

SEPARATION OF VWF AND VWF PROPEPTIDE BY CHROMATOGRAPHIC METHODS

CROSS REFERENCE TO RELATED APPLICATIONS

[0001] This application claims priority to U.S. Provisional No. 62/646,109, filed on March 21, 2018, which is hereby incorporated by reference in its entirety.

FIELD OF THE INVENTION

[0002] The present invention relates to methods for separating mature von Willebrand Factor (VWF) from von Willebrand Factor pro-peptide (VWF-PP).

BACKGROUND OF THE INVENTION

[0003] In the course of protein maturation within a cell, the protein to be matured undergoes posttranslational modifications. These modifications include among others acetylation, methylation, glycosylation and proteolytic cleavage. These modifications are in many cases necessary for the protein function and activity and they may also influence the efficiency of proteins, in particular of enzymes.

[0004] Pro-proteins or protein precursors are inactive proteins that are turned into an active form by one or more of these post-translational modifications, in particular, by the cleavage of a pro-peptide from the pro-protein.

[0005] The active form of these proteins may be useful therapeutic and/or diagnostic proteins. However, the active proteins are usually available at very low amounts in living organisms. As such, the active proteins are produced recombinantly from their pro-proteins which are preferably activated *in vitro* by contacting them with recombinant activation enzymes (e.g., proteases).

[0006] von Willebrand Factor (VWF) is a glycoprotein circulating in plasma as a series of multimers ranging in size from about 500 to 20,000 kD. The full length of cDNA of VWF has been cloned; the propolypeptide corresponds to amino acid residues 23 to 764 of the full length prepro-VWF (Eikenboom et al (1995) Haemophilia, 1, 77-90). Multimeric forms of VWF are composed of 250 kD polypeptide subunits linked together by disulfide bonds.

VWF mediates the initial platelet adhesion to the sub-endothelium of the damaged vessel wall, with the larger multimers exhibiting enhanced hemostatic activity. Multimerized VWF binds to the platelet surface glycoprotein GpIb α , through an interaction in the A1 domain of VWF, facilitating platelet adhesion. Other sites on VWF mediate binding to the blood vessel wall. Thus, VWF forms a bridge between the platelet and the vessel wall that is essential to platelet adhesion and primary hemostasis under conditions of high shear stress. Normally, endothelial cells secrete large polymeric forms of VWF and those forms of VWF that have a lower molecular weight arise from proteolytic cleavage. The multimers of exceptionally large molecular masses are stored in the Weibel-Pallade bodies of the endothelial cells and liberated upon stimulation by agonists such as thrombin and histamine.

[0007] Industrially, VWF, in particular recombinant VWF (rVWF), is synthesized and expressed together with rFVIII in a genetically engineered cell lines, such as an engineered CHO cell line. The function of the co-expressed rVWF is to stabilize rFVIII in the cell culture process. rVWF is synthesized in the cell as pre-propeptide VWF (prepro-VWF), containing a large pro-peptide (VWF-PP) attached to the N-terminus of the mature VWF (matVWF) subunit. Upon maturation in the endoplasmatic reticulum and Golgi apparatus, the VWF-PP is cleaved off by the action of the cellular protease furin and is secreted as a homopolymer of identical subunits, consisting of dimers of the expressed protein. In some cases, furin cleavage produces a heterodimeric complex comprising a mature VWF non-covalently associated with a VWF pro-peptide.

[0008] VWF-PP can be separated from mature VWF by in vitro treatment with furin or furin-like proteases (Schlokot U. *et al.* (1996) *Biotechnol. Appl. Biochem.* 24:257-267; Preininger A. *et al.* (1999) *Cytotechnology* 30:1-15). Furin belongs to the family of the pro-protein convertases and is dependent on Ca²⁺. This enzyme specifically cleaves the C-terminal peptide bond of arginine within a specific sequence, containing arginine at positions -1 and -4. This sequence can be found in numerous human proteins, showing that furin plays a major role in the maturation of a number of human pro-peptide-proteins. Furin used in the method of the present invention is preferably of recombinant origin. Recombinantly produced proteases are advantageously employed because they can be produced in high quantities. In some embodiments, furin is obtained from crude cell culture supernatant of a cell line secreting said protease or cell extract.

[0009] Current conventional methods produce mature VWF by either incubating the pre-propeptide VWF with proteases in a liquid phase whereby the maturation itself (*e.g.*, the cleavage of the pro-peptide from the pro-protein) occurs in an unbound state in free solution, or as described for example in WO2000/049047, by immobilizing the protease on a solid carrier, which is contacted and incubated with a preparation comprising VWF-PP (see, *e.g.*, WO2000/049047). VWF is synthesized by endothelial cells and megakaryocytes as pre-propeptide VWF ("prepro-VWF") that consists to a large extent of repeated domains. Upon cleavage of the signal peptide, prepro-VWF dimerizes through disulfide linkages at the carboxy-terminus region in the endoplasmic reticulum. Additional disulfide linkages are formed near the amino-terminus of the subunits to form multimers in the Golgi. The assembly to multimers is followed by the proteolytic cleavage of the VWF pro-peptide by the pro-peptide processing protease furin. After cleavage, the VWF pro-peptide remains non-covalently associated with the VWF multimer to form a mature VWF / VWF-PP complex. Upon stimulation, the complex is secreted into the blood and the VWF pro-peptide dissociates from the VWF multimers. Therapeutically effective mature VWF multimers can be produced by recombinantly expressing pro-VWF in mammalian cell lines and processing the pro-VWF protein to mature VWF through a series of *in vitro* cleavage and purification steps. However, there remains a need in the art for producing high purity, therapeutically effective mature VWF multimer preparations (mat-rVWF) and the present invention meets this need by providing methods for obtaining high purity, mat-rVWF preparations, where the method comprises, for example, after furin maturation, the addition of a chelating agent and/or increasing the pH to a pH of at least 7 during the purification process to facilitate separation of the VWF-propeptide from mat-rVWF.

BRIEF SUMMARY OF THE INVENTION

[0010] The present invention provides a method for obtaining a composition comprising a high purity, propeptide depleted mature recombinant rVWF (mat-rVWF), said method comprising the steps of:

- a) providing a solution comprising mat-rVWF/ rVWF-PP complex, mat-rVWF, and rVWF propeptide (rVWF-PP);
- b) inducing dissociation of said mat-rVWF/ rVWF-PP complex in said solution in a) into mat-rVWF and rVWF-PP, wherein said dissociation occurs by

disruption of the non-covalently associated mat-rVWF and rVWF-PP, wherein said dissociation is induced by:

- i. addition of at least one chelating agent, or
 - ii. increasing the pH to a pH of at least 7; and
- c) collecting said mat-rVWF to obtain a high purity, mat-rVWF composition, wherein said high purity, mat-rVWF composition comprises at least 95% mature rVWF and less than 5% rVWF-PP.

[0011] In some embodiments, the high purity, mat-rVWF composition comprises at least 96% mat-rVWF and less than 4% rVWF-PP, at least 97% mat-rVWF and less than 3% rVWF-PP, at least 98% mat-rVWF and less than 2% rVWF-PP, at least 99% mat-rVWF and less than 1% rVWF-PP, or at least 99.5% mat-rVWF and less than 0.5% rVWF-PP, or 99.9% mat-rVWF and less than 0.1% rVWF-PP.

[0012] In some embodiments, the solution is selected from the group consisting of a cell culture medium, an antibody column flow-through solution, and a buffered solution.

[0013] In some embodiments, the solution has been treated with furin prior to step a).

[0014] In some embodiments, the solution is an antibody column flow-through solution.

[0015] In some embodiments, the at least one chelating agent is a divalent cation chelating agent. In some embodiments, the divalent cation chelating agent is selected from the group consisting of EDTA, EGTA, CDTA, and citrate.

[0016] In some embodiments, the pH is increased to at least 7.1, 7.2, 7.3, 7.4, 7.5, 7.6, 7.7, 7.8, 7.9, 8.0, 8.1, 8.2, 8.3, 8.4, 8.5, 8.6, 8.7, 8.8, 8.9, or 9.0. In some embodiments, the pH is increased to at least about 7.2 to about 7.8. In some embodiments, the pH is increased to at least about 7.6. In some embodiments, the pH is increased by the addition of basic amino acids, Tris, NaOH, Tricine, or ethanolamine.

[0017] In some embodiments, the collecting in step b) of the method described herein comprises one or more protein separation methods. In some embodiments, the one or more protein separation methods is selected from the group consisting of ion exchange chromatography (IEC), size exclusion chromatography (SEC), physical size separation by membrane technology, and affinity chromatography. In some embodiments, the protein separation method is size exclusion chromatography (SEC). In some embodiments, the one or more protein separation method is ion exchange chromatography (IEC). In some

embodiments, the ion exchange chromatography (IEC) is cation exchange chromatography. In some embodiments, the ion exchange chromatography (IEC) is a combination of anion exchange chromatography and cation exchange chromatography.

[0018] In some embodiments, the one or more protein separation methods comprise a buffer system, wherein said buffer system comprises one or more buffers. In some embodiments, the said one or more buffers includes wash buffers, wherein said one or more wash buffers include one, two, three, four, and/or five wash buffers, wherein when said one or more buffers includes five wash buffers, the first, second, third, and/or fifth wash buffers have a higher pH than the fourth wash buffer, and when said one or more buffers includes four wash buffers, the first, second, and/or fourth wash buffers have a higher pH than the third wash buffer. In some embodiments, the method further comprises a viral inactivation treatment step after the first wash buffer, and optionally the pH of the viral inactivation treatment step has a higher pH than said third and/or fourth wash buffer. In some embodiments, the one or more buffers comprise said one or more chelating agents. In some embodiments, the one or more buffers exhibit a pH of at least 7.

[0019] In some embodiments, the one or more protein separation methods comprise a buffer system, wherein said buffer system comprises one or more loading buffers. In some embodiments, the one or more loading buffers comprise said one or more chelating agents. In some embodiments, the one or more loading buffers exhibit a pH of at least 7.

[0020] In some embodiments, the one or more protein separation methods comprise a buffer system, wherein said buffer system comprises one or more load, wash, and/or elution buffers. In some embodiments, the one or more load, wash, and/or elution buffers comprise said one or more chelating agents. In some embodiments, the one or more load, wash, and/or elution buffers exhibit a pH of at least 7. In some embodiments, the one or more load, wash, and/or elution buffers comprise said one or more chelating agents and exhibit a pH of at least 7.

[0021] In some embodiments, the buffering system is selected from the group consisting of glycine HEPES (4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid), TrisHCl (Tris(hydroxymethyl)-aminomethane), histidine, imidazole, acetate citrate, MES, and 2-(N-morpholino)ethanesulfonic acid.

[0022] In some embodiments, the buffer further comprises one or more monovalent cations. In some embodiments, the one or more monovalent cations are selected from the group consisting of Na⁺, K⁺, Li⁺, and Cs⁺. In some embodiments, the monovalent cation is Na⁺.

[0023] In some embodiments, the buffer further comprises one or more monovalent, divalent and/or trivalent anions. In some embodiments, the one or more monovalent, divalent and/or trivalent anions are selected from the group consisting of Cl⁻, acetate⁻, SO₄²⁻, Br⁻, and citrate³⁻.

[0024] In some embodiments, the buffer system comprises at least one buffer exhibiting a conductivity of ≥ 0.5 mS/cm at 25°C. In some embodiments, the buffer system comprises at least one buffer exhibiting a conductivity of 15.0 ± 0.2 mS/cm at 25°C.

[0025] In some embodiments, the buffer further comprises one or more nonionic detergents. In some embodiments, the nonionic detergent is selected from the group consisting of Triton X100, Tween 80, and Tween 20.

[0026] In some embodiments, the buffer further comprises one or more additional substances selected from the group consisting of non-reducing sugars, sugar alcohols, and polyols.

[0027] In some embodiments, the high purity mat-rVWF composition comprises a host cell (HC) impurity level of ≤ 2.0 %. In some embodiments, the high purity, mat-rVWF composition comprises a host cell (HC) impurity level of ≤ 0.6 %.

[0028] In some embodiments, the solution comprising mat-rVWF/ rVWF-PP complex, mat-rVWF, and rVWF-PP is derived from a capture step for rVWF.

[0029] In some embodiments, the solution comprising mat-rVWF/ rVWF-PP complex, mat-rVWF, and rVWF-PP is derived from a method comprising a FVIII immunoaffinity step and anion exchange chromatography step.

[0030] The present invention also provides a method for obtaining a composition comprising a high purity, propeptide depleted mature recombinant rVWF (high purity mat-rVWF), said method comprising the steps of:

- a) loading a solution comprising pro-rVWF, mat-rVWF/ rVWF-PP complex, mat-rVWF, and/or rVWF propeptide (rVWF-PP) onto an anion exchange column, wherein said pro-rVWF, mat-rVWF/ rVWF-PP complex, and mat-rVWF are bound to said anion exchange column;

- b) washing said anion exchange column in a) containing said bound pro-rVWF, mat-rVWF/ rVWF-PP complex, and mat-rVWF with one or more wash buffers;
- c) treating said column in b) comprising the bound pro-rVWF, mat-rVWF/ rVWF-PP complex, and mat-rVWF with furin, wherein said furin cleaves said pro-rVWF into mat-rVWF and rVWF-PP;
- d) eluting said bound pro-rVWF, mat-rVWF/ rVWF-PP complex, and mat-rVWF from the column in c) with an elution buffer, wherein said elution buffer induces dissociation of said rVWF-PP from mat-rVWF non-covalently associated with said rVWF-PP, and wherein said dissociation is induced by:
 - i. addition of at least one chelating agent into said elution buffer, or
 - ii. increasing the pH of said elution buffer to a pH of at least 7; and
- e) collecting said mat-rVWF separately from said rVWF-PP to obtain a high purity mat-rVWF composition, wherein said high purity mat-rVWF composition comprises at least 95% mature rVWF and less than 5% rVWF-PP.

[0031] In some embodiments, a) and b) occur simultaneously in a single step.

[0032] In some embodiments, the said one or more buffers includes wash buffers, wherein said one or more wash buffers include one, two, three, four, and/or five wash buffers, wherein when said one or more buffers includes five wash buffers, the first, second, third, and/or fifth wash buffers have a higher pH than the fourth wash buffer, and when said one or more buffers includes four wash buffers, the first, second, and/or fourth wash buffers have a higher pH than the third wash buffer. In some embodiments, the method further comprises a viral inactivation treatment step after the first wash buffer, and optionally the pH of the viral inactivation treatment step has a higher pH than said third and/or fourth wash buffer.

[0033] In some embodiments, the solution in a) comprises the flow through from a monoclonal antibody column, wherein said monoclonal antibody is a FVIII monoclonal antibody.

[0034] In some embodiments, the solution in a) is selected from the group consisting of a cell culture medium, an antibody column flow-through solution, and a buffered solution.

[0035] In some embodiments, the at least one chelating agent is a divalent cation chelating agent. In some embodiments, the divalent cation chelating agent is selected from the group consisting of EDTA, EGTA, CDTA, and citrate.

[0036] In some embodiments, the pH is increased to at least 7.1, 7.2, 7.3, 7.4, 7.5, 7.6, 7.7, 7.8, 7.9, or 8.0. In some embodiments, the pH is increased to at least about 7.2 to about 7.8. In some embodiments, the pH is increased to at least about 7.6. In some embodiments, the pH is increased by the addition of basic amino acids. In some embodiments, the one or more wash buffers in b) comprise said one or more chelating agents. In some embodiments, the one or more wash buffers in b) exhibit a pH of at least 7. In some embodiments, the one or more wash buffers in b) comprise said one or more chelating agents and exhibit a pH of at least 7.

[0037] In some embodiments, the method further comprises a step of viral inactivation, wherein said viral inactivation occurs before, after, or concurrently with the washing step and/or the elution step, but before the collecting step. In some embodiments, the viral inactivation treatment inactivates lipid enveloped viruses. In some embodiments, the viral inactivation treatment is a solvent and detergent (S/D) treatment.

[0038] In some embodiments, the one or more buffers comprise a buffer selected from the group consisting of glycine HEPES (4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid), TrisHCl (Tris(hydroxymethyl)-aminomethane), histidine, imidazole, acetate citrate, MES, and 2-(N-morpholino)ethanesulfonic acid.

[0039] In some embodiments, the one or more buffers further comprise one or more monovalent cations. In some embodiments, the one or more monovalent cations are selected from the group consisting of Na⁺, K⁺, Li⁺, and Cs⁺. In some embodiments, the monovalent cation is Na⁺.

[0040] In some embodiments, the one or more buffers further comprise one or more monovalent, divalent, and/or trivalent anions. In some embodiments, the one or more monovalent, divalent and/or trivalent anions are selected from the group consisting of Cl⁻, acetate⁻, SO₄²⁻, Br⁻, and citrate³⁻.

[0041] In some embodiments, the one or more buffers comprise at least one buffer exhibiting a conductivity of ≥ 0.5 mS/cm at 25°C. In some embodiments, the one or more buffers comprise at least one buffer exhibiting a conductivity of 15.0 ± 0.2 mS/cm at 25°C.

[0042] In some embodiments, the one or more buffers further comprise one or more nonionic detergents. In some embodiments, the nonionic detergent is selected from the group consisting of Triton X100, Tween 80, and Tween 20.

[0043] In some embodiments, the said one or more buffers further comprise one or more additional substances selected from the group consisting of non-reducing sugars, sugar alcohols, and polyols.

[0044] In some embodiments, the high purity mat-rVWF composition comprises a host cell (HC) impurity level of ≤ 2.0 %. In some embodiments, the high purity mat-rVWF composition comprises a host cell (HC) impurity level of ≤ 0.6 %.

[0045] In some embodiments, the high purity mat-rVWF composition is used for the production of a pharmaceutical composition.

[0046] The present invention further provides a pharmaceutical composition comprising high purity mat-rVWF generated by the method according to any of the preceding claims and a pharmaceutically acceptable buffer. In some embodiments, the pharmaceutical composition comprises 50 mM Glycine, 10 mM Taurine, 5% (w/w) Sucrose, 5% (w/w) D-Mannitol, 0.1% Polysorbate 80, 2 mM CaCl_2 , 150 mM NaCl, wherein said composition has a pH of about pH 7.4.

[0047] Other objects, advantages and embodiments of the invention will be apparent from the detailed description following.

BRIEF DESCRIPTION OF THE DRAWINGS

[0048] FIG. 1 shows purification of matured rVWF on a cation exchanger as represented in Example 1.

[0049] FIG. 2 provides a table of the purification results.

[0050] FIG. 3 shows a silver stained protein gel and a western blot illustrating the separation of mat-VWF and r-VWF propeptide (rVWF-PP) by the method of Example 1.

[0051] FIG. 4 shows a flow chart of the experimental set-up for Examples 2 and 3.

[0052] FIG. 5 shows a chromatogram for Example 2 and a chromatography scheme used for Examples 2 and 3

[0053] FIG. 6 provides a table of the reagents used and a table of the results for Example 2.

[0054] FIG. 7 shows another chromatogram for Example 2 and a table of the results for Example 3.

[0055] FIG. 8 shows a silver stained protein gel illustrating the separation of mat-rVWF and rVWF propeptide (rVWF-PP) by the method of Example 2 and Example 3.

[0056] FIG. 9 shows a western blot illustrating the separation of rVWF and rVWF propeptide by the method of Example 2 and Example 3.

[0057] FIG. 10 shows a chromatogram, a chromatography scheme, and buffer compositions for Example 4.

[0058] FIG. 11 provides a table of the results for Example 4.

[0059] FIG. 12 shows a silver stained protein gel and a western blot illustrating the separation of mat-rVWF and rVWF propeptide (rVWF-PP) by the method of Example 4.

[0060] FIG. 13 shows a chromatogram, a chromatography scheme, and buffer compositions for Example 5.

[0061] FIG. 14 provides a table of the results for Example 5.

[0062] FIG. 15 shows a chromatogram, a chromatography scheme, and buffer compositions for Example 6.

[0063] FIG. 16 provides a table of the results for Example 6.

[0064] FIG. 17 shows a chromatogram, a chromatography scheme, and buffer compositions for Example 7.

[0065] FIG. 18 provides a table of the results for Example 7.

[0066] FIG. 19 shows a chromatogram, a chromatography scheme, and buffer compositions for Example 8.

[0067] FIG. 20 provides a table of the results for Example 8.

[0068] FIG. 21 shows a silver stained protein gel illustrating the separation of mat-rVWF and rVWF propeptide (rVWF-PP) by the method of Example 8.

[0069] FIG. 22 shows a western blot illustrating the separation of rVWF and rVWF propeptide by the method of Example 8. The 1% agarose gel shows the multimeric pattern of the products.

[0070] FIG. 23 shows a western blot illustrating the separation of mat-rVWF and rVWF propeptide (rVWF-PP) by the method of Example 8.

[0071] FIG. 24 shows a chromatogram, a chromatography scheme, and buffer compositions for Example 9.

[0072] FIG. 25 provides a table of the results for Example 9.

[0073] FIG. 26 provides a table of the products for Example 9.

[0074] FIG. 27 shows a silver stained protein gel illustrating the separation of rVWF and rVWF propeptide by the method of Example 9.

[0075] FIG. 28 shows a western blot illustrating the separation of rVWF and rVWF propeptide by the method of Example 9. The 1% agarose gel shows the multimeric pattern of the products.

[0076] FIG. 29 shows a western blot illustrating the separation of rVWF and rVWF propeptide by the method of Example 9.

[0077] FIG. 30 shows the purity of the product containing fractions obtained for enhanced cation exchange chromatography (CEX) as used for Examples 1, 2, 3, 6, 8, and 9.

[0078] FIG. 31 shows the depletion factor of product related impurities for Examples 1, 2, 3, 6, 8, and 9.

[0079] FIG. 32 shows the purity of the product containing fractions obtained for enhanced size exclusion chromatography (SEC) as used for Examples 4 and 5.

[0080] FIG. 33 shows the depletion factor of product related impurities for Examples 4 and 5.

[0081] FIG. 34 shows the buffer formulations and materials used in the TMAE separation method.

[0082] FIG. 35 shows the loading conditions for the furin-processed mature VWF/VWF-propeptide complex.

[0083] FIG. 36 shows the details of the buffers, conditions, parameters, and flow rates of the chromatography method.

[0084] FIG. 37 shows a chromatogram of the dissociation of furin-processed mature VWF/VWF-propeptide complex into mature VWF and VWF -propeptide (VWF-PP). It shows depletion of VWF-PP from the fraction containing mature VWF.

[0085] FIG. 38 shows another chromatogram of the separation of mature VWF and VWF-propeptide (VWF-PP). It shows depletion of VWF-PP from the fraction containing mature VWF.

[0086] FIGS. 39A and FIG. 39B provide schematic diagrams of exemplary methods for the purification of mature VWF including separation of mature VWF and VWF-PP.

[0087] FIG. 40 provides a table highlighting some of the advantages of the cation exchange chromatography method described herein.

[0088] FIG. 41 shows a schematic of two chromatograms showing the separation of rVWF propeptide using the size exclusion chromatography described herein using either a SQA running buffer or a SQC running buffer that contains citrate. The change in SEC parameters (SEC buffers) did not result in a change in the purification of mature VWF besides increased removal/separation of residual VWF-PP.

[0089] FIG. 42 provides a table highlighting some of the advantages of the optimized SEC buffer (SQC buffer). The SQC buffer includes at least one chelating agent and was shown to reduce the amount of VWF-PP in the purified mature VWF fraction.

[0090] FIG. 43A and FIG. 43B provide flowcharts of downstream processing protocols for rVWF. FIG. 43A shows the currently used process. FIG. 43B shows the process described herein which includes an improved CAT (UNO_S) step.

[0091] FIG. 44 provides a table of the chromatography hardware of step CAT in the first generation (Gen 1) process and the second generation (Gen 2) process.

[0092] FIG. 45 depicts a table of wash and elution conditions of the Gen 2 process.

[0093] FIG. 46 shows a comparison table of the 1st and 2nd generation rVWF small scale polishing steps on UNO_Sphere S (step CAT).

[0094] FIG. 47 depicts a table of the cleaning and sanitization procedure for the UNO_Sphere S column.

[0095] FIG. 48 depicts a table of the composition of buffers for the CAT polishing step.

[0096] FIG. 49A and FIG. 49B show chromatograms of run VW_USS_05. FIG. 49A shows the entire chromatogram, including the CIP procedure. FIG. 49B depicts the 36% buffer B wash and the gradient elution phase. The UV absorption is shown in blue (280nm) and magenta (254nm).

- [0097] FIG. 50 depicts SDS-PAGE silver stain gel and Western blot of run VW_UUS_05.
- [0098] FIG. 51 depicts a multimer agarose gel of run VW_UUS_05.
- [0099] FIG. 52 shows rVWF:Ag data of the different runs of the study.
- [0100] FIG. 53 shows rVWF Risto Co activity data of the different runs of the study.
- [0101] FIG. 54 shows pro-peptide concentration (pro-peptide ($\mu\text{g}/\text{mg}$ rVWF:Ag)) data of the different runs of the study.
- [0102] FIG. 55 shows pro-peptide concentration (pro-peptide (μg PP /1000 U Risto)) data of the different runs of the study.
- [0103] FIG. 56 shows analytical key results in the eluate pools of the different runs of the study.
- [0104] FIG. 57 shows the targeted CAT-E criteria for a successful method development.
- [0105] FIG. 58 provides exemplary embodiments of the anion exchange, cation exchange, and size exclusion chromatography methods for us in separation of mat-rVWF and rVWF-PP.
- [0106] FIG. 59 shows the various VWF forms: pro-VWF (also referred to as pro-rVWF), matVWF/VWF-PP complex (also referred to as mat-rVWF/VWF-PP complex), matVWF (also referred to as mat-rVWF), and VWF-PP (also referred to as rVWF-PP).
- [0107] FIG. 60 shows VWF nucleic acid and amino acid sequences.
- [0108] FIG. 61 shows the DF3338/042 western blot and raw data for analysis.
- [0109] FIG. 62 shows the DF3362/023 western blot and raw data for analysis.
- [0110] FIG. 63 shows the comparison of the data from FIG. 61 and FIG. 62.
- [0111] FIG. 64 shows the amino acid sequence for an exemplary VWF – FVIII fusion protein wherein an active FVIII is embedded in an VWF motif (VWF 764 to 1336 – FVIII heavy chain 24 to 760 – VWF 2218 to 2593 – FVIII light chain 1333 to 2351- VWF 2620 to 2813).

[00112] FIG. 65 shows the amino acid sequence for an exemplary VWF – FVIII fusion protein wherein the n-glycosylation rich domain replaces the FVIII-B-domain (FVIII heavy chain 19 to 760 – vWF 2218 to 2593 – FVIII light chain 1333 to 2351).

[00113] FIG. 66 depicts a table of buffers and compositions used in the variant vWF purification process described in Example 14.

[00114] FIG. 67 shows a chromatogram and chromatogram scheme of the run VW_USS_07.

[00115] FIG. 68 shows analytical key results of the run.

[00116] FIG. 69 shows SDS-PAGE silver stain gel of the representative run. Depletion of rvWF-propeptide was observed during the wash steps Wash 1, WSD, and Wash 2.

[00117] FIG. 70 depicts a table of buffers and compositions used in the variant vWF purification process described in Example 15. This example provides an alternate, variant embodiment for separation of the r-vWF propeptide from the r-VWF polypeptide after furin cleavage in order to test for additional sialylation.

[00118] FIG. 71 shows a chromatogram and chromatogram scheme of the run VW_USS_06.

[00119] FIG. 72 shows analytical key results of the run.

[00120] FIG. 73 shows SDS-PAGE silver stain gel of the representative run.

[00121] FIG. 74 depicts a table of buffers and compositions used in the variant vWF purification process described in Example 16.

[00122] FIG. 75 shows a chromatogram and chromatogram scheme of the run VW_USS_08.

[00123] FIG. 76 shows analytical key results of the run including yield sialylation.

[00124] FIG. 77 shows a SDS-PAGE silver stain gel DFM07247 of the representative run.

[00125] FIG. 78 depicts sialylation profiles of the eluates from the VW_USS_06 and VW_USS_08 runs.

DETAILED DESCRIPTION OF THE INVENTION

I. Introduction

[00126] The method described herein separates mature VWF and VWF propeptide that have been dissociated from the non-covalently linked heterodimeric complex comprising the mature VWF and VWF propeptide. This separation is facilitated (induced) by the addition of at least one chelating agent and/or by increasing the pH to at least 7.0 of the solution comprising the mature VWF and VWF propeptide to a protein separation method. All enhanced anion exchange (AEX), cation exchange (CEX) and/or size exclusion chromatography (SEC) methods as described herein can be combined in any variation to obtain r-vWF with improved properties.

II. Select Definitions

[00127] The term “recombinant” when used with reference, *e.g.*, to a cell, or nucleic acid, protein, or vector, indicates that the cell, nucleic acid, protein or vector, has been modified by the introduction of a heterologous nucleic acid or protein or the alteration of a native nucleic acid or protein, or that the cell is derived from a cell so modified. Thus, for example, recombinant cells express genes that are not found within the native (non-recombinant) form of the cell or express native genes that are otherwise abnormally expressed, under expressed or not expressed at all.

[00128] As used herein, “recombinant VWF” or “rVWF” includes VWF obtained via recombinant DNA technology. In certain embodiments, rVWF proteins of the invention can comprise a construct, for example, as described in US 8,597,910, which is incorporated herein by reference with respect to the methods of producing recombinant VWF. The VWF in the present invention can include all potential forms, including the monomeric and multimeric forms. It should also be understood that the present invention encompasses different forms of VWF to be used in combination. For example, the VWF of the present invention may include different multimers, different derivatives and both biologically active derivatives and derivatives not biologically active.

[00129] In the context of the present invention, the recombinant VWF embraces any member of the VWF family from, for example, a mammal such as a primate, human, monkey, rabbit, pig, rodent, mouse, rat, hamster, gerbil, canine, feline, and biologically active

derivatives thereof. Mutant and variant VWF proteins having activity are also embraced, as are functional fragments and fusion proteins of the VWF proteins. Furthermore, the VWF of the invention may further comprise tags that facilitate purification, detection, or both. The VWF described herein may further be modified with a therapeutic moiety or a moiety suitable for imaging in vitro or in vivo.

[00130] The term “VWF multimer” refers to VWF comprising at least 10 subunits, or 12, 14, or 16 subunits, to about 20, 22, 24 or 26 subunits or more. The term “subunit” refers to a monomer of VWF. As is known in the art, it is generally dimers of VWF that polymerize to form the larger order multimers. (see, e.g., Turecek *et al.*, *Semin. Thromb. Hemost.*, 2010, 36(5): 510-521 which is hereby incorporated by reference in its entirety for all purposes and in particular for all teachings regarding multimer analysis of VWF).

[00131] The term “pre-propeptide VWF,” “prepro-VWF” or “pro-VWF” refers to a non-mature VWF polypeptide comprising a signal peptide of about 22 amino acid residues, a VWF propeptide of about 741 amino acid residues, and a mature VWF subunit of about 2050 amino acid residues. Pro-VWF subunits can dimerize through disulfide bonds near their carboxyl termini in the endoplasmic reticulum to form tail-to tail dimers which are then transported to the Golgi. In the Golgi, additional head-to-head disulfide bonds are formed near the amino-termini of the subunits, thereby forming multimers. Proteolytic cleavage of the VWF propeptide occurs via the processing protease furin, thus producing a mature VWF/VWF-PP complex. When “r” is included prior to the VWF designation, this refers to the recombinant version. In some embodiments, the methods described herein apply to recombinant VWF (rVWF).

[00132] The term “VWF complex” or “mat-VWF/VWF-PP complex” refers to a non-covalently linked heterodimeric structure comprising a mature VWF subunit and VWF propeptide. The VWF complex can be generated as a product of furin cleavage between the propeptide portion and mature VWF portion of the pre-propeptide VWF. When “r” is included prior to the VWF designation, this refers to the recombinant version. In some embodiments, the methods described herein apply to recombinant VWF (rVWF).

[00133] The term “mature VWF” or “mat-VWF,” refers to a mature VWF subunit of about 2050 amino acid residues. A mature VWF subunit can be part of a pre-propeptide VWF or a VWF complex. Mature VWF can be referred to as “free VWF” upon separation (isolation) from a VWF propeptide. When “r” is included prior to the VWF designation, this

refers to the recombinant version. In some embodiments, the methods described herein apply to recombinant VWF (rVWF).

[00134] The term “VWF propeptide” or “VWF-PP,” refers to a VWF propeptide of about 741 amino acid residues. A VWF propeptide can be part of a pre-propeptide VWF or a VWF complex. For instance, in a VWF complex a VWF propeptide is non-covalently associated with a mature VWF subunit. A VWF propeptide can be referred to as “free VWF propeptide” upon separation (isolation) from a mature VWF. When “r” is included prior to the VWF designation, this refers to the recombinant version. In some embodiments, the methods described herein apply to recombinant VWF (rVWF).

[00135] The terms “isolated,” “purified,” or “biologically pure” refer to material that is substantially or essentially free from components that normally accompany it as found in its native state. Purity and homogeneity are typically determined using analytical chemistry techniques such as polyacrylamide gel electrophoresis or high performance liquid chromatography. VWF is the predominant species present in a preparation is substantially purified. The term “purified” in some embodiments denotes that a nucleic acid or protein gives rise to essentially one band in an electrophoretic gel. In other embodiments, it means that the nucleic acid or protein is at least 50% pure, more preferably at least 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99% or more pure. “Purify” or “purification” in other embodiments means removing at least one contaminant from the composition to be purified. In this sense, purification does not require that the purified compound be homogenous, *e.g.*, 100% pure.

[00136] As used herein, the term “about” denotes an approximate range of plus or minus 10% from a specified value. For instance, the language “about 20%” encompasses a range of 18-22%.

III. Detailed Description of Embodiments

[00137] The present invention relates to a method for obtaining a highly pure composition comprising free mature recombinant von Willebrand Factor (rVWF) comprising the steps: dissociating mature rVWF from rVWF pro-peptide using a solution (*e.g.*, dissociation solution) comprising at least one chelating agent or having a pH of at least 7; separating the free mature rVWF from the rVWF pro-peptide; and collecting the free mature

rVWF composition comprising at least 95% free mature rVWF and less than 5% rVWF propeptide.

[00138] The method of the present invention is particularly suited for the in vitro separation of mature VWF from its VWF propeptide. In some embodiments, the separation is induced by adding one or more chelating agents to a solution comprising mature VWF and VWF-PP, increasing the pH of the solution to at least 7.0, or a combination thereof. In some embodiments, the pH is increased to a range from pH 7.0 to pH 9.0.

[00139] The separation method may include using one or more protein separation methods, such as, but not limited to, chromatographic methods for isolating mature VWF from VWF-PP. The method can produce a high purity, free mature rVWF composition. In some embodiments, the free mature rVWF composition comprises at least 95% free mature rVWF and less than 5% free rVWF-PP and/or matVWF/VWF-PP complex. In some cases, the free mature rVWF composition comprises at least 96% free mature rVWF and less than 4% free rVWF-PP and/or matVWF/VWF-PP complex, at least 97% free mature rVWF and less than 3% free rVWF-PP and/or matVWF/VWF-PP complex, at least 98% free mature rVWF and less than 2% free rVWF-PP and/or matVWF/VWF-PP complex, at least 99% free mature rVWF and less than 1% free rVWF-PP and/or matVWF/VWF-PP complex, at least 99.5% free mature rVWF and less than 0.5% free rVWF-PP and/or matVWF/VWF-PP complex.

a. ANION EXCHANGE CHROMATOGRAPHY PURIFICATION

[00140] In one aspect of the present method, mature rVWF (mat-rVWF) is separated from rVWF-PP using anion exchange (AEX) chromatography. In some cases, remaining host cell derived impurities such as CHO host cell proteins, process related impurities such as recombinant furin and low molecular weight viral inactivation reagents, media compounds such as soy peptone, and other product related impurities are removed from the mature VWF

[00141] In another aspect of the present method, mature rVWF is separated from rVWF-PP such as residual rVWF-PP or free rVWF-PP using anion exchange chromatography. For separation, the starting composition, loading solution, or loading composition can comprise a low pH and at least one chelating agent. The loading composition can be applied to an anion exchanger operated in flow through mode. In some

embodiments, the loading solution comprises pro-rVWF, mat-rVWF/ rVWF-PP complex, mat-rVWF, and/or rVWF propeptide (rVWF-PP). In some embodiments, the anion exchanger is operated in binding mode and mature VWF and VWF-PP are separated using a gradient elution buffer comprising at least one chelating agent. In other embodiments, the gradient elution buffer has a neutral to high pH, such as a pH ranging from pH 6.0 to pH 9.0. In another embodiment, the gradient elution buffer comprises one or more chelating agents and has a pH of 7.0 or higher, *e.g.*, pH 7.0 to pH 9.0. For instance, the gradient elution buffer can include EDTA and have a pH of 8.5.

[00142] In some embodiments, the present invention provides a method for obtaining a composition comprising a high purity, propeptide depleted mature recombinant rVWF (high purity mat-rVWF), said method comprising the steps of: (a) loading a solution comprising pro-rVWF, mat-rVWF/ rVWF-PP complex, mat-rVWF, and/or rVWF propeptide (rVWF-PP) onto an anion exchange column, wherein said pro-rVWF, mat-rVWF/ rVWF-PP complex, and mat-rVWF are bound to said anion exchange column; (b) washing said anion exchange column in a) containing said bound pro-rVWF, mat-rVWF/ rVWF-PP complex, and mat-rVWF with one or more wash buffers; (c) treating said column in b) comprising the bound pro-rVWF, mat-rVWF/ rVWF-PP complex, and mat-rVWF with furin, wherein said furin cleaves said pro-rVWF into mat-rVWF and rVWF-PP; (d) eluting said bound pro-rVWF, mat-rVWF/ rVWF-PP complex, and mat-rVWF from the column in c) with an elution buffer, wherein said elution buffer induces dissociation of said rVWF-PP from mat-rVWF non-covalently associated with said rVWF-PP, and wherein said dissociation is induced by: (i) addition of at least one chelating agent into said elution buffer, or (ii) increasing the pH of said elution buffer to a pH of at least 7; and (e) collecting said mat-rVWF separately from said rVWF-PP to obtain a high purity mat-rVWF composition, wherein said high purity mat-rVWF composition comprises at least 95% mature rVWF and less than 5% rVWF-PP.

[00143] In some embodiments, a) and b) occur simultaneously in a single step. In some embodiments, the solution in a) comprises the flow through from an immunoaffinity purification method. In some embodiments, the solution in a) comprises the flow through from a monoclonal antibody column, wherein said monoclonal antibody is a FVIII monoclonal antibody. In some embodiments, the solution in a) is selected from the group consisting of a cell culture medium, an antibody column flow-through solution, and a buffered solution.

[00144] In some embodiments of step (b) washing said anion exchange column in a) containing said bound pro-rVWF, mat-rVWF/ rVWF-PP complex, and mat-rVWF employs washing with one or more wash buffers, wherein one or more wash buffers includes one, two, three, four, and/or five wash buffers. In some embodiments, the second wash buffer comprises components for viral inactivation. In some embodiments, when four or five wash buffers are employed, the second wash buffer comprises components for viral inactivation. In some embodiments, when four or five wash buffers are employed, the second or third wash buffer comprises components for viral inactivation treatment. In some embodiments, the viral inactivation treatment is a solvent and detergent (S/D) treatment. In some embodiments, when five wash buffers are employed the first, second, third, and/or fifth wash buffers have a higher pH than the fourth wash buffer. In some embodiments, when five wash buffers are employed the first, second, third, and fifth wash buffers have a pH of about pH 7 to pH 8, and the fourth wash buffer has a pH of about pH 5 to 6. In some embodiments, when five wash buffers are employed the first, second, third, and/or fifth wash buffers have a pH of around pH 7.4 to pH 7.5, and the fourth wash buffer has a pH of about pH 5.5. In some embodiments, the viral inactivation treatment step occurs with a buffer that has a pH higher than the fourth wash buffer. In some embodiments, when four wash buffers are employed, a viral inactivation treatment step is employed after the first wash buffer. In some embodiments, when four wash buffers are employed, the first, second, and fourth wash buffers have a higher pH than the third wash buffer. In some embodiments, the viral inactivation treatment step occurs with a buffer that has a pH higher than the third wash buffer. In some embodiments, the viral inactivation step occurs with a buffer that has the same pH as the first, second, and/or fourth wash buffers. In some embodiments, when four wash buffers are employed the first, second, and fourth wash buffers have a pH of about pH 7 to about pH 8, and the third wash buffer has a pH of about pH 5 to about pH 6. In some embodiments, when four wash buffers are employed the first, second, and fourth wash buffers have a pH of about pH 7.4 to pH 7.5, and the third wash buffer has a pH of about pH 5.5.

[00145] Anion exchange chromatography can be performed as recognized by those skilled in the art. In some embodiments, the anion exchanger includes, but is not limited to, a STREAMLINE Q XL™, POROS 50 PI™, Q SEPHAROSE™, Emphase™ AEX Hybrid Purifier, Nuvia Q, POROS 50 HQ, Capto Q, Capto Q impress, Unosphere Q, Q Ceramic

HYPERD® F, TOYOPEARL® Q, TOYOPEARL® Super Q, mixed mode AEX resins (*e.g.*, Capto Adhere, Capto adhere impress, or MEP Hypercell), as well as any DEAE, TMAE, tertiary or quaternary amine, or PEI-based resins. In some embodiments, the anion exchanger is a membrane anion exchanger. In some embodiments, the membrane anion exchanger includes, but is not limited to, a Sartobind Q®, Sartobind STIC® PA, Mustang Q®, or ChromaSorb®. In some embodiments, the anion exchanger is a Fractogel TMAE column (Merck - Millipore) or an equivalent thereof.

[00146] In some embodiments, the loading concentration of pro-VWF is from about 90 IU/ml to about 270 IU/ml resin, *e.g.*, about 90 IU/ ml-about 270 IU/ml, about 100 IU/ ml-about 270 IU/ml, about 110 IU/ ml-about 270 IU/ml, about 120 IU/ ml-about 270 IU/ml, about 130 IU/ ml-about 270 IU/ml, about 130 IU/ ml-about 270 IU/ml, about 140 IU/ ml-about 270 IU/ml, about 150 IU/ ml-about 270 IU/ml, about 90 IU/ ml-about 250 IU/ml, about 100 IU/ ml-about 250 IU/ml, about 110 IU/ ml-about 250 IU/ml, about 120 IU/ ml-about 250 IU/ml, about 130 IU/ ml-about 250 IU/ml, about 130 IU/ ml-about 250 IU/ml, about 140 IU/ ml-about 250 IU/ml, about 150 IU/ ml-about 250 IU/ml, about 90 IU/ ml-about 200 IU/ml, about 100 IU/ ml-about 200 IU/ml, about 110 IU/ ml-about 200 IU/ml, about 120 IU/ ml-about 200 IU/ml, about 130 IU/ ml-about 200 IU/ml, about 130 IU/ ml-about 200 IU/ml, about 140 IU/ ml-about 200 IU/ml, about 150 IU/ ml-about 200 IU/ml, about 90 IU/ ml-about 100 IU/ml, about 100 IU/ ml-about 150 IU/ml, about 150 IU/ ml-about 200 IU/ml, about 200 IU/ ml-about 250 IU/ml, or about 250 IU/ ml-about 270 IU/ml resin.

[00147] In some embodiments, the anion exchange method comprises a buffer system. In some embodiments, the buffer system comprised one or more elution buffers. In some embodiments, the buffer system comprises one or more wash buffers. In some embodiments, the buffer system comprises at least one elution buffer and at least one wash buffer. In some embodiments, the buffer system comprises at least two elution buffers and at least two wash buffers.

[00148] In some embodiments, the loading concentration is from about 90 IU/ml to about 270 IU/ml resin, *e.g.*, about 90 IU/ ml-about 270 IU/ml, about 100 IU/ ml-about 270 IU/ml, about 110 IU/ ml-about 270 IU/ml, about 120 IU/ ml-about 270 IU/ml, about 130 IU/ ml-about 270 IU/ml, about 130 IU/ ml-about 270 IU/ml, about 140 IU/ ml-about 270 IU/ml, about 150 IU/ ml-about 270 IU/ml, about 90 IU/ ml-about 250 IU/ml, about 100 IU/ ml-about 250 IU/ml, about 110 IU/ ml-about 250 IU/ml, about 120 IU/ ml-about 250 IU/ml, about 130

IU/ ml-about 250 IU/ml, about 130 IU/ ml-about 250 IU/ml, about 140 IU/ ml-about 250 IU/ml, about 150 IU/ ml-about 250 IU/ml, about 90 IU/ ml-about 200 IU/ml, about 100 IU/ ml-about 200 IU/ml, about 110 IU/ ml-about 200 IU/ml, about 120 IU/ ml-about 200 IU/ml, about 130 IU/ ml-about 200 IU/ml, about 130 IU/ ml-about 200 IU/ml, about 140 IU/ ml-about 200 IU/ml, about 150 IU/ ml-about 200 IU/ml, about 90 IU/ ml-about 100 IU/ml, about 100 IU/ ml-about 150 IU/ml, about 150 IU/ ml-about 200 IU/ml, about 200 IU/ ml-about 250 IU/ml, or about 250 IU/ ml-about 270 IU/ml resin.

[00149] In some embodiments, the pH of the starting composition, loading solution, or loading composition comprises pro-rVWF, mat-rVWF/ rVWF-PP complex, mat-rVWF, and/or rVWF propeptide (rVWF-PP) is from pH 6.0 to pH 9.0, *e.g.*, pH 6.0–pH 9.0, pH 6.3–pH 9.0, pH 6.5–pH 9.0, pH 7.0–pH 9.0, pH 7.5–pH 9.0, pH 7.7.0–pH 9.0, pH 8.0–pH 9.0, pH 6.0–pH 8.5, pH 6.5–pH 8.5, pH 7.0–pH 8.5, pH 7.5–pH 8.5, pH 6.0–pH 8.0, pH 6.5–pH 8.0, pH 7.0–pH 8.0, pH 7.5–pH 8.0, pH 6.0, pH 6.1, pH 6.2, pH 6.3, pH 6.4, pH 6.5, pH 6.6, pH 6.7, pH 6.8, pH 6.9, pH 7.0, pH 7.1, pH 7.2, pH 7.3, pH 7.4, pH 7.5, pH 7.6, pH 7.7, pH 7.8, pH 7.9, pH 8.0, pH 8.1, pH 8.2, pH 8.3, pH 8.4, pH 8.5, pH 8.6, pH 8.7, pH 8.8, pH 8.9, or pH 9.0.

[00150] In some embodiments, the conductivity of the starting composition, loading solution, or loading composition comprises pro-rVWF, mat-rVWF/ rVWF-PP complex, mat-rVWF, and/or rVWF propeptide (rVWF-PP) is from about 5 mS/cm to about 40 mS/cm, *e.g.*, about 5 mS/cm–about 40 mS/cm, about 10 mS/cm–about 40 mS/cm, about 15 mS/cm–about 40 mS/cm, about 20 mS/cm–about 40 mS/cm, about 25 mS/cm–about 40 mS/cm, about 30 mS/cm–about 40 mS/cm, about 10 mS/cm–about 40 mS/cm, about 10 mS/cm–about 30 mS/cm, about 5 mS/cm–about 15 mS/cm, about 15 mS/cm–about 30 mS/cm, or about 20 mS/cm–about 40 mS/cm.

[00151] In some embodiments, the starting composition, loading solution, or loading composition comprises pro-rVWF, mat-rVWF/ rVWF-PP complex, mat-rVWF, and/or rVWF propeptide (rVWF-PP) is diluted with a buffer comprising sodium citrate, such as, but not limited to, 10 mM–80 mM sodium citrate, 15 mM–80 mM sodium citrate, 10 mM–80 mM sodium citrate, 15 mM–60 mM sodium citrate, 20 mM–60 mM sodium citrate, 10 mM sodium citrate, 20 mM sodium citrate, 30 mM sodium citrate, 40 mM sodium citrate, 50 mM sodium citrate, 55 mM sodium citrate, 60 mM sodium citrate, 65 mM sodium citrate, 70 mM sodium citrate, 75 mM sodium citrate, 80 mM sodium citrate, or the like.

[00152] In some embodiments, the first wash buffer comprises at least one chelating agent, and optionally has a pH ranging from pH 6.0 to pH 9.0. In some embodiments, the first wash buffer has a pH ranging from pH 6.0 to pH 9.0, and optionally comprises at least one chelating agent. In some embodiments, the first wash buffer has a pH ranging from pH 6.0 to pH 6.9. In some embodiments, the second wash buffer has a pH ranging from pH 7.0 to pH 9.0. In some embodiments, the first wash buffer can comprise at least one chelating agent and has a pH ranging from pH 6.0 to pH 6.9. In some embodiments, the wash elution buffer has a pH of less than 7. In one embodiment, the second wash buffer has a pH of greater than 7. In some embodiments, when two wash buffers are employed, the first wash buffer has a pH of less than 7 and the second wash buffer has a pH of greater than 7.

[00153] In some embodiments, the one or more wash buffers comprise a NaCl concentration of 120 mM to 200 mM, 130 mM to 200 mM, 140 mM to 200 mM, 150 mM to 200 mM, 120 mM to 190 mM, 130 mM to 190 mM, 140 mM to 190 mM, 150 mM to 190 mM, 120 mM to 180 mM, 130 mM to 180 mM, 140 mM to 180 mM, 150 mM to 180 mM, 120 mM, 125 mM, 130 mM, 135 mM, 140 mM, 145 mM, 150 mM, 155 mM, 160 mM, 165 mM, 170 mM, 175 mM, 180 mM, 185 mM, 190 mM, 195 mM, or 200 mM.

[00154] In some embodiments, the starting composition, loading solution, or loading composition comprising mature VWF and VWF-PP is contacted with a buffer comprising at least one chelating agent, and optionally the buffer has a pH of ranging from pH 6.0 to pH 9.0. In some embodiments, the starting composition, loading solution, or loading composition is contacted with a buffer having a pH ranging from pH 6.0 to pH 9.0, and optionally the buffer comprises at least one chelating agent. In some embodiments, the buffer has a pH ranging from pH 7.0 to pH 9.0. In some embodiments the buffer is a wash buffer. In some embodiments, the buffer is an elution buffer. In some embodiments the buffer is a wash buffer with a pH of 6.0 to 6.9. In some embodiments, the buffer is an elution buffer with a pH of 7.0 to 9.0. In some embodiments, the starting composition, loading solution, or loading composition comprising mature VWF and VWF-PP is contacted first with a wash buffer having a pH from 6.0 to 6.9 and a second with at least one elution buffer having a pH from 7.0 to 9.0.

[00155] In some embodiments, mature VWF is eluted in the anion exchange chromatography step using one elution buffer. In some embodiments, mature VWF is eluted in the anion exchange chromatography step using a gradient elution method comprising more

than one elution buffer. For example, the elution can be performed using two elution buffers, such as, for example, a first elution buffer and a second elution buffer. In some embodiments, the first elution buffer comprises at least one chelating agent, and optionally has a pH ranging from pH 6.0 to pH 9.0. In some embodiments, the first elution buffer has a pH ranging from pH 6.0 to pH 9.0, and optionally comprises at least one chelating agent. In some embodiments, the first elution buffer has a pH ranging from pH 6.0 to pH 6.9. In some embodiments, the second elution buffer has a pH ranging from pH 7.0 to pH 9.0. In some embodiments, the first elution buffer can comprise at least one chelating agent and has a pH ranging from pH 6.0 to pH 6.9. In some embodiments, the first elution buffer has a pH of less than 7. In one embodiment, the second elution buffer has a pH of greater than 7. In some embodiments, when two elution buffers are employed, the first elution buffer has a pH of less than 7 and the second elution buffer has a pH of greater than 7.

[00156] In some embodiments, the pH of the wash buffer for the anion exchange chromatography step is from pH 6.0 to pH 9.0, *e.g.*, pH 6.0–pH 9.0, pH 6.3–pH 9.0, pH 6.5–pH 9.0, pH 7.0–pH 9.0, pH 7.5–pH 9.0, pH 7.7.0–pH 9.0, pH 8.0–pH 9.0, pH 6.0–pH 8.5, pH 6.5–pH 8.5, pH 7.0–pH 8.5, pH 7.5–pH 8.5, pH 6.0–pH 8.0, pH 6.5–pH 8.0, pH 7.0–pH 8.0, pH 7.5–pH 8.0, pH 6.0, pH 6.1, pH 6.2, pH 6.3, pH 6.4, pH 6.5, pH 6.6, pH 6.7, pH 6.8, pH 6.9, pH 7.0, pH 7.1, pH 7.2, pH 7.3, pH 7.4, pH 7.5, pH 7.6, pH 7.7, pH 7.8, pH 7.9, pH 8.0, pH 8.1, pH 8.2, pH 8.3, pH 8.4, pH 8.5, pH 8.6, pH 8.7, pH 8.8, pH 8.9, pH 9.0. In some embodiments, this includes when there are two elution buffers, for example a first and second elution buffer.

[00157] In some embodiments, the pH of the elution buffer for the anion exchange chromatography step is from pH 6.0 to pH 9.0, *e.g.*, pH 6.0–pH 9.0, pH 6.3–pH 9.0, pH 6.5–pH 9.0, pH 7.0–pH 9.0, pH 7.5–pH 9.0, pH 7.7.0–pH 9.0, pH 8.0–pH 9.0, pH 6.0–pH 8.5, pH 6.5–pH 8.5, pH 7.0–pH 8.5, pH 7.5–pH 8.5, pH 6.0–pH 8.0, pH 6.5–pH 8.0, pH 7.0–pH 8.0, pH 7.5–pH 8.0, pH 6.0, pH 6.1, pH 6.2, pH 6.3, pH 6.4, pH 6.5, pH 6.6, pH 6.7, pH 6.8, pH 6.9, pH 7.0, pH 7.1, pH 7.2, pH 7.3, pH 7.4, pH 7.5, pH 7.6, pH 7.7, pH 7.8, pH 7.9, pH 8.0, pH 8.1, pH 8.2, pH 8.3, pH 8.4, pH 8.5, pH 8.6, pH 8.7, pH 8.8, pH 8.9, pH 9.0. In some embodiments, this includes when there are two elution buffers, for example a first and second elution buffer.

[00158] In some embodiments, the pH of the elution buffer is increased as compared to the starting solution in step a), is increased as compared to a first elution buffer when two

elution buffers are employed, and/or is increased as compared to a wash buffer when a wash buffer is employed. In some embodiments, when a wash buffer and an elution buffer is employed, the wash buffer has a pH of less than 7 and the elution buffer has a pH of greater than 7. In some embodiments, when two elution buffers are employed, one elution buffer has a pH of less than 7 and the other elution buffer has a pH of greater than 7. In some embodiments, when a wash buffer and two elution buffers are employed, the wash buffer has a pH of less than 7 and both the elution buffers have a pH of greater than 7. In some embodiments, when a wash buffer and two elution buffers are employed, the wash buffer and the first elution buffer have a pH of less than 7 and the second elution buffer has a pH of greater than 7. In some embodiments, when two wash buffers and two elution buffers are employed, the wash buffers and the first elution buffer have a pH of less than 7 and the second elution buffer has a pH of greater than 7. In some embodiments, when two wash buffers and two elution buffers are employed, both wash buffers have a pH of less than 7 and both the elution buffers have a pH of greater than 7. In some embodiments, when two wash buffers and two elution buffers are employed, the first wash buffer has a pH of less than 7 and the second wash buffer and both elution buffers have a pH of greater than 7.

[00159] In some embodiments, the pH of the one or more wash and/or elution buffers is increased to at least 7.1, 7.2, 7.3, 7.4, 7.5, 7.6, 7.7, 7.8, 7.9, or 8.0, as compared to the loading solution comprising pro-rVWF, mat-rVWF/ rVWF-PP complex, mat-rVWF, and/or rVWF propeptide (rVWF-PP), as recited in step (a) of the method. In some embodiments, the pH of the buffer is increased to at least 7.1, 7.2, 7.3, 7.4, 7.5, 7.6, 7.7, 7.8, 7.9, or 8.0 in order to induce dissociation of the mat-rVWF/ rVWF-PP complex in the solution in step (a) of the method into mat-rVWF and rVWF-PP, wherein said dissociation occurs by disruption of the non-covalently associated mat-rVWF and rVWF-PP. In some embodiments, the pH of the loading solution is increased to at least about 7.2 to about 7.8. In some embodiments, the pH of the loading solution is increased to at least about 7.6. In some embodiments, the pH of the loading solution is increased by the addition of basic amino acids. In some embodiments, the pH of at the loading solution is increased to at least 7. In some embodiments, the pH of the one or more wash buffers is increased to at least about 7.2 to about 7.8. In some embodiments, the pH of the one or more wash buffers is increased to at least about 7.6. In some embodiments, the pH of the one or more wash buffers is increased by the addition of basic amino acids. In some embodiments, the one or more wash buffers exhibit a pH of at

least 7. In some embodiments, the pH of the one or more elution buffers is increased to at least about 7.2 to about 7.8. In some embodiments, the pH of the one or more elution buffers is increased to at least about 7.6. In some embodiments, the pH of the one or more elution buffers is increased by the addition of basic amino acids. In some embodiments, the one or more elution buffers exhibit a pH of at least 7.

[00160] In some embodiments, the one or more buffers (including wash and/or elution buffers) comprise one or more chelating agents. In some embodiments, the elution buffer includes at least one chelating agent. The chelating agent can be a divalent cation chelating agent. In some embodiments, the at least one chelating agent is a divalent cation chelating agent. In some embodiments, the divalent cation chelating agent is selected from the group consisting of EDTA, EGTA, CDTA, and citrate. In some embodiments, the divalent cation chelating agent is selected from the group consisting of NTA, DTPA, EDDS, EDTA, EGTA, CDTA, and citrate. In some embodiments, the chelating agent is NTA. In some embodiments, the chelating agent is DTPA. In some embodiments, the chelating agent is EDDS. In some embodiments, the chelating agent is EDTA. In some embodiments, the chelating agent is EGTA. In some embodiments, the chelating agent is CDTA. In some embodiments, the chelating agent is citrate. In some embodiments, the one or more wash buffers in b) comprise said one or more chelating agents and exhibit a pH of at least 7.

[00161] In some embodiments, the one or more buffers (including wash and/or elution buffers) comprise sodium citrate in a range including but not limited to, 10 mM-80 mM sodium citrate, 15 mM-80 mM sodium citrate, 10 mM-80 mM sodium citrate, 15 mM-60 mM sodium citrate, 20 mM-60 mM sodium citrate, 10 mM sodium citrate, 20 mM sodium citrate, 30 mM sodium citrate, 40 mM sodium citrate, 50 mM sodium citrate, 55 mM sodium citrate, 60 mM sodium citrate, 65 mM sodium citrate, 70 mM sodium citrate, 75 mM sodium citrate, 80 mM sodium citrate, or the like.

[00162] In some embodiments, a first elution buffer further comprises sodium citrate, in a range including but not limited to, 10 mM-60 mM sodium citrate, 15 mM-60 mM sodium citrate, 10 mM-50 mM sodium citrate, 15 mM-50 mM sodium citrate, 20 mM-60 mM sodium citrate, 10 mM sodium citrate, 20 mM sodium citrate, 30 mM sodium citrate, 40 mM sodium citrate, 50 mM sodium citrate, 60 mM sodium citrate, or the like.

[00163] In some embodiments, a second elution buffer further comprises sodium citrate, such as, but not limited to, 10 mM-60 mM sodium citrate, 15 mM-60 mM sodium

citrate, 10 mM-50 mM sodium citrate, 15 mM-50 mM sodium citrate, 20 mM-60 mM sodium citrate, 10 mM sodium citrate, 20 mM sodium citrate, 30 mM sodium citrate, 40 mM sodium citrate, 50 mM sodium citrate, 60 mM sodium citrate, or the like.

[00164] In some embodiments, the elution buffer A and/or elution buffer B of the anion exchange chromatography step comprises about 0.5 mM to about 20 mM EDTA, *e.g.*, about 0.5 mM-about 20 mM, about 1 mM-about 20 mM, about 1.5 mM-about 20 mM, about 2 mM-about 20 mM, about 3 mM-about 20 mM, about 5 mM-about 20 mM, about 0.5 mM-about 15 mM, about 1 mM-about 10 mM, about 1 mM-about 5 mM, about 5 mM, about 0.5 mM, about 1 mM, about 2 mM, about 3 mM, about 4 mM, about 5 mM, about 6 mM, about 7 mM, about 8 mM, about 9 mM, about 10 mM, about 11 mM, about 12 mM, about 13 mM, about 14 mM, about 15 mM, about 16 mM, about 17 mM, about 18 mM, about 19 mM, about 20 mM, or the like.

[00165] In some embodiments, the citrate can be found in the eluent after the rVWF-propeptide has been removed using an anion exchange method. In some embodiments, the citrate can be found in the eluent after the rVWF-propeptide has been removed using a stepwise anion exchange elution method. In some embodiments, the citrate can be found in the eluent after the rVWF-propeptide has been removed using a gradient anion exchange elution method. In some embodiments, the anion exchange counter-ion is citrate³⁻.

[00166] Any of the buffers (buffer systems) described herein can be selected from the group consisting of glycine, HEPES (4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid), TrisHCl (Tris(hydroxymethyl)-aminomethane), histidine, imidazole, acetate citrate, citrate, acetate, MES, phosphate, TrisHCl, Bis-Tris, Histidine, Imidazol, ArgininHCl, LysinHCl, and 2-(N-morpholino)ethanesulfonic acid, as single buffers or as a combination of two or more buffers. In some embodiments, the buffer comprises glycine. In some embodiments, the buffer comprises HEPES (4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid). In some embodiments, the buffer comprises TrisHCl (Tris(hydroxymethyl)-aminomethane). In some embodiments, the buffer comprises histidine. In some embodiments, the buffer comprises imidazole. In some embodiments, the buffer comprises acetate citrate. In some embodiments, the buffer comprises citrate. In some embodiments, the buffer comprises acetate. In some embodiments, the buffer comprises MES. In some embodiments, the buffer comprises phosphate. In some embodiments, the buffer comprises TrisHCl. In some embodiments, the buffer comprises Bis-Tris. In some embodiments, the buffer comprises

Histidine. In some embodiments, the buffer comprises Imidazole. In some embodiments, the buffer comprises Arginine HCl. In some embodiments, the buffer comprises LysinHCl. In some embodiments, the buffer comprises 2-(N-morpholino)ethanesulfonic acid. In some embodiments, the buffer comprises one, two, three, or four of the buffers listed herein.

[00167] In some embodiments, the one or more buffers are selected from the group consisting of glycine HEPES (4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid), TrisHCl (Tris(hydroxymethyl)-aminomethane), histidine, imidazole, acetate citrate, MES, and 2-(N-morpholino)ethanesulfonic acid.

[00168] In some embodiments, the one or more buffers comprise at least one buffer exhibiting a conductivity of ≥ 0.5 mS/cm at 25°C. In some embodiments, the one or more buffers comprise at least one buffer exhibiting a conductivity of 20.0 ± 0.2 mS/cm at 25°C. In some embodiments, the one or more buffers comprise at least one buffer exhibiting a conductivity of 17.0 ± 0.2 mS/cm at 25°C. In some embodiments, the one or more buffers comprise at least one buffer exhibiting a conductivity of 15.0 ± 0.2 mS/cm at 25°C. In some embodiments, the one or more buffers comprise at least one buffer exhibiting a conductivity of 12.0 ± 0.2 mS/cm at 25°C. In some embodiments, the one or more buffers comprise at least one buffer exhibiting a conductivity of 10.0 ± 0.2 mS/cm at 25°C. In some embodiments, the one or more buffers comprise at least one buffer exhibiting a conductivity of 5.0 ± 0.2 mS/cm at 25°C. In some embodiments, the one or more buffers comprise at least one buffer exhibiting a conductivity of 2.0 ± 0.2 mS/cm at 25°C.

[00169] In some embodiments, the flow rate of one or more wash steps of the present method is about 10 cm/h to about 200 cm/h, *e.g.*, about 10 cm/h, about 15 cm/h, about 20 cm/h, about 25 cm/h, about 30 cm/h, about 35 cm/h, about 40 cm/h, about 45 cm/h, about 50 cm/h, about 55 cm/h, about 60 cm/h, about 65 cm/h, about 70 cm/h, about 75 cm/h, about 80 cm/h, about 85 cm/h, about 90 cm/h, about 95 cm/h, about 100 cm/h, about 105 cm/h, about 110 cm/h, about 115 cm/h, about 120 cm/h, about 125 cm/h, about 130 cm/h, about 135 cm/h, about 140 cm/h, about 145 cm/h, about 150 cm/h, about 155 cm/h, about 160 cm/h, about 165 cm/h, about 170 cm/h, about 175 cm/h, about 180 cm/h, about 185 cm/h, about 190 cm/h, about 195 cm/h, or about 200 cm/h. Depending on the resin, in some embodiments the flow rate can be up to 600 cm/h.

[00170] In some embodiments, the flow rate of one or more elution steps of the present method is about 10 cm/h to about 200 cm/h, *e.g.*, about 10 cm/h, about 15 cm/h, about 20

cm/h, about 25 cm/h, about 30 cm/h, about 35 cm/h, about 40 cm/h, about 45 cm/h, about 50 cm/h, about 55 cm/h, about 60 cm/h, about 65 cm/h, about 70 cm/h, about 75 cm/h, about 80 cm/h, about 85 cm/h, about 90 cm/h, about 95 cm/h, about 100 cm/h, about 105 cm/h, about 110 cm/h, about 115 cm/h, about 120 cm/h, about 125 cm/h, about 130 cm/h, about 135 cm/h, about 140 cm/h, about 145 cm/h, about 150 cm/h, about 155 cm/h, about 160 cm/h, about 165 cm/h, about 170 cm/h, about 175 cm/h, about 180 cm/h, about 185 cm/h, about 190 cm/h, about 195 cm/h, or about 200 cm/h. Depending on the resin, in some embodiments the flow rate can be up to 600 cm/h.

[00171] In some embodiments, the one or more buffers further comprise one or more nonionic detergents. In some embodiments, the nonionic detergent is selected from the group consisting of Triton X-100, Tween 80, and Tween 20. In some embodiments, the nonionic detergent is Triton X-100. In some embodiments, the nonionic detergent is Tween 80. In some embodiments, the nonionic detergent is Tween 20.

[00172] In some embodiments, the said one or more buffers further comprise one or more additional substances selected from the group consisting of non-reducing sugars, sugar alcohols, and polyols. In some embodiments, the one or more buffers further comprises one or more non-reducing sugars. In some embodiments, the non-reducing sugar includes but is not limited to sucrose, trehalose, mannitol, sorbitol, galactitol, and/or xylitol. In some embodiments, the one or more buffers further comprises one or more sugar alcohols. In some embodiments, the one or more buffers further comprises one or more polyols. In some embodiments, the sugar alcohol or polyol includes but is not limited to mannitol, xylitol, erythritol, threitol, sorbitol, and/or glycerol. In some embodiments, the buffers further comprise ethylene glycol, propylene glycol, glycerol, 1,2,3-Propanetriol, meso-erythritol, and/or erythritol (meso-1,2,3,4-Butantetrol).

[00173] In some embodiments, the buffer can include one or more monovalent cations. In some embodiments, the one or more monovalent cations are selected from the group consisting of Na^+ , K^+ , Li^+ , Cs^+ , and NH_4^+ . For instance, the monovalent cation can be Na^+ . In other embodiments, the buffer includes one or more monovalent, divalent and/or trivalent anions. The one or more monovalent, divalent and/or trivalent anions can be selected from the group consisting of Cl^- , acetate⁻, SO_4^{2-} , Br^- , citrate³⁻, PO_4^{3-} , and BO_3^{3-} . In some embodiments, the buffer comprises one or more additional substances selected from the group consisting of non-reducing sugars, and sugar alcohols. In some embodiments, the one

or more buffers further comprise one or more monovalent cations. In some embodiments, the one or more monovalent cations are selected from the group consisting of Na^+ , K^+ , Li^+ , and Cs^+ . In some embodiments, the monovalent cation is Na^+ . In some embodiments, the one or more buffers further comprise one or more monovalent, divalent, and/or trivalent anions. In some embodiments, the one or more monovalent, divalent and/or trivalent anions are selected from the group consisting of Cl^- , acetate $^-$, SO_4^{2-} , Br^- , and citrate $^{3-}$.

[00174] The pH of any of the buffers can be adjusted (increased) by adding an amino acid, Tris, NaOH, ethanolamine, and the like.

[00175] In some embodiments, the anion exchange method buffer chelator combination comprises citrate, malate (malic acid), and tartrate (tartaric acid).

b. CATION EXCHANGE CHROMATOGRAPHY PURIFICATION

[00176] In one aspect of the present method, mature VWF(matVWF) is separated from VWF-PP using cation exchange (CEX) chromatography. In some cases, remaining host cell derived impurities such as CHO host cell proteins, process related impurities such as recombinant furin and low molecular weight viral inactivation reagents, media compounds such as soy peptone, and other product related impurities are removed from the mature VWF.

[00177] In another aspect of the present method, mature VWF is separated from VWF-PP such as residual VWF-PP or free VWF-PP using cation exchange chromatography. For separation, the starting composition, loading solution, or loading composition can comprise a low pH and at least one chelating agent. In some embodiments, the starting composition, loading solution, or loading composition comprises pro-rVWF, mat-rVWF/ rVWF-PP complex, mat-rVWF, and/or rVWF propeptide (rVWF-PP). In some embodiments, the cation exchanger is operated in binding mode and mature VWF and VWF-PP are separated using a gradient elution buffer comprising at least one chelating agent. In other embodiments, the gradient elution buffer has a neutral to high pH, such as a pH ranging from pH 6.0 to pH 9.0. In another embodiment, the gradient elution buffer comprises one or more chelating agents and has a pH of 7.0 or higher, *e.g.*, pH 7.0 to pH 9.0. For instance, the gradient elution buffer can include EDTA and have a pH of 8.5.

[00178] In some embodiments, the present invention provides a method for obtaining a composition comprising a high purity, propeptide depleted mature recombinant rVWF (high

purity mat-rVWF), said method comprising the steps of: (a) loading a solution comprising pro-rVWF, mat-rVWF/ rVWF-PP complex, mat-rVWF, and/or rVWF propeptide (rVWF-PP) onto a cation exchange column, wherein said pro-rVWF, mat-rVWF/ rVWF-PP complex, and mat-rVWF are bound to said cation exchange column; (b) washing said cation exchange column in a) containing said bound pro-rVWF, mat-rVWF/ rVWF-PP complex, and mat-rVWF with one or more wash buffers; (c) treating said column in b) comprising the bound pro-rVWF, mat-rVWF/ rVWF-PP complex, and mat-rVWF with furin, wherein said furin cleaves said pro-rVWF into mat-rVWF and rVWF-PP; (d) eluting said bound pro-rVWF, mat-rVWF/ rVWF-PP complex, and mat-rVWF from the column in c) with an elution buffer, wherein said elution buffer induces dissociation of said rVWF-PP from mat-rVWF non-covalently associated with said rVWF-PP, and wherein said dissociation is induced by: (i) addition of at least one chelating agent into said elution buffer, or (ii) increasing the pH of said elution buffer to a pH of at least 7; and (e) collecting said mat-rVWF separately from said rVWF-PP to obtain a high purity mat-rVWF composition, wherein said high purity mat-rVWF composition comprises at least 95% mature rVWF and less than 5% rVWF-PP.

[00179] In some embodiments, a) and b) occur simultaneously in a single step. In some embodiments, the solution in a) comprises the flow through from an immunoaffinity purification method. In some embodiments, the solution in a) comprises the flow through from a monoclonal antibody column, wherein said monoclonal antibody is a FVIII monoclonal antibody. In some embodiments, the solution in a) is selected from the group consisting of a cell culture medium, an antibody column flow-through solution, and a buffered solution.

[00180] The cation exchanger can be operated in binding mode to separate the mature VWF and VWF-PP. Cation exchange chromatography can be performed as recognized by those skilled in the art. In some embodiments, the cation exchanger includes, but is not limited to, POROS® S (Applied Biosystems), Convective Interaction Media (CIM®; BIA Separation), Toyopearl Gigacap S (Tosoh Bioscience, Montgomeryville, PA), Toyopearl Gigacap CM (Tosoh), Toyopearl SP (Tosoh), Toyopearl CM (Tosoh), MacroPrep S (Bio-rad, Hercules, CA), UNOsphereS (Bio-rad, Hercules, CA), MacroPrepCM ((Bio-rad, Hercules, CA), Fractogel EMD SO3 (Merck), Fractogel EMD COO (Merck), Fractogel EMD SE Hicap (Merck), Cellufine Sulfate (JNC), CM and SP Trisacryl (Pall), CM and S HyperD (Pall), S and CM Sepharose CL (GE Healthcare), S and CM Sepharose FF (GE Healthcare),

S and CM CAPTO™ (GE Healthcare), MonoS (GE Healthcare), Source S (GE Healthcare), Nuvia S (Merck), or Cellufine phosphate (JNC). In some embodiments, the cation exchanger is a membrane cation exchanger. In some embodiments, the membrane cation exchanger includes, but is not limited to, Mustang S (Pall) or Sartobind® S. In some embodiments, the cation exchanger is a UNO_Sphere S column (Bio-Rad) or an equivalent thereof.

[00181] In some embodiments of step (b) washing said cation exchange column in a) containing said bound pro-rVWF, mat-rVWF/ rVWF-PP complex, and mat-rVWF employs washing with one or more wash buffers, wherein one or more wash buffers includes one, two, three, four, and/or five wash buffers. In some embodiments, the second wash buffer comprises components for viral inactivation. In some embodiments, when four or five wash buffers are employed, the second wash buffer comprises components for viral inactivation. In some embodiments, when four or five wash buffers are employed, the second or third wash buffer comprises components for viral inactivation treatment. In some embodiments, the viral inactivation treatment is a solvent and detergent (S/D) treatment. In some embodiments, when five wash buffers are employed the first, second, third, and/or fifth wash buffers have a higher pH than the fourth wash buffer. In some embodiments, when five wash buffers are employed the first, second, third, and fifth wash buffers have a pH of about pH 7 to pH 8, and the fourth wash buffer has a pH of about pH 5 to 6. In some embodiments, when five wash buffers are employed the first, second, third, and/or fifth wash buffers have a pH of around pH 7.4 to pH 7.5, and the fourth wash buffer has a pH of about pH 5.5. In some embodiments, the viral inactivation treatment step occurs with a buffer that has a pH higher than the fourth wash buffer. In some embodiments, when four wash buffers are employed, a viral inactivation treatment step is employed after the first wash buffer. In some embodiments, when four wash buffers are employed, the first, second, and fourth wash buffers have a higher pH than the third wash buffer. In some embodiments, the viral inactivation treatment step occurs with a buffer that has a pH higher than the third wash buffer. In some embodiments, the viral inactivation step occurs with a buffer that has the same pH as the first, second, and/or fourth wash buffers. In some embodiments, when four wash buffers are employed the first, second, and fourth wash buffers have a pH of about pH 7 to about pH 8, and the third wash buffer has a pH of about pH 5 to about pH 6. In some embodiments, when four wash buffers are employed the first, second, and fourth wash

buffers have a pH of about pH 7.4 to pH 7.5, and the third wash buffer has a pH of about pH 5.5.

[00182] In some embodiments, the loading concentration of pro-VWF is from about 90 IU/ml to about 270 IU/ml resin, *e.g.*, about 90 IU/ ml-about 270 IU/ml, about 100 IU/ ml-about 270 IU/ml, about 110 IU/ ml-about 270 IU/ml, about 120 IU/ ml-about 270 IU/ml, about 130 IU/ ml-about 270 IU/ml, about 130 IU/ ml-about 270 IU/ml, about 140 IU/ ml-about 270 IU/ml, about 150 IU/ ml-about 270 IU/ml, about 90 IU/ ml-about 250 IU/ml, about 100 IU/ ml-about 250 IU/ml, about 110 IU/ ml-about 250 IU/ml, about 120 IU/ ml-about 250 IU/ml, about 130 IU/ ml-about 250 IU/ml, about 130 IU/ ml-about 250 IU/ml, about 140 IU/ ml-about 250 IU/ml, about 150 IU/ ml-about 250 IU/ml, about 90 IU/ ml-about 200 IU/ml, about 100 IU/ ml-about 200 IU/ml, about 110 IU/ ml-about 200 IU/ml, about 120 IU/ ml-about 200 IU/ml, about 130 IU/ ml-about 200 IU/ml, about 130 IU/ ml-about 200 IU/ml, about 140 IU/ ml-about 200 IU/ml, about 150 IU/ ml-about 200 IU/ml, about 90 IU/ ml-about 100 IU/ml, about 100 IU/ ml-about 150 IU/ml, about 150 IU/ ml-about 200 IU/ml, about 200 IU/ ml-about 250 IU/ml, or about 250 IU/ ml-about 270 IU/ml resin.

[00183] In some embodiments, the cation exchange method comprises a buffer system. In some embodiments, the buffer system comprised one or more elution buffers. In some embodiments, the buffer system comprises one or more wash buffers. In some embodiments, the buffer system comprises at least one elution buffer and at least one wash buffer. In some embodiments, the buffer system comprises at least two elution buffers and at least two wash buffers.

[00184] In some embodiments, the first wash buffer comprises at least one chelating agent, and optionally has a pH ranging from pH 6.0 to pH 9.0. In some embodiments, the first wash buffer has a pH ranging from pH 6.0 to pH 9.0, and optionally comprises at least one chelating agent. In some embodiments, the first wash buffer has a pH ranging from pH 6.0 to pH 6.9. In some embodiments, the second wash buffer has a pH ranging from pH 7.0 to pH 9.0. In some embodiments, the first wash buffer can comprise at least one chelating agent and has a pH ranging from pH 6.0 to pH 6.9. In some embodiments, the wash elution buffer has a pH of less than 7. In one embodiments, the second wash buffer has a pH of greater than 7. In some embodiments, when two wash buffers are employed, the first wash buffer has a pH of less than 7 and the second wash buffer has a pH of greater than 7.

[00185] In some embodiments, the one or more wash buffers comprise a NaCl concentration of 120 mM to 200 mM, 130 mM to 200 mM, 140 mM to 200 mM, 150 mM to 200 mM, 120 mM to 190 mM, 130 mM to 190 mM, 140 mM to 190 mM, 150 mM to 190 mM, 120 mM to 180 mM, 130 mM to 180 mM, 140 mM to 180 mM, 150 mM to 180 mM, 120 mM, 125 mM, 130 mM, 135 mM, 140 mM, 145 mM, 150 mM, 155 mM, 160 mM, 165 mM, 170 mM, 175 mM, 180 mM, 185 mM, 190 mM, 195 mM, or 200 mM.

[00186] In some embodiments, the starting composition, loading solution, or loading composition comprising mature VWF and VWF-PP is contacted with a buffer comprising at least one chelating agent, and optionally the buffer has a pH of ranging from pH 6.0 to pH 9.0. In some embodiments, the starting composition, loading solution, or loading composition is contacted with a buffer having a pH ranging from pH 6.0 to pH 9.0, and optionally the buffer comprises at least one chelating agent. In some embodiments, the buffer has a pH ranging from pH 7.0 to pH 9.0. In some embodiments the buffer is a wash buffer. In some embodiments, the buffer is an elution buffer. In some embodiments the buffer is a wash buffer with a pH of 6.0 to 6.9. In some embodiments, the buffer is an elution buffer with a pH of 7.0 to 9.0. In some embodiments, the starting composition, loading solution, or loading composition comprising mature VWF and VWF-PP is contacted first with a wash buffer having a pH from 6.0 to 6.9 and a second with at least one elution buffer having a pH from 7.0 to 9.0.

[00187] In some embodiments, mature VWF is eluted in the anion exchange chromatography step using one elution buffer. In some embodiments, mature VWF is eluted in the anion exchange chromatography step using a gradient elution method comprising more than one elution buffer. For example, the elution can be performed using two elution buffers, such as, for example, a first elution buffer and a second elution buffer. In some embodiments, the first elution buffer comprises at least one chelating agent, and optionally has a pH ranging from pH 6.0 to pH 9.0. In some embodiments, the first elution buffer has a pH ranging from pH 6.0 to pH 9.0, and optionally comprises at least one chelating agent. In some embodiments, the first elution buffer has a pH ranging from pH 6.0 to pH 6.9. In some embodiments, the second elution buffer has a pH ranging from pH 7.0 to pH 9.0. In some embodiments, the first elution buffer can comprise at least one chelating agent and has a pH ranging from pH 6.0 to pH 6.9. In some embodiments, the first elution buffer has a pH of less than 7. In one embodiment, the second elution buffer has a pH of greater than 7. In

some embodiments, when two elution buffers are employed, the first elution buffer has a pH of less than 7 and the second elution buffer has a pH of greater than 7.

[00188] In some embodiments, the pH of the wash buffer for the cation exchange chromatography step is from pH 6.0 to pH 9.0, *e.g.*, pH 6.0–pH 9.0, pH 6.3–pH 9.0, pH 6.5–pH 9.0, pH 7.0–pH 9.0, pH 7.5–pH 9.0, pH 7.7–pH 9.0, pH 8.0–pH 9.0, pH 6.0–pH 8.5, pH 6.5–pH 8.5, pH 7.0–pH 8.5, pH 7.5–pH 8.5, pH 6.0–pH 8.0, pH 6.5–pH 8.0, pH 7.0–pH 8.0, pH 7.5–pH 8.0, pH 6.0, pH 6.1, pH 6.2, pH 6.3, pH 6.4, pH 6.5, pH 6.6, pH 6.7, pH 6.8, pH 6.9, pH 7.0, pH 7.1, pH 7.2, pH 7.3, pH 7.4, pH 7.5, pH 7.6, pH 7.7, pH 7.8, pH 7.9, pH 8.0, pH 8.1, pH 8.2, pH 8.3, pH 8.4, pH 8.5, pH 8.6, pH 8.7, pH 8.8, pH 8.9, pH 9.0. In some embodiments, this includes when there are two elution buffers, for example a first and second elution buffer.

[00189] In some embodiments, the pH of the elution buffer for the cation exchange chromatography step is from pH 6.0 to pH 9.0, *e.g.*, pH 6.0–pH 9.0, pH 6.3–pH 9.0, pH 6.5–pH 9.0, pH 7.0–pH 9.0, pH 7.5–pH 9.0, pH 7.7–pH 9.0, pH 8.0–pH 9.0, pH 6.0–pH 8.5, pH 6.5–pH 8.5, pH 7.0–pH 8.5, pH 7.5–pH 8.5, pH 6.0–pH 8.0, pH 6.5–pH 8.0, pH 7.0–pH 8.0, pH 7.5–pH 8.0, pH 6.0, pH 6.1, pH 6.2, pH 6.3, pH 6.4, pH 6.5, pH 6.6, pH 6.7, pH 6.8, pH 6.9, pH 7.0, pH 7.1, pH 7.2, pH 7.3, pH 7.4, pH 7.5, pH 7.6, pH 7.7, pH 7.8, pH 7.9, pH 8.0, pH 8.1, pH 8.2, pH 8.3, pH 8.4, pH 8.5, pH 8.6, pH 8.7, pH 8.8, pH 8.9, pH 9.0. In some embodiments, this includes when there are two elution buffers, for example a first and second elution buffer.

[00190] In some embodiments, the pH of the elution buffer is increased as compared to the starting solution in step a), is increased as compared to a first elution buffer when two elution buffers are employed, and/or is increased as compared to a wash buffer when a wash buffer is employed. In some embodiments, when a wash buffer and an elution buffer is employed, the wash buffer has a pH of less than 7 and the elution buffer has a pH of greater than 7. In some embodiments, when two elution buffers are employed, one elution buffer has a pH of less than 7 and the other elution buffer has a pH of greater than 7. In some embodiments, when a wash buffer and two elution buffers are employed, the wash buffer has a pH of less than 7 and both the elution buffers have a pH of greater than 7. In some embodiments, when a wash buffer and two elution buffers are employed, the wash buffer and the first elution buffer have a pH of less than 7 and the second elution buffer has a pH of greater than 7. In some embodiments, when two wash buffers and two elution buffers are

employed, the wash buffers and the first elution buffer have a pH of less than 7 and the second elution buffer has a pH of greater than 7. In some embodiments, when two wash buffers and two elution buffers are employed, both wash buffers have a pH of less than 7 and both the elution buffers have a pH of greater than 7. In some embodiments, when two wash buffers and two elution buffers are employed, the first wash buffer has a pH of less than 7 and the second wash buffer and both elution buffers have a pH of greater than 7. .

[00191] In some embodiments, the pH of the one or more wash and/or elution buffers is increased to at least 7.1, 7.2, 7.3, 7.4, 7.5, 7.6, 7.7, 7.8, 7.9, or 8.0, as compared to the loading solution comprising pro-rVWF, mat-rVWF/ rVWF-PP complex, mat-rVWF, and/or rVWF propeptide (rVWF-PP), as recited in step (a) of the method. In some embodiments, the pH of the buffer is increased to at least 7.1, 7.2, 7.3, 7.4, 7.5, 7.6, 7.7, 7.8, 7.9, or 8.0 in order to induce dissociation of the mat-rVWF/ rVWF-PP complex in the solution in step (a) of the method into mat-rVWF and rVWF-PP, wherein said dissociation occurs by disruption of the non-covalently associated mat-rVWF and rVWF-PP. In some embodiments, the pH of the loading solution is increased to at least about 7.2 to about 7.8. In some embodiments, the pH of the loading solution is increased to at least about 7.6. In some embodiments, the pH of the loading solution is increased by the addition of basic amino acids. In some embodiments, the pH of at the loading solution is increased to at least 7. In some embodiments, the pH of the one or more wash buffers is increased to at least about 7.2 to about 7.8. In some embodiments, the pH of the one or more wash buffers is increased to at least about 7.6. In some embodiments, the pH of the one or more wash buffers is increased by the addition of basic amino acids. In some embodiments, the one or more wash buffers exhibit a pH of at least 7. In some embodiments, the pH of the one or more elution buffers is increased to at least about 7.2 to about 7.8. In some embodiments, the pH of the one or more elution buffers is increased to at least about 7.6. In some embodiments, the pH of the one or more elution buffers is increased by the addition of basic amino acids. In some embodiments, the one or more elution buffers exhibit a pH of at least 7.

[00192] In some embodiments, the pH of the loading solution comprising pro-rVWF, mat-rVWF/ rVWF-PP complex, mat-rVWF, and/or rVWF propeptide (rVWF-PP) is from pH 6.0 to pH 9.0, *e.g.*, pH 6.0–pH 9.0, pH 6.3–pH 9.0, pH 6.5–pH 9.0, pH 7.0–pH 9.0, pH 7.5–pH 9.0, pH 7.7.0–pH 9.0, pH 8.0–pH 9.0, pH 6.0–pH 8.5, pH 6.5–pH 8.5, pH 7.0–pH 8.5, pH 7.5–pH 8.5, pH 6.0–pH 8.0, pH 6.5–pH 8.0, pH 7.0–pH 8.0, pH 7.5–pH 8.0, pH 6.0, pH 6.1,

pH 6.2, pH 6.3, pH 6.4, pH 6.5, pH 6.6, pH 6.7, pH 6.8, pH 6.9, pH 7.0, pH 7.1, pH 7.2, pH 7.3, pH 7.4, pH 7.5, pH 7.6, pH 7.7, pH 7.8, pH 7.9, pH 8.0, pH 8.1, pH 8.2, pH 8.3, pH 8.4, pH 8.5, pH 8.6, pH 8.7, pH 8.8, pH 8.9, or pH 9.0.

[00193] In some embodiments, the conductivity of the starting composition, loading solution, or loading composition comprising pro-rVWF, mat-rVWF/ rVWF-PP complex, mat-rVWF, and/or rVWF propeptide (rVWF-PP) is from about 5 mS/cm to about 40 mS/cm, *e.g.*, about 5 mS/cm–about 40 mS/cm, about 10 mS/cm–about 40 mS/cm, about 15 mS/cm–about 40 mS/cm, about 20 mS/cm–about 40 mS/cm, about 25 mS/cm–about 40 mS/cm, about 30 mS/cm–about 40 mS/cm, about 10 mS/cm–about 40 mS/cm, about 10 mS/cm–about 30 mS/cm, about 5 mS/cm–about 15 mS/cm, about 15 mS/cm–about 30 mS/cm, or about 20 mS/cm–about 40 mS/cm.

[00194] In some embodiments, the loading solution comprising pro-rVWF, mat-rVWF/ rVWF-PP complex, mat-rVWF, and/or rVWF propeptide (rVWF-PP) is diluted with a buffer comprising sodium citrate, such as, but not limited to, 10 mM–80 mM sodium citrate, 15 mM–80 mM sodium citrate, 10 mM–80 mM sodium citrate, 15 mM–60 mM sodium citrate, 20 mM–60 mM sodium citrate, 10 mM sodium citrate, 20 mM sodium citrate, 30 mM sodium citrate, 40 mM sodium citrate, 50 mM sodium citrate, 55 mM sodium citrate, 60 mM sodium citrate, 65 mM sodium citrate, 70 mM sodium citrate, 75 mM sodium citrate, 80 mM sodium citrate, or the like.

[00195] In some embodiments, the pH of the wash buffer for the cation exchange chromatography step is from pH 6.0 to pH 9.0, *e.g.*, pH 6.0–pH 9.0, pH 6.3–pH 9.0, pH 6.5–pH 9.0, pH 7.0–pH 9.0, pH 7.5–pH 9.0, pH 7.7.0–pH 9.0, pH 8.0–pH 9.0, pH 6.0–pH 8.5, pH 6.5–pH 8.5, pH 7.0–pH 8.5, pH 7.5–pH 8.5, pH 6.0–pH 8.0, pH 6.5–pH 8.0, pH 7.0–pH 8.0, pH 7.5–pH 8.0, pH 6.0, pH 6.1, pH 6.2, pH 6.3, pH 6.4, pH 6.5, pH 6.6, pH 6.7, pH 6.8, pH 6.9, pH 7.0, pH 7.1, pH 7.2, pH 7.3, pH 7.4, pH 7.5, pH 7.6, pH 7.7, pH 7.8, pH 7.9, pH 8.0, pH 8.1, pH 8.2, pH 8.3, pH 8.4, pH 8.5, pH 8.6, pH 8.7, pH 8.8, pH 8.9, or pH 9.0.

[00196] In some embodiments, the pH of the elution buffer for the cation exchange chromatography step is from pH 6.0 to pH 9.0, *e.g.*, pH 6.0–pH 9.0, pH 6.3–pH 9.0, pH 6.5–pH 9.0, pH 7.0–pH 9.0, pH 7.5–pH 9.0, pH 7.7.0–pH 9.0, pH 8.0–pH 9.0, pH 6.0–pH 8.5, pH 6.5–pH 8.5, pH 7.0–pH 8.5, pH 7.5–pH 8.5, pH 6.0–pH 8.0, pH 6.5–pH 8.0, pH 7.0–pH 8.0, pH 7.5–pH 8.0, pH 6.0, pH 6.1, pH 6.2, pH 6.3, pH 6.4, pH 6.5, pH 6.6, pH 6.7, pH 6.8, pH

6.9, pH 7.0, pH 7.1, pH 7.2, pH 7.3, pH 7.4, pH 7.5, pH 7.6, pH 7.7, pH 7.8, pH 7.9, pH 8.0, pH 8.1, pH 8.2, pH 8.3, pH 8.4, pH 8.5, pH 8.6, pH 8.7, pH 8.8, pH 8.9, or pH 9.0.

[00197] In some embodiments, the pH of the elution buffer is increased as compared to the starting solution in step a), is increased as compared to a first elution buffer when two elution buffers are employed, and/or is increased as compared to a wash buffer when a wash buffer is employed. In some embodiments, when a wash buffer and an elution buffer is employed, the wash buffer has a pH of less than 7 and the elution buffer has a pH of greater than 7. In some embodiments, when two elution buffers are employed, one elution buffer has a pH of less than 7 and the other elution buffer has a pH of greater than 7. In some embodiments, when a wash buffer and two elution buffers are employed, the wash buffer has a pH of less than 7 and both the elution buffers have a pH of greater than 7. In some embodiments, when a wash buffer and two elution buffers are employed, the wash buffer and the first elution buffer have a pH of less than 7 and the second elution buffer has a pH of greater than 7. In some embodiments, when two wash buffers and two elution buffers are employed, the wash buffers and the first elution buffer have a pH of less than 7 and the second elution buffer has a pH of greater than 7. In some embodiments, when two wash buffers and two elution buffers are employed, both wash buffers have a pH of less than 7 and both the elution buffers have a pH of greater than 7. In some embodiments, when two wash buffers and two elution buffers are employed, the first wash buffer has a pH of less than 7 and the second wash buffer and both elution buffers have a pH of greater than 7. .

[00198] In some embodiments, the pH of the one or more wash and/or elution buffers is increased to at least 7.1, 7.2, 7.3, 7.4, 7.5, 7.6, 7.7, 7.8, 7.9, or 8.0, as compared to the loading solution comprising pro-rVWF, mat-rVWF/ rVWF-PP complex, mat-rVWF, and/or rVWF propeptide (rVWF-PP), as recited in step (a) of the method. In some embodiments, the pH of the buffer is increased to at least 7.1, 7.2, 7.3, 7.4, 7.5, 7.6, 7.7, 7.8, 7.9, or 8.0 in order to induce dissociation of the mat-rVWF/ rVWF-PP complex in the solution in step (a) of the method into mat-rVWF and rVWF-PP, wherein said dissociation occurs by disruption of the non-covalently associated mat-rVWF and rVWF-PP. In some embodiments, the pH of the loading solution is increased to at least about 7.2 to about 7.8. In some embodiments, the pH of the loading solution is increased to at least about 7.6. In some embodiments, the pH of the loading solution is increased by the addition of basic amino acids. In some embodiments, the pH of at the loading solution is increased to at least 7. In some embodiments, the pH of

the one or more wash buffers is increased to at least about 7.2 to about 7.8. In some embodiments, the pH of the one or more wash buffers is increased to at least about 7.6. In some embodiments, the pH of the one or more wash buffers is increased by the addition of basic amino acids. In some embodiments, the one or more wash buffers exhibit a pH of at least 7. In some embodiments, the pH of the one or more elution buffers is increased to at least about 7.2 to about 7.8. In some embodiments, the pH of the one or more elution buffers is increased to at least about 7.6. In some embodiments, the pH of the one or more elution buffers is increased by the addition of basic amino acids. In some embodiments, the one or more elution buffers exhibit a pH of at least 7.

[00199] In some embodiments, the one or more buffers (including wash and/or elution buffers) comprise one or more chelating agents. In some embodiments, the elution buffer includes at least one chelating agent. The chelating agent can be a divalent cation chelating agent. In some embodiments, the at least one chelating agent is a divalent cation chelating agent. In some embodiments, the divalent cation chelating agent is selected from the group consisting of EDTA, EGTA, CDTA, and citrate. In some embodiments, the divalent cation chelating agent is selected from the group consisting of NTA, DTPA, EDDS, EDTA, EGTA, CDTA, and citrate. In some embodiments, the divalent cation chelating agent is selected from the group consisting of citrate, EDTA, DTPA, NTA, and EDDS. In some embodiments, the chelating agent is NTA. In some embodiments, the chelating agent is DTPA. In some embodiments, the chelating agent is EDDS. In some embodiments, the chelating agent is EDTA. In some embodiments, the chelating agent is EGTA. In some embodiments, the chelating agent is CDTA. In some embodiments, the chelating agent is citrate. In some embodiments, the one or more wash buffers in b) comprise said one or more chelating agents and exhibit a pH of at least 7.

[00200] Any of the buffers (buffer systems) described herein can be selected from the group consisting of glycine, HEPES (4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid), TrisHCl (Tris(hydroxymethyl)-aminomethane), histidine, imidazole, acetate citrate, citrate, acetate, MES, phosphate, TrisHCl, Bis-Tris, Histidine, Imidazol, ArgininHCl, LysinHCl, and 2-(N-morpholino)ethanesulfonic acid, as single buffers or as a combination of two or more buffers. In some embodiments, the one or more buffers are selected from the group consisting of glycine HEPES (4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid), TrisHCl (Tris(hydroxymethyl)-aminomethane), histidine, imidazole, acetate citrate, MES, and 2-(N-

morpholino)ethanesulfonic acid. In some embodiments, the buffer comprises citrate, acetate, MES, HEPES, Phosphate, TrisHCl, and/or Bis-Tris. In some embodiments, the buffer comprises glycine. In some embodiments, the buffer comprises HEPES (4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid). In some embodiments, the buffer comprises TrisHCl (Tris(hydroxymethyl)-aminomethane). In some embodiments, the buffer comprises histidine. In some embodiments, the buffer comprises imidazole. In some embodiments, the buffer comprises acetate citrate. In some embodiments, the buffer comprises citrate. In some embodiments, the buffer comprises acetate. In some embodiments, the buffer comprises MES. In some embodiments, the buffer comprises HEPES. In some embodiments, the buffer comprises phosphate. In some embodiments, the buffer comprises Tris-HCl. In some embodiments, the buffer comprises Bis-Tris. In some embodiments, the buffer comprises Histidine. In some embodiments, the buffer comprises Imidazole. In some embodiments, the buffer comprises Arginine HCl. In some embodiments, the buffer comprises Lysine HCl. In some embodiments, the buffer comprises 2-(N-morpholino)ethanesulfonic acid. In some embodiments, the buffer comprises one, two, three, or four of the buffers listed herein.

[00201] In some embodiments, the one or more buffers (including wash and/or elution buffers) comprise sodium citrate in a range including but not limited to, 10 mM-80 mM sodium citrate, 15 mM-80 mM sodium citrate, 10 mM-80 mM sodium citrate, 15 mM-60 mM sodium citrate, 20 mM-60 mM sodium citrate, 10 mM sodium citrate, 20 mM sodium citrate, 30 mM sodium citrate, 40 mM sodium citrate, 50 mM sodium citrate, 55 mM sodium citrate, 60 mM sodium citrate, 65 mM sodium citrate, 70 mM sodium citrate, 75 mM sodium citrate, 80 mM sodium citrate, or the like.

[00202] In some embodiments, a first elution buffer further comprises sodium citrate, in a range including but not limited to, 10 mM-60 mM sodium citrate, 15 mM-60 mM sodium citrate, 10 mM-50 mM sodium citrate, 15 mM-50 mM sodium citrate, 20 mM-60 mM sodium citrate, 10 mM sodium citrate, 20 mM sodium citrate, 30 mM sodium citrate, 40 mM sodium citrate, 50 mM sodium citrate, 60 mM sodium citrate, or the like.

[00203] In some embodiments, a second elution buffer further comprises sodium citrate, such as, but not limited to, 10 mM-60 mM sodium citrate, 15 mM-60 mM sodium citrate, 10 mM-50 mM sodium citrate, 15 mM-50 mM sodium citrate, 20 mM-60 mM sodium citrate, 10 mM sodium citrate, 20 mM sodium citrate, 30 mM sodium citrate, 40 mM sodium citrate, 50 mM sodium citrate, 60 mM sodium citrate, or the like.

[00204] In some embodiments, the one or more buffers (including wash and/or elution buffers) of the cation exchange chromatography step comprise EDTA, so long as the desired rVWF species remains bound to the cation exchange resin. In some embodiments, the one or more buffers (including wash and/or elution buffers) of the cation exchange chromatography step comprises about 0.5 mM to about 20 mM EDTA, *e.g.*, about 0.5 mM-about 20 mM, about 1 mM-about 20 mM, about 1.5 mM-about 20 mM, about 2 mM-about 20 mM, about 3 mM-about 20 mM, about 5 mM-about 20 mM, about 0.5 mM-about 15 mM, about 1 mM-about 10 mM, about 1 mM-about 5 mM, about 5 mM, about 0.5 mM, about 1 mM, about 2 mM, about 3 mM, about 4 mM, about 5 mM, about 6 mM, about 7 mM, about 8 mM, about 9 mM, about 10 mM, about 11 mM, about 12 mM, about 13 mM, about 14 mM, about 15 mM, about 16 mM, about 17 mM, about 18 mM, about 19 mM, about 20 mM, or the like, so long as the desired rVWF species remains bound to the cation exchange resin. In some embodiments, the buffers comprising EDTA are employed as part of a stepwise cation exchange elution. In some embodiments, the buffers comprising EDTA are employed as part of a gradient cation exchange elution. In some embodiments, when EDTA is employed as part of the buffers used in a stepwise cation exchange elution the counter-ion is Na^+ . In some embodiments, when EDTA is employed as part of the buffers used in a gradient cation exchange elution the counter-ion is Na^+ .

[00205] In some embodiments, the citrate can be found in the eluent after the rVWF-propeptide has been removed using a cation exchange method. In some embodiments, the citrate can be found in the eluent after the rVWF-propeptide has been removed using a stepwise cation exchange elution method. In some embodiments, the citrate can be found in the eluent after the rVWF-propeptide has been removed using a gradient cation exchange elution method. In some embodiments, the cation exchange counter-ion is Na^+ .

[00206] In some embodiments, the conductivity of the buffers (including wash and/or elution buffers), ranges from 5 mS/cm to 40 mS/cm, *e.g.*, 5 mS/cm–40 mS/cm, 10 mS/cm–40 mS/cm, 15 mS/cm–40 mS/cm, 20 mS/cm–40 mS/cm, 5 mS/cm–15 mS/cm, 10 mS/cm–25 mS/cm, 15 mS/cm–30 mS/cm, 20 mS/cm–30 mS/cm, or 30 mS/cm–40 mS/cm.

[00207] In some embodiments, the conductivity of at least one wash buffer is from about 5 mS/cm to about 40 mS/cm, *e.g.*, about 5 mS/cm–about 40 mS/cm, about 10 mS/cm–about 40 mS/cm, about 15 mS/cm–about 40 mS/cm, about 20 mS/cm–about 40 mS/cm, about 25 mS/cm–about 40 mS/cm, about 30 mS/cm–about 40 mS/cm, about 10 mS/cm–about 40

mS/cm, about 10 mS/cm–about 30 mS/cm, about 5 mS/cm–about 13 mS/cm, about 5 mS/cm–about 15 mS/cm, about 15 mS/cm–about 30 mS/cm, about 18 mS/cm–about 40 mS/cm, or about 20 mS/cm–about 40 mS/cm. In other embodiments, the conductivity of two or more wash buffers is from about 5 mS/cm to about 40 mS/cm, *e.g.*, about 5 mS/cm–about 40 mS/cm, about 10 mS/cm–about 40 mS/cm, about 15 mS/cm–about 40 mS/cm, about 20 mS/cm–about 40 mS/cm, about 25 mS/cm–about 40 mS/cm, about 30 mS/cm–about 40 mS/cm, about 10 mS/cm–about 40 mS/cm, about 10 mS/cm–about 30 mS/cm, about 5 mS/cm–about 13 mS/cm, about 5 mS/cm–about 15 mS/cm, about 15 mS/cm–about 30 mS/cm, about 18 mS/cm–about 40 mS/cm, or about 20 mS/cm–about 40 mS/cm.

[00208] In some embodiments, the conductivity of at least one elution buffer is from about 5 mS/cm to about 40 mS/cm, *e.g.*, about 5 mS/cm–about 40 mS/cm, about 10 mS/cm–about 40 mS/cm, about 15 mS/cm–about 40 mS/cm, about 20 mS/cm–about 40 mS/cm, about 25 mS/cm–about 40 mS/cm, about 30 mS/cm–about 40 mS/cm, about 10 mS/cm–about 40 mS/cm, about 10 mS/cm–about 30 mS/cm, about 5 mS/cm–about 13 mS/cm, about 5 mS/cm–about 15 mS/cm, about 15 mS/cm–about 30 mS/cm, about 18 mS/cm–about 40 mS/cm, or about 20 mS/cm–about 40 mS/cm. In other embodiments, the conductivity of two or more wash buffers is from about 5 mS/cm to about 40 mS/cm, *e.g.*, about 5 mS/cm–about 40 mS/cm, about 10 mS/cm–about 40 mS/cm, about 15 mS/cm–about 40 mS/cm, about 20 mS/cm–about 40 mS/cm, about 25 mS/cm–about 40 mS/cm, about 30 mS/cm–about 40 mS/cm, about 10 mS/cm–about 40 mS/cm, about 10 mS/cm–about 30 mS/cm, about 5 mS/cm–about 13 mS/cm, about 5 mS/cm–about 15 mS/cm, about 15 mS/cm–about 30 mS/cm, about 18 mS/cm–about 40 mS/cm, or about 20 mS/cm–about 40 mS/cm.

[00209] In some embodiments, the pH of the wash buffer is from pH 6.0 to pH 9.0, *e.g.*, pH 6.0–pH 9.0, pH 6.3–pH 9.0, pH 6.5–pH 9.0, pH 7.0–pH 9.0, pH 7.5–pH 9.0, pH 7.7.0–pH 9.0, pH 8.0–pH 9.0, pH 6.0–pH 8.5, pH 6.5–pH 8.5, pH 7.0–pH 8.5, pH 7.5–pH 8.5, pH 6.0–pH 8.0, pH 6.5–pH 8.0, pH 7.0–pH 8.0, pH 7.5–pH 8.0, pH 6.0, pH 6.1, pH 6.2, pH 6.3, pH 6.4, pH 6.5, pH 6.6, pH 6.7, pH 6.8, pH 6.9, pH 7.0, pH 7.1, pH 7.2, pH 7.3, pH 7.4, pH 7.5, pH 7.6, pH 7.7, pH 7.8, pH 7.9, pH 8.0, pH 8.1, pH 8.2, pH 8.3, pH 8.4, pH 8.5, pH 8.6, pH 8.7, pH 8.8, pH 8.9, pH 9.0.

[00210] In one aspect, the method described herein includes a gradient elution step. The gradient elution step can remove product impurities and process-related impurities to

optimize yield of mature VWF. In some cases, the gradient elution step separates a higher percentage of VWF pro-peptide from mature VWF compared to a prior art method.

[00211] In some embodiments, the conductivity of the one or more elution buffers is from about 5 mS/cm to about 40 mS/cm, *e.g.*, about 5 mS/cm–about 40 mS/cm, about 10 mS/cm–about 40 mS/cm, about 15 mS/cm–about 40 mS/cm, about 20 mS/cm–about 40 mS/cm, about 25 mS/cm–about 40 mS/cm, about 30 mS/cm–about 40 mS/cm, about 10 mS/cm–about 40 mS/cm, about 10 mS/cm–about 30 mS/cm, about 5 mS/cm–about 13 mS/cm, about 5 mS/cm–about 15 mS/cm, about 15 mS/cm–about 30 mS/cm, about 18 mS/cm–about 40 mS/cm, or about 20 mS/cm–about 40 mS/cm. In other embodiments, the conductivity of two or more wash buffers is from about 5 mS/cm to about 40 mS/cm, *e.g.*, about 5 mS/cm–about 40 mS/cm, about 10 mS/cm–about 40 mS/cm, about 15 mS/cm–about 40 mS/cm, about 20 mS/cm–about 40 mS/cm, about 25 mS/cm–about 40 mS/cm, about 30 mS/cm–about 40 mS/cm, about 10 mS/cm–about 40 mS/cm, about 10 mS/cm–about 30 mS/cm, about 5 mS/cm–about 13 mS/cm, about 5 mS/cm–about 15 mS/cm, about 15 mS/cm–about 30 mS/cm, about 18 mS/cm–about 40 mS/cm, or about 20 mS/cm–about 40 mS/cm.

[00212] In some embodiments, the flow rate of one or more wash steps of the present method is about 10 cm/h to about 200 cm/h, *e.g.*, about 10 cm/h, about 15 cm/h, about 20 cm/h, about 25 cm/h, about 30 cm/h, about 35 cm/h, about 40 cm/h, about 45 cm/h, about 50 cm/h, about 55 cm/h, about 60 cm/h, about 65 cm/h, about 70 cm/h, about 75 cm/h, about 80 cm/h, about 85 cm/h, about 90 cm/h, about 95 cm/h, about 100 cm/h, about 105 cm/h, about 110 cm/h, about 115 cm/h, about 120 cm/h, about 125 cm/h, about 130 cm/h, about 135 cm/h, about 140 cm/h, about 145 cm/h, about 150 cm/h, about 155 cm/h, about 160 cm/h, about 165 cm/h, about 170 cm/h, about 175 cm/h, about 180 cm/h, about 185 cm/h, about 190 cm/h, about 195 cm/h, or about 200 cm/h. Depending on the resin, in some embodiments the flow rate can be up to 600 cm/h.

[00213] In some embodiments, the flow rate of one or more elution steps of the present method is about 10 cm/h to about 200 cm/h, *e.g.*, about 10 cm/h, about 15 cm/h, about 20 cm/h, about 25 cm/h, about 30 cm/h, about 35 cm/h, about 40 cm/h, about 45 cm/h, about 50 cm/h, about 55 cm/h, about 60 cm/h, about 65 cm/h, about 70 cm/h, about 75 cm/h, about 80 cm/h, about 85 cm/h, about 90 cm/h, about 95 cm/h, about 100 cm/h, about 105 cm/h, about 110 cm/h, about 115 cm/h, about 120 cm/h, about 125 cm/h, about 130 cm/h, about 135 cm/h, about 140 cm/h, about 145 cm/h, about 150 cm/h, about 155 cm/h, about 160 cm/h, about 165

cm/h, about 170 cm/h, about 175 cm/h, about 180 cm/h, about 185 cm/h, about 190 cm/h, about 195 cm/h, or about 200 cm/h. Depending on the resin, in some embodiments the flow rate can be up to 600 cm/h.

[00214] In some embodiments, the one or more buffers further comprise one or more nonionic detergents. In some embodiments, the nonionic detergent is selected from the group consisting of Triton X-100, Tween 80, and Tween 20. In some embodiments, the nonionic detergent is Triton X-100. In some embodiments, the nonionic detergent is Tween 80. In some embodiments, the nonionic detergent is Tween 20.

[00215] In some embodiments, the said one or more buffers further comprise one or more additional substances selected from the group consisting of non-reducing sugars, sugar alcohols, and polyols. In some embodiments, the one or more buffers further comprises one or more non-reducing sugars. In some embodiments, the non-reducing sugar includes but is not limited to sucrose, trehalose, mannitol, sorbitol, galactitol, and/or xylitol. In some embodiments, the one or more buffers further comprises one or more sugar alcohols. In some embodiments, the one or more buffers further comprises one or more polyols. In some embodiments, the sugar alcohol or polyol includes but is not limited to mannitol, xylitol, erythritol, threitol, sorbitol, and/or glycerol. In some embodiments, the buffers further comprise sorbitol, mannitol, xylitol, sucrose, trehalose, ethylene glycol, propylene glycol, glycerol, 1,2,3-Propanetriol, meso-erythritol, and/or erythritol (meso-1,2,3,4-Butantetrol).

[00216] The pH of any of the buffers can be adjusted (increased) by adding an amino acid, Tris, NaOH, ethanolamine, and the like.

[00217] Any of the buffers (buffer systems) described herein can be selected from the group consisting of Citrate, Acetate, MES, HEPES, Phosphate, TrisHCl, Bis-Tris, as single buffers or as a combination of two or more buffers. In some embodiments, the buffer comprises glycine. In some embodiments, the buffer comprises Citrate. In some embodiments, the buffer comprises Acetate. In some embodiments, the buffer comprises MES. In some embodiments, the buffer comprises HEPES. In some embodiments, the buffer comprises phosphate. In some embodiments, the buffer comprises TrisHCl. In some embodiments, the buffer comprises Bis-Tris. In some embodiments, the buffer comprises one, two, three, or four of the buffers listed herein.

[00218] In some embodiments, the cation exchange method buffer chelator combination comprises citrate, malate (malic acid), and tartrate (tartaric acid).

c. SIZE EXCLUSION CHROMATOGRAPHY PURIFICATION

[00219] In one aspect of the present invention, mature VWF and VWF-PP are separated by way of size exclusion chromatography (SEC). In some cases, remaining host cell derived impurities such as CHO host cell proteins, process related impurities such as recombinant furin and low molecular weight viral inactivation reagents, media compounds such as soy peptone, and other product related impurities are removed from the mature VWF.

[00220] In another aspect of the present method, mature VWF is separated from VWF-PP such as residual VWF-PP or free VWF-PP using size exclusion chromatography. For separation, the starting or loading composition can comprise a low pH and at least one chelating agent. In other embodiments, the gradient elution buffer has a neutral to high pH, such as a pH ranging from pH 6.0 to pH 9.0. In another embodiment, the gradient elution buffer comprises one or more chelating agents and has a pH of 7.0 or higher, *e.g.*, pH 7.0 to pH 9.0. For instance, the gradient elution buffer can include EDTA and have a pH of 8.5.

[00221] In some embodiments, the present invention provides a method for obtaining a composition comprising a high purity, propeptide depleted mature recombinant rVWF (high purity mat-rVWF), said method comprising the steps of: (a) loading a solution comprising pro-rVWF, mat-rVWF/ rVWF-PP complex, mat-rVWF, and/or rVWF propeptide (rVWF-PP) onto an size exclusion column, wherein said pro-rVWF, mat-rVWF/ rVWF-PP complex, and mat-rVWF are bound to said size exclusion column; (b) washing said size exclusion column in a) containing said bound pro-rVWF, mat-rVWF/ rVWF-PP complex, and mat-rVWF with one or more wash buffers; (c) treating said column in b) comprising the bound pro-rVWF, mat-rVWF/ rVWF-PP complex, and mat-rVWF with furin, wherein said furin cleaves said pro-rVWF into mat-rVWF and rVWF-PP; (d) eluting said bound pro-rVWF, mat-rVWF/ rVWF-PP complex, and mat-rVWF from the column in c) with an elution buffer, wherein said elution buffer induces dissociation of said rVWF-PP from mat-rVWF non-covalently associated with said rVWF-PP, and wherein said dissociation is induced by: (i) addition of at least one chelating agent into said elution buffer, or (ii) increasing the pH of said elution buffer to a pH of at least 7; and (e) collecting said mat-rVWF separately from said rVWF-PP

to obtain a high purity mat-rVWF composition, wherein said high purity mat-rVWF composition comprises at least 95% mature rVWF and less than 5% rVWF-PP.

[00222] In some embodiments, a) and b) occur simultaneously in a single step. In some embodiments, the solution in a) comprises the flow through from an immunoaffinity purification method. In some embodiments, the solution in a) comprises the flow through from a monoclonal antibody column, wherein said monoclonal antibody is a FVIII monoclonal antibody. In some embodiments, the solution in a) is selected from the group consisting of a cell culture medium, an antibody column flow-through solution, and a buffered solution.

[00223] In some embodiments, the separation buffer has a neutral to high pH. In other embodiments, the buffer comprises at least one chelating agent. In some embodiments, the buffer comprises at least one chelating agent and has a neutral to high pH. For example, the separation buffer can contain a chelating agent and have a pH of 6.0 or higher, or in some cases, a pH of 7.0 or higher.

[00224] In some embodiments, the loading concentration of pro-VWF is from about 90 IU/ml to about 270 IU/ml resin, *e.g.*, about 90 IU/ ml-about 270 IU/ml, about 100 IU/ ml-about 270 IU/ml, about 110 IU/ ml-about 270 IU/ml, about 120 IU/ ml-about 270 IU/ml, about 130 IU/ ml-about 270 IU/ml, about 130 IU/ ml-about 270 IU/ml, about 140 IU/ ml-about 270 IU/ml, about 150 IU/ ml-about 270 IU/ml, about 90 IU/ ml-about 250 IU/ml, about 100 IU/ ml-about 250 IU/ml, about 110 IU/ ml-about 250 IU/ml, about 120 IU/ ml-about 250 IU/ml, about 130 IU/ ml-about 250 IU/ml, about 130 IU/ ml-about 250 IU/ml, about 140 IU/ ml-about 250 IU/ml, about 150 IU/ ml-about 250 IU/ml, about 90 IU/ ml-about 200 IU/ml, about 100 IU/ ml-about 200 IU/ml, about 110 IU/ ml-about 200 IU/ml, about 120 IU/ ml-about 200 IU/ml, about 130 IU/ ml-about 200 IU/ml, about 130 IU/ ml-about 200 IU/ml, about 140 IU/ ml-about 200 IU/ml, about 150 IU/ ml-about 200 IU/ml, about 90 IU/ ml-about 100 IU/ml, about 100 IU/ ml-about 150 IU/ml, about 150 IU/ ml-about 200 IU/ml, about 200 IU/ ml-about 250 IU/ml, or about 250 IU/ ml-about 270 IU/ml resin.

[00225] In some embodiments, the pH of the starting composition, loading solution, or loading composition comprises pro-rVWF, mat-rVWF/ rVWF-PP complex, mat-rVWF, and/or rVWF propeptide (rVWF-PP) is from pH 6.0 to pH 9.0, *e.g.*, pH 6.0–pH 9.0, pH 6.3–pH 9.0, pH 6.5–pH 9.0, pH 7.0–pH 9.0, pH 7.5–pH 9.0, pH 7.7.0–pH 9.0, pH 8.0–pH 9.0, pH 6.0–pH 8.5, pH 6.5–pH 8.5, pH 7.0–pH 8.5, pH 7.5–pH 8.5, pH 6.0–pH 8.0, pH 6.5–pH 8.0,

pH 7.0–pH 8.0, pH 7.5–pH 8.0, pH 6.0, pH 6.1, pH 6.2, pH 6.3, pH 6.4, pH 6.5, pH 6.6, pH 6.7, pH 6.8, pH 6.9, pH 7.0, pH 7.1, pH 7.2, pH 7.3, pH 7.4, pH 7.5, pH 7.6, pH 7.7, pH 7.8, pH 7.9, pH 8.0, pH 8.1, pH 8.2, pH 8.3, pH 8.4, pH 8.5, pH 8.6, pH 8.7, pH 8.8, pH 8.9, or pH 9.0.

[00226] In some embodiments, the size exclusion method comprises a buffer system. In some embodiments, the buffer system comprises one or more separation buffers. In some embodiments, the buffer system comprises at least one separation buffer. In some embodiments, the buffer system comprises at least two separation buffers. In some embodiments the buffer system comprises at least a first separation buffer and at least a second separation buffer.

[00227] In some embodiments, the first separation buffer comprises at least one chelating agent, and optionally has a pH ranging from pH 6.0 to pH 9.0. In some embodiments, the separation wash buffer has a pH ranging from pH 6.0 to pH 9.0, and optionally comprises at least one chelating agent. In some embodiments, the first separation buffer has a pH ranging from pH 6.0 to pH 6.9. In some embodiments, the second separation buffer has a pH ranging from pH 7.0 to pH 9.0. In some embodiments, the first separation buffer can comprise at least one chelating agent and has a pH ranging from pH 6.0 to pH 6.9. In some embodiments, the first separation buffer has a pH of less than 7. In some embodiments, the second separation buffer has a pH of greater than 7. In some embodiments, when two separation buffers are employed, the first separation buffer has a pH of less than 7 and the second separation buffer has a pH of greater than 7.

[00228] In some embodiments, the starting solution comprising mature rVWF and rVWF-PP is contacted with a separation buffer comprising at least one chelating agent, and optionally the buffer has a pH of ranging from pH 6.0 to pH 9.0. In some embodiments, the starting solution is contacted with a buffer having a pH ranging from pH 6.0 to pH 9.0, and optionally the buffer comprises at least one chelating agent. In some embodiments, the buffer has a pH ranging from pH 7.0 to pH 9.0. In some embodiments the buffer is a first separation buffer with a pH of 6.0 to 6.9. In some embodiments, the buffer is a second separation buffer with a pH of 7.0 to 9.0. In some embodiments, the starting solution comprising mature rVWF and rVWF-PP is contacted first with a first buffer having a pH from 6.0 to 6.9 and a second separation buffer having a pH from 7.0 to 9.0.

[00229] In some embodiments, the pH of the one or more separation buffers is from pH 6.0 to pH 9.0, *e.g.*, pH 6.0–pH 9.0, pH 6.3–pH 9.0, pH 6.5–pH 9.0, pH 7.0–pH 9.0, pH 7.5–pH 9.0, pH 7.7.0–pH 9.0, pH 8.0–pH 9.0, pH 6.0–pH 8.5, pH 6.5–pH 8.5, pH 7.0–pH 8.5, pH 7.5–pH 8.5, pH 6.0–pH 8.0, pH 6.5–pH 8.0, pH 7.0–pH 8.0, pH 7.5–pH 8.0, pH 6.0, pH 6.1, pH 6.2, pH 6.3, pH 6.4, pH 6.5, pH 6.6, pH 6.7, pH 6.8, pH 6.9, pH 7.0, pH 7.1, pH 7.2, pH 7.3, pH 7.4, pH 7.5, pH 7.6, pH 7.7, pH 7.8, pH 7.9, pH 8.0, pH 8.1, pH 8.2, pH 8.3, pH 8.4, pH 8.5, pH 8.6, pH 8.7, pH 8.8, pH 8.9, or pH 9.0.

[00230] In some embodiments, the pH of the elution buffer is increased as compared to the starting solution in step a), is increased as compared to a first separation buffer when two separation buffers are employed, and/or is increased as compared to a first separation buffer when a second separation buffer is employed. In some embodiments, when a first separation buffer and as second separation buffer are employed, the first separation buffer has a pH of less than 7 and the second separation buffer has a pH of greater than 7.

[00231] In some embodiments, the pH of the one or more separation buffers is increased to at least 7.1, 7.2, 7.3, 7.4, 7.5, 7.6, 7.7, 7.8, 7.9, or 8.0, as compared to the loading solution comprising pro-rVWF, mat-rVWF/ rVWF-PP complex, mat-rVWF, and/or rVWF propeptide (rVWF-PP), as recited in step (a) of the method. In some embodiments, the pH of the buffer is increased to at least 7.1, 7.2, 7.3, 7.4, 7.5, 7.6, 7.7, 7.8, 7.9, or 8.0 in order to induce dissociation of the mat-rVWF/ rVWF-PP complex in the solution in step (a) of the method into mat-rVWF and rVWF-PP, wherein said dissociation occurs by disruption of the non-covalently associated mat-rVWF and rVWF-PP. In some embodiments, the pH of the loading solution is increased to at least about 7.2 to about 7.8. In some embodiments, the pH of the loading solution is increased to at least about 7.6. In some embodiments, the pH of the loading solution is increased by the addition of basic amino acids. In some embodiments, the pH of at the loading solution is increased to at least 7. In some embodiments, the pH of the one or more wash buffers is increased to at least about 7.2 to about 7.8. In some embodiments, the pH of the one or more wash buffers is increased to at least about 7.6. In some embodiments, the pH of the one or more separation buffers is increased by the addition of basic amino acids. In some embodiments, the one or more separation buffers exhibit a pH of at least 7.

[00232] In some embodiments, the one or more separation buffers comprise one or more chelating agents. In some embodiments, the elution buffer includes at least one

chelating agent. The chelating agent can be a divalent cation chelating agent. In some embodiments, the at least one chelating agent is a divalent cation chelating agent. In some embodiments, the divalent cation chelating agent is selected from the group consisting of EDTA, EGTA, CDTA, and citrate. In some embodiments, the divalent cation chelating agent is selected from the group consisting of NTA, DTPA, EDDS, EDTA, EGTA, CDTA, and citrate. In some embodiments, the chelating agent is NTA. In some embodiments, the chelating agent is DTPA. In some embodiments, the chelating agent is EDDS. In some embodiments, the chelating agent is EDTA. In some embodiments, the chelating agent is EGTA. In some embodiments, the chelating agent is CDTA. In some embodiments, the chelating agent is citrate. In some embodiments, the one or more wash buffers in b) comprise said one or more chelating agents and exhibit a pH of at least 7.

[00233] In some embodiments, the one or more separation buffers include at least one chelating agent. The chelating agent can be a divalent cation chelating agent. In some embodiments, the divalent cation chelating agent is selected from the group consisting of nitrilo-2,2',2''-triacetic acid (NTA), Diethylenetriaminepentaacetic acid; Diethylenetriamine-N,N,N',N',N''-pentaacetic acid (DTPA), Ethylenediamine-N,N'-disuccinic acid (EDDS), Ethylenediaminetetraacetic acid (EDTA), EGTA, CDTA, and citrate. In some embodiments, the divalent cation chelating agent is selected from the group consisting of NTA, DTPA, EDDS, EDTA, and citrate. In some embodiments, the chelating agent is NTA. In some embodiments, the chelating agent is DTPA. In some embodiments, the chelating agent is EDDS. In some embodiments, the chelating agent is EDTA. In some embodiments, the chelating agent is EGTA. In some embodiments, the chelating agent is CDTA. In some embodiments, the chelating agent is citrate.

[00234] In some embodiments, the elution buffer A and/or elution buffer B of the anion exchange chromatography step comprises about 0.5 mM to about 20 mM EDTA, *e.g.*, about 0.5 mM-about 20 mM, about 1 mM-about 20 mM, about 1.5 mM-about 20 mM, about 2 mM-about 20 mM, about 3 mM-about 20 mM, about 5 mM-about 20 mM, about 0.5 mM-about 15 mM, about 1 mM-about 10 mM, about 1 mM-about 5 mM, about 5 mM, about 0.5 mM, about 1 mM, about 2 mM, about 3 mM, about 4 mM, about 5 mM, about 6 mM, about 7 mM, about 8 mM, about 9 mM, about 10 mM, about 11 mM, about 12 mM, about 13 mM, about 14 mM, about 15 mM, about 16 mM, about 17 mM, about 18 mM, about 19 mM, about 20 mM, or the like.

[00235] In some embodiments, the one or more separation buffers comprise sodium citrate in a range including but not limited to, 10 mM-500 mM sodium citrate, 15 mM-400 mM sodium citrate, 10 mM-400 mM sodium citrate, 15 mM-350 mM sodium citrate, 20 mM-350 mM sodium citrate, 10 mM sodium citrate, 20 mM sodium citrate, 30 mM sodium citrate, 40 mM sodium citrate, 50 mM sodium citrate, 55 mM sodium citrate, 60 mM sodium citrate, 65 mM sodium citrate, 70 mM sodium citrate, 75 mM sodium citrate, 80 mM sodium citrate, 90 mM sodium citrate, 100 mM sodium citrate, 110 mM sodium citrate, 120 mM sodium citrate, 130 mM sodium citrate, 140 mM sodium citrate, 150 mM sodium citrate, 160 mM sodium citrate, 170 mM sodium citrate, 180 mM sodium citrate, 190 mM sodium citrate, 200 mM sodium citrate, 210 mM sodium citrate, 220 mM sodium citrate, 230 mM sodium citrate, 240 mM sodium citrate, 250 mM sodium citrate, 260 mM sodium citrate, 270 mM sodium citrate, 280 mM sodium citrate, 290 mM sodium citrate, 300 mM sodium citrate, 310 mM sodium citrate, 320 mM sodium citrate, 330 mM sodium citrate, 340 mM sodium citrate, 350 mM sodium citrate, 360 mM sodium citrate, 370 mM sodium citrate, 380 mM sodium citrate, 390 mM sodium citrate, 400 mM sodium citrate, 410 mM sodium citrate, 420 mM sodium citrate, 430 mM sodium citrate, 440 mM sodium citrate, 450 mM sodium citrate, 460 mM sodium citrate, 470 mM sodium citrate, 480 mM sodium citrate, 490 mM sodium citrate, 500 mM sodium citrate, 510 mM sodium citrate, 520 mM sodium citrate, 530 mM sodium citrate, 540 mM sodium citrate, 550 mM sodium citrate, 560 mM sodium citrate, 570 mM sodium citrate, 580 mM sodium citrate, 590 mM sodium citrate, or 600 mM sodium citrate, or the like.

[00236] In some embodiments, the one or more separation buffers further comprises sodium citrate, in a range including but not limited to, 10 mM-500 mM sodium citrate, 15 mM-400 mM sodium citrate, 10 mM-400 mM sodium citrate, 15 mM-350 mM sodium citrate, 20 mM-350 mM sodium citrate, 10 mM sodium citrate, 20 mM sodium citrate, 30 mM sodium citrate, 40 mM sodium citrate, 50 mM sodium citrate, 55 mM sodium citrate, 60 mM sodium citrate, 65 mM sodium citrate, 70 mM sodium citrate, 75 mM sodium citrate, 80 mM sodium citrate, 90 mM sodium citrate, 100 mM sodium citrate, 110 mM sodium citrate, 120 mM sodium citrate, 130 mM sodium citrate, 140 mM sodium citrate, 150 mM sodium citrate, 160 mM sodium citrate, 170 mM sodium citrate, 180 mM sodium citrate, 190 mM sodium citrate, 200 mM sodium citrate, 210 mM sodium citrate, 220 mM sodium citrate, 230 mM sodium citrate, 240 mM sodium citrate, 250 mM sodium citrate, 260 mM sodium citrate,

270 mM sodium citrate, 280 mM sodium citrate, 290 mM sodium citrate, 300 mM sodium citrate, 310 mM sodium citrate, 320 mM sodium citrate, 330 mM sodium citrate, 340 mM sodium citrate, 350 mM sodium citrate, 360 mM sodium citrate, 370 mM sodium citrate, 380 mM sodium citrate, 390 mM sodium citrate, 400 mM sodium citrate, 410 mM sodium citrate, 420 mM sodium citrate, 430 mM sodium citrate, 440 mM sodium citrate, 450 mM sodium citrate, 460 mM sodium citrate, 470 mM sodium citrate, 480 mM sodium citrate, 490 mM sodium citrate, 500 mM sodium citrate, 510 mM sodium citrate, 520 mM sodium citrate, 530 mM sodium citrate, 540 mM sodium citrate, 550 mM sodium citrate, 560 mM sodium citrate, 570 mM sodium citrate, 580 mM sodium citrate, 590 mM sodium citrate, or 600 mM sodium citrate, or the like.

[00237] In some embodiments, the one or more separation buffers further comprises sodium citrate, such as, but not limited to, 10 mM-500 mM sodium citrate, 15 mM-400 mM sodium citrate, 10 mM-400 mM sodium citrate, 15 mM-350 mM sodium citrate, 20 mM-350 mM sodium citrate, 10 mM sodium citrate, 20 mM sodium citrate, 30 mM sodium citrate, 40 mM sodium citrate, 50 mM sodium citrate, 55 mM sodium citrate, 60 mM sodium citrate, 65 mM sodium citrate, 70 mM sodium citrate, 75 mM sodium citrate, 80 mM sodium citrate, 90 mM sodium citrate, 100 mM sodium citrate, 110 mM sodium citrate, 120 mM sodium citrate, 130 mM sodium citrate, 140 mM sodium citrate, 150 mM sodium citrate, 160 mM sodium citrate, 170 mM sodium citrate, 180 mM sodium citrate, 190 mM sodium citrate, 200 mM sodium citrate, 210 mM sodium citrate, 220 mM sodium citrate, 230 mM sodium citrate, 240 mM sodium citrate, 250 mM sodium citrate, 260 mM sodium citrate, 270 mM sodium citrate, 280 mM sodium citrate, 290 mM sodium citrate, 300 mM sodium citrate, 310 mM sodium citrate, 320 mM sodium citrate, 330 mM sodium citrate, 340 mM sodium citrate, 350 mM sodium citrate, 360 mM sodium citrate, 370 mM sodium citrate, 380 mM sodium citrate, 390 mM sodium citrate, 400 mM sodium citrate, 410 mM sodium citrate, 420 mM sodium citrate, 430 mM sodium citrate, 440 mM sodium citrate, 450 mM sodium citrate, 460 mM sodium citrate, 470 mM sodium citrate, 480 mM sodium citrate, 490 mM sodium citrate, 500 mM sodium citrate, 510 mM sodium citrate, 520 mM sodium citrate, 530 mM sodium citrate, 540 mM sodium citrate, 550 mM sodium citrate, 560 mM sodium citrate, 570 mM sodium citrate, 580 mM sodium citrate, 590 mM sodium citrate, or 600 mM sodium citrate, or the like.

[00238] In some embodiments, the conductivity of the separation buffer is from about 5 mS/cm to about 40 mS/cm, *e.g.*, about 5 mS/cm–about 40 mS/cm, about 10 mS/cm–about

40 mS/cm, about 15 mS/cm–about 40 mS/cm, about 20 mS/cm–about 40 mS/cm, about 25 mS/cm–about 40 mS/cm, about 30 mS/cm–about 40 mS/cm, about 10 mS/cm–about 40 mS/cm, about 10 mS/cm–about 30 mS/cm, about 5 mS/cm–about 13 mS/cm, about 5 mS/cm–about 15 mS/cm, about 15 mS/cm–about 30 mS/cm, about 18 mS/cm–about 40 mS/cm, or about 20 mS/cm–about 40 mS/cm.

[00239] Any of the buffers (buffer systems) described herein can be selected from the group consisting of Citrate, Acetate, MES, HEPES, phosphate, TrisHCl, Bis-Tris, Histidine, Imidazole, Arginine HCl, Lysine HCl, Glycine, Glycylglycine, borate, MOPS, bicine, tricine, TAPS, TAPSO, and PIPES, as single buffers or as a combination of two or more buffers. In some embodiments, the buffer comprises glycine. In some embodiments, the buffer comprises Citrate, Acetate, MES, HEPES, Phosphate, TrisHCl, Bis-Tris, Histidine, Imidazol, ArgininHCl, LysinHCl, Glycine, Glycylglycine, borate, MOPS, bicine, tricine, TAPS, TAPSO, and/or PIPES. In some embodiments, the buffer comprises Citrate. In some embodiments, the buffer comprises Acetate. In some embodiments, the buffer comprises MES. In some embodiments, the buffer comprises HEPES. In some embodiments, the buffer comprises phosphate. In some embodiments, the buffer comprises Tris-HCl. In some embodiments, the buffer comprises Bis-Tris.

[00240] In some embodiments, the buffer comprises Histidine. In some embodiments, the buffer comprises Imidazole. In some embodiments, the buffer comprises Arginine HCl. In some embodiments, the buffer comprises Lysine HCl. In some embodiments, the buffer comprises Glycine. In some embodiments, the buffer comprises Glycylglycine. In some embodiments, the buffer comprises borate. In some embodiments, the buffer comprises MOPS. In some embodiments, the buffer comprises bicine. In some embodiments, the buffer comprises tricine. In some embodiments, the buffer comprises TAPS. In some embodiments, the buffer comprises TAPSO. In some embodiments, the buffer comprises and PIPES. In some embodiments, the buffer comprises one, two, three, or four of the buffers listed herein.

[00241] In some embodiments, the one or more separation buffers further comprise one or more nonionic detergents. In some embodiments, the nonionic detergent is selected from the group consisting of Triton X-100, Tween 80, and Tween 20. In some embodiments, the nonionic detergent is Triton X-100. In some embodiments, the nonionic detergent is Tween 80. In some embodiments, the nonionic detergent is Tween 20.

[00242] In some embodiments, the flow rate use during the one or more separation buffer steps of the present method is about 10 cm/h to about 200 cm/h, *e.g.*, about 10 cm/h, about 15 cm/h, about 20 cm/h, about 25 cm/h, about 30 cm/h, about 35 cm/h, about 40 cm/h, about 45 cm/h, about 50 cm/h, about 55 cm/h, about 60 cm/h, about 65 cm/h, about 70 cm/h, about 75 cm/h, about 80 cm/h, about 85 cm/h, about 90 cm/h, about 95 cm/h, about 100 cm/h, about 105 cm/h, about 110 cm/h, about 115 cm/h, about 120 cm/h, about 125 cm/h, about 130 cm/h, about 135 cm/h, about 140 cm/h, about 145 cm/h, about 150 cm/h, about 155 cm/h, about 160 cm/h, about 165 cm/h, about 170 cm/h, about 175 cm/h, about 180 cm/h, about 185 cm/h, about 190 cm/h, about 195 cm/h, or about 200 cm/h. Depending on the resin, in some embodiments the flow rate can be up to 600 cm/h.

[00243] In some embodiments, the flow rate use during the one or more separation buffer steps of the present method is about 10 cm/h to about 200 cm/h, *e.g.*, about 10 cm/h, about 15 cm/h, about 20 cm/h, about 25 cm/h, about 30 cm/h, about 35 cm/h, about 40 cm/h, about 45 cm/h, about 50 cm/h, about 55 cm/h, about 60 cm/h, about 65 cm/h, about 70 cm/h, about 75 cm/h, about 80 cm/h, about 85 cm/h, about 90 cm/h, about 95 cm/h, about 100 cm/h, about 105 cm/h, about 110 cm/h, about 115 cm/h, about 120 cm/h, about 125 cm/h, about 130 cm/h, about 135 cm/h, about 140 cm/h, about 145 cm/h, about 150 cm/h, about 155 cm/h, about 160 cm/h, about 165 cm/h, about 170 cm/h, about 175 cm/h, about 180 cm/h, about 185 cm/h, about 190 cm/h, about 195 cm/h, or about 200 cm/h. Depending on the resin, in some embodiments the flow rate can be up to 600 cm/h.

[00244] In some embodiments, the said one or more buffers further comprise one or more additional substances selected from the group consisting of non-reducing sugars, sugar alcohols, and polyols. In some embodiments, the one or more buffers further comprises one or more non-reducing sugars. In some embodiments, the non-reducing sugar includes but is not limited to sucrose, trehalose, mannitol, sorbitol, galactitol, and/or xylitol. In some embodiments, the one or more buffers further comprises one or more sugar alcohols. In some embodiments, the one or more buffers further comprises one or more polyols. In some embodiments, the sugar alcohol or polyol includes but is not limited to mannitol, xylitol, erythritol, threitol, sorbitol, and/or glycerol. In some embodiments, the buffers further comprise sorbitol, mannitol, xylitol, sucrose, trehalose, ethylene glycol, propylene glycol, glycerol, 1,2,3-Propanetriol, meso-erythritol, and/or erythritol (meso-1,2,3,4-Butantetrol).

[00245] In some embodiments, the size exclusion chromatography method buffer chelator combination comprises citrate, malate (malic acid), and tartrate (tartaric acid).

D. IMMUNOAFFINITY PURIFICATION

[00246] In some embodiments, the solution comprising pro-rVWF, mat-rVWF/ rVWF-PP complex, mat-rVWF, and/or rVWF propeptide (rVWF-PP) is obtained from an immunoaffinity purification method, including for example, a monoclonal antibody column. In some embodiments, the monoclonal antibody column comprises a FVIII monoclonal antibody. In some embodiments, the monoclonal antibody column comprises a VWF monoclonal antibody. Such columns and methods are known in the art and have been described. See, for example, Zimmerman *et al.* (U.S. Pat. No. 4,361,509; incorporated by reference herein for all purposes) which describes a method of purifying factor VIII, wherein factor VIII/VWF complex is bound to a monoclonal anti-VWF antibody, and factor VIII is dissociated from the complex by means of CaCl_2 ions. The immunoaffinity carrier to which vWF is still adsorbed is regenerated by means of a chaotropic agent, in particular NaSCN, a vWF/NaSCN solution being incurred as a by-product and being discarded.

[00247] Other methods include those described in U.S. Patent No. 6,579,723, also incorporated by reference herein in its entirety, which describes a method for recovering highly purified vWF or factor VIII/vWF-complex, using an immunoaffinity chromatography procedure. Such method employs recovery of VWF from an immunoaffinity adsorbent by using an eluting agent containing a zwitterionic species. The presence of the zwitterionic species allows for the use of mild conditions throughout the preparation, facilitating retention of molecular integrity, activity, and incorporation of the recovered proteins into pharmaceutical preparations without the need for additional stabilizers or preservatives.

Any such methods can be employed with the current purification method in order to obtain the solution comprising pro-rVWF, mat-rVWF/ rVWF-PP complex, mat-rVWF, and/or rVWF propeptide (rVWF-PP). IN some embodiments, the immunoaffinity purification optionally occurs prior to step (a) in any of the described purification procedures described herein, including those based on cation exchanged, anion exchange, and/or size exclusion chromatography procedures.

E. FREE MATURE VWF

[00248] In some embodiments, the host cell (HC) impurity level of the composition provided herein is equal to or less than 2.0 ppm, *e.g.*, 2.0 ppm, 1.9 ppm, 1.8 ppm, 1.7 ppm, 1.6 ppm, 1.5 ppm, 1.4 ppm, .3 ppm, 1.2 ppm, 1.1 ppm, 1.0 ppm, 0.9 ppm, 0.8 ppm, 0.7 ppm, 0.6 ppm, 0.5 ppm, 0.4 ppm, 0.3 ppm, 0.2 ppm, 0.1 ppm, 0.09 ppm, 0.08 ppm, 0.07 ppm, 0.06 ppm, 0.05 ppm, 0.04 ppm, 0.03 ppm, 0.02 ppm, 0.01 ppm or less. In other embodiments, the host cell impurity level of the composition provided herein is equal to or less than 0.6 ppm, *e.g.*, 0.6 ppm, 0.5 ppm, 0.4 ppm, 0.3 ppm, 0.2 ppm, 0.1 ppm, 0.09 ppm, 0.08 ppm, 0.07 ppm, 0.06 ppm, 0.05 ppm, 0.04 ppm, 0.03 ppm, 0.02 ppm, 0.01 ppm, or less.

[00249] In some embodiments, the host cell (HC) impurity level of the composition provided herein is equal to or less than 5.0% (*e.g.*, $\leq 5.0\%$). In some embodiments, the host cell (HC) impurity level of the composition provided herein is equal to or less than 4.0% (*e.g.*, $\leq 4.0\%$). In some embodiments, the host cell (HC) impurity level of the composition provided herein is equal to or less than 3.0% (*e.g.*, $\leq 3.0\%$). In some embodiments, the host cell (HC) impurity level of the composition provided herein is equal to or less than 2.0% (*e.g.*, $\leq 1.0\%$). In some embodiments, the host cell (HC) impurity level of the composition provided herein is equal to or less than 2.0% (*e.g.*, $\leq 1.0\%$). In some embodiments, the a host cell (HC) impurity level is equal to or less than 0.9% (*e.g.*, $\leq 0.9\%$). In some embodiments, the host cell (HC) impurity level is equal to or less than 0.8% (*e.g.*, $\leq 0.8\%$). In some embodiments, the host cell (HC) impurity level is equal to or less than 0.7% (*e.g.*, $\leq 0.7\%$). In some embodiments, the host cell (HC) impurity level is equal to or less than 0.6% (*e.g.*, $\leq 0.6\%$). In some embodiments, the host cell (HC) impurity level is equal to or less than 0.5% (*e.g.*, $\leq 0.5\%$). In some embodiments, the host cell (HC) impurity level is equal to or less than 0.4% (*e.g.*, $\leq 0.4\%$). In some embodiments, the host cell (HC) impurity level is equal to or less than 0.3% (*e.g.*, $\leq 0.3\%$). In some embodiments, the host cell (HC) impurity level is equal to or less than 0.2% (*e.g.*, $\leq 0.2\%$). In some embodiments, the host cell (HC) impurity level is equal to or less than 0.1% (*e.g.*, $\leq 0.1\%$).

[00250] In some embodiments, the rVWF-PP impurity is less than 15%, less than 10%, less than 5%, less than 4%, less than 3%, less than 2%, less than 1%, less than 0.5%, less than 0.4%, less than 0.3%, less than 0.2%, less than 0.1%, or less than 0.05%. In some embodiments, the rVWF-PP impurity is less than 15%. In some embodiments, the rVWF-PP impurity is less than 10%. In some embodiments, the rVWF-PP impurity is less than 5%. In

some embodiments, the rVWF-PP impurity is less than 4%. In some embodiments, the rVWF-PP impurity is less than 3%. In some embodiments, the rVWF-PP impurity is less than 2%. In some embodiments, the rVWF-PP impurity is less than 1%. In some embodiments, the rVWF-PP impurity is less than 0.5%. In some embodiments, the rVWF-PP impurity is less than 0.4%. In some embodiments, the rVWF-PP impurity is less than 0.3%. In some embodiments, the rVWF-PP impurity is less than 0.2%. In some embodiments, the rVWF-PP impurity is less than 0.1%. In some embodiments, the rVWF-PP impurity is less than 0.05%.

Table 1: Exemplary VWF-PP removal capacity

Step	Load, VWF-PP impurity % (w/w)	Elate, VWF-PP impurity % (w/w)
AEX	~30%*	~12%
CEX	~30%	~<0.1%
SEC	~12%	~<0.1%
*: either pre-matured before load or matured to completion by in-vitro maturation on column (as currently done in the process and part of a claim of a different patent)		

F. RECOMBINANT VWF PRODUCTION

[00251] The free mature recombinant von Willebrand Factor (rVWF) of the present invention can be produced recombinantly. One skilled in the art recognizes useful methods for expressing a recombinant protein in a host cell. In some instances, the method includes expressing a nucleic acid sequence encoding rVWF in a host cell such as a CHO cell and culturing the resulting host cell under certain conditions to produce rVWF, prepro-VWF, pro-VWF, and the like.

[00252] In certain embodiments, the nucleic acid sequence comprising a sequence coding for VWF can be an expression vector. The vector can be delivered by a virus or can be a plasmid. The nucleic acid sequence coding for the protein can be a specific gene or a biologically functional part thereof. In one embodiment, the protein is at least a biologically active part of VWF. The nucleic acid sequence can further comprise other sequences suitable for a controlled expression of a protein such as promoter sequences, enhancers, TATA boxes, transcription initiation sites, polylinkers, restriction sites, poly-A-sequences, protein processing sequences, selection markers, and the like which are generally known to a person of ordinary skill in the art.

[00253] A wide variety of vectors can be used for the expression of the VWF and can be selected from eukaryotic expression vectors. Examples of vectors for eukaryotic expression include: (i) for expression in yeast, vectors such as pAO, pPIC, pYES, pMET, using promoters such as AOX1, GAP, GAL1, AUG1, etc; (ii) for expression in insect cells, vectors such as pMT, pAc5, pIB, pMIB, pBAC, etc., using promoters such as PH, p10, MT, Ac5, OpIE2, gp64, polh, etc., and (iii) for expression in mammalian cells, vectors such as pSVL, pCMV, pRc/RSV, pcDNA3, pBPV, etc., and vectors derived from viral systems such as vaccinia virus, adeno-associated viruses, herpes viruses, retroviruses, etc., using promoters such as CMV, SV40, EF-1, UbC, RSV, ADV, BPV, and β -actin.

[00254] In some aspects, the rVWF used in the methods of the present invention is produced by expression in a mammalian cell culture using methods known in the art. In particular embodiments, the mammalian culture comprises CHO cells. In further embodiments, the rVWF is co-expressed with recombinant Factor VIII (rFVIII) in the same culture. In such embodiments, the rVWF and the rFVIII are purified together (co-purified) or separately using methods known in the art. In other embodiments, the rVWF is expressed in a culture that does not contain rFVIII.

[00255] In some embodiments, rVWF is expressed and isolated from a suitable eukaryotic host system. Examples of eukaryotic cells include, without limitation, mammalian cells, such as CHO, COS, HEK 293, BHK, SK-Hep, and HepG2; insect cells, *e.g.*, SF9 cells, SF21 cells, S2 cells, and High Five cells; and yeast cells, *e.g.*, *Saccharomyces* or *Schizosaccharomyces* cells. In one embodiment, the VWF can be expressed in yeast cells, insect cells, avian cells, mammalian cells, and the like. For example, in a human cell line, a hamster cell line, or a murine cell line. In one particular embodiment, the cell line is a CHO, BHK, or HEK cell line. Typically, mammalian cells, *e.g.*, CHO cell from a continuous cell line, can be used to express the VWF of the present invention. In certain instances, VWF protein is expressed and isolated from a CHO cell expression system.

[00256] VWF can be produced in a cell culture system or according to any cell culture method recognized by those in the art. In some embodiments, the cell cultures can be performed in large bioreactors under conditions suitable for providing high volume-specific culture surface areas to achieve high cell densities and protein expression. One means for providing such growth conditions is to use microcarriers for cell-culture in stirred tank bioreactors. The concept of cell-growth on microcarriers was first described by van Wezel

(van Wezel, A. L., *Nature*, 1967, 216:64-5) and allows for cell attachment on the surface of small solid particles suspended in the growth medium. These methods provide for high surface-to-volume ratios and thus allow for efficient nutrient utilization. Furthermore, for expression of secreted proteins in eukaryotic cell lines, the increased surface-to-volume ratio allows for higher levels of secretion and thus higher protein yields in the supernatant of the culture. Finally, these methods allow for the easy scale-up of eukaryotic expression cultures.

[00257] The cells expressing VWF can be bound to a spherical or a porous microcarrier during cell culture growth. The microcarrier can be a microcarrier selected from the group of microcarriers based on dextran, collagen, plastic, gelatine and cellulose and others as described in Butler (1988. In: Spier & Griffiths, *Animal Cell Biotechnology* 3:283-303). It is also possible to grow the cells to a biomass on spherical microcarriers and subculture the cells when they have reached final fermenter biomass and prior to production of the expressed protein on a porous microcarrier or vice versa. Suitable spherical microcarriers can include smooth surface microcarriers, such as Cytodex™ 1, Cytodex™ 2, and Cytodex™ 3 (GE Healthcare) and macroporous microcarriers such as Cytopore™ 1, Cytopore™ 2, Cytoline™ 1, and Cytoline™ 2 (GE Healthcare).

[00258] In a further embodiment, the VWF propeptide is cleaved from the non-mature VWF in vitro through exposure of the pro-VWF to furin. In some embodiments, the furin used for propeptide cleavage is recombinant furin.

[00259] In certain embodiments, rVWF is expressed in cells cultured in cell culture media that produces high molecular weight rVWF. The terms "cell culture solution," "cell culture medium or media," and "cell culture supernatant" refer to aspects of cell culture processes generally well known in the art. In the context of the present invention, a cell culture solution can include cell culture media and cell culture supernatant. The cell culture media are externally added to the cell culture solution, optionally together with supplements, to provide nutrients and other components for culturing the cells expressing VWF. The cell culture supernatant refers to a cell culture solution comprising the nutrients and other components from the cell culture medium as well as products released, metabolized, and/or excreted from the cells during culture. In further embodiments, the media can be animal protein-free and chemically defined. Methods of preparing animal protein-free and chemically defined culture media are known in the art, for example in US 2006/0094104, US 2007/0212770, and US 2008/0009040, which are both incorporated herein for all purposes.

and in particular for all teachings related to cell culture media. "Protein free" and related terms refers to protein that is from a source exogenous to or other than the cells in the culture, which naturally shed proteins during growth. In another embodiment, the culture medium is polypeptide free. In another embodiment, the culture medium is serum free. In another embodiment the culture medium is animal protein free. In another embodiment the culture medium is animal component free. In another embodiment, the culture medium contains protein, *e.g.*, animal protein from serum such as fetal calf serum. In another embodiment, the culture has recombinant proteins exogenously added. In another embodiment, the proteins are from a certified pathogen free animal. The term "chemically defined" as used herein shall mean, that the medium does not comprise any undefined supplements, such as, for example, extracts of animal components, organs, glands, plants, or yeast. Accordingly, each component of a chemically defined medium is accurately defined. In a preferred embodiment, the media are animal-component free and protein free.

[00260] In certain embodiments, the culture of cells expressing VWF can be maintained for at least about 7 days, or at least about 14 days, 21 days, 28 days, or at least about 5 weeks, 6 weeks, 7 weeks, or at least about 2 months, or 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18 months or longer. The cell density at which a cell-culture is maintained at for production of a recombinant VWF protein will depend upon the culture-conditions and medium used for protein expression. One of skill in the art will readily be able to determine the optimal cell density for a cell-culture producing an VWF. In one embodiment, the culture is maintained at a cell density of between about 0.5×10^6 and 4×10^7 cells/ml for an extended period of time. In other embodiments, the cell density is maintained at a concentration of between about 1.0×10^6 and about 1.0×10^7 cells/ml for an extended period of time. In other embodiments, the cell density is maintained at a concentration of between about 1.0×10^6 and about 4.0×10^6 cells/ml for an extended period of time. In other embodiments, the cell density is maintained at a concentration of between about 1.0×10^6 and about 4.0×10^6 cells/ml for an extended period of time. In yet other embodiments, the cell density may be maintained at a concentration between about 2.0×10^6 and about 4.0×10^6 , or between about 1.0×10^6 and about 2.5×10^6 , or between about 1.5×10^6 and about 3.5×10^6 , or any other similar range, for an extended period of time. After an appropriate time in cell culture, the rVWF can be isolated from the expression system using methods known in the art.

[00261] In a specific embodiment, the cell density of the continuous cell culture for production of rVWF is maintained at a concentration of no more than 2.5×10^6 cells/mL for an extended period. In other specific embodiments, the cell density is maintained at no more than 2.0×10^6 cells/mL, 1.5×10^6 cells/mL, 1.0×10^6 cells/mL, 0.5×10^6 cells/mL, or less. In one embodiment, the cell density is maintained at between 1.5×10^6 cells/mL and 2.5×10^6 cells/mL.

[00262] In one embodiment of the cell cultures described above, the cell culture solution comprises a medium supplement comprising copper. Such cell culture solutions are described, for example, in U.S. Patent No. 8,852,888 and U.S. Patent No. 9,409,971, which is hereby incorporated by reference in its entirety for all purposes and in particular for all teachings related to cell culture methods and compositions for producing recombinant VWF.

[00263] The polynucleotide and amino acid sequences of prepro-VWF are set out in SEQ ID NO:1 and SEQ ID NO:2, respectively, and are available at GenBank Accession Nos. NM_000552 (Homo sapiens von Willebrand factor (VWF) mRNA) and NP_000543, respectively. The amino acid sequence corresponding to the mature VWF protein is set out in SEQ ID NO: 3 (corresponding to amino acids 764-2813 of the full length prepro-VWF amino acid sequence). In some embodiments, the VWF exhibits at least 80%, at least 85%, at least 90%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or at least 100% identity to the sequence of SEQ ID NO:3. In some embodiments, the mat-rVWF of the invention exhibits at least 80%, at least 85%, at least 90%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or at least 100% identity to the sequence of SEQ ID NO:3. See, for example, U.S. Patent No. 8,597,910, U.S. Patent Publication No. 2016/0129090, as well as FIG. 60.

[00264] One form of useful rVWF has at least the property of in vivo-stabilizing, *e.g.* binding, of at least one Factor VIII (FVIII) molecule and having optionally a glycosylation pattern which is pharmacologically acceptable. Specific examples thereof include VWF without the A2 domain thus resistant to proteolysis (Lankhof *et al.*, Thromb. Haemost. 77: 1008-1013, 1997), and a VWF fragment from Val 449 to Asn 730 including the glycoprotein Ib-binding domain and binding sites for collagen and heparin (Pietu *et al.*, Biochem. Biophys. Res. Commun. 164: 1339-1347, 1989). The determination of the ability of a VWF to stabilize at least one FVIII molecule is, in one aspect, carried out in VWF-deficient mammals according to methods known in the state in the art.

[00265] The rVWF of the present invention can be produced by any method known in the art. One specific example is disclosed in WO86/06096 published on Oct. 23, 1986 and U.S. Patent Application No. 07/559,509, filed on Jul. 23, 1990, which is incorporated herein by reference with respect to the methods of producing recombinant VWF. Thus, methods are known in the art for (i) the production of recombinant DNA by genetic engineering, *e.g.* via reverse transcription of RNA and/or amplification of DNA, (ii) introducing recombinant DNA into prokaryotic or eukaryotic cells by transfection, *e.g.* via electroporation or microinjection, (iii) cultivating the transformed cells, *e.g.* in a continuous or batchwise manner, (iv) expressing VWF, *e.g.* constitutively or upon induction, and (v) isolating the VWF, *e.g.* from the culture medium or by harvesting the transformed cells, in order to (vi) obtain purified rVWF, *e.g.* via anion exchange chromatography or affinity chromatography. A recombinant VWF is, in one aspect, made in transformed host cells using recombinant DNA techniques well known in the art. For instance, sequences coding for the polypeptide could be excised from DNA using suitable restriction enzymes. Alternatively, the DNA molecule is, in another aspect, synthesized using chemical synthesis techniques, such as the phosphoramidate method. Also, in still another aspect, a combination of these techniques is used.

[00266] The invention also provides vectors encoding polypeptides of the invention in an appropriate host. The vector comprises the polynucleotide that encodes the polypeptide operatively linked to appropriate expression control sequences. Methods of effecting this operative linking, either before or after the polynucleotide is inserted into the vector, are well known. Expression control sequences include promoters, activators, enhancers, operators, ribosomal binding sites, start signals, stop signals, cap signals, polyadenylation signals, and other signals involved with the control of transcription or translation. The resulting vector having the polynucleotide therein is used to transform an appropriate host. This transformation may be performed using methods well known in the art.

[00267] Any of a large number of available and well-known host cells are used in the practice of this invention. The selection of a particular host is dependent upon a number of factors recognized by the art, including, for example, compatibility with the chosen expression vector, toxicity of the peptides encoded by the DNA molecule, rate of transformation, ease of recovery of the peptides, expression characteristics, bio-safety and costs. A balance of these factors must be struck with the understanding that not all host cells

are equally effective for the expression of a particular DNA sequence. Within these general guidelines, useful microbial host cells include, without limitation, bacteria, yeast and other fungi, insects, plants, mammalian (including human) cells in culture, or other hosts known in the art.

[00268] Transformed host cells are cultured under conventional fermentation conditions so that the desired compounds are expressed. Such fermentation conditions are well known in the art. Finally, the polypeptides are purified from culture media or the host cells themselves by methods well known in the art.

[00269] Depending on the host cell utilized to express a compound of the invention, carbohydrate (oligosaccharide) groups are optionally attached to sites that are known to be glycosylation sites in proteins. Generally, O-linked oligosaccharides are attached to serine (Ser) or threonine (Thr) residues while N-linked oligosaccharides are attached to asparagine (Asn) residues when they are part of the sequence Asn-X-Ser/Thr, where X can be any amino acid except proline. X is preferably one of the 19 naturally occurring amino acids not counting proline. The structures of N-linked and O-linked oligosaccharides and the sugar residues found in each type are different. One type of sugar that is commonly found on both N-linked and O-linked oligosaccharides is N-acetylneuraminic acid (referred to as sialic acid). Sialic acid is usually the terminal residue of both N-linked and O-linked oligosaccharides and, by virtue of its negative charge, in one aspect, confers acidic properties to the glycosylated compound. Such site(s) may be incorporated in the linker of the compounds of this invention and are preferably glycosylated by a cell during recombinant production of the polypeptide compounds (*e.g.*, in mammalian cells such as CHO, BHK, COS). In other aspects, such sites are glycosylated by synthetic or semi-synthetic procedures known in the art.

[00270] In some embodiments, sialylation (also referred to as sialylation), can be performed on the column as part of the purification procedures described herein (including the anion exchange, cation exchange, size exclusion, and/or immunoaffinity methods). In some embodiments, the sialylation results in increased stability of the rVWF as compared to rVWF that has not undergone sialylation. In some embodiments, the sialylation results in increased stability of the rVWF in blood circulation (for example, after administration to a subject) as compared to rVWF that has not undergone sialylation. In some embodiments, the increased stability of sialated rVWF results in an increase of 10%, 20%, 30%, 40%, 50%,

60%, 70%, 80%, 90%, or more as compared rVWF that has not undergone sialylation. In some embodiments, the sialylation results in increased half-life for the rVWF as compared to rVWF that has not undergone sialylation. In some embodiments, the sialylation results in increased half-life for the rVWF in blood circulation (for example, after administration to a subject) as compared to rVWF that has not undergone sialylation. In some embodiments, the increased half-life of sialylated rVWF results in an increase of 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90%, or more as compared rVWF that has not undergone sialylation. In some embodiments, the increased half-life of sialylated rVWF results in rVWF that is stable for 1 hour, 2 hours, 3 hours, 4 hours, 6 hours, 12 hours, 24 hours or more in blood circulation (for example, after administration to a subject) as compared to rVWF that has not undergone sialylation. In some embodiments, sialylation increases the number of 2,3 sialylation and/or 2,6 sialylation. In some embodiments, sialylation is increased by the addition of 2,3 sialyltransferase and/or 2,6 sialyltransferase and CMP-NANA (Cytidine-5'-monophospho-N-acetylneuraminic acid sodium salt) as an additional buffer step. In some embodiments, sialylation is increased by the addition of 2,3 sialyltransferase and CMP-NANA (Cytidine-5'-monophospho-N-acetylneuraminic acid sodium salt) as an additional buffer step. In some embodiments, 2,3 sialylation is increased by the addition of 2,3 sialyltransferase and CMP-NANA (Cytidine-5'-monophospho-N-acetylneuraminic acid sodium salt) as an additional buffer step. In some embodiments, in order to increase sialylation, the bound protein (for example, bound rVWF) is treated with sialidase (e.g., neuraminidase) to remove the 2,3 sialylation and then a wash step is applied to remove the sialidase and introduce 2,6 sialylation. In some embodiments, the 2,6 sialylation is introduced by the addition of 2,6 sialyltransferase and CMP-NANA.

[00271] In some embodiments, 2,6 sialylation is increased by the addition of 2,6 sialyltransferase and CMP-NANA (Cytidine-5'-monophospho-N-acetylneuraminic acid sodium salt) as an additional buffer step. In some embodiments, 2,3 sialylation and/or 2,6 sialylation are increased by the addition of 2,3 sialyltransferase and/or 2,6 sialyltransferase and CMP-NANA (Cytidine-5'-monophospho-N-acetylneuraminic acid sodium salt) as an additional buffer step. In some embodiments, CMP-NANA is chemically or enzymatic modified to transfer modified sialic acid to potential free position. In some embodiments, sialylation is performed by loading rVWF onto the resin, washing with one or more buffers as described herein to deplete unwanted impurities, apply one or more buffers containing

sialyltransferase and CMP-NANA at conditions that allow additional sialylation, and washing with one or more buffers to deplete excess of the sialylation reagents, and eluting with one or more buffers the enhanced rVWF (*e.g.*, the rVWF with increased sialylation). In some embodiments, the sialylation process is performed as part of a cation exchange method, an anion exchange method, a size exclusion method, or an immunoaffinity purification method, as described herein.

[00272] Alternatively, the compounds are made by synthetic methods using, for example, solid phase synthesis techniques. Suitable techniques are well known in the art, and include those described in Merrifield (1973), *Chem. Polypeptides*, pp. 335-61 (Katsoyannis and Panayotis eds.); Merrifield (1963), *J. Am. Chem. Soc.* 85: 2149; Davis *et al.* (1985), *Biochem. Intl.* 10: 394-414; Stewart and Young (1969), *Solid Phase Peptide Synthesis*; U.S. Pat. No. 3,941,763; Finn *et al.* (1976), *The Proteins* (3rd ed.) 2: 105-253; and Erickson *et al.* (1976), *The Proteins* (3rd ed.) 2: 257-527'. Solid phase synthesis is the preferred technique of making individual peptides since it is the most cost-effective method of making small peptides

[00273] Fragments, variants and analogs of VWF can be produced according to methods that are well-known in the art. Fragments of a polypeptide can be prepared using, without limitation, enzymatic cleavage (*e.g.*, trypsin, chymotrypsin) and also using recombinant means to generate a polypeptide fragments having a specific amino acid sequence. Polypeptide fragments may be generated comprising a region of the protein having a particular activity, such as a multimerization domain or any other identifiable VWF domain known in the art.

[00274] Methods of making polypeptide analogs are also well-known. Amino acid sequence analogs of a polypeptide can be substitutional, insertional, addition or deletion analogs. Deletion analogs, including fragments of a polypeptide, lack one or more residues of the native protein which are not essential for function or immunogenic activity. Insertional analogs involve the addition of, *e.g.*, amino acid(s) at a non-terminal point in the polypeptide. This analog may include, for example and without limitation, insertion of an immunoreactive epitope or simply a single residue. Addition analogs, including fragments of a polypeptide, include the addition of one or more amino acids at either or both termini of a protein and include, for example, fusion proteins. Combinations of the aforementioned analogs are also contemplated.

[00275] Substitutional analogs typically exchange one amino acid of the wild-type for another at one or more sites within the protein, and may be designed to modulate one or more properties of the polypeptide without the complete loss of other functions or properties. In one aspect, substitutions are conservative substitutions. “Conservative amino acid substitution” is substitution of an amino acid with an amino acid having a side chain or a similar chemical character. Similar amino acids for making conservative substitutions include those having an acidic side chain (glutamic acid, aspartic acid); a basic side chain (arginine, lysine, histidine); a polar amide side chain (glutamine, asparagine); a hydrophobic, aliphatic side chain (leucine, isoleucine, valine, alanine, glycine); an aromatic side chain (phenylalanine, tryptophan, tyrosine); a small side chain (glycine, alanine, serine, threonine, methionine); or an aliphatic hydroxyl side chain (serine, threonine).

[00276] In one aspect, analogs are substantially homologous or substantially identical to the recombinant VWF from which they are derived. Analogs include those which retain at least some of the biological activity of the wild-type polypeptide, *e.g.* blood clotting activity.

[00277] Polypeptide variants contemplated include, without limitation, polypeptides chemically modified by such techniques as ubiquitination, glycosylation, including polysialation (or polysialylation), conjugation to therapeutic or diagnostic agents, labeling, covalent polymer attachment such as pegylation (derivatization with polyethylene glycol), introduction of non-hydrolyzable bonds, and insertion or substitution by chemical synthesis of amino acids such as ornithine, which do not normally occur in human proteins. Variants retain the same or essentially the same binding properties of non-modified molecules of the invention. Such chemical modification may include direct or indirect (*e.g.*, via a linker) attachment of an agent to the VWF polypeptide. In the case of indirect attachment, it is contemplated that the linker may be hydrolyzable or non-hydrolyzable.

[00278] Preparing pegylated polypeptide analogs will in one aspect comprise the steps of (a) reacting the polypeptide with polyethylene glycol (such as a reactive ester or aldehyde derivative of PEG) under conditions whereby the binding construct polypeptide becomes attached to one or more PEG groups, and (b) obtaining the reaction product(s). In general, the optimal reaction conditions for the acylation reactions are determined based on known parameters and the desired result. For example, the larger the ratio of PEG: protein, the greater the percentage of poly-pegylated product. In some embodiments, the binding construct has a single PEG moiety at the N-terminus. Polyethylene glycol (PEG) may be

attached to the blood clotting factor to, for example, provide a longer half-life in vivo. The PEG group may be of any convenient molecular weight and is linear or branched. The average molecular weight of the PEG ranges from about 2 kiloDalton ("kD") to about 100 kDa, from about 5 kDa to about 50 kDa, or from about 5 kDa to about 10 kDa. In certain aspects, the PEG groups are attached to the blood clotting factor via acylation or reductive alkylation through a natural or engineered reactive group on the PEG moiety (*e.g.*, an aldehyde, amino, thiol, or ester group) to a reactive group on the blood clotting factor (*e.g.*, an aldehyde, amino, or ester group) or by any other technique known in the art.

[00279] Methods for preparing polysialylated polypeptide are described in United States Patent Publication 20060160948, Fernandes et Gregoriadis; *Biochim. Biophys. Acta* 1341: 26-34, 1997, and Saenko *et al.*, *Haemophilia* 12:42-51, 2006. Briefly, a solution of colominic acid (CA) containing 0.1 M NaIO₄ is stirred in the dark at room temperature to oxidize the CA. The activated CA solution is dialyzed against, *e.g.*, 0.05 M sodium phosphate buffer, pH 7.2 in the dark and this solution was added to a rVWF solution and incubated for 18 h at room temperature in the dark under gentle shaking. Free reagents are optionally be separated from the rVWF-polysialic acid conjugate by, for example, ultrafiltration/diafiltration. Conjugation of rVWF with polysialic acid is achieved using glutaraldehyde as cross-linking reagent (Migneault *et al.*, *Biotechniques* 37: 790-796, 2004).

[00280] It is further contemplated in another aspect that a polypeptide of the invention is a fusion protein with a second agent which is a polypeptide. In one embodiment, the second agent which is a polypeptide, without limitation, is an enzyme, a growth factor, an antibody, a cytokine, a chemokine, a cell- surface receptor, the extracellular domain of a cell surface receptor, a cell adhesion molecule, or fragment or active domain of a protein described above. In a related embodiment, the second agent is a blood clotting factor such as Factor VIII, Factor VII, and/or Factor IX. In some embodiments, the second agent is a fusion protein. The fusion protein contemplated is made by chemical or recombinant techniques well-known in the art. In some embodiments, the fusion protein is a rVWF-FVIII fusion protein. In some embodiments, the fusion protein is a rVWF-FVIII fusion protein, wherein an active FVIII is embedded in an VWF motif. In some embodiments, the fusion protein is a rVWF-FVIII fusion protein, wherein an active FVIII is embedded in an VWF motif such that the VWF is full length. In some embodiments, the fusion protein is a rVWF-FVIII fusion protein, wherein an active FVIII is embedded in an VWF motif, wherein parts of the VWF

sequence are deleted and replaced by a FVIII – sequence. In some embodiments of the rVWF-FVIII fusion protein, the FVIII is a B-domain deleted FVIII. In some embodiments of the rVWF-FVIII fusion protein, the N -glycosylation rich domain replaces the FVIII-B-domain. In some embodiments of the rVWF-FVIII fusion protein, the vWF-N glycosylation rich domain is fused to the full length FVIII and/or truncated forms thereof.

[00281] In some embodiments of the rVWF-FVIII fusion protein, the fusion protein comprises:

- a VWF peptide comprising positions 764 to 1336 of the VWF peptide,
- a FVIII peptide comprising positions 24 to 760 of the FVIII heavy chain peptide,
- a VWF peptide comprising positions 2218 to 2593 of the VWF peptide,
- a FVIII peptide comprising positions 1333 to 2351 of the FVIII light chain peptide,
- and
- a VWF peptide comprising positions 2620 to 2813 of the VWF peptide.

In this embodiment of the rVWF-FVIII fusion protein, the position of amino acids is counted from the first position – including Pro and/or signal peptide. In this embodiment of the rVWF-FVIII fusion protein, position 764 in VWF corresponds to position 1 of the mature rVWF (mat-rVWF) and position 20 in FVIII corresponds to position 1 of the mature FVIII peptide. In some embodiments of the rVWF-FVIII fusion protein, the fusion protein sequence is provided in Fig. 64.

[00282] In some embodiments of the rVWF-FVIII fusion protein, the fusion protein comprises:

- a FVIII peptide comprising positions FVIII heavy chain 19 to 760 of the FVIII heavy chain peptide,
- a VWF peptide comprising positions 2218 to 2593 of the VWF peptide, and
- a FVIII peptide comprising positions 1333 to 2351 of the FVIII light chain peptide.

In this embodiment of the rVWF-FVIII fusion protein, the position of amino acids is counted from the first position – including Pro and/or signal peptide. In this embodiment of the rVWF-FVIII fusion protein, position 764 in VWF corresponds to position 1 of the mature rVWF (mat-rVWF) and position 20 in FVIII corresponds to position 1 of the mature FVIII

peptide. In some embodiments of the rVWF-FVIII fusion protein, the fusion protein sequence is provided in Fig. 65.

[00283] It is also contemplated in another aspect that prepro-VWF and pro-VWF polypeptides will provide a therapeutic benefit in the formulations of the present invention. For example, US Patent No. 7,005,502 describes a pharmaceutical preparation comprising substantial amounts of pro-VWF that induces thrombin generation in vitro. In addition to recombinant, biologically active fragments, variants, or other analogs of the naturally-occurring mature VWF, the present invention contemplates the use of recombinant biologically active fragments, variants, or analogs of the prepro-VWF (set out in SEQ ID NO:2) or pro-VWF polypeptides (amino acid residues 23 to 764 of SEQ ID NO: 2) in the formulations described herein.

[00284] Polynucleotides encoding fragments, variants and analogs may be readily generated by a worker of skill to encode biologically active fragments, variants, or analogs of the naturally-occurring molecule that possess the same or similar biological activity to the naturally-occurring molecule. In various aspects, these polynucleotides are prepared using PCR techniques, digestion/ligation of DNA encoding molecule, and the like. Thus, one of skill in the art will be able to generate single base changes in the DNA strand to result in an altered codon and a missense mutation, using any method known in the art, including, but not limited to site- specific mutagenesis. As used herein, the phrase “moderately stringent hybridization conditions” means, for example, hybridization at 42 °C in 50% formamide and washing at 60°C in 0.1 x SSC, 0.1% SDS. It is understood by those of skill in the art that variation in these conditions occurs based on the length and GC nucleotide base content of the sequences to be hybridized. Formulas standard in the art are appropriate for determining exact hybridization conditions. See Sambrook *et al.*, 9.47-9.51 in Molecular Cloning, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York (1989).

G. VIRAL INACTIVATION

[00285] In some embodiments, the method described herein further comprises a step of viral inactivation. The viral inactivation step can occur before, after, or concurrently with the washing step and/or the elution step, but before the collecting step. The viral inactivation treatment can inactivate lipid enveloped viruses. In some embodiments, the viral inactivation treatment is a solvent and detergent (S/D) treatment. In some embodiments, the viral

inactivation treatment includes the use of ethylene glycol, propylenglycol in section alcohols and/or one or more organic solvent(s).

[00286] As used herein, the term “inactivating virus” or “virus inactivation” refers to a process where a virus can no longer infect cells, replicate, and propagate, and per se virus removal. As such, the term “virus inactivation” refers generally to the process of making a fluid disclosed herein completely free of infective viral contaminants. Any degree of viral inactivation using the methods disclosed herein is desirable. However, it is desirable to achieve the degree of viral inactivation necessary to meet strict safety guidelines for pharmaceuticals. These guidelines are set forth by the WHO and well known to those of skill in the art.

[00287] The methods disclosed herein, may further comprise a step of removing a virus from the mixture after incubation. As used herein, the term “removing a virus” or “virus removal” refers to a process that depletes a virus from a mixture disclosed herein, such that the virus particles are effectively extracted from the mixture. The virus can be a viable virus or an inactivated virus. Removal is typically accomplished by size exclusion chromatography or positive adsorption chromatography where the protein of interest binds to a chromatographic resin, including for example, an anion exchange resin or cation exchange resin as described herein. After removal, the amount of a virus remaining is an amount that has substantially no long term or permanent detrimental effect when administered to a subject in need thereof, including for example, a human being.

[00288] In one embodiment, a mixture after removal of virus is essentially free of the virus. As used herein, the term “essentially free of a virus” means that only trace amounts of a virus can be detected or confirmed by the instrument or process being used to detect or confirm the presence or activity of the virus and that such trace amount of the virus is insufficient to be deleterious to the health of the human being. In an aspect of this embodiment, a mixture after removal of virus is entirely free of the virus. As used herein, the term “entirely free of a virus” means that the presence of virus cannot be detected or confirmed within the detection range of the instrument or process being used to detect or confirm the presence or activity of the virus. A protein contained within a mixture that is essentially free or entirely free of a virus can be used to make a pharmaceutical composition that is safe to administer to a human being because the virus is insufficient to be deleterious to the health of the human being.

[00289] In other aspects of this embodiment, a mixture after removal of virus comprises less than 10 PFU/mL of a virus, such as, *e.g.*, less than 1 PFU/mL of a virus, less than 1×10^{-1} PFU/mL of a virus, 1×10^{-2} PFU/mL of a virus, or 1×10^{-3} PFU/mL of a virus.

[00290] In yet other aspects of this embodiment, a mixture after removal of virus comprises less than an ID50 for a virus, such as, *e.g.*, at least 10-fold less than the ID50 for a virus, at least 100-fold less than the ID50 for a virus, at least 200-fold less than the ID50 for a virus, at least 300-fold less than the ID50 for a virus, at least 400-fold less than the ID50 for a virus, at least 500-fold less than the ID50 for a virus, at least 600-fold less than the ID50 for a virus, at least 700-fold less than the ID50 for a virus, at least 800-fold less than the ID50 for a virus, at least 900-fold less than the ID50 for a virus, or at least 1000-fold less than the ID50 for a virus.

[00291] The viral inactivation may be carried out in conjunction with protein purification or not. In some embodiments, the method comprises immobilizing the protein on a support; and treating the immobilized protein with a detergent-solvent mixture comprising a non-ionic detergent and an organic solvent. In some embodiments, the support is a chromatographic resin. In certain embodiments, the detergent-solvent mixture comprises 1% Triton X-100, 0.3% Tri-N-butyl phosphate, and 0.3% Polysorbate 80 (Tween 80). The solvent-detergent mixture treatment can continue for a prolonged time, *e.g.*, for 30 minutes to 1 hour, while the protein remains immobilized on the chromatographic resin, *e.g.*, on a cation exchange resin; and/or solvent-detergent treatment may occur at 2°C to 10°C. This approach to virus inactivation surprisingly can reduce the formation of protein aggregates during treatment with a detergent-solvent mixture by a significant amount, *e.g.*, by more than 50%, as compared to treatment with a solvent-detergent mixture while the protein is not immobilized in solution.

[00292] In some embodiments, the method of inactivating a lipid-coat containing virus comprises the steps of: i) providing a fluid comprising a protein having an activity; ii) mixing an organic solvent and a surfactant with the fluid, thereby creating a mixture; and iii) incubating the mixture for no more than about 120 minutes; wherein both steps (ii) and (iii) are performed at a temperature of no higher than about 20° C.; wherein the mixture after incubation is essentially free of a viable lipid-coat containing virus; and wherein the protein after incubation has an activity of at least 25% of the activity provided in step (i).

[00293] In other embodiments, a protein essentially free of a lipid-coat containing virus can be obtained from a method comprising the steps of: i) providing a fluid comprising a protein having an activity; ii) mixing an organic solvent and a surfactant with the fluid, thereby creating a mixture; and iii) incubating the mixture for no more than about 120 minutes; wherein both steps (ii) and (iii) are performed at a temperature of no higher than about 20° C.; wherein the mixture after incubation is essentially free of a viable lipid-coat containing virus; and wherein the protein after incubation has an activity of at least 25% of the activity provided in step (a).

[00294] In another embodiment, the method of inactivating a lipid-coat containing virus comprises the steps of: i) providing a fluid comprising a blood coagulation protein having an activity (*e.g.*, VWF); ii) mixing an organic solvent and a surfactant with the fluid, thereby creating a mixture; and iii) incubating the mixture for no more than about 120 minutes; wherein both steps (ii) and (iii) are performed at a temperature of no higher than about 20° C.; wherein the mixture after incubation is essentially free of a viable lipid-coat containing virus; and wherein the Factor VIII after incubation has an activity of at least 25% of the activity provided in step (i).

[00295] In some instances, the organic solvent is an ether, an alcohol, a dialkylphosphate or a trialkylphosphate. In certain embodiments, the ether is selected from dimethyl ether, diethyl ether, ethyl propyl ether, methyl-butyl ether, methyl isopropyl ether, and/or methyl isobutyl ether.

[00296] In some embodiments, the alcohol is selected from methanol, ethanol, propanol, isopropanol, n-butanol, isobutanol, n-pentanol, and/or isopentanol. In some embodiments, the dialkylphosphate is selected from di-(n-butyl)phosphate, di-(t-butyl)phosphate, di-(n-hexyl)phosphate, di-(2-ethylhexyl)phosphate, di-(n-decyl)phosphate, and/or ethyl di(n-butyl) phosphate. In some embodiments, the trialkylphosphate is selected from tri-(n-butyl)phosphate, tri-(t-butyl)phosphate, tri-(n-hexyl)phosphate, tri-(2-ethylhexyl)phosphate, and/or tri-(n-decyl)phosphate.

[00297] In some instances, the final concentration of the organic solvent is from about 0.1% (v/v) to about 5.0% (v/v), about 0.1% (v/v) to about 1.0% (v/v), about 0.2% (v/v) to about 0.5% (v/v), or about 0.2% (v/v) to about 0.4% (v/v), about 0.3% (v/v).

[00298] In some instances, the surfactant is selected from an ionic surfactant, a zwitterionic (amphoteric) surfactant, and/or a non-ionic surfactant. The ionic surfactant can be an anion surfactant or cationic surfactant.

[00299] In certain embodiments, the anionic surfactant is selected from an alkyl sulfate, an alkyl ether sulfate, a docusate, a sulfonate fluorosurfactant, an alkyl benzene sulfonate, an alkyl aryl ether phosphate, an alkyl ether phosphate, an alkyl carboxylate, a sodium lauroyl sarcosinate, and/or a carboxylate fluorosurfactant. In some embodiments, the alkyl sulfate is selected from ammonium lauryl sulfate or sodium lauryl sulfate (SDS). In other embodiments, the alkyl ether sulfate is sodium laureth sulfate and/or sodium myreth sulfate. In some embodiments, the docusate is dioctyl sodium sulfosuccinate.

[00300] In some embodiments, the sulfonate fluorosurfactant is selected from perfluorooctanesulfonate (PFOS) and/or perfluorobutanesulfonate. In some embodiments, the alkyl carboxylate is selected from a fatty acid salt and/or sodium stearate. In some embodiments, the carboxylate fluorosurfactant is perfluorononanoate and perfluorooctanoate. In some embodiments, the cationic surfactant is selected from an alkyltrimethylammonium salt, cetylpyridinium chloride (CPC), polyethoxylated tallow amine (POEA), benzalkonium chloride (BAC), benzethonium chloride (BZT), 5-bromo-5-nitro-1,3-dioxane, dimethyldioctadecylammonium chloride, dioctadecyldimethylammonium bromide (DODAB), a pH-dependent primary amine, a pH-dependent secondary amine, and/or a pH-dependent tertiary amine. In some embodiments, the alkyltrimethylammonium salt is selected from cetyl trimethylammonium bromide (CTAB) and/or cetyl trimethylammonium chloride (CTAC). In some embodiments, the primary amine becomes positively charged at pH < 10 or the secondary amine becomes charged at pH < 4.

[00301] In some embodiments, the cationic surfactant is octenidine dihydrochloride.

[00302] In some embodiments, the zwitterionic surfactant is selected from 3-[(3-cholamidopropyl) dimethylammonio]-1-propanesulfonate (CHAPS), a sultaine, a betaine, and/or a lecithin. In some embodiments, the sultaine is cocamidopropyl hydroxysultaine. In some embodiments, the betaine is cocamidopropyl betaine.

[00303] In some embodiments, the non-ionic surfactant is selected from a polyoxyethylene glycol sorbitan alkyl ester, a poloxamer, an alkyl phenol polyglycol ether, a polyethylene glycol alkyl aryl ether, a polyoxyethylene glycol alkyl ether, 2-dodecoxyethanol

(LUBROL®-PX), a polyoxyethylene glycol octylphenol ether, a polyoxyethylene glycol alkylphenol ether, a phenoxypolyethoxyethanol, a glucoside alkyl ether, a maltoside alkyl ether, a thioglucoside alkyl ether, a digitonin, a glycerol alkyl ester, an alkyl aryl polyether sulfate, an alcohol sulfonate, a sorbitan alkyl ester, a cocamide ethanolamine, sucrose monolaurate, dodecyl dimethylamine oxide, and/or sodium cholate. In some embodiments, the polyoxyethylene glycol sorbitan alkyl ester is selected from polysorbate 20 sorbitan monooleate (TWEEN® 20), polysorbate 40 sorbitan monooleate (TWEEN® 40), polysorbate 60 sorbitan monooleate (TWEEN® 60), polysorbate 61 sorbitan monooleate (TWEEN® 61), polysorbate 65 sorbitan monooleate (TWEEN® 65), polysorbate 80 sorbitan monooleate (TWEEN® 80), and/or polysorbate 81 sorbitan monooleate (TWEEN® 81).

[00304] In some embodiments, the poloxamer is selected from Poloxamer 124 (PLURONIC® L44), Poloxamer 181 (PLURONIC® L61), Poloxamer 182 (PLURONIC® L62), Poloxamer 184 (PLURONIC® L64), Poloxamer 188 (PLURONIC® F68), Poloxamer 237 (PLURONIC® F87), Poloxamer 338 (PLURONIC® L108), and/or Poloxamer 407 (PLURONIC® F127).

[00305] In some embodiments, the polyoxyethylene glycol alkyl ether is selected from octaethylene glycol monododecyl ether, pentaethylene glycol monododecyl ether, BRIJ® 30, and/or BRIJ® 35.

[00306] In some cases, the polyoxyethylene glycol octylphenol ether is selected from polyoxyethylene (4-5) p-t-octyl phenol (TRITON® X-45), and/or polyoxyethylene octyl phenyl ether (TRITON® X-100). In some embodiments, the polyoxyethylene glycol alkylphenol ether is nonoxynol-9.

[00307] In some embodiments, the phenoxypolyethoxyethanol is selected from nonyl phenoxypolyethoxyethanol and/or octyl phenoxypolyethoxyethanol.

In some embodiments, the glucoside alkyl ether is octyl glucopyranoside. In some embodiments, the maltoside alkyl ether is dodecyl maltopyranoside. In some embodiments, the thioglucoside alkyl ether is heptyl thioglucopyranoside. In some embodiments, the glycerol alkyl ester is glyceryl laurate. In some embodiments, the cocamide ethanolamine is selected from cocamide monoethanolamine and/or cocamide diethanolamine.

[00308] In some embodiments, the final concentration of the surfactant is from about 0.1% (v/v) to about 10.0% (v/v), or about 0.5% (v/v) to about 5.0% (v/v). In some cases, the surfactant is a plurality of surfactants.

[00309] Useful methods for viral inactivation are described, for example, in U.S. Patent Nos. 6,190,609 and 9,315,560, and U.S. Appl. Publication No. 2017/0327559, the disclosures of which are herein incorporated by reference in their entireties.

[00310] Viral inactivation can be performed as recognized by those skilled in the art. For instance, the solvent tri(n-butyl) phosphate (TNBP) and detergents such as, but not limited to, polysorbate 80 and triton X-100 are effective for inactivating lipid enveloped viruses. Viral inactivation can be performed at room temperatures such as 14°C to about 25°C for about 1 hour or more. In some cases, the incubation time is not longer than two hours.

[00311] In some embodiments, the viral inactivation treatment is stopped by adding a buffer comprising a sodium citrate buffer to the virus inactivated material. In some instances, the sodium citrate buffer comprises from about 40 mM to about 100 mM sodium citrate buffer, *e.g.*, about 40 mM, about 50 mM, about 60 mM, about 70 mM, about 80 mM, about 90 mM, or about 100 mM sodium citrate buffer.

H. VWF MATURATION

[00312] Furin is part of a protein family referred to as SPC (subtilisin-like proprotein convertases), PC (proprotein convertases) or in some cases PACE (paired basic amino acid cleaving enzyme). Members of the furin protein family include but are not limited to Furin, Kex2, PC2, PC1/PC3, PACE4, PC4, PC5 and/or PC7. As part of the present invention, methods provides methods for maturation of pro-VWF (pro-rVWF) into a mat-VWF/VWF-PP (mat-rVWF/rVWF-PP) complex by treatment with furin. Any of these furin family members can be employed in the methods of VWF maturation.

[00313] In some embodiments, the pro-VWF is furin matured on an anion exchange column or resin, on a cation exchange column or resin, or as part of a size separation chromatography method. In some embodiments, the pro-VWF is furin matured on an anion exchange column or resin and/or as part of an anion exchange chromatography procedure. In some embodiments, the pro-VWF is furin matured on a cation exchange column or resin

and/or as part of a cation exchange chromatography procedure. In some embodiments, the pro-VWF is furin matured as part of a size exclusion chromatography procedure. Such methods have been described, for example, in U.S. Patent No. 8,058,411, incorporated by reference herein in its entirety for all purposes.

[00314] In order to facilitate the maturation process and to provide pro-VWF immobilized on the resin at an elevated concentration, in some embodiments of the invention, the chromatographic resin is packed in a chromatographic column. Since the concentration of pro-VWF in the course of its *in vitro* maturation influences the maturation efficiency, it is advantageous to pack the chromatographic resin in a column. Furthermore, the use of chromatographic columns allows the efficient control of the parameters of maturation in a more reproducible manner and makes it simpler to perform the maturation of VWF *in vitro*. In some embodiments, the furin concentration is about 1, about 2, about 3, or about 4 units of recombinant active furin per IU of VWF:Ag (10µg of pro-rVWF). In some embodiments, the furin concentration is about 2-3 units of recombinant active furin per IU of VWF:Ag (10µg of pro-rVWF). In some embodiments, the furin concentration is about 1-2 units of recombinant active furin per IU of VWF:Ag (10µg of pro-rVWF). In some embodiments, the furin concentration is about 2 units of recombinant active furin per IU of VWF:Ag (10µg of pro-rVWF).

[00315] In some embodiments, when the pro-VWF is immobilized on an anion exchange resin and incubated with a solution exhibiting pro-VWF convertase activity, the conductivity measured at 25°C is below 25 mS/cm. In some embodiments, when the pro-VWF is immobilized on an anion exchange resin and incubated with a solution exhibiting pro-VWF convertase activity, the conductivity measured at 25°C is below 20 mS/cm. In some embodiments, when the pro-VWF is immobilized on an anion exchange resin and incubated with a solution exhibiting pro-VWF convertase activity, the conductivity measured at 25°C is below 16 mS/cm. In some embodiments, when the pro-VWF is immobilized on an anion exchange resin and incubated with a solution exhibiting pro-VWF convertase activity, the conductivity measured at 25°C is between 16 mS/cm and 25 mS/cm. In some embodiments, when the pro-VWF is immobilized on an anion exchange resin and incubated with a solution exhibiting pro-VWF convertase activity, the conductivity measured at 25°C is between 20 mS/cm and 25 mS/cm. Pro-rVWF as well as mat-rVWF can be efficiently immobilized on anion exchange resins at these conductivity levels. Consequently, the buffers

applied in the course of the present method have to be adapted correspondingly to maintain the conductivity levels. In some embodiments, the conductivity is such that the furin and/or PACE enzyme is in active form and full or partially in the mobile phase.

[00316] In some embodiments, mat-rVWF is eluted from an anion exchange resin at a conductivity when measured at 25° C of at least 40 mS/cm. In some embodiments, mat-rVWF is eluted from an anion exchange resin at a conductivity when measured at 25° C of at least 60 mS/cm. In some embodiments, mat-rVWF is eluted from an anion exchange resin at a conductivity when measured at 25° C of at least 80 mS/cm. In some embodiments, mat-rVWF is eluted from an anion exchange resin at a conductivity when measured at 25° C of between 40 mS/cm and 80 mS/cm. In some embodiments, mat-rVWF is eluted from an anion exchange resin at a conductivity when measured at 25° C of between 60 mS/cm and 80 mS/cm. In some embodiments, the desired rVWF species starts to elute at a conductivity of between about 12 to 16 mS/cm/25°C with an anion exchange resin (for example with TMAE). In some embodiments the main amount (bulk) of the rVWF desired species was eluted between about 55 to 60 mS/cm/25°C with an anion exchange resin. In some embodiments, the desired rVWF species starts to elute at a conductivity of between about 18 to 24 mS/cm/25°C with a cation exchange resin. In some embodiments the main amount (bulk) of the rVWF desired species was eluted between about 36 to 42 mS/cm/25°C with a cation exchange resin. In some embodiments, the desired rVWF is mature rVWF (e.g., mat-rVWF). In some embodiments, the main amount (bulk) includes at least about 30%, 40%, 50%, 60%, 70%, 80%, 90% or more of the total amount of the desired species that elutes.

[00317] In some embodiments, further washing steps before the mat-rVWF is eluted from the anion exchange resin are employed. In some embodiments, further washing steps before the mat-rVWF is eluted from the cation exchange resin are employed.

[00318] For their proteolytic activity many proteases need co-factors like bivalent metal ions. Furin and furin protein family members require calcium ions for activity. Therefore, if furin is used to mature pro-rVWF *in vitro* calcium salts are employed. In some embodiments, the calcium salt is a soluble calcium salt. In some embodiments, the calcium salt is calcium chloride (CaCl₂). In some embodiments, the calcium salt is calcium acetate. In some embodiments, other bivalent metal ions are employed, including for example, but not limited to, Be²⁺, Ba²⁺, Mg²⁺, Mn²⁺, Sr²⁺, Zn²⁺, Co²⁺, Ni²⁺, Cd²⁺, and/or Cu²⁺. In some embodiments, a combination of two or more bivalent cations are employed. In some

embodiments, Ca^{2+} and Mg^{2+} are employed in combination. In some embodiments, the calcium salt is a soluble magnesium salt. In some embodiments, the magnesium salt is magnesium chloride (MgCl_2). In some embodiments, the furin protein family formulation for use in the maturation comprises a soluble calcium salt at a concentration of 0.01 to 10 mM. In some embodiments, the furin protein family formulation for use in the maturation comprises a soluble magnesium salt at a concentration of 0.01 to 10 mM. In some embodiments, the furin protein family formulation for use in the maturation comprises CaCl_2 at a concentration of 0.01 to 10 mM. In some embodiments, the furin protein family formulation for use in the maturation comprises MgCl_2 at a concentration of 0.01 to 10 mM. In some embodiments, the furin protein family formulation for use in the maturation comprises CaCl_2 at a concentration of 0.1 to 5 mM. In some embodiments, the furin protein family formulation for use in the maturation comprises MgCl_2 at a concentration of 0.1 to 5 mM. In some embodiments, the furin protein family formulation for use in the maturation comprises CaCl_2 at a concentration of 0.2 to 2 mM. In some embodiments, the furin protein family formulation for use in the maturation comprises MgCl_2 at a concentration of 0.2 to 2 mM. In some embodiments, the furin protein family formulation for use in the maturation comprises furin. In some embodiments, the furin concentration is about 1, about 2, about 3, or about 4 units of recombinant active furin per IU of VWF:Ag (10 μg of pro-rVWF). In some embodiments, the furin concentration is about 2-3 units of recombinant active furin per IU of VWF:Ag (10 μg of pro-rVWF). In some embodiments, the furin concentration is about 1-2 units of recombinant active furin per IU of VWF:Ag (10 μg of pro-rVWF). In some embodiments, the furin concentration is about 2 units of recombinant active furin per IU of VWF:Ag (10 μg of pro-rVWF).

[00319] The incubation time of furin with the immobilized pro-rVWF may vary depending on the system used. Also factors like temperature, buffers etc. influence the efficiency of the maturation process. Generally, the maturation process is terminated in less than 48 hours. In some embodiments, the maturation process can occur in less than 1 minute. In some embodiments, the maturation process can occur in less than 40 hours, 36 hours, 30 hours, 24 hours, 20 hours, 16 hours, 10 hours, 5 hours, 2 hours, 1 hour or less. In some embodiments, the incubation for pro-rVWF maturation is performed for less than 1 minute to 48 hours. In some embodiments, the incubation for pro-rVWF maturation is performed for 10 minutes to 42 hours. In some embodiments, the incubation for pro-rVWF maturation is

performed for 20 minutes to 36 hours. In some embodiments, the incubation for pro-rVWF maturation is performed for 30 minutes to 24 hours. In some embodiments, due to the high specificity of furin, “overactivation” of VWF (further proteolytic degradation) does not occur even after prolonged incubation time.

[00320] In some embodiments, the maturation process depends also on the temperature chosen in the course of the incubation. The optimal enzymatic activity of furin varies with the temperature.

[00321] In some embodiments, the incubation for pro-rVWF maturation is performed at a temperature of 2°C to 40°C. In some embodiments, the incubation for pro-rVWF maturation is performed at a temperature of 4°C to 37°C. In some embodiments, the incubation for pro-rVWF maturation is performed at low temperatures such as 2°C. In some embodiments, the maximum temperatures employed are lower than 50°C, in order to avoid and/or prevent protein degradation. In some embodiments, the maximum temperatures employed are lower than 45°C, in order to avoid and/or prevent protein degradation.

[00322] In some embodiments, the pro-VWF (or pro-rVWF) is converted into mat-VWF (or mat-rVWF) by treatment with furin or a furin family member, as described above. In some embodiments, furin treatment results in at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, at least 99.5% conversion of the pro-rVWF into mat-rVWF and rVWF-PP. In some embodiments after size separation in the presence of the addition of at least one chelating agent and/or increasing the pH to a pH of at least 7, there is less than 5% rVWF-PP, less than 4% rVWF-PP, less than 3% rVWF-PP, less than 2% rVWF-PP, less than 1% rVWF-PP, less than 0.5% rVWF-PP, less than 0.4% rVWF-PP, less than 0.1% rVWF, or less than 0.05% rVWF-PP in the eluate.

Table 2: Exemplary pro-VWF removal (based on furin treatment)

Step	Load, VWF-PP impurity	Eluate, VWF-PP impurity
	% (w/w)	% (w/w)
AEX	~70%	~0.5%
CEX	~0.5%	~0.5%
SEC	~0.5%	~0.5%

I. VWF MULTIMERS

[00323] Assessment of the number and percentage of rVWF multimers can be conducted using methods known in the art, including without limitation methods using electrophoresis and size exclusion chromatography methods to separate VWF multimers by size, for example as discussed by Cumming *et al.*, (J Clin Pathol., 1993 May; 46(5): 470-473, which is hereby incorporated by reference in its entirety for all purposes and in particular for all teachings related to assessment of VWF multimers). Such techniques may further include immunoblotting techniques (such as Western Blot), in which the gel is immunoblotted with a radiolabelled antibody against VWF followed by chemiluminescent detection (see, for example, Wen *et al.*, J. Clin. Lab. Anal., 1993, 7: 317-323, which is hereby incorporated by reference in its entirety for all purposes and in particular for all teachings related to assessment of VWF multimers). Further assays for VWF include VWF:Antigen (VWF:Ag), VWF:Ristocetin Cofactor (VWF:RCof), and VWF:Collagen Binding Activity assay (VWF:CBA), which are often used for diagnosis and classification of Von Willebrand Disease (see, for example, Favaloro *et al.*, Pathology, 1997, 29(4): 341-456; Sadler, JE, Annu Rev Biochem, 1998, 67:395-424; and Turecek *et al.*, Semin Thromb Hemost, 2010, 36:510-521, which are hereby incorporated by reference in their entirety for all purposes and in particular for all teachings related to assays for VWF). In some embodiments, the mat-rVWF obtained using the present methods includes any multimer pattern present in the loading sample of the rVWF. In some embodiments, the mat-rVWF obtained using the present methods includes physiological occurring multimer patterns as well as ultralarge VWF-multimer patterns.

J. VWF ASSAYS

[00324] In primary hemostasis VWF serves as a bridge between platelets and specific components of the extracellular matrix, such as collagen. The biological activity of VWF in this process can be measured by different in vitro assays (Turecek *et al.*, Semin Thromb Hemost, 2010, 36: 510-521).

[00325] The VWF:Ristocetin Cofactor (VWF:RCof) assay is based on the agglutination of fresh or formalin-fixed platelets induced by the antibiotic ristocetin in the presence of VWF. The degree of platelet agglutination depends on the VWF concentration and can be measured by the turbidimetric method, *e.g.*, by use of an aggregometer (Weiss *et al.*, J. Clin. Invest., 1973, 52: 2708-2716; Macfarlane *et al.*, Thromb. Diath. Haemorrh.,

1975, 34: 306-308). As provided herein, the specific ristocetin cofactor activity of the VWF (VWF:RCo) of the present invention is generally described in terms of mU/ μ g of VWF, as measured using in vitro assays.

[00326] In some embodiments, the mat-rVWF purified according to the methods of the present invention has a specific activity of at least about 20, 22.5, 25, 27.5, 30, 32.5, 35, 37.5, 40, 42.5, 45, 47.5, 50, 52.5, 55, 57.5, 60, 62.5, 65, 67.5, 70, 72.5, 75, 77.5, 80, 82.5, 85, 87.5, 90, 92.5, 95, 97.5, 100, 105, 110, 115, 120, 125, 130, 135, 140, 145, 150 or more mU/ μ g. In some embodiments, mat-rVWF used in the methods described herein has a specific activity of from 20 mU/ μ g to 150 mU/ μ g. In some embodiments, the mat-rVWF has a specific activity of from 30 mU/ μ g to 120 mU/ μ g. In some embodiments, the mat-rVWF has a specific activity from 40 mU/ μ g to 90 mU/ μ g. In some embodiments, the mat-rVWF has a specific activity selected from variations 1 to 133 found in Table 3, below.

Table 3. Exemplary embodiments for the specific activity of rVWF found in the compositions and used in the methods provided herein.

(mU/ μ g)		(mU/ μ g)		(mU/ μ g)		(mU/ μ g)	
20	Var. 1	110	Var. 35	40-150	Var. 68	70-120	Var. 101
22.5	Var. 2	115	Var. 36	40-140	Var. 69	70-110	Var. 102
25	Var. 3	120	Var. 37	40-130	Var. 70	70-100	Var. 103
27.5	Var. 4	125	Var. 38	40-120	Var. 71	70-90	Var. 104
30	Var. 5	130	Var. 39	40-110	Var. 72	70-80	Var. 105
32.5	Var. 6	135	Var. 40	40-100	Var. 73	80-150	Var. 106
35	Var. 7	140	Var. 41	40-90	Var. 74	80-140	Var. 107
37.5	Var. 8	145	Var. 42	40-80	Var. 75	80-130	Var. 108
40	Var. 9	150	Var. 43	40-70	Var. 76	80-120	Var. 109
42.5	Var. 10	20-150	Var. 44	40-60	Var. 77	80-110	Var. 110
45	Var. 11	20-140	Var. 45	40-50	Var. 78	80-100	Var. 111
47.5	Var. 12	20-130	Var. 46	50-150	Var. 79	80-90	Var. 112
50	Var. 13	20-120	Var. 47	50-140	Var. 80	90-150	Var. 113
52.5	Var. 14	20-110	Var. 48	50-130	Var. 81	90-140	Var. 114
55	Var. 15	20-100	Var. 49	50-120	Var. 82	90-130	Var. 115
57.5	Var. 16	20-90	Var. 50	50-110	Var. 83	90-120	Var. 116
60	Var. 17	20-80	Var. 51	50-100	Var. 84	90-110	Var. 117
62.5	Var. 18	20-70	Var. 52	50-90	Var. 85	90-100	Var. 118
65	Var. 19	20-60	Var. 53	50-80	Var. 86	100-150	Var. 119
67.5	Var. 20	20-50	Var. 54	50-70	Var. 87	100-140	Var. 120
70	Var. 21	20-40	Var. 55	50-60	Var. 88	100-130	Var. 121
72.5	Var. 22	30-150	Var. 56	60-150	Var. 89	100-120	Var. 122
75	Var. 23	30-140	Var. 57	60-140	Var. 90	100-110	Var. 123
77.5	Var. 24	30-130	Var. 58	60-130	Var. 91	110-150	Var. 124

80	Var. 25	30-120	Var. 59	60-120	Var. 92	110-140	Var. 125
82.5	Var. 26	30-110	Var. 60	60-110	Var. 93	110-130	Var. 126
85	Var. 27	30-100	Var. 61	60-100	Var. 94	110-120	Var. 127
87.5	Var. 28	30-90	Var. 62	60-90	Var. 95	120-150	Var. 128
90	Var. 29	30-80	Var. 63	60-80	Var. 96	120-140	Var. 129
92.5	Var. 30	30-70	Var. 64	60-70	Var. 97	120-130	Var. 130
95	Var. 31	30-60	Var. 65	70-150	Var. 98	130-150	Var. 131
97.5	Var. 32	30-50	Var. 66	70-140	Var. 99	130-140	Var. 132
100	Var. 33	30-40	Var. 67	70-130	Var. 100	140-150	Var. 133
105	Var. 34						

Var. = Variation

[00327] The mat-rVWF of the present invention is highly multimeric comprising about 10 to about 40 subunits. In further embodiments, the multimeric rVWF produced using methods of the present invention comprise about 10-30, 12-28, 14-26, 16-24, 18-22, 20-21 subunits. In some embodiments, the rVWF is present in multimers varying in size from dimers to multimers of over 40 subunits (> 10 million Daltons). The largest multimers provide multiple binding sites that can interact with both platelet receptors and subendothelial matrix sites of injury, and are the most hemostatically active form of VWF. In some embodiments, the mat-rVWF of the present invention comprises ultralarge multimers (ULMs). Generally, high and ultralarge multimers are considered to be hemostatically most effective (*see*, for example, Turecek, P., *Hämostaseologie*, (Vol. 37): Supplement 1, pages S15-S25 (2017)). In some embodiments, the mat-rVWF is between 500 kDa and 20,000 kDa. In some embodiments, any desired multimer pattern can be obtained using the methods described. In some embodiments, when anion exchange and/or cation exchanger methods are employed, the pH, conductivity, and/or counterion concentration of the buffers in the one or more wash step(s) or the gradient buffers can be manipulated to obtain the desired multimer pattern. In some embodiments, then size exclusion chromatography methods are employed, the collection criteria can be employed to obtain the desired multimer pattern. In some embodiments, the described multimer pattern comprises ultralarge multimers. In some embodiments, the ultralarge multimers are at least 10,000 kDa, at least 11,000 kDa, at least 12,000 kDa, at least 13,000 kDa, at least 14,000 kDa, at least 15,000 kDa, at least 16,000 kDa, at least 17,000 kDa, at least 18,000 kDa, at least 19,000 kDa, at least 20,000 kDa. In some embodiments, the ultralarge multimers are between about 10,000 kDa and 20,000 kDa. In some embodiments, the ultralarge multimers are between about 11,000 kDa and 20,000

kDa. In some embodiments, the ultralarge multimers are between about 12,000 kDa and 20,000 kDa. In some embodiments, the ultralarge multimers are between about 13,000 kDa and 20,000 kDa. In some embodiments, the ultralarge multimers are between about 14,000 kDa and 20,000 kDa. In some embodiments, the ultralarge multimers are between about 15,000 kDa and 20,000 kDa. In some embodiments, the ultralarge multimers are between about 16,000 kDa and 20,000 kDa. In some embodiments, the ultralarge multimers are between about 17,000 kDa and 20,000 kDa. In some embodiments, the ultralarge multimers are between about 18,000 kDa and 20,000 kDa. In some embodiments, the ultralarge multimers are between about 19,000 kDa and 20,000 kDa. In some embodiments, the mat-rVWF obtained using the present methods includes any multimer pattern present in the loading sample of the rVWF. In some embodiments, the mat-rVWF obtained using the present methods includes physiological occurring multimer patterns as well as ultra large VWF-multimer patterns.

[00328] In some embodiments, the mat-rVWF composition prepared by the purification method described herein has a distribution of rVWF oligomers characterized in that 95% of the oligomers have between 6 subunits and 20 subunits. In some embodiments, the mat-rVWF composition has a distribution of rVWF oligomers characterized in that 95% of the oligomers have a range of subunits selected from variations 458 to 641 found in 4.

Table 4. Exemplary embodiments for the distribution of rVWF oligomers found in the compositions and used in the methods provided herein.

Subunits		Subunits		Subunits		Subunits	
2-40	Var. 458	6-16	Var. 504	12-20	Var. 550	20-28	Var. 596
2-38	Var. 459	6-14	Var. 505	12-18	Var. 551	20-26	Var. 597
2-36	Var. 460	6-12	Var. 506	12-16	Var. 552	20-24	Var. 598
2-34	Var. 461	6-10	Var. 507	12-14	Var. 553	20-22	Var. 599
2-32	Var. 462	6-8	Var. 508	14-40	Var. 554	22-40	Var. 600
2-30	Var. 463	8-40	Var. 509	14-38	Var. 555	22-38	Var. 601
2-28	Var. 464	8-38	Var. 510	14-36	Var. 556	22-36	Var. 602
2-26	Var. 465	8-36	Var. 511	14-34	Var. 557	22-34	Var. 603
2-24	Var. 466	8-34	Var. 512	14-32	Var. 558	22-32	Var. 604
2-22	Var. 467	8-32	Var. 513	14-30	Var. 559	22-30	Var. 605
2-20	Var. 468	8-30	Var. 514	14-28	Var. 560	22-28	Var. 606
2-18	Var. 469	8-28	Var. 515	14-26	Var. 561	22-26	Var. 607
2-16	Var. 470	8-26	Var. 516	14-24	Var. 562	22-24	Var. 608
2-14	Var. 471	8-24	Var. 517	14-22	Var. 563	24-40	Var. 609
2-12	Var. 472	8-22	Var. 518	14-20	Var. 564	24-38	Var. 610
2-10	Var. 473	8-20	Var. 519	14-18	Var. 565	24-36	Var. 611

2-8	Var. 474	8-18	Var. 520	14-16	Var. 566	24-34	Var. 612
4-40	Var. 475	8-16	Var. 521	16-40	Var. 567	24-32	Var. 613
4-38	Var. 476	8-14	Var. 522	16-38	Var. 568	24-30	Var. 614
4-36	Var. 477	8-12	Var. 523	16-36	Var. 569	24-28	Var. 615
4-34	Var. 478	8-10	Var. 524	16-34	Var. 570	24-26	Var. 616
4-32	Var. 479	10-40	Var. 525	16-32	Var. 571	26-40	Var. 617
4-30	Var. 480	10-38	Var. 526	16-30	Var. 572	26-38	Var. 618
4-28	Var. 481	10-36	Var. 527	16-28	Var. 573	26-36	Var. 619
4-26	Var. 482	10-34	Var. 528	16-26	Var. 574	26-34	Var. 620
4-24	Var. 483	10-32	Var. 529	16-24	Var. 575	26-32	Var. 621
4-22	Var. 484	10-30	Var. 530	16-22	Var. 576	26-30	Var. 622
4-20	Var. 485	10-28	Var. 531	16-20	Var. 577	26-28	Var. 623
4-18	Var. 486	10-26	Var. 532	16-18	Var. 578	28-40	Var. 624
4-16	Var. 487	10-24	Var. 533	18-40	Var. 579	28-38	Var. 625
4-14	Var. 488	10-22	Var. 534	18-38	Var. 580	28-36	Var. 626
4-12	Var. 489	10-20	Var. 535	18-36	Var. 581	28-34	Var. 627
4-10	Var. 490	10-18	Var. 536	18-34	Var. 582	28-32	Var. 628
4-8	Var. 491	10-16	Var. 537	18-32	Var. 583	28-30	Var. 629
6-40	Var. 492	10-14	Var. 538	18-30	Var. 584	30-40	Var. 630
6-38	Var. 493	10-12	Var. 539	18-28	Var. 585	30-38	Var. 631
6-36	Var. 494	12-40	Var. 540	18-26	Var. 586	30-36	Var. 632
6-34	Var. 495	12-38	Var. 541	18-24	Var. 587	30-34	Var. 633
6-32	Var. 496	12-36	Var. 542	18-22	Var. 588	30-32	Var. 634
6-30	Var. 497	12-34	Var. 543	18-20	Var. 589	32-40	Var. 635
6-28	Var. 498	12-32	Var. 544	20-40	Var. 590	32-38	Var. 636
6-26	Var. 499	12-30	Var. 545	20-38	Var. 591	32-36	Var. 637
6-24	Var. 500	12-28	Var. 546	20-36	Var. 592	32-34	Var. 638
6-22	Var. 501	12-26	Var. 547	20-34	Var. 593	34-40	Var. 639
6-20	Var. 502	12-24	Var. 548	20-32	Var. 594	36-38	Var. 640
6-18	Var. 503	12-22	Var. 549	20-30	Var. 595	38-40	Var. 641

Var. = Variation

[00329] In some embodiments, the mat-rVWF composition prepared by the methods provided herein can be characterized according to the percentage of mat-rVWF molecules that are present in a particular higher order mat-rVWF multimer or larger multimer. For example, in one embodiment, at least 20% of mat-rVWF molecules in a mat-rVWF composition used in the methods described herein are present in an oligomeric complex of at least 10 subunits. In another embodiment, at least 20% of mat-rVWF molecules in a mat-rVWF composition used in the methods described herein are present in an oligomeric complex of at least 12 subunits. In yet other embodiments, a mat-rVWF composition used in the methods provided herein has a minimal percentage (*e.g.*, has at least X%) of mat-rVWF molecules present in a particular higher-order mat-rVWF multimer or larger multimer (*e.g.*, a

multimer of at least Y subunits) according to any one of variations 134 to 457 found in Table 5 to Table 7.

Table 5. Exemplary embodiments for the percentage of mat-rVWF molecules that are present in a particular higher order mat-rVWF multimer or larger multimer found in the compositions and used in the methods provided herein.

		Minimal Number of Subunits in rVWF Multimer					
		6	8	10	12	14	16
Minimal Percentage of rVWF Molecules	10%	Var. 134	Var. 152	Var. 170	Var. 188	Var. 206	Var. 224
	15%	Var. 135	Var. 153	Var. 171	Var. 189	Var. 207	Var. 225
	20%	Var. 136	Var. 154	Var. 172	Var. 190	Var. 208	Var. 226
	25%	Var. 137	Var. 155	Var. 173	Var. 191	Var. 209	Var. 227
	30%	Var. 138	Var. 156	Var. 174	Var. 192	Var. 210	Var. 228
	35%	Var. 139	Var. 157	Var. 175	Var. 193	Var. 211	Var. 229
	40%	Var. 140	Var. 158	Var. 176	Var. 194	Var. 212	Var. 230
	45%	Var. 141	Var. 159	Var. 177	Var. 195	Var. 213	Var. 231
	50%	Var. 142	Var. 160	Var. 178	Var. 196	Var. 214	Var. 232
	55%	Var. 143	Var. 161	Var. 179	Var. 197	Var. 215	Var. 233
	60%	Var. 144	Var. 162	Var. 180	Var. 198	Var. 216	Var. 234
	65%	Var. 145	Var. 163	Var. 181	Var. 199	Var. 217	Var. 235
	70%	Var. 146	Var. 164	Var. 182	Var. 200	Var. 218	Var. 236
	75%	Var. 147	Var. 165	Var. 183	Var. 201	Var. 219	Var. 237
	80%	Var. 148	Var. 166	Var. 184	Var. 202	Var. 220	Var. 238
	85%	Var. 149	Var. 167	Var. 185	Var. 203	Var. 221	Var. 239
	90%	Var. 150	Var. 168	Var. 186	Var. 204	Var. 222	Var. 240
	95%	Var. 151	Var. 169	Var. 187	Var. 205	Var. 223	Var. 241

Var. = Variation

Table 6. Exemplary embodiments for the percentage of mat-rVWF molecules that are present in a particular higher order mat-rVWF multimer or larger multimer found in the compositions and used in the methods provided herein.

		Minimal Number of Subunits in rVWF Multimer					
		18	20	22	24	26	28
Minimal Percentage of	10%	Var. 242	Var. 260	Var. 278	Var. 296	Var. 314	Var. 332
	15%	Var. 243	Var. 261	Var. 279	Var. 297	Var. 315	Var. 333
	20%	Var. 244	Var. 262	Var. 280	Var. 298	Var. 316	Var. 334
	25%	Var. 245	Var. 263	Var. 281	Var. 299	Var. 317	Var. 335
	30%	Var. 246	Var. 264	Var. 282	Var. 300	Var. 318	Var. 336
	35%	Var. 247	Var. 265	Var. 283	Var. 301	Var. 319	Var. 337
	40%	Var. 248	Var. 266	Var. 284	Var. 302	Var. 320	Var. 338
	45%	Var. 249	Var. 267	Var. 285	Var. 303	Var. 321	Var. 339
	50%	Var. 250	Var. 268	Var. 286	Var. 304	Var. 322	Var. 340

	55%	Var. 251	Var. 269	Var. 287	Var. 305	Var. 323	Var. 341
	60%	Var. 252	Var. 270	Var. 288	Var. 306	Var. 324	Var. 342
	65%	Var. 253	Var. 271	Var. 289	Var. 307	Var. 325	Var. 343
	70%	Var. 254	Var. 272	Var. 290	Var. 308	Var. 326	Var. 344
	75%	Var. 255	Var. 273	Var. 291	Var. 309	Var. 327	Var. 345
	80%	Var. 256	Var. 274	Var. 292	Var. 310	Var. 328	Var. 346
	85%	Var. 257	Var. 275	Var. 293	Var. 311	Var. 329	Var. 347
	90%	Var. 258	Var. 276	Var. 294	Var. 312	Var. 330	Var. 348
	95%	Var. 259	Var. 277	Var. 295	Var. 313	Var. 331	Var. 349

Var. = Variation

Table 7. Exemplary embodiments for the percentage of mat-rVWF molecules that are present in a particular higher order mat-rVWF multimer or larger multimer found in the compositions and used in the methods provided herein.

		Minimal Number of Subunits in rVWF Multimer					
		30	32	34	36	38	40
Minimal Percentage of rVWF Molecules	10%	Var. 350	Var. 368	Var. 386	Var. 404	Var. 422	Var. 440
	15%	Var. 351	Var. 369	Var. 387	Var. 405	Var. 423	Var. 441
	20%	Var. 352	Var. 370	Var. 388	Var. 406	Var. 424	Var. 442
	25%	Var. 353	Var. 371	Var. 389	Var. 407	Var. 425	Var. 443
	30%	Var. 354	Var. 372	Var. 390	Var. 408	Var. 426	Var. 444
	35%	Var. 355	Var. 373	Var. 391	Var. 409	Var. 427	Var. 445
	40%	Var. 356	Var. 374	Var. 392	Var. 410	Var. 428	Var. 446
	45%	Var. 357	Var. 375	Var. 393	Var. 411	Var. 429	Var. 447
	50%	Var. 358	Var. 376	Var. 394	Var. 412	Var. 430	Var. 448
	55%	Var. 359	Var. 377	Var. 395	Var. 413	Var. 431	Var. 449
	60%	Var. 360	Var. 378	Var. 396	Var. 414	Var. 432	Var. 450
	65%	Var. 361	Var. 379	Var. 397	Var. 415	Var. 433	Var. 451
	70%	Var. 362	Var. 380	Var. 398	Var. 416	Var. 434	Var. 452
	75%	Var. 363	Var. 381	Var. 399	Var. 417	Var. 435	Var. 453
	80%	Var. 364	Var. 382	Var. 400	Var. 418	Var. 436	Var. 454
	85%	Var. 365	Var. 383	Var. 401	Var. 419	Var. 437	Var. 455
	90%	Var. 366	Var. 384	Var. 402	Var. 420	Var. 438	Var. 456
	95%	Var. 367	Var. 385	Var. 403	Var. 421	Var. 439	Var. 457

Var. = Variation

[00330] In accordance with the above, the mat-rVWF comprises a significant percentage of high molecular weight (HMW) mat-rVWF multimers. In further embodiments, the HMW rVWF multimer composition comprises at least 10% - 80% mat-rVWF decamers or higher order multimers. In further embodiments, the composition comprises about 10-95%, 20-90%, 30-85%, 40-80%, 50-75%, 60-70% decamers or higher order multimers. In

further embodiments, the HMW mat-rVWF multimer composition comprises at least about 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90% decamers or higher order multimers.

[00331] Assessment of the number and percentage of mat-rVWF multimers can be conducted using methods known in the art, including without limitation methods using electrophoresis and size exclusion chromatography methods to separate mat-rVWF multimers by size, for example as discussed by Cumming et al, (J Clin Pathol. 1993 May; 46(5): 470–473, which is hereby incorporated by reference in its entirety for all purposes and in particular for all teachings related to assessment of mat-rVWF multimers). Such techniques may further include immunoblotting techniques (such as Western Blot), in which the gel is immunoblotted with a radiolabelled antibody against VWF followed by chemiluminescent detection (see for example Wen *et al.*, (1993), J. Clin. Lab. Anal., 7: 317–323, which is hereby incorporated by reference in its entirety for all purposes and in particular for all teachings related to assessment of mat-rVWF multimers). Further assays for VWF include VWF:Antigen (VWF:Ag), VWF:Ristocetin Cofactor (VWF:RCof), and VWF:Collagen Binding Activity assay (VWF:CBA), which are often used for diagnosis and classification of Von Willebrand Disease. (see for example Favaloro *et al.*, Pathology, 1997, 29(4): 341–456, which is hereby incorporated by reference in its entirety for all purposes and in particular for all teachings related to assays for VWF).

[00332] In some embodiments, the ratio of rFVIII procoagulant activity (IU rFVIII:C) to rVWF Ristocetin cofactor activity (IU rVWF:RCo) for the mat-rVWF prepared according to the methods of the present invention is between 3:1 and 1:5. In further embodiments, the ratio is between 2:1 and 1:4. In still further embodiments, the ratio is between 5:2 and 1:4. In further embodiments, the ratio is between 3:2 and 1:3. In still further embodiments, the ratio is about 1:1, 1:2, 1:3, 1:4, 1:5, 2:1, 2:3, 2:4, 2:5, 3:1, 3:2, 3:4, or 3:5. In further embodiments, the ratio is between 1:1 and 1:2. In yet further embodiments, the ratio is 1.1:1, 1.2:1, 1.3:1, 1.4:1, 1.5:1, 1.6:1, 1.7:1, 1.8:1, 1.9:1, or 2:1. In certain embodiments, the ratio of rFVIII procoagulant activity (IU rFVIII:C) to rVWF Ristocetin cofactor activity (IU rVWF:RCo) in a composition useful for a method described herein is selected from variations 1988 to 2140 found in Table 8.

Table 8. Exemplary embodiments for the ratio of rFVIII procoagulant activity (IU rFVIII:C) to rVWF Ristocetin cofactor activity (IU rVWF:RCo) in compositions and used in methods provided herein.

(IU rFVIII:C) to (IU rVWF:RC ₀)		(IU rFVIII:C) to (IU rVWF:RC ₀)		(IU rFVIII:C) to (IU rVWF:RC ₀)		(IU rFVIII:C) to (IU rVWF:RC ₀)	
4:1	Var. 1988	3:1-3:5	Var. 2027	4:3-1:4	Var. 2065	4:5-2:3	Var. 2103
3:1	Var. 1989	3:1-2:3	Var. 2028	4:3-1:3	Var. 2066	4:5-3:4	Var. 2104
2:1	Var. 1990	3:1-3:4	Var. 2029	4:3-2:5	Var. 2067	3:4-1:6	Var. 2105
3:2	Var. 1991	3:1-4:5	Var. 2030	4:3-1:2	Var. 2068	3:4-1:5	Var. 2106
4:3	Var. 1992	3:1-5:6	Var. 2031	4:3-3:5	Var. 2069	3:4-1:4	Var. 2107
1:1	Var. 1993	3:1-1:1	Var. 2032	4:3-2:3	Var. 2070	3:4-1:3	Var. 2108
5:6	Var. 1994	3:1-4:3	Var. 2033	4:3-3:4	Var. 2071	3:4-2:5	Var. 2109
4:5	Var. 1995	3:1-3:2	Var. 2034	4:3-4:5	Var. 2072	3:4-1:2	Var. 2110
3:4	Var. 1996	3:1-2:1	Var. 2035	4:3-5:6	Var. 2073	3:4-3:5	Var. 2111
2:3	Var. 1997	2:1-1:6	Var. 2036	4:3-1:1	Var. 2074	3:4-2:3	Var. 2112
3:5	Var. 1998	2:1-1:5	Var. 2037	1:1-1:6	Var. 2075	2:3-1:6	Var. 2113
1:2	Var. 1999	2:1-1:4	Var. 2038	1:1-1:5	Var. 2076	2:3-1:5	Var. 2114
2:5	Var. 2000	2:1-1:3	Var. 2039	1:1-1:4	Var. 2077	2:3-1:4	Var. 2115
1:3	Var. 2001	2:1-2:5	Var. 2040	1:1-1:3	Var. 2078	2:3-1:3	Var. 2116
1:4	Var. 2002	2:1-1:2	Var. 2041	1:1-2:5	Var. 2079	2:3-2:5	Var. 2117
1:5	Var. 2003	2:1-3:5	Var. 2042	1:1-1:2	Var. 2080	2:3-1:2	Var. 2118
1:6	Var. 2004	2:1-2:3	Var. 2043	1:1-3:5	Var. 2081	2:3-3:5	Var. 2119
4:1-1:6	Var. 2005	2:1-3:4	Var. 2044	1:1-2:3	Var. 2082	3:5-1:6	Var. 2120
4:1-1:5	Var. 2006	2:1-4:5	Var. 2045	1:1-3:4	Var. 2083	3:5-1:5	Var. 2121
4:1-1:4	Var. 2007	2:1-5:6	Var. 2046	1:1-4:5	Var. 2084	3:5-1:4	Var. 2122
4:1-1:3	Var. 2008	2:1-1:1	Var. 2047	1:1-5:6	Var. 2085	3:5-1:3	Var. 2123
4:1-2:5	Var. 2009	2:1-4:3	Var. 2048	5:6-1:6	Var. 2086	3:5-2:5	Var. 2124
4:1-1:2	Var. 2010	2:1-3:2	Var. 2049	5:6-1:5	Var. 2087	3:5-1:2	Var. 2125
4:1-3:5	Var. 2011	3:2-1:6	Var. 2050	5:6-1:4	Var. 2088	1:2-1:6	Var. 2126
4:1-2:3	Var. 2012	3:2-1:5	Var. 2051	5:6-1:3	Var. 2089	1:2-1:5	Var. 2127
4:1-3:4	Var. 2013	3:2-1:4	Var. 2052	5:6-2:5	Var. 2090	1:2-1:4	Var. 2128
4:1-4:5	Var. 2014	3:2-1:3	Var. 2053	5:6-1:2	Var. 2091	1:2-1:3	Var. 2129

4:1-5:6	Var. 2015	3:2-2:5	Var. 2054	5:6-3:5	Var. 2092	1:2-2:5	Var. 2130
4:1-1:1	Var. 2016	3:2-1:2	Var. 2055	5:6-2:3	Var. 2093	2:5-1:6	Var. 2131
4:1-4:3	Var. 2017	3:2-3:5	Var. 2056	5:6-3:4	Var. 2094	2:5-1:5	Var. 2132
4:1-3:2	Var. 2018	3:2-2:3	Var. 2057	5:6-4:5	Var. 2095	2:5-1:4	Var. 2133
4:1-2:1	Var. 2019	3:2-3:4	Var. 2058	4:5-1:6	Var. 2096	2:5-1:3	Var. 2134
4:1-3:1	Var. 2020	3:2-4:5	Var. 2059	4:5-1:5	Var. 2097	1:3-1:6	Var. 2135
3:1-1:6	Var. 2021	3:2-5:6	Var. 2060	4:5-1:4	Var. 2098	1:3-1:5	Var. 2136
3:1-1:5	Var. 2022	3:2-1:1	Var. 2061	4:5-1:3	Var. 2099	1:3-1:4	Var. 2137
3:1-1:4	Var. 2023	3:2-4:3	Var. 2062	4:5-2:5	Var. 2100	1:4-1:6	Var. 2138
3:1-1:3	Var. 2024	4:3-1:6	Var. 2063	4:5-1:2	Var. 2101	1:4-1:5	Var. 2139
3:1-2:5	Var. 2025	4:3-1:5	Var. 2064	4:5-3:5	Var. 2102	1:5-1:6	Var. 2140
3:1-1:2	Var. 2026						

Var. = Variation

[00333] In further embodiments, higher order mat-rVWF multimers of the invention are stable for about 1 to about 90 hours post-administration. In still further embodiments, the higher order mat-rVWF multimers are stable for about 5-80, 10-70, 15-60, 20-50, 25-40, 30-35 hours post-administration. In yet further embodiments, the higher order mat-rVWF multimers are stable for at least 3, 6, 12, 18, 24, 36, 48, 72 hours post-administration. In certain embodiments the stability of the mat-rVWF multimers is assessed *in vitro*.

[00334] In one embodiment, higher order mat-rVWF multimers used in the compositions and methods provided herein have a half-life of at least 12 hour post administration. In another embodiment, the higher order mat-rVWF multimers have a half-life of at least 24 hour post administration. In yet other embodiments, the higher order mat-rVWF multimers have a half-life selected from variations 642 to 1045 found in Table 9.

Table 9. Exemplary embodiments for the half-life of higher order mat-rVWF multimers found in the compositions prepared by the methods provided herein.

Hours		Hours		Hours		Hours	
at least 1	Var. 642	4-22	Var. 743	14-78	Var. 844	24-30	Var. 945
at least 2	Var. 643	4-20	Var. 744	14-72	Var. 845	24-27	Var. 946
at least 3	Var. 644	4-18	Var. 745	14-66	Var. 846	27-90	Var. 947
at least 4	Var. 645	4-16	Var. 746	14-60	Var. 847	27-84	Var. 948

at least 5	Var. 646	4-14	Var. 747	14-54	Var. 848	27-78	Var. 949
at least 6	Var. 647	4-12	Var. 748	14-48	Var. 849	27-72	Var. 950
at least 7	Var. 648	4-10	Var. 749	14-45	Var. 850	27-66	Var. 951
at least 8	Var. 649	4-8	Var. 750	14-42	Var. 851	27-60	Var. 952
at least 9	Var. 650	4-6	Var. 751	14-39	Var. 852	27-54	Var. 953
at least 10	Var. 651	6-90	Var. 752	14-36	Var. 853	27-48	Var. 954
at least 11	Var. 652	6-84	Var. 753	14-33	Var. 854	30-90	Var. 955
at least 12	Var. 653	6-78	Var. 754	14-30	Var. 855	30-84	Var. 956
at least 14	Var. 654	6-72	Var. 755	14-27	Var. 856	30-78	Var. 957
at least 16	Var. 655	6-66	Var. 756	14-24	Var. 857	30-72	Var. 958
at least 18	Var. 656	6-60	Var. 757	14-22	Var. 858	30-66	Var. 959
at least 20	Var. 657	6-54	Var. 758	14-20	Var. 859	30-60	Var. 960
at least 22	Var. 658	6-48	Var. 759	14-18	Var. 860	30-54	Var. 961
at least 24	Var. 659	6-45	Var. 760	14-16	Var. 861	30-48	Var. 962
at least 27	Var. 660	6-42	Var. 761	16-90	Var. 862	30-45	Var. 963
at least 30	Var. 661	6-39	Var. 762	16-84	Var. 863	30-42	Var. 964
at least 33	Var. 662	6-36	Var. 763	16-78	Var. 864	30-39	Var. 965
at least 36	Var. 663	6-33	Var. 764	16-72	Var. 865	30-36	Var. 966
at least 39	Var. 664	6-30	Var. 765	16-66	Var. 866	30-33	Var. 967
at least 42	Var. 665	6-27	Var. 766	16-60	Var. 867	33-90	Var. 968
at least 45	Var. 666	6-24	Var. 767	16-54	Var. 868	33-84	Var. 969
at least 48	Var. 667	6-22	Var. 768	16-48	Var. 869	33-78	Var. 970
at least 54	Var. 668	6-20	Var. 769	16-45	Var. 870	33-72	Var. 971
at least 60	Var. 669	6-18	Var. 770	16-42	Var. 871	33-66	Var. 972
at least 66	Var. 670	6-16	Var. 771	16-39	Var. 872	33-60	Var. 973
at least 72	Var. 671	6-14	Var. 772	16-36	Var. 873	33-54	Var. 974
at least 78	Var. 672	6-12	Var. 773	16-33	Var. 874	33-48	Var. 975
at least 84	Var. 673	6-10	Var. 774	16-30	Var. 875	33-45	Var. 976
at least 90	Var. 674	6-8	Var. 775	16-27	Var. 876	33-42	Var. 977
2-90	Var. 675	8-90	Var. 776	16-24	Var. 877	33-29	Var. 978
2-84	Var. 676	8-84	Var. 777	16-22	Var. 878	33-36	Var. 979
2-78	Var. 677	8-78	Var. 778	16-20	Var. 879	36-90	Var. 980
2-72	Var. 678	8-72	Var. 779	16-18	Var. 880	36-84	Var. 981
2-66	Var. 679	8-66	Var. 780	18-90	Var. 881	36-78	Var. 982
2-60	Var. 680	8-60	Var. 781	18-84	Var. 882	36-72	Var. 983
2-54	Var. 681	8-54	Var. 782	18-78	Var. 883	36-66	Var. 984
2-48	Var. 682	8-48	Var. 783	18-72	Var. 884	36-60	Var. 985
2-45	Var. 683	8-45	Var. 784	18-66	Var. 885	36-54	Var. 986
2-42	Var. 684	8-42	Var. 785	18-60	Var. 886	36-48	Var. 987
2-39	Var. 685	8-39	Var. 786	18-54	Var. 887	36-45	Var. 988
2-36	Var. 686	8-36	Var. 787	18-48	Var. 888	36-42	Var. 989
2-33	Var. 687	8-33	Var. 788	18-45	Var. 889	36-39	Var. 990
2-30	Var. 688	8-30	Var. 789	18-42	Var. 890	39-90	Var. 991
2-27	Var. 689	8-27	Var. 790	18-39	Var. 891	39-84	Var. 992
2-24	Var. 690	8-24	Var. 791	18-36	Var. 892	39-78	Var. 993
2-22	Var. 691	8-22	Var. 792	18-33	Var. 893	39-72	Var. 994

2-20	Var. 692	8-20	Var. 793	18-30	Var. 894	39-66	Var. 995
2-18	Var. 693	8-18	Var. 794	18-27	Var. 895	39-60	Var. 996
2-16	Var. 694	8-16	Var. 795	18-24	Var. 896	39-54	Var. 997
2-14	Var. 695	8-14	Var. 796	18-22	Var. 897	39-48	Var. 998
2-12	Var. 696	8-12	Var. 797	18-20	Var. 898	39-45	Var. 999
2-10	Var. 697	8-10	Var. 798	20-90	Var. 899	39-42	Var. 1000
2-8	Var. 698	10-90	Var. 799	20-84	Var. 900	42-90	Var. 1001
2-6	Var. 699	10-84	Var. 800	20-78	Var. 901	42-84	Var. 1002
2-4	Var. 700	10-78	Var. 801	20-72	Var. 902	42-78	Var. 1003
3-90	Var. 701	10-72	Var. 802	20-66	Var. 903	42-72	Var. 1004
3-84	Var. 702	10-66	Var. 803	20-60	Var. 904	42-66	Var. 1005
3-78	Var. 703	10-60	Var. 804	20-54	Var. 905	42-60	Var. 1006
3-72	Var. 704	10-54	Var. 805	20-48	Var. 906	42-54	Var. 1007
3-66	Var. 705	10-48	Var. 806	20-45	Var. 907	42-48	Var. 1008
3-60	Var. 706	10-45	Var. 807	20-42	Var. 908	42-45	Var. 1009
3-54	Var. 707	10-42	Var. 808	20-39	Var. 909	45-90	Var. 1010
3-48	Var. 708	10-39	Var. 809	20-36	Var. 910	45-84	Var. 1011
3-45	Var. 709	10-36	Var. 810	20-33	Var. 911	45-78	Var. 1012
3-42	Var. 710	10-33	Var. 811	20-30	Var. 912	45-72	Var. 1013
3-39	Var. 711	10-30	Var. 812	20-27	Var. 913	45-66	Var. 1014
3-36	Var. 712	10-27	Var. 813	20-24	Var. 914	45-60	Var. 1015
3-33	Var. 713	10-24	Var. 814	20-22	Var. 915	45-54	Var. 1016
3-30	Var. 714	10-22	Var. 815	22-90	Var. 916	45-48	Var. 1017
3-27	Var. 715	10-20	Var. 816	22-84	Var. 917	48-90	Var. 1018
3-24	Var. 716	10-18	Var. 817	22-78	Var. 918	48-84	Var. 1019
3-22	Var. 717	10-16	Var. 818	22-72	Var. 919	48-78	Var. 1020
3-20	Var. 718	10-14	Var. 819	22-66	Var. 920	48-72	Var. 1021
3-18	Var. 719	10-12	Var. 820	22-60	Var. 921	48-66	Var. 1022
3-16	Var. 720	12-90	Var. 821	22-54	Var. 922	48-60	Var. 1023
3-14	Var. 721	12-84	Var. 822	22-48	Var. 923	48-54	Var. 1024
3-12	Var. 722	12-78	Var. 823	22-45	Var. 924	54-90	Var. 1025
3-10	Var. 723	12-72	Var. 824	22-42	Var. 925	54-84	Var. 1026
3-8	Var. 724	12-66	Var. 825	22-39	Var. 926	54-78	Var. 1027
3-6	Var. 725	12-60	Var. 826	22-36	Var. 927	54-72	Var. 1028
3-4	Var. 726	12-54	Var. 827	22-33	Var. 928	54-66	Var. 1029
4-90	Var. 727	12-48	Var. 828	22-30	Var. 929	54-60	Var. 1030
4-84	Var. 728	12-45	Var. 829	22-27	Var. 930	60-90	Var. 1031
4-78	Var. 729	12-42	Var. 830	22-24	Var. 931	60-84	Var. 1032
4-72	Var. 730	12-39	Var. 831	24-90	Var. 932	60-78	Var. 1033
4-66	Var. 731	12-36	Var. 832	24-84	Var. 933	60-72	Var. 1034
4-60	Var. 732	12-33	Var. 833	24-78	Var. 934	60-66	Var. 1035
4-54	Var. 733	12-30	Var. 834	24-72	Var. 935	66-90	Var. 1036
4-48	Var. 734	12-27	Var. 835	24-66	Var. 936	66-84	Var. 1037
4-45	Var. 735	12-24	Var. 836	24-60	Var. 937	66-78	Var. 1038
4-42	Var. 736	12-22	Var. 837	24-54	Var. 938	66-72	Var. 1039
4-39	Var. 737	12-20	Var. 838	24-48	Var. 939	72-90	Var. 1040

4-36	Var. 738	12-18	Var. 839	24-45	Var. 940	72-84	Var. 1041
4-33	Var. 739	12-16	Var. 840	24-42	Var. 941	72-78	Var. 1042
4-30	Var. 740	12-14	Var. 841	24-39	Var. 942	78-90	Var. 1043
4-27	Var. 741	14-90	Var. 842	24-36	Var. 943	78-84	Var. 1044
4-24	Var. 742	14-84	Var. 843	24-33	Var. 944	84-90	Var. 1045

Var. = Variation

[00335] In some embodiments, the pro-VWF and/or purified mat-rVWF purified in accordance with the present invention is not modified with any conjugation, post-translation or covalent modifications. In particular embodiments, the pro-VWF and/or purified mat-rVWF of the present invention is not modified with a water soluble polymer, including without limitation, a polyethylene glycol (PEG), a polypropylene glycol, a polyoxyalkylene, a polysialic acid, hydroxyl ethyl starch, a poly-carbohydrate moiety, and the like.

[00336] In some embodiments, the pro-VWF and/or purified mat-rVWF purified in accordance with the present invention is modified through conjugation, post-translation modification, or covalent modification, including modifications of the N- or C- terminal residues as well as modifications of selected side chains, for example, at free sulfhydryl-groups, primary amines, and hydroxyl-groups. In one embodiment, a water soluble polymer is linked to the protein (directly or via a linker) by a lysine group or other primary amine. In some embodiments, the pro-VWF and/or purified mat-rVWF of the present invention may be modified by conjugation of a water soluble polymer, including without limitation, a polyethylene glycol (PEG), a polypropylene glycol, a polyoxyalkylene, a polysialic acid, hydroxyl ethyl starch, a poly-carbohydrate moiety, and the like.

[00337] Water soluble polymers that may be used to modify the pro-VWF and/or purified mat-rVWF include linear and branched structures. The conjugated polymers may be attached directly to the coagulation proteins of the invention, or alternatively may be attached through a linking moiety. Non-limiting examples of protein conjugation with water soluble polymers can be found in U.S. Patent Nos. 4,640,835; 4,496,689; 4,301,144; 4,670,417; 4,791,192, and 4,179,337, as well as in Abuchowski and Davis "Enzymes as Drugs," Holcenberg and Roberts, Eds., pp. 367 383, John Wiley and Sons, New York (1981), and Hermanson G., Bioconjugate Techniques 2nd Ed., Academic Press, Inc. 2008.

[00338] Protein conjugation may be performed by a number of well-known techniques in the art, for example, see Hermanson G., Bioconjugate Techniques 2nd Ed., Academic Press, Inc. 2008. Examples include linkage through the peptide bond between a carboxyl

group on one of either the coagulation protein or water-soluble polymer moiety and an amine group of the other, or an ester linkage between a carboxyl group of one and a hydroxyl group of the other. Another linkage by which a coagulation protein of the invention could be conjugated to a water-soluble polymer compound is via a Schiff base, between a free amino group on the polymer moiety being reacted with an aldehyde group formed at the non-reducing end of the polymer by periodate oxidation (Jennings and Lugowski, J. Immunol. 1981; 127:1011-8; Femandes and Gregonradis, Biochim Biophys Acta. 1997; 1341; 26-34). The generated Schiff Base can be stabilized by specific reduction with NaCNBH_3 to form a secondary amine. An alternative approach is the generation of terminal free amino groups on the polymer by reductive amination with NH_4Cl after prior oxidation. Bifunctional reagents can be used for linking two amino or two hydroxyl groups. For example, a polymer containing an amino group can be coupled to an amino group of the coagulation protein with reagents like BS3 (Bis(sulfosuccinimidyl)suberate/Pierce, Rockford, Ill.). In addition, heterobifunctional cross linking reagents like Sulfo-EMCS (N- ϵ -Maleimidocaproyloxy) sulfosuccinimide ester/Pierce) can be used for instance to link amine and thiol groups. In other embodiments, an aldehyde reactive group, such as PEG alkoxide plus diethyl acetal of bromoacetaldehyde; PEG plus DMSO and acetic anhydride, and PEG chloride plus the phenoxide of 4-hydroxybenzaldehyde, succinimidyl active esters, activated dithiocarbonate PEG, 2,4,5-trichlorophenylchloroformate and P-nitrophenylchloroformate activated PEG, may be used in the conjugation of a coagulation protein.

[00339] Another method for measuring the biological activity of VWF is the collagen binding assay, which is based on ELISA technology (Brown and Bosak, Thromb. Res., 1986, 43:303-311; Favaloro, Thromb. Haemost., 2000, 83 127-135). A microtiter plate is coated with type I or III collagen. Then the VWF is bound to the collagen surface and subsequently detected with an enzyme-labeled polyclonal antibody. The last step is a substrate reaction, which can be photometrically monitored with an ELISA reader.

[00340] Immunological assays of von Willebrand factors (VWF:Ag) are immunoassays that measure the concentration of the VWF protein in plasma. They give no indication as to VWF function. A number of methods exist for measuring VWF:Ag and these include both enzyme-linked immunosorbent assay (ELISA) or automated latex immunoassays (LIA.) Many laboratories now use a fully automated latex immunoassay.

Historically laboratories used a variety of techniques including Laurell electroimmunoassay 'Laurell Rockets' but these are rarely used in most labs today.

K. VWF FORMULATIONS/ADMINISTRATION

[00341] The present method also provides for preparation of formulations from the VWF obtained by the purification methods provided herein. In some embodiments, the high purity mat-rVWF composition is used for the production of a pharmaceutical composition. In some embodiments, the mat-rVWF can be formulated into a lyophilized formulation.

[00342] In some embodiments, the formulations comprising a VWF polypeptide of the invention are lyophilized after purification and prior to administration to a subject. Lyophilization is carried out using techniques common in the art and should be optimized for the composition being developed (Tang *et al.*, Pharm Res. 21:191-200, (2004) and Chang *et al.*, Pharm Res. 13:243-9 (1996)).

[00343] A lyophilization cycle is, in one aspect, composed of three steps: freezing, primary drying, and secondary drying (A. P. Mackenzie, Phil Trans R Soc London, Ser B, Biol 278:167 (1977)). In the freezing step, the solution is cooled to initiate ice formation. Furthermore, this step induces the crystallization of the bulking agent. The ice sublimates in the primary drying stage, which is conducted by reducing chamber pressure below the vapor pressure of the ice, using a vacuum and introducing heat to promote sublimation. Finally, adsorbed or bound water is removed at the secondary drying stage under reduced chamber pressure and at an elevated shelf temperature. The process produces a material known as a lyophilized cake. Thereafter the cake can be reconstituted with either sterile water or suitable diluent for injection.

[00344] The lyophilization cycle not only determines the final physical state of excipients but also affects other parameters such as reconstitution time, appearance, stability and final moisture content. The composition structure in the frozen state proceeds through several transitions (*e.g.*, glass transitions, wettings, and crystallizations) that occur at specific temperatures and the structure may be used to understand and optimize the lyophilization process. The glass transition temperature (T_g and/or T_g') can provide information about the physical state of a solute and can be determined by differential scanning calorimetry (DSC). T_g and T_g' are an important parameter that must be taken into account when designing the

lyophilization cycle. For example, Tg' is important for primary drying. Furthermore, in the dried state, the glass transition temperature provides information on the storage temperature of the final product.

i. Pharmaceutical formulations and excipients in general

[00345] Excipients are additives that either impart or enhance the stability and delivery of a drug product (*e.g.*, protein). Regardless of the reason for their inclusion, excipients are an integral component of a formulation and therefore need to be safe and well tolerated by patients. For protein drugs, the choice of excipients is particularly important because they can affect both efficacy and immunogenicity of the drug. Hence, protein formulations need to be developed with appropriate selection of excipients that afford suitable stability, safety, and marketability.

[00346] A lyophilized formulation is, in one aspect, at least comprised of one or more of a buffer, a bulking agent, and a stabilizer. In this aspect, the utility of a surfactant is evaluated and selected in cases where aggregation during the lyophilization step or during reconstitution becomes an issue. An appropriate buffering agent is included to maintain the formulation within stable zones of pH during lyophilization. A comparison of the excipient components contemplated for liquid and lyophilized protein formulations is provided in Table 10.

Table 10: Excipient components of lyophilized protein formulations

Excipient component	Function in lyophilized formulation
Buffer	• Maintain pH of formulation during lyophilization and upon reconstitution
Tonicity agent/ stabilizer	• Stabilizers include cryo and lyoprotectants • Examples include Polyols, sugars and polymers • Cryoprotectants protect proteins from freezing stresses • Lyoprotectants stabilize proteins in the freeze-dried state
Bulking agent	• Used to enhance product elegance and to prevent breakout • Provides structural strength to the lyo cake • Examples include mannitol and glycine
Surfactant	• Employed if aggregation during the lyophilization process is an issue • May serve to reduce reconstitution times • Examples include polysorbate 20 and 80
Anti-oxidant	• Usually not employed, molecular reactions in the lyo cake are greatly retarded
Metal ion/chelating agent	• May be included if a specific metal ion is included only as a co-factor or where the metal is required for protease activity • Chelating agents are generally not needed in lyo formulations
Preservative	• For multi-dose formulations only • Provides protection against microbial growth in formulation • Is usually included in the reconstitution diluent (e.g. WFI)

[00347] The principal challenge in developing formulations for proteins is stabilizing the product against the stresses of manufacturing, shipping and storage. The role of formulation excipients is to provide stabilization against these stresses. Excipients are also be employed to reduce viscosity of high concentration protein formulations in order to enable their delivery and enhance patient convenience. In general, excipients can be classified on the basis of the mechanisms by which they stabilize proteins against various chemical and physical stresses. Some excipients are used to alleviate the effects of a specific stress or to

regulate a particular susceptibility of a specific protein. Other excipients have more general effects on the physical and covalent stabilities of proteins. The excipients described herein are organized either by their chemical type or their functional role in formulations. Brief descriptions of the modes of stabilization are provided when discussing each excipient type.

[00348] Given the teachings and guidance provided herein, those skilled in the art will know what amount or range of excipient can be included in any particular formulation to achieve a biopharmaceutical formulation of the invention that promotes retention in stability of the biopharmaceutical (*e.g.*, a protein). For example, the amount and type of a salt to be included in a biopharmaceutical formulation of the invention is selected based on the desired osmolality (*e.g.*, isotonic, hypotonic or hypertonic) of the final solution as well as the amounts and osmolality of other components to be included in the formulation.

[00349] By way of example, inclusion of about 5% sorbitol can achieve isotonicity while about 9% of a sucrose excipient is needed to achieve isotonicity. Selection of the amount or range of concentrations of one or more excipients that can be included within a biopharmaceutical formulation of the invention has been exemplified above by reference to salts, polyols and sugars. However, those skilled in the art will understand that the considerations described herein and further exemplified by reference to specific excipients are equally applicable to all types and combinations of excipients including, for example, salts, amino acids, other tonicity agents, surfactants, stabilizers, bulking agents, cryoprotectants, lyoprotectants, anti-oxidants, metal ions, chelating agents and/or preservatives.

[00350] Further, where a particular excipient is reported in molar concentration, those skilled in the art will recognize that the equivalent percent (%) w/v (*e.g.*, (grams of substance in a solution sample/mL of solution) X 100%) of solution is also contemplated.

[00351] Of course, a person having ordinary skill in the art would recognize that the concentrations of the excipients described herein share an interdependency within a particular formulation. By way of example, the concentration of a bulking agent may be lowered where, *e.g.*, there is a high protein concentration or where, *e.g.*, there is a high stabilizing agent concentration. In addition, a person having ordinary skill in the art would recognize that, in order to maintain the isotonicity of a particular formulation in which there is no bulking agent, the concentration of a stabilizing agent would be adjusted accordingly (*e.g.*, a “tonicifying” amount of stabilizer would be used). Common excipients are known in the art

and can be found in Powell *et al.*, Compendium of Excipients for Parenteral Formulations (1998), PDA J. Pharm. Sci. Technology, 52:238-311.

ii. Pharmaceutical Buffers and buffering agents

[00352] The stability of a pharmacologically active protein formulation is usually observed to be maximal in a narrow pH range. This pH range of optimal stability needs to be identified early during pre-formulation studies. Several approaches, such as accelerated stability studies and calorimetric screening studies, are useful in this endeavor (Remmele R.L. Jr., *et al.*, Biochemistry, 38(16): 5241-7 (1999)). Once a formulation is finalized, the protein must be manufactured and maintained throughout its shelf-life. Hence, buffering agents are almost always employed to control pH in the formulation.

[00353] The buffer capacity of the buffering species is maximal at a pH equal to the pKa and decreases as pH increases or decreases away from this value. Ninety percent of the buffering capacity exists within one pH unit of its pKa. Buffer capacity also increases proportionally with increasing buffer concentration.

[00354] Several factors need to be considered when choosing a buffer. First and foremost, the buffer species and its concentration need to be defined based on its pKa and the desired formulation pH. Equally important is to ensure that the buffer is compatible with the protein and other formulation excipients, and does not catalyze any degradation reactions. A third important aspect to be considered is the sensation of stinging and irritation the buffer may induce upon administration. For example, citrate is known to cause stinging upon injection (Laursen T, *et al.*, Basic Clin Pharmacol Toxicol., 98(2): 218-21 (2006)). The potential for stinging and irritation is greater for drugs that are administered via the subcutaneous (SC) or intramuscular (IM) routes, where the drug solution remains at the site for a relatively longer period of time than when administered by the IV route where the formulation gets diluted rapidly into the blood upon administration. For formulations that are administered by direct IV infusion, the total amount of buffer (and any other formulation component) needs to be monitored. One has to be particularly careful about potassium ions administered in the form of the potassium phosphate buffer, which can induce cardiovascular effects in a patient (Hollander-Rodriguez JC, *et al.*, Am. Fam. Physician., 73(2): 283-90 (2006)).

[00355] Buffers for lyophilized formulations need additional consideration. Some buffers like sodium phosphate can crystallize out of the protein amorphous phase during freezing resulting in shifts in pH. Other common buffers such as acetate and imidazole may sublime or evaporate during the lyophilization process, thereby shifting the pH of formulation during lyophilization or after reconstitution.

[00356] The buffer system present in the compositions is selected to be physiologically compatible and to maintain a desired pH of the pharmaceutical formulation. In one embodiment, the pH of the solution is between pH 2.0 and pH 12.0. For example, the pH of the solution may be 2.0, 2.3, 2.5, 2.7, 3.0, 3.3, 3.5, 3.7, 4.0, 4.3, 4.5, 4.7, 5.0, 5.3, 5.5, 5.7, 6.0, 6.3, 6.5, 6.7, 7.0, 7.3, 7.5, 7.7, 8.0, 8.3, 8.5, 8.7, 9.0, 9.3, 9.5, 9.7, 10.0, 10.3, 10.5, 10.7, 11.0, 11.3, 11.5, 11.7, or 12.0.

[00357] The pH buffering compound may be present in any amount suitable to maintain the pH of the formulation at a predetermined level. In one embodiment, the pH buffering concentration is between 0.1 mM and 500 mM (1 M). For example, it is contemplated that the pH buffering agent is at least 0.1, 0.5, 0.7, 0.8, 0.9, 1.0, 1.2, 1.5, 1.7, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 25, 30, 40, 50, 60, 70, 80, 90, 100, 200, or 500 mM.

[00358] Exemplary pH buffering agents used to buffer the formulation as set out herein include, but are not limited to organic acids, glycine, histidine, glutamate, succinate, phosphate, acetate, citrate, Tris, HEPES, and amino acids or mixtures of amino acids, including, but not limited to aspartate, histidine, and glycine. In one embodiment of the present invention, the buffering agent is citrate.

[00359] In some embodiments, the formulation comprises 50mM Glycine, 10mM Taurine, 5% (w/w) Sucrose, 5% (w/w) D-Mannitol, 0.1% Polysorbate 80, 2 mM CaCl₂, 150 mM NaCl, and a pH 7.4. In some embodiments, the formulation comprises a high purity mat-rVWF, 50mM Glycine, 10mM Taurine, 5% (w/w) Sucrose, 5% (w/w) D-Mannitol, 0.1% Polysorbate 80, 2 mM CaCl₂, 150 mM NaCl, and a pH 7.4. In some embodiments, the formulation comprises vWF and/or r-vWF/rFVIII and 50mM Glycine, 10mM Taurine, 5% (w/w) Sucrose, 5% (w/w) D-Mannitol, 0.1% Polysorbate 80, 2 mM CaCl₂, 150 mM NaCl, and a pH 7.4.

iii. Pharmaceutical Stabilizers and bulking agents

[00360] In one aspect of the present pharmaceutical formulations, a stabilizer (or a combination of stabilizers) is added to prevent or reduce storage-induced aggregation and chemical degradation. A hazy or turbid solution upon reconstitution indicates that the protein has precipitated or at least aggregated. The term “stabilizer” means an excipient capable of preventing aggregation or physical degradation, including chemical degradation (for example, autolysis, deamidation, oxidation, etc.) in an aqueous state. Stabilizers contemplated include, but are not limited to, sucrose, trehalose, mannose, maltose, lactose, glucose, raffinose, cellobiose, gentiobiose, isomaltose, arabinose, glucosamine, fructose, mannitol, sorbitol, glycine, arginine HCL, poly-hydroxy compounds, including polysaccharides such as dextran, starch, hydroxyethyl starch, cyclodextrins, N-methyl pyrrolidene, cellulose and hyaluronic acid, sodium chloride, (Carpenter *et al.*, Develop. Biol. Standard 74:225, (1991)). In the present formulations, the stabilizer is incorporated in a concentration of about 0.1, 0.5, 0.7, 0.8 0.9, 1.0, 1.2, 1.5, 1.7, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 30, 40, 50, 60, 70, 80, 90, 100, 200, 500, 700, 900, or 1000 mM. In one embodiment of the present invention, mannitol and trehalose are used as stabilizing agents.

[00361] If desired, the formulations also include appropriate amounts of bulking and osmolality regulating agents. Bulking agents include, for example and without limitation, mannitol, glycine, sucrose, polymers such as dextran, polyvinylpyrrolidone, carboxymethylcellulose, lactose, sorbitol, trehalose, or xylitol. In one embodiment, the bulking agent is mannitol. The bulking agent is incorporated in a concentration of about 0.1, 0.5, 0.7, 0.8 0.9, 1.0, 1.2, 1.5, 1.7, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 30, 40, 50, 60, 70, 80, 90, 100, 200, 500, 700, 900, or 1000 mM.

iv. Pharmaceutical Surfactants

[00362] Proteins have a high propensity to interact with surfaces making them susceptible to adsorption and denaturation at air-liquid, vial-liquid, and liquid-liquid (silicone oil) interfaces. This degradation pathway has been observed to be inversely dependent on protein concentration and results in either the formation of soluble and insoluble protein aggregates or the loss of protein from solution via adsorption to surfaces. In addition to container surface adsorption, surface-induced degradation is exacerbated with physical agitation, as would be experienced during shipping and handling of the product.

[00363] Surfactants are commonly used in protein formulations to prevent surface-induced degradation. Surfactants are amphipathic molecules with the capability of out-competing proteins for interfacial positions. Hydrophobic portions of the surfactant molecules occupy interfacial positions (*e.g.*, air/liquid), while hydrophilic portions of the molecules remain oriented towards the bulk solvent. At sufficient concentrations (typically around the detergent's critical micellar concentration), a surface layer of surfactant molecules serves to prevent protein molecules from adsorbing at the interface. Thereby, surface-induced degradation is minimized. Surfactants contemplated herein include, without limitation, fatty acid esters of sorbitan polyethoxylates, *e.g.*, polysorbate 20 and polysorbate 80. The two differ only in the length of the aliphatic chain that imparts hydrophobic character to the molecules, C-12 and C-18, respectively. Accordingly, polysorbate-80 is more surface-active and has a lower critical micellar concentration than polysorbate-20.

[00364] Detergents can also affect the thermodynamic conformational stability of proteins. Here again, the effects of a given detergent excipient will be protein specific. For example, polysorbates have been shown to reduce the stability of some proteins and increase the stability of others. Detergent destabilization of proteins can be rationalized in terms of the hydrophobic tails of the detergent molecules that can engage in specific binding with partially or wholly unfolded protein states. These types of interactions could cause a shift in the conformational equilibrium towards the more expanded protein states (*e.g.* increasing the exposure of hydrophobic portions of the protein molecule in complement to binding polysorbate). Alternatively, if the protein native state exhibits some hydrophobic surfaces, detergent binding to the native state may stabilize that conformation.

[00365] Another aspect of polysorbates is that they are inherently susceptible to oxidative degradation. Often, as raw materials, they contain sufficient quantities of peroxides to cause oxidation of protein residue side-chains, especially methionine. The potential for oxidative damage arising from the addition of stabilizer emphasizes the point that the lowest effective concentrations of excipients should be used in formulations. For surfactants, the effective concentration for a given protein will depend on the mechanism of stabilization.

[00366] Surfactants are also added in appropriate amounts to prevent surface related aggregation phenomenon during freezing and drying (Chang, B, J. Pharm. Sci. 85:1325, (1996)). Thus, exemplary surfactants include, without limitation, anionic, cationic, nonionic, zwitterionic, and amphoteric surfactants including surfactants derived from naturally-

occurring amino acids. Anionic surfactants include, but are not limited to, sodium lauryl sulfate, dioctyl sodium sulfo succinate and dioctyl sodium sulfonate, chenodeoxycholic acid, N-lauroylsarcosine sodium salt, lithium dodecyl sulfate, 1-octanesulfonic acid sodium salt, sodium cholate hydrate, sodium deoxycholate, and glycodeoxycholic acid sodium salt. Cationic surfactants include, but are not limited to, benzalkonium chloride or benzethonium chloride, cetylpyridinium chloride monohydrate, and hexadecyltrimethylammonium bromide. Zwitterionic surfactants include, but are not limited to, CHAPS, CHAPSO, SB3-10, and SB3-12. Non-ionic surfactants include, but are not limited to, digitonin, Triton X-100, Triton X-114, TWEEN-20, and TWEEN-80. Surfactants also include, but are not limited to lauromacrogol 400, polyoxyl 40 stearate, polyoxyethylene hydrogenated castor oil 10, 40, 50 and 60, glycerol monostearate, polysorbate 40, 60, 65 and 80, soy lecithin and other phospholipids such as dioleoyl phosphatidyl choline (DOPC), dimyristoylphosphatidyl glycerol (DMPG), dimyristoylphosphatidyl choline (DMPC), and (dioleoyl phosphatidyl glycerol) DOPG; sucrose fatty acid ester, methyl cellulose and carboxymethyl cellulose. Compositions comprising these surfactants, either individually or as a mixture in different ratios, are therefore further provided. In one embodiment of the present invention, the surfactant is TWEEN-80. In the present formulations, the surfactant is incorporated in a concentration of about 0.01 to about 0.5 g/L. In formulations provided, the surfactant concentration is 0.005, 0.01, 0.02, 0.03, 0.05, 0.06, 0.07, 0.08, 0.09, 0.1, 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, 0.9 or 1.0 g/L.

v. Pharmaceutical Salts

[00367] Salts are often added to increase the ionic strength of the formulation, which can be important for protein solubility, physical stability, and isotonicity. Salts can affect the physical stability of proteins in a variety of ways. Ions can stabilize the native state of proteins by binding to charged residues on the protein's surface. Alternatively, salts can stabilize the denatured state by binding to peptide groups along the protein backbone (-CONH-). Salts can also stabilize the protein native conformation by shielding repulsive electrostatic interactions between residues within a protein molecule. Salts in protein formulations can also shield attractive electrostatic interactions between protein molecules that can lead to protein aggregation and insolubility. In formulations provided, the salt concentration is between 0.1, 1, 10, 20, 30, 40, 50, 80, 100, 120, 150, 200, 300, and 500 mM.

vi. Other common excipient components: Pharmaceutical amino acids

[00368] Amino acids have found versatile use in protein formulations as buffers, bulking agents, stabilizers and antioxidants. Thus, in one aspect histidine and glutamic acid are employed to buffer protein formulations in the pH range of 5.5 - 6.5 and 4.0 - 5.5 respectively. The imidazole group of histidine has a $pK_a = 6.0$ and the carboxyl group of glutamic acid side chain has a pK_a of 4.3 which makes these amino acids suitable for buffering in their respective pH ranges. Glutamic acid is particularly useful in such cases. Histidine is commonly found in marketed protein formulations, and this amino acid provides an alternative to citrate, a buffer known to sting upon injection. Interestingly, histidine has also been reported to have a stabilizing effect, with respect to aggregation when used at high concentrations in both liquid and lyophilized presentations (Chen B, *et al.*, Pharm Res., 20(12): 1952-60 (2003)). Histidine was also observed by others to reduce the viscosity of a high protein concentration formulation. However, in the same study, the authors observed increased aggregation and discoloration in histidine containing formulations during freeze-thaw studies of the antibody in stainless steel containers. Another note of caution with histidine is that it undergoes photo-oxidation in the presence of metal ions (Tomita M, *et al.*, Biochemistry, 8(12): 5149-60 (1969)). The use of methionine as an antioxidant in formulations appears promising; it has been observed to be effective against a number of oxidative stresses (Lam XM, *et al.*, J Pharm Sci, 86(11): 1250-5 (1997)).

[00369] In various aspects, formulations are provided which include one or more of the amino acids glycine, proline, serine, arginine and alanine have been shown to stabilize proteins by the mechanism of preferential exclusion. Glycine is also a commonly used bulking agent in lyophilized formulations. Arginine has been shown to be an effective agent in inhibiting aggregation and has been used in both liquid and lyophilized formulations.

[00370] In formulations provided, the amino acid concentration is between 0.1, 1, 10, 20, 30, 40, 50, 80, 100, 120, 150, 200, 300, and 500 mM. In one embodiment of the present invention, the amino acid is glycine.

vii. Other common excipient components: Pharmaceutical antioxidants

[00371] Oxidation of protein residues arises from a number of different sources. Beyond the addition of specific antioxidants, the prevention of oxidative protein damage involves the careful control of a number of factors throughout the manufacturing process and

storage of the product such as atmospheric oxygen, temperature, light exposure, and chemical contamination. The invention therefore contemplates the use of the pharmaceutical antioxidants including, without limitation, reducing agents, oxygen/free-radical scavengers, or chelating agents. Antioxidants in therapeutic protein formulations are, in one aspect, water-soluble and remain active throughout the product shelf-life. Reducing agents and oxygen/free-radical scavengers work by ablating active oxygen species in solution. Chelating agents such as EDTA are effective by binding trace metal contaminants that promote free-radical formation. For example, EDTA was utilized in the liquid formulation of acidic fibroblast growth factor to inhibit the metal ion catalyzed oxidation of cysteine residues.

[00372] In addition to the effectiveness of various excipients to prevent protein oxidation, the potential for the antioxidants themselves to induce other covalent or physical changes to the protein is of concern. For example, reducing agents can cause disruption of intramolecular disulfide linkages, which can lead to disulfide shuffling. In the presence of transition metal ions, ascorbic acid and EDTA have been shown to promote methionine oxidation in a number of proteins and peptides (Akers MJ, and Defelippis MR. Peptides and Proteins as Parenteral Solutions. In: Pharmaceutical Formulation Development of Peptides and Proteins. Sven Frokjaer, Lars Hovgaard, editors. Pharmaceutical Science. Taylor and Francis, UK (1999)); Fransson J.R., *J. Pharm. Sci.* 86(9): 4046-1050 (1997); Yin J, *et al.*, *Pharm Res.*, 21(12): 2377-83 (2004)). Sodium thiosulfate has been reported to reduce the levels of light and temperature induced methionine-oxidation in rhuMab HER2; however, the formation of a thiosulfate-protein adduct was also reported in this study (Lam XM, Yang JY, *et al.*, *J Pharm Sci.* 86(11): 1250-5 (1997)). Selection of an appropriate antioxidant is made according to the specific stresses and sensitivities of the protein. Antioxidants contemplated in certain aspects include, without limitation, reducing agents and oxygen/free-radical scavengers, EDTA, and sodium thiosulfate.

viii. Other common excipient components: Pharmaceutical metal ions

[00373] In general, transition metal ions are undesired in protein formulations because they can catalyze physical and chemical degradation reactions in proteins. However, specific metal ions are included in formulations when they are co-factors to proteins and in suspension formulations of proteins where they form coordination complexes (*e.g.*, zinc suspension of insulin). Recently, the use of magnesium ions (10 -120 mM) has been proposed to inhibit the isomerization of aspartic acid to isoaspartic acid (WO 2004039337).

[00374] Two examples where metal ions confer stability or increased activity in proteins are human deoxyribonuclease (rhDNase, Pulmozyme®), and Factor VIII. In the case of rhDNase, Ca^{+2} ions (up to 100 mM) increased the stability of the enzyme through a specific binding site (Chen B, *et al.*, / Pharm Sci., 88(4): 477-82 (1999)). In fact, removal of calcium ions from the solution with EGTA caused an increase in deamidation and aggregation. However, this effect was observed only with Ca^{+2} ions; other divalent cations Mg^{+2} , Mn^{+2} and Zn^{+2} were observed to destabilize rhDNase. Similar effects were observed in Factor VIII. Ca^{+2} and Sr^{+2} ions stabilized the protein while others like Mg^{+2} , Mn^{+2} and Zn^{+2} , Cu^{+2} and Fe^{+2} destabilized the enzyme (Fatouros, A., *et al.*, Int. J. Pharm., 155, 121-131 (1997). In a separate study with Factor VIII, a significant increase in aggregation rate was observed in the presence of Al^{+3} ions (Derrick TS, *et al.*, / Pharm. Sci., 93(10): 2549-57 (2004)). The authors note that other excipients like buffer salts are often contaminated with Al^{+3} ions and illustrate the need to use excipients of appropriate quality in formulated products.

ix. Other common excipient components: Pharmaceutical preservatives

[00375] Preservatives are necessary when developing multi-use parenteral formulations that involve more than one extraction from the same container. Their primary function is to inhibit microbial growth and ensure product sterility throughout the shelf- life or term of use of the drug product. Commonly used preservatives include, without limitation, benzyl alcohol, phenol and m-cresol. Although preservatives have a long history of use, the development of protein formulations that includes preservatives can be challenging. Preservatives almost always have a destabilizing effect (aggregation) on proteins, and this has become a major factor in limiting their use in multi-dose protein formulations (Roy S, *et al.*, J Pharm ScL, 94(2): 382-96 (2005)).

[00376] To date, most protein drugs have been formulated for single-use only. However, when multi-dose formulations are possible, they have the added advantage of enabling patient convenience, and increased marketability. A good example is that of human growth hormone (hGH) where the development of preserved formulations has led to commercialization of more convenient, multi-use injection pen presentations. At least four such pen devices containing preserved formulations of hGH are currently available on the market. Norditropin® (liquid, Novo Nordisk), Nutropin AQ® (liquid, Genentech) &

Genotropin (lyophilized - dual chamber cartridge, Pharmacia & Upjohn) contain phenol while Somatrop® (Eli Lilly) is formulated with m-cresol.

[00377] Several aspects need to be considered during the formulation development of preserved dosage forms. The effective preservative concentration in the drug product must be optimized. This requires testing a given preservative in the dosage form with concentration ranges that confer anti-microbial effectiveness without compromising protein stability. For example, three preservatives were successfully screened in the development of a liquid formulation for interleukin-1 receptor (Type I), using differential scanning calorimetry (DSC). The preservatives were rank ordered based on their impact on stability at concentrations commonly used in marketed products (Remmele RL Jr., *et al.*, Pharm Res., 15(2): 200-8 (1998)).

[00378] Development of liquid formulations containing preservatives are more challenging than lyophilized formulations. Freeze-dried products can be lyophilized without the preservative and reconstituted with a preservative containing diluent at the time of use. This shortens the time for which a preservative is in contact with the protein significantly minimizing the associated stability risks. With liquid formulations, preservative effectiveness and stability have to be maintained over the entire product shelf-life (~ 18 -24 months). An important point to note is that preservative effectiveness has to be demonstrated in the final formulation containing the active drug and all excipient components.

[00379] Some preservatives can cause injection site reactions, which is another factor that needs consideration when choosing a preservative. In clinical trials that focused on the evaluation of preservatives and buffers in Norditropin, pain perception was observed to be lower in formulations containing phenol and benzyl alcohol as compared to a formulation containing m-cresol (Kappelgaard A.M., Horm Res. 62 Suppl 3:98-103 (2004)). Interestingly, among the commonly used preservative, benzyl alcohol possesses anesthetic properties (Minogue SC, and Sun DA., AnesthAnalg., 100(3): 683-6 (2005)). In various aspects the use of preservatives provide a benefit that outweighs any side effects.

x. Methods of Preparation of Pharmaceutical Formulations

[00380] The present invention further contemplates methods for the preparation of pharmaceutical formulations.

[00381] The present methods further comprise one or more of the following steps: adding a stabilizing agent as described herein to said mixture prior to lyophilizing, adding at least one agent selected from a bulking agent, an osmolality regulating agent, and a surfactant, each of which as described herein, to said mixture prior to lyophilization.

[00382] The standard reconstitution practice for lyophilized material is to add back a volume of pure water or sterile water for injection (WFI) (typically equivalent to the volume removed during lyophilization), although dilute solutions of antibacterial agents are sometimes used in the production of pharmaceuticals for parenteral administration (Chen, Drug Development and Industrial Pharmacy, 18:1311-1354 (1992)). Accordingly, methods are provided for preparation of reconstituted rVWF compositions comprising the step of adding a diluent to a lyophilized rVWF composition of the invention.

[00383] The lyophilized material may be reconstituted as an aqueous solution. A variety of aqueous carriers, *e.g.*, sterile water for injection, water with preservatives for multi dose use, or water with appropriate amounts of surfactants (for example, an aqueous suspension that contains the active compound in admixture with excipients suitable for the manufacture of aqueous suspensions). In various aspects, such excipients are suspending agents, for example and without limitation, sodium carboxymethylcellulose, methylcellulose, hydroxypropylmethylcellulose, sodium alginate, polyvinylpyrrolidone, gum tragacanth and gum acacia; dispersing or wetting agents are a naturally-occurring phosphatide, for example and without limitation, lecithin, or condensation products of an alkylene oxide with fatty acids, for example and without limitation, polyoxyethylene stearate, or condensation products of ethylene oxide with long chain aliphatic alcohols, for example and without limitation, heptadecaethyl-eneoxycetanol, or condensation products of ethylene oxide with partial esters derived from fatty acids and a hexitol such as polyoxyethylene sorbitol monooleate, or condensation products of ethylene oxide with partial esters derived from fatty acids and hexitol anhydrides, for example and without limitation, polyethylene sorbitan monooleate. In various aspects, the aqueous suspensions also contain one or more preservatives, for example and without limitation, ethyl, or n-propyl, p-hydroxybenzoate.

xi. Exemplary mat-rVWF Formulation for Administration

[00384] In some embodiments, the present methods provide for an enhanced formulation that allows a final product with high potency (high mat-rVWF concentration and enhanced long term stability) in order to reduce the volume for the treatment (100 IU/ml to

10000 IU/ml). In some embodiments, the mat-rVWF concentration in the formulation for administration is about 100 IU/ml to 10000 IU/ml. In some embodiments, the mat-rVWF concentration in the formulation for administration is about 500 IU/ml to 10000 IU/ml. In some embodiments, the mat-rVWF concentration in the formulation for administration is about 1000 IU/ml to 10000 IU/ml. In some embodiments, the mat-rVWF concentration in the formulation for administration is about 2000 IU/ml to 10000 IU/ml. In some embodiments, the mat-rVWF concentration in the formulation for administration is about 3000 IU/ml to 10000 IU/ml. In some embodiments, the mat-rVWF concentration in the formulation for administration is about 4000 IU/ml to 10000 IU/ml. In some embodiments, the mat-rVWF concentration in the formulation for administration is about 5000 IU/ml to 10000 IU/ml. In some embodiments, the mat-rVWF concentration in the formulation for administration is about 6000 IU/ml to 10000 IU/ml. In some embodiments, the mat-rVWF concentration in the formulation for administration is about 7000 IU/ml to 10000 IU/ml. In some embodiments, the mat-rVWF concentration in the formulation for administration is about 8000 IU/ml to 10000 IU/ml. In some embodiments, the mat-rVWF concentration in the formulation for administration is about 9000 IU/ml to 10000 IU/ml. In some embodiments, the mat-rVWF is co-formulated with recombinant coagulation Factor VIII (rFVIII). In some embodiments, the rFVIII is full length FVIII. In some embodiments, the rFVIII is full-length and chemically modified. In some embodiments, the rFVIII comprises a FVIII fusion protein containing FIX-activation peptide instead of B-Domain. In some embodiments, the rFVIII is a FVIII hybrid containing truncated glycosylation rich B-Domain. In some embodiments, the FVIII is a FVIII B-domain-deleted variant. In some embodiments, the FVIII is a chemically modified variant of a FVIII B-domain-deleted variant. In some embodiments, the mat-rVWF with rFVIII co-formulation is made prior to a freeze drying or fill finish step and is stored by mixing the components *in vitro* or in an "on column" procedure (*e.g.*, adding the FVIII during the purification method).

[00385] In some embodiments, the formulation for administration comprises one or more zwitterionic compounds, including for example, amino acids like Histidine, Glycine, Arginine. In some embodiments, the formulation for administration comprises a component with amphipathic characteristic having a minimum of one hydrophobic and one hydrophilic group, including for example polysorbate 80, octylpyranosid, dipeptides, and/or amphipathic peptides. In some embodiments, the formulation for administration comprises a non reducing

sugar or sugar alcohol or disaccharides, including for example, sorbitol, mannitol, sucrose, or trehalose. In some embodiments, the formulation for administration comprises a nontoxic water soluble salt, including for example, sodium chloride, that results in a physiological osmolality. In some embodiments, the formulation for administration comprises a pH in a range from 6.0 to 8.0. In some embodiments, the formulation for administration comprises a pH of about 6.0, about 6.5, about 7, about 7.5 or about 8.0. In some embodiments, the formulation for administration comprises one or more bivalent cations that stabilize rVWF, including for example, Ca^{2+} , Mg^{2+} , Zn^{2+} , Mn^{2+} and/or combinations thereof. In some embodiments, the formulation for administration comprises about 1 mM to about 50 mM Glycine, about 1 mM to about 50 mM Histidine, about zero to about 300 mM sodium chloride (e.g., less than 300 mM sodium), about 0.01 % to about 0.05% polysorbate 20 (or polysorbate 80), and about 0.5 % to about 20% (w/w) sucrose with a pH of about 7.0 and having a physiological osmolality at the time point of administration.

[00386] In some embodiments, the formulation for administration can be freeze dried. In some embodiments, the formulation for administration is stable and can be stored in liquid state at about 2°C to about 8°C, as well as at about 18°C to about 25°C. In some embodiments, the formulation for administration is stable and can be stored in liquid state at about 2°C to about 8°C. In some embodiments, the formulation for administration is stable and can be stored in liquid state at about 18°C to about 25°C.

xii. Administration

[00387] To administer compositions to human or test animals, in one aspect, the compositions comprises one or more pharmaceutically acceptable carriers. The phrases “pharmaceutically” or “pharmacologically” acceptable refer to molecular entities and compositions that are stable, inhibit protein degradation such as aggregation and cleavage products, and in addition do not produce allergic, or other adverse reactions when administered using routes well-known in the art, as described below. “Pharmaceutically acceptable carriers” include any and all clinically useful solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents and the like, including those agents disclosed above.

[00388] The pharmaceutical formulations are administered orally, topically, transdermally, parenterally, by inhalation spray, vaginally, rectally, or by intracranial injection. The term parenteral as used herein includes subcutaneous injections, intravenous,

intramuscular, intracisternal injection, or infusion techniques. Administration by intravenous, intradermal, intramuscular, intramammary, intraperitoneal, intrathecal, retrobulbar, intrapulmonary injection and or surgical implantation at a particular site is contemplated as well. Generally, compositions are essentially free of pyrogens, as well as other impurities that could be harmful to the recipient.

[00389] Single or multiple administrations of the compositions are carried out with the dose levels and pattern being selected by the treating physician. For the prevention or treatment of disease, the appropriate dosage depends on the type of disease to be treated, as defined above, the severity and course of the disease, whether drug is administered for preventive or therapeutic purposes, previous therapy, the patient's clinical history and response to the drug, and the discretion of the attending physician.

xiii. Kits

[00390] As an additional aspect, the invention includes kits which comprise one or more lyophilized compositions packaged in a manner which facilitates their use for administration to subjects. In one embodiment, such a kit includes pharmaceutical formulation described herein (*e.g.*, a composition comprising a therapeutic protein or peptide), packaged in a container such as a sealed bottle or vessel, with a label affixed to the container or included in the package that describes use of the compound or composition in practicing the method. In one embodiment, the pharmaceutical formulation is packaged in the container such that the amount of headspace in the container (*e.g.*, the amount of air between the liquid formulation and the top of the container) is very small. Preferably, the amount of headspace is negligible (*e.g.*, almost none). In one embodiment, the kit contains a first container having a therapeutic protein or peptide composition and a second container having a physiologically acceptable reconstitution solution for the composition. In one aspect, the pharmaceutical formulation is packaged in a unit dosage form. The kit may further include a device suitable for administering the pharmaceutical formulation according to a specific route of administration. Preferably, the kit contains a label that describes use of the pharmaceutical formulations.

xiv. Dosages

[00391] The dosage regimen involved in a method for treating a condition described herein will be determined by the attending physician, considering various factors which

modify the action of drugs, *e.g.* the age, condition, body weight, sex and diet of the patient, the severity of any infection, time of administration and other clinical factors. By way of example, a typical dose of a recombinant VWF of the present invention is approximately 50 U/kg, equal to 500 µg/kg.

[00392] In one aspect, formulations of the invention are administered by an initial bolus followed by a continuous infusion to maintain therapeutic circulating levels of drug product. As another example, the inventive compound is administered as a one-time dose. Those of ordinary skill in the art will readily optimize effective dosages and administration regimens as determined by good medical practice and the clinical condition of the individual patient. The frequency of dosing depends on the pharmacokinetic parameters of the agents and the route of administration. The optimal pharmaceutical formulation is determined by one skilled in the art depending upon the route of administration and desired dosage. See for example, Remington's Pharmaceutical Sciences, 18th Ed. (1990, Mack Publishing Co., Easton, PA 18042) pages 1435-1712, the disclosure of which is hereby incorporated by reference. Such formulations influence the physical state, stability, rate of in vivo release, and rate of in vivo clearance of the administered agents. Depending on the route of administration, a suitable dose is calculated according to body weight, body surface area or organ size. Appropriate dosages may be ascertained through use of established assays for determining blood level dosages in conjunction with appropriate dose-response data. The final dosage regimen is determined by the attending physician, considering various factors which modify the action of drugs, *e.g.* the drug's specific activity, the severity of the damage and the responsiveness of the patient, the age, condition, body weight, sex and diet of the patient, the severity of any infection, time of administration and other clinical factors. As studies are conducted, further information will emerge regarding the appropriate dosage levels and duration of treatment for various diseases and conditions.

EXAMPLES

[00393] The following non-limiting examples are provided for illustrative purposes only in order to facilitate a more complete understanding of representative embodiments.

[00394] These examples should not be construed to limit any of the embodiments described in the present specification including those pertaining to the methods of treating acquired and genetic von Willebrand disease.

Example 1: Purification of matured rVWF on a cation exchanger to separate cVWF propeptide from mature rVWF.

[00395] Example 1 represents a purification of matured rVWF on a cation exchanger (cation exchange (CEX) resin). The rVWF propeptide (rVWF-PP) remains bound to rVWF after furin maturation and was dissociated with sodium citrate as a chelating agent at a neutral pH prior to loading onto a CEX resin. The majority of rVWF propeptide passed through the cation exchange resin. And the remaining rVWF propeptide was depleted after a wash step. Sodium citrate was used as a component of the buffer substance and as a chelating agent.

[00396] Industrially, VWF, in particular recombinant VWF (rVWF), is synthesized and expressed together with rFVIII in a genetically engineered CHO cell line. The function of the co-expressed rVWF is to stabilize rFVIII in the cell culture process. rVWF is synthesized in the cell as the pro-form, containing a large pro-peptide attached to the N-terminus. Upon maturation in the endoplasmic reticulum and Golgi apparatus, the rVWF-PP is cleaved off by the action of the cellular protease furin and is secreted as a homopolymer of identical subunits, consisting of dimers of the expressed protein. However, the maturation is incomplete, leading to a product comprising a mixture of rVWF-PP and mature VWF.

[00397] After a monoclonal antibody step to capture recombinant factor VIII, the flow-through containing rVWF (also referred to as the monoclonal antibody effluent) was loaded onto an anion exchanger (anion exchange (AEX) resin). rVWF was bound on the anion exchanger and was matured with furin in presence of calcium. The rVWF was eluted from the anion exchanger with increasing conductivity. The product containing eluate was conditioned by a 1:2 dilution with 60 mM sodium citrate, pH 7.6 to a conductivity of 13.39 mS/cm and a pH of 7.39. The conditioned aqueous dilution was loaded onto a UNOsphere™ S Cation Exchange Media (Bio Rad, Cat. No.: 156-0115) cation exchanger column with an inner diameter of 15 mm, a bed height of 14.0 cm, and a volume of 24.74 ml with a flow rate of 100 cm/h, and then followed by a wash of 5 CV of 10 mM NaCl, 30 mM Na citrate, pH 7.6±0.2 to remove host cell proteins (HCP) and rVWF-PP. rVWF was eluted by increasing conductivity conducted by a linear gradient with a flow rate of 60 cm/h in 6 CV from 10 mM

NaCl, 30 mM Na Citrate, pH 7.6±0.2 to buffer 500 mM NaCl, 30 mM Na Citrate, pH 7.6±0.2. The main eluate peak was split into two parts to separate low molecular weight rVWF multimers and high molecular weight rVWF multimers.

[00398] FIG. 1 shows purification of matured rVWF on a cation exchanger as represented in Example 1.

[00399] FIG. 2 provides a table of the purification results.

[00400] FIG. 3 shows a silver stained protein gel and a western blot illustrating the separation of rVWF and rVWF-PP by the method described in Example 1.

Examples 2 and 3: Optimized method as described in Example 1 for commercial manufacturing of rVWF.

[00401] Examples 2 and 3 represent an optimized method as described in Example 1 for commercial manufacturing of rVWF.

[00402] For Examples 2 and 3 an experimental setup for fermentation of rVWF and rFVIII was established. The method was used for a simplified purification method to obtain high pure rVWF for biochemical characterization.

[00403] The capture step was performed by tandem chromatography, which combined an affinity chromatography and an anion exchange chromatography in a single process step. rFVIII was bound on an anti FVIII-mAb column at a temperature of 2-8 °C based on immune affinity chromatography technique. This step can separate rFVIII from rVWF. The rVWF containing flow-through was online diluted in the same chromatography system with purified water and loaded directly on an AEX column. Recombinant furin maturation on the AEX column was carried out after increasing the temperature to +15 °C to 28 °C. The furin matured rVWF was eluted with a step elution by increasing conductivity. A polishing step was also performed. The rVWF containing AEX eluate was diluted with 10 mM Na citrate buffer, pH 7.6 and applied onto an UNOsphere™S Cation Exchange Media (Bio Rad, Cat. No.: 156-0115) cation exchanger column having an inner diameter of 15 mm, a bed height of 14.0 cm, and a column volume of 25 ±0.5ml with a flow rate of 100cm/h. After a wash step with 10 mM NaCl, 30 mM Na citrate, 2mM citric acid pH 7.6±0.2, rVWF was eluted with increasing conductivity using a linear gradient with a flow rate of 65 cm/h in 6CV from 10

mM NaCl, 30 mM Na citrate, pH 7.6±0.2 to a buffer of 500 mM NaCl, 30 mM Na citrate, pH 7.6±0.2. The main eluate peak was collected as eluate (pooled eluate) for analytical purposes.

[00404] In the final experimental design the last 30 to 40% of the peak was collected to obtain the rVWF with the highest specific activity.

[00405] FIG. 4 shows a flow chart of the experimental set-up for Examples 2 and 3.

[00406] FIG. 5 shows a chromatogram for Example 2 and a chromatography scheme used for Examples 2 and 3.

[00407] FIG. 6 provides a table of the reagents used and a table of the results for Example 2.

[00408] FIG. 7 shows another chromatogram for Example 2 and a table of the results for Example 3.

[00409] FIG. 8 shows a silver stained protein gel illustrating the separation of rVWF and rVWF propeptide by the method of Example 2 and Example 3.

[00410] FIG. 9 shows a western blot illustrating the separation of rVWF and rVWF propeptide by the method of Example 2 and Example 3.

Example 4: Method for commercial manufacturing of rVWF by separate rVWF and rVWF - PP by size exclusion chromatography.

[00411] Example 4 represents an optimized method for commercial manufacturing of rVWF by separating rVWF and rVWF propeptide (rVWF-PP) via size exclusion chromatography. Sodium citrate is added to the SEC running buffer to provide an efficient split of rVWF and rVWF -PP.

[00412] A rVWF containing ultrafiltrated UNOsphere™ S-eluate was loaded directly onto an array of two Superose 6 prep grade SEC columns in series (GE Healthcare, Cat. No.: 28-9913-16), both with an inner diameter of 16 mm each, a bed height 82.0 cm (2 x 41 cm), and the volume of both columns was approximately 165 ml. The load was applied at a rate of 7 cm/h. The running buffer was 20 mM HEPES free acid, 150 mM NaCl, 15 mM Na citrate

dihydrate pH 7.5 ± 0.2 . The size exclusion chromatography was carried out with isocratic conditions at a linear flow rate of 12 cm/h.

[00413] FIG. 10 shows a chromatogram, a chromatography scheme, and buffer compositions for Example 4.

[00414] FIG. 11 provides a table of the results for Example 4.

[00415] FIG. 12 shows a silver stained protein gel and a western blot illustrating the separation of rVWF and rVWF propeptide by the method of Example 4.

Example 5: Optimized method for commercial manufacturing of mature rVWF by separate rVWF and rVWF-PP by size exclusion chromatography.

[00416] Example 5 represents a method for separating rVWF and rVWF-PP by size exclusion chromatography by applying a pH conditioned rVWF containing start material onto size exclusion chromatography.

[00417] A rVWF containing ultrafiltrated UNOsphereTMS-eluate was conditioned to a pH of 7.5 ± 0.2 with 1 M glycine pH 9.0 prior loading onto the column. This solution was loaded onto an array of two Superose 6 prep grade SEC columns in series (GE Healthcare, Cat. No.: 28-9913-16), both with an inner diameter of 16 mm each, a bed height 82.0 cm (2 x 41 cm), and the volume of both columns was approximately 165 ml. The load was applied at a flow rate of 7 cm/h. The SEC running buffer comprised 20 mM HEPES free acid and 150 mM NaCl, pH 7.5 ± 0.2 . The size exclusion chromatography was carried out with isocratic conditions at a linear flow rate of 12 cm/h.

[00418] FIG. 13 shows a chromatogram, a chromatography scheme, and buffer compositions for Example 5.

[00419] FIG. 14 provides a table of the results for Example 5.

Example 6: CEX method for purification of rVWF from rVWF propeptide without supplementation of chelating agents on a UNOsphereTM S.

[00420] Example 6 represents an CEX method without supplementation of chelating agents on ultrafiltrated UNOsphereTM S. This method is representative of a prior art method

for purifying mature rVWF from rVWF propeptide. The method does not utilize a buffer comprising a chelating agent and/or a buffer having a pH of 7.0 or higher.

[00421] After a monoclonal antibody step to capture recombinant factor VIII, the flow-through, which contains rVWF, was loaded onto an anion exchanger. rVWF was bound on the anion exchanger and was matured with furin in presence of calcium. The rVWF was eluted from the anion exchanger with increasing conductivity. The product containing eluate was then loaded onto a UNOsphere™ S Cation Exchange Media (Bio Rad, Cat. No.: 156-0115) cation exchanger column with an inner diameter of 15 mm, a bed height of 14.2 cm, and a volume of 25.09 ml at a flow rate of 100 cm/h followed by a wash of 10 CV of 10 mM Tris-HCl, 100 mM Na acetate, 85 mM NaCl, pH 6.5 ±0.2 to remove HCP and rVWF-propeptide. rVWF was eluted with a single step by applying 100 mM Na acetate, 500 mM NaCl, 100 mM glycine, 3 mM CaCl₂, pH 7.5 ±0.2 at flow rate of 65 cm/h. The main eluate peak was collected as product containing fraction.

[00422] FIG. 15 shows a chromatogram, a chromatography scheme, and buffer compositions and conditions for Example 6.

[00423] FIG. 16 provides a table of the results for Example 6.

Example 7: SEC method for purification of rVWF from rVWF propeptide without prior supplementation of chelating agents or elevated pH.

[00424] Example 7 represents SEC method without prior supplementation of chelating agents or elevated pH. This method is representative of a prior art method for purifying mature rVWF from rVWF propeptide. The SEC method does not include a buffer comprising a chelating agent and/or a buffer having a pH of 7.0 or higher which is used to condition the starting fraction (material) containing rVWF and residual rVWF propeptide.

[00425] A recombinant VWF containing ultrafiltrated UNOsphere™ S-eluate was loaded directly onto an array of two Superose 6 prep grade SEC columns in series (GE Healthcare, Cat. No.: 28-9913-16), both with an inner diameter of 16 mm each, a bed height of 82.0 cm (2 x 41 cm), and the volume of both columns was approximately 165 ml. The load was applied at a flow rate of 7 cm/h. The running buffer was 20 mM HEPES free acid, 150 mM NaCl, pH7.5 ±0.2. The size exclusion chromatography was carried out with isocratic conditions at a linear flow rate of 12 cm/h.

[00426] FIG. 17 shows a chromatogram, a chromatography scheme, and buffer compositions for Example 7.

[00427] FIG. 18 provides a table of the results for Example 7.

Example 8: Separation of rVWF from rVWF propeptide by anion exchange chromatography and cation exchange chromatography.

[00428] Example 8 represents a purification of matured rVWF on a cation exchanger. The start material was obtained from the current rVWF manufacturing process after the AEX Mustang Q step. The rVWF containing Flow-Through from the AEX Mustang Q step was SD/VI treated and diluted with the chelating agent containing buffer to dissociate rVWF/rVWF-propeptide-complex. The diluted material was applied onto a CEX resin(Unosphere S). The majority of rVWF-PP, host cell proteins(HCPs) and low molecular weight rVWF multimers pass through the cation exchange resin. Remaining rVWF-PP was depleted after a wash step. The bound high molecular weight rVWF multimers were subsequently eluted by increasing the conductivity triggered by sodium ions.

[00429] Industrially, VWF, in particular recombinant VWF (rVWF), is synthesized and expressed together with rFVIII in a genetically engineered CHO cell line. The function of the co-expressed rVWF is to stabilize rFVIII in the cell culture process. rVWF is synthesized in the cell as the pro-form, containing a large pro-peptide attached to the N-terminus. Upon maturation in the endoplasmic reticulum and Golgi apparatus, the pro-peptide is cleaved off by the action of the cellular protease furin and is secreted as a homopolymer of identical subunits, consisting of dimers of the expressed protein. However, the maturation is incomplete, leading to a product comprising a mixture of pro-peptide and mature VWF.

[00430] After a monoclonal antibody step to capture recombinant factor VIII, the flow-through, which contains rVWF, was loaded onto a Fractogel TMAE anion exchanger. rVWF is bound on the anion exchanger and was matured with furin in presence of calcium. The rVWF was eluted from the anion exchanger with increasing conductivity. The TMAE-Eluate was filtrated through a Mustang Q (MUQ) filter unit to remove CHO-DNA and impurities that binds to the filter membrane. The loading material for the CEX step is the effluent of the Mustang Q filtration step (MUQ) that is treated with solvent and detergents to inactivate lipid

enveloped viruses. For virus inactivation the MUQ effluent is incubated with a mix of the two detergents such as Triton-X-100 (1%) and polysorbate 80 (0.3%) and the organic solvent tri-n-butyl phosphate (0.3%) for one hour at room temperature. The product containing MUQ_flow-through was conditioned by a 1:2 dilution with 60 mM sodium citrate pH 7.6 to a conductivity of 21.9 mS/cm and a pH 7.16. The high conductivity was chosen to ensure the removal of rVWF propeptide (rVWF-PP) and low molecular weight rVWF multimers to utilize the capacity of the resin for the desired high molecular weight rVWF multimers. The conditioned dilution was loaded onto a UNOsphere™ S Cation Exchange Media (Bio Rad, Cat. No.: 156-0115) cation exchanger column with an inner diameter of 10 mm, a bed height of 14.3 cm, and volume of 11.23 ml with a flow rate of 100 cm/h. After loading, a first wash (Reequilibration) was performed using 5 CV of 10 mM NaCl, 30 mM Na Citrate, pH 7.6±0.2 to remove weakly bound HCP and rVWF-propeptide.

[00431] The second wash to deplete strong bound HCP and rVWF -propeptide was carried out with a step of 40% 500 mM NaCl, 30 mM Na citrate, pH 7.6±0.2 in 10 mM NaCl, 30 mM Na citrate, pH 7.6±0.2 (Wash 2).

[00432] The elution was carried out in two phases: (1) the first phase included a step of 45% 500 mM NaCl, 30 mM Na citrate, pH 7.6±0.2 in 10 mM NaCl, 30 mM Na citrate, pH 7.6±0.2 (Eluate 1 or E1), and (2) the second phase included a linear gradient from 45% to 100% of 500 mM NaCl, 30 mM Na citrate, pH 7.6±0.2 in 10 mM NaCl, 30 mM Na citrate, pH 7.6±0.2 (Eluate 2 or E2) in 6 column volumes. Wash 2 to the end of the gradient was performed at a flow rate of 65 cm/h.

[00433] FIG. 19 shows a chromatogram, a chromatography scheme, and buffer compositions for Example 8.

[00434] FIG. 20 provides a table of the results for Example 8.

[00435] FIG. 21 shows a silver stained protein gel illustrating the separation of rVWF and rVWF-propeptide by the method of Example 8.

[00436] FIG. 22 shows a western blot illustrating the separation of rVWF and rVWF-propeptide by the method of Example 8. The 1% agarose gel shows the multimeric pattern of the products.

[00437] FIG. 23 shows a western blot illustrating the separation of rVWF and rVWF-propeptide by the method of Example 8.

Example 9: Separation of rVWF from rVWF propeptide by anion exchange chromatography and cation exchange chromatography.

[00438] Example 9 represents an optimized purification of matured rVWF on a cation exchanger. The start material was obtained from the current r-VWF manufacturing process after the AEX Mustang Q step. The rVWF containing Flow-Through from the AEX Mustang Q step was SD/VI treated and diluted with the chelating agent containing buffer to dissociate the rVWF/rVWF-Propeptide-complex. The diluted material was applied onto a CEX resin (Unosphere S). The majority of rVWF-PP, host cell proteins and low molecular weight rVWF multimers pass through the cation exchange resin. Remaining rVWF-PP was depleted after a wash step. The bound high molecular weight rVWF multimers were eluted by a gradient of increasing the conductivity triggered by sodium ions.

[00439] Industrially, VWF, in particular recombinant VWF (rVWF), is synthesized and expressed together with rFVIII in a genetically engineered CHO cell line. The function of the co-expressed rVWF is to stabilize rFVIII in the cell culture process. rVWF is synthesized in the cell as the pro-form, containing a large pro-peptide attached to the N-terminus. Upon maturation in the endoplasmic reticulum and Golgi apparatus, the pro-peptide is cleaved off by the action of the cellular protease furin and is secreted as a homopolymer of identical subunits, consisting of dimers of the expressed protein. However, the maturation is incomplete, leading to a product comprising a mixture of pro-peptide and mature VWF.

[00440] After a monoclonal antibody step to capture recombinant factor VIII, the flow-through, which contains r-VWF, was loaded onto a Fractogel TMAE anion exchanger. rVWF was bound on the anion exchanger and was matured with furin in presence of calcium. The rVWF was eluted from the anion exchanger with increasing conductivity. The TMAE-Eluate was filtrated through a Mustang Q (MUQ) filter unit to remove CHO-DNA and impurities that binds to the filter membrane. The loading material for the CEX step is the effluent of the Mustang Q filtration step (MUQ) that is treated with solvent and detergents to inactivate lipid enveloped viruses.

[00441] For virus inactivation the MUQ effluent is incubated with a mix of the two detergents Triton-X-100 (1%) and polysorbate 80 (0.3%) and the organic solvent tri-n-butyl phosphate (0.3%) for one hour at room temperature. The product containing MUQ_flow

through was conditioned by a 1:2 dilution with 60mM sodium citrate pH 7.6 to a conductivity of 21.9 mS/cm and pH 7.16. The high conductivity is chosen to ensure the removal of rVWF-propeptide and low molecular weight rVWF to utilize the capacity of the resin for the desired high molecular weight r-VWF. The conditioned dilution was loaded onto a UNOsphere™ S Cation Exchange Media (Bio Rad, Cat. No.: 156-0115) cation exchanger column with an inner diameter of 10 mm, a bed height of 14.3 cm, and volume of 11.23 ml with a flow rate of 100 cm/h followed by a first wash (Reequilibration) of 5 CV of 10 mM NaCl, 30 mM Na citrate, pH 7.6±0.2 to remove weakly bound HCP and rVWF-propeptide.

[00442] The second wash (Wash 2) to deplete strong bound HCP and rVWF-propeptide was carried out with a step of 36% 500 mM NaCl, 30 mM Na citrate, pH 7.6±0.2 in 10 mM NaCl, 30 mM Na citrate, pH 7.6±0.2 in 5 column volumes.

[00443] The elution was carried out with a gradient from 36% 500 mM NaCl, 30 mM Na citrate, pH 7.6±0.2 in 10 mM NaCl, 30 mM Na citrate, pH 7.6±0.2 to 100% 500 mM NaCl, 30 mM Na citrate, pH 7.6±0.2 in 10 mM NaCl, 30 mM Na citrate, pH 7.6±0.2 in 8 column volumes. The eluate representing the desired product contains the pool of fractions beginning at >50% of 500 mM NaCl, 30 mM Na citrate, pH 7.6±0.2 in 10 mM NaCl, 30 mM Na citrate, pH 7.6±0.2 to 76% of 500 mM NaCl, 30 mM Na citrate, pH 7.6±0.2 in 10 mM NaCl, 30 mM Na citrate, pH 7.6±0.2. The wash and elution were performed with a flow rate of 50 cm/h.

[00444] FIG. 24 shows a chromatogram, a chromatography scheme, and buffer compositions for Example 9.

[00445] FIG. 25 provides a table of the results for Example 9.

[00446] FIG. 26 provides a table of the products for Example 9.

[00447] FIG. 27 shows a silver stained protein gel illustrating the separation of rVWF and rVWF-propeptide by the method of Example 9.

[00448] FIG. 28 shows a western blot illustrating the separation of rVWF and rVWF propeptide by the method of Example 9. The 1% agarose gel shows the multimeric pattern of the products.

[00449] FIG. 29 shows a western blot illustrating the separation of rVWF and rVWF propeptide by the method of Example 9.

[00450] rVWF purification steps in presence of chelating agents and /or elevated pH showed a high depletion rate of r-VWF propeptide and host cell proteins. The depletion of r-VWF propeptide on cation exchanger is based on the fact that rVWF-PP does not bind onto a cation exchanger at condition in presence of chelating agents and/or elevated pH. The depletion of rVWF propeptide on size exclusion chromatography based on the fact of an efficient size separation in presence of chelating agents and/or elevated pH.

[00451] FIG. 30 shows the purity of the product containing fractions obtained for enhanced cation exchange chromatography (CEX) as used for Examples 1, 2, 3, 6, 8, and 9.

[00452] FIG. 31 shows the depletion factor of product related impurities for Examples 1, 2, 3, 6, 8, and 9.

[00453] FIG. 32 shows the purity of the product containing fractions obtained for enhanced size exclusion chromatography (SEC) as used for Examples 4 and 5.

FIG. 33 shows the depletion factor of product related impurities for Examples 4 and 5.

References

[00454] U.S. Patent No. 8,058,411; Method for producing mature VWF from VWF pro-peptide. Inventors: Wolfgang Mundt, Artur Mitterer, Meinhard Hasslacher, Christa Mayer.

[00455] U.S. Patent No 6,465,624; Purification of von Willebrand factor by cation exchange chromatography. Inventors: Bernhard Fischer, Öyvind L. Schönberger, Artur Mitterer, Christian Fiedler, Friedrich Dörner, Johann Eibl.

Example 10: Separation of rVWF from rVWF propeptide by anion exchange chromatography.

[00456] This study illustrates the dissociation (separation) of furin processed mature VWF/VWF-PP complex into mature VWF and VWF-PP using anion exchange chromatography and a elution buffer with an elevated pH (*e.g.*, pH 8.5) and containing a chelating agent (EDTA). The separation was carried out on an anion exchanger (AEX), in particular, a Fractogel TMAE 650(M). A solvent-detergent treatment for viral inactivation was also performed on the column for about 1 hour. Details of the chromatography experiment are provided in FIGS. 34-36.

[00457] FIG. 34 shows the buffer formulations and materials used in the TMAE separation method.

[00458] FIG. 35 shows the loading conditions for the furin-processed mature VWF/VWF-propeptide complex.

[00459] FIG. 36 shows the details of the buffers, conditions, parameters, and flow rates of the chromatography method.

[00460] FIG. 37 shows a chromatogram of the dissociation of furin-processed mature VWF/VWF-propeptide complex into mature VWF and VWF-propeptide (VWF-PP). It shows depletion of VWF-PP from the fraction containing mature VWF.

[00461] FIG. 38 shows another chromatogram of the separation of mature VWF and VWF-propeptide (VWF-PP). It shows depletion of VWF-PP from the fraction containing mature VWF.

Example 11: Improvements in different chromatography methods for the separation of mature VWF (matVWF) and VWF propeptide (VWF-PP).

[00462] In the first study, two methods for purifying recombinant mature VWF were compared.

[00463] FIG. 39 provides a schematic of the two methods for isolating mature VWF. In one method the downstream processing steps, such as those after obtaining the mAb effluent (MABEffl), the capture step of TMAE anion exchange chromatography and on-column maturation (TMC), and the Mustang Q negative anion exchange chromatography step (MUQ) include solvent-detergent treatment (SDT) for viral inactivation, cation exchange chromatography (CAT), ultrafiltration concentration (UFA), size exclusion chromatography (SEC), and dialysis-ultrafiltration concentration (DUF) to produce a bulk drug substance (mature VWF). In the other method, the downstream processing steps include an improved cation exchange chromatography (CAT) step followed by a dialysis-ultrafiltration (DUF) concentration step to produce a bulk drug substance, and do not include SEC.

[00464] FIG. 40 provides a table highlighting some of the advantages of the improved cation exchange chromatography method (CAT 2.0) described herein and shown in FIG. 39. The improved CAT method can remove: host cell impurities by a reduction factor of greater

than 1000, VWF-PP by a reduction factor of greater than 2000, and residual FVIII by a reduction factor of less than 10. The CAT method can be used to separate and pool VWF multimers. In addition, the method can replace size exclusion chromatography as a polishing step to isolate the active fraction of VWF and to remove remaining host cell derived impurities and VWF-PP.

[00465] In the second study, the conditions of the SEC process were varied to improve the separation mature VWF and VWF-PP. In other words, it was determined that a modified buffer for SEC could increase the purity of mature VWF by reducing the amount of VWF-PP.

[00466] FIG. 41 shows a schematic of two chromatograms showing the separation of r-VWF propeptide using size exclusion chromatography with a standard SEC buffer (SQA buffer) or with a modified SEC buffer (SQC buffer). FIG. 42 provides a table highlighting some of the advantages of using the SQC buffer. For instance, the method using the SQC buffer can remove host cell impurities by a reduction factor of greater than about 100 and residual FVIII by a reduction factor of less than 10. Surprisingly, it can remove VWF-PP such that the impurity levels are less than 2 µg/1000 units.

[00467] As such, described in this example are methods of improving the separation of mature VWF from VWF-PP.

Example 12: Development of an improved CAT (UNO_S) Step.

[00468] The downstream process of recombinant von Willebrand factor(rVWF) 1st generation starting from monoclonal antibody (MAB) flow through includes a polishing step by cation exchange chromatography (CAT) on UNO_Sphere S (UNO_S) resin. The UNO_S Eluate is thereafter concentrated by ultrafiltration and further processed by Size-Exclusion-Chromatography (SEC) to separate high and low molecular weight rVWF multimers and to remove free rVWF pro-peptide, a product related impurity generated in course of the downstream process. The high molecular weight rVWF sub-fraction represents bulk drug substance (BDS) that is finally formulated to obtain final drug product (FDP).

[00469] For the downstream process of 2nd generation rVWF, it was suggested to replace the SEC step by an improved cation exchange chromatography method and to separate high and low and molecular weight rVWF multimers as well as rVWF pro-peptides by an alternative cation exchange (CAT) elution procedure (gradient elution, instead of step

elution). In this example the experiments for the 2nd generation rVWF polishing purification step CAT are outlined. New process parameters like the CAT loading pH and conductivity, the conductivity and length of the column washing steps and the eluate pooling criteria were explored on small scale to obtain a scalable and robust process downstream unit operation step.

1. Objective

[00470] The downstream process of 1st generation rVWF (VONVENDI®) starts with a capture step on TMAE Sepharose (TMC step) using ADVATE® MAB flow through as feed, followed by a Mustang Q filtration step to remove CHO host cell DNA. Next, a Solvent/Detergent (S/D) step is performed to inactivate potential lipid enveloped viruses, followed by a polishing step on UNO_Sphere S (UNO_S) resin a weak cation exchanger (CAT step). The CAT step is dedicated to remove the S/D chemicals introduced during for virus inactivation step. The UNO_S Eluate is thereafter concentrated by ultrafiltration and further processed by Size-Exclusion-Chromatography (SEC) to separate high and low molecular weight rVWF multimers and to remove free rVWF pro-peptide, a product related impurity generated in course of the downstream process. The high molecular weight rVWF sub-fraction represents BDS that is finally formulated to obtain FDP.

[00471] For the downstream process of 2nd generation rVWF it was suggested to cancel the SEC step and to replace it by an improved cation exchange chromatography method.

[00472] In a series of five experiments, the separation of high from low molecular weight rVWF multimers as well as the removal of rVWF pro-peptides was achieved by a gradient CAT elution procedure. The new gradient elution mode was able to replace the step elution procedure that is applied in the 1st generation downstream process. In this example the five experiments for the 2nd generation rVWF polishing purification step CAT are outlined in detail. All experiments were performed according to study plan described herein.

2. Introduction and Background

[00473] The current report describes the development of a 2nd generation (Gen 2) T process, by combining two VWF downstream unit operation steps CAT and SEC as currently applied in the 1st generation (Gen 1) procedure. In a series of experiments, process parameters were explored that had been identified in a risk assessment and that were

considered as important for the performance of the chromatographic step CAT. The current study was based on a scale down model from the current rVWF manufacturing process. This process was established in Orth for the production of Clinical Phase III material and transferred to manufacturing (MFG) scale for commercial production (FIG. 43A). To facilitate an understanding of the introduced changes in the CAT Unit operation step described in the this report, a brief process description of the currently used 1st generation rVWF downstream unit operation steps S/D, CAT and SEC is given below.

[00474] As used in the Gen I process, the rVWF polishing step CAT is a chromatographic cation exchange process on UNO_Sphere S, a macroporous acrylamido based media with a “strong” sulfonic cation exchange ligand. The loading material for the polishing step is the effluent of the anion exchange filtration step MUQ that is treated with solvent and detergents to inactivate lipid enveloped viruses. For virus inactivation the MUQ effluent is incubated with a mix of the two detergents Triton-X-100 (1%) and Polysorbate 80 (0.3%) and the organic solvent tri-n-butyl phosphate (0.3%) for one hour at room temperature. Prior treatment the product solution is filtered through a 0.2 μ m membrane filter to remove potentially present particulates. After virus inactivation, the product solution is diluted with approximately one volume of water to reduce the concentration of the S/D reagents adjust the conductivity for the loading step onto the CAT Column. The pH is not adjusted. The CAT chromatographic step has the main objective to remove the S/D reagents and further reduce process related impurities including media components like soy peptone and other impurities like rFurin, rFVIII polypeptides and CHO derived proteins and DNA. Following the unit operation step CAT, the obtained product fraction (CAT-E) is further processed by Size Exclusion chromatography (SEC) on Superose 6 resin. The loading material for the polishing step SEC is the eluate pool of the Cation Exchange polishing step CAT on UNO_Sphere S. As the loading volume for a SEC column is limited to achieve a reasonable resolution the CAT eluate pool is concentrated by a factor of approximately 15 by ultrafiltration using a cellulose based membrane cassette with a cut-off of 30 kDa (step UFA). At clinical phase III production scale the ultrafiltration concentration (UFA) concentrate is divided in two fractions that are processed separately on the SEC column. This measure was implemented to keep the SEC column volume and column diameters low. The buffer matrix as well as conductivity and pH of the loading material corresponds to the CAT eluate pool and is not adjusted after the concentration step UFA before loading onto the SEC

column. The objective of the step SEC is the final impurity removal for CHO host cell proteins and serves as the major removal step for the product related impurity rVWF pro-peptide generated during the initial capture the step on TMAE Sepharose (TMC step). In addition, the step SEC resolves rVWF multimers based on their size allowing a pooling schema for enrichment of high molecular weight rVWF multimers that contribute to Ristocetin Cofactor activity of the product.

[00475] This report describes the replacement of the current unit operation steps performed in the MFG (FIG. 43A) scale by an improved CAT (UNO_S) step (FIG. 43B). The CAT step improvement was investigated on a small scale. The UDF (concentration/dialysis) step following the CAT step might have to be optimized as well.

3. Materials and Methods

[00476] The materials and the methods as well as the sampling plan are described herein.

3.1 rVWF load materials

[00477] For all experiments, frozen MUQ-E product was used. The material was stored frozen at $\leq -60^{\circ}\text{C}$ in 130 mL aliquots and was thawed overnight at a range from +2 to $+8^{\circ}\text{C}$ on demand. Once the MUQ – Eluate was thawed, S/D reagents were added and the mixture was filtered through a 0.2 μm filter KA02EAVP2S® from Pall. Thereafter, the filtered material was incubated under moderate stirring for 60min at ambient room temperature (about $+25^{\circ}\text{C}$) to inactivate / dissolve potential lipid enveloped viruses. The S/D reaction was stopped by 1:2 dilution with 60 mM Na-Citrate buffer, pH 7.5. Diluted material was used as feed for the following CAT step.

3.2 Chromatography hardware

[00478] For the experiments described in the current report, the small scale chromatography system ÄKTA pure 25 (GE Healthcare) was used. The system was equipped with probes for on-line monitoring UV absorption, conductivity, pressure, temperature and pH with electronic recording. The system was controlled by Unicorn 7.0 operated software. All runs were performed at ambient room temperature.

[00479] The ÄKTA system tubings were PEEK which is different to the large scale where a Millipore process system with stainless steel piping is used. The hardware components are all qualified R&D equipment.

[00480] The lab-scale column that was used for all five experiments was equipped with 10 µm PP frits; the particle size of the UNO_Sphere S resin was about 80 µm in diameter. At large scale stainless steel frits with a mesh size of 20 µm are used. All columns are qualified items designed for R&D purposes.

[00481] A hardware comparison between the current GEN 1 MGF equipment in NE and the small scale GEN 2 equipment used in the current study is shown in FIG. 44.

3.3 Buffers

[00482] The buffers used for the small scale purification runs were made in the laboratory area or were received from the manufacturing area. For the preparation of buffers, qualified chemicals that were also used for the production of buffers for pilot scale clinical production were used. Buffers were 0.2 µm filtered and stored in bags or glass bottles at room temperature before use. A description of the buffer composition is given in FIG. 48.

3.4 Analytical methods

[00483] The rVWF biochemical characterization, potency and impurity assays performed include those to analyze VWF:RistoCo activity, VWF antigen, VWF-propeptide antigen content, FVIII activity chromogenic method, UV absorption profile (280nm, 254nm), polypeptide pattern such as degradation, multimer pattern, and CHO HCP content. In some cases, other analytical test can be performed to determine, such as but not limited to, pro-VWF antigen content, FVIII antigen content, furin activity, furin antigen, total protein (BCA), free sulfhydryl, CHO BIP WB, CHO DNA, murine monoclonal antibody, soy peptone, Triton X-100, polysorbate 80, tri-n-butylphosphate, dynamic light scattering (DLS) (hydrodynamic radius), sialic acids, n-glycan content, VWF collagen binding, and VWF oxidation.

4. Alterations in the CAT process rVWF 2nd generation (GEN 2)

[00484] In order to replace the SEC step in a 2nd generation rVWF downstream process the parameters listed below were explored. Most of the changes introduced are based on R&D feasibility studies. The chromatography resin type (UNO_Sphere S resin by BioRad)

and the composition (not the pH) of the applied buffer was not altered. The 2nd generation CAT process included the following changes: S/D treatment, loading concentration and flow rates, and wash and elution steps.

4.1 S/D treatment

[00485] The S/D treatment was performed in the same way as in the GEN 1 process, except the S/D inactivation was stopped by a 1:2 dilution of the virus inactivated material with 60mM Na-citrate buffer that set the CAT feed to a preferred pH of 7.5 – 8.0 (pH testing range 6.0 – 9.0) and to a preferred conductivity of 10 – 30mS/cm² at +25°C (conductivity testing range 5 – 40mS/cm² at +25°C). In the GEN 1 rVWF CAT step that was performed, the CAT load was set to a pH of 8.9 – 9.2. In the GEN 2 set-up, conductivity and pH were set to a point that minimized the CHO-HCP, CHO-DNA and rVWF pro-peptide binding to the matrix. Similarly low molecular weight (LMW) rVWF molecules were hindered to bind to the column matrix, whereas preferably only high molecular weight (HMW) rVWF molecules were captured. One aim of the present study was to increase the conductivity during the loading phase and to deplete as much LMW rVWF, CHO-HCP, CHO-DNA and rVWF pro-peptide from the feed as possible.

4.2 Loading concentrations and flowrates

[00486] The loading concentration (RU rVWF/mL resin) was increased in course of the study to enable a higher product load without increasing the column volume. At the manufacturing scale (MFG) a loading concentration of 60–140 RU/ml resin is generally applied, in contrast, in the current small scale study 90–270 IU /ml resin were loaded. The equilibration, loading and re-equilibration flow rates of the 2nd generation CAT procedure were the same as in the 1st generation process (100 cm/h). Washing and elution flowrates were altered as shown in FIG. 45.

4.3 Loading concentrations and flowrates

[00487] The washing step preceding the elution phase was altered to optimize the removal of process and product related impurities. The step elution as applied in the 1st generation process was changed to a gradient elution. The gradient length was explored in course of the study. The change of the elution procedure was based on the observation that low molecular weight rVWF molecules elute in early gradient fractions where as high molecular weight rVWF molecules elute in late gradient fractions (see, *e.g.*, US6,465,624).

5. Comparision of the CAT Gen 1 and Gen 2 process

[00488] In both procedures the CAT process includes the following steps: column activation (loading of the anionic ligand with the cationic counter ion sodium) and equilibration (preparing the column for loading in terms of a stable pH and conductivity, monitored at the column outlet), followed by the product loading of the S/D treated and diluted MUQ eluate.

[00489] During loading on MFG scale, the eluate was filtered online through a 0.2 μ m filter to protect the column against particulate matter that could have been formed during the S/D treatment. In the small scale process, this step was omitted. After pumping the product containing solution onto the column, the loading was completed and loosely bound impurities were removed by applying a wash step which removes low molecular weight S/D reagents that were pumped onto the column. The pH and the conductivity of the wash step correspond to the parameters of the equilibration and loading steps. After washing, bound proteins were eluted from the column by applying a step elution using an elution buffer with increased conductivity and counter ion concentration. A product pool of $\leq 3.6^{\circ}$ CV was collected.

[00490] In the small scale Gen 2 process, an alternative gradient elution procedure was used to remove rVWF pro-peptide, small molecular weight rVWF molecules and high molecular weight molecules from the column (FIG. 45). The washing steps preceding the elution were performed as step wash with the same pH and conductivity as the starting point of the gradient elution. Four wash scenarios were tested: 0%B (10 CV), 55%B (10 CV), 40%B (5 CV) & 45%B (5 CV) and 36%B (5 CV). The corresponding gradient elution steps were 0–100%B (12 CV), 55–100%B (6 CV), 45–100% B (6C V) and 36–100%B (6 CV). The elution was completed by a 2–3 CV wash with 100% B.

[00491] The eluates were pooled according to the eluting product related impurities and product sub-species. After elution of the product the column was cleaned and sanitized with basic and acidic solutions. The main objective of this polishing step was the further removal of process related impurities including CHO host cell protein, human rFurin, media compounds like soy peptone), product related impurities (rVWF pro-peptide) and low molecular weight S/D reagents. Only a minor contribution was expected in the removal of rFVIII. Following the improved CAT (UNO_S) step, a protein concentration and buffer exchange step (ultra/diafiltration) can be required. However this ultra/diafiltration step was not part of the study described herein. The differences in the chromatographic procedure

between the 1st GEN MFG scale process and the 2nd generation small scale process are outlined in FIGS. 46–48.

6. Results

[00492] In the following section, the results of the current study are presented. The five experiments conducted at small scale clearly show that the replacement of the SEC unit operation step and the preceding ultrafiltration (buffer exchange) step is possible by the introduction of a modified UNOs (CAT) procedure.

6.1. Chromatograms

[00493] As outlined above, five experiments with different wash and gradient elution procedure were performed. The intention of the UNO S step was to find an optimal method for the removal of product and process related impurities on the one hand and to achieve an optimal yield in terms of VWF Ag and Activity. FIG. 49 shows two chromatograms of the final (5th) run VW_USS_05 are presented. The upper panel of FIG. 49 depicts the total run, including column activation, loading phase (the high UV_{280nm} absorption is caused by the S/D chemicals contained in the feed), re-equilibration, wash, gradient elution, 2M NaCl wash and the CIP procedure. The chromatogram is fused from 2 result files which explains the scale of the x-axis (result file 1: activation until end of load; result file 2: start of re-equilibration, 36% B wash, gradient elution, CIP). The lower panel of FIG. 49 depicts the elution phase in detail (step wash to 36% elution-buffer B, followed by the gradient elution 36% B to 100% B and a 100% elution-buffer B phase).

6.2. SDS-PAGE

[00494] With the variation and optimization of the chromatography conditions applied (e.g., conductivity of washes, start of gradient elution), the separation of pro-peptide and mature rVWF was refined. In addition, the removal of process related impurities and the yield of mature rVWF Ag and activity was improved. SDS-PAGE results (silver stain and anti rVWF Western blot) of the last (5th) run in the series of experiments is presented in FIG. 50.

[00495] The SDS-PAGE was performed on 3–8% Tris-Acetate gels under reducing conditions. The separated polypeptides were visualized by silver staining (top) and Western blot (bottom). Prior to loading, samples were reduced with DTT, thereafter free sulfhydryl groups were blocked with iodo acetamide. For the Western blot, the 1st antibody was a

polyclonal rabbit anti-human-VWF antibody (from Dako; order number A0082; diluted 1:1000), the 2nd antibody was a polyclonal, AP-conjugated goat anti-rabbit-IgG anti body (from Sigma; order number A-8025; diluted 1:2000). The rVWF band runs at above 250 kDa; the VWF pro-peptide runs at about 90 kDa. The pro-peptide is not detected by the antibody used for Western blotting.

[00496] Results of run VWF_USS_05 show a clear separation of pro-peptide and mature rVWF. The eluate sample (lane 16) and a reference sample purified according to the generation 1 procedure (lane 18) are highly comparable.

6.3. Multimer analysis

[00497] To assess the distribution of high and low molecular weight rVWF sub-species multimer analysis by agarose gel and Western blot was performed. Samples from Load, flowthrough (FT), Wash, Elution and high salt wash were tested (FIG. 51) LMW rVWF subspecies are contained in the flow through (FT; effluent fraction) (lane 8) and wash/ pre-elution (lanes 9 and 10). The Elution and post-elution pools (lanes 11 and 12) show a band pattern comparable to the reference sample SEC-F (lane 15). The reference sample was purified according to the generation 1 (Gen 1) process and corresponds roughly to the ascending peak of the SEC eluate pool. The high salt wash (lane 13) contains ultra-large rVWF molecules which is seen by the smear in the upper region of the lane.

[00498] The multimer analyses were performed on 1 % agarose gels according a standard protocol. Approximately 50 ng of rVWF was applied per lane and separated under non-reducing conditions in the presence of urea. The separated polypeptides were visualized by Western Blot using a rabbit anti-human VWF antibody (Dako) as 1st antibody (diluted 1:1000) and an AP-conjugated goat anti-rabbit IgG antibody (Sigma) as 2nd antibody (diluted 1:2000).

[00499] Comparing the rVWF multimer distribution between UNO_S (Gen 2) and SEC (Gen 1) runs, a reverse separation effect can be clearly seen. In the SEC procedure ultra-large and large molecules elute first (void volume), followed by the target molecules and the pro-peptide. In Gen 2 the order of separation is just the opposite (small to large). However, both methods resulted in the same rVWF multimer distribution in the eluate pool. Following the UNO_S step, a UDF (concentration/dialysis) unit operation was required to concentrate the target molecule and to transfer it into formulation buffer.

6.4 Analytical results

[00500] A summary of analytical results is given in FIG. 52-FIG. 55. Each table shows results of one specific analytical assay and contains data of all 5 runs performed in course of the study. A comparative overview of Eluate results is also presented in FIG. 56. Besides of the percentage of rVWF :Ag and Risto Co activity Eluate yields, the table contains calculated ratios to allow a direct comparison between different run setup.

6.5 Match of analytical data to success criteria

[00501] The targeted parameters of the eluate (product fraction) resulting from the modified CAT (UNO_S) unit operation step partly comply with selected BDS product specifications. As the CAT-E product pool needs to be concentrated and dialyzed to obtain BDS material, the development targets (FIG. 57) are mainly (calculated) ratios that are independent of absolute parameter concentrations.

[00502] The fact that most of the development targets were met or nearly reached demonstrates the feasibility of the suggested procedure described herein. Not all analytical assays were performed, yet key results such as rVWF:Ag and Risto yield, CHO HCP and pro-peptide impurity removal, as well as the distribution of rVWF multimers show a comparable performance of the suggested new CAT procedure and the previously applied UNO_S/SEC combination.

7. Discussion

[00503] Five UNO_S runs were performed in the course of the present study to investigate a 2nd generation CAT procedure. The results of the optimized (last) run show a separation of high from low molecular weight rVWF multimers as well as the removal of rVWF pro-peptides and CHO-HCP impurities from the target protein that is comparable to the results achieved with the Gen 1 procedure (*e.g.*, UNO_S in step elution mode + SEC step). The introduced wash step with a conductivity of about 24 mS/cm (36% Elution Buffer B) followed by a gradient elution step to about 50 mS/cm (100% Elution Buffer B) resulted in a CAT Eluate pool of comparable quality to the previously yielded Gen 1 SEC F pool. Although an additional UDF step to concentrate and dialyze the CAT eluate may be used, the Gen 2 CAT procedure described herein shows great potential to replace the UDF and SEC unit operation steps applied in the Gen 1 downstream process to obtain BDS material.

Example 13: Evaluation multimers of DF3338/042 and DF3362/023 Westernblot anti-VWF

[00504] The mat-rVWF obtained from this method was analyzed for the multimeric content. Advantages of the described cation exchange (CEX) methods includes:

- Reduction of unit operations - 1 CEX replaces 3-unit operation of the current process.
- Depletion of r-vWF-Propeptide and depletion of host cell proteins are similar to an affinity step.
- By including the SD-treatment "On colum" on cation exchanger - 4-unit operations are included in one step.
- By including the SD-treatment "On colum" and the furin maturation on cation exchanger - 5 unit operations are included in one step.
- Reduced shear stress that lowers the risk of the generation of thrombotic rVWF (due to less unit operations, filtrations and significant reduced hold times).

[00505] For this analys, western blots were run. The westernblot images were imported into Corel Photo Paint Software and converted into 16 Bit grey scale images. The 16 bit grey scale format is a requirement for the evaluation. The evaluation was made with Image Quant 1D Software.

[00506] The images were vertical flipped to simplify the evaluation (Lane numbers remain the same):

- Band 1-6 = Low molecular weight
- Band 7-12 = Intermediate molecular weight
- Band >12 = High molecularweight

[00507] Densitometric evaluation of vWF multimers of the product obtained from enhanced CEX as described herein as compared to the product obtained from the 3-unit operation process.

Table 11: Densitometric evaluation summary.

	Benchmark	VW USS 04 E	VW USS 05 E
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% Low MW	SUM Band 1-6	40,86	34,91	38,39
% Medium MW	SUM Band 7-12	40,27	39	36,87
% High MW	SUM Band >12	18,87	26,08	24,74

[00508] The raw data showing the multimer percentages is provided in Figures 61-63.

Example 14: Variant vWF Purification Process

I. BACKGROUND

[00509] r-vWF pro-peptide is a product related impurity of CHO Cell derived r-VWF product. The production cell line generates r-VWF which contains about 60% of pro-r-vWF. The r-VWF propeptide is attached to the r-vWF polypeptide covalent by peptide amide bond and additionally non-covalently by divalent cations. The covalent peptide amide bond is cleaved by in-vitro incubation with rFurin. However, the cleaved r-VWF propeptide remains attached to the VWF molecule and a method for separation of these two polypeptides is described in this example. It was discovered that the rVWF/rvWF_PP complex is stabilized by divalent cations and low pH. By applying chelator of divalent cations or high pH in combination with a proper separation method the two molecules can be separated with high efficiency and in a robust manner. As chelator low concentrations of EDTA or citrate were found to be effective and pH greater or equal pH 7 were also be seen effective when applied on cation exchange resin as wash procedure or on size exclusion chromatography when applied in the separation buffer. The same principle should be applicable to all separation technologies including ion exchange or size separation either by resins or membrane technology. In the current production process for rVWF the step SEC is performed with a running buffer containing citrate to support the separation of rVWF and rVWF-PP.

1. Description of Example Scope - VW_USS_07

1. Depletion of r-vWF-Propeptide
2. Example for alternative "SD_VI on column" treatment
3. Generating rFVIII/r-vWF complex "on column"
4. On column pre-formulation during elution of the rFVIII/r-vWF complex in an alternative formulation buffer system

Process Details:

[00510] After a monoclonal antibody step to capture recombinant factor VIII the Flow-through, which contains r-vWF, was loaded onto an Fractogel TMAE anion exchanger. r-vWF was bound on the anion exchanger and was matured with Furin in presence of Calcium. The r-vWF was eluted from the anion exchanger with increasing conductivity. The TMAE-Eluate was filtrated through a Mustang Q (Mustang Q, Pall Part Number XT5000MSTGQP1) filter unit to remove CHO-DNA and impurities that binds to the filter membrane. The product containing MUQ_Flow through was conditioned by a 1:2 dilution with [60mM sodiumcitrate pH 7.6] to a conductivity of 21.9 mS/cm and pH 7.16. The high conductivity was chosen to ensure the removal of r-vWF propeptide and low mol weight r-vWF to utilize the capacity of the resin for the desired high mol weight r-vWF. The conditioned load was loaded onto a UNOsphere™ S Cation Exchange Media (Bio Rad, Art.Nr.: 156-0115) inner diameter=10mm bed height 8.8cm volume 6.91ml with a flow rate of 100cm/h followed by a first wash(Reequilibration) of 2CV with [30mM Na-Citrate, 180mM NaCl, pH 7.5] to deplete strong bound HCP and r-vWF-Propeptide.

[00511] A potential "On column treatment" (WSD) was carried out with [30mM Na-Citrate, 180mM NaCl, pH 7.5 containing 25g/Kg of a mix of 18.0g Polysorbate 80 ,3.5g Dimethylsulfoxide DMSO ,3.5g TnBP] in 12 column volumes and a contact time of approx. 1hour to inactivate lipid enveloped viruses. The components of the "On column treatment" were washed out with Wash 2 in 10 column volumes of [30mM Na-Citrate, 180mM NaCl, pH 7.5]. By applying Wash 3 the buffer was changed from the Sodiumcitrate buffer system to a Glycine/Taurine system by applying [50mM Glycine, 10mM Taurine, 10% Sucrose, 0.1% Polysorbate 80, pH 5.5] in 4 column volumes. At step "FVIII-Con" recombinant human coagulation factor VIII derived from the ADVATE process was loaded onto the bound r-vWF in 10 column volumes.

[00512] The FVIII-Con-buffer consists of [1.57g rFVIII S2 ADV S17B010901B2 diluted in 218.67g of 50 mM Glycine, 10mM Taurine, 5% (w/w) Sucrose, 5% (w/w) D-Mannitol, 0.1% Polysorbate 80, 2 mM CaCl₂, 150 mM NaCl, and a pH 7.4]. Wash 4 was applied to wash out unbound rFVIII and to prepare the buffer matrix for the pre-formulation by applying 5 column volumes of [50 mM Glycine, 10 mM Taurine, 5% (w/w) Sucrose, 5% (w/w) D-Mannitol, 0.1% Polysorbate 80, 2 mM CaCl₂, 150 mM NaCl, pH 7.4]. Both the r-vWF and the rFVIII was eluted with [50 mM Glycine, 10mM Taurine, 5% (w/w) Sucrose,

5% (w/w) D-Mannitol, 0.1% Polysorbate 80, 2mM CaCl₂, 600mM NaCl, pH 7.4± 0.2] from the column to form an eluate. The eluate was diluted to adjust the Sodiumchloride content to approx.150 mM NaCl with [50 mM Glycine, 10 mM Taurine, 5% (w/w) Sucrose, 5% (w/w) D-Mannitol, 0.1% Polysorbate 80, 2mM CaCl₂, pH 7.4].

Process Sequence:

[00513] The sequence of the key steps of this example consists of the following steps (See, also the bottom of Figure 67 for the chromatography scheme.)

1. Mab FVIII capture (FT is the r-vWF containing fraction)
2. Fractogel TMAE capture + maturation
3. Mustang Q in FT mode
4. CEX as described (VW_USS_07)

Result:

[00514] The experiment was successfully carried out in all 4 points:

1. Depletion of r-vWF-Propeptide occurred – during the wash steps Wash 1, WSD ((W)ash with (S)olvent (D)etergent) and Wash 2 (see, Figures 67, 68, and 69.)
2. Example for alternative “SD_VI on column treatment” at step WSD.
3. Generating rFVIII/r-vWF complex “on column” – step FVIII-Con.
4. On column pre-formulation during elution of the rFVIII/r-vWF complex in an alternative formulation buffer system (see, Figure 66, last row).

Example 15: Variant vWF Purification Process – Testing for Sialylation

I. BACKGROUND

[00515] r-vWF pro-peptide is a product related impurity of CHO Cell derived r-VWF product. The production cell line generates r-VWF which contains about 60% of pro-r-vWF. The r-vWF propeptide is attached to the r-VWF polypeptide covalent by peptide amide bond and additionally non-covalently by divalent cations. The covalent peptide amide bond is cleaved by in-vitro incubation with rFurin. However, the cleaved r-VWF propeptide remains

attached to the VWF molecule and a method for separation of these two polypeptides is described in this example. The present example provides an alternate, variant embodiment for separation of the r-vWF propeptide from the r-VWF polypeptide after furin cleavage in order to test for additional sialylation. Additional details and results of the purification process are depicted in FIGS. 70-73 and 78.

1. Experiment Nr.: VW_USS_06

1. Depletion of r-vWF-Propeptide
2. Generate additional 2,6 Sialylation on column on r-vWF

2. Experiment Nr.: VW_USS_06

[00516] After a monoclonal antibody step to capture recombinant factor VIII the Flow-through, which contains r-vWF, was loaded onto an Fractogel TMAE anion exchanger. r-vWF was bound on the anion exchanger and was matured with Furin in presence of Calcium. the r-vWF was eluted from the anion exchanger with increasing conductivity. The TMAE-Eluate was filtrated trough a Mustang Q (Mustang Q, Pall Part Number XT5000MSTGQP1) filter unit to remove CHO-DNA and impurities that binds to the filter membrane. The product containing MUQ_Flow through was conditioned by a 1:2 dilution with [60mM sodiumcitrate pH 7.6] to a conductivity of 18.39mS/cm and pH 7.33. The high conductivity was chosen to ensure the removal of r-vWF propeptide and low molecular weight r-vWF to utilize the capacity of the resin for the desired high mol weight r-vWF. The conditioned load was loaded onto a UNOsphere™ S Cation Exchange Media (Bio Rad, Art. Nr.: 156-0115) inner diameter=10mm bed height 8.8cm volume 6.91ml with a flow rate of 100cm/h followed by a first wash (Reequilibration) of 2CV with [30mM Na-Citrate, 180mM NaCl, pH 7.5] to deplete strong bound HCP and r-vWF-Propeptide. To introduce additional 2,6 Sialylation a mixture of 50% (v/v) CMP-NANA Solution based on [30mM Na-Citrat, 180mM NaCl, pH 7.5] and 50% (v/v) of alpha 2,6 Sialyltransferase based on [30mM Na-Citrat, 180mM NaCl, pH 7.5] was applied onto the column in 10 column volumes and a flow rate of 25cm/h by online mixing. The composition of the CMP-NANA Solution was 11mg CMP_NANA C8271-25mg Lot.Nr.: SLBV 7777 dissolved in 154.29g [30mM Na-Citrate, 180mM NaCl, pH 7.5]. The composition of the alpha 2,6 Sialyltransferase buffer was alpha 2,6 Sialyltransferase S2076-1UN SIGMA, Lot.Nr. SLBV0552 from Photobacterium Damsela dissolved in 1ml purified water-0.5g of the dissolved alpha 2,6 Sialyltransferase was diluted

with 152.10g [30mM Na-Citrat, 180mM NaCl, pH 7.5]. A further wash with 2 column volumes of [30mM Na-Citrate, 180mM NaCl, pH 7.5] was applied to remove excess of CMP_NANA and alpha 2,6 Sialyltransferase. A buffer exchange was provided by applying 4 column volumes of [50 mM HEPES, 150 mM NaCl pH 6.0]. The Elution was performed with [50 mM HEPES, 500mM NaCl, pH 7.5] in 4 column volumes.

3. COMPLETE PURIFICATION SEQUENCE VW_USS_06

[00517] The sequence of the key steps of this example consists of the following steps:

1. Mab FVIII capture (FT is the r-vWF containing fraction)
2. Fractogel TMAE capture + maturation
3. Mustang Q in FT mode
4. CEX as described (VW_USS_06)

Result:

[00518] No additional 2,6 sialylation detected in using the method in the present example. However, 2,3 sialylation was found which is the usual sialylation pattern for r-vWF.

Example 16: Variant vWF Purification Process – Testing for Sialylation

I. BACKGROUND

[00519] r-vWF pro-peptide is a product related impurity of CHO Cell derived r-VWF product. The production cell line generates r-VWF which contains about 60% of pro-r-vWF. The r-vWF propeptide is attached to the r-VWF polypeptide covalent by peptide amide bond and additionally non-covalently by divalent cations. The covalent peptide amide bond is cleaved by in-vitro incubation with rFurin. However, the cleaved r-VWF propeptide remains attached to the VWF molecule and a method for separation of these two polypeptides is described in this example. The present example provides an alternate, variant embodiment for separation of the r-vWF propeptide from the r-VWF polypeptide after furin cleavage in order to test for additional sialylation. Additional details and results of the purification process are depicted in FIGS. 74-78.

1. Experiment Nr.: VW_USS_08

1. Depletion of r-vWF-Propeptide
2. Generate additional 2,6 Sialylation on column on r-vWF

2. Experiment Nr.: VW_USS_08

[00520] After a monoclonal antibody step to capture recombinant factor VIII the Flow-through, which contains r-vWF, was loaded onto an Fractogel TMAE anion exchanger. r-vWF was bound on the anion exchanger and was matured with Furin in presence of Calcium. the r-vWF was eluted from the anion exchanger with increasing conductivity. The TMAE-Eluate was filtrated trough a Mustang Q (Mustang Q, Pall Part Number XT5000MSTGQP1) filter unit to remove CHO-DNA and impurities that binds to the filter membrane. The product containing MUQ_Flow through was conditioned by a 1:2 dilution with [60mM sodium citrate pH 7.6] to a conductivity of 19.97mS/cm and pH 7.33. The high conductivity was chosen to ensure the removal of r-vWF propeptide and low mol weight r-vWF to utilize the capacity of the resin for the desired high mol weight r-vWF. The conditioned load was loaded onto a UNOsphere™ S Cation Exchange Media (Bio Rad, Art.Nr.: 156-0115) inner diameter=10mm bed high 8.8cm volume 6.91ml with a flow rate of 100cm/h followed by a first wash (Reequilibration) of 2CV with [30mM Na-Citrate, 180mM NaCl, pH 7.5] to deplete strong bound HCP and r-vWF-Propeptide. To introduce additional 2,6 Sialylation a mixture of 50%(v/v) CMP-NANA Solution based on [30mM Na-Citrat, 180mM NaCl, pH 7.5] and 50%(v/v) of alpha 2,6 Sialyltransferase based on [30mM Na-Citrat, 180mM NaCl, pH 7.5] was applied onto the column in 10 column volumes and a flow rate of 25cm/h by online mixing. The composition of the CMP-NANA Solution was 14mg CMP_NANA C8271-25mg Lot. Nr.: SLBV 7777 dissolved in 121,57g [30mM Na-Citrat, 180mM NaCl, pH 7.5]. The composition of the alpha 2,6 Sialyltransferase buffer was alpha 2,6 Sialyltransferase S2076-1UN SIGMA, Lot.Nr. SLBV0552 from Photobacterium Damsela dissolved in 121.10g [30mM Na-Citrat, 180mM NaCl, pH 7.5]. A further wash with 2 column volumes of [30mM Na-Citrate, 180mM NaCl, pH 7.5] was applied to remove excess of CMP_NANA and alpha 2,6 Sialyltransferase. A buffer exchange was provided by applying 4 column volumes of [50mM HEPES, 150mM NaCl pH 6.0]. The Elution was performed with [50mM HEPES, 500mM NaCl, pH 7.5] in 4 column volumes.

3. COMPLETE PURIFICATION SEQUENCE VW_USS_08

[00521] The sequence of the key steps of this example consists of the following steps:

1. Mab FVIII capture (FT is the r-vWF containing fraction)
2. Fractogel TMAE capture + maturation
3. Mustang Q in FT mode
4. CEX as described (VW_USS_08)

Result:

[00522] No additional 2,6 sialylation detected using the method in the present example. However, 2,3 sialylation was found which is the usual sialylation pattern for r-vWF.

[00523] The examples set forth above are provided to give those of ordinary skill in the art a complete disclosure and description of how to make and use the embodiments of the compositions, systems and methods of the invention, and are not intended to limit the scope of what the inventors regard as their invention. Modifications of the above-described modes for carrying out the invention that are obvious to persons of skill in the art are intended to be within the scope of the following claims. All patents and publications mentioned in the specification are indicative of the levels of skill of those skilled in the art to which the invention pertains. All references cited in this disclosure are incorporated by reference to the same extent as if each reference had been incorporated by reference in its entirety individually.

[00524] All headings and section designations are used for clarity and reference purposes only and are not to be considered limiting in any way. For example, those of skill in the art will appreciate the usefulness of combining various aspects from different headings and sections as appropriate according to the spirit and scope of the invention described herein.

[00525] All references cited herein are hereby incorporated by reference herein in their entireties and for all purposes to the same extent as if each individual publication or patent or

patent application was specifically and individually indicated to be incorporated by reference in its entirety for all purposes.

[00526] Many modifications and variations of this application can be made without departing from its spirit and scope, as will be apparent to those skilled in the art. The specific embodiments and examples described herein are offered by way of example only, and the application is to be limited only by the terms of the appended claims, along with the full scope of equivalents to which the claims are entitled.

WHAT IS CLAIMED IS:

1. A method for obtaining a composition comprising a high purity, propeptide depleted mature recombinant rVWF (mat-rVWF), said method comprising the steps of:
 - a) providing a solution comprising mat-rVWF/ rVWF-PP complex, mat-rVWF, and rVWF propeptide (rVWF-PP);
 - b) inducing dissociation of said mat-rVWF/ rVWF-PP complex in said solution in a) into mat-rVWF and rVWF-PP, wherein said dissociation occurs by disruption of the non-covalently associated mat-rVWF and rVWF-PP, wherein said dissociation is induced by:
 - i. addition of at least one chelating agent, or
 - ii. increasing the pH to a pH of at least 7; and
 - c) collecting said mat-rVWF to obtain a high purity, mat-rVWF composition, wherein said high purity, mat-rVWF composition comprises at least 95% mature rVWF and less than 5% rVWF-PP.
2. The method of claim 1, wherein said high purity, mat-rVWF composition comprises at least 96% mat-rVWF and less than 4% rVWF-PP, at least 97% mat-rVWF and less than 3% rVWF-PP, at least 98% mat-rVWF and less than 2% rVWF-PP, at least 99% mat-rVWF and less than 1% rVWF-PP, or at least 99.5% mat-rVWF and less than 0.5% rVWF-PP, or 99.9% mat-rVWF and less than 0.1% rVWF-PP.
3. The method according to any of the preceding claims, wherein said solution is selected from the group consisting of a cell culture medium, an antibody column flow-through solution, and a buffered solution.
4. The method according to any of the preceding claims, wherein said solution has been treated with furin prior to step a).
5. The method according to any of the preceding claims, wherein said solution is an antibody column flow-through solution.

6. The method according to any of the preceding claims, wherein said at least one chelating agent is a divalent cation chelating agent.
7. The method according to claim 6, wherein said divalent cation chelating agent is selected from the group consisting of EDTA, EGTA, CDTA, and citrate.
8. The method according to any of the preceding claims, wherein said pH is increased to at least 7.1, 7.2, 7.3, 7.4, 7.5, 7.6, 7.7, 7.8, 7.9, 8.0, 8.1, 8.2, 8.3, 8.4, 8.5, 8.6, 8.7, 8.8, 8.9, or 9.0.
9. The method according to any of the preceding claims, wherein said pH is increased to at least about 7.2 to about 7.8.
10. The method according to any of the preceding claims, wherein said pH is increased to at least about 7.6.
11. The method according to any of the preceding claims, wherein said pH is increased by the addition of basic amino acids, Tris, NaOH, Tricine, or ethanolamine.
12. The method according to any of the preceding claims, wherein said collecting in step b) of claim 1 comprises one or more protein separation methods.
13. The method according to claim 12, wherein said one or more protein separation methods is selected from the group consisting of ion exchange chromatography (IEC), size exclusion chromatography (SEC), physical size separation by membrane technology, and affinity chromatography.
14. The method according to claim 13, wherein said protein separation method is size exclusion chromatography (SEC).
15. The method according to claim 13, wherein said one or more protein separation method is ion exchange chromatography (IEC).

16. The method according to claim 15, wherein said ion exchange chromatography (IEC) is cation exchange chromatography.
17. The method according to claim 15, wherein said ion exchange chromatography (IEC) is a combination of anion exchange chromatography and cation exchange chromatography.
18. The method according to claim 12, wherein said one or more protein separation methods comprise a buffer system, wherein said buffer system comprises one or more buffers.
19. The method according to claim 18, wherein said one or more buffers includes wash buffers, wherein said one or more wash buffers include one, two, three, four, and/or five wash buffers, wherein when said one or more buffers includes five wash buffers, the first, second, third, and/or fifth wash buffers have a higher pH than the fourth wash buffer, and when said one or more buffers includes four wash buffers, the first, second, and/or fourth wash buffers have a higher pH than the third wash buffer.
20. The method according to claim 19, wherein the method further comprises a viral inactivation treatment step after the first wash buffer, and optionally the pH of the viral inactivation treatment step has a higher pH than said third and/or fourth wash buffer.
21. The method according to claim 18, wherein said one or more buffers comprise said one or more chelating agents.
22. The method according to claim 18, wherein said one or more buffers exhibit a pH of at least 7.
23. The method according to claim 12, wherein said one or more protein separation methods comprise a buffer system, wherein said buffer system comprises one or more loading buffers.

24. The method according to claim 23, wherein said one or more loading buffers comprise said one or more chelating agents.
25. The method according to claims 23 or 24, wherein said one or more loading buffers exhibit a pH of at least 7.
26. The method according to claim 12, wherein said one or more protein separation methods comprise a buffer system, wherein said buffer system comprises one or more load, wash, and/or elution buffers.
27. The method according to claim 26, wherein said one or more load, wash, and/or elution buffers comprise said one or more chelating agents.
28. The method according to claims 26 or 27, wherein said one or more load, wash, and/or elution buffers exhibit a pH of at least 7.
29. The method according to any of claims 26 through 28, wherein said one or more load, wash, and/or elution buffers comprise said one or more chelating agents and exhibit a pH of at least 7.
30. The method according to any of claims 18 through 29, wherein said buffering system is selected from the group consisting of glycine HEPES (4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid), TrisHCl (Tris(hydroxymethyl)-aminomethane), histidine, imidazole, acetate citrate, MES, and 2-(N-morpholino)ethanesulfonic acid.
31. The method according to any of claims 18 through 30, wherein said buffer further comprises one or more monovalent cations.
32. The method according to claim 31, wherein said one or more monovalent cations are selected from the group consisting of Na⁺, K⁺, Li⁺, and Cs⁺.
33. The method according to claim 31, wherein said monovalent cation is Na⁺.

34. The method according to any of claims 18 through 33, wherein said buffer further comprises one or more monovalent, divalent and/or trivalent anions.
35. The method according to claim 34, wherein said one or more monovalent, divalent and/or trivalent anions are selected from the group consisting of Cl^- , acetate⁻, SO_4^{2-} , Br^- , and citrate³⁻.
36. The method according to any of claims 18 through 35, wherein said buffer system comprises at least one buffer exhibiting a conductivity of ≥ 0.5 mS/cm at 25°C.
37. The method according to any of claims 18 through 36, wherein said buffer system comprises at least one buffer exhibiting a conductivity of 15.0 ± 0.2 mS/cm at 25°C.
38. The method according to any of claims 18 through 37, wherein said buffer further comprises one or more nonionic detergents.
39. The method according to claim 38, wherein said nonionic detergent is selected from the group consisting of Triton X 100, Tween 80, and Tween 20.
40. The method according to any of claims 18 through 39, wherein said buffer further comprises one or more additional substances selected from the group consisting of non-reducing sugars, sugar alcohols, and polyols.
41. The method according to any of the preceding claims, wherein said high purity mat-rVWF composition comprises a host cell (HC) impurity level of ≤ 2.0 %.
42. The method according to any of the preceding claims, wherein said high purity, mat-rVWF composition comprises a host cell (HC) impurity level of ≤ 0.6 %.
43. The method according to any of the preceding claims, wherein said solution comprising mat-rVWF/ rVWF-PP complex, mat-rVWF, and rVWF-PP is derived from a capture step for rVWF.

44. The method according to any of the preceding claims, wherein said solution comprising mat-rVWF/ rVWF-PP complex, mat-rVWF, and rVWF-PP is derived from a method comprising a FVIII immunoaffinity step and anion exchange chromatography step.
45. A method for obtaining a composition comprising a high purity, propeptide depleted mature recombinant rVWF (high purity mat-rVWF), said method comprising the steps of:
- a) loading a solution comprising pro-rVWF, mat-rVWF/ rVWF-PP complex, mat-rVWF, and/or rVWF propeptide (rVWF-PP) onto an anion exchange column, wherein said pro-rVWF, mat-rVWF/ rVWF-PP complex, and mat-rVWF are bound to said anion exchange column;
 - b) washing said anion exchange column in a) containing said bound pro-rVWF, mat-rVWF/ rVWF-PP complex, and mat-rVWF with one or more wash buffers;
 - c) treating said column in b) comprising the bound pro-rVWF, mat-rVWF/ rVWF-PP complex, and mat-rVWF with furin, wherein said furin cleaves said pro-rVWF into mat-rVWF and rVWF-PP;
 - d) eluting said bound pro-rVWF, mat-rVWF/ rVWF-PP complex, and mat-rVWF from the column in c) with an elution buffer, wherein said elution buffer induces dissociation of said rVWF-PP from mat-rVWF non-covalently associated with said rVWF-PP, and wherein said dissociation is induced by:
 - i. addition of at least one chelating agent into said elution buffer, or
 - ii. increasing the pH of said elution buffer to a pH of at least 7; and
 - e) collecting said mat-rVWF separately from said rVWF-PP to obtain a high purity mat-rVWF composition, wherein said high purity mat-rVWF composition comprises at least 95% mature rVWF and less than 5% rVWF-PP.
46. The method according to claim 45, wherein said one or more buffers includes wash buffers, wherein said one or more wash buffers include one, two, three, four, and/or five wash buffers, wherein when said one or more buffers includes five wash buffers, the first, second, third, and/or fifth wash buffers have a higher pH than the fourth

wash buffer, and when said one or more buffers includes four wash buffers, the first, second, and/or fourth wash buffers have a higher pH than the third wash buffer.

47. The method according to claim 46, wherein the method further comprises a viral inactivation treatment step after the first wash buffer, and optionally the pH of the viral inactivation treatment step has a higher pH than said third and/or fourth wash buffer.
48. The method according to any of claims 45 through 47, wherein a) and b) occur simultaneously in a single step.
49. The method according to any of claims 45 through 48, wherein said solution in a) comprises the flow through from a monoclonal antibody column, wherein said monoclonal antibody is a FVIII monoclonal antibody.
50. The method according to any of claims 45 through 49, wherein said solution in a) is selected from the group consisting of a cell culture medium, an antibody column flow-through solution, and a buffered solution.
51. The method according to any of claims 45 through 50, wherein said at least one chelating agent is a divalent cation chelating agent.
52. The method according to claim 51, wherein said divalent cation chelating agent is selected from the group consisting of EDTA, EGTA, CDTA, and citrate.
53. The method according to any of claims 45 through 52, wherein said pH is increased to at least 7.1, 7.2, 7.3, 7.4, 7.5, 7.6, 7.7, 7.8, 7.9, or 8.0.
54. The method according to any of claims 45 through 53, wherein said pH is increased to at least about 7.2 to about 7.8.
55. The method according to any of claims 45 through 54, wherein said pH is increased to at least about 7.6.

56. The method according to any of claims 45 through 55, wherein said pH is increased by the addition of basic amino acids.
57. The method according to any of claims 45 through 55, wherein said one or more wash buffers in b) comprise said one or more chelating agents.
58. The method according to any of claims 45 through 55, wherein said one or more wash buffers in b) exhibit a pH of at least 7.
59. The method according to any of claims 45 through 55, wherein said one or more wash buffers in b) comprise said one or more chelating agents and exhibit a pH of at least 7.
60. The method according to any of claims 45 through 55, wherein said method further comprises a step of viral inactivation, wherein said viral inactivation occurs before, after, or concurrently with the washing step and/or the elution step, but before the collecting step.
61. The method according to claim 60, wherein said viral inactivation treatment inactivates lipid enveloped viruses.
62. The method according to claims 60 or 61, wherein said viral inactivation treatment is an S/D treatment.
63. The method according to any of claims 45 through 62, wherein said one or more buffers comprise a buffer selected from the group consisting of glycine HEPES (4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid), TrisHCl (Tris(hydroxymethyl)-aminomethane), histidine, imidazole, acetate citrate, MES, and 2-(N-morpholino)ethanesulfonic acid.
64. The method according to any of claims 45 through 63, wherein said one or more buffers further comprise one or more monovalent cations.

65. The method according to claim 64, wherein said one or more monovalent cations are selected from the group consisting of Na⁺, K⁺, Li⁺, and Cs⁺.
66. The method according to claim 65, wherein said monovalent cation is Na⁺.
67. The method according to any of claims 45 through 66, wherein said one or more buffers further comprise one or more monovalent, divalent, and/or trivalent anions.
68. The method according to claim 67, wherein said one or more monovalent, divalent and/or trivalent anions are selected from the group consisting of Cl⁻, acetate⁻, SO₄²⁻, Br⁻, and citrate³⁻.
69. The method according to any of claims 45 through 68, wherein said one or more buffers comprise at least one buffer exhibiting a conductivity of ≥ 0.5 mS/cm at 25°C.
70. The method according to any of claims 45 through 69, wherein said one or more buffers comprise at least one buffer exhibiting a conductivity of 15.0 ± 0.2 mS/cm at 25°C.
71. The method according to any of claims 45 through 70 wherein said one or more buffers further comprise one or more nonionic detergents.
72. The method according to claim 71, wherein said nonionic detergent is selected from the group consisting of Triton X 100, Tween 80, and Tween 20.
73. The method according to any of claims 45 through 72, wherein said one or more buffers further comprise one or more additional substances selected from the group consisting of non-reducing sugars, sugar alcohols, and polyols.
74. The method according to any of claims 45 through 73, wherein said high purity mat-rVWF composition comprises a host cell (HC) impurity level of ≤ 2.0 %.

75. The method according to any of claims 45 through 74, wherein said high purity mat - rVWF composition comprises a host cell (HC) impurity level of $\leq 0.6\%$.
76. The method according to any of the preceding claims, wherein said high purity mat - rVWF composition is used for the production of a pharmaceutical composition.
77. A pharmaceutical composition comprising a high purity mat-rVWF generated by the method according to any of the preceding claims and a pharmaceutically acceptable buffer.
78. The pharmaceutical composition according to claim 77, wherein the composition comprises 50 mM Glycine, 10 mM Taurine, 5% (w/w) Sucrose, 5% (w/w) D-Mannitol, 0.1% Polysorbate 80, 2 mM CaCl_2 , 150 mM NaCl, wherein said composition has a pH of about pH 7.4.