
Competitive Biosensor Having Elevated Sensitivity

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The present invention relates to measures for determining glucose and for diagnosing diseases that are based on impaired glucose metabolism. In particular, the present invention relates to a device comprising a hydrogel having a glucose-binding protein incorporated therein and a ligand of the glucose-binding protein, wherein the hydrogel comprises a first hydrogel matrix made of alginate and a second hydrogel matrix, which forms an interpenetrating network within the first hydrogel matrix. The invention further relates to the use of such a device for determining the glucose content in a sample and to the use of the device for diagnosing impaired glucose metabolism in a test subject.

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The determination of the concentration of glucose by efficient and reliable measurement technology is of great importance in many fields of technology. Not only in pure laboratory analysis, but also in the foodstuffs industry, e.g. in the field of oenology and in the medical field, for example in the diagnosis of diseases which are caused by impaired glucose metabolism, e.g. diabetes mellitus or metabolic syndrome, the rapid and reliable determination of the glucose concentration in given solutions is of central importance. In the field of the diagnosis of diseases that are based on impaired glucose metabolism, glucose sensors are used both in devices which are implantable into the body and can measure the glucose content in a sample taken within the body, and also in sensors that measure the glucose content ex vivo from a test subject's sample.

For determining the glucose content in a solution, systems made of sensor molecule and ligand have been described, wherein the sensor is a glucose-binding protein and the ligand a competitor for glucose, which is initially present in the sensor bound to the glucose sensor molecule. By competition with glucose during the measurement procedure, the competitor is displaced from the glucose-binding protein. The displacement of the competitor from the glucose-binding protein by the glucose during this can be detected by means of a change in a physical or chemical property of the molecules, e.g. by means of fluorescence resonance energy transfer (FRET). The aforesaid systems must of course be present in a spatially demarcated region of the sensor.

For enclosure of the system components, i.e. of the glucose-binding molecule and the ligand that serves as the competitor, hydrogels among others have proved their worth. Suitable hydrogels for this can be polyethylene glycols, but also alginates (e.g. US2007/0105176; US 6,485,703; Russell 1999, Anal Chem 71: 3126-3132). Furthermore, sensors have been described wherein the aforesaid systems in an aqueous medium are enclosed by a semipermeable membrane. Such membranes can for example consist of regenerated cellulose, polyethylene glycol, polyurethane, layer-by-layer (LBL) layers, polyether sulfones, parylene layers or perforated silica (e.g. US2007/0122829).

However, the glucose sensors described in the state of the art exhibit relatively low glucose activity. The glucose activity and hence the sensor performance is predominantly determined by the binding constants for the complex of glucose-binding protein and competitor and glucose-binding protein and glucose.

In the case of ex vivo sensors, in order to obtain as high a sensor sensitivity as possible, an analyte receptor with a binding constant of almost any magnitude can be selected, since the analyte does not
5 have to be liberated again and a very high specificity and hence good sensor performance can mostly be obtained through a very high binding constant.

However, with in vivo sensors, the situation looks
10 different as the sensor must constantly react reversibly to changes in the analyte concentration. Thus the binding constant must not be too high, since otherwise the sensor would already be saturated at low analyte concentrations, and would no longer be able to
15 indicate concentration changes. Moreover, a further problem of the in vivo sensors consists in that the analyte concentration range is fixed and cannot be optimized by dilution or concentration. Hence essentially, in the state of the art, systems
20 consisting of ligand and glucose-binding protein are described wherein medium binding constants are implemented. Adaptation of the measurement sensitivities by alteration of the concentrations of either glucose-binding protein or ligand or both are as
25 a rule limited by the low solubility of the respective molecules. Hence in the sensors described in the aforesaid state of the art, both the measurement sensitivity and also the measurement precision of the sensor are limited (see Rounds 2007, J. Fluorec. 17:
30 57-63).

The purpose of the present invention is to provide a device which enables more efficient determination of the glucose level even in vivo and with which the
35 aforesaid disadvantages are essentially eliminated. The invention is solved by the embodiments described by the claims and the embodiments which are disclosed below.

The invention thus relates to a device comprising a hydrogel having a glucose-binding protein incorporated therein and a ligand of the glucose-binding protein, wherein the hydrogel comprises a first hydrogel matrix
5 made of alginate and a second hydrogel matrix which forms an interpenetrating network within the first hydrogel matrix.

The device according to the invention also concerns a
10 composition consisting of the aforesaid components.

The term "hydrogel" describes a water-containing polymer whose molecules are chemically or physically linked into a three-dimensional network. The polymer
15 molecules can be linked together into the three-dimensional network by covalent or ionic bonds or by entanglement or weaving. The polymers which form the hydrogel preferably contain hydrophilic polymer components which enable the uptake of aqueous
20 solutions, and groups which are capable of interacting with the glucose-binding protein.

The hydrogels according to the invention consist of a first hydrogel matrix made of alginate and a second
25 hydrogel matrix, which is capable of forming an interpenetrating network within the hydrogel matrix set up by the alginate. This second hydrogel matrix preferably consists of a water-soluble polymer having at least one crosslinkable group per molecule and a
30 molecular weight of at most 500,000. Particularly preferable is a molecular weight of at most 250,000, 200,000, 150,000, 100,000 or 50,000. In this, the second hydrogel matrix is preferably selected from the group consisting of: polyvinyl alcohols (PVAs),
35 polyethylene glycols (PEGs), poly(2-oxazolines), polyacrylamides (e.g. dimethylacrylamide), polyhydroxyacrylates (e.g. polyhydroxymethacrylate, polyhydroxyacrylamide, polyvinylpyrrolinones), (2-

methyl-3-ethyl[2-hydroxyethyl]) polymers,
polyhydroxyalkanoates (PHAs), poly (2-methyl-2
oxazolines), poly (2-ethyl-2 oxazolines), poly (2-
hydroxyethyl-2 oxazolines), poly (2-(1-(hydroxymethyl)-
5 ethyl)-2 oxazolines), poly-(hydroxyethyl methacrylate)
(PHEMA), poly-(hydroxyethyl acrylate) (PHEA), poly-
vinylpyrrolidones, poly-(dimethyl)acrylamide, poly-
(hydroxyethyl)acrylamide, polyvinyl alcohols (including
copolymers with vinyl acetates and/or ethylene),
10 poly(ethylene-co-vinyl alcohol), poly(vinyl acetate-co-
vinyl alcohol), poly(ethylene-co-vinyl acetate-co-vinyl
alcohol), polyethylene glycols and poly(ethylene
glycol-co-propylene glycol).

15 An interpenetrating network according to the invention
is preferably obtained by polymerization of the
monomers of the second polymer in the presence of an
already existing first polymer. Particularly
preferably, this can occur by processes which are
20 described in more detail in the practical examples. It
is thus achieved that an interpenetrating network of
the second polymer can form in the already existing
network of the first polymer. The polymer networks are
thus interwoven and/or entangled with one another. In
25 contrast to this, various polymers are present side by
side in mixtures as separate networks not interwoven
and/or entangled with one another.

Particularly preferably, the crosslinking mechanisms of
30 the first and the second polymer differ, so that no
mixed crosslinking arises. The aforesaid alginate which
is to create the first hydrogel matrix is crosslinked
by ionic interactions. The aforesaid second polymers
which are to create the interpenetrating network are
35 all crosslinked by radical or ionic polymerization.

Particularly preferably, the second hydrogel matrix is
formed of polyvinyl alcohol. Particularly preferred is

polyvinyl alcohol having a molecular weight of 10,000 to 100,000, more preferably 10,000 to 50,000, more preferably 10,000 to 20,000, and quite particularly preferably 15,000. Particularly preferably, the
5 polyvinyl alcohol has a crosslinker content of at most 0.5 mmol/g, 0.4 mmol/g, 0.35 mmol/g, or 0.3 mmol/g and quite particularly preferably 0.35 mmol/g. Furthermore, the polyvinyl alcohol particularly preferably has a prepolymer solids content of less than 40 weight
10 percent.

As well as the first hydrogel matrix and the second hydrogel matrix, the hydrogel according to the invention can contain additives, e.g. stabilizers,
15 emulsifiers, antioxidants, UV stabilizers, detergents and/or UV initiators.

The hydrogel used according to the invention can moreover be enclosed by a further, preferably
20 semipermeable, covering material. By means of this enclosure, "leaching" of the sensor components from the hydrogel is prevented. Possible covering materials are semipermeable membranes or other hydrogel matrices. Semipermeable membranes can preferably consist of
25 regenerated cellulose, polyethylene glycol, polyurethane layer-by-layer (LBL) layers, polyether sulfones, parylene layers or perforated silica. Preferably, a further hydrogel matrix can be formed from a polymer selected from the group consisting of
30 alginates, sepharoses, hyaluronic acid, chitosan, polyvinyl alcohols (PVAs), polyethylene glycols (PEGs), carrageenans and polyhydroxalkanoates (PHAs), poly (2-methyl-2 oxazolines), poly (2-ethyl-2 oxazolines), poly (2-hydroxyethyl-2 oxazolines), poly (2-(1-
35 (hydroxymethyl)-ethyl)-2 oxazolines), poly-(hydroxyethyl methacrylate) (PHEMA), poly-(hydroxyethyl acrylate) (PHEA), poly-vinylpyrrolidones, poly-(dimethyl)acrylamide, poly-(hydroxyethyl)acrylamide,

polyvinyl alcohols (including copolymers with vinyl acetates and/or ethylene), poly(ethylene-co-vinyl alcohol), poly(vinyl acetate-co-vinyl alcohol), poly(ethylene-co-vinyl acetate-co-vinyl alcohol),
5 polyethylene glycols and poly(ethylene glycol-co-propylene glycol).

The term "glucose-binding protein" in the context of the invention relates to proteins which are capable of
10 interacting specifically with glucose. Whether a protein is capable of interacting specifically with glucose can readily be determined by those skilled in the art by binding tests known in the state of the art. Particularly preferably, the glucose-binding protein is
15 selected from the group consisting of lectins, enzymes which bind glucose as substrate, and antibodies which specifically recognize glucose. The term "glucose-binding protein" also includes surrogate molecules which can specifically recognize glucose, preferably
20 aptamers which specifically recognize glucose. Quite particularly preferably, the glucose-binding protein is concanavalin A.

Nucleic acid sequences and amino acid sequences which
25 encode the aforesaid glucose-binding proteins are known in the state of the art (Yamauchi 1990, FEBS Letters 260(1): 127-130). Accordingly, the aforesaid proteins can readily be prepared by those skilled in the art. Said proteins can for example be prepared recombinantly
30 or be purified from a biological source. Furthermore, the proteins can also be chemically synthesized. Moreover, most of the aforesaid proteins are commercially available. Antibodies or aptamers which specifically recognize glucose can readily be prepared
35 by those skilled in the art by methods for antibody or aptamer obtention known in the state of the art.

The aforesaid glucose-binding proteins and in particular concanavalin A can preferably also have chemical modifications which mediate increased water solubility compared to unmodified versions of the glucose-binding proteins. Such modifications preferably
5 comprise functionalization with a water-soluble polymer and in a particularly preferable embodiment can be selected from the group consisting of: pegylation, acetylation, polyoxazolinylation and succinylation.

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The detection of glucose in a sample for analysis is effected in the device according to the invention by displacement of the ligand bound to the glucose-binding protein by the glucose contained in the sample
15 (competition between ligand and glucose). Hence the ligand preferably has a lower affinity to the glucose-binding protein than glucose. The displacement can preferably be detected by labeling of the ligand with a dye or another detectable marker molecule. Dyes or
20 other marker molecules which on approach of the molecules bound to them cause a change in at least one measureable physical or chemical property have proved particularly suitable for the detection of a displacement. Suitable systems include those in which a
25 measurable signal is either suppressed or generated by means of energy transfer between the dye molecules. Such energy transfer-based systems are for example described in more detail in WO2001/13783. These can preferably be systems in which a fluorescence signal is
30 suppressed by quenching effects when the dye or marker molecules - and hence the glucose binding protein and its ligand - are in spatial proximity. After the displacement of the ligand by the analyte, the quenching effect is then canceled. This effect is
35 detectable by a change in the fluorescence. For the detection, for example fluorescence photometers as described in WO2002/087429 can be used. Other suitable systems are so-called fluorescence resonance energy

transfer (FRET)-based detection systems. In these, two interacting components, such as the glucose-binding protein and its ligand, are labeled with fluorescent dyes. One component is coupled with an acceptor dye, the other with a donor dye. Through the interaction of the components, the dyes come into spatial proximity, whereby the FRET effect is produced, in which excitation energy is transferred from the donor to the acceptor dye and thus the intensity of the donor dye becomes measurably less. As soon as the interaction of the components is interrupted, the fluorescent intensity of the donor again increases. In the case of the device according to the invention, the glucose can thus be detected via the increase in the intensity of a signal which is generated by a dye or marker molecule after separation of the complex of glucose-binding protein and ligand by the analyte. The dye or the marker molecule is coupled either to the glucose-binding protein or the ligand. For example, a donor dye can be coupled to the glucose-binding protein or the ligand, while the component not coupled to the donor dye is coupled to a suitable acceptor dye. In a thus configured device according to the invention, the FRET effect as a result of the binding of the ligand to the glucose-binding protein can be observed before exposure to a glucose-containing sample. After exposure, the glucose displaces the ligand, so that the measurable intensity of the fluorescence of the donor dye increases, in fact proportionately to the quantity of glucose.

Birch et. al. (Birch 2001, Spectrochimica Acta Part A 57: 2245-2254) have calculated the mathematical solution of the chemical equilibrium for the case of concanavalin A as glucose-binding protein and dextran as ligand. Simulations with varying concanavalin A and dextran concentrations have shown that the ratio (dextran)/(Con A-dextran complex) depends only slightly

on the starting concentrations and that the binding constants K_{Dex} and K_{Gluc} are the main factors for sensor performance.

5 Surprisingly, however, the structure of the dyes also affects the glucose activity. Thus according to the invention a combination of a rhodamine and an oxazine dye is markedly superior to the conventionally used combination of a xanthene and a rhodamine dye (FITC-
10 TMR).

Preferably, the glucose-binding protein which is used in the context of the invention is linked to an oxazine dye. How such a linkage can be effected is well known
15 to those skilled in the art and adequately described in the state of the art. Particularly preferably, the oxazine dye is an oxazine acceptor selected from the group consisting of: ATTO655, ATTO680, EVOblue10, EVOblue30, EVOblue90 and EVOblue100. Quite particularly
20 preferably, ATTO680 is used. Said oxazine dyes are commercially available.

The preferred degree of labeling (DOL) for the glucose-binding protein, e.g. concanavalin A, is 0.1 to 4, more
25 preferably 1 to 4 and particularly preferably 1 to 3. With concanavalin A, a DOL of 1 here corresponds to one mol of dye per mole of concanavalin A tetramer (MW = 104,000). With a high DOL and use of relatively nonpolar dyes (typically long wavelength fluorescent
30 dyes) the glucose-binding protein is preferably functionalized with PEG. The preferred degree of pegylation is 0.1 to 5, preferred molecular weight 200 to 10,000, particularly preferably 800 to 8000, still more preferably 800 to 5000.

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In the context of the experiments on which the present invention is based, it was established that chemical modifications of the glucose-binding proteins which

mediate increased water-solubility in comparison to unmodified forms can advantageously be used in order to achieve a better degree of labeling on the glucose-binding proteins. For the modified glucose-binding proteins according to the present invention, higher degrees of labeling with a dye can also be achieved than for unmodified forms thereof. For such modified concanavalin A proteins, concentrations preferably greater than 0.5 mg/(g matrix) and quite particularly preferably between 2 and 60 mg/(g matrix) can be achieved. Surprisingly, the measured glucose activity of modified concanavalin A proteins here was also comparable with that of native concanavalin A in hydrogel.

The improved solubility is particularly relevant when the glucose-binding proteins are to be labeled with a dye, since glucose-binding proteins labeled with dyes, for example a concanavalin A modified with one of the aforesaid oxazine dyes, have still further reduced solubility in aqueous solution. Here, the higher the degree of labeling is, the lower is the solubility in aqueous solution. However, it is precisely a higher degree of labeling that is needed for glucose-binding proteins as sensor components in the devices according to the invention.

The term "ligand of the glucose-binding protein" means a molecule which is capable of entering into specific bonding with the glucose-binding protein. During this, the molecule essentially interacts with the same binding site as glucose, so that the bound molecule can be displaced from the binding site on the glucose-binding protein by glucose. Hence suitable molecules are structurally related to glucose. The ligand of the glucose-binding protein is preferably an oligosaccharide, a glycosylated macromolecule, e.g. a glycosylated protein or peptide, or a glycosylated

nanoparticle. The aforesaid molecules which can be used as ligands of the glucose-binding protein are known in the state of the art and can readily be prepared by those skilled in the art. Particularly preferably, a
5 dextran is used as ligand of the glucose-binding protein.

The ligand of the glucose-binding protein in the device of the present invention is preferably coupled with a
10 rhodamine dye. Particularly preferably, ATTO590, ATTO610, ROX, TMR, rhodamine G6, Alexa Fluor rhodamine dyes or Dy590, and quite particularly preferably ATTO590, can be used in this.

15 The preferred degree of labeling (DOL) for the ligand of the glucose-binding protein, e.g. dextran, is 0.00003 to 0.016 (mol dye)/(mol subunit), particularly preferably 0.00032 to 0.0065 (mol dye)/(mol subunit) and quite particularly preferably 0.0008 to 0.0035 (mol
20 dye)/(mol subunit). The degree of labeling of the ligand also has an influence on the glucose activity. Excessively low degrees of labeling lead to poor glucose activity, as do excessively high ones.

25 From the aforesaid, it follows that in a preferred embodiment of the device according to the invention, the glucose-binding protein is concanavalin A. The concanavalin A concentration here is particularly preferably greater than 0.5 mg/(g matrix) and quite
30 particularly preferably between 2 and 60 mg/(g matrix). Likewise, through chemical modifications, preferably pegylation, acetylation, polyoxazolinylolation or succinylation, the concanavalin A exhibits increased water-solubility compared to the unmodified
35 concanavalin A. In a preferred embodiment, the glucose-binding protein is linked to an oxazine dye and the ligand of the glucose-binding proteins to a rhodamine dye. Quite particularly preferably, a concanavalin

A/dextran system is used in the device, wherein the dextran is linked to a rhodamine donor dye and the concanavalin A to an oxazine acceptor dye. In the preferred concanavalin A/dextran system, the components
5 are preferably present in a mass ratio (dextran/Con A) from 1:1 to 1:40, with mass ratios close to 1:10 being particularly preferable.

In the context of the present invention, it was
10 ascertained that the combination of oxazine and rhodamine dyes causes a heterodimeric interaction between the dye residues, which intensifies the glucose activity. Thus through the use of oxazine acceptor and rhodamine donor dyes, a 2.1-fold glucose activity can
15 preferably already be achieved in aqueous solution. In the hydrogel used in the context of the present invention, a 2.6-fold increase in the glucose activity could even be produced.

20 In the context of the present invention, it was advantageously ascertained that the use of a hydrogel consisting of a first hydrogel matrix made of alginate and a second hydrogel matrix which forms an interpenetrating network within the first, is capable
25 of creating an environment for the sensor components, namely the glucose-binding protein and the competitive ligands of the glucose-binding protein, which allows efficient determination of the glucose activity. Many approaches for glucose measurement by means of
30 fluorescence are successful in solution, but lose their activity when the sensor components are embedded in hydrogel matrices, since the mobility of the sensor components is restricted (Rounds 2007, J. Fluoresc. 17: 57-63; US 2007/0105176 A1). Surprisingly, however,
35 particularly also in view of the calculations of Birch et al. (Birch 2001, loc cit.), activity increased by up to 2.6-fold compared to aqueous solutions could be detected in the present case. By the selection of

suitable hydrogel matrices, an enrichment of the glucose-binding protein far above its solubility limit in aqueous solution could be achieved. The improved solubility found in the context of the present invention in the hydrogel matrices used according to the invention is attributable to the particular properties of the hydrogel matrices in the interplay with the sensor components and in particular the glucose-binding proteins, such as the concanavalin A. In the case of concanavalin A, for example a more than 10-fold increased concentration compared to a concentration achievable in free solution could be achieved. Usually, the solubility of the receptor component in free solution is further adversely influenced by the addition of the ligand, since the receptor/competitor complex exhibits a lower solubility owing to its size and often also owing to multivalences. While for example a concanavalin A/dextran complex in solution at a mass ratio of 1:10 already begins to precipitate beyond a concanavalin A concentration of 0.5 mg/(g solution), in a suitable hydrogel with the same mass ratio, concanavalin A concentrations of over 50 mg/(g matrix) can be prepared. Hence the usable concentration range can be extended 100-fold. This is of particular importance for applications wherein the analyte concentration is fixed and cannot be adjusted by dilution or concentration. Precisely such difficulties arise with in vivo applications such as the determination of the glucose level in body fluids. In the in vivo situation, the concentration of the analyte glucose cannot be adjusted to the specific assay conditions, but must rather be taken as given.

A further solubility problem arises in particular with in vivo applications of biological sensors. Owing to the higher wavelength, the intrinsic fluorescence of the tissue declines, so that with in vivo applications

long wavelength fluorescent dyes are used. However, these fluorescent dyes are typically apolar owing to their molecular structure and size (conjugated systems). If the glucose-binding protein is now labeled
5 with such a dye, the solubility further decreases, so that high degrees of labeling are also not possible. As already described above, it can therefore be necessary to functionalize the glucose-binding protein with e.g. polyethylene glycol, in order to enable increased
10 solubility and higher degrees of labeling associated therewith. However, functionalization with polyethylene glycol (pegylation) as a rule leads to a markedly reduced glucose activity of for example native concanavalin A (relative glucose activity of 0.4 or
15 less). Surprisingly, in the context of the present invention it was established that in the hydrogel of the device according to the invention this worsening does not occur. Rather, glucose activities for concanavalin A functionalized with polyethylene glycol
20 can be achieved which are comparable with those of native concanavalin A (relative glucose activity = 2.2 or 2.6). Furthermore, it was found that the glucose activity can be still further raised by increasing the degree of labeling on the concanavalin A in a hydrogel
25 such as is used in the device of the present invention. Thus, surprisingly, in the enriching hydrogel, by increasing the degree of labeling on the concanavalin A functionalized with polyethylene glycol, a doubling of the glucose activity could even be achieved. Compared
30 to the measurement in solution, the glucose activity in the hydrogel of the device according to the invention is even increased 4.3-fold. The use of the hydrogel in the device according to the invention thus makes it possible to prepare concentration ratios for the
35 glucose-binding protein and its ligand which with equivalent degrees of labeling allow glucose activities which are increased 4-fold compared to concentrations which are preparable in aqueous solutions. This

advantageously also enables the use of the device according to the invention under conditions wherein the analyte concentration cannot be adapted to the assay conditions, e.g. with in vivo applications.

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For the determination of the glucose concentration under in vivo conditions, e.g. in diabetics, an analyte concentration of 50 to 500 mg/dl has to be resolved. Such resolution can readily be achieved with the
10 devices according to the invention. In this, the devices can be used as a sensor both ex vivo and also in vivo. With in vivo applications, the sensor device can for example be placed subcutaneously, in the eye, e.g. subconjunctivally, or at other sites in the body
15 which enable an assessment of the glucose activities measured.

The invention thus also relates to the use of a device according to the invention as described above for
20 determining the glucose content in a sample.

In the context of the present invention, the term "sample" should be understood to mean a composition, preferably an aqueous composition, which presumably or
25 actually contains glucose. The sample is preferably a biological sample. Quite particularly preferably, the sample is a body fluid, in particular tissue fluid (e.g. interstitial fluid), blood, plasma, serum, lymph, saliva, tear fluid, sweat or urine. Particularly
30 preferably, the sample is tissue fluid, blood, serum or plasma.

Provided that the sample is a biological material, e.g. a body fluid, it can preferably be obtained from a test
35 subject who either actually or presumably has impaired glucose metabolism. The device according to the invention can thus be used for ex vivo diagnosis of diseases or impairments of the glucose metabolism, in

particular for the diagnosis of diabetes mellitus or metabolic syndrome. Furthermore, the device according to the invention can be used not only for diagnosis, but also for monitoring the glucose level. The device
5 thus also enables the support of therapeutic decisions, e.g. insulin doses which have to be administered in response to an altered glucose level.

For ex vivo use, the device according to the invention
10 can for example be introduced into microtiter plates and anchored there. Samples for assay are then applied into the wells of the microtiter plates and can then be assayed with a reader device. Such an approach enables the simultaneous assay of a large number of samples and
15 is thus also economical, in particular in clinical diagnostic practice.

In the context of in vitro use, the invention also relates to a method for determining the quantity of
20 glucose in a sample which actually or presumably contains glucose, comprising the steps:

- (a) Contacting of the device according to the invention with the sample for a period and under conditions which enables the binding of the
25 glucose contained in the samples to the glucose-binding proteins from the device; and
- (b) Determining the quantity of ligand displaced from the glucose in the device, whereby the quantity of glucose is determined.

30 The method according to the invention described above can comprise still further steps. For example, further steps can relate to the processing of the sample, e.g. the obtention of serum from whole blood. Further steps
35 could be performed in order to relate the glucose content determined to pathological changes in the glucose metabolism. For this, the content determined could be compared with reference quantities which are

indicative for certain pathological states, e.g. diabetes mellitus or metabolic syndrome. Such methods can then also be used for in vitro diagnosis of diabetes mellitus or metabolic syndrome. The method
5 according to the invention or individual steps thereof can be performed automatedly, e.g. by computer implementation and/or robot systems.

Suitable samples which can be analyzed with the method
10 described above are described in more detail at another place in the description.

The term "quantity" relates both to the determination of absolute quantities and also of relative quantities.
15 The determination of the absolute quantity can preferably be effected by means of a calibration curve which is created from measured values for known glucose contents with the method according to the invention. Relative quantities in the sense of the invention are
20 quantities which are set in relation to a normalization parameter. It goes without saying that in the context of the method according to the invention parameters can also be determined which can be derived by mathematical operations from the quantity values determined.

25 In the context of the method according to the invention, the contacting should enable penetration of the sample and hence of the glucose contained therein into the device. Further, the contacting should enable
30 the competition of glucose with the ligand on the glucose-binding protein which is embedded in the device.

In the method according to the invention, the detection
35 of the displacement of the ligand is preferably effected by measuring the increase in the intensity of fluorescence which is emitted by a donor dye as described elsewhere herein. The increase results from

before the displacement of the ligand from the glucose-binding protein, since the fluorescent intensity of the donor dye is reduced in the complex with the glucose-binding protein. However, it goes without saying that
5 other techniques for detecting the release of the ligand can also be used.

However, as well as the previously described ex vivo applications and ex vivo methods of the device
10 according to the invention, the invention also relates to the device according to the invention described above for use for diagnosing impaired glucose metabolism in a test subject. Preferably here, the impaired glucose metabolism is caused by diabetes
15 mellitus or metabolic syndrome.

In the in vivo use of the device according to the invention, this is introduced into the body. Here it should be noted that the measurement of the glucose
20 level, which is of course also the basis for the diagnosis, requires that the device comes into contact with a body fluid which contains glucose, wherein the concentration of glucose in the fluid is representative of the glucose level to be determined. Suitable body
25 fluids are enumerated at another place in the description. Particularly preferably, the body fluid is tissue fluid. The device introduced into the body then generates a signal which can be evaluated for making the diagnosis.

30 The device according to the invention is preferably introduced at places in the body which allow optical measurement of the signal generated by the device. Places with either a small tissue thickness between
35 device and body surface or with transparent tissues which can be effectively penetrated by the generated signal are suitable. Particularly preferably, the device is positioned under the skin (subcutaneously) or

in the eye, e.g. subconjunctivally. Appropriate methods for the implantation of the device are known in the state of the art.

5 Alternatively, the signal created by the device according to the invention can also be transferred outside the body by means of a suitable transfer medium. For this, a signal-conducting material can preferably be used as flexible cable, e.g. a glass
10 fiber cable. However, the transfer of the signal can also be effected wirelessly, e.g. as an infrared, radio or wireless signal. It goes without saying that in this case the signal created by the device according to the invention must firstly be read by a detector which must
15 likewise be installed in the device or at least in spatial proximity and be converted into an electromagnetic signal, for example a wireless signal. This electromagnetic signal can then be received by a receiver lying outside the body and evaluated.

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Furthermore, the present invention relates to the device according to the invention, as described above, for use in determining the need for a therapeutic measure in a test subject with impaired glucose
25 metabolism.

Appropriate therapeutic measures comprise those which are used for the treatment of diabetes mellitus or metabolic syndrome. As well as the administration of
30 drugs, e.g. insulin, this also includes the implementation of other therapeutic measures concerning which a decision can be taken on the basis of the impaired glucose metabolism determined, such as therapeutic interventions, e.g. gastric bypass
35 operations, or changes in lifestyle, e.g. the implementation of special diets.

In the context of the previously described use of the device, this can also be coupled with a further device, e.g. a device which controls the delivery of a drug. Here, the device of the present invention can preferably be coupled with a device for the delivery of insulin. The delivery of insulin can then be controlled on the basis of the need determined by the device according to the invention. In this, a change in the glucose level in a sample determined with the device according to the invention is translated, e.g. by a data processing unit in the delivery device, into a command which specifies the requirement for the insulin delivery. The command then mediates the delivery of insulin into the blood as long as needed or in a quantity as needed.

The applications according to the invention of the device according to the invention described above thus allow efficient ex vivo and in vivo diagnosis of the blood sugar level and hence the early recognition of diseases which are associated with impaired glucose metabolism, and also the management of such diseases through its use in the context of clinical monitoring. In this connection, the devices are also suitable for reaching therapeutic decisions based on the diagnostic results determined.

The invention is illustrated by the following practical examples. However, the examples do not limit the protection range.

EXAMPLES

Example 1: Preparation of sensors for determining glucose

Preparation of hydrogel particles (enriching matrix)

1 g of sodium alginate is dissolved in 100 g water with stirring. 66.2 g of $\text{CaCl}_2 \times 2\text{H}_2\text{O}$ are dissolved in 4931.3 g water in a 5L beaker.

The alginate solution is passed into a dual nozzle via a pump. At the same time, compressed air is connected to the second inlet of the nozzle, so that the alginate solution is atomized into fine droplets. The droplets are carried by the air flow into a bath containing the calcium chloride solution, where they gel and sink to the bottom. The gelled beads are then collected.

Preparation of sensors in enriching matrix:

For loading, alginate beads are successively incubated in a dye-labeled concanavalin A solution and a dye-labeled dextran solution. The loaded beads are then centrifuged down and the supernatant solution decanted off. The loaded beads are optionally then incubated overnight in a solution of a second polymer (e.g. PVA or PEG-based) and optionally isolated by centrifugation. The beads are then mixed into an aqueous solution of a photochemically crosslinkable polymer. This mixture is then crosslinked with UV light in order to prevent the sensor components leaching out of the alginate beads.

The quantities for this depend on the concentration of the analyte to be measured and the degree of labeling is selected depending on the desired intensity of the fluorescence signal. As the photochemically crosslinkable polymer, for example Nelfilcon polymer, a polyvinyl alcohol modified with acrylamide groups, can be used. For the photochemical crosslinking, 0.1% Irgacure 2959 is also added. The finished solution is dispensed into suitable molds and cured with UV light.

35

Preparation of sensors in non-enriching matrix:

Dye-labeled concanavalin A solution and dye-labeled dextran solution are successively fed into a water-

based prepolymer mixture and stirred for 3 hours. The quantities for this depend on the concentration of the analyte to be measured and the degree of labeling is selected depending on the desired intensity of the fluorescence signal. As the photochemically crosslinkable polymer, for example Nelfilcon polymer, a polyvinyl alcohol modified with acrylamide groups, can be used. For the photochemical crosslinking, 0.1% Irgacure 2959 is also added. The finished solution is dispensed into suitable molds and cured with UV light.

Example 2: Determination of the glucose activity for the sensors

15 Determination of the glucose activity in sensors:

The fluorescence spectrum of the sensors is determined at various glucose concentrations. The change in the fluorescence intensities of the donor with increasing glucose content serves as a measure of the quality of the glucose sensor. Since with an in vivo glucose sensor glucose concentrations between 50 and 500 mg/dL have to be measured, the glucose activity is calculated as follows:

25
$$GA = (\text{intensity}_{500\text{mg/dL}} - \text{intensity}_{50\text{mg/dL}}) / \text{intensity}_{50\text{mg/dL}}$$

For better comparison, all response values are normalized to the response of the same system in solution. For the determination of the relative glucose activity, the glucose activity of the sensor (in matrix) is divided by the glucose activity in solution.

$$\text{Rel GA} = GA(\text{matrix}) / GA(\text{solution})$$

35 Determination of the glucose activity in solution:

Con A solution and dextran solution are diluted in buffer solution and stirred for several hours. The fluorescence spectrum of the solution is determined at

various glucose concentrations. The change in the fluorescence intensities of the donor with increasing glucose content serves as a measure of the quality of the system. Since with an in vivo glucose sensor glucose concentrations between 50 and 500 mg/dL have to be measured, the glucose activity is calculated as follows:

$$GA = (\text{intensity}_{500\text{mg/dL}} - \text{intensity}_{50\text{mg/dL}}) / \text{intensity}_{50\text{mg/dL}}$$

10

Example 3: Determination of the influence of the matrix

According to the invention, the glucose-binding protein and the ligand are incorporated into a hydrogel matrix which exhibits a certain interaction with the glucose-binding protein. For this, the hydrogel matrix is selected such that firstly the interaction between glucose-binding protein and hydrogel matrix is greater than that between glucose-binding protein and aqueous solution (enrichment of the sensor components). On the other hand, however, the interaction between glucose-binding protein and the analyte (glucose) must be unaffected or not significantly affected by the interaction between glucose-binding protein and hydrogel matrix.

With suitable selection of the hydrogel matrix, through the interaction between glucose-binding protein and hydrogel matrix an enrichment of the glucose-binding proteins in the matrix far above the solubility limit of the glucose-binding protein in aqueous solution is achieved. In the case of Con A, for example up to 10 times higher concentrations can be achieved than in free solution. The solubility problems become still more extreme after addition of the ligand (e.g. dextran), since the glucose-binding protein-ligand complex exhibits lower solubility owing to its size and often also owing to multivalences. While the Con A-

dextran complex in solution at a mass ratio of 1:10 already begins to precipitate beyond a Con A concentration of 0.5 mg/g, Con A concentrations of over 50 mg/g can be prepared in a suitable hydrogel matrix at the same mass ratio. Through the hydrogel matrix, the usable concentration range of the glucose-binding protein, e.g. Con A, can be increased 100-fold.

Since with an in vivo application the range of the analyte concentration is fixed and cannot be adjusted e.g. by dilution or concentration, the concentrations of glucose-binding protein and ligand must be adapted to the in vivo concentration range of the analyte. However, there is often the difficulty that glucose-binding protein concentrations and/or ligand concentrations which exceed the solubility limit would therefore be necessary.

For example, with the Con A-dextran system, in solution a Con A concentration of only 0.5 mg/g can be established at a mass ratio (dex : Con A) of 1:10, since otherwise the Con A-dextran complex begins to precipitate. Here, for the determination of the relative glucose activity (rel GA), the glucose activity (GA) achieved at this concentration is set equal to 1. As expected, in a non-enriching matrix the glucose activity is decreased to half (rel GA = 0.52) owing to the lower mobility of the sensor components. In enriching matrix, with the same concentration almost the same response as in solution is obtained (rel GA = 0.83). However, significantly higher Con A concentrations can also be prepared in the enriching matrix. At a Con A concentration of 10 mg/g, a 2.6-fold increased glucose activity is obtained in enriching hydrogel matrix (see table 1).

Table 1: Influence of the matrix

	in solution	in non- enriching matrix	in enriching matrix	in enriching matrix
Con A concentration	0.5 mg/g	0.5 mg/g	0.5 mg/g	10 mg/g
Rel GA	1	0.52	0.83	2.55

Native Con A with DOL = 1, dextran with 0.001 mol dye (D) per mol dex subunit (SU), mass ratio dex : Con A = 1:10

5

Example 4: Determination of the influence of the concentration of the receptor

In enriching matrix, a dependence of the glucose activity on the starting concentration of the glucose-binding protein and of the ligand is clearly seen. With a rise in the concentration, a rise in the glucose activity is also obtained. At very high receptor concentrations, the glucose activity again declines.

15 The optimal Con A concentration for an in vivo assay lies between 8 and 20 mg Con A/g matrix (see table 2).

Table 2: Influence of the receptor concentration

Con A conc. [mg/g]	0.5	10.0	13.3	30	52.2
Relative Glucose activity (rel GA)	0.83	2.55	2.76	2.14	1.76

Native Con A with DOL = 1, dextran with 0.001 mol dye (D) per mol dex subunit (SU), mass ratio dex : Con A = 1:10

25

Example 5: Determination of the influence of the degree of labeling receptor

Owing to the decreasing intrinsic fluorescence of the

tissue with higher wavelengths, long wavelength fluorescent dyes must be used with in vivo applications. These fluorescent dyes are typically quite nonpolar owing to the larger conjugated system.

5 If the glucose-binding protein is labeled with such dyes, then its solubility decreases, so that high degrees of labeling are often not possible owing to precipitation. In order to increase the solubility of the glucose-binding proteins, it is advantageous to

10 functionalize this e.g. with polyethylene glycol. Thereby, the solubility of the glucose-binding protein increases, as a result of which higher degrees of labeling are possible in the synthesis.

15 However, pegylated Con As in solution lead to only less than half of the glucose activity of native Con A (rel GA = 0.4). Surprisingly, in the hydrogel matrix this worsening does not arise. With PEG-Con A, comparable glucose activity to that with native Con A is obtained

20 (rel GA = 2.2 vs. 2.6) (see table 3).

Table 3: Influence of the degree of labeling (DOL)

DOL Con A		1	1
Type		native	pegylated
Solution, c = 0.5 mg Con A/g	rel GA	1	0.43
Matrix, c = 10 mg Con A/g	rel GA	2.55	2.22

Through the pegylation, it first becomes possible to

25 equip Con A with a high degree of labeling with a long wavelength fluorescent dye, which is in the long-term stable in solution and does not precipitate. Surprisingly, no worsening of the glucose activity due to the pegylation is observed in enriching matrix

30 compared to in solution. Hence, only through the positive effect of the matrix is it possible to use PEG-Con A with a high degree of labeling at high

concentrations in an assay.

Astonishingly, the glucose activity can be further increased by increasing the degree of labeling on the Con A. In the enriching hydrogel matrix, with Con A
5 with DOL = 2.5, almost a doubling of the glucose activity is achieved (rel GA = 4.3 vs. 2.6). In direct comparison to the measurement in solution, the glucose activity in matrix is thus even increased 4.3-fold through the combined effect of concentration and DOL
10 (see table 4).

Table 4: Influence of the degree of labeling (DOL)

DOL Con A		1	1	2.5*	2.5
Type		native	pegylated	native*	pegylated
Solution, c = 0.5 mg Con A/g	rel GA	1	0.43		1.03
Matrix, c = 10 mg Con A/g	rel GA	2.55	2.22		4.28

*native ATTO680-Con A with a DOL of >1.5 is not stable in solution and precipitates.

15

Example 6: Determination of the influence of the fluorescent dyes

The structure of the fluorescent dyes which are bound
20 to glucose-binding protein and ligand respectively also has an influence on the glucose activity. A combination of a rhodamine and an oxazine dye has proved particularly suitable. For the Con A-dextran system, a rhodamine donor (e.g. ATTO590- or ATTO610-dextran or
25 ROX-dextran) and an oxazine acceptor (e.g. ATTO655- or ATTO680-Con A, or Evoblue30-Con A) is particularly preferable. Thereby, in contrast to the conventionally used TMR-Con A/FITC-dextran system (xanthene - rhodamine combination), a 2.1-fold glucose activity in
30 solution and even 2.6-fold in enriching matrix is

obtained. With a rhodamine-oxazine dye pair, a heterodimer interaction can arise between the dyes which intensifies the glucose activity (see tables 5 and 6).

5

Table 5: Influence of the dyes

Con A-dye		ATTO680	TMR
Dextran-dye		ATTO590	FITC
DOL Con A		2.5*	2.3
Type		native*	native
Solution, c = 0.5 mg Con A/g	rel GA	2.4	1.17
Relative activity compared to the conventional system FITC/TMR		2.05	1
Matrix, c = 10 mg Con A/g	rel GA	4.92	1.9
Relative activity compared to the conventional system FITC/TMR		2.59	1

Corresponding values extrapolated on the basis of the experimental values with PEG-Con A:

FITC-dextran with 0.01 mol D/mol UE, mass ratio dex :

10 Con A = 1:10

Table 6: Influence of the dyes

		ATTO590/ATTO680	ROX/Evoblue30
Matrix, c = 10 mg Con A/g	rel GA	2.55	2.1

Native Con A with DOL = 1, dextran with 0.001 mol dye (D) per mol dex subunit (SU), mass ratio dex :

15 1:10

Example 7: Determination of the influence of the nature of the hydrogel matrix

The glucose activity is also dependent on the nature of the hydrogel matrix. The hydrogel matrix can consist of one polymer or of a mixture of several polymers. A hydrogel matrix made of alginate or a mixture of an alginate and a polyvinyl alcohol hydrogel has proved advantageous. Through its interaction with the Con A and the dextran, the alginate hydrogel enables the enrichment of the sensor components in high concentrations.

10

The prepolymer of the 2nd hydrogel (e.g. PVA) can penetrate into the alginate beads and after its crosslinking form an interpenetrating network with the alginate. The mesh width of the interpenetrating network influences the mobility of the molecules of the glucose-binding protein and of the ligand and thus also the glucose activity. The mesh width can be influenced through the nature, the molecular weight and the solids content of the polymer and the content of crosslinker groups which determines the number of junctions.

A lower content of crosslinker groups leads to a higher glucose activity. Through a halving of the crosslinker groups, the activity can even be increased 1.5-fold. Hence, compared to the system in solution, a 4.2-fold improvement in the glucose activity can also be achieved without an increase in the degree of labeling on the Con A (see table 7).

30 Table 7: Influence of the crosslinker content

Crosslinker content (mmol/g)	0.486	0.33	0.26
Rel GA	2.76	4.04	4.24

Native Con A (13 mg/g) with DOL = 1, dextran with 0.001 mol dye (D) per mol dex subunit (SU), mass ratio dex : Con A = 1:10

35 The solids content of the network also has an influence on the glucose activity. The higher the solids content,

the lower is the glucose activity (see table 8).

Table 8: Influence of the solids content

SC Prepolymer	25%	30%	35%	40%
Rel. GA	2.87	2.76	2.48	1.96

Native Con A (13 mg/g) with DOL = 1, dextran with 0.001 mol dye (D) per mol dex subunit (SU), mass ratio dex :

5 Con A = 1:10

EyeSense AG

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Claims

5 1. A device comprising a hydrogel having a glucose-binding protein incorporated therein and a ligand of the glucose-binding protein, wherein the hydrogel comprises a first hydrogel matrix made of alginate and a second hydrogel matrix which forms an
10 interpenetrating network within the first hydrogel matrix.

2. The device as claimed in claim 1, wherein the second hydrogel matrix consists of a water-soluble
15 polymer with at least one crosslinkable group per molecule and a molecular weight of at most 500,000.

3. The device as claimed in claim 1 or 2, wherein the second hydrogel matrix is selected from the group
20 consisting of: polyvinyl alcohols (PVAs; including copolymers with vinyl acetates and/or ethylene), polyethylene glycols (PEGs), and polyhydroxyalkanoates (PHAs), poly (2-methyl-2 oxazolines), poly (2-ethyl-2 oxazolines), poly (2-hydroxyethyl-2 oxazolines), poly
25 (2-(1-(hydroxymethyl)-ethyl)-2 oxazolines), poly-(hydroxyethyl methacrylate) (PHEMA), poly-(hydroxyethyl acrylate) (PHEA), poly-vinylpyrrolidones, poly-(dimethyl)acrylamide, poly-(hydroxyethyl)acrylamide, poly-(ethylene-co-vinyl alcohol), poly(vinyl acetate-co-vinyl alcohol), poly(ethylene-co-vinyl acetate-co-vinyl alcohol) and poly(ethylene glycol-co-propylene glycol).

4. The device as claimed in one of claims 1 to 3,
35 wherein the first and/or the second hydrogel matrix,

which are capable of interacting with the glucose-binding protein.

5 5. The device as claimed in one of claims 1 to 4,
wherein the glucose-binding protein is selected from
the group consisting of: lectins, enzymes which bind
glucose as substrate, antibodies which specifically
recognize glucose, and aptamers which specifically
recognize glucose.

10 6. The device as claimed in one of claims 1 to 5,
wherein the ligand of the glucose-binding protein is an
oligosaccharide, a glycosylated macromolecule or a
glycosylated nanoparticle.

15 7. The device as claimed in one of claims 1 to 6,
wherein the glucose-binding protein is concanavalin A.

20 8. The device as claimed in claim 7, wherein the
concanavalin A concentration is greater than 0.5 mg/(g
matrix).

25 9. The device as claimed in claim 8, wherein the
concanavalin A concentration lies between 2 and 60
mg/(g matrix).

30 10. The device as claimed in one of claims 7 to 9,
wherein the concanavalin A exhibits increased water
solubility compared to unmodified concanavalin A owing
to chemical modification.

35 11. The device as claimed in claim 10, wherein the
modification is selected from the group consisting of:
pegylation, acetylation, succinylation and
polyoxazolinylolation.

12. The device as claimed in one of claims 1 to 11,
wherein the glucose-binding protein is linked to an

oxazine dye and the ligand of the glucose-binding protein to a rhodamine dye.

13. The use of a device as claimed in one of claims 1
5 to 12 for determining the glucose content in a sample.

14. The use as claimed in claim 13, wherein the sample
has been obtained from a test subject who exhibits
impaired glucose metabolism or who presumably exhibits
10 impaired glucose metabolism.

15. The device as claimed in one of claims 1 to 12 for
use in diagnosing impaired glucose metabolism in a test
subject.

15
16. The use as claimed in claim 13 or 14 or the device
as claimed in claim 15, wherein the impaired glucose
metabolism is caused by diabetes mellitus or metabolic
syndrome.

20
17. The device as claimed in one of claims 1 to 12 for
use in determining the need for a therapeutic measure
in a test subject with impaired glucose metabolism.

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