

NEUTRALIZING EPI TOPE-BASED GROWTH ENHANCING VACCINE

CROSS-REFERENCE TO RELATED APPLICATIONS

5 This application is a non-provisional application that claims priority under 35 U.S.C. § 119(e) of provisional application U.S. Serial No. 60/533,719 filed December 31, 2003, the contents of which are hereby incorporated by reference in their entireties.

FIELD OF THE INVENTION

10 The invention relates to the protein growth differentiation factor 8, and to antigenic peptide fragments of growth differentiation factor 8, and related immunogens, vaccines, and methods of treating animals in order to modulate the activity of growth differentiation factor 8.

BACKGROUND OF THE INVENTION

Growth differentiation factor 8 is a protein that is classified with the transforming growth factor- β ("TGF- β ") superfamily. Generally, the proteins of the TGF- β superfamily are initially expressed as precursor (a/k/a prohormone) that undergoes proteolytic cleavage at a cluster of basic residues about 110-140 amino acids from the precursor protein C-terminus. In each case, the active, or mature, TGF- β species is believed to be a disulfide-linked dimer of the cleaved precursor protein C-terminal regions.

25 Growth differentiation factor 8, hereinafter GDF8, is also art-known as GDF-8 or myostatin. The genes encoding the precursor of GDF8 (hereinafter "precursor GDF8") have been cloned from a wide range of organisms. These include the human and murine precursor GDF8 [Nestor *et al.*, 1998, *Proc. Natl. Acad. Sci.* 95:14938-43; U.S. Patent No. 5,827,733, the contents of which are hereby incorporated by reference in their entireties]. It has also been reported that GDF8 immunoreactivity is detectable in human skeletal muscle in both type 1 and type 2 fibers. Antibodies and assays for detecting GDF8 are described, e.g., by U.S. Patent No. 6,096,506.

35 It has further been reported that GDF8 plays a role in down-regulating or inhibiting the growth and development of skeletal muscle, as confirmed by GDF8 knock-out mice (McPherron *et al.*, 1997, *Nature* 387:83-90). For this reason, there

have been previous attempts, particularly in the field of animal husbandry, to modulate GDF8 activity in animals by several means, with the goal of down-regulating GDF8 activity in order to enhance the growth, and/or relative muscle mass, of various food animals.

5 For example, U.S. Patent No. 6,399,312 describes a precursor GDF8 gene promoter and an assay, with the proposal that the assay be used to identify a theoretical inhibitor of that promotor. U.S. Patent No. 6,656,475 describes a method of inhibiting the effect of GDF8 on a cell by contacting the cell with a GDF8 prodomain that competes for a GDF8 receptor, and reports that the C-terminus of mature GDF8
10 may vary. U.S. Patent No. 6,004,937 describes the use of follistatin as a possible antagonist of GDF8. None of these methods has resulted in any practical applications in the fields of animal husbandry or clinical applications (either human or veterinary).

Others have also attempted to employ antibody and vaccine technology for downregulating GDF8 function. For instance, U.S. Patent No. 6,369,201 [the contents
15 of which are hereby incorporated by reference in their entirety], describes peptides, *i.e.*, fragments of GDF8 protein, and a vaccine for eliciting anti-GDF8 antibodies. That patent also reported an unspecified degree of growth or weight gain, relative to controls, in rodents immunized with several of the reported GDF8 peptide fragments.

Other physiological roles for GDF8 have also been described. For example,
20 U.S. Patent No. 6,368,597, [the contents of which are hereby incorporated by reference in their entirety] has suggested that inhibiting GDF8 function is useful for treating Type II diabetes, *e.g.*, by administering an anti-GDF8 antibody or anti-GDF8 vaccine to a patient having this condition.

Nevertheless, there remains a longstanding need in the art for improved
25 antigens and immunogens for eliciting an anti-GDF8 immune response, as well as for improved GDF8 antibodies capable of highly specific binding to GDF8.

The citation of any reference herein should not be construed as an admission that such reference is available as "Prior Art" to the instant application.

30 SUMMARY OF THE INVENTION

The present invention solves these and other shortcomings in the art by providing GDF8 peptides (*e.g.*, peptide fragments of GDF8 of 50 residues or less) comprising a specific neutralizing epitope for GDF8.

In one embodiment of the invention, GDF8 peptides are provided that include, for example, an isolated peptide that comprises from about residue 312 to about residue 361 of natural, human precursor GDF8 (SEQ ID NO:1). Preferably, the inventive GDF8 peptide comprises from about residue 320 to about residue 350, more preferably comprises from about residue 321 to about residue 346 and most preferably comprises from about residue 327 to about residue 346 of natural, human precursor GDF8. The exemplified GDF8 peptide, labeled as DJ5 hereinbelow, is illustrated as follows, in both single and triple letter code, along with residue numbering based on the precursor GDF8 of SEQ ID NO:1, for convenience.

DJ5 (SEQ ID NO: 8)

327	328	329	330	331	332	333	334	335	336	337	338	339	340	341	342	343	344	345	346
V	H	Q	A	N	P	R	G	S	A	G	P	C	C	T	P	T	K	M	S
Val	His	Gln	Ala	Asn	Pro	Arg	Gly	Ser	Ala	Gly	Pro	Cys	Cys	Thr	Pro	Thr	Lys	Met	Ser

In a further embodiment of the invention, the GDF8 peptide optionally includes conservative single amino acid substitutions. Simply by way of example, these can be from one through at least five amino acid positions within the peptide. In a particular embodiment, there is at least one conservative amino acid substitution, *e.g.*, between residues 327 to 346 of GDF8. In another embodiment, the GDF8 peptide includes conservative amino acid substitutions at no more than five amino acid positions within the peptide. In still another embodiment there are two conservative amino acid substitutions between residues 327 to 346 of GDF8. In yet another embodiment, there are three conservative amino acid substitutions between residues 327 to 346 of GDF8. In still another embodiment, there are four conservative amino acid substitutions between residues 327 to 346 of GDF8.

Preferably, the amino acid residue substitutions are at one or more positions, relative to natural, human precursor GDF8 (SEQ ID NO: 1) that are marked by the amino acid variations of the interspecies alignment of Fig. 2. These are at residues 328, 329, 331, 333 and 335, and combinations thereof, wherein,

- (a) amino acid residue 328 is His, Leu or Asn;
- (b) amino acid residue 329 is Gln or Lys;
- (c) amino acid residue 331 is Asn or Ser;
- (d) amino acid residue 333 is Arg or Lys; and/or
- (e) amino acid residue 335 is Ser, Pro or Thr.

Preferably, such modified GDF8 peptides comprise a specific neutralization epitope for an anti-GDF8 antibody and therefore retain the property of specifically binding to an anti-GDF8 antibody, where the antibody is mAb 788 and/or an IgG fraction of goat anti-GDF8 polyclonal antiserum, as exemplified hereinbelow.

5 In still a further embodiment of the invention, nucleic acid molecules, *i.e.*, polynucleotides, encoding the above-mentioned GDF8 peptides are provided.

Preferably, the nucleic acid molecules include a section of the naturally occurring human precursor GDF8 gene, from about nucleotide 1112 to about nucleotide 1171 of the (Genbank accession NM_005259, human GDF8 gene; SEQ
10 ID NO: 2). This section encodes peptide DJ5, as described above. Note that the NM_005259 record includes a large amount of sequence flanking the actual coding region. The artisan will also appreciate that the DJ5 corresponds to nucleotides 979-1038 of the actual coding region of the GDF8 prohormone.

Replicable cloning vectors, and prokaryotic or eukaryotic host cells comprising
15 the nucleic acid molecules are also provided, along with methods of producing a GDF8 peptide that include the steps of: culturing the host cell(s), expressing the encoded peptide, and recovering the peptide. The artisan will also appreciate that the inventive GDF8 peptide will also be readily produced by any standard, art-known chemical synthetic method.

20 In yet a further embodiment of the invention, a vaccine composition that comprises the inventive GDF8 peptide (or fusion protein thereof) is also provided, *e.g.*, that preferably includes one or more adjuvants and other art-standard elements of a peptide or protein-based vaccine composition. Methods of eliciting an anti-GDF8 immune response in an animal, comprising administering to the animal an effective
25 amount of the vaccine composition, are also provided.

In another further embodiment, a screening method is provided for selecting an anti-GDF8 antibody or antibody fragment from among a plurality of antibodies or antibody fragments, comprising contacting the peptide with one or a plurality of antibodies or antibody fragments, and detecting antibody or antibody fragment that
30 selectively binds to the peptide.

In yet another aspect, the present invention provides a method of down-regulating GDF8 activity in an animal. In one such embodiment, the method comprises administering an antibody or antibody fragment to the animal, in an amount and for a duration effective to down-regulate GDF8 activity in the animal, wherein the

antibody binds specifically to the peptide, or by immunizing the animal with a vaccine composition as described herein. The animal is preferably a vertebrate, and more preferably a mammal, avian or fish. Preferably, the mammal is a domesticated animal (e.g., one used in animal husbandry, or alternatively, a companion animal) but can optionally be a human in need of such GDF8 downregulation.

The invention also contemplates fusion proteins incorporating the inventive GDF8 peptides. The fusion proteins can include domains that are signal peptides, for enhanced secretion or cell surface expression of the GDF8 fusion protein and/or to permit purification with a selective binding system. Further, the fusion proteins are contemplated to link one or more GDF8 peptides in a single carrier protein in order to enhance immunogenicity of the GDF8 peptide domain. In a particular embodiment, a fusion protein of the present invention comprises a GDF8 peptide consisting of 50 or fewer amino acid residues that comprises amino acid residues 327 to 346 of SEQ ID NO:1. In a related embodiment of this type, the fusion protein comprises an antigenic subfragment of that GDF8 peptide, e.g., a GDF8 peptide comprising about 10 consecutive amino acid residues from DJ5 (SEQ ID NO:8).

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 illustrates overlapping peptides DJ1 through DJ7, in the GDF8 active region (*i.e.*, mature GDF8), that is from residues 266 - 375 of the precursor GDF8 sequence.

Figure 2 illustrates the alignment of the human DJ5 peptide sequence (SEQ ID NO: 8) compared to the analogous 20-residue peptides, as located in the precursor GDF8 proteins of the recited additional animal species. The amino acid residue positions of 321 through 347 are based on the human precursor GDF8. The Genbank accession numbers (incorporated by reference herein) identify the entire published protein sequence for that species.

The aligned peptides have the following SEQ ID NOs.

Anas platyrhynchos (duck) AAL35275	(SEQ ID NO: 11)
Anser anser (goose) AAL35276	(SEQ ID NO: 12)
Anser anser (goose) AAR18246	(SEQ ID NO: 13)
Bos taurus (cow) AAB86687	(SEQ ID NO: 14)
Canis familiaris (dog) AAR14343	(SEQ ID NO: 15)
Capra hircus (goat) AAR12161	(SEQ ID NO: 16)
Columba livia (pigeon) AAL35277	(SEQ ID NO: 17)
Coturnix chinensis (quail) AAL35278	(SEQ ID NO: 18)
Danio rerio (zebrafish) AAB86693	(SEQ ID NO: 19)
Equus caballus (horse) BAB16046	(SEQ ID NO: 20)
Gallus gallus (chicken) AAK18000	(SEQ ID NO: 21)
Gallus gallus (chicken) AAR18244	(SEQ ID NO: 22)
Homo sapiens (human) NP-005250	(SEQ ID NO: 8)
I. punctatus (catfish) AAK84666	(SEQ ID NO: 23)
Lepus capensis (hare) AAN87890	(SEQ ID NO: 24)
Macaca fascicularis (monkey) AAL17640	(SEQ ID NO: 25)
Meleagris gallopavo (turkey) AAB86692	(SEQ ID NO: 26)
Morone chrysops (white bass) AAK28707	(SEQ ID NO: 27)
Mus musculus (house mouse) AAC531 67	(SEQ ID NO: 28)
O. mykiss (trout) AAK71 707	(SEQ ID NO: 29)
Ovis aries (sheep) AAB86689	(SEQ ID NO: 30)
Papio hamadryas (baboon) AAB86686	(SEQ ID NO: 31)
Rattus norvegicus (rat) AAB86691	(SEQ ID NO: 32)
Salmo salar (salmon) CAC1 9541	(SEQ ID NO: 33)
Sparus aurata (seabream) AAL05943	(SEQ ID NO: 34)
Sus scrofa (pig) AAC08035	(SEQ ID NO: 35)
Sus scrofa (pig) AAR18245	(SEQ ID NO: 36)

DETAILED DESCRIPTION OF THE INVENTION

Accordingly, the present invention identifies GDF8 domains that serve as a specific neutralization epitope for a polyclonal anti-GDF8 goat antiserum and for other certain specific anti-GDF8 antibodies. These epitopes also serve to provide fragments of the GDF8 protein that are useful for eliciting an active and specific immune response against GDF8 proteins, both *in vitro*, e.g., for detection of GDF8 protein, and *in vivo*, for downregulating GDF8 activity. These fragments are generally referred to herein as GDF8 peptides or peptide fragments. The utility of these GDF8 peptides includes use as immunogens for eliciting an anti-GDF8 immune response in animals, and use as highly specific antibody-binding targets in GDF8-related assays.

The specific binding epitopes of GDF8 were identified by contacting anti-GDF8 antiserum with a battery of overlapping GDF8 peptides, and determining the degree of binding activity between the peptides and the antiserum IgG antibodies. The anti-

GDF8 antiserum was obtained from a goat immunized with a precursor GDF8 protein having a structure optimized for expression and antigenicity.

In order to more fully appreciate the instant invention, the following definitions are provided. The use of singular terms for convenience in description is in no way intended to be so limiting. Thus, for example, reference to a composition comprising
5 "a polypeptide" includes reference to one or more of such polypeptides.

As used herein the term "approximately" is used interchangeably with the term "about" and signifies that a value is within twenty percent of the indicated value *i.e.*, a peptide containing "approximately" 50 amino acid residues can contain between 40
10 and 60 amino acid residues.

It is also to be understood that this invention is not limited to the particular configurations, process steps, and materials disclosed herein as such configurations, process steps, and materials may vary somewhat. It is also to be understood that the terminology employed herein is used for the purpose of describing particular
15 embodiments only and is not intended to be limiting, since the scope of the present invention will be limited only by the appended claims and equivalents thereof.

As used herein, the term, "polypeptide" is used interchangeably with the term "protein" and denotes a polymer comprising two or more amino acids connected by peptide bonds. Preferably, unless otherwise stated herein, the term polypeptide is
20 distinguished from the term, "peptide" as employed herein, by size or chain length, wherein a "peptide" refers to a polymer chain of about fifty or fewer amino acids, and a polypeptide or protein refers to polymer chain comprising more than about fifty amino acids. Optionally, a peptide or a polypeptide may lack certain amino acid residues that are encoded by a gene or by an mRNA. For example, a gene or mRNA molecule
25 may encode a sequence of amino acid residues on the N-terminus of a polypeptide (*i.e.*, a signal sequence) that is cleaved from, and therefore, may not be part of the final protein.

A "GDF8 peptide" according to the present invention is a relatively short fragment derived from the GDF8 protein. Even subfragments of such peptides can be
30 termed GDF8 peptides. While not intending to limit the maximum size of a GDF8 peptide according to the invention, it is preferred that the maximum size of the peptide is about 50 residues, more preferably the maximum size is about 40 residues, even more preferably the maximum size is about 30 residues, and still more preferably the maximum size is about 25 residues. More generally, the GDF8 peptide preferably

ranges in size from about 10 to about 50 amino acid residues in length, more preferably from about 15 to about 30 amino acid residues, and in particular, is about 20 amino acid residues in length. GDF8 peptides that are smaller subfragments of other GDF8 peptides of the present invention, for example, a GDF8 peptide
5 comprising about 10 consecutive amino acid residues from DJ5 (SEQ ID NO:8), preferably comprise an antigenic portion (e.g., an epitope) from the larger GDF8 peptide.

In a particular embodiment, a GDF8 peptide comprises a peptide domain that has a degree of homology ranging from about 50% to 100% homology to a peptide
10 defined by residue numbers 327-346 (SEQ ID NO: 8) of the naturally occurring human precursor of GDF8 (SEQ ID NO: 1). The variations in homology noted above are preferably conservative substitutions and/or variations that retain the inventive antigenic structure that is specifically recognized by certain specific anti-GDF8 antibodies. These conserved substitutions represent residue substitutions that are
15 shown by interspecies homology comparisons (e.g., see FIG. 2) to preserve GDF8 function and/or represent residue substitutions between amino acids of analogous chemical (e.g., physical) and electronic structure, thus preserving and/or optimizing the antibody-binding specificity of the inventive GDF8 peptides. Examples of such conservative amino acid substitutions include: replacing one hydrophobic residue
20 such as isoleucine, valine, leucine or methionine for another; or replacing one polar residue of equivalent charge for another, e.g., substituting arginine for lysine, glutamic acid for aspartic acid, or glutamine for asparagine.

In particular, the GDF8 peptide according to the invention will also include a specific neutralization epitope for an anti-GDF8 antibody, *i.e.*, an epitope or antigenic
25 domain that will specifically bind to the **PGA** anti-GDF8 IgG polyclonal antibody described by the Examples below, and/or that binds specifically to commercially available rat monoclonal antibody ("mAb") Cat. No. MAB788 (R&D Systems Inc., Minneapolis, MN).

The terms "purified" or "isolated" as employed herein, refer to materials
30 separated under conditions that reduce or eliminate the presence of unrelated materials, *i.e.*, contaminants, including native materials from which the material is obtained. For example, a purified or isolated protein is preferably free of other proteins or nucleic acids with which it can be found within a cell. A purified material may contain less than about 50%, preferably less than about 75%, and most

preferably less than about 90%, of the cellular components with which it was originally associated. Purity can be evaluated by chromatography, gel electrophoresis, immunoassay, composition analysis, biological assay and other methods known in the art. From a functional aspect, an isolated GDF8 peptide according to the invention is one sufficiently separated from other materials, including precursor GDF8 protein and/or mature GDF8 protein, so as to be capable of eliciting an immune response that is specific for the GDF8 peptide.

Methods for purification are well-known in the art. For example, nucleic acids can be purified by precipitation, chromatography, ultracentrifugation and other means. Proteins and polypeptides, as well as peptides, can be purified by various methods including, without limitation, preparative disc-gel electrophoresis, isoelectric focusing, HPLC, reversed-phase HPLC, gel filtration, ion exchange and partition chromatography, precipitation and salting-out chromatography, extraction and countercurrent distribution. For some purposes, it is preferable to produce the polypeptide in a recombinant system in which the protein contains an additional sequence tag that facilitates purification, such as, but not limited to, a polyhistidine sequence or a sequence that specifically binds to an antibody, such as FLAG[®] and GST. The polypeptide can then be purified from a crude lysate of the host cell by chromatography on an appropriate solid-phase matrix. Alternatively, antibodies, or binding fragments thereof, produced against the polypeptide can be used as purification reagents.

The term "substantially pure" indicates the highest degree of purity which can be achieved using conventional purification techniques known in the art and means a nucleic acid, polypeptide, peptide, or other material that is free from other contaminating proteins, nucleic acids and other biologicals derived from an original source organism or recombinant DNA expression system. Substantial purity may be assayed by standard methods and will typically exceed at least about 75%, preferably at least about 90%, more preferably at least about 95% and most preferably at least about 99% purity. Purity evaluation may be made on a mass or molar basis.

A "polynucleotide" or a "nucleic acid molecule" is a molecule comprising nucleotides including, but is not limited to, RNA, cDNA, genomic DNA and even synthetic DNA sequences. The terms are also contemplated to encompass nucleic acid molecules that include any of the art-known base analogs of DNA and RNA.

A "vector" or "replication vector" is a replicon, such as a plasmid, phage, or cosmid, to which another DNA segment may be attached or incorporated so as to bring about the replication of the attached segment. The term also includes a replicon that includes the incorporated or attached DNA segment of interest.

5 Vectors that can be used in this invention include microbial plasmids, viruses, bacteriophage, integratable DNA fragments and other vehicles that may facilitate integration of the nucleic acids into the genome of the host. Plasmids are the most commonly used form of vector, but all other forms of vectors which serve an equivalent function and which are or become known in the art are suitable for use
10 herein. See, e.g., Pouwels *et al.*, *Cloning Vectors: A Laboratory Manual*, 1985 and Supplements, Elsevier, N.Y., and Rodriguez *et al.* (eds.), *Vectors: A Survey of Molecular Cloning Vectors and Their Uses*, 1988, Butterworth, Boston, MA.

 Insertion of DNA encoding the inventive GDF8 peptide(s) into a vector is easily accomplished when the termini of both the DNA and the vector comprise compatible
15 restriction sites. If this cannot be done, it may be necessary to modify the termini of the DNA and/or vector by digesting back single-stranded DNA overhangs generated by restriction endonuclease cleavage to produce blunt ends, or to achieve the same result by filling in the single-stranded termini with an appropriate DNA polymerase. Alternatively, desired sites may be produced, e.g., by ligating nucleotide sequences
20 (linkers) onto the termini. Such linkers may comprise specific oligonucleotide sequences that define desired restriction sites. Restriction sites can also be generated through the use of the polymerase chain reaction (PCR). See, e.g., Saiki *et al.*, *Science* 239:487 (1988). The cleaved vector and the DNA fragments may also be modified, if required, by homopolymeric tailing.

25 Recombinant expression vectors used in this invention are typically self-replicating DNA or RNA constructs comprising nucleic acids encoding one of the inventive GDF8 peptide(s), usually operably linked to suitable genetic control elements that are capable of regulating expression of the nucleic acids in compatible host cells. Genetic control elements may include a prokaryotic promoter system or a
30 eukaryotic promoter expression control system, and typically include a transcriptional promoter, an optional operator to control the onset of transcription, transcription enhancers to elevate the level of mRNA expression, a sequence that encodes a suitable ribosome binding site, and sequences that terminate transcription and

translation. Expression vectors may also contain an origin of replication that allows the vector to replicate independently of the host cell.

Expression of nucleic acids encoding inventive GDF8 peptide(s) can be carried out by conventional methods in either prokaryotic or eukaryotic cells.

5 A DNA "coding sequence" or a "sequence encoding" a particular protein or peptide, is a DNA sequence which is transcribed and translated into a polypeptide *in vitro* or *in vivo* when placed under the control of appropriate regulatory elements. The boundaries of the coding sequence are determined by a start codon at the 5'-terminus and a translation stop codon at the 3'-terminus. A coding sequence can include, but is not limited to, prokaryotic sequences, cDNA from eukaryotic mRNA, genomic DNA
10 sequences from eukaryotic (e.g., mammalian) DNA, and even synthetic DNA sequences. A transcription termination sequence will usually be located 3' to the coding sequence.

 As used herein the terms "fusion protein" and "fusion peptide" are used
15 interchangeably and encompass "chimeric proteins and/or chimeric peptides" and fusion "intein proteins/peptides". A fusion protein comprises at least a portion of a GDF8 peptide of the present invention joined via a peptide bond to at least a portion of another protein. For example, fusion proteins can comprise a marker protein or peptide, or a protein or peptide that aids in the isolation and/or purification and/or
20 antigenicity of a GDF8 peptide of the present invention. A GDF8 fusion protein can comprise at least a portion of a non-GDF8 protein joined via a peptide bond to at least a portion of a GDF8 polypeptide. In preferred embodiments a portion of the GDF8 is functional, *i.e.*, retains its antigenicity. The non-GDF8 sequences can be amino- or carboxy-terminal to the GDF8 sequences.

25 A recombinant DNA molecule encoding such a fusion protein comprises a sequence encoding at least a portion of a non-GDF8 protein joined in-frame to the GDF8 coding sequence, and can encode a cleavage site for a specific protease, e.g., thrombin or Factor Xa, preferably at or close to the juncture between the GDF8 sequence and the non-GDF8 sequence. In a specific embodiment, the fusion protein
30 is expressed in a CHO cell. Such a fusion protein can be used to isolate the GDF8 peptides of the present invention, through the use of an affinity column that is specific for the protein and/or tag fused to the GDF8 peptide. The purified GDF8 peptide, for example, may then be released from the fusion protein through the use of a proteolytic enzyme and a cleavage site such as has been referred to above.

In one such embodiment, a chimeric GDF8 peptide can be prepared, *e.g.*, a glutathione-S-transferase (GST) fusion protein, a maltose-binding (MBP) protein fusion protein, or a poly-histidine-tagged fusion protein, for expression in any cell, or alternatively in a cell-free system. For example, GST binds glutathione conjugated to a solid support matrix, MBP binds to a maltose matrix, and poly-histidine chelates to a Ni-chelation support matrix. The fusion protein can be eluted from the specific matrix with appropriate buffers, or by treating with a protease specific for a cleavage site usually engineered between the GDF8 peptide and the fusion partner (*e.g.*, GST, MBP, FLAG[®]) as exemplified below, or poly-His as described above.

A "heterologous nucleotide sequence" as used herein is a nucleotide sequence that is added to a nucleotide sequence of the present invention by recombinant methods to form a nucleic acid that is not naturally formed in nature. Such nucleic acids can encode fusion (*e.g.*, chimeric) proteins. Thus the heterologous nucleotide sequence can encode peptides and/or proteins that contain regulatory and/or structural properties. In another such embodiment the heterologous nucleotide sequence can encode a protein or peptide that functions as a means of detecting the protein or peptide encoded by the nucleotide sequence of the present invention after the recombinant nucleic acid is expressed. In still another embodiment the heterologous nucleotide sequence can function as a means of detecting a nucleotide sequence of the present invention. A heterologous nucleotide sequence can comprise non-coding sequences including restriction sites, regulatory sites, promoters and the like.

A "host cell" is a cell that contains, or is capable of containing, and expressing, an exogenous nucleic acid molecule, either transiently or permanently. A cell has been "transformed" by exogenous DNA when such exogenous DNA has been introduced inside the cell membrane. Exogenous DNA may or may not be integrated (covalently linked) into chromosomal DNA making up the genome of the cell. In prokaryotes and yeasts, for example, the exogenous DNA may be maintained on an episomal element, such as a plasmid. With respect to eukaryotic cells, a stably transformed cell is one in which the exogenous DNA has become integrated into the chromosome so that it is inherited by daughter cells through chromosome replication. This stability is demonstrated by the ability of the eukaryotic cell to establish cell lines or clones comprised of a population of daughter cells containing the exogenous DNA.

Prokaryotes include both gram negative and positive organisms, *e.g.*, *E. coli* and *B. subtilis*. Higher eukaryotes include established tissue culture cell lines from animal cells, both of non-mammalian origin, *e.g.*, insect cells, and birds, and mammalian origin, *e.g.*, human, primates, and rodents.

5 Prokaryotic host-vector systems include a wide variety of vectors for many different species. As used herein, *E. coli* and its vectors will be used generically to include equivalent vectors used in other prokaryotes. A representative vector for amplifying DNA is pBR322 or many of its derivatives. Vectors that can be used to express GDF8, and/or GDF8 peptides, include, but are not limited to, those containing
10 the lac promoter (pUC-series); trp promoter (pBR322-trp); lpp promoter (the pIN-series); lambda-pP or pR promoters (pOTS); or hybrid promoters such as ptac (pDR540). See Brosius *et al.*, "Expression Vectors Employing Lambda-, trp-, lac-, and lpp-derived Promoters", in Rodriguez and Denhardt (eds.) *Vectors: A Survey of Molecular Cloning Vectors and Their Uses*, 1988, Butterworth, Boston, pp. 205-236.

15 Yeast, as well as higher eukaryotic tissue culture cells are preferred hosts for the recombinant production of the inventive GDF8 peptides, and/or of anti-GDF8 antibodies and/or fragments of those antibodies. Although any higher eukaryotic tissue culture cell line might be used, including insect baculovirus expression systems, mammalian cells are preferred. Transformation or transfection and propagation of
20 such cells have become a routine procedure. Examples of useful cell lines include HeLa cells, Chinese hamster ovary (CHO) cell lines, baby rat kidney (BRK) cell lines, insect cell lines, bird cell lines, and monkey (COS) cell lines.

Expression vectors for such cell lines usually include, for example, an origin of replication, a promoter, a translation initiation site, RNA splice sites (if genomic DNA is
25 used), a polyadenylation site, and a transcription termination site. These vectors also usually contain a selection gene or amplification gene. Suitable expression vectors may be plasmids, viruses, or retroviruses carrying promoters derived, *e.g.*, from such sources as adenovirus, SV40, parvoviruses, vaccinia virus, or cytomegalovirus. Representative examples of suitable expression vectors include pCR[®]3.1, pCDNA1,
30 pCD [Okayama *et al.*, *Mol. Cell Biol.* 5:1136 (1985)], pMC1neo Poly-A [Thomas *et al.*, *Cell* 51:503 (1987)], pUC19, pREP8, pSVSPORT and derivatives thereof, and baculovirus vectors, such as pAC 373 or pAC 610.

Prokaryotic expression control sequences typically used include promoters, including those derived from the β -lactamase and lactose promoter systems [Chang *et*

al., *Nature*, 198:1056 (1977)], the tryptophan (trp) promoter system [Goeddel *et al.*, *Nucleic Acids Res.* 8:4057 (1980)], the lambda P_L promoter system [Shimatake *et al.*, *Nature*, 292:128 (1981)] and the tac promoter [De Boer *et al.*, *Proc. Natl. Acad. Sci. USA* 292:128 (1983)]. Numerous expression vectors containing such control

5 sequences are known in the art and commercially available.

"Operably linked" refers to an arrangement of elements wherein the components so described are configured so as to perform their usual function. Thus, control elements operably linked to a coding sequence are capable of effecting the expression of the coding sequence. The control elements need not be contiguous with
10 the coding sequence, so long as they function to direct the expression thereof. Thus, for example, intervening untranslated yet transcribed sequences can be present between a promoter and the coding sequence and the promoter can still be considered "operably linked" to the coding sequence.

The invention also includes polyclonal and monoclonal (mAb) antibodies that
15 specifically bind to the inventive GDF8 peptides. As used herein, the term "antibody" refers to an immunoglobulin and/or fragments thereof. A naturally occurring immunoglobulin consists of one or more polypeptides substantially encoded by immunoglobulin genes. The recognized immunoglobulin genes include the kappa, lambda, alpha, gamma, delta, epsilon and mu constant region genes, as well as the
20 myriad immunoglobulin variable region genes. An antibody or antibodies according to the invention also encompass antibody fragments, *i.e.*, antigen-binding fragments, for example, Fv, Fab, and F(ab')₂, engineered single-chain binding proteins, (*e.g.*, Huston *et al.*, *Proc. Natl. Acad. Sci. U.S.A.*, 85, 5879-5883 (1988) and Bird *et al.*, *Science*, 242, 423-426 (1988), hereby incorporated herein by reference in its entirety), as well
25 as bifunctional hybrid antibodies (*e.g.*, Lanzavecchia *et al.*, *Eur. J. Immunol.* 17, 105 (1987)). [See, generally, Hood *et al.*, *Immunology*, Benjamin, N.Y., 2nd ed. (1984), Harlow and Lane, Antibodies. A Laboratory Manual, Cold Spring Harbor Laboratory (1988) and Hunkapiller and Hood, *Nature*, 323, 15-16 (1986), the contents of all of which are hereby incorporated by reference in their entirety.]

30 For example, serum produced from animals immunized by the inventive GDF8 peptides, using standard methods, can be used directly, or the IgG fraction can be separated from the serum using standard methods, such as plasmaphoresis or adsorption chromatography with IgG-specific adsorbents, such as immobilized Protein A or Protein G. Alternatively, monoclonal antibodies can be prepared, and optionally,

antigen binding fragments or recombinant binding proteins derived from such mAbs. Such MAbs or fragments thereof can optionally be humanized by art-known methods.

Hybridomas producing mAbs that selectively bind the GDF8 peptides of the invention, are produced by well-known techniques. Usually, the process involves the fusion of an immortalizing cell line with a B-lymphocyte that produces the desired antibody. Alternatively, non-fusion techniques for generating immortal antibody-producing cell lines can be used, e.g., virally-induced transformation [Casali *et al.*, *Science* 234:476 (1986)]. Immortalizing cell lines are usually transformed mammalian cells, particularly myeloma cells of rodent, bovine, and human origin. Most frequently, rat or mouse myeloma cell lines are employed as a matter of convenience and availability.

Techniques for obtaining antibody-producing lymphocytes from mammals injected with antigens are well known. Generally, peripheral blood lymphocytes (PBLs) are used if cells of human origin are employed, or spleen or lymph node cells are used from non-human mammalian sources. A host animal is injected with repeated dosages of the purified antigen (human cells are sensitized *in vitro*), and the animal is permitted to generate the desired antibody-producing cells before they are harvested for fusion with the immortalizing cell line. Techniques for fusion are also well known in the art, and, in general, involve mixing the cells with a fusing agent, such as polyethylene glycol.

Hybridomas are selected by standard procedures, such as HAT (hypoxanthine-aminopterin-thymidine) selection. Those secreting the desired antibody are selected using standard immunoassays, such as Western blotting, ELISA (enzyme-linked immunosorbent assay), RIA (radioimmunoassay) or the like. Antibodies are recovered from the medium using standard protein purification techniques [Tijssen, *Practice and Theory of Enzyme Immunoassays* (Elsevier, Amsterdam, 1985)].

Many references are available to provide guidance in applying the above techniques [Kohler *et al.*, *Hybridoma Techniques* (Cold Spring Harbor Laboratory, New York, 1980); Tijssen, *Practice and Theory of Enzyme Immunoassays* (Elsevier, Amsterdam, 1985); Campbell, *Monoclonal Antibody Technology* (Elsevier, Amsterdam, 1984); Hurrell, *Monoclonal Hybridoma Antibodies: Techniques and Applications* (CRC Press, Boca Raton, FL, 1982)]. Monoclonal antibodies can also be produced using well-known phage library systems. See, e.g., Huse, *et al.*, *Science* 246:1275 (1989); Ward, *et al.*, *Nature*, 341:544 (1989).

Antibodies thus produced, whether polyclonal or monoclonal, can be used, e.g., in an immobilized form bound to a solid support by well known methods to purify the GDF8 peptides by immunoaffinity chromatography.

Antibodies against the GDF8 peptides can also be used, unlabeled or labeled
5 by standard methods, as the basis for immunoassays to detect or quantify GDF8. The particular label used will depend upon the type of immunoassay. Examples of labels that can be used include, but are not limited to, radiolabels, such as ^{32}P , ^{125}I , ^3H and ^{14}C ; fluorescent labels, such as fluorescein and its derivatives, rhodamine and its derivatives, dansyl and umbelliferone; chemiluminescers, such as luciferia and 2,3-
10 dihydrophthalazinediones; and enzymes, such as horseradish peroxidase, alkaline phosphatase, lysozyme and glucose-6-phosphate dehydrogenase.

The antibodies can be tagged with such labels by known methods. For example, coupling agents such as aldehydes, carbodiimides, dimaleimide, imidates, succinimides, bisdiazotized benzadine and the like may be used to tag the antibodies
15 with fluorescent, chemiluminescent or enzyme labels. The general methods involved are well known in the art and are described, e.g., in *Immunoassay: A Practical Guide*, 1987, Chan (Ed.), Academic Press, Inc., Orlando, FL. Such immunoassays could be carried out, for example, on fractions obtained during purification of the receptors.

The antibodies of the present invention can also be used to identify particular
20 cDNA clones expressing GDF8-related polypeptides in expression cloning systems. Neutralizing antibodies specific for the ligand-binding site of a receptor can also be used as antagonists (inhibitors) to block or downregulate GDF8 function. Such neutralizing antibodies can readily be identified through routine experimentation, as exemplified by the Examples provided below.

Antagonism of GDF8 activity can be accomplished using complete antibody
25 molecules, or well-known antigen binding fragments such as Fab, Fc, F(ab)₂, and Fv fragments. Definitions of such fragments can be found as described hereinabove, or e.g., in Klein, *Immunology* (John Wiley, New York, 1982); Parham, Chapter 14, in Weir, ed. *Immunochemistry*, 4th Ed. (Blackwell Scientific Publishers, Oxford, 1986).
30 The use and generation of antibody fragments has also been described, e.g.: Fab fragments [Tijssen, *Practice and Theory of Enzyme Immunoassays* (Elsevier, Amsterdam, 1985)], Fv fragments [Hochman *et al.*, *Biochemistry* 12:1130 (1973); Sharon *et al.*, *Biochemistry* 15:1591 (1976); Ehrlich *et al.*, U.S. Patent No. 4,355,023] and antibody half molecules (Auditore-Hargreaves, U.S. Patent No. 4,470,925).

Methods for making recombinant Fv fragments based on known antibody heavy and light chain variable region sequences have further been described, e.g., by Moore *et al.* (U.S. Patent No. 4,642,334) and by Plückthun [*Bio/Technology* 9:545 (1991)]. Alternatively, they can be chemically synthesized by standard methods.

5 The present invention also encompasses anti-idiotypic antibodies, both polyclonal and monoclonal, which are produced using the above-described antibodies as antigens. These antibodies are useful because they may mimic the structures of the ligands.

Peptide Synthesis

10 Since the inventive GDF8 peptides, e.g., the DJ5 peptide exemplified hereinbelow, are relatively short (e.g., preferably 50 amino acid residues or less), they may be prepared by art-known methods of peptide synthesis. Synthetic peptides or polypeptides, prepared using the well-known techniques of solid phase, liquid phase, or peptide condensation techniques, or any combination thereof, can include natural
15 and unnatural amino acids. Amino acids used for peptide synthesis may be standard Boc (N_{α} -amino protected N_{α} -t-butyloxycarbonyl) amino acid resin with the standard deprotecting, neutralization, coupling and wash protocols of the original solid phase procedure of Merrifield [*J. Am. Chem. Soc.*, 85:2149-2154 (1963)], or the base-labile N_{α} -amino protected 9-fluorenylmethoxycarbonyl (Fmoc) amino acids first
20 described by Carpino and Han [*J. Org. Chem.*, 37:3403-3409 (1972)]. Both Fmoc and Boc N_{α} -amino protected amino acids can be obtained from Fluka, Bachem, Advanced Chemtech, Sigma, Cambridge Research Biochemical, Bachem, or Peninsula Labs or other chemical companies familiar to those who practice this art. In addition, the method of the invention can be used with other N_{α} -protecting groups
25 that are familiar to those skilled in this art. Solid phase peptide synthesis may be accomplished by techniques familiar to those in the art and provided, for example, in Stewart and Young, SOLID PHASE SYNTHESIS, Second Edition, Pierce Chemical Co., Rockford, Ill. (1984); Fields and Noble, *Int. J. Pept. Protein Res.* 35:161-214 (1990), or using automated synthesizers, such as sold by ABS [Applied Biosystems, 850 Lincoln
30 Centre Drive, Foster City, CA 94404 USA]. Thus, the GDF8 peptides of the invention may comprise D-amino acids, a combination of D- and L-amino acids, and various "designer" amino acids (e.g., *beta*-methyl amino acids, C_{α} -methyl amino acids, and N_{α} -methyl amino acids, etc.) to convey special properties. Synthetic amino acids include ornithine for lysine, fluorophenylalanine for phenylalanine, and norleucine for

leucine or isoleucine. Additionally, by assigning specific amino acids at specific coupling steps, *alpha*-helices, *beta* turns, *beta* sheets, *gamma*-turns, and cyclic peptides can be generated.

Anti-GDF8 Antiserum

5 The methods of the invention included a process of screening GDF8 peptides against a polyclonal anti-GDF8 antiserum. This process identified the epitope that certain anti-GDF8 antibodies bind to in a highly specific way. The anti-GDF8 antiserum was obtained by immunizing an animal with precursor GDF8. The precursor GDF8 gene was modified to provide a form optimized for expression and
10 immunogenicity. For example, the natural DNA sequence of the GDF8 prohormone (SEQ ID NO: 2) was optimized for expression in mammalian and viral expression systems. In addition, changes were made to avoid the negative effects of viral host shutoff mechanisms. Typically viral host shutoff mechanisms involve transcriptional control, RNA stability (splicing) and such. These changes made the nucleic acid less
15 host like and more virus like.

 Further, the DNA sequence was preferably designed to be as divergent from the mammalian nucleic acid sequence as possible. For example, the amino acid sequence of the precursor GDF8 was reverse translated using yeast preferred codons. The resulting sequence was surveyed for codons that retained their homology
20 to the human GDF8 nucleic acid sequence. Where possible these codons were substituted with the next most preferred yeast codons encoding the same amino acid.

 The resulting optimized gene (SEQ ID NO: 3) can be expressed in any suitable host system, including, e.g., art-known insect, mammalian, bacterial, viral and yeast expression systems. For example, insect cell expression systems, such as
25 baculovirus systems, are art-known and described, for instance, by Summers and Smith, Texas Agricultural Experiment Station Bulletin No. 1555 (1987). Materials and methods for baculovirus/insect cell expression systems are commercially available in kit form from, *inter alia*, Invitrogen, San Diego Calif. ("MaxBac" kit). Similarly, bacterial and mammalian cell expression systems are well known in the art and described, for
30 example, by Sambrook *et al.* (MOLECULAR CLONING: A LABORATORY MANUAL; DNA Cloning, Vols. I and II; D. N. Glover ed.). Yeast expression systems are also known in the art and described, for example, by, YEAST GENETIC ENGINEERING (Barr *et al.*, eds., 1989) Butterworths, London. Many other such expression systems are known to the art and are available commercially in kit form. As exemplified herein, the modified

precursor GDF8 gene (SEQ ID NO: 3) was expressed in a Flp-In™ CHO expression system (Invitrogen, Carlsbad, CA) as described in greater detail by Example 1, below.

The precursor GDF8 protein, as well as peptides of the mature GDF8 protein, can be incorporated into any protein- or peptide-compatible vaccine composition.

5 Such vaccine compositions are well known to the art and include, for example, physiologically compatible buffers and saline and the like, as well as adjuvants, as described in greater detail hereinbelow. The vaccine composition including the precursor GDF8 protein is employed for eliciting antiserum for screening and identifying a specific neutralization epitope for an anti-GDF8 antibody.

10 As exemplified herein, purified precursor GDF8 protein expressed by a vector comprising SEQ ID NO:3 was injected into a goat in a vaccine composition that included 1 g of precursor GDF8 protein emulsified into Freund's complete adjuvant (CFA). The vaccine composition was preferably injected subcutaneously (SC) beneath the skin of the goat. Subsequent booster immunizations are preferred. These
15 can be administered at suitable additional intervals with the same or a reduced dosage of the protein, *e.g.*, at intervals ranging from 2-5 weeks after the initial injection.

Beginning from about two weeks after the initial injection, but preferably starting after a longer time period, *e.g.*, from three to fifteen weeks, or longer, serum is
20 collected, as needed, from the immunized animal. The collected serum is then preferably purified and/or fractionated by conventional immunoglobulin purification procedures such as, for example, protein A-Sepharose, protein G-agarose, hydroxylapatite chromatography, gel electrophoresis, dialysis, or affinity chromatography with a suitable ligand. As exemplified herein, the IgG fraction of the
25 serum was further fractionated on a protein G-agarose column.

The anti-GDF8 antiserum IgG fraction is then available for screening against a range of peptides of the mature GDF8, as described in greater detail by Example 3, hereinbelow.

30 GDF8 Binding Epitopes

Suitable anti-GDF8 monoclonal or polyclonal antibodies were contacted with GDF8 protein for a time period sufficient for the antibody to bind selectively to the protein. Thereafter, GDF8 bioassays confirmed that the antibody neutralized substantially all of the GDF8 protein activity. Any GDF8 bioassay can be employed

for this purpose, although, as exemplified hereinbelow by Example 3, an *in vitro* transcriptional activation assay according to Thies *et. al.*, 2001, (*Growth Factors* 18, 251) is preferred.

Generally, a GDF8 peptide useful as an antigen or binding epitope according to the invention includes from about residue 312 to about residue 361 of GDF8 (SEQ ID NO: 1). In particular, a peptide according to the invention includes from about residue 320 to about residue 350 of GDF8 (SEQ ID NO: 1). The peptide preferably includes from about residue 327 to about residue 346 of GDF8 (SEQ ID NO: 1).

The artisan will appreciate that the inventive GDF8 peptide can be readily modified to include at least one conservative amino acid substitution, and at any position. Preferably, that peptide specifically binds to rat monoclonal antibody 788, as exemplified hereinbelow. Such conservative substitutions can include, for example, variations at residues 328, 329 and 335, and combinations thereof, wherein, amino acid residue 328 is His, Leu, Asn or Val; amino acid residue 329 is Lys or Leu; and amino acid residue 335 is Ser or Pro or Thr. Precursor GDF8 residues 328, 329 and 335 vary within the GDF8 protein sequence across species, as illustrated by Fig. 2, but nevertheless, the mature GDF8 remains functional.

Fig. 1 illustrates a map of the GDF8 active region (that forms the mature protein) in the context of its precursor protein. Superimposed on the map of the GDF8 active region are the locations of seven overlapping peptides. These overlapping peptide were designed in order to provide targets for identifying the antibody-binding epitope or epitopes of GDF8. The peptide labeled as DJ5 was identified by screening with the IgG fraction of the exemplified goat anti-GDF8 antiserum as the only significant binding epitope of GDF8 for the exemplified antiserum. This peptide has a sequence (SEQ ID NO: 8) corresponding to residue 327 to residue 346 of precursor GDF8 (SEQ ID NO: 1).

GDF8 Peptide Vaccine Compositions

The GDF8 peptides described above are preferably formulated into vaccine compositions. These vaccine compositions may be employed to immunize an animal in order to elicit a highly specific anti-GDF8 immune response. The result of the immunization will be downregulation of GDF8 function in the immunized animal.

Such vaccine compositions are well known to the art and include, for example, physiologically compatible buffers, preservatives, and saline and the like, as well as adjuvants.

"Adjuvants" are agents that nonspecifically increase an immune response to a particular antigen, thus reducing the quantity of antigen necessary in any given vaccine, and/or the frequency of injection necessary in order to generate an adequate immune response to the antigen of interest. Suitable adjuvants for the vaccination of animals include, but are not limited to, Adjuvant 65 (containing peanut oil, mannide monooleate and aluminum monostearate); Freund's complete or incomplete adjuvant; mineral gels, such as aluminum hydroxide, aluminum phosphate and alum; surfactants, such as hexadecylamine, octadecylamine, lysolecithin, dimethyldioctadecylammonium bromide, N,N-dioctadecyl-N',N'-bis(2-hydroxymethyl)propanediamine, methoxyhexadecylglycerol and pluronic polyols; polyanions, such as pyran, dextran sulfate, poly IC, polyacrylic acid and carbopol; peptides, such as muramyl dipeptide, dimethylglycine and tuftsin; and oil emulsions. The protein or peptides could also be administered following incorporation into liposomes or other microcarriers. Information concerning adjuvants and various aspects of immunoassays are disclosed, e.g., in the series by P. Tijssen, *Practice and Theory of Enzyme Immunoassays*, 3rd Edition, 1987, Elsevier, New York, incorporated by reference herein.

The vaccine composition includes a sufficient amount of the desired immunogen, such as the inventive GDF8 peptides, to elicit an immune response. The amount administered can range from about 0.0001g/kg to about 1.0 g /kg, relative to the mass of the animal. Any suitable vertebrate animal is readily employed to obtain polyclonal antiserum. Preferably, the animal is a mammal, and includes, but is not limited to, rodents, such as a mice, rats, rabbits, horses, canines, felines, bovines, ovines , e.g., goats and sheep, primates, e.g., monkeys, great apes and humans, and the like.

The vaccine composition is readily administered by any standard route, including intravenously, intramuscularly, subcutaneously, intraperitoneally, *in ovo* (particularly for poultry), and/or orally, For fish species, methods of administering a vaccine composition or immunogenic composition include the foregoing, as well as dipping the fish into water comprising an antigenic concentration of the peptide, spraying the fish with an antigenic concentration of the peptide while the fish is briefly

separated from the water, etc. The artisan will appreciate that the vaccine composition is preferably formulated appropriately for each type of recipient animal and route of administration.

Appropriate animal subjects can include those in the wild, livestock (e.g., raised
5 for meat, milk, butter, eggs, fur, leather, feathers and/or wool), beasts of burden, research animals, companion animals, as well as those raised for/in zoos, wild habitats and/or circuses. In a particular embodiment, the animal is a great ape such as a gorilla, or a human.

In one preferred embodiment, the animal is a "food-producing" animal, and the
10 result of immunization is a gain in animal weight, particularly muscle mass, relative to animals not immunized. For purposes of the present invention, the term "food-producing" animal shall be understood to include all animals bred for consumption by humans and/or other animals, or for producing consumables such as eggs or milk. A non-limiting list of such animals include avians (e.g., chickens, turkeys, ducks, geese,
15 ostriches), bovine (e.g., beef/veal cattle, dairy cows, breeding bulls, buffalo), porcine (e.g., hogs or pigs), ovine (e.g., goats or sheep), equine (e.g., horses) as well as aquatic animals, including shellfish and fish such as trout or salmon, and other species raised or harvested for human consumption.

For purposes of the present invention, the term "fish" shall be understood to
20 include without limitation, the *Teleostei* grouping of fish, i.e., teleosts. Both the *Salmoniformes* order (which includes the *Salmonidae* family) and the *Perciformes* order (which includes the *Centrarchidae* family) are contained within the *Teleostei* grouping.

Examples of potential fish recipients include the *Salmonidae* family, the
25 *Serranidae* family, the *Sparidae* family, the *Cichlidae* family, the *Centrarchidae* family, the three-Line Grunt (*Parapristipoma trilineatum*), and the Blue-Eyed Plecostomus (*Plecostomus spp.*).

Salmonidae Family

TAXON NAME	COMMON NAME
<i>Coregonus clupeaformis</i>	Lake whitefish
<i>Coregonus hoyi</i>	Bloater
<i>Oncorhynchus keta</i>	Chum salmon
<i>Oncorhynchus gorbuscha</i>	Pink salmon
<i>Oncorhynchus kisutch</i>	Coho salmon (silver salmon)
<i>Oncorhynchus masou</i>	cherry salmon (masou salmon)
<i>Oncorhynchus nerka</i>	Sockeye salmon
<i>Oncorhynchus tshawytscha</i>	(chinook salmon)
<i>Prosopium cylindraceum</i>	Round whitefish
<i>Oncorhynchus clarki</i>	Cutthroat trout
<i>Oncorhynchus mykiss</i>	Rainbow trout
<i>Salmo salar</i>	Atlantic salmon
<i>Salmo trutta</i>	Brown trout
<i>Salmo trutta</i> X <i>S. fontinalis</i>	Tiger hybrid-trout
<i>Salvelinus alpinus</i>	Arctic charr
<i>Salvelinus confluentus</i>	Bull trout
<i>Salvelinus fontinalis</i>	Brook trout
<i>Salvelinus leucomaenis</i>	Japanese charr (white spotted charr)
<i>Salvelinus malma</i>	Dolly varden (Miyabe charr)
<i>Salvelinus namaycush</i>	Lake trout
<i>Thymallus thymallus</i>	Grayling

Some Members of the Serranidae Family

TAXON NAME	COMMON NAME
<i>Centropomus ocyurus</i>	Bank sea bass
<i>Centropomus philadelphicus</i>	Rock sea bass
<i>Centropomus striata</i>	Black sea bass
<i>Diplectrum bivittatum</i>	Dwarf sandperch
<i>Diplectrum formosum</i>	Sand perch
<i>Epinephelus flavolimbatus</i>	Yellowedge grouper
<i>Epinephelus morio</i>	Red grouper
<i>Serranus phoebe</i>	Tattler
<i>Serranus tortugarum</i>	Chalk bass

5

Some Members of the Sparidae family

TAXON NAME	COMMON NAME
<i>Archosargus probatocephalus</i>	Sheepshead
<i>Archosargus rhomboidalis</i>	Sea bream
<i>Calamus penna</i>	Sheepshead porgy
<i>Lagodon rhomboides</i>	Pinfish
<i>Pagrus Major</i>	Red Sea bream
<i>Sparus aurata</i>	Gilthead Sea bream
<i>Stenotomus chrysops</i>	Scup

Some Members of the *Cichlidae* family

TAXON NAME	COMMON NAME
<i>Aequidens latifrons</i>	Blue acara
<i>Cichlisoma nigrofasciatum</i>	Congo cichlid
<i>Crenichichla</i> sp.	Pike cichlid
<i>Pterophyllum scalare</i>	Angel fish
<i>Tilapia mossambica</i>	Mozambique mouth breeder
<i>Oreochromis</i> sp.	Tilapia
<i>Sarotherodon aurea</i>	Golden Tilapia

Some Members of the *Centrarchidae* family

TAXON NAME	<u>COMMON NAME</u>
<i>Ambloplites rupestris</i>	Rock bass
<i>Centrarchus macropterus</i>	Flier
<i>Elassoma evergladei</i>	Everglades pigmy sunfish
<i>Elassoma okefenokee</i>	Okefenokee pigmy sunfish
<i>Elassoma zonatum</i>	Banded pigmy sunfish
<i>Enneacanthus gloriosus</i>	Bluespotted sunfish
<i>Enneacanthus obesus</i>	Banded sunfish
<i>Lepomis auritus</i>	Redbreast sunfish
<i>Lepomis cyanellus</i>	Green sunfish
<i>Lepomis cyanellus</i> X <i>L. gibbosus</i>	Green x pumpkinseed
<i>Lepomis gibbosus</i>	Pumpkinseed
<i>Lepomis gulosus</i>	Warmouth
<i>Lepomis humilis</i>	Orange-spotted sunfish
<i>Lepomis macrochirus</i>	Bluegill
<i>Lepomis megalotis</i>	Longear sunfish
<i>Micropterus coosae</i>	Shoal bass
<i>Micropterus dolomieu</i>	Smallmouth bass
<i>Micropterus punctulatus</i>	Spotted bass
<i>Micropterus salmoides</i>	Largemouth bass
<i>Pomoxis annularis</i>	White crappie
<i>Pomoxis nigromaculatus</i>	Black crappie

5

In a further preferred embodiment, the animal is a companion animal or a human, and the vaccine is administered to provide long-term downregulation of GDF8 for any veterinary or medical purpose responsive to such GDF8 downregulation. For purposes of the present invention, the term "companion" animal shall be understood to include all animals – horses (equine), cats (feline), dogs (canine), rodents, (including mice, rats, guinea pigs), rabbit species, and avians, such as pigeons, parrots and the like.

10

Other birds receiving such vaccination or antibodies can be associated with either commercial or noncommercial aviculture. These include e.g., *Anatidae*, such as

swans, *Columbidae*, e.g., doves and pigeons, such as domestic pigeons, *Phasianidae*, e.g., partridge, and grouse, *Thesienidae*, *Psittacines*, e.g., parakeets, macaws, and parrots, e.g., raised for the pet or collector market, and members of the Ratite family.

5 In another preferred embodiment, any of the above recited animals (preferably nonhuman) are immunized in order to obtain anti-GDF8 antibodies that specifically bind to the inventive peptides, and the elicited antibodies are harvested for use in assays, and/or in veterinary or human medicine, e.g., to provide downregulation of GDF8 for any veterinary or medical purpose responsive to such GDF8
10 downregulation.

The present invention may be better understood by reference to the following non-limiting examples, which are provided as exemplary of the invention. The following examples are presented in order to more fully illustrate embodiments of the invention and should in no way be construed as limiting the broad scope of the
15 invention.

EXAMPLES

EXAMPLE 1

Materials & Methods

A. Expression and Purification of Precursor GDF8 (GDF8 Prohormone)

The natural DNA sequence of the precursor GDF8 or prohormone (SEQ ID NO: 2) was optimized for expression in mammalian and viral expression systems. To avoid the negative effects of viral host shutoff mechanisms the DNA sequence was designed to be as divergent from the mammalian nucleic acid sequence as possible.
25 To accomplish this the amino acid sequence of the GDF8 prohormone was reversed translated using yeast preferred codons. The resulting sequence was surveyed for codons, which retained their homology to the human GDF8 nucleic acid sequence. Where possible these codons were substituted with the next most preferred yeast codons encoding the same amino acid. The resulting nucleic acid molecule (SEQ ID
30 NO: 3) was commercially synthesized for incorporation into the appropriate expression vectors.

The Flp-In™ CHO expression system (Invitrogen, Carlsbad, CA) was used to express the optimized GDF8 prohormone. Briefly, a GDF8 prohormone construct containing a C-terminal FLAG® (Sigma-Aldrich Corp., St. Louis, MO) epitope fusion

was constructed by inserting the gene encoding the modified GDF8 prohormone into plasmid pCMVtag4B (Stratagene, San Diego, CA.). The FLAG[®] fusion tag facilitates separation of FLAG[®] fusion proteins on an anti-FLAG[®] gel column. A PCR DNA fragment containing the modified GDF8 prohormone-FLAG[®] gene was then cloned
5 into plasmid expression vector pcDNA5/FRT (Invitrogen, Carlsbad, CA). Generation of the Flp-In[™] CHO cell line expressing the GDF8 prohormone-FLAG[®] fusion protein was achieved by cotransfection of the Flp-In[™] CHO cell line with the Flp-In[™] expression vector containing the GDF8-FLAG[®] gene and the Flp recombinase expression plasmid, POG44. Flp recombinase mediates insertion of the Flp-In
10 expression cassette into the genome at an integrated FRT site by site-specific DNA recombination. A stable cell line expressing and secreting the GDF8 prohormone containing the FLAG[®] epitope was obtained using hygromycin B selection.

The stable CHO cell line expressing the GDF8 prohormone containing the FLAG[®] tag was adapted to suspension culture in serum-free media using standard
15 techniques. Conditioned media containing the secreted GDF8 prohormone was generated using the WAVE bioreactor system (WAVE Biotech LLC, Bridgewater, NJ). Purification of the FLAG[®] tagged GDF8 prohormone was achieved by affinity chromatography using an anti-FLAG[®] M2 affinity gel (Sigma-Aldrich Corp., St. Louis, MO).

B. DJ5 Specific Antibody Purification

DJ5 (SEQ ID NO: 8; See Table 2, below) specific antibody fractions were purified by affinity column chromatography. An affinity column was prepared by coupling 10 mg of DJ5 synthetic peptide to 0.8 g of cyanogen bromide-activated
25 Sepharose 4B (Sigma Genosys, Woodlands, TX). The column was washed and equilibrated with PBS. Approximately 11ml of Goat IgG fraction (10 mg/ml) was applied to the affinity column and washed with 25 ml of PBS. Fractions of 1.0 ml were collected and monitored for absorbance at 280 nm. Bound material was eluted with approximately 10 ml of 0.2 M glycine (pH 1.85). Fractions of 1.0 ml were collected and
30 neutralized with 0.25 ml of 0.5 M sodium phosphate, 0.75 M NaCl, pH 7.4. Approximately 25 µl aliquots of unbound fractions 1-10 and bound fractions 25-35 were assayed for ELISA reactivity to DJ5 peptide. Unbound fractions were found to be negative for DJ5 reactivity. Bound fractions exhibited a strong peak of reactivity to the

DJ5 peptide. Unbound fractions 1-11 and bound fractions 26-34 were pooled. Pooled samples were concentrated and their buffer exchanged with phosphate buffered saline (PBS) as indicated below. Sample concentrations were determined by the OD 280 method (CURRENT PROTOCOLS IN IMMUNOLOGY, 2.7.3, John Wiley & Sons, Inc.).

- 5 The unbound sample was adjusted to 10 mg/ml and the bound sample was adjusted to 1 mg/ml, for subsequent use.

EXAMPLE 2

Goat Anti- GDF8 Polyclonal IgG Serum

- 10 Goat anti-precursor GDF8 IgG was obtained from an immunized goat by the following methods.

A. Immunization of Goat

- 15 A Saanen (dairy) goat (approximately 2 year old male) was immunized with purified recombinant GDF8 prohormone (obtained as described by Example 1, above), as follows. One half mg of protein was emulsified in Freund's complete adjuvant (CFA) and injected subcutaneously (SC) beneath the skin of the goat. Subsequent booster immunizations administered SC at weeks three, six, and ten
- 20 contained 0.3 mg of protein emulsified in Freund's incomplete adjuvant (IFA). Blood was collected from the jugular vein with a syringe and needle, and taken with vacuum bottle and tubing. The blood was collected in bottles containing anticoagulant and centrifuged at 2500 RPM for 20 minutes to remove the red blood cells. The plasma was re-calcified to produce serum. The serum sample collected 15 weeks post initial
- 25 immunization was used for further analysis.

B. Collection and Purification of Goat Polyclonal IgG

- 30 Serum was harvested from the goat after 15 weeks, and the IgG fraction was purified from this serum, as follows. The IgG fraction of goat sera was purified on a Protein G agarose column according to the manufactures protocol (Kirkegaard and Perry Laboratories, Inc., Gaithersburg, MD). Eluted fractions were pooled, concentrated, and buffer exchanged with phosphate buffered saline (PBS) utilizing Centriprep centrifugal Filters (Centriprep YM-10, Millipore Corporation, Billerica, MA).
- 35 Sample concentrations were determined by the OD 280 method (CURRENT PROTOCOLS IN IMMUNOLOGY, *Id.*) and adjusted to 10mg/ml.

EXAMPLE 3**Characterization of Goat Antiserum**

The goat antiserum provided by Example 2, above, is designated as **PGA**. It is expected that the **PGA** IgG fraction contains antibodies directed against various epitopes on the GDF8 prohormone molecule. The **PGA** antiserum was characterized by an *in vitro* transcription activation assay, as follows. The *in vitro* transcriptional activation assay used to quantitatively measure GDF8 bio-neutralization is essentially that of Thies *et.al.* (*Growth Factors* 18, 251 (2001)). Ninety-six well tissue culture treated luminometer ViewPlate™ assay plates (PerkinElmer Life and Analytical Sciences, Inc., Boston, MA) were seeded with 1.0×10^5 cells/well of A204 Rhabdomyosarcoma cells (ATCC HTB-82) and incubated in a 37°C, 5% CO₂, humidified chamber. Complete A204 culture media consists of McCoy's 5A medium, 10% fetal bovine serum, 2% L-glutamine, and 1% Penn/Strep. Upon reaching greater than 80% confluence, the cells were transiently transfected with a mixture of plasmid pDPC4-luciferase and HCMV IE-lacZ using the protocol recommended by the manufacturer of the FUGENE transfection reagent (Roche Diagnostics Corporation, Indianapolis, IN) and incubated 16 hours in a 37°C, 5% CO₂, humidified chamber. Plasmid pDPC4-luciferase contains four copies of the CAGA box, derived from the human plasminogen activator inhibitor (PAI-1), which confers GDF8 responsiveness to the heterologous promoter reporter construct.

Plasmid HCMV IE-lacZ contains a *beta*-galactosidase gene under the control of the constitutive human cytomegalovirus immediate early promoter. This gene is added as a control to normalize for transfection efficiencies. Cells were then treated with 100 ng/well GDF8 protein (R&D Systems Inc., Minneapolis, MN) and incubated an additional 16 hours in a 37°C, 5% CO₂, humidified chamber. Luciferase and *beta*-galactosidase were quantified in the treated cells using the Dual-Light Luciferase Assay (Tropix, Applied Biosystems, Foster City, CA).

Each sample was run in duplicate (2 wells). The signal for each well was calculated as the luciferase signal divided by the *beta*-galactosidase signal times 100. The sample signal was calculated as the average of the two wells.

To test the bio-neutralization activity of an antibody sample various concentrations of purified IgG fractions were incubated with the GDF8 protein (approximately 16 hours at 4°C) prior to treatment of the cells. The percent inhibition was calculated as $100 - (100 \times \text{sample signal}) / (\text{signal with GDF8 alone} - \text{signal with$

no GDF8 added). The results of the *in vitro* transcription activation assay are summarized by Table 1, below.

Table 1
GDF8 neutralization titers for Goat Serum PGA

<u>Sample (μg IgG)</u>	<u>% Inhibition of GDF8 Activity</u>
Goat – normal (250)	0
Goat – PGA (250)	95
Goat – PGA (125)	86
Goat – PGA (63)	62
Goat – PGA (31)	22
Goat – PGA (16)	3

- 5 The neutralization assay confirmed that the IgG fraction of the harvested goat serum contains antibodies capable of neutralizing at least 95% of the GDF8 used in this activity assay.

EXAMPLE 4

10 **Goat Polyclonal Antibody Defines**
 A Specific Neutralization Epitope of the GDF8 Protein

- In order to determine the specificity of the neutralizing immune response the **PGA** IgG fraction was assayed for its reactivity with a set of seven overlapping peptides (DJ1-7 see Table 2 and Figure 1) that span the entire coding region of the active GDF8 protein. Reactivity of the Goat **PGA** IgG to each individual peptide was determined by Enzyme-Linked Immunosorbent Assay (ELISA) assay. The GDF8 peptide ELISA was performed essentially as described in Protein Detector™ ELISA Kit HRP, ABTS System (Kirkegaard and Perry Laboratories, Inc., Gaithersburg, MD). The following modifications were used in the assay. Synthetic peptides DJ1-7 (see Table 2, below) were custom synthesized under our direction by ProSci, Inc. (Poway, CA). Plates were coated with synthetic peptides at 500 ng per well and purified GDF8 prohormone at 250 ng per well. Primary antibodies were IgG fractions from various samples. Secondary antibodies were used at a dilution of 1:2000. For goat primary antibody samples the secondary antibody was rabbit peroxidase-labeled antibody to goat IgG. For rat primary antibody samples the secondary antibody was goat peroxidase-labeled antibody to rat IgG. The OD 405 nm was read for 15 minutes with an ELISA plate reader. The ELISA reactivity was calculated as OD 405 per minute times 1000.

Table 2
GDF8 Active Region Peptides

<u>Name</u>	<u>Coordinates*</u>	<u>Amino acid sequence</u>	
DJ1	267-286	DFGLDCDEHSTESRCCRYPL	SEQ ID NO: 4
DJ2	282-301	CRYPLTVDFEAFGWDWIIAP	SEQ ID NO: 5
DJ3	297-316	WIIAPKRYKANYCSGECEV	SEQ ID NO: 6
DJ4	312-331	ECEFVFLQKYPHTHLVHQAN	SEQ ID NO: 7
DJ5	327-346	VHQANPRGSAGPCCTPTKMS	SEQ ID NO: 8
DJ6	342-361	PTKMSPINMLYFNGKEQIIY	SEQ ID NO: 9
DJ7	357-375	EQIIYGKIPAMVVDRCGCS	SEQ ID NO: 10

* relative to Human GDF8 prohormone (Genebank Accession Number NP_005250)

The ELISA results are summarized by Table 3, below.

Table 3
ELISA reactivity of PGA IgG (10 mg/ml)
to GDF8 Active Region Peptides

	<u>OD 405 / minute X 1000</u>		
<u>Antigen</u>	<u>1:20</u>	<u>1:40</u>	<u>1:80</u>
DJ1*	23	10	1
DJ2*	3	0	0
DJ3*	0	10	0
DJ4*	3	0	0
DJ5*	121	37	27
DJ6*	3	0	0
DJ7*	10	1	0
proGDF8**	194	196	199

* peptide, ** prohormone

5 The **PGA** IgG fraction reacted specifically with both the purified GDF8
prohormone and with the DJ5 peptide. Among the GDF8 active region peptides the
IgG fraction reacts specifically and exclusively with the DJ5 peptide. This is a strong
indication that the neutralizing capability of this serum is directed against an epitope
defined by the DJ5 peptide. In order to confirm this hypothesis the DJ5 specific
10 fraction of **PGA** IgG was purified. This was accomplished by affinity chromatography
as described in the materials and methods. The **PGA** antibodies were separated into
DJ5 peptide bound and unbound fractions. Both fractions were assayed for
neutralization activity against the GDF8 protein.

15 The results in Table 4 show that the majority of GDF8 neutralization capacity
resides with the antibody that binds specifically to the DJ5 peptide. This clearly
demonstrates that the DJ5 peptide defines a neutralizing epitope of the GDF8 protein.

Table 4
GDF8 neutralization activity of DJ5 specific antibodies

<u>Sample (µg IgG)</u>	<u>% Inhibition of GDF8 Activity</u>
Goat – normal (250) **	7
DJ5 unbound IgG (250)	26
DJ5 bound IgG (25)	90

** The normal goat IgG was a negative control purified from non-immunized goat sera (commercially purchased).

Curiously, in a preliminary experiment, neutralizing GDF8 antibodies were not obtained when two rabbits were injected with the human DJ5 antigen conjugated to keyhole limpet hemocyanin. As can be seen in Figure 2, the amino acid sequences corresponding to DJ5 for rabbit and human GDF8 are identical, whereas the amino acid sequence of goat DJ5 is different. Therefore, in view of the data provided above for the goat, the preliminary rabbit data suggests that it may be advantageous to use a DJ5 antigen that comprises a different amino acid sequence than that for the corresponding region/portion of the host animal GDF8. Thus, in this case, the ability of the recombinant human GDF8 prohormone to induce bio-neutralizing antibodies in a goat may be due, at least in part, to the fact that the antigen used comprised an amino acid sequence that differs from that of the host sequence by a single amino acid substitution in the DJ5 region/portion of GDF8 [see, amino acid residue 333 in Figure 2]. More particularly, as Figure 2 shows, the Arg₃₃₃ in the human sequence is replaced by a Lys₃₃₃ residue in the goat sequence. This lone conservative amino acid substitution may constitute an alteration that is significant enough to render the protein "foreign" to the goat immunological surveillance system.

EXAMPLE 5

GDF8 Neutralizing Rat mAB 788 Defines A Specific Neutralization Epitope of the GDF8 Protein

Rat monoclonal antibody 788 is reported to neutralize mouse GDF8 bioactivity (R&D Systems Inc., Cat. No. MAB788, Minneapolis, MN). In order to confirm this result we assayed the monoclonal antibody for neutralization activity against the GDF8 protein. The antibody was characterized as described by Example 2, above. The results of this assay are summarized by Table 5, as follows.

Table 5
GDF8 neutralization titers for
monoclonal antibody 788

<u>Sample (µg IgG)</u>	<u>% Inhibition of GDF8 Activity</u>
MAB – 788 (12.5)	47
MAB – 788 (6.3)	17
MAB – 788 (3.1)	7
MAB – 788 (1.8)	0
MAB – 788 (0.1)	0

Table 5 confirms that this antibody is capable of neutralizing the activity of the GDF8 protein. In order to determine the specificity of this neutralizing immune response the rat monoclonal antibody was assayed for its reactivity with a set of seven overlapping peptides (DJ1-7 see table 2 and figure 1) that span the entire coding region of the active GDF8 protein. Reactivity of the monoclonal antibody to each individual peptide was determined by ELISA assay (see materials and methods).

Table 6
ELISA reactivity of Rat MAB 788 (10 mg/ml)
to GDF8 active region peptides

<u>Antigen</u>	<u>OD 405 / minute X 1000</u>		
	<u>1:20</u>	<u>1:40</u>	<u>1:80</u>
DJ1*	4	0	0
DJ2*	0	0	0
DJ3*	0	0	0
DJ4*	0	0	0
DJ5*	133	118	102
DJ6*	0	0	0
DJ7*	0	0	0
proGDF8**	132	127	132
* peptide, ** protein			

The rat monoclonal antibody reacted specifically with both the purified GDF8 prohormone and with the DJ5 peptide. Typically a monoclonal antibody has mono specificity to a single epitope. Among the GDF8 active region peptides this monoclonal antibody reacts specifically and exclusively with the DJ5 peptide. This result provides further independent evidence that the DJ5 peptide defines a neutralizing epitope of the GDF8 protein.

Many modifications and variations of this invention can be made without departing from its spirit and scope, as will be apparent to those skilled in the art. The

specific embodiments described herein are offered by way of example only, and the invention is to be limited only by the terms of the appended claims, together with the full scope of equivalents to which such claims are entitled. Numerous references are cited in the specification, including Genbank accession numbers of published and/or
5 internet-published nucleic acid and polypeptide/protein sequences, the disclosures of which are incorporated by reference in their entireties.

What is claimed is:

1. An isolated peptide consisting of 50 or fewer amino acid residues that comprises amino acid residues 327 to 346 of SEQ ID NO:1.

5

2. The isolated peptide of Claim 1 that comprises amino acid residues 321 to 346 of SEQ ID NO:1.

3. The isolated peptide of claim 2 that comprises amino acid residues 320 to
10 350 of SEQ ID NO:1.

4. The isolated peptide of claim 3 that comprises amino acid residues 312 to residue 361 of SEQ ID NO:1.

15 5. An isolated peptide consisting of 50 or fewer amino acid residues that comprises amino acid residues 327 to 346 of SEQ ID NO:1 comprising an amino acid substitution;

wherein there are no more than five amino acid substitutions between amino acid residues 327 to residue 346; and

20 wherein the peptide specifically binds to rat monoclonal antibody 788.

6. The isolated peptide of claim 5 comprising amino acid substitutions at a position selected from the group consisting of residues 328, 329, 331, 333 and 335, and combinations thereof, wherein,

25 (a) amino acid residue 328 is His, Leu, or Asn;

(b) amino acid residue 329 is Gln or Lys;

(c) amino acid residue 331 is Asn or Ser;

(d) amino acid residue 333 is Arg or Lys; and/or

(e) amino acid residue 335 is Ser, Pro, or Thr.

30

7. The isolated peptide of claim 6 comprising no more than one amino acid substitution between residues 327 to residue 346 of the precursor GDF8, provided that the peptide specifically binds to rat monoclonal antibody 788.

8. The isolated peptide of claim 1 that comprises a specific neutralization epitope for an anti-GDF8 antibody.

9. The isolated peptide of claim 8, wherein the antibody is selected from the group consisting of rat anti-GDF8 monoclonal antibody 788 and an IgG fraction of goat anti-GDF8 polyclonal antiserum.

10. A fusion protein comprising the peptide of claim 1 or an antigenic subfragment of the peptide.

11. A nucleic acid molecule that encodes the fusion protein of claim 10.

12. A nucleic acid molecule that encodes the peptide of claim 6.

13. A nucleic acid molecule that encodes the peptide of claim 1.

14. The nucleic acid molecule of claim 13 that comprises a nucleic acid sequence of nucleotide 1112 to nucleotide 1171 of SEQ ID NO: 2.

15. A replicable cloning vector comprising the nucleic acid molecule of claim 13.

16. A host cell comprising the replicable cloning vector of claim 15.

17. The host cell of claim 16 that is transformed by the cloning vector.

18. The host cell of claim 16 that is a eukaryotic cell.

19. A method of producing a GDF8 peptide comprising the steps of culturing the host cell of claim 18, expressing the encoded peptide, and recovering the peptide.

20. A vaccine composition comprising the peptide of claim 1.

21. A vaccine composition comprising the fusion protein of claim 10.

22. The vaccine composition of claim 20 further comprising an adjuvant.

23. A method of eliciting an anti-GDF8 immune response in an animal,
5 comprising administering to the animal an effective amount of the vaccine composition
of claim 20.

24. A screening method for selecting an anti-GDF8 antibody or antibody
fragment from among a plurality of antibodies or antibody fragments, comprising
10 contacting the peptide of claim 1 with a sample comprising one or a plurality of
antibodies or antibody fragments, and detecting antibody or antibody fragment that
selectively binds to the peptide.

25. A method of down-regulating GDF8 activity in an animal comprising
15 administering an antibody or antibody fragment to the animal, in an amount and for a
duration effective to down-regulate GDF8 activity in the animal, wherein the antibody
binds specifically to the peptide of claim 1.

26. A method of down-regulating GDF8 activity in an animal comprising
20 immunizing the animal with an effective amount of the vaccine composition of claim
20.

27. A method of down-regulating GDF8 activity in an animal comprising
immunizing the animal with an effective amount of the vaccine composition of claim
25 21.