

HUMAN ANTIBODIES TO INFLUENZA HEMAGGLUTININ

FIELD OF THE INVENTION

[0001] The present invention is related to human antibodies and antigen-binding fragments thereof that specifically bind to influenza hemagglutinin (HA), compositions comprising these antibodies and therapeutic and diagnostic methods of using these antibodies.

SEQUENCE LISTING

[0002] An ASCII compliant text file of the sequence listing is filed concurrently with the present specification (37 CFR §1.52(e) and 37 CFR §1.821). The contents of the text file are herein incorporated by reference. The text file containing the sequence listing is named "10119WO01_seqlisting", was created on November 18, 2015, and contains 2.1 MB.

BACKGROUND OF THE INVENTION

[0003] Influenza is a highly contagious disease, which has a long history characterized by waves of pandemics, epidemics, resurgences and outbreaks. In spite of annual vaccination efforts, influenza infections result in substantial morbidity and mortality.

[0004] Influenza viruses consist of three types, A, B and C. Furthermore, influenza A viruses can be classified into subtypes based on allelic variations in antigenic regions of two genes that encode the surface glycoproteins, hemagglutinin (HA) and neuraminidase (NA), which are required for viral attachment and entry into the host cell.

[0005] Hemagglutinin is a trimeric glycoprotein that contains two structural domains, a globular head domain that consists of the receptor-binding site (that is subject to frequent antigenic drift) and the stem region (more conserved among various strains of influenza virus). The HA protein is synthesized as a precursor (HA0), which undergoes proteolytic processing to produce two subunits (HA1 and HA2), which associate with one another to form the stem/globular head structure. The HA1 peptide is responsible for the attachment of virus to the cell surface. The HA2 peptide forms a stem-like structure that mediates the fusion of viral and cell membranes in endosomes, allowing the release of the ribonucleoprotein complex into the cytoplasm.

[0006] Currently, there are eighteen subtypes defined by their hemagglutinin proteins (H1-H18). The 18 HAs can be classified into two groups. Group 1 consists of H1, H2, H5, H6, H8, H9, H11, H12, H13, H16, H17 and H18 subtypes, and group 2 includes H3, H4, H7, H10, H14 and H15 subtypes.

[0007] New strains of the same subtype may arise as a result of a phenomenon called antigenic drift, or mutations in the HA or NA molecules which generate new and different epitopes. A consequence of this is that a new vaccine must be produced every year against viruses that are predicted to emerge, a process that is not only costly, but highly inefficient. While technological advances have improved the ability to produce improved influenza

antigen(s) for vaccine compositions, there remains a need to provide additional sources of protection to address emerging subtypes and strains of influenza.

[0008] While the idea of a vaccine composition comprising the antigen of interest (e.g. the HA and/or NA) to generate broadly neutralizing antibodies in a patient is generally thought to be a good approach, it is not always desirable to use this approach in certain patient populations. For example, in certain patients, a vaccine composition comprising the antigen of interest may not always be effective, such as in the elderly, in the very young, in immunocompromised patients, etc. In these patient populations, or in any patient who is not able to mount an effective immune response, it may be more beneficial to provide a composition already containing broadly neutralizing antibodies that may target epitopes common to a variety of strains within Group 1 and/or Group 2 subtypes.

[0009] To date there has been limited success in identifying such antibodies that broadly neutralize or inhibit influenza viruses. Okuno *et al.* immunized mice with influenza A/Okuda/57 (H2N2) and isolated an antibody designated C179, which bound to a conserved conformational epitope in HA2 and neutralized the Group 1 H2, H1 and H5 subtype influenza A viruses *in vitro* and *in vivo* (Okuno *et al.* (1993) J. Virol. 67(5):2552-2558). Throsby *et al.* identified 13 monoclonal antibodies from human B cells that had broad activity against Group 1 subtypes (Throsby *et al.* (2008), PLOS one 3(2):e3942). Sui *et al.* identified a human monoclonal antibody (F10), which bound H5 and other Group 1 viruses (Sui, *et al.* (2009), Nat. Struct. Mol. Biol. 16(3):265-273).

[0010] However, after decades of research in this area, only a few antibodies are currently in clinical trials to assess their ability to neutralize influenza viruses of different subtypes (See, for example, antibodies under development by Crucell Holland ((US2012/0276115, US2014/0065156, US8470327, US2014/0120113, EP2731967, US8691223, US2013/0243792, US2014/0065165, WO2008/028946 and WO2010/130636); Osaka University (US2011/0319600, EP2380976, US2012/0058124, US2012/0058124), Celltrion (US2013/0004505, EP2545074, WO2014/158001); Vanderbilt University (US2013/0289246), SeaLane Biotechnologies (US2012/0128671), Trellis Bioscience, Inc. (US2012/0020971 EP2582721); Visterra, Inc. (US2013/0302349); Burnham Institute/Dana Farber (US2014/011982, EP2222701, WO2010/027818); Temasek (US8444986, US8574581, US8637644, US8637645, US8383121, US8540996, US8574830, US8540995); HUMABS Biosciences/Institute for Research in Biomedicine (US8871207); MedImmune (WO2015/051010); and Genentech (US2014/0161822), but there are still no marketed antibodies that broadly neutralize or inhibit influenza A virus infection or attenuate the disease caused by various subtypes of this virus. Accordingly, there is still a need in the art to identify new antibodies that neutralize multiple subtypes of influenza A virus, which can be used to prevent or treat an influenza virus infection.

BRIEF SUMMARY OF THE INVENTION

[0011] The present invention provides antibodies and antigen-binding fragments thereof that bind influenza hemagglutinin (HA). The antibodies of the present invention are useful, *inter alia*, for inhibiting or neutralizing the activity of influenza HA. In some embodiments, the antibodies are useful for blocking attachment of the influenza virus to the host cell and/or for preventing the entry of the influenza virus into host cells. In some embodiments, the antibodies function by inhibiting the cell-to-cell transmission of the virus. In certain embodiments, the antibodies are useful in preventing, treating or ameliorating at least one symptom of influenza virus infection in a subject. In certain embodiments, the antibodies may be administered prophylactically or therapeutically to a subject having, or at risk of acquiring, an influenza virus infection. In certain embodiments, compositions containing at least one antibody of the invention may be administered to a subject for whom a vaccine is contra-indicated, or for whom a vaccine is less efficacious, for example, an elderly patient, a very young patient, a patient who may be allergic to any one or more components of a vaccine, or an immunocompromised patient who may be non-responsive to the immunogens in a vaccine. In certain embodiments, compositions containing at least one antibody of the invention may be administered to medical staff, hospitalized patients or nursing home residents or other high-risk patients during an influenza outbreak. In certain embodiments, compositions containing at least one antibody of the invention may be administered as a first line treatment to patients in the event that a predicted yearly vaccine is ineffective, or in the event of a pandemic with a strain that has undergone a major antigenic shift.

[0012] The antibodies of the invention can be full-length (for example, an IgG1 or IgG4 antibody) or may comprise only an antigen-binding portion (for example, a Fab, F(ab')₂ or scFv fragment), and may be modified to affect functionality, e.g., to increase persistence in the host or to increase effector function or eliminate residual effector functions (Reddy *et al.*, 2000, J. Immunol. 164:1925-1933). In certain embodiments, the antibodies may be bispecific.

[0013] In a first aspect, the present invention provides isolated recombinant monoclonal antibodies or antigen-binding fragments thereof that bind specifically to the influenza HA.

[0014] In one embodiment, the present invention provides an isolated recombinant antibody or antigen-binding fragment thereof that specifically binds to influenza A hemagglutinin (HA), wherein the antibody has two or more of the following characteristics:

- (a) is a fully human monoclonal antibody;
- (b) binds to influenza HA with a dissociation constant (K_D) of less than 10^{-9} M, as measured in a real-time bio-layer interferometer based biosensor (Octet HTX assay);
- (c) demonstrates a dissociative half-life ($t_{1/2}$) greater than 370 minutes;
- (d) demonstrates neutralization of group 1 influenza A viruses selected from H1N1, H5N1, H9N2, H13N6 and H16N3, with an IC_{50} of less than 130nM;

(e) demonstrates complement mediated lysis of influenza virus infected cells with an EC_{50} of less than 66nM;

(f) demonstrates protection, as measured by increased survival in an animal model of influenza virus infection when administered either before or after virus challenge; or

(g) wherein the antibody or antigen-binding fragment thereof comprises three heavy chain complementarity determining regions (CDRs) (HCDR1, HCDR2 and HCDR3) contained within any one of the heavy chain variable region (HCVR) sequences listed in Table 1 or Table 12; and three light chain CDRs (LCDR1, LCDR2 and LCDR3) contained within any one of the light chain variable region (LCVR) sequences listed in Table 1 or Table 12.

[0015] In one embodiment, an antibody of the invention demonstrates a dissociative half-life in monkeys that is about 1.5 fold greater than a comparator antibody designated Control I mAb and a dissociative half-life in mice that is about 2 fold greater than the Control I mAb.

[0016] In certain embodiments, an antibody of the invention demonstrates an increase in protection as compared to oseltamivir, when administered at 48 hours post infection, or at 72 hours post infection to a mammal infected with influenza virus.

[0017] In a related embodiment, an antibody of the invention confers an increase in protection in a mammal infected with influenza virus when administered either subcutaneously or intravenously and/or when administered prior to infection, or after infection with influenza virus.

[0018] In one embodiment, an antibody of the invention demonstrates an increase in protection, as compared to an animal administered an isotype (negative) control antibody, when administered to an infected mammal as a single subcutaneous or intravenous dose ranging from about 0.01 mg/kg to about 50 mg/kg.

[0019] In one embodiment, an antibody of the invention demonstrates an increase in protection when administered to an influenza virus infected mammal as a single intravenous dose of about 15 mg/kg compared to oral administration of oseltamivir administered twice daily for 5 days at a dose of about 5 mg/kg to about 25 mg/kg.

[0020] In one embodiment, an antibody of the invention demonstrates a survival rate of greater than about 20% in a mammal infected with influenza virus, when administered prophylactically as a single subcutaneous dose ranging from about 0.01 mg/kg to about 2 mg/kg.

[0021] In one embodiment, an antibody of the invention demonstrates a survival rate of greater than about 30% in a mammal infected with influenza virus, when administered as a single intravenous dose ranging from about 7 mg/kg to about 50 mg/kg by at least 24 hours post infection.

[0022] In one embodiment, an antibody of the invention demonstrates a survival rate of about 30% to about 60% in a mammal infected with influenza virus, when administered as a single intravenous dose of about 7 mg/kg to about 50 mg/kg when administered at 48 hours post infection.

[0023] In one embodiment, an antibody of the invention demonstrates a survival rate of equal

to, or greater than about 60% in a mammal infected with influenza virus, when administered as a single intravenous dose of about 15 mg/kg to about 30 mg/kg at 48 hours or longer post infection.

[0024] In one embodiment, an antibody of the invention demonstrates a survival rate of about 100% in a mammal infected with influenza virus, when administered as a single intravenous dose of about 15 mg/kg at 48 hours, or longer, post infection.

[0025] In one embodiment, an antibody of the invention demonstrates a survival rate of about 100% in a mammal infected with influenza virus, when administered as a single intravenous dose of about 15 mg/kg compared to a 40% survival rate observed with oseltamivir when administered orally twice a day for 5 days at a dose of about 25 mg/kg.

[0026] In one embodiment, an antibody of the invention provides an additive protective effect in a mammal infected with influenza virus when administered with oseltamivir at greater than 48 hours post infection.

[0027] In one embodiment, an antibody of the invention provides an additive protective effect in a mammal infected with influenza virus when administered with oseltamivir at 72 hours post infection.

[0028] In one embodiment, an antibody of the invention provides an additive protective effect when used in combination with oseltamivir when the antibody is administered to an influenza virus infected mammal as a single intravenous dose ranging from about 7 mg/kg to about 15 mg/kg and the oseltamivir is administered orally twice daily for 5 days at a dose of about 25 mg/kg.

[0029] In a related embodiment, an antibody of the invention provides an additive protective effect when used in combination with oseltamivir at 72 hours after influenza virus infection, wherein the antibody is administered as a single intravenous dose ranging from about 7 mg/kg to about 15 mg/kg and the oseltamivir is administered orally twice daily for 5 days at a dose of about 25 mg/kg.

[0030] In one embodiment, an antibody of the invention may be administered intravenously, intranasally, subcutaneously, intradermally, or intramuscularly and the oseltamivir may be administered orally.

[0031] In one embodiment, the oseltamivir is administered prior to, concurrently with, or after administration of an antibody of the invention.

[0032] In one embodiment, the antibody and/or the oseltamivir may be administered as a single dose, or as multiple doses.

[0033] Exemplary anti-influenza HA antibodies of the present invention are listed in Tables 1 and 2 herein. Table 1 sets forth the amino acid sequence identifiers of the heavy chain variable regions (HCVRs), light chain variable regions (LCVRs), heavy chain complementarity determining regions (HCDR1, HCDR2 and HCDR3), and light chain complementarity determining regions (LCDR1, LCDR2 and LCDR3) of exemplary anti-influenza HA antibodies.

Table 2 sets forth the nucleic acid sequence identifiers of the HCVRs, LCVRs, HCDR1, HCDR2, HCDR3, LCDR1, LCDR2 and LCDR3 of the exemplary anti-influenza HA antibodies.

[0034] Further exemplary anti-influenza HA antibodies of the present invention are listed in Tables 12 and 13 herein. Table 12 sets forth the amino acid sequence identifiers of the heavy chain variable regions (HCVRs), light chain variable regions (LCVRs), heavy chain complementarity determining regions (HCDR1, HCDR2 and HCDR3), and light chain complementarity determining regions (LCDR1, LCDR2 and LCDR3) of exemplary anti-influenza HA antibodies. Table 13 sets forth the nucleic acid sequence identifiers of the HCVRs, LCVRs, HCDR1, HCDR2, HCDR3, LCDR1, LCDR2 and LCDR3 of the exemplary anti-influenza HA antibodies.

[0035] The present invention provides antibodies, or antigen-binding fragments thereof, comprising an HCVR comprising an amino acid sequence selected from any of the HCVR amino acid sequences listed in Table 1, or Table 12, or a substantially similar sequence thereof having at least 90%, at least 95%, at least 98% or at least 99% sequence identity thereto.

[0036] In one embodiment the invention provides antibodies, or antigen-binding fragments thereof, which specifically bind influenza HA, comprising a HCVR having an amino acid sequence selected from the group consisting of SEQ ID NOs: 2, 18, 34, 50, 74, 90, 106, 122, 138, 154, 170, 186, 202, 218, 234, 242, 250, 258, 266, 274, 282 and 290.

[0037] The present invention also provides antibodies, or antigen-binding fragments thereof, comprising an LCVR comprising an amino acid sequence selected from any of the LCVR amino acid sequences listed in Table 1, or Table 12, or a substantially similar sequence thereof having at least 90%, at least 95%, at least 98% or at least 99% sequence identity thereto.

[0038] In one embodiment the invention provides antibodies, or antigen-binding fragments thereof, which specifically bind influenza HA, comprising a LCVR having an amino acid sequence selected from the group consisting of SEQ ID NOs: 10, 26, 42, 58, 66, 82, 98, 114, 130, 146, 162, 178, 194, 210 and 226.

[0039] The present invention also provides antibodies, or antigen-binding fragments thereof, comprising an HCVR and an LCVR amino acid sequence pair (HCVR/LCVR) comprising any of the HCVR amino acid sequences listed in Table 1, or Table 12 paired with any of the LCVR amino acid sequences listed in Table 1, or Table 12. According to certain embodiments, the present invention provides antibodies, or antigen-binding fragments thereof, comprising an HCVR/LCVR amino acid sequence pair contained within any of the exemplary anti-influenza HA antibodies listed in Table 1, or Table 12.

[0040] In one embodiment, the isolated antibody or antigen-binding fragment that specifically binds influenza HA comprises a HCVR/LCVR amino acid sequence pair selected from the group consisting of SEQ ID NOs: 2/10, 18/26, 34/42, 50/58, 50/66, 74/82, 74/66, 90/98, 106/114, 122/130, 138/146, 154/162, 170/178, 186/194, 202/210, 218/226, 234/66, 242/66, 250/66, 258/66, 266/66, 274/66, 282/66 and 290/66.

[0041] In certain embodiments, the HCVR/LCVR amino acid sequence pair is selected from the group consisting of SEQ ID NOs: 18/26 (*e.g.*, H1H11729P), 50/58 (*e.g.*, H1H11829N), 50/66 (*e.g.* H1H11829N2), or 106/114 (*e.g.*, H1H14571N).

[0042] In one embodiment, the isolated antibody or antigen-binding fragment comprises:

(a) a HCDR1 domain having an amino acid sequence selected from the group consisting of SEQ ID NOs: 4, 20, 36, 52, 76, 92, 108, 124, 140, 156, 172, 188, 204, 220, 236, 244, 252, 260, 268, 276, 284, and 292;

(b) a HCDR2 domain having an amino acid sequence selected from the group consisting of SEQ ID NOs: 6, 22, 38, 54, 78, 94, 110, 126, 142, 158, 174, 190, 206, 222, 238, 246, 254, 262, 270, 278, 286, and 294;

(c) a HCDR3 domain having an amino acid sequence selected from the group consisting of SEQ ID NOs: 8, 24, 40, 56, 80, 96, 112, 128, 144, 160, 176, 192, 208, 224, 240, 248, 256, 264, 272, 280, 288, and 296;

(d) a LCDR1 domain having an amino acid sequence selected from the group consisting of SEQ ID NOs: 12, 28, 44, 60, 68, 84, 100, 116, 132, 148, 164, 180, 196, 212, and 228;

(e) a LCDR2 domain having an amino acid sequence selected from the group consisting of SEQ ID NOs: 14, 30, 46, 62, 70, 86, 102, 118, 134, 150, 166, 182, 198, 214, and 230; and

(f) a LCDR3 domain having an amino acid sequence selected from the group consisting of SEQ ID NOs: 16, 32, 48, 64, 72, 88, 104, 120, 136, 152, 168, 184, 200, 216 and 232.

[0043] In one embodiment, the isolated antibody or antigen-binding fragment, which specifically binds influenza HA, comprises (a) a HCDR1 of SEQ ID NO: 20, (b) a HCDR2 of SEQ ID NO: 22; (c) a HCDR3 of SEQ ID NO: 24; (d) a LCDR1 of SEQ ID NO: 28; (e) a LCDR2 of SEQ ID NO: 30 and (f) a LCDR3 of SEQ ID NO: 32.

[0044] In one embodiment, the isolated antibody or antigen-binding fragment, which specifically binds influenza HA, comprises (a) a HCDR1 of SEQ ID NO: 52, (b) a HCDR2 of SEQ ID NO: 54; (c) a HCDR3 of SEQ ID NO: 56; (d) a LCDR1 of SEQ ID NO: 68; (e) a LCDR2 of SEQ ID NO: 70 and (f) a LCDR3 of SEQ ID NO: 72.

[0045] In one embodiment, the isolated antibody or antigen-binding fragment, which specifically binds influenza HA, comprises (a) a HCDR1 of SEQ ID NO: 52, (b) a HCDR2 of SEQ ID NO: 54; (c) a HCDR3 of SEQ ID NO: 56; (d) a LCDR1 of SEQ ID NO: 60; (e) a LCDR2 of SEQ ID NO: 62 and (f) a LCDR3 of SEQ ID NO: 64.

[0046] In one embodiment, the isolated antibody or antigen-binding fragment, which specifically binds influenza HA, comprises (a) a HCDR1 of SEQ ID NO: 108, (b) a HCDR2 of SEQ ID NO: 110; (c) a HCDR3 of SEQ ID NO: 112; (d) a LCDR1 of SEQ ID NO: 116; (e) a LCDR2 of SEQ ID NO: 118 and (f) a LCDR3 of SEQ ID NO: 120.

[0047] The present invention also provides antibodies, or antigen-binding fragments thereof, comprising a heavy chain CDR1 (HCDR1) comprising an amino acid sequence selected from any of the HCDR1 amino acid sequences listed in Table 1, or Table 12, or a substantially similar sequence thereof having at least 90%, at least 95%, at least 98% or at least 99% sequence identity.

[0048] The present invention also provides antibodies, or antigen-binding fragments thereof, comprising a heavy chain CDR2 (HCDR2) comprising an amino acid sequence selected from any of the HCDR2 amino acid sequences listed in Table 1, or Table 12, or a substantially similar sequence thereof having at least 90%, at least 95%, at least 98% or at least 99% sequence identity.

[0049] The present invention also provides antibodies, or antigen-binding fragments thereof, comprising a heavy chain CDR3 (HCDR3) comprising an amino acid sequence selected from any of the HCDR3 amino acid sequences listed in Table 1, or Table 12, or a substantially similar sequence thereof having at least 90%, at least 95%, at least 98% or at least 99% sequence identity.

[0050] The present invention also provides antibodies, or antigen-binding fragments thereof, comprising a light chain CDR1 (LCDR1) comprising an amino acid sequence selected from any of the LCDR1 amino acid sequences listed in Table 1, or Table 12, or a substantially similar sequence thereof having at least 90%, at least 95%, at least 98% or at least 99% sequence identity.

[0051] The present invention also provides antibodies, or antigen-binding fragments thereof, comprising a light chain CDR2 (LCDR2) comprising an amino acid sequence selected from any of the LCDR2 amino acid sequences listed in Table 1, or Table 12, or a substantially similar sequence thereof having at least 90%, at least 95%, at least 98% or at least 99% sequence identity.

[0052] The present invention also provides antibodies, or antigen-binding fragments thereof, comprising a light chain CDR3 (LCDR3) comprising an amino acid sequence selected from any of the LCDR3 amino acid sequences listed in Table 1, or Table 12, or a substantially similar sequence thereof having at least 90%, at least 95%, at least 98% or at least 99% sequence identity.

[0053] The present invention also provides antibodies, or antigen-binding fragments thereof, comprising an HCDR3 and an LCDR3 amino acid sequence pair (HCDR3/LCDR3) comprising any of the HCDR3 amino acid sequences listed in Table 1, or Table 12, paired with any of the LCDR3 amino acid sequences listed in Table 1, or Table 12. According to certain embodiments, the present invention provides antibodies, or antigen-binding fragments thereof, comprising an HCDR3/LCDR3 amino acid sequence pair contained within any of the exemplary anti-influenza HA antibodies listed in Table 1, or Table 12. In certain embodiments, the HCDR3/LCDR3 amino acid sequence pair is selected from the group consisting of SEQ ID

NOs: 24/32 (*e.g.*, H1H11729P), 56/64 (*e.g.*, H1H11829N), 56/72 (*e.g.* H1H11829N2) and 112/120 (*e.g.*, H1H14571N).

[0054] The present invention also provides antibodies, or antigen-binding fragments thereof, comprising a set of six CDRs (*i.e.*, HCDR1-HCDR2-HCDR3-LCDR1-LCDR2-LCDR3) contained within any of the exemplary anti-influenza HA antibodies listed in Table 1, or Table 12. In certain embodiments, the HCDR1-HCDR2-HCDR3-LCDR1-LCDR2-LCDR3 amino acid sequence set is selected from the group consisting of SEQ ID NOs: 20-22-24-28-30-32 (*e.g.*, H1H11729P), 52-54-56-60-62-64 (*e.g.*, H1H11829N); 52-54-56-68-70-72 (*e.g.* H1H11829N2) and 108-110-112-116-118 and 120 (*e.g.*, H1H14571N).

[0055] In a related embodiment, the present invention provides antibodies, or antigen-binding fragments thereof, comprising a set of six CDRs (*i.e.*, HCDR1-HCDR2-HCDR3-LCDR1-LCDR2-LCDR3) contained within an HCVR/LCVR amino acid sequence pair as defined by any of the exemplary anti-influenza HA antibodies listed in Table 1, or Table 12. For example, the present invention includes antibodies, or antigen-binding fragments thereof, comprising the HCDR1-HCDR2-HCDR3-LCDR1-LCDR2-LCDR3 amino acid sequences set contained within an HCVR/LCVR amino acid sequence pair selected from the group consisting of SEQ ID NOs: 18/26 (*e.g.*, H1H11729P), 50/58 (*e.g.*, H1H11829N), 50/66 (*e.g.* H1H11829N2) and 106/114 (*e.g.*, H1H14571N). Methods and techniques for identifying CDRs within HCVR and LCVR amino acid sequences are well known in the art and can be used to identify CDRs within the specified HCVR and/or LCVR amino acid sequences disclosed herein. Exemplary conventions that can be used to identify the boundaries of CDRs include, *e.g.*, the Kabat definition, the Chothia definition, and the AbM definition. In general terms, the Kabat definition is based on sequence variability, the Chothia definition is based on the location of the structural loop regions, and the AbM definition is a compromise between the Kabat and Chothia approaches. See, *e.g.*, Kabat, "Sequences of Proteins of Immunological Interest," National Institutes of Health, Bethesda, Md. (1991); Al-Lazikani *et al.*, *J. Mol. Biol.* 273:927-948 (1997); and Martin *et al.*, *Proc. Natl. Acad. Sci. USA* 86:9268-9272 (1989). Public databases are also available for identifying CDR sequences within an antibody.

[0056] The present invention includes anti-influenza HA antibodies having a modified glycosylation pattern. In some embodiments, modification to remove undesirable glycosylation sites may be useful, or an antibody lacking a fucose moiety present on the oligosaccharide chain, for example, to increase antibody dependent cellular cytotoxicity (ADCC) function (see Shield *et al.* (2002) JBC 277:26733). In other applications, modification of galactosylation can be made in order to modify complement dependent cytotoxicity (CDC).

[0057] The present invention also provides for antibodies and antigen-binding fragments thereof that compete for specific binding to influenza HA with an antibody or antigen-binding fragment thereof comprising the CDRs of a HCVR and the CDRs of a LCVR, wherein the HCVR and LCVR each has an amino acid sequence selected from the HCVR and LCVR sequences

listed in Table 1, or Table 12.

[0058] The present invention also provides antibodies and antigen-binding fragments thereof that cross-compete for binding to influenza HA, or that bind the same epitope on influenza HA, as a reference antibody or antigen-binding fragment thereof comprising the CDRs of a HCVR and the CDRs of a LCVR, wherein the HCVR and LCVR each has an amino acid sequence selected from the HCVR and LCVR sequences listed in Table 1, or Table 12.

[0059] The present invention also provides isolated antibodies and antigen-binding fragments thereof that block influenza HA attachment to, and/or entry into a host cell.

[0060] In certain embodiments, the antibodies or antigen-binding fragments of the present invention are bispecific comprising a first binding specificity to a first epitope in the influenza HA and a second binding specificity to another antigen.

[0061] In a second aspect, the present invention provides nucleic acid molecules encoding anti-influenza HA antibodies or portions thereof. For example, the present invention provides nucleic acid molecules encoding any of the HCVR amino acid sequences listed in Table 1, or Table 12; in certain embodiments the nucleic acid molecule comprises a polynucleotide sequence selected from any of the HCVR nucleic acid sequences listed in Table 2, or Table 13, or a substantially similar sequence thereof having at least 90%, at least 95%, at least 98% or at least 99% sequence identity thereto.

[0062] The present invention also provides nucleic acid molecules encoding any of the LCVR amino acid sequences listed in Table 1, or Table 12; in certain embodiments the nucleic acid molecule comprises a polynucleotide sequence selected from any of the LCVR nucleic acid sequences listed in Table 2, or Table 13, or a substantially similar sequence thereof having at least 90%, at least 95%, at least 98% or at least 99% sequence identity thereto.

[0063] The present invention also provides nucleic acid molecules encoding any of the HCDR1 amino acid sequences listed in Table 1, or Table 12; in certain embodiments the nucleic acid molecule comprises a polynucleotide sequence selected from any of the HCDR1 nucleic acid sequences listed in Table 2, or Table 13, or a substantially similar sequence thereof having at least 90%, at least 95%, at least 98% or at least 99% sequence identity thereto.

[0064] The present invention also provides nucleic acid molecules encoding any of the HCDR2 amino acid sequences listed in Table 1, or Table 12; in certain embodiments the nucleic acid molecule comprises a polynucleotide sequence selected from any of the HCDR2 nucleic acid sequences listed in Table 2, or Table 13, or a substantially similar sequence thereof having at least 90%, at least 95%, at least 98% or at least 99% sequence identity thereto.

[0065] The present invention also provides nucleic acid molecules encoding any of the HCDR3 amino acid sequences listed in Table 1, or Table 12; in certain embodiments the nucleic acid molecule comprises a polynucleotide sequence selected from any of the HCDR3 nucleic acid sequences listed in Table 2, or Table 13, or a substantially similar sequence thereof having at least 90%, at least 95%, at least 98% or at least 99% sequence identity thereto.

[0066] The present invention also provides nucleic acid molecules encoding any of the LCDR1 amino acid sequences listed in Table 1, or Table 12; in certain embodiments the nucleic acid molecule comprises a polynucleotide sequence selected from any of the LCDR1 nucleic acid sequences listed in Table 2, or Table 13, or a substantially similar sequence thereof having at least 90%, at least 95%, at least 98% or at least 99% sequence identity thereto.

[0067] The present invention also provides nucleic acid molecules encoding any of the LCDR2 amino acid sequences listed in Table 1, or Table 12; in certain embodiments the nucleic acid molecule comprises a polynucleotide sequence selected from any of the LCDR2 nucleic acid sequences listed in Table 2, or Table 13, or a substantially similar sequence thereof having at least 90%, at least 95%, at least 98% or at least 99% sequence identity thereto.

[0068] The present invention also provides nucleic acid molecules encoding any of the LCDR3 amino acid sequences listed in Table 1, or Table 12; in certain embodiments the nucleic acid molecule comprises a polynucleotide sequence selected from any of the LCDR3 nucleic acid sequences listed in Table 2, or Table 13, or a substantially similar sequence thereof having at least 90%, at least 95%, at least 98% or at least 99% sequence identity thereto.

[0069] The present invention also provides nucleic acid molecules encoding an HCVR, wherein the HCVR comprises a set of three CDRs (*i.e.*, HCDR1-HCDR2-HCDR3), wherein the HCDR1-HCDR2-HCDR3 amino acid sequence set is as defined by any of the exemplary anti-influenza HA antibodies listed in Table 1, or Table 12.

[0070] The present invention also provides nucleic acid molecules encoding an LCVR, wherein the LCVR comprises a set of three CDRs (*i.e.*, LCDR1-LCDR2-LCDR3), wherein the LCDR1-LCDR2-LCDR3 amino acid sequence set is as defined by any of the exemplary anti-influenza HA antibodies listed in Table 1, or Table 12.

[0071] The present invention also provides nucleic acid molecules encoding both an HCVR and an LCVR, wherein the HCVR comprises an amino acid sequence of any of the HCVR amino acid sequences listed in Table 1, or Table 12 and wherein the LCVR comprises an amino acid sequence of any of the LCVR amino acid sequences listed in Table 1, or Table 12. In certain embodiments, the nucleic acid molecule comprises a polynucleotide sequence selected from any of the HCVR nucleic acid sequences listed in Table 2, or Table 13, or a substantially similar sequence thereof having at least 90%, at least 95%, at least 98% or at least 99% sequence identity thereto, and a polynucleotide sequence selected from any of the LCVR nucleic acid sequences listed in Table 2, or a substantially similar sequence thereof having at least 90%, at least 95%, at least 98% or at least 99% sequence identity thereto. In certain embodiments according to this aspect of the invention, the nucleic acid molecule encodes an HCVR and LCVR, wherein the HCVR and LCVR are both derived from the same anti-influenza HA antibody listed in Table 1, or Table 12.

[0072] The present invention provides nucleic acid molecules encoding any of the heavy chain amino acid sequences listed in Table 1, or Table 12. The present invention also provides

nucleic acid molecules encoding any of the light chain amino acid sequences listed in Table 1, or Table 12.

[0073] In a related aspect, the present invention provides recombinant expression vectors capable of expressing a polypeptide comprising a heavy or light chain variable region of an anti-influenza HA antibody. For example, the present invention includes recombinant expression vectors comprising any of the nucleic acid molecules mentioned above, *i.e.*, nucleic acid molecules encoding any of the HCVR, LCVR, and/or CDR sequences as set forth in Table 1, or Table 12. Also included within the scope of the present invention are host cells into which such vectors have been introduced, as well as methods of producing the antibodies or portions thereof by culturing the host cells under conditions permitting production of the antibodies or antibody fragments, and recovering the antibodies and antibody fragments so produced.

[0074] In a third aspect, the invention provides a pharmaceutical composition comprising a therapeutically effective amount of at least one recombinant monoclonal antibody or antigen-binding fragment thereof which specifically binds influenza HA and a pharmaceutically acceptable carrier. In a related aspect, the invention features a composition, which is a combination of an anti-influenza HA antibody and a second therapeutic agent. In one embodiment, the second therapeutic agent is any agent that is advantageously combined with an anti-influenza HA antibody. Exemplary agents that may be advantageously combined with an anti-influenza HA antibody include, without limitation, other agents that bind and/or inhibit influenza HA activity (including other antibodies or antigen-binding fragments thereof, etc.) and/or agents, which do not directly bind influenza HA but nonetheless inhibit viral activity including infectivity of host cells. In certain embodiments, the invention provides for a pharmaceutical composition comprising: (a) a first anti-influenza HA antibody or antigen-binding fragment thereof; (b) a second anti-influenza HA antibody or antigen-binding fragment thereof, wherein the first antibody binds to a first epitope on influenza HA and the second antibody binds to a second epitope on influenza HA wherein the first and second epitopes are distinct and non-overlapping; and (c) a pharmaceutically acceptable carrier or diluent. In certain embodiments, the invention provides for a pharmaceutical composition comprising: (a) a first anti-influenza HA antibody or antigen-binding fragment thereof; (b) a second anti-influenza HA antibody or antigen-binding fragment thereof, wherein the first antibody does not cross-compete with the second antibody for binding to influenza HA; and (c) a pharmaceutically acceptable carrier or diluent. In certain embodiments, the invention provides for a pharmaceutical composition comprising: (a) a first anti-influenza HA antibody or antigen-binding fragment thereof; (b) a second anti-influenza antibody or antigen-binding fragment thereof, which interacts with a different influenza antigen, wherein the first antibody binds to an epitope on influenza HA and the second antibody binds to an epitope on a different influenza antigen; and (c) a pharmaceutically acceptable carrier or diluent. In certain embodiments, the invention provides for a pharmaceutical composition comprising: (a) a first anti-influenza HA antibody or antigen-

binding fragment thereof; (b) a second antibody or antigen-binding fragment thereof, which interacts with a different viral (non-influenza) antigen, wherein the first antibody binds to an epitope on influenza HA and the second antibody binds to an epitope on a different viral (non-influenza) antigen; and (c) a pharmaceutically acceptable carrier or diluent. Additional combination therapies and co-formulations involving the anti-influenza HA antibodies of the present invention are disclosed elsewhere herein.

[0075] In a fourth aspect, the invention provides therapeutic methods for treating a disease or disorder associated with influenza HA (such as viral infection in a subject), or at least one symptom associated with the viral infection, using an anti-influenza HA antibody or antigen-binding portion of an antibody of the invention, wherein the therapeutic methods comprise administering a therapeutically effective amount of a pharmaceutical composition comprising an antibody or antigen-binding fragment of an antibody of the invention to the subject in need thereof. The disorder treated is any disease or condition, which is improved, ameliorated, inhibited or prevented by inhibition of influenza HA activity. In certain embodiments, the invention provides methods to prevent, treat or ameliorate at least one symptom of influenza A infection, the method comprising administering a therapeutically effective amount of an anti-influenza HA antibody or antigen-binding fragment thereof of the invention to a subject in need thereof.

[0076] In some embodiments, the present invention provides methods to ameliorate or reduce the severity, duration, or frequency of occurrence, of at least one symptom of influenza infection in a subject by administering an anti-influenza HA antibody of the invention, wherein the at least one symptom is selected from the group consisting of headache, fever, aches, rhinorrhea (nasal congestion), chills, fatigue, weakness, sore throat, cough, shortness of breath, vomiting, diarrhea, pneumonia, bronchitis, and death.

[0077] In certain embodiments, the invention provides methods to decrease viral load in a subject, the methods comprising administering to the subject an effective amount of an antibody or fragment thereof of the invention that binds influenza HA and blocks influenza HA binding and/or entry into the host cell.

[0078] In certain embodiments, the antibody or antigen-binding fragment thereof may be administered prophylactically or therapeutically to a subject having, or at risk of having, or predisposed to developing an influenza infection. The subjects at risk include, but are not limited to, an immunocompromised person, for example, a person who is immunocompromised because of autoimmune disease, or those persons receiving immunosuppressive therapy (for example, following organ transplant), or those persons afflicted with human immunodeficiency syndrome (HIV) or acquired immune deficiency syndrome (AIDS), certain forms of anemia that deplete or destroy white blood cells, those persons receiving radiation or chemotherapy, or those persons afflicted with an inflammatory disorder. Other subjects at risk for acquiring an influenza infection include an elderly adult (more than 65 years of age), children younger than 2 years of

age, healthcare workers, and people with underlying medical conditions such as pulmonary infection, heart disease or diabetes. Also, any person who comes into physical contact or close physical proximity with an infected individual has an increased risk of developing an influenza virus infection. Moreover, a subject is at risk of contracting an influenza infection due to proximity to an outbreak of the disease, e.g. subject resides in a densely-populated city or in close proximity to subjects having confirmed or suspected infections of influenza virus, or choice of employment, e.g. hospital worker, pharmaceutical researcher, traveler to infected area, or frequent flier.

[0079] In certain embodiments, the antibody or antigen-binding fragment thereof of the invention is administered in combination with a second therapeutic agent to the subject in need thereof. The second therapeutic agent may be selected from the group consisting of an anti-inflammatory drug (such as corticosteroids, and non-steroidal anti-inflammatory drugs), an anti-infective drug, a different antibody to influenza HA, an antibody to a different influenza antigen (e.g. the neuraminidase), an anti-viral drug, a decongestant, an anti-histamine, a vaccine for influenza, a dietary supplement such as anti-oxidants and any other drug or therapy known in the art useful for ameliorating at least one symptom of the influenza infection, or for reducing the viral load in a patient. In certain embodiments, the second therapeutic agent may be an agent that helps to counteract or reduce any possible side effect(s) associated with an antibody or antigen-binding fragment thereof of the invention, if such side effect(s) should occur. The antibody or fragment thereof may be administered subcutaneously, intravenously, intradermally, intraperitoneally, orally, intranasally, intramuscularly, or intracranially. In one embodiment, the antibody may be administered as a single intravenous infusion for maximum concentration of the antibody in the serum of the subject. The antibody or fragment thereof may be administered at a dose of about 0.01 mg/kg of body weight to about 100 mg/kg of body weight of the subject. In certain embodiments, an antibody of the present invention may be administered at one or more doses comprising between 50 mg to 5000 mg.

[0080] The present invention also includes use of an anti-influenza HA antibody or antigen-binding fragment thereof of the invention for treating a disease or disorder that would benefit from the blockade of influenza HA binding and/or activity. The present invention also includes use of an anti- influenza HA antibody or antigen-binding fragment thereof of the invention in the manufacture of a medicament for the treatment of a disease or disorder that would benefit from the blockade of influenza HA binding and/or activity.

[0081] Other embodiments will become apparent from a review of the ensuing detailed description.

BRIEF DESCRIPTION OF THE FIGURES

[0082] Figure 1. Shows that a single dose of H1H11729P at 48 hrs post infection demonstrates greater efficacy than oseltamivir at 48 hrs post infection in treating severe influenza A virus

infection in mice. A single dose of H1H11729P at 15 mg/kg (circles) given 48 h p.i. is more efficacious than oseltamivir (TAMIFLU®) dosed twice daily (BID) for 5 days starting on day 2 post infection at 25 (inverse triangles) or 5 mg/kg (diamonds). Mice were infected intranasally (IN) on day 0 with 10 x MLD₅₀ of A/Puerto Rico/08/1934 (H1N1). Control groups included an uninfected (triangles, dotted line) and infected (hexagons) group that received oral gavage of water and CR8020 as an IgG1 isotype (negative) control.

[0083] Figure 2. Shows that additive efficacy is observed when a single dose of H1H11729P is combined with oseltamivir at 72 hrs post infection to treat severe influenza in mice. Mice received a single sub-efficacious dose of 7 (squares, dotted line) or 15 mg/kg of H1H11729P (circles, dotted line), control IgG (triangles), 25 mg/kg BID oseltamivir for 5 days (diamonds, dotted line) or a combination of a single dose of 7 (squares, solid line) or 15 mg/kg H1H11729P (circles, solid line) and the regimen of oseltamivir for 5 days 72 h after intranasal (IN) infection with 10 x MLD₅₀ of A/Puerto Rico/08/1934 (H1N1). Results of three independent trials (N=15 per group) are shown.

[0084] Figure 3. Shows the HCVR, HCDR1, HCDR2, HCDR3, LCVR, LCDR1, LCDR2 and LCDR3 amino acid sequences for the antibody designated H1H11729P.

[0085] Figure 4. Shows the HCVR, HCDR1, HCDR2, HCDR3, LCVR, LCDR1, LCDR2 and LCDR3 amino acid sequences for the antibody designated H1H11829N2.

DETAILED DESCRIPTION

[0086] Before the present methods are described, it is to be understood that this invention is not limited to particular methods, and experimental conditions described, as such methods and conditions may vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to be limiting, since the scope of the present invention will be limited only by the appended claims.

[0087] Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, preferred methods and materials are now described.

Definitions

[0088] The term “influenza hemagglutinin”, also called “influenza HA” is a trimeric glycoprotein found on the surface of influenza virions, which mediates viral attachment (via HA1 binding to α -2,3- and α -2,6-sialic acids) and entry (through conformational change) into host cells. The HA is comprised of two structural domains: a globular head domain containing the receptor binding site (subject to high frequency of antigenic mutations) and the stem region (more conserved among various strains of influenza virus). The influenza HA is synthesized as a precursor (HA0) that undergoes proteolytic processing to produce two subunits (HA1 and HA2) which

associate with one another to form the stem/globular head structure. The viral HA is the most variable antigen on the virus (18 subtypes can be classified into two groups), but the stem (HA2) is highly conserved within each group.

[0089] The amino acid sequence of full-length Influenza HA is exemplified by the amino acid sequence of influenza isolate H1N1 A/California/04/2009 provided in GenBank as accession number FJ966082.1. The term “influenza-HA” also includes protein variants of influenza HA isolated from different influenza isolates, *e.g.*, GQ149237.1, NC_002017, KM972981.1, etc. The term “influenza-HA” also includes recombinant influenza HA or a fragment thereof. The term also encompasses influenza HA or a fragment thereof coupled to, for example, histidine tag, mouse or human Fc, or a signal sequence.

[0090] The term “influenza infection”, as used herein, also characterized as “flu” refers to the severe acute respiratory illness caused by influenza virus. The term includes respiratory tract infection and the symptoms that include high fever, headache, general aches and pains, fatigue and weakness, in some instances extreme exhaustion, stuffy nose, sneezing, sore throat, chest discomfort, cough, shortness of breath, bronchitis, pneumonia and death in severe cases.

[0091] The term “antibody”, as used herein, is intended to refer to immunoglobulin molecules comprised of four polypeptide chains, two heavy (H) chains and two light (L) chains interconnected by disulfide bonds (*i.e.*, “full antibody molecules”), as well as multimers thereof (*e.g.* IgM) or antigen-binding fragments thereof. Each heavy chain is comprised of a heavy chain variable region (“HCVR” or “V_H”) and a heavy chain constant region (comprised of domains C_{H1}, C_{H2} and C_{H3}). Each light chain is comprised of a light chain variable region (“LCVR” or “V_L”) and a light chain constant region (C_L). The V_H and V_L regions can be further subdivided into regions of hypervariability, termed complementarity determining regions (CDR), interspersed with regions that are more conserved, termed framework regions (FR). Each V_H and V_L is composed of three CDRs and four FRs, arranged from amino-terminus to carboxy-terminus in the following order: FR1, CDR1, FR2, CDR2, FR3, CDR3, FR4. In certain embodiments of the invention, the FRs of the antibody (or antigen binding fragment thereof) may be identical to the human germline sequences, or may be naturally or artificially modified. An amino acid consensus sequence may be defined based on a side-by-side analysis of two or more CDRs.

[0092] Substitution of one or more CDR residues or omission of one or more CDRs is also possible. Antibodies have been described in the scientific literature in which one or two CDRs can be dispensed with for binding. Padlan *et al.* (1995 FASEB J. 9:133-139) analyzed the contact regions between antibodies and their antigens, based on published crystal structures, and concluded that only about one fifth to one third of CDR residues actually contact the antigen. Padlan also found many antibodies in which one or two CDRs had no amino acids in contact with an antigen (see also, Vajdos *et al.* 2002 J Mol Biol 320:415-428).

[0093] CDR residues not contacting antigen can be identified based on previous studies (for example residues H60-H65 in CDRH2 are often not required), from regions of Kabat CDRs

lying outside Chothia CDRs, by molecular modeling and/or empirically. If a CDR or residue(s) thereof is omitted, it is usually substituted with an amino acid occupying the corresponding position in another human antibody sequence or a consensus of such sequences. Positions for substitution within CDRs and amino acids to substitute can also be selected empirically. Empirical substitutions can be conservative or non-conservative substitutions.

[0094] The fully human anti-influenza-HA monoclonal antibodies disclosed herein may comprise one or more amino acid substitutions, insertions and/or deletions in the framework and/or CDR regions of the heavy and light chain variable domains as compared to the corresponding germline sequences. Such mutations can be readily ascertained by comparing the amino acid sequences disclosed herein to germline sequences available from, for example, public antibody sequence databases. The present invention includes antibodies, and antigen-binding fragments thereof, which are derived from any of the amino acid sequences disclosed herein, wherein one or more amino acids within one or more framework and/or CDR regions are mutated to the corresponding residue(s) of the germline sequence from which the antibody was derived, or to the corresponding residue(s) of another human germline sequence, or to a conservative amino acid substitution of the corresponding germline residue(s) (such sequence changes are referred to herein collectively as "germline mutations"). A person of ordinary skill in the art, starting with the heavy and light chain variable region sequences disclosed herein, can easily produce numerous antibodies and antigen-binding fragments which comprise one or more individual germline mutations or combinations thereof. In certain embodiments, all of the framework and/or CDR residues within the V_H and/or V_L domains are mutated back to the residues found in the original germline sequence from which the antibody was derived. In other embodiments, only certain residues are mutated back to the original germline sequence, *e.g.*, only the mutated residues found within the first 8 amino acids of FR1 or within the last 8 amino acids of FR4, or only the mutated residues found within CDR1, CDR2 or CDR3. In other embodiments, one or more of the framework and/or CDR residue(s) are mutated to the corresponding residue(s) of a different germline sequence (*i.e.*, a germline sequence that is different from the germline sequence from which the antibody was originally derived). Furthermore, the antibodies of the present invention may contain any combination of two or more germline mutations within the framework and/or CDR regions, *e.g.*, wherein certain individual residues are mutated to the corresponding residue of a particular germline sequence while certain other residues that differ from the original germline sequence are maintained or are mutated to the corresponding residue of a different germline sequence. Once obtained, antibodies and antigen-binding fragments that contain one or more germline mutations can be easily tested for one or more desired properties such as, improved binding specificity, increased binding affinity, improved or enhanced antagonistic or agonistic biological properties (as the case may be), reduced immunogenicity, etc. Antibodies and antigen-binding fragments obtained in this general manner are encompassed within the present invention.

[0095] The present invention also includes fully human anti-influenza-HA monoclonal antibodies comprising variants of any of the HCVR, LCVR, and/or CDR amino acid sequences disclosed herein having one or more conservative substitutions. For example, the present invention includes anti-influenza-HA antibodies having HCVR, LCVR, and/or CDR amino acid sequences with, *e.g.*, 10 or fewer, 8 or fewer, 6 or fewer, 4 or fewer, etc. conservative amino acid substitutions relative to any of the HCVR, LCVR, and/or CDR amino acid sequences disclosed herein.

[0096] The term "human antibody", as used herein, is intended to include antibodies having variable and constant regions derived from human germline immunoglobulin sequences. The human mAbs of the invention may include amino acid residues not encoded by human germline immunoglobulin sequences (*e.g.*, mutations introduced by random or site-specific mutagenesis *in vitro* or by somatic mutation *in vivo*), for example in the CDRs and in particular CDR3. However, the term "human antibody", as used herein, is not intended to include mAbs in which CDR sequences derived from the germline of another mammalian species (*e.g.*, mouse), have been grafted onto human FR sequences. The term includes antibodies recombinantly produced in a non-human mammal, or in cells of a non-human mammal. The term is not intended to include antibodies isolated from or generated in a human subject.

[0097] The term "recombinant", as used herein, refers to antibodies or antigen-binding fragments thereof of the invention created, expressed, isolated or obtained by technologies or methods known in the art as recombinant DNA technology which include, *e.g.*, DNA splicing and transgenic expression. The term refers to antibodies expressed in a non-human mammal (including transgenic non-human mammals, *e.g.*, transgenic mice), or a cell (*e.g.*, CHO cells) expression system or isolated from a recombinant combinatorial human antibody library.

[0098] The term "specifically binds," or "binds specifically to", or the like, means that an antibody or antigen-binding fragment thereof forms a complex with an antigen that is relatively stable under physiologic conditions. Specific binding can be characterized by an equilibrium dissociation constant of at least about 1×10^{-8} M or less (*e.g.*, a smaller K_D denotes a tighter binding). Methods for determining whether two molecules specifically bind are well known in the art and include, for example, equilibrium dialysis, surface plasmon resonance, and the like. As described herein, antibodies have been identified by real-time, label free bio-layer interferometry assay on an Octet® HTX biosensor, which bind specifically to influenza-HA. Moreover, multi-specific antibodies that bind to one domain in influenza-HA and one or more additional antigens or a bi-specific that binds to two different regions of influenza-HA are nonetheless considered antibodies that "specifically bind", as used herein.

[0099] The term "high affinity" antibody refers to those mAbs having a binding affinity to influenza-HA, expressed as K_D , of at least 10^{-8} M; preferably 10^{-9} M; more preferably 10^{-10} M, even more preferably 10^{-11} M, even more preferably 10^{-12} M, as measured by real-time, label free bio-layer interferometry assay, *e.g.*, an Octet® HTX biosensor, or by surface plasmon

resonance, *e.g.*, BIACORE™, or by solution-affinity ELISA.

[0100] By the term “slow off rate”, “K_{off}” or “k_d” is meant an antibody that dissociates from influenza-HA, with a rate constant of $1 \times 10^{-3} \text{ s}^{-1}$ or less, preferably $1 \times 10^{-4} \text{ s}^{-1}$ or less, as determined by real-time, label free bio-layer interferometry assay, *e.g.*, an Octet® HTX biosensor, or by surface plasmon resonance, *e.g.*, BIACORE™.

[0101] The terms “antigen-binding portion” of an antibody, “antigen-binding fragment” of an antibody, and the like, as used herein, include any naturally occurring, enzymatically obtainable, synthetic, or genetically engineered polypeptide or glycoprotein that specifically binds an antigen to form a complex. The terms “antigen-binding fragment” of an antibody, or “antibody fragment”, as used herein, refers to one or more fragments of an antibody that retain the ability to bind to Influenza HA.

[0102] In specific embodiments, antibody or antibody fragments of the invention may be conjugated to a moiety such a ligand or a therapeutic moiety (“immunoconjugate”), such as an anti-viral drug, a second anti-influenza antibody, or any other therapeutic moiety useful for treating an infection caused by influenza-HA.

[0103] An “isolated antibody”, as used herein, is intended to refer to an antibody that is substantially free of other antibodies (Abs) having different antigenic specificities (*e.g.*, an isolated antibody that specifically binds influenza-HA, or a fragment thereof, is substantially free of Abs that specifically bind antigens other than influenza-HA).

[0104] A “blocking antibody” or a “neutralizing antibody”, as used herein (or an “antibody that neutralizes influenza-HA activity” or “antagonist antibody”), is intended to refer to an antibody whose binding to influenza-HA results in inhibition of at least one biological activity of influenza-HA. For example, an antibody of the invention may prevent or block influenza attachment to, or entry into a host cell. In addition, a “neutralizing antibody” is one that can neutralize, *i.e.*, prevent, inhibit, reduce, impede or interfere with, the ability of a pathogen to initiate and/or perpetuate an infection in a host. The terms “neutralizing antibody” and “an antibody that neutralizes” or “antibodies that neutralize” are used interchangeably herein. These antibodies can be used, alone or in combination, as prophylactic or therapeutic agents with other anti-viral agents upon appropriate formulation, or in association with active vaccination, or as a diagnostic tool.

[0105] The term “surface plasmon resonance”, refers to an optical phenomenon that allows for the analysis of real-time biomolecular interactions by detection of alterations in protein concentrations within a biosensor matrix, for example using the BIACORE™ system (Pharmacia Biosensor AB, Uppsala, Sweden and Piscataway, N.J)

[0106] Bio-layer interferometry is a label-free technology for measuring biomolecular interactions. It is an optical analytical technique that analyzes the interference pattern of white light reflected from two surfaces: a layer of immobilized protein on the biosensor tip, and an internal reference layer. Any change in the number of molecules bound to the biosensor tip

causes a shift in the interference pattern that can be measured in real-time (Abdiche, Y.N., *et al.* Analytical Biochemistry, (2008), 377(2), 209-217). In certain embodiments of the invention, a "real-time bio-layer interferometer based biosensor (Octet HTX assay)" was used to assess the binding characteristics of certain of the anti-influenza HA antibodies.

[0107] The term " K_D ", as used herein, is intended to refer to the equilibrium dissociation constant of a particular antibody-antigen interaction.

[0108] The term "epitope" refers to an antigenic determinant that interacts with a specific antigen-binding site in the variable region of an antibody molecule known as a paratope. A single antigen may have more than one epitope. Thus, different antibodies may bind to different areas on an antigen and may have different biological effects. The term "epitope" also refers to a site on an antigen to which B and/or T cells respond. It also refers to a region of an antigen that is bound by an antibody. Epitopes may be defined as structural or functional. Functional epitopes are generally a subset of the structural epitopes and have those residues that directly contribute to the affinity of the interaction. Epitopes may also be conformational, that is, composed of non-linear amino acids. In certain embodiments, epitopes may include determinants that are chemically active surface groupings of molecules such as amino acids, sugar side chains, phosphoryl groups, or sulfonyl groups, and, in certain embodiments, may have specific three-dimensional structural characteristics, and/or specific charge characteristics.

[0109] The term "cross-competes", as used herein, means an antibody or antigen-binding fragment thereof binds to an antigen and inhibits or blocks the binding of another antibody or antigen-binding fragment thereof. The term also includes competition between two antibodies in both orientations, *i.e.*, a first antibody that binds and blocks binding of second antibody and vice-versa. In certain embodiments, the first antibody and second antibody may bind to the same epitope. Alternatively, the first and second antibodies may bind to different, but overlapping epitopes such that binding of one inhibits or blocks the binding of the second antibody, *e.g.*, *via* steric hindrance. Cross-competition between antibodies may be measured by methods known in the art, for example, by a real-time, label-free bio-layer interferometry assay. Cross-competition between two antibodies may be expressed as the binding of the second antibody that is less than the background signal due to self-self binding (wherein first and second antibodies is the same antibody). Cross-competition between 2 antibodies may be expressed, for example, as % binding of the second antibody that is less than the baseline self-self background binding (wherein first and second antibodies is the same antibody).

[0110] The term "substantial identity" or "substantially identical," when referring to a nucleic acid or fragment thereof, indicates that, when optimally aligned with appropriate nucleotide insertions or deletions with another nucleic acid (or its complementary strand), there is nucleotide sequence identity in at least about 90%, and more preferably at least about 95%, 96%, 97%, 98% or 99% of the nucleotide bases, as measured by any well-known algorithm of sequence identity, such as FASTA, BLAST or GAP, as discussed below. A nucleic acid

molecule having substantial identity to a reference nucleic acid molecule may, in certain instances, encode a polypeptide having the same or substantially similar amino acid sequence as the polypeptide encoded by the reference nucleic acid molecule.

[0111] As applied to polypeptides, the term "substantial similarity" or "substantially similar" means that two peptide sequences, when optimally aligned, such as by the programs GAP or BESTFIT using default gap weights, share at least 90% sequence identity, even more preferably at least 95%, 98% or 99% sequence identity. Preferably, residue positions, which are not identical, differ by conservative amino acid substitutions. A "conservative amino acid substitution" is one in which an amino acid residue is substituted by another amino acid residue having a side chain (R group) with similar chemical properties (e.g., charge or hydrophobicity). In general, a conservative amino acid substitution will not substantially change the functional properties of a protein. In cases where two or more amino acid sequences differ from each other by conservative substitutions, the percent or degree of similarity may be adjusted upwards to correct for the conservative nature of the substitution. Means for making this adjustment are well known to those of skill in the art. (See, e.g., Pearson (1994) *Methods Mol. Biol.* 24: 307-331). Examples of groups of amino acids that have side chains with similar chemical properties include 1) aliphatic side chains: glycine, alanine, valine, leucine and isoleucine; 2) aliphatic-hydroxyl side chains: serine and threonine; 3) amide-containing side chains: asparagine and glutamine; 4) aromatic side chains: phenylalanine, tyrosine, and tryptophan; 5) basic side chains: lysine, arginine, and histidine; 6) acidic side chains: aspartate and glutamate, and 7) sulfur-containing side chains: cysteine and methionine. Preferred conservative amino acids substitution groups are: valine-leucine-isoleucine, phenylalanine-tyrosine, lysine-arginine, alanine-valine, glutamate-aspartate, and asparagine-glutamine. Alternatively, a conservative replacement is any change having a positive value in the PAM250 log-likelihood matrix disclosed in Gonnet *et al.* (1992) *Science* 256: 1443-45. A "moderately conservative" replacement is any change having a nonnegative value in the PAM250 log-likelihood matrix.

[0112] Sequence similarity for polypeptides is typically measured using sequence analysis software. Protein analysis software matches similar sequences using measures of similarity assigned to various substitutions, deletions and other modifications, including conservative amino acid substitutions. For instance, GCG software contains programs such as GAP and BESTFIT which can be used with default parameters to determine sequence homology or sequence identity between closely related polypeptides, such as homologous polypeptides from different species of organisms or between a wild type protein and a mutin thereof. See, e.g., GCG Version 6.1. Polypeptide sequences also can be compared using FASTA with default or recommended parameters; a program in GCG Version 6.1. FASTA (e.g., FASTA2 and FASTA3) provides alignments and percent sequence identity of the regions of the best overlap between the query and search sequences (Pearson (2000) *supra*). Another preferred algorithm when comparing a sequence of the invention to a database containing a large number of

sequences from different organisms is the computer program BLAST, especially BLASTP or TBLASTN, using default parameters. See, e.g., Altschul *et al.* (1990) J. Mol. Biol. 215: 403-410 and (1997) Nucleic Acids Res. 25:3389-3402.

[0113] By the phrase “therapeutically effective amount” is meant an amount that produces the desired effect for which it is administered. The exact amount will depend on the purpose of the treatment, and will be ascertainable by one skilled in the art using known techniques (see, for example, Lloyd (1999) The Art, Science and Technology of Pharmaceutical Compounding).

[0114] As used herein, the term “subject” refers to an animal, preferably a mammal, more preferably a human, in need of amelioration, prevention and/or treatment of a disease or disorder such as viral infection. The subject may have an influenza infection or is predisposed to developing an influenza virus infection. Subjects “predisposed to developing an influenza virus infection”, or subjects “who may be at elevated risk for contracting an influenza virus infection”, are those subjects with compromised immune systems because of autoimmune disease, those persons receiving immunosuppressive therapy (for example, following organ transplant), those persons afflicted with human immunodeficiency syndrome (HIV) or acquired immune deficiency syndrome (AIDS), certain forms of anemia that deplete or destroy white blood cells, those persons receiving radiation or chemotherapy, or those persons afflicted with an inflammatory disorder. Additionally, subject of extreme young or old age are at increased risk. Any person who comes into physical contact or close physical proximity with an infected individual has an increased risk of developing an Influenza virus infection. Moreover, a subject is at risk of contracting an influenza infection due to proximity to an outbreak of the disease, e.g. subject resides in a densely-populated city or in close proximity to subjects having confirmed or suspected infections of Influenza virus, or choice of employment, e.g. hospital worker, pharmaceutical researcher, traveler to infected area, or frequent flier.

[0115] As used herein, the terms “treat”, “treating”, or “treatment” refer to the reduction or amelioration of the severity of at least one symptom or indication of influenza infection due to the administration of a therapeutic agent such as an antibody of the present invention to a subject in need thereof. The terms include inhibition of progression of disease or of worsening of infection. The terms also include positive prognosis of disease, i.e., the subject may be free of infection or may have reduced or no viral titers upon administration of a therapeutic agent such as an antibody of the present invention. The therapeutic agent may be administered at a therapeutic dose to the subject.

[0116] The terms “prevent”, “preventing” or “prevention” refer to inhibition of manifestation of influenza infection or any symptoms or indications of influenza infection upon administration of an antibody of the present invention. The term includes prevention of spread of infection in a subject exposed to the virus or at risk of having influenza infection.

[0117] As used herein a “protective effect” may be demonstrated by any standard procedure known in the art to determine whether an agent such as an anti-viral agent, or an antibody such

as an anti-influenza-HA antibody of the invention can demonstrate any one or more of the following: *e.g.* an increase in survival after exposure to an infectious agent, a decrease in viral load, or amelioration of at least one symptom associated with the infectious agent.

[0118] As used herein, the term “anti-viral drug” refers to any anti-infective drug or therapy used to treat, prevent, or ameliorate a viral infection in a subject. The term “anti-viral drug” includes, but is not limited to TAMIFLU® (Oseltamivir), RELENZA® (Zanamivir), ribavirin, or interferon-alpha2b. In the present invention, the infection to be treated is caused by an influenza virus.

General Description

[0119] Influenza is an infectious disease caused by RNA viruses of the family Orthomyxoviridae (the influenza viruses). Influenza viruses are classified based on core protein into three genera A, B and C that are further divided into subtypes determined by the viral envelope glycoproteins hemagglutinin (HA) and neuraminidase (NA). Influenza A viruses infect a range of mammalian and avian species, whereas type B and C infections are largely restricted to humans. Only types A and B cause human disease of any concern.

[0120] High mutation rates and frequent genetic reassortments of the influenza viruses contribute to great variability of the HA and NA antigens. Minor point mutations causing small changes (“antigenic drift”) occur relatively often. Antigenic drift enables the virus to evade immune recognition, resulting in repeated influenza outbreaks during interpandemic years. Major changes in the HA antigen (“antigenic shift”) are caused by reassortment of genetic material from different influenza A subtypes. Antigenic shifts resulting in new pandemic strains are rare events, occurring through reassortment between animal and human subtypes, for example in co-infected pigs.

[0121] The neutralizing antibody response to Influenza A virus is typically specific for a given viral subtype. There are 18 influenza A subtypes defined by their hemagglutinin (“HA”) proteins. The 18 HAs, H1-H18, can be classified into two groups. Group 1 consists of H1, H2, H5, H6, H8, H9, H11, H12, H13, H16, H17 and H18 subtypes, and group 2 includes H3, H4, H7, H10, H14 and H15 subtypes. For these reasons it would be highly desirable to have a vaccine that induces broadly neutralizing antibodies capable of neutralizing all influenza A virus subtypes as well as their yearly variants. In addition broadly neutralizing heterosubtypic antibodies could be administered as medicaments for prevention or therapy of influenza A infection.

[0122] HA is synthesized as a homo-trimeric precursor polypeptide HA0. Each monomer can be independently cleaved post-translationally to form two polypeptides, HA1 and HA2, linked by a single disulphide bond. The larger N-terminal fragment (HA1 320-330 amino acids) forms a membrane-distal globular domain that contains the receptor-binding site and most determinants recognized by virus-neutralizing antibodies. The HA1 polypeptide of HA is responsible for the attachment of virus to the cell surface. The smaller C-terminal portion (HA2, approximately 180 amino acids) forms a stem-like structure that anchors the globular domain to the cellular or viral membrane. The HA2 polypeptide mediates the fusion of viral and cell membranes in

endosomes, allowing the release of the ribonucleoprotein complex into the cytoplasm.

[0123] There has only been limited success in identifying antibodies that neutralize more than one subtype of influenza A virus. Further, the breadth of neutralization of antibodies identified thus far is narrow and their potency is low. Okuno *et al.*, immunized mice with influenza virus A/Okuda/57 (H2N2) and isolated a monoclonal antibody (C179) that binds to a conserved conformational epitope in HA2 and neutralizes the Group 1 H2, H1 and H5 subtype influenza A viruses in vitro and in vivo in animal models ((Okuno *et al.*, J. Virol. 67:2552-8, 1993).

[0124] Despite decades of research, there are no marketed antibodies that broadly neutralize or inhibit influenza A virus infection or attenuate disease caused by influenza A virus. Therefore, there is a need to identify new antibodies that neutralize multiple subtypes of influenza A virus and can be used as medicaments for prevention or therapy of influenza A infection.

[0125] Passive immunotherapy for prophylaxis or treatment of infectious diseases has been used for more than a century, usually in the form of convalescent human sera that contains high titers of neutralizing antibodies (Good *et al.* 1991; Cancer 68: 1415-1421). Today, multiple purified monoclonal antibodies are currently in preclinical and clinical development for use as anti-microbials (Marasco *et al.* 2007; Nature Biotechnology 25: 1421-1434).

[0126] The inventors have described herein fully human antibodies and antigen-binding fragments thereof that specifically bind to influenza hemagglutinin and modulate the interaction of influenza virus with host cells. The anti-influenza HA antibodies may bind to the influenza virus HA with high affinity. In certain embodiments, the antibodies of the present invention are blocking antibodies wherein the antibodies may bind to influenza HA and block the attachment to and/or entry of the virus into host cells. In some embodiments, the blocking antibodies of the invention may block the binding of influenza virus to cells and as such may inhibit or neutralize viral infectivity of host cells. In some embodiments, the blocking antibodies may be useful for treating a subject suffering from an influenza virus infection. The antibodies when administered to a subject in need thereof may reduce the infection by a virus such as influenza in the subject. They may be used to decrease viral loads in a subject. They may be used alone or as adjunct therapy with other therapeutic moieties or modalities known in the art for treating a viral infection. In certain embodiments, these antibodies may bind to an epitope in the stem region of the viral HA. Furthermore, the identified antibodies can be used prophylactically (before infection) to protect a mammal from infection, or can be used therapeutically (after infection is established) to ameliorate a previously established infection, or to ameliorate at least one symptom associated with the infection.

[0127] The full-length amino acid sequence of an exemplary Influenza HA is shown in GenBank as accession number HC483324.1 (See SEQ ID NO: 62 in PCT publication WO2010/027818).

[0128] In certain embodiments, the antibodies of the invention are obtained from mice immunized with a primary immunogen, such as a full length influenza HA or with a recombinant form of influenza HA or fragments thereof followed by immunization with a secondary

immunogen, or with an immunogenically active fragment of influenza HA. In certain embodiments, the antibodies are obtained from mice immunized with an influenza vaccine composition followed by booster immunization with one or more recombinantly produced HA peptides.

[0129] The immunogen may be a biologically active and/or immunogenic fragment of influenza HA or DNA encoding the active fragment thereof. The fragment may be derived from the stem region of the HA protein. (See Sui *et. al.*, Nature Struct. and Mol. Biol. Published online 22 Feb. 2009; Pages 1-9).

[0130] The peptides may be modified to include addition or substitution of certain residues for tagging or for purposes of conjugation to carrier molecules, such as, KLH. For example, a cysteine may be added at either the N terminal or C terminal end of a peptide, or a linker sequence may be added to prepare the peptide for conjugation to, for example, KLH for immunization.

[0131] Certain anti-influenza-HA antibodies of the present invention are able to bind to and neutralize the activity of influenza-HA, as determined by *in vitro* or *in vivo* assays. The ability of the antibodies of the invention to bind to and neutralize the activity of influenza-HA and thus the attachment and/or entry of the virus into a host cell followed by the ensuing viral infection, may be measured using any standard method known to those skilled in the art, including binding assays, or activity assays, as described herein.

[0132] Non-limiting, exemplary *in vitro* assays for measuring binding activity are illustrated in Example 3, herein. In Example 3, the binding affinity and dissociation constants of anti-influenza-HA antibodies for influenza-HA were determined by real-time bio-layer interferometer based biosensor (Octet HTX assay). In Examples 4 and 5, neutralization assays were used to determine infectivity of diverse group 1 strains of influenza virus. In Example 6, certain antibodies were shown to mediate complement dependent cytotoxicity (CDC) of virus-infected cells *in vitro*. Examples 7 and 10 demonstrate that certain antibodies of the invention are capable of neutralizing an influenza A infection *in vivo* when administered either prophylactically or therapeutically.

[0133] The antibodies specific for influenza-HA may contain no additional labels or moieties, or they may contain an N-terminal or C-terminal label or moiety. In one embodiment, the label or moiety is biotin. In a binding assay, the location of a label (if any) may determine the orientation of the peptide relative to the surface upon which the peptide is bound. For example, if a surface is coated with avidin, a peptide containing an N-terminal biotin will be oriented such that the C-terminal portion of the peptide will be distal to the surface. In one embodiment, the label may be a radionuclide, a fluorescent dye or a MRI-detectable label. In certain embodiments, such labeled antibodies may be used in diagnostic assays including imaging assays.

Antigen-Binding Fragments of Antibodies

[0134] Unless specifically indicated otherwise, the term "antibody," as used herein, shall be understood to encompass antibody molecules comprising two immunoglobulin heavy chains and two immunoglobulin light chains (*i.e.*, "full antibody molecules") as well as antigen-binding fragments thereof. The terms "antigen-binding portion" of an antibody, "antigen-binding fragment" of an antibody, and the like, as used herein, include any naturally occurring, enzymatically obtainable, synthetic, or genetically engineered polypeptide or glycoprotein that specifically binds an antigen to form a complex. The terms "antigen-binding fragment" of an antibody, or "antibody fragment", as used herein, refers to one or more fragments of an antibody that retain the ability to specifically bind to Influenza HA. An antibody fragment may include a Fab fragment, a F(ab')₂ fragment, a Fv fragment, a dAb fragment, a fragment containing a CDR, or an isolated CDR. In certain embodiments, the term "antigen-binding fragment" refers to a polypeptide fragment of a multi-specific antigen-binding molecule. Antigen-binding fragments of an antibody may be derived, *e.g.*, from full antibody molecules using any suitable standard techniques such as proteolytic digestion or recombinant genetic engineering techniques involving the manipulation and expression of DNA encoding antibody variable and (optionally) constant domains. Such DNA is known and/or is readily available from, *e.g.*, commercial sources, DNA libraries (including, *e.g.*, phage-antibody libraries), or can be synthesized. The DNA may be sequenced and manipulated chemically or by using molecular biology techniques, for example, to arrange one or more variable and/or constant domains into a suitable configuration, or to introduce codons, create cysteine residues, modify, add or delete amino acids, etc.

[0135] Non-limiting examples of antigen-binding fragments include: (i) Fab fragments; (ii) F(ab')₂ fragments; (iii) Fd fragments; (iv) Fv fragments; (v) single-chain Fv (scFv) molecules; (vi) dAb fragments; and (vii) minimal recognition units consisting of the amino acid residues that mimic the hypervariable region of an antibody (*e.g.*, an isolated complementarity determining region (CDR) such as a CDR3 peptide), or a constrained FR3-CDR3-FR4 peptide. Other engineered molecules, such as domain-specific antibodies, single domain antibodies, domain-deleted antibodies, chimeric antibodies, CDR-grafted antibodies, diabodies, triabodies, tetrabodies, minibodies, nanobodies (*e.g.* monovalent nanobodies, bivalent nanobodies, etc.), small modular immunopharmaceuticals (SMIPs), and shark variable IgNAR domains, are also encompassed within the expression "antigen-binding fragment," as used herein.

[0136] An antigen-binding fragment of an antibody will typically comprise at least one variable domain. The variable domain may be of any size or amino acid composition and will generally comprise at least one CDR, which is adjacent to or in frame with one or more framework sequences. In antigen-binding fragments having a V_H domain associated with a V_L domain, the V_H and V_L domains may be situated relative to one another in any suitable arrangement. For

example, the variable region may be dimeric and contain $V_H - V_H$, $V_H - V_L$ or $V_L - V_L$ dimers. Alternatively, the antigen-binding fragment of an antibody may contain a monomeric V_H or V_L domain.

[0137] In certain embodiments, an antigen-binding fragment of an antibody may contain at least one variable domain covalently linked to at least one constant domain. Non-limiting, exemplary configurations of variable and constant domains that may be found within an antigen-binding fragment of an antibody of the present invention include: (i) $V_H - C_H1$; (ii) $V_H - C_H2$; (iii) $V_H - C_H3$; (iv) $V_H - C_H1 - C_H2$; (v) $V_H - C_H1 - C_H2 - C_H3$; (vi) $V_H - C_H2 - C_H3$; (vii) $V_H - C_L$; (viii) $V_L - C_H1$; (ix) $V_L - C_H2$; (x) $V_L - C_H3$; (xi) $V_L - C_H1 - C_H2$; (xii) $V_L - C_H1 - C_H2 - C_H3$; (xiii) $V_L - C_H2 - C_H3$; and (xiv) $V_L - C_L$. In any configuration of variable and constant domains, including any of the exemplary configurations listed above, the variable and constant domains may be either directly linked to one another or may be linked by a full or partial hinge or linker region. A hinge region may consist of at least 2 (*e.g.*, 5, 10, 15, 20, 40, 60 or more) amino acids, which result in a flexible or semi-flexible linkage between adjacent variable and/or constant domains in a single polypeptide molecule. Moreover, an antigen-binding fragment of an antibody of the present invention may comprise a homo-dimer or hetero-dimer (or other multimer) of any of the variable and constant domain configurations listed above in non-covalent association with one another and/or with one or more monomeric V_H or V_L domain (*e.g.*, by disulfide bond(s)).

[0138] As with full antibody molecules, antigen-binding fragments may be mono-specific or multi-specific (*e.g.*, bi-specific). A multi-specific antigen-binding fragment of an antibody will typically comprise at least two different variable domains, wherein each variable domain is capable of specifically binding to a separate antigen or to a different epitope on the same antigen. Any multi-specific antibody format, including the exemplary bi-specific antibody formats disclosed herein, may be adapted for use in the context of an antigen-binding fragment of an antibody of the present invention using routine techniques available in the art.

Preparation of Human Antibodies

[0139] Methods for generating human antibodies in transgenic mice are known in the art. Any such known methods can be used in the context of the present invention to make human antibodies that specifically bind to Influenza-HA. An immunogen comprising any one of the following can be used to generate antibodies to Influenza HA. In certain embodiments, the antibodies of the invention are obtained from mice immunized with a full length, native influenza HA (See, for example, GenBank accession number FJ966082.1), or with a live attenuated or inactivated virus, or with DNA encoding the protein or fragment thereof. Alternatively, the influenza-HA protein or a fragment thereof may be produced using standard biochemical techniques and modified and used as immunogen. In one embodiment, the immunogen is a recombinantly produced influenza-HA protein or fragment thereof. In certain embodiments of the invention, the immunogen may be an influenza virus vaccine. In certain embodiments, one

or more booster injections may be administered. In certain embodiments, the booster injections may comprise one or more influenza virus strains, or hemagglutinins derived from these strains, eg., See Protein Sciences H1 A/New Caledonia/20/1999, H5 A/Indonesia/05/2005, H3 A/Victoria/361/2011, H7 A/Netherlands/219/2003, or H9 A/Hong Kong/1073/1988. In certain embodiments, the booster injections may contain a 1:1 mixture of the influenza strains, or a 1:1 mixture of the hemagglutinins derived from the strains. In certain embodiments, the immunogen may be a recombinant Influenza HA peptide expressed in *E. coli* or in any other eukaryotic or mammalian cells such as Chinese hamster ovary (CHO) cells or influenza virus itself.

[0140] Using VELOCIMMUNE® technology (see, for example, US 6,596,541, Regeneron Pharmaceuticals, VELOCIMMUNE®) or any other known method for generating monoclonal antibodies, high affinity chimeric antibodies to influenza-HA are initially isolated having a human variable region and a mouse constant region. The VELOCIMMUNE® technology involves generation of a transgenic mouse having a genome comprising human heavy and light chain variable regions operably linked to endogenous mouse constant region loci such that the mouse produces an antibody comprising a human variable region and a mouse constant region in response to antigenic stimulation. The DNA encoding the variable regions of the heavy and light chains of the antibody are isolated and operably linked to DNA encoding the human heavy and light chain constant regions. The DNA is then expressed in a cell capable of expressing the fully human antibody.

[0141] Generally, a VELOCIMMUNE® mouse is challenged with the antigen of interest, and lymphatic cells (such as B-cells) are recovered from the mice that express antibodies. The lymphatic cells may be fused with a myeloma cell line to prepare immortal hybridoma cell lines, and such hybridoma cell lines are screened and selected to identify hybridoma cell lines that produce antibodies specific to the antigen of interest. DNA encoding the variable regions of the heavy chain and light chain may be isolated and linked to desirable isotypic constant regions of the heavy chain and light chain. Such an antibody protein may be produced in a cell, such as a CHO cell. Alternatively, DNA encoding the antigen-specific chimeric antibodies or the variable domains of the light and heavy chains may be isolated directly from antigen-specific lymphocytes.

[0142] Initially, high affinity chimeric antibodies are isolated having a human variable region and a mouse constant region. As in the experimental section below, the antibodies are characterized and selected for desirable characteristics, including affinity, selectivity, epitope, etc. The mouse constant regions are replaced with a desired human constant region to generate the fully human antibody of the invention, for example wild-type or modified IgG1 or IgG4. While the constant region selected may vary according to specific use, high affinity antigen-binding and target specificity characteristics reside in the variable region.

Bioequivalents

[0143] The anti-influenza-HA antibodies and antibody fragments of the present invention encompass proteins having amino acid sequences that vary from those of the described antibodies, but that retain the ability to bind Influenza HA. Such variant antibodies and antibody fragments comprise one or more additions, deletions, or substitutions of amino acids when compared to parent sequence, but exhibit biological activity that is essentially equivalent to that of the described antibodies. Likewise, the antibody-encoding DNA sequences of the present invention encompass sequences that comprise one or more additions, deletions, or substitutions of nucleotides when compared to the disclosed sequence, but that encode an antibody or antibody fragment that is essentially bioequivalent to an antibody or antibody fragment of the invention.

[0144] Two antigen-binding proteins, or antibodies, are considered bioequivalent if, for example, they are pharmaceutical equivalents or pharmaceutical alternatives whose rate and extent of absorption do not show a significant difference when administered at the same molar dose under similar experimental conditions, either single dose or multiple doses. Some antibodies will be considered equivalents or pharmaceutical alternatives if they are equivalent in the extent of their absorption but not in their rate of absorption and yet may be considered bioequivalent because such differences in the rate of absorption are intentional and are reflected in the labeling, are not essential to the attainment of effective body drug concentrations on, *e.g.*, chronic use, and are considered medically insignificant for the particular drug product studied.

[0145] In one embodiment, two antigen-binding proteins are bioequivalent if there are no clinically meaningful differences in their safety, purity, or potency.

[0146] In one embodiment, two antigen-binding proteins are bioequivalent if a patient can be switched one or more times between the reference product and the biological product without an expected increase in the risk of adverse effects, including a clinically significant change in immunogenicity, or diminished effectiveness, as compared to continued therapy without such switching.

[0147] In one embodiment, two antigen-binding proteins are bioequivalent if they both act by a common mechanism or mechanisms of action for the condition or conditions of use, to the extent that such mechanisms are known.

[0148] Bioequivalence may be demonstrated by *in vivo* and/or *in vitro* methods.

Bioequivalence measures include, *e.g.*, (a) an *in vivo* test in humans or other mammals, in which the concentration of the antibody or its metabolites is measured in blood, plasma, serum, or other biological fluid as a function of time; (b) an *in vitro* test that has been correlated with and is reasonably predictive of human *in vivo* bioavailability data; (c) an *in vivo* test in humans or other mammals in which the appropriate acute pharmacological effect of the antibody (or its target) is measured as a function of time; and (d) in a well-controlled clinical trial that

establishes safety, efficacy, or bioavailability or bioequivalence of an antibody.

[0149] Bioequivalent variants of the antibodies of the invention may be constructed by, for example, making various substitutions of residues or sequences or deleting terminal or internal residues or sequences not needed for biological activity. For example, cysteine residues not essential for biological activity can be deleted or replaced with other amino acids to prevent formation of unnecessary or incorrect intramolecular disulfide bridges upon renaturation. In other contexts, bioequivalent antibodies may include antibody variants comprising amino acid changes, which modify the glycosylation characteristics of the antibodies, *e.g.*, mutations that eliminate or remove glycosylation.

Anti-influenza-HA Antibodies Comprising Fc Variants

[0150] According to certain embodiments of the present invention, anti-influenza-HA antibodies are provided comprising an Fc domain comprising one or more mutations, which enhance or diminish antibody binding to the FcRn receptor, *e.g.*, at acidic pH as compared to neutral pH. For example, the present invention includes anti-influenza-HA antibodies comprising a mutation in the C_H2 or a C_H3 region of the Fc domain, wherein the mutation(s) increases the affinity of the Fc domain to FcRn in an acidic environment (*e.g.*, in an endosome where pH ranges from about 5.5 to about 6.0). Such mutations may result in an increase in serum half-life of the antibody when administered to an animal. Non-limiting examples of such Fc modifications include, *e.g.*, a modification at position 250 (*e.g.*, E or Q); 250 and 428 (*e.g.*, L or F); 252 (*e.g.*, L/Y/F/W or T), 254 (*e.g.*, S or T), and 256 (*e.g.*, S/R/Q/E/D or T); or a modification at position 428 and/or 433 (*e.g.*, H/L/R/S/P/Q or K) and/or 434 (*e.g.*, A, W, H, F or Y [N434A, N434W, N434H, N434F or N434Y]); or a modification at position 250 and/or 428; or a modification at position 307 or 308 (*e.g.*, 308F, V308F), and 434. In one embodiment, the modification comprises a 428L (*e.g.*, M428L) and 434S (*e.g.*, N434S) modification; a 428L, 259I (*e.g.*, V259I), and 308F (*e.g.*, V308F) modification; a 433K (*e.g.*, H433K) and a 434 (*e.g.*, 434Y) modification; a 252, 254, and 256 (*e.g.*, 252Y, 254T, and 256E) modification; a 250Q and 428L modification (*e.g.*, T250Q and M428L); and a 307 and/or 308 modification (*e.g.*, 308F or 308P). In yet another embodiment, the modification comprises a 265A (*e.g.*, D265A) and/or a 297A (*e.g.*, N297A) modification.

[0151] For example, the present invention includes anti-influenza-HA antibodies comprising an Fc domain comprising one or more pairs or groups of mutations selected from the group consisting of: 250Q and 248L (*e.g.*, T250Q and M248L); 252Y, 254T and 256E (*e.g.*, M252Y, S254T and T256E); 428L and 434S (*e.g.*, M428L and N434S); 257I and 311I (*e.g.*, P257I and Q311I); 257I and 434H (*e.g.*, P257I and N434H); 376V and 434H (*e.g.*, D376V and N434H); 307A, 380A and 434A (*e.g.*, T307A, E380A and N434A); and 433K and 434F (*e.g.*, H433K and N434F). All possible combinations of the foregoing Fc domain mutations and other mutations within the antibody variable domains disclosed herein, are contemplated within the scope of the present invention.

[0152] The present invention also includes anti-influenza-HA antibodies comprising a chimeric heavy chain constant (C_H) region, wherein the chimeric C_H region comprises segments derived from the C_H regions of more than one immunoglobulin isotype. For example, the antibodies of the invention may comprise a chimeric C_H region comprising part or all of a C_H2 domain derived from a human IgG1, human IgG2 or human IgG4 molecule, combined with part or all of a C_H3 domain derived from a human IgG1, human IgG2 or human IgG4 molecule. According to certain embodiments, the antibodies of the invention comprise a chimeric C_H region having a chimeric hinge region. For example, a chimeric hinge may comprise an "upper hinge" amino acid sequence (amino acid residues from positions 216 to 227 according to EU numbering) derived from a human IgG1, a human IgG2 or a human IgG4 hinge region, combined with a "lower hinge" sequence (amino acid residues from positions 228 to 236 according to EU numbering) derived from a human IgG1, a human IgG2 or a human IgG4 hinge region. According to certain embodiments, the chimeric hinge region comprises amino acid residues derived from a human IgG1 or a human IgG4 upper hinge and amino acid residues derived from a human IgG2 lower hinge. An antibody comprising a chimeric C_H region as described herein may, in certain embodiments, exhibit modified Fc effector functions without adversely affecting the therapeutic or pharmacokinetic properties of the antibody. (See, e.g., U.S. Provisional Appl. No. 61/759,578, filed February 1, 2013).

Biological Characteristics of the Antibodies

[0153] In general, the antibodies of the present invention function by binding to Influenza HA. For example, the present invention includes antibodies and antigen-binding fragments of antibodies that bind Influenza HA (e.g., at 25°C or at 37°C) with a K_D of less than 10 nM, as measured by real-time bio-layer interferometer based biosensor (Octet HTX assay), or by surface plasmon resonance. In certain embodiments, the antibodies or antigen-binding fragments thereof bind influenza-HA with a K_D of less than about 5nM, less than about 2nM, less than about 1nM, less than about 500pM, less than 250pM, or less than 100pM, as measured by surface plasmon resonance, e.g., using the assay format as described herein, or a substantially similar assay.

[0154] The present invention also includes antibodies and antigen-binding fragments thereof that bind Influenza HA with a dissociative half-life ($t_{1/2}$) of greater than about 100 minutes as measured by surface plasmon resonance at 25°C, e.g., using an assay format as defined herein, or a substantially similar assay. In certain embodiments, the antibodies or antigen-binding fragments of the present invention bind Influenza HA with a $t_{1/2}$ of greater than about 200 minutes, greater than about 300 minutes, greater than about 400 minutes, greater than about 500 minutes, greater than about 600 minutes, greater than about 700 minutes, greater than about 800 minutes, greater than about 900 minutes, or greater than about 1000 minutes as measured by surface plasmon resonance at 25°C, e.g., using an assay format as defined

herein (e.g., mAb-capture or antigen-capture format), or a substantially similar assay. In one embodiment, the antibodies and antigen-binding fragments of the invention bind Influenza HA with a dissociative half-life ($t_{1/2}$) of greater than 300 minutes. In one embodiment, an antibody of the invention provides for about a 1.5 to 2-fold increase in dissociative half life as compared to a comparator antibody designated Control I mAb, when tested in monkeys and mice.

[0155] The present invention also includes antibodies or antigen-binding fragments thereof that neutralize the infectivity of influenza virus for its host cells. In some embodiments, the antibodies exhibit a neutralization potency against various representative group 1 influenza viruses (H1N1 A/Puerto Rico/08/1934; H5N1 A/Vietnam/1203/2004; H1N1 A/California/07/2009; H1N1 A/Wisconsin/1933; H1N1 A/Brisbane/59/1997, H9N2 A/Hong Kong/33982/2009, H13N6 a/gull/Maryland/704/1977 and H16N3 A/shorebird/Delaware/172/2006 with an IC_{50} ranging from about 1.6 nM to about 130 nM in a microneutralization assay, e.g., as shown in Examples 4 and 5, or a substantially similar assay. In one embodiment, the antibodies or antigen-binding fragments thereof that neutralize the infectivity of influenza virus for its host cells do so with an IC_{50} of less than 130 nM.

[0156] The present invention also includes antibodies or antigen-binding fragments thereof that mediate complement-dependent cytotoxicity of infected cells, with an EC_{50} ranging from about 20nM to about 66nM (see example 6). In one embodiment, the antibodies or antigen-binding fragments thereof mediate complement-dependent cytotoxicity of infected cells, with an EC_{50} less than 66 nM.

[0157] The present invention also includes anti-influenza-A HA antibodies that demonstrate an increase in protection, or potent neutralization of influenza A infection *in vivo*. Certain antibodies show potent neutralization when administered either prophylactically (prior to infection) or therapeutically (after infection; see example 7). In certain embodiments, some of the antibodies (H1H11729P and H1H11829N2) demonstrated 100% survival of mice when administered prophylactically as a single dose of 1 mg/kg. Certain antibodies demonstrated significant survival of mice when administered prophylactically at doses as low as 0.5 mg/kg (100% survival using the antibody designated as H1H11729P), at 0.1 mg/kg of H1H11729P (40% survival), or at 0.05 mg/kg of H1H11829N2 (20% survival). Significant survival was also observed when certain exemplary antibodies (H1H11829N2 and H1H11729P) were administered after infection at doses of 15 or 30 mg/kg. In one embodiment, an antibody of the invention demonstrates an additive protective effect in influenza-infected mammals when combined with an anti-viral drug, oseltamivir.

[0158] In one embodiment, the invention provides an isolated recombinant antibody or antigen-binding fragment thereof that binds specifically to Influenza HA, wherein the antibody or fragment thereof exhibits two or more of the following characteristics: (a) is a fully human monoclonal antibody; (b) binds to influenza HA with a dissociation constant (K_D) of less than 10^{-9} M, as measured in a surface plasmon resonance assay; (c) demonstrates a dissociative half-

life ($t_{1/2}$) ranging from about 370 minutes to greater than 1000 minutes; (d) demonstrates neutralization of group 1 influenza A viruses selected from H1N1, H5N1, H9N2, H13N6 and H16N3, with an IC_{50} ranging from about 1.6nM to about 130nM; (e) demonstrates complement mediated lysis of influenza virus infected cells with an EC_{50} of about 20nM to about 66nM; or (f) demonstrates protection, as measured by increased survival in an animal model of influenza virus infection when administered either before or after virus challenge.

[0159] The antibodies of the present invention may possess two or more of the aforementioned biological characteristics, or any combinations thereof. Other biological characteristics of the antibodies of the present invention will be evident to a person of ordinary skill in the art from a review of the present disclosure including the working Examples herein.

Epitope Mapping and Related Technologies

[0160] The present invention includes anti-influenza-HA virus antibodies, which interact with one or more amino acids found within one or more domains of the influenza HA molecule. The epitope to which the antibodies bind may consist of a single contiguous sequence of 3 or more (*e.g.*, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20 or more) amino acids located within the influenza HA molecule (*e.g.* a linear epitope in a domain). Alternatively, the epitope may consist of a plurality of non-contiguous amino acids (or amino acid sequences) located within the influenza-HA molecule (*e.g.* a conformational epitope).

[0161] Various techniques known to persons of ordinary skill in the art can be used to determine whether an antibody "interacts with one or more amino acids" within a polypeptide or protein. Exemplary techniques include, for example, routine cross-blocking assays, such as that described in Antibodies, Harlow and Lane (Cold Spring Harbor Press, Cold Spring Harbor, NY). Other methods include alanine scanning mutational analysis, peptide blot analysis (Reineke (2004) Methods Mol. Biol. 248: 443-63), peptide cleavage analysis crystallographic studies and NMR analysis. In addition, methods such as epitope excision, epitope extraction and chemical modification of antigens can be employed (Tomer (2000) Prot. Sci. 9: 487-496). Another method that can be used to identify the amino acids within a polypeptide with which an antibody interacts is hydrogen/deuterium exchange detected by mass spectrometry. In general terms, the hydrogen/deuterium exchange method involves deuterium-labeling the protein of interest, followed by binding the antibody to the deuterium-labeled protein. Next, the protein/antibody complex is transferred to water and exchangeable protons within amino acids that are protected by the antibody complex undergo deuterium-to-hydrogen back-exchange at a slower rate than exchangeable protons within amino acids that are not part of the interface. As a result, amino acids that form part of the protein/antibody interface may retain deuterium and therefore exhibit relatively higher mass compared to amino acids not included in the interface. After dissociation of the antibody, the target protein is subjected to protease cleavage and mass spectrometry analysis, thereby revealing the deuterium-labeled residues which correspond to

the specific amino acids with which the antibody interacts. See, e.g., Ehring (1999) *Analytical Biochemistry* 267: 252-259; Engen and Smith (2001) *Anal. Chem.* 73: 256A-265A.

[0162] The term "epitope" refers to a site on an antigen to which B and/or T cells respond. B-cell epitopes can be formed both from contiguous amino acids or noncontiguous amino acids juxtaposed by tertiary folding of a protein. Epitopes formed from contiguous amino acids are typically retained on exposure to denaturing solvents, whereas epitopes formed by tertiary folding are typically lost on treatment with denaturing solvents. An epitope typically includes at least 3, and more usually, at least 5 or 8-10 amino acids in a unique spatial conformation.

[0163] Modification-Assisted Profiling (MAP), also known as Antigen Structure-based Antibody Profiling (ASAP) is a method that categorizes large numbers of monoclonal antibodies (mAbs) directed against the same antigen according to the similarities of the binding profile of each antibody to chemically or enzymatically modified antigen surfaces (see US 2004/0101920). Each category may reflect a unique epitope either distinctly different from or partially overlapping with epitope represented by another category. This technology allows rapid filtering of genetically identical antibodies, such that characterization can be focused on genetically distinct antibodies. When applied to hybridoma screening, MAP may facilitate identification of rare hybridoma clones that produce mAbs having the desired characteristics. MAP may be used to sort the antibodies of the invention into groups of antibodies binding different epitopes.

[0164] In certain embodiments, the influenza virus-HA antibodies or antigen-binding fragments thereof bind an epitope within any one or more of the regions exemplified in influenza HA, either in natural form, or recombinantly produced, or to a fragment thereof.

[0165] The present invention includes anti-influenza-HA antibodies that bind to the same epitope, or a portion of the epitope. Likewise, the present invention also includes anti-influenza-HA antibodies that compete for binding to Influenza HA or a fragment thereof with any of the specific exemplary antibodies described herein. For example, the present invention includes anti-influenza-HA antibodies that cross-compete for binding to influenza HA with one or more antibodies obtained from those antibodies described in Tables 1 and 12.

[0166] One can easily determine whether an antibody binds to the same epitope as, or competes for binding with, a reference anti-influenza-HA antibody by using routine methods known in the art. For example, to determine if a test antibody binds to the same epitope as a reference anti-influenza-HA antibody of the invention, the reference antibody is allowed to bind to an Influenza HA or peptide under saturating conditions. Next, the ability of a test antibody to bind to the Influenza HA molecule is assessed. If the test antibody is able to bind to influenza virus HA following saturation binding with the reference anti-influenza-HA antibody, it can be concluded that the test antibody binds to a different epitope than the reference anti-influenza-HA antibody. On the other hand, if the test antibody is not able to bind to the Influenza HA following saturation binding with the reference anti-influenza-HA antibody, then the test antibody may bind to the same epitope as the epitope bound by the reference anti-influenza-HA antibody.

of the invention.

[0167] To determine if an antibody competes for binding with a reference anti-influenza-HA antibody, the above-described binding methodology is performed in two orientations: In a first orientation, the reference antibody is allowed to bind to a Influenza HA under saturating conditions followed by assessment of binding of the test antibody to the influenza-HA molecule. In a second orientation, the test antibody is allowed to bind to an influenza-HA molecule under saturating conditions followed by assessment of binding of the reference antibody to the influenza-HA molecule. If, in both orientations, only the first (saturating) antibody is capable of binding to the influenza-HA molecule, then it is concluded that the test antibody and the reference antibody compete for binding to influenza-HA. As will be appreciated by a person of ordinary skill in the art, an antibody that competes for binding with a reference antibody may not necessarily bind to the identical epitope as the reference antibody, but may sterically block binding of the reference antibody by binding an overlapping or adjacent epitope.

[0168] Two antibodies bind to the same or overlapping epitope if each competitively inhibits (blocks) binding of the other to the antigen. That is, a 1-, 5-, 10-, 20- or 100-fold excess of one antibody inhibits binding of the other by at least 50% but preferably 75%, 90% or even 99% as measured in a competitive binding assay (see, *e.g.*, Junghans *et al.*, Cancer Res. 1990 50:1495-1502). Alternatively, two antibodies have the same epitope if essentially all amino acid mutations in the antigen that reduce or eliminate binding of one antibody reduce or eliminate binding of the other. Two antibodies have overlapping epitopes if some amino acid mutations that reduce or eliminate binding of one antibody reduce or eliminate binding of the other.

[0169] Additional routine experimentation (*e.g.*, peptide mutation and binding analyses) can then be carried out to confirm whether the observed lack of binding of the test antibody is in fact due to binding to the same epitope as the reference antibody or if steric blocking (or another phenomenon) is responsible for the lack of observed binding. Experiments of this sort can be performed using ELISA, RIA, surface plasmon resonance, flow cytometry or any other quantitative or qualitative antibody-binding assay available in the art.

Immunoconjugates

[0170] The invention encompasses a human anti-influenza-HA monoclonal antibody conjugated to a therapeutic moiety ("immunoconjugate"), such as a toxoid or an anti-viral drug to treat influenza virus infection. As used herein, the term "immunoconjugate" refers to an antibody, which is chemically or biologically linked to a radioactive agent, a cytokine, an interferon, a target or reporter moiety, an enzyme, a peptide or protein or a therapeutic agent. The antibody may be linked to the radioactive agent, cytokine, interferon, target or reporter moiety, enzyme, peptide or therapeutic agent at any location along the molecule so long as it is able to bind its target. Examples of immunoconjugates include antibody drug conjugates and

antibody-toxin fusion proteins. In one embodiment, the agent may be a second different antibody to Influenza-HA. In certain embodiments, the antibody may be conjugated to an agent specific for a virally infected cell. The type of therapeutic moiety that may be conjugated to the anti- influenza-HA antibody and will take into account the condition to be treated and the desired therapeutic effect to be achieved. Examples of suitable agents for forming immunoconjugates are known in the art; see for example, WO 05/103081.

Multi-specific Antibodies

[0171] The antibodies of the present invention may be mono-specific, bi-specific, or multi-specific. Multi-specific antibodies may be specific for different epitopes of one target polypeptide or may contain antigen-binding domains specific for more than one target polypeptide. See, *e.g.*, Tutt *et al.*, 1991, J. Immunol. 147:60-69; Kufer *et al.*, 2004, Trends Biotechnol. 22:238-244.

[0172] Any of the multi-specific antigen-binding molecules of the invention, or variants thereof, may be constructed using standard molecular biological techniques (*e.g.*, recombinant DNA and protein expression technology), as will be known to a person of ordinary skill in the art.

[0173] In some embodiments, influenza-HA-specific antibodies are generated in a bi-specific format (a "bi-specific") in which variable regions binding to distinct domains of Influenza HA are linked together to confer dual-domain specificity within a single binding molecule. Appropriately designed bi-specifics may enhance overall influenza-HA-protein inhibitory efficacy through increasing both specificity and binding avidity. Variable regions with specificity for individual domains, (*e.g.*, segments of the N-terminal domain), or that can bind to different regions within one domain, are paired on a structural scaffold that allows each region to bind simultaneously to the separate epitopes, or to different regions within one domain. In one example for a bi-specific, heavy chain variable regions (V_H) from a binder with specificity for one domain are recombined with light chain variable regions (V_L) from a series of binders with specificity for a second domain to identify non-cognate V_L partners that can be paired with an original V_H without disrupting the original specificity for that V_H . In this way, a single V_L segment (*e.g.*, V_{L1}) can be combined with two different V_H domains (*e.g.*, V_{H1} and V_{H2}) to generate a bi-specific comprised of two binding "arms" (V_{H1} - V_{L1} and V_{H2} - V_{L1}). Use of a single V_L segment reduces the complexity of the system and thereby simplifies and increases efficiency in cloning, expression, and purification processes used to generate the bi-specific (See, for example, USSN13/022759 and US2010/0331527).

[0174] Alternatively, antibodies that bind more than one domain and a second target, such as, but not limited to, for example, a second different anti-influenza-HA antibody, may be prepared in a bi-specific format using techniques described herein, or other techniques known to those skilled in the art. Antibody variable regions binding to distinct regions may be linked together with variable regions that bind to relevant sites on, for example, the influenza virus, to confer

dual-antigen specificity within a single binding molecule. Appropriately designed bi-specifics of this nature serve a dual function. Variable regions with specificity for the extracellular domain are combined with a variable region with specificity for outside the extracellular domain and are paired on a structural scaffold that allows each variable region to bind to the separate antigens.

[0175] An exemplary bi-specific antibody format that can be used in the context of the present invention involves the use of a first immunoglobulin (Ig) C_H3 domain and a second Ig C_H3 domain, wherein the first and second Ig C_H3 domains differ from one another by at least one amino acid, and wherein at least one amino acid difference reduces binding of the bi-specific antibody to Protein A as compared to a bi-specific antibody lacking the amino acid difference. In one embodiment, the first Ig C_H3 domain binds Protein A and the second Ig C_H3 domain contains a mutation that reduces or abolishes Protein A binding such as an H95R modification (by IMGT exon numbering; H435R by EU numbering). The second C_H3 may further comprise a Y96F modification (by IMGT; Y436F by EU). Further modifications that may be found within the second C_H3 include: D16E, L18M, N44S, K52N, V57M, and V82I (by IMGT; D356E, L358M, N384S, K392N, V397M, and V422I by EU) in the case of IgG1 antibodies; N44S, K52N, and V82I (IMGT; N384S, K392N, and V422I by EU) in the case of IgG2 antibodies; and Q15R, N44S, K52N, V57M, R69K, E79Q, and V82I (by IMGT; Q355R, N384S, K392N, V397M, R409K, E419Q, and V422I by EU) in the case of IgG4 antibodies. Variations on the bi-specific antibody format described above are contemplated within the scope of the present invention.

[0176] Other exemplary bispecific formats that can be used in the context of the present invention include, without limitation, *e.g.*, scFv-based or diabody bispecific formats, IgG-scFv fusions, dual variable domain (DVD)-Ig, Quadroma, knobs-into-holes, common light chain (*e.g.*, common light chain with knobs-into-holes, etc.), CrossMab, CrossFab, (SEED)body, leucine zipper, Duobody, IgG1/IgG2, dual acting Fab (DAF)-IgG, and Mab² bispecific formats (*see, e.g.*, Klein *et al.* 2012, mAbs 4:6, 1-11, and references cited therein, for a review of the foregoing formats). Bispecific antibodies can also be constructed using peptide/nucleic acid conjugation, *e.g.*, wherein unnatural amino acids with orthogonal chemical reactivity are used to generate site-specific antibody-oligonucleotide conjugates which then self-assemble into multimeric complexes with defined composition, valency and geometry. (*See, e.g.*, Kazane *et al.*, *J. Am. Chem. Soc.* [Epub: Dec. 4, 2012]).

Therapeutic Administration and Formulations

[0177] The invention provides therapeutic compositions comprising the anti-influenza-HA antibodies or antigen-binding fragments thereof of the present invention. Therapeutic compositions in accordance with the invention will be administered with suitable carriers, excipients, and other agents that are incorporated into formulations to provide improved transfer, delivery, tolerance, and the like. A multitude of appropriate formulations can be found in the formulary known to all pharmaceutical chemists: Remington's Pharmaceutical Sciences,

Mack Publishing Company, Easton, PA. These formulations include, for example, powders, pastes, ointments, jellies, waxes, oils, lipids, lipid (cationic or anionic) containing vesicles (such as LIPOFECTIN™), DNA conjugates, anhydrous absorption pastes, oil-in-water and water-in-oil emulsions, emulsions carbowax (polyethylene glycols of various molecular weights), semi-solid gels, and semi-solid mixtures containing carbowax. See also Powell *et al.* "Compendium of excipients for parenteral formulations" PDA (1998) J Pharm Sci Technol 52:238-311.

[0178] The dose of antibody may vary depending upon the age and the size of a subject to be administered, target disease, conditions, route of administration, and the like. When an antibody of the present invention is used for treating a disease or disorder in an adult patient, or for preventing such a disease, it is advantageous to administer the antibody of the present invention normally at a single dose of about 0.1 to about 60 mg/kg body weight, more preferably about 5 to about 60, about 10 to about 50, or about 20 to about 50 mg/kg body weight. Depending on the severity of the condition, the frequency and the duration of the treatment can be adjusted. In certain embodiments, the antibody or antigen-binding fragment thereof of the invention can be administered as an initial dose of at least about 0.1 mg to about 5000 mg, about 1 to about 2000 mg, about 5 to about 1000 mg, or about 10 to about 500 mg, to about 100 mg, or to about 50 mg. In certain embodiments, the initial dose may be followed by administration of a second or a plurality of subsequent doses of the antibody or antigen-binding fragment thereof in an amount that can be approximately the same or less than that of the initial dose, wherein the subsequent doses are separated by at least 1 day to 3 days; at least one week, at least 2 weeks; at least 3 weeks; at least 4 weeks; at least 5 weeks; at least 6 weeks; at least 7 weeks; at least 8 weeks; at least 9 weeks; at least 10 weeks; at least 12 weeks; or at least 14 weeks.

[0179] Various delivery systems are known and can be used to administer the pharmaceutical composition of the invention, *e.g.*, encapsulation in liposomes, microparticles, microcapsules, recombinant cells capable of expressing the mutant viruses, receptor mediated endocytosis (see, *e.g.*, Wu et al. (1987) J. Biol. Chem. 262:4429-4432). Methods of introduction include, but are not limited to, intradermal, transdermal, intramuscular, intraperitoneal, intravenous, subcutaneous, intranasal, epidural and oral routes. The composition may be administered by any convenient route, for example by infusion or bolus injection, by absorption through epithelial or mucocutaneous linings (*e.g.*, oral mucosa, rectal and intestinal mucosa, etc.) and may be administered together with other biologically active agents. Administration can be systemic or local. The pharmaceutical composition can be also delivered in a vesicle, in particular a liposome (see, for example, Langer (1990) Science 249:1527-1533).

[0180] The use of nanoparticles to deliver the antibodies of the present invention is also contemplated herein. Antibody-conjugated nanoparticles may be used both for therapeutic and diagnostic applications. Antibody-conjugated nanoparticles and methods of preparation and use are described in detail by Arruebo, M., et al. 2009 ("Antibody-conjugated nanoparticles for

biomedical applications" in J. Nanomat. Volume 2009, Article ID 439389, 24 pages, doi: 10.1155/2009/439389), incorporated herein by reference. Nanoparticles may be developed and conjugated to antibodies contained in pharmaceutical compositions to target virally infected cells. Nanoparticles for drug delivery have also been described in, for example, US 8257740, or US 8246995, each incorporated herein in its entirety.

[0181] In certain situations, the pharmaceutical composition can be delivered in a controlled release system. In one embodiment, a pump may be used. In another embodiment, polymeric materials can be used. In yet another embodiment, a controlled release system can be placed in proximity of the composition's target, thus requiring only a fraction of the systemic dose.

[0182] The injectable preparations may include dosage forms for intravenous, subcutaneous, intracutaneous, intracranial, intraperitoneal and intramuscular injections, drip infusions, etc. These injectable preparations may be prepared by methods publicly known. For example, the injectable preparations may be prepared, *e.g.*, by dissolving, suspending or emulsifying the antibody or its salt described above in a sterile aqueous medium or an oily medium conventionally used for injections. As the aqueous medium for injections, there are, for example, physiological saline, an isotonic solution containing glucose and other auxiliary agents, etc., which may be used in combination with an appropriate solubilizing agent such as an alcohol (*e.g.*, ethanol), a polyalcohol (*e.g.*, propylene glycol, polyethylene glycol), a nonionic surfactant [*e.g.*, polysorbate 80, HCO-50 (polyoxyethylene (50 mol) adduct of hydrogenated castor oil)], etc. As the oily medium, there are employed, *e.g.*, sesame oil, soybean oil, etc., which may be used in combination with a solubilizing agent such as benzyl benzoate, benzyl alcohol, etc. The injection thus prepared is preferably filled in an appropriate ampoule.

[0183] A pharmaceutical composition of the present invention can be delivered subcutaneously or intravenously with a standard needle and syringe. In addition, with respect to subcutaneous delivery, a pen delivery device readily has applications in delivering a pharmaceutical composition of the present invention. Such a pen delivery device can be reusable or disposable. A reusable pen delivery device generally utilizes a replaceable cartridge that contains a pharmaceutical composition. Once all of the pharmaceutical composition within the cartridge has been administered and the cartridge is empty, the empty cartridge can readily be discarded and replaced with a new cartridge that contains the pharmaceutical composition. The pen delivery device can then be reused. In a disposable pen delivery device, there is no replaceable cartridge. Rather, the disposable pen delivery device comes prefilled with the pharmaceutical composition held in a reservoir within the device. Once the reservoir is emptied of the pharmaceutical composition, the entire device is discarded.

[0184] Numerous reusable pen and autoinjector delivery devices have applications in the subcutaneous delivery of a pharmaceutical composition of the present invention. Examples include, but certainly are not limited to AUTOPEN™ (Owen Mumford, Inc., Woodstock, UK), DISETRONIC™ pen (Disetronic Medical Systems, Burghdorf, Switzerland), HUMALOG MIX

75/25™ pen, HUMALOG™ pen, HUMALIN 70/30™ pen (Eli Lilly and Co., Indianapolis, IN), NOVOPEN™ I, II and III (Novo Nordisk, Copenhagen, Denmark), NOVOPEN JUNIOR™ (Novo Nordisk, Copenhagen, Denmark), BD™ pen (Becton Dickinson, Franklin Lakes, NJ), OPTIPEN™, OPTIPEN PRO™, OPTIPEN STARLET™, and OPTICLIK™ (Sanofi-Aventis, Frankfurt, Germany), to name only a few. Examples of disposable pen delivery devices having applications in subcutaneous delivery of a pharmaceutical composition of the present invention include, but certainly are not limited to the SOLOSTAR™ pen (Sanofi-Aventis), the FLEXPEN™ (Novo Nordisk), and the KWIKPEN™ (Eli Lilly), the SURECLICK™ Autoinjector (Amgen, Thousand Oaks, CA), the PENLET™ (Haselmeier, Stuttgart, Germany), the EPIPEN (Dey, L.P.) and the HUMIRA™ Pen (Abbott Labs, Abbott Park, IL), to name only a few.

[0185] Advantageously, the pharmaceutical compositions for oral or parenteral use described above are prepared into dosage forms in a unit dose suited to fit a dose of the active ingredients. Such dosage forms in a unit dose include, for example, tablets, pills, capsules, injections (ampoules), suppositories, etc. The amount of the antibody contained is generally about 5 to about 5000 mg per dosage form in a unit dose; especially in the form of injection, it is preferred that the antibody is contained in about 5 to about 500 mg and in about 10 to about 250 mg for the other dosage forms.

Therapeutic Uses of the Antibodies

[0186] The antibodies of the present invention are useful for the treatment, and/or prevention of a disease or disorder or condition associated with influenza virus infection and/or for ameliorating at least one symptom associated with such disease, disorder or condition.

[0187] In certain embodiments, the antibodies of the invention are useful to treat subjects suffering from the severe and acute respiratory infection caused by influenza virus. In some embodiments, the antibodies of the invention are useful in decreasing viral titers or reducing viral load in the host. In one embodiment, an antibody or antigen-binding fragment thereof of the invention may be administered at a therapeutic dose to a patient with influenza virus infection.

[0188] One or more antibodies of the present invention may be administered to relieve or prevent or decrease the severity of one or more of the symptoms or conditions of the disease or disorder. The antibodies may be used to ameliorate or reduce the severity of at least one symptom of influenza virus infection including, but not limited to fever, cough, sore throat, headache, body aches, fatigue, extreme exhaustion, shortness of breath, bronchitis, pneumonia, and death.

[0189] It is also contemplated herein to use one or more antibodies of the present invention prophylactically to subjects at risk for developing an influenza virus infection such as immunocompromised individuals, elderly adults (more than 65 years of age), children younger than 2 years of age, healthcare workers, family members in close proximity to a patient suffering from an influenza virus infection, and patients with a medical history (e.g., increased

risk of pulmonary infection, heart disease or diabetes).

[0190] In a further embodiment of the invention the present antibodies are used for the preparation of a pharmaceutical composition for treating patients suffering from an influenza virus infection. In another embodiment of the invention, the present antibodies are used as adjunct therapy with any other agent or any other therapy known to those skilled in the art useful for treating or ameliorating an influenza virus infection.

Combination Therapies

[0191] Combination therapies may include an anti-influenza-HA antibody of the invention and any additional therapeutic agent that may be advantageously combined with an antibody of the invention, or with a biologically active fragment of an antibody of the invention. The antibodies of the present invention may be combined synergistically with one or more drugs or agents (*e.g.* anti-viral agents) used to treat influenza virus.

[0192] For example, exemplary anti-viral agents include, *e.g.*, vaccines, neuraminidase inhibitors or nucleoside analogs. Other exemplary anti-viral agents that may be used in combination with an antibody of the invention can include, *e.g.*, zidovudine, gangcyclovir, vidarabine, idoxuridine, trifluridine, foscarnet, acyclovir, ribavirin, amantadine, remantidine, saquinavir, indinavir, ritonavir, alpha-interferons and other interferons, a neuraminidase inhibitor (*e.g.*, zanamivir (RELENZA®), oseltamivir (TAMIFLU®) laninamivir, peramivir), or rimantadine.

[0193] Other exemplary anti-viral drugs include, but are not limited to, a HA inhibitor, a sialic acid inhibitor and an M2 ion channel inhibitor. In one embodiment, the M2 ion channel inhibitor is amantadine or rimantadine.

[0194] In some embodiments, the antibodies of the invention may be combined with a second therapeutic agent to reduce the viral load in a patient with an influenza virus infection, or to ameliorate one or more symptoms of the infection.

[0195] The antibodies of the present invention may be used in combination with an anti-inflammatory drug (*e.g.*, corticosteroids, and non-steroidal anti-inflammatory drugs), a decongestant, an anti-histamine, an anti-infective drug, a different antibody to Influenza virus, an anti-viral drug, a vaccine for influenza virus, such as FLUMIST® or FLUVIRIN®, a dietary supplement such as anti-oxidants or any other palliative therapy to treat an influenza virus infection.

[0196] In certain embodiments, the second therapeutic agent is another antibody to influenza. In certain embodiments, the second therapeutic agent is another antibody to influenza hemagglutinin. In certain embodiments, the second therapeutic agent is another antibody to a different influenza protein, such as the neuraminidase, or the tetrameric ectodomain of matrix protein 2 (M2e protein). In certain embodiments, the second therapeutic agent is an antibody to a different protein such as the host transmembrane protease, serine 2 (TMPRSS2). The second antibody may be specific for one or more different influenza virus proteins from different

subtypes or strains of the virus. It is contemplated herein to use a combination ("cocktail") of antibodies with broad neutralization or inhibitory activity against influenza virus. In some embodiments, non-competing antibodies may be combined and administered to a subject in need thereof, to reduce the ability of influenza virus to escape due to rapid mutation as a result of selection pressure. In some embodiments, the antibodies comprising the combination bind to distinct non-overlapping epitopes on the HA protein. The antibodies comprising the combination may block the virus attachment and/or entry into and/or fusion with host cells. The antibodies may interact with a hemagglutinin selected from any one or more of the Group 1 influenza A subtypes including H1, H2, H5, H6, H8, H9, H11, H12, H13, H16, H17 or H18 and when used alone, or in combination with any one or more of the agents noted above, may neutralize any one or more of the Group 1 influenza subtypes including, but not limited to the following: H1N1, H5N1, H9N2, H13N6 and H16N3.

[0197] It is also contemplated herein to use a combination of anti-influenza-HA antibodies of the present invention, wherein the combination comprises one or more antibodies that do not cross-compete; In some embodiments, the combination includes a first antibody with broad neutralization activity with a second antibody with activity against a narrow spectrum of isolates and that does not cross-compete with the first antibody.

[0198] As used herein, the term "in combination with" means that additional therapeutically active component(s) may be administered prior to, concurrent with, or after the administration of the anti-influenza-HA antibody of the present invention. The term "in combination with" also includes sequential or concomitant administration of an anti-influenza-HA antibody and a second therapeutic agent.

[0199] The additional therapeutically active component(s) may be administered to a subject prior to administration of an anti-influenza-HA antibody of the present invention. For example, a first component may be deemed to be administered "prior to" a second component if the first component is administered 1 week before, 72 hours before, 60 hours before, 48 hours before, 36 hours before, 24 hours before, 12 hours before, 6 hours before, 5 hours before, 4 hours before, 3 hours before, 2 hours before, 1 hour before, 30 minutes before, 15 minutes before, 10 minutes before, 5 minutes before, or less than 1 minute before administration of the second component. In other embodiments, the additional therapeutically active component(s) may be administered to a subject after administration of an anti-influenza-HA antibody of the present invention. For example, a first component may be deemed to be administered "after" a second component if the first component is administered 1 minute after, 5 minutes after, 10 minutes after, 15 minutes after, 30 minutes after, 1 hour after, 2 hours after, 3 hours after, 4 hours after, 5 hours after, 6 hours after, 12 hours after, 24 hours after, 36 hours after, 48 hours after, 60 hours after, 72 hours after administration of the second component. In yet other embodiments, the additional therapeutically active component(s) may be administered to a subject concurrent with administration of an anti-influenza-HA antibody of the present invention. "Concurrent"

administration, for purposes of the present invention, includes, *e.g.*, administration of an anti-influenza-HA antibody and an additional therapeutically active component to a subject in a single dosage form, or in separate dosage forms administered to the subject within about 30 minutes or less of each other. If administered in separate dosage forms, each dosage form may be administered via the same route (*e.g.*, both the anti-influenza-HA antibody and the additional therapeutically active component may be administered intravenously, etc.); alternatively, each dosage form may be administered via a different route (*e.g.*, the anti-influenza-HA antibody may be administered intravenously, and the additional therapeutically active component may be administered orally). In any event, administering the components in a single dosage form, in separate dosage forms by the same route, or in separate dosage forms by different routes are all considered "concurrent administration," for purposes of the present disclosure. For purposes of the present disclosure, administration of an anti-influenza-HA antibody "prior to", "concurrent with," or "after" (as those terms are defined herein above) administration of an additional therapeutically active component is considered administration of an anti-influenza-HA antibody "in combination with" an additional therapeutically active component.

[0200] The present invention includes pharmaceutical compositions in which an anti-influenza-HA antibody of the present invention is co-formulated with one or more of the additional therapeutically active component(s) as described elsewhere herein.

Administration Regimens

[0201] According to certain embodiments, a single dose of an anti-influenza-HA antibody of the invention (or a pharmaceutical composition comprising a combination of an anti-influenza-HA antibody and any of the additional therapeutically active agents mentioned herein) may be administered to a subject in need thereof. According to certain embodiments of the present invention, multiple doses of an anti-influenza-HA antibody (or a pharmaceutical composition comprising a combination of an anti-influenza-HA antibody and any of the additional therapeutically active agents mentioned herein) may be administered to a subject over a defined time course. The methods according to this aspect of the invention comprise sequentially administering to a subject multiple doses of an anti-influenza-HA antibody of the invention. As used herein, "sequentially administering" means that each dose of anti-influenza-HA antibody is administered to the subject at a different point in time, *e.g.*, on different days separated by a predetermined interval (*e.g.*, hours, days, weeks or months). The present invention includes methods which comprise sequentially administering to the patient a single initial dose of an anti-influenza-HA antibody, followed by one or more secondary doses of the anti-influenza-HA antibody, and optionally followed by one or more tertiary doses of the anti-influenza-HA antibody.

[0202] The terms "initial dose," "secondary doses," and "tertiary doses," refer to the temporal

sequence of administration of the anti-influenza-HA antibody of the invention. Thus, the "initial dose" is the dose which is administered at the beginning of the treatment regimen (also referred to as the "baseline dose"); the "secondary doses" are the doses which are administered after the initial dose; and the "tertiary doses" are the doses which are administered after the secondary doses. The initial, secondary, and tertiary doses may all contain the same amount of anti-influenza-HA antibody, but generally may differ from one another in terms of frequency of administration. In certain embodiments, however, the amount of anti-influenza-HA antibody contained in the initial, secondary and/or tertiary doses varies from one another (*e.g.*, adjusted up or down as appropriate) during the course of treatment. In certain embodiments, two or more (*e.g.*, 2, 3, 4, or 5) doses are administered at the beginning of the treatment regimen as "loading doses" followed by subsequent doses that are administered on a less frequent basis (*e.g.*, "maintenance doses").

[0203] In certain exemplary embodiments of the present invention, each secondary and/or tertiary dose is administered 1 to 48 hours (*e.g.*, 1, 1½, 2, 2½, 3, 3½, 4, 4½, 5, 5½, 6, 6½, 7, 7½, 8, 8½, 9, 9½, 10, 10½, 11, 11½, 12, 12½, 13, 13½, 14, 14½, 15, 15½, 16, 16½, 17, 17½, 18, 18½, 19, 19½, 20, 20½, 21, 21½, 22, 22½, 23, 23½, 24, 24½, 25, 25½, 26, 26½, or more) after the immediately preceding dose. The phrase "the immediately preceding dose," as used herein, means, in a sequence of multiple administrations, the dose of anti-influenza-HA antibody, which is administered to a patient prior to the administration of the very next dose in the sequence with no intervening doses.

[0204] The methods according to this aspect of the invention may comprise administering to a patient any number of secondary and/or tertiary doses of an anti-influenza-HA antibody. For example, in certain embodiments, only a single secondary dose is administered to the patient. In other embodiments, two or more (*e.g.*, 2, 3, 4, 5, 6, 7, 8, or more) secondary doses are administered to the patient. Likewise, in certain embodiments, only a single tertiary dose is administered to the patient. In other embodiments, two or more (*e.g.*, 2, 3, 4, 5, 6, 7, 8, or more) tertiary doses are administered to the patient.

[0205] In certain embodiments of the invention, the frequency at which the secondary and/or tertiary doses are administered to a patient can vary over the course of the treatment regimen. The frequency of administration may also be adjusted during the course of treatment by a physician depending on the needs of the individual patient following clinical examination.

Diagnostic Uses of the Antibodies

[0206] The anti-influenza-HA antibodies of the present invention may be used to detect and/or measure influenza-HA in a sample, *e.g.*, for diagnostic purposes. Some embodiments contemplate the use of one or more antibodies of the present invention in assays to detect a disease or disorder such as viral infection. Exemplary diagnostic assays for influenza-HA may comprise, *e.g.*, contacting a sample, obtained from a patient, with an anti-influenza-HA antibody

of the invention, wherein the anti-influenza-HA antibody is labeled with a detectable label or reporter molecule or used as a capture ligand to selectively isolate influenza-HA from patient samples. Alternatively, an unlabeled anti-influenza-HA antibody can be used in diagnostic applications in combination with a secondary antibody which is itself detectably labeled. The detectable label or reporter molecule can be a radioisotope, such as ^3H , ^{14}C , ^{32}P , ^{35}S , or ^{125}I ; a fluorescent or chemiluminescent moiety such as fluorescein isothiocyanate, or rhodamine; or an enzyme such as alkaline phosphatase, β -galactosidase, horseradish peroxidase, or luciferase. Specific exemplary assays that can be used to detect or measure influenza-HA in a sample include enzyme-linked immunosorbent assay (ELISA), radioimmunoassay (RIA), and fluorescence-activated cell sorting (FACS).

[0207] Samples that can be used in influenza-HA diagnostic assays according to the present invention include any tissue or fluid sample obtainable from a patient, which contains detectable quantities of either influenza HA, or fragments thereof, under normal or pathological conditions. Generally, levels of influenza HA in a particular sample obtained from a healthy patient (*e.g.*, a patient not afflicted with a disease associated with influenza will be measured to initially establish a baseline, or standard, level of influenza-HA. This baseline level of influenza-HA can then be compared against the levels of influenza-HA measured in samples obtained from individuals suspected of having a influenza-HA -associated condition, or symptoms associated with such condition.

[0208] The antibodies specific for influenza HA may contain no additional labels or moieties, or they may contain an N-terminal or C-terminal label or moiety. In one embodiment, the label or moiety is biotin. In a binding assay, the location of a label (if any) may determine the orientation of the peptide relative to the surface upon which the peptide is bound. For example, if a surface is coated with avidin, a peptide containing an N-terminal biotin will be oriented such that the C-terminal portion of the peptide will be distal to the surface.

EXAMPLES

[0209] The following examples are put forth so as to provide those of ordinary skill in the art with a complete disclosure and description of how to make and use the methods and compositions of the invention, and are not intended to limit the scope of what the inventors regard as their invention. Efforts have been made to ensure accuracy with respect to numbers used (*e.g.*, amounts, temperature, etc.) but some experimental errors and deviations should be accounted for. Unless indicated otherwise, parts are parts by weight, molecular weight is average molecular weight, temperature is in degrees Centigrade, room temperature is about 25°C, and pressure is at or near atmospheric.

Example 1: Generation of Human Antibodies to Influenza HA

[0210] Human antibodies to influenza HA were generated in a VELOCIMMUNE[®] mouse comprising DNA encoding human immunoglobulin heavy and kappa light chain variable regions. The mice were immunized with an influenza vaccine composition, followed by a booster dose comprising a mixture of five different recombinant hemagglutinin proteins, at a 1:1 ratio of each. The five recombinant hemagglutinin proteins included in the booster were hemagglutinins from H1 A/New Caledonia/20/1999, H5 A/Indonesia/05/2005, H3 A/Victoria/361/2011, H7 A/Netherlands/219/2003 and H9 A/Hong Kong/1073/1988 (Protein Sciences, Catalog Number 3006). The antibody immune response was monitored by an influenza A HA specific immunoassay. When a desired immune response was achieved, splenocytes were harvested and fused with mouse myeloma cells to preserve their viability and form hybridoma cell lines. The hybridoma cell lines were screened and selected to identify cell lines that produce influenza HA-specific antibodies. Using this technique, and the various immunogens described above, several chimeric antibodies (*i.e.*, antibodies possessing human variable domains and mouse constant domains) were obtained; exemplary antibodies generated in this manner were designated as H1H11820N, H1H11829N, H1H11829N2, H2M11830N, H1H11830N2, H1H11903N and H1H14571N.

[0211] Anti-influenza HA antibodies were also isolated directly from antigen-positive mouse B cells without fusion to myeloma cells, as described in U.S. Patent 7582298. Using this method, several fully human anti-influenza HA antibodies (*i.e.*, antibodies possessing human variable domains and human constant domains) were obtained; exemplary antibodies generated in this manner were designated as H1H11723P, H1H11729P, H1H11704P, H1H11711P, H1H11714P, H1H11717P, H1H11724P, H1H11727P, H1H11730P, H1H11731P2, H1H11734P2, H1H11736P2, H1H11742P2, H1H11744P2, H1H11745P2, H1H11747P2, H1H11748P2.

[0212] The biological properties of the exemplary antibodies generated in accordance with the methods of this Example are described in detail in the Examples set forth below.

Example 2: Heavy and Light Chain Variable Region Amino Acid and Nucleotide Sequences

[0213] Table 1 sets forth the amino acid sequence identifiers of the heavy and light chain variable regions and CDRs of selected anti-influenza HA antibodies of the invention. The corresponding nucleic acid sequence identifiers are set forth in Table 2.

Table 1: Amino Acid Sequence Identifiers

Antibody Designation	SEQ ID NOs:							
	HCVR	HCDR1	HCDR2	HCDR3	LCVR	LCDR1	LCDR2	LCDR3
H1H11723P	2	4	6	8	10	12	14	16
H1H11729P	18	20	22	24	26	28	30	32
H1H11820N	34	36	38	40	42	44	46	48
H1H11829N	50	52	54	56	58	60	62	64
H1H11829N2	50	52	54	56	66	68	70	72
H2aM11829N	50	52	54	56	58	60	62	64

H2M11830N	74	76	78	80	82	84	86	88
H1H11830N2	74	76	78	80	66	68	70	72
H1H11903N	90	92	94	96	98	100	102	104
H1H14571N	106	108	110	112	114	116	118	120
H2a14571N	106	108	110	112	114	116	118	120
H1H11704P	122	124	126	128	130	132	134	136
H1H11711P	138	140	142	144	146	148	150	152
H1H11714P	154	156	158	160	162	164	166	168
H1H11717P	170	172	174	176	178	180	182	184
H1H11724P	186	188	190	192	194	196	198	200
H1H11727P	202	204	206	208	210	212	214	216
H1H11730P2	218	220	222	224	226	228	230	232
H1H11731P2	234	236	238	240	66	68	70	72
H1H11734P2	242	244	246	248	66	68	70	72
H1H11736P2	250	252	254	256	66	68	70	72
H1H11742P2	258	260	262	264	66	68	70	72
H1H11744P2	266	268	270	272	66	68	70	72
H1H11745P2	274	276	278	280	66	68	70	72
H1H11747P2	282	284	286	288	66	68	70	72
H1H11748P2	290	292	294	296	66	68	70	72

Table 2: Nucleic Acid Sequence Identifiers

Antibody Designation	SEQ ID NOs:							
	HCVR	HCDR1	HCDR2	HCDR3	LCVR	LCDR1	LCDR2	LCDR3
H1H11723P	1	3	5	7	9	11	13	15
H1H11729P	17	19	21	23	25	27	29	31
H1H11820N	33	35	37	39	41	43	45	47
H1H11829N	49	51	53	55	57	59	61	63
H1H11829N2	49	51	53	55	65	67	69	71
H2aM11829N	49	51	53	55	57	59	61	63
H2M11830N	73	75	77	79	81	83	85	87
H1H11830N2	73	75	77	79	65	67	69	71
H1H11903N	89	91	93	95	97	99	101	103
H1H14571N	105	107	109	111	113	115	117	119
H2a14571N	105	107	109	111	113	115	117	119
H1H11704P	121	123	125	127	129	131	133	135
H1H11711P	137	139	141	143	145	147	149	151
H1H11714P	153	155	157	159	161	163	165	167
H1H11717P	169	171	173	175	177	179	181	183
H1H11724P	185	187	189	191	193	195	197	199
H1H11727P	201	203	205	207	209	211	213	215
H1H11730P	217	219	221	223	225	227	229	231
H1H11731P2	233	235	237	239	65	67	69	71
H1H11734P2	241	243	245	247	65	67	69	71
H1H11736P2	249	251	253	255	65	67	69	71
H1H11742P2	257	259	261	263	65	67	69	71
H1H11744P2	265	267	269	271	65	67	69	71
H1H11745P2	273	275	277	279	65	67	69	71
H1H11747P2	281	283	285	287	65	67	69	71
H1H11748P2	289	291	293	295	65	67	69	71

[0214] Antibodies are typically referred to herein according to the following nomenclature: Fc prefix (e.g. "H1H," "H2M," etc.), followed by a numerical identifier (e.g. "11723," "11830," etc., as shown in Table 1 or 2), followed by a "P," "P2," "N", N2, or "B" suffix. Thus, according to this nomenclature, an antibody may be referred to herein as, e.g., " H1H11723P," " H2M11830N," etc. The H1H and H2M prefixes on the antibody designations used herein indicate the particular Fc region isotype of the antibody. For example, an "H1M" antibody has a mouse IgG1 Fc, and an "H2M" antibody has a mouse IgG2 Fc (a or b isotype) (all variable regions are fully human as denoted by the first 'H' in the antibody designation). As will be appreciated by a person of ordinary skill in the art, an antibody having a particular Fc isotype can be converted to an antibody with a different Fc isotype (e.g., an antibody with a mouse IgG1 Fc can be converted to an antibody with a human IgG4, etc.), but in any event, the variable domains (including the CDRs) – which are indicated by the numerical identifiers shown in Tables 1, 2, 12 or 13– will remain the same, and the binding properties to antigen are expected to be identical or substantially similar regardless of the nature of the Fc domain.

Antibody Comparators

[0215] Anti-influenza-HA antibody controls were included in some of the following Examples for comparative purposes. Isotype matched negative controls were also used in the Examples. One anti-influenza HA comparator antibody, designated herein as "Control I mAb", is an anti-influenza HA antibody with heavy (HC) and light chain (LC) amino acid sequences as set forth in WO2008/028946 as SEQ ID NO: 65 (HC) and SEQ ID NO: 91 (LC) and also referred to as CR6261. As shown in WO2008/028946, this CR6261 antibody has heavy chain complementarity determining region (HCDR) amino acid sequences (HCDR 1, 2 and 3) as shown as SEQ ID NOs: 1,2 and 3, respectively and light chain complementarity determining region (LCDR) amino acid sequences (LCDR1,2 and 3) as shown in SEQ ID NOs: 13, 14 and 15, respectively.

Example 3: Octet Binding affinities and kinetic constants of monoclonal anti-influenza HA antibodies

[0216] Equilibrium dissociation constants (K_D values) for Influenza hemagglutinin (HA) binding to selected purified anti-HA monoclonal antibodies were determined using a real-time bio-layer interferometer based biosensor (Octet HTX) assay. Octet biosensors coated with either anti-mouse Fc (AMC) or anti-human Fc (AHC) capture were used to capture anti-HA monoclonal antibodies, expressed with mouse Fc (AbPID prefix H1M, H2aM, H2bM) or human IgG1 Fc (AbPID prefix H1H) respectively. All the binding studies were performed in HBS-ET Octet kinetics buffer (10mM HEPES pH 7.4, 0.15M NaCl, 3mM EDTA, 0.05% v/v Surfactant Tween-20, 1mg/mL BSA) at 25°C with plates shaking at a speed of 1000rpm. Antibody captured Octet biosensors were submerged in wells containing different concentrations of diverse strains of HA

(300nM to 11.11nM) for 7 minutes followed by dissociation of antibody bound HA protein in Octet kinetics buffer for 10 minutes. Biosensors were always washed in Octet kinetics buffer in between different steps. Kinetic association (k_a) and dissociation (k_d) rate constants were determined by processing and fitting the data to a 1:1 binding model using Scrubber 2.0c curve fitting software. Binding dissociation equilibrium constants (K_D) and dissociative half-lives ($t_{1/2}$) were calculated from the kinetic rate constants as:

$$K_D (M) = \frac{k_a}{k_d}, \quad \text{and} \quad t_{1/2} (\text{min}) = \frac{\ln(2)}{60 \times k_d}$$

[0217] Binding kinetics parameters for different anti-HA monoclonal antibodies binding to varying Group-1 strains of HA are tabulated in Table 3. In addition, the binding parameters for the same antibodies were determined for binding to group 2 hemagglutinins, however, there was no binding to group 2 HA (data not shown).

Table 3: Binding Kinetics parameters of monoclonal antibodies binding to different HA strains of Group-1 at 25 °C.

mAb Captured	Injected Analyte	Amount of mAb Captured (nm)	300nM HA Bound (nm)	k_a (1/Ms)	k_d (1/s)	K_D (M)	$t_{1/2}$ (min)
H2aM11829N	A/California/07/2009 (H1N1)	0.53 ± 0.01	0.62	1.20E+05	1.00E-05*	8.35E-11	1155*
	A/Puerto Rico/08/34 (H1N1)	0.53 ± 0.01	0.59	1.09E+05	1.16E-05	1.06E-10	999
	A/Vietnam/1203/2004 (H5N1)	0.52 ± 0.01	0.50	3.81E+04	1.99E-05	5.23E-10	580
H1H11729P	A/California/07/2009 (H1N1)	0.65 ± 0.01	0.84	7.35E+04	1.00E-05*	1.36E-10	1155*
	A/Puerto Rico/08/34 (H1N1)	0.67 ± 0.01	0.72	8.26E+04	1.00E-05*	1.21E-10	1155*
	A/Vietnam/1203/2004 (H5N1)	0.66 ± 0.01	0.62	1.99E+04	3.10E-05	1.56E-09	373
H2aM14571N	A/California/07/2009 (H1N1)	0.51 ± 0.01	0.58	1.05E+05	1.18E-05	1.12E-10	978
	A/Puerto Rico/08/34 (H1N1)	0.5 ± 0.02	0.46	6.80E+04	2.44E-05	3.59E-10	474
	A/Vietnam/1203/2004 (H5N1)	0.51 ± 0.01	0.31	1.14E+04	1.00E-05*	8.77E-10	1155*

Example 4: HA Octet cross-competition

[0218] Binding competition between a panel of different anti-influenza HA monoclonal antibodies was determined using a real time, label-free bio-layer interferometry assay on an Octet® HTX biosensor (ForteBio, A Division of Pall Life Sciences). The entire experiment was performed at 25 °C in 0.01M HEPES pH7.4, 0.15M NaCl, 0.05% v/v Surfactant Tween-20, and 1mg/mL BSA (HBS-ET Octet buffer) with the plate shaking at a speed of 1000rpm. To assess whether two antibodies were able to compete with one another for binding to their respective epitopes on HA, a pre-mix assay format was adopted. One Group-1 strain of HA protein (California) was used to study cross-competition between different anti-HA monoclonal antibodies. To achieve this, 100nM of HA reagent (Protein Sciences) was first pre-mixed with 1µM concentration of different anti-HA monoclonal antibodies (subsequently referred to as mAb-2) for at least 2 hours prior to running the binding competition assay. Octet biosensors coated with either an anti-mouse Fc polyclonal antibody (Pall ForteBio Corp., # 18-5088; subsequently referred as AMC) or with an anti-human Fc polyclonal antibody (Pall ForteBio Corp., # 18-5060; subsequently referred as AHC) were first submerged into wells containing 20µg/mL solution of individual anti-HA monoclonal antibodies for 3 minutes to capture anti-HA monoclonal antibodies expressed either with a mouse Fc or with a human Fc, respectively (subsequently referred to as mAb-1). Following the capture step, unoccupied anti-mouse Fc polyclonal antibody and anti-human Fc polyclonal antibody on the Octet biosensors were saturated by submerging into wells containing 1µM solution of a mixture of irrelevant monoclonal antibody expressed with different Fc (human IgG1, mouse IgG2a and mouse IgG2b) for 3 minutes. Finally, the Octet biosensors were immersed for 5 minutes into wells containing the pre-mixed samples of 100nM of HA and 1µM concentration of mAb-2. At the end of each cycle, the captured anti-HA monoclonal antibodies along with the bound pre-complex of HA and mAb-2 were regenerated using three alternate 20 second dips into 20mM H₃PO₄ followed by submerging into HBS-ET Octet buffer. The biosensors were washed in HBS-ET Octet buffer in between every step of the experiment. The real-time binding response was monitored during the entire course of the experiment and the binding response at the end of every step was recorded. During the analysis, the self-self background binding signal (where mAb-1 = mAb-2) caused due to the non-specific binding of anti-HA monoclonal antibody to the unoccupied capture surface was subtracted from the entire column and a cross-competition table was generated (Table 4). The response of mAb-1 binding to the pre-complex of HA and mAb-2 was compared and competitive/non-competitive behavior of different anti-HA monoclonal antibodies was determined.

[0219] As shown in Table 4, the column on the left shows the mAb1 antibodies that are captured using the AHC Octet biosensors and the column on the right demonstrates the antibodies (mAb2) that cross-compete with the mAb1 antibody.

Table 4: Cross-competition between a panel of different anti-HA monoclonal antibodies for binding to H1N1 California strain of HA

First mAb (mAb-1) Captured using AHC Octet Biosensors	mAb-2 Antibodies Shown to Compete with mAb-1
H2aM11829N	H2aM14571N
	H1H11729P
H2aM14571N	H2aM11829N
	H1H11729P
H1H11729P	H2aM11829N
	H2aM14571N

Example 4: Selected Group 1-Specific Influenza A Hemagglutinin Monoclonal Antibodies Show Potent Neutralization of Influenza A *in vitro* across diverse group 1 strains

Cell viability microneutralization assay

[0220] Monoclonal antibodies were tested in a microneutralization assay to evaluate breadth and potency. In brief, Madin-Darby canine kidney (MDCK) cells were plated at 6.0×10^3 cells/well in a 96-well plate. Virus diluent only was added to background control wells. Cells were incubated for 4 to 5 hours at 37 °C with 5% CO₂. Monoclonal antibodies were diluted at 4 times final concentration in virus diluent. The antibodies were diluted 1:3 in duplicate. Virus was thawed on ice and diluted to the appropriate pre-determined concentration. Diluted virus was added to diluted mAbs. The mAb-virus mixture was immediately transferred to the MDCK cells and incubated at 37 °C with 5% CO₂ for 72h. Virus control and uninfected cell control wells were also included. On day 4, the plates were centrifuged at 1200 RPM for 3 minutes. Cells were lysed using 100 µL CelTiter-Glo substrate and ATP release measured using luminescence (Victor X3, PerkinElmer). Percent viability was determined relative to uninfected control. Viability values were analyzed using non-linear 4PL regression to determine IC₅₀ values (GraphPad Prism).

[0221] The mAbs showed potency against a broad range of group 1 influenza A viruses (Table 5), including various H1N1, H5N1, H9N2, H13N6, and H16N3 strains. The IC₅₀ values ranged from 1.68 nM to 48 nM for H2aM11829N and from 2.47 nM to 129.5 nM for H1H11729P.

Table 5. Neutralization potency against various representative group 1 Influenza strains. (average of 3 to 5 runs shown)

PID	IC ₅₀ (nM)						
	H1_ PR34	H1_ WS33	H1_ IVR148	H1_ CA09	H9_ RG26	H13_ MD77	H16_ DE09
H2aM11829N	3.29	1.68	48.0	3.50	10.2	2.11	7.39
H1H11729P	2.82	2.47	129.5	4.54	14.9	33.8	21.6

Example 5. Selected group 1-specific Influenza A hemagglutinin monoclonal antibodies show potent neutralization of Influenza A *in vitro*

[0222] The exemplary monoclonal antibodies (mAb) H1H11829N2 and H1H11729P were selected for *in vitro* microneutralization assays. Briefly, Madin-Darby Canine Kidney (MDCK) cells were plated and incubated overnight to achieve 80 -100% confluency the next day. Monoclonal antibodies were diluted in viral infection medium (VIM) to 50 µg/mL and diluted 1:2 in triplicate or quadruplicate. H5N1 A/Vietnam/1203/2004 or H1N1 A/California/07/2009 was diluted in VIM and added to the diluted antibodies and incubated for 1 h. The samples were then transferred to the MDCKs and incubated for 48 h. After the incubation, 50 µL of the supernatant was transferred to a new 96-well plate. Diluted turkey or horse red blood cells were added to the supernatant and incubated at room temperature for 30 or 60 min. The hemagglutination titer was recorded as the reciprocal of the last dilution that completely inhibited hemagglutination.

[0223] Exemplary antibodies H1H11829N2 and H1H11729P show potent neutralization of H5N1 A/Vietnam/1203/2004 and H1N1 A/California/07/2009 (Table 6). The average IC₉₀ values for exemplary antibodies H1H11829N2 and H1H11729P with respect to neutralization of H5N1 A/Vietnam/1203/2004 were on average 62.50 nM and 26.05 nM (respectively).

Table 6. Hemagglutinin titers after microneutralization assay with H5N1 A/Vietnam/1203/2004 and H1N1 A/California/07/2009 with selected group 1 hemagglutinin-specific antibodies. Shown are the IC₉₀ values (nM).

Antibody Number	IC ₉₀ (nM)			IC ₉₀ (nM)	
	H5N1 A/Vietnam/1203/2004			H1N1 A/California/07/2009	
	Trial 1	Trial 2		Trial 1	
H1H11829N2	83.3	41.7		41.7	
H1H11729P	41.7	10.4		83.3	

Example 6. Complement-dependent Cytotoxicity of Influenza infected cells *in vitro* using influenza A specific monoclonal antibodies

[0224] The exemplary monoclonal antibodies (mAb) H1H11829N2 and H1H11729P were selected for *in vitro* complement-dependent cytotoxicity (CDC) assays. Briefly, Madin-Darby Canine Kidney (MDCK) cells were plated 24 hours prior to infection with H1N1 A/Puerto Rico/08/1934 at an MOI of 3. After an incubation of 20 to 25 hours, cells were harvested and resuspended in CDC assay medium at a concentration of 1 x 10⁶ cells/mL. The diluted target cells (20 µL) were added to each well of a 384-well plate. Monoclonal antibodies were diluted to three times the final starting concentration in CDC assay medium and then diluted 1:2 in triplicate or quadruplicate. CDC assay medium was added to each well not receiving mAb. The plate was shaken at room temperature for ~ 2 min at ~ 500 rpm. Normal human serum complement (NHSC) was prepared at three times the final concentration (15%) in CDC assay medium and 20 µL was added to each well. The plate was incubated for 2 h at room

temperature and the lysis reagent, buffer and substrate was brought to room temperature at this time. The lysis reagent was prepared by dilution of digitonin into CDC assay medium and 5 μ L of this reagent was added to the maximal lysis control wells to establish the maximal signal control. The substrate and buffer were combined and 20 μ L added to all wells and incubated for 10 min at room temperature. The signal was detected on a plate reader (Victor X3, Perkin Elmer). The percent specific lysis was calculated as $((\text{lysis by mAb} + \text{NHSC}) - (\text{lysis by NHSC only})) / ((\text{digitonin maximal lysis}) - (\text{lysis by NHSC only})) \times 100$. Analysis was completed by a four-parameter non-linear regression over an 8-point response curve (GraphPad Prism). Data was plotted for individual trials $\pm$ standard deviation, and data presented as averages of all trails $\pm$ standard error of the mean.

[0225] Exemplary antibodies H1H11829N2 and H1H11729P show potent complement-mediated lysis of infected cells. Three randomly selected trials are shown in Table 7. The average EC_{50} values for exemplary antibodies H1H11829N2 and H1H11729P were on average 41.88 nM ($\pm$ 13.08 SEM) and H1H11729P were 43.25 nM ($\pm$ 3.83 SEM) respectively.

Table 7. Percent specific lysis of H1N1 A/Puerto Rico/08/1934-infected target cells by group 1 hemagglutinin-specific antibodies. Shown are the EC_{50} values (nM) for each of three replicates.

AbPID	EC_{50} (nM)		
	Trial 1	Trial 2	Trial 3
H1H11829N2	65.94	38.75	20.94
H1H11729P	50.83	38.49	40.44
hlgG1 irrelevant mAb	> 333	> 333	> 333

Example 7: Selected Group 1-Specific Influenza A Hemagglutinin Monoclonal Antibodies Show Potent Neutralization of Influenza A Infection *in vivo*

Single dose prophylaxis mouse model

[0226] The exemplary monoclonal antibodies H2aM11829N, H1H11729P and H2a14571N were selected for *in vivo* protection studies using BALB/c mice. Briefly, approximately 6-week old, female mice (approximately 17.5 ± 0.5 g) were injected subcutaneously (SC) with 1 mg/kg of H2aM11829N, H1H11729P, H2a14571N, or mouse or human isotype (negative control) antibody on day -1. Five mice were included per group per experiment. Twenty-four hours post-injection (day 0), mice were challenged intranasally (IN) with $10 \times MLD_{50}$ (800 plaque forming units; PFUs) in 20 μ L saline with H1N1 A/Puerto Rico/08/1934. Mice were weighed daily and were sacrificed when they lost > 20 % of their initial starting weight.

[0227] Exemplary antibodies H2aM11829N, H1H11729P and H2a14571N show potent neutralization of influenza A infection *in vivo* at 1 mg/kg.

Dose response prophylaxis mouse model

[0228] The exemplary monoclonal antibodies H1H11829N2 and H1H11729P were selected for prophylactic *in vivo* protection studies using BALB/c mice. Briefly, 6-week old, female mice (approximately 17.5 ± 0.5 g) were injected SC with 1, 0.5, 0.1 or 0.05 mg/kg or 2, 0.5, 0.2 or 0.05 mg/kg of H1H11829N2, H1H11729P antibody on day -1. Five mice were included per group per experiment. Carrier protein (an irrelevant hlgG1 mAb) was added to all doses of mAb at 1 mg/mL. Twenty-four hours post-injection (day 0), mice were challenged intranasally with $10 \times \text{MLD}_{50}$ (800 plaque forming units; PFUs) in 20 μL saline with H1N1 A/Puerto Rico/08/1934 or $5 \times \text{MLD}_{50}$ (5,000 PFUs) in 20 μL saline with H1N1 A/California/04/2009. Mice were weighed daily and were sacrificed when they lost $> 20\%$ of their initial starting weight.

[0229] When challenged with H1_PR34, exemplary antibodies H1H11829N2 and H1H11729P show potent neutralization of influenza A infection *in vivo* at 1 mg/kg. At 0.5 and 0.1 mg/kg, H1H11729P shows 100% and 40% survival, respectively. (Tables 8, 9 and 10). When challenged with H1_CA09, exemplary antibodies H1H11829N2 and H1H11729P show potent neutralization of influenza A infection *in vivo* at 2, 0.5 and 0.2 mg/kg and showed 60% survival at 0.05 mg/kg. (Tables 8, 9 and 10).

Table 8. Percent survival of mice in experiment 1 after administration of 1 mg/kg group 1 hemagglutinin-specific antibodies, SC prophylactically one day before infection with $10 \times \text{MLD}_{50}$ of H1N1 A/Puerto Rico/08/1934.

PID	Number of mice per group	Percent survival (no. of surviving mice/total no. of mice in the group)
Saline (uninfected)	5	100 (5/5)
H2aM11829N	5	100 (5/5)
H1H11729P	5	100 (5/5)
H2a14571N	ND	ND
hlgG1 isotype control	5	0 (0/5)
mlgG2a isotype control	5	0 (0/5)

Table 9. Percent survival of mice in experiment 2 after administration of 1 mg/kg group 1 hemagglutinin-specific antibodies, SC prophylactically one day before infection with $10 \times \text{MLD}_{50}$ of H1N1 A/Puerto Rico/08/1934.

PID	Number of mice per group	Percent survival (no. of surviving mice/total no. of mice in the group)
Saline (uninfected)	5	100 (5/5)
H2aM11829N	5	100 (5/5)
H1H11729P	5	100 (5/5)
H2a14571N	5	100 (5/5)

Table 10. Percent survival of mice after administration of 1, 0.5, 0.1 or 0.05 mg/kg group 1 hemagglutinin-specific antibodies, SC prophylactically one day before infection with 10 x MLD₅₀ of H1N1 A/Puerto Rico/08/1934 or 2, 0.5, 0.2 or 0.05 mg/kg, SC prophylactically one day before infection with 5 x MLD₅₀ of H1N1 A/California/04/2009.

PID	Challenge virus: H1N1 A/Puerto Rico/08/1934			Challenge virus: H1N1 A/California/04/2009		
	Dose of mAb (mg/kg)	Number of mice per group	Percent survival (no. of surviving mice/total no. of mice in the group)	Dose of mAb (mg/kg)	Number of mice per group	Percent survival (no. of surviving mice/total no. of mice in the group)
Saline with carrier protein (uninfected)	N/A	5	100 (5/5)	N/A	5	100 (5/5)
H1H11829N2	1	5	100 (5/5)	2	5	100 (5/5)
	0.5		60 (3/5)	0.5		100 (5/5)
	0.1		0 (0/5)	0.2		100 (5/5)
	0.05		20 (1/5)	0.05		ND
H1H11729P	1	5	100 (5/5)	2	5	100 (5/5)
	0.5		100 (5/5)	0.5		100 (5/5)
	0.1		40 (2/5)	0.2		100 (5/5)
	0.05		0 (0/5)	0.05		60 (3/5)

Dose response therapeutic mouse model

[0230] The exemplary monoclonal antibodies H1H11829N2 and H1H11729P were selected for therapeutic *in vivo* protection studies using BALB/c mice. Briefly, 6-week old, female mice (approximately 17.5 ± 0.5 g) were infected intranasally with 10 x MLD₅₀ (800 plaque forming units; PFUs) in 20 µL saline with H1N1 A/Puerto Rico/08/1934 or 5 x MLD₅₀ (5,000 PFUs) in 20 µL saline with H1N1 A/California/04/2009. Three, four or five days (in the case of H1_PR34) or one, two or three days (in the case of H1_CA09) post-infection, separate groups of 5 mice were injected IV with 15 mg/kg or 30 mg/kg of H1H11829N2, or H1H11729P antibody. Mice were weighed daily and were sacrificed when they lost > 20 % of their initial starting weight.

[0231] When challenged with both a historical and contemporary H1N1 virus strain, exemplary antibodies H1H11829N2 and H1H11729P show potent neutralization of influenza A infection *in vivo*. (Table 11).

Table 11. Percent survival of mice in a therapeutic model of infection. Mice were infected with 10 x MLD₅₀ of H1N1 A/Puerto Rico/08/1934 or 5 x MLD₅₀ of H1N1 A/California/04/2009 on day 0. On days 3, 4 or 5 (for H1N1 A/Puerto Rico/08/1934) or days 1, 2 or 3 (for H1N1 A/California/04/2009) mice antibodies were administered at 15 or 30 mg/kg, IV.

PID	Challenge virus: H1N1 A/Puerto Rico/08/1934 (15 mg/kg mAb)			Challenge virus: H1N1 A/California/04/2009 (30 mg/kg mAb)		
	Days post - infection	Number of mice per	Percent survival (no. of	Days post-infection	Number of mice per	Percent survival (no. of

		group	surviving mice/total no. of mice in the group		group	surviving mice/total no. of mice in the group
H1H11829N2	3	5	40 (2/5)	1	5	100 (5/5)
	4		ND	2		0 (0/5)
	5		ND	3		20 (1/5)
H1H11729P	3	5	60 (3/5)	1	5	100 (5/5)
	4		0 (0/5)	2		0 (0/5)
	5		0 (0/5)	3		20 (1/5)

Example 8. Selected group 1-specific Influenza A hemagglutinin monoclonal antibodies are generated after immunization with H5 immunogens.

[0232] Additional human antibodies to Influenza A-HA were generated in VELOCIMMUNE® mice comprising DNA encoding human immunoglobulin heavy and kappa light chain variable regions. Naïve mice were divided into four groups. Groups 1 and 2 were immunized with trivalent influenza vaccine (TIV) afluria® 2013-2014 formulation (NDC 3332-110-10). Groups 3 and 4 were left unimmunized. Approximately, four weeks later, groups 1 and 3 were injected intramuscularly with 50 µg H5 DNA (VRC9123 encoding H5 A/Indonesia/05/2005 (see Ledgerwood *et al.* (2011) *Lancet Infect. Dis.* 11: 916-924; Khurana *et al.* (2013) *J. Infect. Dis.* 208:413-417; Ledgerwood *et al.* (2013) *J. Infect. Dis.* 208: 418-422) and groups 2 and 4 were injected intramuscularly with 5 µg H5 monovalent influenza vaccine (MIV; VRC310 containing H5 A/Indonesia/05/2005) (see Ledgerwood *et al.* (2011) *Lancet Infect. Dis.* 11: 916-924, Khurana *et al.* (2013) *J. Infect. Dis.* 208:413-417; Ledgerwood *et al.* (2013) *J. Infect. Dis.* 208: 418-422). Approximately four weeks later, all 4 groups were immunized with 5 µg of H5 monovalent influenza vaccine (MIV; VRC310 containing H5 A/Indonesia/05/2005).

[0233] Anti-influenza A-HA antibodies were isolated directly from receptor-binding site (RBS) mutated, avi-tagged H5 A/Indonesia/05/2005 and/or H1 A/New Caledonia/20/1999 that contained a carboxy-terminal trimerization domain from T4 phage fibrin (foldon)-positive mouse B cells without fusion to myeloma cells, as described in U.S. Patent 7582298. Using this method, several additional fully human anti-influenza A-HA antibodies (*i.e.*, antibodies possessing human variable domains and human constant domains) were obtained; exemplary antibodies generated in this manner can be found in Tables 12 (amino acid sequence identifiers) and 13 (nucleic acid sequence identifiers) below. The samples were evaluated for binding to receptor-binding site (RBS) mutated, foldon, avi-tagged H5 and H1 via Luminex. As expected, all but 16 samples bound to H5 protein with mean fluorescent intensities (MFI) of greater than 15,000. However, for the most part, samples that also bound to H1 protein had high binding to H5 (*i.e.*, greater than 15,000 MFI). Two groups of monoclonal antibodies were observed; those which bound to H1 with greater than 15,000 MFI (20 samples) and those that bound to H5 with greater than 5000 MFI (44 samples).

Table 12: Amino Acid Sequence Identifiers

Amino Acid SEQ ID NOs:								
Antibody Designation	HCVR	HCDR1	HCDR2	HCDR3	LCVR	LCDR1	LCDR2	LCDR3
H1H17952B	298	300	302	304	306	308	310	312
H1H17953B	314	316	318	320	322	324	326	328
H1H17954B	330	332	334	336	338	340	342	344
H1H17955B	346	348	350	352	354	356	358	360
H1H17956B	362	364	366	368	370	372	374	376
H1H17957B	378	380	382	384	386	388	390	392
H1H17958B	394	396	398	400	402	404	406	408
H1H17959B	410	412	414	416	418	420	422	424
H1H17960B	426	428	430	432	434	436	438	440
H1H17961B	442	444	446	448	450	452	454	456
H1H17962B	458	460	462	464	466	468	470	472
H1H17963B	474	476	478	480	482	484	486	488
H1H17964B	490	492	494	496	498	500	502	504
H1H17965B	506	508	510	512	514	516	518	520
H1H17966B	522	524	526	528	530	532	534	536
H1H17967B	538	540	542	544	546	548	550	552
H1H17968B	554	556	558	560	562	564	566	568
H1H17969B	570	572	574	576	578	580	582	584
H1H17970B	586	588	590	592	594	596	598	600
H1H17971B	602	604	606	608	610	612	614	616
H1H17972B	618	620	622	624	626	628	630	632
H1H17973B	634	636	638	640	642	644	646	648
H1H17974B	650	652	654	656	658	660	662	664
H1H17975B	666	668	670	672	674	676	678	680
H1H17976B	682	684	686	688	690	692	694	696
H1H17977B	698	700	702	704	706	708	710	712
H1H17978B	714	716	718	720	722	724	726	728
H1H17979B	730	732	734	736	738	740	742	744
H1H17980B	746	748	750	752	754	756	758	760
H1H17981B	762	764	766	768	770	772	774	776
H1H17982B	778	780	782	784	786	788	790	792
H1H17983B	794	796	798	800	802	804	806	808
H1H17984B	810	812	814	816	818	820	822	824
H1H17985B	826	828	830	832	834	836	838	840
H1H17986B	842	844	846	848	850	852	854	856
H1H17987B	858	860	862	864	866	868	870	872
H1H17988B	874	876	878	880	882	884	886	888
H1H17989B	890	892	894	896	898	900	902	904
H1H17990B	906	908	910	912	914	916	918	920
H1H17991B	922	924	926	928	930	932	934	936
H1H17992B	938	940	942	944	946	948	950	952

H1H17993B	954	956	958	960	962	964	966	968
H1H17994B	970	972	974	976	978	980	982	984
H1H17995B	986	988	990	992	994	996	998	1000
H1H17996B	1002	1004	1006	1008	1010	1012	1014	1016
H1H17997B	1018	1020	1022	1024	1026	1028	1030	1032
H1H17998B	1034	1036	1038	1040	1042	1044	1046	1048
H1H17999B	1050	1052	1054	1056	1058	1060	1062	1064
H1H18000B	1066	1068	1070	1072	1074	1076	1078	1080
H1H18001B	1082	1084	1086	1088	1090	1092	1094	1096
H1H18002B	1098	1100	1102	1104	1106	1108	1110	1112
H1H18003B	1114	1116	1118	1120	1122	1124	1126	1128
H1H18004B	1130	1132	1134	1136	1138	1140	1142	1144
H1H18005B	1146	1148	1150	1152	1154	1156	1158	1160
H1H18006B	1162	1164	1166	1168	1170	1172	1174	1176
H1H18007B	1178	1180	1182	1184	1186	1188	1190	1192
H1H18008B	1194	1196	1198	1200	1202	1204	1206	1208
H1H18009B	1210	1212	1214	1216	1218	1220	1222	1224
H1H18010B	1226	1228	1230	1232	1234	1236	1238	1240
H1H18011B	1242	1244	1246	1248	1250	1252	1254	1256
H1H18012B	1258	1260	1262	1264	1266	1268	1270	1272
H1H18013B	1274	1276	1278	1280	1282	1284	1286	1288
H1H18014B	1290	1292	1294	1296	1298	1300	1302	1304
H1H18015B	1306	1308	1310	1312	1314	1316	1318	1320
H1H18016B	1322	1324	1326	1328	1330	1332	1334	1336
H1H18017B	1338	1340	1342	1344	1346	1348	1350	1352
H1H18018B	1354	1356	1358	1360	1362	1364	1366	1368
H1H18019B	1370	1372	1374	1376	1378	1380	1382	1384
H1H18020B	1386	1388	1390	1392	1394	1396	1398	1400
H1H18021B	1402	1404	1406	1408	1410	1412	1414	1416
H1H18022B	1418	1420	1422	1424	1426	1428	1430	1432
H1H18023B	1434	1436	1438	1440	1442	1444	1446	1448
H1H18024B	1450	1452	1454	1456	1458	1460	1462	1464
H1H18025B	1466	1468	1470	1472	1474	1476	1478	1480
H1H18026B	1482	1484	1486	1488	1490	1492	1494	1496
H1H18027B	1498	1500	1502	1504	1506	1508	1510	1512
H1H18028B	1514	1516	1518	1520	1522	1524	1526	1528
H1H18029B	1530	1532	1534	1536	1538	1540	1542	1544
H1H18030B	1546	1548	1550	1552	1554	1556	1558	1560
H1H18031B	1562	1564	1566	1568	1570	1572	1574	1576
H1H18032B	1578	1580	1582	1584	1586	1588	1590	1592
H1H18033B	1594	1596	1598	1600	1602	1604	1606	1608
H1H18034B	1610	1612	1614	1616	1618	1620	1622	1624
H1H18035B	1626	1628	1630	1632	1634	1636	1638	1640
H1H18037B	1642	1644	1646	1648	1650	1652	1654	1656
H1H18038B	1658	1660	1662	1664	1666	1668	1670	1672
H1H18039B	1674	1676	1678	1680	1682	1684	1686	1688
H1H18040B	1690	1692	1694	1696	1698	1700	1702	1704

H1H18041B	1706	1708	1710	1712	1714	1716	1718	1720
H1H18042B	1722	1724	1726	1728	1730	1732	1734	1736
H1H18043B	1738	1740	1742	1744	1746	1748	1750	1752
H1H18044B	1754	1756	1758	1760	1762	1764	1766	1768
H1H18045B	1770	1772	1774	1776	1778	1780	1782	1784
H1H18046B	1786	1788	1790	1792	1794	1796	1798	1800
H1H18047B	1802	1804	1806	1808	1810	1812	1814	1816
H1H18048B	1818	1820	1822	1824	1826	1828	1830	1832
H1H18049B	1834	1836	1838	1840	1842	1844	1846	1848
H1H18051B	1850	1852	1854	1856	1858	1860	1862	1864
H1H18052B	1866	1868	1870	1872	1874	1876	1878	1880
H1H18053B	1882	1884	1886	1888	1890	1892	1894	1896
H1H18054B	1898	1900	1902	1904	1906	1908	1910	1912
H1H18055B	1914	1916	1918	1920	1922	1924	1926	1928
H1H18056B	1930	1932	1934	1936	1938	1940	1942	1944
H1H18057B	1946	1948	1950	1952	1954	1956	1958	1960
H1H18058B	1962	1964	1966	1968	1970	1972	1974	1976
H1H18059B	1978	1980	1982	1984	1986	1988	1990	1992
H1H18060B	1994	1996	1998	2000	2002	2004	2006	2008
H1H18061B	2010	2012	2014	2016	2018	2020	2022	2024
H1H18062B	2026	2028	2030	2032	2034	2036	2038	2040
H1H18063B	2042	2044	2046	2048	2050	2052	2054	2056
H1H18064B	2058	2060	2062	2064	2066	2068	2070	2072
H1H18065B	2074	2076	2078	2080	2082	2084	2086	2088
H1H18066B	2090	2092	2094	2096	2098	2100	2102	2104
H1H18067B	2106	2108	2110	2112	2114	2116	2118	2120
H1H18068B	2122	2124	2126	2128	2130	2132	2134	2136
H1H18069B	2138	2140	2142	2144	2146	2148	2150	2152
H1H18070B	2154	2156	2158	2160	2162	2164	2166	2168
H1H18071B	2170	2172	2174	2176	2178	2180	2182	2184
H1H18072B	2186	2188	2190	2192	2194	2196	2198	2200
H1H18073B	2202	2204	2206	2208	2210	2212	2214	2216
H1H18074B	2218	2220	2222	2224	2226	2228	2230	2232
H1H18075B	2234	2236	2238	2240	2242	2244	2246	2248
H1H18076B	2250	2252	2254	2256	2258	2260	2262	2264
H1H18077B	2266	2268	2270	2272	2274	2276	2278	2280
H1H18078B	2282	2284	2286	2288	2290	2292	2294	2296
H1H18079B	2298	2300	2302	2304	2306	2308	2310	2312
H1H18080B	2314	2316	2318	2320	2322	2324	2326	2328
H1H18081B	2330	2332	2334	2336	2338	2340	2342	2344
H1H18082B	2346	2348	2350	2352	2354	2356	2358	2360
H1H18083B	2362	2364	2366	2368	2370	2372	2374	2376
H1H18084B	2378	2380	2382	2384	2386	2388	2390	2392
H1H18085B	2394	2396	2398	2400	2402	2404	2406	2408
H1H18086B	2410	2412	2414	2416	2418	2420	2422	2424
H1H18087B	2426	2428	2430	2432	2434	2436	2438	2440
H1H18088B	2442	2444	2446	2448	2450	2452	2454	2456

H1H18089B	2458	2460	2462	2464	2466	2468	2470	2472
H1H18090B	2474	2476	2478	2480	2482	2484	2486	2488
H1H18091B	2490	2492	2494	2496	2498	2500	2502	2504
H1H18092B	2506	2508	2510	2512	2514	2516	2518	2520
H1H18093B	2522	2524	2526	2528	2530	2532	2534	2536
H1H18094B	2538	2540	2542	2544	2546	2548	2550	2552
H1H18095B	2554	2556	2558	2560	2562	2564	2566	2568
H1H18096B	2570	2572	2574	2576	2578	2580	2582	2584
H1H18097B	2586	2588	2590	2592	2594	2596	2598	2600
H1H18098B	2602	2604	2606	2608	2610	2612	2614	2616
H1H18099B	2618	2620	2622	2624	2626	2628	2630	2632
H1H18100B	2634	2636	2638	2640	2642	2644	2646	2648
H1H18101B	2650	2652	2654	2656	2658	2660	2662	2664
H1H18102B	2666	2668	2670	2672	2674	2676	2678	2680
H1H18103B	2682	2684	2686	2688	2690	2692	2694	2696
H1H18104B	2698	2700	2702	2704	2706	2708	2710	2712
H1H18105B	2714	2716	2718	2720	2722	2724	2726	2728
H1H18107B	2730	2732	2734	2736	2738	2740	2742	2744
H1H18108B	2746	2748	2750	2752	2754	2756	2758	2760
H1H18109B	2762	2764	2766	2768	2770	2772	2774	2776
H1H18110B	2778	2780	2782	2784	2786	2788	2790	2792
H1H18111B	2794	2796	2798	2800	2802	2804	2806	2808
H1H18112B	2810	2812	2814	2816	2818	2820	2822	2824
H1H18113B	2826	2828	2830	2832	2834	2836	2838	2840
H1H18114B	2842	2844	2846	2848	2850	2852	2854	2856
H1H18115B	2858	2860	2862	2864	2866	2868	2870	2872
H1H18116B	2874	2876	2878	2880	2882	2884	2886	2888
H1H18117B	2890	2892	2894	2896	2898	2900	2902	2904
H1H18118B	2906	2908	2910	2912	2914	2916	2918	2920
H1H18119B	2922	2924	2926	2928	2930	2932	2934	2936
H1H18120B	2938	2940	2942	2944	2946	2948	2950	2952
H1H18121B	2954	2956	2958	2960	2962	2964	2966	2968
H1H18122B	2970	2972	2974	2976	2978	2980	2982	2984
H1H18123B	2986	2988	2990	2992	2994	2996	2998	3000
H1H18124B	3002	3004	3006	3008	3010	3012	3014	3016
H1H18125B	3018	3020	3022	3024	3026	3028	3030	3032
H1H18126B	3034	3036	3038	3040	3042	3044	3046	3048
H1H18127B	3050	3052	3054	3056	3058	3060	3062	3064
H1H18128B	3066	3068	3070	3072	3074	3076	3078	3080
H1H18129B	3082	3084	3086	3088	3090	3092	3094	3096
H1H18130B	3098	3100	3102	3104	3106	3108	3110	3112
H1H18131B	3114	3116	3118	3120	3122	3124	3126	3128
H1H18132B	3130	3132	3134	3136	3138	3140	3142	3144
H1H18133B	3146	3148	3150	3152	3154	3156	3158	3160
H1H18134B	3162	3164	3166	3168	3170	3172	3174	3176
H1H18135B	3178	3180	3182	3184	3186	3188	3190	3192
H1H18136B	3194	3196	3198	3200	3202	3204	3206	3208

H1H18137B	3210	3212	3214	3216	3218	3220	3222	3224
H1H18138B	3226	3228	3230	3232	3234	3236	3238	3240
H1H18139B	3242	3244	3246	3248	3250	3252	3254	3256
H1H18140B	3258	3260	3262	3264	3266	3268	3270	3272
H1H18141B	3274	3276	3278	3280	3282	3284	3286	3288
H1H18142B	3290	3292	3294	3296	3298	3300	3302	3304
H1H18143B	3306	3308	3310	3312	3314	3316	3318	3320
H1H18144B	3322	3324	3326	3328	3330	3332	3334	3336
H1H18145B	3338	3340	3342	3344	3346	3348	3350	3352
H1H18146B	3354	3356	3358	3360	3362	3364	3366	3368
H1H18147B	3370	3372	3374	3376	3378	3380	3382	3384
H1H18148B	3386	3388	3390	3392	3394	3396	3398	3400
H1H18149B	3402	3404	3406	3408	3410	3412	3414	3416
H1H18150B	3418	3420	3422	3424	3426	3428	3430	3432
H1H18151B	3434	3436	3438	3440	3442	3444	3446	3448
H1H18152B	3450	3452	3454	3456	3458	3460	3462	3464
H1H18153B	3466	3468	3470	3472	3474	3476	3478	3480
H1H18154B	3482	3484	3486	3488	3490	3492	3494	3496
H1H18155B	3498	3500	3502	3504	3506	3508	3510	3512
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H1H18159B	3562	3564	3566	3568	3570	3572	3574	3576
H1H18160B	3578	3580	3582	3584	3586	3588	3590	3592
H1H18161B	3594	3596	3598	3600	3602	3604	3606	3608
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H1H18163B	3626	3628	3630	3632	3634	3636	3638	3640
H1H18164B	3642	3644	3646	3648	3650	3652	3654	3656
H1H18165B	3658	3660	3662	3664	3666	3668	3670	3672
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H1H18167B	3690	3692	3694	3696	3698	3700	3702	3704
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H1H18170B	3738	3740	3742	3744	3746	3748	3750	3752
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H1H18173B	3786	3788	3790	3792	3794	3796	3798	3800
H1H18174B	3802	3804	3806	3808	3810	3812	3814	3816
H1H18175B	3818	3820	3822	3824	3826	3828	3830	3832
H1H18176B	3834	3836	3838	3840	3842	3844	3846	3848
H1H18177B	3850	3852	3854	3856	3858	3860	3862	3864
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H1H18179B	3882	3884	3886	3888	3890	3892	3894	3896
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H1H18188B	4026	4028	4030	4032	4034	4036	4038	4040
H1H18189B	4042	4044	4046	4048	4050	4052	4054	4056
H1H18190B	4058	4060	4062	4064	4066	4068	4070	4072
H1H18191B	4074	4076	4078	4080	4082	4084	4086	4088
H1H18192B	4090	4092	4094	4096	4098	4100	4102	4104
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H1H18197B	4170	4172	4174	4176	4178	4180	4182	4184
H1H18198B	4186	4188	4190	4192	4194	4196	4198	4200
H1H18199B	4202	4204	4206	4208	4210	4212	4214	4216
H1H18200B	4218	4220	4222	4224	4226	4228	4230	4232
H1H18201B	4234	4236	4238	4240	4242	4244	4246	4248
H1H18202B	4250	4252	4254	4256	4258	4260	4262	4264
H1H18203B	4266	4268	4270	4272	4274	4276	4278	4280
H1H18204B	4282	4284	4286	4288	4290	4292	4294	4296
H1H18205B	4298	4300	4302	4304	4306	4308	4310	4312
H1H18206B	4314	4316	4318	4320	4322	4324	4326	4328
H1H18207B	4330	4332	4334	4336	4338	4340	4342	4344
H1H18208B	4346	4348	4350	4352	4354	4356	4358	4360
H1H18209B	4362	4364	4366	4368	4370	4372	4374	4376
H1H18210B	4378	4380	4382	4384	4386	4388	4390	4392
H1H18211B	4394	4396	4398	4400	4402	4404	4406	4408
H1H18212B	4410	4412	4414	4416	4418	4420	4422	4424
H1H18213B	4426	4428	4430	4432	4434	4436	4438	4440
H1H18214B	4442	4444	4446	4448	4450	4452	4454	4456
H1H18216B	4458	4460	4462	4464	4466	4468	4470	4472
H1H18217B	4474	4476	4478	4480	4482	4484	4486	4488
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H1H18219B	4506	4508	4510	4512	4514	4516	4518	4520
H1H18220B	4522	4524	4526	4528	4530	4532	4534	4536
H1H18221B	4538	4540	4542	4544	4546	4548	4550	4552
H1H18222B	4554	4556	4558	4560	4562	4564	4566	4568
H1H18223B	4570	4572	4574	4576	4578	4580	4582	4584
H1H18224B	4586	4588	4590	4592	4594	4596	4598	4600
H1H18225B	4602	4604	4606	4608	4610	4612	4614	4616
H1H18226B	4618	4620	4622	4624	4626	4628	4630	4632
H1H18227B	4634	4636	4638	4640	4642	4644	4646	4648
H1H18228B	4650	4652	4654	4656	4658	4660	4662	4664
H1H18229B	4666	4668	4670	4672	4674	4676	4678	4680
H1H18230B	4682	4684	4686	4688	4690	4692	4694	4696
H1H18231B	4698	4700	4702	4704	4706	4708	4710	4712

H1H18232B	4714	4716	4718	4720	4722	4724	4726	4728
H1H18233B	4730	4732	4734	4736	4738	4740	4742	4744
H1H18234B	4746	4748	4750	4752	4754	4756	4758	4760
H1H18235B	4762	4764	4766	4768	4770	4772	4774	4776
H1H18236B	4778	4780	4782	4784	4786	4788	4790	4792
H1H18237B	4794	4796	4798	4800	4802	4804	4806	4808
H1H18238B	4810	4812	4814	4816	4818	4820	4822	4824
H1H18239B	4826	4828	4830	4832	4834	4836	4838	4840
H1H18240B	4842	4844	4846	4848	4850	4852	4854	4856
H1H18241B	4858	4860	4862	4864	4866	4868	4870	4872
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H1H18243B	4890	4892	4894	4896	4898	4900	4902	4904
H1H18244B	4906	4908	4910	4912	4914	4916	4918	4920
H1H18245B	4922	4924	4926	4928	4930	4932	4934	4936
H1H18246B	4938	4940	4942	4944	4946	4948	4950	4952
H1H18247B	4954	4956	4958	4960	4962	4964	4966	4968
H1H18248B	4970	4972	4974	4976	4978	4980	4982	4984
H1H18249B	4986	4988	4990	4992	4994	4996	4998	5000
H1H18250B	5002	5004	5006	5008	5010	5012	5014	5016
H1H18251B	5018	5020	5022	5024	5026	5028	5030	5032
H1H18252B	5034	5036	5038	5040	5042	5044	5046	5048
H1H18253B	5050	5052	5054	5056	5058	5060	5062	5064
H1H18254B	5066	5068	5070	5072	5074	5076	5078	5080
H1H18255B	5082	5084	5086	5088	5090	5092	5094	5096
H1H18256B	5098	5100	5102	5104	5106	5108	5110	5112
H1H18257B	5114	5116	5118	5120	5122	5124	5126	5128
H1H18258B	5130	5132	5134	5136	5138	5140	5142	5144
H1H18259B	5146	5148	5150	5152	5154	5156	5158	5160
H1H18261B	5162	5164	5166	5168	5170	5172	5174	5176
H1H18262B	5178	5180	5182	5184	5186	5188	5190	5192
H1H18263B	5194	5196	5198	5200	5202	5204	5206	5208
H1H18264B	5210	5212	5214	5216	5218	5220	5222	5224
H1H18265B	5226	5228	5230	5232	5234	5236	5238	5240
H1H18266B	5242	5244	5246	5248	5250	5252	5254	5256
H1H18267B	5258	5260	5262	5264	5266	5268	5270	5272
H1H18268B	5274	5276	5278	5280	5282	5284	5286	5288
H1H18269B	5290	5292	5294	5296	5298	5300	5302	5304
H1H18270B	5306	5308	5310	5312	5314	5316	5318	5320
H1H18271B	5322	5324	5326	5328	5330	5332	5334	5336
H1H18272B	5338	5340	5342	5344	5346	5348	5350	5352
H1H18274B	5354	5356	5358	5360	5362	5364	5366	5368
H1H18275B	5370	5372	5374	5376	5378	5380	5382	5384
H1H18276B	5386	5388	5390	5392	5394	5396	5398	5400
H1H18277B	5402	5404	5406	5408	5410	5412	5414	5416
H1H18278B	5418	5420	5422	5424	5426	5428	5430	5432
H1H18279B	5434	5436	5438	5440	5442	5444	5446	5448
H1H18280B	5450	5452	5454	5456	5458	5460	5462	5464

H1H18281B	5466	5468	5470	5472	5474	5476	5478	5480
H1H18282B	5482	5484	5486	5488	5490	5492	5494	5496
H1H18283B	5498	5500	5502	5504	5506	5508	5510	5512
H1H18284B	5514	5516	5518	5520	5522	5524	5526	5528
H1H18285B	5530	5532	5534	5536	5538	5540	5542	5544
H1H18286B	5546	5548	5550	5552	5554	5556	5558	5560
H1H18287B	5562	5564	5566	5568	5570	5572	5574	5576
H1H18288B	5578	5580	5582	5584	5586	5588	5590	5592
H1H18289B	5594	5596	5598	5600	5602	5604	5606	5608
H1H18290B	5610	5612	5614	5616	5618	5620	5622	5624
H1H18291B	5626	5628	5630	5632	5634	5636	5638	5640
H1H18292B	5642	5644	5646	5648	5650	5652	5654	5656
H1H18293B	5658	5660	5662	5664	5666	5668	5670	5672
H1H18294B	5674	5676	5678	5680	5682	5684	5686	5688
H1H18295B	5690	5692	5694	5696	5698	5700	5702	5704
H1H18297B	5706	5708	5710	5712	5714	5716	5718	5720
H1H18298B	5722	5724	5726	5728	5730	5732	5734	5736
H1H18299B	5738	5740	5742	5744	5746	5748	5750	5752
H1H18300B	5754	5756	5758	5760	5762	5764	5766	5768
H1H18301B	5770	5772	5774	5776	5778	5780	5782	5784
H1H18302B	5786	5788	5790	5792	5794	5796	5798	5800
H1H18303B	5802	5804	5806	5808	5810	5812	5814	5816
H1H18304B	5818	5820	5822	5824	5826	5828	5830	5832
H1H18305B	5834	5836	5838	5840	5842	5844	5846	5848
H1H18306B	5850	5852	5854	5856	5858	5860	5862	5864
H1H18307B	5866	5868	5870	5872	5874	5876	5878	5880
H1H18308B	5882	5884	5886	5888	5890	5892	5894	5896
H1H18309B	5898	5900	5902	5904	5906	5908	5910	5912
H1H18310B	5914	5916	5918	5920	5922	5924	5926	5928
H1H18311B	5930	5932	5934	5936	5938	5940	5942	5944
H1H18312B	5946	5948	5950	5952	5954	5956	5958	5960
H1H18313B	5962	5964	5966	5968	5970	5972	5974	5976
H1H18314B	5978	5980	5982	5984	5986	5988	5990	5992
H1H18315B	5994	5996	5998	6000	6002	6004	6006	6008
H1H18316B	6010	6012	6014	6016	6018	6020	6022	6024
H1H18317B	6026	6028	6030	6032	6034	6036	6038	6040
H1H18318B	6042	6044	6046	6048	6050	6052	6054	6056
H1H18319B	6058	6060	6062	6064	6066	6068	6070	6072
H1H18320B	6074	6076	6078	6080	6082	6084	6086	6088
H1H18321B	6090	6092	6094	6096	6098	6100	6102	6104
H1H18322B	6106	6108	6110	6112	6114	6116	6118	6120
H1H18323B	6122	6124	6126	6128	6130	6132	6134	6136
H1H18324B	6138	6140	6142	6144	6146	6148	6150	6152
H1H18325B	6154	6156	6158	6160	6162	6164	6166	6168
H1H18326B	6170	6172	6174	6176	6178	6180	6182	6184
H1H18327B	6186	6188	6190	6192	6194	6196	6198	6200
H1H18328B	6202	6204	6206	6208	6210	6212	6214	6216

H1H18329B	6218	6220	6222	6224	6226	6228	6230	6232
H1H18330B	6234	6236	6238	6240	6242	6244	6246	6248
H1H18331B	6250	6252	6254	6256	6258	6260	6262	6264
H1H18332B	6266	6268	6270	6272	6274	6276	6278	6280
H1H18333B	6282	6284	6286	6288	6290	6292	6294	6296
H1H18334B	6298	6300	6302	6304	6306	6308	6310	6312
H1H18335B	6314	6316	6318	6320	6322	6324	6326	6328

Table 13 Nucleic Acid Sequence Identifiers

Antibody Designation	Nucleic Acid SEQ ID NOs:							
	HCVR	HCDR1	HCDR2	HCDR3	LCVR	LCDR1	LCDR2	LCDR3
H1H17952B	297	299	301	303	305	307	309	311
H1H17953B	313	315	317	319	321	323	325	327
H1H17954B	329	331	333	335	337	339	341	343
H1H17955B	345	347	349	351	353	355	357	359
H1H17956B	361	363	365	367	369	371	373	375
H1H17957B	377	379	381	383	385	387	389	391
H1H17958B	393	395	397	399	401	403	405	407
H1H17959B	409	411	413	415	417	419	421	423
H1H17960B	425	427	429	431	433	435	437	439
H1H17961B	441	443	445	447	449	451	453	455
H1H17962B	457	459	461	463	465	467	469	471
H1H17963B	473	475	477	479	481	483	485	487
H1H17964B	489	491	493	495	497	499	501	503
H1H17965B	505	507	509	511	513	515	517	519
H1H17966B	521	523	525	527	529	531	533	535
H1H17967B	537	539	541	543	545	547	549	551
H1H17968B	553	555	557	559	561	563	565	567
H1H17969B	569	571	573	575	577	579	581	583
H1H17970B	585	587	589	591	593	595	597	599
H1H17971B	601	603	605	607	609	611	613	615
H1H17972B	617	619	621	623	625	627	629	631
H1H17973B	633	635	637	639	641	643	645	647
H1H17974B	649	651	653	655	657	659	661	663
H1H17975B	665	667	669	671	673	675	677	679
H1H17976B	681	683	685	687	689	691	693	695
H1H17977B	697	699	701	703	705	707	709	711
H1H17978B	713	715	717	719	721	723	725	727
H1H17979B	729	731	733	735	737	739	741	743
H1H17980B	745	747	749	751	753	755	757	759
H1H17981B	761	763	765	767	769	771	773	775
H1H17982B	777	779	781	783	785	787	789	791
H1H17983B	793	795	797	799	801	803	805	807
H1H17984B	809	811	813	815	817	819	821	823

H1H17985B	825	827	829	831	833	835	837	839
H1H17986B	841	843	845	847	849	851	853	855
H1H17987B	857	859	861	863	865	867	869	871
H1H17988B	873	875	877	879	881	883	885	887
H1H17989B	889	891	893	895	897	899	901	903
H1H17990B	905	907	909	911	913	915	917	919
H1H17991B	921	923	925	927	929	931	933	935
H1H17992B	937	939	941	943	945	947	949	951
H1H17993B	953	955	957	959	961	963	965	967
H1H17994B	969	971	973	975	977	979	981	983
H1H17995B	985	987	989	991	993	995	997	999
H1H17996B	1001	1003	1005	1007	1009	1011	1013	1015
H1H17997B	1017	1019	1021	1023	1025	1027	1029	1031
H1H17998B	1033	1035	1037	1039	1041	1043	1045	1047
H1H17999B	1049	1051	1053	1055	1057	1059	1061	1063
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H1H18002B	1097	1099	1101	1103	1105	1107	1109	1111
H1H18003B	1113	1115	1117	1119	1121	1123	1125	1127
H1H18004B	1129	1131	1133	1135	1137	1139	1141	1143
H1H18005B	1145	1147	1149	1151	1153	1155	1157	1159
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H1H18266B	5241	5243	5245	5247	5249	5251	5253	5255
H1H18267B	5257	5259	5261	5263	5265	5267	5269	5271
H1H18268B	5273	5275	5277	5279	5281	5283	5285	5287
H1H18269B	5289	5291	5293	5295	5297	5299	5301	5303
H1H18270B	5305	5307	5309	5311	5313	5315	5317	5319
H1H18271B	5321	5323	5325	5327	5329	5331	5333	5335

H1H18272B	5337	5339	5341	5343	5345	5347	5349	5351
H1H18274B	5353	5355	5357	5359	5361	5363	5365	5367
H1H18275B	5369	5371	5373	5375	5377	5379	5381	5383
H1H18276B	5385	5387	5389	5391	5393	5395	5397	5399
H1H18277B	5401	5403	5405	5407	5409	5411	5413	5415
H1H18278B	5417	5419	5421	5423	5425	5427	5429	5431
H1H18279B	5433	5435	5437	5439	5441	5443	5445	5447
H1H18280B	5449	5451	5453	5455	5457	5459	5461	5463
H1H18281B	5465	5467	5469	5471	5473	5475	5477	5479
H1H18282B	5481	5483	5485	5487	5489	5491	5493	5495
H1H18283B	5497	5499	5501	5503	5505	5507	5509	5511
H1H18284B	5513	5515	5517	5519	5521	5523	5525	5527
H1H18285B	5529	5531	5533	5535	5537	5539	5541	5543
H1H18286B	5545	5547	5549	5551	5553	5555	5557	5559
H1H18287B	5561	5563	5565	5567	5569	5571	5573	5575
H1H18288B	5577	5579	5581	5583	5585	5587	5589	5591
H1H18289B	5593	5595	5597	5599	5601	5603	5605	5607
H1H18290B	5609	5611	5613	5615	5617	5619	5621	5623
H1H18291B	5625	5627	5629	5631	5633	5635	5637	5639
H1H18292B	5641	5643	5645	5647	5649	5651	5653	5655
H1H18293B	5657	5659	5661	5663	5665	5667	5669	5671
H1H18294B	5673	5675	5677	5679	5681	5683	5685	5687
H1H18295B	5689	5691	5693	5695	5697	5699	5701	5703
H1H18297B	5705	5707	5709	5711	5713	5715	5717	5719
H1H18298B	5721	5723	5725	5727	5729	5731	5733	5735
H1H18299B	5737	5739	5741	5743	5745	5747	5749	5751
H1H18300B	5753	5755	5757	5759	5761	5763	5765	5767
H1H18301B	5769	5771	5773	5775	5777	5779	5781	5783
H1H18302B	5785	5787	5789	5791	5793	5795	5797	5799
H1H18303B	5801	5803	5805	5807	5809	5811	5813	5815
H1H18304B	5817	5819	5821	5823	5825	5827	5829	5831
H1H18305B	5833	5835	5837	5839	5841	5843	5845	5847
H1H18306B	5849	5851	5853	5855	5857	5859	5861	5863
H1H18307B	5865	5867	5869	5871	5873	5875	5877	5879
H1H18308B	5881	5883	5885	5887	5889	5891	5893	5895
H1H18309B	5897	5899	5901	5903	5905	5907	5909	5911
H1H18310B	5913	5915	5917	5919	5921	5923	5925	5927
H1H18311B	5929	5931	5933	5935	5937	5939	5941	5943
H1H18312B	5945	5947	5949	5951	5953	5955	5957	5959
H1H18313B	5961	5963	5965	5967	5969	5971	5973	5975
H1H18314B	5977	5979	5981	5983	5985	5987	5989	5991
H1H18315B	5993	5995	5997	5999	6001	6003	6005	6007
H1H18316B	6009	6011	6013	6015	6017	6019	6021	6023
H1H18317B	6025	6027	6029	6031	6033	6035	6037	6039
H1H18318B	6041	6043	6045	6047	6049	6051	6053	6055
H1H18319B	6057	6059	6061	6063	6065	6067	6069	6071
H1H18320B	6073	6075	6077	6079	6081	6083	6085	6087

H1H18321B	6089	6091	6093	6095	6097	6099	6101	6103
H1H18322B	6105	6107	6109	6111	6113	6115	6117	6119
H1H18323B	6121	6123	6125	6127	6129	6131	6133	6135
H1H18324B	6137	6139	6141	6143	6145	6147	6149	6151
H1H18325B	6153	6155	6157	6159	6161	6163	6165	6167
H1H18326B	6169	6171	6173	6175	6177	6179	6181	6183
H1H18327B	6185	6187	6189	6191	6193	6195	6197	6199
H1H18328B	6201	6203	6205	6207	6209	6211	6213	6215
H1H18329B	6217	6219	6221	6223	6225	6227	6229	6231
H1H18330B	6233	6235	6237	6239	6241	6243	6245	6247
H1H18331B	6249	6251	6253	6255	6257	6259	6261	6263
H1H18332B	6265	6267	6269	6271	6273	6275	6277	6279
H1H18333B	6281	6283	6285	6287	6289	6291	6293	6295
H1H18334B	6297	6299	6301	6303	6305	6307	6309	6311
H1H18335B	6313	6315	6317	6319	6321	6323	6325	6327

Example 9. H1H11729P has a Superior Pharmacokinetic Profile in Mice and Non-human Primates (NHPs) Compared to a Comparator Antibody (Control I mAb)

[0234] A study was conducted to compare the pharmacokinetic profile of H1H11729P with a comparator antibody in mice and in non-human primates. One anti-influenza HA comparator antibody designated herein as Control I mAb, is an anti-influenza HA antibody with heavy (HC) and light chain (LC) amino acid sequences as set forth in WO2008/028946 as SEQ ID NO: 65 (HC) and SEQ ID NO: 91 (LC) and is also referred to in WO2008/028946 as CR6261.

[0235] For both the mouse and NHP PK experiments, circulating drug levels were determined by total human antibody analysis using an ELISA immunoassay. Briefly, a goat anti-human IgG polyclonal antibody was coated onto 96-well plates to capture the tested human antibodies in the sera, and then plate bound antibodies were detected using a goat anti-human IgG polyclonal antibody conjugated with horseradish and TMB substrate. The serum samples were in six-dose serial dilutions and reference standards of the respective antibodies in 12-dose serial dilutions. Drug antibody concentrations in the sera were calculated based on the reference standard curve generated using Graphpad Prism software.

A. Mouse Study

[0236] The pharmacokinetic assessment of H1H11729P was conducted in wild-type (WT) C57BL/6 mice. H1H11729P and Control I mAb were administered SC in separated groups of 5 mice each at a dose of 1 mg/kg. Bleeds were collected post-injection at 6h, 1, 2, 3, 4, 7, 11, 14, 22, and 30 days in addition to the bleed collected one day prior to the antibody injection (pre-bleed). Serum fractions from the bleeds were separated and frozen at -80 °C until analysis was conducted.

Results:

[0237] As shown in Table 14, the half-life of H1H11729P in WT mice was 11.1 days, while the half-life of Control I mAb was 5.67 days, demonstrating an advantageous property of the H1H11729P over the comparator antibody.

Table 14: Summary of the Pharmacokinetic Profiles of H1H11729P and Control I mAb in Uninfected Wild-Type Mice

Drug	T _{1/2} (d)	C _{max} (μ/mL)	AUC(d* μ /mL)
H1H11729P	11.1 ± 0.8	12.0 ± 0.9	147 ± 3.4
Control I mAb	5.67 ± 0.6	10.4 ± 0.6	93.8 ± 4.8

B. Cynomolgus NHPs

[0238] Assessment of the pharmacokinetic clearance rate of H1H11729P and Control I mAb was conducted in 3-5 year old, female cynomolgus monkeys. Monkeys were prescreened for influenza A and B antibodies and were found to be negative. The antibody was tested in three animals and was administered subcutaneously at a dose of 3 mg/kg (at a volume of 2 mL/kg and 1.5 mg/mL). Blood samples were collected prior to dosing and post-injection at 1h, 4h, 8h, and on Days 1, 2, 3, 4, 5, 10, 14, 18, 21, 24, 28, 35, 42, 49, 56, 63, 70, 84, and 98. Serum was separated from whole blood and frozen at -80 °C until analysis. Circulating drug concentrations were determined by total human antibody analysis using an ELISA immunoassay. Drug antibody concentrations in the sera were calculated based on the reference standard curve generated using Graphpad Prism software.

Results:

[0239] As shown in Table 15, the half-life of H1H11729P in monkeys was 13.4 days, while the half-life of Control I mAb was 8.05 days, demonstrating an advantageous property of the H1H11729P over the comparator antibody.

Table 15: Summary of the Pharmacokinetic Profiles of H1H11729P and Control I mAb in Cynomolgus Monkeys

Drug	T _{1/2} (d)	C _{max} (μg/mL)	AUC(d* μg /mL)
H1H11729P	13.4 ± 1.46	41.3 ± 7.84	863 ± 145
Control I mAb	8.05 ± 2.17	24.9 ± 1.87	320 ± 70.1

Example 10: H1H11729P Effectively Treats Lethal Influenza Virus Infection in Mice

[0240] There is a substantial unmet need for improved standard of care therapies for treating or preventing of influenza virus infections in humans. Currently, only two classes of drugs are available: the adamantanes and the neuraminidase inhibitors (NAIs). Adamantanes (amantadine and rimantadine) have been associated with the rapid emergence of drug-resistant strains and are no longer recommended for treatment of influenza. NAIs like oseltamivir

(TAMIFLU®) are the front line drugs for treatment and prophylaxis of influenza, however, their window of efficacy is limited: NAIs have been shown to reduce the duration of fever and illness symptoms by about one day in the therapeutic setting if the antiviral is administered within 48 hours of symptom onset with little clinical evidence for efficacy if administered after 48 hours.

[0241] To evaluate the *in vivo* efficacy of H1H1729P in the treatment of severe influenza, experiments were conducted with the following objectives:

[0242] Study 1: To evaluate the efficacy of a single dose of H1H1729P versus the clinical standard of care oseltamivir (TAMIFLU®) given twice per day for 5 days.

[0243] Study 2: To determine the efficacy of H1H1729P administered in combination with oseltamivir.

[0244] The strain used in these studies included a historical [A/Puerto Rico/08/1934 (H1N1)] influenza A virus group 1 isolate. All experiments were performed in 6-week-old wild-type (BALB/c) female mice. Mice were challenged with 10 x mouse LD₅₀ (MLD₅₀) equivalent to 800 plaque-forming units (PFUs) of A/Puerto Rico/08/1934 (H1N1). In the treatment models, mice were challenged intranasally (IN) on day 0 post-infection (p.i.) and fixed doses of mAb were given intravenously (IV) on specific days post-infection (e.g., day 1, 2, 3, 4 or 5). Oseltamivir was resuspended according to the manufacturers instructions and mice were dosed every 12 h (i.e., twice per day; BID) via oral gavage for 5 days, with the first dose administered on day 2 or 3 post-infection. Mice were weighed and observed daily up to day 14 p.i. and were sacrificed when they lost 20% of their starting weight. Results are reported as percent survival.

[0245] In the first experiment the efficacy of either a single dose of H1H1729P (15 mg/kg) or 25 or 5 mg/kg of oseltamivir BID (5 day regimen) initiating 48 hours post-infection (*i.e.* administered therapeutically) was studied to evaluate the effect in the murine model at this timepoint. All mice receiving 5 mg/kg oseltamivir died by day 6, while the dose of 25 mg/kg improved survival to 40%; in contrast, all mice receiving H1H1729P survived (Figure 1 and Table 16).

[0246] In the next experiment, H1H1729P was tested in combination with oseltamivir, beginning 72 hours post-infection. While only approximately 20% of mice treated with 25 mg/kg oseltamivir survived, single doses of H1H1729P at 15 mg/kg and 7 mg/kg resulted in 60% and 36% survival, respectively. An additive effect was observed when the treatments were combined: A single dose of 7 mg/kg H1H1729P combined with a regimen of oseltamivir showed 60% survival and a single dose of 15 mg/kg H1H1729P in combination with a regimen of oseltamivir resulted in 87% survival (Figure 2 and Table 17).

[0247] In summary, H1H1729P displayed robust efficacy in treating mice infected with a severe historical influenza strain, and in fact, demonstrated greater efficacy than oseltamivir at 48 hours post infection. Furthermore, H1H1729P demonstrated additive efficacy when administered in combination with oseltamivir.

Table 16: Results of Study 1: A Single Dose of H1H11729P at 48 hrs p.i. Demonstrates Greater Efficacy than oseltamivir at 48 hrs p.i. in Treating Severe Influenza A Virus Infection in Mice

PID	Number of mice per group	Percent survival (no. of surviving mice/total no. of mice in the group)
Oral gavage control (uninfected)	5	100 (5/5)
H1H11729P	5	100 (5/5)
Oseltamivir (25 mg/kg BID x 5d)	5	40 (2/5)
Oseltamivir (5 mg/kg BID x 5d)	5	0 (0/5)
hlgG1 negative isotype control	5	0 (0/5)
Oral gavage control (infected)	5	0 (0/5)

Table 17: Results of Study 2: Additive Efficacy is Observed when a Single Dose of H1H11729P is Combined with oseltamivir at 72 hrs p.i. to Treat Severe Influenza in Mice

PID	Number of mice per group	Percent survival (no. of surviving mice/total no. of mice in the group)
H1H11729P (15 mg/kg)	15	60 (9/15)
H1H11729P (7 mg/kg)	15	35.7 (5/15)
H1H11729P (15 mg/kg) + Oseltamivir (25 mg/kg BID x 5d)	15	86.7 (13/15)
H1H11729P (7 mg/kg) + Oseltamivir (25 mg/kg BID x 5d)	15	60 (9/15)
Oseltamivir (25 mg/kg BID x 5 d)	15	20 (3/15)
hlgG1 negative isotype control	15	0 (0/15)
Oral gavage control (infected)	15	0 (0/15)

What is claimed is:

1. An isolated antibody or antigen-binding fragment thereof, comprising three heavy chain complementarity determining regions (CDRs) (HCDR1, HCDR2 and HCDR3) contained within a heavy chain variable region (HCVR) comprising the amino acid sequence set forth in SEQ ID NO: 18; and three light chain complementarity determining regions (CDRs) (LCDR1, LCDR2 and LCDR3) contained within a light chain variable region (LCVR) comprising the amino acid sequence set forth in SEQ ID NO: 26.

2. The isolated antibody or antigen-binding fragment thereof of claim 1, comprising a HCVR that comprises an amino acid sequence set forth in SEQ ID NO:18.

3. The isolated antibody or antigen-binding fragment thereof of claim 1, comprising a LCVR that comprises an amino acid sequence set forth in SEQ ID NO: 26.

4. The isolated antibody or antigen-binding fragment thereof of claim 1, comprising a LCVR that comprises an amino acid sequence set forth in SEQ ID NO: 26; and a HCVR that comprises an amino acid sequence set forth in SEQ ID NO: 18.

5. The isolated antibody or antigen-binding fragment thereof of claim 1, comprising:

(a) a HCDR1 that comprises an amino acid sequence set forth in SEQ ID NO: 20;

(b) a HCDR2 that comprises an amino acid sequence set forth in SEQ ID NO: 22;

(c) a HCDR3 that comprises an amino acid sequence set forth in SEQ ID NO: 24;

(d) a LCDR1 that comprises an amino acid sequence set forth in SEQ ID NO: 28;

(e) a LCDR2 that comprises an amino acid sequence set forth in SEQ ID NO: 30; and

(f) a LCDR3 that comprises an amino acid sequence set forth in SEQ ID NO: 32.

6. The isolated antibody or antigen-binding fragment of claim 4 which is an antibody.

7. The isolated antibody of claim 6 which is a recombinant antibody.

8. The isolated antibody of claim 6 which is bispecific.

9. A pharmaceutical composition comprising the isolated antibody of claim 6 and a pharmaceutically acceptable carrier or diluent.

10. The pharmaceutical composition of claim 9 further comprising a second therapeutic agent.

11. An isolated polynucleotide molecule comprising a polynucleotide sequence that encodes a HCVR or an LVCR or both of the antibody of claim 6.

12. A vector comprising the polynucleotide of claim 11.

13. An isolated cell comprising the vector of claim 12.

14. The pharmaceutical composition of claim 10 wherein the second therapeutic agent is an anti-viral drug.

15. The pharmaceutical composition of claim 10 wherein the second therapeutic agent is a neuraminidase inhibitor or an interferon.

16. The pharmaceutical composition of claim 10 wherein the second therapeutic agent is zidovudine, gangcyclovir, vidarabine, idoxuridine, trifluridine, foscarnet, acyclovir, ribavirin, amantadine, remantidine, saquinavir, indinavir, ritonavir, zanamivir, oseltamivir, laninamivir, peramivir or rimantadine.