

A METHOD OF ISOLATING MESENCHYMAL STEM CELLS FROM UMBILICAL CORD AMNIOTIC MEMBRANE USING A CELL CULTURE MEDIUM

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CROSS-REFERENCE TO RELATED APPLICATIONS

The present application claims the benefit of priority of U.S. Provisional Application No. 62/404,582, filed October 5, 2016, the content of which is hereby incorporated by reference it
10 its entirety for all purposes.

FIELD OF THE INVENTION

The present invention relates to a method of isolating mesenchymal stem cells (or such a stem cell population from the amniotic membrane of umbilical cord, as well as a mesenchymal stem cell population isolated from the amniotic membrane of the umbilical cord. The invention is
15 also directed to a cell culture medium for isolating mesenchymal stem cells from the amniotic membrane of the umbilical cord. The invention is also directed to a pharmaceutical composition and uses of the isolated mesenchymal stem cell population. The invention is also directed to methods of treating a disease or disorder comprising administering a mesenchymal stem cell population or a pharmaceutical composition containing such a mesenchymal stem
20 cell population of the invention to a subject in need thereof.

BACKGROUND OF THE INVENTION

Mesenchymal stem cells isolated from the amniotic membrane of the umbilical cord have been first reported in US patent application 2006/0078993 (leading to granted US patents 9,085,755 and 9,737,568) and the corresponding International patent application WO2006/019357. Since
25 then, the umbilical cord tissue has gained attention as a source of multipotent cells; due to its widespread availability, the umbilical cord and in particular stem cells isolated from the amniotic membrane of the umbilical cord (also referred to as “cord lining stem cells”) have been considered as an excellent alternative source of cells for regenerative medicine. See, Jeschke et al. Umbilical Cord Lining Membrane and Wharton’s Jelly-Derived Mesenchymal

Stem Cells: the Similarities and Differences; The Open Tissue Engineering and Regenerative Medicine Journal, 2011, 4, 21-27.

5 A subsequent study compared the phenotype, proliferation rate, migration, immunogenicity, and immunomodulatory capabilities of human mesenchymal stem cells (MSCs) derived from the amniotic membrane of the umbilical cord (umbilical cord lining (CL-MSCs), umbilical cord blood (CB-MSCs), placenta (P-MSCs), and Wharton's jelly (WJ-MSCs) (Stubbendorf et al, Immunological Properties of Extraembryonic Human Mesenchymal Stromal Cells Derived from Gestational Tissue, STEM CELLS AND DEVELOPMENT Volume 22, Number 19, 10 2013, 2619-2629. Stubbendorf et al concluded that extraembryonic gestational tissue-derived MSC populations show a varied potential to evade immune responses as well as exert immunomodulatory effects. The authors also found that CL-MSCs showed the most promising potential for a cell-based therapy, as the cells showed low immunogenicity, but they also showed enhanced proliferative and migratory potential so that future research should 15 concentrate on the best disease models in which CL-MSCs could be administered.

While mesenchymal stem cells of the amniotic membrane can easily be obtained using the protocol as described in US patent application 2006/0078993 and International patent application WO2006/019357, it would be of advantage for clinical trials with these cord lining 20 MSC to have at hand a method that allows to isolate a population of these cord lining MSC's that is highly homogenous and can thus be used for clinical trials.

Accordingly, it is an object of the invention to provide a method of isolating a population of mesenchymal stem cells from the amniotic membrane of umbilical cord that meets this need. It 25 is thus also an object of the invention to provide a highly homogenous population of mesenchymal stem cells isolated from the amniotic membranme of the umbilical cord.

SUMMARY OF THE INVENTION

This object is accomplished by the methods, the mesenchymal stem population, the respective pharmaceutical composition and cell culture medium having the features of the independent 30 claims.

In a first aspect, the invention provides a method of isolating a mesenchymal stem cell population from the amniotic membrane of the umbilical cord, the method comprising

cultivating umbilical cord tissue in a culture medium comprising DMEM (Dulbecco's modified eagle medium), F12 (Ham's F12 Medium), M171 (Medium 171) and FBS (Fetal Bovine Serum).

5 In a second aspect, the invention provides an isolated mesenchymal stem population of the amniotic membrane of the umbilical cord, wherein at least about 90 % or more cells of the stem cell population express each of the following markers: CD73, CD90 and CD105. Preferably, the isolated mesenchymal stem population lack expression of the following markers: CD34, CD45 and HLA-DR. In embodiments of this second aspect, at least about 91
10 % or more, about 92 % or more, about 92 % or more, about 93 % or more, about 94 % or more, about 95 % or more, about 96 % or more, about 97 % or more, about 98 % or more about 99 % or more cells of the isolated mesenchymal stem cell population express each of CD73, CD90 and CD105. In addition, in these embodiments of the second aspect, at least about 91 % or more, about 92 % or more, about 92 % or more, about 93 % or more, about 94
15 % or more, about 95 % or more, about 96 % or more, about 97 % or more, about 98 % or more about 99 % or more cells of the isolated mesenchymal stem cell population preferably lack expression of the markers CD34, CD45 and HLA-DR. The mesenchymal stem cell population may be obtained by a method of isolating a mesenchymal stem cell population of the first aspect.

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In a third aspect, the invention provides a pharmaceutical composition containing a mammalian cell of (the second aspect of) the invention.

In a fourth aspect, the invention provides a method of making a culture medium for isolating
25 the method comprising mixing to obtain a final volume of 500 ml culture medium:

i. 250 ml of DMEM

ii. 118 ml M171

iii. 118 ml DMEM/F12

iv. 12.5 ml Fetal Bovine Serum (FBS) to obtain a final concentration of 2.5% (v/v).

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In a fifth aspect, the invention provides a cell culture medium obtainable by the method of the fourth aspect.

In a sixth aspect, the invention provides a method of isolating mesenchymal stem cells from the amniotic membrane of the umbilical cord, comprising cultivating amniotic membrane tissue in the culture medium prepared by the method of the fourth aspect.

In a seventh aspect, the invention provides a cell culture medium comprising:

- 5 - DMEM in the final concentration of about 55 to 65 % (v/v),
- F12 in a final concentration of about 5 to 15 % (v/v),
- M171 in a final concentration of about 15 to 30 % (v/v) and
- FBS in a final concentration of about 1 to 8 % (v/v).

- 10 In an eight aspect, the invention provides the use of a cell culture medium of the seventh aspect for the isolation of mesenchymal stem cells from the amniotic membrane of umbilical cord.

- 15 In a ninth aspect, the invention provides the use of a cell culture medium of the seventh aspect for the cultivation of mesenchymal stem cells from the amniotic membrane of umbilical cord.

BRIEF DESCRIPTION OF THE DRAWINGS

The invention will be better understood with reference to the detailed description when considered in conjunction with the non-limiting examples and the drawings, in which:

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Fig. 1 shows the technical information sheet of Lonza for Dulbecco's modified eagle medium, including the catalogue number of the DMEM used for the making of the illustrative example of a medium of the invention (PTT-6) in the Experimental Section;

Fig. 2 shows the technical information sheet of Lonza for Ham's F12 medium;

- 25 **Fig. 3** shows the technical information sheet of Lonza for DMEM:F12 (1:1) medium, including the catalogue number of the DMEM:F12 (1:1) medium used for the making of the illustrative example of a medium of the invention (PTT-6) in the Experimental Section;

- 30 **Fig. 4** shows the technical information sheet of Life Technologies Corporation for M171 medium, including the catalogue number of the M171 mediu used for the making of the illustrative example of a medium of the invention (PTT-6) in the Experimental Section;

Fig. 5 shows the list of ingredients, including their commercial supplier and the catalogue number that have been used in the Experimental Section forthe making of the medium PTT-6.

Fig. 6 shows the results of flow cytometry experiments in which mesenchymal stem cells isolated from the umbilical cord have been analysed for the expression of the mesenchymal stem cell markers CD73, CD90 and CD105. For these experiments, mesenchymal stem cells were isolated from umbilical cord tissue by cultivation of the umbilical cord tissue in three different cultivation media, followed by subculturing of the mesenchymal stem cells in the respective medium. The three following culture media were used in these experiments: a) 90% (v/v/ DMEM supplemented with 10 % FBS (v/v), b) the culture medium PTT-4 described in US patent application 2006/0078993 and the corresponding International patent application WO2006/019357 that consist of 90% (v/v) CMRL1066, and 10% (v/v) FBS (see paragraph [0183] of WO2006/019357 and c) the culture medium of the present invention PPT-6 the composition of which is described herein. In this flow cytometry analysis, two different samples of the cord lining mesenchymal stem cell (CLMC) population were analysed for each of the three used culture media. The results are shown in **Fig. 6a** to **Fig.6c**. In more detail, **Fig. 6a** shows the percentage of isolated mesenchymal cord lining stem cells expressing stem cell markers CD73, CD90 and CD105 after isolation from umbilical cord tissue and cultivation in DMEM/10% FBS, **Fig. 6b** shows the percentage of isolated mesenchymal cord lining stem cells expressing stem cell markers CD73, CD90 and CD105 after isolation from umbilical cord tissue and cultivation in PTT-4 and **Fig. 6c** shows the percentage of isolated mesenchymal cord lining stem cells expressing stem cell markers CD73, CD90 and CD105 after isolation from umbilical cord tissue and cultivation in PTT-6.

Fig. 7 shows the results of flow cytometry experiments in which mesenchymal stem cells isolated from the umbilical cord have been analysed for their expression of stem cells markers (CD73, CD90 and CD105, CD34, CD45 and HLA-DR (Human Leukocyte Antigen – antigen D Related) that are used for defining the suitability of multipotent human mesenchymal stem cells for cellular therapy and compared to the expression of these markers by bone marrow mesenchymal stem cells. For this experiment, the mesenchymal stem cells of the amniotic membrane of the umbilical cord were isolated from umbilical cord tissue by cultivation of the umbilical cord tissue in the culture medium of the present invention PPT-6 while the bone marrow mesenchymal stem cells were isolated from human bone marrow using a standard protocol. **Fig. 7a** shows the percentage of isolated mesenchymal cord lining stem cells that express the stem cell markers CD73, CD90 and CD105 and lack expression of CD34, CD45 and HLA-DR after isolation from umbilical cord tissue and cultivation in PTT-6 medium while **Fig. 7b** shows the percentage of isolated bone marrow mesenchymal stem cells that express CD73, CD90 and CD105 and lack expression of CD34, CD45 and HLA-DR.

DETAILED DESCRIPTION OF THE INVENTION

As explained above, in a first aspect the invention is directed to a method of isolating a mesenchymal stem cell population from the amniotic membrane of the umbilical cord, the method comprising cultivating umbilical cord tissue in a culture medium comprising DMEM (Dulbecco's modified eagle medium), F12 (Ham's F12 Medium), M171 (Medium 171) and FBS (Fetal Bovine Serum). It has been surprisingly found in the present application that using such a medium provides for the isolation of a mesenchymal stem cell population from the amniotic membrane of the umbilical cord of which more than 90 %, or even 99 % or more of the cells are positive for the three mesenchymal stem cell markers CD73, CD90 and while at the same these stem cells lack expression of CD34, CD45 and HLA-DR (see the Experimental Section), meaning 99 % or even more cells of this population express the stem cell markers CD73, CD90 and CD105 while not expressing the markers CD34, CD45 and HLA-DR . Such an extremely homogenous and well defined cell population is the ideal candidate for clinical trials and cell based therapies since, they for example, fully meet the criteria generally accepted for human mesenchymal stem cells to be used for cellular therapy as defined, for example, by Dominici et al, "Minimal criteria for defining multipotent mesenchymal stromal cells. The International Society for Cellular Therapy position statement", *Cytotherapy* (2006) Vol. 8, No. 4, 315-317, Sensebe et al., "Production of mesenchymal stromal/stem cells according to good manufacturing practices: a review", *Stem Cell Research & Therapy* 2013, 4:66), Vonk et al., *Stem Cell Research & Therapy* (2015) 6:94, or Kundrotas *Acta Medica Lituanica*. 2012. Vol. 19. No. 2. P. 75–79. Also, using a bioreactor such as a Quantum Cell Expansion System, it is possible to obtain high numbers of mesenchymal stem cells such as 300 to 700 million mesenchymal stem cells per run (see also the Experimental Section). Thus, the present invention allows to provide the amounts of stem cells that are needed for therapeutic applications such as their use in wound healing in a cost efficient manner. In addition, all components used for making the culture medium of the present invention are commercially available in GMP quality. Accordingly, the present invention opens the route to the GMP production of this highly homogenous mesenchymal stem cell population from the amniotic membrane of the umbilical cord.

In this context, it is noted that the culture medium of the present invention allows the isolation of a mesenchymal stem cell population (also referred hereas as "mesenchymal stem cells") from the amniotic membrane under conditions that allow cell proliferation of the

mesenchymal stem/progenitor cells without differentiation of the mesenchymal stem/progenitor cells. Thus, after isolation of the mesenchymal stem cells from the amniotic membrane as described herein the isolated mesenchymal stem/progenitor cell population has the capacity to differentiate into multiple cell types as described in US patent application 5 2006/0078993, US patent 9,085,755, International patent application WO2006/019357, US patent 8,287,854 or WO2007/046775, for instance. As described in US patent application 2006/0078993, for example, the mesenchymal stem cells of the amniotic membrane of the umbilical cord have a spindle shape, express the following genes: POU5f1, Bmi-1, leukemia inhibitory factor (LIF), and secrete Activin A and Follistatin. The mesenchymal stem cells 10 isolated in the present invention can, for example, be differentiated into any type of mesenchymal cell such as, but not limited to, adipocyte, skin fibroblasts, chondrocytes, osteoblasts, tenocytes, ligament fibroblasts, cardiomyocytes, smooth muscle cells, skeletal muscle cells, adipocytes, mucin producing cells, cells derived from endocrine glands such as insulin producing cells (for example, β -islet cells) or neurectodermal cells. The stem cells 15 isolated in the present invention can be differentiated *in vitro* in order to subsequently use the differentiated cell for medical purposes. An illustrative example of such an approach is the differentiation of the mesenchymal stem cells into insulin producing β -islet cells which can then be administered, for example by implantation, to a patient that suffers from an insulin deficiency such as diabetes mellitus (cf. also WO2007/046775 in this respect). Alternatively, 20 the mesenchymal stem cells of the invention can be used in their undifferentiated state for cell based therapy, for example, for wound healing purposes such as treatment of burns or chronic diabetic wounds. In these therapeutic applications the mesenchymal stem cells of the invention can either serve to promote wound healing by interacting with the surrounding diseased tissue or can also differentiate into a respective skin cell (cf., again WO2007/046775, for example).

25 In this context, it is noted that the mesenchymal stem cell population described herein can be isolated and cultivated (i.e. are derived) from any umbilical cord tissue as long as the umbilical cord tissue contains the amniotic membrane (which is also referred to as "cord lining"). Accordingly, the mesenchymal stem cell population can be isolated from (pieces of) the entire umbilical cord as described in the Experimental section of the present application. This 30 umbilical cord tissue may thus contain, in addition to the amniotic membrane, any other tissue/component of the umbilical cord. As shown, for example, in Figure 16 of US patent application 2006/0078993 or International patent application WO2006/019357, the amniotic membrane of the umbilical cord is the outmost part of the umbilical cord, covering the cord. In addition, the umbilical cord contains one vein (which carries oxygenated, nutrient-rich blood

to the fetus) and two arteries (which carry deoxygenated, nutrient-depleted blood away from the fetus). For protection and mechanical support these three blood vessels are embedded in the Wharton's jelly, a gelatinous substance made largely from mucopolysaccharides. Accordingly, the umbilical cord tissue used in the present invention can also comprise this one

5 vein, the two arteries and the Wharton's jelly. The use of such an entire (intact) section of the umbilical cord has the advantage that the amniotic membrane does not need to be separated from the other components of the umbilical cord. This reduces the isolation steps and thus makes the method of the present invention, simpler, faster, less error prone and more economical – which are all important aspects for the GMP production that is necessary for

10 therapeutic application of the mesenchymal stem cells. The isolation of the mesenchymal stem cells can thus start by tissue explant, which may be followed by subsequent subculturing (cultivation) of the isolated mesenchymal stem cells if greater amounts of the mesenchymal stem cells are desired, for example, for use in clinical trials. Alternatively, it is also possible to first separate the amniotic membrane from the other components of the umbilical cord and

15 isolate the mesenchymal cord lining stem cells from the amniotic membrane by cultivation of the amniotic membrane in a culture medium of the present invention. This cultivation can also be carried out by tissue explant, optionally followed by subculturing of the isolated mesenchymal stem cells. In this context, the term “tissue explant” or “tissue explant method” is used in its regular meaning in the art to refer a method in which a tissue, once being

20 harvested, or a piece of the tissue is being placed in a cell culture dish containing culture (growth) medium and by which over time, the stem cells migrate out of the tissue onto the surface of the dish. These primary stem cells can then be further expanded and transferred into fresh dishes through micropropagation (subculturing) as also described here. In this context, it is noted that in terms of production of the cells for therapeutic purposes, in the first step of

25 isolating the amniotic membrane mesenchymal stem cells from the umbilical cord, a master cell bank of the isolated mesenchymal stem cells is obtained, while the subsequent subculturing a working cell bank can be obtained. If a mesenchymal stem cell population of the invention (in particular a population of the mesenchymal stem cells of which at least about 98% or 99 % or express each of the markers CD73, CD90 and CD105 and lack expression of

30 each of the markers: CD34, CD45 and HLA-DR) is used for clinical trials or as an approved therapeutic, a cell population of the working cell bank will be typically used for this purpose. Both the stem cell population of the isolation step (which may make up the master cell bank) and the stem cell population of the subculturing step (which may make up the working cell bank) can, for example, be stored in cryo-preserved form.

As mentioned above, the present method of isolating mesenchymal stem cells from the amniotic membrane of umbilical cord has the advantage that all components used in the culture medium of the invention are available in GMP quality and thus provide the possibility to isolate the mesenchymal stem cells under GMP conditions for subsequent therapeutic administration.

By “DMEM” is meant Dulbecco’s modified eagle medium which was developed in 1969 and is a modification of basal medium eagle (BME) (cf. Fig.1 showing the data sheet of DMEM available from Lonza). The original DMEM formula contains 1000 mg/L of glucose and was first reported for culturing embryonic mouse cells. DMEM has since then become a standard medium for cell culture that is commercially available from various sources such as ThermoFisher Scientific (catalogue number 11965-084), Sigma Aldrich (catalogue number D5546) or Lonza, to new only a few suppliers. Thus, any commercially available DMEM can be used in the present invention. In preferred embodiments, the DMEM used herein is the DMEM medium available from Lonza under catalog number 12-604F. This medium is DMEM supplemented with 4.5 g/L glucose and L-glutamine). In another preferred embodiment the DMEM used herein is the DMEM medium of Sigma Aldrich catalogue number D5546 that contains 1000 mg/L glucose, and sodium bicarbonate but is without L-glutamine.

By “F12” medium is meant Ham’s F12 medium. This medium is also a standard cell culture medium and is a nutrient mixture initially designed to cultivate a wide variety of mammalian and hybridoma cells when used with serum in combination with hormones and transferrin (cf. Fig. 2, showing the data sheet of Ham’s F12 medium from Lonza). Any commercially available Ham’s F12 medium (for example, from ThermoFisher Scientific (catalogue number 11765-054), Sigma Aldrich (catalogue number N4888) or Lonza, to new only a few suppliers) can be used in the present invention. In preferred embodiments, Ham’s F12 medium from Lonza is used.

By “DMEM/F12” or “DMEM:F12” is meant a 1:1 mixture of DMEM with Ham’s F12 culture medium (cf. Fig. 3 showing the data sheet for DMEM: F12 (1:1) medium from Lonza). Also DMEM/F12 (1:1) medium is a widely used basal medium for supporting the growth of many different mammalian cells and is commercially available from various supplier such as ThermoFisher Scientific (catalogue number 11330057), Sigma Aldrich (catalogue number

D6421) or Lonza. Any commercially available DMEM:F12 medium can be used in the present invention. In preferred embodiments, the DMEM:F12 medium used herein is the DMEM/F12 (1:1) medium available from Lonza under catalog number 12-719F (which is DMEM: F12 with L-glutamine, 15 mM HEPES, and 3.151 g/L glucose).

5 By “M171” is meant culture medium 171, which has been developed as basal medium for the culture of for the growth of normal human mammary epithelial cells (cf. Fig. 4 showing the data sheet for M171 medium from Life Technologies Corporation). Also this basal medium is widely used and is commercially available from supplier such as ThermoFisher Scientific or Life Technologies Corporation (catalogue number M171500), for example. Any commercially
10 available M171 medium can be used in the present invention. In preferred embodiments, the M171 medium used herein is the M171 medium available from Life Technologies Corporation under catalogue number M171500.

By “FBS” is meant fetal bovine serum (that is also referred to as “fetal calf serum”), i.e. the blood fraction that remains after the natural coagulation of blood, followed by centrifugation
15 to remove any remaining red blood cells. Fetal bovine serum is the most widely used serum-supplement for *in vitro* cell culture of eukaryotic cells because it has a very low level of antibodies and contains more growth factors, allowing for versatility in many different cell culture applications. The FBS is preferably obtained from a member of the International Serum Industry Association (ISIA) whose primary focus is the safety and safe use of serum
20 and animal derived products through proper origin traceability, truth in labeling, and appropriate standardization and oversight. Suppliers of FBS that are ISIA members include Abattoir Basics Company, Animal Technologies Inc., Biomin Biotechnologia LTDA, GE Healthcare, Gibco by Thermo Fisher Scientific and Life Science Production, to mention only a few. In currently preferred embodiments, the FBS is obtained from GE Healthcare under
25 catalogue number A15-151.

Turning now to the culture medium of the present invention, the culture medium may comprise for the isolation or cultivation of the mesenchymal cord lining stem cells DMEM in a final concentration of about 55 to 65 % (v/v), F12 in a final concentration of about 5 to 15 % (v/v), M171 in a final concentration of about 15 to 30 % (v/v) and FBS in a final concentration
30 of about 1 to 8 % (v/v). The value of “% (v/v)” as used herein refers to the volume of the individual component relative to the final volume of the culture medium. This means, if DMEM is, for example, present in the culture medium a final concentration of about 55 to 65 % (v/v), 1 liter of culture medium contains about 550 to 650 ml DMEM.

In other embodiments, the culture medium may comprise DMEM in a final concentration of about 57.5 to 62.5 % (v/v), F12 in a final concentration of about 7.5 to 12.5 % (v/v), M171 in a final concentration of about 17.5 to 25.0 % (v/v) and FBS in a final concentration of about 1.75 to 3.5 % (v/v). In further embodiments, the culture medium may comprise DMEM in a final concentration of about 61.8 % (v/v), F12 in a final concentration of about 11.8 % (v/v), M171 in a final concentration of about 23.6 % (v/v) and FBS in a final concentration of about 2.5 % (v/v).

In addition to the above-mentioned components, the culture medium may comprise supplements that are advantages for cultivation of the mesenchymal cord lining stem cells. The culture medium of the present invention may, for example, comprises Epidermal Growth Factor (EGF). If present, EGF may be present in the culture medium in a final concentration of about 1 ng/ml to about 20 ng/ml. In some of these embodiments, the culture medium may comprise EGF in a final concentration of about 10ng/ml.

The culture medium of the present invention may also comprises insulin. If present, insulin may be present in a final concentration of about 1 µg/ml to 10 µg/ml. In some of these embodiments, the culture medium may comprise Insulin in a final concentration of about 5µg/ml.

The culture medium may further comprises at least one of the following supplements: adenine, hydrocortisone, and 3,3',5-Triiodo-L-thyronine sodium salt (T3). In such embodiments, the culture medium may comprise all three of adenine, hydrocortisone, and 3,3',5-Triiodo-L-thyronine sodium salt (T3). In these embodiments, the culture medium may comprises may comprise adenine in a final concentration of about 0.05 to about 0.1 µg/ml adenine, hydrocortisone in a final concentration of about 1 to about 10 µg/ml hydrocortisone and/or 3,3',5-Triiodo-L-thyronine sodium salt (T3) in a final concentration of about 0.5 to about 5 ng/ml.

In the method of the invention, the umbilical cord tissue may be cultured till a suitable number of (primary) mesenchymal cord lining stem cells have outgrown from the tissue. In typical embodiments, the umbilical cord tissue is cultivated until cell outgrowth of the mesenchymal stem cells of the amniotic membrane reaches about 70 to about 80% confluency. It is noted here that the term “confluency” or “confluence” is used in its regular meaning in the art of cell culture and is meant as an estimate/indicator of the number of adherent cells in a culture dish or a flask, referring to the proportion of the surface which is covered by cells. For example, 50 percent confluence means roughly half of the surface is covered and there is still room for cells

to grow. 100 percent confluence means the surface is completely covered by the cells, and no more room is left for the cells to grow as a monolayer.

Once a suitable number of primary cells (mesenchymal cord lining stem cells) have been obtained from the cord lining tissue by tissue explant, the mesenchymal stem cells are removed from the cultivation container used for the cultivation. By so doing, a master cell bank containing the (primary) isolated mesenchymal stem cells of the amniotic membrane can be obtained. Typically, since mesenchymal stem cells are adherent cells, removing is carried out using standard enzymatic treatment. For example, the enzymatic treatment may comprise trypsinization as described in International US patent application 2006/0078993, International patent application WO2006/019357 or International patent application WO2007/046775, meaning outgrowing cells can be harvested by trypsinization (0.125% trypsin/0.05% EDTA) for further expansion. If the harvested mesenchymal stem cells are, for example, used for generating a master cell bank, the cells can also be cryo-preserved and stored for further use as explained herein below.

Once being harvested, the mesenchymal stem cells can be transferred to a cultivation container for subculturing. The subculturing can also be started from frozen primary cells, i.e. from the master cell bank. For subculturing any suitable amount of cells can be seeded in a cultivation container such as cell culture plate. The mesenchymal cells can, for this purpose, be suspended in a suitable medium (most conveniently, the culture medium of the present invention) for subculturing at a concentration of, for example, about 0.5×10^6 cells/ml to about 5.0×10^6 cells/ml. In one embodiment the cells are suspended for subcultivation at a concentration of about 1.0×10^6 cells/ml. The subculturing can be carried by cultivation either in simple culture flasks but also, for example, in a multilayer system such as CellStacks (Corning, Corning, NY, USA) or Cellfactory (Nunc, part of Thermo Fisher Scientific Inc., Waltham, MA, USA) that can be stacked in incubators. Alternatively, the subculturing can also be carried out in a closed self-contained system such as a bioreactor. Different designs of bioreactors are known to the person skilled in the art, for example, parallel-plate, hollow-fiber, or micro-fluidic bioreactors. See, for example, Sensebe et al. "Production of mesenchymal stromal/stem cells according to good manufacturing practices: a review", *supra*. An illustrative example of a commercially hollow-fiber bioreactor is the Quantum® Cell Expansion System (Terumo BCT, Inc). that has, for example, been used for the expansion of bone marrow mesenchymal stem cells for clinical trials (cf., Hanley et al, Efficient Manufacturing of Therapeutic Mesenchymal Stromal Cells Using the Quantum Cell Expansion System, *Cytotherapy*. 2014 August ; 16(8): 1048–1058).

Another example of a commercially available bioreactors that can be used for the subculturing of the mesenchymal stem cell population of the present invention is the Xuri Cell Expansion System available from GE Healthcare. The cultivation of the mesenchymal stem cell population in an automated system such as the Quantum® Cell Expansion System is of particular benefit if a working cell bank for therapeutic application is to be produced under GMP conditions and a high number of cells is wanted.

The subculturing of the mesenchymal cord lining stem cells of the invention takes place in a culture medium of the present invention. Accordingly, the culture medium of the present invention can be used both for the isolation of the mesenchymal stem cells from the amniotic membrane and the subsequent cultivation of the isolated primary cells by subcultivation. Also for the subcultivation, the mesenchymal stem cells can be cultured till a suitable amount of cells have grown. In illustrative embodiments the mesenchymal stem cells are subcultured till the mesenchymal stem cells reach about 70 to about 80% confluency.

The isolation/cultivation of the population of mesenchymal cord lining stem cells can be carried out under standard condition for the cultivation of mammalian cells. Typically, the method of the invention of isolating the population of the mesenchymal cord lining stem cells is typically carried out at conditions (temperature, atmosphere) that are normally used for cultivation of cells of the species of which the cells are derived. For example, human umbilical cord tissue and the mesenchymal cord lining stem cells, respectively, are usually cultivated at 37°C in air atmosphere with 5%CO₂. In this context, it is noted that the in present invention the mesenchymal cells may be derived of any mammalian species, such as mouse, rat, guinea pig, rabbit, goat, horse, dog, cat, sheep, monkey or human, with mesenchymal stem cells of human origin being preferred in one embodiment.

Once a desired/suitable number of mesenchymal cord lining stem cells have been obtained from the subculture, the mesenchymal stem cells are harvested by removing them from the cultivation container used for the subcultivation. The harvesting of the mesenchymal stem cells is typically again carried out by enzymatic treatment, including comprises trypsinization of the cells. The isolated mesenchymal stem cells are subsequently collected and are either be directedly used or preserved for further use. Typically, preserving is carried out by cryo-preservation. The term “cryo-preservation” is used herein in its regular meaning to describe a process where the mesenchymal stem cells are preserved by cooling to low sub-zero temperatures, such as (typically) -80°C or -196°C (the boiling point of liquid nitrogen). Cryo-preservation can be carried out as known to the person skilled in the art and can include the

use of cryo-protectors such as dimethylsulfoxide (DMSO) or glycerol, which slow down the formation of ice-crystals in the cells of the umbilical cord.

The isolated population of the mesenchymal cord lining stem cells that is obtained by the isolation method of the present invention is highly defined and homogenous. In typical
 5 embodiments of the method at least about 90 % or more, about 91 % or more, about 92 % or more, about 92 % or more, about 93 % or more, about 94 % or more, about 95 % or more, about 96 % or more, about 97 % or more, about 98 % or more about 99 % or more of the isolated mesenchymal stem cells express the following markers: CD73, CD90 and CD105. In addition, in these embodiments at least about 90 % or more, about 91 % or more, about 92 %
 10 or more, about 92 % or more, about 93 % or more, about 94 % or more, about 95 % or more, about 96 % or more, about 97 % or more, about 98 % or more about 99 % or more of the isolated mesenchymal stem cells may lack expression of the lack expression of the following markers: CD34, CD45 and HLA-DR. In particular embodiments, about 97 % or more, about 98 % or more, or about 99 % or more of the isolated mesenchymal stem cell population
 15 express CD73, CD90 and CD105 while lacking expression of CD34, CD45 and HLA-DR.

Thus, in line with the above disclosure the present invention is also directed to a mesenchymal stem population isolated from the amniotic membrane of the umbilical cord, wherein at least about 90 % or more cells of the stem cell population express each of the following markers: CD73, CD90 and CD105. In preferred embodiments at least about 91 % or more, about 92 %
 20 or more, about 92 % or more, about 93 % or more, about 94 % or more, about 95 % or more, about 96 % or more, about 97 % or more, about 98 % or more about 99 % or more cells of the isolated mesenchymal stem cell population are CD73+, CD90+ and CD105+, meaning that this percentage of the isolate cell population express each of CD73, CD90 and CD105 (cf. the Experimental Section of the present application). In addition, at least about 90 % or more,
 25 about 91 % or more, about 92 % or more, about 92 % or more, about 93 % or more, about 94 % or more, about 95 % or more, about 96 % or more, about 97 % or more, about 98 % or more about 99 % or more of the isolated mesenchymal stem cells may lack expression of the lack expression of the following markers. In particular embodiments about 97 % or more, about 98 % or more, or about 99 % or more of the isolated mesenchymal stem cell population express
 30 CD73, CD90 and CD105 while lacking expressing of CD34, CD45 and HLA-DR. Such a highly homogenous population of mesenchymal stem cells derived from the amniotic membrane of the umbilical cord has been reported here for the first time and meets the criteria for mesenchymal stem cells to be used for cellular therapy (also cf. the Experimental Section

and, for example, Sensebe et al. "Production of mesenchymal stromal/stem cells according to good manufacturing practices: a review", supra). It is noted in this context that this mesenchymal stem cell population can be obtained by either the isolating method of the present invention but also by a different method such as cell sorting, if wanted.

- 5 In line with the above, the present invention is also directed to a pharmaceutical composition comprising a mesenchymal stem population isolated from the amniotic membrane of the umbilical cord, wherein at least about 90 % or more cells of the stem cell population express each of the following markers: CD73, CD90 and CD105 and optionally, lack expression of CD34, CD45 and HLA-DR. The pharmaceutical composition may comprise any
10 pharmaceutically acceptable excipient and may be formulated for any desired pharmaceutical way of administration. The pharmaceutical composition may, for example, be adapted for systemic or topical application.

In a further aspect the invention is directed to a method of making a culture medium for isolating the method comprising, mixing to obtain a final volume of 500 ml culture medium:

- 15 i. 250 ml of DMEM
ii. 118 ml M171
iii. 118 ml DMEM/F12
iv. 12.5 ml Fetal Bovine Serum (FBS) to reach a final concentration of 2.5% (v/v).

As explained above, DMEM/F12 medium is a 1:1 mixture of DMEM and Ham's F12 medium.

- 20 Thus, 118 ml DMEM/F12 medium contain 59 ml DMEM and 59 ml F12. Accordingly, when using this method of making a culture medium, the final concentrations (v/v) mit 500 ml total volume are as follows:

DMEM: $250 \text{ ml} + 59 \text{ ml} = 309 \text{ ml}$, corresponds to $309/500 = 61.8 \%$ (v/v)

M171: 118 ml, corresponds to $118/500 = 23.6 \%$ (v/v)

- 25 F12: 59 ml, corresponds to $59/500 = 11.8 \%$ (v/v).

Embodiments of this method of making a culture medium further comprise adding

- v. 1 ml EGF stock solution (5 µg/ml) to achieve a final EGF concentration of 10ng/ml, and

vi. Insulin 0.175 ml stock solution (14.28 mg/ml) to achieve a final insulin concentration of 5 µg/ml.

It is noted here that in these embodiments, the above-mentioned volumes of these components i. to vi when result in a final volume of 499.675 ml culture medium. If no further components are added to the culture medium, the remaining 0.325 ml (to add up to a volume of 500 ml) can, for example, be any of components i. to iv, that means either DMEM, M171, DMEM/F12 or FBS. Alternatively, the concentration of the stock solution of EGF or Insulin can of course be adjusted such that the total volume of the culture medium is 500 ml. In addition, it is also noted that compoents i. to iv. do not necessarily have to be added in the order in which they are listed but it is of course also possible to use any order to mix these components to arrive at the culture medium of the present invention. This means, that for example, M171 and DMEM/F12 can be mixed together and then combined with DMEM and FBS to reach final concentrations as described here, i.e. a final concentration of DMEM of about 55 to 65 % (v/v), a final concentration of F12 of about 5 to 15 % (v/v), a final concentration of M171 of about 15 to 30 % (v/v) and a final concentration of FBS of about 1 to 8 % (v/v).

In other embodiments, the method further comprises adding to DMEM a volume of 0.325 ml of one or more of the following supplements: adenine, hydrocortisone, 3,3',5-Triiodo-L-thyronine sodium salt (T3), thereby reaching a total volume of 500 ml culture medium. In this embodiments, the final concentration of these supplements in DMEM may be as follows:

- about 0.05 to 0.1 µg/ml adenine, for example about 0.025 µg/ml adenine,
- about 1 to 10 µg/ml hydrocortisone,
- about 0.5 to 5 ng/ml 3,3',5-Triiodo-L-thyronine sodium salt (T3), for example 1.36 ng/ml 3,3',5-Triiodo-L-thyronine sodium salt (T3).

In line with the above disclosure, the invention is also directed to a cell culture medium that is obtainable or that is obtained by the method of making the medium as described here.

In addition, the invention also concerns a method of isolating mesenchymal stem cells from the amniotic membrane of the umbilical cord, wherein this method comprises cultivating amniotic membrane tissue in the culture medium prepared by the method as described here.

Thus, the present invention is also directed to a cell culture medium comprising:

- DMEM in the final concentration of about 55 to 65 % (v/v),

- F12 in a final concentration of about 5 to 15 % (v/v),
- M171 in a final concentration of about 15 to 30 % (v/v) and
- FBS in a final concentration of about 1 to 8 % (v/v).

In certain embodiments of the culture medium described here, the medium comprises DMEM
 5 in the final concentration of about 57.5 to 62.5 % (v/v), F12 in a final concentration of about
 7.5 to 12.5 % (v/v), M171 in a final concentration of about 17.5 to 25.0 % (v/v) and FBS in a
 final concentration of about 1.75 to 3.5 % (v/v). In other embodiments the culture medium
 may comprise DMEM in a final concentration of about 61.8 % (v/v), F12 in a final
 concentration of about 11.8 % (v/v), M171 in a final concentration of about 23.6 % (v/v) and
 10 FBS in a final concentration of about 2.5 % (v/v).

In addition, the culture medium may further comprise Epidermal Growth Factor (EGF) in a
 final concentration of about 1 ng/ml to about 20 ng/ml. In certain embodiments, the culture
 medium comprise EGF in a final concentration of about 10ng/ml. The culture medium
 described herein may further comprise Insulin in a final concentration of about 1 µg/ml to 10
 15 µg/ml. In such embodiments the culture medium may comprise Insulin in a final concentration
 of about 5µg/ml.

The cell culture medium of the invention may further comprise at least one of the following
 supplements: adenine, hydrocortisone, and 3,3',5-Triiodo-L-thyronine sodium salt (T3). In
 certain embodiments the culture medium comprises all three of adenine, hydrocortisone, and
 20 3,3',5-Triiodo-L-thyronine sodium salt (T3). If present, the culture medium may comprise
 adenine in a final concentration of about 0.01 to about 0.1 µg/ml adenine or of about 0.05 to
 about 0.1 µg/ml adenine, hydrocortisone in a final concentration of about 0.1 to about 10
 µg/ml hydrocortisone or of about 1 to about 10 µg/ml hydrocortisone and/or 3,3',5-Triiodo-L-
 thyronine sodium salt (T3) in a final concentration of about 0.5 to about 5 ng/ml.

25 In embodiments of the cell culture medium, 500 ml of the cell culture medium of the present
 invention comprise:

- i. 250 ml of DMEM
- ii. 118 ml M171
- iii. 118 ml DMEM/F12
- 30 iv. 12.5 ml Fetal Bovine Serum (FBS) (final concentration of 2.5%)

In further embodiments, the cell culture medium may further comprise

v. EGF in a final concentration of 10ng/ml, and

vi. Insulin in a final concentration of 5µg/ml.

Both, insulin and and EGF can be added to to the culture medium using a stock solution of
5 choice, such that the total volume of the culture medium does not exceed 500 ml.

In a particular example, the components i. to vi. of the culture medium of the present invention are the components indicated in Figure 5, meaning they are obtained from the respective manufacturers using the catalogue number indicated in Figure 5. The medium that is obtained from mixing the components i. to vi. as indicated in Figure 5 is also referred herein as “PTT-
10 6”. It is again noted in this context that the constituents i. to vi. as well as any other ingredient such as an antibiotic of any other commercial supplier can be used in making the medium of the present invention.

In addition, the cell culture medium of the invention may comprise adenine in a final concentration of about 0.01 to about 0.1 µg/ml adenine or of about 0.05 to about 0.1 µg/ml
15 adenine, hydrocortisone in a final concentration of about 0.1 to 10 µg/ml, of about 0.5 to about 10µg/ml, or of about 1 to about 10 µg/ml hydrocortisone and/or 3,3',5-Triiodo-L-thyronine sodium salt (T3) in a final concentration of about 0.1 to about 5 ng/ml or of about 0.5 to about 5 ng/ml.

Finally, the invention also provides a method of treating a patient having a disease, the method
20 comprising administering to the patient a mesenchymal cord lining stem cell or a pharmaceutical composition containing a stem cell as disclosed herein. The disease can be any disease thas described above. For treating the subject, the mesenchymal stem cell population of the invention may be administered in any suitable way, for example, including but not limited to, topical administration, by implantation or by injection. The stem cell population
25 may, for example, be placed directly onto a wound such as a burn or a diabetic wound (see International patent application WO2007/046775). Alternatively, the stem cell population may also be implanted subcutaneously, for example, directly under the skin, in body fat or the peritoneum.

The invention will be further illustrated by the following non-limiting Experimental Examples.

Experimental Examples

1. Cryopreservation of Umbilical Cord Tissue Prior to Isolation of Mesenchymal Stem Cells

Umbilical cord tissue (the umbilical cords were donated with informed consent of the mother) was processed for the subsequent isolation of the mesenchymal stem cells from the amniotic membrane of the umbilical cord as follows.

1.1 Washing of umbilical cord tissue sample:

- a. Remove scalpels from the protective cover.
- 10 b. Hold the umbilical cord securely using the forceps and cut the cord into a 10 cm length piece using a scalpel. Place the unusable cord back in the original tissue cup.
- c. Transfer the 10 cm long umbilical cord piece into a new 150 mm culture dish. The 150mm culture dish may be used in place of the cups.
- d. Use the cover of the 150 mm culture dish as a resting place for forceps and scalpel.
- 15 e. Remove 25ml Plasmalyte A (Baxter, Catalog # 2B2543Q) with a 30 ml syringe. Hold the syringe at a 45° angle using one hand and dispense the Plasmalyte A directly onto the umbilical cord tissue.
- f. Holding the culture dish at a slight angle remove the Plasmalyte A with a 30 ml syringe and blunt needle.
- 20 g. Collect used Plasmalyte A in a 300 ml transfer bag that serves as a trash container and dispose it in the biohazard bin.
- h. Repeat wash procedure, if necessary using a new culture dish for each wash. Make sure all blood clots on the surface have been removed. More Plasmalyte A can be used if needed to clean the tissue.
- 25 i. Place the tissue into a new labeled tissue culture dish to continue cutting the tissue. Place 20 ml of Plasmalyte A into the dish so the tissue does not dry out while cutting it.
- j. Cut the cords into equal approximately 1-cm sections resulting in 10 sections in total.
- k. Further cut each 1 cm section into smaller pieces with approximately 0.3 cm x 0.3 cm to 0.5 cm x 0.5 cm per section.
- 30 l. Remove any Plasmalyte A that is in the dish.
- m. Pull 25 ml Plasmalyte A with a 30 ml syringe from the original Plasmalyte A bag and dispense directly on the umbilical cord tissue pieces.
- n. Hold culture dish in an angle to collect all Plasmalyte A used for washing the tissue on one side and remove it with a syringe and blunt needle.

o. Repeat wash one more time. There should not be any clots left.

NOTE: If the cord is not frozen right away, the umbilical cord tissue is kept in Plasmalyte A until ready to freeze.

5

1.2 Cryopreservation of umbilical cord tissue:

a. Prepare cryopreservation solution:

i. Prepare 50 ml freezing solution consisting of 60% Plasmalyte A, 30% of 5% Human Serum Albumin, and 10% dimethyl sulfoxide (DMSO).

10 ii. Label a 150 ml transfer bag with “Tissue freeze solution” and attach a plasma transfer set to the port using aseptic technique.

iii. Remove 30 ml Plasmalyte A with a 30 ml Syringe from the original Plasmalyte A bag and transfer it in the transfer bag labeled “tissue freeze solution” with the time and date solution is made.

15 iv. Remove 15 ml of 5% Human Serum Albumin with a 20 ml syringe and transfer it into the labeled transfer bag.

v. Add 5 ml DMSO to the transfer bag.

vi. Mix well and record mixing of freeze solution

b. Remove the Plasmalyte A from the tissue before adding the freeze solution.

20 c. Using a 60 ml syringe, pull all 50 mls of the freeze solution into the syringe add approximately 30 ml freeze solution to the 150 mm cell culture dish containing the umbilical cord tissue. Place a blunt needle on the syringe to keep it sterile.

d. Swirl the culture dish containing the tissue and freezing solution every minute for 10 minutes.

25 e. Using forceps, select 8 randomly chosen sections and place them in each of the four 4 ml cryovials. Select 4 randomly chosen sections and place them into one 1.8 ml cryovial. These sections should be free of blood clots.

f. Fill each cryovial containing the umbilical cord tissue with the remaining freezing solution to the 3.6 ml filling line for the 4 ml tubes and the 1.8 ml line for the 1.8 ml Nunc vial.

30 g. Label one Bactec Lytic/10 - Anaerobic/F and one Bactec Plus Aerobic/F bottle with tissue ID.

h. Remove 20 ml freeze solution from the culture dish with a syringe and a blunt needle, after wiping the Bactec vials with an alcohol swab, switch the blunt needle for an 18g needle and inoculate the aerobic and the anaerobic Bactec bottles with 10 ml each.

- i. Start controlled rate freezer.
- j. After controlled rate freeze is completed place the units in a continuous temperature monitored liquid nitrogen freezer until further use.

5 **2. Isolation of Mesenchymal Cord Lining Stem Cells from umbilical cord tissue**

2.1. Preparing media for processing MSCs from umbilical cord tissue:

a. To make 500 ml PTT6 (culture/growth media) add the following in the order listed:

- i. DMEM, 250 ml
- 10 ii. M171 118 ml
- iii. DMEM F12 118 ml
- iv. FBS 12.5 ml (final concentration of 2.5%)
- v. EGF 1 ml (final concentration of 10ng/ml)
- vi. Insulin 0.175 ml (final concentration of 5µg/ml)

15

The above-mentioned volumes of components i. to vi when result in a final volume of 499.675 ml culture medium. If no further components are added to the culture medium, the remaining 0.325 ml (to add up to a volume of 500 ml) can, for example, be any of components i. to iv, that means either DMEM, M171, DMEM/F12 or FBS. Alternatively, the concentration of the stock solution of EGF or Insulin can of course be adjusted such that the total volume of the culture medium is 500 ml. Alternatively, a stock solution of an antibiotic such as Penicillin-Streptomycin-Amphotericin can be added to result in a final volume of 500 ml. It is also possible to add to the culture medium a volume of 0.325 ml of one or more of the following supplements: adenine, hydrocortisone, 3,3',5-Triiodo-L-thyronine sodium salt (T3), thereby reaching a total volume of 500 ml culture medium.

25

vii. Label the bottle "PTT6" with date media was prepared, initial of the operator, and the phrase "expires on" followed by the expiration date. Expiration date is the earliest expiration date of any of the component or 1 month from the preparation date, whichever comes first.

30

b. To make the rinse media (Hank's Buffered Salt Solution (HBSS) without Calcium or Magnesium and with 5% FBS), add 2.5 ml FBS to 47.5 ml of HBSS in a 50 ml centrifuge tube. Label the tube "Rinse Media" with operator initials and date the media is made.

c. All media will be tested for sterility using Bactec Lytic/10 – Anaerobic/F (Becton Dickinson & Company) and Bactec Pluc + Aerobic/F (Becton Dickinson & Company). Inject 20 ml of prepared media into each bottle.

5 **2.2 Thawing of umbilical cord tissue for MSC harvesting:**

- a. Initiate the thaw once an operator is prepared to process the sample in the clean room. Do not thaw more than 1 vial at a time unless the vials originate from the same donor.
- b. Wipe the water bath with disinfectant followed by 70% isopropanol and fill it with 1 L sterile water. Heat the water bath up to 36-38°C.
- 10 c. Prepare 10 mL of rinse medium consisting of 70% to 90% PlasmaLyte A in the clean room under a biosafety cabinet. Sterile filter the solution with a 0.2-µm syringe filter attached to a 10 ml syringe and keep the solution refrigerated until use.
- d. Place a processing label on a 50 ml conical tube.
- e. Confirm water bath temperature is at 36-38°C.
- 15 f. Take vial(s) of tissue from the liquid nitrogen storage and thaw rapidly in the 37°C water bath filled with 1L of sterile water. The vial holder for the Mr. Frosty Nalgene Cryo 1°C freezing container floats with vials in place and can be used as a floating rack when thawing samples.
- g. Remove the vial from the water bath and spray them with 70% Isopropanol solution. A
- 20 good time to pull the vial from the water bath is when small ice can be seen floating in the vial – suggest internal temperature of the vial is less than 37°C.
- h. Place vial into pass-through and alert the clean room processing technician.

2.3 Preparing for tissue processing:

- 25 a. Umbilical cord tissue processing should be performed in an environmentally monitored (EM) clean room: At the end of each shift, full room and hood cleaning are performed
- b. Prepare/clean the biosafety cabinet.
- c. Perform viable particle counting while working in the biosafety cabinet.
- d. Assemble all necessary supplies in the biosafety cabinet checking each for packaging
- 30 damage and expiration dates. When handling syringes, serological pipets, sterile forceps, scalpels, tissue plates, and needles, make sure not to touch any surface that will come in contact with the sterile product. Only the exterior of the syringe barrel, tubing, plunger tip and/or needle cap or sheath may be safely handled. Discard supply if the surface has been touched or has touched a non-sterile surface.

- e. Record lot numbers and expiration dates (if applicable) of all reagents and supplies to be used.
- f. Receive the thawed vial by cleaning the vial with lint-free wipe moistened with 70% alcohol before transferring into the biosafety cabinet.
- 5 g. Using an aspirating needle with a syringe, withdraw as much liquid from the vial. Avoid suctioning the tissue.
- h. Using sterile forceps, transfer the tissue into a sterile 100 mm petri dish.
- i. Add an aliquot of 5 ml rinse medium to the tissue fragments.
- j. Swirl the contents for 15-30 seconds, then remove the rinse medium with a pipette or
- 10 syringe with aspirating needle. Repeat this rinse process twice.
- k. Add 2 mL of rinse medium to the tissue to avoid drying out the tissue.

2.4. Initiating MSC outgrowth from tissue:

- a. Label the bottom of a 6-well plate “Outgrowth 1” with MSC lot number or umbilical cord
- 15 tissue ID and the date outgrowth is initiated. If 60 mm tissue culture dish is used, divide the plate into 4 quadrants by drawing a grid on the bottom of the dish.
- b. Using sterile, disposable forceps, place one 3 x 3 mm to 5 x 5 mm tissue into each well. If using a 60 mm tissue culture dish, place the tissue into the middle of each quadrant to keep the tissues apart (more than 1 cm from each other).
- 20 c. Fill each well with 3 ml of PTT6.
- d. Using an aspirating needle coupled to 30 ml syringe, withdraw enough media to barely cover the tissue. Do not tilt the plate. Do not touch the bottom of the well with the aspirating needle.
- e. Using an inverted light microscope, observe for cell outgrowth every day (24 ± 6 hrs). Real
- 25 time cell culture imaging system may be used in place of the light microscope.
- f. Change media every day. Be sure to equilibrate the media to room temperature before use.
- i. Aspirate off the medium.
- ii. Add 3 ml of PTT6.
- iii. Aspirate until tissue is barely submerged in the medium.
- 30 g. When cellular outgrowth is observed from the tissue, transplant the tissue to a new 6-well plate using the same procedure as 4.a to 4.e above except label the plate “Outgrowth 2”. Maintain cell outgrowth in “Outgrowth 1” plate by adding 2ml of PTT6 to each well. Observe for confluency every day. Replace media every 2-3 days (be sure to equilibrate the media to room temperature before use).

- h. When cell outgrowth is observed in “Outgrowth 2” plate, repeat step 4.a to 4.e except label the plate “Outgrowth 3.” Maintain cell outgrowth in “Outgrowth 2” plate by adding 2 ml of PTT6 to each well. Observe for confluency every day. Replace media every 2-3 days (be sure to equilibrate the media to room temperature before use).
- 5 i. When outgrowth is observed in “Outgrowth 3” plate, discard the tissue. If the tissues are very small and do not seem to interfere with cell growth, dispose of the tissue when subculturing.
- j. When cells reach 40-50% confluency, observe cells every days to prevent over-expansion.
- k. When cells reach 70-80% confluency, subculture the cells. Do not allow cells to expand
- 10 beyond 80% confluence.

With the size of the tissue explants being about 1-3mm, and the tissue explant/cell culture is performed in 175 mm squared culture dishes, the average number of mesenchymal stem cells harvested from an explant is typically about 4,000 - 6,000 cells/explant. Accordingly, when

15 the mesenchymal stem cells are simultaneously grown out of 48 explants about 300,000 cells can be obtained at harvest. These 300,000 mesenchymal stem cells collected from explants can then be used for subculturing by seeding a 175cm² cell culture flask with such 300,000 cells as described in the following Example 2.5 (this can be referred to as Passage 1). The mesenchymal stem cells obtained from this passage 1 can then be used to seed again 175cm²

20 flasks (Passage 2) and expand the cells as described in the following Example 2.5. The cells obtained from both Passage 1 and Passage 2 can be “banked” by cryo-preservation, with the mesenchymal stem cells obtained after Passage 2 being considered to represent the Master Cell Bank which will be for further expansion of the mesenchymal stem cells, for example, in a bioreactor as explained below in Example 2.7.

25

2.5. Subculturing MSC in cell culture dishes

- a. Perform viable particle while working in the biosafety cabinet. Equilibrate all media to room temperature before use.
- b. When cell outgrowth reaches about 70-80% confluency, subculture cells.
- 30 i. Remove PTT6 from the petri dish.
- ii. Rinse with HBSS without Calcium or Magnesium.
- iii. Add 0.2 ml 1X TrypLE-EDTA and swirl for 1-2 minutes.
- iv. Tilt the dish 30-45° to allow cells to shift down by gravitational flow. Gentle tapping on the side of the plate expedites detachment.

- v. Add 1ml of PTT6. Pipette up and down gently then transfer cells to a 15 ml centrifuge tube. Use clean pipette tip with each well. Cells from all 6 wells can be pooled into a single 15ml tube.
- vi. Centrifuge for 10 minutes at 1200 rpm.
- 5 vii. Remove supernatant and resuspend cells with 5ml PTT6.
- c. Subculturing MSC
 - i. Aliquot 50 μ l of the cell suspension and assay for TNC and viability by Trypan Blue Exclusion Assay.
 - ii. Count cells using a hemocytometer. Expect to count 20-100 cells/square. If the count higher
 - 10 than 100, dilute the original sample 1:5 and repeat Trypan Blue method using a hemocytometer.
 - iii. Calculate viable cells/ml and total viable cells:
 1. Viable cells/ml = viable cell count x dilution factor x 10^4
 2. Total viable cells = viable cell count x dilution factor x total volume x 10^4
 - 15 iv. Calculate % viability:
 1. % viability = viable cell count x 100 / (viable cell count + dead cell count)
 - v. Dilute the cell suspension to 1.0×10^6 cells/ml:
 1. "X" volume = Total viable cells/ 10^6 cells/ml
 2. For example, if total viable cell number is 1.0×10^7 ;
 - 20 3. "X" = $10^7/10^6$ cells/ml or 10 ml, therefore, you would bring your total cell volume up to 10 ml by adding 5 ml to your cell suspension (that is at 5 ml).
 - vi. If the cell suspension is less than 106/ml, determine the volume required to seed 2×10^6 cells for each 150 mm petri dish or 175 cm² flask.
 1. Volume for 2×10^6 cells = 2×10^6 cells $\div$ viable cells/ml
 - 25 2. For example, if viable cells/ml is 8×10^5 cells/ml, 2×10^6 cells $\div$ 8×10^5 cells/ml or 2.5 ml are needed.
 - vii. Set aside 0.5 ml for MSC marker analysis.
 - viii. Seed 2×10^6 cells to each 150 mm petri dish or 175 cm² flask with 30 ml PTT6.
 - ix. Observe cells for attachment, colony formation, and confluence every three days. When
 - 30 cells reach 40-50% confluence, observe cells every one-two days to prevent over-expansion. DO NOT allow cells to expand beyond 80% confluence. A real time cell culturing monitoring system can be used in place of the light microscope.
 - x. Replace media every 2-3 days.

2.6 Cryopreserving MSC cells

- a. Perform viable particle while working in the biosafety cabinet.
- b. When cells reach 70-80% confluence, detach cells using 2 ml 1X TrypLE-EDTA for each 150 mm petri dish or 175 cm² flask.
- 5 i. Remove PTT6 from the petri dish.
- ii. Wash with 5ml HBSS or PBS without calcium or magnesium.
- iii. Add 2 ml 1X TrypLE-EDTA and swirl for 1-2 minutes.
- iv. Tilt the dish 30-45° to allow cells to shift down by gravitational flow. Gentle tapping on the side of the petri dish helps to expedite detachment.
- 10 v. Add 10 ml PTT6 to inactivate TrypLE. Mix well to dissociate cell clumps.
- vi. Transfer cells to 15 ml centrifuge tube using a Pasteur pipette.
- vii. Centrifuge for 10 minutes at 1200 rpm.
- viii. Aspirate medium and resuspend with 10 ml PTT6.
- ix. Aliquot 50 µl and determine total viable cell number and % viability as above. Cell count
- 15 will need to be performed within 15 minutes as the cells may start clumping.
- c. Preparing cells for cryopreservation
- i. Prepare Cell Suspension Media and Cryopreservation Media and freeze the cells

2.7. Subculturing (expansion) of MSC in a Quantum Bioreactor (Terumo BTC, Inc.)

- 20 It is also possible to use a Quantum Bioreactor can used to expand the MSC. The starting cell number for the expansion in the Quantum Bioreactor should range between 20 to 30 million cells per run. The typical yield per run is 300 to 700 million MSC at harvest. The Bioreactor is operated following the protocol of the manufacturer. The so obtained mesenchymal stem cells are typically cryo-preserved (see below) and serve as Working Cell Bank.

25

MATERIALS/ REAGENTS:

1. Quantum Expansion Set
2. Quantum Waste Bag
3. Quantum Media Bag
- 30 4. Quantum Inlet Bag
5. PTT6
6. PBS
7. Fibronectin
8. TrypLE

9. 3ml syringe
10. Glucose test strips
11. Lactate test strips
12. 60ml Cell Culture Plate or equivalent
- 5 13. Medical Grade 5% CO₂ Gas-mix
14. 50ml Combi-tip

EQUIPMENT:

1. Biosafety Cabinet
- 10 2. Glucose Meter (Bayer Healthcare/Ascensia Contour Blood Glucose Meter)
3. Lactate Plus (Nova Biomedical)
4. Peristaltic pump with head
5. Centrifuge, Eppendorf 5810
6. Sterile Tube Connector
- 15 7. M4 Repeat Pipettor
8. RF Sealer

PROCEDURE:

1. Preparing the Quantum Bioreactor

- 20 a) Priming the Quantum Bioreactor
- b) Coating the bioreactor:
 - 1) Prepare the fibronectin solution in the biosafety cabinet.
 - 1) Allow lyophilized fibronectin to acclimate to room temperature (≥ 15 min at room temperature)
 - 25 2) Add 5ml of sterile distilled water; do not swirl or agitate
 - 3) Allow fibronectin to go into solution for 30 min.
 - 4) Using a 10 ml syringe attached with an 18g needle, transfer the fibronectin solution to a Ccell inlet bag containing 95ml of PBS.
 - 2) Attach the bag to the “reagent” line
 - 30 3) Check for bubbles (bubbles may be removed by using “Remove IC Air” or “Remove EC Air” and using “Wash” as the inlet source.
 - 4) Open or set up program for coating the bioreactor (Figure 1. Steps 3-5).
 - 5) Run the program

- 6) While the program is running to coat the bioreactor, prepare a media bag with 4L of PTT6 media.
- 7) Attach the media bag to the IC Media line using a sterile tube connector.
- 8) When the bioreactor coating steps are completed, detach the cell inlet bag used for fibronectin solution using a RF sealer.

5

- c) Washing off excess fibronectin
- d) Conditioning the bioreactor with media

2. Culturing the cells in the Quantum Bioreactor

- a) Loading and attaching the cells with Uniform Suspension:
- 10 b) Feeding and cultivation of the cells
 - 1) Chose media flow rate to feed the cells.
 - 2) Sample for lactate and glucose everyday.
 - 3) Adjust the flow rate of the media as the lactate levels increase. The actual maximal tolerable lactate concentration will be defined by a flask culture from which the cells originate. Determine if adequate PTT6 media is in the media bag.
 - 15 If necessary, replace the PTT6 media bag with a fresh PTT6 media bag.
 - 4) When the flow rate has reached the desired value, measure lactate level every 8-12 hours. If the lactate level does not decrease or if the lactate level continues to increase, harvest the cells.

20 3. Harvesting the cells from the Quantum Bioreactor

- a) When lactate concentration does not decrease, harvest the cells after sampling for lactate and glucose for the last time.
- b) Harvesting the cells:
 - 1) Attach cell inlet bag filled with 100ml TrypLE to the “Reagent” line using a sterile tube connector.
 - 25 2) Confirm sufficient PBS is in the PBS bag. If not, attach a new bag with at least 1.7 liters of PBS to the “Wash” line using a sterile tube connector.
 - 3) Run the Harvest program

4. Cryopreserving the cells

- 30 1) Once the cells have been harvested, transfer the cells to 50ml centrifuge tube to pellet the cells.
- 2) Resuspend using 25ml of cold cell suspension solution. Count the cells using Sysmex or Biorad Cell counter. Attach the cell count report to the respective Quantum Processing Batch Record.

- 3) Adjust cell concentration to $2 \times 10^7/\text{ml}$
- 4) Add equal volume cryopreservation solution and mix well (do not shake or vortex)
- 5) Using a repeat pipettor, add 1ml of the cell suspension in cryopreservative to each 1.8ml vial. Cryopreserve using the CRF program as described in the SOP D6.100
- 5 CB Cryopreservation Using Controlled Rate Freezers
- 6) Store the vials in a designated liquid nitrogen storage space.
- 7) Attach the CRF run report to the form respective MSC P3-Quantum Processing Batch Record.

10 **3. Analysis of Stem Cell Marker Expression in Mesenchymal Cord Lining Stem Populations isolated from umbilical cord tissue, using different culture media**

Flow cytometry experiments were carried out to to analyse mesenchymal stem cells isolated from the umbilical cord for the expression of the mesenchymal stem cell markers CD73, CD90 and CD105.

15

For these experiments, mesenchymal stem cells were isolated from umbilical cord tissue by cultivation of the umbilical cord tissue in three different cultivation media, followed by subculturing of the mesenchymal stem cells in the respective medium as set forth in Example 2.

20

The three following culture media were used in these experiments: a) 90% (v/v) DMEM supplemented with 10 % FBS (v/v), b) the culture medium PTT-4 described in US patent application 2006/0078993 and the corresponding International patent application WO2006/019357 that consist of 90% (v/v) CMRL1066, and 10% (v/v) FBS (see paragraph 25 [0183] of WO2006/019357 and c) the culture medium of the present invention PPT-6 the composition of which is described herein. In this flow cytometry analysis, two different samples of the cord lining mesenchymal stem cell (CLMC) population were analysed for each of the three used culture media.

The following protocol was used for the flow cytometry analysis.

30 **Materials and methods**

Instruments name	Company Name	Serial Name
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BD FACS CANDO	BD	V07300367
Inverted Microscope, CKX41SF	Olympus	4K40846
Centrifuge, Micro spin Tabletop	Biosan	010213-1201-0003

Reagent list	Company Name	CatLog Number
10 X Trypsin	Biowest	X0930-100
10 X PBS	Lonza	17-517Q
DMEM	Lonza	12-604F
Fetal Bovine Serum	GE healthcare	A11-151

Antibody list	Company Name	CatLog Number
Human CD73 Purified AD2 0.1mg	BD	550256
Human CD90 Purified 5E10 1mL	BD	550402
Human CD105 Purified 266 0.1mg	BD	555690
Alexa Fluor 647 goat anti-mouse IgG (H+L) *2 mg/mL*	BD	A21235

Reagents name	Composition
1 X PBS (1L)	100ml of 10 X PBS + 900ml of sterile distilled H2O
1x PBA (50ml)	49.5ml of 1XPBS + 0.5 ml of FBS

Procedure

5 a) *Cell isolation and cultivation from the umbilical cord lining membrane*

1. Explant tissue samples were incubated in a cell culture plate and submerged in the respective medium, then keep it in CO₂ incubator at 37°C as explained in Example 2.
2. The medium was changed every 3 days.

3. Cell outgrowth from tissue culture explants was monitored under light microscopy.
4. At a confluence of about 70%, cells were separated from dish by trypsinization (0.0125% trypsin/0.05% EDTA) and used for flow cytometry experiments.

5 ***b) Trypsinization of cells for experiments***

1. Remove medium from cell culture plate
2. Gently rinse with sterile 1X PBS to remove traces of FBS as FBS will interfere with the enzymatic action of trypsin.
3. Add 1X trypsin to cell culture plate and incubate for 3-5 min in 37 °C.
- 10 4. Observe cells under microscope to ensure that they are dislodged. Neutralize trypsin by adding complete media containing FBS (DMEM with 10% FBS).
5. Use a pipette to break up cell clumps by pipetting cells in media against a wall of the plate. Collect and transfer cell suspension into 50 ml centrifuge tubes
6. Add sterile 1X PBS to plate and rinse it, Collect cell suspension into the same
- 15 centrifuge tube.
7. Centrifuge it at 1800 rpm for 10 mins.
8. Discard supernatant and re-suspend cell pellet with PBA medium.

c) Counting cells

- 20 1. Ensure that the haemocytometer and its cover slip are clean and dry, preferably by washing them with 70% ethanol and letting them dry before wiping them with Kim wipes (lint-free paper).
2. Aliquot a small amount of cells in suspension into a micro centrifuge tube and remove from the BSC hood.
- 25 3. Stain cells in suspension with an equal volume of Trypan Blue, e.g. to 500µl of suspension add 500µl of Trypan Blue (dilution factor = 2X, resulting in 0.2% Trypan Blue solution).
4. Avoid exposure of cells to Trypan Blue for longer than 30 mins as Trypan Blue is toxic and will lead to an increase in non-viable cells, giving a false cell count.
- 30 5. Add 20 µl of the cell suspension mixture to each chamber of a haemocytometer and view under a light microscope.
 - a. Count the number of viable cells (bright cells; non-viable cells take up Trypan Blue readily and thus are dark) in each quadrant of the haemocytometer for a total of 8 quadrants in the upper and lower chamber.

Total cell count is given as (Average number of cells/quadrant) x 10⁴ cells/ml.

d) Staining cells

i. Preparation before staining cells

- 5 • Cell suspension are aliquot into 3 tubes (CD73, CD90, CD105) in duplicates and 2 tubes (negative control), each containing 50,000 cells.

ii. Staining with primary antibody (Ab)

- 10 • Add 1 µl [0.5mg/ml Ab] of primary antibody to 100ul cell suspension. Incubate at 4°C for 45 min.
- Make up to 1ml with PBA.
- Centrifuge 8000 rpm at 4°C for 5 mins.
- Remove supernatant.
- Add 1ml of PBA and re-suspend pellet
- Centrifuge 8000 rpm at 4°C for 5 mins.
- 15 • Remove supernatant.
- Re-suspend in 100ul PBA.

iii. Staining with secondary Ab – in the dark

- 20 • Add 1ul [0.5mg/ml ab] of secondary antibody to 100ul cell suspension. Incubate at 4°C for 30 min.
- Make up to 1ml with PBA.
- Centrifuge 8000 rpm at 4°C for 5 mins.
- Remove supernatant.
- Add 1ml of PBA and re-suspend pellet
- Centrifuge 8000 rpm at 4°C for 5 mins.
- 25 • Remove supernatant
- Re-suspend in 200-300ul PBA for flow cytometry analysis
- Transfer cells to FACS tube for reading in BD FACS CANDO flow cytometry.

30 The results of the flow cytometry analysis are shown in **Fig. 6a to Fig.6c**. **Fig. 6a** shows the percentage of isolated mesenchymal cord lining stem cells expressing stem cell markers CD73, CD90 and CD105 after isolation from umbilical cord tissue and cultivation in DMEM/10% FBS, **Fig. 6b** shows the percentage of isolated mesenchymal cord lining stem

cells expressing stem cell markers CD73, CD90 and CD105 after isolation from umbilical cord tissue and cultivation in PTT-4 and **Fig. 6c** shows the percentage of isolated mesenchymal cord lining stem cells expressing stem cell markers CD73, CD90 and CD105 after isolation from umbilical cord tissue and cultivation in PTT-6. As can be seen from **Fig. 6a**, the population isolated using DMEM/10 % FBS as culture medium cultivation has about 75% CD73+ cells, 78 % 90+ cells and 80 % CD105+ cells (average of two experiments), while after isolation/cultivation of umbilical cord tissue using PPT-4 culture medium (see **Fig. 6b**) the number of mesenchymal stem cells that are CD73-positive, CD90-positive and CD105-positive are about 87 % (CD73+ cells), 93 % /CD90+ cells) and 86 % (CD105+ cells) average of two experiments. The purity of the mesenchymal stem cell population that was obtained by means of cultivation in the PTT-6 medium of the present invention is at least 99.0 % with respect to all three markers (CD73, CD90, CD105), meaning the purity of this cell population is significant higher than for cultivation using PPT-4 medium or DMEM/10 % FBS. In addition and even more importantly, the mesenchymal stem cell population obtained by means of cultivation in PTT-6 is essentially a 100% pure and defined stem cell population. This makes the stem cell population of the present invention the ideal candidate for stem cell based therapies. Thus, this population of mesenchymal cord lining stem cells may become the gold standard for such stem cell based therapeutic approaches.

The findings shown in Fig. 6 are further corroborated by the results of the flow cytometry analysis that are shown in **Fig. 7a** and **Fig.7b**. **Fig. 7a** shows the percentage of isolated mesenchymal cord lining stem cells (mesenchymal stem cells of the amniotic membrane of umbilical cord) that express the stem cell markers CD73, CD90 and CD105 and lack expression of CD34, CD45 and HLA-DR after isolation from umbilical cord tissue and cultivation in PTT-6 medium. As shown in **Fig. 7a**, the mesenchymal stem cell population contained 97.5 % viable cells of which 100 % expressed each of CD73, CD90 and CD105 (see the rows “CD73+CD90+” and “CD73+CD105+”) while 99.2 % of the stem cell population did not express CD45 and 100 % of the stem cell population did not express CD34 and HLA-DR (see the rows “CD34-CD45- and “CD34-HLA-DR-). Thus, the mesenchymal stem cells population obtained by cultivation in PTT-6 medium is essentially a 100% pure and defined stem cell population that meets the criteria that mesenchymal stem cells are to fulfill to be used for cell therapy (95% or more of the stem cell population express CD73, CD90 and CD105, while 98 % or more of the stem cell population lack expression of CD34, CD45 and HLA-DR, see Sensebe et al.”Production of mesenchymal stromal/stem cells according to good

manufacturing practices: a review”, *supra*). It is noted here that the present mesenchymal stem cells of the amniotic membrane adhere to plastic in standard culture conditions and differentiate *in vitro* into osteoblasts, adipocytes and chondroblasts, see US patents 9,085,755, US patent 8,287,854 or WO2007/046775 and thus meet the criteria generally accepted for use
 5 of mesenchymal stem cells in cellular therapy.

Fig. 7b shows the percentage of isolated bone marrow mesenchymal stem cells that express CD73, CD90 and CD105 and lack expression of CD34, CD45 and HLA-DR. As shown in **Fig. 7b**, the bone marrow mesenchymal stem cell population contained 94.3 % viable cells of which 100 % expressed each of CD73, CD90 and CD105 (see the rows “CD73+CD90+” and “CD73+CD105+”) while only 62.8 % of the bone marrow stem cell population lacked expression of CD45 and 99.9 % of the stem cell population lacked expression CD34 and HLA-DR (see the rows “CD34-CD45-” and “CD34-HLA-DR-”). Thus, the bone marrow mesenchymal stem cells that are considered to be gold standard of mesenchymal stem cells are
 15 by far less homogenous/pure in terms of stem cell marker than the mesenchymal stem cells population (of the amniotic membrane of the umbilical cord) of the present application. This finding also shows that the stem cell population of the present invention may be the ideal candidate for stem cell based therapies and may become the gold standard for stem cell based therapeutic approaches.

20 It will be readily apparent to a person skilled in the art that varying substitutions and modifications may be made to the invention disclosed herein without departing from the scope and spirit of the invention.

25 All patents and publications mentioned in the specification are indicative of the levels of those of ordinary skill in the art to which the invention pertains. All patents and publications are herein incorporated by reference to the same extent as if each individual publication was specifically and individually indicated to be incorporated by reference.

30 The inventions illustratively described herein may suitably be practiced in the absence of any element or elements, limitation or limitations, not specifically disclosed herein. Thus, for example, the terms "comprising", "including", "containing", etc. shall be read expansively and without limitation. Additionally, the terms and expressions employed herein have been used as terms of description and not of limitation, and there is no intention in the use of such terms and

expressions of excluding any equivalents of the features shown and described or portions thereof, but it is recognized that various modifications are possible within the scope of the invention claimed. Thus, it should be understood that although the present invention has been specifically disclosed by preferred embodiments and optional features, modification and
5 variation of the inventions embodied therein herein disclosed may be resorted to by those skilled in the art, and that such modifications and variations are considered to be within the scope of this invention. The invention has been described broadly and generically herein. Each of the narrower species and subgeneric groupings falling within the generic disclosure also form part of the invention. This includes the generic description of the invention with a
10 proviso or negative limitation removing any subject matter from the genus, regardless of whether or not the excised material is specifically recited herein. In addition, where features or aspects of the invention are described in terms of Markush groups, those skilled in the art will recognize that the invention is also thereby described in terms of any individual member or subgroup of members of the Markush group. Further embodiments of the invention will
15 become apparent from the following claims.

What is claimed is:

1. A method of isolating a mesenchymal stem cell population from the amniotic membrane of the umbilical cord, the method comprising cultivating umbilical cord tissue in a culture medium comprising DMEM (Dulbecco's modified eagle medium), F12 (Ham's F12 Medium), M171 (Medium 171) and FBS (Fetal Bovine Serum).
2. The method of claim 1, wherein the culture medium comprises DMEM in a final concentration of about 55 to 65 % (v/v), F12 in a final concentration of about 5 to 15 % (v/v) , M171 in a final concentration of about 15 to 30 % (v/v) and FBS in a final concentration of about 1 to 8 % (v/v).
3. The method of claim 2, wherein the culture medium comprises DMEM in a final concentration of about 57.5 to 62.5 % (v/v), F12 in a final concentration of about 7.5 to 12.5 % (v/v) , M171 in a final concentration of about 17.5 to 25.0 % (v/v) and FBS in a final concentration of about 1.75 to 3.5 % (v/v).
4. The method of claim 3, wherein the culture medium comprises DMEM in a final concentration of about 61.8 % (v/v), F12 in a final concentration of about 11.8 % (v/v) , M171 in a final concentration of about 23.6 % (v/v) and FBS in a final concentration of about 2.5 % (v/v).
5. The method of any of claims 1 to 4, wherein the culture medium further comprises Epidermal Growth Factor (EGF) in a final concentration of about 1 ng/ml to about 20 ng/ml.
6. The method of any of claims 1 to 5, wherein the culture medium comprises EGF in a final concentration of about 10ng/ml.
7. The method of any of claims 1 to 6, wherein the culture medium comprises Insulin in a final concentration of about 1 µg/ml to 10 µg/ml.
8. The method of any of claims 1 to 7, wherein the culture medium comprises Insulin in a final concentration of about 5µg/ml.
9. The method of any of the foregoing claims, wherein the culture medium further comprises at least one of the following supplements: adenine, hydrocortisone, and 3,3',5-Triiodo-L-thyronine sodium salt (T3).

10. The method of any of the foregoing claims, wherein the culture medium comprises all three of adenine, hydrocortisone, and 3,3',5-Triiodo-L-thyronine sodium salt (T3).
11. The method of claim 10, wherein the culture medium comprises adenine in a final concentration of about 0.01 to about 0.1 $\mu\text{g/ml}$ adenine, hydrocortisone in a final concentration of about 0.1 to about 10 $\mu\text{g/ml}$ hydrocortisone and/or 3,3',5-Triiodo-L-thyronine sodium salt (T3) in a final concentration of about 0.5 to about 5 ng/ml.
12. The method of any of the foregoing claims, comprising culturing the umbilical cord tissue till the cell outgrowth of the mesenchymal stem cells of the amniotic membrane reaches about 70-80% confluency.
13. The method of claim 12, comprising removing the mesenchymal stem cells from the cultivation container used for the cultivation.
14. The method of claim 13, wherein removing the mesenchymal stem cells from the cultivation container is carried out by enzymatic treatment.
15. The method of claim 14, wherein the enzymatic treatment comprises trypsination.
16. The method of any of claims 13 to 15, wherein the mesenchymal stem cells are transferred for subculturing to a cultivation container for subculturing.
17. The method of claim 16, wherein the mesenchymal cells are suspended for subculturing at a concentration 1.0×10^6 cells/ml.
18. The method of claim 17, wherein the mesenchymal stem cells are subcultured in a culture medium as defined in any of the claims 1 to 10.
19. The method of claim 18, wherein the mesenchymal stem cells are subcultured till the mesenchymal stem cells reach about 70-80% confluency.
20. The method of any of claims 16 to 19, wherein the subculturing is carried out in a self-contained bioreactor.
21. The method of claim 20, wherein the bioreactor is selected from the group consisting of a parallel-plate bioreactor, a hollow-fiber bioreactor and and a micro-fluidic bioreactor.
22. The method of any of the foregoing claims, wherein the umbilical cord tissue is a piece of the entire umbilical cord or the amniotic membrane of the umbilical cord.

23. The method of any of the foregoing claims wherein cultivation is carried out in a CO₂ cell culture incubator at a temperature 37°C.

24. The method of claim 23, comprising removing the mesenchymal stem cells from the cultivation container used for the subcultivation.

25. The method of claim 24, wherein removing the mesenchymal stem cells from the cultivation container is carried out by enzymatic treatment.

26. The method of claim 25, wherein the enzymatic treatment comprises trypsination.

27. The method of claim 26, further comprising collecting the isolated mesenchymal stem cells.

28. The method of any of the foregoing claims, wherein at least about 90 % or more, about 91 % or more, about 92 % or more, about 92 % or more, about 93 % or more, about 94 % or more, about 95 % or more, about 96 % or more, about 97 % or more, about 98 % or more about 99 % or more of the isolated mesenchymal stem cells express the following markers: CD73, CD90 and CD105.

29. The method of any of the foregoing claims, wherein at least about 90 % or more, about 91 % or more, about 92 % or more, about 92 % or more, about 93 % or more, about 94 % or more, about 95 % or more, about 96 % or more, about 97 % or more, about 98 % or more about 99 % or more of the isolated mesenchymal stem cells lack expression of the following markers: CD34, CD45 and HLA-DR (Human Leukocyte Antigen – antigen D Related).

30. The method of any of claims 28 or 29, wherein about 97 % or more, about 98 % or more about 99 % or more of the isolated mesenchymal stem cells express CD73, CD90 and CD105 and lack expression of CD34, CD45 and HLA-DR.

31. The method of any of the foregoing claims, further comprising preserving the isolated stem/progenitor cells for further use.

32. The method of claim 31, wherein preserving is carried out by cryo-preservation.

33. An isolated mesenchymal stem population of the amniotic membrane of the umbilical cord, wherein at least 97 % or more cells of the isolated mesenchymal stem cell population express each of CD73, CD90 and CD105 and lack expression of each of CD34, CD45 and HLA-DR.

34. The mesenchymal stem cell population of claim 33, wherein at least about 98 % or more about 99 % or more cells of the isolated mesenchymal stem cell population express each of CD73, CD90 and CD105 and lack expression of each of CD34, CD45 and HLA-DR.

35. The mesenchymal stem cell population of any of claims 33 to 34, wherein the population is obtainable by the method as defined in any of claims 1 to 30.

36. The mesenchymal stem cell population of any of claims 33 to 34, wherein the population is obtained by the method as defined in any of claims 1 to 30.

37. A pharmaceutical composition comprising an isolated mesenchymal stem population of the amniotic membrane of the umbilical cord, wherein at least 97 % or more cells of the stem cell population express each of the following markers: CD73, CD90 and CD105 and lack expression of each of the following markers: CD34, CD45 and HLA-DR.

38. The pharmaceutical composition of claim 37, wherein the pharmaceutical composition is adapted for systemic or topical application.

39. The pharmaceutical composition of claim 37 or 38, further comprising a pharmaceutically acceptable excipient.

40. A method of making a culture medium suitable for isolating a mesenchymal stem cell population from the amniotic membrane of the umbilical cord, the method comprising mixing to obtain a final volume of 500 ml culture medium:

i. 250 ml of DMEM

ii. 118 ml M171

iii. 118 ml DMEM/F12

iv. 12.5 ml Fetal Bovine Serum (FBS) (final concentration of 2.5%)

41. The method of claim 40, further comprising adding

v. 1 ml EGF stock solution (5 µg/ml) to achieve a final concentration of 10ng/ml)

vi. Insulin 0.175 ml stock solution (14.28 mg/ml) to achieve a final concentration of 5µg/ml.

42. The method of claim 40 or 41, further comprising adding to DMEM one or more of the following supplements: adenine, hydrocortisone, 3,3',5-Triiodo-L-thyronine sodium salt (T3), thereby reaching a total volume of 500 ml culture medium.

43. The method of claim 41, wherein the final concentration of the supplements in DMEM are as follows.

about 0.05 to 0.1 $\mu\text{g/ml}$ adenine, for example about 0.025 $\mu\text{g/ml}$ adenine,

about 1 to 10 $\mu\text{g/ml}$ hydrocortisone,

about 0.5 to 5 ng/ml 3,3',5-Triiodo-L-thyronine sodium salt (T3), for example 1.36 ng/ml 3,3',5-Triiodo-L-thyronine sodium salt (T3).

44. A cell culture medium obtainable by the method of any of claims 40 to 43.

45. A method of isolating mesenchymal stem cells from the amniotic membrane of the umbilical cord, comprising cultivating amniotic membrane tissue in the culture medium prepared by the method as defined in any of claims 40 to 43.

46. A cell culture medium comprising:

- DMEM in the final concentration of about 55 to 65 % (v/v),
- F12 in a final concentration of about 5 to 15 % (v/v),
- M171 in a final concentration of about 15 to 30 % (v/v) and
- FBS in a final concentration of about 1 to 8 % (v/v).

47. The cell culture medium of claim 46, wherein the culture medium comprises DMEM in the final concentration of about 57.5 to 62.5 % (v/v), F12 in a final concentration of about 7.5 to 12.5 % (v/v), M171 in a final concentration of about 17.5 to 25.0 % (v/v) and FBS in a final concentration of about 1.75 to 3.5 % (v/v).

48. The cell culture medium of claim 47, wherein the culture medium comprises DMEM in a final concentration of about 61.8 % (v/v), F12 in a final concentration of about 11.8 % (v/v), M171 in a final concentration of about 23.6 % (v/v) and FBS in a final concentration of about 2.5 % (v/v).

49. The cell culture medium of any of claims 46 to 48, wherein the culture medium further comprises Epidermal Growth Factor (EGF) in a final concentration of about 1 ng/ml to about 20 ng/ml .

50. The cell culture medium of any of claims 46 to 49, wherein the culture medium comprise EGF in a final concentration of about 10 ng/ml .

51. The cell culture medium of any of claims 46 to 50, wherein the culture medium comprises Insulin in a final concentration of about 1 µg/ml to 10 µg/ml.

52. The cell culture medium of claim 51, wherein the culture medium comprises Insulin in a final concentration of about 5µg/ml.

53. The cell culture medium of any of claims 46 to 52, wherein the culture medium further comprises at least one of the following supplements: adenine, hydrocortisone, and 3,3',5-Triiodo-L-thyronine sodium salt (T3).

54. The cell culture medium of claim 53, wherein the culture medium comprises all three of adenine, hydrocortisone, and 3,3',5-Triiodo-L-thyronine sodium salt (T3).

55. The cell culture medium of claim 53 or 54, wherein the culture medium comprises adenine in a final concentration of about 0.05 to about 0.1 µg/ml adenine, hydrocortisone in a final concentration of about 1 to about 10 µg/ml hydrocortisone and/or 3,3',5-Triiodo-L-thyronine sodium salt (T3) in a final concentration of about 0.5 to about 5 ng/ml.

56. The cell culture medium of any of claims 46 to 55, wherein 500 ml of the cell culture medium comprise:

i. 250 ml of DMEM

ii. 118 ml M171

iii. 118 ml DMEM/F12

iv. 12.5 ml Fetal Bovine Serum (FBS) (final concentration of 2.5%)

57. The cell culture medium of claim 56, further comprising

v. EGF in a final concentration of 10ng/ml

vi. Insulin in a final concentration of 5µg/ml.

vi. Insulin 0.175 ml (final concentration of 5µg/ml)

58. The cell culture medium of claim 56 or 57, further comprising adenine in a final concentration of about 0.05 to about 0.1 µg/ml adenine, hydrocortisone in a final concentration of about 1 to about 10 µg/ml hydrocortisone and/or 3,3',5-Triiodo-L-thyronine sodium salt (T3) in a final concentration of about 0.5 to about 5 ng/ml.

59. The use of a cell culture medium as defined in any of claims 46 to 58 for isolation of mesenchymal stem cells from the amniotic membrane of umbilical cord.

60. The use of a cell culture medium as defined in any of claims 46 to 58 for cultivation of mesenchymal stem cells from the amniotic membrane of umbilical cord.

Fig. 1

Dulbecco's modified eagle medium (DMEM)

Product use

Dulbecco's modified eagle medium was developed in 1969 and is a modification of basal medium eagle (BME) that differs from BME and MEM by the following characteristics:

- Vitamins 4X greater than MEM. Vitamins and amino acids greater than BME
- Types and quantities of amino acids greater than MEM and BME
- Iron (ferric nitrate)

Description

- 12-604 With 4.5 g/L glucose, with L-glutamine
12-614 With 4.5 g/L glucose, without L-glutamine
12-707 With 1.0 g/L glucose, without L-glutamine
12-708 With 1.0 g/L glucose, and 25 mM HEPES buffer, without L-glutamine
12-709 With 4.5 g/L glucose, and 25 mM HEPES buffer, without L-glutamine
12-733 With 4.5 g/L glucose, without L-glutamine, without sodium pyruvate
12-741 With 4.5 g/L glucose, with L-glutamine, without sodium pyruvate
12-914 With 4.5 g/L glucose, without L-glutamine, screened to support hybridoma growth
12-917 With 4.5 g/L glucose, without L-glutamine or phenol red
15-604 With 4.5 g/L glucose, with L-glutamine, without sodium pyruvate, powder
15-614 With 4.5 g/L glucose, without L-glutamine or sodium pyruvate, powder
(Powder formulations require the addition of 49.3 mL/L of NaHCO₃ 7.5% solution or 3.70 g/L of NaHCO₃ powder)

Quick reference chart

	Glucose	L-glutamine	Phenol red	HEPES buffer	Sodium pyruvate
12-604	4.5 g/L	+	+	-	+
12-614	4.5 g/L	-	+	-	+
12-707	1.0 g/L	-	+	-	+
12-708	1.0 g/L	-	+	+	+
12-709	4.5 g/L	-	+	+	+
12-733	4.5 g/L	-	+	-	-
12-741	4.5 g/L	+	+	-	-
12-914	4.5 g/L	-	+	-	+
12-917	4.5 g/L	-	-	-	+

Fig. 1 continued

2/12

Specifications

	Sterility	pH	Osmolality (mOsm)	Cell growth promotion (% of control)	Endotoxin (EU/ml)
12-604	Neg.	7.0-7.4	324-352	≥75%	FIO
12-614	Neg.	7.0-7.4	324-352	≥75%	FIO
12-707	Neg.	7.0-7.4	306-346	≥75%	FIO
12-708	Neg.	7.03-7.27	300-326	≥75%	FIO
12-709	Neg.	7.0-7.4	321-351	≥75%	FIO
12-733	Neg.	7.0-7.4	318-360	≥75%	FIO
12-741	Neg.	7.0-7.4	318-360	≥75%	FIO
12-914	Neg.	7.0-7.4	324-352	≥85% **	FIO
12-917	Neg.	7.0-7.4	329-343	≥75%	FIO
15-604 ***	-	5.5-7.0	235-265	≥75%	≤1.0
15-614 ***	-	5.5-7.0	235-265	≥75%	≤1.0

FIO – for information only

** Result of functional test

*** Moisture content #1.0

Storage

2°C to 8°C

Product use statement

THESE PRODUCTS ARE FOR RESEARCH USE ONLY. Not approved for human or veterinary use, for application to humans or animals, or for use in clinical or *in vitro* procedures.

Ordering information

Catalog number	Description	Size
12-604F	DMEM with 4.5 g/L glucose, with L-glutamine	500 ml
12-604Q	DMEM with 4.5 g/L glucose, with L-glutamine	1 L
12-614F	DMEM with 4.5 g/L glucose, without L-glutamine	500 ml
12-614Q	DMEM with 4.5 g/L glucose, without L-glutamine	1 L
12-707F	DMEM with 1.0 g/L glucose, without L-glutamine	500 ml
12-708F	DMEM with 1.0 g/L glucose, and 25 mM HEPES buffer, without L-glutamine	500 ml
12-709F	DMEM with 4.5 g/L glucose, and 25 mM HEPES buffer, without L-glutamine	500 ml
12-733F	DMEM with 4.5 g/L glucose, without L-glutamine, without sodium pyruvate	500 ml
12-733Q	DMEM with 4.5 g/L glucose, without L-glutamine, without sodium pyruvate	1 L
12-741F	DMEM with 4.5 g/L glucose, with L-glutamine, without sodium pyruvate	500 ml
12-914F	DMEM with 4.5 g/L glucose, without L-glutamine, screened to support hybridoma growth.	500 ml
12-917F	DMEM with 4.5 g/L glucose, without L-glutamine or phenol red.	500 ml
15-604D	DMEM with 4.5 g/L glucose, with L-glutamine, without sodium pyruvate, powder	1 x 10L
15-604F	DMEM with 4.5 g/L glucose, without L-glutamine or sodium pyruvate, powder	1 X 50L
15-614D	DMEM with 4.5 g/L glucose, without L-glutamine or sodium pyruvate, powder	1 x 10L

Lonza

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www.lonza.com
biotechserv@lonza.com
Tech Service: 800-521-0390
Document # TS 12-615-2 11/08
Walkersville, MD 21793-0127 USA
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Fig. 2

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Ham's F12 Medium

Product Use

Ham's F12 Medium is a nutrient mixture designed to cultivate a wide variety of mammalian and hybridoma cells when used with serum in combination with hormones and transferrin.

Specifications

Sterility	pH	Osmolality (mOsm)
Negative	7.07-7.40	286-305
Cell Growth Generation	Endotoxin (EU/ml)	
≥75% of control	FIO	

Storage

2°C to 8°C

Product Use Statement

THESE PRODUCTS ARE FOR RESEARCH USE ONLY. Not approved for human or veterinary use, for application to humans or animals, or for use in clinical or *in vitro* procedures.

Ordering Information

Catalog Number	Description	Size
12-615F	Ham's F12 Medium with L-glutamine	500 ml

Lonza

www.lonza.com
U.S. Scientific Support: 800-521-0390
scientific.support@lonza.com
EU/ROW Scientific Support: +49-221-99199-400
scientific.support.eu@lonza.com
Document # TS-12-719-3 08/11
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Fig. 3

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DMEM: F12 (1:1) medium

Product use

DMEM combined with Hams F12 has been extensively used to demonstrate the effect of various hormones and growth factors on target tissues.

Description

- 12-719 with L-glutamine, 15 mM HEPES, and 3.151 g/L glucose
- 15-719 powder – with L-glutamine, 15 mM HEPES, and 3.151 g/L glucose. Powdered medium requires the addition of 16.0 ml/L of NaHCO₃, 7.5% solution or 1.2 g/L NaHCO₃ powder
- 04-687 with 3.151 g/L glucose, with L-glutamine, without HEPES

Specifications

	Sterility	pH	Osmolality (mOsm)
12-719	Negative	7.0-7.4	286-356
15-719*	Negative	4.5-6.5	260-290
04-687	Negative	FIO	FIO
	Cell growth promotion	Endotoxin (EU/ml)	Moisture
12-719	≥ 75% of control	FIO	--
15-719*	≥ 75% of control	≤ 1 EU/ml	≤ 2%
04-687	--	FIO	--

* pH, osmolality, endotoxin, and moisture are done without NaHCO₃; NaHCO₃ is added for cell growth test.

Storage

2°C to 8°C

Product use statement

THESE PRODUCTS ARE FOR RESEARCH USE ONLY. Not approved for human or veterinary use, for application to humans or animals, or for use in clinical or *in vitro* procedures.

Ordering information

Catalog number	Description	Size
12-719F	DMEM: F12 with L-glutamine, 15 mM HEPES, and 3.151 g/L glucose	500 ml
12-719Q	DMEM: F12 with L-glutamine, 15 mM HEPES, and 3.151 g/L glucose	1 L
15-719D	DMEM: F12 powder – with L-glutamine, 15 mM HEPES, and 3.151 g/L glucose. Powdered medium requires the addition of 16.0 ml/L of NaHCO ₃ , 7.5% solution or 1.2 g/L NaHCO ₃ powder	1 x 10L
04-687Q	DMEM: F12 with 3.151 g/L glucose, with L-glutamine, without HEPES	1 L

Medium 171, Medium 171PRF, and MEGS

Medium 171

M-171-500

500 ml

Medium 171PRF (Phenol Red-Free)

M-171PRF-500

500 ml

Product Description

Medium 171 and Medium 171PRF are sterile, liquid tissue culture media intended for use as one component in a complete culture environment for the growth of normal human mammary epithelial cells. Medium 171 is a basal medium containing essential and non-essential amino acids, vitamins, other organic compounds, trace minerals, and inorganic salts. Medium 171PRF is a phenol red-free version of Medium 171. These media do not contain antibiotics, antimycotics, hormones, growth factors, or proteins. These media are HEPES and bicarbonate buffered and are designed for use in an incubator with an atmosphere of 5% CO₂/95% air. To support plating and long-term proliferation of normal human mammary epithelial cells, these media must be supplemented with Mammary Epithelial Growth Supplement (MEGS, cat. no S-015-5).

Intended Use

Medium 171 is intended for use in the routine culture of normal human mammary epithelial cells. Medium 171PRF is intended for use by investigators who wish to culture normal human mammary epithelial cells in the absence of phenol red. When supplemented with MEGS, these media will support the plating and proliferation of normal human mammary epithelial cells at densities between 2.5×10^3 cells/cm² and 8×10^4 cells/cm². ***This product is for research use only. Not for use in animals, humans, or diagnostic procedures.***

Caution: If handled improperly, some components of this product may present a health hazard. Take appropriate precautions when handling this product, including the wearing of protective clothing and eyewear. Dispose of properly.

Storage and Stability

Medium 171 and Medium 171PRF are stored at 4° C in our facility and are shipped at ambient temperature. Upon receipt, these media should be stored at 4° C and should not be frozen. **Protect from light.** Several components of these tissue culture media are light-labile, and we recommend that the media not be exposed to light for lengthy periods of time. If the media are warmed prior to use, do not exceed 37° C. When stored in the dark at 4° C, the product is stable until the expiration date on the label.

Preparation of Supplemented Medium 171

1. Thaw one bottle of MEGS. Take one bottle of Medium from cold storage. Make sure that the caps of the vessels are tight.
2. Gently swirl the bottle of supplement. Avoid splashing the supplement into the cap of the bottle or causing the supplement to foam.
3. Wipe the outside of the containers with a disinfecting solution such as 70% ethanol or isopropanol.
4. Using sterile technique in a laminar flow culture hood, transfer the entire contents of the bottle of supplement to the bottle of Medium.
5. Tightly cap the bottle of supplemented medium and swirl the contents to ensure a homogeneous solution. Avoid causing the medium to foam.

Storage and Stability of Supplemented Medium 171

Once Medium 171 or Medium 171PRF has been supplemented with MEGS, the supplemented medium should be stored in the dark at 4° C and should not be frozen. When stored in the dark at 4° C, the supplemented medium is stable for 1 month.

Selected References

The Medium 171 formulation is based on medium MCDB 170, with modifications.

Hammond SL, Ham RG, Stampfer MR; PNAS 81:5435-5439, 1984

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MEGS Mammary Epithelial Growth Supplement

Cat. no. S-015-5
5 ml

Product Description

Mammary Epithelial Growth Supplement (MEGS) is a sterile, concentrated (100X) solution intended for use as one component in a complete culture environment for the growth of normal human mammary epithelial cells. Each 5 ml bottle of MEGS contains all of the growth factors, hormones, and tissue extracts necessary for the culture of normal human mammary epithelial cells and is the correct amount of supplement for a 500 ml bottle of Medium 171 or Medium 171PRF. MEGS is an ionically-balanced supplement containing bovine pituitary extract (BPE), bovine insulin, hydrocortisone and recombinant human epidermal growth factor. When a 500 ml bottle of Medium 171 or Medium 171PRF is supplemented with MEGS, the final concentrations of the components in the supplemented medium are: BPE, 0.4% v/v; bovine insulin, 5 µg/ml; hydrocortisone, 0.5 µg/ml; and recombinant human epidermal growth factor, 3 ng/ml.

Intended Use

MEGS is intended for use in conjunction with Medium 171 or Medium 171PRF for the routine serum-free culture of normal human mammary epithelial cells. ***This product is for research use only. Not for use in animals, humans, or diagnostic procedures.***

Caution: If handled improperly, some components of this product may present a health hazard. Take appropriate precautions when handling this product, including the wearing of protective clothing and eyewear. Dispose of properly.

Limited Use Label License No. 5: Invitrogen Technology

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MAN0001585

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Fig. 5

PTT6 Medium ingredients list

Medium composition List	Company Name	Catalogue Number
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Basic media

DMEM (also referred to herein as PTT6-basal medium)	Lonza	12-604F
DMEM/F12	Lonza	12-719F
M171	Life technologies	M171500

Serum

Fetal Bovine Serum	GE Healthcare	A15-151
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Antibiotic

Penicillin-Streptomycin-Amphotericin B	Lonza	17-745E
--	-------	---------

Supplements

Adenine (optional)	Sigma	A8626-25G
Hydrocortisone (optional)	Sigma	H-0888
Epidermal Growth Factor	Millipore	GF-144
T3 (3,3',5-Triiodo-L-thyronine sodium salt) (optional)	Sigma	200-223-5
Recombinant Human Insulin AOF	Life Technologies	A11382IJ

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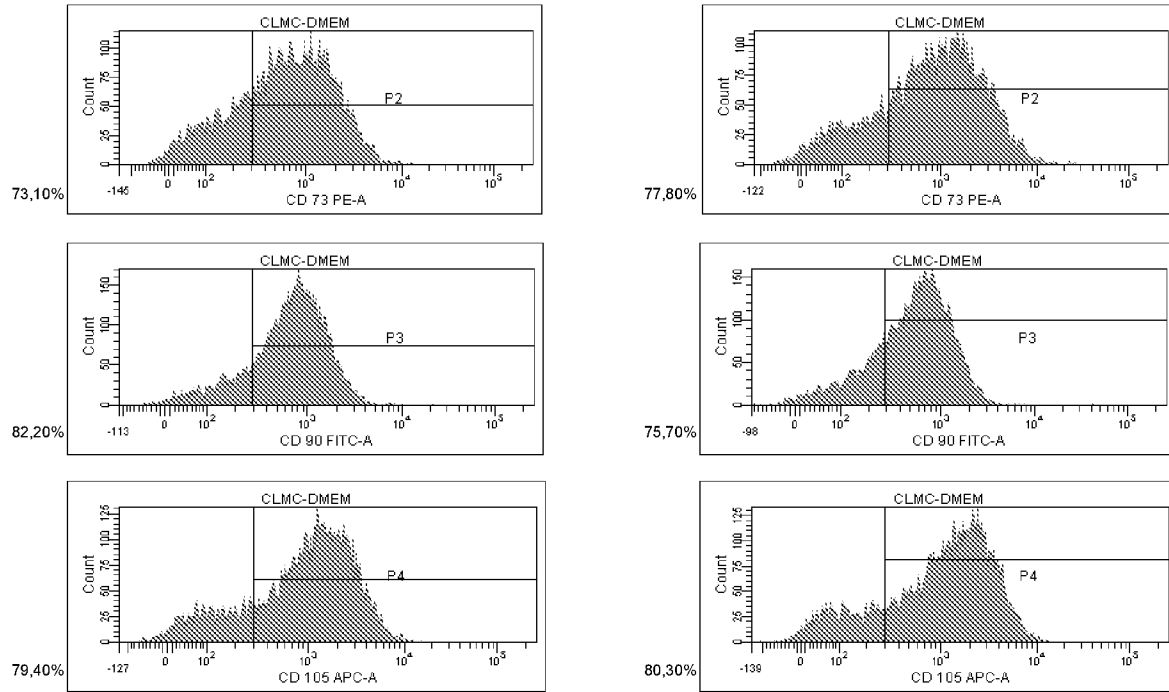


Fig. 6a

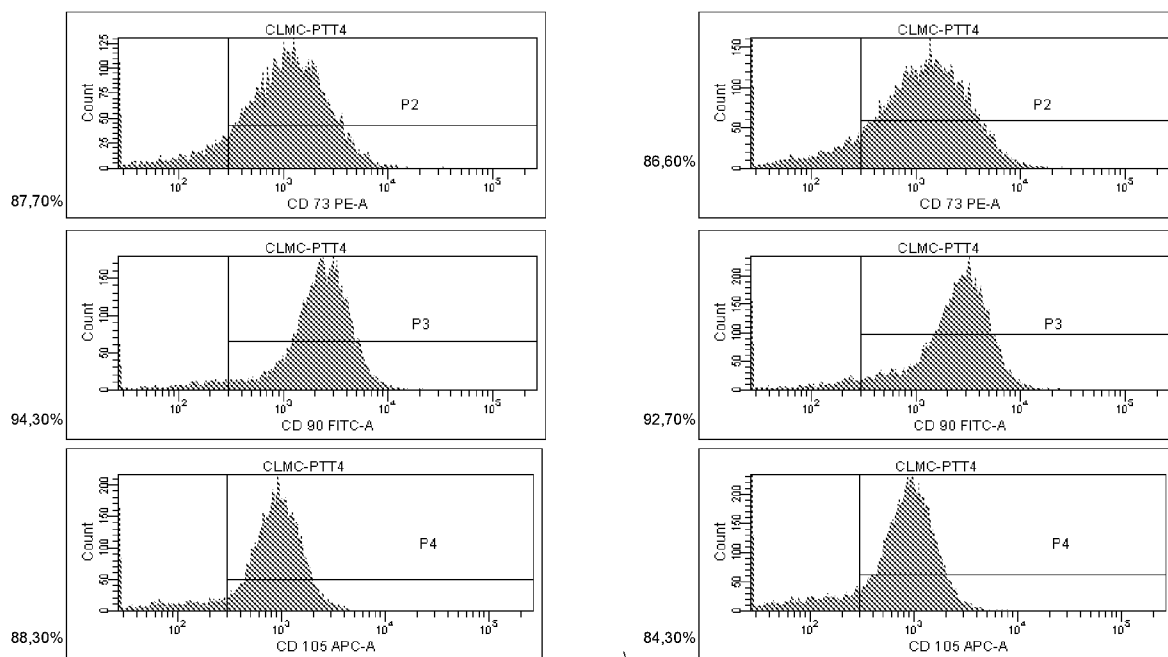


Fig. 6b

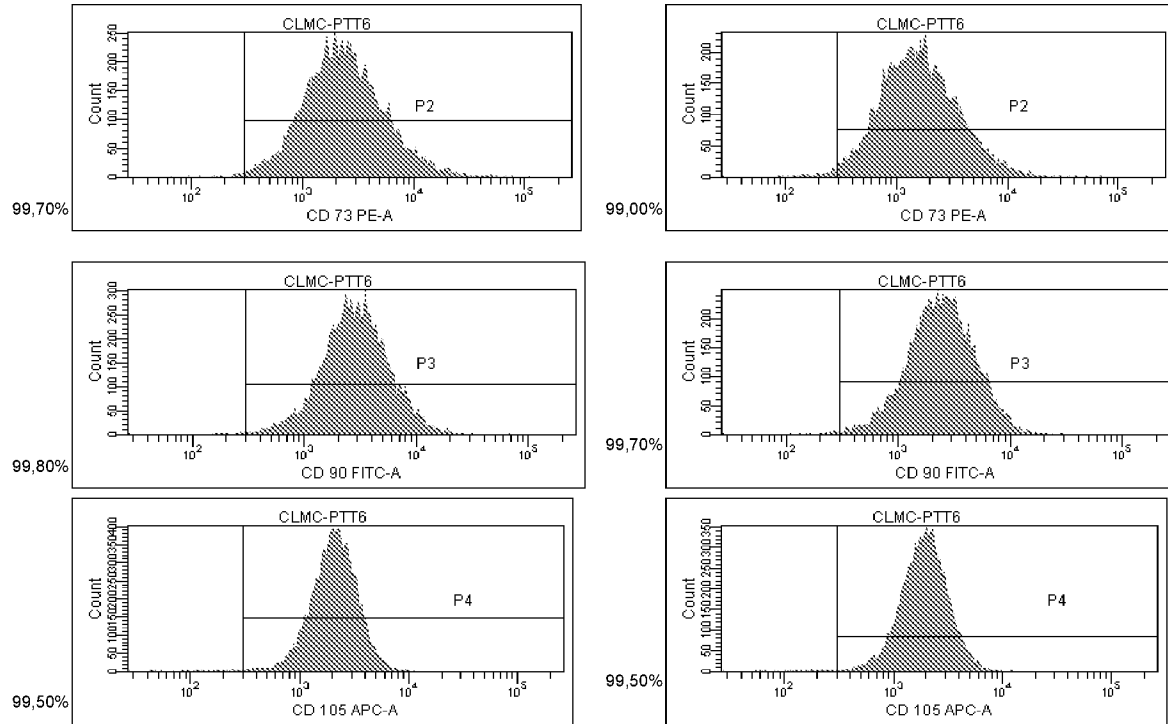
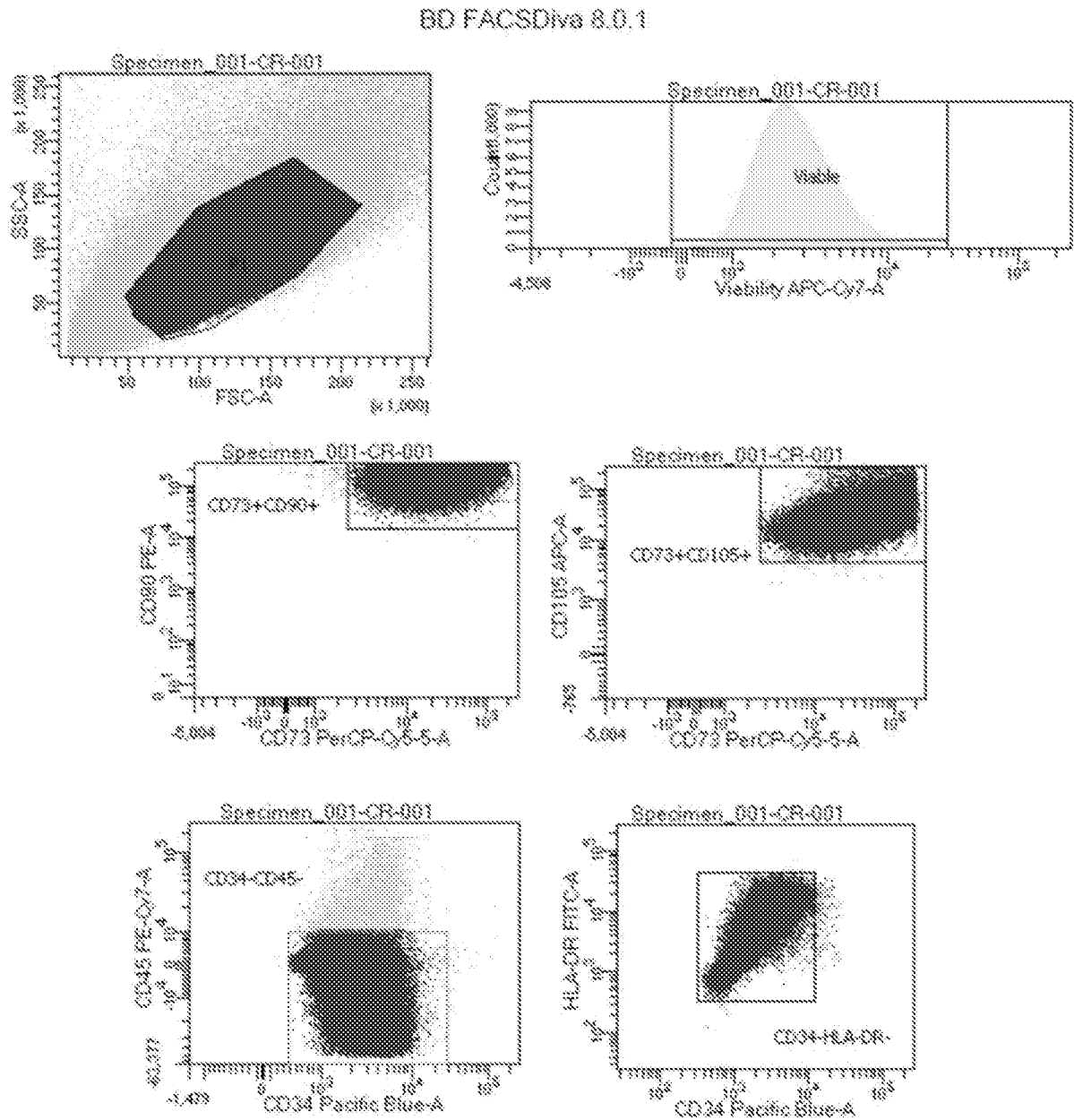


Fig. 6c

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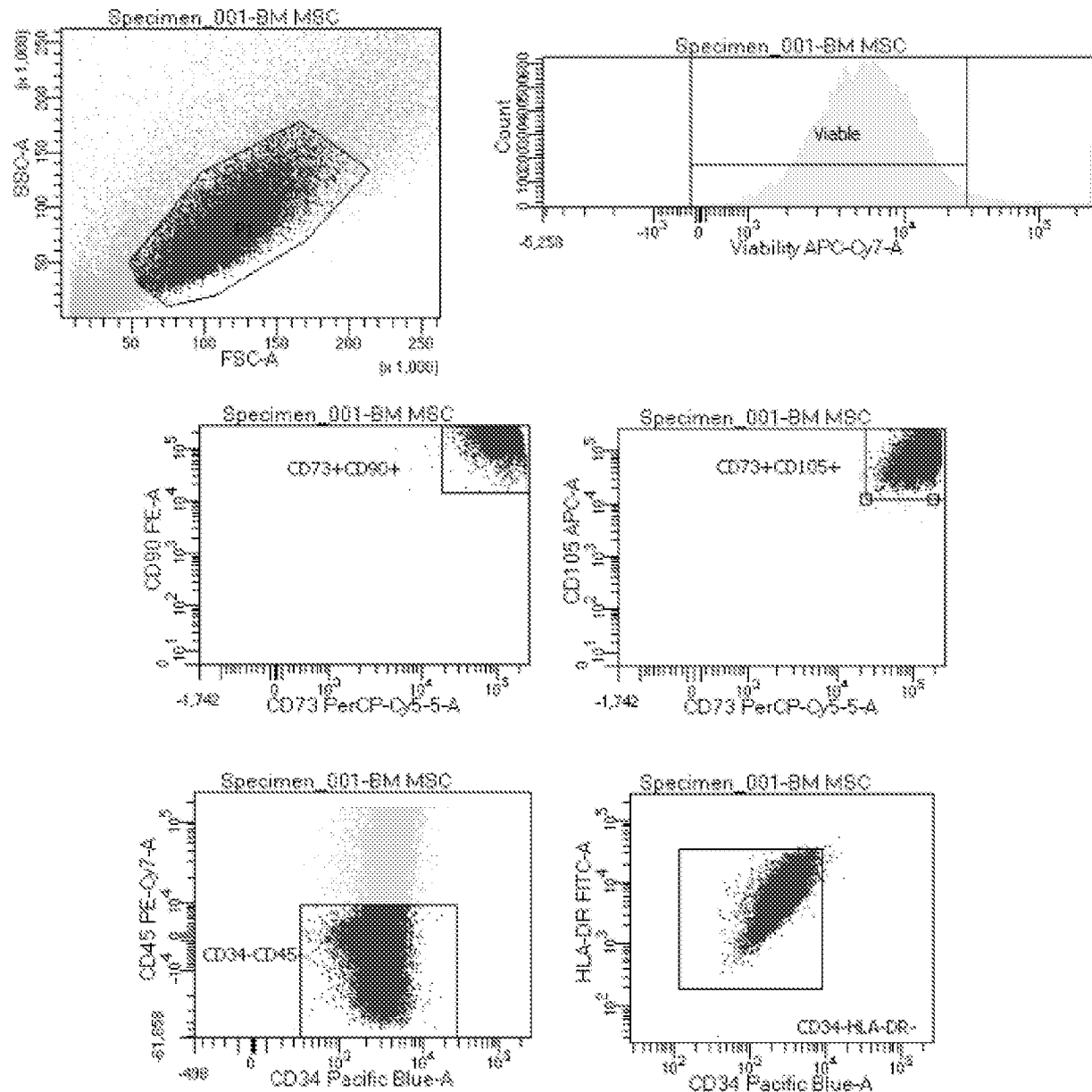
Fig. 7a



Tube: CR-001			
Population	#Events	%Parent	%Total
All Events	540,988	###	100.0
P1	425,093	78.6	78.6
Viable	414,643	97.5	76.6
CD73+CD90+	414,554	100.0	76.6
CD73+CD105+	414,480	100.0	76.6
CD34-CD45-	411,206	99.2	76.0
CD34+HLA-DR-	411,061	100.0	76.0

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Fig. 7b



Experiment Name:	MSC 082216
Specimen Name:	Specimen_001
Tube Name:	BM MSC

Population	#Events	%Parent
CD34-HLA-DR-	21,128	99.9

Population	#Events	%Parent	%Total
All Events	77,022	###	100.0
P1	35,780	46.5	46.5
Viable	33,741	94.3	43.8
CD73+CD90+	33,729	100.0	43.8
CD73+CD105+	33,710	99.9	43.8
CD34-CD45-	21,155	62.8	27.5
CD34-HLA-DR	21,128	99.9	27.4