TNF-α BINDING PROTEINS

This application claims priority to U.S. Provisional Application No. 61/420,999, filed on December 8, 2010.

Field

TNF-α binding proteins, and to their uses in the prevention and/or treatment of acute and chronic immunological diseases such as rheumatoid arthritis, osteoarthritis, psoriasis, multiple sclerosis, and other autoimmune diseases are provided.

Background

There is a need in the art for improved antibodies capable of binding TNF-α.

Summary

A novel family of binding proteins, CDR grafted antibodies, humanized antibodies, and fragments thereof, capable binding TNF-α, binding TNF-α with high affinity, and binding and neutralizing TNF-α are provided. A antibodies, and antigen binding portions thereof, capable of binding TNF-α comprising an amino acid sequence of any one of SEQ ID NO: 31-46 are also provided.

Detailed Description

TNF-α binding proteins, e.g., antibodies or antigen-binding portions thereof, that bind TNF-α are provided. Various aspects relate to antibodies and antibody fragments, and pharmaceutical compositions thereof, as well as nucleic acids, recombinant expression vectors and host cells for making such antibodies and fragments thereof. Methods of using the binding proteins to detect human TNF-α, to inhibit human TNF-α, either in vitro or in vivo, and to regulate gene expression are also encompassed.

Unless otherwise defined herein, scientific and technical terms used herein shall have the meanings that are commonly understood by those of ordinary skill in the art. The meaning and scope of the terms should be clear, however, in the event of any latent ambiguity, definitions provided herein take precedent over any dictionary or extrinsic definition. Further, unless otherwise required by context, singular terms shall include the plural thereof and plural terms shall include the singular thereof. The term "or" includes "and/or" unless stated otherwise. The use of the term "including", "includes" or "included" is not limiting. Also, terms such as "element" or "component" encompass both elements and components comprising one unit and elements and components that comprise more than one subunit unless specifically stated otherwise.

The methods and techniques are generally performed according to conventional methods well known in the arts of cell and tissue culture, molecular biology, immunology, microbiology, genetics, protein and nucleic acid chemistry and hybridization, analytical chemistry, synthetic organic chemistry, medicinal and pharmaceutical chemistry, pharmaceutical preparation, formulation, and delivery and treatment of patients. Such techniques are also described in references cited herein. Enzymatic reactions and purification techniques are performed according to manufacturer's specifications, as commonly known in the art or as otherwise described herein.

The terms "polypeptide," "peptide" and "protein" are used interchangeably to mean a polymeric chain of amino acids. The term "polypeptide" encompasses native or artificial proteins, protein fragments and analogs of a protein sequence. A polypeptide may be monomeric or polymeric.

The term "isolated protein" or "isolated polypeptide" means a protein or polypeptide that is not associated with components that accompany it in its native state, e.g., it is substantially free of other proteins or cellular components from the same species, is expressed by a cell from a different species, or does not occur in nature. Thus, a polypeptide that is chemically synthesized or synthesized in a cellular system different from the cell from which it naturally originates will be "isolated" from its naturally associated components. A protein may also be rendered substantially free of naturally associated components by isolation, using protein purification techniques well known in the art.

The term "recovering" refers to the process of rendering a chemical species such as a polypeptide substantially free of naturally associated components by isolation, e.g., using protein purification techniques well known in the art.

The term "human TNF-α" (abbreviated herein as hTNF-α) includes a dimeric cytokine protein. The term includes a homotrimeric protein comprising three 17.5 kD TNF-α proteins. The homotrimeric protein is referred to as a "TNF-α protein". The term human "TNF-α" is intended to include recombinant human TNF-α (TNF-α) which can be prepared by standard recombinant expression methods. The sequence of human TNF-α is shown in Table 1.

Table 1: Sequence of Human TNF-α

Protein	Sequence Identifier	Sequence
		12345678901234567890123456789**0**
Human TNF-α	**SEQ ID NO.:1**	VRSSSRTPSDKPVAHVVANPQAEGQLQWLN DRANALLANGVELRDNQLVVPSEGLYLIYS QVLFKGQGCPSTHVLLTHTISRIAVSYQTK VNLLSAIKSPCQRETPEGAEAKPWYEPIYL GGVFQLEKGDRLSAEINRPDYLDFAESGQV YFGIIAL

The term “biological activity” refers to all inherent biological properties of a molecule. Biological properties of TNF-α include but are not limited to binding the TNF receptor.

The terms "specific binding" or "specifically binding" in reference to the interaction of an antibody, a protein, or a peptide with a second chemical species, mean that the interaction is dependent upon the presence of a particular structure (e.g., an antigenic determinant or epitope) on the chemical species; for example, an antibody recognizes and binds to a specific protein structure rather than to proteins generally. If an antibody is specific for epitope "A", the presence of a molecule containing epitope A (or free, unlabeled A), in a reaction containing labeled "A" and the antibody, will reduce the amount of labeled A bound to the antibody.

The term “binding protein” includes, but is not limited to, any antibody, or antigen binding portion thereof. Binding proteins also include other constructs that maintain a binding affinity to a target. In some instances, those binding proteins may have structural similarities to antibodies, or antigen binding portions thereof, and they may also have structural differences that would distinguish them from antibodies, or antigen binding portions thereof.

The term "antibody” broadly refers to any immunoglobulin (Ig) molecule comprised of four polypeptide chains, two heavy (H) chains and two light (L) chains, or any functional fragment, mutant, variant, or derivative thereof that retains the essential epitope binding features of an Ig molecule. Such mutant, variant, or derivative anitbody formats are known in the art, nonlimiting embodiments of which are discussed below.

In a full-length antibody, each heavy chain is comprised of a heavy chain variable region (abbreviated herein as HCVR or VH) and a heavy chain constant region. The heavy chain constant region is comprised of three domains, CH1, CH2 and CH3. Each light chain is comprised of a light chain variable region (abbreviated herein as LCVR or VL) and a light chain constant region. The light chain constant region is comprised of one domain, CL. The VH and VL regions can be further subdivided into regions of hypervariability, termed complementarity determining regions (CDR), interspersed with regions that are more conserved, termed framework regions (FR). Each VH and VL is composed of three CDRs and four FRs, arranged from amino-terminus to carboxy-terminus in the following order: FR1, CDR1, FR2, CDR2, FR3, CDR3, FR4. Immunoglobulin

molecules can be of any type (e.g., IgG, IgE, IgM, IgD, IgA and IgY), class (e.g., IgG 1, IgG2, IgG 3, IgG4, IgA1 and IgA2) or subclass.

The term "antigen-binding portion" of an antibody refers to one or more fragments of an antibody that retain the ability to specifically bind to an antigen (e.g., hTNFα). The antigen-binding function of an antibody can have performed by fragments of a full-length antibody. Such antibody embodiments may also be bispecific, dual specific, or multi-specific formats; specifically binding to two or more different antigens. Examples of binding fragments encompassed within the term "antigen-binding portion" of an antibody include (i) an Fab fragment, a monovalent fragment consisting of the VL, VH, CL and CH1 domains; (ii) an $F(ab')_2$ fragment, a bivalent fragment comprising two Fab fragments linked by a disulfide bridge at the hinge region; (iii) a Fd fragment consisting of the VH and CH1 domains; (iv) an Fv fragment consisting of the VL and VH domains of a single arm of an antibody, (v) a dAb fragment (Ward et al., (1989) Nature 341:544-546, Winter et al., PCT Publication No. WO 90/05144), which comprises a single variable domain; and (vi) an isolated complementarity determining region (CDR). Furthermore, although the two domains of the Fv fragment, VL and VH, are coded for by separate genes, they can be joined, using recombinant methods, by a synthetic linker that enables them to be made as a single protein chain in which the VL and VH regions pair to form monovalent molecules (known as single chain Fv (scFv); see e.g., Bird et al. (1988) Science 242:423-426; and Huston et al. (1988) Proc. Natl. Acad. Sci. USA 85:5879-5883). Such single chain antibodies (scFv) are also intended to be encompassed within the term "antigen-binding portion" of an antibody. Other forms of single chain antibodies, such as diabodies are also encompassed. Diabodies are bivalent, bispecific antibodies in which VH and VL domains are expressed on a single polypeptide chain, but using a linker that is too short to allow for pairing between the two domains on the same chain, thereby forcing the domains to pair with complementary domains of another chain and creating two antigen binding sites (see, e.g., Holliger et al. (1993) Proc. Natl. Acad. Sci. USA 90:6444-6448; Poljak et al. (1994) Structure 2:1121-1123). Such antibody binding portions are known in the art (Kontermann and Dubel eds., Antibody Engineering (2001) Springer-Verlag. New York. 790 pp. (ISBN 3-540-41354-5).

The term “antibody construct” refers to a polypeptide comprising one or more the antigen binding portions linked to a linker polypeptide or an immunoglobulin constant domain. Linker polypeptides comprise two or more amino acid residues joined by peptide bonds and are used to link one or more antigen binding portions. Such linker polypeptides are well known in the art (see, e.g., Holliger et al. (1993) Proc. Natl. Acad. Sci. USA 90:6444-6448; Poljak et al. (1994) Structure 2:1121-1123). An immunoglobulin constant domain refers to a heavy (gamma) or light

chain (kappa and delta) constant domain. Human IgG heavy chain and light chain constant domain amino acid sequences are known in the art and represented in Table 2.

Table 2: Sequence Of Human IgG Heavy Chain Constant Domain And Light Chain Constant Domain

Protein	Sequence Identifier	Sequence
		123456789**0**123456789**0**123456789**0**12
Ig gamma-1 constant region	**SEQ ID NO.:2**	ASTKGPSVFFLAPSSKSTSGGTAALGCLVKDY FPEPVTVSWNSGALTSGVHTFPAVLQSSGLYS LSSVVTVPSSSLGTQTYICNVNHKPSNTKVDK KVEPKSCDKTHTCPPCPAPELLGGPSVFLFPP KPKDTLMISRTPEVTCVVVDVSHEDPEVKFNW YVDGVEVHNAKTKPREEQYNSTYRVVSVLTVL HQDWLNGKEYKCKVSNKALPAPIEKTISKAKG QPREPQVYTLPPSREEMTKNQVSLTCLVKGFY PSDIAVEWESNGQPENNYKTTPPVLDSDGSFF LYSKLTVDKSRWQQGNVFSCSVMHEALHNHYT QKSLSLSPGK
Ig gamma-1 constant region mutant	**SEQ ID NO.:3**	ASTKGPSVFPLAPSSKSTSGGTAALGCLVKDY FPEPVTVSWNSGALTSGVHTFPAVLQSSGLYS LSSVVTVPSSSLGTQTYICNVNHKPSNTKVDK KVEPKSCDKTHTCPPCPAPEAAGGPSVFLFPP KPKDTLMISRTPEVTCVVVDVSHEDPEVKFNW YVDGVEVHNAKTKPREEQYNSTYRVVSVLTVL HQDWLNGKEYKCKVSNKALPAPIEKTISKAKG QPREPQVYTLPPSREEMTKNQVSLTCLVKGFY PSDIAVEWESNGQPENNYKTTPPVLDSDGSFF LYSKLTVDKSRWQQGNVFSCSVMHEALHNHYT QKSLSLSPGK
Ig Kappa constant region	**SEQ ID NO.:4**	TVAAPSVFIFPPSDEQLKSGTASVVCLLNNFY PREAKVQWKVDNALQSGNSQESVTEQDSKDST YSLSSTLTLSKADYEKHKVYACEVTHQGLSSP VTKSFNRGEC
Ig Lambda constant region	**SEQ ID NO.:5**	QPKAAPSVTLFPPSSEELQANKATLVCLISDF YPGAVTVAWKADSSPVKAGVETTTPSKQSNNK YAASSYLSLTPEQWKSHRSYSCQVTHEGSTVE KTVAPTECS

Still further, an antibody or antigen-binding portion thereof may be part of a larger immunoadhesion molecule, formed by covalent or noncovalent association of the antibody or antigen binding portion with one or more other proteins or peptides. Examples of such immunoadhesion molecules include use of the streptavidin core region to make a tetrameric scFv molecule (Kipriyanov et al. (1995) Human Antibod. Hybridom. 6:93-101) and use of a cysteine residue, a marker peptide and a C-terminal polyhistidine tag to make bivalent and biotinylated scFv molecules (Kipriyanov et al. (1994) Mol. Immunol. 31:1047-1058). Antigen binding portions of antibodies, such as Fab and $F(ab')_2$ fragments, can be prepared from whole antibodies using conventional techniques, such as papain or pepsin digestion, respectively, of whole antibodies. Moreover, antibodies, antigen binding portions thereof and immunoadhesion molecules can be obtained using standard recombinant DNA techniques, as described herein.

The term "isolated antibody" refers to an antibody that is substantially free of other antibodies having different antigenic specificities (e.g., an isolated antibody that specifically binds hTNFα is substantially free of antibodies that specifically bind antigens other than hTNFα). An isolated antibody that specifically binds hTNFα may, however, have cross-reactivity to other antigens, such as TNFα molecules from other species. Moreover, an isolated antibody may be substantially free of other cellular material and/or chemicals.

The term "human antibody" includes antibodies having variable and constant regions derived from human germline immunoglobulin sequences. The human antibodies may include amino acid residues not encoded by human germline immunoglobulin sequences (e.g., mutations introduced by random or site-specific mutagenesis in vitro or by somatic mutation in vivo), for example in the CDRs and in particular CDR3. However, the term "human antibody" is not intended to include antibodies in which CDR sequences derived from the germline of another mammalian species, such as a mouse, have been grafted onto human framework sequences.

The term "recombinant human antibody" is intended to include all human antibodies that are prepared, expressed, created or isolated by recombinant means, such as antibodies expressed using a recombinant expression vector transfected into a host cell (described further in Section II C, below), antibodies isolated from a recombinant, combinatorial human antibody library (Hoogenboom (1997) TIB Tech. 15:62-70; Azzazy and Highsmith (2002) Clin. Biochem. 35:425-445; Gavilondo and Larrick (2002) BioTechniques 29:128-145; Hoogenboom and Chames (2000) Immunol. Today 21:371-378), antibodies isolated from an animal (e.g., a mouse) that is transgenic for human immunoglobulin genes (see, e.g., Taylor et al. (1992) Nucl. Acids Res. 20:6287-6295; Kellermann and Green (2002) Current Opin. Biotechnol. 13:593-597; Little et al. (2000) Immunol. Today 21:364-370) or antibodies prepared, expressed, created or isolated by any other means that involves splicing of human immunoglobulin gene sequences to other DNA sequences. Such recombinant human antibodies have variable and constant regions derived from human germline immunoglobulin sequences. In certain embodiments, however, such recombinant human antibodies are subjected to in vitro mutagenesis (or, when an animal transgenic for human Ig sequences is used, in vivo somatic mutagenesis) and thus the amino acid sequences of the VH and VL regions of the recombinant antibodies are sequences that, while derived from and related to human germline VH and VL sequences, may not naturally exist within the human antibody germline repertoire in vivo.

The term “chimeric antibody” refers to antibodies that comprise heavy and light chain variable region sequences from one species and constant region sequences from another species, such as antibodies having murine heavy and light chain variable regions linked to human constant regions.

The term "CDR-grafted antibody" refers to antibodies that comprise heavy and light chain variable region sequences from one species but in which the sequences of one or more of the CDR regions of VH and/or VL regions are replaced with CDR sequences of another species, such as antibodies that have human heavy and light chain variable regions in which one or more of the human CDRs (e.g., CDR3) has been replaced with murine CDR sequences.

The term "CDR" refers to the complementarity determining region within antibody variable sequences. There are three CDRs in each of the variable regions of the heavy chain and the light chain, which are designated CDR1, CDR2, and CDR3, for each of the variable regions. The term "CDR set" refers to a group of three CDRs that occur in a single variable region (i.e., VH or VL) of an antigen binding site. The exact boundaries of these CDRs have been defined differently according to different systems. The system described by Kabat (Kabat et al. (1987, 1991) Sequences of Proteins of Immunological Interest (National Institutes of Health, Bethesda, Maryland) not only provides an unambiguous residue numbering system applicable to any variable region of an antibody, but also provides precise residue boundaries defining the three CDRs. These CDRs may be referred to as Kabat CDRs. Chothia and coworkers (Chothia and Lesk (1987) J. Mol. Biol. 196:901-917 and Chothia et al. (1989) Nature 342:877-883) found that certain sub-portions within Kabat CDRs adopt nearly identical peptide backbone conformations, despite having great diversity at the level of amino acid sequence. These sub-portions were designated as L1, L2, and L3 or H1, H2, and H3, where the "L" and the "H" designates the light chain and the heavy chains regions, respectively. These regions may be referred to as Chothia CDRs, which have boundaries that overlap with Kabat CDRs. Other boundaries defining CDRs overlapping with the Kabat CDRs have been described by Padlan et al. (1995) FASEB J. 9:133-139 and MacCallum (1996) J. Mol. Biol. 262(5):732-745). Still other CDR boundary definitions may not strictly follow one of the above systems, but will nonetheless overlap with the Kabat CDRs, although they may be shortened or lengthened in light of prediction or experimental findings that particular residues or groups of residues or even entire CDRs do not significantly impact antigen binding. The methods used herein may utilize CDRs defined according to any of these systems, although certain embodiments use Kabat or Chothia defined CDRs.

The terms "Kabat numbering", "Kabat definition" and "Kabat labeling" are used interchangeably herein. These terms refer to a system of numbering amino acid residues which are more variable (*i.e.* hypervariable) than other amino acid residues in the heavy and light chain variable regions of an antibody, or an antigen binding portion thereof (Kabat et al. (1971) Ann. NY Acad. Sci. 190:382-391 and Kabat et al. (1991) Sequences of Proteins of Immunological Interest, Fifth Edition, U.S. Department of Health and Human Services, NIH Publication No. 91-3242). For the heavy chain variable region, the hypervariable region ranges from amino acid positions 31 to 35 for CDR1, amino acid positions 50 to 65 for CDR2, and amino acid positions

95 to 102 for CDR3. For the light chain variable region, the hypervariable region ranges from amino acid positions 24 to 34 for CDR1, amino acid positions 50 to 56 for CDR2, and amino acid positions 89 to 97 for CDR3.

The growth and analysis of extensive public databases of amino acid sequences of variable heavy and light regions over the past twenty years have led to the understanding of the typical boundaries between framework regions (FR) and CDR sequences within variable region sequences and enabled persons skilled in this art to accurately determine the CDRs according to Kabat numbering, Chothia numbering, or other systems. See, e.g., Martin, In Kontermann and Dübel, eds., Antibody Engineering (Springer-Verlag, Berlin, 2001), chapter 31, pages 432-433. A useful method of determining the amino acid sequences of Kabat CDRs within the amino acid sequences of variable heavy (VH) and variable light (VL) regions is provided below:

To identify a CDR-L1 amino acid sequence:

Starts approximately 24 amino acid residues from the amino terminus of the VL region;

Residue before the CDR-L1 sequence is always cysteine (C);

Residue after the CDR-L1 sequence is always a tryptophan (W) residue, typically Trp-Tyr-Gln (W-Y-Q), but also Trp-Leu-Gln (W-L-Q), Trp-Phe-Gln (W-F-Q), and Trp-Tyr-Leu (W-Y-L);

Length is typically 10 to 17 amino acid residues.

To identify a CDR-L2 amino acid sequence:

Starts always 16 residues after the end of CDR-L1;

Residues before the CDR-L2 sequence are generally Ile-Tyr (I-Y), but also Val-Tyr (V-Y), Ile-Lys (I-K), and Ile-Phe (I-F);

Length is always 7 amino acid residues.

To identify a CDR-L3 amino acid sequence:

Starts always 33 amino acids after the end of CDR-L2;

Residue before the CDR-L3 amino acid sequence is always a cysteine (C);

Residues after the CDR-L3 sequence are always Phe-Gly-X-Gly (F-G-X-G) (SEQ ID NO:6), where X is any amino acid;

Length is typically 7 to 11 amino acid residues.

To identify a CDR-H1 amino acid sequence:

Starts approximately 31 amino acid residues from amino terminus of VH region and always 9 residues after a cysteine (C);

Residues before the CDR-H1 sequence are always Cys-X-X-X-X-X-X-X-X (SEQ ID NO:7), where X is any amino acid;

Residue after CDR-H1 sequence is always a Trp (W), typically Trp-Val (W-V), but also Trp-Ile (W-I), and Trp-Ala (W-A);

Length is typically 5 to 7 amino acid residues.

To identify a CDR-H2 amino acid sequence:

Starts always 15 amino acid residues after the end of CDR-H1;

Residues before CDR-H2 sequence are typically Leu-Glu-Trp-Ile-Gly (L-E-W-I-G) (SEQ ID NO:8), but other variations also;

Residues after CDR-H2 sequence are Lys/Arg-Leu/Ile/Val/Phe/Thr/Ala-Thr/Ser/Ile/Ala (K/R-L/I/V/F/T/A-T/S/I/A);

Length is typically 16 to 19 amino acid residues.

To identify a CDR-H3 amino acid sequence:

Starts always 33 amino acid residues after the end of CDR-H2 and always 3 after a cysteine (C)'

Residues before the CDR-H3 sequence are always Cys-X-X (C-X-X), where X is any amino acid, typically Cys-Ala-Arg (C-A-R);

Residues after the CDR-H3 sequene are always Trp-Gly-X-Gly (W-G-X-G) (SEQ ID NO:9), where X is any amino acid;

Length is typically 3 to 25 amino acid residues.

The terms "acceptor" and "acceptor antibody" refer to the antibody or nucleic acid sequence providing or encoding at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 98% or 100% of the amino acid sequences of one or more of the framework regions. In some embodiments, the term "acceptor" refers to the antibody amino acid or nucleic acid sequence providing or encoding the constant region(s). In yet another embodiment, the term "acceptor" refers to the antibody amino acid or nucleic acid sequence providing or encoding one or more of the framework regions and the constant region(s). In a specific embodiment, the term "acceptor" refers to a human antibody amino acid or nucleic acid sequence that provides or encodes at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 98%, or 100% of the amino acid sequences of one or more of the framework regions. In accordance with this embodiment, an acceptor may contain at least 1, at least 2, at least 3, least 4, at least 5, or at least 10 amino acid residues that does (do) not occur at one or more specific positions of a human antibody. An acceptor framework region and/or acceptor constant region(s) may be, e.g., derived or obtained from a germline antibody gene, a mature antibody gene, a functional antibody (e.g., antibodies well-known in the art, antibodies in development, or antibodies commercially available).

The term "canonical" residue refers to a residue in a CDR or framework that defines a particular canonical CDR structure as defined by Chothia et al. (1987) J. Mol. Biol. 196:901-907 and Chothia et al. (1992) J. Mol. Biol. 227:799. According to Chothia et al., critical portions of the CDRs of many antibodies have nearly identical peptide backbone confirmations despite great

diversity at the level of amino acid sequence. Each canonical structure specifies primarily a set of peptide backbone torsion angles for a contiguous segment of amino acid residues forming a loop.

The terms "donor" and "donor antibody" refer to an antibody providing one or more CDRs. In a particular embodiment, the donor antibody is an antibody from a species different from the antibody from which the framework regions are obtained or derived. In the context of a humanized antibody, the term "donor antibody" refers to a non-human antibody providing one or more CDRs.

The term "framework" or "framework sequence" refers to the remaining sequences of a variable region minus the CDRs. Because the exact definition of a CDR sequence can be determined by different systems, the meaning of a framework sequence is subject to correspondingly different interpretations. The six CDRs (CDR-L1, -L2, and -L3 of light chain and CDR-H1, -H2, and -H3 of heavy chain) also divide the framework regions on the light chain and the heavy chain into four sub-regions (FR1, FR2, FR3 and FR4) on each chain, in which CDR1 is positioned between FR1 and FR2, CDR2 between FR2 and FR3, and CDR3 between FR3 and FR4. Without specifying the particular sub-regions as FR1, FR2, FR3 or FR4, a framework region, as referred by others, represents the combined FR's within the variable region of a single, naturally occurring immunoglobulin chain. A FR represents one of the four sub- regions, and FRs represents two or more of the four sub- regions constituting a framework region.

Human heavy chain and light chain acceptor sequences are known in the art. In one embodiment, the human heavy chain and light chain acceptor sequences are chosen from the sequences listed from V-base (http://vbase.mrc-cpe.cam.ac.uk/) or from IMGT®, the international ImMunoGeneTics information system® (http://imgt.cines.fr/textes/IMGTrepertoire/LocusGenes/). . In another embodiment, the human heavy chain and light chain acceptor sequences are chosen from the sequences described in Table 3 and Table 4.

Table 3: Heavy Chain Acceptor Sequences

SEQ ID No.	Protein region	Sequence
		123456789**0**123456789**0**123456789**0**12
10	VH1-18 FR1	QVQLVQSGAEVKKPGASVKVSCKASGYTFT
11	VH1-18 FR2	WVRQAPGQGLEWMG
12	VH1-18 FR3	RVTMTTDTSTSTAYMELRSLRSDDTAVYYCAR
13	VH7-4.1 FR1	QVQLVQSGSELKKPGASVKVSCKASGYTFT
14	VH7-4.1FR2	WVRQAPGQGLEWMG
15	VH7-4.1 FR3	RFVFSLDTSVSTAYLQISSLKAEDTAVYYCAR
`16	JH1/JH4/JH5 FR4	WGQGTLVTVSS
17	JH2 FR4	WGRGTLVTVSS
18	JH6 FR4	WGQGTTVTVSS
19	JH3 FR4	WGQGTMVTVSS

Table 4: Light Chain Acceptor Sequences

SEQ ID No.	Protein region	Sequence
		123456789**0**123456789**0**123456789**0**12
20	1-39/O12 FR1	DIQMTQSPSSLSASVGDRVTITC
21	1-39/O12 FR2	WYQQKPGKAPKLLIY
22	1-39/O12 FR3	GVPSRFSGSGSGTDFTLTISSLQPEDFATYYC
23	6D-41/A14 FR1	DVVMTQSPAFLSVTPGEKVTITC
24	6D-41/A14 FR2	WYQQKPDQAPKLLIK
25	6D-41/A14 FR3	GVPSRFSGSGSGTDFTFTISSLEAEDAATYYC
26	JK2 FR4	FGQGTKLEIKR
27	JK5 FR4	FGQGTRLEIKR
28	JK1 FR4	FGQGTKVEIKR
29	JK4 FR4	FGGGTKVEIKR
30	JK3 FR4	FGPGTKVDIKR

The term "germline antibody gene" or "gene fragment" refers to an immunoglobulin sequence encoded by non-lymphoid cells that have not undergone the maturation process that leads to genetic rearrangement and mutation for expression of a particular immunoglobulin. (See, e.g., Shapiro et al. (2002) Crit. Rev. Immunol. 22(3): 183-200; Marchalonis et al. (2001) Adv. Exp. Med. Biol. 484:13-30 (2001)). One of the advantages provided by various embodiments stems from the recognition that germline antibody genes are more likely than mature antibody genes to conserve essential amino acid sequence structures characteristic of individuals in the species, hence less likely to be recognized as from a foreign source when used therapeutically in that species.

The term "key" residues refer to certain residues within the variable region that have more impact on the binding specificity and/or affinity of an antibody, in particular a humanized antibody. A key residue includes, but is not limited to, one or more of the following: a residue that is adjacent to a CDR, a potential glycosylation site (can be either N- or O-glycosylation site), a rare residue, a residue capable of interacting with the antigen, a residue capable of interacting with a CDR, a canonical residue, a contact residue between heavy chain variable region and light chain variable region, a residue within the Vernier zone, and a residue in the region that overlaps between the Chothia definition of a variable heavy chain CDR1 and the Kabat definition of the first heavy chain framework.

The term "humanized antibody" refers to antibodies that comprise heavy and light chain variable region sequences from a non-human species (e.g., a mouse) but in which at least a portion of the VH and/or VL sequence has been altered to be more “human-like”, i.e., more similar to human germline variable sequences. One type of humanized antibody is a CDR-grafted antibody, in which non-human CDR sequences are introduced into human VH and VL sequences to replace the corresponding non-human framework (FR) sequences. For example, a "humanized antibody" is an antibody or a variant, derivative, analog or fragment thereof which immunospecifically binds to an antigen of interest and which comprises a framework (FR) region having substantially the

amino acid sequence of a human antibody and a complementary determining region (CDR) having substantially the amino acid sequence of a non-human antibody. The term "substantially" in the context of a CDR refers to a CDR having an amino acid sequence at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 98% or at least about 99% identical to the amino acid sequence of a non-human antibody CDR. A humanized antibody comprises substantially all of at least one, and typically two, variable domains (Fab, Fab', F(ab') 2, FabC, Fv) in which all or substantially all of the CDR regions correspond to those of a non-human immunoglobulin (i.e., donor antibody) and all or substantially all of the framework regions are those of a human immunoglobulin consensus sequence. In an embodiment, a humanized antibody also comprises at least a portion of an immunoglobulin constant region (Fc), typically that of a human immunoglobulin. In some embodiments, a humanized antibody contains both the light chain as well as at least the variable domain of a heavy chain. The antibody also may include the CH1, hinge, CH2, CH3, and CH4 regions of the heavy chain. In some embodiments, a humanized antibody only contains a humanized light chain. In some embodiments, a humanized antibody only contains a humanized heavy chain. In specific embodiments, a humanized antibody only contains a humanized variable domain of a light chain and/or humanized heavy chain.

The humanized antibody can be chosen from any class of immunoglobulins, including IgM, IgG, IgD, IgA and IgE, and any isotype, including without limitation IgG 1, IgG2, IgG3 and IgG4. The humanized antibody may comprise sequences from more than one class or isotype, and particular constant domains may be selected to optimize desired effector functions using techniques well- known in the art.

The framework and CDR regions of a humanized antibody need not correspond precisely to the parental sequences, e.g., the donor antibody CDR or the consensus framework may be mutagenized by substitution, insertion and/or deletion of at least about one amino acid residue so that the CDR or framework residue at that site does not correspond to either the donor antibody or the consensus framework. In a particular embodiment, such mutations will not be extensive. Usually, at least about 80%, at least about 85%, at least about 90%, and at least about 95% of the humanized antibody residues will correspond to those of the parental FR and CDR sequences. The term "consensus framework" refers to the framework region in the consensus immunoglobulin sequence. The term "consensus immunoglobulin sequence" refers to the sequence formed from the most frequently occurring amino acids (or nucleotides) in a family of related immunoglobulin sequences (See e.g., Winnaker, (1987) From Genes to Clones (Verlagsgesellschaft, Weinheim, Germany). A "consensus immunoglobulin sequence" can thus comprise a "consensus variable domain" and/or a "consensus constant domain". A "consensus variable domain" can in turn comprise one or more "consensus framework regions" and/or one or more "consensus CDRs". In a family of immunoglobulins, each position in the consensus sequence is occupied by the amino

acid occurring most frequently at that position in the family. If two amino acids occur equally frequently, either can be included in the consensus sequence.

The term "Vernier" zone refers to a subset of framework residues that may adjust CDR structure and fine-tune the fit to antigen as described by Foote and Winter (1992) J. Mol. Biol. 224:487-499). Vernier zone residues form a layer underlying the CDRs and may impact on the structure of CDRs and the affinity of the antibody.

The term "multivalent binding protein" is used in this specification to denote a binding protein comprising two or more antigen binding sites. In an embodiment, the multivalent binding protein is engineered to have the three or more antigen binding sites, and is generally not a naturally occurring antibody. The term "multispecific binding protein" refers to a binding protein capable of binding two or more related or unrelated targets. Dual variable domain (DVD) binding proteins are binding proteins that comprise two or more antigen binding sites and are tetravalent or multivalent binding proteins. Such DVD binding proteins may be monospecific, i.e capable of binding one antigen or multispecific, i.e. capable of binding two or more antigens. DVD binding proteins comprising two heavy chain DVD polypeptides and two light chain DVD polypeptides are referred to as a DVD-Ig^{TM} molecule. Each half of a DVD-Ig comprises a heavy chain DVD polypeptide, and a light chain DVD polypeptide, and two antigen binding sites. Each binding site comprises a heavy chain variable domain and a light chain variable domain with a total of 6 CDRs involved in antigen binding per antigen binding site. DVD binding proteins and methods of making DVD binding proteins are disclosed in U.S. Patent No. 7,612,181.

One aspect pertains to a DVD binding protein comprising binding proteins capable of binding TNFα. In an embodiment, the DVD binding protein is capable of binding TNF-α and a second target.

The term "neutralizing" refers to neutralization of biological activity of a cytokine when a binding protein specifically binds the cytokine. In an embodiment, a neutralizing binding protein is a neutralizing antibody whose binding to hTNF-α results in inhibition of a biological activity of hTNF-α. In an embodiment, the neutralizing binding protein binds hTNFα and reduces a biologically activity of hTNF-α by at least about 20%, at least about 40%, at least about 60%, at least about 80%, at least about 85% or more. Inhibition of a biological activity of hTNFα by a neutralizing binding protein can be assessed by measuring one or more indicators of hTNF-α biological activity well known in the art. For example neutralization of the cytoxicity of TNFα on L929 cells.

The term "activity" includes activities such as the binding specificity/affinity of an antibody for an antigen, for example, an anti-hTNF-α antibody that binds to an TNF-α antigen and/or the neutralizing potency of an antibody, for example, an anti-hTNF-α antibody whose

binding to hTNF-α inhibits the biological activity of hTNF-α, e.g. neutralization of the cytoxicity of TNFα on L929 cells.

The term "epitope" includes any polypeptide determinant capable of specific binding to an immunoglobulin or T-cell receptor. In certain embodiments, epitope determinants include chemically active surface groupings of molecules such as amino acids, sugar side chains, phosphoryl, or sulfonyl, and, in certain embodiments, may have specific three dimensional structural characteristics, and/or specific charge characteristics. An epitope is a region of an antigen that is bound by an antibody. An epitope thus consists of the amino acid residues of a region of an antigen (or fragment thereof) known to bind to the complementary site on the specific binding partner. An antigenic fragment can contain more than one epitope. In certain embodiments, an antibody is said to specifically bind an antigen when it preferentially recognizes its target antigen in a complex mixture of proteins and/or macromolecules. An epitope thus consists of the amino acid residues of a region of an antigen (or fragment thereof) known to bind to the complementary site on the specific binding partner. An antigenic fragment can contain more than one epitope.

The term "surface plasmon resonance" refers to an optical phenomenon that allows for the analysis of real-time biospecific interactions by detection of alterations in protein concentrations within a biosensor matrix, for example using the BIACORE system (Biacore International AB, a GE Healthcare company, Uppsala, Sweden and Piscataway, New Jersey). For further descriptions, see Jönsson et al. (1993) Ann. Biol. Clin. 51:19-26; Jönsson et al. (1991) Biotechniques 11:620-627; Johnsson et al. (1995) J. Mol. Recognit. 8:125-131; and Johnnson et al. (1991) Anal. Biochem. 198:268-277.

The term "Kon" refers to the on rate constant for association of a binding protein (e.g., an antibody) to the antigen to form the, e.g., antibody/antigen complex as is known in the art. The "Kon" also is known by the terms "association rate constant," or "ka," as used interchangeably herein. This value indicating the binding rate of an antibody to its target antigen or the rate of complex formation between an antibody and antigen also is shown by the equation:

Antibody ("Ab") + Antigen ("Ag")→Ab-Ag.

The term "Koff" refers to the off rate constant for dissociation of a binding protein (e.g., an antibody) from the, e.g., antibody/antigen complex as is known in the art. The "Koff" also is known by the terms "dissociation rate constant" or "kd" as used interchangeably herein. This value indicates the dissociation rate of an antibody from its target antigen or separation of Ab-Ag complex over time into free antibody and antigen as shown by the equation below:

Ab + Ag←Ab-Ag

The terms "equilibrium dissociation constant" or "KD," as used interchangeably herein, refer to the value obtained in a titration measurement at equilibrium, or by dividing the dissociation rate constant (koff) by the association rate constant (kon). The association rate constant, the dissociation rate constant, and the equilibrium dissociation constant are used to represent the binding affinity of an antibody to an antigen. Methods for determining association and dissociation rate constants are well known in the art. Using fluorescence–based techniques offers high sensitivity and the ability to examine samples in physiological buffers at equilibrium. Other experimental approaches and instruments such as a BIACORE (biomolecular interaction analysis) assay can be used (e.g., instrument available from Biacore International AB, a GE Healthcare company, Uppsala, Sweden). Additionally, a KinExA® (Kinetic Exclusion Assay) assay, available from Sapidyne Instruments (Boise, Idaho) can also be used.

The term “labeled binding protein” refers to a protein with a label d that provides for the identification of the binding protein. In an embodiment, the label is a detectable marker, e.g., incorporation of a radiolabeled amino acid or attachment to a polypeptide of biotinyl moieties that can be detected by marked avidin (e.g., streptavidin containing a fluorescent marker or enzymatic activity that can be detected by optical or colorimetric methods). Examples of labels for polypeptides include, but are not limited to, the following: radioisotopes or radionuclides (e.g., ^{3}H, ^{14}C, ^{35}S, ^{90}Y, ^{99}Tc, ^{111}In, ^{125}I, ^{131}I, ^{177}Lu, ^{166}Ho, or and ^{153}Sm); fluorescent labels (e.g., FITC, rhodamine, anddd lanthanide phosphors), enzymatic labels (e.g., horseradish peroxidase, luciferase, alkaline phosphatase); chemiluminescent markers; biotinyl groups; predetermined polypeptide epitopes recognized by a secondary reporter (e.g., leucine zipper pair sequences, binding sites for secondary antibodies, metal binding domains, and epitope tags); and magnetic agents, such as gadolinium chelates.

The term “antibody conjugate” refers to a binding protein, such as an antibody, chemically linked to a second chemical moiety, such as a therapeutic or cytotoxic agent. The term "agent" denotes a chemical compound, a mixture of chemical compounds, a biological macromolecule, or an extract made from biological materials. In an embodiment, the therapeutic or cytotoxic agents include, but are not limited to, pertussis toxin, taxol, cytochalasin B, gramicidin D, ethidium bromide, emetine, mitomycin, etoposide, tenoposide, vincristine, vinblastine, colchicine, doxorubicin, daunorubicin, dihydroxy anthracin dione, mitoxantrone, mithramycin, actinomycin D, 1-dehydrotestosterone, glucocorticoids, procaine, tetracaine, lidocaine, propranolol, and puromycin and analogs or homologs thereof.

The terms “crystal”, and “crystallized” refers to an antibody, or antigen binding portion thereof, that exists in the form of a crystal. Crystals are one form of the solid state of matter,

which is distinct from other forms such as the amorphous solid state or the liquid crystalline state. Crystals are composed of regular, repeating, three-dimensional arrays of atoms, ions, molecules (e.g., proteins such as antibodies), or molecular assemblies (e.g., antigen/antibody complexes). These three-dimensional arrays are arranged according to specific mathematical relationships that are well-understood in the field. The fundamental unit, or building block, that is repeated in a crystal is called the asymmetric unit. Repetition of the asymmetric unit in an arrangement that conforms to a given, well-defined crystallographic symmetry provides the "unit cell" of the crystal. Repetition of the unit cell by regular translations in all three dimensions provides the crystal. See Giege and Ducruix (1999) Crystallization of Nucleic Acids and Proteins, a Practical Approach, 2nd ea., pp. 20 1-16, Oxford University Press, New York, New York.)

The term "polynucleotide" means a polymeric form of two or more nucleotides, either ribonucleotides or deoxynucleotides or a modified form of either type of nucleotide. The term includes single and double stranded forms of DNA or RNA.

The term "isolated polynucleotide" means a polynucleotide (e.g., of genomic, cDNA, or synthetic origin, or a combination thereof) that is not associated with all or a portion of a polynucleotide with which it is associated in nature; with which it is operably linked to a in nature; or with which it occurs in nature as part of a larger sequence.

The term "vector" refers to a nucleic acid molecule capable of transporting another nucleic acid to which it has been linked. One type of vector is a "plasmid", which refers to a circular double stranded DNA loop into which additional DNA segments may be ligated. Another type of vector is a viral vector, wherein additional DNA segments may be ligated into the viral genome. Certain vectors are capable of autonomous replication in a host cell into which they are introduced (e.g., bacterial vectors having a bacterial origin of replication and episomal mammalian vectors). Other vectors (e.g., non-episomal mammalian vectors) can be integrated into the genome of a host cell upon introduction into the host cell, and thereby are replicated along with the host genome. Moreover, certain vectors are capable of directing the expression of genes to which they are operatively linked. Such vectors are referred to herein as "recombinant expression vectors" (or simply, "expression vectors"). In general, expression vectors of utility in recombinant DNA techniques are often in the form of plasmids. In the present specification, "plasmid" and "vector" may be used interchangeably as the plasmid is the most commonly used form of vector. However, the embodiments are intended to include such other forms of expression vectors, such as viral vectors (e.g., replication defective retroviruses, adenoviruses and adeno-associated viruses), which serve equivalent functions.

The term "operably linked" refers to a positioning of components such that they function in their intended manner. A control sequence "operably linked" to a coding sequence is ligated in

such a way that expression of the coding sequence is achieved under conditions compatible with the control sequences. "Operably linked" sequences include expression control sequences that are contiguous with a nucleic acid of interest and expression control sequences that act in trans i.e., are located on a different nucleic acid molecule than a nucleic acid of interest but nevertheless exert control over the nucleic acid of interest, and expression control sequences that are located on the same nucleic acid molecule as, but at a distance from, a nucleic acid of interest. The term "expression control sequence" refers to polynucleotide sequences that are necessary to effect the expression and processing of coding sequences to which they are ligated. Expression control sequences include appropriate transcription initiation, termination, promoter and enhancer sequences; efficient RNA processing signals such as splicing and polyadenylation signals; sequences that stabilize cytoplasmic mRNA; sequences that enhance translation efficiency (i.e., Kozak consensus sequence); sequences that enhance protein stability; and when desired, sequences that enhance protein secretion. The nature of such control sequences differs depending upon the host organism; in prokaryotes, such control sequences generally include promoter, ribosomal binding site, and transcription termination sequence; in eukaryotes, generally, such control sequences include promoters and transcription termination sequence. The term "control sequences" is intended to include components whose presence is essential for expression and processing, and can also include additional components whose presence is advantageous, for example, leader sequences and fusion partner sequences.

"Transformation", as defined herein, refers to any process by which exogenous DNA enters a host cell. Transformation may occur under natural or artificial conditions using various methods well known in the art for the insertion of foreign nucleic acid sequences into a prokaryotic or eukaryotic host cell, for example. The method is selected based on the host cell being transformed and may include, but is not limited to, viral infection, electroporation, lipofection, and particle bombardment. Such "transformed" cells include stably transformed cells in which the inserted DNA is capable of replication either as an autonomously replicating plasmid or as part of the host chromosome. They also include cells which transiently express the inserted DNA or RNA for limited periods of time.

The term "recombinant host cell" (or simply "host cell"), refers to a cell into which exogenous DNA has been introduced. It should be understood that such terms are intended to refer not only to the particular subject cell, but, to the progeny of such a cell. Because certain modifications may occur in succeeding generations due to either mutation or environmental influences, such progeny may not, in fact, be identical to the parent cell, but are still included within the scope of the term "host cell". In an embodiment, host cells include prokaryotic and eukaryotic cells chosen from any of the Kingdoms of life. In an embodiment, eukaryotic cells include protist, fungal, plant and animal cells. In an embodiment, host cells include but are not

limited to the prokaryotic cell line E.coli; mammalian cell lines CHO, HEK 293 and COS; the insect cell line Sf9; and the fungal cell Saccharomyces cerevisiae.

Standard techniques may be used for recombinant DNA, oligonucleotide synthesis, and tissue culture and transformation (e.g., electroporation and lipofection). Enzymatic reactions and purification techniques may be performed according to manufacturer's specifications or as commonly accomplished in the art or as described herein. The foregoing techniques and procedures may be generally performed according to conventional methods well known in the art and as described in various general and more specific references that are cited and discussed throughout the present specification. See e.g., Sambrook et al. Molecular Cloning: A Laboratory Manual (2d ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. (1989)).

The term “transgenic organism” refers to an organism having cells that contain a transgene, wherein the transgene introduced into the organism (or an ancestor of the organism) expresses a polypeptide not naturally expressed in the organism. A “transgene” is a DNA construct that is stably and operably integrated into the genome of a cell from which a transgenic organism develops, directing the expression of an encoded gene product in one or more cell types or tissues of the transgenic organism.

The terms "regulate"and “modulate” are used interchangeably, and refer to a change or an alteration in the activity of a molecule of interest (e.g., the biological activity of hTNF-α). Modulation may be an increase or a decrease in the magnitude of a certain activity or function of the molecule of interest. Exemplary activities and functions of a molecule include, but are not limited to, binding characteristics, enzymatic activity, cell receptor activation, and signal transduction.

Correspondingly, the term "modulator" is a compound capable of changing or altering an activity or function of a molecule of interest (e.g., the biological activity of hTNF-α). For example, a modulator may cause an increase or decrease in the magnitude of a certain activity or function of a molecule compared to the magnitude of the activity or function observed in the absence of the modulator. In certain embodiments, a modulator is an inhibitor, which decreases the magnitude of at least one activity or function of a molecule. Exemplary inhibitors include, but are not limited to, proteins, peptides, antibodies, peptibodies, carbohydrates or small organic molecules. Peptibodies are described, e.g., in WO01/83525.

The term "agonist" refers to a modulator that, when contacted with a molecule of interest, causes an increase in the magnitude of a certain activity or function of the molecule compared to the magnitude of the activity or function observed in the absence of the agonist. Particular agonists of interest may include, but are not limited to, TNF-α polypeptides or polypeptides, nucleic acids, carbohydrates, or any other molecules that bind to hTNF-α.

The term "antagonist" or "inhibitor" refers to a modulator that, when contacted with a molecule of interest causes a decrease in the magnitude of a certain activity or function of the molecule compared to the magnitude of the activity or function observed in the absence of the antagonist. Particular antagonists of interest include those that block or modulate the biological or immunological activity of hTNF-α. Antagonists and inhibitors of hTNF-α may include, but are not limited to, proteins, nucleic acids, carbohydrates, or any other molecules, which bind to hTNF-α .

The term "effective amount" refers to the amount of a therapy which is sufficient to reduce or ameliorate the severity and/or duration of a disorder or one or more symptoms thereof, prevent the advancement of a disorder, cause regression of a disorder, prevent the recurrence, development, onset or progression of one or more symptoms associated with a disorder, detect a disorder, or enhance or improve the prophylactic or therapeutic effect(s) of another therapy (e.g. , prophylactic or therapeutic agent).

The term "biological sample", includes, but is not limited to, any quantity of a substance from a living thing or formerly living thing. Such living things include, but are not limited to, humans, mice, rats, monkeys, dogs, rabbits and other animals. Such substances include, but are not limited to, blood, serum, urine, synovial fluid, cells, organs, tissues, bone marrow, lymph nodes and spleen.

I.Antibodies that Bind Human TNF-α

One aspect provides isolated murine monoclonal antibodies, or antigen-binding portions thereof, that bind to TNF with high affinity, a slow off rate and high neutralizing capacity. A second aspect provides chimeric antibodies that bindTNF-α. A third aspect provides CDR grafted antibodies, or antigen-binding portions thereof, that bind TNF-α. A fourth aspect provides humanized antibodies, or antigen-binding portions thereof, that bind TNF-α. In an embodiment, the antibodies, or portions thereof, are isolated antibodies. In an embodiment, the antibodies are neutralizing human anti-TNF-α antibodies.

A. Method of making anti TNF-α antibodies

Antibodies may be made by any of a number of techniques known in the art.

1. Anti-TNF-α Monoclonal Antibodies Using Hybridoma Technology

Monoclonal antibodies can be prepared using a wide variety of techniques known in the art including the use of hybridoma, recombinant, and phage display technologies, or a combination thereof. For example, monoclonal antibodies can be produced using hybridoma techniques including those known in the art and taught, for example, in Harlow et al. , Antibodies: A Laboratory Manual, 2nd ed., (Cold Spring Harbor Laboratory Press, Cold Spring Harbor, 1988);

Hammerling, et al. eds. , in: Monoclonal Antibodies and T-Cell Hybridomas " In Research Monographs in Immunology, vol. 3 (J.L. Turk, General Editor) (Elsevier, N.Y., 1981). The term "monoclonal antibody" is not limited to antibodies produced through hybridoma technology. The term "monoclonal antibody" refers to an antibody that is derived from a single clone, including any eukaryotic, prokaryotic, or phage clone, and not the method by which it is produced.

Methods for producing and screening for specific antibodies using hybridoma technology are routine and well known in the art. One embodiment provides methods of generating monoclonal antibodies as well as antibodies produced by the method comprising culturing a hybridoma cell secreting an antibody wherein, in an embodiment, the hybridoma is generated by fusing splenocytes isolated from a mouse immunized with an antigen with myeloma cells and then screening the hybridomas resulting from the fusion for hybridoma clones that secrete an antibody able to bind a polypeptide. Briefly, mice can be immunized with an TNF-α antigen. In an embodiment, the TNF-α antigen is administered with an adjuvant to stimulate the immune response. Such adjuvants include complete or incomplete Freund's adjuvant, RIBI (muramyl dipeptides) or ISCOM (immunostimulating complexes). Such adjuvants may protect the polypeptide from rapid dispersal by sequestering it in a local deposit, or they may contain substances that stimulate the host to secrete factors that are chemotactic for macrophages and other components of the immune system. In an embodiment, if a polypeptide is being administered, the immunization schedule will involve two or more administrations of the polypeptide, spread out over several weeks.

After immunization of an animal with an TNF-α antigen, antibodies and/or antibody-producing cells may be obtained from the animal. An anti-TNFα antibody-containing serum is obtained from the animal by bleeding or sacrificing the animal. The serum may be used as it is obtained from the animal, an immunoglobulin fraction may be obtained from the serum, or the anti- TNF-α antibodies may be purified from the serum. Serum or immunoglobulins obtained in this manner are polyclonal, thus having a heterogeneous array of properties.

Once an immune response is detected, e.g., antibodies specific for the antigen TNF-α are detected in the mouse serum, the mouse spleen is harvested and splenocytes isolated. The splenocytes are then fused by well-known techniques to any suitable myeloma cells, for example cells from cell line SP20 available from the ATCC. Hybridomas are selected and cloned by limited dilution. The hybridoma clones are then assayed by methods known in the art for cells that secrete antibodies capable of binding TNF-α. Ascites fluid, which generally contains high levels of antibodies, can be generated by immunizing mice with positive hybridoma clones.

In another embodiment, antibody-producing immortalized hybridomas may be prepared from the immunized animal. After immunization, the animal is sacrificed and the splenic B cells are fused to immortalized myeloma cells as is well known in the art. See, e.g., Harlow and Lane,

supra. In an embodiment, the myeloma cells do not secrete immunoglobulin polypeptides (a non-secretory cell line). After fusion and antibiotic selection, the hybridomas are screened using TNF-α, or a portion thereof, or a cell expressing TNF-α. In an embodiment, the initial screening is performed using an enzyme-linked immunoassay (ELISA) or a radioimmunoassay (RIA). An example of ELISA screening is provided in PCT Publication No. WO 00/37504.

Anti- TNF-α antibody-producing hybridomas are selected, cloned and further screened for desirable characteristics, including robust hybridoma growth, high antibody production and desirable antibody characteristics, as discussed further below. Hybridomas may be cultured and expanded *in vivo* in syngeneic animals, in animals that lack an immune system, e.g., nude mice, or in cell culture *in vitro.* Methods of selecting, cloning and expanding hybridomas are well known to those of ordinary skill in the art.

In an embodiment, the hybridomas are mouse hybridomas, as described above. In another embodiment, the hybridomas are produced in a non-human, non-mouse species such as rats, sheep, pigs, goats, cattle or horses. In another embodiment, the hybridomas are human hybridomas, in which a human non-secretory myeloma is fused with a human cell expressing an anti-TNF-α antibody.

Antibody fragments that recognize specific epitopes may be generated by known techniques. For example, Fab and F(ab')2 fragments may be produced by proteolytic cleavage of immunoglobulin molecules, using enzymes such as papain (to produce Fab fragments) or pepsin (to produce F(ab')2 fragments). F(ab')2 fragments contain the variable region, the light chain constant region and the CH1 domain of the heavy chain.

2. Anti-TNF-α monoclonal antibodies using SLAM

In another aspect, recombinant antibodies are generated from single, isolated lymphocytes using a procedure referred to in the art as the selected lymphocyte antibody method (SLAM), as described in U.S. Patent No. 5,627,052; PCT Publication No. WO 92/02551 and Babcook et al. (1996) Proc. Natl. Acad. Sci. USA 93:7843-7848. In this method, single cells secreting antibodies of interest, *e.g.*, lymphocytes derived from an immunized anima are screened using an antigen-specific hemolytic plaque assay, wherein the antigen TNF-α, a subunit of TNF-α, or a fragment thereof, is coupled to sheep red blood cells using a linker, such as biotin, and used to identify single cells that secrete antibodies with specificity for TNF-α. Following identification of antibody-secreting cells of interest, heavy- and light-chain variable region cDNAs are rescued from the cells by reverse transcriptase-PCR and these variable regions can then be expressed, in the context of appropriate immunoglobulin constant regions (e.g., human constant regions), in mammalian host cells, such as COS or CHO cells. The host cells transfected with the amplified immunoglobulin sequences, derived from *in vivo* selected lymphocytes, can then undergo further

analysis and selection *in vitro*, for example by panning the transfected cells to isolate cells expressing antibodies to TNF-α. The amplified immunoglobulin sequences further can be manipulated *in vitro*, such as by *in vitro* affinity maturation methods such as those described in PCT Publication Nos. WO 97/29131 and WO 00/56772.

3. Anti-TNF-α monoclonal antibodies using transgenic animals

In another embodiment, antibodies are produced by immunizing a non-human animal comprising some, or all, of the human immunoglobulin locus with an TNF-α antigen. In an embodiment, the non-human animal is a XENOMOUSE® transgenic mouse, an engineered mouse strain that comprises large fragments of the human immunoglobulin loci and is deficient in mouse antibody production. See, e.g., Green et al. (1994) Nature Genet. 7:13-21 and U.S. Patent Nos. 5,916,771; 5,939,598; 5,985,615; 5,998,209; 6,075,181; 6,091,001; 6,114,598 and 6,130,364. See also PCT Publication Nos. WO 91/10741; WO 94/02602; WO 96/34096; WO 96/33735; WO 98/16654; WO 98/24893; WO 98/50433; WO 99/45031; WO 99/53049; WO 00/09560; and WO 00/37504. The XENOMOUSE® transgenic mouse produces an adult-like human repertoire of fully human antibodies, and generates antigen-specific human monoclonal antibodies. The XENOMOUSE® transgenic mouse contains approximately 80% of the human antibody repertoire through introduction of megabase sized, germline configuration YAC fragments of the human heavy chain loci and x light chain loci. See, Mendez et al. (1997) Nature Genet. 15:146-156 and Green and Jakobovits (1998) J. Exp. Med. 188:483-495.

4. Anti-TNF-α monoclonal antibodies using recombinant antibody libraries

In vitro methods also can be used to make the antibodies, wherein an antibody library is screened to identify an antibody having the desired binding specificity. Methods for such screening of recombinant antibody libraries are well known in the art and include methods described in, for example, U.S. Patent No. 5,223,409; PCT Publication Nos. WO 92/18619, WO 91/17271, WO 92/20791, WO 92/15679, WO 93/01288, WO 92/01047, WO 92/09690; WO 97/29131; Fuchs et al. (1991) Bio/Technology 9:1369-1372; Hay et al. (1992) Hum. Antibod. Hybridomas 3:81-85; Huse et al. (1989) Science 246:1275-1281; McCafferty et al. (1990) Nature 348:552-554; Griffiths et al. (1993) EMBO J. 12:725-734; Hawkins et al. (1992) J. Mol. Biol. 226:889-896; Clackson et al. (1991) Nature 352:624-628; Gram et al. (1992) Proc. Natl. Acad. Sci. USA 89:3576-3580; Garrard et al. (1991) Bio/Technology 9:1373-1377; Hoogenboom et al. (1991) Nucl. Acids Res. 19:4133-4137; and Barbas et al. (1991) Proc. Natl. Acad. Sci. USA 88:7978-7982, and U.S. Patent Publication No. 2003/0186374..

The recombinant antibody library may be from a subject immunized with TNF-α or a portion of TNF-α. Alternatively, the recombinant antibody library may be from a naïve subject,

i.e., one who has not been immunized with TNF-α, such as a human antibody library from a human subject who has not been immunized with human TNF-α. Antibodies are selected by screening the recombinant antibody library with the peptide comprising human TNF-α to thereby select those antibodies that recognize TNF-α. Methods for conducting such screening and selection are well known in the art, such as described in the references in the preceding paragraph. To select antibodies having particular binding affinities for hTNF-α, such as those that dissociate from human TNF-α with a particular k_{off} rate constant, the art-known method of surface plasmon resonance can be used to select antibodies having the desired k_{off} rate constant. To select antibodies having a particular neutralizing activity for hTNF-α, such as those with a particular an $IC_{50,}$ standard methods known in the art for assessing the inhibition of hTNF-α activity may be used.

One aspect pertains to an isolated antibody, or an antigen-binding portion thereof, that binds human TNF-α. In an embodiment, the antibody is a neutralizing antibody. In various embodiments, the antibody is a recombinant antibody or a monoclonal antibody.

For example, the antibodies can also be generated using various phage display methods known in the art. In phage display methods, functional antibody domains are displayed on the surface of phage particles which carry the polynucleotide sequences encoding them. In a particular, such phage can be utilized to display antigen-binding domains expressed from a repertoire or combinatorial antibody library (e.g., human or murine). Phage expressing an antigen binding domain that binds the antigen of interest can be selected or identified with antigen, e.g., using labeled antigen or antigen bound or captured to a solid surface or bead. Phage used in these methods are typically filamentous phage including fd and M13 binding domains expressed from phage with Fab, Fv or disulfide stabilized Fv antibody domains recombinantly fused to either the phage gene III or gene VIII protein. Examples of phage display methods that can be used to make the antibodies include those disclosed in Brinkmann et al. (1995) J. Immunol. Methods 182:41-50; Ames et al. (1995) J. Immunol. Methods 184:177-186; Kettleborough et al. (1994) Eur. J. Immunol. 24:952-958; Persic et al. (1997) Gene 187 9-18; Burton et al. (1994) Adv. Immunol. 57:191-280; PCT Application No. PCT/GB91/01134; PCT Publication Nos. WO 90/02809; WO 91/10737; WO 92/01047; WO 92/18619; WO 93/11236; WO 95/15982; WO 95/20401; and U.S. Patent Nos. 5,698,426; 5,223,409; 5,403,484; 5,580,717; 5,427,908; 5,750,753; 5,821,047; 5,571,698; 5,427,908; 5,516,637; 5,780,225; 5,658,727; 5,733,743 and 5,969,108.

After phage selection, the antibody coding regions from the phage can be isolated and used to generate whole antibodies including human antibodies or any other desired antigen binding fragment, and expressed in any desired host, including mammalian cells, insect cells, plant cells, yeast, and bacteria, e.g., as described in detail herein. For example, techniques to recombinantly produce Fab, Fab' and F(ab')2 fragments can also be employed using methods

known in the art such as those disclosed in PCT Publication No. WO 92/22324; Mullinax et al. (1992) BioTechniques 12(6):864-869; Sawai et al. (1995) Am. J. Reprod. Immunol. 34:26-34; and Better et al. (1988) Science 240:1041-1043. Examples of techniques that can be used to produce single-chain Fvs and antibodies include those described in U.S. Patent Nos. 4,946,778 and 5,258,498; Huston et al. (1991) Methods Enzymol. 203:46-88; Shu et al. (1993) Proc. Natl. Acad. Sci. USA 90:7995-7999; and Skerra et al. (1988) Science 240:1038-1041.

Alternative to screening of recombinant antibody libraries by phage display, other methodologies known in the art for screening large combinatorial libraries can be applied to the identification of dual specificity antibodies. One type of alternative expression system is one in which the recombinant antibody library is expressed as RNA-protein fusions, as described in PCT Publication No. WO 98/31700 and in Roberts and Szostak (1997) Proc. Natl. Acad. Sci. USA 94:12297-12302. In this system, a covalent fusion is created between an mRNA and the peptide or protein that it encodes by *in vitro* translation of synthetic mRNAs that carry puromycin, a peptidyl acceptor antibiotic, at their 3' end. Thus, a specific mRNA can be enriched from a complex mixture of mRNAs (*e.g.*, a combinatorial library) based on the properties of the encoded peptide or protein, *e.g.*, antibody, or portion thereof, such as binding of the antibody, or portion thereof, to the dual specificity antigen. Nucleic acid sequences encoding antibodies, or portions thereof, recovered from screening of such libraries can be expressed by recombinant means as described above (*e.g.*, in mammalian host cells) and, moreover, can be subjected to further affinity maturation by either additional rounds of screening of mRNA-peptide fusions in which mutations have been introduced into the originally selected sequence(s), or by other methods for affinity maturation *in vitro* of recombinant antibodies, as described above.

In another approach the antibodies can also be generated using yeast display methods known in the art. In yeast display methods, genetic methods are used to tether antibody domains to the yeast cell wall and display them on the surface of yeast. In particular, such yeast can be utilized to display antigen-binding domains expressed from a repertoire or combinatorial antibody library (e. g., human or murine). Examples of yeast display methods that can be used to make the antibodies include those disclosed U.S. Patent No. 6,699,658.

B. Production of recombinant TNF-α antibodies

Antibodies may be produced by any of a number of techniques known in the art. For example, expression from host cells, wherein expression vector(s) encoding the heavy and light chains is (are) transfected into a host cell by standard techniques. The various forms of the term "transfection" are intended to encompass a wide variety of techniques commonly used for the introduction of exogenous DNA into a prokaryotic or eukaryotic host cell, *e.g.*, electroporation, calcium-phosphate precipitation, DEAE-dextran transfection and the like. Although it is possible

to express the antibodies in either prokaryotic or eukaryotic host cells, in an embodiment, mammalian host cells, as such eukaryotic cells (and in particular mammalian cells) are more likely than prokaryotic cells to assemble and secrete a properly folded and immunologically active antibody.

Mammalian host cells for expressing the recombinant antibodies include Chinese Hamster Ovary (CHO cells) (including dhfr- CHO cells, described in Urlaub and Chasin (1980) Proc. Natl. Acad. Sci. USA 77:4216-4220, used with a DHFR selectable marker, e.g., as described in Kaufman and Sharp (1982) J. Mol. Biol. 159:601-621), NS0 myeloma cells, COS cells and SP2 cells. When recombinant expression vectors encoding antibody genes are introduced into mammalian host cells, the antibodies are produced by culturing the host cells for a period of time sufficient to allow for expression of the antibody in the host cells or secretion of the antibody into the culture medium in which the host cells are grown. Antibodies can be recovered from the culture medium using standard protein purification methods.

Host cells can also be used to produce functional antibody fragments, such as Fab fragments or scFv molecules. It will be understood that variations on the above procedure are within the scope of the embodiments. For example, it may be desirable to transfect a host cell with DNA encoding functional fragments of either the light chain and/or the heavy chain of an antibody. Recombinant DNA technology may also be used to remove some, or all, of the DNA encoding either or both of the light and heavy chains that is not necessary for binding to the antigens of interest. The molecules expressed from such truncated DNA molecules are also encompassed by the antibodies. In addition, bifunctional antibodies may be produced in which one heavy and one light chain are an antibody and the other heavy and light chain are specific for an antigen other than the antigens of interest by crosslinking an antibody to a second antibody by standard chemical crosslinking methods.

In an exemplary system for recombinant expression of an antibody, or antigen-binding portion thereof, a recombinant expression vector encoding both the antibody heavy chain and the antibody light chain is introduced into dhfr- CHO cells by calcium phosphate-mediated transfection. Within the recombinant expression vector, the antibody heavy and light chain genes are each operatively linked to CMV enhancer/AdMLP promoter regulatory elements to drive high levels of transcription of the genes. The recombinant expression vector also carries a DHFR gene, which allows for selection of CHO cells that have been transfected with the vector using methotrexate selection/amplification. The selected transformant host cells are cultured to allow for expression of the antibody heavy and light chains and intact antibody is recovered from the culture medium. Standard molecular biology techniques are used to prepare the recombinant expression vector, transfect the host cells, select for transformants, culture the host cells and recover the antibody from the culture medium. Still further is provided a method of synthesizing a

recombinant antibody by culturing a host cell in a suitable culture medium until a recombinant antibody is synthesized. The method can further comprise isolating the recombinant antibody from the culture medium.

1. Anti hTNF-α Antibodies

Table 5 is a list of amino acid sequences of VH and VL regions of murine anti-hTNF-α antibodies.

Table 5: List of Amino Acid Sequences of Murine anti-hTNF-α antibody VH and VL Regions

SEQ ID No.	Protein region		Sequence
			123456789**0**123456789**0**1234567890
31	**VH MAK199**		QIQLVQSGPELKKPGETVMISCKASGYTFT**NYGMN**WVKQAPGKGLKWMG**WINTYTGEPTY****ADDFKG**RFAFSLETSASTAYLQINNLKNED TATYFCAR**KFLTTVVVTDYAMDY**WGQGTSV TVSS
	VH MAK199 CDR-H1	Residues 31-35 of SEQ ID NO.:31	**NYGMN**
	VH MAK199 CDR-H2	Residues 50-66 of SEQ ID NO.:31	**WINTYTGEPTYADDFKG**
	VH MAK199 CDR-H3	Residues 99-113 of SEQ ID NO.:31	**KFLTTVVVTDYAMDY**
32	**VL MAK199**		DIQMTQTTSSLSASLGDRVTISC**RASQDIS****NYLN**WYQQKPDGTVKLLIY**YTSRLQS**GVPS RFSGSGSGTDYSLTISNLEQEDIATYFC**QQ****GNTLPPT**FGVGTKLELK
	VL MAK199 CDR-L1	Residues 24-34 of SEQ ID NO.:32	**RASQDISNYLN**
	VL MAK199 CDR-L2	Residues 50-56 of SEQ ID NO.:32	**YTSRLQS**
	VL MAK199 CDR-L3	Residues 89-97 of SEQ ID NO.:32	**QQGNTLPPT**

2. Anti hTNF-α Chimeric Antibodies

A chimeric antibody is a molecule in which different portions of the antibody are derived from different animal species, such as antibodies having a variable region derived from a murine monoclonal antibody and a human immunoglobulin constant region. Methods for producing chimeric antibodies are known in the art and are discussed in detail in the Examples. See, e.g., Morrison (1985) Science 229:1202; Oi et al. (1986) BioTechniques 4:214; Gillies et al. (1989) J. Immunol. Methods 125:191-202; and U.S. Patent Nos. 5,807,715; 4,816,567; and 4,816,397. In

addition, techniques developed for the production of "chimeric antibodies" by splicing genes from a mouse antibody molecule of appropriate antigen specificity together with genes from a human antibody molecule of appropriate biological activity can be used (Morrison et al. (1984) Proc. Natl. Acad. Sci. 81:851-855; Neuberger et al. (1984) Nature 312:604-608; Takeda et al. (1985) Nature 314:452-454).

In one embodiment, the chimeric antibodies are produced by replacing the heavy chain constant region of the murine monoclonal anti human TNF-α antibodies described in section 1 with a human IgG1 constant region.

3. Anti TNF-α CDR Grafted Antibodies

CDR-grafted antibodies comprise heavy and light chain variable region sequences from a human antibody wherein one or more of the CDR regions of VH and/or VL are replaced with CDR sequences of the murine antibodies. A framework sequence from any human antibody may serve as the template for CDR grafting. However, straight chain replacement onto such a framework often leads to some loss of binding affinity to the antigen. The more homologous a human antibody is to the original murine antibody, the less likely the possibility that combining the murine CDRs with the human framework will introduce distortions in the CDRs that could reduce affinity. Therefore, in an embodiment, the human variable framework that is chosen to replace the murine variable framework apart from the CDRs have at least about 65%, at least about 70%, at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 95%, about 100%, sequence identity with the murine antibody variable region framework. Methods for producing CDR-grafted antibodies are known in the art and described in detail along with humanization of such CDR-grafted antibodies in the Examples (see also, EP Patent No. EP 0 239 400; PCT Publication No. WO 91/09967; U.S. Patent Nos. 5,225,539; 5,530,101; and 5,585,089); veneering or resurfacing (EP Patent Nos. EP 0 592 106 and EP 0 519 596; Padlan (1991) Mol. Immunol. 28(4/5):489-498; Studnicka et al. (1994) Protein Eng. 7(6):805-814; Roguska et al. (1994) Proc. Natl. Acad. Sci. USA 91:969-973), and chain shuffling (U.S. Patent No. 5,565,352).

A specific embodiment provides CDR grafted antibodies with V_H and/or V_L chains as described in Table 6.

Table 6: CDR Grafted Antibodies

SEQ ID No.	Protein region	Sequence
		123456789**0**123456789**0**1234567890

SEQ ID No.	Protein region	Sequence
		12345678901234567890123456789**0**
33	hMAK199VH.1z	QVQLVQSGSELKKPGASVKVSCKASGYTFT **NYGMN**WVRQAPGQGLEWMG**WINTYTGEPTY** **ADDFKG**RFVFSLDTSVSTAYLQISSLKAED TAVYYCAR**KFLTTVVVTDYAMDY**WGQGTTV TVSS
34	hMAk199VH.2z	QVQLVQSGAEVKKPGASVKVSCKASGYTFT **NYGMN**WVRQAPGQGLEWMG**WINTYTGEPTY** **ADDFKG**RVTMTTDTSTSTAYMELRSLRSDD TAVYYCAR**KFLTTVVVTDYAMDY**WGQGTTV TVSS
35	hMAK199VL.1	DIQMTQSPSSLSASVGDRVTITC**RASQDIS** **NYLN**WYQQKPGKAPKLLIY**YTSRLQS**GVPS RFSGSGSGTDFTLTISSLQPEDFATYYC**QQ** **GNTLPPT**FGQGTKLEIK
36	hMAK199VL.2	DVVMTQSPAFLSVTPGEKVTITC**RASQDIS** **NYLN**WYQQKPDQAPKLLIK**YTSRLQS**GVPS RFSGSGSGTDFTFTISSLEAEDAATYYC**QQ** **GNTLPPT**FGQGTKLEIK

4. Anti-hTNF-α Humanized Antibodies

Humanized antibodies are antibody molecules from that have one or more complementarity determining regions (CDRs) from the non-human species and framework regions from a human immunoglobulin molecule. Known human Ig sequences are disclosed, e.g., www.ncbi.nlm.nih.gov/entrez- /query.fcgi; www.atcc.org/phage/hdb.html; www.sciquest.com/; www.abcam.com/; www.antibodyresource.com/onlinecomp.html; www.public.iastate.edu/.about.pedro/research_tools.html; www.mgen.uni-heidelberg.de/SD/IT/IT.html; www.whfreeman.com/immunology/CH- 05/kuby05.htm; www.library.thinkquest.org/12429/Immune/Antibody.html; www.hhmi.org/grants/lectures/1996/vlab/; www.path.cam.ac.uk/.about.mrc7/m- ikeimages.html; www.antibodyresource.com/; mcb.harvard.edu/BioLinks/Immuno-logy.html.www.immunologylink.com/; pathbox.wustl.edu/.about.hcenter/index.- html; www.biotech.ufl.edu/.about.hcl/; www.pebio.com/pa/340913/340913.html- ; www.nal.usda.gov/awic/pubs/antibody/; www.m.ehime-u.acjp/.about.yasuhito- /Elisa.html; www.biodesign.com/table.asp; www.icnet.uk/axp/facs/davies/lin- ks.html; www.biotech.ufl.edu/.about.fccl/protocol.html; www.isac-net.org/sites_geo.html; aximtl.imt.uni-marburg.de/.about.rek/AEP- Start.html; baserv.uci.kun.nl/.about.jraats/linksl.html; www.recab.uni-hd.de/immuno.bme.nwu.edu/; www.mrc-cpe.cam.ac.uk/imt-doc/pu-blic/INTRO.html; www.ibt.unam.mx/vir/V_mice.html; imgt.cnusc.fr:8104/; www.biochem.ucl.ac.uk/.about.martin/abs/index.html; antibody.bath.ac.uk/; abgen.cvm.tamu.edu/lab/wwwabgen.html; www.unizh.ch/.about.honegger/AHOsem-inar/Slide01.html; www.cryst.bbk.ac.uk/.about.ubcg07s/;

www.nimr.mrc.ac.uk/CC/ccaewg/ccaewg.htm; www.path.cam.ac.uk/.about.mrc7/humanisation/TAHHP.html; www.ibt.unam.mx/vir/structure/stat_aim.html; www.biosci.missouri.edu/smithgp/index.html; www.cryst.bioc.cam.ac.uk/.abo- ut.fmolina/Web-pages/Pept/spottech.html; www.jerini.de/fr roducts.htm; www.patents.ibm.com/ibm.html.Kabat et al., Sequences of Proteins of Immunological Interest, U.S. Dept. Health (1983). Such imported sequences can be used to reduce immunogenicity or reduce, enhance or modify binding, affinity, on-rate, off-rate, avidity, specificity, half-life, or any other suitable characteristic, as known in the art.

Framework residues in the human framework regions may be substituted with the corresponding residue from the CDR donor antibody to alter, improve, antigen binding. These framework substitutions are identified by methods well known in the art, e.g., by modeling of the interactions of the CDR and framework residues to identify framework residues important for antigen binding and sequence comparison to identify unusual framework residues at particular positions. (See, e.g., U.S. Patent No. 5,585,089; Riechmann et al. (1988) Nature 332:323-327 Three-dimensional immunoglobulin models are commonly available and are familiar to those skilled in the art. Computer programs are available that illustrate and display probable three-dimensional conformational structures of selected candidate immunoglobulin sequences. Inspection of these displays permits analysis of the likely role of the residues in the functioning of the candidate immunoglobulin sequence, i.e., the analysis of residues that influence the ability of the candidate immunoglobulin to bind its antigen. In this way, FR residues can be selected and combined from the consensus and import sequences so that the desired antibody characteristic, such as increased affinity for the target antigen(s), is achieved. In general, the CDR residues are directly and most substantially involved in influencing antigen binding. Antibodies can be humanized using a variety of techniques known in the art, such as but not limited to those described in Jones et al. (1986) Nature 321:522-525; Verhoeyen et al. (1988) Science 239:1534-1536), Sims et al. (1993) J. Immunol. 151: 2296-2308; Chothia and Lesk (1987) J. Mol. Biol. 196:901-917; Carter et al. (1992) Proc. Natl. Acad. Sci. USA 89:4285-4289; Presta et al. (1993) J. Immunol. 151:2623-2632; Padlan (1991) Mol. Immunol. 28(4/5):489-498; Studnicka et al. (1994) Protein Eng. 7(6):805-814; Roguska. et al. (1994) Proc. Natl. Acad. Sci. USA 91:969-973; PCT Publication Nos. WO 91/09967, WO 99/06834 (PCT/US98/16280), WO 97/20032 (PCT/US96/18978), WO 92/11272 (PCT/US91/09630), WO 92/03461 (PCT/US91/05939), WO 94/18219 (PCT/US94/01234), WO 92/01047 (PCT/GB91/01134), WO 93/06213 (PCT/GB92/01755), WO 90/14443, WO 90/14424, and WO 90/14430; European Publication Nos. EP 0592106, EP 0519596, and EP 0239400; US Patent Nos. 5,565,332; 5,723,323; 5,976,862; 5,824,514; 5,817,483; 5,814,476; 5,763,192; 5,723,323; 5,766,886; 5,714,352; 6,204,023; 6,180,370; 5,693,762; 5,530,101; 5,585,089; 5,225,539; and 4,816,567.

C. Production of Antibodies and Antibody-Producing Cell Lines

In an embodiment, anti-TNF-α antibodies exhibit a high capacity to reduce or to neutralize TNF-α activity, e.g.,as assessed by any one of several in vitro and in vivo assays known in the art. In another embodiment, anti- TNF-α antibodies also exhibit a high capacity to reduce or to neutralize TNF-α activity

In particular embodiments, the isolated antibody, or antigen-binding portion thereof, binds human TNF-α, wherein the antibody, or antigen-binding portion thereof, dissociates from human TNF-α with a k_{off} rate constant of about $0.1s^{-1}$ or less, as determined by surface plasmon resonance, or which inhibits human TNF-α activity with an IC_{50} of about 1×10^{-6}M or less. Alternatively, the antibody, or an antigen-binding portion thereof, may dissociate from human TNF-α with a k_{off} rate constant of about $1 \times 10^{-2}s^{-1}$or less, as determined by surface plasmon resonance, or may inhibit human TNF-α activity with an IC_{50} of about 1×10^{-7}M or less. Alternatively, the antibody, or an antigen-binding portion thereof, may dissociate from human TNF-α with a k_{off} rate constant of about $1 \times 10^{-3}s^{-1}$ or less, as determined by surface plasmon resonance, or may inhibit human TNF-α with an IC_{50} of about 1×10^{-8}M or less. Alternatively, the antibody, or an antigen-binding portion thereof, may dissociate from human TNF-α with a k_{off} rate constant of about $1 \times 10^{-4}s^{-1}$ or less, as determined by surface plasmon resonance, or may inhibit human TNF-α activity with an IC_{50} of about 1×10^{-9}M or less. Alternatively, the antibody, or an antigen-binding portion thereof, may dissociate from human TNF-α with a k_{off} rate constant of about $1 \times 10^{-5}s^{-1}$ or less, as determined by surface plasmon resonance, or may inhibit human TNF-α activity with an IC_{50} of about 1×10^{-10}M or less. Alternatively, the antibody, or an antigen-binding portion thereof, may dissociate from human TNF-α with a k_{off} rate constant of about $1 \times 10^{-5}s^{-1}$or less, as determined by surface plasmon resonance, or may inhibit human TNF-α activity with an IC_{50} of about 1×10^{-11}M or less.

In certain embodiments, the antibody comprises a heavy chain constant region, such as an IgG1, IgG2, IgG3, IgG4, IgA, IgE, IgM or IgD constant region. In an embodiment, the heavy chain constant region is an IgG1 heavy chain constant region or an IgG4 heavy chain constant region. Furthermore, the antibody can comprise a light chain constant region, either a kappa light chain constant region or a lambda light chain constant region. In another embodiment, the antibody comprises a kappa light chain constant region. Alternatively, the antigen binding portion can be, for example, a Fab fragment or a single chain Fv fragment.

Replacements of amino acid residues in the Fc portion to alter antibody effector function are known in the art (U.S. Patent Nos. 5,648,260 and 5,624,821). The Fc portion of an antibody mediates several important effector functions, e.g., cytokine induction, antibody dependent cell-

mediated cytotoxicity (ADCC), phagocytosis, complement dependent cytotoxicity (CDC) and half-life/ clearance rate of antibody and antigen-antibody complexes. In some cases these effector functions are desirable for therapeutic antibody but in other cases might be unnecessary or even deleterious, depending on the therapeutic objectives. Certain human IgG isotypes, particularly IgG1 and IgG3, mediate ADCC and CDC via binding to FcγRs and complement C1q, respectively. Neonatal Fc receptors (FcRn) are the critical components determining the circulating half-life of antibodies. In still another embodiment at least one amino acid residue is replaced in the constant region of the antibody, for example the Fc region of the antibody, such that effector functions of the antibody are altered.

One embodiment provides a labeled binding protein wherein an antibody, or antigen binding portion thereof is derivatized or linked to another functional molecule (e.g., another peptide or protein). For example, a labeled binding protein can be derived by functionally linking an antibody, or antigen binding portion thereof (by chemical coupling, genetic fusion, noncovalent association or otherwise) to one or more other molecular entities, such as another antibody (e.g., a bispecific antibody or a diabody), a detectable agent, a cytotoxic agent, a pharmaceutical agent, and/or a protein or peptide that can mediate association of the antibody, or antigen binding portion thereof, with another molecule (such as a streptavidin core region or a polyhistidine tag).

Useful detectable agents with which an antibody, or antigen binding portion may be derivatized include fluorescent compounds. Exemplary fluorescent detectable agents include fluorescein, fluorescein isothiocyanate, rhodamine, 5-dimethylamine-1-napthalenesulfonyl chloride, phycoerythrin and the like. An antibody may also be derivatized with detectable enzymes, such as alkaline phosphatase, horseradish peroxidase, glucose oxidase and the like. When an antibody is derivatized with a detectable enzyme, it is detected by adding additional reagents that the enzyme uses to produce a detectable reaction product. For example, when the detectable agent horseradish peroxidase is present, the addition of hydrogen peroxide and diaminobenzidine leads to a colored reaction product, which is detectable. An antibody may also be derivatized with biotin, and detected through indirect measurement of avidin or streptavidin binding.

Another provides a crystallized binding protein. Another embodiment relates to crystals of whole anti- TNF-α antibodies and fragments thereof as disclosed herein, and formulations and compositions comprising such crystals. In one embodiment the crystallized binding protein has a greater half-life in vivo than the soluble counterpart of the binding protein. In another embodiment the binding protein retains biological activity after crystallization.

Crystallized binding protein may be produced according methods known in the art and as disclosed in PCT Publication No. WO 02/72636.

Another embodiment provides a glycosylated binding protein wherein an antibody, or antigen-binding portion thereof, comprises one or more carbohydrate residues. Nascent *in vivo* protein production may undergo further processing, known as post-translational modification. In particular, sugar (glycosyl) residues may be added enzymatically, a process known as glycosylation. The resulting proteins bearing covalently linked oligosaccharide side chains are known as glycosylated proteins or glycoproteins. Protein glycosylation depends on the amino acid sequence of the protein of interest, as well as the host cell in which the protein is expressed. Different organisms may produce different glycosylation enzymes (e.g., glycosyltransferases and glycosidases), and have different substrates (e.g., nucleotide sugars) available. Due to such factors, protein glycosylation pattern, and composition of glycosyl residues, may differ depending on the host system in which the particular protein is expressed. Useful glycosyl residues include, but are not limited to, glucose, galactose, mannose, fucose, n-acetylglucosamine and sialic acid. In an embodiment the glycosylated binding protein comprises glycosyl residues such that the glycosylation pattern is human.

It is known to those skilled in the art that differing protein glycosylation may result in differing protein characteristics. For instance, the efficacy of a therapeutic protein produced in a microorganism host, such as yeast, and glycosylated utilizing the yeast endogenous pathway may be reduced compared to that of the same protein expressed in a mammalian cell, such as a CHO cell line. Such glycoproteins may also be immunogenic in humans and show reduced half-life *in vivo* after administration. Specific receptors in humans and other animals may recognize specific glycosyl residues and promote the rapid clearance of the protein from the bloodstream. Other adverse effects may include changes in protein folding, solubility, susceptibility to proteases, trafficking, transport, compartmentalization, secretion, recognition by other proteins or factors, antigenicity, or allergenicity. Accordingly, a practitioner may prefer a therapeutic protein with a specific composition and pattern of glycosylation, for example glycosylation composition and pattern identical, or at least similar, to that produced in human cells or in the species-specific cells of the intended subject animal.

Expressing glycosylated proteins different from that of a host cell may be achieved by genetically modifying the host cell to express heterologous glycosylation enzymes. Using techniques known in the art a practitioner may generate antibodies or antigen-binding portions thereof exhibiting human protein glycosylation. For example, yeast strains have been genetically modified to express non-naturally occurring glycosylation enzymes such that glycosylated proteins (glycoproteins) produced in these yeast strains exhibit protein glycosylation identical to that of animal cells, especially human cells (U.S. Patent Nos. 7,449,308 and 7,029,872).

Further, it will be appreciated by one skilled in the art that a protein of interest may be expressed using a library of host cells genetically engineered to express various glycosylation

enzymes, such that member host cells of the library produce the protein of interest with variant glycosylation patterns. A practitioner may then select and isolate the protein of interest with particular novel glycosylation patterns. In an embodiment, the protein having a particularly selected novel glycosylation pattern exhibits improved or altered biological properties.

D. Uses of Anti-TNF-α Antibodies

Given their ability to bind to human TNF-α, the binding proteins, e.g., anti-human TNF-α antibodies, or antigen binding portions thereof, can be used to detect TNF-α (*e.g.*, in a biological sample, such as whole blood, serum, plasma, urine, saliva, or tissue sample), using any of the vast array of antibody-based immunodetection systems available in the art. Such immunodetection systems include, but are not limited to, immunoprecipitation, immunblotting (Western blot), enzyme-linked immunsorbent assay (ELISA), radioimmunoassay (RIA), tissue immunohistochemistry, surface plasmon resonance (SPR), sandwich immunoassay, antibody-based affinity methods (e.g., affinity beads, affinity columns), immunocompetition assay, immunochip assay (binding protein attached to a silicon chip), and fluorescence activated cell sorting (FACS). For some immunodetection systems, an TNF-α binding protein (or binding portion thereof) (or portion thereof) is attached to a solid substrate using methods available in the art for attaching antibody molecules to the same solid substrate so that the attached binding protein retains its ability to bind human TNF-α during use in the particular immunodetection system. Such solid substrates include, but are not limited to, a cellulose-based filter paper (e.g., cellulose, nitrocellulose, cellulose acetate), a nylon filter, a plastic surface (e.g., microtiter plate, antibody dip stick), a glass substrate (e.g., filters, beads, slides, glass wool), a polymeric particle (e.g., agarose, polyacrylamide), and a silicon chip. For example, an immunodetection system may be used in a method for detecting the presence of TNF-α in a sample in vitro (e.g., a biological sample, such as whole blood, serum, plasma, tissue, urine, saliva, tissue biopsy). Such a method can be used to diagnose a disease or disorder, e.g., an immune cell-associated disorder. The method includes: (i) contacting a test sample or a control sample with an TNF-α binding protein, or TNF-α binding portion thereof, as described herein; and (ii) detecting formation of a complex between the anti- TNF-α binding protein (or binding portion thereof) and TNF-α in the test sample or in the control sample, wherein a statistically significant change in the formation of the complex in the test sample relative to the control sample (or relative to formation of the complex in another test sample taken at an earlier time point) is indicative of the presence of TNF-α in the sample.

As another example, a method may be employed for detecting the presence of human TNF-α in vivo (e.g., in vivo imaging in a subject). The method can be used to diagnose a disease or disorder, e.g., a TNF-α -associated disorder. The method includes: (i) administering an TNF-α

binding protein, or TNF-α binding portion thereof, as described herein to a test subject or a control subject under conditions that allow binding of the binding protein, or TNF-α binding portion thereof, to TNF-α; and (ii) detecting formation of a complex between the binding protein, or binding portion thereof, and TNF-α, wherein a statistically significant change in the formation of the complex in the test subject relative to the control subject, or relative to formation of the complex in the test subject at an earlier time point, is indicative of the presence of TNF-α.

Methods for detecting TNF-α in a sample (e.g., a biological sample) comprise contacting a sample with an TNF-α binding protein (or TNF-α binding portion thereof) described herein and detecting either the binding protein (or binding portion thereof) bound to TNF-α or unbound binding protein (or unbound binding portion thereof) to thereby detect TNF-α in the sample. The binding protein (or portion thereof) is directly or indirectly labeled with a detectable substance to facilitate detection of the bound or unbound binding protein (or portion thereof). Such detectable substances are known in the art and, by way of non-limiting example, include various enzymes, prosthetic groups, fluorescent materials, luminescent materials, and radioactive materials. Examples of suitable enzymes include horseradish peroxidase, alkaline phosphatase, b-galactosidase, or acetylcholinesterase. Examples of suitable prosthetic group complexes include streptavidin/biotin and avidin/biotin. Examples of suitable fluorescent materials include umbelliferone, fluorescein, fluorescein isothiocyanate, rhodamine, dichlorotriazinylamine fluorescein, dansyl chloride, or phycoerythrin. An example of a luminescent material includes luminol. Examples of suitable radioactive materials include the radioisotopes ^{3}H, ^{14}C, ^{35}S, ^{90}Y, ^{99}Tc, ^{111}In, ^{125}I, ^{131}I, ^{177}Lu, ^{166}Ho, and ^{153}Sm.

Alternatively to labeling the binding protein, human TNF-α can be assayed in a sample, e.g., a biological fluid, by a competition immunoassay utilizing recombinant human (rh) TNF-α standards labeled with a detectable substance and an unlabeled TNF-α binding protein. In this assay, the sample, the labeled rh TNF-α standards, and the TNF-α binding protein are combined and the amount of labeled rh TNF-α standard bound to the unlabeled binding protein is determined. The amount of human TNF-α in the sample is inversely proportional to the amount of labeled rhTNF-α standard bound to the anti-TNF-α binding protein. Similarly, human TNF-α can also be assayed in a sample by a competition immunoassay utilizing rhTNF-α standards labeled with a detectable substance and an unlabeled anti-TNF-α binding protein.

In an embodiment, the antibodies and antigen binding portions are capable of neutralizing human TNF-α activity both *in vitro* and *in vivo*. Accordingly, such binding proteins can be used to inhibit TNF-α activity, *e.g.,* in a cell culture containing h TNF-α , in human subjects or in other mammalian subjects having TNF-α with which an antibody cross-reacts. One embodiment provides a method for inhibiting hTNF-α activity comprising contacting hTNF-α with an

antibody or antigen binding portion such that hTNF-α activity is inhibited. For example, in a cell culture containing, or suspected of containing hTNF-α , an antibody or antigen binding portion can be added to the culture medium to inhibit hTNF-α activity in the culture.

Another embodiment provides a method for reducing hTNF-α activity in a subject, advantageously from a subject suffering from a disease or disorder in which TNF-α activity is detrimental. Methods for reducing TNF-α activity in a subject suffering from such a disease or disorder are provided, which method comprises administering to the subject an antibody or antigen binding portion such that TNF-α activity in the subject is reduced. In another embodiment, the TNF-α is human TNF-α, and the subject is a human subject. Alternatively, the subject can be a mammal expressing an TNF-α to which an antibody is capable of binding. Still further the subject can be a mammal into which TNF-α has been introduced (e.g., by administration of TNF-α or by expression of an TNF-α transgene). A binding protein can be administered to a human subject for therapeutic purposes. Moreover, an binding protein can be administered to a non-human mammal expressing an TNF-α with which the binding protein is capable of binding for veterinary purposes or as an animal model of human disease. Regarding the latter, such animal models may be useful for evaluating the therapeutic efficacy of antibodies (e.g., testing of dosages and time courses of administration).

The term "a disorder in which TNF-α activity is detrimental" is intended to include diseases and other disorders in which the presence of TNF-α in a subject suffering from the disorder has been shown to be or is suspected of being either responsible for the pathophysiology of the disorder or a factor that contributes to a worsening of the disorder. Accordingly, a disorder in which TNF-α activity is detrimental is a disorder in which reduction of TNF-α activity is expected to alleviate the symptoms and/or progression of the disorder. Such disorders may be evidenced, for example, by an increase in the concentration of TNF-α in a biological fluid of a subject suffering from the disorder (e.g., an increase in the concentration of TNF-α in serum, plasma, synovial fluid, etc. of the subject), which can be detected, for example, using an anti-TNF-α binding protein as described herein. Non-limiting examples of disorders that can be treated with the antibodies include those disorders discussed in the section below pertaining to pharmaceutical compositions of the binding proteins.

D. Pharmaceutical Composition

Pharmaceutical compositions comprising a binding protein are provided, e.g., an antibody, or antigen-binding portion thereof,and a pharmaceutically acceptable carrier. The pharmaceutical compositions comprising antibodies are for use in, but not limited to, diagnosing, detecting, monitoring, preventing, inhibiting, treating, managing, or ameliorating a disorder or

one or more symptoms thereof, and/or in research. In a specific embodiment, a composition comprises one or more binding proteins. In another embodiment, the pharmaceutical composition comprises one or more binding proteins and one or more prophylactic or therapeutic agents other than antibodies for treating a disorder in which TNF-α activity is detrimental. In yet another embodiment, the prophylactic or therapeutic agents known to be useful for or having been or currently being used in the prevention, treatment, management, or amelioration of a disorder or one or more symptoms thereof. In accordance with these embodiments, the composition may further comprise of a carrier, diluent or excipient.

The binding proteins can be incorporated into pharmaceutical compositions suitable for administration to a subject. Typically, the pharmaceutical composition comprises a binding protein, e.g., an antibody or antigen binding portion thereof, and a pharmaceutically acceptable carrier. The term,"pharmaceutically acceptable carrier" includes any and all solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents, and the like that are physiologically compatible. Examples of pharmaceutically acceptable carriers include one or more of water, saline, phosphate buffered saline, dextrose, glycerol, ethanol and the like, as well as combinations thereof. It may be preferable to include isotonic agents, for example, sugars, polyalcohols such as mannitol, sorbitol, or sodium chloride in the composition. Pharmaceutically acceptable carriers may further comprise minor amounts of auxiliary substances such as wetting or emulsifying agents, preservatives or buffers, which enhance the shelf life or effectiveness of the antibody or antigen binding portion thereof.

Various delivery systems are known and can be used to administer one or more binding proteins or the combination of one or more antibodies and a prophylactic agent or therapeutic agent useful for preventing, managing, treating, or ameliorating a disorder or one or more symptoms thereof, e.g., encapsulation in liposomes, microparticles, microcapsules, recombinant cells capable of expressing the antibody or antibody fragment, receptor- mediated endocytosis (see, e. g., Wu and Wu (1987) J. Biol. Chem. 262:4429-4432), construction of a nucleic acid as part of a retroviral or other vector, etc. Methods of administering a prophylactic or therapeutic agent include, but are not limited to, parenteral administration (e.g., intradermal, intramuscular, intraperitoneal, intravenous and subcutaneous), epidural administration, intratumoral administration, and mucosal admins U.S. Patent Nos. 6,019,968; 5,985,320; 5,985,309; 5,934,272; 5,874,064; 5,855,913; 5,290,540; and 4,880,078; and PCT Publication Nos. WO 92/19244; WO 97/32572; WO 97/44013; WO 98/31346; and WO 99/66903. In one embodiment, a binding protein , combination therapy, or a composition is administered using Alkermes AIR® pulmonary drug delivery technology (Alkermes, Inc., Cambridge, Massachusetts). The prophylactic or therapeutic agents may be administered by any convenient route, for example by infusion or bolus injection, by absorption through epithelial or mucocutaneous linings (e.g., oral mucosa, rectal and

intestinal mucosa, etc.) and may be administered together with other biologically active agents. Administration can be systemic or local.

In a specific embodiment, it may be desirable to administer the prophylactic or therapeutic agents locally to the area in need of treatment, for example, by local infusion, by injection, or by means of an implant. An implant may be of a porous or non-porous material, including membranes and matrices, such as sialastic membranes, polymers, fibrous matrices (e.g., Tissuel®), or collagen matrices. In one embodiment, an effective amount of one or more antibodies is administered locally to the affected area to a subject to prevent, treat, manage, and/or ameliorate a disorder or a symptom thereof. In another embodiment, an effective amount of one or more antibodies is administered locally to the affected area in combination with an effective amount of one or more therapies (e.g., one or more prophylactic or therapeutic agents) other than a binding protein of a subject to prevent, treat, manage, and/or ameliorate a disorder or one or more symptoms thereof.

In another embodiment, the prophylactic or therapeutic agent can be delivered in a controlled release or sustained release system. In one embodiment, a pump may be used to achieve controlled or sustained release (see Langer (1990) Science 249:1527-1533; Sefton (1987) CRC Crit. Rev. Biomed. Eng. 14:201-240; Buchwald et al. (1980) Surgery 88:507-516; Saudek et al. (1989) N. Engl. J. Med. 321:574-579). In another embodiment, polymeric materials can be used to achieve controlled or sustained release of the therapies (see, e.g., Medical Applications of Controlled Release, (Langer and Wise, eds.) (CRC Press, Inc., Boca Raton, 1984); Controlled Drug Bioavailability, Drug Product Design and Performance, (Smolen and Ball, eds.) (Wiley, New York, 1984); Langer and Peppas (1983) J. Macromol. Sci. Rev. Macromol. Chem. Phys. C23:61-126; see also Levy et al. (1985) Science 228:190-192; During et al. (1989) Ann. Neurol. 25:351-356; Howard et al. (1989) J. Neurosurg. 71:105-112); U.S. Patent Nos. 5,679,377; 5,916,597; 5,912,015; 5,989,463; and 5,128,326; and PCT Publication Nos. WO 99/15154 and WO 99/20253. Examples of polymers used in sustained release formulations include, but are not limited to, poly(2-hydroxy ethyl methacrylate), poly(methyl methacrylate), poly(acrylic acid), poly(ethylene-co-vinyl acetate), poly(methacrylic acid), polyglycolides (PLG), polyanhydrides, poly(N- vinyl pyrrolidone), poly(vinyl alcohol), polyacrylamide, poly(ethylene glycol), polylactides (PLA), poly(lactide-co-glycolides) (PLGA), and polyorthoesters. In an embodiment, the polymer used in a sustained release formulation is inert, free of leachable impurities, stable on storage, sterile, and biodegradable. In yet another embodiment, a controlled or sustained release system can be placed in proximity of the prophylactic or therapeutic target, thus requiring only a fraction of the systemic dose (see, e.g., Goodson, J.M., Chapter 6, In Medical Applications of Controlled Release, Vol. II, Applications and Evaluation, (Langer and Wise, eds.)(CRC Press, Inc., Boca Raton, 1984), pp. 115-138).

Controlled release systems are discussed in the review by Langer (1990) Science 249:1527-1533. Any technique known to one of skill in the art can be used to produce sustained release formulations comprising one or more therapeutic agents. See, e.g., U. S. Patent No. 4,526,938; and PCT Publication Nos. WO 91/05548 and WO 96/20698; and Ning et al. (1996) Radiother. Oncol. 39:179-189; Song et al. (1996) PDA J. Pharm. Sci. Technol. 50:372-377; Cleek et al. (1997) Proceed Int'l. Symp. Control. Rel. Bioact. Mater. 24:853-854; and Lam et al. (1997) Proceed. Int'l. Symp. Control Rel. Bioact. Mater. 24:759-760.

In a specific embodiment, where the composition is a nucleic acid encoding a prophylactic or therapeutic agent, the nucleic acid can be administered in vivo to promote expression of its encoded prophylactic or therapeutic agent, by constructing it as part of an appropriate nucleic acid expression vector and administering it so that it becomes intracellular, e.g., by use of a retroviral vector (see U.S. Patent No. 4,980,286), or by direct injection, or by use of microparticle bombardment (e.g., a gene gun; Biolistic, Dupont), or coating with lipids or cell-surface receptors or transfecting agents, or by administering it in linkage to a homeobox-like peptide which is known to enter the nucleus (see, e.g., Joliot et al. (1991) Proc. Natl. Acad. Sci. USA 88:1864-1868). Alternatively, a nucleic acid can be introduced intracellularly and d within host cell DNA for expression by homologous recombination.

A pharmaceutical composition is formulated to be compatible with its intended route of administration. Examples of routes of administration include, but are not limited to, parenteral, e.g., intravenous, intradermal, subcutaneous, oral, intranasal (e.g., inhalation), transdermal (e.g., topical), transmucosal, and rectal administration. In a specific embodiment, the composition is formulated in accordance with routine procedures as a pharmaceutical composition adapted for intravenous, subcutaneous, intramuscular, oral, intranasal, or topical administration to human beings. Typically, compositions for intravenous administration are solutions in sterile isotonic aqueous buffer. Where necessary, the composition may also include a solubilizing agent and a local anesthetic such as lignocamne to ease pain at the site of the injection.

If the compositions are to be administered topically, the compositions can be formulated in the form of an ointment, cream, transdermal patch, lotion, gel, shampoo, spray, aerosol, solution, emulsion, or other form well-known to one of skill in the art. See, e.g., Remington's Pharmaceutical Sciences and Introduction to Pharmaceutical Dosage Forms, 19th ed., (Mack Publishing Co., Easton, Pennsylvania, 1995). For non- sprayable topical dosage forms, viscous to semi-solid or solid forms comprising a carrier or one or more excipients compatible with topical application and having a dynamic viscosity preferably greater than water are typically employed. Other suitable formulations include, without limitation, suspensions, powders, liniments, salves, and the like. In an embodiment, such formulations are sterilized or mixed with auxiliary agents (e.g., preservatives, stabilizers, wetting agents, buffers, or salts) for influencing various properties, such as, for example, osmotic pressure. Other suitable topical dosage forms include sprayable

aerosol preparations wherein the active ingredient, for example, in combination with a solid or liquid inert carrier, is packaged in a mixture with a pressurized volatile (e.g., a gaseous propellant, such as FREON®) or in a squeeze bottle. Moisturizers or humectants can also be added to pharmaceutical compositions and dosage forms if desired. Examples of such additional ingredients are well-known in the art.

If the method comprises intranasal administration of a composition, the composition can be formulated in an aerosol form, spray, mist or in the form of drops. In particular, prophylactic or therapeutic agents can be conveniently delivered in the form of an aerosol spray presentation from pressurized packs or a nebulizer, with the use of a suitable propellant (e.g., dichlorodifluoromethane, trichlorofluoromethane, dichlorotetrafluoroethane, carbon dioxide or other suitable gas). In the case of a pressurized aerosol the dosage unit may be determined by providing a valve to deliver a metered amount. Capsules and cartridges (composed of, e.g., gelatin) for use in an inhaler or insufflator may be formulated containing a powder mix of the compound and a suitable powder base such as lactose or starch.

If the method comprises oral administration, compositions can be formulated orally in the form of tablets, capsules, cachets, gelcaps, solutions, suspensions, and the like. Tablets or capsules can be prepared by conventional means with pharmaceutically acceptable excipients such as binding agents (e.g., pregelatinised maize starch, polyvinylpyrrolidone, or hydroxypropyl methylcellulose); fillers (e.g., lactose, microcrystalline cellulose, or calcium hydrogen phosphate) ; lubricants (e.g., magnesium stearate, talc, or silica); disintegrants (e.g., potato starch or sodium starch glycolate) ; or wetting agents (e.g., sodium lauryl sulphate). The tablets may be coated by methods well-known in the art. Liquid preparations for oral administration may take the form of, but not limited to, solutions, syrups or suspensions, or they may be presented as a dry product for constitution with water or other suitable vehicle before use. Such liquid preparations may be prepared by conventional means with pharmaceutically acceptable additives such as suspending agents (e.g., sorbitol syrup, cellulose derivatives, or hydrogenated edible fats); emulsifying agents (e.g., lecithin or acacia); non-aqueous vehicles (e.g., almond oil, oily esters, ethyl alcohol, or fractionated vegetable oils); and preservatives (e.g., methyl or propyl-p- hydroxybenzoates or sorbic acid). The preparations may also contain buffer salts, flavoring, coloring, and sweetening agents as appropriate. Preparations for oral administration may be suitably formulated for slow release, controlled release, or sustained release of a prophylactic or therapeutic agent(s).

The method may comprise pulmonary administration, e.g., by use of an inhaler or nebulizer, of a composition formulated with an aerosolizing agent. See, e.g., U.S. Patent Nos. 6,019, 968; 5,985,320; 5,985,309; 5,934,272; 5,874,064; 5,855,913; 5,290,540; and 4,880,078; and PCT Publication Nos. WO 92/19244, WO 97/32572, WO 97/44013, WO 98/31346, and WO 99/66903. In a specific embodiment, an antibody, combination therapy, and/or composition is

administered using Alkermes AIR® pulmonary drug delivery technology (Alkermes, Inc., Cambridge, Massachusetts).

The method may comprise administration of a composition formulated for parenteral administration by injection (e.g., by bolus injection or continuous infusion). Formulations for injection may be presented in unit dosage form (e.g., in ampoules or in multi-dose containers) with an added preservative. The compositions may take such forms as suspensions, solutions or emulsions in oily or aqueous vehicles, and may contain formulatory agents such as suspending, stabilizing and/or dispersing agents. Alternatively, the active ingredient may be in powder form for constitution with a suitable vehicle (e.g., sterile pyrogen-free water) before use.

The methods may additionally comprise of administration of compositions formulated as depot preparations. Such long acting formulations may be administered by implantation (e.g., subcutaneously or intramuscularly) or by intramuscular injection. Thus, for example, the compositions may be formulated with suitable polymeric or hydrophobic materials (e.g., as an emulsion in an acceptable oil) or ion exchange resins, or as sparingly soluble derivatives (e.g., as a sparingly soluble salt).

The methods encompass administration of compositions formulated as neutral or salt forms. Pharmaceutically acceptable salts include those formed with anions such as those derived from hydrochloric, phosphoric, acetic, oxalic, tartaric acids, etc., and those formed with cations such as those derived from sodium, potassium, ammonium, calcium, ferric hydroxides, isopropylamine, triethylamine, 2- ethylamino ethanol, histidine, procaine, etc.
Generally, the ingredients of compositions are supplied either separately or mixed together in unit dosage form, for example, as a dry lyophilized powder or water free concentrate in a hermetically sealed container such as an ampoule or sachette indicating the quantity of active agent. Where the mode of administration is infusion, composition can be dispensed with an infusion bottle containing sterile pharmaceutical grade water or saline. Where the mode of administration is by injection, an ampoule of sterile water for injection or saline can be provided so that the ingredients may be mixed prior to administration.

In one embodiment, one or more of the prophylactic or therapeutic agents, or pharmaceutical compositions is packaged in a hermetically sealed container such as an ampoule or sachette indicating the quantity of the agent. In one embodiment, one or more of the prophylactic or therapeutic agents, or pharmaceutical compositions is supplied as a dry sterilized lyophilized powder or water free concentrate in a hermetically sealed container and can be reconstituted (e.g., with water or saline) to the appropriate concentration for administration to a subject. In an embodiment, one or more of the prophylactic or therapeutic agents or pharmaceutical compositions is supplied as a dry sterile lyophilized powder in a hermetically sealed container at a unit dosage of at least about 5 mg, at least about 10 mg, at least about 15 mg, at least about 25 mg, at least about 35 mg, at least about 45 mg, at least about 50 mg, at least about 75 mg, or at least

about 100 mg. The lyophilized prophylactic or therapeutic agents or pharmaceutical compositions should be stored at between 2° C. and 8° C. in its original container and the prophylactic or therapeutic agents, or pharmaceutical compositions should be administered within about 1 week, within about 5 days, within about 72 hours, within about 48 hours, within about 24 hours, within about 12 hours, within about 6 hours, within about 5 hours, within about 3 hours, or within about 1 hour after being reconstituted. In an alternative embodiment, one or more of the prophylactic or therapeutic agents or pharmaceutical compositions is supplied in liquid form in a hermetically sealed container indicating the quantity and concentration of the agent. In an embodiment, the liquid form of the administered composition is supplied in a hermetically sealed container at least about 0.25 mg/ml, at least about 0.5 mg/ml, at least about 1 mg/ml, at least about 2.5 mg/ml, at least about 5 mg/ml, at least about 8 mg/ml, at least about 10 mg/ml, at least about 15 mg/kg, at least about 25 mg/ml, at least about 50 mg/ml, at least about 75 mg/ml or at least about 100 mg/ml. The liquid form should be stored at between 2° C and 8° C in its original container.

The binding proteins can be incorporated into a pharmaceutical composition suitable for parenteral administration. In one aspect, the binding proteins are prepared as an injectable solution containing about 0.1 to about 250 mg/ml antibody. The injectable solution can be composed of either a liquid or lyophilized dosage form in a flint or amber vial, ampoule or pre-filled syringe. The buffer can be L-histidine (about 1 to about 50 mM), optimally about 5 to about 10mM, at pH 5.0 to 7.0 (optimally about pH 6.0). Other suitable buffers include but are not limited to, sodium succinate, sodium citrate, sodium phosphate or potassium phosphate. Sodium chloride can be used to modify the toxicity of the solution at a concentration of about 0 to about 300 mM (e.g., about 150 mM for a liquid dosage form). Cryoprotectants can be included for a lyophilized dosage form, principally about 0 to about 10% sucrose (e.g., about 0.5 to about 1.0%). Other suitable cryoprotectants include trehalose and lactose. Bulking agents can be included for a lyophilized dosage form, principally about 1 to about 10% mannitol (e.g., about 2 to about 4%). Stabilizers can be used in both liquid and lyophilized dosage forms, principally about 1 to about 50 mM L-Methionine (optimally about 5 to about 10 mM). Other suitable bulking agents include glycine, arginine, can be included as about 0 to about 0.05% polysorbate-80 (optimally about 0.005 to about 0.01%). Additional surfactants include but are not limited to polysorbate 20 and BRIJ surfactants.

The compositions may be in a variety of forms. These include, for example, liquid, semi-solid and solid dosage forms, such as liquid solutions (e.g., injectable and infusible solutions), dispersions or suspensions, tablets, pills, powders, liposomes and suppositories. The form depends on the intended mode of administration and therapeutic application. Typical compositions are in the form of injectable or infusible solutions, such as compositions similar to those used for passive immunization of humans with other antibodies. In an embodiment, the mode of

administration is parenteral (e.g., intravenous, subcutaneous, intraperitoneal, intramuscular). In an embodiment, the antibody is administered by intravenous infusion or injection. In another embodiment, the antibody is administered by intramuscular or subcutaneous injection.

Therapeutic compositions typically must be sterile and stable under the conditions of manufacture and storage. The composition can be formulated as a solution, microemulsion, dispersion, liposome, or other ordered structure suitable to high drug concentration. Sterile injectable solutions can be prepared by incorporating the active compound (e.g., antibody or antigen binding portion thereof) in the required amount in an appropriate solvent with one or a combination of ingredients enumerated above, as required, followed by filtered sterilization. Generally, dispersions are prepared by incorporating the active compound into a sterile vehicle that contains a basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile, lyophilized powders for the preparation of sterile injectable solutions, exemplary methods of preparation are vacuum drying and spray-drying that yields a powder of the active ingredient plus any additional desired ingredient from a previously sterile-filtered solution thereof. The proper fluidity of a solution can be maintained, for example, by the use of a coating such as lecithin, by the maintenance of the required particle size in the case of dispersion and by the use of surfactants. Prolonged absorption of injectable compositions can be brought about by including, in the composition, an agent that delays absorption, for example, monostearate salts and gelatin.

The antibodies and antigen binding portions can be administered by a variety of methods known in the art, although for many therapeutic applications, an exemplary route/mode of administration is subcutaneous injection, intravenous injection or infusion. As will be appreciated by the skilled artisan, the route and/or mode of administration will vary depending upon the desired results. In certain embodiments, the active compound may be prepared with a carrier that will protect the compound against rapid release, such as a controlled release formulation, including implants, transdermal patches, and microencapsulated delivery systems. Biodegradable, biocompatible polymers can be used, such as ethylene vinyl acetate, polyanhydrides, polyglycolic acid, collagen, polyorthoesters, and polylactic acid. Many methods for the preparation of such formulations are patented or generally known to those skilled in the art. See, *e.g.*, *Sustained and Controlled Release Drug Delivery Systems*, J.R. Robinson, ed., Marcel Dekker, Inc., New York, 1978.

In certain embodiments, an antibody, or antigen binding portion thereof, may be orally administered, for example, with an inert diluent or an assimilable edible carrier. The compound (and other ingredients, if desired) may also be enclosed in a hard or soft shell gelatin capsule, compressed into tablets, or incorporated directly into the subject's diet. For oral therapeutic administration, the compounds may be incorporated with excipients and used in the form of ingestible tablets, buccal tablets, troches, capsules, elixirs, suspensions, syrups, wafers, and the

like. To administer a compound by other than parenteral administration, it may be necessary to coat the compound with, or co-administer the compound with, a material to prevent its inactivation.

Supplementary active compounds can also be incorporated into the compositions. In certain embodiments, a binding protein (e.g., an antibody, or antigen binding portion thereof) is coformulated with and/or coadministered with one or more additional therapeutic agents that are useful for treating disorders in which TNF-α activity is detrimental. For example, an anti-hTNF-α antibody or antigen binding portion may be coformulated and/or coadministered with one or more additional antibodies that bind other targets (*e.g.*, antibodies that bind other cytokines or that bind cell surface molecules). Furthermore, one or more antibodies may be used in combination with two or more of the foregoing therapeutic agents. Such combination therapies may advantageously utilize lower dosages of the administered therapeutic agents, thus avoiding possible toxicities or complications associated with the various monotherapies.

In certain embodiments, an antibody to TNF-α or fragment thereof is linked to a half-life extending vehicle known in the art. Such vehicles include, but are not limited to, the Fc domain, polyethylene glycol, and dextran. Such vehicles are described, e.g., in U.S. Patent No. 6,660,843 and published PCT Publication No. WO 99/25044.

In a specific embodiment, nucleic acid molecules comprising nucleotide sequences encoding one or more polypeptides of a binding protein or another prophylactic or therapeutic agent are administered to treat, prevent, manage, or ameliorate a disorder or one or more symptoms thereof by way of gene therapy. Gene therapy refers to therapy performed by the administration to a subject of an expressed or expressible nucleic acid. In this embodiment, the nucleic acids produce their encoded binding polypeptide(s) of a binding protein or prophylactic or therapeutic agent that mediates a prophylactic or therapeutic effect.

Any of the methods for gene therapy available in the art can be used. For general reviews of the methods of gene therapy, see Goldspiel et al. (1993) Clin. Pharm. 12:488-505; Wu and Wu (1991) Biotherapy 3:87-95; Tolstoshev (1993) Ann. Rev. Pharmacol. Toxicol. 32:573-596; Mulligan (1993) Science 260:926- 932; and Morgan and Anderson (1993) Ann. Rev. Biochem. 62:191-217; Robinson, C. (1993) Trends Biotechnol. 11(5):155. Methods commonly known in the art of recombinant DNA technology which can be used are described in Ausubel et al. (eds.), Current Protocols in Molecular Biology (John Wiley & Sons, New York, 1993); and Kriegler, Gene Transfer and Expression, A Laboratory Manual, (Stockton Press, New York, 1990). Detailed descriptions of various methods of gene therapy are disclosed in US 2005/0042664.

TNF-α plays a role in the pathology associated with a variety of diseases involving immune and inflammatory elements, such as autoimmune diseases, particularly those assocated with inflammation, including Crohn's disease, psoriasis (including plaque psoriasis), arthritis

(including rheumatoid arthritis, psoratic arthritis, osteoarthritis, or juvenile idiopathic arthritis), multiple sclerosis, systemic lupus erythematosus, ankylosing spondylitis, diabetes (including insulin-dependent diabetes mellitus or autoimmune diabetes), allergy, and autoimmune uveitis. Therefore, the binding proteins herein may be used to treat these disorders. In another embodiment, the disorder is a respiratory disorder; asthma; allergic and nonallergic asthma; asthma due to infection; asthma due to infection with respiratory syncytial virus (RSV); chronic obstructive pulmonary disease (COPD); a condition involving airway inflammation; eosinophilia; fibrosis and excess mucus production; cystic fibrosis; pulmonary fibrosis; an atopic disorder; atopic dermatitis; urticaria; eczema; allergic rhinitis; allergic enterogastritis; an inflammatory and/or autoimmune condition of the skin; an inflammatory and/or autoimmune condition of gastrointestinal organs; inflammatory bowel diseases (IBD); ulcerative colitis; an inflammatory and/or autoimmune condition of the liver; liver cirrhosis; liver fibrosis; liver fibrosis caused by hepatitis B and/or C virus; scleroderma; tumors or cancers; hepatocellular carcinoma; glioblastoma; lymphoma; Hodgkin's lymphoma; a viral infection; a bacterial infection; a parasitic infection; HTLV-1 infection; suppression of expression of protective type 1 immune responses, suppression of expression of a protective type 1 immune response during vaccination, neurodegenerative diseases, neuronal regeneration, and spinal cord injury.

TNF-α may also plays a role in the pathology associated with a variety of diseases involving immune and inflammatory elements. These diseases include, but are not limited to, acquired immunodeficiency disease syndrome; acquired Immunodeficiency related diseases; acquired pernicious anaemia; acute coronary syndromes; acute and chronic pain (different forms of pain); acute idiopathic polyneuritis; acute immune disease associated with organ transplantation; acute or chronic immune disease associated with organ transplantation; acute inflammatory demyelinating polyradiculoneuropathy; acute ischemia; acute liver disease; acute rheumatic fever; acute transverse myelitis; Addison's disease; adult (acute) respiratory distress syndrome; adult Still's disease; alcoholic cirrhosis; alcohol-induced liver injury; allergic diseases; allergy; alopecia; alopecia areata; Alzheimer's disease; anaphylaxis; ankylosing spondylitis; ankylosing spondylitis associated lung disease; anti-phospholipid antibody syndrome; aplastic anemia; arteriosclerosis; arthropathy; asthma; atheromatous disease/arteriosclerosis; atherosclerosis; atopic allergy; atopic eczema; atopic dermatitis; atrophic autoimmune hypothyroidism; autoimmune bullous disease; autoimmune dermatitis; autoimmune diabetes; autoimmune disorder associated with streptococcus infection; autoimmune enteropathy; autoimmune haemolytic anaemia; autoimmune hepatitis; autoimmune hearing loss; autoimmune lymphoproliferative syndrome (ALPS); autoimmune mediated hypoglycaemia; autoimmune myocarditis; autoimmune neutropenia; autoimmune premature ovarian failure; autoimmune thrombocytopenia (AITP); autoimmune thyroid disease; autoimmune uveitis; bronchiolitis

obliterans; Behçet's disease; blepharitis; bronchiectasis; bullous pemphigoid; cachexia; cardiovascular disease; catastrophic antiphospholipid syndrome; celiac disease; cervical spondylosis; chlamydia; choleosatatis; chronic active hepatitis; chronic eosinophilic pneumonia; chronic fatigue syndrome; chronic immune disease associated with organ transplantation; chronic ischemia; chronic liver diseases; chronic mucocutaneous candidiasis; cicatricial pemphigoid; clinically isolated syndrome (CIS) with risk for multiple sclerosis; common varied immunodeficiency (common variable hypogammaglobulinaemia); connective tissue disease associated interstitial lung disease; conjunctivitis; Coombs positive haemolytic anaemia; childhood onset psychiatric disorder; chronic obstructive pulmonary disease (COPD); Crohn's disease; cryptogenic autoimmune hepatitis; cryptogenic fibrosing alveolitis; dacryocystitis; depression ; dermatitis scleroderma; dermatomyositis; dermatomyositis/polymyositis associated lung disease; diabetic retinopathy; diabetes mellitus; dilated cardiomyopathy; discoid lupus erythematosus; disk herniation; disk prolapse; disseminated intravascular coagulation; drug-induced hepatitis; drug-induced interstitial lung disease; drug induced immune hemolytic anemia; endocarditis; endometriosis; endophthalmitis; enteropathic synovitis; episcleritis; erythema multiforme; erythema multiforme major; female infertility; fibrosis; fibrotic lung disease; gestational pemphigoid; giant cell arteritis (GCA); glomerulonephritides; goitrous autoimmune hypothyroidism (Hashimoto's disease); Goodpasture's syndrome; gouty arthritis; graft versus host disease (GVHD); Grave's disease; group B streptococci (BGS) infection; Guillain-Barré syndrome (BGS); haemosiderosis associated lung disease; hay fever; heart failure; hemolytic anemia; Henoch-Schoenlein purpura; hepatitis B; hepatitis C; Hughes syndrome; Huntington's chorea; hyperthyroidism; hypoparathyroidism; idiopathic leucopaenia; idiopathic thrombocytopaenia; idiopathic Parkinson's disease; idiopathic interstitial pneumonia; idiosyncratic liver disease; IgE-mediated allergy; immune hemolytic anemia; inclusion body myositis; infectious diseases; infectious ocular inflammatory disease ; inflammatory bowel disease; inflammatory demyelinating disease; inflammatory heart disease; inflammatory kidney disease; insulin dependent diabetes mellitus; interstitial pneumonitis; IPF/UIP; iritis; juvenile chronic arthritis; juvenile pernicious anaemia; juvenile rheumatoid arthritis (JRA); Kawasaki's disease; keratitis; keratojunctivitis sicca; Kussmaul disease or Kussmaul-Meier disease; Landry's paralysis; Langerhan's cell histiocytosis; linear IgA disease; livedo reticularis; Lyme arthritis; lymphocytic infiltrative lung disease; macular degeneration; male infertility idiopathic or NOS; malignancies; microscopic vasculitis of the kidneys; microscopic polyangiitis; mixed connective tissue disease associated lung disease; Morbus Bechterev; motor neuron disorders; mucous membrane pemphigoid ; multiple sclerosis (all subtypes: primary progressive, secondary progressive, relapsing remitting, etc.); multiple organ failure; myalgic encephalitis/royal free disease; myasthenia gravis; myelodysplastic syndrome; myocardial infarction; myocarditis;

nephrotic syndrome; nerve root disorders; neuropathy; non-alcoholic steatohepatitis; non┤ A non┤ B hepatitis; optic neuritis; organ transplant rejection; osteoarthritis; osteolysis; ovarian cancer; ovarian failure; pancreatitis; parasitic diseases; Parkinson's disease; pauciarticular JRA ; pemphigoid; pemphigus foliaceus; pemphigus vulgaris; peripheral artery occlusive disease (PAOD); peripheral vascular disease (PVD); peripheral artery disease (PAD); phacogenic uveitis; phlebitis; polyarteritis nodosa (or periarteritis nodosa); polychondritis; polymyalgia rheumatica; poliosis; polyarticular JRA; polyendocrine deficiency syndrome; polymyositis; polyglandular deficiency type I and polyglandular deficiency type II; polymyalgia rheumatica (PMR); postinfectious interstitial lung disease; post-inflammatory interstitial lung disease; post-pump syndrome; premature ovarian failure; primary biliary cirrhosis; primary myxoedema; primary Parkinsonism; primary sclerosing cholangitis; primary sclerosing hepatitis; primary vasculitis; prostate and rectal cancer and hematopoietic malignancies (leukemia and lymphoma); prostatitis; psoriasis; psoriasis type 1; psoriasis type 2; psoriatic arthritis; psoriatic arthropathy; pulmonary hypertension secondary to connective tissue disease; pulmonary manifestation of polyarteritis nodosa; pure red cell aplasia; primary adrenal insufficiency; radiation fibrosis; reactive arthritis; Reiter's disease; recurrent neuromyelitis optica; renal disease NOS; restenosis; rheumatoid arthritis; rheumatoid arthritis associated interstitial lung disease; rheumatic heart disease; SAPHO (synovitis, acne, pustulosis, hyperostosis, and osteitis); sarcoidosis; schizophrenia; Schmidt's syndrome; scleroderma; secondary amyloidosis; shock lung; scleritis; sciatica; secondary adrenal insufficiency; sepsis syndrome; septic arthritis; septic shock; seronegative arthropathy; silicone associated connective tissue disease; Sjögren's disease associated lung disease; Sjörgren's syndrome; Sneddon-Wilkinson dermatosis; sperm autoimmunity; spondyloarthropathy; spondylitis ankylosans; Stevens-Johnson syndrome (SJS); Still's disease; stroke; sympathetic ophthalmia; systemic inflammatory response syndrome; systemic lupus erythematosus; systemic lupus erythematosus associated lung disease; systemic sclerosis; systemic sclerosis associated interstitial lung disease; Takayasu's disease/arteritis; temporal arteritis; Th2 Type and Th1 Type mediated diseases; thyroiditis; toxic shock syndrome; toxoplasmic retinitis; toxic epidermal necrolysis; transverse myelitis; TRAPS (Tumor-necrosis factor receptor type 1 (TNFR)-associated periodic syndrome); type B insulin resistance with acanthosis nigricans; type 1 allergic reaction; type-1 autoimmune hepatitis (classical autoimmune or lupoid hepatitis); type-2 autoimmune hepatitis (anti-LKM antibody hepatitis); type II diabetes; ulcerative colitic arthropathy; ulcerative colitis; urticaria; usual interstitial pneumonia (UIP); uveitis; vasculitic diffuse lung disease; vasculitis; vernal conjunctivitis; viral retinitis; vitiligo ; Vogt-Koyanagi-Harada syndrome (VKH syndrome); Wegener's granulomatosis; wet macular degeneration; wound healing; yersinia and salmonella associated arthropathy.

TNF-α binding proteins , or antigen binding portions thereof, can be used alone or in combination to treat such diseases, including in combination with additional therapeutic agents useful in the treatment of autoimmune and inflammatory diseases. It should be understood that the binding proteins or antigen binding portions thereof can be used alone or in combination with an additional agent, e.g., a therapeutic agent, said additional agent being selected by the skilled artisan for its intended purpose. For example, the additional agent can be a therapeutic agent art-recognized as being useful to treat the disease or condition being treated by the antibody. The additional agent also can be an agent that imparts a beneficial attribute to the therapeutic composition, e.g., an agent that affects the viscosity of the composition.

The combinations include the TNF-α binding proteins, or antigen binding fragments thereof, described herein and at least one additional agent listed below. The combination can also include more than one additional agent, e.g., two or three additional agents if the combination is such that the formed composition can perform its intended function.

In one embodiment, combinations include the TNF-α binding proteins, or antigen binding fragments thereof, and an antibody, or fragment thereof, capable of binding human IL-12; PGE2; LPA; NGF; CGRP; SubP; RAGE; histamine; a histamine receptor blocker; bradykinin; IL-1alpha; IL-1beta; VEGF; PLGF; methotrexate; a corticosteroid, a glucocorticoid receptor modulator; cyclosporin, rapamycin, FK506, or a non-steroidal anti-inflammatory agent.

In another embodiment, exemplary combinations include the TNF-α binding proteins, or antigen binding fragments thereof, described herein and a non-steroidal anti-inflammatory drug(s) (NSAIDS), such as, for example, ibuprofen. Other exemplary combinations comprise the antibodies, or antigen binding fragments thereof, described herein and corticosteroids including prednisolone. The side-effects of steroid use can be reduced or eliminated by tapering the steroid dose required when treating patients in combination with the TNF-α binding proteins. Non-limiting examples of therapeutic agents for rheumatoid arthritis with which an antibody, or antibody portion, can be combined include the following: cytokine suppressive anti-inflammatory drug(s) (CSAIDs); antibodies to or antagonists of other human cytokines or growth factors, for example, TNF, LT, IL-1α, IL-2, IL-3, IL-4, IL-5, IL-6, IL-7, IL-8, IL-15, IL-16, IL-18, IL-21, interferons, EMAP-II, GM-CSF, FGF, and PDGF, TNF family members such as, for example, TRAIL, FASL, APRIL, etc., and antibodies to lipid mediators of disease such prostaglandins, e.g., PGE2, S1P, LPA, etc. Other disease mediators include sclerostin, NGF, substance P, CGRP, and other pain mediators. Antibodies, or antigen binding portions thereof, can be combined with antibodies to cell surface molecules such as CD2, CD3, CD4, CD8, CD25, CD28, CD30, CD40, CD45, CD69, CD80 (B7.1), CD86 (B7.2), CD90, CTLA or their ligands including CD154 (gp39 or CD40L).

Exemplary therapeutic agents for combining with the TNF-α binding proteins, or antigen binding fragments thereof, interfere at different points in the autoimmune and subsequent inflammatory cascade, for example, TNF antagonists like chimeric, humanized or human TNF antibodies, D2E7, (PCT Publication No. WO 97/29131), CA2 (REMICADE®), CDP 571, and soluble p55 or p75 TNF receptors, derivatives thereof, (p75TNFR1gG (ENBREL®) or p55TNFR1gG (Lenercept), and also TNF-α converting enzyme (TACE) inhibitors, and other IL-1 inhibitors (Interleukin-1-converting enzyme inhibitors, IL-1RA, etc.). Other agents for combining with the antibodies and antigen binding fragments thereof include Interleukin 11, agents that act parallel to, dependent on, or in concert with TNF-α function such as, for example, IL-18 antagonists (e.g., IL-18 binding proteins such as, for example, antibodies or soluble IL-18 receptors, or antigen binding fragments thereof. Additional agents for combining with the antibodies and antigen binding fragments thereof include non-depleting anti-CD4 inhibitors, antagonists of the co-stimulatory pathway CD80 (B7.1) or CD86 (B7.2) including antibodies, soluble receptors, antagonistic ligands, or antigen binding fragments thereof.

The binding proteins , or antigen binding portions thereof, may also be combined with agents for treatment of rheumatoid arthritis, for example, such as methotrexate, 6-MP, azathioprine sulphasalazine, mesalazine, olsalazine chloroquinine/hydroxychloroquine, pencillamine, aurothiomalate (intramuscular and oral), azathioprine, colchicine, corticosteroids (oral, inhaled and local injection), beta-2 adrenoreceptor agonists (salbutamol, terbutaline, salmeteral), xanthines (theophylline, aminophylline), cromoglycate, nedocromil, ketotifen, ipratropium and oxitropium, cyclosporin, FK506, rapamycin, mycophenolate mofetil, leflunomide, NSAIDs, for example, ibuprofen, corticosteroids such as prednisolone, phosphodiesterase inhibitors, adenosine agonists, antithrombotic agents, complement inhibitors, adrenergic agents, agents that interfere with signaling by proinflammatory cytokines such as TNF-α or IL-1 (e.g., IRAK, NIK, IKK , p38 and MAP kinase inhibitors), IL-1beta converting enzyme inhibitors, TNF-α converting enzyme (TACE) inhibitors, T-cell signaling inhibitors such as kinase inhibitors, metalloproteinase inhibitors, sulfasalazine, azathioprine, 6-mercaptopurines, angiotensin converting enzyme inhibitors, soluble cytokine receptors and derivatives thereof (e.g., soluble p55 or p75 TNF receptors and the derivatives p75TNFRIgG (ENBREL™ and p55TNFRIgG (Lenercept)), sIL-1RI, sIL-1RII, sIL-6R), antiinflammatory cytokines (e.g., IL-4, IL-10, IL-11, IL-13 and TGFβ), celecoxib, folic acid, hydroxychloroquine sulfate, rofecoxib, etanercept, infliximab, naproxen, valdecoxib, sulfasalazine, methylprednisolone, meloxicam, methylprednisolone acetate, gold sodium thiomalate, aspirin, triamcinolone acetonide, propoxyphene napsylate/apap, folate, nabumetone, diclofenac, piroxicam, etodolac, diclofenac sodium, oxaprozin, oxycodone hcl, hydrocodone bitartrate/apap, diclofenac sodium/misoprostol, fentanyl, anakinra, human recombinant, tramadol hcl, salsalate, sulindac,

cyanocobalamin/fa/pyridoxine, acetaminophen, alendronate sodium, prednisolone, morphine sulfate, lidocaine hydrochloride, indomethacin, glucosamine sulf/chondroitin, amitriptyline hcl, sulfadiazine, oxycodone hcl/acetaminophen, olopatadine hcl, misoprostol, naproxen sodium, omeprazole, cyclophosphamide, rituximab, IL-1 TRAP, MRA, CTLA4-IG, IL-18 BP, anti-IL-18, Anti-IL15, BIRB-796, SCIO-469, VX-702, AMG-548, VX-740, Roflumilast, IC-485, CDC-801, and Mesopram.

Non-limiting examples of therapeutic agents for inflammatory bowel disease with which an TNF-α binding protein (e.g., an antibody), or antigen binding portion thereof, can be combined include the following: budenoside; epidermal growth factor, corticosteroids, cyclosporin, sulfasalazine, aminosalicylates, 6-mercaptopurine, azathioprine, metronidazole, lipoxygenase inhibitors, mesalamine, olsalazine, balsalazide, antioxidants, thromboxane inhibitors, IL-1 receptor antagonists, anti-IL-1 monoclonal antibodies, anti-IL-6 monoclonal antibodies, growth factors, elastase inhibitors, pyridinyl-imidazole compounds, antibodies to or antagonists of other human cytokines or growth factors, for example, TNF, LT, IL-1α, IL-2, IL-6, IL-7, IL-8, IL-15, IL-16, IL-17, IL-18, EMAP-II, GM-CSF, FGF, and PDGF. Antibodies, or antigen binding portions thereof, can be combined with antibodies to cell surface molecules such as CD2, CD3, CD4, CD8, CD25, CD28, CD30, CD40, CD45, CD69, CD90 and their ligands. The binding proteins, or antigen binding portions thereof, may also be combined with agents, such as methotrexate, cyclosporin, FK506, rapamycin, mycophenolate mofetil, leflunomide, NSAIDs, for example, ibuprofen, corticosteroids such as prednisolone, phosphodiesterase inhibitors, adenosine agonists, antithrombotic agents, complement inhibitors, adrenergic agents, agents which interfere with signaling by proinflammatory cytokines such as TNF-α or IL-1 (e.g., IRAK, NIK, IKK, p38 or MAP kinase inhibitors), IL-1α converting enzyme inhibitors, TNF-α converting enzyme inhibitors, T-cell signaling inhibitors such as kinase inhibitors, metalloproteinase inhibitors, sulfasalazine, azathioprine, 6-mercaptopurines, angiotensin converting enzyme inhibitors, soluble cytokine receptors and derivatives thereof (e.g., soluble p55 or p75 TNF receptors, sIL-1RI, sIL-1RII, sIL-6R) and antiinflammatory cytokines (e.g., IL-4, IL-10, IL-11, IL-13, and TGFβ).

Exemplary examples of therapeutic agents for Crohn's disease in which an TNF-α binding protein or an antigen binding portion thereof, as described herein, can be combined include the following: TNF antagonists, for example, anti-TNF antibodies, D2E7 (PCT Publication No. WO 97/29131; HUMIRA®), CA2 (REMICADE®), CDP 571, TNFR-Ig constructs, (p75TNFRIgG (ENBREL®) and p55TNFRIgG (Lenercept)) inhibitors, other TNF antagonists such as, for example, Golimumab (Simponi), and PDE4 inhibitors. Binding proteins, or antigen binding portions thereof, can be combined with corticosteroids, for example, budenoside and dexamethasone. Binding proteins, or antigen binding portions thereof, may also be combined with

agents such as sulfasalazine, 5-aminosalicylic acid and olsalazine, and agents that interfere with synthesis or action of proinflammatory cytokines such as IL-1, for example, IL-1α converting enzyme inhibitors and IL-1RA. Binding proteins or antigen binding portion thereof may also be used with T cell signaling inhibitors, for example, tyrosine kinase inhibitors 6-mercaptopurines. Binding proteins, or antigen binding portions thereof, can be combined with IL-11. Binding proteins, or antigen binding portions thereof, can be combined with mesalamine, prednisone, azathioprine, mercaptopurine, infliximab, methylprednisolone sodium succinate, diphenoxylate/atrop sulfate, loperamide hydrochloride, methotrexate, omeprazole, folate, ciprofloxacin/dextrose-water, hydrocodone bitartrate/apap, tetracycline hydrochloride, fluocinonide, metronidazole, thimerosal/boric acid, cholestyramine/sucrose, ciprofloxacin hydrochloride, hyoscyamine sulfate, meperidine hydrochloride, midazolam hydrochloride, oxycodone hcl/acetaminophen, promethazine hydrochloride, sodium phosphate, sulfamethoxazole/trimethoprim, celecoxib, polycarbophil, propoxyphene napsylate, hydrocortisone, multivitamins, balsalazide disodium, codeine phosphate/apap, colesevelam hcl, cyanocobalamin, folic acid, levofloxacin, methylprednisolone, natalizumab and interferon-gamma.

Non-limiting examples of therapeutic agents for multiple sclerosis with which a TNFα binding protein, or antigen binding portion, can be combined include the following: corticosteroids, prednisolone, methylprednisolone, azathioprine, cyclophosphamide, cyclosporine, methotrexate, 4-aminopyridine, tizanidine, interferon-β1a (AVONEX®; Biogen), interferon- β 1b (BETASERON®; Chiron/Berlex), interferonα -n3 (Interferon Sciences/Fujimoto), interferon-α (Alfa Wassermann/J&J), interferon β 1A-IF (Serono/Inhale Therapeutics), Peginterferonα 2b (Enzon/Schering-Plough), Copolymer 1 (Cop-1; COPAXONE®; Teva Pharmaceutical Industries, Inc.), hyperbaric oxygen, intravenous immunoglobulin, clabribine, antibodies to or antagonists or inhibitors of other human cytokines or growth factors and their receptors, for example, TNF, LT, IL-1 β, IL-2, IL-6, IL-7, IL-8, IL-1A, IL-15, IL-16, IL-18, EMAP-II, GM-CSF, FGF, and PDGF. Antibodies, or antigen binding portions thereof, can be combined with antibodies to cell surface molecules such as CD2, CD3, CD4, CD8, CD19, CD20, CD25, CD28, CD30, CD40, CD45, CD69, CD80, CD86, CD90 or their ligands. The antibodies, or antigen binding portions thereof, may also be combined with agents, such as FK506, rapamycin, mycophenolate mofetil, leflunomide, NSAIDs, for example, ibuprofen, phosphodiesterase inhibitors, adensosine agonists, antithrombotic agents, complement inhibitors, adrenergic agents, agents which interfere with signaling by proinflammatory cytokines such as TNFα or IL-1 (e.g., IRAK, NIK, IKK, p38 or MAP kinase inhibitors), IL-1β converting enzyme inhibitors, TACE inhibitors, T-cell signaling inhibitors such as kinase inhibitors, metalloproteinase inhibitors, sulfasalazine, azathioprine, 6-mercaptopurines, angiotensin converting enzyme inhibitors, soluble cytokine receptors and

derivatives thereof (e.g., soluble p55 or p75 TNF receptors, sIL-1RI, sIL-1RII, sIL-6R), antiinflammatory cytokines (e.g., IL-4, IL-10, IL-13 and TGFβ), COPAXONE®, and caspase inhibitors, for example inhibitors of caspase-1.

The TNF-α binding proteins, or antigen binding portions thereof, may also be combined with agents, such as alemtuzumab, dronabinol, Unimed, daclizumab, mitoxantrone, xaliproden hydrochloride, fampridine, glatiramer acetate, natalizumab, sinnabidol, a-immunokine NNSO3, ABR-215062, AnergiX.MS, chemokine receptor antagonists, BBR-2778, calagualine, CPI-1189, LEM (liposome encapsulated mitoxantrone), THC.CBD (cannabinoid agonist) MBP-8298, mesopram (PDE4 inhibitor), MNA-715, anti-IL-6 receptor antibody, neurovax, pirfenidone allotrap 1258 (RDP-1258), sTNF-R1, talampanel, teriflunomide,TGF-β2, tiplimotide, VLA-4 antagonists (for example, TR-14035, VLA4 Ultrahaler, Antegran-ELAN/Biogen), interferon gamma antagonists, IL-4 agonists.

Non-limiting examples of therapeutic agents for the treatment or prevention of angina with which an TNFα binding protein, or antigen binding portion thereof, can be combined include the following: aspirin, nitroglycerin, isosorbide mononitrate, metoprolol succinate, atenolol, metoprolol tartrate, amlodipine besylate, diltiazem hydrochloride, isosorbide dinitrate, clopidogrel bisulfate, nifedipine, atorvastatin calcium, potassium chloride, furosemide, simvastatin, verapamil hcl, digoxin, propranolol hydrochloride, carvedilol, lisinopril, spironolactone, hydrochlorothiazide, enalapril maleate, nadolol, ramipril, enoxaparin sodium, heparin sodium, valsartan, sotalol hydrochloride, fenofibrate, ezetimibe, bumetanide, losartan potassium, lisinopril/hydrochlorothiazide, felodipine, captopril, and bisoprolol fumarate.

Non-limiting examples of therapeutic agents for the treatment or prevention of ankylosing spondylitis with which a binding protein, or antigen binding portion thereof, can be combined include the following: ibuprofen, diclofenac and misoprostol, naproxen, meloxicam, indomethacin, diclofenac, celecoxib, rofecoxib, sulfasalazine, methotrexate, azathioprine, minocyclin, prednisone, etanercept, and infliximab.

Non-limiting examples of therapeutic agents for the treatment or prevention of asthma with which an TNF-α binding protein, or antigen binding portion thereof, can be combined include the following: albuterol, salmeterol/fluticasone, montelukast sodium, fluticasone propionate, budesonide, prednisone, salmeterol xinafoate, levalbuterol hcl, albuterol sulfate/ipratropium, prednisolone sodium phosphate, triamcinolone acetonide, beclomethasone dipropionate, ipratropium bromide, azithromycin, pirbuterol acetate, prednisolone, theophylline anhydrous, methylprednisolone sodium succinate, clarithromycin, zafirlukast, formoterol fumarate, influenza virus vaccine, methylprednisolone, amoxicillin trihydrate, flunisolide, allergy

injection, cromolyn sodium, fexofenadine hydrochloride, flunisolide/menthol, amoxicillin/clavulanate, levofloxacin, inhaler assist device, guaifenesin, dexamethasone sodium phosphate, moxifloxacin hcl, doxycycline hyclate, guaifenesin/d-methorphan, p-ephedrine/cod/chlorphenir, gatifloxacin, cetirizine hydrochloride, mometasone furoate, salmeterol xinafoate, benzonatate, cephalexin, pe/hydrocodone/chlorphenir, cetirizine hcl/pseudoephed, phenylephrine/cod/promethazine, codeine/promethazine, cefprozil, dexamethasone, guaifenesin/pseudoephedrine, chlorpheniramine/hydrocodone, nedocromil sodium, terbutaline sulfate, epinephrine, methylprednisolone, and metaproterenol sulfate.

Non-limiting examples of therapeutic agents for the treatment or prevention of COPD with which an TNF-α binding protein, or antigen binding portion thereof, can be combined include the following: albuterol sulfate/ipratropium, ipratropium bromide, salmeterol/fluticasone, albuterol, salmeterol xinafoate, fluticasone propionate, prednisone, theophylline anhydrous, methylprednisolone sodium succinate, montelukast sodium, budesonide, formoterol fumarate, triamcinolone acetonide, levofloxacin, guaifenesin, azithromycin, beclomethasone dipropionate, levalbuterol hcl, flunisolide, ceftriaxone sodium, amoxicillin trihydrate, gatifloxacin, zafirlukast, amoxicillin/clavulanate, flunisolide/menthol, chlorpheniramine/hydrocodone, metaproterenol sulfate, methylprednisolone, mometasone furoate, p-ephedrine/cod/chlorphenir, pirbuterol acetate, p-ephedrine/loratadine, terbutaline sulfate, tiotropium bromide, (R,R)-formoterol, TgAAT, cilomilast, and roflumilast.

Non-limiting examples of therapeutic agents for the treatment or prevention of HCV with which an TNF-α binding protein, or antigen binding portion thereof, can be combined include the following: interferon-alpha-2a, interferon-alpha-2b, interferon-alpha con1, interferon-alpha-n1, pegylated interferon-alpha-2a, pegylated interferon-alpha-2b, ribavirin, peginterferon alfa-2b + ribavirin, ursodeoxycholic acid, glycyrrhizic acid, thymalfasin, maxamine, VX-497 and any compounds that are used to treat HCV through intervention with the following targets:HCV polymerase, HCV protease, HCV helicase, HCV IRES (internal ribosome entry site).

Non-limiting examples of therapeutic agents for the treatment or prevention of idiopathic pulmonary fibrosis with which a binding protein, or antigen binding portion thereof, can be combined include the following: prednisone, azathioprine, albuterol, colchicine, albuterol sulfate, digoxin, gamma interferon, methylprednisolone sod succ, lorazepam, furosemide, lisinopril, nitroglycerin, spironolactone, cyclophosphamide, ipratropium bromide, actinomycin d, alteplase, fluticasone propionate, levofloxacin, metaproterenol sulfate, morphine sulfate, oxycodone HCl, potassium chloride, triamcinolone acetonide, tacrolimus anhydrous, calcium, interferon-alpha, methotrexate, mycophenolate mofetil, and interferon-gamma-1b.

Non-limiting examples of therapeutic agents for the treatment or prevention of myocardial infarction with which an TNFα binding protein, or antigen binding portion thereof, can be combined include the following: aspirin, nitroglycerin, metoprolol tartrate, enoxaparin sodium, heparin sodium, clopidogrel bisulfate, carvedilol, atenolol, morphine sulfate, metoprolol succinate, warfarin sodium, lisinopril, isosorbide mononitrate, digoxin, furosemide, simvastatin, ramipril, tenecteplase, enalapril maleate, torsemide, retavase, losartan potassium, quinapril hcl/mag carb, bumetanide, alteplase, enalaprilat, amiodarone hydrochloride, tirofiban hcl m-hydrate, diltiazem hydrochloride, captopril, irbesartan, valsartan, propranolol hydrochloride, fosinopril sodium, lidocaine hydrochloride, eptifibatide, cefazolin sodium, atropine sulfate, aminocaproic acid, spironolactone, interferon, sotalol hydrochloride, potassium chloride, docusate sodium, dobutamine hcl, alprazolam, pravastatin sodium, atorvastatin calcium, midazolam hydrochloride, meperidine hydrochloride, isosorbide dinitrate, epinephrine, dopamine hydrochloride, bivalirudin, rosuvastatin, ezetimibe/simvastatin, avasimibe, and cariporide.

Non-limiting examples of therapeutic agents for the treatment or prevention of psoriasis with which an TNFα binding protein, or antigen binding portion thereof, can be combined include the following: calcipotriene, clobetasol propionate, triamcinolone acetonide, halobetasol propionate, tazarotene, methotrexate, fluocinonide, betamethasone diprop augmented, fluocinolone acetonide, acitretin, tar shampoo, betamethasone valerate, mometasone furoate, ketoconazole, pramoxine/fluocinolone, hydrocortisone valerate, flurandrenolide, urea, betamethasone, clobetasol propionate/emoll, fluticasone propionate, azithromycin, hydrocortisone, moisturizing formula, folic acid, desonide, pimecrolimus, coal tar, diflorasone diacetate, etanercept folate, lactic acid, methoxsalen, hc/bismuth subgal/znox/resor, methylprednisolone acetate, prednisone, sunscreen, halcinonide, salicylic acid, anthralin, clocortolone pivalate, coal extract, coal tar/salicylic acid, coal tar/salicylic acid/sulfur, desoximetasone, diazepam, emollient, fluocinonide/emollient, mineral oil/castor oil/na lact, mineral oil/peanut oil, petroleum/isopropyl myristate, psoralen, salicylic acid, soap/tribromsalan, thimerosal/boric acid, celecoxib, infliximab, cyclosporine, alefacept, efalizumab, tacrolimus, pimecrolimus, PUVA, UVB, and sulfasalazine.

Non-limiting examples of therapeutic agents for the treatment or prevention of psoriatic arthritis with which an TNFα binding protein, or antigen binding portion thereof, can be combined include the following: methotrexate, etanercept, rofecoxib, celecoxib, folic acid, sulfasalazine, naproxen, leflunomide, methylprednisolone acetate, indomethacin, hydroxychloroquine sulfate, prednisone, sulindac, betamethasone diprop augmented, infliximab, methotrexate, folate, triamcinolone acetonide, diclofenac, dimethylsulfoxide, piroxicam, diclofenac sodium, ketoprofen, meloxicam, methylprednisolone, nabumetone, tolmetin sodium,

calcipotriene, cyclosporine, diclofenac sodium/misoprostol, fluocinonide, glucosamine sulfate, gold sodium thiomalate, hydrocodone bitartrate/apap, ibuprofen, risedronate sodium, sulfadiazine, thioguanine, valdecoxib, alefacept, and efalizumab.

Non-limiting examples of therapeutic agents for the treatment or prevention of restenosis with which an TNFα binding protein, or antigen binding portion thereof, can be combined include the following: sirolimus, paclitaxel, everolimus, tacrolimus, ABT-578, and acetaminophen.

Non-limiting examples of therapeutic agents for the treatment or prevention of sciatica with which an TNFα binding protein, or antigen binding portion thereof, can be combined include the following: hydrocodone bitartrate/apap, rofecoxib, cyclobenzaprine hcl, methylprednisolone, naproxen, ibuprofen, oxycodone hcl/acetaminophen, celecoxib, valdecoxib, methylprednisolone acetate, prednisone, codeine phosphate/apap, tramadol hcl/acetaminophen, metaxalone, meloxicam, methocarbamol, lidocaine hydrochloride, diclofenac sodium, gabapentin, dexamethasone, carisoprodol, ketorolac tromethamine, indomethacin, acetaminophen, diazepam, nabumetone, oxycodone hcl, tizanidine hcl, diclofenac sodium/misoprostol, propoxyphene napsylate/apap, asa/oxycod/oxycodone ter, ibuprofen/hydrocodone bit, tramadol hcl, etodolac, propoxyphene hcl, amitriptyline hcl, carisoprodol/codeine phos/asa, morphine sulfate, multivitamins, naproxen sodium, orphenadrine citrate, and temazepam.

Non-limiting examples of therapeutic agents for the treatment or prevention of systemic lupus erythematosis (SLE) with which an TNFα binding protein, or an antigen binding portion thereof, can be combined include the following: NSAIDS, for example, diclofenac, naproxen, ibuprofen, piroxicam, indomethacin, COX2 inhibitors, for example, celecoxib, rofecoxib, valdecoxib, anti-malarials, for example, hydroxychloroquine, steroids, for example, prednisone, prednisolone, budenoside, dexamethasone, cytotoxics, for example, azathioprine, cyclophosphamide, mycophenolate mofetil, methotrexate, inhibitors of PDE4 or of purine synthesis inhibitor, for example, CELLCEPT®. Binding proteins , or antigen binding portions thereof, may also be combined with agents such as sulfasalazine, 5-aminosalicylic acid, olsalazine, Imuran and agents which interfere with synthesis, production or action of proinflammatory cytokines such as IL-1, for example, caspase inhibitors like IL-1β converting enzyme inhibitors and IL-1ra. Binding proteins , or antigen binding portion thereof, may also be used with T cell signaling inhibitors, for example, tyrosine kinase inhibitors, or molecules that target T cell activation molecules, for example, CTLA-4-IgG or anti-B7 family antibodies, and anti-PD-1 family antibodies. Binding proteins , or antigen binding portions thereof, can be combined with IL-11 or anti-cytokine antibodies, for example, fonotolizumab (anti-IFNg antibody), or anti-receptor receptor antibodies, for example, anti-IL-6 receptor antibody and antibodies to B-cell surface molecules. Binding proteins, or antigen binding portion thereof, may

also be used with LJP 394 (abetimus), agents that deplete or inactivate B-cells, for example, rituximab (anti-CD20 antibody), lymphostat-B (anti-BlyS antibody), TNF antagonists, for example, anti-TNF antibodies, D2E7 (PCT Publication No. WO 97/29131; HUMIRA®), CA2 (REMICADE®), CDP 571, TNFR-Ig constructs, (p75TNFRIgG (ENBREL®) and p55TNFRIgG (Lenercept)).

The pharmaceutical compositions may include a "therapeutically effective amount" or a "prophylactically effective amount" of an TNFα binding protein, or antigen binding portion thereof. A "therapeutically effective amount" refers to an amount effective, at dosages and for periods of time necessary, to achieve the desired therapeutic result. A therapeutically effective amount of a binding protein, or antigen binding portion thereof, described herein may be determined by a person skilled in the art and may vary according to factors such as the disease state, age, sex, and weight of the individual, and the ability of the antibody, or antigen binding portion thereof, to elicit a desired response in the individual. A therapeutically effective amount is also one in which any toxic or detrimental effects of the antibody, or antigen binding portion thereof, are outweighed by the therapeutically beneficial effects. A "prophylactically effective amount" refers to an amount effective, at dosages and for periods of time necessary, to achieve the desired prophylactic result. Typically, since a prophylactic dose is used in subjects prior to or at an earlier stage of disease, the prophylactically effective amount will be less than the therapeutically effective amount.

Dosage regimens may be adjusted to provide the optimum desired response (*e.g.*, a therapeutic or prophylactic response). For example, a single bolus may be administered, several divided doses may be administered over time or the dose may be proportionally reduced or increased as indicated by the exigencies of the therapeutic situation. It is especially advantageous to formulate parenteral compositions in dosage unit form for ease of administration and uniformity of dosage. Dosage unit form refers to physically discrete units suited as unitary dosages for the mammalian subjects to be treated; each unit containing a predetermined quantity of active compound calculated to produce the desired therapeutic effect in association with the required pharmaceutical carrier. The specification for the dosage unit forms are dictated by and directly dependent on (a) the unique characteristics of the active compound and the particular therapeutic or prophylactic effect to be achieved, and (b) the limitations inherent in the art of compounding such an active compound for the treatment of sensitivity in individuals.

An exemplary, non-limiting range for a therapeutically or prophylactically effective amount of TNFα binding protein, or antigen binding portion thereof, is about 0.1 to about 20 mg/kg, about 1 to about 10 mg/kg. Dosag values may vary with the type and severity of the condition to be alleviated. For any particular subject, specific dosage regimens should be adjusted

over time according to the individual need and the professional judgment of the person administering or supervising the administration of the compositions. Dosage ranges set forth herein are exemplary only and are not intended to limit the scope or practice of the claimed composition.

It will be readily apparent to those skilled in the art that other suitable modifications and adaptations of the compositions and methods described herein are obvious and may be made using suitable equivalents without departing from the scope of the disclosure or the embodiments disclosed herein. The present embodiments will be more clearly understood by reference to the following examples, which are included for purposes of illustration only and are not intended to be limiting.

EXEMPLIFICATION

Example 1: Generation Of Anti-Human TNF-α Monoclonal Antibodies

Mouse anti-human TNF-α monoclonal antibodies are obtained as follows:

Example 1.1: Immunization Of Mice With Human TNF-α Antigen

Twenty micrograms of recombinant purified human TNF-α (R&D Systems, Minneapolis, MN, USA) mixed with complete Freund's adjuvant or Immunoeasy adjuvant (Qiagen, Valencia, CA)is injected subcutaneously into five 6-8 week-old Balb/C, five C57B/6 mice, and five AJ mice on Day 1. On days 24, 38, and 49, twenty micrograms of recombinant purified human TNF-α antigen mixed with incomplete Freund's adjuvant or Immunoeasy adjuvant is injected subcutaneously into the same mice. On day 84 or day 112 or day 144, mice are injected intravenously with 1 μg recombinant purified human TNF-α antigen.

Example 1.2: Generation of Hybridoma

Splenocytes obtained from the immunized mice described in Example 1.1 are fused with SP2/O-Ag-14 cells at a ratio of 5:1 according to the established method described in Kohler and Milstein (1975) Nature 256:495 to generate hybridomas. Fusion products are plated in selection media containing azaserine and hypoxanthine in 96-well plates at a density of 2.5x106 spleen cells per well. Seven to ten days post fusion, macroscopic hybridoma colonies are observed. Supernatant from each well containing hybridoma colonies is tested by ELISA for the presence of antibody to TNF-α. Supernatants displaying TNF-α specific activity are then tested for the ability to neutralize TNF-α in the L929 bioassay (as described in Example 2.7).

Example 1.3: Identification And Characterization Of Anti Human TNF-α Monoclonal Antibodies

Hybridomas producing antibodies that bind TNF-α and are capable of binding TNF-α specifically and particularly those with IC50 values in the L929 bioassay of 5nM or less than 5nM are scaled up and cloned by limiting dilution.

Hybridoma cells are expanded into media containing 10% low IgG fetal bovine serum (Hyclone #SH30151, Logan, UT). On average, 250 mL of each hybridoma supernatant (derived from a clonal population) is harvested, concentrated and purified by protein A affinity chromatography by standard methods. The ability of purified mAbs to inhibit TNF-α activity is determined using the L929 bioassay as described in Example 2.7.

Example 1.4 Determination Of The Amino Acid Sequence Of The Variable Region For Each Murine Anti-Human TNF-α Monoclonal Antibody

For each amino acid sequence determination, approximately 10x106 hybridoma cells are isolated by centrifugation and processed to isolate total RNA with Trizol (Gibco BRL/Invitrogen, Carlsbad, CA.) following manufacturer's instructions. Total RNA is subjected to first strand DNA synthesis using the SuperScript First-Strand Synthesis System (Invitrogen, Carlsbad, CA) per the manufacturer's instructions. Oligo(dT) is used to prime first-strand synthesis to select for poly(A)+ RNA. The first-strand cDNA product is then amplified by PCR with primers designed for amplification of murine immunoglobulin variable regions (Ig-Primer Sets, Novagen, Madison, WI). PCR products are resolved on an agarose gel, excised, purified, and then subcloned with the TOPO Cloning kit into pCR2.1-TOPO vector (Invitrogen, Carlsbad, CA) and transformed into TOP10 chemically competent E. coli (Invitrogen, Carlsbad, CA). Colony PCR is performed on the transformants to identify clones containing insert. Plasmid DNA is isolated from clones containing insert using a QIAprep Miniprep kit (Qiagen, Valencia, CA). Inserts in the plasmids are sequenced on both strands to determine the variable heavy or variable light chain DNA sequences using M13 forward and M13 reverse primers (Fermentas Life Sciences, Hanover MD). Variable heavy and variable light chain sequences of the anti- TNF-α monoclonal antibodies are shown in Table 5.

Example 2: Recombinant anti human TNF-α Antibodies

Example 2.1: Construction And Expression Of Recombinant Chimeric Anti Human TNF-α Antibodies

The DNA encoding the heavy chain constant region of murine anti-human TNF-α monoclonal antibodies MAK199 was replaced by a cDNA fragment encoding the human IgG1 constant region containing 2 hinge-region amino acid mutations by homologous recombination in bacteria. These mutations are a leucine to alanine change at position 234 (EU numbering) and a leucine to alanine change at position 235 (Lund et al. (1991) J. Immunol. 147:2657). The light chain constant region of each of these antibodies was replaced by a human kappa constant region. Full-length chimeric antibodies were transiently expressed in HEK293-6E cells by co-transfection of chimeric heavy and light chain cDNAs ligated into the pHybE expression plasmid (US Patent Publication No. US 20090239259). Cell supernatants containing recombinant chimeric antibody

were purified by Protein A Sepharose chromatography and bound antibody was eluted by addition of acid buffer. Antibodies were neutralized and dialyzed into PBS.

The purified chimeric anti-human TNF-α monoclonal antibodies were then tested for their ability to bind the h TNF-α protein by ELISA to confirm antigen binding.

Example 2.2: Construction of CDR Grafted Anti human TNF-α antibodies

By applying standard methods well known in the art, the CDR sequences of VH and VL chains of monoclonal antibody MAK199 (see Table 5 above) are grafted into different human heavy and light chain acceptor sequences.

Based on sequence VH and VL alignments with the VH and VL sequences of monoclonal antibody MAK199 the following known human sequences are selected:

a) VH1-18 (IGHV1-18) and VH7-4.1 (IGHV7-4-1)and the joining sequences hJH6 for constructing heavy chain acceptor sequences

b) 1-39/O12, and 6-D1/A14 as well as hJK2 for constructing light chain acceptor sequences

By grafting the corresponding VH and VL CDRs of MAK199 into said acceptor sequences, the CDR-grafted, humanized, and modified VH and VL sequences were prepared (see also Table 6, above).

Example 2.3: Construction Of Framework Back Mutations In CDR-Grafted Antibodies

To generate humanized antibody framework back mutations, mutations are introduced into the CDR-grafted antibody sequences as prepared according to Example 2.2, by *de novo* synthesis of the variable domain and/or using mutagenic primers and PCR, and methods well known in the art. Different combinations of back mutations and other mutations are constructed for each of the CDR-grafts as follows. Residue numbers for these mutations are based on the Kabat numbering system.

For heavy chains hMAK199VH.1z, one or more of the following Vernier and VH/VL interfacing residues are back mutated as follows: V2→I, Y91→F.

Additional mutations include the following: Q1→E

For heavy chains hMAK199VH.2z, one or more of the following Vernier and VH/VL interfacing residues are back mutated as follows: V2→I, V67→F, M69→F, T71→L Y91→F.

Additional mutations include the following: Q1→E, R82→S, D85→E.

For light chain hMAK199Vk.1 one or more of the following Vernier and VH/VL interfacing residues are back mutated as follows: A43→T, P44→V, F71→Y, and Y87→F.

For light chain hMAK199Vk.2 one or more of the following Vernier and VH/VL interfacing residues are back mutated as follows: V2→I, A43→T, P44→V, K49→Y, F71→Y, and Y87→F.

Example 2.4: Generation Of Humanized Anti-HTNFα Antibodies Containing Framework Back Mutations In CDR-Grafted Antibodies

The sequences can be used to synthesize nucleic acids using standard DNA synthesis or amplification technologies and assembling the desired antibody fragments into expression vectors, using standard recombinant DNA technology, for expression in cells. For example, nucleic acid codons are determined from amino acids sequences and oligonucleotide DNA is synthesized by Blue Heron Biotechnology, Inc. (www.blueheronbio.com) Bothell, WA USA. The oligonucleotides are assembled into 300-2,000 base pair double-stranded DNA fragments, cloned into a plasmid vector and sequence-verified. Cloned fragments are assembled using an enzymatic process to yield the complete gene and subcloned into an expression vector. (See 7,306,914; 7,297,541; 7,279,159; 7,150,969; 20080115243; 20080102475; 20080081379; 20080075690; 20080063780; 20080050506; 20080038777; 20080022422; 20070289033; 20070287170; 20070254338; 20070243194; 20070225227; 20070207171; 20070150976; 20070135620; 20070128190; 20070104722; 20070092484; 20070037196; 20070028321; 20060172404; 20060162026; 20060153791; 20030215458; and 20030157643).

For example, in silico constructed humanized antibodies described above can be inserted into the multiple cloning site in a pHybE vector (US Patent Publication No. US 2009/0239259). Bacterial colonies are isolated and plasmid DNA extracted; cDNA inserts are sequenced in their entirety. Correct humanized heavy and light chains corresponding to each antibody are co-transfected into HEK 293-6E cells to transiently produce full-length humanized anti-human TNF-α antibodies. pHybE vectors containing the heavy chain grafted cDNA and the light chain grafted cDNA were co-transfected into HEK 293-6E cells. Cell supernatants containing recombinant chimeric antibody are purified by Protein A Sepharose chromatography and bound antibody is eluted by addition of acid buffer. Antibodies are neutralized and dialyzed into PBS. Humanized antibodies are described in Table 7.

Table 7: Humanized Anti-TNFα Antibodies

SEQ ID No.	Protein region	Sequence
		12345678901234567890123456789**0**
37	hMAK199VH.1	EVQLVQSGSELKKPGASVKVSCKASGYTFT **NYGMN**WVRQAPGQGLEWMG**WINTYTGEPTY** **ADDFKG**RFVFSLDTSVSTAYLQISSLKAED TAVYYCAR**KFLTTVVVTDYAMDY**WGQGTTV TVSS
38	hMAK199VH.1a	EIQLVQSGSELKKPGASVKVSCKASGYTFT **NYGMN**WVRQAPGQGLEWMG**WINTYTGEPTY** **ADDFKG**RFVFSLDTSVSTAYLQISSLKAED TAVYFCAR**KFLTTVVVTDYAMDY**WGQGTTV TVSS
39	hMAK199VH.1b	EVQLVQSGSELKKPGASVKVSCKASGYTFT **NYGMN**WVRQAPGQGLEWMG**WINTYTGEPTY** **ADDFKG**RFVFSLDTSVSTAYLQISSLKAED TAVYFCAR**KFLTTVVVTDYAMDY**WGQGTTV TVSS
40	hMAK199VH.2	EVQLVQSGAEVKKPGASVKVSCKASGYTFT **NYGMN**WVRQAPGQGLEWMG**WINTYTGEPTY** **ADDFKG**RVTMTTDTSTSTAYMELSSLRSED TAVYYCAR**KFLTTVVVTDYAMDY**WGQGTTV TVSS
41	hMAK199VH.2a	EIQLVQSGAEVKKPGASVKVSCKASGYTFT **NYGMN**WVRQAPGQGLEWMG**WINTYTGEPTY** **ADDFKG**RFTFTLDTSTSTAYMELSSLRSED TAVYFCAR**KFLTTVVVTDYAMDY**WGQGTTV TVSS
42	hMAK199VH.2b	EIQLVQSGAEVKKPGASVKVSCKASGYTFT **NYGMN**WVRQAPGQGLEWMG**WINTYTGEPTY** **ADDFKG**RVTFTTDTSTSTAYMELSSLRSED TAVYYCAR**KFLTTVVVTDYAMDY**WGQGTTV TVSS
43	hMAK199VL.1a	DIQMTQSPSSLSASVGDRVTITC**RASQDIS** **NYLN**WYQQKPGKTVKLLIY**YTSRLQS**GVPS RFSGSGSGTDYTLTISSLQPEDFATYFC**QQ** **GNTLPPT**FGQGTKLEIK
44	hMAK199VL.1b	DIQMTQSPSSLSASVGDRVTITC**RASQDIS** **NYLN**WYQQKPGKAPKLLIY**YTSRLQS**GVPS RFSGSGSGTDYTLTISSLQPEDFATYYC**QQ** **GNTLPPT**FGQGTKLEIK
45	hMAK199VL.2a	DIVMTQSPAFLSVTPGEKVTITC**RASQDIS** **NYLN**WYQQKPDQTVKLLIY**YTSRLQS**GVPS RFSGSGSGTDYTFTISSLEAEDAATYFC**QQ** **GNTLPPT**FGQGTKLEIK
46	hMAK199VL.2b	DIVMTQSPAFLSVTPGEKVTITC**RASQDIS** **NYLN**WYQQKPDQAPKLLIY**YTSRLQS**GVPS RFSGSGSGTDYTFTISSLEAEDAATYYC**QQ** **GNTLPPT**FGQGTKLEIK

Example 2.5: Humanized Anti-hTNFα hMAK199 Antibody VH/VL Pairings

ABT Unique ID	VH	VL
AB351	hMAK199VH.1	hMAK199VL.1
AB352	hMAK199VH.1	hMAK199VL.1a
AB353	hMAK199VH.1	hMAK199VL.1b
AB354	hMAK199VH.1	hMAK199VL.2
AB355	hMAK199VH.1	hMAK199VL.2a
AB356	hMAK199VH.1	hMAK199VL.2b
AB357	hMAK199VH.1a	hMAK199VL.1
AB358	hMAK199VH.1a	hMAK199VL.1a
AB359	hMAK199VH.1a	hMAK199VL.1b
AB360	hMAK199VH.1a	hMAK199VL.2
AB361	hMAK199VH.1a	hMAK199VL.2a
AB362	hMAK199VH.1a	hMAK199VL.2b
AB363	hMAK199VH.1b	hMAK199VL.1
AB364	hMAK199VH.1b	hMAK199VL.1a
AB365	hMAK199VH.1b	hMAK199VL.1b
AB366	hMAK199VH.1b	hMAK199VL.2
AB367	hMAK199VH.1b	hMAK199VL.2a
AB368	hMAK199VH.1b	hMAK199VL.2b
AB369	hMAK199VH.2	hMAK199VL.1
AB370	hMAK199VH.2	hMAK199VL.1a
AB371	hMAK199VH.2	hMAK199VL.1b
AB372	hMAK199VH.2	hMAK199VL.2
AB373	hMAK199VH.2	hMAK199VL.2a
AB374	hMAK199VH.2	hMAK199VL.2b
AB375	hMAK199VH.2a	hMAK199VL.1
AB376	hMAK199VH.2a	hMAK199VL.1a
AB377	hMAK199VH.2a	hMAK199VL.1b
AB378	hMAK199VH.2a	hMAK199VL.2
AB379	hMAK199VH.2a	hMAK199VL.2a
AB380	hMAK199VH.2a	hMAK199VL.2b
AB381	hMAK199VH.2b	hMAK199VL.1
AB382	hMAK199VH.2b	hMAK199VL.1a
AB383	hMAK199VH.2b	hMAK199VL.1b
AB384	hMAK199VH.2b	hMAK199VL.2
AB385	hMAK199VH.2b	hMAK199VL.2a
AB386	hMAK199VH.2b	hMAK199VL.2b

Example 2.6: Affinity Determination Using BIACORE Technology

Table 8: Reagent Used in Biacore Analyses

Antigen	Vendor Designation	Vendor	Catalog #
TNFα	Recombinant Human TNF-α/TNFSF1A	R&D systems	210-TA

BIACORE Methods:

The BIACORE assay (Biacore, Inc, Piscataway, NJ) determines the affinity of antibodies with kinetic measurements of on-rate and off-rate constants. Binding of antibodies to a target antigen (for example, a purified recombinant target antigen) is determined by surface plasmon resonance-based measurements with a Biacore® 1000 or 3000 instrument (Biacore® AB, Uppsala, Sweden) using running HBS-EP (10 mM HEPES [pH 7.4], 150 mM NaCl, 3 mM EDTA, and 0.005% surfactant P20) at 25° C. All chemicals are obtained from Biacore® AB (Uppsala, Sweden) or otherwise from a different source as described. For example, approximately 5000 RU of goat anti-mouse IgG, (Fcγ), fragment specific polyclonal antibody (Pierce Biotechnology Inc, Rockford, IL) diluted in 10 mM sodium acetate (pH 4.5) is directly immobilized across a CM5 research grade biosensor chip using a standard amine coupling kit according to manufacturer's instructions and procedures at 25 μg/ml. Unreacted moieties on the biosensor surface are blocked with ethanolamine. Modified carboxymethyl dextran surface in flowcell 2 and 4 is used as a reaction surface. Unmodified carboxymethyl dextran without goat anti-mouse IgG in flow cell 1 and 3 is used as the reference surface. For kinetic analysis, rate equations derived from the 1:1 Langmuir binding model are fitted simultaneously to association and dissociation phases of all eight injections (using global fit analysis) with the use of Biaevaluation 4.0.1 software. Purified antibodies are diluted in HEPES-buffered saline for capture across goat anti-mouse IgG specific reaction surfaces. Antibodies to be captured as a ligand (25 μg/ml) are injected over reaction matrices at a flow rate of 5 μl/minute. The association and dissociation rate constants, k_{on} ($M^{-1}s^{-1}$) and k_{off} (s^{-1}), are determined under a continuous flow rate of 25 μl/minute. Rate constants are derived by making kinetic binding measurements at different antigen concentrations ranging from 10 – 200 nM. The equilibrium dissociation constant (M) of the reaction between antibodies and the target antigen is then calculated from the kinetic rate constants by the following formula: $K_D = k_{off}/k_{on}$. Binding is recorded as a function of time and kinetic rate constants are calculated. In this assay, on-rates as fast as $10^6 M^{-1}s^{-1}$ and off-rates as slow as $10^{-6} s^{-1}$ can be measured.

Table 9: BIACORE Analysis of Anti-hTNFα Antibodies

Antibody ID	VH	VL	k_{on} (M-1s-1)	k_{off} (s-1)	k_D (M)
AB351	hMAK199VH.1	hMAK199VL.1	2.00E+06	6.80E-04	3.50E-10
AB352	hMAK199VH.1	hMAK199VL.1a	2.10E+06	9.40E-04	4.50E-10
AB353	hMAK199VH.1	hMAK199VL.1b	2.00E+06	9.20E-04	4.60E-10
AB354	hMAK199VH.1	hMAK199VL.2	2.00E+06	1.00E-03	5.20E-10
AB355	hMAK199VH.1	hMAK199VL.2a	2.40E+06	1.40E-03	5.90E-10
AB356	hMAK199VH.1	hMAK199VL.2b	1.30E+06	8.90E-04	6.70E-10
AB357	hMAK199VH.1a	hMAK199VL.1	1.90E+06	1.30E-03	6.70E-10
AB358	hMAK199VH.1a	hMAK199VL.1a	1.40E+06	9.70E-04	7.00E-10
AB359	hMAK199VH.1a	hMAK199VL.1b	1.30E+06	9.30E-04	7.30E-10
AB360	hMAK199VH.1a	hMAK199VL.2	1.20E+06	9.70E-04	7.80E-10
AB361	hMAK199VH.1a	hMAK199VL.2a	1.20E+06	9.80E-04	7.90E-10
AB362	hMAK199VH.1a	hMAK199VL.2b	1.30E+06	1.10E-03	8.30E-10
AB363	hMAK199VH.1b	hMAK199VL.1	1.40E+06	1.20E-03	8.60E-10
AB364	hMAK199VH.1b	hMAK199VL.1a	1.20E+06	1.10E-03	8.90E-10
AB365	hMAK199VH.1b	hMAK199VL.1b	1.10E+06	1.00E-03	9.10E-10
AB366	hMAK199VH.1b	hMAK199VL.2	1.60E+06	1.50E-03	9.30E-10
AB367	hMAK199VH.1b	hMAK199VL.2a	1.00E+06	1.00E-03	1.00E-09
AB368	hMAK199VH.1b	hMAK199VL.2b	1.10E+06	1.10E-03	1.00E-09
AB369	hMAK199VH.2	hMAK199VL.1	9.40E+05	1.00E-03	1.10E-09
AB370	hMAK199VH.2	hMAK199VL.1a	1.00E+06	1.20E-03	1.10E-09
AB371	hMAK199VH.2	hMAK199VL.1b	1.50E+06	1.70E-03	1.20E-09
AB372	hMAK199VH.2	hMAK199VL.2	9.50E+05	1.10E-03	1.20E-09
AB373	hMAK199VH.2	hMAK199VL.2a	9.40E+05	1.20E-03	1.20E-09
AB374	hMAK199VH.2	hMAK199VL.2b	1.20E+06	1.50E-03	1.30E-09
AB375	hMAK199VH.2a	hMAK199VL.1	9.00E+05	1.10E-03	1.30E-09
AB376	hMAK199VH.2a	hMAK199VL.1a	1.10E+06	1.40E-03	1.30E-09
AB377	hMAK199VH.2a	hMAK199VL.1b	1.10E+06	1.50E-03	1.40E-09
AB378	hMAK199VH.2a	hMAK199VL.2	8.80E+05	1.30E-03	1.40E-09
AB379	hMAK199VH.2a	hMAK199VL.2a	8.90E+05	1.60E-03	1.80E-09
AB380	hMAK199VH.2a	hMAK199VL.2b	8.20E+05	1.50E-03	1.80E-09
AB381	hMAK199VH.2b	hMAK199VL.1	8.60E+05	1.70E-03	1.90E-09
AB382	hMAK199VH.2b	hMAK199VL.1a	7.40E+05	1.50E-03	2.00E-09
AB383	hMAK199VH.2b	hMAK199VL.1b	8.30E+05	1.80E-03	2.10E-09
AB384	hMAK199VH.2b	hMAK199VL.2	6.60E+05	1.40E-03	2.10E-09
AB385	hMAK199VH.2b	hMAK199VL.2a	8.80E+05	1.90E-03	2.20E-09
AB386	hMAK199VH.2b	hMAK199VL.2b	6.60E+05	1.50E-03	2.30E-09

Binding of all humanized constructs characterized by Biacore technology was maintained and comparable to that of the murine parental antibody.

Example 2.7: Neutralization of Human TNFα

L929 cells were grown to a semi-confluent density and harvested using 0.25% tryspin (Gibco#25300). The cells were washed with PBS, counted and resuspended at 1E6 cells/mL in assay media containing 4 μg/mL actinomycin D. The cells were seeded in a 96-well plate (Costar#3599) at a volume of 100 μL and 5E4 cells/well. The anitbodies and control IgG were diluted to a 4x concentration in assay media and serial 1:4 dilutions were performed. The huTNFα was diluted to 400 pg/mL in assay media. Antibody sample (200 μL) was added to the huTNFα (200 μL) in a 1:2 dilution scheme and allowed to incubate for 0.5 hour at room temperature.

The antibody / human TNFα solution was added to the plated cells at 100 μL for a final concentration of 100 pg/mL huTNFα and 150 nM – 0.0001 nM antibody. The plates were incubated for 20 hours at 37ºC, 5 % CO_2. To quantitate viability, 100 μL was removed from the wells and 10 μL of WST-1 reagent (Roche cat# 11644807001) was added. Plates were incubated under assay conditions for 3.5 hours. The plates were read at OD 420-600 nm on a Spectromax 190 ELISA plate reader. An average EC50 from several assays is included in Table 10.

Table 10: Human TNFα Neutralization Assay With Humanized anti-hTNFα Antibodies

Antibody ID	VH	VL	TNFα Neutralization Assay IC50 nM
AB351	hMAK199VH.1	hMAK199VL.1	1.06
AB352	hMAK199VH.1	hMAK199VL.1a	0.35
AB353	hMAK199VH.1	hMAK199VL.1b	0.66
AB354	hMAK199VH.1	hMAK199VL.2	ND
AB355	hMAK199VH.1	hMAK199VL.2a	1.90
AB356	hMAK199VH.1	hMAK199VL.2b	0.87
AB357	hMAK199VH.1a	hMAK199VL.1	1.94
AB358	hMAK199VH.1a	hMAK199VL.1a	0.21
AB359	hMAK199VH.1a	hMAK199VL.1b	3.78
AB360	hMAK199VH.1a	hMAK199VL.2	ND
AB361	hMAK199VH.1a	hMAK199VL.2a	0.82
AB362	hMAK199VH.1a	hMAK199VL.2b	2.11
AB363	hMAK199VH.1b	hMAK199VL.1	0.32
AB364	hMAK199VH.1b	hMAK199VL.1a	0.35
AB365	hMAK199VH.1b	hMAK199VL.1b	1.52
AB366	hMAK199VH.1b	hMAK199VL.2	ND

Antibody ID	VH	VL	TNFα Neutralization Assay IC50 nM
AB367	hMAK199VH.1b	hMAK199VL.2a	0.53
AB368	hMAK199VH.1b	hMAK199VL.2b	1.09
AB369	hMAK199VH.2	hMAK199VL.1	>20
AB370	hMAK199VH.2	hMAK199VL.1a	3.78
AB371	hMAK199VH.2	hMAK199VL.1b	>20
AB372	hMAK199VH.2	hMAK199VL.2	ND
AB373	hMAK199VH.2	hMAK199VL.2a	>20
AB374	hMAK199VH.2	hMAK199VL.2b	>20
AB375	hMAK199VH.2a	hMAK199VL.1	11.45
AB376	hMAK199VH.2a	hMAK199VL.1a	3.65
AB377	hMAK199VH.2a	hMAK199VL.1b	>20
AB378	hMAK199VH.2a	hMAK199VL.2	ND
AB379	hMAK199VH.2a	hMAK199VL.2a	>20
AB380	hMAK199VH.2a	hMAK199VL.2b	>20
AB381	hMAK199VH.2b	hMAK199VL.1	5.90
AB382	hMAK199VH.2b	hMAK199VL.1a	1.19
AB383	hMAK199VH.2b	hMAK199VL.1b	>20
AB384	hMAK199VH.2b	hMAK199VL.2	ND
AB385	hMAK199VH.2b	hMAK199VL.2a	9.76
AB386	hMAK199VH.2b	hMAK199VL.2b	>20

All anti-hTNFα antibodies showed neutralization in the TNFα neutralization assay.

Example 2.8: Physicochemical And In Vitro Stability Analysis Of Humanized Monoclonal Antibodies

Size Exclusion Chromatography

Antibodies ewre diluted to 2.5 mg/mL with water and 20 mL analyzed on a Shimadzu HPLC system using a TSK gel G3000 SWXL column (Tosoh Bioscience, cat# k5539-05k). Samples were eluted from the column with 211 mM sodium sulfate, 92 mM sodium phosphate, pH 7.0, at a flow rate of 0.3 mL/minutes. The HPLC system operating conditions were the following:

Mobile phase: 211 mM Na_2SO_4, 92 mM $Na_2HPO_4*7H_2O$, pH 7.0

Gradient: Isocratic

Flow rate: 0.3 mL/minute

Detector wavelength: 280 nm

Autosampler cooler temp:4°C

Column oven temperature: Ambient

Run time: 50 minutes

Table 11 contains purity data of antibody constructs expressed as percent monomer (unaggregated protein of the expected molecular weight) as determined by the above protocol.

Table 11: Purity of anti-hTNFα Antibodies as Determined by Size Exclusion Chromatography

Antibody ID	VH	VL	% Monomer (purity)
AB351	hMAK199VH.1	hMAK199VL.1	97
AB352	hMAK199VH.1	hMAK199VL.1a	97
AB353	hMAK199VH.1	hMAK199VL.1b	96
AB354	hMAK199VH.1	hMAK199VL.2	97
AB355	hMAK199VH.1	hMAK199VL.2a	98
AB356	hMAK199VH.1	hMAK199VL.2b	96
AB357	hMAK199VH.1a	hMAK199VL.1	96
AB358	hMAK199VH.1a	hMAK199VL.1a	97
AB359	hMAK199VH.1a	hMAK199VL.1b	95
AB360	hMAK199VH.1a	hMAK199VL.2	96
AB361	hMAK199VH.1a	hMAK199VL.2a	96
AB362	hMAK199VH.1a	hMAK199VL.2b	97
AB363	hMAK199VH.1b	hMAK199VL.1	98
AB364	hMAK199VH.1b	hMAK199VL.1a	100
AB365	hMAK199VH.1b	hMAK199VL.1b	100
AB366	hMAK199VH.1b	hMAK199VL.2	99
AB367	hMAK199VH.1b	hMAK199VL.2a	100
AB368	hMAK199VH.1b	hMAK199VL.2b	98
AB369	hMAK199VH.2	hMAK199VL.1	95
AB370	hMAK199VH.2	hMAK199VL.1a	94
AB371	hMAK199VH.2	hMAK199VL.1b	95
AB372	hMAK199VH.2	hMAK199VL.2	88
AB373	hMAK199VH.2	hMAK199VL.2a	88
AB374	hMAK199VH.2	hMAK199VL.2b	91
AB375	hMAK199VH.2a	hMAK199VL.1	97
AB376	hMAK199VH.2a	hMAK199VL.1a	98
AB377	hMAK199VH.2a	hMAK199VL.1b	97
AB378	hMAK199VH.2a	hMAK199VL.2	92
AB379	hMAK199VH.2a	hMAK199VL.2a	98
AB380	hMAK199VH.2a	hMAK199VL.2b	99
AB381	hMAK199VH.2b	hMAK199VL.1	85
AB382	hMAK199VH.2b	hMAK199VL.1a	81
AB383	hMAK199VH.2b	hMAK199VL.1b	83
AB384	hMAK199VH.2b	hMAK199VL.2	87
AB385	hMAK199VH.2b	hMAK199VL.2a	83

AB386	hMAK199VH.2b	hMAK199VL.2b	84

Anti-hTNFα antibodies showed an excellent SEC profile with most showing >95% monomer.

SDS-PAGE

Antibodies are analyzed by sodium dodecyl sulfate - polyacrylamide gel electrophoresis (SDS-PAGE) under both reducing and non-reducing conditions. Adalimumab lot AFP04C is used as a control. For reducing conditions, the samples are mixed 1:1 with 2X tris glycine SDS-PAGE sample buffer (Invitrogen, cat# LC2676, lot# 1323208) with 100 mM DTT, and heated at 60°C for 30 minutes. For non-reducing conditions, the samples are mixed 1:1 with sample buffer and heated at 100°C for 5 minutes. The reduced samples (10 mg per lane) are loaded on a 12% pre-cast tris-glycine gel (Invitrogen, cat# EC6005box, lot# 6111021), and the non-reduced samples (10 mg per lane) are loaded on an 8%-16% pre-cast tris-glycine gel (Invitrogen, cat# EC6045box, lot# 6111021). SeeBlue Plus 2 (Invitrogen, cat#LC5925, lot# 1351542) is used as a molecular weight marker. The gels are run in a XCell SureLock mini cell gel box (Invitrogen, cat# EI0001) and the proteins are separated by first applying a voltage of 75 to stack the samples in the gel, followed by a constant voltage of 125 until the dye front reached the bottom of the gel. The running buffer used is 1X tris glycine SDS buffer, prepared from a 10X tris glycine SDS buffer (ABC, MPS-79-080106)). The gels are stained overnight with colloidal blue stain (Invitrogen cat# 46-7015, 46-7016) and destained with Milli-Q water until the background is clear. The stained gels are then scanned using an Epson Expression scanner (model 1680, S/N DASX003641).

Sedimentation Velocity Analysis

Antibodies are loaded into the sample chamber of each of three standard two-sector carbon epon centerpieces. These centerpieces have a 1.2 cm optical path length and are built with sapphire windows. PBS is used for a reference buffer and each chamber contained 140 μL. All samples are examined simultaneously using a 4-hole (AN-60Ti) rotor in a Beckman ProteomeLab XL-I analytical ultracentrifuge (serial # PL106C01).

Run conditions are programmed and centrifuge control is performed using ProteomeLab (v5.6). The samples and rotor are allowed to thermally equilibrate for one hour prior to analysis (20.0 ± 0.1 °C). Confirmation of proper cell loading is performed at 3000 rpm and a single scan is recorded for each cell. The sedimentation velocity conditions are the following:

Sample Cell Volume: 420 mL

Reference Cell Volume: 420 mL

Temperature: 20°C

Rotor Speed: 35,000 rpm

Time: 8:00 hours

UV Wavelength: 280 nm

Radial Step Size: 0.003 cm

Data Collection: One data point per step without signal averaging.

Total Number of Scans: 100

LC-MS Molecular Weight Measurement Of Intact Antibodies

Molecular weights of intact antibodies are analyzed by LC-MS. Each antibody is diluted to approximately 1 mg/mL with water. An 1100 HPLC (Agilent) system with a protein microtrap (Michrom Bioresources, Inc, cat# 004/25109/03) is used to desalt and introduce 5 mg of the sample into an API Qstar pulsar i mass spectrometer (Applied Biosystems). A short gradient is used to elute the samples. The gradient is run with mobile phase A (0.08% FA, 0.02% TFA in HPLC water) and mobile phase B (0.08% FA and 0.02% TFA in acetonitrile) at a flow rate of 50 mL/minute. The mass spectrometer is operated at 4.5 kvolts spray voltage with a scan range from 2000 to 3500 mass to charge ratio.

LC-MS Molecular Weight Measurement Of Antibody Light And Heavy Chains

Molecular weight measurement of antibody light chains (LC), heavy chains (HC) and deglycosylated HC are analyzed by LC-MS. Aantibody is diluted to 1 mg/mL with water and the sample is reduced to LC and HC with a final concentration of 10 mM DTT for 30 minutes at 37°C. To deglycosylate the antibody, 100 mg of the antibody is incubated with 2 mL of PNGase F, 5 mL of 10% N-octylglucoside in a total volume of 100 mL overnight at 37 °C. After deglycosylation the sample is reduced with a final concentration of 10 mM DTT for 30 minutes at 37°C. An Agilent 1100 HPLC system with a C4 column (Vydac, cat# 214TP5115, S/N 060206537204069) is used to desalt and introduce the sample (5 mg) into an API Qstar pulsar i mass spectrometer (Applied Biosystems). A short gradient is used to elute the sample. The gradient is run with mobile phase A (0.08% FA, 0.02% TFA in HPLC water) and mobile phase B (0.08% FA and 0.02% TFA in acetonitrile) at a flow rate of 50 mL/minute. The mass spectrometer is operated at 4.5 kvolts spray voltage with a scan range from 800 to 3500 mass to charge ratio.

Peptide Mapping

Antibody is denatured for 15 minutes at room temperature with a final concentration of 6 M guanidine hydrochloride in 75 mM ammonium bicarbonate. The denatured samples are reduced with a final concentration of 10 mM DTT at 37°C for 60 minutes, followed by alkylation with 50 mM iodoacetic acid (IAA) in the dark at 37°C for 30 minutes. Following alkylation, the sample is dialyzed overnight against four liters of 10 mM ammonium bicarbonate at 4°C. The

dialyzed sample is diluted to 1 mg/mL with 10 mM ammonium bicarbonate, pH 7.8 and 100 mg of antibody is either digested with trypsin (Promega, cat# V5111) or Lys-C (Roche, cat# 11 047 825 001) at a 1:20 (w/w) trypsin/Lys-C:antibody ratio at 37°C for 4 hrs. Digests are quenched with 1 mL of 1 N HCl. For peptide mapping with mass spectrometer detection, 40 mL of the digests are separated by reverse phase high performance liquid chromatography (RPHPLC) on a C18 column (Vydac, cat# 218TP51, S/N NE9606 10.3.5) with an Agilent 1100 HPLC system. The peptide separation is run with a gradient using mobile phase A (0.02% TFA and 0.08% FA in HPLC grade water) and mobile phase B (0.02% TFA and 0.08% FA in acetonitrile) at a flow rate of 50 mL/minutes. The API QSTAR Pulsar i mass spectromer is operated in positive mode at 4.5 kvolts spray voltage and a scan range from 800 to 2500 mass to charge ratio.

Disulfide Bond Mapping

To denature the antibody, 100 mL of the antibody is mixed with 300 mL of 8 M guanidine HCl in 100 mM ammonium bicarbonate. The pH is checked to ensure that it is between 7 and 8 and the samples are denatured for 15 minutes at room temperature in a final concentration of 6 M guanidine HCl. A portion of the denatured sample (100 mL) is diluted to 600 mL with Milli-Q water to give a final guanidine-HCl concentration of 1 M. The sample (220 mg) is digested with either trypsin (Promega, cat # V5111, lot# 22265901) or Lys-C (Roche, cat# 11047825001, lot# 12808000) at a 1:50 trypsin or 1:50 Lys-C: antibody (w/w) ratios (4.4 mg enzyme: 220 mg sample) at 37°C for approximately 16 hours. An additional 5 mg of trypsin or Lys-C is added to the samples and digestion is allowed to proceed for an additional 2 hours at 37°C. Digestions are stopped by adding 1 mL of TFA to each sample. Digested samples are separated by RPHPLC using a C18 column (Vydac, cat# 218TP51 S/N NE020630-4-1A) on an Agilent HPLC system. The separation is run with the same gradient used for peptide mapping using mobile phase A (0.02% TFA and 0.08% FA in HPLC grade water) and mobile phase B (0.02% TFA and 0.08% FA in acetonitrile) at a flow rate of 50 mL/minute. The HPLC operating conditions are the same as those used for peptide mapping. The API QSTAR Pulsar i mass spectromer is operated in positive mode at 4.5 kvolts spray voltage and a scan range from 800 to 2500 mass-to-charge ratio. Disulfide bonds are assigned by matching the observed MWs of peptides with the predicted MWs of tryptic or Lys-C peptides linked by disulfide bonds.

Free sulfhydryl determination

The method used to quantify free cysteines in an antibody is based on the reaction of Ellman's reagent, 5,5¢- dithio-bis (2-nitrobenzoic acid) (DTNB), with sulfhydryl groups (SH) which gives rise to a characteristic chromophoric product, 5-thio-(2-nitrobenzoic acid) (TNB). The reaction is illustrated in the formula:

DTNB + RSH ® RS-TNB + TNB- + H+

The absorbance of the TNB- is measured at 412 nm using a Cary 50 spectrophotometer. An absorbance curve is plotted using dilutions of 2 mercaptoethanol (b-ME) as the free SH standard and the concentrations of the free sulfhydryl groups in the protein are determined from absorbance at 412 nm of the sample.

The b-ME standard stock is prepared by a serial dilution of 14.2 M b-ME with HPLC grade water to a final concentration of 0.142 mM. Then standards in triplicate for each concentration are prepared. Antibody is concentrated to 10 mg/mL using an amicon ultra 10,000 MWCO centrifugal filter (Millipore, cat# UFC801096, lot# L3KN5251) and the buffer is changed to the formulation buffer used for adalimumab (5.57 mM sodium phosphate monobasic, 8.69 mM sodium phosphate dibasic, 106.69 mM NaCl, 1.07 mM sodium citrate, 6.45 mM citric acid, 66.68 mM mannitol, pH 5.2, 0.1% (w/v) Tween). The samples are mixed on a shaker at room temperature for 20 minutes. Then 180 mL of 100 mM Tris buffer, pH 8.1 is added to each sample and standard followed by the addition of 300 mL of 2 mM DTNB in 10 mM phosphate buffer, pH 8.1. After thorough mixing, the samples and standards are measured for absorption at 412 nm on a Cary 50 spectrophotometer. The standard curve is obtained by plotting the amount of free SH and OD_{412} nm of the b-ME standards. Free SH content of samples are calculated based on this curve after subtraction of the blank.

Weak Cation Exchange Chromatography

Antibody is diluted to 1 mg/mL with 10 mM sodium phosphate, pH 6.0. Charge heterogeneity is analyzed using a Shimadzu HPLC system with a WCX-10 ProPac analytical column (Dionex, cat# 054993, S/N 02722). The samples are loaded on the column in 80% mobile phase A (10 mM sodium phosphate, pH 6.0) and 20% mobile phase B (10 mM sodium phosphate, 500 mM NaCl, pH 6.0) and eluted at a flow rate of 1.0 mL/minute.

Oligosaccharide Profiling

Oligosaccharides released after PNGase F treatment of antibody are derivatized with 2-aminobenzamide (2-AB) labeling reagent. The fluorescent-labeled oligosaccharides are separated by normal phase high performance liquid chromatography (NPHPLC) and the different forms of oligosaccharides are characterized based on retention time comparison with known standards.

The antibody is first digested with PNGaseF to cleave N-linked oligosaccharides from the Fc portion of the heavy chain. The antibody (200 mg) is placed in a 500 mL Eppendorf tube along with 2 mL PNGase F and 3 mL of 10% N-octylglucoside. Phosphate buffered saline is added to bring the final volume to 60 mL. The sample is incubated overnight at 37°C in an Eppendorf

thermomixer set at 700 RPM. Adalimumab lot AFP04C is also digested with PNGase F as a control.

After PNGase F treatment, the samples are incubated at 95°C for 5 minutes in an Eppendorf thermomixer set at 750 RPM to precipitate out the proteins, then the samples are placed in an Eppendorf centrifuge for 2 minutes at 10,000 RPM to spin down the precipitated proteins. The supernatent containing the oligosaccharides are transferred to a 500 mL Eppendorf tube and dried in a speed-vac at 65°C.

The oligosaccharides are labeled with 2AB using a 2AB labeling kit purchased from Prozyme (cat# GKK-404, lot# 132026). The labeling reagent is prepared according to the manufacturer's instructions. Acetic acid (150 mL, provided in kit) is added to the DMSO vial (provided in kit) and mixed by pipeting the solution up and down several times. The acetic acid/DMSO mixture (100 mL) is transferred to a vial of 2-AB dye (just prior to use) and mixed until the dye is fully dissolved. The dye solution is then added to a vial of reductant (provided in kit) and mixed well (labeling reagent). The labeling reagent (5 mL) is added to each dried oligosaccharide sample vial, and mixed thoroughly. The reaction vials are placed in an Eppendorf thermomixer set at 65°C and 700-800 RPM for 2 hours of reaction.

After the labeling reaction, the excess fluorescent dye is removed using GlycoClean S Cartridges from Prozyme (cat# GKI-4726). Prior to adding the samples, the cartridges are washed with 1 mL of milli-Q water followed with 5 ishes of 1 mL 30% acetic acid solution. Just prior to adding the samples, 1 mL of acetonitrile (Burdick and Jackson, cat# AH015-4) is added to the cartridges.

After all of the acetonitrile passed through the cartridge, the sample is spotted onto the center of the freshly washed disc and allowed to adsorb onto the disc for 10 minutes. The disc is washed with 1 mL of acetonitrile followed by five ishes of 1 mL of 96% acetonitrile. The cartridges are placed over a 1.5 mL Eppendorf tube and the 2-AB labeled oligosaccharides are eluted with 3 ishes (400 mL each ish) of milli Q water.

The oligosaccharides are separated using a Glycosep N HPLC (cat# GKI-4728) column connected to a Shimadzu HPLC system. The Shimadzu HPLC system consisted of a system controller, degasser, binary pumps, autosampler with a sample cooler, and a fluorescent detector.

Stability at Elevated Temperatures

The final concentration of the antibodies is adjusted to 2 mg/mL with the appropriate buffers, surfactants, stabilizers, and/or sugars. The antibody solutions are then filter sterized and

0.25 mL aliquots are prepared under sterile conditions. The aliquots are left at either -80°C, 5°C, 25°C, or 40°C for 1, 2 or 3 weeks. At the end of the incubation period, the samples are analyzed by size exclusion chromatography and SDS-PAGE.

The stability samples are analyzed by SDS-PAGE under both reducing and non-reducing conditions. The procedure used is the same as described herein. The gels are stained overnight with colloidal blue stain (Invitrogen cat# 46-7015, 46-7016) and destained with Milli-Q water until the background is clear. The stained gels are then scanned using an Epson Expression scanner (model 1680, S/N DASX003641). To obtain more sensitivity, the same gels are silver stained using silver staining kit (Owl Scientific) and the recommended procedures given by the manufacturer is used.

Example 2.9: Transfection And Expression In HEK 293-6E Cells

The anti-hTNFα antibody vector constructs were tranfected into 293 cells for production of protein. The 293 transient transfection procedure used is a modification of the methods published in Durocher et al. (2002) Nucleic Acids Res. 30(2):E9 and Pham et al. (2005) Biotech. Bioengineering 90(3):332-44. Reagents that were used in the transfection included:

- HEK 293-6E cells (human embryonic kidney cell line stably expressing EBNA1; obtained from National Research Council Canada) cultured in disposable Erlenmeyer flasks in a humidified incubator set at 130 rpm, 37°C and 5% CO_2.

- Culture medium: FreeStyle 293 Expression Medium (Invitrogen 12338-018) plus 25 μg/mL Geneticin (G418) (Invitrogen 10131-027) and 0.1% Pluronic F-68 (Invitrogen 24040-032).

- Transfection medium: FreeStyle 293 Expression Medium plus 10 mM HEPES (Invitrogen 15630-080).

- Polyethylenimine (PEI) stock: 1 mg/mL sterile stock solution, pH 7.0, prepared with linear 25kDa PEI (Polysciences) and stored at less than -15°C.

- Tryptone Feed Medium: 5% w/v sterile stock of Tryptone N1 (Organotechnie, 19554) in FreeStyle 293 Expression Medium.

Cell preparation for transfection: Approximately 2 - 4 hours prior to transfection, HEK 293-6E cells were harvested by centrifugation and resuspended in culture medium at a cell density of

approximately 1 million viable cells per mL. For each transfection, 40 mL of the cell suspension was transferred into a disposable 250-mL Erlenmeyer flask and incubated for 2 - 4 hours.

Transfection: The transfection medium and PEI stock were prewarmed to room temperature (RT). For each transfection, 25µg of plasmid DNA and 50µg of polyethylenimine (PEI) were combined in 5 mL of transfection medium and incubated for 15 – 20 minutes at RT to allow the DNA:PEI complexes to form. For the BR3-Ig transfections, 25µg of BR3-Ig plasmid was used per transfection. Each 5-mL DNA:PEI complex mixture was added to a 40-mL culture prepared previously and returned to the humidified incubator set at 130 rpm, 37°C and 5% CO_2. After 20-28 hours, 5 mL of Tryptone Feed Medium was added to each transfection and the cultures were continued for six days.

Table 12 contains the yield data for parent antibodies expressed as milligrams per liter in HEK 293-6E cells.

Table 12: Transient Expression in Yields of anti-hTNFα Antibodies in HEK 293-6E Cells

Antibody ID	VH	VL	Expression yield (mg/L)
AB351	hMAK199VH.1	hMAK199VL.1	67
AB352	hMAK199VH.1	hMAK199VL.1a	56
AB353	hMAK199VH.1	hMAK199VL.1b	87
AB354	hMAK199VH.1	hMAK199VL.2	81
AB355	hMAK199VH.1	hMAK199VL.2a	54
AB356	hMAK199VH.1	hMAK199VL.2b	22
AB357	hMAK199VH.1a	hMAK199VL.1	78
AB358	hMAK199VH.1a	hMAK199VL.1a	63
AB359	hMAK199VH.1a	hMAK199VL.1b	92
AB360	hMAK199VH.1a	hMAK199VL.2	92
AB361	hMAK199VH.1a	hMAK199VL.2a	52
AB362	hMAK199VH.1a	hMAK199VL.2b	30
AB363	hMAK199VH.1b	hMAK199VL.1	27
AB364	hMAK199VH.1b	hMAK199VL.1a	2
AB365	hMAK199VH.1b	hMAK199VL.1b	4
AB366	hMAK199VH.1b	hMAK199VL.2	4
AB367	hMAK199VH.1b	hMAK199VL.2a	3
AB368	hMAK199VH.1b	hMAK199VL.2b	3
AB369	hMAK199VH.2	hMAK199VL.1	28
AB370	hMAK199VH.2	hMAK199VL.1a	20
AB371	hMAK199VH.2	hMAK199VL.1b	31
AB372	hMAK199VH.2	hMAK199VL.2	107
AB373	hMAK199VH.2	hMAK199VL.2a	73
AB374	hMAK199VH.2	hMAK199VL.2b	59

Antibody ID	VH	VL	Expression yield (mg/L)
AB375	hMAK199VH.2a	hMAK199VL.1	105
AB376	hMAK199VH.2a	hMAK199VL.1a	83
AB377	hMAK199VH.2a	hMAK199VL.1b	106
AB378	hMAK199VH.2a	hMAK199VL.2	120
AB379	hMAK199VH.2a	hMAK199VL.2a	75
AB380	hMAK199VH.2a	hMAK199VL.2b	10
AB381	hMAK199VH.2b	hMAK199VL.1	48
AB382	hMAK199VH.2b	hMAK199VL.1a	55
AB383	hMAK199VH.2b	hMAK199VL.1b	70
AB384	hMAK199VH.2b	hMAK199VL.2	74
AB385	hMAK199VH.2b	hMAK199VL.2a	75
AB386	hMAK199VH.2b	hMAK199VL.2b	42

All antibodies expressed well in HEK 293-6E cells. In most cases >50 mg/L purified antibody could be obtained easily from supernatants of HEK 293-6E cells.

The present disclosure incorporates by reference in their entirety techniques well known in the field of molecular biology. These techniques include, but are not limited to, techniques described in the following publications:

Ausubel et al. eds., Short Protocols In Molecular Biology (4th Ed. 1999) John Wiley & Sons, NY (ISBN 0-471-32938-X).

Lu and Weiner eds., Cloning and Expression Vectors for Gene Function Analysis (2001) BioTechniques Press, Westborough, MA, 298 pp. (ISBN 1-881299-21-X).

Kontermann and Dübel eds., Antibody Engineering (2001) Springer-Verlag, NY, 790 pp. (ISBN 3-540-41354-5).

Old and Primrose, Principles of Gene Manipulation: An Introduction To Genetic Engineering (3d Ed. 1985) Blackwell Scientific Publications, Boston, MA. Studies in Microbiology; V.2:409 pp. (ISBN 0-632-01318-4).

Sambrook et al., Molecular Cloning: A Laboratory Manual, 2d ed. (Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY), Vols. 1-3 (ISBN 0-87969-309-6).

Winnacker, From Genes To Clones: Introduction To Gene Technology (1987) VCH Publishers, NY (translated by Horst Ibelgaufts), 634 pp. (ISBN 0-89573-614-4).

Incorporation by Reference

The contents of all cited references (including literature references, patents, patent applications, and websites) that maybe cited throughout this application are hereby expressly incorporated by reference in their entirety for any purpose, as are the references cited therein. The practice of the present embodiments will employ, unless otherwise indicated, conventional techniques of immunology, molecular biology and cell biology, which are well known in the art.

Equivalents

The embodiments may be carried out in other specific forms without departing from the spirit or essential characteristics thereof. The foregoing embodiments are therefore to be considered in all respects illustrative rather than limiting. Scope is thus indicated by the appended claims rather than by the foregoing description, and all changes that come within the meaning and range of equivalency of the claims are therefore intended to be embraced herein.

We claim:

1. An antibody, or antigen binding portion thereof comprising SEQ ID NO: 31, SEQ ID NO: 32, SEQ ID NO: 33, SEQ ID NO: 34, SEQ ID NO: 35, SEQ ID NO: 36, SEQ ID NO: 37, SEQ ID NO: 38, SEQ ID NO: 39; SEQ ID NO: 40; SEQ ID NO: 41; SEQ ID NO: 42; SEQ ID NO: 43; SEQ ID NO: 44; SEQ ID NO: 45; or SEQ ID NO: 46.

2. An antibody, or antigen binding portion thereof, capable of binding TNF-α comprising at least one CDR comprising residues 31-35 of SEQ ID NO:31 (CDR-H1), residues 50-66 of SEQ ID NO:31 (CDR-H2), residues 99-113 of SEQ ID NO:31 (CDR-H3), residues 24-34 of SEQ ID NO:32 (CDR-L1), residues 50-56 of SEQ ID NO:32 (CDR-L2), or residues 89-97 of SEQ ID NO:32 (CDR-L3).

3. The binding protein according to claim 2, wherein the binding protein comprises at least three CDRs.

4. The binding protein according to claim 3, wherein the at least three CDRs comprise a variable domain CDR set of:
(a) residues 31-35 of SEQ ID NO:31 (CDR-H1), residues 50-66 of SEQ ID NO:31 (CDR-H2), and residues 99-113 of SEQ ID NO:31 (CDR-H3); or
(b) residues 24-34 of SEQ ID NO:32 (CDR-L1), residues 50-56 of SEQ ID NO:32 (CDR-L2), and residues 89-97 of SEQ ID NO:32 (CDR-L3).

5. The binding protein according to claim 4 comprising at least two variable domain CDR sets.

6. The binding protein according to claim 5, wherein the binding protein comprises both of:
(a) residues 31-35 of SEQ ID NO:31 (CDR-H1), residues 50-66 of SEQ ID NO:31 (CDR-H2), and residues 99-113 of SEQ ID NO:31 (CDR-H3); and
(b) residues 24-34 of SEQ ID NO:32 (CDR-L1), residues 50-56 of SEQ ID NO:32 (CDR-L2), and residues 89-97 of SEQ ID NO:32 (CDR-L3).

7. The binding protein according to claim 6, further comprising a human acceptor framework.

8. The binding protein according to claim 7, wherein the human acceptor framework comprises any one of SEQ ID NOS:10-19 or SEQ ID NOS:20-30.

9. The binding protein according to claim 7 or 8, wherein said human acceptor framework comprises at least one framework region amino acid substitution, wherein the amino acid sequence of the framework is at least 65% identifical to the sequence of the human acceptor framework and comprises at least 70 amino acid residues identical to the human acceptor framework.

10. The binding protein according to claim 8, wherein the human acceptor framework, comprises at least one framework region amino acid substitution at a residue adjacent to a CDR, a glycosylation site residue, a rare residue, a residue capable of interacting with human TNF-α, a residue capable of interacting with a CDR, a canonical residue, a contact residue between heavy chain variable region and light chain variable region, a residue within a Vernier zone, and a residue in a region that overlaps between a Chothia-defined variable heavy chain CDR1 and a Kabat-defined first heavy chain framework.

11. The binding protein according to claim 10, wherein the residue is 1H, 2H, 67H, 69H, 71H, 82H, 85H, 91H, and 2L, 43L, 44L, 49L, 71L, or 87L.

12. The binding protein according to claim 11, wherein the binding protein comprises a consensus human acceptor.

13. The binding protein according to claim 1, comprising
(a) a variable heavy chain polypeptide comprising SEQ ID NO: 33, SEQ ID NO: 34, SEQ ID NO: 37, SEQ ID NO: 38, SEQ ID NO: 39, SEQ ID NO: 40, SEQ ID NO: 41, and SEQ ID NO: 42, and
(b) a variable light chain polypeptide comprising SEQ ID NO: 35, SEQ ID NO: 36, SEQ ID NO: 43, SEQ ID NO: 44, SEQ ID NO: 45, and SEQ ID NO: 46.

14. The binding protein of claim 13, wherein said binding protein comprises a variable heavy chain polypeptide and a variable light chain polypeptide comprising the respective amino acid sequences:

SEQ ID NO: 31 and SEQ ID NO: 32;

SEQ ID NO: 33 and SEQ ID NO: 35;
SEQ ID NO: 33 and SEQ ID NO: 43;
SEQ ID NO: 33 and SEQ ID NO: 44;
SEQ ID NO: 33 and SEQ ID NO: 36;

SEQ ID NO: 33 and SEQ ID NO: 45;
SEQ ID NO: 33 and SEQ ID NO: 46;

SEQ ID NO: 34 and SEQ ID NO: 35;
SEQ ID NO: 34 and SEQ ID NO: 43;
SEQ ID NO: 34 and SEQ ID NO: 44;
SEQ ID NO: 34 and SEQ ID NO: 36;
SEQ ID NO: 34 and SEQ ID NO: 45;
SEQ ID NO: 34 and SEQ ID NO: 46;

SEQ ID NO: 37 and SEQ ID NO: 35;
SEQ ID NO: 37 and SEQ ID NO: 43;
SEQ ID NO: 37 and SEQ ID NO: 44;
SEQ ID NO: 37 and SEQ ID NO: 36;
SEQ ID NO: 37 and SEQ ID NO: 45;
SEQ ID NO: 37 and SEQ ID NO: 46;

SEQ ID NO: 38 and SEQ ID NO: 35,
SEQ ID NO: 38 and SEQ ID NO: 43;
SEQ ID NO: 38 and SEQ ID NO: 44;
SEQ ID NO: 38 and SEQ ID NO: 36;
SEQ ID NO: 38 and SEQ ID NO: 45;
SEQ ID NO: 38 and SEQ ID NO: 46;

SEQ ID NO: 39 and SEQ ID NO: 35,
SEQ ID NO: 39 and SEQ ID NO: 43;
SEQ ID NO: 39 and SEQ ID NO: 44;
SEQ ID NO: 39 and SEQ ID NO: 36;
SEQ ID NO: 39 and SEQ ID NO: 45;
SEQ ID NO: 39 and SEQ ID NO: 46;

SEQ ID NO: 40 and SEQ ID NO: 35,
SEQ ID NO: 40 and SEQ ID NO: 43;
SEQ ID NO: 40 and SEQ ID NO: 44;
SEQ ID NO: 40 and SEQ ID NO: 36;
SEQ ID NO: 40 and SEQ ID NO: 45;

SEQ ID NO: 40 and SEQ ID NO: 46;

SEQ ID NO: 41 and SEQ ID NO: 35;
SEQ ID NO: 41 and SEQ ID NO: 43;
SEQ ID NO: 41 and SEQ ID NO: 44;
SEQ ID NO: 41 and SEQ ID NO: 36;
SEQ ID NO: 41 and SEQ ID NO: 45;
SEQ ID NO: 41 and SEQ ID NO: 46;

SEQ ID NO: 42 and SEQ ID NO: 35,
SEQ ID NO: 42 and SEQ ID NO: 43;
SEQ ID NO: 42 and SEQ ID NO: 44;
SEQ ID NO: 42 and SEQ ID NO: 36;
SEQ ID NO: 42 and SEQ ID NO: 45; or
SEQ ID NO: 42 and SEQ ID NO: 46.

15. The binding protein of claim 1, wherein said binding protein is: an immunoglobulin molecule, a disulfide linked Fv, a monoclonal antibody, an scFv, a chimeric antibody, a single domain antibody, a CDR-grafted antibody, a diabody, a humanized antibody, a multispecific antibody, an Fab, a dual specific antibody, a DVD-Ig protein, a Fab', a bispecific antibody, a F(ab')2, or a Fv.

16. The binding protein of claim 1, wherein said binding protein comprises a human IgM constant domain, a human IgG4 constant domain, a human IgG1 constant domain, a human IgE constant domain, a human IgG2 constant domain, a human IgG3 constant domain, or a human IgA constant domain.

17. The binding protein of claim 1, further comprising a heavy chain constant region having an amino acid sequence of SEQ ID NO: 2 or SEQ ID NO: 3.

18. The binding protein of claim 1, further comprising a light chain constant region having an amino acid sequence of SEQ ID NO: 4 or SEQ ID NO: 5.

19. The binding protein of claim 1, wherein said binding protein is capable of neutralizing human TNF-α.

20. The binding protein of claim 1, wherein said binding protein has an on rate constant (K_{on}) to said target of at least about $10^2M^{-1}s^{-1}$; at least about $10^3M^{-1}s^{-1}$; at least about $10^4M^{-1}s^{-1}$; at least about $10^5M^{-1}s^{-1}$; or at least about $106M^{-1}s^{-1}$ as measured by surface plasmon resonance.

21. The binding protein of claim 1, wherein said binding protein has an off rate constant (K_{off}) to said target of at most about $10^{-3}s^{-1}$; at most about $10^{-4}s^{-1}$; at most about $10^{-5}s^{-1}$; or at most about $10^{-6}s^{-1}$, as measured by surface plasmon resonance.

22. The binding protein of claim 1, wherein said binding protein has a dissociation constant (KD) to said target of at most about 10^{-7} M; at most about 10^{-8} M; at most about 10^{-9} M; at most about 10^{-10} M; at most about 10^{-11} M; at most about 10^{-12} M; or at most 10^{-13} M.

23. The binding protein of claim 22, where said binding protein has a dissociation constant (KD) to TNF-α of about at most about 10^{-7} M; at most about 10^{-8} M; at most about 10^{-9} M; at most about 10^{-10} M; at most about 10^{-11} M; at most about 10^{-12} M; or at most 10^{-13} M.

24. The binding protein of claim 1, wherein said binding protein further comprises an immunoadhesion molecule, an imaging agent, a therapeutic agent, or a cytotoxic agent.

25. The binding protein of claim 24, wherein said imaging agent is a radiolabel, an enzyme, a fluorescent label, a luminescent label, a bioluminescent label, a magnetic label, or biotin.

26. The binding protein of claim 25, wherein said radiolabel is: ^{3}H, ^{14}C, ^{35}S, ^{90}Y, ^{99}Tc, ^{111}In, ^{125}I, ^{131}I, ^{177}Lu, ^{166}Ho, or ^{153}Sm.

27. The binding protein of claim 24, wherein said therapeutic or cytotoxic agent is an anti-metabolite, an alkylating agent, an antibiotic, a growth factor, a cytokine, an anti-angiogenic agent, an anti-mitotic agent, an anthracycline, toxin, and an apoptotic agent.

28. The binding protein of claim 1, wherein said binding protein possesses a human glycosylation pattern.

29. The binding protein of claim 1, wherein said binding protein is a crystallized binding protein.

30. An isolated nucleic acid encoding a binding protein amino acid sequence of claim 1.

31. A vector comprising an isolated nucleic acid encoding a binding protein amino acid sequence of claim 1.

32. The vector of claim 31, wherein said vector is pcDNA, pTT, pTT3, pEFBOS, pBV, pJV, pHybE, or pBJ.

32. A host cell comprising the vector of claim 31.

33. The host cell of claim 32, wherein said host cell is a prokaryotic cell.

34. The host cell of 32, wherein said host cell is a eukaryotic cell.

35. The host cell of claim 34, wherein said eukaryotic cell is a protist cell, an animal cell, a plant cell, a fungal cell, a mammalian cell, an avian cell, or an insect cell.

36. The host cell of claim 35, wherein said eukaryotic cell is *S. cerevisiae*, CHO cell, COS cell, or SF9 cell.

37. A method of producing a protein capable of binding TNF-α, the method comprising the steps of culturing the host cell described of claim 32 in culture medium under conditions sufficient to produce a binding protein capable of binding TNF-α.

38. A protein produced according to the method of claim 37.

39. A pharmaceutical composition comprising the binding protein of claim 1 or claim 37, and a pharmaceutically acceptable carrier.

40. The pharmaceutical composition of claim 39, further comprising at least one additional agent for treating a disorder in which TNF-α activity is detrimental.

41. The pharmaceutical composition of claim 40, wherein said additional agent is: a therapeutic agent; an imaging agent; a cytotoxic agent; an angiogenesis inhibitor; a kinase inhibitor; a co-stimulation molecule blocker; an adhesion molecule blocker; an anti-cytokine antibody or functional fragment thereof; methotrexate; cyclosporin; rapamycin; FK506; a detectable label or reporter; a TNF antagonist; an anti-rheumatic; a muscle relaxant; a narcotic; a non-steroid anti-inflammatory drug (NSAID); an analgesic; an anesthetic; a sedative; a local anesthetic; a

neuromuscular blocker; an antimicrobial; an antipsoriatic; a corticosteroid; an anabolic steroid; an erythropoietin; an immunization; an immunoglobulin; an immunosuppressive; a growth hormone; a hormone replacement drug; a radiopharmaceutical; an antidepressant; an antipsychotic; a stimulant; an asthma medication; a beta agonist; an inhaled steroid; an oral steroid; an epinephrine or analog thereof; a cytokine; or a cytokine antagonist.

42. A method for treating a mammal comprising the step of administering to the mammal an effective amount of the composition of claim 39.

43. A method for reducing human TNF-α activity comprising contacting human TNF-α with the binding protein of claim 1 such that human TNF-α activity is reduced.

44. A method for reducing human TNF-α activity in a human subject suffering from a disorder in which TNF-α activity is detrimental, comprising administering to the human subject the binding protein of claim 1 such that human TNF-α activity in the human subject is reduced.

45. A method for treating a subject for a disease or a disorder in which TNF-α activity is detrimental by administering to the subject the binding protein of claim 1 such that treatment is achieved.

46. The method of claim 45, wherein the disorder is an autoimmune and/or inflammatory disorder.

46. The method of claims 46, wherein the disorder is Crohn's disease, plaque psoriasis, rheumatoid arthritis, psoratic arthritis, osteoarthritis, juvenile idiopathic arthritis, multiple sclerosis, systemic lupus erythematosus, ankylosing spondylitis, insulin-dependent diabetes mellitus, autoimmune diabetes, allergy, and autoimmune uveitis.

47. The method of claim 46, wherein the disorder is a respiratory disorder; asthma; allergic and nonallergic asthma; asthma due to infection; asthma due to infection with respiratory syncytial virus (RSV); chronic obstructive pulmonary disease (COPD); a condition involving airway inflammation; eosinophilia; fibrosis and excess mucus production; cystic fibrosis; pulmonary fibrosis; an atopic disorder; atopic dermatitis; urticaria; eczema; allergic rhinitis; allergic enterogastritis; an inflammatory and/or autoimmune condition of the skin; an inflammatory and/or autoimmune condition of gastrointestinal organs; inflammatory bowel diseases (IBD); ulcerative colitis; Crohn's disease; an inflammatory and/or autoimmune condition of the liver; liver cirrhosis; liver fibrosis; liver fibrosis caused by hepatitis B and/or C virus; scleroderma; tumors or cancers;

hepatocellular carcinoma; glioblastoma; lymphoma; Hodgkin's lymphoma; a viral infection; a bacterial infection; a parasitic infection; HTLV-1 infection; suppression of expression of protective type 1 immune responses, and suppression of expression of a protective type 1 immune response during vaccination.

48. A method of treating a patient suffering from a disorder in which TNF-α is detrimental comprising administering the binding protein of claim 1 before, concurrent, or after the administration of a second agent, wherein the second agent is an antibody, or fragment thereof, capable of binding human IL-12; PGE2; LPA; NGF; CGRP; SubP; RAGE; histamine; a histamine receptor blocker; bradykinin; IL-1alpha; IL-1beta; VEGF; PLGF; methotrexate; a corticosteroid, a glucocorticoid receptor modulator; cyclosporin, rapamycin, FK506, or a non-steroidal anti-inflammatory agent.

49. A method of treating a patient suffering from a disorder in which TNF-α is detrimental, the method comprising the step of administering the binding protein of claim 1 before, concurrently, or after the administration of a second agent, wherein the second agent is chosen from TNF antagonists; a soluble fragment of a TNF receptor; ENBREL®; TNF enzyme antagonists; TNF converting enzyme (TACE) inhibitors; muscarinic receptor antagonists; TGF-beta antagonists; interferon gamma; perfenidone; chemotherapeutic agents, methotrexate; leflunomide; sirolimus (rapamycin) or an analog thereof, CCI-779; COX2 or cPLA2 inhibitors; NSAIDs; immunomodulators; p38 inhibitors; TPL-2, MK-2 and NFkB inhibitors; budenoside; epidermal growth factor; corticosteroids; cyclosporine; sulfasalazine; aminosalicylates; 6-mercaptopurine; azathioprine; metronidazole; lipoxygenase inhibitors; mesalamine; olsalazine; balsalazide; antioxidants; thromboxane inhibitors; IL-1 receptor antagonists; anti-IL-1b antibodies; anti-IL-6 antibodies; growth factors; elastase inhibitors; pyridinyl-imidazole compounds; antibodies or agonists of TNF, LT, IL-1β, IL-2, IL-3, IL-4, IL-5, IL-6, IL-7, IL-8, IL-9, IL-10, IL-11, IL-12, IL-14, IL-15, IL-16, IL-17, IL-18, IL-19, IL-20, IL-21, IL-22, IL-23, IL-24, IL-25, IL-26, IL-27, IL-28, IL-29, IL-30, IL-31, IL-32, IL-33, EMAP-II, GM-CSF, FGF, or PDGF; antibodies of CD2, CD3, CD4, CD8, CD25, CD28, CD30, CD40, CD45, CD69, CD90 or their ligands; FK506; rapamycin; mycophenolate mofetil; ibuprofen; prednisolone; phosphodiesterase inhibitors; adensosine agonists; antithrombotic agents; complement inhibitors; adrenergic agents; IRAK, NIK, IKK, p38, or MAP kinase inhibitors; IL-1β converting enzyme inhibitors; TNFa converting enzyme inhibitors; T-cell signaling inhibitors; metalloproteinase inhibitors; 6-mercaptopurines; angiotensin converting enzyme inhibitors; soluble cytokine receptors; soluble p55 TNF receptor; soluble p75 TNF receptor; sIL-1RI; sIL-1RII; sIL-6R; anti-inflammatory cytokines; and TGFb.

50. The method of claims 40-48, wherein said administering to the subject is by at least parenteral, subcutaneous, intramuscular, intravenous, intra-articular, intrabronchial, intraabdominal, intracapsular, intracartilaginous, intracavitary, intracelial, intracerebellar, intracerebroventricular, intracolic, intracervical, intragastric, intrahepatic, intramyocardial, intraosteal, intrapelvic, intrapericardiac, intraperitoneal, intrapleural, intraprostatic, intrapulmonary, intrarectal, intrarenal, intraretinal, intraspinal, intrasynovial, intrathoracic, intrauterine, intravesical, bolus, vaginal, rectal, buccal, sublingual, intranasal, or transdermal.

51. A method of detecting human TNF-α in a sample comprising:
(i) contacting the sample with an TNF-α binding protein as described in claim 1 or an TNF-α binding portion thereof;
and
(ii) detecting formation of a complex between the TNF-α binding protein or binding portion thereof and TNF-α in the sample, wherein a statistically significant change in the formation of the complex in the sample relative to that in a control sample or relative TNF-α in the sample.

52. The method according to claim 51, wherein the sample is whole blood, plasma, serum, urine, saliva, or tissue biopsy.

53. A method of detecting human TNF-α in a human subject comprising:
(i) administering an TNF-α binding protein as described in claim 1, or an TNF-α binding portion thereof, to a test subject or a control subject under conditions that allow binding of the TNF-α binding protein, or TNF-α binding portion thereof, to human TNF-α;
and
(ii) detecting formation of a complex between the binding protein or binding portion thereof and TNF-α, wherein a statistically significant change in the formation of the complex in the test subject relative to the control subject or relative to formation of the complex in the test subject at an earlier time point is indicative of the presence of TNF-α.