

DESCRIPTION

Title of Invention: ANTIBODY

Technical Field

5 [0001] A novel antibody, a use thereof, and the like are disclosed.

Background Art

[0002] Multiple myeloma, which is a typical example of a disease causing neoplastic growth of plasma cells, accounts for about 1%
10 of all cancers, and accounts for a little more than 10% of all hematological malignant tumors. Multiple myeloma is a disease in which a plasma cell present in bone marrow becomes cancerous (becomes an abnormal plasma cell as a result) and undergoes monoclonal growth.

15 [0003] In multiple myeloma, abnormal plasma cells (myeloma cells) spread to the bone marrow throughout the body and grow in every part of the bone marrow throughout the entire body. When the abnormal plasma cells grow, various symptoms including bone breakage appear. The myeloma cells produce M protein, which is an
20 abnormal immunoglobulin, to increase an M protein concentration in blood, and hence the blood becomes viscous.

[0004] The M protein does not function as an original antibody, which recognizes a foreign substance, such as a pathogen, which has entered the body. Accordingly, immunocompetence is reduced.
25 Those phenomena affect many organs, and thus various symptoms occur. Typical symptoms are bone pain and damage, hypercalcaemia, nephropathy and renal failure, anemia, and the like.

[0005] At present, as treatment of multiple myeloma, proteasome inhibitors, iMIDs, such as thalidomide and a derivative thereof,
30 specifically lenalidomide, and chemotherapy using, for example, melphalan in combination with prednisone, and hematopoietic stem cell transplantation are mainly employed.

[0006] However, the myeloma cells eventually acquire resistance to those therapeutic agents in most cases. Accordingly, the reality
35 of the current treatment means is that a

myeloma patient has an unpromising prognosis with a mean survival period after onset of from about 3 years to about 5 years. In addition, those therapeutic agents do not specifically act on only tumor cells serving as targets, and hence have a problem of showing toxicity also to normal cells, consequently causing serious side effects.

[0007] There have been attempts to develop a treatment method for multiple myeloma utilizing a monoclonal antibody. For example, an anti-CS1 antibody, and an anti-CD38 antibody, and the like are considered promising (Non Patent Literatures 1 and 2). In addition, in Patent Literature 1, there is disclosed a therapeutic agent for multiple myeloma or the like, which uses an anti-human CD48 monoclonal antibody as an active ingredient.

[0008] Integrins mainly form a heterodimer of an α -chain and a β -chain to serve a function as a receptor on a cell surface in a living body. There are many combinations of α -chains and β -chains of such integrins.

[0009] In addition, in Non Patent Literatures 4 to 6, there are disclosed chimeric antigen receptor T-cells (CAR-T cells) including an antigen recognition site having an affinity for a certain antigen.

Citation List

Patent Literature

[0010]

PTL 1: WO 2010/117059 A1

Non-patent Literature

[0011]

NPL 1: Journal of Clinical Oncology, 2012 Jun 1; 30(16): 1953-9.

NPL 2: Journal of immunology, 2011 Feb 1; 186(3): 1840-8.

NPL 3: J Biol Chem. 2012 May 4; 287(19): 15749-59.

NPL 4: J Immunol. 2009 Nov 1; 183(9): 5563-74.

NPL 5: N Engl J Med. 2014 Oct 16; 371(16): 1507-17.

NPL 6: Nat Biotechnol. 2002 Jan; 20(1): 70-5.

Summary of Invention

Technical Problem

[0012] The anti-CS1 antibody has relatively high specificity to
5 myeloma cells. However, the antibody alone cannot be said to have
a high anti-myeloma effect, and its effectiveness as a single
agent has not been demonstrated in a clinical test. It has been
found that the anti-tumor effect of the anti-CS1 antibody is
increased through combined use with lenalidomide, and it is
10 considered that an approval is being sought for the combined use.
Meanwhile, CD38 is also expressed in many normal blood cells
including CD34-positive hematopoietic progenitor cells, and hence
is an antigen having low specificity as a therapeutic target of
multiple myeloma. Under such circumstances, an object of the
15 present invention is to provide means that is more effective for
the treatment of, for example, a disease involving neoplastic
growth of plasma cells, such as multiple myeloma.

Solution to Problem

20 [0013] The inventors of the present invention have made extensive
investigations in order to achieve such object, and as a result,
have obtained an MMG49 antibody by performing screening through
use of specific binding to myeloma cells and progenitors thereof
as an indicator. In addition, the inventors have confirmed that
25 such antibody binds to a certain region of human integrin β_7 , and
have found that CAR-T cells generated using an antigen
recognition site of such antibody are extremely useful for the
treatment of myeloma. In addition, the inventors have also
elucidated that an epitope of the MMG49 antibody is present in
30 the region of the amino acid residue positions 20 to 109 of the
human integrin β_7 .

[0014] The present invention has been completed on the basis of
such findings, and encompasses inventions of a wide range of
aspects described below.

35 [0015] (I) Antibody

An antibody (I) encompasses antibodies described in the following items (I-1) to (I-25).

[0016] (I-1)

5 An anti-human integrin β_7 antibody, whose epitope is present in a region of the amino acid residue positions 20 to 109 of human integrin β_7 .

(1-1A)

10 An antibody according to the item (I-1), whose epitope is present in a region of the amino acid residue positions 33 to 109 of the human integrin β_7 .

(1-1B)

An antibody according to the item (I-1), whose epitope is present in a region of the amino acid residue positions 20 to 90 of the human integrin β_7 .

15 (1-1C)

An antibody according to the item (I-1), whose epitope is present in a region of the amino acid residue positions 33 to 90 of the human integrin β_7 .

(I-2)

20 An antibody according to the item (I-1), whose affinity for the epitope is increased in the presence of at least part of a region of the amino acid residue positions 379 to 721 of the human integrin β_7 .

(I-3)

25 An antibody according to the item (I-2), whose affinity for the epitope is increased in the presence of at least part of a region of the amino acid residue positions 417 to 721 of the human integrin β_7 .

(I-4)

30 An antibody according to the item (I-2), whose affinity for the epitope is increased in the presence of at least part of a region of the amino acid residue positions 564 to 721 of the human integrin β_7 .

(I-5)

35 An antibody according to the item (I-2), whose affinity for

light-chain CDR1 having the amino acid sequence set forth in SEQ ID NO: 6;

light-chain CDR2 having the amino acid sequence set forth in SEQ ID NO: 7; and/or

5 light-chain CDR3 having the amino acid sequence set forth in SEQ ID NO: 8.

(I-12)

An antibody according to any one of the items (I-1) to (I-10), the antibody including:

10 a heavy chain variable region having the amino acid sequence set forth in SEQ ID NO: 4; and/or

a light chain variable region having the amino acid sequence set forth in SEQ ID NO: 9.

(I-13)

15 An antibody according to any one of the items (I-1) to (I-12), which is Fv, scFv, a diabody, a triabody, a tetrabody, or a combination thereof.

(I-14)

20 An antibody according to any one of the items (I-1) to (I-11), the antibody including a constant region.

(I-15)

An antibody according to any one of the items (I-1) to (I-12) and (I-14), which is a chimeric antibody.

(I-16)

25 An antibody according to any one of the items (I-1) to (I-12) and (I-14), which is a humanized antibody.

(I-17)

An antibody according to any one of the items (I-1) to (I-12) and (I-14), which is a human antibody.

30 (I-18)

An antibody according to any one of the items (I-1) to (I-12) and (I-14) to (I-17), which is an immunoglobulin, Fab, F(ab')₂, a minibody, scFv-Fc, or a combination thereof.

(I-19)

35 An antibody according to any one of the items (I-1) to (I-

12) and (I-14) to (I-18), which is IgA, IgD, IgE, IgG, or IgM.
(I-20)

An antibody according to any one of the items (I-1) to (I-12) and (I-14) to (I-19), the antibody including a heavy chain
5 having the amino acid sequence set forth in SEQ ID NO: 5 and/or a light chain having the amino acid sequence set forth in SEQ ID NO: 10.

(I-21)

An antibody according to any one of the items (I-1) to (I-20), which has cytotoxic activity.

(I-22)

An antibody according to the item (I-21), in which the cytotoxic activity is ADCC activity and/or CDC activity.

(I-23)

15 An antibody according to any one of the items (I-1) to (I-22), which is a multispecific antibody.

(I-24)

An antibody according to any one of the items (I-1) to (I-23), which has a cytotoxin bound thereto.

20 (I-25)

An antibody according to any one of the items (I-1) to (I-24), which is a monoclonal antibody.

[0017] (II) Polynucleotide

A polynucleotide (II) encompasses a polynucleotide
25 described in the following item (II-1).

(II-1)

A polynucleotide, which has a base sequence encoding the amino acid sequence of the antibody (I).

[0018] (III) Host Cell

30 A host cell (III) encompasses a host cell described in the following item (III-1) or (III-2).

(III-1)

A host cell, which harbors the polynucleotide (II).

(III-2)

35 A host cell according to the item (III-1), which is a

eukaryotic cell.

[0019] (IV) Chimeric Antigen Receptor

A chimeric antigen receptor (IV) encompasses chimeric antigen receptors described in the following items (IV-1) to (IV-5).

(IV-1)

A chimeric antigen receptor, whose epitope is identical to that of the antibody (I).

(IV-2)

A chimeric antigen receptor according to the item (IV-1), the chimeric antigen receptor including an antigen recognition site of the antibody (I).

(IV-3)

A chimeric antigen receptor according to the item (IV-1) or (IV-2), the antigen recognition site including:

a heavy chain variable region including:

heavy-chain CDR1 having the amino acid sequence set forth in SEQ ID NO: 1;

heavy-chain CDR2 having the amino acid sequence set forth in SEQ ID NO: 2; and/or

heavy-chain CDR3 having the amino acid sequence set forth in SEQ ID NO: 3; and/or

a light chain variable region including:

light-chain CDR1 having the amino acid sequence set forth in SEQ ID NO: 6;

light-chain CDR2 having the amino acid sequence set forth in SEQ ID NO: 7; and/or

light-chain CDR3 having the amino acid sequence set forth in SEQ ID NO: 8.

(IV-4)

A chimeric antigen receptor according to any one of the items (IV-1) to (IV-3), in which the antigen recognition site includes:

a heavy chain variable region having the amino acid sequence set forth in SEQ ID NO: 4; and/or

a light chain variable region having the amino acid sequence set forth in SEQ ID NO: 9.

(IV-5)

5 A chimeric antigen receptor according to any one of the items (IV-1) to (IV-4), the chimeric antigen receptor having the amino acid sequence set forth in SEQ ID NO: 21.

[0020] (V) Polynucleotide

A polynucleotide (V) encompasses a polynucleotide described in the following item (V-1) or (V-2) unlike the polynucleotide
10 (II).

(V-1)

A polynucleotide, which encodes the amino acid sequence of the chimeric antigen receptor (IV).

(V-2)

15 A polynucleotide according to the item (V-1), which has the base sequence set forth in SEQ ID NO: 22.

[0021] (VI) Cell

A cell (VI) encompasses a cell described in any one of the following items (VI-1) to (VI-4) unlike the host cell (III).

20 (VI-1)

A cell, which harbors the polynucleotide (V).

(VI-2)

A cell according to the item (VI-1), which is a eukaryotic cell.

25 (VI-3)

A cell according to the item (VI-1) or (VI-2), which is a T-cell or an NK cell.

(VI-4)

30 A cell according to any one of the items (VI-1) to (VI-3), which is a chimeric antigen receptor T-cell or a chimeric antigen receptor NK cell.

[0022] (VII) Pharmaceutical Composition

A pharmaceutical composition (VII) encompasses pharmaceutical compositions described in the following items

35 (VII-1) to (VII-5).

(VII-1)

A pharmaceutical composition, including the antibody (I) or the cell (VI).

(VII-2)

5 A pharmaceutical composition according to the item (VII-1), in which the cell is the chimeric antigen receptor T-cell (VI-4).

(VII-3)

A pharmaceutical composition according to the item (VII-1) or (VII-2), which is for use in treatment of cancer.

10 (VII-4)

A pharmaceutical composition according to the item (VII-3), in which the cancer is blood cancer.

(VII-5)

15 A pharmaceutical composition according to the item (VII-4), in which the blood cancer is a disease causing neoplastic growth of plasma cells.

[0023] (VIII) Treatment or Prevention Method for Disease

A treatment or prevention method (VIII) for a disease encompasses treatment or prevention methods for a disease
20 described in the following items (VIII-1) to (VIII-6).

(VIII-1)

A treatment or prevention method for a disease, including administering a therapeutically effective amount of the antibody (I) or the cell (VI) to a subject.

25 (VIII-2)

A treatment or prevention method according to the item (VIII-1), in which the cell is the chimeric antigen receptor T-cell (VI-4).

(VIII-3)

30 A treatment or prevention method according to the item (VIII-1) or (VIII-2), in which the disease is cancer, and in which the subject is a patient who has developed cancer or an animal having a risk of developing cancer.

(VIII-4) A treatment or prevention method according to the
35 item (VIII-3), in which the cancer is blood cancer.

(VIII-5)

A treatment or prevention method according to the item (VIII-4), in which the blood cancer is a disease causing neoplastic growth of plasma cells.

5 (VIII-6)

A treatment or prevention method for multiple myeloma, targeting active-form human integrin β_7 .

[0024] (IX) Use

10 A use (IX) encompasses uses described in the following items (IX-1) to (IX-5).

(IX-1)

A use of the antibody (I) or the cell (VI), for producing a pharmaceutical composition.

(IX-2)

15 A treatment or prevention method according to the item (IX-1), in which the cell is the chimeric antigen receptor T-cell (VI-4).

(IX-3)

20 A use according to the item (IX-1) or (IX-2), which is for treatment of cancer.

(IX-4)

A use according to the item (IX-3), in which the cancer is blood cancer.

(IX-5)

25 A use according to the item (IX-4), in which the blood cancer is a disease causing neoplastic growth of plasma cells.

[0025] (X) Screening Method

A screening method (X) encompasses screening methods described in the following (X-1) to (X-5).

30 (X-1)

A screening method for an active ingredient of a pharmaceutical composition for treating or preventing cancer, the method including selecting, from a compound library, a candidate substance that specifically binds to human integrin β_7 and binds
35 to a region of the amino acid residue positions 20 to 109 of the

human integrin β_7 .

(X-2)

A screening method according to the item (X-1), further including selecting a substance having cytotoxic activity.

5 (X-3)

A screening method according to the item (X-1) or (X-2), in which the substance to be selected is a monoclonal antibody.

(X-4)

10 A screening method according to any one of the items (X-1) to (X-3), in which the cancer is blood cancer.

(X-5)

A screening method according to the item (X-4), in which the blood cancer is a disease causing neoplastic growth of plasma cells.

15 [0026] (XI) Diagnosis Method

A diagnosis method (XI) encompasses diagnosis methods described in the following items (XI-1) to (XI-5).

(XI-1)

20 A diagnosis method for cancer, including bringing a sample collected from a subject into contact with the antibody (I).

(XI-2)

A diagnosis method according to the item (XI-1), in which the sample collected from a subject is blood or bone marrow fluid.

(XI-3)

25 A diagnosis method according to the item (XI-1) or (XI-2), further including judging that the subject has developed, or has a risk of developing, cancer when cells that bind to the antibody (I) are detected.

(XI-4)

30 A diagnosis method according to the item (XI-3), in which the cancer is blood cancer.

(XI-5)

35 A diagnosis method according to the item (XI-4), in which the cells are plasma cells, and in which the cancer is a disease causing neoplastic growth of plasma cells.

[0027] (XII) Kit

A kit (XII) encompasses kits described in the following items (XII-1) to (XII-3).

(XII-1)

5 A kit for diagnosis of cancer, including the antibody (I).

(XII-2)

A diagnosis method according to the item (XII-1), in which the cancer is blood cancer.

(XII-3)

10 A kit according to the item (XII-2), in which the cancer is a disease causing neoplastic growth of plasma cells.

Advantageous Effects of Invention

[0028] The antibody of the present invention does not recognize
15 normal cells, and hence is useful as an active ingredient of a pharmaceutical composition. In particular, the antibody of the present invention is useful as an active ingredient of a therapeutic agent for cancer (e.g., blood cancer).

[0029] The antibody of the present invention is useful because
20 chimeric antigen receptor T-cells produced by applying its antigen recognition site to a chimeric antigen receptor can be used as an active ingredient of such pharmaceutical composition as described above.

25 Brief Description of Drawings

[0030] FIGS. 1 are results obtained in Example 2 by analyzing the binding of an MMG49 antibody to myeloma patient-derived bone marrow cells through use of FACS. (Left) A diagram for illustrating a method of identifying a myeloma progenitor cell
30 fraction (Myeloma progenitor cells), a myeloma plasma cell fraction (Myeloma plasma cells), and CD45⁺ leukocytes (CD45⁺ leukocytes). (Right) Graphs for showing the binding of the MMG49 antibody to each of the fractions.

FIGS. 2 are graphs for showing results obtained in Example
35 2 by analyzing, by FACS, the binding of the MMG49 antibody to the

myeloma progenitor cell fraction, myeloma plasma cell fraction, and CD45⁺ leukocytes of a plurality of myeloma patient-derived bone marrow cells (UPN1 to UPN5).

FIG. 3 is an illustration of a process of identifying an antigen protein recognized by the MMG49 antibody by an expression cloning method in Example 3. There is illustrated a process of concentrating BaF3 cells that bind to the MMG49 antibody, from an initial concentration of 0.1% or less, by FACS sorting.

FIGS. 4 are graphs for showing results obtained in Example 4 by staining ITGB7-deficient U266 cells generated using a Crisp-cas9 system with the MMG49 antibody or an FIB27 antibody (commercially available anti-integrin β_7 antibody), followed by FACS analysis.

FIG. 5 is an image for showing results obtained in Example 4 by subjecting a product immunoprecipitated from a cell lysate derived from MM1s myeloma cells with the MMG49 antibody or an isotype control antibody, to SDS-PAGE, and then performing western blot with a commercially available anti-integrin β_7 antibody (Abcam plc).

FIGS. 6 are graphs for showing results obtained in Example 5 by analyzing the binding of each of the MMG49 antibody, the FIB27 antibody, and an FIB504 antibody to each cell fraction of healthy person peripheral blood cells (in the figures, B-cells, T-cells, monocytes, neutrophils, red blood cells, and platelets are shown in the stated order from the left-hand side) through use of FACS.

FIGS. 7 are graphs for showing results obtained in Example 5 by analyzing, by FACS, the binding of the MMG49 antibody to each of cell fractions of myeloma patient-derived bone marrow cells. On the left-hand side, a method of identifying each cell fraction is illustrated, and on the right-hand side, graphs for showing the binding of MMG49 to each fraction are shown. In FIGS. 7A, a comparison between hematopoietic stem cell and progenitor cell fractions, and myeloma cells is shown, and in FIGS. 7B, a comparison between B/T lymphocyte fractions, and myeloma

progenitor cell and myeloma plasma cell fractions is shown.

FIGS. 8 are graphs for showing results obtained in Example 6 by analyzing the binding of each of the MMG49 antibody and the FIB27 antibody to each of various myeloma cell lines, and T-cells and B-cells derived from peripheral blood through use of FACS. There are also shown results of confirming the expression of ITGA4 (binding of an anti-integrin α_4 antibody) and the expression of ITGAE (binding of an anti-integrin α_E antibody) in the above-mentioned cells by FACS analysis.

FIGS. 9 are graphs for showing results obtained in Example 6 by analyzing, by FACS, the binding of the MMG49 antibody and the FIB27 antibody to U266 cells and ITGA4 (integrin α_4)-deficient U266 cells. There are also shown results obtained by analyzing, by FACS, the expression of ITGA4 (binding of the anti-integrin α_4 antibody) in the above-mentioned cells.

FIGS. 10 are graphs for showing results obtained in Example 7 by allowing integrin $\alpha_4\beta_7$ -forcibly expressing K562 cells and human normal peripheral blood-derived T-cells treated in the presence of $\text{Ca}^{2+}/\text{Mg}^{2+}$ or Mn^{2+} at 37°C for 20 minutes to react with the MMG49 antibody or an isotype antibody, then staining the cells using an anti-mouse IgG antibody as a secondary antibody, and subjecting the stained cells to FACS analysis.

FIG. 11 is a diagram for illustrating the construction of human/mouse chimeric integrin β_7 proteins and the presence or absence of the binding of the MMG49 antibody to 293T cells caused to transiently express the proteins in Example 8.

FIGS. 12 are graphs for showing results obtained in Example 8 by analyzing, by FACS, the binding of the MMG49 antibody to 293T cells caused to transiently express the human/mouse chimeric integrin β_7 proteins.

FIG. 13 is a graph for summarizing the results shown in FIGS. 12. In the graph in FIG. 13, the axis of ordinate represents the percentage of cells bound to the antibody, and the axis of abscissa represents various human/mouse chimeric integrin β_7 proteins.

FIGS. 14 are graphs for showing results obtained by staining MM1s cells and KMS12BM cells with a chimerized MMG49 antibody generated by linking variable regions of the MMG49 antibody to a human IgG4 antibody constant domain.

5 FIG. 15 is a scheme for illustrating a method of generating a CAR construct using variable regions of the MMG49 antibody.

FIG. 16 are graphs for showing results obtained by staining T-cells caused to express the CAR construct using variable regions of the MMG49 antibody with a PE-anti-human F(ab')₂ antibody.

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FIG. 17 are graphs for showing results obtained in Example 11 by quantitatively determining, by ELISA, the amounts of IFN- γ and IL2 produced through coculture of MMG49 antibody-derived CAR-T cells or T-cells having introduced therein GFP (control) with K562 cells expressing no integrin β_7 or K562 cells caused to forcibly express integrin $\alpha_4\beta_7$. *: $p < 0.05$.

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FIGS. 18 are graphs for showing results obtained in Example 11 by quantitatively determining, by ELISA, the amount of IFN- γ produced through coculture of MMG49 antibody-derived CAR-T cells or T-cells having introduced therein GFP (control) with MMG49 antigen-expressing cells or non-expressing cells.

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FIGS. 19 are graphs for showing results obtained in Example 11 by quantitatively determining, by ELISA, the amount of IL2 produced through coculture of MMG49 antibody-derived CAR-T cells or T-cells having introduced therein GFP (control) with MMG49 antigen-expressing cells or non-expressing cells.

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FIGS. 20 are graphs for showing results obtained in Example 11 by measuring, by ⁵¹Cr killing assay, the degree of cell damage caused by MMG49 antibody-derived CAR-T cells or T-cells having introduced therein GFP (control) with respect to K562 cells expressing no integrin β_7 or K562 cells caused to forcibly express integrin $\alpha_4\beta_7$. The y-axis of each of the graphs in FIGS. 20 represents a cell damage percentage (%).

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FIGS. 21 are graphs for showing results obtained in Example 11 by measuring, by ⁵¹Cr killing assay, the degree of cell damage

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caused by MMG49 antibody-derived CAR-T cells or T-cells having introduced therein GFP (control) with respect to MMG49 antigen-expressing cells or non-expressing cells.

FIGS. 22 are a diagram and graphs for illustrating and showing the design of a therapeutic experiment for a myeloma cell line MM1s engrafted in the bone marrow of an NOG mouse and results thereof in Example 11. Bone marrow cells after 1 week from the transfer of MMG49 antibody-derived CAR-T cells or T-cells having introduced therein GFP (control) were collected and analyzed by FACS. MM1s cells can be identified as human CD138⁺ cells. In an MMG49 antibody-derived CAR-T cell-administered group, MM1s cells in the bone marrow have almost completely disappeared.

FIGS. 23 are a diagram, images, and a graph for illustrating and showing the design of a therapeutic experiment for the myeloma cell line MM1s systemically engrafted to an NOG mouse and results thereof in Example 12. The amounts of myeloma cells before and after the transfer of MMG49 antibody-derived CAR-T cells or T-cells having introduced therein GFP (control) were evaluated by fluorescence intensity measurement based on IVIS imaging. In an MMG49 antibody-derived CAR-T cell-administered group, MM1s cells in the bone marrow have almost completely disappeared.

FIG. 24 is a view for illustrating a comparison between the amino acid sequence of integrin β_7 of human origin and the amino acid sequence of integrin β_7 of mouse origin.

FIGS. 25 are a diagram and graphs for illustrating and showing the construction of human/mouse chimeric integrin β_7 proteins and the presence or absence of the binding of the MMG49 antibody to 293T cells caused to transiently express the proteins in Example 13.

FIG. 26 is a graph for showing results of an experiment for investigating an epitope of the MMG49 antibody in Example 14. MFI on the axis of abscissa represents binding strength to the MMG49 antibody, and a higher numerical value indicates a higher avidity.

Description of Embodiments

[0031] Herein, "include" and "have" are so-called open language, but are each a concept including the closed language "consisting of", and in one embodiment, may be replaced by "consisting of".

[0032] A "myeloma progenitor cell" is a progenitor cell in a stage before differentiating into a myeloma plasma cell, and is characterized by highly expressing CD38, but not expressing CD138, which serves as a marker specific to a mature plasma cell.

Therefore, the myeloma progenitor cell is sometimes referred to as "CD38⁺⁺CD138⁻ cell" or "CD19⁻CD38⁺⁺CD138⁻ cell".

[0033] A "myeloma plasma cell" is generally also called a myeloma cell, and is a cell that produces M protein, which is an abnormal immunoglobulin. The myeloma plasma cell expresses CD138 in addition to highly expressing CD38. Therefore, the myeloma plasma cell is sometimes referred to as "CD38⁺⁺CD138⁺ cell" or "CD19⁻CD38⁺⁺CD138⁺ cell".

[0034] The myeloma progenitor cell and the myeloma plasma cell also mean a tumor progenitor cell and a neoplastic plasma cell, respectively, in a disease causing neoplastic growth of plasma cells other than multiple myeloma.

[0035] A "hematopoietic progenitor cell" is a cell capable of differentiating into various hematopoietic cells. The hematopoietic progenitor cell is characterized by expressing CD34. Therefore, herein, the hematopoietic progenitor cell is sometimes referred to as "CD34⁺ cell".

[0036] (I) Antibody

An antibody (I) is preferably an anti-human integrin β_7 antibody whose epitope is present in the region of the amino acid residue positions 20 to 109 of human integrin β_7 .

[0037] More preferred examples of the antibody (I) may include: an antibody whose epitope is present in the region of the amino acid residue positions 33 to 109 of human integrin β_7 ; and an antibody whose epitope is present in the region of the amino acid residue positions 20 to 90 of human integrin β_7 . The most preferred

example thereof may be an antibody whose epitope is present in the region of the amino acid residue positions 33 to 90 of human integrin β_7 .

[0038] The human integrin β_7 is not particularly limited, and may
5 be a transmembrane protein having the amino acid sequence set forth in SEQ ID NO: 31, the protein being capable of forming a heterodimer with integrin α . Specific examples of the integrin α may include integrin α_4 and integrin α_E .

[0039] Specific examples of the amino acid sequence of the human
10 integrin β_7 may include, in addition to the amino acid sequence set forth in SEQ ID NO: 31, amino acid sequences described in, for example: ACCESSION: EAW96675, VERSION: EAW96675.1, GI: 119617081; ACCESSION: NM000889, VERSION: NM000889.2, GI: 540344585; ACCESSION: XM005268851, VERSION: XM005268851.2, GI: 767974096; ACCESSION: XM006719376, VERSION: XM006719376.2, GI: 767974098; and ACCESSION: XM005268852, VERSION: XM005268852.3, GI: 767974097, listed in the NCBI database.

[0040] The following description regarding the human integrin β_7 is made on the basis of the amino acid sequence set forth in SEQ
20 ID NO: 31. However, for any other amino acid sequence of the human integrin β_7 , a person skilled in the art can easily judge which region or site of the other amino acid sequence of the human integrin β_7 corresponds to a region and/or site of the human integrin β_7 to be described below by determining the homology of
25 the other amino acid sequence to the amino acid sequence set forth in SEQ ID NO: 31 in silico.

[0041] The region of the amino acid residue positions 1 to 19 of the human integrin β_7 is a peptide fragment serving as a signal peptide and being absent when the human integrin β_7 functions as a
30 membrane protein in a living body. Accordingly, when the human integrin β_7 exhibits a function as a membrane protein, its N-terminus is the amino acid residue at position 20 of the above-mentioned amino acid sequence.

[0042] The region of the amino acid residue positions 20 to 109 of
35 the human integrin β_7 includes a PSI domain. The PSI domain of

the human integrin β_7 and the PSI domain of mouse integrin β_7 are known to have a high homology of about 80% or more. However, as illustrated in FIG. 24, when compared to each other, the amino acid residues of the regions of the amino acid residue positions 20 to 109 including the PSI domains of the human integrin β_7 and the mouse integrin β_7 differ from each other at a total of 15 amino acid residues, specifically amino acid residues at position 23, position 26, position 28, position 30, position 32, position 35, position 36, position 38, position 41, position 42, position 48, position 93, position 94, position 102, and position 109 of the human integrin β_7 .

[0043] Therefore, it is preferred that the epitope of the antibody (I) be associated with any one or more, preferably two or more, more preferably three or more of those 15 amino acid residues.

Specifically, the epitope of the antibody (I) is preferably present in the region of the amino acid residue positions 23 to 109 of the human integrin β_7 , more preferably present in the region of the amino acid residue positions 23 to 48 or the region of the amino acid residue positions 93 to 109.

[0044] The epitope of the antibody (I) in another more preferred embodiment may be: the region of the amino acid residue positions 23 to 48; the region of the amino acid residue positions 93 to 109; or a three-dimensional region that is a combination of the region of the amino acid residue positions 23 to 48 and the region of the amino acid residue positions 93 to position 109.

[0045] The epitope of the antibody (I) may be a linear epitope, or may be a conformational epitope (also called a non-linear epitope). It is known to a person skilled in the art that the linear epitope is a case in which consecutive amino acid residues serve as an epitope and the conformational epitope is an epitope formed of non-consecutive amino acid residues.

[0046] For example, the case in which the above-mentioned three-dimensional region that is a combination of the region of the amino acid residue positions 23 to 48 and the region of the amino acid residue positions 93 to 109 serves as the epitope may be

given as an example corresponding to the conformational epitope,
and a case in which a region of non-consecutive amino acid
residues included in the region of the amino acid residue
positions 20 to 109 serves as the epitope is also encompassed in
5 the conformational epitope.

[0047] Of the above-mentioned epitopes, it is preferred that the
amino acid residue at position 48 be strongly related to the
epitope of the antibody (I) or be included in the epitope of the
antibody (I).

10 [0048] A person skilled in the art can understand about specific
linear epitopes and conformational epitopes with reference to,
for example, JP 2011-527572 A, JP 2009-534401 A, or "Dissecting
antibodies with regards to linear and conformational epitopes."
Forsstrom B, Axnas BB, Rockberg J, Danielsson H, Bohlin A, Uhlen
15 M. PLoS One. 2015 Mar 27; 10(3): e0121673. doi:
10.1371/journal.pone.0121673. eCollection 2015.

[0049] In other words, the foregoing means that the antibody (I)
is an antibody that specifically binds to the region of the amino
acid residue positions 20 to 109 of the human integrin β_7 , and in
20 particular, preferably specifically binds to the region at
positions from 23 to 109, more preferably specifically binds to
the region at positions from 23 to 48 and/or positions from 93 to
109.

[0050] In addition, the property of the antibody (I) of binding to
25 the region of the amino acid residue positions 20 to 109 of the
integrin β_7 , which serves as the epitope, is sometimes referred to
as affinity for the epitope. Accordingly, the term "affinity for
the epitope is increased" has the same meaning as "specific
binding capacity for the epitope is increased."

30 [0051] The term "specific" may be distinguished from the term
"selective".

[0052] As another embodiment of the antibody (I), it is preferred
that the affinity of the antibody (I) for the epitope be
increased in the presence of at least part of the region of the
35 amino acid residue positions 379 to 721 of the human integrin β_7 .

[0053] The "at least part of the region of the amino acid residue positions 379 to 721" means that any one of the region of the amino acid residue positions 379 to 721 and a partial region thereof may be adopted. Specific examples of the "partial region
5 thereof" include: at least part of the region of the amino acid residue positions 417 to 721 of the human integrin β_7 ; at least part of the region of the amino acid residue positions 564 to 721 of the human integrin β_7 ; at least part of the region of the amino acid residue positions 379 to 563 of the human integrin β_7 ; at
10 least part of the region of the amino acid residue positions 417 to 563 of the human integrin β_7 ; and at least part of the region of the amino acid residue positions 379 to 416 of the human integrin β_7 . That is, the affinity of the antibody (I) for the epitope can be increased in the presence of any of those regions.

[0054] The term "in the presence of" means that the region of the amino acid residue positions 20 to 109 of the human integrin β_7 and at least part of the region of the amino acid residue positions 379 to 721 of the human integrin β_7 may be present in the same molecule, or the two regions may be present as separate
20 molecules. It is preferred that the two regions be present in the same molecule. The term "in the presence of" may be read as "by".

[0055] A person skilled in the art can easily confirm that the affinity of the above-mentioned antibody (I) for the epitope is increased, by a commonly used immunoassay method described in,
25 for example, Examples to be described below.

[0056] For example, cells caused to express a human/mouse chimeric integrin β_7 protein (#4960), which is various human/mouse chimeric integrin β_7 proteins described in Example 8, and which includes the region of the amino acid residue positions 1 to 109 of
30 integrin β_7 of human origin and includes the region of the amino acid residue positions 722 to 798 of the integrin β_7 of human origin, are prepared, and a human/mouse chimeric integrin β_7 protein (#4961) in which the region of #4960 of the amino acid residue positions 379 to 721 of the integrin β_7 of human origin is
35 replaced with the region of the amino acid residue positions 379

to 721 of integrin β_7 of mouse origin is prepared. In this case, the increase in the affinity of the antibody (I) for the epitope may be confirmed by comparing the degrees of binding of the antibody (I) between cells expressing the latter (#4961) and
5 cells expressing the former (#4960).

[0057] As another embodiment of the antibody (I), it is preferred that the affinity of the antibody (I) for the epitope be increased by activating the human integrin β_7 . Probably because activated human integrin β_7 has a structural feature in a region
10 including the epitope, the affinity of the antibody (I) for the epitope is increased.

[0058] A method of activating the human integrin β_7 is known. For example, by allowing a phorbol ester, such as PMA, a manganese salt, or the like to act on cells expressing the human integrin β_7 ,
15 e.g., cells selected from blood cells and immune cells, such as plasma cells, NK cells, T-cells, B-cells, lymphoblasts, Burkitt lymphoma-derived cells, and dendritic cells, the human integrin β_7 expressed in the cells may be activated. In addition, without being limited to the above-mentioned specific cells, cells caused
20 to express the human integrin β_7 may be used and treated with a phorbol ester, a manganese salt, or the like to activate the human integrin β_7 .

[0059] A person skilled in the art can easily confirm that the affinity of the antibody (I) for the epitope is increased through
25 activation of the human integrin β_7 , by a commonly used immunoassay method described in, for example, Examples to be described below.

[0060] For example, cells caused to express #4960 or #4961, which is the various human/mouse chimeric integrin β_7 proteins described
30 in Example 8 and includes the region of the amino acid residue positions 1 to 109, are prepared, and the cells are subjected to integrin β_7 -activating means as described in Example 7. After that, affinities before and after the activation treatment are compared to each other through measurement using immunoassay
35 means. Thus, the increase in the affinity of the antibody (I) for

the epitope in the cells after the activation may be confirmed.

[0061] As another embodiment of the antibody (I), the antibody (I) may be an anti-human integrin β_7 antibody having a feature of having a higher affinity for human integrin β_7 expressed on myeloma-derived cells than for human integrin β_7 expressed on normal cells.

[0062] The normal cells are not particularly limited as long as the cells are derived from a healthy person, and may be, for example, blood-derived normal cells. Of such normal cells, normal plasma cells are preferred.

[0063] A method of confirming that the antibody has a higher affinity for human integrin β_7 expressed on myeloma cells than for human integrin β_7 expressed on such normal cells can easily be performed by a person skilled in the art by a commonly used immunoassay method described in, for example, Examples to be described below.

[0064] The "commonly used immunoassay method" is not particularly limited as long as the method involves measurement using various antibodies irrespective of the antigen. Examples thereof may include a flow cytometry method (FACS), cell sorting involved therein, western blotting, ELISA, an immunoprecipitation method, a SPR method, and a QCM method.

[0065] As another embodiment of the antibody (I), an epitope of the antibody (I) is preferably identical to that of an MMG49 antibody disclosed in Examples to be described later. An antibody identical to the MMG49 antibody is most preferred. For a method of producing the MMG49 antibody, reference may be made to Examples to be described below.

[0066] As another embodiment of the antibody (I), the antibody (I) is preferably an antibody of an embodiment including a heavy chain variable region and/or a light chain variable region. That is, the antibody (I) may be the heavy chain variable region alone, or may be the light chain variable region alone. The antibody (I) is preferably an antibody including the heavy chain variable region and the light chain variable region.

[0067] A variable region is also called an antigen recognition site, and is understood by a person skilled in the art to be a site important for an antibody to recognize an antigen. Such variable region has three regions called hypervariable regions
5 (also referred to as complementarity determining regions [CDRs]), and it is also known to a person skilled in the art that the CDRs are extremely important regions most involved in the antigen recognition function of an antibody.

[0068] The heavy chain variable region included in the other
10 embodiment of the antibody (I) includes any one or more of heavy-chain CDR1, heavy-chain CDR2, and heavy-chain CDR3. That is, the heavy chain variable region may contain heavy-chain CDR1, heavy-chain CDR2, or heavy-chain CDR3 alone, and preferably includes at least heavy-chain CDR3. A more preferred embodiment includes
15 heavy-chain CDR1, heavy-chain CDR2, and heavy-chain CDR3 in the stated order from the amino-terminus (N-terminus).

[0069] The light chain variable region may be similar to the heavy chain variable region, i.e., includes, for example, any one of light-chain CDR1, light-chain CDR2, and light-chain CDR3,
20 preferably includes at least light-chain CDR3, and preferably includes light-chain CDR1, light-chain CDR2, and light-chain CDR3 in the stated order from the N-terminus of the light chain variable region.

[0070] Regions other than CDR1 to CDR3 in each of the heavy chain
25 variable region and the light chain variable region are sometimes referred to as FRs. More specifically, a region between the N-terminus and the CDR1 is called FR1, a region between the CDR1 and the CDR2 is called FR2, a region between the CDR2 and the CDR3 is called FR3, and a region between the CDR3 and the
30 carboxy-terminus (C-terminus) is called FR4, and the names are designated for each of the heavy chain variable region and the light chain variable region.

[0071] The amino acid sequences of the heavy-chain CDR1 to CDR3 and the light-chain CDR1 to CDR3 are not particularly limited.
35 Examples thereof include heavy-chain CDR1 having the amino acid

sequence set forth in SEQ ID NO: 1, heavy-chain CDR2 having the amino acid sequence set forth in SEQ ID NO: 2, heavy-chain CDR3 having the amino acid sequence set forth in SEQ ID NO: 3, light-chain CDR1 having the amino acid sequence set forth in SEQ ID NO: 6, light-chain CDR2 having the amino acid sequence set forth in SEQ ID NO: 7, and light-chain CDR3 having the amino acid sequence set forth in SEQ ID NO: 8 serving as heavy-chain CDRs 1 to 3 or light-chain CDRs 1 to 3 of the MMG49 antibody.

[0072] As a preferred embodiment of the heavy chain variable region including the heavy-chain CDR1 to CDR3, there may be given, for example, a heavy chain variable region having the amino acid sequence set forth in SEQ ID NO: 4, which is a heavy chain variable region of the MMG49 antibody. In addition, as a preferred embodiment of the light chain variable region including the light-chain CDR1 to CDR3, there may be given, for example, a light chain variable region having the amino acid sequence set forth in SEQ ID NO: 9, which is a light chain variable region of the MMG49 antibody.

[0073] The above-mentioned amino acid sequences of the MMG49 antibody set forth in SEQ ID NOS: 1 to 4 and 6 to 9 are as shown in Table 1 below. Underlined parts in each of the amino acid sequences of the heavy chain and variable regions set forth in SEQ ID NOS: 4 and 9 in Table 1 indicate portions located at the CDR1, the CDR2, and the CDR3 in the stated order from the N-terminus.

[0074]

Table 1

<Amino acid sequences of MMG49 antibody>		
Heavy chain	CDR1 (SEQ ID NO: 1)	GYTFSSYW
	CDR2 (SEQ ID NO: 2)	MLPGSGSS
	CDR3 (SEQ ID NO: 3)	ARGDGNWYFDV
	Variable region (SEQ ID NO: 4)	MEWTWVFLFLLSVTAGVHSQVQLQQSGAELMKPGASVK ISCKASGYTFSSYWIEWVKQRPBGHGLEWIGEMLP SGS <u>SNYNEKFKGKATFTADTSSNTAYMQLSSLTSEDSAVYY</u> <u>CARGDGNWYFDVWGAG</u>
Light chain	CDR1 (SEQ ID NO: 6)	SSVGY
	CDR2 (SEQ ID NO: 7)	ATS
	CDR3 (SEQ ID NO: 8)	QQWSSDPPT

	Variable region (SEQ ID NO: 9)	MDFQVQIFSFLLISASVIMSRGQIVLSQSPAILLSASPG EKVTMTCRASSSVGYMHWFQQKPGSSPKPWIYATSNLA SGVPARFSGSESGTSYSLTISRVEAEDAATYYCQWSS DPPTFGGGTKLEIK
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[0075] The structure of the antibody (I) is not limited. Specific examples of the structure include Fv, scFv, a diabody, a triabody, and a tetrabody, and the structure may also be a structure
5 obtained by appropriately combining these structures. In addition, those structures including the combined structures as well are each sometimes also called a fragment antibody. Such fragment antibody may be an artificially designed recombinant protein including Fv, or may be one fused with a biomolecule, such as a
10 protein.

[0076] The Fv is also called the smallest structural unit of an antibody, and is a structure in which a heavy chain variable region and a light chain variable region are associated with each other through a non-covalent intermolecular interaction. Further,
15 the Fv may be a structure in which the thiol groups of cysteine residues present in the heavy chain variable region and the light chain variable region form a disulfide bond with each other.

[0077] The scFv is a structure in which the C-terminus of a heavy chain variable region and the N-terminus of a light chain
20 variable region are linked through a linker, and is also called a single-chain antibody. In addition, the C-terminus and N-terminus to be linked through a linker may be the C-terminus of the light chain variable region and the N-terminus of the heavy chain variable region. The structure of the scFv may be formed by
25 association based on a non-covalent intermolecular interaction or the like as in the Fv.

[0078] The diabody, the triabody, and the tetrabody are structures in which the above-mentioned scFv forms a dimer, a trimer, and a tetramer, respectively, and is associated in the structurally
30 most stable state through a non-covalent intermolecular interaction or the like between variable regions as in the Fv or the like.

[0079] A person skilled in the art can easily produce the antibody (I) having any of such various structures by: constructing an expression vector through use of commonly used genetic engineering means; and using, with such expression vector, an expression system adopting host cells suited for antibody production, such as prokaryotic cells (such as *Escherichia coli* or actinomycetes) or eukaryotic cells (such as yeast cells, insect cells, or mammalian cells), a commonly used cell-free expression system, or the like. The produced antibody may be appropriately subjected to a commonly used purification process so as to be obtained in a high-purity state.

[0080] As another embodiment of the antibody (I), the antibody (I) may contain a constant region. The constant region is understood by a person skilled in the art to be as follows: a heavy chain constant region includes CH1, CH2, and CH3, and a light chain constant region includes CL. In addition, a region including CH2 and CH3 is sometimes called an Fc domain.

[0081] A specific origin of the constant region is not particularly limited. Examples thereof may include constant regions originating from animal species capable of mass production, animal species closely related to humans, animal species that are less liable to cause immunogenicity in administration to a human, and the like, e.g., constant regions of human origin, mouse origin, rat origin, rabbit origin, monkey origin, and chimpanzee origin.

[0082] In the antibody (I), when the heavy chain variable region and/or the light chain variable region have an amino acid sequence of mouse origin, for example, a constant region of human origin may be combined therewith, to thereby provide the antibody (I) as a chimeric antibody.

[0083] In addition, heavy-chain FR1 to FR4 and/or light-chain FR1 to FR4 in the above-mentioned chimeric antibody may be replaced with amino acid sequences of human origin, to thereby provide the antibody (I) as a humanized antibody.

[0084] Further, heavy-chain CDR1 to CDR3 and/or light-chain CDR1

to CDR3 in the above-mentioned humanized antibody may be replaced with amino acid sequences of human origin to the extent that the functions of the CDRs are not reduced, to thereby provide the antibody (I) as a human antibody. The term "human antibody" is sometimes called a "completely humanized antibody".

[0085] Examples of the structure of the antibody (I) of the embodiment including a constant region may include structures such as Fab, $F(ab')_2$, a minibody, and scFv-Fc, as well as an immunoglobulin having a four-chain structure including a pair of heavy chains each having a heavy chain variable region and a heavy chain constant region, and a pair of light chains each having a light chain variable region and a light chain constant region. Further, a structure obtained by appropriately combining those structures may also be adopted. In addition, those structures including combined structures are each sometimes called a fragment antibody. Such fragment antibody may be an artificially designed recombinant protein including Fv, or may be one fused with a biomolecule, such as a protein.

[0086] The Fab includes a heavy chain fragment including a heavy chain variable region and CH1 in a heavy chain constant region, and a light chain including a light chain variable region and a light chain constant region, and has a structure in which the heavy chain variable region and the light chain variable region are associated with each other through the above-mentioned non-covalent intermolecular interaction, or are bonded to each other through a disulfide bond. Further, the Fab may be such that the CH1 and the CL form a disulfide bond between the thiol groups of cysteine residues respectively present therein.

[0087] The $F(ab')_2$ has a pair of the Fabs, and has a structure in which the CH1s form a disulfide bond between the thiol groups of cysteine residues respectively included therein.

[0088] The minibody has a pair of antibody fragments each including the scFv and CH3, and has a structure in which such antibody fragments are associated with each other through a non-covalent intermolecular interaction between the CH3s.

[0089] The scFv-Fc has a pair of antibody fragments each including the scFv, CH₂, and CH₃, and has a structure in which, as in the minibody, the antibody fragments are associated with each other through a non-covalent intermolecular interaction between the CH₃s, and form a disulfide bond between the thiol groups of cysteine residues included in the respective CH₃s.

[0090] A person skilled in the art can easily produce the antibody (I) including a constant region having any of such various structures as with the antibody (I) including no constant region, by constructing an expression vector through use of commonly used genetic engineering means, and using, with such expression vector, an expression system adopting host cells suited for antibody production. The produced antibody may be appropriately subjected to a commonly used purification process so as to be obtained in a high-purity state.

[0091] The Fab may be obtained by, for example, digesting an immunoglobulin IgG with a protease such as papain. In addition, F(ab')₂, the F(ab')₂ may be obtained by digesting IgG with a protease such as pepsin.

[0092] Of the above-mentioned antibodies (I) each including a constant region, a preferred structure is an immunoglobulin. The subtype of such immunoglobulin is not particularly limited, and examples thereof may include IgA, IgD, IgE, IgG, and IgM. Of those, IgG is preferred, and for example, in the case of IgG of mouse origin, IgG₂ is preferred out of the four subclasses.

[0093] Of the above-mentioned antibodies (I) each including a constant region, an antibody of a more preferred embodiment is an antibody including a heavy chain having the amino acid sequence set forth in SEQ ID NO: 5 and/or a light chain having the amino acid sequence set forth in SEQ ID NO: 10. The most preferred antibody is an antibody including a heavy chain having the amino acid sequence set forth in SEQ ID NO: 5 and a light chain having the amino acid sequence set forth in SEQ ID NO: 10.

[0094] A mutation may be introduced into each of the above-mentioned amino acid sequences depending on the situation. It is

preferred that such mutation be not introduced into heavy-chain CDRs and light-chain CDRs. That is, the mutation is preferably introduced into a heavy-chain FR or a light-chain FR. When the antibody (I) includes a constant region, a mutation may be

5 further introduced in addition to a mutation for adjusting ADCC activity or CDC activity to be described below.

[0095] A specific number of amino acid residues at which mutations are introduced is not particularly limited. For example, identity between an amino acid sequence before mutation introduction and
10 an amino acid sequence after mutation introduction is about 70%, preferably about 75%, more preferably about 80%, more preferably about 85%, more preferably about 90%, more preferably about 95%, more preferably about 96%, more preferably about 97%, more preferably about 98%, most preferably about 99%. Such numerical
15 value is one obtained by rounding.

[0096] The term "identity" refers to the degree of amino acid sequences identical to each other in two or more comparable amino acid sequences. Therefore, as the identity between given two amino acid sequences increases, it can be said that those
20 sequences have not only higher identity but also higher similarity.

[0097] The identity of amino acids may be calculated using an analytical tool that is commercially available or available through the Internet (e.g., software such as FASTA, BLAST, PSI-
25 BLAST, or SSEARCH). For example, main initial conditions to be generally used for a BLAST search are as described below. That is, a value (%) for the identity between amino acid sequences may be calculated by performing a search on Advanced BLAST 2.1 with blastp being used as a program, the Expect value being set to 10, all Filters being turned OFF, BLOSUM62 being used for Matrix, the
30 Gap existence cost, Per residue gap cost, and Lambda ratio being set to 11, 1, and 0.85 (default values), respectively, and the other various parameters being also set to default values.

[0098] The above-mentioned introduction of a mutation into an
35 amino acid sequence refers to substitution, deletion, insertion,

or the like. Specific mutation introduction is not particularly limited as long as the mutation introduction can be achieved by adopting a commonly used method. For example, in the case of the substitution, a conservative substitution technology may be adopted.

[0099] The term "conservative substitution technology" means a technology involving substituting a certain amino acid residue with an amino acid residue having a side chain similar thereto.

[0100] For example, substitution between amino acid residues each having a basic side chain, such as lysine, arginine, and histidine, is a conservative substitution technology. In addition, each of substitutions between: amino acid residues each having an acidic side chain, such as aspartic acid and glutamic acid; amino acid residues each having an uncharged polar side chain, such as glycine, asparagine, glutamine, serine, threonine, tyrosine, and cysteine; amino acid residues each having a non-polar side chain, such as alanine, valine, leucine, isoleucine, proline, phenylalanine, methionine, and tryptophan; amino acid residues each having a β -branched side chain, such as threonine, valine, and isoleucine; and amino acid residues each having an aromatic side chain, such as tyrosine, phenylalanine, tryptophan, and histidine, is similarly a conservative substitution technology.

[0101] As another embodiment of the antibody (I), the antibody (I) may have cytotoxic activity. The cytotoxic activity refers to such activity that the antibody binds to cells, and as a result, causes some damage to the cells bound to the antibody.

[0102] Examples of such cytotoxic activity include ADCC activity and CDC activity. The term "ADCC activity" is an abbreviation of antibody-dependent cytotoxic activity (Antibody-Dependent Cellular Cytotoxicity), and is activity of recruiting cells having cytotoxic activity, such as NK cells expressing a receptor specific to a constant region of an antibody, to the vicinity of the antibody, to thereby induce damage to cells to which the antibody binds through the action of such cells and the like.

[0103] The term "CDC activity" is an abbreviation of complement-

dependent cytotoxic activity (Complement-Dependent Cytotoxicity), and refers to activity that the antibody recruits a complement to its vicinity, to thereby induce an action of causing damage to cells bound to the antibody through the action of such complement.

5 [0104] Here, each of the ADCC activity and the CDC activity may be adjusted by introducing a mutation into a constant region while appropriately referring to the literature, such as Lazar GA et al., Proc Natl Acad Sci USA, 103: 4005-10 (2006), Shields RL et al., J Biol Chem, 276: 6591-604 (2001)), Moore GL et al., J
10 Immunol, 159:3613-21 (1997), An Z et al., MAbs, 1:572-9 (2009).

[0105] For example, when the constant region is human IgG₁, the ADCC activity may be increased by introducing a mutation such as S239D, I332E, S239D/I332E, S239D/I332E/A330L, S298A, K334A, S298A/K334A, or S298A/E333A/K334A.

15 [0106] In addition, when the constant region is human IgG₁ as in the foregoing, the ADCC activity may be decreased by introducing a mutation such as V234A/G237A, or H268Q/V309L/A330S/P331S.

[0107] With regard to the CDC activity, when the constant region is human IgG₁, the activity may be increased by introducing a
20 mutation such as S267E, H268F, S324T, S267E/H268F, S267E/S324T, H268F/S324T, or S267E/H268F/S324T.

[0108] The ADCC activity may be measured in accordance with Brunner K.T., et al.'s method (Brunner, K.T., et al., Immunology, 1968. 14:181-96). For example, myeloma cells are cultured in
25 RPMI1640 medium supplemented with 10% FCS, and are prepared so that the number of cells may be from 0.5×10^4 to 1.0×10^4 . An appropriate amount of Na₂⁵¹CrO₄ is added thereto and allowed to react therewith at 37°C for 1 hour to label the cells with ⁵¹Cr, and the resultant cells are washed and then used as target cells.

30 As effector cells, ones obtained by culturing SCID mouse bone marrow cells for 6 days in RPMI1640 supplemented with 10% FBS, 10 ng/ml mouse GM-CSF, and 40 IU/ml human IL2, or the like may be used. To a 96-well plate, an antibody to be tested or an isotype antibody thereof serving as a control is added at a final
35 concentration from 0.05 µg/mL to 10 µg/mL, and the target cells

(1.0×10^4 cells) and the effector cells (5×10^5 cells) are further added. The mixture is subjected to a reaction at 37°C for 4 hours and centrifuged, and then ^{51}Cr released into the supernatant is measured with a γ -counter. The ADCC activity may be determined on the basis of the following equation.

$$\begin{aligned} \text{[0109] ADCC activity} = & \{ ([^{51}\text{Cr release from target cells}] - \\ & [\text{spontaneous } ^{51}\text{Cr release under antibody-free state}]) / ([\text{maximum} \\ & ^{51}\text{Cr release amount caused by 1\% Triton X-100 addition}] - \\ & [\text{spontaneous } ^{51}\text{Cr release under antibody-free state}]) \} \times 100 \end{aligned}$$

[0110] The CDC activity may also be measured in accordance with Brunner K.T., et al.'s method (Brunner, K.T., et al., Immunology, 1968. 14:181-96). For example, myeloma cells to be used as target cells are cultured in RPMI1640 medium supplemented with 10% FCS, and are prepared so that the number of cells may be from 0.5×10^4 to 1.0×10^4 . An appropriate amount of $\text{Na}_2^{51}\text{CrO}_4$ is added thereto and allowed to react therewith at 37°C for 1 hour to label the cells with ^{51}Cr , and the resultant cells are washed and then used as target cells. An antibody to be tested or an isotype antibody serving as a control suspended in RPMI1640 medium supplemented with fetal bovine serum is added to a 96-well plate at a final concentration of from 0.5 $\mu\text{g/mL}$ to 50 $\mu\text{g/mL}$, and then the target cells and a complement are added, followed by a reaction for 1.5 hours. The reaction liquid is centrifuged, and ^{51}Cr released into the supernatant is measured with a γ -counter. The CDC activity may be determined on the basis of the following equation.

$$\begin{aligned} \text{[0111] CDC activity} = & \{ ([^{51}\text{Cr release from target cells}] - [\text{spontaneous} \\ & ^{51}\text{Cr release under antibody-free state}]) / ([\text{maximum } ^{51}\text{Cr release} \\ & \text{amount caused by 1\% Triton X-100 addition}] - [\text{spontaneous } ^{51}\text{Cr} \\ & \text{release under antibody-free state}]) \} \times 100 \end{aligned}$$

The antibody having cytotoxic activity may be obtained by, for example, evaluating the presence or absence of cytotoxic activity through use of the above-mentioned method, and selecting an antibody having the activity.

[0112] As another embodiment of the antibody (I), the antibody (I) may be a multispecific antibody. That is, the antibody (I) may

have binding capacity with specificity to an antigen other than the region of the amino acid residue positions 20 to 109 of the human integrin β_7 (the antigen is hereinafter referred to as other antigen).

5 [0113] The other antigen is preferably an antigen structurally dissimilar to the region of the amino acid residue positions 20 to 109 of the human integrin β_7 .

[0114] A specific other antigen is not particularly limited. Examples thereof include CD3, CD16, C1q, and Adenovirus knob
10 domain, and at least one of those antigens in appropriate combination may be appropriately adopted as the other antigen. It is preferred that one of the antigens given as examples above be selected as the other antigen. That is, a preferred multispecific antibody is a bispecific antibody.

15 [0115] A person skilled in the art can easily produce such multispecific antibody by appropriately adopting a commonly used technology. For example, the multispecific antibody may be obtained in the following manner: hybridomas generated using antibody-producing cells, such as B-cells, obtained from an
20 animal immunized with cells expressing a peptide fragment corresponding to the region of the amino acid residue positions 20 to 109 of the human integrin β_7 , or chimeric integrin β_7 in which only the region of the amino acid residue positions 20 to 109 of the human integrin β_7 is of human origin and the rest is of
25 non-human origin, such as mouse origin, are prepared; separately, hybridomas are generated using antibody-producing cells, such as B-cells, obtained from an animal immunized with the above-mentioned other antigen; and screening is performed, by a commonly used method, for new hybridomas obtained by cell fusion
30 between the above-mentioned hybridomas (the new hybridomas are also referred to as quadromas in the case of producing a bispecific antibody).

[0116] In addition to the foregoing, for example, in the case of a bispecific antibody, the bispecific antibody may be generated by
35 a procedure described in the following (1) to (4):

(1) An antibody having the structure of the above-mentioned $F(ab')_2$, which uses the region of the amino acid residue positions 20 to 109 of the human integrin β_7 as an epitope, is generated;

5 (2) Meanwhile, an antibody having the structure of $F(ab')_2$ that specifically binds to the other antigen is also similarly generated;

(3) The antibodies of the respective $F(ab')_2$ structures obtained in (1) and (2) are treated with a reducing agent, such as DTT,
10 and then any one of the treated products is further treated with Ellman's reagent; and

(4) The treated antibodies of the $F(ab')_2$ structures obtained in (3) are mixed and allowed to react with each other.

[0117] The bispecific antibody may also be produced by a procedure
15 described in the following (A) to (D).

(A) An antibody using the region of the amino acid residue positions 20 to 109 of the human integrin β_7 as an epitope is generated.

(B) Meanwhile, an antibody that specifically binds to the other
20 antigen is similarly generated.

(C) The amino acid sequences of respective variable regions obtained in (A) and (B) and the base sequences of polynucleotides encoding the variable regions are identified.

(D) An expression vector having incorporated therein
25 polynucleotides having the respective base sequences identified in (C) together with, as necessary, polynucleotides having a base sequence for a constant domain and a linker sequence is generated, and then the expression vector is introduced into host cells suited for antibody production, such as CHO cells.

30 [0118] As another embodiment of the antibody (I), the antibody (I) may have bound thereto a cytotoxin (substance having cytotoxic activity). The cytotoxin is not particularly limited as long as the cytotoxin is a substance that causes some damage to cells, such as the killing of cells or the inhibition of cell growth.

35 [0119] Examples of such cytotoxin may include: alkylating agents,

such as cyclophosphamide hydrate, iphosphamide, thiotepa, busulfan, melphalan, nimustine hydrochloride, ranimustine, dacarbazine, and temozolomide; metabolic antagonists, such as methotrexate, pemetrexed sodium hydrate, fluorouracil, 5 doxifluridine, capecitabine, tegafur, cytarabine, gemcitabine hydrochloride, fludarabine phosphate, nelarabine, cladribine, and calcium levofolinate; antibiotic substances, such as doxorubicin hydrochloride, daunorubicin hydrochloride, pirarubicin, epirubicin hydrochloride, idarubicin hydrochloride, aclarubicin 10 hydrochloride, amrubicin hydrochloride, mitoxantrone hydrochloride, mitomycin C, actinomycin D, bleomycin hydrochloride, peplomycin hydrochloride, zinostatin stimalamer, and calicheamicin; microtubule inhibitors, such as vincristine sulfate, vinblastine sulfate, vindesine sulfate, and paclitaxel; 15 aromatase inhibitors, such as anastrozole, exemestane, letrozole, and fadrozole hydrochloride hydrate; platinating agents, such as cisplatin, carboplatin, nedaplatin, and oxaliplatin; topoisomerase inhibitors, such as irinotecan hydrochloride hydrate, nogitecan hydrochloride, etoposide, and sobuzoxane; 20 adrenal cortex steroids, such as prednisolone and dexamethasone; thalidomide and a derivative thereof, specifically lenalidomide; bortezomib serving as a protease inhibitor; and radioactive isotopes, such as 90-Ittrium.

[0120] Of those, calicheamicin, melphalan, vincristine sulfate, 25 doxorubicin hydrochloride, prednisolone, dexamethasone, thalidomide, lenalidomide, and bortezomib are preferred, and calicheamicin having proven excellence in binding to an antibody is more preferred.

[0121] Each of those cytotoxins is commercially available, and one 30 kind or two or more kinds in appropriate combination may be selected from the above-mentioned cytotoxins.

[0122] The manner of binding between the cytotoxin and the above-mentioned antibody is not particularly limited, and a person skilled in the art can easily bind the cytotoxin to the above- 35 mentioned antibody by, for example, appropriately adopting a

commonly used genetic engineering technology or protein engineering technology. More specifically, there may be given, for example, a method involving binding the cytotoxin to a functional group, such as an amino group, a thiol group, a
5 guanidyl group, a hydroxy group, or a carboxyl group, of an amino acid residue side chain of the antibody (I) via a linker.

[0123] The antibody (I) may be a polyclonal antibody or may be a monoclonal antibody. The antibody (I) is preferably a monoclonal antibody.

10 [0124] The term "monoclonal" means being obtained from a substantially uniform population, and the "monoclonal antibody" means an antibody obtained from such population. That is, it is understood that individual antibodies included in such population are identical to each other except for a naturally occurring
15 mutation that may be present in a minute amount.

[0125] Further, with regard to a specific binding target (epitope) of an antibody, for example, in the case of the antibody (I), the epitope is present in the region of the amino acid residue positions 20 to 109 of the human integrin β_7 . In this regard, in
20 the case of a polyclonal antibody, the epitope is a plurality of sites in the region of the amino acid residue positions 20 to 109 of the human integrin β_7 , whereas in the case of a monoclonal antibody, the epitope is a single site. For this reason, the monoclonal antibody exhibits high specificity, and hence is more
25 advantageous.

[0126] The term "monoclonal" to be used as a modifier is understood to mean being obtained from a substantially uniform population as described above, and should not be understood as a modifier for specifying a production method for the antibody.

30 [0127] Other than the above-mentioned method, a person skilled in the art can easily produce the antibody (I) by adopting a hybridoma method, a recombinant DNA method using a host cell (III) harboring a polynucleotide (II) to be described below, isolation from a phage library, or the like.

35 [0128] For example, there may be given a method involving:

immunizing an animal suited for antibody production, such as a mouse, a rat, or a rabbit, with a peptide corresponding to the region of the amino acid residue positions 20 to 109 of the human integrin β_7 ; then recovering B-cells and subjecting the recovered
5 B-cells to a hybridoma method; and performing screening through use of the function exhibited by the antibody (I) described above as an indicator, to thereby produce the antibody (I).

[0129] In addition to the foregoing, there may be given a method involving: generating cells expressing chimeric integrin β_7 in
10 which only the region of the amino acid residue positions 20 to 109 of integrin β_7 is of human origin and the rest is of non-human origin, such as mouse origin; immunizing an animal suited for antibody production, such as a mouse, a rat, or a rabbit (preferably a mouse), with the cells; then recovering B-cells and
15 subjecting the recovered B-cells to a hybridoma method; and performing screening through use of the function exhibited by the antibody (I) described above as an indicator, to thereby produce the antibody (I).

[0130] Examples of the function exhibited by the antibody (I) may
20 include: an increase in affinity for the region of the amino acid residue positions 20 to 109 of the human integrin β_7 in the presence of at least part of the region of the amino acid residue positions 380 to 721 of the human integrin β_7 ; and an increase in the affinity through activation of the human integrin β_7 .

25 Accordingly, the antibody (I) may be obtained by a method shown in a screening method (X) to be described below, which utilizes such function.

[0131] An epitope for the antibody (I) is present in the region of the amino acid residue positions 20 to 109 of the human
30 integrin β_7 , and hence the antibody (I) is expected to exhibit cytotoxic activity or the like on cells expressing the integrin β_7 by combining one or two or more of not only the above-mentioned ADCC activity and CDC activity, but also apoptosis-inducing activity, survival signal-blocking activity, and the like on such
35 cells. Accordingly, a composition including the antibody (I) is

useful as a pharmaceutical composition (VII) as described in detail below.

[0132] In particular, the antibody (I) uses the region of the amino acid residue positions 20 to 109 of the human integrin β_7 as an epitope, and the affinity of the antibody (I) for the epitope is increased through activation of the integrin β_7 . Active-form integrin β_7 is expressed in blood cells, such as plasma cells, and hence the antibody (I) is used as an active ingredient of a pharmaceutical composition for cancer thereof (e.g., blood cancer). In particular, the antibody (I) is effectively used as a pharmaceutical composition for a disease causing an abnormality in the above-mentioned cells (e.g., myeloma or multiple myeloma).

[0133] (II) Polynucleotide

A polynucleotide (II) is a polynucleotide having a base sequence encoding the amino acid sequence of the antibody (I). The term "polynucleotide" includes, for example, a single-stranded or double-stranded form in which a ribonucleotide, a deoxyribonucleotide, any one nucleotide thereof, or the like is appropriately modified by a known method.

[0134] A person skilled in the art can appropriately determine the base sequence of the polynucleotide (II), for example, in silico on the basis of the amino acid sequence of the antibody (I). The kinds of codons to be used for determining such base sequence are not limited. The base sequence is preferably determined in consideration of the codon frequency of a host in which the polynucleotide is to be used.

[0135] A specific base sequence of the polynucleotide (II) is not particularly limited. Correspondence between each SEQ ID NO in which an amino acid sequence identified as one embodiment of the antibody (I) is set forth and the SEQ ID NO in which a base sequence encoding such amino acid sequence is set forth is shown in Table 2 below. That is, preferred base sequences of the polynucleotide (II) are base sequences set forth in SEQ ID NOS: 11 to 20.

[0136]

Table 2

Amino acid sequence	Base sequence
SEQ ID NO: 1	SEQ ID NO: 11
SEQ ID NO: 2	SEQ ID NO: 12
SEQ ID NO: 3	SEQ ID NO: 13
SEQ ID NO: 4	SEQ ID NO: 14
SEQ ID NO: 5	SEQ ID NO: 15
SEQ ID NO: 6	SEQ ID NO: 16
SEQ ID NO: 7	SEQ ID NO: 17
SEQ ID NO: 8	SEQ ID NO: 18
SEQ ID NO: 9	SEQ ID NO: 19
SEQ ID NO: 10	SEQ ID NO: 20

[0137] The polynucleotide (II) may adopt an embodiment of being incorporated into a vector. The vector is not particularly

5 limited, and may be, for example, a vector for cloning or a vector for expression, and its intended use is not limited.

[0138] In addition, the vector for expression may be a vector for prokaryotic cells, such as *Escherichia coli* or actinomycetes, or may be a vector for eukaryotic cells, such as yeast cells, insect
10 cells, or mammalian cells.

[0139] A base sequence encoding a signal peptide may be appropriately added to the 5'-end side of the polynucleotide (II) (corresponding to the N-terminus side of the antibody (I)).

[0140] A specific method of using the polynucleotide (II) is not
15 particularly limited. For example, the polynucleotide (II) may be used for expressing the antibody (I) by being introduced into the following host cell (III).

[0141] (III) Host Cell

A host cell (III) is a cell harboring the polynucleotide
20 (II). The term "harbor" refers to the keeping of a state in which the polynucleotide (II) is present in the cell, and means that the cell is in a state of not spontaneously discharging the polynucleotide to the outside of the cell irrespective of whether the discharge is active.

25 [0142] The embodiment in which the host cell (III) harbors the polynucleotide (II) is not particularly limited. For example, the polynucleotide may be harbored in the form of a vector in the

cell, or the polynucleotide (II) may be harbored in the form of being integrated into the genome in the cell.

[0143] A specific cell kind of the host cell (III) may be a eukaryotic cell, such as a yeast cell, an insect cell, or a mammalian cell, or may be a prokaryotic cell, such as *Escherichia coli* or an actinomycete, and is not particularly limited.

[0144] (IV) Chimeric Antigen Receptor

A chimeric antigen receptor is an artificial T-cell receptor (TCR)-like protein, and is a protein in which an antigen recognition site expressed on the cell membrane of T-cells (corresponding to an extracellular domain) is replaced with a desired antigen recognition site and which is constructed so as to be able to more effectively exhibit a function, such as cytotoxic activity, of T-cells themselves.

[0145] The chimeric antigen receptor (IV) is a chimeric antigen receptor whose epitope is identical to that of the antibody (I), and more specifically, is a protein including an antigen recognition site of the antibody (I). That is, the epitope present in the antigen recognition site included in the chimeric antigen receptor may be the same as the one described in detail in the antibody (I).

[0146] More specifically, the chimeric antigen receptor (IV) is a protein in which the antigen recognition site of the antibody (I), a spacer sequence, a transmembrane domain, a costimulator, and an TCR intracellular domain are arranged in the stated order from the N-terminus of the chimeric antigen receptor (IV).

[0147] The antigen recognition site of the antibody (I) to be arranged in the chimeric antigen receptor (IV) may be as described in detail in the antibody (I), and specific examples thereof may include a heavy chain variable region and/or a light chain variable region. Of those, the antigen recognition site preferably has the structure of scFv while having a heavy chain variable region and a light chain variable region.

[0148] In such scFv, for example, between the heavy chain variable region and the light chain variable region, a spacer sequence

consisting of about 10 to about 25 amino acid residues may be appropriately arranged. The number of amino acid residues is more preferably from about 15 to about 18. Such spacer sequence may be identical to the above-mentioned spacer sequence to be arranged
5 in the chimeric antigen receptor (IV), or may be different therefrom.

[0149] The spacer sequence to be arranged in the chimeric antigen receptor (IV) is not particularly limited. For example, the spacer sequence may be consisting of about 10 to about 25 amino
10 acid residues. The number of amino acid residues is more preferably from about 15 to about 18.

[0150] The transmembrane domain to be arranged in the chimeric antigen receptor (IV) is not particularly limited. Specifically, a cell transmembrane domain derived from a protein, such as CD28
15 or 4-1BB, expressed in T-cells or the like may be adopted while being allowed to appropriately have a mutation introduced thereinto.

[0151] The costimulator to be arranged in the chimeric antigen receptor (IV) may be a costimulator of T-cells or the like, and
20 is not particularly limited. For example, 4-1BB, OX40, CD28, or the like may be adopted while being allowed to appropriately have a mutation introduced thereinto.

[0152] The TCR intracellular domain to be arranged in the chimeric antigen receptor (IV) is not particularly limited. For example,
25 an intracellular domain derived from CD3, which is also called a TCR ζ chain, or the like may be adopted while being allowed to appropriately have a mutation introduced thereinto. It is preferred that a mutation be introduced into CD3 so as to include an Immunoreceptor Tyrosine-based Activation Motif (ITAM).

30 [0153] The chimeric antigen receptor (IV) preferably has the amino acid sequence set forth in SEQ ID NO: 21.

[0154] A mutation may be appropriately introduced into an amino acid sequence identifying the above-mentioned chimeric antigen receptor. In addition, a mutation may also be similarly
35 introduced into each of the above-mentioned transmembrane domain,

costimulator, and TCR intracellular domain. A specific number of mutations to be introduced is not particularly limited.

[0155] For example, identity between an amino acid sequence before mutation introduction and an amino acid sequence after mutation introduction is about 70%, preferably about 75%, more preferably about 80%, more preferably about 85%, more preferably about 90%, more preferably about 95%, more preferably about 96%, more preferably about 97%, more preferably about 98%, most preferably about 99%. Such numerical value is one obtained by rounding.

[0156] The above-mentioned introduction of a mutation into an amino acid sequence refers to substitution, deletion, insertion, or the like. Specific mutation introduction is not particularly limited as long as the mutation introduction can be achieved by adopting a commonly used method. For example, in the case of the substitution, a conservative substitution technology may be adopted.

[0157] For the production of such chimeric antigen receptor, a person skilled in the art can easily produce the chimeric antigen receptor with reference to a method described in, for example, each of Non Patent Literatures 4 to 6.

[0158] (V) Polynucleotide

A polynucleotide (V) is a polynucleotide encoding the amino acid sequence of the chimeric antigen receptor (IV) unlike the polynucleotide (II).

[0159] As in the polynucleotide (II), the base sequence of the polynucleotide (V) may be appropriately determined, for example, in silico on the basis of the amino acid sequence of the chimeric antigen receptor (IV). The kinds of codons to be used for determining the base sequence are not limited. The base sequence is preferably determined in consideration of the codon frequency of cells serving as a target for which the polynucleotide is to be used.

[0160] A specific base sequence is not particularly limited. For example, there may be given a polynucleotide having the base sequence set forth in SEQ ID NO: 22, which is determined on the

basis of the amino acid sequence of the chimeric antigen receptor (IV) having the amino acid sequence set forth in SEQ ID NO: 21.

Of course, in consideration of the kinds of the codons to be used being not limited, needless to say, the base sequence to be

5 determined on the basis of such amino acid sequence is not limited to the base sequence set forth in SEQ ID NO: 22.

[0161] A base sequence encoding a signal peptide may be appropriately added to the 5'-end side of the polynucleotide (V) (corresponding to the N-terminus side of the chimeric antigen
10 receptor (IV)).

[0162] A specific method of using the polynucleotide (V) is not particularly limited. For example, the polynucleotide (V) may be used for expressing the chimeric antigen receptor (IV) by being introduced into the following cell (VI).

15 [0163] (VI) Cell

A cell (VI) is a cell harboring the polynucleotide (V) unlike the host cell (III). The term "harbor" may be the same as in the host cell (III). A specific kind of the cell may also be the same as in the host cell (III), but the cell (VI) preferably
20 has cytotoxic activity. Examples thereof may include a T-cell, an NK cell, and a K cell. Of those, a killer T-cell (sometimes referred to as cytotoxic T-cell [CTL]), which is a kind of T-cell, is most preferred.

[0164] It is preferred that, when the polynucleotide (V), which
25 encodes the chimeric antigen receptor, included in the cell (VI) is expressed, the antigen recognition site of the antibody (I) serving as a component of the chimeric antigen receptor (IV) be exposed to the outside of the cell, and the transmembrane domain, the costimulator, or the TCR intracellular domain serving as a
30 component of the chimeric antigen receptor (IV) be localized on a cell membrane or inside the cell.

[0165] The costimulator, or the domain to be localized on the cell membrane or inside the cell activates a signal that induces cytotoxic activity in cells when the antigen recognition site of
35 the antibody (I) binds to the region of the amino acid residue

positions 20 to 109 of the human integrin β_7 . In addition, the affinity of the antibody (I) for the region of the amino acid residue positions 20 to 109 of the human integrin β_7 is increased through activation of the human integrin β_7 . Therefore, the antibody (I) attacks, or exhibits cytotoxic activity on, a cell or tissue expressing active-form integrin β_7 serving as a target. [0166] When the cell exhibiting such function is a T-cell, the cell is referred to as chimeric antigen receptor T-cell (VI-4). Like the chimeric antigen receptor T-cell, a cell having the possibility of exhibiting cytotoxic activity, such as an NK cell, can also exhibit an effect similar to that of the chimeric antigen receptor T-cell (such cell is referred to as chimeric antigen receptor NK cell) through cooperation between the binding of the antigen recognition site to the region of the amino acid residue positions 20 to 109 of active-form human integrin β_7 , and the activation of a signal that induces cytotoxic activity at the cell membrane or the intracellular domain. [0167] As described above, the cell (VI) exhibits cytotoxic activity and the like on a cell or tissue expressing active-form integrin β_7 . Accordingly, as with the antibody (I), a composition including the cell (VI) may be said to be useful as such a pharmaceutical composition (IV) as described in detail below. The active-form integrin β_7 is expressed in blood cells, such as plasma cells, and hence the cell (VI) is used as an active ingredient of a pharmaceutical composition for cancer (e.g., blood cancer). In particular, the cell (VI) is effectively used as a pharmaceutical composition for a disease causing an abnormality in the above-mentioned cells (e.g., myeloma or multiple myeloma).

30 [0168] (VII) Pharmaceutical Composition

A pharmaceutical composition (VII) includes the antibody (I) or the cell (VI). The cell (VI) is preferably the chimeric antigen receptor T-cell (VI-4).

[0169] The content of the antibody (I) or the cell (VI) in the pharmaceutical composition (VII) is not particularly limited. For

example, in the case of the antibody (I), the content may be from about 0.001 part by weight to about 10 parts by weight with respect to 100 parts by weight of the pharmaceutical composition. In addition, in the case of the cell (VI), the content may be
5 from about 1 cell/mL to about 10^4 cells/mL.

[0170] An administration method for the pharmaceutical composition (VII) is not particularly limited. Because its active ingredient is an antibody or a cell, parenteral administration or non-enteral administration is preferred. Examples thereof include
10 intravenous administration, intramuscular administration, and subcutaneous administration. Of those, intravenous administration is preferred.

[0171] A dosage form of the pharmaceutical composition (VII) may be prepared together with a pharmacologically acceptable and
15 commonly used carrier depending on the above-mentioned administration method. In consideration of the above-mentioned preferred administration method, the dosage form is preferably an injection.

[0172] A target disease of the pharmaceutical composition (VII) is
20 not particularly limited. A specific example of the target disease is cancer, preferably blood cancer, more preferably a disease causing neoplastic growth of plasma cells. The term "disease causing neoplastic growth of plasma cells" refers to a disease characterized by neoplastic growth of abnormal plasma
25 cells and an increase in abnormal protein secreted therefrom. Examples of such disease may include myeloma, multiple myeloma, plasma cell leukemia, plasmacytoma, H chain disease, and systemic AL amyloidosis. In another embodiment, the target disease of the pharmaceutical composition (VII) may be a different hematologic
30 malignancy, such as malignant lymphoma or leukemia.

[0173] An administration target (subject) of the pharmaceutical composition (VII) may be a patient who has developed the above-mentioned disease, or may be an animal having a risk of developing the above-mentioned disease. Whether "having a risk of
35 developing" may be determined by a diagnosis method (XI) to be

described later. The animal may be, for example, a mammal, and is preferably a human.

[0174] The dose of the pharmaceutical composition (VII) varies depending on various conditions of the administration target, such as the severity of the disease, the degree of a desired effect achieved by the administration, body weight, sex, age, and animal species, and cannot be unconditionally determined. For example, when the active ingredient is the antibody (I), the dose may be generally from about 1 $\mu\text{g/kg}$ (body weight) to about 10 g/kg (body weight) per day. In addition, when the active ingredient is the cell (VI), the dose may be generally from about 10^4 cells/kg (body weight) to about 10^9 cells/kg (body weight).

[0175] As with its dose, the administration schedule of the pharmaceutical composition (VII) also varies depending on various conditions of the administration target, such as the severity of the disease, and cannot be unconditionally determined. For example, the pharmaceutical composition (VII) is preferably administered in the above-mentioned daily dose at a frequency of from once a day to once a month.

20 [0176] (VIII) Treatment or Prevention Method for Disease

A treatment or prevention method (VIII) for a disease is a treatment or prevention method for a disease including a step of administering a therapeutically effective amount of the antibody (I) or the cell (VI) to a subject. The cell (VI) is preferably the chimeric antigen receptor T-cell (VI-4).

[0177] The subject may be the same as in the pharmaceutical composition (VII). When the subject is a patient who has developed a disease, the administration of the therapeutically effective amount of the antibody (I) or the cell (VI) is expected to achieve a therapeutic effect thereon, and when the subject is an animal having a risk of developing a disease, the administration is expected to achieve a preventing effect thereon. As described in a diagnosis method (XI) to be described below, the preventing means keeping of a numerical value measured by a commonly used immunological method from reaching a numerical

value at which it is judged that a disease has developed.

[0178] The disease may be the same as in the pharmaceutical composition (VII), and is exemplified by, for example, cancer. A preferred example of the cancer may be a disease causing
5 neoplastic growth of plasma cells (e.g., multiple myeloma).

[0179] The therapeutically effective amount may be the same as the dose of the pharmaceutical composition (VII), and a formulation of the antibody (I) or the cell (VI) may be the same as the dosage form of the pharmaceutical composition (VII). In addition,
10 an administration method for the antibody (I) or the cell (VI), an administration schedule therefor, and the like may also be as described in detail in the pharmaceutical composition (VII).

[0180] The treatment or prevention method (VIII) for a disease may encompass a treatment or prevention method for multiple myeloma
15 including targeting active-form human integrin β_7 . An example of the targeting may be application of the antibody (I) or the cell (VI).

[0181] (IX) Use

A use (IX) is a use of the antibody (I) or the cell (IV),
20 for producing a pharmaceutical composition.

[0182] The pharmaceutical composition may be the same as the pharmaceutical composition (VII). The cell (IV) is preferably the chimeric antigen receptor T-cell (VI-4).

[0183] In addition, a target disease of the pharmaceutical
25 composition is also the same, and the pharmaceutical composition is used for treating, for example, cancer, preferably blood cancer, more preferably a disease causing neoplastic growth of plasma cells (e.g., myeloma or multiple myeloma).

[0184] Besides, the content of the antibody (I) or the cell (VI)
30 serving as an active ingredient in the pharmaceutical composition, the dosage form thereof, an administration method therefor, an administration schedule therefor, and the like may also be the same as those described in detail in the pharmaceutical composition (VII).

35 [0185] (X) Screening Method

A screening method (X) is a screening method for an active ingredient of a pharmaceutical composition for treating or preventing a disease causing neoplastic growth of plasma cells, including a step of selecting, from a compound library, a
5 candidate substance that specifically binds to human integrin β_7 and binds to a region of the amino acid residue positions 20 to 109 of the human integrin β_7 .

[0186] The pharmaceutical composition may be the same as the pharmaceutical composition (VII), and an example of an active
10 ingredient of the pharmaceutical composition for treating or preventing cancer, preferably blood cancer, more preferably a disease causing neoplastic growth of plasma cells (e.g., myeloma or multiple myeloma) is the antibody (I).

[0187] The compound library is not particularly limited, and an
15 existing library may be used. The compound library is preferably an antibody library, and the library preferably includes hybridomas generated using antibody-producing cells, such as B-cells, obtained from an animal immunized with a desired antigen.

[0188] Here, the desired antigen is not particularly limited, and
20 is preferably, for example, the region of the amino acid residue positions 20 to 109 of the human integrin β_7 . The desired antigen is more preferably active-form human integrin β_7 .

[0189] A method of selecting the candidate substance is not particularly limited. For example, there may be adopted means
25 involving: selecting a candidate substance that specifically binds to human integrin β_7 ; confirming that the candidate substance binds to a peptide fragment corresponding to the region of the amino acid residue positions 20 to 109 of the human integrin β_7 , through use of commonly used immunoassay means; and
30 selecting such candidate substance.

[0190] In addition, there may be adopted means involving:
selecting a candidate substance that specifically binds to the human integrin β_7 ; further confirming a candidate substance that binds to cells expressing the chimeric integrin β_7 in which only
35 the region of the amino acid residue positions 20 to 109 of

integrin β_7 is of human origin and the rest is of non-human origin, such as mouse origin, described in detail in the antibody (I), through use of commonly used immunoassay means; and selecting such candidate substance. Means involving selecting such

5 candidate substance may also be adopted.

[0191] In addition, there may also be adopted means involving: selecting a candidate substance that specifically binds to the human integrin β_7 ; further treating cells expressing the chimeric integrin β_7 in which only the region of the amino acid residue

10 positions 20 to 109 of integrin β_7 is of human origin and the rest is of non-human origin, such as mouse origin, described in detail in the antibody (I) with a phorbol ester, a manganese salt, or the like, and confirming a candidate substance showing an increased degree of binding after the treatment as compared to
15 that before the treatment; and selecting such candidate substance.

[0192] In addition, there may also be adopted means involving: selecting a candidate substance that specifically binds to the human integrin β_7 ; further generating cells expressing the chimera in which a region of the amino acid residue positions 111 to 378
20 of the human integrin β_7 is replaced with one of mouse origin described in detail in the antibody (I), and a chimera in which a region of the amino acid residue positions 110 to 721 of the human integrin β_7 is replaced with one of mouse origin; confirming a candidate substance showing a higher degree of bind for the
25 former; and selecting such candidate substance.

[0193] Further, the screening method (X) may also include a step of selecting a candidate substance through use of the presence of cytotoxic activity as an indicator. Specific cells serving as a target on which the cytotoxic activity is to be confirmed are not
30 particularly limited. Examples thereof may include blood cells expressing active-form integrin β_7 of human origin having a feature in the PSI domain of the human integrin β_7 described above.

[0194] Here, when the candidate substance to be screened is an antibody, the step of selecting a candidate substance through use
35 of the presence of cytotoxic activity as an indicator may be a

step of selecting an antibody having ADCC activity or CDC activity.

[0195] The candidate substance to be selected by the screening method (X) as described above is preferably an antibody, more
5 preferably a monoclonal antibody. The candidate substance to be selected is most preferably the antibody (I).

[0196] (XI) Diagnosis Method

A diagnosis method (XI) is a diagnosis method for cancer, and includes a step of bringing a sample collected from a subject
10 into contact with the antibody (I).

[0197] The subject may be the same as the subject described in detail in the treatment or prevention method (VIII) for a disease.

[0198] The sample collected from a subject may be blood or bone marrow fluid.

15 [0199] A specific diagnosis method is not particularly limited, but may involve, for example, judging that the subject has developed, or has a risk of developing, cancer when cells that bind to the antibody (I) are detected.

[0200] A person skilled in the art can easily determine the degree
20 of binding by adopting a commonly used immunoassay method. Any one of whether the subject has developed cancer and whether the subject has a risk of developing cancer may be determined depending on the degree measured here.

[0201] Specific diagnosis of cancer is not particularly limited,
25 but, for example, the subject may be diagnosed as having developed, or having a risk of developing, blood cancer, more preferably a disease causing neoplastic growth of plasma cells (e.g., myeloma or multiple myeloma) when cells that bind to the antibody (I) are plasma cells.

30 [0202] (XII) Kit

A kit (XII) is a kit for diagnosis of cancer, including the antibody (I).

[0203] The cancer is not particularly limited, and may be as described in detail in the above-mentioned pharmaceutical
35 composition (VII), and is preferably blood cancer, more

preferably a disease causing neoplastic growth of plasma cells (e.g., myeloma or multiple myeloma).

[0204] The kit (XII) may be appropriately accompanied by a manual. In such manual, the method described in detail in the above-
5 mentioned diagnosis method (XI) may be described to serve as a criterion for the diagnosis of cancer.

Examples

[0205] Now, Examples for describing the present invention in more
10 detail are described. Needless to say, the present invention is not limited to Examples described below.

[0206] Test Method: Flow Cytometry and Sorting

In the following Examples, flow cytometry (FACS) used for selecting cells was performed in the following manner.

15 [0207] Bone marrow mononuclear cells collected from an ilium of a myeloma patient who had given informed consent were suspended in an ACK solution (150 mM NH_4Cl and 10 mM KHCO_3), and the whole was left at rest at 4°C for 3 minutes to remove red blood cells. The bone marrow mononuclear cells after the removal were washed with
20 PBS supplemented with 2% fetal bovine serum, and then, in order to prevent the binding of a non-specific antibody, blocking was performed at 4°C for 20 minutes in PBS containing 10% human AB serum.

[0208] After that, each antibody (see below) labeled with a
25 fluorescent dye was added thereto to perform staining at 4°C for 30 minutes. After that, the cells were washed with PBS, and then suspended in PBS containing 1 $\mu\text{g}/\text{ml}$ propidium iodide (PI), followed by FACS analysis. The analysis of the cells and cell
30 sorting were performed using FACS Aria Cell Sorter (manufactured by Becton Dickinson Immunocytometry Systems).

[0209] For the staining of the cells, the following monoclonal antibodies were appropriately selected and used.

- APC-conjugated anti-human CD34 antibody (manufactured by BD Pharmingen)
- 35 •PE-Cy7-conjugated anti-human CD34 antibody (manufactured by BD

Pharmingen)

•APC/Cy7-conjugated anti-human CD19 antibody (manufactured by
Biolegend)

5 •FITC-conjugated anti-human CD38 antibody (manufactured by
eBioscience)

•APC-conjugated anti-human CD138 antibody (manufactured by
Biolegend)

•PE/Cy7-conjugated anti-human CD3 antibody (manufactured by
Biolegend)

10 •FITC-conjugated anti-human CD14 antibody (manufactured by BD
Pharmingen)

•PE/Cy7-conjugated anti-human CD45 antibody (manufactured by
Biolegend)

[0210] [Example 1]

15 Generation of Monoclonal Antibody Library that binds to Myeloma
Cell Line and does not bind to Healthy Person Peripheral Blood

In antibody therapy against multiple myeloma, it is
important to use an antibody that binds to myeloma cells but does
not bind to normal blood cells. In view of this, such antibody
20 was identified by the following method. First, 10,000 clones or
more of monoclonal antibodies that bound to various myeloma cell
lines were generated using the following technique.

[0211] Balb/c mice were immunized at the footpad twice a week for
from 2 weeks to 3 weeks through use of six kinds of human myeloma
25 cell lines (MM.1s cells, RPMI8226 cells, INA6 cells, U266 cells,
OPM2 cells, and KMS12BM cells) as antigens. After that, a lymph
node below the knee was removed, and a cell suspension was
generated and subjected to cell fusion with a SP2/0 mouse myeloma
cell line to generate hybridomas. The cell fusion was performed
30 using a method using polyethylene glycol (PEG method). After that,
the cells were cultured in hypoxanthine-aminopterin-thymidine
medium (HAT medium) to select hybridomas (>10,000 clones).

[0212] Finally, through use of culture supernatants of the
hybridomas, a supernatant including antibodies that bound to the
35 myeloma cell line used for the immunization and did not bind to

healthy person peripheral blood-derived mononuclear cells was selected using FACS. Antibody candidates specific to myeloma cells obtained as a result of the foregoing were about 200 clones, and the hybridomas expressing those antibody candidates were
5 grown and then cryopreserved.

[0213] [Example 2]

Identification of Antibody that specifically binds to Myeloma Cells in Human Multiple Myeloma Patient Bone Marrow

About 200 clones of candidate antibodies obtained in
10 Example 1 described above were used to stain myeloma patient-derived bone marrow cells, followed by analysis using FACS.
[0214] Each candidate antibody was added to multiple myeloma patient-derived bone marrow cells, and the cells were incubated at 4°C for 30 minutes and then washed. A PE-conjugated anti-mouse
15 IgG antibody was added as a secondary antibody, and the cells were further incubated at 4°C for 30 minutes. After washing, finally, the cells were stained using an APC-conjugated anti-human CD138 antibody, FITC-conjugated anti-human CD38, or PE/Cy7-conjugated anti-human CD45. As a negative control, a sample
20 having added thereto an isotype control in place of the candidate antibody was prepared as the same time.

[0215] Those cells were analyzed using FACS to select an antibody that bound to CD45⁻CD38⁺⁺CD138⁺ myeloma plasma cells and CD45⁻CD38⁺⁺CD138⁻ myeloma progenitor cells, but did not bind to CD45⁺
25 blood cells.

[0216] As a result, an MMG49 antibody was identified as an antibody satisfying the above-mentioned condition (FIGS. 1 and FIGS. 2). In each histogram in the figures, the Y-axis represents the number of cells, and the X-axis represents the binding
30 strength of the MMG49 antibody.

[0217] [Example 3]

Identification of Antigen Protein to which MMG49 Antibody binds

An antigen protein to which the MMG49 antibody bound was identified by an expression cloning method.

35 [0218] First, a cDNA library was generated from MM.1s cells, to

which the MMG49 antibody was known to bind, using a superscript choice system for cDNA synthesis (Invitrogen), and was inserted into a pMXs retrovirus vector (donated by Professor Toshio Kitamura at the Institute of Medical Science of the University of Tokyo) using a BstXI adaptor (Invitrogen). The thus generated cDNA library was introduced into plat-E cells (donated by Professor Toshio Kitamura), and BaF3 cells were infected with the resultant retrovirus. Thus, BaF3 cells expressing an MM.1s-derived cDNA library were obtained.

[0219] Next, the cells were repeatedly concentrated by staining the cells with the MMG49 antibody and sorting positive cells by FACS (FIG. 3). After the third sorting, most cells were cells that bound to the MMG49 antibody. Then, the retrovirus insert carried by those cells was amplified by PCR, and then sequenced to identify its base sequence. As a result, it was revealed that the insert carried by the cells was ITGB7.

[0220] [Example 4]

Confirmation that Antigen to which MMG49 Antibody binds is ITGB7-expressed Protein by Generation of ITGB7-deficient Myeloma Cells

An ITGB7-deficient U266 myeloma cell line was generated using a Crisp-cas9 system.

[0221] First, a vector was generated by inserting a double-stranded DNA sequence serving as an ITGB7-specific target sequence into a PX330 (addgene) vector. The vector was introduced together with a linear hygromycin-resistance gene expression vector (Clontech) serving as a vector for drug selection into U266 cells through use of Nucleofector (trademark) II (Lonza). After that, for clones that had grown in a medium supplemented with hygromycin, the expression of ITGB7 was stained using an FIB27 antibody (anti-integrin β_7 antibody; Biolegend), followed by analysis by FACS, to thereby identify ITGB7-deficient cells.

[0222] Next, the resultant ITGB7-deficient cells were stained using the MMG49 antibody and analyzed by FACS. As a result, it was found that the MMG49 antibody bound to wild-type U266 cells, whereas the binding of the MMG49 antibody had completely

disappeared in the ITGB7-deficient strain (FIGS. 4). This shows that MMG49 is bound to only the ITGB7-expressed protein (integrin β_7).

[0223] Next, immunoprecipitation from a lysate of MM1s myeloma cells was performed using the MMG49 antibody, followed by SDS-PAGE. Subsequently, WB was performed using an anti-integrin β_7 antibody (Miltenyi). As a result, integrin β_7 was detected in the product immunoprecipitated with the MMG49 antibody (FIG. 5). This shows that the MMG49 antibody is bound to the integrin β_7 .

[0224] [Example 5]

Measurement of Binding Pattern of MMG49 Antibody in Cell Fractions of Healthy Person Peripheral Blood and Myeloma Patient Bone Marrow

Through use of a commercially available anti-integrin β_7 antibody (FIB27 antibody; Biolegend) and the MMG49 antibody, binding to various cell fractions in healthy person peripheral blood and bone marrow cells was measured.

[0225] Red blood cells were removed from healthy person-derived peripheral blood cells using HES40, and then an Fc receptor blocking reagent (Miltenyi) was added to block non-specific antibody binding. After that, the MMG49 antibody or the FIB27 antibody, or mouse IgG2a serving as an isotype control was added, and the cells were incubated at 4°C for 30 minutes and then washed. A PE-conjugated anti-mouse IgG antibody was added as a secondary antibody, and the cells were further incubated at 4°C for 30 minutes.

[0226] The resultant cells were washed, and then, finally, stained using an APC/Cy7-conjugated anti-human CD19 antibody, an FITC-conjugated anti-human CD14 antibody, or a PE/Cy7-conjugated anti-human CD3 antibody. The cells after the staining were analyzed using FACS, to thereby measure the binding of the MMG49 antibody and the FIB27 antibody in each fraction (FIGS. 6).

[0227] In addition, 100 μ l of PBS (containing EDTA) supplemented with 1 μ l of peripheral blood of a healthy person was similarly stained using the MMG49 antibody or the FIB27 antibody, and

finally stained using a Pacific blue-conjugated anti-human CD235 antibody (BD Pharmingen) or an FITC-conjugated anti-human CD41 antibody (BD Pharmingen), and thus the presence or absence of the binding of each antibody to CD235⁺ red blood cells and platelets was also similarly investigated by FACS analysis (FIGS. 6). The results of the foregoing show that the FIB27 antibody strongly binds to many lymphoid cells, whereas the binding of the MMG49 antibody to the above-mentioned normal blood cells is extremely weak.

[0228] Further, in order to elucidate whether the binding of the MMG49 antibody to each normal cell fraction except for myeloma cells was absent in bone marrow, bone marrow cells of a myeloma patient were also similarly stained using the MMG49 antibody, and finally stained using an APC-conjugated anti-human CD34 antibody (manufactured by BD Pharmingen), Alexa647-conjugated human CD3 (manufactured by BD Pharmingen), a Cy7APC-conjugated anti-CD19 human antibody (manufactured by BD Pharmingen), a PE-Cy7-conjugated anti-CD38 human antibody (manufactured by BD Pharmingen), or an FITC-conjugated anti-CD14 human antibody (manufactured by BD Pharmingen). Those myeloma patient-derived bone marrow cells after the staining were analyzed using FACS, and thus the binding of the MMG49 antibody in each fraction was measured (FIGS. 7). The results of the foregoing show that the MMG49 antibody strongly binds to myeloma cells, but hardly binds to all normal blood cells including hematopoietic stem cell and progenitor cell fractions.

[0229] [Example 6]

Analysis of Binding of MMG49 to Various Cell Lines

The binding of the MMG49 antibody and the FIB27 antibody in various cell lines (MM1s cells, U266 cells, RPMI8226 cells, and JJN3 cells) was analyzed using FACS. A staining method is the same as in the case of peripheral blood or the like described above in Example 5.

[0230] Integrin β_7 is known to form a heterodimer with integrin α_4 or integrin α_E and be expressed on a cell surface. Therefore, the

expression thereof was also analyzed at the same time using FACS, with an Alexa647-cojugated anti-human CD49d antibody (Biolegend) and an APC-conjugated anti-human CD103 antibody (Biolegend).

CD103 represents the integrin α_E , and CD49d represents the

5 integrin α_4 . In the same manner as in Example 5 described above, the expression levels of the integrin α_E and the integrin α_4 in healthy person-derived peripheral blood were also investigated (FIGS. 8).

[0231] As a result, ITGA4 was expressed in most of the myeloma
10 cell lines, and ITGAE was expressed in none of the cell lines. The FIB27 antibody was bound to all myeloma cell lines, but the binding of the MMG49 antibody did not coincide with the expression level of the FIB27 antibody. Further, the binding of the MMG49 antibody or the FIB27 antibody to ITGA4-deficient U266
15 cells generated using a Crisp-cas9 system was investigated by FACS, and as a result, it was found that the binding of both the antibodies to U266 cells had disappeared due to ITGA4 deficiency. That is, it was found that both the MMG49 antibody and the FIB27 antibody recognized β_7 integrin expressed as $\alpha_4\beta_7$ integrin.

20 [0232] [Example 7]

Analysis of Correlation between Activation of Integrin and Binding of MMG49 Antibody

In consideration of the unique binding manner of the MMG49 antibody described above, it was supposed that the MMG49 antibody
25 recognized integrin β_7 that had been changed in structure through activation.

[0233] In view of the foregoing, K562 cells caused to forcibly express $\alpha_4\beta_7$ and human normal peripheral blood CD4T-cells concentrated using a CD4 T-cell enrichment kit (BD pharmingen)
30 were washed with 5 mM EDTA/HBS, and then incubated in 1 mM Ca^{2+} /1 mM Mg^{2+} /HBS (buffer for low activity) or 2 mM Mn^{2+} /HBS (activating buffer) together with the MMG49 antibody or the FIB27 antibody at room temperature for 30 minutes, followed by washing. A PE-conjugated anti-mouse IgG antibody was added as a secondary
35 antibody, and the cells were further incubated at room

temperature for 30 minutes. The resultant cells were analyzed using FACS, to thereby measure the binding of the MMG49 antibody and the FIB27 antibody in cells in which the integrin $\alpha_4\beta_7$ had been activated.

5 [0234] As a result, an enhancement in binding of the MMG49 antibody in the presence of Mn^{2+} was observed (FIGS. 10). Meanwhile, no similar change was observed for the FIB27 antibody. This suggests the possibility that the MMG49 antibody is an antibody specific to activated conformation of integrin β_7 .

10 [0235] [Example 8]

Identification of Epitope essential for Recognition by MMG49 Antibody

In order to identify an epitope recognized by the MMG49 antibody, vectors for expressing eight kinds of human/mouse
15 chimeric integrin β_7 proteins as illustrated in FIG. 11 were generated using an overlapping PCR method.

[0236] Each expression vector was introduced into 293T-cells by a lipofection method, and 48 hours after that, the presence or absence of the binding of the MMG49 antibody was analyzed. The
20 cells were suspended in PBS supplemented with 1% fetal bovine serum, the MMG49 antibody was added, and then the whole was left at rest at room temperature for 30 minutes. After washing, an Alexa488-anti-mouse IgG antibody was added, and the whole was left at rest at room temperature for 30 minutes, followed by
25 analysis by FACS.

[0237] As a result, it was revealed that the MMG49 antibody strongly bound to a chimeric integrin β_7 protein (#4960) in which a region of the amino acid residue positions 110 to 721 was of mouse origin and the rest (region of the amino acid residue
30 positions 20 to 109 and region of the amino acid residue positions 722 to 798) had sequences of human origin in almost the same manner as in the case of a chimeric integrin β_7 protein entirely of human origin (#4927) (FIG. 11 to FIG. 13).

[0238] In view of the fact that the region of the amino acid
35 residue positions 722 to 798 includes a transmembrane domain (TM)

and an intracellular domain (cytoplasmic), and that a region of the amino acid residue positions 1 to 19 is a signal peptide, it was shown that an epitope essential for the binding of the MMG49 antibody was present in the region of the amino acid residue

5 positions 20 to 109 including a PSI domain.

[0239] In addition, it was revealed that the MMG49 antibody had a slightly increased avidity for a chimeric integrin β_7 protein (#4961) in which a region of the amino acid residue positions 110 to 378 was of mouse origin, and in which a region of the amino acid residue positions 20 to 109 and a region of amino acid residue positions 379 to 798 were of human origin as compared to the chimeric integrin β_7 protein (#4960), and the increased avidity was at exactly the same binding level as in the case of the chimeric integrin β_7 protein entirely of human origin (#4927).

10 [0240] Further, it was also revealed that the MMG49 antibody had slightly increased binding capacity for a chimeric integrin β_7 protein (#4944) in which the above-mentioned region of the amino acid residue positions 20 to 109, which had been shown to include the epitope of the MMG49 antibody, and a region of the amino acid residue positions 1 to 378 including a region corresponding to a signal peptide of the amino acid residue positions 1 to 19 were of mouse origin, and in which a region of the amino acid residue positions 379 to 798 was of human origin, and a chimeric integrin β_7 protein (#4945) in which a region of the amino acid residue positions 1 to 416 was of human origin and a region of the amino acid residue positions 417 to 798 was of human origin as compared to a chimeric integrin β_7 protein (#4946) in which a region of the amino acid residue positions 1 to 563 was of mouse origin and a region of the amino acid residue positions 564 to 798 was of human origin, and a chimeric integrin β_7 protein (#4947) in which a region of the amino acid residue positions 1 to 721 was of mouse origin and a region of the amino acid residue positions 722 to 798 was of human origin.

[0241] In view of the above-mentioned experimental results, it was also revealed that the specific avidity, that is, affinity of the

MMG49 antibody for the region of the amino acid residue positions 20 to 109 of the integrin β_7 was increased by the region of the amino acid residue positions 379 to 721 of human integrin β_7 , that is, in the presence of the region of the amino acid residue positions 379 to 721 of human integrin β_7 .

[0242] [Example 9]

Determination of Base Sequence of Antibody Molecule Variable Regions of MMG49 Antibody

The subclass of the MMG49 antibody was confirmed using an Isotyping kit (Roche), and was confirmed to be IgG2a subclass. Further, the base sequences and amino acid sequences of variable regions of the MMG49 antibody were determined.

[0243] A sequence determination method was performed using a Smarter RACE cDNA amplification kit (Clontech). That is, cDNAs generated from mRNAs derived from hybridomas MMG49 producing the MMG49 antibody were used as templates to amplify cDNA fragments of H-chain and κ -chain variable regions by a PCR reaction, and their base sequences were read. The read amino acid sequence and base sequence, and hypervariable regions (CDR1 to CDR3) of the H-chain variable region are shown in Tables 3 and 4 below.

[0244] The read amino acid sequence and base sequence, and hypervariable regions (CDR1 to CDR3) of the L-chain (κ -chain) variable region are also shown in Tables 3 and 4 below.

[0245] In order to confirm the specificity of the isolated variable region sequences of the MMG49 antibody, variable region sequence cDNAs were bound to a human IgG4 constant domain and human IgL κ -chain constant domain sequence to generate a chimerized antibody. Specifically, each variable region sequence was inserted into pFuse-CH-Ig-hG4 or pFuse-CL-Ig-hk (invivogen) using In-Fusion cloning kit (Takara). After that, the resultant was introduced into FreeStyle CHO-S cells (Invitrogen), and a chimerized antibody secreted into the culture supernatant thereof was recovered. Next, MM1s cells to which the MMG49 antibody bound and KMS12BM cells to which the MMG49 antibody did not bind were incubated in a buffer supplemented with MMG49-hIgG4, and were

washed. After that, biotinylated anti-human IgG (Rockland) was added as a secondary antibody, and the cells were washed again, and then stained by adding streptavidin-PE (Biolegend), followed by FACS analysis. As a result, MMG49-hIgG4 showed a staining pattern similar to that of the original MMG49 antibody, suggesting that the obtained variable region sequences were correct (FIGS. 14).

[0246]

<Table 3: Amino acid sequences of MMG49>		
Heavy chain	CDR1 (SEQ ID NO: 1)	GYTFSSYW
	CDR2 (SEQ ID NO: 2)	MLPGSGSS
	CDR3 (SEQ ID NO: 3)	ARGDGNYWYFDV
	Variable region (SEQ ID NO: 4)	MEWTWVFLFLLSVTAGVHSQVQLQQSGAELMKPGAS VKISCKASGYTFSSYWIEWVKQRPBGHLEWIGEMLP GSGSSNYNEKFKGKATFTADTSSNTAYMQLSSLTSE DSAVYYCARGDGNYWYFDVWGAG
Light chain	CDR1 (SEQ ID NO: 6)	SSVGY
	CDR2 (SEQ ID NO: 7)	ATS
	CDR3 (SEQ ID NO: 8)	QQWSSDPPT
	Variable region (SEQ ID NO: 9)	MDFQVQIFSFLLISASVIMSRGQIVLSQSPAILAS PGEKVTMTCRASSSVGYMHWFQQKPGSSPKPWYAT SNLASGVPARFSGSESGTSYSLTISRVEAEDAATYY CQQWSSDPPTFGGGTKLEIK

10 [0247]

<Table 4: Base sequences of MMG49>		
Heavy chain	CDR1 (SEQ ID NO: 11)	GGCTACACATTCAGTAGCTACTGG
	CDR2 (SEQ ID NO: 12)	ATGTTACCTGGAAGTGGTAGTTCT
	CDR3 (SEQ ID NO: 13)	GCAAGGGGGGATGGTAACTACTGGTACTTCGATGTC
	Variable region (SEQ ID NO: 14)	ATGGAATGGACCTGGGTCTTTCTCTTCCTCCTGTCA GTAAGTGCAGGTGTCCACTCCCAGGTTTCAGCTGCAG CAGTCTGGAGCTGAGCTGATGAAGCCTGGGGCCTCA GTGAAGATATCCTGCAAGGCTTCTGGCTACACATTC AGTAGCTACTGGATAGAGTGGGTAAAGCAGAGGCCT GGACATGGCCTTGAGTGGATTGGAGAGATGTTACCT GGAAGTGGTAGTTCTAACTACAATGAGAAGTTCAAG GGCAAGGCCACATTCAGTGCAGATACATCCTCCAAC ACAGCCTACATGCAACTCAGCAGCCTGACATCTGAG GACTCTGCCGTCTATTACTGTGCAAGGGGGGATGGT AACTACTGGTACTTCGATGTCTGGGGCGCAGGG
Light chain	CDR1 (SEQ ID NO: 16)	TCAAGTGTAGTTAC

	CDR2 (SEQ ID NO: 17)	GCCACATCC
	CDR3 (SEQ ID NO: 18)	CAGCAGTGGAGTAGTGACCCACCGACG
	Variable region (SEQ ID NO: 19)	ATGGATTTTCAAGTGCAGATTTTCAGCTTCCTGCTA ATCAGTGCTTCAGTCATAATGTCCAGAGGACAAATT GTTCTCTCCCAGTCTCCAGCAATCCTGTCTGCATCT CCAGGGGAGAAGGTCACAATGACTTGCGGGCCAGC TCAAGTGTAGGTTACATGCACTGGTTCCAGCAGAAG CCAGGATCCTCCCCAAACCCTGGATTTATGCCACA TCCAACCTGGCTTCTGGAGTCCCTGCTCGCTTCAGT GGCAGTGAGTCTGGGACCTCTTACTCTCTCACAATC AGCAGAGTGGAGGCTGAAGATGCTGCCACTTATTAC TGCCAGCAGTGGAGTAGTGACCCACCGACGTTCCGT GGAGGCACCAAGCTGGAAATCAAA

[0248] [Example 10]

Generation of Chimeric Antigen Receptor T-cells using Antibody Molecule Variable Regions of MMG49 Antibody

5 Chimeric antigen receptor T-cells (hereinafter referred to as MMG49 antibody-derived chimeric antigen receptor T-cells) were generated using the MMG49 antibody molecule variable region sequences by the following procedure with reference to Non Patent Literatures 2 to 4 and the like.

10 [0249] (1) Cloning of CD28 and CD3z:

RNAs were collected from Jurkat cells through use of Trizol (Invitrogen), and cDNAs were generated using a Superscript III cDNA synthesis kit (Invitrogen). Then, the cDNAs were used as templates to amplify cDNAs of CD28 and CD3z by PCR, each of which
15 was cloned using a TA cloning kit (Invitrogen), and their base sequences were confirmed by sequencing.

[0250] (2) Binding of Four Fragments, i.e., MMG49 antibody-derived VL/VH, and CD28/CD3z:

Through use of an overlapping PCR method, respective gene
20 fragments of the MMG49 antibody-derived VL region and VH region, and the CD28 and CD3z cloned above were bound to each other to generate chimeric cDNA. A procedure therefor and primers used are illustrated in FIG. 15. The base sequences of the primers used are shown in Table 5 below.

[0251]

Table 5

No.	Primer name	Base sequence
23	49_car_vk-s5	gaattccaccatggattttcaagtgcagatt
24	49_car_vk-as6	gccggaaccgctagtggagccccgtttgatttccagcttgg
25	593_car_vk_as4	gctgccttctccgctgccaggtttgccggaaccgctagtggagcc
26	49_car_vh-s5	aaacctggcagcggagaaggcagccaggttcagctgcagcagtc
27	49_car_vh-as6	tgaggagacggtgaccgtgg
28	49_VKVH28as8	atacataacttcaattgcggccgctgaggagacggtgaccgtgg
29	49carinfus1	ctaggcgccggaattccaccatggattttc
30	tcrzcarinfuas4	aatgtcgacctcgagtggctgttagcgag

[0252] The bound chimeric cDNA was cloned using a ZeroBlunt PCR cloning kit (Invitrogen), and then sequenced to confirm its base sequence. In addition, an amino acid sequence (SEQ ID NO: 21) confirmed on the basis of the confirmed base sequence and the base sequence (SEQ ID NO: 22) are shown in the sequence listing. The amino acid sequence set forth in SEQ ID NO: 21 is one obtained by conversion into an amino acid sequence from the start codon (atg) immediately following the Kozak sequence (gaattccacc) shown above in SEQ ID NO: 23, which was excluded.

[0253] (3) Insertion into Expression Vector:

Subsequently, the chimeric cDNA bound in (2) was cleaved with two restriction enzymes EcoRI/SalI, and inserted into an MSCV-ires-GFP vector.

[0254] The MMG49 antibody-derived chimeric antigen receptor cDNA retrovirus vector generated by the foregoing was introduced into 293T cells together with a gag/pol and VSV-G envelope expression vector through use of lipofectamine 2000 (Invitrogen), to thereby generate a retrovirus. After 48 hours from the gene introduction, a supernatant was recovered and used as a virus solution.

[0255] (4) Introduction into T-cells:

Subsequently, cDNA of an MMG49 antibody-derived chimeric antigen receptor was introduced into human T-cells as described below.

[0256] First, human peripheral blood mononuclear cells were added to a 48-well plate coated with an anti-CD3 antibody (eBioscience)

and cultured for 72 hours. X-VIVO15 (Lonza) supplemented with 10% human AB serum and IL-2 (175 IU/L) was used as a culture medium, and the peripheral blood mononuclear cells were stimulated. After that, the virus solution generated above was added to a 48-well plate coated with Retronectin (Takara), and the virus was adsorbed to Retronectin by centrifugation at 1,700×g for 120 minutes. After that, the peripheral blood mononuclear cells (including T-cells) after the stimulation were added, and the gene was introduced thereinto. After that, culture was continued in the above-mentioned medium to amplify MMG49 antibody-derived chimeric antigen receptor T-cells, which were used in the following investigation. The T-cells caused to express a CAR construct using variable regions of the MMG49 antibody were stained using a PE-anti-human F(ab')₂ antibody (Jackson Laboratory), and as a result, the expression of human F(ab')₂ was detected in proportion to the expression of GFP indicating the introduction of the construct (FIGS. 16). That is, it was confirmed that the introduced CAR was expressed on a cell surface. [0257] [Example 11]

20 Analysis of Recognition of ITGB7-expressing Tumor Cells by MMG49 Antibody-derived Chimeric Antigen Receptor T-cells and Cytotoxic Activity thereof

The MMG49 antibody-derived chimeric antigen receptor T-cells generated by the above-mentioned method or control T-cells having introduced therein only GFP were cocultured with K562 cells expressing no integrin β_7 or K562 cells caused to forcibly express integrin $\alpha_4\beta_7$, and the amount of a produced cytokine was quantitatively determined. Specifically, 1×10^5 each of the T-cells and the target cells were added to a 96-well plate. After 24 hours, a supernatant was recovered, and the amount of production of IFN- γ was measured by ELISA. The measurement was performed using a Quantikine kit (R&D). As a result, only in the coculture of the K562 cells caused to forcibly express integrin $\alpha_4\beta_7$ with the MMG49 antibody-derived chimeric antigen receptor T-cells, higher production of IFN- γ and IL2 than in the control (T-

cells obtained by similarly culturing stimulated peripheral blood mononuclear cells having introduced therein a GFP expression vector) was observed (FIGS. 17).

Next, the MMG49 antibody-derived chimeric antigen receptor
5 T-cells or control T-cells having introduced therein only GFP
were cocultured with myeloma cell lines to which the MMG49
antibody bound (MM.1s cells, RPMI8226 cells, and JJN3 cells) or
cells to which the MMG49 antibody did not bind (KMS12BM, Molt4,
and Raji cells), and the amount of a produced cytokine was
10 similarly quantitatively determined. As a result, only when the
MM.1s, RPMI8226 cells, and JJN3 cells, which were cells to which
the MMG49 antibody bound, were cocultured with the MMG49
antibody-derived chimeric antigen receptor T-cells, higher
production of IFN- γ and IL2 than in the control (T-cells obtained
15 by similarly culturing stimulated peripheral blood mononuclear
cells having introduced therein a GFP expression vector) was
observed (FIGS. 18 and FIGS. 19). The results show that the MMG49
antibody-derived chimeric antigen receptor T-cells are activated
by recognizing an antigen recognized by the MMG49 antibody
20 (sometimes referred to as MMG49 antigen).

[0258] Further, whether the MMG49 antibody-derived chimeric
antigen receptor T-cells damaged a myeloma cell line was
investigated by ^{51}Cr cytotoxicity assay. First, K562 cells
expressing no integrin β_7 or K562 cells caused to forcibly express
25 integrin $\alpha_4\beta_7$ to be used as target cells were cultured in RPMI1640
medium supplemented with 10% FCS, and were prepared so that the
number of cells was from 0.5×10^4 to 1.0×10^4 .

[0259] An appropriate amount of $\text{Na}_2^{51}\text{CrO}_4$ was added thereto and
allowed to react therewith at 37°C for 2 hours to label the cells
30 with ^{51}Cr , and the resultant cells were washed and then used as
target cells. The target cells were mixed with MMG49 antibody-
derived chimeric antigen receptor T-cells suspended in RPMI1640
medium supplemented with fetal bovine serum, and the cells were
cocultured for 4 hours.

35 [0260] After that, ^{51}Cr released into the supernatant was measured

with a γ -counter. A cell damage percentage (%) was determined on the basis of the following expression (1).

$$[0261] \quad (A-B)/(C-D) \times 100 \quad (1)$$

A: Amount of ^{51}Cr released from cells used in experiment

5 B: Spontaneous ^{51}Cr release amount under antibody-free state

C: Maximum ^{51}Cr release amount with addition of 1% Triton X-100

D: Spontaneous ^{51}Cr release amount under antibody-free state.

[0262] As a result, in the K562 cells caused to forcibly express integrin $\alpha_4\beta_7$ to which the MMG49 antibody bound, higher cell
10 damage caused by the MMG49 antibody-derived chimeric antigen receptor T-cells than in the T-cells expressing only GFP serving as a control was observed (FIGS. 20).

[0263] Next, the MMG49 antibody-derived chimeric antigen receptor T-cells or control T-cells having introduced therein only GFP
15 were cocultured with myeloma cell lines to which the MMG49 antibody bound (MM1s cells, RPMI8226 cells, and JJN3 cells) or cells to which the MMG49 antibody did not bind (KMS12BM, Molt4, and Raji cells), and a similar investigation was performed. As a result, only in the K562 cells caused to forcibly express
20 integrin $\alpha_4\beta_7$ to which the MMG49 antibody bound, higher cell damage caused by the MMG49 antibody-derived chimeric antigen receptor T-cells than in the T-cells expressing only GFP serving as a control was observed (FIGS. 21).

[0264] The above-mentioned results show that the MMG49 antibody-
25 derived chimeric antigen receptor T-cells can specifically damage cells expressing an antigen to be recognized by the MMG49 antibody.

[0265] [Example 12]

Analysis of Myeloma Cell-eliminating Ability exhibited by MMG49
30 Antibody-derived Chimeric Antigen Receptor T-cells in vivo

A therapeutic effect on multiple myeloma was investigated in vivo using the MMG49 antibody-derived chimeric antigen receptor T-cells.

[0266] Myeloma cell line MM1s cells (4×10^5 cells) were
35 transplanted into the bone marrow of NOG mice subjected to

radiation exposure at 2.4 Gy. After 5 days, the mice were grouped into an MMG49 antibody-derived chimeric antigen receptor T-cell-administered group and a control T-cell-administered group, and each group was intravenously administered with 5×10^6 cells per mouse. After 7 days from the administration, the bone marrow was analyzed. As a result, marked growth of myeloma cells was clearly observed in all mice of the control T-cell-administered group, whereas the tumor had almost completely disappeared in the MMG49 antibody-derived chimeric antigen receptor T-cell-administered group. The results show that the administration of the MMG49 antibody-derived chimeric antigen receptor T-cells has an ability to eliminate a tumor expressing the MMG49 antigen even in vivo (FIGS. 22).

[0267] Further, a therapeutic effect on multiple myeloma was investigated in vivo using a myeloma-systemically seeded model. [0268] Myeloma cell line MM1s cells (5×10^6 cells) having introduced therein luciferase gene were intravenously transplanted into NOG mice subjected to radiation exposure at 2.4 Gy. After 5 days from the transplantation, the degree of engraftment of the tumor cells was measured using an IVIS imaging system (PerkinElmer). After that, the mice were grouped into an MMG49 antibody-derived chimeric antigen receptor T-cell-administered group and a control T-cell-administered group, and each group was intravenously administered with 3×10^6 cells per mouse 5 days and 7 days after the transplantation. After 7 days from the second T-cell administration, a tumor volume was measured again using the IVIS imaging system. As a result, marked growth of myeloma cells was clearly observed in all mice of the control T-cell-administered group, whereas the tumor had almost completely disappeared in the MMG49 antibody-derived chimeric antigen receptor T-cell-administered group (FIGS. 23). The results show that the administration of the MMG49 antibody-derived chimeric antigen receptor T-cells has an ability to eliminate a tumor expressing the MMG49 antigen even in vivo.

[0269] [Example 13]

For the epitope of the MMG49 antibody, an experiment for investigating, in more detail, the results of the investigation in Example 8 was performed. Vectors for expressing three kinds of human/mouse chimeric integrin β_7 protein as illustrated in FIGS.

5 25 were generated, and each of the expression vectors was introduced into 293T cells by a lipofection method. After 48 hours, the presence or absence of the binding of the MMG49 antibody was analyzed by FACS.

[0270] As a result, it was revealed that the MMG49 antibody
10 strongly bound to a chimeric integrin β_7 protein (ch5.1 in FIGS. 25) in which regions of the amino acid residue positions 1 to 32 and positions 91 to 798 of integrin β_7 protein were of mouse origin, and the rest (region of the amino acid residue positions 33 to 90) had a sequence of human origin in almost the same
15 manner as in the case of the integrin β_7 protein entirely of human origin (#4927 in FIG. 11) (FIGS. 25).

[0271] Thus, it was strongly suggested that the epitope of the MMG49 antibody was included in the amino acid residues at positions 33 to 90 of the human integrin β_7 protein.

20 [0272] [Example 14]

Vectors expressing the human integrin β_7 , the mouse integrin β_7 , and various variants obtained by mutating only one or two amino acids of the human integrin β_7 into an amino acid sequence of mouse origin (R35E/N36D, H38D, M41L/L42Q, and A48V)
25 were introduced into 293T cells by a lipofection method, and an experiment was performed in the same manner as in Example 8 thereafter. As a result, as shown in FIGS. 26, it was revealed that only the A48V variant had a remarkably reduced avidity for the MMG49 antibody as compared to the human integrin β_7 , and had a
30 numerical value close to that of the mouse integrin β_7 . The results revealed that the amino acid residue at position 48 of the human integrin β_7 was strongly related to the epitope of the MMG49 antibody, or included in the epitope of the MMG49 antibody.

[0273] Base sequences and amino acid sequences described
35 herein are shown below.

CLAIMS

[Claim 1]

An anti-human integrin β_7 antibody comprising:

a heavy chain variable region comprising

5 heavy-chain CDR1 having the amino acid sequence set
 forth in SEQ ID NO: 1,

 heavy-chain CDR2 having the amino acid sequence set
 forth in SEQ ID NO: 2, and

 heavy-chain CDR3 having the amino acid sequence set
10 forth in SEQ ID NO: 3; and

a light chain variable region comprising

 light-chain CDR1 having the amino acid sequence set
 forth in SEQ ID NO: 6,

 light-chain CDR2 having the amino acid sequence set
15 forth in SEQ ID NO: 7, and

 light-chain CDR3 having the amino acid sequence set
 forth in SEQ ID NO: 8.

[Claim 2]

20 An antibody according to claim 1, the antibody comprising:

a heavy chain variable region having the amino acid sequence set
forth in SEQ ID NO: 4;

and

a light chain variable region having the amino acid sequence set
25 forth in SEQ ID NO: 9.

[Claim 3]

An antibody according to claim 1 or 2, which is a multispecific antibody.

5

[Claim 4]

A chimeric antigen receptor comprising an antigen recognition site of an antibody according to any one of claims 1 to 3.

10 [Claim 5]

A cell expressing the chimeric antigen receptor of claim 4.

[Claim 6]

15 A polynucleotide, which encodes the chimeric antigen receptor of claim 4.

[Claim 7]

A vector comprising the polynucleotide of claim 6.

20 [Claim 8]

A cell, which harbors the polynucleotide of claim 6 or the vector of claim 7.

[Claim 9]

25 A cell according to claim 5 or 8, which is a chimeric antigen

receptor T cell or NK cell.

[Claim 10]

A pharmaceutical composition for the treatment of multiple myeloma
5 comprising the antibody according to any one of claims 1 to 3.

[Claim 11]

A pharmaceutical composition for the treatment of multiple myeloma
comprising the chimeric antigen receptor T cell or NK cell of
10 claim 9.

[Claim 12]

The pharmaceutical composition of claim 11, which is characterised
in that said chimeric antigen receptor T cell or NK cell is
15 administered at 10^4 cells / kg to 10^9 cells / kg per 1 days.