

REAGENTS FOR USE IN BIOLUMINESCENCEField of the Invention

This invention relates to reagents for use in bioluminescence, and to their use.

5 Background of the Invention

10 The background level of bioluminescence signal is often a problem when using the firefly luciferase reaction to measure adenosine 5'-triphosphate (ATP) levels deriving from prokaryotic cells, eukaryotic cells or both. Particular problems occur when the sample contains growth medium from a natural source, which is often the case in microbiological assays. Since such media are produced from living material, they tend to contain a significant amount of ATP. Most of the background bioluminescence signal
15 arising from them can be removed by reaction with an ATPase enzyme such as potato apyrase or an ATP-utilising enzyme such as a kinase together with its co-substrate. However, a proportion of the signal always remains and can interfere with sensitive bioluminescence measurements. The source
20 has always been thought of as non-microbial or non-cellular ATP which is somehow unavailable to ATPase enzymes, perhaps by reason of being sequestered.

Summary of the Invention

25 The present invention is based on the discovery that the residual background bioluminescence signal can be removed by reaction with certain other enzymes such as phosphodiesterase I from snake venom (E.C. 3.1.4.1; Enzyme Nomenclature 1984, Academic Press), acid phosphatase (E.C. 3.1.3.2) and alkaline phosphatase (E.C. 3.1.3.1). Likely
30 candidates for this "non-degradable ATP" are substances structurally related to ATP that are not attacked by apyrase, or by other enzymes which have ATP as a substrate, yet give rise to light when acted upon by firefly luciferase. They may be true substrates of luciferase, or
35 be acted upon by luciferase to form a true substrate such as ATP, or may break down to form ATP or another luciferase substrate by other unidentified means.



Flavine adenine dinucleotide (FAD) has been identified as one such substance. It occurs in many microbiological growth media and biological samples, is structurally related to ATP, appears to act as a light-emitting substrate for firefly luciferase, but is not degraded by apyrase; it is destroyed by snake venom phosphodiesterase I. Other such substances are adenosine 5'-tetraphosphate (p_4A) and other adenosine polyphosphates, diadenosine 5',5''- P^1, P^4 -tetraphosphate (Ap_4A) and other diadenosine polyphosphates, and other dinucleoside polyphosphates such as Gp_4A and Gp_5A . It is likely that other, as yet unidentified, derivatives of ATP will have similar properties. A few of these have previously been suggested to act as low-efficiency light-emitting luciferase substrates; see Momsen (1978) Biochem. Biophys. Res. Commun. 84:816-822, and Sillero et al (1991) Eur. J. Biochem. 202:507-513.

These compounds mentioned may be destroyed by suitable enzymatic activity, e.g. snake venom phosphodiesterase I or phosphatases. Other enzymes exist which will act upon these nucleotide derivatives with various degrees of specificity. For example, adenosine tetraphosphatase (E.C. 3.6.1.14) can be used in the method of the invention: it hydrolyses the luciferase substrate adenosine 5'-tetraphosphate to give ATP, which will subsequently be destroyed by apyrase. Diadenosine 5',5''- P^1, P^4 -tetraphosphatase (E.C. 3.6.1.17), which has been isolated from yellow lupin seeds (Jakubowski et al (1983) J. Biol. Chem. 258 (16):9982-9989), also produces ATP from its substrates and could be used in combination with apyrase in cases where part of the residual bioluminescence signal arises from Ap_4A .

A reagent to treat liquids, in order to reduce their response in a bioluminescent ATP assay, therefore comprises a combination of an ATP-degrading enzyme, such as potato apyrase, and one or more enzymes that degrade non-ATP luciferase substrates, together with additional divalent



metal ions, such as Mg^{2+} , if required. This combination of enzymes, or of enzyme activities in at least one material, can be used, for example, to treat a growth medium.

Description of the Invention

5 When the reagent of the invention is used with microbiological growth media, the broth is usually treated within a few hours of preparation so that significant microbial growth in the non-sterile solution does not occur. After the enzyme treatment, the broth can be
10 sterilised as normal, e.g. by filtration or by autoclaving. Autoclaving or heat-treatment will have the effect of destroying the enzyme activities, so high levels of the enzymes can be used yet will not interfere with subsequent assays.

15 Alternatively, the broth and enzymes can be sterile-filtered after being mixed together, which will not destroy the activities. On the one hand, when using a sterile mixture, it is possible to use long treatment times and low enzyme levels to ensure that the added enzymes have a
20 negligible effect on subsequent ATP assays. On the other hand, higher levels of active enzymes may be advantageous in microbial enrichment assays where a sample is added to the treated broth and incubated before assay by bioluminescence, since free ATP, FAD or other luciferase
25 substrate which might be present in the sample will be removed during the incubation step.

 When a sample is to be assayed without the enrichment step, and the sample matrix gives rise to a significant background signal (for instance most foods and drinks, or
30 swabs with food or drink residues), the enzyme reagent can be added beforehand to reduce the background in a similar way to above. Microbial and/or cellular ATP subsequently released can then be directly measured without background interference.

The following Examples illustrate the invention.

Example 1

To 250 ml of freshly-prepared Nutrient Broth No. 2 (Oxoid) was added potato apyrase and snake venom phosphodiesterase I to give final activities of 2 and 0.2 International Units per litre respectively. The mixture was then filtered through a 0.22 μ m membrane filter into a sterile vessel and incubated at 25°C for 16 hours.

The response in the firefly luciferase reaction was measured before and after treatment using HS (High Sensitivity) Luciferase-Luciferin Reagent (Celsis) and a Berthold Biolumat luminometer. Before treatment, the response corresponded to 550 pM ATP; after treatment, it corresponded to less than 0.03 pM ATP.

Example 2

Nutrient Broth No. 2 (Oxoid) from a different batch was prepared using sterile water. Portions were put aside and either apyrase alone (10 I.U./litre) or apyrase (10 I.U./litre) plus snake venom phosphodiesterase I (1 I.U./litre) were added, but this time the mixture was not sterilised. After 1 hour at room temperature, the response in the luciferase assay was recorded as in Example 1. The results were as follows:

	Sample	Equivalent [ATP] (pM)
25	Untreated Nutrient Broth	1490
	+ apyrase, 1 hour	8.6
	+ apyrase + PDE I, 1 hour	1.2

Example 3

Sabouraud liquid medium (Oxoid), prepared with sterile water, was treated at room temperature with various commercial grades of potato apyrase (0.25 I.U./ml) with and without acid phosphatase from sweet potato (0.25 I.U./ml). After two hours, the apparent ATP content was measured by bioluminescence, with the following results:

This Sheet Has Been Replaced



5

	Sample	Equivalent [ATP] (pM)	
		- phosphatase	+ phosphatase
	Untreated broth	9722	N/D
	Apyrase Grade I	51	23
5	Apyrase Grade III	57	24
	Apyrase Grade V	233	24
	Apyrase Grade VII	98	23

10 The amount of apyrase present was enough to reduce the level of genuine ATP to below the limit of detection of the assay. The residual substance or substances giving rise to light emission in the assay were removed to varying extents by apyrase alone, and this may represent different levels of contaminant enzyme activities in the preparations. Addition of acid phosphatase reduced the bioluminescence
15 signal further, to a consistent value.

Example 4

Another batch of Sabouraud liquid medium was prepared with sterile water and divided into three samples. One had no additions; to another was added potato apyrase and sweet
20 potato acid phosphatase (0.25 I.U./ml each); to the third was also added snake venom phosphodiesterase I (0.01 I.U./ml). The mixtures were then sterile-filtered and left at room temperature under sterile conditions for 24 hours, at which point they were assayed by bioluminescence. The
25 results were as follows:

	Sample	Equivalent [ATP] (pM)
	Untreated broth	11060
	+ apyrase + acid phosphatase	18.4
30	+ apyrase + acid phosphatase + PDE I	0.8

CLAIMS

1. A composition comprising an ATP-degrading enzyme and one or more enzymes capable of degrading substances, other than ATP, that are substrates for the light-emitting
5 reaction of firefly luciferase.
2. A composition of claim 1, in which a product of reaction of said one or more enzymes with a non-ATP substrate is ATP.
3. A composition according to claim 1 or claim 2, in
10 which said one or more enzymes have FAD-degrading activity.
4. A composition according to any preceding claim, comprising an ATPase and an enzyme having phosphodiesterase I activity.
5. A composition according to any preceding claim, which
15 comprises an ATPase and an enzyme having phosphatase activity.
6. A composition according to any preceding claim, in which the ATPase is potato apyrase.
7. A composition according to any preceding claim, in
20 freeze-dried form.
8. A composition according to any preceding claim, which additionally comprises Mg^{2+} or other divalent metal ion.
9. A composition according to any preceding claim, which additionally comprises one or more other bioluminescence
25 reagents.
10. A composition according to claim 7 or claim 8, in which the components are provided in kit form, in two or more containers.
11. A method for treating a growth medium, which comprises
30 adding to the medium the enzyme activities defined in any of claims 1 to 6.
12. A method for assaying a sample by bioluminescence assay, which comprises the prior step of adding to the sample the enzyme activities as defined in any of claims 1
35 to 6.

This Sheet Has Been Replaced



PCT/GB 94/0116



7

13. A method according to claim 11 or claim 12, in which the enzyme activities are added as a composition as defined in any of claims 1 to 8.