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COMPOSITIONS AND METHODS FOR INCREASING BONE MINERALIZATION

TECHNICAL FIELD

The present invention relates generally to pharmaceutical products and methods and, more specifically, to methods and compositions suitable for increasing the mineral content of bone. Such compositions and methods may be utilized to treat a wide variety of conditions, including for example, osteopenia, osteoporosis, fractures and other disorders in which low bone mineral density are a hallmark of the disease.

BACKGROUND OF THE INVENTION

Two or three distinct phases of changes to bone mass occur over the life of an individual (see Riggs, *West J. Med. 154*:63-77, 1991). The first phase occurs in both men and women, and proceeds to attainment of a peak bone mass. This first phase is achieved through linear growth of the endochondral growth plates, and radial growth due to a rate of periosteal apposition. The second phase begins around age 30 for trabecular bone (flat bones such as the vertebrae and pelvis) and about age 40 for cortical bone (*e.g.*, long bones found in the limbs) and continues to old age. This phase is characterized by slow bone loss, and occurs in both men and women. In women, a third phase of bone loss also occurs, most likely due to postmenopausal estrogen deficiencies. During this phase alone, women may lose an additional 10% of bone mass from the cortical bone and 25% from the trabecular compartment (see Riggs, *supra*).

Loss of bone mineral content can be caused by a wide variety of conditions, and may result in significant medical problems. For example, osteoporosis is a debilitating disease in humans characterized by marked decreases in skeletal bone mass and mineral density, structural deterioration of bone including degradation of bone microarchitecture and corresponding increases in bone fragility and susceptibility to fracture in afflicted individuals. Osteoporosis in humans is preceded by clinical osteopenia (bone mineral density that is greater than one standard deviation but less than 2.5 standard deviations below the mean value for young adult bone), a condition found in approximately 25 million people in the United States. Another 7-8 million patients in the United States have been diagnosed with clinical osteoporosis (defined as bone mineral content greater than 2.5 standard deviations below that of mature young adult bone). Osteoporosis is one of the most expensive diseases for the health care system, costing tens of billions of dollars annually in the United States. In addition to health care-related costs, long-term residential care and lost working days add to the financial and social costs of this disease. Worldwide approximately 75 million people are at risk for osteoporosis.

The frequency of osteoporosis in the human population increases with age, and among

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Caucasians is predominant in women (who comprise 80% of the osteoporosis patient pool in the United States). The increased fragility and susceptibility to fracture of skeletal bone in the aged is aggravated by the greater risk of accidental falls in this population. More than 1.5 million osteoporosis-related bone fractures are reported in the United States each year. Fractured hips, wrists, and vertebrae are among the most common injuries associated with osteoporosis. Hip fractures in particular are extremely uncomfortable and expensive for the patient, and for women correlate with high rates of mortality and morbidity.

Although osteoporosis has been defined as an increase in the risk of fracture due to decreased bone mass, none of the presently available treatments for skeletal disorders can substantially increase the bone density of adults. There is a strong perception among all physicians that drugs are needed which could increase bone density in adults, particularly in the bones of the wrist, spinal column and hip that are at risk in osteopenia and osteoporosis.

Current strategies for the prevention of osteoporosis may offer some benefit to individuals but cannot ensure resolution of the disease. These strategies include moderating physical activity (particularly in weight-bearing activities) with the onset of advanced age, including adequate calcium in the diet, and avoiding consumption of products containing alcohol or tobacco. For patients presenting with clinical osteopenia or osteoporosis, all current therapeutic drugs and strategies are directed to reducing further loss of bone mass by inhibiting the process of bone absorption, a natural component of the bone remodeling process that occurs constitutively.

For example, estrogen is now being prescribed to retard bone loss. There is, however, some controversy over whether there is any long term benefit to patients and whether there is any effect at all on patients over 75 years old. Moreover, use of estrogen is believed to increase the risk of breast and endometrial cancer.

High doses of dietary calcium, with or without vitamin D has also been suggested for postmenopausal women. However, high doses of calcium can often have unpleasant gastrointestinal side effects, and serum and urinary calcium levels must be continuously monitored (see Khosla and Rigss, *Mayo Clin. Proc. 70*:978-982, 1995).

Other therapeutics which have been suggested include calcitonin, bisphosphonates, anabolic steroids and sodium fluoride. Such therapeutics however, have undesirable side effects (*e.g.*, calcitonin and steroids may cause nausea and provoke an immune reaction, bisphosphonates and sodium fluoride may inhibit repair of fractures, even though bone density increases modestly) that may prevent their usage (see Khosla and Rigss, *supra*).

No currently practiced therapeutic strategy involves a drug that stimulates or enhances the growth of new bone mass. The present invention provides compositions and methods which can be utilized to increase bone mineralization, and thus may be utilized to treat a wide variety of conditions where it is desired to increase bone mass. Further, the present invention provides other, related advantages.

SUMMARY OF THE INVENTION

As noted above, the present invention provides a novel class or family of TGF-beta binding-proteins, as well as assays for selecting compounds which increase bone mineral content and bone mineral density, compounds which increase bone mineral content and bone mineral density and methods for utilizing such compounds in the treatment or prevention of a wide variety of conditions.

Within one aspect of the present invention, isolated nucleic acid molecules are provided, wherein said nucleic acid molecules are selected from the group consisting of: (a) an isolated nucleic acid molecule comprising sequence ID Nos. 1, 5, 7, 9, 11, 13, or, 15, or complementary sequence thereof; (b) an isolated nucleic acid molecule that specifically hybridizes to the nucleic acid molecule of (a) under conditions of high stringency; and (c) an isolated nucleic acid that encodes a TGF-beta binding-protein according to (a) or (b). Within related aspects of the present invention, isolated nucleic acid molecules are provided based upon hybridization to only a portion of one of the above-identified sequences (e.g., for (a) hybridization may be to a probe of at least 20, 25, 50, or 100 nucleotides selected from nucleotides 156 to 539 or 555 to 687 of Sequence ID No. 1). As should be readily evident, the necessary stringency to be utilized for hybridization may vary based upon the size of the probe. For example, for a 25-mer probe high stringency conditions could include: 60 mM Tris pH 8.0, 2 mM EDTA, 5x Denhardt's, 6x SSC, 0.1% (w/v) N-laurylsarcosine, 0.5% (w/v) NP-40 (nonidet P-40) overnight at 45 degrees C, followed by two washes with 0.2x SSC / 0.1% SDS at 45-50 degrees. For a 100-mer probe under low stringency conditions, suitable conditions might include the following: 5x SSPE, 5x Denhardt's, and 0.5% SDS overnight at 42-50 degrees, followed by two washes with 2x SSPE (or 2x SSC) /0.1% SDS at 42-50 degrees.

Within related aspects of the present invention, isolated nucleic acid molecules are provided which have homology to Sequence ID Nos. 1, 5, 7, 9, 11, 13, or 15, at a 50%, 60%, 75%, 80%, 90%, 95%, or 98% level of homology utilizing a Wilbur-Lipman algorithm. Representative examples of such isolated molecules include, for example, nucleic acid molecules which encode a protein comprising Sequence ID NOs. 2, 6, 10, 12, 14, or 16, or have

homology to these sequences at a level of 50%, 60%, 75%, 80%, 90%, 95%, or 98% level of homology utilizing a Lipman-Pearson algorithm.

Isolated nucleic acid molecules are typically less than 100kb in size, and, within certain embodiments, less than 50kb, 25kb, 10kb, or even 5kb in size. Further, isolated nucleic acid molecules, within other embodiments, do not exist in a "library" of other unrelated nucleic acid molecules (*e.g.*, a subclone BAC such as described in GenBank Accession No. AC003098 and EMB No. AQ171546). However, isolated nucleic acid molecules can be found in libraries of related molecules (*e.g.*, for shuffling, such as is described in U.S. Patent Nos. 5,837,458; 5,830,721; and 5,811,238). Finally, isolated nucleic acid molecules as described herein do not include nucleic acid molecules which encode Dan, Cerberus, Gremlin, or SCGF (U.S. Patent No. 5,780,263).

Also provided by the present invention are cloning vectors which contain the above-noted nucleic acid molecules, and expression vectors which comprise a promoter (*e.g.*, a regulatory sequence) operably linked to one of the above-noted nucleic acid molecules. Representative examples of suitable promoters include tissue-specific promoters, and viral – based promoters (*e.g.*, CMV-based promoters such as CMV I-E, SV40 early promoter, and MuLV LTR). Expression vectors may also be based upon, or derived from viruses (*e.g.*, a "viral vector"). Representative examples of viral vectors include herpes simplex viral vectors, adenoviral vectors, adenovirus-associated viral vectors and retroviral vectors. Also provided are host cells containing or comprising any of above-noted vectors (including for example, host cells of human, monkey, dog, rat, or mouse origin).

Within other aspects of the present invention, methods of producing TGF-beta binding-proteins are provided, comprising the step of culturing the aforementioned host cell containing vector under conditions and for a time sufficient to produce the TGF-beta binding protein. Within further embodiments, the protein produced by this method may be further purified (*e.g.*, by column chromatography, affinity purification, and the like). Hence, isolated proteins which are encoded by the above-noted nucleic acid molecules (*e.g.*, Sequence ID NOs. 2, 4, 6, 8, 10, 12, 14, or 16) may be readily produced given the disclosure of the subject application.

It should also be noted that the aforementioned proteins, or fragments thereof, may be produced as fusion proteins. For example, within one aspect fusion proteins are provided comprising a first polypeptide segment comprising a TGF-beta binding-protein encoded by a nucleic acid molecule as described above, or a portion thereof of at least 10, 20, 30, 50, or 100 amino acids in length, and a second polypeptide segment comprising a non-TGF-beta binding-

protein. Within certain embodiments, the second polypeptide may be a tag suitable for purification or recognition (*e.g.*, a polypeptide comprising multiple anionic amino acid residues – see U.S. Patent No. 4,851,341), a marker (*e.g.*, green fluorescent protein, or alkaline phosphatase), or a toxic molecule (*e.g.*, ricin).

Within another aspect of the present invention, antibodies are provided which are capable of specifically binding the above-described class of TGF-beta binding proteins (*e.g.*, human BEER). Within various embodiments, the antibody may be a polyclonal antibody, or a monoclonal antibody (*e.g.*, of human or murine origin). Within further embodiments, the antibody is a fragment of an antibody which retains the binding characteristics of a whole antibody (*e.g.*, an $F(ab')_2$, $F(ab)_2$, Fab', Fab, or Fv fragment, or even a CDR). Also provided are hybridomas and other cells which are capable of producing or expressing the aforementioned antibodies.

Within related aspects of the invention, methods are provided detecting a TGF-beta binding protein, comprising the steps of incubating an antibody as described above under conditions and for a time sufficient to permit said antibody to bind to a TGF-beta binding protein, and detecting the binding. Within various embodiments the antibody may be bound to a solid support to facilitate washing or separation, and/or labeled. (*e.g.*, with a marker selected from the group consisting of enzymes, fluorescent proteins, and radioisotopes).

Within other aspects of the present invention, isolated oligonucleotides are provided which hybridize to a nucleic acid molecule according to Sequence ID NOs. 1, 3, 5, 7, 9, 11, 13, 15, 17, or 18 or the complement thereto, under conditions of high stringency. Within further embodiments, the oligonucleotide may be found in the sequence which encodes Sequence ID Nos. 2, 4, 6, 8, 10, 12, 14, or 16. Within certain embodiments, the oligonucleotide is at least 15, 20, 30, 50, or 100 nucleotides in length. Within further embodiments, the oligonucleotide is labeled with another molecule (*e.g.*, an enzyme, fluorescent molecule, or radioisotope). Also provided are primers which are capable of specifically amplifying all or a portion of the above-mentioned nucleic acid molecules which encode TGF-beta binding-proteins. As utilized herein, the term "specifically amplifying" should be understood to refer to primers which amplify the aforementioned TGF-beta binding-proteins, and not other TGF-beta binding proteins such as Dan, Cerberus, Gremlin, or SCGF (U.S. Patent No. 5,780,263).

Within related aspects of the present invention, methods are provided for detecting a nucleic acid molecule which encodes a TGF-beta binding protein, comprising the steps of incubating an oligonucleotide as described above under conditions of high stringency, and

detecting hybridization of said oligonucleotide. Within certain embodiments, the oligonucleotide may be labeled and/or bound to a solid support.

Within other aspects of the present invention, ribozymes are provided which are capable of cleaving RNA which encodes one of the above-mentioned TGF-beta binding-proteins (*e.g.*, Sequence ID NOs. 2, 6, 8, 10, 12, 14, or 16). Such ribozymes may be composed of DNA, RNA (including 2'-O-methyl ribonucleic acids), nucleic acid analogs (*e.g.*, nucleic acids having phosphorothioate linkages) or mixtures thereof. Also provided are nucleic acid molecules (*e.g.*, DNA or cDNA) which encode these ribozymes, and vectors which are capable of expressing or producing the ribozymes. Representative examples of vectors include plasmids, retrotransposons, cosmids, and viral-based vectors (*e.g.*, viral vectors generated at least in part from a retrovirus, adenovirus, or, adeno-associated virus). Also provided are host cells (*e.g.*, human, dog, rat, or mouse cells) which contain these vectors. In certain embodiments, the host cell may be stably transformed with the vector.

Within further aspects of the invention, methods are provided for producing ribozymes either synthetically, or by *in vitro* or *in vivo* transcription. Within further embodiments, the ribozymes so produced may be further purified and / or formulated into pharmaceutical compositions (*e.g.*, the ribozyme or nucleic acid molecule encoding the ribozyme along with a pharmaceutically acceptable carrier or diluent). Similarly, the antisense oligonucleotides and antibodies or other selected molecules described herein may be formulated into pharmaceutical compositions.

Within other aspects of the present invention, antisense oligonucleotides are provided comprising a nucleic acid molecule which hybridizes to a nucleic acid molecule according to Sequence ID NOs. 1, 3, 5, 7, 9, 11, 13, or 15, or the complement thereto, and wherein said oligonucleotide inhibits the expression of TGF-beta binding protein as described herein (e.g., human BEER). Within various embodiments, the oligonucleotide is 15, 20, 25, 30, 35, 40, or 50 nucleotides in length. Preferably, the oligonucleotide is less than 100, 75, or 60 nucleotides in length. As should be readily evident, the oligonucleotide may be comprised of one or more nucleic acid analogs, ribonucleic acids, or deoxyribonucleic acids. Further, the oligonucleotide may be modified by one or more linkages, including for example, covalent linkage such as a phosphorothioate linkage, a phosphotriester linkage, a methyl phosphonate linkage, a methylene(methylimino) linkage, a morpholino linkage, an amide linkage, a polyamide linkage, a short chain alkyl intersugar linkage, a cycloalkyl intersugar linkage, a short chain heteroatomic intersugar linkage and a heterocyclic intersugar linkage. One representative example of a

chimeric oligonucleotide is provied in U.S. Patent No. 5,989,912.

Within yet another aspect of the present invention, methods are provided for increasing bone mineralization, comprising introducing into a warm-blooded animal an effective amount of the ribozyme as described above. Within related aspects, such methods comprise the step of introducing into a patient an effective amount of the nucleic acid molecule or vector as described herein which is capable of producing the desired ribozyme, under conditions favoring transcription of the nucleic acid molecule to produce the ribozyme.

Within other aspects of the invention transgenic, non-human animals are provided. Within one embodiment a transgenic animal is provided whose germ cells and somatic cells contain a nucleic acid molecule encoding a TGF-beta binding-protein as described above which is operably linked to a promoter effective for the expression of the gene, the gene being introduced into the animal, or an ancestor of the animal, at an embryonic stage, with the proviso that said animal is not a human. Within other embodiments, transgenic knockout animals are provided, comprising an animal whose germ cells and somatic cells comprise a disruption of at least one allele of an endogenous nucleic acid molecule which hybridizes to a nucleic acid molecule which encodes a TGF-binding protein as described herein, wherein the disruption prevents transcription of messenger RNA from said allele as compared to an animal without the disruption, with the proviso that the animal is not a human. Within various embodiments, the disruption is a nucleic acid deletion, substitution, or, insertion. Within other embodiments the transgenic animal is a mouse, rat, sheep, pig, or dog.

Within further aspects of the invention, kits are provided for the detection of TGF-beta binding-protein gene expression, comprising a container that comprises a nucleic acid molecule, wherein the nucleic acid molecule is selected from the group consisting of (a) a nucleic acid molecule comprising the nucleotide sequence of SEQ ID NOs: 1, 3, 5, 7, 9, 11, 13, 15, 100, or 101; (b) a nucleic acid molecule comprising the complement of the nucleotide sequence of (a); (c) a nucleic acid molecule that is a fragment of (a) or (b) of at least 15, 20 30, 50, 75, or, 100 nucleotides in length. Also provided are kits for the detection of a TGF-beta binding-protein which comprise a container that comprise one of the TGF-beta binding protein antibodies described herein.

For example, within one aspect of the present invention methods are provided for determining whether a selected molecule is capable of increasing bone mineral content, comprising the steps of (a) mixing one or more candidate molecules with TGF-beta-binding-protein encoded by the nucleic acid molecule according to claim 1 and a selected member of the

TGF-beta family of proteins (*e.g.*, BMP 5 or 6), (b) determining whether the candidate molecule alters the signaling of the TGF-beta family member, or alters the binding of the TGF-beta binding-protein to the TGF-beta family member. Within certain embodiments, the molecule alters the ability of TGF-beta to function as a positive regulator of mesenchymal cell differentiation. Within this aspect of the present invention, the candidate molecule(s) may alter signaling or binding by, for example, either decreasing (*e.g.*, inhibiting), or increasing (*e.g.*, enhancing) signaling or binding.

Within yet another aspect, methods are provided for determining whether a selected molecule is capable of increasing bone mineral content, comprising the step of determining whether a selected molecule inhibits the binding of TGF-beta binding-protein to bone, or an analogue thereof. Representative examples of bone or analogues thereof include hydroxyapatite and primary human bone samples obtained via biopsy.

Within certain embodiments of the above-recited methods, the selected molecule is contained within a mixture of molecules and the methods may further comprise the step of isolating one or more molecules which are functional within the assay. Within yet other embodiments, TGF-beta family of proteins is bound to a solid support and the binding of TGF-beta binding-protein is measured or TGF-beta binding-protein are bound to a solid support and the binding of TGF-beta proteins are measured.

Utilizing methods such as those described above, a wide variety of molecules may be assayed for their ability to increase bone mineral content by inhibiting the binding of the TGF-beta binding-protein to the TGF-beta family of proteins. Representative examples of such molecules include proteins or peptides, organic molecules, and nucleic acid molecules.

Within other related aspects of the invention, methods are provided for increasing bone mineral content in a warm-blooded animal, comprising the step of administering to a warm-blooded animal a therapeutically effective amount of a molecule identified from the assays recited herein. Within another aspect, methods are provided for increasing bone mineral content in a warm-blooded animal, comprising the step of administering to a warm-blooded animal a therapeutically effective amount of a molecule which inhibits the binding of the TGF-beta binding-protein to the TGF-beta super-family of proteins, including bone morphogenic proteins (BMPs). Representative examples of suitable molecules include antisense molecules, ribozymes, ribozyme genes, and antibodies (e.g., a humanized antibody) which specifically recognize and alter the activity of the TGF-beta binding-protein.

Within another aspect of the present invention, methods are provided for increasing bone

mineral content in a warm-blooded animal, comprising the steps of (a) introducing into cells which home to the bone a vector which directs the expression of a molecule which inhibits the binding of the TGF-beta binding-protein to the TGF-beta family of proteins and bone morphogenic proteins (BMPs), and (b) administering the vector-containing cells to a warm-blooded animal. As utilized herein, it should be understood that cells "home to bone" if they localize within the bone matrix after peripheral administration. Within one embodiment, such methods further comprise, prior to the step of introducing, isolating cells from the marrow of bone which home to the bone. Within a further embodiment, the cells which home to bone are selected from the group consisting of CD34+ cells and osteoblasts.

Within other aspects of the present invention, molecules are provided (preferably isolated) which inhibit the binding of the TGF-beta binding-protein to the TGF-beta super-family of proteins.

Within further embodiments, the molecules may be provided as a composition, and can further comprise an inhibitor of bone resorption. Representative examples of such inhibitors include calcitonin, estrogen, a bisphosphonate, a growth factor having anti-resorptive activity and tamoxifen.

Representative examples of molecules which may be utilized in the afore-mentioned therapeutic contexts include, e.g., ribozymes, ribozyme genes, antisense molecules, and/or antibodies (e.g., humanized antibodies). Such molecules may depending upon their selection, used to alter, antagonize, or agonize the signalling or binding of a TGF-beta binding-protein family member as described herein

Within various embodiments of the invention, the above-described molecules and methods of treatment or prevention may be utilized on conditions such as osteoporosis, osteomalasia, periodontal disease, scurvy, Cushing's Disease, bone fracture and conditions due to limb immobilization and steroid usage.

The present invention also provides antibodies that specifically bind to a TGF-beta binding protein, sclerostin (SOST), and provides immunogens comprising sclerostin peptides derived from regions of sclerostin that interact with a member of the TGF-beta superfamily such as a bone morphogenic protein. In one embodiment, the invention provides an antibody, or an antigen-binding fragment thereof, that binds specifically to a sclerostin polypeptide, said sclerostin polypeptide comprising an amino acid sequence set forth in SEQ ID NO:2, 6, 8, 14, 46, or 65, wherein the antibody competitively inhibits binding of the SOST polypeptide to at least one of (i) a bone morphogenic protein (BMP) Type I Receptor binding site and (ii) a BMP

Type II Receptor binding site, wherein the BMP Type I Receptor binding site is capable of binding to a BMP Type I Receptor polypeptide comprising an amino acid sequence set forth in GenBank Acc. Nos. NM_004329 (SEQ ID NO:102); D89675 (SEQ ID NO:103); NM_001203 (SEQ ID NO:104); S75359 (SEQ ID NO:105); NM_030849 (SEQ ID NO:106); D38082 (SEQ ID NO:107); NP_001194 (SEQ ID NO:108); BAA19765 (SEQ ID NO:109); or AAB33865 (SEQ ID NO:110) and wherein the BMP Type II Receptor binding site is capable of binding to a BMP Type II Receptor polypeptide comprising the amino acid sequence set forth in GenBank Acc. NOs. U25110 (SEQ ID NO:111); NM_033346 (SEQ ID NO:112); Z48923 (SEQ ID NO:114); CAA88759 (SEQ ID NO:115); or NM_001204 (SEQ ID NO:113). In another embodiment, the invention provides an antibody, or an antigen-binding fragment thereof, that binds specifically to a sclerostin polypeptide and that impairs formation of a sclerostin homodimer, wherein the sclerostin polypeptide comprises an amino acid sequence set forth in SEQ ID NOs: 2, 6, 8, 14, 46, or 65.

In certain particular embodiments of the invention, the antibody is a polyclonal antibody. In other embodiments, the antibody is a monoclonal antibody, which is a mouse, human, rat, or hamster monoclonal antibody. The invention also provides a hybridoma cell or a host cell that is capable of producing the monoclonal antibody. In other embodiments of the invention, the antibody is a humanized antibody or a chimeric antibody. The invention further provides a host cell that produces the humanized or chimeric antibody. In certain embodiments the antigen-binding fragment of the antibody is a $F(ab')_2$, Fab', Fab, Fd, or Fv fragment. The invention also provides an antibody that is a single chain antibody and provides a host cell that is capable of expressing the single chain antibody. In another embodiment, the invention provides a composition comprising such antibodies and a physiologically acceptable carrier.

In another embodiment, the invention provides an immunogen comprising a peptide comprising at least 21 consecutive amino acids and no more than 50 consecutive amino acids of a SOST polypeptide, said SOST polypeptide comprising an amino acid sequence set forth in SEQ ID NOs: 2, 6, 8, 14, 46, or 65, wherein the peptide is capable of eliciting in a non-human animal an antibody that binds specifically to the SOST polypeptide and that competitively inhibits binding of the SOST polypeptide to at least one of (i) a bone morphogenic protein (BMP) Type I Receptor binding site and (ii) a BMP Type II Receptor binding site, wherein the BMP Type I Receptor binding site is capable of binding to a BMP Type I Receptor polypeptide comprising an amino acid sequence set forth in GenBank Acc. Nos. NM_004329 (SEQ ID NO:102); D89675 (SEQ ID NO:103); NM_001203 (SEQ ID NO:104); S75359 (SEQ ID

NO:105); NM_030849 (SEQ ID NO:106); D38082 (SEQ ID NO:107); NP_001194 (SEQ ID NO:108); BAA19765 (SEQ ID NO:109); or AAB33865 (SEQ ID NO:110) and wherein the BMP Type II Receptor binding site is capable of binding to a BMP Type II Receptor polypeptide comprising the amino acid sequence set forth in GenBank Acc. NOs. U25110 (SEQ ID NO:111); NM_033346 (SEQ ID NO:112); Z48923 (SEQ ID NO:114); CAA88759 (SEQ ID NO:115); or NM_001204 (SEQ ID NO:113). The invention also provides an immunogen comprising a peptide that comprises at least 21 consecutive amino acids and no more than 50 consecutive amino acids of a SOST polypeptide, said SOST polypeptide comprising an amino acid sequence set forth in SEQ ID NO: 2, 6, 8, 14, 46, or 65, wherein the peptide is capable of eliciting in a non-human animal an antibody that binds specifically to the SOST polypeptide and that impairs formation of a SOST homodimer.

In certain particular embodiments, the subject invention immunogens are associated with a carrier molecule. In certain embodiments, the carrier molecule is a carrier polypeptide, and in particular embodiments, the carrier polypeptide is keyhole limpet hemocyanin.

The invention also provides a method for producing an antibody that specifically binds to a SOST polypeptide, comprising immunizing a non-human animal with an immunogen comprising a peptide comprising at least 21 consecutive amino acids and no more than 50 consecutive amino acids of a SOST polypeptide, wherein (a) the SOST polypeptide comprises an amino acid sequence set forth in SEQ ID NO: 2, 6, 8, 14, 46, or 65; (b) the antibody competitively inhibits binding of the SOST polypeptide to at least one of (i) a bone morphogenic protein (BMP) Type I Receptor binding site and (ii) a BMP Type II Receptor binding site; (c) the BMP Type I Receptor binding site is capable of binding to a BMP Type I Receptor polypeptide comprising the amino acid sequence set forth in GenBank Acc. Nos. NM_004329 (SEQ ID NO:102); D89675 (SEQ ID NO:103); NM_001203 (SEQ ID NO:104); S75359 (SEQ ID NO:105); NM_030849 (SEQ ID NO:106); D38082 (SEQ ID NO:107); NP_001194 (SEQ ID NO:108); BAA19765 (SEQ ID NO:109); or AAB33865 (SEQ ID NO:110); and (d) the BMP Type II Receptor binding site is capable of binding to a BMP Type II Receptor polypeptide comprising the amino acid sequence set forth in GenBank Acc. NOs. U25110 (SEQ ID NO:111); NM_033346 (SEQ ID NO:112); Z48923 (SEQ ID NO:114); CAA88759 (SEQ ID NO:115); or NM_001204 (SEQ ID NO:113).

In another embodiment, the invention provides a method for producing an antibody that specifically binds to a SOST polypeptide, said SOST polypeptide comprising an amino acid sequence set forth in SEQ ID NO: 2, 6, 8, 14, 46, or 65, comprising immunizing a non-human

animal with an immunogen comprising a peptide that comprises at least 21 consecutive amino acids and no more than 50 consecutive amino acids of a SOST polypeptide, said SOST polypeptide comprising an amino acid sequence set forth in SEQ ID NO: 2, 6, 8, 14, 46, or 65, wherein the antibody impairs formation of a SOST homodimer.

These and other aspects of the present invention will become evident upon reference to the following detailed description and attached drawings. In addition, documents including various references set forth herein that describe in more detail certain procedures or compositions (*e.g.*, plasmids, etc.), are incorporated by reference in their entirety.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 is a schematic illustration comparing the amino acid sequence of Human Dan; Human Gremlin; Human Cerberus and Human Beer. Arrows indicate the Cysteine backbone.

Figure 2 summarizes the results obtained from surveying a variety of human tissues for the expression of a TGF-beta binding-protein gene, specifically, the Human Beer gene. A semi-quantitative Reverse Transcription-Polymerase Chain Reaction (RT-PCR) procedure was used to amplify a portion of the gene from first-strand cDNA synthesized from total RNA (described in more detail in EXAMPLE 2A).

Figures 3A-3D summarize the results obtained from RNA *in situ* hybridization of mouse embryo sections, using a cRNA probe that is complementary to the mouse Beer transcript (described in more detail in EXAMPLE 2B). Panel 3A is a transverse section of 10.5 dpc embryo. Panel 3B is a sagittal section of 12.5 dpc embryo and panels 3C and 3D are sagittal sections of 15.5 dpc embryos.

Figures 4A-4C illustrate, by western blot analysis, the specificity of three different polyclonal antibodies for their respective antigens (described in more detail in EXAMPLE 4). Figure 4A shows specific reactivity of an anti-H. Beer antibody for H. Beer antigen, but not H. Dan or H. Gremlin. Figure 4B shows reactivity of an anti-H. Gremlin antibody for H. Gremlin antigen, but not H. Beer or H. Dan. Figure 4C shows reactivity of an anti-H. Dan antibody for H. Dan, but not H. Beer or H. Gremlin.

Figure 5 illustrates, by western blot analysis, the selectivity of the TGF-beta binding-protein, Beer, for BMP-5 and BMP-6, but not BMP-4 (described in more detail in EXAMPLE 5).

Figure 6 demonstrates that the ionic interaction between the TGF-beta binding-protein, Beer, and BMP-5 has a dissociation constant in the 15-30 nM range.

Figure 7 presents an alignment of the region containing the characteristic cystine-knot of a SOST (sclerostin) polypeptide and its closest homologues. Three disulphide bonds that form the cystine-knot are illustrated as solid lines. An extra disulphide bond, shown by a dotted line, is unique to this family, which connects two β-hairpin tips in the 3D structure. The polypeptides depicted are SOST: sclerostin (SEQ ID NO:126); CGHB: Human Chorionic Gonadotropin β (SEQ ID NO:127); FSHB: follicle-stimulating hormone beta subunit (SEQ ID NO:128); TSHB: thyrotropin beta chain precursor (SEQ ID NO:129); VWF: Von Willebrand factor (SEQ ID NO:130); MUC2: human mucin 2 precursor (SEQ ID NO:131); CER1: Cerberus 1 (*Xenopus laevis* homolog) (SEQ ID NO:132); DRM: gremlin (SEQ ID NO:133); DAN: (SEQ ID NO:134); CTGF: connective tissue growth factor precursor (SEQ ID NO:135); NOV: NovH (nephroblastoma overexpressed gene protein homolog) (SEQ ID NO:136); CYR6: (SEQ ID NO:137).

Figure 8 illustrates a 3D model of the core region of SOST (SOST_Core).

Figure 9 presents a 3D model of the core region of SOST homodimer.

Figures 10A and 10B provide an amino acid sequence alignment of Noggin from five different animals: human (NOGG_HUMAN (SEQ ID NO:138); chicken (NOGG_CHICK, SEQ ID NO:139); African clawed frog (NOGG_XENLA, SEQ ID NO:140); NOGG_FUGRU, SEQ ID NO:141); and zebrafish (NOGG_ZEBRA, SEQ ID NO:142); and SOST from human (SOST_HUMAN, SEQ ID NO:46), rat (SOST_RAT, SEQ ID NO:65), and mouse (SOST_Mouse, SEQ ID NO:143).

Figure 11 illustrates the Noggin/BMP-7 complex structure. The BMP homodimer is shown on the bottom portion of the figure in surface mode. The Noggin homodimer is shown on top of the BMP dimer in cartoon mode. The circles outline the N-terminal binding region, the core region, and the linker between the N-terminal and core regions.

Figure 12 depicts a 3D model of the potential BMP-binding fragment located at the SOST N-terminal region. A BMP dimer is shown in surface mode, and the potential BMP-binding fragment is shown in stick mode. A phenylalanine residue fitting into a hydrophobic pocket on the BMP surface is noted.

DETAILED DESCRIPTION OF THE INVENTION

DEFINITIONS

Prior to setting forth the invention in detail, it may be helpful to an understanding thereof to set forth definitions of certain terms and to list and to define the abbreviations that will be used hereinafter.

"Molecule" should be understood to include proteins or peptides (*e.g.*, antibodies, recombinant binding partners, peptides with a desired binding affinity), nucleic acids (*e.g.*, DNA, RNA, chimeric nucleic acid molecules, and nucleic acid analogues such as PNA); and organic or inorganic compounds.

"TGF-beta" should be understood to include any known or novel member of the TGF-beta super-family, which also includes bone morphogenic proteins (BMPs).

"TGF-beta receptor" should be understood to refer to the receptor specific for a particular member of the TGF-beta super-family (including bone morphogenic proteins (BMPs)).

"TGF-beta binding-protein" should be understood to refer to a protein with specific binding affinity for a particular member or subset of members of the TGF-beta super-family (including bone morphogenic proteins (BMPs)). Specific examples of TGF-beta binding-proteins include proteins encoded by Sequence ID Nos. 1, 5, 7, 9, 11, 13, 15, 100, and 101.

Inhibiting the "binding of the TGF-beta binding-protein to the TGF-beta family of proteins and bone morphogenic proteins (BMPs)" should be understood to refer to molecules which allow the activation of TGF-beta or bone morphogenic proteins (BMPs), or allow the binding of TGF-beta family members including bone morphogenic proteins (BMPs) to their respective receptors, by removing or preventing TGF-beta from binding to TGF-binding-protein. Such inhibition may be accomplished, for example, by molecules which inhibit the binding of the TGF-beta binding-protein to specific members of the TGF-beta super-family.

"Vector" refers to an assembly that is capable of directing the expression of desired protein. The vector must include transcriptional promoter elements that are operably linked to the gene(s) of interest. The vector may be composed of deoxyribonucleic acids ("DNA"), ribonucleic acids ("RNA"), or a combination of the two (*e.g.*, a DNA-RNA chimeric). Optionally, the vector may include a polyadenylation sequence, one or more restriction sites, as well as one or more selectable markers such as neomycin phosphotransferase or hygromycin phosphotransferase. Additionally, depending on the host cell chosen and the vector employed, other genetic elements such as an origin of replication, additional nucleic acid restriction sites, enhancers, sequences conferring inducibility of transcription, and selectable markers, may also be incorporated into the vectors described herein.

An "isolated nucleic acid molecule" is a nucleic acid molecule that is not integrated in the genomic DNA of an organism. For example, a DNA molecule that encodes a TGF-binding protein that has been separated from the genomic DNA of a eukaryotic cell is an isolated DNA molecule. Another example of an isolated nucleic acid molecule is a chemically-synthesized nucleic acid

molecule that is not integrated in the genome of an organism. The isolated nucleic acid molecule may be genomic DNA, cDNA, RNA, or composed at least in part of nucleic acid analogs.

An "isolated polypeptide" is a polypeptide that is essentially free from contaminating cellular components, such as carbohydrate, lipid, or other proteinaceous impurities associated with the polypeptide in nature. Preferably, such isolated polypeptides are at least about 90% pure, more preferably at least about 95% pure, and most preferably at least about 99% pure. Within certain embodiments, a particular protein preparation contains an isolated polypeptide if it appears nominally as a single band on SDS-PAGE gel with Coomassie Blue staining. The term "isolated" when referring to organic molecules (*e.g.*, organic small molecules) means that the compounds are greater than 90% pure utilizing methods which are well known in the art (*e.g.*, NMR, melting point).

"Sclerosteosis" is a term that was applied by Hansen (1967) (Hansen, H. G., Sklerosteose. *in*: Opitz, H.; Schmid, F., *Handbuch der Kinderheilkunde*. Berlin: Springer (pub.) 6 1967. Pp. 351-355) to a disorder similar to van Buchem hyperostosis corticalis generalisata but possibly differing in radiologic appearance of the bone changes and in the presence of asymmetric cutaneous syndactyly of the index and middle fingers in many cases. The jaw has an unusually square appearance in this condition.

"Humanized antibodies" are recombinant proteins in which murine or other non-human animal complementary determining regions of monoclonal antibodies have been transferred from heavy and light variable chains of the murine or other non-human animal immunoglobulin into a human variable domain.

As used herein, an "antibody fragment" is a portion of an antibody such as $F(ab')_2$, $F(ab)_2$, Fab', Fab, and the like. Regardless of structure, an antibody fragment binds with the same antigen that is recognized by the intact antibody. For example, an anti-TGF-beta binding-protein monoclonal antibody fragment binds to an epitope of TGF-beta binding-protein.

The term antibody fragment or antigen-binding fragment also includes any synthetic or genetically engineered protein that acts like an antibody by binding to a specific antigen to form a complex. For example, antibody fragments include isolated fragments consisting of the light chain variable region, "Fv" fragments consisting of the variable regions of the heavy and light chains, recombinant single chain polypeptide molecules in which light and heavy variable regions are connected by a peptide linker ("sFv proteins"), and minimal recognition units consisting of the amino acid residues that mimic the hypervariable region.

A "detectable label" is a molecule or atom that can be conjugated to a polypeptide moiety

such as an antibody moiety or a nucleic acid moiety to produce a molecule useful for diagnosis. Examples of detectable labels include chelators, photoactive agents, radioisotopes, fluorescent agents, paramagnetic ions, enzymes, and other marker moieties.

As used herein, an "immunoconjugate" is a molecule comprising an anti-TGF-beta binding-protein antibody, or an antibody fragment, and a detectable label or an effector molecule. Preferably, an immunoconjugate has roughly the same, or only slightly reduced, ability to bind TGF-beta binding-protein after conjugation as before conjugation.

Abbreviations: TGF-beta – "Transforming Growth Factor-beta"; TGF-bBP – "Transforming Growth Factor-beta binding-protein" (one representative TGF-bBP is designated "H. Beer"); BMP – "bone morphogenic protein"; PCR – "polymerase chain reaction"; RT-PCR - PCR process in which RNA is first transcribed into DNA using reverse transcriptase (RT); cDNA - any DNA made by copying an RNA sequence into DNA form.

As noted above, the present invention provides a novel class of TGF-beta binding-proteins, as well as methods and compositions for increasing bone mineral content in warm-blooded animals. Briefly, the present inventions are based upon the unexpected discovery that a mutation in the gene which encodes a novel member of the TGF-beta binding-protein family results in a rare condition (sclerosteosis) characterized by bone mineral contents which are one- to four-fold higher than in normal individuals. Thus, as discussed in more detail below this discovery has led to the development of assays which may be utilized to select molecules which inhibit the binding of the TGF-beta binding-protein to the TGF-beta family of proteins and bone morphogenic proteins (BMPs), and methods of utilizing such molecules for increasing the bone mineral content of warm-blooded animals (including for example, humans).

DISCUSSION OF THE DISEASE KNOWN AS SCLEROSTEOSIS

Sclerosteosis is a disease related to abnormal bone mineral density in humans. Sclerosteosis is a term that was applied by Hansen (1967) (Hansen, H. G., Sklerosteose.In: Opitz, H.; Schmid, F., Handbuch der Kinderheilkunde. Berlin: Springer (pub.) 6 1967. Pp. 351-355) to a disorder similar to van Buchem hyperostosis corticalis generalisata but possibly differing in radiologic appearance of the bone changes and differing in the presence of asymmetric cutaneous syndactyly of the index and middle fingers in many cases.

Sclerosteosis is now known to be an autosomal semi-dominant disorder that is characterized by widely disseminated sclerotic lesions of the bone in the adult. The condition is progressive. Sclerosteosis also has a developmental aspect that is associated with syndactyly (two or more fingers are fused together). The Sclerosteosis Syndrome is associated with large

stature and many affected individuals attain a height of six feet or more. The bone mineral content of homozygotes can be 1 to 6 fold greater than observed in normal individuals, and bone mineral density can be 1 to 4 fold above normal values (*e.g.*, from unaffected siblings).

The Sclerosteosis Syndrome occurs primarily in Afrikaaners of Dutch descent in South Africa. Approximately 1/140 individuals in the Afrikaaner population are carriers of the mutated gene (heterozygotes). The mutation shows 100% penetrance. There are anecdotal reports of increased of bone mineral density in heterozygotes with no associated pathologies (syndactyly or skull overgrowth).

No abnormality of the pituitary-hypothalamus axis has been observed in patients with sclerosteosis. In particular, there appears to be no over-production of growth hormone and cortisone. In addition, sex hormone levels are normal in affected individuals. However, bone turnover markers (osteoblast specific alkaline phosphatase, osteocalcin, type 1 procollagen C' propeptide (PICP), and total alkaline phosphatase; (see Comier, C., *Curr. Opin. in Rheu.* 7:243, 1995) indicate that there is hyperosteoblastic activity associated with the disease but that there is normal to slightly decreased osteoclast activity as measured by markers of bone resorption (pyridinoline, deoxypryridinoline, N-telopeptide, urinary hydroxyproline, plasma tartrate-resistant acid phosphatases and galactosyl hydroxylysine (see Comier, *supra*)).

Sclerosteosis is characterized by the continual deposition of bone throughout the skeleton during the lifetime of the affected individuals. In homozygotes the continual deposition of bone mineral leads to an overgrowth of bone in areas of the skeleton where there is an absence of mechanoreceptors (skull, jaw, cranium). In homozygotes with Sclerosteosis, the overgrowth of the bones of the skull leads to cranial compression and eventually to death due to excessive hydrostatic pressure on the brain stem. In all other parts of the skeleton there is a generalized and diffuse sclerosis. Cortical areas of the long bones are greatly thickened resulting in a substantial increase in bone strength. Trabecular connections are increased in thickness which in turn increases the strength of the trabecular bone. Sclerotic bones appear unusually opaque to x-rays.

As described in more detail in Example 1, the rare genetic mutation that is responsible for the Sclerosteosis syndrome has been localized to the region of human chromosome 17 that encodes a novel member of the TGF-beta binding-protein family (one representative example of which is designated "H. Beer"). As described in more detail below, based upon this discovery, the mechanism of bone mineralization is more fully understood, allowing the development of assays for molecules that increase bone mineralization, and use of such molecules to increase

bone mineral content, and in the treatment or prevention of a wide number of diseases.

TGF-BETA SUPER-FAMILY

The Transforming Growth Factor-beta (TGF-beta) super-family contains a variety of growth factors that share common sequence elements and structural motifs (at both the secondary and tertiary levels). This protein family is known to exert a wide spectrum of biological responses that affect a large variety of cell types. Many of the TGF-beta family members have important functions during the embryonal development in pattern formation and tissue specification; in adults the family members are involved, *e.g.*, in wound healing and bone repair and bone remodeling, and in the modulation of the immune system. In addition to the TGF-beta's, the super-family includes the Bone Morphogenic Proteins (BMPs), Activins, Inhibins, Growth and Differentiation Factors (GDFs), and Glial-Derived Neurotrophic Factors (GDNFs). Primary classification is established through general sequence features that bin a specific protein into a general sub-family. Additional stratification within the sub-family is possible due to stricter sequence conservation between members of the smaller group. In certain instances, such as with BMP-5, BMP-6 and BMP-7, the amino acid identity can be as high as 75% among members of the smaller group. This level of identity enables a single representative sequence to illustrate the key biochemical elements of the sub-group that separates it from other members of the larger family.

The crystal structure of TGF-beta2 has been determined. The general fold of the TGF-beta2 monomer contains a stable, compact, cysteine knotlike structure formed by three disulphide bridges. Dimerization, stabilized by one disulfide bridge, is antiparallel.

TGF-beta signals by inducing the formation of hetero-oligomeric complexes of type I and type II receptors. Transduction of TGF-beta signals involves these two distinct type I and type II subfamilies of transmembrane serine/threonine kinase receptors. At least seven type I receptors and five type II receptors have been identified (see Kawabata et al., *Cytokine Growth Factor Rev.* 9:49-61 (1998); Miyazono et al., *Adv. Immunol.* 75:115-57 (2000). TGF-beta family members initiate their cellular action by binding to receptors with intrinsic serine/threonine kinase activity. Each member of the TGF-beta family binds to a characteristic combination of type I and type II receptors, both of which are needed for signaling. In the current model for TGF-beta receptor activation, a TGF-beta ligand first binds to the type II receptor (TbR-II), which occurs in the cell membrane in an oligomeric form with activated kinase. Thereafter, the type I receptor (TbR-I), which cannot bind ligand in the absence of TbR-II, is recruited into the complex to form a ligand/type II/type I ternary complex. TbR-II then phosphorylates TbR-I

predominantly in a domain rich in glycine and serine residues (GS domain) in the juxtamembrane region, and thereby activates TbR-I. The activated type I receptor kinase then phosphorylates particular members of the Smad family of proteins that translocate to the nucleus where they modulate transcription of specific genes.

BONE MORPHOGENIC PROTEINS (BMPS) ARE KEY REGULATORY PROTEINS IN DETERMINING BONE MINERAL DENSITY IN HUMANS

A major advance in the understanding of bone formation was the identification of the bone morphogenic proteins (BMPs), also known as osteogenic proteins (OPs), which regulate cartilage and bone differentiation in vivo. BMPs/OPs induce endochondral bone differentiation through a cascade of events that include formation of cartilage, hypertrophy and calcification of the cartilage, vascular invasion, differentiation of osteoblasts, and formation of bone. As described above, the BMPs/OPs (BMP 2-14, and osteogenic protein 1 and -2, OP-1 and OP-2) *see, e.g.*, GenBank P12643 (BMP-2); GenBank P12645 (BMP3); GenBank P55107 (BMP-3b, Growth/differentiation factor 10) (GDF-10)); GenBank P12644 (BMP4); GenBank P22003 (BMP5); GenBank P22004 (BMP6); GenBank P18075 (BMP7); GenBank P34820 (BMP8); GenBank Q9UK05 (BMP9); GenBank O95393 (BM10); GenBank O95390 (BMP11, Growth/differentiation factor 11 precursor (GDF-11)); GenBank O95972 (BM15)) are members of the TGF-beta super-family. The striking evolutionary conservation between members the BMP/OP sub-family suggests that they are critical in the normal development and function of animals. Moreover, the presence of multiple forms of BMPs/OPs raises an important question about the biological relevance of this apparent redundancy. In addition to postfetal chondrogenesis and osteogenesis, the BMPs/OPs play multiple roles in skeletogenesis (including the development of craniofacial and dental tissues) and in embryonic development and organogenesis of parenchymatous organs, including the kidney. It is now understood that nature relies on common (and few) molecular mechanisms tailored to provide the emergence of specialized tissues and organs. The BMP/OP super-family is an elegant example of nature parsimony in programming multiple specialized functions deploying molecular isoforms with minor variation in amino acid motifs within highly conserved carboxy-terminal regions.

BMPs are synthesized as large precursor proteins. Upon dimerization, the BMPs are proteolyically cleaved within the cell to yield carboxy-terminal mature proteins that are then secreted from the cell. BMPs, like other TGF-beta family members, initiate signal transduction by binding cooperatively to both type I and type II serine/threonine kinase receptors. Type I receptors for which BMPs may act as ligands include BMPR-IA (also known as ALK-3),

BMPR-IB (also known as ALK-6), ALK-1, and ALK-2 (also known as ActR-I). Of the type II receptors, BMPs bind to BMP type II receptor (BMPR-II), Activin type II (ActR-II), and Activin type IIB (ActR-IIB). (*See* Balemans et al., *supra*, and references cited therein). Polynucleotide sequences and the encoded amino acid sequence of BMP type I receptor polypeptides are provided in the GenBank database, for example, GenBank NM_004329 (SEQ ID NO:102 encoded by SEQ ID NO:116); D89675 (SEQ ID NO:103 encoded by SEQ ID NO:117); NM_001203 (SEQ ID NO:104 encoded by SEQ ID NO:118); S75359 (SEQ ID NO:105 encoded by SEQ ID NO:119); NM_030849 (SEQ ID NO:106 encoded by SEQ ID NO:120); and D38082 (SEQ ID NO:107 encoded by SEQ ID NO:121). Other polypeptide sequences of type I receptors are provided in the GenBank database, for example, NP_001194 (SEQ ID NO:108); BAA19765 (SEQ ID NO:109); and AAB33865 (SEQ ID NO:110). Polynucleotide sequences and the encoded amino acid sequence of BMP type II receptor polypeptides are provided in the GenBank database and include, for example, U25110 (SEQ ID NO:111 encoded by SEQ ID NO:122); NM_033346 (SEQ ID NO:112 encoded by SEQ ID NO:123); NM_001204 (SEQ ID NO:113 encoded by SEQ ID NO:124); and Z48923 (SEQ ID NO:114 encoded by SEQ ID NO:125). Additional polypeptide sequences of type II receptors are also provided in the GenBank database, for example, CAA88759 (SEQ ID NO:115).

BMPs, similar to other cystine-knot proteins, form a homodimer structure (Scheufler et al., *J. Mol. Biol.* 287:103-15 (1999)). According to evolutionary trace analysis performed on the BMP/TGF-β family, the BMP type I receptor binding site and type II receptor binding site were mapped to the surface of the BMP structure (Innis et al., *Protein Eng.* 13:839-47 (2000)). The location of the type I receptor binding site on BMP was later confirmed by the x-ray structure of BMP-2/BMP Receptor IA complex (Nickel et al., *J. Joint Surg. Am.* 83A(Suppl 1(Pt 1)):S7-S14 (2001)). The predicted type II receptor binding site is in good agreement with the x-ray structure of TGF-β3/TGF-β Type II receptor complex (Hart et al., *Nat. Struct. Biol.* 9:203-208 (2002)), which is highly similar to the BMP/BMP Receptor IIA system.

BMP ANTAGONISM

The BMP and Activin sub-families are subject to significant post-translational regulation, such as by TGF-beta binding proteins. An intricate extracellular control system exists, whereby a high affinity antagonist is synthesized and exported, and subsequently complexes selectively with BMPs or activins to disrupt their biological activity (W.C. Smith (1999) TIG *15*(1) 3-6). A number of these natural antagonists have been identified, and on the basis of sequence divergence, the antagonists appear to have evolved independently due to the lack of primary

sequence conservation. Earlier studies of these antagonists highlighted a distinct preference for interacting and neutralizing BMP-2 and BMP-4. In vertebrates, antagonists include noggin, chordin, chordin-*like*, follistatin, FSRP, the DAN/Cerberus protein family, and sclerostin (SOST) (*see* Balemans et al., *supra*, and references cited therein). The mechanism of antagonism or inhibition seems to differ for the different antagonists (Iemura et al. (1998) *Proc. Natl. Acad. Sci. USA 95* 9337-9342).

The type I and type II receptor binding sites on the BMP antagonist noggin have also been mapped. Noggin binds to BMPs with high affinity (Zimmerman *et al.*, 1996). A study of the noggin/BMP-7 complex structure revealed the binding interactions between the two proteins (Groppe et al., *Nature* 420:636-42 (2002)). Superposition of the noggin-BMP-7 structure onto a model of the BMP signaling complex showed that noggin binding effectively masks both pairs of binding epitopes (*i.e.*, BMP Type I and Type II receptor binding sites) on BMP-7. The cysteine-rich scaffold sequence of noggin is preceded by an N-terminal segment of about 20 amino acid residues that are referred to as the "clip" (residues 28-48). The type I receptor-binding site is occluded by the N-terminal portion of the clip domain of Noggin, and the type II receptor binding site is occluded by the carboxy terminal portion of the clip domain. Two β-strands in the core region near the C-terminus of noggin also contact BMP-7 at the type II receptor binding site. This binding mode enables a noggin dimer to efficiently block all the receptor binding sites (two type I and two type II receptor binding sites) on a BMP dimer.

NOVEL TGF-BETA BINDING-PROTEINS

As noted above, the present invention provides a novel class of TGF-beta binding-proteins that possess a nearly identical cysteine (disulfide) scaffold when compared to Human DAN, Human Gremlin, and Human Cerberus, and SCGF (U.S. Patent No. 5,780,263) but almost no homology at the nucleotide level (for background information, see generally Hsu, D.R., Economides, A.N., Wang, X., Eimon, P.M., Harland, R.M., "The *Xenopus* Dorsalizing Factor Gremlin Identifies a Novel Family of Secreted Proteins that Antagonize BMP Activities," *Molecular Cell 1*:673-683, 1998).

Representative example of the novel class of nucleic acid molecules encoding TGF-beta binding-proteins are disclosed in SEQ ID NOs: 1, 5, 7, 9, 11, 13, 15, 100, and 101. The polynucleotides disclosed herein encode a polypeptide called Beer, which is also referred to herein as sclerostin or SOST. Representative members of this class of binding proteins should also be understood to include variants of the TGF-beta binding-protein (*e.g.*, SEQ ID NOs: 5 and 7). As utilized herein, a "TGF-beta binding-protein variant gene" (*e.g.*, an isolated nucleic acid

molecule that encodes a TGF-beta binding protein variant) refers to nucleic acid molecules that encode a polypeptide having an amino acid sequence that is a modification of SEQ ID Nos: 2, 10, 12, 14, 16, 46, or 65. Such variants include naturally-occurring polymorphisms or allelic variants of TGF-beta binding-protein genes, as well as synthetic genes that contain conservative amino acid substitutions of these amino acid sequences. A variety of criteria known to those skilled in the art indicate whether amino acids at a particular position in a peptide or polypeptide are similar. For example, a similar amino acid or a conservative amino acid substitution is one in which an amino acid residue is replaced with an amino acid residue having a similar side chain, which include amino acids with basic side chains (*e.g.*, lysine, arginine, histidine); acidic side chains (*e.g.*, aspartic acid, glutamic acid); uncharged polar side chains (*e.g.*, glycine, asparagine, glutamine, serine, threonine, tyrosine, cysteine, histidine); nonpolar side chains (*e.g.*, alanine, valine, leucine, isoleucine, proline, phenylalanine, methionine, tryptophan); beta-branched side chains (*e.g.*, threonine, valine, isoleucine), and aromatic side chains (*e.g.*, tyrosine, phenylalanine, tryptophan). Proline, which is considered more difficult to classify, shares properties with amino acids that have aliphatic side chains (*e.g.*, Leu, Val, Ile, and Ala). In certain circumstances, substitution of glutamine for glutamic acid or asparagine for aspartic acid may be considered a similar substitution in that glutamine and asparagine are amide derivatives of glutamic acid and aspartic acid, respectively.

Additional variant forms of a TGF-beta binding-protein gene are nucleic acid molecules that contain insertions or deletions of the nucleotide sequences described herein. TGF-beta binding-protein variant genes can be identified by determining whether the genes hybridize with a nucleic acid molecule having the nucleotide sequence of SEQ ID Nos: 1, 5, 7, 9, 11, 13, 15, 100, or 101 under stringent conditions. In addition, TGF-beta binding-protein variant genes should encode a protein having a cysteine backbone.

As an alternative, TGF-beta binding-protein variant genes can be identified by sequence comparison. As used herein, two amino acid sequences have "100% amino acid sequence identity" if the amino acid residues of the two amino acid sequences are the same when aligned for maximal correspondence. Similarly, two nucleotide sequences have "100% nucleotide sequence identity" if the nucleotide residues of the two nucleotide sequences are the same when aligned for maximal correspondence. Sequence comparisons can be performed using standard software programs such as those included in the LASERGENE bioinformatics computing suite, which is produced by DNASTAR (Madison, Wisconsin). Other methods for comparing two nucleotide or amino acid sequences by determining optimal alignment are well-known to those

of skill in the art (see, for example, Peruski and Peruski, *The Internet and the New Biology: Tools for Genomic and Molecular Research* (ASM Press, Inc. 1997), Wu et al. (eds.), "Information Superhighway and Computer Databases of Nucleic Acids and Proteins," in Methods in Gene Biotechnology, pages 123-151 (CRC Press, Inc. 1997), and Bishop (ed.), Guide to Human Genome Computing, 2nd Edition (Academic Press, Inc. 1998)).

A variant TGF-beta binding-protein should have at least a 50% amino acid sequence identity to SEQ ID NOs: 2, 6, 10, 12, 14, 16, 46, or 65 and preferably, greater than 60%, 65%, 70%, 75%, 80%, 85%, 90%, or 95% identity. Alternatively, TGF-beta binding-protein variants can be identified by having at least a 70% nucleotide sequence identity to SEQ ID NOs: 1, 5, 9, 11, 13, 15, 100, or 101. Moreover, the present invention contemplates TGF-beta binding-protein gene variants having greater than 75%, 80%, 85%, 90%, or 95% identity to SEQ ID NO:1 or SEQ ID NO:100. Regardless of the particular method used to identify a TGF-beta binding-protein variant gene or variant TGF-beta binding-protein, a variant TGF-beta binding-protein or a polypeptide encoded by a variant TGF-beta binding-protein gene can be functionally characterized by, for example, its ability to bind to and/or inhibit the signaling of a selected member of the TGF-beta family of proteins, or by its ability to bind specifically to an anti-TGF-beta binding-protein antibody.

The present invention includes functional fragments of TGF-beta binding-protein genes. Within the context of this invention, a "functional fragment" of a TGF-beta binding-protein gene refers to a nucleic acid molecule that encodes a portion of a TGF-beta binding-protein polypeptide which either (1) possesses the above-noted function activity, or (2) specifically binds with an anti-TGF-beta binding-protein antibody. For example, a functional fragment of a TGF-beta binding-protein gene described herein comprises a portion of the nucleotide sequence of SEQ ID Nos: 1, 5, 9, 11, 13, 15, 100, or 101.

2. Isolation of the TGF-beta binding-protein gene

DNA molecules encoding a TGF-beta binding-protein can be obtained by screening a human cDNA or genomic library using polynucleotide probes based upon, for example, SEQ ID NO:1. For example, the first step in the preparation of a cDNA library is to isolate RNA using methods well-known to those of skill in the art. In general, RNA isolation techniques provide a method for breaking cells, a means of inhibiting RNase-directed degradation of RNA, and a method of separating RNA from DNA, protein, and polysaccharide contaminants. For example, total RNA can be isolated by freezing tissue in liquid nitrogen, grinding the frozen tissue with a mortar and pestle to lyse the cells, extracting the ground tissue with a solution of phenol/chloroform to remove

proteins, and separating RNA from the remaining impurities by selective precipitation with lithium chloride (*see*, for example, Ausubel et al. (eds.), *Short Protocols in Molecular Biology, 3rd Edition*, pages 4-1 to 4-6 (John Wiley & Sons 1995) ["Ausubel (1995)"]; Wu et al., *Methods in Gene Biotechnology*, pages 33-41 (CRC Press, Inc. 1997) ["Wu (1997)"]). Alternatively, total RNA can be isolated by extracting ground tissue with guanidinium isothiocyanate, extracting with organic solvents, and separating RNA from contaminants using differential centrifugation (see, for example, Ausubel (1995) at pages 4-1 to 4-6; Wu (1997) at pages 33-41).

In order to construct a cDNA library, poly(A)$^+$ RNA is preferably isolated from a total RNA preparation. Poly(A)$^+$ RNA can be isolated from total RNA by using the standard technique of oligo(dT)-cellulose chromatography (see, for example, Ausubel (1995) at pages 4-11 to 4-12).Double-stranded cDNA molecules may be synthesized from poly(A)$^+$ RNA using techniques well-known to those in the art. (*see*, for example, Wu (1997) at pages 41-46). Moreover, commercially available kits can be used to synthesize double-stranded cDNA molecules (for example, Life Technologies, Inc. (Gaithersburg, Maryland); CLONTECH Laboratories, Inc. (Palo Alto, California); Promega Corporation (Madison, Wisconsin); and Stratagene Cloning Systems (La Jolla, California)).

The basic approach for obtaining TGF-beta binding-protein cDNA clones can be modified by constructing a subtracted cDNA library that is enriched in TGF-binding-protein-specific cDNA molecules. Techniques for constructing subtracted libraries are well-known to those of skill in the art (see, for example, Sargent, "Isolation of Differentially Expressed Genes," in *Meth. Enzymol. 152*:423, 1987; and Wu et al. (eds.), "Construction and Screening of Subtracted and Complete Expression cDNA Libraries," in *Methods in Gene Biotechnology*, pages 29-65 (CRC Press, Inc. 1997)).

Various cloning vectors are appropriate for the construction of a cDNA library. For example, a cDNA library can be prepared in a vector derived from bacteriophage, such as a λ gt10 vector (see, for example, Huynh et al., "Constructing and Screening cDNA Libraries in λ gt10 and λgt11," in *DNA Cloning: A Practical Approach Vol. I*, Glover (ed.), page 49 (IRL Press, 1985); Wu (1997) at pages 47-52). Alternatively, double-stranded cDNA molecules can be inserted into a plasmid vector, such as a pBluescript vector (Stratagene Cloning Systems; La Jolla, California), a LambdaGEM-4 (Promega Corp.; Madison, Wisconsin) or other commercially available vectors. Suitable cloning vectors also can be obtained from the American Type Culture Collection (Rockville, Maryland).

In order to amplify the cloned cDNA molecules, the cDNA library is inserted into a

prokaryotic host, using standard techniques. For example, a cDNA library can be introduced into competent *E. coli* DH5 cells, which can be obtained from Life Technologies, Inc. (Gaithersburg, Maryland).

A human genomic DNA library can be prepared by means well-known in the art (see, for example, Ausubel (1995) at pages 5-1 to 5-6; Wu (1997) at pages 307-327). Genomic DNA can be isolated by lysing tissue with the detergent Sarkosyl, digesting the lysate with proteinase K, clearing insoluble debris from the lysate by centrifugation, precipitating nucleic acid from the lysate using isopropanol, and purifying resuspended DNA on a cesium chloride density gradient.

DNA fragments that are suitable for the production of a genomic library can be obtained by the random shearing of genomic DNA or by the partial digestion of genomic DNA with restriction endonucleases. Genomic DNA fragments can be inserted into a vector, such as a bacteriophage or cosmid vector, in accordance with conventional techniques, such as the use of restriction enzyme digestion to provide appropriate termini, the use of alkaline phosphatase treatment to avoid undesirable joining of DNA molecules, and ligation with appropriate ligases. Techniques for such manipulation are well-known in the art (see, for example, Ausubel (1995) at pages 5-1 to 5-6; Wu (1997) at pages 307-327).

Nucleic acid molecules that encode a TGF-beta binding-protein can also be obtained using the polymerase chain reaction (PCR) with oligonucleotide primers having nucleotide sequences that are based upon the nucleotide sequences of the human TGF-beta binding-protein gene, as described herein. General methods for screening libraries with PCR are provided by, for example, Yu et al., "Use of the Polymerase Chain Reaction to Screen Phage Libraries," in *Methods in Molecular Biology, Vol. 15: PCR Protocols: Current Methods and Applications*, White (ed.), pages 211-215 (Humana Press, Inc. 1993). Moreover, techniques for using PCR to isolate related genes are described by, for example, Preston, "Use of Degenerate Oligonucleotide Primers and the Polymerase Chain Reaction to Clone Gene Family Members," in *Methods in Molecular Biology, Vol. 15: PCR Protocols: Current Methods and Applications*, White (ed.), pages 317-337 (Humana Press, Inc. 1993).

Alternatively, human genomic libraries can be obtained from commercial sources such as Research Genetics (Huntsville, AL) and the American Type Culture Collection (Rockville, Maryland). A library containing cDNA or genomic clones can be screened with one or more polynucleotide probes based upon SEQ ID NO:1, using standard methods as described herein and known in the art (*see, for example*, Ausubel (1995) at pages 6-1 to 6-11).

Anti-TGF-beta binding-protein antibodies, produced as described herein, can also be

used to isolate DNA sequences that encode a TGF-beta binding-protein from cDNA libraries. For example, the antibodies can be used to screen λgt11 expression libraries, or the antibodies can be used for immunoscreening following hybrid selection and translation (*see, for example*, Ausubel (1995) at pages 6-12 to 6-16; Margolis et al., "Screening λ expression libraries with antibody and protein probes," in *DNA Cloning 2: Expression Systems, 2nd Edition*, Glover et al. (eds.), pages 1-14 (Oxford University Press 1995)).

The sequence of a TGF-beta binding-protein cDNA or TGF-beta binding-protein genomic fragment can be determined using standard methods. Moreover, the identification of genomic fragments containing a TGF-beta binding-protein promoter or regulatory element can be achieved using well-established techniques,. such as deletion analysis (*see generally* Ausubel (1995), *supra*).

As an alternative, a TGF-beta binding-protein gene can be obtained by synthesizing DNA molecules using mutually priming long oligonucleotides and the nucleotide sequences described herein (*see*, for example, Ausubel (1995) at pages 8-8 to 8-9). Established techniques using the polymerase chain reaction provide the ability to synthesize DNA molecules at least two kilobases in length (Adang et al., *Plant Molec. Biol. 21*:1131, 1993; Bambot et al., *PCR Methods and Applications 2*:266, 1993; Dillon et al., "Use of the Polymerase Chain Reaction for the Rapid Construction of Synthetic Genes," in *Methods in Molecular Biology, Vol. 15: PCR Protocols: Current Methods and Applications*, White (ed.), pages 263-268, (Humana Press, Inc. 1993); Holowachuk et al., *PCR Methods Appl. 4*:299, 1995).

3. Production of TGF-beta binding-protein genes

Nucleic acid molecules encoding variant TGF-beta binding-protein genes can be obtained by screening various cDNA or genomic libraries with polynucleotide probes having nucleotide sequences based upon SEQ ID NO:1, 5, 9, 11, 13, 15, 100, or 101 using procedures described herein. TGF-beta binding-protein gene variants can also be constructed synthetically. For example, a nucleic acid molecule can be devised that encodes a polypeptide having a conservative amino acid change, compared with the amino acid sequence of SEQ ID NOs: 2, 6, 8, 10, 12, 14, 16, 46, or 65. That is, variants can be obtained that contain one or more amino acid substitutions of SEQ ID NOs: 2, 6, 8, 10, 12, 14, 16, 46, or 65, in which an alkyl amino acid is substituted for an alkyl amino acid in a TGF-beta binding-protein amino acid sequence, an aromatic amino acid is substituted for an aromatic amino acid in a TGF-beta binding-protein amino acid sequence, a sulfur-containing amino acid is substituted for a sulfur-containing amino acid in a TGF-beta binding-protein amino acid sequence, a hydroxy-containing amino acid is

substituted for a hydroxy-containing amino acid in a TGF-beta binding-protein amino acid sequence, an acidic amino acid is substituted for an acidic amino acid in a TGF-beta binding-protein amino acid sequence, a basic amino acid is substituted for a basic amino acid in a TGF-beta binding-protein amino acid sequence, or a dibasic monocarboxylic amino acid is substituted for a dibasic monocarboxylic amino acid in a TGF-beta binding-protein amino acid sequence. Among the common amino acids, for example, a "conservative amino acid substitution" is illustrated by a substitution among amino acids within each of the following groups: (1) glycine, alanine, valine, leucine, and isoleucine, (2) phenylalanine, tyrosine, and tryptophan, (3) serine and threonine, (4) aspartate and glutamate, (5) glutamine and asparagine, and (6) lysine, arginine and histidine. In making such substitutions, it is important, when possible, to maintain the cysteine backbone outlined in Figure 1.

Conservative amino acid changes in a TGF-beta binding-protein gene can be introduced by substituting nucleotides for the nucleotides recited in SEQ ID NO:1, 5, 9, 11, 13, 15, 100, or 101. Such "conservative amino acid" variants can be obtained, for example, by oligonucleotide-directed mutagenesis, linker-scanning mutagenesis, mutagenesis using the polymerase chain reaction, and the like (see Ausubel (1995) at pages 8-10 to 8-22; McPherson (ed.), *Directed Mutagenesis: A Practical Approach* (IRL Press 1991)). The functional ability of such variants can be determined using a standard method, such as the assay described herein. Alternatively, a variant TGF-beta binding-protein polypeptide can be identified by the ability to specifically bind anti-TGF-beta binding-protein antibodies.

Routine deletion analyses of nucleic acid molecules can be performed to obtain "functional fragments" of a nucleic acid molecule that encodes a TGF-beta binding-protein polypeptide. As an illustration, DNA molecules having the nucleotide sequence of SEQ ID NO:1 can be digested with *Bal*31 nuclease to obtain a series of nested deletions. The fragments are then inserted into expression vectors in proper reading frame, and the expressed polypeptides are isolated and tested for activity, or for the ability to bind anti-TGF-beta binding-protein antibodies. One alternative to exonuclease digestion is to use oligonucleotide-directed mutagenesis to introduce deletions or stop codons to specify production of a desired fragment. Alternatively, particular fragments of a TGF-beta binding-protein gene can be synthesized using the polymerase chain reaction.

Standard techniques for functional analysis of proteins are described by, for example, Treuter et al., *Molec. Gen. Genet. 240*:113, 1993; Content et al., "Expression and preliminary deletion analysis of the 42 kDa 2-5A synthetase induced by human interferon," in *Biological*

Interferon Systems, Proceedings of ISIR-TNO Meeting on Interferon Systems, Cantell (ed.), pages 65-72 (Nijhoff 1987); Herschman, "The EGF Receptor," in *Control of Animal Cell Proliferation, Vol. 1*, Boynton et al., (eds.) pages 169-199 (Academic Press 1985); Coumailleau et al., *J. Biol. Chem. 270*:29270, 1995; Fukunaga et al., *J. Biol. Chem. 270*:25291, 1995; Yamaguchi et al., *Biochem. Pharmacol. 50*:1295, 1995; Meisel et al., *Plant Molec. Biol. 30*:1, 1996.

The present invention also contemplates functional fragments of a TGF-beta binding-protein gene that have conservative amino acid changes.

A TGF-beta binding-protein variant gene can be identified on the basis of structure by determining the level of identity with nucleotide and amino acid sequences of SEQ ID NOs: 1, 5, 9, 11, 13, 15, 100, or 101 and 2, 6, 10, 12, 14, 16, 46, or 65 as discussed above. An alternative approach to identifying a variant gene on the basis of structure is to determine whether a nucleic acid molecule encoding a potential variant TGF-beta binding-protein gene can hybridize under stringent conditions to a nucleic acid molecule having the nucleotide sequence of SEQ ID Nos: 1, 5, 9, 11, 13, 15, 100, or 101, or a portion thereof of at least 15 or 20 nucleotides in length. As an illustration of stringent hybridization conditions, a nucleic acid molecule having a variant TGF-beta binding-protein sequence can bind with a fragment of a nucleic acid molecule having a sequence from SEQ ID NO:1 in a buffer containing, for example, 5xSSPE (1xSSPE = 180 mM sodium chloride, 10 mM sodium phosphate, 1 mM EDTA (pH 7.7), 5xDenhardt's solution (100xDenhardt's = 2% (w/v) bovine serum albumin, 2% (w/v) Ficoll, 2% (w/v) polyvinylpyrrolidone) and 0.5% SDS incubated overnight at 55-60°C. Post-hybridization washes at high stringency are typically performed in 0.5xSSC (1xSSC = 150 mM sodium chloride, 15 mM trisodium citrate) or in 0.5xSSPE at 55-60°C.

Regardless of the particular nucleotide sequence of a variant TGF-beta binding-protein gene, the gene encodes a polypeptide that can be characterized by its functional activity, or by the ability to bind specifically to an anti-TGF-beta binding-protein antibody. More specifically, variant TGF-beta binding-protein genes encode polypeptides which exhibit at least 50%, and preferably, greater than 60, 70, 80 or 90%, of the activity of polypeptides encoded by the human TGF-beta binding-protein gene described herein.

4. Production of TGF-beta binding-protein in Cultured Cells

To express a TGF-beta binding-protein gene, a nucleic acid molecule encoding the polypeptide must be operably linked to regulatory sequences that control transcriptional expression in an expression vector and then introduced into a host cell. In addition to transcriptional regulatory

sequences, such as promoters and enhancers, expression vectors can include translational regulatory sequences and a marker gene that is suitable for selection of cells that carry the expression vector. Expression vectors that are suitable for production of a foreign protein in eukaryotic cells typically contain (1) prokaryotic DNA elements coding for a bacterial replication origin and an antibiotic resistance marker to provide for the growth and selection of the expression vector in a bacterial host; (2) eukaryotic DNA elements that control initiation of transcription, such as a promoter; and (3) DNA elements that control the processing of transcripts, such as a transcription termination/polyadenylation sequence.

TGF-beta binding-proteins of the present invention are preferably expressed in mammalian cells. Examples of mammalian host cells include African green monkey kidney cells (Vero; ATCC CRL 1587), human embryonic kidney cells (293-HEK; ATCC CRL 1573), baby hamster kidney cells (BHK-21; ATCC CRL 8544), canine kidney cells (MDCK; ATCC CCL 34), Chinese hamster ovary cells (CHO-K1; ATCC CCL61), rat pituitary cells (GH1; ATCC CCL82), HeLa S3 cells (ATCC CCL2.2), rat hepatoma cells (H-4-II-E; ATCC CRL 1548) SV40-transformed monkey kidney cells (COS-1; ATCC CRL 1650) and murine embryonic cells (NIH-3T3; ATCC CRL 1658).

For a mammalian host, the transcriptional and translational regulatory signals may be derived from viral sources, such as adenovirus, bovine papilloma virus, simian virus, or the like, in which the regulatory signals are associated with a particular gene which has a high level of expression. Suitable transcriptional and translational regulatory sequences also can be obtained from mammalian genes, such as actin, collagen, myosin, and metallothionein genes.

Transcriptional regulatory sequences include a promoter region sufficient to direct the initiation of RNA synthesis. Suitable eukaryotic promoters include the promoter of the mouse metallothionein I gene [Hamer et al., *J. Molec. Appl. Genet. 1*:273, 1982], the *TK* promoter of *Herpes* virus [McKnight, *Cell 31*:355, 1982], the *SV40* early promoter [Benoist et al., *Nature 290*:304, 1981], the *Rous* sarcoma virus promoter [Gorman et al., *Proc. Nat'l Acad. Sci. USA 79*:6777, 1982], the cytomegalovirus promoter [Foecking et al., *Gene 45*:101, 1980], and the mouse mammary tumor virus promoter (see, generally, Etcheverry, "Expression of Engineered Proteins in Mammalian Cell Culture," in *Protein Engineering: Principles and Practice*, Cleland et al. (eds.), pages 163-181 (John Wiley & Sons, Inc. 1996)).

Alternatively, a prokaryotic promoter, such as the bacteriophage T3 RNA polymerase promoter, can be used to control TGF-beta binding-protein gene expression in mammalian cells if the prokaryotic promoter is regulated by a eukaryotic promoter (Zhou et al., *Mol. Cell. Biol.*

10:4529, 1990; Kaufman et al., *Nucleic Acids Res. 19*:4485, 1991).

TGF-beta binding-protein genes may also be expressed in bacterial, yeast, insect, or plant cells. Suitable promoters that can be used to express TGF-beta binding-protein polypeptides in a prokaryotic host are well-known to those of skill in the art and include promoters capable of recognizing the T4, T3, Sp6 and T7 polymerases, the P_R and P_L promoters of bacteriophage lambda, the *trp*, *recA*, heat shock, *lacUV5*, *tac*, *lpp-lacSpr*, *phoA*, and *lacZ* promoters of *E. coli*, promoters of *B. subtilis*, the promoters of the bacteriophages of *Bacillus*, *Streptomyces* promoters, the *int* promoter of bacteriophage lambda, the *bla* promoter of pBR322, and the CAT promoter of the chloramphenicol acetyl transferase gene. Prokaryotic promoters have been reviewed by Glick, *J. Ind. Microbiol. 1*:277, 1987, Watson et al., *Molecular Biology of the Gene, 4th Ed.* (Benjamin Cummins 1987), and by Ausubel et al. (1995).

Preferred prokaryotic hosts include *E. coli* and *Bacillus subtilus*. Suitable strains of *E. coli* include BL21(DE3), BL21(DE3)pLysS, BL21(DE3)pLysE, DH1, DH4I, DH5, DH5I, DH5IF', DH5IMCR, DH10B, DH10B/p3, DH11S, C600, HB101, JM101, JM105, JM109, JM110, K38, RR1, Y1088, Y1089, CSH18, ER1451, and ER1647 (see, for example, Brown (Ed.), *Molecular Biology Labfax* (Academic Press 1991)). Suitable strains of *Bacillus subtilus* include BR151, YB886, MI119, MI120, and B170 (see, for example, Hardy, "Bacillus Cloning Methods," in *DNA Cloning: A Practical Approach*, Glover (Ed.) (IRL Press 1985)).

Methods for expressing proteins in prokaryotic hosts are well-known to those of skill in the art (*see*, for example, Williams et al., "Expression of foreign proteins in *E. coli* using plasmid vectors and purification of specific polyclonal antibodies," in *DNA Cloning 2: Expression Systems, 2nd Edition*, Glover et al. (eds.), page 15 (Oxford University Press 1995); Ward et al., "Genetic Manipulation and Expression of Antibodies," in *Monoclonal Antibodies: Principles and Applications*, page 137 (Wiley-Liss, Inc. 1995); and Georgiou, "Expression of Proteins in Bacteria," in *Protein Engineering: Principles and Practice*, Cleland et al. (eds.), page 101 (John Wiley & Sons, Inc. 1996)).

The baculovirus system provides an efficient means to introduce cloned *TGF-beta binding-protein* genes into insect cells. Suitable expression vectors are based upon the *Autographa californica* multiple nuclear polyhedrosis virus (AcMNPV), and contain well-known promoters such as *Drosophila* heat shock protein (hsp) 70 promoter, *Autographa californica* nuclear polyhedrosis virus immediate-early gene promoter (*ie-1*) and the delayed early *39K* promoter, baculovirus p10 promoter, and the *Drosophila* metallothionein promoter. Suitable insect host cells include cell lines derived from IPLB-*Sf*-21, a *Spodoptera frugiperda*

pupal ovarian cell line, such as *Sf*9 (ATCC CRL 1711), *Sf*21AE, and *Sf*21 (Invitrogen Corporation; San Diego, CA), as well as *Drosophila* Schneider-2 cells. Established techniques for producing recombinant proteins in baculovirus systems are provided by Bailey et al., "Manipulation of Baculovirus Vectors," in *Methods in Molecular Biology, Volume 7: Gene Transfer and Expression Protocols*, Murray (ed.), pages 147-168 (The Humana Press, Inc. 1991), by Patel et al., "The baculovirus expression system," in *DNA Cloning 2: Expression Systems, 2nd Edition*, Glover *et al.* (eds.), pages 205-244 (Oxford University Press 1995), by Ausubel (1995) at pages 16-37 to 16-57, by Richardson (ed.), *Baculovirus Expression Protocols* (The Humana Press, Inc. 1995), and by Lucknow, "Insect Cell Expression Technology," in *Protein Engineering: Principles and Practice*, Cleland et al. (eds.), pages 183-218 (John Wiley & Sons, Inc. 1996).

Promoters for expression in yeast include promoters from *GAL1* (galactose), *PGK* (phosphoglycerate kinase), *ADH* (alcohol dehydrogenase), *AOX1* (alcohol oxidase), HIS4 (histidinol dehydrogenase), and the like. Many yeast cloning vectors have been designed and are readily available. These vectors include YIp-based vectors, such as YIp5, YRp vectors, such as YRp17, YEp vectors such as YEp13 and YCp vectors, such as YCp19. One skilled in the art will appreciate that there are a wide variety of suitable vectors for expression in yeast cells.

Expression vectors can also be introduced into plant protoplasts, intact plant tissues, or isolated plant cells. General methods of culturing plant tissues are provided, for example, by Miki et al., "Procedures for Introducing Foreign DNA into Plants," in *Methods in Plant Molecular Biology and Biotechnology*, Glick et al. (eds.), pages 67-88 (CRC Press, 1993).

An expression vector can be introduced into host cells using a variety of standard techniques including calcium phosphate transfection, liposome-mediated transfection, microprojectile-mediated delivery, electroporation, and the like. Preferably, the transfected cells are selected and propagated to provide recombinant host cells that comprise the expression vector stably integrated in the host cell genome. Techniques for introducing vectors into eukaryotic cells and techniques for selecting such stable transformants using a dominant selectable marker are described, for example, by Ausubel (1995) and by Murray (ed.), *Gene Transfer and Expression Protocols* (Humana Press 1991). Methods for introducing expression vectors into bacterial, yeast, insect, and plant cells are also provided by Ausubel (1995).

General methods for expressing and recovering foreign protein produced by a mammalian cell system is provided by, for example, Etcheverry, "Expression of Engineered Proteins in Mammalian Cell Culture," in *Protein Engineering: Principles and Practice*, Cleland et al. (eds.),

pages 163 (Wiley-Liss, Inc. 1996). Standard techniques for recovering protein produced by a bacterial system is provided by, for example, Grisshammer *et al.*, "Purification of over-produced proteins from *E. coli* cells," in *DNA Cloning 2: Expression Systems, 2nd Edition*, Glover *et al.* (eds.), pages 59-92 (Oxford University Press 1995). Established methods for isolating recombinant proteins from a baculovirus system are described by Richardson (ed.), *Baculovirus Expression Protocols* (The Humana Press, Inc., 1995).

More generally, TGF-beta binding-protein can be isolated by standard techniques, such as affinity chromatography, size exclusion chromatography, ion exchange chromatography, HPLC and the like. Additional variations in TGF-beta binding-protein isolation and purification can be devised by those of skill in the art. For example, anti-TGF-beta binding-protein antibodies, obtained as described below, can be used to isolate large quantities of protein by immunoaffinity purification.

5. Production of Antibodies to TGF-beta binding-proteins

The present invention provides antibodies that specifically bind to sclerostin as described herein in detail. Antibodies to TGF-beta binding-protein can be obtained, for example, using the product of an expression vector as an antigen. Antibodies that specifically bind to sclerostin may also be prepared by using peptides derived from any one of the sclerostin polypeptide sequences provided herein (SEQ ID NOs: 2, 6, 8, 10, 12, 14, 16, 46, and 65). Particularly useful anti-TGF-beta binding-protein antibodies "bind specifically" with TGF-beta binding-protein of Sequence ID Nos. 2, 6, 8, 10, 12, 14, 16, 46, or 65 but not to other TGF-beta binding-proteins such as Dan, Cerberus, SCGF, or Gremlin. Antibodies of the present invention (including fragments and derivatives thereof) may be a polyclonal or, especially a monoclonal antibody. The antibody may belong to any immunoglobulin class, and may be for example an IgG, (including isotypes of IgG, which for human antibodies are known in the art as IgG_1, IgG_2, IgG_3, IgG_4); IgE; IgM; or IgA antibody. An antibody may be obtained from fowl or mammals, preferably, for example, from a murine, rat, human or other primate antibody. When desired the antibody may be an internalising antibody.

Polyclonal antibodies to recombinant TGF-beta binding-protein can be prepared using methods well-known to those of skill in the art (see, for example, Green et al., "Production of Polyclonal Antisera," in *Immunochemical Protocols* (Manson, ed.), pages 1-5 (Humana Press 1992); Williams et al., "Expression of foreign proteins in *E. coli* using plasmid vectors and purification of specific polyclonal antibodies," in *DNA Cloning 2: Expression Systems, 2nd Edition*, Glover et al. (eds.), page 15 (Oxford University Press 1995)). Although polyclonal

antibodies are typically raised in animals such as rats, mice, rabbits, goats, or sheep, an anti-TGF-beta binding-protein antibody of the present invention may also be derived from a subhuman primate antibody. General techniques for raising diagnostically and therapeutically useful antibodies in baboons may be found, for example, in Goldenberg et al., international patent publication No. WO 91/11465 (1991), and in Losman et al., *Int. J. Cancer 46*:310, 1990.

The antibody should comprise at least a variable region domain. The variable region domain may be of any size or amino acid composition and will generally comprise at least one hypervariable amino acid sequence responsible for antigen binding embedded in a framework sequence. In general terms the variable (V) region domain may be any suitable arrangement of immunoglobulin heavy (V_H) and/or light (V_L) chain variable domains. Thus for example the V region domain may be monomeric and be a V_H or V_L domain where these are capable of independently binding antigen with acceptable affinity. Alternatively the V region domain may be dimeric and contain V_H-V_H, V_H-V_L, or V_L-V_L, dimers in which the V_H and V_L chains are non-covalently associated (abbreviated hereinafter as F_v). Where desired, however, the chains may be covalently coupled either directly, for example via a disulphide bond between the two variable domains, or through a linker, for example a peptide linker, to form a single chain domain (abbreviated hereinafter as scF_v).

The variable region domain may be any naturally occuring variable domain or an engineered version thereof. By engineered version is meant a variable region domain that has been created using recombinant DNA engineering techniques. Such engineered versions include those created for example from natural antibody variable regions by insertions, deletions or changes in or to the amino acid sequences of the natural antibodies. Particular examples of this type include those engineered variable region domains containing at least one CDR and optionally one or more framework amino acids from one antibody and the remainder of the variable region domain from a second antibody.

The variable region domain may be covalently attached at a C-terminal amino acid to at least one other antibody domain or a fragment thereof. Thus, for example where a V_H domain is present in the variable region domain this may be linked to an immunoglobulin C_H1 domain or a fragment thereof. Similarly a V_L domain may be linked to a C_K domain or a fragment thereof. In this way for example the antibody may be a Fab fragment wherein the antigen binding domain contains associated V_H and V_L domains covalently linked at their C-termini to a CH1 and C_K domain respectively. The CH1 domain may be extended with further amino acids, for example to provide a hinge region domain as found in a Fab' fragment, or to provide further domains,

such as antibody CH2 and CH3 domains.

Another form of an antibody fragment is a peptide coding for a single complementarity-determining region (CDR). CDR peptides ("minimal recognition units") can be obtained by constructing genes encoding the CDR of an antibody of interest. Such genes are prepared, for example, by using the polymerase chain reaction to synthesize the variable region from RNA of antibody-producing cells (see, for example, Larrick et al., *Methods: A Companion to Methods in Enzymology* 2:106, 1991; Courtenay-Luck, "Genetic Manipulation of Monoclonal Antibodies," in *Monoclonal Antibodies: Production, Engineering and Clinical Application*, Ritter et al. (eds.), page 166 (Cambridge University Press 1995); and Ward et al., "Genetic Manipulation and Expression of Antibodies," in *Monoclonal Antibodies: Principles and Applications*, Birch et al., (eds.), page 137 (Wiley-Liss, Inc. 1995)).

Antibodies for use in the invention may in general be monoclonal (prepared by conventional immunisation and cell fusion procedures) or in the case of fragments, derived therefrom using any suitable standard chemical such as reduction or enzymatic cleavage and/or digestion techniques, for example by treatment with pepsin. More specifically, monoclonal anti-TGF-beta binding-protein antibodies can be generated utilizing a variety of techniques. Rodent monoclonal antibodies to specific antigens may be obtained by methods known to those skilled in the art (see, for example, Kohler et al., *Nature 256*:495, 1975; and Coligan et al. (eds.), *Current Protocols in Immunology*, *1*:2.5.1-2.6.7 (John Wiley & Sons 1991) ["Coligan"]; Picksley et al., "Production of monoclonal antibodies against proteins expressed in *E. coli*," in *DNA Cloning 2: Expression Systems, 2nd Edition*, Glover et al. (eds.), page 93 (Oxford University Press 1995)).

Briefly, monoclonal antibodies can be obtained by injecting mice with a composition comprising a TGF-beta binding-protein gene product, verifying the presence of antibody production by removing a serum sample, removing the spleen to obtain B-lymphocytes, fusing the B-lymphocytes with myeloma cells to produce hybridomas, cloning the hybridomas, selecting positive clones which produce antibodies to the antigen, culturing the clones that produce antibodies to the antigen, and isolating the antibodies from the hybridoma cultures.

In addition, an anti-TGF-beta binding-protein antibody of the present invention may be derived from a human monoclonal antibody. Human monoclonal antibodies are obtained from transgenic mice that have been engineered to produce specific human antibodies in response to antigenic challenge. In this technique, elements of the human heavy and light chain locus are introduced into strains of mice derived from embryonic stem cell lines that contain targeted

disruptions of the endogenous heavy chain and light chain loci. The transgenic mice can synthesize human antibodies specific for human antigens, and the mice can be used to produce human antibody-secreting hybridomas. Methods for obtaining human antibodies from transgenic mice are described, for example, by Green et al., *Nature Genet. 7*:13, 1994; Lonberg et al., *Nature 368*:856, 1994; and Taylor et al., *Int. Immun. 6*:579, 1994.

Monoclonal antibodies can be isolated and purified from hybridoma cultures by a variety of well-established techniques. Such isolation techniques include affinity chromatography with Protein-A Sepharose, size-exclusion chromatography, and ion-exchange chromatography (see, for example, Coligan at pages 2.7.1-2.7.12 and pages 2.9.1-2.9.3; Baines et al., "Purification of Immunoglobulin G (IgG)," in *Methods in Molecular Biology, Vol. 10*, pages 79-104 (The Humana Press, Inc. 1992)).

For particular uses, it may be desirable to prepare fragments of anti-TGF-beta binding-protein antibodies. Such antibody fragments can be obtained, for example, by proteolytic hydrolysis of the antibody. Antibody fragments can be obtained by pepsin or papain digestion of whole antibodies by conventional methods. As an illustration, antibody fragments can be produced by enzymatic cleavage of antibodies with pepsin to provide a 5S fragment denoted $F(ab')_2$. This fragment can be further cleaved using a thiol reducing agent to produce 3.5S Fab' monovalent fragments. Optionally, the cleavage reaction can be performed using a blocking group for the sulfhydryl groups that result from cleavage of disulfide linkages. As an alternative, an enzymatic cleavage using pepsin produces two monovalent Fab fragments and an Fc fragment directly. These methods are described, for example, by Goldenberg, U.S. patent No. 4,331,647, Nisonoff et al., *Arch Biochem. Biophys. 89*:230, 1960, Porter, *Biochem. J. 73*:119, 1959, Edelman et al., in *Methods in Enzymology 1*:422 (Academic Press 1967), and by Coligan at pages 2.8.1-2.8.10 and 2.10.-2.10.4.

Other methods of cleaving antibodies, such as separation of heavy chains to form monovalent light-heavy chain fragments, further cleavage of fragments, or other enzymatic, chemical or genetic techniques may also be used, so long as the fragments bind to the antigen that is recognized by the intact antibody.

Alternatively, the antibody may be a recombinant or engineered antibody obtained by the use of recombinant DNA techniques involving the manipulation and re-expression of DNA encoding antibody variable and/or constant regions. Such DNA is known and/or is readily available from DNA libraries including for example phage-antibody libraries (see Chiswell, D J and McCafferty, J. Tibtech. 10 80-84 (1992)) or where desired can be synthesised. Standard

molecular biology and/or chemistry procedures may be used to sequence and manipulate the DNA, for example, to introduce codons to create cysteine residues, to modify, add or delete other amino acids or domains as desired.

One or more replicable expression vectors containing the DNA encoding a variable and/or constant region may be prepared and used to transform an appropriate cell line, e.g. a non-producing myeloma cell line, such as a mouse NSO line or a bacterial, such as *E.coli*, in which production of the antibody will occur. In order to obtain efficient transcription and translation, the DNA sequence in each vector should include appropriate regulatory sequences, particularly a promoter and leader sequence operably linked to a variable domain sequence. Particular methods for producing antibodies in this way are generally well known and routinely used. For example, basic molecular biology procedures are described by Maniatis *et al* (Molecular Cloning, Cold Spring Harbor Laboratory, New York, 1989); DNA sequencing can be performed as described in Sanger *et al* (*Proc. Natl. Acad. Sci. USA* 74: 5463, (1977)) and the Amersham International plc sequencing handbook; site directed mutagenesis can be carried out according to the method of Kramer et al. (*Nucleic Acids Res.* 12, 9441, (1984)); the Anglian Biotechnology Ltd handbook; Kunkel *Proc. Natl. Acad. Sci. USA* 82:488-92 (1985); Kunkel et al., *Methods in Enzymol.* 154:367-82 (1987). Additionally, numerous publications detail techniques suitable for the preparation of antibodies by manipulation of DNA, creation of expression vectors, and transformation of appropriate cells, for example as reviewed by Mountain A and Adair, J R in Biotechnology and Genetic Engineering Reviews (ed. Tombs, M P, 10, Chapter 1, 1992, Intercept, Andover, UK) and in International Patent Specification No. WO 91/09967.

In certain embodiments, the antibody according to the invention may have one or more effector or reporter molecules attached to it and the invention extends to such modified proteins. A reporter molecule may be a detectable moiety or label such as an enzyme, cytotoxic agent or other reporter molecule, including a dye, radionuclide, luminescent group, fluorescent group, or biotin, or the like. The TGF-beta binding protein-specific immunoglobulin or fragment thereof may be radiolabeled for diagnostic or therapeutic applications. Techniques for radiolabeling of antibodies are known in the art. *See, e.g.*, Adams 1998 *In Vivo* 12:11-21; Hiltunen 1993 *Acta Oncol. 32*:831-9. Therapeutic applications are described in greater detail below and may include use of the TGF-beta binding protein specific antibody (or fragment thereof) in conjunction with other therapeutic agents. The effector or reporter molecules may be attached to the antibody through any available amino acid side-chain, terminal amino acid or, where present carbohydrate

functional group located in the antibody, provided that the attachment or the attachment process does not adversely affect the binding properties and the usefulness of the molecule. Particular functional groups include, for example any free amino, imino, thiol, hydroxyl, carboxyl or aldehyde group. Attachment of the antibody and the effector and/or reporter molecule(s) may be achieved via such groups and an appropriate functional group in the effector or reporter molecules. The linkage may be direct or indirect through spacing or bridging groups.

Effector molecules include, for example, antineoplastic agents, toxins (such as enzymatically active toxins of bacterial (such as *P. aeruginosa* exotoxin A) or plant origin and fragments thereof (*e.g.* ricin and fragments thereof; plant gelonin, bryodin from *Bryonia dioica*, or the like. *See, e.g.*, Thrush et al., 1996 *Annu. Rev. Immunol. 14*:49-71; Frankel et al., 1996 *Cancer Res. 56*:926-32); biologically active proteins, for example enzymes; nucleic acids and fragments thereof such as. DNA, RNA and fragments thereof; naturally occurring and synthetic polymers (*e.g.*, polysaccharides and polyalkylene polymers such as poly(ethylene glycol) and derivatives thereof); radionuclides, particularly radioiodide; and chelated metals. Suitable reporter groups include chelated metals, fluorescent compounds, or compounds that may be detected by NMR or ESR spectroscopy. Particularly useful effector groups are calichaemicin and derivatives thereof (*see*, for example, South African Patent Specifications Nos. 85/8794, 88/8127 and 90/2839).

Numerous other toxins, including chemotherapeutic agents, anti-mitotic agents, antibiotics, inducers of apoptosis (or "apoptogens", *see, e.g.*, Green and Reed, 1998, *Science 281*:1309-1312), or the like, are known to those familiar with the art, and the examples provided herein are intended to be illustrative without limiting the scope and spirit of the invention. Particular antineoplastic agents include cytotoxic and cytostatic agents, for example alkylating agents, such as nitrogen mustards (*e.g.*, chlorambucil, melphalan, mechlorethamine, cyclophosphamide, or uracil mustard) and derivatives thereof, triethylenephosphoramide, triethylenethiophosphor-amide, busulphan, or cisplatin; antimetabolites, such as methotrexate, fluorouracil, floxuridine, cytarabine, mercaptopurine, thioguanine, fluoroacetic acid or fluorocitric acid, antibiotics, such as bleomycins (*e.g.*, bleomycin sulphate), doxorubicin, daunorubicin, mitomycins (*e.g.*, mitomycin C), actinomycins (*e.g.*, dactinomycin) plicamycin, calichaemicin and derivatives thereof, or esperamicin and derivatives thereof; mitotic inhibitors, such as etoposide, vincristine or vinblastine and derivatives thereof; alkaloids, such as ellipticine; polyols such as taxicin-I or taxicin-II; hormones, such as androgens (*e.g.*, dromostanolone or testolactone), progestins (*e.g.*, megestrol acetate or medroxyprogesterone

acetate), estrogens (*e.g.*, dimethylstilbestrol diphosphate, polyestradiol phosphate or estramustine phosphate) or antiestrogens (*e.g.*, tamoxifen); anthraquinones, such as mitoxantrone, ureas, such as hydroxyurea; hydrazines, such as procarbazine; or imidazoles, such as dacarbazine.

Chelated metals include chelates of di-or tripositive metals having a coordination number from 2 to 8 inclusive. Particular examples of such metals include technetium (Tc), rhenium (Re), cobalt (Co), copper (Cu), gold (Au), silver (Ag), lead (Pb), bismuth (Bi), indium (In), gallium (Ga), yttrium (Y), terbium (Tb), gadolinium (Gd), and scandium (Sc). In general the metal is preferably a radionuclide. Particular radionuclides include ^{99m}Tc, ^{186}Re, ^{188}Re, ^{58}Co, ^{60}Co, ^{67}Cu, ^{195}Au, ^{199}Au, ^{110}Ag, ^{203}Pb, ^{206}Bi, ^{207}Bi, ^{111}In, ^{67}Ga, ^{68}Ga, ^{88}Y, ^{90}Y, ^{160}Tb, ^{153}Gd, and ^{47}Sc.

The chelated metal may be for example one of the above types of metal chelated with any suitable polydentate chelating agent, for example acyclic or cyclic polyamines, polyethers, (*e.g.*, crown ethers and derivatives thereof); polyamides; porphyrins; and carbocyclic derivatives.In general, the type of chelating agent will depend on the metal in use. One particularly useful group of chelating agents in conjugates according to the invention, however, comprises acyclic and cyclic polyamines, especially polyaminocarboxylic acids, for example diethylenetriaminepentaacetic acid and derivatives thereof, and macrocyclic amines, such as cyclic tri-aza and tetra-aza derivatives (for example, as described in International Patent Specification No. WO 92/22583), and polyamides, especially desferrioxamine and derivatives thereof.

When a thiol group in the antibody is used as the point of attachment this may be achieved through reaction with a thiol reactive group present in the effector or reporter molecule. Examples of such groups include an á-halocarboxylic acid or ester, such as iodoacetamide, an imide, such as maleimide, a vinyl sulphone, or a disulphide. These and other suitable linking procedures are generally and more particularly described in International Patent Specifications Nos. WO 93/06231, WO 92/22583, WO 90/091195, and WO 89/01476.

ASSAYS FOR SELECTING MOLECULES THAT INCREASE BONE DENSITY

As discussed above, the present invention provides methods for selecting and/or isolating compounds that are capable of increasing bone density. For example, within one aspect of the present invention methods are provided for determining whether a selected molecule (*e.g.*, a candidate agent) is capable of increasing bone mineral content, comprising the steps of (a) mixing (or contacting) a selected molecule with TGF-beta binding protein and a selected member of the TGF-beta family of proteins, (b) determining whether the selected molecule

stimulates signaling by the TGF-beta family of proteins, or inhibits the binding of the TGF-beta binding protein to at least one member of the TGF-beta family of proteins. Within certain embodiments, the molecule enhances the ability of TGF-beta to function as a positive regulator of mesenchymal cell differentiation.

Within other aspects of the invention, methods are provided for determining whether a selected molecule (candidate agent) is capable of increasing bone mineral content, comprising the steps of (a) exposing (contacting, mixing, combining) a selected molecule to cells which express TGF-beta binding-protein and (b) determining whether the expression (or activity) of TGF-beta binding-protein in the exposed cells decreases, or whether an activity of the TGF-beta binding protein decreases, and therefrom determining whether the compound is capable of increasing bone mineral content. Within one embodiment, the cells are selected from the group consisting of the spontaneously transformed or untransformed normal human bone from bone biopsies and rat parietal bone osteoblasts. Methods for detecting the level of expression of a TGF-beta binding protein may be accomplished in a wide variety of assay formats known in the art and described herein. Immunoassays may be used for detecting and quantifying the expression of a TGF-beta binding protein and include, for example, Countercurrent Immuno-Electrophoresis (CIEP), radioimmunoassays, radioimmunoprecipitations, Enzyme-Linked Immuno-Sorbent Assays (ELISA), immunoblot assays such as dot blot assays and Western blots, inhibition or competition assays, and sandwich assays (*see* U.S. Patent Nos. 4,376,110 and 4,486,530; *see also Antibodies: A Laboratory Manual, supra*). Such immunoassays may use an antibody that is specific for a TGF-beta binding protein such as the anti-sclerostin antibodies described herein, or may use an antibody that is specific for a reporter molecule that is attached to the TGF-beta binding protein. The level of polypeptide expression may also be determined by quantifying the amount of TGF-beta binding protein that binds to a TGF-beta binding protein ligand. By way of example, binding of sclerostin in a sample to a BMP may be detected by surface plasmon resonance (SPR). Alternatively, the level of expression of mRNA encoding the specific TGF-beta binding protein may be quantified.

Representative embodiments of such assays are provided below in Examples 5 and 6. Briefly, a family member of the TGF-beta super-family or a TGF-beta binding protein is first bound to a solid phase, followed by addition of a candidate molecule. A labeled family member of the TGF-beta super-family or a TGF-beta binding protein is then added to the assay (*i.e.*, the labeled polypeptide is the ligand for whichever polypeptide was bound to the solid phase), the solid phase washed, and the quantity of bound or labeled TGF-beta super-family member or

TGF-beta binding protein on the solid support determined. Molecules which are suitable for use in increasing bone mineral content as described herein are those molecules which decrease the binding of TGF-beta binding protein to a member or members of the TGF-beta super-family in a statistically significant manner. Obviously, assays suitable for use within the present invention should not be limited to the embodiments described within Examples 2 and 3. In particular, numerous parameters may be altered, such as by binding TGF-beta to a solid phase, or by elimination of a solid phase entirely.

Within other aspects of the invention, methods are provided for determining whether a selected molecule is capable of increasing bone mineral content, comprising the steps of (a) exposing (contacting, mixing, combining) a selected molecule (candidate agent) to cells which express TGF-beta and (b) determining whether the activity of TGF-beta from said exposed cells is altered, and therefrom determining whether the compound is capable of increasing bone mineral content. Similar to the methods described herein, a wide variety of methods may be utilized to assess the changes of TGF-beta binding-protein expression due to a selected test compound. In one embodiment of the invention, the candidate agent is an antibody that binds to the TGF-beta binding protein sclerostin disclosed herein.

In a preferred embodiment of the invention, a method is provided for identifying an antibody that modulates a TGF-beta signaling pathway comprising contacting an antibody that specifically binds to a SOST polypeptide with a SOST peptide, including but not limited to the peptides disclosed herein, under conditions and for a time sufficient to permit formation of an antibody plus (+) SOST (antibody/SOST) complex and then detecting the level (*e.g.*, quantifying the amount) of the SOST/antibody complex to determine the presence of an antibody that modulates a TGF-beta signaling pathway. The method may be performed using SPR or any number of different immunoassays known in the art and disclosed herein, including an ELISA, immunoblot, or the like. A TGF-beta signaling pathway includes a signaling pathway by which a BMP binds to a type I and a type II receptor on a cell to stimulate or induce the pathway that modulates bone mineral content. In certain preferred embodiments of the invention, an antibody that specifically binds to SOST stimulates or enhances the pathway for increasing bone mineral content. Such an antibody may be identified using the methods disclosed herein to detect binding of an antibody to SOST specific peptides.

The subject invention methods may also be used for identifying antibodies that impair, inhibit (including competitively inhibit), or prevent binding of a BMP to a SOST polypeptide by detecting whether an antibody binds to SOST peptides that are located in regions or portions of

regions on SOST to which a BMP binds, such as peptides at the amino terminal end of SOST and peptides that include amino terminal amino acid residues and a portion of the core region (docking core) of SOST (*e.g.*, SEQ ID NOs:47-64, 66-73, and 92-95). The methods of the present invention may also be used to identify an antibody that impairs, prevents, or inhibits, formation of SOST homodimers. Such an antibody that binds specifically to SOST may be identified by detecting binding of the antibody to peptides that are derived from the core or the carboxy terminal region of SOST (*e.g.*, SEQ ID NOs: 74-91 and 96-99).

Within another embodiment of the present invention, methods are provided for determining whether a selected molecule is capable of increasing bone mineral content, comprising the steps of (a) mixing or contacting a selected molecule (candidate agent) with a TGF-beta-binding-protein and a selected member of the TGF-beta family of proteins, (b) determining whether the selected molecule up-regulates the signaling of the TGF-beta family of proteins, or inhibits the binding of the TGF-beta binding-protein to the TGF-beta family of proteins. Within certain embodiments, the molecule enhances the ability of TGF-beta to function as a positive regulator of mesenchymal cell differentiation.

Similar to the above described methods, a wide variety of methods may be utilized to assess stimulation of TGF-beta due to a selected test compound. One such representative method is provided below in Example 6 (see also Durham et al., *Endo. 136*:1374-1380.

Within yet other aspects of the present invention, methods are provided for determining whether a selected molecule (candidate agent) is capable of increasing bone mineral content, comprising the step of determining whether a selected molecule inhibits the binding of TGF-beta binding-protein to bone, or an analogue thereof. As utilized herein, it should be understood that bone or analogues thereof refers to hydroxyapatite, or a surface composed of a powdered form of bone, crushed bone or intact bone. Similar to the above described methods, a wide variety of methods may be utilized to assess the inhibition of TGF-beta binding-protein localization to bone matrix. One such representative method is provided below in Example 7 (*see also* Nicolas et al., *Calcif. Tissue Int.* 47:206-12 (1995)).

In one embodiment of the invention, an antibody or antigen-binding fragment thereof that specifically binds to a sclerostin polypeptide is capable of competitively inhibiting binding of a TGF-beta family member to the sclerostin polypeptide. The capability of the antibody or antibody fragment to impair or blocking binding of a TGF-beta family member, such as a BMP, to sclerostin may be determined according to any of the methods described herein. The antibody or fragment thereof that specifically binds to sclerostin may impair, block, or prevent binding of

a TGF-beta family member to sclerostin by impairing sclerostin homodimer formation. An antibody that specifically binds to sclerostin may also be used to identify an activity of sclerostin by inhibiting or impairing sclerostin from binding to a BMP. Alternatively, the antibody or fragment thereof may be incorporated in a cell-based assay or in an animal model in which sclerostin has a defined activity to determine whether the antibody alters (increases or decreases in a statistically significant manner) that activity. An antibody or fragment thereof that specifically binds to sclerostin may be used to examine the effect of such an antibody in a signal transduction pathway and thereby modulate (stimulate or inhibit) the signaling pathway. Preferably, binding of an antibody to SOST results in a stimulation or induction of a signaling pathway.

While the methods recited herein may refer to the analysis of an individual test molecule, that the present invention should not be so limited. In particular, the selected molecule may be contained within a mixture of compounds. Hence, the recited methods may further comprise the step of isolating a molecule that inhibits the binding of TGF-beta binding-protein to a TGF-beta family member.

CANDIDATE MOLECULES

A wide variety of molecules may be assayed for their ability to inhibit the binding of TGF-beta binding-protein to a TGF-beta family member. Representative examples discussed in more detail below include organic molecules (*e.g.*, organic small molecules), proteins or peptides, and nucleic acid molecules. Although it should be evident from the discussion below that the candidate molecules described herein may be utilized in the assays described herein, it should also be readily apparent that such molecules can also be utilized in a variety of diagnostic and therapeutic settins.

1. Organic Molecules

Numerous organic small molecules may be assayed for their ability to inhibit the binding of TGF-beta binding-protein to a TGF-beta family member. For example, within one embodiment of the invention suitable organic molecules may be selected from either a chemical library, wherein chemicals are assayed individually, or from combinatorial chemical libraries where multiple compounds are assayed at once, then deconvoluted to determine and isolate the most active compounds.

Representative examples of such combinatorial chemical libraries include those described by Agrafiotis et al., "System and method of automatically generating chemical compounds with desired properties," U.S. Patent No. 5,463,564; Armstrong, R.W., "Synthesis of

combinatorial arrays of organic compounds through the use of multiple component combinatorial array syntheses," WO 95/02566; Baldwin, J.J. et al., "Sulfonamide derivatives and their use," WO 95/24186; Baldwin, J.J. et al., "Combinatorial dihydrobenzopyran library," WO 95/30642; Brenner, S., "New kit for preparing combinatorial libraries," WO 95/16918; Chenera, B. et al., "Preparation of library of resin-bound aromatic carbocyclic compounds," WO 95/16712; Ellman, J.A., "Solid phase and combinatorial synthesis of benzodiazepine compounds on a solid support," U.S. Patent No. 5,288,514; Felder, E. et al., "Novel combinatorial compound libraries," WO 95/16209; Lerner, R. et al., "Encoded combinatorial chemical libraries," WO 93/20242; Pavia, M.R. et al., "A method for preparing and selecting pharmaceutically useful non-peptide compounds from a structurally diverse universal library," WO 95/04277; Summerton, J.E. and D.D. Weller, "Morpholino-subunit combinatorial library and method," U.S. Patent No. 5,506,337; Holmes, C., "Methods for the Solid Phase Synthesis of Thiazolidinones, Metathiazanones, and Derivatives thereof," WO 96/00148; Phillips, G.B. and G.P. Wei, "Solid-phase Synthesis of Benzimidazoles," *Tet. Letters 37*:4887-90, 1996; Ruhland, B. et al., "Solid-supported Combinatorial Synthesis of Structurally Diverse β-Lactams," *J. Amer. Chem. Soc. 111*:253-4, 1996; Look, G.C. et al., "The Indentification of Cyclooxygenase-1 Inhibitors from 4-Thiazolidinone Combinatorial Libraries," *Bioorg and Med. Chem. Letters 6*:707-12, 1996.

2. Proteins and Peptides

A wide range of proteins and peptides may likewise be utilized as candidate molecules for inhibitors of the binding of TGF-beta binding-protein to a TGF-beta family member.

a. Combinatorial Peptide Libraries

Peptide molecules which are putative inhibitors of the binding of TGF-beta binding-protein to a TGF-beta family member may be obtained through the screening of combinatorial peptide libraries. Such libraries may either be prepared by one of skill in the art (*see e.g.*, U.S. Patent Nos. 4,528,266 and 4,359,535, and Patent Cooperation Treaty Publication Nos. WO 92/15679, WO 92/15677, WO 90/07862, WO 90/02809, or purchased from commercially available sources (*e.g.*, New England Biolabs Ph.D.™ Phage Display Peptide Library Kit).

b. Antibodies

The present invention provides antibodies that specifically bind to a sclerostin polypeptide methods for using such antibodies. The present invention also provides sclerostin polypeptide immunogens that may be used for generation and analysis of these antibodies. The

antibodies may be useful to block or impair binding of a sclerostin polypeptide, which is a TGF-beta binding protein, to a ligand, particularly a bone morphogenic protein, and may also block or impair binding of the sclerostin polypeptide to one or more other ligands.

A molecule such as an antibody that inhibits the binding of the TGF-beta binding protein to one or more members of the TGF-beta family of proteins, including one or more bone morphogenic proteins (BMPs), should be understood to refer to, for example, a molecule that allows the activation of a TGF-beta family member or BMP, or allows binding of TGF-beta family members including one or more BMPs to their respective receptors by removing or preventing the TGF-beta member from binding to the TGF-binding-protein.

The present invention also provides peptide and polypeptide immunogens that may be used to generate and/or identify antibodies or fragments thereof that are capable of inhibiting, preventing, or impairing binding of the TGF-beta binding protein sclerostin to one or more BMPs. The present invention also provides peptide and polypeptide immunogens that may be used to generate and/or identify antibodies or fragments thereof that are capable of inhibiting, preventing, or impairing (*e.g.*, decreasing in a statistically significant manner) the formation of sclerostin homodimers. The antibodies of the present invention are useful for increasing the mineral content and mineral density of bone, thereby ameliorating numerous conditions that result in the loss of bone mineral content, including for example, disease, genetic predisposition, accidents that result in the lack of use of bone (*e.g.*, due to fracture), therapeutics that effect bone resorption or that kill bone forming cells, and normal aging.

Polypeptides or peptides useful for immunization and/or analysis of sclerostin-specific antibodies may also be selected by analyzing the primary, secondary, and tertiary structure of a TGF-beta binding protein according to methods known to those skilled in the art and described herein, in order to determine amino acid sequences more likely to generate an antigenic response in a host animal. *See, e.g.*, Novotny, *Mol. Immunol.* 28:201-207 (1991); Berzofsky, *Science* 229:932-40 (1985)). Modeling and x-ray crystallography data may also be used to predict and/or identify which portions or regions of a TGF-beta binding protein interact with which portions of a TGF-beta binding protein ligand, such as a BMP. TGF-beta binding protein peptide immunogens may be designed and prepared that include amino acid sequences within or surrounding the portions or regions of interaction. These antibodies may be useful to block or impair binding of the TGF-beta binding protein to the same ligand and may also block or impair binding of the TGF-beta binding protein to one or more other ligands.

Antibodies or antigen binding fragments thereof contemplated by the present invention

include antibodies that are capable of specifically binding to sclerostin and competitively inhibiting binding of a TGF-beta polypeptide, such as a BMP, to sclerostin. For example, the antibodies contemplated by the present invention competitively inhibit binding of the sclerostin polypeptide to the BMP Type I receptor site on a BMP, or to the BMP Type II receptor binding site, or may competitively inhibit binding of sclerostin to both the Type I and Type II receptor binding sites on a BMP. Without wishing to be bound by theory, when an anti-sclerostin antibody competitively inhibits binding of the Type I and/or Type II binding sites of the BMP polypeptide to sclerostin, thus blocking the antagonistic activity of sclerostin, the receptor binding sites on BMP are available to bind to the Type I and Type II receptors, thereby increasing bone mineralization. The binding interaction between a TGF-beta binding protein such as sclerostin and a TGF-beta polypeptide such as a BMP generally occurs when each of the ligand pairs forms a homodimer. Therefore instead of or in addition to using an antibody specific for sclerostin to block, impair, or prevent binding of sclerostin to a BMP by competitively inhibiting binding of sclerostin to BMP, a sclerostin specific antibody may be used to block or impair sclerostin homodimer formation.

By way of example, one dimer of human Noggin, which is a BMP antagonist that has the ability to bind a BMP with high affinity (Zimmerman et al., *supra*), was isolated in complex with one dimer of human BMP-7 and analyzed by multiwavelength anomalous diffraction (MAD) (Groppe et al., *Nature* 420:636-42 (2002)). As discussed herein, this study revealed that Noggin dimer may efficiently block all the receptor binding sites (two type I and two type II receptor binding sites) on a BMP dimer. The location of the amino acids of Noggin that contact BMP-7 may be useful in modeling the interaction between other TGF-beta binding proteins, such as sclerostin (SOST), and BMPs, and thus aiding the design of peptides that may be used as immunogens to generate antibodies that block or impair such an interaction.

In one embodiment of the present invention, an antibody, or an antigen-binding fragment thereof, that binds specifically to a SOST polypeptide competitively inhibits binding of the SOST polypeptide to at least one or both of a bone morphogenic protein (BMP) Type I Receptor binding site and a BMP Type II Receptor binding site that are located on a BMP. The epitopes on SOST to which these antibodies bind may include or be included within contiguous amino acid sequences that are located at the N-terminus of the SOST polypeptide (amino acids at about positions1-56 of SEQ ID NO:46). The polypeptides may also include a short linker peptide sequence that connects the N-terminal region to the core region, for example, polypeptides as provided in SEQ ID NO:92 (human) and SEQ ID NO:93 (rat). Shorter representative N-

terminus peptide sequences of human SOST (*e.g.*, SEQ ID NO:46) include SEQ ID NOS:47-51 , and representative rat SOST (*e.g.*, SEQ ID NO:65) peptide sequences include SEQ ID NOS:57-60.

Antibodies that specifically bind to a SOST polypeptide and block or competitively inhibit binding of the SOST polypeptide to a BMP, for example, by blocking or inhibiting binding to amino acids of a BMP corresponding to one or more of the Type I and Type II receptor binding sites may also specifically bind to peptides that comprise an amino acid sequence corresponding to the core region of SOST (amino acids at about positions 57-146 of SEQ ID NO:46). Polypeptides that include the core region may also include additional amino acids extending at either or both the N-terminus and C-terminus, for example, to include cysteine residues that may be useful for conjugating the polypeptide to a carrier molecule. Representative core polypeptides of human and rat SOST, for example, comprise the amino acid sequences set forth in SEQ ID NO:94 and SEQ ID NO:95 , respectively. Such antibodies may also bind shorter polypeptide sequences. Representative human SOST core peptide sequences are provided in SEQ ID NOs:66-69 and representative rat SOST core sequences are provided in SEQ ID NOs:70-73.

In another embodiment, antibodies that specifically bind to a SOST polypeptide impair (inhibit, prevent, or block, *e.g.*, decrease in a statistically significant manner) formation of a SOST homodimer. Because the interaction between SOST and a BMP may involve a homodimer of SOST and a homodimer of the BMP, an antibody that prevents or impairs homodimer formation of SOST may thereby alter bone mineral density, preferably increasing bone mineral density. In one embodiment, antibodies that bind to the core region of SOST prevent homodimer formation. Such antibodies may also bind to peptides that comprise contiguous amino acid sequences corresponding the core region, for example, SEQ ID NOs: 74, 75, and 98 (human SOST) and SEQ ID NOs:76 and 99 (rat SOST). Antibodies that bind to an epitope located on the C-terminal region of a SOST polypeptide (at about amino acid positions 147-190 of either SEQ ID NO:46 or 65) may also impair homodimer formation. Representative C-terminal polypeptides of human and rat SOST, for example, comprise the amino acid sequences set forth in SEQ ID NO:96 and SEQ ID NO:97, respectively. Such antibodies may also bind shorter polypeptide sequences. Representative human SOST C-terminal peptide sequences are provided in SEQ ID NOs:78-81 and representative rat SOST C-terminal sequences are provided in SEQ ID NOs:86-88.

The SOST polypeptides and peptides disclosed herein to which antibodies may

specifically bind are useful as immunogens. These immunogens of the present invention may be used for immunizing an animal to generate a humoral immune response that results in production of antibodies that specifically bind to a Type I or Type II receptor binding site or both located on a BMP include peptides derived from the N-terminal region of SOST or that may prevent SOST homodimer formation.

Such SOST polypeptides and peptides that are useful as immunogens may also be used in methods for screening samples containing antibodies, for example, samples of purified antibodies, antisera, or cell culture supernatants or any other biological sample that may contain one or more antibodies specific for SOST. These peptides may also be used in methods for identifying and selecting from a biological sample one or more B cells that are producing an antibody that specifically binds to SOST (*e.g.*, plaque forming assays and the like). The B cells may then be used as source of a SOST specific antibody-encoding polynucleotide that can be cloned and/or modified by recombinant molecular biology techniques known in the art and described herein.

A "biological sample" as used herein refers in certain embodiments to a sample containing at least one antibody specific for a SOST polypeptide, and a biological sample may be provided by obtaining a blood sample, biopsy specimen, tissue explant, organ culture, or any other tissue or cell preparation from a subject or a biological source. A sample may further refer to a tissue or cell preparation in which the morphological integrity or physical state has been disrupted, for example, by dissection, dissociation, solubilization, fractionation, homogenization, biochemical or chemical extraction, pulverization, lyophilization, sonication, or any other means for processing a sample derived from a subject or biological source. The subject or biological source may be a human or non-human animal, a primary cell culture (*e.g.*, B cells immunized in vitro), or culture adapted cell line including but not limited to genetically engineered cell lines that may contain chromosomally integrated or episomal recombinant nucleic acid sequences, immortalized or immortalizable cell lines, somatic cell hybrid cell lines, differentiated or differentiatable cell lines, transformed cell lines, and the like.

SOST peptide immunogens may also be prepared by synthesizing a series of peptides that, in total, represent the entire polypeptide sequence of a SOST polypeptide and that each have a portion of the SOST amino acid sequence in common with another peptide in the series. This overlapping portion would preferably be at least four amino acids, and more preferably 5, 6, 7, 8, 9, or 10 amino acids. Each peptide may be used to immunize an animal, the sera collected from the animal, and tested in an assay to identify which animal is producing antibodies that

impair or block binding of SOST to a TGF-beta protein. Antibodies are then prepared from such identified immunized animals according to methods known in the art and described herein.

Antibodies which inhibit the binding of TGF-beta binding-protein to a TGF-beta family member may readily be prepared given the disclosure provided herein. Particularly useful are anti-TGF-beta binding-protein antibodies that "specifically bind" TGF-beta binding-protein of SEQ ID NOs: 2, 6, 8, 10, 12, 14, 16, 46, or 65, but not to other TGF-beta binding-proteins such as Dan, Cerberus, SCGF, or Gremlin. Within the context of the present invention, antibodies are understood to include monoclonal antibodies, polyclonal antibodies, single chain, chimeric, CDR-grafted immunoglobulings, anti-idiotypic antibodies, and antibody fragments thereof (*e.g.*, Fab, Fd, Fab', and $F(ab')_2$, F_V variable regions, or complementarity determining regions). As discussed above, antibodies are understood to be specific against TGF-beta binding-protein, or against a specific TGF-beta family member, if they bind with a K_a of greater than or equal to 10^7 M^{-1}, preferably greater than or equal to 10^8 M^{-1}, and do not bind to other TGF-beta binding-proteins, or bind with a K_a of less than or equal to 10^6 M^{-1}. Affinity of an antibody for its cognate antigen is also commonly expressed as a dissociation constant K_D, and an anti-SOST antibody specifically binds to a TGF-beta family member if it binds with a K_D of less than or equal to about 10^{-5} M, more preferably less than or equal to about 10^{-6} M, still more preferably less than or equal to 10^{-7} M, and still more preferably less than or equal to 10^{-8} M. Furthermore, antibodies of the present invention preferably block, impair, or inhibit (*e.g.*, decrease with statistical significance) the binding of TGF-beta binding-protein to a TGF-beta family member. The affinity of a monoclonal antibody or binding partner, as well as inhibition of binding can be readily determined by one of ordinary skill in the art (*see* Scatchard, *Ann. N.Y. Acad. Sci. 51*:660-672, 1949). Affinity may also be determined by surface plasmon resonance (SPR; BIAcore, Biosensor, Piscataway, NJ). For surface plasmon resonance, target molecules are immobilized on a solid phase and exposed to ligands in a mobile phase running along a flow cell. If ligand binding to the immobilized target occurs, the local refractive index changes, leading to a change in SPR angle, which can be monitored in real time by detecting changes in the intensity of the reflected light. The rates of change of the SPR signal can be analyzed to yield apparent rate constants for the association and dissociation phases of the binding reaction. The ratio of these values gives the apparent equilibrium constant (affinity) (*see, e.g.*, Wolff et al., *Cancer Res.* 53:2560-65 (1993)).

An antibody according to the present invention may belong to any immunoglobulin class, for example IgG, IgE, IgM, IgD, or IgA, and may be any one of the different isotypes that may

comprise a class (such as IgG1, IgG2, IgG3, and IgG4 of the human IgG class). It may be obtained from or derived from an animal, for example, fowl (*e.g.,* chicken) and mammals, which includes but is not limited to a mouse, rat, hamster, rabbit, or other rodent, a cow, horse, sheep, goat, camel, human, or other primate. The antibody may be an internalising antibody.

Methods well known in the art may be used to generate antibodies, polyclonal antisera, or monoclonal antibodies that are specific for a TGF-beta binding protein such as SOST. Antibodies also may be produced as genetically engineered immunoglobulins (Ig) or Ig fragments designed to have desirable properties. For example, by way of illustration and not limitation, antibodies may include a recombinant IgG that is a chimeric fusion protein having at least one variable (V) region domain from a first mammalian species and at least one constant region domain from a second, distinct mammalian species. Most commonly, a chimeric antibody has murine variable region sequences and human constant region sequences. Such a murine/human chimeric immunoglobulin may be "humanized" by grafting the complementarity determining regions (CDRs) derived from a murine antibody, which confer binding specificity for an antigen, into human-derived V region framework regions and human-derived constant regions. Fragments of these molecules may be generated by proteolytic digestion, or optionally, by proteolytic digestion followed by mild reduction of disulfide bonds and alkylation. Alternatively, such fragments may also be generated by recombinant genetic engineering techniques.

Certain preferred antibodies are those antibodies that inhibit or block a TGF-beta binding protein activity within an *in vitro* assay, as described herein. Binding properties of an antibody to a TGF-beta binding protein may generally be assessed using immunodetection methods including, for example, an enzyme-linked immunosorbent assay (ELISA), immunoprecipitation, immunoblotting, countercurrent immunoelectrophoresis, radioimmunoassays, dot blot assays, inhibition or competition assays, and the like, which may be readily performed by those having ordinary skill in the art (*see, e.g.,* U.S. Patent Nos. 4,376,110 and 4,486,530; Harlow et al., *Antibodies: A Laboratory Manual,* Cold Spring Harbor Laboratory (1988)).

An immunogen may be comprised of cells expressing a TGF-beta binding protein, purified or partially purified TGF-beta binding polypeptides, or variants or fragments (*i.e.,* peptides) thereof, or peptides derived from a TGF-beta binding protein. Such peptides may be generated by proteolytic cleavage of a larger polypeptide, by recombinant molecular methodologies, or may be chemically synthesized. For instance, nucleic acid sequences encoding TGF-beta binding proteins are provided herein, such that those skilled in the art may

routinely prepare TGF-beta binding proteins for use as immunogens. Peptides may be chemically synthesized by methods as described herein and known in the art. Alternatively, peptides may be generated by proteolytic cleavage of a TGF-beta binding protein, and individual peptides isolated by methods known in the art such as polyacrylamide gel electrophoresis or any number of liquid chromatography or other separation methods. Peptides useful as immunogens typically may have an amino acid sequence of at least 4 or 5 consecutive amino acids from a TGF-beta binding protein amino acid sequence such as those described herein, and preferably have at least 6, 7, 8, 9, 10, 11, 12, 14, 15, 16, 18, 19 or 20 consecutive amino acids of a TGF-beta binding protein. Certain other preferred peptide immunogens comprise at least 6 but no more than 12 or more consecutive amino acids of a TGF-beta binding protein sequence, and other preferred peptide immunogens comprise at least 21 but no more than 50 consecutive amino acids of a SOST polypeptide. Other preferred peptide immunogens comprise 21-25, 26-30, 31-35, 36-40, 41-50, or any whole integer number of amino acids between and including 21 and 100 consecutive amino acids, and between 100 and 190 consecutive amino acids of a TGF-beta binding protein sequence.

As disclosed herein, polyclonal antibodies may be readily generated by one of ordinary skill in the art from a variety of warm-blooded animals such as horses, cows, various fowl, rabbits, mice, sheep, goats, baboons, or rats. Typically, the TGF-beta binding-protein or unique peptide thereof of 13-20 amino acids or as described herein (preferably conjugated to keyhole limpet hemocyanin by cross-linking with glutaraldehyde) is used to immunize the animal through intraperitoneal, intramuscular, intraocular, intradermal, or subcutaneous injections, along with an adjuvant such as Freund's complete or incomplete adjuvant, or the Ribi Adjuvant System (Corixa Corporation, Seattle, WA). *See also, e.g.*, Harlow et al., *supra*. In general, after the first injection, animals receive one or more booster immunizations according to a preferred schedule that may vary according to, *inter alia*, the antigen, the adjuvant (if any), and/or the particular animal species. The immune response may be monitored by periodically bleeding the animal and preparing and analyzing sera in an immunoassay, such as an ELISA or Ouchterlony diffusion assay, or the like, to determine the specific antibody titer. Particularly preferred polyclonal antisera will give a detectable signal on one of these assays, such as an ELISA, that is preferably at least three times greater than background. Once the titer of the animal has reached a plateau in terms of its reactivity to the protein, larger quantities of antisera may be readily obtained either by weekly bleedings, or by exsanguinating the animal.

Polyclonal antibodies that bind specifically to the TGF-beta binding protein or peptide

may then be purified from such antisera, for example, by affinity chromatography using protein A. Alternatively, affinity chromatography may be performed wherein the TGF-beta binding protein or peptide or an antibody specific for an Ig constant region of the particular immunized animal species is immobilized on a suitable solid support.

Antibodies for use in the invention include monoclonal antibodies that are prepared by conventional immunization and cell fusion procedures as described herein an known in the art. Monoclonal antibodies may be readily generated using conventional techniques (*see, e.g.,* Kohler et al., *Nature* 256:495, 1975; Coligan et al. (eds.), *Current Protocols in Immunology*, 1:2.5.1-2.6.7 (John Wiley & Sons 1991) ["Coligan"]; U.S. Patent Nos. RE 32,011, 4,902,614, 4,543,439, and 4,411,993 which are incorporated herein by reference; *see also Monoclonal Antibodies, Hybridomas: A New Dimension in Biological Analyses*, Plenum Press, Kennett, McKearn, and Bechtol (eds.), 1980, and *Antibodies: A Laboratory Manual*, Harlow and Lane (eds.), Cold Spring Harbor Laboratory Press, 1988, which are also incorporated herein by reference; Picksley et al., "Production of monoclonal antibodies against proteins expressed in *E. coli*," in *DNA Cloning 2: Expression Systems, 2nd Edition*, Glover et al. (eds.), page 93 (Oxford University Press 1995)). Antibody fragments may be derived therefrom using any suitable standard technique such as proteolytic digestion, or optionally, by proteolytic digestion (for example, using papain or pepsin) followed by mild reduction of disulfide bonds and alkylation. Alternatively, such fragments may also be generated by recombinant genetic engineering techniques.

Briefly, within one embodiment a subject animal such as a rat or mouse or hamster is immunized with TGF-beta binding-protein or a portion of a region thereof, including peptides within a region, as described herein. The protein may be admixed with an adjuvant such as Freund's complete or incomplete adjuvant or Ribi adjuvant in order to increase the resultant immune response. Between one and three weeks after the initial immunization the animal may be reimmunized with another booster immunization, and tested for reactivity to the protein using assays described herein. Once the animal has reached a plateau in its reactivity to the injected protein, it is sacrificed, and organs which contain large numbers of B cells such as the spleen and lymph nodes are harvested. The harvested spleen and/or lymph node cell suspensions are fused with a suitable myeloma cell that is drug-sensitized in order to create a "hybridoma" which secretes monoclonal antibody. Suitable myeloma lines include, for example, NS-0, SP20, NS-1 (ATCC No. TIB 18), and P3X63 - Ag 8.653 (ATCC No. CRL 1580).

The lymphoid (*e.g.*, spleen) cells and the myeloma cells may be combined for a few

minutes with a membrane fusion-promoting agent, such as polyethylene glycol or a nonionic detergent, and then plated at low density on a selective medium that supports the growth of hybridoma cells but not unfused myeloma cells. Following the fusion, the cells may be placed into culture plates containing a suitable medium, such as RPMI 1640, or DMEM (Dulbecco's Modified Eagles Medium) (JRH Biosciences, Lenexa, Kansas), as well as additional ingredients, such as fetal bovine serum (FBS, *i.e.*, from Hyclone, Logan, Utah, or JRH Biosciences). Additionally, the medium should contain a reagent which selectively allows for the growth of fused spleen and myeloma cells such as HAT (hypoxanthine, aminopterin, and thymidine) (Sigma Chemical Co., St. Louis, Missouri). After about seven days, the resulting fused cells or hybridomas may be screened in order to determine the presence of antibodies which are reactive with TGF-beta binding-protein (depending on the antigen used), and which block, impair, or inhibit the binding of TGF-beta binding-protein to a TGF-beta family member. Hybridomas that produce monoclonal antibodies that specifically bind to sclerostin or a variant thereof are preferred.

A wide variety of assays may be utilized to determine the presence of antibodies which are reactive against the proteins of the present invention, including for example countercurrent immuno-electrophoresis, radioimmunoassays, radioimmunoprecipitations, enzyme-linked immuno-sorbent assays (ELISA), dot blot assays, western blots, immunoprecipitation, inhibition or competition assays, and sandwich assays (*see* U.S. Patent Nos. 4,376,110 and 4,486,530; *see also Antibodies: A Laboratory Manual*, Harlow and Lane (eds.), Cold Spring Harbor Laboratory Press, 1988). The hybridomas are cloned, for example, by limited dilution cloning or by soft agar plaque isolation, and reassayed. Thus, a hybridoma producing antibodies reactive against the desired protein may be isolated.

The monoclonal antibodies from the hybridoma cultures may be isolated from the supernatants of hybridoma cultures. An alternative method for production of a murine monoclonal antibody is to inject the hybridoma cells into the peritoneal cavity of a syngeneic mouse, for example, a mouse that has been treated (*e.g.*, pristane-primed) to promote formation of ascites fluid containing the monoclonal antibody. Monoclonal antibodies can be isolated and purified by a variety of well-established techniques. Such isolation techniques include affinity chromatography with Protein-A Sepharose, size-exclusion chromatography, and ion-exchange chromatography (*see*, for example, Coligan at pages 2.7.1-2.7.12 and pages 2.9.1-2.9.3; Baines et al., "Purification of Immunoglobulin G (IgG)," in *Methods in Molecular Biology, Vol. 10,* pages 79-104 (The Humana Press, Inc. 1992)). Monoclonal antibodies may be purified by

affinity chromatography using an appropriate ligand selected based on particular properties of the antibody (*e.g.*, heavy or light chain isotype, binding specificity, etc.). Examples of a suitable ligand, immobilized on a solid support, include Protein A, Protein G, an anti-constant region (light chain or heavy chain) antibody, an anti-idiotype antibody, and a TGF-beta binding protein, or fragment or variant thereof.

In addition, an anti-TGF-beta binding-protein antibody of the present invention may be a human monoclonal antibody. Human monoclonal antibodies may be generated by any number of techniques with which those having ordinary skill in the art will be familiar. Such methods include, but are not limited to, Epstein Barr Virus (EBV) transformation of human peripheral blood cells (*e.g.*, containing B lymphocytes), *in vitro* immunization of human B cells, fusion of spleen cells from immunized transgenic mice carrying inserted human immunoglobulin genes, isolation from human immunoglobulin V region phage libraries, or other procedures as known in the art and based on the disclosure herein. For example, human monoclonal antibodies may be obtained from transgenic mice that have been engineered to produce specific human antibodies in response to antigenic challenge. Methods for obtaining human antibodies from transgenic mice are described, for example, by Green et al., *Nature Genet.* 7:13, 1994; Lonberg et al., *Nature* 368:856, 1994; Taylor et al., *Int. Immun.* 6:579, 1994; .S. Patent No. 5,877,397; Bruggemann et al., 1997 *Curr. Opin. Biotechnol. 8*:455-58; Jakobovits et al., 1995 *Ann. N. Y. Acad. Sci. 764*:525-35. In this technique, elements of the human heavy and light chain locus are introduced into strains of mice derived from embryonic stem cell lines that contain targeted disruptions of the endogenous heavy chain and light chain loci. (*See also* Bruggemann et al., *Curr. Opin. Biotechnol.* 8:455-58 (1997)). For example, human immunoglobulin transgenes may be mini-gene constructs, or transloci on yeast artificial chromosomes, which undergo B cell-specific DNA rearrangement and hypermutation in the mouse lymphoid tissue. Human monoclonal antibodies may be obtained by immunizing the transgenic mice, which may then produce human antibodies specific for the antigen. Lymphoid cells of the immunized transgenic mice can be used to produce human antibody-secreting hybridomas according to the methods described herein. Polyclonal sera containing human antibodies may also be obtained from the blood of the immunized animals.

Another method for generating human TGF-beta binding protein specific monoclonal antibodies includes immortalizing human peripheral blood cells by EBV transformation. *See, e.g.*, U.S. Patent No. 4,464,456. Such an immortalized B cell line (or lymphoblastoid cell line) producing a monoclonal antibody that specifically binds to a TGF-beta binding protein (or a variant or fragment thereof) can be identified by immunodetection methods as provided herein,

for example, an ELISA, and then isolated by standard cloning techniques. The stability of the lymphoblastoid cell line producing an anti-TGF-beta binding protein antibody may be improved by fusing the transformed cell line with a murine myeloma to produce a mouse-human hybrid cell line according to methods known in the art (*see, e.g.*, Glasky et al., *Hybridoma* 8:377-89 (1989)). Still another method to generate human monoclonal antibodies is *in vitro* immunization, which includes priming human splenic B cells with antigen, followed by fusion of primed B cells with a heterohybrid fusion partner. *See, e.g.*, Boerner et al., 1991 *J. Immunol.* *147*:86-95.

In certain embodiments, a B cell that is producing an anti-SOST antibody is selected and the light chain and heavy chain variable regions are cloned from the B cell according to molecular biology techniques known in the art (WO 92/02551; US patent 5,627,052; Babcook et al., *Proc. Natl. Acad. Sci. USA* 93:7843-48 (1996)) and described herein. Preferably B cells from an immunized animal are isolated from the spleen, lymph node, or peripheral blood sample by selecting a cell that is producing an antibody that specifically binds to SOST. B cells may also be isolated from humans, for example, from a peripheral blood sample. Methods for detecting single B cells that are producing an antibody with the desired specificity are well known in the art, for example, by plaque formation, fluorescence-activated cell sorting, in vitro stimulation followed by detection of specific antibody, and the like. Methods for selection of specific antibody producing B cells include, for example, preparing a single cell suspension of B cells in soft agar that contains SOST or a peptide fragment thereof. Binding of the specific antibody produced by the B cell to the antigen results in the formation of a complex, which may be visible as an immunoprecipitate. After the B cells producing the specific antibody are selected, the specific antibody genes may be cloned by isolating and amplifying DNA or mRNA according to methods known in the art and described herein.

For particular uses, fragments of anti-TGF-beta binding protein antibodies may be desired. Antibody fragments, $F(ab')_2$, Fab, Fab', Fv, Fc, Fd, retain the antigen binding site of the whole antibody and therefore bind to the same epitope. These antigen-binding fragments derived from an antibody can be obtained, for example, by proteolytic hydrolysis of the antibody, for example, pepsin or papain digestion of whole antibodies according to conventional methods. As an illustration, antibody fragments can be produced by enzymatic cleavage of antibodies with pepsin to provide a 5S fragment denoted $F(ab')_2$. This fragment can be further cleaved using a thiol reducing agent to produce 3.5S Fab' monovalent fragments. Optionally, the cleavage reaction can be performed using a blocking group for the sulfhydryl groups that result from

cleavage of disulfide linkages. As an alternative, an enzymatic cleavage using papain produces two monovalent Fab fragments and an Fc fragment directly. These methods are described, for example, by Goldenberg, U.S. patent No. 4,331,647, Nisonoff et al., *Arch. Biochem. Biophys.* *89*:230, 1960; Porter, *Biochem. J.* *73*:119, 1959; Edelman et al., in *Methods in Enzymology* *1*:422 (Academic Press 1967); and by Coligan at pages 2.8.1-2.8.10 and 2.10.-2.10.4. Other methods for cleaving antibodies, such as separating heavy chains to form monovalent light-heavy chain fragments (Fd), further cleaving of fragments, or other enzymatic, chemical, or genetic techniques may also be used, so long as the fragments bind to the antigen that is recognized by the intact antibody.

An antibody fragment may also be any synthetic or genetically engineered protein that acts like an antibody in that it binds to a specific antigen to form a complex. For example, antibody fragments include isolated fragments consisting of the light chain variable region, "Fv" fragments consisting of the variable regions of the heavy and light chains, recombinant single chain polypeptide molecules in which light and heavy variable regions are connected by a peptide linker (scFv proteins), and minimal recognition units consisting of the amino acid residues that mimic the hypervariable region. The antibody of the present invention preferably comprises at least one variable region domain. The variable region domain may be of any size or amino acid composition and will generally comprise at least one hypervariable amino acid sequence responsible for antigen binding and which is adjacent to or in frame with one or more framework sequences. In general terms, the variable (V) region domain may be any suitable arrangement of immunoglobulin heavy (V_H) and/or light (V_L) chain variable domains. Thus, for example, the V region domain may be monomeric and be a V_H or V_L domain, which is capable of independently binding antigen with acceptable affinity. Alternatively, the V region domain may be dimeric and contain V_H-V_H, V_H-V_L, or V_L-V_L, dimers. Preferably, the V region dimer comprises at least one V_H and at least one V_L chain that are non-covalently associated (hereinafter referred to as F_v). If desired, the chains may be covalently coupled either directly, for example via a disulphide bond between the two variable domains, or through a linker, for example a peptide linker, to form a single chain Fv (scF_v).

The variable region domain may be any naturally occurring variable domain or an engineered version thereof. By engineered version is meant a variable region domain that has been created using recombinant DNA engineering techniques. Such engineered versions include those created, for example, from a specific antibody variable region by insertions, deletions, or changes in or to the amino acid sequences of the specific antibody. Particular examples include engineered

variable region domains containing at least one CDR and optionally one or more framework amino acids from a first antibody and the remainder of the variable region domain from a second antibody.

The variable region domain may be covalently attached at a C-terminal amino acid to at least one other antibody domain or a fragment thereof. Thus, for example, a V_H domain that is present in the variable region domain may be linked to an immunoglobulin C_H1 domain, or a fragment thereof. Similarly a V_L domain may be linked to a C_K domain or a fragment thereof. In this way, for example, the antibody may be a Fab fragment wherein the antigen binding domain contains associated V_H and V_L domains covalently linked at their C-termini to a CH1 and C_K domain, respectively. The CH1 domain may be extended with further amino acids, for example to provide a hinge region or a portion of a hinge region domain as found in a Fab' fragment, or to provide further domains, such as antibody CH2 and CH3 domains.

Another form of an antibody fragment is a peptide comprising for a single complementarity-determining region (CDR). CDR peptides ("minimal recognition units") can be obtained by constructing polynucleotides that encode the CDR of an antibody of interest. Such polynucleotides are prepared, for example, by using the polymerase chain reaction to synthesize the variable region using mRNA of antibody-producing cells as a template (*see*, for example, Larrick et al., *Methods: A Companion to Methods in Enzymology* 2:106, 1991; Courtenay-Luck, "Genetic Manipulation of Monoclonal Antibodies," in *Monoclonal Antibodies: Production, Engineering and Clinical Application*, Ritter et al. (eds.), page 166 (Cambridge University Press 1995); and Ward et al., "Genetic Manipulation and Expression of Antibodies," in *Monoclonal Antibodies: Principles and Applications*, Birch et al., (eds.), page 137 (Wiley-Liss, Inc. 1995)).

Alternatively, the antibody may be a recombinant or engineered antibody obtained by the use of recombinant DNA techniques involving the manipulation and re-expression of DNA encoding antibody variable and/or constant regions. Such DNA is known and/or is readily available from DNA libraries including for example phage-antibody libraries (*see* Chiswell and McCafferty, *Tibtech*. 10:80-84 (1992)) or if desired can be synthesized. Standard molecular biology and/or chemistry procedures may be used to sequence and manipulate the DNA, for example, to introduce codons to create cysteine residues, or to modify, add or delete other amino acids or domains as desired.

Chimeric antibodies, specific for a TGF-beta binding protein, and which include humanized antibodies, may also be generated according to the present invention. A chimeric

antibody has at least one constant region domain derived from a first mammalian species and at least one variable region domain derived from a second, distinct mammalian species (*see, e.g.*, Morrison et al., *Proc. Natl. Acad. Sci. USA*, 81:6851-55 (1984)). In preferred embodiments, a chimeric antibody may be constructed by cloning the polynucleotide sequence that encodes at least one variable region domain derived from a non-human monoclonal antibody, such as the variable region derived from a murine, rat, or hamster monoclonal antibody, into a vector containing a nucleotide sequence that encodes at least one human constant region (*see, e.g.*, Shin et al., *Methods Enzymol.* 178:459-76 (1989); Walls et al., *Nucleic Acids Res.* 21:2921-29 (1993)). By way of example, the polynucleotide sequence encoding the light chain variable region of a murine monoclonal antibody may be inserted into a vector containing a nucleotide sequence encoding the human kappa light chain constant region sequence. In a separate vector, the polynucleotide sequence encoding the heavy chain variable region of the monoclonal antibody may be cloned in frame with sequences encoding a human IgG constant region, for example, the human IgG1 constant region. The particular human constant region selected may depend upon the effector functions desired for the particular antibody (*e.g.*, complement fixing, binding to a particular Fc receptor, etc.). Preferably, the constructed vectors will be transfected into eukaryotic cells for stable expression of the chimeric antibody. Another method known in the art for generating chimeric antibodies is homologous recombination (*e.g.*, U.S. Patent No. 5,482,856).

A non-human/human chimeric antibody may be further genetically engineered to create a "humanized" antibody. Such a humanized antibody may comprise a plurality of CDRs derived from an immunoglobulin of a non-human mammalian species, at least one human variable framework region, and at least one human immunoglobulin constant region. Useful strategies for designing humanized antibodies may include, for example by way of illustration and not limitation, identification of human variable framework regions that are most homologous to the non-human framework regions of the chimeric antibody. Without wishing to be bound by theory, such a strategy may increase the likelihood that the humanized antibody will retain specific binding affinity for a TGF-beta binding protein, which in some preferred embodiments may be substantially the same affinity for a TGF-beta binding protein or variant or fragment thereof, and in certain other preferred embodiments may be a greater affinity for TGF-beta binding protein. *See, e.g.*, Jones et al., 1986 *Nature* 321:522-25; Riechmann et al., 1988 *Nature* 332:323-27. Designing such a humanized antibody may therefore include determining CDR loop conformations and structural determinants of the non-human variable regions, for example,

by computer modeling, and then comparing the CDR loops and determinants to known human CDR loop structures and determinants. *See, e.g.*, Padlan et al., 1995 *FASEB* 9:133-39; Chothia et al., 1989 *Nature*, 342:377-383. Computer modeling may also be used to compare human structural templates selected by sequence homology with the non-human variable regions. *See, e.g.*, Bajorath et al., 1995 *Ther. Immunol.* 2:95-103; EP-0578515-A3. If humanization of the non-human CDRs results in a decrease in binding affinity, computer modeling may aid in identifying specific amino acid residues that could be changed by site-directed or other mutagenesis techniques to partially, completely or supra-optimally (*i.e.*, increase to a level greater than that of the non-humanized antibody) restore affinity. Those having ordinary skill in the art are familiar with these techniques, and will readily appreciate numerous variations and modifications to such design strategies.

One such method for preparing a humanized antibody is called veneering. As used herein, the terms "veneered FRs" and "recombinantly veneered FRs" refer to the selective replacement of FR residues from, *e.g.*, a rodent heavy or light chain V region, with human FR residues in order to provide a xenogeneic molecule comprising an antigen-binding site that retains substantially all of the native FR polypeptide folding structure. Veneering techniques are based on the understanding that the ligand binding characteristics of an antigen-binding site are determined primarily by the structure and relative disposition of the heavy and light chain CDR sets within the antigen-binding surface. Davies et al., *Ann. Rev. Biochem.* 59:439-73, 1990. Thus, antigen binding specificity can be preserved in a humanized antibody only wherein the CDR structures, their interaction with each other, and their interaction with the rest of the V region domains are carefully maintained. By using veneering techniques, exterior (*e.g.*, solvent-accessible) FR residues that are readily encountered by the immune system are selectively replaced with human residues to provide a hybrid molecule that comprises either a weakly immunogenic, or substantially non-immunogenic veneered surface.

The process of veneering makes use of the available sequence data for human antibody variable domains compiled by Kabat et al., in *Sequences of Proteins of Immunological Interest*, 4th ed., (U.S. Dept. of Health and Human Services, U.S. Government Printing Office, 1987), updates to the Kabat database, and other accessible U.S. and foreign databases (both nucleic acid and protein). Solvent accessibilities of V region amino acids can be deduced from the known three-dimensional structure for human and murine antibody fragments. Initially, the FRs of the variable domains of an antibody molecule of interest are compared with corresponding FR sequences of human variable domains obtained from the above-identified sources. The most

homologous human V regions are then compared residue by residue to corresponding murine amino acids. The residues in the murine FR that differ from the human counterpart are replaced by the residues present in the human moiety using recombinant techniques well known in the art. Residue switching is only carried out with moieties which are at least partially exposed (solvent accessible), and care is exercised in the replacement of amino acid residues that may have a significant effect on the tertiary structure of V region domains, such as proline, glycine, and charged amino acids.

In this manner, the resultant "veneered" antigen-binding sites are thus designed to retain the rodent CDR residues, the residues substantially adjacent to the CDRs, the residues identified as buried or mostly buried (solvent inaccessible), the residues believed to participate in non-covalent (*e.g.*, electrostatic and hydrophobic) contacts between heavy and light chain domains, and the residues from conserved structural regions of the FRs which are believed to influence the "canonical" tertiary structures of the CDR loops. These design criteria are then used to prepare recombinant nucleotide sequences that combine the CDRs of both the heavy and light chain of a antigen-binding site into human-appearing FRs that can be used to transfect mammalian cells for the expression of recombinant human antibodies that exhibit the antigen specificity of the rodent antibody molecule.

An additional method for selecting antibodies that specifically bind to a TGF-beta binding protein or variant or fragment thereof is by phage display. *See, e.g.*, Winter et al., 1994 *Annu. Rev. Immunol.* 12:433-55; Burton et al., 1994 *Adv. Immunol.* 57:191-280. Human or murine immunoglobulin variable region gene combinatorial libraries may be created in phage vectors that can be screened to select Ig fragments (Fab, Fv, sFv, or multimers thereof) that bind specifically to TGF-beta binding protein or variant or fragment thereof. *See, e.g.*, U.S. Patent No. 5,223,409; William D. Huse et al., "Generation of a Large Combinational Library of the Immunoglobulin Repertoire in Phage Lambda," *Science 246*:1275-1281, December 1989; *see also* L. Sastry et al., "Cloning of the Immunological Repertoire in *Escherichia coli* for Generation of Monoclonal Catalytic Antibodies: Construction of a Heavy Chain Variable Region-Specific cDNA Library," *Proc. Natl. Acad. Sci. USA 86:*5728-5732, August 1989; *see also* Michelle Alting-Mees et al., "Monoclonal Antibody Expression Libraries: A Rapid Alternative to Hybridomas," *Strategies in Molecular Biology 3*:1-9, January 1990; Kang et al., 1991 *Proc. Natl. Acad. Sci. USA* 88:4363-66; Hoogenboom et al., 1992 *J. Molec. Biol.* 227:381-388; Schlebusch et al., 1997 *Hybridoma* 16:47-52 and references cited therein). A commercial system is available from Stratagene (La Jolla, California) which enables the production of

antibodies through recombinant techniques. Briefly, mRNA is isolated from a B cell population, and utilized to create heavy and light chain immunoglobulin cDNA expression libraries in the λ ImmunoZap(H) and λImmunoZap(L) vectors. Positive plaques may subsequently be converted to a non-lytic plasmid which allows high level expression of monoclonal antibody fragments from *E. coli*. Alternatively, a library containing a plurality of polynucleotide sequences encoding Ig variable region fragments may be inserted into the genome of a filamentous bacteriophage, such as M13 or a variant thereof, in frame with the sequence encoding a phage coat protein. A fusion protein may be a fusion of the coat protein with the light chain variable region domain and/or with the heavy chain variable region domain. According to certain embodiments, immunoglobulin Fab fragments may also be displayed on a phage particle (*see, e.g.,* U.S. Patent No. 5,698,426). These vectors may be screened individually or co-expressed to form Fab fragments or antibodies (*see* Huse et al., *supra*; *see also* Sastry et al., *supra*).

Similarly, portions or fragments, such as Fab and Fv fragments, of antibodies may also be constructed utilizing conventional enzymatic digestion or recombinant DNA techniques to incorporate the variable regions of a gene which encodes a specifically binding antibody. Within one embodiment, the genes which encode the variable region from a hybridoma producing a monoclonal antibody of interest are amplified using nucleotide primers for the variable region. These primers may be synthesized by one of ordinary skill in the art, or may be purchased from commercially available sources. Stratagene (La Jolla, California) sells primers for mouse and human variable regions including, among others, primers for V_{Ha}, V_{Hb}, V_{Hc}, V_{Hd}, C_{H1}, V_L and C_L regions. These primers may be utilized to amplify heavy or light chain variable regions, which may then be inserted into vectors such as ImmunoZAP™ H or ImmunoZAP™ L (Stratagene), respectively. These vectors may then be introduced into *E. coli*, yeast, or mammalian-based systems for expression. Utilizing these techniques, large amounts of a single-chain protein containing a fusion of the V_H and V_L domains may be produced (*see* Bird et al., *Science 242*:423-426, 1988). In addition, such techniques may be utilized to change a "murine" antibody to a "human" antibody, without altering the binding specificity of the antibody.

In certain particular embodiments of the invention, combinatorial phage libraries may also be used for humanization of non-human variable regions. *See, e.g.,* Rosok et al., 1996 *J. Biol. Chem. 271*:22611-18; Rader et al., 1998 *Proc. Natl. Acad. Sci. USA 95*:8910-15. A phage library may be screened to select an Ig variable region fragment of interest by immunodetection methods known in the art and described herein, and the DNA sequence of the inserted immunoglobulin gene in the phage so selected may be determined by standard techniques. *See,*

Sambrook et al., 2001 *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Press. The selected Ig-encoding sequence may then be cloned into another suitable vector for expression of the Ig fragment or, optionally, may be cloned into a vector containing Ig constant regions, for expression of whole immunoglobulin chains.

In certain other embodiments, the invention contemplates SOST-specific antibodies that are multimeric antibody fragments. Useful methodologies are described generally, for example in Hayden et al. 1997, *Curr Opin. Immunol.* 9:201-12; Coloma et al., 1997 *Nat. Biotechnol.* 15:159-63). For example, multimeric antibody fragments may be created by phage techniques to form miniantibodies (U.S. Patent No. 5,910 573) or diabodies (Holliger et al., 1997, *Cancer Immunol. Immunother.* 45:128-130).

In certain embodiments of the invention, an antibody specific for SOST may be an antibody that is expressed as an intracellular protein. Such intracellular antibodies are also referred to as intrabodies and may comprise an Fab fragment, or preferably comprise a scFv fragment (*see, e.g.*, Lecerf et al., *Proc. Natl. Acad. Sci. USA* 98:4764-49 (2001). The framework regions flanking the CDR regions can be modified to improve expression levels and solubility of an intrabody in an intracellular reducing environment (*see, e.g.*, Worn et al., *J. Biol. Chem.* 275:2795-803 (2000). An intrabody may be directed to a particular cellular location or organelle, for example by constructing a vector that comprises a polynucleotide sequence encoding the variable regions of an intrabody that may be operatively fused to a polynucleotide sequence that encodes a particular target antigen within the cell (*see, e.g.*, Graus-Porta et al., *Mol. Cell Biol.* 15:1182-91 (1995); Lener et al., *Eur. J. Biochem.* 267:1196-205 (2000)). An intrabody may be introduced into a cell by a variety of techniques available to the skilled artisan including via a gene therapy vector, or a lipid mixture (*e.g.*, Provectin™ manufactured by Imgenex Corporation, San Diego, CA), or according to photochemical internalization methods.

Introducing amino acid mutations into an immunoglobulin molecule specific for a TGF-beta binding protein may be useful to increase the specificity or affinity for TGF-beta binding protein or to alter an effector function. Immunoglobulins with higher affinity for TGF-beta binding protein may be generated by site-directed mutagenesis of particular residues. Computer assisted three-dimensional molecular modeling may be employed to identify the amino acid residues to be changed, in order to improve affinity for the TGF-beta binding protein. *See, e.g.*, Mountain et al., 1992, *Biotechnol. Genet. Eng. Rev.* 10: 1-142. Alternatively, combinatorial libraries of CDRs may be generated in M13 phage and screened for immunoglobulin fragments with improved affinity. *See, e.g.*, Glaser et al., 1992, *J. Immunol.* 149:3903-3913; Barbas et al.,

1994 *Proc. Natl. Acad. Sci. USA* 91:3809-13; U.S. Patent No. 5,792, 456.

Effector functions may also be altered by site-directed mutagenesis. *See, e.g.*, Duncan et al., 1988 *Nature* 332:563-64; Morgan et al., 1995 *Immunology* 86:319-24; Eghtedarzedeh-Kondri et al., 1997 *Biotechniques* 23:830-34. For example, mutation of the glycosylation site on the Fc portion of the immunoglobulin may alter the ability of the immunoglobulin to fix complement. *See, e.g.,* Wright et al., 1997 *Trends Biotechnol.* 15:26-32. Other mutations in the constant region domains may alter the ability of the immunoglobulin to fix complement, or to effect antibody-dependent cellular cytotoxicity. *See, e.g.*, Duncan et al., 1988 *Nature* 332:563-64; Morgan et al., 1995 *Immunology 86*:319-24; Sensel et al., 1997 *Mol. Immunol.* 34:1019-29.

According to certain embodiments, non-human, human, or humanized heavy chain and light chain variable regions of any of the Ig molecules described herein may be constructed as single chain Fv (scFv) polypeptide fragments (single chain antibodies). *See, e.g.*, Bird et al., 1988 *Science* 242:423-426; Huston et al., 1988 *Proc. Natl. Acad. Sci. USA* 85:5879-5883. Multi-functional scFv fusion proteins may be generated by linking a polynucleotide sequence encoding an scFv polypeptide in-frame with at least one polynucleotide sequence encoding any of a variety of known effector proteins. These methods are known in the art, and are disclosed, for example, in EP-B1-0318554, U.S. Patent No. 5,132,405, U.S. Patent No. 5,091,513, and U.S. Patent No. 5,476,786. By way of example, effector proteins may include immunoglobulin constant region sequences. *See, e.g.*, Hollenbaugh et al., 1995 *J. Immunol. Methods* 188:1-7. Other examples of effector proteins are enzymes. As a non-limiting example, such an enzyme may provide a biological activity for therapeutic purposes (*see, e.g.,* Siemers et al., 1997 *Bioconjug. Chem.* 8:510-19), or may provide a detectable activity, such as horseradish peroxidase-catalyzed conversion of any of a number of well-known substrates into a detectable product, for diagnostic uses. Still other examples of scFv fusion proteins include Ig-toxin fusions, or immunotoxins, wherein the scFv polypeptide is linked to a toxin.

The scFv or any antibody fragment described herein may, in certain embodiments, be fused to peptide or polypeptide domains that permits detection of specific binding between the fusion protein and antigen (*e.g.*, a TGF-beta binding protein). For example, the fusion polypeptide domain may be an affinity tag polypeptide for detecting binding of the scFv fusion protein to a TGF-beta binding protein by any of a variety of techniques with which those skilled in the art will be familiar. Examples of a peptide tag, include avidin, streptavidin or His (*e.g.*, polyhistidine). Detection techniques may also include, for example, binding of an avidin or streptavidin fusion protein to biotin or to a biotin mimetic sequence (*see, e.g.*, Luo et al., 1998 *J.*

Biotechnol. 65:225 and references cited therein), direct covalent modification of a fusion protein with a detectable moiety (*e.g.*, a labeling moiety), non-covalent binding of the fusion protein to a specific labeled reporter molecule, enzymatic modification of a detectable substrate by a fusion protein that includes a portion having enzyme activity, or immobilization (covalent or non-covalent) of the fusion protein on a solid-phase support. Other useful affinity polypeptides for construction of scFv fusion proteins may include streptavidin fusion proteins, as disclosed, for example, in WO 89/03422, U.S. 5,489,528, U.S. 5,672,691, WO 93/24631, U.S. 5,168,049, U.S. 5,272,254; avidin fusion proteins (*see, e.g.,* EP 511,747); an enzyme such as glutathione-S-transferase; and *Staphylococcus aureus* protein A polypeptide.

The polynucleotides encoding an antibody or fragment thereof that specifically bind a TGF-beta binding protein, as described herein, may be propagated and expressed according to any of a variety of well-known procedures for nucleic acid excision, ligation, transformation, and transfection using any number of known expression vectors. Thus, in certain embodiments expression of an antibody fragment may be preferred in a prokaryotic host, such as *Escherichia coli* (*see, e.g.,* Pluckthun et al., 1989 *Methods Enzymol.* 178:497-515). In certain other embodiments, expression of the antibody or a fragment thereof may be preferred in a eukaryotic host cell, including yeast (*e.g., Saccharomyces cerevisiae, Schizosaccharomyces pombe,* and *Pichia pastoris*), animal cells (including mammalian cells) or plant cells. Examples of suitable animal cells include, but are not limited to, myeloma (such as a mouse NSO line), COS, CHO, or hybridoma cells. Examples of plant cells include tobacco, corn, soybean, and rice cells.

Once suitable antibodies have been obtained, they may be isolated or purified by many techniques well known to those of ordinary skill in the art (*see Antibodies: A Laboratory Manual*, Harlow and Lane (eds.), Cold Spring Harbor Laboratory Press, 1988). Suitable techniques include peptide or protein affinity columns (including use of anti-constant region antibodies attached to the column matrix), HPLC or RP-HPLC, purification on protein A or protein G columns, or any combination of these techniques.

c. Mutant TGF-beta binding-proteins

As described herein and below in the Examples (e.g., Examples 8 and 9), altered versions of TGF-beta binding-protein which compete with native TGF-beta binding-protein's ability to block the activity of a particular TGF-beta family member should lead to increased bone density. Thus, mutants of TGF-beta binding-protein which bind to the TGF-beta family member but do not inhibit the function of the TGF-beta family member would meet the criteria. The mutant versions must effectively compete with the endogenous inhibitory functions of TGF-

beta binding-protein .

d. Production of proteins

Polypeptides described herein include the TGF binding protein sclerostin and variants thereof and antibodies or fragments thereof that specifically bind to sclerostin. The polynucleotides that encode these polypeptides include derivatives of the genes that are substantially similar to the genes and isolated nucleic acid molecules, and, when appropriate, the proteins (including peptides and polypeptides) that are encoded by the genes and their derivatives. As used herein, a nucleotide sequence is deemed to be "substantially similar" if (a) the nucleotide sequence is derived from the coding region of the above-described genes and nucleic acid molecules and includes, for example, portions of the sequence or allelic variations of the sequences discussed above, or alternatively, encodes a molecule which inhibits the binding of TGF-beta binding-protein to a member of the TGF-beta family; (b) the nucleotide sequence is capable of hybridization to nucleotide sequences of the present invention under moderate, high or very high stringency (*see* Sambrook et al., *Molecular Cloning: A Laboratory Manual*, 2nd ed., Cold Spring Harbor Laboratory Press, NY, 1989); and/or (c) the DNA sequences are degenerate as a result of the genetic code to the DNA sequences defined in (a) or (b). Further, the nucleic acid molecule disclosed herein includes both complementary and non-complementary sequences, provided the sequences otherwise meet the criteria set forth herein. Within the context of the present invention, high stringency means standard hybridization conditions (*e.g.*, 5XSSPE, 0.5% SDS at 65°C, or the equivalent).

The structure of the proteins encoded by the nucleic acid molecules described herein may be predicted from the primary translation products using the hydrophobicity plot function of, for example, P/C Gene or Intelligenetics Suite (Intelligenetics, Mountain View, California), or according to the methods described by Kyte and Doolittle (*J. Mol. Biol. 157*:105-132, 1982).

Proteins of the present invention may be prepared in the form of acidic or basic salts, or in neutral form. In addition, individual amino acid residues may be modified by oxidation or reduction. Furthermore, various substitutions, deletions, or additions may be made to the amino acid or nucleic acid sequences, the net effect of which is to retain or further enhance or decrease the biological activity of the mutant or wild-type protein. Moreover, due to degeneracy in the genetic code, for example, there may be considerable variation in nucleotide sequences encoding the same amino acid sequence.

Other derivatives of the proteins disclosed herein include conjugates of the proteins along

with other proteins or polypeptides. This may be accomplished, for example, by the synthesis of N-terminal or C-terminal fusion proteins which may be added to facilitate purification or identification of proteins (*see* U.S. Patent No. 4,851,341, *see also,* Hopp et al., *Bio/Technology* 6:1204, 1988.) Alternatively, fusion proteins such as Flag®/TGF-beta binding-protein be constructed in order to assist in the identification, expression, and analysis of the protein.

Proteins of the present invention may be constructed using a wide variety of techniques described herein. Further, mutations may be introduced at particular loci by synthesizing oligonucleotides containing a mutant sequence, flanked by restriction sites enabling ligation to fragments of the native sequence. Following ligation, the resulting reconstructed sequence encodes a derivative having the desired amino acid insertion, substitution, or deletion.

Alternatively, oligonucleotide-directed site-specific (or segment specific) mutagenesis procedures may be employed to provide an altered gene or nucleic acid molecule having particular codons altered according to the substitution, deletion, or insertion required. Exemplary methods of making the alterations set forth above are disclosed by Walder et al. (*Gene 42*:133, 1986); Bauer et al. (*Gene 37*:73, 1985); Craik (*BioTechniques*, January 1985, 12-19); Smith et al. (*Genetic Engineering: Principles and Methods*, Plenum Press, 1981); and Sambrook et al. (*supra*). Deletion or truncation derivatives of proteins (*e.g.,* a soluble extracellular portion) may also be constructed by utilizing convenient restriction endonuclease sites adjacent to the desired deletion. Subsequent to restriction, overhangs may be filled in and the DNA religated. Exemplary methods of making the alterations set forth above are disclosed by Sambrook et al. (*Molecular Cloning: A Laboratory Manual*, 2d Ed., Cold Spring Harbor Laboratory Press, 1989).

Mutations which are made in the nucleic acid molecules of the present invention preferably preserve the reading frame of the coding sequences. Furthermore, the mutations will preferably not create complementary regions that when transcribed could hybridize to produce secondary mRNA structures, such as loops or hairpins, that would adversely affect translation of the mRNA. Although a mutation site may be predetermined, it is not necessary that the nature of the mutation *per se* be predetermined. For example, in order to select for optimum characteristics of mutants at a given site, random mutagenesis may be conducted at the target codon and the expressed mutants screened for gain or loss or retention of biological activity. Alternatively, mutations may be introduced at particular loci by synthesizing oligonucleotides containing a mutant sequence, flanked by restriction sites enabling ligation to fragments of the native sequence. Following ligation, the resulting reconstructed sequence encodes a derivative

having the desired amino acid insertion, substitution, or deletion.

Nucleic acid molecules which encode proteins of the present invention may also be constructed utilizing techniques such as PCR mutagenesis, chemical mutagenesis (Drinkwater and Klinedinst, *PNAS 83*:3402-3406, 1986), by forced nucleotide misincorporation (*e.g.*, Liao and Wise *Gene 88*:107-111, 1990), or by use of randomly mutagenized oligonucleotides (Horwitz et al., *Genome 3*:112-117, 1989).

The present invention also provides for the manipulation and expression of the above described genes and nucleic acid molecules by culturing host cells containing a vector capable of expressing the above-described genes. Such vectors or vector constructs include either synthetic or cDNA-derived nucleic acid molecules encoding the desired protein, which are operably linked to suitable transcriptional or translational regulatory elements. Suitable regulatory elements may be derived from a variety of sources, including bacterial, fungal, viral, mammalian, insect, or plant genes. Selection of appropriate regulatory elements is dependent on the host cell chosen, and may be readily accomplished by one of ordinary skill in the art. Examples of regulatory elements include a transcriptional promoter and enhancer or RNA polymerase binding sequence, a transcriptional terminator, and a ribosomal binding sequence, including a translation initiation signal.

Nucleic acid molecules that encode any of the proteins described above may be readily expressed by a wide variety of prokaryotic and eukaryotic host cells, including bacterial, mammalian, yeast or other fungi, viral, insect, or plant cells. Methods for transforming or transfecting such cells to express foreign DNA are well known in the art (*see, e.g.*, Itakura et al., U.S. Patent No. 4,704,362; Hinnen et al., *Proc. Natl. Acad. Sci. USA 75*:1929-1933, 1978; Murray et al., U.S. Patent No. 4,801,542; Upshall et al., U.S. Patent No. 4,935,349; Hagen et al., U.S. Patent No. 4,784,950; Axel et al., U.S. Patent No. 4,399,216; Goeddel et al., U.S. Patent No. 4,766,075; and Sambrook et al. *Molecular Cloning: A Laboratory Manual*, 2nd ed., Cold Spring Harbor Laboratory Press, 1989; for plant cells see Czako and Marton, *Plant Physiol. 104*:1067-1071, 1994; and Paszkowski et al., *Biotech. 24*:387-392, 1992).

Bacterial host cells suitable for carrying out the present invention include *E. coli*, *B. subtilis*, *Salmonella typhimurium*, and various species within the genera *Pseudomonas*, *Streptomyces*, and *Staphylococcus*, as well as many other bacterial species well known to one of ordinary skill in the art and described herein. A representative example of a bacterial host cell includes *E. coli* DH5α (Stratagene, LaJolla, California).

Bacterial expression vectors preferably comprise a promoter which functions in the host

cell, one or more selectable phenotypic markers, and a bacterial origin of replication. Representative promoters include the β-lactamase (penicillinase) and lactose promoter system (*see* Chang et al., *Nature 275*:615, 1978), the T7 RNA polymerase promoter (Studier et al., *Meth. Enzymol. 185*:60-89, 1990), the lambda promoter (Elvin et al., *Gene 87*:123-126, 1990), the *trp* promoter (Nichols and Yanofsky, *Meth. in Enzymology 101*:155, 1983), and the *tac* promoter (Russell et al., *Gene 20*:231, 1982). Representative selectable markers include various antibiotic resistance markers such as the kanamycin or ampicillin resistance genes. Many plasmids suitable for transforming host cells are well known in the art, including among others, pBR322 (*see* Bolivar et al., *Gene 2*:95, 1977), the pUC plasmids pUC18, pUC19, pUC118, pUC119 (*see* Messing, *Meth. in Enzymology 101*:20-77, 1983 and Vieira and Messing, *Gene 19*:259-268, 1982), and pNH8A, pNH16a, pNH18a, and Bluescript M13 (Stratagene, La Jolla, California).

Yeast and fungi host cells suitable for carrying out the present invention include, among others, *Saccharomyces pombe*, *Saccharomyces cerevisiae*, the genera *Pichia* or *Kluyveromyces* and various species of the genus *Aspergillus* (McKnight et al., U.S. Patent No. 4,935,349). Suitable expression vectors for yeast and fungi include, among others, YCp50 (ATCC No. 37419) for yeast, and the amdS cloning vector pV3 (Turnbull, *Bio/Technology 7*:169, 1989), YRp7 (Struhl et al., *Proc. Natl. Acad. Sci. USA 76*:1035-1039, 1978), YEp13 (Broach et al., *Gene 8*:121-133, 1979), pJDB249 and pJDB219 (Beggs, *Nature 275*:104-108, 1978) and derivatives thereof.

Preferred promoters for use in yeast include promoters from yeast glycolytic genes (Hitzeman et al., *J. Biol. Chem. 255*:12073-12080, 1980; Alber and Kawasaki, *J. Mol. Appl. Genet. 1*:419-434, 1982) or alcohol dehydrogenase genes (Young et al., in *Genetic Engineering of Microorganisms for Chemicals*, Hollaender et al. (eds.), p. 355, Plenum, New York, 1982; Ammerer, *Meth. Enzymol. 101*:192-201, 1983). Examples of useful promoters for fungi vectors include those derived from *Aspergillus nidulans* glycolytic genes, such as the *adh3* promoter (McKnight et al., *EMBO J. 4*:2093-2099, 1985). The expression units may also include a transcriptional terminator. An example of a suitable terminator is the *adh3* terminator (McKnight et al., *supra*, 1985).

As with bacterial vectors, the yeast vectors will generally include a selectable marker, which may be one of any number of genes that exhibit a dominant phenotype for which a phenotypic assay exists to enable transformants to be selected. Preferred selectable markers are those that complement host cell auxotrophy, provide antibiotic resistance, or enable a cell to

utilize specific carbon sources, and include *leu2* (Broach et al., *ibid.*), *ura3* (Botstein et al., *Gene 8*:17, 1979), or *his3* (Struhl et al., *ibid.*). Another suitable selectable marker is the *cat* gene, which confers chloramphenicol resistance on yeast cells.

Techniques for transforming fungi are well known in the literature and have been described, for instance, by Beggs (*ibid.*), Hinnen et al. (*Proc. Natl. Acad. Sci. USA 75*:1929-1933, 1978), Yelton et al. (*Proc. Natl. Acad. Sci. USA 81*:1740-1747, 1984), and Russell (*Nature 301*:167-169, 1983). The genotype of the host cell may contain a genetic defect that is complemented by the selectable marker present on the expression vector. Choice of a particular host and selectable marker is well within the level of ordinary skill in the art.

Protocols for the transformation of yeast are also well known to those of ordinary skill in the art. For example, transformation may be readily accomplished either by preparation of spheroplasts of yeast with DNA (*see* Hinnen et al., *PNAS USA 75*:1929, 1978) or by treatment with alkaline salts such as LiCl (*see* Itoh et al., *J. Bacteriology 153*:163, 1983). Transformation of fungi may also be carried out using polyethylene glycol as described by Cullen et al. (*Bio/Technology 5*:369, 1987).

Viral vectors include those that comprise a promoter that directs the expression of an isolated nucleic acid molecule that encodes a desired protein as described above. A wide variety of promoters may be utilized within the context of the present invention, including for example, promoters such as MoMLV LTR, RSV LTR, Friend MuLV LTR, adenoviral promoter (Ohno et al., *Science 265*:781-784, 1994), neomycin phosphotransferase promoter/enhancer, late parvovirus promoter (Koering et al., *Hum. Gene Therap. 5*:457-463, 1994), Herpes TK promoter, SV40 promoter, metallothionein IIa gene enhancer/promoter, cytomegalovirus immediate early promoter, and the cytomegalovirus immediate late promoter. Within particularly preferred embodiments of the invention, the promoter is a tissue-specific promoter (*see e.g.*, WO 91/02805; EP 0,415,731; and WO 90/07936). Representative examples of suitable tissue specific promoters include neural specific enolase promoter, platelet derived growth factor beta promoter, bone morphogenic protein promoter, human alpha1-chimaerin promoter, synapsin I promoter and synapsin II promoter. In addition to the above-noted promoters, other viral-specific promoters (*e.g.*, retroviral promoters (including those noted above, as well as others such as HIV promoters), hepatitis, herpes (*e.g.*, EBV), and bacterial, fungal or parasitic (*e.g.*, malarial) -specific promoters may be utilized in order to target a specific cell or tissue which is infected with a virus, bacteria, fungus, or parasite.

Mammalian cells suitable for carrying out the present invention include, among others

COS, CHO, SaOS, osteosarcomas, KS483, MG-63, primary osteoblasts, and human or mammalian bone marrow stroma. Mammalian expression vectors for use in carrying out the present invention will include a promoter capable of directing the transcription of a cloned gene, nucleic acid molecule, or cDNA. Preferred promoters include viral promoters and cellular promoters. Bone specific promoters include the promoter for bone sialo-protein and the promoter for osteocalcin. Viral promoters include the cytomegalovirus immediate early promoter (Boshart et al., *Cell 41*:521-530, 1985), cytomegalovirus immediate late promoter, SV40 promoter (Subramani et al., *Mol. Cell. Biol. 1*:854-864, 1981), MMTV LTR, RSV LTR, metallothionein-1, adenovirus E1a. Cellular promoters include the mouse metallothionein-1 promoter (Palmiter et al., U.S. Patent No. 4,579,821), a mouse V_K promoter (Bergman et al., *Proc. Natl. Acad. Sci. USA 81*:7041-7045, 1983; Grant et al., *Nucleic Acids Res. 15*:5496, 1987) and a mouse V_H promoter (Loh et al., *Cell 33*:85-93, 1983). The choice of promoter will depend, at least in part, upon the level of expression desired or the recipient cell line to be transfected.

Such expression vectors may also contain a set of RNA splice sites located downstream from the promoter and upstream from the DNA sequence encoding the peptide or protein of interest. Preferred RNA splice sites may be obtained from adenovirus and/or immunoglobulin genes. Also contained in the expression vectors is a polyadenylation signal located downstream of the coding sequence of interest. Suitable polyadenylation signals include the early or late polyadenylation signals from SV40 (Kaufman and Sharp, *ibid.*), the polyadenylation signal from the Adenovirus 5 E1B region and the human growth hormone gene terminator (DeNoto et al., *Nucleic Acids Res. 9*:3719-3730, 1981). The expression vectors may include a noncoding viral leader sequence, such as the Adenovirus 2 tripartite leader, located between the promoter and the RNA splice sites. Preferred vectors may also include enhancer sequences, such as the SV40 enhancer. Expression vectors may also include sequences encoding the adenovirus VA RNAs. Suitable expression vectors can be obtained from commercial sources (*e.g.*, Stratagene, La Jolla, California).

Vector constructs comprising cloned DNA sequences can be introduced into cultured mammalian cells by, for example, calcium phosphate-mediated transfection (Wigler et al., *Cell 14*:725, 1978; Corsaro and Pearson, *Somatic Cell Genetics 7*:603, 1981; Graham and Van der Eb, *Virology 52*:456, 1973), electroporation (Neumann et al., *EMBO J. 1*:841-845, 1982), or DEAE-dextran mediated transfection (Ausubel et al. (eds.), *Current Protocols in Molecular Biology*, John Wiley and Sons, Inc., NY, 1987). To identify cells that have stably transfected

with the vector or have integrated the cloned DNA, a selectable marker is generally introduced into the cells along with the gene or cDNA of interest. Preferred selectable markers for use in cultured mammalian cells include genes that confer resistance to drugs, such as neomycin, hygromycin, and methotrexate. The selectable marker may be an amplifiable selectable marker. Preferred amplifiable selectable markers are the DHFR gene and the neomycin resistance gene. Selectable markers are reviewed by Thilly (*Mammalian Cell Technology*, Butterworth Publishers, Stoneham, Massachusetts, which is incorporated herein by reference).

Mammalian cells containing a suitable vector are allowed to grow for a period of time, typically 1-2 days, to begin expressing the DNA sequence(s) of interest. Drug selection is then applied to select for growth of cells that are expressing the selectable marker in a stable fashion. For cells that have been transfected with an amplifiable, selectable marker, the drug concentration may be increased in a stepwise manner to select for increased copy number of the cloned sequences, thereby increasing expression levels. Cells expressing the introduced sequences are selected and screened for production of the protein of interest in the desired form or at the desired level. Cells that satisfy these criteria can then be cloned and scaled up for production.

Protocols for the transfection of mammalian cells are well known to those of ordinary skill in the art. Representative methods include calcium phosphate mediated transfection, electroporation, lipofection, retroviral, adenoviral and protoplast fusion-mediated transfection (*see* Sambrook et al., *supra*). Naked vector constructs can also be taken up by muscular cells or other suitable cells subsequent to injection into the muscle of a mammal (or other animals).

Methods for using insect host cells and plant host cells for production of polypeptides are known in the art and described herein. Numerous insect host cells known in the art can also be useful within the present invention,. For example, the use of baculoviruses as vectors for expressing heterologous DNA sequences in insect cells has been reviewed by Atkinson et al. (*Pestic. Sci. 28*:215-224,1990). Numerous vectors and plant host cells known in the art can also be useful within the present invention, for example, the use of *Agrobacterium rhizogenes* as vectors for expressing genes and nucleic acid molecules in plant cells (see review by Sinkar et al., *J. Biosci.* (*Bangalore 11*:47-58, 1987).

Within related aspects of the present invention, proteins of the present invention may be expressed in a transgenic animal whose germ cells and somatic cells contain a gene which encodes the desired protein and which is operably linked to a promoter effective for the expression of the gene. Alternatively, in a similar manner transgenic animals may be prepared

that lack the desired gene (*e.g.*, "knock-out" mice). Such transgenics may be prepared in a variety of non-human animals, including mice, rats, rabbits, sheep, dogs, goats, and pigs (*see* Hammer et al., *Nature 315*:680-683, 1985, Palmiter et al., *Science 222*:809-814, 1983, Brinster et al., *Proc. Natl. Acad. Sci. USA 82*:4438-4442, 1985, Palmiter and Brinster, *Cell 41*:343-345, 1985, and U.S. Patent Nos. 5,175,383, 5,087,571, 4,736,866, 5,387,742, 5,347,075, 5,221,778, and 5,175,384). Briefly, an expression vector, including a nucleic acid molecule to be expressed together with appropriately positioned expression control sequences, is introduced into pronuclei of fertilized eggs, for example, by microinjection. Integration of the injected DNA is detected by blot analysis of DNA from tissue samples. It is preferred that the introduced DNA be incorporated into the germ line of the animal so that it is passed on to the animal's progeny. Tissue-specific expression may be achieved through the use of a tissue-specific promoter, or through the use of an inducible promoter, such as the metallothionein gene promoter (Palmiter et al., 1983, *supra*), which allows regulated expression of the transgene.

Proteins can be isolated by, among other methods, culturing suitable host and vector systems to produce the recombinant translation products as described herein. Supernatants from such cell lines, or protein inclusions, or whole cells from which the protein is not excreted into the supernatant, can then be treated by a variety of purification procedures in order to isolate the desired proteins. For example, the supernatant may be first concentrated using commercially available protein concentration filters, such as an Amicon or Millipore Pellicon ultrafiltration unit. Following concentration, the concentrate may be applied to a suitable purification matrix such as, for example, an anti-protein antibody (*e.g.*, an antibody that specifically binds to the polypeptide to be isolated) bound to a suitable support. Alternatively, anion or cation exchange resins may be employed in order to purify the protein. As a further alternative, one or more reverse-phase high performance liquid chromatography (RP-HPLC) steps may be employed to further purify the protein. Other methods of isolating the proteins of the present invention are well known in the art.

The purity of an isolated polypeptide may be determined by techniques known in the art and described herein, such as gel electrophoresis and chromatography methods. Preferably, such isolated polypeptides are at least about 90% pure, more preferably at least about 95% pure, and most preferably at least about 99% pure. Within certain specific embodiments, a protein is deemed to be "isolated" within the context of the present invention if no other undesired protein is detected pursuant to SDS-PAGE analysis followed by Coomassie blue staining. Within other embodiments, the desired protein can be isolated such that no other undesired protein is detected

pursuant to SDS-PAGE analysis followed by silver staining.

3. Nucleic Acid Molecules

Within other aspects of the invention, nucleic acid molecules are provided which are capable of inhibiting TGF-beta binding-protein binding to a member of the TGF-beta family. For example, within one embodiment antisense oligonucleotide molecules are provided which specifically inhibit expression of TGF-beta binding-protein nucleic acid sequences (*see generally*, Hirashima et al. in *Molecular Biology of RNA: New Perspectives* (M. Inouye and B. S. Dudock, eds., 1987 Academic Press, San Diego, p. 401); *Oligonucleotides: Antisense Inhibitors of Gene Expression* (J.S. Cohen, ed., 1989 MacMillan Press, London); Stein and Cheng, *Science 261*:1004-1012, 1993; WO 95/10607; U.S. Patent No. 5,359,051; WO 92/06693; and EP-A2-612844). Briefly, such molecules are constructed such that they are complementary to, and able to form Watson-Crick base pairs with, a region of transcribed TGF-beta binding-protein mRNA sequence. The resultant double-stranded nucleic acid interferes with subsequent processing of the mRNA, thereby preventing protein synthesis (see Example 10).

Within other aspects of the invention, ribozymes are provided which are capable of inhibiting the TGF-beta binding-protein binding to a member of the TGF-beta family. As used herein, "ribozymes" are intended to include RNA molecules that contain anti-sense sequences for specific recognition, and an RNA-cleaving enzymatic activity. The catalytic strand cleaves a specific site in a target RNA at greater than stoichiometric concentration. A wide variety of ribozymes may be utilized within the context of the present invention, including for example, the hammerhead ribozyme (for example, as described by Forster and Symons, *Cell 48*:211-220, 1987; Haseloff and Gerlach, *Nature 328*:596-600, 1988; Walbot and Bruening, *Nature 334*:196, 1988; Haseloff and Gerlach, *Nature 334*:585, 1988); the hairpin ribozyme (for example, as described by Haseloff et al., U.S. Patent No. 5,254,678, issued October 19, 1993 and Hempel et al., European Patent Publication No. 0 360 257, published March 26, 1990); and *Tetrahymena* ribosomal RNA-based ribozymes (see Cech et al., U.S. Patent No. 4,987,071). Ribozymes of the present invention typically consist of RNA, but may also be composed of DNA, nucleic acid analogs (*e.g.*, phosphorothioates), or chimerics thereof (*e.g.*, DNA/RNA/RNA).

4. Labels

The gene product or any of the candidate molecules described above and below, may be labeled with a variety of compounds, including for example, fluorescent molecules, toxins, and radionuclides. Representative examples of fluorescent molecules include fluorescein, *Phycobili*

proteins, such as phycoerythrin, rhodamine, Texas red and luciferase. Representative examples of toxins include ricin, abrin diphtheria toxin, cholera toxin, gelonin, pokeweed antiviral protein, tritin, *Shigella* toxin, and *Pseudomonas* exotoxin A. Representative examples of radionuclides include Cu-64, Ga-67, Ga-68, Zr-89, Ru-97, Tc-99m, Rh-105, Pd-109, In-111, I-123, I-125, I-131, Re-186, Re-188, Au-198, Au-199, Pb-203, At-211, Pb-212 and Bi-212. In addition, the antibodies described above may also be labeled or conjugated to one partner of a ligand binding pair. Representative examples include avidin-biotin, streptavidin-biotin, and riboflavin-riboflavin binding protein.

Methods for conjugating or labeling the molecules described herein with the representative labels set forth above may be readily accomplished by one of ordinary skill in the art (*see* Trichothecene Antibody Conjugate, U.S. Patent No. 4,744,981; Antibody Conjugate, U.S. Patent No. 5,106,951; Fluorogenic Materials and Labeling Techniques, U.S. Patent No. 4,018,884; Metal Radionuclide Labeled Proteins for Diagnosis and Therapy, U.S. Patent No. 4,897,255; and Metal Radionuclide Chelating Compounds for Improved Chelation Kinetics, U.S. Patent No. 4,988,496; *see also* Inman, *Methods In Enzymology*, Vol. 34, *Affinity Techniques, Enzyme Purification: Part B*, Jakoby and Wilchek (eds.), Academic Press, New York, p. 30, 1974; *see also* Wilchek and Bayer, "The Avidin-Biotin Complex in Bioanalytical Applications," *Anal. Biochem. 171*:1-32, 1988).

PHARMACEUTICAL COMPOSITIONS

As noted above, the present invention also provides a variety of pharmaceutical compositions, comprising one of the above-described molecules which inhibits the TGF-beta binding-protein binding to a member of the TGF-beta family along with a pharmaceutically or physiologically acceptable carrier, excipients or diluents. Generally, such carriers should be nontoxic to recipients at the dosages and concentrations employed. Ordinarily, the preparation of such compositions entails combining the therapeutic agent with buffers, antioxidants such as ascorbic acid, low molecular weight (less than about 10 residues) polypeptides, proteins, amino acids, carbohydrates including glucose, maltose, sucrose or dextrins, chelating agents such as EDTA, glutathione and other stabilizers and excipients. Neutral buffered saline or saline mixed with nonspecific serum albumin are exemplary appropriate diluents.

The pharmaceutical compositions of the present invention may be prepared for administration by a variety of different routes. In general, the type of carrier is selected based on the mode of administration. Pharmaceutical compositions may be formulated for any appropriate manner of administration, including, for example, topical, oral, nasal, intrathecal,

rectal, vaginal, sublingual or parenteral administration, including subcutaneous, intravenous, intramuscular, intrasternal, intracavernous, intrameatal, or intraurethral injection or infusion. A pharmaceutical composition (*e.g.*, for oral administration or delivery by injection) may be in the form of a liquid (*e.g.*, an elixir, syrup, solution, emulsion or suspension). A liquid pharmaceutical composition may include, for example, one or more of the following: sterile diluents such as water for injection, saline solution, preferably physiological saline, Ringer's solution, isotonic sodium chloride, fixed oils that may serve as the solvent or suspending medium, polyethylene glycols, glycerin, propylene glycol or other solvents; antibacterial agents; antioxidants; chelating agents; buffers such as acetates, citrates or phosphates and agents for the adjustment of tonicity such as sodium chloride or dextrose. A parenteral preparation can be enclosed in ampoules, disposable syringes or multiple dose vials made of glass or plastic. The use of physiological saline is preferred, and an injectable pharmaceutical composition is preferably sterile.

The compositions described herein may be formulated for sustained release (*i.e.*, a formulation such as a capsule or sponge that effects a slow release of compound following administration). Such compositions may generally be prepared using well known technology and administered by, for example, oral, rectal or subcutaneous implantation, or by implantation at the desired target site. Sustained-release formulations may contain an agent dispersed in a carrier matrix and/or contained within a reservoir surrounded by a rate controlling membrane. Carriers for use within such formulations are biocompatible, and may also be biodegradable; preferably the formulation provides a relatively constant level of active component release. The amount of active compound contained within a sustained release formulation depends upon the site of implantation, the rate and expected duration of release and the nature of the condition to be treated or prevented. Illustrative carriers useful in this regard include microparticles of poly(lactide-co-glycolide), polyacrylate, latex, starch, cellulose, dextran and the like. Other illustrative delayed-release carriers include supramolecular biovectors, which comprise a non-liquid hydrophilic core (*e.g.*, a cross-linked polysaccharide or oligosaccharide) and, optionally, an external layer comprising an amphiphilic compound, such as a phospholipid (*see e.g.*, U.S. Patent No. 5,151,254 and PCT applications WO 94/20078, WO/94/23701 and WO 96/06638).

In another illustrative embodiment, biodegradable microspheres (*e.g.*, polylactate polyglycolate) are employed as carriers for the compositions of this invention. Suitable biodegradable microspheres are disclosed, for example, in U.S. Patent Nos. 4,897,268; 5,075,109; 5,928,647; 5,811,128; 5,820,883; 5,853,763; 5,814,344, 5,407,609 and 5,942,252.

Modified hepatitis B core protein carrier systems, such as described in WO/99 40934, and references cited therein, will also be useful for many applications. Another illustrative carrier/delivery system employs a carrier comprising particulate-protein complexes, such as those described in U.S. Patent No. 5,928,647, which are capable of inducing a class I-restricted cytotoxic T lymphocyte responses in a host.

In another illustrative embodiment, calcium phosphate core particles are employed as carriers or as controlled release matrices for the compositions of this invention. Exemplary calcium phosphate particles are disclosed, for example, in published patent application No. WO/0046147.

For pharmaceutical compositions comprising a polynucleotide encoding an anti-SOST antibody and/or modulating agent (such that the polypeptide and/or modulating agent is generated *in situ*), the polynucleotide may be present within any of a variety of delivery systems known to those of ordinary skill in the art, including nucleic acid, and bacterial, viral and mammalian expression systems. Techniques for incorporating DNA into such expression systems are well known to those of ordinary skill in the art. The DNA may also be "naked," as described, for example, in Ulmer et al., *Science 259*:1745-1749, 1993 and reviewed by Cohen, *Science 259*:1691-1692, 1993. The uptake of naked DNA may be increased by coating the DNA onto biodegradable beads, which are efficiently transported into the cells.

The development of suitable dosing and treatment regimens for using the particular compositions described herein in a variety of treatment regimens, including *e.g.*, oral, parenteral, intravenous, intranasal, and intramuscular administration and formulation, is well known in the art, some of which are briefly discussed below for general purposes of illustration.

In certain applications, the pharmaceutical compositions disclosed herein may be delivered *via* oral administration to an animal. As such, these compositions may be formulated with an inert diluent or with an assimilable edible carrier, or they may be enclosed in hard- or soft-shell gelatin capsule, or they may be compressed into tablets, or they may be incorporated directly with the food of the diet.

In certain circumstances it will be desirable to deliver the pharmaceutical compositions disclosed herein parenterally, intravenously, intramuscularly, or even intraperitoneally. Such approaches are well known to the skilled artisan, some of which are further described, for example, in U.S. Patent No. 5,543,158; U.S. Patent No. 5,641,515 and U.S. Patent No. 5,399,363. In certain embodiments, solutions of the active compounds as free base or pharmacologically acceptable salts may be prepared in water suitably mixed with a surfactant,

such as hydroxypropylcellulose. Dispersions may also be prepared in glycerol, liquid polyethylene glycols, and mixtures thereof and in oils. Under ordinary conditions of storage and use, these preparations generally will contain a preservative to prevent the growth of microorganisms.

Illustrative pharmaceutical forms suitable for injectable use include sterile aqueous solutions or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersions (for example, see U.S. Patent No. 5,466,468). In all cases the form must be sterile and must be fluid to the extent that easy syringability exists. It must be stable under the conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms, such as bacteria and fungi. The carrier can be a solvent or dispersion medium containing, for example, water, ethanol, polyol (*e.g.*, glycerol, propylene glycol, and liquid polyethylene glycol, and the like), suitable mixtures thereof, and/or vegetable oils. Proper fluidity may be maintained, for example, by the use of a coating, such as lecithin, by the maintenance of the required particle size in the case of dispersion and/or by the use of surfactants. The prevention of the action of microorganisms can be facilitated by various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, sorbic acid, thimerosal, and the like. In many cases, it will be preferable to include isotonic agents, for example, sugars or sodium chloride. Prolonged absorption of the injectable compositions can be brought about by the use in the compositions of agents delaying absorption, for example, aluminum monostearate and gelatin.

In one embodiment, for parenteral administration in an aqueous solution, the solution should be suitably buffered if necessary and the liquid diluent first rendered isotonic with sufficient saline or glucose. These particular aqueous solutions are especially suitable for intravenous, intramuscular, subcutaneous and intraperitoneal administration. In this connection, a sterile aqueous medium that can be employed will be known to those of skill in the art in light of the present disclosure. For example, one dosage may be dissolved in 1 ml of isotonic NaCl solution and either added to 1000 ml of hypodermoclysis fluid or injected at the proposed site of infusion, (see for example, *Remington's Pharmaceutical Sciences*, 15th ed., pp. 1035-1038 and 1570-1580). Some variation in dosage will necessarily occur depending on the condition of the subject being treated. Moreover, for human administration, preparations will of course preferably meet sterility, pyrogenicity, and the general safety and purity standards as required by FDA Office of Biologics standards.

In another embodiment of the invention, the compositions disclosed herein may be

formulated in a neutral or salt form. Illustrative pharmaceutically-acceptable salts include the acid addition salts (formed with the free amino groups of the protein) and which are formed with inorganic acids such as, for example, hydrochloric or phosphoric acids, or such organic acids as acetic, oxalic, tartaric, mandelic, and the like. Salts formed with the free carboxyl groups can also be derived from inorganic bases such as, for example, sodium, potassium, ammonium, calcium, or ferric hydroxides, and such organic bases as isopropylamine, trimethylamine, histidine, procaine and the like. Upon formulation, solutions will be administered in a manner compatible with the dosage formulation and in such amount as is therapeutically effective.

The carriers can further comprise any and all solvents, dispersion media, vehicles, coatings, diluents, antibacterial and antifungal agents, isotonic and absorption delaying agents, buffers, carrier solutions, suspensions, colloids, and the like. The use of such media and agents for pharmaceutical active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the active ingredient, its use in the therapeutic compositions is contemplated. Supplementary active ingredients can also be incorporated into the compositions. The phrase "pharmaceutically-acceptable" refers to molecular entities and compositions that do not produce an allergic or similar untoward reaction when administered to a human.

In certain embodiments, liposomes, nanocapsules, microparticles, lipid particles, vesicles, and the like, are used for the introduction of the compositions of the present invention into suitable host cells/organisms. In particular, the compositions of the present invention may be formulated for delivery either encapsulated in a lipid particle, a liposome, a vesicle, a nanosphere, or a nanoparticle or the like. Alternatively, compositions of the present invention can be bound, either covalently or non-covalently, to the surface of such carrier vehicles.

The formation and use of liposome and liposome-like preparations as potential drug carriers is generally known to those of skill in the art (see for example, Lasic, *Trends Biotechnol.* *16*(7):307-21, 1998; Takakura, *Nippon Rinsho* *56*(3):691-95, 1998; Chandran et al., *Indian J. Exp. Biol.* *35*(8):801-09, 1997; Margalit, *Crit. Rev. Ther. Drug Carrier Syst.* *12*(2-3):233-61, 1995; U.S. Patent No. 5,567,434; U.S. Patent No. 5,552,157; U.S. Patent No. 5,565,213; U.S. Patent No. 5,738,868 and U.S. Patent No. 5,795,587, each specifically incorporated herein by reference in its entirety).

Liposomes have been used successfully with a number of cell types that are normally difficult to transfect by other procedures, including T cell suspensions, primary hepatocyte cultures and PC 12 cells (Renneisen et al., *J. Biol. Chem.* *265*(27):16337-42, 1990; Muller et al.,

DNA Cell Biol. 9(3):221-29, 1990). In addition, liposomes are free of the DNA length constraints that are typical of viral-based delivery systems. Liposomes have been used effectively to introduce genes, various drugs, radiotherapeutic agents, enzymes, viruses, transcription factors, allosteric effectors and the like, into a variety of cultured cell lines and animals. Furthermore, he use of liposomes does not appear to be associated with autoimmune responses or unacceptable toxicity after systemic delivery.

In certain embodiments, liposomes are formed from phospholipids that are dispersed in an aqueous medium and spontaneously form multilamellar concentric bilayer vesicles (also termed multilamellar vesicles (MLVs).

Alternatively, in other embodiments, the invention provides for pharmaceutically-acceptable nanocapsule formulations of the compositions of the present invention. Nanocapsules can generally entrap compounds in a stable and reproducible way (see, for example, Quintanar-Guerrero et al., *Drug Dev. Ind. Pharm.* 24(12):1113-28, 1998). To avoid side effects due to intracellular polymeric overloading, such ultrafine particles (sized around 0.1 μm) may be designed using polymers able to be degraded *in vivo*. Such particles can be made as described, for example, by Couvreur et al., *Crit. Rev. Ther. Drug Carrier Syst.* 5(1):1-20, 1988; zur Muhlen et al., *Eur. J. Pharm. Biopharm.* 45(2):149-55, 1998; Zambaux et al., *J. Controlled Release* 50(1-3):31-40, 1998; and U.S. Patent No. 5,145,684.

In addition, pharmaceutical compositions of the present invention may be placed within containers, along with packaging material that provides instructions regarding the use of such pharmaceutical compositions. Generally, such instructions will include a tangible expression describing the reagent concentration, as well as within certain embodiments, relative amounts of excipient ingredients or diluents (*e.g.*, water, saline or PBS) which may be necessary to reconstitute the pharmaceutical composition.

METHODS OF TREATMENT

The present invention also provides methods for increasing the mineral content and mineral density of bone. Briefly, numerous conditions result in the loss of bone mineral content, including for example, disease, genetic predisposition, accidents which result in the lack of use of bone (*e.g.*, due to fracture), therapeutics which effect bone resorption, or which kill bone forming cells and normal aging. Through use of the molecules described herein which inhibit the TGF-beta binding-protein binding to a TGF-beta family member such conditions may be treated or prevented. As utilized herein, it should be understood that bone mineral content has been increased if bone mineral content has been increased in a statistically significant manner

(*e.g.*, greater than one-half standard deviation), at a selected site.

A wide variety of conditions that result in loss of bone mineral content may be treated with the molecules described herein. Patients with such conditions may be identified through clinical diagnosis utilizing well known techniques (see, *e.g.*, Harrison's Principles of Internal Medicine, McGraw-Hill, Inc.). Representative examples of diseases that may be treated included dysplasias, wherein there is abnormal growth or development of bone. Representative examples of such conditions include achondroplasia, cleidocranial dysostosis, enchondromatosis, fibrous dysplasia, Gaucher's Disease, hypophosphatemic rickets, Marfan's Syndrome, multiple hereditary exotoses, neurofibromatosis, osteogenesis imperfecta, osteopetrosis, osteopoikilosis, sclerotic lesions, fractures, periodontal disease, pseudoarthrosis, and pyogenic osteomyelitis.

Other conditions which may be treated or prevented include a wide variety of causes of osteopenia (*i.e.*, a condition that causes greater than one standard deviation of bone mineral content or density below peak skeletal mineral content at youth). Representative examples of such conditions include anemic states, conditions caused by steroids, conditions caused by heparin, bone marrow disorders, scurvy, malnutrition, calcium deficiency, idiopathic osteoporosis, congenital osteopenia or osteoporosis, alcoholism, chronic liver disease, senility, postmenopausal state, oligomenorrhea, amenorrhea, pregnancy, diabetes mellitus, hyperthyroidism, Cushing's disease, acromegaly, hypogonadism, immobilization or disuse, reflex sympathetic dystrophy syndrome, transient regional osteoporosis, and osteomalacia.

Within one aspect of the present invention, bone mineral content or density may be increased by administering to a warm-blooded animal a therapeutically effective amount of a molecule that inhibits binding of the TGF-beta binding-protein to a TGF-beta family member. Examples of warm-blooded animals that may be treated include both vertebrates and mammals, including for example humans, horses, cows, pigs, sheep, dogs, cats, rats and mice. Representative examples of therapeutic molecules include ribozymes, ribozyme genes, antisense oligonucleotides, and antibodies (*e.g.*, humanized antibodies or any other antibody described herein).

Within other aspects of the present invention, methods are provided for increasing bone density, comprising the steps of introducing into cells which home to bone, a vector that directs the expression of a molecule which inhibits binding of the TGF-beta binding-protein to a member of the TGF-beta family, and administering the vector-containing cells to a warm-blooded animal. Briefly, cells that home to bone may be obtained directly from the bone of

patients (*e.g.*, cells obtained from the bone marrow such as CD34+, osteoblasts, osteocytes, and the like), from peripheral blood, or from cultures.

A vector that directs the expression of a molecule that inhibits the binding of TGF-beta binding-protein to a member of the TGF-beta family may be introduced into cells. Representative examples of suitable vectors include viral vectors such as herpes viral vectors (*e.g.*, U.S. Patent No. 5,288,641), adenoviral vectors (*e.g.*, WO 94/26914, WO 93/9191; Kolls et al., *PNAS 91*(1):215-219, 1994; Kass-Eisler et al., *PNAS 90*(24):11498-502, 1993; Guzman et al., *Circulation 88*(6):2838-48, 1993; Guzman et al., *Cir. Res. 73*(6):1202-1207, 1993; Zabner et al., *Cell 75*(2):207-216, 1993; Li et al., *Hum Gene Ther. 4*(4):403-409, 1993; Caillaud et al., *Eur. J. Neurosci. 5*(10:1287-1291, 1993; Vincent et al., *Nat. Genet. 5*(2):130-134, 1993; Jaffe et al., *Nat. Genet. 1*(5):372-378, 1992; and Levrero et al., *Gene 101*(2):195-202, 1991), adeno-associated viral vectors (WO 95/13365; Flotte et al., *PNAS 90*(22):10613-10617, 1993), baculovirus vectors, parvovirus vectors (Koering et al., *Hum. Gene Therap. 5*:457-463, 1994), pox virus vectors (Panicali and Paoletti, *PNAS 79*:4927-4931, 1982; and Ozaki et al., *Biochem. Biophys. Res. Comm. 193*(2):653-660, 1993), and retroviruses (*e.g.*, EP 0,415,731; WO 90/07936; WO 91/0285, WO 94/03622; WO 93/25698; WO 93/25234; U.S. Patent No. 5,219,740; WO 93/11230; WO 93/10218). Viral vectors may likewise be constructed which contain a mixture of different elements (*e.g.*, promoters, envelope sequences, and the like) from different viruses, or non-viral sources. Within various embodiments, either the viral vector itself, or a viral particle which contains the viral vector may be utilized in the methods and compositions described below.

Within other embodiments of the invention, nucleic acid molecules which encode a molecule which inhibits binding of the TGF-beta binding-protein to a member of the TGF-beta family may be administered by a variety of techniques, including, for example, administration of asialoosomucoid (ASOR) conjugated with poly-L-lysine DNA complexes (Cristano et al., *PNAS* 92122-92126, 1993), DNA linked to killed adenovirus (Curiel et al., *Hum. Gene Ther. 3*(2):147-154, 1992), cytofectin-mediated introduction (DMRIE-DOPE, Vical, California), direct DNA injection (Acsadi et al., *Nature 352*:815-818, 1991); DNA ligand (Wu et al., *J. of Biol. Chem. 264*:16985-16987, 1989); lipofection (Felgner et al., *Proc. Natl. Acad. Sci. USA 84*:7413-7417, 1989); liposomes (Pickering et al., *Circ. 89*(1):13-21, 1994; and Wang et al., *PNAS 84*:7851-7855, 1987); microprojectile bombardment (Williams et al., *PNAS 88*:2726-2730, 1991); and direct delivery of nucleic acids which encode the protein itself either alone (Vile and Hart, *Cancer Res. 53*: 3860-3864, 1993), or utilizing PEG-nucleic acid complexes. Representative

examples of molecules that may be expressed by the vectors of present invention include ribozymes and antisense molecules, each of which are discussed in more detail above.

Determination of increased bone mineral content may be determined directly through the use of X-rays (*e.g.*, Dual Energy X-ray Absorptometry or "DEXA"), or by inference through bone turnover markers (such as osteoblast specific alkaline phosphatase, osteocalcin, type 1 procollagen C' propeptide (PICP), and total alkaline phosphatase; see Comier, C., *Curr. Opin. in Rheu.* 7:243, 1995), or by markers of bone resorption (pyridinoline, deoxypryridinoline, N-telopeptide, urinary hydroxyproline, plasma tartrate-resistant acid phosphatases and galactosyl hydroxylysine; see Comier, *supra*). The amount of bone mass may also be calculated from body weights or by other methods known in the art (see Guinness-Hey, *Metab. Bone Dis. and Relat. Res.* 5:177-181, 1984).

As will be evident to one of skill in the art, the amount and frequency of administration will depend, of course, on such factors as the nature and severity of the indication being treated, the desired response, the condition of the patient, and so forth. Typically, the compositions may be administered by a variety of techniques, as noted above.

The following examples are offered by way of illustration and not by way of limitation.

EXAMPLES

EXAMPLE 1

SCLEROSTEOSIS MAPS TO THE LONG ARM OF HUMAN CHROMOSOME 17

Genetic mapping of the defect responsible for sclerosteosis in humans localized the gene responsible for this disorder to the region of human chromosome 17 that encodes a novel TGF-beta binding-protein family member. In sclerosteosis, skeletal bone displays a substantial increase in mineral density relative to that of unafflicted individuals. Bone in the head displays overgrowth as well. Sclerosteosis patients are generally healthy although they may exhibit variable degrees of syndactyly at birth and variable degrees of cranial compression and nerve compression in the skull.

Linkage analysis of the gene defect associated with sclerosteosis was conducted by applying the homozygosity mapping method to DNA samples collected from 24 South African Afrikaaner families in which the disease occurred. (Sheffield et al., 1994, *Human Molecular Genetics* 3:1331-1335. "Identification of a Bardet-Biedl syndrome locus on chromosome 3 and evaluation of an efficient approach to homozygosity mapping"). The Afrikaaner population of South Africa is genetically homogeneous; the population is descended from a small number of founders who colonized the area several centuries ago, and it has been isolated by geographic

and social barriers since the founding. Sclerosteosis is rare everywhere in the world outside the Afrikaaner community, which suggests that a mutation in the gene was present in the founding population and has since increased in numbers along with the increase in the population. The use of homozygosity mapping is based on the assumption that DNA mapping markers adjacent to a recessive mutation are likely to be homozygous in affected individuals from consanguineous families and isolated populations.

A set of 371 microsatellite markers (Research Genetics, Set 6) from the autosomal chromosomes was selected to type pools of DNA from sclerosteosis patient samples. The DNA samples for this analysis came from 29 sclerosteosis patients in 24 families, 59 unaffected family members and a set of unrelated control individuals from the same population. The pools consisted of 4-6 individuals, either affected individuals, affected individuals from consanguineous families, parents and unaffected siblings, or unrelated controls. In the pools of unrelated individuals and in most of the pools with affected individuals or family members analysis of the markers showed several allele sizes for each marker. One marker, D17S1299, showed an indication of homozygosity: one band in several of the pools of affected individuals.

All 24 sclerosteosis families were typed with a total of 19 markers in the region of D17S1299 (at 17q12-q21). Affected individuals from every family were shown to be homozygous in this region, and 25 of the 29 individuals were homozygous for a core haplotype; they each had the same alleles between D17S1787 and D17S930. The other four individuals had one chromosome which matched this haplotype and a second which did not. In sum, the data compellingly suggested that this 3 megabase region contained the sclerosteosis mutation. Sequence analysis of most of the exons in this 3 megabase region identified a nonsense mutation in the novel TGF-beta binding-protein coding sequence (C>T mutation at position 117 of Sequence ID No. 1 results in a stop codon). This mutation was shown to be unique to sclerosteosis patients and carriers of Afrikaaner descent. The identity of the gene was further confirmed by identifying a mutation in its intron (A>T mutation at position +3 of the intron) which results in improper mRNA processing in a single, unrelated patient with diagnosed sclerosteosis.

EXAMPLE 2

TISSUE-SPECIFICITY OF TGF-BETA BINDING-PROTEIN GENE EXPRESSION

A. Human Beer Gene Expression by RT-PCR:

First-strand cDNA was prepared from the following total RNA samples using a commercially available kit ("Superscript Preamplification System for First-Strand cDNA

Synthesis", Life Technologies, Rockville, MD): human brain, human liver, human spleen, human thymus, human placenta, human skeletal muscle, human thyroid, human pituitary, human osteoblast (NHOst from Clonetics Corp., San Diego, CA), human osteosarcoma cell line (Saos-2, ATCC# HTB-85), human bone, human bone marrow, human cartilage, vervet monkey bone, saccharomyces cerevisiae, and human peripheral blood monocytes. All RNA samples were purchased from a commercial source (Clontech, Palo Alto, CA), except the following which were prepared in-house: human osteoblast, human osteosarcoma cell line, human bone, human cartilage and vervet monkey bone. These in-house RNA samples were prepared using a commercially available kit ("TRI Reagent", Molecular Research Center, Inc., Cincinnati, OH).

PCR was performed on these samples, and additionally on a human genomic sample as a control. The sense Beer oligonucleotide primer had the sequence 5'-CCGGAGCTGGAGAACAACAAG-3' (SEQ ID NO:19). The antisense Beer oligonucleotide primer had the sequence 5'-GCACTGGCCGGAGCACACC-3' (SEQ ID NO:20). In addition, PCR was performed using primers for the human beta-actin gene, as a control. The sense beta-actin oligonucleotide primer had the sequence 5'-AGGCCAACCGCGAGAAGATGA CC -3' (SEQ ID NO:21). The antisense beta-actin oligonucleotide primer had the sequence 5'-GAAGT CCAGGGCGACGTAGCA-3' (SEQ ID NO:22). PCR was performed using standard conditions in 25 ul reactions, with an annealing temperature of 61 degrees Celsius. Thirty-two cycles of PCR were performed with the Beer primers and twenty-four cycles were performed with the beta-actin primers.

Following amplification, 12 ul from each reaction were analyzed by agarose gel electrophoresis and ethidium bromide staining. See Figure 2A.

B. RNA In-situ Hybridization of Mouse Embryo Sections:

The full length mouse *Beer* cDNA (Sequence ID No. 11) was cloned into the pCR2.1 vector (Invitrogen, Carlsbad, CA) in the antisense and sense direction using the manufacturer's protocol. ^{35}S-alpha-GTP-labeled cRNA sense and antisense transcripts were synthesized using in-vitro transcription reagents supplied by Ambion, Inc (Austin, TX). In-situ hybridization was performed according to the protocols of Lyons et al. (*J. Cell Biol. 111*:2427-2436, 1990).

The mouse *Beer* cRNA probe detected a specific message expressed in the neural tube, limb buds, blood vessels and ossifying cartilages of developing mouse embryos. Panel A in Figure 3 shows expression in the apical ectodermal ridge (aer) of the limb (l) bud, blood vessels (bv) and the neural tube (nt). Panel B shows expression in the 4th ventricle of the brain (4). Panel C shows expression in the mandible (ma) cervical vertebrae (cv), occipital bone (oc),

palate (pa) and a blood vessel (bv). Panel D shows expression in the ribs (r) and a heart valve (va). Panel A is a transverse section of 10.5 dpc embryo. Panel B is a sagittal section of 12.5 dpc embryo and panels C and D are sagittal sections of 15.5 dpc embryos.

ba=branchial arch, h=heart, te=telencephalon (forebrain), b=brain, f=frontonasal mass, g=gut, h=heart, j=jaw, li=liver, lu=lung, ot=otic vesicle, ao=, sc=spinal cord, skm=skeletal muscle, ns=nasal sinus, th=thymus , to=tongue, fl=forelimb, di=diaphragm

EXAMPLE 3

EXPRESSION AND PURIFICATION OF RECOMBINANT BEER PROTEIN

A. Expression in COS-1 Cells:

The DNA sequence encoding the full length human Beer protein was amplified using the following PCR oligonucleotide primers: The 5' oligonucleotide primer had the sequence 5'-**AAGCTT**GGTACCATGCAGCTCCCAC-3' (SEQ ID NO:23) and contained a HindIII restriction enzyme site (in bold) followed by 19 nucleotides of the *Beer* gene starting 6 base pairs prior to the presumed amino terminal start codon (ATG). The 3' oligonucleotide primer had the sequence 5'-**AAGCTT**CTACTTGTCATCGTCGTCCTTGTAGTCGTAGGCGTTCTC CAGCT-3' (SEQ ID NO:24) and contained a HindIII restriction enzyme site (in bold) followed by a reverse complement stop codon (CTA) followed by the reverse complement of the FLAG epitope (underlined, Sigma-Aldrich Co., St. Louis, MO) flanked by the reverse complement of nucleotides coding for the carboxy terminal 5 amino acids of the Beer. The PCR product was TA cloned ("Original TA Cloning Kit", Invitrogen, Carlsbad, CA) and individual clones were screened by DNA sequencing. A sequence-verified clone was then digested by HindIII and purified on a 1.5% agarose gel using a commercially available reagents ("QIAquick Gel Extraction Kit", Qiagen Inc., Valencia, CA). This fragment was then ligated to HindIII digested, phosphatase-treated pcDNA3.1 (Invitrogen, Carlsbad, CA) plasmid with T4 DNA ligase. DH10B *E. coli* were transformed and plated on LB, 100 μg/ml ampicillin plates. Colonies bearing the desired recombinant in the proper orientation were identified by a PCR-based screen, using a 5' primer corresponding to the T7 promoter/priming site in pcDNA3.1 and a 3' primer with the sequence 5'- GCACTGGCCGGAGCACACC-3' (SEQ ID NO:25) that corresponds to the reverse complement of internal BEER sequence. The sequence of the cloned fragment was confirmed by DNA sequencing.

COS-1 cells (ATCC# CRL-1650) were used for transfection. 50 μg of the expression plasmid pcDNA-Beer-Flag was transfected using a commercially available kit following protocols supplied by the manufacturer ("DEAE-Dextran Transfection Kit", Sigma Chemical

Co., St. Louis, MO). The final media following transfection was DMEM (Life Technologies, Rockville, MD) containing 0.1% Fetal Bovine Serum. After 4 days in culture, the media was removed. Expression of recombinant BEER was analyzed by SDS-PAGE and Western Blot using anti-FLAG® M2 monoclonal antibody (Sigma-Aldrich Co., St. Louis, MO). Purification of recombinant BEER protein was performed using an anti-FLAG M2 affinity column ("Mammalian Transient Expression System", Sigma-Aldrich Co., St. Louis, MO). The column profile was analyzed via SDS-PAGE and Western Blot using anti-FLAG M2 monoclonal antibody.

B. Expression in SF9 insect cells:

The human *Beer* gene sequence was amplified using PCR with standard conditions and the following primers:

Sense primer: 5'-GTCGTCGGATCCATGGGGTGGCAGGCGTTCAAGAATGAT-3' (SEQ ID NO:26)

Antisense primer: 5'-GTCGTCAAGCTTCTACTTGTCATCGTCCTTGTAGTCGTA GGCGTTCTCCAGCTCGGC-3' (SEQ ID NO:27)

The resulting cDNA contained the coding region of Beer with two modifications. The N-terminal secretion signal was removed and a FLAG epitope tag (Sigma) was fused in frame to the C-terminal end of the insert. BamH1 and HindIII cloning sites were added and the gene was subcloned into pMelBac vector (Invitrogen) for transfer into a baculoviral expression vector using standard methods.

Recombinant baculoviruses expressing Beer protein were made using the Bac-N-Blue transfection kit (Invitrogen) and purified according to the manufacturers instructions.

SF9 cells (Invitrogen) were maintained in TNM_FH media (Invitrogen) containing 10% fetal calf serum. For protein expression, SF9 cultures in spinner flasks were infected at an MOI of greater than 10. Samples of the media and cells were taken daily for five days, and Beer expression monitored by western blot using an anti-FLAG M2 monoclonal antibody (Sigma) or an anti-Beer rabbit polyclonal antiserum.

After five days the baculovirus-infected SF9 cells were harvested by centrifugation and cell associated protein was extracted from the cell pellet using a high salt extraction buffer (1.5 M NaCl, 50 mM Tris pH 7.5). The extract (20 ml per 300 ml culture) was clarified by centrifugation, dialyzed three times against four liters of Tris buffered saline (150 mM NaCl, 50 mM Tris pH 7.5), and clarified by centrifugation again. This high salt fraction was applied to Hitrap Heparin (Pharmacia; 5 ml bed volume), washed extensively with HEPES buffered saline

(25 mM HEPES 7.5, 150 mM Nacl) and bound proteins were eluted with a gradient from 150 mM NaCl to 1200 mM NaCl. Beer elution was observed at aproximately 800 mM NaCl. Beer containing fractions were supplemented to 10% glycerol and 1 mM DTT and frozen at – 80 degrees C.

EXAMPLE 4

PREPARATION AND TESTING OF POLYCLONAL ANTIBODIES TO BEER, GREMLIN, AND DAN

A. Preparation of antigen:

The DNA sequences of Human *Beer*, Human *Gremlin*, and Human *Dan* were amplified using standard PCR methods with the following oligonucleotide primers:

H. Beer

Sense: 5' -GACTTGGATCCCAGGGGTGGCAGGCGTTC- 3' (SEQ ID NO:28)

Antisense 5' -AGCATAAGCTTCTAGTAGGCGTTCTCCAG- 3' (SEQ ID NO:29)

H. Gremlin

Sense: 5' -GACTTGGATCCGAAGGGAAAAAGAAAGGG- 3' (SEQ ID NO:30)

Antisense: 5' -AGCATAAGCTTTTAATCCAAATCGATGGA- 3' (SEQ ID NO:31)

H. Dan

Sense: 5' -ACTACGAGCTCGGCCCCACCACCCATCAACAAG- 3' (SEQ ID NO:32)

Antisense: 5' -ACTTAGAAGCTTTCAGTCCTCAGCCCCCTCTTCC-3' (SEQ ID NO:33)

In each case the listed primers amplified the entire coding region minus the secretion signal sequence. These include restriction sites for subcloning into the bacterial expression vector pQE-30 (Qiagen Inc., Valencia, CA) at sites BamHI/HindIII for Beer and Gremlin, and sites SacI/HindIII for Dan. pQE30 contains a coding sequence for a 6x His tag at the 5' end of the cloning region. The completed constructs were transformed into *E. coli* strain M-15/pRep (Qiagen Inc) and individual clones verified by sequencing. Protein expression in M-15/pRep and purification (6xHis affinity tag binding to Ni-NTA coupled to Sepharose) were performed as described by the manufacturer (Qiagen, The QIAexpressionist).

The *E. coli*-derived Beer protein was recovered in significant quantity using solubilization in 6M guanidine and dialyzed to 2-4M to prevent precipitation during storage. Gremlin and Dan protein were recovered in higher quantity with solubilization in 6M guanidine and a post purification guanidine concentration of 0.5M.

B. Production and testing of polyclonal antibodies:

Polyclonal antibodies to each of the three antigens were produced in rabbit and in

chicken hosts using standard protocols (R & R Antibody, Stanwood, WA; standard protocol for rabbit immunization and antisera recovery; Short Protocols in Molecular Biology. 2nd edition. 1992. 11.37-11.41. Contributors Helen M. Cooper and Yvonne Paterson; chicken antisera was generated with Strategic Biosolutions, Ramona, CA).

Rabbit antisera and chicken egg Igy fraction were screened for activity via Western blot. Each of the three antigens was separated by PAGE and transferred to 0.45um nitrocellulose (Novex, San Diego, CA). The membrane was cut into strips with each strip containing approximately 75 ng of antigen. The strips were blocked in 3% Blotting Grade Block (Bio-Rad Laboratories, Hercules, CA) and washed 3 times in 1X Tris buffer saline (TBS) /0.02% TWEEN buffer. The primary antibody (preimmunization bleeds, rabbit antisera or chicken egg IgY in dilutions ranging from 1:100 to 1:10,000 in blocking buffer) was incubated with the strips for one hour with gentle rocking. A second series of three washes 1X TBS/0.02%TWEEN was followed by an one hour incubation with the secondary antibody (peroxidase conjugated donkey anti-rabbit, Amersham Life Science, Piscataway, NJ; or peroxidase conjugated donkey anti-chicken, Jackson ImmunoResearch, West Grove, PA). A final cycle of 3X washes of 1X TBS/0.02%TWEEN was performed and the strips were developed with Lumi-Light Western Blotting Substrate (Roche Molecular Biochemicals, Mannheim, Germany).

C. Antibody cross-reactivity test:

Following the protocol described in the previous section, nitrocellulose strips of Beer, Gremlin or Dan were incubated with dilutions (1:5000 and 1:10,000) of their respective rabbit antisera or chicken egg IgY as well as to antisera or chicken egg Igy (dilutions 1:1000 and 1:5000) made to the remaining two antigens. The increased levels of nonmatching antibodies was performed to detect low affinity binding by those antibodies that may be seen only at increased concentration. The protocol and duration of development is the same for all three binding events using the protocol described above. There was no antigen cross-reactivity observed for any of the antigens tested.

EXAMPLE 5

INTERACTION OF BEER WITH TGF-BETA SUPER-FAMILY PROTEINS

The interaction of Beer with proteins from different phylogenetic arms of the TGF-β superfamily were studied using immunoprecipitation methods. Purified TGFβ–1, TGFβ–2, TGFβ–3, BMP-4, BMP-5, BMP-6 and GDNF were obtained from commerical sources (R&D systems; Minneapolis, MN). A representative protocol is as follows. Partially purified Beer

was dialyzed into HEPES buffered saline (25 mM HEPES 7.5, 150 mM NaCl). Immunoprecipitations were done in 300 ul of IP buffer (150 mM NaCl, 25 mM Tris pH 7.5, 1mM EDTA, 1.4 mM β-mercaptoethanol, 0.5 % triton X 100, and 10% glycerol). 30 ng recombinant human BMP-5 protein (R&D systems) was applied to 15 ul of FLAG affinity matrix (Sigma; St Louis MO)) in the presence and absence of 500 ng FLAG epitope-tagged Beer. The proteins were incubated for 4 hours @ 4°Cand then the affinity matrix-associated proteins were washed 5 times in IP buffer (1 ml per wash). The bound proteins were eluted from the affinity matrix in 60 microliters of 1X SDS PAGE sample buffer. The proteins were resolved by SDS PAGE and Beer associated BMP-5 was detected by western blot using anti-BMP-5 antiserum (Research Diagnostics, Inc) (see Figure 5).

BEER Ligand Binding Assay:

FLAG-Beer protein (20 ng) is added to 100 ul PBS/0.2% BSA and adsorbed into each well of 96 well microtiter plate previously coated with anti-FLAG monoclonal antibody (Sigma; St Louis MO) and blocked with 10% BSA in PBS. This is conducted at room temperature for 60 minutes. This protein solution is removed and the wells are washed to remove unbound protein. BMP-5 is added to each well in concentrations ranging from10 pM to 500 nM in PBS/0.2% BSA and incubated for 2 hours at room temperature. The binding solution is removed and the plate washed with three times with 200ul volumes of PBS/0.2% BSA. BMP-5 levels are then detected using BMP-5 anti-serum via ELISA (F.M. Ausubel et al (1998) Current Protocols in Mol Biol. Vol 2 11.2.1-11.2.22). Specific binding is calculated by subtracting non-specific binding from total binding and analyzed by the LIGAND program (Munson and Podbard, Anal. Biochem., 107, p220-239, (1980).

In a variation of this method, Beer is engineered and expressed as a human Fc fusion protein. Likewise the ligand BMP is engineered and expressed as mouse Fc fusion. These proteins are incubated together and the assay conducted as described by Mellor et al using homogeneous time resolved fluorescence detection (G.W. Mellor et al., *J of Biomol Screening,* 3(2) 91-99, 1998).

EXAMPLE 6

SCREENING ASSAY FOR INHIBITION OF TGF-BETA BINDING-PROTEIN BINDING TO TGF-BETA FAMILY MEMBERS

The assay described above is replicated with two exceptions. First, BMP concentration is held fixed at the Kd determined previously. Second, a collection of antagonist candidates is added at a fixed concentration (20 uM in the case of the small organic molecule collections and

1 uM in antibody studies). These candidate molecules (antagonists) of TGF-beta binding-protein binding include organic compounds derived from commercial or internal collections representing diverse chemical structures. These compounds are prepared as stock solutions in DMSO and are added to assay wells at $\leq$ 1% of final volume under the standard assay conditions. These are incubated for 2 hours at room temperature with the BMP and Beer, the solution removed and the bound BMP is quantitated as described. Agents that inhibit 40% of the BMP binding observed in the absence of compound or antibody are considered antagonists of this interaction. These are further evaluated as potential inhibitors based on titration studies to determine their inhibition constants and their influence on TGF-beta binding-protein binding affinity. Comparable specificity control assays may also be conducted to establish the selectivity profile for the identified antagonist through studies using assays dependent on the BMP ligand action (e.g. BMP/BMP receptor competition study).

EXAMPLE 7

INHIBITION OF TGF-BETA BINDING-PROTEIN LOCALIZATION TO BONE MATRIX

Evaluation of inhibition of localization to bone matrix (hydroxyapatite) is conducted using modifications to the method of Nicolas (Nicolas, V. *Calcif Tissue Int 57*:206, 1995). Briefly, ^{125}I-labelled TGF-beta binding-protein is prepared as described by Nicolas (*supra*). Hydroxyapatite is added to each well of a 96 well microtiter plate equipped with a polypropylene filtration membrane (Polyfiltroninc, Weymouth MA). TGF-beta binding-protein is added to 0.2% albumin in PBS buffer. The wells containing matrix are washed 3 times with this buffer. Adsorbed TGF-beta binding-protein is eluted using 0.3M NaOH and quantitated.

Inhibitor identification is conducted via incubation of TGF-beta binding- protein with test molecules and applying the mixture to the matrix as described above. The matrix is washed 3 times with 0.2% albumin in PBS buffer. Adsorbed TGF-beta binding-protein is eluted using 0.3 M NaOH and quantitated. Agents that inhibit 40% of the TGF-beta binding-protein binding observed in the absence of compound or antibody are considered bone localization inhibitors. These inhibitors are further characterized through dose response studies to determine their inhibition constants and their influence on TGF-beta binding-protein binding affinity.

EXAMPLE 8

CONSTRUCTION OF TGF-BETA BINDING-PROTEIN MUTANT

A. Mutagenesis:

A full-length TGF-beta binding-protein cDNA in pBluescript SK serves as a template for

mutagenesis. Briefly, appropriate primers (see the discussion provided above) are utilized to generate the DNA fragment by polymerase chain reaction using Vent DNA polymerase (New England Biolabs, Beverly, MA). The polymerase chain reaction is run for 23 cycles in buffers provided by the manufacturer using a 57°C annealing temperature. The product is then exposed to two restriction enzymes and after isolation using agarose gel electrophoresis, ligated back into pRBP4-503 from which the matching sequence has been removed by enzymatic digestion. Integrity of the mutant is verified by DNA sequencing.

B. Mammalian Cell Expression and Isolation of Mutant TGF-beta binding-protein:

The mutant TGF-beta binding-protein cDNAs are transferred into the pcDNA3.1 mammalian expression vector described in EXAMPLE 3. After verifying the sequence, the resultant constructs are transfected into COS-1 cells, and secreted protein is purified as described in EXAMPLE 3.

EXAMPLE 9

ANIMAL MODELS -I

GENERATION OF TRANSGENIC MICE OVEREXPRESSING THE *BEER* GENE

The ~200 kilobase (kb) BAC clone 15G5, isolated from the CITB mouse genomic DNA library (distributed by Research Genetics, Huntsville, AL) was used to determine the complete sequence of the mouse *Beer* gene and its 5' and 3' flanking regions. A 41 kb SalI fragment, containing the entire gene body, plus ~17 kb of 5' flanking and ~20 kb of 3' flanking sequence was sub-cloned into the BamHI site of the SuperCosI cosmid vector (Stratagene, La Jolla, CA) and propagated in the *E. coli* strain DH10B. From this cosmid construct, a 35 kb MluI - AviII restriction fragment (Sequence No. 6), including the entire mouse *Beer* gene, as well as 17 kb and 14 kb of 5' and 3' flanking sequence, respectively, was then gel purified, using conventional means, and used for microinjection of mouse zygotes (DNX Transgenics; US Patent No. 4,873,191). Founder animals in which the cloned DNA fragment was integrated randomly into the genome were obtained at a frequency of 5-30% of live-born pups. The presence of the transgene was ascertained by performing Southern blot analysis of genomic DNA extracted from a small amount of mouse tissue, such as the tip of a tail. DNA was extracted using the following protocol: tissue was digested overnight at 55°C in a lysis buffer containing 200 mM NaCl, 100 mM Tris pH8.5, 5 mM EDTA, 0.2% SDS and 0.5 mg/ml Proteinase K. The following day, the DNA was extracted once with phenol/chloroform (50:50), once with chloroform/isoamylalcohol (24:1) and precipitated with ethanol. Upon resuspension in TE (10mM Tris pH7.5, 1 mM EDTA) 8-10 ug of each DNA sample were digested with a restriction endonuclease, such as

EcoRI, subjected to gel electrophoresis and transferred to a charged nylon membrane, such as HyBondN+ (Amersham, Arlington Heights, IL). The resulting filter was then hybridized with a radioactively labelled fragment of DNA deriving from the mouse *Beer* gene locus, and able to recognize both a fragment from the endogenous gene locus and a fragment of a different size deriving from the transgene. Founder animals were bred to normal non-transgenic mice to generate sufficient numbers of transgenic and non-transgenic progeny in which to determine the effects of *Beer* gene overexpression. For these studies, animals at various ages (for example, 1 day, 3 weeks, 6 weeks, 4 months) are subjected to a number of different assays designed to ascertain gross skeletal formation, bone mineral density, bone mineral content, osteoclast and osteoblast activity, extent of endochondral ossification, cartilage formation, etc. The transcriptional activity from the transgene may be determined by extracting RNA from various tissues, and using an RT-PCR assay which takes advantage of single nucleotide polymorphisms between the mouse strain from which the transgene is derived (129Sv/J) and the strain of mice used for DNA microinjection [(C57BL5/J x SJL/J)F2].

ANIMAL MODELS – II

DISRUPTION OF THE MOUSE BEER GENE BY HOMOLOGOUS RECOMBINATION

Homologous recombination in embryonic stem (ES) cells can be used to inactivate the endogenous mouse *Beer* gene and subsequently generate animals carrying the loss-of-function mutation. A reporter gene, such as the *E. coli β-galactosidase* gene, was engineered into the targeting vector so that its expression is controlled by the endogenous *Beer* gene's promoter and translational initiation signal. In this way, the spatial and temporal patterns of *Beer* gene expression can be determined in animals carrying a targeted allele.

The targeting vector was constructed by first cloning the drug-selectable phosphoglycerate kinase (PGK) promoter driven *neomycin-resistance* gene (*neo*) cassette from pGT-N29 (New England Biolabs, Beverly, MA) into the cloning vector pSP72 (Promega, Madson, WI). PCR was used to flank the PGK*neo* cassette with bacteriophage P1 loxP sites, which are recognition sites for the P1 Cre recombinase (Hoess et al., PNAS USA, 79:3398, 1982). This allows subsequent removal of the neo-resistance marker in targeted ES cells or ES cell-derived animals (US Patent 4,959,317). The PCR primers were comprised of the 34 nucleotide (ntd) loxP sequence, 15-25 ntd complementary to the 5' and 3' ends of the PGKneo cassette, as well as restriction enzyme recognition sites (BamHI in the sense primer and EcoRI in the anti-sense primer) for cloning into pSP72. The sequence of the sense primer was 5'-AATCTGGATCCATAACTTCGTATAGCATACATTATACGAAGTTATCTGCAG

GATTCGAGGGCCCCT-3' (SEQ ID NO:34); sequence of the anti-sense primer was 5'-AATCTGAATTCCACCGGTGTTAATTAAATAACTTCGTATAATGTATGCTATACGA AGTTATAGATCTAGAG TCAGCTTCTGA-3' (SEQ ID NO:35).

The next step was to clone a 3.6 kb XhoI-HindIII fragment, containing the *E. coli β-galactosidase* gene and SV40 polyadenylation signal from pSVβ (Clontech, Palo Alto, CA) into the pSP72-PGKneo plasmid. The "short arm" of homology from the mouse *Beer* gene locus was generated by amplifying a 2.4 kb fragment from the BAC clone 15G5. The 3' end of the fragment coincided with the translational initiation site of the *Beer* gene, and the anti-sense primer used in the PCR also included 30 ntd complementary to the 5' end of the *β-galactosidase* gene so that its coding region could be fused to the Beer initiation site in-frame. The approach taken for introducing the "short arm" into the pSP72-βgal-PGKneo plasmid was to linearize the plasmid at a site upstream of the *β-gal* gene and then to co-transform this fragment with the "short arm" PCR product and to select for plasmids in which the PCR product was integrated by homologous recombination. The sense primer for the "short arm" amplification included 30 ntd complementary to the pSP72 vector to allow for this recombination event. The sequence of the sense primer was 5'-ATTTAGGTGACACTATAGAACTCGAGCAGCTGAAGCTTAAC CACATGGTGGCTCACAACCAT-3' (SEQ ID NO:36) and the sequence of the anti-sense primer was 5'-AACGACGGCCAGTGAATCCGTAATCATGGTCATGCTGCCAGGTGGAG GAGGGCA-3' (SEQ ID NO:37).

The "long arm" from the *Beer* gene locus was generated by amplifying a 6.1 kb fragment from BAC clone 15G5 with primers which also introduce the rare-cutting restriction enzyme sites SgrAI, FseI, AscI and PacI. Specifically, the sequence of the sense primer was 5'-ATTACCACCGGTGACACCCGCTTCCTGACAG-3' (SEQ ID NO:38); the sequence of the anti-sense primer was 5'-ATTACTTAATTAAACATGGCGCGCCATATGGCCGGCCCCT AATTGCGGCGCATCGTTAATT-3' (SEQ ID NO:39). The resulting PCR product was cloned into the TA vector (Invitrogen, Carlsbad, CA) as an intermediate step.

The mouse *Beer* gene targeting construct also included a second selectable marker, the *herpes simplex virus I thymidine kinase* gene (HSVTK) under the control of rous sarcoma virus long terminal repeat element (RSV LTR). Expression of this gene renders mammalian cells sensitive (and inviable) to gancyclovir; it is therefore a convenient way to select against neomycin-resistant cells in which the construct has integrated by a non-homologous event (US Patent 5,464,764). The RSVLTR-HSVTK cassette was amplified from pPS1337 using primers that allow subsequent cloning into the FseI and AscI sites of the "long arm"-TA vector plasmid.

For this PCR, the sequence of the sense primer was 5'-ATTACGGCCGGCCGCAAA GGAATTCAAGA TCTGA-3' (SEQ ID NO:40); the sequence of the anti-sense primer was 5'-ATTACGGCGCGCCCCTCACAGGCCGCACCCAGCT-3' (SEQ ID NO:41).

The final step in the construction of the targeting vector involved cloning the 8.8 kb SgrAI-AscI fragment containing the "long arm" and RSVLTR-HSVTK gene into the SgrAI and AscI sites of the pSP72-"short arm"-βgal-PGKneo plasmid. This targeting vector was linearized by digestion with either AscI or PacI before electroporation into ES cells.

EXAMPLE 10

ANTISENSE-MEDIATED BEER INACTIVATION

17-nucleotide antisense oligonucleotides are prepared in an overlapping format, in such a way that the 5' end of the first oligonucleotide overlaps the translation initiating AUG of the Beer transcript, and the 5' ends of successive oligonucleotides occur in 5 nucleotide increments moving in the 5' direction (up to 50 nucleotides away), relative to the Beer AUG. Corresponding control oligonucleotides are designed and prepared using equivalent base composition but redistributed in sequence to inhibit any significant hybridization to the coding mRNA. Reagent delivery to the test cellular system is conducted through cationic lipid delivery (P.L. Felgner, *Proc. Natl. Acad. Sci. USA 84*:7413, 1987). 2 ug of antisense oligonucleotide is added to 100 ul of reduced serum media (Opti-MEM I reduced serum media; Life Technologies, Gaithersburg MD) and this is mixed with Lipofectin reagent (6 ul) (Life Technologies, Gaithersburg MD) in the 100 ul of reduced serum media. These are mixed, allowed to complex for 30 minutes at room temperature and the mixture is added to previously seeded MC3T3E21 or KS483 cells. These cells are cultured and the mRNA recovered. Beer mRNA is monitored using RT-PCR in conjunction with Beer specific primers. In addition, separate experimental wells are collected and protein levels characterized through western blot methods described in Example 4. The cells are harvested, resuspended in lysis buffer (50 mM Tris pH 7.5, 20 mM NaCl, 1mM EDTA, 1% SDS) and the soluble protein collected. This material is applied to 10-20 % gradient denaturing SDS PAGE. The separated proteins are transferred to nitrocellulose and the western blot conducted as above using the antibody reagents described. In parallel, the control oligonucleotides are added to identical cultures and experimental operations are repeated. Decrease in Beer mRNA or protein levels are considered significant if the treatment with the antisense oligonucleotide results in a 50% change in either instance compared to the control scrambled oligonucleotide. This methodology enables selective gene inactivation and subsequent phenotype characterization of the mineralized nodules in the tissue culture model.

EXAMPLE 11

MODELING OF SCLEROSTIN CORE REGION

Homology recognition techniques (*e.g.*, PSI-BLAST (Altschul et al., *Nucleic Acids Res.* 25:3389-402 (1997)), FUGUE (Shi et al., *J. Mol. Biol.* 310:243-57 (2001)) suggested that the core region of SOST (SOST_Core) adopts a cystine-knot fold. FUGUE is a sensitive method for detecting homology between sequences and structures. Human Chorionic Gonadotropin β (hCG-β), for which an experimentally determined 3D structure is known, was identified by FUGUE (Shi et al., *supra*) as the closest homologue of SOST_Core. Therefore, hCG-β was used as the structural template to build 3D models for SOST_Core.

An alignment of SOST_Core and its close homologues is shown in Figure 7. Among the homologues shown in the alignment, only hCG-β (CGHB) had known 3D structure. The sequence identity between SOST_Core and hCG-β was approximately 25%. Eight CYS residues were conserved throughout the family, emphasizing the overall structural similarity between SOST_Core and hCG-β. Three pairs of cystines (1-5, 3-7, 4-8) formed disulfide bonds (shown with solid lines in Figure 7) in a "knot" configuration, which was characteristic to the cystine-knot fold. An extra disulfide bond (2-6), shown as a dotted line in Figure 7, was unique to this family and distinguished the family of proteins from other cystine-knot families (*e.g.*, TGF-β, BMP).

SOST_Core was modeled using PDB (Berman et al., *Acta Crystallogr. D. Biol. Crystallogr.* 58(Pt 6 Pt1):899-907 (2002)) entry 1HCN, the 3D structure of hCG-β (Wu et al., *Structure* 2:545-58 (1994)), as the structural template. Models were calculated with MODELER (Sali & Blundell, *J. Mol. Biol.* 234:779-815 (1993)). A snapshot of the best model is shown in Figure 8.

Most of the cystine-knot proteins form dimers because of the lack of hydrophobic core in a monomer (Scheufler et al., *supra*; Schlunegger and Grutter, *J. Mol. Biol.* 231:445-58 (1993)); Wu et al., *supra*). SOST likely follows the same rule and forms a homodimer to increase its stability. Constructing a model for the dimerized SOST_Core region presented several challenges because (1) the sequence similarity between SOST_Core and hCG-β was low (25%); (2) instead of a homodimer, hCG-β formed a heterodimer with hCG-α; and (3) a number of different relative conformations of monomers have been observed in dimerized cystine-knot proteins from different families (*e.g.*, PDGF, TGF-β, Neurotrophin, IL-17F, Gonadotropin), which suggested that the dimer conformation of SOST could deviate significantly from the hCG-α/β heterodimer conformation. In constructing the model, hCG-α was replaced with hCG-β

from the heterodimer structure (1HCN) using structure superimposition techniques combined with manual adjustment, and then a SOST_Core homodimer model was built according to the pseudo hCG-β homodimer structure. The final model is shown in Figure 9.

EXAMPLE 12

MODELING SOST-BMP INTERACTION

This example describes protein modeling of type I and type II receptor binding sites on BMP that are involved with interaction between BMP and SOST.

Competition studies demonstrated that SOST competed with both type I and type II receptors for binding to BMP. In an ELISA-based competition assay, BMP-6 selectively interacted with the sclerostin-coated surface (300 ng/well) with high affinity (K_D = 3.4 nM). Increasing amounts of BMP receptor IA (FC fusion construct) competed with sclerostin for binding to BMP-6 (11 nM) (IC_{50} = 114nM). A 10-fold molar excess of the BMP receptor was sufficient to reduce binding of sclerostin to BMP-6 by approximately 50%. This competition was also observed with a BMP receptor II-FC fusion protein (IC_{50} = 36nM) and DAN (IC_{50} = 43nM). Specificity of the assay was shown by lack of competition for binding to BMP-6 between sclerostin and a rActivin R1B-FC fusion protein, a TGF-β receptor family member that did not bind BMP.

The type I and type II receptor binding sites on a BMP polypeptide have been mapped and were spatially separated (Scheufler et al., *supra*; Innis et al., *supra*; Nickel et al., *supra*; Hart et al. *supra*). Noggin, another BMP antagonist that binds to BMP with high affinity, contacts BMP at both type I and type II receptor binding sites via the N-terminal portion of Noggin (Groppe et al., *supra*). The two β-strands in the core region near the C-terminal also contact BMP at the type II receptor binding site.

A manually tuned alignment of Noggin and SOST indicated that the two polypeptides shared sequence similarity between the N-terminal portions of the proteins and between the core regions. An amino acid sequence alignment is presented in Figure 10. The cysteine residues that form the characteristic cys-knot were conserved between Noggin and SOST. The overall sequence identity was 24%, and the sequence identity within the N-terminal binding region (alignment positions 1-45) was 33%. Two residues in the Noggin N-terminal binding region, namely Leu (L) at alignment position 21 and Glu (E) at position 23, were reported to play important roles in BMP binding (Groppe et al., *supra*). Both residues were conserved in SOST as well. The sequence similarity within the core region (alignment positions 131-228) was about

20%, but the cys-knot scaffold was maintained and a sufficient number of key residues was conserved, supporting homology between Noggin and SOST.

The Noggin structure was compared to SOST also to understand how two SOST monomers dimerize. As shown in Figure 11, the Noggin structure suggested that the linker between the N-terminal region and the core region not only played a role in connecting the two regions, but also formed part of the dimerization interface between two Noggin monomers. One major difference between Noggin and SOST was that the linker between the N-terminal region and the core region was much shorter in SOST.

The C-terminal region of SOST may play a role in SOST dimerization. The sequence of Noggin ended with the core region, while SOST had an extra C-terminal region. In the Noggin structure a disulfide bond connected the C-termini of two Noggin monomers. Thus, the C-terminal region of SOST started close to the interface of two monomers and could contribute to dimerization. In addition, secondary structure prediction showed that some portions of the C-terminal region of SOST had a tendency to form helices. This region in SOST may be responsible for the dimerization activity, possibly through helix-helix packing, which mimicked the function of the longer linker in Noggin. Another difference between the structure of Noggin and SOST was the amino acid insertion in the SOST core region at alignment positions 169-185 (see Figure 10). This insertion extended a β-hairpin, which pointed towards the dimerization interface in the Noggin structure (shown in Figure 11 as a loop region in the middle of the monomers and above the C-terminal Cys residue). This elongated β-hairpin could also contribute to SOST dimerization.

EXAMPLE 13

DESIGN AND PREPARATION OF SOST PEPTIDE IMMUNOGENS

This Example describes the design of SOST peptide immunogens that are used for immunizing animals and generating antibodies that block interactions between BMP and SOST and prevent dimer formation of SOST monomers.

BMP Binding Fragments

The overall similarity between SOST and Noggin and the similarity between the N-terminal regions of the two polypeptides suggest that SOST may interact with BMP in a similar manner to Noggin. That is, the N-terminal region of SOST may interact with both the type I and type II receptor binding sites on BMP, and a portion of the core region (amino acid alignment positions 190-220 in Figure 10) may interact with the type II receptor binding site such that

antibodies specific for these SOST regions may block or impair binding of BMP to SOST.

The amino acid sequences of these SOST polypeptide fragments for rat and human SOST are provided as follows.

SOST_N_Linker: The N-terminal region (includes the short linker that connects to the core region)

Human: QGWQAFKNDATEIIPELGEYPEPPPELENNKTMNRAE NGGRPPHHPFETKDVSEYS (SEQ ID NO:92)

Rat: QGWQAFKNDATEIIPGLREYPEPPQELENNQTMNRAEN GGRPPHHPYDTKDVSEYS (SEQ ID NO:93)

SOST_Core_Bind: Portion of the core region that is likely to contact BMP at its type II receptor binding site (extended slightly at both termini to include the CYS residue anchors):

Human: CIPDRYRAQRVQLLCPGGEAPRARKVRLVASC (SEQ ID NO:94)

Rat: CIPDRYRAQRVQLLCPGGAAPRSRKVRLVASC (SEQ ID NO:95)

SOST Dimerization Fragments

The C-terminal region of SOST is likely to be involved in the formation of SOST homodimers (see Example 12). The elongated β-hairpin may also play a role in homodimer formation. Antibodies that specifically bind to such regions may prevent or impair dimerization of SOST monomers, which may in turn interfere with interaction between SOST and BMP. Polypeptide fragments in rat and human SOST corresponding to these regions are as follows.

SOST_C: the C-terminal region

Human: LTRFHNQSELKDFGTEAARPQKGRKPRPRARSAKANQA ELENAY (SEQ ID NO:96)

Rat: LTRFHNQSELKDFGPETARPQKGRKPRPRARGAKANQAE LENAY (SEQ ID NO:97)

SOST_Core_Dimer: Portion of the core region that is likely involved in SOST dimerization (extended slightly at both termini to include the Cys residue anchors):

Human: CGPARLLPNAIGRGKWWRPSGPDFRC (SEQ ID NO:98)

Rat: CGPARLLPNAIGRVKWWRPNGPDFRC (SEQ ID NO:99)

BMP Binding Fragment at SOST N-terminus

The key N-terminal binding region of SOST (alignment positions 1-35 in Figure 10) was modeled on the basis of the Noggin/BMP-7 complex structure (Protein Data Bank Entry No: 1M4U) and the amino acid sequence alignment (see Figure 10) to identify amino acid residues of the SOST N-terminus that likely interact with BMP. The model of SOST is presented in Figure 12. In the comparative model, phenylalanine (Phe, F) at alignment position 8 (see arrow and accompanying text) in the SOST sequence projects into a hydrophobic pocket on the surface of the BMP dimer. The same "knob-into-hole" feature has been observed in the BMP and type I receptor complex structure (Nickel et al., *supra*), where Phe85 of the receptor fits into the same pocket, which is a key feature in ligand-type I receptor recognition for TGF-β superfamily members (including, for example, TGF-β family, BMP family, and the like). According to the model, a proline (Pro) directed turn is also conserved, which allows the N-terminal binding fragment to thread along the BMP dimer surface, traveling from type I receptor binding site to type II receptor binding site on the other side of the complex. Also conserved is another Pro-directed turn near the carboxy end of the binding fragment, which then connects to the linker region. Extensive contacts between SOST and BMP are evident in Figure 12.

Peptide Immunogens

Peptides were designed to encompass the SOST N-terminal region predicted to make contact with BMP proteins. The peptide sequences are presented below. For immunizing animals, the peptide sequences were designed to overlap, and an additional cysteine was added to the C-terminal end to facilitate crosslinking to KLH. The peptides were then used for immunization. The peptide sequences of the immunogens are as follows.

Human SOST:

QGWQAFKNDATEIIPELGEY (SEQ ID NO:47)

TEIIPELGEYPEPPPELENN (SEQ ID NO:48)

PEPPPELENNKTMNRAENGG (SEQ ID NO:49)

KTMNRAENGGRPPHHPFETK (SEQ ID NO:50)

RPPHHPFETKDVSEYS (SEQ ID NO:51)

Human SOST Peptides with Additional Cys:

QGWQAFKNDATEIIPELGEY-C (SEQ ID NO:52)

TEIIPELGEYPEPPPELENN-C (SEQ ID NO:53)

PEPPPELENNKTMNRAENGG-C (SEQ ID NO:54)

KTMNRAENGGRPPHHPFETK-C (SEQ ID NO:55)

RPPHHPFETKDVSEYS-C (SEQ ID NO:56)

Rat SOST:

QGWQAFKNDATEIIPGLREYPEPP (SEQ ID NO:57)

PEPPQELENNQTMNRAENGG (SEQ ID NO:58)

ENGGRPPHHPYDTKDVSEYS (SEQ ID NO:59)

TEIIPGLREYPEPPQELENN (SEQ ID NO:60)

Rat SOST Peptides with Additional Cys:

QGWQAFKNDATEIIPGLREYPEPP-C (SEQ ID NO:61)

PEPPQELENNQTMNRAENGG-C (SEQ ID NO:62)

ENGGRPPHHPYDTKDVSEYS-C (SEQ ID NO:63)

TEIIPGLREYPEPPQELENN-C (SEQ ID NO:64)

The following peptides were designed to contain the amino acid portion of core region that was predicted to make contact with BMP proteins. Cysteine was added at the C-terminal end of each peptide for conjugation to KLH, and the conjugated peptides were used for immunization. In the Docking Core N-terminal Peptide an internal cysteine was changed to a serine to avoid double conjugation to KLH.

For Human SOST:

Amino acid sequence without Cys residues added:

Docking_Core_N-terminal_Peptide: IPDRYRAQRVQLLCPGGEAP (SEQ ID NO:66)

Docking_Core_Cterm_Peptide: QLLCPGGEAPRARKVRLVAS (SEQ ID NO:67)

Docking_Core_N-terminal_Peptide: IPDRYRAQRVQLLCPGGEAP-C (SEQ ID NO:68)

Docking_Core_Cterm_Peptide: QLLCPGGEAPRARKVRLVAS-C (SEQ ID NO:69)

For Rat SOST:

Amino acid sequence without Cys residues added or substituted:

Docking_Core_N-terminal_Peptide: IPDRYRAQRVQLLSPGG (SEQ ID NO:70)

Docking_Core_Cterm_Peptide: PGGAAPRSRKVRLVAS (SEQ ID NO:71)

Peptide immunogens with Cys added and substituted:

Docking_Core_N-terminal_Peptide: IPDRYRAQRVQLLSPGG-C (SEQ ID NO:72)

Docking_Core_Cterm_Peptide: PGGAAPRSRKVRLVAS-C (SEQ ID NO:73)

Two regions within SOST that potentially interact to form SOST homodimers include the amino acids with the SOST core region that are not present in Noggin. Human SOST peptides designed to contain this sequence had a C-terminal or N-terminal Cys that was conjugated to KLH. For the rat SOST peptide, a cysteine was added to the carboxy terminus of the sequence (SEQ ID NO:76). The KLH conjugated peptides were used for immunization.

For Human SOST:

CGPARLLPNAIGRGKWWRPS (SEQ ID NO:74)

IGRGKWWRPSGPDFRC (SEQ ID NO:75)

For Rat SOST:

PNAIGRVKWWRPNGPDFR (SEQ ID NO:76)

Rat SOST peptide with cysteine added

PNAIGRVKWWRPNGPDFR-C (SEQ ID NO:77)

The second region within SOST that potentially interacts to form SOST homodimers includes the C-terminal region. Peptide immunogens were designed to include amino acid sequences within this region (see below). For conjugation to KLH, a cysteine residue was added to the C-terminal end, and the conjugated peptides were used for immunization.

For Human SOST:

KRLTRFHNQS ELKDFGTEAA (SEQ ID NO:78)

ELKDFGTEAA RPQKGRKPRP (SEQ ID NO:79)

RPQKGRKPRP RARSAKANQA (SEQ ID NO:80)

RARSAKANQA ELENAY (SEQ ID NO:81)

Peptide immunogens with Cys added at C-terminus:

KRLTRFHNQS ELKDFGTEAA-C (SEQ ID NO:82)

ELKDFGTEAA RPQKGRKPRP-C (SEQ ID NO:83)

RPQKGRKPRP RARSAKANQA-C (SEQ ID NO:84)

RARSAKANQA ELENAY-C (SEQ ID NO:85)

For Rat SOST:

KRLTRFHNQSELKDFGPETARPQ (SEQ ID NO:86)

KGRKPRPRARGAKANQAELENAY (SEQ ID NO:87)

SELKDFGPETARPQKGRKPRPRAR (SEQ ID NO:88)

Peptide immunogens with Cys added at C-terminus:

KRLTRFHNQSELKDFGPETARPQ-C (SEQ ID NO:89)

KGRKPRPRARGAKANQAELENAY-C (SEQ ID NO:90)

SELKDFGPETARPQKGRKPRPRAR-C (SEQ ID NO:91)

EXAMPLE 14

ASSAY FOR DETECTING BINDING OF ANTIBODIES TO A TGF-BETA BINDING-PROTEIN

This example describes an assay for detecting binding of a ligand, for example, an antibody or antibody fragment thereof, to sclerostin.

A FLAG®-sclerostin fusion protein was prepared according to protocols provided by the manufacturer (Sigma Aldrich, St. Louis, MO) and as described in U.S. Patent No. 6,395,511. Each well of a 96 well microtiter plate is coated with anti-FLAG® monoclonal antibody (Sigma Aldrich) and then blocked with 10% BSA in PBS. The fusion protein (20 ng) is added to 100 µl PBS/0.2% BSA and adsorbed onto the 96-well plate for 60 minutes at room temperature. This protein solution is removed and the wells are washed to remove unbound fusion protein. A BMP, for example, BMP-4, BMP-5, BMP-6, or BMP-7, is diluted in PBS/0.2% BSA and added to each well at concentrations ranging from 10 pM to 500 nM. After an incubation for 2 hours at room temperature, the binding solution is removed and the plate is washed three times with 200 µl volumes of PBS/0.2% BSA. Binding of the BMP to sclerostin is detected using polyclonal antiserum or monoclonal antibody specific for the BMP and an appropriate enzyme-conjugated second step reagent according to standard ELISA techniques (*see, e.g.,* Ausubel et al., *Current Protocols in Mol Biol.* Vol 2 11.2.1-11.2.22 (1998)). Specific binding is calculated by subtracting non-specific binding from total binding and analyzed using the LIGAND program (Munson and Podbard, *Anal. Biochem.* 107:220-39 (1980)).

Binding of sclerostin to a BMP is also detected by homogeneous time resolved fluorescence detection (Mellor et al., *J Biomol. Screening,* 3:91-99 (1998)). A polynucleotide

sequence encoding sclerostin is operatively linked to a human immunoglobulin constant region in a recombinant nucleic acid construct and expressed as a human Fc-sclerostin fusion protein according to methods known in the art and described herein. Similarly, a BMP ligand is engineered and expressed as a BMP-mouse Fc fusion protein. These two fusion proteins are incubated together and the assay conducted as described by Mellor et al.

EXAMPLE 15

SCREENING ASSAY FOR ANTIBODIES THAT INHIBIT BINDING OF TGF-BETA FAMILY MEMBERS TO TGF-BETA BINDING PROTEIN

This example describes a method for detecting an antibody that inhibits binding of a TGF-beta family member to sclerostin. An ELISA is performed essentially as described in Example 14 except that the BMP concentration is held fixed at its Kd (determined, for example, by BIAcore analysis). In addition, an antibody or a library or collection of antibodies is added to the wells to a concentration of 1 μM. Antibodies are incubated for 2 hours at room temperature with the BMP and sclerostin, the solution removed, and the bound BMP is quantified as described (see Example 14). Antibodies that inhibit 40% of the BMP binding observed in the absence of antibody are considered antagonists of this interaction. These antibodies are further evaluated as potential inhibitors by performing titration studies to determine their inhibition constants and their effect on TGF-beta binding-protein binding affinity. Comparable specificity control assays may also be conducted to establish the selectivity profile for the identified antagonist using assays dependent on the BMP ligand action (*e.g.*, a BMP/BMP receptor competition study).

EXAMPLE 16

INHIBITION OF TGF-BETA BINDING-PROTEIN LOCALIZATION TO BONE MATRIX

Evaluation of inhibition of localization to bone matrix (hydroxyapatite) is conducted using modifications to the method of Nicolas (*Calcif. Tissue Int.* 57:206-12 (1995)). Briefly, ^{125}I-labelled TGF-beta binding-protein is prepared as described by Nicolas (*supra*). Hydroxyapatite is added to each well of a 96-well microtiter plate equipped with a polypropylene filtration membrane (Polyfiltroninc, Weymouth MA). TGF-beta binding-protein diluted in 0.2% albumin in PBS buffer is then added to the wells. The wells containing matrix are washed 3 times with 0.2% albumin in PBS buffer. Adsorbed TGF-beta binding-protein is eluted using 0.3 M NaOH and then quantified.

An antibody that inhibits or impairs binding of the sclerostin TGF-beta binding protein to

the hydroxyapatite is identified by incubating the TGF-beta binding protein with the antibody and applying the mixture to the matrix as described above. The matrix is washed 3 times with 0.2% albumin in PBS buffer. Adsorbed sclerostin is eluted with 0.3 M NaOH and then quantified. An antibody that inhibits the level of binding of sclerostin to the hydroxyapatite by at least 40% compared to the level of binding observed in the absence of antibody is considered a bone localization inhibitor. Such an antibody is further characterized in dose response studies to determine its inhibition constant and its effect on TGF-beta binding-protein binding affinity.

From the foregoing, although specific embodiments of the invention have been described herein for purposes of illustration, various modifications may be made without deviating from the spirit and scope of the invention. Accordingly, the invention is not limited except as by the appended claims.

CLAIMS

We claim the following:

1. An antibody, or an antigen-binding fragment thereof, that binds specifically to a sclerostin polypeptide, said sclerostin polypeptide comprising an amino acid sequence set forth in SEQ ID NO:2, 6, 8, 14, 46, or 65, wherein the antibody competitively inhibits binding of the sclerostin polypeptide to at least one of (i) a bone morphogenic protein (BMP) Type I Receptor binding site and (ii) a BMP Type II Receptor binding site, wherein the BMP Type I Receptor binding site is capable of binding to a BMP Type I Receptor polypeptide comprising an amino acid sequence set forth in a sequence selected from the group consisting of GenBank Acc. Nos. NM_004329 (SEQ ID NO:102); D89675 (SEQ ID NO:103); NM_001203 (SEQ ID NO:104); S75359 (SEQ ID NO:105); NM_030849 (SEQ ID NO:106); D38082 (SEQ ID NO:107); NP_001194 (SEQ ID NO:108); BAA19765 (SEQ ID NO:109); and AAB33865 (SEQ ID NO:110) and wherein the BMP Type II Receptor binding site is capable of binding to a BMP Type II Receptor polypeptide comprising the amino acid sequence set forth in a sequence selected from the group consisting of GenBank Acc. NOs. U25110 (SEQ ID NO:111); NM_033346 (SEQ ID NO:112); Z48923 (SEQ ID NO:114); CAA88759 (SEQ ID NO:115); and NM_001204 (SEQ ID NO:113).

2. An antibody, or an antigen-binding fragment thereof, that binds specifically to a sclerostin polypeptide and that impairs formation of a sclerostin homodimer, wherein the sclerostin polypeptide comprises an amino acid sequence set forth in SEQ ID NOs: 2, 6, 8, 14, 46, or 65.

3. The antibody of either claim 1 or claim 2, wherein the antibody is a polyclonal antibody.

4. The antibody of either claim 1 or claim 2, wherein the antibody is a monoclonal antibody.

5. The antibody of claim 4 wherein the monoclonal antibody is selected from the group consisting of a mouse monoclonal antibody, a human monoclonal antibody, a rat monoclonal antibody, and a hamster monoclonal antibody.

6. A hybridoma cell producing the antibody of claim 4.

7. A host cell that is capable of expressing the antibody of claim 4.

8. The antibody of either claim 1 or claim 2, wherein the antibody is a humanized antibody or a chimeric antibody.

9. A host cell that is capable of expressing the antibody of claim 8.

10. The antibody of either claim 1 or claim 2, wherein the antigen-binding fragment is selected from the group consisting of $F(ab')_2$, Fab', Fab, Fd, and Fv.

11. The antibody of either claim 1 or claim 2 that comprises a single chain antibody.

12. A host cell that is capable of expressing the antibody of claim 11.

13. A composition comprising an antibody, or antigen-binding fragment thereof, according to either claim 1 or claim 2 and a physiologically acceptable carrier.

14. An immunogen comprising a peptide comprising at least 21 consecutive amino acids and no more than 50 consecutive amino acids of a SOST polypeptide, said SOST polypeptide comprising an amino acid sequence set forth in SEQ ID NOs: 2, 6, 8, 14, 46, or 65, wherein the peptide is capable of eliciting in a non-human animal an antibody that binds specifically to the SOST polypeptide and that competitively inhibits binding of the SOST polypeptide to at least one of (i) a bone morphogenic protein (BMP) Type I Receptor binding site and (ii) a BMP Type II Receptor binding site, wherein the BMP Type I Receptor binding site is capable of binding to a BMP Type I Receptor polypeptide comprising an amino acid sequence set forth in a sequence selected from the group consisting of GenBank Acc. Nos. NM_004329 (SEQ ID NO:102); D89675 (SEQ ID NO:103); NM_001203 (SEQ ID NO:104); S75359 (SEQ ID NO:105); NM_030849 (SEQ ID NO:106); D38082 (SEQ ID NO:107); NP_001194 (SEQ ID NO:108); BAA19765 (SEQ ID NO:109); and AAB33865 (SEQ ID NO:110) and wherein the BMP Type II Receptor binding site is capable of binding to a BMP Type II Receptor polypeptide comprising the amino acid sequence set forth in a sequence selected from the group consisting of GenBank Acc. NOs. U25110 (SEQ ID NO:111); NM_033346 (SEQ ID NO:112); Z48923 (SEQ ID NO:114); CAA88759 (SEQ ID NO:115); and NM_001204 (SEQ ID NO:113).

15. An immunogen comprising a peptide that comprises at least 21 consecutive amino acids

and no more than 50 consecutive amino acids of a SOST polypeptide, said SOST polypeptide comprising an amino acid sequence set forth in SEQ ID NO: 2, 6, 8, 14, 46, or 65, wherein the peptide is capable of eliciting in a non-human animal an antibody that binds specifically to the SOST polypeptide and that impairs formation of a SOST homodimer.

16. The immunogen of either claim 14 or claim 15 wherein the peptide is associated with a carrier molecule.

17. The immunogen of claim 16 wherein the carrier molecule is carrier polypeptide.

18. The immunogen of claim 17 wherein the carrier polypeptide is keyhole limpet hemocyanin.

19. A method for producing an antibody that specifically binds to a SOST polypeptide, comprising immunizing a non-human animal with an immunogen according to claim 14, wherein (a) the SOST polypeptide comprises an amino acid sequence set forth in SEQ ID NO: 2, 6, 8, 14, 46, or 65; (b) the antibody competitively inhibits binding of the SOST polypeptide to at least one of (i) a bone morphogenic protein (BMP) Type I Receptor binding site and (ii) a BMP Type II Receptor binding site; (c) the BMP Type I Receptor binding site is capable of binding to a BMP Type I Receptor polypeptide comprising the amino acid sequence set forth in a sequence selected from the group consisting of GenBank Acc. Nos. NM_004329 (SEQ ID NO:102); D89675 (SEQ ID NO:103); NM_001203 (SEQ ID NO:104); S75359 (SEQ ID NO:105); NM_030849 (SEQ ID NO:106); D38082 (SEQ ID NO:107); NP_001194 (SEQ ID NO:108); BAA19765 (SEQ ID NO:109); and AAB33865 (SEQ ID NO:110); and (d) the BMP Type II Receptor binding site is capable of binding to a BMP Type II Receptor polypeptide comprising the amino acid sequence set forth in a sequence selected from the group consisting of GenBank Acc. NOs. U25110 (SEQ ID NO:111); NM_033346 (SEQ ID NO:112); Z48923 (SEQ ID NO:114); CAA88759 (SEQ ID NO:115); and NM_001204 (SEQ ID NO:113).

20. A method for producing an antibody that specifically binds to a SOST polypeptide, said SOST polypeptide comprising an amino acid sequence set forth in SEQ ID NO: 2, 6, 8, 14, 46, or 65, comprising immunizing a non-human animal with an immunogen according to claim 15, wherein the antibody impairs formation of a SOST homodimer.