WHAT IS CLAIMED IS:

1. A method comprising the steps of:

(a) subjecting harvested Dupilumab to affinity chromatography;

(b) subjecting said Dupilumab pooled from eluate of step (a) to viral inactivation at a pH from about 3 to about 4 and then adjusting the pH to from about 5 to about 8;

(c) subjecting said Dupilumab pooled from step (b) to cation exchange chromatography in bind and elute mode;

(d) subjecting said Dupilumab pooled from eluate of step (c) to anion exchange chromatography in flowthrough mode; and

(e) subjecting said Dupilumab pooled from flowthrough fractions of step (d) to virus retentive filtration to produce Dupilumab.

2. The method of claim 1, further comprising a harvest pre-treatment step prior to step (a).

3. The method of claim 2, wherein said harvest pre-treatment step includes adjusting said Dupilumab to a transient pH level from about 4 to 5.5.

4. The method of claim 1, further comprising subjecting said Dupilumab to ultrafiltration and diafiltration (UF/DF) after step (e).

5. The method of claim 4, wherein said UF/DF includes a diafiltration buffer having a pH between 4.0 and 4.5.

6. The method of claim 4, wherein a concentrated Dupilumab pool following UF/DF has a pH of about 5.3.

7. The method of claim 4, wherein said UF/DF includes a diafiltration buffer comprising between about 4 mM acetate and about 6 mM acetate.

8. The method of claim 1, wherein said affinity chromatography is Protein A chromatography.

9. The method of claim 8, wherein a Protein A resin is selected from the group consisting of MabSelect PrismA, MabSelect SuRe, MabSelect SuRe LX, MabSelect, MabSelect SuRe pcc, MabSelect Xtra, rProtein A Sepharose, ProSep HC, ProSep Ultra, ProSep Ultra Plus, MabCapture, and Amsphere A3.

10. The method of claim 8, wherein a Protein A resin is selected that is capable of receiving a load at a concentration above 55 g/L.

11. The method of claim 8, wherein a Protein A column load pH is between 6 and 8.

12. The method of claim 1, wherein said Dupilumab comprises three heavy chain complementarity determining region (HCDR) sequences comprising SEQ ID NOs: 3, 4, and 5, and three light chain complementarity determining region (LCDR) sequences comprising SEQ ID NOs: 6, 7, and 8.

13. The method of claim 1, wherein said Dupilumab comprises a heavy chain variable region (HCVR) comprising the amino acid sequence of SEQ ID NO: 1 and a light chain variable region (LCVR) comprising the amino acid sequence of SEQ ID NO: 2.

14. A method comprising the steps of:

(a) culturing cells expressing Dupilumab;

(b) subjecting said cells to transient pH levels from about 4 to 5.5, then raising pH levels to from about 5.5 to 6.5;

(c) harvesting said cells by centrifugation to separate cell debris from clarified media comprising said Dupilumab;

(d) subjecting said clarified media to affinity chromatography;

(e) subjecting said Dupilumab pooled from eluate of step (d) to viral inactivation at a pH from about 3 to about 4 and then adjusting the pH to from about 5 to about 8;

(f) subjecting said Dupilumab pooled from step (e) to cation exchange chromatography in bind and elute mode;

(g) subjecting said Dupilumab pooled from eluate of step (f) to anion exchange chromatography in flowthrough mode; and

(h) subjecting said Dupilumab pooled from flowthrough fractions of step (g) to virus retentive filtration to produce Dupilumab.

15. The method of claim 14, further comprising subjecting said Dupilumab to ultrafiltration and diafiltration (UF/DF) after step (h).

16. The method of claim 14, wherein said affinity chromatography is Protein A chromatography.

17. The method of claim 16, wherein a Protein A resin is selected from the group consisting of MabSelect PrismA, MabSelect SuRe, MabSelect SuRe LX, MabSelect, MabSelect SuRe pcc, MabSelect Xtra, rProtein A Sepharose, ProSep HC, ProSep Ultra, ProSep Ultra Plus, MabCapture, and AmsphereA3.

18. The method of claim 14, wherein an anion exchange resin is selected from the group consisting of Q Sepharose Fast Flow, Poros 50PI, Poros 50HQ, Capto Q Impres, Capto DEAE, Toyopearl QAE-550, Toyopearl DEAE-650, Toyopearl GigaCap Q-650, Fractogel EMD TMAE Hicap, Sartobind STIC PA nano, Sartobind Q nano, CUNO BioCap, and XOHC.

19. The method of claim 14, wherein a cation exchange resin is selected from the group consisting of Fractogel Hicap, Capto SP ImpRes, Capto S ImpAc, CM Hyper D grade F, Eshmuno S, Nuvia C Prime, Nuvia S, Poros HS, and Poros XS.

20. The method of claim 14, further comprising passing said Dupilumab through a LifeAssure filter after the viral inactivation of step (e) and prior to the cation exchange chromatography of step (f).

21. The method of claim 15, wherein said UF/DF step comprises a membrane filter device selected from the group consisting of Pellicon 2, Pellicon 3 cassettes with 10 kD, 30 kD or 50 kD membranes, Kvick 10 kD, 30 kD or 50 kD membrane cassettes, and Centramate and Centrasette 10 kD, 30 kD or 50 kD cassettes.

22. The method of claim 15, wherein said UF/DF step does not include addition of arginine.

23. The method of claim 14, wherein said Dupilumab comprises three heavy chain complementarity determining region (HCDR) sequences comprising SEQ ID NOs: 3, 4, and 5, and three light chain complementarity determining region (LCDR) sequences comprising SEQ ID NOs: 6, 7, and 8.

24. The method of claim 14, wherein said Dupilumab comprises a heavy chain variable region (HCVR) comprising the amino acid sequence of SEQ ID NO: 1 and a light chain variable region (LCVR) comprising the amino acid sequence of SEQ ID NO: 2.

25. A method for producing an anti-IL4Rα antibody or antigen-binding fragment thereof, comprising:

(a) subjecting a harvested antibody or antigen-binding fragment thereof to affinity chromatography;

(b) subjecting said antibody or antigen-binding fragment thereof pooled from eluate of step (a) to viral inactivation;

(c) subjecting said antibody or antigen-binding fragment thereof pooled from step (b) to cation exchange chromatography in bind and elute mode;

(d) subjecting said antibody or antigen-binding fragment thereof pooled from eluate of step (c) to anion exchange chromatography in flowthrough mode;

(e) subjecting said antibody or antigen-binding fragment thereof pooled from flowthrough fractions of step (d) to virus retentive filtration; and

(f) subjecting said antibody or antigen-binding fragment thereof pooled from step (e) to ultrafiltration and diafiltration (UF/DF) to produce an anti-IL4Rα antibody or antigen-binding fragment thereof, wherein said UF/DF step does not include addition of arginine.

26. The method of claim 25, wherein said UF/DF step includes a diafiltration buffer having a pH between 4.0 and 4.5.

27. The method of claim 25, wherein said UF/DF step includes a diafiltration buffer comprising between about 4 mM acetate and about 6 mM acetate.

28. The method of claim 25, wherein a concentrated pool following UF/DF has a pH of about 5.3.

29. The method of claim 25, wherein said UF/DF step comprises a membrane filter device selected from the group consisting of Pellicon 2, Pellicon 3 cassettes with 10 kD, 30 kD or 50 kD membranes, Kvick 10 kD, 30 kD or 50 kD membrane cassettes, and Centramate and Centrasette 10 kD, 30 kD or 50 kD cassettes.

30. A method comprising the steps of:

(a) subjecting a harvested antibody to affinity chromatography;

(b) subjecting said antibody pooled from eluate of step (a) to viral inactivation at a pH from about 3 to about 4 and then adjusting the pH to from about 5 to about 8;

(c) subjecting said antibody pooled from step (b) to cation exchange chromatography in bind and elute mode;

(d) subjecting said antibody pooled from eluate of step (c) to anion exchange chromatography in flowthrough mode; and

(e) subjecting said antibody pooled from flowthrough fractions of step (d) to virus retentive filtration to produce an anti-IL4Rα antibody.

31. The method of claim 30, further comprising a harvest pre-treatment step prior to step (a).

32. The method of claim 31, wherein said harvest pre-treatment step includes adjusting said antibody to a transient pH level from about 4 to 5.5.

33. The method of claim 30, further comprising subjecting said antibody to ultrafiltration and diafiltration (UF/DF) after step (e).

34. The method of claim 33, wherein said UF/DF includes a diafiltration buffer having a pH between 4.0 and 4.5.

35. The method of claim 33, wherein a concentrated antibody pool following UF/DF has a pH of about 5.3.

36. The method of claim 33, wherein said UF/DF includes a diafiltration buffer comprising between about 4 mM acetate and about 6 mM acetate.

37. The method of claim 30, wherein said affinity chromatography is Protein A chromatography.

38. The method of claim 37, wherein a Protein A resin is selected from the group consisting of MabSelect PrismA, MabSelect SuRe, MabSelect SuRe LX, MabSelect, MabSelect SuRe pcc, MabSelect Xtra, rProtein A Sepharose, ProSep HC, ProSep Ultra, ProSep Ultra Plus, MabCapture, and Amsphere A3.

39. The method of claim 37, wherein a Protein A resin is selected that is capable of receiving a load at a concentration above 55 g/L.

40. The method of claim 37, wherein a Protein A column load pH is between 6 and 8.

41. The method of claim 30, wherein said anti-IL-4Rα antibody comprises three heavy chain complementarity determining region (HCDR) sequences comprising SEQ ID NOs: 3, 4, and 5, and three light chain complementarity determining region (LCDR) sequences comprising SEQ ID NOs: 6, 7, and 8.

42. The method of claim 30, wherein said anti-IL-4Rα antibody comprises a heavy chain variable region (HCVR) comprising the amino acid sequence of SEQ ID NO: 1 and a light chain variable region (LCVR) comprising the amino acid sequence of SEQ ID NO: 2.

43. The method of claim 30, wherein said anti-IL-4Rα antibody is Dupilumab.