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METHOD OF DETECTION OF INFLUENZA VIRUS AND COMPOUNDS FOR USE THEREIN

This invention relates to diagnostic methods, and in particular to methods for diagnosis of influenza virus infection. The invention provides a novel diagnostic method which can be used for detection of all strains of influenza virus Type A and Type B. The method is based on the use of compounds able to bind specifically to the active site of influenza virus neuraminidase, or functionalised derivatives of such compounds, as binding and/or detecting agents to identify influenza virus in clinical specimens. The term "neuraminidase binders" is used hereinafter to refer to these compounds and their functionalized derivatives. The method and compounds of the invention can function either in the presence or the absence of compounds binding non-specifically to influenza virus neuraminidase.

The invention particularly relates to a new class of chemical compounds and their use as diagnostic agents for the visual or instrumental detection of influenza A and B. More specifically the invention concerns a group of novel neuraminic acid (sialic acid) derivatives which bind to influenza A and B neuraminidase, and which also have a functionality which allows the compounds to be bound to a surface or to a detectable label.

Background of the Invention

Influenza A and B viruses are major causes of acute respiratory disease, with an estimated 30-50 million infections annually in the United States alone. Influenza A has been responsible for major epidemics, such as the

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"Spanish Flu" of 1919 which killed millions of people.

Many viral and bacterial infections may present with symptoms similar to those of influenza. The rapid identification of respiratory viruses would enable
5 physicians to use the most appropriate therapy early in the illness. For example, an early and accurate diagnosis would allow decisions regarding the use of antibacterial therapy and hospitalisation of children and the elderly.

Laboratory tests for the identification of
10 viruses in clinical material are widely used, and a variety of different detection methodology is available. The textbook, "Laboratory Diagnosis of Viral Infections", Marcel Dekker 1992, Ed E.H. Lennette generally discusses methods which are used for a wide range of viruses,
15 including influenza virus.

A number of tests are available for the diagnosis of influenza A and B. The traditional method of identifying influenza viruses has been the use of cell culture, which is highly sensitive and specific.
20 Unfortunately, the time required for culture, isolation and identification of influenza virus can range between 2 and 10 days, thus making it virtually useless in guiding the physician to an appropriate therapy. Since influenza virus infection is normally self-limited, diagnosis must be rapid
25 if therapy is to be effective. In other words, such cell culture methods are normally only of value in providing retrospective epidemiological information.

In addition to the cell culture methods for detecting influenza, there have recently become available a
30 few rapid direct tests, which are specific for influenza A. Thus, a monoclonal immunofluorescence assay (IFA) has been

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reported (Spada, B. et al, J. Virol. Methods, 1991 33 305) and at least one enzyme immunoassay (EIA) is available (Ryan-Poirier, K.A. et al, J. Clin. Microbiol., 1992 30 1072). A number of comparisons of these rapid detection
5 methods for influenza A have been reported; see for example Leonardi, G.P. et al, J. Clin. Microbiol., 1994 32 70, who recommended that direct specimen testing be used together with culture isolation, so as to permit both identification of the virus, in time to institute therapy and infection
10 control measures, and to monitor the antigenic constitution of influenza strains prevalent in the community for epidemiological purposes. The IFA method is reported to be labor-intensive, and requires considerable technical expertise, with the results often being difficult to
15 interpret. On the other hand, the EIA method (Directigen FLU-A; Becton Dickinson Microbiology Systems) gave a high level of false-positive results, and it has been recommended that this assay should be used in laboratories only in addition to or as a substitute for direct
20 immunofluorescence tests (Waner, J.L. et al, J. Clin. Microbiol., 1991 29 479).

As well as the problems mentioned above with the available rapid assays for influenza, there are other fundamental deficiencies in some of these methods.
25 Firstly, none of the available assays can detect influenza B, which means that even a negative test result would leave the physician uncertain about the type of therapy that should be used. Secondly, if a rapid immunoassay method depends on the use of antibodies to one
30 of the influenza A proteins, there may be a serious problem in detecting new strains of the virus which have undergone

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a drift or shift in the structure of the antigenic proteins. Influenza A is notorious for its propensity to undergo such changes.

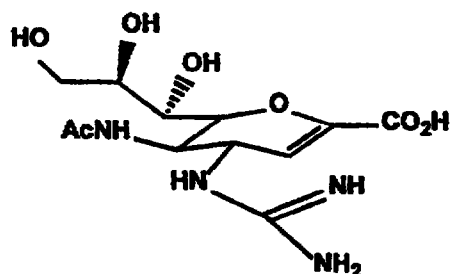
Another type of rapid assay for influenza viruses has been described in a series of patent specifications (see for example Liav, A. et al, International Patent Application No. WO 92/12256). The method involves the use of a chromogenic substrate for the influenza neuraminidase enzyme. In other words the assay depends on visualising a dye, which is formed when the influenza neuraminidase cleaves a special sialic acid-dye conjugate molecule. This technique appears to offer limited specificity, because it could not readily distinguish between the presence of viral neuraminidase and other forms of the enzyme, particularly bacterial neuraminidase. It may also have low sensitivity because of the relatively slow activity of viral neuraminidase.

Neuraminidase is one of the key proteins present on the surface of the influenza virus, and it plays an important role in the ability of the virus to infect human cells. It has long been thought that agents which bind to the neuraminidase enzyme might prevent infection by influenza, and much effort has gone into seeking such binders. Whilst many compounds have shown in vitro activity against influenza neuraminidase, only recently has it been established that it is possible to achieve protection from influenza infection in vivo by the use of a powerful neuraminidase binder which binds to the active site of the enzyme (see von Itzstein, M. et al, Nature 1993 363 418 and our earlier patent applications, International Patent Applications No. WO 92/06691 and No. WO 91/16320,

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the entire disclosures of which are herein incorporated by this reference). In particular it has been found that 2,3-didehydro-2,4-dideoxy-4-guanidinyl-N-acetylneuraminic acid (Compound I, designated GG167) is a potent binder of influenza neuraminidase, and also shows potent *in vivo* antiviral activity in animals (Ryan, D.M. et al, Antimicrobial Agents and Chemotherapy, 1994 38 2270) and in human volunteers (Hayden, F.G. et al, J. American Medical Assoc., 1996 275 295).

Compound (I) GG167



More recently it has been found that certain substituted cyclohexene derivatives of sialic acid are also potent binders of influenza virus neuraminidase (Kim, C.U. et al, J. Amer. Chem. Soc., 1997 119 681), and specifically the compound (3R,4R,5S)-4-acetamido-5-amino-3-(1-ethylpropoxy)-1-cyclohexene-1-carboxylic acid (GS 4071)

It is the purpose of the present invention to overcome some of the problems inherent in the prior art methods and to provide a simple and sensitive means of detecting influenza viruses.

Accordingly, we have now found that biologically active substances which possess stereochemical complementarity to the active site of influenza virus

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neuraminidase, as disclosed in the publication by von Itzstein et al referred to above, such that the IC_{50} for binding is 10^{-6} M or less, and in particular certain derivatives of Compound I can be used to detect influenza virus through their ability both to selectively bind the influenza virus and at the same time to be attached to a surface or to a detectable linking group.

Summary of the Invention

The invention therefore provides in a first aspect a method of detection of influenza virus, comprising the step of exposing a sample suspected to comprise said virus to a compound (neuraminidase binder) able to bind specifically to the active site of influenza virus neuraminidase.

The method of the invention is applicable to all known strains of influenza A and influenza B.

Preferably the neuraminidase binder is linked via a spacer group to a surface or to a detectable label. The spacer group should be sufficiently long that the detectable label is exposed on the surface of the virus particle.

The method of the invention may use selective capture and concentration of the virus, followed by detection of the virus using any convenient conventional method; the detection method need not have inherent selectivity. For example, the neuraminidase binder may be attached to a support material, such as a membrane or polymer, such that virus particles will be selectively captured and concentrated when a sample is passed over or through the support. Therefore in one preferred embodiment

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of the invention, the spacer group terminates in a functionality able to bind to a surface. Many suitable functionalities are known in the art. For example the terminal functionality may be a biotinyl group which can be used to attach the binder to a surface coated with avidin, streptavidin, or an antibody directed against biotin.

Alternatively the terminal functionality may for example be an amino group which can be used to conjugate the neuraminidase binder to a carboxy-comprising surface.

Alternatively, a selective detection approach may be used; the virus particles in a sample may for example be exposed to a neuraminidase binder coupled to a detectable label, under conditions such that the binder binds selectively to the viral neuraminidase on the surface of the viral particle. Preferably the detectable label is covalently coupled to the neuraminidase binder. The detectable label is then detected using any convenient method. For some detection systems, it is convenient to focus the sample into a confined area, for example a spot or a line on a surface. This may be achieved by a variety of methods; for example, the sample may be suspended or non-selectively captured on to a filter or other support material, and then exposed to the labelled neuraminidase binder as described above. There are several known methods for the non-selective capture of influenza virus, for example on a fetuin-coated surface (J. Virological Methods, 1992 39 111) or on a suitable membrane (J. Virological Methods, 1992 40 77). Another suitable detection system could be an optical assay device as described by Miller, B.J. et al, United States Patent Application No. 5,418,136.

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In another, alternative, aspect the invention may use a combination of selective capture and selective detection to provide a simple and sensitive two-stage method of detecting influenza virus. This makes use of the fact that each influenza virus particle typically has about one hundred neuraminidase molecules spread over its spherical surface (White, D.O., Curr. Top. Microbiol. Immunol., 1974 63 1-48), and can therefore attach to more than one neuraminidase binder molecule at the same time.

Thus, a neuraminidase binder compound may be attached to a support, for example as a narrow band across a length of porous membrane. The test sample is then applied at the other end of the membrane and allowed to flow across the band of bound compound. Any influenza virus particles in the test sample will be trapped by the membrane-bound compound and thus retained in the narrow band. In the second stage of the test a detectable label attached to the neuraminidase binder is allowed to flow through the membrane across the band of bound influenza virus particles. The presence of influenza virus is then shown by an observable change in the membrane at the site of the bound compound. Suitable chromatographic assay devices have been described, for example by Chandler, H.M., International Patent Application No. WO 92/21977.

A very large number of suitable detection systems is known in the art, for example biotin-streptavidin, enzymic systems such as horseradish peroxidase or alkaline phosphatase, fluorescence systems, chemiluminescence systems, colloidal gold, radioactive labels and agglutination systems. It is contemplated that colloidal gold linked to a neuraminidase binder via a spacer group

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will be a particularly convenient detectable label.

Another convenient detection system would utilize the neuraminidase binder covalently coupled to horseradish peroxidase. The skilled person will readily be able to
5 select a suitable detection system and to optimise conditions for detection, using normal trial and error experimentation.

The neuraminidase binder may be any agent which binds to the active site of the influenza virus
10 neuraminidase, provided that it does not comprise a detectable label or spacer group which is cleaved by the neuraminidase. The binding need not be irreversible. The IC_{50} for binding is suitably 10^{-6} M or better.

Suitable agents include, but are not limited to,
15 compounds of suitable IC_{50} disclosed in our earlier patent applications, International Patent Applications No. WO/92/06691 and WO/91/16320, and also U.S. Patent No. 5,453,533 by Luo et al, U.S. Patent No. 5,512,596 by Gilead Sciences, Inc., and International Patent Application No.
20 WO/96/26933 by Gilead Sciences, Inc. Although these specifications disclose the use of the compounds disclosed therein for treatment or prophylaxis of influenza virus infection, it is not suggested that the compounds are useful for diagnostic purposes, using assays of the type
25 described herein.

According to a second aspect, the invention provides a neuraminidase binder suitable for use in the method of the invention, comprising an analogue of neuraminic acid (sialic acid) substituted at the C₇ or
30 equivalent position with a spacer group suitable for coupling to a detectable label or to a surface.

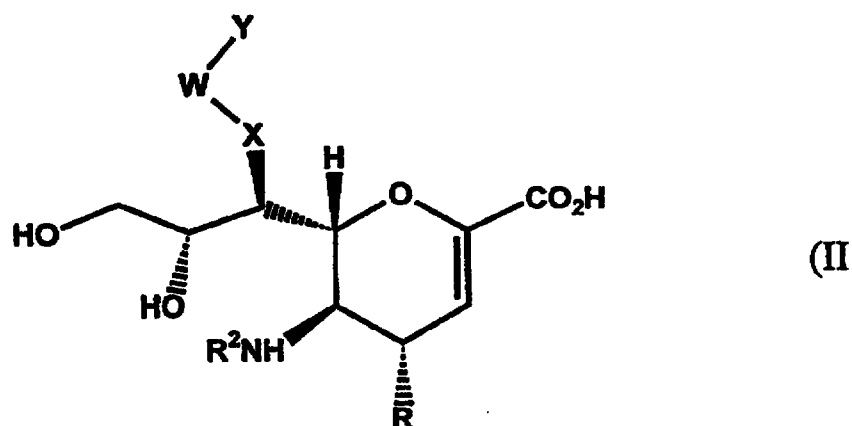
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In a preferred embodiment, this aspect provides compounds of formula (II):



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wherein

R represents an azido group, an unsubstituted or substituted guanidino group, or an unsubstituted or substituted amino group;

10

R² represents COCH₃, COCF₃, or SO₂CH₃;

X represents O or NH;

W represents a spacer group made up of a chain of atoms having a total length of between four and one hundred atoms, and optionally also comprising substituted carbon and/or nitrogen atoms and optionally including oxygen and/or sulphur atoms;

15

Y represents OH, SH, NH₂, CH=O, CH=CH₂, CO₂H, CONHNH₂, or NH-biotinyl, or a protected form of one of these end functionalities;

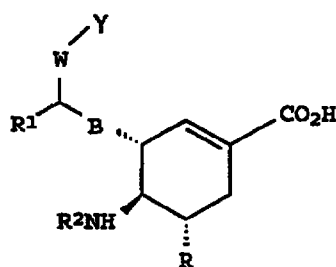
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and wherein the substituent on the amino or guanidino group, when present, is a substituted or unsubstituted C₁₋₆ alkyl, such as a methyl or ethyl group or an aryl group such as phenyl; or an amino, hydroxy, cyano or alkoxycarbonyl group;

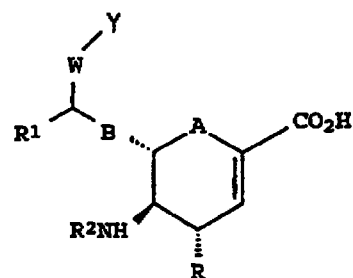
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with the proviso that X-W-Y does not include the group $OC(=Z)NR^5R^6$, wherein Z represents O or S, and R^5 and R^6 independently represents H or a hydrocarbon group optionally substituted by NH_2 , OH or SH.

5 In another preferred embodiment, the invention provides neuraminidase binders of formula (IIA) or (IIB)



(IIA)



(IIB)

10 wherein R, R^2 , W and Y are as defined above for formula (II), A and B are oxygen or CH_2 or may represent a single bond, and R^1 is a lipophilic C_1 - C_{12} alkyl group which is optionally substituted by one or more halogen atoms, cycloalkyl, alkoxy, haloalkoxy or optionally substituted
15 aryl groups.

Suitable spacer groups W include, but are not limited to, linear peptides, oligosaccharides, polyols, polyethylene glycol groups, hydrocarbon groups and hydrocarbon groups linked together with oxygen or sulphur
20 atoms or with carbonyl, amido, urea or hydrazide functionalities. Spacer groups W may also comprise combinations of these various groups.

Suitable protecting groups for the end functionality Y include, but are not limited to, esters of



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the OH, SH and CO₂H groups, carbamates of the NH₂ and CONHNH₂ groups, and acetals of the CH=O group.

As used herein, the term "hydrocarbon group" includes saturated and unsaturated straight or branched hydrocarbon groups, including aryl groups, and combinations of such groups.

Preferred compounds of the invention include compounds of formula (II) wherein R represents guanidino and R² represents acetyl.

Another group of preferred compounds of the invention of formula (II) are those wherein X is O and Y is NH₂ or NH-biotinyl.

A third preferred group of compounds of the invention includes neuraminidase binders of formula (IIA) in which R is amino, R² is acetyl, B is oxygen and R¹ is an alkyl group.

Further preferred compounds are those of formula (II) wherein W is a spacer group selected from the group consisting of CONH(CH₂)_n, CONH(CH₂)_nNHCONH(CH₂)_m, CONH(CH₂)_n[NHCO(CH₂)₅]_m, CO(CH₂)_n, CO(CH₂)_nNHCONH(CH₂)_m, CO(CH₂)_nNHCO(CH₂)_q, CONH(CH₂)_nNH(COCH₂NH)_qCOCH₂, CONH(CH₂CH₂O)_nCH₂CH₂NHCO(CH₂)_q, and CONH(CH₂)_nNHCO(CH₂)_q, wherein n and m are integers between 2 and 12, and q is 0 or an integer between 1 and 12.

The person skilled in the art will be able to select a suitable detectable label and detection system, and to test the compounds referred to above for their suitability using routine methods. Thus the skilled person would be able to choose a suitable detection system, such as selective capture or selective detection as described above. For selective capture systems, the spacer group

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will terminate in a functionality able to bind to a solid support. Many suitable functionalities are known in the art.

The substitution by the group W-Y at C₇ (or
5 equivalent) should not decrease the ability of the parent compound to bind to neuraminidase by a significant extent; preferably the binding is not decreased more than 100-fold. The ability to bind to neuraminidase may be assessed using any convenient method; one suitable method is the *in vitro*
10 bioassay of neuraminidase activity described by Warner and O'Brien, *Biochemistry*, 1979 18 2783-2787. *In vitro* bioassay is described in our Application No. WO 91/16320. Alternatively binding of a radioactively-labelled binder compound to neuraminidase can be assessed by methods known
15 in the art.

Suitable types of clinical samples for use in the method of the invention include throat swabs, nasal swabs, naso-pharyngeal washings, nasal washes and gargles, or combinations of any of these; the gargles may optionally be
20 concentrated, for example by ultracentrifugation, if necessary, but it is expected that this will only be the case when the number of viral particles is low.

It will be clearly understood that the label which is linked to the spacer group may be an epitope
25 suitable for use in antibody detection kits, such as those described in U.S. Patent No. 4943522 by Eisinger et al (assigned to Quidel); in optical assay devices having an active receptive surface, such as those described in U.S. Patent No. 5,418,136 by Miller et al (assigned to Biostar,
30 Inc.); or in agglutination detection systems such as those described in U.S. Patent No. 4,894,347 and International

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Patent Application No WO 91/04492, both by Agen Limited.

It is also contemplated that biosensor systems are suitable for use in the method of the invention. For example, a carbohydrate biosensor surface for detection of bacteria

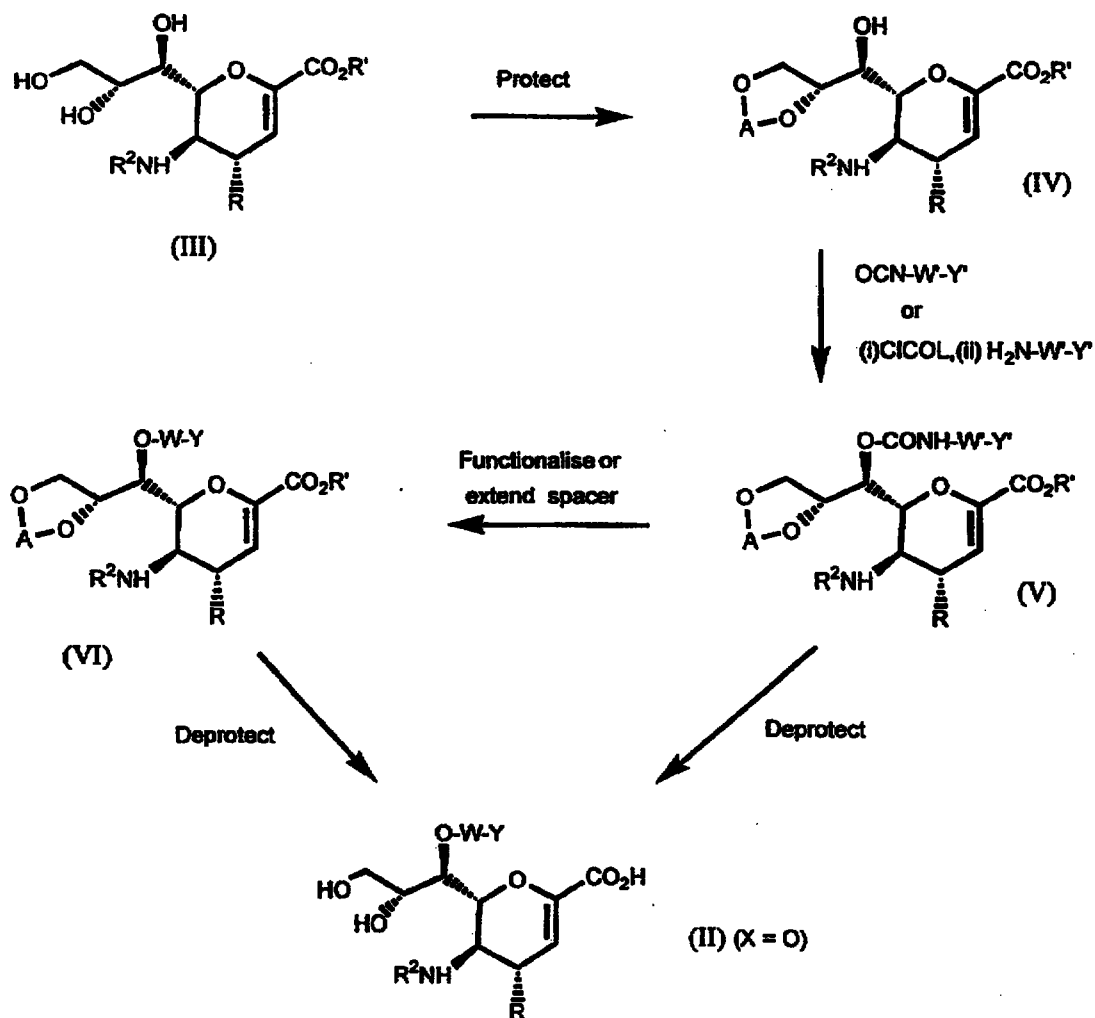
5 has been described (Nilsson, K.G.I. and C.F. Mandenius; BioTechnology, 1994 12 1376-1378).

Some compounds according to the invention bind particularly strongly to influenza virus neuraminidase, with IC_{50} of 10^{-8} M or better, and these compounds may also
10 be therapeutically useful.

The compounds of the invention may be prepared by a variety of methods, and in a further aspect of the invention provides methods for the preparation of compounds of formula (II). In one type of approach the preparation
15 of the compounds of the invention wherein X is O may be carried out as shown in the reaction schemes below in which R, R² and Y are as defined for formula (II), R' is an alkyl group and W' is a spacer group. Thus, a sialic acid derivative of formula (III) is converted into the 8,9-
20 protected form of formula (IV), which may be for example a cyclic carbonate (A = C=O) or an acetonide (A = CMe₂). The remaining free 7-hydroxy group is acylated with either an isocyanate group OCN-W'-Y or an acyl chloride (ClCOL), which has a second leaving group L, the group L then being
25 displaced by an amine H₂N-W'-Y. The group W' is a spacer group, generally of 4 to 10 atoms length, generally within the definition of W, and W may be equivalent to CONHW'. The spacer group W' can be further extended or functionalised, for example by the addition of biotin to
30 the end of the chain as in (VI).

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Finally, the protecting groups A, R' and any that are present on the substituents R and Y, can be removed under standard conditions to give the compounds of formula (II). A variety of protecting groups suitable for protection of the hydroxy and carbonyl groups is known in the art; see for example "Protecting Groups in Organic Synthesis" by T.H. Greene (Wiley & Sons, 1981).



10

Compounds of formulae (III) and (IV) are known, and their preparation has been described, for example, in International Patent Application No. WO 91/16320.

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Another type of approach for the preparation of compounds of the invention wherein X is NH involves starting with a sialic acid derivative and carrying out reactions which give a double inversion at the C7 position.

5 For example, an intermediate of general formula (IV) may be reacted under Mitsunobu Reaction conditions to give a suitable leaving group ester with inverted stereochemistry at C7 (Anderson, N.G. et al, J. Org. Chem., 1996 61 7955). Displacement of the C7 ester with a nucleophile such as
10 azide then gives a C7 nitrogen substituted derivative with the correct stereochemistry. An alternative method for the introduction of nitrogen to the C7 position of sialic acid derivatives has been described (Kong and von Itzstein, Tetrahedron Letters, 1995 36 957).

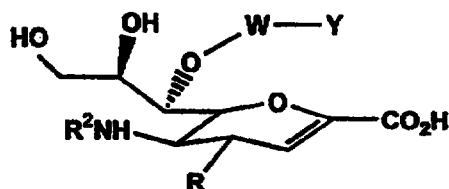
15 Most of the intermediates of formula (V) and (VI) are novel compounds, and these comprise a further aspect of the present invention.

Detailed Description of the Invention

20 The invention will now be described in detail by way of reference only to the following non-limiting examples.

Examples of compounds of the invention of formula (II) (X=O) are shown in Table 1. Those examples of
25 substituent Y which include biotin have the biotin attached via an amide linkage to its carboxy group.

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Table 1



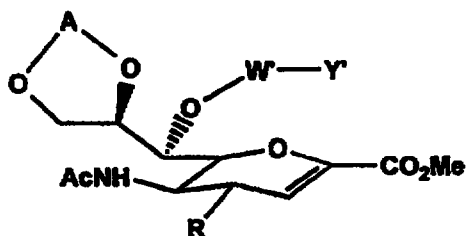
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Compound No.	R	R ²	W	Y
8	NHC(=NH)NH ₂	Ac	CONH(CH ₂) ₆ NHCONH(CH ₂) ₆	NH ₂
11	NHC(=NH)NH ₂	Ac	CONH(CH ₂) ₆	NH ₂
13	NHC(=NH)NH ₂	Ac	CONH(CH ₂) ₆	NHBoc
17	NHC(=NH)NH ₂	Ac	CONH(CH ₂ CH ₂ O) ₂ CH ₂ CH ₂	NH ₂
19	NHC(=NH)NH ₂	Ac	CONH(CH ₂) ₆	NH-Biotin
20	NH ₂	Ac	CONH(CH ₂) ₆ NHCONH(CH ₂) ₆	NHBoc
21	NHC(=NH)NH ₂	Ac	CONH(CH ₂) ₆ (NHCOCH ₂) ₃	NH-Biotin
22	NHC(=NH)NH ₂	Ac	CONH(CH ₂) ₆ NHCO(CH ₂) ₁₁	NH-Biotin
23	NHC(=NH)NH ₂	Ac	CONH(CH ₂) ₆ NHCO(CH ₂) ₅	NH-Biotin
25	NHC(=NH)NH ₂	Ac	CONH(CH ₂) ₆ [NHCO(CH ₂) ₅] ₄	NH ₂
26	NHC(=NH)NH ₂	Ac	CONH(CH ₂) ₆ [NHCO(CH ₂) ₅] ₄	NH-Biotin
27	NHC(=NH)NH ₂	Ac	CONH(CH ₂) ₆ NHCO(CH ₂) ₅ NHCO CH ₂ (OCH ₂ CH ₂) ₁₆	NH-Biotin
28	NHC(=NH)NH ₂	Ac	CONH(CH ₂) ₆ [NHCO(CH ₂) ₅] ₂	NH ₂
29	NHC(=NH)NH ₂	Ac	CONH(CH ₂) ₆ [NHCO(CH ₂) ₅] ₄ N H-COCH ₂ CH ₂	CONHNHBoc
30	NHC(=NH)NH ₂	Ac	CONH(CH ₂) ₆ [NHCO(CH ₂) ₅] ₄ N H-COCH ₂ CH ₂	CONHNH ₂
31	NHC(=NH)NH ₂	Ac	CONH(CH ₂) ₆ NHCOCH ₂ CH ₂	CO ₂ H

and examples of intermediates of formula (V) and (VI) are given in Table 2. Again Boc represents butoxycarbonyl.

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Table 2



Compound No.	A	R	W'	Y'
3	C=O	N ₃	CONH(CH ₂) ₆	NHBoc
4	C=O	N ₃	CONH(CH ₂) ₆	NH ₂
5	C=O	N ₃	CONH(CH ₂) ₆ NHCONH(CH ₂) ₆	NHBoc
6	C=O	NH ₂	CONH(CH ₂) ₆ NHCONH(CH ₂) ₆	NHBoc
7	C=O	NHC(=NH)NH ₂	CONH(CH ₂) ₆ NHCONH(CH ₂) ₆	NHBoc
9	C=O	NH ₂	CONH(CH ₂) ₆	NHBoc
10	C=O	NHC(=NBoc)NHBoc	CONH(CH ₂) ₆	NHBoc
12	C=O	NHC(=NH)NH ₂	CONH(CH ₂) ₆	NHBoc
14	C=O	N ₃	CONH(CH ₂ CH ₂ O) ₂ CH ₂ CH ₂	NHBoc
15	C=O	NH ₂	CONH(CH ₂ CH ₂ O) ₂ CH ₂ CH ₂	NHBoc
16	C=O	NHC(=NBoc)NHBoc	CONH(CH ₂ CH ₂ O) ₂ CH ₂ CH ₂	NHBoc
18	C=O	NHC(=NH)NH ₂	CONH(CH ₂) ₆	NH ₂
24	C=O	NHC(=NH)NH ₂	CONH(CH ₂) ₆ [NHCO(CH ₂) ₃] ₄	NHBoc
32	C=O	NHC(=NH)NH ₂	CONH(CH ₂) ₆ NHCOCH ₂ CH ₂	CO ₂ H

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Example 1 **Preparation of 5-acetamido-7-[6'-(6"-
aminohexylureido)-hexyl 1-carbamoyloxy-4-
guanidino-2,3,4,5-tetradecoxy-D-glycero-D-
galacto-non-2-enopyranosonic acid (8)**

- 5 (A) Preparation of methyl 5-acetamido-4-azido-8,9-
monocarbonyldioxy-2,3,4,5-tetradecoxy-D-glycero-D-galacto-
non-2-enopyranosonate (2).

To a suspension of methyl 5-acetamido-4-azido-
2,3,4,5-tetradecoxy-D-glycero-D-galacto-non-2-
10 enopyranosonate (1) (1.32 g, 4.03 mmol) in dry
dichloromethane (32 ml) and acetonitrile (16 ml) containing
N,N-dimethylaniline (1.6 ml, 12.46 mmol) in an ice-bath
under argon, was added dropwise a 20% solution of phosgene
in toluene (3 ml, 6.06 mmol) over a period of 20 min. The
15 clear reaction mixture was stirred at ice-bath temperature
for 2 hr, then left at room temperature for 16 hr before
being evaporated to dryness. The residue was partitioned
between ethyl acetate (300 ml) and water (50 ml). The
organic layer was washed successively with 0.5 M HCl
20 solution (32 ml) at 5-10°C, saturated sodium chloride
solution (3x30 ml), dried over anhydrous Na₂SO₄, then
evaporated to dryness. The residue was subjected to flash
column chromatography (silica gel, ethyl
acetate/hexane=8/1) to afford compound (2) (1.035 g, 71%)
25 as a white foam.

MS (FAB): 357 (M+1)⁺

i.r. (CHCl₃) cm⁻¹: 3314, 2101, 1774, 1734, 1654.

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¹H-nmr (CDDl₃) δ (ppm): 2.13 (s, 3H), 3.81 (s, 3H), 3.82 (d, 1H) 3.90 (d, 1H), 4.08 (dd, 1H), 4.21 (dd, 1H), 4.72 (d, 2H), 4.97 (ddd, 1H), 5.95 (d, 1H), 6.55 (d, 1H).

5 (B) Preparation of methyl 5-acetamido-4-azido-7-(6'-tert-butoxycarbonylaminohexyl)-carbamoyloxy-8,9-monocarbonyldioxy-2,3,4,5-tetradecoxy-D-glycero-D-galactonon-2-enopyranosonate (3)

To a solution of compound (2) (577 mg, 1.62 mmol) in anhydrous pyridine (8 ml) under argon, was added 4-dimethylaminopyridine (DMAP) (275 mg, 2.25 mmol) and 4-nitrophenyl chloroformate (424 mg, 2.11 mmol). The reaction mixture was stirred at 30-35°C for 3.5 hr. To this mixture was added 6-tert-butoxycarbonylamino-
15 hexylamine (567 mg, 2.63 mmol) and DMAP (405 mg, 3.32 mmol). The mixture was stirred at room temperature under argon for 24 hr then diluted with ethyl acetate (400 ml). The ethyl acetate solution was washed successively with water (3x40 ml), 2M HCl (40 ml) at 5-10°C and then water
20 (3x20 ml). The organic layer was evaporated to dryness to give the crude product (1.103 g) which was then flash chromatographed (silica gel, firstly, ethyl acetate/hexane = 2/1, secondly ethyl acetate) to afford the title compound (3) (726 mg, 75%) as a white foam.

25

MS (FAB): 599(M+1)⁺, 607(m+9)⁺, (M+1+2H₂O-N₂)

i.r. (CHCl₃) cm⁻¹: 3328, 2734, 2099, 1799, 1732, 1682.

30 ¹H-nmr (CDCl₃) δ (ppm): 1.20-1.40 (m, 4H), 1.40-1.60 (m, 13H), 2.07 (s, 3H), 3.01-3.42 (m, 5H), 3.84 (s, 3H), 4.50-

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4.85 (m, 4H), 4.93 (m, 2H), 5.03 (m, 1H), 5.41 (m, 1H),
5.95 (d, 1H), 6.48 (d, 1H).

(C) Preparation of methyl 5-acetamido-7-[6'-(6"-
5 tert-butoxycarbonylaminohexylureido)-hexyl]-carbamoyloxy-4-
azido-8,9-monocarbonyldioxy-2,3,4,5-tetradecoxy-D-glycero-D-
galacto-non-2-enopyranosonate (5).

A solution of compound (3) (200 mg, 0.334 mmol)
in trifluoroacetic acid (2 ml) was stirred under argon at
10 room temperature for 1 hr, then evaporated under reduced
pressure to dryness. The residue was taken up in methanol
(5 ml) and evaporated to dryness again to give compound (4)
as a white foam.

15 ¹H-nmr (CDCl₃) δ (ppm): 1.25-1.70 (m, 8H), 2.08 (s, 3H),
3.0-3.5 (m, 4H), 3.81 (s, 3H), 3.88 (dd, 1H), 4.4-4.8 (m,
4H), 5.01 (ddd, 1H), 5.30 (dd, 1H), 5.51 (dd, 1H), 5.86 (d,
1H), 6.73 (d, 1H), 7.34 (d, 1H).

20 Compound (4) was then dissolved in pyridine
(2 ml) containing a mixture of DMAP (30 mg, 0.246 mmol) and
6-tert-butoxycarbonylaminoethyl isocyanate (0.15 ml,
0.434 mmol). The reaction mixture was stirred under argon
at 18-20°C for 72 hr, then stirred with methanol (0.2 ml)
25 at room temperature for 3 hr. The resulting reaction
mixture was evaporated to dryness and partitioned between
ethyl acetate (200 ml) and 1M NaH₂PO₄ solution (50 ml).
The organic layer was washed successively with 1M NaH₂PO₄
(2x50 ml), water (3x25 ml) and vacuum evaporated to
30 dryness. The residue was flash-chromatographed (silica
gel, using ethyl acetate/hexane = 8/1, then ethyl

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acetate/methanol = 10/1) to afford the title compound (5) (180 mg, 72%) as a white foam.

MS (FAB): 741(m+1)⁺, 749(m+9)⁺

5

i.r. (CHCl₃) cm⁻¹: 3356, 2934, 2098, 1801, 1732, 1654.

¹H-nmr (CDCl₃+CD₃OD) δ(ppm): 1.12-1.42 (m, 25H), 1.80 (s, 3H), 2.90-3.18 (m, 9H), 3.70 (s, 3H), 3.90 (dd, 1H), 4.18 (dd, 1H), 4.32 (dd, 1H), 4.55 (d, 2H), 4.91 (m, 1H), 5.21 (br, 1H), 5.35 (dd, 1H), 5.45 (br d, 1H), 5.82 (d, 1H), 6.05 (dd, 1H), 7.82 (d, 1H).

(D) Preparation of methyl 5-acetamido-4-amino-7-[6'-(6"-tert-butoxycarbonyl aminohexylureido)-hexyl]-carbamoyloxy-8,9-monocarbonyldioxy-2,3,4,5-tetradecoxy-D-glycero-D-galacto-non-2-enopyranosonate (6).

A mixture of compound (5) (180 mg, 0.246 mmol) acetic acid (35 mg, 0.58 mmol) and Pd/C (10%) (30 mg) was agitated in a mixture of methanol (10 ml) and toluene (6 ml) under hydrogen at room temperature for 1 hr. The reaction mixture was filtered through Celite, the filter-cake was washed with methanol (30 ml) and the filtrate and washings were combined and vacuum evaporated to dryness. The residue was subjected to flash column chromatography (silica gel, ethyl acetate/2-propanol/water = 5/2/1) to afford compound (6) (46 mg, 47.5%) as a white foam, as well as recovered starting material (80 mg, 44%).

¹H-nmr (CD₃OD) δ(ppm): 1.15-1.55 (m, 25H), 1.87 (s, 3H, AcOH), 1.92 (s, 3H), 2.90-3.10 (m, 8H), 3.56 (dd, 1H), 3.69

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(s, 3H), 3.91 (dd, 1H), 4.20 (dd, 1H), 4.62 (d, 2H), 5.05 (dd, 1H), 5.46 (dd, 1H), 5.85 (dd, 1H).

(E) Preparation of methyl 5-acetamido-7-[6'-(6"-
5 tert-butoxycarbonylaminohexylureido) -hexyl]-carbamoyloxy-
4-[2,3-bis(tert-butoxycarbonyl]-guanidino-8,9-monocarbonyl-
dioxo-2,3,4,5-tetradecoxy-D-glyco-D-galacto-non-2-
enopyranosonate(7).

A solution of compound (6) (46 mg, 0.065 mmol) in
10 methanol (10 ml) was treated with Amberlite IRA-400 (OH)
resin (100 mg) for 30 seconds and filtered off. The resin
was washed with methanol (20 ml). The filtrate and washing
were combined, evaporated to dryness to give 41mg solid
substance which was then mixed with N,N'-bis-tert-
15 butoxycarbonyl-1H-pyrazole-1-carboxamidine (103 mg,
0.332 mmol) in methanol (5 ml). The mixture was stirred
under argon at 30-35°C for 5 days, then evaporated to
dryness. The residue was flash chromatographed (silica gel,
first ethyl acetate/hexane = 8/1, then ethyl acetate) to
20 give compound (7) (47 mg, 88.8 %) as a white foam.

i.r. (CHCl₃) cm⁻¹: 3312, 2933, 1802, 1728, 1643, 1564.

¹H-nmr (CDCl₃) δ(ppm): 1.20-1.60 (m, 43H), 1.87 (s, 3H),
25 2.19 (s, 1H), 2.90-3.20 (m, 8H), 3.73 (s, 3H), 4.17 (dd,
1H), 4.33 (br d, 1H), 4.53-5.20 (m, 8H), 5.51 (br, 1H), 5.88
(d, 1H), 6.66 (br, 1H), 8.38 (d, 1H).

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(F) Preparation of 5-acetamido-7-[6'-(6"-aminohexylureido)-hexyl]-carbamoyloxy-4-guanidino-2,3,4,5-tetradecoxy-D-glycero-D-galacto-non-2-enopyranosonic acid (8).

5 A solution of compound (7) (47 mg, 0.058 mmol) in a mixture of methanol (5 ml) and water (5 ml) containing triethylamine (0.8 ml) was stirred under argon at room temperature for 6 hr, then evaporated under reduced pressure to dryness. The residue was dissolved in a
10 mixture of methanol (5 ml) and water (5 ml), then stirred with Dowex 50WX8(H⁺) resin (1 g) at room temperature for 1 hr. The resin was filtered and washed with a mixture of methanol and water (1:1), then stirred with 2M NH₄OH solution (20 ml) at room temperature for 2 hr and filtered
15 off. The filtrate was evaporated to dryness. The residue was redissolved in water (5 ml) and evaporated to dryness again. The resulting solid was dissolved in water (2 ml) and freeze-dried to afford compound (8) (16 mg, 45.7%) as a white solid with a positive guanidine (Sakaguchi) reaction.

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MS (FAB): 617 (M+1)⁺

¹H-nmr (D₂O) δ(ppm): 1.10-1.17 (m, 18H), 2.07 (s, 3H), 2.85-3.20 (m, 6H), 3.50 (dd, 1H), 3.70 (dd, 1H), 4.10 (m, 2H),
25 4.45 (dd, 1H), 4.52 (dd, 1H), 4.95 (dd, 1H), 5.65 (d, 1H).

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Example 2 Preparation of 5-acetamido-7-(6'-aminohexyl)-carbamoyloxy-4-guanidino-2,3,4,5-tetradecoxy-D-glycero-D-galacto-non-2-enopyranosonic acid as trifluoroacetic acid salt (11)

(A) Preparation of methyl 5-acetamido-4-amino-7-(6'-tert-butoxycarbonylaminohexyl)-carbamoyloxy-8,9-monocarbonyldioxy-2,3,4,5-tetradecoxy-D-glycero-D-galacto-non-2-enopyranosonate (9).

Compound (3) (Example 1, part (B) (500 mg, 0.836 mmol) was hydrogenated, according to the procedure described in Example 1, part (D) to afford compound (9) (285 mg, 59.6%) as a white foam.

MS (FAB): 573 (M+1)⁺

i.r. (CHCl₃) cm⁻¹: 3385, 2935, 1794, 1725, 1670, 1542.

¹H-nmr (CD₃OD) δ (ppm): 1.15-1.48 (m, 17H), 1.91 (s, 3H), 2.96 (m, 4H), 3.51 (dd, 1H), 3.70 (s, 3H), 3.88 (dd, 1H), 4.21 (dd, 1H), 4.62 (d, 2H), 5.05 (ddd, 1H), 5.45 (dd, 1H), 5.87 (d, 1H).

(B) Preparation of methyl 5-acetamido-7-(6'-tert-butoxycarbonylaminohexyl)-carbamoyloxy-4-[2',3"-bis(tert-butoxycarbonyl)]-guanidino-8,9-monocarbonyldioxy-2,3,4,5-tetradecoxy-D-glycero-D-galacto-non-2-enopyranosonate (10).

The title compound (10) (149 mg, 83.4%) was obtained as a white foam from compound (9) (125 mg, 0.219 mmol) following the procedure described in Example 1, part (E).

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MS (FAB): 815 (M+1)⁺

i.r. (CHCl₃) cm⁻¹: 3313, 2978, 1805, 1727, 1643, 1610,
5 1558.

¹H-nmr (CDCl₃) δ (ppm): 1.11-1.82 (m, 35H), 1.91 (s, 3H),
2.98-3.21 (m, 4H), 4.13 (dd, 1H), 4.32 (dd, 1H), 4.50-4.80
(m, 3H), 4.91-5.22 (m, 3H), 5.45 (dd, 1H), 5.89 (d, 1H),
10 6.69 (br d, 1H), 7.63 (d, 1H), 8.48 (d, 1H).

(C) Preparation of 5-acetamido-7-(6'-aminohexyl)-
carbamoyloxy-4-guanidino-2,3,4,5-tetradecoxy-D-glycero-D-
galacto-non-2-enopyranosonic acid as trifluoroacetic acid
15 salt (11).

Compound (10) (54 mg, 0.066 mmol) was stirred in
a mixture of methanol (25 ml) and water (25 ml) containing
triethylamine (5 ml) under argon at room temperature for
6hr then evaporated under vacuum to dryness. The residue
20 was chromatographed (silica gel, ethyl acetate/2-
propanol/water = 5/2/1) and the fractions with R_f value of
0.4 were combined and evaporated to dryness. The residue
was stirred in trifluoroacetic acid (2 ml) under argon at
room temperature for 1hr then evaporated to dryness. The
25 residue was dissolved in water (3 ml) and freeze-dried to
afford the title compound (11) (27 mg, 69 %).

MS (FAB): 475 (M+1)⁺, 589 (M+1+TFA)⁺.

30 ¹H-nmr (D₂O) δ (ppm): 1.27-1.81 (m, 8H), 1.98 (s, 3H),
2.89-3.21 (m, 4H), 3.51 (dd, 1H), 3.69 (dd, 1H), 4.04 (m,

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1H), 4.18 (dd, 1H), 4.46 (dd, 1H), 4.57 (dd, 1H), 4.96 (dd, 1H), 6.01 (d, 1H).

Example 3

Preparation of 5-acetamido-7-(6'-tert-butoxycarbonylaminohexyl)-carbamoyloxy-4-guanidino-2,3,4,5-tetradecoxy-D-glycero-D-galacto-non-2-enopyranosonic acid (13).

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To a solution of compound (9) (24 mg, 0.042 mmol) in water (5 ml) was added 1H-pyrazole-1-carboxamidine (44 mg, 0.30 mmol) and imidazole (43 mg, 0.63 mmol) at room temperature. The resulting mixture was stirred under argon at 90-95°C for 22 hr, then evaporated to dryness to give crude compound (12) which gave a strong positive Sakaguchi reaction, indicating the presence of a guanidine group.

The residue was stirred in water (5 ml) containing triethylamine (0.5 ml) under argon at room temperature for 16 hr before evaporating to dryness. The residue was subjected to chromatography (silica gel, 2-propanol/water = 3/1). The fractions with R_f value of 0.2 were combined and evaporated to dryness, then freeze-dried to afford the title compound (13) (9 mg, 37%) as a white solid.

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MS (FAB): 575 (M+1)⁺

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¹H-nmr (D₂O) δ (ppm): 1.25-1.83 (m, 17H), 1.95 (s, 3H), 2.95-3.25 (m, 4H), 3.52 (dd, 1H), 3.63 (dd, 1H), 3.72 (dd, 1H), 3.98 (dd, 1H), 4.10 (m, 1H), 4.46 (dd, 1H), 4.52 (dd, 1H), 5.62 (d, 1H).

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Example 4 Preparation of 5-acetamido-7-{2-[2-(2-aminoethoxy)-ethoxy]-ethyl}-carbamoyloxy-4-guanidino-2,3,4,5-tetradecoxy-D-glycero-D-galacto-non-2-enopyranosonic acid (17)

- 5 (A) Preparation of methyl 5-acetamido-4-azido-7-{2'-[2''-(2'''-tert-butoxycarbonylaminoethoxy)-ethoxy]-ethyl}-carbamoyloxy-8,9-monocarbonyldioxy-2,3,4,5-tetradecoxy-D-glycero-D-galacto-non-2-enopyranosonate (14).

Following a similar procedure to that described
10 in Example I, part (B) compound (14) (420 mg, 67%) was obtained as a white foam from compound (2) (356 mg, 1 mmol) and 2-[2'-(2''-tert-butoxycarbonylamino-ethoxy)ethoxy]-ethylamine.

15 MS (FAB): 631 (M+1)⁺ 639 (M+9)⁺

i.r. (CHCl₃) cm⁻¹: 3328, 2934, 2099, 1802, 1734, 1658, 1537.

20 ¹H-nmr (CDCl₃) δ (ppm): 1.41 (s, 9H), 1.97 (s, 3H), 3.12-3.64 (m, 12H), 3.78 (s, 3H), 4.41-4.71 (m, 4H), 5.01 (d, 2H), 5.12 (m, 1H), 5.43 (dd, 1H) 5.67 (br, 1H), 5.91 (d, 1H), 6.83 (br d, 1H).

- 25 (B) Preparation of methyl 5-acetamido-4-amino-7-{2'-[2''-(2'''-tert-butoxycarbonylaminoethoxy)-ethoxy]-ethyl}carbamoyloxy-8,9-monocarbonyldioxy-2,3,4,5-tetradecoxy-D-glycero-D-galacto-non-2-enopyranosonate (15).

Catalytic-hydrogenation of compound (14) (400 mg,
30 0.634 mmol), as described in Example I, part (D) afforded compound (15) (144 mg, 37.5%) as a white foam after

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chromatography (silica gel, ethyl acetate/2-propanol/water = 5/2/1).

¹H-nmr (CD₃OD) δ(ppm): 1.32 (s, 9H), 1.82 (s, 3H), 1.89
5 (s, 3H, AcOH), 3.04-3.22 (m, 4H), 3.36-3.53 (m, 8H), 3.68
(s, 3H), 3.72 (br d, 1H), 3.98 (dd, 1H), 4.25 (dd, 1H),
4.62 (d, 2H), 5.03 (ddd, 1H), 5.43 (dd, 1H), 5.84 (d, 1H).

(C) Preparation of methyl 5-acetamido-7-{2'-[2''-(2'''-
10 tert-butoxycarbonylaminoethoxy)-ethoxy]-ethyl}-
carbamoyloxy-4-[2',3''-bis(tert-butoxycarbonyl)]-guanidino-
8,9-mono-carbonyldioxy-2,3,4,5-tetradecoxy-D-glycero-D-
galacto-non-2-enopyranosonate (16).

Compound (15) (101 mg), 0.167 mmol) was
15 guanidinated by the procedure described in Example I, part
(E) to give compound (16) (72 mg, 51%) as a white foam.

¹H-nmr (CDCl₃) δ(ppm): 1.42 (s, 9H), 1.49 (br s, 18H), 1.89
(s, 3H), 3.12-3.65 (m, 12H), 3.78 (s, 3H), 4.07 (m, 1H),
20 4.32 (br d, 1H), 4.55-4.75 (m, 2H), 4.90-5.35 (m, 3H), 5.40-
5.68 (m, 2H), 5.91 (d, 1H), 6.38 (d, 1H), 8.42 (d, 1H).

(D) Preparation of 5-acetamido-7-{2'-[2''-(2'''-
aminoethoxy)-ethoxy]-ethyl}-carbamoyloxy-4-guanidino-
25 2,3,4,5-tetradecoxy-D-glycero-D-galacto-non-2-enopyranosonic
acid (17).

De-protection of compound (16) (70 mg,
0.082 mmol) by the procedure described in Example I, part
(F) afforded the title compound (17) (25 mg, 60%) as a
30 white solid which gave a positive Sakaguchi reaction.

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MS (FAB): 507 (M+1)⁺

¹H-nmr (D₂O) δ (ppm): 1.96 (s, 3H), 3.14-3.78 (m, 14H), 4.12 (m, 2H), 4.41 (dd, 1H), 4.52 (dd, 1H), 4.93 (dd, 1H), 5.68 (d, 1H).

Example 5 Preparation of 5-acetamido-7-(6'-biotinylaminoheptyl)-carbamoyloxy-4-guanidino-2,3,4,5-tetradecoxy-D-glycero-D-galacto-non-2-enopyranosonic acid (19)

Compound (10) (176 mg, 0.216 mmol) was stirred in trifluoroacetic (5ml) under argon at room temperature for 1hr, then vacuum evaporated to dryness to afford compound (18) which was then allowed to react with N-hydroxy-succinimide ester of biotin (110 mg, 0.324 mmol) in aqueous acetone (7.5 ml of water and 15 ml of acetone) containing a mixture of NaHCO₃ (27 mg, 0.32 mmol) and Na₂CO₃ (34 mg, 0.32 mmol) at room temperature for 3 hr. The resulting mixture was evaporated to dryness. The residue was stirred in a mixture of methanol (25 ml) and water (25 ml) containing triethylamine (4 ml) under argon at room temperature for 16 hr. The mixture was vacuum evaporated to dryness and the residue was subjected to chromatography (firstly, ethyl acetate/2-propanol/water = 5/2/1, then 2-propanol/water = 3/1). The fractions with R_f value of 0.25 in the second solvent system were collected and evaporated to dryness.

The residue was dissolved in water and treated with Dowex 50X8(H⁺) resin (0.5 g) for 0.5 hr at room temperature. The resin was washed successively with water (50 ml), methanol (20 ml), water (10 ml), then eluted with

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2MNH₄OH. The eluate was evaporated to dryness and freeze-dried to afford the title compound (19), (60 mg, 39.7%) as a white foam, which gave a positive Sakaguchi reaction.

5 MS (FAB): 701 (M+1)⁺

i.r. cm⁻¹: 3330, 2934, 1674, 1616, 1429.

¹H-nmr (D₂O) δ (ppm): 1.25-1.80 (m, 14H), 2.01(s, 3H), 2.23
10 (dd, 2H), 2.78 (d, 1H), 2.98 (dd, 1H), 3.08-3.26 (m, 4H),
3.35 (m, 1H), 3.51 (dd, 1H), 3.68 (dd, 1H), 4.02-4.28 (m,
2H), 4.32-4.08 (m, 4H), 4.92 (dd, 1H), 5.76 (d, 1H).

Example 6 Preparation of 5-acetamido-4-amino-7-[6'-
15 (6"-tert-butoxycarbonyl-aminohexylureido)-
 hexyl-carbamoyloxy-2,3,4,5-tetradecy-D-
 glycero-D-galacto-non-2-enopyranosonic acid
 (20)

Compound (6) (50 mg, 0.07 mmol) was stirred in a
20 mixture of methanol (30 ml) and water (15 ml) containing
triethylamine (4 ml) under argon at room temperature for
16 hr. The resulting mixture was vacuum evaporated to
dryness. The residue was chromatographed (silica gel
2-propanol/water = 3/1) to afford the title compound (20)
25 (30 mg, 63.5%) as a white foam.

MS (FAB): 675 (M+1)⁺

i.r. cm⁻¹: 3333, 2932, 1710, 1633, 1557.

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¹H-nmr (D₂O) δ(ppm): 1.22-1.57 (m, 25H), 1.98 (s, 3H), 2.98-3.18 (m, 8H), 3.49 (dd, 1H), 3.64 (dd, 1H), 4.02-4.28 (m, 3H), 4.53 (br d, 1H), 4.92 (dd, 1H), 5.72 (d, 1H).

5 Example 7 Preparation of 5-acetamido-7-(6'-biotinylamino-triglycinamido-hexyl)-carbamoyloxy-4-guanidino-2,3,4,5-tetradecoxy-D-glycero-D-galacto-non-2-enopyranosonic acid (21)

10 To a solution of 6-biotinylaminotriglycine (101.2 mg, 0.24 mmol) in a mixture of water (0.4 ml) and acetone (4 ml) at -10° were added successively triethylamine (24.6 mg, 0.24 mmol), N-methylmorpholine (7 mg, 0.06 mmol) and isobutyl chloroformate (35 mg, 0.256 mmol). The whole mixture was stirred under argon at -10° for 12 min. before being combined with a solution of compound (18) (92 mg, 0.156 mmol) in a mixture of water (1 ml) and acetone (1 ml) containing triethylamine (24.6 mg, 0.24 mmol) at 0-5°. The resulting reaction mixture was allowed to stir at 15-20° for 4 hr and then evaporated under reduced pressure to dryness. The residue was taken up in acetone (2 x 25 ml) and filtered to give a solid (69 mg) which was then stirred in a mixture of methanol (10 ml) and water (10 ml) containing triethylamine (5 ml) under argon at room temperature for 16 hr. The solution was evaporated to dryness and redissolved in water (20 ml) and then evaporated to dryness again. This procedure was repeated five times to produce crude compound (21) (60 mg), which was then subjected to flash column chromatography (silica gel, firstly 2-propanol/water = 5/1, and then secondly 2-propanol/water = 3/1) to afford the

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title compound (21) (17 mg, 12.5%) as a white solid which gave a positive Sakaguchi reaction (guanidine colour reaction) and a positive biotin colour reaction.

5 MS (FAB): 872 (M+1)⁺, 871 (M⁺)

i.r. cm⁻¹: 3287 (br), 1651 (br).

¹H-nmr (D₂O) δ(ppm): 1.22-1.81 (m, 14H), 1.98 (s, 3H),
10 2.37 (m, 2H), 2.78 (d, 1H), 2.95-3.55 (m, 7H), 3.61-4.16
(m, 9H), 4.39-4.67 (m, 4H), 4.92 (dd, 1H), 5.67 (d, 1H).

Example 8 5-Acetamido-7-(6'-biotinylamino-dodecanoyl-
 aminohexyl)-carbamoyl-4-guanidino-2,3,4,5-
15 tetradecyloxy-D-glycero-D-galacto-non-2-
 enopyranosonic acid (22)

This compound was prepared by coupling of biotinylaminododecanoic acid and compound (18) using carbonyl diimidazole, as described in Methods in
20 Enzymology, 1990 184 664.

¹H-nmr (CD₃OD) δ (ppm): 1.28-1.85 (m, 33H), 1.98 (s, 3H),
2.22 (m, 4H), 2.75 (d, 1H), 2.97 (dd, 1H), 3.03-3.28 (m,
6H), 3.49 (dd, 1H), 3.64 (dd, 1H), 4.07 (m, 1H), 4.20 (m,
25 1H), 4.34 (m, 2H), 4.50 (m, 2H), 5.57 (d, 1H).

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Example 9 5-Acetamido-7-(6'-biotinylamino-caproyl-aminohexyl)-carbamoyl-4-guanidino-2,3,4,5-tetradecoxy-D-glycero-D-galacto-non-2-enopyranosonic acid (23)

5 This compound was prepared by coupling of biotinylaminocaproic acid and compound (18) using similar conditions to those described in Example 8.

¹H-nmr (CD₃OD) δ (ppm): 1.30-1.85 (m, 21H), 1.99 (s, 3H),
10 2.24 (m, 4H), 2.76 (d, 1H), 2.98 (dd, 1H), 3.05-3.3 (m, 7H), 3.50 (dd, 1H), 3.66 (m, 1H), 4.07 (m, 1H), 4.21 (m, 1H), 4.37 (m, 2H), 4.52 (m, 2H), 5.58 (d, 1H).

Example 10 Preparation of 5-acetamido-7-(6'-(6''-(6'''-aminocaproyl)-triaminocaproyl)-aminohexyl)-carbamoyloxy-4-guanidino-2,3,4,5-tetradecoxy-D-glycero-D-galacto-non-2-enopyranosonic acid (25)

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(A) Preparation of methyl 5-acetamido-7-{6'-[6''-(6'''-tert-butoxycarbonylaminocaproyl)-tri-amino-caproyl]-aminohexyl}-carbamoyloxy-4-guanidino-8,9-monocarbonyldioxy-2,3,4,5-tetradecoxy-D-glycero-D-galacto-non-2-enopyranosonate (24).

20

To a solution of 6-[6'-(6''-tert-butoxycarbonyl-aminocaproyl)-di-aminocaproyl]-aminocaproic acid (150 mg, 0.263 mmol) in methanol (5 ml) at -20°C were added successively potassium t-butoxide (30 mg, 0.267 mmol), N-methylmorpholine (15 mg, 0.148 mmol), and isobutyl chloroformate (40 mg, 0.293 mmol). The whole mixture was
25 stirred under argon at -15--20°C for 20 min. before being
30

- 35 -

combined with a solution of compound (18) (140 mg, 0.238 mmol) in a mixture of methanol (1.5 ml) and water (1.5 ml) containing triethylamine (38 mg, 0.372 mmol) at 0-5°C. The resulting reaction mixture was allowed to
5 agitate at 15-20°C for 4 hrs, then evaporated under reduced pressure to dryness. The residue was taken up in acetone (50 ml x 2) and filtered to give a white solid (224 mg after air-drying) which was then chromatographed on silica gel (ethyl acetate/2-propanol/water=5/2/1) to afford
10 compound (24) (80mg, 31.5%).

MS (FAB) 1068 (M+1)⁺ 1067 (M⁺)

¹H-nmr (CD₃OD) δ 1.21-1.72 (m, 41H), 1.99 (s, 3H), 2.18 (t, 8H), 2.92-3.24 (m, 12H), 3.79 (s, 3H), 4.14 (dd, 1H), 4.51-4.78 (m, 4H), 5.19 (ddd, 1H), 5.59 (dd, 1H) 5.92 (d, 1H).
15

(B) Preparation of compound (25)

Compound (24) (29.5 mg, 0.027 mmol) was dissolved
20 in trifluoroacetic acid (2 ml) under argon. This mixture was stirred at room temperature for 1 hr, then evaporated under reduced pressure to dryness. The residue was redissolved in water (5 ml) and vacuum evaporated to dryness again before redissolving in water (2 ml) and
25 freeze-drying to form a white solid. This solid was dissolved in a mixture of methanol (10 ml), water (10 ml) and triethylamine (5 ml). The resulting solution was allowed to agitate under argon at room temperature for 16 hr before being evaporated to dryness. It was
30 redissolved in water (10 ml) and evaporated to dryness

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again. The residue was triturated successively in acetone (20 ml x 3) and ethanol (20 ml), then freeze-dried to afford compound (25) (14 mg, 56%) as a white solid, which gave a positive Sakaguchi reaction and positive ninhydrin
5 reaction.

MS (FAB) 927 (M+1)⁺

¹H-nmr (D₂O) δ 1.15-1.69 (m, 32H), 1.90 (s, 3H), 2.11-2.23
10 (m, 8H), 2.91-3.21 (m, 12H), 3.41-3.70 (m, 2H), 3.85-4.12 (m, 2H), 4.48-4.52 (m, 2H), 4.92 (dd, 1H), 5.58 (d, 1H).

Example 11 Preparation of 5-acetamido-7-{6'-[6"-{6"'-biotinylaminocaproyl)-tri-aminocaproyl]-aminohexyl}-carbamoyloxy-4-guanidino-2,3,4,5-tetradecoxy-D-glycero-D-galacto-non-2-enopyranosonic acid (26).
15

A solution of compound (24) (71.4 mg, 0.0667 mmol) in trifluoroacetic acid (2 ml) was stirred
20 under argon at room temperature for 1hr, then vacuum evaporated to dryness. The residue was dissolved in water (5 ml) and evaporated under reduced pressure to dryness. The resulting residue was dissolved in pyridine (6 ml) containing biotin-N-hydroxysuccinimide ester (36 mg,
25 0.105 mmol). The reaction mixture was stirred under argon at 40-50°C for 48 hrs, then evaporated under reduced pressure to dryness. The resulting residue was stirred in acetone (30 ml) at room temperature for 4 hr and then filtered. The precipitate was washed with acetone (10 ml x
30 2) and air-dried to afford a white solid (72 mg). This

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solid was stirred in a mixture of methanol (25 ml) and water (25 ml) containing triethylamine (5 ml) under argon at room temperature for 16 hrs before being vacuum evaporated to dryness. The residue was subjected to chromatography (silica gel, the first run ethyl acetate/2-propanol/water=5/2/1 and the second run 2-propanol/water = 3/1) to afford the title compound (26) (30.4 mg, 39.5%) as white solid, which gave a positive Sakaguchi reaction and a positive biotin colour reaction.

10

MS (FAB): 1153 (M+1)⁺

infrared spectrum: 3292, 2931, 1638, 1541 cm⁻¹.

15 ¹H-nmr (D₂O) δ (ppm) 1.12-1.62 (m, 38H), 1.91 (s, 3H), 2.12 (m, 10H), 2.65 (d, 1H), 2.82-3.15 (m, 13H), 3.30-3.62 (m, 3H), 3.81-4.18 (m, 3H), 4.25-4.56 (m, 4H), 5.51 (d, 1H).

20 Example 12 Preparation of 5-acetamido 7-{6'-[6"-{6'''-hydrazidosuccinylaminocaproyl}-triaminocaproyl]-aminohexyl}-carbamoyloxy-4-guanidino-2,3,4,5-tetra-deoxy-D-glycero-D-galacto-non-2-enopyranosonic acid (30)

To a solution of 6-{6'-[6"-{N-Boc-hydrazido-succinyl)-aminocaproyl]-diaminocaproyl}-amino-caproic acid (100mg, 0.146mmol) in anhydrous methanol (4ml) at -20°C were successively added potassium tert-butoxide (16.4mg, 0.146mmol), N-methylmorpholine (14.7mg, 0.146mmol), and isobutyl chloroformate (24mg, 0.176mmol). The whole mixture was stirred at -15°C for 12 min. before combining

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with a solution of 5-acetamido-7-(6'-aminohexyl)-
carbamoyloxy-4-guanidino-2,3,4,5-tetradecy-D-glycero-D-
galacto-non-2-enopyranosonic acid trifluoroacetic acid salt
(11) (76.6mg, 0.13mmol) in a mixture of methanol (1.5ml)
5 and water (1.5ml) containing triethylamine (16mg,
0.158mmol) at 5°C. The resulting reaction mixture was
allowed to stir at room temperature for 2 hrs, then
evaporated under reduced pressure to dryness. The residue
was taken up in acetone (50ml) and filtered. The solid was
10 washed with acetone (10ml), then air-dried. It was
subjected to column chromatography (silica gel, firstly
ethyl acetate/2-propanol/water=5/2/1, then 2-
propanol/water=3/1) to afford the compound (29) (63mg
42.5%) as a white solid.

15

¹H-nmr (CD₃OD) δ (ppm)

1-21~1.72 (m, 41H), 1.95 (s, 3H), 2.17 (m, 8H), 2.48 (AB,
4H), 3.0~3.28 (m, 12H), 3.51 (dd, 1H), 3.62 (dd, 1H), 4.13
20 (m, 1H), 4.17 (dd, 1H), 4.35 (dd, 1H), 4.46 (dd, 1H), 4.98
(dd, 1H), 5.56 (d, 1H).

Compound (29) (60mg, 0.0526mmol) was stirred in
trifluoroacetic acid (2ml) under argon at room temperature
25 for 1 hr, then vacuum evaporated to dryness. The residue
was dissolved in water (5ml) and evaporated under reduced
pressure to dryness, then lyophilized to afford compound
(34) trifluoroacetate as a white solid. This was dissolved
in water (10ml), treated with Amberlite IRA-400(OH-) resin
30 to remove trifluoroacetic acid, and filtered off. The
filtrate was evaporated to dryness. The residue was

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trituated in acetone (15ml), then freeze-dried to afford the title compound (30) (40mg, 73%) as a white solid which was stored under argon.

5 MS(FAB) 1041 (M+1)⁺ 1042 (M+2)⁺

¹H-nmr (D₂O) δ(ppm) (Trifluoroacetic acid salt)

1.15~1.68 (m, 32H), 1.93 (s, 3H) 2.18 (m, 8H), 2.52 (AB,
10 4H), 2.98-3.23 (m, 12H), 3.40~4.20 (m, 4H), 4.42 (dd, 1H),
4.52 (dd, 1H), 4.92 (dd, 1H), 5.88 (d, 1H).

Example 13 Preparation of methyl 5-acetamido-7-[6'-
(3"-hydroxycarbonylpropionyl)-aminoethyl-
15 carbamoyloxy-4-guanidino-8,9-
monocarbonyldioxy-2,3,4,5-tetradecoxy-D-
glycero-D-galacto-non-2-enopyranosonate
(32)

To a solution of methyl 5-acetamido-7-(6'-
20 aminoethyl)-carbamoyloxy-4-guanidino-8,9-monocarbonyldioxy-
2,3,4,5-tetradecoxy-D-glycero-D-galacto-non-2-
enopyranosonate trifluoroacetate (18) (100mg, 0.159mmol) in
anhydrous pyridine (2.5ml) were added dimethylaminopyridine
(24mg, 0.195mmol) and succinic anhydride (19.5mg,
25 0.195mmol). The mixture was stirred under argon at 45°C
for 20 hrs then vacuum evaporated to remove pyridine. The
residue was dissolved in methanol (10ml) and acidified to
pH 2 with 2N HCl, then the acidic solution was evaporated
to dryness. The resulting residue was subjected to flash-
30 chromatography (firstly ethyl acetate/methanol=10/1, then

- 40 -

ethyl acetate/2-propanol/water=5/2/1, silica gel) to afford compound (32) (40mg, 40%).

MS (FAB) 615 (M+1)⁺

5

¹H-nmr (CD₃OD) δ(ppm)

1.25~1.62 (m, 8H), 1.93 (s, 3H), 2.48 (AB, 4H), 2.95~3.21
(m, 4H), 3.80 (s, 3H), 4.10~4.30 (m, 1H), 4.50~4.80 (m,
10 3H), 5.0~5.25 (m, 2H), 5.59 (dd, 1H), 5.92 (dd, 1H).

Example 14 Determination of the binding of the
compounds of the invention to influenza
virus

15 Two influenza A virus strains and one influenza B
virus were used to test the ability of the compounds to
bind to whole virus influenza neuraminidase. The influenza
A strains were NWS/G70C and NWS/Tokyo and the influenza B
strain was B/Vic/02/87. The neuraminidase assay was
20 carried out following a published procedure (Potier, M. et
al, Anal. Biochem., 1979 94 287, and the measured
inhibition constants (IC₅₀) are summarised in Table 3.

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Table 3

**Binding Constants (M) for Binding Influenza
Viruses Compounds of the Invention**

Compound No.	NWS/Tokyo	NWS/G70C	B/Vic/02/87
8	4×10^{-8}	1×10^{-8}	7×10^{-8}
11	2×10^{-8}	2×10^{-8}	not done
13	2×10^{-8}	2×10^{-8}	8×10^{-8}
17	2×10^{-8}	2×10^{-8}	5×10^{-7}
19	1×10^{-8}	5×10^{-9}	2×10^{-8}
20	1×10^{-5}	1×10^{-5}	not done
21	4×10^{-8}	2×10^{-8}	1×10^{-7}
22	3×10^{-8}	5×10^{-9}	5×10^{-8}
23	2×10^{-8}	5×10^{-9}	6×10^{-8}
25	5×10^{-8}	4×10^{-8}	1×10^{-7}
26	2×10^{-8}	2×10^{-8}	4×10^{-8}
DANA	4×10^{-5}	1×10^{-5}	2×10^{-5}
GG167 (I)	1×10^{-9}	1×10^{-9}	3×10^{-9}

5

Example 15 Capture of Influenza Virus on to AvidinCoated ELISA Wells Using Compound No. 21

A 96 well ELISA plate was coated overnight with
 10 an avidin (Sigma) solution (10 µg/ml). The plate wells
 were then blocked with a PBS(Phosphate buffered saline)-
 Tween 20 solution. Compound No. 21 was added at 1 and
 10 µM to all wells in each of two rows down the plate.
 Compound No. 8 (control) was added at 10 µM to all the
 15 wells in another row. After incubation for 1 hour at room
 temperature, unbound conjugate was removed and the plate

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was washed in PBS-Tween 20. NWS/G70C Influenza A virus (H1N9 subtype) was added, starting from 2.5×10^6 virus particles in the first well (equivalent to 5×10^7 pfu/ml), and diluting in serial two-fold dilutions across the plate.

5 The virus was allowed to incubate for approximately 30 minutes and unbound virus was removed by washing. Bound virus was reacted with a polyclonal rabbit anti-hemagglutinin antibody and then the bound anti-hemagglutinin antibody was detected with a sheep anti-

10 rabbit antibody-horseradish peroxidase (Ab-HRPO) conjugate, with washings at each step. Virus bound Ab-HRPO was detected by the addition of ABTS (Sigma) as the HRPO substrate and incubation for about thirty minutes.

Both concentrations (1 and 10 μ M) of Compound No.

15 21 gave similar results; a strong absorption signal at the highest virus concentration and then a decreasing signal with virus dilution, thus clearly capturing virus and allowing detection down to at least 1.6×10^5 pfu per test. In contrast, the non-biotinylated control Compound No. 8

20 showed only a weak, unchanging absorption at all virus concentrations.

Example 16 Detection of Influenza Virus on ELISA
Plates Using Compound No. 21

25 Starting with a virus solution of approximately 1×10^6 pfu/ml, serial two-fold dilutions of NWS/G70C influenza A virus in PBS were added directly on to an ELISA (Dynatech) plate and allowed to bind by standing overnight (all the ELISA wells contained 50 μ l). After washing, the

30 plates were blocked with PBS-Tween 20 and then Compound No. 21 was added at 1 and 10 μ M to two separate rows, with

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control Compound No. 8 added at 10 μ M to another row. After incubation for 1 hour the plates were washed to remove unbound compound and then the virus was detected with Streptavidin-HRPO (Boehringer-Mannheim), again using
5 ABTS as the chromogenic substrate, with about thirty minutes incubation.

Both concentrations of Compound No. 21 allowed detection of the virus, giving an increasingly strong absorption in parallel with increasing virus concentration.
10 A minimum of about 10^5 virus particles per well could readily be detected with Compound No. 21. In contrast, the non-biotin control Compound No. 8 gave no signal at any concentration of virus.

15 Example 17 Capture and detection of a complex between
influenza virus and Compound No. 21

To allow coating of virus particles with the GG167-biotin derivative, 10, 2.5 and 0.625 μ l of a solution of the NWS/G70C influenza A virus (1×10^8 pfu/ml) was pre-
20 incubated for 1 hour in each well of separate rows of an ELISA plate. Half $\log_{10}$ dilutions of Compound No. 21 were added, starting from 1 μ M and going down to 0.00001 μ M. The virus-compound complexes were then transferred to an ELISA plate which had been pre-coated with avidin (as in
25 Example 15) and incubated for 1 hour to allow capture. The plates were washed in PBS-Tween 20 and the captured virus was detected with a polyclonal rabbit antibody directed to the virus hemagglutinin (as in Example 15).

The best result was found with a 0.01 μ M
30 concentration of Compound No. 21, which clearly allowed detection of virus down to 0.625×10^5 pfu. At higher

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concentrations of compound the signal was weaker, possibly due to the blocking of some avidin sites by free Compound No. 21, whilst at lower concentrations ($< 0.0001 \mu\text{M}$) the detection signal was also weaker, probably due to there
5 being insufficient compound to fully bind to all of the virus particles.

Direct detection of bound virus with streptavidin-HRPO was also found to be possible, though there was a lower level of signal in this case.

10

Example 18 Capture and detection of a complex between
 influenza virus and Compound No. 26

Following the same procedure given in Example 17, but using the GG167-derivative Compound No. 26, it was
15 found that the two subtypes of influenza A, NWS/G70C and NWS/Tokyo (H1N2), were both readily detected when present at 2.5×10^5 pfu/well. The concentrations of Compound No. 26 tested were from 0.001 nM to $10 \mu\text{M}$; the best results were obtained at $0.1 \mu\text{M}$ for both subtypes of virus.

20

It will be apparent to the person skilled in the art that while the invention has been described in some detail for the purpose of clarity and understanding, various modifications and alterations to the embodiments and methods described herein may be made without departing
25 from the scope of the inventive concept disclosed in this specification.



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CLAIMS

1. A method of detection of influenza virus, comprising the step of exposing a sample suspected to comprise said virus to a compound (neuraminidase binder) able to bind specifically to the active site of influenza virus neuraminidase.
2. A method according to Claim 1, in which the neuraminidase binder is attached to a support material such that virus particles will be selectively captured and concentrated when a sample is passed over or through the support.
3. A method according to Claim 1 or Claim 2, in which the neuraminidase binder is linked via a spacer group to a surface.
4. A method according to Claim 3, in which the spacer group terminates in a functionality able to bind to a surface.
5. A method according to Claim 4, in which the terminal functionality is a biotinyl group and the surface is coated with avidin, streptavidin, or an antibody directed against biotin.
6. A method according to Claim 4, in which the terminal functionality is an amino group and the surface comprises carboxy groups.
7. A method according to Claim 1, in which the neuraminidase binder is linked to a detectable label.
8. A method according to Claim 7, in which the detectable label is covalently coupled to the neuraminidase binder.
9. A method according to Claim 7 or Claim 8 in which the virus particles in a sample are exposed to a

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neuraminidase binder coupled to a detectable label, under conditions such that the binder binds selectively to the viral neuraminidase on the surface of the viral particle.

10. A method according to Claim 7 or Claim 8, which
5 comprises the steps of selective capture and concentration of the virus.

11. A method according to Claim 1, comprising the step of selective capture and selective detection of influenza virus.

10 12. A method according to Claim 11, comprising the steps of:

a) exposing the sample to a neuraminidase binder bound to a support, and

b) exposing influenza virus particles retained
15 on the support to a neuraminidase binder linked to a detectable label.

13. A method according to any one of the preceding claims, in which the neuraminidase binder has an IC_{50} for binding of less than 10^{-6} M.

20 14. A neuraminidase binder comprising an analogue of neuraminic acid (sialic acid) substituted at the C₇ or equivalent position with a spacer group suitable for coupling to a detectable label or to a surface, in which said analogue does not comprise a detectable label or
25 spacer group which is cleaved by the neuraminidase.

15. A neuraminidase binder according to Claim 14, in which the analogue of neuraminic acid is a compound of formula (II):

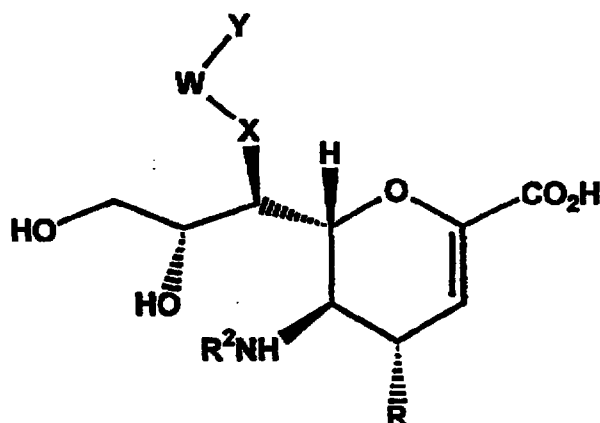
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(II)

wherein

5 R represents an azido group, an unsubstituted or substituted guanidino group, or an unsubstituted or substituted amino group;

 R² represents COCH₃, COCF₃ or SO₂CH₃;

 X represents O or NH;

10 W represents a spacer group made up of a chain of atoms having a total length of between four and one hundred atoms, and optionally also comprising substituted carbon and/or nitrogen atoms and optionally including oxygen and/or sulphur atoms;

15 Y represents OH, SH, NH₂, CH=O, CH=CH₂, CO₂H, CONHNH₂, or NH-biotinyl, or a protected form of one of these end functionalities;

 and wherein the substituent on the amino or guanidino group, when present, is a substituted or
20 unsubstituted C₁₋₆ alkyl group, or an amino, hydroxy, cyano or alkoxy carbonyl group;

 with the proviso that X-W-Y does not include the group OC(=Z)NR⁵R⁶, wherein Z represents O or S, and R⁵ and



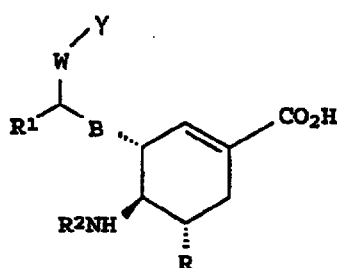


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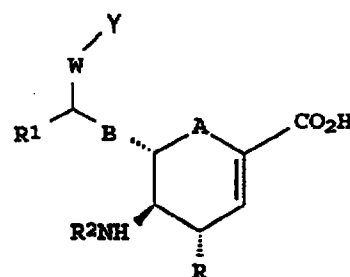
R^6 independently represents H or a hydrocarbon group optionally substituted by NH_2 , OH or SH.

16. A neuraminidase binder according to Claim 15, in which the X-W-Y group attached at the 7-position is a terminally functionalized N-substituted-alkyl carbamate.

17. A neuraminidase binder according to Claim 14, in which the analogue of neuraminic acid is a compound of formula (IIA) or (IIB)



(IIA)



(IIB)

10

wherein R, R^2 , W and Y are as defined for formula (II) in Claim 14, A and B are oxygen or CH_2 , or may represent a single bond, and R^1 is a lipophilic C_1 - C_{12} alkyl group which is optionally substituted by one or more halogen atoms, cycloalkyl, alkoxy, haloalkoxy or optionally substituted aryl groups.

18. A neuraminidase binder according to Claim 17, in which the spacer group W is selected from the group consisting of linear peptides; oligosaccharides; polyols; polyethylene glycol groups; and hydrocarbon groups linked together with oxygen or sulphur atoms or with carbonyl, amido, urea or hydrazide functionalities, or combinations of these groups.

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19. A neuraminidase binder according to any one of Claims 15 to 18, in which the protecting group for the end functionality Y is selected from the group consisting of esters of the OH, SH and CO₂H groups, carbamates of the NH₂ and CONHNH₂ groups, and acetals of the CH=O group.
20. A neuraminidase binder according to Claim 15, in which R is guanidino and R² is acetyl.
21. A neuraminidase binder according to Claim 15 or Claim 20, in which X is O and Y is NH₂ or NH-biotinyl.
22. A neuraminidase binder according to any one of Claims 15, 20 and 21, in which W is a spacer group selected from the group consisting of CONH(CH₂)_n, CONH(CH₂)_nNHCONH(CH₂)_m, CONH(CH₂)_n[NHCO(CH₂)₅]_m, CO(CH₂)_n, CO(CH₂)_nNHCONH(CH₂)_m, CO(CH₂)_nNHCO(CH₂)_q, CONH(CH₂)_nNH(COCH₂NH)_qCOCH₂, CONH(CH₂CH₂O)_nCH₂CH₂NHCO(CH₂)_q, and CONH(CH₂)_nNHCO(CH₂)_q, wherein n and m are integers between 2 and 12, and q is 0 or an integer between 1 and 12.
23. A neuraminidase binder according to any one of Claims 14 to 18, which has an IC₅₀ of less than 10⁻⁶ M.
24. A neuraminidase binder according to Claim 23, which has an IC₅₀ of less than 10⁻⁸ M.
25. A neuraminidase binder according to any one of Claims 14 to 17, selected from the compounds of Table 1, other than Compound 11.
26. 5-acetamido-7-(6'-biotinylamino-triglycinamido-hexyl)-carbamoyloxy-4-guanidino-2,3,4,5-tetradecoxy-D-glycero-D-galacto-non-2-enopyranosonic acid (21).
27. 5-acetamido-7-(6'-(6''-(6'''-aminocaproyl)-triaminocaproyl)-aminohexyl)-carbamoyloxy-4-guanidino-



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2,3,4,5-tetradecoxy-D-glycero-D-galacto-non-2-enopyranosonic acid (25).

28. 5-acetamido-7-{6'-[6"- (6"'-biotinylaminocaproyl)-tri-aminocaproyl]-aminohexyl}-carbamoxyloxy-4-guanidino-

5 2,3,4,5-tetradecoxy-D-glycero-D-galacto-non-2-enopyranosonic acid (26).

29. 5-acetamido 7-{6'-[6"- (6"'-hydrazidosuccinylaminocaproyl)-triaminocaproyl]-aminohexyl}-carbamoxyloxy-4-guanidino-2,3,4,5-tetradecoxy-D-
10 glycero-D-galacto-non-2-enopyranosonic acid (30).

30. A neuraminidase binder according to any one of Claims 14 to 29, linked to a detectable label.

31. A neuraminidase binder according to Claim 30, in which the link is covalent.

15 32. A neuraminidase binder according to Claim 30 or Claim 31, in which the detectable label is an epitope suitable for use in an antibody detection kit, in an optical assay device having an active receptive surface, or in an agglutination detection system.

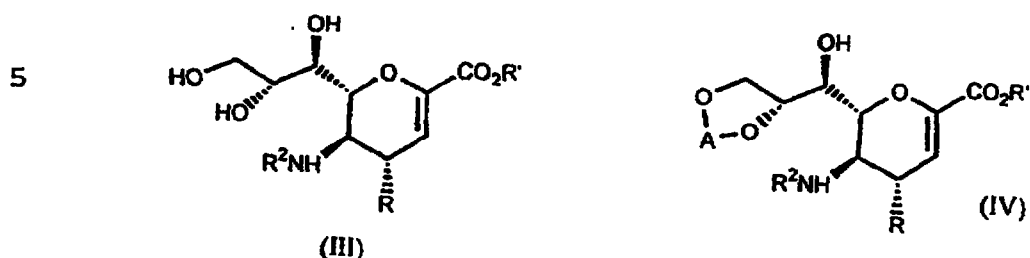
20 33. A neuraminidase binder according to Claim 14 which is linked to a detectable label, wherein the neuraminidase binder is a compound disclosed in International Patent Applications No. WO/92/06691, No. WO/91/16320 or No. WO/96/26933, or in U.S. Patent No.
25 5,453,533.

34. A diagnostic composition comprising a compound according to any one of Claims 14 to 33, together with a diagnostically-suitable carrier.

35. A method of synthesis of a neuraminidase binder
30 of formula (II) according to Claim 15, in which X is oxygen, comprising the steps of:

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a) converting a sialic acid derivative of formula (III) to an 8,9-protected compound of formula (IV),



10 in which R and R² are as defined for formula (II), R¹ is an alkyl group, and A is a protecting group;

b) acylating the 7-hydroxy group;
c) optionally extending or functionalising the spacer group W', and

15 d) removing the protecting groups.

36. A method of synthesis of a neuraminidase binder of formula (II) according to Claim 15, in which X is NH, comprising the steps of

a) reacting a compound of formula (IV) as defined in Claim 35 under Mitsunobu reaction conditions to give a leaving group ester with inverted stereochemistry at C7; and

b) displacing the C7 ester with a nucleophile.

37. A compound of general formula (V) or (VI)

25



30



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in which A is a protecting group;

W and W' are spacer groups as defined for W in
Claim 15 or Claim 18;

R and R² are as defined in Claim 15;

5 R¹ is an alkyl group; and

Y' is as defined in Claim 15 or Claim 19;

38. A method according to Claim 37, in which A is C=O
or CMe₂.

39. A method according to Claim 35, in which A in
10 formulae (IV) to (VI) is C=O or CMe₂.

40. A method according to Claim 35 or Claim 36, in
which the 7-hydroxy group is acylated with an isocyanate
group OCN-W'-Y, and where W' is a spacer group as defined
for W in Claim 15 or Claim 18 and Y is as defined in Claim
15 or Claim 19.

41. A method according to Claim 35 or Claim 36, in
which the 7-hydroxy group is acylated with an acyl chloride
of formula ClCOL, in which L is a leaving group, and L is
then displaced by an amine of formula H₂N-W'-Y, where W' is
20 as defined for W in Claim 15 or Claim 18, and Y is as
defined in Claim 15 or Claim 19.

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"Spanish Flu" of 1919 which killed millions of people.

Many viral and bacterial infections may present with symptoms similar to those of influenza. The rapid identification of respiratory viruses would enable
5 physicians to use the most appropriate therapy early in the illness. For example, an early and accurate diagnosis would allow decisions regarding the use of antibacterial therapy and hospitalisation of children and the elderly.

Laboratory tests for the identification of
10 viruses in clinical material are widely used, and a variety of different detection methodology is available. The textbook, "Laboratory Diagnosis of Viral Infections", Marcel Dekker 1992, Ed E.H. Lennette generally discusses methods which are used for a wide range of viruses,
15 including influenza virus.

A number of tests are available for the diagnosis of influenza A and B. The traditional method of identifying influenza viruses has been the use of cell culture, which is highly sensitive and specific.
20 Unfortunately, the time required for culture, isolation and identification of influenza virus can range between 2 and 10 days, thus making it virtually useless in guiding the physician to an appropriate therapy. Since influenza virus infection is normally self-limited, diagnosis must be rapid
25 if therapy is to be effective. In other words, such cell culture methods are normally only of value in providing retrospective epidemiological information.

In addition to the cell culture methods for detecting influenza, there have recently become available a
30 few rapid direct tests, which are specific for influenza A. Thus, a monoclonal immunofluorescence assay (IFA) has been

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neuraminidase, as disclosed in the publication by von Itzstein et al referred to above, such that the IC_{50} for binding is 10^{-6} M or less, and in particular certain derivatives of Compound I can be used to detect influenza virus through their ability both to selectively bind the influenza virus and at the same time to be attached to a surface or to a detectable linking group.

Summary of the Invention

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The invention therefore provides in a first aspect a method of detection of influenza virus, comprising the step of exposing a sample suspected to comprise said virus to a compound (neuraminidase binder) which is able to bind specifically to the active site of influenza virus neuraminidase, but which is not cleaved by neuraminidase.

The method of the invention is applicable to all known strains of influenza A and influenza B.

For the purposes of this specification it is to be clearly understood that the word "comprising" is to be understood to mean "including but not limited to". The words "comprise" and "comprises" are to be similarly understood.

Preferably the neuraminidase binder is linked via a spacer group to a surface or to a detectable label. The spacer group should be sufficiently long that the detectable label is exposed on the surface of the virus particle.

The method of the invention may use selective capture and concentration of the virus, followed by detection of the virus using any convenient conventional method; the detection method need not have inherent selectivity. For example, the neuraminidase binder may be attached to a support material, such as a membrane or polymer, such that virus particles will be selectively captured and concentrated when a sample is passed over or through the support. Therefore in one preferred embodiment

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will be a particularly convenient detectable label. Another convenient detection system would utilize the neuraminidase binder covalently coupled to horseradish peroxidase. The skilled person will readily be able to
5 select a suitable detection system and to optimise conditions for detection, using normal trial and error experimentation.

The neuraminidase binder may be any agent which binds to the active site of the influenza virus
10 neuraminidase, provided that it does not comprise a detectable label or spacer group which is cleaved by the neuraminidase. The binding need not be irreversible. The IC_{50} for binding is suitably 10^{-6} M or better.

Suitable agents include, but are not limited to,
15 compounds of suitable IC_{50} disclosed in our earlier patent applications, International Patent Applications No. WO/92/06691 and WO/91/16320, and also U.S. Patent No. 5,453,533 by Luo et al, U.S. Patent No. 5,512,596 by Gilead Sciences, Inc., and International Patent Application No.
20 WO/96/26933 by Gilead Sciences, Inc. Although these specifications disclose the use of the compounds disclosed therein for treatment or prophylaxis of influenza virus infection, it is not suggested that the compounds are useful for diagnostic purposes, using assays of the type
25 described herein.

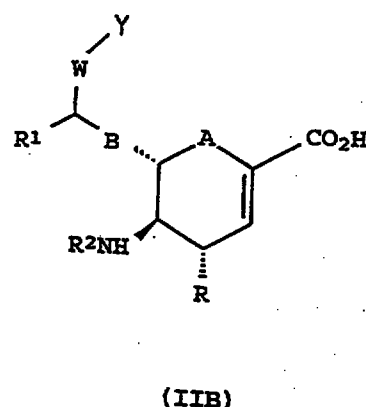
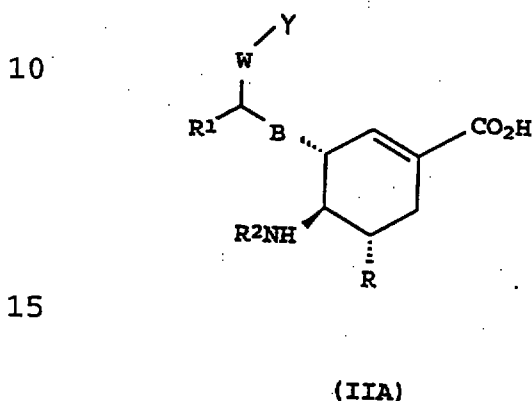
According to a second aspect, the invention provides a neuraminidase binder suitable for use in the method of the invention, comprising an analogue of neuraminic acid (sialic acid) substituted at the C_7 or at
30 an equivalent position on a branched side chain attached to the ring system of said analogue with a spacer group suitable for coupling to a detectable label or to a surface.

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with the proviso that X-W-Y does not include the group $OC(=Z)NR^5R^6$, wherein Z represents O or S, and R^5 and R^6 independently represents H or a hydrocarbon group optionally substituted by NH_2 , OH or SH, but R^5 and R^6 are not both H.

In another preferred embodiment, the invention provides neuraminidase binders of formula (IIA) or (IIB)



wherein R, R^2 , W and Y are as defined above for formula (II), A and B are oxygen or CH_2 or may represent a single bond, and R^1 is a lipophilic C_1 - C_{12} alkyl group which is optionally substituted by one or more halogen atoms, cycloalkyl, alkoxy, haloalkoxy or optionally substituted aryl groups.

Suitable spacer groups W include, but are not limited to, linear peptides, oligosaccharides, polyols, polyethylene glycol groups, hydrocarbon groups and hydrocarbon groups linked together with oxygen or sulphur atoms or with carbonyl, amido, urea or hydrazide functionalities. Spacer groups W may also comprise combinations of these various groups.

Suitable protecting groups for the end functionality Y include, but are not limited to, esters of



THE CLAIMS DEFINING THE INVENTION ARE AS FOLLOWS:

1. A method of detection of influenza virus, comprising the step of exposing a sample suspected to
5 comprise said virus to a compound (neuraminidase binder) which is able to bind specifically to the active site of influenza virus neuraminidase, but which is not cleaved by neuraminidase.
2. A method according to Claim 1, in which the
10 neuraminidase binder is attached to a support material such that virus particles will be selectively captured and concentrated when a sample is passed over or through the support.
3. A method according to Claim 1 or Claim 2, in
15 which the neuraminidase binder is linked via a spacer group to a surface.
4. A method according to Claim 3, in which the spacer group terminates in a functionality able to bind to a surface.
- 20 5. A method according to Claim 4, in which the terminal functionality is a biotinyl group and the surface is coated with avidin, streptavidin, or an antibody directed against biotin.
6. A method according to Claim 4, in which the
25 terminal functionality is an amino group and the surface comprises carboxy group.
7. A method according to Claim 1, in which the neuraminidase binder is linked to a detectable label.
8. A method according to Claim 7, in which the
30 detectable label is covalently coupled to the neuraminidase binder.
9. A method according to Claim 7 or Claim 8 in which the virus particles in a sample are exposed to a neuraminidase binder coupled to a detectable label, under
35 conditions such that the binder binds selectively to the viral neuraminidase on the surface of the viral particle.
10. A method according to Claim 7 or Claim 8, which



comprises the steps of selective capture and concentration of the virus.

11. A method according to Claim 1, comprising the step of selective capture and selective detection of influenza virus.

12. A method according to Claim 11, comprising the steps of:

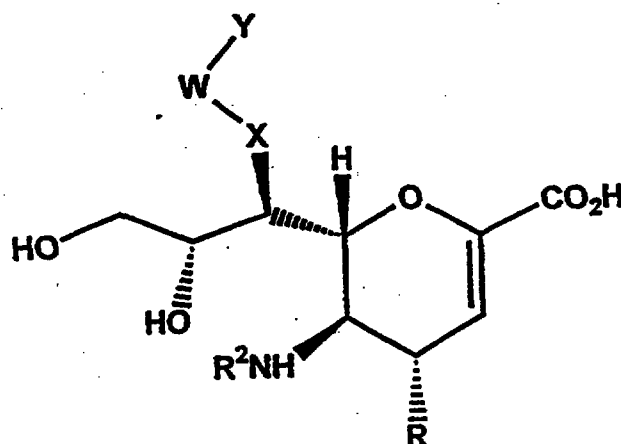
a) exposing the sample to a neuraminidase binder bound to a support, and

b) exposing influenza virus particles retained on the support to a neuraminidase binder linked to a detectable label.

13. A method according to any one of the preceding claims, in which the neuraminidase binder has an IC_{50} for binding of less than 10^{-6} M.

14. A neuraminidase binder comprising an analogue of neuraminic acid (sialic acid) substituted at the C₇ or at an equivalent position on a branched side chain attached to the ring system of said analogue with a spacer group suitable for coupling to a detectable label or to a surface, in which said analogue does not comprise a detectable label or spacer group which is cleaved by the neuraminidase.

15. A neuraminidase binder according to Claim 14, in which the analogue of neuraminic acid is a compound of formula (II):



(II)



wherein

R represents an azido group, an unsubstituted or substituted guanidino group, or an unsubstituted or substituted amino group;

5 R^2 represents COCH_3 , COCF_3 , OR SO_2CH_3 ;

X represents O or NH;

W represents a spacer group made up of a chain of atoms having a total length of between four and one hundred atoms, and optionally also comprising substituted carbon and/or nitrogen atoms and optionally including oxygen and/or sulphur atoms;

Y represents OH, SH, NH_2 , $\text{CH}=\text{O}$, $\text{CH}=\text{CH}_2$, CO_2H , CONHNH_2 , OR NH-biotinyl, or a protected form of one of these end functionalities;

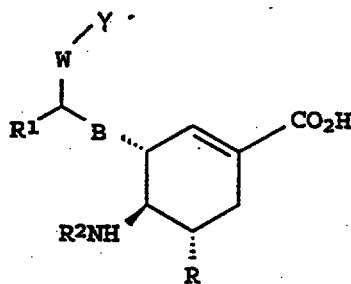
15 and wherein the substituent on the amino or guanidino group, when present, is a substituted or unsubstituted C_{1-6} alkyl group, or an amino, hydroxy, cyano or alkoxy carbonyl group;

with the proviso that X-W-Y does not include the group $\text{OC}(=\text{Z})\text{NR}^5\text{R}^6$, wherein Z represents O or S, and R^5 and R^6 independently represents H or a hydrocarbon group optionally substituted by NH_2 , OH or SH, but R^5 and R^6 are not both H.

16. A neuraminidase binder according to Claim 15, in which the X-W-Y group attached at the 7-position is a terminally functionalized N-substituted-alkyl carbamate.

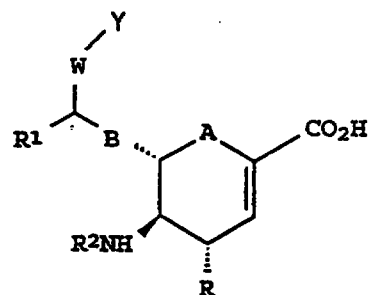
17. A neuraminidase binder according to Claim 14, in which the analogue of neuraminic acid is a compound of formula (IIA) or (IIB)

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(IIA)



(IIB)



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wherein R, R¹, W and Y are as defined for formula (II) in Claim 14, A and B are oxygen or CH₂, or may represent a single bond, and R¹ is a lipophilic C₁-C₁₂ alkyl group which is optionally substituted by one or more

5 halogen atoms, cycloalkyl, alkoxy, haloalkoxy or optionally substituted aryl groups.

18. A neuraminidase binder according to Claim 17, in which the spacer group W is selected from the group consisting of linear peptides; oligosaccharides; polyols; 10 polyethylene glycol groups; and hydrocarbon groups linked together with oxygen or sulphur atoms or with carbonyl, amido, urea or hydrazide functionalities, or combinations of these groups.



in which A is a protecting group;

W and W' are spacer groups as defined for W in
Claim 15 or Claim 18;

R and R² are as defined in Claim 15;

5 R¹ is an alkyl group; and

Y' is as defined in Claim 15 or Claim 19.

38. A method according to Claim 35, in which A is C=O
or CMe₂.

39. A compound according to Claim 37, in which A in
10 formulae (IV) to (VI) is C=O or CMe₂.

40. A method according to Claim 35 or Claim 36, in
which the 7-hydroxy group is acylated with an isocyanate
group OCN-W'-Y, and where W' is a spacer group as defined
for W in Claim 15 or Claim 18 and Y is as defined in Claim
15 15 or Claim 19.

41. A method according to Claim 35 or Claim 36, in
which the 7-hydroxy group is acylated with an acyl chloride
of formula ClCOL, in which L is a leaving group, and L is
then displaced by an amine of formula H₂N-W'-Y, where W' is
20 as defined for W in Claim 15 or Claim 18, and Y is as
defined in Claim 15 or Claim 19.

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(54) Title: METHOD OF DETECTION OF INFLUENZA VIRUS AND COMPOUNDS FOR USE THEREIN (57) Abstract The invention provides a method of detection of influenza virus which utilises compounds able to bind specifically to the active site of influenza virus neuraminidase, and novel compounds for use in the method.		