

CAG, EF1 α , SV40, CMV, UBC, PGK, and CBA. In some instances, the regulatory element results in selective expression of the transgene in PV neurons at a level that is at least 1.5 fold, at least 2 fold, at least 3 fold, at least 4 fold, at least 5 fold, at least 6 fold, at least 7 fold, at least 8 fold, at least 9 fold, at least 10 fold, at least 15 fold, or at least 20 fold as compared to selective expression of the transgene in PV neurons when operably linked to a non-selective regulatory element, as measured by the co-localization assay. In some cases, the regulatory element results in selective expression in PV neurons that is at least 2%, at least 5%, at least 10%, at least 15%, at least 20%, at least 25%, at least 30%, at least 35%, at least 40%, at least 45%, at least 50%, at least 55%, at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, or at least 95% higher than expression in PV neurons when the transgene is operably linked to a non-selective regulatory element. In some cases, the regulatory element results in selective expression in PV neurons that is about 1.5 times, 2 times, 2.5 times, 3 times, 3.5 times, 4 times, 4.5 times, 5 times, 5.5 times, 6 times, 6.5 times, 7 times, 7.5 times, 8 times, 8.5 times, 9 times, 9.5 times, 10 times, 15 times, 20 times, 25 times, 30 times, 40 times, 50 times, or 100 times higher than expected for natural distribution of PV neurons in the CNS. In some cases, the co-localization assay is an immunohistochemical assay. In some cases, the immunohistochemical assay comprises an anti-PV antibody. In some cases, the co-localization assay is performed as shown in Example 5 below. In some cases, the transgene encodes an ion channel subunit, a neurotransmitter regulator, a DNA binding domain, a gene editing protein, or a variant or a functional fragment thereof. In some cases, the ion channel subunit is an alpha subunit or a beta subunit of a sodium ion channel or a subunit of a potassium ion channel. In some cases, the transgene comprises any one of (i) SEQ ID NOs: 37-43; (ii) a functional fragment thereof; or (iii) a sequence having at least 80% sequence identity to (i) or (ii). In some cases, sequence identity is determined using BLAST. In some cases, the transgene comprises (i) SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, or KV3.3; (ii) a functional fragment thereof; or (iii) a sequence having at least 80% sequence identity to (i) or (ii). In some cases, the transgene is a neurotransmitter regulator that comprises (i) STXBP1, (ii) a functional fragment thereof, or (iii) a sequence having at least 80% sequence identity to (i) or (ii). In some cases, the transgene comprises a DNA binding protein that modulates expression of an endogenous gene. In some cases, the endogenous gene is SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.2, KV3.3, or STXBP1. In some cases, the transgene comprises a DNA binding protein that comprises a DNA binding domain of a DNA binding protein or a DNA cleaving protein (e.g., a nuclease, a restriction enzyme, a recombinase, etc.) wherein the DNA cleaving

or a combination thereof, or (iii) a sequence with at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to (i) or (ii). In some cases, sequence identity is determined using BLAST. In some cases, the regulatory elements result in selective expression of the transgene in PV neurons that is greater than expression of the same transgene when operably linked to a non-selective regulatory element, as measured by a co-localization assay. In some cases, the immunohistochemical assay comprises an anti-PV antibody (e.g., as described in Example 5 below). In some cases, the non-selective regulatory element is a constitutive promoter. In some cases, the non-selective regulatory element is any one of CAG, EF1 α , SV40, CMV, UBC, PGK, and CBA. In some cases, the regulatory elements result in selective expression of the transgene in PV neurons at a level that is at least 1.5 fold, at least 2 fold, at least 3 fold, at least 4 fold, at least 5 fold, at least 6 fold, at least 7 fold, at least 8 fold, at least 9 fold, at least 10 fold, at least 15 fold, or at least 20 fold as compared to a non-selective regulatory element when operably linked to the transgene, as measured by a co-localization assay. In some cases, the regulatory elements result in selective expression in PV neurons that is at least 2%, at least 5%, at least 10%, at least 15%, at least 20%, at least 25%, at least 30%, at least 35%, at least 40%, at least 45%, at least 50%, at least 55%, at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, or at least 95% higher than expression in PV neurons when the transgene is operably linked to a non-selective regulatory element. In some cases, the regulatory elements result in selective expression in PV neurons that is about 1.5 times, 2 times, 2.5 times, 3 times, 3.5 times, 4 times, 4.5 times, 5 times, 5.5 times, 6 times, 6.5 times, 7 times, 7.5 times, 8 times, 8.5 times, 9 times, 9.5 times, 10 times, 15 times, 20 times, 25 times, 30 times, 40 times, 50 times, or 100 times higher than expected for natural distribution of PV neurons in CNS. In some cases, the transgene is any one of SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.3, STXBP1, a DNA binding protein, a gene editing protein, or a functional fragment thereof. In some cases, the regulatory elements and the transgene are in an AAV. In some cases, the AAV is AAV1, AAV8, AAV9, scAAV1, scAAV8, or scAAV9.

[0011] In another example, the present disclosure contemplates a method of treating a neurological condition or disorder in a subject in need thereof, the method comprising contacting a cell with a nucleic acid cassette comprising: one or more regulatory elements operably linked to a transgene that result in selective expression of the transgene in PV neurons over one or more non-PV cells in CNS. In some cases, each of the regulatory elements comprises (i) a sequence of SEQ ID NOs: 1-32, (ii) a functional fragment or a

combination thereof, or (iii) a sequence with at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to (i) or (ii). In some cases, sequence identity is determined using BLAST. In some cases, the transgene is a voltage-gated ion channel subunit, or a variant or a functional fragment thereof. In some cases, the subunit is a beta subunit of a sodium ion channel. In some cases, the subunit is an alpha subunit of a sodium ion channel. In some cases, the subunit is of a potassium ion channel. In some cases, the transgene is any one of (i) SCN1A, SCN1B, SCN2B, KV3.1, or KV3.3; (ii) a functional fragment thereof; or (iii) a sequence having at least 80% sequence identity to (i) or (ii). In some cases, the transgene is a DNA binding protein. In some cases, the DNA binding protein modulates an endogenous gene. In some cases, the endogenous gene is SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.3, or STXBP1. In some cases, the transgene is a DNA binding protein that comprises a DNA binding domain of a DNA binding protein or a DNA cleaving protein (e.g., a nuclease, a restriction enzyme, a recombinase, etc.) wherein the DNA cleaving domain or nuclease domain has been deactivated, e.g., a nuclease-deactivated Cas (dCas), a deactivated transcription activator-like effector nuclease, or a nuclease-deactivated zinc finger protein. In some cases, the transgene comprises a DNA binding domain linked to a transcriptional modulating domain (e.g., a transcriptional activator or repressor domain). In some cases, the transgene is a gene editing protein. In some cases, the gene editing protein is a Cas protein, e.g., Cas9. In some cases, the neurological condition or disorder is associated with a haploinsufficiency or a mutation in any of SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.3, or STXBP1. In some cases, the neurological condition or disorder is epilepsy, neurodegeneration, tauopathy, or neuronal hypoexcitability. In some cases, the neurological condition or disorder is Dravet syndrome. In some cases, the neurological condition or disorder is Alzheimer's disease. In some cases, the neurological condition or disease is a psychiatric disorder (e.g., schizophrenia, obsessive compulsive disorder, addiction, depression, anxiety, psychosis); an autism spectrum disorder (e.g., Fragile X syndrome, Rett syndrome); epilepsy (e.g., chronic traumatic encephalopathy, generalized epilepsy with febrile seizures plus (GEFS+), epileptic encephalopathy, temporal lobe epilepsy, focal epilepsy, tuberous sclerosis); or neurodegeneration (e.g., Alzheimer's disease, Parkinson's disease). In some cases, the neurological condition or disease is any seizure and/or epilepsy related condition or disease wherein PV neurons are implicated. In some cases, the regulatory elements of this disclosure result in selective expression of the transgene in PV neurons that is greater than expression of the same transgene when operably

linked to a non-selective regulatory element, as measured by a co-localization assay. In some cases, the non-selective regulatory element is a constitutive promoter. In some cases, the non-selective regulatory element is any one of CAG, EF1 α , SV40, CMV, UBC, PGK, and CBA. In some cases, the regulatory elements result in selective expression in PV neurons at a level that is at least 1.5 fold, at least 2 fold, at least 3 fold, at least 4 fold, at least 5 fold, at least 6 fold, at least 7 fold, at least 8 fold, at least 9 fold, at least 10 fold, at least 15 fold, or at least 20 fold as compared to a non-selective regulatory element when operably linked to the transgene, as measured by a co-localization assay. In some cases, the regulatory elements result in selective expression in PV neurons that is at least 2%, at least 5%, at least 10%, at least 15%, at least 20%, at least 25%, at least 30%, at least 35%, at least 40%, at least 45%, at least 50%, at least 55%, at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, or at least 95% higher than expression in PV neurons when the transgene is operably linked to a non-selective regulatory element. In some cases, the regulatory elements result in selective expression in PV neurons that is about 1.5 times, 2 times, 2.5 times, 3 times, 3.5 times, 4 times, 4.5 times, 5 times, 5.5 times, 6 times, 6.5 times, 7 times, 7.5 times, 8 times, 8.5 times, 9 times, 9.5 times, 10 times, 15 times, 20 times, 25 times, 30 times, 40 times, 50 times, or 100 times higher than expected for natural distribution of PV neurons in CNS. In some cases, the nucleic acid cassette is in an AAV. In some cases, the AAV is AAV1, AAV8, AAV9, scAAV1, scAAV8, or scAAV9.

[0012] In one example, the present disclosure provides a method of treating Dravet syndrome, comprising contacting a cell with an AAV comprising a transgene, wherein the transgene is any one of (i) SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, or a DNA binding protein, (ii) a functional fragment thereof, or (iii) a sequence having at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to (i) or (ii). In some cases, sequence identity is measured using BLAST. In some cases, the DNA binding protein modulates an endogenous gene. In some cases, the DNA binding protein is a transcriptional modulator. In some cases, the transgene is a DNA binding protein that comprises a DNA binding domain of a DNA binding protein or a DNA cleaving protein (e.g., a nuclease, a restriction enzyme, a recombinase, etc.) wherein the DNA cleaving domain or nuclease domain has been deactivated, e.g., a nuclease-deactivated Cas (dCas), a deactivated transcription activator-like effector nuclease, or a nuclease-deactivated zinc finger protein. In some cases, the DNA binding domain is linked to a transcriptional modulating domain (e.g., a transcriptional activator or repressor domain). In some cases, the transgene comprises a gene editing protein,

e.g., a Cas protein, Cas9. In some cases, the endogenous gene is SCN1A, SCN2A, SCN8A, SCN1B, or SCN2B. In some cases, the AAV further comprises one or more PV neuron selective regulatory elements or one or more regulatory elements disclosed herein operably linked to the transgene. In some cases, each of the regulatory elements independently comprises (i) a sequence of SEQ ID NOs: 1-32, (ii) a functional fragment or a combination thereof, or (iii) a sequence with at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to (i) or (ii).

[0013] In another example, the present disclosure provides a method of treating Alzheimer's disease, comprising contacting a cell with an AAV comprising a transgene, wherein the transgene is any one of (i) SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.3, STXBP1, or a DNA binding protein; (ii) a functional fragment thereof; or (iii) a sequence having at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to (i) or (ii). In some cases, sequence identity is measured using BLAST. In some cases, the DNA binding protein modulates an endogenous gene. In some cases, the endogenous gene is SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.3, or STXBP1. In some cases, the transgene is a DNA binding protein comprising a transcriptional modulator. In some cases, the transgene is a DNA binding protein that comprises a DNA binding domain of a DNA binding protein or a DNA cleaving protein (e.g., a nuclease, a restriction enzyme, a recombinase, etc.) wherein the DNA cleaving domain or nuclease domain has been deactivated, e.g., a nuclease-deactivated Cas (dCas), a deactivated transcription activator-like effector nuclease, or a nuclease-deactivated zinc finger protein. In some cases, the DNA binding domain is linked to a transcriptional modulating domain (e.g., a transcriptional activator or repressor domain). In some cases, the transgene comprises a gene editing protein, e.g., a Cas protein, Cas9. In some cases, the AAV further comprises one or more PV neuron selective regulatory elements or one or more regulatory elements disclosed herein operably linked to the transgene. In some cases, each of the regulatory elements independently comprises each of the regulatory elements independently comprises (i) a sequence of SEQ ID NOs: 1-32, (ii) a functional fragment or a combination thereof, or (iii) a sequence with at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to (i) or (ii).

[0013a] In one aspect, the present disclosure provides a nucleic acid cassette comprising a parvalbumin (PV) neuron sensitive regulatory element operably linked to a heterologous transgene, wherein the regulatory element is less than 2.5 kb and comprises SEQ ID NO: 1 or SEQ ID NO: 8.

[0013b] In one aspect, the present disclosure provides a nucleic acid cassette comprising a parvalbumin (PV) neuron selective regulatory element of less than 2.5 kb operably linked to a heterologous transgene, wherein the regulatory element comprises a sequence with at least 95% sequence identity to SEQ ID NO: 1 or SEQ ID NO: 8, and wherein the PV neuron selective regulatory element increases selective expression of the heterologous transgene in parvalbumin (PV) neurons in a central nervous system (CNS).

INCORPORATION BY REFERENCE

[0014] All publications, patents, and patent applications mentioned in this specification are herein incorporated by reference to the same extent as if each individual publication, patent, or patent application was specifically and individually indicated to be incorporated by reference.

BRIEF DESCRIPTION OF THE DRAWINGS

[0015] The novel features of the invention are set forth with particularity in the appended claims. A better understanding of the features and advantages of the present invention will be obtained by reference to the following detailed description that sets forth illustrative embodiments, in which the principles of the invention are utilized, and the accompanying drawings of which:

[0016] FIG. 1 illustrates the frequency of seizures (seizures per 12hr interval) in SCN1A heterozygous mice after treatment with a recombinant AAVDJ vector comprising either SCN1B or eGFP operably linked to a regulatory element comprising a sequence of SEQ ID NO: 32. The graph illustrates the mean values at each day of recording with error bars representing the standard error of the mean.

[0017] FIG. 2 illustrates high gamma power (50-100 Hz) of different mice: wild-type control (WT), untreated transgenic APP/PS1 mice (APP/PS1), or transgenic APP/PS1 mice treated

with rAAV comprising SCN1B operably linked to a regulatory element comprising a sequence of SEQ ID NO: 32 (APP/PS1 + SCN1B).

[0018] FIG. 3A illustrates immunofluorescence co-localization assay of CNS cells from pups following neonatal systemic injections of AAV9 comprising eGFP transgene operably linked to a regulatory element comprising a sequence of SEQ ID NO: 1 or SEQ ID NO: 8. AAV9 comprising eGFP transgene operably linked to CAG was used as a control. Lower row images illustrate eGFP+ cells. Middle row images illustrate PV+ cells, which were stained with an anti-PV antibody. Top row images (merge) illustrate an overlay of PV+, eGFP+ fluorescence (with representative eGFP+ and PV+ cells which are shown as white or light grey cells indicated by arrowheads) and DAPI+.

[0019] FIG. 3B illustrates the quantification of immunofluorescence co-localization studies illustrated in **FIG. 3A**, wherein selective expression in PV cells is expressed as the

channel or a subunit thereof. In some cases, the transgene is SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.2, KV3.3, STXBP1, a DNA binding protein (e.g., a DNA binding protein that modulates expression of an endogenous gene), or a variant or functional fragment thereof. In some cases, the transgene comprises a sequence having at least 80%, at least 85%, at least 90%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to any one of SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.2, KV3.3, STXBP1, a DNA binding protein, or a variant or functional fragment thereof. In some cases, the transgene is a DNA binding protein that modulates expression of an endogenous gene, such as any one of SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.2, KV3.3, and STXBP1.

[0032] As used herein, the singular forms “a”, “an” and “the” are intended to include the plural forms as well, unless the context clearly indicates otherwise. Furthermore, to the extent that the terms “including”, “includes”, “having”, “has”, “with”, or variants thereof are used in either the detailed description and/or the claims, such terms are intended to be inclusive in a manner similar to the term “comprising”.

[0033] The term “about” or “approximately” means within an acceptable error range for the particular value as determined by one of ordinary skill in the art, which will depend in part on how the value is measured or determined, i.e., the limitations of the measurement system. For example, “about” can mean within one or more than one standard deviation, per the practice in the art. Alternatively, “about” can mean a range of up to 20%, up to 15%, up to 10%, up to 5%, or up to 1%) of a given value.

[0034] The terms “determining”, “measuring”, “evaluating”, “assessing”, “assaying”, “analyzing”, and their grammatical equivalents can be used interchangeably herein to refer to any form of measurement, and include determining if an element is present or not (for example, detection). These terms can include both quantitative and/or qualitative determinations. Assessing may be relative or absolute.

[0035] The term “expression” refers to the process by which a nucleic acid sequence or a polynucleotide is transcribed from a DNA template (such as into mRNA or other RNA transcript) and/or the process by which a transcribed mRNA is subsequently translated into peptides, polypeptides, or proteins. Transcripts and encoded polypeptides may be collectively referred to as “gene product.” If the polynucleotide is derived from genomic DNA, expression may include splicing of the mRNA in a eukaryotic cell.

[0036] As used herein, “operably linked”, “operable linkage”, “operatively linked”, or grammatical equivalents thereof refer to juxtaposition of genetic elements, e.g., a promoter,

enzymatic reaction in each of these assays preferentially targets open/accessible chromatin states, a state which is thought to be predictive of regulatory elements. To discover cell-type selective regulatory elements, one can assay for open chromatin sequence for target cell-type of interest (e.g., parvalbumin neurons) and compare that to open chromatin sequences for non-target cell types (e.g., excitatory neurons). Additional filters can be applied to further refine target selection, including proximity to a cell-type selective gene, species conservation, and/or sequence motifs, such as transcription factor binding sites. DNA sequences that are uniquely identified in the target cell type can be synthesized and cloned into an expression vector. The selectivity of a regulatory element can be determined using immunohistochemical methods to quantify co-localization to known cell-type selective proteins.

[0054] For example, one method of isolating a cell-type selective regulatory element includes isolating nuclei from a brain tissue or cell type of interest from an animal model, which can be achieved by using an affinity purification method that isolates the tissue or cell type of interest (e.g., using beads coated to an anti-PV antibody for isolating PV neurons), using high-throughput natural priming and DNA synthesis to generate a pool of sequences from open chromatin regions in the nuclei, sequencing the pool of sequences to identify putative sequences that drive gene expression in the tissue or cell type of interest, and verifying selective expression in a reporter system in a cell line in vitro and/or in an animal model.

[0055] Another method for identifying candidate regulatory elements that are selective in a tissue or cell type of interest include using R26-CAG-LSL-Sun1-sfGFP-Myc knockin mouse for harvesting the tissue or cell type of interest, isolating GFP+/Myc+ nuclei from the mouse neocortex of this strain using affinity purification, e.g., using anti-GFP or anti-Myc antibodies and protein G-coated magnetic beads to isolate nuclei from the neocortex. Nuclear RNA from purified nuclei or whole neocortical nuclei can be converted to cDNA and amplified with the Nugen Ovation RNA-seq System V2 (Nugen 7102), followed by sequencing using the Illumina HISEQ® 2500. Genomic DNA from purified nuclei can be fragmented and used to make MethylC-seq libraries, which can be sequenced using the Illumina HISEQ® 2000. To generate an ATAC-seq library, nuclei bound to beads are transposed using Tn5 transposase (Illumina FC-121–1030). After 9–12 cycles of PCR amplification, libraries are sequenced using an Illumina HISEQ® 2500. To generate a ChIP-seq library, excitatory neuron nuclei can be digested to mononucleosomes using micrococcal nuclease, followed by salt extraction of chromatin, and native ChIP and library construction, which can be sequenced on an Illumina HISEQ® 2500. After sequencing these libraries, the sequences are mapped to identify, for example, correlation in cell-type-specific hypo-methylation in CG-rich regions, histone

thereof, or sequences having at least 80%, at least 90%, at least 95%, or at least 99% sequence identity thereto.

[0065] In some cases, one or more regulatory elements of the present disclosure result in selective gene expression in a PV cell. In some cases, regulatory elements that show selective activity or function in a target cell type also show minimal activity or function in one or more off-target cell-types, e.g., non-PV CNS cell-types, non-inhibitory neurons or excitatory neurons, non-PV cells.

[0066] In some cases, one or more regulatory elements operably linked to a gene modulates gene expression in a cell, including but not limited to, selective expression in a target cell-type over non-target cell-types. Selective expression in a target cell or cell type can also be referred to as cell-selective expression or cell-type selective expression.

[0067] Selective expression generally refers to expression in a high fraction of cells of the cell type of interest (or the target cell type) as compared to other cells (or non-target cell type). Selective expression can also be viewed as preferential expression in a target cell or target cell type over one or more non-target cells or cell-types. In some cases, selective expression of one or more regulatory elements of this disclosure is compared to CAG, EF1 α , a constitutive promoter (e.g., SV40, CMV, UBC, PGK, and CBA), or a non-selective regulatory element that is known to drive expression in any cell or cell type without selectivity. In some cases, selective expression of one or more regulatory elements of this disclosure is compared to CAG, EF1 α , a constitutive promoter (e.g., SV40, CMV, UBC, PGK, and CBA), a non-selective regulatory element, or an expression cassette without the regulatory elements.

[0068] Non-target cell types can include a different subset, subtype, or type of cells as compared to the target cell or target cell type, or all non-target cell types. In some cases, one or more regulatory elements operably linked to a gene result in selective expression in a target cell type over at least one type of non-target cells, or at least two, at least three, at least four, at least five, or more than five types of non-target cells. In some cases, non-target cell types refer to all other cell types not including the target cell type. In some cases, non-target cell types are all other cell types within a relevant tissue or organ not including the target cell type, e.g., all non-target cell types in the CNS, all non-target cell types in the hippocampus. In some cases, a non-target cell or non-target cell type encompasses a subset or subtype of cells that is not the target cell. For example, non-PV CNS cell-types can include GABAergic cells that express calretinin and/or somatostatin instead of parvalbumin, or all GABAergic cells that do not express parvalbumin. In some cases, cell types are distinguished by having a

cassette. In some cases, an expression cassette further comprises a promoter, e.g., a hSyn1 promoter, CBA promoter, a CMV promoter, an EF1 α promoter, a polyA signal (e.g., SV40 polyA signal), or a post-transcriptional regulatory element such as woodchuck hepatitis virus post-transcriptional regulatory element (WPRE).

[0100] In some aspects, one or more regulatory elements described herein are operably linked to a transgene in an expression cassette. In some cases, a gene therapy comprises an expression cassette comprising a transgene operably linked to one or more, two or more, three or more, four or more, or five or more regulatory elements of the present disclosure to result in selective expression of the transgene in a target tissue or cell type, such as PV neurons. In some cases, an expression cassette comprises one or more PV cell selective regulatory elements or one or more regulatory elements disclosed herein operably linked to a transgene, e.g., a reporter gene, eGFP, SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.2, KV3.3, STXBP1, a DNA binding protein, or a variant or a fragment thereof, or sequences having at least 80%, at least 85%, at least 90%, at least 95%, or at least 99% sequence identity thereto.

[0101] In some cases, an expression cassette is adapted for delivery via gene therapy. In some cases, an expression cassette is a linear or a circular construct. In some cases, an expression cassette is part of a plasmid, vector, a viral vector, or rAAV.

[0102] In some cases, a gene therapy is administered directly to the CNS of a subject in need thereof or systematically via injection and/or infusion. In some cases, such subject has been diagnosed with a disease or condition associated with a haploinsufficiency or a genetic mutation, such as a haploinsufficiency or a mutation in any one of the following genes: SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.2, KV3.3, or STXBP1. In some cases, the subject is at risk for or has Dravet syndrome, Alzheimer's disease, epilepsy, neurodegeneration, tauopathy, neuronal hypoexcitability and/or seizures. In some cases, such gene therapy is delivered using a virus or a viral vector, such as rAAV. In some cases, an AAV serotype with a tropism for CNS cells and/or ability to cross the blood brain barrier is used, such as AAV9 or a variant thereof.

[0103] In some cases, one or more regulatory elements (e.g., one or more of SEQ ID NOs: 1-32, or a fragment or a combination thereof, or sequences having at least 80%, at least 85%, at least 90%, at least 95%, or at least 99% sequence identity thereto) operably linked to a transgene in an expression cassette result in selective gene expression in PV cells as compared to non-PV CNS cells, or as compared to a control element, such as a CAG, EF1 α , a constitutive promoter (e.g., SV40, CMV, UBC, PGK, and CBA), or a non-selective

selective regulatory elements or one or more regulatory elements disclosed herein operably linked to a transgene, wherein the transgene is SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KAV3.2, KV3.3, STXBP1, DNA binding protein (e.g., transcriptional modulator of an endogenous gene), or a variant or a functional fragment thereof, or a sequence having at least 80%, at least 85%, at least 90%, at least 95%, or at least 99% sequence identity thereto. In some cases, such regulatory elements increase the selective expression of the transgene in PV neurons as compared to non-PV CNS cell-types. In some cases, such regulatory elements increase the expression of the transgene selectively in a target cell type, such as a PV neuron. In some cases, the target cell type is a PV cell.

[0106] Techniques contemplated herein for gene therapy of somatic cells include delivery via a viral vector (e.g., retroviral, adenoviral, AAV, helper-dependent adenoviral systems, hybrid adenoviral systems, herpes simplex, pox virus, lentivirus, and Epstein–Barr virus), and non-viral systems, such as physical systems (naked DNA, DNA bombardment, electroporation, hydrodynamic, ultrasound, and magnetofection), and chemical system (cationic lipids, different cationic polymers, and lipid polymers).

[0107] The cloning capacity of vectors or viral expression vectors is a particular challenge for expression of large transgenes. For example, AAV vectors typically have a packaging capacity of ~4.8kb, lentiviruses typically have a capacity of ~8kb, adenoviruses typically have a capacity of ~7.5kb and alphaviruses typically have a capacity of ~7.5 kb. Some viruses can have larger packaging capacities, for example herpesvirus can have a capacity of >30kb and vaccinia a capacity of ~25kb. Advantages of using AAV for gene therapy include low pathogenicity, very low frequency of integration into the host genome, and the ability to infect dividing and non-dividing cells.

[0108] To address the size constraints of certain viral vectors or to improve expression from viral vectors, the present disclosure contemplates the use of regulatory elements that are shorter than 2.5 kb, 2 kb, 1.5 kb, 1 kb, 900bp, 800bp, 700bp, 600bp, 500bp, 400bp, 300bp, 200bp, 150bp, or 110bp, but at least 10bp, 50bp or 100bp in length. In some cases, the size of the combined regulatory element is about 2500bp, 2000bp, 1500bp, 1400bp, 1300bp, 1200bp, 1100bp, or 1000bp. In some cases, each combined regulatory element has a total length of about 100bp, 200bp, 300bp, 400bp, 500bp, 600bp, 700bp, 800bp, 900bp, 1000bp, 1100bp, 1200bp, 1300bp, 1400bp, 1500bp, 1600bp, 1700bp, 1800bp, 1900bp, 2000bp, 2100bp, 2200bp, 2300bp, 2400bp, or 2500bp. In some cases, the size of a combined RE has a total length of about 200bp–3000bp, 200bp–2500bp, 200bp–2100bp, 500bp–2500bp, 1000bp–2500bp, 1500bp–2500bp, 1500b–2000bp, or 2000bp–2500bp.

deactivated, e.g., a nuclease-deactivated Cas (dCas), a deactivated transcription activator-like effector nuclease, or a nuclease-deactivated zinc finger protein. In some cases, the DNA binding domain is linked to a transcriptional modulating domain (e.g., a transcriptional activator or repressor domain). In some cases, the transgene comprises a gene editing protein, e.g., a Cas protein, Cas9.

[0118] In some aspects, the transgene is a voltage-gated ion channel or a subunit thereof, such as SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.2, or KV3.3, or a functional fragment or variant thereof. In some cases, the transgene is an alpha subunit of a sodium ion channel. In some cases, the transgene is a beta subunit of a sodium ion channel. In some aspects, the neurotransmitter regulator is STXBP1 or a functional fragment or variant thereof.

[0119] In some aspects, an expression cassette is delivered as a viral vector, such as AAV. In some aspects, the AAV is AAV1, AAV8, AAV9, AAV-DJ, scAAV1, scAAV8, or scAAV9. In some aspects, a gene therapy comprising an expression cassette of this disclosure is administered to a subject in need thereof (e.g., a human patient, a mammal, a transgenic animal, or an animal model). In some cases, the subject in need thereof has symptoms of, has been diagnosed with, or is at risk of developing Alzheimer's disease, Dravet syndrome, epilepsy, neurodegeneration, tauopathy, neuronal hypoexcitability, and/or seizures. In some cases, the subject in need thereof has an insufficient gene expression or a mutation in any one or more of SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.2, KV3.3, and STXBP1.

[0120] In some cases, a gene therapy, such as rAAV9, is used to deliver an expression cassette comprising one or more PV cell selective regulatory elements or one or more regulatory elements disclosed herein operably linked a transgene, wherein the transgene is SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.2, KV3.3, STXBP1, a DNA binding protein, or a functional fragment thereof, or a sequence having at least 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% sequence identity to any one of SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.2, KV3.3, STXBP1, a DNA binding protein, or a functional fragment thereof. In some cases, the transgene comprises a sequence having at least 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 100% sequence identity to any one of SEQ ID NOs: 37-43, or a functional fragment thereof, as provided in **TABLE 2** below.

basal promoter, a transgene, one or more post-transcriptional RNA regulatory elements, and a 3' ITR.

[0124] An exemplary AAV expression cassette is illustrated in **FIG. 7**. In some cases, the expression cassette contains a 5' AAV ITR, an enhancer (e.g., PV cell selective enhancer or one or more combined regulatory elements), a promoter (e.g., one or more PV cell selective promoters or regulatory elements), a transgene (e.g., SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.2, KV3.3, STXBP1, or a DNA binding protein), a post-transcriptional regulatory element, and a 3' AAV ITR. The promoter can be PV cell selective, or a constitutive promoter. In some cases, the transgene is a reporter gene, e.g., a coding sequence for eGFP, RFP, or a fluorescent marker. In other cases, the transgene is a DNA binding protein that modulates gene expression.

[0125] In some cases, the transgene is a therapeutic transgene, e.g., a coding sequence for SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.2, KV3.3, STXBP1, or a DNA binding protein, or a functional fragment or a sequence having at least 80%, at least 85%, at least 90%, at least 95%, or at least 99% sequence identity thereto. The post-transcriptional regulatory element can be any sequence which influences the expression of a protein from an mRNA or stability of an RNA, for example, an intron, an internal ribosome entry site (IRES), or a woodchuck hepatitis virus post-transcriptional regulatory element. In some cases, the post-transcriptional regulatory element is a combination of two or more post-transcriptional regulatory elements.

[0126] The expression cassette can be designed for delivery by an optimized therapeutic retroviral vector, e.g., a lentiviral vector. The retroviral vector can be a lentiviral vector comprising a left (5') LTR; sequences which aid packaging and/or nuclear import of the virus, at least one cell-type selective regulatory element, optionally a lentiviral reverse response element (RRE); optionally a promoter or active portion thereof; a transgene operably linked to one or more regulatory elements; optionally an insulator; and a right (3') retroviral LTR.

[0127] In some cases, the expression cassette comprises one or more cell-type selective regulatory elements disclosed herein. In some cases, the expression cassette comprises two or more regulatory elements combined. In some examples, the expression cassette comprises two or more regulatory elements that are not combined, for example, a promoter upstream of the transgene and an enhancer or stability element located downstream of the transgene.

[0128] In some cases, the expression cassette contains a putative cell-type selective regulatory element that has selective activity in a cell type of interest, for example, a putative

cell-type selective expression, or preferential expression, of the transgene in a target cell type over at least one, at least two, at least three, at least four, at least five, or more than five non-target cell types. In some cases, an expression cassette comprises one or more regulatory elements operably linked to a transgene to result in cell-type selective expression or preferential expression in a target cell subtype over at least one, at least two, at least three, at least four, at least five, or more than five non-target subtypes, or all other known subtypes of the cell. In some cases, the transgene is any one of SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.3, and STXBP1, or a functional fragment thereof, or sequences having at least 80%, at least 85%, at least 90%, at least 95%, or at least 99% sequence identity thereto. In some cases the transgene is a DNA binding protein that modulates an endogenous gene (e.g., an endogenous SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.2, KV3.3, or STXBP1). In some cases, the transgene is any one of SEQ ID NOs: 36-43, or a functional fragment thereof, or sequences having at least 80%, at least 85%, at least 90%, at least 95%, or at least 99% sequence identity thereto. In some cases, the transgene is a transcriptional modulator. In some cases, the transgene is a DNA binding protein that comprises a DNA binding domain of a DNA binding protein or a DNA cleaving protein (e.g., a nuclease, a restriction enzyme, a recombinase, etc.) wherein the DNA cleaving domain or nuclease domain has been deactivated, e.g., a nuclease-deactivated Cas (dCas), a deactivated transcription activator-like effector nuclease, or a nuclease-deactivated zinc finger protein. In some cases, the DNA binding domain is linked to a transcriptional modulating domain (e.g., a transcriptional activator or repressor domain). In some cases, the transgene is a gene editing protein, such as a Cas family protein, Cas9, a zinc finger nuclease, a zinc finger nuclease, or a transcription activator-like effector nuclease. In some cases, the transgene is a reporter gene or a fluorescent marker. In some cases, an expression cassette disclosed herein is in a viral vector. In some cases, an expression cassette disclosed herein is packaged in an rAAV, such as rAAV9 or rAAVDJ. In some cases, an expression cassette disclosed herein is delivered into a cell as a gene therapy. In some cases, a gene therapy disclosed herein is delivered into a subject, preferably a human or a mammal. In some cases, an expression cassette disclosed herein is used to treat a neurological condition or disease, such as epilepsy, a neurodegenerative disease, tauopathy, neuronal hypoexcitability, Dravet syndrome or Alzheimer's disease.

[0136] In some cases, an expression cassette (e.g., gene therapy, viral vector, vector, or plasmid) comprises any one or more of SEQ ID NOs: 1-32, or a functional fragment or a combination thereof, or sequences having at least 80%, at least 85%, at least 90%, at least

that does not express parvalbumin. In some instances, other CNS cells refer to CNS cell types that have never expressed any of PV, GAD2, GAD1, NKX2.1, DLX1, DLX5, SST and VIP.

[0152] In some instances, a regulatory element disclosed herein is selective for a PV expressing cell over at least one, two, three, four, five, or more than five non-PV CNS cell-types. In some cases, non-PV cell types include non-PV GABAergic cells. In some cases, the cell type of interest is a PV cell. In some cases, REs selective for PV cells are referred to as PV cell selective regulatory elements.

[0153] In some cases, the PV cell selective regulatory elements disclosed herein include the sequences of SEQ ID NOs: 1-32, or any combination thereof.

[0154] In some cases, one or more PV cell selective regulatory elements or one or more regulatory elements disclosed herein are used to increase expression of a transgene in PV-expressing cells by at least 2, 5, 10, 15, 20, 30, 40, 50, 60, 70, 80, 90, 100, or more fold as compared to expression without the regulatory element. In some cases, a RE in an expression cassette increases gene expression by at least 1.5%, 2%, 5%, 10%, 15%, 20%, or 50%, or more than 1.5%, 2%, 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, or 100% as compared to expression without the regulatory elements. In some cases, compositions and methods of use thereof comprise expression cassettes containing one or more regulatory elements that result in a 10-500% increase in transgene expression, e.g., expression of SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KCNC1 (also known as KV3.1), KCNC3 (also known as KV3.3), STXBP1, a DNA binding protein, or a variant or functional fragment thereof, or a protein thereof, as compared to the level without the regulatory elements or as compared to a non-selective regulatory element (e.g., CAG, EF1 α , a constitutive promoter, or SEQ ID NO: 34, or a functional fragment thereof, or a sequence having at least 80%, at least 85%, at least 90%, at least 95%, or at least 99% sequence identity thereto). In some cases, the increase in gene expression and/or protein level of any one of SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KCNC1, KCNC3, and STXBP1 is 1.5-5%, 5%-10%, 10-15%, 15-20%, 20-30%, 30-40%, 40-50%, 50-60%, 60-70%, 70-80%, 80-90%, 90-100%, 100-150%, 150-200%, 250-300%, 300-350%, 350-400%, 400-450%, 450-500%, or 1.5-20%, 20%-50%, 50%-100%, 100-200%, 200-300%, 300-400%, or 400-500% as compared to the level without the expression cassette or regulatory elements. In some cases, such gene or protein expression is selective in PV cells as compared to an expression cassette comprising a control (e.g., CAG, EF1 α , a constitutive promoter (e.g., SV40, CMV, UBC, PGK, and CBA), or a non-selective regulatory element) or a non-cell type selective regulatory element (e.g., SEQ ID NO: 34, or a functional

[0169] In some cases, a transgene is useful to treat a disease associated with a specific cell type of interest. In some cases, a cell type of interest is a neuron, an inhibitory neuron, a GABAergic neuron, or a PV neuron. In some cases, a transgene is any one or more of SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.3, and STXBP1. In some cases, a transgene is a DNA binding protein that modulates expression of a gene (e.g., a transcriptional activator or a transcriptional repressor that modulates expression of an endogenous gene). In some cases, a transgene is a gene editing protein, such as a zinc finger nuclease, a transcription activator-like effector nuclease, a Cas family protein. In some cases, a transgene is a reporter gene or a detectable marker, such as eGFP, tdTomato, or RFP. In some cases, a transgene is a Cas protein, such as Cas9.

[0170] Transgenes useful to treat a condition associated with PV neuron cells can be incorporated in a vector, nucleic acid cassette, or method as described herein. Transgenes used herein generally do not contain introns, or do not contain more than one intron. A transgene can be obtained from a cDNA sequence rather than from genomic sequence. In some cases, transgenes can contain some, or all, of their endogenous introns. In some examples, such a transgene encodes for a DNA binding domain or an ion channel. Examples of DNA binding domains that can be encoded for in the expression cassettes of this disclosure include zinc fingers, Cas9, a Cas family protein, dCas9, a dCas family protein or a transcriptional activator like effector (TALE). In some cases, the transgene is a DNA binding protein that comprises a DNA binding domain of a DNA binding protein or a DNA cleaving protein (e.g., a nuclease, a restriction enzyme, a recombinase, etc.) wherein the DNA cleaving domain or nuclease domain has been deactivated, e.g., a nuclease-deactivated Cas (dCas), a deactivated transcription activator-like effector nuclease, or a nuclease-deactivated zinc finger protein. In some cases, the DNA binding domain is linked to a transcriptional modulating domain (e.g., a transcriptional activator or repressor domain). In some cases, the transgene comprises a gene editing protein, e.g., a Cas protein, Cas9. In some cases, the transgene is a subunit or a component of an ion channel or a membrane protein, or a gene associated with a neurological condition or disease disclosed herein. Examples of ion channel transgenes which can be used in the expression cassettes of this disclosure include voltage gated and ligand gated ion channels. Voltage gated ion channels include sodium channels, calcium channels, potassium channels, and proton channels. In some instances, the transgene codes for a subunit of a voltage gated sodium channel. Examples of voltage gated sodium channel subunits include SCN1B (NM_001037.4), SCN1A (NM_001165963.1), and SCN2B, (NM_004588.4).

[0171] In some instances, the transgene codes for a subunit of a voltage gated potassium channel. Examples of voltage gated sodium channel subunits include KCNC1 (NM_001112741.1), and KCNC3 (NM_004977.2). In some cases, a transgene is any one or more of SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.3, STXBP1, a variant and a functional fragment thereof. In some cases, a transgene is a sequence having at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or at least 99.5% sequence identity to any one of SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.3, STXBP1, a variant or a functional fragment thereof. In some cases, such sequence identity is measured using BLAST.

[0172] In some cases, an expression cassette disclosed herein comprises one or more PV-selective regulatory elements or one or more regulatory elements of this disclosure operably linked to a transgene having at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or at least 99.5% sequence identity to a sequence encoding any one of SEQ ID NOs: 37-43, or a functional fragment or variant thereof, or the GenBank sequences corresponding to SEQ ID NOs: 37-43. In some cases, an expression cassette disclosed herein comprises a transgene having a sequence according to (i) a sequence of SEQ ID NOs: 37-43, or (ii) a functional fragment thereof, or (iii) a sequence having at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or at least 99.5% sequence identity to (i) or (ii).

[0173] In some examples, the transgene is a neurotransmitter regulator, or a variant or functional fragment thereof. A neurotransmitter regulator may be involved in regulating production or release of a neurotransmitter in the CNS. For example, a neurotransmitter regulator may assist with synaptic fusion to release neurotransmitters. An example of a neurotransmitter regulator is STXBP1 (NM_001032221.3) or a functional fragment thereof, or a sequence having at least 80%, at least 85%, at least 90%, at least 95%, or at least 99% sequence identity thereto. The transgene may also be a subunit of a neurotransmitter regulator.

[0174] In some case, an expression cassette of this disclosure can contain an AAV2 5' ITR, a PV-selective enhancer, a PV-selective promoter, or a combination of one or more PV-selective promoters and enhancers, a cDNA of SCN1B, a WPRE, a hGH polyA signal, PV-selective regulatory element, and an AAV2 3' ITR. In one example, the expression cassette comprises an AmpR promoter, and AmpR coding sequence, a bacterial origin of replication,

subject can be a patient suffering from a form of epilepsy. In some instances, the subject is a patient with Dravet syndrome. The subject can be a patient suffering from a neurodegenerative disease, for example, a patient with Alzheimer's disease. In some instances, epilepsy, encephalopathy, and/or seizures are associated with a genetic mutation in SCN8A. In some cases, a genetic mutation in SCN8A can give rise to epilepsy syndromes, e.g., Dravet syndrome. In some instances, a genetic mutation in STXBP1 is associated with encephalopathy with epilepsy, characterized by recurrent seizures.

[0182] In some instances, a subject treated with one or more compositions described herein is one diagnosed with a mutation or genetic aberration in an ion channel or a neurotransmitter regulator (e.g., a syntaxin binding protein). Examples of such mutations include mutations in SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KCNC1, KCNC3, and/or STXBP1, or combination thereof. The expression cassette containing a cell-type selective regulatory element as described herein can be delivered to a subject to treat or prevent a disease with symptoms associated with a specific cell type. For example, an expression cassette comprising a transgene operably linked to one or more PV cell selective regulatory element is delivered to a subject who has symptoms, or is at risk of developing symptoms, associated with PV neurons.

[0183] In some cases, the treatment can be administered to a subject with, or at risk of developing, Dravet syndrome. Symptoms associated with Dravet syndrome include seizures, memory defects, developmental delay, poor muscle tone and/or cognitive problems. Treatment with an expression cassette of this disclosure can result in an improvement of one or more symptoms, such as a reduction in number, duration, and/or intensity of seizures. Administration of a gene therapy as described herein to a subject at risk of developing Dravet syndrome can prevent the development of or slow the progression of one or more symptoms.

[0184] In another example the treatment may be administered to a subject suffering from Alzheimer's disease. Symptoms associated with Alzheimer's disease include short term memory loss, cognitive difficulties, seizures, and difficulties with language, executive functions, perception (agnosia), and execution of movements (apraxia). Treatment with an expression cassette of this disclosure can result in an improvement of one or more Alzheimer's disease symptoms, such as a reduction in progression of memory loss, or the prevention of one or more symptoms. In some cases, the treatment can result in a correction of high gamma power brain activity. The treatment can result in a decrease in seizure frequency and/or seizure severity, or a decrease in high gamma power activity by 10%, 20%, 30%, 40%, 50%, 60%, or 70%. In some cases, the treatment can result in an improvement in

transgene (e.g., a reporter transgene or a therapeutic transgene) such that the regulatory elements drive selective expression or preferential expression in at least one target cell type at a level that is statistically significantly higher than the expression driven by CAG or EF1 α or a non-selective regulatory element when operably linked to the same transgene, or by the same construct without the regulatory elements. In some cases, statistically significantly higher means the regulatory elements drive selective expression in the target cell type at a level that is at least 1.1, 1.2, 1.3, 1.4, 1.5, 2, 2.5, 3, 3.5, 4, 4.5, 5, 5.5, 6, 6.5, 7, 7.5, 8, 8.5, 9, 9.5, 10, 10.5, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 25, 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95, or 100 times the expression level by the CAG, EF1 α , a constitutive promoter, or a non-selective regulatory element when operably linked to the same transgene in the target cell type, or by the same construct without the regulatory elements. In some cases, such cell-type selective expression is assayed using a co-localization assay as described herein. In some cases, the transgene is any one or more of SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.3, STXBP1, and a functional fragment thereof. In some cases, the transgene is a DNA binding protein that modulates expression of an endogenous gene, such as a transcriptional modulator, a transcriptional activator, or a transcriptional repressor. In some cases, the transgene is a DNA binding protein that comprises a DNA binding domain of a DNA binding protein or a DNA cleaving protein (e.g., a nuclease, a restriction enzyme, a recombinase, etc.) wherein the DNA cleaving domain or nuclease domain has been deactivated, e.g., a nuclease-deactivated Cas (dCas), a deactivated transcription activator-like effector nuclease, or a nuclease-deactivated zinc finger protein. In some cases, the DNA binding domain is linked to a transcriptional modulating domain (e.g., a transcriptional activator or repressor domain). In some cases, the transgene is a gene editing protein, such as a zinc finger nuclease or a transcription activator-like effector nuclease. In some cases, a transgene is a reporter gene or a detectable marker, such as eGFP, tdTomato, or RFP. In some cases, a transgene is a Cas protein, such as Cas9.

[0190] In some cases, a gene therapy comprising an expression cassette disclosed herein is used to treat a neurological condition or disease. In some cases, a gene therapy comprising an expression cassette disclosed herein is used to treat a neurological condition or disease, wherein the expression cassette comprises any one or more, two or more, three or more, four or more, five or more, six or more, seven or more, eight or more, nine or more, or ten or more of SEQ ID NOs: 1-32, or a functional fragment or a combination thereof, or sequences having at least 80%, at least 85%, at least 90%, at least 95%, or at least 99% sequence identity thereto, operably linked to a transgene. In some cases, the transgene is any one or

more of SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.3, STXBP1, a DNA binding protein and a functional fragment thereof. In some cases, a gene therapy comprising an expression cassette disclosed herein is used to treat Dravet syndrome. In some aspects, a gene therapy comprising an expression cassette disclosed herein is used to treat Alzheimer's disease. In some cases, a gene therapy comprising an expression cassette disclosed herein is used to treat epilepsy and/or seizure symptoms associated with Dravet syndrome and/or Alzheimer's disease. In some cases, treating any one of Dravet syndrome, Alzheimer's disease, epilepsy, neurodegeneration, tauopathy, neuronal hypoexcitability and/or seizures comprises delivering or administering a gene therapy of this disclosure to a cell of a subject in need thereof. In some cases, the subject in need thereof is at risk for or has any one of Dravet syndrome, Alzheimer's disease, epilepsy, and/or seizures. In some cases, the subject is a child or a minor. In some cases, a gene therapy comprising an expression cassette disclosed herein is used to treat an infant, a child, or a minor diagnosed with or is at risk of developing Dravet syndrome. In some cases, a gene therapy comprising an expression cassette disclosed herein is used to treat a subject comprising a mutation or a genetic defect in any one or more of SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.3, and STXBP1.

[0191] In some aspects, the present disclosure provides a method of treating any one of Dravet syndrome, Alzheimer's disease, epilepsy, neurodegeneration, tauopathy, neuronal hypoexcitability and/or seizures, comprising administering a gene therapy into a cell of a subject, wherein the gene therapy comprises an expression cassette disclosed herein. In some cases, such expression cassette comprises any one or more of SEQ ID NOs: 1-32, or a functional fragment or a combination thereof, or sequences having at least 80%, at least 85%, at least 90%, at least 95%, or at least 99% sequence identity thereto, operably linked to a transgene, wherein the transgene is any one of SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.3, STXBP1, and a functional fragment thereof. In some cases, the transgene is a subunit of a sodium ion channel or a potassium ion channel. In some cases, the transgene is a syntaxin binding protein. In some cases, the transgene is a transcriptional modulator, e.g., a transcriptional activator or a transcriptional repressor. In some cases, the transgene is a transcriptional modulator that modulates the expression of an endogenous gene (e.g., SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.3, or STXBP1). In some cases, a transgene is a gene editing protein, such as a zinc finger nuclease or a transcription activator-like effector nuclease. In some cases, the transgene is a Cas protein, such as Cas9.

[0192] In other aspects, the present disclosure provides a method for modifying any gene therapy designed for treating Dravet syndrome, Alzheimer's disease, epilepsy, neurodegeneration, tauopathy, neuronal hypoexcitability and/or seizures by adding one or more regulatory elements disclosed herein to improve the cell-type selectivity of the gene therapy. In some cases, the gene therapy is an rAAV gene therapy.

[0193] In some cases, treatment with an expression cassette disclosed herein reduces seizure duration and/or frequency, e.g., seizures associated with Dravet syndrome, by at least 1%, 2%, 3%, 4%, 5%, 6%, 7%, 8%, 9%, 10%, 11%, 12%, 13%, 14%, 15%, 16%, 17%, 18%, 19%, 20%, 21%, 22%, 23%, 24%, 25%, 26%, 27%, 28%, 29%, 30%, 31%, 32%, 33%, 34%, 35%, 36%, 37%, 38%, 39%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, or 95% as compared to an untreated control or as compared to the level before treatment.

[0194] In some cases, treatment with an expression cassette disclosed herein reduces high gamma power activity (e.g., high gamma power activity associated with Alzheimer's disease) by at least 1%, 2%, 3%, 4%, 5%, 6%, 7%, 8%, 9%, 10%, 11%, 12%, 13%, 14%, 15%, 16%, 17%, 18%, 19%, 20%, 21%, 22%, 23%, 24%, 25%, 26%, 27%, 28%, 29%, 30%, 31%, 32%, 33%, 34%, 35%, 36%, 37%, 38%, 39%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, or 95% as compared to an untreated control or as compared to the level before treatment.

[0195] In one aspect, the present disclosure provides a nucleic acid cassette of this disclosure comprises one or more regulatory elements operably linked to a transgene that result in selective expression in a target cell type over one or more non-target cell types, e.g., selective expression in PV neurons in the CNS over one or more non-PV CNS cell types. In some cases, each of the regulatory elements comprises (i) a sequence of SEQ ID NOs: 1-32, (ii) a functional fragment or a combination thereof, or (iii) a sequence with at least 80% sequence identity to (i) or (ii). In some cases, the percent sequence identity can be measured using BLAST. In some cases, at least one of the regulatory elements is human derived. In some cases, at least one of the regulatory elements is derived from a non-human mammal. In some cases, the regulatory elements are non-naturally occurring. In some cases, the regulatory elements result in a selectivity of expression in PV cells that is greater than expression of the transgene operably linked to CAG or EF1 α or a non-selective regulatory element as measured by a co-localization assay. In some cases, a transgene is any one or more of SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.3, and STXBPI, or a functional fragment hereof, or sequences having at least 80%, at least 85%, at least 90%, at least 95%, or at least 99% sequence identity thereto. In some cases, a transgene is a DNA binding protein that modulates

expression of a gene (e.g., an endogenous gene such as SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.3, and STXBP1), such as transcriptional modulator, transcriptional activator, or transcriptional repressor. In some cases, a transgene is a gene editing protein, such as a zinc finger nuclease or a transcription activator-like effector nuclease. In some cases, a transgene is a reporter gene or a detectable marker, such as eGFP, tdTomato, or RFP. In some cases, a transgene is a Cas protein, such as Cas9. In some cases, the regulatory elements result in selective expression in PV cells at a level that is at least 0.5 fold, at least 0.6 fold, at least 0.7 fold, at least 0.8 fold, at least 0.9 fold, at least 1.1 fold, at least 1.2 fold, at least 1.3 fold, at least 1.4 fold, at least 1.5 fold, at least 2 fold, at least 3 fold, at least 4 fold, at least 5 fold, at least 6 fold, at least 7 fold, at least 8 fold, at least 9 fold, at least 10 fold, at least 11 fold, at least 12 fold, at least 13 fold, at least 14 fold, at least 15 fold, at least 16 fold, at least 17 fold, at least 18 fold, at least 19 fold, at least 20 fold, at least 25 fold, at least 30 fold, at least 40 fold, at least 50 fold, at least 60 fold, at least 70 fold, at least 80 fold, at least 90 fold, at least 100 fold as compared to that of a CAG or EF1 α or a non-selective regulatory element, as measured by the co-localization assay. In some cases, a fold difference refers to the fold difference between the percentage of eGFP+, PV+ cells that result from one or more regulatory elements and that of a non-selective regulatory element. In some cases, the co-localization assay is an immunohistochemical assay, as described below in Example 5. In some instances, a co-localization assay is performed using a commercially available anti-PV antibody. In some cases, the transgene encodes an ion channel subunit, a neurotransmitter regulator, or a variant or a functional fragment thereof. In some cases, the ion channel subunit is an α subunit or a β subunit of a sodium ion channel or a subunit of a potassium ion channel. In some cases, the transgene is any one of (i) SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.3, or a DNA binding protein; (ii) a functional fragment thereof; or (iii) a sequence having at least 80% sequence identity to (i) or (ii). In some cases, the neurotransmitter regulator is (i) STXBP1, (ii) a functional fragment thereof, or (iii) a sequence having at least 80% sequence identity to (i) or (ii). In some cases, the regulatory elements and the operably linked transgene are located on different chromosomes. In some cases, the regulatory elements combined are less than 2.5kb, less than 1.5 kb, less than 1 kb, or less than 500 bp in size. In some cases, the non-PV cells comprise any one or more of non-PV CNS cell types, including but not limited to excitatory neurons, dopaminergic neurons, astrocytes, microglia, or motor neurons. In some cases, the nucleic acid cassette is a linear construct. In some cases, the nucleic acid cassette is a vector. In some cases, the nucleic acid cassette is a plasmid. In some cases, the vector is a viral vector. In some cases, the viral

[0198] In one aspect, a neurological condition described herein is treated with a gene therapy, preferably one that results in preferential expression in one tissue type or cell type over another, e.g., a PV neuron as determined via a co-localization assay. In some cases, the gene therapy is an AAV.

[0199] In one aspect, a method of targeting expression of any transgene to PV neurons in the CNS comprises operably linking one or more of PV neuron selective regulatory elements to a transgene. In some cases, the regulatory elements comprise one or more sequences of SEQ ID NOs: 1-32, or sequences with at least 80% sequence identity to SEQ ID NOs: 1-32, or a functional fragment thereof. In some cases, the regulatory elements result in selective expression in PV neurons at a level that is at least 2 fold, at least 5 fold, or at least 7 fold, or at least 10 fold as compared to CAG or EF1 α or a non-selective regulatory element operably linked to the transgene, as measured by a co-localization assay. In some cases, the transgene is SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.3, STXBP1, a DNA binding protein, or a functional fragment thereof. In some cases, the regulatory elements and the transgene are in an AAV. In some cases, the AAV is AAV9.

[0200] In one aspect, a method of treating a neurological condition or disorder in a subject in need thereof comprises contacting a cell with a nucleic acid cassette comprising one or more regulatory elements operably linked to a transgene that results in selective expression in PV neurons over one or more non-PV CNS cells. In some cases, the regulatory elements comprise one or more of SEQ ID NOs: 1-32, or a functional fragment or a combination thereof, or sequences having at least 80%, at least 85%, at least 90%, at least 95%, or at least 99% sequence identity thereto. In some cases, the transgene is a voltage-gated ion channel subunit, or a variant or a functional fragment thereof. In some cases, the subunit is a beta subunit of a sodium ion channel. In some cases, the subunit is an alpha subunit of a sodium ion channel. In some cases, the subunit is of a potassium ion channel. In some cases, the transgene is any one of (i) SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.3, a DNA binding protein or STXBP1; (ii) a functional fragment thereof; or (iii) a sequence having at least 80% sequence identity to (i) or (ii). In some cases, the neurological condition or disorder is associated with a haploinsufficiency or a mutation in any of SCN1A, SCN1B, SCN2B, KV3.1, and KV3.3. In some cases, the neurological condition or disorder is Dravet syndrome. In some cases, the neurological condition or disorder is Alzheimer's disease. In some cases, the neurological condition or disease is a psychiatric disorder (e.g., schizophrenia, obsessive compulsive disorder, addiction, depression, anxiety, psychosis); an autism spectrum disorder (e.g., Fragile X syndrome, Rett syndrome); epilepsy (e.g., chronic

EXAMPLES

[0203] These examples are provided for illustrative purposes only and not to limit the scope of the claims provided herein.

EXAMPLE 1

Identifying Putative PV-Selective Regulatory Elements

[0204] To identify and screen putative regulatory element that are selective for PV cells, one can harvest PV cells from a R26-CAG-LSL-Sun1-sfGFP-Myc knockin mouse using affinity purification, e.g., using anti-GFP or anti-Myc antibodies and protein G-coated magnetic beads. PV cells can be enriched by using anti-PV antibody coated beads or affinity purification matrix. Nuclei are then isolated from the PV cells. Nuclear RNA can be purified from the nuclei and converted to cDNA, and amplified with the Nugen Ovation RNA-seq System V2 (Nugen 7102), followed by sequencing using the Illumina HISEQ® 2500. Genomic DNA can be purified from nuclei, fragmented, and used to make methylC-seq libraries, which can be sequenced using the Illumina HISEQ® 2000. To generate an ATAC-seq library, nuclei bound to beads are transposed using Tn5 transposase (Illumina FC-121–1030). After 9–12 cycles of PCR amplification, libraries are sequenced using an Illumina HISEQ® 2500. To generate a ChIP-seq library, nuclei of PV cells are digested to mononucleosomes using micrococcal nuclease, followed by salt extraction of chromatin, and native ChIP and library construction, which can be sequenced on an Illumina HISEQ® 2500. After sequencing these libraries, the sequences are mapped to identify correlations and patterns in hypo-methylation in CG-rich regions, histone modifications, transcriptional factor binding sites, and patterns associated with highly expressed transcriptional factors in PV cells. Overlapping features and correlations from multiple assays and/or libraries described above provide convergent evidence for identifying candidate sequences that are putative PV-selective regulatory elements. Putative PV-selective regulatory elements can be further tested using a co-localization assay as described in Example 5 below. Putative PV-selective regulatory elements can also be tested in B6 PV-Cre mouse (Jackson Laboratory), which is a B6 PV-Cre knock-in mouse that expresses Cre recombinase in parvalbumin-expressing, as described in Example 2 below. After validating PV-selectivity of the regulatory elements, the regulatory elements can be operably linked to a transgene to target expression selectively to PV cells over at least one, two, three, four, five, or more than five non-PV cells.

EXAMPLE 2

Selectivity for PV Neurons in PV-Cre Mouse

[0205] Selectivity for PV neurons can be determined using fluorescent imaging. AAV9 vectors containing eGFP operably linked to (i) a control promoter (EF1a); or (ii) a PV-selective RE identified in Example 1 above; or (iii) a PV-selective RE selected SEQ ID NOs: 1-32; and AAV9 vectors containing a Cre dependent tdTomato are co-injected into a B6 PV-Cre mouse (Jackson Labs). PV-Cre is a knock-in mouse that expresses Cre recombinase in parvalbumin-expressing neurons (such as interneurons in the brain and proprioceptive afferent sensory neurons in the dorsal root ganglia), without disrupting endogenous Pvalb expression.

[0206] Mice are infused bilaterally with 1.5 μ L of AAV9 vector (5^{12} to 1^{13} gc/ml) into the dorsal and ventral hippocampus at a rate of 0.3 μ L/min with a 4 min rest period following injection. Mice are anesthetized for the injection. The animals are placed in a stereotaxic frame (Kopf instruments, USA), using the following coordinates for the dorsal hippocampus (AP -2.0 mm, lateral $\pm$ 1.5, DV -1.4 mm from dura) and the ventral hippocampus (AP -3.1 mm, lateral $\pm$ 2.8, DV -3.8 mm from dura). A Hamilton syringe (model # 80308; 10 μ L syringe with corresponding 30 ga blunt tip needle) can be used with the stereotactic micromanipulator, to designate and drill the bur holes. The drill is only used to penetrate the bone. Following drilling, the infusion cannula is lowered into the brain to the depth of the desired location for injection, e.g., injection volume: 1.5 μ L; injection rate: 0.3 μ L/min. Prior to infusion, the needle is allowed to equilibrate for 1 minute. Once delivery is completed, the needle is left for 4 min and then withdrawn over approximately 1 min. Once all infusions are complete, the skin incision is closed with sutures and administered post-surgery analgesics. The treated mice undergo daily health checks for the remainder of the study and are weighed once weekly to monitor body weight.

[0207] For tissue collection, mice are euthanized via isoflurane overdose and perfused with 4% Paraformaldehyde (PFA). A piece of brain tissue containing the hippocampus is extracted and placed in 4% PFA at 4°C for at least 12 hours. The brain tissue is then dehydrated in 30% sucrose (in phosphate buffered saline) at 4°C until the tissue sinks to the bottom of the tube. Brain tissue is embedded in TISSUE-TEK® OCT for sectioning in a cryostat. Sectioned brain tissue is stained for eGFP and tdTomato using standard immunohistochemistry procedures with anti-RFP polyclonal rabbit antibody (Rockland Antibodies and Assay) and anti-eGFP polyclonal chicken antibody (Aves Labs). Fluorescence microscope imaging is used to visualize the cells. eGFP, or green fluorescence, corresponds to all gene expression. Red

fluorescence from tdTomato corresponds to PV+ cells. An overlap of the two fluorescence signals, which can be visualized as yellow or white cells, represents PV+ cells that express the eGFP transgene. AAV9 vectors comprising a PV-selective regulatory element is expected to yield higher number of cells that are eGFP+ and PV+ as compared to the control promoter (EF1a). For example, fluorescence imaging of cells from mice injected with AAV9s comprising any one of PV-selective REs (e.g., SEQ ID NOs: 1-32 or putative REs identified in Example 1) are expected to show higher number of eGFP+ cells that are also PV+. Selectivity for PV cells can be quantified as percentage of all eGFP+ cells that are also PV+.

EXAMPLE 3

Reduction of Seizures in Dravet Mouse Model

[0208] B6(Cg)-*Scn1a*^{tm1.1Dsf/J} mice were obtained from the Dravet syndrome European Federation via the Jackson Laboratories. These mice contain a Dravet syndrome associated mutation in exon 24 of SCN1A (A to V at position 1783). The mice also contain a floxed exon 24 with wildtype sequence. When not manipulated, this strain of mice expresses two copies of the WT allele of SCN1A. However, upon delivery of an AAV expressing Cre recombinase, any cell targeted by the AAV will switch to expressing one copy of the mutant allele. Upon expression of the mutant SCN1A subunit, mice develop spontaneous seizures within 10 days.

[0209] B6(Cg)-*Scn1a*^{tm1.1Dsf/J} and control C57Bl/6 mice were injected, as in Example 2, with AAVs expressing CRE recombinase under the control of the EF1 α promoter and an AAV comprising PV cell selective regulatory element SEQ ID NO: 32 driving expression of either eGFP (SEQ ID NO: 36) or SCN1B (SEQ ID NO: 37). Once all four infusions were complete, telemetry implantation was performed immediately, (F20-EET, Data Sciences International). Electrocorticogram data was monitored continuously for 14 days from 10 days after the surgery. Electrocorticogram data was analyzed and all seizure events were recorded, annotated with date, time start, time stop, duration of the seizures, and severity score. **FIG. 1** illustrates the frequency of seizures in 12 hour windows over 14 days following treatment. The mice treated with SCN1B showed a trend towards lower seizure frequency compared to the control animals.

[0210] This observation was consistent with the notion that the beta unit of the sodium ion channel, e.g., SCN1B, can contribute to the trafficking and assembly of the sodium ion channel and that increasing the expression of the beta unit selectively in PV neurons can

result in increased trafficking and assembly of the Nav1.1 channel, thus leading to a trend towards lower seizure frequency and duration in the mice treated with SCN1B gene therapy.

EXAMPLE 4

Treating Alzheimer's Disease in a Mouse Model

[0211] Female APP/PS1 and WT mice bred at PSYCHOGENICS® were used in the study. APP/PS1 mice contain human transgenes for both Amyloid Beta Precursor Protein (APP) bearing the Swedish mutation (670 G-T and 671 A-C) and Presenilin 1 (PSEN1) containing an L166P mutation, both under the control of the Thy1 promoter. These mice develop symptoms of Alzheimer's disease, including amyloid plaques and memory defects. Further description of these mice can be found in Radde et al., 2006 (Radde, Rebecca, et al., "A β 42-driven cerebral amyloidosis in transgenic mice reveals early and robust pathology." EMBO reports 7.9 (2006): 940-946).

[0212] APP/PS1 mice were used as a model to determine the effect of treatment with SCN1B under the control of a RE on symptoms of Alzheimer's disease. APP/PS1 mice and non-transgenic controls were injected with either a control vector expressing eGFP or a treatment vector expressing SCN1B, both under the control of SEQ ID NO: 32; and implanted with an EET transmitter as in Example 3. Brain activity was assessed over 24 hours at 4 weeks after surgery. Electrocorticogram data was automatically analyzed and power levels in the different frequency bands were compared. **FIG. 2** illustrates the high gamma power (50-100Hz) in non-transgenic controls (WT), APP/PS1, and APP/PS1 mice treated with SCN1B. Increased high gamma power activity is associated with seizures in Alzheimer's patients and epilepsy patients. The APP/PS1 mice showed a higher level of high gamma power activity than the control mice. However, the increase was absent in the treated mice indicating effective treatment with the vector.

EXAMPLE 5

Selectivity for PV Neurons in C57BL/6J (WT) Mouse

[0213] The selectivity of various REs disclosed herein were tested for selective gene expression in PV neurons using immunohistochemical methods. C57BL/6J (WT) mouse line was used for the PV immunohistochemical assays. Expression cassettes comprising reporter transgene eGFP operably linked to a regulatory element (SEQ ID NO: 1 or SEQ ID NO: 8) or a CAG promoter in an AAV9 construct.

[0214] Pup systemic infusions: Postnatal day 1 C57BL/6J mice were infused via facial vein injection with AA9 vector (1×10^{12} to 3×10^{12}) using a 300 U insulin syringe with a 31 G needle. For tissue collection, mice were euthanized 21 days post-infusion via overdose of sodium pentobarbital (i.p.) and perfused with heparinized (2.5 IU/ml) saline followed by perfusion with 4% formaldehyde. Brains were removed and subsequently immersion-fixed in 4% formaldehyde for 24-48 hours at 4 degrees Celsius. The brain was then placed into PBS containing 30% sucrose and allowed to sink at 4 degrees Celsius (~2-3 days). Upon sinking the individual brain hemispheres were frozen in TISSUE-TEK® OCT with the midline facing down. Frozen brains were processed for sagittal sections on a cryostat and placed free-floating into PBS. Sections were stained for eGFP and parvalbumin (PV) using standard immunohistochemistry procedures with chicken anti-GFP (Aves Lab, GFP-1020) and mouse anti-PV (Sigma, P3088).

[0215] Adult systemic infusions: 4-week-old, C57BL/6 mice were infused via tail vein injection with 60µL of AAV9 vector (4.9×10^{13} to 1×10^{14} gc/ml) expressing eGFP. For tissue collection mice were euthanized 21 days post-infusion via isoflurane overdose and whole brains were extracted, washed with PBS and placed into separate 5ml tubes containing ice cold 4% formaldehyde. Tissue was fixed at 4 degrees Celsius overnight. The following day, the brain was placed into PBS containing 30% sucrose and allowed to sink at 4 degrees Celsius. Upon sinking the individual brain hemispheres were frozen in Tissue-Tek OCT with the midline facing down. Frozen brains were processed for sagittal sections on a cryostat and placed free-floating into PBS. Sections were stained for EGFP and parvalbumin (PV) using standard immunohistochemistry procedures with chicken anti-GFP (Aves Lab, GFP-1020) and mouse anti-PV (Sigma, P3088).

[0216] Immunohistochemistry protocol: Immunohistochemistry was used to analyze the co-localization of eGFP signal and PV signal using the anti-PV antibody, wherein overlay of the signals exhibited as white or light gray spots in the top panel images (merge), wherein representative overlay was indicated by arrowheads. Overlay of the eGFP and PV fluorescence is indicative of expression in PV cells. Such experiments can be used to determine the selective expression of expression in PV cells. To perform the immunohistochemical experiments, tissues obtained from each mouse were blocked with a Blocking Buffer Solution (comprising 3% BSA, 3%NGS, 0.3% Triton X-100, 0.2% Tween-20 in 1X PBS) for 1 hour at room temperature. The tissues were then incubated with primary antibodies in blocking buffer overnight at 4C, washed three times with 1mL 1X PBS, each with 5 minutes interval. Then the tissues were incubated with secondary antibodies in

blocking buffer for 1 hour at room temperature, followed by washing three times, each time with 1mL 1X PBS and with 5 minutes interval. The tissues were incubated DAPI (1:1000) in PBS buffer for 5 minute and wash twice with 1mL 1X PBS. Tissues were mounted onto slides, imaged, and analyzed using a fluorescence microscope. Images were taken using a Vectra 3 imaging system (Perkin Elmer) and quantified for co-labeling of eGFP and PV staining using inform-Tissue finder, advanced image analysis software or hand scored. At least 80 GFP positive cells were counted in each panel before determining the percentage of co-localization.

[0217] FIGS. 3A-3C illustrates the results of the immunohistochemistry experiments performed in pups after systemic AAV9 injections. **FIGS. 4A-4C** illustrates results of similar immunohistochemistry experiments performed in adult mice following AAV9 injections.

[0218] FIG. 3A illustrates the overlay of the immunohistochemistry experiments performed in pups after systemic AAV9 injections. **FIG. 3B** illustrates the quantification of the co-localization of the immunohistochemistry experiments, wherein selectivity for PV cells was measured as percentage of GFP+ cells that were also PV+, as compared to eGFP expression under the control of the CAG promoter.

[0219] FIG. 4A illustrates the overlay of the immunohistochemistry experiments performed in adult mice after systemic AAV9 injections. **FIG. 4B** illustrates the quantification of the co-localization of the immunohistochemistry experiments, wherein selectivity for PV cells was measured as percentage of GFP+ cells that were also PV+, as compared to eGFP expression under the control of the EF1 α .

[0220] It is estimated that GABAergic neurons constitute about 20% of CNS, while PV cells constitute about 40% of GABAergic neurons, which means that PV cells make up approximately 8% of all neurons in the CNS. See, Pelkey, KA et al., 2017; and Lee, S. et al., 2010. Thus, one would predict that about 8% of the cells labeled by a non-selective regulatory element (e.g., CAG, EF1 α , or a constitutive promoter) would be PV positive, or within this range. Therefore, expression in PV cells above 8% is indicative of increased selectivity in PV cells. Notably, AAV9 injections comprising regulatory element SEQ ID NO: 8 resulted in about 60% of cells as PV positive, which was 7.5 times higher than what was expected by the distribution of PV cells.

[0221] Similar immunohistochemistry experiments as described above were performed to determine the selective expression of additional regulatory elements, SEQ ID NOs: 2-7 and 9-22 as compared to a non-selective regulatory element having a sequence of SEQ ID NO: 34, except AAVDJ viral vector was used to deliver eGFP operably linked to a regulatory

element into C57BL/6J (WT) mice. Such AAVDJ virus was injected directly into the CNS in the hippocampus of adult mice. At least 80 GFP positive cells were counted in each experiment before calculating the percentage of co-localization, or selectivity, as percentage of GFP positive cells that were also PV positive. **FIGS. 5A-5F** illustrate the fluorescence imaging used for determining co-localization, or selectivity, measured as percentage of eGFP positive cells that were also PV positive and in comparison to the signal of non-selective regulatory element SEQ ID NO: 34. Cells that were positive for a marker appear as white/gray cells in the images. Merge images illustrate the overlap between the corresponding eGFP and anti-PV images. Cells that were positive for both eGFP and PV appear as white/light gray cells in the merge image. **FIG. 6** illustrates the quantification of the co-localization analysis, measured as percentage of eGFP+ cells that were also PV+.

EXAMPLE 6

Treatment of Dravet Syndrome in Different Mouse Lines

[0222] Treatment of Dravet syndrome and/or symptoms thereof using the expression cassettes described herein can be tested in various mouse lines, such as B6(Cg)-*Scn1a*^{tm1.1Dsf/J} as described above, *Scn1a*^{tm1Kea}, and *Scn1a*-R1470X mouse lines. These mouse lines are established mouse models for Dravet syndrome. *Scn1a*^{tm1Kea} and *Scn1a*-R1470X mouse lines do not require CRE recombinase.

[0223] The *Scn1a*^{tm1Kea} mouse (available from the Jackson Laboratory; described in Hawkins et al., Scientific Reports, vol. 7: 15327 (2017)) comprises a deletion of the first coding exon of SCN1A. Mice homozygous for the SCN1A knockout allele are characterized by tremors, ataxia, seizures, and die by postnatal day 16. Heterozygous mice on the C57BL/6 background develop spontaneous seizures and die within weeks. Such mouse strain can be used to study safety and efficacy of treatment of epilepsy and Dravet syndrome. See, Miller et al., Genes Brain Behav. 2014 Feb;13(2):163-72 for additional information.

[0224] The *Scn1a*-R1470X mouse is a knock-in mouse carrying a premature stop codon, R1407X, in exon 21 of the SCN1A gene. The same mutation has been identified as a pathogenic mutation in three unrelated SMEI patients. *Scn1a*^{RX/RX} pups are characterized by recurrent spontaneous seizures at 12 postnatal days, including tonic-clonic and clonic seizures at 12–16 postnatal days, and rhythmic jerking movements and involuntary muscle contraction. See, Ogiwara et al., Journal of Neuroscience, May 30, 2007, 27 (22) 5903-5914 for additional information.

[0225] To test the compositions described herein, such as AAV gene therapy and treatment using such gene therapy, Dravet mice of each of the mouse strains described above and control mice (e.g., a wild-type mouse or an untreated Dravet mouse for the strain) are injected (e.g., administered by intraperitoneal injection) with AAVs expressing either eGFP or another reporter gene, or an expression cassette comprising one or more PV-selective REs (e.g., SEQ ID NOs: 1-32) as described herein operably linked to a transgene disclosed herein, such as SCN1A, SCN1B, or SCN2B, or any of SEQ ID NOs: 37-39, or a variant or functional fragment thereof. Following AAV injections, mouse survival is monitored over time. All mice are monitored daily for general health (e.g. weight, hydration, grooming, and mobility) and deaths were recorded. Telemetry implantation can be performed immediately after AAV injections (F20-EET, Data Sciences International). Electrocorticogram data can be recorded and monitored continuously for at least 14 days from 10 days after the surgery. All seizure events can be recorded for at least 14 days following AAV treatment, annotated with date, time start, time stop, duration, and severity score. A reduction in the frequency and/or duration of seizures following treatment with an AAV as described above as compared to the eGFP control or an untreated control is indicative of the efficacy of the gene therapy in reducing the symptoms and/or severity of Dravet syndrome.

[0226] After treatment of the mice with AAV, the expression levels of the transgene (e.g., SCN1A; SCN1B; SCN2B; a DNA binding protein, such as a transcriptional activator, that modulates an endogenous SCN1A, SCN1B, or SCN2B; any of SEQ ID NOs: 37-39; or any variant or functional fragment thereof) can be monitored over time using various PCR and/or sequencing methods to show AAV treatment can result in an increase in gene expression in PV cells. Northern blot analysis and in situ hybridization can also be used to analyze transgene expression in vivo. The level of the protein expressed from the protein can also be monitored after treatment to show an increase in transgene expression correlates with an increase in the corresponding protein in vivo. Protein levels can be assayed using various methods, including, but not limited to, Western blot analysis, immunohistochemistry, immunofluorescence histochemistry, and/or ELISA assays. Formation of functional voltage-gated sodium ion channels can also be assayed using current-clamp analysis.

[0227] Hyperthermia-induced seizures can be evaluated to compare the wild-type mice and/or untreated Dravet mice with Dravet mice treated with AAV gene therapy comprising an expression cassette described herein (e.g., an expression cassette comprising one or more REs of this disclosure operably linked to a transgene of this disclosure, such as SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, a functional fragment thereof, or a DNA binding protein

that modulates an endogenous SCN1A, SCN2A, SCN8A, SCN1B, or SCN2B). In such experiments, the core body temperature is monitored with a RET-3 rectal temperature probe (Physitemp Instruments, Inc., New Jersey, USA) and controlled by a heat lamp connected to a rodent temperature regulator (TCAT-2DF, Physitemp Instruments, Inc.) reconfigured with a Partlow 1160 + controller (West Control Solutions, Brighton, UK). Body temperature is raised 0.5 °C every two minutes until the onset of the first clonic convulsion. As compared to the untreated Dravet mice, Dravet mice treated with an AAV gene therapy are expected to have a higher threshold temperature before the onset of first clonic convulsion and/or have a higher proportion of mice that remain seizure free at the maximum temperature tested.

[0228] Different doses of AAV comprising an expression cassette can also be administered to mice to determine the safety and efficacy profile of each gene therapy treatment. These preclinical studies can also inform the optimal dose(s) of the gene therapy to use for treating Dravet syndrome.

EXAMPLE 7

Treatment of Alzheimer's Disease in Mouse

[0229] Female APP/PS1 and wild-type (WT) mice, which are bred at PSYCHOGENICS® and are established mouse model of Alzheimer's disease, can be used to study the safety and efficacy of the compositions described herein in treating Alzheimer's disease, comprising one or more PV-selective REs. APP/PS1 mice is describe above in Example 4.

[0230] APP/PS1 mice and non-transgenic controls are injected with either a control AAV vector expressing eGFP or a treatment AAV vector comprising one or more PV-selective REs disclosed herein, e.g., SEQ ID NOs: 1-32, operably linked to a transgene that is deficient or impaired in Alzheimer's disease, such as SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.3, STXBP1, a DNA binding protein that modulates an endogenous gene (e.g., SCN1A, SCN2A, SCN8A, SCN1B, SCN2B, KV3.1, KV3.3, or STXBP1), or any one of SEQ ID NOs: 37-43, or a functional fragment thereof.

[0231] Following AAV injections, mouse survival is monitored over time. All mice are monitored daily for general health (e.g. weight, hydration, grooming, and mobility) and deaths were recorded. After injections of the AAVs, mice are also implanted with an EET transmitter as described in Example 3 above. Brain activity can be recorded and monitored over 24 hours for at least 4 weeks after surgery. Electrocorticogram data can be automatically analyzed, and power levels in the different frequency bands (50-100Hz) can be compared across different groups: WT mice, untreated APP/PS1 mice, and AAV-treated APP/PS1

CLAIMS

WHAT IS CLAIMED IS:

1. A nucleic acid cassette comprising a parvalbumin (PV) neuron selective regulatory element operably linked to a heterologous transgene, wherein the regulatory element is less than 2.5 kb and comprises SEQ ID NO: 1 or SEQ ID NO: 8.
2. The nucleic acid cassette of claim 1, wherein the heterologous transgene comprises a nucleic acid sequence encoding a DNA binding protein.
3. The nucleic acid cassette of claim 2, wherein the DNA binding protein is a transcriptional modulator of an endogenous gene.
4. The nucleic acid cassette of claim 2, wherein the DNA binding protein comprises a zinc finger.
5. The nucleic acid cassette of claim 2, wherein the DNA binding protein is a nuclease-deactivated zinc finger protein.
6. The nucleic acid cassette of claim 2, wherein the DNA binding protein is linked to a transcriptional activator domain.
7. The nucleic acid cassette of claim 2, wherein the DNA binding protein is linked to a transcriptional repressor domain.
8. The nucleic acid cassette of claim 2, wherein the DNA binding protein is a transcriptional activator that modulates an endogenous SCN1A gene.
9. The nucleic acid cassette of claim 1, wherein the nucleic acid cassette is an adeno-associated virus (AAV) vector.
10. The nucleic acid cassette of claim 9, wherein the AAV vector is an AAV9 or a scAAV9.
11. The nucleic acid cassette of claim 8, wherein the nucleic acid cassette is an adeno-associated virus (AAV) vector.
12. The nucleic acid cassette of claim 11, wherein the AAV vector is an AAV9 or a scAAV9.
13. The nucleic acid cassette of claim 1, wherein the PV neuron selective regulatory element comprises SEQ ID NO: 31 or SEQ ID NO: 32.

14. The nucleic acid cassette of claim 13, wherein the heterologous transgene comprises a nucleic acid sequence encoding a DNA binding protein.
15. The nucleic acid cassette of claim 14, wherein the DNA binding protein is a transcriptional activator that modulates an endogenous SCN1A gene.
16. The nucleic acid cassette of claim 13, wherein the nucleic acid cassette is an adeno-associated virus (AAV) vector.
17. The nucleic acid cassette of claim 16, wherein the AAV vector is an AAV9 or a scAAV9.
18. The nucleic acid cassette of claim 15, wherein the nucleic acid cassette is an adeno-associated virus (AAV) vector.
19. The nucleic acid cassette of claim 18, wherein the AAV vector is an AAV9.
20. The nucleic acid cassette of claim 18, wherein the AAV vector is a scAAV9.
21. A nucleic acid cassette comprising a parvalbumin (PV) neuron selective regulatory element of less than 2.5 kb operably linked to a heterologous transgene, wherein the regulatory element comprises a sequence with at least 95% sequence identity to SEQ ID NO: 1 or SEQ ID NO: 8, , and wherein the PV neuron selective regulatory element increases selective expression of the heterologous transgene in parvalbumin (PV) neurons in a central nervous system (CNS).
22. The nucleic acid cassette of claim 21, wherein the heterologous transgene comprises a nucleic acid sequence encoding a DNA binding protein.
23. The nucleic acid cassette of claim 22, wherein the DNA binding protein is a transcriptional modulator of an endogenous gene.
24. The nucleic acid cassette of claim 22, wherein the DNA binding protein comprises a zinc finger.
25. The nucleic acid cassette of claim 22, wherein the DNA binding protein is linked to a transcriptional activator domain.
26. The nucleic acid cassette of claim 22, wherein the DNA binding protein is a transcriptional activator that modulates an endogenous SCN1A gene.
27. The nucleic acid cassette of claim 21, wherein the nucleic acid cassette is an adeno-associated virus (AAV) vector.

28. The nucleic acid cassette of claim 27, wherein the AAV vector is an AAV9 or a scAAV9.

29. The nucleic acid cassette of claim 21, wherein the regulatory element comprises a sequence of SEQ ID NO: 31 or SEQ ID NO: 32.

30. The nucleic acid cassette of claim 29, wherein the heterologous transgene comprises a nucleic acid sequence encoding a DNA binding protein.

31. The nucleic acid cassette of claim 30, wherein the DNA binding protein is a transcriptional activator that modulates an endogenous SCN1A gene.

32. The nucleic acid cassette of claim 29, wherein the nucleic acid cassette is an adeno-associated virus (AAV) vector.

33. The nucleic acid cassette of claim 32, wherein the AAV vector is an AAV9 or a scAAV9.

34. The nucleic acid cassette of claim 31, wherein the nucleic acid cassette is an adeno-associated virus (AAV) vector.

35. The nucleic acid cassette of claim 34, wherein the AAV vector is an AAV9 or a scAAV9.

36. The nucleic acid cassette of claim 1, wherein the regulatory element comprises SEQ ID NO: 1.

37. The nucleic acid cassette of claim 1, wherein the regulatory element comprises SEQ ID NO: 8.

38. The nucleic acid cassette of claim 21, wherein the regulatory element comprises a sequence that is at least 95% identical to SEQ ID NO: 1.

39. The nucleic acid cassette of claim 21, wherein the regulatory element comprises a sequence that is at least 95% identical to SEQ ID NO: 8.