

**RECOGNITION OF CELLULAR TARGET BINDING BY A BIOACTIVE AGENT
USING INTRACELLULAR BIOLUMINESCENCE RESONANCE ENERGY
TRANSFER**

CROSS-REFERENCE TO RELATED APPLICATIONS

The present application claims priority to U.S. Provisional Patent Application Serial No. 61/736,429 filed December 12, 2012; U.S. Provisional Patent Application Serial No. 61/794,461 filed March 15, 2013; and U.S. Provisional Patent Application Serial No. 61/880,048 filed September 19, 2013; each of which is hereby incorporated by reference in its entirety.

FIELD

The present invention provides compositions and methods for detection and analysis of intracellular binding of a bioactive agent to a cellular target. In particular, provided herein are bioactive agents tethered to fluorophores, cellular targets fused to bioluminescent reporters, or portions, components, or subunits of bioluminescent reporters, and methods of detecting and analyzing the interaction of bioactive agents with cellular targets therewith.

BACKGROUND

The interaction of molecular species with cellular targets is critically important in understanding cellular physiology and developing therapeutic interventions, such as new synthetic drugs and biopharmaceuticals. Methods are needed for accurately and efficiently determining target engagement, particularly within living cells where these interactions mediate their phenotypic responses. The ability to affectively interrogate target engagement has broad implications to the discovery process, ranging from high-throughput screening, optimization of screening hit into drug leads, and the discovery and characterization of therapeutically relevant cellular targets.

Phenotypic-based screening with a small molecule library plays an important role in the drug discovery field. Using such screening approaches, compound libraries, without prior knowledge of their underlying cellular targets, are screened for their ability to elicit a phenotypic response, e.g., mitigate disease symptoms. While this approach can be used to identify bioactive agents, e.g., small molecules, that are able to modulate cellular physiology, determining the biological relevant targets of these small molecule hits is a major technical challenge. In addition, small molecules promoting some desirable phenotypic responses may pose *in vivo* liabilities due to off-target interactions. In order to predict drug selectivity and minimize potential side effects,

it is important to also identify off-target interactions (e.g., lower affinity). Most methods used today for identifying the targets of bioactive agents rely on the enrichment of these targets from complex cellular lysate using “bi-functionalized” compounds that contain a selective moiety (e.g., the bioactive agent or related compound) and a sorting moiety (e.g., affinity tag or solid support). As the enrichment is based on the binding properties of the compounds, where the inherent affinity of these compounds for their target is insufficient, compound analogs are designed to covalently bind to the target (e.g., photo-crosslink). By either approach, the efficacy and specificity of target isolation is essential, and the failure rate of these methods is high. Failure of these approaches can be due to insufficient capture of the target or high background due to non-specific capture. Contributing factors to this failures include: compounds binding multiple targets with low to moderate affinity with these relative weak interactions difficult to detect; lack of robust, straightforward, unbiased technologies to characterize the detected interactions; inability to perform target isolation within the native cellular environment upon which interactions may depend; limited information provided about the binding potency of targets in the cell; and high background of false positive interactions due to non-specific binding to the solid support or functionalized small molecule.

SUMMARY

In some embodiments, the present disclosure provides BRET assay systems comprising: (a) a bioactive agent tethered to a chromophore (e.g., fluorophore); (b) a cellular target fused to a bioluminescent reporter; and (c) a substrate for the bioluminescent reporter. In an aspect, the present disclosure provides an assay system comprising: (a) a small molecule bioactive agent tethered to a small molecule fluorophore; (b) a cell that expresses a protein fusion of: a cellular target and a bioluminescent reporter, wherein the small molecule bioactive agent is capable of non-covalently binding to the cellular target upon interaction therewith, wherein the emission spectrum of the bioluminescent reporter overlaps with the absorption spectrum of the fluorophore; and (c) a substrate for the bioluminescent reporter. In some embodiments, the bioactive agent is a small molecule. In some embodiments, the bioactive agent is an inhibitor of protein function, e.g., an enzyme inhibitor or a receptor inhibitor. In some embodiments, the chromophore is a fluorophore. In some embodiments, the fluorophore is a carboxy rhodamine analog. In some embodiments, the bioluminescent reporter comprises a polypeptide with at least 70% sequence identity (e.g., 75% identity... 80% identity... 85% identity... 90% identity, 95% identity... 98% identity... 99% identity) with SEQ ID NO: 1. In some embodiments, (b) is a cellular target fused to a portion, or subunit, or component of a bioluminescent reporter. In some embodiments, (b) is a cellular target fused to a polypeptide that requires interaction with another polypeptide to produce luminescence. In some embodiments, (a) and (b) are within a cell. In some embodiments, (b) is expressed intracellularly as a fusion protein with a protein of interest, e.g., cellular target. In some embodiments, the cellular target is composed of more than one component, subunit or polypeptide, e.g., the cellular target is a protein complex. In some embodiments, the bioluminescent reporter is composed of more than one component, subunit or polypeptide, e.g., the bioluminescent reporter is a protein complex. In

some embodiments, (a) is added extracellularly and enters the cell. In some embodiments, (a) is present both within a cell and in the medium surrounding the cell. In some embodiments, (a) is present both bound to a cell and in the medium surrounding the cell. In some embodiments, the amount of (a) present in the surrounding medium is significantly greater than the amount in the cell or bound to the cell, e.g., greater by at least 2 fold, at least 5 fold, 10 fold, 30 fold, or 100 fold. In some embodiments, the cellular target is a binding partner of the bioactive agent. In some embodiments, the emission spectrum of the bioluminescent reporter overlaps with the absorption spectrum of a fluorophore. In some embodiments, upon binding of the bioactive agent to the cellular target, conversion of the substrate to a reaction product by the bioluminescent reporter results in excitation of a fluorophore by BRET and light emission from a fluorophore. In some embodiments, (a) is one of a library of agents or compounds tethered to a chromophore. In some embodiments, (a) is one of a library of agents or compounds tethered to fluorophores. In some embodiments, (b) is one of a plurality of potential cellular targets fused to a bioluminescent reporter. In some embodiments, (b) is a library of cells expressing one of a plurality of potential cellular targets fused to a bioluminescent reporter.

In some embodiments, the present disclosure provides methods of detecting, analyzing, characterizing, etc. the binding of a bioactive agent to a cellular target. In an embodiment, the present disclosure provides a method of detecting the binding of a bioactive agent to a cellular target comprising the system as disclosed herein. In some embodiments, the cellular target may be the primary drug target. In some embodiments, the cellular target is an off-target liability that may cause undesirable side effects *in vivo*. In some embodiments, binding by the bioactive agent to the cellular target may have no discernible biological effect. In some embodiments, the present disclosure provides cells comprising one or more of (e.g., each of) (a) a bioactive agent tethered to a chromophore (e.g., fluorophore); (b) a cellular target fused to a bioluminescent reporter; and (c) a substrate for the bioluminescent reporter. In an embodiment, the present disclosure provides a cell comprising the system as disclosed herein. In some embodiments, the binding of a bioactive agent to a cellular target is non-covalent. In some embodiments, the chromophore is a fluorophore. In an aspect, the present disclosure provides methods for detection of an interaction between a small molecule bioactive agent and a cellular target comprising: (a) expressing in a cell a fusion of said cellular target and a bioluminescent reporter, the bioluminescent reporter having a first emission spectrum; (b) contacting said cell extracellularly with a conjugate of said small molecule bioactive agent tethered to a small molecule fluorophore, said small molecule fluorophore having an excitation spectrum that overlaps said first emission spectrum and having a second emission spectrum that is distinct from said first emission spectrum, wherein the conjugate is cell permeable, and wherein the small molecule bioactive agent is capable of binding non-covalently to the cellular target upon interaction therewith; (c) contacting said cell with a substrate for said bioluminescent reporter, wherein the substrate is cell permeable; (d) detecting fluorescence at within said second emission spectrum, wherein the intensity of said fluorescence within the second emission spectrum correlates with binding between said small molecule bioactive agent with said cellular target.

In some embodiments, the present disclosure provides methods for detection of an interaction between a bioactive agent and a cellular target comprising: (a) providing a fusion of said cellular target and a bioluminescent reporter; (b) contacting said fusion with said bioactive agent tethered to a chromophore (e.g., fluorophore); (c) contacting said fusion with a substrate for said reporter; (d) detecting a change in the spectral distribution of emitted light relative to said fusion contacted with said substrate in the absence of said chromophore. In some embodiments, the present disclosure provides a method for detection of an interaction of a second bioactive agent and a cellular target comprising: (a) providing a fusion of said cellular target and a bioluminescent reporter; (b) contacting said fusion with both a first bioactive agent tethered to a chromophore (e.g., fluorophore) and with said second bioactive agent; (c) contacting said fusion with a substrate for said reporter; (d) detecting a change in the spectral distribution of emitted light relative to said fusion contacted with said first bioactive agent and said substrate in the absence of said second bioactive agent. In some embodiments, the change in spectral distribution of emitted light is caused by displacement of said first bioactive agent by said second bioactive agent. In some embodiments, said displacement is competitive displacement. In some embodiments, the change in spectral distribution of emitted light is used to estimate binding affinity of the bioactive agent to cellular target. In some embodiments, said second bioactive agent is one of a plurality of bioactive agents. In some embodiments, the change in spectral distribution of emitted light is used to estimate the relative binding affinity of a plurality of bioactive agents to a cellular target. In some embodiments, said first and said second bioactive agents are synthetic molecules. In some embodiments, said cellular targets are in living cells, permeabilized cells or cell lysates. In some embodiments, the change in the spectral distribution of emitted light is determined by measuring the ratio of light intensity at two different wavelengths, or two different ranges of wavelengths. In some embodiments, the change in spectral distribution over time is detected. In some embodiments, the chromophore is a fluorophore.

In some embodiments, the present disclosure provides a BRET assay system comprising: (a) a bioactive agent tethered to a fluorophore; (b) a first interaction partner fused to a structurally complementary peptide of a bioluminescent reporter; (c) a second interaction partner fused to a structurally complementary polypeptide of a bioluminescent reporter; and (d) a substrate for the bioluminescent reporter, wherein the first and second interaction partners interact to form an interaction complex, and wherein the first interaction partner, second interaction partner, and/or interaction complex are binding partners for the bioactive agent. In some embodiments, the first interaction partner and the second interaction partner are proteins or polypeptides that interact to form a protein complex. In some embodiments, the first interaction partner and the second

interaction partner are brought together by the interaction of the structurally complementary polypeptides. In some embodiments, the structurally complementary polypeptides are brought together by the interaction of first interaction partner and the second interaction partner. In some embodiments, the interaction of the first interaction partner and the second interaction partner is determined by an increase in luminescence. In some embodiments, the first and second interaction partners form the interaction complex in the presence or absence of the bioactive agent. In some embodiments, the interaction complex is a binding partner for the bioactive agent, but neither the first or second interaction partners alone are binding partners for the bioactive agent. In some embodiments, one of the interaction partners, but not the other is a binding partner for the bioactive agent. In some embodiments, interaction complex formation requires binding of the bioactive agent to an interaction partner. In some embodiments, interaction complex formation is independent of binding of the bioactive agent. In some embodiments, the present disclosure provides BRET assay systems comprising: (a) a bioactive agent tethered to a fluorophore; (b) a cellular target fused to a structurally complementary peptide of a bioluminescent reporter; (c) a structurally complementary polypeptide of a bioluminescent reporter; and (d) a substrate for the bioluminescent reporter. In some embodiments, the complementary peptide and polypeptide of the bioluminescent reporter associate to form active bioluminescent reporter enzyme. In some embodiments, upon binding of the bioactive agent to the cellular target and association of the complementary peptide and polypeptide, conversion of the substrate to a reaction product by the bioluminescent reporter enzyme results in excitation of the fluorophore by BRET and fluorescence emission from the fluorophore.

In some embodiments, the present disclosure provides an assay system comprising: (a) a bioactive agent tethered to a fluorophore; (b) a first binding partner fused to a complementary peptide of a bioluminescent reporter; (c) a second binding partner fused to a complementary polypeptide of a bioluminescent reporter; and (c) a substrate for the bioluminescent reporter. In some embodiments, the complementary peptide and polypeptide of the bioluminescent reporter associate to form active bioluminescent reporter enzyme. In some embodiments, when the first and second binding partners interact, the bioactive agent binds to the interacting partners and association of the complementary peptide and polypeptide occurs, conversion of the substrate to a reaction product by the bioluminescent reporter enzyme results in excitation of the fluorophore by BRET and fluorescence emission from the fluorophore.

In some embodiments, the bioactive agent is tethered to a fluorescence or luminescence quencher (e.g., Dabcyl). In such embodiments, titration of the untethered bioactive agent produces a gain in signal.

In some embodiments, the present disclosure provides imaging (e.g., BRET imaging, via charge couple device camera, etc.) of any of the systems described herein to confirm and/or identify the location (e.g., intracellular, extracellular, etc.) of fluorescence and/or BRET signals resulting from the presence and/or interactions of system components (e.g., bioactive agent, cellular target, etc.).

In some embodiments systems and methods described herein (See, e.g., Example 23 for an exemplary embodiment) are useful for the measurement of intracellular selectivity and affinity of a bioactive agent against a panel of putative cellular targets. In some embodiments, the effect on BRET is monitored as the bioactive agent/cellular target complex is competitively disrupted. In some embodiments, affinities are inferred by the IC₅₀ value generated via competitive disruption. In some embodiments, the affinities of inhibitors to individual domains of a cellular target are determined via genetic fusion of a bioluminescent reporter to a segregated domain of the cellular target.

DRAWINGS

Figure 1 shows a schematic representation of an embodiment of the present invention: (A) fluorophore-tethered bioactive agent and bioluminescent-reporter-tethered cellular target; (B) binding of bioactive agent to cellular target; (C) addition of reporter substrate results in BRET; and (D) displacement by excess untethered bioactive agent results in loss of BRET and fluorescence.

Figure 2 shows graphs comparing the BRET profiles of two different fluorophore-SAHA derivatives as BRET acceptors for NANOLUC-HDAC6 fusions in live cells.

Figure 3 shows graphs depicting the effect of dilution of expression on assay performance. The first graph demonstrates the effect of lowered expression on the BRET assay window using competitive displacement of fluorophore-tethered SAHA from NANOLUC-fused HDAC6. The second graph shows the effect of lowered expression on both BRET assay window, as well as observed compound potency (EC₅₀) for a drug tracer binding to NanoLuc-Histamine H1 receptor.

Figure 4 shows graphs demonstrating the requirement of the cellular milieu for pro-drug processing.

Figure 5 shows graphs demonstrating the ability to use permeabilization agents to potentiate the entry of impermeable drug tracers.

Figure 6 shows a graph depicting competitive displacement of fluorophore-tethered BIRB-derivative from NANOLUC-fused p38.

Figure 7 shows graphs depicting measurement of the kinetics of fluorophore-tethered BIRB derivative from NANOLUC-fused p38.

Figure 8 shows exemplary fluorophore-tethered bioactive agents and other compounds that find use in embodiments described herein.

Figure 9 shows a comparison of p38 fusions with Renilla luciferase and NanoLuc luciferase within an embodiment described herein.

Figure 10 shows that high affinity interaction with a drug tracer with optimal signal to background is achieved when expression of the NANOLUC fusion is close to endogenous level.

Figure 11 shows dose-response curves for NanoLuc-p38alpha binding to a BIRB-TOM conjugate compared to a BIRB-TMR conjugate.

Figure 12 shows dose-response curves for PKCalpha-NanoLuc binding to a BIM-TOM conjugate compared to a BIM-TMR conjugate.

Figure 13 shows a graph depicting detection of specific BRET response in live cells.

Figure 14 shows affinity of PBI-4838 to Jnk2, p38beta, and p38alpha.

Figure 15 shows affinity of PBI-4838 to Jnk2, p38beta, and p38alpha.

Figure 16 shows relative affinities of PBI-4966 to wild-type BRD4 versus the mutant BRD4.

Figure 17 shows structurally-distinct compounds can be identified by competitive displacement of drug tracers using the method of the present invention.

Figure 18 shows the ability to monitor drug/target interactions using fluorophore drug conjugate in cells.

Figure 19 shows graphs depicting a specific BRET response in TNT.

Figure 20 shows graphs depicting measurement of binding of fluorescently-labeled cytokines to NanoLuc-EGFR. Affinities of the therapeutic antibodies, Vectibix, Erbitux and Herceptin, are also demonstrated by competitive displacement of the cytokine.

Figure 21 contains wavelength scans for a complementary polypeptide (SEQ ID NO: 5) complemented with either a complementary peptide alone (SEQ ID NO: 4) or the complementary peptide (SEQ ID NO: 4) conjugated to TMR demonstrating efficient energy transfer.

Figure 22 contains wavelength scans for NanoLuc fused to HaloTag (NL-HT) and a complementary polypeptide (SEQ ID NO: 6) complemented with a complementary peptide linked to a fluorescent Non-ChloroTOM (NCT) dye (PBI-5074).

Figure 23 shows a schematic of a tertiary interaction wherein the energy transfer with a complex of a structurally complementary peptide and polypeptide of a bioluminescent protein can also be used to measure interaction of interaction partners. In the schematic, a GPCR (first interaction partner) fused to a complementary polypeptide of a bioluminescent protein and a GPCR interacting protein (second interaction partner) fused to a complementary peptide of a bioluminescent protein form a bioluminescent complex when they interact (to form an interaction complex). This allows measurement of the binary interaction. If a small molecule GPCR ligand bearing a fluorescent moiety for energy transfer interacts with this system, energy transfer occurs. Therefore, the binary protein-protein interaction and the ternary drug-protein-protein interaction can be measured in the same experiment.

Figure 24 shows graphs demonstrating that BRET data between a cellularly expressed fusion of 11S peptide and BRD4 and exogenously added NCT conjugated to BRD4 ligand is dependent upon the presence of PEP-80 (the structural complement to 11S); (A) donor intensity, (B) acceptor intensity.

Figure 25 shows graphs demonstrating that BRET data between a cellularly expressed fusion of 11S peptide and BRD4 and exogenously added NCT conjugated to iBET is dependent upon the presence of PEP-80 (the structural complement to 11S); (A) 1 μ M PEP-80, (B) no PEP-80 control.

Figure 26 shows BRET signals (acceptor and donor intensities) dependent on facilitated 11S-BRD4/114-Histone H3.3 complex.

Figure 27 shows that a fluorescent BRD4 ligand (iBET-NCT/PBI-4966) can displace the 11S-BRD4/114-Histone H3.3 complex.

Figure 28 shows non-fluorescent iBET-151 IC50 competitive displacement of the 11S-BRD4/114-Histone H3.3 complex.

Figure 29 shows that Class I HDACs (HDAC1, 2, 3, and 8) generate specific BRET signals with SAHA-TOM, as predicted based on the reported affinity and selectivity of SAHA to these individual targets.

Figure 30 shows that Class IIb HDACs (HDAC6 and 10), as well as segregated domains of HDAC6, generate specific BRET with SAHA-TOM, as predicted based on the reported affinity and selectivity of SAHA to these individual targets.

Figure 31 shows that Class IIa HDACs do not generate significant BRET with SAHA-TOM, as predicted based on the reported selectivity of SAHA to Class I and IIb HDACs.

Figure 32 shows competitive displacement results of SAHA against individual HDAC-NanoLuc fusions complexed with SAHA-TOM, as determined via BRET. From these concentration response curves, IC-50 values can be calculated, as shown in table next to the graph.

Figure 33 (left) shows a conversion between IC-50 value and Ki value for individual HDAC-NanoLuc fusions, as determined via BRET. The Cheng-Prusoff equation was used for this conversion. Figure 33 (right) shows a diagram representing relative affinities of SAHA against individual HDAC/NanoLuc fusion proteins, as determined via BRET.

Figure 34 shows BRET imaging of intracellular HDAC10-NLuc / PBI-4968 +/- SAHA complexes in the presence or absence of SAHA as a specificity control.

Figure 35 shows BRET imaging of intracellular NLuc-HDAC6 / PBI-4968 +/- SAHA complexes in the presence or absence of SAHA as a specificity control.

Figure 36 shows dot plot analysis of SAHA binding intracellular SAHA-TOM (PBI-4968) BRET imaging experiments with HDAC10 and HDAC6.

DETAILED DESCRIPTION

The present invention provides compositions and methods for detection and analysis of intracellular binding of a bioactive agent to a cellular target. In particular, provided herein are bioactive agents tethered to a chromophore, e.g., fluorophores, potential cellular targets fused to bioluminescent reporter proteins and methods of detecting and analyzing the interaction of bioactive agents with cellular targets therewith (See Figure 1).

The interaction of a first entity (e.g., bioactive agent) and a second entity (e.g., cellular target) is detected, characterized, quantified, analyzed, etc. through the detection/measurement of

a signal produced by signal transfer (e.g., transfer of energy (e.g., fluorescence, light energy, resonance, by BRET, etc.)) between a third entity (e.g., fluorophore) connected, fused, tethered, linked, etc. to the first entity and a fourth entity (e.g., bioluminescent reporter protein) connected, fused, tethered, linked, etc. to the second entity. The interaction and/or binding of the first and second entities bring the third and fourth entities into close enough proximity to allow signal transfer (e.g., energy transfer) from one to another. In some embodiments, the fourth entity (e.g., bioluminescent reporter protein) emits energy (e.g., upon interaction with its substrate), and that emitted energy is absorbed by the third entity (e.g., fluorophore), causing the third entity to emit measurably different energy form the fourth entity (e.g., light at a different wavelength). In such embodiments, detection of the energy emitted from the third entity (e.g., light at the emission maximum of the third entity) upon addition to the system of the substrate for the fourth entity (e.g., bioluminescent reporter) indicates interaction of the first and second entities. In some embodiments, the duration, kinetics, affinity, strength, and/or specificity, of the binding of the first and second entities is detected, measured, quantified, determined, interrogated, etc. based on measurement of the signal output of the fourth entity (e.g., bioluminescent reporter protein) under various conditions.

In various embodiments, a cellular target fused to a bioluminescent reporter protein and a bioactive agent tethered to a chromophore, e.g., fluorophore, are provided (e.g., intracellularly, extracellularly, in lysate, *in vitro*, etc.). Substrate for the bioluminescent reporter protein is added to the system. If an interaction has occurred (e.g., binding) between the bioactive agent and the cellular target, the bioluminescent reporter protein and chromophore (e.g., fluorophore) are brought into close enough proximity for BRET to occur, and a detectable signal to be emitted from the chromophore (e.g., fluorophore).

In some embodiments, a complementary peptide and polypeptide that can interact (e.g., form a complex) to form a bioluminescent reporter protein (or protein complex) are used. In some embodiments, a complementary peptide is fused to a first interaction partner and a complementary polypeptide is fused to a second interaction partner. In some embodiments, the first and second interaction partners form a complex (e.g., by binding to each other). In some embodiments, the first and second interaction partners form an interaction complex when one or both interacts with a bioactive agent. In some embodiments, the first and second interaction partners form an interaction complex in the presence or absence of a bioactive agent. In some embodiments, formation of the interaction complex brings the complementary peptide and polypeptide together to form the bioluminescent reporter. In some embodiments, formation of the bioluminescent reporter allows detection of formation of the interaction complex. In some

embodiments, a fluorophore is tethered to the bioactive agent. In some embodiments, energy is transferred from the bioluminescent reporter to the fluorophore when the interaction complex is formed and the bioactive agent is bound to one of the interaction partners or the interaction complex. In some embodiments, the fluorophore allows detection or measurement of the interaction of the first and second interaction partners. In some embodiments, the fluorophore allows detection or measurement of the binding of the bioactive agent to its binding partner (e.g., the first interaction partner, the second interaction partner, and/or the interaction complex).

In some embodiments, a complementary peptide of a bioluminescent reporter is fused to a target of interest. A complementary polypeptide and a bioactive agent tethered to a fluorophore are provided to detect or measure the interaction of the bioactive agent to the target of interest. In some embodiments, the complementary peptide of a bioluminescent reporter is fused to a target of interest, a complementary polypeptide of the bioluminescent reporter protein and a bioactive agent tethered to a fluorophore is applied to detecting proximity of the fluorescently labeled ligand, e.g., a bioactive agent, to protein complexes (e.g. detecting ligand binding selectively to heterodimeric or homodimeric receptors) by linking members of the complex to components of a multicomponent luminescent donor. This application could be thus used to monitor target engagement within protein complexes that may have a role in disease pathology.

In some embodiments, the cellular target and bioluminescent reporter fusion is expressed in cells in which an assay is to be performed. In some embodiments, the fusion is expressed at or near the native abundance for the cellular target. In some embodiments, the fluorophore-tethered bioactive agent is added extracellularly (e.g., added to the culture medium) and enters the cell via diffusion, active transport, passive transport, endocytosis, or any suitable mechanism. In some embodiments, varying amount of fluorophore-tethered bioactive agent is added to the cells to assay binding kinetics, assay binding affinity, provide sufficient signal, etc.

In certain embodiments, the present invention provides compositions, methods, and systems for detection of intracellular interactions between a bioactive agent and cellular target (e.g., known or unknown). In some embodiments, a fusion of a bioluminescent reporter and a cellular target are expressed within a cell. The bioactive agent, tethered to a fluorophore, is introduced to the cell (e.g., conjugate of fluorophore and bioactive agent is cell permeable, the cell is permeabilized, etc.). A substrate for the bioluminescent reporter protein is added to the cell prior, concurrently or subsequent to the addition of the bioactive agent. Detection of a fluorescent signal from the fluorophore (as the result of BRET) indicates an intracellular interaction (e.g., binding) between the bioactive agent and cellular target. In some embodiments, a cellular target fused to a bioluminescent reporter is expressed at a natural cellular abundance

(e.g., relative to the native cellular target, or at a level appropriate for proper biological function of the fused target). In some embodiments, interaction of a bioactive agent with a cellular target is detected intracellularly.

In some embodiments, characteristics of the interaction between the bioactive agent and cellular target are interrogated by altering the cellular conditions or the conditions of the system. For example, in some embodiments, a competitive binder of the cellular target (e.g., untethered bioactive agent) is added to the cell to compete with the fluorophore-tethered bioactive agent.

In some embodiments, a library of bioluminescent reporter protein-tagged cellular targets is provided (e.g., in solution, in a lysate, immobilized on a surface, expressed within a cell, etc.). In some embodiments, a bioactive agent is provided which lacks a known cellular target or where knowledge of the cellular target is uncertain or incomplete. The cellular target of a bioactive agent is determined by adding the bioactive agent to the library and determining which cellular-target fusion produces BRET induced fluorescence of the bioactive agent-tethered fluorophore. In some embodiments, the library of bioluminescent reporter-tagged cellular targets is provided as a collection of nucleic acids or vectors comprising nucleic acids (e.g., plasmids, BacMam viruses, Lentiviruses, etc.) encoding the protein fusions. In some embodiments, bioluminescent reporter protein-tagged cellular targets are expressed within cells. In some embodiments, the library of bioluminescent reporter-tagged cellular targets is provided by translating nucleic acids in cell-free translation reactions. In some embodiments, a library of cellular target fusions, or cells expressing cellular target fusions, are provided in a microplate format. In such embodiments, the interaction of a bioactive agent (e.g., one identified through a phenotypic assay or screen) with the entire library of cellular targets can be interrogated (e.g., in solution, in lysates, intracellularly, etc.) in a high-throughput manner. In some embodiments, bioluminescent reporter-tagged cellular targets are immobilized on a solid surface to create a protein array. For example, in some embodiments, in addition to the bioluminescent reporter, cellular targets are also expressed as fusions with a tag or coupled to a protein (e.g., HALOTAG, Promega) that allows for the proteins to be covalently immobilized on a solid surface (e.g., a surface displaying appropriate ligands (e.g., HALOTAG Ligand)). In certain embodiments, a library of potential bioactive agents (e.g., hit compounds or drug-like small molecules) is added to the system (e.g., array) and any pairs capable of producing BRET are identified. In some embodiments, cellular targets for all or a portion of a library of bioactive agents are unknown.

In certain embodiments, compositions, methods, and systems herein provide a conjugate of a bioactive agent and an energy acceptor (e.g., fluorophore, chromophore). In some embodiments, a bioactive agent is any small molecule (e.g., > 2000 daltons, >1000 daltons, >500

daltons, etc.), macromolecule, synthetic molecule or molecular complex capable of interacting with the biology of a cell. In some embodiments, an energy acceptor is an entity capable of generating, exhibiting, and/or emitting a signal (e.g., light, heat, chemical reaction, fluorescence, resonance energy, etc.) when triggered by energy absorption (e.g., resonance energy transfer). In some embodiments, a bioactive agent and energy acceptor (e.g., fluorophore, chromophore) are fused, tethered, connected, etc., by any suitable structure or mechanism (e.g., expressed as a fusion construct (e.g., with or without peptide linker), chemically linked (e.g., directly or indirectly), enzymatically linked, linked by a linker (e.g., peptide, nucleic acid, polymer, ester linkage, PEG linker, carbon chain, etc.)). In some embodiments, the conjugate of a bioactive agent and an energy acceptor (e.g., bioactive agent tethered to a fluorophore) is produced by non-natural chemical synthesis (e.g., a purposeful execution of chemical reactions not present in natural cells). The type of linkage should not be viewed as limiting.

As used herein, the term “bioactive agent” refers generally to any physiologically or pharmacologically active substance or a substance suitable for detection. In some embodiments, a bioactive agent is a potential therapeutic compound (e.g., small molecule, peptide, nucleic acid, etc.), or drug-like molecule. In some embodiments, the bioactive agent is produced by non-natural chemical synthesis (e.g., a purposeful execution of chemical reactions not present in natural cells). Bioactive agents for use in embodiments described herein are not limited by size or structure.

In certain embodiments, libraries of bioactive agents (e.g., >10 agents, >50 agents, >100 agents, >500 agents, >1000 agents, >5000 agents, >10,000 agents, >50,000 agents, etc.) are provided. In some embodiments, systems, methods, and compositions are provided for screening libraries of bioactive agents. In some embodiments, the present invention provides means of identifying the bioactive agent in a library responsible for producing, eliciting, inducing, etc. phenotypic effect and/or activity. In some embodiments, the present invention provides means of identifying the cellular target of a bioactive agent (e.g., a bioactive agent responsible for the phenotypic effect and/or activity).

As used herein, the term “energy acceptor” refers to any small molecule (e.g., chromophore), macromolecule (e.g., autofluorescent protein, phycobiliproteins, nanoparticle, surface, etc.), or molecular complex that produces a readily detectable signal in response to energy absorption (e.g., resonance energy transfer). In certain embodiments, an energy acceptor is a fluorophore or other detectable chromophore. Suitable fluorophores include, but are not limited to: xanthene derivatives (e.g., fluorescein, rhodamine, Oregon green, eosin, Texas red, etc.), cyanine derivatives (e.g., cyanine, indocarbocyanine, oxacarbocyanine, thiocarbocyanine,

merocyanine, etc.), naphthalene derivatives (e.g., dansyl and prodan derivatives), oxadiazole derivatives (e.g., pyridyloxazole, nitrobenzoxadiazole, benzoxadiazole, etc.), pyrene derivatives (e.g., cascade blue), oxazine derivatives (e.g., Nile red, Nile blue, cresyl violet, oxazine 170, etc.), acridine derivatives (e.g., proflavin, acridine orange, acridine yellow, etc.), arylmethine derivatives (e.g., auramine, crystal violet, malachite green, etc.), tetrapyrrole derivatives (e.g., porphyrin, phthalocyanine, bilirubin, etc.), CF dye (Biotium), BODIPY (Invitrogen), ALEXA FLUOR (Invitrogen), DYLIGHT FLUOR (Thermo Scientific, Pierce), ATTO and TRACY (Sigma Aldrich), FluoProbes (Interchim), DY and MEGASTOKES (Dyomics), SULFO CY dyes (CYANDYE, LLC), SETAU AND SQUARE DYES (SETA BioMedicals), QUASAR and CAL FLUOR dyes (Biosearch Technologies), SURELIGHT DYES (APC, RPE, PerCP, Phycobilisomes)(Columbia Biosciences), APC, APCXL, RPE, BPE (Phyco-Biotech), autofluorescent proteins (e.g., YFP, RFP, mCherry, mKate), quantum dot nanocrystals, etc. In some embodiments, a fluorophore is a rhodamine analog (e.g., carboxy rhodamine analog), such as those described in U.S. Pat. App. Ser. No. 13/682,589, herein incorporated by reference in its entirety. Some such fluorophores are described herein in Example 8. In some embodiments, BRET efficiency is significantly enhanced by the technical features of rhodamine analog (e.g., carboxy rhodamine analog) as an energy acceptor compared to other fluorophores. For example, the left shifted EC50 and reduced nonspecific background of these dyes is advantageous for use in some embodiments.

As used herein, the terms “TOM” and “NonChloroTOM” (or “NCT”) refer to the same type of fluorophore and are used interchangeably throughout the application.

In certain embodiments, compositions, methods, and systems herein provide a fusion of a cellular target and a bioluminescent reporter protein (e.g., luciferase). In some embodiments, a cellular target and bioluminescent reporter protein are fused, tethered, connected, etc. by any suitable structure or mechanism (e.g., expressed as a fusion construct (e.g., with or without peptide linker), chemically linked (e.g., through covalent or non-covalent bonds), enzymatically linked, linked by a linker (e.g., peptide, nucleic acid, other polymer (e.g., ester linkage, PEG linker, carbon chain, etc.)). In some embodiments, an amino acid chain (e.g., 3-100 amino acids) is used to connect the cellular target and a bioluminescent reporter protein. In some embodiments, the structure and/or function of neither the cellular target nor the bioluminescent reporter are impacted (e.g., significantly impacted) by fusion or the presence of the linker. In certain embodiments, a linker allows fusion without loss of activity or one or both of the elements. In other embodiments, an amino acid linker properly spaces and/or orients the bioluminescent reported for energy transfer with the fluorophore.

In some embodiments, a cellular target comprises any suitable binding/interaction partner (e.g., receptor, enzyme, protein complex) for a bioactive agent (e.g., small molecule, protein, nucleic acid, lipid, etc.). However, in some embodiments, knowledge of the interaction between a cellular target and bioactive agent is not required to practice the invention. In particular embodiments, a cellular target is a protein or protein complex that binds to or otherwise interacts with (e.g., has a binding affinity to) a bioactive agent. In more particular embodiments, a cellular target is a receptor protein or an enzyme that binds to or otherwise interacts with (e.g., has a binding affinity to) a small molecule bioactive agent. The present invention is not limited by the identity, type, or class of cellular targets. In certain embodiments, libraries of hundreds, thousands, tens of thousands, more different cellular targets find use in the present invention. Examples of cellular targets may include nucleic acids (e.g., DNA or RNA), polysaccharides or a complex comprising any of these with polypeptides. In some embodiments, a cellular target is a G-protein coupled receptor or a protein kinase.

In certain embodiments, the bioluminescent reporter is a luciferase. In some embodiments, a luciferase is selected from those found in *Gaussia*, *Coleoptera*, (e.g., fireflies), *Renilla*, *Vargula*, *Oplophorus*, *Aequoria*, mutants thereof, portions thereof, variants thereof, and any other luciferase enzymes suitable for the systems and methods described herein. In some embodiments, the bioluminescent reporter protein is a modified, enhanced luciferase enzyme from *Oplophorus* (e.g., NANOLUC enzyme from Promega Corporation, SEQ ID NO: 1, or a sequence with at least 70% identity (e.g., >70%, >80%, >90%, >95%) thereto). In some embodiments, the bioluminescent reporter protein is a thermostable *Photuris pennsylvanica* luciferase or a sequence with at least 70% identity (e.g., >70%, >80%, >90%, >95%) thereto). Exemplary bioluminescent reporters are described, for example, in U.S. Pat. App. No. 2010/0281552 and U.S. Pat. App. No. 2012/0174242, both of which are herein incorporated by reference in their entireties.

In some embodiments, the bioluminescent reporter protein comprises NANOLUC (See U.S. Pat. App. Nos. 2010/0281552 and 2012/0174242, herein incorporated by reference in their entireties). In some embodiments, the bioluminescent reporter protein comprises a polypeptide with at least 70% identity (e.g., >70%, >80%, >90%, >95%) to SEQ ID NO: 1 that retains bioluminescent characteristics. In certain embodiments, the use of the NANOLUC enzyme, or a variant thereof, provides features (e.g., signal intensity, brightness, high light output, narrow spectrum, etc.) that enable the use of the BRET assays described herein. In some embodiments, the high light output of NANOLUC enables the low concentration (e.g., <1 μ M, <100 nM, <10 nm, <1 nm, etc.) of assay components, e.g., DNA encoding NANOLUC, useful to carry out

assays under physiologically relevant conditions. In some embodiments, NANOLUC enables the use of BRET in characterizing cellular targets identified in a phenotypic screen.

In some embodiments, a substrate for the bioluminescent reporter protein is provided. In some embodiments, the bioluminescent reporter protein converts the substrate into a reaction product and releases light energy as a by-product. In some embodiments, the substrate is a substrate for a luciferase enzyme. In some embodiments, the substrate is a structural variant or derivative of coelenterazine (e.g., furimazine). In some embodiments, the substrate is a substrate for a modified, enhanced luciferase enzyme from *Oplophorus*, e.g., NANOLUC enzyme from Promega Corporation (e.g., SEQ ID NO: 1). In some embodiments, a pro-substrate for the bioluminescent reporter protein is provided, which produces a substrate through a chemical or physical process (e.g., hydrolysis, enzymatic reaction, photo-cleavage, etc.). In some embodiments, the pro-substrate comprises coelenterazine, a coelenterazine derivative, a structural or functional equivalent of coelenterazine, a molecule substantially equivalent to coelenterazine (e.g., structurally and/or functionally), or molecule functionally or structurally similar to coelenterazine. In some embodiments, the bioluminescent reporter protein converts the coelenterazine, coelenterazine derivative, structural or functional equivalent of coelenterazine, or substantial equivalent to coelenterazine into coelenteramide, a coelenteramide derivative, a structural or functional equivalent of coelenteramide, or a substantial equivalent to coelenteramide and releases light energy as a by-product.

In some embodiments, fluorophore and bioluminescent reporter are selected that exhibit sufficient overlap of emission (e.g., of bioluminescent reporter) and excitation (e.g., of fluorophore) spectra to provide efficient energy transfer between the two (e.g., by non-radiative dipole–dipole coupling). In some embodiments, the peak emission of the bioluminescent reporter is substantially separated from the peak emission of the fluorophore, for example by at least 80 nm, 100 nm, 120 nm, 140 nm, etc., in wavelength. In particular embodiments, the Forster distance of the fluorophore and bioluminescent reporter pair is small (e.g., <20 nm, <10 nm, <5 nm, <3 nm, etc.). In such embodiments, the short Forster distance results in the requirement that the fluorophore and bioluminescent reporter must be brought into very close proximity for energy transfer to occur. Therefore, the short Forster distance reduces aberrant and/or background signal (e.g., created by diffusing fluorophore and/or reporter).

In certain embodiments, a fluorophore and bioluminescent reporter pair are selected that are sufficiently bright to allow detection of the transferred signal at a native abundance (or near native abundance) of the protein of interest and/or the cellular target fused to the bioluminescent reporter. In some embodiments, should either the selected fluorophore or bioluminescent

reporter produce insufficient energy (light) emission, either the fusion of the cellular target and bioluminescent reporter will need to be overexpressed (e.g., beyond native abundance, beyond a biologically relevant level, etc.), and/or the amount of fluorophore-tethered bioactive agent will have to be increased (e.g., to a potentially toxic level, beyond a physiologically relevant level, above the amount when kinetic experiments can be performed, etc.). In some embodiments, sufficient brightness of the bioluminescent reporter and fluorophore allows detection of bioactive agent and cellular target interaction at a range of concentrations and ratios.

In some embodiments, compositions, methods, and systems are provided for identification of the cellular targets of hit compounds emerging from a phenotypic assay or phenotypic screen. In some embodiments, following identification of bioactive agents capable of eliciting a phenotype, “hit compounds” tethered to fluorophores are used to identify cellular targets, through their linkage to a bioluminescence reporter protein (e.g., binding of bioactive agent to cellular target results in BRET between bioluminescence reporter protein and fluorophore). In such embodiments, although a phenotype has been associated with a particular bioactive agent, the interaction partner (e.g., cellular target) for that bioactive agent is unknown or is uncertain. In some embodiments, a bioactive agent tethered to a fluorophore is able to regenerate a phenotype, thereby validating that tethering the bioactive agent to the fluorophore (or the cellular target to the bioluminescent reporter) does not affect the cellular binding patterns and/or disrupt its biological activity. In some embodiments, the use of a library of cellular targets, each fused to bioluminescent reporter protein (e.g., NANOLUC), provides less interference due to poor solubility or low native abundance of the cellular target compared to other detection methods (e.g., mass spectrometry analysis). In some embodiments, such a detection method provides a greater degree of sensitivity or specificity. In some embodiments, target identification through linkage to a bioluminescence reporter protein enables detection through energy transfer, even when binding of the target is inefficient or incomplete. In some embodiments, binding of a bioactive agent (e.g., small molecule) to a cellular target is characterized using a competitive binding assay (SEE Figure 1C and 1D). In some embodiments, BRET allows for analysis of the hit compound’s binding affinity in live cells using competitive displacement of a fluorescent tracer (e.g., fluorophore) that binds to the same site on the cellular target. In some embodiments, systems and methods described herein provide the ability to use two distinct methods for target identification, thus providing higher stringency for identifying cellular targets, complementary methods addressing limitations with other approaches (e.g., BRET provides the advantage of maintaining the bioactive agent in equilibrium with the target throughout the experiment).

Some embodiments described herein find use in drug discovery, drug validation, drug target discovery, or drug target validation. In certain embodiments, the binding interaction between a bioactive agent (e.g., drug-like small molecule) and a cellular target can be detected, validated, and/or characterized. In some embodiments, the relative binding affinity of bioactive agents for a cellular target (e.g., in solution, in a lysate, on a surface, in a cell, etc.), can be determined by their ability to displace a bioactive agent that has been tethered to a fluorophore. Specifically, higher binding affinity of a first bioactive agent relative to a second bioactive agent is indicated by requiring a lower concentration of the first bioactive agent to displace a tethered bioactive agent relative to the second bioactive agent. Displacement of the tethered bioactive agent is determined by the loss or reduction of energy transfer from the bioluminescent reporter protein fused to a cellular target. In some embodiments, the concentration of bioactive agent needed to displace a tethered bioactive agent is used to estimate the binding EC50 or the inhibition constant (K_i) for bioactive agent. In some embodiments, the development of new or modified bioactive agents is guided by their ability to displace one or more bioactive agents, each tethered to a fluorophore, from one or more cellular targets, each fused to a bioluminescent reporter protein.

In some embodiments, a collection of compounds which may have unknown binding affinity to a cellular target may be screened for their ability to bind the target fused to a bioluminescent protein by determining their ability to displace a bioactive agent tethered to a fluorophore. In some embodiments, compounds may be screened for their ability to bind a first cellular target preferentially relative to a second cellular target by their ability to displace a first tethered bioactive agent from the first cellular target relative to a second tethered bioactive agent from the second bioactive target. In some embodiments, the first and second tethered bioactive agents are the same.

In some embodiments, systems and methods described herein provide the ability to determine the affinity of a bioactive agent (e.g., hit compound, research lead, lead compound, etc.) for wild-type and mutant version(s) of a cellular target, e.g., target protein. In some embodiments, characterization of the affinity and selectivity of the fluorescently-labeled bioactive agent (e.g., drug) to a disease-relevant mutant protein may be performed in cells. Such systems and methods may be useful to identify bioactive agent(s) (e.g., drug) that selectively bind a wild-type or mutant protein differentially.

In addition to primary targets, intended targets, and/or known targets, bioactive agents (e.g., hit compounds) may also bind to unexpected and/or unintended cellular targets (off targets). In some cases, off target binding of a bioactive agent is responsible for a portion of the

therapeutic and/or adverse effects associated with administration of the bioactive agent. In some embodiments, systems and methods described herein provide the ability to identify off targets of a bioactive agent. Understanding the identity and extent of off-target bioactive agent binding provides valuable information regarding the pharmacology of the agent.

In some embodiments, systems and methods described herein provide the ability to estimate the binding characteristics of a bioactive agent tethered with a fluorophore to a cellular target fused to a bioluminescent reporter (e.g., EC₅₀, K_d, binding rate, environmental influences, etc.) In some embodiments, the binding characteristics are correlated to biochemical, physical, or phenotypic characteristics related to the cellular target. In some embodiments, the kinetic profile of the formation or dissolution of the BRET complex can be used to infer association or dissociation rates of untethered bioactive agents. In some embodiments, these association/dissociation rates can be used to estimate drug residence times for individual targets within intact cells. In some embodiments, systems and methods are useful in thermodynamic molecular mechanism of action (MMOA) studies on entropic vs. enthalpic interactions. In some embodiments, displacement of the tethered bioactive agent by a separate agent may occur through separate sites on the cellular target. In some embodiments, the binding characteristics of the tethered bioactive agent may be used to determine the influence of post-translational modification (e.g., cleavage, phosphorylation, methylation, acetylation, lipidation, etc.), intracellular translocation (e.g., movement to a nucleus, mitochondria, membrane, etc.), or protein interaction (e.g., interactions with other proteins, nucleic acids, lipids, etc.) on the cellular target. An example is determining the binding characteristics of an antibody to a cellular target by its influence on the binding characteristics of the tethered bioactive agent. In some embodiments, the binding characteristics of the tethered bioactive agent may be used to determine the influence of chemical modifications or transformation of the bioactive agent or the tethered fluorophore (e.g., intracellular metabolism, change in ionic state, etc.).

In some embodiments, the cellular target may comprise more than one molecular component. For example, the target may comprise more than one polypeptide, and may further comprise other natural or synthetic molecules (e.g., prosthetic groups, cofactors, metabolites, nucleic acids, lipids, carbohydrates, etc.). In some embodiments, a bioactive agent tethered to a fluorophore binds to a first molecular component, and a bioluminescent reporter is fused to a second molecular component, so that a signal from the tethered bioactive agent is produced when bound to the first molecular component and the first molecular component is in close proximity to the second molecular component.

A priori knowledge of the existence of an interaction between a bioactive agent and cellular target is not needed to practice the invention. In some embodiments, detection and/or characterization of an unknown or previously unidentified interaction by energy transfer is provided. Advantages of the systems, compositions, and methods described herein, over other methods of target discovery may include: a broad range of possible bioactive agent concentrations because (in some embodiments) it need not be expressed within cells (e.g., allows for addition of enough acceptor fluorophore to generate detectable signal), natural protein concentration of cellular target (e.g., no need for overexpression to gain enough signal for detection), signal detectable on a plate reader (e.g., high throughput detection, no imaging necessary), detection of interactions within a cell, etc.

In some embodiments, bioactive agents (e.g., hit compounds) are conjugated to a fluorescent energy acceptor dye, and therefore the binding of the modified agent to its luciferase-fused (e.g., NANOLUC-fused) cellular target results in energy transfer from NANOLUC to the acceptor dye. Such a system provides a homogenous assay that can be performed in living cells. In some embodiments, the labeled bioactive agent remains in equilibrium with the cellular target throughout the experiment allowing detection of targets that interact with the hit compounds with low affinity. In some embodiments, BRET enables measurements of binding affinity in living cells by the competitive displacement of a fluorescent tracer designed for the same binding site.

In some embodiments, BRET efficiency is significantly enhanced by the technical features of NANOLUC as an energy donor compared to other luciferases. For example, NANOLUC is significantly brighter than other luciferases commonly used for BRET, thereby allowing energy transfer to be quantitated at lower expression levels which are more suitable for maintaining relevant biology within a cell. In some embodiments, the narrow emission spectrum of NANOLUC increases the dynamic range by reducing spectral cross-over in the acceptor channel. In some embodiments, the dynamic range can be further increased by using long-wavelength acceptors that emit in the near-red region of the spectrum (600 to 650 nm). In some embodiments, evaluation of multiple dye ligands during development of embodiments of the present invention revealed that rhodamine analogs (e.g., carboxy rhodamine analog), such as those described in U.S. Pat. App. Ser. No. 13/682,589, herein incorporated by reference in its entirety, delivers an optimal dynamic range for use with NANOLUC and/or in the BRET applications described herein.

EXPERIMENTAL

Example 1

Experiments were conducted during development of embodiments of the present invention to demonstrate the improved performance of PBI tracer conjugates and compared to conjugates with standard dyes for drug tracer applications. In this example, conjugates of suberoylanilide hydroxamic acid (SAHA), an inhibitor of histone deacetylase 6 (HDAC6), and a PBI dye (SAHA-TOM (PBI-4968); See Figure 8) or standard dye (SAHA-TAMRA (PBI-4967); See Figure 8) were utilized in an intracellular BRET assay using a NanoLuc-HDAC fusion protein (SEQ ID NO: 3).

HEK293 cells were transfected using FuGene HD (Promega Corp.) with plasmid DNA encoding a NanoLuc-HDAC6 fusion protein. NanoLuc-HDAC6 DNA was diluted 1:1000 with a promoterless carrier DNA (pGEM3Z) to yield a final concentration of 50ng/well total DNA in 96-well plate format (at a seeding density of 20,000 cells/well). Twenty-four hours post-transfection, cells were then incubated with serially-diluted tracer in the presence or absence of a molar excess of unlabeled SAHA (as a specificity control). Following equilibration with the drug tracers, furimazine (a coelenterazine derivative substrate for NanoLuc; Promega Corp.) was added at a concentration of 20uM, and BRET was quantified on a Varioskan luminometer. The specific BRET signal was calculated by subtracting the non-specific signal (in the presence of unlabeled SAHA) from the signal generated from the SAHA tracer alone.

Results in Figure 2 demonstrate that the SAHA-TOM (PBI-4968) conjugate/tracer generated a superior, specific BRET signal compared to SAHA-TMR (PBI-4967) and also generated a left-shifted EC₅₀ value for binding within living cells. As a drug conjugate for BRET applications, the benefit of the PBI-4968 TOM dye over other commonly used fluorophores can be applied to competitive binding assays or target identification (chemical proteomics) screening using NANOLUC fusion protein libraries.

While this example demonstrates the binding of a drug tracer to a known high-affinity target, in other embodiments such a tracer is combined with a library of NANOLUC fusion proteins to profile relative drug affinities and specificities across target families. In such embodiments, profiling efforts aide in identification of drugs with promiscuous binding profiles, which may correlate with undesirable drug side effects in vivo.

Example 2

Experiments were conducted during development of embodiments of the present invention to demonstrate the unexpected benefit of low expression levels of NanoLuc for BRET applications.

A) HEK293 cells were transfected using Fugene HD with varying amounts of NanoLuc-HDAC6 DNA and seeded in a 96-well plate format. For the transfection, NanoLuc-HDAC6 DNA was diluted with promoterless carrier DNA (pGEM3Z). The final DNA concentration/well remained 50ng/well; however the NanoLuc-HDAC6 DNA was diluted 1:10, 1:100, and 1:1000 (at a seeding density of 20,000 cells/well in 96-well format). Twenty-four hours post-transfection, cells were treated with serially-diluted SAHA in the presence of a fixed concentration (1uM) of SAHA-TOM (PBI-4968; See Figure 8) conjugate. After two hours of incubation, furimazine was added to 20uM, and BRET detected on a Varioskan luminometer. The results in Figure 3 demonstrate that an unexpectedly high dilution of DNA encoding the NanoLuc fusion was required to achieve proper signal to background for BRET applications.

B) DNA encoding a NanoLuc-Histamine H1 (GPCR) fusion protein was transfected into HEK293 cells using varying amounts of DNA ranging from undiluted to 1:10,000 (diluted with promoterless carrier DNA (pGEM3Z as described above) to maintain a constant amount of DNA/transfection as described above). Twenty-four hours post-transfection, cells were treated with serially-diluted with mepyramine-bodipy-633 (CellAura) and equilibrated for 2 hours. Cells were then treated with furimazine to a concentration of 20uM, and BRET detected on a Varioskan luminometer.

The results in Figure 3B indicate that a large dilution of DNA encoding the NanoLuc fusion was required to generate a high affinity interaction with optimal signal-to-background. The ability to dilute NanoLuc fusion proteins to exceedingly low levels is beneficial in various BRET applications, including competitive binding assays or target identification (chemical proteomics) screening of NanoLuc fusion proteins.

Example 3

Experiments were conducted during development of embodiments of the present invention to demonstrate the predictive value of intracellular BRET measurement for quantifying the binding of pro-drugs compared to traditional biochemical (activity-based) formats. The example includes the natural pro-drug, FK228, which requires the reducing environment of the cell cytoplasm to become activated and capable of binding to HDAC6.

HEK293 cells were transfected with DNA encoding NanoLuc-HDAC6 at a 1:1000 dilution with carrier DNA as previously described. Twenty-four hours post-transfection, cells were incubated with serially-diluted natural pro-drug, FK-228; in the presence of a fixed concentration (1uM) of SAHA-TOM (PBI-4968) conjugate. For comparison, a biochemical, activity-based HDAC6 assay was run in parallel. Briefly, a 3-fold serial dilution of FK-228

(Selleckchem catalog # S3020) was performed at 100X in 100% DMSO in wells of a 96-well plate. A 5 μ L aliquot of this 100X/100% DMSO titration series was added to 245 μ L of HDAC-GloTM I/II assay buffer alone (Promega Corp.) or 245 μ L of HDAC-GloTM I/II assay buffer supplemented with 2X (0.5mM) DTT to make a 2X/2% DMSO master intermediate titration series of FK-228 in wells of a 96-well plate. From this master intermediate titration series, 5 μ L replicates were transferred to wells of a white 384-well assay plate (Corning 3673). A 5 μ L addition of 2X (2nM) HDAC 6 (BPS Bioscience catalog # 50006) was added to all wells for a final concentration of 1nM HDAC6/well. The 10 μ L enzyme/inhibitor mixes was allowed to pre-incubate for 45 minutes at room temperature. An equal volume (10 μ L) of HDAC-GloTM I/II Final Detection Reagent (Promega Corp.) was added to all wells (20 μ L final assay volume), and luminescence was measured after a 10 minute incubation at room temperature. For all test conditions, the final concentration of HDAC-GloTM I/II substrate was 50 μ M. The data was plotted (sigmoidal dose response – variable slope) using PrismTM software from GraphPad (no DTT: closed circles; 0.25 mM DTT: open squares).

The results in Figures 4A and 4B indicate the requirement of the cellular milieu for measuring binding of pro-drugs that must be processed by the cell to become activated.

Example 4

Experiments were conducted during development of embodiments of the present invention to demonstrate the use of permeabilization agents to introduce impermeable drug tracers to cells.

HEK293 cells were transfected with DNA encoding PKC α (PKC α)-NanoLuc or NanoLuc-PKC α at a 1:1000 dilution of DNA with promoterless carrier DNA as described previously. Twenty-four hours post-transfection, cells were treated with or without digitonin to a final concentration of 50 μ g/mL. Cells were then treated with serially-diluted a staurosporine-PBI-dye conjugate (PBI-5129; See Figure 8). Cells were co-incubated in the presence or absence of 5 μ M unconjugated staurosporine as a specificity control for binding. Following two hours of equilibration with a tracer, furimazine was added to a final concentration of 20 μ M, and BRET ratios were measured on a Varioskan luminometer (Figures 5A and 5B).

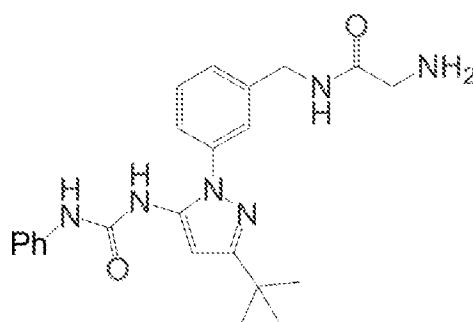
In a second experiment, identically-transfected PKC α -NanoLuc cells were treated with serially diluted digitonin prior to treatment with a fixed concentration of staurosporine-TOM (PBI-5129; See Figure 8) tracer (5 μ M final concentration) in the presence or absence of 5 μ M unlabeled staurosporine (specificity control) (Figures 5C).

Figure 5A-C demonstrate the ability to use permeabilization agent to potentiate the entry of impermeable drug tracers and applied to BRET-based chemical proteomics screens of NanoLucTM fusion protein libraries.

Example 5

Experiments were conducted during development of embodiments of the present invention to demonstrate the use of NanoLuc/BRET to measure relative drug affinities in living cells. In some embodiments, similar experiments are configured to optimize leads from high-throughput chemical screens.

HEK293 cells were transfected with DNA encoding a NanoLuc-p38 fusion protein (to a final concentration of 50ng/well DNA in 96-well format). Twenty-four hours post-transfection, cells were treated with serially diluted BIRB-796 or PBI-4835 in the presence of 0.5uM PBI-4838 (BIRB conjugate derivative; see, e.g., US 13/682,589; herein incorporated by reference in its entirety).



PBI 4835

After two hours of equilibration, furimazine was added to a concentration of 20uM, and BRET was measured on a Varioskan luminometer.

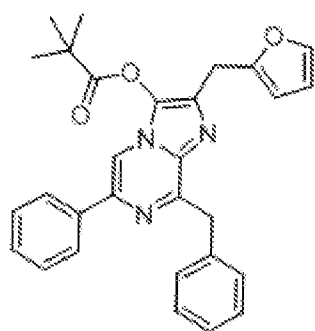
Figure 6 demonstrates that the dose-response BRET curves support the higher known affinity of BIRB-796 over PBI-4835 ("BIRB Derivative"). The ability to measure the relative affinities of unlabeled drugs can be applied to HTS screens, lead optimization or chemical proteomics applications. In some embodiments, similar experiments are configured to characterize optimized hit compounds from high-throughput chemical screens (e.g., compounds with lower IC₅₀ values for tracer displacement would indicate higher binding efficiency to the target of interest).

Example 6

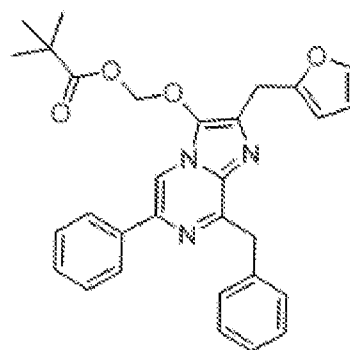
Experiments were conducted during development of embodiments of the present invention to demonstrate the use of NanoLuc/BRET to monitor the kinetics of drug binding in living cells. In some embodiments, similar experiments are configured to optimize leads from high-throughput chemical screens.

HEK293 cells were transfected with DNA encoding a NanoLuc-p38 fusion (to a final concentration of 50ng/well DNA in 96-well format). Twenty-four hours post-transfection, cells were pretreated for 2 hours with protected furimazine (PBI-4378; See Figure 8) to a final concentration of 20uM. BRET was measured over time on a Varioskan luminometer set to 37°C. After a short pre-read, cells were stimulated with varying concentrations of PBI-4838 (in the presence or absence of 1uM BIRB796 as a specificity control). Dose and time-dependent increases in BRET were then monitored kinetically over 4 hours.

In a separate experiment, transfected cells were pretreated with both 20uM PBI-4377 and 1uM PBI-4838 to generate a steady BRET signal.



PBI 4378



PBI 4377

After a short pre-read, cells were stimulated with 1uM unlabeled BIRB-796, and competitive displacement was monitored by BRET in real-time.

The results in Figure 7A-B support use of BRET for kinetic measurements of target engagement in living cells. Such formats can be exploited for HTS or chemical proteomics screening of NanoLuc fusion proteins in live cells. In some embodiments, similar experiments are configured to characterize optimized lead compounds from high-throughput chemical screens (e.g., compounds with altered kinetics of association or dissociation to the target of interest). In addition to providing a measure of relative compound affinities, such kinetic BRET measurements could enable quantitative assessments of target residence time biochemically or within intact cells

Example 7

The following example relates to the use of specific NANOLUC substrates (SEE FIG. 8) or enzyme components to discriminate the subcellular localization of a drug binding event using BRET.

The physical separation of complementary NANOLUC enzyme polypeptides or peptide allows the isolation of NANOLUC (donor) signal to the extracellular space. For example, the

small signal peptide is genetically tethered to a cell surface receptor. Upon exogenous addition of the large signal polypeptide to the cell medium, the donor signal is isolated to the extracellular space. This signal isolation increases the signal/background generated in various BRET applications including drug tracer binding/displacement.

In some embodiments, full length NANOLUC protein fusions are utilized if impermeable NANOLUC substrate is applied.

Example 8

The following provides synthesis schemes for exemplary compounds that find use in embodiments of the present invention.

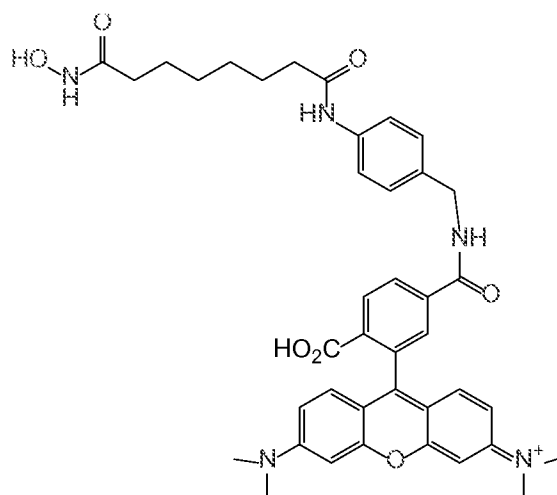
Boc-protected SAHA amine

7-Trityloxycarbamoyl heptanoic acid (200 mg, 463 μmol) was combined with 4-[(N-Boc)aminomethyl]aniline (113 mg, 510 μmol), HBTU (352 mg, 927 μmol) and triethylamine (194 μL , 1.4 mmol) in 3 mL of DMF. The reaction was stirred overnight, then adsorbed onto Celite, and the product was obtained by column chromatography eluting with a gradient of 0- >100% EtOAc in heptanes. Calcd for $\text{M}+\text{H}$: 635.3; found 635.9

SAHA amine

Suberoyl(4-[(N-Boc)aminomethyl]anilide) hydroxamic acid (286 mg, 450 μmol) was dissolved in 2 mL of DCM to which was added 0.25 mL of TIS. Trifluoroacetic acid (0.9 mL) was then added, and the reaction was stirred for 30 min. Solvents were removed under reduced pressure, and the crude reaction product could be purified by preparative HPLC or used without further purification.

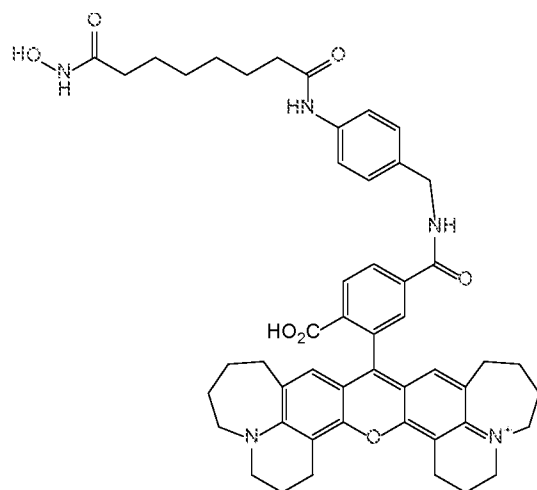
PBI-4967 SAHA-TAMRA



Suberoyl[4-(aminomethyl)anilide] hydroxamic acid crude reaction product (27 mg) was combined with 4 mg (7.6 μ mol) of tetramethylrhodamine6-succinimidyl ester in 1 mL of DMF with 5 drops of TEA. After 30 min, the reaction was diluted with H₂O and MeCN, and the product was isolated by preparative HPLC (5->60% MeCN in 0.1% aqueous TFA).

Lyophilization of the appropriate fractions yielded the desired product as a magenta solid. Calcd for M+H: 706.3; found 706.6.

PBI-4968 SAHA-TOM

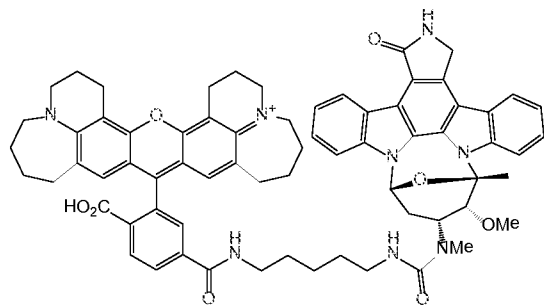


Suberoyl[4-(aminomethyl)anilide] hydroxamic acid crude reaction product (8 mg) was combined with 5 mg (7.6 μ mol) of TOM 6-succinimidyl ester in 0.8 mL of DMF with 3 drops of TEA. After 30 min, the reaction was diluted with H₂O and MeCN, and the product was isolated by preparative HPLC (5->60% MeCN in 0.1% aqueous TFA). Lyophilization of the appropriate fractions yielded the desired product. Calcd for M+H: 838.4; found 838.7

Staurosporine-amine

Staurosporine p-nitrophenylcarbamate (3 mg, 4.8 μ mol) was dissolved in 0.5 mL of DMF and treated with excess cadaverine. The reaction was warmed in a 70°C oil bath for 2 h, then diluted with H₂O, acidified with formic acid, and subjected to preparative HPLC eluting with 25%->75% MeCN in 10 mM aqueous NH₄OAc. The appropriate fractions were lyophilized to afford a slightly yellow solid. Calcd for M+H: 595.3; found 595.5

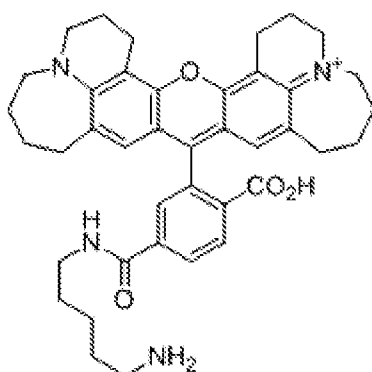
PBI-5129 Staurosporine-TOM



Staurosporine 5-aminopentylcarboxamide (5 mg, 8.4 μmol) was dissolved in 1 mL DMF and treated with triethylamine and TOM 6-SE (4 mg, 6 μmol). The reaction was monitored by analytical HPLC. Upon completion of the reaction, MeCN and H_2O were added, and the TEA was neutralized by addition of a small amount of AcOH. Preparative HPLC (25% $\rightarrow$ 100% MeOH in 10 mM NH_4OAc) and subsequent concentration yielded 1.8 mg of a dark blue solid. Calcd for $\text{M}+\text{H}$: 1139.5; found 1139.8.

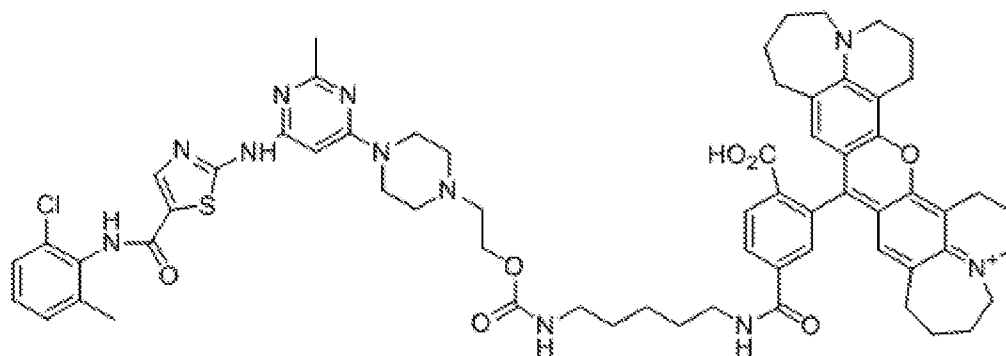
Dasatinib-TOM

TOM-pentylamine



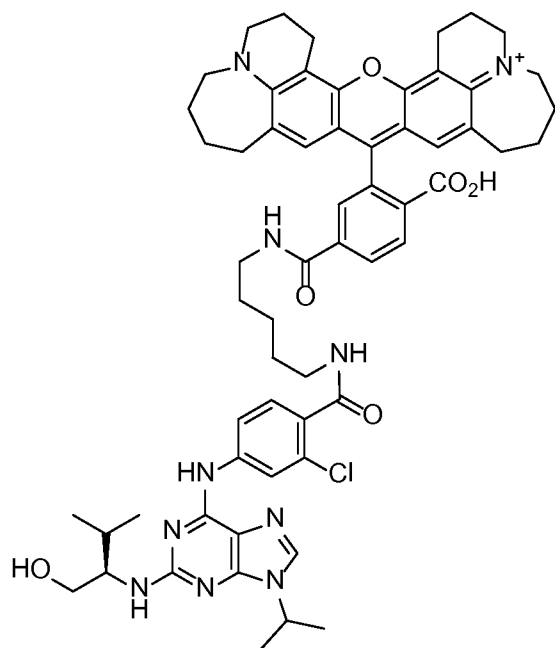
TOM 6-carboxylic acid (26 mg, 46 μmol) was stirred in 1 mL of DMF with 2 equiv of triethylamine and treated with TSTU (17 mg, 56.5 μmol , 1.2 equiv), and the reaction was monitored by HPLC. After 40 min, the reaction was added to a solution of cadaverine (94 mg, 0.92 mmol, 20 equiv) in 0.5 mL DMF and stirred for 20 min. The reaction was then neutralized by addition of TFA and diluted with MeCN and water. Preparative HPLC (25% $\rightarrow$ 100% MeCN in 0.1 % aqueous TFA) and subsequent lyophilization afforded the desired product as a purple solid. LCMS: Calcd for $((\text{M}+2\text{H})/2)$ 324.4; found 324.3.

Dasatinib-TOM



Dasatinib (25 mg, 51 μ mol) was combined with p-nitrophenyl chloroformate (14 mg, 69 μ mol, 1.36 equiv) and 30 μ L TEA in 0.9 mL of 2:1 DMF:THF. The reaction was stirred overnight and then 35 mg additional p-nitrophenyl chloroformate added. After stirring for an additional 24 h, $\frac{1}{2}$ the volume of the reaction was added to a solution of 7 mg (10 μ mol) of TOM-pentylamine in 0.5 mL of DMF and 30 μ L TEA. The reaction was then stirred for 1 h, followed by dilution with MeCN and water and purification by preparative HPLC. Lyophilization afforded the desired product as a purple solid. MS: Calcd for ((M+2H)/2) 580.8; found 581.0.

Purvalanol-TOM PBI 5077



Purvalanol (10 mg, 23 μ mol) was stirred in 1 mL of DMF with 20 mg of TEA (198 μ mol) and treated with 7.7 mg (25.4 μ mol, 1.1 equiv) of TSTU. After 30 min, the reaction was diluted with Et₂O and $\frac{1}{2}$ of the total reaction volume was added to a vial containing 6 mg (9 μ mol) of TOM (5-aminopentyl)-6-carboxamide. The reaction was stirred for 3 h, and the volatile organics were removed under reduced pressure. The resulting DMF solution was diluted with water and MeCN and acidified with TFA, then the product was isolated by preparative HPLC (25->100%

MeCN in 0.1% aqueous TFA) and subsequent lyophilization to yield 3.5 mg of a purple solid. MS: Calcd for M+H: 1061.5, found 1061.6.

Example 9

Use of Renilla Luciferase or NanoLuc luciferase in the compound identification method of the present invention

HEK293 cells were transfected with plasmid DNA encoding NanoLuc-p38alpha or Renilla Luc-p38alpha using Fugene HD(Promega Corporation) at a 3:1 lipid:DNA ratio and seeded into 96-well plates at a density of 20,000 cells per well (to yield 50ng/well of DNA). 24h post-transfection, cell medium was replaced with serum-free medium (Opti-MEM) and incubated with serially-diluted PBI-4838 (BIRB796 derivative conjugated to TOM dye) in the presence/absence of BIRB796 at 1uM. Cells were incubated at 37°C for two hours. To the NanoLuc-expressing samples, furimazine was added to a final concentration of 20uM. To the Renilla Luc-expressing samples, native coelenterazine was added to a final concentration of 20uM. BRET was then measured on a Varioskan luminometer equipped with 450nm bandpass and 630nm longpass filters. The BRET ratio was determined by dividing the signal in the 630 channel by the signal in the 450 channel.

Figure 9 demonstrates that both Renilla luciferase and NanoLuc luciferase can be used in the method of the present invention. Upon treatment of cells expressing NanoLuc-p38alpha or Renilla Luc-p38alpha with serially-diluted PBI-4838, a dose-dependent increase in BRET is observed. Saturating concentrations of unlabeled BIRB-796 are able to block this BRET signal, indicating specificity of binding using either luciferase fusion to p38alpha.

Example 10

Comparison of TOM-BIRB vs. TMR-BIRB against NanoLuc-p38 alpha

As described above, HEK293 cells were transfected with plasmid DNA encoding NanoLuc-p38alpha using Fugene HD and seeded into 96-well plates (to yield 50ng/well of DNA). 24h post-transfection, cell medium was replaced with serum-free medium (Opti-MEM) and incubated with serially-diluted PBI-4838 (BIRB796 derivative conjugated to TOM dye) or PBI-4836 (BIRB derivative conjugated to TMR dye) in the presence/absence of BIRB796 at 1uM. Cells were incubated at 37°C for two hours. To the NanoLuc-expressing samples, furimazine was added to a final concentration of 20uM. BRET was then measured on a Varioskan luminometer equipped with 450nm bandpass and 630nm longpass filters. The BRET ratio was determined by dividing the signal in the 630 channel by the signal in the 450 channel.

Figure 11 demonstrates another example of the increased potency of the TOM adduct over the TMR adduct for binding to a target protein in a cell-based format. Dose-response curves for NanoLuc-p38alpha binding indicate an EC₅₀ of 38nM for the BIRB-TOM conjugate, compared to an EC₅₀ of 450nM for the BIRB-TMR conjugate. The reported affinity of BIRB-TOM is more consistent with literature reports compared to the BIRB-TMR (Chem Biol Drug Des 2009; 74: 547–559).

Example 11

Comparison of BIM-TOM vs. BIM-TMR against PKCalpha-NanoLuc

As described above, HEK293 cells were transfected with plasmid DNA encoding PKCalpha-NanoLuc (diluted 1:1000 into pGEM3Z carrier DNA) using Fugene HD and seeded into 96-well plates (to yield 50ng/well of total DNA). 24h post-transfection, cell medium was replaced with serum-free medium (Opti-MEM) and incubated with serially-diluted PBI-5075 (BIM conjugated to TOM dye) or PBI-5051 (BIM conjugated to TMR dye) in the presence/absence of staurosporine at 5uM. Cells were incubated at 37°C for two hours. To the NanoLuc-expressing samples, furimazine was added to a final concentration of 20uM. BRET was then measured on a Varioskan luminometer equipped with 450nm bandpass and 630nm longpass filters. The BRET ratio was determined by dividing the signal in the 630 channel by the signal in the 450 channel. To determine specific BRET signals, the BRET ratio at each concentration of tracer + unlabeled staurosporine was subtracted from the BRET ratio at each concentration of tracer without unlabeled staurosporine.

Figure 12 demonstrates another example of the increased potency of the TOM adduct over the TMR adduct for binding to a target protein in a permeabilized cell format. Dose-response curves for PKCalpha-NanoLuc binding indicate an EC₅₀ of 483nM for the BIM-TOM conjugate, compared to dramatically right-shifted potency for the BIM-TMR conjugate. The reported affinity of BIM-TOM is more consistent with literature reports compared to the BIM-TMR (J Biol Chem. 1991 Aug 25; 266(24):15771-81.).

Example 12

Target Identification Across p38/MAPK Pathway-Determination of Selectivity

The following example serves to demonstrate the ability to profile the selectivity of a drug tracer against a panel of putative targets within a given phylogenetic target family. Similar experimental configurations may be used to identify the target of fluorescently labeled drugs using BRET in cells. This configuration may lead to the identification of the primary drug target,

as well as off-target interactors. Engagement of multiple targets may be indicative of drug promiscuity and potential drug side effects. This example serves to demonstrate the ability to measure both primary, as well as secondary target interactions via BRET.

HEK293 cells were transfected with plasmid DNA encoding N or C-terminal NanoLuc fusions to various members of the MAPK pathway (Jnk1, Jnk2, Jnk3, p38alpha, p38beta, p38gamma, p38 delta, or PKCalpha or MAPK 8, 9, 10, 14, 11, 12, or 13, PKCalpha respectively) using Fugene HD (Promega Corporation) at a 3:1 lipid:DNA ratio and seeded into 96-well plates at a density of 20,000 cells per well (yield 50ng/well of DNA). Twenty-four hours post-transfection, cell medium was replaced with serum-free medium (Opti-MEM) and incubated with 2uM PBI-4838 in the presence or absence of BIRB796 at 4uM. Cells were incubated at 37°C for two hours. To the NanoLuc-expressing samples, furimazine was added to a final concentration of 20uM. BRET was then measured on a Varioskan luminometer equipped with 450nm bandpass and 630nm longpass filters. The BRET ratio was determined by dividing the signal in the 630 channel by the signal in the 450 channel. To determine response ratios, the BRET values with tracer alone were divided by the BRET values with tracer + unmodified BIRB796.

Figure 13 indicates selectivity of PBI-4838 to Jnk2, p38beta and p38alpha, consistent with literature reports. P38alpha is recognized as the primary target of BIRB796. However, interactions with targets such as Jnk2 and p38beta could indicate potential off-target liabilities. As expected, PKCalpha-NanoLuc fusion combined with PBI-4838 showed relatively little specific BRET signal owing to poor affinity of BIRB796 toward this target.

Example 13

Target Identification Across p38/MAPK Pathway-Determination of Affinity

The following example serves to demonstrate the ability to profile the affinity of a drug tracer against a panel of putative targets within a given phylogenetic target family. Similar experimental configurations may be used to characterize the affinity of the fluorescently labeled drug to a given target using BRET in cells.

HEK293 cells were transfected with plasmid DNA encoding NanoLuc-p38alpha, NanoLuc-p38beta, Jnk2-NanoLuc, PKCalpha-NanoLuc, or NanoLuc-HDAC6 using Fugene HD (Promega Corporation) at a 3:1 lipid:DNA ratio and seeded into 96-well plates at a density of 20,000 cells per well (yield 50ng/well of DNA). Twenty-four hours post-transfection, cell medium was replaced with serum-free medium (Opti-MEM) and incubated with serially-diluted PBI-4838 in the presence or absence of BIRB796 at 4uM. Cells were incubated at 37°C for two hours. To a separate set of samples, transfected cells were treated with serially-diluted BIRB796

in the presence of 1uM PBI-4838. To the NanoLuc-expressing samples, furimazine was added to a final concentration of 20uM. BRET was then measured on a Varioskan luminometer equipped with 450nm bandpass and 630nm longpass filters. The BRET ratio was determined by dividing the signal in the 630 channel by the signal in the 450 channel.

Figures 14 and 15 demonstrate the apparent affinity of PBI-4838 to Jnk2, p38beta, and p38alpha. As expected, PKCalpha and HDAC6 showed relatively little specific BRET signal owing to poor affinity of PBI-4838 to these targets. Furthermore, the high affinity of BIRB796 toward Jnk2, p38alpha, and p38beta is demonstrated by competitive displacement in cells. Similar experiments could be configured to rank-order the affinity of drug tracers to a given set of targets (by an increase in BRET) or unmodified drugs (by competitive displacement). As indicated above, high-affinity interactions with targets such as Jnk2 and p38beta could indicate potential off-target liabilities.

Example 14

Affinity Determination of a Drug Tracer for Wild-type versus Mutant Version of Target Protein

The following example serves to demonstrate the ability to measure the relative affinity of a drug tracer to a wild-type versus mutant target protein. In this example, the affinity of a fluorescently-labeled iBET compound (small-molecule inhibitor that binds BRD4 preventing its interaction with acetylated histones) was determined for wild-type and mutant version (N140A/N433A; lacks full binding capacity to iBET) of BRD4 (Bromodomain-containing protein 4). Similar experimental configurations may be used to characterize the affinity and selectivity of the fluorescently-labeled drug to a disease-relevant mutant protein in cells. Such experiments may be useful to identify drugs that selectively bind to wild-type or mutant proteins differentially.

HEK293 cells were transfected with plasmid DNA encoding NanoLuc-BRD4 or NanoLuc-BRD4 (N140A/N433A) using Eugene HD (Promega Corporation) at a 3:1 lipid:DNA ratio and seeded into 96-well plates at a density of 20,000 cells per well (yield 50ng/well of DNA). Twenty-four hours post-transfection, cell medium was replaced with serum-free medium (Opti-MEM) and incubated with serially-diluted PBI-4966 (iBET conjugated to TOM dye) in the presence/absence of iBET at 10uM. Cells were incubated at 37°C for two hours. To the NanoLuc-expressing samples, furimazine was added to a final concentration of 20uM. BRET was then measured on a Varioskan luminometer equipped with 450nm bandpass and 630nm longpass filters. The BRET ratio was determined by dividing the signal in the 630 channel by the signal in the 450 channel.

Figure 16 indicates relative affinities of PBI-4966 to wild-type BRD4 versus the mutant BRD4. As expected, a right-shifted potency is observed with the mutant BRD4 indicating a lowered affinity for PBI-4966. Similar experiments could be configured to rank the order of the affinity of drug tracers to disease-relevant mutants of a target. This principle could be expanded to measuring relative affinities of drugs toward wild-type or mutant proteins by competitive tracer displacement. Since increased BRET signal is only indicative of binding to the NanoLuc fusion, this method enables measurements of target engagement amidst a complex mixture of similar antigens present in the assay sample. These antigens could include targets of similar properties, but non-identical to the target of interest (for instance, to discriminate heterologous vs. endogenous analyte or mutant vs. wt. analyte, etc.).

Example 15

Screening Compound Panel to Determine Target Engagement

The following example demonstrates that the method of the present invention can be used to screen compound panels to determine target engagement. This screening method could be expanded to larger compound libraries (e.g. LOPAC) as well.

HEK293 cells were transfected with NanoLuc-HDAC6 DNA using Fugene HD and seeded into wells of a 96-well plate. For the transfection, NanoLuc-HDAC6 DNA was diluted with promoterless carrier DNA (pGEM3Z). The final DNA concentration/well remained 50ng/well; however the NanoLuc-HDAC6 DNA was diluted 1:1000 (at a seeding density of 20,000 cells/well in 96-well format). Twenty-four hours post-transfection, cells were treated with serially-diluted inhibitors (SEE Figure 17) in the presence of a fixed concentration (1uM) of SAHA-TOM conjugate (PBI-4968). After two hours of incubation, furimazine was added to 20uM, and BRET detected on a Varioskan luminometer.

The results in Figure 17 demonstrate that structurally-distinct compounds can be identified by competitive displacement of drug tracers using the method of the present invention. Furthermore, relative potency of various compound classes can be confirmed. The results indicate little/no binding efficiency of apicidin to HDAC6 compared to high binding efficiency of panabinoestat or SAHA to HDAC6. This is consistent with literature reports using orthogonal assay formats. This screening method could be expanded to include high-throughput screening of large chemical libraries to identify/characterize new chemical matter with inhibitory potential against a given target.

Example 16

Monitoring Drug-Target Interactions

It is commonly accepted that a high degree of non-specific BRET occurs in medium containing a luciferase in the presence of a fluorophore of significant spectral overlap (Couturier 2012). This has been described as a liability to using BRET for high-throughput chemical screens. The following example serves to demonstrate the ability to monitor drug/target interactions using fluorophore-drug conjugates in the presence of a luciferase, NanoLuc, in medium at concentration ranges commonly recognized as problematic for BRET.

HEK293 cells were transfected with plasmid DNA encoding CDK2-NanoLuc (diluted 1:100 into pGEM3Z carrier DNA) using Fugene HD (Promega Corporation) at a 3:1 lipid:DNA ratio and seeded into wells of 96-well plates at a density of 20,000 cells per well (yield 50ng/well of total DNA). Twenty-four hours post-transfection, cell medium was replaced with serum-free medium (Opti-MEM) and incubated with serially-diluted PBI-5077 (purvalanol B conjugated to TOM dye) in the presence/absence of Purvalanol B at 10uM. Cells were incubated at 37°C for two hours.

To the NanoLuc-expressing samples, furimazine was added to a final concentration of 20uM. BRET was then measured on a Varioskan luminometer equipped with 450nm bandpass and 630nm longpass filters. The BRET ratio was determined by dividing the signal in the 630 channel by the signal in the 450 channel (Figure 18).

This example supports use of fluorophore tracers in medium for monitoring drug/target interactions via BRET. By co-incubating the sample (+ drug tracer) with purvalanol B (unmodified drug), the non-specific BRET signal between the NanoLuc and the tracer can be measured. Over the tracer concentration range tested, this background BRET signal was low. These results demonstrate that specific binding between the drug tracer and the NanoLuc-target fusion can be monitored in cell medium despite significant spectral overlap between the NanoLuc (donor) and fluorophore (acceptor).

Example 17

Target ID Screening using Protein Arrays

The following example demonstrates the ability to generate NanoLuc fusion arrays using cell-free expression and measure drug binding and relative affinity by BRET.

A panel of 11 kinases (N- and C- terminal NanoLuc fusions) including putative targets of BIRB796 (such as members of the MAPK and Jnk families), putative targets of Dasatinib (such as Src and LCK) as well as an irrelevant HDAC6 negative control was expressed in TnT® T7 Quick Coupled Reticulocyte Transcription/Translation System (Promega Corporation) as recommended by the manufacturer to create an array with 23 different NanoLuc fusions. Each TNT® reaction was diluted 1:100 in PBS and plated in replicates into wells of 96-well plates.

The NanoLuc fusion array was screened with 2 different TOM-drug tracer (BIRB-TOM; **PBI-4838** and Dasatinib-TOM; **PBI-5170**) in the presence and absence of excess relevant free drug. For each TNT® reaction, 4 replicates with 2uM BIRB TOM/4 replicates with 2uM BIRB-TOM +4uM BIRB796 (total of 2x 96-well plates) and 4 replicas with 1uM Dasatinib-TOM/ 4 replicates with 1uM Dasatinib-TOM+5uM Dasatinib (total of 2x 96-well plates) were performed. The reactions were incubated with constant mixing for 2h at room temperature. Following the incubation, furimazine was added to a final concentration of 20uM. BRET was then measured on a Varioskan luminometer equipped with 450nm bandpass and 610nm longpass filters. The BRET ratio was determined by dividing the signal in the 610 channel by the signal in the 450 channel. To determine response ratios, the BRET values with tracer alone were divided by the BRET values with tracer + unmodified.

The results (Figure 19) demonstrate selectivity of the BIRB-TOM tracer to JNk2, JNk3, MAPK14, MAPK11 and MAPK13 as well as selectivity of the Dasatinib–TOM tracer to ABL1, Src, LCK, MEK5, MAPK11 and MAPK14 consistent with literature reports. These results demonstrate the ability to profile the drug tracer's against a panel of putative targets expressed in cell free expression system and to identify hits. This no wash protein NanoLuc fusion array configuration may lead to the identification of the primary drug target, as well as off-target interactors.

Following the above screen, the BIRB-TOM hits were further analyzed for binding affinity to the BIRB-TOM tracer and competitive displacement of the BIRB TOM Tracer with the BIRB796. Binding affinity experiments to the BIRB-TOM tracer were preformed by incubating 3 replicates of the diluted TNT reactions with serially-diluted BIRB-TOM tracer in the presence/absence of BIRB796 at 10uM. Reactions were incubated at room temperature with constant mixing for two hours and after adding furimazine to a final concentration of 20uM analyzed as described above. Competitive displacement experiments were preformed by incubating 3 replicates of the diluted TNT reactions with serially-diluted BIRB796 in the presence of BIRB-TOM tracer at 2uM. Reactions were incubated at room temperature with constant mixing for two hours and after adding furimazine to a final concentration of 20uM analyzed as described above. The competitive displacement experiments results in Figure 19 indicates that BIRB796 had highest affinity to MAPK14 and Jnk2 with lower affinity to MAPK11, MAPK13, and Jnk3. Consistent with literature reports, BIRB796 binds efficiently to MAPK14, but also engages other MAPK and Jnk family members indicating off-target liabilities for BIRB-796. These results demonstrate that hits expressed in cell free expression systems and identified in the protein array screen can be further analyzed by BRET for binding affinity to the

drug. Furthermore these results support the use of cell-free expression systems to generate NanoLuc protein fusions as suitable analytes for target engagement applications.

Example 18

Expression Level of NanoLuc Fusions

The following example demonstrates that high affinity interaction with the drug tracer with optimal signal to background is achieved when the expression of the NanoLuc fusion is close to endogenous level. As previously described, it was demonstrated that a 1:1000 dilution of the NanoLuc HDAC6 DNA with a carrier DNA was required to achieve a high affinity interaction. In this example, the expression of serial dilutions of the NanoLuc HDAC6 DNA was compared to the expression of the endogenous HDAC6 using western blot analysis.

HEK293 cells were seeded into wells of 6-well plates at 2.5×10^5 cells/well and transfected with PEI with varying amounts of NanoLuc HDAC6 DNA. The NanoLuc HDAC6 DNA was diluted with a non-expressing carrier DNA (pCI neo). The final concentration of DNA remained 2ug/well while the NanoLuc HDAC6 DNA was diluted 1:0, 1:10, 1:100, 1:1000 and 1:10000. The control was un-transfected cells. 24 hours post transfection, the media was removed, and cells washed with PBS and lysed for 10 min with 400ul of mammalian lysis buffer (Promega) +1:50 RQ1 DNase (Promega) +1x RQ1 buffer. Following lysis, 133ul of 4X SDS-loading buffer was added to each well, and the cell lysates were collected, analyzed on a SDS-PAGE gel and electro-transferred onto a PVDF membrane (Invitrogen). The membrane was blocked for 1 hour with 5 % BSA (Promega) in TBS buffer and probed overnight at 4°C with 1:500 dilution of HDAC6 antibody (Millipore) in TBS supplemented with 0.1% tween (TBST). After 3 x washes in TBST, the membrane was incubated with Anti-Rabbit HRP antibody (Jackson) in TBST for 1 hour, then washed 5 x with TBST and 1 x with TBS. The membrane was then incubated for 1min with the ECL substrate from Promega and scanned on the Image Quant LAS4000 (GE). The results (Figure 10) indicate that dilution of 1:1000 of the NanoLuc HDAC6 DNA results with expression level that is similar to the expression level of endogenous HDAC6. This is the same dilution that provided the high affinity interaction with the drug tracer with optimal signal to background.

Example 19

Quantification of binding of fluorescent cytokines to a fusion of a cell surface receptor and NanoLuc

Experiments were conducted during development of embodiments of the present invention to demonstrate the ability to measure the relative affinities of therapeutic antibodies against a cell surface receptor by competitive displacement of a fluorescently-labeled cytokine.

In this example, commercially-available TAMRA-epidermal growth factor (TMR-EGF; Life Technologies) was applied in a cellular BRET assay using a stable cell line expressing a NanoLuc-EGFR (Her1) fusion protein. The BRET signal generated between TMR-EGF and NanoLuc-EGFR can be inhibited upon treatment with therapeutic antibodies known to interfere with binding of EGF.

A HEK293 stable cell line was generated by transfection of pF5 NanoLuc-EGFR plasmid DNA (Promega Corp.) and FuGene HD (Promega Corp.). After transfection, the stable cell line was generated by G418 selection, followed by limited dilution cloning. Clonally-derived cells expressing NanoLuc-EGFR were seeded into 96-well plates at a density of 20,000 cells per well. 20 hours after seeding, medium was replaced with serum-free OptiMEM and cells were serum starved. After starvation, cells were treated with serially-diluted TMR-EGF (Life Technologies). Following equilibration with the TMR-EGF, furimazine (a coelenterazine derivative substrate for NanoLuc; Promega Corp.) was added at a concentration of 20uM, and BRET was quantified on a Varioskan luminometer. In subsequent experiments, antibody-mediated displacement of TMR-EGF was measured by pretreating cells with Vectibix, Erbitux, or Herceptin (negative control) prior to stimulation with TMR-EGF (10ng/mL final concentration).

Results in Figure 20 demonstrate that the BRET can be used to quantify binding of fluorescent cytokines to cell surface receptors tethered to NanoLuc. Furthermore, the affinities of therapeutic antibodies capable of inhibiting cytokine binding can be measured by competitive displacement of the fluorescent cytokine. Vectibix and Erbitux were capable of inhibiting the binding of TMR-EGF to NanoLuc-EGFR, whereas Herceptin (a Her2 binder with negligible affinity to Her1) was incapable of inhibiting TMR-EGF binding. This example demonstrates that the binding of a fluorescent cytokine to a known high-affinity target can be quantified by BRET, in a format that can be readily reconfigured to monitor binding of therapeutic antibodies (e.g. Vectibix, Erbitux, etc.) or other binders (e.g. cytokine biologics) by competitive displacement of the fluorescent cytokine. This demonstrates the ability to use BRET as a technique to measure the relative potencies of biologic drugs in a cellular format.

Example 20

BRET Applications

In some embodiments, the present invention enables the detection/measurement of three moieties coming together: (1) A peptide of a bioluminescent reporter protein and (2) a polypeptide of a bioluminescent reporter protein which interact through structural complementation to form a complex, and (3) a fluorescent third moiety (e.g., fluorescent small molecule).

A) This example demonstrates energy transfer from a bioluminescent complex formed from a structurally complementary peptide (SEQ ID NO: 6) and polypeptide (SEQ ID NO: 7), which form a bioluminescent complex. A fluorescent dye was attached to the complementary peptide sequence. Alternatively, a fluorescent protein could be fused (e.g., a fusion protein) with a peptide or polypeptide (e.g., created from a genetic construct).

E. coli clarified lysate expressing the complementary polypeptide sequence was prepared (poly lysate). 40uL of the poly lysate was mixed with 10uL of complementary peptide (pep) or fluorescently-labeled complementary peptide (pep-TMR) and incubated for 10min at RT. 50uL 100uM substrate for the complex (furimazine) in 50mM HEPES pH 7.4 was added and incubated for 30min at RT. Luminescence was measured over 400-700nm on TECAN M1000.

Figure 21 illustrates very efficient energy transfer from the Poly/Pep complex (donor) to TMR (acceptor), and the corresponding red shift in the wavelength of light being emitted.

B) This example demonstrates using BRET in detection, such as detecting small molecule concentration or enzymatic activity. Because energy transfer is strongly dependent on distance, the magnitude of energy transfer can often be related to the conformation of the system. For instance, insertion of a polypeptide that chelates calcium can be used to measure calcium concentration through modulation of energy transfer.

An enzyme that also changes the distance, either through causing a conformational change of the sensor as above or through cleavage of the sensor from the fluorescent moiety, can be measured through a system as described herein. One structurally complementary member of a complex is bound to a fluorescent moiety which transfers its energy when the structurally complementary peptide and polypeptide of a bioluminescent reporter protein interact. One example of this is a peptide sensor that has been made wherein the peptide is conjugated to a fluorescent dye via a linker (e.g., DEVD Caspase-3 cleavage site). When exposed to the complementary polypeptide, energy transfer is observed. When exposed to Caspase-3, the fluorescent moiety is released from the complex, and energy transfer is eliminated but luminescence at 460nm remains.

A complementary polypeptide (SEQ ID NO: 8) and NL-HT (NanoLuc fused to HaloTag) were purified. 20uL of 8pM NL-HT was mixed with 20uL of 100nM PBI-3781 (See, e.g., U.S. Pat. App. Ser. No. 13/682,589, herein incorporated by reference in its entirety) and incubated for 10min at RT. 40uL NanoGlo+100uM furimazine was added, and luminescence measured over 300-800nm on TECAN M1000.

20uL of 33ng/uL of a complementary polypeptide (SEQ ID NO: 6) was mixed with 20uL of ~500uM PBI-5074 (fluorescent dye-linker-complementary peptide). 40uL NanoGlo+100uM furimazine was added, and luminescence measured over 300-800nm on TECAN M1000.

Figure 22 illustrates energy transfer from the complementary peptide/polypeptide complex (donor) to the TOM-dye of PBI-5074 (acceptor), and the corresponding red shift in the wavelength of light being emitted.

C) Ternary Interactions

The energy transfer with a structurally complementary pair of a bioluminescent reporter protein can also be used to measure three molecules interacting. One example is a GPCR fused to a complementary polypeptide of a bioluminescent reporter protein and a GPCR interacting protein fused with a complementary peptide of the bioluminescent reporter protein that form a bioluminescent complex when they interact. This allows measurement of the binary interaction. If a small molecule GPCR ligand bearing a fluorescent moiety for energy transfer interacts with the complex, energy transfer occurs. Therefore, the binary protein-protein interaction and the ternary drug-protein-protein interaction are measured in the same experiment. Also, the fluorescent molecule only causes a signal when interacting with the protein pair, which removes any signal from the ligand interacting with an inactive protein (Figure 23).

Example 21

BRET with multi-component donor

The following example demonstrates that ligand-receptor interactions can be monitored via energy transfer from a multi-component luminescent donor, where light emission requires simultaneous presence of the donor components. In this example, the donor is comprised of two complementary subunits, an 11S polypeptide and a PEP-80 peptide. The 11S polypeptide was genetically tethered to the N-terminus of BRD4, and the fusion protein expressed in HEK293 cells. Luminescence was generated from the fusion protein only in the presence of both furimazine () and the PEP-80 complementary polypeptide (Figure 24A). Cell lysates incubated with furimazine alone (no Pep-80 peptide) did not generate significant donor signal or acceptor signal (Figures 24A, 25A). When a solution of fluorescent BRD4 ligand (iBET-NCT) was applied, dose-dependent BRET signal was generated only in the presence of the PEP-80 peptide (Figure 24B, 25A, 25B). Furthermore, this BRET signal could be diminished through competitive binding of by a non-fluorescent BRD4 ligand demonstrating signal specificity (Figure 25A).

HEK293 cells were transfected with 11S-BRD4 DNA using Fugene HD and seeded in a 96-well plate format. For the transfection, 11S-BRD4 DNA was diluted with promoterless carrier DNA (pGEM3Z). The final DNA concentration/well remained 50ng/well; however the

11S-BRD4 DNA was diluted 1:10 (at a seeding density of 20,000 cells/well in 96-well format). Twenty hours post-transfection, cells were lysed in OptiMEM + 50ug/mL digitonin in the presence of absence of 1uM PEP-80 peptide. Cells were then treated with serially-diluted iBET-NCT (PBI-4966) in the presence of or absence of 10uM iBET as a specificity control. After 1 hour of incubation, furimazine was added to 10uM, and BRET detected on a Varioskan luminometer.

The concept illustrated by this example can be applied to detecting proximity of a fluorescently labeled ligand to protein complexes (e.g. detecting ligand binding selectively to heterodimeric or homodimeric receptors) by linking members of the complex to components of a multicomponent luminescent donor. This application could be thus used to monitor target engagement within disease-relevant protein complexes.

Example 22

Experiments were conducted during development of embodiments of the present invention to demonstrate that intracellular ligand-receptor interactions can be monitored via energy transfer from a protein complex, where light emission requires proximity of the luminescent donor components, facilitated by the interaction between to proteins. In this example, the donor is comprised of two subunits, an 11S peptide and a 114 peptide. The 11S peptide was genetically tethered to the N-terminus of BRD4, and the 114 peptide was genetically tethered to the C-terminus of Histone H3.3. These fusion proteins were simultaneously expressed in HEK293 cells. Luminescence was generated from the fusion protein only in the presence of both fusion constructs (Figure 26A). Cells expressing 11S-BRD4 alone, H3.3-114 alone or various control constructs did not generate significant donor signal or acceptor signal (Figures 26). When a solution of fluorescent BRD4 ligand (iBET-NCT/PBI-4966) was applied, dose-dependent acceptor signal was generated only in samples co-transfected with 11S-BRD4 and Histone H3.3-114 (Figure 27). When expressed as a BRET ratio (610/450), specific BRET was observed only from a facilitated complex of the 11S and 114 subunits and iBET-NCT or NANOLUC-BRD4 and iBET-NCT. Furthermore, this BRET signal was diminished in a dose-dependent manner through competitive binding of a non-fluorescent BRD4 ligand (IBET 151) demonstrating signal specificity (Figure 28).

HEK293 cells were transfected using Fugene HD with BRD4 DNA fusion constructs +/- Histone H3.3 DNA fusion constructs and seeded in a 96-well plate format. For the transfection, the reporter constructs were diluted with promoterless carrier DNA (pGEM3Z). The final DNA concentration/well remained 50ng/well; however each DNA expression construct was diluted

1:10 (at a seeding density of 20,000 cells/well in 96-well format). Twenty hours post-transfection, cells were treated with serially-diluted iBET-NCT (PBI-4966) in the presence or absence of 10uM iBET 151 as a specificity control. To determine if iBET-151 could compete with iBET-NCT tracer in a dose-dependent manner, transfected cells were treated with serially diluted iBET-151 in the presence of a fixed concentration (2uM) of iBET-NCT/PBI-4966 tracer. After 1 hour of incubation, furimazine was added to 10uM, and BRET detected on a Varioskan luminometer.

The concept illustrated by this example can be applied to detecting proximity of a fluorescently-labeled ligand to protein complexes (e.g., detecting ligand binding selectively to heterodimeric or homodimeric receptors) by linking members of the complex to components of a multicomponent luminescent donor. This application could be used to monitor target engagement within protein complexes. Moreover, this technique could enable the identification or characterization of compounds selective to disease-relevant intracellular protein complexes.

Example 23

Experiments were conducted during development of embodiments of the present invention to demonstrate the ability to profile the intracellular selectivity and affinity of a drug against a panel of putative targets within a given phylogenetic target family using BRET. In this example, engagement of Vorinostat (SAHA) is profiled over the entire panel of Class I/II/IV (non-sirtuin) HDACs within cells. First, the fluorescent SAHA derivative (SAHA-NCT) is profiled against various NANOLUC/HDAC fusions using BRET. Once energy transfer is confirmed between individual NANOLUC/HDAC fusions and the SAHA tracer, affinity of SAHA is determined by competitive disruption of each energy transfer complex. Affinities can then be inferred by the IC50 value generated in each experiment. Furthermore, the affinities of inhibitors to individual domains can be determined via genetic fusion of NANOLUC to the segregated domain of the target protein.

Determination of SAHA tracer affinities to individual HDACs within cells. HeLa cells were transfected with plasmid DNA encoding NANOLUC fusions with the entire family of non-sirtuin histone deacetylases (HDACs 1-11). Cells were transfected by diluting each DNA construct at a mass ratio of 1:100 into pGEM3Z carrier DNA and forming lipid DNA complexes using Fugene HD (Promega Corporation) at a 3:1 lipid:DNA ratio. Transfected cells were then seeded into 96-well plates at a density of 20,000 cells per well (total yield 50ng/well of DNA). Twenty-four hours post-transfection, cell medium was replaced with serum-free medium (Opti-MEM) and incubated with serially-diluted PBI-4968 in the presence or absence of SAHA at

20uM (the latter samples used to quantify non-specific BRET). Cells were incubated at 37°C for two hours. To the NANOLUC-expressing samples, furimazine was added to a final concentration of 10uM. BRET was then measured on a BMG Clariostar luminometer equipped with 450nm bandpass and 610nm longpass filters. The BRET ratio was determined by dividing the signal in the 630 channel by the signal in the 450 channel. To determine specific, background corrected BRET values, the nonspecific BRET values at each concentration of tracer were subtracted from BRET values with tracer alone.

Example 24

The following example serves to demonstrate the ability to measure the binding of a compound to an intracellular target with single cell resolution by using BRET imaging. This example shows the binding of SAHA and its fluorescent derivative, SAHA-NCT (PBI-4968) to NanoLuc-HDAC6 and HDAC10-NanoLuc in living cells.

Measurement of SAHA and SAHA-NCT binding to HDAC6 and HDAC10 in living cells by BRET imaging. HeLa cells were transfected with plasmid DNA encoding NanoLuc fusions with HDAC6 and HDAC10 respectively. Cells were transfected by diluting each DNA construct at a mass ratio of 1:100 into pGEM3Z carrier DNA and forming lipid DNA complexes by adding Eugene HD (Promega Corporation) at a 3:1 lipid:DNA ratio. The transfection complex was mixed with a cell suspension (1e5 cells per ml) followed by plating into glass-bottom 35 mm dishes at a density of 200,000 cells per dish (total yield 50ng/well of DNA). Twenty-four hours post-transfection, cell medium was replaced with serum-free medium (Opti-MEM) and incubated with 2 mM PBI-4968 in the presence or absence of SAHA at 10 mM. Cells were incubated at 37°C for two hours. To the NanoLuc-expressing samples, furimazine was added to a final concentration of 10uM. The samples were imaged using an Olympus LV200 Bioluminescent Microscope. In order to measure the binding and displacement of PBI-4968 to either NanoLuc-HDAC6 or HDAC10-Nanoluc by BRET, sequential images were acquired using a 495 shortpass and a 590 nm longpass filter respectively (Figure 34 and 35). Image sections representing individual cells were quantitatively analyzed using the image analysis program ImageJ. The BRET ratio was determined by dividing the value obtained for the 590 channel by the value of the 450 channel. A minimum of 100 individual cells was analyzed for each sample. All data are summarized in a dot plot format (Figure 36). Specific binding of SAHA and SAHA-NCT was demonstrated by comparative analysis of BRET values obtained from three different samples:

Sample 1 – negative control (untreated),

Sample2 – positive control (2 mM PBI-4968),

Sample 3 – Tracer displacement (2 mM PBI-4968 + 10 mM SAHA)

The results demonstrate the cellular uptake and specific binding of PBI-4968 as well as SAHA to different members of the HDAC family (HDAC6 and 10) on a single cell level. The results suggest that the degree of uptake and/or binding varies considerably with in a cell population. The image based BRET assay format enables the analysis of target specific compound binding on a single cell level to determine differences based on cell type, cell density, cell cycle status and other physiological parameters that require single cell resolution for analysis.

All publications and patents mentioned in the present application and/or listed below are herein incorporated by reference. Various modification and variation of the described methods and compositions of the invention will be apparent to those skilled in the art without departing from the scope and spirit of the invention. Although the invention has been described in connection with specific preferred embodiments, it should be understood that the invention as claimed should not be unduly limited to such specific embodiments. Indeed, various modifications of the described modes for carrying out the invention that are obvious to those skilled in the relevant fields are intended to be within the scope of the following claims.

CLAIMS

1. An assay system comprising:
 - (a) a small molecule bioactive agent tethered to a small molecule fluorophore;
 - (b) a cell that expresses a protein fusion of: a cellular target and a bioluminescent reporter, wherein the small molecule bioactive agent is capable of binding non-covalently to the cellular target upon interaction therewith, wherein the emission spectrum of the bioluminescent reporter overlaps with the absorption spectrum of the fluorophore; and
 - (c) a substrate for the bioluminescent reporter.
2. The system of claim 1, wherein the fluorophore is a carboxy rhodamine analog.
3. The system of claim 1, wherein the bioluminescent reporter comprises a polypeptide with at least 70% sequence identity with SEQ ID NO.: 1.
4. The system of claim 1, wherein upon binding of the small molecule bioactive agent to the cellular target, conversion of the substrate to a reaction product by the bioluminescent reporter results in excitation of the small molecule fluorophore by BRET and fluorescence emission from the fluorophore.
5. The system of claim 1, wherein the small molecule bioactive agent is one of a library of small molecule bioactive agents tethered to fluorophores.
6. The system of claim 1, wherein the cellular target is one of a plurality of cellular targets fused to bioluminescent reporters.
7. A method for detection of an interaction between a small molecule bioactive agent and a cellular target comprising:
 - (a) expressing in a cell a fusion of said cellular target and a bioluminescent reporter, the bioluminescent reporter having a first emission spectrum ;

- (b) contacting said cell extracellularly with a conjugate of said small molecule bioactive agent tethered to a small molecule fluorophore, said small molecule fluorophore having an excitation spectrum that overlaps said first emission spectrum and having a second emission spectrum that is distinct from said first emission spectrum, wherein the conjugate is cell permeable, and wherein the small molecule bioactive agent is capable of binding non-covalently to the cellular target upon interaction therewith;
- (c) contacting said cell with a substrate for said bioluminescent reporter, wherein the substrate is cell permeable;
- (d) detecting fluorescence at within said second emission spectrum, wherein the intensity of said fluorescence within the second emission spectrum correlates with binding between said small molecule bioactive agent with said cellular target.