

DESCRIPTION

Title of Invention

METHOD FOR PRODUCING ANTIBODY FUSION PROTEIN

Technical Field

5 [0001] The present invention relates to a method for producing a fusion protein in which an antibody is fused with a lysosomal enzyme, for example, a method for purifying a recombinant fusion protein, which has been obtained by culturing a host cell into which an expression vector incorporating the gene encoding the fusion protein is introduced,
10 to such a purity as permits its direct use as medical drug.

Background Art

[0002] Currently, many medicines containing a recombinant protein as an active ingredient are commercially available. Such recombinant proteins are obtained in the culture supernatant by culturing a host cell
15 into which an expression vector incorporating the gene encoding the protein of interest is introduced. The recombinant proteins obtained in the culture supernatant cannot be used as medicines as they are, because they contain contaminants. For using them as medicines, it is necessary to purify the recombinant proteins contained in the culture supernatant.

20 [0003] Reported are methods for purifying the recombinant proteins to such a level as permits their usages as medical drugs, wherein the proteins are obtained in the culture supernatants by culturing the host cells, that host cells are mammalian cells. For example, a method has been reported in which human erythropoietin (hEPO), a glycoprotein
25 that effects on erythroblast progenitor cells to differentiate them into erythrocytes and promotes the production of erythrocytes, is expressed

as a recombinant protein using CHO cells as a host cell, and purified from the supernatant by using various kinds of chromatography including dye affinity column chromatography to such a level as permits its use as medical drug (Patent Document 1). Further, for example, a method has been reported in which human follicle stimulating hormone (hFSH), which is one of gonadotropic hormones having the activity to promote the production and secretion of estrogen in the ovary, is expressed as a recombinant protein using CHO cells as a host cell, and purified from the supernatant by using various kinds of chromatography including cation exchange column chromatography to such a level as permits its use as medical drug (Patent Document 2). Further, for example, a method has been reported in which human iduronate-2-sulfatase (hI2S), which is one of lysosome enzymes having the activity of hydrolyzing sulfate bond in glycosaminoglycan (GAG) molecule such as heparan sulfate and dermatan sulfate, is expressed as a recombinant protein using CHO cells as a host cell, and purified from the supernatant by using various kinds of chromatography including cation exchange column chromatography to such a level as permits its use as medical drug (Patent Document 3). And, for example, human α -galactosidase A (h α -Gal A), which is one of lysosome enzymes having the activity of hydrolyzing terminal α -galactosyl bonds of glycolipids and glycoproteins, is expressed as a recombinant protein using CHO cells as a host cell, and purified from the supernatant by using various kinds of chromatography including anion exchange column chromatography to such a level as permits its use as medical drug (Patent Document 4 and 5). Further, for example, human DNase I,

having the activity of degrading DNA nonspecifically in a base sequence, is expressed as a recombinant protein using CHO cells as a host cell, and purified from the supernatant by using various kinds of chromatography including dye ligand affinity column chromatography to such a level as permits its use as medical drug (Patent Document 6).

[0004] As such in order to obtain a recombinant protein that can be used as a medicine, unique purification methods have been developed for each one of recombinant proteins.

Citation List

Patent Literature

[0005] [Patent Document 1] JP 2010-511378

[Patent Document 2] JP 2009-273427

[Patent Document 3] JP 2014-508506

[Patent Document 4] WO 2014/017088

[Patent Document 5] WO 2016/117341

[Patent Document 6] WO 2016/067944

Summary of Invention

Technical Problem

[0006] An objective of the present invention is to provide a method for expressing a fusion protein in which an antibody is fused to another protein as a recombinant protein, and purifying the protein to such a purity as permits its distribution to the market as a medical drug.

Solution to Problem

[0007] In a study for the above-mentioned object, as a result of intense studies, the present inventors found that a fusion protein, in which an anti-transferrin receptor antibody has been fused to human

iduronate-2-sulfatase (hI2S), can be purified effectively and at high purity by culturing in a serum-free medium mammalian cells introduced with the expression vector incorporated a gene encoding the fusion protein, and purifying the fusion protein obtained in the culture supernatant by using by using a column chromatography employing as a solid phase a material having affinity for the fusion protein, a column chromatography employing as a solid phase a material having affinity for the phosphate group, and a size exclusion column chromatography. The present invention was completed based on these findings. Thus the present invention provides what follows:

1. A method for production of a fusion protein in which an antibody and a human lysosomal enzyme are fused, the method comprising;

(a) a step of culturing mammalian cells producing the fusion protein in a serum-free medium to let the mammalian cells secrete the fusion protein in the culture medium,

(b) a step of collecting culture supernatant by removing the mammalian cells from the culture medium, and

(c) a step of purifying the fusion protein from the culture supernatant by a column chromatography employing as a solid phase a material to which a substance having affinity for the fusion protein has been bound, a column chromatography employing as a solid phase a material having affinity for the phosphate group, and a size exclusion column chromatography.

2. The method for production according to (1) above, wherein, in the step (c), the column chromatography employing as a

solid phase a material to which a substance having affinity for the fusion protein has been bound, the column chromatography employing as a solid phase a material having affinity for the phosphate group, and the size exclusion column chromatography are conducted in this order.

5 3. The method for production according to (1) or (2) above, wherein the substance having affinity for the fusion protein is selected from the group consisting of Protein A, Protein G, Protein L, Protein A/G, an antigen against said antibody, an antibody recognizing said antibody as an antigen, and an antibody against the lysosomal enzyme.

10 4. The method for production according to any one of (1) to (3) above, wherein the material having affinity for a phosphate group is fluoroapatite or hydroxyapatite.

 5. The method for production according to any one of (1) to (3) above, wherein the material having affinity for phosphate group is hydroxyapatite.

15 6. The method for production according to any one of (1) to (5) above, wherein said antibody fused to the human lysosomal enzyme is a humanized antibody or a human antibody.

 7. The method for production according to any one of (1) to (5) above, wherein said antibody fused to the human lysosomal enzyme is a humanized antibody.

20 8. The method for production according to any one of (1) to (7) above, wherein said antibody fused to the human lysosomal enzyme recognizes a molecule present on the surface of vascular endothelial cells as an antigen.

25 9. The method for production according to (8) above,

wherein the molecule present on the surface of vascular endothelial cells is selected from the group consisting of transferrin receptor (TfR), insulin receptor, leptin receptor, lipoprotein receptor, IGF receptor, OATP-F, organic anion transporter, MCT-8, monocarboxylic acid transporter, and an Fc receptor.

10. The method for production according to (8) above, wherein the vascular endothelial cells are cerebral vascular endothelial cells.

11. The method for production according to (10) above, wherein the molecule present on the surface of the cerebrovascular endothelial cell is selected from the group consisting of transferrin receptor (TfR), insulin receptor, leptin receptor, lipoprotein receptor, IGF receptor, OATP-F, organic anion transporter, MCT-8, and monocarboxylic acid transporter.

12. The method for production according to any one of (8) to (11) above, wherein the vascular endothelial cells are human vascular endothelial cells.

13. The method for production according to any one of (1) to (12) above, wherein said antibody is an anti-human transferrin receptor antibody.

14. The method for production according to any one of (1) to (13) above, wherein said antibody and the human lysosomal enzyme are fused via a linker in the fusion protein, and wherein the linker is selected from the group consisting of polyethylene glycol, polypropylene glycol, copolymer of ethylene glycol and propylene glycol, polyoxyethylated polyol, polyvinyl alcohol, polysaccharide,

dextran, polyvinyl ether, biodegradable polymer, lipid polymer, chitin, hyaluronic acid, biotin-streptavidin, and a derivative thereof.

15. The method for production according to any one of (1) to (13) above, wherein the human lysosomal enzyme is linked, by peptide bonds directly or via a linker sequence, to the heavy chain of said antibody on the C-terminal side or the N-terminal side thereof in the fusion protein.

16. The method for production according to any one of (1) to (13) above, wherein the human lysosomal enzyme is linked, by peptide bonds directly or via a linker sequence, to the light chain of said antibody on the C-terminal side or the N-terminal side thereof in the fusion protein.

17. The method for production according to (15) or (16) above, wherein the linker sequence consists of 1 to 50 amino acid residues.

18. The method for production according to (17), wherein the linker sequence comprises an amino acid sequence selected from the group consisting of a single glycine, a single serine, the amino acid sequence of Gly-Ser, the amino acid sequence of Gly-Gly-Ser, the amino acid sequence set forth as SEQ ID NO:1, the amino acid sequence set forth as SEQ ID NO:2, the amino acid sequence set forth as SEQ ID NO:3, and the amino acid sequences consisting of 1 to 10 thereof that are consecutively linked.

19. The method for production according to (17) or (18) above, wherein the linker sequence is represented by the amino acid sequence of Gly-Ser.

20. The method for production according to any one of (1) to (19) above, wherein the human lysosomal enzyme is human iduronate-2-sulfatase (human I2S).

21. The method for production according to (20) above, wherein the human I2S comprises the amino acid sequence set forth as SEQ ID NO:5.

22. The method for production according to (20) above, wherein the human I2S has at least 80% amino acid sequence identity to the amino acid sequence set forth as SEQ ID NO:5, and has an activity as human I2S.

23. The method for production according to (20) above, wherein the human I2S has at least 90% amino acid sequence identity to the amino acid sequence set forth as SEQ ID NO:5, and has an activity as human I2S.

24. The method for production according to (20) above, wherein the human I2S has the amino acid sequence introduced 1 to 10 amino acid substitutions, deletions or additions relative to the amino acid sequence set forth as SEQ ID:NO5, and has an activity as human I2S.

25. The method for production according to (20) above, wherein the human I2S has the amino acid sequence introduced 1 to 5 amino acid substitutions, deletions or additions relative to the amino acid sequence set forth as SEQ ID:NO5, and has an activity as human I2S.

26. The method for production according to (20) above, wherein the human I2S has the amino acid sequence introduced 1 to 3

amino acid substitutions, deletions or additions relative to the amino acid sequence set forth as SEQ ID:NO5, and has an activity as human I2S.

27. The method for production according to any one of (1) to (26) above, wherein said antibody is a human anti-hTfR antibody, and the human anti-hTfR antibody is selected from the group consisting of (a) to (c) below;

(a) the human anti-hTfR antibody, wherein the light chain and the heavy chain thereof comprise the amino acid sequences set forth as SEQ ID NO:6 and SEQ ID NO:7, respectively,

(b) the human anti-hTfR antibody, wherein the light chain and the heavy chain thereof comprise the amino acid sequences set forth as SEQ ID NO:8 and SEQ ID NO:9, respectively, and

(c) the human anti-hTfR antibody, wherein the light chain and the heavy chain thereof comprise the amino acid sequences set forth as SEQ ID NO:10 and SEQ ID NO:11, respectively.

28. The method for production according to any one of (1) to (26) above, wherein said antibody is a human anti-hTfR antibody, and the human anti-hTfR antibody is selected from the group consisting of (a) to (c) below;

(a) the human anti-hTfR antibody, wherein the light chain and the heavy chain thereof have at least 80% amino acid sequence identity to the amino acid sequence set forth as SEQ ID NO:6 and SEQ ID NO:7, respectively,

(b) the human anti-hTfR antibody, wherein the light chain and the heavy chain thereof have at least 80% amino acid sequence identity

to the amino acid sequence set forth as SEQ ID NO:8 and SEQ ID NO:9, respectively, and

5 (c) the human anti-hTfR antibody, wherein the light chain and the heavy chain thereof have at least 80% amino acid sequence identity to the amino acid sequence set forth as SEQ ID NO:10 and SEQ ID NO:11, respectively.

29. The method for production according to any one of (1) to (26) above, wherein said antibody is a human anti-hTfR antibody, and the human anti-hTfR antibody is selected from the group consisting of (a) to (c) below;

(a) the human anti-hTfR antibody, wherein the light chain and the heavy chain thereof have at least 90% amino acid sequence identity to the amino acid sequence set forth as SEQ ID NO:6 and SEQ ID NO:7, respectively,

15 (b) the human anti-hTfR antibody, wherein the light chain and the heavy chain thereof have at least 90% amino acid sequence identity to the amino acid sequence set forth as SEQ ID NO:8 and SEQ ID NO:9, respectively, and

20 (c) the human anti-hTfR antibody, wherein the light chain and the heavy chain thereof have at least 90% amino acid sequence identity to the amino acid sequence set forth as SEQ ID NO:10 and SEQ ID NO:11, respectively.

30. The method for production according to any one of (1) to (26) above, wherein said antibody is a human anti-hTfR antibody, and the human anti-hTfR antibody is selected from the group consisting of (a) to (c) below;

(a) the human anti-hTfR antibody, wherein the light chain thereof has the amino acid sequence introduced 1 to 10 amino acid substitutions, deletions or additions relative to the amino acid sequence set forth as SEQ ID NO:6, and wherein the heavy chain thereof has the amino acid sequence introduced 1 to 10 amino acid substitutions, deletions or additions relative to the amino acid sequence set forth as SEQ ID NO:7,

(b) the human anti-hTfR antibody, wherein the light chain thereof has the amino acid sequence introduced 1 to 10 amino acid substitutions, deletions or additions relative to the amino acid sequence set forth as SEQ ID NO:8, and wherein the heavy chain thereof has the amino acid sequence introduced 1 to 10 amino acid substitutions, deletions or additions relative to the amino acid sequence set forth as SEQ ID NO:9, and

(c) the human anti-hTfR antibody, wherein the light chain thereof has the amino acid sequence introduced 1 to 10 amino acid substitutions, deletions or additions relative to the amino acid sequence set forth as SEQ ID NO:10, and wherein the heavy chain thereof has the amino acid sequence introduced 1 to 10 amino acid substitutions, deletions or additions relative to the amino acid sequence set forth as SEQ ID NO:11.

31. The method for production according to any one of (1) to (26) above, wherein said antibody is a human anti-hTfR antibody, and the human anti-hTfR antibody is selected from the group consisting of (a) to (c) below;

(a) the human anti-hTfR antibody, wherein the light chain

thereof has the amino acid sequence introduced 1 to 3 amino acid substitutions, deletions or additions relative to the amino acid sequence set forth as SEQ ID NO:6, and wherein the heavy chain thereof has the amino acid sequence introduced 1 to 3 amino acid substitutions, deletions or additions relative to the amino acid sequence set forth as SEQ ID NO:7,

(b) the human anti-hTfR antibody, wherein the light chain thereof has the amino acid sequence introduced 1 to 3 amino acid substitutions, deletions or additions relative to the amino acid sequence set forth as SEQ ID NO:8, and wherein the heavy chain thereof has the amino acid sequence introduced 1 to 3 amino acid substitutions, deletions or additions relative to the amino acid sequence set forth as SEQ ID NO:9, and

(c) the human anti-hTfR antibody, wherein the light chain thereof has the amino acid sequence introduced 1 to 3 amino acid substitutions, deletions or additions relative to the amino acid sequence set forth as SEQ ID NO:10, and wherein the heavy chain thereof has the amino acid sequence introduced 1 to 3 amino acid substitutions, deletions or additions relative to the amino acid sequence set forth as SEQ ID NO:11.

32. The method for production according to (20) above, wherein said antibody is a human anti-hTfR antibody, the human lysosomal enzyme is human iduronate-2-sulfatase, and the fusion protein is selected from the group consisting of (a) to (c) below;

(a) the fusion protein comprising the light chain of the humanized anti-hTfR antibody having the amino acid sequence set forth

as SEQ ID NO:6, and the heavy chain of the humanized anti-hTfR antibody having the amino acid sequence set forth as SEQ ID NO:12 to which the human iduronate-2-sulfatase set forth as SEQ ID NO:1 is linked on the C-terminal side thereof and via a linker sequence of Gly-Ser,

(b) the fusion protein comprising the light chain of the humanized anti-hTfR antibody having the amino acid sequence set forth as SEQ ID NO:8, and the heavy chain of the humanized anti-hTfR antibody having the amino acid sequence set forth as SEQ ID NO:13 to which the human iduronate-2-sulfatase set forth as SEQ ID NO:1 is linked on the C-terminal side thereof and via a linker sequence of Gly-Ser, and

(c) the fusion protein comprising the light chain of the humanized anti-hTfR antibody having the amino acid sequence set forth as SEQ ID NO:10, and the heavy chain of the humanized anti-hTfR antibody having the amino acid sequence set forth as SEQ ID NO:14 to which the human iduronate-2-sulfatase set forth as SEQ ID NO:1 is linked on the C-terminal side thereof and via a linker sequence of Gly-Ser.

33. The method for production according to (20) above, wherein said antibody is a human anti-hTfR antibody, the human lysosomal enzyme is human iduronate-2-sulfatase (human I2S), and the fusion protein is selected from the group consisting of (a) to (c) below;

(a) the fusion protein comprising;

the light chain of the humanized anti-hTfR antibody having the amino acid sequence set forth as SEQ ID NO:6, and

the heavy chain of the humanized anti-hTfR antibody to which the human iduronate-2-sulfatase is linked on the C-terminal side thereof via a linker sequence of Gly-Ser, and having the amino acid sequence set forth as SEQ ID NO:12 as a whole,

5 (b) the fusion protein comprising;

the light chain of the humanized anti-hTfR antibody having the amino acid sequence set forth as SEQ ID NO:8, and

the heavy chain of the humanized anti-hTfR antibody to which the human iduronate-2-sulfatase is linked on the C-terminal side thereof via a linker sequence of Gly-Ser, and having the amino acid sequence set forth as SEQ ID NO:13 as a whole, and

(c) the fusion protein comprising;

the light chain of the humanized anti-hTfR antibody having the amino acid sequence set forth as SEQ ID NO:10, and

15 the heavy chain of the humanized anti-hTfR antibody to which the human iduronate-2-sulfatase is linked on the C-terminal side thereof via a linker sequence of Gly-Ser, and having the amino acid sequence set forth as SEQ ID NO:14 as a whole.

Effects of Invention

20 [0008] The present invention enables to provide a fusion protein of an anti-transferrin receptor antibody and a lysosomal enzyme, that fusion protein has been purified to such a purity as permits its clinical usage as a therapeutic agent for a lysosomal disease accompanied with central nervous system disorders. In particular, it enables to provide a fusion
25 protein of anti-transferrin receptor antibody and human I2S purified to such a purity as permits its clinical usage as a therapeutic agent for

Hunter syndrome accompanied with central nervous system disorders.

Brief Description of Drawings

[0009] [FIG. 1] FIG. 1 shows the SE-HPLC chart of the purified product of I2S-anti-hTfR antibody obtained in Example 6. The vertical axis shows absorbance at 215 nm, and the horizontal axis shows retention time. (A): a peak corresponding to monomer of I2S-anti-hTfR antibody 3, (B): a peak corresponding to multimer of I2S-anti-hTfR antibody 3.

[FIG. 2] FIG. 2 shows the SE-HPLC chart of the purified product of I2S-anti-hTfR antibody obtained in Example 12. The vertical axis shows absorbance at 215 nm, and the horizontal axis shows retention time. (A): a peak corresponding to monomer of I2S-anti-hTfR antibody 3, (B): a peak corresponding to multimer of I2S-anti-hTfR antibody 3.

Description of Embodiments

[0010] The present invention relates to a method for producing a protein in which an anti-transferrin receptor antibody (anti-TfR antibody) is bound to a human lysosomal enzyme. Here, the antibody to be bound to the lysosomal enzyme is not particularly limited as to the animal species of the antibody, as long as it has a property to specifically bind to the antigen, but particularly, it is a human antibody or a humanized antibody. For example, the antibody may be an antibody of a mammal other than human, or it may be a chimeric antibody of a human antibody and a mammalian antibody other than human.

[0011] The term "human antibody" refers to an antibody whose entirety is encoded by a gene originating from human. However, the term "human antibody", however, also includes an antibody encoded by a gene obtained by introducing a mutation into an original human gene

for a purpose of enhancing expression efficiency of the gene, for example, without modifying the original amino acid sequence. The term "human antibody" also includes an antibody which is produced by combining two or more genes encoding human antibodies and replacing
5 a certain part of a human antibody with a part of another human antibody. A human antibody includes three complementarity determining regions (CDRs) in the light chain of the immunoglobulin and three complementarity determining regions (CDRs) in the heavy chain of the immunoglobulin. The three CDRs in the light chain of the
10 immunoglobulin are called, from the N-terminal side, CDR1, CDR2 and CDR3, respectively. The three CDRs in the heavy chain of the immunoglobulin are also called, from the N-terminal side, CDR1, CDR2 and CDR3, respectively. The term "human antibody" also includes human antibody produced by replacing a CDR of a human
15 antibody with a CDR of another human antibody to modify such properties as the antigen specificity and the affinity of the original human antibodies, etc.

[0012] In the present invention, the term "human antibody" also includes an antibody which is produced through modification of the
20 gene of the original human antibody by introducing a mutation, such as substitution, deletion, addition, to the amino acid sequence of the original antibody. When replacing one or more amino acids of the amino acid sequence of the original antibody with other amino acids, the number of amino acid replaced may preferably be 1 to 20, more
25 preferably 1 to 10, still more preferably 1 to 5, and still more preferably 1 to 3. When deleting one or more amino acids of the amino acid

sequence of the original antibody, the number of amino acids deleted may preferably be 1 to 20, more preferably 1 to 10, still more preferably 1 to 5, and still more preferably 1 to 3. An antibody produced by a combined mutation of these substitution and deletion of amino acids is also a "human antibody". In some cases, one or more amino acids, preferably 1 to 20, more preferably 1 to 10, still more preferably 1 to 5, and still more preferably 1 to 3 amino acids may be added inside the amino acid sequence of the original antibody or on its N- or C-terminus. An antibody produced by a combined mutation of addition, substitution, and deletion of amino acids is also a "human antibody". The amino acid sequence of such a mutated antibody has an identity of preferably not lower than 80%, more preferably not lower than 90%, still more preferably not lower than 95%, and even more preferably not lower than 98%, to the amino acid sequence of the original antibody. Thus, in the present invention, the term "gene originating from human" includes not only the unmutated gene originating from human but also a gene produced by modifying it.

[0013] In the present invention, the term "humanized antibody" refers to an antibody in which part of the amino acid sequence of its variable region (e.g., especially the whole or part of its CDRs) originates from a non-human mammal while the rest originates from human. An example of humanized antibody is an antibody produced by replacing the three complementarity determining regions (CDRs) of the light chain of the immunoglobulin and the three complementarity determining regions (CDRs) of the heavy chain of the immunoglobulin constituting a human antibody, with CDRs from a non-human mammal.

As far as it originates from a non-human mammal, there is no particular limitation as to the biological species from which those CDRs originate that are grafted into a proper position of the human antibody, though preferred are mouse, rat, rabbit, horse or non-human primate, for example mouse.

[0014] A detailed explanation will be given below regarding the case where the antibody is a humanized antibody or human antibody. In human antibody light chain, there are λ and κ chains. The light chain constituting the antibody may either be λ and κ chain. And in heavy chain of human or humanized antibody, there are γ , μ , α , σ , and ϵ chains, which correspond to IgG, IgM, IgA, IgD and IgE, respectively. Though the heavy chain constituting the antibody may be any of γ , μ , α , σ , and ϵ chains, preferred is a γ chain. Further, in γ chain of antibody heavy chain, there are γ_1 , γ_2 , γ_3 and γ_4 chains, which correspond to IgG1, IgG2, IgG3 and IgG4, respectively. Where the heavy chain constituting the antibody is a γ chain, though the γ chain may be any of γ_1 , γ_2 , γ_3 and γ_4 chains, preferred is a γ_1 or γ_4 chain. In the case where the antibody is a humanized antibody or human antibody and IgG, the antibody light chain may either be λ chain or κ chain, and though the antibody heavy chain may either be γ_1 , γ_2 , γ_3 and γ_4 chains, preferred is a γ_1 or γ_4 chain. For example, a preferable embodiment of the antibody includes an antibody whose light chain is a λ chain and heavy chain is a γ_1 chain.

[0015] In the present invention, the term "chimeric antibody" refers to an antibody produced by connecting fragments of two or more different antibodies originating from two or more different species.

[0016] A chimeric antibody between a human antibody and a non-human mammalian antibody is an antibody provided by replacing part of a human antibody with part of a non-human mammalian antibody. As explained below, an antibody is made of an Fc region, a Fab region and a hinge region. A specific example of such chimeric antibodies is a chimeric antibody whose Fc region originates from a human antibody while its Fab region originates from a non-human mammalian antibody. The hinge region either originates from a human antibody or from a non-human mammalian antibody. On the contrary, the term chimeric antibody also includes one whose Fc region originates from a non-human mammalian antibody while its Fab region originates from a human antibody. In such a case also, the hinge region either originates from a human antibody or from a non-human mammalian antibody.

[0017] An antibody can be viewed as composed of a variable region and a constant region. Additional examples of chimeric antibodies include an antibody in which the heavy chain constant region (C_H) and the light chain constant region (C_L) both originate from a human antibody while the heavy chain variable region (V_H) and the light chain variable region (V_L) both originate from an antibody of a non-human mammal, and conversely, an antibody in which the heavy chain constant region (C_H) and the light chain constant region (C_L) both originate from an antibody of a non-human mammal, while the heavy chain variable region (V_H) and the light chain variable region (V_L) both originate from a human antibody. In these, there is no particular limitation as to the biological species of the non-human mammal, as far as it is a

non-human mammal, though preferred are mouse, rat, rabbit, horse or non-human primate, and mouse, more preferably mouse.

[0018] A chimeric antibody between a human antibody and a mouse antibody is designated in particular "human/mouse chimeric antibody".

5 Examples of human/mouse chimeric antibodies include a chimeric antibody in which the Fc region originates from a human antibody while the Fab region originates from a mouse antibody, and conversely, a chimeric antibody whose Fc region originates from mouse antibody, while its Fab region originates from a human antibody. A hinge region
10 either originate from a human antibody or a mouse antibody. Additional specific examples of human/mouse chimeric antibodies include those whose heavy chain constant region (C_H) and light chain constant region (C_L) originate from a human antibody while its heavy chain variable region (V_H) and light chain variable region (V_L) originate from a mouse
15 antibody, and conversely, those whose heavy chain constant region (C_H) and light chain constant region (C_L) originate from a mouse antibody while its heavy chain variable region (V_H) and light chain variable region (V_L) originate from a human antibody.

[0019] Originally, an antibody is of the basic structure having four
20 polypeptide chains in total consisting of two immunoglobulin light chains and two immunoglobulin heavy chains. However, in the present invention the term "antibody" refers, besides an antibody having this basic structure, also to:

(1) one consisting of two polypeptide chains: a single
25 immunoglobulin light chain and a single immunoglobulin heavy chain, and also, as explained later,

(2) a single-chain antibody consisting of an immunoglobulin light chain which is linked, on the C-terminal side thereof, to a linker sequence which in turn is linked, on the C-terminal side thereof, to an immunoglobulin heavy chain,

5 (3) single-chain antibodies consisting of an immunoglobulin heavy chain which is linked, on the C-terminal side thereof, to a linker sequence which in turn is linked, on the C-terminal side thereof, to an immunoglobulin light chain, and

(4) one consisting of a Fab region, i.e., a structure left behind by
10 removal of the Fc region from an antibody having the basic structure, as the original meaning, and one consisting of the Fab region and the whole or part of the hinge region (including Fab, F(ab'), and F(ab')₂) also are included in the term "antibody" in the present invention. Furthermore, scFv in which the variable region of the light chain and
15 the variable region of the heavy chain are linked via a linker sequence to form a single chain antibody is also included in the antibody of the present invention.

[0020] In the present invention, the term "single-chain antibody" refers to a protein in which an amino acid sequence comprising the whole or
20 part of an immunoglobulin light chain variable region linked, on the C-terminal side thereof, to a linker sequence, which in turn is linked, on the C-terminal side thereof, to the amino acid sequence of the whole or part of an immunoglobulin heavy chain variable region, and having an ability to specifically bind a certain antigen. Further, a protein in
25 which an amino acid sequence comprising the whole or part of an immunoglobulin heavy chain variable region is linked, on the

C-terminal side thereof, to a linker sequence, which in turn is further linked, on the C-terminal side thereof, to the amino acid sequence of the whole or part of an immunoglobulin light chain variable region, and which has an ability to specifically bind to a certain antigen, is also included in the term "single-chain antibody" in the present invention. For example, those described in (2) and (3) are included in "single-chain antibody". In a single-chain antibody in which an immunoglobulin heavy chain is linked, on the C-terminal side thereof and via a linker sequence, to an immunoglobulin light chain, the immunoglobulin heavy chain generally lacks the Fc region. An immunoglobulin light chain variable region has three complementarity determining regions (CDRs) which participate in determining the antigen specificity of an antibody. Likewise, an immunoglobulin heavy chain variable region also has three CDRs. Those CDRs are the primary regions that determine the antigen specificity of an antibody. Therefore, a single-chain antibody preferably contains all the three CDRs of the immunoglobulin heavy chain and all the three CDRs of the immunoglobulin light chain. However, it is also possible to provide a single-chain antibody in which one or more of those CDRs are deleted, insofar as the antigen-specific affinity of the antibody is retained.

[0021] In a single-chain antibody, the linker sequence placed between the light chain and the heavy chain of the immunoglobulin is preferably a peptide chain consisting of preferably 2 to 50, more preferably 8 to 50, still more preferably 10 to 30, even more preferably 12 to 18, or 15 to 25, for example 15 or 25 amino acid residues. While there is no particular limitation as to the specific amino acid sequence of such a

linker sequence insofar as the anti-hTfR antibody comprising the both chains linked thereby retains the affinity to hTfR, it is preferably made of glycine only, or of glycine and serine. For example, there are the amino acid sequence Gly-Ser, the amino acid sequence of Gly-Gly-Ser, the amino acid sequence of Gly-Gly-Gly, the amino acid sequence of Gly-Gly-Gly-Gly-Ser (SEQ ID NO:1), the amino acid sequence of Gly-Gly-Gly-Gly-Gly-Ser (SEQ ID NO:2), the amino acid sequence of Ser-Gly-Gly-Gly-Gly-Gly (SEQ ID NO:3), or a sequence which includes a sequence corresponding to 2 to 10 or 2 to 5 of any of those amino acid sequences consecutively linked. For example, in linking the amino acid sequence of the entire immunoglobulin heavy chain variable region on the C-terminal side thereof and via a linker sequence, to immunoglobulin light chain variable region to produce ScFV a preferable linker sequence comprises a linker sequence consisting of a total of 15 amino acids corresponding to three of the amino acid sequence of Gly-Gly-Gly-Gly-Ser (SEQ ID NO:1) consecutively linked. [0022] In the present invention, the antigen specifically recognized by the antibody is, for example, a molecule present on the surface of vascular endothelial cells (surface antigen). Examples of such surface antigens include transferrin receptor (TfR), insulin receptor, leptin receptor, lipoprotein receptor, IGF receptor, organic anion transporters such as OATP-F, monocarboxylic acid transporters such as MCT-8, Fc receptors, and the like, but are not limited to these. Antigens are preferably these molecules (surface antigens) present on the surface of human vascular endothelial cells.

[0023] Among the surface antigens described above, organic anion

transporters such as transferrin receptor (TfR), insulin receptor, leptin receptor, lipoprotein receptor, IGF receptor, OATP-F and the like, monocarboxylic acid transduct such as MCT-8 Porter is present on the surface of brain capillary endothelial cells (cerebral vascular endothelial cells) forming the blood brain barrier (Blood Brain Barrier). Antibodies capable of recognizing these antigens can bind to brain capillary endothelial cells via antigens. And antibodies bound to brain capillary endothelial cells can cross the blood brain barrier and reach the central nervous system. Therefore, by binding the protein of interest to such an antibody, it is possible to reach the central nervous system. Protein of interest may be a protein having a function to exert a drug effect in the central nervous system. For example, lysosomal enzymes that are deficient or dysfunctional in lysosomal disease patients with central nervous system disorders are mentioned as proteins of interest. Such a lysosomal enzyme cannot reach the central nervous system as it is and does not show a drug effect against a central nervous system disorder of a patient, but by allowing it to bind with these antibodies, it can pass through the blood brain barrier As a result, the central nervous system disorder found in lysosomal disease patients can be improved.

[0024] In the present invention, the term "human transferrin receptor" or "hTfR" refers to a membrane protein having the amino acid sequence set forth as SEQ ID NO:4. The anti-hTfR antibody of the present invention is, in one of its embodiments, that which binds also to the region from the cysteine residue at the position 89th from the N-terminus to the phenylalanine at the C-terminus in the amino acid sequence set forth as SEQ ID NO:4 (i.e., the extracellular region of the

monkey TfR), though it is not limited to this embodiment.

[0025] A method for preparing an antibody is described below, an antibody against hTfR taken as an example. For preparation of an antibody to hTfR, there is known a general method according to which a recombinant human transferrin receptor (rhTfR) is produced using cells which have an introduced expression vector having an incorporated hTfR gene, and then animals such as mice are immunized with this rhTfR. By collecting those cells which produce antibodies to hTfR from the immunized animals and fusing them with myeloma cells, hybridoma cells can be obtained having an ability to produce the anti-hTfR antibody.

[0026] Further, cells producing an antibody to hTfR can also be obtained by collecting immunocompetent cells from an animal such as mouse, and immunizing them with rhTfR by in vitro immunization. In conducting immunization by vitro immunization, there is no particular limitation as to the animal species from which the immunocompetent cells are derived, though preferred are mouse, rat, rabbit, guinea pig, dog, cat, horse, and primates including human, and more preferred are mouse, rat and human, and still more preferably mouse and human. As mouse immunocompetent cells, spleen cells prepared from mouse spleen may be used, for example. As human immunocompetent cells, such cells can be used as prepared from human peripheral blood, bone marrow, spleen, and the like. By immunizing human immunocompetent cells according to in vitro immunization, a human antibody to hTfR can be obtained.

[0027] In the present invention, there is no particular limitation as to the

human lysosomal enzyme to be linked to the anti-hTfR antibody. As such lysosomal enzymes, included are α -L-iduronidase, iduronate-2-sulfatase, glucocerebrosidase, β -galactosidase, GM2 activator protein, β -hexosaminidase A, β -hexosaminidase B, N-acetylglucosamine-1-phosphotransferase, α -mannosidase, β -mannosidase, galactosylceramidase, saposin C, arylsulfatase A, α -L-fucosidase, aspartylglucosaminidase, α -N-acetylgalactosaminidase, acidic sphingomyelinase, α -galactosidase A, β -glucuronidase, heparan N-sulfatase, α -N-acetylglucosaminidase, acetyl CoA: α -glucosaminide N-acetyltransferase, N-acetylglucosamine-6-sulfate sulfatase, acid ceramidase, amylo-1,6-glucosidase, sialidase, aspartylglucosaminidase (PPT1), tripeptidyl-peptidase 1 (TPP-1), hyaluronidase 1, CLN1, and CLN2, and the like.

[0028] When the antibody specifically recognizes a molecule present on the surface of the vascular endothelial cell (surface antigen), the human lysosomal enzyme linked to the antibody can be used as a therapeutic agent for central nervous system disorders, i.e. α -L-iduronidase as a therapeutic agent for central nervous system disorders in Hurler syndrome or Hurler-Scheie syndrome; iduronate-2-sulfatase as a therapeutic agent for central nervous system disorders in Hunter syndrome; glucocerebrosidase as a therapeutic agent for central nervous system disorders in Gaucher's disease; β -galactosidase as a therapeutic agent for central nervous system disorders in GM1 gangliosidosis Types 1 to 3; GM2 activator protein as a therapeutic agent for central nervous system disorders in GM2-gangliosidosis, AB variant; β -hexosaminidase A as a therapeutic

agent for central nervous system disorders in Sandhoff's disease and
 Tay-Sachs disease; β -hexosaminidase B as a therapeutic agent for
 central nervous system disorders in Sandhoff's disease;
 N-acetylglucosamine-1-phosphotransferase as a therapeutic agent for
 5 central nervous system disorders in I-cell disease; α -mannosidase as a
 therapeutic agent for central nervous system disorders in
 α -mannosidosis; β -mannosidase as a therapeutic agent for central
 nervous system disorders in β -mannosidosis; galactosylceramidase as a
 therapeutic agent for central nervous system disorders in Krabbe
 10 disease; saposin C as a therapeutic agent for central nervous system
 disorders in Gaucher's disease-like storage disease; arylsulfatase A as a
 therapeutic agent for central nervous system disorders in metachromatic
 white matter degeneration (metachromatic leukodystrophy);
 α -L-fucosidase as a therapeutic agent for central nervous system
 15 disorders in fucosidosis; aspartylglucosaminidase as a therapeutic agent
 for central nervous system disorders in aspartylglucosaminuria;
 α -N-acetylgalactosaminidase as a therapeutic agent for central nervous
 system disorders in Schindler disease and Kawasaki disease; acidic
 sphingomyelinase as a therapeutic agent for central nervous system
 20 disorders in Niemann-Pick disease; α -galactosidase A as a therapeutic
 agent for central nervous system disorders in Fabry disease;
 β -glucuronidase as a therapeutic agent for central nervous system
 disorders in Sly syndrome; heparan N-sulfatase,
 α -N-acetylglucosaminidase, acetyl CoA: α -glucosaminide
 25 N-acetyltransferase and N-acetylglucosamine-6-sulfate sulfatase as
 therapeutic agents for central nervous system disorders in Sanfilippo

syndrome; acid ceramidase as a therapeutic agent for central nervous system disorders in Farber disease; amylo-1,6-glucosidase as a therapeutic agent for central nervous system disorders in Cori's disease (Forbes-Cori's disease); sialidase as a therapeutic agent for central nervous system disorders in sialidase deficiency; aspartylglucosaminidase as a therapeutic agent for central nervous system disorders in aspartylglucosaminuria; palmitoyl protein thioesterase 1 (PPT-1) as a therapeutic agent for central nervous system disorders in neuronal ceroid lipofuscinosis or Santavuori-Haltia disease; tripeptidyl-peptidase 1 (TPP-1) as a therapeutic agent for central nervous system disorders in neuronal ceroid lipofuscinosis or Jansky-Bielschowsky disease; hyaluronidase 1 as a therapeutic agent for central nervous system disorders in hyaluronidase deficiency; CLN1 and CLN2 as therapeutic agents for central nervous system disorders in Batten disease. In particular, the anti-hTfR antibody of the present invention, after passing through the blood-brain barrier, reaches the brain parenchyma and the hippocampus neuron-like cells of the cerebrum, and Purkinje cells of the cerebellum, and is expected further to reach neuron-like cells of the striatum of the cerebrum and the neuron-like cells of the substantia nigra of the mesencephalon. Therefore, the anti-hTfR antibody can be fused with proteins which need to exhibit their functions in those tissues or cells to strength the pharmacological effects of the proteins. Medical applications of it, however, are not limited thereto.

[0029] In the case where the antibody specifically recognizes a molecule present on the surface of vascular endothelial cells (surface

antigen), lysosomal enzymes to be preferably linked to the antibody include human iduronate-2-sulfatase (hI2S). I2S is one of lysosome enzyme having an activity for hydrolyzing sulfate bonds present in glycosaminoglycan (GAG) molecules such as heparan sulfate and dermatan sulfate. Hunter syndrome is a genetic disorder in which this enzyme is congenitally deleted. In the patients of Hunter syndrome, heparan sulfate and dermatan sulfate accumulate in the tissues, resulting in various symptoms such as corneal opacity, mental development delay, and so on. However, in the mild cases, mental developmental delay may not be observed. Since the fusion protein between the antibody and hI2S can degrade GAG accumulated in brain tissues by passing through BBB, it can be used as a therapeutic agent for central nervous system disorders by administered to a patient with Hunter syndrome showing mental developmental delay.

[0030] In the present invention, the term "human I2S" or "hI2S" refers to hI2S particularly having the same amino acid sequence as wild type hI2S. The wild type hI2S has an amino acid sequence consisting of 525 amino acids set forth as SEQ ID NO: 5. However, not limited to this, a hI2S containing a mutation such as substitution, deletion, addition and so on added to the amino acid sequence of the wild type hI2S is also included in hI2S, as long as it has I2S activity. When amino acids of the amino acid sequence of hI2S are substituted with other amino acids, the number of amino acids to be substituted is preferably 1 to 10, more preferably 1 to 5, still more preferably 1 to 3, and still more preferably 1 to 2. When amino acids in the amino acid sequence of hI2S are deleted, the number of amino acids to be deleted is preferably 1 to 10, more

preferably 1 to 5, still more preferably 1 to 3, and still more preferably 1 to 2. A combined mutation of these addition, substitution, and deletion of amino acids can also be carried out. When adding one or more amino acids to the hI2S, they may be added inside, or on the N-terminal side or C-terminal side thereof, and the number of amino acids added is preferably 1 to 10, more preferably 1 to 5, still more preferably 1 to 3, and even more preferably 1 or 2. A combined mutation of these substitution and deletion of amino acids can also be carried out. The amino acid sequence of such a mutated hI2S has an identity of preferably not lower than 80%, more preferably not lower than 90%, still more preferably not lower than 95%, to the amino acid sequence of the original hI2S.

[0031] The statement that hI2S has the I2S activity herein means that the hI2S fused to an antibody has an activity not lower than 3% of the activity that the natural-type hI2S intrinsically has. However, the activity is preferably not lower than 10%, more preferably not lower than 20%, still more preferably not lower than 50%, even more preferably not lower than 80% of the activity that the natural-type hI2S intrinsically has. The same also applies when the I2S has one or more of mutations. The antibody is, for example, an anti-hTfR antibody.

[0032] In the present invention, the term "fusion protein" refers to a substance obtained by binding an antibody and a human lysosomal enzyme directly, or via a non-peptide linker or a peptide linker. Methods for conjugating antibodies and human lysosomal enzymes are described in detail below.

[0033] For binding an antibody to a lysosomal enzyme, a method is

available to bind them together via a non-peptide linker or a peptide linker. As non-peptide linkers, there can be used polyethylene glycol, polypropylene glycol, copolymer of ethylene glycol and propylene glycol, polyoxyethylated polyol, polyvinyl alcohol, polysaccharides, dextran, polyvinyl ether, biodegradable polymer, polymerized lipid, chitins, and hyaluronic acid, or derivatives thereof, or combinations thereof. A peptide linker is a peptide chain consisting of 1 to 50 amino acids linked by peptide bonds or a derivative thereof, whose N-terminus and C-terminus are to be covalently bonded either to the antibody or the lysosomal enzyme, respectively, to bind the antibody to such a c lysosomal enzyme.

[0034] When biotin-streptavidin is used as the non-peptide linker, the antibody and a human lysosomal enzyme may be bound to each other via binding between biotin and streptavidin, where the antibody is bound to the biotin, and the human lysosomal enzyme is bound to the streptavidin. Conversely, the antibody and a human lysosomal enzyme may be bound to each other via binding between biotin and streptavidin, where the antibody is bound to the streptavidin, and the human lysosomal enzyme is bound to the biotin.

[0035] In particular, a conjugate which is formed by binding the antibody of the present invention to human lysosomal enzyme via PEG as a non-peptide linker, is designated "anti-antibody-PEG-human lysosomal enzyme". An anti-antibody-PEG-human lysosomal enzyme can be prepared by first binding the antibody to PEG to form antibody-PEG, and then binding the antibody-PEG to the human lysosomal enzyme. Alternatively, an anti-antibody-PEG-human

lysosomal enzyme can be prepared by first binding the human lysosomal enzyme to PEG to form "human lysosomal enzyme-PEG", and then binding the "human lysosomal enzyme-PEG" to the antibody. In order to bind PEG to the antibody and the human lysosomal enzyme, a PEG is employed which is modified with such functional groups as carbonate, carbonylimidazole, active ester of carboxylic acid, azlactone, cyclic imide thione, isocyanate, isothiocyanate, imidate, aldehyde or the like. Such functional groups introduced to PEG react mainly with amino groups in the antibody and the human lysosomal enzyme to covalently bind PEG to the antibody and the human lysosomal enzyme. Though there is no particular limitation as to the molecular weight and the configuration of PEG employed here, its mean molecular weight (MW) is as follows: preferably MW=500 to 60000, more preferably MW=500 to 20000. For example, such PEG whose mean molecular weight is about 300, about 500, about 1000, about 2000, about 4000, about 10000, about 20000, and the like. PEG is preferably used as a non-peptide linker.

[0036] For example, "antibody-PEG" can be prepared by mixing the antibody with a polyethylene glycol having aldehyde groups as functional groups (ALD-PEG-ALD) so that the molar ratio of ALD-PEG-ALD to the antibody is 11, 12.5, 15, 110, 120 and the like, and then adding to the mixture a reducing agent such as NaCNBH_3 to let a reaction take place. Then, by reacting "anti-hTfR antibody-PEG" with a human lysosomal enzyme in the presence of a reducing agent such as NaCNBH_3 , "antibody-PEG-protein" is obtained. On the contrary, it is also possible to obtain "antibody-PEG-protein" by first

binding a human lysosomal enzyme to ALD-PEG-ALD to prepare "human lysosomal enzyme-PEG", and then binding the "human lysosomal enzyme-PEG" to the antibody.

[0037] The antibody and a human lysosomal enzyme can also be bound together through peptide bonds by linking the antibody heavy chain or light chain, on the C-terminal side or the N-terminal side thereof, either via a linker sequence or directly, to the N-terminus or the C-terminus of the human lysosomal enzyme, respectively. Thus the fusion protein between the antibody and a human lysosomal enzyme can be obtained by incorporating into a mammalian expression vector a DNA fragment in which a cDNA encoding the human lysosomal enzyme is placed in-frame directly, or via a DNA fragment encoding a linker sequence, on the 3'-end or 5'-end side of a cDNA encoding the heavy chain or light chain of the antibody, and culturing mammalian cells into which the above expression vector has been introduced. Where the DNA fragment encoding a human lysosomal enzyme is linked to the heavy chain, a mammalian expression vector in which a cDNA fragment encoding the antibody light chain is also introduced into the same host cells, whereas if DNA fragment encoding a human lysosomal enzyme is linked to the light chain, a mammalian expression vector in which a cDNA fragment encoding the antibody heavy chain is also incorporated into the same host cells. In the case where the antibody is a single-chain antibody, the fusion protein comprising the antibody and a human lysosomal enzyme combined can be obtained by incorporating, into an expression vector (for eukaryotic cells such as mammalian and yeast, or for prokaryotic cells such as E. coli.), a DNA fragment which is formed by linking the

cDNA encoding a human lysosomal enzyme, on the 5'-end side or on the 3'-end side thereof, directly or via a DNA fragment encoding a linker sequence, to the cDNA encoding the single-chain antibody, and allowing the fusion protein be expressed in those cells into which the expression vector has been introduced.

[0038] In a fusion protein of the type in which a human lysosomal enzyme is linked to the antibody light chain on the C-terminal side thereof, the antibody comprises an amino acid sequence including the whole or part of the light chain variable region and an amino acid sequence including the whole or part of the heavy chain variable region, and the human lysosomal enzyme is linked to the light chain of this antibody on the C-terminal side thereof. Here, the antibody light chain and a human lysosomal enzyme may be linked together, directly or via a linker.

[0039] In a fusion protein of the type in which a human lysosomal enzyme is linked to the antibody heavy chain on the C-terminal side thereof, the antibody comprises an amino acid sequence including the whole or part of the light chain variable region and an amino acid sequence including the whole or part of the heavy chain variable region, and the human lysosomal enzyme is linked to the heavy chain of this antibody on the C-terminal side thereof. Here, the antibody heavy chain and a human lysosomal enzyme may be linked together, directly or via a linker.

[0040] In a fusion protein of the type in which a human lysosomal enzyme is linked to the antibody light chain on the N-terminal side thereof, the antibody comprises an amino acid sequence including the

whole or part of the light chain variable region and an amino acid sequence including the whole or part of the heavy chain variable region, and the human lysosomal enzyme is linked to the light chain of this antibody on the N-terminal side thereof. Here, the antibody light chain and a human lysosomal enzyme may be linked together, directly or via a linker.

[0041] In a fusion protein of the type in which a human lysosomal enzyme is linked to the antibody heavy chain on the N-terminal side thereof, the antibody comprises an amino acid sequence including the whole or part of the light chain variable region and an amino acid sequence including the whole or part of the heavy chain variable region, and the human lysosomal enzyme is linked to the heavy chain of this antibody on the N-terminal side thereof. Here, the antibody heavy chain and a human lysosomal enzyme may be linked together, directly or via a linker.

[0042] In the above, when the linker sequence is placed between the antibody and a human lysosomal enzyme, the linker sequence may be a peptide chain consisting preferably of 1 to 50, more preferably of 1 to 17, still more preferably of 1 to 10, even more preferably of 1 to 5 amino acids, and in accordance with the human lysosomal enzyme to be linked to the anti-hTfR antibody, the number of amino acids of the linker sequence may be adjusted to 1, 2, 3, 1 to 17, 1 to 10, 10 to 40, 20 to 34, 23 to 31, 25 to 29, etc., as desired. Though there is no particular limitation as to amino acid sequence of the linker sequence insofar as the antibody linked by it retains the affinity to hTfR and a human lysosomal enzyme linked by the linker sequence also exhibits the

protein's own physiological activity under a physiological condition, the linker may preferably be composed of glycine and serine. Examples of such linkers include one consisting of a single amino acid either glycine or serine, the amino acid sequence of Gly-Ser, the amino acid sequence of Gly-Gly-Ser, the amino acid sequence of Gly-Gly-Gly-Gly-Ser (SEQ ID NO:1), the amino acid sequence of Gly-Gly-Gly-Gly-Gly-Ser (SEQ ID NO:2), the amino acid sequence of Ser-Gly-Gly-Gly-Gly-Gly (SEQ ID NO:3), or a sequence which includes 1 to 10 or 2 to 5 of any of those amino acid sequences consecutively linked. They have sequences consisting of 1 to 50, 2 to 17, 2 to 10, 10 to 40, 20 to 34, 23 to 31, or 25 to 29 amino acids. For example, those comprising the amino acid sequence of Gly-Ser may preferably be used as linker sequences. Same can be applied when the antibody is a single strand antibody.

[0043] Besides, in the present invention, when a peptide chain includes a plurality of linker sequences, each of those linker sequences is designated, from the N-terminal side, the first linker sequence, the second linker sequence, and so on, for convenience.

[0044] Preferred embodiments of the antibody, that antibody is a humanized antibody and an anti-human transferrin receptor antibody, include the following (x) to (z) below,

(X) the light chain comprises the amino acid sequence set forth as SEQ ID NO: 6, and the heavy chain comprises the amino acid sequence set forth as SEQ ID NO: 7;

(Y) the light chain comprises the amino acid sequence set forth as SEQ ID NO: 8, and the heavy chain comprises the amino acid sequence set forth as SEQ ID NO: 9;

(Z) the light chain comprises the amino acid sequence set forth as SEQ ID NO: 10, and the heavy chain comprises the amino acid sequence set forth as SEQ ID NO: 11. Here, (x), (y) and (z) correspond to a humanized anti-hTfR antibody No. 1, a humanized anti-hTfR antibody No. 2, and a humanized anti-hTfR antibody No. 3, respectively, that antibodies are described in the examples.

[0045] However, preferred embodiments of the antibody are not limited to the (x) to (z) above, when the antibody is a humanized antibody and an anti-human transferrin receptor antibody. For example, the antibody can be used in the present invention, whose amino acid sequence of the light chain has an identity not lower than 80% to the amino acid sequence of each one of light chain in the above (x) to (z), and whose amino acid sequence of the heavy chain has an identity not lower than 80% to the amino acid sequence of each one of heavy chain in the above (x) to (z) insofar as that antibody has affinity for hTfR. For example, the antibody can be used in the present invention, whose amino acid sequence of the light chain has an identity not lower than 90% to the amino acid sequence of each one of light chain in the above (x) to (z), and whose amino acid sequence of the heavy chain has an identity not lower than 90% to the amino acid sequence of each one of heavy chain in the above (x) to (z) insofar as that antibody has affinity for hTfR. For example, the antibody can be used in the present invention, whose amino acid sequence of the light chain has an identity not lower than 95% to the amino acid sequence of each one of light chain in the above (x) to (z), and whose amino acid sequence of the heavy chain has an identity not lower than 95% to the amino acid sequence of each one of

heavy chain in the above (x) to (z) insofar as that antibody has affinity for hTfR.

[0046] Further, the antibody can be used in the present invention, which has in the light chain the amino acid sequence corresponding to the amino acid sequence introduced 1 to 10 of amino acid substitution, deletion, or addition in each one of the amino acid sequence of the light chain set forth in (x)~(z) above, and has in the heavy chain the amino acid sequence corresponding to the amino acid sequence introduced 1 to 10 of amino acid substitution, deletion, or addition in each one of the amino acid sequence of the light chain set forth in (x)~(z) above.

Further, the antibody can be used in the present invention, which has in the light chain the amino acid sequence corresponding to the amino acid sequence introduced 1 to 5 of amino acid substitution, deletion, or addition in each one of the amino acid sequence of the light chain set forth in (x)~(z) above, and has in the heavy chain the amino acid sequence corresponding to the amino acid sequence introduced 1 to 5 of amino acid substitution, deletion, or addition in each one of the amino acid sequence of the light chain set forth in (x)~(z) above. Further, the

antibody can be used in the present invention, which has in the light chain the amino acid sequence corresponding to the amino acid sequence introduced 1 to 3 of amino acid substitution, deletion, or addition in each one of the amino acid sequence of the light chain set forth in (x)~(z) above, and has in the heavy chain the amino acid sequence corresponding to the amino acid sequence introduced 1 to 3 of amino acid substitution, deletion, or addition in each one of the amino acid sequence of the light chain set forth in (x)~(z) above.

[0047] In the preferred embodiment (x) of the above antibody, the amino acid sequence set forth as SEQ ID NO: 15 corresponds to a variable region in the amino acid sequence of the light chain set forth as SEQ ID NO: 6, and the amino acid sequence set forth as SEQ ID NO: 16 corresponds to a variable region in the amino acid sequence of the light chain set forth as SEQ ID NO: 7. In the preferred embodiment (x) of the above antibody, the amino acid sequence set forth as SEQ ID NO: 17 corresponds to a variable region in the amino acid sequence of the light chain set forth as SEQ ID NO: 8, and the amino acid sequence set forth as SEQ ID NO: 18 corresponds to a variable region in the amino acid sequence of the light chain set forth as SEQ ID NO: 9. In the preferred embodiment (x) of the above antibody, the amino acid sequence set forth as SEQ ID NO: 19 corresponds to a variable region in the amino acid sequence of the light chain set forth as SEQ ID NO: 10, and the amino acid sequence set forth as SEQ ID NO: 20 corresponds to a variable region in the amino acid sequence of the light chain set forth as SEQ ID NO: 11. In the preferred embodiments (x) to (z) of these antibodies, the substitution, deletion or addition into the amino acid sequence constituting the amino acid sequence of the heavy chain or/and the light chain is particularly introduced into these variable regions .

[0048] In the present invention, the identity between the amino acid sequence of an unmutated antibody and the amino acid sequence of an antibody produced by introducing a mutation into it may be readily calculated using well-known homology calculator algorithms. As such algorithms, there are, for example, BLAST (Altschul SF. J Mol. Biol.

215. 403-10 (1990)), a similarity search by Pearson and Lipman (Proc. Natl. Acad. Sci. USA. 85. 2444 (1988)), and the local homology algorithm of Smith and Waterman (Adv. Appl. Math. 2. 482-9 (1981)), and the like.

5 [0049] Preferred embodiments of the fusion protein between the antibody and a lysosomal enzyme, that antibody is the humanized anti-hTfR antibody and that lysosomal enzyme is human iduronate-2-sulfatase (human I2S), include the following (a) to (c) below,

10 (a) a fusion protein comprising a light chain of the humanized anti-hTfR antibody having the amino acid sequence set forth as SEQ ID NO: 6, and a heavy chain of humanized anti-hTfR antibody having the amino acid sequence set forth as SEQ ID NO: 7 and linked, on the C-terminus thereof via a linker sequence of Gly-Ser, to human
15 iduronate-2-sulfatase set forth as SEQ ID NO: 5,

(b) a fusion protein comprising a light chain of the humanized anti-hTfR antibody having the amino acid sequence set forth as SEQ ID NO: 8, and a heavy chain of humanized anti-hTfR antibody having the amino acid sequence set forth as SEQ ID NO: 9 and linked, on the
20 C-terminus thereof via a linker sequence of Gly-Ser, to human iduronate-2-sulfatase set forth as SEQ ID NO: 5,

(c) a fusion protein comprising a light chain of the humanized anti-hTfR antibody having the amino acid sequence set forth as SEQ ID NO: 10, and a heavy chain of humanized anti-hTfR antibody having the
25 amino acid sequence set forth as SEQ ID NO: 11 and linked, on the C-terminus thereof via a linker sequence of Gly-Ser, to human

iduronate-2-sulfatase set forth as SEQ ID NO: 5.

[0050] Further preferred embodiments of the fusion protein between the antibody and a lysosomal enzyme, that antibody is the humanized anti-hTfR antibody and that lysosomal enzyme is human iduronate-2-sulfatase (human I2S), include the following (a) to (c) below,

(a) a fusion protein comprising a light chain of the humanized anti-hTfR antibody having the amino acid sequence set forth as SEQ ID NO: 6, and

a heavy chain of the humanized anti-hTfR antibody linked, on the C-terminal side thereof and via a linker sequence of Gly-Ser, to the human iduronate-2-sulfatase, and having the amino acid sequence set forth as SEQ ID NO:12 as the whole linked heavy chain.

(b) a fusion protein comprising a light chain of the humanized anti-hTfR antibody having the amino acid sequence set forth as SEQ ID NO: 8, and

a heavy chain of the humanized anti-hTfR antibody linked, on the C-terminal side thereof and via a linker sequence of Gly-Ser, to the human iduronate-2-sulfatase, and having the amino acid sequence set forth as SEQ ID NO:13 as the whole linked heavy chain,

(c) a fusion protein comprising a light chain of the humanized anti-hTfR antibody having the amino acid sequence set forth as SEQ ID NO: 10, and

a heavy chain of the humanized anti-hTfR antibody linked, on the C-terminal side thereof and via a linker sequence of Gly-Ser, to the human iduronate-2-sulfatase, and having the amino acid sequence set

forth as SEQ ID NO:14 as the whole linked heavy chain,

[0051] In the present invention, the fusion protein between the antibody and a human lysosomal enzyme may be produced by culturing a mammalian cell, which is artificially manipulated so as to produce the fusion protein by expression or strong expression of a gene encoding the fusion protein. In this end, the gene to be strongly expressed in the mammalian cells producing the fusion protein is generally introduced into the mammalian cell by transformation with an expression vector introduced with the gene. Examples of the means for artificially modifying an intrinsic gene to let it be strongly expressed include, but not limited to, replacing the promoter upstream of the intrinsic gene with a promoter which strongly induces expression of the gene. Further, though there is no particular limitation on the mammalian cells, cells derived from human, mouse, Chinese hamster are preferable, and CHO cells, the cells derived from Chinese hamster ovary cells, are particularly preferable. In the present invention, the term "fusion protein" means particularly the fusion protein secreted into the medium when mammalian cells producing the fusion protein are cultured.

[0052] A fusion protein between the antibody and a human lysosomal enzyme can also be produced by producing the antibody and a human lysosomal enzyme, respectively, and then binding these together via a non-peptide linker or a peptide linker. For this, the antibody and the human lysosomal enzyme can be produced as recombinant proteins by culturing genetically manipulated mammalian cells so as to produce these by expressing or strongly expressing the genes encoding them.

[0053] There is no particular limitation as to the expression vector for

incorporating and expressing the gene encoding the fusion protein, the antibody, or a lysosomal enzyme, provided that it has the potential to let the gene express when introduced into mammalian cells. The gene incorporated in the expression vector is placed downstream of a DNA sequence (gene expression regulatory site) capable of regulating the frequency of gene transcription in mammalian cells. The gene expression regulatory site that can be used in the present invention includes, for example, cytomegalovirus-derived promoter, SV40 early promoter, human elongation factor-1 alpha (EF-1 alpha) promoter, human ubiquitin C promoter and the like.

[0054] Mammalian cells having such an introduced expression vector come to express a desired protein incorporated in the expression vector, but the expression levels vary in each of the cells. Therefore, in order to efficiently produce a recombinant protein, it is necessary to select a cell having a high expression level of the desired protein among from mammalian cells into which the expression vector has been introduced. In order to perform this selection step, the expression vector incorporates a gene that serves as a selectable marker.

[0055] The most common selection marker is an enzyme (drug resistance marker) that degrades drugs such as puromycin, neomycin, and the like. Mammalian cells will be killed in the presence of these drugs beyond certain concentrations. Mammalian cells into which an expression vector has been introduced, however, become viable in the presence of those drugs because such cells can decompose the drugs with the drug selection markers incorporated in the expression vector and thus detoxify them or weaken their toxicity. When those cells,

which have been introduced with an expression vector incorporated with a drug resistance marker, are successively cultured in a selective medium containing the drug corresponding to the drug resistance marker while gradually increasing the concentration of the drug, cells that can proliferate in the presence of the drug at relatively higher concentrations are obtained. Such cells that express a drug selection marker at high levels also tend to express, at high levels, a gene encoding a protein of interest incorporated together into the expression vector, and as a result, mammalian cells thus will be obtained that express the protein of interest at high levels.

[0056] As a selection marker, glutamine synthetase (GS) may also be used. Glutamine synthetase is an enzyme synthesizing glutamine from glutamic acid and ammonia. Mammalian cells die, when cultured in a selective medium which contains an inhibitor of glutamine synthetase, such as methionine sulfoximine (MSX), but not glutamine. But when the mammalian cells have been introduced with an expression vector incorporated with glutamine synthetase, the cells become to be capable to grow in the presence of MSX at higher concentrations. At this time, if the cells are successively cultured while gradually increasing the concentration of the MSX, the result was that the cells capable of proliferating in the presence of the MSX at relatively higher concentrations are obtained. The cells selected as such a manner generally tend to express, at high levels, a gene encoding a protein of interest incorporated in the expression vector concomitantly with glutamine synthetase, and as a result, mammalian cells thus will be obtained that express the protein of interest at high levels.

[0057] As a selection marker, dihydrofolate reductase (DHFR) may also be used. When DHFR is used as the selection marker, mammalian cells are cultured in a selective medium which contains a DHFR inhibitor such as methotrexate and aminopterin. If the cells have been successively cultured while gradually increasing the concentration of the DHFR inhibitor, the cells that can proliferate in the presence of the DHFR inhibitor at relatively higher concentrations are obtained. The cells selected as such a manner generally tend to express, at high levels, a gene encoding a protein of interest incorporated in the expression vector concomitantly with DHFR, and as a result, the mammalian cells thus will be obtained that express the protein of interest at high levels.

[0058] An expression vector has been known in which glutamine synthetase (GS), as a selection marker, is located downstream of a gene encoding a protein of interest via internal ribosome entry site (IRES), (International Patent Gazette; WO2012/063799, WO2013/161958). Expression vectors described in these literatures may be particularly preferable for the use in the method of production of the present invention.

[0059] For examples, an expression vector for expression of the protein can be preferably used in the method for production of the present inventions, that vector comprises a gene expression regulatory site, and a gene encoding the protein downstream thereof, an internal ribosome entry site further downstream thereof, a gene encoding a glutamine synthetase still further downstream thereof, and additionally a dihydrofolate reductase gene or a drug resistance gene downstream of either the same gene expression regulatory site or another gene

expression regulatory site in addition to the former. In this expression vector, as a gene expression regulatory site or another gene expression regulatory site, a cytomegalovirus-derived promoter, an SV40 early promoter, a human elongation factor-1 alpha promoter (hEF-1 alpha promoter), and a human ubiquitin C promoter are preferable, and the hEF-1 alpha promoter is particularly preferable.

[0060] As an internal ribosome entry site, those derived from 5' untranslated regions of viruses or genes selected from the group consisting of viruses of Picornaviridae, Picornaviridae Aphthovirus, hepatitis A virus, hepatitis C virus, coronavirus, bovine enterovirus, Theiler's murine encephalomyelitis virus, Coxsackie B virus, human immunoglobulin heavy chain binding protein gene, drosophila antennapedia gene, and drosophila Ultrabithorax gene may be preferably used. The internal ribosome entry site derived from 5' untranslated regions of mouse encephalomyocarditis virus may be particularly preferably used. When an internal ribosome binding site derived from the 5' untranslated region of the mouse encephalomyocarditis virus is used, not only the wild type internal ribosome binding site, but also those of which part of the multiple start codons contained in the wild type internal ribosome binding site has been disrupted may be preferably used. Further, as a drug resistance gene to be preferably used in this expression vector, puromycin or neomycin resistance gene is preferable, and puromycin resistance gene is more preferable.

[0061] Further, for examples, an expression vector for expressing the protein can be preferably used in the method for production of the

present inventions, that vector comprises hEF-1 α promoter, and a gene encoding the protein downstream thereof, an internal ribosome entry site derived from 5' untranslated regions of mouse encephalomyocarditis virus further downstream thereof, a gene
5 encoding a glutamine synthetase still further downstream thereof, and further another gene expression regulatory site and a dihydrofolate reductase gene thereof, wherein the internal ribosome binding site is that of which part of the multiple start codons contained in the wild type internal ribosome binding site has been disrupted. The expression
10 vectors described in WO2013/161958 are the examples of such vectors.

[0062] Further, for examples, an expression vector for expressing the protein can be preferably used in the method for production of the present inventions, that vector comprises human hEF-1 α promoter, and a gene encoding the protein downstream thereof, an internal ribosome
15 entry site derived from 5' untranslated regions of mouse encephalomyocarditis virus further downstream thereof, a gene encoding a glutamine synthetase still further downstream thereof, and further another gene expression regulatory site and a drug resistance gene downstream thereof, wherein the internal ribosome binding site is
20 that of which part of the multiple start codons contained in the wild type internal ribosome binding site has been disrupted. pE-mIRES-GS-puro described in WO2012/063799 and pE-mIRES-GS-mNeo described in WO2013/161958 are the examples of such vectors.

[0063] In the present invention, mammalian cells into which an
25 expression vector incorporating a gene encoding the fusion protein, an antibody, or a lysosomal enzyme has been introduced are subjected to a

selective culture in a selective medium to select the cells expressing the fusion protein, the antibody, or the lysosomal enzyme at high levels.

[0064] In performing a selective culture using DHFR as a selective marker, the concentration of the DHFR inhibitor in a selective medium is increased in a stepwise manner. When the DHFR inhibitor is methotrexate, the maximum concentration is preferably 0.25 to 5 μ M, more preferably 0.5 to 1.5 μ M, still more preferably about 1.0 μ M.

[0065] When GS used as a selection marker, the concentration of the GS inhibitor in a selective medium is increased in a stepwise manner.

When the GS inhibitor is MSX, the maximum concentration is preferably 100 to 1000 μ M, more preferably 200 to 500 μ M, and still more preferably about 300 μ M. And performing this, a medium not containing glutamine is generally used as the selective medium.

[0066] When using an enzyme degrading puromycin as a selection marker, the maximum concentration of puromycin contained in a selective medium is preferably 3 to 30 μ g/mL, more preferably 5 to 20 μ g/mL, and still more preferably about 10 μ g/mL.

[0067] When using an enzyme degrading neomycin as a selection marker, the maximum concentration of G418 contained in a selective medium is preferably 0.1 to 2 mg/mL, more preferably 0.5 to 1.5 mg/mL, and still more preferably about 1 mg/mL.

[0068] In addition, as a medium for culturing mammalian cells including the medium used for selective culture and the medium used for producing the fusion protein, an antibody, or a lysosomal protein (recombinant protein-production medium), both described later in detail, any medium can be used without particular limitation, as long as

they can be used for culturing and growing mammalian cells, but a serum-free medium is preferably used.

[0069] The cells selected by the selective culture and showing a high expression level of the fusion protein, an antibody, or a lysosomal protein are used for their production as producing cells thereof. The production of the fusion protein, an antibody, or a lysosomal protein is carried out by culturing producing cells thereof in a recombinant protein-production medium. This culture is called production culture.

[0070] In the present invention, an example of serum-free media which is to be used as a recombinant protein-production medium is the medium which contains; 3 to 700 mg/mL of amino acids, 0.001 to 50 mg/L of vitamins, 0.3 to 10 g/L of monosaccharides, 0.1 to 10000 mg/L of inorganic salts, 0.001 to 0.1 mg/L of trace elements, 0.1 to 50 mg/L of nucleosides, 0.001 to 10 mg/L of fatty acids, 0.01 to 1 mg/L of biotin, 0.1 to 20 micrograms/L of hydrocortisone, 0.1 to 20 mg/L of insulin, 0.1 to 10 mg/L of vitamin B₁₂, 0.01 to 1 mg/L of putrescine, 10 to 500 mg/L of sodium pyruvate, and water-soluble iron compounds. As desired, it may also include thymidine, hypoxanthine, a conventional pH indicator, and antibiotics.

[0071] Further, as a serum-free medium used for the production of recombinant protein, DMEM/F12 medium, a mixed medium comprising DMEM and F12, may also be used as a basic medium. Each of these media is well known to those skilled in the art. Furthermore, as a serum-free medium, DMEM(HG)HAM modified (R5) medium may be used, too, which contains sodium hydrogen carbonate, L-glutamine, D-glucose, insulin, sodium selenite, diaminobutane, hydrocortisone,

ferric (II) sulfate, asparagine, aspartic acid, serine, and polyvinyl alcohol. Furthermore, a commercially available serum-free medium may also be used as a basic medium, including CD OptiCHO™ medium, CHO-S-SFM II medium, or CD CHO medium (Thermo Fisher Scientific Inc.), EX-CELL™ 302 medium or EX-CELL™ 325-PF medium (SAFC Biosciences Inc). For example, EX-CELL™ Advanced CHO Fed-batch medium (SAFC Biosciences) which is a serum-free medium containing 16 μmol/L thymidine, 100 μmol/L hypoxanthine, and 4 mmol/L L-alanyl-L-glutamine may be preferably used for culturing the fusion protein-producing cells.

[0072] In the production culture of the cells producing the fusion protein, an antibody, or a lysosomal protein, the density of the producing cells thereof in the medium for recombinant protein-production is preferably adjusted to 0.2×10^5 to 5×10^5 cells/mL, more preferably 1×10^5 to 4×10^5 cells/mL, still more preferably about 2×10^5 cells/mL, when starting the culture.

[0073] Production culture has been performed while observing the cell viability (%) over time, so that the cell survival rate during the production culture is maintained preferably at 85% or more, more preferably 90% or more.

[0074] During the production culture, the culture temperature is maintained preferably at 33.5 to 37.5°C, and the dissolved oxygen saturation level during the production medium is maintained preferably at 38 to 42%, more preferably at about 40%. Here, the term “dissolved oxygen saturation level” means the dissolution amount of oxygen when the saturated dissolution amount of oxygen is taken as 100% under

same conditions.

[0075] During the production culture, the production medium is stirred with an impeller (impeller). At this time, the rotational speed of the impeller is adjusted preferably to 67 to 72 rotations per minute, more preferably to 70 rotations per minute, but the rotational speed may be changed as needed depending on the shape of the impeller or the like.

[0076] Suitable culture conditions for the production culture at the early stage include, for example, such a condition in that the density of the recombinant protein-producing cells in the medium for recombinant protein production is 2×10^5 cells/mL; the culture temperature during the production culture period is maintained at 34 to 37°C; the dissolved oxygen saturation level in the production medium is 40%; and the medium is agitated with an impeller rotating at a speed of about 89 rpm.

[0077] After completion of the production culture, the culture medium is collected. The culture supernatant is obtained by centrifuging or filtrating the collected culture. The desired fusion protein contained in the culture supernatant can be purified by a process using various chromatographies. The purification process can be carried out at room temperature or low temperature environment, but carried out preferably under a low temperature environment, and particularly preferably at a temperature of 1 to 10°C.

[0078] Hereinafter, the purification method of the fusion protein between the antibody and a human lysosomal enzyme contained in the culture supernatant is described in detail.

[0079] A step of the purification process is a column chromatography employing as solid phase a substance having affinity for the fusion

protein. In this step, there is no particular limitation as to the material having an affinity for the fusion protein, but preferable are protein A, protein G, protein L, protein A/G, and an antibody that recognizes the antibody constituting the fusion protein as an antigen, an antigen which is recognized by the antibody constituting the fusion protein, more preferable is protein A. The combination thereof can also be used. By loading the culture supernatant, the fusion protein contained in the culture supernatant is let bind to the column, and after washing the column, the fusion protein is eluted from the column. Thus, most of the contaminants can be removed. When the antibody constituting the fusion protein is human IgG, the antibody recognizing the antibody constituting the fusion protein as an antigen is an anti-human IgG antibody.

[0080] Among substances having an affinity for the fusion protein described above, protein A, protein G, protein L, protein A/G, and an antibody that recognizes the antibody constituting the fusion protein as an antigen can be viewed as substances having an affinity for the fusion protein, but also as substances having an affinity for antibodies.

[0081] Protein A is a protein having a molecular weight of about 42 kD, present on the cell wall of *Staphylococcus aureus*.

Protein A can specifically bind to the Fc region of human antibodies (or humanized antibodies) of the IgG1, IgG2 and IgG4 type. Protein A can also bind to the Fab region of IgG belonging to the VH3 subfamily. Accordingly, protein A can be used when an antibody constituting a part of the fusion protein to be purified has an Fc region and is a human antibody (or humanized antibody) of IgG1, IgG2 and IgG4 type. When

the antibody constituting a part of the fusion protein to be purified is Fab, F(ab'), or F(ab')² of a human antibody (or humanized antibody) belonging to the VH3 subfamily, Protein A can also be used.

[0082] Protein A used herein is not limited to wild-type protein A as long as it has the desired affinity for the antibody, but includes a mutant type protein A, wherein 1 to 10 amino acids substitution, deletion, or addition have been introduced into the amino acid sequence of the wild-type protein A. Furthermore, it may be a peptide containing a partial sequence of the amino acid sequence of wild-type or mutant type protein A as long as it has the desired affinity for the antibody. Such a partial sequence includes a domain that binds to the antibody.

[0083] Protein Gs are the proteins constituting *streptococcus* itself, and of those, "G148 protein G" having a molecular weight of about 65 kD and "C40 protein G" having a molecular weight of about 58 kD are particularly well known. Protein G can specifically bind to human antibodies (or humanized antibodies) of the IgG1, IgG2, IgG3 and IgG4 type. Accordingly, when the antibody constituting a part of the fusion protein to be purified is of the IgG3 type which does not bind to Protein A, the fusion protein can be purified by using Protein G.

[0084] The protein G used here is not limited to the wild type protein G, but a mutant type protein G containing 1 to 10 of amino acid substitution, deletion, or addition in the amino acid sequence may also be used as long as it has the desired affinity for the antibody. Furthermore, a peptide containing a partial sequence of the amino acid sequence of wild-type or a mutant type protein A may also be used as long as it has the desired affinity for the antibody. Such a partial

sequence contains an antibody-binding region. The wild-type protein G has an albumin binding region, and a mutant type protein G in which such a region is deleted can be particularly suitably used in the present invention.

5 [0085] Protein L is one of a protein constituting the bacterial body of *Peptostreptococcus magnus*. Protein L can bind specifically to the κ chain of a light chain of a human antibody (or a humanized antibody) belonging to κ I, κ III and κ IV subtypes. Therefore, when an antibody constituting a part of the fusion protein to be purified has a light chain
10 belonging to these subtypes, the protein L can be used even if the antibody is Fab or ScFv.

[0086] The protein L used here is not limited to the wild-type protein L, but a mutant type protein L containing 1 to 10 of amino acid substitution, deletion, or addition in the amino acid sequence may also
15 be used as long as it has the desired affinity for the antibody. Furthermore, a peptide containing a partial sequence of the amino acid sequence of wild-type or a mutant type protein A may also be used as long as it has the desired affinity for the antibody. Such a partial sequence contains an antibody-binding region. The wild-type protein L
20 has an albumin binding region, and a mutant type protein L in which such a region is deleted can be particularly suitably used in the present invention.

[0087] Protein A/G is an artificial protein produced by combining four Fc binding regions of protein A and two Fc binding regions of protein G.
25 Protein A/G has both properties of Protein G and Protein A, and it is possible to purify not only an antibody that can be purified by Protein A

and but also an antibody that can be purified by Protein G.

[0088] When the antibody constituting a part of the fusion protein is human IgG, the antibody which recognizes an antibody constituting a part of a fusion protein as an antigen is an anti-human IgG antibody specifically binding to this human IgG. Such an anti-human IgG antibody can be prepared as a monoclonal antibody or as a polyclonal antibody by immunizing an animal with an antibody constituting a fusion protein or a part thereof as an antigen.

[0089] A substance that is recognized by the antibody constituting a part of the fusion protein as an antigen is an extracellular region of TfR, insulin receptor, leptin receptor, Lipoprotein receptor, IGF receptor, OATP-F, organic anion transporter, MCT-8, and Fc receptor, when the antibody is an anti-transferrin receptor (TfR) antibody, an anti-insulin receptor antibody, an anti-leptin receptor antibody, an anti-lipoprotein receptor antibody, an anti-IGF receptor antibody, an anti-OATP-F antibody, an anti-organic anion transporter antibody, an anti-MCT-8 antibody, an anti-monocarboxylic acid transporter antibody, and an Fc receptor antibody.

[0090] Another step of the purification process is a column chromatography employing as solid phase a substance having affinity for the phosphate group. There is no particular limitation as to the solid phase having an affinity for the phosphate group used for this, but hydroxyapatite and fluoroapatite are preferable, and hydroxyapatite is particularly preferable. It is preferable that the pH of the solution containing the fusion protein and loaded on the column chromatography is adjusted to 6.8 to 7.8 before loaded.

[0091] In the step of the purification process above, the fusion protein is let bound to the solid phase equilibrated by a buffer solution having pH near neutral and containing a salt and phosphate. The buffer solution used for this is preferably MES buffer, and its pH is preferably 6.8 to 7.8. Though there is no particular limitation for the salt contained in the buffer solution, sodium chloride is preferable, and its concentration is preferably 70 to 230 mM, more preferably 160 to 220 mM. The concentration of phosphate contained in the buffer solution is preferably 0.2 to 4.0 mM, more preferably 1 to 2.5 mM.

[0092] After washing the column to which the fusion protein is bound, the fusion protein is eluted from the column with a buffer solution having pH near neutral and containing a salt, and the fraction containing the fusion protein is recovered. The buffer solution used for this is preferably a phosphate buffer solution, and its pH is preferably 6.8 to 7.8. The concentration of phosphate contained in the buffer solution is preferably 10 to 50 mM, more preferably 20 to 40 mM. Though there is no particular limitation for the salt contained in the buffer solution, sodium chloride is preferable, and its concentration is preferably 70 to 230 mM, more preferably 160 to 220 mM. The concentration of phosphate contained in the buffer solution is preferably 0.2 to 4.0 mM.

[0093] Further another step of the purification process is a size exclusion column chromatography, which is a step for removing low molecular-weight impurities such as endotoxin, as well as multimeric complexes or decomposition products of the fusion protein. Thus, substantially pure fusion protein is obtained through this.

[0094] In the purification process of the fusion protein, a step for

inactivating the virus that may be brought from the culture supernatant may optionally be added. Such an additional step for virus inactivation may be conducted prior the purification, and may be interposed between any two adjacent steps of the purification process. For example, when
5 the purification process includes a column chromatography employing as solid phase a material coupled with a substance having affinity for the fusion protein (the first step of the purification process), a column chromatography employing as solid phase a material having affinity for phosphate group (the second step of the purification process), and a size
10 exclusion column chromatography (the third step of the purification process) in this order, the step for virus inactivation preferably be interposed between the first and the second step of the purification processes.

[0095] The virus inactivation step is conducted by adding a nonionic
15 surfactant to a solution containing the fusion protein and stirring at 20 to 60°C for 2 to 6 hours. Preferable examples of the nonionic surfactant used for this include polysorbate 20, 80, and tri n-butylphosphate, or a mixture thereof.

[0096] The virus inactivation step can also be carried out using a virus
20 removal membrane. Viruses contained in the solution can be removed by passing the solution containing the fusion protein through the virus removal membrane with a pore size of 35 nm or 20 nm.

[0097] The purified product of the fusion protein obtained by using the
25 production method of the present invention is of such purity as is sufficient for its direct use as a medical drug. The concentration of host cell-derived proteins (HCP) contained in the purified product of the

fusion protein is preferably less than 300 ppm, more preferably less than 100 ppm, for example less than 60 ppm. In addition, the proportion of the multimer in the whole fusion protein contained in the purified product of the fusion protein is preferably less than 1%.

5 [0098] When a purified product of a fusion protein obtained by using the production method of the present invention is provided as a medical drug, it can be provided in such a form as an aqueous preparation or a lyophilized preparation, containing an appropriate excipient. In the case of preparing the aqueous preparation, it may be filled into a vial, or it
10 may be provided as a prefilled-type preparation filled in a syringe in advance. In the case of a freeze-dried preparation, it has been dissolved with an aqueous solution before use.

[0099] When the purified product of the fusion protein is administered as a medicament to a human, it may be administered, for example,
15 intravenously, intramuscularly, subcutaneously, intraperitoneally, intraarterially, or intralesionally, but preferably administered intravenously.

[0100] Further, as the purified product of the fusion protein can pass through the BBB when administered to a human, it can be used as a
20 therapeutic agent for various diseases accompanying central nervous system disorder. By administering the fusion protein, the central nervous system disorder can be prevented, ameliorated, or its progression can be delayed.

[0101] Hereinafter, described in detail is a fusion protein (humanized
25 anti-hTfR antibody-I2S), wherein the light chain of the humanized anti-hTfR antibody has the amino acid sequence set forth as SEQ ID

NO:10, and wherein the heavy chain of the humanized anti-hTfR antibody is linked, on the C-terminal side thereof and via a linker sequence set forth as Gly-Ser, to the human iduronate-2-sulfatase, and the whole linked heavy chain has the amino acid sequence set forth as SEQ ID NO:14. The antibody constituting a part of this fusion protein is of the IgG1 type.

[0102] The first step of the purification process is column chromatography employing as a solid phase a material letting a substance having an affinity for the fusion protein bound thereto. Substances having an affinity for the antibody used for this are not particularly limited, but preferred are protein A, protein G, protein L, protein A/G, anti-human IgG1 antibody, hTfR which is an antigen of the antibody, or anti-human I2S antibody, more preferably Protein A. In the case where the substance is hTfR, it is an extracellular region thereof.

[0103] When Protein A is employed as a substance having an affinity for the fusion protein in the first step, the culture supernatant containing the fusion protein is bound to a column equilibrated in advance with a buffer containing a neutral solution containing a salt. The buffer solution used for this is preferably a trometamol buffer, and its pH is preferably 6.5 to 7.5, more preferably about 7.0. Although there is no particular limitation as to the salt contained in the buffer, sodium chloride is preferable, and its concentration is preferably 60 to 180 mM, more preferably 100 to 150 mM, and still more preferably about 140 mM.

[0104] After washing the column to which the fusion protein is bound, the fusion protein is eluted with an acidic buffer containing salt, and the

fraction containing the fusion protein is collected. The buffer solution used for this is preferably a glycine buffer, and its pH is preferably 3.2 to 3.8, more preferably 3.5. Although there is no particular limitation as to the salt contained in the buffer, sodium chloride is preferable, and its concentration is preferably 60 to 180 mM, more preferably 100 to 150 mM, and still more preferably about 140 mM. The pH of the solution containing the recovered fusion protein is rapidly adjusted so as to become around neutral.

[0105] The second step of the purification process is column chromatography using a material having affinity for the phosphate group as the solid phase. There is no particular limitation as to the solid phase having an affinity for the phosphoric acid group employed for this, but hydroxyapatite and fluoroapatite are preferable, and hydroxyapatite is particularly preferable.

[0106] In the second step of the purification process, when hydroxyapatite is employed as the solid phase having affinity for the phosphate group, the fusion protein is let bound to the solid phase equilibrated with a buffer at or near neutral pH containing salt and phosphoric acid. The buffer solution used for this is preferably an MES buffer, the pH of which is preferably 6.8 to 7.8, more preferably 7.3. Although there is no particular limitation as to the salt contained in the buffer, sodium chloride is preferred, and its concentration is preferably 150 to 230 mM, more preferably 215 mM. The concentration of phosphoric acid contained in the buffer is preferably 1.0 to 4.0 mM, more preferably 2.0 mM.

[0107] After washing the column to which the fusion protein is bound,

the fusion protein is eluted from the column with a buffer at or near neutral pH containing a salt, and the fraction containing the fusion protein is collected. The buffer solution used for this is preferably a phosphate buffer, the pH of which is preferably 6.8 to 7.8, more preferably pH 7.3. The concentration of phosphoric acid contained in the buffer is preferably 30 to 50 mM, more preferably about 35 mM. Although there is no particular limitation as to the salt contained in the buffer, sodium chloride is preferred, and its concentration is preferably 150 to 230 mM, more preferably 215 mM.

[0108] The third step of the purification process is size exclusion column chromatography. This step is for removing low-molecular impurities such as endotoxin, multimers and degradation products of the fusion protein and the like, whereby a substantially pure fusion protein can be obtained.

[0109] In the purification process of the humanized anti-hTfR antibody-I2S, a step for inactivating the virus possibly brought from the culture supernatant may be added. This virus inactivation step may be carried out before the first step of the purification process, between any of each step in the purification process, or after completion of the purification process. For example it can be carried out prior to the first step of the purification processor between the first step and the second step of the purification process.

[0110] The virus inactivation step is carried out by adding a nonionic surfactant to a solution containing humanized anti-hTfR antibody-I2S and stirring at 20 to 60°C for 2 to 6 hours. Preferable examples of the nonionic surfactant used for this include polysorbate 20, 80, and tri

n-butylphosphate, or a mixture thereof.

[0111] The virus inactivation step may also be carried out using a virus removal membrane. By passing a solution containing humanized anti-hTfR antibody-I2S through a virus removal membrane with a pore size of 35 nm or 20 nm, the virus contained in the solution can be removed.

[0112] A purified product of the humanized anti-hTfR antibody-I2S obtained by using the method for production of the present invention is of purity that can be used as it is as a medicine. The concentration of the host cell-derived protein (HCP) contained in the purified product of the humanized anti-hTfR antibody-I2S is less than 100 ppm, for example less than 60 ppm, less than 40 ppm, or the like. Also, the proportion of the polymer in the whole humanized anti-hTfR antibody-I2S contained in the purified product of the humanized anti-hTfR antibody-I2S is less than 1%, for example less than 0.8%, less than 0.6%, less than 0.5%, and so on.

[0113] When a purified product of the humanized anti-hTfR antibody-I2S obtained by using the method for production of the present invention is supplied as a medicine, it can be supplied as an aqueous liquid preparation or a freeze-dried preparation containing an appropriate excipient. In the case of preparing an aqueous liquid preparation, it may be filled into a vial, or it may be supplied as a prefilled type preparation filled in advance in a syringe. In the case of a freeze-dried preparation, it is used by being dissolved in an aqueous medium before use.

[0114] When the purified humanized anti-hTfR antibody-I2S is

administered as a pharmaceutical to humans, it can be administered, for example, intravenously, intramuscularly, subcutaneously, intraperitoneally, intraarterially, or intralesionally. For example, the purified product can be intravenously administered by drip infusion.

5 [0115] Further, the purified product of humanized anti-hTfR antibody-I2S can be used as a therapeutic agent for Hunter's syndrome, particularly Hunter's syndrome accompanied by central nervous disorder. The humanized anti-hTfR antibody-I2S administered to patients with Hunter's syndrome degrades glycosaminoglycans (GAG)
10 accumulated in organs of patients, and furthermore degrade GAG accumulated in the brain tissues by passing through BBB. Therefor it can prevent, ameliorate, or delay the progress of central nervous disorders accompanying Hunter's syndrome.

Examples

15 [0116] While the present invention will be described in further detail below referring to examples, it is not intended that the present invention be limited to the examples.

[0117] [Example 1] Construction of expression vector for hI2S-humanized anti-hTfR antibody fusion protein

20 An expression vector for hI2S-humanized anti-hTfR antibody fusion protein was constructed using genes encoding three types of humanized anti-hTfR antibodies (Nos. 1 to 3). The antibody No. 1 comprises a light chain having the amino acid sequence set forth as SEQ ID NO:6 and a heavy chain having the amino acid sequence set forth as
25 SEQ ID NO:7, the antibody No. 2 comprises a light chain having the amino acid sequence set forth as SEQ ID NO: 8 and a heavy chain

having the amino acid sequence set forth as SEQ ID NO: 9, the antibody No. 3 comprises a light chain having the amino acid sequence set forth as SEQ ID NO: 10 and a heavy chain having the amino acid sequence set forth as SEQ ID NO: 11, respectively.

5 [0118] A pEF/myc/nuc vector (Invitrogen Inc.) was digested with KpnI and NcoI to cut out the region containing the EF-1 α promoter and its first intron, and the region was blunt-ended with T4 DNA polymerase. A pCI-neo vector (Invitrogen) was digested with BglIII and EcoRI to cut out the region containing the enhancer/promoter and intron of CMV,
10 and then the region was blunt-ended with T4 DNA polymerase. The above region containing the EF-1 α promoter and its first intron was inserted into this to construct a pE-neo vector. The pE-neo vector was digested with SfiI and BstXI and a region of approximately 1 kbp containing the neomycin resistance gene was cut out. Amplification of
15 hygromycin gene was carried out by PCR reaction using primers Hyg-Sfi5' (SEQ ID NO:27) and Hyg-BstX3' (SEQ ID NO:28) and using pcDNA 3.1/Hygro(+) vector (Invitrogen Inc.) as a template. The amplified hygromycin gene was digested with SfiI and BstXI and inserted into the pE-neo vector from which the above neomycin
20 resistance gene has been cut out to construct a pE-hygr vector. A method for constructing the pE-hygr vector is also disclosed in Patent Document (JP2009-273427A).

[0119] A DNA fragment set forth as SEQ ID NO:21 and containing the gene encoding the full length of the light chain of the humanized
25 anti-hTfR antibody No. 1 having the amino acid sequence set forth as SEQ ID NO:6 was artificially synthesized. A MluI sequence was

introduced on the 5' side of this DNA fragment and a NotI sequence on the 3' side thereof. This DNA fragment was digested with MluI and NotI and incorporated between MluI and NotI of the pE-neo vector. The obtained vector was designated pE-hygr(LC1) which is a vector for expressing the light chain of humanized anti-hTfR antibody No. 1.

[0120] A DNA fragment (SEQ ID NO: 22) containing a gene encoding the full length of the light chain of humanized anti-hTfR antibody No. 2 having the amino acid sequence set forth as SEQ ID NO:8 was artificially synthesized. The MluI sequence was introduced on the 5' side of this DNA fragment and the NotI sequence on the 3' side thereof. This DNA fragment was digested with MluI and NotI and incorporated between MluI and NotI of the pE-neo vector. The resulting vector was designated pE-hygr(LC 2) which is a vector for expressing the light chain of humanized anti-hTfR antibody No. 2.

[0121] A DNA fragment (SEQ ID NO: 23) containing a gene encoding the full length of the light chain of humanized anti-hTfR antibody No. 3 having the amino acid sequence set forth as SEQ ID NO: 10 was artificially synthesized. The MluI sequence was introduced on the 5' side of this DNA fragment and the NotI sequence on the 3' side thereof. This DNA fragment was digested with MluI and NotI and incorporated between MluI and NotI of the pE-neo vector. The obtained vector was defined as pE-hygr(LC3) which is a vector for expressing the light chain of humanized anti-hTfR antibody No. 3.

[0122] A DNA fragment was artificially synthesized, having a nucleotide sequence set forth as SEQ ID NO:24 containing a gene encoding a protein in which hI2S having an amino acid sequence set

forth as SEQ ID NO:5 is linked to the C-terminal side of the heavy chain of the humanized anti-hTfR antibody No.1 having an amino acid sequence set forth as SEQ ID NO:7 via a linker having an amino acid sequence set forth as (Gly-Ser). This DNA fragment encodes a protein having the amino acid sequence set forth as SEQ ID NO:12, in which a heavy chain of humanized anti-hTfR antibody No.1 binds to hI2S. This DNA fragment was digested with MluI and NotI and inserted between MluI and NotI of the pE-neo vector to construct pE-neo (HC-I2S-1).

[0123] A DNA fragment having a nucleotide sequence set forth as SEQ ID NO:25 containing a gene encoding a protein in which hI2S having an amino acid sequence set forth as SEQ ID NO:5 is linked to the C-terminal side of the heavy chain of the humanized anti-hTfR antibody No. 2 having an amino acid sequence set forth as SEQ ID NO:9 via a linker having an amino acid sequence set forth as (Gly-Ser) was artificially synthesized. This DNA fragment encodes a protein having the amino acid sequence set forth as SEQ ID NO:13, in which a heavy chain of humanized anti-hTfR antibody No.2 binds to hI2S. This DNA fragment was digested with MluI and NotI and integrated between MluI and NotI of the pE-neo vector to construct pE-neo (HC-I2S-2).

[0124] A DNA fragment having a nucleotide sequence set forth as SEQ ID NO:26 containing a gene encoding a protein in which hI2S having an amino acid sequence set forth as SEQ ID NO:5 is linked to the C-terminal side of the heavy chain of the humanized anti-hTfR antibody No. 3 having an amino acid sequence set forth as SEQ ID NO:11 via a linker having an amino acid sequence set forth as (Gly-Ser) was artificially synthesized. This DNA fragment encodes a protein having

the amino acid sequence shown in SEQ ID NO:14, in which a heavy chain of humanized anti-hTfR antibody No.3 binds to hI2S. This DNA fragment was digested with MluI and NotI and integrated between MluI and NotI of the pE-neo vector to construct pE-neo (HC-I2S-3).

5 [0125] [Example 2-1] Preparation of a high expression cell lines of hI2S-humanized anti-hTfR antibody fusion proteins

CHO cells (CHO-K1 obtained from American Type Culture Collection) were transformed with combinations of pE-hygr (LC1) and pE-neo (HC-I2S-1) constructed in Example 1, pE-hygr (LC2) and
10 pE-neo (HC-I2S-2) constructed in Example 1 and pE-hygr (LC3) and pE-neo (HC-I2S-3) constructed in Example 1, respectively, using the GenePulser (Bio-Rad Inc.).

[0126] Transformation of cells was in brief carried out by the following method. 5×10^5 CHO-K1 cells were seeded in a 3.5 cm culture dish to which CD OptiCHO™ medium (Thermo Fisher Scientific Inc.) was
15 added and cultured overnight at 37 °C under 5% CO₂. After the culture, the cells were suspended in Opti-MEM™I medium (Thermo Fisher Scientific Inc.) to a density of 5×10^6 cells/mL. 100 µL of the cell suspension was collected, and thereto 5 µL each of the pE-hygr (LC1) and pE-neo (HC-I2S-1) plasmid DNA solutions both having been
20 diluted to 100 µg/mL with CD OptiCHO™ medium was added. Electroporation was performed using GenePulser (Bio-Rad Inc.), and plasmids were introduced into the cells. After overnight culture under the condition of 37 °C, 5% CO₂, the cells were selectively cultured in
25 CD OptiCHO™ medium supplemented with 0.5 mg/mL of hygromycin and 0.8 mg/mL of G418. For the combination of pE-hygr (LC2) and

pE-neo (HC-I2S-2) and the combination of pE-hygr (LC3) and pE-neo (HC-I2S-3), the transformations of the cells were conducted by the same method.

[0127] Then, the cells selected above through the selection culture were seeded on 96-well plates so that not more than one cell might be seeded per well by limiting dilution. The cells then were cultured for about 10 days so that monoclonal colonies were formed. Respective culture supernatants of the wells in which monoclonal colony was formed were collected, the amount of the humanized antibody contained in culture supernatants was determined by ELISA, and the hI2S-humanized anti-hTfR antibody fusion protein high-expressing cell lines were selected.

[0128] The ELISA above was conducted as follows in general. To each well of 96-well microtiter plates (Nunc Inc.) were added 100 μ L, of a goat anti-human IgG polyclonal antibody solution diluted with 0.05 M sodium bicarbonate buffer (pH 9.6) to 4 μ g/mL, and the plate was left to stand for at least one hour at room temperature so as to allow the antibody to be adsorbed by the plates. Then, after each well was washed three times with a phosphate-buffered saline (pH 7.4) supplemented with 0.05% Tween20 (PBS-T), 200 μ L, of Starting Block (PBS) Blocking Buffer (Thermo Fisher Scientific Inc.) was added to each well, and the plates were left to stand for 30 minutes at room temperature. After each well was washed with PBS-T three times, the culture supernatant or the human IgG reference standard product which had been diluted with a phosphate buffer saline (pH 7.4) supplemented with 0.5% BSA and 0.05% Tween20 (PBS-BT) to appropriate

concentrations, was added to each well, in the amount of 100 μ L, and the plates were left to stand for at least one hour at room temperature. After the plates were washed three times with PBS-T, 100 μ L of HRP-labeled anti-human IgG polyclonal antibody solution which had been diluted with PBS-BT, was added to each well, and the plates were left to stand for at least one hour at room temperature. After the wells were washed three times with PBS-T, 0.4 mg/mL o-phenylenediamine in citrate-phosphate buffer (pH 5.0) was added to each well, in the amount of 100 μ L, and the wells were left to stand for 8 to 20 minutes at room temperature. Then, 1 mol/L sulfuric acid was added to each well, in the amount of 100 μ L to terminate the reaction, and the absorbance for each well was measured at 490 nm using a 96-well plate reader. The cells corresponding to the wells which exhibited the higher measurements were regarded as a high-expressing cell line for hI2S-humanized anti-hTfR antibody fusion protein.

[0129] A high-expressing cell line of a hI2S-humanized anti-hTfR antibody fusion protein obtained by transformation with combination of pE-hygr(LC1) and pE-neo(HC-I2S-1) was designated as a hI2S-anti-hTfR antibody expressing strain 1. The fusion protein of hI2S and humanized anti-hTfR antibody expressed by this cell line was designated as I2S-anti-hTfR antibody 1.

[0130] A high-expressing cell line of a hI2S-humanized anti-hTfR antibody fusion protein obtained by transformation with combination of pE-hygr(LC2) and pE-neo(HC-I2S-2) was designated as a hI2S-anti-hTfR antibody expressing strain 2. The fusion protein of hI2S and humanized anti-hTfR antibody expressed by this cell line was

designated as I2S-anti-hTfR antibody 2.

[0131] A high-expressing cell line of a hI2S-humanized anti-hTfR antibody fusion protein obtained by transformation with combination of pE-hygr(LC3) and pE-neo(HC-I2S-3) was designated as a
5 hI2S-anti-hTfR antibody expressing strain 3. The fusion protein of hI2S and humanized anti-hTfR antibody expressed by this cell line was designated as I2S-anti-hTfR antibody 3.

[0132] [Example 3] Preparation of high expression cell line of hI2S-Humanized anti-hTfR antibody fusion protein

10 The hI2S-anti-hTfR antibody expressing strains 1 to 3 obtained in Example 2 were suspended in CD OptiCHO™ medium containing 10 mg/L insulin, 40 mg/mL thymidine, and 10%(v/v) DMSO, and dispensed into cryotubes and stored as seed cells in liquid nitrogen.

[0133] [Example 4] Culture of hI2S-anti-hTfR antibody expressing
15 strain

The hI2S-anti-hTfR antibodies were produced by the method described below. The hI2S-anti-hTfR antibody expressing strain 3 obtained in Example 2-1 was suspended in about 200 L of serum-free medium (EX-CELL Advanced CHO Fed-batch Medium, Sigma Aldrich
20 Inc.) containing 4 mM L-alanyl-L-glutamine, 100 μmol/L hypoxanthine and 16 μmol/L thymidine to the density of about 2×10^5 cells/mL. 140 L of this cell suspension was transferred to a culture tank. The medium was stirred with an impeller at a rate of 89 rpm, the dissolved oxygen saturation of the medium was kept at about 40%, and the cells were
25 cultured for about 11 days at a temperature range of 34 to 37 °C. During the culture period, cell number, cell viability, medium glucose

concentration, and lactate concentration were monitored. When the glucose concentration of the medium became less than 15 mmol/L, the glucose solution was added to the medium immediately so that the glucose concentration became 37.89 mmol/L. After completion of the culture, the medium was collected. The recovered medium was filtered with Millistak+HC Pod Filter grade D0HC (Merck Inc.) and further filtered with Millistak+HC grade X0HC (Merck Inc.) to obtain a culture supernatant containing I2S-anti-hTfR antibody 3. The culture supernatant was subjected to ultrafiltration using a Pellicon™ 3 Cassette w/Ultracel PLCTK Membrane (pore size: 30 kDa, membrane area: 1.14 m², Merck Inc.) and concentrated until the liquid volume was about 1/17. The concentrate was then filtered using OpticapXL600 (0.22 μm, Merck Inc.). The obtained solution was used as a concentrated culture supernatant.

[0134] [Example 5] Virus inactivation

To the concentrated culture supernatant obtained in Example 4, tri-n-butyl phosphate (TNBP) and polysorbate 80 were added so that the final concentrations were 0.3% (v/v) and 1% (w/v), respectively, and gently stirred at room temperature for 4 hours. This procedure is conducted for inactivating the virus contaminating the culture supernatant. However, as long as culturing is carried out using a serum-free medium not containing biological components, there is little possibility that viruses harmful to the human body are contaminated in the culture supernatant.

[0135] [Example 6] Purification of hI2S-anti-hTfR antibody

The concentrated culture supernatant after the virus inactivation

was added to a Millipak-200 Filter Unit (pore size: 0.22 μ m, Merck Inc.) after adding thereto 20 mM Tris-HCl buffer (pH 7.0) containing 0.5 volume of 140 mM NaCl. The solution after filtration was loaded onto a MabSelect SuRe LX column (column volume: about 3.2 L, bed height: about 20 cm, GE Healthcare Inc.), which was a protein A affinity column, equilibrated with 4 column volumes of 20 mM Tris-HCl buffer (pH 7.0) containing 140 mM NaCl, at a constant flow rate of 200 cm/hr to adsorb I2S-anti-hTfR antibody 3 to protein A.

[0136] Subsequently, the column was washed with 5 column volumes of 10 mM Tris-HCl buffer (pH 7.0) containing 500 mM NaCl and 450 mM arginine at the same flow rate. Then the column was further washed with 2.5 column volumes of 20 mM Tris-HCl buffer (pH 7.0) containing 140 mM NaCl at the same flow rate. Then I2S-anti-hTfR antibody 3 adsorbed to Protein A was eluted with 5 column volumes of 100 mM glycine buffer (pH 3.5) containing 140 mM NaCl. The eluate was immediately neutralized by receiving in a container containing 1 M Tris-HCl buffer (pH 7.5) in advance.

[0137] To the above eluate from the Protein A affinity column, 200 mM phosphate buffer (pH 7.0), 10 mM MES buffer (pH 7.3) containing 4 M NaCl and 2 mM phosphate buffer, and 1 M Tris-HCl buffer solution (pH 8.0) were added in the order, and the concentrations of sodium phosphate and NaCl contained in the eluate were adjusted to 2 mM and 215 mM, respectively, and the pH of the eluate was adjusted to 7.3. The eluate was then filtered through Opticap XL 600 (pore size: 0.22 μ m, Merck Inc.). The solution after filtration was applied to a CHT Type II 40 μ m column, a hydroxyapatite column (Column volume: about 3.2 L,

bed height: about 20 cm, Bio-Rad Inc.), equilibrated with 4 column volumes of 10 mM MES buffer solution (pH 7.3) containing 215 mM NaCl and 2 mM sodium phosphate at a constant flow rate of 200 cm/hr to adsorb I2S-anti-hTfR antibody 3 to hydroxyapatite.

5 [0138] Subsequently, the column was washed with 5 column volumes of the same buffer at the same flow rate. Then I2S-anti-hTfR antibody 3 adsorbed on hydroxyapatite was eluted with 5 column volumes of 35 mM phosphate buffer (pH 7.3) containing 215 mM NaCl. Purification by the hydroxyapatite column was carried out in two portions using half
10 volume of the eluate from the protein A affinity column.

[0139] To the above eluate from the hydroxyapatite column, dilute hydrochloric acid was added to adjust the pH to 6.5. Then, ultrafiltration was carried out using Pellicon™ 3 Cassette w/Ultracel PLCTK Membrane (pore size: 30 kDa, membrane area: 1.14 m², Merck Inc.) to
15 concentrate I2S-anti-hTfR antibody 3 in the solution at the concentration of about 2 mg/mL. The concentrate was then filtered using Opticap XL 600 (0.22 µm, Merck Inc.).

[0140] The above concentrated solution was applied to a Superdex 200 column, size exclusion column (Column volume: about 12.6 L, bed
20 height: 40 cm, GE Healthcare Inc.) equilibrated with 5 column volumes of 20 mM phosphate buffer (pH 6.5) containing 0.8 mg/mL NaCl and 75 mg/mL sucrose at a constant flow rate of 19 cm/hr, and the same buffer was supplied at the same flow rate. At this time, an absorbance photometer for continuously measuring the absorbance of the eluate was
25 placed in the flow path of the eluate from the size exclusion column, and the absorbance at 280 nm was monitored, the fractions which

corresponded to an absorption peak at 280 nm were collected as a fractions containing I2S-anti-hTfR antibody 3, which was used as a purified product of I2S-anti-hTfR antibody. Purification on the size exclusion column was carried out in two portions using half volume of the eluate from the hydroxyapatite column.

[0141] [Example 7] Measurement of recovery rate of I2S-anti-hTfR antibody in each purification step

The amount of I2S-anti-hTfR antibody 3 loaded and recovered in the eluate in each purification step were measured using the ELISA method described in Example 2. The results are shown in Table 1. 30.6 g of I2S-anti-hTfR antibody 3, corresponding to approximately 76.5% of 36.7 g of I2S-anti-hTfR antibody 3 contained initially in the culture supernatant, was recovered as a purified product. These results indicate that the purification method described in the above examples is very efficient as a purification method of I2S-anti-hTfR antibody 3. In Table 1, the process recovery rate (%) means the ratio of the recovered rhI2S amount to the loaded amount of rhI2S in each purification process, and the total recovery rate (%) means the ratio of the amount of rhI2S recovered in each purification step to the initial amount of rhI2S used in the purification step.

[0142] [TABLE 1]

(Table 1) Recovery rate of I2S-anti-hTfR antibody 3 in each purification step

Purification Step	I2S-anti-hTfR antibody 3			
	loaded amount (g)	Eluted amount (g)	process recovery rate (%)	total recovery rate (%)
Protein A affinity column	36.7	34.6	100.1	100.1
Hydroxyapatite column	34.1	31.4	92.1	90.9
Gel filtration column	30.6	26.5	86.4	76.5

[0143] [Example 8] Analysis of purified product of I2S-anti-hTfR antibody (quantification of HCP)

The amount of host cell-derived protein (HCP) contained in the purified product of I2S-anti-hTfR antibody was quantified by ELISA method. At first, 100 μ L of anti-CHO cell-derived protein antibody was added to each well of a 96-well plate (Nunc Inc.), and let stand overnight to adsorb the antibody. After washing each well three times, 200 μ L of a blocking solution containing casein was added to each well and the plate was shaken at 25°C for 60 minutes. After washing each well three times, 100 μ L each of a solution (sample solution) containing a purified product of I2S-anti-hTfR antibody or HCP standard solution was added to each well, followed by shaking at 25°C for 2 hours. After washing each well three times, 100 μ L of biotinylated anti-CHO cell-derived protein antibody was added to each well and the plate was shaken at 25°C for 60 minutes. After washing each well three times, 100 μ L of HRP-conjugated streptavidin (Jackson Immuno Research Laboratories Inc.) was added and the plate was shaken at 25°C for 60 minutes. After washing each well three times, 100 μ L of TMB substrate solution was added to each well and the plate was shaken at 25°C to develop color. As the TMB substrate solution, a mixture of TMB

peroxidase substrate and peroxidase substrate solution B, both included in TMB microwell peroxidase substrate system (KPL Inc.), in equal amount was used. After color development, 100 μ L of 1 mol/L sulfuric acid was added to each well to stop the enzymatic reaction, and the absorbance at 450 nm in each well was measured using a 96-well plate reader. A standard curve was produced on the measurement value of the HCP standard solution, and the value of the sample solution was interpolated to the standard curve to quantify the HCP contained in the purified product of the I2S-anti-hTfR antibody. The HCP contained in the purified product of the I2S-anti-hTfR antibody was quantified from the quantified value of HCP thus determined and the quantitative value of the purified product of I2S-anti-hTfR antibody measured by the ELISA method described in Example 2. As a result, it was found that the amount of HCP contained in the purified product of I2S-anti-hTfR antibody was about 35 ppm (ie, about 35 ng of HCP per 1 mg of I2S-anti-hTfR antibody purified product).

[0144] [Example 9] Analysis of purified product of I2S-anti-hTfR antibody (SE-HPLC analysis)

TSK gel UltraSW Aggregate column (inner diameter 7.8 mm $\times$ height 30 cm, Tosoh Corporation Inc.) was set in UV/VIS detector of SPD-20 AV, the LC-20A system (Shimadzu Corporation). The column was equilibrated with 200 mM phosphate buffer (pH 6.5) containing 5% propanol and 20 mM NaCl. To this column, 10 μ L of a solution containing the purified product of I2S-anti-hTfR antibody obtained in Example 6 at a concentration of 1 mg/mL was loaded at a constant flow rate of 0.5 mL/min, and the same buffer was supplied at the same flow

rate. Figure 1 shows the elution profile obtained by measuring the absorbance at 215 nm. The obtained profile showed almost only a single peak corresponding to I2S-anti-hTfR antibody 3. However, a peak (peak B in the figure) derived from the polymer of I2S-anti-hTfR antibody 3 detected prior to the main peak (peak A in the figure) was observed. From the ratio of the area of the peak B to the area of the whole peak, the ratio of the polymer to the whole I2S-anti-hTfR antibody 3 was calculated to be about 0.49%.

[0145] [Example 10] Analysis of purified product of I2S-anti-hTfR antibody (Summary)

The results of analysis of the purified product of the I2S-anti-hTfR antibody described above indicate that the purified product of the I2S-anti-hTfR antibody obtained in Example 6 contains almost no impurities including HCP and that the abundance ratio of the polymer is extremely low. That is, it is concluded that the purified product of I2S-anti-hTfR antibody has the quality to permit its use as a medicine as it is, for example, as an intravenously, intramuscularly, subcutaneously, intraperitoneally, intraarterially or intralesionally administered medicine.

[0146] [Example 11] Culture of hI2S-anti-hTfR antibody expressing strain (alternative method)

The seed cells of the hI2S-anti-hTfR antibody expressing strain 3 obtained in Example 3 were thawed in a 37°C water bath. The cells were cultured with shaking in serum-free medium (EX-CELL Advanced CHO Fed-batch Medium, Sigma Aldrich Inc.) containing 4 mM L-alanyl-L-glutamine, 100 µmol/L hypoxanthine, 16 µmol/L thymidine,

500 µg/mL hygromycin B, and 10 µg/mL puromycin at a density of 4×10^5 cells/mL for 3 days under the conditions of 37°C and 5% CO₂. This culture was repeated until the cell number grew to at least 5×10^{11} cells.

[0147] Subsequently, the cells were suspended in serum-free medium

(EX-CELL Advanced CHO Fed-batch Medium, Sigma Aldrich Inc.) supplemented with 4 mM L-alanyl-L-glutamine, 100 µmol/L hypoxanthine, and 16 µmol/L thymidine so that the cell density became

about 2×10^5 cells/mL. About 1400 L of this cell suspension was transferred to a culture tank and stirred with an impeller at a rate of 80

rpm, and the pH of the medium was kept at 6.9 and the dissolved oxygen saturation at about 40%, and cells were cultured for about 11 days while adjusting the culture temperature at the range of 34~37°C.

Further, 70 L of EX-CELL Advanced CHO Feed 1 containing 35 g/L glucose was added daily from day 3 to day 10. Sampling was carried out

every day during culturing, and cell number, viability, glucose concentration, lactic acid concentration were measured. The expression level of anti-hTfRAb-I2S was measured from day 5 to day 11. When the glucose concentration became less than 15 mmol/L, glucose was immediately added so as to have its concentration to be 37.89 mmol/L.

[0148] After completion of the culture, the medium was collected. The collected medium was filtered through Millistak+HC Pod Filter grade D0HC (Merck Inc.) and further filtered through Millistak+ HCgrade X0HC (Merck Inc.) to obtain a culture supernatant containing I2S-anti-hTfR antibody 3. The culture supernatant was subjected to ultrafiltration using a Pellicon™ 3 Cassette w/Ultracel PLCTK Membrane (pore size: 30 kDa, membrane area: 9.12 m², Merck Inc.)

and concentrated until the liquid volume was about 1/13. The concentrate was then filtered using Opticap XL 600 (0.22 µm, Merck Inc.). The obtained solution was used as a concentrated culture supernatant.

5 [0149] [Example 12] Purification of hI2S-anti-hTfR Antibody (Alternative Method)

10 A 1/3 volume of the concentrated culture supernatant obtained in Example 11 was loaded on a MabSelect SuRe LX column (column volume: about 9.8 L, bed height: about 20 cm, GE Healthcare Inc.), a Protein A affinity column, equilibrated with 140 column volumes of 20 mM Tris-HCl buffer (pH 7.0) containing 140 mM NaCl at a constant flow rate of 200 cm/hr to adsorb I2S-anti-hTfR antibody 3 to Protein A. [0150] Subsequently, the column was washed by supplying 5 column volumes of 10 mM Tris-HCl buffer (pH 7.0) containing 500 mM NaCl and 450 mM arginine at the same flow rate. Then the column was washed with 2.5 column volumes of 20 mM Tris-HCl buffer (pH 7.0) containing 140 mM NaCl at the same flow rate. Then I2S-anti-hTfR antibody 3 adsorbed on Protein A was eluted with 5 column volumes of 100 mM glycine buffer (pH 3.5) containing 140 mM NaCl. The eluate was immediately neutralized by collecting in a container containing 100 mM MES buffer (pH 7.0).

20 [0151] Polysorbate 80 was added to the above eluate from the protein A affinity column so that the final concentration was to be 1% (w/v), and the mixture was gently stirred at room temperature for 3 hours or more. 25 This step is for inactivating viruses that may be contaminating the eluate.

[0152] To the above solution after the virus inactivation, 200 mM phosphate buffer (pH 7.0), 10 mM MES buffer (pH 7.3) containing 4 M NaCl and 2 mM phosphate buffer, and 1 M Tris-HCl buffer solution (pH 8.8) were added in the order, and the concentrations of sodium phosphate and NaCl contained in the eluate were adjusted to 2 mM and 215 mM, respectively, and the pH of the eluate was adjusted to 7.3. The eluate was then filtered through OPTICAP SHC XL 3 (0.22 μ m, Merck Inc.). The solution after filtration was applied to a hydroxyapatite column, CHT Type II 40 μ m column (volume of column: about 19.2 L, bed height: about 20 cm, Bio-Rad Inc.), equilibrated with 4 column volumes of 10 mM MES buffer (pH 7.3) containing 215 mM NaCl and 2 mM phosphate buffer at a constant flow rate of 200 cm/hr to a column to adsorb I2S-anti-hTfR antibody 3 on hydroxyapatite.

[0153] Subsequently, the column was washed by supplying 5 column volumes of same buffer at the same flow rate. Then I2S-anti-hTfR antibody 3 adsorbed on hydroxyapatite was eluted with 5 column volumes of 35 mM phosphate buffer (pH 7.3) containing 215 mM NaCl.

[0154] To the above eluate from the hydroxyapatite column, dilute hydrochloric acid was added to adjust the pH to 6.5. Subsequently, ultrafiltration was carried out using Pellicon™ 3 Cassette w/Ultracel PLCTK Membrane (pore size: 30 kDa, membrane area: 2.85 m², Merck Inc.) to concentrate I2S-anti-hTfR antibody 3 in the solution at the concentration of about 20 mg/mL. The concentrate was then filtered using Opticap XL 600 (0.22 μ m, Merck Inc.).

[0155] The above concentrated solution was applied to a Superdex 200 column, size exclusion column (column volume: about 38.5 L, bed

height: 40 cm, GE Healthcare Inc.) equilibrated with 5 column volumes of 20 mM phosphate buffer (pH 6.5) containing 0.8 mg/mL NaCl and 75 mg/mL sucrose at a flow rate of 24 cm/hr or less, and the same buffer was supplied at the same flow rate. At this time, an absorbance photometer for continuously measuring the absorbance of the eluate was placed in the flow path of the eluate from the size exclusion column, and the absorbance at 280 nm was monitored, the fractions which corresponded to an absorption peak at 280 nm were collected as a fraction containing I2S-anti-hTfR antibody 3. The recovered solution was filtered with Planova 20N (size: 0.3 m², Asahi Kasei Medical Inc.) and Millipak-100 Filter Unit (pore diameter: 0.22 μm, Merck Inc.). The solution after filtration was designated as purified product of I2S-anti-hTfR antibody.

[0156] [Example 13] Measurement of recovery rate of I2S-anti-hTfR antibody in each purification step (alternative method)

The amount of I2S-anti-hTfR antibody 3 loaded and recovered in the eluate in each purification step (alternative method) were measured using the ELISA method described in Example 2. The results are shown in Table 2. 68.8 g of I2S-anti-hTfR antibody 3, corresponding to approximately 82% of 96.8 g of I2S-anti-hTfR antibody 3 contained initially in the culture supernatant, was recovered as a purified product. These results indicate that the purification method (alternative method) described in the above Example 12 is very efficient as a purification method of the I2S-anti-hTfR antibody. Meanings of process recovery rate (%) and total recovery rate (%) in Table 3 are the same as those used in Table 1.

[0157] [TABLE 2]

(Table 2) Recovery rate of I2S-anti-hTfR antibody 3 in each purification step

Purification Step	I2S-anti-hTfR antibody 3			
	loaded amount (g)	Eluted amount (g)	process recovery rate (%)	total recovery rate (%)
Protein A affinity column	96.8	100.1	103.3	103.3
Hydroxyapatite column	93.7	81.1	86.6	89.5
Gel filtration column	68.8	62.8	91.3	81.7

[0158] [Example 14] Analysis of purified product of I2S-anti-hTfR antibody obtained by alternative method (quantification of HCP)

5 For the purified product of I2S-anti-hTfR antibody obtained in Example 12, The amount of HCP was quantified by the method described in Example 8. The results indicate that the amount of HCP contained in the purified product of I2S-anti-hTfR antibody was about 20 ppm (ie, about 20 ng of HCP per 1 mg of I2S-anti-hTfR antibody purified product).

10 [0159] [Example 15] Analysis of purified product of I2S-anti-hTfR antibody obtained by alternative method (SE-HPLC analysis)

SE-HPLC analysis was performed for the purified product of I2S-anti-hTfR antibody obtained in Example 12 by the method described in Example 9. The analytical result is shown in figure 2. The obtained profile showed almost only a single peak corresponding to I2S-anti-hTfR antibody 3. However, a peak (peak B in the figure) derived from the polymer of I2S-anti-hTfR antibody 3 detected prior to the main peak (peak A in the figure) was observed. From the ratio of the area of the peak B to the area of the whole peak, the ratio of the polymer to the whole I2S-anti-hTfR antibody 3 was calculated to be about

0.37%.

[0160] [Example 16] Analysis of purified product of I2S-anti-hTfR antibody obtained by alternative (Summary)

5 The results of analysis of the purified product of the I2S-anti-hTfR antibody described above indicate that the purified product of the I2S-anti-hTfR antibody obtained in Example 12 contains almost no impurities including HCP and that the abundance ratio of the polymer is extremely low. That is, it is concluded that the purified product of I2S-anti-hTfR antibody has the quality to permit its use as a
10 medicine as it is, for example, as an intravenously, intramuscularly, subcutaneously, intraperitoneally, intraarterially or intralesionally administered medicine.

Industrial Applicability

[0161] According to the present invention, for example, a fusion protein
15 in which an antibody is fused with another protein can be provided, that fusion protein is purified to such a purity as permits its direct use as a medicine.

Sequence Listing Free Text

[0162] SEQ ID NO:1 = Amino acid sequence of an exemplified linker 1
20 SEQ ID NO:2 = Amino acid sequence of an exemplified linker 2
SEQ ID NO:3 = Amino acid sequence of an exemplified linker 3
SEQ ID NO:6 = Amino acid sequence of the light-chain of humanized anti-hTfR antibody No.1
SEQ ID NO:7 = Amino acid sequence of the heavy-chain of
25 humanized anti-hTfR antibody No.1
SEQ ID NO:8 = Amino acid sequence of the light-chain of

humanized anti-hTfR antibody No.2

SEQ ID NO:9 = Amino acid sequence of the heavy-chain of humanized anti-hTfR antibody No.2

5 SEQ ID NO:10 = Amino acid sequence of the light-chain of humanized anti-hTfR antibody No.3

SEQ ID NO:11 = amino acid sequence of the heavy-chain of humanized anti-hTfR antibody No.3

SEQ ID NO:12 = Amino acid sequence of fusion protein of the heavy-chain of humanized anti-hTfR antibody No.1 and hI2S

10 SEQ ID NO:13 = Amino acid sequence of fusion protein of the heavy-chain of humanized anti-hTfR antibody No.2 and hI2S

SEQ ID NO:14 = Amino acid sequence of fusion protein of the heavy-chain of humanized anti-hTfR antibody No.3 and hI2S

15 SEQ ID NO:15 = Amino acid sequence of the light-chain variable region of humanized anti-hTfR antibody No.1

SEQ ID NO:16 = Amino acid sequence of the heavy-chain variable region of humanized anti-hTfR antibody No.1

SEQ ID NO:17 = Amino acid sequence of the light-chain variable region of humanized anti-hTfR antibody No.2

20 SEQ ID NO:18 = Amino acid sequence of the heavy-chain variable region of humanized anti-hTfR antibody No.2

SEQ ID NO:19 = Amino acid sequence of the light-chain variable region of humanized anti-hTfR antibody No.3

25 SEQ ID NO:20 = Amino acid sequence of the heavy-chain variable region of humanized anti-hTfR antibody No.3

SEQ ID NO:21 = Nucleotide sequence encoding amino acid

sequence of the light-chain of humanized anti-hTfR antibody No.1,
synthetic sequence

5 SEQ ID NO:22 = Nucleotide sequence encoding amino acid
sequence of the light-chain of humanized anti-hTfR antibody No.2,
synthetic sequence

 SEQ ID NO:23 = Nucleotide sequence encoding amino acid
sequence of the light-chain of humanized anti-hTfR antibody No.3,
synthetic sequence

10 SEQ ID NO:24 = Nucleotide sequence encoding amino acid
sequence of fusion protein of the heavy-chain of humanized anti-hTfR
antibody No.1 and hI2S, synthetic sequence

 SEQ ID NO:25 = Nucleotide sequence encoding amino acid
sequence of fusion protein of the heavy-chain of humanized anti-hTfR
antibody No.2 and hI2S, synthetic sequence

15 SEQ ID NO:26 = Nucleotide sequence encoding amino acid
sequence of fusion protein of the heavy-chain of humanized anti-hTfR
antibody No.3 and hI2S, synthetic sequence

 SEQ ID NO:27 = Primer Hyg-Sfi5', synthetic sequence

 SEQ ID NO:28 = Primer Hyg-BstX3', synthetic sequence

20 **Sequence Listing**

CLAIMS

Claim 1. A method for production of a fusion protein in which an anti-transferrin receptor (anti-TfR) antibody and a human iduronate-2-sulfatase (human I2S) are fused, the method comprising;

5 (a) a step of culturing mammalian cells producing the fusion protein in a serum-free medium to let the mammalian cells secrete the fusion protein in the culture medium,

(b) a step of collecting culture supernatant by removing the mammalian cells from the culture medium, and

10 (c) a step of purifying the fusion protein from the culture supernatant, by three column chromatographies consisting of

(i) a column chromatography employing as a solid phase a material to which a substance having affinity for the fusion protein has been bound, wherein the substance having affinity for the fusion protein is Protein A,

15 (ii) a column chromatography employing as a solid phase a material having affinity for the phosphate group, wherein the material having affinity for the phosphate group is hydroxyapatite, and

20 (iii) a size exclusion column chromatography, wherein the three column chromatographies (i) to (iii) are performed in this order, and wherein the light chain and the heavy chain of the anti-TfR antibody comprise the amino acid sequences set forth as SEQ ID NO:10 and SEQ ID NO:11, respectively.

25 Claim 2. The method for production according to claim 1, wherein the anti-TfR antibody fused to the human I2S is a humanized

anti-TfR antibody or a human anti-TfR antibody.

5 Claim 3. The method for production according to claim 1, wherein the anti-TfR antibody fused to the human I2S is a humanized anti-TfR antibody.

10 Claim 4. The method for production according to any one of claims 1 to 3, wherein the anti-TfR antibody is an anti-human transferrin receptor (anti-hTfR) antibody.

15 Claim 5. The method for production according to any one of claims 1 to 4, wherein the anti-TfR antibody and the human I2S are fused via a linker in the fusion protein, and wherein the linker is selected from the group consisting of polyethylene glycol, polypropylene glycol, copolymer of ethylene glycol and propylene glycol, polyoxyethylated polyol, polyvinyl alcohol, polysaccharide, dextran, polyvinyl ether, biodegradable polymer, lipid polymer, chitin, hyaluronic acid, and biotin-streptavidin,.

20 Claim 6. The method for production according to any one of claims 1 to 4, wherein the human I2S is linked, by peptide bonds directly or via a linker sequence, to the heavy chain of the anti-TfR antibody on the C-terminal side or the N-terminal side thereof in the fusion protein.

25 Claim 7. The method for production according to any one of

claims 1 to 4, wherein the human I2S is linked, by peptide bonds directly or via a linker sequence, to the light chain of the anti-TfR antibody on the C-terminal side or the N-terminal side thereof in the fusion protein.

5

Claim 8. The method for production according to claim 6 or 7, wherein the linker sequence consists of 1 to 50 amino acid residues.

10 Claim 9. The method for production according to claim 8, wherein the linker sequence comprises an amino acid sequence selected from the group consisting of a single glycine, a single serine, the amino acid sequence of Gly-Ser, the amino acid sequence of Gly-Gly-Ser, the amino acid sequence set forth as SEQ ID NO:1, the amino acid sequence set forth as SEQ ID NO:2, the amino acid sequence set forth as SEQ ID NO:3, and the amino acid sequences consisting of 1 to 10 thereof that are consecutively linked.

20 Claim 10. The method for production according to claim 8 or 9, wherein the linker sequence is represented by the amino acid sequence of Gly-Ser.

Claim 11. The method for production according to any one of claims 1 to 10, wherein the human I2S comprises the amino acid sequence set forth as SEQ ID NO:5.

25

Claim 12. The method for production according to any one of

claims 1 to 10, wherein the human I2S has the amino acid sequence introduced 1 to 3 amino acid substitutions, deletions or additions relative to the amino acid sequence set forth as SEQ ID:NO5, and has an activity as human I2S.

5

Claim 13. The method for production according to claim 1, wherein the anti-TfR antibody is a humanized anti-hTfR antibody, and the fusion protein comprises the light chain of the humanized anti-hTfR antibody having the amino acid sequence set forth as SEQ ID NO:10, and the heavy chain of the humanized anti-hTfR antibody having the amino acid sequence set forth as SEQ ID NO:11 to which the human iduronate-2-sulfatase set forth as SEQ ID NO:5 is linked on the C-terminal side thereof and via a linker sequence of Gly-Ser.

10

Claim 14. The method for production according to claim 1, wherein the anti-TfR antibody is a humanized anti-hTfR antibody, and the fusion protein comprises;

15

the light chain of the humanized anti-hTfR antibody having the amino acid sequence set forth as SEQ ID NO:10, and

20

the heavy chain of the humanized anti-hTfR antibody to which the human iduronate-2-sulfatase is linked on the C-terminal side thereof via a linker sequence of Gly-Ser, and having the amino acid sequence set forth as SEQ ID NO:14 as a whole.