SERUM-FREE CELL CULTURE MEDIUM

FIELD

[0001] The invention relates to media for the culturing of cells and for the production of recombinant proteins. The invention specifically relates to serum-free media for the culturing of recombinant CHO cells for the production of protein biotherapeutics.

BACKGROUND

[0002] Cell culture media comprising serum or protein hydrolysate components (i.e., peptones and tryptones) have a long history of use in the production of recombinant proteins from cultured cells. These components contain growth factors and a wide variety of other uncharacterized elements beneficial to cell growth and culture. However, they also contain uncharacterized elements that reduce growth or otherwise negatively impact recombinant protein production. They can also be an unwelcome potential source of variability. Despite their drawbacks, the benefits of using sera and hydrolysates have outweighed some of the disadvantages and they have been widely used in many cell culture applications.

[0003] Human biological therapeutics (biopharmaceuticals) are generally produced in mammalian cell culture, particularly CHO cell culture. The presence of uncharacterized or partly characterized components in those cell cultures are highly undesirable for the manufacture of biopharmaceuticals for human use. The use of such uncharacterized or partly characterized components not only introduces production and regulatory inconsistencies, it raises the possibility of viral or fungal infection of the production culture.

[0004] The reduction of lot-to-lot variability in drug product yield and composition is another important factor in selecting a culturing process. Sera, hydrolysates, and other undefined elements introduce variability in the yield, composition, and quality of biopharmaceutical production lots. The quality and purity of media elements may also affect yield, since drug titers often rely in part on maintaining a particular balance of nutrients. Where the relative amounts of nutrients vary from one media lot to another, drug yield can vary and this variance can be unacceptable or uneconomical.

[0005] Using serum containing or hydrolysate-based media introduces downstream processing challenges. The concentration of a desired biopharmaceutical in culture is generally on the order of grams per liter. The presence of serum and hydrolysates in media can add more than 10 g/L of uncharacterized peptides and proteins, which must be removed in subsequent processing steps. Serum and hydrolysates can also introduce variability in the amount of metals and other trace elements in the media. The elimination of serum and hydrolysates from culture media therefore eliminates these variations and potential encumbrances to the production and processing of drug substance.

[0006] Among others, the benefits to using serum-free and hydrolysate-free media include reduction in cost, reduction in variablity between drug lots, and minimization of the risk of introducing adventitious agents from undefined and unrefined components. Furthermore, where media are defined and uniform between batch runs, qualifying runs to test new media batches against current media are likewise minimized. Thus, there is a need in the art for media for culturing mammalian cells, wherein the media are chemically-defined and free of sera and hydrolysates, or that are serum-free and contain low manageable levels of hydrolysates, and yet allow for healthy and robust cell growth and maintenance, and high-titer production of biopharmaceutical drug substance.

SUMMARY

[0007] The inventors have made the surprising discovery that the inclusion of ornithine, either with or without putrescine, in cell culture media that is free of sera ("OS" media) increases cell viability and density, reduces cell doubling time, and permits high titer protein production by those cells. The inventors have also discovered that OS media that contains low or trace amounts of protein hydrolysates or is chemically defined (*i.e.,* contains no protein hydrolysates) in particular provides restored cell viability and density, cell doubling time, and high titer protein production.

[0008] In one aspect, the invention provides a cell culture medium, which is serum-free and comprises at least 0.09 mM $\pm$ 0.014 mM ornithine. In one embodiment the ornithine is present in the medium at a concentration ranging from 0.09 $\pm$ 0.014 mM to 0.9 $\pm$ 0.14 mM, such

as 0.09 ± 0.014 mM, 0.3 ± 0.05 mM, 0.6 ± 0.09 mM, or 0.9 ± 0.14 mM ornithine. In some embodiments, the medium also contains at least 0.20 ± 0.03 mM putrescine. In some embodiments, the additional putrescine is at a concentration ranging from 0.20 ± 0.03 mM to 0.714 ± 0.11 mM, such as 0.20 ± 0.03 mM, 0.35 ± 0.06, or 0.714 ± 0.11 mM putrescine. In some embodiments, the medium contains ≤ 7.5 g/L hydrolysate. In some embodiments, the medium is free of any hydrolysate.

[0009] In one embodiment, the medium contains a base medium that is chemically defined, such as a custom formulation or a commercially available base medium. In one embodiment, the complete medium is chemically defined, free of sera and free of hydrolysate.

[00010] In some embodiments, the medium, which is at its useful concentration (*i.e.,* 1X) contains at least 40 ± 6 mM or at least 70 ± 10.5 mM of a mixture of amino acids or amino acid salts. In one embodiment, the medium contains at least 40 mM of a mixture of amino acids. In this or another embodiment, the medium contains at least 70 mM of a mixture of amino acids. In one embodiment, the mixture of amino acids (with the notable exception of glutamine, which may be added back to the medium as a point of use addition) contains alanine, arginine, asparagine, aspartic acid, cysteine, glutamic acid, glycine, histidine, isoleucine, leucine, lysine, methionine, phenylalanine, proline, serine, threonine, tryptophan, tyrosine, and valine.

[00011] In some embodiments, the medium contains one or more fatty acids. In one particular embodiment, the medium contains a mixture of fatty acids (or fatty acid derivatives) and alpha tocopherol. Fatty acids or fatty acid derivatives are selected from the group consisting of linoleic acid, linolenic acid, thioctic acid, oleic acid, palmitic acid, stearic acid, arachidic acid, acid, lauric acid, behenic acid, decanoic acid, dodecanoic acid, hexanoic acid, lignoceric acid, myristic acid, and octanoic acid.

[00012] In some embodiments, the medium contains a mixture of nucleosides. In one embodiment, the medium contains adenosine, guanosine, cytidine, uridine, thymidine, and hypoxanthine.

[00013] In some embodiments, the medium contains a mixture of salts. Salts include divalent cations, such as calcium and magnesium. In one embodiment, the medium contains calcium chloride and magnesium sulfate. Other salts may include those of phosphate.

[00014] In a specific embodiment, the medium (1) contains ≤ 7.5 g/L of a hydrolysate, (2) is serum-free, (3) contains 0.09 ± 0.014 mM, 0.3 ± 0.05 mM, 0.6 ± 0.09 mM, or 0.9 ± 0.14 mM ornithine, (4) optionally additionally contains 0.20 ± 0.03 mM, 0.35 ± 0.06, or 0.714 ± 0.11 mM putrescine, (5) contains at least about 40 mM or at least about 70 mM of a mixture of amino acids including alanine, arginine, asparagine, aspartic acid, cysteine, glutamic acid, glycine, histidine, isoleucine, leucine, lysine, methionine, phenylalanine, proline, serine, threonine, tryptophan, tyrosine, and valine, (6) contains tocopherol and a mixture of fatty acids, (7) contains a mixture of nucleosides including adenosine, guanosine, cytidine, uridine, thymidine, and hypoxanthine, and (8) contains salts of calcium, magnesium, and phosphate.

[00015] In another aspect, the invention provides a method for cultivating cells in a cell culture medium, such as any embodiment of the medium described in the foregoing aspect. In one embodiment, the method employs the steps of propagating or maintaining a cell or cells in a medium that (1) contains ≤ 7.5 g/L hydrolysate, or no hydrolysate, (2) is free of sera, (3) contains ornithine at a concentration of at least 0.09 mM ± 0.014 mM, (4) and optionally contains putrescine, such as at least 0.20 ± 0.03 mM.

[00016] In some embodiments, the cell or cells are mammalian cells, avian cells, insect cells, yeast cells, or bacteria cells. In one embodiment, the cells are mammalian cells useful in the production of recombinant proteins, such as CHO cells or the derivative CHO-K1. In some embodiments, the cells express a protein of interest, such as a biotherapeutic protein. The biotherapeutic protein may be an antigen binding protein, which may contain an Fc domain. In some embodiments, the protein of interest is a receptor-Fc-fusion protein, such as an ScFv molecule or a trap molecule. Trap molecules include the VEGF trap and IL-1 Trap proteins. In some embodiments, the protein of interest is an antibody, such as a humanized monoclonal antibody, a bispecific antibody, or an antibody fragment.

[00017] Given the positive effects on cell growth by including ornithine or a combination of ornithine and putrescine in serum-free media, the cells cultured according to this method have an average doubling time that is no more than 30 hours. In one embodiment, the cell doubling time is no more than 24 hours. In one embodiment, when compared to cell growth in media that contains less than 0.09 ± 0.014 mM ornithine (or less than 0.09 ± 0.014 mM ornithine and less than 0.2 ± 0.03 mM putrescine), the cells grown according to this method have an average doubling time that is at least one third the doubling time of the comparator control culture.

[00018] Likewise, the inclusion of ornithine alone or a combination of ornithine and putrescine in serum-free media allows cultured cells to reach a higher viable cell count density than without the inclusion of ornithine or the combination of ornithine and putrescine. In one serum-free and hydrolysate-free embodiment of the OS medium, the cell culture is capable of attaining a viable cell count density that is at least 15% greater than a similar cell culture in a similar cell culture medium that contains less than 0.09 ± 0.014 mM ornithine (or less than 0.09 ± 0.014 mM ornithine and less than 0.2 ± 0.03 mM putrescine). In another serum-free and hydrolysate-free embodiment of the OS medium, the cell culture is capable of attaining a viable cell count density that is at least 3-fold greater than a similar cell culture in a similar cell culture medium that contains less than 0.09 ± 0.014 mM ornithine (or less than 0.09 ± 0.014 mM ornithine and less than 0.2 ± 0.03 mM putrescine).

[00019] In another embodiment, the method includes the step of adding one or more point-of-use additions to the cell culture medium. In some embodiments, the point-of-use addition is any one or more of $NaHCO_3$, glutamine, insulin, glucose, $CuSO_4$, $ZnSO_4$, $FeCl_3$, $NiSO_4$, Na_4 EDTA, and Na_3 Citrate. In one embodiment, the method employs the step of adding each of the following point-of-use chemicals to the cell culture medium: $NaHCO_3$, glutamine, insulin, glucose, $CuSO_4$, $ZnSO_4$, $FeCl_3$, $NiSO_4$, Na_4 EDTA, and Na_3 Citrate. In some embodiments, the point-of-use additions can be included in the medium at the outset.

[00020] In a specific embodiment, the aspect provides a method for cultivating cells in a serum-free medium which contains (1) ornithine at either 0.09 ± 0.014 mM, 0.3 ± 0.05 mM, 0.6 ± 0.09 mM, or 0.9 ± 0.14 mM cell culture medium; (2) optionally additionally putrescine at either

0.20 ± 0.03 mM, 0.35 ± 0.06, or 0.714 ± 0.11 mM; (3) at least about 40 mM or at least about 70 mM of a mixture of amino acids including alanine, arginine, asparagine, aspartic acid, cysteine, glutamic acid, glycine, histidine, isoleucine, leucine, lysine, methionine, phenylalanine, proline, serine, threonine, tryptophan, tyrosine, and valine; (4) tocopherol and a mixture of fatty acids; (6) a mixture of nucleosides including adenosine, guanosine, cytidine, uridine, thymidine, and hypoxanthine, and (9) salts of calcium, magnesium, and phosphate, wherein the cells cultured according to this method have an average doubling time that is no more than 24 hours or at least one third the doubling time of the comparator control culture; and the cells cultured are capable of attaining a viable cell count density that is at least 15% greater or at least 3-fold greater than a similar cell culture in a similar cell culture medium that contains less than 0.09 ± 0.015 mM ornithine (or less than 0.09 ± 0.014 mM ornithine and less than 0.2 ± 0.03 mM putrescine). In another embodiment, the cell culture is capable of attaining a viable cell count density that is at least 3-fold greater than a similar cell culture in a similar cell culture medium that contains less than 0.09 ± 0.014 mM ornithine (or less than 0.09 ± 0.014 mM ornithine and less than 0.2 ± 0.03 mM putrescine). In one embodiment, the medium contains ≤ 7.5 g/L hydrolysate; and in another embodiment, free of hydrolysates.

[00021] In another aspect, the invention provides a method for producing a protein of interest by employing the steps of (1) introducing into a cell a nucleic acid sequence that encodes a protein of interest; (2) selecting a cell carrying that nucleic acid sequence; (3) culturing the selected cell in an embodiment of the serum-free cell culture medium described in the first aspect or according to any embodiment of the method described in the second aspect; and (4) expressing the protein of interest in the cell, wherein the protein of interest is secreted into the medium. In some embodiments, the cell used in the production of the protein is a mammalian cell capable of producing a biotherapeutic, such as CHO, 293, and BHK cell, or any derivatives of them. In one embodiment, the cell is a CHO cell, such as a CHO-K1 cell.

[00022] In some embodiments the protein of interest is an antigen binding protein. In some embodiments, the protein of interest is a protein that has an Fc domain. In some cases, those two proteins of interest may overlap, such as in the case of a receptor-Fc-fusion protein, an antibody, and a ScFv protein for example. Thus, in some embodiments, the protein of

interest is an antibody, such as a human antibody or a humanized antibody, an antibody fragment, such as an Fab or $F(ab')_2$, a bispecific antibody, a trap molecule, such as a VEGF-Trap or an IL-1-Trap, an ScFv molecule, a soluble TCR-Fc fusion protein, or the like.

[00023] In one embodiment, the protein of interest is capable of being produced at an average seven day titer that is at least 7% greater, at least 14% greater, at least 80% greater, at least two fold greater, or at least three fold greater than the average seven day titer produced by a similar cell in a serum-free cell culture medium that contains less than 0.09 ± 0.014 mM ornithine (or less than 0.09 ± 0.014 mM ornithine and less than 0.2 ± 0.03 mM putrescine) ("non-OS" media).

[00024] In a specific embodiment, the protein of interest is produced by (1) introducing into a CHO cell a nucleic acid sequence that encodes a protein of interest, such as an antibody or other antigen-binding protein; (2) selecting a cell carrying that nucleic acid sequence; (3) culturing the selected cell in a serum-free cell culture medium which contains (a) ornithine at either 0.09 ± 0.014, 0.3 ± 0.05 mM, 0.6 ± 0.09 mM, or 0.9 ± 0.14 mM; (b) optionally additionally putrescine at either 0.20 ± 0.03 mM, 0.35 ± 0.06, or 0.714 ± 0.11 mM; (c) at least 40 mM or at least 70 mM of a mixture of amino acids including: alanine, arginine, asparagine, aspartic acid, cysteine, glutamic acid, glycine, histidine, isoleucine, leucine, lysine, methionine, phenylalanine, proline, serine, threonine, tryptophan, tyrosine, and valine; (d) tocopherol and a mixture of fatty acids; (e) a mixture of nucleosides including adenosine, guanosine, cytidine, uridine, thymidine, and hypoxanthine, and (f) salts of calcium, magnesium, and phosphate; and (d) expressing the protein of interest in the CHO cell, wherein the protein of interest is secreted into the medium. In some embodiments, the serum-free cell culture medium may include ≤ 7.5 g/L hydrolysates; or in other embodiments no hydrolysates at all.

DETAILED DESCRIPTION

[00025] The applicants have made the surprising discovery that the addition of ornithine, or a combination of ornithine and putrescine ("OS medium") improves viable cell density, cell doubling time, and protein production by a cell in a cell culture relative to a serum-free medium that contains very little or no ornithine or little or none of a combination of ornithine and putrescine ("non-OS medium").

[00026] Before the present cell cultures and methods are described, it is to be understood that this invention is not limited to particular methods and experimental conditions described, as such methods and conditions may vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to be limiting.

[00027] Unless defined otherwise, all technical and scientific terms used in this application have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any methods and materials similar or equivalent to those described in this application can be used in the practice or testing of the present invention, certain specific methods and materials are now described. Units, prefixes, and symbols may be denoted in their SI accepted form. Numeric ranges recited herein are open-bracketed, meaning that they include the numbers defining the range. Unless otherwise noted, the terms "a" or "an" are to be construed as meaning "at least one of". The section headings used herein are for organizational purposes only and are not to be construed as limiting the subject matter described. The methods and techniques described herein are generally performed according to conventional methods known in the art and as described in various general and more specific references that are cited and discussed throughout the present specification unless otherwise indicated. See, e.g., Sambrook et al., Molecular Cloning: A Laboratory Manual, 3rd ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N. Y. (2001) and Ausubel et al., Current Protocols in Molecular Biology, Greene Publishing Associates (1992), Harlow and Lane Antibodies: A Laboratory Manual, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. (1990), and Julio E. Celis, Cell Biology: A Laboratory Handbook, 2nd ed., Academic Press, New York, N.Y. (1998), and Dieffenbach and

Dveksler, PCR Primer: A Laboratory Manual, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. (1995). All publications mentioned throughout this disclosure are incorporated herein by reference in their entirety.

DEFINITIONS

[00028] As used herein "peptide," "polypeptide" and "protein" are used interchangeably throughout and refer to a molecule comprising two or more amino acid residues joined to each other by a peptide bond. Peptides, polypeptides and proteins may also include modifications such as glycosylation, lipid attachment, sulfation, gamma-carboxylation of glutamic acid residues, alkylation, hydroxylation and ADP-ribosylation. Peptides, polypeptides, and proteins can be of scientific or commercial interest, including protein-based drugs. Peptides, polypeptides, and proteins include, among other things, antibodies and chimeric or fusion proteins. Peptides, polypeptides, and proteins are produced by recombinant animal cell lines using cell culture methods.

[00029] The term “heterologous polynucleotide sequence”, as used herein refers to nucleic acid polymers encoding proteins of interest, such as chimeric proteins (like trap molecules), antibodies or antibody portions (e.g., VH, VL, CDR3) that are produced as a biopharmaceutical drug substance. The heterologous polynucleotide sequence may be manufactured by genetic engineering techniques (*e.g.,* such as a sequence encoding a chimeric protein, or a codon-optimized sequence, an intronless sequence, *et cetera*) and introduced into the cell, where it may reside as an episome or be intergrated into the genome of the cell. The heterologous polynucleotide sequence may be a naturally occurring sequence that is introduced into an ectopic site within the production cell genome. The heterologous polypeptide sequence may be a naturally occurring sequence from another organism, such as a sequence encoding a human ortholog.

[00030] “Antibody” refers to an immunoglobulin molecule consisting of four polypeptide chains, two heavy (H) chains and two light (L) chains inter-connected by disulfide bonds. Each heavy chain has a heavy chain variable region (HCVR or VH) and a heavy chain constant region. The heavy chain constant region contains three domains, CH1, CH2 and CH3. Each

light chain has a light chain variable region and a light chain constant region. The light chain constant region consists of one domain (CL). The VH and VL regions can be further subdivided into regions of hypervariability, termed complementarity determining regions (CDR), interspersed with regions that are more conserved, termed framework regions (FR). Each VH and VL is composed of three CDRs and four FRs, arranged from amino-terminus to carboxy-terminus in the following order: FR1, CDR1, FR2, CDR2, FR3, CDR3, FR4. The term "antibody" includes reference to both glycosylated and non-glycosylated immunoglobulins of any isotype or subclass. The term "antibody" includes antibody molecules prepared, expressed, created or isolated by recombinant means, such as antibodies isolated from a host cell transfected to express the antibody. The term antibody also includes bispecific antibody, which includes a heterotetrameric immunoglobulin that can bind to more than one different epitope. Bispecific antibodies are generally described in US Patent Application Publication No. 2010/0331527, which is incorporated by reference into this application.

[00031] The term "antigen-binding portion" of an antibody (or “antibody fragment”), refers to one or more fragments of an antibody that retain the ability to specifically bind to an antigen. Examples of binding fragments encompassed within the term "antigen-binding portion" of an antibody include (i) a Fab fragment, a monovalent fragment consisting of the VL, VH, CL and CH1 domains; (ii) a F(ab')2 fragment, a bivalent fragment comprising two Fab fragments linked by a disulfide bridge at the hinge region; (iii) a Fd fragment consisting of the VH and CH1 domains; (iv) a Fv fragment consisting of the VL and VH domains of a single arm of an antibody, (v) a dAb fragment (Ward et al. (1989) Nature 241:544-546), which consists of a VH domain, (vi) an isolated CDR, and (vii) an scFv, which consists of the two domains of the Fv fragment, VL and VH, joined by a synthetic linker to form a single protein chain in which the VL and VH regions pair to form monovalent molecules. Other forms of single chain antibodies, such as diabodies are also encompassed under the term “antibody” (see e.g., Holliger et al. (1993) PNAS USA 90:6444-6448; Poljak et al. (1994) Structure 2:1121-1123).

[00032] Still further, an antibody or antigen-binding portion thereof may be part of a larger immunoadhesion molecule, formed by covalent or noncovalent association of the antibody or antibody portion with one or more other proteins or peptides. Examples of such

immunoadhesion molecules include use of the streptavidin core region to make a tetrameric scFv molecule (Kipriyanov et al. (1995) Human Antibodies and Hybridomas 6:93-101) and use of a cysteine residue, a marker peptide and a C-terminal polyhistidine tag to make bivalent and biotinylated scFv molecules (Kipriyanov et al. (1994) Mol. Immunol. 31:1047-1058). Antibody portions, such as Fab and F(ab')2 fragments, can be prepared from whole antibodies using conventional techniques, such as via papain or pepsin digestion of whole antibodies. Moreover, antibodies, antibody portions and immunoadhesion molecules can be obtained using standard recombinant DNA techniques commonly known in the art (see Sambrook *et al.,* 1989).

[00033] The term "human antibody" is intended to include antibodies having variable and constant regions derived from human germline immunoglobulin sequences. The human antibodies of the invention may include amino acid residues not encoded by human germline immunoglobulin sequences (e.g., mutations introduced by random or site-specific mutagenesis *in vitro* or by somatic mutation *in vivo*), for example in the CDRs and in particular CDR3. However, the term "human antibody", as used herein, is not intended to include antibodies in which CDR sequences derived from the germline of another mammalian species, such as a mouse, have been grafted onto human framework sequences.

[00034] The term "recombinant human antibody", as used herein, is intended to include all human antibodies that are prepared, expressed, created or isolated by recombinant means, such as antibodies expressed using a recombinant expression vector transfected into a host cell, antibodies isolated from a recombinant, combinatorial human antibody library, antibodies isolated from an animal (e.g., a mouse) that is transgenic for human immunoglobulin genes (see e.g., Taylor et al. (1992) Nucl. Acids Res. 20:6287-6295) or antibodies prepared, expressed, created or isolated by any other means that involves splicing of human immunoglobulin gene sequences to other DNA sequences. Such recombinant human antibodies have variable and constant regions derived from human germline immunoglobulin sequences. In certain embodiments, however, such recombinant human antibodies are subjected to *in vitro* mutagenesis (or, when an animal transgenic for human Ig sequences is used, *in vivo* somatic mutagenesis) and thus the amino acid sequences of the VH and VL regions of the recombinant

antibodies are sequences that, while derived from and related to human germline VH and VL sequences, may not naturally exist within the human antibody germline repertoire *in vivo*.

[00035] “Fc fusion proteins” comprise part or all of two or more proteins, one of which is an Fc portion of an immunoglobulin molecule, which are not otherwise found together in nature. Preparation of fusion proteins comprising certain heterologous polypeptides fused to various portions of antibody-derived polypeptides (including the Fc domain) has been described, *e.g.,* by Ashkenazi et al., Proc. Natl. Acad. ScL USA 88: 10535, 1991; Byrn et al., Nature 344:677, 1990; and Hollenbaugh et al., "Construction of Immunoglobulin Fusion Proteins", in Current Protocols in Immunology, Suppl. 4, pages 10.19.1 - 10.19.11, 1992. “Receptor Fc fusion proteins” comprise one or more extracellular domain(s) of a receptor coupled to an Fc moiety, which in some embodiments comprises a hinge region followed by a CH2 and CH3 domain of an immunoglobulin. In some embodiments, the Fc-fusion protein contains two or more distinct receptor chains that bind to a one or more ligand(s). For example, an Fc-fusion protein is a trap, such as for example an IL-1 trap (*e.g.,* rilonacept, which contains the IL-1RAcP ligand binding region fused to the IL-1R1 extracellular region fused to Fc of hIgG1; see U.S. Pat. No. 6,927,004), or a VEGF trap (*e.g.,* aflibercept, which contains the Ig domain 2 of the VEGF receptor Flt1 fused to the Ig domain 3 of the VEGF receptor Flk1 fused to Fc of hIgG1; see U.S. Pat. Nos. 7,087,411 and 7,279,159).

MEDIA

[00036] The present invention provides a serum-free medium that is useful in culturing cells and producing a biopharmaceutical drug substance. "Serum- free" applies to a cell culture medium that does not contain animal sera, such as fetal bovine serum. The serum-free media may contain ≤ 7.5 g/L of hydrolysates, such as soy hydrolysate. The present invention also provides chemically defined media, which is not only serum-free, but also hydrolysate-free. "Hydrolysate-free" applies to cell culture media that contains no exogenous protein hydrolysates such as animal or plant protein hydrolysates such, for example peptones, tryptones and the like.

[00037] The elimination of serum and reducing or eliminating hydrolysates from cell culture media, while reducing lot-to-lot variability and enhancing downstream processing steps,

unfortunately diminishes cell growth, viability and protein expression. Thus, chemically defined serum-free and low to no hydrolysate media requires additional ingredients to improve cell growth and protein production. The cell culture media of the invention may be supplemented with additional ingredients such as polyamines or increased concentrations of components like amino acids, salts, sugars, vitamins, hormones, growth factors, buffers, antibiotics, lipids, trace elements and the like, depending on the requirements of the cells to be cultured or the desired cell culture parameters. Specifically, the cell culture medium here is supplemented with ornithine, putrescine, or both ("OS media") to improve cell growth, cell viability, and recombinant protein production.

[00038] In some embodiments, the OS medium contains ornithine at a concentration (expressed in micromoles per liter) of at least about 90, 100, 150, 200, 250, 300, 350, 400, 450, 500, 540, 545, 550, 555, 560, 565, 568, 567, 568, 569, 570, 571, 572, 573, 574, 575, 576, 577, 578, 579, 580, 581, 582, 583, 584, 585, 586, 587, 588, 589, 590, 591, 592, 593, 594, 595, 596, 597, 598, 599, 600, 601, 602, 603, 604, 605, 606, 607, 608, 609, 610, 611, 612, 613, 614, 615, 616, 617, 618, 620, 625, 630, 635, 640, 645, 650, 700, 750, 800, 850, or 900 μM.

[00039] In some embodiments, the media contains ornithine at a concentration of about 85, 90, 95, 100, 105, 110, 113, or 115 μM. In one embodiment, the medium contains 100 μM ± 15 μM ornithine. In one embodiment, the medium contains 15 mg/L ± 2.25 mg/L ornithine • HCl.

[00040] In some embodiments, the media contains ornithine at a concentration of about 255, 260, 265, 270, 275, 280, 285, 290, 295, 300, 305, 310, 315, 320, 325, 330, 335, 340, or 345 μM. In one embodiment, the medium contains 300 μM ± 45 μM ornithine. In one embodiment, the medium contains 50 mg/L ± 7.5 mg/L ornithine • HCl.

[00041] In some embodiments, the media contains ornithine at a concentration of about 510, 515, 520, 525, 530, 535, 540, 545, 550, 555, 560, 565, 570, 575, 580, 585, 590, 595, 600, 605, 610, 615, 620, 625, 630, 635, 640, 645, 650, 655, 660, 665, 670, 675, 680, 685, or 690 μM. In one embodiment, the medium contains 600 μM ± 90 μM ornithine. In one embodiment, the medium contains 100 mg/L ± 15 mg/L ornithine • HCl.

[00042] In some embodiments, the media contains ornithine at a concentration of 765, 770, 775, 780, 785, 790, 795, 800, 805, 810, 815, 820, 825, 830, 835, 840, 845, 850, 855, 860,

865, 870, 875, 880, 885, 890, 895, 900, 905, 910, 915, 920, 925, 930, 935, 940, 945, 950, 955, 960, 965, 970, 975, 980, 985, 990, 995, 1,000, 1,005, 1,010, 1,015, 1,020, 1,025, 1,030, or 1,035 µM. In one embodiment, the medium contains 900 µM ± 135 µM ornithine. In one embodiment, the medium contains 150 mg/L ± 22.5 mg/L ornithine • HCl.

[00043] Putrescine may optionally be added to the ornithine supplemented media. Putrescine has been included, at very low concentrations, as a component in some cell culture media formulations; see for example WO 2005/028626, which describes 0.02-0.08 mg/L putrescine; US Patent No. 5,426,699 (0.08 mg/L); US Patent No. RE30,985 (0.16 mg/L); US Patent No. 5,811,299 (0.27 mg/L); US Patent No. 5,122,469 (0.5635 mg/L); US Patent No. 5,063,157 (1 mg/1); WO 2008/154014 (~100 µM - ~ 1000 µM); US Pat. App. No. 2007/0212770 (0.5 – 30 mg/L polyamine; 2 mg/L putrescine; 2 mg/L putrescine + 2 mg/L ornithine; 2 mg/L putrescine + 10 mg/L ornithine).

[00044] In some embodiments, the media contains a combination of ornithine and putrescine, wherein the putrescine can be at a concentration of at least about 150, 155, 160, 165, 170, 175, 180, 185, 190, 195, 200, 205, 210, 215, 220, 225, 230, 235, 240, 245, 250, 255, 260, 265, 270, 275, 280, 285, 290, 295, 300, 305, 310, 315, 320, 325, 330, 335, 340, 345, 350, 355, 260, 365, 370, 375, 380, 385, 390, 395, 400, 405, or 410 µM.

[00045] In some embodiments, the media contains putrescine at a concentration of about 170, 175, 180, 185, 190, 195, 200, 205, 210, 215, 220, 225, or 230 µM. In one embodiment, the medium contains 200 µM ± 30 µM putrescine in addition to ≥ 90 µM ± 14 µM ornithine. In one embodiment, the medium contains 30 mg/L ± 4.5 mg/L putrescine • 2HCl in addition to ≥ 15 mg/L ± 2.25 mg/L ornithine • HCl.

[00046] In some embodiments, the media contains putrescine at a concentration of about 295, 300, 305, 310, 315, 320, 325, 330, 335, 340, 345, 350, 355, 360, 365, 370, 375, 380, 385, 390, 395, 400, or 405 µM. In one embodiment, the medium contains 350 µM ± 52.5 µM putrescine in addition to ≥ 90 µM ± 14 µM ornithine. In one embodiment, the medium contains 57 mg/L ± 8.55 mg/L putrescine • 2HCl in addition to ≥ 15 mg/L ± 2.25 mg/L ornithine • HCl. .

[00047] In some embodiments, the media contains putrescine at a concentration of about 595, 600, 605, 610, 615, 620, 625, 630, 635, 640, 645, 650, 655, 660, 665, 670, 675, 680, 685,

690, 695, 700, 705, 710, 715, 720, 725, 730, 735, 740, 745, 750, 755, 760, 765, 770, 775, 780, 785, 790, 795, 800, or 805 μM. In one embodiment, the medium contains 714 μM ± 105 μM putrescine in addition to ≥ 90 μM ± 14 μM ornithine. In one embodiment, the medium contains 115 mg/L ± 17.25 mg/L putrescine • 2HCl in addition to ≥ 15 mg/L ± 2.25 mg/L ornithine • HCl.

[00048] In some embodiments, the media contains a pair-wise combination of any concentration of putrescine and ornithine listed above. In some embodiments, the media contains any pair-wise combination of about 595, 600, 605, 610, 615, 620, 625, 630, 635, 640, 645, 650, 655, 660, 665, 670, 675, 680, 685, 690, 695, 700, 705, 710, 715, 720, 725, 730, 735, 740, 745, 750, 755, 760, 765, 770, 775, 780, 785, 790, 795, 800, or 805 μM putrescine, and about 510, 515, 520, 525, 530, 535, 540, 545, 550, 555, 560, 565, 570, 575, 580, 585, 590, 595, 600, 605, 610, 615, 620, 625, 630, 635, 640, 645, 650, 655, 660, 665, 670, 675, 680, 685, or 690 μM ornithine. For example, the media in one embodiment contains about 700 μM putrescine plus any one of 510, 511, 512 μM, *et sequens* ornithine; or 701 μM putrescine plus any one of 510, 511, 512 μM, *et sequens* ornithine; *et cetera*. Also for example, the media in one embodiment contains about 600 μM ornithine plus any one of 700, 701, 702 μM, *et sequens* putrescine; or 601 μM ornithine plus any one of 700, 701, 702 μM, *et sequens* putrescine; *et cetera*. In some embodiments, the media contains 702 μM ± 106 μM purescine + 593 μM ± 89 μM ornithine. In one particular embodiment, the media contains about 714 μM putrescine and 593 μM ornithine. In one embodiment, the media contains 115 mg/L ± 17 mg/L putrescine • 2HCl and 100 mg/L ± 15 mg/L ornithine • HCl. In one particular embodiment, the media contains 115 mg/L putrescine • 2HCl and 100 mg/L ornithine • HCl.

[00049] In one embodiment and in addition to the inclusion of ornithine or putrescine, the media contains a mixture of nucleosides in a cumulative concentration of at least 50 μM, at least 60 μM, at least 70 μM, at least 80 μM, at least 90 μM, at least 100 μM, at least 110 μM, at least 115 μM, at least 120 μM, at least 125 μM, at least 130 μM, at least 135 μM, at least 140 μM, at least 145 μM, at least 150 μM, at least 155 μM, at least 160 μM, at least 165 μM, or at least 170 μM. In one embodiment, the media contains about 174 μM ± 26 μM nucleoside. In one embodiment, the media contains purine derivatives in a cumulative concentration of at least

40 μM, at least 45 μM, at least 50 μM, at least 55 μM, at least 60 μM, at least 65 μM, at least 70 μM, at least 75 μM, at least 80 μM, at least 85 μM, at least 90 μM, at least 95 μM, at least 100 μM, or at least 105 μM. In one embodiment, the media contains about 106 μM ± 5 μM of purine derivatives. Purine derivatives include hypoxanthine and the nucleosides adenosine and guanosine. In one embodiment, the media contains pyrimidine derivatives in a cumulative concentration of at least 30 μM, at least 35 μM, at least 40 μM, at least 45 μM, at least 50 μM, at least 55 μM, at least 60 μM, or at least 65 μM. In one embodiment, the media contains about 68 μM ± 5 μM of pyrimidine derivatives. Pyrimidine derivatives include the nucleosides thymidine, uridine, and cytidine. In one particular embodiment, the media contains adenosine, guanosine, cytidine, uridine, thymidine and hypoxanthine.

[00050] In addition to the inclusion of ornithine or putrescine, in one embodiment, the media also contains amino acids in a cumulative concentration of at least 40 mM, wherein the amount of glutamine is not included in the calculation of the cumulative total. In one embodiment, glutamine is not included in the media, but may be supplied as a "point-of-use addition" to the media during the culturing of cells such as during the production of protein. Thus, in some embodiments, such as in the method to culture cells or the method to produce a protein of interest, the media may be supplemented with glutamine as a point-of-use addition. In one such embodiment, glutamine is added in an amount less than about 40 mM, less than about 35 mM, less than about 30 mM, less than about 25 mM, less than about 20 mM, less than about 15 mM, less than about 10 mM, less than about 8 mM, less than about 7 mM, less than about 6 mM, less than about 5 mM, less than about 4 mM, less than about 3 mM, or less than about 2.5 mM. In one embodiment the amount of glutamine in the media that was supplemented with glutamine is about 2 mM ± 0.5 mM.

[00051] In one embodiment, in addition to the inclusion of ornithine or a combination of both ornithine and putrescine, the media also contains amino acids having a non-polar side group in a concentration of at least 15 mM, at least 24 mM, at least 25 mM, at least 26 mM, at least 27 mM, at least 28 mM, at least 29 mM, or at least 30 mM. In one embodiment, the media contains about 30 mM of amino acids having a non-polar side group. In one

embodiment, of the total amount of amino acids by mole contained within the media, at least 32%, at least 33%, at least 34%, at least 35%, at least 36%, at least 37%, at least 38%, at least 39%, at least 40%, or at least 41% are amino acids having non-polar side groups. In one embodiment, about 42% ± 1% by mole of the amino acids in the media are amino acids having a non-polar side group. Amino acids having a non-polar side group include alanine, valine, leucine, isoleucine, proline, phenylalanine, tryptophan, and methionine.

[00052] In one embodiment, in addition to the inclusion of ornithine or a combination of both ornithine and putrescine, the media also contains amino acids having an uncharged polar side group in a concentration of about 10 mM to 34 mM, about 11 mM to 33 mM, about 12 mM to 32 mM, about 13 mM to 31 mM, about 14 mM to 30 mM, about 15 mM to 29 mM, about 16 mM to 28 mM, about 17 mM to 27 mM, about 18 mM to 26 mM, about 19 mM to 25 mM, about 20 mM to 24 mM, about 21 mM to 23 mM, or about 22 mM. In one embodiment, the medium contains about 22 mM of amino acids having an uncharged polar side group. In another embodiment, the medium contains about 12 mM amino acids having an uncharged polar side group. In one embodiment, of the total amount by mole of amino acids contained within the media, about 14% to 46%, about 15% to 45%, about 16% to 44%, about 17% to 43%, about 18% to 42%, about 19% to 41%, about 20% to 40%, about 21% to 39%, about 22% to 38%, about 23% to 37%, about 24% to 36%, about 25% to 35%, about 26% to 34%, about 27% to 33%, about 28% to 32%, about 29% to 31%, or about 30% are amino acids having uncharged polar side groups. In one embodiment, about 30%± 3% by mole of the amino acids in the media are amino acids having an uncharged polar side group. Amino acids having an uncharged polar side group include glycine, serine, threonine, cysteine, tyrosine, asparagine, and glutamine.

[00053] In one embodiment, in addition to the inclusion of ornithine or a combination of both ornithine and putrescine, the media also contains amino acids having a negative charge at pH 6 (*i.e.,* acidic amino acids) in a concentration of about 4 mM to 14 mM, about 5 mM to 13 mM, about 6 mM to 12 mM, about 7 mM to 11 mM, about 8 mM to 10 mM, about 9 mM, or about 4 mM. In one embodiment, the media contains about 9 mM of acidic amino acids. In one

embodiment, the media contains 9 mM ± 1 mM of acidic amino acids. In one embodiment, of the total amount by mole of amino acids contained within the media, about 8% to 18%, about 9% to 17%, about 10% to 16%, about 11% to 15%, about 12% to 14%, or about 13% are acidic amino acids. In one embodiment, about 12.6% ± 1% by mole of the amino acids in the media are acidic amino acids. Acidic amino acids include aspartic acid and glutamic acid.

[00054] In one embodiment, in addition to the inclusion of ornithine or a combination of both ornithine and putrescine, the media also contains amino acids having a positive charge at pH 6 (*i.e.,* basic amino acids) in a concentration of at least 3.5 mM, at least 4 mM, at least 5 mM, at least 6 mM, at least 7 mM, at least 8 mM, at least 9 mM, at least 10 mM, or at least 11 mM. In one embodiment, the media contains about 11 mM of basic amino acids. In one embodiment, the media contains about 11.42 mM ± 1 mM of basic amino acids. In one embodiment, of the total amount by mole of amino acids contained within the media, at least 5%, at least 6%, at least 7%, at least 8%, at least 9%, at least 10%, at least 11%, at least 12%, at least 13%, at least 14%, or at least 15% are basic amino acids. In one embodiment, about 16% by mole of the amino acids in the media are basic amino acids. In one embodiment, about 15.8% ± 2.4% by mole of the amino acids in the media are basic amino acids. In one embodiment, about 21% ± 3.2% by mole of the amino acids in the media are basic amino acids. Basic amino acids include lysine, arginine, and histidine.

[00055] In one embodiment, in addition to the inclusion of ornithine or a combination of both ornithine and putrescine, the media also contains about 30 mM non-polar amino acids, about 22 mM uncharged polar amino acids, about 9 mM acidic amino acids, and about 11 mM basic amino acids. In one embodiment, of the amino acids in the media about 42% by mole are non-polar amino acids, about 30% by mole are uncharged polar amino acids, about 13% by mole are acidic amino acids, and about 16% by mole are basic amino acids.

[00056] In addition to the inclusion of ornithine or a combination of both ornithine and putrescine, in one embodiment, the media contains micromolar amounts of fatty acids (or fatty acid derviatives) and tocopherol. In one embodiment, the fatty acids include any one or more of linoleic acid, linolenic acid, thioctic acid, oleic acid, palmitic acid, stearic acid, arachidic acid,

arachidonic acid, lauric acid, behenic acid, decanoic acid, dodecanoic acid, hexanoic acid, lignoceric acid, myristic acid, and octanoic acid. In one embodiment, the media contains tocopherol, linoleic acid, and thioctic acid.

[00057] In one embodiment, the media also contains a mixture of vitamins, which includes other nutrients and essential nutrients, at a cumulative concentration of at least about 700 µM or at least about 2 mM. In one embodiment, the mixture of vitamins contains one or more of D-biotin, choline chloride, folic acid, myo-inositol, niacinamide, pyridoxine HCl, D-pantothenic acid (hemiCa), riboflavin, thiamine HCl, vitamin B12, and the like. In one embodiment, the mixture of vitamins includes all of D-biotin, choline chloride, folic acid, myo-inositol, niacinamide, pyridoxine HCl, D-pantothenic acid (hemiCa), riboflavin, thiamine HCl, and vitamin B12.

[00058] Various embodiments of the media of the invention include any of the combinations of the above described embodiments, including chemically defined, hydrolysate-free serum-free media comprising ornithine or putrescine in the indicated amounts, plus *inter alia* (a) amino acids; (b) optionally nucleosides; (c) salts of divalent cations; (d) fatty acids and tocopherol; and (e) vitamins. In some embodiments, all small amounts of hydrolysates may be added to the OS media.

[00059] The applicants envision that in the practice of this invention any one or more of a variety of base media or combinations thereof, to which the ornithine or a combination of both ornithine and putrescine are added, may be used. Base media are generally known in the art and include *inter alia* Eagle's MEME (minimal essential media) (Eagle, Science, 1955, 112(3168):501-504), Ham's F12 (Ham, Proc. Nat'l. Acad. Sci. USA, 1965, 53:288-293), F-12 K medium, Dulbecco's medium, Dulbecco's Modified Eagle Medium (Proc. Natl. Acad. Sci. USA., 1952 August; 38(8): 747–752), DMEM/ Ham's F12 1:1, Trowell's T8, A2 media Holmes and Wolf, Biophys. Biochem. Cytol., 1961, 10:389-401), Waymouth media (Davidson and Waymouth, Biochem. J., 1945, 39(2):188–199), Williams E media (William's et al., Exp. Cell Res., 1971, 69:105 *et seq.*), RPMI 1640 (Moore et al., J. Amer. Med. Assoc., 1967, 199:519-524), MCDB 104/110 media (Bettger et al., Proc. Nat'l. Acad. Sci. USA, 1981, 78(9):5588-5592), Ventrex HL-1 media, albumin-globulin media (Orr et al., Appl. Microbiol., 1973, 25(1):49-

54), RPMI- 1640 Medium, RPMI- 1641 Medium, Iscove's Modified Dulbecco's Medium, McCoy's 5 A Medium, Leibovitz's L- 15 Medium, and serum-free media such as EX-CELL™ 300 Series (JRH Biosciences, Lenexa, Kansas), protamine-zinc-insulin media (Weiss et al., 1974, US 4,072,565), biotin-folate media (Cartaya, 1978, US Re30,985), Transferrin-fatty acid media (Baker, 1982, US 4,560,655), transferrin-EGF media (Hasegawa, 1982, US 4,615,977; Chessebeuf, 1984, US 4,786,599), and other media permutations (see Inlow, US 6,048,728; Drapeau, US 7,294,484; Mather, US 5,122,469; Furukawa, US 5,976,833; Chen, US 6,180,401; Chen, US 5,856,179; Etcheverry, US 5,705,364; Etcheverry, US 7,666,416; Ryll, US 6,528,286; Singh, US 6,924,124; Luan, US 7,429,491; and the like).

[00060] In a particular embodiment, the media is chemically defined and contains in addition to the ornithine or combination of both ornithine and putrescine: $CaCl_2$ $2H_2O$; HEPES buffer, KCl; $MgSO_4$; NaCl; Na_2HPO_4 or other phosphate salts; pyruvate; L-alanine; L-arginine HCl; L-asparagine H_2O; L-aspartic acid; L-cysteine HCl H_2O; L-glutamic acid; Glycine; L-histidine HCl H_2O; L-isoleucine; L-leucine; L-lysine HCl; L-methionine; L-ornithine HCl; L-phenylalanine; L-proline; L-serine; L-threonine; L-tryptophan; L-tyrosine 2Na 2 H_2O; L-valine; D-biotin; choline chloride; folic acid; myo-inositol; niacinamide; pyridoxine HCl; D-pantothenic acid; riboflavin; thiamine HCl; vitamin B12; ρ-aminobenzoic acid; ethanolamine HCl; Pluronic F68; DL-a-tocopherol phosphate; linoleic acid; Na_2SeO_3; thioctic acid; and glucose; and optionally adenosine; guanosine; cytidine; uridine; thymidine; and hypoxanthine 2Na.

[00061] In one embodiment, the starting osmolarity of the media of the invention is 200-500, 250-400, 275-350, or about 300 mOsm. During growth of the cells in the media of the invention, and in particular following any feedings according to a fed batch protocol, the osmolarity of the culture may increase up to about 350, 400, 450, or as high as 500 mOsm.

[00062] In some embodiments wherein the osmolarity of the defined medium is less than about 300, the osmolarity is brought to about 300 with the addition of one or more salts in excess of the amount specified. In one embodiment, osmolarity is increased to a desired level by adding one or more of an osmolyte selected from sodium chloride, potassium chloride, a magnesium salt, a calcium salt, an amino acid salt, a salt of a fatty acid, sodium bicarbonate, sodium carbonate, potassium carbonate, a chelator that is a salt, a sugar (*e.g.,* galactose,

glucose, sucrose, fructose, fucose, *etc.*), and a combination thereof. In one embodiment, the osmolyte is added over and above its concentration in a component already present in the defined medium (e.g., a sugar is added over and above the concentration specified for a sugar component).

[00063] Each and every embodiment of the media described above, as well as any other serum-free media containing at least about 90 μM ornithine (or containing a combination of at least about 100 μM ornithine plus at least about 200 μM putrescine) are herinafter refered to as ornithine supplemented ("OS") media. Conversely, media containing no ornithine (or no ornithine/putrescine combination), or media containing less than 100 μM ornithine (or media containing less than 100 μM ornithine and less than 200 μM putrescine), are hereinafter refered to as non-ornithine supplemented ("non-OS") media.

CELL CULTURE

[00064] The present invention provides a cell culture comprising a cell line expressing a protein of interest in an OS medium as described above. In one embodiment, the cell culture contains insulin, which can be added as a point-of-use ingredient to the media, or can be included in the media formulation. In one embodiment, the cell line comprises cells capable of producing a biotherapeutic protein. Examples of cell lines that are routinely used to produce protein biotherapeutics include *inter alia* primary cells, BSC cells, HeLa cells, HepG2 cells, LLC-MK cells, CV-1 cells, COS cells, VERO cells, MDBK cells, MDCK cells, CRFK cells, RAF cells, RK cells, TCMK-1 cells, LLCPK cells, PK15 cells, LLC-RK cells, MDOK cells, BHK cells, BHK-21 cells, CHO cells, CHO-K1 cells, NS-1 cells, MRC-5 cells, WI-38 cells, BHK cells, 3T3 cells, 293 cells, RK cells, Per.C6 cells and chicken embryo cells. In one embodiment, the cell line is a CHO cell line or one or more of several specific CHO cell variants optimized for large-scale protein production, *e.g.,* CHO-K1.

[00065] "Cell culture" or "culture" means the growth and propagation of cells outside of a multicellular organism or tissue. Suitable culture conditions for mammalian cells are known in the art. See *e.g.* Animal cell culture: A Practical Approach, D. Rickwood, ed., Oxford University Press, New York (1992). Mammalian cells may be cultured in suspension or while attached to a solid substrate. Fluidized bed bioreactors, hollow fiber bioreactors, roller bottles, shake flasks, or stirred tank bioreactors, with or without microcarriers, and operated in a batch, fed batch, continuous, semi-continuous, or perfusion mode are available for mammalian cell culture. Cell culture media or concentrated feed media may be added to the culture continuously or at intervals during the culture. For example, a culture may be fed once per day, every other day, every three days, or may be fed when the concentration of a specific medium component, which is being monitored, falls outside a desired range.

[00066] Animal cells, such as CHO cells, may be cultured in small scale cultures, such as in 125 ml containers having about 25 ml of media, 250 ml containers having about 50 to 100 ml of media, 500 ml containers having about 100 to 200 ml of media. Alternatively, the cultures can be large scale such as for example 1000 ml containers having about 300 to 1000 ml of media, 3000 ml containers having about 500 ml to 3000 ml of media, 8000 ml containers having

about 2000 ml to 8000 ml of media, and 15000 ml containers having about 4000 ml to 15000 ml of media. Cultures for manufacturing can contain 10,000 L of media or more. Large scale cell cultures, such as for clinical manufacturing of protein therapeutics, are typically maintained for days, or even weeks, while the cells produce the desired protein(s). During this time the culture can be supplemented with a concentrated feed medium containing components, such as nutrients and amino acids, which are consumed during the course of the culture. Concentrated feed medium may be based on any cell culture media formulation. Such a concentrated feed medium can contain most of the components of the cell culture medium at, for example, about 5X, 6X, 7X, 8X, 9X, 10X, 12X, 14X, 16X, 20X, 30X, 50X, 100x, 200X, 400X, 600X, 800X, or even about 1000X of their normal useful amount. Concentrated feed media are often used in fed batch culture processes.

[00067] In some embodiments, the cell culture media is supplemented with “point-of-use additions”, also known as additions, point-of-use ingredients, or point-of-use chemicals, during the course of cell growth or protein production. Point-of-use additions include any one or more of a growth factor or other proteins, a buffer, an energy source, a salt, an amino acid, a metal, and a chelator. Other proteins include transferrin and albumin. Growth factors, which include cytokines and chemokines, are generally known in the art and are known to stimulate cell growth, or in some cases, cellular differentialtion. A growth factor is usually a protein (*e.g.,* insulin), a small peptide, or a steroid hormone, such as estrogen, DHEA, testosterone, and the like. In some cases, a growth factor may be a non-natural chemical that promotes cell proliferation or protein production, such as *e.g*., tetrahydrofolate (THF), methotrexate, and the like. Non-limiting examples of protein and peptide growth factors include angiopoietins, bone morphogenetic proteins (BMPs), brain-derived neurotrophic factor (BDNF), epidermal growth factor (EGF), erythropoietin (EPO), fibroblast growth factor (FGF), glial cell line-derived neurotrophic factor (GDNF), granulocyte colony-stimulating factor (G-CSF), granulocyte macrophage colony-stimulating factor (GM-CSF), growth differentiation factor-9 (GDF9), hepatocyte growth factor (HGF), hepatoma-derived growth factor (HDGF), insulin, insulin-like growth factor (IGF), migration-stimulating factor, myostatin (GDF-8), nerve growth factor (NGF) and other neurotrophins, platelet-derived growth factor (PDGF), thrombopoietin (TPO),

transforming growth factor alpha(TGF-α), transforming growth factor beta(TGF-β), tumor necrosis factor-alpha(TNF-α), vascular endothelial growth factor (VEGF), wnt signaling pathway agonists, placental growth factor (PlGF), fetal Bovine somatotrophin (FBS), interleukin-1 (IL-1), IL-2, IL-3, IL-4, IL-5, IL-6, IL-7, and the like. In one embodiment, the cell culture media is supplemented with the point-of-use addition growth factor insulin. In one embodiment, the concentration of insulin in the media, *i.e.,* the amount of insulin in the cell culture media after addition, is from about 0.1 μM to 10 μM. One or more the point-of-use additions can also be included in the media formulation of some embodiments.

[00068] Buffers are generally known in the art. The invention is not restricted to any particular buffer or buffers, and any one of ordinary skill in the art can select an appropriate buffer or buffer system for use with a particular cell line producing a particular protein. In one embodiment, a point-of-use addition buffer is $NaHCO_3$. In one embodiment, the point-of-use addition buffer comprises $NaHCO_3$. In another embodiment, the buffer is HEPES.

[00069] Energy sources for use as a point-of-use addition in cell culture are also well known in the art. Without limitation, in one embodiment, the point-of-use addition energy source is glucose. Given the particular and specific requirements of a particular cell line and the protein to be produced, in one embodiment the glucose can be added to a concentration of about 1 to 20 mM in the media. In some cases, glucose can be added at high levels up to 10 g/L.

[00070] Chelators are likewise well known in the art of cell culture and protein production. Tetrasodium EDTA dehydrate and citrate are two common chelators used in the art, although other chelators may be employed in the practice of this invention. In one embodiment, a point-of-use addition chelator is tetrasodium EDTA dihydrate. In one embodiment, a point-of-use addition chelator is citrate, such as $Na_3C_6H_5O_7$.

[00071] In one embodiment, the cell culture may be supplemented with one or more point-of-use addition amino acids, such as *e.g.,* glutamine. In one embodiment, the cell culture media is supplemented with the point-of-use addition glutamine at a final concentration of about 1 mM to 13 mM.

[00072] Other point-of-use additions include one or more of various metal salts, such as salts of iron, nickel, zinc and copper. In one embodiment, the cell culture media is supplemented with any one or more of copper sulfate, zinc sulfate, ferric chlroide; and nickel sulfate.

[00073] In one embodiment, the cell culture media is supplemented with any one or more or all of the following point-of-use additions: about 29.8 mM NaHCO3, about 2 mM glutamine, about 0.86 μM insulin, about 11.1 mM glucose, about 6.54 μM zinc sulfate, about 0.168 μM copper sulfate, about 75 μM ferric chloride, about 0.639 μM nickel sulfate, about 85 μM EDTA, and about 50 μM citrate.

[00074] In one embodiment, the media is supplemented at intervals during cell culture according to a fed-batch process. Fed-batch culturing is generally known in the art and employed to optimized protein production (see Y.M. Huang *et al.,* Biotechnol Prog. 2010 Sep-Oct;26(5):1400-10).

[00075] Cell viability, viable cell density, and cell doubling are improved relative to cells grown in culture without ornithine or putrescine. Regarding cell viability, cells grown in OS media show a viability that is at least 10%, at least 15%, at least 20%, at least 25%, at least 30%, at least 35%, at least 40%, at least 45%, at least 50%, at least, 55%, at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 99%, at least 100%, or at least 3-fold greater than the viability of similar or identical cells grown in non-OS media.

[00076] In some embodiments, the doubling rate of viable mammalian cells in OS media is at least 5%, at least 6%, at least 7%, at least 8%, at least 9%, at least 10%, at least 11%, at least 12%, at least 13%, at least 14%, at least 15%, at least 16%, at least 17%, at least 18%, at least 19%, at least 20%, at least 21%, at least 22%, at least 23%, at least 24%, at least 25%, at least 26%, at least 27%, at least 28%, at least 29%, at least 30%, at least 35%, at least 40%, at least 45%, at least 50%, at least 55%, at least 60%, at least 70%, at least 80%, at least 90%, at least 100%, or at least 3-fold greater than the doubling rate of mammalian cells cultured in non-OS media. In some embodiments, the doubling rate of viable mammalian cells in OS media is about 10%, 11%, 12%, 13%, 14%, 15%, 16%, 17%, 18%, 19%, 20%, 21%, 22%, 23%, 24%,

25%, 26%, 27%, 28%, 29%, or 30% greater than the doubling rate of mammalian cells in non-OS media.

[00077] In some embodiments, the doubling time of actively cycling mammalian cells is less than 30 hours, less than 29 hours, less than 28 hours, less than 27 hours, less than 26 hours, less than 25 hours, less than 24 hours, less than 23 hours, less than 22 hours, less than 21 hours, less than 20 hours, less than 19 hours, or less than 18 hours in OS media. In some embodiments, the doubling time of actively growing mammalian cells is less than 28 hours in OS media. In some embodiments, the doubling time of mammalian cells is about 27 ± 1 hours, about 26 ± 1 hours, about 25 ± 1 hours, about 24 ± 1 hours, about 23 ± 1 hours, about 22 ± 1 hours, or about 21 ± 1 hours in OS media. In some embodiments, the doubling time of actively cycling mammalian cells is about 24 ± 1 hours in OS media. In some embodiments, the doubling time of actively dividing cells cultured in OS media is at least 15%, at least 16%, at least 17%, at least 18%, at least 19%, at least 20%, or at least 25% shorter than the doubling time of actively cycling cells cultured in a non-OS media.

PROTEIN PRODUCTION

[00078] In addition to chemically defined OS media and methods of culturing cells in OS media, the present invention provides methods of producing a protein, such as a therapeutically effective antibody or other biopharmaceutical drug substance, in a cell cultured in OS media.

[00079] In some embodiments, the rate of production of protein by mammalian cells cultured in OS media is at least 5%, 10%, 15%, or 20% greater than the rate of production of protein by an identical mammalian cell cultured in non-OS media. In some embodiments the rate of production of protein in cells cultured in OS media is at least 1 ρg/cell/day ("PCD"), at least 2 PCD, at least 3 PCD, at least 4 PCD, at least 5 PCD, at least 6 PCD, at least 7 PCD, at least 8 PCD, at least 9 PCD, at least 10 PCD, at least 15 PCD, at least 20 PCD, at least 25 PCD, at least 30 PCD, at least 35 PCD, at least 40 PCD, at least 45 PCD, at least 50 PCD, at least 75 PCD, or at least 100 PCD.

[00080] In some embodiments the protein production yield or titer, which can be expressed in grams of protein product per liter of culture media, from cells cultured in OS media

is at least 100 mg/L, at least 1 g/L, at least 1.2 g/L, at least 1.4 g/L, at least 1.6 g/L, at least 1.8 g/L, at least 2 g/L, at least 2.5 g/L, at least 3 g/L, at least, 3.5 g/L, at least 4 g/L, at least 4.5 g/L, at least 5 g/L, at least 5.5 g/L, at least 6 g/L, at least 6.5 g/L, at least 7 g/L, at least 7.5 g/L, at least 8 g/L, at least 8.5 g/L, at least 9 g/L, at least 9.5 g/L, at least 10 g/L, or at least 20 g/L.

[00081] In some embodiments, the protein product (protein of interest) is an antibody, a human antibody, a humanized antibody, a chimeric antibody, a monoclonal antibody, a multispecific antibody, a bispecific antibody, an antigen binding antibody fragment, a single chain antibody, a diabody, triabody or tetrabody, a Fab fragment or a F(ab')2 fragment, an IgD antibody, an IgE antibody, an IgM antibody, an IgG antibody, an IgG1 antibody, an IgG2 antibody, an IgG3 antibody, or an IgG4 antibody. In one embodiment, the antibody is an IgG1 antibody. In one embodiment, the antibody is an IgG2 antibody. In one embodiment, the antibody is an IgG4 antibody.

[00082] In some embodiments, the protein of interest is a recombinant protein that contains an Fc moiety and another domain, (*e.g.,* an Fc-fusion protein). In some embodiments, an Fc-fusion protein is a receptor Fc-fusion protein, which contains one or more of one or more extracellular domain(s) of a receptor coupled to an Fc moiety. In some embodiments, the Fc moiety comprises a hinge region followed by a CH2 and CH3 domain of an IgG. In some embodiments, the receptor Fc-fusion protein contains two or more distinct receptor chains that bind to either a single ligand or multiple ligands. For example, an Fc-fusion protein is a trap, such as for example an IL-1 trap (*e.g.,* rilonacept, which contains the IL-1RAcP ligand binding region fused to the Il-1R1 extracellular region fused to Fc of hIgG1; see U.S. Pat. No. 6,927,004, which is herein incorporated by reference in its entirety), or a VEGF trap (*e.g.,* aflibercept, which contains the Ig domain 2 of the VEGF receptor Flt1 fused to the Ig domain 3 of the VEGF receptor Flk1 fused to Fc of hIgG1; see U.S. Pat. Nos. 7,087,411 and 7,279,159).

[00083] The present invention is not limited to any particular type of cell for protein production. Examples of cell types suitable for protein production include mammalian cells, insect cells, avian cells, bacterial cells, and yeast cells. The cells may be stem cells or recombinant cells transformed with a vector for recombinant gene expression, or cells transfected with a virus for producing viral products. The cells may contain a recombinant

heterologous polynucleotide construct that encodes a protein of interest. That construct can be an episome or it can be an element that is physically integrated into the genome of the cell. The cells may also produce a protein of interest without having that protein encoded on a heterologous polypeptide construct. In other words, the cell may naturally encode the protein of interest, such as a B-cell producing an antibody. The cells may also be primary cells, such as chicken embryo cells, or primary cell lines. Examples of useful cells include BSC cells, LLC-MK cells, CV-1 cells, COS cells, VERO cells, MDBK cells, MDCK cells, CRFK cells, RAF cells, RK cells, TCMK-1 cells, LLCPK cells, PK15 cells, LLC-RK cells, MDOK cells, BHK-21 cells, chicken embryo cells, NS-1 cells, MRC-5 cells, WI-38 cells, BHK cells, 293 cells, RK cells, Per.C6 cells and CHO cells. In various embodiments, the cell line is a CHO cell derviative, such as CHO-K1, CHO DUX B-11, CHO DG-44, Veggie-CHO, GS-CHO, S-CHO, or CHO lec mutant lines.

[00084] In one embodiment, the cell, which is a CHO cell, ectopically expresses a protein. In one embodiment, the protein comprises an immunoglobulin heavy chain region, such as a CH1, CH2, or CH3 region. In one embodiment, the protein comprises a human or rodent immunoglobulin CH2 and CH3 region. In one embodiment, the protein comprises a human or rodent immunoglobulin CH1, CH2, and CH3 region. In one embodiment, the protein comprises a hinge region and a CH1, CH2, and CH3 region. In a specific embodiment, the protein comprises an immunoglobulin heavy chain variable domain. In a specific embodiment, the protein comprises an immunoglobulin light chain variable domain. In a specific embodiment, the protein comprises an immunoglobulin heavy chain variable domain and an immunoglobulin light chain variable domain. In a specific embodiment, the protein is an antibody, such as a human antibody, a rodent antibody, or a chimeric human/rodent antibody (*e.g.,* human/mouse, human/rat, or human hamster).

[00085] A production phase can be conducted at any scale of culture, from individual flasks and shaker flasks or wave bags, to one-liter bioreactors, and to large scale industrial bioreactors. A large scale process can be conducted in a volume of about 100 liters to 20,000 liters or more. One or more of several means may be used to control protein production, such as temperature shift or chemical induction. A growth phase may occur at a higher temperature than a production phase. For example, a growth phase may occur at a first temperature of

about 35°C to 38°C, and a production phase may occur at a second temperature of about 29°C to 37°C, optionally from about 30°C to 36°C or from about 30°C to 34°C. In addition, chemical inducers of protein production, such as caffeine, butyrate, tamoxifen, estrogen, tetracycline, doxycycline, and hexamethylene bisacetamide (HMBA), may be added concurrent with, before, or after a temperature shift. If inducers are added after a temperature shift, they can be added from one hour to five days after the temperature shift, such as from one to two days after the temperature shift. Production cell cultures may be run as continuous feed culture system, as in a chemostat (see C. Altamirano *et al.,* Biotechnol Prog. 2001 Nov-Dec; 17(6):1032-41), or according to a fed-batch process (Huang, 2010).

[00086] The invention is useful for improving protein production via cell culture processes. The cell lines used in the invention can be genetically engineered to express a polypeptide of commercial or scientific interest. Genetically engineering the cell line involves transfecting, transforming or transducing the cells with a recombinant polynucleotide molecule, or otherwise altering (*e.g.,* by homologous recombination and gene activation or fusion of a recombinant cell with a non-recombinant cell) so as to cause the host cell to express a desired recombinant polypeptide. Methods and vectors for genetically engineering cells or cell lines to express a polypeptide of interest are well known to those of skill in the art; for example, various techniques are illustrated in Current Protocols in Molecular Biology. Ausubel et al., eds. (Wiley & Sons, New York, 1988, and quarterly updates); Sambrook et al., Molecular Cloning: A Laboratory Manual (Cold Spring Laboratory Press, 1989); Kaufman, R. J., Large Scale Mammalian Cell Culture, 1990, pp. 15-69. A wide variety of cell lines suitable for growth in culture are available from the American Type Culture Collection (Manassas, Va.) and commercial vendors. Examples of cell lines commonly used in the industry include VERO, BHK, HeLa, CVl (including Cos), MDCK, 293, 3T3, myeloma cell lines (e.g., NSO, NSl), PC12, WI38 cells, and Chinese hamster ovary (CHO) cells. CHO cells are widely used for the production of complex recombinant proteins, such as cytokines, clotting factors, and antibodies (Brasel et al. (1996), Blood 88:2004-2012; Kaufman et al. (1988), J.Biol Chem 263:6352-6362; McKinnon et al. (1991), J MoI Endocrinol 6:231-239; Wood et al. (1990), J Immunol.

145:3011-3016). The dihydrofolate reductase (DHFR)-deficient mutant cell lines (Urlaub et al. (1980), Proc Natl Acad Sci USA 77: 4216-4220), DXBl 1 and DG-44, are desirable CHO host cell lines because the efficient DHFR selectable and amplifiable gene expression system allows high level recombinant protein expression in these cells (Kaufman RJ. (1990), Meth Enzymol 185:537-566). In addition, these cells are easy to manipulate as adherent or suspension cultures and exhibit relatively good genetic stability. CHO cells and the proteins recombinantly expressed by them have been extensively characterized and have been approved for use in clinical and commercial manufacturing by regulatory agencies. In some embodiments, the CHO cell lines are cell lines as described in U.S. Patent Application Publications No. 2010/0304436 A1, 2009/0162901 A1 and 2009/0137416 A1, and U.S. Patents No. 7,455,988 B2, 7,435,553 B2, and 7,105,348 B2.

[00087] The present invention is not limited in scope by the specific embodiments described herein, which are intended as illustrations of individual aspects or embodiments of the invention. Functionally equivalent methods and components are within the scope of the invention. Various modifications of the invention, in addition to those described here, are apparent to those skilled in the art from the foregoing description and accompanying drawings. Such modifications fall within the scope of the invention.

[00088] The invention is based, in part, on the discovery that addition of ornithine or a combination of ornithine and putrescine to serum free cell culture media results in increased cell growth, viability and polypeptide production from a recombinantly engineered animal cell line (or natural cell) expressing a protein of interest, thereby enhancing culture robustness, improving the yield of the polypeptide of interest.

EXAMPLE 1: IMPROVED VIABLE CELL CULTURE DENSITY

[00089] A 250mL shake flask was inoculated from a seed culture of a recombinant antibody producing cell line derived from CHO K1. The inoculated cells were grown at 36.5ºC for seven days and fed glucose on days three and five. Cells were grown in each of two separate chemically defined (hydrolysate-free and serum-free) media. The first medium contained about 75 mM amino acids (Medium 1), the second medium contained about 40 mM

amino acids (Medium 2), and both formulations contained no more than 2.5 μM (0.4 mg/L) putrescine. Another group of medium conditions was generated by adding soy hydrolysate at a concentration of 7.5 g/L to Medium 2. To each of the three control media, about 593 μM ornithine (as 100 mg/L L-ornithine • HCl), or a combination of about 593 μM ornithine (as 100 mg/L L-ornithine • HCl) and about 714 μM putrescine (as 115 mg/L putrescine • 2HCl) were added. Aliquots of 3 mL culture were removed on days 3, 5, and 7 and viable cell counts were conducted using trypan blue exclusion on a BioProfile FLEX™ instrument (Nova Biomedical). At day zero, all cultures contained $0.8x10^6$ viable cells per mL. For a given medium (Medium 1, Medium 2, or Medium 2 + Soy), viable cell counts over a seven-day period revealed that CHO cells grown in media supplemented with ornitihine or ornithine plus putrescine had increased viable cell densities. The effect was especially pronounced in the hydrolysate free media (*i.e.,* 2-fold to 4-fold or more increase in viable cell density) during the seven-day period. Hydrolysate free OS Medium 2 performed comparably to soy containing non-OS Medium 2 indicating that the cell growth benefit of soy hydrolysate can be replicated by ornithine replacement. Increased cell density by adding ornithine or ornithine and putrescine to Medium 2 + soy was also observed. Results are presented in Table 1.

[00090] TABLE 1: AVERAGE VIABLE CELL CULTURE DENSITY (10^6 CELLS PER MILLILITER) AND X FOLD INCREASE OVER BASELINE*

Supplement:	Time	Unsupplemented	Ornitihine	Ornithine + putrescine
Medium 1	3 days	2.4 / 1X	6.1 / 2.5X	5.0 / 2.1X
	5 days	3.4 / 1X	12.6 / 3.7X	12.4 / 3.6X
	7 days	3.6 / 1X	7.0 / 1.9X	6.8 / 1.9X
Medium 2	3 days	1.7 / 1X	5.1 / 3.0X	5.2 / 3.1X
	5 days	2.0 / 1X	7.6 / 3.8X	8.0 / 4.0X
	7 days	1.6 / 1X	5.9 / 3.7X	5.8 / 3.6X
Medium 2 + soy hydrolysate	3 days	5.2 / 1X	5.4 / 1X	4.7 / 0.9X
	5 days	7.7 / 1X	9.3 / 1.2X	9.3 / 1.2X
	7 days	ND	9.6 / ND	9.1 / ND

*Base line is unsupplemented media for a given medium formulation on a given day.

[00091] We also examined the effect of various amounts of ornithine • HCl (*i.e.,* 50 mg/mL, 100 mg/mL, and 150 mg/mL) on viable cell density in Medium 3, which contains about 75 mM of amino acids and 0.4 mg/L putrescine HCl ("Medium 3"). A single seed train culture of a recombinant antibody producing cell line derived from CHO K1 was used to inoculate 50 mL

TubeSpin® Bioreactors (TPP) at 0.4 X 10^6 cells/mL at a 15 mL working volume. The cells were grown in a 37°C incubator for three days. Aliquots of 3 mL culture were removed on day 3 and viable cell counts were conducted using trypan blue exclusion on a BioProfile FLEX™ instrument (Nova Biomedical). All three levels of ornithine improved cell density on average (N=3) by slightly more than two fold. The results are depicted in Table 2.

[00092] TABLE 2: VIABLE CELL CULTURE DENSITY (10^6 CELLS PER MILLILITER) AND X FOLD INCREASE OVER BASELINE*

	Control	Ornithine HCl		
Concentration (mg/mL)	0	50	100	150
Viable cell density (10^6 cells/mL)	1.3	3.2	3.1	3.1
Fold increase over control	1X	2.5X	2.4X	2.4X

*Base line is unsupplemented Medium 3.

EXAMPLE 2: IMPROVED CELL CULTURE DOUBLING TIME

[00093] The doubling time of a recombinant antibody producing cell line derived from CHO K1 cells in logarithmic growth phase was determined under various cell culture media conditions. Seed train cultures were passaged at 36.5ºC in 250 mL shaker flasks over a period of 14 days in each of three separate media: Medium 1, Medium 2, and Medium 2 containing soy hydrolysate (Medium 2 + Soy). Aliquots of 1 mL were removed from each condition on Day 0 and at the time of seed train passage (every 2 or 3 days), and viable cell counts were conducted using trypan blue exclusion on a CDV™ instrument (Nova Biomedical). Medium 1 was tested unsupplemented or supplemented with ornithine • HCl at 100 mg/L or both putrescine • 2HCl at 115 mg/L and ornithine • HCl at 100 mg/L. Medium 2 with low putrescine • 2HCl (0.4 mg/L) was tested unsupplemented or supplemented with ornithine • HCl at 100 mg/L or both putrescine • 2HCl at 115 mg/L and ornithine • HCl at 100 mg/L. The results are depicted in Tables 3 and 4. Ornithine supplementation, either with or without putrescine, to Medium 1 was required to achieve significant growth. Supplementing hydrolysate free Medium 2 with ornithine or ornithine + putrescine decreased the cell doubling time by about 25% to 30%.

Doubling times were also reduced to a lesser extent upon the addition of ornithine or ornithine + putrescine to hydrolysate containing Medium 2.

[00094] TABLE 3: CELL DOUBLING TIME (HOURS) AND APPROXIMATE PERCENT DOUBLING TIME DECREASE RELATIVE TO BASELINE* IN MEDIUM 1

Supplement	Medium 1	
*None	75	
Ornithine	23	69%
Putrescine + ornithine	21	72%

*Baseline is unsupplemented Medium1.

[00095] TABLE 4: CELL DOUBLING TIME (HOURS) AND APPROXIMATE PERCENT DOUBLING TIME DECREASE RELATIVE TO BASELINE* IN MEDIUM 2

Supplement	Medium 2		Medium 2 + Soy	
*Unsupplemented	27		22.5	
Ornithine	21	22%	20.5	8.9%
Putrescine + ornithine	19.5	28%	21	6.7%

*Baseline is unsupplemented Medium 2.

EXAMPLE 3: IMPROVED ANTIBODY TITERS

[00096] Having established that the inclusion of ornithine or ornitihine + putrescine improves cell proliferation and viable cell density in culture, we further investigated the effect of those conditions on recombinant protein production titers. We examined the expression and secretion of recombinant IgG by a CHO-K1 derived cell line. In this experiment, the average antibody titer was determined at day seven in culture under various media formats. As above, Medium 1 with low putrescine (0.4mg/L putrescine • 2HCl), ornithine (100 mg/L ornithine • HCl), and both ornithine and putrescine (100 mg/L ornithine • HCl /115 mg/L putrescine • 2HCl) were tested. Medium 2 and Medium 2 + Soy with low putrescine (0.4mg/L putrescine • 2HCl), ornithine (100 mg/L ornithine • HCl), and both ornithine and putrescine (100 mg/L ornithine • HCl /115 mg/L putrescine • 2HCl) were also tested. In all cases, the inclusion of ornithine or ornithine and putrescine at a level above 0.4 mg/L resulted in a significantly larger protein titer, *i.e.,* at least about two-fold higher titers. The results are depicted in Table 5.

[00097] TABLE 5: AVERAGE SEVEN-DAY ANTIBODY TITERS AND APPROXIMATE FOLD INCREASE (X) IN TITER RELATIVE TO BASELINE*

Supplement	Medium 1		Medium 2		Medium 2 + Soy	
Unsupplemented*	0.31 g/L	1X	0.29 g/L	1X	0.54 g/L	1X
Ornithine	0.94 g/L	3.0X	0.65 g/L	2.2X	0.98 g/L	1.8X
Putrescine + ornithine	0.95 g/L	3.1X	0.64 g/L	2.2X	1.07 g/L	2X

* Baseline is set at titer in for unsupplemeted media of each type.

[00098] We also examined the effect of various amounts of ornithine • HCl (*i.e.,* 50 mg/mL, 100 mg/mL, and 150 mg/mL) in Medium 3 on antibody production. A single seed train culture of a recombinant antibody producing cell line derived from CHO K1 was used to seed 50 mL TubeSpin® Bioreactors (TPP) at 0.4 X 10^6 cells/mL at a 15 mL working volume. The cells were grown in a 37°C incubator for three days. All three levels of ornithine supplementation improved antibody titer on the average (N=3) by slightly more than 50%. The results are depicted in Table 6.

[00099] TABLE 6: ANTIBODY TITERS (GRAMS PER LITER) AND X FOLD INCREASE IN TITER OVER BASELINE*

	Control	Ornithine HCl		
Supplement concentration (mg/mL)	0	50	100	150
Antibody titer (mg/mL)	79	120	127	124
Fold increase over control	1X	1.5X	1.6X	1.6X

*Base line is unsupplemented Medium 3.

WHAT IS CLAIMED IS:

1. A cell culture medium, which is serum-free, comprising ≥ 0.09 mM ± 0.014 mM ornithine.
2. The cell culture medium of claim 1 comprising ≥ 0.20 ± 0.03 mM putrescine.
3. The cell culture medium of any one of claims 1 and 2 comprising 0.09 ± 0.014 mM to 0.9 ± 0.14 mM ornithine.
4. The cell culture medium of any one of claims 1-3 comprising ornithine at 0.09 ± 0.014 mM, 0.3 ± 0.05 mM, 0.6 ± 0.09 mM, or 0.9 ± 0.14 mM.
5. The cell culture medium of any one of claims 1-4 comprising 0.20 ± 0.03 mM to 0.714 ± 0.11 mM putrescine.

6. The cell culture of any one of claims 1-4 comprising putrescine at 0.20 ± 0.03 mM, 0.35 ± 0.06, or 0.714 ± 0.11 mM.
7. The cell culture medium of any one of claims 1-6, wherein the medium is hydrolysate-free.
8. The cell culture medium of any one of claims 1-6, wherein the medium is chemically defined.
9. The cell culture medium of any one of claims 1-8 comprising ≥ 40 ± 6 mM of a mixture of amino acids or salts thereof.
10. The cell culture medium of claim 9, wherein the mixture of amino acids consists of alanine, arginine, asparagine, aspartic acid, cysteine, glutamic acid, glycine, histidine, isoleucine, leucine, lysine, methionine, phenylalanine, proline, serine, threonine, tryptophan, tyrosine, and valine.
11. The cell culture medium of any one of claims 1-10 comprising one or more fatty acids.
12. The cell culture medium of claim 11, wherein the one or more fatty acids are selected from the group consisting of linoleic acid, linolenic acid, thioctic acid, oleic acid, palmitic acid, stearic acid, arachidic acid, arachidonic acid, lauric acid, behenic acid, decanoic acid, dodecanoic acid, hexanoic acid, lignoceric acid, myristic acid, and octanoic acid.
13. The cell culture medium of any one of claims 1-12 comprising a mixture of nucleosides.
14. The cell culture medium of claim 13, wherein the mixture of nucleosides comprises one or more of adenosine, guanosine, cytidine, uridine, thymidine, and hypoxanthine.
15. The cell culture medium of any one of claims 1-14 comprising adenosine, guanosine, cytidine, uridine, thymidine, and hypoxanthine.
16. The cell culture medium of any one of claims 1-15 comprising one or more divalent cations.
17. The cell culture medium of claim 16, wherein the divalent cation is magnesium, calcium, or both.
18. The cell culture medium of any one of claims 1-17 comprising Ca^{2+} and Mg^{2+}.
19. A method for cultivating cells, comprising the steps of: (a) providing a cell culture medium according to any one of claims 1-18, and (b) propagating or maintaining a cell in the cell culture medium to form a cell culture.
20. The method of claim 19, wherein the cell is selected from the group consisting of mammalian cell, avian cell, insect cell, bacterial cell, and yeast cell.
21. The method of claim 19 or claim 20, wherein the cell is a CHO cell.
22. The method of any one of claims 19-21, wherein the cell expresses a protein of interest.
23. The method of claim 22, wherein the protein of interest is an antigen binding protein.
24. The method of claims 22 or 23, wherein the protein of interest comprises an Fc domain.
25. The method of any one of claims 22-24, wherein the protein of interest is a receptor-Fc-fusion protein.
26. The method of claim 25, wherein the receptor-Fc-fusion protein is a trap protein.
27. The method of claim 26, wherein the trap protein is an IL-1 antagonist or a VEGF antagonist.

28. The method of claims 22 or 23, wherein the protein of interest is an antibody or an antibody fragment.
29. The method of claim 28, wherein the antibody or the antibody fragment is a recombinant human antibody or fragment thereof.
30. The method of any one of claims 19-29, wherein the cells have an average doubling time of ≤ 30 hours.
31. The method of any one of claims 19-30, wherein the cells have an average doubling time of ≤ 24 hours.
32. The method of any one of claims 19-31, wherein the cells have an average doubling time that is at least one third that of cells grown in a cell culture medium that contains < 0.3 ± 0.045 mM ornithine and < 0.2 ± 0.03 mM putrescine.
33. The method of any one of claims 19-32, wherein the cell culture is capable of attaining a viable cell count density that is at least 15% greater than a similar cell culture in media that contains < 0.09 ± 0.014 mM ornithine and < 0.2 ± 0.03 mM putrescine.
34. The method of any one of claims 19-33, wherein the cell culture is capable of attaining a viable cell count density that is at least 3-fold greater than a similar cell culture in a similar cell culture medium that contains < 0.09 ± 0.014 mM ornithine and < 0.2 ± 0.03 mM putrescine.
35. The method of any one of claims 19-34 comprising the step of adding one or more point-of-use additions to the cell culture medium.
36. The method of claim 35, wherein the point-of-use additions comprise one of more of $NaHCO_3$, glutamine, insulin, glucose, $CuSO_4$, $ZnSO_4$, $FeCl_3$, $NiSO_4$, Na_4 EDTA, and Na_3 Citrate.
37. The method of claim 35 or 36, wherein each of $NaHCO_3$, glutamine, insulin, glucose, $CuSO_4$, $ZnSO_4$, $FeCl_3$, $NiSO_4$, Na_4 EDTA, and Na_3 Citrate are added to the medium as point-of-use additions.
38. A method for producing a protein comprising the steps of: (a) introducing into a cell a nucleic acid comprising a sequence encoding a protein of interest; (b) selecting a cell carrying the nucleic acid; (c) culturing the selected cell in a cell culture medium of any one of claims 1-18 or according to the method of any one of claims 19-37; and (d) expressing the protein of interest in the cell, wherein the protein of interest is secreted into the medium.
39. The method of claim 38, wherein the cell is a CHO cell, 293 cell or BHK cell.
40. The method of claim 38 or 39, wherein the protein of interest is an antigen-binding protein.
41. The method of any one of claims 38-40, wherein the protein of interest comprises an Fc domain.
42. The method of any one of claims 38-41, wherein the protein of interest is selected from the group consisting of receptor-Fc-fusion protein (TRAP), soluble TCR-Fc fusion protein, antibody, Fc-fusion protein, and ScFv protein.

43. The method according to any one of claims 38-42, wherein the protein-of-interest is produced at an average 7-day titer that is at least 7% greater than the average 7-day titer produced by a similar cell in a cell culture medium that contains less than 0.09 ± 0.014 mM ornithine and less than 0.2 ± 0.03 mM putrescine.
44. The method according to any one of claims 38-43, wherein the protein-of-interest is produced at an average 7-day titer that is at least 14% greater than the average 7-day titer produced by a similar cell in a cell culture medium that contains less than 0.09 ± 0.014 mM ornithine and less than 0.2 ± 0.03 mM putrescine.
45. The method according to any one of claims 38-44, wherein the protein-of-interest is produced at an average 7-day titer that is at least 80% greater than the average 7-day titer produced by a similar cell in a cell culture medium that contains less than 0.09 ± 0.014 mM ornithine and less than 0.2 ± 0.03 mM putrescine.
46. The method according to any one of claims 38-45, wherein the protein-of-interest is produced at an average 7-day titer that is at least 2-fold greater than the average 7-day titer produced by a similar cell in a cell culture medium that contains less than 0.09 ± 0.014 mM ornithine and less than 0.2 ± 0.03 mM putrescine.
47. The method according to any one of claims 38-46, wherein the protein-of-interest is produced at an average 7-day titer that is at least 3-fold greater than the average 7-day titer produced by a similar cell in a cell culture medium that contains less than 0.09 ± 0.014 mM ornithine and less than 0.2 ± 0.03 mM putrescine.
48. The method according to any one of claims 38-47, wherein the protein-of-interest is a recombinant human antibody.