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CHEMICAL & ENGINEERING NEWS, vol. 57, no. 1, January 1, 1979, J.L. FOX "Antibody reagents revolutionizing immunology", pages 15-17
TRANSPLANTATION PROCEEDINGS, vol. 12, no. 3, Suppl. 1, September 1980, P.C. KUNG et al. "Strategies for generating monoclonal antibodies defining human T-lymphocyte differentiation antigens", pages 141-146

Description

Field of the invention

This invention relates generally to new hybrid cell lines and more specifically to hybrid cell lines for production of monoclonal antibody to an antigen found on approximately 95% of normal human thymocytes, to the antibody so produced, to therapeutic and diagnostic compositions and to diagnostic methods employing this antibody.

Description of the prior art

The fusion of mouse myeloma cells to spleen cells from immunized mice by Kohler and Milstein in 1975 [*Nature* 256, 495—497 (1975)] demonstrated for the first time that it was possible to obtain a continuous cell line making homogeneous (so-called "monoclonal") antibody. Since this seminal work, much effort has been directed to the production of various hybrid cells (called "hybridomas") and to the use of the antibody made by these hybridomas for various scientific investigations. See, for example, *Current Topics in Microbiology and Immunology*, Volume 81—"Lymphocyte Hybridomas", F. Melchers, M. Potter, and N. Warner, Editors, Springer-Verlag, 1978, and references contained therein; C. J. Barnstable, et al., *Cell*, 14, 9—20 (May, 1978); P. Parham and W. F. Bodmer, *Nature* 276, 397—399 (November, 1978); *Handbook of Experimental Immunology*, Third Edition, Volume 2, D. M. Wier, Editor, Blackwell, 1978, Chapter 25; and *Chemical and Engineering News*, January 1, 1979, 15—17. These references simultaneously indicate the rewards and complications of attempting to produce monoclonal antibody from hybridomas. While the general technique is well understood conceptually, there are many difficulties met and variations required for each specific case. In fact, there is no assurance, prior to attempting to prepare a given hybridoma, that the desired hybridoma will be obtained, that it will produce antibody if obtained, or that the antibody so produced will have the desired specificity. The degree of success is influenced principally by the type of antigen employed and the selection technique used for isolating the desired hybridoma.

The attempted production of monoclonal antibody to human lymphocyte cell-surface antigens has been reported only in a few instances. See, for example, *Current Topics in Microbiology and Immunology*, *ibid*, 66—69 and 164—169. The antigens used in these reported experiments were cultured human lymphoblastoid leukemia and human chronic lymphocytic leukemia cell lines. Many hybridomas obtained appeared to produce antibody to various antigens on all human cells. None of the hybridomas produced antibody against a predefined class of human lymphocytes.

More recently, the present applicants and others have authored articles disclosing the preparation and testing of hybridomas making antibody to certain T-cell antigens. See, for

example, Reinherz, E. L., et al., *J. Immunol.* 123, 1312—1317 (1979); Reinherz, E. L., et al., *Proc. Natl. Acad. Sci.*, 76, 4061—4065 (1979); and Kung, P. C., et al., *Science*, 206, 347—349 (1979).

It should be understood that there are two principal classes of lymphocytes involved in the immune system of humans and animals. The first of these (the thymus-derived cell or T cell) is differentiated in the thymus from haemopoietic stem cells. While within the thymus, the differentiating cells are termed "thymocytes." The mature T cells emerge from the thymus and circulate between the tissues, lymphatics, and the bloodstream. These T cells form a large proportion of the pool of recirculating small lymphocytes. They have immunological specificity and are directly involved in cell-mediated immune responses (such as graft rejection) as effector cells. Although T cells do not secrete humoral antibodies, they are sometimes required for the secretion of these antibodies by the second class of lymphocytes discussed below. Some types of T cells play a regulating function in other aspects of the immune system. The mechanism of this process of cell cooperation is not yet completely understood.

The second class of lymphocytes (the bone marrow-derived cells or B cells) are those which secrete antibody. They also develop from haemopoietic stem cells, but their differentiation is not determined by the thymus. In birds, they are differentiated in an organ analogous to the thymus, called the Bursa of Fabricius. In mammals, however, no equivalent organ has been discovered, and it is thought that these B cells differentiate within the bone marrow.

It is now recognized that T cells are divided into at least several subtypes, termed "helper", "suppressor", and "killer" T cells, which have the function of (respectively) promoting a reaction, suppressing a reaction, or killing (lysing) foreign cells. These subclasses are well understood for murine systems, but they have only recently been described for human systems. See, for example, R. L. Evans, et al., *Journal of Experimental Medicine*, Volume 145, 221—232, 1977; and L. Chess and S. F. Schlossman—"Functional Analysis of Distinct Human T-Cell Subsets Bearing Unique Differentiation Antigens", in *Contemporary Topics in Immunobiology*, O. Stutman, Editor, Plenum Press, 1977, Volume 7, 363—379.

The ability to identify or suppress classes or subclasses of T cells is important for diagnosis or treatment of various immunoregulatory disorders or conditions.

For example, certain leukemias and lymphomas have differing prognosis depending on whether they are of B cell or T cell origin. Thus, evaluation of the disease prognosis depends upon distinguishing between these two classes of lymphocytes. See, for example, A. C. Aisenberg and J. C. Long, *The American Journal of Medicine*, 58:300 (March, 1975); D. Belpomme, et al., in *Immunological Diagnosis of Leukemias and Lymphomas*, S. Thierfelder, et al., eds,

Springer, Heidelberg, 1977, 33—45; and D. Bel-
pomme, *et al.*, *British Journal of Haematology*,
1978, 38, 85.

Certain disease states (e.g., juvenile rheu-
matoid arthritis, malignancies, and agammaglo-
bulinemia) are associated with an imbalance of T
cell subclasses. It has been suggested that auto-
immune diseases generally are associated with
an excess of "helper" T cells or a deficiency of
certain "suppressor" T cells, while agammaglo-
bulinemia is associated with an excess of certain
"suppressor" T cells or a deficiency of "helper" T
cells. Malignancies generally are associated with
an excess of "suppressor" T cells.

In certain leukemias, excess T cells are
produced in an arrested stage of development.
Diagnosis may thus depend on the ability to
detect this imbalance or excess and to determine
which developmental stage is in excess. See, for
example, J. Kersey, *et al.*, "Surface Markers
Define Human Lymphoid Malignancies with
Differing Prognoses" in *Haematology and Blood
Transfusion*, Volume 20, Springer-Verlag, 1977,
17—24, and references contained therein; and E.
L. Reinherz, *et al.*, *J. Clin. Invest.*, 64, 392—397
(1979).

Acquired agammaglobulinemia, a disease state
in which no immunoglobulin is produced, com-
prises at least two distinct types. In type I the
failure to produce immunoglobulin is due to an
excess of suppressor T cells, while in type II it is
due to a lack of helper T cells. In both types, there
appears to be no defect or lack in the patients' B
cells, the lymphocytes which are responsible for
the actual secretion of the antibody; however,
these B cells are being either suppressed or "not
helped", resulting in greatly decreased or absent
immunoglobulin production. The type of acquired
agammaglobulinemia may thus be determined
by testing for an excess of suppressor T cells or
an absence of helper T cells.

On the therapeutic side, there is some sugges-
tion, as yet not definitely proven, that adminis-
tration of antibodies against the subtype of T cell
in excess may have therapeutic benefit in auto-
immune disease or malignancies. For example, a
helper T cell cancer (certain cutaneous T cell
lymphomas and certain T cell acute lympho-
blastic leukemias) may be treated by an antibody
to a helper T cell antigen. Treatment of auto-
immune disease caused by an excess of helper
cells may also be accomplished in the same
fashion. Treatment of diseases (e.g., malig-
nancies or type I acquired agammaglobulinemia)
due to an excess of suppressor T cells may be
treated by administration of an antibody to a
suppressor T cell antigen.

Antisera against the entire class of human T
cells (so-called antihuman thymocyte globulin or
ATG) has been reported useful therapeutically in
patients receiving organ transplants. Since the
cell-mediated immune response (the mechanism
whereby transplants are rejected) depends upon
T cells, administration of antibody to T cells
prevents or retards this rejection process. See, for

example, Cosimi, *et al.*, "Randomized Clinical
Trial of ATG in Cadaver Renal Allograft Reci-
pients: Importance of T Cell Monitoring", *Surgery*
40:155—163 (1976) and references contained
therein.

The identification and suppression of human T
cell classes and subclasses has previously been
accomplished by the use of spontaneous auto-
antibodies or selective antisera for human T cells
obtained by immunizing animals with human T
cells, bleeding the animals to obtain serum, and
adsorbing the antiserum with (for example) auto-
logous but not allogeneic B cells to remove
antibodies with unwanted reactivities. The
preparation of these antisera is extremely diffi-
cult, particularly in the adsorption and purification
steps. Even the adsorbed and purified antisera
contain many impurities in addition to the desired
antibody, for several reasons. First, the serum
contains millions of antibody molecules even
before the T cell immunization. Second, the
immunization causes production of antibodies
against a variety of antigens found on all human T
cells injected. There is no selective production of
antibody against a single antigen. Third, the titer
of specific antibody obtained by such methods is
usually quite low, (e.g., inactive at dilutions
greater than 1:100) and the ratio of specific to
non-specific antibody is less than 1/10⁶.

See, for example, the Chess and Schlossman
article referred to above (at pages 365 and
following) and the Chemical and Engineering
News article referred to above, where the defi-
ciencies of prior art antisera and the advantages
of monoclonal antibody are described.

Summary of invention

There has now been discovered a novel
hybridoma (designated OKT10) which is capable
of producing novel monoclonal antibody against
an antigen found on approximately 95% of
normal human thymocytes, 5% of normal human
peripheral T cells, 10% of E⁻ peripheral mono-
nuclear cells (B cells and null cells) and 10—20%
of bone marrow cells.

The antibody so produced is monospecific for a
single determinant on approximately 95% of
normal human thymocytes and contains essen-
tially no other anti-human immunoglobulin, in
contrast to prior art antisera (which are inherently
contaminated with antibody reactive to numerous
human antigens) and to prior art monoclonal
antibodies (which are not monospecific for a
human thymocyte antigen). Moreover, this
hybridoma can be cultured to produce antibody
without the necessity of immunizing and killing
animals, followed by the tedious adsorption and
purification steps necessary to obtain even the
impure antisera of the prior art.

It is accordingly one object of this invention to
provide hybridomas which produce antibodies
against an antigen found on about 95% of normal
human thymocytes.

It is a further aspect of the present invention to
provide methods for preparing these hybridomas.

A further object of the invention is to provide essentially homogeneous antibody against an antigen found on about 95% of normal human thymocytes.

A still further object is to provide compositions for treatment or diagnosis of disease or for identification of T cell or thymocyte subclasses employing this antibody.

Other objects and advantages of the invention will become apparent from the examination of the present disclosure.

In satisfaction of the foregoing objects and advantages, there is provided by this invention a novel hybridoma producing novel antibody to an antigen found on approximately 95% of normal human thymocytes, the antibody itself, and diagnostic and therapeutic compositions employing the antibody. The hybridoma was prepared generally following the method of Milstein and Kohler. Following immunization of mice with leukemic cells from humans with T-cell acute lymphoblastic leukemia (T-ALL), the spleen cells of the immunized mice were fused with cells from a mouse myeloma line and the resultant hybridomas screened for those with supernatants containing antibody which gave selective binding to normal E rosette positive human T cells and/or thymocytes. The desired hybridomas were subsequently cloned and characterized. As a result, a hybridoma was obtained which produces antibody (designated OKT10) against an antigen on approximately 95% of normal human thymocytes. Not only does this antibody react with about 95% of normal human thymocytes, but it also reacts with about 5% of normal human peripheral T cells, 10% of E⁻ peripheral mononuclear cells (B cells and null cells), and 10—20% of bone marrow cells.

In view of the difficulties indicated in the prior art and the lack of success reported using malignant cell lines as the antigen, it was surprising that the present method provided the desired hybridoma. It should be emphasized that the unpredictable nature of hybrid cell preparation does not allow one to extrapolate from one antigen or cell system to another. In fact, the present applicants have discovered that the use of a T cell malignant cell line or purified antigens separated from the cell surface as the antigen were generally unsuccessful.

Both the subject hybridoma and the antibody produced thereby are identified herein by the designation "OKT10", the particular material referred to being apparent from the context. The subject hybridoma was deposited on November 21, 1979 at the American Type Culture Collection, 12301 Parklawn Drive, Rockville, Maryland 20852, and was given the ATCC accession number CRL 8022.

According to the present invention there is provided mouse monoclonal antibody which reacts with

- i) approximately 95% of normal human thymocytes,

- ii) 5% of normal human peripheral T cells.
- iii) 10% of E⁻ peripheral mononuclear cells, and
- iv) 10—20% of bone marrow cells.

Preferably the antibody exhibits the pattern of reactivity to normal human thymocytes shown in Figure 1, and may also exhibit the pattern of reactivity to peripheral T cells and thymocytes shown in Table 2.

The antibody preferably is reactive with an antigen on leukemic T-ALL cells.

Advantageously the antibody is produced by a hybridoma formed by fusion of cells from a mouse myeloma line and spleen cells from a mouse previously immunized with human leukemic T-ALL cells.

Preferably the antibody is produced by hybridoma ATCC CRL 8022 (OKT 10).

The present invention also provides a hybridoma for producing antibody according to the invention, formed by fusion of spleen cells from a mouse previously immunized with human leukemic T-ALL cells and cells from a mouse myeloma line.

Preferably, the hybridoma is ATCC CRL 8022 (OKT 10).

According to a further aspect of the present invention, there is provided a method for preparing an antibody of the invention, which comprises the steps of:

- i) immunizing mice with human leukemic T-ALL cells;
- ii) removing the spleens from said mice and making a suspension of spleen cells;
- iii) fusing said spleen cells with mouse myeloma cells in the presence of a fusion promoter;
- iv) diluting and culturing the fused cells in separate wells in a medium which will not support the unfused myeloma cells;
- v) evaluating the supernatant in each well containing a hybridoma for the presence of an antibody having any of the properties specified above;
- vi) selecting and cloning hybridomas producing the desired antibody; and
- vii) recovering the antibody from the supernatant above said clones.

Alternatively, the selected hybridoma is transferred intraperitoneally to a mouse and the antibody is harvested from the malignant ascites or serum of said mouse.

In another aspect, the invention provides a method for preparing mouse monoclonal antibody, which comprises culturing the hybridoma ATCC CRL 8022 in a suitable medium and recovering the antibody from the supernatant above said hybridoma.

Alternatively, the method comprises injecting into a mouse the hybridoma ATCC CRL 8022 and recovering the antibody from the malignant ascites or serum of said mouse.

The preparation and characterization of the hybridoma and the resultant antibody will be

better understood by reference to the following description and Examples.

Detailed description of the invention

The method of preparing the hybridoma generally comprises the following steps:

A. Immunizing mice with leukemic T-ALL cells. While it has been found that female CAF₁ mice are preferred, it is contemplated that other mouse strains could be used. The immunization schedule and thymocyte concentration should be such as to produce useful quantities of suitably primed splenocytes. Three immunizations at fourteen day intervals with 2×10^7 cells/mouse/injection in 0.2 ml phosphate buffered saline has been found to be effective.

B. Removing the spleens from the immunized mice and making a spleen suspension in an appropriate medium. About one ml of medium per spleen is sufficient. These experimental techniques are well-known.

C. Fusing the suspended spleen cells with mouse myeloma cells from a suitable cell line by the use of a suitable fusion promoter. The preferred ratio is about 5 spleen cells per myeloma cell. A total volume of about 0.5—1.0 ml of fusion medium is appropriate for about 10^8 splenocytes. Many mouse myeloma cell lines are known and available, generally from members of the academic community or various deposit banks, such as the Salk Institute Cell Distribution Center, La Jolla, CA. The cell line used should preferably be of the so-called "drug resistant" type, so that unfused myeloma cells will not survive in a selective medium, while hybrids will survive. The most common class is 8-azaguanine resistant cell lines, which lack the enzyme hypoxanthine guanine phosphoribosyl transferase and hence will not be supported by HAT (hypoxanthine, aminopterin, and thymidine) medium. It is also generally preferred that the myeloma cell line used be of the so-called "non-secreting" type, in that it does not itself produce any antibody, although secreting types may be used. In certain cases, however, secreting myeloma lines may be preferred. While the preferred fusion promoter is polyethylene glycol having an average molecular weight from about 1000 to about 4000 (commercially available as PEG 1000, etc.), other fusion promoters known in the art may be employed.

D. Diluting and culturing in separate containers, the mixture of unfused spleen cells, unfused myeloma cells, and fused cells in a selective medium which will not support the unfused myeloma cells for a time sufficient to allow death of the unfused cells (about one week). The dilution may be a type of limiting one, in which the volume of diluent is statistically calculated to isolate a certain number of cells (e.g., 1—4) in each separate container (e.g., each well of a microtiter plate). The medium is one (e.g., HAT medium) which will not support the drug resistant (e.g., 8-azaguanine resistant) unfused myeloma cell line. Hence, these myeloma cells perish. Since the unfused spleen cells are non-malignant, they

have only a finite number of generations. Thus, after a certain period of time (about one week) these unfused spleen cells fail to reproduce. The fused cells, on the other hand, continue to reproduce because they possess the malignant quality of the myeloma parent and the ability to survive in the selective medium of the spleen cell parent.

E. Evaluating the supernatant in each container (well) containing a hybridoma for the presence of antibody to E rosette positive purified human T cells or thymocytes.

F. Selecting (e.g., by limiting dilution) and cloning hybridomas producing the desired antibody.

Once the desired hybridoma has been selected and cloned, the resultant antibody may be produced in one of two ways. The purest monoclonal antibody is produced by *in vitro* culturing of the desired hybridoma in a suitable medium for a suitable length of time, followed by recovery of the desired antibody from the supernatant. The suitable medium and suitable length of culturing time are known or are readily determined. This *in vitro* technique produces essentially monospecific monoclonal antibody, essentially free from other specific antihuman immunoglobulin. There is a small amount of other immunoglobulin present since the medium contains xenogeneic serum (e.g., fetal calf serum). However, this *in vitro* method may not produce a sufficient quantity or concentration of antibody for some purposes, since the concentration of monoclonal antibody is only about 50 µg/ml.

To produce a much greater concentration of slightly less pure monoclonal antibody, the desired hybridoma may be injected into mice, preferably syngenic or semi-syngenic mice. The hybridoma will cause formation of antibody-producing tumors after a suitable incubation time, which will result in a high concentration of the desired antibody (about 5—20 mg/ml) in the bloodstream and peritoneal exudate (ascites) of the host mouse. Although these host mice also have normal antibodies in their blood and ascites, the concentration of these normal antibodies is only about 5% of the monoclonal antibody concentration. Moreover, since these normal antibodies are not antihuman in their specificity, the monoclonal antibody obtained from the harvested ascites or from the serum is essentially free of any contaminating antihuman immunoglobulin. This monoclonal antibody is high titer (active at dilutions of 1:50,000 or higher) and high ratio of specific to non-specific immunoglobulin (about 1/20). Immunoglobulin produced incorporating the light myeloma chains are non-specific, "nonsense" peptides which merely dilute the monoclonal antibody without detracting from its specificity.

Example 1

Production of monoclonal antibodies

A. Immunization and somatic cell hybridization
Female CAF₁ mice (Jackson Laboratories; 6—8

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weeks old) were immunized intraperitoneally with 2×10^7 human leukemic T-ALL cells in 0.2 ml of phosphate buffered saline at 14-day intervals. Four days after the third immunization, spleens were removed from the mice, and a single cell suspension was made by pressing the tissue through a stainless steel mesh.

Cell fusion was carried out according to the procedure developed by Kohler and Milstein. 1×10^6 splenocytes were fused in 0.5 ml of a fusion medium comprising 35% polyethylene glycol (PEG 1000) and 5% dimethylsulfoxide in RPMI 1640 medium (Gibco, Grand Island, NY) with 2×10^7 P3X63Ag8U1 myeloma cells supplied by Dr. M. Scharff, Albert Einstein College of Medicine, Bronx, NY. These myeloma cells secrete IgG₁κ light chains.

B. Selection and growth of hybridoma

After cell fusion, cells were cultured in HAT medium (hypoxanthine, aminopterin, and thymidine) at 37°C with 5% CO₂ in a humid atmosphere. Several weeks later, 40 to 100 μl of supernatant from cultures containing hybridomas were added to a pellet of 10^6 peripheral lymphocytes separated into E rosette positive (E⁺) and E rosette negative (E⁻) populations, which were prepared from blood of healthy human donors as described by Mendes (*J. Immunol.* 111:860, 1973). Detection of mouse hybridoma antibodies binding to these cells was determined by indirect immunofluorescence. Cells incubated with culture supernatants were stained with a fluoresceinated goat-anti-mouse IgG (G/M FITC) (Meloy Laboratories, Springfield, VA; F/p=2.5) and the fluorescent antibody-coated cells were subsequently analyzed on the Cytofluorograf FC200/4800A (Ortho Instruments, Westwood, MA) as described in Example III. Hybridoma cultures containing antibodies reacting specifically with E⁺ lymphocytes (T cells) and/or thymocytes were selected and cloned twice by limiting dilution methods in the presence of feeder cells. Subsequently, the clones were transferred intraperitoneally by injecting 1×10^7 cells of a given clone (0.2 ml volume) into CAF₁ mice primed with 2,6,10,14-tetramethylpentadecane, sold by Aldrich Chemical Company under the name Pristine. The malignant ascites from these mice were then used to characterize lymphocytes as described below in Example II. The subject hybrid antibody OKT10 was demonstrated by standard techniques to be of IgG₁ subclass.

Example II

Characterization of OKT10 reactivity

A. Isolation of lymphocyte populations

Human peripheral blood mononuclear cells were isolated from healthy volunteer donors (ages 15–40) by Ficoll-Hypaque density gradient centrifugation (Pharmacia Fine Chemicals, Piscataway, NJ) following the technique of Boyum, *Scand. J. Clin. Lab. Invest.* 21 (Suppl. 97): 77, 1968. Unfractionated mononuclear cells were separated into surface Ig⁺ (B) and Ig⁻ (T plus Null)

populations by Sephadex G-200 anti-F (ab')₂ column chromatography as previously described by Chess, *et al.*, *J. Immunol.* 113:1113 (1974). T cells were recovered by E rosetting the Ig⁻ population with 5% sheep erythrocytes (Microbiological Associates, Bethesda, MD). The rosetted mixture was layered over Ficoll-Hypaque and the recovered E⁺ pellet treated with 0.155M NH₄Cl (10 ml per 10^8 cells). The T cell population so obtained was <2% EAC rosette positive and >95% E rosette positive as determined by standard methods. In addition, the non-rosetting Ig⁻ (Null cell) population was harvested from the Ficoll interface. This latter population was <5% E⁺ and ≤2% slg⁺. The surface Ig⁺ (B) population was obtained from the Sephadex G-200 column following elution with normal human gamma globulin as previously described. This population was >95% surface Ig⁺ and <5% E⁺.

Normal human bone marrow cells were obtained from the posterior iliac crest of normal human volunteers by needle aspiration.

B. Isolation of thymocytes

Normal human thymus gland was obtained from patients aged two months to 14 years undergoing corrective cardiac surgery. Freshly obtained portions of the thymus gland were immediately placed in 5% fetal calf serum in medium 199 (Gibco), finely minced with forceps and scissors, and subsequently made into single cell suspensions by being pressed through wire mesh. The cells were next layered over Ficoll-Hypaque and spun and washed as previously described in section A above. The thymocytes so obtained were >95% viable and ≥90% E rosette positive.

C. Cell lines of T lineage and T acute lymphoblastic leukemia cells

T cell lines CEM, HSB-2, and MOLT-4 were provided by Dr. H. Lazarus (Sidney Farber Cancer Institute, Boston, MA). Leukemic cells were obtained from 25 patients with the diagnosis of T cell ALL. These individual tumors had been previously determined to be of T cell lineage by their spontaneous rosette formation with sheep erythrocytes (>20% E⁺), and reactivity with T cell specific heteroantisera anti-HTL (B.K.) and A99, as previously described. Tumor populations were cryopreserved at -196°C by vapor phase liquid nitrogen with 10% DMSO and 20% AB human serum until the time of surface characterization. All tumor populations analyzed were more than 90% blasts by Wright-Giemsa morphology of cytocentrifuge preparations.

Example III

Cytofluorographic analysis and cell separation
Cytofluorographic analysis of monoclonal antibodies with all cell populations was performed by indirect immunofluorescence with fluorescein-conjugated goat anti-mouse IgG (G/M FITC) (Meloy Laboratories) utilizing a Cytofluorograf FC200/4800A (Ortho Instruments). In brief,

1x10⁶ cells were treated with 0.15 ml OKT10 at a 1:500 dilution, incubated at 4°C for 30 minutes, and washed twice. The cells were then reacted with 0.15 ml of a 1:40 dilution G/M FITC at 4°C for 30 minutes, centrifuged, and washed three times. Cells were then analyzed on the Cytofluorograf, and the intensity of fluorescence per cell was recorded on a pulse height analyzer. A similar pattern of reactivity was seen at a dilution of 1:10,000, but further dilution caused loss of reactivity. Background staining was obtained by substituting a 0.15 ml aliquot of 1:500 ascites from a CAF₁ mouse intraperitoneally injected with a non-producing hybrid clone.

In experiments involving antibody and complement mediated lympholysis, thymocytes and peripheral T cells were cultured overnight following selective lysis and then subsequently analyzed on the Cytofluorograf.

Example IV

Lysis of lymphoid populations with monoclonal antibody and complement

Forty x10⁶ peripheral T cells or thymocytes were placed in a 15 ml plastic tube (Falcon, Oxnard, CA). Cell pellets were incubated with 0.8 cc of OKT3, OKT4, OKT8, or normal ascites control diluted 1:200 in PBS, resuspended, and incubated at 20°C for 60 minutes. Subsequently, 0.2 cc of fresh rabbit complement was added to the antibody treated populations, resuspended, and further incubated at 37°C in a shaking water bath for 60 minutes. At the end of this time, cells were spun down and viable cells enumerated by Trypan blue exclusion. After counting, cells were washed two additional times in 5% FCS and placed in final media [RPMI 1640 (Grand Island Biological Company, Grand Island, NY) containing 20% AB⁺ human serum, 1% penicillin-streptomycin, 200 mM L-glutamine, 25 mM HEPES buffer, and 0.5% sodium bicarbonate] and incubated overnight in a humid atmosphere with 5% CO₂ at 37°C.

Brief description of the drawings

Figure 1 shows the fluorescence pattern obtained on the Cytofluorograf after reacting normal human thymocytes with OKT10 and other monoclonal antibodies at a 1:500 dilution and G/M FITC. Background fluorescence staining was obtained by incubating each population with a 1:500 dilution of ascitic fluid from a mouse injected with a non-producing clone.

Figure 2 shows the stages of intrathymic differentiation in man.

The production of the hybridoma and the production and characterization of the resulting monoclonal antibody were conducted as described in the above Examples. Although large quantities of the subject antibody were prepared by injecting the subject hybridoma intraperitoneally into mice and harvesting the malignant ascites, it is clearly contemplated that the hybridoma could be cultured *in vitro* by tech-

niques well-known in the art and the antibody removed from the supernatant.

Table 1 shows the reactivity of OKT6, OKT8, OKT9, and OKT10 with various human lymphoid cell populations. The OKT10 monoclonal antibody is reactive with approximately 95% of normal human thymocytes, 5% of normal human peripheral T cells, 10% of E⁻ peripheral mononuclear cells (B cells and null cells), and 10-20% of bone marrow cells. This pattern of reactivity is one test by which the subject antibody OKT10 may be detected and distinguished from other antibodies.

Figure 1 shows representative fluorescence pattern obtained on the Cytofluorograf after reacting normal human thymocyte suspensions with a 1:500 dilution of OKT3, OKT4, OKT5, OKT6, OKT8, OKT9, OKT10, and G/M FITC. Similar patterns of reactivity were seen with 12 additional normal human thymocyte populations tested. As shown, significant differences exist in both the percentage of reactivity and fluorescence intensity with each of these monoclonal antibodies. For example, OKT9 reacts with approximately 10% of thymocytes with low fluorescence intensity while OKT5, OKT6, OKT8 and OKT10 react with approximately 70% of thymocytes at a higher fluorescence intensity. OKT4, which reacts with 75% of thymocytes, is intermediate between OKT9 and the monoclonal antibodies which give a pattern of greater fluorescence intensity. In addition, Figure 1 shows that approximately 15% of thymocytes are detected with OKT3 by indirect immunofluorescence. Not shown is OKT1, whose pattern of reactivity is virtually identical to OKT3 on thymocytes. The pattern of reactivity in Figure 1 is another test by which the subject antibody OKT10 may be detected and distinguished from other antibodies.

Table 2 shows the distribution of antigens defined by various monoclonal antibodies on human peripheral T cells and lymphocytes, as determined by the series of lysis experiments described in Example IV. Since only OKT3, OKT4, and OKT8 were complement fixing monoclonal antibodies, these three were utilized.

As shown in Table 2A, the entire T cell population reacts with OKT3 while OKT4, OKT5, and OKT8 react with 60%, 25%, and 34% of T cells, respectively. Lysis with OKT4 and complement diminished the total number by 62% and specifically deleted the OKT4⁺ population. In addition, the percentage of OKT5⁺ and OKT8⁺ cells increased and there was no effect on the absolute number of OKT5⁺ and OKT8⁺ T cells. These experiments suggested that OKT4⁺ was distinct from the OKT5⁺ and OKT8⁺ populations. Further support for this conclusion was obtained by lysis of T cells with OKT8 and complement. In this case, the percentage of OKT4⁺ T cells increased, the absolute number remained the same, and OKT8⁺ and OKT5⁺ populations were eliminated. Moreover, these results demonstrated that the OKT8⁺ population was reciprocal to the OKT4⁺ population and contained the entire OKT5⁺ T cell subset.

Similar experiments with human thymocyte

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populations gave different results. As shown in Table 2B, approximately 75% of thymocytes were OKT4⁺ or OKT8⁺. Moreover, following lysis with either OKT4 or OKT8, only 25% of thymocytes remained. The majority of residual thymocytes were reactive with OKT3, whereas only a minority was reactive with OKT6. These findings demonstrate that a major population of human thymocytes bear the OKT4, OKT5, OKT6, and OKT8 surface antigens on the same cell. In addition, Table 2 demonstrates that following treatment with OKT8 or OKT4, there is a marked increase in the mature thymocytes bearing the OKT3 antigen. Thus, the majority of OKT3 reactive thymocytes have already segregated into OKT4⁺ or OKT8⁺ subsets, since the major proportion of residual cells following OKT4 or OKT8 lysis are OKT3⁺. If the OKT3⁺ subpopulation were both OKT4⁺ and OKT8⁺, then lysis with either monoclonal antibody should have removed the OKT3 reactive thymocytes.

To further determine the relationship of OKT3 reactive thymocyte subpopulations to the other monoclonal antibody defined thymocyte fractions, thymocytes were treated with OKT3 and complement and the residual cells were then compared to untreated thymocyte populations. As shown in Table 2B, OKT3 and complement removed 25% of thymocytes. Moreover, there was no major loss of OKT4, OKT5, OKT6, or OKT8 reactive populations. These findings suggest that the vast majority of thymocytes bearing the OKT6 marker are contained in the OKT3⁺ population. In addition, they further suggest that thymocytes simultaneously expressing antigens defined by OKT4, OKT5, and OKT8 are likewise restricted to the OKT3⁺ population. It should also be noted that the OKT9 reactive population of thymocytes was not diminished following OKT3 and complement treatment of the unfractionated thymocytes, thus showing that the OKT9⁺ subpopulation is largely restricted to the OKT3⁺ thymocyte population.

Based upon these results, it has been possible to describe the stages of intrathymic development of human thymocytes. As shown in Figure 2, virtually all thymocytes bear the OKT10 marker. In addition, thymocytes acquire at an early stage the OKT9 marker (Thy1 and Thy2, respectively). This stage defines the minority of thymocytes and accounts for approximately 10% of the unfractionated population. Subsequently, human thymocytes acquire a thymocyte unique antigen defined by OKT6 and concurrently express OKT4, OKT5, and OKT8 (Thy4). This latter subpopulation represents the majority of thymocytes and accounts for upwards of 70–80% of the thymic population. With further maturation, thymocytes lose OKT6 reactivity, acquire OKT3 (and OKT1) reactivity, and segregate into OKT4⁺ and OKT5⁺/OKT8⁺ subsets (Thy7 and Thy8). Lastly, it appears that as the thymocyte is exported into the peripheral T cell compartment, it loses the OKT10 marker since this antigen is lacking on virtually all peripheral T lymphocytes. Possible transitional states between these three major stages of

thymic development are designated by Thy3, Thy5, and Thy6 in Figure 2.

Since acute lymphoblastic leukemia of T lineage is thought to be derived from immature thymocytes, the relationship between tumor cells from individuals with T-ALL and these proposed stages of intrathymic differentiation was determined. Twenty-five tumor cell populations from individuals with T-ALL and three T cell lines previously studied with conventional anti-T cell reagents and E rosetting were investigated. As shown in Table 3, the majority of T-ALL leukemic cells were reactive with either OKT10 alone or OKT9 and OKT10 and failed to react with the other monoclonal antibodies. Thus, 15/25 cases studied appeared to possess early thymocyte antigens (Stage I).

In contrast, 5/25 cases were reactive with OKT6, suggesting derivation from a more mature thymus population (Stage II). This T-ALL group was itself heterogeneous with respect to OKT4, OKT8, and OKT9 reactivity as shown in Table 3. Cells from 2/5 patients possess most of the common thymocyte antigens including OKT4, OKT6, and OKT8. It is worthy of note that OKT5 is not present on any of these 5 Stage II tumors even though OKT8 reactivity was observed. This latter result clearly suggests that OKT5 and OKT8 define different antigens or different determinants on the same antigen. Finally, 1/25 patients' tumors came from a mature thymocyte population (Stage III) as defined by its reactivity with OKT3. This individual's tumor, in addition, was reactive with OKT4, OKT8, and OKT10. Of the 25 leukemic populations analyzed, only four tumors could not be clearly categorized. Three were positive with OKT4 and OKT8, but lacked OKT3 and OKT6 and most likely represented transitions from Thy4 and Thy7, 8. One of 25 cases appeared to be a transition from Thy3 to Thy4 since it possessed OKT8 and OKT10 reactivity.

T cell lines derived from T-ALL tumor populations also represented cells from a specific state of intrathymic differentiation. As shown in Table 4, HSB was reactive with OKT9 and OKT10 exclusively and would therefore define a tumor population derived from Stage I. In contrast, CEM was reactive with OKT4, OKT6, OKT8, OKT9 and OKT10 and appeared to derive from a Stage II thymocyte. Finally, MOLT-4 seems to represent a leukemic transformation at a stage between HSB-2 and CEM since it expressed OKT6, OKT8, OKT9, and OKT10.

Since all patients studied with T cell acute lymphoblastic leukemia have been shown to have OKT10⁺ cells, the use of OKT10 antibody allows diagnosis of T-cell ALL. Patients with null cell ALL also exhibited OKT10⁺ cells.

The relationships shown in Tables 2–4 are a further way in which OKT10 antibody may be detected and distinguished from other antibodies.

Other monoclonal antibody producing hybridomas prepared by the present applicants (designated OKT1, OKT3, OKT4, and OKT5) are described and claimed in the following European

patent applications: 80300829.1, 80301356.4, 80301357.2, and 80303272.1.

Still other monoclonal antibody producing hybridomas prepared by the present applicants (designated OKT6, OKT8, and OKT9) are described and claimed in European patent applications filed on even date herewith and entitled:

Hybrid Cell Line For Producing Complement-Fixing Monoclonal Antibody to Human Suppressor T Cells, Antibody, and Methods; Hybrid Cell Line For Producing Monoclonal Antibody to Human Early Thymocyte Antigen, Antibody, and Methods; and Hybrid Cell Line for Producing Monoclonal Antibody to a Human Thymocyte Antigen, Antibody, and Methods and numbered

respectively, 80304347.0, 80304348.8 and 80304349.6.

These applications are incorporated herein by reference.

According to the present invention there are provided a hybridoma capable of producing antibody against an antigen found on approximately 95% of normal human thymocytes, a method for producing this hybridoma, monoclonal antibody against an antigen found on approximately 95% of normal human thymocytes, methods for producing the antibody, compositions for treatment or diagnosis and methods of diagnosis of disease or identification of T cell or thymocyte subclasses employing this antibody.

TABLE 1
Reactivity of monoclonal antibodies on human lymphoid populations

Monoclonal antibody	Peripheral blood (30)*		Bone marrow (6)	Thymus (22)
	E ⁺	E ⁻		
OKT6	0%	0%	0%	70%
OKT8	30%	0%	<2%	80%
OKT9	0%	0%	0%	≤10%
OKT10	<5%	10%	≤20%	95%

* Numbers in parentheses represent the number of samples tested; % values are means.

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TABLE 2
Differences in distribution or antigens defined by
monoclonal antibody on human peripheral T cells and thymocytes

Lymphoid population	Total cell number recovered	Percent reactivity of residual cells with monoclonal antibodies:							
		OKT3	OKT4	OKT5	OKT6	OKT8	OKT9	OKT10	
A. Peripheral T Cells									
Untreated*	40×10 ⁶	98	60	25	0	34	—	—	
OKT4+C'†	15.2×10 ⁶	95	0	70	0	95	—	—	
OKT8+C'	25.2×10 ⁶	95	92	0	0	0	—	—	
B. Thymocytes									
Untreated*	40×10 ⁶	30	75	70	65	75	8	92	
OKT4+C'	10×10 ⁶	80	0	85	10	55	6	75	
OKT8+C'	9.5×10 ⁶	85	65	0	13	0	5	85	
OKT3+C'	30×10 ⁶	0	60	80	82	90	12	86	

* Untreated populations and populations treated with complement alone were indistinguishable on re-analysis. Non-specific lysis was ≤5% in all cases. Results are representative of 6 experiments.

† C' = complement

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TABLE 3
Cell surface characteristics of acute lymphoblastic leukemia of T-lineage

Differentiative status	Reactivity with monoclonal antibodies:							Number of T-ALL tested (n=25)
	OKT3	OKT4	OKT5	OKT6	OKT8	OKT9	OKT10	
Stage I								
A. Prothymocyte (Thy1)†	-	-	-	-	-	-	++	7
B. Early thymocyte (Thy2)	-	-	-	-	-	+	+	8
Stage II								
Common thymocyte								
(Thy3)	-	+	-	+	+	+	+	1
(Thy4)	-	+	-	+	+	-	+	1
(Thy3-4)	-	-	-	+	-	-	+	2
(Thy4-6)	-	-	-	+	+	-	+	1
Stage III								
Late thymocyte (Thy8)	+	-	+	-	+	-	+	1
								21*

* An additional four tumors could not be easily categorized into Stage I-III. See text for details of their characterization.

† Thy designation refers to Figure 2

‡ Positive (+) reactivity was defined as $\geq 30\%$ specific fluorescence above background control while negative (-) reactivity was indistinguishable from background staining on tumor cell suspensions.

TABLE 4
Reactivity with monoclonal antibodies

Cell line	OKT3	OKT4	OKT5	OKT6	OKT8	OKT9	OKT10
HSB-2	-*	-	-	-	-	+	+
CEM	-	+	-	+	+	+	+
MOLT-4	-	-	-	+	+	+	+

* Criteria for - and + reactivity was the same as in Table 3.

Although only a single hybridoma producing a single monoclonal antibody against a human thymocyte antigen is described, it is contemplated that the present invention encompasses all monoclonal antibodies exhibiting the characteristics described herein. It is determined that the subject antibody OKT10 belongs to the subclass IgG₁, which is one of four subclasses of murine IgG. These subclasses of immunoglobulin G differ from one another in the so-called "fixed" regions, although an antibody to a specific antigen will have a so-called "variable" region which is functionally identical regardless of which subclass of immunoglobulin G it belongs to. That is, a monoclonal antibody exhibiting the characteristic described herein may be of subclass IgG₁, IgG_{2a}, IgG_{2b}, or IgG₃, or of classes IgM, IgA, or other known Ig classes. The differences among these classes or subclasses will not affect the selectivity of the reaction pattern of the antibody, but may affect the further reaction of the antibody with other materials, such as (for example) complement or anti-mouse antibodies. Although the subject antibody is specifically IgG₁, it is contemplated that antibodies having the patterns of reactivity illustrated herein are included within the subject invention regardless of the immunoglobulin class or subclass to which they belong.

Further included within the subject invention are methods for preparing the monoclonal antibodies described above employing the hybridoma technique illustrated herein. Although only one example of a hybridoma is given herein, it is contemplated that one skilled in the art could follow the immunization, fusion, and selection methods provided herein and obtain other hybridomas capable of producing antibodies having the reactivity characteristics described herein. Since the individual hybridoma produced from a known mouse myeloma cell line and spleen cells from a known species of mouse cannot be further identified except by reference to the antibody produced by the hybridoma, it is contemplated that all hybridomas producing antibody having the reactivity characteristics described above are included within the subject invention, as are methods for making this antibody employing the hybridoma.

Further aspects of the invention are methods of diagnosis of disease employing the monoclonal antibody OKT10 or any other monoclonal anti-

body exhibiting the pattern of reactivity provided herein. The subject antibody may be used to detect and study intrathymic differentiation as summarized in Figure 2. Moreover, the subject antibody may be employed to diagnose disease states involving a defect of excess of OKT10⁺ cells. These techniques may be employed using OKT10 antibody alone or in combination with other antibodies (e.g., OKT3—OKT10). Patterns of reactivity with a panel of antibodies to T cells and T cell subsets will allow more precise detection of certain disease states then is possible using prior diagnostic methods.

Treatment of disease states (e.g., malignancies such as null cell ALL or T-ALL) manifesting themselves as an excess of OKT10⁺ cells may be accomplished by administration of a therapeutically effective amount of OKT10 antibody to an individual in need of such treatment. By selective reaction with OKT10⁺ antigen, the effective amount of OKT10 antibody will reduce the excess of OKT10⁺ cells, thus ameliorating the effects of the excess. Diagnostic and therapeutic compositions comprising effective amounts of OKT10 antibody in admixture with diagnostically or pharmaceutically acceptable carriers, respectively, are also included within the present invention.

Claims

1. Mouse monoclonal antibody which reacts with

- i) approximately 95% of normal human thymocytes,
- ii) 5% of normal human peripheral T cells.
- iii) 10% of E⁻ peripheral mononuclear cells, and
- iv) 10—20% of bone marrow cells.

2. Mouse monoclonal antibody according to claim 1, which exhibits the pattern of reactivity to normal human thymocytes shown in Figure 1.

3. Mouse monoclonal antibody according to claim 1 or claim 2, which exhibits the pattern of reactivity to peripheral T cells and thymocytes shown in Table 2.

4. Mouse monoclonal antibody according to any one of claims 1 to 3, which is reactive with an antigen on leukemic T-ALL cells.

5. Mouse monoclonal antibody according to any one of claims 1 to 4, of class IgG.

TABLE 4
Reactivity with monoclonal antibodies

Cell line	OKT3	OKT4	OKT5	OKT6	OKT8	OKT9	OKT10
HSB-2	-	-	-	-	-	+	+
CEM	-	+	-	+	+	+	+
MOLT-4	-	-	-	+	+	+	+

* Criteria for - and + reactivity was the same as in Table 3.

Although only a single hybridoma producing a single monoclonal antibody against a human thymocyte antigen is described, it is contemplated that the present invention encompasses all monoclonal antibodies exhibiting the characteristics described herein. It was determined that the subject antibody OKT10 belongs to the subclass IgG₁, which is one of four subclasses of murine IgG. These subclasses of immunoglobulin G differ from one another in the so-called "fixed" regions, although an antibody to a specific antigen will have a so-called "variable" region which is functionally identical regardless of which subclass of immunoglobulin G it belongs to. That is, a monoclonal antibody exhibiting the characteristic described herein may be of subclass IgG₁, IgG_{2a}, IgG_{2b}, or IgG₃, or of classes IgM, IgA, or other known Ig classes. The differences among these classes or subclasses will not affect the selectivity of the reaction pattern of the antibody, but may affect the further reaction of the antibody with other materials, such as (for example) complement or anti-mouse antibodies. Although the subject antibody is specifically IgG₁, it is contemplated that antibodies having the patterns of reactivity illustrated herein are included within the subject invention regardless of the immunoglobulin class or subclass to which they belong.

Further included within the subject invention are methods for preparing the monoclonal antibodies described above employing the hybridoma technique illustrated herein. Although only one example of a hybridoma is given herein, it is contemplated that one skilled in the art could follow the immunization, fusion, and selection methods provided herein and obtain other hybridomas capable of producing antibodies having the reactivity characteristics described herein. Since the individual hybridoma produced from a known mouse myeloma cell line and spleen cells from a known species of mouse cannot be further identified except by reference to the antibody produced by the hybridoma, it is contemplated that all hybridomas producing antibody having the reactivity characteristics described above are included within the subject invention, as are methods for making this antibody employing the hybridoma.

Further aspects of the invention are methods of diagnosis of disease employing the monoclonal antibody OKT10 or any other monoclonal anti-

body exhibiting the pattern of reactivity provided herein. The subject antibody may be used to detect and study intrathymic differentiation as summarized in Figure 2. Moreover, the subject antibody may be employed to diagnose disease states involving a defect of excess of OKT10⁺ cells. These techniques may be employed using OKT10 antibody alone or in combination with other antibodies (e.g., OKT3—OKT10). Patterns of reactivity with a panel of antibodies to T cells and T cell subsets will allow more precise detection of certain disease states then is possible using prior diagnostic methods.

Treatment of disease states (e.g., malignancies such as null cell ALL or T-ALL) manifesting themselves as an excess of OKT10⁺ cells may be accomplished by administration of a therapeutically effective amount of OKT10 antibody to an individual in need of such treatment. By selective reaction with OKT10⁺ antigen, the effective amount of OKT10 antibody will reduce the excess of OKT10⁺ cells, thus ameliorating the effects of the excess. Diagnostic and therapeutic compositions comprising effective amounts of OKT10 antibody in admixture with diagnostically or pharmaceutically acceptable carriers, respectively, are also included within the present invention.

Claims

1. Mouse monoclonal antibody which reacts with

- i) approximately 95% of normal human thymocytes,
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- iii) 10% of E⁻ peripheral mononuclear cells, and
- iv) 10—20% of bone marrow cells.

2. Mouse monoclonal antibody according to claim 1, which exhibits the pattern of reactivity to normal human thymocytes shown in Figure 1.

3. Mouse monoclonal antibody according to claim 1 or claim 2, which exhibits the pattern of reactivity to peripheral T cells and thymocytes shown in Table 2.

4. Mouse monoclonal antibody according to any one of claims 1 to 3, which is reactive with an antigen on leukemic T-ALL cells.

5. Mouse monoclonal antibody according to any one of claims 1 to 4, of class IgG.



(21)

6. Mouse monoclonal antibody according to any one of claims 1 to 5, produced by a hybridoma formed by fusion of cells from a mouse myeloma line and spleen cells from a mouse previously immunized with human leukemic T-ALL cells.

7. Mouse monoclonal antibody produced by hybridoma ATCC CRL 8022 (OKT10).

8. A hybridoma which produces monoclonal antibody according to any one of claims 1 to 6 formed by fusion of spleen cells from a mouse previously immunized with human leukemic T-ALL cells and cells from a mouse myeloma line.

9. Hybridoma ATCC CRL 8022 (OKT10).

10. A method for preparing mouse monoclonal antibody according to any one of claims 1 to 6, which comprises the steps of:

- i) immunizing mice with human leukemic T-ALL cells;
- ii) removing the spleens from said mice and making a suspension of spleen cells;
- iii) fusing said spleen cells with mouse myeloma cells in the presence of a fusion promoter;
- iv) diluting and culturing the fused cells in separate wells in a medium which will not support the unfused myeloma cells;
- v) evaluating the supernatant in each well containing a hybridoma for the presence of an antibody having the properties specified in any of claims 1 to 6;
- vi) selecting and cloning hybridomas producing the desired antibody; and
- vii) recovering the antibody from the supernatant above said clones.

11. A method for preparing mouse monoclonal antibody according to any one of claims 1 to 7, which comprises the steps of:

- i) immunizing mice with human leukemic T-ALL cells;
- ii) removing the spleens from said mice and making a suspension of the spleen cells;
- iii) fusing said spleen cells with mouse myeloma cells in the presence of a fusion promoter;
- iv) diluting and culturing the fused cells in separate wells in a medium which will not support the unfused myeloma cells;
- v) evaluating the supernatant in each well containing a hybridoma for the presence of an antibody having the properties specified in any one of claims 1 to 6;
- vii) selecting and cloning hybridomas producing the desired antibody;
- viii) transferring said clones intraperitoneally into mice; and
- ix) harvesting the malignant ascites or serum from said mice.

12. A method for preparing mouse monoclonal antibody, which comprises culturing the hybridoma ATCC CRL 8022 in a suitable medium

and recovering the antibody from the supernatant above said hybridoma.

13. A method for preparing mouse monoclonal antibody, which comprises injecting into a mouse the hybridoma ATCC CRL 8022 and recovering the antibody from the malignant ascites or serum of said mouse.

14. A diagnostic composition of matter for detection of OKT10⁺ cell excess or deficiency comprising, in admixture with a diagnostically acceptable carrier, an amount of mouse monoclonal antibody according to any one of claims 1 to 7, effective to detect OKT10⁺ excess or deficiency.

15. A therapeutic composition of matter comprising, in admixture with a pharmaceutically acceptable carrier, an amount of mouse monoclonal antibody according to any one of claims 1 to 7, effective to reduce the amount of OKT10⁺ cells in an individual having an excess of said OKT10⁺ cells.

16. A method for detection of a deficiency or excess of OKT10⁺ cells in an individual which comprises reacting a T cell or thymocyte composition from said individual with a diagnostically-effective amount of mouse monoclonal antibody according to any one of claims 1 to 7 or a composition according to claim 14 and measuring the percentage of the total peripheral T cell and thymocyte population which reacts with said antibody.

17. The method of claim 16 wherein the excess is T cell ALL or null cell ALL.

18. Mouse monoclonal antibody according to any one of claims 1 to 7, or a composition according to claim 15, for the treatment of an OKT10⁺ cell excess in an individual in need of such treatment.

19. A method for preparing a composition according to claim 14 or claim 15 comprising mixing a diagnostically- or pharmaceutically-acceptable carrier with a diagnostically- or pharmaceutically- effective amount of mouse monoclonal antibody according to any one of claims 1 to 7.

Patentansprüche

1. Monoklonaler Maus-Antikörper, welcher mit

- i) annähernd 95% normalen humanen Thymocyten,
- ii) 5% normalen humanen peripheren T-Zellen,
- iii) 10% E⁻-peripheren mononuklearen Zellen und
- iv) 10 bis 20% Knochenmarkzellen reagiert.

2. Monoklonaler Maus-Antikörper nach Anspruch 1, welcher das in Figur 1 dargestellte Reaktivitätsmuster gegenüber normalen humanen Thymocyten aufweist.

3. Monoklonaler Maus-Antikörper nach Anspruch 1 oder 2, welcher das in Tabelle 2 dargestellte Reaktivitätsmuster gegenüber peripheren T-Zellen und Thymocyten aufweist.



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4. Monoklonaler Maus-Antikörper nach einem der Ansprüche 1 bis 3, welcher mit einem Antigen auf leukämischen T-ALL-Zellen reaktiv ist.

5. Monoklonaler Maus-Antikörper nach einem der Ansprüche 1 bis 4 der Klasse IgG.

6. Monoklonaler Maus-Antikörper nach einem der Ansprüche 1 bis 5, welcher durch ein Hybridom gebildet wird, das durch Fusion von Zellen von einer Maus-Myelom-Linie und von Milzzellen von einer zuvor mit humanen leukämischen T-ALL-Zellen immunisierten Maus gebildet ist.

7. Monoklonaler Maus-Antikörper, welcher von einem Hybridom ATCC CRL 8022 (OKT10) gebildet wird.

8. Ein Hybridom, welches einen monoklonalen Antikörper nach einem der Ansprüche 1 bis 6 produziert, gebildet durch Fusion von Zellen von einer zuvor mit humanen leukämischen T-ALL-Zellen immunisierten Maus und Zellen aus einer Maus-Myelom-Linie.

9. Das Hybridom ATCC CRL 8022 (OKT10).

10. Ein Verfahren zur Herstellung von monoklonalem Antikörper nach einem der Ansprüche 1 bis 6, umfassend die Stufen:

- i) Immunisieren von Mäusen mit humanen leukämischen T-ALL-Zellen;
- ii) Entfernen der Milz aus den genannten Mäusen und Bereiten einer Suspension der Milzzellen;
- iii) Fusion der genannten Milzzellen mit Maus-Myelom-Zellen in Gegenwart eines Fusionspromotors;
- iv) Verdünnen und Vermehren der vereinigten Zellen in getrennten Vertiefungen in einem Medium, welches die nicht-vereinigten Myelom-Zellen nicht erhält;
- v) Auswerten des Überstandes in jeder Vertiefung, die ein Hybridom enthält, hinsichtlich des Vorliegens eines Antikörpers mit den in einem der Ansprüche 1 bis 6 angegebenen Eigenschaften;
- vi) Auswählen und Klonen von Hybridomen, welche den gewünschten Antikörper bilden; und
- vii) Gewinnen des Antikörpers aus dem Überstand über den genannten Klonen.

11. Ein Verfahren zur Herstellung von monoklonalem Antikörper nach einem der Ansprüche 1 bis 7, umfassend die Stufen:

- i) Immunisieren von Mäusen mit humanen leukämischen T-ALL-Zellen;
- ii) Entfernen der Milz aus den genannten Mäusen und Bereiten einer Suspension der Milzzellen;
- iii) Fusion der genannten Milzzellen mit Maus-Myelom-Zellen in Gegenwart eines Fusionspromotors;
- iv) Verdünnen und Vermehren der vereinigten Zellen in getrennten Vertiefungen in einem Medium, welches die nicht-vereinigten Myelom-Zellen nicht erhält;

v) Auswerten des Überstandes in jeder Vertiefung, die ein Hybridom enthält, hinsichtlich des Vorliegens eines Antikörpers mit den in einem der Ansprüche 1 bis 6 angegebenen Eigenschaften;

vi) Auswählen und Klonen von Hybridomen, welche den gewünschten Antikörper bilden;

vii) Überführen der genannten Klone auf intraperitonealem Wege in Mäuse; und

viii) Ernten der malignen Asciten oder von Serum aus den genannten Mäusen.

12. Ein Verfahren zur Herstellung von monoklonalem Maus-Antikörper, welches ein Vermehren des Hybridoms ATCC CRL 8022 in einem geeigneten Medium und das Gewinnen des Antikörpers aus dem Überstand über dem genannten Hybridom umfaßt.

13. Ein Verfahren zur Herstellung von monoklonalem Maus-Antikörper, welches ein Injizieren des Hybridoms ATCC CRL 8022 in eine Maus und das Gewinnen des Antikörpers aus den malignen Asciten oder aus dem Serum der genannten Maus umfaßt.

14. Eine diagnostische Zusammensetzung zur Feststellung eines OKT10⁺-Zellen-Überschusses oder -Mangels, umfassend, im Gemisch mit einem diagnostisch annehmbaren Träger, eine zur Feststellung eines OKT10⁺-Zellen-Überschusses oder -Mangels wirksame Menge eines monoklonalen Maus-Antikörpers nach einem der Ansprüche 1 bis 7.

15. Eine therapeutische Zusammensetzung, enthaltend, im Gemisch mit einem pharmazeutisch annehmbaren Träger, eine zur Verminderung der Menge an OKT10⁺-Zellen in einem Individuum mit einem Überschuß an den genannten OKT10⁺-Zellen wirksame Menge des monoklonalen Maus-Antikörpers nach einem der Ansprüche 1 bis 7.

16. Ein Verfahren zur Feststellung eines Mangels oder eines Überschusses an OKT10⁺-Zellen in einem Individuum, umfassend ein Reagieren einer T-Zellen- oder Thymocyten-Zusammensetzung des genannten Individuums mit einer diagnostisch wirksamen Menge des monoklonalen Maus-Antikörpers nach einem der Ansprüche 1 bis 7 oder mit einer Zusammensetzung nach Anspruch 14 und Messen des Prozentsatzes der gesamten peripheren T-Zellen- und Thymocyten-Population, die mit dem Antikörper reagiert.

17. Das Verfahren nach Anspruch 16, worin der Überschuß T-Zellen-ALL oder Null-Zellen-ALL ist.

18. Monoklonaler Maus-Antikörper nach einem der Ansprüche 1 bis 7, oder eine Zusammensetzung nach Anspruch 15, zur Behandlung eines OKT10⁺-Zellen-Überschusses in einem Individuum, das eine solche Behandlung benötigt.

19. Ein Verfahren zur Herstellung einer Zusammensetzung nach Anspruch 14 oder Anspruch 15, umfassend das Vermischen eines diagnostisch oder pharmazeutisch annehmbaren Trägers mit einer diagnostisch oder pharmazeutisch wirksamen Menge von monoklonalem



Maus-Antikörper nach einem der Ansprüche 1 bis 7.

Revendications

1. Anticorps monoclonal de souris qui réagit avec:

- i) environ 95% des thymocytes humains normaux;
- ii) 5% des cellules T périphériques humaines normales;
- iii) 10% des cellules mononucléaires périphériques E⁻ et
- iv) 10—20% des cellules de la moelle osseuse.

2. Anticorps monoclonal de souris selon la revendication 1 qui présente le profil de réactivité vis-à-vis des thymocytes humains normaux illustré par la Figure 1.

3. Anticorps monoclonal de souris selon la revendication 1 ou la revendication 2 qui présente le profil de réactivité vis-à-vis des cellules périphériques T et des thymocytes illustré dans le Tableau 2.

4. Anticorps monoclonal de souris selon l'une quelconque des revendications 1 à 3 qui réagit avec un antigène présent sur les cellules leucémiques de la leucémie lymphoblastique aiguë à cellules T.

5. Anticorps monoclonal de souris selon l'une quelconque des revendications 1 à 4 appartenant à la fraction IgG.

6. Anticorps monoclonal de souris selon l'une quelconque des revendications 1 à 5, produit par un hybridome formé par fusion de cellules d'une lignée de myélome de souris et de cellules spléniques d'une souris préalablement immunisée avec des cellules leucémiques de sujets humains atteints de leucémie lymphoblastique aiguë à cellules T.

7. Anticorps monoclonal de souris produit par l'hybridome ATCC CRL 8022 (OKT10).

8. Un hybridome qui produit un anticorps monoclonal selon l'une des revendications 1 à 6, formé par fusion de cellules spléniques d'une souris préalablement immunisée avec des cellules leucémiques d'un sujet humain atteint de leucémie lymphoblastique aiguë à cellules T et des cellules d'une lignée de myélome de souris.

9. Hybridome ATCC CRL 8022 (OKT10).

10. Un procédé pour la préparation d'un anticorps monoclonal de souris selon l'une quelconque des revendications 1 à 6 qui comprend les stades de:

- i) immunisation de souris avec des cellules leucémiques d'un sujet humain atteint de leucémie lymphoblastique aiguë à cellules T;
- ii) prélèvement des rates desdites souris et formation d'une suspension des cellules spléniques;
- iii) fusion desdites cellules spléniques avec des cellules de myélome de souris en présence d'un promoteur de fusion;

iv) dilution et culture des cellules fusionnées dans des cupules séparées dans un milieu qui n'entretient pas les cellules de myélome non fusionnées;

v) évaluation du surnageant dans chaque cupule contenant un hybridome pour déterminer la présence d'un anticorps ayant les propriétés spécifiées dans l'une quelconque des revendications 1 à 6;

vi) sélection et clonage des hybridomes produisant l'anticorps désiré; et

vii) récupération de l'anticorps dans le surnageant au-dessus desdits clones.

11. Un procédé pour la préparation d'un anticorps monoclonal de souris selon l'une quelconque des revendications 1 à 7 qui comprend les stades:

i) immunisation de souris avec des cellules leucémiques d'un sujet humain atteint de leucémie lymphoblastique aiguë à cellules T;

ii) prélèvement des rates desdites souris et formation d'une suspension des cellules spléniques;

iii) fusion desdites cellules spléniques avec des cellules de myélome de souris en présence d'un promoteur de fusion;

iv) dilution et culture des cellules fusionnées dans des cupules séparées dans un milieu qui n'entretient pas les cellules de myélome non fusionnées;

v) évaluation du surnageant dans chaque cupule contenant un hybridome pour déterminer la présence d'un anticorps ayant les propriétés spécifiées dans l'une quelconque des revendications 1 à 6;

vi) sélection et clonage des hybridomes produisant l'anticorps désiré;

vii) transfert intrapéritonéal desdits clones à des souris; et

viii) récolte du liquide d'ascite maligne ou du sérum desdites souris.

12. Un procédé pour la préparation d'anticorps monoclonal de souris qui comprend la culture de l'hybridome ATCC CRL 8022 dans un milieu approprié et la récupération de l'anticorps dans le surnageant au-dessus dudit hybridome.

13. Un procédé pour la préparation d'un anticorps monoclonal de souris qui comprend l'injection à une souris de l'hybridome ATCC CRL 8022 et la récupération de l'anticorps à partir du liquide d'ascite maligne ou du sérum de ladite souris.

14. Une composition diagnostique pour la détection d'un excès ou d'un défaut de cellules OKT10⁺ comprenant, en mélange avec un support convenant au diagnostic, une quantité d'un anticorps monoclonal de souris selon l'une quelconque des revendications 1 à 7, efficace pour détecter un excès ou un défaut de cellules OKT10⁺.

15. Une composition thérapeutique comprenant, en mélange avec un support acceptable en pharmacie, une quantité d'un anticorps mono-



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clonal de souris selon l'une quelconque des revendications 1 à 7, efficace pour réduire la quantité des cellules OKT10⁺ chez un sujet présentant un excès desdites cellules OKT10⁺.

16. Un procédé pour la détection d'un excès ou d'un défaut de cellules OKT10⁺ chez un sujet, qui comprend la réaction d'une composition de cellules T ou de thymocytes dudit sujet avec une quantité efficace pour le diagnostic d'un anticorps monoclonal de souris selon l'une quelconque des revendications 1 à 7 ou d'une composition selon la revendication 14 et la mesure du pourcentage de la population totale des cellules périphériques T et des thymocytes réagissant avec ledit anticorps.

17. Le procédé de la revendication 16 dans

lequel l'excès est constitué de cellules T de leucémie lymphoblastique aiguë ou de cellules nulles de leucémie lymphoblastique aiguë.

18. Anticorps monoclonal de souris selon l'une quelconque des revendications 1 à 7 ou composition selon la revendication 15 pour le traitement d'un excès de cellules OKT10⁺ chez un sujet nécessitant un tel traitement.

19. Un procédé pour préparer une composition selon la revendication 14 ou la revendication 15 comprenant le mélange d'un support acceptable pour le diagnostic ou acceptable en pharmacie avec une quantité efficace pour le diagnostic ou efficace en pharmacie d'un anticorps monoclonal de souris selon l'une quelconque des revendications 1 à 7.

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