COMBINATION OF BLyS AND/OR APRIL INHIBITION AND IMMUNNOSUPPRESSANTS FOR TREATMENT OF AUTOIMMUNE DISEASE

FIELD OF THE INVENTION

[1] The invention relates to novel combination therapies involving BLyS or BLyS/APRIL inhibition and immunosuppressants for the treatment of autoimmune diseases.

BACKGROUND OF THE INVENTION

[2] Lymphocytes are one of several populations of white blood cells; they specifically recognize and respond to foreign antigen. The three major classes of lymphocytes are B lymphocytes (B cells), T lymphocytes (T cells) and natural killer (NK) cells. B lymphocytes are the cells responsible for antibody production and provide humoral immunity. B cells mature within the bone marrow and leave the marrow expressing an antigen-binding antibody on their cell surface. When a naive B cell first encounters the antigen for which its membrane-bound antibody is specific, the cell begins to divide rapidly and its progeny differentiate into memory B cells and effector cells called plasma cells. Memory B cells have a longer life span and continue to express membrane-bound antibody with the same specificity as the original parent cell. Plasma cells do not produce membrane-bound antibody but instead produce secreted form of the antibody. Secreted antibodies are the major effector molecules of humoral immunity.

[3] A group of tumor necrosis factor (TNF) receptors found on the surface of B cells under various conditions are among the cellular regulators of B cell function in the immune system. In particular, three TNF receptors: transmembrane activator and CAML interactor (TACI), B cell activator belonging to the TNF family receptor (BAFF-R), and B cell maturation protein (BCMA) are known to bind one or both TNF ligands – B Lymphocyte stimulator (BLyS also known as BAFF, TALL-1, ztnf4 and THANK) and a proliferation-inducing ligand (APRIL). Specifically, TACI and BCMA are known to bind both BLyS and APRIL and BAFF-R binds only BLyS.

[4] A number of BLyS and/or APRIL antagonists have been developed in order to block the various functions of BLyS, which include but should not be limited to B cell co-stimulation, plasmablast and plasma cell survival, Ig class switching, enhanced B-cell antigen presenting cell function, survival of malignant B cells, development of B-1 cell function, B

cell development beyond the T-1 stage, and complete germinal centre formation Some of these molecules can also bind to and block the effect of APRIL on B cells and other components of the immune system (Dillon et al. (2006) Nat. Rev. Drug Dis. 5, 235-246). Molecules that have been developed to affect B cell function by interfering with BLyS and/or APRIL binding include BLyS antibodies such as Lymphostat-B (Belimumab) (Baker et al, (2003) Arthritis Rheum, 48, 3253-3265 and WO 02/02641); receptor-extracellular domain/Fc domain fusions proteins such as TACI-Ig, including one particular embodiment, atacicept (U.S. Patent Application No. 20060034852), BAFF-R-Fc (WO 05/0000351), and BCMA-Ig or other fusion proteins utilizing receptor extracellular domains. A further class of BLyS and/or APRIL antagonists include other molecules relying on BLyS binding ability to block binding to its receptors such as AMG 623, receptor antibodies, and other molecules disclosed in WO 03/035846 and WO 02/16312

[5] The current approach for the treatment of autoimmune diseases is suppression of the unwanted immune reaction. For example, in the treatment of lupus nephritis (LN), a serious complication involving the kidney of patients suffering from systemic lupus nephritis (SLE), several immunosuppressive drugs have proven beneficial. These drugs include cyclophosphamide (CYC), azathioprine (AZA), cyclosporine A (CSA), and mycophenolate mofetil (MMF) (for general reviews see, Mok et al. (2003) Ann Rheum Dis 62, 799-804 and Iaccarino et al. (2007) Autoimmunity Reviews 6, 190-195). Although these drugs are beneficial, there remains a need in the art to improve the response to immune suppressive drugs in order to effectively treat autoimmune disease.

SUMMARY OF THE INVENTION

[6] One aspect of the present invention is a method of reducing immunoglobulin levels in a mammal comprising administering a BLyS and/or APRIL antagonist and an immunosuppressive drug. One preferred method is where the BLyS and/or APRIL antagonist is a Fc-fusion protein which can be a TACI-Fc-fusion protein comprising the extracellular domain of TACI or a functional fragment thereof, a BAFF-R-Fc-fusion protein comprising the extracellular domain of BAFF-R or a functional fragment thereof, or a BCMA-Fc-fusion protein comprising the extracellular domain of BCMA or a functional fragment thereof. In particular, the Fc-fusion protein comprises the polypeptide sequences of SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, or SEQ ID NO: 26.

[7] In another embodiment, the BLyS and/or APRIL antagonist is a BLyS antibody, preferably that binds BLyS within a region comprising amino acids 162-275 of SEQ ID NO: 8, or the BLyS antibody known as LymphoStat-B. In further embodiment, the BLyS and/or APRIL antagonist is a TACI antibody, preferably that binds TACI within a region comprising 72-109 of SEQ ID NO:2 or 82-222 of SEQ ID NO:2.

[8] In the methods of the present invention some of the immunosuppressive drugs contemplated include cyclophosphamide (CYC), azathioprine (AZA), cyclosporine A (CSA), or mycophenolate mofetil (MMF), although any drug that suppresses the immune system may be suitable. The immunoglobulin level that is reduced by the present method can be IgM, IgG, IgA, IgD or IgE or combinations thereof

[9] The present invention also encompasses a method of alleviating a B-cell regulated autoimmune disorders comprising administering to a patient suffering from the disorder, a therapeutically effective amount of an immunosuppressive drug and of a BLyS and/or APRIL antagonist. In one embodiment, the autoimmune disorder is selected from the group consisting of rheumatoid arthritis, juvenile rheumatoid arthritis, systemic lupus erythematosus (SLE), lupus nephritis (LN), Wegener's disease, inflammatory bowel disease, idiopathic thrombocytopenic purpura (ITP), thrombotic throbocytopenic purpura (TTP), autoimmune thrombocytopenia, multiple sclerosis, psoriasis, IgA nephropathy, IgM polyneuropathies, myasthenia gravis, vasculitis, diabetes mellitus, Reynaud's syndrome, Sjorgen's syndrome and glomerulonephritis. One disease specifically treated in this manner is lupus nephritis.

[10] Particularly when the autoimmune disorder is rheumatoid arthritis, systemic lupus erythematosus, or lupus nephritis, in one embodiment, the BLyS and/or APRIL antagonist and the immunosuppressive drug can be administered in further conjunction with therapy using a second immunosuppressive drug such as nonsteroidal anti-inflammatory drugs (NSAIDs), glucocorticoid, prednisone, and disease-modifying antirheumatic drug (DMARD).

[11] The methods of the present invention utilize immunosuppressive drugs which have been shown to be effective in treating autoimmune disease in combination with the BLyS and/or APRIL antagonist. Among the immunosuppressive drugs contemplated for use in the methods of the present invention are cyclophosphamide (CYC), azathioprine (AZA), cyclosporine A (CSA), and mycophenolate mofetil (MMF).

[12] In any of the methods of treatment or alleviation of a disorder where the immunosuppressive drug and the BLyS and/or APRIL antagonist are administered to a patient, the immunosuppressive drug and BLyS and/or APRIL antagonist can be administered concurrently or sequentially. In a specific embodiment, the immunosuppressive agent is administered before BLyS and/or APRIL antagonist

[13] A composition comprising an immunosuppressive agent and a BLyS and/or APRIL antagonist is also provided.

[14] Further provided by the invention is an article of manufacture comprising an immunosuppressive agent, a BLyS and/or APRIL antagonist, and a label wherein the label indicates that the composition is for treating a B cell regulated autoimmune disorder.

[15] In any of the embodiments of the methods, compositions and articles of manufacture of the invention, the BLyS and/or APRIL antagonist, if an antibody, includes chimeric and humanized antibodies.

[16] In any of the embodiments of the methods, compositions and articles of manufacture of the invention, the BLyS and/or APRIL antagonist, in one embodiment, is a fusion protein between the extracellular domain of a receptor that binds BLyS and the Fc domain of an immunoglobulin, or an Fc-fusion protein. In specific embodiments, the Fc-fusion protein selected from the group consisting of TACI Fc-fusion protein comprising the extracellular domain of TACI, in particular atacicept, BAFF-R Fc-fusion protein comprising the extracellular domain of BAFF-R, in particular BR3-Ig, and BCMA Fc-fusion protein comprising the extracellular domain of BCMA. In other embodiments, the BLyS and/or APRIL antagonist is a BLyS antibody, in particular, a BLyS antibody that binds BLyS within a region of BLyS comprising residues 162-275, in particular Lymphostat B. In another embodiment, the BLyS and/or APRIL antagonist is a BAFF-R antibody including one that binds in a region comprising residues 23-38 of human BAFF-R. In another embodiment, the BLyS and/or APRIL antagonist is a TACI antibody, a BCMA antibody, or an antibody that binds both molecules as described in U.S. Patent Application No. 2003-0012783.

BRIEF DESCRIPTION OF THE FIGURES

[17] FIGURE 1 graphs the mean value (± standard deviation) of IgG for the four treatment groups – vehicle, MMF only, atacicept only, and atacicept and MMF. All pairwise comparisons of these values were statistically significant ($p < 0.05$) using log-transformed data other than vehicle vs. MMF alone.

[18] FIGURE 2 graphs the mean value (± standard deviation) of IgM for the four treatment groups – vehicle, MMF only, atacicept only, and atacicept and MMF. All pairwise comparisons of these values were statistically significant ($p < 0.05$) using log-transformed data other than vehicle vs. MMF alone.

[19] FIGURE 3 graphs the mean value (± standard deviation) of IgA for the four treatment groups – vehicle, MMF only, atacicept only, and atacicept and MMF. All pairwise comparisons of these values were statistically significant ($p < 0.05$) using log-transformed data other than vehicle vs. MMF alone.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

[20] While immunosuppressive treatment appears useful in the treatment of autoimmune disease, it was discovered from the experiments described herein that administration of a combination of immunosuppressives with a BLyS and/or APRIL antagonist is a method of treatment that will block multiple signal pathways in B cells believed responsible for the production of antibodies directed at self-antigens, thereby triggering and/or perpetuating the autoimmune condition. This results in a reduction of circulating immunoglobulin in a mammal undergoing such treatment. As such circulating immunoglobulin is believed at least partially responsible for triggering the negative symptoms of autoimmune disease, the combination of immunesuppressive drugs and therapies directed against the BLyS pathway therefore provides a novel method of treating B cell-mediated diseases such as B cell-based autoimmune diseases. The combination therapy of immunesuppressive drugs with a BLyS and/or APRIL antagonist may offer more effective alternatives to existing treatments for certain diseases, e. g., lupus nephritis.

[21] An "autoimmune disease" herein is any non-malignant disease or disorder arising from antibodies that are produced directed against an individual's own (self) antigens and/or tissues.

[22] As used herein, "B cell depletion" refers to a reduction in B cell levels in an animal or human after drug or antibody treatment, as compared to the level before treatment. B cell levels are measurable using well known assays such as by getting a complete blood count, by FACS analysis staining for known B cell markers, and by methods such as described in the Experimental Examples. B cell depletion can be partial or complete. In a

patient receiving a B cell depleting drug, B cells are generally depleted for the duration of time when the drug is circulating in the patient's body and the time for recovery of B cells.

[23] "Immunosuppressive drugs" are any molecules that interfere with the immune system and blunt its response to foreign or self antigens. Cyclophosphamide (CYC) and mycophenolate mofetil (MMF) are two such kinds of molecules. This term is intended to encompass any drug or molecule useful as a therapeutic agent in downregulating the immune system. This method particularly contemplates drugs that have been used to treat autoimmune diseases such as rheumatoid arthritis, juvenile rheumatoid arthritis, systemic lupus erythematosus (SLE), lupus nephritis (LN), Wegener's disease, inflammatory bowel disease, idiopathic thrombocytopenic purpura (ITP), thrombotic throbocytopenic purpura (TTP), autoimmune thrombocytopenia, multiple sclerosis, psoriasis, IgA nephropathy, IgM polyneuropathies, myasthenia gravis, vasculitis, diabetes mellitus, Reynaud's syndrome, Sjorgen's syndrome and glomerulonephritis.

[24] The terms "BLyS" or "BLyS polypeptide," "TALL-1" or "TALL-1 polypeptide," or "BAFF" or "BAFF polypeptide" when used herein encompass "native sequence BLyS polypeptides" and "BLyS variants." "BLyS" is a designation given to those polypeptides which are encoded by the Human BLyS sequence (SEQ ID NO: 7) or the Mouse BLyS sequence (SEQ ID NO: 9). Polypeptides which show BLyS biological activity are encompassed within this designation as well. For example, a biologically active BLyS potentiates any one or combination of the following events in vitro or in vivo : an increased survival of B cells, an increased level of IgGand/or IgM, an increased numbers of plasma cells, and processing of NF- Kb2/100 to p52NF-Kb in splenic B cells (e.g., Batten, M et al. , (2000) J. Exp. Med. 192: 1453-1465; Moore, et al. , (1999) Science 285: 260-263; Kayagaki, et al. , (2002) 10: 515-524). Several assays useful for testing BLyS and/or APRIL antagonists such as the B cell proliferation assay described in WO 00/40716 among others are well known to one of ordinary skill in the art.

[25] Briefly, human B cells are isolated from peripheral blood mononuclear cells using CD19 magnetic beads and the VarioMacs magnetic separation system (Miltenyi Biotec Auburn, CA) according to the manufacturer's instructions. Purified B cells are mixed with soluble BLyS (25 ng/ml) and recombinant human IL-4 (10 ng/ml Pharmingen), and the cells are plated onto round bottom 96 well plates at 1 x 10^5 cells per well. The BLyS and/or APRIL antagonist to be tested can be diluted from about 5 μg/ml to about 6 ng/ml, and incubated with the B cells for five days, pulsing overnight on day four with 1μCi ^{3}H-

thymidine per well. As a control, BLyS and/or APRIL antagonist can also be incubated with B cells and IL-4 without BLyS. Plates are harvested using Packard plate harvester, and counted using the Packard reader.

[26] A "native sequence" polypeptide comprises a polypeptide having the same amino acid sequence as the corresponding polypeptide derived from nature. Such native sequence polypeptides can be isolated from nature or can be produced by recombinant and/or synthetic means. The term "native sequence" specifically encompasses naturally-occurring truncated, soluble or secreted forms (e. g., an extracellular domain sequence), naturally-occurring variant forms (e. g., alternatively spliced forms) and naturally-occurring allelic variants of the polypeptide.

[27] In general, "variant" polypeptides for any of the polypeptides disclosed in the present specification include polypeptides wherein one or more amino acid residues are added or deleted at the N-and/or C-terminus, as well as within one or more internal domains, of the full-length or "native sequence" amino acid sequence. When discussing extracellular domains of receptors, fragments that bind a native sequence BlyS polypeptide are also contemplated. Conversely, when discussing BLyS fragments, fragments that bind any one or more of the three BLyS receptors are contemplated. Ordinarily, a variant polypeptide will have at least about 80% amino acid sequence identity, more preferably at least about 81% amino acid sequence identity, more preferably at least about 82% amino acid sequence identity, more preferably at least about 83% amino acid sequence identity, more preferably at least about 84% amino acid sequence identity, more preferably at least about 85% amino acid sequence identity, more preferably at least about 86% amino acid sequence identity, more preferably at least about 87% amino acid sequence identity, more preferably at least about 88% amino acid sequence identity, more preferably at least about 89% amino acid sequence identity, more preferably at least about 90% amino acid sequence identity, more preferably at least about 91% amino acid sequence identity, more preferably at least about 92% amino acid sequence identity, more preferably at least about 93% amino acid sequence identity, more preferably at least about 94% amino acid sequence identity, more preferably at least about 95% amino acid sequence identity, more preferably at least about 96% amino acid sequence identity, more preferably at least about 97% amino acid sequence identity, more preferably at least about 98% amino acid sequence identity and yet more preferably at least about 99% amino acid sequence identity with the polypeptide or a specified fragment thereof. Generally, variant polypeptides do not encompass the native polypeptide sequence. Ordinarily, variant

polypeptides are at least about 10 amino acids in length, often at least about 20 amino acids in length, more often at least about 30 amino acids in length, more often at least about 40 amino acids in length, more often at least about 50 amino acids in length, more often at least about 60 amino acids in length, more often at least about 70 amino acids in length, more often at least about 80 amino acids in length, more often at least about 90 amino acids in length, more often at least about 100 amino acids in length, more often at least about 150 amino acids in length, more often at least about 200 amino acids in length, more often at least about 250 amino acids in length, more often at least about 300 amino acids in length, or more.

[28] As mentioned above, a BLyS and/or APRIL antagonist can function in a direct or indirect manner to partially or fully block, inhibit or neutralize BLyS signaling, in vitro or in vivo. For instance, the BLyS and/or APRIL antagonist can directly bind BLyS. For example, a direct binder is a polypeptide comprising the extracellular domain (ECD) of a BLyS receptor such as TACI, BAFF-R, and BCMA.

[29] The BLyS receptors involved in the present invention can be described as follows. The TACI polypeptides of the invention include TACI polypeptides comprising or consisting of amino acids 1-246 of SEQ ID NO: 2. The general term "TACI" includes the TACI polypeptides described in WO 98/39361, WO 00/40716, WO 01/85782, WO 01/87979, WO 01/81417, and WO 02/094852. The TACI polypeptides of the invention can be isolated from a variety of sources, such as from human tissue types or from another source, or prepared by recombinant and/or synthetic methods. The BAFF-R polypeptides of the invention include the BAFF-R polypeptide comprising or consisting of the contiguous sequence of amino acid residues 1 to 184 of SEQ ID NO:4. The general term "BAFF-R" includes the BAFF-R polypeptides described in WO 02/24909 and WO 03/14294. The BAFF-R polypeptides of the invention can be isolated from a variety of sources, such as from human tissue types or from another source, or prepared by recombinant and/or synthetic methods. The BCMA polypeptide of the invention include BCMA polypeptides comprising or consisting of amino acid residues 1-184 of SEQ ID NO:6. The general term "BCMA" includes the BCMA polypeptides described in Laabi et al. , EMBO J., 11: 3897-3904 (1992); Laabi et al. , Nucleic Acids Res., 22: 1147-1154 (1994); Gras etal., Int. Immunology, 7: 1093-1106 (1995); and Madry etal., Int. Immunology, 10: 1693-1702 (1998). The BCMA polypeptides of the invention can be isolated from a variety of sources, such as from human tissue types or from another source, or prepared by recombinant and/or synthetic methods.

[30] For the purposes of functioning as a BLyS and/or APRIL antagonist, the ECD of these receptors is a polypeptide essentially free of the transmembrane or cytoplasmic domains that generally retains the ability to bind BLyS. Specifically, the extracellular domain of TACI can comprise amino acids 1 to 154 of the TACI polypeptide sequence (SEQ ID NO: 2). Additionally, the ECD can be fragments or variants of this sequence, such as ECD forms of TACI as described in von Bulow et al., supra, WO 98/39361, WO 00/40716, WO 01/85782, WO 01/87979, and WO 01/81417. In particular, these ECD forms can comprise amino acids 1-106 of SEQ ID NO:2, amino acids 1-142 of SEQ ID NO:2, amino acids 30-154 of SEQ ID NO:2, amino acids 30-106 of SEQ ID NO:2, amino acids 30-110 of SEQ ID NO:2, amino acids 30-119 of SEQ ID NO:2, amino acids 1-166 of SEQ ID NO:2, amino acids 1-165 of SEQ ID NO:2, amino acids 1-114 of SEQ ID NO: 2, amino acids 1-119 of SEQ ID NO:2, amino acids 1-120 of SEQ ID NO:2, and amino acids 1-126 of SEQ ID NO:2. In addition, the TACI ECD can comprise those molecules having only one cysteine rich domain

[31] ECD forms of BAFF-R include those comprising amino acids 1–71 of the BAFF-R polypeptide sequence (SEQ ID NO: 4). Additionally, the ECD can be fragments or variants of this sequence such as ECD forms of BAFF-R as described in WO 02/24909, WO 03/14294, and WO 02/38766. In particular, these ECD forms can comprise amino acids 1-77 of SEQ ID NO: 4, amino acids 7-77 of SEQ ID NO:4, amino acids 1-69 of SEQ ID NO:4, amino acids 7-69 of SEQ ID NO:4, amino acids 2-62 of SEQ ID NO:4, amino acids 2-71 of SEQ ID NO:4, amino acids 1-61 of SEQ ID NO:4 and amino acids 2-63 of SEQ ID NO:4, amino acids 1-45 of SEQ ID NO:4, amino acids 1-39 of SEQ ID NO:4, amino acids 7-39 of SEQ ID NO:4, amino acids 1-17 of SEQ ID NO:4, amino acids 39-64 of SEQ ID NO:4, amino acids 19-35 of SEQ ID NO:4, and amino acids 17-42 of SEQ ID NO:4. In addition, the BAFF-R ECD can comprise those molecules having a cysteine rich domain.

[32] ECD forms of BCMA include those comprising amino acids 1-48 of the BCMA polypeptide sequence (SEQ ID NO: 6). Additionally, the ECD can be fragments or variants of this sequence, such as ECD forms of BCMA as described in WO 00/40716 and WO 05/075511. In particular, these ECD forms can comprise amino acids 1-150 of SEQ ID NO:6, amino acids 1-48 of SEQ ID NO:6, amino acids 1-41 of SEQ ID NO:6, amino acids 8-41 of SEQ ID NO:6, amino acids 8-37 of SEQ ID NO:6, amino acids 8-88 of SEQ ID NO:6, amino acids 41-88 of SEQ ID NO:6, amino acids 1-54 of SEQ ID NO:6, amino acids 4-55 of SEQ ID NO:6, amino acids 4-51 of SEQ ID NO:6, and amino acids 21-53 of SEQ ID NO:6.

In addition, the BCMA ECD can comprise those molecules having only a partial cysteine rich domain.

[33] In a further embodiment, the BLyS binding region of a BLyS receptor (e. g., an extracellular domain or fragment thereof of BAFF-R, BCMA or TACI) can be fused to an Fc portion of an immunoglobulin molecule to facilitate its solubility in vivo. According to one embodiment, the BLyS and/or APRIL antagonist binds to a BLyS polypeptide with a binding affinity of 100nM or less. According to another embodiment, the BLyS and/or APRIL antagonist binds to a BLyS polypeptide with a binding affinity of 10nM or less. According to yet another embodiment, the BLyS and/or APRIL antagonist binds to a BLyS polypeptide with a binding affinity of 1nM or less.

[34] In another example, BLyS and/or APRIL antagonists include BLyS binding polypeptides that are not native sequences or varients thereof. Some examples of such polypepeptides are those having the sequence of Formula I, Formula II, Formula III as described in WO 05/000351. In particular, some binding polypeptides include ECFDLLVRAWVPCSVLK (SEQ ID NO:13), ECFDLLVRHWVPCGLLR (SEQ ID NO:14), ECFDLLVRRWVPCEMLG (SEQ ID NO:15), ECFDLLVRSWVPCHMLR (SEQ ID NO:16), ECFDLLVRHWVACGLLR (SEQ ID NO:17), or sequences listed in FIG. 32 of WO 05/000351.

[35] Alternatively, the BLyS and/or APRIL antagonist can bind an extracellular domain of native sequence TACI, BAFF-R, or BCMA at its BLyS binding region to partially or fully block, inhibit or neutralize BLyS binding in vitro, in situ, or in vivo. For example, such indirect antagonist is a TACI antibody that binds in a region of TACI such that the binding of BLyS is sterically hindered. For example, binding at amino acids 72-109 or a neighboring region is believed to block BLyS binding. It could also be advantageous to block APRIL binding to this molecule, which is believed to occur in the region of amino acids 82-222. Another BLyS and/or APRIL antagonist is a BAFF-R antibody that binds in a region of BAFF-R such that binding of human BAFF-R to BLyS is sterically hindered. For example, binding at amino acids 23-38 or amino acids 17-42 or a neighboring region is believed to block BLyS binding. Finally, a further indirect antagonist would be a BCMA antibody that binds in a rgion of BCMA such that the binding of BLyS is sterically hindered. For example, binding at amino acids 5-43 or a neighborhing region is believed to block BLyS (or APRIL) binding.

[36] In some embodiments, a BLyS and/or APRIL antagonist according to this invention includes BLyS antibodies. The term "antibody" when referring to is used in the broadest sense and specifically covers, for example, monoclonal antibodies, polyclonal antibodies, antibodies with polyepitopic specificity, single chain antibodies, and fragments of antibodies. According to some embodiments, a polypeptide of this invention is fused into an antibody framework, for example, in the variable region or in a CDR such that the antibody can bind to and inhibit BLyS binding to TACI, BAFF-R, or BCMA or inhibits BLyS signaling. The antibodies comprising a polypeptide of this invention can be chimeric, humanized, or human. The antibodies comprising a polypeptide of this invention can be an antibody fragment. Alternatively, an antibody of this invention can be produced by immunizing an animal with a polypeptide of this invention. Thus, an antibody directed against a polypeptide of this invention is contemplated.

[37] In particular, antibodies specific for BLyS that bind within a region of human BLyS (SEQ ID NO: 8) comprising residues 162-275 and/or a neighboring amino acid of amino acids selected from the group consisting of 162, 163, 206, 211, 231, 233, 264 and 265 of human BLyS are contemplated. The binding of the antibodies are such that the antibody sterically hinders BLyS binding to one or more of its receptors. Such antibodies are described in WO 02/02641 and WO 03/055979. A particularly preferred antibody is the one described as Lyphostat-B (Baker et al. (2003) Arthritis Rheum, 48, 3253-3265).

[38] The term "monoclonal antibody" as used herein refers to an antibody obtained from a population of substantially homogeneous antibodies, i. e., the individual antibodies comprising the population are identical except for possible naturally occurring mutations that can be present in minor amounts.

[39] Monoclonal antibodies are highly specific, being directed against a single antigenic site. Furthermore, in contrast to conventional (polyclonal) antibody preparations which typically include different antibodies directed against different determinants (epitopes), each monoclonal antibody is directed against a single determinant on the antigen. In addition to their specificity, the monoclonal antibodies are advantageous in that they are synthesized by the hybridoma culture, uncontaminated by other immunoglobulins. The modifier "monoclonal" indicates the character of the antibody as being obtained from a substantially homogeneous population of antibodies, and is not to be construed as requiring production of the antibody by any particular method. For example, the monoclonal antibodies to be used in accordance with the present invention may be made by the hybridoma method first described

by Kohler etal., Nature, 256: 495 (1975), or may be made by recombinant DNA methods (see, e. g. , U. S. Patent No. 4,816, 567). The "monoclonal antibodies"may also be isolated from phage antibody libraries using the techniques described in Clackson etal., Nature, 352: 624-628 (1991) and Marks et al. , J. Mol.Biol., 222: 581-597 (1991), for example.

[40] The monoclonal antibodies herein specifically include "chimeric" antibodies (immunoglobulins) in which a portion of the heavy and/or light chain is identical with or homologous to corresponding sequences in antibodies derived from a particular species or belonging to a particular antibody class or subclass, while the remainder of the chain (s) is identical with or homologous to corresponding sequences in antibodies derived from another species or belonging to another antibody class or subclass, as well as fragments of such antibodies, so long as they exhibit the desired biological activity (U. S. Patent No. 4,816, 567; Morrison et al., Proc. Natl. Acad. Sci. USA, 81: 6851-6855 (1984)). Methods of making chimeric antibodies are known in the art.

[41] "Humanized" forms of non-human (e. g. , murine) antibodies are chimeric immunoglobulins, immunoglobulin chains or fragments thereof (such as Fv, Fab, Fab', F (ab') 2 or other antigen-binding subsequences of antibodies) which contain minimal sequence derived from non-human immunoglobulin.

[42] For the most part, humanized antibodies are human immunoglobulins (recipient antibody) in which residues from a complementarity-determining region (CDR) of the recipient are replaced by residues from a CDR of a non-human species (donor antibody) such as mouse, rat or rabbit having the desired specificity, affinity, and capacity. In some instances, Fv framework region (FR) residues of the human immunoglobulin are replaced by corresponding non-human residues. Furthermore, humanized antibodies may comprise residues which are found neither in the recipient antibody nor in the imported CDR or framework sequences. These modifications are made to further refine and maximize antibody performance. In general, the humanized antibody will comprise substantially all of at least one, and typically two, variable domains, in which all or substantially all of the hypervariable loops correspond to those of a non-human immunoglobulin and all or substantially all of the FR regions are those of a human immunoglobulin sequence although the FR regions may include one or more amino acid substitutions that improve binding affinity. The number of these amino acid substitutions in the FR are typically no more than 6 in the H chain, and in the L chain, no more than 3. The humanized antibody optimally also will comprise at least a portion of an immunoglobulin constant region (Fc), typically that of a human

immunoglobulin. For further details, see Jones et al., Nature, 321: 522-525 (1986); Reichmann et al. , Nature, 332: 323-329 (1988); and Presta, Curr. Op. Struct. Biol., 2: 593-596 (1992). The humanized antibody includes a PRIMATIZED antibody wherein the antigen-binding region of the antibody is derived from an antibody produced by, e. g. , immunizing macaque monkeys with the antigen of interest. Methods of making humanized antibodies are known in the art.

[43] Human antibodies can also be produced using various techniques known in the art, including phage-display libraries. Hoogenboom and Winter, J. Mol.Biol., 227: 381 (1991); Marks et al. , J. Mol. Biol., 222: 581 (1991). The techniques of Cole et al. and Boerner et al. are also available for the preparation of human monoclonal antibodies. Cole et al. , Monoclonal Antibodies and Cancer Therapy, Alan R. Liss, p. 77 (1985); Boerner et al. , J. Immunol., 147(1) : 86-95 (1991).

[44] "Functional fragments" of the binding antibodies of the invention are those fragments that retain binding to BLyS, TACI, BAFF-R, or BCMA with substantially the same affinity as the intact full chain molecule from which they are derived and may be able to deplete B cells as measured by in vitro or in vivo assays such as those described herein.

[45] Antibody "effector functions" refer to those biological activities attributable to the Fc region (a native sequence Fc region or amino acid sequence variant Fc region) of an antibody, and vary with the antibody isotype. Examples of antibody effector functions include: Clq binding and complement dependent cytotoxicity; Fc receptor binding; antibody-dependent cell-mediated cytotoxicity (ADCC); phagocytosis; down regulation of cell surface receptors (e. g. B cell receptor); and B cell activation.

[46] "Antibody-dependent cell-mediated cytotoxicity" or "ADCC" refers to a form of cytotoxicity in which secreted Ig bound onto Fc receptors (FcRs) present on certain cytotoxic cells (e. g. Natural Killer (NK) cells, neutrophils, and macrophages) enable these cytotoxic effector cells to bind specifically to an antigen-bearing-target cell and subsequently kill the-target cell with cytotoxins. The antibodies-"arm"the cytotoxic cells and are absolutely required for such killing. The primary cells for mediating ADCC, NK cells, express FcyRIII only, whereas monocytes express FcyRI, FcyRII and FcyRIII. FcR expression on hematopoietic cells is summarized in Table 3 on page 464 of Ravetch and Kinet, Ann. Rev. Immunol 9: 457-92 (1991). To assess ADCC activity of a molecule of interest, an in vitro ADCC assay, such as that described in US Patent No. 5,500, 362 or 5,821, 337 may be performed. Useful effector cells for such assays include peripheral blood mononuclear cells

(PBMC) and Natural Killer (NK) cells. Alternatively, or additionally, ADCC activity of the molecule of interest may be assessed in vivo, e. g. , in a animal model such as that disclosed in Clynes et al. PNAS (USA) 95: 652-656 (1998).

[47] "Complement dependent cytotoxicity" or "CDC" refers to the lysis of a target cell in the presence of complement. Activation of the classical complement pathway is initiated by the binding of the first component of the complement system (Clq) to antibodies (of the appropriate subclass) which are bound to their cognate antigen. To assess complement activation, a CDC assay, e. g. as described in Gazzano- Santoroetal., J. Immunol. Methods 202: 163 (1996), may be performed.

[48] An "isolated" antibody is one which has been identified and separatedand/or recovered from a component of its natural environment. Contaminant components of its natural environment are materials which would interfere with diagnostic or therapeutic uses for the antibody, and may include enzymes, hormones, and other proteinaceous or nonproteinaceous solutes. In preferred embodiments, the antibody will be purified (1) to greater than 95% by weight of antibody as determined by the Lowry method, and most preferably more than 99% by weight, (2) to a degree sufficient to obtain at least 15 residues of N- terminal or internal amino acid sequence by use of a spinning cup sequenator, or (3) to homogeneity by SDS-PAGE under reducing or nonreducing conditions using Coomassie blue or, preferably, silver stain.

[49] Isolated antibody includes the antibody insitu within recombinant cells since at least one component of the antibody's natural environment will not be present. Ordinarily, however, isolated antibody will be prepared by at least one purification step.

[50] Amino acids may be grouped according to similarities in the properties of their side chains (in A. L. Lehninger, in Biochemistry, second ed. , pp. 73-75, Worth Publishers, New York (1975)) : (1) non-polar: Ala (A), Val (V), Leu (L), Ile (I), Pro (P), Phe (F), Trp (W), Met (M) (2) uncharged polar: Gly (G), Ser (S), Thr (T), Cys (C), Tyr (Y), Asn (N),Gln (Q) (3) acidic: Asp (D), Glu (E) (4) basic : Lys(K), Arg (R), His(H-) Alternatively, naturally occurring residues may be divided into groups based on common side- chain properties : (1) hydrophobic: Norleucine, Met, Ala, Val, Leu, Ile ; (2) neutral hydrophilic : Cys, Ser, Thr, Asn,Gln ; (3) acidic: Asp, Glu; (4) basic: His, Lys, Arg; (5) residues that influence chain orientation: Gly, Pro; (6) aromatic: Trp, Tyr, Phe.

[51] The term "conservative" amino acid substitution as used within this invention is meant to refer to amino acid substitutions which substitute functionally equivalent amino

acids. Conservative amino acid changes result in silent changes in the amino acid sequence of the resulting peptide. For example, one or more amino acids of a similar polarity act as functional equivalents and result in a silent alteration within the amino acid sequence of the peptide. In general, substitutions within a group may be considered conservative with respect to structure and function. However, the skilled artisan will recognize that the role of a particular residue is determined by its context within the three-dimensional structure of the molecule in which it occurs. For example, Cys residues may occur in the oxidized (disulfide) form, which is less polar than the reduced (thiol) form. The long aliphatic portion of the Arg side chain may constitute a critical feature of its structural or functional role, and this may be best conserved by substitution of a nonpolar, rather than another basic residue. Also, it will be recognized that side chains containing aromatic groups (Trp, Tyr, and Phe) can participate in ionic-aromatic or"cation-pi"interactions. In these cases, substitution of one of these side chains with a member of the acidic or uncharged polar group may be conservative with respect to structure and function. Residues such as Pro, Gly, and Cys (disulfide form) can have direct effects on the main chain conformation, and often may not be substituted without structural distortions.

[52] "Percent (%) amino acid sequence identity"with respect to the ligand or receptor polypeptide sequences identified herein is defined as the percentage of amino acid residues in a candidate sequence that are identical with the amino acid residues in such a ligand or receptor sequence identified herein, after aligning the sequences and introducing gaps, if necessary, to achieve the maximum percent sequence identity, and not considering any conservative substitutions as part of the sequence identity. Alignment for purposes of determining percent amino acid sequence identity can be achieved in various ways that are within the skill in the art, for instance, using publicly available computer software such as BLAST, BLAST-2, ALIGN, ALIGN-2 or Megalign (DNASTAR) software. Those skilled in the art can determine appropriate parameters for measuring alignment, including any algorithms needed to achieve maximal alignment over the full-length of the sequences being compared. For purposes herein, however, % amino acid sequence identity values are obtained as described below by using the sequence comparison computer program ALIGN-2, wherein the complete source code for the ALIGN-2 program is provided in the table below. The ALIGN-2 sequence comparison computer program was authored by Genentech, Inc. and the source code shown in the table below has been filed with user documentation in the U. S. Copyright Office, Washington D. C. , 20559, where it is registered under U. S. Copyright

Registration No.TXU510087. The ALIGN-2 program is-publicly available through Genentech, Inc., South San Francisco, California or can be compiled from the source code provided in the table below. The ALIGN-2 program should be compiled for use on a UNIX operating system, preferably digital UNIX V4.0D. All sequence comparison parameters are set by the ALIGN-2 program and do not vary.

[53] A useful method for identification of certain residues or regions in a protein that are preferred locations for mutagenesis is called"alanine scanning mutagenesis"as described by Cunningham and Wells Science, 244: 1081-1085 (1989). A residue or group of target residues are identified (e. g. , charged residues such as arg, asp, his, lys, and glu) and replaced by a neutral or negatively charged amino acid (most preferably alanine or polyalanine) to affect the interaction of the amino acids with a binding target. Those amino acid locations demonstrating functional sensitivity to the substitutions then are refined by introducing further or other variants at, or for, the sites of substitution. Thus, while the site for introducing an amino acid sequence variation is predetermined, the nature of the mutation per se need not be predetermined. For example, to analyze the performance of a mutation at a given site, ala scanning or random mutagenesis is conducted at the target codon or region and the expressed variants are screened for the desired activity.

[54] The term, "dihedral angle" refers to a rotation about a bond. See e. g. , Creighton, T. E. , (1993) Protein: Structures and Molecular Properties, 2 ed. , W. H. Freeman and Company, New York, NY. The term,"phi,"is a dihedral angle that denotes a rotation about theN-C bond of an amino acid. See e. g. , Creighton, T. E. , (1993) Protein: Structures and Molecular Properties, 2 ed. , W. H. Freeman and Company, New York, NY. Type I beta turns are described in Hutchinson, E. G. & Thornton, J. M. (1994) A revised set of potentials for beta turn formation in proteins. Protein Science 3, 2207-2216.

[55] A "fusion protein" and a "fusion polypeptide" refer to a polypeptide having two portions covalently linked together, where each of the portions is a polypeptide having a different property. The property may be a biological property, such as activity in vitro or in vivo. The property may also be a simple chemical or physical property, such as binding to a target molecule, catalysis of a reaction, etc. The two portions may be linked directly by a single peptide bond or through a peptide linker containing one or more amino acid residues. Generally, the two portions and the linker will be in reading frame with each other.

[56] A "conjugate" refers to any hybrid molecule, including fusion proteins and as well as molecules that contain both amino acid or protein portions and non-protein portions.

Conjugates may be synthesized by a variety of techniques known in the art including, for example, recombinant DNA techniques, solid phase synthesis, solution phase synthesis, organic chemical synthetic techniques or a combination of these techniques. The choice of synthesis will depend upon the particular molecule to be generated. For example, a hybrid molecule not entirely"protein"in nature may be synthesized by a combination of recombinant techniques and solution phase techniques.

[57] As used herein, the term "Fc-fusion protein" designates antibody-like molecules which combine the binding specificity of a heterologous protein with the effector functions of immunoglobulin constant domains. Structurally, the Fc-fusion proteins comprise a fusion of an amino acid sequence with the desired binding specificity which is other than the antigen recognition and binding site of an antibody (i. e., is "heterologous"), and an immunoglobulin constant domain sequence. The Fc-fusion protein molecule typically includes a contiguous amino acid sequence comprising at least the binding site of a receptor or a ligand. The immunoglobulin constant domain sequence in the Fc-fusion protein can be obtained from any immunoglobulin, such as IgG-1, IgG-2, IgG-3, or IgG-4 subtypes, IgA (includingIgA-1 and IgA-2), IgE, IgD or IgM. For example, useful Fc-fusion proteins according to this invention are polypeptides that comprise the BLyS binding portions of a BLyS receptor without the transmembrane or cytoplasmic sequences of the BLyS receptor. In one embodiment, the extracellular domain of BAFF-R, TACI or BCMA is fused to a constant domain of an immunoglobulin sequence.

The term “mammal” refers to any animal classified as a mammal, including humans, domestic and farm animals, and zoo, sports, or pet animals, such as dogs, horses, cats, cows, etc. Preferably, the mammal herein is human.

[58] The term "therapeutically effective amount" refers to an amount of an antibody or a antagonist drug effective to "alleviate" or "treat" a disease or disorder in a subject or mammal. In the case of cancer, the therapeutically effective amount of the drug may reduce the number of cancer cells; reduce the tumor size; inhibit(i. e., slow to some extent and preferably stop) cancer cell infiltration into peripheral organs; inhibit (i. e. , slow to some extent and preferably stop) tumor metastasis; inhibit, to some extent, tumor growth; and/or relieve to some extent one or more of the symptoms associated with the cancer. See the definition of"treated"below. To the extent the drug may prevent growth and/or kill existing cancer cells, it may be cytostatic and/or cytotoxic.

[59] The BLyS or BLyS receptor antibodies of the invention can be produced by transient or stable transfection eukaryotic host cells such as CHO cells.

"Carriers" as used herein include pharmaceutically acceptable carriers, excipients, or stabilizers which are nontoxic to the cell or mammal being exposed thereto at the dosages and concentrations employed. Often the physiologically acceptable carrier is an aqueous pH buffered solution. Examples of physiologically acceptable carriers include buffers such as phosphate, citrate, and other organic acids; antioxidants including ascorbic acid; low molecular weight (less than about 10 residues) polypeptide; proteins, such as serum albumin, gelatin, or immunoglobulins; hydrophilic polymers such as polyvinylpyrrolidone ; amino acids such as glycine, glutamine, asparagine, arginine or lysine ; monosaccharides, disaccharides, and other carbohydrates including glucose, mannose, or dextrins; chelating agents such as EDTA; sugar alcohols such as mannitol or sorbitol; salt-forming counterions such as sodium; and/or nonionic surfactants such as TWEEN polyethylene glycol(PEG), and PLURONICSTM.

Polynucleotides, Vectors, Host Cells

[60] According to a number of embodiments disclosed herein, the BLyS and/or APRIL antagonist can comprise specific polypeptides that are produced using specific polynucleotides in specific vectors and using specific host cells. The various types of polypeptides of the present invention can be broadly described and are selected from the group consisting of receptor-based seqences, antibody-based sequences, and artifical (i.e., non-native) binding sequences. Examples of the receptor-based sequences are those sequences that bind BLyS that were isolated from or derived from domains of the receptors that bind BLyS in vivo, such as TACI, BAFF-R, or BCMA. Antibody-based sequences are those that are produced using antibody development technology and maintain the general structure of an antibody molecule. Examples of antibody-based sequences are LymphoStat-B, or antibodies to receptors of BLyS. Examples of the artificial binding sequences include the 17mer peptides described herein, polypeptides incorporating one or more 17mer peptides as core regions, and covalently modified forms of the 17mer peptides and polypeptides (e. g., Fc-fusion proteins, labeled polypeptides, protected polypeptides, conjugated polypeptides, fusion proteins, etc.). Various techniques that are employed for making these forms of polypeptides are described herein. Methods for labeling polypeptides and conjugating molecules to polypeptides are known in the art.

[61] Compositions of the invention can be prepared using recombinant techniques known in the art.

[62] The description below relates to methods of producing such specific polypeptides by culturing host cells transformed or transfected with a vector containing the encoding nucleic acid and recovering the polypeptide from the cell culture. (See, e.g., Sambrook etal., Molecular Cloning: A Laboratory Manual (New York: Cold Spring Harbor Laboratory Press, 1989); Dieffenbach et al., PCR Primer: A Laboratory Manual (Cold Spring Harbor Laboratory Press, 1995)).

[63] The nucleic acid (e. g., cDNA orgenomic DNA) encoding the desired polypeptide may be inserted into a replicable vector for further cloning (amplification of the DNA) or for expression. Various vectors are publicly available. The vector components generally include, but are not limited to, one or more of the following: a signal sequence, an origin of replication, one or more marker genes, an enhancer element, a promoter, and a transcription termination sequence, each of which is described below. Optional signal sequences, origins of replication, marker genes, enhancer elements and transcription terminator sequences that may be employed are known in the art and described in further detail in W0 97/25428.

[64] Expression and cloning vectors usually contain a promoter that is recognized by the host organism and is operably linked to the encoding nucleic acid sequence. Promoters are untranslated sequences located upstream (5') to the start codon of a structural gene (generally within about 100 to 1000 bp) that control the transcription and translation of a particular nucleic acid sequence, to which they are operably linked. Such promoters typically fall into two classes, inducible and constitutive. Inducible promoters are promoters that initiate increased levels of transcription from DNA under their control in response to some change in culture conditions, e. g. , the presence or absence of a nutrient or a change in temperature. At this time a large number of promoters recognized by a variety of potential host cells are well known. These promoters are operably linked to the encoding DNA by removing the promoter from the source DNA by restriction enzyme digestion and inserting the isolated promoter sequence into the vector.

[65] Construction of suitable vectors containing one or more of the above-listed components employs standard ligation techniques. Isolated plasmids or DNA fragments are cleaved, tailored, andre-ligated in the form desired to generate the plasmids required. For analysis to confirm correct sequences in plasmids constructed, the ligation mixtures can be

used to transform E. coli K12 strain 294 (ATCC 31,446) and successful transformants selected by ampicillin or tetracycline resistance where appropriate. Plasmids from the transformants are prepared, analyzed by restriction endonuclease digestion, and/or sequenced using standard techniques known in the art. [See, e. g. , Messing etal., Nucleic Acids Res. , 9: 309 (1981); Maxam et al. , Methods in Enzymology, 65: 499 (1980)].

[66] Expression vectors that provide for the transient expression in mammalian cells of the encoding DNA may be employed. In general, transient expression involves the use of an expression vector that is able to replicate efficiently in a host cell, such that the host cell accumulates many copies of the expression vector and, in turn, synthesizes high levels of a desired polypeptide encoded by the expression vector [Sambrook et al. , supra]. Transient expression systems, comprising a suitable expression vector and a host cell, allow for the convenient positive identification of polypeptides encoded by clonedDNAs, as well as for the rapid screening of such polypeptides for desired biological or physiological properties.

[67] Other methods, vectors, and host cells suitable for adaptation to the synthesis of the desired polypeptide in recombinant vertebrate cell culture are described in Gething etal., Nature, 293: 620-625 (1981); Mantei etal., Nature, 281: 40-46 (1979); EP 117,060 ; and EP 117,058.

[68] Suitable host cells for cloning or expressing the DNA in the vectors herein include prokaryote, yeast, or higher eukaryote cells. Suitable prokaryotes for this purpose include but are not limited toeubacteria, such as Gram-negative or Gram-positiveorganisms, for-example, Enterobacteriaceae such as Escherichia, e. g. , E. coli, Enterobacter, Erwinia,Klebsiella, Proteus, Salmonella, e. g. , Salmonella typhimurium, Serratia, e. g. , Serratia marcescans, and Shigella, as well as Bacilli such as B. subtilis and B.licheniformis (e. g. , B.licheniformis 41P disclosed in DD 266,710 published 12 April 1989), Pseudomonas such as P. aeruginosa, and Streptomyces. Preferably, the host cell should secrete minimal amounts of proteolytic enzymes.

[69] In addition to prokaryotes, eukaryotic microbes such as filamentous fungi or yeast are suitable cloning or expression hosts for vectors. Suitable host cells for the expression of glycosylated polypeptide are derived from multicellular organisms. Examples of all such host cells are described further in W097/25428.

[70] Host cells are transfected and preferably transformed with the above-described expression or cloning vectors and cultured in nutrient media modified as appropriate for

inducing promoters, selecting transformants, or amplifying the genes encoding the desired sequences.

[71] Transfection refers to the taking up of an expression vector by a host cell whether or not any coding sequences are in fact expressed. Numerous methods of transfection are known to the ordinarily skilled artisan, for example,CaP04 and electroporation. Successful transfection is generally recognized when any indication of the operation of this vector occurs within the host cell.

[72] Transformation means introducing DNA into an organism so that the DNA isreplicable, either as an extrachromosomal element or by chromosomal integrant. Depending on the host cell used, transformation is done using standard techniques appropriate to such cells. The calcium treatment employing calcium chloride, as described in Sambrook et al. , supra, or electroporation is generally used for prokaryotes or other cells that contain substantial cell-wall barriers. Infection with Agrobacterium tumefaciens is used for transformation of certain plant cells, as described by Shaw et al., Gene, 23: 315 (1983) and WO 89/05859 published 29 June 1989. In addition, plants may be transfected using ultrasound treatment as described in WO 91/00358 published 10 January1991.

[73] For mammalian cells without such cell walls, the calcium phosphate precipitation method of Graham and van der Eb, Virology, 52: 456-457 (1978) may be employed. General aspects of mammalian cell host system transformations have been described in U. S. Pat. No. 4,399, 216. Transformations into yeast are typically carried out according to the method of Van Solingen et al., J. Bact. , 130: 946 (1977) and Hsiao et al. , Proc. Natl. Acad. Sci. (USA), 76: 3829 (1979). However, other methods for introducing DNA into cells, such as by nuclear microinjection, electroporation, bacterial protoplast fusion with intact cells, or polycations, e. g., polybrene, polyornithine, may also be used. For various techniques for transforming mammalian cells, see Keown et al., Methods in Enzymology, 185: 527-537 (1990) and Mansour et al., Nature, 336: 348-352 (1988).

[74] Prokaryotic cells can be cultured in suitable culture media as described generally in Sambrook et al., supra. Examples of commercially available culture media include Ham's F10 (Sigma), Minimal Essential Medium ("MEM", Sigma), RPMI-1640 (Sigma), and Dulbecco's Modified Eagle's Medium ("DMEM", Sigma). Any such media may be supplemented as necessary with hormones and/or other growth factors (such as insulin, transferrin, or epidermal growth factor), salts (such as sodium chloride, calcium, magnesium, and phosphate), buffers (such as HEPES), nucleosides (such as adenosine andthymidine),

antibiotics (such as gentamycin) ,-trace elements (defined as inorganic compounds usually present at final concentrations in themicromolar range), and glucose or an equivalent energy source. Any other necessary supplements may also be included at appropriate concentrations that would be known to those skilled in the art. The culture conditions, such as temperature, pH, and the like, Ore those previously used with the host cell selected for expression, and will be apparent to the ordinarily skilled artisan.

[75] In general, principles, protocols, and practical techniques for maximizing the productivity of mammalian cell cultures can be found in Mammalian Cell Biotechnology: A Practical Approach, M. Butler, ed. (IRL Press, 1991).

[76] The expressed polypeptides may be recovered from the culture medium as a secreted polypeptide, although may also be recovered from host cell lysates when directly produced without a secretory signal. If the polypeptide is membrane-bound, it can be released from the membrane using a suitable detergent solution (e. g. Triton-X 100) or its extracellular region may be released by enzymatic cleavage.

[77] When the polypeptide is produced in a recombinant cell other than one of human origin, it is free of proteins or polypeptides of human origin. However, it is usually necessary to recover or purify the polypeptide from recombinant cell proteins or polypeptides to obtain preparations that are substantially homogeneous. As a first step, the culture medium or lysate may be centrifuged to remove particulate cell debris. The following are procedures exemplary of suitable purification procedures: by fractionation on an ion-exchange column; ethanol precipitation; reverse phase HPLC; chromatography on silica or on a cation-exchange resin such as DEAE; chromatofocusing; SDS-PAGE; ammonium sulfate precipitation; gel filtration using, for example, Sephadex G-75; and protein A Sepharose columns to remove contaminants such as IgG.

Phage Display

[78] According to some embodiments, the polypeptides of this invention selected from the group consisting of: Formula I, Formula II, Formula III, ECFDLLVRAWVPCSVLK (SEQ ID NO :13), ECFDLLVRHWVPCGLLR (SEQ ID NO:14), ECFDLLVRRWVPCEMLG (SEQ ID NO:15), ECFDLLVRSWVPCHMLR (SEQ ID NO:16), ECFDLLVRHWVACGLLR (SEQ ID NO:17), and sequences listed in FIG. 32 of WO 05/000351, may utilized in phage display.

[79] Using the techniques of phage display allows the generation of large libraries of protein variants which can be rapidly sorted for those sequences that bind to a target molecule with high affinity. Nucleic acids encoding variant polypeptides are fused to a nucleic acid sequence encoding a viral coat protein, such as the gene III protein or the gene VIII protein. Monovalent phage display systems where the nucleic acid sequence encoding the protein or polypeptide is fused to a nucleic acid sequence encoding a portion of the gene III protein have been developed. (Bass, S., Proteins, 8: 309 (1990); Lowman and Wells, Methods: A Companion to Methods in Enzymology, 3 : 205 (1991)). In a monovalent phage display system, the gene fusion is expressed at low levels and wild type gene III proteins are also expressed so that infectivity of the particles is retained. Methods of generating peptide libraries and screening those libraries have been disclosed in many patents (e. g. U. S. Patent No. 5,723,286; U. S. Patent No. 5,432,018; U. S. Patent No. 5,580,717; U. S. Patent No. 5,427,908; and U. S. Patent No. 5,498,530).

[80] In some embodiments, Formula I, Formula II or Formula III are expressed as peptide libraries on phage. The phage expressing the library of polypeptides of FormulaI, Formula II or Formula III are then subjected to selection based onBLyS binding. In some embodiments, the selection process involves allowing some phage bind to biotinylatedBLyS which is subsequently bound to a neutravidin plate. Phage bound to the plate through the BLyS-biotin-neutravidin binding are recovered and propogated. In some embodiments, the phage are subject to several rounds of selection. In some embodiments, the phage is incubated with BLyS-biotin, followed by the addition of unbiotinylated BLyS as a competitive binder.

[81] Additional guidance of use of phage display in the context of the present invention is provided in the Examples.

Polypeptides fused or conjugated to Heterologous polypeptides

[82] Fc-fusion protein molecules comprising the polypeptides of this invention are further contemplated for use in the methods herein. In some embodiments, the molecule comprises a fusion of a polypeptide of this invention with an immunoglobulin or a particular region of an immunoglobulin. For a bivalent form of the Fc-fusion protein, such a fusion usefully comprises the Fc region of an IgG molecule. In a further embodiment, the Fc region is from a human IgG1 molecule. In some embodiments, the immunoglobulin fusion includes the hinge, CH2 and CH3, or the hinge, CH1, CH2 and CH3 regions of anIgGl molecule.

[83] For the production of immunoglobulin fusions, see also US Patent No. 5,428,130, US Patent No. 5,843,725, U.S. Patent No. 6,018,026, and Chamow et al., TIBTECH, 14: 52-60 (1996).

[84] The simplest and most straightforward Fc-fusion protein design often combines the binding domain(s) of an antagonist polypeptide of this invention, preferably a native sequence, with the Fc region of an immunoglobulin heavy chain. In another embodiment, the polypeptide can be artificial, such as a polypeptide comprising a sequence of Formula I, Formula II, Formula III, ECFDLLVRAWVPCSVLK (SEQ ID NO: 13), ECFDLLVRHWVPCGLLR (SEQ ID NO: 14), ECFDLLVRRWVPCEMLG (SEQ ID NO: 15), ECFDLLVRSWVPCHMLR (SEQ ID NO: 16), ECFDLLVRHWVACGLLR (SEQ ID NO: 17), or sequences listed in FIG. 32 can be covalently linked to an Fc portion of an immunoglobulin. In addition, one or more of these polypeptides can be linked to one another and linked to an Fc portion of an immunoglobulin.

[85] Ordinarily, when preparing the Fc-fusion proteins of the present invention, nucleic acid encoding the binding domain will be fused C-terminally to nucleic acid encoding the N-terminus of an immunoglobulin constant domain sequence, however N-terminal fusions are also possible.

[86] Typically, in such fusions the encoded chimeric polypeptide will retain at least functionally active hinge, CH2 and CH3 domains of the constant region of an immunoglobulin heavy chain. Fusions are also made to the C-terminus of the Fc portion of a constant domain, or immediately N-terminal to the CH1 of the heavy chain or the corresponding region of the light chain. The precise site at which the fusion is made is not critical; particular sites are well known and may be selected in order to optimize the biological activity, secretion, or binding characteristics of the Fc-fusion protein.

[87] In a preferred embodiment, the binding domain sequence is fused to the N-terminus of the Fc region of immunoglobulin Gl(IgGl). It is possible to fuse the entire heavy chain constant region to the binding domain sequence. However, more preferably, a sequence beginning in the hinge region just upstream of the papain cleavage site which defines IgG Fc chemically (i. e. residue 216, taking the first residue of heavy chain constant region to be 114), or analogous sites of other immunoglobulins is used in the fusion. In a particularly preferred embodiment, the binding domain amino acid sequence is fused to (a) the hinge region and CH2 and CH3 or (b) the CH1, hinge, CH2 and CH3 domains, of an IgG heavy chain.

[88] For bispecific Fc-fusion proteins, the Fc-fusion proteins are assembled as multimers, and particularly as heterodimers or heterotetramers. Generally, these assembled immunoglobulins will have known unit structures. A basic four chain structural unit is the form in which IgG, IgD, and IgE exist. A four chain unit is repeated in the higher molecular weight immunoglobulins; IgM generally exists as a pentamer of four basic units held together by disulfide bonds. IgA globulin, and occasionally IgG globulin, may also exist in multimeric form in serum. In the case of multimer, each of the four units may be the same or different.

[89] Various exemplary assembled Fc-fusion proteins within the scope herein are schematically diagrammed below: (a) ACL-ACL; (b)ACH- (ACH, ACL-ACH, ACL-VHCH, or VLCL-ACH); (c) ACL-ACH- (ACL-ACH, ACL-VHCH, VLCL-ACH, or VLCL-VHCH) (d) ACL-VHCH- (ACH, or ACL-VHCH, or VLCL-ACH); (e) VLCL-ACH- (ACL-VHCH, or VLCL-ACH); and(f) (A-Y) n- (VLCL-VHCH) 2, wherein each A represents identical or different polypeptides comprising an amino acid sequence of sequences derived from BLyS receptor domains, sequences derived from antibodies to BLyS or to receptors to BLyS, or artifical sequences such as Formula I, Formula II, Formula III, ECFDLLVRAWVPCSVLK (SEQ ID NO 5), ECFDLLVRHWVPCGLLR (SEQ ID NO 6), ECFDLLVRRWVPCEMLG (SEQ ID NO 7), ECFDLLVRSWVPCHMLR (SEQ ID NO 8), ECFDLLVRHWVACGLLR (SEQ ID NO 9), or sequences listed in FIG. 32 or combinations thereof;

[90] VL is an immunoglobulin light chain variable domain; VH is an immunoglobulin heavy chain variable domain; CL is an immunoglobulin light chain constant domain; CH is an immunoglobulin heavy chain constant domain; n is an integer greater than 1; Y designates the residue of a covalent cross-linking agent.

[91] In the interests of brevity, the foregoing structures only show key features; they do not indicate joining (J) or other domains of the immunoglobulins, nor are disulfide bonds shown. However, where such domains are required for binding activity, they shall be constructed to be present in the ordinary locations which they occupy in the immunoglobulin molecules.

[92] Alternatively, the Fc sequences can be inserted between immunoglobulin heavy chain and light chain sequences, such that an immunoglobulin comprising a chimeric heavy chain is obtained. In this embodiment, the Fc sequences are fused to the 3'end of an immunoglobulin heavy chain in each arm of an immunoglobulin, either between the hinge and the CH2 domain, or between the CH2 and CH3 domains. Similar constructs have been reported by Hoogenboom et al., Mol. Immunol., 28: 1027-1037 (1991).

[93] Although the presence of an immunoglobulin light chain is not required in the Fc-fusion proteins of the present invention, an immunoglobulin light chain might be present either covalently associated to an binding domain-immunoglobulin heavy chain fusion polypeptide, or directly fused to the bdining domain. In the former case, DNA encoding an immunoglobulin light chain is typically coexpressed with the DNA encoding the binding domain-immunoglobulin heavy chain fusion protein. Upon secretion, the hybrid heavy chain and the light chain will be covalently associated to provide an immunoglobulin-like structure comprising two disulfide- linked immunoglobulin heavy chain-light chain pairs. Methods suitable for the preparation of such structures are, for example, disclosed in U. S. Patent No. 4,816,567, issued 28 March 1989.

[94] Fc-fusion proteins are most conveniently constructed by fusing the cDNA sequence encoding the binding domain portion in-frame to an immunoglobulin cDNA sequence. However, fusion to genomic immunoglobulin fragments can also be used (see, e. g. Aruffo et al., Cell, 61: 1303-1313 (1990); and Stamenkovic et al., Cell, 66: 1133-1144 (1991)). The latter type of fusion requires the presence of Ig regulatory sequences for expression. cDNAs encoding IgG heavy-chain constant regions can be isolated based on published sequences fromcDNA libraries derived from spleen or peripheral blood lymphocytes, by hybridization or by polymerase chain reaction (PCR) techniques. The cDNAs encoding the binding domain and the immunoglobulin parts of the Fc-fusion protein are inserted in tandem into a plasmid vector that directs efficient expression in the chosen host cells.

[95] Particular modifications have been made to produce Fc sequences useful for creating fusion molecules such as the BLyS and/or APRIL antagonist Fc fusion molecules for use in the present invention. Specifically, six versions of a modified human IgG1 Fc were generated for creating Fc fusion proteins and are named Fc-488, as well as Fc4, Fc5, Fc6, Fc7, and Fc8. Fc-488 (SEQ ID NO: 39) was designed for convenient cloning of a fusion protein containing the human γ1 Fc region, and it was constructed using the wild-type human immunoglobulin γ1 constant region as a template. Concern about potential deleterious effects due to an unpaired cysteine residue led to the decision to replace the cysteine that normally disulfide bonds with the immunoglobulin light chain constant region with a serine residue. An additional change was introduced at the codon encoding EU index position 218 to introduce a BglII restriction enzyme recognition site for ease of future DNA manipulations. These changes were introduced into the PCR product encoded on the PCR primers. Due to the location of the BglII site and in order to complete the Fc hinge region,

codons for EU index positions 216 and 217 were incorporated in the fusion protein partner sequences.

[96] Fc4, Fc5, and Fc6 contain mutations to reduce effector functions mediated by the Fc by reducing FcγRI binding and complement C1q binding. Fc4 contains the same amino acid substitutions that were introduced into Fc-488. Additional amino acid substitutions were introduced to reduce potential Fc mediated effector functions. Specifically, three amino acid substitutions were introduced to reduce FcγRI binding. These are the substitutions at EU index positions 234, 235, and 237. Substitutions at these positions have been shown to reduce binding to FcγRI (Duncan et al., Nature 332:563 (1988)). These amino acid substitutions may also reduce FcγRIIa binding, as well as FcγRIII binding (Sondermann et al., Nature 406:267 (2000); Wines et al., J. Immunol. 164:5313 (2000)).

[97] Several groups have described the relevance of EU index positions 330 and 331 (amino acid residues 134 and 135 of SEQ ID NO:6) in complement C1q binding and subsequent complement fixation (Canfield and Morrison, J. Exp. Med. 173:1483 (1991); Tao et al., J. Exp. Med. 178:661 (1993)). Amino acid substitutions at these positions were introduced in Fc4 to reduce complement fixation. The C_{H3} domain of Fc4 is identical to that found in the corresponding wild-type polypeptide, except for the stop codon, which was changed from TGA to TAA to eliminate a potential dam methylation site when the cloned DNA is grown in dam plus strains of E. coli.

[98] In Fc5, the arginine residue at EU index position 218 was mutated back to a lysine, because the BglII cloning scheme was not used in fusion proteins containing this particular Fc. The remainder of the Fc5 sequence matches the above description for Fc4.

[99] Fc6 is identical to Fc5 except that the carboxyl terminal lysine codon has been eliminated. The C-terminal lysine of mature immunoglobulins is often removed from mature immunoglobulins post-translationally prior to secretion from B-cells, or removed during serum circulation. Consequently, the C-terminal lysine residue is typically not found on circulating antibodies. As in Fc4 and Fc5 above, the stop codon in the Fc6 sequence was changed to TAA.

[100] Fc7 is identical to the wild-type γ1 Fc except for an amino acid substitution at EU index position 297 located in the C_{H2} domain. EU index position Asn-297 is a site of N-linked carbohydrate attachment. N-linked carbohydrate introduces a potential source of variability in a recombinantly expressed protein due to potential batch-to-batch variations in the carbohydrate structure. In an attempt to eliminate this potential variability, Asn-297 was

mutated to a glutamine residue to prevent the attachment of N-linked carbohydrate at that residue position. The carbohydrate at residue 297 is also involved in Fc binding to the FcRIII (Sondermann et al., Nature 406:267 (2000)). Therefore, removal of the carbohydrate should decrease binding of recombinant Fc7 containing fusion proteins to the FcγRs in general. As above, the stop codon in the Fc7 sequence was mutated to TAA.

[101] Fc8 is identical to the wild-type immunoglobulin γ1 region shown in SEQ ID NO:6, except that the cysteine residue at EU index position 220 was replaced with a serine residue. This mutation eliminated the cysteine residue that normally disulfide bonds with the immunoglobulin light chain constant region. The use of any of these specific Fc domains for formation of the BLyS and/or APRIL antagonist is within the scope of the present invention.

[102] Leucine zipper forms of these molecules are also contemplated by the invention. "Leucine zipper" is a term in the art used to refer to a leucine rich sequence that enhances, promotes, or drives dimerization ortrimerization of its fusion partner (e. g. , the sequence or molecule to which the leucine zipper is fused or linked to). Various leucine zipper polypeptides have been described in the art. See, e. g., Landschulz et al., Science, 240: 1759 (1988); US Patent 5,716,805; WO 94/10308; Hoppe et al., FEBS Letters, 344: 1991 (1994); Maniatis et al., Nature, 341: 24 (1989). Those skilled in the art will appreciate that a leucine zipper sequence may be fused at either the 5'or 3'end of the polypeptide of this invention.

[103] The polypeptides of the present invention can also be modified in a way to form chimeric molecules by fusing the polypeptide to another, heterologous polypeptide or amino acid sequence.

[104] According to some embodiments, such heterologous polypeptide or amino acid sequence is one which acts to oligimerize the chimeric molecule. In some embodiments, such a chimeric molecule comprises a fusion of the polypeptide with a tag polypeptide which provides an epitope to which a tag antibody can selectively bind. The epitope tag is generally placed at the amino-or carboxyl-terminus of the polypeptide.

[105] The presence of such epitope-tagged forms of the polypeptide can be detected using an antibody against the tag polypeptide. Also, provision of the epitope tag enables the polypeptide to be readily purified by affinity purification using a tag antibody or another type of affinity matrix that binds to the epitope tag.

[106] Various tag polypeptides and their respective antibodies are well known in the art. Examples include poly- histidine (poly-his) or poly-histidine-glycine (poly-his-gly) tags;

the flu HA tag polypeptide and its antibody 12CA5 [Field et al. , Mol. Cell. Biol. , 8: 2159-2165 (1988)] ; the c-myc tag and the 8F9,3C7,6E10, G4, B7 and9E10 antibodies thereto [Evan etal., Molecular and Cellular Biology, 5: 3610-3616 (1985)] ; and the Herpes Simplex virus glycoprotein D (gD) tag and its antibody [Paborsky etal., Protein Engineering, 3 (6): 547-553 (1990)]. Other tag polypeptides include the Flag-peptide [Hopp et al., BioTechnology, 6: 1204-1210 (1988)] ; the KT3 epitope peptide [Martin etal., Science, 255: 192-194 (1992)] ; an"-tubulin epitope peptide [Skinner et al. , J. Biol. Chem. , 266: 15163-15166 (1991)] ; and the T7 gene 10 protein peptide tag [Lutz-Freyermuth et al. , Proc. Natl. Acad.Sci : USA, 87 : 6393-6397 (1990)].

Construction of Peptide-Polymer Conjugates

[107] In some embodiments the strategy for the conjugation of a polymer, (e. g, PEGylation) of synthetic peptides consists of combining, through forming a conjugate linkage in solution, a peptide and a PEG moiety, each bearing a special functionality that is mutually reactive toward the other. The peptides can be easily prepared with conventional solid phase synthesis. The peptides are "preactivated" with an appropriate functional group at a specific, site. The precursors are purified and fully characterized prior to reacting with the PEG moiety. Ligation of the peptide with PEG usually takes place in aqueous phase and can be easily monitored by reverse phase analytical HPLC. The PEGylated peptides can be easily purified by preparative HPLC and characterized by analytical HPLC, amino acid analysis and laser desorption mass spectrometry.

[108] In some embodiments, a peptide is covalently bonded via one or more of the amino acid residues of the peptide to a terminal reactive group on the polymer, depending mainly on the reaction conditions, the molecular weight of the polymer, etc. The polymer with the reactive group (s) is designated herein as activated polymer. The reactive group selectively reacts with free amino or other reactive groups on the peptide. Potential reactive sites include: N-terminal amino group, epsilon amino groups on lysine residues, as well as other amino, imino, carboxyl, sulfhydryl, hydroxyl, and other hydrophilic groups. It will be understood, however, that the type and amount of the reactive group chosen, as well as the type of polymer employed, to obtain optimum results, will depend on the particular peptide employed to avoid having the reactive group react with too many particularly active groups on the peptide. In some embodiments, a reactive residue, (e. g., lysine (K), a modified, non-

natural amino acid, or other small molecule) may be substituted at a position suitable for conjugation.

[109] In some embodiments, the peptide comprises the sequence of Formula I, FormulaII, Formula III, ECFDLLVRAWVPCSVLK (SEQ ID NO 5), ECFDLLVRHWVPCGLLR (SEQ ID NO 6), ECFDLLVRRWVPCEMLG (SEQ ID NO 7), ECFDLLVRSWVPCHMLR (SEQ ID NO 8), ECFDLLVRHWVACGLLR (SEQ ID NO 9), or sequences listed in FIG. 32 of WO 05/000351 have a terminal reactive group.

[110] In some embodiments, the peptide comprises at least one and more preferably, more than one of a polypeptide comprising a sequence of Formula I, FormulaII, Formula III, ECFDLLVRAWVPCSVLK (SEQ ID NO 5), ECFDLLVRHWVPCGLLR (SEQ ID NO 6), ECFDLLVRRWVPCEMLG (SEQ ID NO 7), ECFDLLVRSWVPCHMLR (SEQ ID NO 8), ECFDLLVRHWVACGLLR (SEQ ID NO 9), or sequences listed in FIG. 32 of WO 05/000351. The polypeptides that are linked together can have the same sequence or have different sequences and a terminal reactive group. In some embodiments, these polypeptides can be joined to one another, optionally, through the use of a linker.

[111] While conjugation may occur at any reactive amino acid on the polypeptide, in some embodiments, the reactive amino acid is lysine, which is linked to the reactive group of the activated polymer through its free epsilon-amino group, or glutamic oraspartic acid, which is linked to the polymer through an amide bond. In some embodiments, the reactive amino acids of the peptide are not cysteine residues at positions X2 and Xlz.

[112] The degree of polymer conjugation with each peptide will vary depending upon the number of reactive sites on the peptide, the molecular weight, hydrophilicity and other characteristics of the polymer, and the particular peptide derivatization sites chosen. In some embodiments, the conjugate has a final molar ratio of 1 to 10 polymer molecules per peptide molecule, but greater numbers of polymer molecules attached to the peptides of the invention are also contemplated. Insomeiembodiments, each conjugate contains one polymer molecule. The desired amount of derivatization is easily achieved by using an experimental matrix in which the time, temperature and other reaction conditions are varied to change the degree of substitution, after which the level of polymer substitution of the conjugates is determined by size exclusion chromatography or other means known in the art.

[113] In some embodiments, the polymer contains only a single group which is reactive. This helps to avoid cross-linking of protein molecules. However, it is within the scope herein to maximize reaction conditions to reduce cross-linking, or to purify the reaction

products through gel filtration or ion exchange chromatography to recover substantially homogenous derivatives. In other embodiments, the polymer contains two or more reactive groups for the purpose of linking multiple peptides to the polymer backbone.

[114] Again, gel filtration or ion exchange chromatography can be used to recover the desired derivative in substantially homogeneous form. In some embodiments, the polymer is covalently bonded directly to the peptide without the use of a multifunctional (ordinarily bifunctional) crosslinking agent. In some embodiments, there is a 1: 1 molar ratio of PEG chain to peptide.

[115] The covalent modification reaction may take place by any appropriate method generally used for reacting biologically active materials with inert polymers, preferably at about pH 5-9, more preferably 7-9 if the reactive groups on the peptide are lysine groups. Generally, the process involves preparing an activated polymer (the polymer typically having at least one terminal hydroxyl group to be activated), preparing an active substrate from this polymer, and thereafter reacting the peptide with the active substrate to produce the peptide suitable for formulation. The above modification reaction can be performed by several methods, which may involve one or more steps. Examples of modifying agents that can be used to produce the activated polymer in a one-step reaction include cyanuric acid chloride (2,4,6-trichloro-S- triazine) and cyanuric acid fluoride.

[116] In some embodiments, the modification reaction takes place in two steps wherein the polymer is reacted first with an acid anhydride such as succinic or glutaric anhydride to form a carboxylic acid, and the carboxylic acid is then reacted with a compound capable of reacting with the carboxylic acid to form an activated polymer with a reactive ester group that is capable of reacting with the peptide. Examples of such compounds include N-hydroxysuccinimide, 4-hydroxy-3-nitrobenzene sulfonic acid, and the like, and preferably N-hydroxysuccinimide or 4-hydroxy-3-nitrobenzene sulfonic acid is used. For example, monomethyl substituted PEG may be reacted at elevated temperatures, preferably about100-110 C for four hours, with glutaric anhydride. The monomethyl PEG-glutaric acid thus produced is then reacted with N- hydroxysuccinimide in the presence of a carbodiimide reagent such as dicyclohexyl or isopropyl carbodiimide to produce the activated polymer, methoxypolyethylene glycolyl-N-succinimidyl glutarate, which can then be reacted with the GH. This method is described in detail in Abuchowski etal., Cancer Biochem. Biophys. , 7: 175-186 (1984). In another example, the monomethyl substituted PEG may be reacted with glutaric anhydride followed by reaction with 4-hydroxy-3-nitrobenzene sulfonic acid

(HNSA) in the presence of dicyclohexyl carbodiimide to produce the activated polymer. HNSA is described by Bhatnagar et al. , Peptides:Synthesis-Structure-Func-tion. Proceedings of the Seventh American Peptide Symposium, Rich et al. (eds.) (Pierce Chemical Co. , RockfordIll., 1981), p. 97-100, and in Nitecki et al., High-Technology Route to Virus Vaccines (American Society for Microbiology: 1986) entitled"Novel Agent for Coupling Synthetic Peptides to Carriers and Its Applications."

[117] In some embodiments, covalent binding to amino groups is accomplished by known chemistries based upon cyanuric chloride, carbonyl diimidazole, aldehyde reactive groups (PEG alkoxide plus diethyl acetal of bromoacetaldehyde; PEG plus DMSO and acetic anhydride, or PEG chloride plus the phenoxide of 4-hydroxybenzaldehyde, activated succinimidyl esters, activated dithiocarbonate PEG, 2,4, 5- trichlorophenylcloroformate or P-nitrophenylcloroformate activated PEG. Carboxyl groups are derivatized by coupling PEG-amine using carbodiimide.Sulfhydryl groups are derivatized by coupling to maleimido-substituted PEG (e. g. alkoxy-PEG amine plus sulfosuccinimidyl 4- (N-maleimidomethyl) cyclohexane-l-carboxylate) as described in WO97/10847 published Mar. 27,1997, or PEG-maleimide commercially available from Nektar Technologies, San Carlos, CA (formerly Shearwater Polymers, Inc.). Alternatively, free amino groups on the peptide (e. g. epsilon amino groups on lysine residues) may be coupled to N-hydroxysucciminidyl substituted PEG (PEG-NHS available from Nektar Technologies;) or can be thiolated with 2-imino-thiolane (Traut's reagent) and then coupled to maleimide- containing derivatives of PEG as described in Pedley et al. , Br. J. Cancer, 70: 1126-1130 (1994).

[118] Many inert polymers, including but not limited to PEG, are suitable for use in pharmaceuticals. See, e. g. , Davis et al. , Biomedical Polymers: Polymeric Materials and Pharmaceuticals for Biomedical Use, pp. 441-451 (1980). In some embodiments of the invention, a non-proteinaceous polymer is used. The nonproteinaceous polymer is typicially a hydrophilic synthetic polymer, i. e. , a polymer not otherwise found in nature. However, polymers which exist in nature and are produced by recombinant or in vitro methods are also useful, as are polymers which are isolated from native sources. Hydrophilic polyvinyl polymers fall within the scope of this invention, e. g. polyvinylalcohol and polyvinylpyrrolidone. Particularly useful are polyalkylene ethers such as polyethylene glycol (PEG); polyoxyalkylenes such as polyoxyethylene, polyoxypropylene, and block copolymers of polyoxyethylene and polyoxypropylene (Pluronics) ; polymethacrylates; carbomers; branched or unbranched polysaccharides which comprise the saccharide monomers D-

mannose, D-and L-galactose, fucose, fructose, D-xylose, L-arabinose, D-glucuronic acid, sialic acid, D-galacturonic acid, D-mannuronic acid (e. g. polymannuronic acid, or alginic acid), D- glucosamine, D-galactosamine, D-glucose andneuraminic acid including homopolysaccharides and heteropolysaccharides such as lactose, amylopectin, starch, hydroxyethyl starch, amylose, dextrane sulfate, dextran, dextrins, glycogen, or the polysaccharide subunit of acid mucopolysaccharides, e. g. hyaluronic acid; polymers of sugar alcohols such as polysorbitol and polymannitol; heparin orheparon.

The polymer prior to conjugation need not be, but preferably is, water soluble, but the final conjugate is preferably water-soluble. Preferably, the conjugate exhibits a water solubility of at least about 0.01 mg/ml, and more preferably at least about 0.1 mg/ml, and still more preferably at least about 1 mg/ml.

[119] In addition, the polymer should not be highly immunogenic in the conjugate form, nor should it possess viscosity that is incompatible with intravenous infusion, injection, or inhalation if the conjugate is intended to be administered by such routes.

[120] The molecular weight of the polymer can range up to about 100, 000 D, and preferably is at least about 500 D, or at least about 1,000 D, or at least about 5,000 D. In some embodiments, the PEG or other polymer has a molecular weight in the range of 5000 to 20,000 D. The molecular weight chosen can depend upon the effective size of the conjugate to be achieved, the nature (e.g. structure, such as linear or branched) of the polymer, and the degree of derivatization, i. e. the number of polymer molecules per peptide, and the polymer attachment site or sites on the peptide. In some embodiments, branched PEG's may used to induce a large increase in effective size of the peptides. PEG or other polymer conjugates may be utilized to increase half-life, increase solubility, stabilize against proteolytic attack, and reduceimmunogenicity.

[121] Functionalized PEG polymers to modify the peptides of the invention are available from Nektar Technologies of San Carlos, CA (formerly Shearwater Polymers, Inc.). Such commercially available PEG derivatives include, but are not limited to, amino-PEG, PEG amino acid esters. PEG-N-hydroxysuccinamide chemistry(NHS), PEG-hydrazide, PEG-thiol, PEG-succinate, carboxymethylated PEG, PEG-propionic acid, PEG amino acids, PEG succinimidylsuccinate, PEG succinimidyl propionate, succinimidyl ester of carboxymethylated PEG, succinimidyl carbonate of PEG, succinimidyl esters of amino acid PEGs, PEG-xycarbonylimidazole, PEG-nitrophenyl carbonate, PEG tresylate, PEG-glycidyl ether, PEG-aldehyde, PEG vinylsulfone, PEG-maleimide, PEG-orthopyridyl-disulfide,

heterofunctional PEGs, PEG vinyl derivatives, PEG silanes, and PEG phospholides. The reaction conditions for coupling these PEG derivatives will vary depending on the protein, the desired degreeof PEGylation, and the PEG derivative utilized. Some factors involved in the choice of PEG derivatives include: the desired point of attachment (such as lysine or cysteine R-groups), hydrolytic stability and reactivity of the derivatives, stability, toxicity and antigenicity of the linkage, suitability for analysis, etc. Specific instructions for the use of any particular derivative are available from the manufacturer.

[122] The conjugates may be characteized by SDS-PAGE, gel filtration, NMR, tryptic mapping, liquid chromatrography-mass spectrophotometry, and in vitro biological assays. For example, the extent of PEG conjugation may be shown by SDS-PAGE and gel filtration, and then analyzed by NMR, which has a specific resonance peak for the methylene hydrogens of PEG. The number of PEG groups on each molecule can be calculated from the NMR spectrum or mass spectrometry. Polyacrylamide gelelectrophoresis in 10% SDS is appropriately run in 10 mMTris-HCI pH 8.0, 100 mMNaCI as elution buffer. To demonstrate which residue is PEGylated, tryptic mapping can be performed. Thus, PEGylated peptides are digested with trypsin at the protein/enzyme ratio of 100 to 1 in mg basis at37 C for 4 hours in 100 mM sodium acetate, 10 mMTris-HCI, 1 mM calcium chloride, pH 8.3, and acidified to pH < 4 to stop digestion before separating on HPLC NucleosilC-18 (4.6 mm. times.150 mm,5. mu. ,100A). Thechromatogram is compared to that ofnon-PEGylated starting material. Each peak can then be analyzed by mass spectrometry to verify the size of the fragment in the peak. The fragment (s) that carried PEG groups are usually not retained on the HPLC column after injection and disappear from the chromatograph. Such disappearance from the chromatograph is an indication ofPEGylation on that particular fragment that should contain at least one lysine residue. PEGylated peptides may then be assayed for ability to bind to the BLyS by conventional methods.

[123] In some embodiments, conjugates are purified by ion-exchange chromatography, (e. g, ion- exchange HPLC. The chemistry of many of the electrophilically activated PEG's results in a reduction of amino group charge of the PEGylated product. Thus, high resolution ion exchange chromatography can be used to separate the free and conjugated proteins, and to resolvespecies-', with different levels ofPEGylation.

[124] In fact, the resolution of different species (e. g. containing one or two PEG residues) is also possible due to the difference in the ionic properties of the unreacted amino acids. In one embodiment, species with difference levels ofPEGylation are resolved

according to the methods described in WO 96/34015 (International Application No.PCT/US96/05550 published Oct. 31, 1996). Heterologous species of the conjugates are purified from one another in the same fashion.

[125] In some embodiments, PEG-N-hydroxysuccinamide (NHS) reacts with a primary amine (e. g. lysines and the N-terminus). In some embodiments, PEG-NHS reacts with a C-terminal lysine (K) of the polypeptide. In some embodiments, the lysine residue is added to the C-terminus of the 17-mer polypeptide, while in other embodiments, Xi is substituted with lysine. In some embodiments, the polymer reacts with the N-terminus. In a preferred embodiment, the conjugate is generated by utilizing the derivatization and purification methods described in the Examples below.

[126] In one aspect, the invention provides any of the above-described conjugates formed by its component parts, i. e. one or more peptide (s) covalently attached to one or more polymer molecule (s), without any extraneous matter in the covalent molecular structure of the conjugate.

[127] The methods and articles of manufacture of the present invention use, or incorporate, an antibody which binds to BLyS or one or more of its three receptors. Accordingly, methods for generating such antibodies will be described here.

[128] The BLyS or BLyS receptor to be used for production of, or screening for, antibodies may be, e. g., a soluble form of the antigen or a portion thereof, containing the desired epitope. As described above, the BLyS sequence and the sequence of the BLyS receptors are known as are the boundaries of the various domains of these polypeptides. Peptide fragments of the extracellular domain (ECD) can be used as immunogens. Based on these known sequences and domain delineations, one of skill in the art can express the BLyS or BLyS receptors polypeptides and fragments thereof for use to produce antibodies.

[129] To generate antibodies to BLyS or its receptors, full length polypeptides or peptide fragments of 6 or greater residues in length can be used as immunogens to raise antibodies in rodents including mice, hamsters, and rats, in rabbit, goat, or other suitable animal. Soluble BLyS or BLyS receptor polypeptide or immunogenic fragments thereof can be expressed is suitable host cells such as bacteria or eukaryotic cells. In one embodiment, human and murine detergent-solubilized full-length BLyS are produced in E. coli and used to immunize and screen for hybridomas producing BLyS antibodies.

[130] Alternatively, or additionally, B cells or cell lines expressing BLyS or BLyS receptors at their cell surface can be used to generate, and/or screen for, antibodies. Other

forms of BLyS useful for generating antibodies will be apparent to those skilled in the art, such as phage display methodology can also be used to produce BLyS binding antibody. The antibodies that bind BLyS or the BLyS receptors may be chimeric, humanized, or human. Such antibodies and methods of generating them are described in more detail below.

[131] Polyclonal antibodies are preferably raised in animals by multiple subcutaneous (sc) or intraperitoneal (ip) injections of the relevant antigen and an adjuvant. It may be useful to conjugate the relevant antigen to a protein that is immunogenic in the species to be immunized, e.g., keyhole limpet hemocyanin, serum albumin, bovine thyroglobulin, or soybean trypsin inhibitor using a bifunctional or derivatizing agent, for example, maleimidobenzoylsulfosuccinimide ester (conjugation through cysteine residues), N-hydroxysuccinimide (through lysine residues), glutaraldehyde, succinic anhydride,SOC12, orR1N=C=NR, where R and R'are different alkyl groups.

[132] Animals are immunized against the antigen, immunogenic conjugates, or derivatives by combining, e. g., 100wu or 5g of the protein or conjugate (for rabbits or mice, respectively) with 3 volumes of Freund's complete adjuvant and injecting the solution intradermally at multiple sites. One month later the animals are boosted with 1/5 to 1/10 the original amount of peptide or conjugate in Freund's complete adjuvant by subcutaneous injection at multiple sites. Seven to 14 days later the animals are bled and the serum is assayed for antibody titer. Animals are boosted until the titer plateaus. Preferably, the animal is boosted with the conjugate of the same antigen, but conjugated to a different protein and/or through a different cross-linking reagent. Conjugates also can be made in recombinant cell culture as protein fusions. Also, aggregating agents such as alum are suitably used to enhance the immune response.

[133] Monoclonal antibodies are obtained from a population of substantially homogeneous antibodies, i. e., the individual antibodies comprising the population are identical except for possible naturally occurring mutations that may be present in minor amounts. Thus, the modifier "monoclonal" indicates the character of the antibody as not being a mixture of discrete antibodies.

[134] For example, the monoclonal antibodies may be made using the hybridoma method first described by Kohler etal., Nature, 256: 495 (1975), or may be made by recombinant DNA methods (U. S. Patent No. 4,816,567). In the hybridoma method, a mouse or other appropriate host animal, such as a hamster, is immunized as hereinabove described to elicit lymphocytes that produce or are capable of producing antibodies that will specifically

bind to the protein used for immunization. Alternatively, lymphocytes may be immunized in vitro. Lymphocytes then are-fused with myeloma cells using a suitable fusing agent, such as polyethylene glycol, to form a hybridoma cell (Goding, Monoclonal Antibodies: Principles and Practice, pp. 59-103 (Academic Press, 1986)).

[135] The hybridoma cells thus prepared are seeded and grown in a suitable culture medium that preferably contains one or more substances that inhibit the growth or survival of the unfused, parental myeloma cells. For example, if the parental myeloma cells lack the enzyme hypoxanthine guanine phosphoribosyl transferase (HGPRT or HPRT), the culture medium for the hybridomas typically will include hypoxanthine, aminopterin, and thymidine (HAT medium), which substances prevent the growth of HGPRT-deficient cells.

[136] Preferred myeloma cells are those that fuse efficiently, support stable high-level production of antibody by the selected antibody-producing cells, and are sensitive to a medium such as HAT medium.

[137] Among these, preferred myeloma cell lines are murine myeloma lines, such as those derived from MOPC- 21 andMPC-11 mouse tumors available from the Salk Institute Cell Distribution Center, San Diego, California USA, and SP-2 or X63-Ag8-653 cells available from the American Type Culture Collection, Rockville, Maryland USA. Human myeloma and mouse-human heteromyeloma cell lines also have been described for the production of human monoclonal antibodies (Kozbor, J.Immunol., 133: 3001 (1984); Brodeur etal., Monoclonal Antibody Production Techniques and Applications, pp. 51-63 (Marcel Dekker, Inc. , New York, 1987)).

[138] Culture medium in which hybridoma cells are growing is assayed for production of monoclonal antibodies directed against the antigen. Preferably, the binding specificity of monoclonal antibodies produced by hybridoma cells is determined by immunoprecipitation or by an in vitro binding assay, such as radioimmunoassay (RIA) or enzyme-linked immunoabsorbent assay (ELISA). The binding affinity of the monoclonal antibody can, for example, be determined by the Scatchard analysis of Munson et al., Anal. Biochem., 107: 220 (1980).

[139] After hybridoma cells are identified that produce antibodies of the desired specificity, affinity, and/or activity, the clones may besubcloned by limiting dilution procedures and grown by standard methods (Goding, Monoclonal Antibodies Principles and Practice, pp. 59-103 (Academic Press, 1986)). Suitable culture media for this purpose

include, for example, D-MEM or RPMI-1640 medium. In addition, the hybridoma cells may be grown in vivo as ascites tumors in an animal.

[140] The monoclonal antibodies secreted by the subclones are suitably separated from the culture medium, ascites fluid, or serum by conventional immunoglobulin purification procedures such as, for example, protein A-Sepharose, hydroxylapatite chromatography, gel electrophoresis, dialysis, or affinity chromatography. DNA encoding the monoclonal antibodies is readily isolated and sequenced using conventional procedures (e. g. , by using oligonucleotide probes that are capable of binding specifically to genes encoding the heavy and light chains of murine antibodies). The hybridoma cells serve as a preferred source of such DNA. Once isolated, the DNA may be placed into expression vectors, which are then transfected into host cells such as E. coli cells, simian COS cells, Chinese Hamster Ovary (CHO) cells, or myeloma cells that do not otherwise produce immunoglobulin protein, to obtain the synthesis of monoclonal antibodies in the recombinant host cells. Review articles on recombinant expression in bacteria of DNA encoding the antibody include Skerra et al., Curr. Opinion in Immunol., 5: 256-262 (1993) and Pluckthun, Immunol Revs., 130: 151-188 (1992).

[141] In a further embodiment, antibodies or antibody fragments can be isolated from antibody phage libraries generated using the techniques described in McCafferty et al., Nature, 348:552-554 (1990).

[142] Clacksonet al., Nature, 352: 624-628 (1991) and Markset al., J. Mol. Biol., 222: 581-597 (1991) describe the isolation of murine and human antibodies, respectively, using phage libraries. Subsequent publications describe the production of high affinity (nM range) human antibodies by chain shuffling (Marks et al.,BiolZ'echraology, 10: 779-783 (1992)), as well as combinatorial infection and in vivo recombination as a strategy for constructing very large phage libraries (Waterhouse et aL, Nuc. Acids. Res., 21: 2265-2266 (1993)). Thus, these techniques are viable alternatives to traditional monoclonal antibody hybridoma techniques for isolation of monoclonal antibodies.

[143] The DNA also may be modified, for example, by substituting the coding sequence for human heavy-and light-chain constant domains in place of the homologous murine sequences (U. S. Patent No. 4,816,567; Morrison, et al., Proc. Natl Acad. Sci. USA, 81: 6851 (1984)), or by covalently joining to the immunoglobulin coding sequence all or part of the coding sequence for a non-immunoglobulin polypeptide.

[144] Typically such non-immunoglobulin polypeptides are substituted for the constant domains of an antibody, or they are substituted for the variable domains of one antigen-combining site of an antibody to create a chimeric bivalent antibody comprising one antigen-combining site having specificity for an antigen and another antigen-combining site having specificity for a different antigen.

[145] Methods for humanizing non-human antibodies have been described in the art. Preferably, a humanized antibody has one or more amino acid residues introduced into it from a source which is non- human. These non-human amino acid residues are often referred to as "import" residues, which are typically taken from an "import" variable domain. Humanization can be essentially performed following the method of Winter and co-workers (Jones et al., Nature, 321: 522-525 (1986); Riechmann etal., Nature, 332: 323-327 (1988); Verhoeyen et aL, Science, 239: 1534-1536 (1988)), by substituting hypervariable region sequences for the corresponding sequences of a human antibody. Accordingly, such"humanized" antibodies are chimeric antibodies (U. S. Patent No. 4,816, 567) wherein substantially less than an intact human variable domain has been substituted by the corresponding sequence from a non-human species. In practice, humanized antibodies are typically human antibodies in which some hypervariable region residues and possibly some FR residues are substituted by residues from analogous sites in rodent antibodies.

[146] The choice of human variable domains, both light and heavy, to be used in making the humanized antibodies is very important to reduce antigenicity. According to the so-called "best-fit" method, the sequence of the variable domain of a rodent antibody is screened against the entire library of known human variable-domain sequences. The human sequence which is closest to that of the rodent is then accepted as the human framework region (FR) for the humanized antibody (Sims etal., J. Immunol., 151: 2296 (1993); Chothia etal., J. Mol. Biol., 196: 901 (1987)). Another method uses a particular framework region derived from the consensus sequence of all human antibodies of a particular subgroup of light or heavy chains. The same framework may be used for several different humanized antibodies (Carter etal., Proc. Natl. Acad. Sci. USA, 89: 4285 (1992); Presta et al., J. Immunol., 15-1 :2623 (1993)).

[147] It is further important that antibodies be humanized with retention of high affinity for the antigen and other favorable biological properties. To achieve this goal, according to a preferred method, humanized antibodies are prepared by a process of analysis

of the parental sequences and various conceptual humanized products using three-dimensional models of the parental and humanized sequences.

[148] Three-dimensional immunoglobulin models are commonly available and are familiar to those skilled in the art. Computer programs are available which illustrate and display probable three-dimensional conformational structures of selected candidate immunoglobulin sequences. Inspection of these displays permits analysis of the likely role of the residues in the functioning of the candidate immunoglobulin sequence, i.e., the analysis of residues that influence the ability of the candidate immunoglobulin to bind its antigen. In this way, FR residues can be selected and combined from the recipient and import sequences so that the desired antibody characteristic, such as increased affinity for the target antigen (s), is achieved. In general, the hypervariable region residues are directly and most substantially involved in influencing antigen binding.

[149] As an alternative to humanization, human antibodies can be generated. For example, it is now possible to produce transgenic animals (e. g., mice) that are capable, upon immunization, of producing a full repertoire of human antibodies in the absence of endogenous immunoglobulin production. For example, it has been described that the homozygous deletion of the antibody heavy-chain joining region (JH) gene in chimeric and germ-line mutant mice results in complete inhibition of endogenous antibody production.

[150] Transfer of the human germ-line immunoglobulin gene array in such germ-line mutant mice will result in the production of human antibodies upon antigen challenge. See, e. g., Jakobovits et al., Proc. Natl. Acad. Sci. USA, 90: 2551 (1993); Jakobovits etal., Nature, 362: 255-258 (1993); Bruggermann etal., Year in Immuno., 7: 33 (1993); and US Patent Nos. 5,591, 669,5, 589,369 and 5,545, 807.

[151] Alternatively, phage display technology (McCafferty etal., Nature 348: 552-553 (1990)) can be used to produce human antibodies and antibody fragments in vitro, from immunoglobulin variable (V) domain gene repertoires from unimmunized donors. According to this technique, antibody V domain genes are cloned in-frame into either a major or minor coat protein gene of a filamentous bacteriophage, such as M13 or fd, and displayed as functional antibody fragments on the surface of the phage particle. Because the filamentous particle contains a single-stranded DNA copy of the phage genome, selections based on the functional properties of the antibody also result in selection of the gene encoding the antibody exhibiting those properties. Thus, the phage mimics some of the properties of the B cell.

[152] Phage display can be performed in a variety of formats; for their review see, e. g., Johnson, Kevin S. and Chiswell, David J., Current Opinion in Structural Biology 3: 564-571 (1993). Several sources of V-gene segments can be used for phage display. Clackson et al., Nature, 352: 624-628 (1991) isolated a diverse array of anti-oxazolone antibodies from a small random combinatorial library of V genes derived from the spleens of immunized mice. A repertoire of V genes from unimmunized human donors can be constructed and antibodies to a diverse array of antigens (including self-antigens) can be isolated essentially following the techniques described by Marks etal., J. Mol. Biol. 222: 581-597 (1991), or Griffith etal., EMBO J. 12: 725-734 (1993). See, also, US Patent-Nos. 5,565, 332 and 5,573, 905. Human antibodies may also be generated by in vitro activated B cells (see US Patents 5,567, 610 and 5,229, 275).

[153] Various techniques have been developed for the production of antibody fragments. Traditionally, these fragments were derived via proteolytic digestion of intact antibodies (see, e. g., Morimoto etal., Journal of Biochemical and Biophysical Methods 24: 107-117 (1992) and Brennan et al., Science, 229: 81 (1985)). However, these fragments can now be produced directly by recombinant host cells. For example, the antibody fragments can be isolated from the antibody phage libraries discussed above. Alternatively, Fab'-SH fragments can be directly recovered from E. coli and chemically coupled to form F (ab') 2 fragments (Carter et al., Bio/Technology 10: 163-167 (1992)). According to another approach, F (ab')2 fragments can be isolated directly from recombinant host cell culture. Other techniques for the production of antibody fragments will be apparent to the skilled practitioner. In other embodiments, the antibody of choice is a single chain Fv fragment (scFv). See WO 93/16185; US Patent No. 5,571,894; and US Patent No. 5,587, 458. The antibody fragment may also be a "linear antibody,", e. g., as described in US Patent 5,641,870 for example. Such linear antibody fragments may be monospecific or bispecific.

[154] Bispecific antibodies are antibodies that have binding specificities for at least two different epitopes. Exemplary bispecific antibodies may bind to two different epitopes of the B cell surface marker. Other such antibodies may bind a first B cell marker and further bind a second B cell surface marker. Alternatively, an anti-B cell marker binding arm may be combined with an arm which binds to a triggering molecule on a leukocyte such as a T-cell receptor molecule (e. g. CD2 or CD3), or Fc receptors for IgG (FcyR), such as FcyRI (CD64),FcyRII (CD32) and FcyRIII (CD16) so as to focus cellular defense mechanisms to the B cell. Bispecific antibodies may also be used to localize cytotoxic agents to the B cell.

[155] These antibodies possess a B cell marker-binding arm and an arm which binds the cytotoxic agent (e. g. saporin, anti-interferon-, vinca alkaloid, ricin A chain, methotrexate or radioactive isotope hapten).

[156] Bispecific antibodies can be prepared as full length antibodies or antibody fragments (e. g. F (ab') 2 bispecific antibodies). Methods for making bispecific antibodies are known in the art. Traditional production of full length bispecific antibodies is based on the coexpression of two immunoglobulin heavy chain-light chain pairs, where the two chains have different specificities (Millstein et aL, Nature, 305: 537-539 (1983)).

[157] Because of the random assortment of immunoglobulin heavy and light chains, these hybridomas (quadromas) produce a potential mixture of 10 different antibody molecules, of which only one has the correct bispecific structure. Purification of the correct molecule, which is usually done by affinity chromatography steps, is rather cumbersome, and the product yields are low. Similar procedures are disclosed in WO93/08829, and in Traunecker et al., EMBO J. , 10: 3655-3659 (1991).

[158] According to a different approach, antibody variable domains with the desired binding specificities (antibody-antigen combining sites) are fused to immunoglobulin constant domain sequences. The fusion preferably is with an immunoglobulin heavy chain constant domain, comprising at least part of the hinge, CH2, and CH3 regions. It is preferred to have the first heavy-chain constant region(CH1) containing the site necessary for light chain binding, present in at least one of thefusions. DNAs-encoding the immunoglobulin heavy chain fusions and, if desired, the immunoglobulin light chain, are inserted into separate expression vectors, and are co-transfected into a suitable host organism. This provides for great flexibility in adjusting the mutual proportions of the three polypeptide fragments in embodiments when unequal ratios of the three polypeptide chains used in the construction provide the optimum yields. It is, however, possible to insert the coding sequences for two or all three polypeptide chains in one expression vector when the expression of at least two polypeptide chains in equal ratios results in high yields or when the ratios are of no particular significance.

[159] In a preferred embodiment of this approach, the bispecific antibodies are composed of a hybrid immunoglobulin heavy chain with a first binding specificity in one arm, and a hybrid immunoglobulin heavy chain-light chain pair (providing a second binding specificity) in the other arm. It was found that this asymmetric structure facilitates the separation of the desired bispecific compound from unwanted immunoglobulin chain

combinations, as the presence of an immunoglobulin light chain in only one half of the bispecific molecule provides for a facile way of separation. This approach is disclosed in WO 94/04690. For further details of generating bispecific antibodies see, for example, Suresh et al., Methods in Enzymology, 121: 210 (1986). According to another approach described in US Patent No. 5,731,168, the interface between a pair of antibody molecules can be engineered to maximize the percentage of heterodimers which are recovered from recombinant cell culture. The preferred interface comprises at least a part of the CH3 domain of an antibody constant domain. In this method, one or more small amino acid side chains from the interface of the first antibody molecule are replaced with larger side chains (e. g. tyrosine or tryptophan). Compensatory"cavities"of identical or similar size to the large side chain (s) are created on the interface of the second antibody molecule by replacing large amino acid side chains with smaller ones (e. g. alanine or threonine). This provides a mechanism for increasing the yield of the heterodimer over other unwanted end-products such as homodimers.

[160] Bispecific antibodies include cross-linked or "heteroconjugate" antibodies. For example, one of the antibodies in the heteroconjugate can be coupled to avidin, the other to biotin. Such antibodies have, for example, been proposed to target immune system cells to unwanted cells (US Patent No. 4,676, 980), and for treatmentof HIV infection (WO 91/00360, WO 92/200373, and EP 03089). Heteroconjugate antibodies may be made using any convenient cross-linking methods. Suitable cross-linking agents are well known in the art, and are disclosed in US Patent No. 4,676,980, along with a number of cross-linking techniques.

[161] Techniques for generating bispecific antibodies from antibody fragments have also been described in the literature. For example, bispecific antibodies can be prepared using chemical linkage. Brennan et al, Science, 229: 81 (1985) describe a procedure wherein intact antibodies are proteolytically cleaved to generate F (ab') 2 fragments. These fragments are reduced in the presence of the dithiol complexing agent sodium arsenite to stabilize vicinal dithiols and prevent intermolecular disulfide formation. The Fab' fragments generated are then converted to thionitrobenzoate (TNB) derivatives. One of the Fab'-TNB derivatives is then reconverted to the Fab'-thiol by reduction with mercaptoethylamine and is mixed with an equimolar amount of the other Fab'-TNB derivative to form the bispecific antibody. The bispecific antibodies produced can be used as agents for the selective immobilization of enzymes.

[162] Recent progress has facilitated the direct recovery of Fab'-SH fragments from E. coli, which can be chemically coupled to form bispecific antibodies. Shalaby et al., J. Exp. Med., 175: 217-225 (1992) describe the production of a fully humanized bispecific antibody F (ab') 2 molecule. Each Fab'fragment was separately secreted from E. coli and subjected to directed chemical coupling in vitro to form the bispecific antibody. The bispecific antibody thus formed was able to bind to cells overexpressing the ErbB2 receptor and normal human T cells, as well as trigger the lytic activity of human cytotoxic lymphocytes against human breast tumor targets.

[163] Various techniques for making and isolating bispecific antibody fragments directly from recombinant cell culture have also been described. For example, bispecific antibodies have been produced using leucine zippers. Kostelny et al., J. Immunol., 148 (5): 1547-1553 (1992). The leucine zipper peptides from the Fos and Jun proteins were linked to the Fab'portions of two different antibodies by gene fusion.

[164] The antibody homodimers were reduced at the hinge region to form monomers and then re-oxidized to form the antibody heterodimers. This method can also be utilized for the production of antibody homodimers.

[165] The "diabody" technology described by Hollinger et al, Proc. Natl. Acad. Sci. USA, 90: 6444-6448 (1993) has provided an alternative mechanism for making bispecific antibody fragments. The fragments comprise a heavy-chain variable domain (VH) connected to a light-chain variable domain (VL) by a linker which is too short to allow pairing between the two domains on the same chain. Accordingly, the VH and VL domains of one fragment are forced to pair with the complementary VL and VH domains of another fragment, thereby forming two antigen-binding sites. Another strategy for making bispecific antibody fragments by the use of single-chain Fv (sFv) dimers has also been reported. See Gruber etal.,J. ImmunoL, 152: 5368 (1994).

[166] Antibodies with more than two valencies are contemplated. For example, trispecific antibodies can be prepared. Tuttet al., J. Immunol. 147: 60(1991).

[167] Amino acid sequence modification (s) of protein or peptide antagonists and antibodies described herein are contemplated. For example, it may be desirable to improve the binding affinity and/or other biological properties of the BLYS binding antibody or antagonist. Amino acid sequence variants of the antagonist are prepared by introducing appropriate nucleotide changes into the antagonist nucleic acid, or by peptide synthesis. Such modifications include, for example, deletions from,and/or insertions intoand/or substitutions

of, residues within the amino acid sequences of the antagonist. Any combination of deletion, insertion, and substitution is made to arrive at the final construct, provided that the final construct possesses the desired characteristics. The amino acid changes also may alter post-translational processes of the antagonist, such as changing the number or position of glycosylation sites.

[168] A useful method for identification of certain residues or regions of the antagonist that are preferred locations for mutagenesis is called "alanine scanning mutagenesis" as described by Cunningham and Wells Science, 244: 1081-1085 (1989). Here, a residue or group of target residues are identified(e. g. , charged residues such as arg, asp, his, lys, and glu) and replaced by a neutral or negatively charged amino acid (most preferably alanine or polyalanine) to affect the interaction of the amino acids with antigen. Those amino acid locations demonstrating functional sensitivity to the substitutions then are refined by introducing further or other variantsat, or for,-the sites of substitution. Thus, while the site for introducing an amino acid sequence variation is predetermined, the nature of the mutation per se need not be predetermined. For example, to analyze the performance of a mutation at a given site, tala scanning or random mutagenesis is conducted at the target codon or region and the expressed antagonist variants are screened for the desired activity.

[169] Amino acid sequence insertions include amino-and/or carboxyl-terminal fusions ranging in length from one residue to polypeptides containing a hundred or more residues, as well as intrasequence insertions of single or multiple amino acid residues. Examples of terminal insertions include an antagonist with an N- terminal methionyl residue or the antagonist fused to a cytotoxic polypeptide. Other insertional variants of the antagonist molecule include the fusion to the N-or C-terminus of the antagonist of an enzyme, or a polypeptide which increases the serum half-life of the antagonist.

[170] Another type of variant is an amino acid substitution variant. These variants have at least one amino acid residue in the antagonist molecule replaced by different residue. The sites of greatest interest for substitutional mutagenesis of antibody antagonists include the hypervariable regions, but FR alterations are also contemplated. Conservative substitutions are shown in Table 1 under the heading of "preferred substitutions". If such substitutions result in a change in biological activity, then more substantial changes, denominated"exemplary substitutions"in Table 1, or as further described below in reference to amino acid classes, may be introduced and the products screened.

[171] Substantial modifications in the biological properties of the antagonist are accomplished by selecting substitutions that differ significantly in their effect on maintaining (a) the structure of the polypeptide backbone in the area of the substitution, for example, as a sheet or helical conformation, (b) the charge or hydrophobicity of the molecule at the target site, or (c) the bulk of the side chain. Naturally occurring residues are divided into groups based on common side-chain properties: (1) hydrophobic: norleucine, met, ala, val, leu, ile; (2) neutral hydrophilic: cys, ser, thr; (3) acidic: asp, glu; (4) basic: asn, gln, his, lys, arg; (5) residues that influence chain orientation: gly, pro; and (6) aromatic: trp, tyr, phe. Non-conservative substitutions will entail exchanging a member of one of these classes for another class.

[172] Any cysteine residue not involved in maintaining the proper conformation of the antagonist also may be substituted, generally with serine, to improve the oxidative stability of the molecule and prevent aberrant crosslinking. Conversely, cysteine bond (s) may be added to the antagonist to improve its stability (particularly where the antagonist is an antibody fragment such as an Fv fragment).

[173] A particularly preferred type of substitutional variant involves substituting one or more hypervariable region residues of a parent antibody. Generally, the resulting variant (s) selected for further development will have improved biological properties relative to the parent antibody from which they are generated. A convenient way for generating such substitutional variants is affinity maturation using phage display. Briefly, several hypervariable region sites (e. g. 6-7 sites) are mutated to generate all possible amino substitutions at each site. The antibody variants thus generated are displayed in a monovalent fashion from filamentous phage particles as fusions to the gene III product ofM13 packaged within each particle. The phage-displayed variants are then screened for their biological activity (e. g. binding affinity) as herein disclosed. In order to identify candidate hypervariable region sites for modification, alanine scanning mutagenesis can be performed to identify hypervariable region residues contributing significantly to antigen binding. Alternatively, or in additionally, it may be beneficial to analyze a crystal structure of the antigen-antibody complex to identify contact points between the antibody and antigen. Such contact residues and neighboring residues are candidates for substitution-according to the techniques elaborated herein. Once such variants are generated, the panel of variants is subjected to screening as described herein and antibodies with superior properties in one or more relevant assays may be selected for further development.

[174] Another type of amino acid variant of the antagonist alters the original glycosylation pattern of the antagonist. By altering is meant deleting one or more carbohydrate moieties found in the antagonist, and/or adding one or more glycosylation sites that are not present in the antagonist.

[175] Glycosylation of polypeptides is typically either N-linked or 0-linked. N-linked refers to the attachment of the carbohydrate moiety to the side chain of an asparagine residue. The tripeptide sequences asparagine-X- serine and asparagine-X-threonine, where X is any amino acid except proline, are the recognition sequences for enzymatic attachment of the carbohydrate moiety to the asparagine side chain. Thus, the presence of either of these tripeptide sequences in a polypeptide creates a potential glycosylation site. 0-linked glycosylation refers to the attachment of one of the sugars N-aceylgalactosamine, galactose, or xylose to a hydroxyamino acid, most commonly serine or threonine, although 5-hydroxyproline or 5-hydroxylysine may also be used.

[176] Addition of glycosylation sites to the antagonist is conveniently accomplished by altering the amino acid sequence such that it contains one or more of the above-described tripeptide sequences (forN- linked glycosylation sites). The alteration may also be made by the addition of, or substitution by, one or more serine or threonine residues to the sequence of the original antagonist (for 0-linked glycosylation sites).

[177] Nucleic acid molecules encoding amino acid sequence variants of the antagonist are prepared by a variety of methods known in the art. These methods include, but are not limited to, isolation from a natural source (in the case of naturally occurring amino acid sequence variants) or preparation by oligonucleotide- mediated (or site-directed) mutagenesis, PCR mutagenesis, and cassette mutagenesis of an earlier prepared variant or a non-variant version of the antagonist.

[178] It may be desirable to modify the antagonist of the invention with respect to effector function, e. g. so as to enhance antigen-dependent cell-mediatedcyotoxicity (ADCC) and/or complement dependent cytotoxicity (CDC) of the antagonist. This may be achieved by introducing one or more amino acid substitutions in an Fc region of an antibody antagonist. Alternatively or additionally, cysteine residue (s) may be introduced in the Fc region, thereby allowing interchain disulfide bond formation in this region.

[179] The homodimeric antibody thus generated may have improved internalization capability and/or increased complement-mediated cell killing and antibody-dependent cellular cytotoxicity (ADCC). See Caron et aL, J. Exp Med. 176: 1191-1195 (1992) and

Shopes, B.J. Immunol. 148: 2918-2922 (1992). Homodimeric antibodies with enhanced anti-tumor activity may also be prepared using heterobifunctional cross-linkers as described in Wolff etal. Cancer Research 53: 2560-2565 (1993). Alternatively, an antibody can be engineered which has dual Fc regions and may thereby have enhanced complement lysis and ADCC capabilities. See Stevenson etal. Anti-Cancer Drug Design 3: 219-230 (1989).

[180] To increase the serum half life of the antagonist, one may incorporate a salvage receptor binding epitope into the antagonist (especially an antibody fragment) as described in US Patent 5,739, 277, for example. As used herein, the term"salvage receptor binding epitope"refers to an epitope of the Fc region of an IgG molecule (e.g.,-IgGl, IgG2, IgG3, or IgG4) that-is responsible for increasing the in vivo serum half- life of the IgG molecule.

Assays

[181] Peripheral B-cell concentrations are determined by a FACS method that count CD3-/CD40+ cells.

[182] The percent of CD3-CD40+ B cells of total lymphocytes in samples can be obtained by the following gating strategy. The lymphocyte population is marked on the forward scatter/side scatter scattergram to define Region 1(Ri). Using events inRI, fluorescence intensity dot plots are displayed for CD40 and CD3 markers. Fluorescently labeled isotype controls are used to determine respective cutoff points for CD40 and CD3 positivity.

FACS analysis

[183] Half million cells are washed and resuspended in 1001 of FACS buffer, which is phosphate buffered saline with 1% BSA, containing5, ul of staining or control antibody. All the staining antibodies, including isotype controls, are obtained from PharMingen, San Diego, CA. Human BLYS expression is assessed by staining withRituxan@ along with FITC-conjugated anti-humanIgGl secondary antibody.

[184] FACS analysis is conducted using FACScan and Cell Quest (Becton Dickinson Immunocytometry Systems, San Jose, CA). All the lymphocytes are defined in the forward and side light scatterings, while all the B lymphocytes are defined with the expression of B220 on the cell surface.

[185] B cell depletion and recovery are assessed by analyzing peripheral B cell counts and analysis of hBLYS+ B cells by FACS in the spleen, lymph node and bone marrow

on a daily basis for the first week after injection and thereafter on a weekly basis. Serum levels of the injected 2H7 variant antibody are monitored.

Pharmaceutical Formulations

[186] Therapeutic formulations of the BLyS and/or APRIL antagonists such as BLyS-binding antibodies used in accordance with the present invention are prepared for storage by mixing an antibody having the desired degree of purity with optional pharmaceutically acceptable carriers, excipients or stabilizers (Remitgtorz's Phamamaceutical Science 16th edition, Osol, A. Ed. (1980)), in the form of lyophilized formulations or aqueous solutions. Acceptable carriers, excipients, or stabilizers are nontoxic to recipients at the dosages and concentrations employed, and include buffers such as phosphate, citrate, and other organic acids; antioxidants including ascorbic acid and methionine; preservatives (such as octadecyldimethylbenzyl ammonium chloride; hexamethonium chloride; benzalkonium chloride, benzethonium chloride; phenol, butyl or benzyl alcohol; alkyl parabens such as methyl or propyl paraben; catechol; resorcinol; cyclohexanol; 3-pentanol; and m-cresol); low molecular weight (less than about 10 residues) polypeptides; proteins, such as serum albumin, gelatin, or immunoglobulins; hydrophilic polymers such as olyvinylpyrrolidone; amino acids such as glycine, glutamine, asparagine, histidine, arginine, or lysine ; monosaccharides, disaccharides, and other carbohydrates including glucose, mannose, or dextrins; chelating agents such as EDTA; sugars such as sucrose, mannitol, trehalose or sorbitol; salt-forming counter-ions such as sodium; metal complexes (e. g. Zn-protein complexes);and/or non-ionic surfactants such as TWEEN, PLURONICSTM or polyethylene glycol (PEG).

[187] The formulation herein may also contain more than one active compound as necessary for the particular indication being treated, preferably those with complementary activities that do not adversely affect each other. For example, it may be desirable to further provide a cytotoxic agent, chemotherapeutic agent, cytokine or immunosuppressive agent (e. g. one which acts on T cells, such as cyclosporin or an antibody that binds T cells, e. g. one which binds LFA-1). The effective amount of such other agents depends on the amount of antibody present in the formulation, the type of disease or disorder or treatment, and other factors discussed above. These are generally used in the same dosages and with administration routes as described herein or about from 1 to 99% of the heretofore employed dosages.

[188] The active ingredients may also be entrapped in microcapsules prepared, for example, by coacervation techniques or by interfacial polymerization, for example, hydroxymethylcellulose or gelatin- microcapsules and poly- (methylmethacylate) microcapsules, respectively, in colloidal drug delivery systems (for example, liposomes, albumin microspheres, microemulsions, nano-particles and nanocapsules) or in macroemulsions. Such techniques are disclosed in Remington's Pharmaceutical Sciences 16th edition, Osol, A. Ed. (1980).

[189] Sustained-release preparations may be prepared. Suitable examples of sustained-release preparations include semi-permeable matrices of solid hydrophobic polymers containing the antagonist, which matrices are in the form of shaped articles, e. g. films, or microcapsules. Examples of sustained- release matrices include polyesters, hydrogels (for example, poly (2-hydroxyethyl-methacrylate), or poly (vinylalcohol)), polylactides (U. S. Pat. No. 3,773, 919), copolymers of L-glutamic acid and.ethyl-L-glutamate, non-degradable ethylene-vinyl acetate, degradable lactic acid-glycolic acid copolymers such as the LUPRON DEPOT (injectablemicrospheres composed of lactic acid-glycolic acid copolymer and leuprolide acetate), and poly-D- (-)-3-hydroxybutyric acid.

[190] The formulations to be used for in vivo administration must be sterile. This is readily accomplished by filtration through sterile filtration membranes.

Disease Treatment

Diseases

[191] The immunosuppressive drugs and the BLyS and/or APRIL antagonists of the invention are useful to treat B-cell regulated autoimmune disorders. B-cell regulated autoimmune diseases include arthritis (rheumatoid arthritis, juvenile rheumatoid arthritis, osteoarthritis, psoriatic arthritis), psoriasis, dermatitis including atopic dermatitis; chronic autoimmune urticaria, polymyositis/dermatomyositis, toxic epidermal necrolysis, systemic scleroderma and sclerosis, responses associated with inflammatory bowel disease (IBD) (Crohn's disease, ulcerative colitis), respiratory distress syndrome, adult respiratory distress syndrome (ARDS), meningitis, allergic rhinitis, encephalitis, uveitis, colitis, glomerulonephritis, allergic conditions, eczema, asthma, conditions involving infiltration of T cells and chronic inflammatory responses, atherosclerosis, autoimmune myocarditis, leukocyte adhesion deficiency, systemic lupus erythematosus (SLE), lupus (including nephritis, non-renal, discoid, alopecia), juvenile onset diabetes, multiple sclerosis, allergic

encephalomyelitis, immune responses associated with acute and delayed hypersensitivity mediated by cytokines and T-lymphocytes, tuberculosis, sarcoidosis, granulomatosis including Wegener's granulomatosis, agranulocytosis, vasculitis (including ANCA), aplastic anemia, Coombs positive anemia, Diamond Blackfan anemia, immune hemolytic anemia including autoimmune hemolytic anemia (AIHA), pernicious anemia, pure red cell aplasia (PRCA), Factor VIII deficiency, hemophilia A, autoimmune neutropenia, pancytopenia, leukopenia, diseases involving leukocyte diapedesis, CNS inflammatory disorders, multiple organ injury syndrome, myasthenia gravis, antigen-antibody complex mediated diseases, anti-glomerular basement membrane disease, anti- phospholipid antibody syndrome, allergic neuritis, Bechet disease, Castleman's syndrome,Goodpasture's Syndrome,Lambert-Eaton Myasthenic Syndrome, Reynaud's syndrome, Sjorgen's syndrome, Stevens- Johnson syndrome, solid organ transplant rejection (including pretreatment for high panel reactive antibody titers, IgA deposit in tissues, etc), graft versus host disease (GVHD), pemphigoid bullous, pemphigus (all including vulgaris, foliaceus), autoimmune polyendocrinopathies, Reiter's disease, stiff-man syndrome, giant cell arteritis, immune complex nephritis, IgA nephropathy, IgM polyneuropathies or IgM mediated neuropathy, idiopathic thrombocytopenic purpura (ITP), thrombotic throbocytopenic purpura(TTP), autoimmune thrombocytopenia, autoimmune disease of the testis and ovary including autoimune orchitis and oophoritis, primary hypothyroidism; autoimmune endocrine diseases including autoimmune thyroiditis, chronic thyroiditis (Hashimoto's Thyroiditis), subacute thyroiditis, idiopathic hypothyroidism, Addison's disease, Grave's disease, autoimmune polyglandular syndromes (or polyglandular endocrinopathy syndromes), Type I diabetes also referred to as insulin-dependent diabetes mellitus (IDDM) and Sheehan's syndrome; autoimmune hepatitis, Lymphoid interstitial pneumonitis (HIV), bronchiolitis obliterans (non- transplant) vs NSIP,Guillain-Barre'Syndrome, Large Vessel Vasculitis (including Polymyalgia Rheumatica and Giant Cell (Takayasu's) Arteritis), Medium Vessel Vasculitis (includingKawasaki's Disease and Polyarteritis Nodosa), ankylosing spondylitis, Berger's Disease (IgA nephropathy), Rapidly Progressive Glomerulonephritis, Primary biliary cirrhosis, Celiac sprue (gluten enteropathy), Cryoglobulinemia, ALS, coronary artery disease.

[192] The desired level of B-cell depletion will depend on the disease. Preferably, the B cell depletion is sufficient to prevent progression of disease for at least 2 months, more preferably 3 months, even more preferably 4 months, more preferably 5 months, even more preferably 6 or more months. In even more preferred embodiments, the B cell depletion is

sufficient to increase the time in remission by at least 6 months, more preferably 9 months, more preferably one year, more preferably 2 years, more preferably 3 years, even more preferably 5 or more years. In a most preferred embodiment, the B cell depletion is sufficient to cure the disease. In preferred embodiments, the B cell depletion in the autoimmune patient is at least transiently about 75% and more preferably, 80%, 85%, 90%, 95%, 99% and even 100% of the baseline level before treatment.

[193] For treatment of an autoimmune disease, it may be desirable to modulate the extent of B cell depletion depending on the disease and/or the severity of the condition in the individual patient, by adjusting the dosage of the immunosuppressive drug or the BLyS and/or APRIL antagonist. Thus, B-cell depletion can but does not have to be complete. Or, total B cell-depletion may be desired in initial treatment but in subsequent treatments, the dosage may be adjusted to achieve only partial depletion. In one embodiment, the B cell depletion is at least 20%, i. e., 80% or less of B-cells remain as compared to the baseline level before treatment. In other embodiments, B-cell depletion is 25%, 30%, 40%, 50%, 60%, 70% or greater.

[194] Preferably, the B cell depletion is sufficient to halt progression of the disease, more preferably to alleviate the signs and symptoms of the particular disease under treatment, even more preferably to cure the disease.

[195] For therapeutic applications, the immunosuppressive drug and BLyS and/or APRIL antagonist compositions of the invention can be used in combination therapy with additional drugs such as anti-inflammatory drugs, including DMARDS and other biologics. The preceding treatment methods can be administered in conjunction with other forms of conventional therapy for autoimmune disease, either consecutively with, pre-or post-conventional therapy.

[196] A patient is alleviated or successfully treated of a B cell regulated autoimmune diseases by the present methods of the invention if there is a measurable-improvement in the symptoms or other applicable criteria after administration of the compositions of the invention compared to before treatment. The effect of treatment may be apparent within 3-10 weeks after administration of the compositions of the invention. The applicable criteria for each disease will be well known to the physician of skill in the appropriate art. For example, the physician can monitor the treated patient for clinical, or serologic evidence of disease such as serologic markers of disease, complete blood count including B cell count, and serum immunoglobulin levels. Serum levels of IgG and IgM are reduced in BLyS and/or APRIL

antagonist, such as atacicept, treated mice. Human patients responding to actacicept treatment likewise show a reduction in serum IgG and IgM levels.

[197] The parameters for assessing efficacy or success of treatment of an autoimmune or autoimmune related disease will be known to the physician of skill in the appropriate disease. Generally, the physician of skill will look for reduction in the signs and symptoms of the specific disease. The following are by way of examples.

[198] Rheumatoid arthritis (RA) is an autoimmune disorder of unknown etiology. Most RA patients suffer a chronic course of disease that, even with therapy, may result in progressive joint destruction, deformity, disability and even premature death. The goals of RA therapy are to prevent or control joint damage, prevent-loss of function and decreasepain. Initial therapy of RA usually involves administration of one or more of the following drugs: nonsteroidal anti-inflammatory drugs (NSAIDs), glucocorticoid (via joint injection), and low-dose prednisone. See"Guidelines for the management of rheumatoid arthritis"Arthritis & Rhemmatism 46 (2): 328-346 (February, 2002). The majority of patients with newly diagnosed RA are started with disease-modifying antirheumatic drug (DMARD) therapy within 3 months of diagnosis.

[199] DMARDs commonly used in RA are hydroxycloroquine, sulfasalazine, methotrexate, leflunomide, etanercept, infliximab (plus oral and subcutaneous methrotrexate), azathioprine, D-penicillamine, Gold (oral), Gold (intramuscular), minocycline, cyclosporine, Staphylococcal protein A immunoadsorption.

[200] Because the body produces tumor necrosis factor alpha(TNFa) during RA,TNFa inhibitors have used for therapy of that disease. Etanercept (ENBREL) is an injectable drug approved in the US for therapy of active RA. Etanercept binds toTNFa and serves to remove mostTNFa from joints and blood, thereby preventingTNFa from promoting inflammation and other symptoms of rheumatoid arthritis.

[201] Etanercept is an "Fc-fusion protein" fusion protein consisting of the extracellular ligand binding portion of the human 75 kD (p75) tumor necrosis factor receptor (TNFR) linked to the Fc portion of a humanIg Gl.

[202] Infliximab, sold under the trade name REMICADE, is an immune-suppressing drug prescribed to treat RA and Crohn's disease. Infliximab is a chimeric monoclonal antibody that binds to TNF and reduces inflammation in the body by targeting and binding to TNFα which produces inflammation.

[203] Adalimumab(HUMIRATM, Abbott Laboratories), previously known as D2E7, is a human monoclonal antibody that binds toTNFa and is approved for reducing the signs and symptoms and inhibiting the progression of structural damage in adults with moderately to severely active RA who have had insufficient response to one or more traditional disease modifyingDMARDs.

[204] Treatment of rheumatoid arthritis by administering immunosuppressive drugs and a BLyS and/or APRIL antagonist can be preformed in conjunction with therapy with one or more of the aforementioned drugs for RA. For rheumatoid arthritis, for example, measurements for progress in treatment may include the number of swollen and tender joints and the length of morning stiffness. Patients may be examined for how much the joint in the hands and feet have eroded by using X-rays and a scoring system known as the Sharp score. Another scoring system is based on the American College of Rheumatology criteria for assessing response to therapies.

[205] One method of evaluating treatment efficacy in RA is based on American College of Rheumatology (ACR) criteria, which measures the percentage of improvement in tender and swollen joints, among other things. The RA patient can be scored at for example, ACR 20 (20 percent improvement) compared with no antibody treatment (e.g,, baseline before treatment) or treatment with placebo. Other ways of evaluating the efficacy of antibody treatment include X-ray scoring such as the Sharp X-ray score used to score structural damage such as bone erosion and joint space narrowing. Patients can also be evaluated for the prevention of or improvement in disability based on Health Assessment Questionnaire [HAQ] score, AIMS score, SF-36 at time periods during or after treatment. The ACR 20 criteria may include 20% improvement in both tender (painful) joint count and swollen joint count plus a20% improvement in at least 3 of 5 additional measures:

1. patient's pain assessment by visual analog scale (VAS),
2. patient's global assessment of disease activity (VAS),
3. physician's global assessment of disease activity (VAS),4. patient's self-assessed disability measured by the Health Assessment Questionnaire, and
5. acute phase reactants, CRP or ESR.

[206] The ACR 50 and 70 are defined analogously. Preferably, the patient is administered an amount of a BLYS binding antibody of the invention effective to achieve at least a score of ACR 20, preferably at least ACR 30, more preferably at least ACR50, even more preferably at least ACR70, most preferably at least ACR 75 and higher.

[207] Psoriatic arthritis has unique and distinct radiographic features. For psoriatic arthritis, joint erosion and joint space narrowing can be evaluated by the Sharp score as well. The humanized BLYS binding antibodies disclosed herein can be used to prevent the joint damage as well as reduce disease signs and symptoms of the disorder.

[208] Yet another aspect of the invention is a method of treating Lupus or SLE by administering to the patient suffering from SLE, a therapeutically effective amount of a BLyS and/or APRIL antagonist in combination with an immunosuppressive drug of the invention. SLEDAI scores provide a numerical quantitation of disease activity. The SLEDAI is a weighted index of 24 clinical and laboratory parameters known to correlate with disease activity, with a numerical range of 0-103. See Bryan Gescuk & John Davis, "Novel therapeutic agent for systemic lupus erythematosus" in Current Opinion in Rheumatology 2002,14 : 515-521. Antibodies to double-stranded DNA are believed to cause renal flares and other manifestations of lupus. Patients undergoing antibody treatment can be monitored for time to renal flare, which is defined as a significant, reproducible increase in serum creatinine, urine protein or blood in the urine. Alternatively or in addition, patients can be monitored for levels of antinuclear antibodies and antibodies to double-stranded DNA. Treatments for SLE include high-dose corticosteroids and/or cyclophosphamide (HDCC). For systemic lupus erythematosus, patients can be monitored for levels of antinuclear antibodies and antibodies to double-stranded DNA.

[209] A particular aspect of the present invention is the treatment of lupus nephritis with a combination of immunosuppressive drugs and a BLyS and/or APRIL antagonist, such as atacicept.

[210] Spondyloarthropathies are a group of disorders of the joints, including ankylosing spondylitis, psoriatic arthritis and Crohn's disease. Treatment success can be determined by validated patient and physician global assessment measuring tools.

[211] A further autoimmune disease that can be treated using the methods of the present invention is psoriasis. Various medications are presently used to treat psoriasis; treatment differs directly in relation to disease severity. Patients with a more mild form of psoriasis typically utilize topical treatments, such as topical steroids, anthralin, calcipotriene, clobetasol, and tazarotene, to manage the disease while patients with moderate and severe psoriasis are more likely to employ systemic (methotrexate, retinoids, cyclosporine, PLTVA and UVB) therapies. Tars are also used. These therapies have a combination of safety concerns, time consuming regimens, or inconvenient processes of treatment. Furthermore,

some require expensive equipment and dedicated space in the office setting. Systemic medications can produce serious side effects, including hypertension, hyperlipidemia, bone marrow suppression, liver disease, kidney disease and gastrointestinal upset. Also, the use of phototherapy can increase the incidence of skin cancers. In addition to the inconvenience and discomfort associated with the use of topical therapies, phototherapy and systemic treatments require cycling patients on and off therapy and monitoring lifetime exposure due to their side effects.

[212] Treatment efficacy for psoriasis is assessed by monitoring changes in clinical signs and symptoms of the disease including Physician's Global Assessment (PGA) changes and Psoriasis Area and Severity Index (PASI) scores, Psoriasis Symptom Assessment (PSA), compared with the baseline condition. The patient can be measured periodically throughout treatment on the Visual analog scale used to indicate the degree of itching experienced at specific time points.

[213] Various other inflammatory diseases that have autoimmune causation are also within the scope of the present treatment method. Among the diseases specifically contemplated are autoimmune hepatitis and autoimmune thyroiditis. Autoimmune hepatitis involves inflammation of the liver due to self-attack by the patient's own immune system. Similarily, self-attack and resulting inflammation of the thyroid is the biological cause of autoimmune thyroiditis. Diseases such as this are expected to be eleviated using by the combination of BLyS and/or APRIL antagonist with an immunesuppressive drug such as MMF.

Dosing

[214] Depending on the indication to be treated and factors relevant to the dosing that a physician of skill in the field would be familiar with, the BLyS and/or APRIL antagonists and immunosuppressive drugs of the invention will be administered at a dosage that is efficacious for the treatment of that indication while minimizing toxicity and side effects. Generally, the BLyS and/or APRIL antagonist of the present invention invention will be administered to a human patient at a dosage range of about 0.25 mg/kg to about 25 mg/kg body weight, preferably at about 1 mg/kg to about 10 mg/kg. Alternatively expressed, a preferred range of dosages for the BLyS and/or APRIL antagonist is about 75 to about 190 mg per dose. In a preferred embodiment, the dosage is about 150 mg per dose for the BLyS and/or APRIL antagonist.

[215] The treatment methods of the invention comprises a combination of concurrently and sequentially administering the immunosuppressive drugs and the BLyS

and/or APRIL antagonist (both referred to herein as the treatment moieties). In sequential administration, the treatment moeities can be administered in either order, i. e., immunosuppressive drugs first followed by BLyS and/or APRIL antagonist. The patient can be treated with one drug and monitored for efficacy before treatment with the one drug. For example, if the immunosuppressive drug produces a partial response, treatment can be followed with the BLyS and/or APRIL antagonist to achieve a full response, and vice versa. Alternatively, the patient can be initially administered both drugs and subsequent dosing can be with only one or the other drug.

[216] To condition the patient to tolerate the drugs and/or to reduce the occurrence of adverse effects such as infusion-related symptoms which arise from the initial and subsequent administrations of the therapeutic compound, the mammal in need thereof can be administered a first or initial conditioning dose of one or both drugs and then administered at least a second therapeutically effective dose of one or both drugs wherein the second and any subsequent doses are higher than the first dose. The first dose serves to condition the mammal to tolerate the higher second therapeutic dose. In this way, the mammal is able to tolerate higher doses of the therapeutic compound than could be administered initially. A "conditioning dose" is a dose which attenuates or reduces the frequency or the severity of first dose adverse side effects associated with administration of a therapeutic compound. The conditioning dose may be a therapeutic dose, a sub-therapeutic dose, a symptomatic dose or a sub-symptomatic dose. A therapeutic dose is a dose which exhibits a therapeutic effect on the patient and a sub-therapeutic dose is a dose which dose not exhibit a therapeutic effect on the patient treated. A symptomatic dose is a dose which induces at least one adverse effect on administration and a sub-symptomatic dose is a dose which does not induce an adverse effect. Some adverse effects are fever, headache, nausea, vomiting, breathing difficulties, myalgia, and chills.

[217] Beyond a conditioning dose regimen, there are a number of other regimens that can be followed to achieve the disease alleviation of the presention invention. One such approach is a short treatment course of one immunosuppressive drug, followed by tapering, followed by treatment with the combination of another immunosuppressive drug and the BLyS and/or APRIL antagonist. For example, corticosteroids can be given 1000-1500 mg orally, twice daily for four weeks, followed by tapering of 5 mg/week down to 10 mg/day over the course of 10 weeks. Once it is time to begin the BLyS and/or APRIL antagonist, a loading dose regimen may be appropriate. In particular, atacicept can be administed twice

per week for four weeks followed by weekly administration for an extended period of time, such as 48 weeks. In contrast, the immunosuppressive drug will tend to be administered in an escalating dose regimen. For example, MMF can be administered at a dose of 500 mg twice daily and increased to 750 mg twice daily after two weeks. Weekly advances in dosage can be made up to a maximum dose of 1000 mg 3 times daily if the patient's white blood cell count remains at acceptable levels (i.e., above 3.0 X 10^9/L or 3000/mm^2). In particular, it would be expected that the maximal range of administration of MMF would be from 2000 to 3000 mg per patient per day.

Route of administration

[218] The BLyS and/or APRIL antagonists and the immunosuppressive drugs are administered to a human patient in accord with known methods, such as by intravenous administration, e. g., as a bolus or by continuous infusion over a period of time, by subcutaneous, intramuscular, intraperitoneal, intracerobrospinal, intra-articular, intrasynovial, intrathecal, or inhalation routes. Immunosuppressive drugs with appropriate formulation may also be administered orally or topically. The BLyS and/or APRIL antagonist and some immunosuppressive drugs will generally be administered by intravenous or subcutaneous administration. The different treatment moeities can be administered by the same or different routes.

Articles of Manufacture and Kits

[219] Another embodiment of the invention is an article of manufacture comprising a BLyS and/or APRIL antagonist and immunosuppressive drugs to treat a B-cell regulated autoimmune disorder as disclosed above. In a specific embodiment, the article of manufacture contains actacicept and MMF for the treatment of lupus nephritis.

[220] The article of manufacture comprises at least one container and a label or package insert on or associated with the container. Suitable containers include, for example, bottles, vials, syringes, etc. The containers may be formed from a variety of materials such as glass or plastic. The container holds a composition of the invention which is effective for treating the condition and may have a sterile access port (for example the container may be an intravenous solution bag or a vial having a stopper pierceable by a hypodermic injection needle). The label or package insert indicates that the composition is used for treating the

particular condition, e. g., lupus nephritis or rheumatoid arthritis. The label or package insert will further comprise instructions for administering the composition to the patient. Additionally, the article of manufacture may further comprise a second container comprising a pharmaceutically-acceptable buffer, such as bacteriostatic water for injection (BWFI), phosphate-buffered saline, Ringer's solution and dextrose solution. It may further include other materials desirable from a commercial and user standpoint, including other buffers, diluents, filters, needles, and syringes.

[221] Kits are also provided that are useful for various purposes, e. g., for B-cell killing assays. As with the article of manufacture, the kit comprises a container and a label or package insert on or associated with the container. The container holds a composition comprising at least one immunosuppressive drug and one BLyS and/or APRIL antagonist of the invention. Additional containers may be included that contain, e. g., diluents and buffers, control antibodies. The label or package insert may provide a description of the composition as well as instructions for the intended in vitro or diagnostic use.

Experimental Examples

Example 1

Production of BLyS and/or APRIL antagonist

[222] Four amino terminal truncated versions of TACI-Fc were generated. All four had a modified human tissue plasminogen activator signal sequence as disclosed in WO 02/094852 (SEQ ID NO: 41) fused to amino acid residue number 30 of SEQ ID NO:2. However, the four proteins differed in the location of point in which the Fc5 was fused to the TACI amino acid sequence of SEQ ID NO:2. Table 1 outlines the structures of the four fusion proteins.

Table 1

TACI Fc Fusion Proteins

Designation of TACI-Fc	TACI amino acid residues
TACI(d1-29)-Fc5	30 to 154 of SEQ ID NO:2
TACI(d1-29, d107-154)-Fc5	30 to 106 of SEQ ID NO:2
TACI(d1-29, d111-154)-Fc5	30 to 110 of SEQ ID NO:2
TACI(d1-29, d120-154)-Fc5	30 to 119 of SEQ ID NO:2

[223] Protein encoding expression cassettes were generated by overlap PCR using standard techniques (see, for example, Horton et al., Gene 77:61 (1989)). A nucleic acid molecule encoding TACI and a nucleic acid molecule encoding Fc5 were used as PCR templates. Oligonucleotide primers are identified in Tables 2 and 3.

Table 2
Oligonucleotide Primers Used to Produce TACI Fusion Proteins

Designation of TACI-Fc	Oligonucleotide Designations			
	5' TACI	3' TACI	5' Fc5	3' Fc5
TACI(d1-29)-Fc5	ZC24,903	ZC24,955	ZC24,952	ZC24,946
TACI(d1-29, d107-154)-Fc5	ZC24,903	ZC24,951	ZC24,949	ZC24,946
TACI(d1-29, d111-154)-Fc5	ZC24,903	ZC28,978	ZC28,979	ZC24,946
TACI(d1-29, d120-154)-Fc5	ZC24,903	ZC28,981	ZC28,980	ZC24,946

Table 3
Oligonucleotide Sequences

Primer	Nucleotide Sequence	SEQ ID NO.
ZC24,903	5' TATTAGGCCGGCCACCATGGATGCAATGA 3'	27
ZC24,955	5' TGAAGATTTGGGCTCCTTGAGACCTGGGA 3'	28
ZC24,952	5' TCCCAGGTCTCAAGGAGCCCAAATCTTCA 3'	29
ZC24,946	5' TAATTGGCGCGCCTCTAGATTATTTACCCGGAGACA 3'	30
ZC24,951	5' TGAAGATTTGGGCTCGTTCTCACAGAAGTA 3'	31
ZC24,949	5' ATACTTCTGTGAGAACGAGCCCAAATCTTCA 3'	32
ZC28,978	5' TTTGGGCTCGCTCCTGAGCTTGTTCTCACA 3'	33
ZC28,979	5' CTCAGGAGCGAGCCCAAATCTTCAGACA 3'	34
ZC28,981	5' TTTGGGCTCCCTGAGCTCTGGTGGAA 3'	35
ZC28,980	5' GAGCTCAGGGAGCCCAAATCTTCAGACA 3'	36

[224] The first round of PCR amplifications consisted of two reactions for each of the four amino terminal truncated versions. The two reactions were performed separately using the 5'and 3' TACI oligonucleotides in one reaction, and the 5' and 3' Fc5

oligonucleotides in another reaction for each version. The conditions of the first round PCR amplification were as follows. To a 25 µl final volume was added approximately 200 ng template DNA, 2.5 µl 10x Pfu reaction Buffer (Stratagene), 2 µl of 2.5 mM dNTPs, 0.5 µl of 20 µM each 5' oligonucleotide and 3' oligonucleotide, and 0.5 µl Pfu polymerase (2.5 units, Stratagene). The amplification thermal profile consisted of 94°C for 3 minutes, 35 cycles at 94°C for 15 seconds, 50°C for 15 seconds, 72°C for 2 minutes, followed by a 2 minute extension at 72°C. The reaction products were fractionated by agarose gel electrophoresis, and the bands corresponding to the predicted sizes were excised from the gel and recovered using a QIAGEN QIAQUICK Gel Extraction Kit (Qiagen), according to the manufacturer's instructions.

[225] The second round of PCR amplification, or overlap PCR amplification reaction, was performed using the gel purified fragments from the first round PCR as DNA template. The conditions of the second round PCR amplification were as follows. To a 25 µl final volume was added approximately 10 ng template DNA each of the TACI fragment and the Fc5 fragment, 2.5 µl 10x Pfu reaction Buffer (Stratagene), 2 µl of 2.5 mM dNTPs, 0.5 µl of 20 µM each ZC24,903 (SEQ ID NO:27) and ZC24,946 (SEQ ID NO:30) and 0.5 µl Pfu polymerase (2.5 units, Stratagene). The amplification thermal profile consisted of 94°C for 1 minute, 35 cycles at 94°C for 15 seconds, 55°C for 15 seconds, 72°C for 2 minutes, followed by a 2 minute extension at 72°C. The reaction products were fractionated by agarose gel electrophoresis, and the bands corresponding to the predicted sizes were excised from the gel and recovered using a QIAGEN QIAQUICK Gel Extraction Kit (Qiagen), according to the manufacturer's instructions.

[226] Each of the four versions of the amino terminal truncated TACI-Fc PCR products were separately cloned using Invitrogen's ZEROBLUNT TOPO PCR Cloning Kit following the manufacturer's recommended protocol. Table 4 identifies the nucleotide and amino acid sequences of these TACI-Fc constructs.

Table 4
Sequences of TACI-Fc Variants

Designation of TACI-Fc	SEQ ID Nos.	
	Nucleotide	Amino Acid
TACI(d1-29)-Fc5	18	19
TACI(d1-29, d107-154)-Fc5	20	21
TACI(d1-29, d111-154)-Fc5	22	23
TACI(d1-29, d120-154)-Fc5	24	25

[227] After the nucleotide sequences were verified, plasmids comprising each of the four versions of the amino terminal truncated TACI-Fc fusions were digested with FseI and AscI to release the amino acid encoding segments. The FseI - AscI fragments were ligated into a mammalian expression vector containing a CMV promoter and an SV40 poly A segment. Expression vectors were introduced into Chinese hamster ovary cells as described below.

Example 2
Production of TACI-Fc Proteins by Chinese Hamster Ovary Cells

[228] The TACI-Fc expression constructs were used to transfect, via electroporation, suspension-adapted Chinese hamster ovary (CHO) DG44 cells grown in animal protein-free medium (Urlaub et al., Som. Cell. Molec. Genet. 12:555 (1986)). CHO DG44 cells lack a functional dihydrofolate reductase gene due to deletions at both dihydrofolate reductase chromosomal locations. Growth of the cells in the presence of increased concentrations of methotrexate results in the amplification of the dihydrofolate reductase gene, and the linked recombinant protein-encoded gene on the expression construct.

[229] CHO DG44 cells were passaged in PFCHO media (JRH Biosciences, Lenexa, KS), 4 mM L-Glutamine (JRH Biosciences), and 1x hypothanxine-thymidine supplement (Life Technologies), and the cells were incubated at 37°C and 5% CO_2 in Corning shake flasks at 120 RPM on a rotating shaker platform. The cells were transfected separately with linearized expression plasmids. To ensure sterility, a single ethanol precipitation step was performed on ice for 25 minutes by combining 200 µg of plasmid DNA in an Eppendorf tube with 20 µl of sheared salmon sperm carrier DNA (5' → 3' Inc. Boulder, CO, 10 mg/ml), 22 µl of 3M NaOAc (pH 5.2), and 484 µl of 100% ethanol (Gold Shield Chemical Co., Hayward,

CA). After incubation, the tube was centrifuged at 14,000 RPM in a microfuge placed in a 4°C cold room, the supernatant removed and the pellet washed twice with 0.5 ml of 70% ethanol and allowed to air dry.

[230] The CHO DG44 cells were prepared while the DNA pellet was drying by centrifuging 10^6 total cells (16.5 ml) in a 25 ml conical centrifuge tube at 900 RPM for 5 minutes. The CHO DG44 cells were resuspended into a total volume of 300 µl of PFCHO growth media, and placed in a Gene-Pulser Cuvette with a 0.4 cm electrode gap (Bio-Rad). The DNA, after approximately 50 minutes of drying time, was resuspended into 500 µl of PFCHO growth media and added to the cells in the cuvette so that the total volume did not exceed 800 µl and was allowed to sit at room temperature for 5 minutes to decrease bubble formation. The cuvette was placed in a BioRad Gene Pulser II unit set at 0.296 kV (kilovolts) and 0.950 HC (high capacitance) and electroporated immediately.

[231] The cells were incubated 5 minutes at room temperature before placement in 20 ml total volume of PFCHO media in a CoStar T-75 flask. The flask was placed at 37°C and 5% CO_2 for 48 hours when the cells were then counted by hemocytometer utilizing trypan blue exclusion and put into PFCHO selection media without hypothanxine-thymidine supplement and containing 200 mM methotrexate (Cal Biochem).

[232] Upon recovery of the methotrexate selection process, the conditioned media containing the secreted TACI-Fc proteins were examined by Western Blot analysis.

Example 3
Combination of Atacicept and MMF in mice

[233] Atacicept and MMF (CellCept) co-administration was tested in a 28-day study in CD-1 mice. One hundred fifty animals were distributed among four treatment groups. Group 1 received vehicle control articles used for both atacicept and MMF using the same route and regimen used for each test article. Group 2 received MMF (351 mg/kg) via daily oral gavage. Group 3 received atacicept (20 mg/kg) three times weekly via subcutaneous injection. Group 4 received atacicept (20 mg/kg, 3x/week, SC) and MMF (351 mg/kg, daily, oral), typically administered within 30 minutes of each other. The administered dose of MMF was chosen as the maximum feasible dose in mice, and mimicked the human equivalent dose for a 70 kg human of 29 mg/kg used for liver transplant patients (approximately 2 g/day) using body surface area to scale the human dose to mice.

[234] Animals were sacrificed at baseline (Day -1) and Days 14, 28 (last day of treatment), 42, 56, and 112. Mice were checked daily for mortality, abnormalities, and signs of pain or distress. Body weights were measured and recorded on Day -1 of the study and weekly thereafter, and prior to sacrifice. Blood was collected from sacrificed animals for comprehensive hematology and serum chemistry analyses, flow cytometric analyses of lymphocyte subtypes, and total serum IgG, IgM, and IgA concentrations. Total serum IgG, IgM, and IgA were measured using a Comprehensive necropsies were conducted on sacrificed animals for evaluation of organ weight changes, gross pathology, and histopathology.

[235] Total IgG levels were evaluated using a mouse IgG ELISA quantitation kit (Catalog #E90-131, Bethyl Laboratories, Montgomery, TX), combined with the ELISA Starter Accessory Package (Catalog #E101, Bethyl Laboratories, Montgomery, TX). All liquid handling was performed using the integrated Freedom Evo 150 liquid handling system and Columbus Washer (TECAN, Denmark). Microtiter wells were coated with affinity purified goat anti-mouse IgG and incubated at room temperature for 1 hour. The wells were aspirated, washed with wash solution, and then blocked with blocking solution for 30 minutes. The blocking solution was removed, the plates washed, and samples/standards added in duplicate. After 1 hour, the samples/standards were aspirated and the wells washed with Wash Solution. Horseradish peroxidase-conjugated goat anti-mouse IgG was added, and the wells were incubated at room temperature for 1 hour. Unbound conjugated antibody was removed by washing, and the amount of conjugate remaining in the well was measured by incubation with TMB substrate (Bethyl Laboratories, Montgomery, TX) for 10 minutes. The color reaction was stopped with addition of 2 M H_2SO_4, and the resulting absorbance (OD-450) was measured on a Spectramax 190 microplate reader.

[236] Data calculations for total IgG were performed as follows: the average zero standard (plate blank) OD-450 value was subtracted from each sample and standard, and the duplicate readings were averaged. A standard curve was created by plotting the mean OD for each standard vs. concentration using a four-parameter logistic curve fit. Sample concentrations were obtained by interpolation from the standard curve. The mean sample concentration was multiplied by the dilution factor. Data analyzed using SoftMax®Pro software included OD values, the standard curve(s), the standard deviation and %CV between replicates, and the interpolated value of the unknown(s).

[237] Total IgM levels were evaluated using a mouse IgM ELISA quantitation kit (Catalog #E90-101, Bethyl Laboratories, Montgomery, TX), combined with the ELISA Starter Accessory Package (Catalog #E101, Bethyl Laboratories, Montgomery, TX). All liquid handling was performed using the integrated Freedom Evo 150 liquid handling system and Columbus Washer (TECAN, Denmark). Microtiter wells were coated with affinity purified goat anti-mouse IgM and incubated at room temperature for 1 hour. The wells were aspirated, washed with wash solution, and then blocked with blocking solution for 30 minutes. The blocking solution was removed, the plates washed, and samples/standards added in duplicate. After 1 hour, the samples/standards were aspirated and the wells washed with wash solution. Horseradish peroxidase-conjugated goat anti-mouse IgM was added, and the wells were incubated at room temperature for 1 hour. Unbound conjugated antibody was removed by washing, and the amount of conjugate remaining in the well was measured by incubation with TMB substrate (Bethyl Laboratories, Montgomery, TX) for 10 minutes. The color reaction was stopped with addition of 2 M H_2SO_4, and the resulting absorbance (OD-450) was measured on a Spectramax 190 microplate reader.

Data calculations for total IgM were performed as follows:

[238] The average zero standard (plate blank) OD-450 value was subtracted from each sample and standard, and the duplicate readings were averaged. A standard curve was created by plotting the mean OD for each standard vs. concentration using a four-parameter logistic curve fit. Sample concentrations were obtained by interpolation from the standard curve. The mean sample concentration was multiplied by the dilution factor. Data analyzed with SoftMax®Pro software, included OD values, the standard curve(s), the standard deviation and %CV between replicates, and the interpolated value of the unknown(s).

[239] Total IgA levels were evaluated using a mouse IgA ELISA quantitation kit (Catalog #E90-103, Bethyl Laboratories, Montgomery, TX), combined with the ELISA Starter Accessory Package (Catalog #E101, Bethyl Laboratories, Montgomery, TX). All liquid handling was performed using the integrated Freedom Evo 150 liquid handling system and Columbus Washer (TECAN, Denmark). Microtiter wells were coated with affinity purified goat anti-mouse IgA and incubated at room temperature for 1 hour. The wells were aspirated, washed with wash solution, and then blocked with blocking solution for 30 minutes. The blocking solution was removed, the plates washed, and samples/standards added in duplicate. After 1 hour, the samples/standards were aspirated and the wells washed with wash solution. Horseradish peroxidase-conjugated goat anti-mouse IgM was added, and

the wells were incubated at room temperature for 1 hour. Unbound conjugated antibody was removed by washing, and the amount of conjugate remaining in the well was measured by incubation with TMB substrate (Bethyl Laboratories, Montgomery, TX) for 10 minutes. The color reaction was stopped with addition of 2 M H_2SO_4, and the resulting absorbance (OD-450) was measured on a Spectramax 190 microplate reader.

Data calculations for total IgA were performed as follows:

[240] The average zero standard (plate blank) OD-450 value was subtracted from each sample and standard, and the duplicate readings were averaged. A standard curve was created by plotting the mean OD for each standard vs. concentration using a four-parameter logistic curve fit. Sample concentrations were obtained by interpolation from the standard curve. The mean sample concentration was multiplied by the dilution factor. Data analyzed with SoftMax®Pro software, included OD values, the standard curve(s), the standard deviation and %CV between replicates, and the interpolated value of the unknown(s).

[241] The mean results of the change in immunoglobulin measurements are recorded in Table 5 below and graphically in Figures 1, 2, and 3.

Table 5

Numeric Mean IgG, IgM, and IgA results for Day 28

and Difference from Vehicle (μg/mL)

	Vehicle (V)	MMF	Actacicept (A)	Atacicept + MMF (A + M)
IgG	179	185	75	31
IgM	238	317	76	19
IgA	498	506	105	33

	Δ1 (V – MMF)	Δ2 (V – A)	Δ1 + Δ2	Δ3 (V – (A + M))	Δ3 Compared to Δ1 + Δ2
IgG	-6	104	98	148	148 > 98
IgM	-79	162	83	219	219 > 83
IgA	-8	393	385	464	464 > 385

[242] As can be seen from these calculations, the combination has a synergistic effect on the reduction of immunoglobin for all three types (i.e., the mean change from vehicle with the combination (Δ3) is more than the addition of the mean change from vehicle to MMF alone (Δ1) with the mean change from vehicle to Atacicept alone (Δ2) (or, mathematically, Δ3 > Δ1 + Δ2)).

Conclusion

[243] The experiments herein demonstrated surprising results in that the combination of immunosuppressive drugs and BLyS and/or APRIL antagonists, such as MMF and atacicept, resulted in a synergistic depletion of Ig levels compared to the level of reduction with immunosuppressive drugs and BLyS and/or APRIL antagonists alone.

Example 4

Repeat and Extension of Atacicept and MMF Combination Experiment

[244] Using similar procedures to those described in Example 3 and dosing until day 91, the results of Example 3 were replicated and extended in time to days 35, 91, 126, and 182. The results of this experiment confirmed the synergistic effect of Atacicept and MMF on the mean levels of immunoglobulin for at least one of the three types of immunoglobulin at each time point. The synergistic results are summarized in Tables 6 below.

Table 6

Synergistic Results of the Numeric Mean for IgG, IgM, and IgA results for Extended Experiment

and Difference from Vehicle (μg/mL)

Sample Day	Immunoglobulin Type	Vehicle (V)	MMF	Actacicept (A)	Atacicept + MMF (A + M)
35	IgG	519.7	940.13	261.35	148.59
35	IgM	105.7	141.34	5.54	5.11
91	IgG	697.01	1192.87	191.84	128.22
91	IgA	924.36	1764.55	37.53	0
126	IgA	1113.48	1467.05	817.22	778.05
182	IgM	73.95	53.77	73.75	47.63

Sample Day	Immunoglobulin Type	Δ1 (V – MMF)	Δ2 (V – A)	Δ1 + Δ2	Δ3 (V – (A + M))	Δ3 Compared to Δ1 + Δ2
35	IgG	-420.43	258.35	-162.08	371.11	371.11> -162.08
35	IgM	-35.64	100.16	64.52	100.59	100.59>64.52
91	IgG	-495.86	505.17	9.31	568.79	568.79>9.31
91	IgA	-840.19	886.83	46.64	924.36	924.36>46.64
126	IgA	-353.57	296.26	57.31	335.43	335.43>57.31
182	IgM	20.18	.20	20.38	26.32	26.32>20.38

[245] It should be noted that as selected immunoglobulin subtypes reached maximal suppression, the ability to observe the synergistic effect is masked because of the low absolute numbers. This "masking effect" explains why the synergy was less consistently seen across all three immunoglobulin subtypes at later time points. In sum, these results confirm the synergistic effects of the combination of Atacicept and MMF on the numeric mean of the levels of the indicated immunoglobulin types seen in Example 3 and extend that conclusion over a time course from 35, 91, 126, and 182 days.

[246] For the sake of completeness, the following describes the experimental groups, doses, volumes administered and concentrations used in this repeat and extension experiment:

Table 7

Group	*No of animals*	*Dose (mg/kg/day)*	*Dose route*	*Dose schedule*
1 Vehicle 1 + Vehicle 2	20M + 20F	0 0	s.c. oral	3/week daily
2 Atacicept	20M + 20F	20	s.c.	3/week
3 MMF	20M + 20F	350	oral	daily
4 Atacicept + MMF	20M + 20F	20 350	s.c. oral	3/week daily

Administration regimen

ATACICEPT: every other day, for 3 days/week, by subcutaneous route.
MMF: daily, by oral route.

[247] Group 1 received both the vehicle for ATACICEPT and the vehicle for CellCept at the same treatment frequency as the treated groups. Group 2 was treated with only ATACICEPT

[248] Group 3 received only CellCept. Group 4 received CellCept oral gavage followed by ATACICEPT s.c. injection within a period of approximately 30 minutes. The day of the first treatment was considered day 1 of the study. Throughout the study period clinical observations, hematological and blood chemistry tests, toxicokinetics and additional investigations were carried out.

Total IgG, IgM and IgA determinations/Satellite groups 9, 10, 11 and 12

[249] Additional satellite groups of animals were used for total immunoglobulin determinations and antibody determination. They were treated and weighed according to the same procedures as the main study.

Table 8

Group/sex (N° of animals)	ATACICEPT Doses (mg/kg/day) Subcutaneous	MMF Doses (mg/kg/day) Oral
9 M (15)	0	0
9 F (15)		
10 M (12)	20	n.a.
10 F (12)		
11 M (12)	n.a.	350
11 F (12)		
12 M (12)	20	350
12 F (12)		

Animals were housed in 3 mice/sex/cage.

[250] Blood samples were collected from 3 animals/sex/group (in progressive numerical order) for the determination of IgG, IgM and IgA serum levels at the following times:

from animals of the control group (gr.9): Pre-dose
from animals of groups 9, 10, 11 and 12:

- on day 34 of the 5-week treatment period
- on day 90 of the 13-week treatment period
- on day 34 of the 5-week recovery period
- on day 90 of the 13-week recovery period

[251] The blood (at least 0.8 -1 mL) was collected at sacrifice from the abdominal aorta of mice after they have been completely anaesthetized with diethyl ether.

[252] Samples were collected in tubes without any anticoagulant and allowed to clot for about 1 hour at room temperature. The clot were spun down by centrifugation at 3000 rpm at +4°C for 15 minutes. Blood cells were discarded and the serum banked in 100 μL aliquots (four aliquots for every single analysis) and stored at –80°C until analysis. Total IgG, IgM and IgA levels were determined by validated ELISA methods.

[253] Sacrifice was performed at the end of the 5-week treatment period on 4 animals/sex/group (the first ones in numerical order); at the end of the 13-week treatment period on 8 animals/sex/group (the second ones in numerical order); at the end of the 5-week recovery period on 4 animals/sex/group (the first recovered animals in numerical order); and at the end of the 13-week recovery period on 4 animals/sex/group (the remaining animals).

Conclusion

[254] The extension experiment reported herein support the results of Example 3 where it was surprisingly shown that the combination of immunosuppressive drugs and BLyS and/or APRIL antagonists, such as MMF and atacicept, resulted in a synergistic depletion of Ig levels compared to the level of reduction with immunosuppressive drugs and BLyS and/or APRIL antagonists alone.

References

[255] References cited within this application, including patents, published applications and other publications are herein incorporated by reference.

We claim:

1. A method of reducing immunoglobulin levels in a mammal comprising administering a BLyS antagonist and an immunosuppressive drug.

2. The method of claim 1 wherein the BLyS antagonist is a Fc-fusion protein.

3. The method of claim 2, wherein the Fc-fusion protein is selected from the group consisting of a TACI-Fc-fusion protein comprising the extracellular domain of TACI or a functional fragment thereof, a BAFF-R-Fc-fusion protein comprising the extracellular domain of BAFF-R or a functional fragment thereof, and a BCMA-Fc-fusion protein comprising the extracellular domain of BCMA or a functional fragment thereof.

4. The method of claim 3, wherein the Fc-fusion protein comprises the polypeptide sequence selected from the group consisting of SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, and SEQ ID NO: 25.

5. The method of claim 3, wherein the Fc-fusion protein comprises the polypeptide sequence of SEQ ID NO: 26.

6. The method of claim 1, wherein the BLyS antagonist is a BLyS antibody.

7. The method of claim 6, wherein the BLyS antibody binds BLyS within a region comprising amino acids 162-275 of SEQ ID NO: 8.

8. The method of claim 6, wherein the BLyS antibody is LymphoStat-B.

9. The method of claim 1, wherein the BLyS antagonist is a TACI antibody.

10. The method of claim 1, wherein the TACI antibody binds TACI within a region comprising amino acids selected from the group consisting of 72-109 of SEQ ID NO:2 and 82-222 of SEQ ID NO:2.

11. The method of claim 10, wherein the TACI antibody binds both TACI and BCMA.

12. The method of claim 1, wherein the immunosuppressive drug is selected from the group consisting of cyclophosphamide (CYC), azathioprine (AZA), cyclosporine A (CSA), and mycophenolate mofetil (MMF).

13. The method of claim 12, wherein the immunosuppressive drug is mycophenolate mofetil (MMF).

14. The method of claim 1 wherein the BLyS antagonist and the immunosuppressive drug act synergistically to reduce immunoglobulin levels.

15. The method of claim 1 wherein the immunoglobulin level that is reduced is selected from the group consisting of IgM, IgG, IgA, IgD and IgE.

16. A method of alleviating a B-cell regulated autoimmune disorder comprising administering to a patient suffering from the disorder a therapeutically effective amount of a BLyS antagonist and an immunosuppressive drug.

17. The method of claim 16 wherein the BLyS antagonist is a Fc-fusion protein.

18. The method of claim 17, wherein the Fc-fusion protein is selected from the group consisting of a TACI-Fc-fusion protein comprising the extracellular domain of TACI or a functional fragment thereof, a BAFF-R-Fc-fusion protein comprising the extracellular domain of BAFF-R or a functional fragment thereof, and a BCMA-Fc-fusion protein comprising the extracellular domain of BCMA or a functional fragment thereof.

19. The method of claim 18, wherein the Fc-fusion protein comprises the polypeptide sequence selected from the group consisting of SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, and SEQ ID NO: 25.

20. The method of claim 18, wherein the Fc-fusion protein comprises the polypeptide sequence of SEQ ID NO: 26.

21. The method of claim 16, wherein the BLyS antagonist is a BLyS antibody.

22. The method of claim 16, wherein the BLyS antibody binds BLyS within a region comprising amino acids 162-275 of SEQ ID NO: 8.

23. The method of claim 16, wherein the BLyS antibody is LymphoStat-B.

24. The method of claim 16, wherein the BLyS antagonist is a TACI antibody.

25. The method of claim 16, wherein the TACI antibody binds TACI within a region comprising amino acids selected from the group consisting of 72-109 of SEQ ID NO:2 and 82-222 of SEQ ID NO:2.

26. The method of claim 24, wherein the TACI antibody binds both TACI and BCMA.

27. The method of claim 16, wherein the immunosuppressive drug is selected from the group consisting of cyclophosphamide (CYC), azathioprine (AZA), cyclosporine A (CSA), and mycophenolate mofetil (MMF).

28. The method of claim 27, wherein the immunosuppressive drug is mycophenolate mofetil (MMF).

29. The method of claim 16 wherein the BLyS antagonist and the immunosuppressive drug act synergistically to reduce immunoglobulin levels.

30. The method of claim 16 wherein the immunoglobulin level that is reduced is selected from the group consisting of IgM, IgG, IgA, IgD and IgE.

31. The method of claim 16 wherein the autoimmune disease is selected from the group consisting of rheumatoid arthritis, juvenile rheumatoid arthritis, systemic lupus erythematosus (SLE), lupus nephritis (LN), Wegener's disease, inflammatory bowel disease, idiopathic thrombocytopenic purpuria (ITP), thrombotic thrombocytopenic purpura (TTP), autoimmune thrombocytopenia, multiple sclerosis, psoriasis, IgA nephropathy, IgM polyneuropathies, myasthenia gravis, vasculitis, diabetes, mellitus, Reynauld's syndrome, Sjorgen's syndrome, glomerulonephritis, autoimmune hepatitis, and autoimmune thyroiditis.

32. The method of claim 31 wherein the autoimmune disease is lupus nephritis.

33. The method of claim 16 the BLyS antagonist is administered at a dosage of about 1 to about 2.5 mg/kg and the immunosuppressive drug is administered at a dosage of about 1 to about 4 mg/kg.

34. The method of claim 16 wherein the BLyS antagonist and the immunosuppressive drug is administered in conjunction with therapy using a second immunosuppressive drug selected from the group consisting of nonsteroidal anti-inflammatory drugs (NSAIDs), glucocorticoid, prednisone, and disease-modifying antirheumatic drugs (DMARDs).

35. A composition comprising a BLyS antagonist and an immunosuppressive drug.

36. An article of manufacture comprising a BLyS antagonist and an immunosuppressive drug, and a label wherein the label indicates that the composition is for treating a B cell regulated autoimmune disorder.

37. The method of claim 1 wherein said BLyS antagonist is also an APRIL antagonist.

38. The method of claim 16 wherein said BLyS antagonist is also an APRIL antagonist.

39. The composition of claim 34 wherein said BLyS antagonist is also an APRIL antagonist.

40. The article of manufacture of claim 35 wherein said BLyS antagonist is also an APRIL antagonist.

41. A method of reducing immunoglobulin levels in a mammal comprising administering an APRIL antagonist and an immunosuppressive drug.

42. A method of alleviating a B-cell regulated autoimmune disorder comprising administering to a patient suffering from the disorder a therapeutically effective amount of an APRIL antagonist and an immunosuppressive drug.

43. A composition comprising an APRIL antagonist and an immunosuppressive drug.

44. An article of manufacture comprising an APRIL antagonist and an immunosuppressive drug, and a label wherein the label indicates that the composition is for treating a B cell regulated autoimmune disorder.