Human anti-B7RP1 Neutralizing Antibodies

This application claims priority to U.S. provisional patent application, Serial No. 60/700,265, filed July 18, 2005, the disclosure of which is explicitly incorporated by reference herein.

FIELD OF THE INVENTION

The invention relates to human monoclonal antibodies that bind B7 related protein-1 (B7RP1). Compositions and methods for treating diseases and disorders related to immunosuppression and immune activation are also described.

BACKGROUND OF THE INVENTION

T-cells initiate the immune response, mediate antigen-specific effector functions, and regulate the activity of other leukocytes by secreting cytokines. For the generation of a proper T-lymphocyte (T-cell) immune response, two signals must be provided to the T-cell by antigen presenting cells (APC). Antigen must be presented to the T-cell receptor (TCR) via a major histocompatibility complex (MHC), in an event that determines specificity. T-cells can only recognize antigen presented on an APC. In addition to the antigen receptor, proper T-cell activation also requires the interaction of other cell-surface molecules on both the T-cell and the APC. These molecules, referred to as co-stimulatory molecules, consist of a receptor on the responding cell and a ligand present on the inducer cell. This antigen independent, co-stimulatory signal must be delivered by engagement of members of the B7 family on the APC with their receptors on T-cells. A productive immune response leads to proliferation, differentiation, clonal expansion, and effector function. In the absence of the second, co-stimulatory signal, T-cells undergo a state of long-lasting antigen-specific unresponsiveness, termed anergy. Phase II clinical experiments have demonstrated that blocking one co-stimulation pathway is efficacious in the treatment of psoriasis (Abrams *et al.*, 2000, *J Exp Med.* 192:681-94; Abrams *et al.*, 1999, *J. Clin. Invest.* 103:1243-52) and rheumatoid arthritis (Kremer *et al.*, 2003, *New*

England Journal of Medicine 349:1907-15), indicating that this general strategy is a good target for immunomodulatory therapy.

A particular co-stimulatory B7 molecule, B7 related protein-1 (B7RP1), is a type 1 transmembrane protein with a signal sequence and extracellular domain at the amino-terminus, an extracellular domain comprising two Ig loops, a transmembrane domain, and a carboxy terminal intracellular domain (PCT Application Publication No. WO 00/46240). B7RP1 preferentially binds to ICOS (which stands for "inducible costimulator"; Yoshinga *et al.*, 2000, *Int. Immun.* 12:1439-1447) expressed on the cell surface of T-cells. ICOS plays an important role in the production of both type 1 and type 2 cytokines by activated T-cells (Coyle *et al.*, 2000, *Immunity* 13:95-105).

B7RP1 is the sole ligand expressed constitutively on APCs (Yoshinaga *et al.*, 1999, *Nature.* 402:827-32), while ICOS is expressed only on activated T-cells (McAdam *et al.*, 2000, *Journal of Immunology* 165:5035-40). B7RP1-dependent signaling is required for the activation of the effector (*i.e.* fully activated) T-cell, as well as its maturation from its naïve precursor (Dong *et al.*, 2003, *Journal of Autoimmunity.* 21:255-60; Coyle *et al.*, 2000, *Immunity.* 13:95-105). Consequently, the B7RP1/ICOS interaction is required for proper T-cell-dependent recall immune responses (Dong *et al.*, 2003, *Journal of Autoimmunity.* 21:255-60).

Current attempts to interfere with the co-stimulatory T-cell pathway have focused primarily on co-stimulatory polypeptides that block T-cell activation only, but have not focused on activation and maturation. Consequently, these therapies provide general inhibition of T-cell function. In contrast, blocking the B7RP1/ICOS interaction provides a more specific inhibition of T-cell function by affecting only mature effector T-cells. Thus, blocking the B7RP1/ICOS interaction in a clinical setting is highly desirable because it would provide a more limited side-effect profile than co-stimulation therapies that block naïve T-cell activation only.

SUMMARY OF THE INVENTION

In a first aspect, the present invention refers to an isolated antibody that binds specifically to B7RP1, wherein the antibody
comprises a heavy chain having a heavy chain variable region and a light chain having a light chain variable region, wherein the heavy chain variable region comprises an amino acid sequence that has at least 98% sequence identity to the amino acid sequence set forth in SEQ ID NO: 9, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof, and the light chain variable region comprises an amino acid sequence that has at least 98% sequence identity to the amino acid sequence set forth in SEQ ID NO: 2, or an antigen-binding or immunogenic fragment thereof.

In a second aspect, the present invention refers to the use of a therapeutically effective amount of the antibody of the present invention, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof, in the manufacture of a medicament for treating an autoimmune disease or inflammatory response in a patient.

In a third aspect, the present invention refers to the use of a therapeutically effective amount of the antibody of the present invention, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof, in the manufacture of a medicament for inhibiting co-stimulatory signal T-cell activation in a patient.

In a fourth aspect, the present invention refers to a pharmaceutical composition comprising a pharmaceutically acceptable carrier and a therapeutically effective amount of the antibody of the present invention, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof.

In a fifth aspect, the present invention refers to a method for detecting B7RP1 in a biological sample comprising:

a) contacting the sample with the antibody of the present invention, under conditions that allow for binding of the antibody to B7RP1; and

b) measuring the level of bound antibody in the sample.

In a sixth aspect, the present invention refers to a nucleic acid molecule that encodes the antibody of the present invention.

In a seventh aspect, the present invention refers to a vector comprising the nucleic acid molecule of the present invention.

In an eighth aspect, the present invention refers to a host cell comprising the nucleic acid of the present invention or the vector of the present invention.

In a ninth aspect, the present invention refers to a host cell comprising a first vector and a second vector, wherein the first vector comprises a nucleic acid molecule encoding an amino acid sequence that has at least 98% sequence identity to the amino acid sequence set forth in SEQ ID NO: 9, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof, and wherein the second vector comprises a nucleic acid molecule encoding an amino acid sequence that has at least 98% sequence identity to the amino acid sequence set forth in SEQ ID NO: 2, or an antigen-binding or immunogenic fragment thereof.

In a tenth aspect, the present invention refers to an isolated cell line that produces an antibody according to the present invention.

In an eleventh aspect, the present invention refers to a method of producing an antibody comprising culturing the host cell of the present invention, or the cell line of the present invention, and collecting the antibody from the culture medium or the host cell.

In a twelfth aspect, the present invention refers to the use of the pharmaceutical composition of the present invention in the manufacture of a medicament for treating a condition caused by increased expression of B7RP1 or increased sensitivity to B7RP1 in a patient.

Disclosed herein are monoclonal antibodies that bind to B7 related protein-1 (B7RP1). In one example, the monoclonal antibodies are human monoclonal antibodies that neutralize biological activities of B7RP1 and are particularly useful for inhibiting partially or completely the immune co-stimulatory activity of B7RP1. Also

provided are cells, particularly hybridoma cells that produce the monoclonal antibodies described herein. In particular aspects, the antibodies described herein bind specifically to the H or D region of B7RP1 as described herein.

Also disclosed are fusion proteins comprising the sequence of an antibody Fc region and one or more sequences identified as SEQ ID NO: 1 through SEQ ID NO. 40. Such molecules can be prepared using methods as described, for example, in International Patent Application, Publication No. WO 00/24782, which is hereby incorporated by reference. Such molecules can be expressed, for example, in mammalian cells (*e.g.* Chinese Hamster Ovary cells) or bacterial cells (*e.g. E. coli* cells).

In certain aspects, the disclosure provides antibodies comprising a heavy chain and a light chain, wherein the heavy chain comprises an heavy chain constant region selected from IgG1, IgG2, IgG3, IgG4, IgM, IgA and IgE heavy chain constant regions or any allelic variation thereof (as discussed in Kabat *et al.*, 1991, SEQUENCES OF PROTEINS OF IMMUNOLOGICAL INTEREST, Fifth Edition, U.S. Department of Health and Human Services, NIH Publication No. 91-3242), incorporated herein by reference, and the variable region of the heavy chain comprises an amino acid sequence as set forth in any of SEQ ID NO: 7 through SEQ ID NO. 14, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof. An antibody described herein comprises either an amino acid sequence of the IgG2 heavy chain constant region as set forth in SEQ ID NO: 41 or an antigen-binding or an immunologically functional immunoglobulin fragment thereof, or an amino acid sequence of the IgG1 heavy chain constant region as set forth in SEQ ID NO: 42 or an antigen-binding or an immunologically functional immunoglobulin fragment thereof. In certain examples, the antibodies are monoclonal antibodies, human antibodies, or preferably human monoclonal antibodies.

In certain aspects, the disclosure provides antibodies comprising a heavy chain and a light chain, wherein the light chain comprises a constant region having an amino acid sequence as set forth in SEQ ID NO: 43 or an antigen-binding or an immunologically functional immunoglobulin fragment thereof, and the light chain variable region comprises an amino acid sequence as set forth in any of SEQ ID NO: 1 through SEQ ID NO. 6, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof. In certain examples, the antibodies are

monoclonal antibodies, human antibodies, or preferably human monoclonal antibodies.

In certain aspects, antibodies described herein comprise a heavy chain and a light chain, wherein the variable region of the heavy chain comprises an amino acid sequence as set forth in SEQ ID NO: 7 or SEQ ID NO: 8, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof. In other aspects, the light chain variable region comprises an amino acid sequence as set forth in SEQ ID NO: 1, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof.

In other aspects, antibodies described herein comprise a heavy chain and a light chain, wherein the variable region of the heavy chain comprises an amino acid sequence as set forth in SEQ ID NO: 9, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof. In other aspects, the light chain variable region comprises an amino acid sequence as set forth in SEQ ID NO: 2, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof.

In additional aspects, the heavy chain comprises at least one complementarity determining region (CDR) having an amino acid sequence as set forth in any of SEQ ID NO: 27 through SEQ ID NO. 40, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof. In still further aspects, the light chain comprises at least one CDR having an amino acid sequence as set forth in any of SEQ ID NO: 15 through SEQ ID NO. 26, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof.

The disclosure also provides antibodies that bind specifically to B7RP1, wherein the heavy chain comprises a variable region comprising an amino acid sequence as set forth in SEQ ID NO: 7 or SEQ ID NO: 8, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof, and the light chain comprises a variable region comprising an amino acid sequence as set forth in SEQ ID NO: 1, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof.

In addition, described herein are antibodies that bind specifically to B7RP1, wherein the heavy chain comprises a variable region comprising an amino acid sequence as set forth in SEQ ID NO: 9, or an antigen-binding or an immunologically

functional immunoglobulin fragment thereof, and the light chain comprises a variable region comprising an amino acid sequence as set forth in SEQ ID NO: 2, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof.

In certain aspects, the disclosure also provides antibodies, comprising a heavy chain and a light chain, wherein the heavy chain comprises a heavy chain variable region, and wherein the heavy chain variable region comprises a sequence that has at least about 75%, at least about 80%, at least about 85%, at least about 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or at least about 99% identity to the amino acid sequence as set forth in any of SEQ ID NO: 7 through SEQ ID NO. 14, and wherein the light chain comprises a light chain variable region, and wherein the light chain variable region comprises a sequence that has at least about 80%, at least about 85%, at least about 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or at least about 99% identity to the amino acid sequence as set forth in any of SEQ ID NO: 1 through SEQ ID NO. 6, wherein the antibody binds specifically to B7RP1.

The disclosure also provides antibodies that bind specifically to B7RP1, wherein the heavy chain comprises an amino acid sequence as set forth in SEQ ID NO: 44 or SEQ ID NO: 46, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof, and the light chain comprises an amino acid sequence as set forth in SEQ ID NO: 45, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof.

The disclosure also provides antibodies that bind specifically to B7RP1, wherein the heavy chain comprises an amino acid sequence as set forth in SEQ ID NO: 47, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof, and the light chain comprises an amino acid sequence as set forth in SEQ ID NO: 48, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof.

In certain aspects, the disclosure provides antibodies, comprising a heavy chain and a light chain, wherein the heavy chain comprises a heavy chain variable region, and wherein the heavy chain variable region comprises at least one CDR having a sequence that has at least about 75%, at least about 80%, at least about 85%, at least about 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or at least about 99% identity to the amino acid sequence as set forth in any of SEQ ID NO: 27 through

SEQ ID NO. 40, and wherein the light chain comprises a light chain variable region, and wherein the light chain variable region comprises at least one CDR having an amino acid sequence that has as least about 80%, at least about 85%, at least about 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or at least about 99% identity to the amino acid sequence as set forth in SEQ ID NO: 15 through SEQ ID NO. 26, wherein the antibody binds specifically to B7RP1.

Also described herein are single chain antibodies, single chain Fv antibodies, F(ab) antibodies, F(ab)' antibodies and $(Fab')_2$ antibodies.

In particular aspects, the disclosure provides a light chain comprising an amino acid sequence as set forth in SEQ ID NO: 15 through SEQ ID NO. 26, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof.

In addition, the disclosure provides a heavy chain comprising an amino acid sequence as set forth in any of SEQ ID NO: 27 through SEQ ID NO. 40, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof.

The disclosure also relates to isolated human antibodies that specifically bind B7RP1, wherein the antibody comprises: (a) human heavy chain framework regions, a human heavy chain CDR1 region, a human heavy chain CDR2 region, and a human heavy chain CDR3 region; and (b) human light chain framework regions, a human light chain CDR1 region, a human light chain CDR2 region, and a human light chain CDR3 region. In certain aspects, the human heavy chain CDR1 region can be the heavy chain CDR1 region as shown in any of SEQ ID NO: 27, 30, or 35 and the human light chain CDR1 region can be the light chain CDR1 region shown in any of SEQ ID NO: 15, 18, or 24. In other aspects, the human heavy chain CDR2 region can be the heavy chain CDR2 region as shown in any of SEQ ID NO: 28, 31, 33, 36, or 39, and the human light chain CDR2 region can be the light chain CDR2 as shown in any of SEQ ID NO: 16, 19, or 21. In still other aspects, the human heavy chain CDR3 region is the heavy chain CDR3 region as shown in any of SEQ ID NO: 29, 32, 34, 37, 38 or 40, and the human light chain CDR3 region is the light chain CDR3 region as shown in any of SEQ ID NO: 17, 20, 22, 23, 25, or 26.

The antibodies described herein are characterized by the ability to bind specifically to B7RP1. Furthermore, antibodies described herein have the capacity to antagonize at least one *in vitro* and/or *in vivo* activity associated with B7RP1

polypeptides. The disclosure provides isolated anti-human B7RP1 human antibodies with high affinity binding to B7RP1 polypeptides, wherein the antibodies bind to a human B7RP1 polypeptide and dissociates from the human B7RP1 polypeptide with a dissociation constant (K_D) of about 10^{-6} M, 10^{-7} M, 10^{-8} M, 10^{-9} M, 10^{-10} M, 10^{-11} M, 10^{-12} M, or less, as determined using KinExA, or which inhibit B7RP1 induced survival in an *in vitro* neutralization assay with an EC_{50} of about 10^{-6} M, 10^{-7} M, 10^{-8} M, 10^{-9}M, 10^{-10} M, 10^{-11}M, 10^{-12} M, or less.

The disclosure also provides isolated human antibodies or an antigen-binding or immunologically functional immunoglobulin fragments thereof that bind specifically to B7RP1, wherein the antibodies or fragments comprise a heavy chain variable region comprising a heavy chain CDR1, CDR2, and CDR3, wherein:

a) the heavy chain CDR1 has an amino acid sequence as set forth in SEQ ID NO: 27, the heavy chain CDR2 has an amino acid sequence as set forth in SEQ ID NO: 28, and the heavy chain CDR 3 has an amino acid sequence as set forth in SEQ ID NO: 29;

b) the heavy chain CDR1 has an amino acid sequence as set forth in SEQ ID NO: 30, the heavy chain CDR2 has an amino acid sequence as set forth in SEQ ID NO: 31, and the heavy chain CDR 3 has an amino acid sequence as set forth in SEQ ID NO: 32;

c) the heavy chain CDR1 has an amino acid sequence as set forth in SEQ ID NO: 27, the heavy chain CDR2 has an amino acid sequence as set forth in SEQ ID NO: 33, and the heavy chain CDR 3 has an amino acid sequence as set forth in SEQ ID NO: 34;

d) the heavy chain CDR1 has an amino acid sequence as set forth in SEQ ID NO: 35, the heavy chain CDR2 has an amino acid sequence as set forth in SEQ ID NO: 36, and the heavy chain CDR 3 has an amino acid sequence as set forth in SEQ ID NO: 37;

e) the heavy chain CDR1 has an amino acid sequence as set forth in SEQ ID NO: 27, the heavy chain CDR2 has an amino acid sequence as set forth in SEQ ID NO: 33, and the heavy chain CDR 3 has an amino acid sequence as set forth in SEQ ID NO: 38; or

f) the heavy chain CDR1 has an amino acid sequence as set forth in SEQ ID NO: 35, the heavy chain CDR2 has an amino acid sequence as set forth in SEQ ID NO: 39, and the heavy chain CDR 3 has an amino acid sequence as set forth in SEQ ID NO: 40.

The disclosure also provides an isolated human antibody or an antigen-binding or an immunologically functional immunoglobulin fragment thereof that binds specifically to B7RP1, wherein the antibody or fragment comprises a light chain variable region comprising a light chain CDR1, CDR2, and CDR3, wherein:

a) the light chain CDR1 has an amino acid sequence as set forth in SEQ ID NO: 15, the light chain CDR2 has an amino acid sequence as set forth in SEQ ID NO: 16, and the light chain CDR 3 has an amino acid sequence as set forth in SEQ ID NO: 17;

b) the light chain CDR1 has an amino acid sequence as set forth in SEQ ID NO: 18, the light chain CDR2 has an amino acid sequence as set forth in SEQ ID NO: 19, and the light chain CDR 3 has an amino acid sequence as set forth in SEQ ID NO: 20;

c) the light chain CDR1 has an amino acid sequence as set forth in SEQ ID NO: 15, the light chain CDR2 has an amino acid sequence as set forth in SEQ ID NO: 21, and the light chain CDR 3 has an amino acid sequence as set forth in SEQ ID NO: 22;

d) the light chain CDR1 has an amino acid sequence as set forth in SEQ ID NO: 18, the light chain CDR2 has an amino acid sequence as set forth in SEQ ID NO: 19, and the light chain CDR 3 has an amino acid sequence as set forth in SEQ ID NO: 23;

e) the light chain CDR1 has an amino acid sequence as set forth in SEQ ID NO: 24, the light chain CDR2 has an amino acid sequence as set forth in SEQ ID NO: 16, and the light chain CDR 3 has an amino acid sequence as set forth in SEQ ID NO: 25; or

f) the light chain CDR1 has an amino acid sequence as set forth in SEQ ID NO: 24, the light chain CDR2 has an amino acid sequence as set forth in SEQ ID NO: 16, and the light chain CDR 3 has an amino acid sequence as set forth in SEQ ID NO: 26.

The disclosure also provides antibodies that compete with binding of the antibodies described herein to B7RP1. In certain aspects, a competitive antibody described herein competes with binding of an antibody that comprises any of SEQ ID NO: 1-40 to human B7RP1.

Also part of the disclosure are polynucleotide sequences that encode anti-human B7RP1 human antibodies, vectors comprising the polynucleotide sequences encoding anti-human B7RP1 human antibodies, host cells transformed with vectors incorporating polynucleotides that encode anti-human B7RP1 human antibodies, formulations comprising anti-human B7RP1 human antibodies and methods of making and using same.

The disclosure also provides methods for detecting B7RP1 in a biological sample, comprising the step of contacting the sample with an antibody described herein or antigen-binding fragment thereof. An anti-B7RP1 antibody described herein may be employed in any known assay method, such as competitive binding assays, direct and indirect sandwich assays, immunoprecipitation assays and enzyme-linked immunosorbent assays (ELISA) (*See*, Sola, 1987, Monoclonal Antibodies: A Manual of Techniques, pp. 147-158, CRC Press, Inc.) for the detection and quantitation of B7RP1. The antibodies can bind B7RP1 with an affinity that is appropriate for the assay method being employed.

In addition, the disclosure provides methods for treating a disease associated with increased production of B7RP1, increased sensitivity to B7RP1, and/or diseases related to control of T-cell responses, comprising the step of administering a pharmaceutically effective amount of a pharmaceutical composition comprising at least one antibody described herein or an antigen-binding or an immunologically functional immunoglobulin fragment thereof to an individual in need thereof.

Embodiments of the invention will become evident from the following detailed description and the claims.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1A depicts the 16H antibody variable region sequence (SEQ ID NO: 7) and the corresponding 16H variable region germline (16Hg) sequence (SEQ ID NO: 8).

Figure 1B depicts results of co-stimulation assays using anti-CD3 and hB7RP1-Fc fusion protein demonstrating that 16Hg retains its biological activities compared with 16H.

Figure 2 shows the results of Biacore® binding assays with 16H, 16Hg, and 5D antibodies.

Figure 3 shows the results of KinExA binding assay with 5D antibody.

Figure 4 shows the results of KinExA binding assay with 2H antibody.

Figure 5 shows the results of KinExA binding assay with 2H germline (2Hg) antibody.

Figure 6 depicts the results of binding-competition assays showing that 16H antibody competes away binding of ICOS-Fc on B7RP-1, analyzed by flow cytometry.

Figure 7 depicts a summary of a B7RP-1 single nucleotide polymorphism (SNP) analysis.

Figure 8 depicts a summary of the analysis of a set of anti-human B7RP-1 monoclonal antibodies in ELISA competition assays. Values shown are IC_{50}s for inhibition of binding of an ICOS-Fc fusion protein.

Figure 9A shows fluorescent staining of B7RP1 extracellular domain (ECD) with labeled 16H, 5D, and ICOS antibodies.

Figure 9B shows similar binding efficacy of 16H and 5D antibodies to a B7RP1 SNP variant.

Figure 9C depicts the results of co-stimulation assays with 16H or 5D antibodies and SNP variants.

Figure 10A shows plate co-stimulation assay results with 1B7v2 monoclonal antibodies compared with a number of different anti-murine B7RP-1 monoclonal antibodies.

Figures 10B, 10C, and 10D show the results of antigen challenge experiments, analyzed for antigen-specific serum IgM (Figure 10B), IgG2a (Figure 10C), and IgG1 (Figure 10D).

Figure 11 depicts ELISA results demonstrating that serum IL-5 levels are repressed by 1B7v2 antibodies.

Figure 12A shows that 16H antibodies can bind to cynomolgus monkey B7RP1 (right panel) and human B7RP1 (left panel).

Figure 12B shows that 16H, 16Hg, and 5D antibodies can inhibit cynomolgus monkey B7RP1/ICOS-dependent T cell activation.

Figure 13A depicts individual cynomolgus monkey and group mean titer values at day 53 and day 57 after secondary challenge with tetanus toxoid on day 42 in animals treated with 16H antibodies.

Figure 13B depicts individual cynomolgus monkey and group mean titer values at day 53 and day 57 after secondary challenge with tetanus toxoid on day 42 in animals treated with 5D antibodies.

DETAILED DESCRIPTION OF CERTAIN EMBODIMENTS

The section headings used herein are for organizational purposes only and are not to be construed as limiting the subject matter described. All references cited in this application are expressly incorporated by reference herein for any purpose.

Definitions

Conventional techniques may be used for recombinant DNA, oligonucleotide synthesis, and tissue culture and transformation (*e.g.*, electroporation, lipofection). Enzymatic reactions and purification techniques may be performed according to manufacturer's specifications or as commonly accomplished in the art or as described herein. The foregoing techniques and procedures may be generally performed according to methods well known in the art and as described in various general and more specific references that are cited and discussed throughout the present specification. *See e.g.*, Sambrook *et al.*, 2001, MOLECULAR CLONING: A LABORATORY MANUAL, 3d ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., which is incorporated herein by reference for any purpose. Unless specific definitions are provided, the nomenclature utilized in connection with, and the laboratory procedures and techniques of, analytical chemistry, synthetic organic

chemistry, and medicinal and pharmaceutical chemistry described herein are those well known and commonly used in the art. Similarly, conventional techniques may be used for chemical syntheses, chemical analyses, pharmaceutical preparation, formulation, and delivery, and treatment of patients.

As utilized in accordance with the present disclosure, the following terms, unless otherwise indicated, shall be understood to have the following meanings. The phrases "biological property", "biological characteristic", and the term "activity" in reference to an antibody of the present invention are used interchangeably herein and include, but are not limited to, epitope affinity and specificity (*e.g.*, anti-human B7RP1 human antibody binding to human B7RP1), ability to antagonize the activity of the targeted polypeptide (*e.g.*, B7RP1 activity), the *in vivo* stability of the antibody, and the immunogenic properties of the antibody. Other identifiable biological properties or characteristics of an antibody recognized in the art include, for example, cross-reactivity, (*i.e.*, with non-human homologs of B7RP1, or with other proteins or tissues, generally), and ability to preserve high expression levels of protein in mammalian cells. The aforementioned properties or characteristics can be observed or measured using art-recognized techniques including, but not limited to ELISA, competitive ELISA, surface plasmon resonance analysis, *in vitro* and *in vivo* neutralization assays (*e.g.*, Example 2), and immunohistochemistry with tissue sections from different sources including human, primate, or any other appropriate source. Particular activities and biological properties of anti-human B7RP1 human antibodies are described in further detail in the Examples below.

The term "isolated polynucleotide" as used herein shall mean a polynucleotide of genomic DNA, cDNA, RNA, or synthetic origin or some combination thereof, which by virtue of its origin the isolated polynucleotide (1) is not associated with all or a portion of a polynucleotide with which the isolated polynucleotide is found in nature, (2) is linked to a polynucleotide to which it is not linked in nature, or (3) does not occur in nature as part of a larger sequence.

The term "polynucleotide" as referred to herein means single-stranded or double-stranded nucleic acid polymers of at least 10 nucleotides in length. In certain embodiments, the nucleotides comprising the polynucleotide can be ribonucleotides or deoxyribonucleotides or a modified form of either type of nucleotide. Said modifications include base modifications such as bromuridine, ribose modifications

such as arabinoside and 2',3'-dideoxyribose and internucleotide linkage modifications such as phosphorothioate, phosphorodithioate, phosphoroselenoate, phosphorodiselenoate, phosphoroanilothioate, phoshoraniladate and phosphoroamidate. The term "polynucleotide" specifically includes single and double stranded forms of DNA or RNA.

The term "oligonucleotide" referred to herein includes naturally occurring, and modified nucleotides linked together by naturally occurring, and/or non-naturally occurring oligonucleotide linkages. Oligonucleotides are a polynucleotide subset comprising members that are generally single-stranded and have a length of 200 nucleotides or fewer. In certain embodiments, oligonucleotides are 10 to 60 nucleotides in length. In certain embodiments, oligonucleotides are 12, 13, 14, 15, 16, 17, 18, 19, or 20 to 40 nucleotides in length. Oligonucleotides may be single stranded or double stranded, *e.g.* for use in the construction of a genetic mutant. Oligonucleotides of the invention may be sense or antisense oligonucleotides with reference to a protein-coding sequence.

The term "naturally occurring nucleotides" includes deoxyribonucleotides and ribonucleotides. The term "modified nucleotides" includes nucleotides with modified or substituted sugar groups and the like. The term "oligonucleotide linkages" includes oligonucleotide linkages such as phosphorothioate, phosphorodithioate, phosphoroselenoate, phosphorodiselenoate, phosphoroanilothioate, phoshoraniladate, phosphoroamidate, and the like. *See, e.g.*, LaPlanche *et al.*, 1986, *Nucl. Acids Res.*, 14:9081; Stec *et al.*, 1984, *J. Am. Chem. Soc.*, 106:6077; Stein *et al.*, 1988, *Nucl. Acids Res.*, 16:3209; Zon *et al.*, 1991, *Anti-Cancer Drug Design*, 6:539; Zon *et al.*, 1991, OLIGONUCLEOTIDES AND ANALOGUES: A PRACTICAL APPROACH, pp. 87-108 (F. Eckstein, Ed.), Oxford University Press, Oxford England; Stec *et al.*, U.S. Pat. No. 5,151,510; Uhlmann and Peyman, 1990, *Chemical Reviews*, 90:543, the disclosures of which are hereby incorporated by reference for any purpose. An oligonucleotide can include a detectable label to enable detection of the oligonucleotide or hybridization thereof.

The term "isolated protein" referred to herein means that a subject protein (1) is free of at least some other proteins with which it would be found in nature, (2) is essentially free of other proteins from the same source, *e.g.*, from the same species, (3) is expressed by a cell from a different species, (4) has been separated from at least

about 50 percent of polynucleotides, lipids, carbohydrates, or other materials with which it is associated in nature, (5) is not associated (by covalent or noncovalent interaction) with portions of a protein with which the "isolated protein" is associated in nature, (6) is operably associated (by covalent or noncovalent interaction) with a polypeptide with which it is not associated in nature, or (7) does not occur in nature. Such an isolated protein can be encoded by genomic DNA, cDNA, mRNA or other RNA, of synthetic origin, or any combination thereof. In one embodiment, the isolated protein is substantially free from proteins or polypeptides or other contaminants that are found in its natural environment that would interfere with its use (therapeutic, diagnostic, prophylactic, research or otherwise).

An "isolated" antibody is one that has been identified and separated and/or recovered from a component of its natural environment. Contaminant components of its natural environment are materials that would interfere with diagnostic or therapeutic uses for the antibody, and may include enzymes, hormones, and other proteinaceous or non-proteinaceous substances. In certain embodiments, the antibody is purified (1) to greater than 95% or greater than 99% by weight of antibody as determined by the Lowry method, (2) to a degree sufficient to obtain at least 15 residues of N-terminal or internal amino acid sequence by use of a spinning cup sequenator, or (3) to homogeneity by SDS-PAGE under reducing or nonreducing conditions using Coomassie blue or silver stain. Isolated antibody includes the antibody *in situ* within recombinant cells since at least one component of the antibody's natural environment will not be present.

The terms "polypeptide" or "protein" means molecules having the sequence of native proteins, that is, proteins produced by naturally-occurring and specifically non-recombinant cells, or genetically-engineered or recombinant cells, and comprise molecules having the amino acid sequence of the native protein, or molecules having deletions from, additions to, and/or substitutions of one or more amino acids of the native sequence. The terms "polypeptide" and "protein" specifically encompass anti-B7RP1 antibodies, or sequences that have deletions from, additions to, and/or substitutions of one or more amino acid of an anti- B7RP1 antibody.

The term "polypeptide fragment" refers to a polypeptide that has an amino-terminal deletion, a carboxyl-terminal deletion, and/or an internal deletion. In certain embodiments, fragments are at least 5 to about 500 amino acids long. It will be

appreciated that in certain embodiments, fragments are at least 5, 6, 8, 10, 14, 20, 50, 70, 100, 110, 150, 200, 250, 300, 350, 400, or 450 amino acids long. Particularly useful polypeptide fragments include functional domains, including binding domains particularly antigen-binding domains, especially wherein the antigen is an epitope of human B7RP1. In the case of an anti-B7RP1 antibody, useful fragments include but are not limited to a CDR region, a variable domain of a heavy or light chain, a portion of an antibody chain or just its variable region including two CDRs, and the like.

The term "specific binding agent" refers to a naturally occurring or non-naturally occurring molecule that specifically binds to a target. Examples of specific binding agents include, but are not limited to, proteins, peptides, nucleic acids, carbohydrates, and lipids. In certain embodiments, a specific binding agent is an antibody.

The term "specific binding agent to B7RP1" refers to a specific binding agent that specifically binds any portion of B7RP1. In certain embodiments, a specific binding agent to B7RP1 is an antibody that binds specifically to B7RP1.

By way of example, an antibody "binds specifically" to a target if the antibody, when labeled, can be competed away from its target by the corresponding non-labeled antibody.

The term "immunologically functional immunoglobulin fragment" as used herein refers to a polypeptide fragment that contains at least the CDRs of the immunoglobulin heavy and light chains. An immunologically functional immunoglobulin fragment of the invention is capable of binding to an antigen. In certain embodiments, the antigen is a ligand that specifically binds to a receptor. In these embodiments, binding of an immunologically functional immunoglobulin fragment of the invention prevents binding of the ligand to its receptor, interrupting the biological response resulting from ligand binding to the receptor. In one embodiment, an immunologically functional immunoglobulin fragment of the invention binds specifically to B7RP1. Preferably, the fragment binds specifically to human B7RP1.

The term "naturally-occurring" or "native" as used herein and applied to an object refers to the fact that the object can be found in nature. For example, a polypeptide or polynucleotide sequence that is present in an organism (including

viruses) that can be isolated from a source in nature and that has not been intentionally modified by man is naturally-occurring. The term "non-naturally occurring" or "non-native" as used herein refers to a material that is not found in nature or that has been structurally modified or synthesized by man. For example, "non-naturally occurring" can refer to a variant, such as a polynucleotide variant that can be produced using art-known mutagenesis techniques, or a polypeptide variant produced by such a polynucleotide variant. Such variants include, for example, those produced by nucleotide substitutions, deletions or additions that may involve one or more nucleotides. Polynucleotide variants can be altered in coding or non-coding regions or both. Alterations in the coding regions may produce conservative or non-conservative amino acid substitutions, deletions, or additions. Especially certain among these are silent substitutions, additions, deletions, and conservative substitutions, which do not alter the properties and activities of a B7RP1 antibody of the invention. One of skill in the art can readily determine how to generate such a variant using methods well known in the art.

The term "operably linked" means that the components to which the term is applied are in a relationship that allows them to carry out their inherent functions under suitable conditions. For example, a control sequence "operably linked" to a protein coding sequence is ligated thereto so that expression of the protein coding sequence is achieved under conditions compatible with the transcriptional activity of the control sequences.

The term "control sequence" as used herein refers to polynucleotide sequences that can effect expression, processing or intracellular localization of coding sequences to which they are operably linked. The nature of such control sequences may depend upon the host organism. In particular embodiments, control sequences for prokaryotes may include a promoter, ribosomal binding site, and transcription termination sequence. In other particular embodiments, control sequences for eukaryotes may include promoters comprising one or a plurality of recognition sites for transcription factors, transcription enhancer sequences, transcription termination sequences and polyadenylation sequences. In certain embodiments, "control sequences" can include leader sequences and/or fusion partner sequences.

The term "vector" includes a nucleic acid molecule capable of carrying into a cell another nucleic acid to which it has been linked. One type of vector is a

"plasmid", which refers to a circular double stranded DNA loop into which additional DNA segments may be ligated. Another type of vector is a viral vector, wherein additional DNA segments may be ligated into the viral genome. Certain vectors are capable of autonomous replication in a host cell into which they are introduced (*e.g.*, bacterial vectors having a bacterial origin of replication and episomal mammalian vectors). Other vectors (*e.g.*, non-episomal mammalian vectors) can be integrated into the genome of a host cell upon introduction into the host cell and thereby are replicated along with the host genome. Moreover, certain vectors are capable of directing the expression of genes to which they are operatively linked. Such vectors are referred to herein as "recombinant expression vectors" (or simply, "expression vectors"). In general, expression vectors useful in the practice of recombinant DNA techniques are often in the form of plasmids. In the present specification, "plasmid" and "vector" may be used interchangeably as the plasmid is the most commonly used form of vector. However, the invention is intended to include such other forms of expression vectors, such as viral vectors (*e.g.*, replication defective retroviruses, adenoviruses and adeno-associated viruses), which serve equivalent functions.

The phrase "recombinant host cell" (or simply "host cell") includes a cell into which a recombinant expression vector has been introduced. It will be understood by those of skill in the art that such terms are intended to refer not only to the particular subject cell but to the progeny of such a cell. Because certain modifications may occur in succeeding generations due to either mutation or environmental influences, such progeny may not, in fact, be identical to the parent cell, but are still included within the scope of the term "host cell" as used herein. A wide variety of host expression systems can be used to express the antibodies of the present invention including bacterial, yeast, baculoviral and mammalian expression systems (as well as phage display expression systems). An example of a suitable bacterial expression vector is pUC19. To express an antibody recombinantly, a host cell is transfected with one or more recombinant expression vectors carrying DNA fragments encoding the immunoglobulin light and heavy chains of the antibody such that the light and heavy chains are expressed in the host cell and can be secreted into the medium in which the host cells are cultured, from which medium the antibodies can be recovered. Standard recombinant DNA methodologies are used to obtain antibody heavy and light chain genes, incorporate these genes into recombinant expression

vectors and introduce the vectors into host cells, such as those described in Sambrook *et al.*, 2001, MOLECULAR CLONING, A LABORATORY MANUAL, Cold Spring Harbor Laboratories, Ausubel, F.M. et al. (eds.), CURRENT PROTOCOLS IN MOLECULAR BIOLOGY, Greene Publishing Associates, (1989) and in U.S. Patent No. 4,816,397 to Boss et al.

The term "transduction" is used to refer to the transfer of genes from one bacterium to another, usually by a phage. "Transduction" also refers to the acquisition and transfer of eukaryotic cellular sequences by retroviruses.

The term "transfection" is used to refer to the uptake of foreign or exogenous DNA by a cell, and a cell has been "transfected" when the exogenous DNA has been introduced inside the cell membrane. A number of transfection techniques are well known in the art and are disclosed herein. *See, e.g.*, Graham *et al.*, 1973, *Virology* 52: 456; Sambrook *et al.*, 2001, MOLECULAR CLONING, A LABORATORY MANUAL, Cold Spring Harbor Laboratories; Davis *et al.*, 1986, BASIC METHODS IN MOLECULAR BIOLOGY, Elsevier; and Chu *et al.*, 1981, *Gene* 13: 197. Such techniques can be used to introduce one or more exogenous DNA moieties into suitable host cells.

The term "transformation" as used herein refers to a change in a cell's genetic characteristics, and a cell has been transformed when it has been modified to contain a new DNA. For example, a cell is transformed where it is genetically modified from its native state. Following transfection or transduction, the transforming DNA may recombine with DNA from the cell by physically integrating into a chromosome of the cell, or may be maintained transiently as an episomal element without being replicated, or may replicate independently as a plasmid. A cell is considered to have been stably transformed when the DNA is replicated with the division of the cell.

The term "antigen" refers to a molecule or a portion of a molecule capable of being bound by a selective binding agent, such as an antibody, and additionally capable of being used in an animal to produce antibodies capable of binding to an epitope of that antigen. An antigen may have one or more epitopes.

In certain embodiments, antibody variants include glycosylation variants wherein the number and/or type of glycosylation site has been altered compared to the amino acid sequences of the parent polypeptide. In certain embodiments, protein variants comprise a greater or a lesser number of N-linked glycosylation sites than the

native protein. An N-linked glycosylation site is characterized by the sequence: Asn-Xaa-Ser or Asn-Xaa-Thr, wherein the amino acid residue designated as Xaa may be any amino acid residue except proline. The substitution of amino acid residues to create this sequence provides a potential new site for the addition of an N-linked carbohydrate chain. Alternatively, substitutions that eliminate this sequence will remove an existing N-linked carbohydrate chain. Also provided is a rearrangement of N-linked carbohydrate chains wherein one or more N-linked glycosylation sites (typically those that are naturally occurring) are eliminated and one or more new N-linked sites are created. Additional antibody variants include cysteine variants wherein one or more cysteine residues are deleted from or substituted for another amino acid (*e.g.*, serine) compared to the parent amino acid sequence. Cysteine variants may be useful when antibodies must be refolded into a biologically active conformation such as after the isolation of insoluble inclusion bodies. Cysteine variants generally have fewer cysteine residues than the native protein, and typically have an even number to minimize interactions resulting from unpaired cysteines.

In additional embodiments, antibody variants can include antibodies comprising a modified Fc fragment or a modified heavy chain constant region. An Fc fragment, which stands for "fragment that crystallizes," or a heavy chain constant region can be modified by mutation to confer on an antibody altered binding characteristics. *See, for example*, Burton and Woof, 1992, *Advances in Immunology* 51: 1-84; Ravetch and Bolland, 2001, *Annu. Rev. Immunol.* 19: 275-90; Shields *et al.*, 2001, *Journal of Biol. Chem* 276: 6591-6604; Telleman and Junghans, 2000, *Immunology* 100: 245-251; Medesan *et al.*, 1998, *Eur. J. Immunol.* 28: 2092-2100; all of which are incorporated herein by reference). Such mutations can include substitutions, additions, deletions, or any combination thereof, and are typically produced by site-directed mutagenesis using one or more mutagenic oligonucleotide(s) according to methods described herein, as well as according to methods known in the art (see, *for example*, Sambrook *et al.*, MOLECULAR CLONING: A LABORATORY MANUAL, 3rd Ed., 2001, Cold Spring Harbor, N.Y.and Berger and Kimmel, METHODS IN ENZYMOLOGY, Volume 152, Guide to Molecular Cloning Techniques, 1987, Academic Press, Inc., San Diego, CA., which are incorporated herein by reference).

According to certain embodiments, amino acid substitutions may (1) reduce susceptibility to proteolysis, (2) reduce susceptibility to oxidation, (3) alter binding affinity, and/or (4) confer or modify other physicochemical or functional properties on such polypeptides. According to certain embodiments, single or multiple amino acid substitutions (in certain embodiments, conservative amino acid substitutions) may be made in the naturally occurring sequence (in certain embodiments, in the portion of the polypeptide outside the domain(s) forming intermolecular contacts). In certain embodiments, a conservative amino acid substitution typically does not substantially change the structural characteristics of the parent sequence (*e.g.*, a replacement amino acid should disrupt or tend to disrupt secondary structure that characterizes a parent sequence, such as a helix). Examples of art-recognized polypeptide secondary and tertiary structures are described in PROTEINS, STRUCTURES AND MOLECULAR PRINCIPLES, (Creighton, Ed.), 1984, W. H. Freeman and Company, New York; INTRODUCTION TO PROTEIN STRUCTURE (C. Branden and J. Tooze, eds.), 1991, Garland Publishing, New York, N.Y.; and Thornton *et al.*, 1991, *Nature* 354:105, each of which are incorporated herein by reference.

"Antibody" or "antibody peptide(s)" refer to an intact antibody, or a binding fragment thereof that competes with the intact antibody for specific binding. In certain embodiments, binding fragments are produced by recombinant DNA techniques. In additional embodiments, binding fragments are produced by enzymatic or chemical cleavage of intact antibodies. Binding fragments include, but are not limited to, F(ab), F(ab'), $F(ab')_2$, Fv, and single-chain antibodies.

The invention provides antibodies that comprise a heavy chain and a light chain, wherein the heavy and light chains together form an antigen binding structure capable of specifically binding B7RP1. A full-length heavy chain includes a variable region domain, V_H, and three constant region domains, C_H1, C_H2, and C_H3. The V_H domain is at the amino-terminus of the polypeptide, and the C_H3 domain is at the carboxyl-terminus. The term "heavy chain", as used herein, encompasses a full-length heavy chain and fragments thereof. A full-length light chain includes a variable region domain, V_L, and a constant region domain, C_L. Like the heavy chain, the variable region domain of the light chain is at the amino-terminus of the polypeptide. The term "light chain", as used herein, encompasses a full-length light chain and fragments thereof. A F(ab) fragment is comprised of one light chain and

the C_H1 and variable regions of one heavy chain. The heavy chain of a F(ab) molecule cannot form a disulfide bond with another heavy chain molecule. A F(ab') fragment contains one light chain and one heavy chain that contains more of the constant region, between the C_H1 and C_H2 domains, such that an interchain disulfide bond can be formed between two heavy chains to form a $F(ab')_2$ molecule. The Fv region comprises the variable regions from both the heavy and light chains, but lacks the constant regions. Single-chain antibodies are Fv molecules in which the heavy and light chain variable regions have been connected by a flexible linker to form a single polypeptide chain, which forms an antigen-binding region. Single chain antibodies are discussed in detail in International Patent Application Publication No. WO 88/01649 and U.S. Patent Nos. 4,946,778 and 5,260,203.

A bivalent antibody other than a "multispecific" or "multifunctional" antibody, in certain embodiments, is understood to comprise binding sites having identical antigenic specificity.

In assessing antibody binding and specificity according to the invention, an antibody substantially inhibits adhesion of a ligand to a receptor when an excess of antibody reduces the quantity of ligand bound to receptor by at least about 20%, 40%, 60%, 80%, 85%, or more (as measured, *inter alia*, using an *in vitro* competitive binding assay).

By "neutralizing antibody" is meant an antibody molecule that is able to block or substantially reduce an effector function of a target antigen to which it binds. Accordingly, a "neutralizing" anti-B7RP1 antibody is capable of blocking or substantially reducing an effector function, such as receptor binding and/or elicitation of a cellular response, of B7RP1. "Substantially reduce" is intended to mean at least about 60%, at least about 70%, at least about 75%, at least about 80%, at least about 85%, or at least about 90% reduction of an effector function of the target antigen (e.g., human B7RP1).

The term "epitope" includes any site on an antigen that is capable of specific binding to an immunoglobulin or T-cell receptor. In certain embodiments, epitope determinants include chemically active surface groupings of molecules such as amino acids, sugar side chains, phosphoryl groups, or sulfonyl groups, and, in certain embodiments, may have specific three-dimensional structural characteristics, and/or

specific charge characteristics. An epitope is a region of an antigen that is bound by an antibody. In certain embodiments, an antibody is said to specifically bind an antigen when it preferentially recognizes its target antigen in a complex mixture of proteins and/or macromolecules. In certain embodiments, an antibody is said to specifically bind an antigen when the equilibrium dissociation constant is about 10^{-6} M, 10^{-7} M, 10^{-8} M, 10^{-9} M, 10^{-10} M, 10^{-11} M, 10^{-12} M, or less than about 10^{-12}M.

An antibody binds "essentially the same epitope" as a reference antibody, when the two antibodies recognize identical or sterically overlapping epitopes. The most widely used and rapid methods for determining whether two antibodies bind to identical or sterically overlapping epitopes are competition assays, which can be configured in all number of different formats, using either labeled antigen or labeled antibody. Usually, the antigen is immobilized on a substrate, and the ability of unlabeled antibodies to block the binding of labeled antibodies is measured using radioactive isotopes or enzyme labels.

The term "agent" is used herein to denote a chemical compound, a mixture of chemical compounds, a biological macromolecule, or an extract made from biological materials.

As used herein, the terms "label" or "labeled" refers to incorporation of a detectable marker, *e.g.*, by incorporation of a radiolabeled amino acid or attachment to a polypeptide of biotin moieties that can be detected by labeled avidin (*e.g.*, streptavidin comprising a detectable marker such as a fluorescent marker, a chemiluminescent marker or an enzymatic activity that can be detected by optical or colorimetric methods). In certain embodiments, the label can also be therapeutic. Various methods of labeling polypeptides and glycoproteins are known in the art and may be used advantageously in the methods disclosed herein. Examples of labels for polypeptides include, but are not limited to radioisotopes or radionuclides such as ^{3}H, ^{14}C, ^{15}N, ^{35}S, ^{90}Y, ^{99m}Tc, ^{111}In, ^{125}I, and ^{131}I, fluorescent labels (*e.g.*, fluorescein isothiocyanate or FITC, rhodamine, or lanthanide phosphors), enzymatic labels (*e.g.*, horseradish peroxidase, β-galactosidase, luciferase, alkaline phosphatase), chemiluminescent labels, hapten labels such as biotinyl groups, and predetermined polypeptide epitopes recognized by a secondary reporter (*e.g.*, leucine zipper pair sequences, binding sites for secondary antibodies, metal binding domains, or epitope

tags). In certain embodiments, labels are attached by spacer arms (such as $(CH_2)_n$, where n < about 20) of various lengths to reduce potential steric hindrance.

The term "biological sample", as used herein, includes, but is not limited to, any quantity of a substance from a living thing or formerly living thing. Such living things include, but are not limited to, humans, mice, monkeys, rats, rabbits, and other animals. Such substances include, but are not limited to, blood, serum, urine, cells, organs, tissues, bone, bone marrow, lymph nodes, and skin.

The term "pharmaceutical agent or drug" as used herein refers to a chemical compound or composition capable of inducing a desired therapeutic effect when properly administered to a patient. The expression "pharmaceutically effective amount" in reference to a pharmaceutical composition comprising one or a plurality of the antibodies of the invention is understood to mean, according to the invention, an amount of the said pharmaceutical composition that is capable of abolishing, in a patient, the decrease in the sensitivity threshold to external stimuli with a return of this sensitivity threshold to a level comparable to that observed in healthy subjects.

A "disorder" is any condition that would benefit from treatment according to the present invention. "Disorder" and "condition" are used interchangeably herein and include chronic and acute immune system disorders or immune system diseases associated with inappropriate immune response, including those pathological conditions which predispose the mammal to the disorder in question. A number of conditions and disorders that would benefit from the treatment according to the present invention are described, for example, in International Patent Application No. PCT/US00/01871 (Publication No. WO 00/46240), the disclosure of which is incorporated by reference in its entirety.

The terms "immune system disease" and "immune system condition" encompass any medical condition or disorder associated with increased levels of B7RP1, increased sensitivity to B7RP1, or T-cell mediated diseases, including, but not limited to, autoimmune disease, graft survival, bone marrow and organ transplantation, allosensitization due to blood transfusions, toxic shock syndrome, T-cell dependent B-cell mediated diseases, chronic inflammatory diseases associated with chronic immune cell dysfunction, lymphoproliferative disorders (such as multiple myeloma, Waldenstom's macroglobulinemia, and crioglobulinemias), and

cancer. Non-limiting examples of autoimmune diseases include systemic lupus erythematosis, rheumatoid arthritis, immune thrombocytopenic purpura (ITP), multiple sclerosis, diabetes, and psoriasis. Non-limiting examples of chronic inflammatory diseases include inflammatory bowel disease (such as Crohn's disease and ulcerative colitis), Grave's disease, Hashimoto's thyroiditis, and diabetes mellitus.

The terms "immune system disease" and "immune system condition" also encompass any clinical condition that would be ameliorated by the inhibition of antibody production, such as hypersensitivity reactions. Hypersensitivity reactions can be caused, for example, by hay fever, allergies, asthma, atopy, and acute edema. Non-limiting examples of diseases that cause antibody-mediated hypersensitivity reactions include systemic lupus erythematosis, arthritis (such as rheumatoid arthritis, reactive arthritis, psoriatic arthritis), nephropathies (such as glomerulo-nephritis, membranous, mesangiocapillary, focal segmental, focal necrotizing, crescentic, and proliferative nephropathies such as tubulopathies), skin disorders (such as pemphigus and pemphigoid, erythema nodosum), endocrinopathies (such as thyroiditis, Grave's disease, Hashimoto's disease, insulin dependent diabetes mellitus), various pneumopathies (such as extrinsic alveolitis), various vasculopathies, coeliac disease, diseases with aberrant production of IgA, many anemias and thrombocytopenias, Guillain-Barre Syndrome, and myasthenia gravis.

As used herein, the terms "effective amount" and "therapeutically effective amount" when used with reference to a vehicle- or a pharmaceutical composition comprising one or more anti-human B7RP1 human antibodies refers to an amount or dosage sufficient to produce a desired result (i.e., where for therapy with the vehicle- or anti-human B7RP1 human antibodies of the present invention the desired result is the desired modulation of T-cell responses, for example) or to support an observable decrease in the level of one or more biological activities of B7RP1. More specifically, a therapeutically effective amount is an amount of the anti-human B7RP1 human antibody(ies) sufficient to inhibit, for some period of time, one or more of the clinically defined pathological processes associated with the condition at issue, e.g. immune disorders and diseases, in a subject treated *in vivo* with the agent. In the present invention, an "effective amount" of an anti-B7RP1 antibody may modulate T-cell responses in a patient. In the methods of the present invention, the term "control"

and grammatical variants thereof, are used to refer to the prevention, partial or complete inhibition, reduction, delay or slowing down of an unwanted event, e.g. immune response. The effective amount may vary depending on the specific vehicle- or anti-human B7RP1 human antibody(ies) selected, and is also dependent on a variety of factors and conditions related to the subject to be treated and the severity of the disorder. For example, if the vehicle- or anti-human B7RP1 human antibody(ies) is to be administered *in vivo*, factors such as the age, weight and health of the patient as well as dose response curves and toxicity data obtained in preclinical animal work would be among those considered. If the agent is to be contacted with the cells *in vitro*, one would also design a variety of pre-clinical *in vitro* studies to assess such parameters as uptake, half-life, dose, toxicity, etc. The determination of an effective amount or a therapeutically effective amount for a given agent is well within the ability of those skilled in the art.

As used herein, the terms "B7 related protein-1" and "B7RP1" are defined as all mammalian species of native sequence B7RP1, which is described in International Patent Application Publication No. WO 00/46240, which is incorporated herein by reference.

As used herein, "substantially pure" or "substantially purified" means a compound or species that is the predominant species present (*i.e.*, on a molar basis it is more abundant than any other individual species in the composition). In certain embodiments, a substantially purified fraction is a composition wherein the species comprises at least about 50 percent (on a molar basis) of all macromolecular species present. In certain embodiments, a substantially pure composition will comprise more than about 80%, 85%, 90%, 95%, or 99% of all macromolar species present in the composition. In certain embodiments, the species is purified to essential homogeneity (contaminant species cannot be detected in the composition by conventional detection methods) wherein the composition consists essentially of a single macromolecular species.

The term "patient" includes human and animal subjects.

"Treatment" or "treat" refers to both therapeutic treatment and prophylactic or preventative measures. Those in need of treatment include those already with the

disorder as well as those prone to have the disorder or those in which the disorder is to be prevented.

Unless otherwise required by context, singular terms shall include pluralities and plural terms shall include the singular.

According to certain embodiments of the invention, antibodies directed to B7RP1 may be used to treat immune system disorders and immune system diseases, including but not limited to, those mentioned above.

In one aspect of the invention are provided fully human monoclonal antibodies raised against and having biological and immunological specificity for binding to human B7RP1. In another aspect the invention provides nucleic acids comprising nucleotide sequences encoding amino acid sequences for heavy and light chain immunoglobulin molecules, particularly sequences corresponding to the variable regions thereof. Particular embodiments of this aspect of the invention are sequences corresponding to complementarity determining regions (CDRs), specifically from CDR1 through CDR3, of the heavy and light chains provided by the invention. In yet another aspect the invention provides hybridoma cells and cell lines that express the immunoglobulin molecules and antibodies, such as monoclonal antibodies of the invention. The invention also provides biologically and immunologically purified preparations of antibodies, such as monoclonal antibodies raised against and having biological and immunological specificity for binding to human B7RP1.

The ability to clone and reconstruct megabase-sized human loci in yeast artificial chromosomes (YACs) and to introduce them into the mouse germline provides an advantageous approach to elucidating the functional components of very large or crudely mapped loci as well as generating useful models of human disease. Furthermore, the utilization of such technology for substitution of mouse loci with their human equivalents provides unique insights into the expression and regulation of human gene products during development, their communication with other systems, and their involvement in disease induction and progression.

An important practical application of such a strategy is the "humanization" of the mouse humoral immune system. Introduction of human immunoglobulin (Ig) loci into mice in which the endogenous Ig genes have been inactivated offers the opportunity to study mechanisms underlying programmed expression and assembly of

antibodies as well as their role in B-cell development. Furthermore, such a strategy provides a source for production of fully human monoclonal antibodies (MAbs).

The term "human antibody" includes antibodies having variable and constant regions substantially corresponding to human germline immunoglobulin sequences. In certain embodiments, human antibodies are produced in non-human mammals, including, but not limited to, rodents, such as mice and rats, and lagomorphs, such as rabbits. In certain embodiments, human antibodies are produced in hybridoma cells. In certain embodiments, human antibodies are produced recombinantly.

The term "recombinant" in reference to an antibody includes antibodies that are prepared, expressed, created or isolated by recombinant means. Representative examples include antibodies expressed using a recombinant expression vector transfected into a host cell, antibodies isolated from a recombinant, combinatorial human antibody library, antibodies isolated from an animal (*e.g.*, a mouse) that is transgenic for human immunoglobulin genes (*see e.g.*, Taylor, *et al.*, 1992, *Nucl. Acids Res.* 20:6287-6295); or antibodies prepared, expressed, created or isolated by any means that involves splicing of human immunoglobulin gene sequences to other DNA sequences. Such recombinant human antibodies have variable and constant regions derived from human germline immunoglobulin sequences.

Human antibodies have at least three advantages over non-human and chimeric antibodies for use in human therapy:

1) because the effector portion of the antibody is human, it may interact better with the other parts of the human immune system (e.g., destroy the target cells more efficiently by complement-dependent cytotoxicity (CDC) or antibody-dependent cellular cytotoxicity (ADCC));

2) the human immune system should not recognize the human antibody as foreign, and, therefore the antibody response against such an injected antibody should be less than against a totally foreign non-human antibody or a partially foreign chimeric antibody;

3) injected non-human antibodies have been reported to have a half-life in the human circulation much shorter than the half-life of human antibodies. Injected human antibodies will have a half-life essentially identical to naturally occurring human antibodies, allowing smaller and less frequent doses to be given.

Thus, fully human antibodies are expected to minimize the immunogenic and allergic responses intrinsic to mouse or mouse-derivatized MAbs, and to thereby increase the efficacy and safety of the administered antibodies. Fully human antibodies of the invention, therefore, can be used in the treatment of diseases and disorders associated with inappropriate immune response, the treatment thereof requiring repeated antibody administration. Thus, one particular advantage of the anti-B7RP1 antibodies of the invention is that the antibodies are fully human and can be administered to patients in a non-acute manner while minimizing adverse reactions commonly associated with human anti-mouse antibodies or other previously described non-fully human antibodies from non-human species.

One skilled in the art can engineer mouse strains deficient in mouse antibody production with large fragments of the human Ig loci so that such mice produce human antibodies in the absence of mouse antibodies. Large human Ig fragments may preserve the large variable gene diversity as well as the proper regulation of antibody production and expression. By exploiting the mouse cellular machinery for antibody diversification and selection and the lack of immunological tolerance to human proteins, the reproduced human antibody repertoire in these mouse strains yields high affinity antibodies against any antigen of interest, including human antigens. Using the hybridoma technology, antigen-specific human MAbs with the desired specificity may be produced and selected.

Transgenic animals (e.g., mice) can also be used to produce human antibodies in the absence of endogenous immunoglobulin production. For example, transfer of the human germ-line immunoglobulin gene array in such germ-line mutant mice will result in the production of human antibodies upon antigen challenge (see, *e.g.*, Jakobovits *et al.*, 1993, *Proc. Natl. Acad. Sci. USA* 90:2551-2555; Jakobovits *et al.*, 1993, *Nature* 362:255-258; Bruggemann *et al.*, 1993, *Year in Immun.* 7:33, 1994, *Nature* 148:1547-1553) and, 1996, *Nature Biotechnology* 14:826; Gross *et al.*, 2000, *Nature* 404:995-999; and U.S. Patents Nos. 5,877,397, 5,874,299, 5,814,318, 5,789,650, 5,770,429, 5,661,016, 5,633,425, 5,625,126, 5,569,825, and 5,545,806, (each of which is incorporated herein by reference in its entirety for all purposes)). Human antibodies can also be produced in phage display libraries (Hoogenboom and Winter, 1992, *J. Mol. Biol* 227:381; Marks *et al.*, 1991, *J. Mol. Biol.* 222:581). The techniques of Cole *et al.* and Boerner *et al.* are also available for the preparation of

human monoclonal antibodies (Cole *et al.*, 1985, MONOCLONAL ANTIBODIES AND CANCER THERAPY Alan R. Liss, p. 77; and Boerner *et al.*, 1991, *J. Immunol.* 147:86-95).

Recombinant human antibodies may also be subjected to *in vitro* mutagenesis (or, when an animal transgenic for human Ig sequences is used, *in vivo* somatic mutagenesis) and, thus, the amino acid sequences of the V_H and V_L regions of the recombinant antibodies are sequences that, while derived from those related to human germline V_H and V_L sequences, may not naturally exist within the human antibody germline repertoire *in vivo*.

In certain embodiments, the skilled artisan can use constant regions from species other than human along with the human variable region(s) in such mice to produce chimeric antibodies.

A bispecific or bifunctional antibody typically is an artificial hybrid antibody having two different heavy chain/light chain pairs and two different binding sites. Bispecific antibodies may be produced by a variety of methods including, but not limited to, fusion of hybridomas or linking of F(ab') fragments. *See, e.g.*, Songsivilai & Lachmann, 1990, *Clin. Exp Immunol.* 79: 315-321; Kostelny *et al.*, 1992, *J. Immunol.* 148:1547-1553.

The invention provides antibodies that bind to human B7RP1. These antibodies can be produced by immunization with full-length B7RP1 or fragments thereof. The antibodies of the invention can be polyclonal or monoclonal, and/or may be recombinant antibodies. In preferred embodiments, antibodies of the invention are human antibodies prepared, for example, by immunization of transgenic animals capable of producing human antibodies (see, for example, International Patent Application, Publication W0 93/12227).

The complementarity determining regions (CDRs) of the light chain and heavy chain variable regions of anti- B7RP1 antibodies of the invention can be grafted to framework regions (FRs) from the same, or another, species. In certain embodiments, the CDRs of the light chain and heavy chain variable regions of anti-B7RP1 antibody may be grafted to consensus human FRs. To create consensus human FRs, FRs from several human heavy chain or light chain amino acid sequences are aligned to identify a consensus amino acid sequence. The FRs of the anti-B7RP1 antibody heavy chain

or light chain can be replaced with the FRs from a different heavy chain or light chain. Rare amino acids in the FRs of the heavy and light chains of anti-B7RP1 antibody typically are not replaced, while the rest of the FR amino acids can be replaced. Rare amino acids are specific amino acids that are in positions in which they are not usually found in FRs. The grafted variable regions from anti-B7RP1 antibodies of the invention can be used with a constant region that is different from the constant region of anti-B7RP1 antibody. Alternatively, the grafted variable regions are part of a single chain Fv antibody. CDR grafting is described, *e.g.*, in U.S. Patent Nos. 6,180,370, 5,693,762, 5,693,761, 5,585,089, and 5,530,101, which are hereby incorporated by reference for any purpose.

Antibodies of the invention can be prepared using transgenic mice that have a substantial portion of the human antibody producing locus inserted in antibody-producing cells of the mice, and that are further engineered to be deficient in producing endogenous, murine, antibodies. Such mice are capable of producing human immunoglobulin molecules and antibodies and do not produce or produce substantially reduced amounts of murine immunoglobulin molecules and antibodies. Technologies utilized for achieving this result are disclosed in the patents, applications, and references disclosed in the specification herein. In certain embodiments, the skilled worker may employ methods as disclosed in International Patent Application Publication No. WO 98/24893, which is hereby incorporated by reference for any purpose. *See also* Mendez *et al.*, 1997, *Nature Genetics* 15:146-156, which is hereby incorporated by reference for any purpose.

The monoclonal antibodies (mAbs) of the invention can be produced by a variety of techniques, including conventional monoclonal antibody methodology, *e.g.*, the standard somatic cell hybridization technique of Kohler and Milstein (1975, *Nature* 256:495). Other techniques for producing monoclonal antibodies may be employed, *e.g.*, viral or oncogenic transformation of B-lymphocytes.

An exemplary animal system for preparing hybridomas is the mouse. Hybridoma production in the mouse is known in the art and immunization protocols and techniques for isolation of immunized splenocytes for fusion are also known in the art. Fusion partners (*e.g.*, murine myeloma cells) and fusion procedures are also known.

In a certain embodiment, human monoclonal antibodies directed against B7RP1 can be generated using transgenic mice carrying parts of the human immune system rather than the mouse system. These transgenic mice, referred to herein as "HuMab" mice, contain a human immunoglobulin gene minilocus that encodes unrearranged human heavy (μ and γ) and κ light chain immunoglobulin sequences, together with targeted mutations that inactivate the endogenous μ and κ chain loci (Lonberg *et al.*, 1994, *Nature* 368:856-859). Accordingly, the mice exhibit reduced expression of mouse IgM or κ and in response to immunization, the introduced human heavy chain and light chain transgenes undergo class switching and somatic mutation to generate high affinity human IgG κ monoclonal antibodies (Lonberg *et al.*, *supra.*; Lonberg and Huszar, 1995, *Intern. Rev. Immunol.* 13:65-93; Harding and Lonberg, 1995, *Ann.N.Y.Acad.Sci.* 764:536-546). The preparation of HuMab mice is described in detail in Taylor *et al.*, 1992, *Nucleic Acids Res.* 20:6287-6295; Chen *et al.*, 1993, *International Immunology* 5:647-656; Tuaillon *et al.*, 1994, *J. Immunol.* 152:2912-2920; Lonberg *et al.*, 1994, *Nature* 368:856-859; Lonberg, 1994, *Handbook of Exp. Pharmacology* 113:49-101; Taylor *et al.*, 1994, *International Immunology* 6:579-591; Lonberg & Huszar, 1995, *Intern. Rev. Immunol.* 13:65-93; Harding & Lonberg, 1995, *Ann. N.Y. Acad. Sci* 764:536-546; Fishwild *et al.*, 1996, *Nature Biotechnology* 14:845-851, the contents of all of which are hereby incorporated by reference in their entirety. *See further* U.S. Patent Nos. 5,545,806; 5,569,825; 5,625,126; 5,633,425; 5,789,650; 5,877,397; 5,661,016; 5,814,318; 5,874,299; and 5,770,429; all to Lonberg and Kay, as well as U.S. Patent No. 5,545,807 to Surani *et al.*; International Patent Application Publication Nos. WO 93/1227, published June 24, 1993; WO 92/22646, published December 23, 1992; and WO 92/03918, published March 19, 1992, the disclosures of all of which are hereby incorporated by reference in their entirety. Alternatively, transgenic mice strains described in the Examples below can be used to generate human anti-B7RP1 antibodies.

The present invention provides human monoclonal antibodies that are specific for and neutralize bioactive human B7RP1 polypeptides. Also provided are antibody heavy and light chain amino acid sequences which are highly specific for and neutralize B7RP1 polypeptides when they are bound to them. This high specificity enables the anti-human B7RP1 human antibodies, and human monoclonal antibodies with like specificity, to be effective immunotherapy for B7RP1 associated diseases.

In one aspect, the invention provides isolated human antibodies that bind the same or essentially the same epitope as the 16H antibody provided herein.

In one aspect, the invention provides isolated human antibodies comprising at least one of the amino acid sequences shown in SEQ ID NOS: 1-40 or 44-58 that binds a B7RP1 polypeptide epitope with high affinity and has the capacity to antagonize B7RP1 polypeptide activity. These antibodies may bind the same or essentially the same epitope as the anti-B7RP1 antibodies shown in the Examples herein.

In certain embodiments, the isolated antibodies bind to B7RP1 polypeptide with a dissociation constant (K_D) of about 10^{-6} M, 10^{-7} M, 10^{-8} M, 10^{-9} M, 10^{-10} M, 10^{-11} M or less and inhibits B7RP1 induced survival in an *in vitro* neutralization assay with an EC_{50} of about 10^{-6} M, 10^{-7} M, 10^{-8} M, 10^{-9} M or less. Examples of anti-human B7RP1 human antibodies that meet the aforementioned binding and neutralization criteria are provided herein.

In certain embodiments, anti-human B7RP1 human antibodies of the invention are referred to herein as 16H, 16Hg (germline), 5D, 2H, 2Hg (germline), 15H, 41H, and 43H. Antibody 16H comprises V_L and V_H polypeptide sequences as shown in SEQ ID NO: 7 and SEQ ID NO: 1, respectively. Antibody 16Hg comprises a variable light chain (V_L) and variable heavy chain (V_H) polypeptide sequences as shown in SEQ ID NO: 1 and SEQ ID NO: 8, respectively. Antibody 5D comprises V_L and V_H polypeptide sequences as shown in SEQ ID NO: 2 and SEQ ID NO: 9, respectively. Antibody 2H comprises V_L and V_H polypeptide sequences as shown in SEQ ID NO: 3 and SEQ ID NO: 10, respectively. Antibody 2Hg comprises V_L and V_H polypeptide sequences as shown in SEQ ID NO: 3 and SEQ ID NO: 11, respectively. Antibody 15H comprises V_L and V_H polypeptide sequences as shown in SEQ ID NO: 4 and SEQ ID NO: 12, respectively. Antibody 41H comprises V_L and V_H polypeptide sequences as shown in SEQ ID NO: 5 and SEQ ID NO: 13, respectively. Antibody 43H comprises V_L and V_H polypeptide sequences as shown in SEQ ID NO: 6 and SEQ ID NO: 14, respectively. The properties of the anti-human B7RP1 human antibodies of the present invention are specifically disclosed in the Examples. Particularly notable is the high affinity for B7RP1 polypeptide and high capacity to antagonize B7RP1 polypeptide activity demonstrated herein.

The dissociation constant (K_D) of an anti-human B7RP1 human antibody can be determined by surface plasmon resonance as generally described in the Examples below. Generally, surface plasmon resonance analysis measures real-time binding interactions between ligand (recombinant B7RP1 polypeptide immobilized on a biosensor matrix) and analyte (antibodies in solution) by surface plasmon resonance (SPR) using the BIAcore® system (Pharmacia Biosensor, Piscataway, NJ). Surface plasmon analysis can also be performed by immobilizing the analyte (antibodies on a biosensor matrix) and presenting the ligand (recombinant V in solution). The dissociation constant (K_D) of an anti-human B7RP1 human antibody can also be determined by using KinExA methodology. In certain embodiments of the invention, the antibodies bind to B7RP1 with a K_D of approximately 10^{-5} M, 10^{-6} M, 10^{-7} M, 10^{-8} M, 10^{-9} M, 10^{-10} M, 10^{-11} M, or 10^{-12} M. The term "K_D", as used herein, is intended to refer to the dissociation constant of a particular antibody-antigen interaction. For purposes of the present invention K_D was determined as shown in the Examples below.

In certain embodiments, the antibodies of the invention are of the IgG1, IgG2, IgG3, or IgG4 isotype. The antibodies may be of the IgG2 or IgG1 isotype. In other embodiments, the antibodies of the invention may be of the IgM, IgA, IgE, or IgD isotype. In certain embodiments of the invention, the antibodies comprise a human kappa light chain and a human IgG1, IgG2, IgG3, or IgG4 heavy chain. Expression of antibodies of the invention comprising an IgG1 or an IgG2 heavy chain constant region is described in the Examples below. In particular embodiments, the variable regions of the antibodies are ligated to a constant region other than the constant region for the IgG1, IgG2, IgG3, or IgG4 isotype. In certain embodiments, the antibodies of the invention have been cloned for expression in mammalian cells.

In certain embodiments, conservative modifications to the heavy chains and light chains of anti-B7RP1 antibodies (and corresponding modifications to the encoding nucleotides) will produce anti-B7RP1 antibodies having functional and chemical characteristics similar to those of the anti-B7RP1 antibodies disclosed herein. In contrast, substantial modifications in the functional and/or chemical characteristics of anti-B7RP1 antibodies may be accomplished by selecting substitutions in the amino acid sequence of the heavy and light chains that differ significantly in their effect on maintaining (a) the structure of the molecular backbone

in the area of the substitution, for example, as a sheet or helical conformation, (b) the charge or hydrophobicity of the molecule at the target site, or (c) the bulk of the side chain.

For example, a "conservative amino acid substitution" may involve a substitution of a native amino acid residue with a nonnative residue such that there is little or no effect on the polarity or charge of the amino acid residue at that position. Furthermore, any native residue in the polypeptide may also be substituted with alanine, as has been previously described for "alanine scanning mutagenesis."

Amino acid substitutions (whether conservative or non-conservative) can be determined by those skilled in the art at the time such substitutions are desired. In certain embodiments, amino acid substitutions can be used to identify those amino acid residues of an anti-B7RP1 antibody that are involved in binding specificity and/or affinity of the antibody for B7RP1 (*e.g.* residues that are involved in binding of the antibody to a particular epitope), such as amino acid residues in CDR1, CDR2, and/or CDR3 regions of the light or heavy chains as described herein. Such amino acid substitutions may increase or decrease the affinity of the anti-B7RP1 antibodies described herein.

Minor changes in an amino acid sequence such as deletion, addition or substitution of one, a few or even several amino acids may lead to an allelic form of the original protein which has substantially identical properties. Therefore, in addition to the antibodies specifically described herein, other "substantially homologous" antibodies can be readily designed and manufactured utilizing various recombinant DNA techniques well known to those skilled in the art. In general, modifications of the genes may be readily accomplished by a variety of well-known techniques, such as site-directed mutagenesis. Therefore, the present invention contemplates "variant" or "mutant" anti-B7RP1 human antibodies having substantially similar characteristics to the anti-B7RP1 human antibodies disclosed herein (See, for example, WO 00/56772, all of which is hereby incorporated herein by reference). Thus, by the term "variant" or "mutant" in reference to an anti-B7RP1 human antibody is meant any binding molecule (molecule X) (i) in which the hypervariable regions CDR1, CDR2, and CDR3 of the heavy chain or the hypervariable regions CDR1, CDR2, and CDR3 of the light chain taken as a whole are at least about 80% homologous, at least about 90% homologous, or at least about

95% homologous to the hypervariable regions as shown in SEQ ID NO: 15 through SEQ ID NO. 26 or SEQ ID NO: 27 through SEQ ID NO: 40, respectively, and (ii) wherein the variant or mutant is capable of inhibiting the activity of human B7RP1 to the same extent as a reference anti-B7RP1 human antibody having framework regions identical to those of molecule X. Such antibodies may bind to human B7RP1 or to mouse B7RP1 or both. The mouse B7RP1 sequence is described in WO 00/46240, which is incorporated by reference.

Ordinarily, an anti-B7RP1 human antibody variant will have light and/or heavy chain CDRs, when taken as a whole, that are at least about 80% amino acid sequence identity, at least about 85% sequence identity, at least about 90% sequence identity, at least about 91% sequence identity, at least about 92% sequence identity, at least about 93% sequence identity, at least about 94% sequence identity, at least about 95% sequence identity, at least about 96% sequence identity, at least about 97% sequence identity, at least about 98% sequence identity, or at least about 99% amino acid sequence identity to the amino acid sequence as shown in SEQ ID NOS: 15 through SEQ ID NO. 26 and/or SEQ ID NOS: 27 through SEQ ID NO. 40, respectively. Such antibodies may bind to human B7RP1 or to mouse B7RP1 or to both.

An anti-B7RP1 human antibody variant will have a light chain variable region, when taken as a whole, that has at least about 80% amino acid sequence identity, at least about 81% sequence identity, at least about 82% sequence identity, at least about 83% sequence identity, at least about 84% sequence identity, at least about 85% sequence identity, at least about 86% sequence identity, at least about 87% sequence identity, at least about 88% sequence identity, at least about 89% sequence identity, at least about 90% sequence identity, at least about 91% sequence identity, at least about 92% sequence identity, at least about 93% sequence identity, at least about 94% sequence identity, at least about 95% sequence identity, at least about 96% sequence identity, at least about 97% sequence identity, at least about 98% sequence identity, at least about 99% amino acid sequence identity to the amino acid sequence as shown in SEQ ID NOS: 1 through SEQ ID NO. 6, and/or a heavy chain variable region, when taken as a whole, that has at least about 70% amino acid sequence identity, at least about 75% sequence identity, at least about 80% sequence identity, at least about 81% sequence identity, at least about 82% sequence identity, at least about 83% sequence

identity, at least about 84% sequence identity, at least about 85% sequence identity, at least about 86% sequence identity, at least about 87% sequence identity, at least about 88% sequence identity, at least about 89% sequence identity, at least about 90% sequence identity, at least about 91% sequence identity, at least about 92% sequence identity, at least about 93% sequence identity, at least about 94% sequence identity, at least about 95% sequence identity, at least about 96% sequence identity, at least about 97% sequence identity, at least about 98% sequence identity, or at least about 99% amino acid sequence identity to the amino acid sequence as shown in SEQ ID NOS: 7 through SEQ ID NO. 14. Such antibodies may bind to human B7RP1 and/or mouse B7RP1.

As will be appreciated by those of skill in the art, many of the potential CDR-contact residues are amenable to substitution by other amino acids and still allow the antibody to retain substantial affinity for the antigen. Likewise, many of the framework residues not in contact with the CDRs in the heavy and light chains can accommodate substitutions of amino acids from the corresponding positions from other human antibodies, by human consensus amino acids, or from other mouse antibodies, without significant loss of the affinity or non-immunogenicity of the human antibody. Selection of various alternative amino acids may be used to produce versions of the disclosed anti-B7RP1 antibodies and fragments thereof that have varying combinations of affinity, specificity, non-immunogenicity, ease of manufacture, and other desirable properties.

A "variant" in reference to a polynucleotide is intended to refer to a nucleic acid molecule having at least about 75% nucleic acid sequence identity with a polynucleotide sequence of the present invention. Ordinarily, a polynucleotide variant will have at least about 75% nucleic acid sequence identity, at least about 80% nucleic acid sequence identity, at least about 81% nucleic acid sequence identity, at least about 82% nucleic acid sequence identity, at least about 83% nucleic acid sequence identity, at least about 84% nucleic acid sequence identity, at least about 85% nucleic acid sequence identity, at least about 86% nucleic acid sequence identity, at least about 87% nucleic acid sequence identity, at least about 88% nucleic acid sequence identity, at least about 89% nucleic acid sequence identity, at least about 90% nucleic acid sequence identity, at least about 91% nucleic acid sequence identity, at least about 92% nucleic acid sequence identity, at least about 93% nucleic acid

sequence identity, at least about 94% nucleic acid sequence identity, at least about 95% nucleic acid sequence identity, at least about 96% nucleic acid sequence identity, at least about 97% nucleic acid sequence identity, at least about 98% nucleic acid sequence identity, or at least about 99% nucleic acid sequence identity with a novel nucleic acid sequence disclosed herein.

In alternative embodiments, antibodies of the invention can be expressed in cell lines other than hybridoma cell lines. In these embodiments, sequences encoding particular antibodies can be used for transformation of a suitable mammalian host cell. According to these embodiments, transformation can be achieved using any known method for introducing polynucleotides into a host cell, including, for example packaging the polynucleotide in a virus (or into a viral vector) and transducing a host cell with the virus (or vector) or by transfection procedures known in the art, as exemplified by U.S. Pat. Nos. 4,399,216, 4,912,040, 4,740,461, and 4,959,455 (all of which are hereby incorporated herein by reference for any purpose). Generally, the transformation procedure used may depend upon the host to be transformed. Methods for introducing heterologous polynucleotides into mammalian cells are well known in the art and include, but are not limited to, dextran-mediated transfection, calcium phosphate precipitation, polybrene mediated transfection, protoplast fusion, electroporation, encapsulation of the polynucleotide(s) in liposomes, and direct microinjection of the DNA into nuclei.

A nucleic acid molecule encoding the amino acid sequence of a heavy chain constant region, a heavy chain variable region, a light chain constant region, or a light chain variable region of an anti-B7RP1 antibody of the invention is inserted into an appropriate expression vector using standard ligation techniques. In one embodiment, the anti-B7RP1 antibody heavy chain or light chain constant region is appended to the C-terminus of the appropriate variable region and is ligated into an expression vector. The vector is typically selected to be functional in the particular host cell employed (*i.e.*, the vector is compatible with the host cell machinery such that amplification of the gene and/or expression of the gene can occur). For a review of expression vectors, *see* METHODS IN. ENZYMOLOGY 185 (Goeddel, ed.), 1990, Academic Press.

Typically, expression vectors used in any of the host cells will contain sequences for plasmid maintenance and for cloning and expression of exogenous nucleotide sequences. Such sequences, collectively referred to as "flanking

sequences" in certain embodiments will typically include one or more of the following nucleotide sequences: a promoter, one or more enhancer sequences, an origin of replication, a transcriptional termination sequence, a complete intron sequence containing a donor and acceptor splice site, a sequence encoding a leader sequence for polypeptide secretion, a ribosome binding site, a polyadenylation sequence, a polylinker region for inserting the nucleic acid encoding the polypeptide to be expressed, and a selectable marker element. Each of these sequences is discussed below.

Optionally, the vector may contain a "tag"-encoding sequence, *i.e.*, an oligonucleotide molecule located at the 5' or 3' end of the anti-B7RP1 antibody polypeptide coding sequence; the oligonucleotide sequence encodes polyHis (such as hexaHis), or another "tag" such as FLAG, HA (hemaglutinin influenza virus), or *myc* for which commercially available antibodies exist. This tag is typically fused to the polypeptide upon expression of the polypeptide, and can serve as a means for affinity purification or detection of the anti-B7RP1 antibody from the host cell. Affinity purification can be accomplished, for example, by column chromatography using antibodies against the tag as an affinity matrix. Optionally, the tag can subsequently be removed from the purified anti-B7RP1 antibody polypeptide by various means such as using certain peptidases for cleavage.

Flanking sequences may be homologous (*i.e.*, from the same species and/or strain as the host cell), heterologous (*i.e.*, from a species other than the host cell species or strain), hybrid (*i.e.*, a combination of flanking sequences from more than one source), synthetic or native. As such, the source of a flanking sequence may be any prokaryotic or eukaryotic organism, any vertebrate or invertebrate organism, or any plant, provided that the flanking sequence is functional in, and can be activated by, the host cell machinery.

Flanking sequences useful in the vectors of this invention may be obtained by any of several methods well known in the art. Typically, flanking sequences useful herein will have been previously identified by mapping and/or by restriction endonuclease digestion and can thus be isolated from the proper tissue source using the appropriate restriction endonucleases. In some cases, the full nucleotide sequence of a flanking sequence may be known. Here, the flanking sequence may be synthesized using the methods described herein for nucleic acid synthesis or cloning.

Whether all or only a portion of the flanking sequence is known, it may be obtained using polymerase chain reaction (PCR) and/or by screening a genomic library with a suitable probe such as an oligonucleotide and/or flanking sequence fragment from the same or another species. Where the flanking sequence is not known, a fragment of DNA containing a flanking sequence may be isolated from a larger piece of DNA that may contain, for example, a coding sequence or even another gene or genes. Isolation may be accomplished by restriction endonuclease digestion to produce the proper DNA fragment followed by isolation using agarose gel purification, Qiagen® column chromatography (Chatsworth, CA), or other methods known to the skilled artisan. The selection of suitable enzymes to accomplish this purpose will be readily apparent to one of ordinary skill in the art.

An origin of replication is typically a part of those prokaryotic expression vectors purchased commercially, and the origin aids in the amplification of the vector in a host cell. If the vector of choice does not contain an origin of replication site, one may be chemically synthesized based on a known sequence, and ligated into the vector. For example, the origin of replication from the plasmid pBR322 (New England Biolabs, Beverly, MA) is suitable for most gram-negative bacteria, and various viral origins (*e.g.*, SV40, polyoma, adenovirus, vesicular stomatitus virus (VSV), or papillomaviruses such as HPV or BPV) are useful for cloning vectors in mammalian cells. Generally, the origin of replication component is not needed for mammalian expression vectors (*for example*, the SV40 origin is often used only because it also contains the virus early promoter).

A transcription termination sequence is typically located 3' to the end of a polypeptide coding region and serves to terminate transcription. Usually, a transcription termination sequence in prokaryotic cells is a G-C rich fragment followed by a poly-T sequence. While the sequence is easily cloned from a library or even purchased commercially as part of a vector, it can also be readily synthesized using methods for nucleic acid synthesis such as those described herein.

A selectable marker gene encodes a protein necessary for the survival and growth of a host cell grown in a selective culture medium. Typical selection marker genes encode proteins that (a) confer resistance to antibiotics or other toxins, *e.g.*, ampicillin, tetracycline, or kanamycin for prokaryotic host cells; (b) complement auxotrophic deficiencies of the cell; or (c) supply critical nutrients not available from

complex or defined media. Exemplary selectable markers are the kanamycin resistance gene, the ampicillin resistance gene, and the tetracycline resistance gene. Advantageously, a neomycin resistance gene may also be used for selection in both prokaryotic and eukaryotic host cells.

Other selectable genes may be used to amplify the gene that will be expressed. Amplification is the process wherein genes that are required for production of a protein critical for growth or cell survival are reiterated in tandem within the chromosomes of successive generations of recombinant cells. Examples of suitable selectable markers for mammalian cells include dihydrofolate reductase (DHFR) and promoterless thymidine kinase genes. Mammalian cell transformants are placed under selection pressure wherein only the transformants are uniquely adapted to survive by virtue of the selectable gene present in the vector. Selection pressure is imposed by culturing the transformed cells under conditions in which the concentration of selection agent in the medium is successively increased, thereby leading to the amplification of both the selectable gene and the DNA that encodes another gene, such as an antibody that binds to B7RP1 polypeptide. As a result, increased quantities of a polypeptide such as an anti-B7RP1 antibody are synthesized from the amplified DNA.

A ribosome-binding site is usually necessary for translation initiation of mRNA and is characterized by a Shine-Dalgarno sequence (prokaryotes) or a Kozak sequence (eukaryotes). The element is typically located 3' to the promoter and 5' to the coding sequence of the polypeptide to be expressed.

In some cases, such as where glycosylation is desired in a eukaryotic host cell expression system, one may manipulate the various pre- or prosequences to improve glycosylation or yield. For example, one may alter the peptidase cleavage site of a particular signal peptide, or add pro-sequences, which also may affect glycosylation. The final protein product may have, in the -1 position (relative to the first amino acid of the mature protein) one or more additional amino acids incident to expression, which may not have been totally removed. For example, the final protein product may have one or two amino acid residues found in the peptidase cleavage site, attached to the amino-terminus. Alternatively, use of some enzyme cleavage sites may result in a slightly truncated form of the desired polypeptide, if the enzyme cuts at such an area within the mature polypeptide.

Expression and cloning vectors of the invention will typically contain a promoter that is recognized by the host organism and operably linked to the molecule encoding the anti-B7RP1 antibody. Promoters are untranscribed sequences located upstream (*i.e.*, 5') to the start codon of a structural gene (generally within about 100 to 1000 bp) that control transcription of the structural gene. Promoters are conventionally grouped into one of two classes: inducible promoters and constitutive promoters. Inducible promoters initiate increased levels of transcription from DNA under their control in response to some change in culture conditions, such as the presence or absence of a nutrient or a change in temperature. Constitutive promoters, on the other hand, uniformly transcribe genes to which they are operably linked, that is, with little or no control over gene expression. A large number of promoters, recognized by a variety of potential host cells, are well known. A suitable promoter is operably linked to the DNA encoding heavy chain or light chain comprising an anti-B7RP1 antibody of the invention by removing the promoter from the source DNA by restriction enzyme digestion and inserting the desired promoter sequence into the vector.

Suitable promoters for use with yeast hosts are also well known in the art. Yeast enhancers are advantageously used with yeast promoters. Suitable promoters for use with mammalian host cells are well known and include, but are not limited to, those obtained from the genomes of viruses such as polyoma virus, fowlpox virus, adenovirus (such as Adenovirus 2), bovine papilloma virus, avian sarcoma virus, cytomegalovirus, retroviruses, hepatitis-B virus and Simian Virus 40 (SV40). Other suitable mammalian promoters include heterologous mammalian promoters, for example, heat-shock promoters and the actin promoter.

Additional promoters which may be of interest include, but are not limited to: SV40 early promoter (Bernoist and Chambon, 1981, *Nature* 290:304-10); CMV promoter (Thomsen *et al.*, 1984, *Proc. Natl. Acad. Sci. USA* 81:659-663); the promoter contained in the 3' long terminal repeat of Rous sarcoma virus (Yamamoto, *et al.*, 1980, *Cell* 22:787-97); herpes thymidine kinase promoter (Wagner *et al.*, 1981, *Proc. Natl. Acad. Sci. U.S.A.* 78:1444-45); promoter and regulatory sequences from the metallothionine gene (Brinster *et al.*, 1982, *Nature* 296:39-42); and prokaryotic promoters such as the beta-lactamase promoter (Villa-Kamaroff *et al.*, 1978, *Proc. Natl. Acad. Sci U.S.A.*, 75:3727-31); or the tac promoter (DeBoer *et al.*, 1983, *Proc.*

Natl. Acad Sci. U.S.A., 80:21-25). Also of interest are the following animal transcriptional control regions, which exhibit tissue specificity and have been utilized in transgenic animals: the elastase I gene control region that is active in pancreatic acinar cells (Swift *et al.*, 1984, *Cell* 38:639-46; Ornitz *et al.*, 1986, *Cold Spring Harbor Symp. Quant Biol.* 50:399-409 (1986); MacDonald, 1987, *Hepatology* 7:425-515); the insulin gene control region that is active in pancreatic beta cells (Hanahan, 1985, *Nature* 315:115-22); the immunoglobulin gene control region that is active in lymphoid cells (Grosschedl *et al.*, 1984, *Cell* 38:647-58; Adames *et al.*, 1985, *Nature* 318:533-38; Alexander *et al.*, 1987, *Mol. Cell. Biol.*, 7:1436-44); the mouse mammary tumor virus control region that is active in testicular, breast, lymphoid and mast cells (Leder *et al.*, 1986, *Cell* 45:485-95); the albumin gene control region that is active in liver (Pinkert *et al.*, 1987, *Genes and Devel.* 1:268-76); the alpha-feto-protein gene control region that is active in liver (Krumlauf *et al.*, 1985, *Mol. Cell. Biol.*, 5:1639-48; Hammer *et al.*, 1987, *Science* 235:53-58); the alpha 1-antitrypsin gene control region that is active in liver (Kelsey *et al.*, 1987, *Genes and Devel.* 1:161-71); the beta-globin gene control region that is active in myeloid cells (Mogram *et al.*, 1985, *Nature* 315:338-40; Kollias *et al.*, 1986, *Cell* 46:89-94); the myelin basic protein gene control region that is active in oligodendrocyte cells in the brain (Readhead *et al.*, 1987, *Cell* 48:703-12); the myosin light chain-2 gene control region that is active in skeletal muscle (Sani, 1985, *Nature* 314:283-86); and the gonadotropic releasing hormone gene control region that is active in the hypothalamus (Mason *et al.*, 1986, *Science* 234:1372-78).

An enhancer sequence may be inserted into the vector to increase transcription of DNA encoding light chain or heavy chain comprising an anti-B7RP1 antibody of the invention by higher eukaryotes. Enhancers are *cis*-acting elements of DNA, usually about 10-300 bp in length, that act on the promoter to increase transcription. Enhancers are relatively orientation and position independent, having been found at positions both 5' and 3' to the transcription unit. Several enhancer sequences available from mammalian genes are known (*e.g.*, globin, elastase, albumin, alpha-feto-protein and insulin). Typically, however, an enhancer from a virus is used. The SV40 enhancer, the cytomegalovirus early promoter enhancer, the polyoma enhancer, and adenovirus enhancers known in the art are exemplary enhancing elements for the activation of eukaryotic promoters. While an enhancer may be positioned in the

vector either 5' or 3' to a coding sequence, it is typically located at a site 5' from the promoter.

Expression vectors of the invention may be constructed from a starting vector such as a commercially available vector. Such vectors may or may not contain all of the desired flanking sequences. Where one or more of the flanking sequences described herein are not already present in the vector, they may be individually obtained and ligated into the vector. Methods used for obtaining each of the flanking sequences are well known to one skilled in the art.

After the vector has been constructed and a nucleic acid molecule encoding light chain, a heavy chain, or a light chain and a heavy chain comprising an anti-B7RP1 antibody has been inserted into the proper site of the vector, the completed vector may be inserted into a suitable host cell for amplification and/or polypeptide expression. The transformation of an expression vector for an anti-B7RP1 antibody into a selected host cell may be accomplished by well known methods including transfection, infection, calcium phosphate co-precipitation, electroporation, microinjection, lipofection, DEAE-dextran mediated transfection, or other known techniques. The method selected will in part be a function of the type of host cell to be used. These methods and other suitable methods are well known to the skilled artisan, and are set forth, for example, in Sambrook *et al.*, *supra*.

A host cell, when cultured under appropriate conditions, synthesizes an anti-B7RP1 antibody that can subsequently be collected from the culture medium (if the host cell secretes it into the medium) or directly from the host cell producing it (if it is not secreted). The selection of an appropriate host cell will depend upon various factors, such as desired expression levels, polypeptide modifications that are desirable or necessary for activity (such as glycosylation or phosphorylation) and ease of folding into a biologically active molecule

Mammalian cell lines available as hosts for expression are well known in the art and include, but are not limited to, immortalized cell lines available from the American Type Culture Collection (ATCC), including but not limited to Chinese hamster ovary (CHO) cells, HeLa cells, baby hamster kidney (BHK) cells, monkey kidney cells (COS), human hepatocellular carcinoma cells (*e.g.*, Hep G2), and a number of other cell lines. In certain embodiments, cell lines may be selected through

determining which cell lines have high expression levels and constitutively produce antibodies with B7RP1 binding properties. In another embodiment, a cell line from the B cell lineage that does not make its own antibody but has a capacity to make and secrete a heterologous antibody can be selected.

Antibodies of the invention are useful for detecting B7RP1 in biological samples and identification of cells or tissues that produce B7RP1 protein. Antibodies of the invention that specifically bind to B7RP1 may be useful in treatment of B7RP1 mediated diseases. Said antibodies can be used in binding assays to detect B7RP1 and to inhibit B7RP1 from forming a complex with B7RP1 receptors. Said antibodies that bind to B7RP1 and block interaction with other binding compounds may have therapeutic use in modulating B7RP1 mediated diseases. In certain embodiments, antibodies to B7RP1 may block B7RP1 binding to its receptor, which may result in disruption of the B7RP1 induced signal transduction cascade.

The present invention also relates to the use of one or more of the antibodies of the present invention in the manufacture of a medicament for the treatment of a disorder or condition caused by increased expression of B7RP1 or increased sensitivity to B7RP1 in a patient such as any one of disorders or conditions disclosed herein.

In certain embodiments, the invention provides pharmaceutical compositions comprising a therapeutically effective amount of one or a plurality of the antibodies of the invention together with a pharmaceutically acceptable diluent, carrier, solubilizer, emulsifier, preservative and/or adjuvant. Acceptable formulation materials are nontoxic to recipients at the dosages and concentrations employed. In preferred embodiments, pharmaceutical compositions comprising a therapeutically effective amount of anti-B7RP1 antibodies are provided.

In certain embodiments, acceptable formulation materials are nontoxic to recipients at the dosages and concentrations employed.

In certain embodiments, the pharmaceutical composition may contain formulation materials for modifying, maintaining or preserving, for example, the pH, osmolarity, viscosity, clarity, color, isotonicity, odor, sterility, stability, rate of dissolution or release, adsorption or penetration of the composition. In such embodiments, suitable formulation materials include, but are not limited to, amino

acids (such as glycine, glutamine, asparagine, arginine or lysine); antimicrobials; antioxidants (such as ascorbic acid, sodium sulfite or sodium hydrogen-sulfite); buffers (such as borate, bicarbonate, Tris-HCl, citrates, phosphates or other organic acids); bulking agents (such as mannitol or glycine); chelating agents (such as ethylenediamine tetraacetic acid (EDTA)); complexing agents (such as caffeine, polyvinylpyrrolidone, beta-cyclodextrin or hydroxypropyl-beta-cyclodextrin); fillers; monosaccharides; disaccharides; and other carbohydrates (such as glucose, mannose or dextrins); proteins (such as serum albumin, gelatin or immunoglobulins); coloring, flavoring and diluting agents; emulsifying agents; hydrophilic polymers (such as polyvinylpyrrolidone); low molecular weight polypeptides; salt-forming counterions (such as sodium); preservatives (such as benzalkonium chloride, benzoic acid, salicylic acid, thimerosal, phenethyl alcohol, methylparaben, propylparaben, chlorhexidine, sorbic acid or hydrogen peroxide); solvents (such as glycerin, propylene glycol or polyethylene glycol); sugar alcohols (such as mannitol or sorbitol); suspending agents; surfactants or wetting agents (such as pluronics, PEG, sorbitan esters, polysorbates such as polysorbate 20, polysorbate 80, triton, tromethamine, lecithin, cholesterol, tyloxapal); stability enhancing agents (such as sucrose or sorbitol); tonicity enhancing agents (such as alkali metal halides, for example, sodium or potassium chloride, mannitol sorbitol); delivery vehicles; diluents; excipients and/or pharmaceutical adjuvants. *See* REMINGTON'S PHARMACEUTICAL SCIENCES, 18th Edition, (A.R. Gennaro, ed.), 1990, Mack Publishing Company.

In certain embodiments, the optimal pharmaceutical composition will be determined by one skilled in the art depending upon, for example, the intended route of administration, delivery format and desired dosage. *See, for example,* REMINGTON'S PHARMACEUTICAL SCIENCES, *supra.* In certain embodiments, such compositions may influence the physical state, stability, rate of *in vivo* release and rate of *in vivo* clearance of the antibodies of the invention.

In certain embodiments, the primary vehicle or carrier in a pharmaceutical composition may be either aqueous or non-aqueous in nature. For example, a suitable vehicle or carrier may be water for injection, physiological saline solution or artificial cerebrospinal fluid, possibly supplemented with other materials common in compositions for parenteral administration. Neutral buffered saline or saline mixed

with serum albumin are further exemplary vehicles. In certain embodiments, pharmaceutical compositions of the present invention comprise Tris buffer of about pH 7.0-8.5, or acetate buffer of about pH 4.0-5.5, and may further include sorbitol, sucrose, Tween-20 and/or a suitable substitute therefor. In certain embodiments of the invention, anti-B7RP1 antibody compositions may be prepared for storage by mixing the selected composition having the desired degree of purity with optional formulation agents (REMINGTON'S PHARMACEUTICAL SCIENCES, *supra*) in the form of a lyophilized cake or an aqueous solution. Further, in certain embodiments, the anti-B7RP1 antibody product may be formulated as a lyophilizate using appropriate excipients such as sucrose.

The pharmaceutical compositions of the invention can be selected for parenteral delivery. Alternatively, the compositions may be selected for inhalation or for delivery through the digestive tract, such as orally. Preparation of such pharmaceutically acceptable compositions is within the skill of the art.

The formulation components are present in concentrations that are acceptable to the site of administration. In certain embodiments, buffers are used to maintain the composition at physiological pH or at a slightly lower pH, typically within a pH range of from about 5 to about 8.

When parenteral administration is contemplated, the therapeutic compositions for use in this invention may be provided in the form of a pyrogen-free, parenterally acceptable aqueous solution comprising the desired anti-B7RP1 antibody in a pharmaceutically acceptable vehicle. A particularly suitable vehicle for parenteral injection is sterile distilled water in which the anti-B7RP1 antibody is formulated as a sterile, isotonic solution, properly preserved. In certain embodiments, the preparation can involve the formulation of the desired molecule with an agent, such as injectable microspheres, bio-erodible particles, polymeric compounds (such as polylactic acid or polyglycolic acid), beads or liposomes, that may provide controlled or sustained release of the product which can be delivered *via* depot injection. In certain embodiments, hyaluronic acid may also be used, having the effect of promoting sustained duration in the circulation. In certain embodiments, implantable drug delivery devices may be used to introduce the desired antibody molecule.

Pharmaceutical compositions of the invention can be formulated for inhalation. In these embodiments, anti-B7RP1 antibodies are advantageously formulated as a dry, inhalable powder. In certain embodiments, anti-B7RP1 antibody inhalation solutions may also be formulated with a propellant for aerosol delivery. In certain embodiments, solutions may be nebulized. Pulmonary administration and formulation methods therefore are further described in International Patent Application No. PCT/US94/001875, which is incorporated by reference and describes pulmonary delivery of chemically modified proteins.

It is also contemplated that formulations can be administered orally. Anti-B7RP1 antibodies that are administered in this fashion can be formulated with or without carriers customarily used in the compounding of solid dosage forms such as tablets and capsules. In certain embodiments, a capsule may be designed to release the active portion of the formulation at the point in the gastrointestinal tract when bioavailability is maximized and pre-systemic degradation is minimized. Additional agents can be included to facilitate absorption of the anti-B7RP1 antibody. Diluents, flavorings, low melting point waxes, vegetable oils, lubricants, suspending agents, tablet disintegrating agents, and binders may also be employed.

A pharmaceutical composition of the invention is provided to comprise an effective quantity of one or a plurality of anti-B7RP1 antibodies in a mixture with non-toxic excipients that are suitable for the manufacture of tablets. By dissolving the tablets in sterile water, or another appropriate vehicle, solutions may be prepared in unit-dose form. Suitable excipients include, but are not limited to, inert diluents, such as calcium carbonate, sodium carbonate or bicarbonate, lactose, or calcium phosphate; or binding agents, such as starch, gelatin, or acacia; or lubricating agents such as magnesium stearate, stearic acid, or talc.

Additional pharmaceutical compositions will be evident to those skilled in the art, including formulations involving anti-B7RP1 antibodies in sustained- or controlled-delivery formulations. Techniques for formulating a variety of other sustained- or controlled-delivery means, such as liposome carriers, bio-erodible microparticles or porous beads and depot injections, are also known to those skilled in the art. *See, for example*, International Patent Application No. PCT/US93/00829, which is incorporated by reference and describes controlled release of porous polymeric microparticles for delivery of pharmaceutical compositions. Sustained-

release preparations may include semipermeable polymer matrices in the form of shaped articles, *e.g.* films, or microcapsules. Sustained release matrices may include polyesters, hydrogels, polylactides (as disclosed in U.S. Patent No. 3,773,919 and European Patent Application Publication No. EP 058481, each of which is incorporated by reference), copolymers of L-glutamic acid and gamma ethyl-L-glutamate (Sidman *et al.*, 1983, *Biopolymers* 22:547-556), poly (2-hydroxyethyl-methacrylate) (Langer *et al.*, 1981, *J. Biomed. Mater. Res.* 15:167-277 and Langer, 1982, *Chem. Tech.* 12:98-105), ethylene vinyl acetate (Langer *et al.*, *supra*) or poly-D(-)-3-hydroxybutyric acid (European Patent Application Publication No. EP 133,988). Sustained release compositions may also include liposomes that can be prepared by any of several methods known in the art. See *e.g.*, Eppstein *et al.*, 1985, *Proc. Natl. Acad. Sci. USA* 82:3688-3692; European Patent Application Publication Nos. EP 036,676; EP 088,046 and EP 143,949, incorporated by reference.

Pharmaceutical compositions used for *in vivo* administration are typically provided as sterile preparations. Sterilization can be accomplished by filtration through sterile filtration membranes. When the composition is lyophilized, sterilization using this method may be conducted either prior to or following lyophilization and reconstitution. Compositions for parenteral administration can be stored in lyophilized form or in a solution. Parenteral compositions generally are placed into a container having a sterile access port, for example, an intravenous solution bag or vial having a stopper pierceable by a hypodermic injection needle.

Once the pharmaceutical composition has been formulated, it may be stored in sterile vials as a solution, suspension, gel, emulsion, solid, or as a dehydrated or lyophilized powder. Such formulations may be stored either in a ready-to-use form or in a form (*e.g.*, lyophilized) that is reconstituted prior to administration.

The invention also provides kits for producing a single-dose administration unit. The kits of the invention may each contain both a first container having a dried protein and a second container having an aqueous formulation. In certain embodiments of this invention, kits containing single and multi-chambered pre-filled syringes (*e.g.*, liquid syringes and lyosyringes) are provided.

The effective amount of an anti-B7RP1 antibody-containing pharmaceutical composition to be employed therapeutically will depend, for example, upon the

therapeutic context and objectives. One skilled in the art will appreciate that the appropriate dosage levels for treatment will vary depending, in part, upon the molecule delivered, the indication for which the anti-B7RP1 antibody is being used, the route of administration, and the size (body weight, body surface or organ size) and/or condition (the age and general health) of the patient. In certain embodiments, the clinician may titer the dosage and modify the route of administration to obtain the optimal therapeutic effect. A typical dosage may range from about 0.1 μg/kg to up to about 30 mg/kg or more, depending on the factors mentioned above. In certain embodiments, the dosage may range from 0.1 μg/kg up to about 30 mg/kg; from 1 μg/kg up to about 30 mg/kg; or from 5 μg/kg up to about 30 mg/kg.

Dosing frequency will depend upon the pharmacokinetic parameters of the particular anti-B7RP1 antibody in the formulation used. Typically, a clinician administers the composition until a dosage is reached that achieves the desired effect. The composition may therefore be administered as a single dose, or as two or more doses (which may or may not contain the same amount of the desired molecule) over time, or as a continuous infusion via an implantation device or catheter. Further refinement of the appropriate dosage is routinely made by those of ordinary skill in the art and is within the ambit of tasks routinely performed by them. Appropriate dosages may be ascertained through use of appropriate dose-response data. In certain embodiments, the antibodies of the invention can be administered to patients throughout an extended time period. Chronic administration of an antibody of the invention minimizes the adverse immune or allergic response commonly associated with antibodies that are raised against a human antigen in a non-human animal, for example, a non-fully human antibody produced in a non-human species.

The route of administration of the pharmaceutical composition is in accord with known methods, *e.g.* orally, through injection by intravenous, intraperitoneal, intracerebral (intra-parenchymal), intracerebroventricular, intramuscular, intra-ocular, intraarterial, intraportal, or intralesional routes; by sustained release systems or by implantation devices. In certain embodiments, the compositions may be administered by bolus injection or continuously by infusion, or by implantation device.

The composition also may be administered locally via implantation of a membrane, sponge or another appropriate material onto which the desired molecule

has been absorbed or encapsulated. In certain embodiments, where an implantation device is used, the device may be implanted into any suitable tissue or organ, and delivery of the desired molecule may be *via* diffusion, timed-release bolus, or continuous administration.

It also may be desirable to use anti-B7RP1 antibody pharmaceutical compositions according to the invention *ex vivo*. In such instances, cells, tissues or organs that have been removed from the patient are exposed to anti-B7RP1 antibody pharmaceutical compositions after which the cells, tissues and/or organs are subsequently implanted back into the patient.

In particular, anti-B7RP1 antibodies can be delivered by implanting certain cells that have been genetically engineered, using methods such as those described herein, to express and secrete the polypeptide. In certain embodiments, such cells may be animal or human cells, and may be autologous, heterologous, or xenogeneic. In certain embodiments, the cells may be immortalized. In other embodiments, in order to decrease the chance of an immunological response, the cells may be encapsulated to avoid infiltration of surrounding tissues. In further embodiments, the encapsulation materials are typically biocompatible, semi-permeable polymeric enclosures or membranes that allow the release of the protein product(s) but prevent the destruction of the cells by the patient's immune system or by other detrimental factors from the surrounding tissues.

EXAMPLES

The following examples, including the experiments conducted and results achieved are provided for illustrative purposes only and are not to be construed as limiting the invention.

Example 1

Production of Human Monoclonal Antibodies Against B7 related protein-1 (B7RP1)

Antigen

Purified recombinant human B7RP-1 (hB7RP-1) prepared as described in International Patent Application Publication No. WO 00/46240, which is incorporated herein by reference, or CHO cells transfected to express hB7RP-1 were used as the antigen. Mature human B7RP-1 has the amino acid sequence of residues X to 302 in the sequence shown in WO 00/46240 as SEQ ID NO: 17, wherein X can be 19, 20, 21, 22, 24 or 28.

Transgenic HuMab Mice

Fully human monoclonal antibodies to B7RP-1 were prepared using HCo7 and HCo12 strains of HuMab transgenic mice, both of which express human antibody genes. In both of these mouse strains, the endogenous mouse kappa light chain gene has been homozygously disrupted as described in Chen *et al.* (1993) *EMBO J.* 12:811-820 and the endogenous mouse heavy chain gene has been homozygously disrupted as described in Example 1 of PCT Publication WO 01/09187. Each of these mouse strains carries a human kappa light chain transgene, KCo5, as described in Fishwild *et al.* (1996) *Nature Biotechnology* 14:845-851. The HCo7 strain carries the HCo7 human heavy chain transgene as described in U.S. Patent Nos. 5,545,806; 5,625,825; and 5,545,807. The HCo12 strain carries the HCo12 human heavy chain transgene as described in Example 2 of PCT Publication WO 01/09187.

HuMab Immunizations:

To generate fully human monoclonal antibodies to B7RP-1, HuMab mice of the HCo7 or HCo12 strain were immunized with purified recombinant B7RP-1 or CHO cells transfected to express B7RP-1. General immunization schemes for HuMab mice are described in Lonberg *et al* (1994) *Nature* 368(6474): 856-859; Fishwild *et al.* (1996) *Nature Biotechnology* 14: 845-851 and PCT Publication WO 98/24884. The mice were 6-16 weeks of age upon the first infusion of antigen. A purified recombinant preparation of B7RP-1 antigen (50 μg) or a preparation of transfected CHO cells ($3.5 \times 10^6 - 1 \times 10^7$ cells) was used to immunize the HuMab mice intraperitonealy.

Transgenic mice were immunized twice with purified antigen in complete Freund's adjuvant intraperitonealy, followed by 2-4 weeks of IP immunizations (up to a total of 8 immunizations) with the purified antigen in incomplete Freund's adjuvant.

Immunization with CHO cells transfected to express B7RP-1 was the same except that complete Freund's adjuvant and incomplete Freund's adjuvant were not used with the cells. The immune response was monitored by retroorbital bleeds. The plasma was screened by ELISA (as described below), and mice with sufficient titers of anti-B7RP-1 human immunogolobulin were used for fusions. Mice were boosted intravenously with antigen 3 and 2 days before sacrifice and removal of the spleen. Typically, 10-20 fusions for each antigen were performed. Several dozen mice were immunized for each antigen. A total of 28 mice of the HCo7 and HCo12 mice strains were immunized with B7RP-1.

Selection of HuMab Mice Producing Anti-B7RP-1 Antibodies:

To select HuMab mice producing antibodies that bound B7RP-1, sera from immunized mice was tested by ELISA as described by Fishwild *et al.* (1996). Briefly, microtiter plates were coated with purified recombinant B7RP-1 at 1-2 µg /ml in PBS, 50 µl/wells incubated 4 °C overnight then blocked with 200 µl/well of 5% chicken serum in PBS/Tween (0.05%). Dilutions of plasma from B7RP-1-immunized mice were added to each well and incubated for 1-2 hours at ambient temperature. The plates were washed with PBS/Tween and then incubated with a goat-anti-human IgG Fc polyclonal antibody conjugated with horseradish peroxidase (HRP) for 1 hour at room temperature. After washing, the plates were developed with ABTS substrate (Sigma, A-1888, 0.22 mg/ml) and analyzed by spectrophotometer at OD 415-495. Mice that developed the highest titers of anti-B7RP-1 antibodies were used for fusions. Fusions were performed as described below and hybridoma supernatants were tested for anti-B7RP-1 activity by ELISA.

Generation of Hybridomas Producing Human Monoclonal Antibodies to B7RP-1:

The mouse splenocytes, isolated from the HuMab mice, were fused with PEG to a mouse myeloma cell line based upon standard protocols. The resulting hybridomas were then screened for the production of antigen-specific antibodies. Single cell suspensions of splenic lymphocytes from immunized mice were fused to one-fourth the number of SP2/0 nonsecreting mouse myeloma cells (ATCC, CRL 1581) with 50% PEG (Sigma). Cells were plated at approximately $1x10^{5}$/well in flat

bottom microtiter plate, followed by about two week incubation in selective medium containing 10% fetal bovine serum, 10% P388D1 (ATCC, CRL TIB-63) conditioned medium, 3-5% origen (IGEN) in DMEM (Mediatech, CRL 10013, with high glucose, L-glutamine and sodium pyruvate) plus 5 mM HEPES, 0.055 mM 2-mercaptoethanol, 50 mg/ml gentamycin and 1x HAT (Sigma, CRL P-7185). After 1-2 weeks, cells were cultured in medium in which the HAT was replaced with HT. Individual wells were then screened by ELISA (described above) for human anti-B7RP-1 monoclonal IgG antibodies. Once extensive hybridoma growth occurred, medium was monitored usually after 10-14 days. The antibody secreting hybridomas were replated, screened again and, if still positive for human IgG, anti-B7RP-1 monoclonal antibodies were subcloned at least twice by limiting dilution. The stable subclones were then cultured *in vitro* to generate small amounts of antibody in tissue culture medium for further characterization.

Example 2

Cloning the anti-B7RP1 Antibody Heavy and Light Chains

The hybridoma expressing the B7RP1 binding monoclonal antibody 16H was used as a source to isolate total RNA using TRIzol® reagent (Invitrogen). A 5' RACE (rapid amplification of cDNA ends) oligonucleotide (5'- CGA CUG GAG CAC GAG GAC ACU GAC AUG GAC UGA AGG AGU AGA AA-3'; SEQ ID NO: 69) was ligated to the RNA using the GeneRacer™ Kit (Invitrogen) components and protocol. First strand cDNA was synthesized using a random primer with an extension adapter (5'-_ GGC CGG ATA GGC CTC CAN NNN NNT-3') (SEQ ID NO: 59) and a 5' RACE (rapid amplification of cDNA ends) preparative assay was performed using the GeneRacer™ Kit (Invitrogen) according to instructions from the manufacturer. For preparing complete light chain encoding cDNA, the forward primer was the GeneRacer™ nested primer, and the reverse primer was (5'- GGG GTC AGG CTG GAA CTG AGG-3') (SEQ ID NO: 60). For preparing cDNA encoding the variable region of the heavy chain, the forward primer was the GeneRacer™ nested primer and the reverse primer was (5'- TGA GGA CGC TGA CCA CAC G-3') (SEQ ID NO: 61). RACE products were cloned into pCR4-TOPO

(Invitrogen) and the sequences determined. Consensus sequences were used to design primers for full-length antibody chain PCR amplification.

For preparing cDNA encoding anti-B7RP1 16H kappa light chain, the 5' PCR primer encoded the amino terminus of the signal sequence, an *Xba*I restriction enzyme site, and an optimized Kozak sequence (5'-CAG CAG AAG CTT CTA GAC CAC CAT GGA CAT GAG GGT CCT CGC TCA GCT CCT GGG-3') (SEQ ID NO: 62). The 3' primer encoded the carboxyl terminus and termination codon, as well as a *Sal*I restriction site (5'-CTT GTC GAC TCA ACA CTC TCC CCT GTT GAA GCT C-3') (SEQ ID NO: 63). The resulting PCR product fragment was purified, digested with *Xba*I and *Sal*I, and then gel isolated and ligated into the mammalian expression vector pDSRα20 (*see* International Application, Publication No. WO 90/14363, which is herein incorporated by reference for any purpose. pDSRα20 was produced by changing nucleotide 2563 in pDSRa19 from a "Guanosine" to an "Adenosine" by site directed mutagenesis.).

For preparing cDNA encoding anti- B7RP1 16H heavy chain the 5' PCR primer encoded the amino terminus of the signal sequence, an *Xba*I restriction enzyme site, and an optimized Kozak sequence (5'-ACA ACA AAG CTT CTA GAC CAC CAT GGA GTT GGG GCT GAA CTG G-3') (SEQ ID NO: 64). The 3' primer encoded the carboxyl end of the variable region, including a naturally occurring sense strand *BsmB*I site (5'- GTG GAG GCA CTA GAG ACG GTG ACC AGG ATT CC - 3'; SEQ ID NO: 65). The resulting product was purified, digested with *Xba*I and *BsmB*I, gel isolated and ligated into the pDSRα20 vector containing the human IgG1 constant region and also into the pDSRα20 vector containing the human IgG2 constant region. All of the hybridoma derived anti-B7RP1 heavy chain variable regions, regardless of the native constant region associated, were cloned as described above into both the pDSRα20 vectors containing the human IgG1 and the human IgG2 constant regions.

Example 3

Expression of Anti-B7RP1 Antibodies in Chinese Hamster Ovary (CHO) Cells

Stable expression of the 16H anti-B7RP1 mAb was achieved by co-transfection of 16H-heavy chain/pDSRα19 IgG2 B7RP1-kappa/pDSRα19 plasmids

into dihydrofolate reductase deficient (DHFR$^-$) serum-free adapted Chinese hamster ovary (CHO) cells using a calcium phosphate method (the full length 16H heavy chain sequence is shown in SEQ ID NO: 44; the 16H kappa chain sequence is shown in SEQ ID NO: 45). Transfected cells were selected in medium containing dialyzed serum but not containing hypoxanthine-thymidine to ensure the growth of cells expressing the DHFR enzyme. Transfected clones were screened using assays such as ELISA in order to detect the expression of 16H anti-B7RP1 mAb in the conditioned medium. The highest expressing clones were subjected to increasing concentrations of methotrexate (MTX) for DHFR amplification. MTX amplified clones were screened using assays such as ELISA in order to detect higher expression of 16H anti-B7RP1 mAb in the conditioned medium. The highest expressing clones were subjected to subcloning to obtain a homogeneous population and creation of cell banks.

Other recombinant anti-B7RP1 antibodies of the invention can be generated in Chinese hamster ovary cells deficient in DHFR using the same protocol as described above for the anti-B7RP1 monoclonal antibody. The DNA sequences encoding the complete heavy chain or light chain of each anti-B7RP1 antibody of the invention are cloned into expression vectors. CHOd-cells are co-transfected with an expression vector capable of expressing a complete heavy chain and an expression vector expressing the complete light chain of the appropriate anti-B7RP1 antibody. For example, to generate a 5D anti-B7RP1 antibody, cells are co-transfected with a vector capable of expressing a complete heavy chain comprising the amino acid sequence as set forth in SEQ ID NO: 47 and a vector capable of expressing a complete light chain comprising the amino acid sequence set forth in SEQ ID NO: 48. Table 2 summarizes exemplary complete light chains and exemplary complete heavy chains for anti-B7RP1 antibodies having human IgG heavy chain constant regions. One of skill in the art will recognize that the IgG1 or IgG2 could be substituted for each other (*i.e.* where IgG1 is listed in the table, IgG2 could be present, and vice versa). Alternatively, any other immunoglobulin (e.g., IgM, IgA, IgE or IgH) could be used to generate antibodies of the invention.

Table 2

Antibody	Heavy Chain Variable Region + Heavy Chain Constant Region	Complete Heavy Chain
16H(IgG2)	SEQ ID NO: 7 + SEQ ID NO: 41	SEQ ID NO: 44
16H(IgG1)	SEQ ID NO: 7 + SEQ ID NO: 42	SEQ ID NO: 70
16Hg(IgG2)	SEQ ID NO: 8 + SEQ ID NO: 41	SEQ ID NO: 46
16Hg(IgG1)	SEQ ID NO: 8 + SEQ ID NO: 42	SEQ ID NO: 71
5D(IgG1)	SEQ ID NO: 9 + SEQ ID NO: 42	SEQ ID NO: 47
5D(IgG2)	SEQ ID NO: 9 + SEQ ID NO: 41	SEQ ID NO: 72
2H(IgG2)	SEQ ID NO: 10 + SEQ ID NO: 41	SEQ ID NO: 49
2H(IgG1)	SEQ ID NO: 10 + SEQ ID NO: 42	SEQ ID NO: 73
2Hg(IgG2)	SEQ ID NO: 11 + SEQ ID NO: 41	SEQ ID NO: 51
2Hg(IgG1)	SEQ ID NO: 11 + SEQ ID NO: 42	SEQ ID NO: 74
43H(IgG2)	SEQ ID NO: 14 + SEQ ID NO: 41	SEQ ID NO: 52
43H(IgG1)	SEQ ID NO: 14 + SEQ ID NO: 42	SEQ ID NO: 75
41H(IgG2)	SEQ ID NO: 13 + SEQ ID NO: 41	SEQ ID NO: 54
41H(IgG1)	SEQ ID NO: 13 + SEQ ID NO: 42	SEQ ID NO: 76
15H(IgG2)	SEQ ID NO: 12 + SEQ ID NO: 41	SEQ ID NO: 56
15H(IgG1)	SEQ ID NO: 12 + SEQ ID NO: 42	SEQ ID NO: 57

Antibody	Light Chain Variable Region + Light Chain Constant Region	Complete Light Chain
16H	SEQ ID NO: 1 + SEQ ID NO: 43	SEQ ID NO: 45
5D	SEQ ID NO: 2 + SEQ ID NO: 43	SEQ ID NO: 48
2H	SEQ ID NO: 3 + SEQ ID NO: 43	SEQ ID NO: 50
43H	SEQ ID NO: 6 + SEQ ID NO: 43	SEQ ID NO: 53
41H	SEQ ID NO: 5 + SEQ ID NO: 43	SEQ ID NO: 55
15H	SEQ ID NO: 4 + SEQ ID NO: 43	SEQ ID NO: 58

Example 4

Production of anti-B7RP1 Antibody

Anti-B7RP1 antibody is produced by expression in a clonal line of CHO cells. For each production run, cells from a single vial are thawed into serum-free cell culture media. The cells are grown initially in a T-flask followed by spinner flasks and then grown in stainless steel reactors of increasing scale up to a 2000L bioreactor. Production is carried out in a 2000L bioreactor using a fed batch culture, in which a nutrient feed containing concentrated media components is added to maintain cell growth and culture viability. Production lasts for approximately two weeks during which time anti-B7RP1 antibody is constitutively produced by the cells and secreted into the cell culture medium.

The production reactor is controlled at a predetermined pH, temperature, and dissolved oxygen level: pH is controlled by carbon dioxide gas and sodium carbonate addition; dissolved oxygen is controlled by air, nitrogen, and oxygen gas flows.

At the end of production, the cell broth is fed into a disk stack centrifuge and the culture supernatant is separated from the cells. The concentrate is further clarified through a depth filter followed by a 0.2 μm filter. The clarified conditioned media is then concentrated by tangential flow ultrafiltration. The conditioned media is concentrated 15- to 30- fold. The resulting concentrated conditioned medium is then either processed through purification or frozen for purification at a later date.

Example 5

Germlining the 16H mAb

Sequence alignment of the 16H antibody with human germline sequences showed that the framework sequence in the variable region of the 16H antibody was most identical to the V_H 3-07 and JH4 germline sequences, with only three amino acid differences (**Figure** 1A). The framework sequence for the VK region of the 16H antibody was found to be identical to the VK1-L15 germline sequence. It is theoretically possible that somatic hypermutations are recognized as foreign by the immune response of a patient; in which case the patient would generate an anti-idiotype response that could neutralize the therapeutic. To reduce this possibility, the three amino acid changes in the VH framework region were converted back to the V_H 3-07 and JH4 germline sequences (**Figure** 1A). Since the germline V_H and J_H gene segments are present in every human genome, the germline version of 16H is not likely to be recognized as foreign by the immune response of a dosed patient. Plate co-stimulation bioassays were conducted to determine if the germlined antibodies could induce T-cell proliferation with an IC_{50} similar to the IC_{50} of the non-germlined antibodies. The co-stimulation assays were conducted as described below using anti-CD3 and hB7RP-1-Fc fusion protein confirmed that this germlined antibody, referred to as 16Hgermline or 16Hg, retains its biological activities (**Figure** 1B).

Example 6

Affinity Measurement of Monoclonal Antibodies by Biacore® and KinExA

Three antibodies (5D and 16H, prepared as described in Example 1, and 16H germline, prepared as described in Example 5) were purified and submitted to binding

affinity analysis. B7RP1-Fc was immobilized at a high density on a CM5 sensor chip using standard amine coupling chemistry. A fixed concentration of mAb was then incubated with varying concentrations of B7RP-1 or B7RP1-Fc for at least eight hours at room temperature to allow them to reach equilibrium. The samples were then injected over the B7RP1-Fc surface, and the binding signal observed represented free antibody remaining in solution at equilibrium. By using two different antibody concentration (0.2nM and 1nM), the K_D of the interaction between a particular mAb and ligand was calculated from nonlinear regression analysis of the competition curves using a dual-curve one-site homogeneous binding model (Adamczyk *et al.*, 1999, *Bioconjugate Chem.* 10:1032-37; Adamczyk *et al.*, 2000, *Methods* 20:319-28). As shown in Figure 2 and Table 3, the 16H, 16Hg, and 5D mAbs all bound both soluble B7RP-1 and B7RP-1-Fc proteins at high affinities. In addition, the results indicated that the 16H (non-germline) and the 16Hg (germline) reacted similarly, demonstrating that germlining did not significantly affect binding between antibody and ligand.

Table 3

Summary of K_D Values		
	B7RP-1	B7RP1-Fc
5D	37 pM	1.6 pM
16H	1.9 nM	27 pM
16H (germline)	2.7 nM	17 pM

Binding of 5D, 2H, and 2H germline antibodies was also tested using KinExA (kinetic exclusion assay) technology. In this assay, hB7RP-1 was coupled to agarose beads. The beads were used to create a bead column. Samples containing antibody at a fixed concentration, which were allowed to come to equilibrium with varying concentrations of hB7RP-1, were then passed over the bead column. Antibody not complexed with ligand bound to the coated beads.

A fluorescent tagged anti-human Fc secondary antibody was used to detect bound test antibody. The signal obtained was proportional to free antibody in solution at a given ligand concentration. Using two different antibody concentrations, the K_D of the interaction was calculated from nonlinear regression analysis of the competition curves using a dual-curve one-site homogeneous binding model (Adamczyk *et al.*,

1999, *Bioconjugate Chem.* 10:1032-37; Adamczyk *et al.*, 2000, *Methods* 20:319-28). Figures 3, 4, and 5 show the dual-curve fits for antibodies 5D, 2H and 2H(germline). Using this technique, an approximately 10-fold difference was seen in the K_Ds for antibodies 5D and 2H.

The results of the Biacore® and KinExA assays demonstrated that antibody 5D has a higher affinity for hB7RP-1 than do either 2H or 16H. Also, the germline version of antibody 2H does not show a significant difference from the non-germline construct.

Example 7

Functional Characteristics of anti-B7RP1 Antibodies

The functional characteristics of B7RP-1 antibodies of the invention were evaluated using binding-competition assays, *in vitro* co-stimulation assays and *in vitro* tetanus toxoid assays.

Binding-competition studies

Binding-competition studies were conducted with the 16H mAbs to demonstrate that they can compete for ICOS binding for B7RP-1. CHO cells transfected with a gene encoding the full-length human B7RP-1 were first incubated with decreasing amounts of unlabeled 16H mAb and subsequently stained with a fluorescently-labeled ICOS-Fc fusion protein. The cells were then analyzed using flow cytometry. As shown in Figure 6, ICOS-Fc stained the B7RP-1-transfected CHO cells; 0.4μg/ml of 16H mAb did not affect ICOS-Fc binding. However, 6 and 25 μg/ml of 16H efficiently competed away ICOS-Fc binding, indicating that the 16H mAb indeed competed for ICOS binding on B7RP-1.

Co-stimulation Assays

Cell culture plates (Falcon, Cat No.353077, U bottom) were coated with 1 μg/ml anti-human CD3 antibodies (PharMingen Cat No.555336) and 10ug/ml anti-human IgG (Fc specific, Sigma Cat No.I3391). The anti-CD3 antibodies and anti-

human immunoglobulin in phosphate buffered saline (PBS) were added to each well (100µl/well). The coated plates were incubated at 4°C overnight or at room temperature for 2 hours. The plates were then washed with PBS twice. After washing, 1µg/ml human B7-2Fc (R&D System, Cat No.141-B2) or 5µg/ml hB7RP1Fc, each diluted in PBS, were added to each well (100µl per well). The plates were then incubated at room temperature for 3 hours and washed twice with PBS thereafter. Purified human T cells were added ($1x10^5$ per well) in 200µl volume of media (RPMI 1640 supplemented with 10% fetal calf serum (FCS), penicillin-streptomycin-L-glutamine (PSG), β-mercpatoethanol (2-ME), N-Acetyl aspartate (NAA) and Napyruvate) and incubated at 37°C, 5% CO_2 for 48 hours. ^{3}H thymidine (ICN Cat No.2404205) was added at 1µCi/well and the cells were incubated overnight at 37°C, 5% CO_2. The cells were then harvested and counted.

Cell culture plates (Falcon, Cat No.353077, U bottom) were coated with 0.1 µg/ml anti-human CD3 as above. hB7RP1 transfected CHO cells (5000RAD irradiated) were added at $2x10^4$ per well followed by purified human T cells at $1x10^5$ per well in 200µl volume. Plates were incubated at 37°C, 5% CO_2 for 48 hours as above. ^{3}H thymidine was added at 1µCi/well. Cells were incubated overnight, harvested, and counted as above.

Tetanus Toxoid Assays

PBMC were purified from human blood using a Ficoll-Paque (Amersham Biosciences) gradient as follows. Blood was diluted 1:2 with PBS, diluted blood was layered on top of the Ficoll (1/3 room temp Ficoll + 2/3 diluted blood), centrifuged at 2500 rpm for 30 minutes at room temperature, the top layer (plasma & platelets) was aspirated off, and the mononuclear cell layer was transferred to a fresh 50 ml tube. The isolated PBMC were washed with PBS (3x the volume of the mononuclear cell layer) and centrifuged for 10 minutes at 1300 rpm at room temperature and washed as above. The PBMC were resuspended in media (RPMI 1640 + 10% heat-inactivated FBS + 1X PSG + 1X NEAA + 55 µM 2-ME) and the cells were counted.

PMBC were added to wells of a 96-well round bottom plate at 100 µl PBMC/well ($3x10^6$/ml). Tetanus toxoid (20 µg/ml; University of Massachusetts) was added for a final concentration of 5 µg/ml. The cells were incubated for 3 days at

37°C; 100 µl supernatant were collected and incubated for an addition 6 to 8 hours in the presence of 1 µCi/well ^{3}H-thymidine (MP Biomedicals). The cells were then harvested and counted.

Table 4 summarizes the functional characteristics of certain antibodies of the invention as determined using the assays described above.

Table 4

	Plate	Biacore Fc	Biacore mono	CHO	Tetanus Toxoid
2H	43	89		1445	15
15H	36				141
16H	53	27	1900	276	27
16Hg	32	17	2700	523	
41H	52				115
43H	46				35
5D	55	1.6	37	1456	15
ICOS-Fc	200-1000	1000		10,672	
α-CD86					40

*EC_{50}/KD values in pM

Example 8

Epitope Mapping

Experiments were conducted to identify the region on B7RP-1 to which the 16H/16Hg and 5D monoclonal antibodies bind. To do this, a novel Fluorescence-Activated Cell Sorter (FACS) binding assay was developed. The human extracellular domain (ECD) of B7RP1 (SEQ ID NO: 66) as well as truncated forms of B7RP-1 containing either the Ig1 (IgV-like; SEQ ID NO: 67) or the Ig2 (IgC-like; SEQ ID NO: 68) were expressed as N-terminal, in-frame fusions with chicken avidin.

SEQ ID NO: 66 (ECD):

DTQEKEVRAMVGSDVELSCACPEGSRFDLNDVYVYWQTSESKTVVTYHIPQNSSLENVDSRYRNRALMS
PAGMLRGDFSLRLFNVTPQDEQKFHCLVLSQSLGFQEVLSVEVTLHVAANFSVPVVSAPHSPSQDELTF
TCTSINGYPRPNVYWINKTDNSLLDQALQNDTVFLNMRGLYDVVSVLRIARTPSVNIGCCIENVLLQQN
LTVGSQTGNDIGERDKITENP

SEQ ID NO: 67 (IgV-like):

DTQEKEVRAMVGSDVELSCACPEGSRFDLNDVYVYWQTSESKTVVTYHIPQNSSLENVDSRYRNRALMS
PAGMLRGDFSLRLFNVTPQDEQKFHCLVLSQSLGFQEVLSVEVTLHVAANFSVPVVSAPHSPSQDELTF
T

SEQ ID NO: 68 (IgC-like):

LGFQEVLSVEVTLHVAANFSVPVVSAPHSPSQDELTFTCTSINGYPRPNVYWINKTDNSLLDQALQNDT
VFLNMRGLYDVVSVLRIARTPSVNIGCCIENVLLQQNLTVGSQTGNDIGERDKITENP

Expression vectors containing genes encoding these fusion proteins were individually transiently transfected into 293T cells and the conditioned media from these cell lines were used as the source of fusion protein. The avidin-tag was used to capture the B7RP1 fusion proteins from solution using a biotin-coated bead. Fusion proteins were incubated with either fluorescently-labeled 16H or 5D mAbs or a fluorescently-labeled ICOS-Fc fusion protein, and incubated with biotin-coated beads. The beads were recovered and analyzed using flow cytometry on a Becton-Dickinson Bioscience FACScan (BD, Franklin Lakes, NJ). As shown in **Figure 9A**, fluorescent staining of the beads was detected with the 16H, 5D, and the ICOS reagents when the full ECD of B7RP-1 was attached, indicating that all three of these reagents could bind to the ECD of B7RP-1. Similarly, all three reagents bound to the avidin fusion protein containing only the Ig1 domain, indicating that both ICOS and the blocking anti-B7RP-1 mAbs could bind to this region. In contrast, neither ICOS nor the anti-B7RP-1 mAbs could bind to the fusion protein containing only the membrane-proximal Ig2 domain. Thus, the ICOS, 16H, and 5D binding regions on B7RP-1 were located in the Ig1 domain.

The antibodies generated as described above in Example 1 and tested for binding using the avidin fusion binding assay, could be divided into two epitope classes, H and D, as shown in Table 5. Of the 100 antibodies initially selected based on their ability to bind B7RP1, 15 failed to bind in the avidin fusion binding assay, most likely because of degradation.

Table 5

Classification of mAbs by epitope	
Class	#
H epitope	75
D epitope	10
New epitope, ICOS blocker	0

New epitope, not an ICOS blocker	0
No detectable binding	15

Example 9

SNP identification and functional analysis

One major single nucleotide polymorphism (SNP) variant was identified in B7RP-1 that is present in the population with an allele frequency of 28.4% (**Figure 7**). The variant was identified within the mature protein coding sequence. A search of the National Center for Biotechnology Information (NCBI) databank revealed a second potential SNP variant; the second variant was identified in a 1.5 individual (three chromosome) analysis. The first SNP variant (V128I) was located in the first IgV-like domain, whereas the NCBI SNP variant (L221F) was located in the second IgC-like domain.

As discussed above, both the 16H and 5D monoclonal antibodies bind to the first IgV-like domain, this it is unlikely that the latter L221F variant affects either 16H or 5D mAb binding or function. Nonetheless, to determine if either of these SNP variants affects 16H or 5D binding and/or function, two different experiments were conducted. In the first set of experiments, avidin fusion proteins were constructed with the two SNP variants and tested for binding to 16H or 5D antibodies in the flow cytometric assay as described above. These representative mAbs from the H and D epitope classes bound to the SNP variants with similar efficacy as the wild-type B7RP-1 (**Figure 9B**). These data suggested that antibodies from both the H and D epitope classes bind to the B7RP-1 SNP variants.

In the second approach, Fc fusion proteins were constructed using the B7RP-1 SNP variant sequences and compared for the ability of these proteins to stimulate T cells in the plate co-stimulation assay (**Figure 9C**). Both the 16H and 5D antibodies inhibited co-stimulation mediated by the SNP variant Fc fusion proteins with similar EC_{50}s as the wild-type fusion protein. Taken together these data indicated that the two potential B7RP-1 SNP variants were recognized by the antibodies of the invention. Thus, the antibodies of the invention can bind to target in patients containing these SNP variants.

Example 10

In vivo Animal Efficacy Models

The ability of B7RP-1 antibodies to inhibit immune response was analyzed using a murinized rat anti-murine B7RP-1 monoclonal antibody (1B7v2) and challenging BALB/c mice with keyhole-limpet hemocyanin (KLH).

Generation of the murinized rat anti-murine B7RP-1 monoclonal antibody 1B7v2

A Chinese-Hamster-Ovary cell line that overexpressed a full-length murine B7RP-1 was injected into rats as a primary immunization, and subsequently with a murine B7RP-1-Fc fusion protein to boost the immune response. Spleens were harvested 3 or 4 days post-intravenous boost and the splenic B cells fused with the Y3-Ag1.2.3 rat myeloma line (ATCC CRL-1631). Cells were then selected in media supplemented with hypoxanthine-aminopterin-thymidine (HAT) for 2 weeks and subsequently single-cell subcloned by limiting dilution. These procedures are described in "Practical Immunology, 2nd ed." Leslie Hudson and Frank C. Hay; Blackwell Scientific Publications 1980.

Genes encoding the 1B7 immunoglobulin were cloned from the 1B7 cell line using standard procedures (Sambrook *et al.*, 2001, MOLECULAR CLONING: A LABORATORY MANUAL, 3d ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y). The isotype switch of the human anti-huB7RP1 MAbs was accomplished by cloning the variable region fragments containing *Xba*I and *Bsm*BI restriction site cohesive ends into the pDSRa vector with the human IgG1 or huIgG2 constant region which also had *Xba*I and *Bsm*BI ends. For the 1B7 rat anti-muB7RP1 the chimera was formed by a three step overlapping PCR process. The rat variable region was PCR amplified with a 3' primer that contained part, ~25-35 nucleotides, of the murine constant region. The murine constant region was amplified with a 5' primer that contained part, ~25-35 nucleotides, of the rat variable region. The two fragments were then used as template and the 5' rat variable region (*Xba*I containing) and the murine 3' constant region (*Sal*I containing) primers were used to generate a complete light chain or heavy chain. The light chain and heavy chain PCR products were then digested with *Xba*I and *Sal*I and cloned into pDSRa19. A total of 25µg of

linearized DNA (12.5 μg pDC323B LC + 12.5 μg pDC324 HC) were transfected into CS-9 cells using electroporation and selected on DHFR-supplemented medium.

To test the efficacy of the 1B7v2 mAb, plate co-stimulation assays were conducted with this mAb. The results were compared with other anti-murine B7RP-1 mAbs (**Figure 10A**). As discussed above, 1B7 is the original hybridoma-produced mAb; two different preparations (labeled 1.33 and 7.4) were tested. 5E1 and 11G10 were other anti-mB7RP-1 monoclonals generated in the fusions described above. Finally, HK5.3 was a commercially-available anti-mB7RP-1 (ebiosciences # 16-5985-85).

The 1B7v2 mAb blocked T cell activation in this assay equal to or better than any of the other mAbs, and thus was selected as the surrogate therapeutic for further studies.

Antigen Challenge in Mice

Keyhole Limpet Hemocyanin (KLH) was purchased from Pierce Biotechnology (Rockford, Illinois). Dosing solution #1 (KLH 5mg/kg in 1mg/mouse ALUM) was prepared with equal parts of 2x ALUM (500mg of ALUM plus 50ml PBS (phosphate buffered saline)) and 2x KLH (2.0ml dH20 (RNAse-Free) mixed with 20mg of lyophilized KLH, brought to 20ml with 1x PBS). Dosing solution #2 (KLH 1mg/kg in 1mg/mouse ALUM) was prepared with 1 part 2x KLH mixed with 4 parts 1x phosphate buffered saline.

Female BALB/c mice were primed either with 1 mg/kg of KLH/alum and re-immunized on day 21 with 5mg/kg KLH only, introduced by intraperitoneal injection. Mice were treated by intraperitoneal injection with 1B7v.2, the isotype control antibody (anti-AGP3 PB) or the vehicle (PBS) alone, starting on day 1 (one day prior to priming with KLH/alum) in a final volume of 200μl every 5 days.

The mice were bled every 7 days retro-orbitally (approximately 200μl) to obtain approximately 50-100μl of serum for analysis of antigen-specific serum IgM (**Figure 10B**), IgG2a (**Figure 10C**), and IgG1 (**Figure 10D**). Both the isotype-control and vehicle-treated mice showed significant primary and secondary immune responses. The IgM response was not affected by treatment, whereas, blockade of

B7RP-1-ICOS with 1B7v2 decreased both primary and secondary IgG2a and IgG1 responses in a statistically-significant manner.

IL-5 is a cytokine released by T cells in response to antigen stimulation that induces B cell differentiation and function. As the B7RP-1/ICOS interaction is believed to be critical for T-cell-dependent B cell function, measuring serum IL-5 levels was used to determine if interdiction of the B7RP-1/ICOS axis was indeed affecting T cell function. As expected, blockade of B7RP-1 also inhibited antigen-induced serum IL-5 levels. Sera were harvested from the mice from the antigen challenge experiment outlined above 24 hours after the antigen challenge on day 21, and serum IL-5 levels were determined by ELISA. As shown in **Figure 11**, elevated IL-5 levels were detected in the test mice as early as 9 hours after challenge; levels began to decline by 48 hours and returned to baseline by 72 hours. Treatment of the mice with 1B7v2 mAb lead to a statistically significant repression of IL-5 levels at the 24-hour time point.

Example 11

Binding to cynomolgus monkey B7RP-1

To determine if the anti-hB7RP-1 mAbs also bind to cynomolgus monkey B7RP-1, flow cytometric staining experiments were conducted with the 16H mAb and B cells purified from cynomolgus monkeys and humans. As shown in **Figure 12A**, addition of fluorescently-labeled 16H to cyno B cells lead to staining, indicating that 16H was indeed binding to cyno B7RP-1 (right panel). As expected, 16H also stained human B cells (left panel). In addition, 16H, 16Hg, and 5D were tested in plate co-stimulation assays using cyno T cells, cyno B7RP-1-Fc, and anti-CD3 mAb. As shown in **Figure 12B**, all three mAbs inhibited cyno B7RP-1-dependent cyno T cell activation, indicating that these mAbs functionally block the cyno ICOS-B7RP-1 interaction.

Example 12

T-Cell Dependent Antigen Responses in the Cynomolgus Monkey Following Administration of the Anti-B7RP-1 Antibodies

A cynomolgus monkey study was conducted with two anti-B7RP-1 monoclonal antibodies, 16H and 5D, to assess the ability of these antibodies to inhibit a T-cell dependent B cell antigen response as determined by serum levels of antigen-specific antibody. Briefly, the anti-keyhole limpet hemocyanin (KLH) and anti-tetanus toxoid antibody responses were examined following antigen challenge in the presence of B7RP-1 antibodies in the cynomolgus monkey.

Test Article 1 was 16H and Test Article 2 was 5D. The Control Article was the vehicle for B7RP-1 antibody (0.01 sodium acetate, pH 5.0, 5% sorbitol, 0.004% Tween 20). Keyhole Limpet Hemocyanin (KLH) was purchased from Pierce Biotechnology (Rockford, Illinois).

The KLH was prepared by reconstitution with sterile water to yield a 10 mg/mL stock solution. The stock solution was diluted with sterile water to yield a 1 mg/mL dosing solution. Tetanus Toxoid used for these experiments was Super-Tet® Tetanus Toxoid w/Havlogen®, purchased from Intervet™ Inc. (Milsboro, Delaware). The dose level for these experiments was 75 IU (0.5 mL of 150 IU/mL).

Table 6 shows the treatment group distribution of 28 cynomolgus monkeys.

Table 6

Group No.	Number of Males/Females	Test Article	Route	Dose Level (mg/kg)	Dose Volume (mL/kg)	Dose Solution Conc. (mg/mL)
1	2/2	Control	IV	0	1	0
2	2/2	B7RP-1 5D	IV	0.1	1	0.1
3	2/2	B7RP-1 5D	IV	1.0	1	1.0
4	2/2	B7RP-1 5D	IV	8.0	1	10.0
5	2/2	B7RP-1 16H	IV	0.1	1	0.1
6	2/2	B7RP-1 16H	IV	1.0	1	1.0
7	2/2	B7RP-1 16H	IV	8.0	1	10.0

Test article doses were administered via intravenous injection to all animals on Days 1, 8, 15, 22, 29, 36, 43, and 50. Animals scheduled for necropsy in Groups 1-4 (1/sex/group) received an additional dose on Day 57. Evaluation of immune response was conducted on all animals via immunization with KLH and tetanus toxoid antigens followed by blood sampling for antigen-specific immunoglobulins (IgM and IgG).

Titer values were present following primary administration of both the KLH and tetanus antigens. For KLH, primary titer values ranged from 0 to 900 for both IgM and IgG. As the primary KLH challenge was administered prior to test article

administration, no effect of the B7RP-1 antibodies was evaluated. For tetanus toxoid, primary titer values ranged from 0 to 50 for IgM and from 0 to 4050 for IgG. There were no differences in the primary response to tetanus toxoid between the B7RP-1 antibody groups and the control group.

As expected, titer values for IgG were increased following secondary administration of both the KLH and tetanus antigens, when compared to the primary titer values. For KLH, secondary titer values ranged from 0 to 300 for IgM and from 0 to 8100 for IgG. However, there was no evidence of inhibition of the KLH secondary response attributed to administration of the B7RP-1 antibodies.

For tetanus toxoid, secondary titer values were below 50 for IgM and ranged from 1350 to 36450 for IgG. Results for individual animal and group mean values are presented in the **Figure 13A** (16H antibody) and **Figure 13B** (5D antibody) for Days 53 and 57 following the secondary challenge with tetanus toxoid on Day 42.

On Day 53, the number of animals reaching peak response was 3/4, 1/4, and 1/4 at the 0.1, 1, and 8 mg/kg dose levels of 16H, respectively, and 1/4, 1/4, and 1/4 at the 0.1, 1, and 8 mg/kg dose levels of 5D respectively, compared to 2/4 control animals. Thus, in general, the number of animals reaching a high titer on Day 53 was reduced in the B7RP-1 antibody-treated groups. On Day 57, titer values were maintained in the control animals, while titer values for several of the B7RP-1 antibody-treated animals declined from the Day 53 values. The number of animals with high titers on Day 57 was 0/4, 0/4, and 1/4 at the 0.1, 1, and 8 mg/kg dose levels of 16H, respectively, and 0/4, 0/4, and 0/4 at the 0.1, 1, and 8 mg/kg dose levels of 5D respectively, compared to 2/4 control animals.

These results demonstrated that the two B7RP-1 antibodies 16H and 5D inhibited a T-cell dependent B cell antigen response in cynomolgus monkeys, as determined by serum levels of tetanus toxoid-specific antibody. In addition, the presence of the B7RP-1 antibodies was important for blockage of the B7RP-1-ICOS interaction during the primary response in order to detect an effect following the secondary challenge.

These results and the results from Example 10 demonstrated that both the surrogate therapeutic and the therapeutic candidates blocked T and B cell- dependent immune responses in murine and monkey model systems, which indicated that

blocking this co-stimulatory axis may be efficacious in the treatment of B-cell-mediated diseases such as Systemic Lupus Erythematosus (SLE), asthma, and Rheumatoid Arthritis (RA).

It should be understood that the foregoing disclosure emphasizes certain specific embodiments of the invention and that all modifications or alternatives equivalent thereto are within the spirit and scope of the invention as set forth in the appended claims.

We claim:

1. An isolated antibody that binds specifically to B7RP1, wherein the antibody comprises a heavy chain having a heavy chain variable region and a light chain having a light chain variable region, wherein the heavy chain variable region comprises an amino acid sequence that has at least 98% sequence identity to the amino acid sequence set forth in SEQ ID NO: 9, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof, and the light chain variable region comprises an amino acid sequence that has at least 98% sequence identity to the amino acid sequence set forth in SEQ ID NO: 2, or an antigen-binding or immunogenic fragment thereof.
2. The antibody of claim 1, wherein the antibody binds specifically to human B7RP1 but not to mouse B7RP1.
3. The antibody of claim 1, wherein the antibody binds specifically to human B7RP1 and mouse B7RP1.
4. The antibody of claim 1, wherein the heavy chain variable region comprises an amino acid sequence as set forth in SEQ ID NO: 9, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof.
5. The antibody of claim 1, wherein the heavy chain comprises a human heavy chain CDR1 that has an amino acid sequence as set forth in SEQ ID NO: 30, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof.
6. The antibody of claim 1, wherein the heavy chain comprises a human heavy chain CDR3 that has an amino acid sequence as set forth in SEQ ID NO: 32, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof.
7. The antibody of claim 1 wherein the heavy chain comprises a human heavy chain CDR2 that has an amino acid sequence as set forth in SEQ ID NO: 31, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof.
8. The antibody of claim 1 comprising a heavy chain having a heavy chain variable region comprising an amino acid sequence as set forth in SEQ ID NO: 9, an antigen-binding or an immunologically functional immunoglobulin

fragment thereof, and a light chain having a light chain variable region comprising an amino acid sequence as set forth in SEQ ID NO: 2, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof.

9. The antibody of claim 1, wherein the light chain variable region comprises an amino acid sequence as set forth in SEQ ID NO: 2, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof.

10. The antibody of claim 1, wherein the light chain comprises a human light chain CDR1 that has an amino acid sequence as set forth in SEQ ID NO: 18, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof.

11. The antibody of claim 1, wherein the light chain comprises a human light chain CDR3 that has an amino acid sequence as set forth in SEQ ID NO: 20, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof.

12. The antibody of claim 1, wherein the light chain comprises a human light chain CDR2 that has an amino acid sequence as set forth in SEQ ID NO: 19, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof.

13. The antibody of claim 1 wherein the heavy chain comprises a CDR1 of SEQ ID NO: 30, a CDR2 of SEQ ID NO: 31 and a CDR3 of SEQ ID NO: 32.

14. The antibody of claim 1 wherein the light chain comprises a CDR1 of SEQ ID NO: 18, a CDR2 of SEQ ID NO: 19 and a CDR3 of SEQ ID NO: 20.

15. The antibody of claim 1, wherein the heavy chain and light chain comprise a single-chain antibody.

16. The antibody of claim 15, which is a single-chain Fv antibody.

17. The antibody of claim 1or 35, which is a Fab antibody.

18. The antibody of claim 1or 35, which is Fab' antibody.

19. The antibody of claim 1or 35, which is a $(Fab')_2$ antibody.

20. The antibody of claim 1or 35, wherein the antibody is a fully human antibody.

21. The antibody of claim 1or 35, wherein the antibody inhibits B7RP1 activity.

22. Use of a therapeutically effective amount of the antibody of any one of claims 1 to 14, 35, 36 or 37, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof, in the manufacture of a medicament for treating an autoimmune disease or inflammatory response in a patient.

23. The use of claim 22, wherein the autoimmune disease is rheumatoid arthritis, or systemic Lupus erythematosus.

24. The use of claim 22, wherein the inflammatory response is asthma.

25. Use of a therapeutically effective amount of the antibody of any one of claims 1 to 14, 35, 36 or 37, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof, in the manufacture of a medicament for inhibiting co-stimulatory signal T-cell activation in a patient.

26. A pharmaceutical composition comprising a pharmaceutically acceptable carrier and a therapeutically effective amount of the antibody of any one of claims 1 to 14, 35, 36 or 37, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof.

27. Use of the pharmaceutical composition of claim 26 in the manufacture of a medicament for treating an autoimmune disease or inflammatory response in a patient.

28. A method for detecting B7RP1 in a biological sample comprising:

a) contacting the sample with the antibody of claim 1, under conditions that allow for binding of the antibody to B7RP1; and

b) measuring the level of bound antibody in the sample.

29. A nucleic acid molecule that encodes the antibody of claim 1 to 14, 35, 36 or 37.

30. A vector comprising the nucleic acid molecule of claim 29.

31. A host cell comprising the nucleic acid of claim 29 or the vector of claim 30.

32. A host cell comprising a first vector and a second vector, wherein the first vector comprises a nucleic acid molecule encoding an amino acid sequence that has at least 98% sequence identity to the amino acid sequence set forth in SEQ ID NO: 9, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof, and wherein the second vector comprises a

nucleic acid molecule encoding an amino acid sequence that has at least 98% sequence identity to the amino acid sequence set forth in SEQ ID NO: 2, or an antigen-binding or immunogenic fragment thereof.

33. An isolated cell line that produces an antibody according to any of claims 1 to 14, 35, 36 or 37.

34. A method of producing an antibody comprising culturing the host cell of claim 31 or 32, or the cell line of claim 33, and collecting the antibody from the culture medium or the host cell.

35. The antibody of claim 1 comprising amino acid sequences set forth in SEQ ID NO: 18, 19, 20, 30, 31 and 32.

36. The antibody of claim 1 or 35, that binds specifically to:

a) the D region of B7RP1, or

b) the H region of B7RP1.

37. The antibody of claim 1 or 35, wherein the antibody comprises an amino acid sequence as set forth in any of SEQ ID NO: 47or 72, and an amino acid sequence as set forth in SEQ ID NO: 48, or an antigen-binding or an immunologically functional immunoglobulin fragment thereof.

38. Use of the pharmaceutical composition of claim 26 in the manufacture of a medicament for treating a condition caused by increased expression of B7RP1 or increased sensitivity to B7RP1 in a patient.