Method for promoting and improving properties of adipose tissue, tissue and cells obtained by said method.

The present invention relates to a method for promoting anti-inflammatory properties and for improving repair ability of a tissue containing mesenchymal stem cells (MSCs), to the tissue and to mesenchymal stem cells obtained by said method.

It is known to prepare tissues that can repair, replace or restore structures and/or impaired biological functions resulting from congenital defects, ageing, diseases, or destruction thereof (Haseltine, 2003; Gutmann et al., 2005, Greenwood. et al., 2006).

Mesenchymal stem cells (MSCs) are multi-potent cells, able to self-renew and able to differentiate, in vitro, into cells of mesenchymal origin, including osteoblasts, adipocytes and chondrocytes, and able to give rise to tissues such as bone, fat, cartilage and muscle, in vivo (Pittenger et al., 1999). Recent studies have proved that the capacity of tissue regeneration can be improved by using mesenchymal stem cells derived from human adipose tissue (so called hADSCs) (Gimble et al., 2011).

The number of mesenchymal stem cells seems to be considerably higher in adipose tissue than in other tissues, such as bone marrow (Bieback et al., 2004).

The autologous transplantation of adipose tissue is a well-known therapeutic procedure, used for repairing several damages to tissues.

Moreover several recent publications disclose that human adipose-derived stem cells (hADSCs) are able, being properly stimulated, to differentiate in other different types of cells, such as neurons, cardiomyocytes, hepatocytes, and pancreatic cells (Schäffler et al., 2007; Strem et al., 2005) and have a clinical potential for vasculogenesis (Madonna et al., 2009), osteogenesis (Shenaq et al., 2010) and neuronal repair models (Wakabayashi et al., 2010).

It is also known that mechanical forces can be very important for adult tissue plasticity, as it occurs during embryogenesis, tissue remodeling and mitotic and motility behavior of cells (Vogel et al., 2006).

Cells perceive and respond to forces applied from the outside and to forces exerted on the cell matrix and on cell-cell contacts.

Ion channels activated by stretch, growth factors, cytoskeletal strands, extracellular matrixes, focal adhesions, and tyrosine kinase are involved in the conversion of mechanical stimuli into biochemical signals in cells (mechanotransduction) that lead to downstream events and therefore guide key messages on fate to the nucleus (El Haj et al., 1999, Na et al., 2008; Walker et al., 2000, Venkatesan Iyer et al., 2012, Chen et al., 2013).

An important challenge in regenerative medicine is the minimal manipulation of stem cells, such to easily transform a specific cell line in cell therapy.

"Minimal manipulation" means a processing that

does not alter the original relevant characteristics of the tissue relating to the tissue's utility for reconstruction, repair, or replacement (www.ema.europe.eu; www.fda.gov).

Regulatory agencies (FDA and EMA) and the International Society for Stem Cell Research (ISSCR) state that cell therapy should be considered as a medicine when there is more than one "minimal manipulation" of cells intended for clinical application or when the intended use of the cells is different from their usual function in the body (www.ema.europe.eu; www.fda.gov; Hyun et al., 2008).

Regulatory agencies state that patient protection is the core of such rules. In addition to the application of the same safety and efficacy rules, as for all the medicines, the quality and production of cell-based products declared in the requirements of good-manufacturing practice (GMP). These are standards recognized overall for guaranteeing the quality in producing and controlling medicines. Any use of such cell-based medicines is subjected to authorization and controls, including the procedure for manufacturing them (www.ema.europe.eu; www.fda.gov). According to regulatory agencies, in the cell culture in vitro, it is necessary to consider said rules to guarantee the growth and manipulation of isolated cells as acceptable. The processing steps shall be properly designed to preserve integrity and to control the function of the cells. Any manipulation procedures shall be documented in details and closely monitored

according to specific process controls. The duration of cell culture and maximum number of cell passages shall be clearly specified and validated. The relevant genotypic and phenotypic characteristics of primary cell cultures, of established cell lines and the derived cell clones shall be defined and their stability, with respect to culture longevity, determined. Consistency and repeatability of the cell culture process shall be demonstrated and the culture conditions, including the media and the duration shall be optimized with respect to the intended clinical function of the cells. For such reasons, research on stem cells applied to regenerative medicine has to adhere to the main concept of "minimal manipulation".

It is also known that as people get older, the handling of Parkinson's disease is a more and more important and demanding aspect of medical practice for neurologists and general practice doctors.

Parkinson's disease (PD) is a common neurodegenerative disease, a synucleinopathy, with a prevalence of 160/100 000 units in Western Europe, till involving about 4% of the population over 80 years old.

The comprehension of the pathogenesis of the disease has improved in the last ten years and for example important progresses have been made in creating cells producing dopamine from stem cells.

The general object of the present invention is to activate at molecular and cellular level the adipose tissue, for example removed by liposuction, in a very simple, cheap and particularly functional manner.

A further object of the present invention is to have a minimal manipulation of cells, particularly of mesenchymal stem cells, derived from adipose tissue, without chemical digestion of said tissue.

Another object is to change the inflammatory condition of a adipose tissue, particularly to trigger anti-inflammatory properties and to suppress the production of inflammatory cytokines such as TNF-alpha (tumor necrosis factor-alpha) in human adipose tissue obtained for example by liposuction, and to improve the tissue repair ability, by a minimal manipulation procedure, such to use mesenchymal stem cells obtained from human adipose tissue (called as hADSCs) for therapeutic purposes.

Considering the above objects, according to the present invention, we have thought of providing a method for promoting anti-inflammatory properties and for improving the repair capacity of a tissue containing mesenchymal stem cells (MSCs), a adipose tissue obtained by said method and mesenchymal stem cells derived from said tissue, having the characteristics disclosed in the annexed main claims and in the subclaims.

According to the present invention the method comprises at least one mechanical activation step performed by applying an orbital shaking force on a adipose tissue lipoaspirated or removed from the body.

According to the present invention the force applied on the adipose tissue lipoaspirated or removed from the body has an intensity ranging from 40 to 120 x

g.

In a preferred embodiment such applied force has intensity equal to 97 x g.

Said force is applied for a time ranging from 3 to 20 minutes.

For example said force is applied for 10 minutes.

It is possible to provide to apply the force to the adipose tissue lipoaspirated or anyway removed from the human or animal body by any known technique, and contained in a suitable container, only once or it is possible to provide said force to be applied two or more times in a manner repeated over time.

For each repetition, the force intensity as well as the duration of each application can be the same or different.

The method of the present invention can be applied both to fresh adipose tissue and to cryopreserved tissue.

The orbital shaking force can be applied to the adipose tissue lipoaspirated or removed from the body by a device composed of a motor with an orbital movement provided with a disposable and sterile container, inside which the adipose tissue lipoaspirated or removed from the body is contained, which has a capacity up to 500 ml.

The present invention relates also to a tissue obtained by said method and stem cells obtained from said tissue by a further step of

isolating and expanding said mesenchymal stem cells contained in said adipose tissue lipoaspirated or

removed from the body (hADSCs).

Particularly the adipose tissue and the mesenchymal stem cells obtained by the method of the present invention have improved anti-inflammatory properties and improved repair capacity, since in said tissue and mesenchymal stem cells:

- the expression of inflammatory cytokines is eliminated such as tumor necrosis factor-alpha (TNF-alpha);
- the expression of its natural inhibitor TSG-6 and of interleukin 15 is considerably enhanced;
- the expression of leptin is considerably improved;

and the parameters that define the stemness are increased such as the expression of pluripotency-related genes (SOX2, OCT4, Nanog), of beta-tubulin III and TSG6 mRNA.

The mechanically activated adipose tissue, that is removed, preserved in a container and subjected to orbital shaking, is completely similar, from a structural point of view, to the tissue obtained by biopsy.

The object of the present invention is also the use of said tissue and mesenchymal stem cells in cell therapy.

Particularly and not as a limitation, stem cells present in the adipose tissue, for example said mesenchymal stem cells activated according to one or more of the characteristics and of the embodiments described above, are used in activation of endogenous

stem cells of spinal cord.

Characteristics of the present invention and its advantages will be more clear and evident from an examination of the following description, referred to annexed figures showing experimental results.

In the figures:

- figures 1A, 1B and 1C are the differential expression of cytokines in lipoaspirated adipose tissue and in lipoaspirated adipose tissue after mechanical activation according to the present invention: TSG6 is already observable after 3 minutes of stimulation and it has maximum expression within 6-10 minutes; TNF-alpha is suppressed in 6 minutes. Figure 1D shows that such effects are observable also in the cryopreserved adipose tissue; TQ= time 0, PZA=patient A, PZB= patient B, TNF-α=tumor necrosis factor-alpha, LEPT=leptin, IL15= interleukin 15;

- figure 2 is the expression of pluripotency genes (SOX2, OCT4, Nanog), beta-tubulin III and TSG6 tested with real-time RT PCR in samples from biopsy of human adipose tissue, lipoaspirated and lipoaspirated mechanically activated by the method of the present invention (10 minutes). All the investigated genes are hardly activated in the regular bioptic tissue, that is non processed tissue, according to the present invention (biopsy), and are slightly activated by performing the known Coleman procedure after liposuction (lipoaspirate). Differently the upregulation is incomparably higher after the specific activation according to the present invention for 10

minutes;

- figure 3 is the results of a real-time quantitative PCT measuring the expression levels of mRNA of pluripotency genes; TQ= time 0, PZA=patient A, PZB= patient B;

- figure 4 shows that the application of a mechanical force to the adipose tissue modifies the interaction of SOX2 and HuR with the promoter of specific genes (bTUBIII=beta-tubulin III, TSG6, IL15=interleukin15 and TNF-α=tumor necrosis factor-alpha);

- figures 5A and 5B are the morphology in vivo of hADSCs obtained from adipose tissue treated by orbital shaking according to the present invention (6 and 10 minutes, respectively);

- figures 6A and 6B are the expression of beta-tubulin III (TUJ 1) and of vimentin in hADSCs obtained from adipose tissue mechanically treated with orbital shaking according to the present invention;

- figure 7 shows FACS analyses of the expression of mesenchymal markers in mesenchymal stem cells obtained from adipose tissue treated or not treated with a force 97 x g; TQ=time 0;

- figures 8A and 8B are the analysis of the karyotype unchanged in hADSCs (passages 2 to 9) obtained from adipose tissue treated with mechanical force according to the present invention. The micrograph relates to hADSCs of a case and it is representative of the same analysis performed in other three cases; PZ59= patient 59;

- figure 9 shows the spheroids formed by hADSCs derived from adipose tissue mechanically activated according to the present invention, after having been cultured in a neurosphere medium for 21 days; NPCs= Adult murine neural precursor cells;

- figure 10 shows that SOX2 interacts with the promoter of specific genes (bTUBIII=beta-tubulin III, TSG6, IL15=interleukin 15 and TNF-α=tumor necrosis factor-alpha) also in mesenchymal stem cells (hADSCs) isolated from adipose tissue treated with force 97 x g;

- figure 11 shows that mesenchymal stem cells (MSCs) isolated from adipose tissue mechanically activated according to the present invention (hADSCs) are able to reduce the release of IL-1 beta from THP1 cells; LPS= bacterial lipopolysaccharide endotoxin;

- figure 12 shows that hADSCs isolated from adipose tissue mechanically activated according to the present invention efficaciously reduce the release of IL-1 beta from THP1 cells that have a LPS-mediated increase, unlike those from lipoaspirate (LS);

- figure 13 shows the levels of mRNA of proinflammatory and anti-inflammatory cytokines. The experiment was performed by real-time RT-PCR. Results are the mean of two different experiments of real-time RT-PCR. hADSCs from mechanically activated adipose tissue knock down inflammatory cytokines IL1α and TNF-α=tumor necrosis factor-alpha and increase TSG6;

- figure 14 shows the evaluation of the percentage of errors in the use of forelimbs. Such data were obtained by using the horizontal grid test. hADSCs

obtained by the method according to the present invention reduce the percentage of error in Parkinson's animal model after transplantation;

- figure 15 shows the evaluation of the time necessary to climb down from the top of the vertical grid in the vertical grid test. MPTP-treated mice employed longer time to turn head down and to climb down from the vertical grid. Time is considerably reduced by the intrastriatal injection of hADSCs obtained by the method according to the present invention;

- figures 16A-16D are the expression of lncRNA-COX2 in areas of the brain of mice. The expression was studied 4 days after intrastriatal injection of hADSCs obtained by the method of the present invention in mice with Parkinson's disease induced by MPTP. MPTP causes the appearance of Linc-COX2 in cortex and striatum (areas involved by MPTP-induced Parkinson's disease), that is eliminated by the intrastriatal treatment of the mouse with mesenchymal stem cells from adipose tissue activated according to the present invention. This is related to the improvement of the limb function;

- figure 17 is the localization in the brain of LnRNA COX2 in health mouse (NT); in mouse suffering from Parkinson's disease (MPTP) and in mice with Parkinson's disease treated by hADSCs obtained by the method of the present invention (MPTP + MSCs). The analysis was performed 4 days after intrastriatal injection of hADSCs. LnRNA-COX2 is shown in green color

and hADSCs in red color (marked with PKH26);

- figure 18 is the expression of IL-6 mRNA: mice suffering from Parkinson's disease (MPTP) and mice with Parkinson's disease treated by hADSCs obtained by the method of the present invention (MPTP + MSCs). The analysis was performed 4 days after intrastriatal injection of hADSCs and shows IL-6 being knocked down;

- figure 19A and 19B are the treatment by MPTP and LPS-activated primary microglial cells;

- figure 20 is the real-time RT-PCR of IL6 mRNA, lnRNA COX2 and ln RNA CCND1 (used as negative control) of microglia in culture and treated with LPS or MPTP. The culture medium that has been exposed to hADSC cells obtained from activated fat (SUP), knocks down the production of IL-6 and Ln-Cox2;

- figures 21A-21C are the anti-inflammatory action with cytokines being knocked down exerted by the tissue of the present invention in spinal cord at 48 hours from the traumatic injury;

- figure 22 is the functional recovery (BMS scale) of hind limbs in mice with spinal injury transplanted with lipoaspirated adipose tissue (LS) and with adipose tissue mechanically activated according to the present invention (MALS) compared with a saline treatment. MALS promotes a progressive and significant functional recovery;

- figure 23 is the graft in the injured spinal cord of adipose tissue mechanically activated according to the present invention.

- figure 24 is the treatment of a traumatic injury

at 70Kdyne of the spinal cord, figure 24a a control of the expression of Nestin without treatment, figure 24b the contusion at 70Kdyne, figure 24c the contusion at 70Kdyne + LS and figure 24d the contusion at 70Kdyne + MALS respectively;

- figures 25a to 25d are enlarged details of ependymal stem cells activated by activated adipose cells that is application of MALS;

- figure 26 is the situation after 4 weeks from the activation by MALS;

- figures 27a to 27c are the formation of new neurons in the grey matter of the injured spinal cord due to migration of ependymal stem cells activated by MALS.

The method of the present invention allows adipose tissue removed from the human body to be mechanically activated, particularly mesenchymal stem cells present therein.

According to one of the known techniques, the adipose tissue to be processed by the method of the present invention can be obtained by elective liposuction procedures under local anesthesia.

The final product will be called as lipoaspirated adipose tissue (lipoaspirate).

It is obviously possible to use also other procedures for extracting adipose tissue.

In the present description reference is made to human adipose tissue but obviously the method of the present invention can be applied also to animal adipose tissue.

An example of a procedure involves an infiltration step, where a saline solution containing the vasoconstrictor epinephrine (2 mg/ml) is infused in the adipose tissue to minimize blood loss and contamination of the tissue by cells of peripheral blood.

According to one embodiment of the present invention, for mechanical activation, 10 ml of washed lipoaspirated adipose tissue was harvested in a 50 ml tube and subjected to orbital shaking with a force equal to 97 x g for different time that is 3, 6, 10 and 20 minutes, at ambient temperature.

According to the present invention the anti-inflammatory properties of the adipose tissue, particularly of human adipose tissue containing mesenchymal stem cells, can be activated by applying a specific movement, having a specific force (intensity) and duration.

Considering that accumulation of adipose tissue is actually considered as an inflammatory process, proinflammatory and anti-inflammatory factors have been evaluated in lipoaspirated adipose tissue obtained by the same group of patients and subjected to specific mechanical forces for a variable amount of minutes.

The non processed lipoaspirated fat was used as control.

Such as shown in figure 1, the protein TSG6 is already observable after 3 minutes of stimulation and it has the maximum expression within 6-10 minutes; TNF-alpha is suppressed in 6 minutes.

Experimental results show that the application of

a force of 97 x g ("x g" that is, as known, a multiple of the gravitational force of the Earth):

- eliminates the typical expression of adipose tissue of inflammatory cytokines such as tumor necrosis factor-alpha (TNF-alpha);

- considerably enhances its natural inhibitor TSG6 and interleukin 15, that are present at marginal level in the normal adipose tissue that is not processed tissue,

- considerably improves the expression of leptin, the suppressor of the desire for food produced by fat.

Said improved expression of leptin proves that said force can be also applied in vivo on the surfaces of the body where said hormone plays an important role.

With reference to figure 1 data show that:

- the maximum expression of TSG6 is reached within 6 and 10 minutes of mechanical activation with force 97 x g (Figure 1b),

- the production of TNF-alpha is inhibited within 6 minutes in the same conditions (Figure 1c).

Moreover the maximum expression of beta-tubulin III is reached within 20 minutes of mechanical activation with force 97 x g.

Lower forces are less efficacious.

According to the present invention in order to achieve a promotion of anti-inflammatory properties and an improvement of repair capacity of the lipoaspirated adipose tissue it is possible to apply a force with intensity ranging from 40 to 120 x g.

The mechanical activation of anti-inflammatory

processes is efficacious even when the adipose tissue is cryopreserved at -80°C for several months, such as shown in figure 1D.

When the force 97 x g is applied for three minutes on the adipose tissue that has been cryopreserved, TNF-alpha is cancelled while TSG6 and leptin considerably increase as in fresh tissue.

The simple lipoaspiration procedure produces a soft mechanical activation that is observed by the expression of some pluripotency-related genes (SOX2, OCT4, Nanog), of beta-tubulin III and of TSG6 mRNA, as determined by real-time RT-PCR.

The results obtained in the human adipose tissue from biopsy, in lipoaspirated human adipose tissue, and human adipose tissue lipoaspirated and mechanically activated (10 minutes) are shown in figure 2.

Figure 2 shows the expression of pluripotency genes (SOX2, OCT4, Nanog), beta-tubulin III and TSG6 tested with real-time RT PCR in samples from biopsy of human adipose tissue, lipoaspirated and lipoaspirated mechanically activated by the method of the present invention (10 minutes). All the investigated genes are hardly activated in the regular bioptic tissue, that is non processed tissue, according to the present invention (biopsy), and are slightly activated by performing the known Coleman procedure after liposuction (lipoaspirate). Differently the upregulation is incomparably higher after the specific activation according to the present invention for 10 minutes.

All the investigated genes are hardly active in the regular bioptic tissue, not mechanically manipulated according to the present invention, and such activation is slightly improved by the known Coleman procedure after liposuction (lipoaspirate). However the application of the specific mechanical force, here described and claimed, for 10 minutes causes an incomparably higher up-regulation.

Pluripotency regulator genes SOX2, NANOG and OCT4 are completely activated within 6 minutes of mechanical activation with force 97 x g.

Therefore in addition to anti-inflammatory properties, also the parameters that define stemness of mesenchymal stem cells present in the adipose tissue are increased by the mechanical activation, in a manner independent from force and time (Figure 3).

It is known that Sox2 has a critical role in the maintenance of embryonic and neural stem cells.

Furthermore it has been noted that (Go et al., 2008) Sox2 is not only essential for pluripotency and self-renewal of embryonic stem cells, but is also expressed in somatic stem cells with superior expansion and differentiation potential (Vai et al., 2008).

Sox2 is a transcription factor (34kDa) containing a HMG box for DNA binding and it forms trimeric complexes with Oct4 controlling the expression of a given number of genes.

According to the present invention the specific mechanical activation of the adipose tissue directly regulates the gene expression guided by the

transcription factor SOX2 on the region of the promoter of different genes coding for the above described cytokines.

Moreover Chip-Seq on immunoprecipited SOX2 shows a high binding efficiency of SOX2 on the promoter IL-6 in glioblastoma or ES cells (Fang et al., 2011). The region of the promoter -3000bp of beta-tubulin III, TSG6, TNF-alpha, IL1B, IL10, IL15 and leptin was analysed in silico, to search for consensus sequence for SOX2 and ChIP tests (chromatine immunoprecipitation) were designed to detect binding of the transcription factor in liquid phase. The Chip test shows that SOX2 and HuR bind to the region of the promoter of beta-tubulin III, IL15 and TSG6 and that the mechanical application of 97 x g regulates in differential manner such interaction (figure 4).

SOX2 does not bind anymore the beta-tubulin III promoter after 20 minutes of application of the above mentioned force, therefore the transcription of beta-tubulin III is improved.

The increased transcription of TSG6 is regulated by the loss of HuR binder, while IL15 is likely regulated by other factors different from Sox2 and HuR.

Stem cells from adipose tissue were purified from human fat harvested through microliposuction under tumescent local anesthesia by Coleman microcannulas.

It is obviously possible to use also other procedures for removing the adipose tissue.

Following this procedure, the lipoaspirated adipose tissue was centrifuged in a sterile closed

system at 2000 rpm for 10 minutes and the pellet was subjected to collagenase digestion for half an hour at 37°C.

The digest is centrifuged for separating the floating population of mature adipocytes from the stromal vascular fraction (SVF).

After 24 hours plating in a standard medium (Dulbecco Modified Eagle Medium supplemented with 10% fetal bovine), non-adherent cells were removed.

The plastic-adherent cell population define the stromal cells derived from adipose tissue (ADSCs).

Starting from the adipose tissue subjected to a force 97 x g, an efficacious method, that avoids enzymatic treatment, and reproducible for isolation and expansion of hADSCs (human adipose derivated stem cells) has been developed by modifying the methods described by Matsumoto et al., 2007 and Perrini S. et al., 2013.

The adipose tissue lipoaspirated and treated with a force of 97 x g was placed in 25cm^2 culture flasks (NUNC, Roskilde, Denmark) completely filled with culture medium. This allows the tissue to adhere to plate top.

After 2 weeks of culture the adipose tissue was removed, the flask turned upside down and cells were kept in culture.

Cells reached a confluence of 90% in 30 days.

The yield of cells was about 5.5-6 x 105 cells for each plate containing 2 ml of mechanically treated adipose tissue.

Morphology in vitro (captured by means of EVOS® microscope apparatus, AMG, USA) of these cells exhibits a fibroblast-like phenotype comparable to that of cells obtained by means of classical methods (Figure 5A and 5b).

Such mesenchymal stem cells have the typical expression of beta-tubulin III (TUJ1) and vimentin (Figure 6).

hADSCs cells (human adipose derivated stem cells) isolated by means of the protocol described above express mesenchymal markers according to FACS analysis (figure 7) with high values (90-100%) for the known typical mesenchymal markers (such as CD44, CD73, CD90 and CD166), a mean value for endothelial tissue (CD31, CD34, CD144, CD146, KDR) and a minimal representation for hematopoietic tissue (CD45, CD133, CD56) (Figure 7). This can suggest that these cells can also be subjected to endothelial fate.

In order to study the chromosomal stability of hADSCs obtained from the adipose tissue mechanically activated according to the present invention, a QFQ banding has been carried out. Cells did not present any chromosomal rearrangement and the chromosome number was normal in 9 passages in culture (Figure 8A and 8B).

The generation of spheroids is normally limited to neural stem cells, here it has been demonstrated that also hADSCs from fat activated according to the present invention form neurospheres.

hADSCs (human adipose derivated stem cells) were incubated with serum-free medium usually used for

forming neurospheres (Gritti et al., 2002).

The test forming the spheres was carried out with 5x103/cm^2 plating. About 50% of hADSCs obtained from mechanically activated adipose tissue, according to the present invention, formed large spheroids reaching an average diameter of 100 micron at 21 culture days (Figure 9).

About 50% of hADScs obtained from mechanically activated adipose tissue formed large spheroids reaching an average diameter of 100 micron at 21 culture days, and forming a mean of 100.95 ± 10.89 spheres/cm^2 within 21 days (Figure 9).

Adult murine neural precursor cells (NPCs) obtained from the subventricular zone were used as positive control: in this case the formation of spheres was obviously higher (Figure 9).

The involvement of SOX2 in the regular expression of cytokines mentioned above was also studied in cells isolated from adipose tissue treated with a force 97 x g.

Chip assay shows that SOX2 binds with the promoting region of beta-tubulin III, IL15 and TSG6 (Figure 10). Thus the activation in vivo is maintained also by hADSCs derived from the method of the present invention.

Recent reports suggest that MSCs (Mesenchymal stem cells) are able to modulate inflammation (Prockop et al., 2013).

THP-1 is a category of rounded and single cell suspension, with distinct monocytes markers resembling

primary monocytes and macrophages in morphology and differentiation property (Tsuchiya et al., 1980).

Interleukyn-1β (IL-1β) is a powerful mediator of inflammation and immune response, mainly produced by activated monocytes (Dinarello 1994).

The stimulation of monocytes with bacterial lipopolysaccharide endotoxin (LPS) leads to the production of IL-1β by activation of the pro-IL-1 gene, expression of pro-IL-1β mRNA, and following translation of pro-Il-1β message.

Pro-IL-1β is processed to the mature and biologically active IL-1β of the cysteine protease interleukin-1 converting enzyme (Kostura et al., 1989).

Anti-inflammatory action of hADSCs obtained from mechanically activated adipose tissue was evaluated in a co-culture test (trans-well®) with THP1 cells activated with LPS (10 micrograms/ml; for 3 hours).

As shown in figure 11 the release of IL-1β is maximum after 3 hours from the stimulation with LPS, and the presence of hADSCs obtained from mechanically activated adipose tissue considerably reduced the secretion of chemokine (Figure 11).

A similar experiment based on the release reaction of IL-1beta from THP1 monocytes activated by LPS ((10 micrograms/ml; for 3 hours) (CTRL+) was performed to compare the action of hADSCs obtained from activated adipose tissue (cell 10) with those from lipoaspirated adipose tissue (LS).

Figure 12 shows that THP1 releases IL-1beta and such release is doubled by the addition of LPS. The

increase in the release of IL-1beta is eliminated by mesenchymal stem cells derived from adipose tissue mechanically activated according to the method of the present invention, while those from lipoaspirate have a negligible marginal effect (Figure 12).

Anti-inflammatory activity of hADSCs from activated adipose tissue according to the method of the present invention is also highlighted from the following experiments shown below.

Figure 13 shows that monocytes activated by LPS (THP1 + LPS) express high levels of inflammatory cytokines, also in presence of stem cells derived from lipoaspirate (LS); such levels drastically are reduced when such monocytes are cultured in presence of hADSCs obtained from adipose tissue mechanically activated by the method of the present invention (Figure 13). It also demonstrates that the production of anti-inflammatory peptide TSG6 is strongly implemented by hADSCs derived from adipose tissue activated by the method of the present invention.

The present invention relates to hADSCs that is mesenchymal stem cells obtained from human or animal adipose tissue mechanically activated by the method of the present invention for the use as a medicine.

In particular they are applied in cell therapy both for human beings and for animals.

The present invention relates also to hADSCs that is mesenchymal stem cells obtained from human or animal adipose tissue mechanically activated by the method of the present invention for the use in the treatment of

motor or coordination disorders, for example in patients suffering from Parkinson's disease.

The recovery of motor dysfunctions, in a Parkinson's disease model was investigated by means of horizontal and vertical grid tests ("horizontal grid test" and "vertical grid test").

The graph shown in figure 14 shows that mice rapidly loose coordination of forelimbs and hind limbs, after exposition to MPTP, such dysfunction is consolidated at 10^{th} day post-injury, when the cell intrastriatal graft is performed.

3 days after the transplantation (13^{th} day) hADSCs obtained by the method of the present invention already cause a gradual improvement in limb coordination, showing a decrease in the percentage of movement errors of forelimbs, that after 10 days is 60-70% in MPTP animals and about 25% in animals treated with hADSCs obtained from adipose tissue mechanically activated by the method of the present invention.

These positive results were observed also by performing the "vertical grid" test.

A considerable longer time was detected in MPTP treated mice with respect to the control (Figure 15) for turning around and climbing down on the vertical grid.

Mice treated with hADSCs obtained from adipose tissue mechanically treated by the method according to the present invention show an improvement already 3 days after the injection (Figure 15).

Performances of animals considerably improved also

in the following days.

It is known that microglia cells, the resident innate immune cells in the brain, have been implicated as active contributors to neuron damage in neurodegenerative diseases, in which the overactivation and dysregulation of microglia might result in disastrous and progressive neurotoxic consequences.

Inflammation is a base component of a wide range of neurodegenerative diseases and of their associated neuropathology, and more and more evidences suggest that microglia is a causal key factor in such process.

Activated microglia is present close to degenerating neurons in substantia nigra of patients suffering from Parkinson's disease and patients suffering from other Parkinsonian disorders (McGeer et al., 1988, Langston et al., 1999, Inamura et al., 2003).

Activation of microglia in such disease is not limited to substantia nigra, but it is found also in putamen, hippocampus, transentorhinal cortex, cingulate cortex and temporal cortex (Inamura et al.,2003).

The selective loss of dopaminergic neurons (DA) in substantia nigra may be due to many negative events, such as depletion of glutathione for dopaminergic neurons (resulting in a reduced antioxidant effect), high content of DA (redox active molecule) in neurons of substantia nigra (Zigmund et al., 2002), high iron concentrations (Zecca et al., 2004) (redox active elements) and an increase of microglia in substantia nigra (Lawson et al., 1990) with respect to other

regions.

Therefore dopaminergic neurons in substantia nigra could be particularly vulnerable to inflammatory attacks due to their precarious redox balance and colocalization with a large microglia population.

The transplantation in mice of hADSCs obtained from adipose tissue mechanically activated by the method of the present invention changes the genetic profile of mouse brain.

Particularly the effect of transplantation on the regulation of long non-coding RNA involved in inflammatory processes, such as lincRNA-Cox2 was observed (Gomez et al., 2013).

The mammalian genome "junk DNA" encodes many thousands of large non-coding transcripts, including a class of large intergenic non-coding RNAs (lincRNAs), few among them have been functionally characterized for their biological role by Guttman and contributors (Guttman et al., 2011) that performed loss-of-function studies on lincRNAs expressed in mouse embryonic stem cells (ESCs) and characterized the effects on gene expression.

Recent studies have found thousands of long non-coding RNAs (LncRNAs) in mammalian genomes (Birney et al., 2007, Guttman et al., 2009-2012) that regulate the gene expression in different biological processes (Guttman et al., 2011). LncRNAs are differentially regulated in virus-infected cells and in dendritic cells following lipopolysaccharide (LPS) stimulation.

The lincRNA-COX2 expression was studied by RT-PCR,

in a different area of the brain (cortex, striatum, hippocampus of animals with MPTP-induced Parkinson's disease and 4 days after injection of stem cells). The expression of lncRNA-COX2 is induced in mice suffering from Parkinson's disease and its expression seems to be related with the presence of HuR as demonstrated by its co-immunoprecipitation in RIP assay (RNA immunoprecipitation) performed on homogenized striatum.

The expression of lncRNA-COX2 is deeply down-regulated by intrastratial injection of hADSCs obtained from adipose tissue mechanically activated by the method of the present invention (MPTP/MSC; figure 16A-D). Down-regulation of lncRNA-COX2 is strictly related to the recovery of the function by the animal, such as shown by behavioral tests (figures 16A-D).

The expression of lncRNA-COX2 was investigated also by FISH (Fluorescent in situ hybridization) an approach able to detect and localize specific RNA targets (mRNA, lncRNA and miRNA) in cells, circulating tumor cells, and tissue samples (Figure 17).

Recently it has been demonstrated that linRNA COX2 up-regulates the expression of IL-6, by acting on its promoter (Carpenter et al., 2013).

Levels of expression of IL-6 mRNA have been studied by real-time RT-PCR in cerebral areas of mice suffering from Parkinson's disease and mice affected by Parkinson's disease treated by hADSCs of the present invention.

As shown in figure 18 the expression of IL-6 is considerably increased and it is deeply opposed by the

intrastriatal injection of hADSCs of the present invention.

The mechanism with which hADSCs can down-regulate the expression of lincRNA-COX2 after having been grafted in the damaged brain of the recipient and can guide the recovery process has been studied.

LincRNA-COX2 has been studied in experiments in vitro on primary glia activated with MPTP.

The expression of lnc-RNA-COX2 has been studied by RT-PCR and it has been induced by the MPTP treatment of primary glia cultures (3 hours at 37°).

FISH analyses confirmed that lncRNA-COX2 is expressed in positive GFAP cells (that is glial cells) and its cell localization is related to the HuR cell distribution (RNA immunoprecipitation).

The RNA binding protein HuR/ELAVL1 binds to AU-rich elements (AREs) promoting the stabilization and translation of a given number of mRNAs into the cytoplasm, dictating their fate (D'Agostino et al., 2013).

The performed experiments show that the interaction with HuR seems necessary also for the activation of microglia, as denoted by IBA1 staining.

IBA1 is a protein specifically expressed in macrophages/microglia and is upregulated in microglia following nerve injury, central nervous system ischemia, and several other brain diseases. Incubation of glial cell primary cultures with MPTP or LPS for 3 hours considerably enhances staining with specific anti-IBA1 antibodies (Figures 19A-B).

The treatment with diidrotansione (DI), an inhibitor of HuR binding RNAs that leads to its cytoplasm mislocalization causes the lnRNA-COX2 down-regulation in cells treated with MPTP or LPS (Figures 19A-B). Such reduction corresponds to the contraction of IL-6 mRNA increase (Figures 19A-B).

Also BAY11-7083 treatment, a specific lnRNA-COX2 inhibitor by blocking NFkB, opposes the increase of levels of IL6 mRNA.

Moreover in the same experimental conditions, microglial cells were incubated with the supernatant of hADSCs cultures.

Figure 20 shows that the supernatant conditioned with hADSCs, derived from the mechanically activated fat, is able to particularly oppose the expression of lnRNA COX2 and IL-6 in MPTP or LPS activated primary glial cells.

The expression of lnRNA CCND1, a long non coding RNA referred to the promoter of cyclin D1, used as negative control, was not modified in any of the experimented conditions.

The recovery of motor functions of hind limbs after spinal injury was observed in mice after the transplantation of mechanically activated adipose tissue (MALS), while the lipoaspirate is inefficacious (figures 21 and 22).

The spinal injury, induced by a contusion made by an impactor with a diameter of 1 mm that hits the exposed thoracic spinal cord at level T9 with a force of 70 Kdyne for 1 second, causes paralysis of the hind

limbs that partially improves, less than 3 points according to BMS (Basso Mouse Scale) (figure 22). The transplantation of lipoaspirated fat (LS) does not cause considerable improvement, while the treatment with MALS causes a great improvement exceeding 6 points according to BMS. 9 points means a normal ambulation.

The mechanically activated adipose tissue (MALS) of the present invention was applied on the surface of the injured spinal cord, where it gradually penetrates while filling the injury area and by joining the two portions of the injured spinal cord and allowing for the nerve regeneration.

The anti-inflammatory action of the mechanically activated adipose tissue (MALS) is rapid and it suppresses the production of inflammatory cytokines in the injured spinal cord. After 48 hours from the injury, the factors released from the injured spinal cord knock down the production of Il-1 alpha, IL-1 beta and IL-6 in the injury site and in caudal and rostral portions of the injured spinal cord.

From what described above and with reference to the figures, it is clear how the application of a orbital shaking force with an intensity and for the time described and claimed in the present invention allows anti-inflammatory properties and stem properties of mesenchymal stem cells present in the adipose tissue lipoaspirated or removed with any other procedure from the human body to be activated with a minimal manipulation of cells.

The adipose tissue and/or stem cells of the

present invention can be therefore used for therapeutic purposes, or also cosmetic purposes, for preparing medicines to be used during cell therapies.

The tissue and/or mesenchymal stem cells obtained by the method of the present invention can be:

- used as a medicine,

- used for treating motor and coordination dysfunctions,

- used for treating Parkinson's disease,

- used for treating spinal cord injuries.

In particular the transplantation of tissue and/or mesenchymal stem cells derived from human adipose tissue hADSCs, mechanically activated as described and claimed herein, can promote the rapid recovery of motor functionalities in patients that have lost or reduced such mobility, such as for example patients suffering from Parkinson's disease. Moreover the activated adipose tissue is very efficacious in promoting functional recovery after spinal cord injury due to its anti-inflammatory properties and to its ability to penetrate in the injury and to promote nerve regeneration.

Experimental procedures

Cell culture

Two ml of mechanically activated adipose tissue were inoculated in 25 cm^2 flasks completely filled with culture medium known as "Dulbecco's modified Eagle's Medium" (DMEM, Invitrogen, Carlsbad, CA) supplemented with 20% fetal bovine serum (FBS, JRH Bioscience, Lenexa, KS, Lot 6G2146) and incubated at 37°C in 5%

CO_2. Flasks were turned upside-down such that cells floated up and adhered to the top inner ceiling surface of the flask. After 15 days, the medium and tissue were removed and the flasks were turned upside-down so that the cells were oriented on the bottom. The medium was replaced every 4 days until the cells reached confluence (90-95% after about 30 days).

Histological control analyses and lipoaspirate mechanically treated tissues

Histological analyses were performed on fresh adipose tissue (with or without mechanical treatment) on samples from the same subject. Briefly, formalin-fixed paraffin-embedded tissue samples were processed for conventional histopathological examination and immunohistochemistry. Standard 4 micron thick tissue sections stained with hematoxylin and eosin (H&E) were examined by direct wide field light microscopy. For immunohistochemistry, four paraffin samples were sectioned (4µm thickness), de-paraffined, and re-hydrated in xylene and graded concentrations of ethanol to distilled water. Then they were rinsed with PBS, treated with blocking solution (PBS + 1% V/V fetal bovine serum for 1 hour at room temperature) and incubated with primary antibodies overnight at 4°C. After treatment with primary antibodies, the sections were washed with PBS and incubated with appropriate secondary antibodies. Endogenous peroxidase activity was quenched with 3% hydrogen peroxide in distilled water for 10 min. Staining was performed with 3,3′ diaminobenzidine (DAB) as a chromogen (DAKO EnVision

detection kit). In control determinations, primary antibodies were omitted and replaced with equivalent concentrations of unrelated IgG of the same subclass. The following primary antibodies were used: Vimentin (1:5000; Dako Cytomatic); S100 (1: 2000; Novocastra); β-Tubulin III (1:6000; Covance); human Nestin (1:2000; Millipore); Nanog (1:1000; Novus).

For immunofluorescence studies, cells were washed with PBS, fixed with paraformaldehyde (4% in PBS) treated with blocking solution and incubated with primary antibody anti β-Tubulin III (1:6000; Covance) overnight at 4°C.

After treatment with primary antibody, sections were washed with PBS and incubated with secondary antibody (Alexa Fluor® 555 goat anti-mouse 1 : 200 Molecular Probes®, Invitrogen, Life Technologies Italia, Monza, Italy) for 2 hours at room temperature. Nuclei were stained with DAPI (Hoechst 1/1000) and then mounted using the FluorSave Reagent (Calbiochem, Merck Chemical, Darmstadt, Germany) and analyzed by immunofluorescence microscopy (Leica 5500). As negative reference for the confocal analysis we used a consecutive section that was stained by omitting primary antibody anti β-Tubulin III and replacing it with equivalent concentrations of unrelated IgG of the same subclass.

Extraction of RNA and qRT-PCR analysis

Cell RNAs were extracted using TRI Reagent® (Sigma-Aldrich, St. Luis, MO, USA) according to manufacturer's instructions. RNA purity and quantity

was assessed by Nanodrop (Fisher Scientific) (A260/A280 1.8–2 was considered suitable for further analysis), possible contaminating DNA was removed and cDNA was prepared from 1 mg of RNA using "high Capacity RNA-to-cDNA" Kit (Applied Biosystems, Foster City, CA, USA).

Gene expression of adipose tissues

Quantitative results were obtained by the real-time SYBR-Green assay (Euroclone spa Milano, ITALIA) on total RNA extracted using TRI Reagent® (Sigma-Aldrich, St- Luis, MO, USA) treated with DNasi I – Rnasi free (Ambion inc. Foster City, CA, USA) and reverse-transcripted using the "high Capacity cDNA Reverse kit Transcription" kit (Ambion Inc. Foster City, CA, USA) as suggested by the manufacturer. Differentially expressed genes were recognized by applying a significance threshold on t test at unequal variance.

ChIP on adipose tissue

Chromatin Immunoprecipitation was performed on 1 ml of mechanically treated or not treated lipoaspirated adipose tissue, as described in Haim et al., 2013.

G-banding karyotype Analysis

Cytogenetic analyses were performed on "in situ" cultures obtained by inoculating hADSCs directly into a coverslip inside Petri dishes containing 2 ml of medium (αMEM). Cells were treated with Colcemid (0.02 ml/ml) (Life Technologies a Carlsbad, California, United States of America) for 90 minutes, hypotonic solution (1:1 Na citrate 1%: NaCl 0,3%) (Sigma-Aldrich, St. Louis, MO, USA) and fixative solution of 3:1 methanol:acetic acid (VWR International Radnor,

Pennsylvania, USA), replaced twice. At least twenty-five QFQ banding metaphases were observed for each sample. The image was acquired using a fluorescence microscope (BX 60 Olympus) and analyzed with Powergene PSI system.

Formation of spheroids

hADSCs derived from mechanically activated lipoaspirated tissue were cultured in alpha-MEM medium with fetal bovine serum (20%) to reach 85% confluence at passage 1. Alpha-MEM medium was removed cells were washed twice with sterile PBS, therefore medium for neurospheres was added containing bFGF (20 ng/ml) and FEG (10 ng/ml) with no serum (Gritti et al., 2002).

Cell culture THP-1 culture cells and activation with LPS

THP-1 cells (202-TIB; ATTC, Rockville, MD) were grown in suspension in RPMI 1640 media supplemented with 10% fetal calf serum (FCS), 50 U/mL penicillin, and 50 µg/mL streptomycin in 150 cm^2 flasks. Cell cultures and all experiments were carried out at 37°C in 5% CO2 and 95% room air. Cells were passaged every 3 days and used between passages 3 and 12. Anti-inflammatory action of hADSCs cells was analysed in trans-well® culture system on 6 plates.

hADSCs were plated in well below at density of 1.5/104 cells/well in alpha-MEM medium. Twenty-four hours after their adhesion THP-1 cells were added in the well above at density of 75 x 104 cells/well. To induce release of IL-1 beta, lipopolysaccharide (LPS) was administered at the final concentration of 10

micrograms/ml. After three hours of incubation, supernatants were harvested in ice and immediately frozen. Release of IL-1 beta was studied by immunoblotting.

Experimental animal model for Parkinson's disease and cell injection

MPTP (1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine) is a neurotoxin precursor to MPP+, which causes permanent symptoms of Parkinson's disease by destroying dopaminergic neurons in the substantia nigra of the brain. It has been used to study disease models in various animal studies.

Injection of MPTP causes rapid onset of Parkinsonism, hence users of MPPP contaminated with MPTP will develop these symptoms. After two months, C5BL/6J mice were subjected of a first intraperitoneal injection of MPTP (36 mg/kg). Motor dysfunctions were evaluated every 2 days for the following 7 days, then animals were subjected to a second injection (20mg/kg). After 3 days (day 10) cells were injected by stereotaxic surgery into the striatum. Coordinates, in relation to bregma, were: 0.1 mm posterior, 2.4 mm mediolateral, and 3.6 mm dorsal. Control group was injected only with 5 ml of sterile PBS or with hADSCs obtained from mechanically activated adipose tissue.

Labeling of PKH26 cells

"PKH26 Cell Linker" Kits provide fluorescent labeling of live cells over an extended period of time, with no apparent toxic effects. "Fluorescent Linker" Kits are effective for a variety of cell types and

exhibit no significant leaking, or transfer from cell to cell. They provide stable, clear, intense, accurate and reproducible fluorescent labeling of cells. For transplantation experiments 1x105 cells were injected in each animal.

Before each transplantation hADSCs were labelled with PKH26 and staining was performed according to the manufacturer protocol (Sigma-Aldrich ®). For the injection PKH26 positive cells were dissolved in sterile PBS at a concentration of 1 x 105 cells/5µl.

Evaluation of motor dysfunctions

Changes in the level of motor dysfunction can be used as a non invasive manner for evaluating alterations in the number of DArgic neurons and/or in the amount of DA in PD animal models, as mice systemically administered with 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP). In this study DA-associated motor dysfunctions were evaluated by the "vertical grid test" and "horizontal grid test".

In the "horizontal grid test" device once the mouse firmly grabbed on the grids with all four paws, the apparatus was turned upside down and the animal was videotaped for a maximum duration of 30 seconds.

With each step, the paw may fall or slip between the wire of the grid. This was recorded as a "foot fault". The total number of steps (movement of each forelimb) that the mouse used to cross the grid was counted, and the total number of foot faults for each forelimb were recorded.

The Vertical grid test was performed by placing

the mouse 3 cm from the top of a vertical grid, facing upward, and was videotaped while it turned around and climbed down the grid. The time required to climb down starting from said top edge was measured.

The recovery of motor functions of hind limbs after spinal cord injury, induced by a contusion made by an impactor with a diameter of 1 mm that hits the exposed thoracic spinal cord at level T9 with a force of 70 Kdyne for 1 second, was evaluated by observing the use of hind limbs in the "free locomotion" test. The used scale (BMS, Basso Mouse Scale) describes the several recovery levels of motility from 0 that means a paralysis of hind limbs to 9 that means a normal locomotion. The value reached by the treatment with saline and lipoaspirate was about 3 points (plantar positioning of the paw with poor ability in supporting the body) while MALS led to overcome 6 points in the scale corresponding to a frequent or consistent plantar positioning and mainly coordinated positioning of the paws with support of the body.

Figures 24 to 27 show a further embodiment of the application of the invention wherein activated adipose cells that is MALS allow endogenous stem cells of the spinal cord to be activated for the treatment of the spinal cord. As it is clear in presence of an injury, as a contusion of the spinal cord at 70 Kdyne. Figure 24d shows the contusion treated with MALS. Figures 25a to 25d show enlarged details of ependymal stem cells activated by activated adipose cells that is the application of MALS.

After 4 weeks from the activation by MALS the treatment provided the results visible in figures 26 and 27. It is noted the formation of new neurons in the grey matter of the injured spinal cord due to the migration of ependymal stem cells activated by MALS.

CLAIMS

1. Method for promoting anti-inflammatory properties and for improving the repair ability of a tissue containing mesenchymal stem cells (MSCs) characterized in that it comprises at least one step of mechanical activation performed by applying a orbital shaking force on a adipose tissue lipoaspirated or removed from the body, characterized in that the applied force has an intensity ranging from 40 to 120 x g, characterized in that said force is applied for a time ranging from 3 to 20 minutes.

2. Method according to claim 1 characterized in that the applied force has an intensity equal to 97 x g.

3. Method according to one or more of the preceding claims, characterized in that said force is applied for 10 minutes.

4. Method according to one or more of the preceding claims, characterized in that said mechanical activation step applying an orbital shaking force is repeated two or more times with an equal or different force intensity for each step and/or with an equal or different force application duration for each step.

5. Method according to one or more of the preceding claims, characterized in that said lipoaspirated adipose tissue is cryopreserved.

6. Method according to one or more of the preceding claims, characterized in that it provides a step isolating and expanding the mesenchymal stem cells

contained in said adipose tissue lipoaspirated or removed from the body (ADSCs).

7. Method according to one or more of the preceding claims, characterized in that the step isolating and expanding the mesenchymal stem cells contained in said adipose tissue lipoaspirated or removed from the body (ADSCs) comprises:

- inoculation of 2 ml of mechanically activated adipose tissue in 25 cm^2 flask completely filled with culture medium "Dulbecco's modified Eagle's Medium" (DMEM) supplemented with 20% fetal bovine serum;

- incubation at 37°C in 5% CO2;

- turning upside-down the flask such that cells float up and adhere to the top inner ceiling surface of the flask;

- removing the culture medium and tissue after 15 days and turning upside-down the flask so that the cells are on the bottom;

- replacing the culture medium every 4 days until reaching cell confluence of 90-95%.

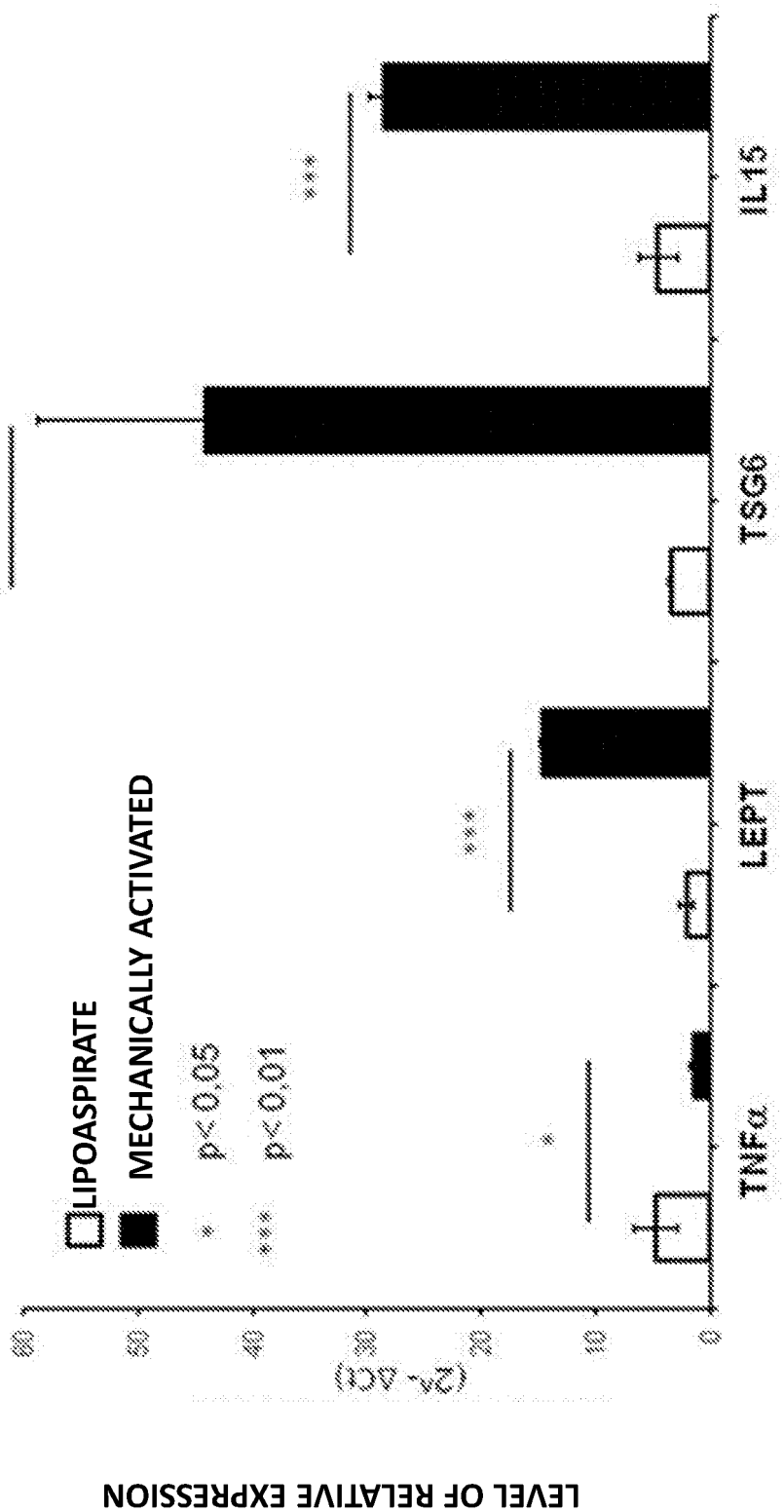

LEVEL OF RELATIVE EXPRESSION
(2^- ΔCt)
0
20
40
60
80
LIPOASPIRATE
MECHANICALLY ACTIVATED
* p<0,05
*** p<0,01
TNFα
LEPT
TSG6
IL15
***
***
*
***

Fig.1A

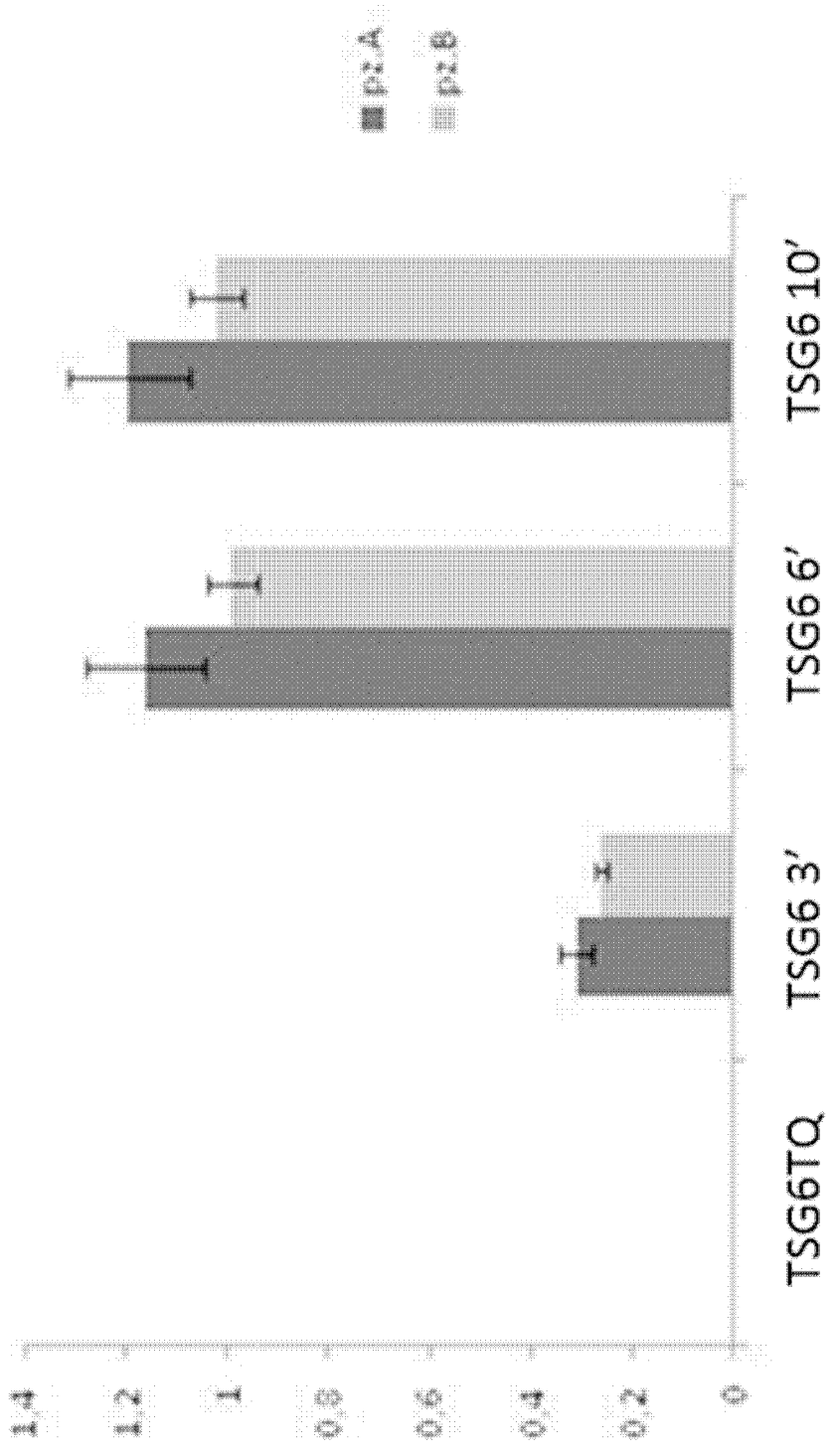

p2.A
p2.B
1.4
1.2
1
0.8
0.6
0.4
0.2
0
TSG6TQ
TSG6 3'
TSG6 6'
TSG6 10'
RELATIVE EXPRESSION

Fig.1B

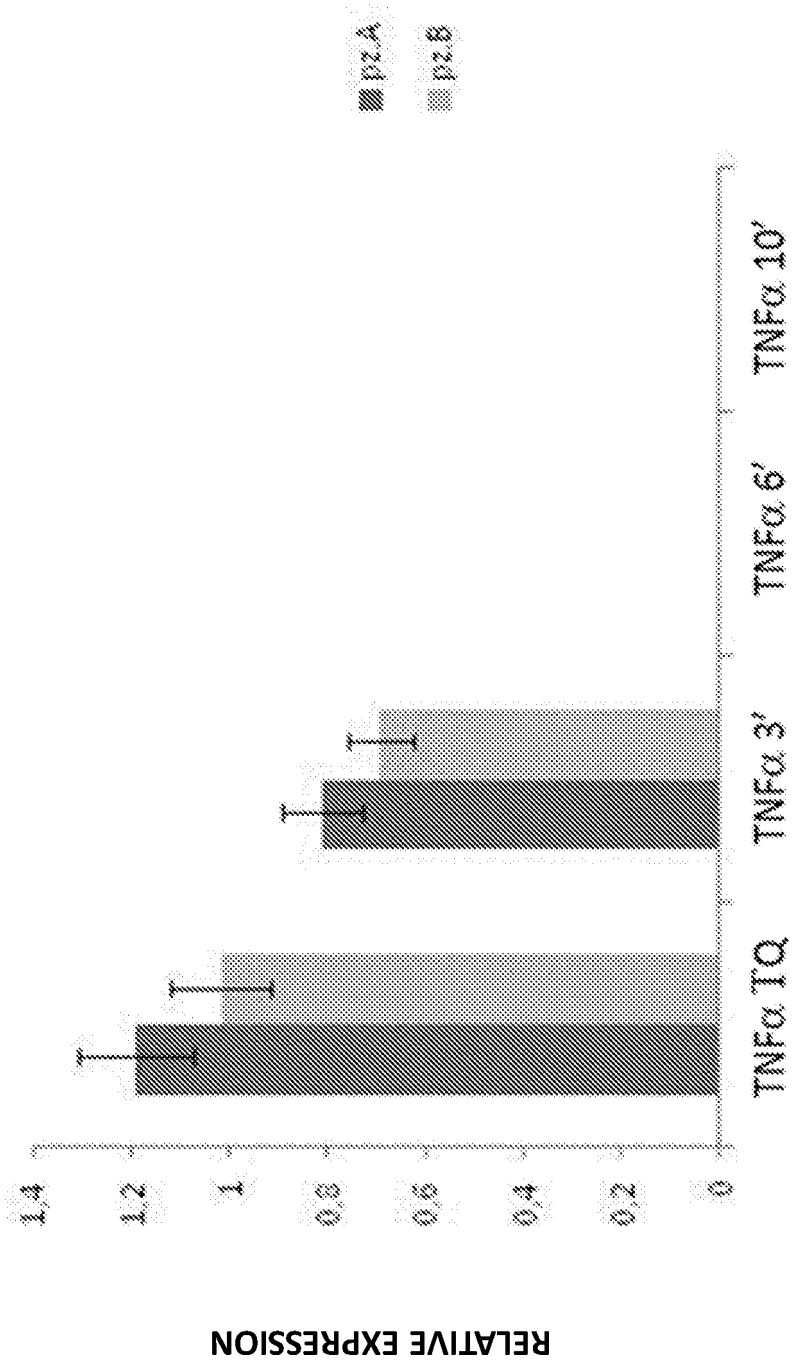

p2.A
p2.B
RELATIVE EXPRESSION
1,4
1,2
1
0,8
0,6
0,4
0,2
0
TNFα TQ
TNFα 3'
TNFα 6'
TNFα 10'

Fig.1C

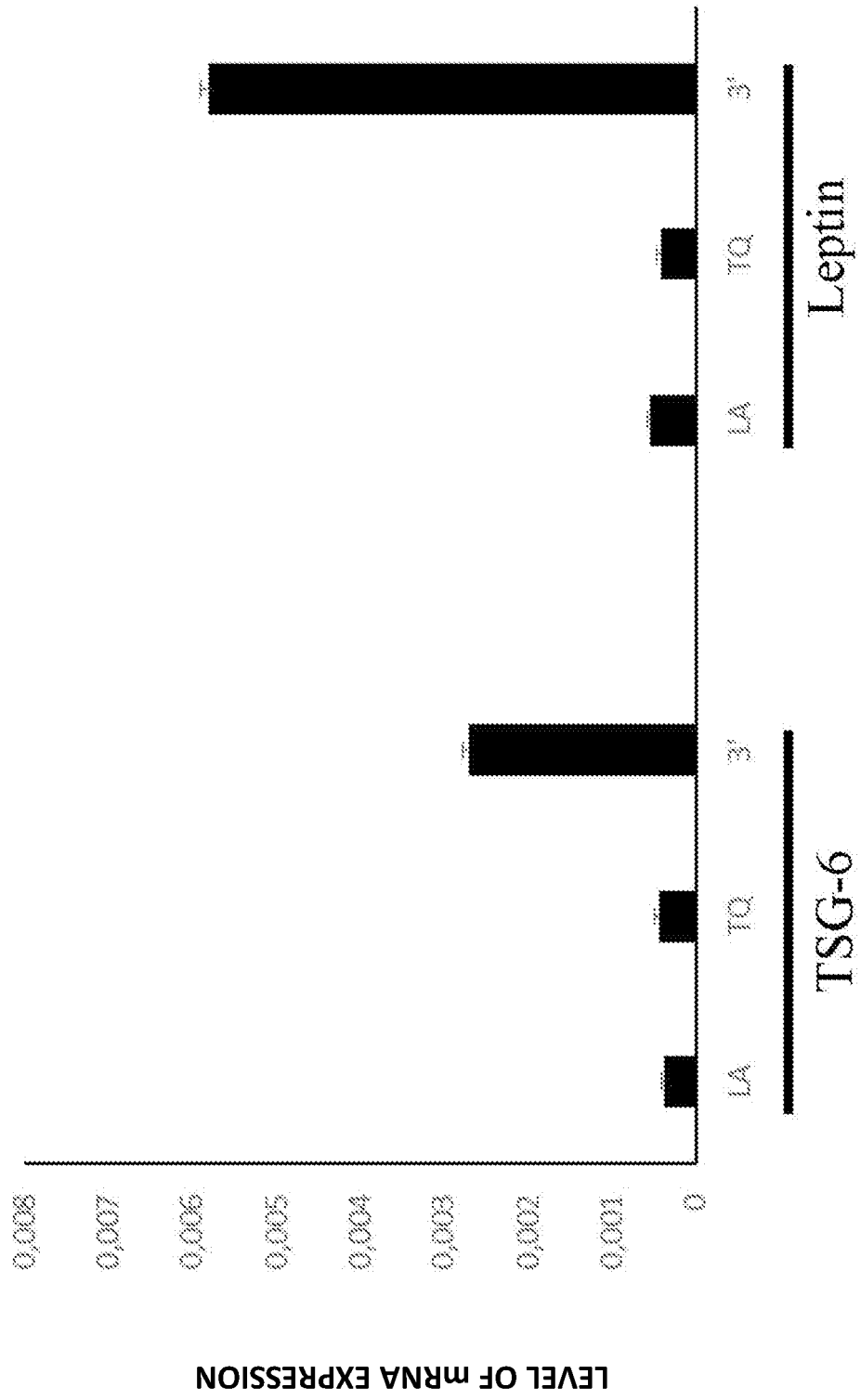

LEVEL OF mRNA EXPRESSION
0,008
0,007
0,006
0,005
0,004
0,003
0,002
0,001
0
LA
TQ
3'
TSG-6
LA
TQ
3'
Leptin

Fig.1D

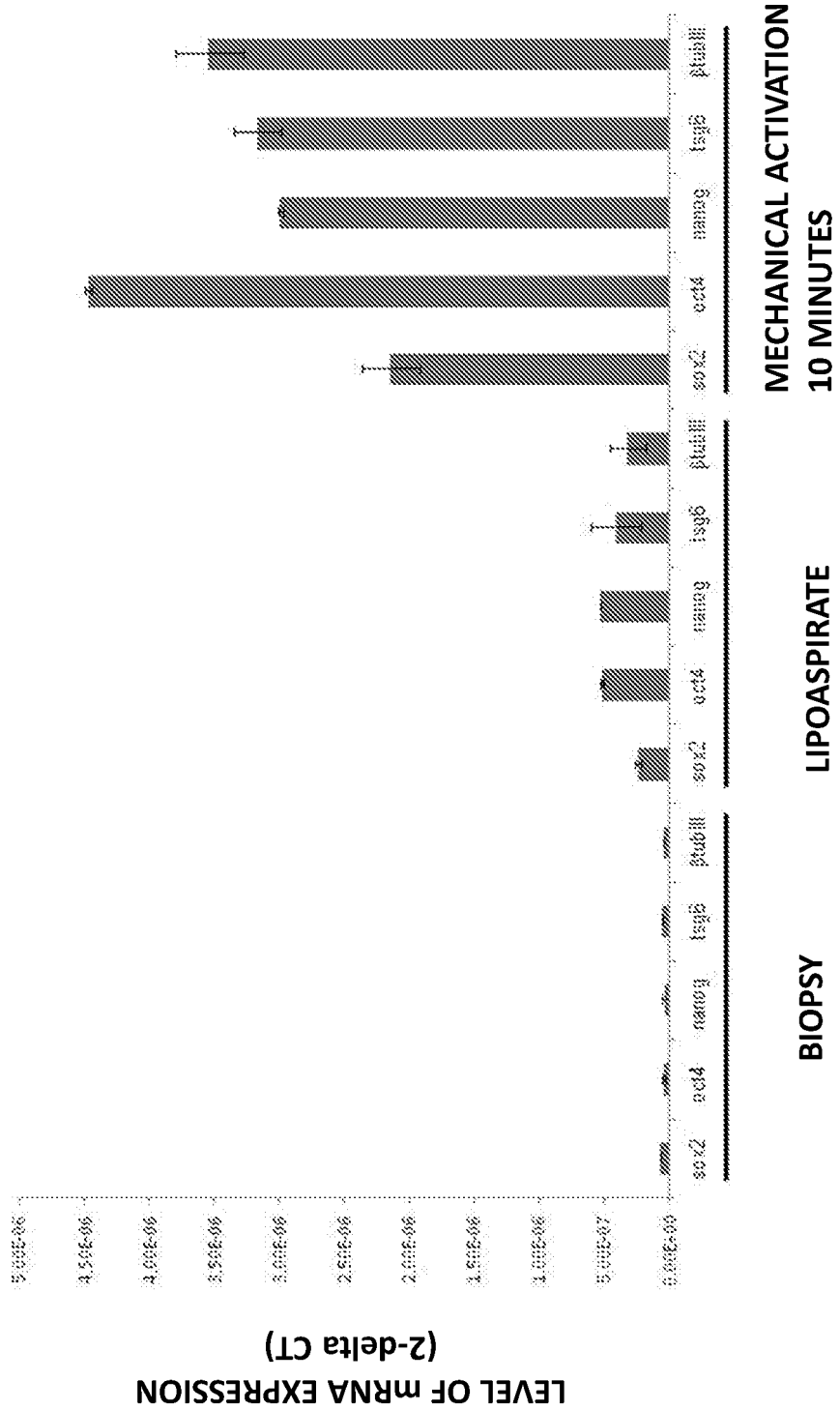

LEVEL OF mRNA EXPRESSION
(2-delta CT)
BIOPSY
LIPOASPIRATE
MECHANICAL ACTIVATION
10 MINUTES
sox2
oct4
nanog
tsg6
βtubIII

Fig.2

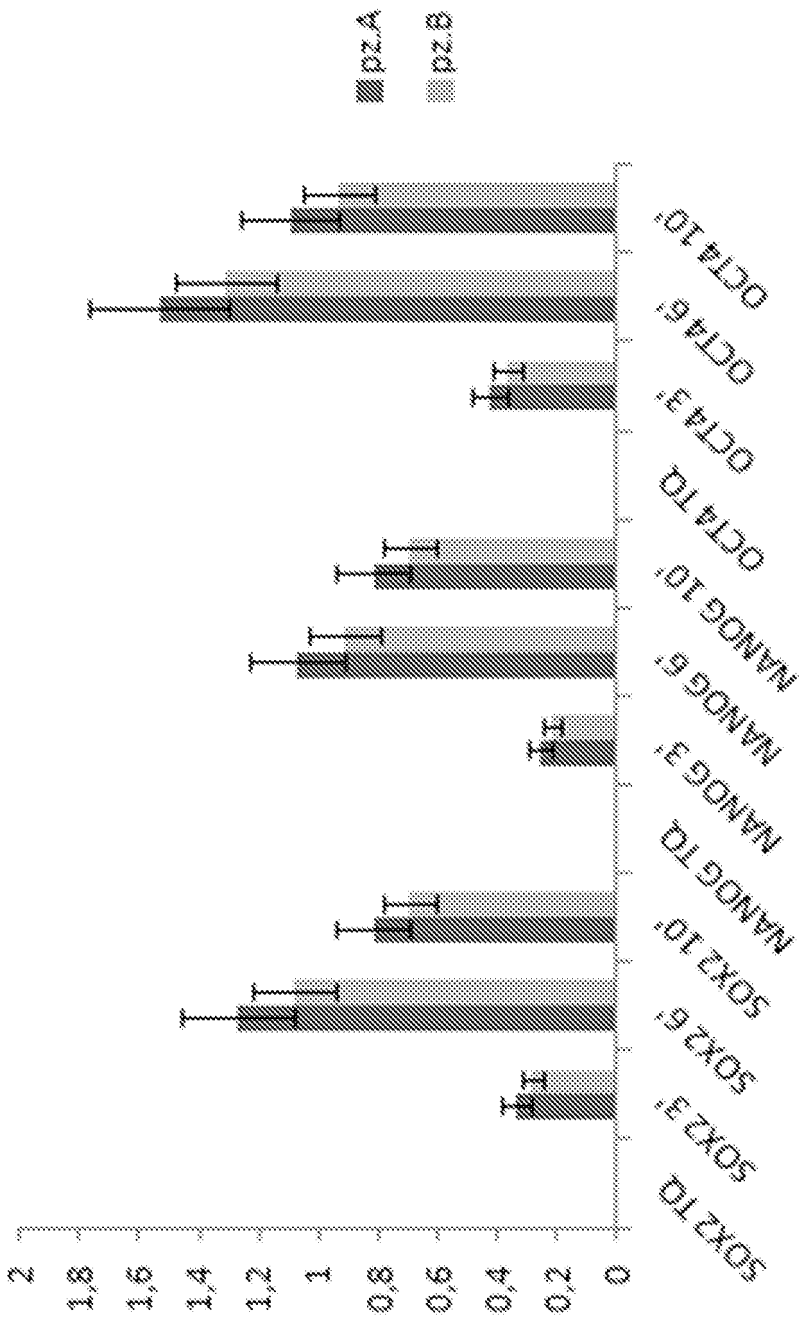

pz.A
pz.B
2
1,8
1,6
1,4
1,2
1
0,8
0,6
0,4
0,2
0
SOX2 TG
SOX2 3'
SOX2 6'
SOX2 10'
NANOG TG
NANOG 3'
NANOG 6'
NANOG 10'
OCT4 TG
OCT4 3'
OCT4 6'
OCT4 10'

Fig.3

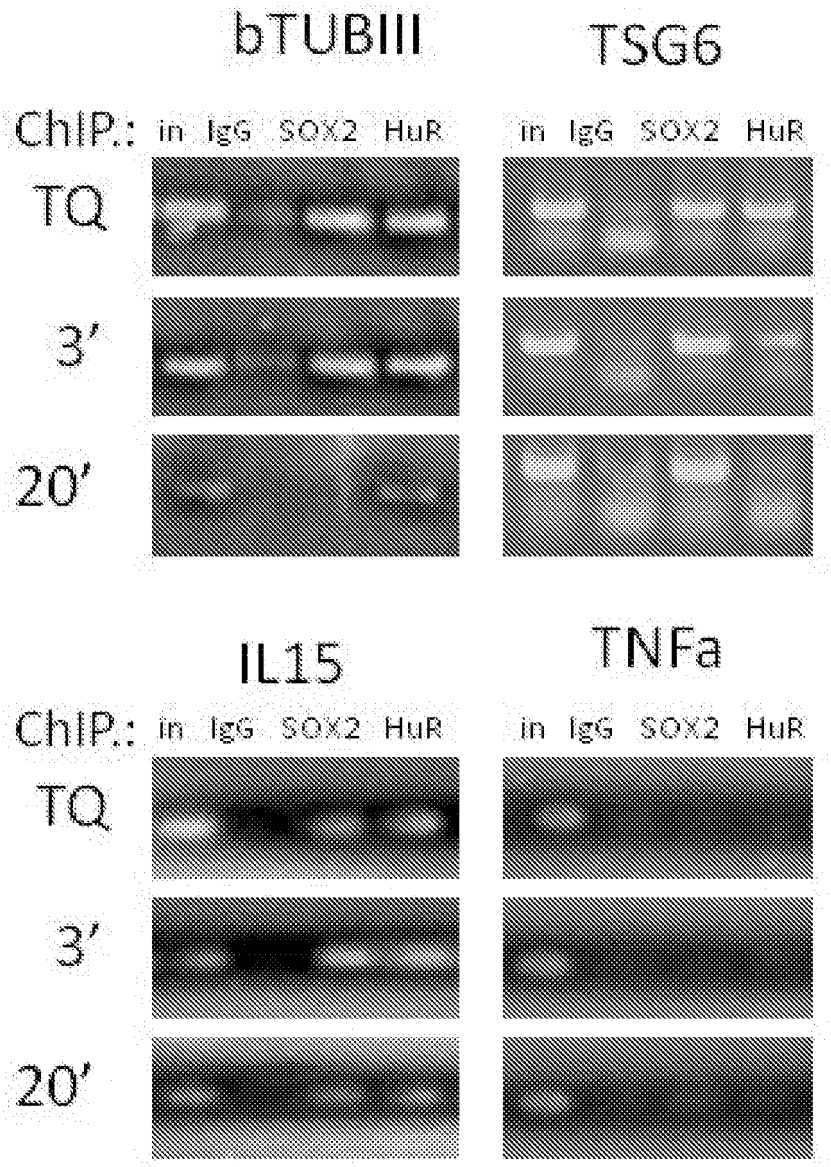

bTUBIII
TSG6
ChIP.: in IgG SOX2 HuR
in IgG SOX2 HuR
TQ
3'
20'
IL15
TNFa
ChIP.: in IgG SOX2 HuR
in IgG SOX2 HuR
TQ
3'
20'

Fig.4

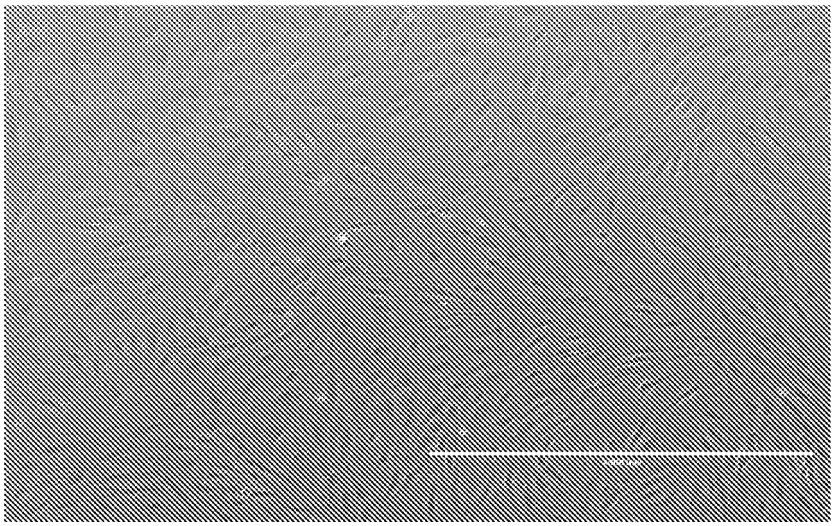


Fig.5A

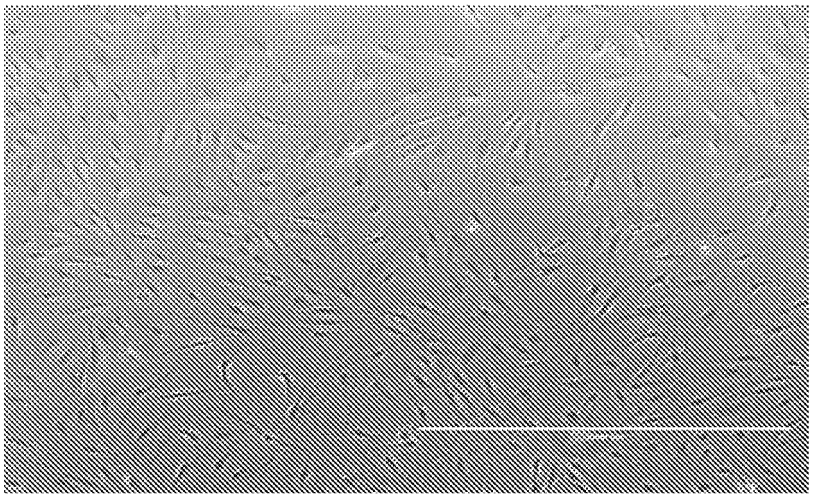


Fig.5B

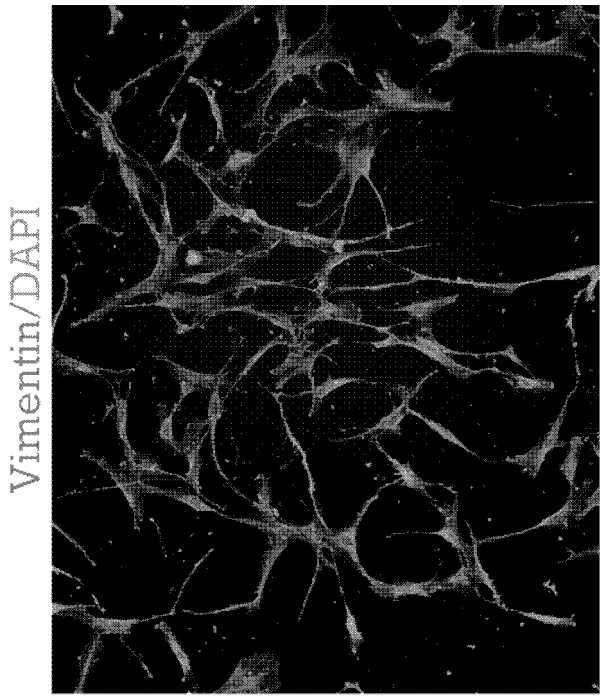


Fig.6B

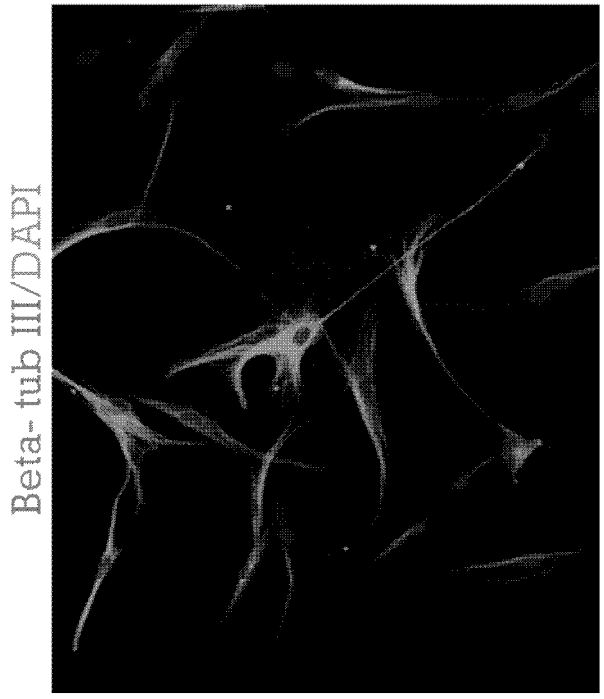


Fig.6A

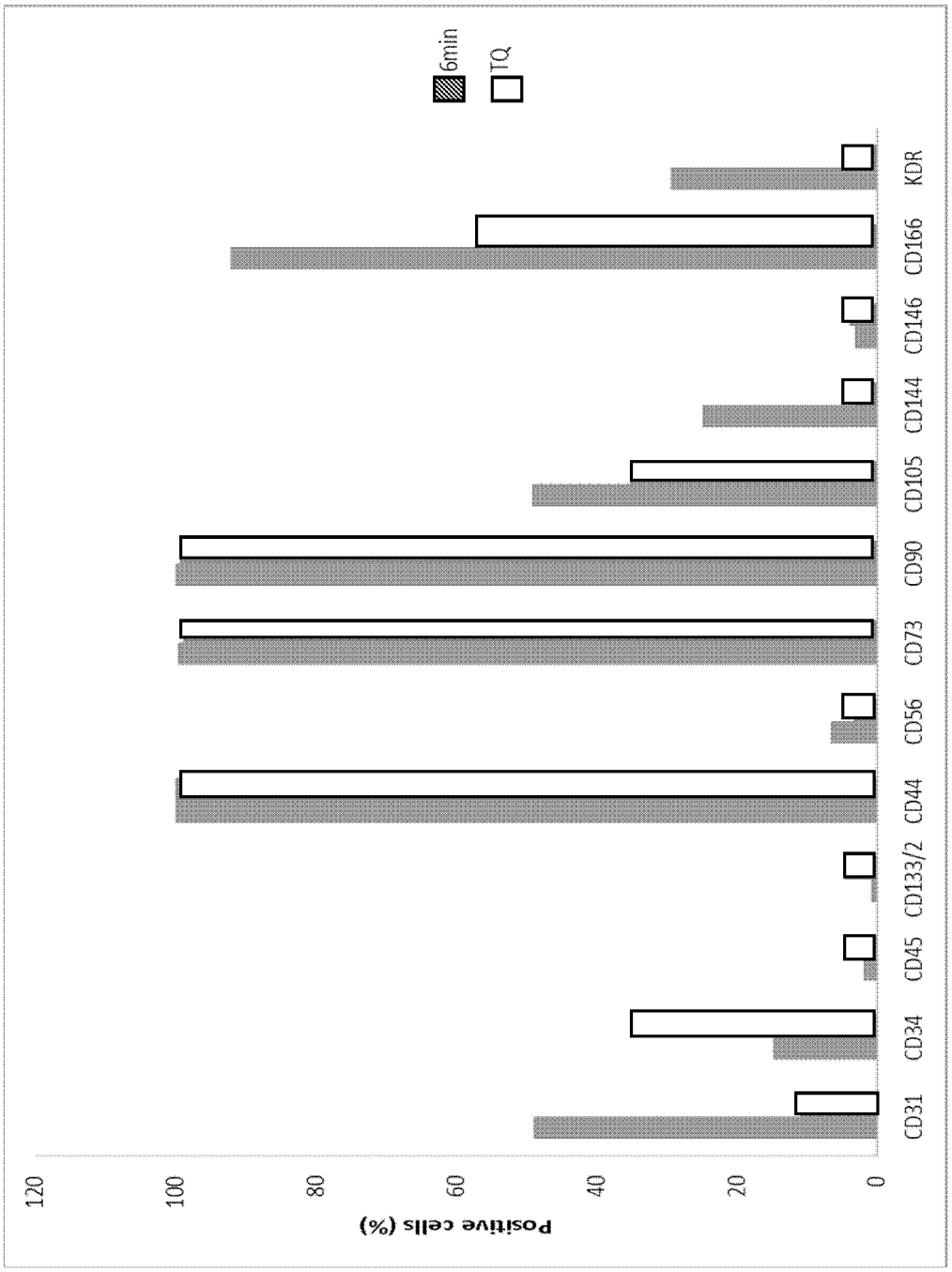

6min
TQ
Positive cells (%)
120
100
80
60
40
20
0
CD31
CD34
CD45
CD133/2
CD44
CD56
CD73
CD90
CD105
CD144
CD146
CD166
KDR
POSITIVE CELLS (%)

Fig.7

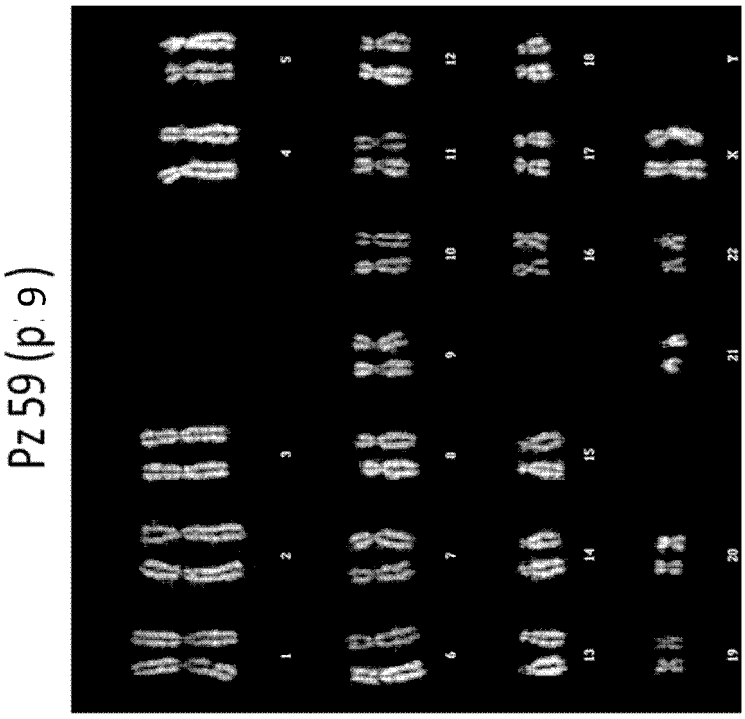


Fig.8B

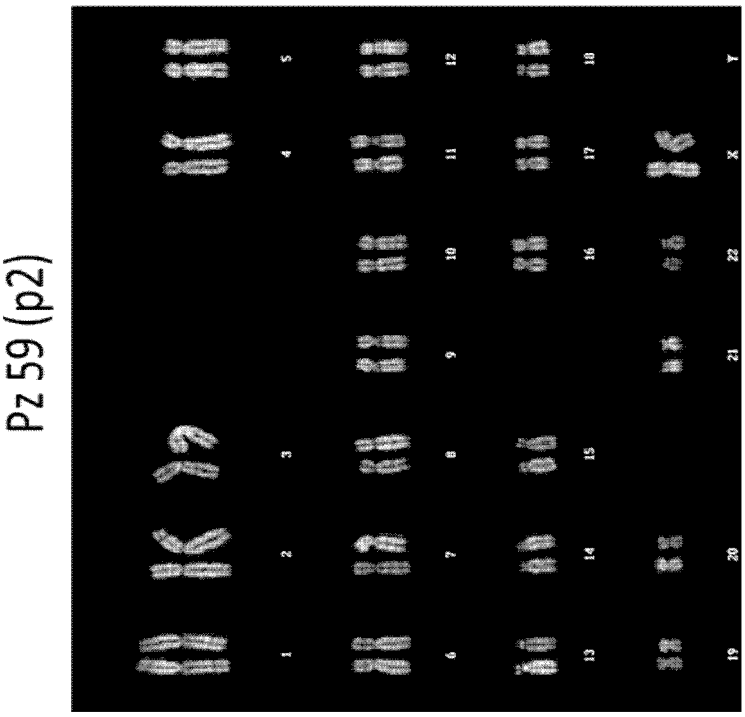
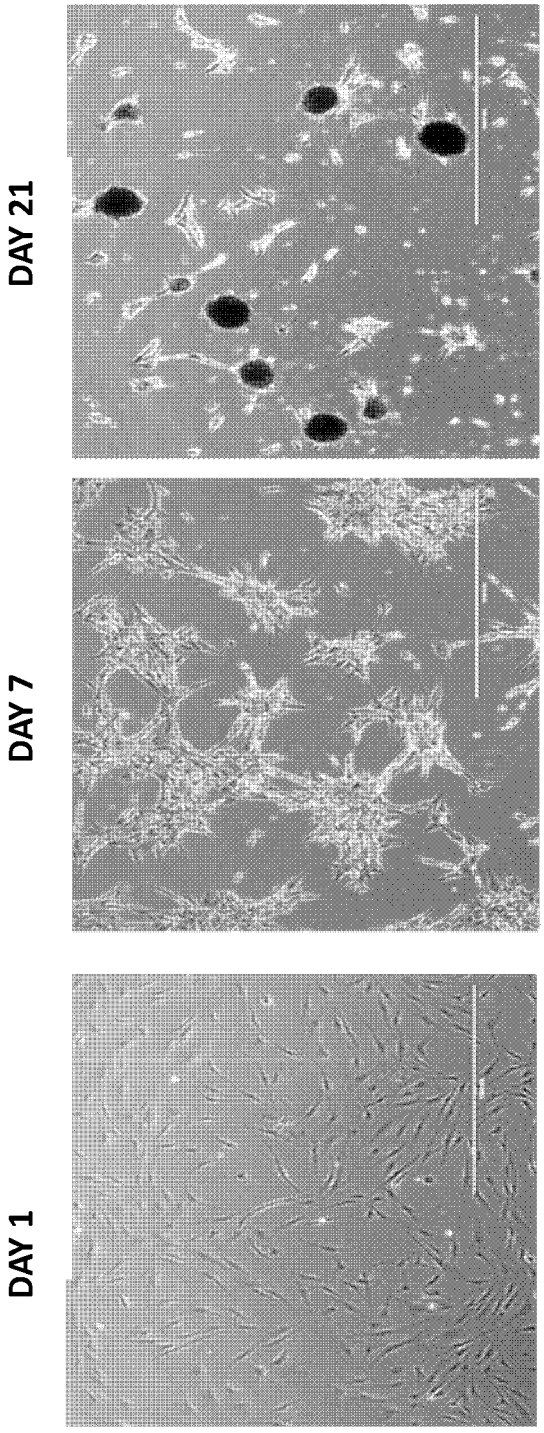


Fig.8A


DAY 1
DAY 7
DAY 21

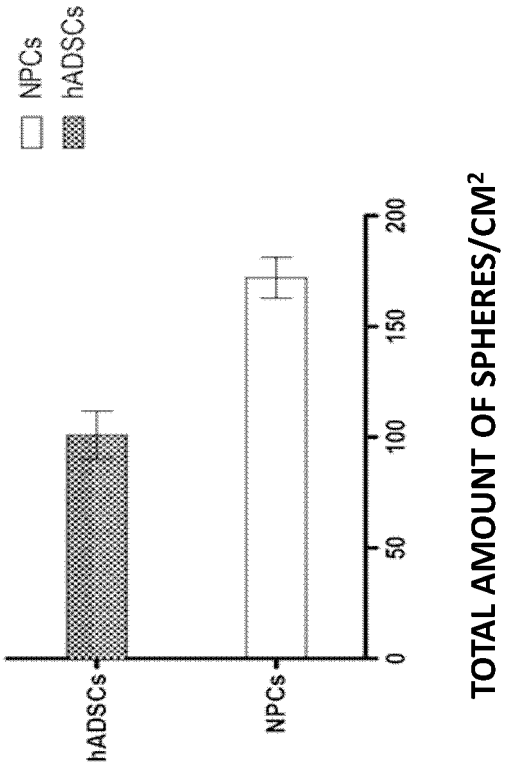

NPCs
hADSCs
0
50
100
150
200
hADSCs
NPCs
TOTAL AMOUNT OF SPHERES/CM²

Fig.9

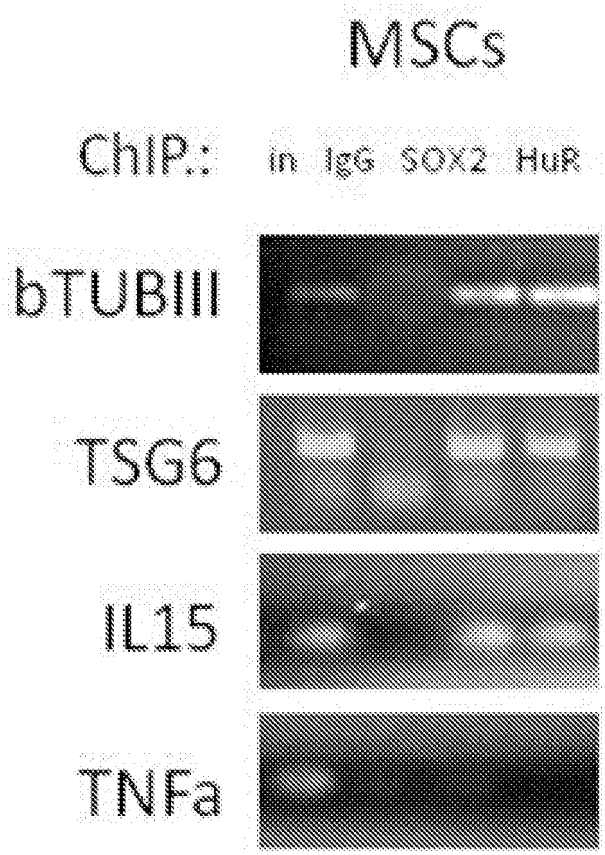


Fig.10

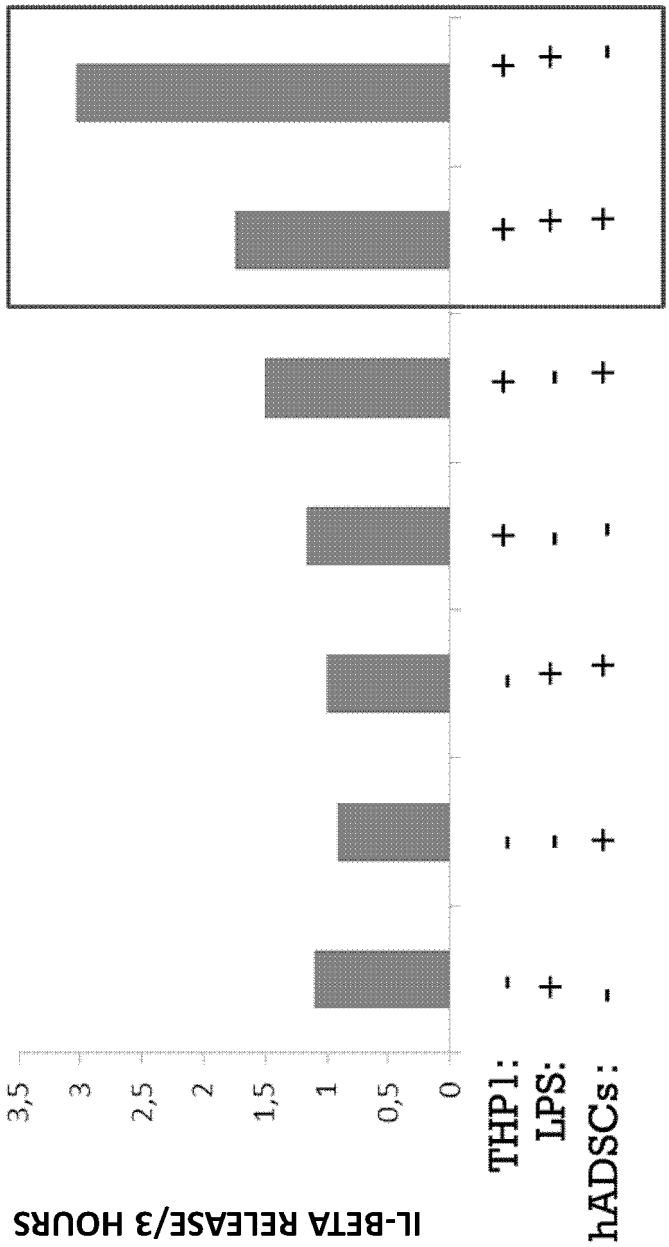


Fig.11

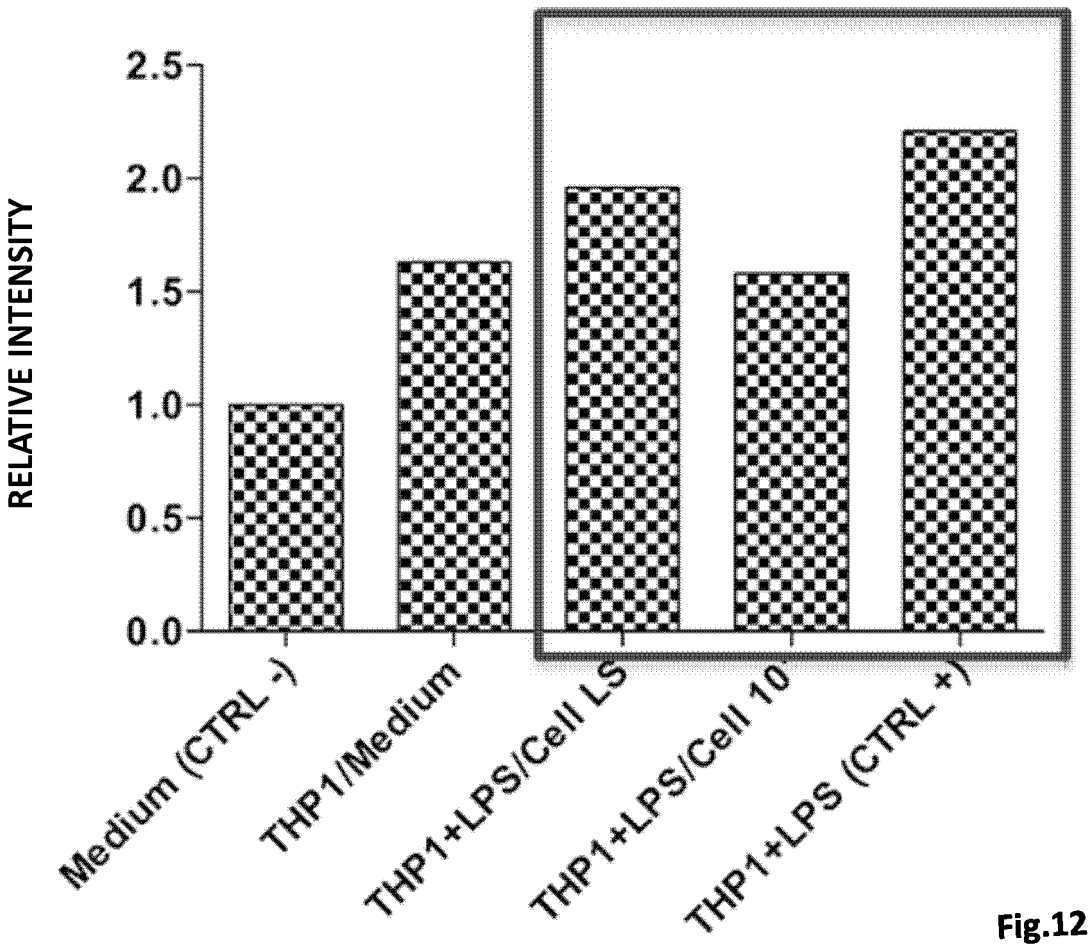
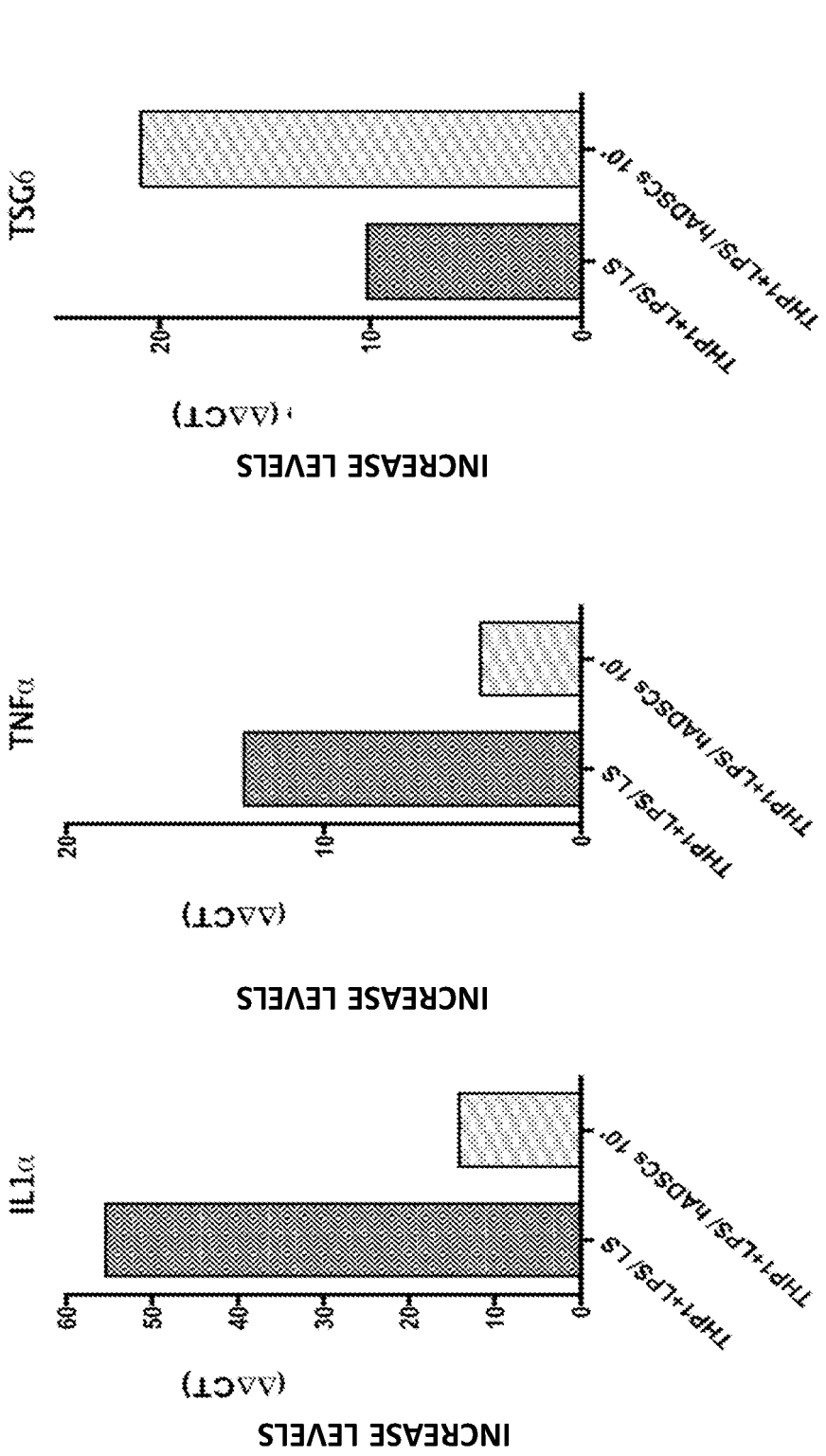

RELATIVE INTENSITY
2.5
2.0
1.5
1.0
0.5
0.0
Medium (CTRL -)
THP1/Medium
THP1+LPS/Cell LS
THP1+LPS/Cell 10
THP1+LPS (CTRL +)

Fig.12

Fig.13


TSG6
INCREASE LEVELS
(ΔΔCT)
THP1+LPS/ LS
THP1+LPS/ hADSCs 10:
TNFα
INCREASE LEVELS
(ΔΔCT)
THP1+LPS/ LS
THP1+LPS/ hADSCs 10:
IL1α
INCREASE LEVELS
(ΔΔCT)
THP1+LPS/ LS
THP1+LPS/ hADSCs 10:

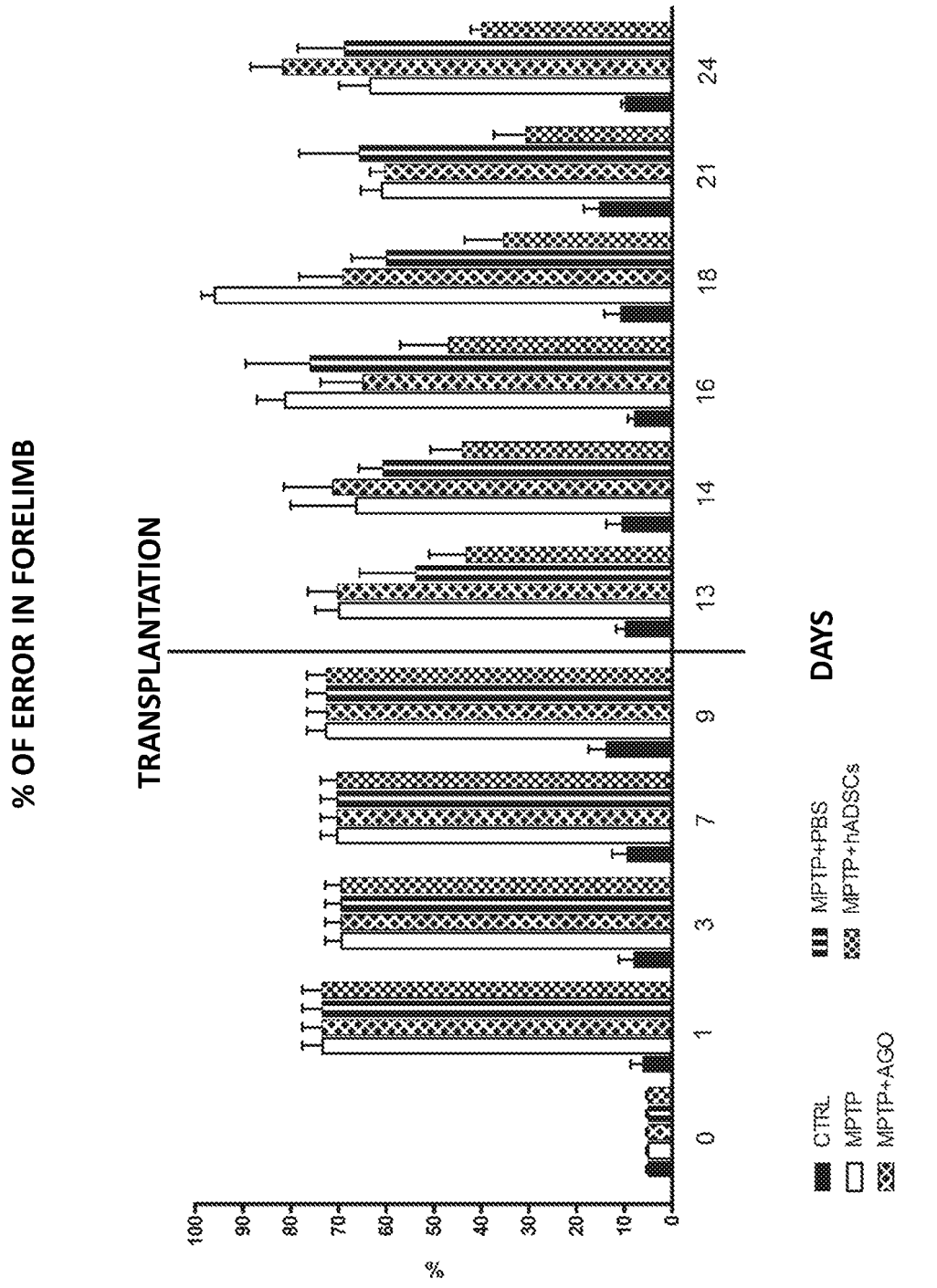

% OF ERROR IN FORELIMB
TRANSPLANTATION
DAYS
%
100
90
80
70
60
50
40
30
20
10
0
0
1
3
7
9
13
14
16
18
21
24
CTRL
MPTP
MPTP+AGO
MPTP+PBS
MPTP+hADSCs

Fig.14

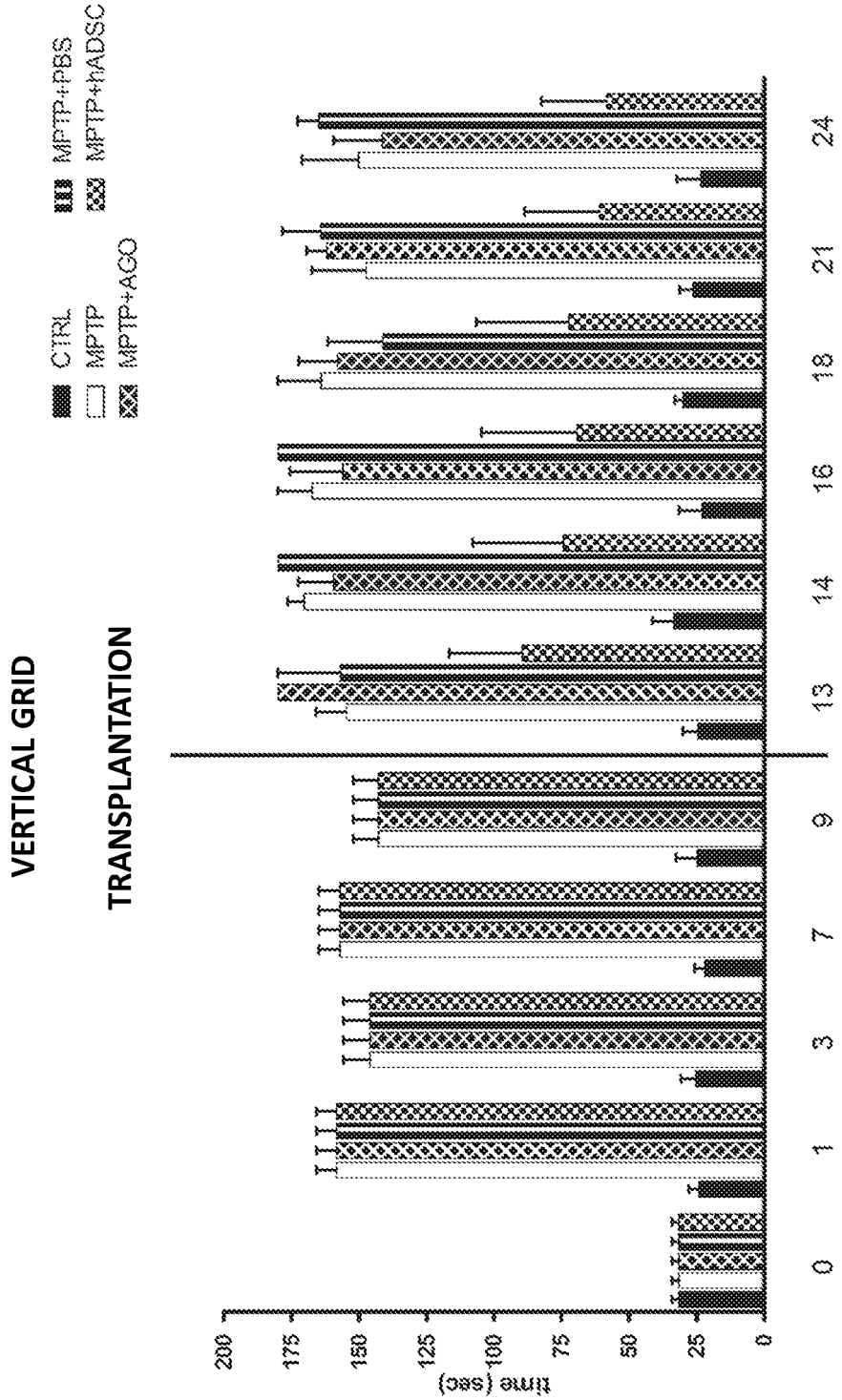

VERTICAL GRID
TRANSPLANTATION
CTRL
MPTP
MPTP+AGO
MPTP+PBS
MPTP+hADSC
time (sec)
200
175
150
125
100
75
50
25
0
0
1
3
7
9
13
14
16
18
21
24
DAYS

Fig.15

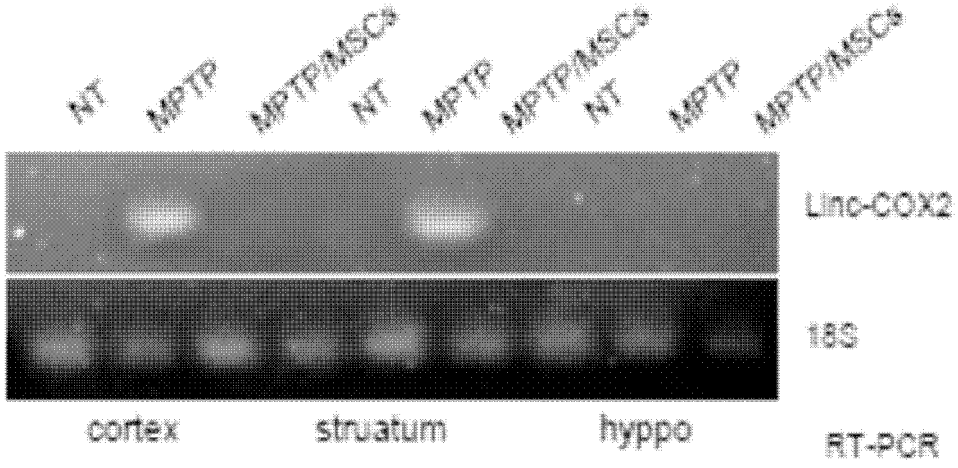


Fig.16A

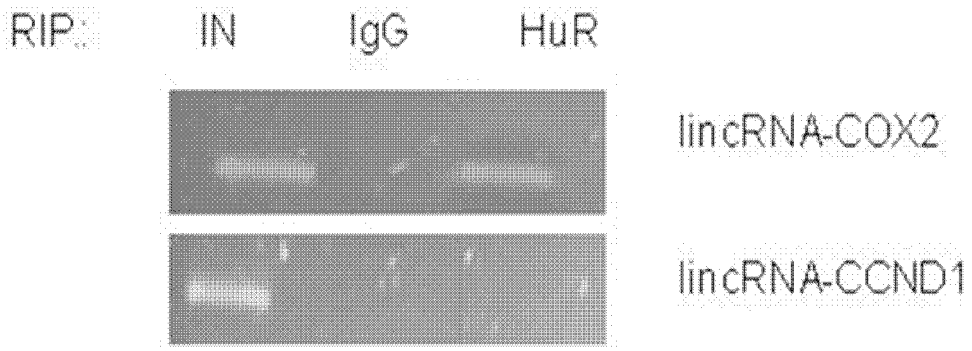


Fig.16B

ERROR IN FORELIMB

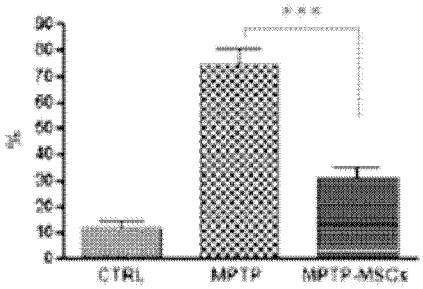


Fig.16C

VERTICAL GRID

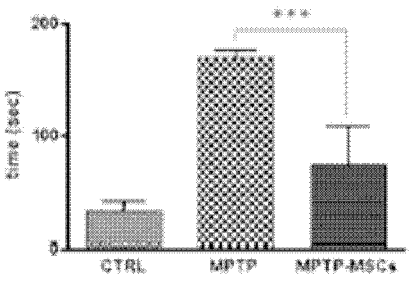


Fig.16D

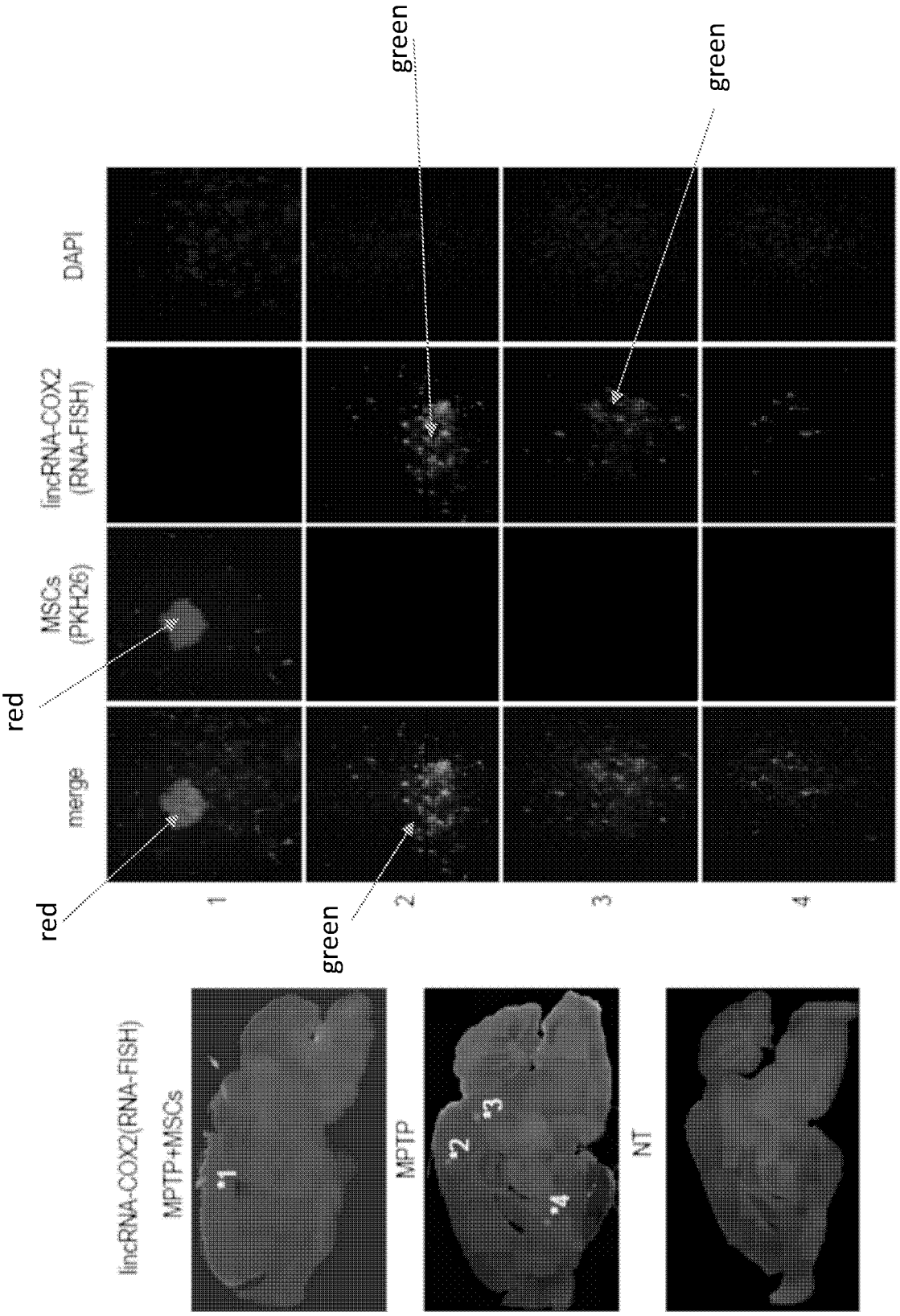

lincRNA-COX2/(RNA-FISH)
MPTP+MSCs
1
MPTP
2
3
4
NT
green
red
merge
MSCs (PKH26)
lincRNA-COX2 (RNA-FISH)
DAPI
green
green
red
1
2
3
4
1= MSCs Injection site

Fig. 17

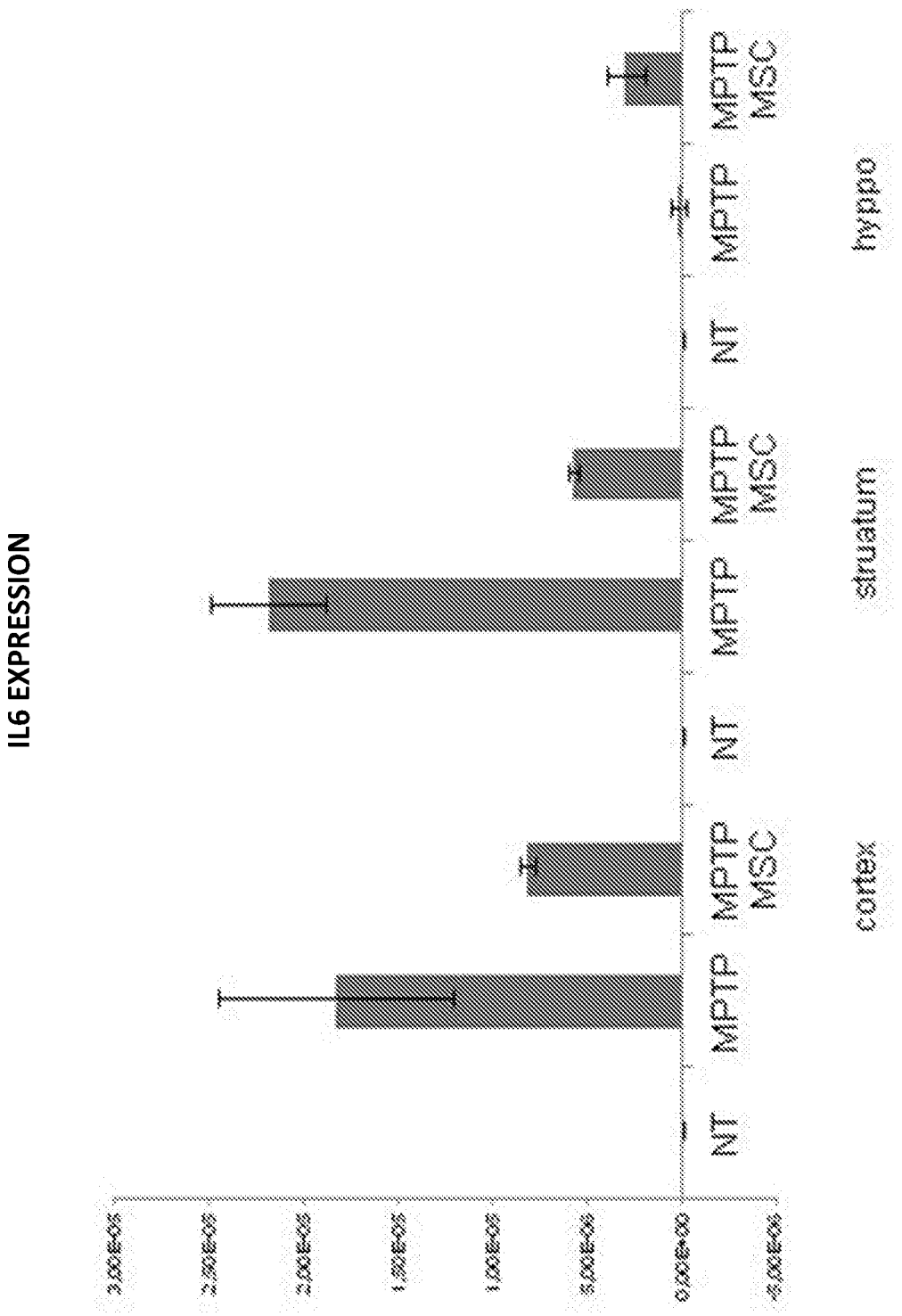

IL6 EXPRESSION
3,00E-05
2,50E-05
2,00E-05
1,50E-05
1,00E-05
5,00E-06
0,00E+00
-5,00E-06
NT
MPTP
MPTP MSC
NT
MPTP
MPTP MSC
NT
MPTP
MPTP MSC
cortex
striatum
hyppo

Fig. 18

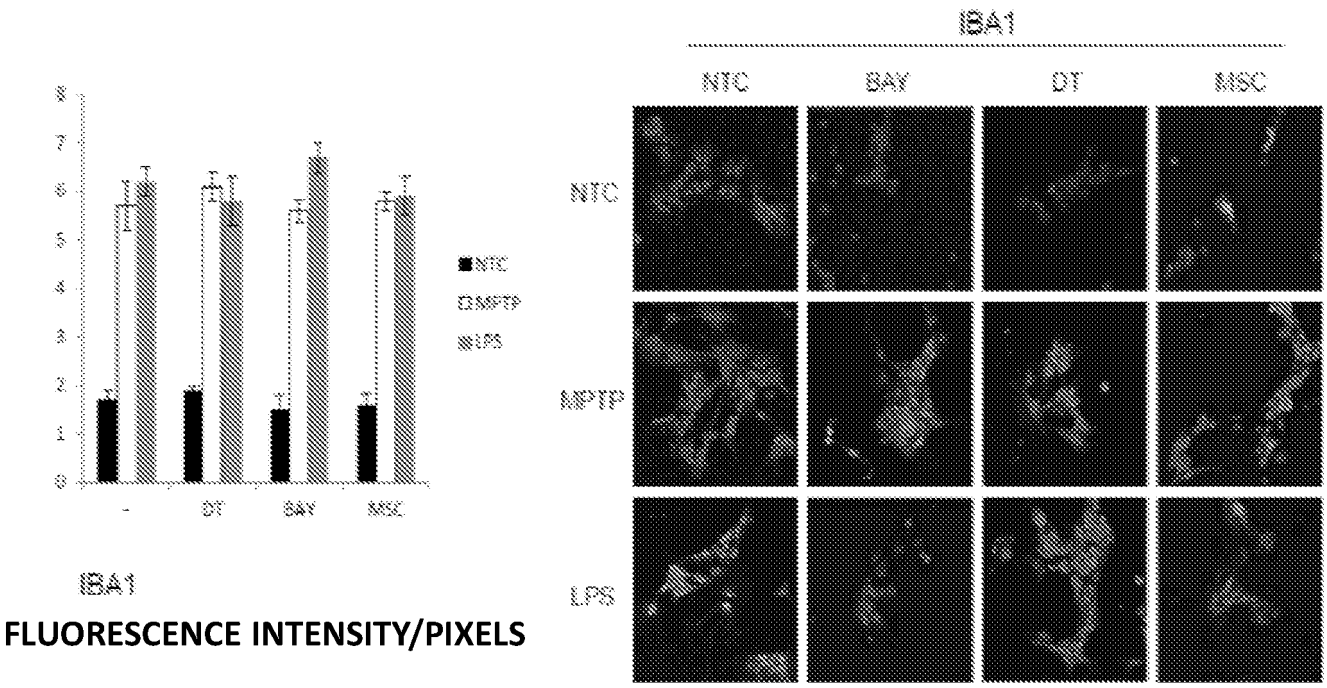


Fig. 19A

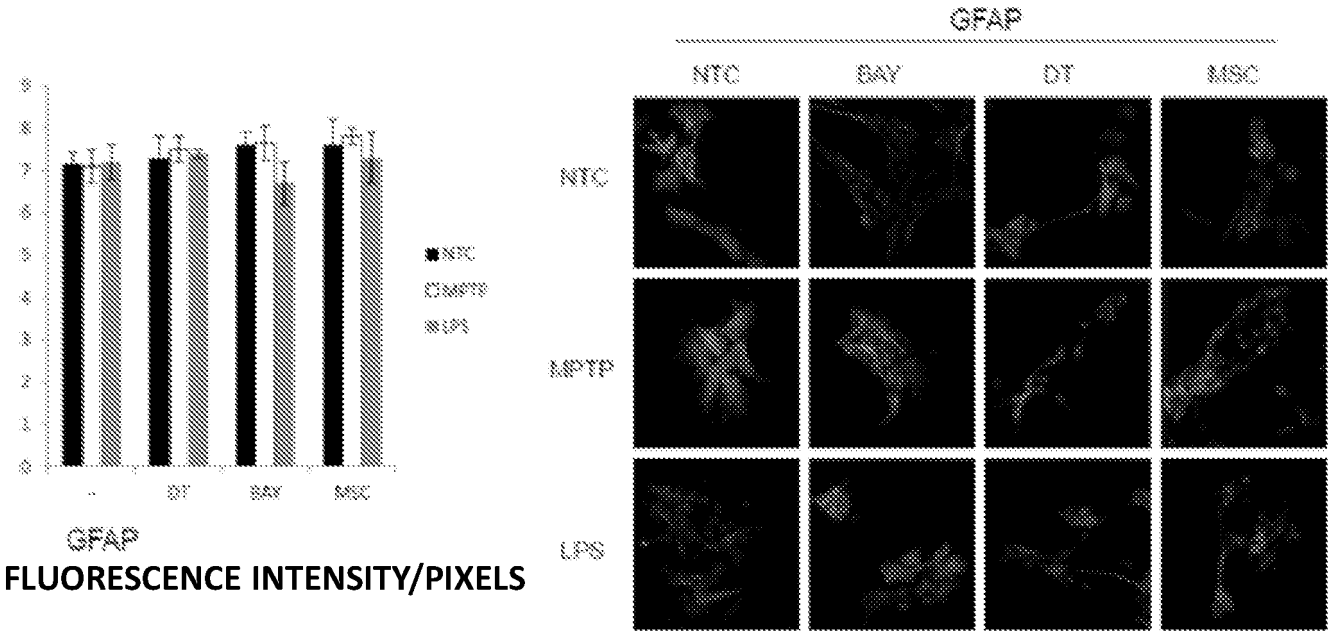


Fig. 19B

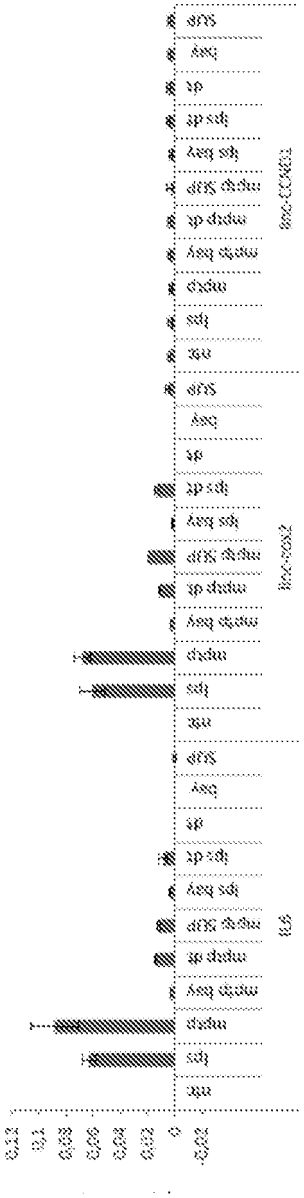

LEVEL OF mRNA EXPRESSION
2exp (-deltaCt)
0.12
0.1
0.08
0.06
0.04
0.02
0
mtc
tps
mpcp
mpcp dt
mpcp beg
mpcp dt
mpcp SUP
tps beg
tps dt
dt
beg
SUP
mtc
tps
mpcp
mpcp beg
mpcp dt
mpcp SUP
tps beg
tps dt
dt
beg
SUP
mtc
tps
mpcp
mpcp dt
mpcp beg
mpcp dt
mpcp SUP
tps beg
tps dt
dt
beg
SUP
Wnt-CCND1
Wnt-TCF4
Wnt-CTNNB1

Fig. 20

CYTOKINE EXPRESSION (48 HOURS)

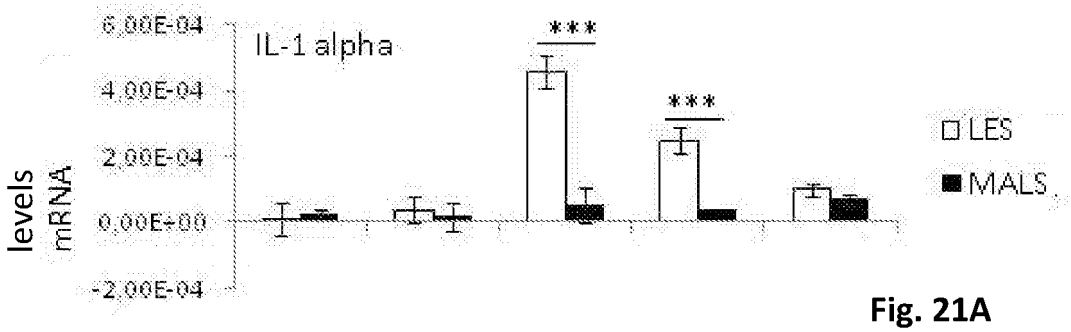


Fig. 21A

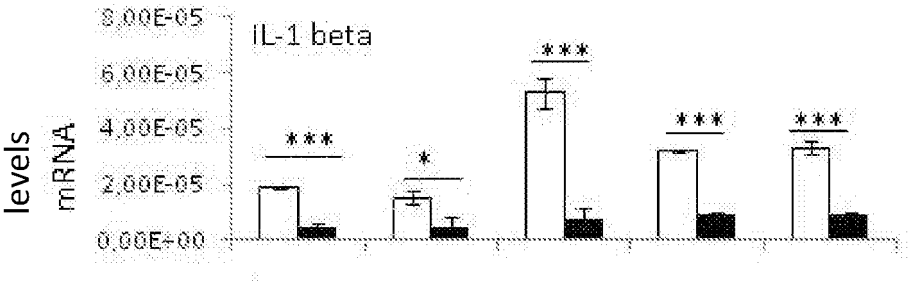
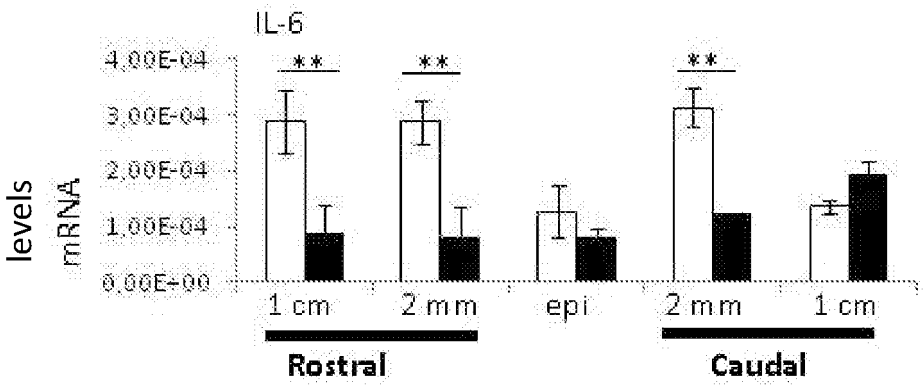


Fig. 21B



* p< 0.05; ** p<0.01; *** p<0.001 vs LES

Fig. 21C

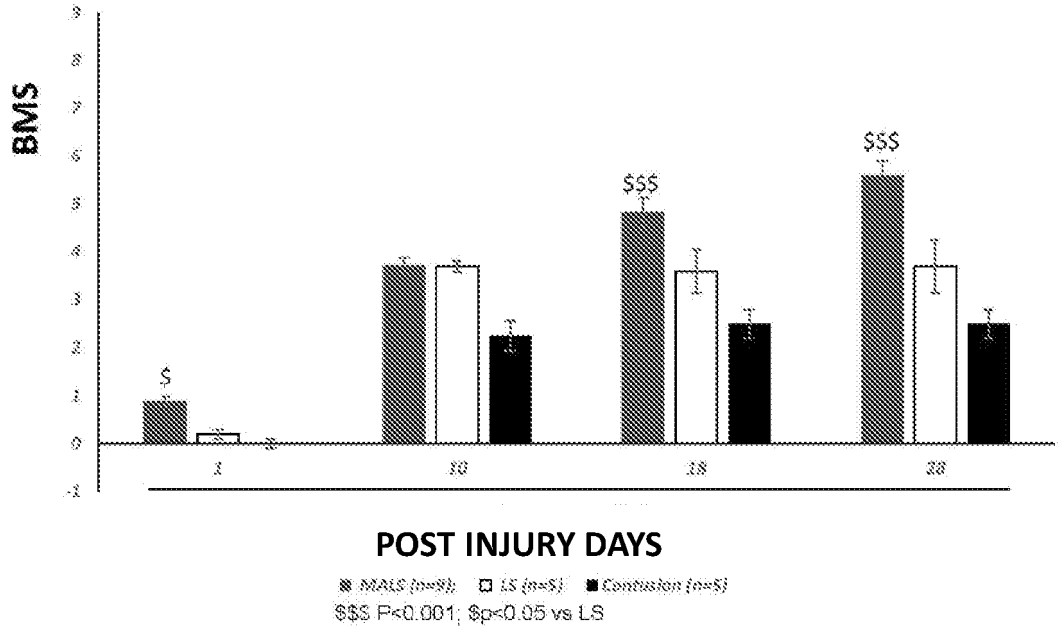

BMS
$
$$$
$$$
1
10
18
28
POST INJURY DAYS
MALS (n=9)
LS (n=5)
Contusion (n=5)
$$$ P<0.001; $p<0.05 vs LS

Fig. 22

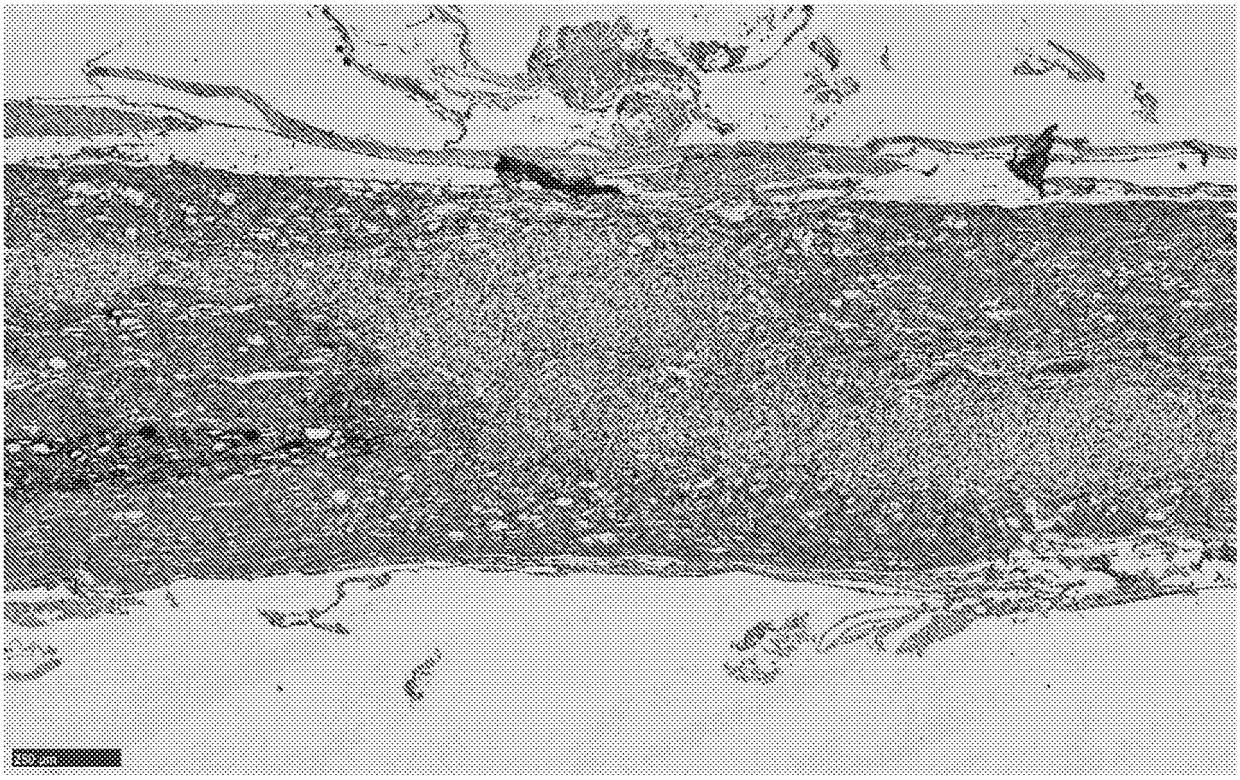
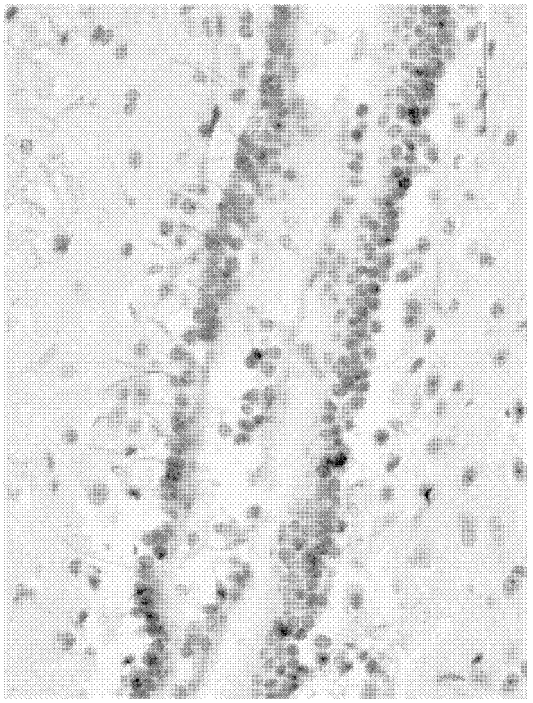


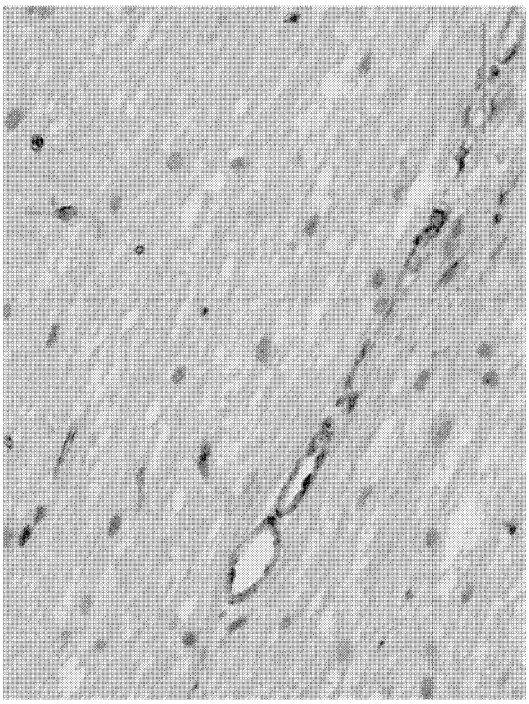
Fig. 23



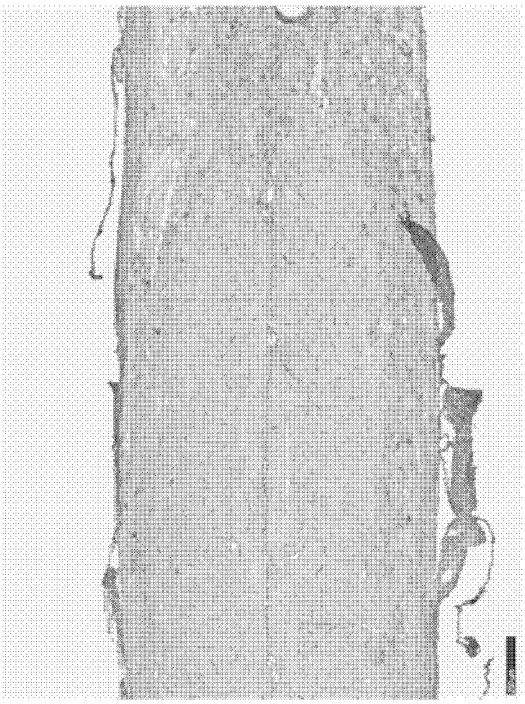
Contusion 70Kdyne Fig. 24 b



Contusion 70Kdyne + MALS
Fig. 24 d



CTRL NESTIN
Fig. 24 a



Contusion 70Kdyne + LS
Fig. 24 c

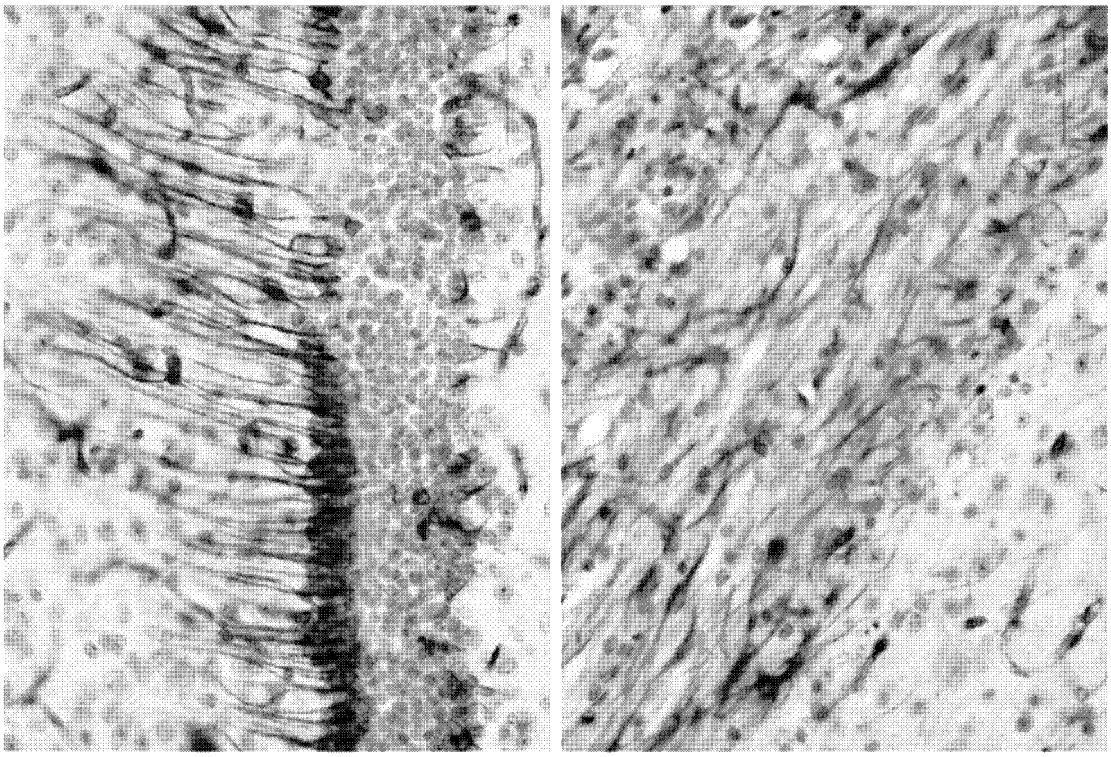


Fig. 25 a

Fig. 25 b

Fig. 25 c

Fig. 25 d

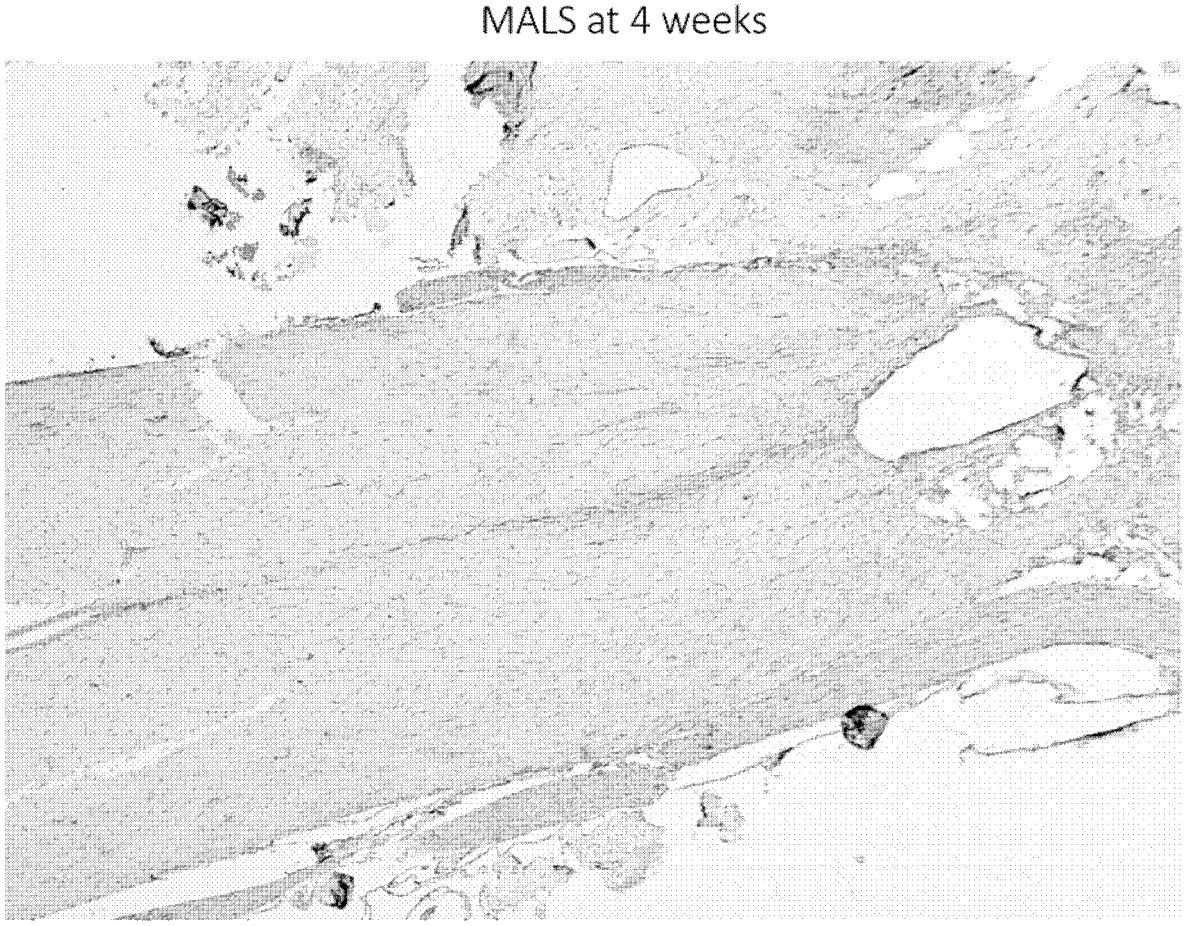


Fig. 26

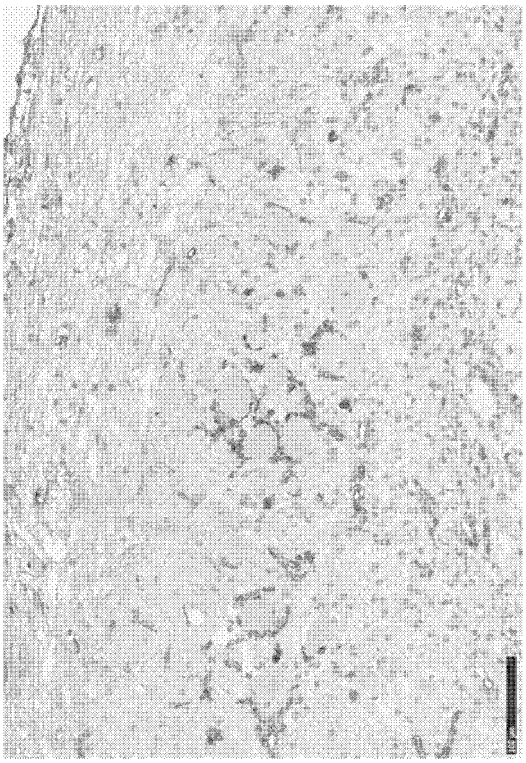


Fig. 27b

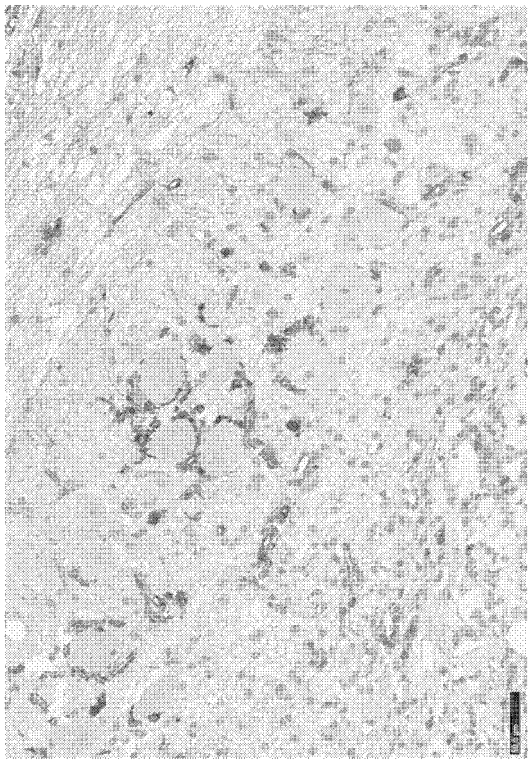


Fig. 27 c



Fig. 27 a