

METHOD OF RECONSTITUTING LIPOSOMAL ANNAMYCIN

[0001] This application claims the benefit of priority of United States Provisional Application No.

62/868,190, filed June 28, 2019, the disclosure of which is hereby incorporated by reference as if written herein in its entirety.

[0002] Annamycin is a cancer chemotherapeutic agent of the anthracene structural family. Because of its physical and pharmacological properties, compositions of Annamycin for dosage forms have been described in which the drug is formulated as a liposome (See US 7,238,366). Clinical trials have been described for liposomal Annamycin in adult patients with relapsed refractory Acute lymphoblastic leukemia (M. Wetzler, et al., Clinical Lymphoma, Myeloma and Leukemia, 13 (4), 430-434 August 2013) and in the treatment of doxorubicin-resistant breast cancer (D. J. Booser et al., Cancer Chemother. Pharmacol. 50; 6-8, 2002).

[0003] In drug delivery systems, it is desirable to be able to produce a drug formulation that is of consistent purity and quality in order to ensure that the correct dosage is administered, thus providing the most efficacious treatment, while avoiding unwanted side effects caused by impurities. Furthermore, poorly maintained or improperly formulated drug material can decompose prior to administration, rendering it ineffective, resulting in waste of costly active pharmaceutical ingredients. Regulatory standards also necessitate that a consistent level of purity of drug and amount of drug be provided by following the procedures specified on the label.

[0004] Annamycin presents particularly difficult formulation challenges because of its inherent physical properties, and because of inherent instability of both the parent drug and of liposomal formulation under certain conditions. It would therefore be advantageous to find a consistent method for preparing a dosage form of liposomal Annamycin that assures purity and avoids decomposition.

SUMMARY

[0005] Provided is a method of reconstituting a preliposomal Annamycin lyophilizate to form a liposomal Annamycin suspension.

[0006] Also provide is a method of reconstituting Annamycin, such method comprising:

- adding pre-warmed saline to one or more containers of preliposomal Annamycin lyophilizate, to form suspended liposomal Annamycin with a concentration of about 0.5 -1.5 mg/mL;
- maintaining said one or more containers at 24 -45 °C;
- diluting said suspended liposomal Annamycin, with pre-warmed saline to provide liposomal Annamycin at a concentration in saline of about 0.5– 1.5 mg/mL;
- and
- maintaining said one or more containers of suspended liposomal Annamycin and said diluted liposomal Annamycin at 24- 45°C during and after the dilution.

[0007] Also provided is a method of preparing an effective dose of a reconstituted liposomal formulation of Annamycin, such method comprising:

- warming syringes and non-PVC IV bags, each containing saline (about) 0.9% aqueous injection USP sodium chloride at $37^{\circ}\text{C} \pm 4^{\circ}\text{C}$ for at least about three hours, in an oven or bag warmer, wherein

- the volume of saline in mL in a syringe is equal to the weight in mg of preliposomal lyophilizate Annamycin to be reconstituted, and

- the volume of saline within said IV bag equivalent to the total amount of saline in all syringes, and

- the volume of said IV bag is of sufficient capacity to contain the total volume of saline and liposomal Annamycin. for dilution of the total dose of liposomal Annamycin;

- removing a sufficient number of vials containing preliposomal Annamycin lyophilizate to provide the desired total dose of Annamycin from the freezer and allowing the vials to stand at room temperature for 30 minutes to 2 hours;

- vigorously shaking each vial of preliposomal lyophilizate Annamycin to break up any dry cake;

- injecting air into each vial from air filled syringes until pressure in each vial is equalized;

- adding a volume of $37^{\circ}\text{C} \pm 4^{\circ}\text{C}$ saline from a syringe kept at $37^{\circ}\text{C} \pm 4^{\circ}\text{C}$ in an oven to a vial containing a weight equivalent vial of preliposomal lyophilizate Annamycin such that the final Annamycin concentration is approximately 1 mg/mL, maintaining negative pressure throughout the addition of saline;

- immediately shaking the vial back and forth vigorously by hand for 15 seconds, then placing the vial in a resealable polyethylene storage bag;

- repeating the addition of saline, shaking the vial, and placing the vial in the storage bag until all vials required for the total dose of Annamycin are similarly prepared;

- shaking, for a second time, all vials, 2-4 at a time for 1-2 minutes;

- storing all vials of reconstituted liposomal Annamycin at $37^{\circ}\text{C} \pm 4^{\circ}\text{C}$ until ready for dilution, for 30 minutes up to 1 hour;

- removing vials of reconstituted liposomal Annamycin from storage at $37^{\circ}\text{C} \pm 4^{\circ}\text{C}$ no more than two at a time, and after gently inverting each vial once, withdrawing a foam-free suspension from the vial while maintaining negative pressure throughout the withdrawal of the suspension;

- introducing the withdrawn foam-free suspension of reconstituted liposomal Annamycin slowly into a pre-warmed non-PVC IV bag of saline, prepared as described above and repeating this process for the second vial;

- repeating the steps of removing the vials, withdrawing the suspension, and introducing the suspension into the prewarmed IV bag until all vials of reconstituted liposomal Annamycin

required for the total dose of Annamycin have been introduced into the non-PVC IV bag, providing an IV bag containing the final reconstituted liposomal Annamycin with a concentration of about 0.5 mg/mL; and

agitating the saline and reconstituted liposomal Annamycin in the IV bag by gently inverting the container several times, without generating foam.

[0008] Also provided is a method of treating cancer comprising administering to a patient in need thereof, an effective dose of diluted liposomal Annamycin prepared by

adding pre-warmed saline to one or more containers of preliposomal Annamycin lyophilizate, to form a liposomal Annamycin suspension with a concentration of about 0.5 -1.5 mg/mL;

maintaining said one or more containers at 24 -45 °C;

diluting said suspended liposomal Annamycin, with pre-warmed saline to provide liposomal Annamycin at a concentration in saline of about 0.3 – 0.7 mg/mL; and

maintaining said one or more containers of suspended liposomal Annamycin and said diluted liposomal Annamycin at 24- 45°C during and after the dilution.

[0009] Also provided is a use of diluted reconstituted liposomal Annamycin prepared according to the above methods, in the manufacture of a medicament for the treatment of cancer.

[0010] Also provided is a diluted reconstituted liposomal Annamycin prepared according to the above methods, for use in the treatment of cancer.

BRIEF DESCRIPTION OF THE DRAWINGS

[0011] Fig. 1. A schematic representation, exemplifying the method for reconstituting liposomal Annamycin from preliposomal Annamycin lyophilizate.

[0012] Fig. 2. A histogram result overlay for 20 – 25°C samples.

[0013] Fig. 3. A histogram result overlay for 28 – 42°C samples.

[0014] Fig. 4. A histogram result overlay for 20 – 42°C samples.

[0015] DETAILED DESCRIPTION

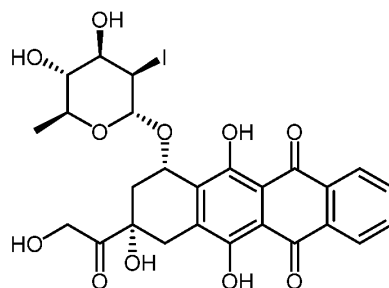
[0016] Provided is a method of making a formulation of liposomal Annamycin with improved stability and high purity.

Definitions

[0017] Unless defined otherwise, all technical and scientific terms used herein have the same meaning as is commonly understood by one of skill in the art to which the invention(s) belong.

[0018] As used herein, the following terms have the meaning as defined below.

[0019] The term “Annamycin” shall mean the compound (7S,9S)-7-(((2R,3R,4R,5R,6S)-4,5-dihydroxy-3-iodo-6-methyltetrahydro-2H-pyran-2-yl)oxy)-6,9,11-trihydroxy-9-(2-hydroxyacetyl)-7,8,9,10-tetrahydrotetracene-5,12-dione, having the following structure:



[0020] The term "liposomes," "liposomal," and the like shall mean generally spherical structures comprising lipids, fatty acids, lipid bilayer type structures, unilamellar vesicles and amorphous lipid vesicles. Classically, liposomes are completely closed lipid bilayer membranes containing an entrapped aqueous volume. Liposomes include non-classical forms where the Annamycin may be inside the bilayer, part of the bilayer, and absorbed onto the bilayer. Liposomes may be unilamellar vesicles (possessing a single bilayer membrane) or multilamellar vesicles (onion-like structures characterized by multiple membrane bilayers, each separated from the next by an aqueous layer). The bilayer is composed of two lipid monolayers having a hydrophobic "tail" region and a hydrophilic "head" region. The structure of the membrane bilayer is such that the hydrophobic (nonpolar) "tails" of the lipid monolayers orient toward the center of the bilayer while the hydrophilic "head" orient towards the aqueous phase.

[0021] The term "preliposome-lyophilizate" and "preliposomal lyophilizate" shall mean a non-aqueous material that will form liposomes upon addition of aqueous solution. In some embodiments the non-aqueous material is dry (as in non-liquid, non-gel) material. Lyophilizate is used expansively to include the dry residue of sublimation of frozen liquids from non-volatile materials, the residue of roto evaporation and similar procedures, and dry compositions that, upon addition of an aqueous phase (with or without agitation) will result in liposomes. It is particularly to be understood that "preliposome-lyophilizate" is not in liposomal form after lyophilization.

[0022] The term "preliposomal Annamycin lyophilizate" is one preliposome-lyophilizate where the drug is Annamycin. In some embodiments, the lyophilizate is prepared according to the procedures set forth in U.S. Patent Publication 2005/0238707, which is incorporated herein by reference for all purposes.

[0023] The term "pre-warmed" means that the article or solution or suspension is warmed to a temperature of about 24- 45°C, such as 30-45 °C, for example, 37 °C ± 4 °C, or 37 °C ± 2 °C, 37 °C ± 1 °C.

[0024] The term "high purity Annamycin preliposomal lyophilizate" shall mean purity of material which is no less than 95% Annamycin as analyzed by HPLC using a verified standard sample. In some embodiments, the Annamycin is at least 96% pure, or at least 97% pure, or at least 98% pure, or at least 99% pure.

[0025] In the foregoing sections, a number of abbreviations and acronyms were used, and the full description of these are provided as follows:

DMPC	dimyristoyl phosphatidylcholine
DMPG	1,2-dimyristoyl-sn-glycero-3-phospho-(1'-rac-glycerol) (sodium salt)
DMSO	dimethyl sulfoxide
IV	intravenous
DEHP	bis(2-ethylhexyl) phthalate
PLA	preliposomal lyophilized Annamycin
PVC	polyvinyl chloride
WFI	water for injection; in compliance with USP

[0026] Throughout this specification and in the claims that follow, unless the context requires otherwise, the word “comprise”, or variations such as “comprises” or “comprising”, should be understood to imply the inclusion of a stated integer or step or group of integers or steps but not the exclusion of any other integer or step or group of integers or steps. It is intended herein that by recitation of such specified ranges, the ranges recited also include all those specific integer amounts between the cited ranges. For example, the range of about 24-45 °C, it is intended to also encompass 24, 30, 33, 34, 35, 36, 37, 38, 39, 40, and 45 °C.

[0027] The entire disclosure of each United States patent and international patent application mentioned in this patent specification is fully incorporated by reference herein for all purposes.

[0028] Also provided is a method of making a liposomal Annamycin that has superior uniformity and stability. Provided is a novel method of making a liposomal Annamycin that has superior uniformity and stability. In particular, provided is a method of reconstituting Annamycin, such method comprising:

adding pre-warmed saline to one or more containers of preliposomal Annamycin lyophilizate, to form suspended liposomal Annamycin with a concentration of about 0.5 -1.5 mg/mL;
 maintaining said one or more containers at 24 -45 °C;
 diluting said suspended liposomal Annamycin, with pre-warmed saline to provide liposomal Annamycin at a concentration in saline of about 0.5 – 1.5 mg/mL; and
 maintaining said one or more containers of suspended liposomal Annamycin and said diluted liposomal Annamycin at 24- 45°C during and after the dilution.

[0029] In some embodiments, the liposomal Annamycin suspension has a concentration of 0.5 – 1.5 mg/mL. In some embodiments, the liposomal Annamycin suspension has a concentration of 0.8 – 1.2 mg/mL. In some embodiments, the liposomal Annamycin suspension has a concentration of about 1 mg/mL.

[0030] In some embodiments, the total weight of the Annamycin in the diluted suspension added into a pre-warmed saline containing IV bag is equal to the effective dose required.

[0031] In some embodiments, the suspended liposomal Annamycin is diluted by adding it into a pre-warmed saline-containing IV bag. In some embodiments, the saline-containing IV bag is at a temperature of 30 -45 °C. In some embodiments, the saline-containing IV bag is at a temperature of 37 °C ± 4 °C.

[0032] In some embodiments, the reconstituted liposomal Annamycin is formed at or above 24 °C. In some embodiments, the reconstituted liposomal Annamycin is maintained at or above 24 °C prior to and during dilution, and prior to and during infusion. It was surprisingly found that liposomal Annamycin was unstable below 24 °C.

[0033] In some embodiments, the reconstituted liposomal Annamycin is maintained at a temperature of 30 - 45 °C. In some embodiments, the reconstituted liposomal Annamycin is maintained at a temperature of 30 - 42 °C. In some embodiments, the reconstituted liposomal Annamycin is maintained at a temperature of 37 °C $\pm$ 4 °C. In some embodiments, the suspension of liposomal Annamycin is maintained at or near 37 °C and the diluted liposomal Annamycin is maintained at or near 37°C.

[0034] In some embodiments, the diluted liposomal Annamycin has a concentration of about 0.5 mg/mL.

[0035] In some embodiments, a protective sleeve is used on the IV drip line to ensure that the diluted liposomal Annamycin is delivered at a temperature of at or above 37 °C $\pm$ 4 °C. In some embodiments, the sleeve is heated. In some embodiments, the sleeve is insulated.

[0036] In some embodiments, the method of reconstituting Annamycin, comprises:

- adding saline at 37 °C $\pm$ 4 °C to one or more containers of preliposomal Annamycin lyophilizate, to form suspended liposomal Annamycin with a concentration of about 1 mg/mL;
- maintaining said one or more containers at 37 °C $\pm$ 4 °C;
- diluting said suspended liposomal Annamycin, with saline at 37 °C $\pm$ 4 °C to provide liposomal Annamycin at a concentration in saline of about 0.5 mg/mL; and
- maintaining said one or more containers of suspended liposomal Annamycin and said diluted liposomal Annamycin at 37 °C $\pm$ 4 °C during and after the dilution.

[0037] In some embodiments, the diluted liposomal Annamycin is maintained at 37 °C $\pm$ 4 °C prior to and during administration.

[0038] In some embodiments, the container of the lyophilized Annamycin is a vial having at least 4 mL of headspace after the addition of the pre-warmed saline to allow sufficient mixing to form the liposomes. In some embodiments, the vial is a 50 mL size.

[0039] In some embodiments, the method of preparing an effective dose of a reconstituted liposomal formulation of Annamycin comprises:

- warming syringes and non-PVC IV bags, each containing saline (about) 0.9% aqueous injection USP sodium chloride at 37 °C $\pm$ 4 °C for at least about three hours, in an oven or bag warmer, wherein the volume of saline in mL in a syringe is equal to the weight in mg of preliposomal lyophilizate Annamycin to be reconstituted, and
- the volume of saline within said IV bag equivalent to the total amount of saline in all syringes, and
- the volume of said IV bag is of sufficient capacity to contain the total volume of saline and liposomal Annamycin. for dilution of the total dose of liposomal Annamycin;

removing a sufficient number of vials containing preliposomal Annamycin lyophilizate to provide the desired total dose of Annamycin from the freezer and allowing the vials to stand at room temperature for 30 minutes to 2 hours;

vigorously shaking each vial of preliposomal lyophilizate Annamycin to break up any dry cake;

injecting air into each vial from air filled syringes until pressure in each vial is equalized;

adding a volume of $37^{\circ}\text{C} \pm 4^{\circ}\text{C}$ saline from a syringe kept at $37^{\circ}\text{C} \pm 4^{\circ}\text{C}$ in an oven to a vial containing a weight equivalent vial of preliposomal lyophilizate Annamycin such that the final Annamycin concentration is approximately 1 mg/mL, maintaining negative pressure throughout the addition of saline;

immediately shaking the vial back and forth vigorously by hand for 15 seconds, then placing the vial in a resealable polyethylene storage bag;

repeating the steps of adding saline, shaking the vial, and placing the vial in the storage bag, until all vials required for the total dose of Annamycin are similarly prepared;

shaking, for a second time, all vials, 2-4 at a time for 1-2 minutes;

storing all vials of reconstituted liposomal Annamycin at $37^{\circ}\text{C} \pm 4^{\circ}\text{C}$ until ready for dilution, for 30 minutes up to 1 hour;

removing vials of reconstituted liposomal Annamycin from storage at $37^{\circ}\text{C} \pm 4^{\circ}\text{C}$ no more than two at a time, and after gently inverting each vial once, withdrawing a foam-free suspension from the vial while maintaining negative pressure throughout the withdrawal of the suspension;

introducing the withdrawn foam-free suspension of reconstituted liposomal Annamycin slowly into a pre-warmed non-PVC IV bag of saline, prepared as described in above and repeating this process for the second vial;

repeating the steps of removing the vials, withdrawing the suspension, and introducing the suspension into the pre-warmed IV bag, until all vials of reconstituted liposomal Annamycin required for the total dose of Annamycin have been introduced into the non-PVC IV bag, providing an IV bag containing the final reconstituted liposomal Annamycin with a concentration of about 0.5 mg/mL; and

agitating the saline and reconstituted liposomal Annamycin in the IV bag by gently inverting the container several times, without generating foam.

[0040] In some embodiments, the preliposomal Annamycin lyophilizate is comprised of Annamycin, one or more lipids, and one or more non-ionic surfactants. In some embodiments, the lipids comprise DMPC and DMPG, and the non-ionic surfactant comprises a polysorbate-type surfactant formed from the ethoxylation of sorbitan followed by the addition of a carboxylic acid. In some embodiments, the non-ionic surfactant comprises polyoxyethylene sorbitan monolaurate. In some embodiments, the preliposomal Annamycin lyophilizate comprises 1.8 – 2.2 wt % Annamycin; 3.0 – 3.4 wt. % Polysorbate 20; and 94.4 - 95.2 wt. % of lipids selected from DMPC and DMPG. In some embodiments, the DMPC is 65.3 -67.3 wt. % and the DMPG is 27.1 – 29.9 wt. %.

[0041] Also provided is a method of treatment via infusion to patients in need thereof, of an effective dose of a reconstituted liposomal Annamycin, prepared from preliposomal Annamycin lyophilizate by the above described method wherein the temperature of the liposomal formulation was maintained at 37 °C $\pm$ 4 °C throughout reconstitution and administration and used within 24 hours.

[0042] Also provided is the use of the above described reconstituted liposomal Annamycin for the treatment of cancer comprising administering to a patient in need thereof an effective dose of diluted liposomal Annamycin prepared as described above. In some embodiments, the cancer is selected from breast cancer, leukemia, acute myeloid leukemia, acute lymphoblastic leukemia, gastric cancer, Hodgkin and non-Hodgkin lymphoma, ovarian cancer, thyroid cancer, colon cancer, traditional cell bladder cancer, and Wilms tumor.

[0043] The reconstituted liposomal Annamycin prepared as described herein is useful for treatment of certain human cancer, such as breast cancer; leukemia, including acute myeloid leukemia (AML) and acute lymphoblastic leukemia; gastric cancer; Hodgkin and non-Hodgkin lymphoma; ovarian cancer; thyroid cancer; colon cancer; traditional cell bladder cancer; and Wilms tumor.

Dosages and routes of administration.

[0044] In general, the reconstituted liposomal formulation is used within 24 h of reconstitution for the treatment of patients with the conditions for which treatment would be beneficial. Administering the drug intravenously is achieved by standard means, and depending on dosage amount, is completed within 110 to 130 min.

[0045] Liposomal Annamycin is used in methods of treating cancer and in methods of inhibiting the growth of tumors in mammals, particularly in humans. Cancer that may be treated include leukemias and lymphomas. In particular, Annamycin may be used in the treatment of leukemias including acute myeloid leukemia (AML), chronic myeloid leukemia (CML), acute lymphocytic leukemia (ALL), and chronic lymphocytic leukemia (CLL). Liposomal Annamycin may be used in the treatment of lymphomas including Hodgkin and non-Hodgkin lymphoma. The methods involve administering to a mammal an effective amount of the drug composition.

[0046] The administering step can suitably be parenteral and by intravenous, intraarterial, intramuscular, intralymphatic, intraperitoneal, subcutaneous, intrapleural, intrathecal injection, or by topical application dosage. In some embodiments the Annamycin is administered intravenously. In some embodiments, such administration is a repeated dose regimen until tumor regression or disappearance is achieved, and may be used in conjunction with forms of tumor therapy such as surgery or chemotherapy with different agents. In some embodiments, the dose administered of a composition is between approximately 120 and 250 mg/m² with respect the mammalian subject to which it is administered.

[0047] The invention is further illustrated by the following examples, which are not to be construed in any way as imposing limitations upon the scope thereof. In the contrary, it is to be clearly understood that resort

may be had to various other embodiments, modifications, and equivalents thereof which, after reading the description herein, may suggest themselves to those skilled in the art without departing from the spirit of the present invention and/or scope of the appended claims.

EXAMPLE 1

General Methods

A. Materials

[0048] Preliposomal Annamycin lyophilizate was prepared as a powder comprising DMPC, DMPG, and Polysorbate 20, and Annamycin.

[0049] Dimyristoyl phosphatidylcholine (DMPC) and 1,2-dimyristoyl-sn-glycero-3-[phosphor-rac-(1-glycerol)] sodium salt (DMPG) were obtained as a dry powder from Nippon Fine Chemicals, Inc., Osaka, Japan.

[0050] Annamycin, as a THF solvate (>95% pure) (MW=640.39) was synthesized as previously described with slight modifications (Horton, D., Priebe, W. 4-demethoxy-3'-desamino-2'-halo-anthracyclines and pharmaceutical compositions containing same. U.S. Pat. No. 4,537,882, 1985.)

[0051] Normal saline was obtained from Abbott Laboratories, North Chicago, Ill.

[0052] In the following example, the reconstitution of pre-liposomal Annamycin was carried out according to the following method:

Step 1: Preparation of Saline Syringes and IV Bag

[0053] All procedures were carried out in a laminar flow hood or clean room utilizing sterile technique.

[0054] Syringes (45 mL) of saline (0.9% sodium chloride, injection USP) were prepared, one for each 45 mg vial of liposomal Annamycin to be used. These syringes of saline were used for reconstitution of the preliposomal Annamycin. For example, for a 360 mg of total dose of liposomal Annamycin, eight syringes were prepared.

[0055] The syringes were placed in a 37 °C oven or IV bag warmer for at least 2-3 hours.

[0056] A non-PVC IV bag was prepared containing an additional volume of saline, equal to the total volume of saline in the syringes used for reconstitution of preliposomal Annamycin. The size of the IV bag was of sufficient volume to contain twice the volume of saline in the syringes. For example, if the total dose of study drug was 360 mg, eight (8) 45-mg vials of preliposomal Annamycin were required, each requiring 45 mL of saline or 360 mL total saline. Thus, a non-PVC IV bag was prepared which also contains 360 mL saline. This was accomplished by removal of 640 mL of saline from a standard 1,000 mL non-PVC IV bag of saline. A 1000 mL IV bag was of sufficient size to hold twice the volume of the saline remaining in the bag, i.e., $2 \times 360 \text{ mL} = 720 \text{ mL}$.

[0057] The prepared saline IV bag was warmed, in a commercially available IV bag warmer which had been preheated per manufacturer instructions to approximately 37 °C or an oven heated to 37 °C. The IV bag

was placed in the bag warmer or oven prior to use in the subsequent steps for approximately 2-3 h, so that the IV bag reached a temperature of about 37 °C.

Step 2: Reconstitution of Preliposomal Annamycin Lyophilizate

[0058] Vials of preliposomal Annamycin lyophilizate (45 mg of Annamycin, not including THF solvate weight, each), with a combined weight sufficient to provide the total required dose of drug, were removed from the freezer and allowed to stand at room temperature from about 30 minutes to about 2 hours. The vials were vigorously shaken to break up any dry cake of preliposomal lyophilizate Annamycin that may have formed.

[0059] The vials were moved to a laminar flow hood, and air was injected into each by means of air-filled syringe until the air pressure was equalized. Syringes of 45 mL saline were removed one at a time and utilized immediately for reconstitution from the 37 °C oven or IV bag warmer preheated to 37 °C. The bag warmer or oven was positioned in the hood or next to the hood where the lyophilized preliposomal Annamycin was reconstituted in order to minimize possible cooling. The saline used for reconstitution was at 37 °C (± 4 °C) when mixed with the lyophilized drug product in the vials.

[0060] The syringes were removed from the bag warmer or oven one at a time, immediately prior to mixing with the preliposomal Annamycin lyophilizate. For example, a syringe of 45 mL saline was removed from the 37 °C IV bag warmer or oven, and 45 mL of 37 °C saline was added to the first 45-mg vial of preliposomal Annamycin lyophilizate, providing a final Annamycin concentration of approximately 1 mg/mL. Following addition normal pressure was achieved by use of a needle just piercing the septum. The needle was removed and the vial was immediately shaken vigorously by hand for two minutes, placed in a zip-locked plastic bag and placed back into the oven or bag warmer at 37 °C.

[0061] In a similar manner, additional syringes of 45 mL saline was then removed, one at a time, from the 37 °C IV bag warmer or oven, and 45 mL of 37 °C saline was added to each additional 45-mg vial of preliposomal Annamycin lyophilizate, shaken and placed in the sealed plastic bag. Each vial was placed in its own plastic bag and placed in the oven or bag warmer until all vials required for the total dose of preliposomal Annamycin were reconstituted.

[0062] When all vials have been reconstituted and placed in the oven or bag warmer, two to four vials were removed and shaken for an additional 1-2 minutes and then placed back in the oven or bag warmer. This was done until all vials were shaken a second time. The vials were allowed to stand for 30 min to one hour at 37 °C degrees before commencing dilution into the final IV bag as described in Step 2 below.

[0063] Particle Size Analysis of samples of the reconstituted Annamycin were carried out and determined to consistently have a mean particle size of about 8 μ m, 50 % distributed in the range of from about 5-10 μ m and 90% distributed in the range of about 2-20 μ m.

Step 3: Dilution of Liposomal Annamycin into the IV Bag

[0065] Dilution of the liposomal Annamycin was completed within approximately one hour of being reconstituted. Vials of reconstituted liposomal Annamycin, prepared as described in Step 2, were maintained

at about 37 °C prior to transfer to the IV bag for dilution, and were not to be taken out of the 37 °C oven until immediately prior to the dilution.

[0066] A vial (45 mg in 45 mL saline, one at a time) of reconstituted liposomal Annamycin, prepared as described above in Step 2, containing the measured amount of saline at about 37°C for dilution. was removed from the 37 °C oven. The vial was gently inverted, and the liposomal suspension was withdrawn via syringe. If necessary to equalize the pressure, the septum was pierced for 1 second, the needle was removed, and the withdrawal of the drug was continued. The suspension was immediately transferred slowly (so as to not generate foam) into the pre-warmed non-PVC IV bag prepared as in Step 1. Care was taken to avoid transferring any foam.

[0067] In similar fashion, the additional vials, sufficient to provide the required total dose of preliposomal Annamycin, were removed from the oven, one at a time, gently inverted, the liposomal Annamycin suspension was withdrawn via syringe and immediately transferred slowly to the contents of the pre-warmed non-PVC IV bag until all vials were transferred to the non-PVC IV bag containing saline for dilution. The saline and liposomal Annamycin in the IV bag were mixed by gently inverting the container several times, avoiding the generation of foam. The time was recorded on the IV bag. The final liposomal Annamycin concentration was approximately 0.5 mg/mL.

[0068] Throughout this step, the reconstituted liposomal Annamycin diluted IV bag was maintained at about 37 °C by immediately transferring the IV bag to the prewarmed IV bag warmer.

[0069] The IV bag warmer, when ready to be transferred to the patient infusion room was unplugged, transferred in 30 minutes – 60 min and plugged in immediately near the patient for infusion. The diluted liposomal Annamycin suspension was maintained at about 37 °C leading up to and during the patient infusion.

[0070] The reconstituted liposomal Annamycin diluted with saline was stable for 24 hours when held at about 37 °C after which drug was not acceptable for administration to subjects. Only non-PVC and non-DEHP tubing were used for drug administration.

[0071] Care was taken to keep track of the time from the start of reconstitution until the completion of the 2 hour infusion. No more than 24 hours was allowed to lapse from the time of reconstitution to the time of completing the infusion.

EXAMPLE 2

Temperature Studies

[0072] An appearance test was performed on each sample. Additional appearance observations were made immediately after reconstitution of sample material at each temperature. Any changes in appearance that occurred over the duration of the particle size testing were also noted.

[0073] The appearance results obtained prior to and immediately after reconstitution of the sample material are shown below along with appearance after particle size testing. The appearances prior to reconstitution

results for all samples were identical to the target limit. However, the appearance following reconstitution was dependent on temperature. As the sample temperature increases, the likelihood of undispersed particles decreases. Clusters of undispersed particles were seen at temperatures between 20 – 25°C, while temperatures between 28 – 42°C showed no evidence of these clusters. Additionally the table shows changes in appearance following reconstitution at the end of particle size testing were limited to those at lower temperatures. These lower temperature samples had a tendency to disperse the particle clusters that were present immediately after reconstitution, though not always entirely. Unsurprisingly, the higher temperature sample's appearance remained the same because there were no undispersed clusters of particles present immediately after reconstitution.

Temperature Variation	Appearance Prior to Reconstitution	Appearance Following Reconstitution	Additional Observations*
Nominal (37°C)	Orange powder, free from visible contamination	Orange milky suspension; free from visible contamination of foreign material	No significant change
20° C	Orange powder, free from visible contamination	Orange milky suspension; free from visible contamination of foreign material, clusters of undispersed, un-suspended sample particles present	Fewer cluster present
23° C	Orange powder, free from visible contamination	Orange milky suspension; free from visible contamination of foreign material, one cluster of undispersed, un-suspended sample particles present	No significant change
25° C	Orange powder, free from visible contamination	Orange milky suspension; free from visible contamination of foreign material, one cluster of undispersed, un-suspended sample particles present	No clusters present
28° C	Orange powder, free from visible contamination	Orange milky suspension; free from visible contamination of foreign mate	No significant change
30° C	Orange powder, free from visible contamination	Orange milky suspension; free from visible contamination of foreign material	No significant change
42° C	Orange powder, free from visible contamination	Orange milky suspension; free from visible contamination of foreign mater	No significant change

*These observations were made at the time of sample analysis, between approximately 1 – 2 hours after appearance following reconstitution

[0074] The particle size study was performed using a laser diffraction particle size analyzer. A single sample was prepared and analyzed for each of the temperatures 37 (Nominal), 20, 23, 25, 28, 30, and 42 °C. Each step of the sample preparation (i.e. incubation temperature, etc...) and analysis (i.e. instrument dispersant temperature, etc...) was performed at the specified temperature.

[0075] The results obtained from the particle size analysis temperature study are shown below. Histogram result overlays are shown in Figures 2 – 4. Overall, particle size results for samples between 20 – 25°C were

significantly different than the target limits, while results for samples between 28 – 42°C were similar. The extremely large particle size results for the 20°C sample suggests the sample material did not form liposomes upon reconstitution. Results for samples between 23 – 25°C were significantly smaller than the target limits, signifying the liposomes are in the processes of being formed. Results for samples between 28 – 42°C are much more stable, and are similar to target limits. This indicates the liposomes have fully formed and are fairly stable in this temperature range. However, it should be noted that the 28°C sample results do not fall within the target limits. This highlights that while there is a particle size transition that occurs in the temperature range of 25 - 28°C, the liposomes may not have entirely formed until a temperature of approximately 30°C has been reached. Finally, it should be noted that when viewing the results for the 28 – 42°C samples, the Dv90 result increases as temperature increases.

Vial	Target Temp. (°C)	Actual Analysis Temp. (°C)	Dv10 (µm)		Dv50 (µm)		Dv90(µm)	
			Rounded	Full Precision	Rounded	Full Precision	Rounded	Full Precision
1	37*	37.2	3	2.77	6	5.66	10	9.90
4	20	20.3	17	17.2	69	68.9	165	165
% Difference			467	521	1050	1117	1550	1567
5	23	22.8	0	0.383	1	0.872	8	7.67
% Difference			100	86	83	85	20	23
7	25	25.2	0	0.435	1	1.01	3	3.33
% Difference			100	84	83	82	70	66
6	28	28.0	1	0.796	4	4.27	8	7.63
% Difference			67	71	33	25	20	23
3	30	29.6	3	2.53	5	5.19	9	8.66
% Difference			0	9	17	8	10	13
2	42	41.9	2	2.15	6	5.60	10	10.4
% Difference			33	22	0	1	0	5

*Nominal

Note: Full precision values are shown to 3 significant figures for improved data interpretation

[0076] The appearance and particle size results both show temperature has an impact on reconstituted drug product liposomes particle size distribution. Reconstitution at temperatures between 20 – 25°C leaves clusters of particles undispersed, while the particle size results for these temperatures vary greatly. Appearance and particle size results for temperatures between 28 – 42°C are much more stable, indicating a transitional temperature range of approximately 25 – 28°C. All of this data indicates the drug product should be reconstituted and maintained at a temperature between approximately 30 – 42°C.

[0077] While the invention has been described with reference to particular embodiments and examples, those skilled in the art recognize that various modifications may be made to the invention without departing from the spirit and scope thereof.

[0078] All references cited herein, including patents, patent applications, and publications, are incorporated herein by reference, in their entirety. Whether previously specifically incorporated or not.

[0079] The various features and embodiments of the present invention, referred to in individual section above apply, as appropriate, to other sections, mutatis mutandis. Consequently, features specified in one section may be combined with features specified in other sections, as appropriate.

[0080] The foregoing description of some specific embodiments provides sufficient information that others can, by applying current knowledge, readily modify or adapt for various applications such specific embodiments without departing from the generic concept, and, therefore, such adaptations and modifications should and are intended to be comprehended within the meaning and range of equivalents of the disclosed embodiments. It is to be understood that the phraseology or terminology employed herein is for the purpose of description and not of limitation. In the drawings and the description, there have been disclosed exemplary embodiments and, although specific terms may have been employed, they are unless otherwise stated used in a generic and descriptive sense only and not for the purposes of limitation of the scope of the claims. Moreover, one skilled in the art will appreciate that certain steps of the methods discussed herein may be sequenced in alternative order or steps may be combined. Therefore, it is intended that the appended claims not be limited to the particular embodiment disclosed herein. Those skilled in the art will recognize, or be able to ascertain using no more than routine experimentation, many equivalents to the embodiments of the invention described herein. Such equivalents are encompassed by the following claims.