TITLE OF THE INVENTION
PRODUCTION OF GLYCOPROTEINS WITH MODIFIED FUCOSYLATION

BACKGROUND OF THE INVENTION

(1) Field of the Invention

The present invention relates to the field of glycobiology, and in particular to methods for genetically engineering host cells that lack an endogenous pathway for fucosylating *N*-glycans of glycoproteins to be able to produce glycoproteins with fucosylated *N*-glycans.

(2) Description of Related Art

Therapeutic proteins intended for use in humans that are glycosylated should have complex, human *N*-glycosylation patterns. In general, it would be advantageous to produce therapeutic proteins using bacterial or eukaryotic microorganisms because of (a) the ability to rapidly produce high concentrations of protein; (b) the ability to use sterile, well-controlled production conditions (for example, GMP conditions); (c) the ability to use simple, chemically defined growth media; (d) ease of genetic manipulation; (e) the absence of contaminating human or animal pathogens; (f) the ability to express a wide variety of proteins, including those poorly expressed in cell culture owing to toxicity etc.; and, (g) ease of protein recovery (for example, via secretion into the medium). However, prokaryotes and lower eukaryotes do not normally produce proteins having complex *N*-glycosylation patterns. Therefore, animal cells are generally used to produce therapeutic proteins where it is desirable that the protein have a complex, human-like *N*-glycosylation pattern. But, there are a number of significant drawbacks to using animal cells for producing therapeutic proteins.

Only certain therapeutic proteins are suitable for expression in animal cells (for example, those lacking in any cytotoxic effect or other effect adverse to growth). Animal cell culture systems are usually very slow, frequently requiring over a week of growth under carefully controlled conditions to produce any useful quantity of the protein of interest. Protein yields nonetheless compare unfavorably with those from microbial fermentation processes. In addition, cell culture systems typically require complex and expensive nutrients and cofactors, such as bovine fetal serum. Furthermore, growth may be limited by programmed cell death (apoptosis).

Moreover, animal cells (particularly mammalian cells) are highly susceptible to viral infection or contamination. In some cases the virus or other infectious agent may compromise the growth of the culture, while in other cases the agent may be a human pathogen rendering the therapeutic protein product unfit for its intended use. Furthermore, many cell culture processes require the use of complex, temperature-sensitive, animal-derived growth media components, which may carry pathogens such as bovine spongiform encephalopathy

(BSE) prions. Such pathogens are difficult to detect and/or difficult to remove or sterilize without compromising the growth medium. In any case, use of animal cells to produce therapeutic proteins necessitates costly quality controls to assure product safety.

Recently, it has been shown that lower eukaryotes, particularly yeast, can be genetically modified so that they express proteins having complex *N*-glycosylation patterns that are human-like or humanized. Such genetically modified lower eukaryotes can be achieved by eliminating selected endogenous glycosylation enzymes that are involved in producing high mannose *N*-glycans and introducing various combinations of exogenous enzymes involved in making complex *N*-glycans. Methods for genetically engineering yeast to produce complex *N*-glycans has been described in U.S. Patent No. 7,029,872 and U.S. Published patent Application Nos. 2004/0018590, 2005/0170452, 2006/0286637, 2004/0230042, 2005/0208617, 2004/0171826, 2005/0208617, and 2006/0160179. For example, a host cell can be selected or engineered to be depleted in 1,6-mannosyl transferase activities, which would otherwise add mannose residues onto the *N*-glycan on a glycoprotein, and then further engineered to include each of the enzymes involved in producing complex, human-like *N*-glycans.

Animal and human cells have a fucosyltransferase pathway that adds a fucose residue to the GlcNAc residue at the reducing end the *N*-glycans on a protein. The fucosylation pathway in humans consists of a GDP-mannose dehydratase and GDP-keto-deoxy-mannose-epimerase/GDP-keto-deoxy-galactose-reductase (FX protein), both located in the cytoplasm, which in concert converts GDP-mannose to GDP-fucose; a GDP-fucose transporter located in the membrane of the Golgi apparatus, which transports the GDP-fucose into the Golgi apparatus; and a fucosyltransferase (Fut8), which transfers the fucose residue by means of an 1,6-linkage to the 6 position of the GlcNAc residue at the reducing end of the *N*-glycan. In contrast to higher eukaryotes, many lower eukaryotes, for example yeast, lack the enzymes involved in the fucosyltransferase pathway, produce glycoproteins that do not contain fucose (*See* for example, Bretthauer/Catellino, Biotechnol. Appl. Biochem. 30: 193-200 (1999); Rabina *et al.*, Anal. Biochem. 286: 173-178 (2000)). However, the lack of fucose on glycoproteins has been shown to have advantages in certain cases. For example, in the production of monoclonal antibodies, immunoglobulin molecules, and related molecules, it has been shown that removal of the fucose sugar from the *N*-glycan of immunoglobulins increases or alters its binding to selected Ig receptors, which effects changes in properties such as antibody-dependent cellular cytotoxicity, or ADCC. (*See*, for example, U.S. Published Patent Application Nos. 2005/0276805 and US2003/0157108)

However, while removal of fucose from the *N*-glycans of immunoglobulins appears to enhance ADCC activity of the immunoglobulins, fucosylated *N*-glycans appear to be important for other glycoproteins. For example, the deletion of the fucosyltransferase gene in mice induces severe growth retardation, early death during post-natal development, and

emphysema-like changes in the lung. These *Fut8*$^{-/-}$ null mice were rescued from the emphysema-like phenotype by administration of exogenous TGF-beta1. Additionally, impaired receptor-mediated signaling was rescued by reintroduction of the Fut8 gene, showing that core fucosylation is crucial for proper functioning of growth factor receptors such as TGF-beta1 and EGF (Wang *et al.*, Meth. Enzymol. 417: 11-22 (2006). In lung tissue derived from *Fut8*$^{-/-}$ mice, the loss of core fucosylation impairs the function of low-density lipoprotein (LDL) receptor-related protein-1 (LRP-1), resulting in a reduction in the endocytosis of insulin like growth factor (IGF)-binding protein-3 (IGFBP-3) (Lee *et al.*, J. Biochem. (Tokyo) 139: 391-8 (2006). In *Fut8*$^{-/-}$ mouse embryonic fibroblast cells, α3β1 integrin-mediated cell migration is abolished and cell signaling is decreased, identifying the core fucose as essential for protein function (Zhao *et al.*, J. Biol. Chem., 281: 38343-38350 (2006). In addition, there may be situations where it is desirable to produce antibody compositions where at least a portion of the antibodies in the compositions are fucosylated in order to decrease ADCC activity. Therefore, in particular cases it will be advantageous to provide lower eukaryotic organisms and cells capable of producing fucosylated glycopeptides. Accordingly, development of methods and materials for the production of lower eukaryotic host cells, such as fungi and yeast, and particularly yeasts such as *Pichia pastoris*, *K. lactis*, and others, would facilitate development of genetically enhanced yeast strains for the recombinant production of fucosylated glycoproteins.

BRIEF SUMMARY OF THE INVENTION

Accordingly, the present invention provides methods and materials for making lower eukaryotic expression systems that can be used to produce recombinant, fucosylated glycoproteins. In particular, provided are vectors containing genes encoding one or more of the enzymes involved in the mammalian fucosylation pathway and lower eukaryote host cells that have been transformed with the vectors to produce host cells that are capable of producing fucosylated glycoproteins. The vectors, host cells, and methods are particularly well adapted to use in expression systems based on yeast and fungal host cells, such as *Pichia pastoris*.

In one embodiment, the present invention provides methods and materials for transforming lower eukaryotic host cells with one or more vectors encoding the enzymatic activities for conversion of GDP-mannose into GDP-fucose and for attachment of fucose to an *N*-glycan produced by the host cell. In further embodiments, the present invention comprises hybrid vectors encoding a fusion protein comprising the catalytic domain of a fucosylation pathway enzyme fused to a non-native leader sequence, which encodes a targeting sequence that targets the fusion peptide to the appropriate location in the endoplasmic reticulum, the early Golgi apparatus, or the late Golgi apparatus. For example, the catalytic domain for the fucosyltransferase is fused to a leader peptide that targets the catalytic domain to a location within the endoplasmic reticulum, the early Golgi apparatus, or the late Golgi apparatus. In

further embodiments, the lower eukaryote host cell is transformed with a vector encoding a GDP-fucose transferase which transports GDP-fucose from the cytoplasm to the interior of the Golgi.

The present invention provides a recombinant lower eukaryote host cell comprising a fucosylation pathway. In particular aspects, the host cell is yeast or filamentous fungus, for example, a yeast of the *Pichia* sp. such as *Pichia pastoris*.

In further aspects, the host cell further does not display α1,6-mannosyltransferase activity with respect to the *N*-glycan on a glycoprotein and includes an α1,2-mannosidase catalytic domain fused to a cellular targeting signal peptide not normally associated with the catalytic domain and selected to target α1,2-mannosidase activity to the ER or Golgi apparatus of the host cell whereby, upon passage of a recombinant glycoprotein through the ER or Golgi apparatus of the host cell, a recombinant glycoprotein comprising a fucosylated $Man_5GlcNAc_2$ glycoform is produced.

In further aspects, the above host cell further includes a GlcNAc transferase I catalytic domain fused to a cellular targeting signal peptide not normally associated with the catalytic domain and selected to target GlcNAc transferase I activity to the ER or Golgi apparatus of the host cell; whereby, upon passage of the recombinant glycoprotein through the ER or Golgi apparatus of the host cell, a recombinant glycoprotein comprising a fucosylated $GlcNAcMan_5GlcNAc_2$ glycoform is produced.

In further aspects, the above host cell further includes a mannosidase II catalytic domain fused to a cellular targeting signal peptide not normally associated with the catalytic domain and selected to target mannosidase II activity to the ER or Golgi apparatus of the host cell; whereby, upon passage of the recombinant glycoprotein through the ER or Golgi apparatus of the host cell, a recombinant glycoprotein comprising a fucosylated $GlcNAcMan_3GlcNAc_2$ glycoform is produced.

In further aspects, the above host cell further includes a GlcNAc transferase II catalytic domain fused to a cellular targeting signal peptide not normally associated with the catalytic domain and selected to target GlcNAc transferase II activity to the ER or Golgi apparatus of the host cell; whereby, upon passage of the recombinant glycoprotein through the ER or Golgi apparatus of the host cell, a recombinant glycoprotein comprising a fucosylated $GlcNAc_2Man_3GlcNAc_2$ glycoform is produced.

In further aspects, the above host cell further includes a Galactose transferase II catalytic domain fused to a cellular targeting signal peptide not normally associated with the catalytic domain and selected to target Galactose transferase II activity to the ER or Golgi apparatus of the host cell; whereby, upon passage of the recombinant glycoprotein through the ER or Golgi apparatus of the host cell, a recombinant glycoprotein comprising a fucosylated $Gal_2GlcNAc_2Man_3GlcNAc_2$ glycoform is produced.

In further aspects, the above host cell further includes a sialytransferase catalytic domain fused to a cellular targeting signal peptide not normally associated with the catalytic domain and selected to target sialytransferase activity to the ER or Golgi apparatus of the host cell; whereby, upon passage of the recombinant glycoprotein through the ER or Golgi apparatus of the host cell, a recombinant glycoprotein comprising a fucosylated $NANA_2Gal_2GlcNAc_2Man_3GlcNAc_2$ glycoform is produced.

Transforming the above host cells with a nucleic acid encoding a particular glycoprotein, compositions of the glycoprotein can be produced that comprise a plurality of glycoforms, each glycoform comprising at least one *N*-glycan attached thereto, wherein the glycoprotein composition thereby comprises a plurality of *N*-glycans in which a predominant glycoform comprises a desired fucosylated *N*-glycan. Depending upon the specific glycoprotein desired, the methods of the present invention can be used to obtain glycoprotein compositions in which the predominant *N*-glycoform is present in an amount between 5 and 80 mole percent greater than the next most predominant *N*-glycoform; in further embodiments, the predominant *N*-glycoform may be present in an amount between 10 and 40 mole percent; 20 and 50 mole percent; 30 and 60 mole percent; 40 and 70 mole percent; 50 and 80 mole percent greater than the next most predominant *N*-glycoform. In other embodiments, the predominant *N*-glycoform is a desired fucosylated *N*-glycoform and is present in an amount of greater than 25 mole percent; greater than 35 mole percent; greater than 50 mole percent; greater than 60 mole percent; or greater than 75 mole percent of the total number of *N*-glycans.

Thus, are provided host cells for producing glycoprotein compositions comprising a plurality of glycoforms, each glycoform comprising at least one *N*-glycan attached thereto, wherein the glycoprotein composition thereby comprises a plurality of fucosylated *N*-glycans in which the predominant *N*-glycan is selected from the group consisting of $Man_5GlcNAc_2$, $GlcNAcMan_5GlcNAc_2$, $Man_3GlcNAc_2$, $GlcNAcMan_3GlcNAc_2$, $GlcNAc_2Man_3GlcNAc_2$, $GalGlcNAc_2Man_3GlcNAc_2$, $Gal_2GlcNAc_2Man_3GlcNAc_2$, $NANAGal_2GlcNAc_2Man_3GlcNAc_2$, and $NANA_2Gal_2GlcNAc_2Man_3GlcNAc_2$.

In further aspects, greater than 25 mole percent of the plurality of fucosylated *N*-glycans consists essentially of a fucosylated glycoform in which the glycoform is selected from the group consisting of $Man_5GlcNAc_2$, $GlcNAcMan_5GlcNAc_2$, $Man_3GlcNAc_2$, $GlcNAcMan_3GlcNAc_2$, $GlcNAc_2Man_3GlcNAc_2$, $GalGlcNAc_2Man_3GlcNAc_2$, $Gal_2GlcNAc_2Man_3GlcNAc_2$, $NANAGal_2GlcNAc_2Man_3GlcNAc_2$, and $NANA_2Gal_2GlcNAc_2Man_3GlcNAc_2$.

In further still aspects, greater than 25 mole percent; greater than 35 mole percent; greater than 50 mole percent; greater than 60 mole percent; greater than 75 mole percent; or greater than 90 mole percent of the plurality of *N*-glycans consists essentially of a fucosylated glycoform in which the glycoform is selected from the group consisting of $Man_5GlcNAc_2$,

$GlcNAcMan_5GlcNAc_2$, $Man_3GlcNAc_2$, $GlcNAcMan_3GlcNAc_2$, $GlcNAc_2Man_3GlcNAc_2$, $GalGlcNAc_2Man_3GlcNAc_2$, $Gal_2GlcNAc_2Man_3GlcNAc_2$, $NANAGal_2GlcNAc_2Man_3GlcNAc_2$, and $NANA_2Gal_2GlcNAc_2Man_3GlcNAc_2$.

In the above glycoprotein composition, the fucose is in an α1,3-linkage with the GlcNAc at the reducing end of the *N*-glycan, an α1,6-linkage with the GlcNAc at the reducing end of the *N*-glycan, an α1,2-linkage with the Gal at the non-reducing end of the *N*-glycan, an α1,3-linkage with the GlcNac at the non-reducing end of the *N*-glycan, or an α1,4-linkage with a GlcNAc at the non-reducing end of the *N*-glycan.

Therefore, in particular aspects of the above the glycoprotein compositions, the glycoform is in an α1,3-linkage or α1,6-linkage fucose to produce a glycoform selected from the group consisting of $Man_5GlcNAc_2(Fuc)$, $GlcNAcMan_5GlcNAc_2(Fuc)$, $Man_3GlcNAc_2(Fuc)$, $GlcNAcMan_3GlcNAc_2(Fuc)$, $GlcNAc_2Man_3GlcNAc_2(Fuc)$, $GalGlcNAc_2Man_3GlcNAc_2(Fuc)$, $Gal_2GlcNAc_2Man_3GlcNAc_2(Fuc)$, $NANAGal_2GlcNAc_2Man_3GlcNAc_2(Fuc)$, and $NANA_2Gal_2GlcNAc_2Man_3GlcNAc_2(Fuc)$; in an α1,3-linkage or α1,4-linkage fucose to produce a glycoform selected from the group consisting of $GlcNAc(Fuc)Man_5GlcNAc_2$, $GlcNAc(Fuc)Man_3GlcNAc_2$, $GlcNAc_2(Fuc_{1-2})Man_3GlcNAc_2$, $GalGlcNAc_2(Fuc_{1-2})Man_3GlcNAc_2$, $Gal_2GlcNAc_2(Fuc_{1-2})Man_3GlcNAc_2$, $NANAGal_2GlcNAc_2(Fuc_{1-2})Man_3GlcNAc_2$, and $NANA_2Gal_2GlcNAc_2(Fuc_{1-2})Man_3GlcNAc_2$; or in an α1,2-linkage fucose to produce a glycoform selected from the group consisting of $Gal(Fuc)GlcNAc_2Man_3GlcNAc_2$, $Gal_2(Fuc_{1-2})GlcNAc_2Man_3GlcNAc_2$, $NANAGal_2(Fuc_{1-2})GlcNAc_2Man_3GlcNAc_2$, and $NANA_2Gal_2(Fuc_{1-2})GlcNAc_2Man_3GlcNAc_2$.

In other aspects, the glycoprotein composition of the present invention comprise compositions wherein the above *N*-glycoform is present at a level from about 5 to 80 mole percent; 10 to 40 mole percent; 20 to 50 mole percent; 30 to 60 mole percent; 40 to 70 mole percent; or 50 to 80 mole percent greater than the next most predominant *N*-glycoform.

Definitions

As used herein, the terms "*N*-glycan" and "glycoform" are used interchangeably and refer to an *N*-linked oligosaccharide, for example, one that is attached by an asparagine-*N*-acetylglucosamine linkage to an asparagine residue of a polypeptide. *N*-linked glycoproteins contain an *N*-acetylglucosamine residue linked to the amide nitrogen of an asparagine residue in the protein. The predominant sugars found on glycoproteins are glucose (Glc), galactose (Gal), mannose (Man), fucose (Fuc), *N*-acetylgalactosamine (GalNAc), *N*-acetylglucosamine (GlcNAc) and sialic acid (for example, *N*-acetyl-neuraminic acid (NANA)). The processing of the sugar groups occurs co-translationally in the lumen of the ER and continues in the Golgi apparatus for *N*-linked glycoproteins.

N-glycans have a common pentasaccharide core of $Man_3GlcNAc_2$. *N*-glycans differ with respect to the number of branches (antennae) comprising peripheral sugars (for example, GlcNAc, galactose, fucose, and sialic acid) that are added to the $Man_3GlcNAc_2$ core structure which is also referred to as the "trimannose core", the "pentasaccharide core", or the "paucimannose core". *N*-glycans are classified according to their branched constituents (for example, high mannose, complex or hybrid). A "high mannose" type *N*-glycan has five or more mannose residues. A "complex" type *N*-glycan typically has at least one GlcNAc attached to the 1,3 mannose arm and at least one GlcNAc attached to the 1,6 mannose arm of a "trimannose" core. Complex *N*-glycans may also have galactose or *N*-acetylgalactosamine residues that are optionally modified with sialic acid or derivatives (for example, "NANA" or "NeuAc", where "Neu" refers to neuraminic acid and "Ac" refers to acetyl). Complex *N*-glycans may also have intrachain substitutions comprising "bisecting" GlcNAc and core fucose ("Fuc"). As an example, when a *N*-glycan comprises a bisecting GlcNAc on the trimannose core, the structure can be represented as $Man_3GlcNAc_2(GlcNAc)$ or $Man_3GlcNAc_3$. When an *N*-glycan comprises a core fucose attached to the trimannose core, the structure may be represented as $Man_3GlcNAc_2(Fuc)$. Complex *N*-glycans may also have multiple antennae on the "trimannose core," often referred to as "multiple antennary glycans." A "hybrid" *N*-glycan has at least one GlcNAc on the terminal of the 1,3 mannose arm of the trimannose core and zero or more mannoses on the 1,6 mannose arm of the trimannose core. The various *N*-glycans are also referred to as "glycoforms."

Abbreviations used herein are of common usage in the art, see, for example, abbreviations of sugars, above. Other common abbreviations include "PNGase", or "glycanase" or "glucosidase" which all refer to peptide *N*-glycosidase F (EC 3.2.2.18).

The term "expression control sequence" as used herein refers to polynucleotide sequences that are necessary to affect the expression of coding sequences to which they are operatively linked. Expression control sequences are sequences that control the transcription, post-transcriptional events, and translation of nucleic acid sequences. Expression control sequences include appropriate transcription initiation, termination, promoter, and enhancer sequences; efficient RNA processing signals such as splicing and polyadenylation signals; sequences that stabilize cytoplasmic mRNA; sequences that enhance translation efficiency (for example, ribosome binding sites); sequences that enhance protein stability; and when desired, sequences that enhance protein secretion. The nature of such control sequences differs depending upon the host organism; in prokaryotes, such control sequences generally include promoter, ribosomal binding site, and transcription termination sequence. The term "control sequences" is intended to include, at a minimum, all components whose presence is necessary for expression, and can also include additional components whose presence is advantageous, for example, leader sequences and fusion partner sequences.

The term "recombinant host cell" ("expression host cell", "expression host system", "expression system" or simply "host cell"), as used herein, is intended to refer to a cell into which a recombinant vector has been introduced. It should be understood that such terms are intended to refer not only to the particular subject cell but to the progeny of such a cell. Because certain modifications may occur in succeeding generations due to either mutation or environmental influences, such progeny may not, in fact, be identical to the parent cell, but are still included within the scope of the term "host cell" as used herein. A recombinant host cell may be an isolated cell or cell line grown in culture or may be a cell which resides in a living tissue or organism.

The term "eukaryotic" refers to a nucleated cell or organism, and includes insect cells, plant cells, mammalian cells, animal cells, and lower eukaryotic cells.

The term "lower eukaryotic cells" includes yeast, fungi, collar-flagellates, microsporidia, alveolates (for example, dinoflagellates), stramenopiles (for example, brown algae, protozoa), rhodophyta (for example, red algae), plants (for example, green algae, plant cells, moss) and other protists. Yeast and fungi include, but are not limited to: *Pichia pastoris, Pichia finlandica, Pichia trehalophila, Pichia koclamae, Pichia membranaefaciens, Pichia minuta (Ogataea minuta, Pichia lindneri), Pichia opuntiae, Pichia thermotolerans, Pichia salictaria, Pichia guercuum, Pichia pijperi, Pichia stiptis, Pichia methanolica, Pichia* sp., *Saccharomyces cerevisiae, Saccharomyces* sp., *Hansenula polymorpha, Kluyveromyces sp., Kluyveromyces lactis, Candida albicans, Aspergillus nidulans, Aspergillus niger, Aspergillus oryzae, Trichoderma reesei, Chrysosporium lucknowense, Fusarium* sp., *Fusarium gramineum, Fusarium venenatum, Physcomitrella patens* and *Neurospora crassa. Pichia sp.,* any *Saccharomyces sp., Hansenula polymorpha,* any *Kluyveromyces sp., Candida albicans,* any *Aspergillus sp., Trichoderma reesei, Chrysosporium lucknowense,* any *Fusarium sp.,* and *Neurospora crassa.*

The term "peptide" as used herein refers to a short polypeptide, for example, one that is typically less than about 50 amino acids long and more typically less than about 30 amino acids long. The term as used herein encompasses analogs and mimetics that mimic structural and thus biological function.

As used herein, the term "predominantly" or variations such as "the predominant" or "which is predominant" will be understood to mean the glycan species that has the highest mole percent (%) of total *N*-glycans after the glycoprotein has been treated with PNGase and released glycans analyzed by mass spectroscopy, for example, MALDI-TOF MS. In other words, the phrase "predominantly" is defined as an individual entity, such as a specific glycoform, is present in greater mole percent than any other individual entity. For example, if a composition consists of species A in 40 mole percent, species B in 35 mole percent and species C in 25 mole percent, the composition comprises predominantly species A.

Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention pertains. Exemplary methods and materials are described below, although methods and materials similar or equivalent to those described herein can also be used in the practice of the present invention and will be apparent to those of skill in the art. All publications and other references mentioned herein are incorporated by reference in their entirety. In case of conflict, the present specification, including definitions, will control. The materials, methods, and examples are illustrative only and not intended to be limiting.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 illustrates the fucosylation pathway present in many higher eukaryotic cells.

Figure 2 shows the glyco-engineering steps required to produce a recombinant yeast capable of producing fucosylated glycoproteins. Endogenous GDP-Mannose, present in the yeast cytoplasm, is converted to GDP-Fucose by GDP-mannose-dehydratase (GMD) and the bifunctional enzyme FX. Subsequently, the product is translocated into the Golgi apparatus by the GDP-Fucose transporter (GFTr) and fucose is transferred onto the acceptor glycan by α-1,6-fucosyltransferase (*FUT8*). Enzymes are indicated by blue text and metabolic intermediates by black text. GDP-kdMan (GDP-4-keto-6-deoxy-mannose) and GDP-kdGal (GDP-4-keto-6-deoxy-galactose) are intermediates in the conversion of GDP-mannose to GDP-fucose.

Figure 3A shows the vectors used in engineering yeast strains to produce fucosylated glycoproteins. Represented is the expression vector pSH995 into which the fucose biosynthetic and transfer genes are introduced. Introduction of the genes required for biosynthesis and transfer of fucose into pSH995 produced the vector pSH1022.

Figure 3B shows the vector pSH1022. Shown in (B) are the flanking regions of the *TRP2* loci used to integrate the genes into the *Pichia* genome; the dominant selection marker NATr; the GAPDH-CYC expression cassette; and the pUC19 plasmid backbone.

Figure 4A shows a MALDI-TOF scan of the *N*-glycans released from the rat EPO demonstrating that *Pichia pastoris* strain YSH661 (strain RDP974 transformed with vector pSH1022 containing the fucosylation pathway genes) produced rEPO comprising $Gal_2GlcNAc_2Man_3GlcNAc_2(Fuc)$ and $Gal_2GlcNAc_2Man_3GlcNAc_2$ *N*-glycans. The $Gal_2GlcNAc_2Man_3GlcNAc_2(Fuc)$ *N*-glycans are within the box.

Figure 4B shows a MALDI-TOF scan of the *N*-glycans released from the rat EPO control strain YSH660 (strain RDP974 transformed with control vector pSH995) produced afucosylated or fucose-free $Gal_2GlcNAc_2Man_3GlcNAc_2$ *N*-glycans only.

DETAILED DESCRIPTION OF THE INVENTION

The present invention provides methods and materials for the genetically engineering host cells capable of producing glycoproteins proteins that have fucosylated *N*-glycans. While the methods and materials are exemplified in the yeast *Pichia pastoris*, which does not possess an endogenous fucosylation pathway, the methods and materials may also be used to genetically engineer other lower eukaryotes such as fungi, prokaryote, and those higher eukaryotes, which do not have an endogenous fucosylation pathways, for example, insect cells. In other embodiments, the methods and materials may used to genetically engineer higher eukaryote cells that have an endogenous fucosylation pathway but where it is desirable to increase the amount of fucosylation present in glycoproteins produced by such host cells.

In general, the method of the present invention involves producing a host cell capable of producing fucosylated glycoproteins by introducing into the host cell nucleic acids encoding those enzymes or enzymatic activities involved in the fucosylation pathway that when introduced into the host cell will render the cell capable of producing fucosylated glycoproteins. These nucleic acids include, for example, nucleic acids encoding a GDP-mannose-4,6-dehydratase activity, a GDP-keto-deoxy-mannose-epimerase activity/GDP-keto-deoxy-galactose-reductase activity, a GDP-fucose transporter protein, and a fucosyltransferase activity. An overview of the fucosylation pathway in higher eukaryotes is shown in Figure 1.

GDP-mannose-4,6-dehydratase (GMD) (EC 4.2.1.47) converts GDP-mannose to GDP-4-keto-6-deoxy-mannose in the presence of NAD has been identified in a number of species. The human GMP (hGMD) is encoded by the nucleotide sequence shown in SEQ ID NO: 1 and has the amino acid sequence shown in SEQ ID No: 2. Homologous genes with GDP-mannose-dehydratase activity include the porcine GMD, (Broschat *et al.*, Eur. J. Biochem., 153(2):397-401 (1985)),*Caenorhabditis elegans* GMD and *Drosophila melanogaster* GMD (*See*, for example, Rhomberg *et al.*, FEBS J.; 273:2244-56 (2006)); *Arabidopsis thaliana*; (*See*, e.g., Nakayama *et al.*, Glycobiology; 13:673-80 (2003)); and *E. coli*, (Somoza *et al.*,Structure, 8:123-35 (2000)).

GDP-keto-deoxy-mannose-epimerase/GDP-keto-deoxy-galactose-reductase (GDP-L-fucose synthase, EC 1.1.1.271) is a bifunctional enzyme, which has been identified in both eukaryotes and prokaryotes. The human GDP-keto-deoxy-mannose-epimerase/GDP-keto-deoxy-galactose-reductase is called the FX protein (also known as hFX or GER). The nucleotide sequence encoding the hFX is shown in SEQ ID NO: 3. The hFX protein has the amino acid sequence shown in SEQ ID NO: 4.

The GDP-fucose transporter has been identified in several species. The human GDP-fucose transporter (hGFTr) has been identified as related to congenital disorders of glycosylation-II (CDG-II) (Lubke *et al.*, Nat. Genet. 28: 73-6 (2001)). Also known as Leukocyte Adhesion Deficiency II (LAD II), it appears that the disorder results from a disturbance in fucosylation of selectin ligands. Roos and Law, Blood Cells Mol. Dis. 27: 1000-4 (2001). The

nucleotide sequence encoding the hGFTr is shown in SEQ ID NO: 5 and the amino acid sequence of the hGFTr is shown in SEQ ID NO: 6. Homologous genes with GDP-fucose transporter activity have been identified in other species, such as *Drosophila melanogaster* (Ishikawa *et al.*, Proc. Natl. Acad. Sci. U S A. 102:18532-7 (2005)), rat liver (Puglielli and Hirschberg; J. Biol. Chem. 274:35596-60 (1999)), and a putative CHO homolog (Chen *et al.*, Glycobiology;15:259-69 (2005)).

A number of fucosyltransferases have been identified (See Breton *et al.*, Glycobiol. 8: 87-94 (1997); Becker , Lowe, Glycobiol. 13: 41R-53R (2003); Ma *et al.*, Glycobiol. 16: 158R-184R (2006)), for example, α1,2-fucosyltransferase (EC 2.4.1.69; encoded by *FUT1* and *FUT2*), α1,3-fucosyltransferase (glycoprotein 3-α-L-fucosyltransferase, EC 2.4.1.214; encoded by *FUT3-FUT7* and *FUT9*), α1,4-fucosyltransferase (EC 2.4.1.65; encoded by *FUT3*), and α1,6-fucosyltransferase (glycoprotein 6-α-L-fucosyltransferase, EC 2.4.1.68; encoded by *FUT8*). In general, α1,2-fucosyltransferase transfer fucose to the terminal galactose residue in an *N*-glycan by way of an α1,2 linkage. In general, the α1,3-fucosyltransferase and α1,4-fucosyltransferases transfer fucose to a GlcNAc residue at the non-reducing end of the *N*-glycan.

In general, α1,6-fucosyltransferases transfer fucose by way of an α1,6-linkage to the GlcNAc residue at the reducing end of *N*-glycans (asparagine-linked GlcNAc). Typically, α1,6-fucosyltransferase requires a terminal GlcNAc residue at the non-reducing end of at least one branch of the trimannose core to be able to add fucose to the GlcNAc at the reducing end. However, an α1,6-fucosyltransferase has been identified that requires a terminal galactoside residue at the non-reducing end to be able add fucose to the GlcNAc at the reducing end (Wilson *et al.*, Biochm. Biophys. Res. Comm. 72: 909-916 (1976)) and Lin *et al.* (Glycobiol. 4: 895-901 (1994)) has shown that in Chinese hamster ovary cells deficient for GlcNAc transferase I, the α1,6-fucosyltansferase will fucosylate $Man_4GlcNAc_2$ and $Man_5GlcNAc_2$ *N*-glycans. Similarly, α1,3-fucosyltransferase transfers to the GlcNAc residue at the reducing end of *N*-glycans but by way of an α1,3-linkage, generally with a specificity for *N*-glycans with one unsubstituted non-reducing terminal GlcNAc residue. The *N*-glycan products of this enzyme are present in plants, insects, and some other invertebrates (for example, *Schistosoma, Haemonchus, Lymnaea*). However, U.S. Patent No. 7,094,530 describes an α1,3-fucosyltransferase isolated from human monocytic cell line THP-1.

The human α1,6-fucosyltransferase (h*FUT8*) has been identified by Yamaguchi *et al.*, (Cytogenet. Cell. Genet. 84: 58-6 (1999)). The nucleotide sequence encoding the human *FUT8* is shown in SEQ ID NO:7. The amino acid sequence of the h*FUT8* is shown in SEQ ID NO: 8. Homologous genes with *FUT8* activity have been identified in other species, such as a rat *FUT8* (r*FUT8*) having the amino acid sequence shown in SEQ ID NO:10 and encoded by the nucleotide sequence shown in SEQ ID NO: 9; a mouse Fut8 (m*FUT8*) having the amino acid

sequence shown in SEQ ID NO:12 and encoded by the nucleotide sequence shown in SEQ ID NO:11, and a porcine *FUT8* (p*FUT8*) having amino acid sequence shown in SEQ ID NO:14 and encoded by the nucleotide sequence shown in SEQ ID NO:13. *FUT8* has also been identified in CHO cells (Yamane-Ohnuki *et al.*, Biotechnol. Bioeng. 87: 614-622 (2004)), monkey kidney COS cells (Clarke and Watkins, Glycobiol. 9: 191-202 (1999)), and chicken cells (Coullin *et al.*, Cytogenet. Genome Res. 7: 234-238 (2002)). Paschinger *et al.*, Glycobiol. 15: 463-474 (2005) describes the cloning and characterization of fucosyltransferases from *C. elegans* and *D. melanogaster*. *Ciona intestinalis*, *Drosophila pseudoobscura*, *Xenopus laevis*, and *Danio rerio* putative α1,6-fucosyltransferases have been identified (GenBank accession numbers AJ515151, AJ830720, AJ514872, and AJ781407, respectively).

The aforementioned fucosylation pathway enzymes or activities are encoded by nucleic acids. The nucleic acids can be DNA or RNA, but typically the nucleic acids are DNA because it preferable that the nucleic acids encoding the fucosylation pathway enzymes or activities are stably integrated into the genome of the host cells. The nucleic acids encoding the fucosylation pathway enzymes or activities are each operably linked to regulatory sequences that allow expression of the fucosylation pathway enzymes or activities. Such regulatory sequences include a promoter and optionally an enhancer upstream of the nucleic acid encoding the fucosylation pathway enzyme or activity and a transcription termination site downstream of the fucosylation pathway enzyme or activity. The nucleic acid also typically further includes a 5' untranslated region having a ribosome binding site and a 3' untranslated region having a polyadenylation site. The nucleic acid is often a component of a vector such as a plasmid, which is replicable in cells in which the fucosylation pathway enzyme or activity is expressed. The vector can also contain a marker to allow selection of cells transformed with the vector. However, some cell types, in particular yeast, can be successfully transformed with a nucleic acid that lacks vector sequences.

In general, the host cells transformed with the nucleic acids encoding the one or more fucosylation pathway enzymes or activities further includes one or more nucleic acids encoding desired glycoproteins. Like for the fucosylation pathway enzymes, the nucleic acids encoding the glycoproteins are operably linked to regulatory sequences that allow expression of the glycoproteins. The nucleic acids encoding the glycoproteins can be amplified from cell lines known to express the glycoprotein using primers to conserved regions of the glycoprotein (*See*, for example, Marks *et al.*, J. Mol. Biol. : 581-596 (1991)). Nucleic acids can also be synthesized *de novo* based on sequences in the scientific literature. Nucleic acids can also be synthesized by extension of overlapping oligonucleotides spanning a desired sequence (*See*, for example, Caldas *et al.*, Protein Engineering, 13: 353-360 (2000)).

The type of fucosylated N-glycan structure produced by the host cell will depend on the glycosylation pathway in the host cell and the particular fucosyltransferase. For example,

α1,2-fucosyltransferases in general add a fucose to the terminal galactose on an N-glycan. As such, a pathway that utilizes an α1,2-fucosyltransferase would preferably be introduced into a host cell that is capable of producing N-glycans having a $Gal_2GlcNAc_2Man_3GlcNAc_2$ glycoform. The N-glycans produced will have fucose in an α1,2 linkage to the terminal galactose residues. Both α1,3-fucosyltransferases and α1,4-fucosyltransferases add fucose to one or more GlcNAc residues at or near the non-reducing end by way of an α1,3 or α1,4 linkage, respectively, or for some α1,3-fucosyltransferases, by way of an α1,3 linkage to the core GlcNAc linked to the asparagine residue of the glycoprotein. As such, a pathway that utilizes an α1,3/4-fucosyltransferase would preferably be introduced into a host cell that is capable of producing N-glycans having at least a $GlcNAcMan_5GlcNAc_2$ glycoform. Finally, α1,6-fucosyltransferases in general transfer fucose by way of an α1,6 linkage to the core GlcNAc linked to the asparagine residue of the glycoprotein. In general, a pathway that utilizes an α1,6-fucosyltransferase would preferably be introduced into a host cell that is capable of producing N-glycans having at least a $GlcNAcMan_5GlcNAc_2$, $Man_5GlcNAc_2$, or $Man_4GlcNAc_2$ glycoform.

The glycoproteins that can be produced in accordance using the methods disclosed herein include any desired protein for therapeutic or diagnostic purposes, regardless of the origin of the nucleic acid sequence for producing the glycoprotein. For example, monoclonal antibodies in which the *N*-glycan is not fucosylated have increased ADCC activity; however, increased ADCC activity is undesirable for monoclonal antibodies intended to bind receptor ligands as a treatment for a disorder but not elicit ADCC activity. Monoclonal antibodies produced in the host cells disclosed herein comprising the fucosylation pathway will have fucosylated *N*-glycans and will be expected to have decreased ADCC activity. As another example, immunoadhesions (*See*, U.S. Patent Nos. 5,428,130, 5,116,964, 5,514,582, and 5,455,165; Capon *et al.* Nature 337:525 (1989); Chamow and Ashkenazi, Trends Biotechnol. 14: 52-60 (1996); Ashkenazi and Chamow, Curr. Opin. Immunol. 9: 195-200 (1997)), which comprise the extracellular portion of a membrane-bound receptor fused to the Fc portion of an antibody produced in the host cells disclosed herein comprising the fucosylation pathway will have fucosylated *N*-glycans and will be expected to have decreased ADCC activity. Examples of glycoproteins that can be produced according to the methods herein to have fucosylated *N*-glycans include, but are not limited to, erythropoietin (EPO); cytokines such as interferon-α, interferon-β., interferon-γ, interferon-ω, and granulocyte-CSF; coagulation factors such as factor VIII, factor IX, and human protein C; monoclonal antibodies, soluble IgE receptor α-chain, IgG, IgM, IgG, urokinase, chymase, and urea trypsin inhibitor, IGF-binding protein, epidermal growth factor, growth hormone-releasing factor, annexin V fusion protein, angiostatin, vascular endothelial growth factor-2, myeloid progenitor inhibitory factor-1, osteoprotegerin tissue, plasminogen activator, G-CSF, GM-CSF, and TNF-receptor.

In particular embodiments, one or more of the nucleic acids encode fusion proteins comprising the catalytic domain of a fucosylation pathway protein fused to a targeting peptide, which targets the fusion protein to a particular region within the cell. Typically, the targeting peptide will target the fusion protein to a location within the secretory pathway. The term "secretory pathway" thus refers to organelles and components within the cell where glycoproteins are modified in preparation for secretion. The secretory pathway includes the endoplasmic reticulum (ER), the Golgi apparatus, the trans-Golgi network, and the secretory vesicles. For example, suitable cellular targeting peptides may target the catalytic domain to the ER, the Golgi apparatus, the trans-Golgi network, or secretory vesicles. Targeting peptides which may be useful in the present invention include those described in U.S. Patent No. 7,029,872. In one embodiment, the catalytic domain of the fucosyltransferase is fused to a targeting peptide that directs the fusion protein to the Golgi apparatus. The particular targeting peptide fused to the fucosyltransferase catalytic domain will depend on the host cell, the particular fucosyltransferase, and the glycoprotein being produced. Examples of targeting peptides that can be used for targeting the fucosyltransferase have been disclosed in, for example, U.S. Patent No. 7,029,872 and U.S. Published Patent Application Nos. 2004/0018590, 2004/0230042, 2005/0208617, 2004/0171826, 2006/0286637, and 2007/0037248.

The nucleic acids encoding the enzymes or activities involved in the fucosylation pathway are ligated into vectors, which are capable of being used to transfect host cells. Typically, the vectors will include regulatory elements, which have been isolated from the same species of cell as the intended host cell, or which have been isolated from other species, but which are known to be functional when inserted into the intended host cell. Typically, these regulatory elements include 5' regulatory sequences, such as promoters, as well as 3' regulatory sequences, such as transcription terminator sequences. Vectors will typically also include at least one selectable marker element that allows for selection of host cells that have been successfully transfected with the vector. The vectors are transferred into the intended host cells, and the resulting cells are screened for the presence of the selectable marker, to identify those host cells which have been successfully transfected with the vector, and which will therefore also carry the vector encoding the fusion protein.

Lower eukaryotes such as yeast are often preferred for expression of glycoproteins because they can be economically cultured, give high yields of protein, and when appropriately modified are capable of producing glycoproteins with particular predominant *N*-glycan structures.. Yeast, in particular, offers established genetics allowing for rapid transformations, tested protein localization strategies and facile gene knock-out techniques. Various yeasts, such as *K. lactis*, *Pichia pastoris*, *Pichia methanolica*, and *Hansenula polymorpha* are commonly used for cell culture and production of proteins because they are able to grow to high cell densities and secrete large quantities of recombinant protein at an industrial scale. Likewise, filamentous

fungi, such as *Aspergillus niger*, *Fusarium* sp, *Neurospora crassa* and others can be used to produce glycoproteins at an industrial scale.

Lower eukaryotes, particularly yeast, can be genetically modified so that they express glycoproteins in which the glycosylation pattern is complex or human-like or humanized. Such genetically modified lower eukaryotes can be achieved by eliminating selected endogenous glycosylation enzymes that are involved in producing high mannose *N*-glycans and introducing various combinations of exogenous enzymes involved in making complex *N*-glycans. Methods for genetically engineering yeast to produce complex *N*-glycans has been described in U.S. Patent No. 7,029,872 and U.S. Published patent Application Nos. 2004/0018590, 2005/0170452, 2006/0286637, 2004/0230042, 2005/0208617, 2004/0171826, 2005/0208617, and 2006/0160179. For example, a host cell is selected or engineered to be depleted in 1,6-mannosyl transferase activities, which would otherwise add mannose residues onto the *N*-glycan on a glycoprotein. For example, in yeast, the *OCH1* gene encodes 1,6-mannosyl transferase activity. The host cells is then further engineered to include one or more of the enzymes involved in producing complex, human-like *N*-glycans.

In one embodiment, the host cell further includes an α1,2-mannosidase catalytic domain fused to a cellular targeting signal peptide not normally associated with the catalytic domain and selected to target the α1,2-mannosidase activity to the ER or Golgi apparatus of the host cell. Passage of a recombinant glycoprotein through the ER or Golgi apparatus of the host cell produces a recombinant glycoprotein comprising a fucosylated $Man_5GlcNAc_2$ glycoform, for example a $Man_5GlcNAc_2(Fuc)$ glycoform. U.S. Patent No, 7,029,872 and U.S. Published Patent Application Nos. 2004/0018590 and 2005/0170452 disclose lower eukaryote host cells capable of producing a glycoprotein comprising a $Man_5GlcNAc_2$ glycoform.

In a further embodiment, the immediately preceding host cell further includes a GlcNAc transferase I (GnTI) catalytic domain fused to a cellular targeting signal peptide not normally associated with the catalytic domain and selected to target GlcNAc transferase I activity to the ER or Golgi apparatus of the host cell. Passage of the recombinant glycoprotein through the ER or Golgi apparatus of the host cell produces a recombinant glycoprotein comprising a fucosylated $GlcNAcMan_5GlcNAc_2$ glycoform, for example a $GlcNAcMan_5GlcNAc_2(Fuc)$ glycoform. U.S. Patent No, 7,029,872 and U.S. Published Patent Application Nos. 2004/0018590 and 2005/0170452 disclose lower eukaryote host cells capable of producing a glycoprotein comprising a $GlcNAcMan_5GlcNAc_2$ glycoform. The glycoprotein produced in the above cells can be treated *in vitro* with a hexaminidase to produce a recombinant glycoprotein comprising a fucosylated $Man_5GlcNAc_2(Fuc)$ glycoform.

In a further still embodiment, the immediately preceding host cell further includes a mannosidase II catalytic domain fused to a cellular targeting signal peptide not normally associated with the catalytic domain and selected to target mannosidase II activity to the ER or

Golgi apparatus of the host cell. Passage of the recombinant glycoprotein through the ER or Golgi apparatus of the host cell produces a recombinant glycoprotein comprising a fucosylated $GlcNAcMan_3GlcNAc_2$ glycoform, for example a $GlcNAcMan_3GlcNAc_2(Fuc)$ glycoform. U.S. Published Patent Application No. 2004/0230042 discloses lower eukaryote host cells that express mannosidase II enzymes and are capable of producing glycoproteins having predominantly a $GlcNAc_2Man_3GlcNAc_2$ glycoform. The glycoprotein produced in the above cells can be treated *in vitro* with a hexaminidase to produce a recombinant glycoprotein comprising a $Man_3GlcNAc_2(Fuc)$ glycoform.

In a further still embodiment, the immediately preceding host cell further includes GlcNAc transferase II (GnTII) catalytic domain fused to a cellular targeting signal peptide not normally associated with the catalytic domain and selected to target GlcNAc transferase II activity to the ER or Golgi apparatus of the host cell. Passage of the recombinant glycoprotein through the ER or Golgi apparatus of the host cell produces a recombinant glycoprotein comprising a fucosylated $GlcNAc_2Man_3GlcNAc_2$ glycoform, for example a $GlcNAc_2Man_3GlcNAc_2(Fuc)$ glycoform. U.S. Patent No, 7,029,872 and U.S. Published Patent Application Nos. 2004/0018590 and 2005/0170452 disclose lower eukaryote host cells capable of producing a glycoprotein comprising a $GlcNAc_2Man_3GlcNAc_2$ glycoform. The glycoprotein produced in the above cells can be treated *in vitro* with a hexaminidase to produce a recombinant glycoprotein comprising a $Man_3GlcNAc_2(Fuc)$ glycoform.

In a further still embodiment, the immediately preceding host cell further includes a Galactose transferase II catalytic domain fused to a cellular targeting signal peptide not normally associated with the catalytic domain and selected to target Galactose transferase II activity to the ER or Golgi apparatus of the host cell. Passage of the recombinant glycoprotein through the ER or Golgi apparatus of the host cell produces a recombinant glycoprotein comprising a fucosylated $Gal_2GlcNAc_2Man_3GlcNAc_2$ glycoform, for example a $Gal_2GlcNAc_2Man_3GlcNAc_2(Fuc)$ glycoform. U.S. Published Patent Application No. 2006/0040353 discloses lower eukaryote host cells capable of producing a glycoprotein comprising a $Gal_2GlcNAc_2Man_3GlcNAc_2$ glycoform. The glycoprotein produced in the above cells can be treated *in vitro* with a galactosidase to produce a recombinant glycoprotein comprising a fucosylated $GlcNAc_2Man_3GlcNAc_2$ glycoform, for example a $GlcNAc_2Man_3GlcNAc_2(Fuc)$ glycoform.

In a further still embodiment, the immediately preceding host cell further includes a sialytransferase catalytic domain fused to a cellular targeting signal peptide not normally associated with the catalytic domain and selected to target sialytransferase activity to the ER or Golgi apparatus of the host cell. Passage of the recombinant glycoprotein through the ER or Golgi apparatus of the host cell produces a recombinant glycoprotein comprising a fucosylated $NANA_2Gal_2GlcNAc_2Man_3GlcNAc_2$ glycoform, for example, a

$NANA_2Gal_2GlcNAc_2Man_3GlcNAc_2(Fuc)$ glycoform. For lower eukaryote host cells such as yeast and filamentous fungi, it is preferred that the host cell further include a means for providing CMP-sialic acid for transfer to the *N*-glycan. U.S. Published Patent Application No. 2005/0260729 discloses a method for genetically engineering lower eukaryotes to have a CMP-sialic acid synthesis pathway and U.S. Published Patent Application No. 2006/0286637 discloses a method for genetically engineering lower eukaryotes to produce sialylated glycoproteins. The glycoprotein produced in the above cells can be treated *in vitro* with a neuraminidase to produce a recombinant glycoprotein comprising a fucosylated $Gal_2GlcNAc_2Man_3GlcNAc_2$ glycoform, for example, a $Gal_2GlcNAc_2Man_3GlcNAc_2(Fuc)$ glycoform.

Any one of the preceding host cells can further include one or more GlcNAc transferase selected from the group consisting of GnTIII, GnTIV, GnTV, GnTVI, and GnTIX to produce glycoproteins having bisected and/or multiantennary *N*-glycan structures such as disclosed in U.S. Published Patent Application Nos. 2004/074458 and 2007/0037248. Various of the preceding host cells further include one or more sugar transporters such as UDP-GlcNAc transporters (for example, *Kluyveromyces lactis* and *Mus musculus* UDP-GlcNAc transporters), UDP-galactose transporters (for example, *Drosophila melanogaster* UDP-galactose transporter), and CMP-sialic acid transporter (for example, human sialic acid transporter). Because lower eukaryote host cells such as yeast and filamentous fungi lack the above transporters, it is preferable that lower eukaryote host cells such as yeast and filamentous fungi be genetically engineered to include the above transporters.

In further embodiments of the above host cells, the host cells are further genetically engineered to eliminate glycoproteins having α-mannosidase-resistant *N*-glycans by deleting or disrupting the β-mannosyltransferase gene (*BMT2*)(*See*, U.S. Published Patent Application No. 2006/0211085) and glycoproteins having phosphomannose residues by deleting or disrupting one or both of the phosphomannosyl transferase genes *PNO1* and *MNN4B* (*See* for example, U.S. Published Patent Application Nos. 2006/0160179 and 2004/0014170). In further still embodiments of the above host cells, the host cells are further genetically modified to eliminate O-glycosylation of the glycoprotein by deleting or disrupting one or more of the Dol-P-Man:Protein (Ser/Thr) Mannosyl Transferase genes (*PMT*s) (See U.S. Patent No. 5,714,377).

It has been shown that codon optimization of genes or transcription units coding for particular polypeptides leads to increased expression of the encoded polypeptide, that is increased translation of the mRNA encoding the polypeptide. Therefore, in the case of the host cells disclosed herein, increased expression of the encoded enzymes will produce more of the encoded enzymes, which can lead to increased production of *N*-glycans that are fucosylated. In the context of codon optimization, the term "expression" and its variants refer to translation of the mRNA encoding the polypeptide and not to transcription of the polynucleotide encoding the

polypeptide. The term "gene" as used herein refers to both the genomic DNA or RNA encoding a polypeptide and to the cDNA encoding the polypeptide.

Codon optimization is a process that seeks to improve heterologous expression of a gene when that gene is moved into a foreign genetic environment that exhibits a different nucleotide codon usage from the gene's native genetic environment or improve ectopic expression of a gene in its native genetic environment when the gene naturally includes one or more nucleotide codons that are not usually used in genes native to the genetic environment that encode highly expressed genes. In other words, codon optimization involves replacing those nucleotide codons of a gene that are used at a relatively low frequency in a particular genetic environment or organism with nucleotide codons that are used in genes that are expressed at a higher frequency in the genetic environment or organism. In that way, the expression (translation) of the gene product (polypeptide) is increased. The assumption is that the nucleotide codons that appear with high frequency in highly expressed genes are more efficiently translated than nucleotide codons that appear at low frequency.

In general, methods for optimizing nucleotide codons for a particular gene depend on identifying the frequency of the nucleotide codons for each of the amino acids used in genes that are highly expressed in an organism and then replacing those nucleotide codons in a gene of interest that are used with low frequency in the highly expressed genes with nucleotide codons that are identified as being used in the highly expressed genes (*See* for example Lathe, Synthetic Oligonucleotide Probes Deduced from Amino Acid Sequence Data: Theoretical and Practical Considerations, J. Molec. Biol.: 183: 1-12 (1985); Nakamura *et al.*, Nuc. Acid Res. 28: 292 (2000); Fuglsang, Protein Expression & Purification 31: 247-249 (2003)). There are numerous computer programs that will automatically analyze the nucleotide codons of a nucleic acid of an organism encoding a gene and suggest nucleotide codons to replace nucleotide codons, which occur with low frequency in the organism, with nucleotide codons that are found in genes that are highly expressed in the organism.

The following examples are intended to promote a further understanding of the present invention.

EXAMPLE 1

This Example shows the construction of a *Pichia pastoris* strain capable of producing glycoproteins that include fucose in the *N*-glycan structure of the glycoprotein.

Escherichia coli strains TOP10 or XL10-Gold are used for recombinant DNA work. PNGase-F, restriction and modification enzymes are obtained from New England BioLabs (Beverly, MA), and used as directed by the manufacturer. α-1,6-Fucosidase is obtained from Sigma-Aldrich (St. Louis, MO) and used as recommended by the manufacturer. Oligonucleotides are obtained from Integrated DNA Technologies (Coralville, IA). Metal

chelating "HisBind" resin is obtained from Novagen (Madison, WI). 96-well lysate-clearing plates are from Promega (Madison, WI). Protein-binding 96-well plates are from Millipore (Bedford, MA). Salts and buffering agents are from Sigma-Aldrich (St. Louis, MO).

Amplification of fucosylation pathway genes.

An overview of the fucosylation pathway is shown in Figure 1. The open reading frame (ORF) of hGMD is amplified from human liver cDNA (BD Biosciences, Palo Alto, CA) using Advantage 2 polymerase following the procedure recommended by the manufacturer. Briefly, the primers SH415 and SH413 (5'- GGCGG CCGCC ACCAT GGCAC ACGCA CCGGC ACGCT GC-3' (SEQ ID NO:15) and 5'- TTAAT TAATC AGGCA TTGGG GTTTG TCCTC ATG-3' (SEQ ID NO:16), respectively) are used to amplify a 1,139 bp product from human liver cDNA using the following conditions: 97°C for 3 minutes; 35 cycles of 97°C for 30 seconds, 50°C for 30 seconds, 72°C for 2 minutes; and 72°C for 10 minutes. Subsequently the product is cloned into pCR2.1 (Invitrogen, Carlsbad, CA), sequenced, and the resultant construct designated pSH985.

Using the conditions outlined above, the primers SH414 and SH411 (5'-GGCGG CCGCC ACCAT GGGTG AACCC CAGGG ATCCA TG-3' (SEQ ID NO:17) and 5'-TTAAT TAATC ACTTC CGGGC CTGCT CGTAG TTG-3' (SEQ ID NO:18), respectively) are used to amplify a 986 bp fragment from human kidney cDNA (BD Biosciences, Palo Alto, CA), which corresponds to the ORF of the human FX gene. Subsequently, this fragment is cloned into pCR2.1, sequenced, and designated pSH988.

The ORF of the human GFTr is amplified from human spleen cDNA (BD Biosciences, Palo Alto, CA) using the conditions outlined above, and the primers RCD679 and RCD680 (5'-GCGGC CGCCA CCATG AATAG GGCCC CTCTG AAGCG G-3' (SEQ ID NO:19) and 5'-TTAAT TAATC ACACC CCCAT GGCGC TCTTC TC-3' (SEQ ID NO:20), respectively). The resultant 1,113 bp fragment is cloned into pCR2.1, sequenced, and designated pGLY2133.

A truncated form of the mouse *FUT8* ORF, encoding amino acids 32 to 575 and lacking the nucleotides encoding the endogenous transmembrane domain, is amplified from mouse brain cDNA (BD Biosciences, Palo Alto, CA) using the conditions outlined above and the primers SH420 and SH421 (5'- GCGGC GCGCC GATAA TGACC ACCCT GATCA CTCCA G-3' (SEQ ID NO:21) and 5'- CCTTA ATTAA CTATT TTTCA GCTTC AGGAT ATGTG GG-3' (SEQ ID NO:22), respectively). The resultant 1,654 bp fragment is cloned into pCR2.1, sequenced, and designated pSH987.

Generation of fucosylation genes in yeast expression cassettes.

Open reading frames for GMD, FX, and GFTr are generated by digesting the above vectors with *Not*I and *Pac*I restriction enzymes to produce DNA fragments with a *Not*I compatible 5' end and a *Pac*I compatible 3' end. The FUT8 fragment is generated by digesting with *Asc*I and *Pac*I restriction enzymes to produce a DNA with an *Asc*I compatible 5' end and a *Pac*I compatible 3' end.

To generate the GMD expression cassette, GMD is cloned into yeast expression vector pSH995, which contains a *P. pastoris* GAPDH promoter and *S. cerevisiae* CYC transcription terminator sequence and is designed to integrate into the *Pichia* genome downstream of the *Trp2* ORF, using the nourseothricin resistance marker. This vector is illustrated in Figure 3A. The vector pSH985 is digested with *Not*I and *Pac*I to excise a 1.1 Kb fragment containing the GMD ORF, which is then subcloned into pSH995 previously digested with the same enzymes. The resultant vector, containing GMD under the control of the GAPDH promoter, is designated pSH997. A.

To generate the FX expression cassette, the vector pSH988 is digested with *Not*I and *Pac*I to excise a 1.0 Kb fragment containing the FX ORF, which is treated with T4 DNA polymerase to remove single strand overhangs (J. Sambrook , D. W. Russell, Molecular Cloning: A laboratory Manual (Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, ed. 3rd, 2001)). Subsequently this fragment is subcloned into the vector pGLY359 (Hamilton *et al.*, Science 313, 1441 (2006)) previously digested with *Not*I and *Asc*I, and treated with T4 DNA polymerase. The resultant vector, pSH994, contains an FX expression cassette consisting of the FX ORF operably linked at the 5' end to a *P. pastoris* PMA1 promoter (PpPMA1prom) and at the 3' end to a *P. pastoris* PMA transcription terminator sequence (PpPMA1tt). The expression cassette is flanked by *Swa*I restriction sites.

The GFTr expression cassette is generated by digesting pGLY2133 with *Not*I and *Pac*I to excise a 1.1 Kb fragment containing the GFTr ORF, which is treated with T4 DNA polymerase. Subsequently this fragment is subcloned into the vector pGLY363 (Hamilton, *supra.*), previously digested with *Not*I and *Pac*I, and treated with T4 DNA polymerase. The resultant vector, pGLY2143, contains a GFTr expression cassette consisting of the GFTr ORF operably linked at the 5' end to a PpPMA1prom and at the 3' end to a PpPMA1tt. The expression cassette is flanked by *Rsr*II restriction sites.

To generate the FUT8 catalytic domain fused to a yeast localization signal, the first 36 amino acids of *S. cerevisiae* targeting region of *Mnn2* are analyzed by the GeneOptimizer software and codon-optimized for *P. pastoris* expression (GeneArt, Regensburg, Germany). The resultant synthetic DNA for *ScMnn2* amino acids 1 to 36 is generated with 5' *Not*I and 3' *Asc*I restriction enzyme compatible ends, cloned into a shuttle vector to produce plasmid vector pSH831. Subsequently, the vector pSH987 is digested with *Asc*I and *Pac*I to liberate a 1.6 Kb fragment encoding the FUT8 catalytic domain ORF, which is then subcloned in-frame to the

DNA encoding the *ScMnn2* targeting peptide in the vector pSH831, previously digested with the same enzymes. The resultant vector is designated pSH989. To generate the FUT8-ScMnn2 expression cassette, pSH989 is digested with *Not*I and *Pac*I to release a 1.8 Kb fragment, which is subcloned into the vector pGLY361 (Hamilton *et al.*, Science 313, 1441 (2006)) digested with the same enzymes. The resultant vector, pSH991, contains a FUT8-Mnn2 fusion protein consisting of the FUT8-Mnn2 fusion ORF operably linked at the 5' end to a *P. pastoris* TEF promoter (PpTEFprom) and at the 3' end to a *P. pastoris* TEF transcription terminator sequence (PpTEFtt). The expression cassette is flanked by *Sgf*I restriction sites.

Generation of fucosylation engineering vector.

Vector pSH994 is digested with *Swa*I to release a 2.5 Kb fragment containing the FX expression cassette, which is subcloned into pSH997 (contains the GMD expression cassette) digested with *Pme*I. The resultant vector in which the PMA-FX and GAPDH-GMD expression cassettes are aligned in the same direction is designated pSH1009. The 2.7 Kb fragment containing the PMA-GFTr expression cassette is excised from pGLY2143 using the restriction enzyme *RsrII* and subcloned into pSH1009 digested with the same enzyme. The resultant vector in which the PMA-GFTr and GAPDH expression cassettes are aligned in the same direction is designated pSH1019. Finally, the 1.8 Kb TEF-FUT8 cassette is excised from pSH991 using *Sgf*I and subcloned into pSH1019 digested with the same enzyme. The resultant vector in which the TEF-FUT8 and the GAPDH expression cassettes are aligned in the same direction is designated pSH1022. This vector is illustrated in Figure 2B.

Generation of rat EPO expression vector.

A truncated form of *Rattus norvegicus* erythropoietin gene (rEPO), encoding amino acids 27 to 192, is amplified from rat kidney cDNA (BD Biosciences, Palo Alto, CA) using Advantage 2 polymerase as recommended by the manufacturer. Briefly, the primers rEPO-forward and rEPO-reverse (5'-GGGAA TTCGC TCCCC CACGC CTCAT TTGCG AC-3' (SEQ ID NO:23) and 5'-CCTCT AGATC ACCTG TCCCC TCTCC TGCAG GC-3' (SEQ ID NO:24), respectively) are used to amplify a 516 bp product from rat kidney cDNA using the following cycling conditions: 1 cycle at 94°C for 1 minute; 5 cycles at 94°C for 30 seconds, 72°C for 1 minute; 5 cycles at 94°C for 30 seconds, 70°C for 1 minute; 25 cycles at 94°C for 20 seconds, 68°C for 1 minute. Subsequently, the product is cloned into pCR2.1 (Invitrogen, Carlsbad, CA), sequenced, and the resultant construct designated pSH603. To generate the yeast expression vector, pSH603 is digested with *Eco*RI and *Xba*I to liberate a 506 bp fragment which was subcloned into pPICZαA (Invitrogen, Carlsbad, CA), which has previously been digested with the same enzymes. The resultant expression vector is designated pSH692. The rEPO in pSH692 is under the under the control of the AOX methanol-inducible promoter.

Generation of yeast strains and production of rat EPO.

A *P. pastoris* glycoengineered cell line, YGLY1062, which is capable of producing recombinant glycoproteins having predominantly $Gal_2GlcNAc_2Man_3GlcNAc_2$ *N*-glycans (similar to the strains described in U.S. Published Patent Application No. 2006/0040353, which produce glycoproteins having $Gal_2GlcNAc_2Man_3GlcNAc_2$ *N*-glycans) is transformed with vector PSH692 to produce strain RDP974, which produces recombinant rat EPO (rEPO) with $Gal_2GlcNAc_2Man_3GlcNAc_2$ *N*-glycans. Strain RDP974 is similar to stain RDP762 described in Hamilton *et al.*, Science 313, 1441-1443 (2006), which produces rat EPO having $Gal_2GlcNAc_2Man_3GlcNAc_2$ *N*-glycans.

The RPD974 strain has deletions in the *OCH1*, *PNO1*, *MNN4B*, and *BMT2* genes and includes DNA encoding the full-length *Kluyveromyces lactis* UDP-GlcNAc transporter, *M. musculus* UDP-GlcNAc transporter, *S. cerevisiae* UDP-galactose 4-epimerase, and *D. melanogaster* UDP-Galactose transporter; and DNA encoding a *M. musculus* α1,2-Mannosidase I catalytic domain fused to DNA encoding amino acids 1-36 of an *S. cerevisiae* MNN2 leader sequence; DNA encoding the *H. sapiens* β1,2-GlcNAc transferase I (GnTI) catalytic domain fused to DNA encoding amino acids 1-36 of an *S. cerevisiae* MNN2 leader sequence; DNA encoding a *Drosophila melanogaster* Mannosidase II catalytic domain fused to DNA encoding amino acids 1-36 of an *S. cerevisiae* MNN2 leader sequence, DNA encoding a *Rattus norvegicus* β1,2-GlcNAc transferase II (GnTII) catalytic domain fused to DNA encoding amino acids 1-97 of an *S. cerevisiae* MNN2 leader sequence, and DNA encoding an *H. sapiens* β1,4-galactosyltransferase (GalTI) catalytic domain fused to DNA encoding amino acids 1-58 of an *S. cerevisiae* KRE2 (MNTI) leader sequence. U.S. Published Patent Application No. 2006/0040353 discloses methods for producing *Pichia pastoris* cell lines that produce galactosylated glycoproteins in lower yeast (*See* also, U.S. Patent No. 7,029,872, U.S. Published Patent Application Nos. 2004/0018590, 2004/0230042, 2005/0208617, 2004/0171826, 2006/0286637, and 2007/0037248, and Hamilton *et al.*, Science 313, 1441-1443 (2006).

Strain RDP974 is then used as the host strain for introducing the fucosylation pathway in vector pSH1022. Briefly, 10 μg of the control plasmid pSH995 or the fucosylation pathway plasmid pSH1022 is digested with the restriction enzyme *Sfi*I to linearize the vector and transformed by electroporation into the host strain RDP974. The transformed cells are plated on YPD containing 100 ng/mL nourseothricin and incubated at 26°C for five days. Subsequently several clones are picked and analyzed for fucose transfer onto the *N*-glycans of rEPO. A strain transformed with the control vector is designated YSH660, while a strain transformed with pSH1022 and demonstrating fucose transfer is designated YSH661.

Typically, protein expression is carried out by growing the transformed strains at 26°C in 50 mL buffered glycerol-complex medium (BMGY) consisting of 1% yeast extract, 2%

peptone, 100 mM potassium phosphate buffer, pH 6.0, 1.34% yeast nitrogen base, 4 X 10-5% biotin, and 1% glycerol as a growth medium. Induction of protein expression is performed in 5 mL of buffered methanol-complex medium (BMMY), consisting of 1.5% methanol instead of glycerol in BMGY.

Recombinant rEPO is expressed as described above and Ni-chelate column purified as described in Choi *et al.* (Proc. Natl. Acad. Sci. U S A 100, 5022 (2003) and Hamilton *et al.* (Science 301, 1244 (2003)). The resultant protein is analyzed by SDS-PAGE (Laemmli, Nature 227, 680 (1970)) and stained for visualization with coomassie blue. Fucose is removed by *in vitro* digestion with α-1,6-fucosidase (Sigma-Aldrich, St. Louis, MO) treatment, as recommended by the manufacturer.

For glycan analysis, the glycans are released from rEPO by treatment with PNGase-F (Choi et al. (2003); Hamilton *et al.* (2003)). Released glycans are analyzed by MALDI/Time-of-flight (TOF) mass spectrometry to confirm glycan structures (Choi *et al.* (2003)). To quantitate the relative amount of fucosylated glycans present, the *N*-glycosidase F released glycans are labeled with 2-aminobenzidine (2-AB) and analyzed by HPLC (Choi *et al.* (2003)). The percentage of fucosylated and non-fucosylated glycans is calculated by comparing the peak area of each species before and after fucosidase treatment.

Analysis of the *N*-glycans produced in strain YSH661 produced essentially as described above showed that the strain produced recombinant rEPO comprising $Gal_2GlcNAc_2Man_3GlcNAc_2(Fuc)$ *N*-glycans. Figure 4A, which shows the results of a MALDI-TOF analysis of the *N*-glycans on rEPO produced in strain YSH661, shows that the strain produced *N*-glycans comprised $Gal_2GlcNAc_2Man_3GlcNAc_2(Fuc)$ $Gal_2GlcNAc_2Man_3GlcNAc_2$ *N*-glycans. The $Gal_2GlcNAc_2Man_3GlcNAc_2(Fuc)$ *N*-glycans are within the box. Figure 4B, which shows a MALDI-TOF analysis of the *N*-glycans on rEPO produced in control strain YSH660 (without fucosylation pathway), shows that the strain produced only afucosylated *N*-glycans comprising only $Gal_2GlcNAc_2Man_3GlcNAc_2$.

EXAMPLE 2

A *Pichia pastoris* strain capable of producing glycoproteins having $NANA_2Gal_2GlcNAc_2Man_3GlcNAc_2(Fuc)$ *N*-glycans can be made by introducing the vector pSH1022 into a *Pichia pastoris* strain capable of producing glycoproteins having $NANA_2Gal_2GlcNAc_2Man_3GlcNAc_2$ *N*-glycans. For example, vector pSH1022 containing the genes encoding the components of the fucosylation pathway can be transformed into the strain YSH597, which produces rat EPO having $NANA_2Gal_2GlcNAc_2Man_3GlcNAc_2$ N-glycans and is disclosed in U.S. Provisional Application No. 60/801,688 and Hamilton *et al.* Science 313, 1441-1443 (2006). The rat EPO produced in the strain upon induction will include $NANA_2Gal_2GlcNAc_2Man_3GlcNAc_2(Fuc)$ *N*-glycans.

The following provides a prophetic method for introducing the enzymes encoding the sialylation pathway into strain YSH661 of Example 1.

Open reading frames for *Homo sapiens* UDP-N-acetylglucosamine-2-epimerase/N-acetylmannosamine kinase (GNE), *H. sapiens* N-acetylneuraminate-9-phosphate synthase (SPS), *H. sapiens* CMP-sialic acid synthase (CSS), *Mus musculus* CMP-sialic acid transporter (CST), and amino acids 40 to 403 of *M. musculus* α-2,6-sialyltransferase (ST) are analyzed by the GeneOptimizer software and codon-optimized for *P. pastoris* expression (GeneArt, Regensburg, Germany). The resultant synthetic DNAs for GNE, SPS, CSS and CST are generated with 5' *Bsa*I and 3' *Hpa*I restriction sites, cloned into a shuttle vector and designated pGLY368, 367, 366 and 369, respectively. The synthetic DNA for ST is generated with 5' *Asc*I and 3' *Pac*I restriction sites, cloned into a shuttle vector, and designated pSH660. To generate the SPS, CSS and CST expression cassettes, the vectors pGLY367, 366 and 369 are digested with *Bsa*I and *Hpa*I to excise 1.1, 1.3, and 1.0 Kb fragments, which are treated with T4 DNA polymerase to remove single strandoverhangs. Subsequently, these fragments are subcloned into the vectors pGLY359, 17, and 363 previously digested with *Not*I and *Asc*I for the former, and *Not*I and *Pac*I for the latter two, and treated with T4 DNA polymerase. The resultant vectors pSH819, containing SPS in a PpPMA1prom-PpPMA1tt cassette flanked by *Pac*I restriction sites; pSH824, containing CSS in a PpGAPDH-ScCYCtt cassette flanked by 5' *Bgl*II and 3' *Bam*HI restriction sites; and pGLY372, containing CST in a PpPMA1prom-PpPMA1tt cassette flanked by *Rsr*II restriction sites. To generate the ST catalytic domain fused to a yeast localization signal, the *S. cerevisiae* targeting region of *Mnt1* is amplified from genomic DNA using Taq DNA polymerase (Promega, Madison, WI) and the primers ScMnt1-for and ScMnt1-rev (5'-GGGCGGCCGCCACCATGGCCCTCTTTCTC AGTAAGAGACT GTTGAG-3' (SEQ ID NO:25) and 5'-CCGGCGCGCCCGATGACTTGTTG TTCAGGGGATATAGATCCTG-3' (SEQ ID NO:26), respectively). The conditions used are: 94°C for 3 minutes, 1 cycle; 94°C for 30 seconds, 55°C for 20 seconds, 68°C for 1 minute, 30 cycles; 68°C for 5 minutes, 1 cycle. The resultant 174 bp fragment containing 5' *Not*I and 3' *Asc*I restriction sites is subcloned in-frame 5' to the codon-optimized ST, creating the vector pSH861. Subsequently this vector is digested with *Not*I and *Pac*I to excise a 1.3Kb fragment, containing the ST-fusion, treated with T4 DNA polymerase and subcloned into pGLY361 prepared by digestion with *Not*I and *Pac*I, and treated with T4 DNA polymerase. The resultant vector, containing the ST-fusion in a PpTEFprom-PpTEFtt cassette flanked by *Sgf*I restriction sites, is designated pSH893.

A yeast expression vector pSH823, containing a *P. pastoris* GAPDH promoter and *S. cerevisiae* CYC transcription terminator, is designed to integrate into the *Pichia* genome downstream of the *Trp2* ORF. The 2.6Kb fragment encoding the PMA-CST expression cassette is excised from pGLY372 using the restriction enzyme *Rsr*II and subcloned into pSH823 digested with the same enzyme. The resultant vector in which the PMA-CST and GAPDH expression cassettes are aligned in the same direction was designated pSH826. Subsequently this vector is digested with the restriction enzymes *Not*I and *Pac*I and the single strand overhangs removed with T4 DNA polymerase. Into this linearized

construct, the 2.2Kb fragment of GNE, isolated from pGLY368 by digestion with *Bsa*I and *Hpa*I, and treated with T4 DNA polymerase to remove single strand overhangs, is subcloned. This vector is designated pSH828. Subsequently this vector is digested with *Pac*I, into which the 2.7 Kb *Pac*I fragment of pSH819, encoding the PMA-SPS expression cassette, is subcloned. The vector produced, in which the PMA-SPS expression cassette is aligned in the opposite orientation to the GAPDH expression cassette, is designated pSH830. At this stage the *URA5* marker is replaced with *HIS1* by excising the 2.4 Kb URA5 fragment from pSH830 using *Xho*I and replacing it with the 1.8 Kb fragment of *HIS1* from pSH842 digested with the same enzyme. The resultant vector in which the *HIS1* ORF is aligned in the same direction as GAPDH-GNE expression cassette is designated pSH870. Subsequently, this vector is digested with *Bam*HI and the 2.1 Kb fragment from pSH824 isolated by digestion with *Bam*HI and *Bgl*II, containing the GAPDH-CSS expression cassette, is subcloned. The vector generated, in which the newly introduced expression cassette is orientated in the opposite direction as the GAPDH-GNE cassette, is designated pSH872. Next, the 2.2 Kb expression cassette containing the TEF-ST is digested with *Sgf*I from pSH893 and subcloned into pSH872 digested with the same enzyme. The vector generated, in which the TEF-ST cassette is orientated in the opposite direction as the GAPDH-GNE cassette, is designated pSH926.

The pSH926 vector is transformed into strain YSH661, which is then capable of producing rat EPO having $NANA_2Gal_2GlcNAc_2Man_3GlcNAc_2$ *N*-glycans.

EXAMPLE 3

A *Pichia pastoris* strain capable of producing a human EPO having $NANA_2Gal_2GlcNAc_2Man_3GlcNAc_2(Fuc)$ *N*-glycans can be made by introducing the vector pSH1022 into a *Pichia pastoris* strain capable of producing human EPO having $NANA_2Gal_2GlcNAc_2Man_3GlcNAc_2$ *N*-glycans. For example, vector pSH1022 containing the genes encoding the components of the fucosylation pathway can be transformed into a strain that is capable of producing glycoproteins having $NANA_2Gal_2GlcNAc_2Man_3GlcNAc_2$ N-glycans, such as strain YSH597 disclosed in Hamilton *et al.*, Science 313, 1441-1443 (2006) or YSH661 of Example 2 comprising the genes encoding the sialylation pathway enzymes but replacing the DNA encoding rat EPO with DNA encoding the human EPO. The strain will then produce human EPO having $NANA_2Gal_2GlcNAc_2Man_3GlcNAc_2(Fuc)$ N-glycans.

While the present invention is described herein with reference to illustrated embodiments, it should be understood that the invention is not limited hereto. Those having ordinary skill in the art and access to the teachings herein will recognize additional modifications and embodiments within the scope thereof. Therefore, the present invention is limited only by the claims attached herein.

SEQUENCES

SEQ ID NO:1

DNA

Homo sapian

GDP-Mannose-dehydratase (hGMD)

ATGGCACACGCACCGGCACGCTGCCCCAGCGCCCGGGGCTCCGGGGACGGCGAGATGGGCA
AGCCCAGGAACGTGGCGCTCATCACCGGTATCACAGGCCAGGATGGTTCCTACCTGGCTGAG
TTCCTGCTGGAGAAAGGCTATGAGGTCCATGGAATTGTACGGCGGTCCAGTTCATTTAATAC
GGGTCGAATTGAGCATCTGTATAAGAATCCCCAGGCTCACATTGAAGGAAACATGAAGTTG
CACTATGGCGATCTCACTGACAGTACCTGCCTTGTGAAGATCATTAATGAAGTAAAGCCCAC
AGAGATCTACAACCTTGGAGCCCAGAGCCACGTCAAAATTTCCTTTGACCTCGCTGAGTACA
CTGCGGACGTTGACGGAGTTGGCACTCTACGACTTCTAGATGCAGTTAAGACTTGTGGCCTT
ATCAACTCTGTGAAGTTCTACCAAGCCTCAACAAGTGAACTTTATGGGAAAGTGCAGGAAAT
ACCCCAGAAGGAGACCACCCCTTTCTATCCCCGGTCACCCTATGGGGCAGCAAAACTCTATG
CCTATTGGATTGTGGTGAACTTCCGTGAGGCGTATAATCTCTTTGCAGTGAACGGCATTCTCT
TCAATCATGAGAGTCCCAGAAGAGGAGCTAATTTCGTTACTCGAAAAATTAGCCGGTCAGTA
GCTAAGATTTACCTTGGACAACTGGAATGTTTCAGTTTGGGAAATCTGGATGCCAAACGAGA
TTGGGGCCATGCCAAGGACTATGTGGAGGCTATGTGGTTGATGTTGCAGAATGATGAGCCG
GAGGACTTCGTTATAGCTACTGGGGAGGTCCATAGTGTCCGGGAATTTGTCGAGAAATCATT
CTTGCACATTGGAAAAACCATTGTGTGGGAAGGAAAGAATGAAAATGAAGTGGGCAGATGT
AAAGAGACCGGCAAAGTTCACGTGACTGTGGATCTCAAGTACTACCGGCCAACTGAAGTGG
ACTTTCTGCAGGGCGACTGCACCAAAGCGAAACAGAAGCTGAACTGGAAGCCCCGGGTCGC
TTTCGATGAGCTGGTGAGGGAGATGGTGCACGCCGACGTGGAGCTCATGAGGACAAACCCC
AATGCCTGA

SEQ ID NO:2

Protein

Homo sapian

GDP-Mannose-dehydratase (hGMD)

MAHAPARCPSARGSGDGEMGKPRNVALITGITGQDGSYLAEFLLEKGYEVHGIVRRSSSFNTGRI
EHLYKNPQAHIEGNMKLHYGDLTDSTCLVKIINEVKPTEIYNLGAQSHVKISFDLAEYTADVDGV
GTLRLLDAVKTCGLINSVKFYQASTSELYGKVQEIPQKETTPFYPRSPYGAAKLYAYWIVVNFRE
AYNLFAVNGILFNHESPRRGANFVTRKISRSVAKIYLGQLECFSLGNLDAKRDWGHAKDYVEAM
WLMLQNDEPEDFVIATGEVHSVREFVEKSFLHIGKTIVWEGKNENEVGRCKETGKVHVTVDLK
YYRPTEVDFLQGDCTKAKQKLNWKPRVAFDELVREMVHADVELMRTNPNA

SEQ ID NO:3

DNA

Homo sapian

GDP-keto-deoxy-mannose-epimerase/GDP-keto-deoxy-galactose-reductase (FX protein)

ATGGGTGAACCCCAGGGATCCATGCGGATTCTAGTGACAGGGGGCTCTGGGCTGGTAGGCA
AAGCCATCCAGAAGGTGGTAGCAGATGGAGCTGGACTTCCTGGAGAGGACTGGGTGTTTGT
CTCCTCTAAAGACGCCGATCTCACGGATACAGCACAGACCCGCGCCCTGTTTGAGAAGGTCC
AACCCACACACGTCATCCATCTTGCTGCAATGGTGGGGGGCCTGTTCCGGAATATCAAATAC
AATTTGGACTTCTGGAGGAAAAACGTGCACATGAACGACAACGTCCTGCACTCGGCCTTTGA
GGTGGGGGCCCGCAAGGTGGTGTCCTGCCTGTCCACCTGTATCTTCCCTGACAAGACGACCT
ACCCGATAGATGAGACCATGATCCACAATGGGCCTCCCCACAACAGCAATTTTGGGTACTCG
TATGCCAAGAGGATGATCGACGTGCAGAACAGGGCCTACTTCCAGCAGTACGGCTGCACCT
TCACCGCTGTCATCCCCACCAACGTTTTCGGGCCCCACGACAACTTCAACATCGAGGATGGC
CACGTGCTGCCTGGCCTCATCCACAAGGTGCACCTGGCCAAGAGCAGCGGCTCGGCCCTGAC
GGTGTGGGGTACAGGGAATCCGCGGAGGCAGTTCATATACTCGCTGGACCTGGCCCAGCTCT
TTATCTGGGTCCTGCGGGAGTACAATGAAGTGGAGCCCATCATCCTCTCCGTGGGCGAGGAA
GATGAGGTCTCCATCAAGGAGGCAGCCGAGGCGGTGGTGGAGGCCATGGACTTCCATGGGG
AAGTCACCTTTGATACAACCAAGTCGGATGGGCAGTTTAAGAAGACAGCCAGTAACAGCAA
GCTGAGGACCTACCTGCCCGACTTCCGGTTCACACCCTTCAAGCAGGCGGTGAAGGAGACCT
GTGCTTGGTTCACTGACAACTACGAGCAGGCCCGGAAGTGA

SEQ ID NO:4

Protein

Homo sapian

GDP-keto-deoxy-mannose-epimerase/GDP-keto-deoxy-galactose-reductase (FX protein)

MGEPQGSMRILVTGGSGLVGKAIQKVVADGAGLPGEDWVFVSSKDADLTDTAQTRALFEKVQP
THVIHLAAMVGGLFRNIKYNLDFWRKNVHMNDNVLHSAFEVGARKVVSCLSTCIFPDKTTYPID
ETMIHNGPPHNSNFGYSYAKRMIDVQNRAYFQQ
YGCTFTAVIPTNVFGPHDNFNIEDGHVLPGLIHKVHLAKSSGSALTVWGTGNPRRQFIYSLDLAQ
LFIWVLREYNEVEPIILSVGEEDEVSIKEAAEAVVEAMDFHGEVTFDTTKSDGQFKKTASNSKLR
TYLPDFRFTPFKQAVKETCAWFTDNYEQARK

SEQ ID NO:5

DNA
Homo sapian
Human GDP-fucose transporter
ATGAATAGGGCCCCTCTGAAGCGGTCCAGGATCCTGCACATGGCGCTGACCGGGGCCTCAG
ACCCCTCTGCAGAGGCAGAGGCCAACGGGGAGAAGCCCTTTCTGCTGCGGGCATTGCAGAT
CGCGCTGGTGGTCTCCCTCTACTGGGTCACCTCCATCTCCATGGTGTTCCTTAATAAGTACCT
GCTGGACAGCCCCTCCCTGCGGCTGGACACCCCCATCTTCGTCACCTTCTACCAGTGCCTGGT
GACCACGCTGCTGTGCAAAGGCCTCAGCGCTCTGGCCGCCTGCTGCCCTGGTGCCGTGGACT
TCCCCAGCTTGCGCCTGGACCTCAGGGTGGCCCGCAGCGTCCTGCCCCTGTCGGTGGTCTTC
ATCGGCATGATCACCTTCAATAACCTCTGCCTCAAGTACGTCGGTGTGGCCTTCTACAATGT
GGGCCGCTCACTCACCACCGTCTTCAACGTGCTGCTCTCCTACCTGCTGCTCAAGCAGACCA
CCTCCTTCTATGCCCTGCTCACCTGCGGTATCATCATCGGGGGCTTCTGGCTTGGTGTGGACC
AGGAGGGGGCAGAAGGCACCCTGTCGTGGCTGGGCACCGTCTTCGGCGTGCTGGCTAGCCT
CTGTGTCTCGCTCAACGCCATCTACACCACGAAGGTGCTCCCGGCGGTGGACGGCAGCATCT
GGCGCCTGACTTTCTACAACAACGTCAACGCCTGCATCCTCTTCCTGCCCCTGCTCCTGCTGC
TCGGGGAGCTTCAGGCCCTGCGTGACTTTGCCCAGCTGGGCAGTGCCCACTTCTGGGGGATG
ATGACGCTGGGCGGCCTGTTTGGCTTTGCCATCGGCTACGTGACAGGACTGCAGATCAAGTT
CACCAGTCCGCTGACCCACAATGTGTCGGGCACGGCCAAGGCCTGTGCCCAGACAGTGCTG
GCCGTGCTCTACTACGAGGAGACCAAGAGCTTCCTCTGGTGGACGAGCAACATGATGGTGCT
GGGCGGCTCCTCCGCCTACACCTGGGTCAGGGGCTGGGAGATGAAGAAGACTCCGGAGGAG
CCCAGCCCCAAAGACAGCGAGAAGAGCGCCATGGGGGTGTGA

SEQ ID NO:6
Protein
Homo sapian
Human GDP-fucose transporter
MNRAPLKRSRILHMALTGASDPSAEAEANGEKPFLLRALQIALVVSLYWVTSISMVFLNKYLLDS
PSLRLDTPIFVTFYQCLVTTLLCKGLSALAACCPGAVDFPSLRLDLRVARSVLPLSVVFIGMITFN
NLCLKYVGVAFYNVGRSLTTVFNVLLSYLLLKQTTSFYALLTCGIIIGGFWLGVDQEGAEGTLS
WLGTVFGVLASLCVSLNAIYTTKVLPAVDGSIWRLTFYNNVNACILFLPLLLLLGELQALRDFAQ
LGSAHFWGMMTLGGLFGFAIGYVTGLQIKFTSPLTHNVSGTAKACAQTVLAVLYYEETKSFLW
WTSNMMVLGGSSAYTWVRGWEMKKTPEEPSPKDSEKSAMGV

SEQ ID NO:7
DNA

Homo sapian

alpha 1,6 fucosyltransferase (hFuT8)

ATGCGGCCATGGACTGGTTCCTGGCGTTGGATTATGCTCATTCTTTTTGCCTGGGGGACCTTG
CTGTTTTATATAGGTGGTCACTTGGTACGAGATAATGACCATCCTGATCACTCTAGCCGAGA
ACTGTCCAAGATTCTGGCAAAGCTTGAACGCTTAAAACAACAGAATGAAGACTTGAGGCGA
ATGGCCGAATCTCTCCGGATACCAGAAGGCCCTATTGATCAGGGGCCAGCTATAGGAAGAG
TACGCGTTTTAGAAGAGCAGCTTGTTAAGGCCAAAGAACAGATTGAAAATTACAAGAAACA
GACCAGAAATGGTCTGGGGAAGGATCATGAAATCCTGAGGAGGAGGATTGAAAATGGAGCT
AAAGAGCTCTGGTTTTTCCTACAGAGTGAATTGAAGAAATTAAAGAACTTAGAAGGAAATG
AACTCCAAAGACATGCAGATGAATTTCTTTTGGATTTAGGACATCATGAAAGGTCTATAATG
ACGGATCTATACTACCTCAGTCAGACAGATGGAGCAGGTGATTGGCGGGAAAAAGAGGCCA
AAGATCTGACAGAACTGGTTCAGCGGAGAATAACATATCTTCAGAATCCCAAGGACTGCAG
CAAAGCCAAAAAGCTGGTGTGTAATATCAACAAAGGCTGTGGCTATGGCTGTCAGCTCCATC
ATGTGGTCTACTGCTTCATGATTGCATATGGCACCCAGCGAACACTCATCTTGGAATCTCAG
AATTGGCGCTATGCTACTGGTGGATGGGAGACTGTATTTAGGCCTGTAAGTGAGACATGCAC
AGACAGATCTGGCATCTCCACTGGACACTGGTCAGGTGAAGTGAAGGACAAAAATGTTCAA
GTGGTCGAGCTTCCCATTGTAGACAGTCTTCATCCCCGTCCTCCATATTTACCCTTGGCTGTA
CCAGAAGACCTCGCAGATCGACTTGTACGAGTGCATGGTGACCCTGCAGTGTGGTGGGTGTC
TCAGTTTGTCAAATACTTGATCCGCCCACAGCCTTGGCTAGAAAAAGAAATAGAAGAAGCC
ACCAAGAAGCTTGGCTTCAAACATCCAGTTATTGGAGTCCATGTCAGACGCACAGACAAAG
TGGGAACAGAAGCTGCCTTCCATCCCATTGAAGAGTACATGGTGCATGTTGAAGAACATTTT
CAGCTTCTTGCACGCAGAATGCAAGTGGACAAAAAAAGAGTGTATTTGGCCACAGATGACC
CTTCTTTATTAAAGGAGGCAAAAACAAAGTACCCCAATTATGAATTTATTAGTGATAACTCT
ATTTCCTGGTCAGCTGGACTGCACAATCGATACACAGAAAATTCACTTCGTGGAGTGATCCT
GGATATACATTTTCTCTCTCAGGCAGACTTCCTAGTGTGTACTTTTTCATCCCAGGTCTGTCG
AGTTGCTTATGAAATTATGCAAACACTACATCCTGATGCCTCTGCAAACTTCCATTCTTTAGA
TGACATCTACTATTTTGGGGGCCAGAATGCCCACAATCAAATTGCCATTTATGCTCACCAAC
CCCGAACTGCAGATGAAATTCCCATGGAACCTGGAGATATCATTGGTGTGGCTGGAAATCAT
TGGGATGGCTATTCTAAAGGTGTCAACAGGAAATTGGGAAGGACGGGCCTATATCCCTCCTA
CAAAGTTCGAGAGAAGATAGAAACGGTCAAGTACCCCACATATCCTGAGGCTGAGAAATAA

SEQ ID NO:8

Protein

Homo sapian

alpha 1,6 fucosyltransferase (hFuT8)

MRPWTGSWRWIMLILFAWGTLLFYIGGHLVRDNDHPDHSSRELSKILAKLERLKQQNEDLRRM
AESLRIPEGPIDQGPAIGRVRVLEEQLVKAKEQIENYKKQTRNGLGKDHEILRRRIENGAKELWFF
LQSELKKLKNLEGNELQRHADEFLLDLGHHERSIMTDLYYLSQTDGAGDWREKEAKDLTELVQ
RRITYLQNPKDCSKAKKLVCNINKGCGYGCQLHHVVYCFMIAYGTQRTLILESQNWRYATGGW
ETVFRPVSETCTDRSGISTGHWSGEVKDKNVQVVELPIVDSLHPRPPYLPLAVPEDLADRLVRVH
GDPAVWWVSQFVKYLIRPQPWLEKEIEEATKKLGFKHPVIGVHVRRTDKVGTEAAFHPIEEYMV
HVEEHFQLLARRMQVDKKRVYLATDDPSLLKEAKTKYPNYEFISDNSISWSAGLHNRYTENSLR
GVILDIHFLSQADFLVCTFSSQVCRVAYEIMQTLHPDASANFHSLDDIYYFGGQNAHNQIAIYAHQ
PRTADEIPMEPGDIIGVAGNHWDGYSKGVNRKLGRTGLYPSYKVREKIETVKYPTYPEAEK

SEQ ID NO:9
DNA
Rat
Alpha-1,6-fucosyltransferase (rFuT8)
ATGCGGGCATGGACTGGTTCCTGGCGTTGGATTATGCTCATTCTTTTTGCCTGGGGGACCTTG
TTGTTTTATATAGGTGGTCATTTGGTTCGAGATAATGACCACCCTGATCACTCTAGCAGAGA
ACTCTCCAAGATTCTTGCAAAGCTTGAACGCTTAAAACAACAAAATGAAGACTTGAGGCGA
ATGGCTGAGTCTCTACGAATACCAGAAGGCCCCATTGACCAGGGGACGGCTACGGGAAGAG
TCCGTGTTTTAGAAGAACAGCTTGTTAAGGCCAAAGAACAGATTGAAAATTACAAGAAACA
AGCCAGAAATGGTCTGGGGAAGGATCATGAACTCTTAAGGAGGAGGATTGAAAATGGAGCT
AAAGAGCTCTGGTTTTTTCTACAAAGTGAACTGAAGAAATTAAAGCATCTAGAAGGAAATG
AACTCCAAAGACATGCAGATGAAATTCTTTTGGATTTAGGACACCATGAAAGGTCTATCATG
ACGGATCTATACTACCTCAGTCAAACAGATGGAGCAGGGGATTGGCGTGAAAAAGAGGCCA
AAGATCTGACAGAGCTGGTCCAGCGGAGAATAACTTATCTCCAGAATCCCAAGGACTGCAG
CAAAGCCAGGAAGCTGGTGTGTAACATCAATAAGGGCTGTGGCTATGGTTGCCAACTCCATC
ACGTGGTCTACTGTTTCATGATTGCTTATGGCACCCAGCGAACACTCATCTTGGAATCTCAG
AATTGGCGCTATGCTACTGGTGGATGGGAGACTGTGTTTAGACCTGTAAGTGAGACATGCAC
AGACAGATCTGGCCTCTCCACTGGACACTGGTCAGGTGAAGTGAATGACAAAAATATTCAA
GTGGTGGAGCTCCCCATTGTAGACAGCCTCCATCCTCGGCCTCCTTACTTACCACTGGCTGTT
CCAGAAGACCTTGCAGATCGACTCGTAAGAGTCCATGGTGATCCTGCAGTGTGGTGGGTGTC
CCAGTTCGTCAAATATTTGATTCGTCCACAACCTTGGCTAGAAAAGGAAATAGAAGAAGCC
ACCAAGAAGCTTGGCTTCAAACATCCAGTCATTGGAGTCCATGTCAGACGCACAGACAAAG
TGGGAACAGAGGCAGCCTTCCATCCCATCGAAGAGTACATGGTACATGTTGAAGAACATTTT
CAGCTTCTCGCACGCAGAATGCAAGTGGATAAAAAAAAGAGTATATCTGGCTACCGATGACC
CTGCTTTGTTAAAGGAGGCAAAGACAAAGTACTCCAATTATGAATTTATTAGTGATAACTCT
ATTTCTTGGTCAGCTGGATTACACAATCGGTACACAGAAAATTCACTTCGGGGCGTGATCCT

GGATATACACTTTCTCTCTCAGGCTGACTTCCTAGTGTGTACTTTTTCATCCCAGGTCTGTCG
GGTTGCTTATGAAATCATGCAAACCCTGCATCCTGATGCCTCTGCAAACTTCCACTCTTTAGA
TGACATCTACTATTTTGGAGGCCAAAATGCCCACAACCAGATTGCCGTTTATCCTCACAAAC
CTCGAACTGATGAGGAAATTCCAATGGAACCTGGAGATATCATTGGTGTGGCTGGAAACCA
TTGGGATGGTTATTCTAAAGGTGTCAACAGAAAACTTGGAAAAACAGGCTTATATCCCTCCT
ACAAAGTCCGAGAGAAGATAGAAACAGTCAAGTATCCCACATATCCTGAAGCTGAAAAATA
G

SEQ ID NO:10
Protein
Rat
Alpha-1,6-fucosyltransferase (rFuT8)
MRAWTGSWRWIMLILFAWGTLLFYIGGHLVRDNDHPDHSSRELSKILAKLERLKQQNEDLRRM
AESLRIPEGPIDQGTATGRVRVLEEQLVKAKEQIENYKKQARNGLGKDHELLRRRIENGAKELW
FFLQSELKKLKHLEGNELQRHADEILLDLGHHE
RSIMTDLYYLSQTDGAGDWREKEAKDLTELVQRRITYLQNPKDCSKARKLVCNINKGCGYGCQ
LHHVVYCFMIAYGTQRTLILESQNWRYATGGWETVFRPVSETCTDRSGLSTGHWSGEVNDKNI
QVVELPIVDSLHPRPPYLPLAVPEDLADRLVRVH
GDPAVWWVSQFVKYLIRPQPWLEKEIEEATKKLGFKHPVIGVHVRRTDKVGTEAAFHPIEEYMV
HVEEHFQLLARRMQVDKKRVYLATDDPALLKEAKTKYSNYEFISDNSISWSAGLHNRYTENSLR
GVILDIHFLSQADFLVCTFSSQVCRVAYEIMQTLHPDASANFHSLDDIYYFGGQNAHNQIAVYPH
KPRTDEEIPMEPGDIIGVAGNHWDGYSKGVNRKLGKTGLYPSYKVREKIETVKYPTYPEAEK

SEQ ID NO:11
DNA
Mouse
Alpha-1,6-fucosyltransferase (mFuT8)
ATGCGGGCATGGACTGGTTCCTGGCGTTGGATTATGCTCATTCTTTTTGCCTGGGGGACCTTG
TTATTTTATATAGGTGGTCATTTGGTTCGAGATAATGACCACCCTGATCACTCCAGCAGAGA
ACTCTCCAAGATTCTTGCAAAGCTTGAACGCTTAAAACAGCAAAATGAAGACTTGAGGCGA
ATGGCTGAGTCTCTCCGAATACCAGAAGGCCCCATTGACCAGGGGACAGCTACAGGAAGAG
TCCGTGTTTTAGAAGAACAGCTTGTTAAGGCCAAAGAACAGATTGAAAATTACAAGAAACA
AGCTAGAAATGGTCTGGGGAAGGATCATGAAATCTTAAGAAGGAGGATTGAAAATGGAGCT
AAAGAGCTCTGGTTTTTTCTACAAAGCGAACTGAAGAAATTAAAGCATTTAGAAGGAAATG
AACTCCAAAGACATGCAGATGAAATTCTTTTGGATTTAGGACACCATGAAAGGTCTATCATG

ACAGATCTATACTACCTCAGTCAAACAGATGGAGCAGGGGATTGGCGTGAAAAAGAGGCCA
AAGATCTGACAGAGCTGGTCCAGCGGAGAATAACATATCTCCAGAATCCTAAGGACTGCAG
CAAAGCCAGGAAGCTGGTGTGTAACATCAATAAAGGCTGTGGCTATGGTTGTCAACTCCATC
ACGTGGTCTACTGTTTCATGATTGCTTATGGCACCCAGCGAACACTCATCTTGGAATCTCAG
AATTGGCGCTATGCTACTGGTGGATGGGAGACTGTGTTTAGACCTGTAAGTGAGACATGTAC
AGACAGATCTGGCCTCTCCACTGGACACTGGTCAGGTGAAGTAAATGACAAAAACATTCAA
GTGGTCGAGCTCCCCATTGTAGACAGCCTCCATCCTCGGCCTCCTTACTTACCACTGGCTGTT
CCAGAAGACCTTGCAGACCGACTCCTAAGAGTCCATGGTGACCCTGCAGTGTGGTGGGTGTC
CCAGTTTGTCAAATACTTGATTCGTCCACAACCTTGGCTGGAAAAGGAAATAGAAGAAGCC
ACCAAGAAGCTTGGCTTCAAACATCCAGTTATTGGAGTCCATGTCAGACGCACAGACAAAG
TGGGAACAGAAGCAGCCTTCCACCCCATCGAGGAGTACATGGTACACGTTGAAGAACATTT
TCAGCTTCTCGCACGCAGAATGCAAGTGGATAAAAAAAAGAGTATATCTGGCTACTGATGATC
CTACTTTGTTAAAGGAGGCAAAGACAAAGTACTCCAATTATGAATTTATTAGTGATAACTCT
ATTTCTTGGTCAGCTGGACTACACAATCGGTACACAGAAAATTCACTTCGGGGTGTGATCCT
GGATATACACTTTCTCTCACAGGCTGACTTTCTAGTGTGTACTTTTTCATCCCAGGTCTGTCG
GGTTGCTTATGAAATCATGCAAACCCTGCATCCTGATGCCTCTGCGAACTTCCATTCTTTGGA
TGACATCTACTATTTTGGAGGCCAAAATGCCCACAATCAGATTGCTGTTTATCCTCACAAAC
CTCGAACTGAAGAGGAAATTCCAATGGAACCTGGAGATATCATTGGTGTGGCTGGAAACCA
TTGGGATGGTTATTCTAAAGGTATCAACAGAAAACTTGGAAAAACAGGCTTATATCCCTCCT
ACAAAGTCCGAGAGAAGATAGAAACAGTCAAGTATCCCACATATCCTGAAGCTGAAAAATA
G

SEQ ID NO:12

Protein

Mouse

Alpha-1,6-fucosyltransferase (mFuT8)

MRAWTGSWRWIMLILFAWGTLLFYIGGHLVRDNDHPDHSSRELSKILAKLERLKQQNEDLRRM
AESLRIPEGPIDQGTATGRVRVLEEQLVKAKEQIENYKKQARNGLGKDHEILRRRIENGAKELWF
FLQSELKKLKHLEGNELQRHADEILLDLGHHERSIMTDLYYLSQTDGAGDWREKEAKDLTELVQ
RRITYLQNPKDCSKARKLVCNINKGCGYGCQLHHVVYCFMIAYGTQRTLILESQNWRYATGGW
ETVFRPVSETCTDRSGLSTGHWSGEVNDKNIQVVELPIVDSLHPRPPYLPLAVPEDLADRLLRVH
GDPAVWWVSQFVKYLIRPQPWLEKEIEEATKKLGFKHPVIGVHVRRTDKVGTEAAFHPIEEYMV
HVEEHFQLLARRMQVDKKRVYLATDDPTLLKEAKTKYSNYEFISDNSISWSAGLHNRYTENSLR
GVILDIHFLSQADFLVCTFSSQVCRVAYEIMQTLHPDASANFHSLDDIYYFGGQNAHNQIAVYPH
KPRTEEEIPMEPGDIIGVAGNHWDGYSKGINRKLGKTGLYPSYKVREKIETVKYPTYPEAEK

SEQ ID NO:13
DNA
Porcine
Alpha-1,6-fucosyltransferase (pFuT8)

ATGCGGCCATGGACTGGTTCGTGGCGTTGGATTATGCTCATTCTTTTTGCCTGGGGGACCTTG
CTATTTTACATAGGTGGTCACTTGGTACGAGATAATGACCACTCTGATCACTCTAGCCGAGA
ACTGTCCAAGATTTTGGCAAAGCTGGAACGCTTAAAACAACAAAATGAAGACTTGAGGAGA
ATGGCTGAATCTCTCCGAATACCAGAAGGCCCCATTGATCAGGGGCCAGCTTCAGGAAGAG
TTCGTGCTTTAGAAGAGCAATTTATGAAGGCCAAAGAACAGATTGAAAATTATAAGAAACA
AACTAAAAATGGTCCAGGGAAGGATCATGAAATCCTAAGGAGGAGGATTGAAAATGGAGCT
AAAGAGCTCTGGTTTTTTCTACAAAGTGAGTTGAAGAAATTAAAGAATTTAGAAGGAAATG
AACTCCAAAGACATGCAGATGAATTTCTATCAGATTTGGGACATCATGAAAGGTCTATAATG
ACGGATCTATACTACCTCAGTCAAACAGATGGGGCAGGTGATTGGCGTGAAAAGGAGGCCA
AAGATCTGACAGAGCTGGTCCAGCGGAGAATAACATATCTTCAGAATCCCAAGGACTGCAG
CAAAGCCAAGAAGCTAGTGTGTAATATCAACAAAGGCTGTGGCTATGGCTGTCAGCTCCATC
ATGTAGTGTACTGCTTTATGATTGCATATGGCACCCAGCGAACACTCGCCTTGGAATCTCAC
AATTGGCGCTACGCTACTGGGGGATGGGAAACTGTGTTTAGACCTGTAAGTGAGACGTGCA
CAGACAGATCTGGCAGCTCCACTGGACATTGGTCAGGTGAAGTAAAGGACAAAAATGTTCA
GGTGGTTGAGCTCCCCATTGTAGACAGTGTTCATCCTCGTCCTCCATATTTACCCCTGGCTGT
CCCAGAAGACCTTGCAGATCGACTTGTACGAGTCCATGGTGATCCTGCAGTGTGGTGGGTAT
CCCAGTTTGTCAAGTACTTGATTCGCCCACAACCCTGGCTGGAAAAGGAAATAGAAGAGGC
CACCAAGAAGCTAGGCTTCAAACATCCAGTTATTGGAGTCCATGTTAGACGCACAGACAAA
GTGGGAGCGGAAGCAGCCTTCCATCCCATTGAGGAATACACGGTGCACGTTGAAGAAGACT
TTCAGCTTCTTGCTCGCAGAATGCAAGTGGATAAAAAAAGGGTGTATTTGGCCACAGATGAC
CCTGCTTTGTTAAAAGAGGCAAAAACAAAGTACCCCAGTTATGAATTTATTAGTGATAACTC
TATCTCTTGGTCAGCTGGACTACATAATCGATATACAGAAAATTCACTTCGGGGTGTGATCC
TGGATATACACTTTCTCTCCCAGGCAGACTTCCTAGTGTGTACTTTTTCATCGCAGGTCTGTA
GAGTTGCTTATGAAATCATGCAAGCGCTGCATCCTGATGCCTCTGCGAACTTCCGTTCTTTGG
ATGACATCTACTATTTTGGAGGCCCAAATGCCCACAACCAAATTGCCATTTATCCTCACCAA
CCTCGAACTGAAGGAGAAATCCCCATGGAACCTGGAGATATTATTGGTGTGGCTGGAAATC
ACTGGGATGGCTATCCTAAAGGTGTTAACAGAAAACTGGGAAGGACGGGCCTATATCCCTC
CTACAAAGTTCGAGAGAAGATAGAAACAGTCAAGTACCCCACATATCCCGAGGCTGACAAG
TAA

SEQ ID NO:13

Protein
Porcine
Alpha-1,6-fucosyltransferase (pFuT8)
MRPWTGSWRWIMLILFAWGTLLFYIGGHLVRDNDHSDHSSRELSKILAKLERLKQQNEDLRRM
AESLRIPEGPIDQGPASGRVRALEEQFMKAKEQIENYKKQTKNGPGKDHEILRRRIENGAKELWF
FLQSELKKLKNLEGNELQRHADEFLSDLGHHERSIMTDLYYLSQTDGAGDWREKEAKDLTELV
QRRITYLQNPKDCSKAKKLVCNINKGCGYGCQLHHVVYCFMIAYGTQRTLALESHNWRYATGG
WETVFRPVSETCTDRSGSSTGHWSGEVKDKNVQVVELPIVDSVHPRPPYLPLAVPEDLADRLVR
VHGDPAVWWVSQFVKYLIRPQPWLEKEIEEATKKLGFKHPVIGVHVRRTDKVGAEAAFHPIEEY
TVHVEEDFQLLARRMQVDKKRVYLATDDPALLKEAKTKYPSYEFISDNSISWSAGLHNRYTENS
LRGVILDIHFLSQADFLVCTFSSQVCRVAYEIMQALHPDASANFRSLDDIYYFGGPNAHNQIAIYP
HQPRTEGEIPMEPGDIIGVAGNHWDGYPKGVNRKLGRTGLYPSYKVREKIETVKYPTYPEADK

SEQ ID NO:15
DNA
Artificial
PCR primer SH415
GGCGG CCGCC ACCAT GGCAC ACGCA CCGGC ACGCT GC

SEQ ID NO:16
DNA
Artificial
PCR primer SH413
TTAAT TAATC AGGCA TTGGG GTTTG TCCTC ATG

SEQ ID NO:17
DNA
Artificial
PCR primer SH414
GGCGG CCGCC ACCAT GGGTG AACCC CAGGG ATCCA TG

SEQ ID NO:18
DNA

Artificial
PCR primer SH411
TTAAT TAATC ACTTC CGGGC CTGCT CGTAG TTG

SEQ ID NO:19
DNA
Artificial
PCR primer RCD679
GCGGC CGCCA CCATG AATAG GGCCC CTCTG AAGCG G

SEQ ID NO:20
DNA
Artificial
PCR primer RCD680
TTAAT TAATC ACACC CCCAT GGCGC TCTTC TC

SEQ ID NO:21
DNA
Artificial
PCR primer SH420
GCGGC GCGCC GATAA TGACC ACCCT GATCA CTCCA G

SEQ ID NO:22
DNA
Artificial
PCR primer SH421
CCTTA ATTAA CTATT TTTCA GCTTC AGGAT ATGTG GG-

SEQ ID NO:23
DNA
Artificial
PCR primer rEPO-forward

GGGAA TTCGC TCCCC CACGC CTCAT TTGCG AC

SEQ ID NO:24
DNA
Artificial
PCR primer rEPO-reverse
CCTCT AGATC ACCTG TCCCC TCTCC TGCAG GC

SEQ ID NO:25
DNA
Artificial
PCR primer ScMnt1-for
GGGCGGCCGCCACCATGGCCCTCTTTCTC AGTAAGAGACT GTTGAG

SEQ ID NO:26
DNA
Artificial
PCR primer ScMnt1-rev
CCGGCGCGCCCGATGACTTGTTG TTCAGGGGATATAGATCCTG

WHAT IS CLAIMED:

1. A recombinant lower eukaryote host cell comprising a fucosylation pathway.

2. The host cell of Claim 1 which is yeast or filamentous fungus.

3. The host cell of Claim 2 wherein the yeast is a *Pichia* sp.

4. The host cell of Claim 3 wherein the *Pichia* sp. is *Pichia pastoris*.

5. The host cell of claim 1 wherein the host cell further does not display α1,6-mannosyltransferase activity with respect to the *N*-glycan on a glycoprotein and includes an α1,2-mannosidase catalytic domain fused to a cellular targeting signal peptide not normally associated with the catalytic domain and selected to target α1,2-mannosidase activity to the ER or Golgi apparatus of the host cell whereby, upon passage of a recombinant glycoprotein through the ER or Golgi apparatus of the host cell, a recombinant glycoprotein comprising a fucosylated $Man_5GlcNAc_2$ glycoform is produced.

6. The host cell of Claim 5 further including a GlcNAc transferase I catalytic domain fused to a cellular targeting signal peptide not normally associated with the catalytic domain of and selected to target GlcNAc transferase I activity to the ER or Golgi apparatus of the host cell; whereby, upon passage of the recombinant glycoprotein through the ER or Golgi apparatus of the host cell, a recombinant glycoprotein comprising a fucosylated $GlcNAcMan_5GlcNAc_2$ glycoform is produced.

7. The host cell of Claim 6 further including a mannosidase II catalytic domain fused to a cellular targeting signal peptide not normally associated with the catalytic domain and selected to target mannosidase II activity to the ER or Golgi apparatus of the host cell; whereby, upon passage of the recombinant glycoprotein through the ER or Golgi apparatus of the host cell, a recombinant glycoprotein comprising a fucosylated $GlcNAcMan_3GlcNAc_2$ glycoform is produced.

8. The host cell of Claim 7 further including a GlcNAc transferase II catalytic domain fused to a cellular targeting signal peptide not normally associated with the catalytic domain and selected to target GlcNAc transferase II activity to the ER or Golgi apparatus of the host cell; whereby, upon passage of the recombinant glycoprotein through the ER or Golgi

apparatus of the host cell, a recombinant glycoprotein comprising a fucosylated $GlcNAc_2Man_3GlcNAc_2$ glycoform is produced.

9. The host cell of Claim 8 further including a Galactose transferase II catalytic domain fused to a cellular targeting signal peptide not normally associated with the catalytic domain and selected to target Galactose transferase II activity to the ER or Golgi apparatus of the host cell; whereby, upon passage of the recombinant glycoprotein through the ER or Golgi apparatus of the host cell, a recombinant glycoprotein comprising a fucosylated $Gal_2GlcNAc_2Man_3GlcNAc_2$ glycoform is produced.

10. The host cell of Claim 9 further including a sialytransferase catalytic domain fused to a cellular targeting signal peptide not normally associated with the catalytic domain and selected to target sialytransferase activity to the ER or Golgi apparatus of the host cell; whereby, upon passage of the recombinant glycoprotein through the ER or Golgi apparatus of the host cell, a recombinant glycoprotein comprising a fucosylated $NANA_2Gal_2GlcNAc_2Man_3GlcNAc_2$ glycoform is produced.

11. A glycoprotein composition comprising a plurality of glycoforms, each glycoform comprising at least one *N*-glycan attached thereto, wherein the glycoprotein composition thereby comprises a plurality of *N*-glycans in which greater than 25 mole percent of the plurality of *N*-glycans consist essentially of a fucosylated *N*-glycan selected from the group consisting of $Man_5GlcNAc_2$, $GlcNAcMan_5GlcNAc_2$, $Man_3GlcNAc_2$, $GlcNAcMan_3GlcNAc_2$, $GlcNAc_2Man_3GlcNAc_2$, $GalGlcNAc_2Man_3GlcNAc_2$, $Gal_2GlcNAc_2Man_3GlcNAc_2$, $NANAGal_2GlcNAc_2Man_3GlcNAc_2$, and $NANA_2Gal_2GlcNAc_2Man_3GlcNAc_2$.

12. The glycoprotein composition of Claim 11 wherein the fucosylated *N*-glycans are greater than 50 mole percent of the plurality of *N*-glycans.

13. The glycoprotein composition of Claim 11 wherein the fucosylated *N*-glycans are greater than 75 mole percent of the plurality of *N*-glycans.

14. The glycoprotein composition of Claim 11 wherein the fucosylated *N*-glycans are greater than 80 mole percent of the plurality of *N*-glycans.

15. The glycoprotein composition of Claim 11 wherein the fucose is in an α1,3-linkage with the GlcNAc at the reducing end of the *N*-glycan.

16. The glycoprotein composition of Claim 11 wherein the fucose is in an α1,6-linkage with the GlcNAc at the reducing end of the *N*-glycan.

17. The glycoprotein composition of Claim 11 wherein the fucose is in an α1,2-linkage with the Gal at the non-reducing end of the *N*-glycan.

18. The glycoprotein composition of Claim 11 wherein the fucose is in an α1,3-linkage with the GlcNac at the non-reducing end of the *N*-glycan.

19. The glycoprotein composition of Claim 11 wherein the fucose is in an α1,4-linkage with a GlcNAc at the non-reducing end of the *N*-glycan.

20. A hybrid vector comprising (a) DNA regulatory elements which are functional in a lower eukaryotic host cell operatively linked to (b) DNA coding sequence encoding a fusion protein encoding (i) a targeting sequence; and (b) a catalytic domain of a fucosylation pathway enzyme.

21. The vector of Claim 17 wherein the fucosylation pathway enzyme is a fucosyltransferase.

SEQUENCE LISTING

<110> Stephen Hamilton

<120> PRODUCTION OF GLYCOPROTEINS WITH
MODIFIED FUCOSYLATION

<130> GF0028Y

<150> 60/905,345
<151> 2007-03-07

<160> 26

<170> FastSEQ for Windows Version 4.0

<210> 1
<211> 1119
<212> DNA
<213> Homo sapians

<220>
<221> CDS
<222> (1)...(1119)
<223> GDP-Mannose-dehydratase (hGMD)

<400> 1

```
atg gca cac gca ccg gca cgc tgc ccc agc gcc cgg ggc tcc ggg gac   48
Met Ala His Ala Pro Ala Arg Cys Pro Ser Ala Arg Gly Ser Gly Asp
 1               5                  10                  15

ggc gag atg ggc aag ccc agg aac gtg gcg ctc atc acc ggt atc aca   96
Gly Glu Met Gly Lys Pro Arg Asn Val Ala Leu Ile Thr Gly Ile Thr
            20                  25                  30

ggc cag gat ggt tcc tac ctg gct gag ttc ctg ctg gag aaa ggc tat   144
Gly Gln Asp Gly Ser Tyr Leu Ala Glu Phe Leu Leu Glu Lys Gly Tyr
        35                  40                  45

gag gtc cat gga att gta cgg cgg tcc agt tca ttt aat acg ggt cga   192
Glu Val His Gly Ile Val Arg Arg Ser Ser Ser Phe Asn Thr Gly Arg
    50                  55                  60

att gag cat ctg tat aag aat ccc cag gct cac att gaa gga aac atg   240
Ile Glu His Leu Tyr Lys Asn Pro Gln Ala His Ile Glu Gly Asn Met
65                  70                  75                  80

aag ttg cac tat ggc gat ctc act gac agt acc tgc ctt gtg aag atc   288
Lys Leu His Tyr Gly Asp Leu Thr Asp Ser Thr Cys Leu Val Lys Ile
                85                  90                  95

att aat gaa gta aag ccc aca gag atc tac aac ctt gga gcc cag agc   336
Ile Asn Glu Val Lys Pro Thr Glu Ile Tyr Asn Leu Gly Ala Gln Ser
            100                 105                 110

cac gtc aaa att tcc ttt gac ctc gct gag tac act gcg gac gtt gac   384
His Val Lys Ile Ser Phe Asp Leu Ala Glu Tyr Thr Ala Asp Val Asp
        115                 120                 125

gga gtt ggc act cta cga ctt cta gat gca gtt aag act tgt ggc ctt   432
Gly Val Gly Thr Leu Arg Leu Leu Asp Ala Val Lys Thr Cys Gly Leu
    130                 135                 140

atc aac tct gtg aag ttc tac caa gcc tca aca agt gaa ctt tat ggg   480
Ile Asn Ser Val Lys Phe Tyr Gln Ala Ser Thr Ser Glu Leu Tyr Gly
145                 150                 155                 160
```

```
aaa gtg cag gaa ata ccc cag aag gag acc acc cct ttc tat ccc cgg   528
Lys Val Gln Glu Ile Pro Gln Lys Glu Thr Thr Pro Phe Tyr Pro Arg
                165                 170                 175

tca ccc tat ggg gca gca aaa ctc tat gcc tat tgg att gtg gtg aac   576
Ser Pro Tyr Gly Ala Ala Lys Leu Tyr Ala Tyr Trp Ile Val Val Asn
            180                 185                 190

ttc cgt gag gcg tat aat ctc ttt gca gtg aac ggc att ctc ttc aat   624
Phe Arg Glu Ala Tyr Asn Leu Phe Ala Val Asn Gly Ile Leu Phe Asn
        195                 200                 205

cat gag agt ccc aga aga gga gct aat ttc gtt act cga aaa att agc   672
His Glu Ser Pro Arg Arg Gly Ala Asn Phe Val Thr Arg Lys Ile Ser
    210                 215                 220

cgg tca gta gct aag att tac ctt gga caa ctg gaa tgt ttc agt ttg   720
Arg Ser Val Ala Lys Ile Tyr Leu Gly Gln Leu Glu Cys Phe Ser Leu
225                 230                 235                 240

gga aat ctg gat gcc aaa cga gat tgg ggc cat gcc aag gac tat gtg   768
Gly Asn Leu Asp Ala Lys Arg Asp Trp Gly His Ala Lys Asp Tyr Val
                245                 250                 255

gag gct atg tgg ttg atg ttg cag aat gat gag ccg gag gac ttc gtt   816
Glu Ala Met Trp Leu Met Leu Gln Asn Asp Glu Pro Glu Asp Phe Val
            260                 265                 270

ata gct act ggg gag gtc cat agt gtc cgg gaa ttt gtc gag aaa tca   864
Ile Ala Thr Gly Glu Val His Ser Val Arg Glu Phe Val Glu Lys Ser
        275                 280                 285

ttc ttg cac att gga aaa acc att gtg tgg gaa gga aag aat gaa aat   912
Phe Leu His Ile Gly Lys Thr Ile Val Trp Glu Gly Lys Asn Glu Asn
    290                 295                 300

gaa gtg ggc aga tgt aaa gag acc ggc aaa gtt cac gtg act gtg gat   960
Glu Val Gly Arg Cys Lys Glu Thr Gly Lys Val His Val Thr Val Asp
305                 310                 315                 320

ctc aag tac tac cgg cca act gaa gtg gac ttt ctg cag ggc gac tgc  1008
Leu Lys Tyr Tyr Arg Pro Thr Glu Val Asp Phe Leu Gln Gly Asp Cys
                325                 330                 335

acc aaa gcg aaa cag aag ctg aac tgg aag ccc cgg gtc gct ttc gat  1056
Thr Lys Ala Lys Gln Lys Leu Asn Trp Lys Pro Arg Val Ala Phe Asp
            340                 345                 350

gag ctg gtg agg gag atg gtg cac gcc gac gtg gag ctc atg agg aca  1104
Glu Leu Val Arg Glu Met Val His Ala Asp Val Glu Leu Met Arg Thr
        355                 360                 365

aac ccc aat gcc tga                                             1119
Asn Pro Asn Ala  *
    370
```

<210> 2
<211> 372
<212> PRT
<213> Homo sapians

<400> 2

```
Met Ala His Ala Pro Ala Arg Cys Pro Ser Ala Arg Gly Ser Gly Asp
 1               5                  10                  15
Gly Glu Met Gly Lys Pro Arg Asn Val Ala Leu Ile Thr Gly Ile Thr
            20                  25                  30
Gly Gln Asp Gly Ser Tyr Leu Ala Glu Phe Leu Leu Glu Lys Gly Tyr
```

```
        35                  40                  45
Glu Val His Gly Ile Val Arg Arg Ser Ser Ser Phe Asn Thr Gly Arg
    50                  55                  60
Ile Glu His Leu Tyr Lys Asn Pro Gln Ala His Ile Glu Gly Asn Met
65                  70                  75                  80
Lys Leu His Tyr Gly Asp Leu Thr Asp Ser Thr Cys Leu Val Lys Ile
                85                  90                  95
Ile Asn Glu Val Lys Pro Thr Glu Ile Tyr Asn Leu Gly Ala Gln Ser
            100                 105                 110
His Val Lys Ile Ser Phe Asp Leu Ala Glu Tyr Thr Ala Asp Val Asp
        115                 120                 125
Gly Val Gly Thr Leu Arg Leu Leu Asp Ala Val Lys Thr Cys Gly Leu
    130                 135                 140
Ile Asn Ser Val Lys Phe Tyr Gln Ala Ser Thr Ser Glu Leu Tyr Gly
145                 150                 155                 160
Lys Val Gln Glu Ile Pro Gln Lys Glu Thr Thr Pro Phe Tyr Pro Arg
                165                 170                 175
Ser Pro Tyr Gly Ala Ala Lys Leu Tyr Ala Tyr Trp Ile Val Val Asn
            180                 185                 190
Phe Arg Glu Ala Tyr Asn Leu Phe Ala Val Asn Gly Ile Leu Phe Asn
        195                 200                 205
His Glu Ser Pro Arg Arg Gly Ala Asn Phe Val Thr Arg Lys Ile Ser
    210                 215                 220
Arg Ser Val Ala Lys Ile Tyr Leu Gly Gln Leu Glu Cys Phe Ser Leu
225                 230                 235                 240
Gly Asn Leu Asp Ala Lys Arg Asp Trp Gly His Ala Lys Asp Tyr Val
                245                 250                 255
Glu Ala Met Trp Leu Met Leu Gln Asn Asp Glu Pro Glu Asp Phe Val
            260                 265                 270
Ile Ala Thr Gly Glu Val His Ser Val Arg Glu Phe Val Glu Lys Ser
        275                 280                 285
Phe Leu His Ile Gly Lys Thr Ile Val Trp Glu Gly Lys Asn Glu Asn
    290                 295                 300
Glu Val Gly Arg Cys Lys Glu Thr Gly Lys Val His Val Thr Val Asp
305                 310                 315                 320
Leu Lys Tyr Tyr Arg Pro Thr Glu Val Asp Phe Leu Gln Gly Asp Cys
                325                 330                 335
Thr Lys Ala Lys Gln Lys Leu Asn Trp Lys Pro Arg Val Ala Phe Asp
            340                 345                 350
Glu Leu Val Arg Glu Met Val His Ala Asp Val Glu Leu Met Arg Thr
        355                 360                 365
Asn Pro Asn Ala
    370
```

<210> 3
<211> 966
<212> DNA
<213> Homo sapians

<220>
<221> CDS
<222> (1)...(966)
<223> GDP-ketoxy-deoxy-mannose-epimerase/GDP-keto-deoxy-galactose-reductase (FX protein)

<400> 3

```
atg ggt gaa ccc cag gga tcc atg cgg att cta gtg aca ggg ggc tct   48
Met Gly Glu Pro Gln Gly Ser Met Arg Ile Leu Val Thr Gly Gly Ser
 1               5                  10                  15

ggg ctg gta ggc aaa gcc atc cag aag gtg gta gca gat gga gct gga   96
Gly Leu Val Gly Lys Ala Ile Gln Lys Val Val Ala Asp Gly Ala Gly
            20                  25                  30

ctt cct gga gag gac tgg gtg ttt gtc tcc tct aaa gac gcc gat ctc   144
Leu Pro Gly Glu Asp Trp Val Phe Val Ser Ser Lys Asp Ala Asp Leu
        35                  40                  45
```

```
acg gat aca gca cag acc cgc gcc ctg ttt gag aag gtc caa ccc aca   192
Thr Asp Thr Ala Gln Thr Arg Ala Leu Phe Glu Lys Val Gln Pro Thr
     50                  55                  60

cac gtc atc cat ctt gct gca atg gtg ggg ggc ctg ttc cgg aat atc   240
His Val Ile His Leu Ala Ala Met Val Gly Gly Leu Phe Arg Asn Ile
 65                  70                  75                  80

aaa tac aat ttg gac ttc tgg agg aaa aac gtg cac atg aac gac aac   288
Lys Tyr Asn Leu Asp Phe Trp Arg Lys Asn Val His Met Asn Asp Asn
                 85                  90                  95

gtc ctg cac tcg gcc ttt gag gtg ggg gcc cgc aag gtg gtg tcc tgc   336
Val Leu His Ser Ala Phe Glu Val Gly Ala Arg Lys Val Val Ser Cys
            100                 105                 110

ctg tcc acc tgt atc ttc cct gac aag acg acc tac ccg ata gat gag   384
Leu Ser Thr Cys Ile Phe Pro Asp Lys Thr Thr Tyr Pro Ile Asp Glu
        115                 120                 125

acc atg atc cac aat ggg cct ccc cac aac agc aat ttt ggg tac tcg   432
Thr Met Ile His Asn Gly Pro Pro His Asn Ser Asn Phe Gly Tyr Ser
    130                 135                 140

tat gcc aag agg atg atc gac gtg cag aac agg gcc tac ttc cag cag   480
Tyr Ala Lys Arg Met Ile Asp Val Gln Asn Arg Ala Tyr Phe Gln Gln
145                 150                 155                 160

tac ggc tgc acc ttc acc gct gtc atc ccc acc aac gtt ttc ggg ccc   528
Tyr Gly Cys Thr Phe Thr Ala Val Ile Pro Thr Asn Val Phe Gly Pro
                165                 170                 175

cac gac aac ttc aac atc gag gat ggc cac gtg ctg cct ggc ctc atc   576
His Asp Asn Phe Asn Ile Glu Asp Gly His Val Leu Pro Gly Leu Ile
            180                 185                 190

cac aag gtg cac ctg gcc aag agc agc ggc tcg gcc ctg acg gtg tgg   624
His Lys Val His Leu Ala Lys Ser Ser Gly Ser Ala Leu Thr Val Trp
        195                 200                 205

ggt aca ggg aat ccg cgg agg cag ttc ata tac tcg ctg gac ctg gcc   672
Gly Thr Gly Asn Pro Arg Arg Gln Phe Ile Tyr Ser Leu Asp Leu Ala
    210                 215                 220

cag ctc ttt atc tgg gtc ctg cgg gag tac aat gaa gtg gag ccc atc   720
Gln Leu Phe Ile Trp Val Leu Arg Glu Tyr Asn Glu Val Glu Pro Ile
225                 230                 235                 240

atc ctc tcc gtg ggc gag gaa gat gag gtc tcc atc aag gag gca gcc   768
Ile Leu Ser Val Gly Glu Glu Asp Glu Val Ser Ile Lys Glu Ala Ala
                245                 250                 255

gag gcg gtg gtg gag gcc atg gac ttc cat ggg gaa gtc acc ttt gat   816
Glu Ala Val Val Glu Ala Met Asp Phe His Gly Glu Val Thr Phe Asp
            260                 265                 270

aca acc aag tcg gat ggg cag ttt aag aag aca gcc agt aac agc aag   864
Thr Thr Lys Ser Asp Gly Gln Phe Lys Lys Thr Ala Ser Asn Ser Lys
        275                 280                 285

ctg agg acc tac ctg ccc gac ttc cgg ttc aca ccc ttc aag cag gcg   912
Leu Arg Thr Tyr Leu Pro Asp Phe Arg Phe Thr Pro Phe Lys Gln Ala
    290                 295                 300

gtg aag gag acc tgt gct tgg ttc act gac aac tac gag cag gcc cgg   960
Val Lys Glu Thr Cys Ala Trp Phe Thr Asp Asn Tyr Glu Gln Ala Arg
305                 310                 315                 320
```

```
aag tga                                                                966
Lys  *
```

<210> 4
<211> 321
<212> PRT
<213> Homo sapians

<400> 4

```
Met Gly Glu Pro Gln Gly Ser Met Arg Ile Leu Val Thr Gly Gly Ser
 1               5                  10                  15
Gly Leu Val Gly Lys Ala Ile Gln Lys Val Val Ala Asp Gly Ala Gly
            20                  25                  30
Leu Pro Gly Glu Asp Trp Val Phe Val Ser Ser Lys Asp Ala Asp Leu
        35                  40                  45
Thr Asp Thr Ala Gln Thr Arg Ala Leu Phe Glu Lys Val Gln Pro Thr
    50                  55                  60
His Val Ile His Leu Ala Ala Met Val Gly Gly Leu Phe Arg Asn Ile
65                  70                  75                  80
Lys Tyr Asn Leu Asp Phe Trp Arg Lys Asn Val His Met Asn Asp Asn
                85                  90                  95
Val Leu His Ser Ala Phe Glu Val Gly Ala Arg Lys Val Val Ser Cys
            100                 105                 110
Leu Ser Thr Cys Ile Phe Pro Asp Lys Thr Thr Tyr Pro Ile Asp Glu
        115                 120                 125
Thr Met Ile His Asn Gly Pro Pro His Asn Ser Asn Phe Gly Tyr Ser
    130                 135                 140
Tyr Ala Lys Arg Met Ile Asp Val Gln Asn Arg Ala Tyr Phe Gln Gln
145                 150                 155                 160
Tyr Gly Cys Thr Phe Thr Ala Val Ile Pro Thr Asn Val Phe Gly Pro
                165                 170                 175
His Asp Asn Phe Asn Ile Glu Asp Gly His Val Leu Pro Gly Leu Ile
            180                 185                 190
His Lys Val His Leu Ala Lys Ser Ser Gly Ser Ala Leu Thr Val Trp
        195                 200                 205
Gly Thr Gly Asn Pro Arg Arg Gln Phe Ile Tyr Ser Leu Asp Leu Ala
    210                 215                 220
Gln Leu Phe Ile Trp Val Leu Arg Glu Tyr Asn Glu Val Glu Pro Ile
225                 230                 235                 240
Ile Leu Ser Val Gly Glu Glu Asp Glu Val Ser Ile Lys Glu Ala Ala
                245                 250                 255
Glu Ala Val Val Glu Ala Met Asp Phe His Gly Glu Val Thr Phe Asp
            260                 265                 270
Thr Thr Lys Ser Asp Gly Gln Phe Lys Lys Thr Ala Ser Asn Ser Lys
        275                 280                 285
Leu Arg Thr Tyr Leu Pro Asp Phe Arg Phe Thr Pro Phe Lys Gln Ala
    290                 295                 300
Val Lys Glu Thr Cys Ala Trp Phe Thr Asp Asn Tyr Glu Gln Ala Arg
305                 310                 315                 320
Lys
```

<210> 5
<211> 1095
<212> DNA
<213> Homo sapians

<220>
<221> CDS
<222> (1)...(1095)
<223> GDP-fucose transporter

<400> 5

```
atg aat agg gcc cct ctg aag cgg tcc agg atc ctg cac atg gcg ctg   48
```

```
Met Asn Arg Ala Pro Leu Lys Arg Ser Arg Ile Leu His Met Ala Leu
 1               5                  10                  15

acc ggg gcc tca gac ccc tct gca gag gca gag gcc aac ggg gag aag   96
Thr Gly Ala Ser Asp Pro Ser Ala Glu Ala Glu Ala Asn Gly Glu Lys
            20                  25                  30

ccc ttt ctg ctg cgg gca ttg cag atc gcg ctg gtg gtc tcc ctc tac  144
Pro Phe Leu Leu Arg Ala Leu Gln Ile Ala Leu Val Val Ser Leu Tyr
        35                  40                  45

tgg gtc acc tcc atc tcc atg gtg ttc ctt aat aag tac ctg ctg gac  192
Trp Val Thr Ser Ile Ser Met Val Phe Leu Asn Lys Tyr Leu Leu Asp
    50                  55                  60

agc ccc tcc ctg cgg ctg gac acc ccc atc ttc gtc acc ttc tac cag  240
Ser Pro Ser Leu Arg Leu Asp Thr Pro Ile Phe Val Thr Phe Tyr Gln
65                  70                  75                  80

tgc ctg gtg acc acg ctg ctg tgc aaa ggc ctc agc gct ctg gcc gcc  288
Cys Leu Val Thr Thr Leu Leu Cys Lys Gly Leu Ser Ala Leu Ala Ala
                85                  90                  95

tgc tgc cct ggt gcc gtg gac ttc ccc agc ttg cgc ctg gac ctc agg  336
Cys Cys Pro Gly Ala Val Asp Phe Pro Ser Leu Arg Leu Asp Leu Arg
            100                 105                 110

gtg gcc cgc agc gtc ctg ccc ctg tcg gtg gtc ttc atc ggc atg atc  384
Val Ala Arg Ser Val Leu Pro Leu Ser Val Val Phe Ile Gly Met Ile
        115                 120                 125

acc ttc aat aac ctc tgc ctc aag tac gtc ggt gtg gcc ttc tac aat  432
Thr Phe Asn Asn Leu Cys Leu Lys Tyr Val Gly Val Ala Phe Tyr Asn
    130                 135                 140

gtg ggc cgc tca ctc acc acc gtc ttc aac gtg ctg ctc tcc tac ctg  480
Val Gly Arg Ser Leu Thr Thr Val Phe Asn Val Leu Leu Ser Tyr Leu
145                 150                 155                 160

ctg ctc aag cag acc acc tcc ttc tat gcc ctg ctc acc tgc ggt atc  528
Leu Leu Lys Gln Thr Thr Ser Phe Tyr Ala Leu Leu Thr Cys Gly Ile
                165                 170                 175

atc atc ggg ggc ttc tgg ctt ggt gtg gac cag gag ggg gca gaa ggc  576
Ile Ile Gly Gly Phe Trp Leu Gly Val Asp Gln Glu Gly Ala Glu Gly
            180                 185                 190

acc ctg tcg tgg ctg ggc acc gtc ttc ggc gtg ctg gct agc ctc tgt  624
Thr Leu Ser Trp Leu Gly Thr Val Phe Gly Val Leu Ala Ser Leu Cys
        195                 200                 205

gtc tcg ctc aac gcc atc tac acc acg aag gtg ctc ccg gcg gtg gac  672
Val Ser Leu Asn Ala Ile Tyr Thr Thr Lys Val Leu Pro Ala Val Asp
    210                 215                 220

ggc agc atc tgg cgc ctg act ttc tac aac aac gtc aac gcc tgc atc  720
Gly Ser Ile Trp Arg Leu Thr Phe Tyr Asn Asn Val Asn Ala Cys Ile
225                 230                 235                 240

ctc ttc ctg ccc ctg ctc ctg ctg ctc ggg gag ctt cag gcc ctg cgt  768
Leu Phe Leu Pro Leu Leu Leu Leu Leu Gly Glu Leu Gln Ala Leu Arg
                245                 250                 255

gac ttt gcc cag ctg ggc agt gcc cac ttc tgg ggg atg atg acg ctg  816
Asp Phe Ala Gln Leu Gly Ser Ala His Phe Trp Gly Met Met Thr Leu
            260                 265                 270

ggc ggc ctg ttt ggc ttt gcc atc ggc tac gtg aca gga ctg cag atc  864
```

```
Gly Gly Leu Phe Gly Phe Ala Ile Gly Tyr Val Thr Gly Leu Gln Ile
        275                 280                 285

aag ttc acc agt ccg ctg acc cac aat gtg tcg ggc acg gcc aag gcc   912
Lys Phe Thr Ser Pro Leu Thr His Asn Val Ser Gly Thr Ala Lys Ala
    290                 295                 300

tgt gcc cag aca gtg ctg gcc gtg ctc tac tac gag gag acc aag agc   960
Cys Ala Gln Thr Val Leu Ala Val Leu Tyr Tyr Glu Glu Thr Lys Ser
305                 310                 315                 320

ttc ctc tgg tgg acg agc aac atg atg gtg ctg ggc ggc tcc tcc gcc   1008
Phe Leu Trp Trp Thr Ser Asn Met Met Val Leu Gly Gly Ser Ser Ala
                325                 330                 335

tac acc tgg gtc agg ggc tgg gag atg aag aag act ccg gag gag ccc   1056
Tyr Thr Trp Val Arg Gly Trp Glu Met Lys Lys Thr Pro Glu Glu Pro
            340                 345                 350

agc ccc aaa gac agc gag aag agc gcc atg ggg gtg tga               1095
Ser Pro Lys Asp Ser Glu Lys Ser Ala Met Gly Val  *
        355                 360


<210> 6
<211> 364
<212> PRT
<213> Homo sapians

<400> 6
Met Asn Arg Ala Pro Leu Lys Arg Ser Arg Ile Leu His Met Ala Leu
 1               5                  10                  15
Thr Gly Ala Ser Asp Pro Ser Ala Glu Ala Glu Ala Asn Gly Glu Lys
            20                  25                  30
Pro Phe Leu Leu Arg Ala Leu Gln Ile Ala Leu Val Val Ser Leu Tyr
        35                  40                  45
Trp Val Thr Ser Ile Ser Met Val Phe Leu Asn Lys Tyr Leu Leu Asp
    50                  55                  60
Ser Pro Ser Leu Arg Leu Asp Thr Pro Ile Phe Val Thr Phe Tyr Gln
65                  70                  75                  80
Cys Leu Val Thr Thr Leu Leu Cys Lys Gly Leu Ser Ala Leu Ala Ala
                85                  90                  95
Cys Cys Pro Gly Ala Val Asp Phe Pro Ser Leu Arg Leu Asp Leu Arg
            100                 105                 110
Val Ala Arg Ser Val Leu Pro Leu Ser Val Val Phe Ile Gly Met Ile
        115                 120                 125
Thr Phe Asn Asn Leu Cys Leu Lys Tyr Val Gly Val Ala Phe Tyr Asn
    130                 135                 140
Val Gly Arg Ser Leu Thr Thr Val Phe Asn Val Leu Leu Ser Tyr Leu
145                 150                 155                 160
Leu Leu Lys Gln Thr Thr Ser Phe Tyr Ala Leu Leu Thr Cys Gly Ile
                165                 170                 175
Ile Ile Gly Gly Phe Trp Leu Gly Val Asp Gln Glu Gly Ala Glu Gly
            180                 185                 190
Thr Leu Ser Trp Leu Gly Thr Val Phe Gly Val Leu Ala Ser Leu Cys
        195                 200                 205
Val Ser Leu Asn Ala Ile Tyr Thr Thr Lys Val Leu Pro Ala Val Asp
    210                 215                 220
Gly Ser Ile Trp Arg Leu Thr Phe Tyr Asn Asn Val Asn Ala Cys Ile
225                 230                 235                 240
Leu Phe Leu Pro Leu Leu Leu Leu Leu Gly Glu Leu Gln Ala Leu Arg
                245                 250                 255
Asp Phe Ala Gln Leu Gly Ser Ala His Phe Trp Gly Met Met Thr Leu
            260                 265                 270
Gly Gly Leu Phe Gly Phe Ala Ile Gly Tyr Val Thr Gly Leu Gln Ile
        275                 280                 285
Lys Phe Thr Ser Pro Leu Thr His Asn Val Ser Gly Thr Ala Lys Ala
    290                 295                 300
```

```
Cys Ala Gln Thr Val Leu Ala Val Leu Tyr Tyr Glu Glu Thr Lys Ser
305                 310                 315                 320
Phe Leu Trp Trp Thr Ser Asn Met Met Val Leu Gly Gly Ser Ser Ala
                325                 330                 335
Tyr Thr Trp Val Arg Gly Trp Glu Met Lys Lys Thr Pro Glu Glu Pro
            340                 345                 350
Ser Pro Lys Asp Ser Glu Lys Ser Ala Met Gly Val
        355                 360


<210> 7
<211> 1728
<212> DNA
<213> Homo sapians

<220>
<221> CDS
<222> (1)...(1728)
<223> alpha-1,6-fucosyltransferase (hFuT8)

<400> 7
atg cgg cca tgg act ggt tcc tgg cgt tgg att atg ctc att ctt ttt   48
Met Arg Pro Trp Thr Gly Ser Trp Arg Trp Ile Met Leu Ile Leu Phe
 1               5                  10                  15

gcc tgg ggg acc ttg ctg ttt tat ata ggt ggt cac ttg gta cga gat   96
Ala Trp Gly Thr Leu Leu Phe Tyr Ile Gly Gly His Leu Val Arg Asp
            20                  25                  30

aat gac cat cct gat cac tct agc cga gaa ctg tcc aag att ctg gca   144
Asn Asp His Pro Asp His Ser Ser Arg Glu Leu Ser Lys Ile Leu Ala
        35                  40                  45

aag ctt gaa cgc tta aaa caa cag aat gaa gac ttg agg cga atg gcc   192
Lys Leu Glu Arg Leu Lys Gln Gln Asn Glu Asp Leu Arg Arg Met Ala
    50                  55                  60

gaa tct ctc cgg ata cca gaa ggc cct att gat cag ggg cca gct ata   240
Glu Ser Leu Arg Ile Pro Glu Gly Pro Ile Asp Gln Gly Pro Ala Ile
65                  70                  75                  80

gga aga gta cgc gtt tta gaa gag cag ctt gtt aag gcc aaa gaa cag   288
Gly Arg Val Arg Val Leu Glu Glu Gln Leu Val Lys Ala Lys Glu Gln
                85                  90                  95

att gaa aat tac aag aaa cag acc aga aat ggt ctg ggg aag gat cat   336
Ile Glu Asn Tyr Lys Lys Gln Thr Arg Asn Gly Leu Gly Lys Asp His
            100                 105                 110

gaa atc ctg agg agg agg att gaa aat gga gct aaa gag ctc tgg ttt   384
Glu Ile Leu Arg Arg Arg Ile Glu Asn Gly Ala Lys Glu Leu Trp Phe
        115                 120                 125

ttc cta cag agt gaa ttg aag aaa tta aag aac tta gaa gga aat gaa   432
Phe Leu Gln Ser Glu Leu Lys Lys Leu Lys Asn Leu Glu Gly Asn Glu
    130                 135                 140

ctc caa aga cat gca gat gaa ttt ctt ttg gat tta gga cat cat gaa   480
Leu Gln Arg His Ala Asp Glu Phe Leu Leu Asp Leu Gly His His Glu
145                 150                 155                 160

agg tct ata atg acg gat cta tac tac ctc agt cag aca gat gga gca   528
Arg Ser Ile Met Thr Asp Leu Tyr Tyr Leu Ser Gln Thr Asp Gly Ala
                165                 170                 175

ggt gat tgg cgg gaa aaa gag gcc aaa gat ctg aca gaa ctg gtt cag   576
Gly Asp Trp Arg Glu Lys Glu Ala Lys Asp Leu Thr Glu Leu Val Gln
            180                 185                 190
```

```
cgg aga ata aca tat ctt cag aat ccc aag gac tgc agc aaa gcc aaa   624
Arg Arg Ile Thr Tyr Leu Gln Asn Pro Lys Asp Cys Ser Lys Ala Lys
        195                 200                 205

aag ctg gtg tgt aat atc aac aaa ggc tgt ggc tat ggc tgt cag ctc   672
Lys Leu Val Cys Asn Ile Asn Lys Gly Cys Gly Tyr Gly Cys Gln Leu
    210                 215                 220

cat cat gtg gtc tac tgc ttc atg att gca tat ggc acc cag cga aca   720
His His Val Val Tyr Cys Phe Met Ile Ala Tyr Gly Thr Gln Arg Thr
225                 230                 235                 240

ctc atc ttg gaa tct cag aat tgg cgc tat gct act ggt gga tgg gag   768
Leu Ile Leu Glu Ser Gln Asn Trp Arg Tyr Ala Thr Gly Gly Trp Glu
                245                 250                 255

act gta ttt agg cct gta agt gag aca tgc aca gac aga tct ggc atc   816
Thr Val Phe Arg Pro Val Ser Glu Thr Cys Thr Asp Arg Ser Gly Ile
            260                 265                 270

tcc act gga cac tgg tca ggt gaa gtg aag gac aaa aat gtt caa gtg   864
Ser Thr Gly His Trp Ser Gly Glu Val Lys Asp Lys Asn Val Gln Val
        275                 280                 285

gtc gag ctt ccc att gta gac agt ctt cat ccc cgt cct cca tat tta   912
Val Glu Leu Pro Ile Val Asp Ser Leu His Pro Arg Pro Pro Tyr Leu
    290                 295                 300

ccc ttg gct gta cca gaa gac ctc gca gat cga ctt gta cga gtg cat   960
Pro Leu Ala Val Pro Glu Asp Leu Ala Asp Arg Leu Val Arg Val His
305                 310                 315                 320

ggt gac cct gca gtg tgg tgg gtg tct cag ttt gtc aaa tac ttg atc  1008
Gly Asp Pro Ala Val Trp Trp Val Ser Gln Phe Val Lys Tyr Leu Ile
                325                 330                 335

cgc cca cag cct tgg cta gaa aaa gaa ata gaa gaa gcc acc aag aag  1056
Arg Pro Gln Pro Trp Leu Glu Lys Glu Ile Glu Glu Ala Thr Lys Lys
            340                 345                 350

ctt ggc ttc aaa cat cca gtt att gga gtc cat gtc aga cgc aca gac  1104
Leu Gly Phe Lys His Pro Val Ile Gly Val His Val Arg Arg Thr Asp
        355                 360                 365

aaa gtg gga aca gaa gct gcc ttc cat ccc att gaa gag tac atg gtg  1152
Lys Val Gly Thr Glu Ala Ala Phe His Pro Ile Glu Glu Tyr Met Val
    370                 375                 380

cat gtt gaa gaa cat ttt cag ctt ctt gca cgc aga atg caa gtg gac  1200
His Val Glu Glu His Phe Gln Leu Leu Ala Arg Arg Met Gln Val Asp
385                 390                 395                 400

aaa aaa aga gtg tat ttg gcc aca gat gac cct tct tta tta aag gag  1248
Lys Lys Arg Val Tyr Leu Ala Thr Asp Asp Pro Ser Leu Leu Lys Glu
                405                 410                 415

gca aaa aca aag tac ccc aat tat gaa ttt att agt gat aac tct att  1296
Ala Lys Thr Lys Tyr Pro Asn Tyr Glu Phe Ile Ser Asp Asn Ser Ile
            420                 425                 430

tcc tgg tca gct gga ctg cac aat cga tac aca gaa aat tca ctt cgt  1344
Ser Trp Ser Ala Gly Leu His Asn Arg Tyr Thr Glu Asn Ser Leu Arg
        435                 440                 445

gga gtg atc ctg gat ata cat ttt ctc tct cag gca gac ttc cta gtg  1392
Gly Val Ile Leu Asp Ile His Phe Leu Ser Gln Ala Asp Phe Leu Val
    450                 455                 460
```

```
tgt act ttt tca tcc cag gtc tgt cga gtt gct tat gaa att atg caa   1440
Cys Thr Phe Ser Ser Gln Val Cys Arg Val Ala Tyr Glu Ile Met Gln
465                 470                 475                 480

aca cta cat cct gat gcc tct gca aac ttc cat tct tta gat gac atc   1488
Thr Leu His Pro Asp Ala Ser Ala Asn Phe His Ser Leu Asp Asp Ile
                485                 490                 495

tac tat ttt ggg ggc cag aat gcc cac aat caa att gcc att tat gct   1536
Tyr Tyr Phe Gly Gly Gln Asn Ala His Asn Gln Ile Ala Ile Tyr Ala
            500                 505                 510

cac caa ccc cga act gca gat gaa att ccc atg gaa cct gga gat atc   1584
His Gln Pro Arg Thr Ala Asp Glu Ile Pro Met Glu Pro Gly Asp Ile
        515                 520                 525

att ggt gtg gct gga aat cat tgg gat ggc tat tct aaa ggt gtc aac   1632
Ile Gly Val Ala Gly Asn His Trp Asp Gly Tyr Ser Lys Gly Val Asn
    530                 535                 540

agg aaa ttg gga agg acg ggc cta tat ccc tcc tac aaa gtt cga gag   1680
Arg Lys Leu Gly Arg Thr Gly Leu Tyr Pro Ser Tyr Lys Val Arg Glu
545                 550                 555                 560

aag ata gaa acg gtc aag tac ccc aca tat cct gag gct gag aaa taa   1728
Lys Ile Glu Thr Val Lys Tyr Pro Thr Tyr Pro Glu Ala Glu Lys  *
                565                 570                 575
```

<210> 8
<211> 575
<212> PRT
<213> Homo sapians

<400> 8

```
Met Arg Pro Trp Thr Gly Ser Trp Arg Trp Ile Met Leu Ile Leu Phe
 1               5                  10                  15
Ala Trp Gly Thr Leu Leu Phe Tyr Ile Gly Gly His Leu Val Arg Asp
            20                  25                  30
Asn Asp His Pro Asp His Ser Ser Arg Glu Leu Ser Lys Ile Leu Ala
        35                  40                  45
Lys Leu Glu Arg Leu Lys Gln Gln Asn Glu Asp Leu Arg Arg Met Ala
    50                  55                  60
Glu Ser Leu Arg Ile Pro Glu Gly Pro Ile Asp Gln Gly Pro Ala Ile
65                  70                  75                  80
Gly Arg Val Arg Val Leu Glu Glu Gln Leu Val Lys Ala Lys Glu Gln
                85                  90                  95
Ile Glu Asn Tyr Lys Lys Gln Thr Arg Asn Gly Leu Gly Lys Asp His
            100                 105                 110
Glu Ile Leu Arg Arg Arg Ile Glu Asn Gly Ala Lys Glu Leu Trp Phe
        115                 120                 125
Phe Leu Gln Ser Glu Leu Lys Lys Leu Lys Asn Leu Glu Gly Asn Glu
    130                 135                 140
Leu Gln Arg His Ala Asp Glu Phe Leu Leu Asp Leu Gly His His Glu
145                 150                 155                 160
Arg Ser Ile Met Thr Asp Leu Tyr Tyr Leu Ser Gln Thr Asp Gly Ala
                165                 170                 175
Gly Asp Trp Arg Glu Lys Glu Ala Lys Asp Leu Thr Glu Leu Val Gln
            180                 185                 190
Arg Arg Ile Thr Tyr Leu Gln Asn Pro Lys Asp Cys Ser Lys Ala Lys
        195                 200                 205
Lys Leu Val Cys Asn Ile Asn Lys Gly Cys Gly Tyr Gly Cys Gln Leu
    210                 215                 220
His His Val Val Tyr Cys Phe Met Ile Ala Tyr Gly Thr Gln Arg Thr
225                 230                 235                 240
Leu Ile Leu Glu Ser Gln Asn Trp Arg Tyr Ala Thr Gly Gly Trp Glu
```

```
                245                 250                 255
Thr Val Phe Arg Pro Val Ser Glu Thr Cys Thr Asp Arg Ser Gly Ile
            260                 265                 270
Ser Thr Gly His Trp Ser Gly Glu Val Lys Asp Lys Asn Val Gln Val
        275                 280                 285
Val Glu Leu Pro Ile Val Asp Ser Leu His Pro Arg Pro Pro Tyr Leu
    290                 295                 300
Pro Leu Ala Val Pro Glu Asp Leu Ala Asp Arg Leu Val Arg Val His
305                 310                 315                 320
Gly Asp Pro Ala Val Trp Trp Val Ser Gln Phe Val Lys Tyr Leu Ile
                325                 330                 335
Arg Pro Gln Pro Trp Leu Glu Lys Glu Ile Glu Glu Ala Thr Lys Lys
            340                 345                 350
Leu Gly Phe Lys His Pro Val Ile Gly Val His Val Arg Arg Thr Asp
        355                 360                 365
Lys Val Gly Thr Glu Ala Ala Phe His Pro Ile Glu Glu Tyr Met Val
    370                 375                 380
His Val Glu Glu His Phe Gln Leu Leu Ala Arg Arg Met Gln Val Asp
385                 390                 395                 400
Lys Lys Arg Val Tyr Leu Ala Thr Asp Asp Pro Ser Leu Leu Lys Glu
                405                 410                 415
Ala Lys Thr Lys Tyr Pro Asn Tyr Glu Phe Ile Ser Asp Asn Ser Ile
            420                 425                 430
Ser Trp Ser Ala Gly Leu His Asn Arg Tyr Thr Glu Asn Ser Leu Arg
        435                 440                 445
Gly Val Ile Leu Asp Ile His Phe Leu Ser Gln Ala Asp Phe Leu Val
    450                 455                 460
Cys Thr Phe Ser Ser Gln Val Cys Arg Val Ala Tyr Glu Ile Met Gln
465                 470                 475                 480
Thr Leu His Pro Asp Ala Ser Ala Asn Phe His Ser Leu Asp Asp Ile
                485                 490                 495
Tyr Tyr Phe Gly Gly Gln Asn Ala His Asn Gln Ile Ala Ile Tyr Ala
            500                 505                 510
His Gln Pro Arg Thr Ala Asp Glu Ile Pro Met Glu Pro Gly Asp Ile
        515                 520                 525
Ile Gly Val Ala Gly Asn His Trp Asp Gly Tyr Ser Lys Gly Val Asn
    530                 535                 540
Arg Lys Leu Gly Arg Thr Gly Leu Tyr Pro Ser Tyr Lys Val Arg Glu
545                 550                 555                 560
Lys Ile Glu Thr Val Lys Tyr Pro Thr Tyr Pro Glu Ala Glu Lys
                565                 570                 575
```

<210> 9
<211> 1728
<212> DNA
<213> Rattus norvegicus

<220>
<221> CDS
<222> (1)...(1728)
<223> alpha-1,6-fucosyltransferase (rFuT8)

<400> 9

```
atg cgg gca tgg act ggt tcc tgg cgt tgg att atg ctc att ctt ttt   48
Met Arg Ala Trp Thr Gly Ser Trp Arg Trp Ile Met Leu Ile Leu Phe
 1               5                  10                  15

gcc tgg ggg acc ttg ttg ttt tat ata ggt ggt cat ttg gtt cga gat   96
Ala Trp Gly Thr Leu Leu Phe Tyr Ile Gly Gly His Leu Val Arg Asp
            20                  25                  30

aat gac cac cct gat cac tct agc aga gaa ctc tcc aag att ctt gca   144
Asn Asp His Pro Asp His Ser Ser Arg Glu Leu Ser Lys Ile Leu Ala
        35                  40                  45

aag ctt gaa cgc tta aaa caa caa aat gaa gac ttg agg cga atg gct   192
Lys Leu Glu Arg Leu Lys Gln Gln Asn Glu Asp Leu Arg Arg Met Ala
```

```
     50                  55                  60

gag tct cta cga ata cca gaa ggc ccc att gac cag ggg acg gct acg   240
Glu Ser Leu Arg Ile Pro Glu Gly Pro Ile Asp Gln Gly Thr Ala Thr
 65                  70                  75                  80

gga aga gtc cgt gtt tta gaa gaa cag ctt gtt aag gcc aaa gaa cag   288
Gly Arg Val Arg Val Leu Glu Glu Gln Leu Val Lys Ala Lys Glu Gln
                 85                  90                  95

att gaa aat tac aag aaa caa gcc aga aat ggt ctg ggg aag gat cat   336
Ile Glu Asn Tyr Lys Lys Gln Ala Arg Asn Gly Leu Gly Lys Asp His
            100                 105                 110

gaa ctc tta agg agg agg att gaa aat gga gct aaa gag ctc tgg ttt   384
Glu Leu Leu Arg Arg Arg Ile Glu Asn Gly Ala Lys Glu Leu Trp Phe
        115                 120                 125

ttt cta caa agt gaa ctg aag aaa tta aag cat cta gaa gga aat gaa   432
Phe Leu Gln Ser Glu Leu Lys Lys Leu Lys His Leu Glu Gly Asn Glu
    130                 135                 140

ctc caa aga cat gca gat gaa att ctt ttg gat tta gga cac cat gaa   480
Leu Gln Arg His Ala Asp Glu Ile Leu Leu Asp Leu Gly His His Glu
145                 150                 155                 160

agg tct atc atg acg gat cta tac tac ctc agt caa aca gat gga gca   528
Arg Ser Ile Met Thr Asp Leu Tyr Tyr Leu Ser Gln Thr Asp Gly Ala
                165                 170                 175

ggg gat tgg cgt gaa aaa gag gcc aaa gat ctg aca gag ctg gtc cag   576
Gly Asp Trp Arg Glu Lys Glu Ala Lys Asp Leu Thr Glu Leu Val Gln
            180                 185                 190

cgg aga ata act tat ctc cag aat ccc aag gac tgc agc aaa gcc agg   624
Arg Arg Ile Thr Tyr Leu Gln Asn Pro Lys Asp Cys Ser Lys Ala Arg
        195                 200                 205

aag ctg gtg tgt aac atc aat aag ggc tgt ggc tat ggt tgc caa ctc   672
Lys Leu Val Cys Asn Ile Asn Lys Gly Cys Gly Tyr Gly Cys Gln Leu
    210                 215                 220

cat cac gtg gtc tac tgt ttc atg att gct tat ggc acc cag cga aca   720
His His Val Val Tyr Cys Phe Met Ile Ala Tyr Gly Thr Gln Arg Thr
225                 230                 235                 240

ctc atc ttg gaa tct cag aat tgg cgc tat gct act ggt gga tgg gag   768
Leu Ile Leu Glu Ser Gln Asn Trp Arg Tyr Ala Thr Gly Gly Trp Glu
                245                 250                 255

act gtg ttt aga cct gta agt gag aca tgc aca gac aga tct ggc ctc   816
Thr Val Phe Arg Pro Val Ser Glu Thr Cys Thr Asp Arg Ser Gly Leu
            260                 265                 270

tcc act gga cac tgg tca ggt gaa gtg aat gac aaa aat att caa gtg   864
Ser Thr Gly His Trp Ser Gly Glu Val Asn Asp Lys Asn Ile Gln Val
        275                 280                 285

gtg gag ctc ccc att gta gac agc ctc cat cct cgg cct cct tac tta   912
Val Glu Leu Pro Ile Val Asp Ser Leu His Pro Arg Pro Pro Tyr Leu
    290                 295                 300

cca ctg gct gtt cca gaa gac ctt gca gat cga ctc gta aga gtc cat   960
Pro Leu Ala Val Pro Glu Asp Leu Ala Asp Arg Leu Val Arg Val His
305                 310                 315                 320

ggt gat cct gca gtg tgg tgg gtg tcc cag ttc gtc aaa tat ttg att   1008
Gly Asp Pro Ala Val Trp Trp Val Ser Gln Phe Val Lys Tyr Leu Ile
```

```
                325                 330                 335

cgt cca caa cct tgg cta gaa aag gaa ata gaa gaa gcc acc aag aag   1056
Arg Pro Gln Pro Trp Leu Glu Lys Glu Ile Glu Glu Ala Thr Lys Lys
            340                 345                 350

ctt ggc ttc aaa cat cca gtc att gga gtc cat gtc aga cgc aca gac   1104
Leu Gly Phe Lys His Pro Val Ile Gly Val His Val Arg Arg Thr Asp
        355                 360                 365

aaa gtg gga aca gag gca gcc ttc cat ccc atc gaa gag tac atg gta   1152
Lys Val Gly Thr Glu Ala Ala Phe His Pro Ile Glu Glu Tyr Met Val
    370                 375                 380

cat gtt gaa gaa cat ttt cag ctt ctc gca cgc aga atg caa gtg gat   1200
His Val Glu Glu His Phe Gln Leu Leu Ala Arg Arg Met Gln Val Asp
385                 390                 395                 400

aaa aaa aga gta tat ctg gct acc gat gac cct gct ttg tta aag gag   1248
Lys Lys Arg Val Tyr Leu Ala Thr Asp Asp Pro Ala Leu Leu Lys Glu
                405                 410                 415

gca aag aca aag tac tcc aat tat gaa ttt att agt gat aac tct att   1296
Ala Lys Thr Lys Tyr Ser Asn Tyr Glu Phe Ile Ser Asp Asn Ser Ile
            420                 425                 430

tct tgg tca gct gga tta cac aat cgg tac aca gaa aat tca ctt cgg   1344
Ser Trp Ser Ala Gly Leu His Asn Arg Tyr Thr Glu Asn Ser Leu Arg
        435                 440                 445

ggc gtg atc ctg gat ata cac ttt ctc tct cag gct gac ttc cta gtg   1392
Gly Val Ile Leu Asp Ile His Phe Leu Ser Gln Ala Asp Phe Leu Val
    450                 455                 460

tgt act ttt tca tcc cag gtc tgt cgg gtt gct tat gaa atc atg caa   1440
Cys Thr Phe Ser Ser Gln Val Cys Arg Val Ala Tyr Glu Ile Met Gln
465                 470                 475                 480

acc ctg cat cct gat gcc tct gca aac ttc cac tct tta gat gac atc   1488
Thr Leu His Pro Asp Ala Ser Ala Asn Phe His Ser Leu Asp Asp Ile
                485                 490                 495

tac tat ttt gga ggc caa aat gcc cac aac cag att gcc gtt tat cct   1536
Tyr Tyr Phe Gly Gly Gln Asn Ala His Asn Gln Ile Ala Val Tyr Pro
            500                 505                 510

cac aaa cct cga act gat gag gaa att cca atg gaa cct gga gat atc   1584
His Lys Pro Arg Thr Asp Glu Glu Ile Pro Met Glu Pro Gly Asp Ile
        515                 520                 525

att ggt gtg gct gga aac cat tgg gat ggt tat tct aaa ggt gtc aac   1632
Ile Gly Val Ala Gly Asn His Trp Asp Gly Tyr Ser Lys Gly Val Asn
    530                 535                 540

aga aaa ctt gga aaa aca ggc tta tat ccc tcc tac aaa gtc cga gag   1680
Arg Lys Leu Gly Lys Thr Gly Leu Tyr Pro Ser Tyr Lys Val Arg Glu
545                 550                 555                 560

aag ata gaa aca gtc aag tat ccc aca tat cct gaa gct gaa aaa tag   1728
Lys Ile Glu Thr Val Lys Tyr Pro Thr Tyr Pro Glu Ala Glu Lys  *
                565                 570                 575
```

<210> 10
<211> 575
<212> PRT
<213> Rattus norvegicus

<400> 10

```
Met Arg Ala Trp Thr Gly Ser Trp Arg Trp Ile Met Leu Ile Leu Phe
 1               5                  10                  15
Ala Trp Gly Thr Leu Leu Phe Tyr Ile Gly Gly His Leu Val Arg Asp
            20                  25                  30
Asn Asp His Pro Asp His Ser Ser Arg Glu Leu Ser Lys Ile Leu Ala
        35                  40                  45
Lys Leu Glu Arg Leu Lys Gln Gln Asn Glu Asp Leu Arg Arg Met Ala
    50                  55                  60
Glu Ser Leu Arg Ile Pro Glu Gly Pro Ile Asp Gln Gly Thr Ala Thr
65                  70                  75                  80
Gly Arg Val Arg Val Leu Glu Glu Gln Leu Val Lys Ala Lys Glu Gln
                85                  90                  95
Ile Glu Asn Tyr Lys Lys Gln Ala Arg Asn Gly Leu Gly Lys Asp His
            100                 105                 110
Glu Leu Leu Arg Arg Arg Ile Glu Asn Gly Ala Lys Glu Leu Trp Phe
        115                 120                 125
Phe Leu Gln Ser Glu Leu Lys Leu Lys His Leu Glu Gly Asn Glu
    130                 135                 140
Leu Gln Arg His Ala Asp Glu Ile Leu Leu Asp Leu Gly His His Glu
145                 150                 155                 160
Arg Ser Ile Met Thr Asp Leu Tyr Tyr Leu Ser Gln Thr Asp Gly Ala
                165                 170                 175
Gly Asp Trp Arg Glu Lys Glu Ala Lys Asp Leu Thr Glu Leu Val Gln
            180                 185                 190
Arg Arg Ile Thr Tyr Leu Gln Asn Pro Lys Asp Cys Ser Lys Ala Arg
        195                 200                 205
Lys Leu Val Cys Asn Ile Asn Lys Gly Cys Gly Tyr Gly Cys Gln Leu
    210                 215                 220
His His Val Val Tyr Cys Phe Met Ile Ala Tyr Gly Thr Gln Arg Thr
225                 230                 235                 240
Leu Ile Leu Glu Ser Gln Asn Trp Arg Tyr Ala Thr Gly Gly Trp Glu
                245                 250                 255
Thr Val Phe Arg Pro Val Ser Glu Thr Cys Thr Asp Arg Ser Gly Leu
            260                 265                 270
Ser Thr Gly His Trp Ser Gly Glu Val Asn Asp Lys Asn Ile Gln Val
        275                 280                 285
Val Glu Leu Pro Ile Val Asp Ser Leu His Pro Arg Pro Pro Tyr Leu
    290                 295                 300
Pro Leu Ala Val Pro Glu Asp Leu Ala Asp Arg Leu Val Arg Val His
305                 310                 315                 320
Gly Asp Pro Ala Val Trp Trp Val Ser Gln Phe Val Lys Tyr Leu Ile
                325                 330                 335
Arg Pro Gln Pro Trp Leu Glu Lys Glu Ile Glu Glu Ala Thr Lys Lys
            340                 345                 350
Leu Gly Phe Lys His Pro Val Ile Gly Val His Val Arg Arg Thr Asp
        355                 360                 365
Lys Val Gly Thr Glu Ala Ala Phe His Pro Ile Glu Glu Tyr Met Val
    370                 375                 380
His Val Glu Glu His Phe Gln Leu Leu Ala Arg Arg Met Gln Val Asp
385                 390                 395                 400
Lys Lys Arg Val Tyr Leu Ala Thr Asp Asp Pro Ala Leu Leu Lys Glu
                405                 410                 415
Ala Lys Thr Lys Tyr Ser Asn Tyr Glu Phe Ile Ser Asp Asn Ser Ile
            420                 425                 430
Ser Trp Ser Ala Gly Leu His Asn Arg Tyr Thr Glu Asn Ser Leu Arg
        435                 440                 445
Gly Val Ile Leu Asp Ile His Phe Leu Ser Gln Ala Asp Phe Leu Val
    450                 455                 460
Cys Thr Phe Ser Ser Gln Val Cys Arg Val Ala Tyr Glu Ile Met Gln
465                 470                 475                 480
Thr Leu His Pro Asp Ala Ser Ala Asn Phe His Ser Leu Asp Asp Ile
                485                 490                 495
Tyr Tyr Phe Gly Gly Gln Asn Ala His Asn Gln Ile Ala Val Tyr Pro
            500                 505                 510
His Lys Pro Arg Thr Asp Glu Glu Ile Pro Met Glu Pro Gly Asp Ile
        515                 520                 525
```

```
Ile Gly Val Ala Gly Asn His Trp Asp Gly Tyr Ser Lys Gly Val Asn
    530                 535                 540
Arg Lys Leu Gly Lys Thr Gly Leu Tyr Pro Ser Tyr Lys Val Arg Glu
545                 550                 555                 560
Lys Ile Glu Thr Val Lys Tyr Pro Thr Tyr Pro Glu Ala Glu Lys
                565                 570                 575
```

<210> 11
<211> 1728
<212> DNA
<213> Mus musculus

<220>
<221> CDS
<222> (1)...(1728)
<223> alpha-1,6-fucosyltransferase (mFuT8)

<400> 11

```
atg cgg gca tgg act ggt tcc tgg cgt tgg att atg ctc att ctt ttt   48
Met Arg Ala Trp Thr Gly Ser Trp Arg Trp Ile Met Leu Ile Leu Phe
 1               5                  10                  15

gcc tgg ggg acc ttg tta ttt tat ata ggt ggt cat ttg gtt cga gat   96
Ala Trp Gly Thr Leu Leu Phe Tyr Ile Gly Gly His Leu Val Arg Asp
            20                  25                  30

aat gac cac cct gat cac tcc agc aga gaa ctc tcc aag att ctt gca   144
Asn Asp His Pro Asp His Ser Ser Arg Glu Leu Ser Lys Ile Leu Ala
        35                  40                  45

aag ctt gaa cgc tta aaa cag caa aat gaa gac ttg agg cga atg gct   192
Lys Leu Glu Arg Leu Lys Gln Gln Asn Glu Asp Leu Arg Arg Met Ala
    50                  55                  60

gag tct ctc cga ata cca gaa ggc ccc att gac cag ggg aca gct aca   240
Glu Ser Leu Arg Ile Pro Glu Gly Pro Ile Asp Gln Gly Thr Ala Thr
65                  70                  75                  80

gga aga gtc cgt gtt tta gaa gaa cag ctt gtt aag gcc aaa gaa cag   288
Gly Arg Val Arg Val Leu Glu Glu Gln Leu Val Lys Ala Lys Glu Gln
                85                  90                  95

att gaa aat tac aag aaa caa gct aga aat ggt ctg ggg aag gat cat   336
Ile Glu Asn Tyr Lys Lys Gln Ala Arg Asn Gly Leu Gly Lys Asp His
            100                 105                 110

gaa atc tta aga agg agg att gaa aat gga gct aaa gag ctc tgg ttt   384
Glu Ile Leu Arg Arg Arg Ile Glu Asn Gly Ala Lys Glu Leu Trp Phe
        115                 120                 125

ttt cta caa agc gaa ctg aag aaa tta aag cat tta gaa gga aat gaa   432
Phe Leu Gln Ser Glu Leu Lys Lys Leu Lys His Leu Glu Gly Asn Glu
    130                 135                 140

ctc caa aga cat gca gat gaa att ctt ttg gat tta gga cac cat gaa   480
Leu Gln Arg His Ala Asp Glu Ile Leu Leu Asp Leu Gly His His Glu
145                 150                 155                 160

agg tct atc atg aca gat cta tac tac ctc agt caa aca gat gga gca   528
Arg Ser Ile Met Thr Asp Leu Tyr Tyr Leu Ser Gln Thr Asp Gly Ala
                165                 170                 175

ggg gat tgg cgt gaa aaa gag gcc aaa gat ctg aca gag ctg gtc cag   576
Gly Asp Trp Arg Glu Lys Glu Ala Lys Asp Leu Thr Glu Leu Val Gln
            180                 185                 190

cgg aga ata aca tat ctc cag aat cct aag gac tgc agc aaa gcc agg   624
```

```
Arg Arg Ile Thr Tyr Leu Gln Asn Pro Lys Asp Cys Ser Lys Ala Arg
        195                 200                 205

aag ctg gtg tgt aac atc aat aaa ggc tgt ggc tat ggt tgt caa ctc   672
Lys Leu Val Cys Asn Ile Asn Lys Gly Cys Gly Tyr Gly Cys Gln Leu
    210                 215                 220

cat cac gtg gtc tac tgt ttc atg att gct tat ggc acc cag cga aca   720
His His Val Val Tyr Cys Phe Met Ile Ala Tyr Gly Thr Gln Arg Thr
225                 230                 235                 240

ctc atc ttg gaa tct cag aat tgg cgc tat gct act ggt gga tgg gag   768
Leu Ile Leu Glu Ser Gln Asn Trp Arg Tyr Ala Thr Gly Gly Trp Glu
                245                 250                 255

act gtg ttt aga cct gta agt gag aca tgt aca gac aga tct ggc ctc   816
Thr Val Phe Arg Pro Val Ser Glu Thr Cys Thr Asp Arg Ser Gly Leu
            260                 265                 270

tcc act gga cac tgg tca ggt gaa gta aat gac aaa aac att caa gtg   864
Ser Thr Gly His Trp Ser Gly Glu Val Asn Asp Lys Asn Ile Gln Val
        275                 280                 285

gtc gag ctc ccc att gta gac agc ctc cat cct cgg cct cct tac tta   912
Val Glu Leu Pro Ile Val Asp Ser Leu His Pro Arg Pro Pro Tyr Leu
    290                 295                 300

cca ctg gct gtt cca gaa gac ctt gca gac cga ctc cta aga gtc cat   960
Pro Leu Ala Val Pro Glu Asp Leu Ala Asp Arg Leu Leu Arg Val His
305                 310                 315                 320

ggt gac cct gca gtg tgg tgg gtg tcc cag ttt gtc aaa tac ttg att  1008
Gly Asp Pro Ala Val Trp Trp Val Ser Gln Phe Val Lys Tyr Leu Ile
                325                 330                 335

cgt cca caa cct tgg ctg gaa aag gaa ata gaa gaa gcc acc aag aag  1056
Arg Pro Gln Pro Trp Leu Glu Lys Glu Ile Glu Glu Ala Thr Lys Lys
            340                 345                 350

ctt ggc ttc aaa cat cca gtt att gga gtc cat gtc aga cgc aca gac  1104
Leu Gly Phe Lys His Pro Val Ile Gly Val His Val Arg Arg Thr Asp
        355                 360                 365

aaa gtg gga aca gaa gca gcc ttc cac ccc atc gag gag tac atg gta  1152
Lys Val Gly Thr Glu Ala Ala Phe His Pro Ile Glu Glu Tyr Met Val
    370                 375                 380

cac gtt gaa gaa cat ttt cag ctt ctc gca cgc aga atg caa gtg gat  1200
His Val Glu Glu His Phe Gln Leu Leu Ala Arg Arg Met Gln Val Asp
385                 390                 395                 400

aaa aaa aga gta tat ctg gct act gat gat cct act ttg tta aag gag  1248
Lys Lys Arg Val Tyr Leu Ala Thr Asp Asp Pro Thr Leu Leu Lys Glu
                405                 410                 415

gca aag aca aag tac tcc aat tat gaa ttt att agt gat aac tct att  1296
Ala Lys Thr Lys Tyr Ser Asn Tyr Glu Phe Ile Ser Asp Asn Ser Ile
            420                 425                 430

tct tgg tca gct gga cta cac aat cgg tac aca gaa aat tca ctt cgg  1344
Ser Trp Ser Ala Gly Leu His Asn Arg Tyr Thr Glu Asn Ser Leu Arg
        435                 440                 445

ggt gtg atc ctg gat ata cac ttt ctc tca cag gct gac ttt cta gtg  1392
Gly Val Ile Leu Asp Ile His Phe Leu Ser Gln Ala Asp Phe Leu Val
    450                 455                 460

tgt act ttt tca tcc cag gtc tgt cgg gtt gct tat gaa atc atg caa  1440
```

```
Cys Thr Phe Ser Ser Gln Val Cys Arg Val Ala Tyr Glu Ile Met Gln
465                 470                 475                 480

acc ctg cat cct gat gcc tct gcg aac ttc cat tct ttg gat gac atc   1488
Thr Leu His Pro Asp Ala Ser Ala Asn Phe His Ser Leu Asp Asp Ile
                485                 490                 495

tac tat ttt gga ggc caa aat gcc cac aat cag att gct gtt tat cct   1536
Tyr Tyr Phe Gly Gly Gln Asn Ala His Asn Gln Ile Ala Val Tyr Pro
            500                 505                 510

cac aaa cct cga act gaa gag gaa att cca atg gaa cct gga gat atc   1584
His Lys Pro Arg Thr Glu Glu Glu Ile Pro Met Glu Pro Gly Asp Ile
        515                 520                 525

att ggt gtg gct gga aac cat tgg gat ggt tat tct aaa ggt atc aac   1632
Ile Gly Val Ala Gly Asn His Trp Asp Gly Tyr Ser Lys Gly Ile Asn
    530                 535                 540

aga aaa ctt gga aaa aca ggc tta tat ccc tcc tac aaa gtc cga gag   1680
Arg Lys Leu Gly Lys Thr Gly Leu Tyr Pro Ser Tyr Lys Val Arg Glu
545                 550                 555                 560

aag ata gaa aca gtc aag tat ccc aca tat cct gaa gct gaa aaa tag   1728
Lys Ile Glu Thr Val Lys Tyr Pro Thr Tyr Pro Glu Ala Glu Lys  *
                565                 570                 575
```

<210> 12
<211> 575
<212> PRT
<213> Mus musculus

<400> 12

```
Met Arg Ala Trp Thr Gly Ser Trp Arg Trp Ile Met Leu Ile Leu Phe
 1               5                  10                  15
Ala Trp Gly Thr Leu Leu Phe Tyr Ile Gly Gly His Leu Val Arg Asp
            20                  25                  30
Asn Asp His Pro Asp His Ser Ser Arg Glu Leu Ser Lys Ile Leu Ala
        35                  40                  45
Lys Leu Glu Arg Leu Lys Gln Gln Asn Glu Asp Leu Arg Arg Met Ala
    50                  55                  60
Glu Ser Leu Arg Ile Pro Glu Gly Pro Ile Asp Gln Gly Thr Ala Thr
65                  70                  75                  80
Gly Arg Val Arg Val Leu Glu Glu Gln Leu Val Lys Ala Lys Glu Gln
                85                  90                  95
Ile Glu Asn Tyr Lys Lys Gln Ala Arg Asn Gly Leu Gly Lys Asp His
            100                 105                 110
Glu Ile Leu Arg Arg Arg Ile Glu Asn Gly Ala Lys Glu Leu Trp Phe
        115                 120                 125
Phe Leu Gln Ser Glu Leu Lys Lys Leu Lys His Leu Glu Gly Asn Glu
    130                 135                 140
Leu Gln Arg His Ala Asp Glu Ile Leu Leu Asp Leu Gly His His Glu
145                 150                 155                 160
Arg Ser Ile Met Thr Asp Leu Tyr Tyr Leu Ser Gln Thr Asp Gly Ala
                165                 170                 175
Gly Asp Trp Arg Glu Lys Glu Ala Lys Asp Leu Thr Glu Leu Val Gln
            180                 185                 190
Arg Arg Ile Thr Tyr Leu Gln Asn Pro Lys Asp Cys Ser Lys Ala Arg
        195                 200                 205
Lys Leu Val Cys Asn Ile Asn Lys Gly Cys Gly Tyr Gly Cys Gln Leu
    210                 215                 220
His His Val Val Tyr Cys Phe Met Ile Ala Tyr Gly Thr Gln Arg Thr
225                 230                 235                 240
Leu Ile Leu Glu Ser Gln Asn Trp Arg Tyr Ala Thr Gly Gly Trp Glu
                245                 250                 255
Thr Val Phe Arg Pro Val Ser Glu Thr Cys Thr Asp Arg Ser Gly Leu
```

```
            260                 265                 270
Ser Thr Gly His Trp Ser Gly Glu Val Asn Asp Lys Asn Ile Gln Val
        275                 280                 285
Val Glu Leu Pro Ile Val Asp Ser Leu His Pro Arg Pro Pro Tyr Leu
    290                 295                 300
Pro Leu Ala Val Pro Glu Asp Leu Ala Asp Arg Leu Leu Arg Val His
305                 310                 315                 320
Gly Asp Pro Ala Val Trp Trp Val Ser Gln Phe Val Lys Tyr Leu Ile
                325                 330                 335
Arg Pro Gln Pro Trp Leu Glu Lys Glu Ile Glu Glu Ala Thr Lys Lys
            340                 345                 350
Leu Gly Phe Lys His Pro Val Ile Gly Val His Val Arg Arg Thr Asp
        355                 360                 365
Lys Val Gly Thr Glu Ala Ala Phe His Pro Ile Glu Glu Tyr Met Val
    370                 375                 380
His Val Glu Glu His Phe Gln Leu Leu Ala Arg Arg Met Gln Val Asp
385                 390                 395                 400
Lys Lys Arg Val Tyr Leu Ala Thr Asp Asp Pro Thr Leu Leu Lys Glu
                405                 410                 415
Ala Lys Thr Lys Tyr Ser Asn Tyr Glu Phe Ile Ser Asp Asn Ser Ile
            420                 425                 430
Ser Trp Ser Ala Gly Leu His Asn Arg Tyr Thr Glu Asn Ser Leu Arg
        435                 440                 445
Gly Val Ile Leu Asp Ile His Phe Leu Ser Gln Ala Asp Phe Leu Val
    450                 455                 460
Cys Thr Phe Ser Ser Gln Val Cys Arg Val Ala Tyr Glu Ile Met Gln
465                 470                 475                 480
Thr Leu His Pro Asp Ala Ser Ala Asn Phe His Ser Leu Asp Asp Ile
                485                 490                 495
Tyr Tyr Phe Gly Gly Gln Asn Ala His Asn Gln Ile Ala Val Tyr Pro
            500                 505                 510
His Lys Pro Arg Thr Glu Glu Glu Ile Pro Met Glu Pro Gly Asp Ile
        515                 520                 525
Ile Gly Val Ala Gly Asn His Trp Asp Gly Tyr Ser Lys Gly Ile Asn
    530                 535                 540
Arg Lys Leu Gly Lys Thr Gly Leu Tyr Pro Ser Tyr Lys Val Arg Glu
545                 550                 555                 560
Lys Ile Glu Thr Val Lys Tyr Pro Thr Tyr Pro Glu Ala Glu Lys
                565                 570                 575


<210> 13
<211> 1728
<212> DNA
<213> Sus scrofa

<220>
<221> CDS
<222> (1)...(1728)
<223> alpha-1,6-fucosyltransferase (pFuT8)

<400> 13
atg cgg cca tgg act ggt tcg tgg cgt tgg att atg ctc att ctt ttt   48
Met Arg Pro Trp Thr Gly Ser Trp Arg Trp Ile Met Leu Ile Leu Phe
 1               5                  10                  15

gcc tgg ggg acc ttg cta ttt tac ata ggt ggt cac ttg gta cga gat   96
Ala Trp Gly Thr Leu Leu Phe Tyr Ile Gly Gly His Leu Val Arg Asp
            20                  25                  30

aat gac cac tct gat cac tct agc cga gaa ctg tcc aag att ttg gca   144
Asn Asp His Ser Asp His Ser Ser Arg Glu Leu Ser Lys Ile Leu Ala
        35                  40                  45

aag ctg gaa cgc tta aaa caa caa aat gaa gac ttg agg aga atg gct   192
Lys Leu Glu Arg Leu Lys Gln Gln Asn Glu Asp Leu Arg Arg Met Ala
    50                  55                  60
```

```
gaa tct ctc cga ata cca gaa ggc ccc att gat cag ggg cca gct tca   240
Glu Ser Leu Arg Ile Pro Glu Gly Pro Ile Asp Gln Gly Pro Ala Ser
 65                  70                  75                  80

gga aga gtt cgt gct tta gaa gag caa ttt atg aag gcc aaa gaa cag   288
Gly Arg Val Arg Ala Leu Glu Glu Gln Phe Met Lys Ala Lys Glu Gln
                85                  90                  95

att gaa aat tat aag aaa caa act aaa aat ggt cca ggg aag gat cat   336
Ile Glu Asn Tyr Lys Lys Gln Thr Lys Asn Gly Pro Gly Lys Asp His
            100                 105                 110

gaa atc cta agg agg agg att gaa aat gga gct aaa gag ctc tgg ttt   384
Glu Ile Leu Arg Arg Arg Ile Glu Asn Gly Ala Lys Glu Leu Trp Phe
        115                 120                 125

ttt cta caa agt gag ttg aag aaa tta aag aat tta gaa gga aat gaa   432
Phe Leu Gln Ser Glu Leu Lys Lys Leu Lys Asn Leu Glu Gly Asn Glu
    130                 135                 140

ctc caa aga cat gca gat gaa ttt cta tca gat ttg gga cat cat gaa   480
Leu Gln Arg His Ala Asp Glu Phe Leu Ser Asp Leu Gly His His Glu
145                 150                 155                 160

agg tct ata atg acg gat cta tac tac ctc agt caa aca gat ggg gca   528
Arg Ser Ile Met Thr Asp Leu Tyr Tyr Leu Ser Gln Thr Asp Gly Ala
                165                 170                 175

ggt gat tgg cgt gaa aag gag gcc aaa gat ctg aca gag ctg gtc cag   576
Gly Asp Trp Arg Glu Lys Glu Ala Lys Asp Leu Thr Glu Leu Val Gln
            180                 185                 190

cgg aga ata aca tat ctt cag aat ccc aag gac tgc agc aaa gcc aag   624
Arg Arg Ile Thr Tyr Leu Gln Asn Pro Lys Asp Cys Ser Lys Ala Lys
        195                 200                 205

aag cta gtg tgt aat atc aac aaa ggc tgt ggc tat ggc tgt cag ctc   672
Lys Leu Val Cys Asn Ile Asn Lys Gly Cys Gly Tyr Gly Cys Gln Leu
    210                 215                 220

cat cat gta gtg tac tgc ttt atg att gca tat ggc acc cag cga aca   720
His His Val Val Tyr Cys Phe Met Ile Ala Tyr Gly Thr Gln Arg Thr
225                 230                 235                 240

ctc gcc ttg gaa tct cac aat tgg cgc tac gct act ggg gga tgg gaa   768
Leu Ala Leu Glu Ser His Asn Trp Arg Tyr Ala Thr Gly Gly Trp Glu
                245                 250                 255

act gtg ttt aga cct gta agt gag acg tgc aca gac aga tct ggc agc   816
Thr Val Phe Arg Pro Val Ser Glu Thr Cys Thr Asp Arg Ser Gly Ser
            260                 265                 270

tcc act gga cat tgg tca ggt gaa gta aag gac aaa aat gtt cag gtg   864
Ser Thr Gly His Trp Ser Gly Glu Val Lys Asp Lys Asn Val Gln Val
        275                 280                 285

gtt gag ctc ccc att gta gac agt gtt cat cct cgt cct cca tat tta   912
Val Glu Leu Pro Ile Val Asp Ser Val His Pro Arg Pro Pro Tyr Leu
    290                 295                 300

ccc ctg gct gtc cca gaa gac ctt gca gat cga ctt gta cga gtc cat   960
Pro Leu Ala Val Pro Glu Asp Leu Ala Asp Arg Leu Val Arg Val His
305                 310                 315                 320

ggt gat cct gca gtg tgg tgg gta tcc cag ttt gtc aag tac ttg att   1008
Gly Asp Pro Ala Val Trp Trp Val Ser Gln Phe Val Lys Tyr Leu Ile
                325                 330                 335
```

```
cgc cca caa ccc tgg ctg gaa aag gaa ata gaa gag gcc acc aag aag   1056
Arg Pro Gln Pro Trp Leu Glu Lys Glu Ile Glu Glu Ala Thr Lys Lys
            340                 345                 350

cta ggc ttc aaa cat cca gtt att gga gtc cat gtt aga cgc aca gac   1104
Leu Gly Phe Lys His Pro Val Ile Gly Val His Val Arg Arg Thr Asp
        355                 360                 365

aaa gtg gga gcg gaa gca gcc ttc cat ccc att gag gaa tac acg gtg   1152
Lys Val Gly Ala Glu Ala Ala Phe His Pro Ile Glu Glu Tyr Thr Val
    370                 375                 380

cac gtt gaa gaa gac ttt cag ctt ctt gct cgc aga atg caa gtg gat   1200
His Val Glu Glu Asp Phe Gln Leu Leu Ala Arg Arg Met Gln Val Asp
385                 390                 395                 400

aaa aaa agg gtg tat ttg gcc aca gat gac cct gct ttg tta aaa gag   1248
Lys Lys Arg Val Tyr Leu Ala Thr Asp Asp Pro Ala Leu Leu Lys Glu
                405                 410                 415

gca aaa aca aag tac ccc agt tat gaa ttt att agt gat aac tct atc   1296
Ala Lys Thr Lys Tyr Pro Ser Tyr Glu Phe Ile Ser Asp Asn Ser Ile
            420                 425                 430

tct tgg tca gct gga cta cat aat cga tat aca gaa aat tca ctt cgg   1344
Ser Trp Ser Ala Gly Leu His Asn Arg Tyr Thr Glu Asn Ser Leu Arg
        435                 440                 445

ggt gtg atc ctg gat ata cac ttt ctc tcc cag gca gac ttc cta gtg   1392
Gly Val Ile Leu Asp Ile His Phe Leu Ser Gln Ala Asp Phe Leu Val
    450                 455                 460

tgt act ttt tca tcg cag gtc tgt aga gtt gct tat gaa atc atg caa   1440
Cys Thr Phe Ser Ser Gln Val Cys Arg Val Ala Tyr Glu Ile Met Gln
465                 470                 475                 480

gcg ctg cat cct gat gcc tct gcg aac ttc cgt tct ttg gat gac atc   1488
Ala Leu His Pro Asp Ala Ser Ala Asn Phe Arg Ser Leu Asp Asp Ile
                485                 490                 495

tac tat ttt gga ggc cca aat gcc cac aac caa att gcc att tat cct   1536
Tyr Tyr Phe Gly Gly Pro Asn Ala His Asn Gln Ile Ala Ile Tyr Pro
            500                 505                 510

cac caa cct cga act gaa gga gaa atc ccc atg gaa cct gga gat att   1584
His Gln Pro Arg Thr Glu Gly Glu Ile Pro Met Glu Pro Gly Asp Ile
        515                 520                 525

att ggt gtg gct gga aat cac tgg gat ggc tat cct aaa ggt gtt aac   1632
Ile Gly Val Ala Gly Asn His Trp Asp Gly Tyr Pro Lys Gly Val Asn
    530                 535                 540

aga aaa ctg gga agg acg ggc cta tat ccc tcc tac aaa gtt cga gag   1680
Arg Lys Leu Gly Arg Thr Gly Leu Tyr Pro Ser Tyr Lys Val Arg Glu
545                 550                 555                 560

aag ata gaa aca gtc aag tac ccc aca tat ccc gag gct gac aag taa   1728
Lys Ile Glu Thr Val Lys Tyr Pro Thr Tyr Pro Glu Ala Asp Lys  *
                565                 570                 575
```

<210> 14
<211> 575
<212> PRT
<213> Sus scrofa

<400> 14

```
Met Arg Pro Trp Thr Gly Ser Trp Arg Trp Ile Met Leu Ile Leu Phe
 1               5                  10                  15
Ala Trp Gly Thr Leu Leu Phe Tyr Ile Gly Gly His Leu Val Arg Asp
            20                  25                  30
Asn Asp His Ser Asp His Ser Ser Arg Glu Leu Ser Lys Ile Leu Ala
        35                  40                  45
Lys Leu Glu Arg Leu Lys Gln Gln Asn Glu Asp Leu Arg Arg Met Ala
    50                  55                  60
Glu Ser Leu Arg Ile Pro Glu Gly Pro Ile Asp Gln Gly Pro Ala Ser
65                  70                  75                  80
Gly Arg Val Arg Ala Leu Glu Glu Gln Phe Met Lys Ala Lys Glu Gln
                85                  90                  95
Ile Glu Asn Tyr Lys Lys Gln Thr Lys Asn Gly Pro Gly Lys Asp His
            100                 105                 110
Glu Ile Leu Arg Arg Arg Ile Glu Asn Gly Ala Lys Glu Leu Trp Phe
        115                 120                 125
Phe Leu Gln Ser Glu Leu Lys Lys Leu Lys Asn Leu Glu Gly Asn Glu
    130                 135                 140
Leu Gln Arg His Ala Asp Glu Phe Leu Ser Asp Leu Gly His His Glu
145                 150                 155                 160
Arg Ser Ile Met Thr Asp Leu Tyr Tyr Leu Ser Gln Thr Asp Gly Ala
                165                 170                 175
Gly Asp Trp Arg Glu Lys Glu Ala Lys Asp Leu Thr Glu Leu Val Gln
            180                 185                 190
Arg Arg Ile Thr Tyr Leu Gln Asn Pro Lys Asp Cys Ser Lys Ala Lys
        195                 200                 205
Lys Leu Val Cys Asn Ile Asn Lys Gly Cys Gly Tyr Gly Cys Gln Leu
    210                 215                 220
His His Val Val Tyr Cys Phe Met Ile Ala Tyr Gly Thr Gln Arg Thr
225                 230                 235                 240
Leu Ala Leu Glu Ser His Asn Trp Arg Tyr Ala Thr Gly Gly Trp Glu
                245                 250                 255
Thr Val Phe Arg Pro Val Ser Glu Thr Cys Thr Asp Arg Ser Gly Ser
            260                 265                 270
Ser Thr Gly His Trp Ser Gly Glu Val Lys Asp Lys Asn Val Gln Val
        275                 280                 285
Val Glu Leu Pro Ile Val Asp Ser Val His Pro Arg Pro Pro Tyr Leu
    290                 295                 300
Pro Leu Ala Val Pro Glu Asp Leu Ala Asp Arg Leu Val Arg Val His
305                 310                 315                 320
Gly Asp Pro Ala Val Trp Trp Val Ser Gln Phe Val Lys Tyr Leu Ile
                325                 330                 335
Arg Pro Gln Pro Trp Leu Glu Lys Glu Ile Glu Glu Ala Thr Lys Lys
            340                 345                 350
Leu Gly Phe Lys His Pro Val Ile Gly Val His Val Arg Arg Thr Asp
        355                 360                 365
Lys Val Gly Ala Glu Ala Ala Phe His Pro Ile Glu Glu Tyr Thr Val
    370                 375                 380
His Val Glu Glu Asp Phe Gln Leu Leu Ala Arg Arg Met Gln Val Asp
385                 390                 395                 400
Lys Lys Arg Val Tyr Leu Ala Thr Asp Asp Pro Ala Leu Leu Lys Glu
                405                 410                 415
Ala Lys Thr Lys Tyr Pro Ser Tyr Glu Phe Ile Ser Asp Asn Ser Ile
            420                 425                 430
Ser Trp Ser Ala Gly Leu His Asn Arg Tyr Thr Glu Asn Ser Leu Arg
        435                 440                 445
Gly Val Ile Leu Asp Ile His Phe Leu Ser Gln Ala Asp Phe Leu Val
    450                 455                 460
Cys Thr Phe Ser Ser Gln Val Cys Arg Val Ala Tyr Glu Ile Met Gln
465                 470                 475                 480
Ala Leu His Pro Asp Ala Ser Ala Asn Phe Arg Ser Leu Asp Asp Ile
                485                 490                 495
Tyr Tyr Phe Gly Gly Pro Asn Ala His Asn Gln Ile Ala Ile Tyr Pro
            500                 505                 510
His Gln Pro Arg Thr Glu Gly Glu Ile Pro Met Glu Pro Gly Asp Ile
        515                 520                 525
Ile Gly Val Ala Gly Asn His Trp Asp Gly Tyr Pro Lys Gly Val Asn
    530                 535                 540
```

Arg Lys Leu Gly Arg Thr Gly Leu Tyr Pro Ser Tyr Lys Val Arg Glu
545 550 555 560
Lys Ile Glu Thr Val Lys Tyr Pro Thr Tyr Pro Glu Ala Asp Lys
565 570 575

<210> 15
<211> 37
<212> DNA
<213> Artificial Sequence

<220>
<223> PCR primer SH415

<400> 15
ggcggccgcc accatggcac acgcaccggc acgctgc 37

<210> 16
<211> 33
<212> DNA
<213> Artificial Sequence

<220>
<223> PCR primer SH413

<400> 16
ttaattaatc aggcattggg gtttgtcctc atg 33

<210> 17
<211> 37
<212> DNA
<213> Artificial Sequence

<220>
<223> PCR primer SH414

<400> 17
ggcggccgcc accatgggtg aaccccaggg atccatg 37

<210> 18
<211> 33
<212> DNA
<213> Artificial Sequence

<220>
<223> PCR primer SH411

<400> 18
ttaattaatc acttccgggc ctgctcgtag ttg 33

<210> 19
<211> 36
<212> DNA
<213> Artificial Sequence

<220>
<223> PCR primer RCD679

<400> 19
gcggccgcca ccatgaatag ggcccctctg aagcgg 36

<210> 20
<211> 32
<212> DNA
<213> Artificial Sequence

<220>
<223> PCR primer RCD680

<400> 20
ttaattaatc acacccccat ggcgctcttc tc 32

<210> 21
<211> 36
<212> DNA
<213> Artificial Sequence

<220>
<223> PCR primer SH420

<400> 21
gcggcgcgcc gataatgacc accctgatca ctccag 36

<210> 22
<211> 37
<212> DNA
<213> Artificial Sequence

<220>
<223> PCR primer SH421

<400> 22
ccttaattaa ctatttttca gcttcaggat atgtggg 37

<210> 23
<211> 32
<212> DNA
<213> Artificial Sequence

<220>
<223> PCR primer rEPO-forward

<400> 23
gggaattcgc tcccccacgc ctcatttgcg ac 32

<210> 24
<211> 32
<212> DNA
<213> Artificial Sequence

<220>
<223> PCR primer rEPO-reverse

<400> 24
cctctagatc acctgtcccc tctcctgcag gc 32

<210> 25
<211> 46
<212> DNA
<213> Artificial Sequence

<220>
<223> PCR primer ScMntI-forward

<400> 25
gggcggccgc caccatggcc ctctttctca gtaagagact gttgag 46

<210> 26
<211> 43
<212> DNA
<213> Artificial Sequence

<220>
<223> PCR primer ScMnt1-reverse

<400> 26

ccggcgcgcc cgatgacttg ttgttcaggg gatatagatc ctg 43