HUMAN MONOCLONAL ANTIBODY NEUTRALIZING VASCULAR ENDOTHELIAL GROWTH FACTOR RECEPTOR AND USE THEREOF

TECHNICAL FIELD

The present invention relates to human monoclonal antibodies neutralizing vascular endothelial growth factor receptor and the use thereof, and more particularly to human ScFv molecules neutralizing vascular endothelial growth factor receptor, and a composition for inhibiting angiogenesis and a composition for treating cancer, which contain the human ScFv molecules.

BACKGROUND ART

Angiogenesis means the formation of new blood vessels from pre-existing vessels by the growth, differentiation and migration of endothelial cells and does not occur in healthy adults, except for some special occasions, including wound healing, menstruation, etc. However, the excessive formation of new blood vessels in diseases, such as tumor growth and metastasis, age-related macular degeneration, rheumatoid arthritis, diabetic retinopathy, psoriasis and chronic inflammation, has been reported (Cameliet and Jain, *Nature*, 407:249, 2000). For this reason, many efforts to treat diseases, particularly tumors, using angiogenesis inhibitors, have been made.

Factors involved in angiogenesis include vascular endothelial growth factor (VEGF), epithelial growth factor (EGF), platelet-derived growth factor (PDGF), transforming growth factor-b (TGFb), fibroblast growth factor (FGF), etc. Among them, the vascular endothelial growth factor is an endothelial cell-specific factor which is

involved directly in the growth, differentiation and migration of endothelial cells, and there are four different isoforms (VEGF165, VEGF121, VEGF189 and VEGF206). Among the four isoforms, VEGF165 is the most abundant isoform in all human tissues except placenta (Tisher *et al.*, *J. Biol. Chem.*, 266:11947, 1991).

Vascular endothelial growth factor (VEGF) regulates new blood vessel formation resulting from the differentiation of endothelial precursors (angioblasts) *in situ*, is expressed in embryonic tissues (Breier *et al.*, *Development (Camb)*, 114:521, 1992), macrophages, and proliferating epithelial keratinocytes during wound healing (Brown *et al.*, *J. Exp. Med.*, 176:1375, 1992), and may be responsible for tissue edema associated with inflammation (Ferrara *et al.*, *Endocr. Rev.*, 13:18, 1992). *In situ* hybridization studies have demonstrated high VEGF expression in a number of human tumor lines including glioblastoma multiforme, hemangioblastoma, central nervous system neoplasms and AIDS-associated Kaposi's sarcoma (Plate *et al.*, *Nature,* 359:845, 1992; Plate *et al.*, *Cancer Res.*, 53: 5822, 1993; Berkman *et al.*, *J. Clin. Invest.*, 91:153, 1993; Nakamura *et al.*, *AIDS Weekly,* 13(1), 1992). High levels of VEGF were also observed in hypoxia induced angiogenesis (Shweiki *et al.*, *Nature,* 359:843, 1992).

The biological function of VEGF is mediated through its high affinity VEGF receptors which are selectively expressed in endothelial cells during embryogenesis (Millauer *et al.*, *Cell*, 72:835, 1993) and during tumor formation. VEGF receptors (VEGFR) typically are class III receptor-type tyrosine kinases characterized by having several, typically 5 or 7, immunoglobulin-like loops in their amino-terminal extracellular ligand-binding domain of a receptor (Kaipainen *et al.*, *J. Exp. Med.*, 178:2027, 1993). The other two regions include a transmembrane region and a carboxy-terminal intracellular catalytic domain interrupted by an insertion of hydrophilic interkinase sequences of variable lengths, called the kinase insert domain (Terman *et al.*, *Oncogene*, 6:1677, 1991). VEGF receptors include fms-like tyrosine kinase receptor (Flt-1), or VEGFR-1 (Shibuya *et al.*, *Oncogene*, 5:519,

1990; WO 92/14248; Terman *et al.*, *Oncogene*, 6:1677, 1991), kinase insert domain-containing receptor/fetal liver kinase (KDR/Flk-1), or VEGFR-2 (Matthews *et al., PNAS*, 88:9026, 1991), although other receptors such as neuropilin-1 and neuropilin-2 can also bind VEGF. Another tyrosine kinase receptor, VEGFR-3 (Flt-4), binds the VEGF homologues VEGF-C and VEGF-D and is important in the development of lymphatic vessels.

High levels of Flk-1 are expressed by endothelial cells that infiltrate gliomas (Plate *et al., Nature,* 359:845, 1992). Flk-1 levels are specifically upregulated by VEGF produced by human glioblastomas (Plate *et al., Cancer Res.*, 53:5822, 1993).

The finding of high levels of Flk-1 expression in glioblastoma associated endothelial cells (GAEC) indicates that receptor activity is probably induced during tumor formation since Flk-1 transcripts are barely detectable in normal brain endothelial cells. This upregulation is confined to the vascular endothelial cells in close proximity to the tumor. Blocking VEGF activity with neutralizing anti-VEGF monoclonal antibodies (mAbs) resulted in inhibition of the growth of human tumor xenografts in nude mice (Kim, K. *et al.*, *Nature*, 362:841-844, 1993), indicating a direct role for VEGF in tumor-related angiogenesis.

Although VEGF ligands are upregulated in tumor cells, and the receptors thereof are upregulated in tumor infiltrated vascular endothelial cells, the expression levels of VEGF ligands and the receptors thereof are low in normal cells that are not associated with angiogenesis. Therefore, such normal cells would block the interaction between VEGF and the receptors thereof to inhibit angiogenesis, thus inhibiting tumor growth.

High levels of VEGFR-2 are expressed by endothelial cells that infiltrate gliomas, and are specifically upregulated by VEGF produced by human glioblastomas (Plate *et al.*, *Nature,* 359:845, 1992; Plate *et al.*, *Cancer Res.*, 53:5822, 1993). The

finding of high levels of VEGR-2 expression in glioblastoma associated endothelial cells (GAEC) suggests that receptor activity is induced during tumor formation, since VEGFR-2 transcripts are barely detectable in normal brain endothelial cells.

Therefore, studies focused on inhibiting the activity of VEGF, which is expressed in tumor growth sites, to inhibit angiogenesis so as to inhibit tumor growth, are being actively conducted. Typically, methods of inhibiting VEGF receptors on the membrane of cancer cells to prevent VEGF from entering cells have been developed. Examples of cell lines producing VEGFR antibodies include a hybridoma cell line producing rat anti-mouse VEGFR-2 monoclonal antibody (DC101; ATCC HB 11534), a hybridoma cell line (M25, 18A1; ATCC HB 12152) producing mouse anti-mouse VEGFR-2 monoclonal antibody mAb 25, and a hybridoma cell line producing mouse anti-mouse VEGFR-2 monoclonal antibody mAb 73 [(M73,24; ATCC HB 12153), KM1730(FERM BP-5697; WO 98/22616; WO 99/59636), KM1731 (FERM BP-5718), KM1732 (FERM BP-5698), KM1748 (FERM BP-5699), KM1750 (FERM BP-5700)].

There has been a continuous development of humanized antibodies against VEGF receptors. These humanized antibodies against VEGF receptors, developed to date, showed high competition with VEGF *in vitro*, but had problems in that their ability to neutralize VEGF receptors in cells is reduced and in that the antibodies do not show cross-reactivity in mice or rats, such that animal tests cannot be carried out.

Accordingly, the present inventors have constructed a library of non-immunized fully human antibodies, screened single chain variable fragment (ScFv) antibodies against VEGF receptor (KDR), and found that the antibodies exhibit an excellent KDR-neutralizing effect not only *in vitro*, but also in cells and *in vivo*, and show cross-reactivity even in mice and rats, thereby completing the present invention.

SUMMARY OF THE INVENTION

It is an object of the present invention to provide a fully human single chain variable fragment (ScFv) antibodies 6A6-ScFv and 6A6-IgG, which have an excellent ability to neutralize VEGF receptor in cells and *in vivo*.

Another object of the present invention is to provide a fully human single chain variable fragment (ScFv), which is a light chain variant of 6A6, which shows a more excellent ability to neutralize VEGF receptor compared to that of 6A6-ScFv.

Still another object of the present invention is to provide a composition for inhibiting angiogenesis, which contains a fully human ScFv or IgG having the ability to neutralize VEGF receptor.

Yet another object of the present invention is to provide a composition for treating cancer, which contains a fully human ScFv or IgG having the ability to neutralize VEGF receptor.

To achieve the above objects, in one aspect, the present invention provides a single chain variable fragment (ScFv) molecule, which contains a light chain variable region represented by an amino acid sequence of any one of SEQ ID NOS: 1 to 19 and functions to neutralize vascular endothelial growth factor receptor. In the present invention, the ScFv (single chain variable fragment) molecule and a construct thereof preferably have a heavy chain variable region represented by an amino acid sequence of SEQ ID NO: 20.

In another aspect, the present invention provides a DNA encoding said ScFv (single chain variable fragment) molecule, a vector containing said DNA, and recombinant cells transformed with said vector. In the present invention, the cells are preferably bacterial or animal cells.

In still another aspect, the present invention provides a composition for inhibiting angiogenesis, which contains said ScFv molecule, and a composition for treating cancer, which contains said ScFv molecule.

In still another aspect, the present invention provides an IgG, which contains a light chain variable region represented by an amino acid sequence of any one of SEQ ID NOS: 1 to 19 and functions to neutralize vascular endothelial growth factor receptor. In the present invention, said IgG preferably has a heavy chain variable region represented by an amino acid sequence of SEQ ID NO: 20.

In yet another aspect, the present invention provides a composition for inhibiting angiogenesis, which contains said IgG, and a composition for treating cancer, which contains said IgG.

Other features and aspects of the present invention will be apparent from the following detailed description and the appended claims.

BRIEF DESCRIPTION OF DRAWINGS

FIG. 1 shows the amino acid sequence and function of a gene inserted into a pCDNA3-KDR D123tFcm vector.

FIG. 2 shows a schematic diagram of KDR(ECD1-2) and KDR(ECD2-3)-Fc for epitope mapping according to the present invention.

FIG. 3 shows the results of SDS-PAGE of KDR(ECD1-3)-Fc purified in the present invention.

FIG. 4 shows the results of VEGF competition assays for anti-KDR phage and anti-KDR-SvFc according to the present invention.

FIG. 5 shows the nucleic acid sequence, amino acid sequence and CDR sequence of 6A6 ScFv phage according to the present invention.

FIG. 6 shows SDS-PAGE results for purified 6A6 ScFv.

FIG. 7 shows the results of VEGF competition assays using anti-KDR-ScFv.

FIG. 8 shows the results of epitope mapping of anti-KDR-ScFv according to the present invention.

FIG. 9 shows a cleavage map of pIGHD-6A6Hvy that is a vector containing the invariable region and heavy chain region of 6A6 IgG.

FIG. 10 shows a cleavage map of pIgGLD-6A6Lgt that is a vector containing the constant region and light chain region of IgG.

FIG. 11 shows the results of SDS-PAGE of purified 6A6 IgG.

FIG. 12 shows the results of VEGF competition assays using anti-KDR 6A6 IgG.

FIG. 13 shows the results of competition assays of anti-KDR 6A6 IgG with VEGF families.

FIG. 14 shows the results of FACS analysis for the binding affinity of the inventive anti-KDR IgG antibody to HUVEC cells.

FIG. 15 shows FACS assay results for the competition of the inventive anti-KDR IgG antibody with VEGF165 in HUVEC cells.

FIG. 16 shows the results of Western blot analysis for the expression of KDR in K562 cells (ATCC CCL-243).

FIG. 17 shows the binding affinity of the inventive anti-KDR IgG antibody to the K562 cell line.

FIG. 18 shows FACS assay results for the competition of the inventive anti-KDR IgG antibody with VEGF in the K562 cell line.

FIG. 19 shows FACS analysis results for the binding affinity of an anti-KDR antibody to a Gleevec-resistant cell line.

FIG. 20 shows analysis results for the cell proliferation inhibition of the anti-KDR-IgG according to the present invention.

FIG. 21 shows the results of Western blot analysis for the ability of the inventive anti-KDR antibody to inhibit KDR phosphorylation and ERK phosphorylation, which is induced by VEGF.

FIG. 22 shows the ability of an IgG-type KDR antibody to inhibit the migration of endothelial cells by VEGF.

FIG. 23 shows that an IgG-type KDR antibody inhibits endothelial cell tube formation induced by VEGF.

FIG. 24 shows the inhibition of VEGF-KDR internalization through the binding between the IgG-type KDR antibody and cell surface KDR.

FIG. 25 shows the inhibitory effect of the IgG-type KDR antibody on rat aortic ring sprouting induced by VEGF.

FIG. 26 shows analysis results for the inhibitory effect of the IgG-type KDR antibody on angiogenesis induced by VEGF.

FIG. 27 shows analysis results for the inhibitory effect of a 6A6 antibody on tumor growth in the HCT116 cell line in a colon cancer mouse xenograft model.

FIG. 28 is a photograph of tumors excised after treatment with the 6A6 antibody in the colon cancer mouse xenograft model.

FIG. 29 shows analysis results for the inhibitory effect of the 6A6 antibody on tumor growth in the A549 cell line in a lung cancer mouse xenograft model.

FIG. 30 shows the results of labeling of the IgG-type 6A6 antibody with radioactive isotope iodine.

FIG. 31 shows color images of the IgG-type 6A6 antibody labeled with iodine-123 in a mouse tumor model of chronic myelogenous leukemia.

FIG. 32 is a schematic diagram showing the preparation of light-chain shuffling.

FIG. 33 shows the results of VEGF competition assays of anti-KDR antibodies obtained through light chain shuffling.

FIG. 34 shows the results of VEGF competition assays of anti-KDR antibodies obtained through light chain shuffling.

FIG. 35 shows the results of VEGF competition assays of anti-KDR antibodies obtained through light chain shuffling.

DETAILED DESCRIPTION OF THE INVENTION, AND PPRFERRED EMBODIMENTS

In one aspect, the present invention relates to a fully human ScFv (single chain variable fragment) antibody 6A6 (TTAC-0001)-ScFv, which neutralizes vascular endothelial growth factor.

The 6A6-ScFv was screened in the following manner. First, a library of fully human antibodies was constructed, and a cell line expressing a fusion protein composed of each of extracellular domains 1-3 of KDR(VEGFR-2), fused to Fc, was constructed. Anti-KDR-ScFv antibodies neutralizing KDR were screened from the fully human antibody library in the cell line using purified recombinant KDR D1-D3-Fc fusion proteins.

The screened ScFv antibodies were expressed and purified in bacteria with V5 tagging, and human KDR D1-D3-Fc binding assays and VEGF competition assays were performed in a ScFv-phage particle state. Also, BIAcore analysis was carried out to measure the ScFv-antibody affinity, and 6A6-ScFv having constantly maintained affinity was obtained and converted in the form of IgG.

Also, in the present invention, it was confirmed through Western blotting that 6A6-ScFv inhibited the phosphorylation of an angiogenesis signaling factor ERK in primary cultured HUVEC cells and that this inhibition was dependent on the concentration of 6A6-ScFv.

In another aspect, the present invention relates to a fully human antibody 6A6-IgG neutralizing vascular endothelial growth factor receptor.

FACS analysis revealed that the 6A6-IgG was strongly bound to endogenous human KDR, which was expressed on the surface of living HUVEC cells (ATCC, USA), compared to a commercially available IMC-1C11 IgG chimeric antibody (Imclone, USA), and that, even when the cells were also treated with a competitively binding human $VEGF_{165}$, the 6A6-IgG more effectively neutralized KDR, expressed on the surface of the HUVEC cells, compared to the IMC-1C11 IgG chimeric antibody.

This suggests that the results of the VEGF competition assays in ELISA differ from the results indicating that 6A6-IgG and the IMC-1C11 IgG chimeric antibody neutralized KDR at similar levels. That is, the *in vitro* assay results and the *in vivo* assay results can differ from each other and there is a limitation in screening highly efficient antibodies based on the *in vitro* assay results.

Also, in the present invention, it could be observed that the 6A6-IgG antibody was more strongly bound to KDR, expressed in the human acute myeloid eukemia cell line K562 (ATCC, USA), compared to the IMC-1C11 IgG.

Moreover, in the present invention, it was confirmed through Western blotting that 6A6-IgG inhibited the phosphorylation of the angiogenesis signaling factor ERK in primary cultured HUVEC cells and this inhibition was increased according to concentration dependent manner of 6A6-IgG.

Also, in the present invention, it was observed that the 6A6-IgG according to the present invention inhibited the chemotactic motility of HUVEC cells moving to an environment having VEGF present therein and that the 6A6-IgG also inhibited the tube formation of HUVEC cells, which is direct angiogenesis action.

Furthermore, in the present invention, in order to confirm that the inhibitory effect of 6A6-IgG on VEGF effects on HUVEC cells is because 6A6-IgG blocks the entrance of VEGF receptors into HUVEC cells, observation with a confocal microscope was performed in an experiment using a KDR antibody labeled with FITC. As a result, it was obserbed that the VEGF receptor (KDR) could not enter the cells, when cells were treated with 6A6-IgG.

Also, through an *ex vivo* rat aortic ring assay, it was found that, in rat aortic rings treated with 6A6-IgG, vascular sprouting did not occur. Also, angiogenesis was analyzed through a metrigel plug assay by injecting matrigel subcutaneously into mice. As a result, in a group treated with VEGF, angiogenesis in plugs was observed, but in a group treated with VEGF along with 6A6-IgG, angiogenesis was not observed, suggesting that 6A6-IgG had an angiogenesis inhibitory effect *in vivo*.

In still another aspect, the present invention relates to variants obtained by mutating the light chain sequence of 6A6-ScFv through light chain shuffling.

Through the light chain shuffling, 18 light chain variants of 6A6-ScFv were obtained, and the light chain shuffling was performed in the following manner.

(1) In order to prevent 6A6 from being selected again during a biopanning process, DNA of a 6A6 light chain shuffling library was treated with a restriction enzyme *Spe*I having a recognition site at the CDR3 of 6A6. After the DNA was transfected into ETB cells, a sub-library was constructed, and KDR affinity and VEGF competition assays in ELISA were performed to select clones having excellent KDR neutralizing ability.

(2) In a washing step in the biopanning process, the antibody clones were allowed to compete with soluble KDR to select clones having excellent KDR neutralizing ability.

(3) In a step of allowing phages to bind to the antigen KDR in the biopanning

process, IMC-1121B IgG (ImClone, USA) was also added in order to select clones having KDR neutralizing ability which was superior or similar to that of the 1121B IgG.

In still another aspect, the present invention relates to a composition for inhibiting angiogenesis and a composition for treating cancer, which contain said ScFv or IgG.

As used herein, the term "angiogenesis" includes angiogenesis involved in tumor growth and metastasis, age-related macular degeneration, rheumatoid arthritis, diabetic retinopathy, psoriasis and chronic inflammation, but the scope of the present invention is not limited thereto.

In the present invention, said cancer includes, but is not limited to, colon cancer, pancreas cancer, rectal cancer, colorectal cancer, prostate cancer, renal cancer, melanoma, prostate cancer metastasized to bone, ovarian cancer and blood cancer.

The composition of the present invention can be administered by any route suitable for a specific molecule. The composition of the present invention may be provided to animals, including humans, by any suitable means, directly (e.g., locally, such as by injection, subcutaneous injection or topical administration to a tissue locus) or systemically (e.g., parenterally or orally). Where the composition of the present invention is to be provided parenterally, such as by intravenous, subcutaneous, opthalmic, intraperitoneal, intramuscular, buccal, rectal, vaginal, intraorbital, intracerebral, intracranial, intraspinal, intraventricular, intrathecal, intracisternal, intracapsular, intranasal administratiom or by aerosol administration, the composition preferably comprises part of an aqueous or physiologically compatible fluid suspension or solution. Thus, the carrier or excipient is physiologically acceptable so that in addition to delivery of the desired agent to the subject, the solution does not otherwise adversely affect the subject's electrolyte and/or volume balance. The aqueous medium for the agent thus may comprise

normal physiologic saline.

The ScFv or IgG protein of the present invention may be administered for therapeutic treatments to a cancer patient in an amount sufficient to prevent, inhibit, or reduce the progression of the tumor, e.g., the growth, invasiveness, metastases and/or recurrence of the tumor. An amount adequate to accomplish this is defined as a "therapeutically effective dose." Amounts effective for this use will depend upon the severity of the disease and the general state of the patient's own immune system.

The dose of the protein according to the present invention is preferably 0.01-100 mg/kg, and more preferably 0.1-10 mg/m^2.

However, the optimal dose will depend upon a disease being treated and the existence of side effects and can be determined using routine experimentation. The administration of the antibody may be by periodic bolus injections, or by continuous intravenous or intraperitoneal administration from an external reservoir (for example, from an intravenous bag) or an internal reservoir (for example, from a bioerodable implant). Furthermore, the antibody proteins of the present invention also may be administered to the intended recipient together with a plurality of different biologically active molecules. However, the optimal combination of fusion protein and other molecules, modes of administration, dosages may be determined by routine experimentation well within the level of skill in the art.

The composition according to the present invention may be used in combination with other therapeutic agents associated with the relevant disease.

There is synergy when tumors, including human tumors, are treated with the VEGF receptor antibody in combination with radiation, chemotherapy, an additional receptor antagonist or a combination thereof. In other words, the inhibition of

tumor growth by a VEGF receptor antagonist is enhanced more than expected when combined with chemotherapeutic agents, radiation, or an additional receptor antagonist or combinations thereof. Synergy may be shown, for example, by greater inhibition of tumor growth with combined treatment than it would be expected from the additive effect of treatment with a VEGF receptor antagonist and a chemotherapeutic agent, radiation, or an additional receptor antagonist. Preferably, synergy is demonstrated by remission of the cancer where remission is not expected from treatment with a combination of a VEGF receptor antagonist and a chemotherapeutic agent, radiation, or an additional receptor antagonist.

The VEGF receptor antagonist is administered before, during, or after commencing chemotherapy or radiation therapy, as well as any combination thereof, i.e. before and during, before and after, during and after, or before, during, and after commencing the chemotherapy and/or radiation therapy. For example, when the VEGF receptor antagonist is an antibody, the antibody is typically administered between 1 and 30 days, preferably between 3 and 20 days, more preferably between 5 and 12 days before commencing radiation therapy and/or chemotherapy.

VEGF receptor antibody

In one embodiment, the VEGF receptor antibody binds specifically to an epitope on the extracellular domain of a VEGF receptor. The extracellular domain of a VEGF receptor is the ligand-binding domain. The ligand-binding domain may be found at either end of the receptor, but is normally found at the amino-terminal end.

Some examples of VEGF receptors include the protein tyrosine kinase receptors referred to in the literature as Flt-1 (VEGFR-1), KDR and Flk-1 (VEGFR-2). Unless otherwise stated or clearly suggested otherwise by context, this specification will follow the customary literature nomenclature of VEGF receptors. KDR will be referred to as the human form of a VEGF receptor having MW 180 kD (Terman

et al., *Oncogene*, 6:1677, 1991). Flk-1 (VEGFR-2) is will be referred to as the murine homolog of KDR (Matthews *et al.*, *PNAS*, 88:9026, 1991). Flt-1 (VEGFR-1) is referred to as a form of VEGF receptor different from, but related to, the KDR/Flk-1 receptor (Shibuya *et al.*, *Oncogene*, 5:519, 1990).

Other VEGF receptors include those that can be cross-link with labeled VEGF, or that can be co-immunoprecipitated with KDR. Some known forms of these VEGF receptors have molecular weights of approximately 170 kD, 150 kD, 130-135 kD, 120-125 kD and 85 kD (Quinn *et al.*, *PNAS*, 90:7533, 1993; Scher *et al.*, *J. Biol. Chem.*, 271:5761, 1996).

The VEGF receptor is usually bound to a cell, such as an endothelial cell. The VEGF receptor may also be bound to a non-endothelial cell, such as a tumor cell. Alternatively, the VEGF receptor may be free from the cell, preferably in soluble form.

The antagonist of the present invention neutralizes VEGF receptors. In this specification, neutralizing a receptor means inactivating the intrinsic kinase activity of the receptor to transduce a signal. A reliable assay for VEGF receptor neutralization is the inhibition of receptor phosphorylation.

The present invention is not limited by any particular mechanism of VEGF receptor neutralization. The mechanism caused by one antagonist is not necessarily the same as that caused by another antagonist. Some possible mechanisms include preventing binding of the VEGF ligand to the extracellular binding domain of the VEGF receptor, and preventing dimerization or oligomerization of receptors. Other mechanisms cannot, however, be ruled out.

A VEGF receptor (or VEGFR) antibody, in the context of the present invention, inhibits activation of the VEGFR subfamily of receptors. By inhibition of

activation of the VEGFR subfamily of receptors is meant any decrease in the activation of the VEGFR. That is, the prevention of activation need not completely stop activation of the VEGFR. Moreover, inhibition of VEGFR activation, as defined by the present invention, means inhibition of the VEGFR following interaction of the VEGFR antibody and VEGFR. By association is meant sufficient physical or chemical interaction between the VEGFR and VEGFR antibody which inhibits tyrosine kinase activity of the receptor. One of skill in the art would appreciate those examples of such chemical interactions, which include association or bonding, are known in the art and include covalent bonding, ionic bonding, hydrogen bonding, etc. Accordingly, the VEGFR antagonist of the present invention inhibits the tyrosine kinase activity of the receptor, which prevents autophosphorylation of the receptor and phosphorylation of various other proteins involved in the VEGFR signaling pathways. Such pathways, which are involved in regulation of vasculogenesis and angiogenesis, include any of the following: the phospholipase Cy (PLCy) pathway or the phosphatidylinositol 3' kinase (PI3-K)/Akt and mitogen activated protein kinase (MAPK) pathway (Larrivee *et al.*, *Int. J. Med.*, 5:447, 2000).

The VEGFR subfamily of receptors is characterized by the presence of seven immunoglobulin-like loops in the extracellular domain, a single transmembrane region and a split tyrosine kinase domain in the intracellular region (class III receptor tyrosine kinases). There are several known members of the VEGFR subfamily of receptors, examples of which include VEGFR-1, VEGFR-2, and VEGFR-3.

It is generally believed that KDR (VEGFR-2) is the main VEGF signal transducer that results in endothelial cell proliferation, migration, differentiation, tube formation, increase of vascular permeability, and maintenance of vascular integrity. VEGFR-1 possesses a much weaker kinase activity, and is unable to generate a mitogenic response when stimulated by VEGF, although it binds to VEGF with an

affinity that is approximately 10-fold higher than KDR (VEGFR-2). VEGFR-1 has also been implicated in VEGF- and placenta growth factor (P1GF)-induced migration of monocytes and macrophages and production of tissue factor.

As is the case with VEGFR described above, increased VEGFR activation can result from higher levels of ligand, VEGFR gene amplification, increased transcription of the receptor or mutations that cause unregulated receptor signaling.

In one embodiment of the present invention, the VEGFR antibody inhibits binding of VEGFR to its ligand. Binding of a ligand to an external, extracellular domain of VEGFR stimulates receptor dimerization, autophosphorylation of VEGFR, activation of the receptor's internal, cytoplasmic tyrosine kinase domain, and initiation of multiple signal transduction pathways involved in regulation of vasculogenesis and angiogenesis.

Ligands for VEGFR include VEGF and its homologues P1GF, VEGF-B, VEGF-C, VEGF-D, and VEGF-E. For example, P1GF, which is a dimeric secreted factor and only binds VEGFR-1, is produced in large amounts by villous cytotrophoblast, sincytiotrophoblast and extravillous trophoblast and has close amino acid homology to VEGF. Three isoforms exist in humans, P1GF-1, P1GF-2, and P1GF-3. Studies with P1GF- deficient mice demonstrate that this growth factor is not involved in angiogenesis per se, but rather, specifically modulates the angiogenic and permeability effects of VEGF during pathological situations. Also, VEGF-D is closely related to VEGF-C by virtue of the presence of N-and C-terminal extensions that are not found in other VEGF family members. In adult human tissues, VEGF-D mRNA is most abundant in heart, lung, skeletal muscle, colon, and small intestine. Analyses of VEGF-D receptor specificity revealed that VEGF-D is a ligand for both VEGFR-2 (Flkl) and VEGFR-3 (Flt4) and can activate these receptors; however, VEGF-D does not bind to VEGFR-1. In addition, VEGF-D is a mitogen for endothelial cells.

In another embodiment of the present invention, the VEGFR antibody binds specifically to VEGFR. It should be appreciated that the VEGFR antibody can bind externally to the extracellular portion of VEGFR, which may or may not inhibit binding of the ligand, or internally to the tyrosine kinase domain. Preferably, the VEGFR antagonist of the present invention is an antibody, or functional equivalent thereof, specific for VEGFR, details of which are described in more detail below.

In one preferred embodiment, the VEGF receptor antibody binds specifically to KDR. Particularly preferred are antigen-binding proteins that bind to the extracellular domain of KDR and block binding by one or both of its ligands, VEGF and P1GF, and/or neutralize VEGF-induced or PIGF-induced activation of KDR.

There also exist various hybridomas that produce VEGFR-2 antibodies. For example, a hybridoma cell line producing rat anti-mouse VEGFR-2 monoclonal antibody (DC101) was deposited as ATCC HB 11534; a hybridoma cell line (M25. 18A1) producing mouse anti-mouse VEGFR-2 monoclonal antibody mAb 25 was deposited as ATCC HB 12152; and a hybridoma cell line (M73.24) producing mouse anti-mouse VEGFR-2 monoclonal antibody mAb 73 was deposited as ATCC HB 12153.

In addition, various hybridomas that produce anti-VEGFR-1 antibodies exist and include, but are not limited to, hybridomas KM1730 (deposited as FERM BP-5697), KM1731 (deposited as FERM BP-5718), KM1732 (deposited as FERM BP-5698), KM1748 (deposited as FERM BP-5699), KM1750 (deposited as FERM BP-5700) disclosed in WO 98/22616, WO 99/59636, AU 5066698 B2, and CA 2328893.

Many other VEGFR antagonists are known in the art. Some examples of VEGFR antagonists are described in US 5,185,438; 5,621,090; 5,283,354; 5,270,458;

5,367,057; 5,548,065; 5,747,651; 5,912,133; 6,677,434; 6,960,446; 5,840,301; 5,861,499; 6,365,157; 5,955,311; 6,365,157; 6,811,779; and WO 2001/66063. US 5,861,301, Terman *et al., Oncogene,* 6:1677, 1991, WO 94/10202, and WO 95/21865, disclose VEGFR antagonists and, specifically, anti-VEGFR-2 antibodies. In addition, anti-VEGFR-2 antibodies are disclosed in US. 6,177,401 and 5,712,395. US 5,981,569 discloses VEGFR antagonists that are organic molecules. Also, bi-specific antibodies (BsAbs), which are antibodies that have two different antigen-binding specificities or sites, directed against KDR (VEGFR-2) and VEGFR-1 are known. Also, Hennequin *et al., J. Med. Chem.,* 42:5369, 1999 discloses certain quinazolines, quinolines and cinnolines as being useful as VEGF receptor antagonists (Annie *et al., J. Acqu. Immune Defic. Syn. and Hum. Retrovirol.*, 17: A41, 1998).

Furthermore, assays for the determination of VEGFR antibodies are known in the art. The VEGFR antibodies of the present invention inhibit the tyrosine kinase activity of VEGFR, which generally involves phosphorylation events. Accordingly, phosphorylation assays are useful in determining VEGFR antibodies in the present invention. Some assays for tyrosine kinase activity are described in Panek *et al., J. Pharmacol. Exp. Thera.*, 283:1433, 1997 and Batley *et al., Life Sci.*, 62:143, 1998. In addition, methods specific for detection of VEGFR expression can be utilized.

Antibodies

The antibodies of the present invention may be produced by methods known in the art (Kohler and Milstein, *Nature*, 256:495, 1975; Campbell, Monoclonal Antibody Technology, The Production and Characterization of Rodent and Human Hybridomas; Burdon *et al.*, Eds., Laboratory Techniques in Biochemistry and Molecular Biology, Vol. 13, Elsevier Science Publishers, Amsterdam, 1985; Huse *et al., Science*, 246:1275, 1989). The antibodies of the present invention can be

monoclonal or polyclonal antibodies or any other suitable type of an antibody, such as a fragment or a derivative of an antibody, a single chain variable fragment (scFv) or a synthetic homolog of the antibody, provided that the antibody has the same binding characteristics as, or that has binding characteristics comparable to, those of the whole antibody. As used herein, unless otherwise indicated or clear from the context, antibody domains, regions and fragments follow standard definitions as are well known in the art (Abbas *et al.*, Cellular and Molecular Immunology, W.B. Saunders Company, Philadelphia, PA, 1991). Preferably, the antibodies of the present invention are monoclonal antibodies.

Antibody fragments can be produced by cleaving a whole antibody, or by expressing DNA that encodes the fragment. Fragments of antibodies may be prepared by methods described in the published literature (Lamoyi *et al., J. Immunol. Methods*, 56:235, 1983; Parham, *J. Immunol.,* 131:2895, 1983). Such fragments may contain one or both of an Fab fragment and an F(ab')2 fragment. Such fragments may also contain single chain variable fragment antibodies, i.e. scFv, dibodies, or other antibody fragments. Methods of producing such functional equivalents are disclosed in WO 93/21319, EP 239,400, WO 89/09622, EP 338,745 and EP 332,424.

Single chain variable fragments (scFv) are polypeptides that consist of the variable region of a heavy chain of an antibody linked to the variable region of a light chain a short peptide linker. Thus, the scFv comprises the entire antibody-combining site. These chains may be produced in bacteria, or in eukaryotic cells. A typical example of a single chain antibody in the present invention is 6A6-ScFv (TTAC-0001). 6A6-ScFv was shown to block VEGF-KDR (VEGF-VEGFR-2) interaction and inhibit VEGF-stimulated receptor phosphorylation. This 6A6-ScFv binds both to soluble KDR (VEGFR-2) and cell surface-expressed KDR (VEGFR-2) on HUVEC cells and K562 cells. The 6A6-ScFv has a light chain sequence of SEQ ID NO: 35 and a heavy chain sequence of SEQ ID NO: 36. The 6A6-ScFv

antibody is a fully human antibody and can be constructed with Fab', F(ab')2, bivalent ScFv, bivalent recombiant ScFv or human IgG antibodies.

Preferably, although the antibody fragments contain all six complementarity-determining regions (CDRs) of the whole antibody, fragments containing fewer than all of such regions, such as three, four or five CDRs, may also be functional. If the antibody fragment is too short to be immunogenic, it may be conjugated to a carrier molecule. Some suitable carrier molecules include keyhole limpet hemocyanin and bovine serum albumen. Conjugation may be carried out by methods known in the art.

Antibodies of the present invention also include antibodies whose binding characteristics can be improved by direct mutation, methods of affinity maturation, phage display, or chain shuffling. Affinity and specificity may be modified or improved by mutating CDRs and screening for antigen binding sites having the desired characteristics (Yang *et al., J. Mol. Biol.*, 254:392, 1995). CDRs are mutated in a variety of ways. One way is to randomize individual residues or combinations of residues so that in a population of otherwise identical antigen binding sites, all twenty amino acids are found at particular positions. Alternatively, mutations are induced over a range of CDR residues by error prone PCR methods ((Hawkins *et al., J. Mol. Biol.*, 226:889, 1992). Phage display vectors containing heavy and light chain variable region genes are propagated in mutator strains of *E. coli* (Low *et al., J. Mol. Biol.*, 250:359, 1996). These methods of mutagenesis are illustrative of the many methods known to one of skill in the art.

Antibodies, and particularly monoclonal antibodies, can be produced by methods known in the art. Examples for production of antibodies include, but are not limited to, production in hybridoma cells and transformation of mammalian cells with DNA encoding the receptor antagonist. These methods are described in

various publications (Kohler and Milstein, *Nature*, 256:495, 1975; Campbell in "Monoclonal Antibody Technology, The Production and Chracterization of Rodent and Human Hybridomas" in Burdon *et al*, Eds., Laboratory Techniques in Biochemistry and Molecular Biology, Vol.13, Elservier Science Publishers, Amsterdam, 1985; Huse *et al., Science*, 246:1275, 1989).

Equivalents of antibodies are also prepared by methods known in the art. For example, fragments of antibodies may be prepared enzymatically from whole antibodies. Preferably, equivalents of antibodies are prepared from DNA encoding such equivalents. DNA encoding fragments of antibodies may be prepared by deleting all but the desired portion of the DNA that encodes the full-length antibody. DNA encoding chimerized antibodies may be prepared by recombining DNA encoding human constant regions, derived substantially or exclusively from the corresponding human antibody regions, and DNA encoding variable regions, derived substantially or exclusively from the sequence of the variable region of a mammal other than a human. DNA encoding humanized antibodies may be prepared by recombining DNA encoding constant regions and variable regions other than the complementarity determining regions (CDRs), derived substantially or exclusively from the corresponding human antibody regions, and DNA encoding CDRs, derived substantially or exclusively from a mammal other than a human.

Suitable sources of DNA molecules that encode fragments of antibodies include cells, such as hybridomas, that express the full-length antibody. The fragments may be used by themselves as antibody equivalents, or may be recombined into equivalents, as described above. The DNA deletions and recombinations described in this section may be carried out by known methods, such as those described in the published patent applications listed above in the section entitled "Functional Equivalents of Antibodies" and/or other standard recombinant DNA techniques, such as those described below.

Preferred host cells for transformation of vectors and expression of the antibodies of the present invention are mammalian cells, e.g., COS-7 cells, Chinese hamster ovary (CHO) cells, and cell lines of lymphoid origin such as lymphoma, myeloma, or hybridoma cells. Other eukaryotic host, such as yeasts, can be alternatively used. For example, mouse fetal liver stromal cell line 2018 binds to APtag-Flk 1 and APtag-Flk-2 fusion proteins, i.e., contains ligands of VEGFR-2 and Flk-2 (ATCC, Manassas, VA, CRL 10907), human fetal spleen cell line Fsp 62891 contains Flk-2 ligand (ATCC CRL 10935), and human stromal fetal thymus cell line, F. thy 62891, contains VEGFR-2 ligand (ATCC CRL 10936).

As used herein, the term "vector" means any nucleic acid comprising a competent nucleotide sequence to be incorporated into a host cell and to be recombined with and integrated into the host cell genome, or to replicate autonomously as an episome. Such vectors include linear nucleic acids, plasmids, phagemids, cosmids, RNA vectors, viral vectors and the like. Examples of a viral vector include, but are not limited to, a retrovirus, an adenovirus and an adeno-associated virus.

As used herein, the term “gene expression” or “expression of a target protein” is understood to mean the transcription of a DNA sequence, the translation of an mRNA transcript and the secretion of an Fc fusion protein product.

In the present invention, suitable host cells can be transformed or transfected with DNA and can be used to express and/or secrete target proteins. Preferred host cells for use in the present invention include immortalized hybridoma cells, NS/O myeloma cells, 293 cells, Chinese hamster ovary (CHO) cells, HELA cells and COS cells.

The transformed host cells are cultured by methods known in the art in a liquid medium containing assimilable sources of carbon (carbohydrates such as glucose or lactose), nitrogen (amino acids, peptides, proteins or their degradation products such

as peptones, ammonium salts or the like), and inorganic salts (sulfates, phosphates and/or carbonates of sodium, potassium, magnesium and calcium). The medium additionally contains, for example, growth-promoting substances, such as trace elements, for example iron, zinc, manganese and the like.

Where it is desired to express a gene construct in yeast, a suitable selection gene for use in yeast is the trpl gene present in the yeast plasmid YRp7 (Stinchcomb *et al.*, *Nature*, 282:39, 1979; Kingsman *et al.*, *Gene*, 7:141, 1979). The trpl gene provides a selection marker for a mutant strain of yeast lacking the ability to grow in tryptophan, for example, ATCC 44076 or PEP4-1 (Jones, *Genetics*, 85:12, 1977). The presence of the trpl lesion in the yeast host cell genome then provides an effective environment for detecting transformation by growth in the absence of tryptophan. Similarly, Leu2-deficient yeast strains (ATCC 20622 or 38626) are complemented by known plasmids bearing the Leu2 gene.

Alternatively, the DNA encoding the receptor antagonist can be cloned into vectors derived from viruses such as adenovirus, adeno-associated virus, herpesvirus, retrovirus or lentivirus. Gene expression is controlled by inducible or uninducible regulatory sequences.

Briefly, a suitable source of cells containing nucleic acid molecules that express the desired DNA, such as an antibody, antibody equivalent or VEGF receptor, is selected. Total RNA is prepared by standard procedures from a suitable source. The total RNA is used to direct cDNA synthesis. Standard methods for isolating RNA and synthesizing cDNA are provided in standard manuals of molecular biology such as, for example, those described above.

The cDNA may be amplified by known methods. For example, the cDNA may be used as a template for amplification by polymerase chain reaction (PCR) (Saiki *et al.*, *Science*, 239:487, 1988; US 4,683,195). The sequences of the oligonucleotide

primers for the PCR amplification are derived from the known sequence to be amplified. The oligonucleotides are synthesized by methods known in the art (Caruthers, *Science,* 230:281, 1985).

A mixture of upstream and downstream oligonucleotides is used in the PCR amplification. The conditions are optimized for each particular primer pair according to standard procedures. The PCR product is analyzed, for example, by electrophoresis for cDNA having the correct size, corresponding to the sequence between the primers. Alternatively, the coding region may be amplified in two or more overlapping fragments. The overlapping fragments are designed to include a restriction site permitting the assembly of the intact cDNA from the fragments.

In order to isolate the entire protein-coding regions for the VEGF receptors, for example, the upstream PCR oligonucleotide primer is complementary to the sequence at the 5' end, preferably encompassing the ATG start codon and at least 5-10 nucleotides upstream of the start codon. The downstream PCR oligonucleotide primer is complementary to the sequence at the 3' end of the desired DNA sequence. The desired DNA sequence preferably encodes the entire extracellular portion of the VEGF receptor, and optionally encodes all or part of the transmembrane region, and/or all or part of the intracellular region, including the stop codon.

The DNA to be amplified, such as that encoding antibodies, antibody equivalents, or VEGF receptors, may also be replicated in a wide variety of cloning vectors in a wide variety of host cells. The host cell may be prokaryotic or eukaryotic.

The vector into which the DNA is spliced may comprise segments of chromosomal, non-chromosomal and synthetic DNA sequences. Some suitable prokaryotic cloning vectors include plasmids derived from *E. coli*, such as colEl, pCRI, pBR322, pMB9, pKSM, and RP4. Prokaryotic vectors also include derivatives of phage DNA such as M13 and other filamentous single-stranded DNA phages. A

preferred vector for cloning nucleic acid encoding the VEGF receptor is the Baculovirus vector.

The vector containing the DNA to be expressed is transfected into a suitable host cell. The host cell is maintained in an appropriate culture medium, and subjected to conditions under which the cells and the vector replicate. The vector may be recovered from the cell. The DNA to be expressed may be recovered from the vector.

The DNA to be expressed, such as that encoding antibodies, antibody equivalents, or receptors, may be inserted into a suitable expression vector and expressed in a suitable prokaryotic or eucaryotic host cell.

For example, the DNA inserted into a host cell may encode the entire extracellular portion of the VEGF receptor, or a soluble fragment of the extracellular portion of the VEGF receptor. The extracellular portion of the VEGF receptor encoded by the DNA is optionally attached at either, or both, the 5' end or the 3' end to additional amino acid sequences. The additional amino acid sequences may be attached to the VEGF receptor extracellular region, such as the leader sequence, the transmembrane region and/or the intracellular region of the VEGF receptor. The additional amino acid sequences may also be sequences not attached to the VEGF receptor in nature. Preferably, such additional amino acid sequences serve a particular purpose, such as to improve expression levels, secretion, solubility, or immunogenicity.

Vectors for expressing proteins in bacteria, especially *E. coli*, are known (Dieckmann and Tzagoloff, *J. Biol. Chem.*, 260:1513, 1985). These vectors contain DNA sequences that encode anthranilate synthetase (TrpE) followed by a polylinker at the carboxy terminus. Other expression vector systems are based on beta-galactosidase (pEX); lambda PL; maltose binding protein (pMAL); and

glutathione S-transferase (pGST) (*Gene*, 67:31, 1988; *Peptide Research,* 3:167, 1990).

Suitable vectors for expression in mammalian cells are also known. Such vectors include well-known derivatives of SV-40, adenovirus, retrovirus-derived DNA sequences and shuttle vectors derived from combination of functional mammalian vectors, such as those described above, and functional plasmids and phage DNA.

Additional vectors for expression of eukaryotic cells are known in the art (Southern, P. J. and Berg, P., *J. Mol. Appl. Genet.*, 1:327, 1982; Subramani *et al, Mol. Cell. Biol.*, 1:854, 1981; Kaufinann and Sharp, *J. Mol. Biol.*, 159:601, 1982; Kaufinann and Sharp, *Mol. Cell. Biol.*, 1982; Scahill *et al., PNAS*, 80:4654, 1983; Urlaub and Chasin, *PNAS*, 77:4216, 1980). The expression vectors useful in the present invention contain at least one expression control sequence that is operatively linked to the DNA sequence or fragment to be expressed. The control sequence is inserted in the vector in order to control and to regulate the expression of the cloned DNA sequence. Examples of useful expression control sequences include the lac system, the trp system, the tac system, the trc system, major operator and promoter regions of phage lambda, the control region of fd coat protein, the glycolytic promoters of yeast, e.g., the promoter for 3-phosphoglycerate kinase, the promoters of yeast acid phosphatase, e.g., Pho5, the promoters of the yeast alpha-mating factors, and promoters derived from polyoma, adenovirus, retrovirus, and simian virus, e.g., the early and late promoters of SV40, and other sequences known to control the expression of genes of prokaryotic or eukaryotic cells and their viruses.

Vectors containing the control signals and DNA to be expressed, such as that encoding antibodies, antibody equivalents, or VEGF receptors, are inserted into a host cell for expression. Some useful expression host cells include well-known prokaryotic and eukaryotic cells. Some suitable prokaryotic hosts include, for example, *E. coli,* such as *E. coli* SG-936, *E. coli* HB 101, *E. coli* W3110, *E. coli*

X1776, *E. coli* X2282, *E. coli* DHI, and *E. coli* MRCI, *Pseudomonas*, *Bacillus*, such as *Bacillus subtilis*, and *Streptomyces*. Suitable eukaryotic cells include yeast and other fungi, insect, animal cells, such as COS cells and CHO cells, human cells and plant cells in tissue culture.

Following expression in a host cell maintained in a suitable medium, the polypeptide or peptide to be expressed, such as antibodies, antibody equivalents, or VEGF receptors, may be isolated from the medium, and purified by methods known in the art. If the polypeptide or peptide is not secreted into the culture medium, the host cells are lysed prior to isolation and purification.

In addition, the antibodies of the invention may be prepared by immunizing a mammal with a soluble receptor. The soluble receptors themselves may be used as immunogens, or may be attached to a carrier protein or to other objects, such as beads, i.e., sepharose beads. After the mammal has produced antibodies, a mixture of antibody-producing cells, such as the splenocytes, is isolated. Monoclonal antibodies may be produced by isolating individual antibody-producing cells from the mixture and making the cells immortal by, for example, fusing them with tumor cells, such as myeloma cells. The resulting hybridomas are preserved in culture, and express monoclonal antibodies, which are harvested from the culture medium.

The antibodies may also be prepared from receptors bound to the surface of cells that express the specific receptor of interest. The cell to which the receptors are bound may be a cell that naturally expresses the receptor, such as a vascular endothelial cell for VEGFR. Alternatively, the cell to which the receptor is bound may be a cell into which the DNA encoding the receptor has been transfected, such as 3T3 cells, which have been transfected with VEGFR.

A receptor may be used as an immunogen to raise an antibody of the present invention. The receptor peptide may be obtained from natural sources, such as

from cells that express the receptors. For example, the VEGF receptor peptide may be obtained from vascular endothelial cells. Alternatively, synthetic receptor peptides may be prepared using commercially available machines. In such an embodiment, the VEGF receptor amino acid sequence can be provided through the published literatures (Shibuya *et al., Oncogene*, 5:519, 1990; PCT/US92/01300; Terman *et al., Oncogene,* 6:1677, 1991; Matthews *et al., PNAS*, 88:9026, 1991).

As an alternative, DNA encoding a receptor, such as a cDNA or a fragment thereof, is cloned and expressed, and the resulting polypeptide is recovered and thus it may be used as an immunogen to raise an antibody of the present invention. For example, in order to prepare the VEGF receptors against which the antibodies are made, nucleic acid molecules that encode the VEGF receptors of the present invention, or portions thereof, especially the extracellular portions thereof, may be inserted into known vectors for expression in host cells using standard recombinant DNA techniques, such as those described below. Suitable sources of such nucleic acid molecules include cells that express VEGF receptors, i.e., vascular endothelial cells.

The antibody may be prepared in any mammal; suitable mammals other than human include, for example, a rabbit, rat, mouse, horse, goat, or primate. Mice are frequently used to prepare monoclonal antibodies. The antibody may be a member of one of the following immunoglobulin classes: IgG, IgM, IgA, IgD, or IgE, and the subclasses thereof, and preferably is an IgG1 antibody. The antibodies of the present invention and their functional equivalents may be or may combine members of any of the immunoglobulin classes.

Neutralization of VEGF activation of VEGF receptors

Neutralization of activation of a VEGF receptor in a sample of endothelial or non-endothelial cells, such as tumor cells, may be performed *in vitro* or *in vivo*.

Neutralizing VEGF activation of a VEGF receptor in a sample of VEGF-receptor expressing cells comprises contacting the cells with the antibody of the present invention. The cells are contacted *in vitro* with the antibody, before, simultaneously with, or after, adding VEGF to the cell sample.

In vivo, the antibody of the present invention is contacted with a VEGF receptor by administration to a mammal. Methods of administration to a mammal include, for example, oral, intravenous, intraperitoneal, subcutaneous, or intramuscular administration.

This *in vivo* neutralization method is useful for inhibiting angiogenesis in a mammal. Angiogenesis inhibition is a useful therapeutic method, such as for preventing or inhibiting angiogenesis associated with pathological conditions such as tumor growth. Accordingly, the the antibody of the present invention is an anti-angiogenic and anti-tumor immunotherapeutic agent.

As used herein the term "mammal" means any mammal. Some examples of mammals include pet animals, such as dogs and cats; farm animals, such as pigs, cattle, sheep, and goats; laboratory animals, such as mice and rats; primates, such as monkeys, apes, and chimpanzees; and humans.

VEGF receptors are found on some non-endothelial cells, such as tumor cells, indicating the unexpected presence of an autocrine and/or paracrine loop in these cells. The antagonists, e.g., the antibodies, of this invention are useful in neutralizing the activity of VEGF receptors on such cells, thereby blocking the autocrine and/or paracrine loop, and inhibiting tumor growth. The methods of inhibiting angiogenesis and of inhibiting pathological conditions such as tumor growth in a mammal comprise administering an effective amount of any one of the inventive antagonists, e.g., antibodies, including any of the functional equivalents thereof, systemically to a mammal, or directly to a tumor within the mammal. The

mammal is preferably human. This method is effective for treating subjects with both solid tumors, preferably highly vascular tumors, and non-solid tumors.

The inhibition or reduction of tumor growth includes the prevention or inhibition of the progression of a tumor, including cancerous and noncancerous tumors. The progression of a tumor includes the invasiveness, metastasis, recurrence and increase in size of the tumor. The inhibition or reduction of tumor growth also includes the destruction of a tumor.

All types of tumors may be treated by the methods of the present invention. The tumors may be solid or non-solid.

Some examples of solid tumors that can be treated with the antagonists of the present invention include carcinomas, sarcomas, blastomas or gliomas. Some examples of such tumors include epidermoid tumors, squamous tumors, such as head and neck tumors, colorectal tumors, prostate tumors, breast tumors, lung tumors, including small cell and non-small cell lung tumors, pancreatic tumors, thyroid tumors, ovarian tumors, and liver tumors. Other examples include Kapos's sarcoma, CNS neoplasms, neuroblastomas, capillary hemangioblastomas, meningiomas and cerebral metastases, melanoma, gastrointestinal and renal carcinomas and sarcomas, rhabdomyosarcoma, glioblastoma, preferably glioblastoma multiforme, and leiomyosarcoma. Examples of vascularized skin cancers for which the antagonists of this invention are effective include squamous cell carcinoma, basal cell carcinoma and skin cancers that can be treated by suppressing the growth of malignant keratinocytes, such as human malignant keratinocytes.

Some examples of non-solid tumors include leukemias, multiple myelomas and lymphomas. Some examples of leukemias include acute myelocytic leukemia (AML), chronic myelocytic leukemia (CML), acute lymphocytic leukemia (ALL),

chronic lymphocytic leukemia (CLL), erythrocytic leukemia or monocytic leukemia. Some examples of lymphomas include lymphomas associated with Hodgkin's disease and non- Hodgkin's disease.

Preventing or inhibiting angiogenesis is also useful to treat non-neoplastic pathologic conditions characterized by excessive angiogenesis, such as neovascular glaucoma, proliferative retinopathy including proliferative diabetic retinopathy, arthritis, macular degeneration, hemangiomas, angiofibromas, and psoriasis.

Using Inventive Antibodies to Isolate and Purify VEGF Receptor

The antagonists of the present invention may be used to isolate and purify the VEGF receptor using conventional methods such as affinity chromatography (Dean *et al.*, Affinity Chromatography: A Practical Approach, IRL Press, Arlington, VA, 1985). Other methods well known in the art include magnetic separation with antibody-coated magnetic beads, "panning" with an antibody attached to a solid matrix, and flow cytometry (FACS).

The source of the VEGF receptor is typically vascular cells, and especially vascular endothelial cells, that express the VEGF receptor. Suitable sources of vascular endothelial cells are blood vessels, such as umbilical cord blood cells, especially, human umbilical cord vascular endothelial cells (HUVEC).

The VEGF receptors may be used as a starting material to produce other materials, such as antigens for making additional monoclonal and polyclonal antibodies that recognize and bind to the VEGF receptor or other antigens on the surface of VEGF-expressing cells.

Using Inventive Antibodies to Isolate and Purify KDR Positive Tumor Cells

The antibodies of the present invention may be used to isolate and purify Flk-1 KDR (VEGFR-2) positive tumor cells, i.e., tumor cells expressing KDR, using conventional methods such as affinity chromatography (Dean, P.D.G. *et al.*, Affinity Chromatography: A Practical Approach, IRL Press, Arlington, VA, 1985). Other methods well known in the art include magnetic separation with antibody-coated magnetic beads, cytotoxic agents, such as complement, conjugated to the antibody, "panning" with an antibody attached to a solid matrix, and flow cytometry (FACS).

Monitoring Levels of VEGF and VEGF Receptors *In Vitro* or *In Vivo*

The antibodies of the present invention may be used to monitor the levels of VEGF or VEGF receptors *in vitro* or *in vivo* in biological samples using standard assays and methods known in the art. Some examples of biological samples include bodily fluids, such as blood. Standard assays involve, for example, labeling the antibodies and conducting standard immunoassays, such as radioimmunoassays, as is well know in the art.

Standard recombinant DNA techniques useful in carrying out the present invention are described in the literature (Sambrook *et al.*, "Molecular Cloning, "Second Edition, Cold Spring Harbor Laboratory Press, 1987; Ausubel *et al,* (Eds) "Current Protocols in Molecular Biology, "Green Publishing Associates/Wiley-Interscience, New York, 1990).

Administration

The receptor antibodies of the present invention can be administered for therapeutic treatments to a patient suffering from a tumor in an amount sufficient to prevent, inhibit, or reduce the progression of the tumor, e.g., the growth, invasiveness, metastases and/or recurrence of the tumor. An amount adequate to accomplish this

purpose is defined as a therapeutically effective dose. Amounts effective for this use will depend upon the severity of the disease and the general state of the patient's own immune system. Dosing schedules will also vary depending on the disease state and status of the patient, and will typically range from a single bolus dosage or continuous infusion to multiple administrations per day (e.g., every 4-6 hours), or as indicated by the treating physician and the patient's condition. It should be noted, however, that the present invention is not limited to any particular dose.

The present invention can be used to treat any suitable tumor, including, for example, tumors of the breast, heart, lung, small intestine, colon, spleen, kidney, bladder, head and neck, ovary, prostate, brain, pancreas, skin, bone, bone marrow, blood, thymus, uterus, testicles, cervix or liver. Preferably, the methods of the present invention are used when the tumor is a tumor of the colon or when the tumor is a non-small cell lung carcinoma (NSCLC).

Furthermore, the tumors of the present invention preferably have aberrant expression or signaling of VEGFR. Enhanced signaling by VEGFR has been observed in many different human cancers. High levels of VEGFR-2 are expressed by endothelial cells that infiltrate gliomas (Plate *et al.*, *Nature,* 359:845, 1992). VEGFR-2 levels are specifically upregulated by VEGF produced by human glioblastomas (Plate *et al.*, *Cancer Res.*, 53:5822, 1993). The finding of high levels of VEGFR-2 expression in glioblastoma associated endothelial cells (GAEC) indicates that receptor activity is probably induced during tumor formation since VEGFR-2 transcripts are barely detectable in normal brain endothelial cells. This upregulation is confined to the vascular endothelial cells in close proximity to the tumor.

The present invention is useful for inhibition or reduction of tumor growth. By inhibition or reduction of tumor growth is meant prevention, inhibition, or reduction of the progression of the tumor, e. g, the growth, invasiveness, metastases and/or

recurrence of the tumor. In addition, the present invention can also be useful in treating an angiogenic condition, such as atherosclerosis, arthritis, macular degeneration and psoriasis. The identification of those patients that have conditions for which the present invention is useful is well within the ability and knowledge of one skilled in the art.

In the present invention, any suitable method or route can be used to administer the VEGFR antibodies, for example, oral, intravenous, intraperitoneal, subcutaneous, or intramuscular administration. The dose of antagonist administered depends on numerous factors, including, for example, the type of antibodies, the type and severity of tumor to be treated and the route of administration of the antibodies. It should be emphasized, however, that the present invention is not limited to any particular method or route of administration.

In one alternative embodiment, the VEGFR antagonist and can be administered in combination with one or more antineoplastic agents (US 6,217,866). Any suitable antineoplastic agent can be used, such as a chemotherapeutic agent or radiation. Examples of chemotherapeutic agents include, but are not limited to, cisplatin, doxorubicin, paclitaxel, irinotecan (CPT-11), topotecan or a combination thereof. When the antineoplastic agent is radiation, the source of the radiation can be either external (external beam radiation therapy-EBRT) or internal (brachytherapy-BT) to the patient being treated. The dose of antineoplastic agent administered depends on numerous factors, including, for example, the type of agent, the type and severity of tumor being treated and the route of administration of the agent. It should be emphasized, however, that the present invention is not limited to any particular dose.

In an additional alternative embodiment, the VEGFR antibody of the present invention can be administered in combination with one or more suitable adjuvants, such as, for example, cytokines (for example, IL-10 and IL-13) or other immune stimulators. See, for example, Larrivee *et al.*, supra. It should be appreciated,

however, that administration of only the VEGFR antagonist is sufficient to prevent, inhibit, or reduce the progression of the tumor in a therapeutically effective manner.

In addition, the VEGFR antibody can be administered as a ligand conjugate, which binds specifically to the receptor and delivers a toxic, lethal payload following ligand-toxin internalization. Conjugates between toxins and the receptors were designed with the aim of developing toxic agents specific for EGFR- or VEGFR-overexpressing tumor cells while minimizing nonspecific toxicity. For example, a conjugate composed of EGF and Pseudomonas endotoxin (PE) was shown to be toxic toward EGFR-expressing HeLa cells *in vitro*. Various agents, including thioridazine and adenovirus, can enhance cellular uptake of the conjugate, as well as increase the cytotoxicity of the conjugate.

It is understood that the VEGFR antibodies of the present invention, where used in a mammal for the purpose of prophylaxis or treatment, will be administered in the form of a composition additionally comprising a pharmaceutically acceptable carrier. Suitable pharmaceutically acceptable carriers include, for example, one or more of water, saline, phosphate buffered saline, dextrose, glycerol, ethanol and the like, as well as combinations thereof. Pharmaceutically acceptable carriers may further comprise minor amounts of auxiliary substances such as wetting or emulsifying agents, preservatives or buffers, which enhance the shelf life or effectiveness of the binding proteins. The compositions for the injection may, as is well known in the art, be formulated so as to provide quick, sustained or delayed release of the active ingredient after administration to the mammal.

The VEGFR antibodies of the present invention may be in a variety of forms. These include, for example, solid, semi-solid and liquid dosage forms, such as tablets, pills, powders, liquid solutions, dispersions or suspensions, liposomes, suppositories, injectable and infusible solutions. The preferred form depends on the intended mode of administration and therapeutic application.

Such antibodies can be prepared in a manner well known in the pharmaceutical art. In making the composition, the active ingredient will usually be mixed with a carrier, or diluted by a carrier, and/or enclosed within a carrier which may, for example, be in the form of a capsule, sachet, paper or other container. When the carrier serves as a diluent, it may be a solid, semi-solid, or liquid material, which acts as a vehicle, excipient or medium for the active ingredient. Thus, the composition may be in the form of tablets, lozenges, sachets, cachets, elixirs, suspensions, aerosols (as a solid or in a liquid medium), ointments containing for example up to 10% by weight of the active compound, soft and hard gelatin capsules, suppositories, injection solutions, suspensions, sterile packaged powders, and a topical patch.

Radiation

The source of radiation, used in combination with a VEGF receptor antagonist, can be either external or internal to the patient being treated. When the source is external to the patient, the therapy is known as external beam radiation therapy (EBRT). When the source of radiation is internal to the patient, the treatment is called brachytherapy (BT).

The radiation is administered in accordance with well known standard techniques using standard equipment manufactured for this purpose, such as AECL Theratron and Varian Clinac. The dose of radiation depends on numerous factors as is well known in the art. Such factors include the organ being treated, the healthy organs in the path of the radiation that might inadvertently be adversely affected, the tolerance of the patient to radiation therapy, and the area of the body in need of treatment. The dose will typically be between 1 and 100 Gy, and more particularly between 2 and 80 Gy. Some doses that have been reported include 35 Gy to the spinal cord, 15 Gy to the kidneys, 20 Gy to the liver, and 65-80 Gy to the prostate.

It should be emphasized, however, that the present invention is not limited to any particular dose. The dose will be determined by treating physician in accordance with particular factors in a given situation, including the factors mentioned above.

The distance between the source of the external radiation and the point of entry into the patient may be any distance that represents an acceptable balance between killing target cells and minimizing side effects. Typically, the source of the external radiation is between 70 cm and 100 cm from the point of entry into the patient.

Brachytherapy is generally carried out by placing the source of radiation in the patient. Typically, the source of radiation is placed approximately 0-3 cm from the tissue being treated. Known techniques include interstitial, intercavitary, and surface brachytherapy. The radioactive seeds can be implanted permanently or temporarily. Some typical radioactive atoms that have been used in permanent implants include iodine-125 and radon. Some typical radioactive atoms that have been used in temporary implants include radium, cesium-137, and iridium-192. Some additional radioactive atoms that have been used in brachytherapy include americium-241 and gold-198.

The dose of radiation for brachytherapy can be the same as that mentioned above for external beam radiation therapy. In addition to the factors mentioned above for determining the dose of external beam radiation therapy, the nature of the radioactive atom used is also taken into account in determining the dose of brachytherapy.

Chemotherapy

Chemotherapeutic agents include all chemical compounds that are effective in inhibiting tumor growth. The administration of chemotherapeutic agents can be

accomplished in a variety of ways including systemically by the parenteral and enteral routes. In one embodiment, the VEGF receptor antagonist and the chemotherapeutic agent are administered as separate molecules. In another embodiment, the VEGF receptor antagonist is attached, for example, by conjugation, to a chemotherapeutic agent.

Examples of chemotherapeutic agents include alkylating agents, for example, nitrogen mustards, ethyleneimine compounds and alkyl sulphonates; antimetabolites, for example, folic acid, purine or pyrimidine antagonists; mitotic inhibitors, for example, vinca alkaloids and derivatives of podophyllotoxin; cytotoxic antibiotics; compounds that damage or interfere with DNA expression.

Additionally, chemotherapeutic agents include antibodies, biological molecules and small molecules, as described herein. Particular examples of chemotherapeutic agents or chemotherapy include cisplatin, dacarbazine (DTIC), dactinomycin, mechlorethamine (nitrogen mustard), streptozocin, cyclophosphamide, carmustine (BCNU), lomustine (CCNE), doxorubicin (adriamycin), daunorubicin, procarbazine, mitomycin, cytarabine, etoposide, methotrexate, 5- fluorouracil, vinblastine, vincristine, bleomycin, paclitaxel (taxol), docetaxel (taxotere), aldesleukin, asparaginase, busulfan, carboplatin, cladribine, dacarbazine, floxuridine, fludarabine, hydroxyurea, ifosfamide, interferon alpha, leuprolide, megestrol, melphalan, mercaptopurine, plicamycin, mitotane, pegaspargase, pentostatin, pipobroman, plicamycin, streptozocin, tamoxifen, teniposide, testolactone, thioguanine, thiotepa, uracil mustard, vinorelbine, chlorambucil, taxol and combinations thereof.

The present invention also includes kits for inhibiting tumor growth comprising a therapeutically effective amount of an EGFR antagonist and a therapeutically effective amount of a VEGFR antagonist. The EGFR or VEGFR antagonist of the inventive kits can be any suitable antagonist, examples of which have been

described above. Preferably, the EGFR antagonist of the kit comprises an antibody or functional equivalent thereof, specific for EGFR. Alternatively, and also preferably, the EGFR antagonist of the kit comprises a small molecule specific for EGFR. The VEGFR antagonist of the kit preferably comprises an antibody or functional equivalent thereof, specific for VEGFR. Alternatively, the VEGFR antagonist of the kit preferably comprises a small molecule specific for VEGFR. In addition, the kits of the present invention can further comprise an antineoplastic agent. Examples of suitable antineoplastic agents in the context of the present invention have been described herein. The kits of the present invention can further comprise an adjuvant, examples of which have also been described above.

Accordingly, the receptor antibodies of the present invention can be used *in vivo* and *in vitro* for investigative, diagnostic, prophylactic, or treatment methods, which are well known in the art. Of course, it is to be understood and expected that variations in the principles of invention herein disclosed can be made by one skilled in the art and it is intended that such modifications are to be included within the scope of the present invention.

Examples

Hereinafter, the present invention will be described in further detail. It is to be understood, however, that these examples are illustrative purpose only and are not to be construed to limit the scope of the present invention.

Example 1: Establishment of KDR-Fc secreting cell line

The gene corresponding to the extracellular domains (ECDs) 1-3 of the KDR gene (accession no. AF063658 in GenBank) was amplified from a human placental cDNA library (Clonetech, USA). The amplification was carried out using the

primers KDR 1F (SEQ ID NO: 21) and KDR 3R (SEQ ID NO: 22) having *Bam*HI and *Nhe*I digestion sites respectiely.

SEQ ID NO: 21: 5'-CGC GGATCC ATGGAG AGCAA-3'

SEQ ID NO: 22: 5'-CCGCTAGC TTTTTCATGGACCCTGACA-3'

To produce a KDR(ECD1-3)-Fc chimeric protein, a pcNA3-BACE-Fc vector (Korean Patent Publication 10-2005-0032177) composed of a BACE-Fc protein gene inserted into a pcDNA3 vector (Invitrogen, USA) was digested with *Bam*HI and *Nhe*I, and then ligated with the PCR fragment digested with the same restriction enzymes. The Fc domain was amplified by PCR using the primers ThFc-F (SEQ ID NO: 23) and MycFc-R (SEQ ID NO: 24) so as to have a thrombin digestion site and a myc tag, and the amplified fragment was ligated with the vector using *Nhe*I and *Xho*I sites, thus constructing pcDNA3-KDR D123tFcm.

SEQ ID NO: 23: 5'-CCGCTAGCAGCGGCCTGGTGCCGCGCGGCAGCG ACAAAACTCAC-3':

SEQ ID NO: 24: 5'-GGCTCGAGTCACAGGTCTTCCTCAGAGATCA GC TTCTGCTCTTACCCGGAGAC-3'

The pcDNA3-KDR D123tFcm consists of a base sequence encoding amino acid residues 1-327 comprising the secretion signal sequence and extracellular domain of human KDR, a base sequence encoding a thrombin recognition site (SSGLVPRGS), a base sequence encoding 227 amino acids corresponding to the Fc domain of human immunoglobulin G (hIgG), and a base sequence (EQKLISEEDL) encoding the myc tag (FIG. 1).

For epitope mapping of antibodies, KDR (ECD1-2)-Fc (amino acid residues 1-222) and KDR(ECD2-3)-Fc (amino acid residues 1-327 (Δ24-116)) were prepared. To clone KDR (ECD1-2), the prepared sequence was amplified by PCR using a primer

KDR 1F (5'-CGC GGATCC ATGGAG AGCAA: SEQ ID NO: 25) and a primer KDR 12R (5'-CTA GCTAGC CCTAT ACCCT ACAAC GACA-3': SEQ ID NO: 26), and then the PCR amplified fragment was inserted into pcDNA3-KDR D123tFcm digested with *Bam*HI and *Nhe*I, thus preparing pcDNA3-KDR D12tFcm. To clone KDR (ECD2-3), a PCR fragment of the primer KDR 1F and the primer KDR 23SR (SEQ ID NO: 26) and a PCR fragment of the primer KDR 23SF (SEQ ID NO: 27) and the primer KDR 23R (SEQ ID NO: 28) were amplified by overlap PCR. The resulting PCR fragment was inserted into pcDNA3-KDR D123tFcm using *Bam*HI and *Nhe*I sites, thus preparing pcDNA3-KDR D23tFcm (FIG. 2).

SEQ ID NO: 26: 5'-ACA TAACCC ACAG AGGCG GCCCGGG TCTCCA-3'

SEQ ID NO: 27: 5'-GACCCGGGCCGCCTCTGTGGGTTATGTTCAAGATTACAGA-3'

SEQ ID NO: 28: 5'-CTA GCTAGC TTTTTCA TGGACCCTGACA-3'

To produce a KDR(ECD)-Fc chimeric protein, the above-prepared pcDNA3-KDR D123tFcm vector was transfected into CHO-DG44 cells (Aprogen, Korea), and the cells were cultured in α-MEM(GibCo, USA), containing 10% dFBS (Gibco, USA) and 500 μg/ml G418 (geneticin; Sigma, USA). To optimize the expression of the KDR(ECD)-Fc chimeric protein, the cells were cultured in CHO-SFM2 medium (Gibco) in the presence of MTX (methotrexate, Sigma), while the MTX concentration was increased. As a result, it was confirmed that the protein was optimally expressed at 700 nM MTX.

The produced protein was purified using protein A affinity chromatography (protein A-Sepharose, GE healthcare) and size exclusion chromatography (Hiload superdex 200, GE healthcare) and stored in 10 mM phosphate buffer (pH 7.0) containing 150 mM NaCl. FIG. 3 shows the results of SDS-PAGE of KDR(ECD1-3)-Fc purified according to the above method.

Example 2: Preparation of complete human (naïve) single chain antibody (ScFv)

phage display library

Total RNA was obtained from five healthy bone-marrow donors using TRI reagents (Sigma), and based on the total RNA, mRNA was purified using an mRNA purification kit (oligotex mRNA preparation kit, Qiagen, USA). The mRNA was treated using an RT-PCR system (ThermoScript RT-PCR system, Gibco-BRL, USA) to obtain cDNA. To obtain a VH gene, each of a V gene fragment and a DJ fragment was amplified using the primers shown in Table 1, and each of the amplified DNA fragments was amplified by 2nd PCR using primers (SEQ ID NOS: 29-61) having *Sfi*I restriction enzyme sites at the 5' end and the 3' end.

Table 1: Primer sequence for amplifying VH and DJ gene fragments

VH gene-forward		SEQ ID NO:
H05	GARGTGCAGCTGGTGGAGTC	29
H06	CAGSTGCAGCTGCAGGAGTC	30
H08	CAGGTACAGCTGCAGCAGTC	31
H09	CAGRTGCAGCTGGTGCAGTCTGGGG	32
H11	GAGGTGCAGCTGGTGCAGTCTGGAGCA	33
H12	CAGGTTCAGCTGGTGCAGTCTGGAG	34
H13	CAGGTTCAGCTGGTGCAGTCTGGGG	35
H14	CAGGTCCAGCTGGTACAGTCTGGGG	36
H15	CAGGTCACCTTGAAGGAGTCTGGTCCTGT	37
H16	CAGATCACCTTGAAGGAGTCTGGTCCTAC	38
H17	CAGGTCACCTTGAGGGAGTCTGGTCCTGC	39
H25	GAGGTGCAGCTGGTGGAGTCTGGGGGAGGTG	40
H32	CAGGTGCAGCTACAGCAGTGGGGCG	41
VH gene-back		
H210	AATACACGGCCGTGTCCTCAGATC	42
H210L	AATACACGGCCGTGTCCTCAGATCTCAGGCTG CTCAGCTCCATGTAGGCTGAG	43
H211	AGCTCCATGTAGGCTGTGTCT	44
H212	AGCTCCATGTAGGCTGTGCTCATAGACC	45
H213	AGCTCCATGTAGGCTGTGCTTGTGGACA	46
H214	AGCTCCATGTAGGCTGTGCTTATGGAG	47
H220	AAGGACCACCTGCTTTTGGAGG	48
H230	AATACACGGCCGTGTCCTCGGCTCTCAGACTG TTCATT	49
H240	AATACACGGCCTGTCCACGGCGG	50
H250	AATACATGGCGGTGTCCGAGGCCT	51
DJ gene-forward		
CDR3-1	GATCTGAGGACACGGCCGTGTATTACTGT	52

CDR3-2	CCTCCAAAAGCCAGGTGGTCCTT	53
CDR3-3	GAGCCGAGGACACGGCCGTGTATTACTGT	54
CDR3-4	CCGCCGTGGACACGGCCGTGTATTACTGT	55
CDR3-5	AGGCCTCGGACACCGCCATGTATTACTGT	56
DJ gene-back		
JH-U1	CTGAGGAGACGGTGACC	57
V-DJ fusion		
H48SfiI-for	GCGATGGCCCAGCCGGCCATGGCCCAGRTGCA GCTGGTRSAGTC	58
H49SfiI-for	GCGATGGCCCAGCCGGCCATGGCCCAGRTCAC CTTGARGGAGTC	59
H50SfiI-for	GCGATGGCCCAGCCGGCCATGGCCCAGGTRCA GCTRCAGSAGT	60
H47SfiI-back	GGAATTCGGCCCCCGAGGCCTGARGAGACRGT GACC	61

cf.) R: A or G; S: C or G

To obtain a VL gene, 1st PCR was performed using each of primers (Table 2) for lambda gene amplification and primers (Table 3) for kappa gene amplification, and each of the amplified fragments was subjected to 2nd PCR using primers (lambda: SEQ ID NOS. 76-81, and kappa: SEQ ID NOS. 106-108) having a *Bst*XI digestion site at the 5' end and the 3' end.

Table 2: Primer sequence for amplifying Lambda gene fragment

Vλ forward		SEQ ID NO:
L01	CAGYCTGTGCTGACTCAG	62
L03	CAGCCTGTGCTGACTCAAT	63
L06	TCCTATGAGCTGACWCAG	64
L15	CAGYCTGTGCTGACTCAGCCGT	65
L20	CAGTCTGTGCTGACGCAGCCG	66
L23	CAGTCTGCCCTGACTCAGCCTC	67
L24	CAGTCTGCCCTGACTCAGCCTG	68
L25	CAGRCTGTGGTGACYCAGGAGCCCTCAC	69
L26	CAGRCTGTGGTGACYCAGGAGCCATCGT	70
L28	TCCTATGAGCTGACWCAGCCACT	71
L34B	AATTTTATGCTGACTCAGCCC	72
Vλ back		
L35	CCTCCTCCACCTAGGACGGTGACCTTGG TCCCAGTT	73
L36	CCTCCTCCACCTAGGACGGTCAGCTTGG TCCCTCCG	74
L37	CCTCCTCCACCGAGGGCGGTCAGCTGGG TGCCTCCT	75

Vλ 2nd PCR (*Bst*XI)		
L34BstXI-for	GGTGGATCCAGCGGTGTGGGTTCCAATT TTATGCTGACTCAGCCC	76
L40BStXI-for	GGTGGATCCAGCGGTGTGGGTTCCCAGY CTGTGCTGACTCAGCC	77
L41BstXI-for	GGTGGATCCAGCGGTGTGGGTTCCCAGC CTGTGCTGACTCAATC	78
L42BstXI-for	GGTGGATCCAGCGGTGTGGGTTCCCAGT CTGCCCTGACTCAGCC	79
L43BstXI-for	GGTGGATCCAGCGGTGTGGGTTCCCAGR CTGTGGTGACYCAGGA	80
L44BstXI-for	GGTGGATCCAGCGGTGTGGGTTCCTCCT ATGAGCTGACWCAG	81
L38BstXI-back	GAATTCCACGAGGCTGGCTCCTCCACCK AGGRCGGT	82

cf) K: G or T; R: A or G; Y: T or C; W: A or T

Table 3: Primer sequence for amplifying Kappa gene fragment

Vκ forward		SEQ ID NO:
K12	GACATCCAGATGACCCAGTCTCCATCCTCCC	83
K13	GACATCCAGATGACCCAGTCTCCATCCTCA	84
K14	GACATCCAGATGACCCAGTCTCCATCTTCYG	85
K15	GACATCCAGATGACCCAGTCTCCTTCCA	86
K16	AACATCCAGATGACCCAGTCTCCATCTGCCA	87
K17	AACATCCAGATGACCCAGTCTCCATCCTT	88
K18	GCCATCCAGTTGACCCAGTCTCCAT	89
K19	GCCATCCGGATGACCCAGTCTCCATTCTCC	90
K20	GTCATCTGGATGACCCAGTCTCCATCCTTA	91
K21	GATATTGTGATGACCCAGACTCCACTCTCTCTGT	92
K22	GATATTGTGATGACCCAGACTCCACTCTCCCTGC	93
K23	GATATTGTGATGACCCAGACTCCACTCTCCTCA	94
K24	GATRTTGTGATGACTCAGTCTCCACTCTC	95
K25	GAAATTGTGTTGACRCAGTCTCCAG	96
K27	GACATCGTGATGACCCAGTCTCCAG	97
VKA1	GATGTTGTGATGACTCAGTCTCCACTCTCCCTGCC CGTCACCCTTGGAC	98
VK10	GAAATTGTGCTGACTCAGTCTCCAGACTTT	99
VK30	GACATCCAGATGACCCAGTCTCCATCCTCCCTGTC TGCATCTGTAGGAG	100
Vκ back		
K28	TCCTCCACGTTTGATTTCCACCTTGGTCCCTTG	101
K29	TCCTCCACGTTTGATCTCCAGCTTGGTCCCC	102
K30	TCCTCCACGTTTGATATCCACTTTGGTCCCAG	103
K31	TCCTCCACGTTTGATCTCCACCTTGGTCCCTCC	104
K32	TCCTCCACGTTTAATCTCCAGTCGTGTCCCT	105
Vk *Bst*XI		

K33BstXI-for	GGTGGATCCAGCGGTGTGGGTTCCGACATCCAGA TGACCCAGTCTCC	106
K34BstXI-for	GGTGGATCCAGCGGTGTGGGTTCCGATATTGTGA TGACCCAGWCTCC	107
K36BstXI-back	GAATTCCACGAGGCTGGCTCCTCCACGTTTGATH TCCA	108

cf) H: A, C or T; W: A or T

To introduce the VH gene fragment and the VL gene fragment into a phagemid vector, a pAK100 vector (Krebber, A. *et al.*, *J. Immunol. Method.*, 201:35, 1997) was used. To introduce the VL gene fragment, three *Bst*XI domains (236, 365 and 488) present in the lac repressor gene (*lacI*) of the pAK100 vector were mutated using the Quikchange site-specific mutagenesis kit (Stratagene, USA). Using the modified pAK100 vector, to prepare a backbone vector for the construction of the ScFv library, the heavy chain V gene, amplified using the H05 primer (SEQ ID NO: 29) and the H230 primer (SEQ ID NO: 49), and the DJ gene fragment, amplified using the CDR3-3 primer (SEQ ID NO: 54) and the JH-U1 primer (SEQ ID NO: 57), were subjected to 2nd PCR with H48*Sfi*I (SEQ ID NO: 58)/H47*Sfi*I (SEQ ID NO: 61), and the resulting heavy chain variable region was digested with *Sfi*I and ligated into the modified pAK100 vector digested with the same enzyme. To introduce a light chain and a linker, the heavy chain region was amplified using primers (forward: SEQ ID NO: 109; and backward: SEQ ID NO: 110), and the light chain sequence of the human 4-1BB antibody (LB506) (Korean Patent Publication 2000-0034847) was amplified using each of primers (forward: SEQ ID NO: 111; and backward: SEQ ID NO: 112). The amplified fragments were inserted into the modified pAK100 vector having the heavy chain variable domain introduced therein, using *Xba*I/*Eco*RI, thus preparing an antibody library backbone vector.

SEQ ID NO: 109: 5'-CGAATTTCTAGATAACGA-3'

SEQ ID NO: 110: 5'-CCTCCGCCACTACCTCCTCCTCCGAGGCCCCCGAGGCCTGA-3'

SEQ ID NO: 111: 5'-GGTAGTGGCGGAGGAGGCTCCGGTGGA TCCAGCGGTGTGG GTTCCGATATTGTG-3'

SEQ ID NO: 112: 5'-CTCGAATTCCCACGAGGCTGGCTCCTCCACGTTTGATTTC-3'

In order to introduce light chain variable regions, each of amplified light chain (κ, λ) variable regions was digested with *Bst*XI and inserted into the antibody library backbone vector. The resulting plasmid was digested with a *Sfi*I restriction enzyme and ligated with a heavy chain variable region-amplified PCR fragment previously digested with *Sfi*I. The ligated plasmid was transfected into ElectroTen-Blue competent cells (Stratagene, USA). As a result, a ScFv phage library having a diversity of about 10^{11} was collected from the colony.

Example 3: Biopanning

The library stock constructed in Example 2 was grown to the log phase and rescued with the M13K07 helper phage (GE healthcare, USA). The resulting library was amplified in 2xYT medium (2xYT/C/K; containing 34 μg/ml of chloramphenicol and 70 μg/ml of kanamycin and supplemented with 1 mM IPTG) at 30℃ overnight.

Phage stock was precipitated in 20% PEG6000/2.5 M NaCl and resuspended in PBS. Resuspened phage stock was incubated in 2% skimmed milk/PBS solution containing 500 ug/ml of a human Fc protein at 37℃ for 1 hour in order to remove phages showing anti-human Fc.

The KDR (human VEGFR-2) used as an antigen was KDR(ECD1-3)-Fc comprising IgG-like domains 1, 2 and 3 of the extracellular domain of KDR. The KDR(ECD1-3)-Fc stable cell line prepared in Example 1 was cultured, and KDR(ECD1-3)-Fc was purified from the cultured cell line.

Maxisorb Star tubes (Nunc, Denmark) coated with KDR(ECD1-3)-Fc (10 ㎍/㎖) were first blocked with 2% skimmed milk/PBS at room temperature for 2 hours, and then inoculated with 5.4 x 10^{12} pfu of the phage stock at room temperature for 1

hour. The tubes were washed 10 times with PBST (PBS containing 0.1% Tween 20), and then washed 10 times with PBS. The bound phage was eluted with 1 ml of 100 mM fresh triethylamine solution at room temperature for 10 minutes. The eluted phage was left to stand together with 10 ml of mid-log-phase XL1-Blue cells at 37℃ for 30 minutes, and then shake-cultured for 30 minutes. Then, the infected XL1-Blue cells were cultured in a 1% glucose-containing 2x YT/C plate at 30℃ overnight. Following the first panning, the second and third panning processes were performed by coating KDR(ECD1-3)-Fc into a 96-well plate (Nunc, USA) instead of the maxisorp tube. After the third panning was performed, the KDR neutralizing ability of the obtained phage was analyzed through VEGF competition assays.

For VEGF competition assays, a microplate coated with 200 ng of VEGF165 (R&D system) overnight was allowed to react with 2% skimmed milk/PBS at 37℃ for 2 hours. The microplate was washed with PBS, and then a mixture, obtained by reacting 10 ng of KDR (ECD1-3)-Fc with various amounts of phage at room temperature for 1 hour, was placed in each well of the plate and allowed to react at room temperature for 2 hours. The reaction solution was washed with PBS, allowed to react with a rabbit anti-KDR antibody (Reliatech, Germany) at 37℃ for 1 hour, and allowed to react with an HRP (horse radish peroxidase)-conjugated goat anti-rabbit antibody (Abcam, UK) at 37 ℃ for 1 hour. After the completion of the reaction, each well was color-developed with a TMB solution (Sigma), and then measured for absorbance at 450 nm (FIG. 4).

As a result, it was seen that 6A6, 6H1, 6G1 and 6C1 could all inhibit the binding of VEGF to KDR, and among them, 6A6 and 6H1 showed the highest ability to neutralize VEGF. Also, 6A6 and 6H1 were shown to have a binding affinity similar to that of a reconstructed 1C11 (hereinafter referred to as 1C11) phage obtained in Example 4. The DNA sequences, amino acid sequences and CDR sequences of 6A6 (TTAC-0001) ScFv are shown in FIG. 5.

Also, the base sequences and amino acid sequences of 6A6 (TTAC-0001) ScFv were expressed as heavy chain CDR 1 (SEQ ID NO: 113 and SEQ ID NO: 114), heavy chain CDR 2 (SEQ ID NO: 115 and SEQ ID NO: 116), heavy chain CDR 3 (SEQ ID NO: 117 and SEQ ID NO: 118), light chain CDR 1 (SEQ ID NO: 119 and SEQ ID NO: 120), light chain CDR 2 (SEQ ID NO: 121 and SEQ ID NO: 122), light chain CDR 3 (SEQ ID NO: 123 and SEQ ID NO: 124), heavy chain variable regions (SEQ ID NO: 125 and SEQ ID NO: 20), light chain variable regions (SEQ ID NO: 126 and SEQ ID NO: 1), IgG heavy chain regions (SEQ ID NO: 127 and SEQ ID NO: 128), and IgG light chain regions (SEQ ID NO: 129 and SEQ ID NO: 130).

Example 4: Construction of reconstructed IMC-1121(rIMC-1121) and IMC-1C11(rIMC-1C11) phage vectors

In order to obtain IMC-1C11 ScFv (PCT/US2001/10504) and IMC-1121 ScFv (PCT/US2002/006762) phage particles (Imclone) to be used as positive control groups, the ScFv region of each antibody was cloned into the pAK vector.

For IMC-1C11, a light chain variable gene was cloned using, as a template, a pTA-d9-07 clone (LG Life Sciences) obtained from a mouse na△ve antibody library (LG Life Sciences). The clone was amplified by PCR using the LR and LF primers shown in Table 4, and the amplified light chain variable gene was digested with *Bst*XI and ligated into a library backbone vector pretreated with *Bst*XI. A heavy chain variable gene was amplified by PCR using a pTA-A5N2-10 clone (LG Life Sciences) as a template with the primers shown in Table 4. After each of the PCR reactions was performed using each of the primer pairs HF1-RI(A), HF2-HR2(B), HF3-HR3(C) and HF4-HR4(D), and each of the amplified fragments was amplified by overlap PCR using A-B (HF1-HR2 primer set) and C-D (HF3-HR4 primer pair), and then amplified by overlap PCR using A-B-C-D (HF1-HR4 primer

pair). Then, each of the amplified fragments was treated with *Sfi*I and ligated with the 1C11 light chain gene-containing library backbone vector treated with *Sfi*I. Table 4 shows the PelB signal sequence and DNA sequence to amber (TGA) codon of the phage vector (pAK-r1c11).

5

Table 4: LR and LF primer

Name	Sequence	SEQ ID NO:
pTA-d9_07 light chain	GACATTGTTC TCATCCAGTC TCCAGCAATC ATGTCTGCAT CTCCAGGGGA GAAGGTCACC ATAACCTGCA GTGCCAGCTC AAGTGTAAGT TACATGCACT GGTTCCAGCA GAAGCCAGGC ACTTCTCCCA AACTCTGGAT TTATAGCACA TCCAACCTGG CTTCTGGAGT CCCTGCTCGC TTCAGTGGCA GTGGATCTGG GACCTCTTAC TCTCTCACAA TCAGCCGAAT GGAGGCTGAA GATGCTGCCA CTTATTACTG CCAGCAAAGG AGTAGTTACC CATTCACGTT CGGCTCGGGG ACAAAGTTGG AAATAAAA	131
pTA-A5N^2-10 heavy chain	CAGGTTCAGC TCCAGCAGTC TGGGGCAGAG CTTGTGAGGT CAGGGGCCTC AGTCAAGTTG TCCTGCACAG CTTCTGGCTT CAACATTAAA GACTACTATA TGCACTGGGT GAAGCAGAGG CCTGAACAGG GCCTGGAGTG GATTGGATGG ATTGATCCTG CGAATGGTAA TACTAAATATGACCCGAAGT TCCAGGGCAA GGCCACTATA ACAGCAGACA CATCCTCCAA CACAGCCTAC CTGCAGCTCA GCAGCCTGAC ATCTGAGGAC ACTGCCGTCT ATTACTGTGC TAGATGGGAC TGGTACTTCG ATGTCTGGGG CGCAGGGACC ACGGTCACCG TTTCC	132
LF	CTGCAGAACC AGCGGTGTGG GTTCCGACAT CGAGCTCACT CAGTCTCCAT G	133
LR	CTGCAGAACC ACGAGGCTGG CTCCTCCACG TTTTATTTCC AGCTTGGTCC CCG	134
HF1	CGGCCCAGCC GGCCATGGCC CAGGTCAAGC TGCAGCAGTC TGGGGCAGAG CTTGTGGGGT CAGGGGCC	135
HF2	GGCTTCAACA TTAAAGACTT CTATATGCA	136
HF3	GATTATGCCC CGAAGTTCCA GGGCAAGGCC ACCATGACTG CAGACTCATC CTCCA	137
HF4	TACTGTAATG CATACTATGG TGACTACGAA GGCTACTGGG GCCAA	138
HR1	GTCTTTAATG TTGAAGCCAG AAGTTGTGCA G	139
HR2	ACTTCGGGGC ATAATCAGAA TCACCATTCT CAGGATCAAT CCATCCAATC	140
HR3	GTATGCATTA CAGTAATAG	141
HR4	CCGAGGCCCC CGAGGCCTGA GGAGACGGTG ACCGTGGTCC CTTGGCCCCA GTAGCCTTCG TA	142
r1C11-ScFv DNA	ATGAAATACC TATTGCCTAC GGCAGCCGCTGGATTGTTAT TACTCGCGGC CCAGCCGGCC ATGGCCCAGG TCAAGCTGCA GCAGTCTGGG GCAGAGCTTG TGGGGTCAGG GGCCTCAGTC AAATTGTCCT GCACAACTTC TGGCTTCAAC ATTAAAGACT TCTATATGCA CTGGGTGAAG CAGAGGCCTG AACAGGGCCT GGAGTGGATT GGATGGATTG TCCTGAGAA TGGTGATTCT GATTATGCCC CGAAGTTCCA GGGCAAGGCC ACCATGACTG CAGACTCATC CTCCAACACA GCCTACCTGC AGCTCAGCAG CCTGACATCT GAGGACACTG CCGTCTATTA CTGTAATGCA TACTATGGTG ACTACGAAGG CTACTGGGGC CAAGGGACCA CGGTCACCGT CTCCTCAGGC CTCGGGGGCC TCGGAGGAGG AGGTAGTGGC GGAGGAGGCT CCGGTGGATC CAGCGGTGTG GGTTCCGACA TCGAGCTCAC TCAGTCTCCA GCAATCATGT	143

	CTGCATCTCC AGGGGAGAAG GTCACCATAA CCTGCAGTGC CAGCTCAAGT GTAAGTTACA TGCACTGGTT CCAGCAGAAG CCAGGCACTT CTCCCAAACT CTGGATTTAT AGCACATCCA ACCTGGATTA TGGAGTCCCT GCTCGCTTCA GTGGCAGTGG ATCTGGGACC TCTTACTCTC TCACAATCAG CCGAATGGAG GCTGAAGATG CTGCCACTTA TTACTGCCAG CAAAGGAGTA GTTACCCATT CACGTTCGGC TCGGGGACCA AGCTGGAAAT AAAACGTGGA GGAGCCAGCC TCGTGGAATT CGAGCAGAAG CTGATCTCTG AGGAAGACCT GTAG	

In order to obtain IMC-1121, 6G1 was used as a template to clone a light chain variable region. The 6G1 template was amplified by PCR using the primer pairs LF-KR1(A), LF1-LR2(B), LF2-LR3(C) and LF3-LR4(D), was amplified by overlap PCR using A-B (LF-LR2 primer set) and C-D (LF2-LR primer pair), and was then amplified by overlap PCR using A-B-C-D (LF-LR primer pair). The obtained PCR fragment was treated with *Bst*XI and inserted into the library backbone vector (reconstructed IMC-1121; hereinafter referred to as IMC-1121). The primers used herein are shown in Table 5.

For the heavy chain variable region, the YGKL-136 clone having a sequence closest to IMC-1121 among the clone sequences obtained from the human na△ve scFv library (Example 2) was used as a template. The YGKL-136 clone was amplified by PCR using each of the primer pairs HF-HR1(A), HF1-HR2(B) and HF2-HR(C), was subjected to A+B overlap PCR (HF-HR2 primer pair), and was then subjected to A+B and C overlap PCR (HF-HR primer pair). The produced PCR fragment was treated with *Sfi*I and ligated into a light chain-containing library backbone vector.

Table 5: LF and HF primer for cloning light chain variable region

Name	Sequence	SEQ ID NO:
YGKL-136 heavy chain (template)	GAGGTGCAGC TGGTGGAGTC TGGGGGAGGC CTGGTCAAGC CTGGGGGGTC CCTGAGACTC TCCTGTGCAG CCTCTGGATT CACCTTCAGT AGCTATAGCA TGAACTGGGT CCGCCAGGCT CCAGGGAAGG GGCTGGAGTG GGTCTCATCC ATTAGTAGTA GTAGTAGTTA CATACACTAC GCAGACTCAG TGAAGGGCCG ATTCACCATC TCCAGAGACA ACGCCAAGAA CTCACTGTAT CTGCAAATGA ACAGTCTGAG AGCCGAGGAC ACGGCCGTGT ATTACTGTGC GAGAGTCACA GATGCTTTTG ATATCTGGGG CCCCGGAACC CTGGTCACCG TCTCCTCA	144
6G1 light chain (template)	GACATCCAGA TGACCCAGTC TCCATCTTCC GTGTCTGCAT CTGTAGGAGA CAGAGTCACC ATCACTTGTC GGGCGAGTCA GGGTATTAGC AGCTATTTAG GCTGGTATCA GCAGAAACCA GGGAAAGCCC CTAAGCTCCT GATCTATGCT GCATCCAATT TGCAAACAGG GGTCCCGCCA AGGTTCAGCG GCAGTGGATC CGGGACAAGT TTCACTCTCA CCCTCAATAA TGTGCAGCCT GAAGATTCTG CAACTTACTA TTGTCAACAG GCTGACAGTT TCCCTCTTTC GGCGGAGGGA CCAAAGTGGA AATCAAACGT GAGGAGCC	145
LF primer	CCCCAGCGGT GTGGGTTCCG ACA	146
LR1 primer	TGGTGACTCT GTCTCCTATA GATGCAGACA CGGATGAT	147
LF1 primer	TCTATAGGAG ACAGAGTCAC CA	148
LR2 primer	TACCAGCCTA ACCAGTTGTC AATACCCTGA CTCGCCCG	149
LF2 primer	TTGACAACTG GTTAGGCTGG TATCAGCAGA AACCAGGG AAA	150
LR3 primer	ACCTTGATGG GACCCCTGTG TCCAAATTGG ATGCATCATA GATCAGGAGC TT	151
LR primer	CCCCACGAGG CTGGCTCCTC CA	152
HF primer	CCGGCCCAGC CGGCCATGGC CGAGGTGCAG CTGGTGCAGT CTGGGGGAGG CCTGGTCA	153
HF1 primer	GTAGTAGTAG TAGTTACATA TACTACGCAG ACTCAGTGA	154
HF2 primer	TTACTGTGCG AGAGTCACAG ATGCTTTTGA TATCTGGGGC CAAGGGACAA	155
HR1 primer	TCACTGAGTC TGCGTAGTAT ATGTAACTAC TACTACT	156
HR2 primer	CTGTGACTCT CGCACAGTAA TACA	157
HR primer	CCGGCCCCCG AGGCCTGAGG AGACGGTGAC CATTGTCCCT TGGCCCCAG	158
r1121-ScFv DNA	ATGAAATACC TATTGCCTAC GGCAGCCGCT GGATTGTTAT TACTCGCGGC CCAGCCGGCC ATGGCCGAGG TGCAGCTGGT GCAGTCTGGG GGAGGCCTGG TCAAGCCTGG GGGGTCCCTG AGACTCTCCT GTGCAGCCTC TGGATTCACC TTCAGTAGCT ATAGCATGAA CTGGGTCCGC CAGGCTCCAG GGAAGGGGCT GGAGTGGGTC TCATCCATTA GTAGTAGTAG TAGTTACATA TACTACGCAG ACTCAGTGAA GGGCCGATTC ACCATCTCCA GAGACAACGC CAAGAACTCA CTGTATCTGC AAATGAACAG TCTGAGAGCC GAGGACACGG CCGTGTATTA CTGTGCGAGA GTCACAGATG CTTTTGATAT CTGGGGCCAA GGGACAATGG TCACCGTCTC CTCAGGCCTC GGGGGCCTCG GAGGAGGAGG	159

	TAGTGGCGGA GGAGGCTCCG GTGGATCCAG CGGTGTGGGT TCCGACATCC AGATGACCCA GTCTCCATCT TCCGTGTCTG CATCTATAGG AGACAGAGTC ACCATCACTT GTCGGGCGAG TCAGGGTATT GACAACTGGT TAGGCTGGTA TCAGCAGAAA CCTGGGAAAG CCCCTAAACT CCTGATCTAC GATGCATCCA ATTTGGACAC AGGGGTCCCA TCAAGGTTCA GTGGAAGTGG ATCTGGGACA TATTTTACTC TCACCATCAG TAGCCTGCAA GCTGAAGATT TTGCAGTTTA TTTCTGTCAA CAGGCTAAAG CTTTTCCTCC CACTTTCGGC GGAGGGACCA AGGTGGACAT CAAACGTGGA GGAGCCAGCC TCGTGGAATT CGAGCAGAAG CTGATCTCTG AGGAAGACCT GTGA	

Example 5: Production and purification of soluble ScFv

To prepare soluble 6A6 ScFv, pAK-6A6, having a pelB sequence and an ScFv sequence, was digested with *Eco*RI and *Xba*I to obtain a fragment, having the pelB sequence and the ScFv sequence. The fragment was inserted into the pET21b vector (Novagen, USA) using the same restriction enzymes. To add an myc tag to the vector inserted with the fragment having the pelB sequence and the ScFv sequence, the pET21b vector, inserted with the pelB sequence and the ScFv sequence, as a template, was amplified by PCR using primers (mycFor: SEQ ID NO: 160, and mycRev: SEQ ID NO: 161), and the PCR fragment was ligatged into the vector having the pelB sequence and the ScFv sequence, using *Eco*RI and *Xho*I, thus constructing pET21b-KDR 6A6.

SEQ ID NO: 160: 5'-GAGCCAGCCTCGTGGAATTCGAACAAAAA-3'

SEQ ID NO: 161: 5'-TGCTCGAGAT TCAGATCCTC TTCTGAGATG AGTTTTTGTT GAATTCCACG AGGCT-3'

For kinetic measurement, a V5 tag sequence (GKPIPNPLLGLDST) was inserted in an *Xho*I site downstream of the myc tag in the following manner. The resulting sequence was amplified by PCR using primers (V5-For: SEQ ID NO: 162, and V5-Rev: SEQ ID NO: 163) amplifying the *Eco*RI and V5 tag sequence-containing *Xho*I digestion sites of the pET21b-KDR 6A6, and the amplified frasgment was digested

with *Eco*RI and *Xho*I and ligated into the pET21b-KDR 6A6 digested with the same restriction enzymes, thus constructing pETV-KDR 6A6. The constructed pETV-KDR 6A6 was transformed into *E. coli* BL21(DE3).

SEQ ID NO: 162: 5'-CCAGCCTCGTGGAATTC GAAC-3'

SEQ ID NO: 163: 5'-CCGCTCGAG GGTGGAGTC CAGACCTAATAG AGGGTT TGGGATCGG CTTTCCATTCAGATC CTCTTCTGA-3'

The *E. coli* BL21(DE3) cells transformed with the pETV-KDR 6A6 were cultured to express the soluble ScFv protein and centrifuged, and a periplasmic fraction was collected from the cells using 50 mM Tris (pH 8.0) solution containing 20% sucrose and 200 μg/ml of a lysozyme and protease inhibitor cocktail (Roche, Swiss). The obtained fraction was purified using Ni-NTI affinity chromatography (Hisprep, GE healthcare, USA) and ion exchange chromatography (Q-sepharose, SP-sepharose, GE healthcare, USA), thus obtaining an ScFv protein.

For 6A6, the Hisprep column was equilibrated using a solution containing 20 mM imidazole, 0.4M NaCl and 1x PBS, and the periplasmic fraction was placed in the column and eluted with 300 mM imidazole-containing solution (300 mM imidazole, 0.4M NaCl/1xPBS). The eluted protein was dialyzed with 50 mM imidazole (pH 6.7) solution, and then eluted using cation exchange chromatography while increasing the concentration of NaCl to 0.5M. The eluted protein was concentrated (centriprep YM10, milipore, USA), and then dialyzed and stored in PBS solution. FIG. 6 shows the results of SDS-PAGE of the purified 6A6 ScFv.

Example 6: VEGF competition assays with VEGF

In order to examine whether the isolated ScFv can inhibit the binding of KDR to VEGF, competition assays were performed. For this purpose, 20 ng of VEGF165 was coated into a 96-well microtiter plate at room temperature overnight, and then

allowed to react with 2% skimmed milk/PBS at 37℃ for 2 hours. After the completion of the reaction, the plate was washed with PBS, and then mixture solutions, obtained by allowing 100 ng of Fc-digested KDR(ECD1-3) to react with various amounts of ScFv at room temperature for 1 hour, were placed in the microtiter plate and allowed to react at room temperature for 2 hours. After the completion of the reaction, the plate was washed with PBS, and then an anti-KDR mouse antibody (5 μg/ml, Reliatech, Germany) was added thereto and allowed to react at 37℃ for 1 hour. Then, a 1:5000 dilution of an HRP-conjugated goat anti-mouse antibody (Abcam, UK) was added thereto and allowed to react for 1 hours, and then TMB solution was added thereto and allowed to react. Then, the cells in each well of the plate were measured for absorbance at 450 nm and 650 nm. FIG. 7 shows the results of VEGF competitive assays with the anti-KDR-ScFv purified in Example 5. As shown in FIG. 7, it can be seen that only 6A6-ScFv shows a potent ability to neutralize KDR.

Example 7: Epitope mapping with anti-KDR ScFv antibody

In order to examine which anti-KDR ScFv antibodies bind to which domain of extracellular domains 1-3 of KDR, 3 μg/ml of each of the KDR(ECD1-2), KDR(ECD2-3) and KDR(ECD 1-3)-Fc prepared according to the method of Example 1 was coated into a 96-well plate by reaction at 37℃ for 2 hours. After the completion of the reaction, the plate was washed with PBS, and then the portion of the plate, which has not been coated with the KDR protein, was blocked with 2% skimmed milk/PBS. Then, the plate was washed again with PBS, and then 330 nM anti-KDR ScFv antibody was added thereto and allowed to react at 37℃ for 1 hour and 30 minutes. After the completion of the reaction, the plate was washed again with PBS, and then a 1:500 dilution of an HRP-conjugated rabbit anti-6xHis antibody (Abcam, UK) was added thereto and allowed to react at 37℃ for 1 hour. Then, the cells in each well were color-developed with TMB solution and measured for absorbance at 450 nm.

As a result, it could be seen that 6A6 was bound to the extracellular domain 3 of KDR in the same manner as IMC-1121 (FIG. 8). However, 6G1 and 6C1 were more strongly bound to the domain 1, even though the absorbance was low.

Example 8: Expression and purification of IgG

For expression in the form of whole IgG, heavy chain and light chain expression vectors, each comprising a whole constant region, were prepared. For the heavy chain expression vector, a pIgGHD vector (Aprogen, Korea) having a heavy chain backbone of human 4-1 bb was treated with *Sfi*I, and then ligated with a fragment, obtained by treating the heavy chain variable region of pAK-ScFv with *Sfi*I, thus constructing an expression vector pIgGHD-6A6Hvy, comprising a whole constant region and a heavy chain region (FIG. 9).

For the light chain expression vector, a pIgGLD vector (Aprogen, Korea) having a light chain backbone of human 4-1 bb was treated with *Bst*XI, and then ligated with a fragment, obtained by treating the light chain variable region of pAK-ScFv with *Bst*XI, thus constructing an expression vector pIgGLD-6A6Lgt, comprising a whole constant region and a light chain region (FIG. 10). In the case of IMC-1C11 and 1121, IgG expression vectors were constructed in the same manner as described above.

For the expression of IgG, the same amount of the light chain expression vector (pIgGHD-6A6Hvy for the 6A6 clone) and the heavy chain expression vector (pIgGLD-6A6Lgt for the 6A6 clone) were co-transfected into CHO DG44 cells (Aprogen, Korea). The co-transfected cells were cultured in α-MEM medium containing 10% dFBS and 500 μg/ml of G418, and then a clone having the highest protein expression level was selected, while MTX was added thereto at a concentration ranging from 10 nM to 700 nM.

For the expression of antibodies, the cells were cultured in CHO-SF2 medium, containing 700 nM MTX, at 37℃, and the culture was collected. 6A6-IgG was purified from the combined supernatant by affinity chromatography using a protein A column (GE healthcare, USA) according to the manufacturer's protocol. The supernatant was poured into the protein A column equilibrated with a solution containing 20 mM sodium phosphate (pH 7.0) and 100 mM NaCl and was washed with a solution containing 20 mM sodium phosphate (pH 7.0), 1 mM EDTA and 500 mM NaCl. Then, the protein was eluted with a 0.1 M glycine-HCl (pH 3.3) solution containing 100 mM NaCl. The eluted protein was neutralized with 1 M Tris. The eluted protein was mixed with 5 mM sodium phosphate buffer (pH 6.0) at a ratio of 1:1, and then poured into a prepacked SP-Sepharose column (GE healthcare) equilibrated with 5 mM sodium phosphate (pH 6.0) containing 50 mM NaCl. The protein bound to the column was eluted with a sodium phosphate buffer (pH 7.0) containing 50 mM NaCl and was poured into a prepacked Q-sepharose column (GE healthcare) equilibrated with an elution buffer, and unbound protein was collected. The collected protein was concentrated with 30 Kd vivaspin 20 (Sartorius) and dialyzed with PBS. FIG. 11 shows the results of SDS-PAGE of the 6A6 IgG protein purified according to the above-described method.

Example 9: Competition assays of anti-KDR IgG with various VEGFs

Competition assays of anti-KDR IgG with VEGF were carried out in the same manner as the VEGF competition assays of KDR ScFv, conducted in Example 6 using VEGF165. As a result, 6A6 IgG among anti-IgGs showed the highest ability to neutralize KDR, similar to the results of the competition assays conducted with ScFv, and it showed KDR neutralizing ability similar to that of the IMC-1121 anti-KDR IgG reconstructed on the basis of the amino acid sequence (FIG. 12).

Also, in order to examine the binding and competition of 6A6 IgG with isotypes and

VEGF families other than VEGF165, 200 ng of each of VEGF121, VEGF165, VEGF-C, VEGF-D and VEGF-E was coated into a 96-well plate, and then competition assays were carried out in the same manner as in Example 6. As a result, 6A6 IgG showed VEGF neutralizing ability by binding to VEGF121, VEGF165 and VEGF-E, which belong to VEGF-A, and it did not bind to VEGF-C and VEGF-D (FIG. 13).

Example 10: Analysis of binding affinity of anti-KDR ScFv and IgG

The binding affinity of the antibodies to KDR (VEGFR-2) was measured with BIAcore (GE healthcare). In the case of ScFv, KDR(ECD1-3)-Fc was immobilized onto a CM5 chip (GE healthcare, Sweden) according to the manufacturer's manual, and in the case of V5-tagged ScFv, a V5 antibody (Abchem, UK) was immobilized onto the chip. The V5-tagged ScFv was bound to the CM5 chip having the V5 antibody immobilized thereon, and then Fc-free KDR(ECD1-3) was allowed to run on the chip surface, thus obtaining sensorgrams. In the case of IgG, as in the case of the ScFv having no V5, KDR(ECD1-3)-Fc was immobilized onto the CM5 chip, and then various amounts of the antibody was allowed to run on the chip surface, thus obtaining sensorgrams. Based on the sensorgram obtained at each concentration, the kinetic constants k_{on} and k_{off} were measured, and K_d was calculated from the ratio of the kinetic constants k_{off}/k_{on} (Table 6).

As a result, it was confirmed that, among various ScFvs, ScFv having a high binding affinity for KDR was 6A6. Also, when 6A6 was converted in the form of IgG, the Kd value thereof was about 2-fold lower than that of IMC-1121. This suggests that 6A6 IgG was more strongly bound to KDR compared to IMC-1121.

Table 6: K_d (M) value of anti-KDR ScFv, IgG

	ScFv	ScFv-V5	IgG		
			k_{on} (1/Ms)	k_{off} (1/s)	K_d (M)
6A6	1.11E-08	6.93E-09	**3.17E+05**	**7.3E-05**	**2.3E-10**
6H1	N/A	N/A	5.02E+04	7.20E-03	1.43E-07
6G1	4.11E-07	3.31E-08	9.06E+04	5.48E-03	6.05E-08
6C1	4.31E-08	N/A	1.38E+05	9.58E-03	6.95E-08
IMC-1121b	N/A	N/A	**2.27E+05**	**8.75E-05**	**3.85E-10**

* N/A (not applicable)

Example 11: Analysis of KDR neutralizing ability of anti-KDR-IgG in HUVEC cells using FACS analysis

Primary-cultured HUVEC cells were cultured in serum-free medium overnight to induce the overexpression of KDR, and then the cells were harvested, washed three times with PBS. The washed cells were allowed to react with 6A6 or IMC-1C11 IgG (10μg/ml) at 4℃ for 1 hour, and then allowed to react with an FITC-labeled rabbit anti-human IgG antibody (Abchem, UK) for 60 minutes. After completion of the reaction, the cells were washed and analyzed with a flow cytometer (FACS; model EPICS9, Coulter Corp., USA).

As a result, as shown in FIG. 14, IMC-1C11 and 6A6 recognized the KDR of HUVEC cells at the same level.

Also, in order to examine competitive inhibitory ability against VEGF, HUVEC cells were cultured in a serum-free condition overnight to induce the expression of KDR, and then the cells were harvested and washed three times with PBS. The washed cells were allowed to react with 20 ng/ml of VEGF at room temperature for 30 minutes. After the completion of the reaction, the cells were allowed to react

with 6A6-IgG and IMC-1C11 IgG at 4℃ for 1 hour, and then allowed to react with an FITC-labeled rabbit anti-human IgG antibody at 4℃ for 30 minutes.

As a result, as shown in FIG. 15, it was observed that the two antibodies all showed a signal of binding to VEGF165, indicating that 6A6-IgG was competitively bound to VEGF. Also, in the VEGF competition assays, 6A6-IgG and IMC-1C11 IgG showed the same level of KDR-neutralizing ability, but the KDR neutralizing ability in actual living cells was about two-fold higher for 6A6-IgG than for IMC-1C11 IgG.

Example 12: Analysis of KDR neutralizing ability of anti KDR-IgG in K562 cells

In order to examine the KDR binding affinity of the antibodies in KDR expressing cells lines other than HUVEC cells, the expression of KDR in the leukemia cell line K562 (ATCC CCL-243) was analyzed. FIG. 16 shows the results of Western blot analysis for the expression of KDR in K562 (ATCC CCL-243) cells. As shown in FIG. 16, K562 and HUVEC cells expressed KDR regardless of the presence or absence of serum. Thus, the K562 cells (ATCC CCL-243) were treated in the same manner as the HUVEC cells of Example 11 and analyzed by FACS in order to examine whether KDR-IgG could bind to the K562 cells.

As a result, as shown in FIG. 17, only 6A6-IgG was bound to the K562 cells at a significant level, unlike the HUVEC cells. The results of FACS assays through VEGF competition are shown in FIG. 18. As shown in FIG. 18, the K562 cells did not show a great change in the rate of positive cells, unlike the HUVEC cells. Although the reason is unclear, this is thought to be because the 6A6 antibody strongly binds to KDR expressed on the surface of the K562 cells or regulates the growth of the cells using the autocrine loop mechanism of VEGF/KDR(VEGFR-2), and thus if VEGF is externally treated, the expression of KDR in the K562 cells is induced, so that an increased amount of the KDR protein is expressed on the cell

surface, and the 6A6-IgG signal is increased compared to before the VEGF is externally treated. Also, as shown in FIG. 19, it was seen that 6A6-IgG could bind the KDR of gleevec-resistant K562 cells (The Catholic University of Korea).

Example 13: Analysis of inhibition of HUVEC cell proliferation by 6A6 antibody

The inhibition of HUVEC cell proliferation by anti-KDR-IgG was analyzed using WST-1 reagent (Roche, Swiss). HUVEC cells were dispensed into each well of a gelatin-coated, 24-well culture plate at a concentration of 2 x 10^4 cells/well and cultured for 18 hours. Then, the cells were further cultured in serum-free M199 medium (Sigma-aldrich, USA) for 4 hours, and then 20 ng/ml of VEGF and various concentrations of 6A6 were added thereto. Then, the WST-1 reagent was added thereto according to the manufacturer's manual, and after 1 hour and 4 hours, the cells were measured for absorbance at 450 nm and 690 nm (FIG. 20).

As a result, when the HUVEC cells were treated with VEGF, the proliferation thereof was increased by about three times, but when the 6A6 antibody was added to the HUVEC cells, the proliferation of the cells was reduced in a concentration-dependent manner.

Example 14: Analysis of effect of 6A6 antibody on inhibition of KDR and ERK phosphorylation

Sufficiently grown HUVEC cells were cultured in 1% FBS-containing M199 medium for 6 hours, and then treated with VEGF and 6A6, IMC-1121 and 6C1 antibodies at various concentrations for 10 minutes. Then, the cells were lysed with 1 ml of lysis buffer (20 mM Tris-HCl, pH 8.0, 2 mM EDTA, 137 mM NaCl, 1 mM Na_3VO_4, 1 mM PMSF, 10% glycerol, 1% Triton X-100) and centrifuged, and the supernatant was treated with 1 μg/ml of an anti-KDR/Flk-1 antibody (Santa cruz

Biotechnology, USA) at 4℃ for 3 hours. The treated supernatant was incubated on protein-A agarose beads (Sigma-aldrich, USA) for 1 hour, and the immunoprecipitated protein was electrophoresed on SDS-PAGE, and then analyzed by Western blot (FIG. 21A).

As a result, it was observed that, when the cells were treated with VEGF, the phosphorylation of KDR was increased as expected, but the cells were treated with 6A6 or IMC-1121, the phosphorylation of KDR by VEGF was inhibited. Also, the 6C1 antibody had no effect on the neutralization of VEGF.

According to the above-described method, a test was carried out to examine whether the phosphorylation of the kinase ERK known to receive the signal of KDR would be inhibited. As a result, it was confirmed that 6A6 and IMC-1121 inhibited the phosphorylation of ERK, but 6C1 did not substantially inhibit the phosphorylation of ERK as expected (FIG. 21B).

Example 15: Analysis of inhibitory effect of anti-KDR-6A6 IgG on the chemotaxis of endothelial cells induced by VEGF

In order to examine the inhibitory effect of 6A6-IgG on the migration of HUVEC cells induced by VEGF, a transwell (Corning costar, USA) having a 6.5-mm-diameter polycarbonate filter (8 μM pore size) was used. The surface of the lower layer of the filter was coated with 10 μg of gelatin, and fresh M199 medium (containing 1% FBS) and VEGF were placed in the lower layer well of the filter. HUVEC cells were diluted in M199 medium (containing 1% FBS) at a concentration of 1 x 10^6/ml, various concentrations of the anti-KDR antibodies were added thereto and allowed to react at room temperature for 30 minutes. 100 μl of the reaction solution was placed in the upper layer well and allowed to react at 37℃ for 4 hours. Then, the cells were stained with hematoxylin and eosin. Non-migrated cells were removed with cotton, and cells migrated into the lower layer

well were observed with a microscope to measure the number of the migrated cells. As a result, it was observed that the migration of HUVEC cells induced by VEGF was inhibited by 6A6 in a concentration-dependent manner (FIG. 22).

Example 16: Analysis of inhibitory effect of anti-KDR-6A6 IgG on endothelial cell tube formation induced by VEGF

In order to examine whether the 6A6 antibody inhibits HUVEC tube formation induced by VEGF, 250 μl of growth factor-reduced matrigel (Collaborative biomedical products, USA) was placed in each well of a 16-mm-diameter tissue culture plate and polymerized at 37℃ for 30 minutes. HUVEC cells were suspended in M199 medium (containing 1% FBS), and various amounts of the antibody was mixed and allowed to react with the cells. After 30 minutes, the cells were plated on the matrigel at a concentration of 2 x 10^5 cells/well, 10 ng/ml of VEGF was added thereto, and the cells were cultured for 20 hours. The cultured cells were observed with a microscope and imaged with an Image-Pro plus (Media cybernetics, USA). As a result, it was observed that HUVEC tube formation induced by VEGF was inhibited by 6A6-IgG (FIG. 23).

Example 17: Inhibition of VEGF-KDR internalization by binding of 6A6 antibody to KDR on cell surface

HUVEC cells were placed on a gelatin-coated cover slip at a concentration of $2x10^4$ cells/well, after 24 hours, the cells were washed twice with M199 medium and cultured in M199 medium (containing 1% FBS) for 6 hours. The HUVEC cells were allowed to react with various concentrations of the antibody for 30 minutes and allowed to react with 10 ng/ml of VEGF for 10 minutes. After the completion of the reaction, the cells were immobilized and infiltrated with methanol or 2% formaldehyde for 10 minutes and washed with PBS. Then, the cells were blocked with 0.1% Triton X-100 and 2% BSA/PBS for 30 minutes, and the cells were

allowed to react with a mouse KDR antibody for 1 hour, and then allowed to react with an FITC-labeled anti-mouse antibody at room temperature for 45 minutes. The cover slip was mounted with SloFade (Molecular Probe) and observed with a confocal microscope (Zeiss, Germany) at 488 nm (excitation wavelength).

As a result, as shown in FIG. 24, it was observed that the 6A6 antibody inhibited the infiltration of KDR into cells, and 6G1 did not substantially inhibit the infiltration of KDR into cells.

Example 18: *ex vivo* analysis of inhibitory effect of anti-KDR-IgG on angiogenesis

In order to examine whether 6A6-IgG inhibits aortic ring vessel sprouting induced by VEGF, an aortic ring assay was performed. First, arteries were separated from 6-week-old rats (Sprague Dawley), and then cut to a size of about 0.5 mm. The cut artery was placed on 120 μl of a matrigel-coated, 48-well plate and covered with 50 μl of matrigel. VEGF (10 ng/ml) and each of 6A6-IgG, 6C1-IgG and 1121-IgG were mixed with human endothelial serum-free medium (Invitrogen) to a final volume of 200 μl, and the mixture was placed in each well of the plate. After 6 days, the cells were immobilized and stained with Diff-Quick (Baxter Diagnostics). Data were rated on a scale of 0 (least positive) to 5 (most positive), and six independent tests were performed. FIG. 25A shows a vessel sprouting image, and FIG. 25B shows the statistical results of scores for vessel sprouting. The 6A6 antibody inhibited vessel sprouting induced by VEGF, but the 6C1 antibody did not show the inhibitory ability. The above-described *ex vivo* rat aortic ring assay results have a very important meaning in addition to the simple fact that 6A6 inhibits angiogenesis induced by VEGF. That is, the results revealed that 6A6 could bind to Flk-1 to neutralize the human KDR homologue Flk-1 expressed in rats, although it was prepared for the purpose of neutralizing human KDR. In other words, it can be seen that the 6A6 antibody has cross-reactivity between humans and rats.

Example 19: Effect of 6A6 on inhibition of *in vivo* angiogenesis induced by VEGF (*in vivo* mouse matrigel plug assay)

Following the rat aortic ring assay, a mouse matrigel plug assay was performed in order to examine whether the human KDR neutralizing antibody 6A6 can inhibit angiogenesis induced by VEGF *in vivo* in mice. For this purpose, 6-8-week-old C57/BL6 mice were subcutaneously injected with 0.6 ml of matrigel, containing 200 μg of the antibody, 100 ng of VEGF and 10 units of heparin. After 7 days, the matrigel plug was taken out by surgery, and the image thereof was photographed (FIG. 26A). Then, the plug was rapidly frozen with liquid nitrogen in the presence of an OCT (optimum cutting temperature) compound and cut to a thickness of 8-12 μm. The cut plug was fixed with 4% neutral buffered paraformaldehyde, and the density of the microvessels was measured with an anti-CD31 antibody (FIG. 26B). It was observed that the 6A6 antibody could inhibit blood vessel formation induced by VEGF *in vivo* in mice. Like the rat *ex vivo* experiment, it was confirmed again that the 6A6 antibody could neutralize the mouse KDR homologue Flk-1 and had cross-reactivity between humans and mice. Among therapeutic antibodies, a KDR antibody having cross-reactivity between humans and mice has not yet been reported. When the species cross-reactivity of the 6A6 antibody is used, the *in vivo* effect of the antibody can be confirmed using mice or rats.

Example 20: Anti-cancer effect of 6A6 antibody in colon cancer xenograft animal model

The anticancer effect of the 6A6 antibody in colon cancer xenograft animal models was analyzed using T cell-, B cell- and NK cell-deleted NOD/SCID IL-2R null mice (female, 11-week-old, weighed 25 g, The Jackson Laboratories, USA) known to have an advantage that the cells more easily receive human cancer cells, compared to NOD/SCID mice.

As human cancer cells, human colon cancer cells known as HCT116 (ATCC, USA) were used, and the injection of the tumor cells was performed by injecting the cells subcutaneously into the left side of the mice at a concentration of 2 x 10^5 cells (serum-free DMEM)/10 μl at day 0.

The 6A6 antibody was injected intravenously into the mice from day 1 (24 hours from day 0 at which the tumor cells were injected) three times a week. The mice were divided into three groups, each consisting of 5 animals. The group 1 was a PBS-injected group (control group), the group 2 was injected with 100 μg/ea (= 4 mg/kg) of the 6A6 antibody, and the group 3 was injected with 200 μg/ea (= 8 mg/kg) of the 6A6 antibody. The size of a tumor occurring in the mice was measured according to the following equation on alternate days for 26 days:

Tumor volume = 1/2 x (length x area x height).

At day 30, the mice were sacrificed, and the weight of a tumor was measured. As a result, it was observed that the tumor size was dose-dependently reduced in the groups administered with the 6A6 antibody, compared to the control group injected with PBS (FIG. 27 and FIG. 28).

Example 21: Anti-cancer effect of 6A6 antibody in lung cancer xenograft animal models

Human lung cancer A549 cells (ATCC, USA) were injected subcutaneously into nude mice (Japan SLC, Japan) at a concentration of 7 x 10^7 cells to form tumors. 10 days after the injection of the cancer cells, the tumors could be visually observed, and then the 6A6 antibody was injected intraperitoneally into the mice three times a week.

The mice were divided into three groups, each consisting of five animals. The

group 1 was a PBS-injected group (control group), the group 2 was injected with 1 mg/kg of the 6A6 antibody, and the group 3 was injected with 1 mg/kg of Avastin (Genentech, USA). As a result, as can be seen in FIG. 29, the growth of the tumor was inhibited in the group injected with the 6A6 antibody and the positive control group injected with Avastin, compared to the control group injected with PBS.

Example 22: *In vivo* tumor targeting of radioactive iodine-labeled 6A6 antibody

The tumor targeting of the 6A6 antibody was analyzed using the binding affinity of the antibody to CML (chronic myelogenous leukemia) K562 cells. The antibody was labeled with radioactive iodine-125 using an iodobead method to label more than 90% of the antibody with iodine (FIG. 30A), and an anti-KDR (6A6) antibody labeled with radioactive iodine having a purity of more than 98% was prepared (FIG. 30B). A CML tumor model was prepared by injecting K562 cells subcutaneously into Balb/c nude mice, and when the tumor size reached 1 cm at 21-28 days after the injection of the K562 cells, the iodine-125-labeled antibody (100 μg) was injected into the tail vein of the K562 tumor model nude mice. 2 hour and 24 hours after the injection of the antibody, gamma-camera images of animals having tumors formed therein were obtained. It was observed that the introduction of the antibody into the tumors showed similar patterns at 2 hours and 24 hours, and the background radioactivity was reduced after 24 hours. It was observed that the antibody was localized to the tumor, suggesting that KDR was expressed on the K562 tumor. Thus, due to the therapeutic effect of the antibody itself by localization, as well as due to an increase in the therapeutic effect caused by beta-rays emitted from the radioactive isotope, the antibody can possibly be used as a radioimmunotherapy agent (FIG. 31).

Example 23: Affinity maturation of 6A6-IgG using light chain shuffling

In order to identify antibodies having an affinity higher than that of 6A6, a heavy

chain was removed from the DNA of the complete human antibody library, prepared in Example 2, using restriction enzyme *Sfi*I. Into the site from which the heavy chain has been removed, a heavy chain of pAK-6A6 treated with *Sfi*I restriction enzyme was inserted. The resulting DNA was transformed into ETB (Electro Ten blue) cells (Stratagene, USA), and the cells were cultured in SOB medium for 1 hour. Then, the cells were spread on a 2 x YT (Cm) square plate, and the next day, the colony was collected and stored at -70℃. As a result, a 6A6 light chain shuffling library having a diversity of $4x10^6$ was constructed (FIG. 32).

In order to examine whether the light chain shuffling of the library has been successfully achieved, 48 clones were randomly selected, and the light chain sequences thereof were analyzed. As a result, there was no overlap in the light chain sequences of the 48 clones.

From the library, clones having a binding affinity higher than that of 6A6 were screened in the same manner as in Example 3 through a biopanning process using a phage display.

18 candidates were finally obtained through the following procedures.

(1) In order to prevent 6A6 from being selected again during the biopanning process, the DNA of the 6A6 light chain shuffling library was treated with a restriction enzyme *Spe*I having a recognition site at the CDR3 of 6A6. After the DNA was transformed into ETB cells, a sub-library was constructed based on the cultured cells, and the KDR affinity of the sub-library was analyzed in ELISA. Among candidates resulting from the fourth panning, 94 candidates were selected and subjected to KDR binding assays in the same manner as in Example 5. Among candidates showing positive responses, 4 candidates were randomly selected, and the DNA sequences thereof were determined. Also, the candidates were subjected to VEGF competition assays using the respective phages (FIG. 33). As a result,

4SD5, 4SC3 and 4SC5, which had KDR neutralizing ability similar to equal to that of the positive control group 6A6, were selected.

(2) In a washing step in the biopanning process, KE3, KE6, 2KG8, 3KE11, 3KF11, 3KG3 and K3F1 were selected through competition with soluble KDR. The biopanning process used herein was as follows.

Maxisorp Star tubes (Nunc, Denmark) were coated with 4 ml of KDR (5 μg/ml) and blocked with 2% skimmed milk/PBS at 37℃ for 2 hours. Then, 500 μl of the 6A6 light chain shuffling library phage suspended in 2% skimmed milk was allowed to bind to the KDR, and then allowed to react in 0.1% PBS-T (tween20) for 1 hour. Then, the tubes were washed 10 times with 0.1% PBST and washed 10 times with PBS buffer. Then, 4 ml of soluble KDR (25μg/ml/PBS) was added and allowed to bind thereto for 30 minutes, and the tubes were treated with 100 mM triethylamine for 10 minutes to elute the phage. The eluted phage was neutralized with 500 μl of 1M Tris-Cl (pH7.5) and transformed into *E. coli* XL1-Blue cells for 50 minutes, and then the cells were cultured.

The antibodies selected in each of the three panning steps were subjected to VEGF competition assays in a ScFv-phage particle state. Among four candidate antibodies obtained through the first panning, KE3 and KE6, having high VEGF competitive power compared to those of the other candidate antibodies, were selected, and KC7 and KQ11 were excluded (FIG. 34A). Among three candidate antibodies obtained through the second panning, 2KE5 having low VEGF competitive power was excluded, and the remaining 2KG4 and 2KG8, having VEGF competitive power similar to that of 6A6, were selected. The 2KG4 showed the same sequence as 3KG3, an antibody selected later, and thus it was substituted with 3KG3 (FIG. 34B).

Among five candidate antibodies obtained through the third panning, 3KG2 and

3KF7, having low competitive power, were excluded, and only 3KE11, 3KF11 and 3KG3 were selected (FIG. 34C). Similarly, K3F1 obtained through the third panning was selected, and K3F1 showed a significantly high VEGF competitive power compared to that of 6A6 (FIG. 34D).

(3) In a step of allowing phage to bind to the antigen KDR in the biopanning process, IMC-1121 IgG obtained in Example 8 was also added and, as a result, IE4, 3IG11, 3IG12, 3IE1, 3IH2, I2F2, I3A12 and I3F2 clones were selected. The biopanning process used herein was as follows.

Maxisorp Star tubes (Nunc, Denmark) were coated with 4 ml of KDR (5μg/ml) and blocked with 2% skimmed milk/PBS at 37℃ for 2 hours. Then, 500 μl of the 6A6 light chain shuffling library phage suspended in 2% skimmed milk containing 21 μg/ml of IMC-1121 IgG (0.14 μM) was allowed to bind thereto for 1 hour, and then allowed to react in 0.1% PBST for 1 hour. Then, the tubes were washed 10 times with 0.1% PBST and washed with 10 times with PBS buffer. Then, 4 ml of soluble KDR (25 μg/ml/PBS) was added to the tubes and allowed to bind for 30 minutes, and then treated with 100 mM triethylamine for 10 minutes to elute the phage. The eluted phage was neutralized with 500 μl of 1M Tris-Cl (pH 7.5), and then transformed into *E. coli* XL1-Blue cells for 50 minutes, and the cells were cultured.

The antibodies selected in each of the three panning steps were subjected to VEGF competition assays in a ScFv-phage particle state. Among three candidate antibodies obtained through the first panning, only IE4 having VEGF competitive power similar to that of 6A6 was selected (FIG. 35A). IE4 was analyzed for the DNA sequence thereof and, as a result, 28 amino acids in IE4 were different from those in 6A6. In addition, 6A6 had 108 light chain amino acids, whereas IE4 had 107 amino acids, indicating that one amino acid was deleted in the CDR3 of 6A6.

FIG. 35B shows the results of VEGF competition assays of three candidate antibodies obtained through the third panning. In DNA sequencing, 3IG8 had a light chain sequence completely different from that of 6A6, and in the results of FACS, 3IG8 did not bind to living cells, indicating that it was not converted in the form of IgG. 3IG11, 3IG12, 3IE1 and 3IH2 were selected, and 3IA7 was excluded, because it had a stop codon in the light chain sequence. FIG. 35C shows the results of VEGF competition assays of candidate antibodies obtained through the second panning and the third panning. 3IA12, I3F2 and I2F2 were all selected.

The light chain DNA sequences of the 18 selected clones are shown in SEQ ID NO: 164 to SEQ ID NO: 181, and amino acid sequences deduced from the DNA sequences are shown in SEQ ID NO: 2 to SEQ ID NO: 19. Regions substituted compared to the light chain amino acid of 6A6 (TTAC-0001) are shown in Table 7. Also, the clones were renamed "TTAC-0002 to TTAC-0019" (Table 8).

Table 7: Mutation site of 18 selected clones

Clone name	Mutation site
KE3 (TTAC-0002)	S13A, R23G, L27I, D29S, V30Q, N31S
KE6 (TTAC-0003)	S13A, R19G, R23G, N26D, L27I, D29S, V30K, N31S, R38K, M47I, A51S
IE4 (TTAC-0004)	N1S, F2Y, M3E, V12S, S13A, R19T, R23E, D25K, L27I, D29S, V30K, N31S, R38K, V46L, M47I, A51Q, G56A, G67D, T69M, G76R, E78A, D91G, R92N, T93G, S94K, E95V, T99G, V103L
SD5 (TTAC-0005)	S13A, T100A
2KG8 (TTAC-0006)	S13A
3KE11 (TTAC-0007)	T71I
3KF11 (TTAC-0008)	P8S
3KG3 (TTAC-0009)	P8H
3IG11 (TTAC-0010)	V46I
3IG12	R23M

(TTAC-0011)	
3IE1 (TTAC-0012)	P8S, S13P, P39R, Y96F
3IH2 (TTAC-0013)	V12L, K16Q
K3F1 (TTAC-0014)	S13A, K16Q
I2F2 (TTAC-0015)	S9A
I3A12 (TTAC-0016)	E59K
I3F2 (TTAC-0017)	M47I
4SC3 (TTAC-0018)	S94N
4SC5 (TTAC-0019)	N1Q, M3V, S13A, R23G, D25N, L27I, D29S, V30K, N31S, R38K, M47I, A51S, S66F, G76R, R92S, T93S, S94R, E95D

Table 8: New name of antibodies developed

Name of antibodies developed	New name	Name of antibodies developed	New name
6A6	TTAC-0001	3IG12	TTAC-0011
KE3	TTAC-0002	3IE1	TTAC-0012
KE6	TTAC-0003	3IH2	TTAC-0013
IE4	TTAC-0004	K3F1	TTAC-0014
SD5	TTAC-0005	I2F2	TTAC-0015
2KG8	TTAC-0006	I3A12	TTAC-0016
3KE11	TTAC-0007	I3F2	TTAC-0017
3KF11	TTAC-0008	4SC3	TTAC-0018
3KG3	TTAC-0009	4SC5	TTAC-0019
3IG11	TTAC-0010		

INDUSTRIAL APPLICABILITY

As described in detail above, the present invention provides a fully human antibody, which has an excellent ability to neutralize VEGF receptor in cells and *in vivo*, and a composition for inhibiting angiogenesis and a composition for treating cancer, which contain said antibody. The inventive 6A6 antibody neutralizing vascular endothelial growth factor receptor shows excellent neutralizing ability in living cells, compared to that of a commercially available antibody against vascular endothelial

growth factor receptor, and shows the ability to neutralize vascular endothelial growth factor receptor not only in humans, but also in mice and rats. Thus, the 6A6 antibody will be useful in anticancer studies and will be highly effective in cancer treatment.

Although the present invention has been described in detail with reference to the specific features, it will be apparent to those skilled in the art that this description is only for a preferred embodiment and does not limit the scope of the present invention. Thus, the substantial scope of the present invention will be defined by the appended claims and equivalents thereof.

THE CLAIMS

What is Claimed is:

1. A single chain variable fragment (ScFv) molecule neutralizing vascular endothelial growth factor receptor, wherein the ScFv molecule contains a light chain variable region represented by an amino acid sequence of any one of SEQ ID NOS: 1 to 19 and a heavy chain variable region represented by an amino acid sequence of SEQ ID NO: 20.

2. A DNA encoding the ScFv (single chain variable fragment) molecule of claim 1.

3. A vector containing the DNA of claim 2.

4. A Recombinant cells transformed with the vector of claim 3.

5. The recombinant cells according to claim 4, wherein the cells are bacterial or animal cells.

6. A composition for inhibiting angiogenesis, which contains the ScFv molecule of claim 1.

7. A composition for treating cancer, which contains the ScFv molecule of claim 1.

8. An IgG neutralizing vascular endothelial growth factor receptor, wherein the IgG contains a light chain variable region represented by an amino acid sequence of any one of SEQ ID NOS: 1 to 19 and a heavy chain variable region represented by an amino acid sequence of SEQ ID NO:20.

9. A composition for inhibiting angiogenesis, which contains the IgG of claim 8.

10. A composition for treating cancer, which contains the IgG of claim 8.