

DRUG REGULATED TRANSGENE EXPRESSION**CROSS-REFERENCE TO RELATED APPLICATIONS**

[0001] The present application claims the benefit of priority to U.S. Provisional Patent Application No. 62/058,973, filed October 2, 2014, U.S. Provisional Patent Application No. 61/977,751, filed April 10, 2014, U.S. Provisional Patent Application No. 61/986,479, filed April 30, 2014, U.S. Provisional Patent Application No. 62/089,730 filed December 9, 2014, U.S. Provisional Patent Application No. 62/090845, filed December 11, 2014, and U.S. Provisional Patent Application No. 62/088,363, filed December 5, 2014. The entire disclosures of the aforementioned applications are expressly incorporated by reference in their entireties.

REFERENCE TO SEQUENCE LISTING, TABLE, OR COMPUTER PROGRAM LISTING

[0002] The present application is being filed along with a Sequence Listing in electronic format. The Sequence Listing is provided as a file entitled SCRI-53WO_SEQUENCE_LISTING.TXT, created April 7, 2015, which is 65kb in size. The information in the electronic format of the Sequence Listing is incorporated herein by reference in its entirety.

BACKGROUND OF THE INVENTION

[0003] The adoptive transfer of human T lymphocytes that are engineered by gene transfer to express chimeric antigen receptors (CARs) specific for surface molecules expressed on tumor cells has the potential to effectively treat cancer. Chimeric receptors are synthetic receptors that include an extracellular ligand binding domain, most commonly a single chain variable fragment of a monoclonal antibody (scFv) linked to intracellular signaling components, most commonly CD3 ζ alone or combined with one or more costimulatory domains. Much of the research in the design of chimeric receptors has focused on defining scFvs and other ligand binding elements that target malignant cells without causing serious toxicity to essential normal tissues, and on defining the optimal composition of intracellular signaling modules to activate T cell effector functions.

[0004] Although, T cell (CAR-T) adoptive therapy clinical trials (chimeric antigen receptor expressing T cells) are demonstrating potent anti-tumor activity, it is apparent that

significant toxicities can arise, for example, engraftment-induced cytokine storm, tumor lysis syndromes and ongoing B cell cytopenias, each of which are attributable to unregulated functional outputs of constitutively expressed CARs. Such toxicities can in some context threaten to limit the applicability of CAR-T cell adoptive therapy. Clinical trials using transgene-modified adoptive T cell immunotherapies have only tested T cells that constitutively express the transgene, or are always in the “ON” state, contributing in large part to transgene associated side-effects. Suicide gene-mediated elimination of CAR-T cells can ameliorate such toxicities; however, this approach risks premature attenuation of anti-tumor activity and significantly impacts curative potential.

[0005] Current small molecule-regulated transgene expression technologies rely on a variety of drug inputs including macrolides, ecdysones and rapamycin analogs. Clinical applicability of these systems is limited due to toxic off target effects, unfavorable biodistribution and pharmacodynamics profiles, limited output dynamic range, and/or limited availability as FDA-approved commercially available pharmaceuticals. Furthermore, many of these systems use chimeric transcriptional regulators built from xenogeneic components, thus introducing the complication of immunogenicity when applying these systems to human therapeutics.

[0006] There is a need to identify methods for determining elements of chimeric receptor design that are important for therapeutic activity and cell populations to genetically modify and adoptively transfer that will provide enhanced survival and efficacy *in vivo* while minimizing adverse side effects. There is also a need for expression systems and methods for modulating cells for use in cell therapy, such as for modulating expression of recombinant antigen receptors such as CARs and/or other molecules expressed by such cells, such as to improve therapeutic activity, enhanced survival and/or efficacy *in vivo* and/or minimize adverse side effects.

SUMMARY OF THE INVENTION

[0007] One aspect of the disclosure includes a genetic system to deliver drug-regulated transgene expression in cells. In an alternative, regulated transgene expression is targeted to cells, such as lymphocytes designed for use in adoptive immunotherapy. This system provides rigorous safety attributes to chimeric antigen receptor (CAR) redirected adoptive therapeutic strategies without sacrificing curative intent that permits real-time clinician control of CAR expression *in vivo*. By engineering vectors that enable drug responsive transcriptional control of CAR expression, the activity of CARs and other cell

mediators can be turned “ON” and “OFF” *in vivo*, based on a clinician prescribed pharmaceutical drug input that exhibits clinically permissive pharmacokinetics, tissue distribution, and partitioning between the extracellular space and cytosol of lymphocytes. The genetic system provides for drug regulated transgene expression to enforce a functional “OFF” state in the absence of the drug and a functional “ON” state transgene expression in the presence of the drug.

[0008] One alternative of such a drug is tamoxifen. Tamoxifen is an estrogen antagonist/partial agonist that is an FDA-approved and commercially available drug. It is taken orally and can be administered on a daily basis over an extended period of time. Tamoxifen has a proven safety record, favorable pharmacokinetic profile, excellent tissue distribution and a low partition coefficient between the extracellular space and cytosol. Other drugs can be selected based on safety record, favorable pharmacokinetic profile, and excellent tissue distribution, a low partition coefficient between the extracellular space and cytosol, and/or low toxicities.

[0009] In some alternatives, the system employs a synthetic transcriptional regulator, which, in the presence of tamoxifen, binds a synthetic promoter upstream of a transgene to induce expression. The tamoxifen regulated transcription factor (“TamR-tf”, also designated “HEA3”) is a chimeric transcription factor composed of human subunits including the N-terminal DNA binding domain of Hepatocyte Nuclear Factor 1-alpha (HNF-1 α) fused in frame to the mutant tamoxifen-specific ligand binding domain of the estrogen receptor ligand binding domain (ER-LBD), that is in turn fused to the p65 activation domain of NF- κ B (p65). An exemplary amino acid sequence is provided in Table 10 and is identified as SEQ ID NO: 40. The mutant tamoxifen-specific ligand binding domain of the estrogen receptor ligand binding domain (ER-LBD) is found at amino acids 282-595 of the TamR-tf and has a mutation at position 521. The p65 activation domain of NF- κ B (p65) is found at amino acids 596-862 of SEQ ID NO: 40. Further changes can be made to the transcriptional activator to increase the properties of the transcription factor including, without limitation, altering one or more amino acids in the estrogen receptor ligand binding domain to increase the affinity of the factor for estrogen analogs and altering one or more amino acids in the p65 transactivating domain.

[0010] In the absence of tamoxifen, TamR-tf is excluded from the nucleus by binding of cytosolic heat-shock protein 90 (HSP90) to the tamoxifen binding active site and transgene expression is in the “OFF” state. Nanomolar concentrations of cytosolic tamoxifen

actively out competes HSP90 for ER-LBD binding, resulting in TamR-tf translocation to the nucleus. Upon nuclear translocation, TamR-tf is readily available to bind its restricted synthetic promoter (e.g. 7xHBD/EF1 α p). In the presence of tamoxifen, binding of TamR-tf to 7xHBD/EF1 α p promoter induces the “ON” state of transgene expression. In some alternatives, this transcriptional regulator can be modified to provide for a varying level of control of transgene expression. Amino acid substitutions in the LBD of TamR-tf permit selective responsiveness to tamoxifen and its metabolites, where 4-hydroxy tamoxifen (4-OHT) is the most pharmacologically active metabolite, in regards to TamR-tf activity, while lacking interaction with endogenous estrogen.

[0011] In one aspect, the present disclosure relates to methods and compositions to confer and/or augment immune responses mediated by cellular immunotherapy, such as, by adoptively transferring tumor-specific, genetically modified subsets of CD8+ or CD4+ T cells alone, or in combination. The disclosure provides for chimeric receptor nucleic acids, and vectors and host cells including such nucleic acids. The nucleic acid sequence that encodes the chimeric receptor links together a number of modular components that can be excised and replaced with other components in order to customize the chimeric receptor for efficient T cell activation and recognition of a specific target molecule or an epitope on the target molecule and provide for regulated transcription as described herein.

[0012] In some alternatives, a system for inducible expression of a chimeric antigen receptor comprises: a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain; and d) a polynucleotide coding for an intracellular signaling domain; and a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the drug is tamoxifen and/or its metabolites. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor. In some alternatives, the second promoter is constitutive or inducible.

[0013] In some alternatives, a system for inducible expression of a chimeric antigen receptor comprises: a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a cytokine, a chemokine, a polypeptide that regulates apoptosis and/or a polypeptide that modulates checkpoint signaling; and a second nucleic acid comprising a second promoter operably linked to a nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the second promoter is constitutive or inducible. In an exemplary alternative, the second nucleic acid further comprises a polynucleotide coding for a chimeric antigen receptor, under the control of a constitutive promoter. In some alternatives, the drug is tamoxifen and/or its metabolites.

[0014] In another aspect, the present disclosure provides compositions to confer and/or augment immune responses mediated by cellular immunotherapy, such as by adoptively transferring tumor-specific, subset specific genetically modified CD4⁺ T cells, wherein the CD4⁺ T cells confer and/or augment the ability of CD8⁺ T cells to sustain anti-tumor reactivity and increase and/or maximize tumor-specific proliferation. In some alternatives, the CD4⁺ cells are genetically modified to express a chimeric receptor nucleic acid and/or chimeric receptor polypeptide under the control of a regulated promoter, as described herein.

[0015] In another aspect, the present disclosure provides compositions to confer and/or augment immune responses mediated by cellular immunotherapy, such as by adoptively transferring tumor-specific, subset specific genetically modified T cells. In some alternatives, the cells are precursor T cells. In some alternatives, the cells are hematopoietic stem cells. In some alternatives, the cells are CD8⁺ T cells. In some alternatives, the CD8⁺ T cells express a chimeric receptor nucleic acid and/or chimeric receptor polypeptide under the control of a regulated promoter, as described herein.

[0016] Some alternatives concern methods of performing cellular immunotherapy in a subject having a disease or disorder by administering to the subject a genetically modified T lymphocyte cell preparation that provides a cellular immune response and administering a drug that induces a transgene in the genetically modified T lymphocyte cells. In some alternatives, the genetically modified T lymphocyte cell preparation comprises precursor T cells. In some alternatives, the genetically modified T lymphocyte cell preparation comprises hematopoietic stem cells. In some alternatives, the genetically modified CD8⁺ and genetically modified CD4⁺ cell population are co-administered. In some alternatives, the drug is

tamoxifen and/or its metabolites. In some alternatives, the T cells are autologous or allogeneic T cells. Various modifications of the above method are possible. For example, the chimeric receptor that is expressed by the CD4⁺ T cell and the CD8⁺ T cell can be the same or different.

[0017] In some alternatives, a system for inducible expression of a chimeric antigen receptor is provided, wherein the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain; and d) a polynucleotide coding for an intracellular signaling domain; and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the drug is tamoxifen and/or its metabolites. In some alternatives, the second promoter is inducible. In some alternatives, the second promoter is constitutive. In some alternatives, the first promoter comprises a nucleic acid sequence of SEQ ID NO: 41. In some alternatives, the second promoter is a constitutive promoter. In some alternatives, the second promoter is the EF1 α p. In some alternatives, the transcriptional activator comprises a sequence of SEQ ID NO: 40. In some alternatives, the first nucleic acid further comprises a first vector and the second nucleic acid further comprises a second vector. In some alternatives, both vectors are packaged in a viral vector. In some alternatives, the viral vector is a lentivirus. In some alternatives, the first and second nucleic acid comprise a vector. In some alternatives, the first nucleic acid further comprises a nucleic acid sequence coding for a selectable marker. In some alternatives, the second nucleic acid further comprises a nucleic acid coding for a selectable marker. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor.

[0018] In some alternatives, a system for inducible expression of a chimeric antigen receptor is provided, wherein the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain,

wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized a polynucleotide coding for a transmembrane domain; and d) a polynucleotide coding for an intracellular signaling domain and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter.. In some alternatives, the second promoter is constitutive or inducible. In some alternatives, the drug is tamoxifen and/or its metabolites. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the drug is tamoxifen and/or its metabolites. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment. In some alternatives, the ligand binding domain is single chain variable fragment. In some alternatives, the tumor specific molecule is CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, or MAGE A3 TCR or combinations thereof. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor. In some alternatives, the second promoter is inducible. In some alternatives, the second promoter is constitutive.

[0019] In some alternatives, a chimeric receptor polypeptide is provided, wherein the chimeric receptor polypeptide is coded for by a system. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide

spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain; and d) a polynucleotide coding for an intracellular signaling domain; and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the second promoter is constitutive or inducible. In some alternatives, the drug is tamoxifen and/or its metabolites. In some alternatives, the first promoter comprises a nucleic acid sequence of SEQ ID NO: 41. In some alternatives, the second promoter is a constitutive promoter. In some alternatives, the second promoter is the EF1 α p. In some alternatives, the transcriptional activator comprises a sequence of SEQ ID NO: 40. In some alternatives, the first nucleic acid further comprises a first vector and the second nucleic acid further comprises a second vector. In some alternatives, both vectors are packaged in a viral vector. In some alternatives, the viral vector is a lentivirus. In some alternatives, the first and second nucleic acid comprise a vector. In some alternatives, the first nucleic acid further comprises a nucleic acid sequence coding for a selectable marker. In some alternatives, the second nucleic acid further comprises a nucleic acid coding for a selectable marker. In some alternatives, the second promoter is an inducible promoter. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor.

[0020] In some alternatives, a system for inducible expression of chimeric antigen receptor is provided, wherein the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a cytokine, a chemokine receptor, a polypeptide that regulates apoptosis, or a polypeptide that modulates checkpoint signaling and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the second promoter is constitutive or inducible. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some

alternatives, the ligand binding domain is an antibody fragment, preferably a binding domain thereof. In some alternatives, the tumor specific molecule is CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, or MAGE A3 TCR or combinations thereof. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a cytokine, a chemokine receptor, a polypeptide that regulates apoptosis, or a polypeptide that modulates checkpoint signaling; and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the second promoter is constitutive or inducible. In some alternatives, the drug is tamoxifen and/or its metabolites. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for chimeric antigen receptor comprising a ligand binding domain, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment, preferably a binding fragment thereof. In some alternatives, the ligand binding domain is single chain variable fragment. In some alternatives, the tumor specific molecule is CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, or MAGE A3 TCR or combinations thereof. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor.

[0021] In some alternatives, a host cell is provided, wherein the host cell comprises a system. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a

transmembrane domain; and d) a polynucleotide coding for an intracellular signaling domain; and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the second promoter is constitutive or inducible. In some alternatives, the drug is tamoxifen and/or its metabolites. In some alternatives, the first promoter comprises a nucleic acid sequence of SEQ ID NO: 41. In some alternatives, the second promoter is a constitutive promoter. In some alternatives, the second promoter is the EF1 α p. In some alternatives, the transcriptional activator comprises a sequence of SEQ ID NO: 40. In some alternatives, the first nucleic acid further comprises a first vector and the second nucleic acid further comprises a second vector. In some alternatives, both vectors are packaged in a viral vector. In some alternatives, the viral vector is a lentivirus. In some alternatives, the first and second nucleic acid comprise a vector. In some alternatives, the first nucleic acid further comprises a nucleic acid sequence coding for a selectable marker. In some alternatives, the second nucleic acid further comprises a nucleic acid coding for a selectable marker. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment, preferably a binding fragment thereof. In some alternatives, the tumor specific molecule is CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, or MAGE A3 TCR or combinations thereof. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a cytokine, a chemokine receptor, a polypeptide that regulates apoptosis, or a polypeptide that modulates checkpoint signaling; and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the second promoter is constitutive or inducible. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for chimeric antigen receptor comprising a ligand binding domain, wherein the ligand is a tumor specific molecule, viral

molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment, preferably a binding fragment thereof. In some alternatives, the ligand binding domain is single chain variable fragment. In some alternatives, the tumor specific molecule is CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, or MAGE A3 TCR or combinations thereof. In some alternatives, the host cell is a CD8+ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8+ T cells, central memory CD8+ T cells, effector memory CD8+ T cells and bulk CD8+ T cells. In some alternatives, the host cell is a CD4+ T helper lymphocyte cell that is selected from the group consisting of naïve CD4+ T cells, central memory CD4+ T cells, effector memory CD4+ T cells, and bulk CD4+ T cells. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor. In some alternatives, the host cell is precursor T cell. In some alternatives, the precursor T cell is a hematopoietic stem cell.

[0022] In some alternatives, a composition is provided, wherein the composition comprises a host cell in a pharmaceutically acceptable excipient. In some alternatives, the host cell comprises a system. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain; and d) a polynucleotide coding for an intracellular signaling domain; and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the second promoter is constitutive or inducible. In some alternatives, the drug is tamoxifen and/or its metabolites. In some alternatives, the first promoter comprises a nucleic acid sequence of SEQ ID NO: 41. In some alternatives, the second promoter is a constitutive

promoter. In some alternatives, the second promoter is the EF1 α p. In some alternatives, the transcriptional activator comprises a sequence of SEQ ID NO: 40. In some alternatives, the first nucleic acid further comprises a first vector and the second nucleic acid further comprises a second vector. In some alternatives, both vectors are packaged in a viral vector. In some alternatives, the viral vector is a lentivirus. In some alternatives, the first and second nucleic acid comprise a vector. In some alternatives, the first nucleic acid further comprises a nucleic acid sequence coding for a selectable marker. In some alternatives, the second nucleic acid further comprises a nucleic acid coding for a selectable marker. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment. In some alternatives, the tumor specific molecule is CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, or MAGE A3 TCR or combinations thereof. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a cytokine, a chemokine receptor, a polypeptide that regulates apoptosis, or a polypeptide that modulates checkpoint signaling; and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the second promoter is constitutive or inducible. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment, preferably a binding

fragment thereof. In some alternatives, the ligand binding domain is single chain variable fragment. In some alternatives, the tumor specific molecule is CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, or MAGE A3 TCR or combinations thereof. In some alternatives, the host cell is precursor T cell. In some alternatives, the precursor T cell is a hematopoietic stem cell. In some alternatives, the host cell is a CD8⁺ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8⁺ T cells, central memory CD8⁺ T cells, effector memory CD8⁺ T cells and bulk CD8⁺ T cells. In some alternatives, the host cell is a CD4⁺ T helper lymphocyte cell that is selected from the group consisting of naïve CD4⁺ T cells, central memory CD4⁺ T cells, effector memory CD4⁺ T cells, and bulk CD4⁺ T cells. In some alternatives, the composition comprises a host cell wherein the host cell is a CD8⁺ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8⁺ T cells, central memory CD8⁺ T cells, effector memory CD8⁺ T cells and bulk CD8⁺ T cells and further comprises another host cell wherein the host cell is a CD4⁺ T helper lymphocyte cell that is selected from the group consisting of naïve CD4⁺ T cells, central memory CD4⁺ T cells, effector memory CD4⁺ T cells, and bulk CD4⁺ T cells. In some alternatives, the host cell is precursor T cell. In some alternatives, the precursor T cell is a hematopoietic stem cell. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor. In some alternatives, the composition comprises a host cell wherein the host cell is a CD8⁺ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8⁺ T cells, central memory CD8⁺ T cells, effector memory CD8⁺ T cells and bulk CD8⁺ T cells or the host cell is a CD4⁺ T helper lymphocyte cell that is selected from the group consisting of naïve CD4⁺ T cells, central memory CD4⁺ T cells, effector memory CD4⁺ T cells, and bulk CD4⁺ T cells and a second host cell, wherein the second host cell is a precursor T cell or a hematopoietic stem cell.

[0023] In some alternatives, an *in vitro* method for preparing a host cell is provided wherein the method comprises a) providing a system and b) introducing the system into a separate isolated T lymphocyte population and expanding each T lymphocyte population *in vitro*. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a

tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain; and d) a polynucleotide coding for an intracellular signaling domain; and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the second promoter is constitutive or inducible. In some alternatives, the drug is tamoxifen and/or its metabolites. In some alternatives, the second promoter is constitutive or inducible. In some alternatives, the first promoter comprises a nucleic acid sequence of SEQ ID NO: 41. In some alternatives, the second promoter is a constitutive promoter. In some alternatives, the second promoter is the EF1 α p. In some alternatives, the transcriptional activator comprises a sequence of SEQ ID NO: 40. In some alternatives, the first nucleic acid further comprises a first vector and the second nucleic acid further comprises a second vector. In some alternatives, both vectors are packaged in a viral vector. In some alternatives, the viral vector is a lentivirus. In some alternatives, the first and second nucleic acid comprise a vector. In some alternatives, the first nucleic acid further comprises a nucleic acid sequence coding for a selectable marker. In some alternatives, the second nucleic acid further comprises a nucleic acid coding for a selectable marker. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment, preferably a binding fragment thereof. In some alternatives, the tumor specific molecule is CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, or MAGE A3 TCR or combinations thereof. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a cytokine, a chemokine receptor, a polypeptide that regulates apoptosis, or a

polypeptide that modulates checkpoint signaling; and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the second promoter is constitutive or inducible. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for chimeric antigen receptor comprising a ligand binding domain, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment, preferably a binding fragment thereof. In some alternatives, the ligand binding domain is single chain variable fragment. In some alternatives, the tumor specific molecule is CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, or MAGE A3 TCR or combinations thereof. In some alternatives, wherein the T lymphocytes are expanded, the method further comprises culturing the cells in the presence of anti-CD3 and/or anti CD28, and at least one homeostatic cytokine until the cells expand sufficiently for use as a cell infusion. In some alternatives, the lymphocyte is CD8⁺ or CD4⁺. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor. In some alternatives, the host cell is precursor T cell. In some alternatives, the precursor T cell is a hematopoietic stem cell.

[0024] In some alternatives, a use of a host cell or a composition in combination with a drug that induces expression of a transgene in the host cell or composition for the treatment of cancer or a viral infection is provided. In some alternatives, the host cell comprises a system. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain; and d) a

polynucleotide coding for an intracellular signaling domain; and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the second promoter is constitutive or inducible. In some alternatives, the drug is tamoxifen and/or its metabolites. In some alternatives, the first promoter comprises a nucleic acid sequence of SEQ ID NO: 41. In some alternatives, the second promoter is a constitutive promoter. In some alternatives, the second promoter is the EF1 α p. In some alternatives, the transcriptional activator comprises a sequence of SEQ ID NO: 40. In some alternatives, the first nucleic acid further comprises a first vector and the second nucleic acid further comprises a second vector. In some alternatives, both vectors are packaged in a viral vector. In some alternatives, the viral vector is a lentivirus. In some alternatives, the first and second nucleic acid comprise a vector. In some alternatives, the first nucleic acid further comprises a nucleic acid sequence coding for a selectable marker. In some alternatives, the second nucleic acid further comprises a nucleic acid coding for a selectable marker. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment. In some alternatives, the tumor specific molecule is CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, or MAGE A3 TCR or combinations thereof. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a cytokine, a chemokine receptor, a polypeptide that regulates apoptosis, or a polypeptide that modulates checkpoint signaling; and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the second promoter is constitutive or inducible. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for chimeric antigen receptor comprising a ligand binding domain, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell

population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment, preferably a binding fragment thereof. In some alternatives, the ligand binding domain is single chain variable fragment. In some alternatives, the tumor specific molecule is CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, or MAGE A3 TCR or combinations thereof. In some alternatives, the host cell is a CD8⁺ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8⁺ T cells, central memory CD8⁺ T cells, effector memory CD8⁺ T cells and bulk CD8⁺ T cells. In some alternatives, the host cell is a CD4⁺ T helper lymphocyte cell that is selected from the group consisting of naïve CD4⁺ T cells, central memory CD4⁺ T cells, effector memory CD4⁺ T cells, and bulk CD4⁺ T cells. In some alternatives, the cells are precursor T cells. In some alternatives, the cells are hematopoietic stem cells. In some alternatives, the composition comprises a host cell in a pharmaceutically acceptable excipient. In some alternatives, the host cell comprises a system. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the second promoter is constitutive or inducible. In some alternatives, the drug is tamoxifen and/or its metabolites. In some alternatives, the first promoter comprises a nucleic acid sequence of SEQ ID NO: 41. In some alternatives, the second promoter is a constitutive promoter. In some alternatives, the second promoter is the EF1 α p. In some alternatives, the transcriptional activator comprises a sequence of SEQ ID NO: 40. In some alternatives, the first nucleic acid further comprises a first vector and the second nucleic acid further comprises a second vector. In some alternatives, both vectors are packaged in a viral vector. In some alternatives, the viral vector is a lentivirus. In some alternatives, the first and

second nucleic acid comprise a vector. In some alternatives, the first nucleic acid further comprises a nucleic acid sequence coding for a selectable marker. In some alternatives, the second nucleic acid further comprises a nucleic acid coding for a selectable marker. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment, preferably a binding fragment thereof. In some alternatives, the tumor specific molecule is CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, or MAGE A3 TCR or combinations thereof. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a cytokine, a chemokine receptor, a polypeptide that regulates apoptosis, or a polypeptide that modulates checkpoint signaling; and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the second promoter is constitutive or inducible. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment, preferably a binding fragment thereof. In some alternatives, the ligand binding domain is single chain variable fragment. In some alternatives, the tumor specific molecule is CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, or MAGE A3 TCR or combinations thereof. In some alternatives, the host cell is a CD8⁺ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8⁺ T cells, central

memory CD8+ T cells, effector memory CD8+ T cells and bulk CD8+ T cells. In some alternatives, the host cell is a CD4+ T helper lymphocyte cell that is selected from the group consisting of naïve CD4+ T cells, central memory CD4+ T cells, effector memory CD4+ T cells, and bulk CD4+ T cells. In some alternatives, the composition comprises a host cell wherein the host cell is a CD8+ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8+ T cells, central memory CD8+ T cells, effector memory CD8+ T cells and bulk CD8+ T cells and another host cell wherein the host cell is a CD4+ T helper lymphocyte cell that is selected from the group consisting of naïve CD4+ T cells, central memory CD4+ T cells, effector memory CD4+ T cells, and bulk CD4+ T cells. In some alternatives, the cancer is a solid tumor or hematologic malignancy. In some alternatives, the solid tumor is selected from the group consisting of a breast cancer, brain cancer, lung cancer, colon cancer, renal cancer, pancreatic cancer, prostate cancer, and ovarian cancer. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor. In some alternatives, the host cell is precursor T cell. In some alternatives, the precursor T cell is a hematopoietic stem cell. In some alternatives, the host cell is precursor T cell. In some alternatives, the precursor T cell is a hematopoietic stem cell. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor. In some alternatives, the composition comprises a host cell wherein the host cell is a CD8+ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8+ T cells, central memory CD8+ T cells, effector memory CD8+ T cells and bulk CD8+ T cells or the host cell is a CD4+ T helper lymphocyte cell that is selected from the group consisting of naïve CD4+ T cells, central memory CD4+ T cells, effector memory CD4+ T cells, and bulk CD4+ T cells and a second host cell, wherein the second host cell is a precursor T cell or a hematopoietic stem cell.

[0025] In some alternatives, a method of performing cellular immunotherapy in a subject having cancer or a viral infection is provided wherein the method comprises administering a composition or a host cell to the subject and administering a drug that induces expression of a transgene in the composition or the host cells. In some alternatives, the host cell comprises a system. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a

tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain; and d) a polynucleotide coding for an intracellular signaling domain; and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the second promoter is constitutive or inducible. In some alternatives, the drug is tamoxifen and/or its metabolites. In some alternatives, the first promoter comprises a nucleic acid sequence of SEQ ID NO: 41. In some alternatives, the second promoter is a constitutive promoter. In some alternatives, the second promoter is the EF1 α p. In some alternatives, the transcriptional activator comprises a sequence of SEQ ID NO: 40. In some alternatives, the first nucleic acid further comprises a first vector and the second nucleic acid further comprises a second vector. In some alternatives, both vectors are packaged in a viral vector. In some alternatives, the viral vector is a lentivirus. In some alternatives, the first and second nucleic acid comprise a vector. In some alternatives, the first nucleic acid further comprises a nucleic acid sequence coding for a selectable marker. In some alternatives, the second nucleic acid further comprises a nucleic acid coding for a selectable marker. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment, preferably a binding fragment thereof. In some alternatives, the tumor specific molecule is CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, or MAGE A3 TCR or combinations thereof. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a cytokine, a chemokine receptor, a polypeptide that regulates apoptosis, or a polypeptide that modulates checkpoint signaling; and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid

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transcriptional activator comprises a sequence of SEQ ID NO: 40. In some alternatives, the first nucleic acid further comprises a first vector and the second nucleic acid further comprises a second vector. In some alternatives, both vectors are packaged in a viral vector. In some alternatives, the viral vector is a lentivirus. In some alternatives, the first and second nucleic acid comprise a vector. In some alternatives, the first nucleic acid further comprises a nucleic acid sequence coding for a selectable marker. In some alternatives, the second nucleic acid further comprises a nucleic acid coding for a selectable marker. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment, preferably a binding fragment thereof. In some alternatives, the tumor specific molecule is CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, or MAGE A3 TCR or combinations thereof. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a cytokine, a chemokine receptor, a polypeptide that regulates apoptosis, or a polypeptide that modulates checkpoint signaling; and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the second promoter is constitutive or inducible. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment, preferably a binding fragment thereof. In some alternatives, the ligand binding domain is single chain variable fragment. In some

alternatives, the tumor specific molecule is CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, or MAGE A3 TCR or combinations thereof. In some alternatives, the host cell is a CD8+ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8+ T cells, central memory CD8+ T cells, effector memory CD8+ T cells and bulk CD8+ T cells. In some alternatives, the host cell is a CD4+ T helper lymphocyte cell that is selected from the group consisting of naïve CD4+ T cells, central memory CD4+ T cells, effector memory CD4+ T cells, and bulk CD4+ T cells. In some alternatives, the composition comprises a host cell wherein the host cell is a CD8+ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8+ T cells, central memory CD8+ T cells, effector memory CD8+ T cells and bulk CD8+ T cells and another host cell wherein the host cell is a CD4+ T helper lymphocyte cell that is selected from the group consisting of naïve CD4+ T cells, central memory CD4+ T cells, effector memory CD4+ T cells, and bulk CD4+ T cells. In some alternatives, the host cell is precursor T cell. In some alternatives, the precursor T cell is a hematopoietic stem cell. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor. In some alternatives, the composition comprises a host cell wherein the host cell is a CD8+ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8+ T cells, central memory CD8+ T cells, effector memory CD8+ T cells and bulk CD8+ T cells or the host cell is a CD4+ T helper lymphocyte cell that is selected from the group consisting of naïve CD4+ T cells, central memory CD4+ T cells, effector memory CD4+ T cells, and bulk CD4+ T cells and a second host cell, wherein the second host cell is a precursor T cell or a hematopoietic stem cell. In some alternatives, the cancer is selected from a solid tumor or hematologic malignancy. In some alternatives, the solid tumor is selected from the group consisting of a breast cancer, brain cancer, lung cancer, colon cancer, renal cancer, pancreatic cancer, prostate cancer, and ovarian cancer. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor. In some alternatives, the host cell is a precursor T cell. In some alternatives, the precursor T cell is a hematopoietic stem cell. In some alternatives, the isolated T lymphocyte population comprises precursor T cells. In some alternatives, the precursor T cells are hematopoietic stem cells. In some alternatives, the administering of the drug is performed after administering of the composition or host cells, wherein administering is performed 1

day, 2 days, 3 days, 4 days, 5 days, 6 days, 7 days, 2 weeks, 4 weeks or two months, or any time in between any two values of time listed.

BRIEF DESCRIPTION OF THE DRAWINGS

[0026] **Figure 1A** shows the expression of ZsGreen and EGFRt as determined by flow cytometry in Jurkat cells transduced with a dual plasmid lentiviral construct including constructs A and B in the presence or absence of 4 hydroxy tamoxifen (4OHT). The results show the expression of EGFRt in the presence or absence of 4-OHT indicating the cells carry construct A. The results also show that the cells carry construct B as expression of ZsGreen is induced in the presence of 4-OHT. **Figure 1B** shows construct A, which comprises the constitutive promoter EF1 α p linked to TamR-tf (HEA3) linked to EGFRt, and construct B comprising a synthetic promoter 7xHBD/mE1b linked to a polynucleotide coding for ZsGreen.

[0027] **Figure 2** shows the expression of ZsGreen in transduced Jurkat cells contacted with different doses of 4-OHT ranging from 50 to 1000nM.

[0028] **Figure 3** shows the on and off rate kinetics of expression of ZsGreen in transduced Jurkat cells with a single 48 hour treatment of 4-OHT followed by a washout ($\circ$) and transduced Jurkat cells with a 24 hour treatment of 4-OHT followed by washout and then a re-stimulation with 4-OHT at day 15 ($\blacksquare$).

[0029] **Figure 4A-D** shows the expression of ZsGreen in CD4 central memory cells transduced with dual package lentiviral constructs A and B. Cells were divided into 3 treatment groups, 4OHT alone (**Figure 4A**), 4OHT combined with CD3/CD28 bead co-treatment (**Figure 4B**) or 4OHT alone for 48 hours, followed by addition of CD3/CD28 beads (**Figure 4C**). Expression of ZsGreen was monitored over 96 hours. **Figure 4D** shows construct A, which comprises the constitutive promoter EF1 α p linked to TamR-tf (HEA3) linked to EGFRt, and construct B comprising a synthetic promoter 7xHBD/mE1b linked to a polynucleotide coding for ZsGreen.

[0030] **Figures 5A-C** shows the expression of ZsGreen in CD8 central memory cells transduced with dual package lentiviral constructs A and B as shown in **Figure 4**. Cells were divided into 3 treatment groups, 4OHT alone (**Figures 5A**), 4OHT combined with CD3/CD28 bead co-treatment (**Figures 5B**) or 4OHT alone for 48 hours, followed by addition of CD3/CD28 beads (**Figures 5C**). Expression of ZsGreen was monitored over 72 hours.

[0031] **Figure 6A** shows expression of EGFRt and Her2t in Human Jurkat T cells transduced with constructs A and B as shown in **Figure 6C**. Expression of EGFRt and Her2t were monitored in the presence or absence of 4OHT. Samples were stained with EGFRt-APC antibody and Herceptin-biotin, followed by SA-PE. **Figure 6B** shows a western blot of parental Jurkat cells and transduced CD19CAR transduced cells stained with mouse antiCD247 which recognizes the intracellular CD3 zeta chain on CD19 CAR (about 48 kDA), the endogenous CD3 zeta chain has migrates at 23 kDA. The endogenous CD3 zeta chain was detected in all of the cells. The CD19 CAR CD3 zeta chain was only detected in transduced Jurkat cells exposed to 4-OHT. **Figure 6C** shows construct A, which comprises the constitutive promoter EF1 α p linked to TamR-tf (HEA3) linked to EGFRt, and construct B comprising a synthetic promoter 7xHBD/mE1b linked to a polynucleotide coding for CD19CAR linked to a polynucleotide coding for Her2t.

[0032] **Figure 7A** shows expression of EGFRt and Her2t in Human Jurkat T cells transduced with TamR CD19CAR LV including an additional selective marker, DHFRdm in the presence or absence of 4OHT. Expression of EGFRt and Her2t were monitored in the presence or absence of 4OHT. Samples were stained with EGFRt-APC antibody and Herceptin-biotin, followed by SA-PE. **Figure 7B** shows a western blot of parental Jurkat cells and transduced CD19CAR transduced cells stained with mouse antiCD247 which recognizes the intracellular CD3 zeta chain on CD19 CAR (about 48 kDA), the endogenous CD3 zeta chain has migrates at 23 kDA. The endogenous CD3 zeta chain was detected in all of the cells. The CD19 CAR CD3 zeta chain was only detected in transduced Jurkat cells exposed to 4-OHT. **Figure 7C** shows construct A, which comprises the constitutive promoter EF1 α p linked to TamR-tf(HEA3) linked to EGFRt, and construct B comprising a synthetic promoter 7xHBD/mE1b linked to a polynucleotide coding for CD19CAR linked to a polynucleotide coding for Her2t linked to a polynucleotide coding for DHFRdm.

[0033] **Figure 8A** shows expression of EGFRt and Her2t in Human CD4 central memory T cells transduced with TamR CD19CAR LV including an additional selective marker, DHFRdm in the presence or absence of 4OHT and antiCD3/CD28 beads. Expression of EGFRt and Her2t were monitored in the presence or absence of 4OHT and in the presence or absence of antiCD3/CD28 beads. Samples were stained with EGFRt-APC antibody and Herceptin-biotin, followed by SA-PE. **Figure 8B** shows construct A, which comprises the constitutive promoter EF1 α p linked to TamR-tf(HEA3) linked to EGFRt, and construct B comprising a synthetic promoter 7xHBD/mE1b linked to a polynucleotide coding for

CD19CAR linked to a polynucleotide coding for Her2t linked to a polynucleotide coding for DHFRdm.

DETAILED DESCRIPTION

Definitions.

[0034] Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which the invention pertains.

[0035] "About" as used herein when referring to a measurable value is meant to encompass variations of $\pm 20\%$ or $\pm 10\%$, more preferably $\pm 5\%$, even more preferably $\pm 1\%$, and still more preferably $\pm 0.1\%$ from the specified value.

[0036] "Antigen" or "Ag" as used herein refers to a molecule that provokes an immune response. This immune response can involve either antibody production, or the activation of specific immunologically-competent cells, or both. It is readily apparent that an antigen can be generated synthesized, produced recombinantly or can be derived from a biological sample. Such a biological sample can include, but is not limited to a tissue sample, a tumor sample, a cell or a biological fluid such, for example, blood, plasma or ascites fluid.

[0037] "Anti-tumor effect" as used herein, refers to a biological effect, which can be manifested by a decrease in tumor volume, a decrease in the number of tumor cells, a decrease in the number of metastases, an increase in life expectancy, or a decrease of various physiological symptoms associated with the cancerous condition. An "anti-tumor effect" can also be manifested by a decrease in recurrence or an increase in the time before recurrence.

[0038] "Chimeric receptor" as used herein refers to a synthetically designed receptor comprising a ligand binding domain of an antibody or other protein sequence that binds to a molecule associated with the disease or disorder and is linked via a spacer domain to one or more intracellular signaling domains of a T cell or other receptors, such as a costimulatory domain. Chimeric receptor can also be referred to as artificial T cell receptors, chimeric T cell receptors, chimeric immunoreceptors, and chimeric antigen receptors (CARs). These CARs are engineered receptors that can graft an arbitrary specificity onto an immune receptor cell. The term Chimeric antigen receptors or "CARs" are also considered by some investigators to include the antibody or antibody fragment, the spacer, signaling domain, and transmembrane region. However, due to the surprising effects of modifying the different components or domains of the CAR described herein, such as the epitope binding region (for

example, antibody fragment, scFv, or portion thereof), spacer, transmembrane domain, and/or signaling domain), the components of the CAR are frequently distinguished throughout this disclosure in terms of independent elements. The variation of the different elements of the CAR can, for example, lead to stronger binding affinity for a specific epitope or antigen. "Co-stimulatory domain," as the term is used herein refers to a signaling moiety that provides to T cells a signal which, in addition to the primary signal provided by for instance the CD3 zeta chain of the TCR/CD3 complex, mediates a T cell response, including, but not limited to, activation, proliferation, differentiation, cytokine secretion, and the like. A co-stimulatory domain can include all or a portion of, but is not limited to, CD27, CD28, 4-1BB, OX40, CD30, CD40, , ICOS, lymphocyte function-associated antigen-1 (LFA-1), CD2, CD7, LIGHT, NKG2C, B7-H3, or a ligand that specifically binds with CD83. In some alternatives, the co-stimulatory domain is an intracellular signaling domain that interacts with other intracellular mediators to mediate a cell response including activation, proliferation, differentiation and cytokine secretion, and the like. "Coding for" are used herein refers to the property of specific sequences of nucleotides in a polynucleotide, such as a gene, a cDNA, or an mRNA, to serve as templates for synthesis of other macromolecules such as a defined sequence of amino acids. Thus, a gene codes for a protein if transcription and translation of mRNA corresponding to that gene produces the protein in a cell or other biological system. A "nucleic acid sequence coding for a polypeptide" includes all nucleotide sequences that are degenerate versions of each other and that code for the same amino acid sequence.

[0039] "Conditional" or "Inducible" as used herein refer to the nucleic acid construct that includes a promoter that provides for gene expression in the presence of an inducer and does not substantially provide for gene expression in the absence of the inducer.

[0040] "Constitutive" as used herein refer to the nucleic acid construct that includes a promoter that is constitutive providing for expression of a polypeptide that is continuously produced.

[0041] "Specific" or "Specificity" can refer to the characteristic of a ligand for the binding partner or alternatively, the binding partner for the ligand, and can include complementary shape, charge and hydrophobic specificity for binding. Specificity for binding can include stereospecificity, regioselectivity and chemoselectivity. In some alternatives, a method of making a nucleic acid encoding a chimeric antigen receptor is provided such that a nucleic acid encoding a chimeric antigen receptor is generated.

[0042] “Regulate” or “modulate” as described herein, refers to the act of controlling a biological process, or to exert a modifying or controlling influence on a biological or cellular process or pathway. In some alternatives, a system for inducible expression of a chimeric antigen receptor is provided, wherein the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte, a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized, a polynucleotide coding for a transmembrane domain, and d) a polynucleotide coding for an intracellular signaling domain, and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, modulation comprises modulation of cellular differentiation and apoptosis. In some alternatives, modulation comprises modulation of cellular proliferation by regulation of activity of proteins. In some alternatives, the proteins are cell cycle regulators and transcription factors. In some alternatives, the cell cycle regulators are cyclin D1, p21, p27 and/or cdc25A. In some alternatives, the transcription factors are c-Myc.

[0043] “Cytotoxic T lymphocyte “(CTL) as used herein refers to a T lymphocyte that expresses CD8 on the surface thereof (e.g., a CD8⁺ T cell). In some alternatives such cells are preferably “memory” T cells (T_M cells) that are antigen-experienced.

“Central memory” T cell (or “T_{CM}”) as used herein refers to an antigen experienced CTL that expresses CD62L, CCR-7 and/or CD45RO on the surface thereof, and does not express or has decreased expression of CD45RA, as compared to naïve cells. In some alternatives, central memory cells are positive for expression of CD62L, CCR7, CD28, CD127, CD45RO, and/or CD95, and may have decreased expression of CD54RA, as compared to naïve cells. “Effector memory” T cell (or “T_{EM}”) as used herein refers to an antigen experienced T cell that does not express or has decreased expression of CD62L on the surface thereof, as compared to central memory cells, and does not express or has a decreased expression of CD45RA, as compared to naïve cell. In some alternatives, effector memory cells are negative for expression of CD62L and/or CCR7, as compared to naïve cells or central memory cells, and may have variable expression of CD28 and/or CD45RA.

[0044] “Naïve “ T cells as used herein refers to a non-antigen experienced T lymphocyte that expresses CD62L and/or CD45RA, and does not express CD45RO-, as compared to central or effector memory cells. In some alternatives, naïve CD8+ T lymphocytes are characterized by the expression of phenotypic markers of naïve T cells including CD62L, CCR7, CD28, CD127, and/or CD45RA.

[0045] “Effector “ “T_E” T cells as used herein refers to antigen experienced cytotoxic T lymphocyte cells that do not express or have decreased expression of CD62L, CCR7, and/or CD28, and are positive for granzyme B and/or perforin, as compared to central memory or naïve T cells.

“Enriched” and “depleted” as used herein to describe amounts of cell types in a mixture refers to the subjecting of the mixture of the cells to a process or step, which results in an increase in the number of the “enriched” type and a decrease in the number of the “depleted” cells. Thus, depending upon the source of the original population of cells subjected to the enriching process, a mixture or composition may contain 60, 70, 80, 90, 95, or 99 percent or more (in number or count) of the “enriched” cells and/or 40, 30, 20, 10, 5 or 1 percent or less (in number or count) of the “depleted” cells.

[0046] “Epitope” as used herein refers to a part of an antigen or molecule that is recognized by the immune system including antibodies, T cells, and/ or B cells. Epitopes usually have at least 7 amino acids and can be linear or conformational.

“Isolated,” when used to describe the various polypeptides disclosed herein, means polypeptide or nucleic acid that has been identified and separated and/or recovered from a component of its natural environment. Preferably, the isolated polypeptide or nucleic acid is free of association with all components with which it is naturally associated. Contaminant components of its natural environment are materials that would typically interfere with diagnostic or therapeutic uses for the polypeptide or nucleic acid, and can include enzymes, hormones, and other proteinaceous or non-proteinaceous solutes.

[0047] “Intracellular signaling domain” as used herein refers to all or a portion of one or more domains of a molecule (here the chimeric receptor molecule) that provides for activation of a lymphocyte. Intracellular domains of such molecules mediate a signal by interacting with cellular mediators to result in proliferation, differentiation, activation and other effector functions. In some alternatives, such molecules include all or portions of CD28, CD3, or 4-1BB, or combinations thereof.

[0048] "Ligand" as used herein refers to a substance that binds specifically to another substance to form a complex. Examples of ligands include epitopes on antigens, molecules that bind to receptors, substrates, inhibitors, hormones, and/or activators. "Ligand binding domain" as used herein refers to substance or portion of a substance that binds to a ligand. Examples of ligand binding domains include antigen binding portions of antibodies, extracellular domains of receptors, and/or active sites of enzymes. "Operably linked" as used herein refers to functional linkage between a regulatory sequence and a heterologous nucleic acid sequence resulting in expression of the latter. For example, a first nucleic acid sequence is operably linked with a second nucleic acid sequence when the first nucleic acid sequence is placed in a functional relationship with the second nucleic acid sequence. For instance, a promoter is operably linked to a coding sequence if the promoter affects the transcription or expression of the coding sequence. Generally, operably linked DNA sequences are contiguous and, where necessary to join two protein coding regions, in the same reading frame.

[0049] "Percent (%) amino acid sequence identity" with respect to the chimeric receptor polypeptide sequences identified herein is defined as the percentage of amino acid residues in a candidate sequence that are identical with the amino acid residues in the reference sequence for each of the ligand binding domain, spacer, transmembrane domain, and/or the lymphocyte activating domain, after aligning the sequences and introducing gaps, if necessary, to achieve the maximum percent sequence identity, and not considering any conservative substitutions as part of the sequence identity. Alignment for purposes of determining percent amino acid sequence identity can be achieved in various ways that are within the skill in the art, for instance, using publicly available computer software such as BLAST, BLAST-2, ALIGN, ALIGN-2 or Megalign (DNASTAR) software. Those skilled in the art can determine appropriate parameters for measuring alignment, including any algorithms needed to achieve maximal alignment over the full-length of the sequences being compared. For example, % amino acid sequence identity values generated using the WU-BLAST-2 computer program [Altschul et al., *Methods in Enzymology*, 266:460-480 (1996)] uses several search parameters, most of which are set to the default values. Those that are not set to default values (i.e., the adjustable parameters) are set with the following values: overlap span=1, overlap fraction=0.125, word threshold (T) =11 and scoring matrix=BLOSUM62. A % amino acid sequence identity value is determined by dividing (a) the number of matching identical amino acid residues between the each or all of the polypeptide amino acid sequence of the reference chimeric receptor sequence provided in Table 2 and the comparison amino

acid sequence of interest as determined by WU-BLAST-2 by (b) the total number of amino acid residues of the polypeptide of interest. In some alternatives, the percent sequence identity of amino acids or nucleic acids are determined by computer software.

[0050] "Chimeric receptor variant polynucleotide" or "chimeric receptor variant nucleic acid sequence" as used herein refers to a polypeptide-encoding nucleic acid molecule as defined below having at least 80%, 85%, 90%, or 95% nucleic acid sequence identity (or a percentage nucleic acid sequence identity within a range defined by any two of the aforementioned percentages) with the polynucleotide acid sequence shown in **Table 1** or a specifically derived fragment thereof, such as polynucleotide coding for an antigen binding domain, a polynucleotide encoding a spacer domain, a polynucleotide coding for a transmembrane domain and/ or a polynucleotide coding for a lymphocyte stimulatory domain. Ordinarily, a chimeric receptor variant of polynucleotide or fragment thereof will have at least 80% nucleic acid sequence identity, more preferably at least 81% nucleic acid sequence identity, more preferably at least 82% nucleic acid sequence identity, more preferably at least 83% nucleic acid sequence identity, more preferably at least 84% nucleic acid sequence identity, more preferably at least 85% nucleic acid sequence identity, more preferably at least 86% nucleic acid sequence identity, more preferably at least 87% nucleic acid sequence identity, more preferably at least 88% nucleic acid sequence identity, more preferably at least 89% nucleic acid sequence identity, more preferably at least 90% nucleic acid sequence identity, more preferably at least 91% nucleic acid sequence identity, more preferably at least 92% nucleic acid sequence identity, more preferably at least 93% nucleic acid sequence identity, more preferably at least 94% nucleic acid sequence identity, more preferably at least 95% nucleic acid sequence identity, more preferably at least 96% nucleic acid sequence identity, more preferably at least 97% nucleic acid sequence identity, more preferably at least 98% nucleic acid sequence identity and yet more preferably at least 99% nucleic acid sequence identity with the nucleic acid sequence as shown in Table or a derived fragment thereof. Variants do not encompass the native nucleotide sequence. In this regard, due to the degeneracy of the genetic code, one of ordinary skill in the art will immediately recognize that a large number of chimeric receptor variant polynucleotides having at least 80% nucleic acid sequence identity to the nucleotide sequence of **Table 1** will encode a polypeptide having an amino acid sequence which is identical to the amino acid sequence of **Table 2**.

[0051] "Substantially purified" refers to a molecule that has 10%, 9%, 8%, 7%, 6%, 5%, 4%, 3%, 2%, or 1% or less other molecule types or other cell types. A substantially

purified cell also refers to a cell, which has been separated from other cell types with which it is normally associated in its naturally occurring state. In some instances, a population of substantially purified cells refers to a homogenous population of cells.

[0052] “Not substantially found” when used in reference the presence of a tumor antigen or other molecules on normal cells refers to the percentage of a normal cell type that has the antigen or molecule, and / or the density of the antigen on the cells. In some alternatives, not substantially found means that the antigen or molecule is found on less than 50% of normal cell type and/or at a 50% less density as compared to the amount of cells or antigen found on a tumor cell or other diseased cell.

[0053] “T cells” or “T lymphocytes” as used herein can be from any mammalian, preferably primate, species, including monkeys, dogs, and humans. In some alternatives the T cells are allogeneic (from the same species but different donor) as the recipient subject; in some alternatives the T cells are autologous (the donor and the recipient are the same); in some alternatives the T cells are syngeneic (the donor and the recipients are different but are identical twins).

[0054] “Vector” or “construct” is a nucleic acid used to introduce heterologous nucleic acids into a cell that has regulatory elements to provide expression of the heterologous nucleic acids in the cell. Vectors include but are not limited to plasmid, minicircles, yeast, and viral genomes. In some alternatives, the vectors are plasmid, minicircles, yeast, or viral genomes.

[0055] “Apoptosis” as described herein, refers to the process of programmed cell death (PCD) that can occur in multicellular organisms. Biochemical events lead to characteristic cell changes (morphology) and death. These changes include blebbing, cell shrinkage, nuclear fragmentation, chromatin condensation, and chromosomal DNA fragmentation. In apoptosis, a cell initiates intracellular apoptotic signaling in response to a stress, which can bring about cell suicide. The binding of nuclear receptors by glucocorticoids, heat, radiation, nutrient deprivation, viral infection, hypoxia and increased intracellular calcium concentration, for example, by damage to the membrane, can all trigger the release of intracellular apoptotic signals by a damaged cell. A number of cellular components, such as poly ADP ribose polymerase, can also help regulate apoptosis.

[0056] Before the actual process of cell death is precipitated by enzymes, apoptotic signals must cause regulatory proteins to initiate the apoptosis pathway. This step allows apoptotic signals to cause cell death, or the process to be stopped, should the cell no

longer need to die. Several proteins are involved, but two main methods of regulation have been identified: targeting mitochondria functionality, or directly transducing the signal via adaptor proteins to the apoptotic mechanisms. Another extrinsic pathway for initiation identified in several toxin studies is an increase in calcium concentration within a cell caused by drug activity, which also can cause apoptosis via a calcium binding protease calpain.

[0057] Apoptosis can be regulated by many factors. These factors can include but are not limited to genes that can express IL-2, IL-15, Chemokine receptors, Bcl2, CA-Akt, dn-TGFBetaRIII, dn-SHP1/2, and/or PD-1CD28 chimeras. IL-15 regulates T and natural killer cell activation and proliferation. In rodent lymphocytes, IL15 was shown to prevent apoptosis by inducing an apoptosis inhibitor, BCL2L1/BCL-X(L). In humans with celiac disease, IL-15 similarly suppresses apoptosis in T-lymphocytes by inducing Bcl-2 and/or BCL-xL. Bcl-2 (B-cell lymphoma 2), encoded in humans by the BCL2 gene, is the founding member of the Bcl-2 family of regulator proteins that regulate cell death (apoptosis), by either inducing (pro-apoptotic) it or inhibiting it (anti-apoptotic). Bcl-2 is specifically considered as an important anti-apoptotic protein and is, thus classified as an oncogene. Protein kinase B (PKB), also known as Akt, is a serine/threonine-specific protein kinase that plays a key role in multiple cellular processes such as glucose metabolism, apoptosis, cell proliferation, transcription and cell migration. In some alternatives, a system for inducible expression of chimeric antigen receptor is provided, wherein the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a cytokine, a chemokine receptor, a polypeptide that regulates apoptosis, or a polypeptide that modulates checkpoint signaling and b) a second nucleic acid comprising a second constitutive or inducible promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the polypeptide that regulates apoptosis or modulates checkpoint signaling comprises IL-2, IL-15, Chemokine receptors, Bcl2, CA-Akt, dn-TGFBetaRIII, dn-SHP1/2 or PD-1CD28 chimeras.

[0058] “Checkpoint signaling” as described herein, blocks the cell cycle at specific transition points, checkpoints to ensure that the events of the cell cycle take place in the correct order. Checkpoint signaling can also be activated. By way of example and not of limitation, checkpoint signaling can occur by damage to the DNA so that the cell cycle does not have to proceed until the damage is repaired. “Cell cycle checkpoints” are control mechanisms in eukaryotic cells which ensure proper division of the cell. Each checkpoint serves as a potential halting point along the cell cycle, during which the conditions of the cell

are assessed, with progression through the various phases of the cell cycle occurring when favorable conditions are met. Currently, there are three known checkpoints: the G1 checkpoint, also known as the restriction or start checkpoint; the G2/M checkpoint; and the metaphase checkpoint, also known as the spindle checkpoint. The biochemical pathways that restrain cell cycle transition and/or induce cell death after stress are known as cell cycle checkpoints. These checkpoints maintain the fidelity of DNA replication, repair, and division. Polypeptides that can regulate checkpoint signaling can include but are not limited to p53, p107, p130, and transcriptional repressor Rb.

[0059] “Negative checkpoint regulators” as described herein, refers to factors that can restrict the ability of T-cell responses to effectively attack tumors. They are also referred to as negative checkpoint signaling. In some alternatives, a system for inducible expression of chimeric antigen receptor is provided, wherein the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a cytokine, a chemokine receptor, a polypeptide that inhibits apoptosis, or a polypeptide that modulates checkpoint signaling and b) a second nucleic acid comprising a second constitutive or inducible promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the polypeptide that modulates checkpoint signaling inhibits negative checkpoint regulators. In some alternatives, the negative checkpoint regulator comprises VISTA, LAG-3 and/or TIM3.

[0060] In another example, cell cycle inhibitors mediating the growth inhibitory cues of upstream signaling pathways, the cyclin-CDK inhibitors of the Cip/Kip family p21Cip1, p27Kip1, and p57Kip2 have emerged as multifaceted proteins with functions beyond cell cycle regulation. In addition to regulating the cell cycle, Cip/Kip proteins can also play important roles in apoptosis, transcriptional regulation, cell fate determination, cell migration and cytoskeletal dynamics. A complex phosphorylation network modulates Cip/Kip protein functions by altering their subcellular localization, protein-protein interactions, and stability. These functions are essential for the maintenance of normal cell and tissue homeostasis, in processes ranging from embryonic development to tumor suppression. In some alternatives, a system for inducible expression of chimeric antigen receptor is provided, wherein the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a cytokine, a chemokine receptor, a polypeptide that regulates apoptosis, or a polypeptide that

modulates checkpoint signaling and b) a second nucleic acid comprising a second constitutive or inducible promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the polypeptide modulates checkpoint signaling. In some alternatives, the polypeptide that modulates checkpoint signaling inhibits negative checkpoint regulators. In some alternatives, the negative checkpoint regulator comprises VISTA, LAG-3 and/or TIM3.

[0061] “T cell precursors” as described herein refers to lymphoid precursor cells that can migrate to the thymus and become T cell precursors, which do not express a T cell receptor. All T cells originate from hematopoietic stem cells in the bone marrow. Hematopoietic progenitors (lymphoid progenitor cells) from hematopoietic stem cells populate the thymus and expand by cell division to generate a large population of immature thymocytes. The earliest thymocytes express neither CD4 nor CD8, and are therefore classed as double-negative ($CD4^-CD8^-$) cells. As they progress through their development, they become double-positive thymocytes ($CD4^+CD8^+$), and finally mature to single-positive ($CD4^+CD8^-$ or $CD4^-CD8^+$) thymocytes that are then released from the thymus to peripheral tissues.

[0062] About 98% of thymocytes die during the development processes in the thymus by failing either positive selection or negative selection, whereas the other 2% survive and leave the thymus to become mature immunocompetent T cells.

[0063] The double negative (DN) stage of the precursor T cell is focused on producing a functional β -chain whereas the double positive (DP) stage is focused on producing a functional α -chain, ultimately producing a functional $\alpha\beta$ T cell receptor. As the developing thymocyte progresses through the four DN stages (DN1, DN2, DN3, and DN4), the T cell expresses an invariant α -chain but rearranges the β -chain locus. If the rearranged β -chain successfully pairs with the invariant α -chain, signals are produced which cease rearrangement of the β -chain (and silence the alternate allele) and result in proliferation of the cell. Although these signals require this pre-TCR at the cell surface, they are dependent on ligand binding to the pre-TCR. These thymocytes will then express both CD4 and CD8 and progresses to the double positive (DP) stage where selection of the α -chain takes place. If a rearranged β -chain does not lead to any signaling (e.g. as a result of an inability to pair with the invariant α -chain), the cell may die by neglect (lack of signaling).

[0064] “Hematopoietic stem cells” or “HSC” as described herein, are precursor cells that can give rise to myeloid cells such as, for example, macrophages, monocytes,

macrophages, neutrophils, basophils, eosinophils, erythrocytes, megakaryocytes/platelets, dendritic cells and lymphoid lineages (such as, for example, T-cells, B-cells, NK-cells). HSCs have a heterogeneous population in which three classes of stem cells exist, which are distinguished by their ratio of lymphoid to myeloid progeny in the blood (L/M).

[0065] This disclosure provides for a system that has an inducible component for expression of transgenes and a constitutive component for expression of transgenes. The system can be tailored to provide for regulated expression of one or more transgenes to provide for functional characteristics in the transduced cells.

[0066] In some alternatives, a transgene under the control of the inducible promoter is a chimeric antigen receptor (CAR). The inducible promoter provides for the capacity to terminate CAR expression in cells while providing for reactivation of the cells at a later date (e.g. in the case of relapse). In addition, the cycling of CAR T cells through on and off periods can minimize exhaustion and/or anergy due to chronic stimulation of the T cell receptors.

[0067] The design of the vectors also provides for additional transgenes that can enhance one or more functional characteristics of transduced cells, such as enhanced tumor potency, survival and proliferation of transduced cells. In some alternatives, these transgenes are under the control of an inducible promoter. Such transgenes include, without limitation, genes that promote survival and proliferation, genes that prevent apoptosis, and genes that regulate checkpoint signaling. Such genes include genes encoding IL-2, IL-15, Chemokine receptors, Bcl2, CA-Akt, dn-TGFbetaRIII, dn-SHP1/2, and/or PD-1CD28 chimeras. In some alternatives, the transgenes are genes encoding IL-2, IL-15, Chemokine receptors, Bcl2, CA-Akt, dn-TGFbetaRIII, dn-SHP1/2, and/or PD-1CD28 chimeras. In some alternatives, the gene that modulates checkpoint signaling, encodes a polypeptide that inhibits negative checkpoint regulators. In some alternatives, the negative checkpoint regulator comprises VISTA, LAG-3 and/or TIM3.

[0068] The disclosure provides for a system comprising first and second nucleic acids, and vectors and host cells including such nucleic acids. Each of the first and second nucleic acids comprise a number of modular components that can be excised and replaced with other components in order to customize the system for a specific target cell. In some alternatives, the first nucleic acid includes an inducible promoter for control of the expression of the genes (e.g polynucleotide coding for a chimeric antigen receptor) in an on and off manner as needed. In other alternatives, the second nucleic acid comprises a constitutive

promoter that provides for expression of a transcriptional activator. In some alternatives, the gene encodes for a chimeric antigen receptor.

Inducible System.

[0069] The disclosure provides a system useful for providing regulated expression of transgenes in cells. Such transgenes include, without limitation, T cell receptors, affinity matured T cell receptors, chimeric antigen receptors, chemokine receptors, cytokines, genes that inhibit apoptosis, and/or genes that modulate checkpoint signaling. In some alternatives, the polypeptide that modulates checkpoint signaling inhibits negative checkpoint regulators. In some alternatives, the negative checkpoint regulator comprises VISTA, LAG-3 and/or TIM3. In some alternatives, the system contains a number of modular components that provide for easy substitution of elements of the nucleic acid. In some alternatives of the system, the system provides regulation of expression of transgenes in cells. In some alternatives, the transgenes code for T cell receptors, affinity matured T cell receptors, chimeric antigen receptors, chemokine receptors, cytokines, genes that regulate apoptosis, and/or genes that modulate checkpoint signaling. In some alternatives, the gene that modulates checkpoint signaling encodes a polypeptide inhibits negative checkpoint regulators. In some alternatives, the negative checkpoint regulator comprises VISTA, LAG-3 and/or TIM3.

[0070] In some alternatives, a system for inducible expression of chimeric antigen receptor comprises: a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a chimeric antigen receptor, the chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; and d) a polynucleotide coding for an intracellular signaling domain; and a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional modulator for the inducible promoter. In some alternatives, the second promoter is constitutive or inducible.

[0071] In some alternatives, a polynucleotide coding for a chimeric antigen receptor comprises a polynucleotide coding for a ligand binding domain, wherein the target

molecule is a tumor specific antigen, a polynucleotide coding for a polypeptide spacer wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain; and a polynucleotide coding for an intracellular signaling domain. In some alternatives, an expression vector comprises a first and/or second nucleic acid, as described herein. Polypeptides encoded by all of or a portion of the chimeric receptor nucleic acids are also included herein.

[0072] In other alternatives, a first nucleic acid comprises a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a gene that promotes cell survival and proliferation, a gene that regulates apoptosis, and/or a gene that modulates checkpoint signaling. Such genes include genes encoding IL-2, IL-15, Chemokine receptors, Bcl2, CA-Akt, dn-TGFbetaRIII, dn-SHP1/2, and/or PD-1CD28 chimeras. In some alternatives, the gene that modulates checkpoint signaling encodes a polypeptide that inhibits negative checkpoint regulators. In some alternatives, the negative checkpoint regulator comprises VISTA, LAG-3 and/or TIM3.

Inducible Promoters.

[0073] A system comprises a first nucleic acid comprising a first promoter inducible by a drug. By utilizing an inducible promoter, transgene expression can be turned on and off in order to avoid toxic side effects and/or to allow the cells to rest during remission. Although several inducible promoter systems are known, clinical applicability of these systems is limited due to toxic off target effects, unfavorable biodistribution and pharmacodynamics profiles, limited output dynamic range, and/or limited availability as FDA-approved commercially available pharmaceuticals. Furthermore, many of these systems use chimeric transcriptional regulators built from xenogeneic components, thus introducing the complication of immunogenicity when applying these systems to human therapeutics.

[0074] In some alternatives, a first promoter is inducible by a drug. The drug is selected based on safety record, favorable pharmacokinetic profile, tissue distribution, a low partition coefficient between the extracellular space and cytosol, low immunogenicity, low toxicities, and/or high expression in lymphocytes. In a specific alternative, a drug is selected that is FDA approved, provides for transgene expression in lymphocytes, does not activate other undesirable gene expression, and induces a promoter that does not contain any xenogeneic components. In some alternatives, the inducible promoter is activated by a

transcriptional activator that interacts with a drug. The transcriptional activator is activated or able to bind to and activate the inducible promoter in the presence of the drug.

[0075] A specific alternative of a drug is a drug that binds to an estrogen receptor ligand binding domain of a transcriptional activator. In some alternatives, the drug includes tamoxifen, its metabolites, analogs, and pharmaceutically acceptable salts and/or hydrates or solvates thereof.

[0076] Tamoxifen, CAS RN: 10540-29-1, is also known as 2-(4-((1Z)-1,2-diphenyl-1-butenyl)phenoxy)-N,N-dimethyl-ethanamine, or (Z)-2-(para-(1,2-Diphenyl-1-butenyl)phenoxy)-N,N-dimethylamine (IUPAC), and has a molecular formula of $C_{26}H_{29}NO$, M.W. 371.52. Tamoxifen is a Selective Estrogen Receptor Modulator with tissue-specific activities. Tamoxifen acts as an anti-estrogen (inhibiting agent) agent in the mammary tissue, but as an estrogen (stimulating agent) in cholesterol metabolism, bone density, and cell proliferation in the endometrium. Tamoxifen is frequently administered orally as a pharmaceutically acceptable salt. For example, Tamoxifen citrate (RN 54965-24-1, M.W. 563.643) is indicated for treatment of metastatic breast cancer, and as an adjuvant for the treatment of breast cancer in women following mastectomy axillary dissection, and breast irradiation. Tamoxifen citrate is also indicated to reduce incidence of breast cancer in women at high risk for breast cancer.

[0077] Metabolites of tamoxifen in rat, mouse and human breast cancer patients, including major metabolites N-desmethyltamoxifen (RN 31750-48-8, M.W. 357.494) and 4-hydroxytamoxifen (4-OHT) (RN 68392-35-8, M.W. 387.52, Afimoxifene), are disclosed in Robinson et al., Metabolites, pharmacodynamics, and pharmacokinetics of tamoxifen in rats and mice compared to the breast cancer patient. Drug Metab Dispos January 1991 19:36-43, which is incorporated by reference herein in its entirety. Additional cytochrome P-450 metabolites are disclosed in Crewe et al., 2002, including cis-4-hydroxytamoxifen (RN 174592, M.W. 387.52; Afimoxifene, E-isomer), and 4'-hydroxytamoxifen ((Z)-4-(1-(4-(2-(dimethylamino)ethoxy)phenyl)-1-phenylbut-1-en-2-yl)phenol). See Crewe et al., 2002, Metabolism of Tamoxifen by recombinant human cytochrome P-450 enzymes: Formation of the 4-hydroxy, 4'-hydroxy and N-desmethyl metabolites and isomerization of trans-4-hydroxytamoxifen, Drug Metab Dispos, 30(8): 869-874, FIG. 1, which is incorporated herein by reference.

[0078] Compounds with structural similarity to tamoxifen include, but are not limited to, cis-tamoxifen (RN 13002-65-8, M.W. 371.521), 4-methyltamoxifen (RN 73717-

95-5, M.W. 385.548), N-desmethyltamoxifen (RN 31750-48-8, M.W. 357.494), (Z)-desethyl methyl tamoxifen (RN 15917-50-7, M.W. 357.494), (E)-desethyl methyl tamoxifen (RN 31750-45-5, M.W. 357.494), trans-4-hydroxytamoxifen (RN 68047-06-3, M.W. 387.52), Afimoxifene (RN 68392-35-8, M.W. 387.52, 4-hydroxytamoxifen), Afimoxifene, E-isomer (RN 174592-47-3, M.W. 387.52), 4-chlorotamoxifen (RN 77588-46-6, M.W. 405.966), 4-fluorotamoxifen (RN 73617-96-6, M.W. 389.511), Toremifene (RN 89778-26-7, M.W. 405.966), desethyl tamoxifen (RN 19957-51-8, M.W. 343.47), (E)-desethyl tamoxifen (RN 97151-10-5, M.W. 343.47), (Z)-desethyl tamoxifen (RN 97151-11-6, M.W. 343.47), Miproxifene (RN 129612-87-9, M.W. 429.6), 2-(p-(beta-ethyl-alpha-phenylstyryl)phenoxy)triethylamine (RN 749-86-0, M.W. 399.575), Droloxifene (RN 82413-20-5, M.W. 387.52), 4-iodo-tamoxifen (RN 116057-68-2, M.W. 497.413), dihydrotamoxifen (RN 109640-20-2, M.W. 373.537), (E)-N,N-dimethyl-2-(4-(1-(2-methylphenyl)-2-phenyl-1-butenyl)phenoxy)ethanamine (RN 97150-96-4, M.W. 385.548), or 4-hydroxytoremifene (RN 110503-62-3, M.W. 421.965); and/or pharmaceutically acceptable salts and/or hydrates or solvates thereof.

[0079] For example, citrate salts of tamoxifen, or citrate salts of compounds with structural similarity to tamoxifen, include, but are not limited to tamoxifen citrate (RN 54965-24-1, M.W. 563.64), 2-(p-(1,2-diphenyl-1-butenyl)phenoxy)-N,N-dimethylethylamine citrate (RN 7244-97-5, 563.64), (E)-tamoxifen citrate (RN 76487-65-5, M.W. 563.64), Toremifene citrate (RN 89778-27-8, M.W. 598.088), Droloxifene citrate (RN 97752-20-0, M.W. 579.64), 2-(p-(1,2-bis(p-methoxyphenyl)-1-butenyl)phenoxy)triethylamine citrate (RN 42920-39-8, M.W. 651.748), 2-(4-(1,2-diphenylethenyl)phenoxy)-N,N-diethyl-ethanamine 2-hydroxy-1,2,3-propanetricarboxylate (RN 40297-42-5, M.W. 563.643), 2-(p-(alpha-phenylstyryl)phenoxy)triethylamine citrate (RN 102433-95-4, M.W. 563.64), 2-(p-(2-(p-methoxyphenyl)-1-phenyl-1-butenyl)phenoxy)triethylamine citrate (1:1) (RN 42824-34-0, M.W. 637.72), 2-(p-(1-(p-methoxyphenyl)-2-phenylpropenyl)phenoxy)triethylamine citrate (RN 13554-24-0, M.W. 607.696), 2-(p-(alpha-(p-methoxyphenyl)styryl)phenoxy)triethylamine citrate monohydrate (RN 13542-71-7, M.W. 593.669), 2-(p-(p-methoxy-alpha-phenylphenethyl) phenoxy)triethylamine citrate (RN 16421-72-0, M.W. 595.685), alpha-(p-(2-(diethylamino)ethoxy)phenyl)-beta-ethyl-p-methoxy-alpha-phenylphenethyl alcohol citrate (1:1) (RN 35263-93-5, M.W. 639.737), 1-(p-(2-(diethylamino)ethoxy)phenyl)-2-(p-methoxyphenyl)-1-phenylethanol citrate (M.W. 611.68), alpha-p-(2-(diethylamino)ethoxy)phenyl)-beta-ethyl-alpha-(p-hydroxyphenyl)-p-

methoxyphenethyl alcohol citrate (RN 35263-96-8, M.W. 655.737), and/or 2-(p-(p-methoxy-alpha-methylphenethyl)phenoxy)-triethylamine citrate (RN 15624-34-7, M.W. 533.614).

[0080] In some alternatives, an effective amount of the drug for inducing expression is an amount that provides for an increase in transgene expression over uninduced and/or basal level of expression. In some alternatives, this amount can be readily determined using known dosages and pharmacokinetic profile of the drug.

[0081] In some alternatives, the inducible promoter has a low level of basal activity. When a lentiviral vector is used, the level of basal activity in uninduced cells is 20%, 15%, 10%, 5%, 4%, 3%, 2%, 1% or less, as compared to when cells are induced to express the gene. The level of basal activity can be determined by measuring the amount of the expression of the transgene (e.g. marker gene) in the absence of the inducer (e.g. drug) using flow cytometry.

[0082] In some alternatives, the inducible promoter provides for a high level of induced activity, as compared to uninduced or basal activity. In some alternatives, the level of activity in the induced state is 2, 4, 6, 8, or 10 fold or greater than the activity level in the uninduced state. In some alternatives, transgene expression under control of the inducible promoter is turned off in the absence of a transactivator in less than 10, 8, 6, 4, 2, or 1 days excluding 0 days.

[0083] In some alternatives, an inducible promoter can be designed and/or modified to provide for a low level of basal activity, a high level of inducibility, and/or a short time for reversibility. In some alternatives, the inducible promoter is the 7xHBD/mE1b promoter. An exemplary sequence for the promoter is found in **Table 12** (SEQ ID NO: 41). For example, in the 7xHBD/mE1b promoter, mutations can be made to enhance the binding of the transcriptional activator.

[0084] In some alternatives, the system employs a synthetic transcriptional activator which, in the presence of the drug (e.g. tamoxifen), binds a synthetic promoter upstream of a transgene to induce expression. In some alternatives, the transcriptional activator is TamR-tf (HEA3). The tamoxifen regulated transcription factor ("TamR-tf", also designated "HEA3") is a chimeric transcription factor composed of human subunits including the N-terminal DNA binding domain of Hepatocyte Nuclear Factor 1-alpha (HNF-1 α) (e.g. amino acids 1-281 of SEQ ID NO: 40) fused in frame to the mutant tamoxifen-specific ligand binding domain of the estrogen receptor ligand binding domain (ER-LBD), that is in turn fused to the p65 activation domain of NF- κ B (p65). An exemplary amino acid sequence is

provided in **Table 10** and is identified as SEQ ID NO: 40. The mutant tamoxifen-specific ligand binding domain of the estrogen receptor ligand binding domain (ER-LBD) is found at amino acids 282-595 of the TamR-tf and has a mutation at position 521. The p65 activation domain of NF- κ B (p65 or TAD) is found at amino acids 596 to 862.

[0085] In some alternatives, a system for inducible expression of a chimeric antigen receptor is provided, wherein the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain; and d) a polynucleotide coding for an intracellular signaling domain; and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the drug is tamoxifen and/or its metabolites. In some alternatives, the second promoter is constitutive or inducible. In some alternatives, the first promoter comprises a nucleic acid sequence of SEQ ID NO: 41. In some alternatives, the second promoter is a constitutive promoter. In some alternatives, the second promoter is the EF1 α p. In some alternatives, the transcriptional activator comprises a sequence of SEQ ID NO: 40. In some alternatives, the first nucleic acid further comprises a first vector and the second nucleic acid further comprises a second vector. In some alternatives, both vectors are packaged in a viral vector. In some alternatives, the viral vector is a lentivirus. In some alternatives, the first and second nucleic acid comprise a vector. In some alternatives, the first nucleic acid further comprises a nucleic acid sequence coding for a selectable marker. In some alternatives, the second nucleic acid further comprises a nucleic acid coding for a selectable marker. In some alternatives, a system for inducible expression of chimeric antigen receptor is provided, wherein the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug operably linked to a polynucleotide coding for a cytokine, a chemokine receptor, a polypeptide that regulates apoptosis, or a polypeptide that modulates checkpoint signaling; and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the polypeptide that modulates checkpoint signaling inhibits negative

checkpoint regulators. In some alternatives, the negative checkpoint regulator comprises VISTA, LAG-3 and/or TIM3. In some alternatives, the second promoter is constitutive or inducible. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment, preferably a binding fragment thereof. In some alternatives, the ligand binding domain is single chain variable fragment. In some alternatives, the tumor specific molecule is CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, or MAGE A3 TCR or combinations thereof. In some alternatives, the system employs a synthetic transcriptional activator which, in the presence of a drug binds a synthetic promoter upstream of a transgene to induce expression. In some alternatives, the transcriptional activator is TamR-tf (HEA3). In some alternatives, the drug is tamoxifen.

[0086] Additional changes can be made to the transcriptional activator to increase the properties of the transcription factor including, without limitation, altering one or more amino acids in the estrogen receptor ligand binding domain and/or altering one or more amino acids in the p65 transactivating domain. Altering amino acids in the estrogen receptor binding domain can provide for more specific binding of the drug to the transcriptional activator. An example of a transcriptional activator with altered amino acid sequence in the ER-LBD is shown in **Table 11** (SEQ ID NO:43). Mutations are made at amino acid position 400, 543, and 544 of SEQ ID NO: 40. The transcriptional activator with altered sequence has increased affinity for tamoxifen or 4-OHT. Altering amino acids in the p65 transactivating domain can provide for increased expression of the transgene in the absence of activation of the transduced cells.

[0087] In the absence of tamoxifen, TamR-tf is excluded from the nucleus by binding of cytosolic heat-shock protein 90 (HSP90) to the tamoxifen binding active site and transgene expression is in the “OFF” state. Nanomolar concentrations of cytosolic tamoxifen actively outcompete HSP90 for ER-LBD binding, resulting in TamR-tf translocation to the

nucleus. Upon nuclear translocation, TamR-tf is readily available to bind its restricted synthetic promoter. In the presence of tamoxifen, binding of TamR-tf to 7xHBD/EF1ap promoter induces the “ON” state of transgene expression. In some alternatives, this transcriptional regulator can be modified to provide for varying level of control of transgene expression. Amino acid substitutions in the LBD of TamR-tf (HEA-3) permit selective responsiveness to tamoxifen and its metabolites, where 4-hydroxy tamoxifen (4-OHT) is the most pharmacologically active metabolite, in regards to TamR-tf (HEA-3) activity, while lacking interaction with endogenous estrogen.

[0088] In some alternatives, the inducible promoter functions in a lentiviral construct and/or in lymphocytes.

Chimeric Antigen Receptors.

[0089] A system for expression of chimeric antigen receptor comprises: a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a chimeric antigen receptor, the chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain; and d) a polynucleotide coding for an intracellular signaling domain. In other alternatives, another polynucleotide coding for a chimeric antigen receptor is under the control of a constitutive promoter. In some alternatives, the drug is tamoxifen.

Ligand binding domain.

[0090] In some alternatives, the chimeric receptor nucleic acid comprises a polynucleotide coding for a ligand binding domain. In some alternatives, the ligand binding domain specifically binds to a tumor or viral specific antigen and said ligand binding domain may be humanized. In some alternatives, a ligand binding domain, includes without limitation, receptors or portions thereof, small peptides, peptidomimetics, substrates, cytokines, and the like. In some alternatives, the ligand binding domain is an antibody or fragment thereof, preferably a binding fragment thereof, any of which may be humanized. A nucleic acid sequence coding for an antibody or antibody fragment can readily be determined. In a specific alternative, the polynucleotide codes for a single chain Fv that specifically binds

CD19. In other specific alternatives, the polynucleotide codes for a single chain Fv that specifically binds HER2, CE7, hB7H3, or EGFR and, optionally, said polynucleotide encodes a humanized version thereof. The sequences of these antibodies and binding domains thereof are known to or can readily be determined by those of skill in the art.

[0091] Tumor antigens are proteins that are produced by tumor cells that elicit an immune response. The selection of the ligand binding domain of the invention will depend on the type of cancer to be treated, and can target tumor antigens or other tumor cell surface molecules. A tumor sample from a subject can be characterized for the presence of certain biomarkers or cell surface markers. For example, breast cancer cells from a subject can be positive or negative for each of Her2Neu, Estrogen receptor, and/or the Progesterone receptor. A tumor antigen or cell surface molecule is selected that is found on the individual subject's tumor cells. Tumor antigens and cell surface molecules are well known in the art and include, for example, carcinoembryonic antigen (CEA), prostate specific antigen, PSMA, Her2/neu, estrogen receptor, progesterone receptor, ephrinB2, CD19, CD20, CD22, CD23, CD123, CS-1, CE7, hB7H3, ROR1, mesothelin, c-Met, GD-2, and/or MAGE A3 TCR. In some alternatives, a target molecule is a cell surface molecule that is found on tumor cells and is not substantially found on normal tissues, or restricted in its expression to non-vital normal tissues.

[0092] In one alternative, the target molecule on the tumor comprises one or more epitopes associated with a malignant tumor. Malignant tumors express a number of proteins that can serve as target antigens for T cell receptor or chimeric receptor mediated recognition. Other target molecules belong to the group of cell transformation-related molecules such as the oncogene HER-2/Neu/ErbB2. In some alternatives, the tumor antigen is selectively expressed or overexpressed on the tumor cells as compared to control cells of the same tissue type. In other alternatives, the tumor antigen is a cell surface polypeptide.

[0093] Once a tumor cell surface molecule that might be targeted with a chimeric receptor is identified, an epitope of the target molecule is selected and characterized. Antibodies that specifically bind a tumor cell surface molecule can be prepared using methods of obtaining monoclonal antibodies, methods of phage display, methods to generate human or humanized antibodies, or methods using a transgenic animal or plant engineered to produce human antibodies. Phage display libraries of partially or fully synthetic antibodies are available and can be screened for an antibody or fragment thereof that can bind to the target molecule. Phage display libraries of human antibodies are also available. In some alternatives,

antibodies specifically bind to a tumor cell surface molecule and do not cross react with nonspecific components such as bovine serum albumin or other unrelated antigens. Once identified, the amino acid sequence or polynucleotide sequence coding for the antibody can be isolated and/or determined.

[0094] Antibodies or antigen binding fragments include all or a portion of polyclonal antibodies, a monoclonal antibody, a human antibody, a humanized antibody, a synthetic antibody, a chimeric antibody, a bispecific antibody, a minibody, and a linear antibody. "Antibody fragments" comprise a portion of an intact antibody, preferably the antigen binding or variable region of the intact antibody and can readily be prepared. Examples of antibody fragments include Fab, Fab', F(ab')₂, and Fv fragments; diabodies; linear antibodies; single-chain antibody molecules; and multispecific antibodies formed from antibody fragments. In some alternatives the antibody fragments are Fab, Fab', F(ab')₂, and Fv fragments; diabodies; linear antibodies; single-chain antibody molecules; or multispecific antibodies formed from antibody fragments. Any of such aforementioned antibodies or antibody fragments can be humanized and used with the compositions and methods described herein.

[0095] In some alternatives, a number of different antibodies that bind to a particular tumor cell surface molecules can be isolated and characterized. In some alternatives, the antibodies are characterized based on epitope specificity of the targeted molecule. In addition, in some cases, antibodies that bind to the same epitope can be selected based on the affinity of the antibody for that epitope. In some alternatives, an antibody has an affinity of at least 1 mM, and preferably <50 nM. In some alternatives, the antibody has an affinity of 50 nM, 100 nM, 200 nM, 300nM 400 nM, 500 nM, 1uM, 100uM, 200uM, 300uM, 400uM, 500uM, 600uM, 700 uM, 800uM, 900uM or 1mM or an affinity within a range defined by any two of the aforementioned values. In some alternatives, an antibody is selected that has a higher affinity for the epitope, as compared to other antibodies. For example, an antibody is selected that has at least a 2 fold, at least a 5 fold, at least a 10 fold, at least a 20 fold, at least a 30 fold, at least a 40 fold, or at least a 50 fold greater affinity than a reference antibody that binds to the same epitope or an affinity that is greater than a reference antibody within a range defined by any two of the aforementioned values.

[0096] In some alternatives, target molecules are CD19, CD20, CD22, CD23, CE7, hB7H3, EGFR, CD123, CS-1, ROR1, mesothelin, Her2, c-Met, PSMA, GD-2, or

MAGE A3 TCR or combinations thereof. In some alternatives, the antibody or binding fragment thereof specific for these target molecules is humanized.

[0097] In specific alternatives, the target antigen is CD19. A number of antibodies specific for CD19 are known to those of skill in the art and can be readily characterized for sequence, epitope binding, and affinity. In a specific alternative, the chimeric receptor construct includes a scFv sequence from a FMC63 antibody. In other alternatives, the scFv is a human or humanized ScFv comprising a variable light chain comprising a CDRL1 sequence of RASQDISKYLN (SEQ ID NO: 88), CDRL2 sequence of SRLHSGV (SEQ ID NO: 89), and a CDRL3 sequence of GNTLPYTTFG (SEQ ID NO: 90). In other alternatives, the scFv is a human or humanized ScFv comprising a variable heavy chain comprising CDRH1 sequence of DYGVSV (SEQ ID NO: 91), CDRH2 sequence of VIWGSETTYNSALKS (SEQ ID NO: 92), and a CDRH3 sequence of YAMDYWG (SEQ ID NO: 93). The disclosure also contemplates variable regions that have at least 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% amino acid sequence identity to that of the scFv for FMC63 and that have at least the same affinity for CD19.

[0098] In some alternatives, CDR regions are found within antibody regions as numbered by Kabat as follows: for the light chain; CDRL1 amino acids 24-34; CDRL2 amino acids 50-56; CDRL3 at amino acids 89-97; for the heavy chain at CDRH1 at amino acids 31-35; CDRH2 at amino acids 50-65; and for CDRH3 at amino acids 95-102. CDR regions in antibodies can be readily determined.

[0099] In specific alternatives, the target antigen is Her2. A number of antibodies specific for Her2 are known to those of skill in the art and can be readily characterized for sequence, epitope binding, and affinity. In a specific alternative, the chimeric receptor construct includes a scFv sequence from a Herceptin antibody. In other alternatives, the scFv is a human or humanized ScFv comprising a variable light chain comprising a CDRL1 sequence, CDRL2 sequence and a CDRL3 sequence of the Herceptin antibody. In other alternatives, the scFv is a human or humanized ScFv comprising a variable heavy chain comprising CDRH1 sequence, CDRH2, and a CDRH3 sequence of Herceptin. The CDR sequences can readily be determined from the amino acid sequence of Herceptin. The disclosure also contemplates variable regions that have at least 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% amino acid sequence identity to that of the scFv for Herceptin and that have at least the same affinity for Her2.

[0100] “Humanized antibodies,” as described herein, refers to antibodies from non-human species whose protein sequences have been modified to increase their similarity to antibody variants produced naturally in humans. The process of “humanization” can be applied to monoclonal antibodies developed for administration to humans (for example, antibodies developed as anti-cancer drugs). Humanization can be desirable when the process of developing a specific antibody involves utilization of a non-human immune system (such as that in mice). The protein sequences of antibodies produced in this way are partially distinct from homologous antibodies occurring naturally in humans, and are therefore potentially immunogenic when administered to human patients. Humanized antibodies are distinct from chimeric antibodies, in that they have the protein sequences made more similar to human antibodies but can carry a larger stretch of non-human protein. A derivative of a humanized antibody can refer to a segment of an antibody or sequence that is derived from a humanized antibody. In some alternatives, the ligand binding domain comprises a humanized antibody or portion thereof. In some alternatives, the ligand binding domain comprises a scFv. In some alternatives, the scFv is a humanized scFv.

[0101] Humanization can be desirable in some alternatives for reducing the immunogenicity of monoclonal antibodies that are derived from xenogeneic sources, such as, for example, rodents. Humanization is also desirable in some alternatives so as to improve the interaction of the antibody or a fragment thereof with the human immune system. Due to the development of hybridoma technology, a large number of xenogeneic antibodies are highly immunogenic in humans, which can ultimately limit their clinical applications especially when administration may need to be repeated. Additionally, they can be rapidly removed from the circulation and can cause systemic inflammatory effects as well. Therefore humanization strategies are desirable in some alternatives to circumvent these situations. Techniques for antibody humanization are known to those skilled in the art.

[0102] In some alternatives, a polynucleotide coding for a ligand binding domain is operably linked to a polynucleotide coding for a spacer region. In some alternatives, the polynucleotide coding for a ligand binding domain can also have one or more restriction enzyme sites at the 5’ and/or 3’ ends of the coding sequence in order to provide for easy excision and replacement of the polynucleotide with another polynucleotide coding for a ligand binding domain coding for a different antigen or that has different binding characteristics. For example, a restriction site, NheI, is encoded upstream of the leader sequence; and a 3’ RsrII located within the hinge region allows subcloning of any desirable

scFv into a chimeric receptor vector. In some alternatives, the polynucleotide is codon optimized for expression in mammalian cells.

[0103] In some alternatives, the polynucleotide coding for a ligand binding domain is operably linked to a signal peptide. In some alternatives, the signal peptide is a signal peptide for granulocyte colony stimulating factor. Polynucleotides coding for other signal peptides such as CD8 alpha can be utilized. In some alternatives, the polynucleotide codes for CD8 alpha.

[0104] In some alternatives, the polynucleotide coding for a ligand binding domain is operably linked to a promoter. A promoter is selected that provides for expression of the chimeric antigen receptor in a mammalian cell. In a specific alternative, the promoter is an inducible promoter.

[0105] A specific alternative of a polynucleotide coding for a ligand binding domain is shown in **Table 1** as the scFv from an antibody that specifically binds CD19, such as FMC63. A polynucleotide encoding for a flexible linker including the amino acids GSTSGSGKPGSGEGSTKG (SEQ ID NO: 94) separates the VH and VL chains in the scFv. The amino acid sequence of the scFv including the linker is shown in **Table 2**. (SEQ ID NO:11) Other CD19-targeting antibodies such as SJ25C1 and HD37 are known. (SJ25C1: Bejcek et al. Cancer Res 2005, PMID 7538901; HD37: Pezutto et al. JI 1987, PMID 2437199).

Spacer.

[0106] In some alternatives, the chimeric receptor nucleic acid comprises a polynucleotide coding for a spacer region. Typically a spacer region is found between the ligand binding domain and the transmembrane domain of the chimeric receptor. In some alternatives, a spacer region provides for flexibility of the ligand binding domain and allows for high expression levels in lymphocytes. A CD19-specific chimeric receptor having a spacer domain of 229 amino acids had less antitumor activity than a CD19-specific chimeric receptor with a short spacer region comprised of the modified IgG4 hinge only. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor.

[0107] In some alternatives, a spacer region has at least 10 to 229 amino acids, 10 to 200 amino acids, 10 to 175 amino acids, 10 to 150 amino acids, 10 to 125 amino acids, 10 to 100 amino acids, 10 to 75 amino acids, 10 to 50 amino acids, 10 to 40 amino acids, 10 to

30 amino acids, 10 to 20 amino acids, or 10 to 15 amino acids, or a length within a range defined by any two of the aforementioned lengths. In some alternatives, a spacer region has 12 amino acids or less, 119 amino acids or less, or 229 amino acids or less but greater than 1 or 2 amino acids. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor.

[0108] In some alternatives, the spacer region is derived from a hinge region of an immunoglobulin like molecule. In some alternatives, a spacer region comprises all or a portion of the hinge region from a human IgG1, human IgG2, a human IgG3, or a human IgG4, and can contain one or more amino acid substitutions. Exemplary sequences of the hinge regions are provided in **Table 8**. In some alternatives, a portion of the hinge region includes the upper hinge amino acids found between the variable heavy chain and the core, and the core hinge amino acids including a polyproline region. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor.

[0109] In some alternatives, hinge region sequences can be modified in one or more amino acids in order to avoid undesirable structural interactions such as dimerization. In a specific alternative, the spacer region comprises a portion of a modified human hinge region from IgG4, for example, as shown in **Table 2** or **Table 8** (SEQ ID NO: 21). A representative of a polynucleotide coding for a portion of a modified IgG4 hinge region is provided in **Table 1**. (SEQ ID NO: 4). In some alternatives, a hinge region can have at least 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% sequence identity with a hinge region amino acid sequence identified in **Table 2** or **Table 8**, or any other percent sequence identity between any two of the percent sequence identities listed. In a specific alternative, a portion of a human hinge region from IgG4 has an amino acid substitution in the core amino acids from CPSP to CPPC.

[0110] In some alternatives, all or a portion of the hinge region is combined with one or more domains of a constant region of an immunoglobulin. For example, a portion of a hinge region can be combined with all or a portion of a CH2 or CH3 domain or variant thereof. In some alternatives, the spacer region does not include the 47-48 amino acid hinge region sequence from CD8 α or the spacer region consisting of an extracellular portion of the CD28 molecule.

[0111] In some alternatives, a short spacer region has 12 amino acids or less and comprises all or a portion of a IgG4 hinge region sequence or variant thereof, an intermediate spacer region has 119 amino acids or less and comprises all or a portion of a IgG4 hinge region sequence and a CH3 region or variant thereof, and a long spacer has 229 amino acids or less and comprises all or a portion of a IgG4 hinge region sequence, a CH2 region, and a CH3 region or variant thereof. In some alternatives, a short spacer region has 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11 or 12 amino acids or a size within a range defined by any two of the aforementioned amino acid lengths. In some alternatives, a medium spacer region has 13, 20, 30, 40, 50, 60, 70, 80, 90, 100, 110 or 119 amino acids or a size within a range defined by any two of the aforementioned amino acid lengths. In some alternatives, a spacer region has 120, 130, 140, 150, 160, 170, 180, 190, 200, 210 or 219 amino acids or a size within a range defined by any two of the aforementioned amino acid lengths.

[0112] A polynucleotide coding for a spacer region can be readily prepared by synthetic or recombinant methods from the amino acid sequence. In some alternatives, a polynucleotide coding for a spacer region is operably linked to a polynucleotide coding for a transmembrane region. In some alternatives, the polynucleotide coding for the spacer region can also have one or more restriction enzyme sites at the 5' and/or 3' ends of the coding sequence in order to provide for easy excision and replacement of the polynucleotide with another polynucleotide coding for a different spacer region. In some alternatives, the polynucleotide coding for the spacer region is codon optimized for expression in mammalian cells.

[0113] In an alternative, the spacer region is a hinge region sequence from IgG1, IgG2, IgG3, or IgG4 or a portion thereof, a hinge region sequence from IgG1, IgG2, IgG3, or IgG4 in combination with all or a portion of a CH2 region or variant thereof, a hinge region sequence from IgG1, IgG2, IgG3, or IgG4 in combination with all or a portion of a CH3 region or variant thereof, and a hinge region sequence from IgG1, IgG2, IgG3, or IgG4 in combination with all or a portion of a CH2 region or variant thereof, and/or a CH3 region or variant thereof. In some alternatives, a short spacer region is a modified IgG4 hinge sequence (SEQ ID NO: 4) having 12 amino acids or less but greater than one or two amino acids, an intermediate sequence is a IgG4 hinge sequence with a CH3 sequence having 119 amino acids or less but greater than one or two amino acids (SEQ ID NO: 62); or a IgG4 hinge sequence with a CH2 and CH3 region having 229 amino acids or less but greater than one or two amino acids (SEQ ID NO: 50).

Transmembrane domain.

[0114] In some alternatives, the chimeric receptor nucleic acid comprises a polynucleotide coding for a transmembrane domain. The transmembrane domain provides for anchoring of the chimeric receptor in the membrane.

[0115] In an alternative, the transmembrane domain that naturally is associated with one of the domains in the chimeric receptor is used. In some cases, the transmembrane domain can be selected or modified by amino acid substitution to avoid binding of such domains to the transmembrane domains of the same or different surface membrane proteins to minimize interactions with other members of the receptor complex.

[0116] The transmembrane domain can be derived either from a natural or a synthetic source. When the source is natural, the domain can be derived from any membrane-bound or transmembrane protein.

[0117] Transmembrane regions comprise at least the transmembrane region(s) of the alpha, beta or zeta chain of the T-cell receptor, CD28, CD3, CD45, CD4, CD8, CD9, CD16, CD22; CD33, CD37, CD64, CD80, CD86, CD134, CD137 and/or CD154. In a specific alternative, the transmembrane domain comprises the amino acid sequence of the CD28 transmembrane domain as shown in **Table 2**. A representative polynucleotide sequence coding for the CD28 transmembrane domain is shown in **Table 1** (SEQ ID NO: 5).

[0118] A transmembrane domain can be synthetic or a variant of a naturally occurring transmembrane domain. In some alternatives, synthetic or variant transmembrane domains comprise predominantly hydrophobic residues such as leucine and valine. In some alternatives, a transmembrane domain can have at least 80%, 85%, 90%, 95%, or 100% amino acid sequence identity with a transmembrane domain as shown in **Table 2** or **Table 6** or an amino acid sequence identity within a range defined by any two of the aforementioned values. Variant transmembrane domains preferably have a hydrophobic score of at least 50 as calculated by Kyte Doolittle.

[0119] A polynucleotide coding for a transmembrane domain can be readily prepared by synthetic or recombinant methods. In some alternatives, a polynucleotide coding for a transmembrane domain is operably linked to a polynucleotide coding for an intracellular signaling region. In some alternatives, the polynucleotide coding for a transmembrane domain can also have one or more restriction enzyme sites at the 5' and/or 3' ends of the coding sequence in order to provide for easy excision and replacement of the polynucleotide coding

for a transmembrane domain with another polynucleotide coding for a different transmembrane domain. In some alternatives, the polynucleotide coding for a transmembrane domain is codon optimized for expression in mammalian cells. In some alternatives, the mammalian cells are human cells.

Intracellular signaling domain.

[0120] In some alternatives, the chimeric receptor nucleic acid comprises a polynucleotide coding for an intracellular signaling domain. The intracellular signaling domain provides for activation of one function of the transduced cell expressing the chimeric receptor upon binding to the ligand expressed on tumor cells. In some alternatives, the intracellular signaling domain contains one or more intracellular signaling domains. In some alternatives, the intracellular signaling domain is a portion of and/or a variant of an intracellular signaling domain that provides for activation of at least one function of the transduced cell.

[0121] Examples of intracellular signaling domains for use in a chimeric receptor of the disclosure include the cytoplasmic sequences of the CD3 zeta chain, and/or co-receptors that act in concert to initiate signal transduction following chimeric receptor engagement, as well as any derivative or variant of these sequences and any synthetic sequence that has the same functional capability. T cell activation can be said to be mediated by two distinct classes of cytoplasmic signaling sequence: those that initiate antigen-dependent primary activation and provide a T cell receptor like signal (primary cytoplasmic signaling sequences) and those that act in an antigen-independent manner to provide a secondary or co-stimulatory signal (secondary cytoplasmic signaling sequences). Primary cytoplasmic signaling sequences that act in a stimulatory manner can contain signaling motifs which are known as receptor tyrosine-based activation motifs or ITAMs. Examples of ITAM containing primary cytoplasmic signaling sequences include those derived from CD3 zeta, FcR gamma, CD3 gamma, CD3 delta, CD3 epsilon, CD5, CD22, CD79a, CD79b, and/or CD66d. In some alternatives, the primary signaling intracellular domain can have at least 80%, 85%, 90%, or 95% sequence identity to CD3zeta having a sequence provided in **Table 2** or at least a percent sequence identity that is within a range defined by any two of the percent sequence identities listed. In some alternatives of the variants, of CD3 zeta retain at least one, two, three or all ITAM regions as shown in **Table 7**.

[0122] In a preferred alternative, the intracellular signaling domain of the chimeric receptor can be designed to comprise the CD3-zeta signaling domain by itself or combined with any other desired cytoplasmic domain(s). For example, the intracellular signaling domain of the chimeric receptor can comprise a CD3zeta chain and a costimulatory signaling region.

[0123] The costimulatory signaling region refers to a portion of the chimeric receptor comprising the intracellular domain of a costimulatory molecule. A costimulatory molecule is a cell surface molecule other than an antigen receptor or their ligands that is required for a response of lymphocytes to an antigen. Examples of such molecules include CD27, CD28, 4-1BB (CD 137), OX40, CD30, CD40, lymphocyte function-associated antigen-1 (LFA-1), CD2, CD7, LIGHT, NKG2C, B7-H3, zeta chain associated protein kinase (ZAP70), and/or a ligand that specifically binds with CD83. In some alternatives, the costimulatory signaling domain can have at least 80%, 85%, 90%, or 95% amino acid sequence identity to the intracellular domain of CD28 as shown in **Table 5** or to 4-1BB having a sequence provided in **Table 2** or at least a percent sequence identity that is within a range defined by any two of the percent sequence identities listed. In an alternative, a variant of the CD28 intracellular domain comprises an amino acid substitution at positions 186-187, wherein LL is substituted with GG.

[0124] The intracellular signaling sequences of the chimeric receptor can be linked to each other in a random or specified order. In some alternatives, a short oligo- or polypeptide linker, preferably between 2 and 10 amino acids in length can form the linkage. In one alternative, the intracellular signaling domains comprises all or a portion of the signaling domain of CD3-zeta or variant thereof and all or a portion of the signaling domain of CD28 or a variant thereof. In another alternative, the intracellular signaling domain comprises all or a portion of the signaling domain of CD3-zeta or variant thereof and all or a portion of the signaling domain of 4-1BB or variant thereof. In yet another alternative, the intracellular signaling domain comprises all or a portion of the signaling domain of CD3-zeta or variant thereof, all or a portion of the signaling domain of CD28 or variant thereof, and all or a portion of the signaling domain of 4-1BB or variant thereof. In a specific alternative, the amino acid sequence of the intracellular signaling domain comprising a variant of CD3zeta and a portion of the 4-1BB intracellular signaling domain is provided in **Table 2**. A representative nucleic acid sequence is provided in **Table 1** (SEQ ID NO: 6; SEQ ID NO: 7). In some alternatives, the nucleic acid sequence comprises the sequence set forth in SEQ ID

NO: 6. In some alternatives, the nucleic acid sequence comprises the sequence set forth in SEQ ID NO: 7.

[0125] In an alternative, a polynucleotide coding for an intracellular signaling domain comprises a 4-1BB intracellular domain linked to a portion of a CD3zeta domain. In other alternatives, a 4-1BB intracellular domain and a CD28 intracellular domain are linked to a portion of a CD3 zeta domain.

[0126] A polynucleotide coding for an intracellular signaling domain can be readily prepared by synthetic or recombinant methods from the amino acid sequence. In some alternatives, the polynucleotide coding for an intracellular signaling domain can also have one or more restriction enzyme sites at the 5' and/or 3' ends of the coding sequence in order to provide for easy excision and replacement of the polynucleotide coding for an intracellular signaling domain with another polynucleotide coding for a different intracellular signaling domain. In some alternatives, the polynucleotide coding for an intracellular signaling domain is codon optimized for expression in mammalian cells. In some alternatives, the mammalian cells are human cells.

Marker sequences.

[0127] In some alternatives, the system further comprises one or more marker sequences under the control of an inducible promoter. A marker sequence can provide for selection of transduced cells, and/or identification of transduced cells. In some alternatives, the marker sequence is for a selection of transduced cells and/or identification of transduced cells. In some alternatives, the marker sequence is operably linked to a polynucleotide sequence coding for a linker sequence. In some alternatives, the linker sequence is a cleavable linker sequence. In some alternatives, the linker is a cleavable T2A linker.

[0128] A number of different marker sequences can be employed. Typically a marker sequence has a functional characteristic that allows for selection of transduced cells and/or detection of transduced cells. In some alternatives, the marker sequence is compatible with transduction of human lymphocytes. In some alternatives, the marker sequence allows for selection of transduced cells and/or detection of transduced cells.

[0129] The positive selectable marker can be a gene, which upon being introduced into the host cell, expresses a dominant phenotype permitting positive selection of cells carrying the gene. Genes of this type are known in the art, and include, inter alia, hygromycin-B phosphotransferase gene (hph) which confers resistance to hygromycin B, the amino

glycoside phosphotransferase gene (neo or aph) from Tn5, which codes for resistance to the antibiotic G418, the dihydrofolate reductase (DHFR) gene, which provides resistance to methotrexate, DHFR dm (exemplary polynucleotide and amino acid sequences in **Table 14**, SEQ ID NO: 46 and SEQ ID NO: 47, the pac gene that provides resistance to puromycin, Sh ble gene which inactivates zeocin, the adenosine deaminase gene (ADA), and the multi-drug resistance (MDR) gene. Transduced cells cultured in the presence of these agents will survive and be selected.

[0130] In an alternative, a first nucleic acid further comprises a polynucleotide coding for a marker sequence. In an alternative, the marker sequence is a truncated epidermal growth factor receptor as shown in **Table 2**. An exemplary polynucleotide for the truncated epidermal growth factor receptor is shown in Table 1. (SEQ ID NO: 9). In some alternatives, the marker sequence is a truncated Her2 sequence. An exemplary polynucleotide and amino acid for the truncated Her2 sequences is shown in **Table 13** and provided by SEQ ID NO: 44 and SEQ ID NO: 45, respectively.

[0131] In some alternatives, the polynucleotide coding for the marker sequence is operably linked to a polynucleotide coding for a linker sequence. In a specific alternative, the linker sequence is a cleavable linker sequence T2A, as shown in **Table 2**. An exemplary polynucleotide sequence coding for the T2A linker is provided in **Table 1**. (SEQ ID NO:8).

[0132] A polynucleotide coding for marker sequence can be readily prepared by synthetic or recombinant methods from the amino acid sequence. In some alternatives, a polynucleotide coding for a marker sequence is operably linked to a polynucleotide coding for an intracellular signaling domain. In some alternatives, the polynucleotide coding for a marker sequence can also have one or more restriction enzyme sites at the 5' and/or 3' ends of the coding sequence in order to provide for easy excision and replacement of the polynucleotide coding for a marker sequence with another polynucleotide coding for a different marker sequence. In some alternatives, the polynucleotide coding for a marker sequence is codon optimized for expression in mammalian cells, preferably humans.

[0133] In some alternatives, two or more marker sequences can be employed. In some alternatives, a first marker sequence is under control of a constitutive promoter and provides for an indication that the transduced cell is expressing the transgene. In other alternatives, a second marker sequence is under the control of the inducible promoter and provides an indication that the transgene expression has been induced. In some alternatives, the marker under the control of the inducible promoter can be used to select for cells in which

noninduced or basal expression is much lower than in other cells by selecting cells that have a lower expression of the marker sequence under the control of the inducible promoter and expand those cells for further applications.

Other Genetic Components under control of Inducible Promoter.

[0134] In some alternatives, the first nucleic acid comprises a polynucleotide sequence coding for genes that promote survival and proliferation, genes that prevent apoptosis, and/or genes that inhibit negative checkpoint signaling under the control of an inducible promoter. Such genes include genes encoding IL-2, IL-15, Chemokine receptors, Bcl2, CA-Akt, dn-TGFbetaRIII, dn-SHP1/2, and/or PD-1CD28 chimeras. These genes are also placed under the control of an inducible promoter as described herein. In some alternatives, the genes encode IL-2, IL-15, Chemokine receptors, Bcl2, CA-Akt, dn-TGFbetaRIII, dn-SHP1/2, and/or PD-1CD28 chimeras. In some alternatives, the gene that modulates checkpoint signaling encodes a polypeptide that inhibits negative checkpoint regulators. In some alternatives, the negative checkpoint regulator comprises VISTA, LAG-3 and/or TIM3.

[0135] In some alternatives, a first nucleic acid comprises a first inducible promoter linked to a polynucleotide coding for a cytokine or chemokine receptor. Chemokines, also referred to as chemotactic cytokines, are a group of structurally related proteins that regulate cell trafficking of lymphocytes. In some alternatives, the chemokines are homeostatic or inflammatory. Chemokine receptors include CCR2, CCR7, or CCR15. Cytokines include interleukins such as IL2, IL-12, IL-7, and/or IL-15, interferons, such as interferon δ , tumor necrosis factor, and a TLR4 agonist. In some alternatives, the chemokine receptors comprise CCR2, CCR7, and/or CCR15. In some alternatives, the chemokine receptors include CCR2, CCR7, or CCR15. In some alternatives, the cytokines include interleukins, wherein the interleukins are IL2, IL-12, IL-7 and/or IL-15 or interferons, wherein the interferons comprise interferon δ , tumor necrosis factor, or a TLR4 agonist.

[0136] In some alternatives, a first nucleic acid comprises a first inducible promoter linked to a polynucleotide coding for a polypeptide that regulates apoptosis. In some alternatives, genes that inhibit apoptosis include, for example, Bcl2, and/or CA-Akt. In some alternatives, the polypeptide is Bcl2 or CA-Akt.

[0137] In some alternatives, a first nucleic acid comprises a first inducible promoter linked to a polynucleotide coding for a polypeptide that modulates checkpoint

signaling. Such genes include dn-TGFbetaRIII, dn-SHP1/2, and/or PD-1CD28 chimeras. In some alternatives, the polypeptide is dn-TGFbetaRIII, dn-SHP1/2, and/or PD-1CD28 chimeras. In some alternatives, the polypeptide that modulates checkpoint signaling inhibits negative checkpoint regulators. In some alternatives, the negative checkpoint regulator comprises VISTA, LAG-3 and/or TIM3.

[0138] Exemplary sequences for polynucleotides encoding these genes are found at:

Gene	A.A.	A.A.	N.A.	N.A.
	Accession	gI	Accession	gI
CCR2	P41597	1168965	NM_001123041	183979979
CCR7	P32248	1352335	NM_001838	299473754
IL-2	AAH70338	47682793	BC070338	47682792
IL-12	AAD16432	4323579	AF101062	4323578
IL-7	AAC63047	386824	NM_000880	315467865
IL-15	CAG46804	49456967	CR542007	49456966
IFN-g	EAW97180	119617586	EAW97180	119617586
TNF	NP_000585	25952111	NM_000594	395132451
Bcl2	AAH27258	20072668	BC027258	20072667
CA-Akt	NP_001014432	62241015	NM_001014432	62241014
TGFbeta Receptor III	Q03167	311033535	NM_003243	307574689
SHP1	NP_002822	18104989	NM_002831	166064064
SHP2	Q06124	84028248	NM_002834	33356176
PD-1	NP_001129245	209413749	NM_001135773	209413748
CD28	AAA51945	180092	NM_006139	340545506

[0139] Any number of nucleic acids can be placed under the control of an inducible promoter including those coding for chimeric antigen receptor, a marker sequences, a cytokine, a chemokine, an inhibitor of apoptosis, and/ or an inhibitor of negative checkpoint signaling. In some alternatives, one or more inducible promoters can be utilized to provide for an adequate expression level of each of the nucleic acids. In some alternatives, constructs can

be prepared with a gene such as a cytokine under the control of an inducible promoter and a construct comprising a chimeric antigen receptor under the control of a constitutive promoter. Such constructs are useful to provide for cell survival and proliferation of transduced cells, for example lymphocytes expressing a chimeric antigen receptor.

Constitutive Promoter Systems.

[0140] In other alternatives, a system comprises a second nucleic acid that comprises a constitutive promoter or a second inducible promoter linked to a transcriptional activator. In other alternatives, a system comprises a second nucleic acid that comprises a promoter linked to a transcriptional activator. In some alternatives, the promoter is a constitutive promoter or an inducible promoter. In some alternatives, a constitutive promoter includes EF1 α promoter, actin promoter, the myosin promoter, the hemoglobin promoter, and the creatine kinase promoter. In some alternatives, viral promoters such as the CMV promoter are excluded. In some alternatives, the constitutive promoter can be linked to one or more of a polynucleotide coding for marker, or a chimeric antigen receptor as described herein.

Constitutive Promoters.

[0141] A constitutive promoter provides for continuous gene expression of the gene under the control of the promoter. In some alternatives, the constitutive promoter is a promoter that provides for gene expression in a lentiviral construct and/or in lymphocytes. In some alternatives, the promoter is not derived from a xenogenic source such as a plant or a virus.

[0142] In a specific alternative, the constitutive promoter comprises EF1 α promoter, actin promoter, the myosin promoter, the hemoglobin promoter, and/or the creatine kinase promoter. In some alternatives, viral promoters such as the CMV promoter are excluded.

Transcriptional Activators.

[0143] In some alternatives, the constitutive promoter is operably linked to a transcriptional activator. In some alternatives, the transcriptional activator activates an inducible promoter in the presence of the inducer (e.g. drug).

[0144] In some alternatives, an inducible promoter is induced in the presence of a transcriptional activator. In some alternatives, the transcriptional activator preferentially binds

to the promoter in the presence of the drug. In some alternatives, the transcriptional activator is TamR-tf(HEA3). Modification of the transcriptional activator can be made in the amino acid sequence that can affect the ability of activator to bind to the drug, the promoter, or both. For example, binding in the ER ligand binding domain would affect the binding of the drug to the transcriptional activator.

[0145] In some alternatives, the system employs a synthetic transcriptional activator which, in the presence of the drug (e.g. tamoxifen), binds a synthetic promoter upstream of a transgene to induce expression. In some alternatives, the transcriptional activator is TamR-tf (HEA3). The tamoxifen regulated transcription factor ("TamR-tf", also designated "HEA3") is a chimeric transcription factor composed of human subunits including the N-terminal DNA binding domain of Hepatocyte Nuclear Factor 1-alpha (HNF-1 α) (e.g. amino acids 1-281 of SEQ ID NO: 40) fused in frame to the mutant tamoxifen-specific ligand binding domain of the estrogen receptor ligand binding domain (ER-LBD), that is in turn fused to the p65 activation domain of NF- κ B (p65). An exemplary amino acid sequence is provided in **Table 10** and is identified as SEQ ID NO: 40. The mutant tamoxifen-specific ligand binding domain of the estrogen receptor ligand binding domain (ER-LBD) is found at amino acids 282-595 of the TamR-tf and has a mutation at position 521. The p65 activation domain of NF- κ B (p65 or TAD) is found at amino acids 596 to 862.

[0146] Additional changes can be made to the transcriptional activator to increase the properties of the transcription factor including, without limitation, altering one or more amino acids in the estrogen receptor ligand binding domain and/or altering one or more amino acids in the p65 transactivating domain. Altering amino acids in the estrogen receptor binding domain can provide for more specific binding of the drug to the transcriptional activator. An example of a transcriptional activator with altered sequence in the ER-LBD is shown in **Table 11** (SEQ ID NO:43). Mutations are made at amino acid position 400, 543, and 544 of SEQ ID NO: 40. The transcriptional activator with altered sequence has increased affinity for tamoxifen or 4-OHT. Altering amino acids in the p65 transactivating domain can provide for increased expression of the transgene in the absence of activation of the transduced cells.

Marker.

[0147] In some alternatives, the constitutive promoter is operably linked to a polynucleotide coding for a marker polypeptide. Such marker polypeptides are described herein, and include EGFRt, Her2t, and/or DHFRdm.

Chimeric Antigen Receptor.

[0148] In some alternatives, the constitutive promoter is operably linked to a polynucleotide coding for a chimeric antigen receptor. In some alternatives, the chimeric antigen receptor comprises a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the spacer is optimized to provide for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor. Examples of chimeric antigen receptors are described herein.

Vectors.

[0149] A variety of vector combinations can be constructed to provide for efficiency of transduction and transgene expression. In some alternatives, the vector is a dual packaged or single (all in one) viral vector. In other alternatives, the vectors can include a combination of viral vectors and plasmid vectors. Other viral vectors include foamy virus, adenoviral vectors, retroviral vectors, and lentiviral vectors. In some alternatives, the vector is a lentiviral vector. In some alternatives, the vector is a foamy viral vector, adenoviral vectors, retroviral vectors or lentiviral vectors.

[0150] In some alternatives, a plasmid vector or a viral vector comprises a first nucleic acid comprising an inducible promoter linked to a polynucleotide coding for a chimeric antigen receptor. In some alternatives, a plasmid vector or viral vector comprises a first nucleic acid sequence comprising a polynucleotide coding for a gene that enhances cell survival or proliferation, a gene that regulates apoptosis, and/or a gene that modulates

checkpoint signaling. In some alternatives, the modulation of checkpoint signaling inhibits negative checkpoint regulators. In some alternatives, the negative checkpoint regulator comprises VISTA, LAG-3 and/or TIM3. Such polynucleotides code for a cytokine, or a chemokine receptor. In some alternatives, a plasmid vector or a viral vector comprises a first nucleic acid comprising an inducible promoter linked to a polynucleotide coding for a marker sequence. Marker sequences are described herein. In some alternatives, the marker sequence is compatible with transduction of human lymphocytes. In some alternatives, the marker sequence allows for selection of transduced cells and/or detection of transduced cells. In some alternatives, the marker is a gene that can include inter alia, hygromycin-B phosphotransferase gene (hph), which confers resistance to hygromycin B, the amino glycoside phosphotransferase gene (neo or aph) from Tn5, which codes for resistance to the antibiotic G418, the dihydrofolate reductase (DHFR) gene, which provides resistance to methotrexate, DHFR dm (exemplary polynucleotide and amino acid sequences in **Table 14**, SEQ ID NO:46 and SEQ ID NO:47, the pac gene that provides resistance to puromycin, Sh ble gene, which inactivates zeocin, the adenosine deaminase gene (ADA), and/or the multi-drug resistance (MDR) gene. A first nucleic acid can include a number of different polynucleotide sequences all under the control of the inducible promoter. For example, a polynucleotide coding for a chimeric antigenic receptor can be linked to a polynucleotide coding for a marker polypeptide and/or a polynucleotide coding for cytokine or chemokine receptor.

[0151] In some alternatives, a lentiviral vector comprises a second nucleic acid comprising a constitutive promoter linked to a nucleic acid sequence coding for transcriptional activator that binds to drug and activates expression of an inducible promoter. In some alternatives, a lentiviral vector with a constitutive promoter can also include a nucleic acid sequence including a marker gene, piggyback transposase, and/or a polynucleotide coding for a chimeric antigen receptor. Each element of the nucleic acid can be separated from one another with a sequence such as a T2A self-cleaving sequence. In some alternatives, the elements of the nucleic acid are separated from one another with a sequence self-cleaving sequence. In some alternatives, the self-cleaving sequence is T2A.

[0152] In other alternatives, the heterogeneous (heterogeneous to the vector, e.g., lentiviral vector) nucleic acid sequence is limited by the amount of additional genetic components that can be packaged in the vector. In some alternatives, a construct contains at least two genes heterogenous to the viral vector. In some alternatives, the construct contains

no more than 4 genes heterogenous to the viral vector. The number of genes heterogenous to the viral vector that can be packaged in the vector can be determined by detecting the expression of one or more transgenes, and selecting vector constructs that provide for transduction of at least 10% of the cells and/or detectable expression levels of the transgene in at least 10% of the cells.

[0153] In some alternatives, a lentivirus is a dual packaged virus. A dual packaged virus contains at least one conditional construct comprising an inducible promoter operably linked to a polynucleotide coding for a chimeric antigen receptor. Optionally the conditional construct comprises a marker gene, a nucleic acid for a cytokine, a nucleic acid for a chemokine receptor. In some alternatives, a dual packaged lentivirus contains a constitutive construct comprising a constitutive promoter. In an alternative, the constitutive construct comprises a constitutive promoter linked to a transcriptional activator for the inducible promoter. In some alternatives, the constitutive construct also includes a marker gene and/or a polynucleotide encoding a cytokine or chemokine. In some alternatives of a system with two constructs, each construct can be packaged in a separate viral vector and the viral vectors can be mixed together for transduction in a cell population.

[0154] When the constitutive and conditional constructs both contain a marker gene, the marker gene on each construct is the same or different from one another. In some alternatives, when the constitutive and conditional constructs both contain a polynucleotide coding for a chimeric antigen receptor, the chimeric antigen receptor can be targeted to the same antigen but have different ligand binding domains, can be targeted to the same antigen but different epitopes, or can be targeted to different antigens.

[0155] In some alternatives, the vector is a minicircle. Minicircles are episomal DNA vectors that are produced as circular expression cassettes devoid of any bacterial plasmid DNA backbone. Their smaller molecular size enables more efficient transfections and offers sustained expression over a period of weeks as compared to standard plasmid vectors that only work for a few days. In some alternatives, a minicircle contains a drug inducible promoter linked to a polynucleotide coding for a chimeric antigen receptor. In some alternatives, the inducible promoter can be linked to chemokine receptor, a marker gene, and/or a cytokine. One or more minicircles can be employed. In some alternatives, a minicircle comprises an inducible promoter linked to a polynucleotide coding for a first chimeric antigen receptor, another minicircle comprises an inducible promoter linked to a polynucleotide coding for a second and different chimeric antigen receptor, and/ or a

minicircle comprises an inducible promoter linked to a polynucleotide coding for a chemokine receptor, a chimeric antigen receptor, and a marker gene. Each element of the constructs is separated by a nucleic acid, such as that coding for a self-cleaving T2A sequence. In some alternatives, each element of the constructs is separated by a nucleic acid, such as that coding for a self-cleaving T2A sequence. In some alternatives each minicircle differs from one another in the chimeric antigen receptor including but not limited to the spacer length and sequence, the intracellular signaling domain, and/or the marker sequence. The minicircle vector can be used with a constitutive lentivirus vector coding for a transcriptional activator for the inducible promoter. In some alternatives, the minicircle vector is used with a constitutive lentivirus vector coding for a transcriptional activator for the inducible promoter.

[0156] In some alternatives, the vector is a piggy bac transposon. The PiggyBac (PB) transposon is a mobile genetic element that efficiently transposes between vectors and chromosomes via a "cut and paste" mechanism. During transposition, the PB transposase recognizes transposon-specific inverted terminal repeat sequences (ITRs) located on both ends of the transposon vector and efficiently moves the contents from the original sites and efficiently integrates them into TTAA chromosomal sites. The powerful activity of the PiggyBac transposon system enables genes of interest between the two ITRs in the PB vector to be easily mobilized into target genomes.

[0157] In some alternatives, a PB contains a drug inducible promoter linked to a polynucleotide coding for a chimeric antigen receptor. In some alternatives, the inducible promoter can be linked to chemokine receptor, a marker gene, and/or a cytokine. One or more PB transposons can be employed. In some alternatives, a PB comprises an inducible promoter linked to a polynucleotide coding for a first chimeric antigen receptor, another PB comprises an inducible promoter linked to a polynucleotide coding for a second and different chimeric antigen receptor, and/ or a PB comprises an inducible promoter linked to a polynucleotide coding for a chemokine receptor, a chimeric antigen receptor, and a marker gene. Each element of the constructs is separated by a nucleic acid, such as that coding for a self-cleaving T2A sequence. In some alternatives each PB differs from one another in the chimeric antigen receptor including but not limited to the spacer length and sequence, the intracellular signaling domain, and/or the marker sequence. The PB vector can be used with a constitutive lentivirus vector coding for a transcriptional activator for the inducible promoter and constitutive vector comprising the piggyback transposase linked to a constitutive promoter.

[0158] In some alternatives, a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; and d) a polynucleotide coding for an intracellular signaling domain; and a second nucleic acid comprising a second constitutive or inducible promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the first and second nucleic acid are in a single lentivirus vector. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor.

[0159] In some alternatives, the first nucleic acid further comprises a marker gene. In some alternatives, the second nucleic acid further comprises polynucleotide coding for a second and different chimeric antigen receptor. The first and second chimeric antigen receptor can differ from one another in the ligand binding domain, the target antigen, an epitope of the target antigen, the spacer domain in length and sequence (short medium or long), and in the intracellular signaling domains.

[0160] In some alternatives, in a single lentivirus construct the first and second nucleic acids can be separated by a genomic insulator nucleic acid such as the sea urchin insulator chromatin domain. In other alternatives, the inducible promoter of the first nucleic acid and the constitutive promoter of the second nucleic acid are in opposite orientation.

[0161] One or more of these vectors can be used in conjunction with one another to transduce target cells and provide for inducible expression of a chimeric antigen receptor.

Host Cells and Compositions: T lymphocyte populations.

[0162] The compositions described herein provide for genetically modified host cells with the vectors and/or constructs as described herein. In some alternatives, the host cells are CD4+ and/or CD8+ T lymphocytes.

[0163] T lymphocytes can be collected in accordance with known techniques and enriched or depleted by known techniques such as affinity binding to antibodies such as flow cytometry and/or immunomagnetic selection. After enrichment and/or depletion steps, *in vitro*

expansion of the desired T lymphocytes can be carried out in accordance with known techniques or variations thereof that will be apparent to those skilled in the art. In some alternatives, the T cells are autologous T cells obtained from the patient.

[0164] For example, the desired T cell population or subpopulation can be expanded by adding an initial T lymphocyte population to a culture medium in vitro, and then adding to the culture medium feeder cells, such as non-dividing peripheral blood mononuclear cells (PBMC), (e.g., such that the resulting population of cells contains at least 5, 10, 20, or 40 or more PBMC feeder cells for each T lymphocyte in the initial population to be expanded); and incubating the culture (e.g. for a time sufficient to expand the numbers of T cells). The non-dividing feeder cells can comprise gamma-irradiated PBMC feeder cells. In some alternatives, the PBMC are irradiated with gamma rays in the range of 3000 to 3600 rads to prevent cell division. In some alternatives, the PBMC are irradiated with gamma rays of 3000, 3100, 3200, 3300, 3400, 3500 or 3600 rads or any value of rads between any two endpoints of any of the listed values to prevent cell division. The order of addition of the T cells and feeder cells to the culture media can be reversed if desired. The culture can typically be incubated under conditions of temperature and the like that are suitable for the growth of T lymphocytes. For the growth of human T lymphocytes, for example, the temperature will generally be at least 25 degrees Celsius, preferably at least 30 degrees, more preferably 37 degrees. In some alternatives, the temperature for the growth of human T lymphocytes is 22, 24, 26, 28, 30, 32, 34, 36, 37 degrees Celsius or any other temperature between any two endpoints of any of the listed values.

[0165] The T lymphocytes expanded include CD8⁺ cytotoxic T lymphocytes (CTL) and CD4⁺ helper T lymphocytes that can be specific for an antigen present on a human tumor or a pathogen. In some alternatives, the cells include precursor T cells. In some alternatives, the cells are hematopoietic stem cells.

[0166] In some alternatives, the expansion method can further comprise adding non-dividing EBV-transformed lymphoblastoid cells (LCL) as feeder cells. LCL can be irradiated with gamma rays in the range of 6000 to 10,000 rads. In some alternatives, the LCL are irradiated with gamma rays in of 6000, 6500, 7000, 7500, 8000, 8500, 9000, 9500 or 10,000 rads or any amount of rads between two endpoints of any of the listed values. The LCL feeder cells can be provided in any suitable amount, such as a ratio of LCL feeder cells to initial T lymphocytes of at least 10:1.

[0167] In some alternatives, the expansion method can further comprise adding anti-CD3 and/or anti CD28 antibody to the culture medium (*e.g.*, at a concentration of at least 0.5 ng/ml). In some alternatives, the expansion method can further comprise adding IL-2 and/or IL-15 to the culture medium (*e.g.*, wherein the concentration of IL-2 is at least 10 units/ml).

[0168] After isolation of T lymphocytes both cytotoxic and helper T lymphocytes can be sorted into naïve, memory, and effector T cell subpopulations either before or after expansion.

[0169] CD8⁺ cells can be obtained by using standard methods. In some alternatives, CD8⁺ cells are further sorted into naïve, central memory, and effector memory cells by identifying cell surface antigens that are associated with each of those types of CD8⁺ cells. In some alternatives, memory T cells are present in both CD62L⁺ and CD62L⁻ subsets of CD8⁺ peripheral blood lymphocytes. PBMC are sorted into CD62L-CD8⁺ and CD62L⁺CD8⁺ fractions after staining with anti-CD8 and anti-CD62L antibodies. In some alternatives, the expression of phenotypic markers of central memory T_{CM} include CD45RO, CD62L, CCR7, CD28, CD3, and/or CD127 and are negative or low for granzyme B. In some alternatives, central memory T cells are CD45RO⁺, CD62L⁺, and/or CD8⁺ T cells. In some alternatives, effector T_E are negative for CD62L, CCR7, CD28, and/or CD127, and positive for granzyme B and/or perforin. In some alternatives, naïve CD8⁺ T lymphocytes are characterized by the expression of phenotypic markers of naïve T cells including CD62L, CCR7, CD28, CD3, CD127, and/or CD45RA.

[0170] CD4⁺ T helper cells are sorted into naïve, central memory, and effector cells by identifying cell populations that have cell surface antigens. CD4⁺ lymphocytes can be obtained by standard methods. In some alternatives, naïve CD4⁺ T lymphocytes are CD45RO⁻, CD45RA⁺, CD62L⁺, and/or CD4⁺ T cells. In some alternatives, central memory CD4⁺ cells are CD62L⁺ and/or CD45RO⁺. In some alternatives, effector CD4⁺ cells are CD62L⁻ and/or CD45RO⁻.

[0171] Whether a cell or cell population is positive for a particular cell surface marker can be determined by flow cytometry using staining with a specific antibody for the surface marker and an isotype matched control antibody. A cell population negative for a marker refers to the absence of significant staining of the cell population with the specific antibody above the isotype control, positive refers to uniform staining of the cell population above the isotype control. In some alternatives, a decrease in expression of one or markers

refers to loss of 1 log₁₀ in the mean fluorescence intensity and/or decrease of percentage of cells that exhibit the marker of at least 20% of the cells, 25% of the cells, 30% of the cells, 35% of the cells, 40% of the cells, 45% of the cells, 50% of the cells, 55% of the cells, 60% of the cells, 65% of the cells, 70% of the cells, 75% of the cells, 80% of the cells, 85% of the cells, 90% of the cell, 95% of the cells, and 100% of the cells or any % between 20 and 100% when compared to a reference cell population. In some alternatives, a cell population positive for one or markers refers to a percentage of cells that exhibit the marker of at least 50% of the cells, 55% of the cells, 60% of the cells, 65% of the cells, 70% of the cells, 75% of the cells, 80% of the cells, 85% of the cells, 90% of the cell, 95% of the cells, or 100% of the cells or any % between 50 and 100% when compared to a reference cell population.

[0172] Whether a cell or cell population is positive for a particular cell surface marker can be determined by flow cytometry using staining with a specific antibody for the surface marker and an isotype matched control antibody. A cell population negative for a marker refers to the absence of significant staining of the cell population with the specific antibody above the isotype control, positive refers to uniform staining of the cell population above the isotype control. In some alternatives, a decrease in expression of one or markers refers to loss of 1 log₁₀ in the mean fluorescence intensity and/or decrease of percentage of cells that exhibit the marker of at least 20% of the cells, 25% of the cells, 30% of the cells, 35% of the cells, 40% of the cells, 45% of the cells, 50% of the cells, 55% of the cells, 60% of the cells, 65% of the cells, 70% of the cells, 75% of the cells, 80% of the cells, 85% of the cells, 90% of the cell, 95% of the cells, and 100% of the cells or any % between 20 and 100% when compared to a reference cell population. In some alternatives, an increase refers to an increase in mean fluorescence intensity and/or to an increase in the number of cells in a cell population that are positive for one or a given marker, such as a population in which s refers to a percentage of cells that exhibit the marker of at least 50% of the cells, 55% of the cells, 60% of the cells, 65% of the cells, 70% of the cells, 75% of the cells, 80% of the cells, 85% of the cells, 90% of the cell, 95% of the cells, or 100% of the cells or any % between 50 and 100% exhibit the marker, e.g., when compared to a reference cell population.

[0173] In some alternatives, populations of CD4⁺ and CD8⁺ that are antigen specific can be obtained by stimulating naïve or antigen specific T lymphocytes with antigen. For example, antigen-specific T cell lines or clones can be generated to Cytomegalovirus antigens by isolating T cells from infected subjects and stimulating the cells in vitro with the same antigen. Naïve T cells can also be used. Any number of antigens from tumor cells can

be utilized as targets to elicit T cell responses. In some alternatives, the adoptive cellular immunotherapy compositions are useful in the treatment of a disease or disorder including a solid tumor, hematologic malignancy, breast cancer or melanoma.

Modification of T lymphocyte populations.

[0174] In some alternatives it can be desired to introduce functional genes into the T cells to be used in immunotherapy in accordance with the present disclosure. For example, the introduced gene or genes can improve the efficacy of therapy by promoting the viability and/or function of transferred T cells; or they can provide a genetic marker to permit selection and/or evaluation of *in vivo* survival or migration; or they can incorporate functions that improve the safety of immunotherapy, for example, by making the cell susceptible to controlled expression of the transgene. This can be carried out in accordance with known techniques that will be apparent to those skilled in the art based upon the present disclosure.

[0175] In some alternatives, T cells are modified with a vector coding for drug inducible chimeric receptors as described herein. In some alternatives, cells are modified with a vector comprising a polynucleotide coding for a chimeric antigen receptor under control of an inducible promoter. In other alternatives, cells are modified with a vector comprising a polynucleotide coding for a cytokine, chemokine receptor, a gene that regulates apoptosis, or a gene that modulates checkpoint signaling under the control of an inducible promoter. In some alternatives, the T cells are obtained from the subject to be treated, in other alternatives, the lymphocytes are obtained from allogeneic human donors, preferably healthy human donors. In some alternatives, the modulation of checkpoint signaling inhibits negative checkpoint regulators. In some alternatives, the negative checkpoint regulator comprises VISTA, LAG-3 and/or TIM3.

[0176] Chimeric receptors can be constructed with a specificity for any cell surface marker by utilizing antigen binding fragments or antibody variable domains of, for example, antibody molecules. The antigen binding molecules can be linked to one or more cell signaling modules. In some alternatives, cell signaling modules include CD3 transmembrane domains, CD3 intracellular signaling domains, and/or CD28 transmembrane domains. In some alternatives, the intracellular signaling domain comprises a CD28 transmembrane and signaling domain linked to a CD3 zeta intracellular domain. In some alternatives, a chimeric receptor can also include a transduction marker such as tEGFR.

[0177] In some alternatives, the same or a different chimeric receptor can be introduced into each of population of CD4+ and/or CD8+ T lymphocytes. In some alternatives, the chimeric receptor in each of these populations has a ligand binding domain that specifically binds to the same ligand on the tumor or infected cell or a different antigen or epitope. The cellular signaling modules can differ. In some alternatives, the intracellular signaling domain of the CD8+ cytotoxic T cells is the same as the intracellular signaling domain of the CD4+ helper T cells. In other alternatives, the intracellular signaling domain of the CD8+ cytotoxic T cells is different than the intracellular signaling domain of the CD4+ helper T cells.

[0178] In some alternatives each of the CD4 or CD8 T lymphocytes can be sorted into naïve, central memory, effector memory or effector cells prior to transduction, as described herein. In some alternatives, each of the CD4 or CD8 T lymphocytes can be sorted into naïve, central memory, effector memory, or effector cells after transduction.

[0179] As described herein, in some alternatives, naïve CD4+ cells are CD45RO-, CD45RA+, CD62L+, and/or CD4+ positive T cells. In some alternatives, central memory CD4+ cells are CD62L positive and/or CD45RO positive. In some alternatives, effector CD4+ cells are CD62L negative and/or CD45RO positive. Each of these populations can be independently modified with a chimeric receptor.

[0180] As described, in some alternatives, memory T cells are present in both CD62L+ and CD62L- subsets of CD8+ peripheral blood lymphocytes. PBMC are sorted into CD62L-CD8+ and CD62L+CD8+ fractions after staining with anti-CD8 and anti-CD62L antibodies. In some alternatives, the expression of phenotypic markers of central memory T cells (TCM) include CD62L, CCR7, CD28, CD3, and/or CD127 and are negative or low for granzyme B. In some alternatives, central memory T cells are CD45RO+, CD62L+, and/or CD8+ T cells. In some alternatives, effector T cells (T_E) are negative for CD62L, CCR7, CD28, and/or CD127, and positive for granzyme B and/or perforin. In some alternatives, naïve CD8+ T lymphocytes are characterized by CD8+, CD62L+, CD45RO+, CCR7+, CD28+ CD127+, and/or CD45RO+. Each of these populations can be independently modified with a chimeric receptor.

[0181] Various transduction techniques have been developed which utilize recombinant infectious virus particles for gene delivery. This represents a currently preferred approach to the transduction of T lymphocytes of the present invention. The viral vectors, which have been used in this way include virus vectors derived from simian virus 40,

adenoviruses, adeno-associated virus (AAV), lentiviral vectors, and/or retroviruses. Thus, gene transfer and expression methods are numerous but essentially function to introduce and express genetic material in mammalian cells. Several of the above techniques have been used to transduce hematopoietic or lymphoid cells, including calcium phosphate transfection, protoplast fusion, electroporation, and infection with recombinant adenovirus, adeno-associated virus and retrovirus vectors. Primary T lymphocytes have been successfully transduced by electroporation and by retroviral or lentiviral infection.

[0182] Retroviral and lentiviral vectors provide a highly efficient method for gene transfer into eukaryotic cells. Moreover, retroviral or lentiviral integration takes place in a controlled fashion and results in the stable integration of one or a few copies of the new genetic information per cell.

[0183] It is contemplated that overexpression of a stimulatory factor (for example, a lymphokine or a cytokine) can be toxic to the treated individual. Therefore, it is within the scope of the invention to include gene segments that cause the T cells of the invention to be susceptible to negative selection *in vivo*. By "negative selection" is meant that the infused cell can be eliminated as a result of a change in the *in vivo* condition of the individual. The negative selectable phenotype can result from the insertion of a gene that confers sensitivity to an administered agent, for example, a compound. Negative selectable genes are known in the art, and include, *inter alia* the following: the Herpes simplex virus type I thymidine kinase (HSV-I TK) gene, which confers ganciclovir sensitivity; the cellular hypoxanthine phosphoribosyltransferase (HPRT) gene, the cellular adenine phosphoribosyltransferase (APRT) gene, and bacterial cytosine deaminase.

[0184] In some alternatives it can be useful to include in the T cells a positive marker that enables the selection of cells of the negative selectable phenotype *in vitro*. The positive selectable marker can be a gene that upon being introduced into the host cell expresses a dominant phenotype permitting positive selection of cells carrying the gene. Genes of this type are known in the art, and include, *inter alia*, hygromycin-B phosphotransferase gene (hph), which confers resistance to hygromycin B, the amino glycoside phosphotransferase gene (neo or aph) from Tn5, which codes for resistance to the antibiotic G418, the dihydrofolate reductase (DHFR) gene, the adenosine deaminase gene (ADA), and/or the multi-drug resistance (MDR) gene.

[0185] A variety of methods can be employed for transducing T lymphocytes, as is well known in the art. In some alternatives, transduction is carried out using lentiviral vectors.

[0186] In some alternatives, CD4⁺ and CD8⁺ cells each can separately be modified with an expression vector encoding a chimeric receptor to form defined populations. In some alternatives, cells can be separately modified with a vector comprising a polynucleotide under the control of a constitutive promoter and a vector comprising a polynucleotide coding for a cytokine or chemokine receptor under control of an inducible promoter.

[0187] In some alternatives, these cells are then further sorted into subpopulations of naïve, central memory and effector cells as described above, by sorting for cell surface antigens unique to each of those cell populations. In addition, CD4⁺ or CD8⁺ cell populations can be selected by their cytokine profile or proliferative activities. For example, CD4⁺ T lymphocytes that have enhanced production of cytokines such as IL-2, IL-4, IL-10, TNF α , and/or IFN γ , as compared to sham transduced cells or transduced CD8⁺ cells when stimulated with antigen can be selected. In other alternatives, naïve or central memory CD4⁺ T cells that have enhanced production of IL-2 and/or TNF α are selected. Likewise, CD8⁺ cells that have enhanced IFN γ production are selected, as compared to sham transduced CD8⁺ cells.

[0188] In some alternatives, CD4⁺ and CD8⁺ cells are selected that are cytotoxic for antigen bearing cells. In some alternatives, CD4⁺ are expected to be weakly cytotoxic as compared to CD8⁺ cells. In a preferred alternative, transduced lymphocytes, such as CD8⁺ central memory cells, are selected that provide for tumor cell killing *in vivo* using an animal model established for the particular type of cancer.

[0189] In yet other alternatives, transduced chimeric receptor expressing T cells are selected that can persist *in vivo* using an animal model established for the particular type of cancer. In some alternatives, transduced chimeric receptor CD8⁺ central memory cells with a short spacer region have been shown to persist *in vivo* after introduction into the animal for 3 days or more, 10 days or more, 20 days or more, 30 days or more, 40 days or more, or 50 days or more.

[0190] The disclosure contemplates that combinations of CD4⁺ and CD8⁺ T cells will be utilized in the compositions. In one alternative, combinations of chimeric receptor transduced CD4⁺ cells can be combined with chimeric receptor transduced CD8⁺ cells of the

same ligand specificity or combined with CD8⁺ T cells that are specific for a distinct tumor ligand. In other alternatives, chimeric receptor transduced CD8⁺ cells are combined with chimeric receptor transduced CD4⁺ cells specific for a different ligand expressed on the tumor. In yet another alternative, chimeric receptor modified CD4⁺ and CD8⁺ cells are combined. In some alternatives CD8⁺ and CD4⁺ cells can be combined in different ratios for example, a 1:1 ratio of CD8⁺ and CD4⁺, a ratio of 10:1 of CD8⁺ to CD4⁺, or a ratio of 100:1 of CD8⁺ to CD4⁺, or any other ratio of CD8⁺ to CD4⁺ that is between any of the listed ratios. In some alternatives, the combined population is tested for cell proliferation in vitro and/or in vivo, and the ratio of cells that provides for proliferation of cells is selected.

[0191] After transduction and/or selection for chimeric receptor bearing cells, the cell populations are preferably expanded in vitro until a sufficient number of cells are obtained to provide for at least one infusion into a human subject, typically around 10⁴ cells/kg to 10⁹ cells/kg. In some alternatives, the transduced cells are cultured in the presence of antigen bearing cells, anti CD3, anti CD28, and IL 2, IL-7, IL 15, or IL-21 or combinations thereof.

[0192] In some alternatives, CD4⁺ and CD8⁺ cells that proliferate in response to cytokine stimulation, antigen or tumor targets in vitro or in vivo are selected. For example, CD4⁺ or CD8⁺ transduced cells that proliferate vigorously when stimulated with antiCD3 and/ or anti-CD28 are selected. In some alternatives, stimulation of transduced cells provides for enhanced transgene expression in the presence of an inducer (e.g. drug) of those trans genes under the control of an inducible promoter.

[0193] Each of the subpopulations of CD4⁺ and CD8⁺ cells can be combined with one another. In a specific alternative, modified naïve or central memory CD4⁺ cells are combined with modified central memory CD8⁺ T cells to provide a synergistic cytotoxic effect on antigen bearing cells, such as tumor cells.

Compositions.

[0194] The disclosure provides for an adoptive cellular immunotherapy composition comprising a genetically modified T lymphocyte cell preparation as described herein. In some alternatives, the T lymphocyte cell preparation comprises CD4⁺ T cells that have a chimeric receptor comprising an extracellular antibody variable domain specific for a ligand associated with the disease or disorder, a spacer region, a transmembrane domain, and an intracellular signaling domain of a T cell receptor or other receptors under the control of a

drug inducible promoter as described herein. In other alternatives, an adoptive cellular immunotherapy composition further comprises a chimeric receptor modified tumor-specific CD8⁺ cytotoxic T lymphocyte cell preparation that provides a cellular immune response, wherein the cytotoxic T lymphocyte cell preparation comprises CD8⁺ T cells that have a chimeric receptor comprising an extracellular single chain antibody specific for a ligand associated with the disease or disorder, a spacer region, a transmembrane domain, and an intracellular signaling domain of a T cell receptor under the control of a drug inducible promoter as described herein. In some alternatives, the chimeric receptor modified T cell population of the disclosure can persist *in vivo* for at least 3 days or longer. In an alternative, each of these populations can be combined with one another or other cell types to provide a composition.

[0195] In some alternatives, the CD4⁺ T helper lymphocyte cell is naïve CD4⁺ T cells, central memory CD4⁺ T cells, effector memory CD4⁺ T cells, or bulk CD4⁺ T cells. In some alternatives, CD4⁺ helper lymphocyte cell is a naïve CD4⁺ T cell, wherein the naïve CD4⁺ T cell comprises a CD45RO⁻, CD45RA⁺, and/or is a CD62L⁺ CD4⁺ T cell.

[0196] In some alternatives, the CD8⁺ T cytotoxic lymphocyte cell is a naïve CD8⁺ T cell, central memory CD8⁺ T cell, effector memory CD8⁺ T cell and/or bulk CD8⁺ T cell. In some alternatives, the CD8⁺ cytotoxic T lymphocyte cell is a central memory T cell, wherein the central memory T cell comprises a CD45RO⁺, CD62L⁺, and/or CD8⁺ T cell. In yet other alternatives, the CD8⁺ cytotoxic T lymphocyte cell is a central memory T cell and the CD4⁺ helper T lymphocyte cell is a naïve or central memory CD4⁺ T cell.

[0197] In some alternatives, the compositions comprise T cell precursors. In some alternatives, the compositions comprise hematopoietic stem cells. In some alternatives, the composition comprises a host cell wherein the host cell is a CD8⁺ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8⁺ T cells, central memory CD8⁺ T cells, effector memory CD8⁺ T cells and bulk CD8⁺ T cells or a CD4⁺ T helper lymphocyte cell that is selected from the group consisting of naïve CD4⁺ T cells, central memory CD4⁺ T cells, effector memory CD4⁺ T cells, and bulk CD4⁺ T cells and a second host cell, wherein the second host cell is a precursor T cell. In some alternatives, the precursor T cell is a hematopoietic stem cell.

Methods.

[0198] The disclosure provides methods of making adoptive immunotherapy compositions and uses or methods of using these compositions for performing cellular immunotherapy in a subject having a disease or disorder. In some alternatives, a method of manufacturing the compositions comprises obtaining a modified naïve or central memory CD4⁺ T helper cell, wherein the modified helper T lymphocyte cell preparation comprises CD4⁺ T cells that have a chimeric receptor comprising a ligand binding domain specific for a tumor cell surface molecule, a spacer domain, a transmembrane domain, and an intracellular signaling domain under control of an inducible promoter as described herein. In other alternatives, CD4⁺ cells have a cytokine or chemokine receptor under the control of an inducible promoter.

[0199] In another alternative, a method further comprises obtaining a modified CD8⁺ central memory T cell, wherein the modified central memory CD8 T lymphocyte cell preparation comprises CD8⁺ cells that have a chimeric receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte, a spacer domain, a transmembrane domain, and an intracellular signaling domain under control of the inducible promoter as described herein. In other alternatives, CD4⁺ cells have a cytokine or chemokine receptor under the control of an inducible promoter.

[0200] The drug inducible promoter in both modified CD4⁺ T cells and modified CD8⁺ cytotoxic T cells can be the same or different. In some alternatives, in one population of cells the promoter linked to the chimeric antigen receptor is a constitutive promoter and in the other population it is an inducible promoter. For example, modified CD4⁺ T cells that have a chimeric receptor comprising a ligand binding domain specific for a tumor cell surface molecule, a spacer domain, a transmembrane domain, and an intracellular signaling domain under control of an constitutive promoter, while the CD8⁺ cytotoxic T cell comprises CD8⁺ cells that have a chimeric receptor comprising a ligand binding domain specific for a tumor cell surface molecule, a spacer domain, a transmembrane domain, and an intracellular signaling domain under control of the inducible promoter.

[0201] In some alternatives, the polynucleotide can code for a chimeric antigen receptor that differs in the CD4⁺ versus the CD8⁺ cell population. The difference between the two constructs can include the specificity or affinity of the ligand binding domain for an

antigen or epitope, the length and sequence of the spacer region, and the intracellular signaling components.

[0202] The preparation of the CD4+ and CD8+ cells that are modified with a chimeric receptor is described throughout this disclosure. Antigen specific T lymphocytes can be obtained from a patient having the disease or disorder or can be prepared by in vitro stimulation of T lymphocytes in the presence of antigen. Subpopulations of CD4+ and/or CD8+ T lymphocytes that are not selected for antigen specificity can also be isolated as described herein and combined in the methods of manufacturing.

[0203] In some alternatives, the combination of cell populations can be evaluated for uniformity of cell surface makers, the ability to proliferate through at least two generations, to have a uniform cell differentiation status. Quality control can be performed by co-culturing a cell line expressing the target ligand with chimeric receptor modified T cells and the drug that induces expression of the chimeric antigen receptor to determine if the chimeric receptor modified T cells recognize the cell line using cytotoxicity, proliferation, or cytokine production assays in the presence of the inducer that are known in the field. Cell differentiation status and cell surface markers on the chimeric receptor modified T cells can be determined by flow cytometry. In some alternatives, the markers and cell differentiation status on the CD8+ cells include CD3, CD8, CD62L, CD28, CD27, CD69, CD25, PD-1, CTLA-4, CD45RO, and/or CD45RA. In some alternatives, the markers and the cell differentiation status on the CD4+ cells include CD3, CD4, CD62L, CD28, CD27, CD69, CD25, PD-1, CTLA-4 CD45RO, and/or CD45RA.

[0204] In some alternatives, the chimeric receptor modified T cells as described herein are able to persist *in vivo* for at least 3 days, or at least 10 days. In some alternatives, the chimeric receptor modified T cells as described herein, can proliferate *in vivo* through at least 2, or at least 3 generations as determined by CFSE dye dilution. Proliferation and persistence of the chimeric receptor modified T cells can be determined by using an animal model of the disease or disorder and administering the cells and determining persistence and/or proliferative capacity of the transferred cells. In other alternatives, proliferation and activation can be tested in vitro by going through multiple cycles of activation with antigen bearing cells.

[0205] The disclosure also provides methods of performing cellular immunotherapy in a subject having a disease or disorder comprising: administering a

composition of lymphocytes expressing a chimeric receptor under the control of a drug inducible promoter as described herein, and administering the drug.

[0206] In some alternatives, the drug is tamoxifen, variants, derivatives, pharmaceutical salts, solvates, and hydrates thereof as described herein. In some alternatives, the drug is delivered prior to, at the same time as the composition, or at later time points after the composition has been administered.

[0207] In some alternatives, the drug is administered with the composition, and if a toxic effect of the composition is observed the drug is withdrawn until the toxic effects diminish. After the symptoms of toxicity diminish, the drug is administered again. In some alternatives, a drug can be administered again once symptoms of toxicity diminish.

[0208] In some alternatives, the drug is administered with the composition but once the subject has a decrease in the tumor load or cancer cells, the drug is withdrawn for a period of time to allow the modified cells to rest and if there is no activity of the modified cells, the modified cells are not needed because of remission of the cancer.

[0209] In other alternatives, a method comprises administering to the subject a genetically modified cytotoxic T lymphocyte cell preparation that provides a cellular immune response, wherein the cytotoxic T lymphocyte cell preparation comprises CD8 + T cells that have a chimeric receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte, a spacer domain, a transmembrane domain, and an intracellular signaling domain under the control of a drug inducible promoter as described herein, and/or a genetically modified helper T lymphocyte cell preparation that elicits direct tumor recognition and augments the genetically modified cytotoxic T lymphocyte cell preparations ability to mediate a cellular immune response, wherein the helper T lymphocyte cell preparation comprises CD4+ T cells that have a chimeric receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte, a spacer domain, a transmembrane domain, and an intracellular signaling domain under control of a constitutive or drug inducible promoter as described herein and administering the drug that induces the inducible promoter. In some alternatives, the administering of the drug is performed after

administering of the composition or host cells, wherein administering is performed 1 day, 2 days, 3 days, 4 days, 5 days, 6 days, 7 days, 2 weeks, 4 weeks or two months, or any time in between any two values of time listed.

[0210] In other alternatives, a method comprises administering to the subject a genetically modified cytotoxic T lymphocyte cell preparation that provides a cellular immune response, wherein the cytotoxic T lymphocyte cell preparation comprises CD8 + T cells that have a chimeric receptor comprising a ligand binding domain specific for a tumor cell surface molecule, a spacer domain, a transmembrane domain, and an intracellular signaling domain under the control of a constitutive promoter as described herein, and/or a genetically modified helper T lymphocyte cell preparation that elicits direct tumor recognition and augments the genetically modified cytotoxic T lymphocyte cell preparations ability to mediate a cellular immune response, wherein the helper T lymphocyte cell preparation comprises CD4+ T cells that have a chimeric receptor comprising a ligand binding domain specific for a tumor cell surface molecule, a spacer domain, a transmembrane domain, and an intracellular signaling domain under control of a constitutive or drug inducible promoter as described herein and administering the drug that induces the inducible promoter. In some alternatives, the tumor specific molecule is a tumor surface molecule.

[0211] In other alternatives, a method comprises administering to the subject a genetically modified cytotoxic T lymphocyte cell preparation that provides a cellular immune response, wherein the cytotoxic T lymphocyte cell preparation comprises CD8 + T cells that express a cytokine, chemokine receptor, a polypeptide that regulates apoptosis, and/or a polypeptide that modulates checkpoint signaling under the control of an inducible promoter as described herein, and/or a genetically modified helper T lymphocyte cell preparation that elicits direct tumor recognition and augments the genetically modified cytotoxic T lymphocyte cell preparations ability to mediate a cellular immune response, wherein the helper T lymphocyte cell preparation comprises CD4+ T cells that express a cytokine, chemokine receptor, a polypeptide that regulates apoptosis, and/or a polypeptide that modulates checkpoint signaling under control of a constitutive or drug inducible promoter as described herein and administering the drug that induces the inducible promoter. In some alternatives, one or more of the cell populations expresses a chimeric antigen receptor under the control of a constitutive promoter. In some alternatives, the modulation of checkpoint signaling inhibits negative checkpoint regulators. In some alternatives, the negative checkpoint regulator comprises VISTA, LAG-3 and/or TIM3.

[0212] An effective amount of the drug for induction is an amount of the drug that provides for induction of the chimeric antigen receptor in at least 5 %, at least 10%, at least 20%, at least 30%, at least 40%, at least 50%, at least 60%, at least 70%, at least 80%, at least 90%, at least 100% or any number in between any of the listed percent values of the transduced cells.

[0213] Another alternative describes a method of performing cellular immunotherapy in a subject having a disease or disorder comprising: analyzing a biological sample of the subject for the presence of a target molecule associated with the disease or disorder and administering the adoptive immunotherapy compositions described herein and administering the drug that induces the inducible promoter, wherein the chimeric receptor specifically binds to the target molecule.

[0214] Subjects that can be treated by the present invention are, in general, human and other primate subjects, such as monkeys and apes for veterinary medicine purposes. The subjects can be male or female and can be any suitable age, including infant, juvenile, adolescent, adult, and geriatric subjects.

[0215] The methods are useful in the treatment or inhibition of, for example, hematologic malignancy, melanoma, breast cancer, brain cancer, and other epithelial malignancies or solid tumors. In some alternatives, the molecule associated with the disease or disorder is an orphan tyrosine kinase receptor ROR1, Her2, EGFR, CE7, hB7H3, CD19, CD20, CD22, mesothelin, CEA, or a hepatitis B surface antigen.

[0216] Subjects that can be addressed using the methods described herein include subjects identified or selected as having cancer, including but not limited to colon, lung, liver, breast, renal, prostate, ovarian, skin (including melanoma), bone, and brain cancer, etc. Such identification and/or selection can be made by clinical or diagnostic evaluation. In some alternatives the tumor associated antigens or molecules are known, such as melanoma, breast cancer, brain cancer, squamous cell carcinoma, colon cancer, leukemia, myeloma, and/or prostate cancer. In other alternatives the tumor associated molecules can be targeted with genetically modified T cells expressing an engineered chimeric receptor. Examples include but are not limited to B cell lymphoma, breast cancer, brain cancer, prostate cancer, and/or leukemia.

[0217] Cells prepared as described above can be utilized in methods and compositions for adoptive immunotherapy in accordance with known techniques, or

variations thereof that will be apparent to those skilled in the art based on the instant disclosure.

[0218] In some alternatives, the cells are formulated by first harvesting them from their culture medium, and then washing and concentrating the cells in a medium and container system suitable for administration (a "pharmaceutically acceptable" carrier) in a treatment-effective amount. Suitable infusion medium can be any isotonic medium formulation, typically normal saline, Normosol R (Abbott) or Plasma-Lyte A (Baxter), but also 5% dextrose in water or Ringer's lactate can be utilized. The infusion medium can be supplemented with human serum albumin, fetal bovine serum or other human serum components.

[0219] In some alternatives, a treatment or inhibitory effective amount of cells in the composition is a transduced CD4 or CD8 cell or at least 2 cell subsets (for example, 1 CD8+ central memory T cell subset and 1 CD4+ helper T cell subset) or is more typically greater than 10^2 cells, and up to 10^6 , up to and including 10^8 or 10^9 cells and can be more than 10^{10} cells. The number of cells will depend upon the ultimate use for which the composition is intended as will the type of cells included therein. For example, if cells that are specific for a particular antigen are desired, then the population will contain greater than 70%, generally greater than 80%, 85% and 90-95% of such cells. For uses provided herein, the cells are generally in a volume of a liter or less, can be 500 mls or less, even 250 mls or 100 mls or less or a volume in between any two listed volume values. Hence the density of the desired cells is typically greater than 10^4 cells/ml and generally is greater than 10^7 cells/ml, generally 10^8 cells/ml or greater. The clinically relevant number of immune cells can be apportioned into multiple infusions that cumulatively equal or exceed 10^6 , 10^7 , 10^8 , 10^8 , 10^9 , 10^{10} or 10^{11} cells or any amount of cells defined between any two endpoints of any of the listed values.

[0220] In some alternatives, the lymphocytes of the invention can be used to confer immunity to individuals. By "immunity" is meant a lessening of one or more physical symptoms associated with a response to infection by a pathogen, or to a tumor, to which the lymphocyte response is directed. The amount of cells administered is usually in the range present in normal individuals with immunity to the pathogen. Thus, the cells are usually administered by infusion, with each infusion in a range of from 2 cells, up to at least 10^6 to 3×10^{10} cells, preferably in the range of at least 10^7 to 10^9 cells. The T cells can be administered by a single infusion, or by multiple infusions over a range of time. However, since different individuals are expected to vary in responsiveness, the type and amount of

cells infused, as well as the number of infusions and the time range over which multiple infusions are given are determined by the attending physician, and can be determined by routine examination. The generation of sufficient levels of T lymphocytes (including cytotoxic T lymphocytes and/or helper T lymphocytes) is readily achievable using the rapid expansion method of the present invention, as exemplified herein.

[0221] In some alternatives, a composition as described herein is administered to an identified or selected subject, such as a subject identified or selected as having melanoma, breast cancer, brain cancer, squamous cell carcinoma, colon cancer, leukemia, myeloma, and/or prostate cancer, intravenously, intraperitoneally, intratumorally, into the bone marrow, into the lymph node, and /or into cerebrospinal fluid. In some alternatives, the chimeric receptor engineered compositions are delivered to the site of the tumor. Alternatively, the compositions as described herein can be combined with a compound that targets the cells to the tumor or the immune system compartments and avoid sites such as the lung.

[0222] In some alternatives, the compositions as described herein are administered with chemotherapeutic agents and/or immunosuppressants. In an alternative, a patient is first administered a chemotherapeutic agent that inhibits or destroys other immune cells followed by the compositions described herein. In some cases, chemotherapy can be avoided entirely.

[0223] In some alternatives, a method comprising administering the modified T cells as described herein in combination with the inducer (e.g. inducible drug) until the tumor burden is diminished. Once the tumor burden is diminished, the inducer drug can be withdrawn in order to switch the expression of the chimeric antigen receptor off and decrease the number of T cell expressing the receptor. In other alternatives, the inducer drug can be administered at different time in order to switch the expression of the chimeric antigen receptor on in the event of a relapse or increase in tumor growth.

[0224] In other alternatives, the inducer drug can be given for a period of time of days, weeks, or months, and then withdrawn for days, weeks or months, followed by re-administration of the inducer drug for days, weeks or months to allow for cycling of the expression of the chimeric antigen receptor to avoid anergy or nonresponsiveness due to chronic stimulation of the cells.

Vector construction and preparation of dual packaged lentivirus.

[0225] An inducible lentiviral vector encoding 7xHBD/mE1b-CD19t-her2t-T2A-epHIV7) was constructed. CD19t specific chimeric receptors were constructed using: (1) the VL and VH chain segments of the CD19-specific mAb FMC63 (SEQ ID NO: 3), linked by a (G₄S)₃ linker (SEQ ID NO: 12) peptide (VL-linker-VH); (2) a spacer domain derived from IgG4-Fc Hinge only (12 AA encoded by (SEQ ID NO: 4)). Spacers contained a S → P substitution within the Hinge domain located at position 108 of the native IgG4-Fc protein; the 27 AA transmembrane domain of human CD28 (Uniprot Database: P10747, (SEQ ID NO: 14)); (4) a signaling module comprising either (i) the 41 AA cytoplasmic domain of human CD28 with an LL → GG substitution located at position 186-187 of the native CD28 protein (SEQ ID NO: 14) ; and/or (ii) the 42 AA cytoplasmic domain of human 4-1BB (Uniprot Database: Q07011, (SEQ ID NO: 15)); linked to (iii) the 112 AA cytoplasmic domain of isoform 3 of human CD3ζ (Uniprot Database: P20963, (SEQ ID NO: 16)).

[0226] The nucleic acid sequences coding for the CD19t were linked with sequences coding for Her2t (SEQ ID NO: 44); and the self-cleaving T2A sequence (SEQ ID NO: 8).

[0227] A conditional lentiviral vector encoding 7xHBD/mEF1ap-ZsGreen-epHIV7 was constructed. The synthetic promoter 7xHBD/mEF1ap was constructed by combining seven minimal hepatocyte nuclear family-1 (HNF-1) binding sites cloned from the human albumin promoter and the huEF1α promoter TATA box and has a sequence of (SEQ ID NO: 41). In this way, only in the presence of tamoxifen does binding of HEA-3 to 7xHBD/EF1mp promoter induce the “ON” state of transgene expression.

[0228] A conditional lentiviral vector encoding 7xHBD/mEF1ap-CD19t-T2A-DHFRdm_epHIV7 was constructed. CD19t specific chimeric receptors were constructed using: (1) the VL and VH chain segments of the CD19-specific mAb FMC63 (SEQ ID NO: 3), linked by a (G₄S)₃ linker (SEQ ID NO: 12) peptide (VL-linker-VH); (2) a spacer domain derived from IgG4-Fc Hinge only (12 AA encoded by (SEQ ID NO: 4)). Spacers contained a S → P substitution within the Hinge domain located at position 108 of the native IgG4-Fc protein; the 27 AA transmembrane domain of human CD28 (Uniprot Database: P10747, (SEQ ID NO: 14)); (4) a signaling module comprising either (i) the 41 AA cytoplasmic domain of human CD28 with an LL → GG substitution located at position 186-

187 of the native CD28 protein (SEQ ID NO: 14) ; and/or (ii) the 42 AA cytoplasmic domain of human 4-1BB (Uniprot Database: Q07011, (SEQ ID NO: 15)); linked to (iii) the 112 AA cytoplasmic domain of isoform 3 of human CD3 ζ (Uniprot Database: P20963, (SEQ ID NO: 16)).

[0229] The nucleic acid sequences coding for the CD19t were linked with sequences coding for the self-cleaving T2A sequence (SEQ ID NO: 8); and DHFRdm (SEQ ID NO: 46)

[0230] The transcriptional regulator, HEA-3, is a chimeric transcription factor composed of human subunits including the N-terminal DNA binding domain of Hepatocyte Nuclear Factor 1-alpha (HNF-1 α) fused in frame to the mutant tamoxifen-specific ligand binding domain of the estrogen receptor ligand binding domain (ER-LBD), that is in turn fused to the p65 activation domain of NF- κ B (p65). In the absence of tamoxifen, HEA-3 is excluded from the nucleus by binding of cytosolic heat-shock protein 90 (HSP90) to the tamoxifen binding active site and transgene expression is in the "OFF" state. Nanomolar concentrations of cytosolic tamoxifen actively outcompete HSP90 for ER-LBD binding, resulting in HEA-3 translocation to the nucleus. Upon nuclear translocation, HEA-3 is readily available to bind its restricted synthetic promoter. Transcriptional responsiveness to HEA-3 in the presence of tamoxifen is achieved when transgenes are placed behind an HEA-3 responsive synthetic promoter (7xHBD/EF1mp).

[0231] A constitutive construct was constructed with a constitutive promoter EF-1 α linked to a polynucleotide coding for a transcriptional activator HEA3 (SEQ ID NO: 39) and a marker sequence EGFRt (SEQ ID NO: 9).

[0232] Human codon-optimized nucleotide sequences encoding each transgene were synthesized (LifeTechnologies, Carlsbad, CA) and cloned into the ePHIV7 lentiviral vector using NheI and NotI restriction sites. The ePHIV7 lentiviral vector had been derived from the pHIV7 vector by replacing the cytomegalovirus promoter of pHIV7 with an EF-1 promoter.

[0233] The inducible CD19 chimeric receptor-encoding and the constitutive lentivirus was produced in 293T cells co-transfected with the lentiviral vector and the packaging vectors pCHGP-2, pCMV-Rev2 and pCMV-G using Calphos transfection reagent (Clontech). Medium was changed 16 hours after transfection, and lentivirus collected after 24, 48 and 72 hours.

Generation of Jurkat T-cell lines expressing the CD19 chimeric receptors and ZsGreen when induced with tamoxifen.

[0234] Jurkat cells were transduced with lentiviral supernatant (MOI = 3) supplemented with 1 µg/mL polybrene (Millipore) on day 3 after activation by centrifugation at 2,100 rpm for 45 minutes at 32°C. T cells were expanded in RPMI, 10% human serum, 2 mM L-glutamine and 1 % penicillin-streptomycin (CTL medium), supplemented with recombinant human (rh) IL-2 to a final concentration of 50 U/mL every 48 hours.

Additional Alternatives.

[0235] In some alternatives, system for inducible expression of a chimeric antigen receptor is provided, wherein the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized a polynucleotide coding for a transmembrane domain; and d) a polynucleotide coding for an intracellular signaling domain; and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. . In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor. In some alternatives, the drug is tamoxifen and/or its metabolites. In some alternatives, the first promoter comprises a nucleic acid sequence of SEQ ID NO: 41. In some alternatives, the second promoter is a constitutive promoter. In some alternatives, the second promoter is the EF1αp. In some alternatives, the transcriptional activator comprises a sequence of SEQ ID NO: 40. In some alternatives, the first nucleic acid further comprises a first vector and the second nucleic acid further comprises a second vector. In some alternatives, both vectors are packaged in a viral vector. In some alternatives, the viral vector is a lentivirus. In some alternatives, the first and second nucleic acid comprise a vector. In some alternatives, the first nucleic acid further comprises a nucleic acid sequence coding for a

selectable marker. In some alternatives, the second nucleic acid further comprises a nucleic acid coding for a selectable marker.

[0236] In some alternatives, a system for inducible expression of chimeric antigen receptor comprises: a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a cytokine, a chemokine, a polypeptide that regulates apoptosis and/or a polypeptide that modulates checkpoint signaling; and a second nucleic acid comprising a second promoter operably linked to a nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the second promoter is constitutive or inducible. In an exemplary alternative, the second nucleic acid further comprises a polynucleotide coding for a chimeric antigen receptor, under the control of a constitutive promoter.

[0237] In another aspect, the present disclosure provides compositions to confer and/or augment immune responses mediated by cellular immunotherapy, such as by adoptively transferring tumor-specific, subset specific genetically modified CD4⁺ T cells, wherein the CD4⁺ T cells confer and/or augment the ability of CD8⁺ T cells to sustain anti-tumor reactivity and increase and/or maximize tumor-specific proliferation. In some alternatives, the CD4⁺ cells are genetically modified to express a chimeric receptor nucleic acid and/or chimeric receptor polypeptide under the control of a regulated promoter as described herein.

[0238] In another aspect, the present disclosure provides compositions to confer and/or augment immune responses mediated by cellular immunotherapy, such as by adoptively transferring tumor-specific, subset specific genetically modified CD8⁺ T cells. In some alternatives, the CD8⁺ T cells express a chimeric receptor nucleic acid and/or chimeric receptor polypeptide under the control of a regulated promoter, as described herein.

[0239] In one alternative, the present invention provides a method of performing cellular immunotherapy in a subject having a disease or disorder by administering to the subject a genetically modified T lymphocyte cell preparation that provides a cellular immune response and administering a drug that induces a transgene in the genetically modified T lymphocyte cells.

[0240] In some alternatives, the genetically modified CD8⁺ and genetically modified CD4⁺ cell population are co-administered. In some alternatives, the T cells are autologous or allogeneic T cells. Various modifications of the above method are possible. For

example, the chimeric receptor that is expressed by the CD4⁺ T cell and the CD8⁺ T cell can be the same or different.

[0241] In some alternatives, a system for inducible expression of a chimeric antigen receptor is provided, wherein the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain; and d) a polynucleotide coding for an intracellular signaling domain; and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the second promoter is constitutive or inducible. In some alternatives, the drug is tamoxifen and/or its metabolites. In some alternatives, the first promoter comprises a nucleic acid sequence of SEQ ID NO: 41. In some alternatives, the second promoter is a constitutive promoter. In some alternatives, the second promoter is the EF1 α p. In some alternatives, the transcriptional activator comprises a sequence of SEQ ID NO: 40. In some alternatives, the first nucleic acid further comprises a first vector and the second nucleic acid further comprises a second vector. In some alternatives, both vectors are packaged in a viral vector. In some alternatives, the viral vector is a lentivirus. In some alternatives, the first and second nucleic acid comprise a vector. In some alternatives, the first nucleic acid further comprises a nucleic acid sequence coding for a selectable marker. In some alternatives, the second nucleic acid further comprises a nucleic acid coding for a selectable marker. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor.

[0242] In some alternatives, a system for inducible expression of chimeric antigen receptor is provided, wherein the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a cytokine, a chemokine receptor, a polypeptide that regulates apoptosis, or a polypeptide that modulates checkpoint signaling; and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional

activator for the inducible promoter. In some alternatives, the polypeptide that modulates checkpoint signaling inhibits negative checkpoint regulators. In some alternatives, the negative checkpoint regulator comprises VISTA, LAG-3 and/or TIM3. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for chimeric antigen receptor comprising a ligand binding domain, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor; a polynucleotide coding for a transmembrane domain. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment. In some alternatives, the ligand binding domain is single chain variable fragment. In some alternatives, the tumor specific molecule is CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, or MAGE A3 TCR or combinations thereof. In some alternatives, the second promoter is an inducible promoter or a constitutive promoter.

[0243] In some alternatives, a chimeric receptor polypeptide is provided, wherein the chimeric receptor polypeptide is coded for by a system. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte, a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain, and d) a polynucleotide coding for an intracellular signaling domain, and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter.. In some alternatives, the drug is tamoxifen and/or its metabolites. In some alternatives, the first promoter comprises a nucleic acid sequence of SEQ ID NO: 41. In some alternatives, the second promoter is a constitutive promoter. In some alternatives, the second promoter is the EF1 α p. In some alternatives, the

transcriptional activator comprises a sequence of SEQ ID NO: 40. In some alternatives, the first nucleic acid further comprises a first vector and the second nucleic acid further comprises a second vector. In some alternatives, both vectors are packaged in a viral vector. In some alternatives, the viral vector is a lentivirus. In some alternatives, the first and second nucleic acid comprise a vector. In some alternatives, the first nucleic acid further comprises a nucleic acid sequence coding for a selectable marker. In some alternatives, the second nucleic acid further comprises a nucleic acid coding for a selectable marker. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a cytokine, a chemokine receptor, a polypeptide that regulates apoptosis, or a polypeptide that modulates checkpoint signaling and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the polypeptide that modulates checkpoint signaling inhibits negative checkpoint regulators. In some alternatives, the negative checkpoint regulator comprises VISTA, LAG-3 and/or TIM3. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for chimeric antigen receptor comprising a ligand binding domain, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment. In some alternatives, the tumor specific molecule is selected from the group consisting of CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, and MAGE A3 TCR or combinations thereof. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a cytokine, a chemokine receptor, a polypeptide that regulates apoptosis, or a polypeptide that modulates checkpoint signaling; and b) a second nucleic acid comprising a second constitutive or inducible promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In

some alternatives, the polypeptide that modulates checkpoint signaling inhibits negative checkpoint regulators. In some alternatives, the negative checkpoint regulator comprises VISTA, LAG-3 and/or TIM3. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for chimeric antigen receptor comprising a ligand binding domain, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor; a polynucleotide coding for a transmembrane domain. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment. In some alternatives, the ligand binding domain is single chain variable fragment. In some alternatives, the tumor specific molecule is selected from the group consisting of CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, and MAGE A3 TCR or combinations thereof. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor. In some alternatives, the second promoter is an inducible promoter. In some alternatives, the second promoter is constitutive promoter.

[0244] In some alternatives, a host cell is provided, wherein the host cell comprises a system. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte, a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain, and d) a polynucleotide coding for an intracellular signaling domain, and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter.. In some alternatives, the drug is tamoxifen and/or its metabolites. In some alternatives, the first promoter comprises a nucleic acid sequence of

SEQ ID NO: 41. In some alternatives, the second promoter is a constitutive promoter. In some alternatives, the second promoter is the EF1 α p. In some alternatives, the transcriptional activator comprises a sequence of SEQ ID NO: 40. In some alternatives, the first nucleic acid further comprises a first vector and the second nucleic acid further comprises a second vector. In some alternatives, both vectors are packaged in a viral vector. In some alternatives, the viral vector is a lentivirus. In some alternatives, the first and second nucleic acid comprise a vector. In some alternatives, the first nucleic acid further comprises a nucleic acid sequence coding for a selectable marker. In some alternatives, the second nucleic acid further comprises a nucleic acid coding for a selectable marker. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a cytokine, a chemokine receptor, a polypeptide that regulates apoptosis, or a polypeptide that modulates checkpoint signaling and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the polypeptide that modulates checkpoint signaling inhibits negative checkpoint regulators. In some alternatives, the negative checkpoint regulator comprises VISTA, LAG-3 and/or TIM3. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for chimeric antigen receptor comprising a ligand binding domain, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment. In some alternatives, the tumor specific molecule is selected from the group consisting of CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, and MAGE A3 TCR or combinations thereof. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a cytokine, a chemokine receptor, a polypeptide that regulates apoptosis, or a polypeptide that modulates checkpoint signaling; and b) a second nucleic acid comprising a

second constitutive or inducible promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the polypeptide that modulates checkpoint signaling inhibits negative checkpoint regulators. In some alternatives, the negative checkpoint regulator comprises VISTA, LAG-3 and/or TIM3. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for chimeric antigen receptor comprising a ligand binding domain, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor; a polynucleotide coding for a transmembrane domain. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment. In some alternatives, the ligand binding domain is single chain variable fragment. In some alternatives, the tumor specific molecule is selected from the group consisting of CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, and MAGE A3 TCR or combinations thereof. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor. In some alternatives, the second promoter is an inducible promoter. In some alternatives, the second promoter is constitutive promoter. In some alternatives, the tumor specific molecule is selected from the group consisting of CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, and MAGE A3 TCR or combinations thereof. In some alternatives, the host cell is a CD8⁺ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8⁺ T cells, central memory CD8⁺ T cells, effector memory CD8⁺ T cells and bulk CD8⁺ T cells. In some alternatives, the host cell is a CD4⁺ T helper lymphocyte cell that is selected from the group consisting of naïve CD4⁺ T cells, central memory CD4⁺ T cells, effector memory CD4⁺ T cells, and bulk CD4⁺ T cells. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor. In some alternatives, the host cell is a precursor T cell. In some alternatives, the precursor T cell is a hematopoietic stem cell.

[0245] In some alternatives, a composition is provided, wherein the composition comprises a host cell in a pharmaceutically acceptable excipient. In some alternatives, the host cell comprises a system. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte, a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain, and d) a polynucleotide coding for an intracellular signaling domain, and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter.. In some alternatives, the drug is tamoxifen and/or its metabolites. In some alternatives, the first promoter comprises a nucleic acid sequence of SEQ ID NO: 41. In some alternatives, the second promoter is a constitutive promoter. In some alternatives, the second promoter is the EF1 α p. In some alternatives, the transcriptional activator comprises a sequence of SEQ ID NO: 40. In some alternatives, the first nucleic acid further comprises a first vector and the second nucleic acid further comprises a second vector. In some alternatives, both vectors are packaged in a viral vector. In some alternatives, the viral vector is a lentivirus. In some alternatives, the first and second nucleic acid comprise a vector. In some alternatives, the first nucleic acid further comprises a nucleic acid sequence coding for a selectable marker. In some alternatives, the second nucleic acid further comprises a nucleic acid coding for a selectable marker. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a cytokine, a chemokine receptor, a polypeptide that regulates apoptosis, or a polypeptide that modulates checkpoint signaling and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the polypeptide that modulates checkpoint signaling inhibits negative checkpoint regulators. In some alternatives, the negative checkpoint regulator comprises VISTA, LAG-3 and/or TIM3. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for chimeric antigen receptor comprising a ligand binding domain, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population,

wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment. In some alternatives, the tumor specific molecule is selected from the group consisting of CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, and MAGE A3 TCR or combinations thereof. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a cytokine, a chemokine receptor, a polypeptide that regulates apoptosis, or a polypeptide that modulates checkpoint signaling; and b) a second nucleic acid comprising a second constitutive or inducible promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the polypeptide that modulates checkpoint signaling inhibits negative checkpoint regulators. In some alternatives, the negative checkpoint regulator comprises VISTA, LAG-3 and/or TIM3. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for chimeric antigen receptor comprising a ligand binding domain, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor; a polynucleotide coding for a transmembrane domain. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment. In some alternatives, the ligand binding domain is single chain variable fragment. In some alternatives, the tumor specific molecule is selected from the group consisting of CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, and MAGE A3 TCR or combinations thereof. In some alternatives, the spacer is optimized for increased T cell proliferation and/or

cytokine production in response to the ligand as compared to a reference chimeric receptor. In some alternatives, the second promoter is an inducible promoter. In some alternatives, the second promoter is constitutive promoter. In some alternatives, the host cell is a CD8⁺ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8⁺ T cells, central memory CD8⁺ T cells, effector memory CD8⁺ T cells and bulk CD8⁺ T cells. In some alternatives, the host cell is a CD4⁺ T helper lymphocyte cell that is selected from the group consisting of naïve CD4⁺ T cells, central memory CD4⁺ T cells, effector memory CD4⁺ T cells, and bulk CD4⁺ T cells. In some alternatives, the composition comprises a host cell wherein the host cell is a CD8⁺ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8⁺ T cells, central memory CD8⁺ T cells, effector memory CD8⁺ T cells and bulk CD8⁺ T cells and further comprises another host cell wherein the host cell is a CD4⁺ T helper lymphocyte cell that is selected from the group consisting of naïve CD4⁺ T cells, central memory CD4⁺ T cells, effector memory CD4⁺ T cells, and bulk CD4⁺ T cells. In some alternatives, the composition comprises a host cell wherein the host cell is a CD8⁺ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8⁺ T cells, central memory CD8⁺ T cells, effector memory CD8⁺ T cells and bulk CD8⁺ T cells or a CD4⁺ T helper lymphocyte cell that is selected from the group consisting of naïve CD4⁺ T cells, central memory CD4⁺ T cells, effector memory CD4⁺ T cells, and bulk CD4⁺ T cells and a second host cell, wherein the second host cell is a precursor T cell. In some alternatives, the precursor T cell is a hematopoietic stem cell.

[0246] In some alternatives, an in vitro method for preparing a host cell is provided wherein the method comprises a) providing a system and b) introducing the system into a separate isolated T lymphocyte population and expanding each T lymphocyte population in vitro. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte, a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain, and d) a polynucleotide coding for an intracellular signaling domain, and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional

activator for the inducible promoter.. In some alternatives, the drug is tamoxifen and/or its metabolites. In some alternatives, the first promoter comprises a nucleic acid sequence of SEQ ID NO: 41. In some alternatives, the second promoter is a constitutive promoter. In some alternatives, the second promoter is the EF1 α p. In some alternatives, the transcriptional activator comprises a sequence of SEQ ID NO: 40. In some alternatives, the first nucleic acid further comprises a first vector and the second nucleic acid further comprises a second vector. In some alternatives, both vectors are packaged in a viral vector. In some alternatives, the viral vector is a lentivirus. In some alternatives, the first and second nucleic acid comprise a vector. In some alternatives, the first nucleic acid further comprises a nucleic acid sequence coding for a selectable marker. In some alternatives, the second nucleic acid further comprises a nucleic acid coding for a selectable marker. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a cytokine, a chemokine receptor, a polypeptide that regulates apoptosis, or a polypeptide that modulates checkpoint signaling and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the polypeptide that modulates checkpoint signaling inhibits negative checkpoint regulators. In some alternatives, the negative checkpoint regulator comprises VISTA, LAG-3 and/or TIM3. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for chimeric antigen receptor comprising a ligand binding domain, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment. In some alternatives, the tumor specific molecule is selected from the group consisting of CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, and MAGE A3 TCR or combinations thereof. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide

coding for a cytokine, a chemokine receptor, a polypeptide that regulates apoptosis, or a polypeptide that modulates checkpoint signaling; and b) a second nucleic acid comprising a second constitutive or inducible promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the polypeptide that modulates checkpoint signaling inhibits negative checkpoint regulators. In some alternatives, the negative checkpoint regulator comprises VISTA, LAG-3 and/or TIM3. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for chimeric antigen receptor comprising a ligand binding domain, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor; a polynucleotide coding for a transmembrane domain. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment. In some alternatives, the ligand binding domain is single chain variable fragment. In some alternatives, the tumor specific molecule is selected from the group consisting of CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, and MAGE A3 TCR or combinations thereof. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor. In some alternatives, the second promoter is an inducible promoter. In some alternatives, the second promoter is constitutive promoter. In some alternatives, wherein the T lymphocytes are expanded, the method further comprises culturing the cells in the presence of anti-CD3 and/or anti CD28, and at least one homeostatic cytokine until the cells expand sufficiently for use as a cell infusion. In some alternatives, the lymphocyte is CD8+ or CD4+. In some alternatives, the cells are precursor T cells. In some alternatives, the cells are hematopoietic stem cells. In some alternatives, the composition comprises a host cell wherein the host cell is a CD8+ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8+ T cells, central memory CD8+ T cells, effector memory CD8+ T cells and bulk CD8+ T cells or a CD4+ T helper lymphocyte cell that is selected from the group consisting of naïve CD4+ T cells, central memory CD4+ T cells, effector memory CD4+ T cells, and bulk CD4+ T cells

and a second host cell, wherein the second host cell is a precursor T cell. In some alternatives, the precursor T cell is a hematopoietic stem cell.

[0247] In some alternatives, a use of a host cell or a composition in combination with a drug that induces expression of a transgene in the host cell or composition for the treatment of cancer or a viral infection is provided. In some alternatives, the host cell comprises a system. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte, a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain, and d) a polynucleotide coding for an intracellular signaling domain, and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter.. In some alternatives, the drug is tamoxifen and/or its metabolites. In some alternatives, the first promoter comprises a nucleic acid sequence of SEQ ID NO: 41. In some alternatives, the second promoter is a constitutive promoter. In some alternatives, the second promoter is the EF1 α p. In some alternatives, the transcriptional activator comprises a sequence of SEQ ID NO: 40. In some alternatives, the first nucleic acid further comprises a first vector and the second nucleic acid further comprises a second vector. In some alternatives, both vectors are packaged in a viral vector. In some alternatives, the viral vector is a lentivirus. In some alternatives, the first and second nucleic acid comprise a vector. In some alternatives, the first nucleic acid further comprises a nucleic acid sequence coding for a selectable marker. In some alternatives, the second nucleic acid further comprises a nucleic acid coding for a selectable marker. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a cytokine, a chemokine receptor, a polypeptide that regulates apoptosis, or a polypeptide that modulates checkpoint signaling and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the polypeptide that modulates checkpoint signaling inhibits negative checkpoint regulators. In some alternatives, the negative checkpoint regulator comprises VISTA, LAG-3 and/or TIM3. In

some alternatives, the second nucleic acid further comprises a polynucleotide coding for chimeric antigen receptor comprising a ligand binding domain, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment. In some alternatives, the tumor specific molecule is selected from the group consisting of CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, and MAGE A3 TCR or combinations thereof. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a cytokine, a chemokine receptor, a polypeptide that regulates apoptosis, or a polypeptide that modulates checkpoint signaling; and b) a second nucleic acid comprising a second constitutive or inducible promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the polypeptide that modulates checkpoint signaling inhibits negative checkpoint regulators. In some alternatives, the negative checkpoint regulator comprises VISTA, LAG-3 and/or TIM3. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for chimeric antigen receptor comprising a ligand binding domain, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor; a polynucleotide coding for a transmembrane domain. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment. In some alternatives, the ligand binding domain is single chain variable fragment. In some alternatives, the tumor specific molecule is selected from

the group consisting of CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, and MAGE A3 TCR or combinations thereof. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor. In some alternatives, the second promoter is an inducible promoter. In some alternatives, the second promoter is constitutive promoter. In some alternatives, the host cell is a CD8+ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8+ T cells, central memory CD8+ T cells, effector memory CD8+ T cells and bulk CD8+ T cells. In some alternatives, the host cell is a CD4+ T helper lymphocyte cell that is selected from the group consisting of naïve CD4+ T cells, central memory CD4+ T cells, effector memory CD4+ T cells, and bulk CD4+ T cells. In some alternatives, the composition comprises a host cell in a pharmaceutically acceptable excipient. In some alternatives, the host cell comprises a system. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain; and d) a polynucleotide coding for an intracellular signaling domain; and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the second promoter is constitutive or inducible. In some alternatives, the drug is tamoxifen and/or its metabolites. In some alternatives, the first promoter comprises a nucleic acid sequence of SEQ ID NO:41. In some alternatives, the second promoter is a constitutive promoter. In some alternatives, the second promoter is the EF1 α p. In some alternatives, the transcriptional activator comprises a sequence of SEQ ID NO:40. In some alternatives, the first nucleic acid further comprises a first vector and the second nucleic acid further comprises a second vector. In some alternatives, both vectors are packaged in a viral vector. In some alternatives, the viral vector is a lentivirus. In some alternatives, the first and second nucleic acid comprise a vector. In some alternatives, the first nucleic acid further comprises a nucleic acid sequence coding for a selectable marker. In some alternatives, the second nucleic acid further comprises a nucleic acid coding for a

selectable marker. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment. In some alternatives, the tumor specific molecule is selected from the group consisting of CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, and MAGE A3 TCR or combinations thereof. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a cytokine, a chemokine receptor, a polypeptide that regulates apoptosis, or a polypeptide that modulates checkpoint signaling; and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the second promoter is constitutive or inducible. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment. In some alternatives, the ligand binding domain is single chain variable fragment. In some alternatives, the tumor specific molecule is selected from the group consisting of CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, and MAGE A3 TCR or combinations thereof. In some alternatives, the host cell is a precursor T cell. In some alternatives, the precursor T cell is a hematopoietic stem cell. In some alternatives, the isolated T lymphocyte population

comprises precursor T cells. In some alternatives, the precursor T cells are hematopoietic stem cells. In some alternatives, the host cell is a CD8+ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8+ T cells, central memory CD8+ T cells, effector memory CD8+ T cells and bulk CD8+ T cells. In some alternatives, the host cell is a CD4+ T helper lymphocyte cell that is selected from the group consisting of naïve CD4+ T cells, central memory CD4+ T cells, effector memory CD4+ T cells, and bulk CD4+ T cells. In some alternatives, the composition comprises a host cell wherein the host cell is a CD8+ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8+ T cells, central memory CD8+ T cells, effector memory CD8+ T cells and bulk CD8+ T cells and another host cell wherein the host cell is a CD4+ T helper lymphocyte cell that is selected from the group consisting of naïve CD4+ T cells, central memory CD4+ T cells, effector memory CD4+ T cells, and bulk CD4+ T cells. In some alternatives, the cancer is a solid tumor or hematologic malignancy. In some alternatives, the solid tumor is selected from the group consisting of a breast cancer, brain cancer, lung cancer, colon cancer, renal cancer, pancreatic cancer, prostate cancer, and ovarian cancer. In some alternatives, the composition comprises a host cell wherein the host cell is a CD8+ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8+ T cells, central memory CD8+ T cells, effector memory CD8+ T cells and bulk CD8+ T cells or a CD4+ T helper lymphocyte cell that is selected from the group consisting of naïve CD4+ T cells, central memory CD4+ T cells, effector memory CD4+ T cells, and bulk CD4+ T cells and a second host cell, wherein the second host cell is a precursor T cell. In some alternatives, the precursor T cell is a hematopoietic stem cell.

[0248] In some alternatives, a method of performing cellular immunotherapy in a subject having cancer or a viral infection is provided wherein the method comprises administering a composition or a host cell to the subject and administering a drug that induces expression of a transgene in the composition or the host cells. In some alternatives, the host cell comprises a system. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte, a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain, and d) a

polynucleotide coding for an intracellular signaling domain, and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter.. In some alternatives, the drug is tamoxifen and/or its metabolites. In some alternatives, the first promoter comprises a nucleic acid sequence of SEQ ID NO: 41. In some alternatives, the second promoter is a constitutive promoter. In some alternatives, the second promoter is the EF1 α p. In some alternatives, the transcriptional activator comprises a sequence of SEQ ID NO: 40. In some alternatives, the first nucleic acid further comprises a first vector and the second nucleic acid further comprises a second vector. In some alternatives, both vectors are packaged in a viral vector. In some alternatives, the viral vector is a lentivirus. In some alternatives, the first and second nucleic acid comprise a vector. In some alternatives, the first nucleic acid further comprises a nucleic acid sequence coding for a selectable marker. In some alternatives, the second nucleic acid further comprises a nucleic acid coding for a selectable marker. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a cytokine, a chemokine receptor, a polypeptide that regulates apoptosis, or a polypeptide that modulates checkpoint signaling and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the polypeptide that modulates checkpoint signaling inhibits negative checkpoint regulators. In some alternatives, the negative checkpoint regulator comprises VISTA, LAG-3 and/or TIM3. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for chimeric antigen receptor comprising a ligand binding domain, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment. In some alternatives, the tumor specific molecule is selected from the group consisting of CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, and MAGE A3 TCR or combinations thereof. In

some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a cytokine, a chemokine receptor, a polypeptide that regulates apoptosis, or a polypeptide that modulates checkpoint signaling; and b) a second nucleic acid comprising a second constitutive or inducible promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the polypeptide that modulates checkpoint signaling inhibits negative checkpoint regulators. In some alternatives, the negative checkpoint regulator comprises VISTA, LAG-3 and/or TIM3. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for chimeric antigen receptor comprising a ligand binding domain, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor; a polynucleotide coding for a transmembrane domain. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment. In some alternatives, the ligand binding domain is single chain variable fragment. In some alternatives, the tumor specific molecule is selected from the group consisting of CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, and MAGE A3 TCR or combinations thereof. In some alternatives, the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor. In some alternatives, the second promoter is an inducible promoter. In some alternatives, the second promoter is constitutive promoter.. In some alternatives, the host cell is a precursor T cell. In some alternatives, the host cell is a hematopoietic stem cell. In some alternatives, the host cell is a CD8⁺ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8⁺ T cells, central memory CD8⁺ T cells, effector memory CD8⁺ T cells and bulk CD8⁺ T cells. In some alternatives, the host cell is a CD4⁺ T helper lymphocyte cell that is selected from the group consisting of naïve CD4⁺ T cells, central memory CD4⁺ T cells, effector memory CD4⁺ T cells, and bulk CD4⁺ T cells. In some alternatives, the composition comprises a host cell in a pharmaceutically acceptable excipient. In some alternatives, the

host cell comprises a system. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain; and d) a polynucleotide coding for an intracellular signaling domain; and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the second promoter is constitutive or inducible. In some alternatives, the drug is tamoxifen and/or its metabolites. In some alternatives, the first promoter comprises a nucleic acid sequence of SEQ ID NO: 41. In some alternatives, the second promoter is a constitutive promoter. In some alternatives, the second promoter is the EF1 α p. In some alternatives, the transcriptional activator comprises a sequence of SEQ ID NO: 40. In some alternatives, the first nucleic acid further comprises a first vector and the second nucleic acid further comprises a second vector. In some alternatives, both vectors are packaged in a viral vector. In some alternatives, the viral vector is a lentivirus. In some alternatives, the first and second nucleic acid comprise a vector. In some alternatives, the first nucleic acid further comprises a nucleic acid sequence coding for a selectable marker. In some alternatives, the second nucleic acid further comprises a nucleic acid coding for a selectable marker. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment. In some alternatives, the tumor specific molecule is selected from the group consisting of CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, and MAGE A3 TCR or

combinations thereof. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a cytokine, a chemokine receptor, a polypeptide that regulates apoptosis, or a polypeptide that modulates checkpoint signaling; and b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter. In some alternatives, the second promoter is constitutive or inducible. In some alternatives, the polypeptide that modulates checkpoint signaling inhibits negative checkpoint regulators. In some alternatives, the negative checkpoint regulator comprises VISTA, LAG-3 and/or TIM3. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for chimeric antigen receptor comprising a ligand binding domain, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain; and d) a polynucleotide coding for an intracellular signaling domain. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment. In some alternatives, the ligand binding domain is single chain variable fragment. In some alternatives, the tumor specific molecule is selected from the group consisting of CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, and MAGE A3 TCR or combinations thereof. In some alternatives, the host cell is a precursor T cell. In some alternatives, the host cell is a hematopoietic stem cell. In some alternatives, the host cell is a CD8⁺ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8⁺ T cells, central memory CD8⁺ T cells, effector memory CD8⁺ T cells and bulk CD8⁺ T cells. In some alternatives, the host cell is a CD4⁺ T helper lymphocyte cell that is selected from the group consisting of naïve CD4⁺ T cells, central memory CD4⁺ T cells, effector memory CD4⁺ T cells, and bulk CD4⁺ T cells. In some alternatives, the composition comprises a host cell wherein the host cell is a CD8⁺ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8⁺ T cells, central memory CD8⁺ T cells, effector memory CD8⁺ T cells and bulk CD8⁺ T cells and another host cell wherein the host cell is a CD4⁺ T helper lymphocyte cell that is selected from the group consisting of naïve CD4⁺ T cells, central memory CD4⁺ T cells, effector memory CD4⁺ T cells, and bulk CD4⁺ T cells. In some alternatives, the cancer is selected from a solid

tumor or hematologic malignancy. In some alternatives, the solid tumor is selected from the group consisting of a breast cancer, brain cancer, lung cancer, colon cancer, renal cancer, pancreatic cancer, prostate cancer, and ovarian cancer. In some alternatives, the host cell is a precursor T cell. In some alternatives, the precursor T cell is a hematopoietic stem cell. In some alternatives, the isolated T lymphocyte population comprises precursor T cells. In some alternatives, the precursor T cells are hematopoietic stem cells. In some alternatives, the composition comprises a host cell wherein the host cell is a CD8⁺ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8⁺ T cells, central memory CD8⁺ T cells, effector memory CD8⁺ T cells and bulk CD8⁺ T cells or a CD4⁺ T helper lymphocyte cell that is selected from the group consisting of naïve CD4⁺ T cells, central memory CD4⁺ T cells, effector memory CD4⁺ T cells, and bulk CD4⁺ T cells and a second host cell, wherein the second host cell is a precursor T cell. In some alternatives, the precursor T cell is a hematopoietic stem cell.

More Alternatives.

[0249] In some alternatives, a system for inducible expression of a chimeric antigen receptor, the system is provided, wherein the system comprises a) a first nucleic acid comprising a first promoter, which is an inducible promoter, operably linked to a polynucleotide coding for a chimeric antigen receptor and b) a second nucleic acid comprising a second promoter operably linked to a polynucleotide coding for a transcriptional activator, which is capable of activating transcription from the first promoter in the presence of a drug or metabolite thereof. In some alternatives, a system for inducible expression is provided, wherein the system comprises a) an inducible promoter, b) a polynucleotide coding for a chimeric antigen receptor and c) a polynucleotide coding for a transcriptional activator, which transcriptional activator is capable of activating transcription from the inducible promoter in the presence of a drug or metabolite thereof. In some alternatives, the polynucleotide coding for the chimeric receptor is operably linked to the inducible promoter. In some alternatives, the system further comprises further comprising a polynucleotide encoding a recombinant protein, which polynucleotide is operably linked to the inducible promoter. In some alternatives, the drug or metabolite thereof comprises: (i) a drug tolerated when administered to a human subject daily or weekly, or a metabolite thereof; (ii) a molecule that specifically binds to a human receptor, optionally the estrogen receptor, or a metabolite thereof; and/or (iii) tamoxifen and/or a metabolite or analog of tamoxifen. In

some alternatives, the transcriptional activator comprises: (a) a DNA-binding domain; (b) a ligand-binding domain that specifically binds to the drug or metabolite thereof; and (c) a transactivation domain, optionally linked and/or fused in that order. In some alternatives of the system, (a) the DNA-binding domain comprises DNA binding sites not present in a protein naturally expressed in a lymphocyte or not present in a protein naturally expressed in a T cell; and/or (b) the drug or metabolite is the molecule that specifically binds to the human receptor, optionally estrogen receptor, or metabolite thereof, and the binding between the drug or metabolite and the ligand-binding domain is selective for the ligand-binding domain over the human receptor, whereby binding by the ligand-binding domain to the drug or metabolite is greater, optionally at least 1.5, 2, 3, or 4 times as strong, as the binding by the human receptor; and/or (c) the transactivation domain comprises a p65 transactivation domain or functional variant thereof; and/or (d) the first promoter comprises one or more binding sites for the DNA binding domain. In some alternatives, the first promoter does not comprise another binding site for any human DNA binding domain other than a DNA-binding domain or domains present in the transcriptional activator; and/or wherein the first promoter is a synthetic chimeric promoter and/or the transcriptional activator is a synthetic chimeric transcriptional activator; and/or wherein the DNA binding domain comprises a DNA binding domain present in a hepatocyte nuclear factor, which is optionally HNF1-alpha or HNF1-beta. In some alternatives of the system, the first promoter comprises the nucleic acid sequence of SEQ ID NO: 41. In some alternatives, the second promoter is a constitutive promoter. In some alternatives, the second promoter is or comprises an EFl α promoter or functional portion thereof. In some alternatives, the transcriptional activator comprises a polypeptide having the sequence of SEQ ID NO: 40. In some alternatives, the first nucleic acid is comprised within a first vector, which is further comprised by the system and the second nucleic acid is comprised within a second vector, which is further comprised by the system. In some alternatives, the first nucleic acid and the second nucleic acid or the first promoter, polynucleotide encoding the chimeric antigen receptor, second promoter, and polynucleotide encoding the transactivator, are comprised within a vector, which is further comprised by the system. In some alternatives of the system, the system is comprised in a single viral packaging vector. In some alternatives, the viral vector is a lentiviral vector. In some alternatives, the first nucleic acid further comprises a nucleic acid sequence coding for a selectable marker or wherein the system further comprises a selectable marker operably linked to the first promoter. In some alternatives, the second nucleic acid further comprises a

nucleic acid coding for a selectable marker or wherein the system further comprises a selectable marker operably linked to the second promoter.

[0250] In some alternatives, a system for inducible expression is provided, wherein the system comprises a) a first nucleic acid comprising a first promoter, which is an inducible promoter and is operably linked to a polynucleotide coding for a cytokine, a chemokine receptor, a polypeptide that inhibits apoptosis, or a polypeptide that inhibits negative checkpoint signaling and b) a second nucleic acid comprising a second promoter, which is a constitutive or inducible promoter, operably linked to a polynucleotide coding for a transcriptional activator capable of inducing transcription from the first promoter in the presence of a drug or metabolite or analog thereof. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for a recombinant antigen receptor, which optionally is a chimeric antigen receptor. In some alternatives, the chimeric antigen receptor comprises a) a ligand binding domain, which binds to a ligand that is optionally a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population that is suitable to mediate recognition and elimination by a lymphocyte b) a polypeptide spacer, wherein the spacer optionally provides for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor, c) a transmembrane domain and d) an intracellular signaling domain. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment. In some alternatives, the ligand binding domain is single chain variable fragment. In some alternatives, the tumor specific molecule is selected from the group consisting of CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, MAGE A3 TCR and combinations thereof.

[0251] In some alternatives, a chimeric receptor polypeptide encoded by the system is provided. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter, which is an inducible promoter, operably linked to a polynucleotide coding for a chimeric antigen receptor and b) a second nucleic acid comprising a second promoter operably linked to a polynucleotide coding for a transcriptional activator, which is capable of activating transcription from the first promoter in the presence of a drug or metabolite thereof. In some alternatives, a system for inducible expression is provided, wherein the system comprises a) an inducible promoter, b) a polynucleotide coding for a chimeric antigen receptor and c) a polynucleotide coding for a transcriptional activator,

which transcriptional activator is capable of activating transcription from the inducible promoter in the presence of a drug or metabolite thereof. In some alternatives, the polynucleotide coding for the chimeric receptor is operably linked to the inducible promoter. In some alternatives, the system further comprises further comprising a polynucleotide encoding a recombinant protein, which polynucleotide is operably linked to the inducible promoter. In some alternatives, the drug or metabolite thereof comprises: (i) a drug tolerated when administered to a human subject daily or weekly, or a metabolite thereof; (ii) a molecule that specifically binds to a human receptor, optionally the estrogen receptor, or a metabolite thereof; and/or (iii) tamoxifen and/or a metabolite or analog of tamoxifen. In some alternatives, the transcriptional activator comprises: (a) a DNA-binding domain; (b) a ligand-binding domain that specifically binds to the drug or metabolite thereof; and (c) a transactivation domain, optionally linked and/or fused in that order. In some alternatives of the system, (a) the DNA-binding domain comprises DNA binding sites not present in a protein naturally expressed in a lymphocyte or not present in a protein naturally expressed in a T cell; and/or (b) the drug or metabolite is the molecule that specifically binds to the human receptor, optionally estrogen receptor, or metabolite thereof, and the binding between the drug or metabolite and the ligand-binding domain is selective for the ligand-binding domain over the human receptor, whereby binding by the ligand-binding domain to the drug or metabolite is greater, optionally at least 1.5, 2, 3, or 4 times as strong, as the binding by the human receptor; and/or (c) the transactivation domain comprises a p65 transactivation domain or functional variant thereof; and/or (d) the first promoter comprises one or more binding sites for the DNA binding domain. In some alternatives, the first promoter does not comprise another binding site for any human DNA binding domain other than a DNA-binding domain or domains present in the transcriptional activator; and/or wherein the first promoter is a synthetic chimeric promoter and/or the transcriptional activator is a synthetic chimeric transcriptional activator; and/or wherein the DNA binding domain comprises a DNA binding domain present in a hepatocyte nuclear factor, which is optionally HNF1-alpha or HNF1-beta. The system of any of claims 1-8, wherein the first promoter comprises the nucleic acid sequence of SEQ ID NO: 41. In some alternatives, the second promoter is a constitutive promoter. In some alternatives, the second promoter is or comprises an EF1a promoter or functional portion thereof. In some alternatives, the transcriptional activator comprises a polypeptide having the sequence of SEQ ID NO: 40. In some alternatives, the first nucleic acid is comprised within a first vector, which is further comprised by the system and the

second nucleic acid is comprised within a second vector, which is further comprised by the system. In some alternatives, the first nucleic acid and the second nucleic acid or the first promoter, polynucleotide encoding the chimeric antigen receptor, second promoter, and polynucleotide encoding the transactivator, are comprised within a vector, which is further comprised by the system. In some alternatives of the system, the system is comprised in a single viral packaging vector. In some alternatives, the viral vector is a lentiviral vector. In some alternatives, the first nucleic acid further comprises a nucleic acid sequence coding for a selectable marker or wherein the system further comprises a selectable marker operably linked to the first promoter. In some alternatives, the second nucleic acid further comprises a nucleic acid coding for a selectable marker or wherein the system further comprises a selectable marker operably linked to the second promoter. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter, which is an inducible promoter and is operably linked to a polynucleotide coding for a cytokine, a chemokine receptor, a polypeptide that inhibits apoptosis, or a polypeptide that inhibits negative checkpoint signaling and b) a second nucleic acid comprising a second promoter, which is a constitutive or inducible promoter, operably linked to a polynucleotide coding for a transcriptional activator capable of inducing transcription from the first promoter in the presence of a drug or metabolite or analog thereof. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for a recombinant antigen receptor, which optionally is a chimeric antigen receptor. In some alternatives, the chimeric antigen receptor comprises a a) ligand binding domain, which binds to a ligand that is optionally a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population that is suitable to mediate recognition and elimination by a lymphocyte b) a polypeptide spacer, wherein the spacer optionally provides for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor, c) a transmembrane domain and d) an intracellular signaling domain. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment. In some alternatives, the ligand binding domain is single chain variable fragment. In some alternatives, the tumor specific molecule is selected from the group consisting of CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, and MAGE A3 TCR and combinations thereof.

[0252] In some alternatives, a host cell comprising a system or viral packaging vector is provided. In some alternatives, the system comprises a) a first nucleic acid

comprising a first promoter, which is an inducible promoter, operably linked to a polynucleotide coding for a chimeric antigen receptor and b) a second nucleic acid comprising a second promoter operably linked to a polynucleotide coding for a transcriptional activator, which is capable of activating transcription from the first promoter in the presence of a drug or metabolite thereof. In some alternatives, a system for inducible expression is provided, wherein the system comprises a) an inducible promoter, b) a polynucleotide coding for a chimeric antigen receptor and c) a polynucleotide coding for a transcriptional activator, which transcriptional activator is capable of activating transcription from the inducible promoter in the presence of a drug or metabolite thereof. In some alternatives, the polynucleotide coding for the chimeric receptor is operably linked to the inducible promoter. In some alternatives, the system further comprises further comprising a polynucleotide encoding a recombinant protein, which polynucleotide is operably linked to the inducible promoter. In some alternatives, the drug or metabolite thereof comprises: (i) a drug tolerated when administered to a human subject daily or weekly, or a metabolite thereof; (ii) a molecule that specifically binds to a human receptor, optionally the estrogen receptor, or a metabolite thereof; and/or (iii) tamoxifen and/or a metabolite or analog of tamoxifen. In some alternatives, the transcriptional activator comprises: (a) a DNA-binding domain; (b) a ligand-binding domain that specifically binds to the drug or metabolite thereof; and (c) a transactivation domain, optionally linked and/or fused in that order. In some alternatives of the system, (a) the DNA-binding domain comprises DNA binding sites not present in a protein naturally expressed in a lymphocyte or not present in a protein naturally expressed in a T cell; and/or (b) the drug or metabolite is the molecule that specifically binds to the human receptor, optionally estrogen receptor, or metabolite thereof, and the binding between the drug or metabolite and the ligand-binding domain is selective for the ligand-binding domain over the human receptor, whereby binding by the ligand-binding domain to the drug or metabolite is greater, optionally at least 1.5, 2, 3, or 4 times as strong, as the binding by the human receptor; and/or (c) the transactivation domain comprises a p65 transactivation domain or functional variant thereof; and/or (d) the first promoter comprises one or more binding sites for the DNA binding domain. In some alternatives, the first promoter does not comprise another binding site for any human DNA binding domain other than a DNA-binding domain or domains present in the transcriptional activator; and/or wherein the first promoter is a synthetic chimeric promoter and/or the transcriptional activator is a synthetic chimeric transcriptional activator; and/or wherein the DNA binding domain comprises a DNA binding

domain present in a hepatocyte nuclear factor, which is optionally HNF1-alpha or HNF1-beta. In some alternatives, the first promoter comprises the nucleic acid sequence of SEQ ID NO: 41. In some alternatives, the second promoter is a constitutive promoter. In some alternatives, the second promoter is or comprises an EF1 α promoter or functional portion thereof. In some alternatives, the transcriptional activator comprises a polypeptide having the sequence of SEQ ID NO: 40. In some alternatives, the first nucleic acid is comprised within a first vector, which is further comprised by the system and the second nucleic acid is comprised within a second vector, which is further comprised by the system. In some alternatives, the first nucleic acid and the second nucleic acid or the first promoter, polynucleotide encoding the chimeric antigen receptor, second promoter, and polynucleotide encoding the transactivator, are comprised within a vector, which is further comprised by the system. In some alternatives, the system described above is comprised in a single viral packaging vector. In some such alternatives, viral vector is a lentiviral vector. In some alternatives, the first nucleic acid further comprises a nucleic acid sequence coding for a selectable marker or wherein the system further comprises a selectable marker operably linked to the first promoter. In some alternatives, the second nucleic acid further comprises a nucleic acid coding for a selectable marker or wherein the system further comprises a selectable marker operably linked to the second promoter. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter, which is an inducible promoter and is operably linked to a polynucleotide coding for a cytokine, a chemokine receptor, a polypeptide that inhibits apoptosis, or a polypeptide that inhibits negative checkpoint signaling and b) a second nucleic acid comprising a second promoter, which is a constitutive or inducible promoter, operably linked to a polynucleotide coding for a transcriptional activator capable of inducing transcription from the first promoter in the presence of a drug or metabolite or analog thereof. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for a recombinant antigen receptor, which optionally is a chimeric antigen receptor. In some alternatives, the chimeric antigen receptor comprises a a) ligand binding domain, which binds to a ligand that is optionally a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population that is suitable to mediate recognition and elimination by a lymphocyte b) a polypeptide spacer, wherein the spacer optionally provides for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor, c) a transmembrane domain and d) an intracellular signaling domain. In some alternatives, the first promoter is in opposite orientation to the

second promoter. In some alternatives, the ligand binding domain is an antibody fragment. In some alternatives, the ligand binding domain is single chain variable fragment. In some alternatives, the tumor specific molecule is selected from the group consisting of CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, and MAGE A3 TCR and combinations thereof.

[0253] In some alternatives, a host cell is provided, wherein the host cell is a primary human lymphocyte, wherein the host cell comprises a) a nucleic acid comprising a first promoter, which is an inducible synthetic promoter containing a binding site for a DNA binding domain not naturally present in the primary human lymphocyte and b) a polynucleotide coding for a transcriptional activator, the transcriptional activator comprising i) the DNA binding domain, wherein the DNA binding domain does not specifically bind to a DNA sequence naturally present in the primary human lymphocyte; ii) a domain that specifically binds to a drug or metabolite thereof and does not bind or does not bind with as great a degree of affinity to any molecule naturally present in the primary human lymphocyte and iii) a transactivation domain, wherein the transcriptional activator is capable of inducing transcription from the first promoter in the presence of the drug or metabolite thereof and/or upon binding of the drug or metabolite thereof to the domain in (ii). In some alternatives, the cell further comprises a polynucleotide encoding a chimeric antigen receptor, which optionally is operably linked to the first promoter. In some alternatives, In some alternatives, the cell further comprises a second promoter, which is operably linked to the polynucleotide encoding the transcriptional activator, wherein the second promoter is optionally constitutive. In some alternatives, the host cell is a CD8⁺ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8⁺ T cells, central memory CD8⁺ T cells, effector memory CD8⁺ T cells and bulk CD8⁺ T cell. In some alternatives, the host cell is a CD4⁺ T helper lymphocyte cell that is selected from the group consisting of naïve CD4⁺ T cells, central memory CD4⁺ T cells, effector memory CD4⁺ T cells, and bulk CD4⁺ T cells.

[0254] In some alternatives, a composition is provided, wherein the composition comprises a host cell in a pharmaceutically acceptable excipient. In some alternatives, the host cell the host cell is a primary human lymphocyte, wherein the host cell comprises a) a nucleic acid comprising a first promoter, which is an inducible synthetic promoter containing a binding site for a DNA binding domain not naturally present in the primary human lymphocyte and b) a polynucleotide coding for a transcriptional activator, the transcriptional activator comprising i) the DNA binding domain, wherein the DNA binding domain does not

specifically bind to a DNA sequence naturally present in the primary human lymphocyte; ii) a domain that specifically binds to a drug or metabolite thereof and does not bind or does not bind with as great a degree of affinity to any molecule naturally present in the primary human lymphocyte and iii) a transactivation domain, wherein the transcriptional activator is capable of inducing transcription from the first promoter in the presence of the drug or metabolite thereof and/or upon binding of the drug or metabolite thereof to the domain in (ii). In some alternatives, the cell further comprises a polynucleotide encoding a chimeric antigen receptor, which optionally is operably linked to the first promoter. In some alternatives, In some alternatives, the cell further comprises a second promoter, which is operably linked to the polynucleotide encoding the transcriptional activator, wherein the second promoter is optionally constitutive. In some alternatives, the host cell is a CD8+ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8+ T cells, central memory CD8+ T cells, effector memory CD8+ T cells and bulk CD8+ T cell. In some alternatives, the host cell is a CD4+ T helper lymphocyte cell that is selected from the group consisting of naïve CD4+ T cells, central memory CD4+ T cells, effector memory CD4+ T cells, and bulk CD4+ T cells. In some alternatives, the composition comprises a CD8+ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8+ T cells, central memory CD8+ T cells, effector memory CD8+ T cells and bulk CD8+ T cell and a CD4+ T helper lymphocyte cell that is selected from the group consisting of naïve CD4+ T cells, central memory CD4+ T cells, effector memory CD4+ T cells, and bulk CD4+ T cells.

[0255] In some alternatives, an *in vitro* method for preparing a host cell is provided and comprises introducing a system into a separate isolated T lymphocyte population and expanding each T lymphocyte population *in vitro*. In some alternatives, the host cell the host cell is a primary human lymphocyte, wherein the host cell comprises a) a nucleic acid comprising a first promoter, which is an inducible synthetic promoter containing a binding site for a DNA binding domain not naturally present in the primary human lymphocyte and b) a polynucleotide coding for a transcriptional activator, the transcriptional activator comprising i) the DNA binding domain, wherein the DNA binding domain does not specifically bind to a DNA sequence naturally present in the primary human lymphocyte; ii) a domain that specifically binds to a drug or metabolite thereof and does not bind or does not bind with as great a degree of affinity to any molecule naturally present in the primary human lymphocyte and iii) a transactivation domain, wherein the transcriptional activator is capable of inducing transcription from the first promoter in the presence of the drug or metabolite

thereof and/or upon binding of the drug or metabolite thereof to the domain in (ii). In some alternatives, the cell further comprises a polynucleotide encoding a chimeric antigen receptor, which optionally is operably linked to the first promoter. In some alternatives, the cell further comprises a second promoter, which is operably linked to the polynucleotide encoding the transcriptional activator, wherein the second promoter is optionally constitutive. In some alternatives, the host cell is a CD8+ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8+ T cells, central memory CD8+ T cells, effector memory CD8+ T cells and bulk CD8+ T cell. In some alternatives, the host cell is a CD4+ T helper lymphocyte cell that is selected from the group consisting of naïve CD4+ T cells, central memory CD4+ T cells, effector memory CD4+ T cells, and bulk CD4+ T cells. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter, which is an inducible promoter, operably linked to a polynucleotide coding for a chimeric antigen receptor and b) a second nucleic acid comprising a second promoter operably linked to a polynucleotide coding for a transcriptional activator, which is capable of activating transcription from the first promoter in the presence of a drug or metabolite thereof. In some alternatives, a system for inducible expression is provided, wherein the system comprises a) an inducible promoter, b) a polynucleotide coding for a chimeric antigen receptor and c) a polynucleotide coding for a transcriptional activator, which transcriptional activator is capable of activating transcription from the inducible promoter in the presence of a drug or metabolite thereof. In some alternatives, the polynucleotide coding for the chimeric receptor is operably linked to the inducible promoter. In some alternatives, the system further comprises further comprising a polynucleotide encoding a recombinant protein, which polynucleotide is operably linked to the inducible promoter. In some alternatives, the drug or metabolite thereof comprises: (i) a drug tolerated when administered to a human subject daily or weekly, or a metabolite thereof; (ii) a molecule that specifically binds to a human receptor, optionally the estrogen receptor, or a metabolite thereof; and/or (iii) tamoxifen and/or a metabolite or analog of tamoxifen. In some alternatives, the transcriptional activator comprises: (a) a DNA-binding domain; (b) a ligand-binding domain that specifically binds to the drug or metabolite thereof; and (c) a transactivation domain, optionally linked and/or fused in that order. In some alternatives of the system, (a) the DNA-binding domain comprises DNA binding sites not present in a protein naturally expressed in a lymphocyte or not present in a protein naturally expressed in a T cell; and/or (b) the drug or metabolite is the molecule that specifically binds to the human receptor, optionally estrogen receptor, or

metabolite thereof, and the binding between the drug or metabolite and the ligand-binding domain is selective for the ligand-binding domain over the human receptor, whereby binding by the ligand-binding domain to the drug or metabolite is greater, optionally at least 1.5, 2, 3, or 4 times as strong, as the binding by the human receptor; and/or (c) the transactivation domain comprises a p65 transactivation domain or functional variant thereof; and/or (d) the first promoter comprises one or more binding sites for the DNA binding domain. In some alternatives, the first promoter does not comprise another binding site for any human DNA binding domain other than a DNA-binding domain or domains present in the transcriptional activator; and/or wherein the first promoter is a synthetic chimeric promoter and/or the transcriptional activator is a synthetic chimeric transcriptional activator; and/or wherein the DNA binding domain comprises a DNA binding domain present in a hepatocyte nuclear factor, which is optionally HNF1-alpha or HNF1-beta. In some alternatives, the first promoter comprises the nucleic acid sequence of SEQ ID NO: 41. In some alternatives, the second promoter is a constitutive promoter. In some alternatives, the second promoter is or comprises an EF1 α promoter or functional portion thereof. In some alternatives, the transcriptional activator comprises a polypeptide having the sequence of SEQ ID NO: 40. In some alternatives, the first nucleic acid is comprised within a first vector, which is further comprised by the system and the second nucleic acid is comprised within a second vector, which is further comprised by the system. In some alternatives, the first nucleic acid and the second nucleic acid or the first promoter, polynucleotide encoding the chimeric antigen receptor, second promoter, and polynucleotide encoding the transactivator, are comprised within a vector, which is further comprised by the system. In some alternatives, the system is comprised in a single viral packaging vector. In some alternatives, the viral vector is a lentiviral vector. In some alternatives, the first nucleic acid further comprises a nucleic acid sequence coding for a selectable marker or wherein the system further comprises a selectable marker operably linked to the first promoter. In some alternatives, the second nucleic acid further comprises a nucleic acid coding for a selectable marker or wherein the system further comprises a selectable marker operably linked to the second promoter. In some alternatives, the system comprises a) a first nucleic acid comprising a first promoter, which is an inducible promoter and is operably linked to a polynucleotide coding for a cytokine, a chemokine receptor, a polypeptide that inhibits apoptosis, or a polypeptide that inhibits negative checkpoint signaling and b) a second nucleic acid comprising a second promoter, which is a constitutive or inducible promoter, operably linked to a polynucleotide coding for a

transcriptional activator capable of inducing transcription from the first promoter in the presence of a drug or metabolite or analog thereof. In some alternatives, the second nucleic acid further comprises a polynucleotide coding for a recombinant antigen receptor, which optionally is a chimeric antigen receptor. In some alternatives, the chimeric antigen receptor comprises a a) ligand binding domain, which binds to a ligand that is optionally a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population that is suitable to mediate recognition and elimination by a lymphocyte b) a polypeptide spacer, wherein the spacer optionally provides for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor, c) a transmembrane domain and d) an intracellular signaling domain. In some alternatives, the first promoter is in opposite orientation to the second promoter. In some alternatives, the ligand binding domain is an antibody fragment. In some alternatives, the ligand binding domain is single chain variable fragment. In some alternatives, the tumor specific molecule is selected from the group consisting of CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, MAGE A3 TCR and combinations thereof. In some alternatives, the T lymphocytes in the population are expanded and wherein the method further comprises culturing the cells in the presence of anti-CD3 and/or anti CD28, and at least one homeostatic cytokine until the cells expand sufficiently for use as a cell infusion. In some alternatives, culturing in the presence of anti-CD3 and/or anti CD28, and at least one homeostatic cytokine can be performed before or after the introduction of the system.

[0256] In some alternatives, a use of the host cell or a composition in combination with the drug or a metabolite thereof for the treatment of cancer or a viral infection is provided. In some alternatives, the host cell is a primary human lymphocyte, wherein the host cell comprises a) a nucleic acid comprising a first promoter, which is an inducible synthetic promoter containing a binding site for a DNA binding domain not naturally present in the primary human lymphocyte and b) a polynucleotide coding for a transcriptional activator, the transcriptional activator comprising i) the DNA binding domain, wherein the DNA binding domain does not specifically bind to a DNA sequence naturally present in the primary human lymphocyte; ii) a domain that specifically binds to a drug or metabolite thereof and does not bind or does not bind with as great a degree of affinity to any molecule naturally present in the primary human lymphocyte and iii) a transactivation domain, wherein the transcriptional activator is capable of inducing transcription from the first promoter in the presence of the

drug or metabolite thereof and/or upon binding of the drug or metabolite thereof to the domain in (ii). In some alternatives, the cell further comprises a polynucleotide encoding a chimeric antigen receptor, which optionally is operably linked to the first promoter. In some alternatives, In some alternatives, the cell further comprises a second promoter, which is operably linked to the polynucleotide encoding the transcriptional activator, wherein the second promoter is optionally constitutive. In some alternatives, the host cell is a CD8+ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8+ T cells, central memory CD8+ T cells, effector memory CD8+ T cells and bulk CD8+ T cell. In some alternatives, the host cell is a CD4+ T helper lymphocyte cell that is selected from the group consisting of naïve CD4+ T cells, central memory CD4+ T cells, effector memory CD4+ T cells, and bulk CD4+ T cells. In some alternatives, the composition comprises a host cell in a pharmaceutically acceptable excipient. In some alternatives, the host cell the host cell is a primary human lymphocyte, wherein the host cell comprises a) a nucleic acid comprising a first promoter, which is an inducible synthetic promoter containing a binding site for a DNA binding domain not naturally present in the primary human lymphocyte and b) a polynucleotide coding for a transcriptional activator, the transcriptional activator comprising i) the DNA binding domain, wherein the DNA binding domain does not specifically bind to a DNA sequence naturally present in the primary human lymphocyte; ii) a domain that specifically binds to a drug or metabolite thereof and does not bind or does not bind with as great a degree of affinity to any molecule naturally present in the primary human lymphocyte and iii) a transactivation domain, wherein the transcriptional activator is capable of inducing transcription from the first promoter in the presence of the drug or metabolite thereof and/or upon binding of the drug or metabolite thereof to the domain in (ii). In some alternatives, the cell further comprises a polynucleotide encoding a chimeric antigen receptor, which optionally is operably linked to the first promoter. In some alternatives, the cell further comprises a second promoter, which is operably linked to the polynucleotide encoding the transcriptional activator, wherein the second promoter is optionally constitutive. In some alternatives, the host cell is a CD8+ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8+ T cells, central memory CD8+ T cells, effector memory CD8+ T cells and bulk CD8+ T cell. In some alternatives, the host cell is a CD4+ T helper lymphocyte cell that is selected from the group consisting of naïve CD4+ T cells, central memory CD4+ T cells, effector memory CD4+ T cells, and bulk CD4+ T cells. In some alternatives, the composition comprises a CD8+ T cytotoxic lymphocyte cell selected from the group

consisting of naïve CD8+ T cells, central memory CD8+ T cells, effector memory CD8+ T cells and bulk CD8+ T cell and a CD4+ T helper lymphocyte cell that is selected from the group consisting of naïve CD4+ T cells, central memory CD4+ T cells, effector memory CD4+ T cells, and bulk CD4+ T cells.

[0257] In some alternatives, a cell and a drug for use in treatment or inhibition of cancer or a viral infection is provided, wherein the cell comprises: (a) a polynucleotide encoding a chimeric antigen receptor that specifically binds to an antigen associated with the cancer or the viral infection, (b) an inducible synthetic promoter, and (c) a transcriptional activator containing a DNA binding domain that specifically binds to the synthetic promoter and a domain that specifically binds to the drug or a metabolite thereof and is capable of inducing transcription from the synthetic promoter in the presence of the drug or a metabolite thereof. In some alternatives, the cancer is a solid tumor or hematologic malignancy. In some alternatives, the solid tumor is selected from the group consisting of a breast cancer, lung cancer, colon cancer, renal cancer, pancreatic cancer, prostate cancer, and ovarian cancer.

[0258] In some alternatives, a method of performing cellular immunotherapy in a subject having cancer or a viral infection is provided wherein the method comprises administering a composition or a host cell to the subject and administering the drug or metabolite thereof, thereby inducing expression from the promoter. In some alternatives, the host cell is a primary human lymphocyte, wherein the host cell comprises a) a nucleic acid comprising a first promoter, which is an inducible synthetic promoter containing a binding site for a DNA binding domain not naturally present in the primary human lymphocyte and b) a polynucleotide coding for a transcriptional activator, the transcriptional activator comprising i) the DNA binding domain, wherein the DNA binding domain does not specifically bind to a DNA sequence naturally present in the primary human lymphocyte; ii) a domain that specifically binds to a drug or metabolite thereof and does not bind or does not bind with as great a degree of affinity to any molecule naturally present in the primary human lymphocyte and iii) a transactivation domain, wherein the transcriptional activator is capable of inducing transcription from the first promoter in the presence of the drug or metabolite thereof and/or upon binding of the drug or metabolite thereof to the domain in (ii). In some alternatives, the cell further comprises a polynucleotide encoding a chimeric antigen receptor, which optionally is operably linked to the first promoter. In some alternatives, In some alternatives, the cell further comprises a second promoter, which is operably linked to the polynucleotide encoding the transcriptional activator, wherein the second promoter is

optionally constitutive. In some alternatives, the host cell is a CD8⁺ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8⁺ T cells, central memory CD8⁺ T cells, effector memory CD8⁺ T cells and bulk CD8⁺ T cell. In some alternatives, the host cell is a CD4⁺ T helper lymphocyte cell that is selected from the group consisting of naïve CD4⁺ T cells, central memory CD4⁺ T cells, effector memory CD4⁺ T cells, and bulk CD4⁺ T cells. In some alternatives, the composition comprises a host cell in a pharmaceutically acceptable excipient. In some alternatives, the host cell the host cell is a primary human lymphocyte, wherein the host cell comprises a) a nucleic acid comprising a first promoter, which is an inducible synthetic promoter containing a binding site for a DNA binding domain not naturally present in the primary human lymphocyte and b) a polynucleotide coding for a transcriptional activator, the transcriptional activator comprising i) the DNA binding domain, wherein the DNA binding domain does not specifically bind to a DNA sequence naturally present in the primary human lymphocyte; ii) a domain that specifically binds to a drug or metabolite thereof and does not bind or does not bind with as great a degree of affinity to any molecule naturally present in the primary human lymphocyte and iii) a transactivation domain, wherein the transcriptional activator is capable of inducing transcription from the first promoter in the presence of the drug or metabolite thereof and/or upon binding of the drug or metabolite thereof to the domain in (ii). In some alternatives, the cell further comprises a polynucleotide encoding a chimeric antigen receptor, which optionally is operably linked to the first promoter. In some alternatives, the cell further comprises a second promoter, which is operably linked to the polynucleotide encoding the transcriptional activator, wherein the second promoter is optionally constitutive. In some alternatives, the host cell is a CD8⁺ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8⁺ T cells, central memory CD8⁺ T cells, effector memory CD8⁺ T cells and bulk CD8⁺ T cell. In some alternatives, the host cell is a CD4⁺ T helper lymphocyte cell that is selected from the group consisting of naïve CD4⁺ T cells, central memory CD4⁺ T cells, effector memory CD4⁺ T cells, and bulk CD4⁺ T cells. In some alternatives, the composition comprises a CD8⁺ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8⁺ T cells, central memory CD8⁺ T cells, effector memory CD8⁺ T cells and bulk CD8⁺ T cell and a CD4⁺ T helper lymphocyte cell that is selected from the group consisting of naïve CD4⁺ T cells, central memory CD4⁺ T cells, effector memory CD4⁺ T cells, and bulk CD4⁺ T cells. In some alternatives, the cancer is selected from a solid tumor or hematologic malignancy. In some alternatives, the solid tumor is selected from the group consisting of a

breast cancer, lung cancer, colon cancer, renal cancer, pancreatic cancer, prostate cancer, and ovarian cancer.

[0259] Another aspect of the disclosure includes a genetic system to deliver drug-regulated transgene expression in cells, such as drug-regulated expression of a recombinant protein such as a recombinant antigen receptor and/or a molecule expressed by a cell expressing a recombinant antigen receptor. In an alternative, regulated transgene expression is engineered into and/or contained within cells, such as lymphocytes, for example, for use in adoptive cell therapy, such as adoptive immunotherapy. Such systems provide rigorous safety attributes to cell therapies, such as chimeric antigen receptor (CAR) adoptive therapeutic strategies, generally without sacrificing curative intent. In some aspects, such features permit real-time clinician control of recombinant protein expression, e.g., CAR expression, *in vivo*. By engineering vectors that enable drug responsive transcriptional control of recombinant gene, e.g., CAR, expression, the activity of the recombinant gene, e.g., CARs, and/or other cell mediators can be turned “ON” and “OFF” *in vivo*, for example, based on a clinician prescribed pharmaceutical drug input that exhibits clinically permissive pharmacokinetics, tissue distribution, and partitioning between the extracellular space and cytosol of lymphocytes. The genetic system provides for drug regulated transgene expression to enforce a functional “OFF” state in the absence of the drug and a functional “ON” state transgene expression in the presence of the drug.

[0260] One alternative of such a drug is tamoxifen. Tamoxifen is an estrogen antagonist/partial agonist that is an FDA-approved and commercially available drug. It is taken orally and can be administered on a daily basis over an extended period of time. Tamoxifen has a proven safety record, favorable pharmacokinetic profile, excellent tissue distribution and a low partition coefficient between the extracellular space and cytosol. Functional analogs of tamoxifen also may be used. Other drugs can be selected, for example, based on safety record, favorable pharmacokinetic profile, excellent tissue distribution, a low partition coefficient between the extracellular space and cytosol, and/or low toxicities.

[0261] In some alternatives, the system employs a synthetic transcriptional regulator, e.g., synthetic transcriptional activator, which, in the presence of tamoxifen, can be induced to bind to a synthetic promoter operably linked to, e.g., upstream of, a transgene, to induce expression from the promoter, e.g., of the transgene. In some alternatives provided herein, the transcriptional activator is regulated by a drug or metabolite thereof, such as a tamoxifen-regulated transcriptional activator or transcription factor, which may be regulated,

e.g., induced, by tamoxifen or an analog or metabolite thereof. Exemplary tamoxifen-regulated transcription factors are chimeric transcription factors, such as those comprising a DNA binding domain specific for a synthetic promoter of the system, a domain that specifically binds tamoxifen and/or metabolite(s) thereof, for example, with affinity higher than the affinity of the domain for a natural molecule such as estrogen, and a transactivating domain, such as a strong transactivating domain. One tamoxifen regulated transcription factor ("TamR-tf", also designated "HEA3") is a chimeric transcription factor composed of human subunits including the N-terminal DNA binding domain of Hepatocyte Nuclear Factor 1-alpha (HNF-1 α) fused in frame to a mutant (G521R) tamoxifen-specific form of an estrogen receptor ligand binding domain (ER-LBD), which binds to tamoxifen metabolites with high affinity as compared to estrogen, which is in turn fused to the p65 activation domain of NF- κ B (p65). An exemplary amino acid sequence of a TamR-tf is provided in Table 10 and is identified as SEQ ID NO: 40. In this sequence, the mutant tamoxifen-specific form of the ligand binding domain of the estrogen receptor ligand binding domain (ER-LBD) is found at amino acids 282-595 of the TamR-tf and has a mutation at position 521, as compared to wild-type estrogen receptor ligand binding domain. The p65 activation domain of NF- κ B (p65) is found at amino acids 596-862 of SEQ ID NO: 40. Changes can be made to the transcriptional activator, for example, to increase the properties of the transcription factor including, without limitation, altering one or more amino acids in the estrogen receptor ligand binding domain to increase the affinity of the factor for estrogen analogs and altering one or more amino acids in the p65 transactivating domain. In some alternatives, changes are made to the transcriptional activator that result in an altered transcription factor which retains or substantially retains one or more functions of tamR-tf, such as tamoxifen-specific binding and/or the same or substantially the same or at least the same specificity for tamoxifen or metabolite as compared to any natural molecule, at least the same or about the same specific DNA binding function, and/or the same or at least the same degree of transactivation activity.

[0262] In the absence of tamoxifen, the transcriptional activator, e.g., TamR-tf, is generally excluded from the nucleus by binding of cytosolic heat-shock protein 90 (HSP90) to the tamoxifen-binding active site, resulting in the expression of a transgene operably linked to the tamR-tf-inducible promoter being in the "OFF" state. Nanomolar concentrations of cytosolic tamoxifen actively out-compete HSP90 for ER-LBD binding, resulting in TamR-tf translocation to the nucleus. Upon nuclear translocation, TamR-tf is readily available to bind to a restricted synthetic promoter (e.g. 7xHBD/EF1ap). In the presence of tamoxifen, binding

of TamR-tf to the synthetic promoter, e.g., the 7xHBD/EF1 α p promoter, induces the “ON” state of expression for a transgene operably linked to the synthetic promoter. In some alternatives, this transcriptional regulator can be modified to provide for a varying level of control of transgene expression. Amino acid substitutions in the LBD of TamR-tf permit selective responsiveness to tamoxifen and its metabolites, where 4-hydroxy tamoxifen (4-OHT) is the most pharmacologically active metabolite, in regards to TamR-tf activity, while lacking interaction with endogenous estrogen. In some alternatives, a system for inducible expression of chimeric antigen receptor is provided, wherein the system comprises: a first nucleic acid comprising a first promoter inducible by a drug operably linked to a polynucleotide, such as one coding for a chimeric antigen receptor, the chimeric antigen receptor optionally comprising a ligand binding domain, wherein the ligand binding domain binds to a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population that is suitable to mediate recognition and elimination by a lymphocyte, a polypeptide spacer, wherein the spacer optionally provides for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor, a transmembrane domain, and an intracellular signaling domain. In some alternatives, the system further includes a second nucleic acid encoding a transcriptional modulator for the inducible promoter, which is capable of modulating, e.g., activating, transcription from the first promoter, such as in the presence of the drug or metabolite thereof; typically the system comprises a second constitutive or inducible promoter operably linked to a nucleic acid coding for the transcriptional modulator.

[0263] In some alternatives of the inducible system, the first promoter is operably linked to a polynucleotide coding for a gene that promotes cell survival and proliferation, a gene that prevents apoptosis, and/or a gene that inhibits negative checkpoint signaling. Such genes include genes encoding IL-2, IL-15, Chemokine receptors, Bcl2, CA-Akt, dn-TGF β RIII, dn-SHP1/2, and/or PD-1CD28 chimeras.

[0264] In some alternatives, the system employs a synthetic transcriptional activator which, in the presence of the drug (e.g. tamoxifen) or a metabolite thereof, such as following administration of the drug, is induced to bind a synthetic promoter upstream of a transgene to induce expression. In some alternatives, the transcriptional activator is TamR-tf (HEA3) or other tamoxifen-inducible transcriptional activator with analogous domains. The tamoxifen regulated transcription factor (“TamR-tf”, also designated “HEA3”) is a chimeric transcription factor composed of human subunits including the N-terminal DNA binding

domain of Hepatocyte Nuclear Factor 1-alpha (HNF-1 α)(e.g. amino acids 1-281 of SEQ ID NO: 40) fused in frame to the mutant tamoxifen-specific ligand binding domain of the estrogen receptor ligand binding domain (ER-LBD), that is in turn fused to the p65 activation domain of NF- κ B (p65).

[0265] In some alternatives of the compositions herein, the CD4+ T helper lymphocyte cell is naïve CD4+ T cells, central memory CD4+ T cells, effector memory CD4+ T cells, or bulk CD4+ T cells. In some alternatives, CD4+ helper lymphocyte cell is a naïve CD4+ T cell, wherein the naïve CD4+ T cell comprises a CD45RO-, CD45RA+, and/or is a CD62L+ CD4+ T cell. In some alternatives, at least 20, 30, 40, 50, 60, 70, 80, 90, 95, 99, or 100 % of the cells in the composition are CD4+; in some alternatives, at least 20, 30, 40, 50, 60, 70, 80, 90, 95, 99, or 100 % of the cells in the composition or the CD4+ cells in the composition are naïve CD4+ T cells, central memory CD4+ T cells, effector memory CD4+ T cells, or bulk CD4+ T cells.

[0266] In some alternatives of the compositions herein, the CD8+ T cytotoxic lymphocyte cell is a naïve CD8+ T cell, central memory CD8+ T cell, effector memory CD8+ T cell and/or bulk CD8+ T cell. In some alternatives, the CD8+ cytotoxic T lymphocyte cell is a central memory T cell, wherein the central memory T cell comprises a CD45RO+, CD62L+, and/or CD8+ T cell. In yet other alternatives, the CD8+ cytotoxic T lymphocyte cell is a central memory T cell and the CD4+ helper T lymphocyte cell is a naïve or central memory CD4+ T cell. In some alternatives, at least 20, 30, 40, 50, 60, 70, 80, 90, 95, 99, or 100 % of the cells in the composition are CD8+; in some alternatives, at least 20, 30, 40, 50, 60, 70, 80, 90, 95, 99, or 100 % of the cells in the composition or the CD8+ cells in the composition are naïve CD8+ T cells, central memory CD8+ T cells, effector memory CD8+ T cells, or bulk CD8+ T cells. In some embodiments, at least 20, 30, 40, 50, 60, 70, 80, 90, 95, 99, or 100 % of the cells in the composition, and/or of the CD4+ and/or CD8+ cells in the composition, express the transgene or recombinant molecule, such as the CAR, and/or contain the expression system.

[0267] Additionally provided are methods of making compositions including adoptive immunotherapy compositions, such as those containing the systems, and uses or methods of using these compositions, such as for performing cellular immunotherapy in a subject having a disease or disorder.

[0268] In some alternatives, a method of manufacturing the compositions comprises obtaining generating a modified naïve or naïve-derived or central memory or

central memory-derived CD4⁺ T helper cell or population containing the same, wherein the modified helper T lymphocyte cell preparation comprises CD4⁺ T cells that have a chimeric receptor, such as one comprising a ligand binding domain specific for a tumor cell surface molecule, a spacer domain, a transmembrane domain, and an intracellular signaling domain under control of an inducible promoter as described herein. In other some alternatives, CD4⁺ cells have a cytokine or chemokine receptor under the control of an inducible promoter.

[0269] In some alternatives of the methods described herein, the T-cells are administered with other T-cells that are non-CAR expressing, and/or produce Tam-inducible proteins. In some alternatives, the transcriptional activator for the CAR comprises a DNA-binding domain, a tamoxifen/metabolite binding domain, or a transactivation domain. In some alternatives, one or more, generally all, of the domains from the synthetic transcriptional activator is or are derived from human proteins, such as human or substantially human domains. Such features can reduce immunogenicity of the constructs upon administration to human subjects, for example, in cell therapy.

Expression in Jurkat Cells.

[0270] The “ON” and “OFF” state of Jurkat T cells expressing TamR ZsGreen was studied.

Constructs.

[0271] In some alternatives, this system involves two components: 1) constitutive expression of HEA-3 linked to a single transgene or set of transgenes by T2A skip-linker domains and 2) conditional expression of a transgene that is under control of the HEA-3 restricted synthetic promoter 7xHBD/mEF1ap whereby induction of the transgene occurs in response to tamoxifen. Depending on the desired use of TamRLV genetic control, a combination of delivery systems and vector compositions can be used. Construction of constructs is described above.

[0272] An alternative of the TamR-LV system allows for constitutive HEA3 expression and inducible expression of either ZsGreen or chimeric antigen receptors, permitting kinetic analysis and biological effect controlled in an “ON” and “OFF” manner by the presence or absence of tamoxifen. For this, two constructs were used: 1) HEA-3 is driven by the human EF1a promoter and linked to the constructed truncated EGFR (EGFRt)

transmembrane marker protein by a T2A skip-linker sequence and cloned into a third generation ePHIV7 self-inactivating lentivirus packaging plasmid (construct A). (See **Table 10**; SEQ ID NO: 39; **Table 1**: SEQ ID NO: 8 and SEQ ID NO: 9) and 2) 7xHBD/mEF1ap controls tamoxifen-dependent expression of ZsGreen cloned into pcDNA3.1(-) which was engineered to lack the commercially used CMV promoter (construct B) (See **Table 12**; SEQ ID NO: 41). ZsGreen1 is a human codon-optimized ZsGreen variant that encodes the brightest commercially available green fluorescent protein. (Available from Clontech).

Methods.

[0273] Jurkat cells were transduced at a MOI of 5 with lentivirus packaging constructs described above in which a dual packaging approach in which each plasmid is co-transfected into 293T cells during lentiviral production. This construct utilized molar ratio of 1:2 of construct A to construct B. Construct A encodes the chimeric transcription factor HEA3 linked via a ribosomal cleavage sequence, T2A, to the tracking and selection marker EGFRt. Construct B encodes the synthetic, HEA3-responsive promoter, 7xHBD/mE1b which regulates expression of the transgene ZsGreen-DR1, a short half-life green fluorescent reporter gene.

[0274] After transduction, cells were expanded in culture, enriched for EGFRt expression utilizing Miltenyi magnetic bead selection, to a purity >99%. After further expansion in culture, cells were harvested and treated with Ethanol (Vehicle) as a negative control or 500 nM 4-hydroxytamoxifen (4OHT) for 24 hours, and then were harvested, stained with Erbitux-biotin followed by SA-APC, and analyzed for APC expression and ZsGreen expression via flow cytometry. The results are shown in **Figure 1**.

Dose response.

[0275] Cells were harvested and were then subjected to treatment with 4OHT with concentrations ranging from 0 nM-1000 nM, as indicated. Samples were harvested, washed, and stained with EGFRt-biotin, followed by streptavidin-APC, and then analyzed by flow cytometry for ZsGreen expression. The results are shown in **Figure 2**.

On and Off rate Kinetics.

[0276] The population was subjected to 500 nM 4OHT stimulation for 48 hours, and then sorted for ZsGreen + cells utilizing FACS, yielding a >99% ZsGreen + population on immediate post-sort analysis. Cells were subsequently expanded in culture for 3 weeks, and then prepared for kinetic studies. Cells were divided into 3 treatment groups: (1) no 4OHT (2) 200 nM 4OHT (added every 48 hours) (3) 24 hour 200 nM 4OHT, followed by washout 1x in PBS, then culture in 4OHT free media for 14 days, followed by a 24 hour re-stimulation with 200 nm 4OHT, and then washout in 1x PBS. At each time point, samples were harvested and analyzed via flow cytometry for ZsGreen expression. Results are shown in **Figure 3** and are presented as % ZsGreen+.

Results.

[0277] The results in **Figure 1** show that in the presence of 4-hydroxy tamoxifen (4-OHT), about 50% of the cells express ZsGreen, indicating these cells have both constructs A and B and that 4-OHT induced expression of construct B. The results of the dose response curve show that a concentration of 200 nM or greater of 4-OHT was effective to induce expression of the transgene in transduced cells. (**Figure 2**). The results in **Figure 3** show that as the 4-OHT is washed out of the culture, the Zs Green expression drops off to less than 10% of the max ZsGreen activity within 5 days. When 4-OHT is added back, ZsGreen activity returns to about 100% max activity within 2 days.

Discussion.

[0278] These experiments show that human T Jurkat cells can be transduced with a dual packaged lentivirus with a constitutive component and an inducible component. Expression of the ZsGreen gene is induced in the presence of 4-OHT in a dose responsive manner. In addition, washing out of 4-OHT from the cell culture resulted in a decrease in expression of ZsGreen that could be re-stimulated by addition of 4-OHT back to the cells.

Expression in Primary CD4 Central Memory Cells and CD8 central Memory cells.

[0279] “ON” and “OFF” state of CD4 and CD8 Central memory T cells expressing TamR ZsGreen was studied.

Constructs.

[0280] In some alternatives, this system involves two components: 1) constitutive expression of HEA-3 linked to a single transgene or set of transgenes by T2A skip-linker domains (construct A) (See **Table 10**; SEQ ID NO: 39; **Table 1** SEQ ID NO: 8 and SEQ ID NO: 9) and 2) conditional expression of a transgene that is under control of the HEA-3 restricted synthetic promoter 7xHBD/mEF1ap whereby induction of the transgene occurs in response to tamoxifen (construct B) (See **Table 12**; SEQ ID NO: 41). Depending on the desired use of TamRLV genetic control a combination of delivery systems and vector compositions can be used. Constructs were prepared as described above.

Methods.

[0281] CD4 central memory cells were obtained from peripheral blood by selecting cells through flow cytometry for markers CD4, and CD62L and negative for CD45RO. Cells were cultured with anti-CD3/anti-CD28 beads for 3 days.

[0282] CD8 central memory cells were obtained from peripheral blood by selecting cells through flow cytometry for markers CD8, and CD62L and negative for CD45RO. Cells were cultured with anti-CD3/anti-CD28 beads for 3 days.

[0283] After 3 days, CD4 or CD8 central memory cells were transduced at a MOI of 5 with lentivirus packaging constructs described above in which a dual packaging approach in which each plasmid is co-transfected into 293T cells during lentiviral production. This construct utilized molar ratio of 1:2 of construct A to construct B. Construct A encodes the chimeric transcription factor HEA3 linked via a ribosomal cleavage sequence, T2A, to the tracking and selection marker EGFRt. Construct B encodes the synthetic, HEA3-responsive promoter, 7xHBD/mE1b which regulates expression of the transgene ZsGreen-DR1, a short half-life green fluorescent reporter gene.

[0284] On day 21 after transduction, cells were enriched for EGFRt, and subsequently expanded with feeder cells, IL2, and IL15. Following expansion, cells were divided into 3 treatment groups, 4OHT alone (A), 4OHT combined with CD3/CD28 bead cotreatment (B) or 4OHT alone for 48 hours, followed by addition of CD3/CD28 beads (C). Corresponding samples without 4OHT were also obtained for comparison. All samples were harvested at indicated time points following 4OHT treatment, stained with EGFRt-biotin

antibody, followed by SA-APC, and analyzed for flow cytometry. The results are shown in **Figure 4** (CD4 central memory) and **Figure 5** (CD8 central memory).

Results.

[0285] The results show that the CD4 cells transduced with the dual plasmid vector required the presence of activation stimulus in order to express the ZsGreen even in the presence of tamoxifen. See **Figure 4B**. About 70% of the primary transduced CD4 cells in the presence of tamoxifen and antiCD3/anti-CD28 beads expressed ZsGreen. Gene expression was seen when activation occurred after a 48 hour treatment with tamoxifen. See **Figure 4C**.

[0286] The results show that the CD8 cells transduced with the dual plasmid vector also required the presence of activation stimulus in order to express the ZsGreen even in the presence of tamoxifen. See **Figure 5B**. About 37% of the primary transduced CD8 cells in the presence of tamoxifen and antiCD3/anti-CD28 beads expressed ZsGreen. Gene expression was seen when activation occurred after a 48 hour treatment with tamoxifen. See **Figure 5C**.

[0287] These results show that primary central memory T cells transduced with an inducible construct can express the transgene in the presence of the inducer upon activation with antiCD3/CD28 cells. However, nonactivated cells expressing the transgene can be readily isolated using immunomagnetic or flow cytometry sorting.

Construction of TamR---CD19CAR LV.

[0288] Construction of the vector was accomplished by dual packaging of transfer plasmids housing constructs as described above at a plasmid molar ratio of 1:1. (See **Figure 6C and 7C**) Construct A under the constitutive EF-1alpha promoter encodes TamR-tf (HEA3) linked via a ribosomal cleavage sequence, T2A, to the tracking and selection marker EGFRt. (See **Table 10**; SEQ ID NO: 39, **Table 1**; SEQ ID NO: 8 and SEQ ID NO: 9) Construct B contains the 7xHBD/mE1b promoter with downstream transgene cDNA including CD19CAR linked via 2a cleavage sequence to Her2t, a tracking and selection marker. (See **Table 12**; SEQ ID NO: 41; **Table 2** SEQ ID NO: 10; **Table 13**; SEQ ID NO: 44). Another construct was prepared adding an additional selective marker DHFRdm (See **Table 14**; SEQ ID NO: 46).

[0289] Human Jurkat cells were transduced at a MOI of 2, then were expanded in culture, and selected for EGFRt+ cells via magnetic bead selection. After further expansion, cells were either treated with vehicle alone (EtOH) or 500 nM 4OHT, and harvested for flow

cytometry. Samples were stained with EGFRt-APC antibody and Herceptin-biotin, followed by SA PE. Samples were also prepared for western blot, the primary antibody used is an anti-CD247 mouse mAb which recognizes the intracellular CD3 zeta chain on the CD19CAR (~48 kDa). The endogenous CD3 zeta migrates at ~23 kDa. The results are shown in **Figure 6**.

Jurkat T cells transduced with TamR CD19CAR LV containing a selective marker.

[0290] Construction of TamR CD19CAR LV was done through dual packaging of constructs A and B. Construct A contains HEA3 linked via a ribosomal cleavage sequence, T2A, to the tracking and selection marker EGFRt. Construct B contains the 7xHBD/mE1b promoter with downstream transgene consisting of CD19CAR linked via 2a cleavage sequence to Her2t, a tracking and selection marker, linked via another 2a cleavage sequence (P2A) to DHFRdm, a methotrexate selection gene.

[0291] Cells were transduced with TamR CD19CAR LV at a MOI of 1, expanded in culture, selected for EGFRt via magnetic bead selection. Following further expansion cells were treated with vector alone (EtOH) or 500 nM 4OHT, and harvested for flow cytometry, (upper panel). **Figure 7**. Samples were stained with EGFRt-APC antibody and Herceptin-biotin, followed by SA- PE. Parental (untransduced Jurkat) is shown for comparison. Samples were also prepared for western blot, the primary antibody used is an anti-CD247 mouse mAb which recognizes the intracellular CD3 zeta chain on the CD19CAR (~48 kDa). The endogenous CD3 zeta migrates at ~23 kDa.

Results.

[0292] The results show that EGFRt is detected in 94.6% of cells transduced with a construct not including the additional selective marker DHFRdm. See **Figure 6B**. **Figure 6C** shows that in the presence of tamoxifen, about 42.6% of the cells transduced with a construct not including DHFRdm expressed both EGFRt and Her2t. The cells expressing both markers expressed the CD3zeta chain as part of the CAR construct (48 kDa) as detected in Western blot. See **Figure 6B**.

[0293] **Figure 7** shows that 85.9% Jurkat cells transduced with a construct including another 2a cleavage sequence (P2A) to DHFRdm in the absence of tamoxifen express EGFRt. In the presence of tamoxifen, about 7.5% of the cells expressed both EGFRt

and Her2t. The cells expressing both markers expressed the CD3zeta chain as part of the CAR construct (48 kDa) as detected in Western blot. See **Figure 7B**.

[0294] Inducible expression of CD19 CAR is shown in transduced Jurkat cells. Induction can be measured by detecting expression of the marker Her2t as well as detecting expression of CD19CAR using western blot. Adding an additional selectable marker, DHFRdm, to the inducible construct did not improve inducible gene expression.

TamR CD19CAR LV Transduced human CD4 TCM T cells.

[0295] CD4 central memory cells were obtained from peripheral blood by selecting cells through flow cytometry for markers CD4, and CD62L and negative for CD45RO. Cells were cultured anti-CD3/anti-CD28 beads for 3 days. After 3 days, CD4 Central memory cells were transduced at a MOI of 5 with a lentivirus which packages constructs A and B. (**Figure 8B**). Construct A encodes the chimeric transcription factor HEA3 linked via a ribosomal cleavage sequence, T2A, to the tracking and selection marker EGFRt. (See **Table 10**; SEQ ID NO: 39; **Table 1**; SEQ ID NO: 8 and SEQ ID NO: 9). Construct B encodes the synthetic, HEA3-responsive promoter, 7xHBD/mE1b which regulates expression of the transgene CD19CAR- linked via a T2A to Her2t for tracking and selection, and via a second 2a sequence, P2A, to the methotrexate selection gene DHFRdm. (See **Table 12**; SEQ ID NO: 41; **Table 2** SEQ ID NO: 10; **Table 13**; SEQ ID NO: 44; **Table 14**; SEQ ID NO: 46).

[0296] On day 21 after transduction, cells were enriched for EGFRt utilizing magnetic bead selection, and subsequently expanded with irradiated feeder cells, IL2, and IL15. Two weeks after expansion, cells were cryopreserved. In the experiment depicted above, CD4+ CEM cells which were mock transduced, or CD4 +TamR CD19CAR LV were thawed, placed in culture with CD3/CD28 beads, IL-2, IL-15. CD4+TamRCD19CAR LV were treated or not with 500 nM 4OHT for 24 hours. All samples were harvested washed, then stained with an EGFRt-APC antibody and Her2t-biotin, followed by SA-PE, and analyzed via flow cytometry. The results are shown in **Figure 8**.

Results.

[0297] The results in **Figure 8** show that in the absence of tamoxifen 75.6% of transduced primary CD4 central memory cells express EGFRt, indicating the constitutive

expression of construct A. In the presence of tamoxifen and an activating stimulus (e.g. antiCD3/ antiCD28 beads), about 14.7 % of the primary cells express both EGFRt and Her2t.

[0298] Primary CD4 cells transduced with an inducible vector in the presence of the inducer and an activating stimulus express the transgene under control of the inducible promoter as detected by the expression of the marker gene Her2t. However, nonactivated cells expressing the transgene can be readily isolated using immunomagnetic or flow cytometry sorting. Further characterization of these cells will involve detecting of expression of CD19CAR.

[0299] The foregoing is illustrative of the present invention, and is not to be construed as limiting thereof. The invention is defined by the following claims, with equivalents of the claims to be included therein. All references and documents referred to herein are hereby incorporated by reference.

Table 1

Sequence of anti-CD19 short spacer chimeric receptor

GMCSRss-CD19scFv-IgG4hinge-CD28tm-41BB-Zeta-T2A-EGFRt

Atgtctgtctgtgtgaccagctgtctgtgtgcgagctgccccaccccgctttctgtgatcccc (GMCSFRss) (SEQ ID NO:2)

Gacatccagatgaccagaccactccagctgagcgccagcctggggcaccgggtgaccatcagctgccgggcccagccaggaca
tcagcaagtaacctgaactggtafcagcagaagcccgcacgcaccgtcaagctgctgatctaccaccagccggctgcacagcgcg
tgcccagccggttttagcggcagcgctccggcaccgactacagcctgacatctccaaacctggaacaggagaatatacgcacctactt
ttgccagcaggcaacacactgcccacacctttggcgcggaacaagaactggaatcaccggcagcacctcggcgacggcgaagc
ctggcagcgcgagggcagcaccaaggcgaggtgaagctgcaggaaagcgccctggcctggttgggcccccagccagagcctga
gcgtgacctgcaccgtgagcgcgctgagcctgcccgactacggcgctgagctggaatccggcagccccccaggaaagggcctggaatg
gctggcgctgactctggggcagcgagaccactactacaacagcgccctgaagagccggctgaccatcatcaaggacaacagcaaga
gccaggtgttctgaagatgaacagcctgcagaccgacgacaccgccatctactactgcgcaagcactactactacggggcagcta
cgccatggactactggggccagggcaccagcgtgaccgtgagcagc (CD19scFv) (SEQ ID NO:3)

Gaatctaagtacggaccgacctgcccccttgcct (IgG4hinge) (SEQ ID NO:4)

Atgttctgggtgctgggtggtcgaggcgctggcctgtacagcctgctggtcaccgtggcctcatcatcttttgggtg
(CD28tm-)(SEQ ID NO:5)

Aaacggggcagaaagaaactcctgtatatattcaacaaccatttatgagaccagtacaaactactcaagaggaagatggctgtagctg
ccgatttcagaagaagaagaaggaggatgtgaactg (41BB) (SEQ ID NO:6)

Cgggtgaagttcagcagaagcgcgacgcccctgcctaccagcagggccagaatcagctgtacaacgagctgaacctgggcagaa
gggaagagttacgacgtctctggataagcggagagggccggaccctgagatgggcgggcaagcctcgccggaagaacccccaggaag

gcctgtataacgaactgcagaaagacaagatggccgaggcctacagcgagatcgcatgaaggcgagcggaggcgggcaagg
gccacgacggcctgtatcagggcctgtccaccgccaccaaggatacctacgacgcctgcacatgcaggccctgcccccaagg
(CD3Zeta)- (SEQ ID NO:7)

Ctcgaggcgccggagaggcgaggaagtcttctaactgcggtagctggaggagaatcccgccctagg (T2A) (SEQ
ID NO:8)

Atgcttctctgggtgacaagccttctgctctgtgagttaccacaccagcattctctctgatccacgaaaagtgtgtaacggaataggta
ttggtgaattaaagactcactctccataaatgctacgaatattaacacttcaaaaactgcacctccatcagtggcgatctccacatcctgc
cgggtggcatttaggggtgactccttcacacatactcctcctggatccacaggaaactggatattctgaaaaccgtaagggaatcacag
ggtttttctgattcaggccttggcctgaaaacaggacggacctccatgcctttgagaacctagaaatcatagcggcaggaccaagcaa
catggtcagttttcttctgcagtcgtcagcctgaacataacatccttgggattacgctccctcaaggagataagtgatggagatgtgataatt
tcaggaaacaaaaatttggctatgcaaatacaataaactggaaaaaactgttgggacctccggtcagaaaacaaaaattataagcaac
agagggtgaaaacagctgcaaggccacagccaggtctgcatgcttctctccccgagggtgctggggcccgaggccacggg
actgcgtctcttggcgaatgtcagccgaggcagggaatgcgtggacaagtgcaccttctggagggtgagccaaggagtttggga
gaactctgagtcatacagtgccaccagagtgctgcctcaggccatgaacatcacctgcacaggacggggaccagacaactgtatc
cagtggtccactacattgacggccccactgcgtcaagacctgcccggcaggagtcattgggagaaaacaacacctggtctggaag
tacgcagacgccggccatgtgtgccactgtgccatccaaactgcacctacggatgcactgggcccagggtcttgagggtgtccaacga
atgggcctaagatcccgatccatgccactgggatggtggggccctcctcttctgctgtgtgtgtggccctggggatggcctctcatg/
ga (EGFR) (SEQ ID NO:9)

Table 2**GMCSFRss**

DNA: ATGCTGTGCTGGTGACCAGCCTGCTGCTGTGCGAGCTGCCCCACCCCGCC
 AA: M L L L V T S L L L C E L P H P A

CD19scFv

DNA: TTTCTGTGATCCCC:GACATCCAGATGACCCAGACCACCTCCAGCCTGAGC
 AA: F L L I P D I Q M T Q T T S S L S

DNA: GCCAGCCTGGGCGACCGGGTGACCATCAGCTGCCGGGCCAGCCAGGACATC
 AA: A S L G D R V T I S C R A S Q D I

DNA: AGCAAGTACCTGAAGTGGTATCAGCAGAAGCCCGACGGCACCGTCAAGCTG
 AA: S K Y L N W Y Q Q K P D G T V K L

DNA: CTGATCTACCACACCAGCCGGCTGCACAGCGGCGTCCCCAGCCGGTTTAGC
 AA: L I Y H T S R L H S G V P S R F S

DNA: GGCAGCGGCTCCGGCACCGACTACAGCCTGACCATCTCCAACCTGGAACAG
 AA: G S G S G T D Y S L T I S N L E Q

DNA: GAAGATATCGCCACCTACTTTTGCCAGCAGGGCAACACTGCCTACACC
 AA: E D I A T Y F C Q Q G N T L P Y T

DNA: TTTGGCGGCGGAACAAAGCTGGAAATCACCGGCAGACCTCCGGCAGCGGC
 AA: F G G G T K L E I T G S T S G S G

DNA: AAGCCTGGCAGCGGCGAGGGCAGCACCAAGGGCGAGGTGAAGCTGCAGGAA
 AA: K P G S G E G S T K G E V K L Q E

DNA: AGCGGCCCTGGCCTGGTGGCCCCAGCCAGAGCCTGAGCGTGACCTGCACC
 AA: S G P G L V A P S Q S L S V T C T

DNA: GTGAGCGGCGTGAGCCTGCCCGACTACGGCGTGAGCTGGATCCGGCAGCCC
 AA: V S G V S L P D Y G V S W I R Q P

DNA: CCCAGGAAGGCCTGGAATGGCTGGGCGTGATCTGGGGCAGCGAGACCACC
 AA: P R K G L E W L G V I W G S E T T

DNA: TACTACAACAGCGCCCTGAAGAGCCGGCTGACCATCATCAAGGACAACAGC
 AA: Y Y N S A L K S R L T I I K D N S

DNA: AAGAGCCAGGTGTTCCCTGAAGATGAACAGCCTGCAGACCGACGACCCGCC
 AA: K S Q V F L K M N S L Q T D D T A

DNA: ATCTACTACTGCGCCAAGCACTACTACTACGGCGGCAGCTACGCCATGGAC
 AA: I Y Y C A K H Y Y Y G G S Y A M D

IgG4hinge

DNA: TACTGGGGCCAGGGCACCAGCGTGACCGTGAGCAGC:GAGAGCAAGTACGGA
 AA: Y W G Q G T S V T V S S E S K Y G

CD28tm

DNA: CCGCCCTGCCCCCTTGCCCT:ATGTTCTGGGTGCTGGTGGTGGTGGGAGGC
 AA: P P C P P C P M F W V L V V V G G

DNA: GTGCTGGCCTGCTACAGCCTGCTGGTCACCGTGGCCTTCATCATCTTTTGG
 AA: V L A C Y S L L V T V A F I I F W

41BB

DNA: GTG:AAACGGGGCAGAAAGAAACTCCTGTATATATTCAAACAACCATTTATG
 AA: V K R G R K K L L Y I F K Q P F M

DNA: AGACCAGTACAACTACTCAAGAGGAAGATGGCTGTAGCTGCCGATTTC
 AA: R P V Q T T Q E E D G C S C R F P

CD3Zeta

DNA: GAAGAAGAAGAAGGAGGATGTGAACTGCGGGTGAAG: TTCAGCAGAAGCGCC
 AA: E E E E G G C E L R V K F S R S A

DNA: GACGCCCTGCCTACCAGCAGGGCCAGAATCAGCTGTACAACGAGCTGAAC
 AA: D A P A Y Q Q G Q N Q L Y N E L N

DNA: CTGGGCAGAAGGGAAGAGTACGACGTCCTGGATAAGCGGAGAGGCCGGGAC
 AA: L G R R E E Y D V L D K R R G R D

DNA: CCTGAGATGGGCGGCAAGCCTCGGCGGAAGAACCCAGGAAGGCCTGTAT
 AA: P E M G G K P R R K N P Q E G L Y

DNA: AACGAACTGCAGAAAGACAAGATGGCCGAGGCCTACAGCGAGATCGGCATG
 AA: N E L Q K D K M A E A Y S E I G M

DNA: AAGGGCGAGCGGAGGCGGGCAAGGGCCACGACGGCCTGTATCAGGGCCTG
 AA: K G E R R R G K G H D G L Y Q G L

DNA: TCCACCGCCACCAAGGATACCTACGACGCCCTGCACATGCAGGCCCTGCC
 AA: S T A T K D T Y D A L H M Q A L P

T2A

DNA: CCAAGG:CTCGAGGCGGCGGAGAGGGCAGAGGAAGTCTTCTAACATGCGGT
 AA: P R L E G G G E G R G S L L T C G

EGFRt

DNA: GACGTGGAGGAGAATCCCGGCCCTAGG:ATGCTTCTCCTGGTGACAAGCCTT
 AA: D V E E N P G P R M L L L V T S L

DNA: CTGCTCTGTGAGTTACCACACCCAGCATTCTCCTGATCCCACGCAAAGTG
 AA: L L C E L P H P A F L L I P R K V

DNA: TGTAAACGGAATAGGTATTGGTGAATTTAAAGACTCACTCTCCATAAATGCT
 AA: C N G I G I G E F K D S L S I N A

DNA: ACGAATATTAAACACTTCAAAAACATGCACCTCCATCAGTGGCGATCTCCAC
 AA: T N I K H F K N C T S I S G D L H

DNA: ATCCTGCCGGTGGCATTTAGGGGTGACTCCTTCACACATACTCCTCCTCTG
 AA: I L P V A F R G D S F T H T P P L

DNA: GATCCACAGGAACCTGGATATTCTGAAAACCGTAAAGGAAATCACAGGGTTT
 AA: D P Q E L D I L K T V K E I T G F

DNA: TTGCTGATTACAGGCTTGGCCTGAAAACAGGACGGACCTCCATGCCTTTGAG
 AA: L L I Q A W P E N R T D L H A F E

DNA: AACCTAGAAATCATACGCGGCAGGACCAAGCAACATGGTCAGTTTTCTCTT
 AA: N L E I I R G R T K Q H G Q F S L

DNA: GCAGTCGTCAGCCTGAACATAACATCCTTGGGATTACGCTCCCTCAAGGAG
AA: A V V S L N I T S L G L R S L K E

DNA: ATAAGTGATGGAGATGTGATAATTCAGGAAACAAAATTGTGCTATGCA
AA: I S D G D V I I S G N K N L C Y A

DNA: AATACAATAAACTGGAAAACTGTTTGGGACCTCCGGTCAGAAAACCAA
AA: N T I N W K K L F G T S G Q K T K

DNA: ATTATAAGCAACAGAGGTGAAAACAGCTGCAAGGCCACAGGCCAGGTCTGC
AA: I I S N R G E N S C K A T G Q V C

DNA: CATGCCTTGTGCTCCCCGAGGGCTGCTGGGGCCCGAGCCAGGGACTGC
AA: H A L C S P E G C W G P E P R D C

DNA: GTCTCTTGCCGGAATGTCAGCCGAGGCAGGGAATGCGTGACAAGTGCAAC
AA: V S C R N V S R G R E C V D K C N

DNA: CTTCTGGAGGTGAGCCAAGGGAGTTTGTGGAGAACTCTGAGTGCATACAG
AA: L L E G E P R E F V E N S E C I Q

DNA: TGCCACCCAGAGTGCCTGCCTCAGGCCATGAACATCACCTGCACAGGACGG
AA: C H P E C L P Q A M N I T C T G R

DNA: GGACCAGACAACCTGTATCCAGTGTGCCCCACTACATTGACGGCCCCCACTGC
AA: G P D N C I Q C A H Y I D G P H C

DNA: GTCAAGACCTGCCCGCAGGAGTCATGGGAGAAAACAACACCCTGGTCTGG
AA: V K T C P A G V M G E N N T L V W

DNA: AAGTACGCAGACGCCGCCATGTGTGCCACCTGTGCCATCCAACTGCACC
AA: K Y A D A G H V C H L C H P N C T

DNA: TACGGATGCACTGGGCCAGGTCTTGAAGGCTGTCCAACGAATGGGCCTAAG
AA: Y G C T G P G L E G C P T N G P K

DNA: ATCCCGTCCATCGCCACTGGGATGGTGGGGCCCTCCTCTTGCTGCTGGTG
AA: I P S I A T G M V G A L L L L L V

DNA: GTGGCCCTGGGGATCGGCCTCTTCATGTGA (SEQ ID NO:10)
AA: V A L G I G L F M * (SEQ ID NO:11)

Table 3

ZXR-014 Nucleotide and amino acid sequences (map of sections)

GMCSFRss: nt2084-2149 (SEQ ID NO: 50)

CD19scFv: nt2150-2884 (SEQ ID NO: 51)

Igg4Hinge: nt2885-2920 (SEQ ID NO: 52)

CD28tm: nt2921-3004 (SEQ ID NO: 53)

41BB: nt3005-3130 (SEQ ID NO: 54)

Zeta: nt3131-3466 (SEQ ID NO: 55)

T2A: nt3467-3538 (SEQ ID NO: 56)

EGFRt: nt3539-4612 (SEQ ID NO: 57)

Primers for sequencing:

Oligo name	Sequence	Region
oJ02649	ATCAAAAGAATAGACCGAGATAGGGT	pre-U5 (SEQ ID NO:22)
oJ02648	CCGTACCTTTAAGACCAATGACTTAC	delU3 (SEQ ID NO:23)
oJ02650	TTGAGAGTTTTCGCCCCG	mid-Ampr (SEQ ID NO:24)
oJ02651	AATAGACAGATCGCTGAGATAGGT	post-Ampr (SEQ ID NO:25)
oJ02652	CAGGTATCCGGTAAGCGG	CoE1 ori (SEQ ID NO:26)
oJ02653	CGACCAGCAACCATAGTCC	SV40 (SEQ ID NO:27)
oJ02654	TAGCGGTTTGACTCACGG	CMV (SEQ ID NO:28)
oJ02655	GCAGGGAGCTAGAACGATTC	psi (SEQ ID NO:29)
oJ02656	ATTGTCTGGTATAGTGCAGCAG	RRE (SEQ ID NO:30)
oJ02657	TCGCAACGGGTTTGCC	EF1p (SEQ ID NO:31)
oJ02658	AGGAAGATATCGCCACCTACT	CD19Rop (SEQ ID NO:32)
oJ02601	CGGGTGAAGTTCAGCAGAAG	Zeta (SEQ ID NO:33)
oJ02735	ACTGTGTTTGCTGACGCAAC	WPRES (SEQ ID NO:34)
oJ02715	ATGCTTCTCCTGGTGACAAG	EGFRt (SEQ ID NO:35)

Table 4 Uniprot P0861 IgG4-Fc(SEQ ID NO:13)

10	20	30	40	50	60
ASTKGPSVFP	LAPCSRSTSE	STAALGCLVK	DYFPEPVTVS	WNSGALTSGV	HTFPAVLQSS
70	80	90	100	110	120
GLYSLSSVVT	VPSSSLGTKT	YTCNVDHKPS	NTKVDKRVES	KYGPSPCPSCP	APEFLGGPSV
130	140	150	160	170	180
FLFPPKPKDT	LMISRTPEVT	CVVVDVSQED	PEVQFNWYVD	GVEVHNAKTK	PREEQFNSTY
190	200	210	220	230	240
RVVSVLTIVLH	QDWLNGKEYK	CKVSNKGLPS	SIEKTISKAK	GQPREPQVYT	LPDSQEEEMTK
250	260	270	280	290	300
NQVSLTCLVK	GFYPDSIAVE	WESNGQPENN	YKTPPVLDSD	DGSFFLYSRL	TVDKSRWQEG
310	320				
NVFSCSVME	ALHNHYTQKS	LSLSLGK			
1-98 CH1 (SEQ ID NO: 59)					
99-110 Hinge (SEQ ID NO: 60)					
111-220 CH2 (SEQ ID NO: 61)					
221-327 CH3 (SEQ ID NO: 62)					
Position 108 S→P (SEQ ID NO: 63)					

Table 5 Uniprot P10747 CD28(SEQ ID NO:14)

<u>10</u>	<u>20</u>	<u>30</u>	<u>40</u>	<u>50</u>	<u>60</u>
MLRLLALNL	FFSIQVTGNK	ILVKQSPMLV	AYDNAVNLS	C KSYNLF	SRE FRASLHKGLD
<u>70</u>	<u>80</u>	<u>90</u>	<u>100</u>	<u>110</u>	<u>120</u>
SAVEVCVVYG	NYSQQIQVYS	KTGFNCDGKL	GNESVTFYIQ	NLYVNQTDIY	FCKIEVMYPP
<u>130</u>	<u>140</u>	<u>150</u>	<u>160</u>	<u>170</u>	<u>180</u>
PYLDNEKSNG	TIIHVKGKHL	CPSPLFPGPS	KPFWVLVVVG	GVLACYSLIV	TVAFIIFWVR
<u>190</u>	<u>200</u>	<u>210</u>	<u>220</u>		
SKRSRL	LHSD YNMTPRRPG	PTRKHYQPYA	PPRDFAAAYRS		

- 1-18 signal peptide (SEQ ID NO: 64)
- 19-152 extracellular domain (SEQ ID NO: 65)
- 153-179 transmembrane domain (SEQ ID NO: 66)
- 180-220 intracellular domain (SEQ ID NO: 67)
- Position 186-187 LL→GG (SEQ ID NO: 68)

Table 6 Uniprot Q07011 4-1BB(SEQ ID NO:15)

10	20	30	40	50	60
MGNSCYNIVA	TLLLVLNFER	TRSLQDPCSN	CPAGTFCDNN	RNQICSPCPP	NSFSSAGGQR
70	80	90	100	110	120
TCDICRQCKG	VFRTRKECSS	TSNAECDCTP	GFHCLGAGCS	MCEQDCKQGQ	ELTKKGCKDC
130	140	150	160	170	180
CFGTNDQKR	GICRPWTNCS	LDGKSVLVNG	TKERDVVCGP	SPADLSPGAS	SVTPPAPARE
190	200	210	220	230	240
PGHSPQIISF	FIALTSTALL	FLLFFLTLRF	SVVKRGRKKL	LYIFKQPFMR	PVQTTQEEDG
250					
CSCRFPEEEE GGCEL					
1-23 signal peptide (SEQ ID NO: 69)					
24-186 extracellular domain (SEQ ID NO: 70)					
187-213 transmembrane domain (SEQ ID NO: 71)					
214-255 intracellular domain (SEQ ID NO: 72)					

Table 7 Uniprot P20963 human CD3ζ isoform 3 (SEQ ID NO:16)

10	20	30	40	50	60
MKWKALFTAA	ILQAQLPITE	AQSFGLLDPK	LCYLLDGILF	IYGVILTALF	LRVKFSRSAD
70	80	90	100	110	120
APAYQQGQNO	LYNELNLGRR	EEYDVLDKRR	GRDPEMGGKP	QRRKNPQEGL	YNELQKDKMA
130	140	150	160		
EAYSEIGMKG	ERRRGKGHDG	LYQGLSTATK	DTYDALHMQA	LPPR	

- 1-21 signal peptide (SEQ ID NO: 73)
- 22-30 extracellular (SEQ ID NO: 74)
- 31-51 transmembrane (SEQ ID NO: 75)
- 52-164 intracellular domain (SEQ ID NO: 76)
- 61-89 ITAM1 (SEQ ID NO: 77)
- 100-128 ITAM2 (SEQ ID NO: 78)
- 131-159 ITAM3 (SEQ ID NO: 79)

Table 8 Exemplary Hinge region Sequences

Human IgG1 EPKSCDKTHTCPPCP (SEQ ID NO:17)

Human IgG2 ERKCCVECPPEP (SEQ ID NO:18)

Human IgG3 ELKTPLGDTHTCPPEP (EPKSCDTPPCPPEP)₃ (SEQ ID NO:19)

Human IgG4 ESKYGPPCPPEP (SEQ ID NO:20)

Modified Human IgG4 ESKYGPPCPPEP (SEQ ID NO: 21)

Modified Human IgG4 YGPPCPPEP (SEQ ID NO:36)

Modified Human IgG4 KYGPPCPPEP (SEQ ID NO:37)

Modified Human IgG4 EVVKYGPPCPPEP (SEQ ID NO:38)

Table 9 Her 2 construct-short spacer (SEQ ID NO:1)

GMCSFss-Her2scFv-IgG4hinge-CD28tm-41BB-Zeta-T2A-EGFRt

Leader5 Atgcttctcctgggacaaagccttctgctctgtgagttaccacaccagcattctcctgatccca (SEQ ID NO: 80)**Her2scFV**

Gatatccagatgacccagtcctccgagctccctgtccgcctctgtgggcgatagggtcaccatcacctgccgtgccagtcaggatgt
 gaatactgctgtagcctggatcaacagaaacaggaaaaagctccgaaactactgatttactcggcatccttcttactctggagtc
 cttctcgttctctgtgttcagatctgggacggatttactctgaccatcagcagctcgcagccggaagacttcgcaacttattactgtca
 10 gcaacattatactactctcccacgttcggacagggtaccaaggtggagatcaaaaggcagtagtagcggcggtggctccgggggc
 ggatccgggtggggcgccagcagcgagggtcagctgggtggagctggcggtggcctgggtgcagccagggggctcactccgtttgt
 cctgtgcagcttctggtctcaacattaaagacacctatatactgggtgcgtcagccccgggtaaggcctggaatgggtgcaa
 ggatttactctacgaatggttatactagatagccgatagcgtcaaggccgtttcactataagcgcagacacatccaaaaacacagc
 ctacctgcagatgaacagcctgcgtgctgaggacactgccgtctattattgttctagatggggaggggacggcttctatgctatggact
 15 actggggtcaaggaaacccgtggtcaccgtctcaggt (SEQ ID NO: 81)

Hinge spacer

Gagagcaagtacggaccgctgcccccttgcct (SEQ ID NO: 82)

CD28tm

Atgttctgggtgctggtggtgctggaggcgtgctggcctgtacagcctgctgggtcaccgtggccttcatcttttgggtg
 20 (SEQ ID NO: 83)

4-1BB

Aaacggggcagaaagaaactcctgtatataattcaacaacatttatgagaccagtacaaactactcaaggaagatggctgtag
 ctgccgatttcagaagaagaagaaggaggtggaactg (SEQ ID NO:84)

CD3 zeta

Cgggtgaagtacagcagaagcggcagccccctgcctaccagcagggccagaatcagctgtacaacgagctgaacctggggcag
 aagggaagagtacgactccttgataagcggagggccgggaccctgagatggcgggcgaagcctcggcggaagaaacccccag
 gaaggcctgtataacgaactgcagaagacaagatggccgagcctacagcgagatcgcatgaaaggcgagcggagcgggg
 gcaaggccacgacggcctgtatcaggccgtgtccaccgcccacaaaggatacctacgacgcctgcacatgacggccctcccc
 caagg (SEQ ID NO: 85)

30 **T2A**

Ctcgagggcgggcggagaggcagaggaagtcttctaactgcgggtgacgtggaggagaatcccgccctagg (SEQ ID
 NO: 86)

tEGFR

atgcttctcctgggtgacaagccttctgctctgtgagttaccacaccagcattctcctgatccacgcaaagtgtgaacggaatagg
 35 tattggtgaatttaagactactctccataaatgctacgaatattaacacttcaaaaactgcacctccatcagtgccgatctccacatc
 ctgccgggtgacatttaggggtgactccttcacatactcctcctgtggtccacaggaactggatattctgaaaacgtaaaaggaaat
 cacagggttttctgattcaggttggcctgaaaacaggacggacccatgcctttgagaacctagaatcatacggcgaggac
 caagcaacatggtcagtttctctgagtcgtcagcctgaacataacatccttgggttacgtccctcaaggagataagtgaggag
 atgtgataatttcaggaacaaaaatttctgctatgcaatacaataaactggaaaaaactgttgggacctccggtcagaaaacaaa
 40 attataagcaacagagggtgaaaacagctgcaaggccacagggcaggtctgcatgcttctgctccccgagggtgctggggcc
 cggagcccaaggactgctgtcttggcgaatgtcagccgaggcaggggaatgctgtgacaagtgaaccttctggagggtgagc
 caagggaagtttgggagaactctgagtcatacagtgccaccagagtgctgctcagggccatgaacatcacctgcacaggacgg
 ggaccagacaactgtatccagtggtccactacattgacggccccactgctcaagacctgcccggcagggagtcattgggagaaa
 acaacacctgggtggaagtacgacagcggccatgtgtccacctgtgcatccaaactgcacctacggatgactggggcc

aggctctgaaggctgtccaacgaatgggcctaagatcccgccatgccactgggatgggtggggccctcctcttgctgctgggtgt
ggcctggggatcgcccttcatgtga (SEQ ID NO: 87)

Table 10 TamR-tf (HEA3) nucleic acid (SEQ ID NO: 39)**HEA3 nucleotide (SEQ ID NO:39) and amino acid sequence (SEQ ID NO:40)**

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      M V S K L S Q L Q T E L L A A L L E S G L S K E A L I Q A .
ATGGTG TCCAAGCTGT CCCAGCTGCA GACAGAACTG CTGGCAGCAC TGCTGGAAG OGGCCTGAGC AAGAGGGGCC TGATTGAGG
TACCAC AGGTGAGACA GGTGGAGCT CTGTCTGAG GACGCTGCTG ACGACCTTC GCGSACTCG TTTCTCGGG ACTAAGTGG

· L G E P G P Y L L A G E G P L D K G E S C G G G R G E L A E L P N
AGTCGGGAA CTGGAGCTT ATCTGCTGC TGGGAAAGC CCTCTGGATA AGGGCAGAG CTGTGGCGA GGAAGAGGAG AGCTGGCGA GCTGCTAAC
TGAGCCGCTT GGACCTGGA TAGACGAGG ACCGCTTCG GAGAGCTAT TCCGCTCTG GACACCGCT CTTTCTCTC TCGACCGCT CGAGGGATTG

G L G E T R G S E D E T D D D G E D F T F P I L K E L E N L S P E E
GGCTGGGGG AGACAGAGG CAGCGAGGAC GAGACAGAG ACGACGGCGA GGACTTCAC CCCCCATCC TGAAGAGCT GGAAGACCTG AGCCCGGAGG
CCGACCGGCG TGTCTCTCC GTGCTCTCG CTCTGCTGC TGTGCGCGT CCTGAAGTGG GGGGGGTAGG ACTTTCTCGA CTTTTTGGAC TCGGGGCTCC

· A A H Q K A V V E T L L Q E D P W R V A K M V K S Y L Q Q H N I P .
AAGCGGCCA CCAGAAAGCC GTGGTGGAGA CACTGCTGCA GGAAGATCC TGGGGGTGG CCAAGATGGT CAAGAGCTAC CTGACGAGC ACAACATGCC
TTCGGGGGT GGTCTTTGG CACGACCTGT GTGACGAGT CTTCTTAGG ACCGCCGAG GGTCTACCA GTTCTCGATG GACCTGCTG TTTTGTAGGG

· Q R E V V D T T G L N Q S H L S Q H L N K G T P M K T Q K R A A L
CCAGCGGAG GTGTGGAGA CCACGGGCT GAACGAGAG CACCTGAGC AGCAGCTGAA CAAGGGCACC CCAATGAAA CCGAGAGAG AGCCCGCTG
GGTCGCCCTC CACCACTGT GGTGGCGGA CTTGCTCTG GTGACTCGG TCGTGACTT GTTCCCGTGG GGGTACTTTT GGGTCTCTC TCGGGGGAC

Y T W Y V R K Q R E V A Q Q F T H A G Q G G L I E E P T G D E L P T
TACACTTGT ACGTGGGAA GCAGAGAGAG GTGGCCGAG AGTTTACCA GCGCGGCGG GCGCGCTGA TGGAGGAGC TACCGGAGC GAGTGGCCA
ATGTGAACCA TGCACGCTT CTTCTCTCTC CACCGGGTGG TCAATGTGT GCGGCGGCT CCGCGGACT AGTCTCTGG ATGGCGGCTG CTGACGGGT

· K K G R R N R F K W G P A S Q Q I L F Q A Y E R Q K N P S K E E R .
CCAGAGGGG CAGACGAGC CGTTTAAAT GGGCTCTGC ATCTGAGCAG ATCTGTCTC AGGCTACGA GCGCAGAG AACCCAGCA AAGAGGAGG
GGTCTCTCC GTCTGCTTG GCAATTTCA CCGCGGAGG TAGAGTGTG TAGAGAGAG TCGGATGCT CCGCTCTTC TTGGGTGTG TTTCTCTG

· E T L V E E C N R A E C I O R G V S P S Q A Q G L G S N L V T E V
GGAGACACTG GTGGAGAGT GCAACGGGC CGAGTGATC CAGAGAGGCG TGAGCCCTC TCAGGCTCAG GGCTCGGCA GCAATCTGT CACCGAAGTG
CCTCTGTGAC CACTTCTCA GGTGGGCGG GTCACTGAG GTCTCTCGC ACTGGGAG AGTCCGAGT CCGAGCCGT GTTAGACCA GTGGCTTCA

R V Y N W F A N R R K E E A F R H K L S A G D M R A A N L W P S P L
CGGGTGTACA ATTGGTTGC CAACCGGGG AAGAGGAGG CCTTCGGCA CAGCTGTCT GTGTGGGATA TGAGAGCCG CAGCTGTGG CCGAGCCCC
GCCACATGT TAACCAAGG GTTGGCGGC TTTCTCTTC GGAAGCGGT GTTCAACGA CAGCGCTAT ATTCTCGCG GTTGAACAC GGTCTGGGG

· M I K R S K K N S L A L S L T A D Q M V S A L L D A R P P I L Y S .
TGATGATCA GGGAGCAAG AAGAAGACC TGGCCCTGAG CCGAGCGCC GATCAGATGG TGTGCGCTGT GTTGAGGCC GAGCCCCCTA TCGTGTACG
ACTACTAGT CCGCTGCTC TTTCTGTGG ACCGCGACT GACTGGGCG CTAGTCTACC CAGCGGAGA CAGCTCGCG CTGCGGGAT AGGACATGT

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· E Y D P T R P F S E A S M M G L L T N L A D R E L V H M I N W A K
 CGAGTAGGAC CCGACACGAC CTTTCAGCGA GGCACAGATG ATGGGCGTGC TGACCAACCT GGCCGACCGG GAGCTGGTGC ACATGATCAA CTGGGCGAAG
 GCTCATGCTG GGGTGGTCTG GGAAGTCGCT CCGGTCTGAC TACCCGGACG ACTGGTTGGA CCGGCTGGCC CTGACCAAG TGTACTAGTT GACCCGGTTC

 R V P G F V D L T L H D Q V H L L E C A W L E I L M I G L V W R S M
 CGGGTCCCGG CGATCGTGGG CTTGACCCCTG CACGACAGG TCGACCTGCT GSAATGTCG TGGCTGGAAA TCTGTATGAT CGGCTCGG TGAGAGGGA
 GCCCACGGG CCAAGCACCT GACTGGGAC GTGCTGGTCC AGGTGGACGA CTTTACACGG ACCGACCTTT AGGACTACTA GCGGAGGAC ACCTCTTGGT

 · E H P G K L L F A P N L L L D R N Q G K C V E G M V E I F D M L L
 TGGAACACCC CCGCAAGCTG CTGTTGCCC CCAACCTGCT CTTGACCGG AACAGGGAA ACTGCTGGGA GGGCATGSGT GAGATCTCTG ACATGCTGCT
 ACCTTGTGGG CGGTTGGAC GACAAGCGGG GGTGGACGA GACCTGGCC TTGCTCCCTT TCAAGCACCT CCGTACCAAG CTCTASAGC TGTACAGGA

 · A T S S R F R M M N L Q G E E F V C L K S I I L L N S G V Y T F L
 GGGCAGCTCC AGCGGTTCC GGTATGAA CTTGACGGG GAGGAATTCG TGTGCTGAA GTCCATCATC CTGCTGAACA GCGGCGTGA CACCTTCCTG
 CCGGTGGAGG TCGGCGAAG CTTACTACTT GAGCTGCGG CTCCTTAAGC ACAGGACTT CAGGTAGTAG GAGGACTTGT CCGCGACAT GTGAGAGGAC

 S S T L K S L E E K D H I H R V L D K I T D T L I H L M A K A G L T
 TCAACACCGT TGAAGTCCCT GGAGAGAGG GAACACATCC ACCGGGTGCT GGACAGATC ACGACACCC TGATCCACCT GATGGCCAAG GCTGGCTGA
 AGTAGTGGG ACTTCAGGA CTTCTCTTC CTGCTTAGG TGGCCACGA CTTGTTCTAG TGGCTGTGG ACTAGTGGG CTACCGGTC CAGCGGACT

 · L Q Q Q H Q R L A Q L L L I L S H I R H M S N K R M E H L Y S M K
 CACTCCAGCA CCGACACGAG AGACTGGGCC AGCTGCTGCT GATCTGAGC CACATCGGC ACATGAGCAA CAAGCGGAT GACACCTGT ACAGCATGAA
 GTAGGTCTGT CTGCTGGCT TCTGACCGG TCGACAGCA CTAGGACTCG GTGTAGGCG TGTACTGCT GTTCGCTAC CTGTGAGA TGTGCTACT

 · C K N V V P L Y D L L L E M L D A H R L H A P T S R G G A S V E E
 GTGCAAGAAC GTGGTGCCCC TGTAGACCT GCTGCTGAG ATGCTGGATG CCCACAGACT GCAAGCCCC ACAGCAGAG GCGGAGCCAG CTTGGAGGAA
 CAGTTCCTTG CACCAAGGG ACATGCTGA CAGCAGCTC TACGACCTAC GGTGTCTGA CTTGCGGGA TGTCTGCTC CCGCTGGCT GCACCTCCTT

 T D Q S H L A T A G S T S S H S L Q K Y Y I T G E A S G E P A T V E
 ACCGACAGT CTCACCTGGC CACCGCGGC AGCAGACGA GCCACAGCT GCAGAGTAC TACATCACG GCGAGGCGA GGGATTCCCT GCCACGCTG
 TGGTGTGTA GAGTGACCG GTGGGCGCG TGTGTTCTGT CCGTGTGGA CTTCTTCATG ATGTAGTGG CCGTCCGCT CCGTAAAGGA CCGTGGCAGC

 · F Q Y L P D T D D R H R I E E K R K R T Y E T F E K S I M K K S P F
 AGTTCCAGTA CCGCCGACG ACGAGGACG CCGACCGGAT CGAGGAAAG CTAAGAGAG ATTCAAGAG ATCATGAGA GATCCGCTT CAGCCGCTT
 TCAAGGTGAT GGAAGGCTG TGGTGTGCG CCGTGGCTA GTCCTTTTC GCTTGGCTT GATGCTCTG TAAGTTCTG TAGTACTTCT TACGGGGAA

 · S G P T D P R P P P R R I A V E S R S S A S V E K P A P Q P Y P F
 CAGGCGCCC ACGATCCCA GACCCGCGC TAGAAGATC GCGGTGCGCA GCAGATCTAG CCGCAGCTG CCGCAGCTG CCGCCAGCC CTACCTTTC
 GTGCGCGGG TGGTAGGGT CTGGGGGGG ATCTCTTAG CCGCAGGGT CTTCTAGAT GGGTGGAC GGGTGGAC GGGGGTGG GATGGGAAAG

 T S S L S T I N Y D E E P T M V F P S G Q I S Q A S A L A P A P P Q
 ACCAGAGCC TGAGCAGCAT CAATACGAC GAGTCCCTA CCAATGCTT CCGCAGGCG CAGATCTCT AGGCTCTCT TGTGGCAGT GCTCCAGTC
 TGGTGTGG ACTGTGGA GTTGATGCT CTAAGGGAT GGTACCAA GGGGTGGCG GTCTAGAGG TCGGAGAG AGACCGTGA CAGGTTGAG

 · V L P Q A P A P A P A P A M V S A L A Q A P A P V P P V L A P G P P

AGAGTCTGCT TCAAGGCTCT GCTGACGCG CAGGCGCTGC CATGCTGTCT GCACTGGGCG AGGCTGAGC TGTGCTGCT GTGTGGGCG CTGACTTTC
 TCGAGGAGG AGTCCGGA CAGGTGGG GTGCGGAGG GTACACAGA CTTGACGCG TCGAGATTC AGGACAGA CAGGACCGG GAGCTGGAG

 · Q A V A P P A P K P T Q A G E G T L E F A L L Q L Q F D D N D L G
 TCGAGCTG GCGCTGCT CCGTAAAC TACCTAGCG GCGAGGAA CACTGTGGA GCGCTGCT GAGCTCAGT TCGAGAGA GATCTGGA
 AGTCCAGAC CCGGAGGAG GCGGATTC ATGGTGGG CCGTCTCT GTGACATCT CCGGAGAG GTGAGTGA ACTGTGCT CCGGAGCT

 A L L G N E T D P A V F T D L A S V D N S E F Q Q L L N Q G I P V A
 GAGCTGCT GCAATACAC CAGGCGGCT CTCTTACG ACCTGGCTG CTTGACAC AGCAGCTTC AGCAGCTCT CAACAGGCT ATCCCTGCT
 CTTGAGGAG CTTATGCT GTGGGCGG CACAATGCT TGGAGGAG GCACTGTT TCTCTAAG TGTGAGGA GTTGCTCGG TAGGAGAG

 · P H T T E P M L N E Y F R A I T R L V T G A Q R P P D P A P A P L
 CCGCAGAC CAGGAGGCT ATGCTGAG AATACCGCA GCGATCAC AACTGTTA CAGGCGGGA GAGGCTTCA GATCAGAG CAGTCTACT
 GGGTGTGCT GTGGTGGG TAGACTAC TTATGGGCT CCGGATGCT TCTGACAGT GTGCGGCT CCGGAGCT CTAGGCTG GTGAGTGA

 · G A F G L P N G L L S G D E D F S S I A D M D F S A L L S Q I S S
 GAGAGGCT GAGTCTGTA ATGGCTGCT GTTGGGAG GAGGACTCT CCAGGATTC CAGATGAG TTCAAGGCG TGTGTGCA GATCAGAG
 CCGTGGGA CCGAGGAT TACCGAGA CAGAGGCTG CTGCGAGA GGTGTAAG GTGTACTG AGTGTGCG AGGAGAGCT CTAGTCTG

Table 11 TamR-tf (HEA4) nucleic acid (SEQ ID NO: 42) amino acid sequence (SEQ ID NO: 43)**HEA4 Nucleotide(SEQ ID NO: 42) and Amino Acid Sequence**

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      M V S K L S Q L Q T E L L A A L L E S G L S K E A L I Q A
      ATGGTG TCGAAGCTCT CCGAGCTGCA GACAGAATG CAGCAGACAG TGTGGAAAG CAGCTGAGC AAGAGAGCC TGATTGAGG
      TACGAC AGATTGACA GCGTGAGCT GTGTGTGAG GACGCTGGG AGAGCTTTG GCGAGACTG TTTTCTGGG ATTAAGTGG
      * L G E F G G P Y L L A G E G P L D K G E S C G G G R G E L A E L F N
      ACTGGGGA CCGTGACTT ATCTGCTCC TGGCAGAGC CCGTGGATA AAGGCGAAG CTTGGGAGA GAAGAGAGG AGCTGGCGA GGTGCTAAC
      TGAGCGCTT GAGCTGAA CAGAGAGCG AGCGTTCCG GAGAGCTAT TCGGCTCTC GACAGCGCT CTTCTCTC TCGAGCGGT GAGAGATTG
      G L G E T R G S E L E T D D D G E D F T P P I L K E L E N L S P E E
      GCGTGGGG AGAGAAGAG CAGCAGAGC GAGAGAGAG AGCAGCGGA GAGCTTACG CCGCGATCG TGAAGAGCT GAGAAAGCT AGCGCGAGG
      CAGAGCTGG TGTCTCTC GTGCTCTG GTCTCTCTG TCGTGGCTT CCGAGAGGG GAGGAGTAG ACTTTTGA CTTTGGAG TCGAGCTCT
      * A A R Q K A V V E T L L Q E D P W R Y A K M V K S Y L Q Q H N I P
      AAGCTCTCA CAGAAAGC GTGCTGAGA CACTCTGCA GAGAGATCC TGGAGCTCG CCAAGATGT CAGAGTAC CTTGAGAGC ACAATCTCC
      TTGGCGGCT GTCTTTGAG CAGAGCTCT GTGAGAGCT CTTTAGGG AGCTCGAGC GGTCTACA GTTCTGATG GAGCTCTCG TGTGTAGG
      * Q R E V V D T T G L N Q S H L S Q H L N K G T P M K T Q K R A A L
      CAGCGGGG GTGCTGACA CAGAGAGCT GAGCAGAGC CAGCAGAGC AGCAGCTGA CAGGCTAGC CCGATGAAA CCGAGAGAG AGCGCGCTG
      GGTGGCTTC CAGAGCTGT GTGCGCGGA CTTGCTCTG GTGAGCTGG TGTGAGCTT GTTCTCTCG GAGTACTTT GGTCTCTCT TCGAGAGAG
      Y T W Y V R K Q R E V A Q Q F T H A G Q G G L I E E P T G D E L P T
      TACTCTGCT AATGCGAAA GAGAGAGAG GTGCTGAGC AGTTTACCA CCGTGGTAG GCGCTGTA TGAAGAGAG TACAGAGAG GAGCTCTCA
      AZTGAGACA TCGAGCTCT CTTCTCTCT CAGCGCTCT TGAATGTGT GCGCTGAGT CCGCGAGCT AGCTCTTG AGCGCGCTG CTTGAGAGT
      * K E G E R N K F N W G P A S Q Q I L F Q A Y S R Q E N P S K E E R
      CAGAGAGGG CAGAGAGAG CCGTTTAACT GCGCGCTCTC ATCTGAGAG ATCTCTCTC AGCGCTAGA GCGCGAGAG AAGCGAGAG AAGAGAGAG
      GTTCTCTCT GTGCTCTG GCGAATGCA CCGCGGAGC TGAAGCTCT CAGAGAGAG TGTGAGAGT CCGCTCTCT TGAAGTGT TTTCTCTG
      * E T L V S E C N R A E C I Q R G V S E S Q A Q G L G S N L V T E V
      GAGAGACTG CTTGAGAGT CAGAGCGCG CAGTGGCATC GAGAGAGCG TGAAGCTCT TGAAGCTCTG GCGCTGCGA CCAATCTGT CAGCAGAGT
      CTTCTCTAG CAGCTCTCA CTTGCGCG GCGAGCTAG GTTCTCTCT ACTCGAGAG AGTCTGAGT CCGAGAGCT CTTAGAGCA GTGCTCTAG
      R V Y N W P A N K R K E E A P R H K L S A G D M R A A N L W P S P L
      CAGCTGTACA ATCTCTCT CAGCGCGCG AAGAGAGAG CTTCTGCGA CAGCTCTCT GTTCTGATA TGAAGCTCT CAGCTGTCT CAGAGCGCG
      GCGAGAGAG TGAAGAGAG TTTCTCTCT GAGAGCGCT GTTCTGAGA CAGAGCTAT ACTCTGAGG GTTCTGAGC GGTCTGAGG
      * N I K E S K K N S L A L S L T A D Q M V S A L L D A K P P I L Y S
      TGATGTCAA CCGAGAGAG AAGAGAGCT TGGCTGAG CTTGAGCTCT GATGAGATG TGTCTCTCT GTTCTGAGT GAGCGCTCT TCTCTGAG
      ATCTACTTT CCGCTCTCT TTTCTCTCT AGCGAGCT GAGTCTGAG CAGCTTAC AGAGAGAG GAGCTCTCT CTTGAGAGT AGGAGAGT

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· E Y D P T R P F S E A S M M G L L T N L A D R E L V H M I N W A K
CGAGTACGAC CCCACCAGAC CCTTCAGCGA GGCCAGCATG ATGGGCTTGC TGACCAACCT GGCCGACCGG GAGCTGGTGC ACATGATCAA CTGGGCCAAG
GCTCATGTG GGGTGGTCTG GGAAGTCGCT CGGCTGTAC TACCGGAGC ACTGGTTGGA CGGCTGGCC CTGACACAG TGTACTAGTT GACCGGTTCT

R V P G F V D L T L H D Q V H L L E C A W L E I L M I G L V W R S M
CGGCTGCCCG GCTTGTGGA COTGACCCTG CACGACCAGG TCCACCTGCT GGAATGTGCG TGGCTGGAAA TCCTGATGAT CGGCTCTGTG TGGAGAAGCA
GCCACGCGGC GGAAGCACT GACTGAGAC GTGCTGTCC AGGTGGAAGA CTTTACACGG ACCGACCTTT AGGACTACTA GCGGAGACAC ACCTCTTCTG

· E H P V K L L F A P N L L L D R N Q G K C V E G M V E I F D M L L ·
TGGAACACC CGTGAAGCTG CTGTTGCCCC CCAACCTGCT CCTGGACCGG AACGAGGGA AGTGCGTGA GGSCATGGTG GAGATCTTCG ACATGCTGCT
ACCTTGTGG GACTTCGAC GACAAGGGG GGTGGAAGA GGACCTGGCC TTGGTCCCTT TCACGCACCT CCGTAACAC CTCTAGAAGC TGTACGAAGA

· A T S S R F R M M N L Q G E E F V C L K S I I L L N S G V Y T F L
GGCCAGCTCC AGCGGTTCC GGATGATGAA CTTGACGGGC GAGGAATTGG TGTGCTGAA GTCCATCATC CTGCTGAACA GCGGCTGTA CACCTTCCTG
CCGTTGGAGG TCGGCCAAGG CTTACTACTT GGAAGTCCCG CTCCTTAAGC ACACGGACTT CAGGTAGTAG GAAGACTTGT CGCGCACAT GTGGAAGGAC

S S T L K S L E E K D H I H R V L D K I T D T L I H L M A K A G L T
TCATCCACCC TGAAGTCCCT GGAAGAGAAG GACCACATCC ACCGGTGTCT GGACAAGATC ACCGACACCC TGATCCACCT GATGGCCAG GCTGGCTGA
AGTAGTGGG ACTTCAGGA CTTCTCTTC CTGTTGTAGG TGGCCCAAGA CTTGTTCTAG TGGCTGTGGG ACTAGGTGA CTACCGGTTT CGACGGGACT

· L Q Q Q H Q R L A Q L L L I L S H I R H M S N K G M E H L Y S M K ·
CACTCCAGCA GCAGCACAG AGACTGGGCC AGCTGCTGCT GATCCTGAGC CACATCCGCC ACATGAGCAA CAGGGATG GAACACCTGT ACAGCATGAA
GTGAGTGTG GTGCTGGTC TGTGACCGG TOGACGAAGA CTAGGACTCG GTGTAGGCCG TGTACTCGTT GTTCCCTTAC CTTGTGGACA TGTCTACTTT

· C K N V V P L Y D L L L E A A D A H R L H A P T S R G G A S V E E
GTSCAAGAAC GTGGTGGCC TGTACGACT GTGCTGAGG GCTGCGGATG CCAACAGACT GCAAGCCCT ACAAGCAGAG GCGAGCCAG CTTGGAAGAA
CACGTTCTTG CACACGGGG ACATGCTGA CAGGAGCTC CAGCGCTAC GGTGTCTGA CGTGCGGGA TGTGCTCTC CGCTCGGTC GCACCTCTCT

T D Q S H L A T A G S T S S H S L Q K Y Y I T G E A E G F P A T V E
ACCGACAGT CTCACCTGGC CACGCGCGC AGCACAGCA GCCACAGCCT GCAGAAGTAC TACATCACCG GCGAGGCGA GGGATTCCCT GCCACGCTGG
TGCTGTGCA GAGTGAAGG GTGGCGCGC TGTGTGCTG CGTGTGGA CGTCTTCATG ATGTAGTGC CGCTCGGCT CCTAAGGGA CGTGGCACC

· F Q Y L P D T D D R H R I E E K R K R T Y E T F K S I M K K S P F ·
AGTTCCAGTA CTTGCCGAC ACCGACGACC GGCACCGAT CAGGAAAAG GGAAGGGA CCTACGAGAC ATTCAAGAGC ATCATGAAGA AGTCCCTCTT
TCAAGTCAAT GGACGGGCTG TGGCTGCTG CCGTGGCCTA GCTCTTTTC GCCTTGGCT GATGCTCTG TAAGTTCTG TGTACTTCT TCAGGGGGA

· S G P T D P R P P P R R I A V P S R S S A S V P K E A P Q P Y P F
CAGCGGCCCC ACCGATCCCA GACCCCCC TAGAAGAATC GCGGTGCCA GCAGATCTAG CGCCAGCGTG CCAAGCCTG CCCCCAGCC CTACCTTTTC
GTGCGCGGG TGGTAGGGT CTGGGGGGG ATCTTCTTAG CGGACGGGT GGTCTAGATC GGGTGGCAC GGGTGGAC GGGGGGTGG GATGGGAAAG

T S S L S T I N Y D E F P T M V F P S G Q I S Q A S A L A P A P P Q
ACGAGGAGC TGAGCACCAT CAACTACGAC GAGTCCCTA CCAATGTTT CCGAGGGC CAGATCTCT AGGCTCTG TCTGGCACCT GCTCCACCTC
TGGTCTGG ACTGTGTA GTGATGCTG CTCAGGGAT GSTACCAAA GGGTGGCG GTCTAGAGG TCGGAGAG AGACCGTGA GAGGTGGAG

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· V L P Q A P A P A P A P A M V S A L A Q A P A P V P V L A F G P P ·
AGGTGTGTC TCAGGCCCC GTCCAGCCC CAGCCCTTGC CATGTGTCT GCACGTGGCC AGGCTCCAG TCCTGTGCT GTGCTGGCC CTGGACCTCC
TCCAGACGG AGTCGGGGG CAGGCTCGG GTCCGGGACG GTACACAGA CGTGACCGG TCCGAGGTG AAGACACGA CACGACCGG GACCTGGAGG

· Q A V A P P A P K P T Q A G E G T L S E A L L Q L Q F D D E D L G
TCAGGTGTG GCCCCCTCTG CCCCCTAAC TCACCAGGCC GGGGAGGAA CACTGTGTGA GCCCCCTGT CAGCTCCAGT TCGACACGA GGATCTGGGA
AGTCCGACAC CGGGGAGGAC GGGGATTGG ATGGGTCCG CCCCCTCTT GTGACAGACT CCGGGACGAC GTGAGGTGA AGTGTGTCT CCTAGACCTT

A L L G N S T D P A V F T D L A S V D N S E F Q Q L L N Q G I P V A
GCATGTGG GCAATAGCAC CGACCCCGCC GTGTTTACG ACCTGGCTTC CGTGGACAC AGCGAGTTC AGCAGCTCT CACCTAGGCG ATCCCTGTG
CGTAGACAC GTTATCTGT GCTGGGGGG CACCAATGCG TGGACCGGAG GCACCTGTG TCGTCAAGG TGTGTAGGA GTTGTCCCG TAGGGACAGC

· P H T T E P M L M E Y P E A I T R L V T G A Q R P P D P A P A P L ·
CCCCACAC CACCGAGGCC ATGCTGATGG AATACCCGGA GGCAATCACC AGACTGTGA CAGGCGGCCA GAGGCTTCA GATCCAGCAC CAGCTCCACT
GGGTGTGTG GTGGCTGGG TAGACTACG TTATGGGCT CCGGTAGTGG TCTGACCACT GTCCGGGGT CTCGGAGGT CTAGGTGTG GTGAGGTGA

· G A P G L P N G L L S G D E D F S S I A D M D F S A L L S Q I S S
GGGAGCCCT GGCCTGCTA ATGGGTGTG GTCTGGGAC GAGGACTTCT CCAGCATTC CACATGGAC TTCAGCGCC TGCTGTCCA GATCAGCAGC
CCCTCGGGA CCGACGGAT TACCGACGA CAGACCGCT CTCTGAAGA GTTCTAAGC GCTGTACCT AAGTGGGG ACGACGGGT CTAGTGTG

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Table 12 7xHBD/mEF1α nucleic acid sequence (SEQ ID NO: 41)

Tagttaataatcacaatagttaataatctacaatagttaataatctacaatagttaataatctacaatagttaataatctacaat
agttaataatctacaa

Table 13 Her2t nucleic acid (SEQ ID NO: 44) and amino acid sequence (SEQ ID NO: 45)**HER2t (CHP) Nucleotide and Amino Acid Sequence**

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M L L L V T S L L L C E L P H
ATGCT TCTCTGGTG ACAAGCCTTC TGCTCTGTGA GTTACCACAC
TAGGA AGAGGACCAC TGTGGGAG ACGAGACACT CAATGGTGTG

P A F L L I P C H P E C Q P Q N G S V T C F G P E A D Q C V A C A H
CCAGCATTC TCCTGATCCC ATGCCACCT GAGTGTGAG CCCAGAAAG CTCAGTGACC TGTTTTGGAC CGGAGGCTGA CCAGTGTGTG GCCTGTGCCC
GGTGTGTAAGG AGGACTAGGG TAGGGTGGGA CTCACAGTCG GGGTCTTACC GAGTCACTGG ACAAAACCTG GCCTCCGACT GGTACACAC CCGACCGGG

· Y K D P P F C V A R C P S G V K P D L S Y M P I W K F P D E E G A ·
ACTATAAGGA CCTCCCTTC TGCGTGGGCC GCTGCCCGAG CGGTGTGAAA CCTGACCTCT CCTACATGCC CATCTGAGG TTTCAGATG AGGAGGGCGC
TGATATTCCT GGGAGGGAG ACGACCGGG CAGCGGGTC SCCACACTTT GGA CTGGAGA GGATGTACGG GTAGACCTTC AAAGTCTAC TCCTCCCGG

· C Q P C P I N C T H S C V D L D D K G C P A E Q R A S P L T S I I
ATGCCAGCT TGCCCATCA ACTGCACGA CTCCTGTGTG GACTCGATG AGAAGGGCTG CCGCGCGAG CAGAGAGCCA GCGCTGTGAC CTCATCATG
TAGGTGGA ACGGGGTAGT TGACGTGGT GAGGACACAC CTGGACTTAC TGTTCCTGAC GGGCGCGTC GTCTCTGCT CCGGAGACTG CAGGTAGTAG

S A V V G I L L V V V L G V V F G I L I *
TCTCGGTGG TTGCATTCT GCTGGTGTG GTCTGGGGG TGCTCTTTGG GATCCTCATG TGA
AGACGCCACC AACGTAGA CGACAGCAC CAGAACCCCG ACCAGAAACC CTAGGATAG ACT

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Table 14 DHFRdm nucleic acid (SEQ ID NO: 46) and amino acid (SEQ ID NO: 47)**DHFRdm Nucleotide and Amino Acid Sequence**

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M V G S L N ·
ATGGTTG GTTCGCTAAA
TACCAAC CAAGOGATTT

· C I V A V S Q N M G I G K N G D F P W P P L R N E S R Y F Q R M T
CTGCATCGTC GCTGTGTCCC AGAACATGGG CATCGGCAAG AACGGGGACT TCCCTGGCC ACCGCTCAGG AATGAATCCA GATATTTCOA GAGAATGACC
GACGTAGCAG CGACACAGGG TCTTGTAACC GTAGCCGTTT TTGCCCCGTA AGGGGACCGG TGGCGAGTCC TTACTTAGGT CTATAAAGGT CTCCTACTGG

T T S S V E G K Q N L V I M G K K T W F S I P E K N R P L K G R I N
ACAACCTCTT CAGTAGAAGG TAAACAGAAAT CTGGTGATTA TGGGTAAGAA GACCTGGTTC TCCATTCTCG AGAAGAATCG ACCTTTAAAG GGTAGAATTA
TGTTGGAGAA GTCATCTTCC ATTTGTCTTA GAOCCTAAT ACCGATTCTT CTGGACCAAG AGGTAAGGAC TCTTCTTAGC TGGAAATTTT CCATCTTAAT

· L V L S R E L K E P P Q G A H F L S R S L D D A L K L T E Q P E L ·
ATTTAGTTCT CAGCAGAGAA CTCAGGAAC CTCACAAAG AGCTCATTTT CTTTCCAGAA GTCTAGATGA TGGCTTAAAA CTTACTGAAC AACCAGAATT
TAAATCAAGA GTCTCTCTT GAGTCTCTTG GAGGTGTTCC TCGAGTAAAA GAAAGGTCTT CAGATCTACT ACGGAATTTT GAATGACTTG TTGGTCTTAA

· A N K V D M V W I V G G S S V Y K E A M N H P G H L K L F V T R I
AGCAANTAAA GTAGACATGG TCTGGATAGT TGGTGGCAGT TCTGTTTATA AGGAAGCCAT GAATCACCCA GGGCATCTTA AACTATTGT GACAAGGATC
TCGTTTTATT CATCTGTACC AGACCTATCA ACCACCGTCA AGACAAATAT TCCTTCGGTA CTTAGTGGGT CCGGTAGAAT TTGATAAACA CTGTTCTAG

M Q D F E S D T F P P E I D L E K Y K L L P E Y P G V L S D V Q E E
ATGCAAGACT TTGAAAGTGA CAGCTTTTTT CCAGAAATTG ATTTGGAGAA ATATAAACTT CTGCCAGAAAT ACCCAGGTGT TCTCTCTGAT GTCCAGGAGG
TAGGTTCTGA AACTTTCACT GTGCAAAAAA GGTCCTTAAAC TAAAGCTCTT TATATTTGAA GACGGTCTTA TGGGTCCACA AGAGAGACTA CAGGTCTTCC

· K G I K Y K F E V Y E K N D
AGAAAGGCAT TAAGTACAAA TTGAAGTAT ATGAGAAGAA TGAT
TCTTTCGTA ATTCATGTTT AAACCTCATA TACTCTTCTT ACTA

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WHAT IS CLAIMED IS:

1. A system for inducible expression of a chimeric antigen receptor comprising:
 - a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a chimeric antigen receptor comprising a ligand binding domain, wherein the ligand binding domain is specific for a ligand, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain; and d) a polynucleotide coding for an intracellular signaling domain; and
 - b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter.
2. The system of claim 1, wherein the drug is tamoxifen and/or its metabolites.
3. The system of claim 2, wherein the first promoter comprises a nucleic acid sequence of SEQ ID NO: 41.
4. The system of any one of claims 1-3, wherein the second promoter is a constitutive promoter.
5. The system of claim 4, wherein the second promoter is the EF1 α p.
6. The system of any one of claims 1-5, wherein the transcriptional activator comprises a sequence of SEQ ID NO: 40.
7. The system of any one of claims 1-6, wherein the first nucleic acid further comprises a first vector and the second nucleic acid further comprises a second vector.
8. The system of claim 7, wherein both vectors are packaged in a viral vector.
9. The system of claim 8, wherein the viral vector is a lentivirus.

10. The system of any one of claims 1-6, wherein the first and second nucleic acid comprise a vector.
11. The system of any one of claims 1-10, wherein the first nucleic acid further comprises a nucleic acid sequence coding for a selectable marker.
12. The system of anyone of claims 1-11, wherein the second nucleic acid further comprises a nucleic acid coding for a selectable marker.
13. A system for inducible expression of chimeric antigen receptor comprising:
 - a) a first nucleic acid comprising a first promoter inducible by a drug, wherein the first nucleic acid is operably linked to a polynucleotide coding for a cytokine, a chemokine receptor, a polypeptide that regulates apoptosis, or a polypeptide that modulates checkpoint signaling; and
 - b) a second nucleic acid comprising a second promoter operably linked to nucleic acid coding for a transcriptional activator for the inducible promoter.
14. The system of any one of claims 1-13, wherein the second nucleic acid further comprises a polynucleotide coding for chimeric antigen receptor comprising a ligand binding domain, wherein the ligand is a tumor specific molecule, viral molecule, or any other molecule expressed on a target cell population, wherein the ligand can elicit recognition, modulation, inhibition, and/or elimination by a lymphocyte; a polynucleotide coding for a polypeptide spacer, wherein the spacer is optimized; a polynucleotide coding for a transmembrane domain; and d) a polynucleotide coding for an intracellular signaling domain.
15. The system of any one of claims 1-14 wherein the first promoter is in opposite orientation to the second promoter.
16. The system of any one of claims 1-15, wherein the ligand binding domain is an antibody fragment.

17. The system of claim 16, wherein the ligand binding domain is single chain variable fragment.
18. The system of any one of claim 1-17, wherein the tumor specific molecule is selected from the group consisting of CD19, CD20, CD22, CD23, CD123, CS-1, ROR1, CE7, EGFR, hB7H3, mesothelin, c-Met, PSMA, Her2, GD-2, MAGE A3 TCR and combinations thereof.
19. A chimeric receptor polypeptide coded for by the first second and/or second nucleic acid of the system of any one of claims 1-18 or 34-38.
20. A host cell comprising a system of any one of claims 1-18 or 34-38.
21. The host cell of claim 20, wherein the host cell is a CD8⁺ T cytotoxic lymphocyte cell selected from the group consisting of naïve CD8⁺ T cells, central memory CD8⁺ T cells, effector memory CD8⁺ T cells and bulk CD8⁺ T cells.
22. The host cell according to claim 21, wherein the host cell is a CD4⁺ T helper lymphocyte cell that is selected from the group consisting of naïve CD4⁺ T cells, central memory CD4⁺ T cells, effector memory CD4⁺ T cells, and bulk CD4⁺ T cells.
23. A composition comprising a host cell of any one of claims 20-22 or 39-40, in a pharmaceutically acceptable excipient.
24. The composition of claim 23 comprising a host cell of claim 21 and a host cell of claim 22.
25. An *in vitro* method for preparing a host cell of any one of claims 20-22 or 39-40 comprising:
 - a) providing a system of any one of claims 1-18 or 34-38; and
 - b) introducing the system into a separate isolated T lymphocyte population and expanding each T lymphocyte population *in vitro*.

26. The method of claim 25, wherein the T lymphocytes are expanded, and wherein the method further comprises culturing the cells in the presence of anti-CD3 and/or anti CD28, and at least one homeostatic cytokine until the cells expand sufficiently for use as a cell infusion.
27. The method of any one of claims 25 or 26, wherein the lymphocyte is CD8⁺ or CD4⁺.
28. A use of the host cell of any one of claims 20-22 or 39-40 or the composition of claims 23-24 or 41 in combination with a drug that induces expression of a transgene in the host cell or composition for the treatment of cancer or a viral infection.
29. The use of claim 28, wherein the cancer is a solid tumor or hematologic malignancy.
30. The use of claim 29, wherein the solid tumor is selected from the group consisting of a breast cancer, brain cancer, lung cancer, colon cancer, renal cancer, pancreatic cancer, prostate cancer, and ovarian cancer.
31. A method of performing cellular immunotherapy in a subject having cancer or a viral infection comprising: administering the composition of any one of claims 23 to 24 or a host cell of any one of claims 20-22 or 39-40 to the subject and administering a drug that induces expression of a transgene in the composition or the host cells.
32. The method of claim 31, wherein the cancer is selected from a solid tumor or hematologic malignancy.
33. The method of claim 32, wherein the solid tumor is selected from the group consisting of a breast cancer, brain cancer, lung cancer, colon cancer, renal cancer, pancreatic cancer, prostate cancer, and ovarian cancer.
34. The system of any one of claims 1-3 or 4-12 wherein the second promoter is an inducible promoter.

35. The system of any one of claim 1-12 or 34, wherein the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor.

36. The system of any one of claims 13-18, wherein the second promoter is an inducible promoter.

37. The system of any one of claims 13-18, wherein the second promoter is constitutive promoter.

38. The system of any one of claims 14-18 or 36-37, wherein the spacer is optimized for increased T cell proliferation and/or cytokine production in response to the ligand as compared to a reference chimeric receptor.

39. The host cell of claim 20, wherein the host cell is a precursor T cell.

40. The host cell of claim 39, wherein the precursor T cell is a hematopoietic stem cell.

41. The composition of claim 23 comprising a host cell of claim 21 or 22 and a host cell of claim 39 or 40.

42. The method of claim 25 or 26, wherein the isolated T lymphocyte population comprises precursor T cells.

43. The method of claim 42, wherein the precursor T cells are hematopoietic stem cells.

44. The method of claim 31, wherein the administering of the drug is performed with the administering the composition or host cells.

45. The method of claim 31, wherein the administering of the drug is performed after administering of the composition or host cells, wherein administering is performed 1 day, 2 days, 3 days, 4 days, 5 days, 6 days, 7 days, 2 weeks, 4 weeks or two months, or any time in between any two values of time listed.