenced by the lack of green seen in the bean. Some chlorogenic acid was left in the bean, though much of it is obviously and evidently removed. Removal of chlorogenic acid from green coffee beans allowed good myceliation at moisture contents of 30% and greater, whereas green coffee beans that did not have a chlorogenic acid removal step required a moisture content of 60% for good myceliation.

[00202] Liquid culture: The culture comprising fungi for use in inoculating the prepared green coffee beans was agitated with sparged air and a motorized paddle to create turbulent environment and to shear hyphae with pure mechanical force. The dual agitation method was superior to either method individually, since sparged air created the most turbulence at the top half of the culture, while affecting the bottom less, which was agitated by a motorized paddle. In return the paddle could be run at a lower RPM and still obtain the hyphal sphere

size obtained by a faster RPM in the absence of sparging. The hyphal size was about 2-5 micron in diameter). Undamaged mycelium and proper morphology in the prepared fungi were prepared by this method and used for culturing and/or myceliation.

[00203] EXAMPLE 9

[00204] Black Italian Winter Perigord Truffles [Truffle 1 (T1), and Truffle 2 (T2)], were aseptically manipulated in the mycology laboratory environment to obtain a number of new truffle cultivars for commercial work, including opposite mating type isolates, by the following 4 methods.

[00205] Method 1: fresh (less than one week old) intact whole pieces of T1 (truffle gleba chunks of approximately 10mm by 20mm in size) were placed into oak-agar (a general undefined agar media based on oak leaves and oak twigs), in 50 petri plates, which were sealed with parafilm to allow cultures to incubate at room temperature (23 ° C) for 5 weeks. After 5 weeks, pieces of T1 were seen to regenerate into mycelia described by the following cultivars: (a) vigorous white mycelium having purple hues, on 6 petri plates, (b) vigorous golden mycelium with purple hues, on 3 plates, (c) vigorous white and gold mycelium, on 6 plates, (d) non-vigorous wispy purple and gold mycelium on 2 plates, and (e) non-vigorous wispy white and gold mycelium on 2 plates. The remainder of the petri plates (37 plates) were discarded after becoming overgrown by (bacterial and fungal) contamination. Isolates (a), (b), and (c) were determined to have commercial potential. The (a) vigorous white isolate is the mating type mycelium maternal to the original truffle mushroom, while isolate (b) is the paternal mating type mycelium to the original truffle mushroom, and (c) is a cultivar derived from non-sexual genetic recombination or anastomosis of both the (a) and the (b) mating types. After 5 to 6 weeks growing on oak-agar, cultures of the (c) vigorous white and gold mycelium begin to produce tiny (0.1mm to 2.0 mm) truffle fruits. After the structures were cultured to fresh petri plates, and onto similar agricultural media, these cultures produced ever-growing black truffle mushrooms.

[00206] Method 2: fresh (less than one week old) intact spores of T1 (0.01 g to 0.1 g clumps of spores of a mass approximately 0.1mm by 0.1mm in size) were placed into seasoned oak-agar medium in petri plates (40 plates), which were sealed with parafilm for 5 weeks to allow spores to sprout at room temperature (23 degrees C). After 5 weeks, spores were seen to sprout into mycelia on 10 plates; however, all mycelium that sprouted from the spores, though viable, was extremely wispy and grew slowly.

[00207] Method 3: fresh (less than one week old) intact whole pieces of T1 (40 g of gleba chunks of approximately 10mm by 20mm in size) were placed into 2 bottles each containing

400ml sterile water and 50 ug/ml of each the following antibiotics: Ampicillin, Penicillin, Chloramphenicol, Erythromycin, Neomycin, Streptomycin, with closures on but not tight, where they were allowed to incubate undisturbed for 5 weeks at room temperature (23 ° C) with an occasional gentle shaking. After 5 weeks, intact pieces were removed from antibiotic buffer and cast into general undefined agar media (containing the same levels of antibiotics), based on Oak leaves and twigs, into 50 petri plates which were then sealed with parafilm to allow cultures to incubate for 2 weeks at room temperature (23 °C). After 2 weeks of regeneration in agar, pieces of T1 regenerated into the following mycelial cultivars: (a) white mycelium on 12 petri plates, (b) golden mycelium on 4 plates, (c) white and gold mycelium on 2 plates. The remainder of the petri plates (32 plates) were discarded after showing no growth or after becoming overgrown by fungal contamination. The isolates obtained via Method 3, though very similar to the isolates from Method 1, were much less vigorous.

[00208] The isolates (a), (b) and (c) from Method 1 were prepared as described herein and used for myceliation of substrates. Substrates included prepared green coffee beans. The myceliation of the prepared green coffee was allowed to proceed until the desired coffee flavor for the resultant roasted coffee was achieved, about 14 days. The resultant myceliated green coffee beans were roasted, brewed, cupped, and tasted by trained tasters. It was found that when the isolate described for method 1, isolate (a) was used, a pleasant flavorful aroma/taste in the myceliated coffee, reminiscent of flowers was achieved. When the isolate described for method 1, isolate (c) was used, a pleasant flavorful aroma/taste, reminiscent of truffles, was achieved.

[00209] EXAMPLE 10

[00210] The myceliated coffee products including myceliated green coffee beans and grounds produced by the methods of the instant invention contained added polysaccharides and beta glucans. An analysis showed that Robusta coffee grounds produced by the methods of the invention had 30.54 mg dextran per gram of coffee grounds. This result provided the total polysaccharide amount in the substrate through a spectrophotometric method based on a modified phenol-sulfuric acid approach. The analysis also showed that Robusta coffee grounds produced by the methods of the invention had beta glucans at 0.432%, as measured by the MYBG method utilizing strong hydrolysis conditions to hydrolyze beta glucan with quantification by spectrophotometric method. This represents an advantage over consuming beta-glucans from Reishi mushrooms, as these mushrooms are a non-culinary mushroom for reasons of bitterness, woodiness, and hardness, or in pill form.

[00211] EXAMPLE 11

[00212] Taste Comparison 1, Sumatran, Peruvian, and Honduran Arabica beans

[00213] A coffee professional and owner of a coffee roasting business owner (taster) and a trained employee taste-tested, in a double-blind trial, a comparison of standard premium coffee beans using Sumatran, Peruvian, and Honduran Arabica beans (control beans) with coffee beans produced by the methods of the present invention (myceliated beans). Both myceliated beans and control beans were roasted by on the day of the trial. These were cupped side-by-side with the control, using standard coffee tasting techniques.

[00214] The flavor-enhancing effects of myceliation were confirmed. The tasters sampled myceliated and normal brews of each variety in this blind taste test. Notes were taken. At the conclusion of the tasting, the coffee beans used for each cup were identified.

[00215] Commenting first on the myceliated Sumatra it was described as having a fuller body, more complex and less bitter flavor than the control Sumatra. The tasters stated that this was the only process they were aware of that actually removed a taste defect and actually enhanced the flavor.

[00216] The myceliated Peruvian showed a noticeable flavor-enhancement as well, being a less bitter, more sweet, and a markedly “brighter” cup when myceliated. Despite being a high-quality bean, the control Peruvian tasted “flat” by comparison.

[00217] Out of the two tasters, one taster was able to taste a difference in the Honduran brew. The methods of the invention resulted in removing the bitter compounds found in coffee resulting in a better tasting cup of coffee.

[00218] Taste Comparison 2 Arabica beans

[00219] A myceliated coffee taste test was held at a coffee house. The barista/roaster (taster) delivered the formal cupping of their in-house Sulawesi Arabica coffee (Indonesian in origin). The beans were selected from inventory and both the myceliated and control beans (Arabica) were roasted the day of the cupping. The results showed that myceliated coffee (produced by the methods of the invention) had an improved flavor profile.

[00220] The taster described the myceliated coffee as less acidic, sweeter, fuller in body, more complex and overall a better taste than the original bean. Several other participants in the taste test and also noticed the flavor-enhancement found in myceliated coffee.

[00221] Taste Comparison 3 Robusta beans

[00222] A coffee professional and owner of a coffee roasting business owner (taster) and a trained employee taste-tested, in a double-blind trial, a comparison of control Robusta coffee beans with Robusta coffee beans produced by the methods of the present invention. Coffee made from 100% Robusta beans is generally considered undrinkable due to the high acidity

and bitterness of Robusta. Thus, Robusta coffee beans are not typically used alone, instead, Robusta beans are typically pre-processed (e.g. steamed), then blended with more expensive beans to make it palatable.

[00223] This lower quality bean is considered undrinkable by some due to its bitterness, but Robusta is used commercially in coffee blends to bring the price down. Robusta is growing in its use and the Robusta segment of the coffee market has grown from 39% of the market in 2008 to 41% of the market in 2013.

[00224] The tasters agreed that myceliated Robusta is “hands-down” a better cup of coffee than non-myceliated. One taster commented that “you have proved beyond a doubt that your technology works”; the other taster commented that he is a self-proclaimed coffee snob and “I would drink this myceliated Robusta on a daily basis”, simultaneously remarking that the non-myceliated coffee was unpalatable. More specifically, he noticed a conspicuous lack of bitterness and acidity in the cup, with a fuller body in the taste. The tasters remarked that a non-professional coffee taster would be able to taste and appreciate the difference, and that myceliated Robusta holds great value in the marketplace. Other employees of the roasting company also noticed the difference.

[00225] The taste testing results clearly demonstrate that the instant processes enhance the taste of coffee. The results showed that the processes of the invention removed taste defects like bitterness from both Arabica and Robusta coffee beans and enhanced their flavor and value. The processes result in a sweeter, fuller bodied and more complex tasting coffee with either Robusta or Arabica beans.

CLAIMS

1. A method for preparation of a myceliated coffee product, comprising:
 - a) providing prepared green coffee beans comprising the steps of:
 - i. providing green coffee beans;
 - ii. sterilizing the green coffee beans to provide prepared green coffee beans;
 - b) providing a prepared fungal component consisting of a fungus selected from the group consisting of *Ganoderma lucidum*, *Cordyceps sinensis*, *Trametes versicolor*, *Volvariella volvacea* and *Grifola frondosa* in liquid tissue culture as a submerged fungal liquid tissue culture;
 - c) inoculating the prepared green coffee beans with the prepared fungal component;and
 - d) culturing the prepared green coffee beans and prepared fungal component to prepare the myceliated coffee product, wherein the myceliated coffee beans are capable of being used to prepare a palatable coffee beverage having reduced bitterness for human consumption.
2. The method of claim 1, further comprises reducing the amount of chlorogenic acid.
3. The method of claim 1, wherein the step of providing prepared green coffee beans further comprises hydrating the green coffee beans.
4. The method of claim 3, wherein the green coffee beans are hydrated to about a 60% moisture level or to about a 30% moisture level.
5. The method of claim 1, wherein the prepared fungal component comprises the fungal species *G. lucidum*.
6. The method of claim 5, wherein the prepared fungal component comprises the fungal species *G. lucidum* strain 806.

7. The method of claim 1, wherein the prepared fungal component comprises the fungal species *C. sinensis*.
8. The method of claim 1, wherein the culturing step is carried out for about 7 days.
9. The method of claim 1, wherein the green coffee beans are from *Coffea arabica*.
10. The method of claim 1, wherein the green coffee beans are from *Coffea robusta*.