GENERATION AND SELECTION OF PROTEIN LIBRARY IN SILICO

BACKGROUND OF THE INVENTION

Cross Reference to Related Applications:

This application is a continuation-in-part of U.S. Application No. 10/153,159, filed May 20, 2002, entitled "Structure-Based Selection And Affinity Maturation of Antibody Library, and is also a continuation-in-part of Application No. 10/153,176, filed May 20, 2002, entitled "Generation Affinity Maturation of Antibody Library in Silico", both of which are a continuation-in-part of U.S. Patent Application Serial No: 10/125,687 entitled "Structure-based construction of human antibody library" filed April 17, 2002, which claims the benefit of U.S. Provisional Application Serial No: 60/284,407 entitled "Structure-based construction of human antibody library" filed April 17, 2001. These applications are incorporated herein by reference.

Field of the Invention

The present invention relates generally to a computer-aided design of a protein with binding affinity to a target molecule and, more particularly, relates to methods for screening and identifying antibodies (or immunoglobulins) with diverse sequences and high affinity to a target antigen by combining computational prediction and experimental screening of a biased library of antibodies.

Description of Related Art

Antibodies are made by vertebrates in response to various internal and external stimuli (antigens). Synthesized exclusively by the B cells, antibodies are produced in millions of forms, each with a different amino acid sequence and a different binding site for an antigen. Collectively called immunoglobulins (abbreviated as Ig), they are among the most abundant protein components in the blood, constituting about 20% of the total plasma protein by weight.

A naturally occurring antibody molecule consists of two identical "light" (L) protein chains and two identical "heavy" (H) protein chains, all held together by both hydrogen bonding and precisely located disulfide linkages. Chothia et al. (1985) J. Mol. Biol. 186:651-663; and Novotny and Haber (1985) Proc. Natl. Acad. Sci. USA 82:4592-4596. The N-terminal domains of the L and H chains together form the antigen recognition site of each antibody.

The mammalian immune system has evolved unique genetic mechanisms that enable it to generate an almost unlimited number of different light and heavy chains in a remarkably economical way by joining separate gene segments together before they are transcribed. For each type of Ig chain—κ light chains, λ light chains, and heavy chain—there is a separate pool of gene segments from which a single peptide chain is eventually synthesized. Each pool is on a different chromosome and usually contains a large number of gene segments encoding the V region of an Ig chain and a smaller number of gene segments encoding the C region. During B cell development a complete coding sequence for each of the two Ig chains to be synthesized is assembled by site-specific genetic recombination, bringing together the entire coding sequences for a V region and the coding sequence for a C region. In addition, the V region of a light chain is encoded by a DNA sequence assembled from two gene segments— a V gene segment and short joining or J gene segment. The V region of a heavy chain is encoded by a DNA sequence assembled from three gene segments— a V gene segment, a J gene segment and a diversity or D segment.

The large number of inherited V, J and D gene segments available for encoding Ig chains makes a substantial contribution on its own to antibody diversity, but the combinatorial joining of these segments greatly increases this contribution. Further, imprecise joining of gene segments and somatic mutations introduced during the V-D-J segment joining at the pre-B cell stage greatly increases the diversity of the V regions.

After immunization against an antigen, a mammal goes through a process known as affinity maturation to produce antibodies with higher affinity toward the antigen. Such antigen-driven somatic hypermutation fine-tunes antibody responses to a given antigen,

presumably due to the accumulation of point mutations specifically in both heavy-and light-chain V region coding sequences and a selected expansion of high-affinity antibody-bearing B cell clones.

Structurally, various functions of an antibody are confined to discrete protein domains (regions). The sites that recognize and bind antigen consist of three hyper-variable or complementarity-determining regions (CDRs) that lie within the variable (V_H and V_L) regions at the N-terminal ends of the two H and two L chains. The constant domains are not involved directly in binding the antibody to an antigen, but are involved in various effector functions, such as participation of the antibody in antibody-dependent cellular cytotoxicity.

The domains of natural light and heavy chains have the same general structures, and each domain comprises four framework regions, whose sequences are somewhat conserved, connected by three CDRs. The four framework regions largely adopt a β-sheet conformation and the CDRs form loops connecting, and in some cases forming part of, the β-sheet structure. The CDRs in each chain are held in close proximity by the framework regions and, with the CDRs from the other chain, contribute to the formation of the antigen binding site.

Generally all antibodies adopt a characteristic "immunoglobulin fold". Specifically, both the variable and constant domains of an antigen binding fragment (Fab, consisting of V_L and C_L of the light chain and V_H and C_H1 of the heavy chain) consist of two twisted antiparallel β-sheets which form a β- sandwich structure. The constant regions have three- and four-stranded β-sheets arranged in a Greek key-like motif, while variable regions have a further two short β strands producing a five-stranded β-sheet.

The V_L and V_H domains interact via the five-stranded β sheets to form a nine-stranded β barrel of about 8.4Å radius, with the strands at the domain interface inclined at approximately 50° to one another. The domain pairing brings the CDR loops into close proximity. The CDRs themselves form some 25% of the V_L/V_H domain interface.

The six CDRs, (CDR-L1, -L2 and -L3 for the light chain, and CDR-H1, -H2 and -H3 for the heavy chain), are supported on the β barrel framework, forming the antigen binding site. While their

sequences are hypervariable in comparison with the rest of the immunoglobulin structure, some of the loops show a relatively high degree of both sequence and structural conservation. In particular, CDR-L2 and CDR-H1 are highly conserved in conformation.

Chothia and co-workers have shown that five of the six CDR loops (all except CDR-H3) adopt a discrete, limited number of main-chain conformations (termed canonical structures of the CDRs) by analysis of conserved key residues. Chothia and Lesk (1987) J. Mol. Biol. 196:901-917; Chothia et al. (1989) Nature (London) 342:877; and Chothia et al. (1998) J. Mol. Biol. 278:457-479. The adopted structure depends on both the CDR length and the identity of certain key amino acid residues, both in the CDR and in the contacting framework, involved in its packing. The canonical conformations were determined by specific packing, hydrogen bonding interactions, and stereochemical constraints of only these key residues which serve as structural determinants.

Various methods have been developed for modeling the three dimensional structures of the antigen binding site of an antibody. Other than x-ray crystallography, nuclear magnetic resonance (NMR) spectroscopy has been used in combination with computer model building to study the atomic details of antibody-ligand interactions. Dwek et al. (1975) Eur. J. Biochem. 53:25-39. Dwek and coworkers used spin-labeled hapten to deduce the combining site of the MoPC 315 myeloma protein for dinitrophenyl. Similar analysis has also been done using anti-spin labeled monoclonal antibodies (Anglister et al. (1987) Biochem. 26: 6958-6064) and on the anti-2-phenyloxazolone Fv fragments (McManus and Riechmann (1991) Biochem. 30:5851-5857).

Computer-implemented analysis and modeling of antibody combining site (or antigen binding site) are based on homology analysis comparing the target antibody sequence with those of antibodies with known structures or structural motifs in existing data bases (e.g. the Brookhaven Protein Data Bank). By using such homology-based modeling methods approximate three-dimensional structure of the target antibody is constructed. Early antibody modeling was based on the conjecture that CDR loops with identical length and different sequence may adopt similar conformations. Kabat and Wu (1972) Proc.

Natl. Acad. Sci. USA 69: 960-964. A typical segment match algorithm is as follows: given a loop sequence, the Protein Data Bank can be searched for short, homologous backbone fragments (e.g. tripeptides) which are then assembled and computationally refined into a new combining site model.

More recently, the canonical loop concept has been incorporated into the computer-implemented structural modeling of an antibody combining site. In its most general form, the canonical structure concept assumes that (1) sequence variation at other than canonical positions is irrelevant for loop conformation, (2) canonical loop conformations are essentially independent of loop-loop interactions, and (3) only a limited number of canonical motifs exist and these are well represented in the database of currently known antibody crystal structures. Based on this concept, Chothia predicted all six CDR loop conformations in the lysozyme-binding antibody D1.3 and five canonical loop conformations in four other antibodies. Chothia (1989), *supra.* It is also possible to improve the modeling of CDRs of antibody structures by combining the homology-based modeling with conformational search procedures. Martin, A.C.R. (1989) PNAS 86, 9268-72.

Besides modeling a specific antibody structure, efforts have been made in generating artificial (or synthetic) libraries of antibodies which are screened against a specific target antigen. A fully synthetic combinatorial antibody library has been designed based on modular consensus frameworks and CDRs randomized with trinucleotides. Knappik et al. (2000) J. Mol. Biol. 296:57-86. In this study, the human antibody repertoire was analyzed in terms of structure, amino acid sequence diversity and germline usage. Modular consensus framework sequences with seven V_H and seven V_L were derived to cover 95% of variable germline families and optimized for expression in E. coli. After cloning the genes in all 49 combinations into a phagemid vector, a set of antibody phage display libraries were created, totaling $2x10^9$ members in the libraries.

Phage display technology has been used extensively to generate large libraries of antibody fragments by exploiting the capability of bacteriophage to express and display biologically functional protein molecule on its surface. Combinatorial libraries of antibodies have been

generated in bacteriophage lambda expression systems which may be screened as bacteriophage plaques or as colonies of lysogens (Huse et al. (1989) Science 246: 1275; Caton and Koprowski (1990) Proc. Natl. Acad. Sci. (U.S.A.) 87: 6450; Mullinax et al (1990) Proc. Natl. Acad. Sci. (U.S.A.) 87: 8095; Persson et al. (1991) Proc. Natl. Acad. Sci. (U.S.A.) 88: 2432). Various embodiments of bacteriophage antibody display libraries and lambda phage expression libraries have been described (Kang et al. (1991) Proc. Natl. Acad. Sci. (U.S.A.) 88: 4363; Clackson et al. (1991) Nature 352: 624; McCafferty et al. (1990) Nature 348: 552; Burton et al. (1991) Proc. Natl. Acad. Sci. (U.S.A.) 88: 10134; Hoogenboom et al. (1991) Nucleic Acids Res. 19: 4133; Chang et al. (1991) J. Immunol. 147: 3610; Breitling et al. (1991) Gene 104: 147; Marks et al. (1991) J. Mol. Biol. 222: 581; Barbas et al. (1992) Proc. Natl. Acad. Sci. (U.S.A.) 89: 4457; Hawkins and Winter (1992) J. Immunol. 22: 867; Marks et al. (1992) Biotechnology 10: 779; Marks et al. (1992) J. Biol. Chem. 267: 16007; Lowman et al (1991) Biochemistry 30: 10832; Lerner et al. (1992) Science 258: 1313). Also see review by Rader, C. and Barbas, C. F. (1997) "Phage display of combinatorial antibody libraries" Curr. Opin. Biotechnol. 8:503-508.

Generally, a phage library is created by inserting a library of random oligonucleotides or a cDNA library encoding antibody fragment such as V_L and V_H into gene 3 of M13 or fd phage. Each inserted gene is expressed at the N-terminal of the gene 3 product, a minor coat protein of the phage. As a result, peptide libraries that contain diverse peptides can be constructed. The phage library is then affinity screened against immobilized target molecule of interest, such as an antigen, and specifically bound phage particles are recovered and amplified by infection into *Escherichia coli* host cells. Typically, the target molecule of interest such as a receptor (e.g., polypeptide, carbohydrate, glycoprotein, nucleic acid) is immobilized by a covalent linkage to a chromatography resin to enrich for reactive phage particles by affinity chromatography and/or labeled for screening plaques or colony lifts. This procedure is called biopanning. Finally, high affinity phage clones can be amplified and sequenced for deduction of the specific peptide sequences.

A method for humanizing antibody by using computer modeling

has also been developed by Queen et al. US Patent No. 5,693,762. The structure of a non-human, donor antibody (e.g., a mouse monoclonal antibody) is predicted based on computer modeling and key amino acids in the framework are predicted to be necessary to retain the shape, and thus the binding specificity of the CDRs. These few key murine donor amino acids are selected based on their positions and characters within a few defined categories and substituted into a human acceptor antibody framework along with the donor CDRs. For example, category 1: The amino acid position is in a CDR as defined by Kabat et al. Kabat and Wu (1972) Proc. Natl. Acad. Sci. USA 69: 960-964. Category 2: If an amino acid in the framework of the human acceptor immunoglobulin is unusual, and if the donor amino acid at that position is typical for human sequences, then the donor amino acid rather than the acceptor many be selected. Category 3: In the position immediately adjacent to one or more of the 3 CDR's in the primary sequence of the humanized immunoglobulin chain, the donor amino acid(s) rather than the acceptor amino acid may be selected. Based on these criteria, a series of elaborate selections of individual amino acids from the donor antibody is conducted. The resulting humanized antibody usually includes about 90% human sequence. The humanized antibody designed by computer modeling is tested for antigen binding. Experimental results such as binding affinity are fed back to the computer modeling process to fine-tune the structure of the humanized antibody. The redesigned antibody can then be tested for improved biological functions. Such a reiterate fine tuning process can be labor intensive and unpredictable.

SUMMARY OF THE INVENTION

Disclosed herein is an innovative methodology for efficiently generating and screening protein libraries for optimized proteins with desirable biological functions, such as improved binding affinity towards biologically and/or therapeutically important target molecules. The process may be carried out computationally in a high throughput manner by mining the ever-expanding databases of protein sequences for all organisms, especially human. The evolutionary data of proteins may be utilized to expand both sequence and structure space of the protein libraries for functional screening in vitro or in vivo. By using the inventive methodology, an expanded and yet functionally biased library of proteins such as antibodies may be constructed based on computational evaluation of extremely diverse protein sequences and functionally relevant structures in silico.

In one embodiment, a method is provided for designing and selecting protein(s) with desirable function(s). The method is preferably implemented in a computer through in silico selection of protein sequences based on the amino acid sequence of a target structural/functional motif or domain in a lead protein, herein after referred to as the "lead sequence". The lead sequence may be employed to search databases of protein sequences. The choice of the database depends on the specific functional requirement of the designed motifs. For example, if the lead protein is an enzyme and the target motif includes the active site of the enzyme, databases of proteins/peptides of a particular origin, organism, species or combinations thereof, may be queried using various search criteria to yield a hit list of sequences each of which can substitute the target motif in the lead protein. A similar approach may be used for designing other motifs or domains of the lead protein. The designed sequences for each individual motif/domain may be combined to generate a library of designed proteins. In addition, to reduce immunogenicity of the designed proteins for human applications such as therapeutics or diagnosis, databases of proteins of human origin or humanized proteins are preferably searched to yield the hit list of sequences, especially for motifs derived from sites of the lead protein that may not be structurally or functionally critical. The library of designed

proteins may be tested experimentally to yield proteins with improved biological function(s) over the lead protein.

In another embodiment, the method comprises the steps of:

providing an amino acid sequence derived from a lead protein, the amino acid sequence being designated as a lead sequence;

comparing the lead sequence with a plurality of tester protein sequences; and

selecting from the plurality of tester protein sequences at least two peptide segments that have at least 15% sequence identity with the lead sequence, the selected peptide segments forming a hit library; and

forming a library of designed proteins by substituting the lead sequence with the hit library.

Optionally, the method further comprises the steps of:

building an amino acid positional variant profile of the hit library;

combining the amino acid variants in the hit library to produce a combination of hit variants which form a hit variant library; and

selecting proteins with a desirable function from the hit variant library.

Also optionally, the method further comprises the steps of:

determining if a member of the hit library or the hit variant library is structurally compatible with a three-dimensional structure of the lead sequence or the lead protein by using a scoring function; and

selecting the members that score equal to or better than the lead sequence or the lead protein.

Also optionally, the method further comprises the steps of:

constructing a nucleic acid library comprising DNA segments encoding the amino acid sequences of the hit library, the hit variant library or the selected members based on the structural evaluation described above;

expressing the nucleic acid library to generate a library of recombinant proteins; and

selecting proteins with the desired function from the library of recombinant proteins .

Also optionally, the method further comprises the steps of:

building an amino acid positional variant profile of the hit library;

converting amino acid positional variant profile of the hit library into a nucleic acid positional variant profile by back-translating the amino acid positional variants into their corresponding genetic codons;

constructing a degenerate nucleic acid library of DNA segments by combinatorially combining the nucleic acid positional variants;

expressing the degenerate nucleic acid library to generate a library of recombinant proteins; and

selecting proteins with the desired function from the library of recombinant proteins .

Optionally, the genetic codons may be the ones that are preferred for expression in cells of a particular organism, such as mammalian cells, insect, plant, yeast, or bacteria. Optionally, genetic codons may be the ones that can reduce the size chosen such that the diversity of the degenerate nucleic acid library of DNA segments is within the experimentally coverable diversity without undue experimental efforts, for example, to be below $1x10^7$, and preferably below $1x10^6$.

The lead protein may be a protein whose function is desired to be improved or altered, preferably a biological function in vitro or in vivo. The lead protein may be a full-length protein, oligopeptide or peptide, and may also be an unnatural protein or peptide. Optionally, the lead protein may be a fragment or domain of a known protein, including but not limited to structural and/or functional domains such as enzymatic domains, binding domains, and smaller fragments or motifs, such as turns, helixes and loops. In addition, protein variants, i.e. non-naturally occurring protein analog structures, may be used.

The lead protein is preferably a protein used in industry, therapeutics and/or diagnosis. The type of lead protein may be a ligand, cell surface receptor, antigen, antibody, cytokine, hormone, transcription factor, signaling module, cytoskeletal protein and enzyme.

Specific classes of enzymes include, but are not limited to, hydrolases such as proteases, carbohydrases, lipases; isomerases such as racemases, epimerases, tautomerases, or mutases; transferases, kinases, oxidoreductases, and phophatases. Particular examples of enzymes are listed in the Swiss-Prot enzyme database.

Other examples of the lead protein cytokines include, but are not limited to, IL-1, IL-2, IL-3, IL-4, IL-5, IL6, IL-8, IL-10, IFN-β, INF-γ, IFN-α-2a; IFN α-2B, TNF- α; CD40 ligand (chk), Human Obesity Protein Leptin, Granulocyte-Macrophage Colony-Stimulating Factor (GMCSF), Bone Morphogenetic Protein-7, Ciliary Neurotrophic Factor, Granulocyte-Macrophage Colony-Stimulating Factor, Monocyte Chemoattractant Protein 1, Macrophage Migration Inhibitory Factor, Human Glycosylation-Inhibiting Factor, Human Rantes, Human Macrophage Inflammatory Protein 1 Beta, human growth hormone, Leukemia Inhibitory Factor, Human Melanoma Growth Stimulatory Activity, neutrophil activating peptide-2, Cc-Chemokine Mcp-3, Platelet Factor M2, Neutrophil Activating Peptide 2, Eotaxin, Stromal Cell-Derived Factor-1, Insulin, Insulin-like Growth Factor I, Insulin-like Growth Factor II, Transforming Growth Factor B1, Transforming Growth Factor B2, Transforming Growth Factor B3, Transforming Growth Factor A, Vascular Endothelial growth factor (VEGF), acidic Fibroblast growth factor, basic Fibroblast growth factor, Endothelial growth factor, Nerve growth factor, Brain Derived Neurotrophic Factor, Ciliary Neurotrophic Factor, Platelet Derived Growth Factor, Human Hepatocyte Growth Factor, Glial Cell-Derived Neurotrophic Factor, Erythropoietin; coaguation factors including, but not limited to, TPA and Factor VIIa; receptors, including, but not limited to, the extracellular Region Of Human Tissue Factor Cytokine-Binding Region Of Gp130, G-CSF receptor, erythropoietin receptor, Fibroblast Growth Factor receptor, TNF receptor, IL-1 receptor, IL-1 receptor/IL1ra complex, IL4 receptor, INF-γ, receptor alpha chain, MHC Class I, MHC Class II, T Cell Receptor, Insulin receptor, insulin receptor tyrosine kinase and human growth hormone receptor.

In yet another embodiment, a method is provided for in silico design and selection of protein sequences based on a lead structural template. Ensembles of different sequences having substantially similar structures to the structural template may be employed as lead sequences to search databases of protein structures for remote homologues of the lead sequences having low sequence identity and yet structurally similar. By using the method, a library of

diverse protein sequences may be constructed and screened experimentally in vitro or in vivo for protein mutants with improved or desired function(s).

In a further embodiment, the inventive methodology is implemented in designing antibodies that are diverse in sequence and yet functionally related to each other. Based on the designed antibody sequences, a library of antibodies may be constructed to include diverse sequences in the complementary determining regions (CDRs) and/or humanized frameworks (FRs) of a non-human antibody in a high throughput manner. This library of antibodies may be screened against a wide variety of target molecules for novel or improved functions.

In a yet further embodiment, a method is provided for in silico selection of antibody sequences based on the amino acid sequence of a region in a lead antibody, herein after referred to as the "lead sequence". The lead sequence may be employed to search databases of protein sequences. The choice of the database may depend on the specific functional requirement of the designed motifs. For example: in order to design the framework regions of variable chains for therapeutic application, collections of protein sequences that are evolutionarily related such as fully human immunoglobulin sequences and human germline immunoglobulin sequences might be used except for a few structurally critical sites. This would reduce the immunogenic response by preserving the origin of the sequences by introducing as few foreign mutants as possible in this highly conserved region (for framework regions). On the other hand, diverse sequence databases such as immunoglobulin sequences of various species or even unrelated sequence in genbank may be used to design the CDRs in order to improve binding affinity with antigens in this highly variable region. By using the method, a library of diverse antibody sequences may be constructed and screened experimentally in vitro or in vivo for antibody mutants with improved or desired function(s).

According to a first aspect, the method for constructing a library of antibody sequences comprises the steps of:

providing an amino acid sequence of the variable region of the heavy chain (V_H) or light chain (V_L) of a lead antibody;

identifying the amino acid sequences in the CDRs of the lead antibody;

selecting one of the CDRs in the V_H or V_L region of the lead antibody;

providing an amino acid sequence that comprises at least 3 consecutive amino acid residues in the selected CDR, the selected amino acid sequence being a lead sequence;

comparing the lead sequence with a plurality of tester protein sequences; and

selecting from the plurality of tester protein sequences at least two peptide segments that have at least 15% sequence identity with the lead sequence, the selected peptide segments forming a hit library.

The method may further comprise the step of:

constructing a nucleic acid library comprising DNA segments encoding the amino acid sequences of the hit library.

Optionally, the method may further comprise the steps of:

building an amino acid positional variant profile of the hit library;

converting amino acid positional variant profile of the hit library into a nucleic acid positional variant profile by back-translating the amino acid positional variants into their corresponding genetic codons; and

constructing a degenerate nucleic acid library of DNA segments by combinatorially combining the nucleic acid positional variants.

Optionally, the genetic codons may be the ones that are preferred for expression in bacteria. Optionally, genetic codons may be the ones that can reduce the size chosen such that the diversity of the degenerate nucleic acid library of DNA segments is within the experimentally coverable diversity without undue experimental efforts, for example, to be below $1x10^7$, and preferably below $1x10^6$.

In another embodiment, the method comprises the steps of:

providing an amino acid sequence of the variable region of the heavy chain (V_H) or light chain (V_L) of a lead antibody;

identifying the amino acid sequences in the CDRs and FRs of the lead antibody;

selecting one of the CDRs in the V_H or V_L region of the lead antibody;

providing a first amino acid sequence that comprises at least 3 consecutive amino acid residues in the selected CDR, the selected amino acid sequence being a CDR lead sequence;

comparing the CDR lead sequence with a plurality of CDR tester protein sequences;

selecting from the plurality of CDR tester protein sequences at least two peptide segments that have at least 15% sequence identity with the CDR lead sequence, the selected peptide segments forming a CDR hit library;

selecting one of the FRs in the V_H or V_L region of the lead antibody;

providing a second amino acid sequence that comprises at least 3 consecutive amino acid residues in the selected FR, the selected amino acid sequence being a FR lead sequence;

comparing the FR lead sequence with a plurality of FR tester protein sequences; and

selecting from the plurality of FR tester protein sequences at least two peptide segments that have at least 15% sequence identity with the FR lead sequence, the selected peptide segments forming a FR hit library; and

combining the CDR hit library and the FR hit library to form a hit library.

According to the method, the plurality of CDR tester protein sequences may comprise amino acid sequences of human or non-human antibodies.

Also according to the method, the plurality of FR tester protein sequences may comprise amino acid sequences of human origins, preferably human or humanized antibodies (e.g., antibodies with at least 50% human sequence, preferably at least 70% human sequence, more preferably at least 90 % human sequence, and most preferably at least 95% human sequence in V_H or V_L), more preferably fully human antibodies, and most preferably human germline antibodies.

Also according to the method, at least one of the plurality of CDR tester protein sequences may be different from the plurality of FR tester protein sequences.

Also according to the method, the plurality of CDR tester protein sequences may be human or non-human antibody sequences and the plurality of FR tester protein sequences may be human antibody sequences, preferably human germline antibody sequences.

The method may further comprise the step of:

constructing a nucleic acid library comprising DNA segments encoding the amino acid sequences of the hit library.

Optionally, the method may further comprise the steps of:

building an amino acid positional variant profile of the CDR hit library;

converting the amino acid positional variant profile of the CDR hit library into a first nucleic acid positional variant profile by back-translating the amino acid positional variants into their corresponding genetic codons; and

constructing a degenerate CDR nucleic acid library of DNA segments by combinatorially combining the nucleic acid positional variants.

Optionally, the genetic codons may be the ones that are preferred for expression in bacteria. Optionally, genetic codons may be the ones that can reduce the size chosen such that the diversity of the degenerate nucleic acid library of DNA segments is within the experimentally coverable diversity without undue experimental efforts, such as diversity below $1x10^7$, preferably below $1x10^6$.

In yet another embodiment, the method comprises the steps of:

providing an amino acid sequence of the variable region of the heavy chain (V_H) or light chain (V_L) of a lead antibody;

identifying the amino acid sequences in the FRs of the lead antibody;

selecting one of the FRs in the V_H or V_L region of the lead antibody;

providing a first amino acid sequence that comprises at least 3 consecutive amino acid residues in the selected FR, the selected amino acid sequence being a first FR lead sequence;

comparing the first lead FR sequence with a plurality of FR tester protein sequences; and

selecting from the plurality of FR tester protein sequences at least two peptide segments that have at least 15% sequence identity with the first FR lead sequence, the selected peptide segments forming a first FR hit library.

The method may further comprise the steps of

providing a second amino acid sequence that comprises at least 3 consecutive amino acid residues in a FR that is different from the selected FR, the selected amino acid sequence being a second FR lead sequence;

comparing the second FR lead sequence with the plurality of FR tester protein sequences; and

selecting from the plurality of FR tester protein sequences at least two peptide segments that have at least 15% sequence identity with the second FR lead sequence, the selected peptide segments forming a second FR hit library; and

combining the first FR hit library and the second FR hit library to form a hit library.

According to the method, the lead CDR sequence may comprise at least 5 consecutive amino acid residues in the selected CDR. The selected CDR may be selected from the group consisting of V_H CDR1, V_H CDR2, V_H CDR3, V_L CDR1, V_L CDR2, and V_L CDR3 of the lead antibody.

Also according to the method, the lead FR sequence may comprise at least 5 consecutive amino acid residues in the selected FR. The selected FR may be selected from the group consisting of V_H FR1, V_H FR2, V_H FR3, V_H FR4, V_L FR1, V_L FR2, V_L FR3 and V_L FR4 of the lead antibody.

The method may further comprise the step of:

constructing a nucleic acid or degenerate nucleic acid library comprising DNA segments encoding the amino acid sequences of the hit library.

In a further embodiment, a method is provided for in silico selection of antibody sequences based on the amino acid sequence of a region in a lead antibody, i.e., the "lead sequence", and its 3D structure. The structure of the lead sequence may be employed to search databases of protein structures for segments having similar 3D structures. These segments may be aligned to yield a sequence profile,

herein after referred to as the "lead sequence profile". The lead sequence profile may be employed to search databases of protein sequences for remote homologues of the lead sequence having low sequence identity and yet structurally similar. By using the method, a library of diverse antibody sequences may be constructed and screened experimentally in vitro or in vivo for antibody mutants with improved or desired function(s).

In a yet further embodiment, the method comprises the steps of:

providing an amino acid sequence of the variable region of the heavy chain (V_H) or light chain (V_L) of a lead antibody;

identifying the amino acid sequences in the CDRs of the lead antibody;

selecting one of the CDRs in the V_H or V_L region of the lead antibody;

providing an amino acid sequence that comprises at least 3 consecutive amino acid residues in the selected CDR, the selected amino acid sequence being a lead sequence;

providing a three-dimensional structure of the lead sequence;

building a lead sequence profile based on the structure of the lead sequence;

comparing the lead sequence profile with a plurality of tester protein sequences; and

selecting from the plurality of tester protein sequences at least two peptide segments that have at least 10% sequence identity with lead sequence, the selected peptide segments forming a hit library.

According to the method, the three-dimensional structure of the lead sequence may be a structure derived from X-crystallography, nuclear magnetic resonance (NMR) spectroscopy or theoretical structural modeling.

According to the method, the step of building a lead sequence profile may include:

comparing the structure of the lead sequence with the structures of a plurality of tester protein segments;

determining the root mean square difference of the main chain conformations of the lead sequence and the tester protein segments;

selecting the tester protein segments with root mean square difference of the main chain conformations less than 5 Å, preferably less than 4 Å, more preferably less than 3 Å, and most preferably less than 2 Å; and

aligning the amino acid sequences of the selected tester protein segments with the lead sequence to build the lead sequence profile.

Optionally, the structures of the plurality of tester protein segments may be retrieved from the protein data bank.

Optionally, the step of building a lead sequence profile may include:

comparing the structure of the lead sequence with the structures of a plurality of tester protein segments;

determining the Z-score of the main chain conformations of the lead sequence and the tester protein segments;

selecting the segments of the tester protein segments with the Z-score higher than 2, preferably higher than 3, more preferably higher than 4, and most preferably higher than 5; and

aligning the amino acid sequences of the selected tester protein segments with the lead sequence to build the lead sequence profile.

Optionally, the step of building a lead sequence profile may be implemented by an algorithm selected from the group consisting of CE, MAPS, Monte Carlo and 3D clustering algorithms.

The method may further comprise the step of:

constructing a nucleic acid library comprising DNA segments encoding the amino acid sequences of the hit library.

Optionally, the method may further comprise the steps of:

building an amino acid positional variant profile of the hit library;

converting amino acid positional variant profile of the hit library into a nucleic acid positional variant profile by back-translating the amino acid positional variants into their corresponding trinucleotide codons; and

constructing a degenerate nucleic acid library of DNA segments by combinatorially combining the nucleic acid positional variants.

Any of the above methods may further comprise the following steps:

introducing the DNA segments in the nucleic acid or degenerate nucleic acid library into cells of a host organism;

expressing the DNA segments in the host cells such that recombinant antibodies containing the amino acid sequences of the hit library are produced in the cells of the host organism; and

selecting the recombinant antibody that binds to a target antigen with affinity higher than 10^6 M^{-1}, preferably 10^7 M^{-1}, more preferably 10^8 M^{-1}, and most preferably 10^9 M^{-1}.

In yet another embodiment, a method is provided for in silico selection of antibody sequences based on 3D structure of a lead antibody. A lead sequence or sequence profile from a specific region of the lead antibody may be employed to search databases of protein sequences for remote homologues of the lead sequence having low sequence identity and yet structurally similar. These remote homologues form a hit library. The sequences in the hit library may be subjected to evaluation for their structural compatibility with a 3D structure of the lead antibody, hereinafter referred to as the "lead structural template". Sequences in the hit library that are structurally compatible with the lead structural template may be selected and screened experimentally in vitro or in vivo for antibody mutants with improved or desired function(s).

According to a second aspect, the method for constructing a library of antibodies based on a structure of a lead antibody comprises the steps of:

providing an amino acid sequence of the variable region of the heavy chain (V_H) or light chain (V_L) of a lead antibody, the lead antibody having a known three dimensional structure which is defined as a lead structural template;

identifying the amino acid sequences in the CDRs of the lead antibody;

selecting one of the CDRs in the V_H or V_L region of the lead antibody;

providing an amino acid sequence that comprises at least 3 consecutive amino acid residues in the selected CDR, the selected amino acid sequence being a lead sequence;

comparing the lead sequence profile with a plurality of tester protein sequences;

selecting from the plurality of tester protein sequences at least two peptide segments that have at least 10% sequence identity with lead sequence, the selected peptide segments forming a hit library;

determining if a member of the hit library is structurally compatible with the lead structural template using a scoring function; and

selecting the members of the hit library that score equal to or better than or equal to the lead sequence.

According to the method, the scoring function may be an energy scoring function selected from the group consisting of electrostatic interactions, van der Waals interactions, electrostatic solvation energy, solvent-accessible surface solvation energy, and conformational entropy.

Optionally, the scoring function may be one incorporating a forcefield selected from the group consisting of the Amber forcefield, Charmm forcefield, the Discover cvff forcefields, the ECEPP forcefields, the GROMOS forcefields, the OPLS forcefields, the MMFF94 forcefield, the Tripos forcefield, the MM3 forcefield, the Dreiding forcefield,and UNRES forcefield,and other knowledge-based statistical forcefield (mean field) and structure-based thermodynamic potential functions.

Also according to the method, the step of selecting the members of the hit library includes selecting the members of the hit library that may have a lower or equal total energy than that of the lead sequence calculated based on a formula of

$$\Delta E_{total} = E_{vdw} + E_{bond} + E_{angel} + E_{electrostatics} + E_{solvation}$$

Also according to the method, the step of selecting the members of the hit library may include selecting the members of the hit library that have a lower binding free energy than that of the lead sequence calculated as the difference between the bound and unbound states using a refined scoring function

$$\Delta G_b = \Delta G_{MM} + \Delta G_{sol} - T\Delta S_{ss}$$

where

$$\Delta G_{MM} = \Delta G_{ele} + \Delta G_{vdw} \quad (1)$$

$$\Delta G_{sol} = \Delta G_{ele\text{-}sol} + \Delta G_{ASA} \quad (2)$$

The method may further comprise the step of:

constructing a nucleic acid library comprising DNA segments encoding the amino acid sequences of the hit library.

Optionally, the method may further comprise the steps of:

building an amino acid positional variant profile of the hit library;

converting amino acid positional variant profile of the hit library into a nucleic acid positional variant profile by back-translating the amino acid positional variants into their corresponding trinucleotide codons; and

constructing a degenerate nucleic acid library of DNA segments by combinatorially combining the nucleic acid positional variants.

In one embodiment, a method is provided for in silico selection of antibody sequences based on a 3D structure or structure ensemble of a lead antibody, or a structure emsemble of multiple antibodies, hereinafter collectively referred to as the lead structural template. A lead sequence or sequence profile from a specific region of the lead antibody may be employed to search databases of protein sequences for remote homologues of the lead sequence having low sequence identity and yet structurally similar. These remote homologues form a hit library. An amino acid positional variant profile (AA-PVP) of the hit library may be built based on frequency of amino acid variant appearing at each position of the lead sequence. Based on the AA-PVP, a hit variant library may be constructed by combinatorially combining the amino acid variant at each position of the lead sequence with or without cutoff of low frequency variants. The sequences in the hit variant library may be subjected to evaluation for their structural compatibility with the lead structural template. Sequences in the hit library that may be structurally compatible with the lead structural template are selected and screened experimentally in vitro or in vivo for antibody mutants with improved or desired function(s).

In another embodiment, the method comprises the steps of:

providing an amino acid sequence of the variable region of the heavy chain (V_H) or light chain (V_L) of a lead antibody, the lead antibody

having a known three dimensional structure which is defined as a lead structural template;

identifying the amino acid sequences in the CDRs of the lead antibody;

selecting one of the CDRs in the V_H or V_L region of the lead antibody;

providing an amino acid sequence that comprises at least 3 consecutive amino acid residues in the selected CDR, the selected amino acid sequence being a lead sequence;

comparing the lead sequence with a plurality of tester protein sequences;

selecting from the plurality of tester protein sequences at least two peptide segments that have at least 10% sequence identity with lead sequence, the selected peptide segments forming a hit library;

building an amino acid positional variant profile of the hit library based on frequency of amino acid variant appearing at each position of the lead sequence;

combining the amino acid variants in the hit library to produce a combination of hit variants which form a hit variant library;

determining if a member of the hit variant library is structurally compatible with the lead structural template using a scoring function; and

selecting the members of the hit variant library that score equal to or better than the lead sequence.

According to the method, the step of combining the amino acid variants in the hit library may include:

selecting the amino acid variants with frequency of appearance higher than 4 times, preferably 6 times, more preferably 8 times, and most preferably 10 times (2% to 10% and preferably 5% of the frequency for the cutoff and then include some of the amino acids from the lead sequence if they are missed after cutoff); and

combining the selected amino acid variants in the hit library to produce a combination of hit variants which form a hit variant library.

According to the method, the scoring function may be an energy scoring function selected from the group consisting of electrostatic interactions, van der Waals interactions, electrostatic solvation energy,

solvent-accessible surface solvation energy, and conformational entropy.

Optionally, the scoring function may be one incorporating a forcefield selected from the group consisting of the Amber forcefield, Charmm forcefield, the Discover cvff forcefields, the ECEPP forcefields, the GROMOS forcefields, the OPLS forcefields, the MMFF94 forcefield, the Tripos forcefield, the MM3 forcefield, the Dreiding forcefield,and UNRES forcefield,and other knowledge-based statistical forcefield (mean field) and structure-based thermodynamic potential functions.

The method may further comprise the step of:

constructing a nucleic acid library comprising DNA segments encoding the amino acid sequences of the selected members of the hit variant library.

Optionally, the method may further comprise the steps of:

parsing the selected members of hit variant library into at least two sub-hit variant libraries;

selecting a sub-hit variant library;

building an amino acid positional variant profile of the selected sub-hit variant library;

converting the amino acid positional variant profile of the selected sub-hit variant library into a nucleic acid positional variant profile by back-translating the amino acid positional variants into their corresponding trinucleotide codons; and

constructing a degenerate nucleic acid library of DNA segments by combinatorially combining the nucleic acid positional variants.

The step of parsing the hit variant library may include:

randomly selecting 10-30 members of the hit variant library that score equal to or better than the lead sequence, the selected members forming a sub-variant library.

Optionally, the step of parsing the hit variant library may include:

building an amino acid positional variant profile of the hit variant library, resulting a hit variant profile;

parsing the hit variant profile into segments of sub-variant profile based on the contact maps of the Cα, or Cβ or heavy atoms of the structure or structure ensembles of a lead sequence within certain

distance cutoff (8A to 4.5 A). A structural model or lead structural template within a distance of 4.5 Å, preferably within 5 Å, more preferably within 6 Å, and most preferably within 8 Å.

In yet another embodiment, the method comprises the steps of:

providing an amino acid sequence of the variable region of the heavy chain (V_H) or light chain (V_L) of a lead antibody, the lead antibody having a known three dimensional structure;

providing 3D structures of one or more antibodies with different sequences in V_H or V_L region than that of the lead antibody;

forming a structure ensemble by combining the structures of the lead antibody and the one or more antibodies; the structure ensemble being defined as a lead structural template;

identifying the amino acid sequences in the CDRs of the lead antibody;

selecting one of the CDRs in the V_H or V_L region of the lead antibody;

providing an amino acid sequence that comprises at least 3 consecutive amino acid residues in the selected CDR, the selected amino acid sequence being a lead sequence;

comparing the lead sequence with a plurality of tester protein sequences;

selecting from the plurality of tester protein sequences at least two peptide segments that have at least 10% sequence identity with lead sequence, the selected peptide segments forming a hit library;

building an amino acid positional variant profile of the hit library based on frequency of amino acid variant appearing at each position of the lead sequence;

combining the amino acid variants in the hit library to produce a combination of hit variants which form a hit variant library;

determining if a member of the hit variant library is structurally compatible with the lead structural template using a scoring function; and

selecting the members of the hit variant library that score equal to or better than the lead sequence.

[Route VII. Claim the sequential steps by using a lead sequence from sequence to structure to functional space shown in **Figure 2B**]

In a particular embodiment, the method comprises the steps of:

a) providing an amino acid sequence of the variable region of the heavy chain (V_H) or light chain (V_L) of a lead antibody, the lead antibody having a known three dimensional structure;

b) identifying the amino acid sequences in the CDRs of the lead antibody;

c) selecting one of the CDRs in the V_H or V_L region of the lead antibody;

d) providing an amino acid sequence that comprises at least 3 consecutive amino acid residues in the selected CDR, the selected amino acid sequence being defined as a lead sequence;

e) comparing the lead sequence with a plurality of tester protein sequences;

f) selecting from the plurality of tester protein sequences at least two peptide segments that have at least 10% sequence identity with lead sequence, the selected peptide segments forming a hit library;

g) building an amino acid positional variant profile of the hit library based on frequency of amino acid variant appearing at each position of the lead sequence;

h) combining the amino acid variants in the hit library to produce a combination of hit variants which form a hit variant library;

i) determining if a member of the hit variant library is structurally compatible with the lead structural template using a scoring function;

j) selecting the members of the hit variant library that score equal to or better than the lead sequence;

k) constructing a degenerate nucleic acid library comprising DNA segments encoding the amino acid sequences of the selected members of the hit variant library;

l) determining the diversity of the nucleic acid library, if the diversity is higher than $1x10^6$, repeating steps j) through l) until the diversity of the diversity of the nucleic acid library is equal to or lower than $1x10^6$;

m) introducing the DNA segments in the degenerate nucleic acid library into cells of a host organism;

n) expressing the DNA segments in the host cells such that recombinant antibodies containing the amino acid sequences of the hit library are produced in the cells of the host organism;

o) selecting the recombinant antibody that binds to a target antigen with affinity higher than 10^6 M^{-1}; and

p) repeating steps e) through o) if no recombinant antibody is found to bind to the target antigen with affinity higher than 10^6 M^{-1}.

In another particular embodiment, the method comprises the steps of:

a) providing an amino acid sequence of the variable region of the heavy chain (V_H) or light chain (V_L) of a lead antibody, the lead antibody having a known three dimensional structure which is defined as a lead structural template;

b) identifying the amino acid sequences in the CDRs of the lead antibody;

c) selecting one of the CDRs in the V_H or V_L region of the lead antibody;

d) providing an amino acid sequence that comprises at least 3 consecutive amino acid residues in the selected CDR, the selected amino acid sequence being defined as a lead sequence;

e) mutating the lead sequence by substituting one or more of the amino acid residues of the lead sequence with one or more different amino acid residues, resulting in a lead sequence mutant library;

f) determining if a member of the lead sequence mutant library is structurally compatible with the lead structural template using a first scoring function;

g) selecting the lead sequence mutants that score equal to or better than the lead sequence;

h) comparing the lead sequence with a plurality of tester protein sequences;

i) selecting from the plurality of tester protein sequences at least two peptide segments that have at least 10% sequence identity with lead sequence, the selected peptide segments forming a hit library;

j) building an amino acid positional variant profile of the hit library based on frequency of amino acid variant appearing at each position of the lead sequence;

k) combining the amino acid variants in the hit library to produce a combination of hit variants;

l) combining the selected lead sequence mutants with the combination of hit variants to produce a hit variant library;

m) determining if a member of the hit variant library is structurally compatible with the lead structural template using a second scoring function;

n) selecting the members of the hit variant library that score equal to or better than the lead sequence;

o) constructing a degenerate nucleic acid library comprising DNA segments encoding the amino acid sequences of the selected members of the hit variant library;

p) determining the diversity of the nucleic acid library, and if the diversity is higher than $1x10^6$, repeating steps n) through p) until the diversity of the diversity of the nucleic acid library is equal to or lower than $1x10^6$;

q) introducing the DNA segments in the degenerate nucleic acid library into cells of a host organism;

r) expressing the DNA segments in the host cells such that recombinant antibodies containing the amino acid sequences of the hit library are produced in the cells of the host organism;

s) selecting the recombinant antibody that binds to a target antigen with affinity higher than 10^6 M^{-1}; and

t) repeating steps e) through s) if no recombinant antibody is found to bind to the target antigen with affinity higher than 10^6 M^{-1}.

According to a third aspect, a computer-implemented method is provided for constructing a library of mutant antibodies based on a lead antibody. The method comprises the steps of:

taking as an input an amino acid sequence that comprises at least 3 consecutive amino acid residues in a CDR region of the lead antibody, the amino acid sequence being a lead sequence;

employing a computer executable logic to compare the lead sequence with a plurality of tester protein sequences;

selecting from the plurality of tester protein sequences at least two peptide segments that have at least 15% sequence identity with lead sequence; and

generating as an output the selected peptide segments which form a hit library.

According to any of the above methods, the length of the lead sequence is preferably between 5-100 aa, more preferably between 6-80 aa, and most preferably between 8-50 aa.

According to any of the above methods, the step of identifying the amino sequences in the CDRs may be carried out by using Kabat criteria or Chothia criteria.

Also according to any of the above methods, the lead sequence may comprise an amino acid sequence from a particular region within the V_H or V_L of the lead antibody, CDR1, CDR2 or CDR3, or from a combination of the CDR and FRs, such as CDR1-FR2, FR2-CDR2-FR3, and the full-length V_H or V_L sequence. The lead sequence preferably comprises at least 6 consecutive amino acid residues in the selected CDR, more preferably at least 7 consecutive amino acid residues in the selected CDR, and most preferably all of the amino acid residues in the selected CDR.

Also according to any of the above methods, the lead sequence may further comprise at least one of the amino acid residues immediately adjacent to the selected CDR.

Also according to any of the above methods, the lead sequence may further comprise at least one of the FRs flanking the selected CDR.

Also according to any of the above methods, the lead sequence may further comprise one or more CDRs or FRs adjacent the C-terminus or N-terminus of the selected CDR.

Also according to any of the above methods, the lead structural template may be a 3D structure of a fully assembled lead antibody, or a heavy chain or light chain variable region of the lead antibody (e.g., CDR, FR and a combination thereof).

Also according to any of the above methods, the plurality of tester protein sequences includes preferably antibody sequences, more preferably human antibody sequences, and most preferably human

germline antibody sequences (V-database), especially for the framework regions.

Also according to any of the above methods, the plurality of tester protein sequences may be retrieved from genbank of the NIH or Swiss-Prot database or the Kabat database for CDRs of antibodies.

Also according to any of the above methods, the step of comparing the lead sequence with the plurality of tester protein sequences may be implemented by an algorithm selected from the group consisting of BLAST, PSI-BLAST, profile HMM, and COBLATH.

Also according to any of the above methods, the sequence identity of the selected peptide segments in the hit library with the lead sequence is preferably at least 25%, preferably at least 35%, and most preferably at least 45%.

According to any of the above method, the method may further comprises the following steps:

introducing the DNA segments in the nucleic acid or degenerate nucleic acid library into cells of a host organism;

expressing the DNA segments in the host cells such that recombinant antibodies containing the amino acid sequences of the hit library encoded by the nucleic acid or degenerate nucleic acid library are produced in the cells of the host organism; and

selecting the recombinant antibody that binds to a target antigen with affinity higher than 10^6 M^{-1}, preferably 10^7 M^{-1}, more preferably 10^8 M^{-1}, and most preferably 10^9 M^{-1}.

The recombinant antibodies may be fully assembled antibodies, Fab fragments, Fv fragments, or single chain antibodies.

The host organism may include any organism or its cell line that is capable of expressing transferred foreign genetic sequence, including but not limited to bacteria, yeast, plant, insert, and mammals.

The recombinant antibodies may be fully assembled antibodies, Fab fragments, Fv fragments, or single chain antibodies. For example, the recombinant antibodies may be expressed in bacterial cells and displayed on the surface of phage particles. The recombinant antibodies displayed on phage particles may be a double-chain heterodimer formed between V_H and V_L. The heterodimerization of V_H and V_L chains may be facilitated by a heterodimer formed between two

non-antibody polypeptide chains fused to the V_H and V_L chains, respectively. For example, these two non-antibody polypeptide may be derived from a heterodimeric receptors $GABA_B$ R1 (GR1) and R2 (GR2), respectively.

Alternatively, the recombinant antibodies displayed on phage particles may be a single-chain antibody containing V_H and V_L linked by a peptide linker. The display of the single chain antibody on the surface of phage particles may be facilitated by a heterodimer formed between a fusion of the single chain antibody with GR1 and a fusion of phage pIII capsid protein with GR2.

The target antigen to be screened against includes small molecules and macromolecules such as proteins, peptides, nucleic acids and polycarbohydrates.

According to a fourth aspect, there is provided a computer-readable medium comprising logic for constructing a library of mutant antibodies based on a lead antibody, the logic comprising:

logic which

takes as an input an amino acid sequence that comprises at least 3 consecutive amino acid residues in a CDR of the lead antibody, the amino acid sequence being a lead sequence;

compares the lead sequence with a plurality of tester protein sequences;

selects from the plurality of tester protein sequences at least two peptide segments that have at least 15% sequence identity with lead sequence; and

generates as an output the selected peptide segments which form

a hit library.

Disclosed herein, monoclonal antibodies are provided that are capable of binding to human vascular endothelial growth factor (VEGF) with a binding affinity higher than 10^6 M^{-1}. The monoclonal antibody may be a fully assembled antibody, a Fab fragment, a Fv fragment or a single chain antibody (scFv).

In one embodiment, the heavy chain CDR3 of the monoclonal antibody comprises an amino acid sequence selected from group consisting of SEQ ID Nos: 36-48 and 63-125.

In another embodiment, the heavy chain CDR1 of the monoclonal antibody comprises an amino acid sequence selected from group consisting of SEQ ID Nos: 19-30.

In yet another embodiment, the heavy chain CDR2 of the monoclonal antibody comprises an amino acid sequence selected from group consisting of SEQ ID Nos: 31-35.

Optionally, the heavy chain CDR3 of the monoclonal antibody comprises an amino acid sequence selected from group consisting of SEQ ID Nos: 36-48 and 63-125, and the heavy chain CDR1 of the monoclonal antibody comprises an amino acid sequence selected from group consisting of SEQ ID Nos: 19-30.

Also optionally, the heavy chain CDR3 of the monoclonal antibody comprises an amino acid sequence selected from group consisting of SEQ ID Nos: 36-48 and 63-125, and the heavy chain CDR2 of the monoclonal antibody comprises an amino acid sequence selected from group consisting of SEQ ID Nos: 31-35.

Also optionally, the heavy chain CDR1 of the monoclonal antibody comprises an amino acid sequence selected from group consisting of SEQ ID Nos: 19-30, and the heavy chain CDR2 of the monoclonal antibody comprises an amino acid sequence selected from group consisting of SEQ ID Nos: 31-35.

In another embodiment, the heavy chain variable region (V_H) of the monoclonal antibody against VEGF comprises an amino acid sequence of SEQ ID No: 126, and the light chain variable region (V_L) of the monoclonal antibody against VEGF comprises an amino acid sequence of SEQ ID No: 127.

In yet another embodiment, the heavy chain variable region (V_H) of the monoclonal antibody against VEGF comprises an amino acid sequence selected from group consisting of SEQ ID Nos: 126, 128, 129, 130, and 131, and the light chain variable region (V_L) of the monoclonal antibody against VEGF comprises an amino acid sequence of SEQ ID No: 127.

The antibodies designed by using the methods disclosed

herein may be used for diagnosing or therapeutic treatment of various diseases, including but not limited to, cancer, autoimmune diseases such as multiple sclerosis, rheumatoid arthritis, systemic lupus erythematosus, Type I diabetes, and myasthenia gravis, graft-versus-host disease, cardiovascular diseases, viral infection such as HIV, hepatitis viruses, and herpes simplex virus, bacterial infection, allergy, Type II diabetes, hematological disorders such as anemia. The antibodies may also be used as conjugates that are linked with diagnostic or therapeutic moieties, or in combination with chemotherapeutic or biological agents. The antibodies may also be formulated for delivery via a wide variety of routes of administration. For example, the antibodies may be administered or coadministered orally, topically, parenterally, intraperitoneally, intravenously, intraarterially, transdermally, sublingually, intramuscularly, rectally, transbuccally, intranasally, via inhalation, vaginally, intraoccularly, via local delivery (for example by a catheter or a stent), subcutaneously, intraadiposally, intraarticularly, or intrathecally.

According to any of the above embodiments, the designed proteins (e.g.. antibodies) may be synthesized, or expressed in cells of any organism, including but not limited to bacteria, yeast, plant, insect, and mammal. Particular types of cells may include, but are not limited to, *Drosophila melanogaster* cells, *Saccharomyces cerevisiae* and other yeasts, E. coli, Bacillus subtilis, SF9 cells, C129 cells, 293 cells, Neurospora, BHK, CHO, COS, and HeLa cells, fibroblasts, Schwanoma cell lines, immortalized mammalian myeloid and lymphoid cell lines, Jurkat cells, mast cells and other endocrine and exocrine cells, and neuronal cells. Examples of mammalian cells may include, but are not limited to, tumor cells of all types (particularly melanoma, myeloid leukemia, carcinomas of the lung, breast, ovaries, colon, kidney, prostate, pancreas and testes), cardiomyocytes, endothelial cells, epithelial cells, lymphocytes (T-cell and B cell), mast cells, eosinophils, vascular intimal cells, hepatocytes, leukocytes including mononuclear leukocytes, stem cells such as haemopoetic, neural, skin, lung, kidney, liver and myocyte stem cells, osteoclasts, chondrocytes and other connective tissue cells, keratinocytes, melanocytes, liver cells, kidney cells, and adipocytes.

Preferably, the designed protein is purified or isolated after expression according to methods known to those skilled in the art. Examples of purification methods may include electrophoretic, molecular, immunological and chromatographic techniques, including ion exchange, hydrophobic, affinity, and reverse-phase HPLC chromatography, and chromatofocusing. The degree of purification necessary may vary depending on the use of the designed protein. In some instances no purification may be necessary.

Also according to any of the embodiments described above, the designed proteins may be screened for a desired function, preferably a biological function such as their binding to a known binding partner, physiological activity, stability profile (pH, thermal, buffer conditions), substrate specificity, immunogenicity, toxicity, etc.

In the screening using a cell-based assay, the designed protein may be selected based on an altered phenotype of the cell, preferably in some detectable and/or measurable way. Examples of phenotypic changes may include, but are not limited to, gross physical changes such as changes in cell morphology, cell growth, cell viability, adhesion to substrates or other cells, and cellular density; changes in the expression of one or more RNAs, proteins, lipids, hormones, cytokines, or other molecules; changes in the equilibrium state (i.e. half-life) or one or more RNAs, proteins, lipids, hormones, cytokines, or other molecules; changes in the localization of one or more RNAs, proteins, lipids, hormones, cytokines, or other molecules; changes in the bioactivity or specific activity of one or more RNAs, proteins, lipids, hormones, cytokines, receptors, or other molecules; changes in the secretion of ions, cytokines, hormones, growth factors, or other molecules; alterations in cellular membrane potentials, polarization, integrity or transport; changes in infectivity, susceptability, latency, adhesion, and uptake of viruses and bacterial pathogens.

According to any of the above embodiments, the designed proteins (e.g.. antibodies) may be synthesized, or expressed as fusion proteins with a tag protein or peptide. The tag protein or peptide may be used to identify, isolate, signal, stabilize, increase flexibility of, increase degradation of, increase secretion, translocation or

intracellular retention of or enhance expression of the designed proteins.

BRIEF DESCRIPTION OF THE FIGURES

Figures 1A-D illustrate four embodiments of the method that can be used in the present invention to select for proteins with desired functions. Lead in **Figures 1A-D** can be either the lead sequence or sequence profile from multiple structure-based alignment. The hit library, hit variant library I and II are defined in the definition section.

Figures 1E-H illustrate four of the possible embodiments of the method that can be used in the present invention to select for proteins with desired functions. Here, the lead refers to a structure or structure model or structure ensemble or profile (multiple superimposed structures), the corresponding sequence or sequence profile from the lead structure or structure ensemble can be then used to screen all possible sequences or random combinations for the hit sequence library based on structure-based screening. The resulting hit variant libraries can be used for direct experimental screening or compared with the sequence hit profile derived from the corresponding lead sequence or sequence profile (see **Figures 2A-C**). The structure template referes to structure, structure ensemble (more than 2 structures) from experimental determination and/or modeling.

Figure 2A is a schematic overview of the in silico protein evolution system provided by the present invention. The triangular relationship among sequence, structure and function spaces is shown to illustrate potential paths traversing from the lead structure/lead structural profile or lead sequence/lead sequence profile to candidate sequences through sequence, structure and function spaces.

In sequence space, the lead sequence(s) or profile is used to search the specific database for evolutionarily related sequences. Sequence profile based on the structural alignment of the lead structure can be used to search for remote homologues of the lead sequence. The variant profile of the hit library describes the positional frequency and entropy of the amino acid sequence. The variant profile can be filtered and re-profiled at a given cutoff to give the evolutionally preferred variant profile. This procedure can be iterated with various searching methods on related sequence database.

In structure space, an in silico variant profile is generated using a structure-based screening of random or evolutionally pooled sequence library. The variant profile can be filtered and refined to give the structurally preferred variant profile. This procedure can be iterated and refined with better scoring functions and representative structure ensemble.

The variant profile generated using either evolutionally- or structurally-based approaches can be used in sequential (2B: from sequence to structure to function space; 2C: from structure to sequence to function space) or parallel fashion (from sequence space to function space and from structure space to function space) to give an overall variant profile or library of amino acids. The resulting variant library of amino acids is back-translated into nucleic acid library by using preferred or optimized codons. This procedure can be iterated with different filtering and partitioning procedure to adjust the library size to within experimentally manageable range.

To select for functional mutants in function space, the synthesized nucleic acid library is introduced into vectors by transformation and functionally expressed or displayed, for example, on phage particles. Rounds of selection and enrichment against immobilized antigen are carried out. The whole or part of the procedure can be iterated and refined until the desired candidates are selected experimentally.

Figure 2B. A schematic diagram of an embodiment of the methodology provided in the present invention for antibody library design. A sequential procedure moves from **sequence** first to **structure** and to **function** space. The design starts from a lead sequence or sequence profile (multiple aligned sequences from structure-based alignment). A hit library is generated by searching the sequence database. The hit profile given by the hit library at certain cutoff will give the hit variant library. Either the hit library or hit variant libraries can be screened computationally using the lead structure or structure ensemble as the template structure. The resulting sequence library is ranked based on their compatibility with the template structure or structure ensemble. Sequences with scores better than or equal to the

lead sequence are selected and profiled to generate nucleic acid (NA) library. The in silico NA library size is evaluated and passed on to oligonucleotide synthesis if the library size is acceptable. Otherwise, the hit variant library is repartitioned into smaller segments and smaller NA libraries are generated. In the function space, the nucleic acid library is experimentally screened and positive sequences are fed back into the computational cycle for library refinement. Strong positive clones are passed on for further evaluation and potential therapeutic development. If no hits occur in the experimental screening, the lead or its new lead profile is selected for the target system and the process is reiterated.

Figure 2C. A schematic diagram of another embodiment of the methodology provided in the present invention for antibody library design. An alternative sequential procedure moves from the **structure** first to **sequence** and to **function** space. The design starts from a lead structure or structure ensemble. A combination of random mutations at target positions is screened computationally for their compatibility with the structure template. A variant profile of the sequences that score better than or equal to the lead sequence is generated. This variant profile can be compared and/or combined with those given by searching the sequence database. Novel mutants might be included or excluded based on the consensus frequency shown in sequence and structure space to generate a nucleic acid library. The rest of the procedure is similar to those described in Figure 2B. This approach emphasizes the importance of finding novel mutants by structure-based computational screening without relying on the evolutionary sequence information. The sequence profile from searching database will help to assess the variant profile obtained from computational screening that lies on the accuracy of the scoring function as well as on the sampling algorithm used.

Figure 3 illustrates a process for constructing a hit library in silico via database search using either the single lead or the lead profile based on structural alignment. The search results are sorted and redundant sequences (even if the background is different) are removed to produce a list of unique sequences in the hit library. Impact of the

lead sequence/sequence profile, sequence searching methods, and various database are shown in Figure 4-6.

Figure 4 illustrates a process for constructing a hit variant library I based on the variant profile from the hit library that is used to analyze the evolutionary positional preferences for amino acids. A refined variant profile is derived by filtering based on selection criteria that include frequency, variation entropy, and energy score of the amino acid variants at each position. The hit variant library II is combinatorially enumerated from the refined variant profile.

Figure 5 illustrates a process for structural evaluation and selection of a hit variant library I or II to create a structurally screened version of hit variant library II. The computational selection uses simple as well as custom energy function to score and rank the hit variant library I or II sequences applied to a lead structural template. For each sequence, the side chains are generated using a backbone-dependent rotamer library and the side chains and backbone are energy minimized against the template background to relieve any local strain. The fitness of the hit variant library I or II in the template structure is scored and ranked using simple as well as custom energy functions. Several ensembles of the "best" sequences are selected to build a new hit variant library II for translation into a nucleic acid (NA) library. The selection criteria may include sequence clustering, structural considerations or functional considerations. The ensembles of amino acid sequences are re-profiled for generating the nucleic acid library within experimentally manageable limit (Figure 6).

Figure 6 illustrates a process for constructing a nucleic acid (NA) library by back-translation from hit variant library II. The back translation of amino acids into nucleic acids is intended to keep the size of the nucleic acid library within experimentally manageable limit while optimizing the prefered codon usage. The size of the nucleic acid library is calculated and kept within the experimental limit or the hit variant profile is modified by reducing the variant number or partitioned into shorter segments. Partitioning may be accomplished either by using

structurally correlated segments or series of overlapping sequentially correlated segments.

Figure 7 is an overview of a strategy of sampling a library at several regions of the fitness landscape. The fitness landscape of the selected peptide sequences can be expanded to cover a larger fitness landscape if the combinatorial amino acid or its degenerate nucleic acid libraries can be designed to sample a larger function space. Strategic sampling from a designed library leads to overlapping and expanded diversity that can include significant evolutionary jumps in the fitness landscape of the function space.

Figure 8 shows modular elements of a typical library plasmid for antibody engineering. The libraries of framework and CDR sequences can be designed, respectively or combinatorially in iteration. FR= framework region. CDR=complementarity determining region. RE=restriction enzyme site.

Figure 9A is a sequence comparison between the parental and matured anti-VEGF antibody in V_H CDRs. "c" indicates where atoms of the antigen-antibody complex contact within 4.5 A in the X-ray structure. Bold letters highlight the differences in amino acids between the parental and matured antibody in V_H CDRs (CDR1 and CDR3). The numbering for V_H CDRs follows the convention by kabat and a sequential scheme (100, 101 rather than 100, 100a etc).

Figure 9B is a sequence comparison between the parental and matured anti-VEGF antibody in V_H CDR3 with its adjacent regions. The sequence (SEQ ID NO: 5) from parental antibody is the lead sequence used for searching database. The numbering for V_H CDRs are both Kabat and a sequencetial scheme used here also.

Figure 10A is a plot showing the distribution of the frequency of a hit library versus their sequence identity (in %) relative to the lead sequence of V_H CDR3 of parental anti-VEGF antibody. The lead sequence is shown in Figure 9B and the profile HMM (HAMMER2.1.1)

was used to search the Kabat database (Johnson, G and Wu, TT (2001) Nucleic Acids Research, 29, 205-206).

Figure 10B illustrates the phylogenetic tree of the sequences of a hit library shown in Figure 10A in order to show the phylogenetic diversity of the hit library resulting from the database search in Figure 10A.

Figure 11 shows a variant profile for the 107 sequences of the hit library generated based on the lead sequence of V_H CDR3 of parental anti-VEGF antibody. The upper portion shows a table listing the amino acid frequency of 20 amino acids at each position of the lead sequence. The variant profile at the bottom shows the amino acid positional diversity. A complete enumeration of a combinatorial library with no selective control of amino acid diversity (shown in lower left portion of the figure) will require a library size on the order of 10^{19}. The lower right portion of the figure shows a filtered variant profile obtained by using a cutoff frequency of 10. All positional amino acids occurring 10 or less times among the 107 members of the hit list are filtered. This filtered variant profile can be further screened computationally to reflect the ranking order of the structural compatibility if only the antibody structure is used, or binding affinity with the antigen if the complex structure between antibody and antigen is used. The variant profile shows no correlation with the contact sites between antigen and antibody as indicated in Figure 9A.

Figures 12A and 12B show a typical plot of the scores of an anti-VEGF antibody variant library in the parental(1bj1) and matured(1cz8) antibody structure, respectively, in the absence **(A)** and presence of VEGF antigen **(B)**, using a scoring function of the total energy of the Amber94 forcefield implemented in CONGEN. The scores of the matured (M) and parental (P) sequences are marked by the arrows. The mature sequence scores better than that of the parental sequence in the absence and presence of the antigen in both template structures. **Figure 12C** shows the correlation between the scores of the variant library in the presence and absence of the antigen.

Figures 12D and E show that the simple scoring function used here is also in general correlated with a refined scoring function for the hit library (Figures 10 & 11) using the template structure of the matured antibody (1cz8), although some scattering in the correlation plot suggest that some terms involving the solvation etc should be added into the simple scoring function to improve the correlation.

Figure 13A shows how the present inventive methods can select the top ten sequences from a computational screening of an anti-VEGF V_H CDR3 hit variant library for experimental screening, to demonstrate that diverse, functional sequences, different from the parental or matured ones, can be selected. The amino acid variant profile and the corresponding variant library in the degenerate nucleic acids are listed. An energy diagram at the upper right portion of the figure shows from left to right the energy distribution of the 10 selected sequences from computational screening, their variant amino acid combinatorial library, nucleic acid combinatorial library and positive clones selected from experimental screening in vitro. The sequence library that corresponds to each of sequence pools shown in the energy diagram is indicated with arrows. **Figure 13B & C** show the top 10 sequences from computational screening of the variant libraries for V_H CDR1 and CDR2, respectively, the amino acid variant profile and corresponding variant library in degenerate nuclei acids for V_H CDR1 and CDR2 libraries of anti-VEGF antibodies.

Figure 14A shows UV reading of the ELISA positive clones identified in round 1 and round 3 selections of functional anti-VEGF ccFv antibodies with V_H CDR3 encoded by the designed nucleic acid library (Figure 13A). The bottom numbers indicate the column numbers in a 96-well (8x12) ELISA plate. Different bar shadings indicated different rows.

Figure 14B shows V_H CDR3 sequences of the positive clones from round 1 and 3 selection via phage display of the nucleic acid library shown in Figure 13A. It is clear that many diverse sequences are selected with large variations at several positions that are different

from V_H CDR3 of parental and matured anti-VEGF antibody (Figure 9A & B).

Figure 14C illustrates a phylogenetic tree of the positive clones showing the diversity of the screened sequences. The sequence identities of the selected positive clones from V_H CDR3 shown in Figures 14A & B ranged from 57 to 73 percent relative to the parental V_H CDR3 sequence, with N-terminal CAK and C-terminal WG residues included (see Figure 9B).

Figures 15A-B are pie charts showing the breakdown of the origins of the screened sequences in the first and third rounds into three groups: designed amino acid sequences, combinatorial amino acid sequences from the designed sequences, and the novel combinatorial amino acid sequences encoded by the synthesized degenerate nucleic acid library. **A**: V_H CDR3 clones from the first round screening in vitro with distribution of experimentally selected sequences from positive clones in 3 libraries. **B**: V_H CDR3 clones from the third round screening in vitro with distribution of experimentally selected sequences from positive clones in 3 libraries. Because only limited number of positive clones from each round are selected for sequence analysis, the figures are only used to illustrate rough percentages of the selected sequences from designed, its combinatorial amino acid and nucleic acid libraries.

Figure 16A is a table that lists the experimentally selected amino acids sequences from V_H CDR1, CDR2 and CDR3 libraries of degenerate nucleic acids shown in Figures 13A-C. **Figure 16B** shows the distribution of the sequence identities of selected sequences from V_H CDR1, CDR2 and CDR3 libraries relative to the corresponding parental sequence of anti-VEGF V_H CDR1, 2, and 3 respectively. It is clear that functional, diverse sequences different from the corresponding parental sequences can be selected experimentally.

Figure 17A shows the schematic relationship among 4 different libraries (designed amino acid sequences, the combinatorial library of amino acid variant of the designed sequences, and combinatorial

degenerate nucleic acid libraries encoding the unique amino acid sequences and the entire degenerate nucleic acid library) and the distribution of the experimentally selected positive clones shown in X. The innermost (striped) circle represents the designed amino acid sequence library selected, for example, based on energy scores of the hit variant library. The shaded circle represents combinatorial amino acid library of the selected sequences from computational screening of a hit variant library. The third (stippled) circle represents the combinatorial amino acid library encoding the unique combinatorial amino acid library. The outermost circle represents the degenerate nucleic acid library for all amino acid sequences derived from the back-translation of the amino acid library. The relative size of the outermost versus the third (stippled circle) depends on the efficiency of the back-translation procedure from amino acids to nucleic acid sequences with consideration for other factors such as the codon usage. "X" indicates experimentally selected sequences. For example, anti-VEGF V_H CDR3 library from round 3 is shown here (see table in Figure 17B). The distribution among different libraries depends on selection conditions, the effectiveness of library design, the relative size of the selected clones versus library or number of sequenced clones etc.

Figure 17B shows a table delineating the relationships among the four libraries (Figure 17A) and the distribution of the experimentally selected sequences of the positive clones for anti-VEGF V_H CDR1, 2, and 3 libraries. The "AA_Seq/Comb" column indicates the number of selected amino acid sequences by computational screening (designed library I) and the number of recombinant sequences of the selected sequences (variant library II). The "NN_seqs/peptide seq" column indicates the number of nucleic acid sequences of the degenerate nucleic acid library, and the unique amino acid sequences encoded by the degenerate nucleic acid library. The "exp_seq" column shows the number of the experimentally selected, unique sequences from positive clones. The "distribution of the selected sequences" column indicates the numbers of unique sequences from designed amino acid sequences, their combinatorial library of amino acid variants and the combinatorial library of the degenerate nucleic acids encoding unique peptide

sequences.

Figure 18 shows the evolution of the sequence fitness scores for anti-VEGF V_H CDR3 libraries at various stages in the procedure, starting from left to right: a lead sequence, hit library, hit variant library I, selected sequences from computational screening (shaded band), the combinatorial library of selected sequences (hit variant library II), combinatorial nucleic acid library encoding the combinatorial amino acid sequences, and experimentally selected sequences. A lead sequence was used to identify evolutionary hit library from a database of sequences. An in silico combinatorial library was designed based on the diversity of the hit library. A subset of the computationally screened sequences with scores better than the lead was used to generate a combinatorial amino acid library. A degenerate nucleic acid library coding the combinatorial amino acid library was generated using degenerate nucleic acid synthesis strategy to expand the diversity. Experimental screening of the library led to sequences with potentially improved function.

Figure 19A shows the lead profile generated from structure-based mutiple seqeuce alignment. The structural motif of the lead sequence is used to search protein structure database (PDB databank) for similar structures within certain distance cutoff. The five structures are superimposed using C_α atoms of the V_H CDR3. The average root mean square deviation (RMSD) between each structure and V_H CDR3 structural motif (colored in magenta) is about 2 Å. The corresponding mutiple sequence alignment is shown to the right, together with their PDB IDs and corresponding colors.

Figure 19B shows a variant profile for the 251 unique sequences of the hit library generated based on the lead sequence profile of V_H CDR3 of parental anti-VEGF antibody. The upper portion shows a table listing the amino acid frequency of 20 amino acids at each position of the lead sequence. The lower portion of the figure shows a filtered variant profile obtained by using a 5% cutoff of the frequency or 12 in

this case. All positional amino acids occurring 12 or less times among the 251 members of the hit list are removed. This filtered variant profile can be further screened computationally using the structure ensembles.

Figure 19C shows the distribution of the sequences from the hit library relative to the parental V_H CDR3 sequence (Figure 9B). The circles indicate that the sequence identity up to 36% can be identified using the single parental sequence for HMM search. The triangles indicate that even lower sequence identity up to ~20% can be found using the lead sequence profile from a structure-based multiple sequence alignment. The sequence searching strategy used here can find diverse hits with remote homology (as low as 20%) to the lead sequence.

Figure 19D shows the general strategy in generating a focused library that lies within the intersection of the sequence, structure and function spaces. As shown in Figure 19A-C, the diversity of the hit sequences is increased by using a structure-based mutiple alignment. It is possible to expand the diversity in both sequence and structure spaces, good hits can be identified in the intersection of all three spaces.

Figure 20 is a schematic representation depicting various antigen-binding unit (Abu) configurations. Note two novel display systems employed in the current inventive methods: ccFv system, heterodimeric coiled-coil stabilized Fv with a disulfide bond between GR1 and GR2, and GMCT system, adapter-mediated scFv display system.

Figure 21 depicts the nucleotide and amino acid sequences of $GABA_b$ receptor 1 and 2 that were used in constructing the subject ccFv Abu. The coiled-coil sequences are derived from human $GABA_b$-R1 and $GABA_b$-R2 receptors. The coding amino acid sequences from $GABA_b$

receptors are written as bold letters. A flexible GlyGlyGlyGly spacer was added to the amino-terminus of R1 and R2 heterodimerization sequences to favor the functional Fv heterodimer formation. To further stabilize the heterodimer, we introduced a ValGlyGlyCys spacer to lock the heterodimeric coiled-coil pair by a disulfide bond. The additional SerArg coding sequences at N-terminus of GGGG spacer provides XbaI or XhoI sites for the fusion of the GR1 and GR2 domains to the carboxy-terminus of V_H and V_L fragment, respectively.

Figures 22 A-B depict the nucleotide and amino acid sequences of V_H and V_L of anti-VEGF ccFv antibody AM2, respectively.

Figure 23A is a schematic representation of the phagemid vectors pABMD12.

Figure 23B depicts the sequence of pABMD12 vector.

Figure 24 depicts a comparison of the binding capability of phage displayed AM2 ccFv and scFv to the immobilized VEGF antigen. The results demonstrate that ccFv can be assembled and displayed on phage particles.

Figure 25A depicts the results of an ELISA using AM2-ccFv phages from model library pannings. The results demonstrate the enrichment of phages displaying AM2-ccFv antibody in panning of model libraries.

Figure 25B show the PCR results from $1/10^7$ model library panning which shows that the test sequence can be selected from the model library.

Figure 26 depicts the results of ELISA using phages from library panning. The results show that the VEGF-binding phages were selected out from V_H CDR1, CDR2 libraries (see Figure 14A for V_H CDR3).

Figure 27 (same as Figure 16A) is a table listing the amino

acids sequences of experimentally selected clones encoding designed for anti-VEGF VH CDR1, CDR2 and CDR3 libraries (see Figures 13A-C).

Figure 28A show the sequence library of a composite anti-VEGF V_H CDR3 library. Because the library size is too big to be covered by one or several degenerate nucleic acid library, the variant profile is parsed into 3 segments with their variant profiles shown in Figure 28A. The segments are parsed based on the contact map of C_α atoms within 8Å shown on the right side of Figure 28A. Figure 28A also shows the ribbon diagram of the anti-VEGF V_H CDR3 as well as contact distances among C_α atoms within 8Å. The approach provide a general way to parse a large variant profile into smaller segments based on the topology of the structure. Low resolution structure or structure model can serve the purpose here because only structural constraints from topological features is required for sequence segmentation in order to capture covariants distant in primary sequence such as N- and C-termini residues close in the loop.

Figure 28B covers the N- and C-termini that might contain coupled variants (1-3). The variant profiles of both amino acid library and nucleic acid library are listed, together with the combinatorial size of the libraries and final synthesized degenerate oligonucleotides. **Figure 28C** contains segment (4) and **Figure 28D** contains another segment (5). All three segments are covered by nucleic acid libraries with sizes less than 10^6: (1-3) in figure 28B are targeted by 3 degenerate nucleic acid libraries, whereas (4) and (5) in figures 28 C-D are targeted by a separate degenerate nucleic acid library.

Figure 29 summarizes the procedures and conditions used for panning ccFv library L14 as well as the enrichment factor from each panning. L14 library is constructed in Figure28A-D by pooling together all 5 degenerate oligonucleotides shown in Figure 28B-D.

Figure 30 shows the amino acid sequences of the V_H CDR3 variants selected from panning 5 and 7 of library L14 using ccFv display

platform. Note that after panning 5, all variants are located at position 101. Only two variants, S101R and S101T, are selected after round 7.

Figure 31 shows the enrichment of HR (H97, S101R) phage from panning of library L14 for V_H CDR3. The enrichment for HR and parental antibody WT (see also Figure 9B) at round 0, 5 and 7 were highlighted.

Figure 32 shows a simple diagram of a novel Coiled-coil Domain Interaction Mediated Display (CDIM) adapter-directed display system for single chain antibody library. Transformation Infectionof expression vector pGDH1alone in E. coli bacteria permits expression and production of soluble proteins fused with GR1 in bacterial periplasmic space. Additional superinfection of the same bacteria with the UltraHelper phage vector expressing the engineered coat protein fused with GR2 and other phage proteins permits the display of antibody fragments (or other proteins) on the surface of filamentous phage following synthesis of phage particles in periplasmic space of bacteria.

Figure 33A shows the map of the GMCT-UltraHelper phage plasmid. The construct contains a nucleotide sequence encoding an additional copy of the engineered gene III fused to adaptor GR2 and myc protein tag in KO7kpn phage vector, and ribosome binding sequence-OmpA leader sequence adjacent to the wild-type gene III sequence. **Figure 33B** shows the genetically modified region of KO7Kpn to produce GMCT-UltraHelper phage at the nucleotide and amino acid sequence level.

Figure 34A & B show the protein expression vector map (A) and the complete nucleotide sequence (B) for pABMX14, which includes an ampicillin-resistance gene for antibiotic selection (Amp), a plasmid origin of replication (ColE1 ori), a f1 phage origin of replication (f1 ori), lac promoter/lac O1 controlled protein expression cassette (plac-RBS-pelB-GR1-DH), and restriction endonuclease sites are also shown. The

NcoI/XbaI or NcoI/NotI or XbaI/NotI restriction sites can be used to insert nucleotide sequence encoding proteins of interest.

Figure 35A summarizes the procedure and conditions used for panning scFv library L17, together with the enrichment factor from each round (A). The sequences of L17 library in V_H CDR3 region are exactly the same as those of L14 (see Figure 28A-D). **Figure 35B** shows the flowchart of the panning process.

Figure 36 shows the amino acid sequences of the V_H CDR3 variants selected from library L17 by off-rate panning from two parallel steps 4 and 5, respectively, using the adapter-mediated phage display system. Note in off-rate panning 4, sequences were selected with variants located at positions 97 and/or 101 (100a in Kabat nomenclature). In off-rate panning 5, sequences were selected with variants located at 101(100a) and/or 102 (100b) and/or 103 (100c). Two important mutants YS (H97Y-S101) and HT (H97-S101T or H97-S100aT) in the mature sequence were selected from panning 4 and panning 5, separately. The combination of variants at these two positions might give the mature sequence H97Y and S100aT in V_H CDR3 (Figure 9B). But this combination is deliberately avoided in the parsed segments (see Figure 28A-D). Also, note that HR (H97-S100aR) is again shown in higher frequency (3/1) than HT (H97-S100aT), the mature sequence (Figure 9B), consistent with the similar observation (7/3) in panning 7 of Figure 30.

Figure 37 shows the affinity data of 4 antibodies containing the VH CDR3 (FR123) of anti-VEGF antibody selected via ccFv display format from designer libraries using BIAcore biosensor. The measurement is done by measuring the change of SPR units (y-axis) vs time (x-axis) when a purified antibody binds its antigen (VEGF) immobilized on the CM5 biochip at 25°C. Both the on-rate and off-rate changes were determined from the data fitting using 1:1 Langmuir binding model. The X50 is in ccFv format and contain the parental sequences for VH and VL shown in Figure 22A and 22B. X63 contains

H97Y and S101T in VH CDR3 with 6.3-fold improvement in Kd (**see Figure 9B**) and the rest is the same as X50. X64 contains S101R mutant in VH CDR3 with 2.5-fold improvement relative the reference X50; the improvement comes almost exclusively from the on-rate increase. The X65 contains H97Y and S101R, showing 10-fold improvement relative to X50 using the ccFv format under the same condition, which is stronger in binding affinity than the best reported mutant combination X63 (H97Y and S101T) of the affinity-matured VH CDR3 sequence (see Chen et al supra (1999) J. Mol Biol 293, 865-881).

Figure 38A shows the framework regions FR123 of heavy chain variable regions defined based on the Kabat nomenclature, together the random libraries used for humanization reported (Baca et al. supra, 1997) for comparison. The murine anti-VEGF VH framework FR123 sequence is shown in A4.6.1 are shown in Figure 9B. The humanized antibody used as the parental and reference framework fr123 here (therein after referred to as "humanized anti-VEGF antibody") reported in the literature (see Presta et al. supra, 1997). The sequence number annotated above the FR123 sequence is based on the kabat nomenclature (kabataa) and its consecutive order including and amino acids in its CDRs. **Figure 38B** shows the variant profiles for the hit library generated using the human VH germline sequences based on the lead sequence of V_H FR123 of the murine anti-VEGF antibody. The variant profile at the bottom shows the amino acid positional diversity. The lower portion of the figure shows the filtered variant profiles obtained by using a cutoff frequency of 5 and 13, repsectively. All positional amino acids occurring 5 or less times or (13 or less) among the members of the hit list are filtered. **Figure 38B**-continuous shows that the reprofiled variant profile for the hit library generated using the human VH germline sequences based on the lead sequence of V_H FR123 of the murine anti-VEGF antibody without cutoff but the variant at each position is ranked based on its structural compatibility with the antibody structure using total energy or van der waals energy. This ranking highlights certain amino acids at low occurrence frequency are important structurally in stabilizing the scaffolding of the framework, kept for optimization. **Figure 38C** shows the variant profiles for the hit

library generated using the Kabat-derived human VH sequences based on the lead sequence of V_H FR123 of the murine anti-VEGF antibody with a filtered variant profile at a cutoff of 19. The murine VH FR123 sequence is listed as the reference above the dotted line with position annotated using consecutive number. All the variants of amino acids are listed below the dotted line. The dot in the variant represent the same amino acid as in the reference. Figure 38D shows the designer libraries using the filtered variant profile from human VH germline sequences at cutoff 5 (see Figure 38B). The sequence number annotated above the FR123 sequence is based on the kabat nomenclature (kabataa) and its consecutive order including and amino acids in its CDRs. This filtered variant profile can be further screened computationally to reflect the ranking order of the structural compatibility if only the antibody structure is used. Two amino acids, F70(F69) and L72(L71), missing from the filtered variant profile at cutoff 5 were also included because they are among the best preferred amino acids at these positons based on structure-based scoring. The final submitted library for top 100 ranked sequences from structure-based screening also include F70(F69), L72(L71), S77(S76) and K98(K94) (the number in the bracket representing sequence number based on kabat nomenclature), because some amino acids such as R is over predicted in the computation for both L72(L71) and K98(K94) as discussed previously for K94R in the VH CDR3 affinity maturation.

Figure 39A depicts the distribution of scoring diagram for VH framework fr123 hit sequences of murine anti-VEGF using the human VH germline sequences in relatively densely populated blue strips in column 1 in x-axis, together with the murine and humanized framework fr123 (see Presta et al. supra) sequence and a widely used human VH germline DP47 in the relatively sparsely populated blue strips in column 0 in x-axis, using 1bj1 (upper panel) and 1cz8 (lower panel) as the template structures in the absence (leftmost column) and presence(middle column) of the VEGF antigen. The scores of sequences in the presence and absence of antigen is correlated (in the rightmost column), indicating the antibody structure for framework optimization is sufficient for most of the framework optimization because they have

minimal contact with antigen. The scoring digrams for the combinatorial sequence libraries are not shown here.

Figure 39B depicts the ranking scoring in the left panel based on the difference between sequences in the library and the reference murine VH FR123 sequence and the phylogenetic distances in x-axis (distance connecting them (see Figure 14C also) for the reference, murine VH FR123, humanized VH FR123 reported (Presta et al., supra 1997 and Chen et al. supra 1999) and the top ranked 200 designer sequences and human VH3 germlines including a widely used VH human germline called DP47. The top 200 ranking sequences from structure-based screening of one variant profile (AA-PVP) of human germlines are clustered with the human VH3 germline family in phylogenentic analysis (red cycle), whereas the lead murine antibody framework is genetically distant in its phylogentic distance from the designed (when only human germline VH sequences at high occurrence frequency are included and the humanized sequence from 1bj1 (see Presta et al., supra), although the phylogenetic distance would change slightly by including amino acids with relatively low occurrence frequency such as F70(F69) and K98(K94) (see Figure 42C and D). The y-axis shows most of the designed framework VH fr123 have good structural compatibility with the structure relative to the murine reference and humanized framework VH fr123, close to DP47. These support the human-like features of the framework optimization for the inventive method described here as defined partly by its database used.

Figure 40A & B show the overlapping oligos used for library assembly, nucleic acid and amino acid sequences of the heavy chain variable region (VH) library of anti-VEGF. Degenerative positions of the DNA sequence are indicated by S (C or G), R (A or G), M (A or C), Y (C or T), K (G or T), W (A or T), respectively; and the corresponding amino acid residues encoded are labeled by "X". CDR regions are expressed in bold letters. HindIII and StyI are upstream and downstream cloning sites for the library, respectively.

Figure 41 Summary of panning of the phage display library for anti-VEGF VH. P1 to P8 indicates the 1st to the 8th rounds of

panning. VEGF concentration for coating and the amount of phages of the library (input) were decreased with the advancement of the panning. All wash conditions began with 10 times of brief rinse in PBST and ended with 10 times brief rinse in PBS before elution of bound phages took place. The incubation was performed at 37°C for 2 hours in all cases. In the 8th panning, the library was mixed with competitive phages in a ratio of 5 in the incubation.

Figure 42A Full-length sequences of hit clones from panning of the phage display library of the anti-VEGF VH. Sequencing data were obtained from clones isolated from 7th and the 8th pannings of the phage display library, respectively. Sequences of CDR regions (CDR1, 2, and 3) adopted remain to be the same as in the murine anti-VEGF antibody sequences (see Figure 9B) in library construction as described in the text. Hit rate is the occurrence of a particular clone in the indicated panning stage.

Figure 42B Summary of hit positions from panning of the phage display library of the anti-VEGF VH. The letters represent the amino acid residues in a particular position (indicated by numbers behind the letters, which was based upon linear order of amino acid sequence of variable region of heavy chain of anti-VEGF as illustrated in Figure 38A in both sequential and kabat nomenclature annotated). The published murine sequence of anti-VEGF VH and its corresponding humanized version were listed in the first and second columns on the left, respectively, in alignment with dominant residues at the same positions of human immunoglobin family III. Sequencing data were obtained from clones isolated from the 5th, 6th, 7th, and the 8th pannings of the phage display library, respectively. The numbers in front of a letter indicates the hit rate (in %) of the particular residue in sampling. (* generated by PCR error).

Figure 42C Phylogenetic analysis of top hit VH sequences from panning of the phage display libraries of the anti-VEGF, together with human germline VH3 families, murine anti-VEGF VH framework FR123 and humanized VH framework fr123 as annotated. As shown in Figure

42C, the human germline VH3 family is clustered together in phylogenetic distance as expected. The selected optimized VH frameworks also cluster together with the humanized VH sequence (see annotation), very close in phylogenetic distance to the human germline VH3 family, while the murine VH framework is very distant from the optimized VH frameworks and human germlines. This supports the conclusion that the present inventive method in designing optimized frameworks with fully human or human-like sequences of the optimized antibodies, depending on the fine balance between human-like and compatibility with structure template or templates from ensemble structure or structure average. **Figure 42D** shows the phylogenetic distances of these sequences in another tree view with annotation for a few well characterized sequences D36, D40 and D42 and related sequences. The D36 is as human as or a little better than the humanized sequence reported in its phylogenetic distance.

Figure 43A shows the sequences of the optimized VH frameworks (FR123) of anti-VEGF antibodies selected from the designer VH optimization libraries using ccFv phage display system (see description in Figures 23-25 above). The VH fr123 of D36, D40 and D42, together with the original murine antibody VH FR123 and humanized sequence (Presta et al supra) with the same CDRs from murine antibody. The dots the lower panel indicate the amino acids are the same as the reference (murine VH framework fr123).

Figure 43B shows the affinity data of 5 antibodies, parental antibody (X50) and the optimized frameworks (D36, D40, D41 and D42) of anti-VEGF antibody selected from designer libraries using BIAcore biosensor (see Figure 43A and notes in Figure 43B for their sequences). The measurement is done by measuring the change of SPR units (y-axis) vs time (x-axis) when a purified antibody binds its antigen (VEGF) immobilized on the CM5 biochip at 25°C. Both the on-rate and off-rate changes were determined from the data fitting using 1:1 Langmuir binding model. 2 humanized frameworks D36 and D40 are ~4-folder higher in binding affinity (in ccFv format) upon framework optimization than the parental/reference anti-VEGF antibody sequence (see **Figure**

22A & B for the humanized anti-VEGF antibody framework reported in the literature (Presta LG, Chen H, O'Connor SJ, Chisholm V, Meng YG, Krummen L, Winkler M, Ferrara N (1997) Cancer Res. 57, 4593-4599), while D42 is sabout the same as the reference antibody. Because the reported humanized anti-VEGF antibody (**Figures 22 A & B**) is ~ 2 times weaker than its corresponding murine antibody, these two humanized antibodies should have ~2-fold higher binding affinity upon humanization than the corresponding murine antibody.

Figure 44 shows the increased stability of the optimized VH frameworks (D36 and D40). The y-axis shows the percentage of the antibody remain active in binding to the immobilized VEGF antigen using BIAcore at 25C after the purified antibody is incubated at 4, 37 and 42C for 17 hours for the parental X50 and optimized frameworks (D36 and D40). It shows that the optimized frameworks have higher stability than the humanized VH framework reported (Presta et al. supra, 1997).

Figure 45 shows the improved expression of the optimized VH frameworks. The optimized frameworks (D36, D40 and D42) also show the improved expression relative to the parental/wild type antibody (X50) as shown in the yield expression detected by SDS-PAGE/coomassie blue staining.

Figure 46 shows amino acid sequences of VH and VL of selected antibodies against human VEGF.

DEFINITION

Structural cluster: a group of structures that are clustered into a family based on some empirically chosen cutoff values of the root mean square deviation (RMSD) (for example, of the C_α atoms of the aligned residues) and statistical significance (Z-score). These values are empirically decided after an overall comparison among structures of interest. Several programs can be used for searching structural clusters. For CE (combinatorial extension) algorithms (Shindyalov IN, Bourne PE (1998) Protein Engineering 11, 739-747), the criteria used are RMSD < 2 Å and Z-score >4. MAPS (Multiple Alignment of Protein Structures) is an automated program for comparisons of multiple protein structures. The program can automatically superimpose the 3d models of common structural similarities, detect which residues are structural equivalent among all the structures and provide the residue-to-residue alignment. The structurally equivalent residues are defined according to the approximate position of both main-chain and side-chain atoms of all the proteins. According to structure similarity, the program calculate a score of structure diversity, which can be used to build a phylogenetic tree (Lu, G. (1998) "An Approach for Multiple Alignment of Protein Structures"). In structural clustering, members within a structural cluster are analyzed to understand some consensus information about the distribution of all structural templates within a family and constraints on their sequences or sequence profiles within a structural family.

Ensemble structures: It is well-known in the structural determination by NMR (nuclear magnetic resonance), the ensemble of structures rather than a single structure, with perhaps several members, all of which fit the NMR data and retain good stereochemistry, is deposited with the Protein Data Bank. Comparisons between the models in this ensemble provide some information on how well the protein conformation was determined by the NMR constraints. It should be pointed out that all the sequences corresponding to NMR-determined ensemble structures have the same sequences (one protein with variable conformations). The structural ensemble here, additionally,

refers to different proteins with variations in sequence and/or length but have similar main chain conformations, in addition to those structures, such as from NMR determinations or from molecular dynamics simulations, have the same sequence but differ structurally due to natural shape fluctuations.

Ensemble sequences: A population of sequences that statistically defines a certain property of a target protein such as stability or binding affinity.

Ensemble average or representative structure: If all members within a structural cluster has the same length of amino acids, the positions of atoms in the main chain atoms of all structures are averaged, and the average model is then adjusted to obey normal bond distances and angles ("restrained minimization"), similar to NMR-determined average structure. If all members within a structural cluster vary in the length of amino acids, a member, which is representative of the average characteristics of all other members within the cluster, will be chosen as the representative structure.

Canonical structures: the commonly occurring main-chain conformations of the hypervariable regions.

Structural repertoire: the collection of all structures populated by a class of proteins such as the modular structures and canonical structures observed for antibody framework and CDRs.

Sequence repertoire: collection of sequences for a protein family.

Functional repertoire: the collection of all functions performed by proteins, which is related here, for example for antibodies, to the diverse functional CDRs that are capable of binding to various antigens.

Germline gene segments: refers to the genes from the germline (the haploid gametes and those diploid cells from which they are formed). The germline DNA contains multiple gene segments that encode a single

immunoglubin heavy or light chains. These gene segments are carried in the germ cells but cannot be transcribed and translated into heavy and light chains until they are arranged into functional genes. During B-cell differentiation in the bone marrow, these gene segments are randomly shuffled by a dynamic genetic system capable of generating more than 108 specificities. Most of these gene segment sequences are accessible from the germline database. The variable heavy and light chains called V-gene database are classified into subfamilies based on sequence homology.

Rearranged immunoglobulin sequences: the functional immunoglobulin gene sequences in heavy and light chains that are generated by transcribing and translating the germline gene segments during B-cell differentiation and maturation process. Most of the rearranged immunoglobulin sequences used here are from Kabat-Wu database.

BLAST: Basic Local Alignment Search Tool for pairwise sequence analysis. Blast uses a heuristic algorithm with position-independent scoring parameters to detect similarity between two sequences, the default parameters are used with Expect at 10, Word Size 3 Scoring matrix BLOSUM62, Gap costs for existence 11 and extension 1.

PSI-BLAST: The Position-Specific Iterated BLAST, or PSI-BLAST program performs an iterative search in which sequences found in one round of searching are used to build a score model for the next round of searching. In PSI-BLAST the algorithm is not tied to a specific score matrix. Traditionally, it has been implemented using an AxA substitution matrix where A is the alphabet size. PSI-BLAST instead uses a QxA matrix, where Q is the length of the query sequence; at each position the cost of a letter depends on the position with respect to the query and the letter in the subject sequence. Two PSI-BLAST parameters have been adjusted: the pseudocount constant default has been changed from 10 to 7, and the E-value threshold for including matches in the PSI-BLAST model has been changed from 0.001 to 0.002.

Energy landscape: An energy distribution where peaks and wells define ensemble states of a molecule. It is believed that an energy landscape can provide a complete description of the folding process as well as descritions of local structural states, whereas the common optimized or minimized structure describes only a single structural species out of a collection of many possible states within a local energy minimum.

Fitness/Fitness score: A measure of an experimentally observable property of a molecule such as stability, activity and affinity.

Fitness landscape: A distribution of a fitness score defined by other intrinsic parameters of the molecule, such as sequence.

Sequence space: See sequence repertoire.

Structure Space: See structure repertoire.

Functional Space: See functional repertoire

Lead sequence: the sequence used for searching sequence database.

Variant profile/sequence profile/positional variant profile (PVP): description of the amino acid entropy at each position for a set of peptide sequences. This includes both the range and frequency of the amino acids (AA-PVP) or nucleic acids (NA-PVP).

Hit library/Hit list: the collection of sequences found by searching the sequence database using the lead sequence or sequence profile.

Hit variant library I/Library I: An in silico amino acid sequence library derived from the combinatorial enumeration of the variant profile of the hit library.

Hit variant library II/Library II/Designed amino acid library/Refined amino acid library: An in silico amino acid sequence

library derived from the hit variant library I as a result of a re-profiling or specific design. Re-profiling of the variants can be accomplished 1) by selecting a sequence cluster(s) based energy ranking with a specific cut off value or a window of sequences containing key amino acid residues, 2) by including specific positional residues indentified by functional screening, and/or 3) by inclusion or exclusion of residues or sequence clusters as determined by those trained in the arts using any other means available for making such determinations.

Hit variant library III/Library III: An amino acid sequence library that is expressed in vitro by the degenerate oligonucleotide library (below) for functional screening. Library III expands the sequence space of Library II due to back translation, optimized codon usage, recombination at the nucleotide level and expression of the resulting combinatorial nucleic acid library.

Degenerate nucleic acid/oligonucleotide library: The library of mixed oligonucleotides that is used to target an amino acid variant profile that corresponds to a designed amino acid library (library II above). It is derived from the combinatorial enumeration of the corresponding nucleic acid positional variant profile that is back translated from the amino acid positional variant profile of library II using optimized codon(s).

Combinatorial amino acid/peptide library: Library generated from the complete combinatorial enumeration of an amino acid positional variant profile. Library I and II are such libraries.

Combinatorial nucleic acid /oligonucleotide library: Library generated from the complete combinatorial enumeration of a nucleic acid positional variant profile.

DNA shuffling: A method of generating recombinant oligonucleotides from a mixture of parental sequences through multiple iterations of oligonucleotide fragmentation and homologous recombination (Stemmer WP (1994) Nature 370, 389-391)

In silico rational library design: a method of designing a digital amino acid or nucleic acid library that incorporates evolutionary, structural, and functional data in order to define and efficiently sample ensembles in the sequence and structure spaces in order to identify those that have a desired fitness.

Profile Hidden Markov Model (profile HMM): A statistical model of the primary structure consensus of a sequence family based on the sequence profile of proteins. It uses position-specific scores for amino acids and for opening and extending an insertion and deletion to detect remote sequence homologues based on the statistical description of the consensus of a multiple sequence alignment. The multiple sequence alignments are given either by the multiple sequence alignment program such as ClustalW or structure-based multiple sequence alignment given by structural clustering.

Threading: a process of assigning the folding of the protein by threading its sequence to a library of potential structural templates by using a scoring function that incorporates the sequence as well as the local parameters such as secondary structure and solvent exposure. The threading process starts from prediction of the secondary structure of the amino acid sequence and solvent accessibility for each residue of the query sequence. The resulting one-dimensional (1D) profile of the predicted structure is threaded into each member of a library of known 3D structures. The optimal threading for each sequence-structure pair is obtained using dynamic programming. The overall best sequence-structure pair constitutes the predicted 3D structure for the query sequence.

Reverse threading: a process of searching for the optimal sequence(s) from sequence database by threading them onto a given target structure and/or structure cluster. Various scoring functions may be used to select for the optimal sequence(s) from the library comprising protein sequences with various lengths.

Side chain rotamer: the conformation of an amino acid side chain defined in terms of the dihedral angels or chi angles of side chains.

Rotamer library: a distribution of side chain rotamers either based on the backbone dihedral angles phi and psi called backbone-dependent rotamer library or independent of backbone dihedral angles called backbone-independent rotamer library for all amino acids derived from the analysis of side chain conformations in the protein structural database

See Dunbrack RL and Karplus M (1993) JMB 230, 543-574.

DETAILED DESCRIPTION OF THE INVENTION

The present invention provides a system and method for efficiently generating and screening protein libraries for optimized proteins with improved biological functions, such as improved binding affinity towards biologically and/or therapeutically important target molecules. The process is carried out computationally in a high throughput manner by mining the ever-expanding databases of protein sequences of all organisms, especially human. With a combination of database-mining of evolutionary sequences from nature with computational design of structurally relevant variants of the natural sequences, the method of the present invention represents a distinct departure from other approaches in computational design and functional screening of protein libraries.

By using this innovative method, a biased library of proteins such as antibodies can be constructed based on computational evaluation of extremely diverse protein sequences and functionally relevant structures in silico. This ensemble-based statistical method of library construction and screening in silico efficiently maps out the distribution of the fitness and energy landscapes in protein sequence and structure spaces, a goal practically unachievable for in vitro or in vivo screening. Following screening in silico, an expanded nucleic acid library based on the sequences encoding the selected proteins is constructed, introduced into an expression system, and screened for proteins with improved or novel functions in vitro or vivo.

Figure 1 is a series of flowcharts outlining various embodiments of the method of the present invention. Based on a lead protein with known sequence and/or structure, libraries of proteins can be constructed and screened for candidates with desired functions following at least four different routes (Route I-IV) shown in **Figure 1**.

In one embodiment, the method comprises the steps of:

providing an amino acid sequence of the variable region of the heavy chain (V_H) or light chain (V_L) of a lead antibody;

identifying the amino acid sequences in the CDRs of the lead antibody;

selecting one of the CDRs in the V_H or V_L region of the lead antibody;

providing an amino acid sequence that comprises at least 3 consecutive amino acid residues in the selected CDR, the selected amino acid sequence being a lead sequence;

comparing the lead sequence with a plurality of tester protein sequences; and

selecting from the plurality of tester protein sequences at least two peptide segments that have at least 15% sequence identity with the lead sequence, the selected peptide segments forming a hit library.

The method may further comprise the step of:

constructing a nucleic acid library comprising DNA segments encoding the amino acid sequences of the hit library.

Route I in **Figure 1A** schematically represents this embodiment. According to this embodiment, a lead protein (e.g., an antibody), with known sequence and structure, is provided. A rich pool of protein sequences (e.g., human antibody repertoire) is screened for varying identity with a selected segment of the lead protein (herein after referred to as "the lead sequence"). From this screening, a list of protein sequences can selected with varying degrees of homology (herein after referred to as the "hit library") using a sequence alignment method such as Hidden Markov Model or HMM. Amino acid sequences of the hit library are then profiled against the lead sequence to show variance of amino acid residues in each position of the lead sequence. As will be described in more detail in Section 7 below, some or all of the profiled sequences in the hit library are selected and translated back to a library of nucleic acid for functional screening in vitro or vivo.

Optionally, the method may further comprise the steps of:

building an amino acid positional variant profile of the hit library;

converting amino acid positional variant profile of the hit library into a nucleic acid positional variant profile by back-translating the amino acid positional variants into their corresponding genetic codons; and

constructing a degenerate nucleic acid library of DNA segments by combinatorially combining the nucleic acid positional variants.

Route II in **Figure 1B** schematically represents this embodiment. According to this embodiment, after amino acid sequences of the hit library are profiled against the lead sequence, a combinatorial library (herein after referred to as "hit variant library I" or "library I") is constructed based on the frequency of an amino acid in each residue position (also called amino acid positional variant profile or AA-PVP). Using this approach the hit variant library I is substantially larger than the hit library. By modifying (e.g., filtering) the AA-PVP to bias towards preferred mutants for each position, based on those observed at higher frequencies, indicating evolutionary preference, a reduced variant profile is generated and its combinatorial enumeration leads to hit variant library II. Hit variant library II profile is translated back to a library of nucleic acid for functional screening in vitro or vivo.

Optionally, the genetic codons may be the ones that are preferred for expression in bacteria. Optionally, genetic codons may bethe ones that can reduce the sizechosen such that the diversity of the degenerate nucleic acid library of DNA segments is within the experimentally coverable diversity without undue experimental effort, preferably below $1x10^7$ and more preferably below $1x10^6$.

In another embodiment, the method comprises the steps of:

providing an amino acid sequence of the variable region of the heavy chain (V_H) or light chain (V_L) of a lead antibody;

identifying the amino acid sequences in the CDRs and FRs of the lead antibody;

selecting one of the CDRs in the V_H or V_L region of the lead antibody;

providing a first amino acid sequence that comprises at least 3 consecutive amino acid residues in the selected CDR, the selected amino acid sequence being a CDR lead sequence;

comparing the CDR lead sequence with a plurality of CDR tester protein sequences;

selecting from the plurality of CDR tester protein sequences at least two peptide segments that have at least 15% sequence identity with the CDR lead sequence, the selected peptide segments forming a CDR hit library;

selecting one of the FRs in the V_H or V_L region of the lead antibody;

providing a second amino acid sequence that comprises at least 3 consecutive amino acid residues in the selected FR, the selected amino acid sequence being a FR lead sequence;

comparing the FR lead sequence with a plurality of FR tester protein sequences; and

selecting from the plurality of FR tester protein sequences at least two peptide segments that have at least 15% sequence identity with the FR lead sequence, the selected peptide segments forming a FR hit library; and

combining the CDR hit library and the FR hit library to form a hit library.

According to the method, the plurality of CDR tester protein sequences may comprise amino acid sequences of human or non-human antibodies.

Also according to the method, the plurality of FR tester protein sequences may comprise amino acid sequences of human origins, preferably human or humanized antibodies (e.g., antibodies with at least 50% human sequence, preferably at least 70% human sequence, more preferably at least 90 % human sequence, and most preferably at least 95% human sequence in V_H or V_L), more preferably fully human antibodies, and most preferably human germline antibodies.

Also according to the method, at least one of the plurality of CDR tester protein sequences is different from the plurality of FR tester protein sequences.

Also according to the method, the plurality of CDR tester protein sequences are human or non-human antibody sequences and the plurality of FR tester protein sequences are human antibody sequences, preferably human germline antibody sequences.

The method may further comprise the step of:

constructing a nucleic acid library comprising DNA segments encoding the amino acid sequences of the hit library.

Optionally, the method may further comprise the steps of:

building an amino acid positional variant profile of the CDR hit library;

converting the amino acid positional variant profile of the CDR hit library into a first nucleic acid positional variant profile by back-translating the amino acid positional variants into their corresponding genetic codons; and

constructing a degenerate CDR nucleic acid library of DNA segments by combinatorially combining the nucleic acid positional variants.

Optionally, the genetic codons may be the ones that are preferred for expression in bacteria. Optionally, genetic codons may bethe ones that can reduce the sizechosen such that the diversity of the degenerate nucleic acid library of DNA segments within the experimentally coverable diversity (< 10^6 or 7) without undue experimental effort.is below $1x10^7$, preferably below $1x10^6$.

In yet another embodiment, the method comprises the steps of:

providing an amino acid sequence of the variable region of the heavy chain (V_H) or light chain (V_L) of a lead antibody;

identifying the amino acid sequences in the FRs of the lead antibody;

selecting one of the FRs in the V_H or V_L region of the lead antibody;

providing a first amino acid sequence that comprises at least 3 consecutive amino acid residues in the selected FR, the selected amino acid sequence being a first FR lead sequence;

comparing the first lead FR sequence with a plurality of FR tester protein sequences; and

selecting from the plurality of FR tester protein sequences at least two peptide segments that have at least 15% sequence identity with the first FR lead sequence, the selected peptide segments forming a first FR hit library.

The method may further comprise the steps of

providing a second amino acid sequence that comprises at least 3 consecutive amino acid residues in a FR that is different from the selected FR, the selected amino acid sequence being a second FR lead sequence;

comparing the second FR lead sequence with the plurality of FR tester protein sequences; and

selecting from the plurality of FR tester protein sequences at least two peptide segments that have at least 15% sequence identity with the second FR lead sequence, the selected peptide segments forming a second FR hit library; and

combining the first FR hit library and the second FR hit library to form a hit library.

According to the method, the lead CDR sequence may comprise at least 5 consecutive amino acid residues in the selected CDR. The selected CDR may be selected from the group consisting of V_H CDR1, V_H CDR2, V_H CDR3, V_L CDR1, V_L CDR2, and V_L CDR3 of the lead antibody.

Also according to the method, the lead FR sequence may comprise at least 5 consecutive amino acid residues in the selected FR. The selected FR may be selected from the group consisting of V_H FR1, V_H FR2, V_H FR3, V_H FR4, V_L FR1, V_L FR2, V_L FR3 and V_L FR4 of the lead antibody.

The method may further comprise the step of:

constructing a nucleic acid or degenerate nucleic acid library comprising DNA segments encoding the amino acid sequences of the hit library.

In another aspect of the invention, a method is provided for in silico selection of antibody sequences based on the amino acid sequence of a region in a lead antibody, i.e., the "lead sequence", and its 3D structure. The structure of the lead sequence is employed to search databases of protein structures for segments having similar 3D structures. These segments are aligned to yield a sequence profile, herein after referred to as the "lead sequence profile". The lead sequence profile is employed to search databases of protein sequences for remote homologues of the lead sequence having low sequence identity and yet structurally similar. By using the method, a library of diverse antibody sequences can be constructed and screened experimentally in vitro or in vivo for antibody mutants with improved or desired function(s).

In one embodiment, the method comprises the steps of:

providing an amino acid sequence of the variable region of the heavy chain (V_H) or light chain (V_L) of a lead antibody;

identifying the amino acid sequences in the CDRs of the lead antibody;

selecting one of the CDRs in the V_H or V_L region of the lead antibody;

providing an amino acid sequence that comprises at least 3 consecutive amino acid residues in the selected CDR, the selected amino acid sequence being a lead sequence;

providing a three-dimensional structure of the lead sequence;

building a lead sequence profile based on the structure of the lead sequence;

comparing the lead sequence profile with a plurality of tester protein sequences; and

selecting from the plurality of tester protein sequences at least two peptide segments that have at least 10% sequence identity with lead sequence, the selected peptide segments forming a hit library.

According to the method, the three-dimensional structure of the lead sequence may be a structure derived from X-crystallography, nuclear magnetic resonance (NMR) spectroscopy or theoretical structural modeling.

According to the method, the step of building a lead sequence profile may include:

comparing the structure of the lead sequence with the structures of a plurality of tester protein segments;

determining the root mean square difference of the main chain conformations of the lead sequence and the tester protein segments;

selecting the tester protein segments with root mean square difference of the main chain conformations less than 5 Å, preferably less than 4 Å, more preferably less than 3 Å, and most preferably less than 2 Å; and

aligning the amino acid sequences of the selected tester protein segments with the lead sequence to build the lead sequence profile.

Optionally, the structures of the plurality of tester protein segments are retrieved from the protein data bank.

Optionally, the step of building a lead sequence profile may include:

comparing the structure of the lead sequence with the structures of a plurality of tester protein segments;

determining the Z-score of the main chain conformations of the lead sequence and the tester protein segments;

selecting the segments of the tester protein segments with the Z-score higher than 2, preferably higher than 3, more preferably higher than 4, and most preferably higher than 5; and

aligning the amino acid sequences of the selected tester protein segments with the lead sequence to build the lead sequence profile.

Optionally, the step of building a lead sequence profile may be implemented by an algorithm selected from the group consisting of CE, MAPS, Monte Carlo and 3D clustering algorithms.

The method may further comprise the step of:

constructing a nucleic acid library comprising DNA segments encoding the amino acid sequences of the hit library.

Optionally, the method may further comprise the steps of:

building an amino acid positional variant profile of the hit library;

converting amino acid positional variant profile of the hit library into a nucleic acid positional variant profile by back-translating the amino acid positional variants into their corresponding trinucleotide codons; and

constructing a degenerate nucleic acid library of DNA segments by combinatorially combining the nucleic acid positional variants.

Any of the above methods may further comprise the following steps:

introducing the DNA segments in the nucleic acid or degenerate nucleic acid library into cells of a host organism;

expressing the DNA segments in the host cells such that recombinant antibodies containing the amino acid sequences of the hit library are produced in the cells of the host organism; and

selecting the recombinant antibody that binds to a target antigen with affinity higher than 10^6 M^{-1}, preferably 10^7 M^{-1}, more preferably 10^8 M^{-1}, and most preferably 10^9 M^{-1}.

In one embodiment, the method comprises the steps of:

providing an amino acid sequence of the variable region of the heavy chain (V_H) or light chain (V_L) of a lead antibody, the lead antibody having a known three dimensional structure which is defined as a lead structural template;

identifying the amino acid sequences in the CDRs of the lead antibody;

selecting one of the CDRs in the V_H or V_L region of the lead antibody;

providing an amino acid sequence that comprises at least 3 consecutive amino acid residues in the selected CDR, the selected amino acid sequence being a lead sequence;

comparing the lead sequence profile with a plurality of tester protein sequences;

selecting from the plurality of tester protein sequences at least two peptide segments that have at least 10% sequence identity with lead sequence, the selected peptide segments forming a hit library;

determining if a member of the hit library is structurally compatible with the lead structural template using a scoring function; and

selecting the members of the hit library that score equal to or better thanor equal to the lead sequence.

According to the method, the scoring function is an energy scoring function selected from the group consisting of electrostatic interactions, van der Waals interactions, electrostatic solvation energy, solvent-accessible surface solvation energy, and conformational entropy.

Optionally, the scoring function is one incorporating a forcefield selected from the group consisting of the Amber forcefield, Charmm forcefield, the Discover cvff forcefields, the ECEPP forcefields, the GROMOS forcefields, the OPLS forcefields, the MMFF94 forcefield, the Tripos forcefield, the MM3 forcefield, the Dreiding forcefield,and UNRES forcefield,and other knowledge-based statistical forcefield (mean field) and structure-based thermodynamic potential functions.

Also according to the method, the step of selecting the members of the hit library includes selecting the members of the hit library that

have a lower or equal total energy than that of the lead sequence calculated based on a formula of

$\Delta E_{total} = E_{vdw} + E_{bond} + E_{angel} + E_{electrostatics} + E_{solvation}$

Also according to the method, the step of selecting the members of the hit library includes selecting the members of the hit library that have a lower binding free energy than that of the lead sequence calculated as the difference between the bound and unbound states using a refined scoring function

$$\Delta G_b = \Delta G_{MM} + \Delta G_{sol} - T\Delta S_{ss}$$

where

$$\Delta G_{MM} = \Delta G_{ele} + \Delta G_{vdw} \quad (1)$$

$$\Delta G_{sol} = \Delta G_{ele\text{-}sol} + \Delta G_{ASA} \quad (2)$$

The method may further comprise the step of:

constructing a nucleic acid library comprising DNA segments encoding the amino acid sequences of the hit library.

Route III in **Figure 1C** schematically represents this embodiment. According to this embodiment, sequences of the hit library are built into the 3D structure of the lead protein by substituting side chains from a rotamer database, and scored for their structural compatibility with the 3D structure of the lead protein (herein after referred to as "the lead structural template". Based on the structural evaluation, the hit library is reprofiled by ranking according to the score in energy function. Some of the sequences in the hit library with a desired energy function are selected and translated back to a library of nucleic acid for functional screening in vitro or vivo. There is no amino acid sequence combinatorial step in this embodiment.

Optionally, the method may further comprise the steps of:

building an amino acid positional variant profile of the hit library;

converting amino acid positional variant profile of the hit library into a nucleic acid positional variant profile by back-translating the

amino acid positional variants into their corresponding trinucleotide codons; and

constructing a degenerate nucleic acid library of DNA segments by combinatorially combining the nucleic acid positional variants.

In yet another embodiment, the method comprises:

In one embodiment, the method comprises the steps of:

providing an amino acid sequence of the variable region of the heavy chain (V_H) or light chain (V_L) of a lead antibody, the lead antibody having a known three dimensional structure which is defined as a lead structural template;

identifying the amino acid sequences in the CDRs of the lead antibody;

selecting one of the CDRs in the V_H or V_L region of the lead antibody;

providing an amino acid sequence that comprises at least 3 consecutive amino acid residues in the selected CDR, the selected amino acid sequence being a lead sequence;

comparing the lead sequence with a plurality of tester protein sequences;

selecting from the plurality of tester protein sequences at least two peptide segments that have at least 10% sequence identity with lead sequence, the selected peptide segments forming a hit library;

building an amino acid positional variant profile of the hit library based on frequency of amino acid variant appearing at each position of the lead sequence;

combining the amino acid variants in the hit library to produce a combination of hit variants which form a hit variant library;

determining if a member of the hit variant library is structurally compatible with the lead structural template using a scoring function; and

selecting the members of the hit variant library that score equal to or better than the lead sequence.

According to the method, the step of combining the amino acid variants in the hit library includes:

selecting the amino acid variants with frequency of appearance higher than 4 times, preferably 6 times, more preferably 8 times, and

most preferably 10 times (2% to 10% and preferably 5% of the frequency for the cutoff and then include some of the amino acids from the lead sequence if they are missed after cutoff); and

combining the selected amino acid variants in the hit library to produce a combination of hit variants which form a hit variant library.

According to the method, the scoring function is an energy scoring function selected from the group consisting of electrostatic interactions, van der Waals interactions, electrostatic solvation energy, solvent-accessible surface solvation energy, and conformational entropy.

Optionally, the scoring function is one incorporating a forcefield selected from the group consisting of the Amber forcefield, Charmm forcefield, the Discover cvff forcefields, the ECEPP forcefields, the GROMOS forcefields, the OPLS forcefields, the MMFF94 forcefield, the Tripos forcefield, the MM3 forcefield, the Dreiding forcefield,and UNRES forcefield,and other knowledge-based statistical forcefield (mean field) and structure-based thermodynamic potential functions.

The method may further comprise the step of:

constructing a nucleic acid library comprising DNA segments encoding the amino acid sequences of the selected members of the hit variant library.

Route IV in **Figure 1D** schematically represents this embodiment. According to this embodiment, after amino acid sequences of the hit library are profiled against the lead sequence, a combinatorial library of hit variants, i.e., hit variant library I. Hit variant library II is constructed based on the frequency of appearance of an amino acid in each residue position (as in Route III). Sequences of hit variant library II are built into the 3D structure of the template protein by substituting side chains from a rotamer database, and scored for their structural compatibility with the lead structural template. Based on the structural evaluation, the hit variant library II is re-profiled by ranking according to the score in energy function. Some of the sequences in the re-profiled hit variant library II with a desired energy function are selected and translated back to a library of nucleic acid for functional screening in vitro or in vivo. Additional modifications to the variant profile of library II can be applied based on other selective

factors determined by those trained in the arts. Thus library II is a designed library based on evolutionary, structural, and/or functional data.

Based on the sequences of the selected hit list or hit variant library II that are generated in silico, a synthetic library of antibody can be constructed in the lab and screened against the target antigen. A wide variety of biological assays can be used for high throughput screening, such as phage display (Smith and Scott (1993) Method Enzymol. 217: 228-257), ribosome display (Hanes and Pluckthun (1997) Proc. Natl. Acad. Sci. USA 94:4937-4942), yeast display (Kieke et al. (1997) Protein Eng. 10:1303-1310), and other extra- or intra-cellular expression systems.

In another embodiment, the method comprises the steps of:

providing an amino acid sequence of the variable region of the heavy chain (V_H) or light chain (V_L) of a lead antibody, the lead antibody having a known three dimensional structure;

providing 3D structures of one or more antibodies with different sequences in V_H or V_L region than that of the lead antibody;

forming a structure ensemble by combining the structures of the lead antibody and the one or more antibodies; the structure ensemble being defined as a lead structural template;

identifying the amino acid sequences in the CDRs of the lead antibody;

selecting one of the CDRs in the V_H or V_L region of the lead antibody;

providing an amino acid sequence that comprises at least 3 consecutive amino acid residues in the selected CDR, the selected amino acid sequence being a lead sequence;

comparing the lead sequence with a plurality of tester protein sequences;

selecting from the plurality of tester protein sequences at least two peptide segments that have at least 10% sequence identity with lead sequence, the selected peptide segments forming a hit library;

building an amino acid positional variant profile of the hit library based on frequency of amino acid variant appearing at each position of the lead sequence;

combining the amino acid variants in the hit library to produce a combination of hit variants which form a hit variant library;

determining if a member of the hit variant library is structurally compatible with the lead structural template using a scoring function; and

selecting the members of the hit variant library that score equal to or better than the lead sequence.

Such a process, i.e., computational prediction of a digital antibody library and experimental screening of the synthetic antibody library, can be reiterated to improve the binding affinity of selected antibodies. After the first round of screening, the three-dimensional structure of the selected antibody or antibodies can be modeled computationally. Also, the structure can be modified by expanding the sequence and conformation space and by subjecting it to soft docking by the target antigen to create a second generation of the digital antibody library. The second generation of the digital antibody library can then be screened experimentally to select for the antibodies with higher affinity than the first generation of selected antibodies. Such a reiterating process of structural modification and screening against the antigen effectively mimics the natural process of antibody maturation in vertebrates.

The conceptual framework and practical applications of the present invention are described in detail in the following sections.

1. Conceptual Framework of the Present Invention

The present invention provides innovative solutions to problems long existing in the field of molecular biology, in particular, protein folding and design. The approach developed by the inventors combines the best ideas in protein folding and design into a powerful integrated system that can develop novel protein products for practical applications in a high throughput and cost-effective manner.

The inventors believe that a central issue in molecular biology is to map out the functional repertoire of biopolymers such as proteins, RNA and DNA molecules in terms of their sequence and structure. The functional repertoire of biopolymers is shaped by a complex interplay of selective pressures during the course of evolution and by physical

constraints on the folding and stability of biopolymers under various environmental conditions. What is the difference between the natural biopolymers and the random polymers? What is the best strategy to exploit the rich diversity of function, sequence and structure spaces of naturally occurring biopolymers to create novel biopolymers with stable structures and proper biological functions? Answers to these questions are of fundamental interest in molecular design and evolution, especially in the discovery of novel proteins with enhanced binding and catalytic activities.

The present invention addresses these issues in the following three steps: 1) discuss the general conceptual framework underlying protein folding and evolution to provide the basic knowledge needed for understanding the present invention; 2) describe the current experimental and theoretical methods used in protein folding and design and the problems related to these approaches; and 3) outline the inventive approaches to solve some of the longstanding problems in protein design and engineering.

1) Protein Folding and Evolution

Proteins are essential molecules for performing a diverse array of biological functions. Proteins acquire their biological functions by folding their linear sequences into unique three-dimensional structures. Predicting protein structure from sequence still remains an unsolved problem. However, important progress has been made in understanding the mechanisms of protein folding, especially with the advent of the statistical interpretation of the ensembles of intermediates and transition states in folding pathways.

The dynamics nature of protein conformation in solution has been well documented in both experimental and theoretical studies. Dynamic fluctuation in protein conformation can be essential for carrying out some of their biological functions such as in allosteric regulation (Monod, J., Wyman, J., and Changeux, J. P. (1965) J. Mol. Biol., 12:88-118) in protein-protein and protein-nucleic acid interactions, and conformational gating (Zhou, H-X, Wlodek, S.T., McCammon, J.A. (1998) PNAS 95, 9280-9283.) in enzymatic activities.

The continuous ensemble approach is favored over the classical

discrete-state approach for describing protein folding mechanism because it provides, not only a more realistic view of biopolymers, as compared to the static x-ray structure, but a general framework for describing a growing body of experimental observations that would difficult to interpret, otherwise (Hong Qian (2002) Protein Science 11, 1-5). This view emphasizes the importance of using the statistical properties of the continuous distribution of conformational ensembles on an energy landscape in understanding biological functions of macromolecules (Baldwin RL (1995) 5, 103-109 J Biomol. NMR; Pande VJ etc (1998) Curr. Opin. Struct. Biol., 8, 68-79).

The random energy model (REM) used to study heteropolymer freezing and design provides an excellent approximate physical model for protein folding and design (see Vijay S. Pande, Alexander Yu. Grosberg, and Toyoichi Tanaka, Review of Modern Physics, Vol. 72, No. 1, 2000 and references within). Much has been learned from the quantitative studies of simple models of protein folding and design based on the statistical properties of the freezing transition for heteropolymers. The phase transition between conformational states of ensembles distributed in continuous energy spectra provides a more realistic description of the folding and binding properties of proteins compared to the traditional view of a few discrete states populating a set of well-defined energy wells. The REM landscape suggests that a necessary and sufficient condition for any designed sequences to fold into a kinetically accessible and thermodynamic stable conformation is an energy distribution that shows a continuous energy spectrum in the upper portion and a pronounced energy minimum in the lower portion (See Vijay S. Pande, Alexander Yu. Grosberg, and Toyoichi Tanaka, Review of Modern Physics, Vol. 72, No. 1, 2000 and references within; Shakhnovich and Gutin, 1993 PNAS, 90, 7195-7199). Therefore, sequences should be designed to enlarge the energy gap between the ground state of the designed sequence and the bottom of the REM continuous energy spectrum. The energy gap is enlarged either by pulling down the energy of the native conformation of sequences (positive design for stability) or by pushing up the energy of alternative conformation of a sequence (negative design for specificity).

The general rules derived from this simple model of protein

folding was strictly followed in a recent de novo computational protein design: the composition of amino acids is kept unchanged while the energy is minimized (Koehl P & Levitt M (1999) J Mol Biol 293, 1161-1181). It is argued that defining the ensemble characteristics of the sequences compatible to a given structure is more important than finding the specific optimal sequence (Koehl P & Levitt M (1999) J Mol Biol 293, 1183-1193). The multiple alignment of the designed sequences defines a sequence space that is measured by information entropy; a subset of this sequence space is similar in size to the sequence space derived from the same structural alignment observed in Nature (Koehl P; Levitt M (2001) PNAS 1-6). This work shows that topology and stability defines the sequence space of a given fold, while a subset of the sequence space can be defined by the functional fitness. However, this method poses too much restriction on the choice of amino acids at each position by keeping the composition of amino acids unchanged.

The dynamic nature of protein evolution has been actively pursued by theoretical and evolutionary biologists (Maynard-Smith, J (1970) Nature, 225, 563-564). Mapping sequences (genotypes) into values measuring the fitness landscape is a core issue of evolutionary biology. Although the relationship between genotype and phenotype is too complicated to be analyzed in general by a quantitative method, this relationship can be, however, simplified to relations between sequence (genotype) and structure (phenotype) and therefore, fitness values can be used to score the fitness of sequences to a given shape of biopolymers as shown below:

Genotype (sequences) ←fitness score→ Phenotype (structure)

Proteins observed in nature have evolved under selective pressures to perform specific functions. Interestingly, fitness landscape of functional proteins has been mapped and simulated using similar tools as in protein folding field. The fitness landscape is mapped out in sequence space in order to define the mutant ensemble that would enhance the functional property of a protein. Statistical properties of the sequence ensemble have been used to describe the neutral network

in sequence space of the target protein (Stadler P F. Journal of Molecular Structure (Theochem) 463, 7-19 (1999); J Theor Biol 2001, 212, 35-46).

There are three essential ingredients embedded in the landscape theory: a set of configurations; a fitness function assigned to each configuration; and the connectivity between configurations that define the distance or relation between configurations. A fitness function can be broadly defined as a property of a protein such as the binding affinity between two proteins (receptor and ligand; antigen and antibody), the catalytic activity of an enzyme, or the structural stability of a target scaffold.

From the perspective of evolution, the fitness landscapes arising from mapping the sequence-structure relations of natural RNA and proteins predict the existence of neutral networks in sequence space evolved under partially correlated landscapes, providing an efficient route to adaptive evolution toward a new fitness function. In contrast, the random sequences evolved under rugged fitness landscapes without neutral neighbors are trapped in local optima, leading to localized populations in sequence space. The natural sequence has undergone evolutionary optimization under selective pressure through a mountain climbing process. An effective route to a new fitness function via sequence alteration is to follow the neutral networks in sequence space rather than by random mutation. (Stadler P F. Journal of Molecular Structure (Theochem) 463, 7-19 (1999); J Theor Biol 2001, 212, 35-46; Aderonke Babajide etc (1997) Folding & Design 2, 261-269). The relative efficiency of searching the fitness landscape via point-mutation versus gene recombination in protein space can be simulated and compared using the REM as well as heterpolymer-based model (Bogarad L, Deem MW (1999) PNAS 96, 2591-2595; Cui Y, Wong WH, Bornberg-Bauer E, Chan HS (2002) 99, 809-814).

The above-described theoretical studies of protein folding and evolution using simplified models have provided some insights into the statistical properties of ensemble states of protein structures and sequences during folding and evolution. The inventors believe that a theory that combines the concepts in molecular biology, physics of spin glass and physics of heteropolymer should provide a unified framework

for the dynamic properties of biopolymers. The question now becomes how to turn such a conceptual framework based on models of proteins into a practical approach to map out the functional landscape of proteins in both sequence and structure spaces.

2) Current Experimental and Theoretical Methods for Protein Sequence Design in the Art and Problems That Lie Therein

A major goal in protein engineering is to generate proteins with novel or improved function. To this end, two alternative approaches have been used to obtain proteins, mainly enzymes, with desired properties: in vitro directed molecular evolution and structure-based computational design. The approach of in vitro directed evolution employs homologous sequences, random mutagenesis and gene shuffling to generate diverse sequence library. Mutants with desirable properties are selected in a high throughput screening and re-shuffled. This procedure is iterated until a desired level of functional enhancement is attained.

The first law in directed evolution that states, "You get what you screen for," underscores the importance of the screening method in evaluating the functional fitness of the protein libraries (Wintrode, P & Arnold, FH (2000) Adv Protein Chem. 55, 161-226). The availability and improved sensitivity of high throughput enzymatic screenings have led to some successes of directed evolution. Compared to rational engineering, the directed evolution requires little or no additional information such as the structure of the target enzyme, and can screen directly for biological activities from a large pool of molecules under defined selective pressure.

The dependence on the screening ability imposes an upper limit on the size of the generated combinatorial library and therefore the size of the sampled functional space. Because random mutagenesis by using error-prone PCR is biased and inefficient process for generating a diverse library, the probability of a significant functional improvement by any single random mutation is small and drops rapidly for multiple simultaneous random mutations. It is also difficult to generate several mutants simultaneously at a single codon position at the nucleic acid level.

Furthermore, the dependence of the DNA shuffling on homologous recombination of sequences with high homology (>70%) limits the sequence space that the resulting library can span. As a result, each successive iteration of shuffling and screening leads to sampling in a shrinking local sequence space. This may be efficient for identifying new homologous sequences with enhanced properties but may not be adequate for identifying truly novel sequences with potentially greater functional improvements.

Nonetheless, beneficial amino acid substitutions are generated and identified by incorporating random mutagenesis. Accumulating beneficial point mutations has been used successfully to evolve and screen a number of important enzymes with desired properties. Besides the simple random mutagenesis strategy, gene recombination by DNA shuffling, including family shuffling approach that combines genes from multiple parents of the same or different species, creates highly improved biocatalysts (Ness J E Del Cardayre, SB Minshull, J & Stemmer, WPC (2000) Adv Protein Chem 55, 261-292).

As a closely related problem to protein folding, protein design is considered as the inverse folding problem (Drexler, KE (1981) PNAS 78, 5275-5278; Pabo, C. (1983) Nature 301, 200): finding the sequences that give rise to the target structure. Designing protein sequences that would give rise to the target scaffold is considered to be an important step in engineering proteins with improved properties for a wide range of applications.

A major issue related to the inverse folding protocol is the necessity of maintaining a rigid protein backbone. Because conformational space needed to be sampled is enormous, for practical reasons, the static X-ray structure of a protein is still widely used as a starting point in rational structure-based protein or drug design. The inverse protein folding approach tries to compute the optimal sequence compatible with the protein structure based on semi-empirical all-atom energy functions describing the interactions between amino acids. While the native protein is known to tolerate small perturbation with robust conformational adaptation, the computational ground state of a rigid protein backbone is, however, not sufficiently adaptable to small perturbation in protein backbone or side chain rotamers to provide an

accurate measure of stability.

Some efforts in backbone parameterization have been made to address these issues by adjusting the relative orientation between regular secondary structures (Harbury, PB, Tidor B. & Kim, PS (1995) Protein Science 92, 8408-8412; Su A & Mayo SL (1997) Prot Sci. 6, 1701-1707; Harbury PB, Plecs JJ, Tidor B, Alber T, Kim PS (1998) Science 282, 1462-1467). The inventors believe that a simple but efficient solution to relieve the local constraints is energy minimization including backbone and side chains (Keating AE, Malashkevich VN, Tidor B, Kim PS (2001) PNAS 98, 14825-30) for any structure type of a protein as demonstrated in the present invention for protein loops, which are irregular and whose backbone movements are hard to parameterize in general.

Apart from a few cases with regular secondary structures (see below), most of the protein design strategy strictly follows the inverse folding protocols in sequence selection in order to reduce the immense task of searching the conformational space. Even with backbone fixed, powerful searching algorithms, including stochastic Monte Carlo or genetic algorithm and deterministic dead end elimination, are needed to search for the best solution to an empirical energy function that incorporates various factors in stabilizing a protein assembled from a rotamer library of protein side chains (Ponder, J.W. & Richards, F.M. (1983) J. Mol. Biol. 193, 775-791; Hellinga, H. W., Richards, F.M. (1994) PNAS 91, 5803-5807; Desjarlais, J.R. & Handel, T.M. (1995) Prot Sci. 4, 2006-2018; Dahiyat, B.I. & Mayo, S.L. (1996) Prot. Sci. 5, 895-903).

For amino acids exposed on the surface, evolutionary pressure may play a greater role in determining the sequence selection than in the core regions where packing constraints lead to conserved amino acid selections. But having fewer physical constraints on the surface and highly variable charge and polar solvation interactions poses a challenging design problem for exposed side chains. This limitation restricts most protein design methods to the core of proteins because the steric constraints are major determinants in designing amino acids in these positions.

Some algorithms try to divide proteins into discontinuous regions

such as core, boundary and surface residues in order to have different scoring functions for different sites of protein structures (Dahiyat, B.I. & Mayo, S.L. (1996) Prot. Sci. 5, 895-903). However, for protein-protein interactions, the important residues are located on the surface of proteins, and most likely on the loops of proteins, the most difficult or irregular structure class of proteins. Upon interaction between proteins, some of the interacting residues become buried or half-exposed, making it difficult to model their interactions as specific class of residues in discrete regions of proteins. The inventors believe that although protein loops are widely involved in mediating protein-proteins interactions such as interactions between CDRs of antibodies and antigens or cytokines and their receptors, the methods existing in the art are still far from being capable of predicting the interactions, with high accuracy, of the loop structures of proteins by using force field-based approach alone, unless it is combined with a good homology model and database information (van Vlijmen HW, Karplus M (1997) J Mol Biol 267, 975-1001).

Given the inability of current force fields in predicting protein folding, a perpetual problem in protein folding and design is to develop an energy function that captures all factors known to contribute to protein stability, whose predictions compare favorably with experimental data. No matter how elaborate this procedure may be, calculating the small difference between two large numbers of stabilities for the folded and unfolded states of a protein is intrinsically difficult and error-prone. This difficulty becomes even greater if the region of interest lies at the interface between two proteins with the polar and charge residues whose forcefield parameters are still under active investigation for an accurate evaluation. The scoring function may also overfit the experimental feedback from a specific test system. In short, compared to the core packing inside proteins, accurate calculation of interactions between proteins that are dominated by polar and charged residues still remains a difficult task in this field. The inventor believes that side chain placing algorithms shown to be so effective in packing the hydrophobic core of proteins may not provide an effective solution to this standing problem.

The inventors stress that using the fixed backbone in the inverse

folding protocol also over-restricts the positioning of the side chain rotamers and the steric repulsion between them. Such stiff constraints on the side chain rotamers are unrealistic. A real protein would accommodate side chain mutations or rotamers through dynamic fluctuations in solution that is reminiscent of an altered ensemble of conformational states. It is noted that a parametric representation between regular secondary structural elements has been used to drive the systematic folding of protein backbones (Harbury, P.B., Tidor, B. & Kim, P.S (1995); Su & Mayo (1997) Prot Sci.; Harbury P.B. etc (1999) Science 282, 1462-1467). However, it is still difficult to use such an approach on non-regular secondary structural element such as a loop to account for the fluctuating ensemble states.

Given the limitations of the computational methods, impatient evolutionary protein designers have chosen to avoid the rational structure-based approach altogether and to invent a set of powerful experimental tools. But no matter how powerful, creating a diverse library by random mutagenesis and screening them by experiment is a highly inefficient process. On the other hand, recombination of homologous genes by DNA shuffling allows only a limited sampling of the sequence and structure space.

The inventors believe that a computational method that has no a priori physical limitations can search a much larger sequence space. In addition, a key advantage and the main driving force of the rational approach is the ability to design and control the sequence library at every stage prior to experimental screening. This allows the protein designer to make greater virtual jumps in protein sequence space that sample greater distances which might lead to discovery of novel sequences and structures that has little or no homology to the starting sequences. Additionally, the virtual size and direction of these "jumps" can be controlled in accordance with experimental feedback to follow the functional landscape to a new peak. This capability is expected to increase dramatically with increasing computational power and development of novel algorithms and new software tools.

Obviously, computational power will not by itself make the computational protein design superior to in vitro protein evolution experimental method unless the subtle but important structural

perturbations resulting from the directed evolution can be understood and captured. For example, it has been shown that the beneficial mutations are generally not localized to the catalytic sites but are distributed over large parts of proteins with perturbed protein backbone (Spiller B, Gershenson A, Arnold FH, Stevens R. (1999) PNAS 96, 12305-12310)

In the current art, the experimental screening for biological activities is still the only reliable approach available to evaluate the biological functions of molecules that are controlled by complicated competing factors under experimental conditions. It is extremely hard to correctly capture all the details simultaneously in a computational method and to pin point the answer without extensive experimental test. In addition, most of the scoring functions can only calculate the stability rather than activity or specificity.

Some statistics-based approaches have been developed that shed light on the evolutionary sequence design. Using a simplified model similar to the random energy model in protein folding, Bogarad and Deem have shown that DNA swapping of nonhomologous DNA segments with low energy structures is much more efficient in searching the fitness landscape in protein space than gene recombination of homologous DNA by DNA shuffling, which in turn is better than point mutations (Bogarad L, Deem MW (1999) PNAS 96, 2591-2595). Recently, a heteropolymer-based model has been used to explicitly map out the sequence-structure relationship in the fitness landscape in a structure-based evolutionary approach (Cui Y, Wong WH, Bornberg-Bauer E, Chan HS (2002) 99, 809-814). The point mutations are found to lead to diffusive walks on the evolutionary landscape, where crossovers can tunnel through barriers of diminished fitness. The smoothness of the energy or fitness landscape, together with the ratio between crossover and point-mutation rates, determines the effectiveness of crossovers in sampling the protein sequence and structure space. Thus, the inventors believe that evolutionary sequence design should not be limited to point mutations and homologous gene recombinations.

Experimental feedback is also essential to show any of the expected improvement in protein properties and to improve the

agreement between theoretical prediction and experimental test (Desjarlais, J.R. & Handel, T.M. (1995) Prot Sci. 4, 2006-2018; Dahiyat, B.I. & Mayo, S.L. (1996) Prot. Sci. 5, 895-903; Keating AE, Malashkevich VN, Tidor B, Kim PS (2001) PNAS 98, 14825-30). Thus, the inventors believe that unless the agreement between experimental and computational values are confirmed (Keating AE, Malashkevich VN, Tidor B, Kim PS (2001) PNAS 98, 14825-30) and demonstrated extensively, including polar and charged residues at various regions of different kinds of proteins, experimental library should not be limited to sequences around the global optimal or suboptimal solution from computation. Instead, the experimental library should be constructed to cover a wide range of distributions over the energy landscapes that score as good as or better than the lead sequence.

Some convergence between in vitro directed evolution and computational sequence design has begun to emerge. For example, the structure-based de novo designed enzymes are usually not very active (Benson, DE, Wisz, MS & Hellinga HW (2000) PNAS 97, 6292-6297; Bolon DN, Mayo SL (2001) PNAS 98, 14274-14279). But these de novo design of the sequences in a different scaffold can serve as a starting point and subject to directed evolution for activity improvement (Altamirano, MM, Blackburn, JM, Aguayo C, Fersht AR (2000) Nature 403, 617-622). Conversely, structure-based computational method can be used to identify potential sites for concentrated point mutations in evolutionary design in order to reduce the search space in directed evolution, although these sites are found to be different from those from sequence profiling. (Voigt CA, Mayo S, Arnold, FH & Wang Z-G (2001) PNAS 98, 3778-3783).

However, the inventors believe that the strategies for directed evolution should be analyzed and measured in quantitative terms before launching the laborious experimental work. Some steps have been taken to simulate the DNA shuffling computationally to optimize the possible experimental conditions and possible limits for enhancement (Moore, GL, Maranas CD, Lutz S, Benkovic S (2001) PNAS 98, 3226-3231). Given the huge protein space that can be searched by various approaches, it is important to compare the efficiency and limitations inherent to each experimental or computational approach in

order to determine the best route for the specific problem at hand.

The inventors also believe that, for structure-based protein design, the heart of the problem lies in the deterministic approach to a complicated problem with unrealistic assumptions. It is well known that interactions that stabilize a protein are very complex. The static structure used for design is an ensemble average of the dynamic fluctuations observed in solution that can change upon interacting with another protein or a ligand. Therefore, the idea of looking for the optimal solution to a target function is an interesting theoretical challenge but might be of little interest or practical relevance to real biological problems. Either the defect in energy function or the stringent restriction of using rigid backbone or both would contaminate the "optimal solution" to the design problem. Thus, again, the inventors believe that experimental library should not be limited to sequences around the global optimal or suboptimal solution from computation that might be biased by the assumption and parameters used in the computation. Instead, the sequences covering a preferred range that, for example, scores better than or equal to the lead sequence should be used for experimental screening.

For evolutionary protein design, current approaches to the design of proteins as biocatalysts (e.g., enzymes) still remain more art than science. But some methods are robust enough to be directly applied to solve real world problems in commercial biocatalyst design. Although DNA recombination by DNA shuffling and random mutagenesis have provided diverse protein libraries for functional screening, more efficient ways of library generation should be explored and the process should become predictable and routine rather than relying exclusively on the final screening results. So far, directed evolution has been applied most successfully to solve the biocatalyst design because it is easier to do high throughput screening for enzymatic activities where chemical reactions can be readily detected.

However, the inventors believe that the unexpected solution provided by directed evolution with mutations distributed throughout the entire protein sequence also poses problems for evolving certain proteins of pharmaceutical interest. In therapeutic antibody design, the mutations need to be limited to certain regions such as the CDR and

modifications to a previously inert framework regions may render the protein potentially immunogenic. Such undesireable mutants during experimental shuffling has to be minimized or reduced by tedious backcrossing procedure; hopefully removal of these immunogenic mutants will not negate the activity improvement earned by hard experimental effort.

The rational structure-based protein design has undergone fast evolution in its development and has begun to deliver some impressive results. Over the years, exciting progress has been made in computationally designing protein variants possessing the target scaffold (Dahiyat, B.I. & Mayo, S.L. (1997) Science 278, 82-87) and markedly improved thermal stability by repacking the hydrophobic core (Malakauskas, S.M. & Mayo, S.L. (1998) Nature Struct. Biol. 5, 470-475) and discovering novel scaffold not yet observed in nature (Harbury P.B. etc (1998) Science 282, 1462-1467). For biological activity and affinity design, some interesting progress has been made to extend this rational approach to affect binding affinity by designing residues around the binding sites in three different conformational states with open, apo- and closed ligand-binding states that can modulate the binding activity through an allosteric effect on the binding sites (Marvin, J.S. & Hellinga H.W. (2001) Nat Struct Biol 8, 795-798.). However, for most proteins of biological and medical interest, the structural information required for such design is still unavailable or at a low resolution insufficient for such design, although structural genomic project is promised to increase the structural information at an accelerated pace.

3) The Inventive Approach

The present invention provides an innovative approach to efficiently map out the distribution of the fitness and energy landscape in protein sequence and structure space by using ensemble-based statistical methods.

Given the incomplete knowledge of principles underlying protein folding and design, the ensemble-based statistical approach to protein combinatorial library seeks to design sequence ensembles that are compatible to a given structure or structure family, that cover a distribution of the energy landscape with scores better than that of the

lead sequence. It is statistical because it is the distribution of sequences or structures rather than a specific optimal solution to a given fixed structure that are designed. It is ensemble-based because it is structure/sequence ensembles that are targeted by nucleic acid libraries rather than a specific sequence or structure.

The inventors believe that partitioning of the energy distribution function into different ensemble states in sequence space allows for an effective sampling by subsequent experimental methods. This statistical approach to mapping the functional space of selected protein sequences provides a means to select protein sequences of real biological interest in the context of a fitness landscape described above. By defining the ensemble statistical properties rather than a single optimized sequence or a group of sub-optimal sequences, a protein designer is more likely to avoid getting trapped in a biased solution or move in a wrong direction resulting from the limitations inherent in current computational methods.

The inventive approach is developed by combining insights gleaned from theoretical studies of the simple models of protein folding and evolution based on the inventors' understanding of the problems associated with methods existing in the art. Through investigation and diligent experimentation, the inventors have developed practical solutions to the problems in the areas of protein folding, engineering and design, especially in the exciting field of antibody engineering.

Figure 2A schematically outlines an in silico biopolymer evolution system developed by the inventors. As shown in **Figure 2A-C**, the path from the initial target biopolymer (e.g., a protein) to the final candidate sequences with desired function(s) traverses in three spaces of biological importance: the sequence, structure and function spaces.

In the sequence space, the lead sequence(s) is employed to search the database(s) for evolutionarily related sequences. It is noted that this search may be applied to the structure space to obtain more distant sequences when structural alignment is used. The variant profile of the hit library describes the amino acid frequency and variants at the each position.

In the structure space, a hit variant library is generated in silico based on a reduced variant profile and partitioning (Figures 1C, 1D and

2A-C) or a complete sequence library or their random combinations (see Figures 1E-H, 2A and C). This hit variant library or random/complete sequence library is scored using a structural template, and preferred sequence ensembles are selected and re-profiled for the generation of an expanded nucleic acid (NA) library in silico. The size of the in silico NA library is evaluated and passed on for oligonucleotide synthesis if the library size is acceptable. Otherwise, the hit variant library is re-partitioned into smaller segments and smaller NA libraries are generated with overlapping sequences to maintain sequence and structural correlation among the resulting libraries (see Example section below and Figures 28A-C).

In the function space, the NA library is experimentally screened and positive sequences are input back into the computational cycle for library refinement. Strong positive clones are passed on for further evaluation and potential therapeutic development. If no hits occur in the experimental screening, new lead sequence ensembles in structure-based scoring and/or variant profile are selected for the target system and the process is restarted.

As can be appreciated from the depiction in Figure 2A, an important distinction between the approach described here from other methods in the field of computational and evolutionary sequence design is that the present invention combines the best from both worlds to explore the fitness landscape in sequence and structure spaces more efficiently. Our approach combines the evolutionary information in protein sequence database with the physical constraints such as compatibility of the sequences with the 3D structure of a protein. The biological function of proteins can be computationally evaluated through sampling of a limited set of sequences that satisfies both evolutionary selection in sequence space and physical constraints in structure space.

In a particular application of the inventive methodology, antibodies are utilized as a model system for both experimental and computational tests. Antibodies are widely used in research, diagnostics and medical application. Antibodies can bind a variety of targets with good specificity and affinity. Catalytic antibodies are also being developed to catalyze chemical reactions.

In a more particular application, antibody hypervariable loops or complementarity determining regions (CDRs) as well as the framework regions (FRs) are targeted. The CDRs determine antibody-antigen binding and specificity, whereas the framework regions provide the scaffold on which the CDRs are correctly positioned for biological function. The antibody molecule is well suited for engineering because of its modular structure, with CDRs and framework regions that are well defined sequentially and structurally.

As outlined in **Figure 1A** (Route I), polypeptide segments in an expressed protein database are computationally screened against a specific region (e.g., V_H CDR3) of a lead antibody to be optimized and those that match in their sequence patterns with that of the lead antibody are selected. The selected sequences form a hit library.

Furthermore, as outlined in **Figure 1B** (Route II), a variant profile can be generated by listing amino acid variants at each sequence position from the hit library, together with the number of the occurrence in the hit library. The combinatorial enumeration of this profile represents the hit variant library I. This variant profile can be edited either by including amino acids from the lead sequence or sequence profile at the corresponding positions where they are missed from the hit library or by eliminating amino acid variants that occur below a certain cut off frequency, or both. The resulting variant profile defines the hit variant library II, the designed library.

As outlined in **Figures 1C** and **1D**, each member of the hit variant library I or II is "grafted" onto the corresponding region of the lead antibody template structure or model, if available, and selected, using a scoring function, for ones that are structurally compatible with the rest of the 3D structure. Optionally, the hit variant library can be evaluated in the presence or absence of a target antigen. Antibodies with favorable scores are selected and screened experimentally in a laboratory for their actual binding affinity towards the antigen. As will be shown in the EXAMPLE section, a large number of antibodies against human vascular endothelial growth factor (VEGF) are selected using this approach and proven to be able to bind to the target antigen VEGF. Some of them show affinity higher than that of the lead antibody (see **Figures 30 & 36**).

As will become more apparent with further disclosure in the sections below, the approach provided by the present invention is not only conceptually distinguishable from those in the art but also possesses many practical advantages in antibody engineering.

By exploiting the expressed protein sequences compiled in the protein databases, this approach, not only effectively mimics the natural process of affinity maturation in silico, but can potentially drastically hasten the evolution of proteins with improved binding affinity. For example, any set of amino acid sequences, including but not limited to sequences of immunological interest, from various species can be used to maximize the library diversity for profiling against a lead sequence for CDR affinity maturation. However, sequences of human germlines and/or origins should be used for profiling against a lead sequence for framework regions for humanization or framework design in order to minimize potential immunogenicity. Thus, the choice of the database, based on their application, sizes and origins of species such as human, mouse, etc. or all species available, permits the flexibility and control on the design proteins.

Further, the approach optionally includes modeling of the protein mutant (e.g., a mutant of the lead antibody) in the presence of the target molecule (e.g., the antigen of the lead antibody) if the complex structure or a model is available. By including the interaction between the antibody and antigen in the calculation, the screening process more closely mimics the natural process of affinity maturation, as an antigen-directed process, and the calculated binding affinities may correlate better with experimental values.

Moreover, the method of the present invention combines computational prediction of an antibody library, which is biased toward a specific target molecule or antigen, if the complex structure or structure model is available, with experimental screening of the library to select for those with high binding affinity to the antigen. Such a process can be reiterated to improve the binding affinity of selected antibodies. Given the availability of a high affinity complex structure as a template, the hit variant library can be computationally pre-screened to reduce the library size, yet remain functionally highly focused compared to traditional libraries generated through complete

randomization of amino acids in each position of the lead antibody. Through prediction and construction of the hit variant library in silico, the whole process of protein evolution can be hastened, effectively mimicking the natural process of antibody affinity maturation in a high throughput manner.

In a preferred embodiment, the lead protein is an antibody or immunoglobulin and the target molecule is an antigen that binds to the template antibody. It should be noted that the lead protein may be any protein, preferably a protein with known three-dimensional structure which may be resolved using X-ray crystallography or nuclear magnetic resonance spectroscopy. Alternatively, the 3D structure or structure ensembles of the template protein may be provided by computer modeling using algorithms known in the art.

4) Comparison of the Inventive Methods with Others in Antibody Selection and Engineering

It is understood that selection of antibodies from a highly diverse library allows for a broad coverage of sequences, thereby maximizing the chance of finding the optimal sequence(s). However, for antibody sequences that are derived from random mutagenesis of the lead antibody, for example, in the CDRs, not all structures of the randomized CDRs are compatible with the 3D structure of the lead antibody. By using expressed proteins sequences as opposed to those from random mutagenesis and filtering out the incompatible sequences using the inventive method, a fewer number of sequences are selected. As a result, the sequence space of an antibody to be screened is reduced in size without losing sequences that may be highly relevant to affinity binding maturation and stabilization of the mutant antibody.

In contrast, the current methods in the art for constructing an antibody library involve *in vitro* isolation of cDNA libraries from immunized human antibody gene pool, naive B-cell Ig repertoire, or particular germline sequences. Barbas and Burton (1996), supra; De Haard et al. (1999), supra; and Griffiths et al (1994), supra. These libraries are very large and extremely diverse in terms of antibody sequences. Such a conventional approach attempts to create a library of

antibody as large, and as diverse as possible to mimic immunological response to antigen in vivo. Typically, these large libraries of antibody are displayed on phage surface and screened for antibodies with high binding affinity to a target molecule. Such a "fishing in a large pond" or "finding a needle in a huge hay stack" approach is based on the assumption that a simple increase in the size of sequence repertoire should make it more likely to fish out the antibody that can bind to a target antigen with high affinity, but, in practice, is inefficient for affinity maturation due to inadequate sampling, insufficient diversity and indeterminate library composition.

The inventors believe that there are several problems associated with such a conventional approach. A simple increase in the size of sequence library may not necessarily correlate with an effective increase in functional diversity. Further, due to the physical limit on making an extremely large experimental library, it may be very difficult to construct a library with diversity over 10^{11} in vitro. The library that is actually screened experimentally probably presents only a fraction of the sequence repertoire at the theoretically predicted size. In addition, there is a legitimate concern that, with the difficulties and the under-representation problems associated with handling and manipulation of an extremely large library in vitro, time and money may be lost in an effort trying to increase the size of the library and yet not significantly increasing the functional diversity.

Another approach existing in the art is to design an artificial antibody library computationally and then construct a synthetic antibody library which is expressed in bacteria. Knappik et al., *supra*. The artificial antibody library was designed based the consensus sequence of each subgroup of the heavy chain and light chain sequences according to the germline families. The consensus was automatically weighted according to the frequency of usage. The most homologous rearranged sequences for each consensus sequence was identified by searching against the compilation of rearranged sequences, and all positions where the consensus differed from this nearest rearranged sequence were inspected. Furthermore, models for the seven V_H and seven V_L consensus sequences were built and analyzed according to their structural properties.

However, there are a few problems concerning such an approach as far as therapeutic applications of the selected antibody are concerned. The definition of consensus sequence may be too arbitrary and such artificial sequences defined may not be representative of a natural, functional structure, although experimental test and structural analysis may eliminate some unfavorable amino acid combinations. Although the consensus sequences may be designed to cover mainly those human germline sequences that are highly used in rearranged human sequences, it might bias the consensus sequence library toward a limited number of antigens exposed to human being so far in the course of evolution. Although these library construction method is mainly focused on finding a lead antibody or hit from a large antibody library, for the affinity maturation, most of the approach described above still quite limited for antibody affinity maturation. More tranditional approach such as CDR walk, random mutagenesis, or stepwise saturated mutagenesis at each position of CDRs etc are used for antibody affinity maturation. The present invention is specifically tailored to designing biased library for affinity maturation.

The inventors believe that sampling the functional space by mapping structures from different species covers a wider range of functional CDRs in an antibody library and will expand the range of antigens it can bind. This approach would be very important in the design of antibody libraries to target novel antigens. The method of the present invention typically relies on structural constraints derived from antibodies or from other natural sources. According to the present invention, a complete sequence space of all proteins available, preferably antibodies, including those from both human and other species, can be analyzed by fitting each library sequence into the 3D structural framework of the lead antibody.

Based on this analysis, the resulting mutant antibodies are not only novel in their sequences but also possess higher affinity than that of the lead antibody. As shown in the section of EXAMPLE below, a large number of mutant antibodies are selected using the inventive method and experimentally proven to bind to human VEGF with affinity similar to or higher than the lead anti-VEGF antibody.

2. General Description of Procedures Employed to Implement Protein Design Strategies of the Present Invention

The procedures involve the exploration of sequence, structure and functional spaces and the evaluation of the relationships among them (**Figures 1A-D, 1E-H, 2A-C**). Starting point can be either a lead structure or a lead sequence or both, if available. The procedure systematically explores both the sequence space and structure space in order to identify variant profiles optimized for functional screening. There are three modes of information exchange: i) separate evaluation of information in sequence and/or structure space and then combined, ii) consecutive evaluation from sequence to structure, or from structure to sequence, or iii) from sequence or structure alone. While the sequence design can be explored in sequence and structure spaces separately (two separate cycles), the variant profiles from these two separate cycles can be compared and combined in order to arrive at the optimal overall variant profile with good consensus variant profile that is likely to produce strong candidates in the functional screen.

The two starting points are interrelated operationally because a sequence profile may be derived as a result of comparing the target sequence with homologous sequences or through structural alignment of known homologous structures. Sequence profiles may also be derived from mutational data that suggest functional or structural information. Similarly structure ensembles may be generated through molecular dynamic simulations but can also be derived from sequence alignments of know structures or from homology-based modeling.

The two filtering and refining cycles in sequence and structure spaces are further linked during the filtering and evaluative steps because the variant profiles arrived by each cycle are compared and/or passed to the other cycle for further refinement. For the sequence-derived variant profile, it is structurally evaluated on a known template in structure space in order to rank and refine the variant profile. Conversely, the structure-derived variant profile can be passed on to the sequence space to evaluate if they belong to the same superfamily of the hit or variant library or for comparison and partitioning to control the final library size.

1) Sequence Space

In sequence space, the goal is to determine the variant profile that is optimized for the target function. The cycle begins with the identification of the hit library through database sequence search and alignment using the sequence profile. This may be a simple BLAST search or a probabilistic approach such as profile HMM. Based on the variations within the hit library, the sequence can be filtered and partitioned. This is achieved by evaluating the amino acid frequency and distribution at each position. Commonly, the residues with the highest frequencies at each position as well as the residues from the target sequence are included in the variant profile. A cutoff value, such 5% or higher, depending on the distribution of the variant frequency, or amino acids ranked relatively higher at each position can be included in the variant profile.

Partitioning may be necessary to set a practical limit on the final size of the oligonucleotide library. Partitioning can be determined by calculating the size of the oligonucleotide library as a function of the degenerate nucleic acid library of the various variant profile segments. Thus, a highly variable variant profile can be partitioned so that the size of the resulting oligonucleotide library can be set within the limit for effective and efficient experimental synthesis, transformation and screening.

An alternative partitioning scheme is to employ structural correlation information. Since peptides folding in three-dimensions interact among sequentially distant segments, a structural template or a model can be used to assign structurally correlated sequences for partitioning. For instance, the ends of the loop may be correlated while the apex itself is relatively free of interactions with the ends. In such a case, the variant profile can be partitioned into at least two profiles: one for the two ends and one for the apex.

Either or both approaches can be employed in partitioning a highly variant profile. When partitioning, there should be at least 2, preferably 3 or more residue overlaps between the segments so that some structural correlation is maintained between adjacent segments. Either or both approaches can be employed to achieve operationally

optimized oligonucleotide library sizes.

Once the sequence variant profile is determined, its library is computationally screened using a known structural template or a homology-based model and a scoring function (see below). This ranking is used to filter and reduce the variant profile by identifying favorable variants while filtering out unfavorable variants, thereby simultaneously enriching and reducing the size of the experimental library.

2) Structure Space

In structure space, the goal also is to determine the variant profile optimized for the target function but starting with one structure or an ensemble of structures and then scoring the sequences based on the average of the ensemble of structures. The cycle begins with a set of structures and associated sequences that can be computationally screened and evaluated using a scoring function.

For a theoretical, ideal scoring function that accounts for all physicochemical variables, the energy score ranking would correlate perfectly with the functional ranking. This is neither possible nor computationally practical and one must use an imperfect scoring function that will coarsely correlate structure or sequence with function. Since the goal of the design protocol is to identify a set of probable sequences that will possess the desired function, an imperfect scoring function that, nevertheless correlates sequence and structure with function, can be used.

Such a scoring function can involve any combination of computational terms that correlates or maps functional values to a sequence or structural value. A simple case is that of a van der Waals energy that correlates hydrophobic packing function with sequences containing the appropriate density of aliphatic or aromatic sidechains. Another might be an enzymatic hydrolytic activity that correlates with the existence of a nucleophilic sidechain group at a particular position in a sequence.

In general, the scoring function will be based on thermodynamic energy sum that incorporates some or all of the contributing terms that correlate with the structural stability and function of the protein. Most commonly, these will include the electrostatic solvation energy,

nonpolar solvation energy and sidechain and backbone entropy. MM-PBSA or MM-GBSA is such a method that combines standard terms calculated using molecular mechanical (MM) forcefields with the solvation terms including electrostatic solvation with continuous solvent model, calculated either by solving the Poisson-Boltzmann (BP) equation or using the Generalized Born (GB) approximation, and solvent-accessible solvation term, based on proportionality to the surface area (SA), together with contribution from the conformational entropy, including backbone and side chains. Good correlation between experimental and MM-PBSA calculated values based on the ensemble structures derived from molecular dynamic simulation has been reported (Wang W, Donini O, Reyes CM, Kollman PA. (2001) Annu Rev Biophys Biomol Struct 30, 211-43). The refined scoring function based on the MM-PBSA was used to evaluate the simple scoring function based on the total energy of Amber94 forcefield implemented in CONGEN, which was used to scan a sequence library for its compatibility with a template structure (see for example, **Figure 12**). The comparison between the simple scoring function used here and the refined scoring function for a hit library of the lead sequence using one template structure (1cz8) (**Figures 12D & E**) suggests that the simple scoring function is correlated with the refined scoring function, although significant scattering in the correlation map suggests that some refinement in the simple scoring function can be done to improve its agreement with the refined scoring function.

Compared to other scoring functions used in protein and drug design, MM-PBSA or MM-GBSA is a better physical model for scoring and would handle various problems on an uniform basis, although it is computationally expensive because multiple trajectories from molecular dynamic simulation in explicit water is required to calculate the ensemble averages for the system. This method is useful for studying some of the difficult mutants beyond the simple scoring procedure, and can serve as a control to validate the procedure used in high-throughput computational screening.

3) *Optimized Variant Profile*

The first result of the design protocol is the optimal variant

profile. It embodies the results of both the sequence and structure evaluations so that evolutionary and structural preferences are incorporated into the design. Subsequent steps in the functional space aim to evaluate and refine this profile, and, if necessary, modify earlier steps, so that cyclic enrichment of the resulting library can be accomplished at various steps in the design protocol.

In a preferred embodiment, the method comprises:

the method comprises the steps of:

a) providing an amino acid sequence of the variable region of the heavy chain (V_H) or light chain (V_L) of a lead antibody, the lead antibody having a known three dimensional structure;

b) identifying the amino acid sequences in the CDRs of the lead antibody;

c) selecting one of the CDRs in the V_H or V_L region of the lead antibody;

d) providing an amino acid sequence that comprises at least 3 consecutive amino acid residues in the selected CDR, the selected amino acid sequence being defined as a lead sequence;

e) comparing the lead sequence with a plurality of tester protein sequences;

f) selecting from the plurality of tester protein sequences at least two peptide segments that have at least 10% sequence identity with lead sequence, the selected peptide segments forming a hit library;

g) building an amino acid positional variant profile of the hit library based on frequency of amino acid variant appearing at each position of the lead sequence;

h) combining the amino acid variants in the hit library to produce a combination of hit variants which form a hit variant library;

i) determining if a member of the hit variant library is structurally compatible with the lead structural template using a scoring function;

j) selecting the members of the hit variant library that score equal to or better than the lead sequence;

k) constructing a degenerate nucleic acid library comprising DNA segments encoding the amino acid sequences of the selected members of the hit variant library;

l) determining the diversity of the nucleic acid library, if the diversity is higher than $1x10^6$, repeating steps j) through l) until the diversity of the diversity of the nucleic acid library is equal to or lower than $1x10^6$;

m) introducing the DNA segments in the degenerate nucleic acid library into cells of a host organism;

n) expressing the DNA segments in the host cells such that recombinant antibodies containing the amino acid sequences of the hit library are produced in the cells of the host organism;

o) selecting the recombinant antibody that binds to a target antigen with affinity higher than 10^6 M^{-1}; and

p) repeating steps e) through o) if no recombinant antibody is found to bind to the target antigen with affinity higher than 10^6 M^{-1}.

As shown in **Figure 2B,** the method is executed starting from the target sequence or sequence profile based on structure-based multiple alignment, searching for variant profile based on evolutionary enriched sequence database, and then evaluating their compatibility with structure template or ensembles, and then selecting sequence ensembles that can be targeted experimentally. This procedure has been exemplified in our examples. First, it utilizes the evolutionary information encoded in sequences or their combinations including expression, folding, etc. that are not yet captured in theoretical calculations. Second, after removing a lot of unrelated random sequences, structure-based screening for the resulting library is amenable to refined computational screening. Also refined computational scoring such as MM-PBSA can be applied to some of them using ensemble structures. The inventors believe this procedure tends to give highly refined sequence library for experimental screening with significant savings in time and cost.

Figure 2C illustrates another embodiment of the method. The method comprises the steps of:

a) providing an amino acid sequence of the variable region of the heavy chain (V_H) or light chain (V_L) of a lead antibody, the lead antibody having a known three dimensional structure which is defined as a lead structural template;

b) identifying the amino acid sequences in the CDRs of the lead antibody;

c) selecting one of the CDRs in the V_H or V_L region of the lead antibody;

d) providing an amino acid sequence that comprises at least 3 consecutive amino acid residues in the selected CDR, the selected amino acid sequence being defined as a lead sequence;

e) mutating the lead sequence by substituting one or more of the amino acid residues of the lead sequence with one or more different amino acid residues, resulting in a lead sequence mutant library;

f) determining if a member of the lead sequence mutant library is structurally compatible with the lead structural template using a first scoring function;

g) selecting the lead sequence mutants that score equal to or better than the lead sequence;

h) comparing the lead sequence with a plurality of tester protein sequences;

i) selecting from the plurality of tester protein sequences at least two peptide segments that have at least 10% sequence identity with lead sequence, the selected peptide segments forming a hit library;

j) building an amino acid positional variant profile of the hit library based on frequency of amino acid variant appearing at each position of the lead sequence;

k) combining the amino acid variants in the hit library to produce a combination of hit variants;

l) combining the selected lead sequence mutants with the combination of hit variants to produce a hit variant library;

m) determining if a member of the hit variant library is structurally compatible with the lead structural template using a second scoring function;

n) selecting the members of the hit variant library that score equal to or better than the lead sequence;

o) constructing a degenerate nucleic acid library comprising DNA segments encoding the amino acid sequences of the selected members of the hit variant library;

p) determining the diversity of the nucleic acid library, and if the diversity is higher than $1x10^6$, repeating steps n) through p) until the diversity of the diversity of the nucleic acid library is equal to or lower than $1x10^6$;

q) introducing the DNA segments in the degenerate nucleic acid library into cells of a host organism;

r) expressing the DNA segments in the host cells such that recombinant antibodies containing the amino acid sequences of the hit library are produced in the cells of the host organism;

s) selecting the recombinant antibody that binds to a target antigen with affinity higher than 10^6 M^{-1}; and

t) repeating steps e) through s) if no recombinant antibody is found to bind to the target antigen with affinity higher than 10^6 M^{-1}.

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4) Function Space

In functional space, the goal is to express and screen the library derived from the optimized variant profile. There are two components that comprise the function cycle. An operational component that may not directly affect function but is important in the expression of the protein is the optimization of the oligonucleotide. The determination of the practical limit on the size of the oligonucleotide library is used as a guide to sequence partitioning and reprofiling of the variants.

The other component is the functional screen that directly reflects the results of all previous steps and is the final evaluative portion of the design strategy. The results of the experimental functional screen determine whether the library candidates can be passed on for further evaluation or used to enrich and refine the libraries from previous steps. For instance, a set of sequences exhibiting varying levels of function can be used to narrow the variant profile or to give weights to different residues at indicated positions. In addition, sequence space jumps through the use of degenerate oligonucleotide design may lead to the identification of a novel functional variant that can be used to further enrich the optimized variant profile. Alternatively, the frequency of a particular set of amino acid may reflect either a functional preference of expressional preference. In the latter option, a low expressing sequence that, nevertheless, exhibit good

function may prompt a modification in the codon usage that can improve expression levels while maintaining function. It is important to select some second or third "tier" variants, ones that occur at lower frequencies, since selecting only the highest frequency variants only leads closer to concensus and likely leads to "average" functioning sequences. It is possible that exceptional variants are likely to come from combinations not observed in nature. While we use natural evolutionary patterns as our guide, we look for combinations not observed in nature, either because they are unfavorable in evolutionary time scale but possibly useful for our more immediate applications, or, perhaps, because nature has yet to try them out. In this regard, structure-based sreening of random mutants or their combinations would potentially yield those mutants are yet observed in nature but nevertheless preferred structurally, although this puts stringent requirements on accuracy of the structure and potential functions as well as computational speed.

5) Iteration, Refinement, and Enrichment

The design protocol is divided according to different spaces that are evaluated but all the operational cycles are inter-related and integrated so that information can be exchanged and cycled freely to and from any space in order to continually refine and enrich the library based on the optimized variant profile. As a result, the pathway from target sequence or structure to candidate sequences is not a single pathway but a series of oscillations among the three cycles, each improving the selection in the optimized variant profile.

In addition, functional evaluation and iterative nature of the design protocol not only help improve the variant selection but also help increase the accuracy of the scoring function, at least for the range of sequences and structures examined. A missed prediction may indicate incompatible template. It may also indicate that a particular contribution may need to be more heavily weighed, for instance, backbone entropy in the context of glycine preference in functional screen. A particular charged residue such as Arg versus Lys in V_H CDR3 may be favored because of its role in orientating a specific conformation (see example section below).

6) Re-profiling of sequences according to scores and ranking

As described above, sequences in the hit variant library can be evaluated based on their structural compatibility with the lead antibody in the presence and absence of the antigen. According to the scores and rankings obtained from the structural evaluation, the sequences in the hit variant library are re-profiled to optimize the sampling of the sequence and structure space for functional sequences. This step involves the selection of a sub-population of the hit variant library that scores better than the lead sequence(s) and re-profiling them to generate an optimized library. One option is to re-profile all of the sequences scoring better than the leads. However, this is likely to lead to too large a library for experimental screen. A preferred way is to select a subset of sequences in a certain low energy window or several such subsets (**Figure 7**). This will reduce the eventual size of the experimental nucleic acid library as will be described in the section below and outlined in **Figure 6**. When combined with rational selection and design, this step should enrich the library with better scoring sequences.

The modification and optimization of the profile must take into account the ultimate size of the physical nucleic acid library (**Figure 6**). One strategy is to re-profile the best scoring 10-20% of the hit variant library to limit the number of positional variants within certain limit that can be easily targeted in experiments (preferably < 10^6 for degenerate nucleic acid library). Similarly we might select a set of low energy sequences that contain desired amino acids in certain positions.

7) Partitioning of sequences into fragments

Another size controlling strategy is to partition the sequences based on structurally correlated and uncorrelated fragments in structure space. These parsed sequences with the smaller variant profiles can be used for generating several smaller libraries. The rationale for this is that, to a first approximation, structurally distant segments are often uncorrelated so that mutations widely separated can be treated independently, whereas those fragments that couple with

each other in space should be targeted simultaneously by the combinatorial nucleic acid libraries. In the case of loops, the sequences forming the base of the loop are generally correlated due to loop closure, but the apex is often uncorrelated from the base of the loop. In such a case the amino acid sequence variant profile is partitioned into three segments and the first and third segments (base of the loop) are used for one profile and library design and the second segment (apex of the loop) is used for the second profile and library design. There should be 2 or 3 positional overlaps between the fragments to maintain a small level of structural correlation among the resulting libraries. In a similar fashion, a longer profile can be partitioned into a chain of overlapping segments to span the length of the sequence and corresponding libraries generated. Simple criteria such as the C_α or C_β distance matrix can be examined to identify correlated segments (Figure 28A). Optionally, a more detailed interaction matrix can be mapped out to explore numbers and types of interactions, but the underlying principle is the same for identifying correlated segments.

The resulting re-profiling can be further modified and enhanced based on observed experimental or structural criteria. These can include varying positions with known hydrogen bonds with additional polar amino acids, region of high van der Waals contacts with bulky aliphatic or aromatic groups, or region which might benefit from increased flexibility with glycine. In an experimental feedback, variants may be added based on assay results from earlier screening as a basis for subsequent design improvement. A more sophisticated analysis might take into account the coupling of amino acid groups such as salt bridges or hydrogen bonds within the sequence. Additional design constraints might include solvent accessible surface area of nonpolar groups of proteins.

With the modified and optimized profile, we generate a new amino acid sequence library designated the "hit variant library II" or a group of libraries (hit variant library IIA, IIB, IIC, etc) and score these using the same energy function. The energy distribution should expand beyond the original energy window since variant recombination and profile modification are intended to expand the sequence and structure

space covered (**Figures 7, 13A, 17A, & 18**).

Various embodiments of the inventive methodology are described in detail as follows.

3. Construction of Hit Antibody Library in silico

As illustrated in **Figure 1A**, a hit library can be constructed in silico based on the lead sequence from a region of the lead antibody. Sequences from a database of protein sequences, such as genbank of the NIH or the Kabat database for CDRs of antibodies, are searched based on their alignment with the lead sequence by using a variety of sequence alignment algorithms.

Figure 3 illustrates an exemplary procedure for constructing the hit library, which begins with a search of a protein sequence database of varying identity with the lead sequence or sequence profiles. The lead sequence profile is generated by aligning sequences within the same family of a structural motif. This lead sequence profile can be used to build the HMM to search the sequence database for hit libraries of remote homology to the lead sequence. This approach is taken to find a rich pool of diverse hit sequences (i.e., the hit library) to ensure that all available variants of the lead sequence from the database are included.

The database screened against the lead sequence(s) preferably includes expressed protein sequences, including sequences of all organisms. More preferably, the protein sequences originate from mammals including humans and rodents if the frameworks are targeted. Optionally, the protein sequences may originate from a specific species or a specific population of the same species. For example, the protein sequences collected from a human immunoglobulin sequence database can be used to construct the library of polypeptide segments. Compared to the conventional way of building the library using completely random protein sequences, this approach of the present invention takes advantage of the sequence information derived from the evolution of proteins, thus more closely mimicking the natural process of antibody generation and affinity maturation.

Depending on the region/domain of the protein to be designed, databases of proteins with different evolutionary origins may be exploited. For example, to reduce human immunogenicity of the design antibody, sequences of human origins, more preferably germline sequences, are used for the design purpose. On the other hand, to increase the diversity in the CDRs, extensive sequence search and selection from a wide range of databases and/or structure-based design procedures may be employed to increase the structural and/or functional diversity. Through such sequence and structure-based selections, rare combination of sequences may be found in the CDRs while the sequences in the framework regions are kept as close to the human sequence family as possible.

In addition, some combinations of amino acid residues from sequences of diverse species including human or other non-human species including but not limited to mouse, rabbit, etc., may be preferred at certain regions such as the boundaries between CDRs and frameworks in antibodies. This approach may be taken in order to maintain or optimize the relative orientations among various motifs.

Many sequence alignment methods can be used to align sequences from the database with the lead sequence (or lead sequence profiles) ranging from a high to low sequence identity. A number of sequence-based alignment programs have been developed, including but not limited to Smith-Waterman algorithm, Needleman-Wunsch algorithms, Fasta, Blast, Psi-Blast, Clustalx, and profile Hidden Markov Model.

Optionally, a simple sequence search method such as BLAST (Basic Local Alignment Search Tool) can be used for searching closely related sequences (e.g., > 50% sequence homology). BLAST uses a heuristic algorithm with position-independent scoring parameters (e.g., BLOSUM62 etc) to detect similarity between two sequences and is widely used in routine sequence alignment (Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ (1990) J Mol Biol 215, 403-410). However, the BLAST analysis may be too restrictive to detect remote homologues of the lead sequence. More advanced tools for sequence alignment can be used to search for remote homologues of the lead sequence.

A profile-based sequence alignment method may be used to

search for the variants for the lead sequence, such as PSI-BLAST (Position-Specific Iterated BLAST) and HMM. These profile-based sequence alignment methods can detect more remote homologues of the lead sequence (Altschul, SF, Madden, TL, Schaffer AA, Zhang J, Zhang Z, Miller W, Lipman DJ (1997) Nucleic Acids Res 25, 3389-3402; Krogh, A, Brown M, Mian SI, Sjolander Km Haussler D (1994) J. Mol. Biol 235, 1501-1531).

PSI-BLAST is a new generation BLAST program belonging to the profile-based sequence searching methods (Altschul, SF, Madden, TL, Schaffer AA, Zhang J, Zhang Z, Miller W, Lipman DJ (1997) Nucleic Acids Res 25, 3389-3402). PSI-BLAST automatically combines the statistically significant alignments produced by BLAST into a position-specific matrix to score sequence alignment in the database. The newly searched sequences are incorporated into the position-specific scoring matrix to start another round of sequence search in the database. This procedure is iterated until no new hits are found or the pre-set criteria are met. Although PSI-BLAST may not be as sensitive as the Profile Hidden Markov Models (HMM), it can be used in the present invention because of its speed and ease of operation in the absence of a pre-built motif profile. The

Profile Hidden Markov Models or HMM are statistical models of the primary sequence consensus of a given sequence or sequence alignment family. The sequence family is defined as the multiple sequence alignment resulting from the corresponding multiple sequence and/or structure alignment. The formal probabilistic basis underlying HMM makes it possible to use Bayesian probability theory to guide the setting of the scoring parameters based on the profile of aligned sequences. This same feature also allows the HMM to use a consistent approach, using the position-dependent scores, to score the alignment for both amino acids and gaps. These features in HMM make it a powerful method to search for remote homologues compared to the traditional heuristic methods (Eddy S.R (1996) Curr Opin Struct. Biol 6, 361-365). The pattern in the primary sequence can be detected by the pattern recognition algorithms and therefore can be used to pull out more members related to the target sequence (when one sequence is used) or sequence profile (when multiple sequence alignment is used).

To capture the higher order correlation in a sequence, or the interactions between amino acids in three-dimensional space, the multiple sequence alignment resulting from multiple structural alignment is a preferred method to be used in the present invention to generate the hit library.

Optionally, a structure-based sequence alignment may be used to search for a highly diverse hit library. This method is advantageous because it is a gold standard that can be used for comparing various multiple sequence alignments in the absence of any detectable sequence homology (Sauder JM, Arthur JW, Dunbrack RL Jr (2000) Proteins 40, 6-22). The multiple structure alignment can directly yield the corresponding multiple sequence alignment. Alternatively, these closely related structures can be used as structural templates for sequence threading to generate the multiple sequence alignment profile (Jones DT (1999) J Mol Biol 1999, 797-815). Methods combining multiple sequence and structure alignments have been reported to annotate the structural and functional properties of known protein sequences (Al-Lazikani B, Sheinerman FB, Honig B (2001) PNAS 98, 14796-14801).

Also optionally, a reverse threading process may be used to search for of a highly diverse hit library. A reverse threading process is the counter part of the threading process. Threading is a process of assigning the folding of a protein by threading its sequence (i.e., the query sequence) to a library of potential structural templates by using a scoring function that incorporates the sequence side chain interactions as well as the local parameters such as secondary structure and solvent exposure. The threading process starts with a prediction of the secondary structure of an amino acid sequence and solvent accessibility for each residue of the query sequence. The resulting one-dimensional (1D) profile of the predicted structure is threaded into each member of a library of known 3D structures. The optimal threading for each sequence-structure pair is obtained using dynamic programming. The overall best sequence-structure pair constitutes the predicted 3D structure for the query sequence.

In contrast, reverse threading is a process of searching for the optimal sequence(s) from sequence database by threading them onto a

given target structure or structure cluster ensembles of the target structure. Various scoring functions may be used to select for the optimal sequence(s) from the library comprising protein sequences with various lengths.

For example, amino acid sequences from a human germline immunoglobulin database can be threaded onto the 3D structure of the lead antibody to search for the sequences with acceptable scores. The selected sequences constitute the hit library. The reverse threading process is the opposite of the threading process in that the former tries to find the best sequences fitting to the target structural template whereas the latter finds the best 3D structure structures that fit the target structure profile.

Additionally, the top hits of the sequences found for the lead antibody may be profiled by reverse threading multiple amino acids at each position in a combinatorial approach to select for the best "consensus" combinatorial sequences compatible with the 3D structure of the lead antibody. This process of searching for a consensus sequence is different from the method of using simple sequence average at each position described in Knappik, et al (2000). The consensus sequence according to the present invention is created using the structurally-based reverse engineering approach using all possible combination of amino acids that are allowed at each position, based on the retrieved sequences and optimized by scoring their compatibility with the structural template.

In addition to the methods used for sequence alignment, the sequence motif and the corresponding database used in the sequence alignment are also of critical importance in the present inventive method. The sequence or sequence profile used here are defined based on structural analysis of the protein functions for antibody regions, such as the CDR motifs (CDR1, CDR2 and CDR3) for antigen binding and the framework regions (FR1, FR2, FR3 and FR4) for supporting the antibody scaffold. As an example, Genbank and Kabat databases can be used to search for sequence hits from various species to increase the diversity of the hit library matching the CDRs of antibodies in order to maximize the binding affinity of a designed antibody. On the other hand, human or even human germline sequence database is preferably

used to search for sequence hits for framework design in order to decrease the chance of creating immunogenic epitopes of non human origins in a designed framework. This sequence selection step allows for maximum flexibility and control of the sequence source for design, especially when considering the eventual therapeutic application of the designed antibody.

The hit library can be refined further by eliminating redundant sequences and re-profiled to get a more accurate HMM or PSI-BLAST profile. As described in detail in the Example section, the V_H CDR3 sequence, according to the Kabat classification (and also the structure motif) of a humanized anti-VEGF antibody with or without a few residues flanking them at N- or C-termini, was used as the lead sequence. The utilities (hmmbuild, hmmcalibrate, hmmsearch, hmmalign) in HMMER 2.1.1 software package with default setting (Eddy S in http://hmmer.wustl.eduhttp://hmmer.wustl.edu) were used to build the HMM model, to calibrate HMM model against synthesized random sequences, to search the database for hit sequences and align them. Only hit sequences with the same length to the lead sequence are used for alignment and variant profile. Insertion or deletion in aligned sequences can be also used to profile the variants at aligned positions.

As illustrated in **Figure 3**, when a single lead sequence of the V_H CDR3 sequence of the anti-VEGF antibody was used as HMM to search the Kabat database, 108 unique sequences were found with sequence identity ranging from 40 to 100% relative to the lead sequence (**Figure 10A & 19C**). When a multiple aligned sequence profile of this lead sequence was used as HMM to search the same Kabat database, 251 unique sequence hits were found with sequence identity ranging from 15 to 100% to the lead sequence (**Figure 19C**). These results show that a profile HMM can find sequences with remote homology to the lead sequence. Thus, a sequence profile derived from the multiple structure-alignment would extend the diversity of the hit library.

Sequences of the hit library also depend on the database used. For example, by replacing the Kabat database with Genpept in the above, hits that are different from those in Kabat database were found either when the single lead sequence was used as HMM or when the

structure-based sequence profile was used as HMM.

The sequences in the hit library constructed by searching the databases can be analyzed (e.g., by profiling based on the positional frequency of each amino acid residue) and used directly for screening in vitro or in vivo for the desired function(s). See Route I in **Figure 1A** and **Figure 3**.

Optionally, the sequences in the hit library are profiled and used to construct a hit variant library I which is then screened in vitro or in vivo for the desired function(s). See Route II in **Figure 1B** and **Figure 4**.

Also optionally, the hit library is filtered based on the scoring of their compatibility with the lead structural template using methods such as reverse threading or forcefield-based full atom representation. Based on the resulting ranking of the scores, a hit variant library II is selected for screening in vitro or in vivo for the desired function(s). See Route III in **Figure 1C** and Figure 5.

Also optionally, the hit variant library I is filtered based on the scoring of their compatibility with the lead structural template using methods such as threading or forcefield-based full atom representation. Based on the the relative ranking of the hits, a subset of multiply aligned sequences are selected to create hit variant library II and screened in vitro or in vivo for the desired function(s). See Route IV in **Figure 1D** and Figure 5.

4. Construction of the Hit Variant Library

To further explore the rich diversity encoded in the structure and sequence spaces of proteins, the hits that are selected based on sequence alignment are profiled at each amino acid position of the sequences to generate a variant profile. A hit variant library is combinatorially enumerated using this variant profile. **Figure 4** illustrates an exemplary process for constructing a hit variant library. The variant profile generated from the hit library (i.e., sequence hits or filtered sequence hits) is listed based on frequency of amino acid appearing at each position in the hit sequences (**Figures 11 & 19B**). The variants profiled provides an excellent starting point for

constructing combinatorial libraries.

Some cutoff values based on the frequencies (e.g., a frequency of over 5% or higher) or preferred variants of amino acids at each position, and/or computational results can be applied to reduce the size of this hit variant library (see the lower portion of **Figure 11** for a cutoff at 10% of the totoal number of hits; **Figure 19B** uses 5%). The variants based on these highly preferred amino acid residues at each position should offer a good pool of recombinant sequences for fishing out sequences with high affinity or other desired functions.

The informational sequence entropy, calculated based on the variant frequency at each position, provides a quantitative means to measure how significant the residue identities in aligned sequences deviates from a random distribution of amino acid residues. A relative entropy can be used in the present invention to take into account highly variable mutagenesis probabilities of the sequences involving protein variants (Plaxco KW, Larson S, Ruczinski, Riddle DS, Thayer EC, Buchwitz B, Davidson AR, Baker D (2000) J Mol Biol 298, 303-312). The inventors believe that the relative site entropies provide a good guide for the positions and mutants that should be targeted for computational and experimental screening since they are based on real evolutionary data from databases of expressed proteins.

The relative site entropy measures the diversity at each position of amino acid residues accumulated during evolution while maintaining structure and function of the hit sequences. These sites are chosen to recombine for computational and experimental screening. Because the size of the resulting combinatorial hit variant library is much smaller than that generated by a random combination of all 20 amino acids at each position, it is possible to carry out more accurate and detailed computational or even direct experimental screening.

The sequence entropies resulting from the hit library in the present invention are not related to the site entropies which others in the field have used to measure the structural tolerance toward amino acid substitution, using force-field based computational method (Voigt CA, Mayo SL, Arnold FH, Wang ZG (2001) PNAS 98, 3778-3783). Although a forcefield-based method would provide some novel mutants that may not yet have been sampled by evolution, the site entropy

derived from the evolutionary sequences (i.e., the sequence entropy) should provide more meaningful statistics on the variation and preferred mutants at each position with all information including structural, kinetic, expression and biological activities incorporated. This may be important for targeting difficult structures such as loop regions in antibodies that are not yet fully understood or predicted by forcefield-based methods, but they can be modeled with some confidence using the database-based methods of the present invention. The homology-based method that relies on the evolutionary information is still one of the most reliable ways to model loop structures that can be augmented with forcefield-based simulations.

As will be described in detail in the Example section, the variant profile for an anti-VEGF antibody (the lead antibody) was searched by using several different approaches. Based on a sequence of V_H CDR3 of this lead antibody, the variant profiles of the hit lists from Kabat, genpept and a non-redundant database, combining Kabat, genpept, imgt, and others, are listed. Important mutants observed by others in affinity matured sequences from this antibody also appear with high frequency in the variant profile searched using the methods of the present invention. For example, it was believed that the single most important mutant was H97 in the lead sequence replaced by Y97 in the matured sequence (**Figure 9B**) which is almost 50% in the amino acid variants at this position (**Figure 11**). The above-described methods of the present invention have several advantages in protein design and engineering. In any recombinant library, the diversity is necessarily limited by the ability to screen, which means that allocation and, thus design, of diversity is an important factor in the creation of a functionally relevant library. The inventive method is an *in silico* rational design of protein, in particular, antibody. It begins with the selection of functionally similar "natural" polypeptide fragments from databases of expressed proteins to form the hit library. Analysis of specific positional variations in the "naturally" occurring peptide fragments yields evolutionary data about preferred residues and positions—the variant profile. A critical analysis of the variants can identify important residues and combinations. Combinatorial enumeration of the reduced set of select variants leads to the

generation of a hit variant library that is focused on the functionally relevant sequences.

Starting with the variant profile, the *in silico* rational library design of the present invention generates a focused library or libraries of protein fragments based on functional and structural data. To some extent, in silico recombination is similar in principle to DNA shuffling of a family of homologous sequences. But the present inventive approach is a highly efficient sequence recombination procedure for a family of protein sequences with widely distributed sequence homology. Furthermore, in the present invention, the recombinations occur at the amino acid level and can be localized to specific functional region to generate a library whose members are designed rather than randomly recombined. It is not constrained by a homology requirement and can be selectively modified according to structural or experimental data. For example, the sequences in the hit library have sequence identities relative to the lead sequence ranging from 100 to 20, or even lower depending on the searching method and database used. In comparison, the DNA shuffling is DNA recombination process between closely related sequence homologues with stringent requirement on the sequence homology between recombined nucleic acid sequence; DNA shuffling is inefficient in generating beneficial mutant recombination and it is prone to random mutations during experimental recombination.

5. Structure-Based Evaluation of Antibody Variant Library

The hit library or a hit variant library, derived from the recombination of the variant profile from the hit library as described above, may be evaluated based on their structural compatibility with the lead protein. For structure-based evaluation of the antibody variant library, the present invention addresses the following questions: i) how to model conformations of noncanonical loops in the presence of antigen which forms a protein complex with the antibody; (ii) how to place side chains on CDR loop backbones to best fit the antibody and/or antigen structure; and (iii) how to combine CDR loops with the best framework model to allow formation of stable antibody-antigen

complex with high affinity. Implementing procedures are described in detail as follows.

1) Antibody structures and structure models

A structural template of the lead antibody can either be taken directly from an X-ray or NMR structure or modeled using structural computational engines described below. As shown in the EXAMPLE section, the structural templates for anti-VEGF antibody are taken from PDB databank, 1BJ1 for the parental antibody and 1CZ8 for the matured antibody. Both templates were used in the presence and absence of the antigen VEGF. The scoring listed in the examples is from 1CZ8 in the presence of the antigen VEGF.

2) Evaluation based on structural template of the lead antibody

As an example, an antibody with a known 3D structure serves as the lead protein. This requirement for a well-defined structure (such as one obtained by X-ray crystallography) is not absolute since alternative techniques, such as homology-based modeling, may be applied to generate a reasonably defined template structure for a target protein to be engineered. Generation of the hit variant library requires the determination, modification, and optimization of the amino acid positional variant profile. The lead sequence and sequences in the hit library and the hit variant library are scored in the context of the 3D structure of the lead antibody and scored to obtain the ranking distribution for these sequences. It is noted that, although the scoring in the EXAMPLE section is based on an empirical all-atom energy function, any computationally tractable scoring or fitness function may be applied to structurally evaluate these sequences.

Figure 5 illustrates an exemplary procedure for structural evaluation of sequences from the lead, the hit library and the hit variant library. For scoring and ranking, these sequences are built into the lead structural template by substituting side chains from a backbone-dependent/independent rotamer library (Dunbrack RL Jr, Karplus M (1993) J Mol Biol 230:543-574). The side chains and the backbone of the substituted segment are then locally energy minimized to relieve local strain. Each structure is scored using a custom energy function

that measures the relative stability of the sequence in the lead structural template.

Comparison of the energies for sequences from the lead, the hit library and the hit variant library indicates the degree of structural compatibility of the various sequences with the lead structural template. It is not unreasonable to obtain a very broad distribution with many sequences scoring better or worse than the lead sequence. The focus is not to identify specific sequences (although permissible) but to identify a population of sequences or a sequence ensemble with average scores equal to or better than the lead sequences and share ensemble properties in sequence that can be targeted simultaneously using degenerate nucleic acid libraries. The amino acid sequence ensemble represents a sequence space that is likely to show good structural compatibility with better binding sites and orientation for epitope recognition than a single, specific sequence. The combinatorial libraries of the sequence ensembles distributed around the statistical ensemble average should be targeted experimentally in order to increase the chance of finding good candidates with improved affinity.

3) Evaluation based on lead structural template in the presence of its ligand

Optionally, sequences from the lead, the hit library and the hit variant library can be evaluated based on the lead structural template in the presence of its ligand or antigen, for example, a lead anti-VEGF antibody in complex with VEGF. This approach is useful when structure of the complex formed by the lead protein and its ligand is known or readily ascertained.

In the presence of the antigen, the complete thermodynamic cycle of complex formation between an antibody and an antigen may be included in the calculation. The conformation of the antibody, especially in the combining site, may be modeled based on individual CDR loop conformation from its canonical family with preferred side-chain rotamers as well as the interactions between CDR loops. A wide range of conformations, including those of the side chains of amino acid residues and those of the CDR loops in the antigen combining site, can be sampled and incorporated into a main framework (or a scaffold) of an

antibody. With the antigen present, such conformational modeling assures higher physical relevancy in the scoring, using physical-chemical force fields as well as semi-empirical and knowledge-based parameters, and better representation of the natural process of antibody production and maturation in the body.

4) Correlation of the scores of antibody sequences in the presence and absence of an antigen

It is desirable to have the complex structure between an antigen and its antibody to focus the antibody library towards sequences with good probability of binding the antigen. Unfortunately, for most antibodies of biomedical interest, the complex structure between the antibody and antigen is not yet available.

The inventors found that many sequences that are favorable in stabilizing target antibody scaffold are also among the selected candidates that can stabilize the specific antibody-antigen complex even for the V_H CDR3 that is involved directly in binding to the antigen. Correlation analysis shows that there is a general correlated trend in the scores of the antibody sequences in the presence and absence of the antigen (Figure 12C). Further, a large population of sequences selected with good scores is favorable in stabilizing the scaffolding of the binding motifs such as V_H CDR3 for anti-VEGF used here.

It should be noted that, without the complex structure, the antibody structure alone can still give a population of sequences that stabilize the target scaffold while possessing the right binding site for the antigen. Although conformational change upon antigen binding has been observed, it is not clear if conformation change is only one of many possible solutions or is an absolute requirement for the antigen-antibody interaction. The goal is to identify an ensemble of sequences likely to form a functional proteins so the bound structure is not a requirement as long as it does not undergo major conformational shifts. Based on the available structures of antibodies in both bound and unbound states, this is a good assumption. At least, some structure fluctuations are allowed in the approach taken here (see 19A) as far as they belong to the same family of ensemble structures.

Alternatively, if the structure of the lead antibody is not

available, a template may be generated by modeling. Antibody structure or structure motifs are among some of the best known examples of proteins for which structural models can be generated, using homology modeling, with a relatively high degree of confidence. Thus, it is still possible to target a sequence library for the lead sequence without using the lead structure template. As will be shown in the EXAMPLE section, stretches of sequence libraries that cover the target motifs can be synthesized and used to screen for antibody with high affinity without relying on the structure of the lead antibody.

5) Structural computational engines

Many programs are available for modeling and evaluating the libraries against the lead structural template. For example, a molecular mechanics software may be employed for these purposes, examples of which include, but are not limited to CONGEN, SCWRL, UHBD, GENPOL and AMBER.

CONGEN (**CON**formation **GEN**erator) is a program for performing conformational searches on segments of proteins (R. E. Bruccoleri (1993) *Molecular Simulations* 10, 151-174 (1993); R. E. Bruccoleri, E. Haber, J. Novotny, (1988) Nature 335, 564-568 (1988); R. Bruccoleri, M. Karplus. (1987) Biopolymers 26, 137-168. It is most suited to problems where one needs to construct undetermined loops or segments in a known structure, i.e. homology modeling. The program is a modification of CHARMM version 16, and has most of the capabilities of that version of CHARMM (Brooks BR, Bruccoleri BE, Olafson BD, States DJ, Swaminathan S, Karplus M. (1983) J. Comput. Chem. 4, 187-217).

The basic energy function used includes terms for bonds, angles, torsional angels, improper angles, van der Waals and electrostatic interactions with distance dependent dielectric constant using Amber94 forcefield which can be determined using CONGEN. (see EXAMPLE section).

CONGEN program is used to search for low-energy conformers that are close or correspond to the naturally occurring structure with lowest free energy (Bruccoleri and Karplus (1987) Biopolymers 26:137-168; and Bruccoleri and Novotny (1992) Immunomethods 96-106).

Given an accurate Gibbs function and a short loop sequence, all of the stereochemically acceptable structures of the loop can be generated and their energies calculated. The one with the lower energy is selected.

The program can be used to perform both conformational searches and structural evaluation using basic or refined scoring function. The program can calculate other properties of the molecules such as the solvent accessible surface area and conformational entropies, given steric constraints. Each one of these properties in combination with other properties described below can be used to score the digital libraries.

According to the present invention, the defined canonical structures for five of the CDRs (V_L CDR1, 2, and 3, and V_H CDR1, and 2) except for V_H CDR3. V_H CDR3 is known to show large variation in its length and conformations, although progress has been made in modeling its conformation with increasing number of antibody structures becoming available in the PDB (protein data bank) database. CONGEN may be used to generate conformations of a loop region (e.g., V_H CDR3) if no canonical structure is available, to replace the side chains of the template sequence with the corresponding side chain rotamers of the target amino acids. Third, the model can be further optimized by energy minimization or molecular dynamics simulation or other protocols to relieve the steric clashes and strains in the structure model.

SCWRL is a side chain placing program that can be used to generate side chain rotamers and combinations of rotamers using the backbone dependent rotamer library (Dunbrack RL Jr, Karplus M (1993) J Mol Biol 230:543-574; Bower, MJ, Cohen FE, Dunbrack RL (1997) J Mol Biol 267, 1268-1282). The library provides lists of chi1-chi2-chi3-chi4 values and their relative probabilities for residues at given phi-psi values. The program can further explore these conformations to minimize sidechain-backbone clashes and sidechain-sidechain clashes. Once the steric clash is minimized, the side chains and the backbone of the substituted segment can be energy minimized to relieve local strain using CONGEN (Bruccoleri and Karplus (1987) Biopolymers 26:137-168).

Several automatic programs that are developed specifically for building antibody structures may be used for structural modeling of antibody in the present invention. The ABGEN program is an automated antibody structure generation algorithm for obtaining structural models of antibody fragments. Mandal et al. (1996) Nature Biotech. 14:323-328. ABGEN utilizes a homology based scaffolding technique and includes the use of invariant and strictly conserved residues, structural motifs of known Fab, canonical features of hypervariable loops, torsional constraints for residue replacements and key inter-residue interactions. Specifically, the ABGEN algorithm consists of two principal modules, ABalign and ABbuild. ABalign is the program that provides the alignment of an antibody sequence with all the V-region sequences of antibodies whose structures are known and computes alignment score scores. The highest scoring library sequence is considered to be the best fit to the test sequence. ABbuild then uses this best fit model output by ABalign to generate the three-dimensional structure and provides Cartesian coordinates for the desired antibody sequence.

WAM (Whitelegg NRJ and Rees, AR (2000) Protein Engineering 13, 819-824) is an improved version of ABM which uses a combined algorithm (Martin, ACR, Cheetham, JC, and Rees AR (1989) PNAS 86, 9268-9272) to model the CDR conformations using the canonical conformations of CDRs loops from x-ray PDB database and loop conformations generated using CONGEN. In short, the modular nature of antibody structure makes it possible to model its structure using a combination of protein homology modeling and structure predictions.

In a preferred embodiment, the following procedure will be used to model antibody structure. Because antibody is one of the most conserved proteins in both sequence and structure, homology models of antibodies are relatively straightforward, except for certain CDR loops that are not yet determined within existing canonical structures or those with insertion or deletions. However, these loops can be modeled using algorithms that combine homology modeling with conformational search (for example, CONGEN can be used for such purpose).

The defined canonical structures for five of the CDRs (L1,2,3 and H1,2) are used. H3 in variable heavy chain (i.e., V_H CDR3) is known to

show a large variation in its length and conformation, although progress has been made in modeling its conformation as more antibody structures became available. The modeling methods include protein structure prediction methods such as threading, and comparative modeling, which aligns the sequence of unknown structure with at least one known structure based on the similarity modeled sequence. The de novo or *ab initio* methods also show increasing promise in predicting the structure from sequence alone. The unknown loop conformations can be sampled using CONGEN if no canonical structure is available (Bruccoleri RE, Haber E, Novotny J (1988) Nature 355, 564-568). Alternatively, *ab initio* methods, including but not limited to Rosetta ab initio method, can be used to predict antibody CDR structures (Bonneau R, Tsai J, Ruczinski I, Chivian D, Rohl C, Strauss CE, Baker D (2001) Proteins Suppl 5, 119-126) without relying on similarity at the fold level between the modeled sequence and any known structures. A more accurate method that uses the state-of-the-art explicit solvent molecular dynamics and implicit solvent free energy calculations can be used to refine and select for native-like structures from models generated from either CONGEN or Rossetta *ab initio* method (Lee MR, Tsai J, Baker D, Kollman PA (2001) J Mol Biol 313, 417-430).

Either the X-ray structures as used here (1BJ1 and/or 1CZ8) or the modeled structure as described above can be used as the structural template for designing antibody library for experimental screening described below.

6) Scoring functions for structural evaluation

In one embodiment of the present invention, computational analysis is used for structural evaluation of the selected sequences from the sequence evaluation processes described above in Sections 3 and 4. The structural evaluation is based on an empirical and parameterized scoring function and is intended to reduce the number of subsequent *in vitro* screenings necessary.

This approach uses an existing structural template to score all the amino acid libraries generated. The use of a known structure as a template to assess antibody-antigen interaction assumes that (i) the

structures of the antibody and antigen molecules do not change significantly between bound and free states, (ii) the mutations in the CDRs do not significantly alter the global as well as local structures and (iii) the energetic effects due to mutations in the CDRs are localized and can be scored to assess functions directly related to the mutations. An advantage of having a known structure as a template is that it can serve as a good starting point for design improvements rather than compared to the more challenging approach using modeled structures. The energy distribution of these sequence hits should reveal how well they cover the fitness function of the target scaffold in terms of their structural compatibility with the target.

Since the above-described assumptions necessarily introduce errors due to uncertainties in the structures of the mutants, it is likely that a sophisticated scoring function would still fail to give meaningful prediction if the mutant has altered the structure. A generic but well tested forcefield (see below) was used in the initial calculations in the model system of anti-VEGF antibody as shown in the Example section. It may avoid the bias built into the specific systems in general if the preferred region of the fitness landscape can be explored by sampling the ensemble sequences implemented experimentally. However, the present invention does not preclude the use of more sophisticated scoring functions for the structural evaluation.

Many energy functions can be used to score the compatibility between sequences and structures. Typically, four kinds of energy functions can be used: (1) empirical physical chemistry forcefields such as standard molecular mechanic forcefields discussed below that are derived from simple model compounds; (2) knowledge-based statistical forcefields extracted from protein structures, the so called potential of mean force (PMF) or the threading score derived from the structure-based sequence profiling; (3) parameterized forcefield by fitting the forcefield parameters using experimental model system; (4) combinations of one or several terms from (1) to (3) with various weighting factor for each term.

The following are some well-tested physical-chemistry forcefields that can be used or incorporated into the scoring functions. For example, amber 94 forcefield was used in CONGEN to score the

sequence-structure compatibility in the examples below. The forcefields include but are not limited to the following forcefields which are widely used by those skilled in the art: **Amber 94** (Cornell, WD, Cieplak P, Bayly CI, Gould IR, Merz KM Jr, Ferguson DM, Spellmeyer DC, Fox T, Caldwell JW and Kollman PA. *JACS* (1995) 117, 5179-5197 (1995); **CHARMM** (Brooks, B.R., Bruccoleri, R.E., Olafson, B.D., States, D.J., Swaminathan, S., Karplus, M. (1983) J. Comp. Chem. 4, 187-217.; MacKerell, A D ; Bashford, D; Bellott, M; Dunbrack, R L; Eva seck, J D; Field, M J; Fischer, S; Gao, J; Guo, H; Ha, S; JosephMcCarthy, D; Kuc nir, L; Kuczera, K; Lau, F T K; Mattos, C; Michnick, S; Ngo, T; Nguyen, D T; Pro hom, B; Reiher, W E; Roux, B; Schlenkrich, M; Smith, J C; Stote, R; Straub, J; Watanabe, M; WiorkiewiczKuczera, J; Yin, D; Karplus, M (1998) J. Phys. Chem., B 102, 3586-3617); **Discover CVFF** (Dauber-Osguthorpe, P.; Roberts, V. A.; Osguthorpe, D. J.; Wolff, J.; Genest, M.; Hagler, A. T. (1988) Proteins: Structure, Function and Genetics, 4, 31-47.); **ECEPP** (Momany, F. A., McGuire, R. F., Burgess, A. W., & Scheraga, H. A., (1975) J. Phys. Chem. 79, 2361-2381.; Nemethy, G., Pottle, M. S., & Scheraga, H. A., (1983) J. Phys. Chem. 87, 1883-1887.); **GROMOS** (Hermans, J., Berendsen, H. J. C., van Gunsteren, W. F., & Postma, J. P. M., (1984) Biopolymers 23, 1); **MMFF94** (Halgren, T. A. (1992) J. Am. Chem. Soc. 114, 7827-7843.; Halgren, T. A. (1996) J. Comp. Chem 17, 490-519.; Halgren, T. A. (1996) J. Comp. Chem. 17, 520-552.; Halgren, T. A. (1996) J. Comp. Chem. 17, 553-586.; Halgren, T. A., and Nachbar, R. B. (1996) J. Comp. Chem. 17, 587-615.; Halgren, T. A. (1996) J. Comp. Chem. 17, 616-641.); **OPLS** (see Jorgensen, W. L., & Tirado-Rives, J.,(1988) J. Am. Chem. Soc. 110, 1657-1666.; Damm, W., A. Frontera, J. Tirado-Rives and W. L. Jorgensen (1997) J. Comp. Chem. 18, 1955-1970.); **Tripos** (Clark, M., Cramer III, R. D., van Opdenhosch, N., (1989) Validation of the General Purpose Tripose 5.2 Force Field, J. Comp. Chem. 10, 982-1012.); **MM3** (Lii, J-H., & Allinger, N. L. (1991) J. Comp. Chem. 12, 186-199). Other generic forcefields such as **Dreiding** (Mayo SL, Olafson BD, Goddard (1990) J Phy Chem 94, 8897-8909) or specific forcefield used for protein folding or simulations like **UNRES** (United Residue Forcefield; Liwo et al., (1993) Protein Science 2, 1697-1714; Liwo et al., (1993) Protein Science 2, 1715-1731; Liwo et al., (1997) J. Comp. Chem. 18, 849-873;

Liwo et al., (1997) J. Comp. Chem. 18:874-884; Liwo et al., (1998) J. Comp. Chem. 19:259-276.), may also be used.

The statistical potentials derived from protein structures can be also used to assess the compatibility between sequences and protein structure using. These potential include but not limited to residue pair potentials (Miyazawa S, Jernigan R (1985) Macromolecules 18, 534-552; Jernigan RL, Bahar, I (1996) Curr. Opin. Struc. Biol. 6, 195-209). The potentials of mean force (Hendlich et al., (1990) J. Mol. Biol. 216, 167-180) has been used to calculate the conformational ensembles of proteins (Sippl M (1990) J Mol Biol. 213, 859-883). However, some limitations of these forcefields are also discussed (Thomas PD, Dill KA (1996) J Mol Biol 257, 457-469; Ben-Naim A (1997) J Chem Phys 107, 3698-3706).

Another methods to score the compatibility between sequences and structure is to use sequence profiling (Bowie JU, Luthy R, Eisenberg DA (1991) Science 253, 164-170) or threading scores (Jones DT, Taylor WR, Thornton JM (1992) Nature 358, 86-89; Bryant, SH, Lawrence, CE (1993) Proteins 16, 92-112; Rost B, Schneider R, Sander C (1997) J Mol Biol 270, 471-480; Xu Y, Xu D (2000) Proteins 40, 343-354). These statistical forcefields based on the quasichemical approximation or Boltzmann statistics or Bayes theorem (Simons KT, Kooperberg C, Huang E, Baker D (1997) J Mol Biol 268, 209-225) are used to assess the goodness of the fit between a sequence and a structure or for protein design (Dima RI, Banavar J R, Maritan A (2000) Protein Science 9, 812-819).

Furthermore, the structure-based thermodynamic parameters related to the thermodynamic stability of the protein structures can be also used to evaluate the fitness between a sequence and a structure. In the structure-based thermodynamic methods, the thermodynamic quantities such as heat capacity, enthalpy, entropy can be calculated based on the structure of a protein to explain the temperature-dependence of the thermal unfolding using the thermodynamic data from model compounds or protein calorimetry studies (Spolar RS, Livingstone JR, Record MT (1992) Biochemistry 31, 3947-3955; Spolar RS, Record MT (1994) Science 263, 777-784; Murphy KP, Freire E (1992) Adv Protein Chem 43, 313-361; Privalov PL, Makhatadze GI

(1993) J Mol Biol 232, 660-679; Makhatadze GI, Privalov PL (1993) J Mol Biol 232, 639-659). The structure-based thermodynamic parameters can be used to calculate structural stability of mutant sequences and hydrogen exchange protection factors using ensemble-based statistical thermodynamic approach (Hilser VJ, Dowdy D, Oas TG, Freire E (1998) PNAS 95, 9903-9908). Thermodynamic parameters relating to statistical thermodynamic models of the formation of the protein secondary structures have also been determined using experimental model systems with excellent agreement between predictions and experimental data (Rohl CA, Baldwin RL (1998) Methods Enzymol 295, 1-26; Serrano L (2000) Adv Protein Chem 53, 49-85).

A combination of various terms from molecular mechanic forcefields plus some specific components has been used in most protein design programs. In a preferred embodiment, the forcefield is composed of one or several terms such as the vdw, hydrogen bonding and electrostatic interactions from the standard molecular mechanics forcefields such as Amber, Charmm, OPLS, cvff, ECEPP, plus one or several terms that are believed to control the stability of proteins.

To improve the scoring function, additional energy terms are included in later steps that allow tuning of the scoring function to better address deviations from experimental results and influence of specific antibody-antigen interactions of interest. For example, one energy term can penalize arginine mutation to reduce its contribution to the overall score due to the uncertainty of prediction its sidechain conformation and to compensate for the bias in the current scoring function that favors arginine. Another energy term can score the charged and polar group solvent exposure based on surface area calculation so that mutations that lead to charge burial are penalized according to exposed surface.

In practice, there are many scoring functions that can be used to score the compatibility of sequences with a template structure or structure ensemble. The refined scoring function is composed of several terms including contributions from electrostatic and van der Waals interactions, ΔG_{MM} calculated using molecular mechanics forcefield, contribution from solvation including electrostatic solvation and

solvent-accessible surface, ΔG_{sol}, and contribution from the conformational entropy (Sharp KA. (1998) Proteins 33, 39-48; Novotny J, Bruccoleri RE, Davis M, Sharp KA (1997) J Mol Biol 268, 401-411).

A simple fast way for computational screening is to calculate structural stability of a sequence using the total or combination of energy terms using a basic scoring function that includes terms from molecular mechanic forcefield such as Amber94 as implemented in CONGEN.

$$\Delta E_{total} = E_{bond} + E_{angel} + E_{dihed} + E_{impr} + E_{vdw} + E_{elec} + E_{solvation} + E_{other}$$

or alternatively, the binding free energy is calculated as the difference between the bound and unbound states using a refined scoring function

$$\Delta G_b = \Delta G_{MM} + \Delta G_{sol} - T\Delta S_{ss}$$

where:

$$\Delta G_{MM} = \Delta G_{ele} + \Delta G_{vdw} \quad (1)$$

$$\Delta G_{sol} = \Delta G_{ele\text{-}sol} + \Delta G_{ASA} \quad (2)$$

The ΔG_{ele} and ΔG_{vdw} electrostatic and van der Waals interaction energy are calculated using Amber94 parameters implemented in CONGEN for ΔG_{MM}, whereas the $\Delta G_{ele\text{-}sol}$ is electrostatic solvation energy required to move a heterogeneously distributed charges in a protein with no dielectric boundary into an aqueous phase with dielectric boundary defined by the shape of a protein. This is calculated by solving the Poisson-Boltzmann equation for the electrostatic potential for the reference and mutant structures. ΔG_{ASA}, the nonpolar energy is the energetic cost of moving nonpolar solute groups into an aqueous solvent, resulting in the reorganization of the solvent molecules. This has been shown to correlate linearly with the solvent accessible surface area of the molecule (Sitkoff D, Sharp, KA, Honig B (1994) J Phys Chem 98, 1978-1988; Pascual-Ahir & Silla (1990) J Comp Chem 11, 1047-1060).

The change in the side chain entropy (ΔS_{ss}) is a measure of the

effect on the local side chain conformational space, particularly at the binding interface. This is calculated from the ratio of the number of allowed side chain conformations in the bound and unbound states. For general scoring purposes, the independent side chain approximation is applied to the mutated side chains in order to avoid the huge computational demand imposed by sampling conformational space of multiple side chains in various backbone conformations.

The sequences in the hit library or hit variant library are evaluated for their structural compatibility with the target structure and are mapped out on the energy landscape of the target fold. For the anti-VEGF antibody, the scores for the antibody sequences in the presence and absence of antigen are correlated in general trend because a large number of variants are capable of stabilizing the antibody scaffold (see **Figure 12C**). Among them, there is a significant fraction of the sequences that are capable of binding the target epitope. As shown in the EXAMPLE section, CDR library sequences are ranked based on their fitness scores, based on the relative stability of the template antibody-antigen complex (1CZ8), and experimentally selected sequences are identified (**Figure 13A**).

It is beneficial, if possible, to determine the scores in both the antigen bound and unbound states to eliminate any grossly unfavorable sequences in either state. By doing so, we can avoid the need to accurately score the differences between the bound and unbound states while still effectively reducing the search space.

The scoring function is used to score the sequences in the hit library, hit variant library I or hit variant library II and, optionally, the differences between the lead sequence or lead structural template sequence and the library sequence is calculated to complete a thermodynamic cycle. Consequently, sequences can be selected for further experimental screening based on any of the following criteria: 1) sequences that score better than the lead sequence in stabilizing the antibody structure are selected; 2) sequences that score better than the lead sequence in stabilizing the antibody-antigen complex structure are selected; 3) the difference in the score between the bound and unbound states is better than the lead sequence, provided the scoring function is sensitive enough to discriminate small differences between

large numbers. The last criterion should be used only if highly refined scoring functions or high quality ensemble based scoring function is available and prefereably with systems where high quality mutant data are available for calibration of the scoring function.

Sequences that score better than the lead sequence(s) are analyzed and sorted into distinct clusters. A combination of the clusters should cover sufficient sequence and structure space that covers desired regions in the fitness landscape (**Figure 7**). This approach of selecting a scoring window by clustering the sequences is taken as an effort to reduce the physical library size. Another benefit of the clustering approach is that combination of the subsequent nucleic acid libraries (e.g., nucleic acid library I, II, III, etc., **Figure 7**) from several disjointed scoring windows may still cover a large portion of sequence and structure space with better scores than the lead sequence. A desirable result of this clustering process is that since each of these clusters of sequences requires a much smaller physical library size than the combined library, the nucleic acid library encoding each of the clusters is small enough for a thorough screening in vitro or in vivo.

In one embodiment of the present invention, the scoring of the hit variant library is used to select a population of sequences optimized for the desired function and to formulate the starting design for hit variant library II. Scoring of the resulting hit variant library II is used to determine the effects of modification and design enhancements on variant profile. Hit variant library III, derived from the nucleic acide library (described in detail in Section 7 below), is also scored to determine the fitness of the library and to evaluate the effectiveness of the scoring function in mapping the sequence and structure space onto the fitness landscape of the molecular target.

In a particular embodiment, standard terms from MM terms have been combined with the solvation terms including electrostatic solvation and solvent-accessible solvation term calculated with continuous solvent model for electrostatic solvation; these MM-PBSA or MM-GBSA method, together with contribution from the conformational entropy including backbone and side chains, have shown good correlation between experimental and calculated values in the free energy change

(Wang W, Kollman P (2000) J Mol Biol 303, 567-582). Compared to other scoring functions used in protein and drug design, MM-PBSA or MM-GBSA is better physical model for scoring and would handle various problems with a consistent approach, although it is computational expensive because multiple trajectories from molecular dynamic simulation in explicit water are required to calculate the ensemble averages for the system and continuous solvent model is still computationally slow. These accurate methods should provide a benchmark for calibrating the simple scoring function used for library screening or for studying some challenging mutations that elude simple calculations.

7) Examples of forcefields for protein design

An important interaction for scoring the correct packing interactions inside the core of proteins, van der Waals (vdw) interaction was used to design the protein core sequences by testing allowed rotamer sequences in enumeration (Ponder JW, Richards FM (1987) J Mol Biol 193, 775-791. A group of sequences can be selected under a potential function using simulated evolution with stochastic algorithm; the ranking order of the energies of selected sequences for residues in the hydrophobic cores of proteins correlates well with their biological activities (Hellinga HW, Richards FM (1994) PNAS 91, 5803-5807).

Similar approaches were also used to design proteins using stochastic algorithm (Desjarlais J, Handel T, (1995) Protein Science 4, 2006-2018; Kono H, Doi J (1994) Proteins, 19, 244-255). Effect of potential function on the designed sequences of a target scaffold has been evaluated by including van der Waals, electrostatics, and surface-dependent semi-empirical environmental free energy or combinations of terms in an automatic protein design method that keeps the composition of amino acid sequence unchanged. It was shown that each additional term of the energy function increases progressively the performance of the designed sequences with vdw for packing, electrostatics for folding specificity and environmental solvation term for burial of the hydrophobic residues and for exposure of the hydrophilic residue (Koehl P, Levitt M (1999) J Mol Biol 293, 1161-1181).

The self-consistent mean field approach was used to sample the energy surface in order to find the optimal solution, (Delarue M, Koehl. (1997) Pac. Symp. Biocomput. 109-121; Koehl P, Delarue M, (1994) J. Mol. Biol. 239, 249-275; Koehl P, Delarue M (1995) Nat. Struct. Biol. 2,163-170; Koehl P, Delarue M (1996) Curr. Opin. Struct. Biol. 6:222-226; Lee J. (1994) Mol. Biol. 236, 918-939; Vasquez (1995) Biopolymers 36, 53-70). Combination of terms from molecular forcefield, knowledge-based statistical forcefield and other empirical correction has been also used to design protein sequences that are close to the native sequence of the target scaffold (Kuhlman B, Baker D (2000) PNAS 97, 10383-10388). The structure-based thermodynamic terms were included in addition to the steric repulsion in the protein core design (Jiang X, Farid H, Pistor E, Farid RS (2000) Protein Science 9, 403-416). Knowledge-based potentials have been used to design proteins (Rossi A, Micheletti C, Seno F, Maritan A (2001) Biophysical Journal 80, 480-490).

Forcefields have been also optimized specifically for protein design purpose in combination with the dead end elimination algorithm (Dahiyat BI, Mayo SL (1996) Protein Science 5, 895-903). The energy function is decomposed into pairwise functional forms that combine molecular mechanic energy terms with specific solvation term is used for residues at the core, boundary and surface positions; dead end elimination algorithm is used to sip through huge number of combinatorial rotameric sequences. The stringency of force fields and rigid inverse folding protocol with fixed backbone used in protein design has inevitably resulted a significant rate of false negative: rejection of many sequences that might be acceptable if soft energy function or flexible backbone is allowed. Moreover, the energy function used for protein design is quite different from general forcefields such as Amber or Charmm that are widely used and tested for studying protein folding or stability (Gordon DB, Marshall SA, Mayo SL (1999) Curr Opin Stru Biol 9, 509-513). Cautions should be exercised in comparing the sequences designed using specific protocol with others from alternative methods because a direct comparison among them may not be possible due to the false negative issues involved in protein design protocols.

The inventors believe that, although a high false negative rate in protein design is not a problem for designing proteins with few restriction, this will pose serious problems for designing proteins for pharmaceutical application for which only small restrictive region is allowed to have altered sequences to improve protein function. For example, many variants are acceptable for V_H CDR3, even though only one or two residues in the V_H CDR3 in the VEGF antibody would actually improve its binding affinity, but for the framework regions, only a few mutants can be tolerated for humanization. Therefore, it is accuracy rather than the scale or speed of computational screening that matters the most for functional improvement in order to identify those few mutants in the targeted region.

Optionally, molecular dynamics or other computational methods can be used to generate structure ensembles and the ensemble average scores used to rank sequences (Kollman PA, Massova I, Reyes C, Kuhn B, Huo SH, Chong LT, Lee M, Lee TS, Duan Y, Wang W, Donini O, Cieplak P, Srinivasan P, Case DA, and Cheatham TE (2000) Acc. Chem Res. 33, 889-897). The average properties calculated from ensemble structures show better correlation with corresponding data from experimental measurement.

6. Construction of Mutant Antibody Library Based on Lead Structural Template

Alternatively, a mutant antibody library may be constructed directly based on the 3D structure of the lead antibody and then screened for desired function in vitro or vivo. This approach takes a short cut by avoiding the construction of the hit variant library and directly evaluates sequences from the hit library constructed by screening protein databases. This approach is depicted as Route III in **Figures 1C or 1E-H**.

As described in detail in section 3, there are several ways to construct the hit library. One way of building the hit library is to search in a protein database to find those segments that match in sequence pattern with the amino acid sequence of the region to be mutated, for example, CDR3 of the heavy chain (CDR H3) of the lead

antibody. A conventional BLAST analysis may be employed to search for sequences with high homology to the CDR H3 sequence.

Optionally, PSI-BLAST may be used to search for sequence homologues of the CDR H3 sequence of the template antibody.

Also optionally, single target sequence and/or multiple sequence alignment can be used to build a profile Hidden Markov Model (HMM). This HMM is then be used to search for both close and remote human homologues from a protein sequence database such as Kabat database of proteins and the human germline immunoglobulin database for frameworks. The Kabat database of proteins of immunological interest from various species can be used for designing diverse sequences for CDRs.

The sequences in the hit library selected by using any of the above methods for sequence alignment or combinations thereof can be profiled to compare the type of amino acid and its frequency of appearance in each position of the corresponding region in the template antibody (e.g., CDR H3).

Each member of this hit library is grafted onto the corresponding region in the template antibody (e.g., CDR H3) and tested for its structural compatibility with the rest of the antibody by using scoring functions described in section 5 above.

Using similar approaches, hit libraries can be constructed based on lead sequences from different regions of the lead antibody, such as CDR1, CDR2 of the heavy chain and light chain, and tested for structural compatibility with the rest of the lead antibody. These libraries may be combined to allow simultaneous mutations to different regions of the lead antibody, thereby increasing the diversity of the mutant antibody library.

All of the mutant antibody sequences selected in these processes are pooled and screened for high affinity binding to the target antigen in vitro or in vivo.

7. Construction of Nucleic Acid Library for Experimental Screening

To facilitate functional screening in vitro or in vivo, nucleic acid libraries are constructed to encode the amino acid sequences that are selected by using the above-described methods of the present invention. The size of the nucleic acid library may vary depending on the particular method of selecting and profiling the amino acid sequences. For example, the size of the nucleic acid may reach $>10^6$ if too many amino acid sequences are chosen and recombined. Partitioning and re-profiling of the amino acid sequences may be performed to reduce the size of the nucleic acid library to facilitate efficient and thorough screening experimentally. As described in Section 5 above, the profile used to generate the hit variant library II, for example, is also used to determine the size of the nucleic acid library for experimental screening in vitro or in vivo.

Figure 6 illustrates an exemplary procedure for constructing a nucleic acid library to encode the amino acid sequences of the selected amino acid variants, e.g., hit variant library II (**Figures 4 & 5**). To construct the nucleic acid library, the variants in the amino acid profile are back translated into corresponding nucleic acids by taking into account of the library size and codon usages (**Figure 6**).

For example, to obtain the simplest and smallest nucleic acid library covering the diversity of a given amino acid library, only the preferred codons used in the expression system (e.g., E. coli) are selected to encode the amino acid library. The corresponding nucleotide positional variant profile (NT-PVP) is obtained from the back translation of the AA-PVP and the nucleic acid library size is determined from the nucleotide combinatorial enumeration. See example in **Figure 13A-C**. If this size is less than 10^6, synthesis of the nucleic acid library or libraries (e.g., nucleic acid library I, II, III, etc., **Figure 7**) is performed and experimental screening is then conducted. If the size is greater than 10^6, the hit variant library II is partitioned into a shorter library or the scoring distribution is resampled to generate a new AA-PVP to generate a smaller library size, as described in section 2 under sequence space or reprofile.

By using NT-PVP, a degenerate nucleic acid library can be constructed without synthesizing each of the selected nucleic acid sequences individually. This approach reduces cost and time because

the synthesis of the nucleic acid libraries can be accomplished in one pass for each library (e.g., nucleic acid I, II, III, etc., **Figure 7**) by programming an automated oligonucleotide synthesizer with different mixtures of nucleotides for each position. As a result, the sequence space of the degenerate nucleic acid library is significantly expanded with an increase in diversity. Although the size of the nucleic acid library (translated as hit variant library III) is larger than the one faithfully encoding the designed amino acid sequences (e.g., hit variant library II), this approach of degenerate library construction not only guarantees to include the designed sequences but also promises to increase the chance of finding novel sequences with equivalent or better functions than the originally designed ones.

For reassurance, the nucleic acid library generated by using NT-PVP is translated back to an amino acid sequence library to generate hit variant library III and scored using an energy function to evaluate the sequence and structure space covered by the hit variant library II and the fitness of the library (**Figure 13A**). The ultimate comparison requires experimental selection data to validate the fitness of the libraries and the effectiveness of the scoring function in mapping the sequence and structure space onto the fitness landscape.

8. Construction of Mutant Libraries with No Structure Available.

Mutant libraries can be constructed by partitioning sequence libraries into smaller segments. This is advantegious if only low resultion structure or no structure is available. A composite library is designed by partitioning sequences into overlapping consecutive sequence segments. Each fragment can be targeted with a degenerate nucleic acid library. It should be noted that if even low resolution structural model or other structural information is available, the variants that are determined to be structurally coupled should be targeted simultaneously using degenerate nucleic acid libraries (see example below). The idea has been described in 7) of Section **2** and is illustrated in Example below (see **Figures 28A-D** for design and **Figures 30 and 36** for experimental results).

In brief, sequence variant library can be parsed into smaller fragments as follows: the structurally distant segments are often uncorrelated so that mutations widely separated can be treated independently, whereas those fragments that couple with each other in space should be targeted simultaneously by the combinatorial nucleic acid libraries. It should be noted that the structural information is desirable but not absolutely necessary in this case. (see details in Example below and **Figures 28A-D**).

Advantages of the present invention

By sampling a large combinatorial space of amino acid sequence and structural motifs and scoring the intermolecular interactions between proteins, a library of amino acid sequences can be screened computationally. For the specific antibody-antigen complex used here, several libraries of the antibody are designed and constructed based on the lead sequence alone, the antibody structure and the complex structure between the antibody and antigen, respectively. All of the libraries are biased towards the lead antibody, either its sequence and/or structure; some of them are directed towards the specific antigen in the complex. Thus, the antibody libraries are more focused and relevant than a collection of antibodies from a cDNA library or from a random mutagenesis of a specific antibody lead. These libraries are screened experimentally for affinity maturation with the specific antigen. Various sequences different from the lead antibody sequence in CDRs were selected (see **Figures 16A and 27**). Some of the selected sequences show slower off-rate (suggesting higher affinity) than that of the lead antibody (or parental antibody). Among them, two of the mutants (see **Figures 30 & 36**) are identical to the critical mutants in the affinity-matured V_H CDR3 sequence reported in the literature such as (H97Y and/or S101T), whereas one novel mutant (S101R) was found to be even better in off-rate panning, determined by two independent experimental systems, than S101T reported in the literature (Chen Y, Wiesmann C, Fuh G, Li B, Christinger HW, McKay P, de Vos AM (1999) J Mol Biol 293, 865-881).

The present inventions are believed to be advantageous in several aspects. First, this approach utilizes evolutionary data of proteins to expand the hit library in both sequence and structure spaces. The sequence searching methods, ranging from a simple BLAST to the increasingly powerful profile based approaches, such as PSI-BLAST and/or HAMMER, are employed to search for close as well as remote homologues of a lead sequence from the evolutionarily enriched sequence database. The use of sequence profile based on the multiple structure alignment of the available lead structure allows the sampling of a larger sequence space than by traditional, multiple sequence alignment approaches. The methods used here, therefore, increase the diversity as well as the chance to find novel hits or combination of mutants with enhanced binding affinity.

Second, the sampling in sequence space also emphasizes the choice of sequence database suitable for the specific purpose. For example, the use of the diverse sequence database for designing CDRs and the use of the human germlines or sequences of human origins for the framework regions should be exploited in designing proteins for pharmaceutical applications where immunogenicity is a major concern.

Third, sequence design using existing sequences from various databases is simple and highly efficient since only evolutionally enriched sequences or their combinations are used. A refined, yet computationally expensive scoring function can be applied to score the resulting sequence pool of manageable size, that incorporates, implicitly, the information involving folding and expression.

Fourth, the implementation of the structural template and optimized scoring function can efficiently filter and reduce the size of the combinatorial hit variant library prior to any experimental screening. Thus, a large virtual sequence space can be computationally sampled and subsequent selection of ensembles of favorable sequences can direct the experimental synthesis of several small libraries that cover a diverse sequence space.

Fifth, the control of the library size (which is usually around 10^3 to 10^7 for nucleic acid library) may make it easier to implement experimentally for direct functional screening. Because the direct

functional screening is the ultimate test on the validity and accuracy of the in silico methods, some intrinsic limits related to scoring function and structure template in the computational screening can be tested experimentally.

Sixth, the use of simple structural correlation to partition long sequences allows the control of the library size so that it is experimentally manageable without a significant loss of diversity. It also makes it possible to design sequence libraries for a lead sequence with little structural information available.

Finally, the adaptability and parameterization of the scoring function permits refinement with each experimental cycle. The experimentally screened clones represent an actual positional variant in a profile that can be used as a feedback for refining the scoring function by refining the various scoring terms.

In summary, exploring the function space by combining direct experimental screening, within experimental limit, with indirect computational screening in sequence and structure space of a target protein is a powerful approach to protein engineering and design as we demonstrate here for antibodies.

EXAMPLE

Methods of the present invention were used for in silico construction of antibody libraries. The vascular endothelial growth factor (VEGF) is chosen as the antigen for the present proof-of-principle experiments in order to demonstrate the present invention in antibody design. A rich collection of sequence and structure information is available for VEGF and it receptor (Muller YA, Christinger HW, Keyt BA, de Vos AM (1997) Structure 5, 1325-1338; Wiesmann C, Fuh G, Christinger HW, Eigenbrot C, Wells JA, de Vos AM (1997) Cell 91, 695-704), a complex between VEGF and its humanized antibody (Muller YA, Christinger HW, Li B, Cunningham BC, Lowman HB, de Vos AM (1998) Structure 6, 1153-1167, and a complex between VEGF and its matured antibody (Chen Y, Wiesmann C, Fuh G, Li B, Christinger HW, McKay P, de Vos AM (1999) J Mol Biol 293, 865-881). These provide a good platform for testing the methods of the present invention. By using the

methods provided by the present invention, several digital libraries of anti-VEGF antibodies were designed in silico by utilizing incrementally enriched information from an antibody sequence, the structure of an antibody, the complex structure between an antibody and its antigen. Populations of the antibody libraries were screened in vitro for high affinity binding to VEGF via two independent novel phage display systems with antibody binding unit in single or double chains.

1. In silico Design of Anti-VEGF Antibody Libraries

VEGF is a key angiogenic factor in development and is involved in the growth of solid tumor by stimulating endothelial cells. A murine monoclonal antibody was found to block VEGF-dependent cell proliferation and slow the tumor growth in vivo (Kim KJ, Li B, Winer J, Armanini M, Gillett N, Phillips HS, Ferrara N (1993) Nature 362, 841-844). This murine antibody was humanized (Presta LG, Chen H, O'Connor SJ, Chisholm V, Meng YG, Krummen L, Winkler M, Ferrara N (1997) Cancer Res. 57, 4593-4599; Baca M, Presta LG, O'Connor SJ, Wells JA (1997) J Biol Chem 272, 10678-10684) and affinity-matured by using phage-display and off-rate selection (Chen Y, Wiesmann C, Fuh G, Li B, Christinger HW, McKay P, de Vos AM (1999) J Mol Biol 293, 865-881). X-ray structure for the complex formed between VEGF and the parental antibody was reported (Muller YA, Chen Y, Christinger HW, Li B, Cunningham, BC, Lowman HB, de Vos AM (1998) Structure 6, 1153-1167.), as well as the one formed between VEGF and the matured antibody (Chen Y, Wiesmann C, Fuh G, Li B, Christinger HW, McKay P, de Vos AM, Lowman HB (1999) J. Mol Biol 293, 865-881).

Figure 9A shows the amino acid sequences of the variable regions of the humanized anti-VEGF antibody (therein after referred to as "parental anti-VEGF antibody") and the antibody affinity matured from the humanized anti-VEGF antibody (therein after referred to as "matured anti-VEGF antibody"). Each of the amino acid residues in the V_H CDRs that were observed to be in contact with the antigen is labeled as "c" underneath. **Figure 9B** is an alignment of the parental and matured anti-VEGF antibody in the V_H CDRs. The framework and CDRs are designated according to the Kabat criteria (Kabat EA, Redi-

Miller M, Perry HM, Gottesman KS (1987) Sequences of Proteins of Immunological Interest 4th edit, National Institutes of Health, Bethesda, MD). Differences in amino acid residues are highlighted in bold letter. As shown in **Figure 9B**, the matured antibody only has two amino acid residues that are different from the parental one in both V_H CDR1(T28D and N31H) and V_H CDR3 (H97Y and S100aT). There is no change in CDR2 after the affinity maturation.

The matured anti-VEGF antibody has a 135 times higher binding affinity to VEGF than the parental one with 4 mutations in the V_H chain (T28D, N31H, H97Y, and S100aT). The two of the mutations in V_H CDR3 individually improve binding affinity by 14-fold (from H97Y) and 2-fold (from S100aT) relative to the parental antibody (see Table 6 of Chen Y, Wiesmann C, Fuh G, Li B, Christinger HW, McKay P, de Vos AM, Lowman HB (1999) J. Mol Biol 293, 865-881). The 14-fold affinity improvement by H97Y alone in V_H CDR3 makes it the single most important mutation for affinity maturation, which is consistent with observation in the x-ray complex structure that two additional H-bonds are made by H97Y mutant between the antigen and antibody.

According to the present invention, each motif such as CDR and framework of the antibody can be targeted using a modular in silico evolutionary design approach. This modular design is depicted in **Figure 8**. It has been understood that there are only a limited number of conformations (called canonical structures) for each CDR. These structural features of an antibody provide an excellent system for testing the evolutionary sequence design by using structured motifs at various regions of an antibody, such as CDR1, CDR2, and CDR3 in V_L & V_H as well as the framework regions from the extensive analysis of antibody structures. These structure and sequence conservation are observed across different species. In fact, the scaffolding of antibodies, or the immunoglobin fold, is one of the most abundant structure observed in nature and is highly conserved among various antibodies and related molecules.

The inventors believe that parental anti-VEGF antibody described above can serve as a lead protein in a model system for directed antibody affinity maturation using the methods of the present invention. The matured anti-VEGF antibody (Chen et al., *supra*) can

serve as a reference or positive control to validate the results obtained by using the inventive methods.

In addition, structural superposition revealed that the structure of the complex formed between VEGF and the parental antibody almost overlaps with that formed between VEGF and the matured antibody. Since the antibody structures before and after affinity maturation remain substantially the same, structures of both parental and matured antibodies were used in the design of digital libraries of anti-VEGF antibodies using the inventive methods. The inventive method can be also used to design antibodies with induced fit upon antigen binding using sequence-based approach or structure ensembles that contain the induced structure changes.

Using parental anti-VEGF antibody as the lead protein and its V_H CDR3 as the lead sequence, digital libraries of V_H CDR3 were constructed by following the procedure outlined as Route IV in **Figure 1D** and the diagram in **Figure 2**.

The lead sequence included V_H CDR3 of parental anti-VEGF antibody and a few amino acid residues from the adjacent framework regions (**Figure 9B**). As an overview, a hit library was constructed by searching and selecting hit amino acid sequences with remote homology to V_H CDR3. Variant profile was built to list all variants at each position based on the hit library and filtered with certain cutoff value to reduce of the size of the resulting hit variant library within computational or experimental limit. Variant profiles were also built in order to facilitate i) the sampling of the sequence space that covers the preferred region in the fitness landscape; ii) the partitioning and synthesis of degenerate nucleic acid libraries that target the preferred peptide ensemble sequences; iii) the experimental screening of the antibody libraries for the desired function; and iv) the analysis of experimental results with feedback for further design and optimization.

The lead structural templates were obtained from the available X-ray structures of the complexes formed between VEGF and anti-VEGF antibodies. The complex structure of VEGF and parental anti-VEGF antibody is designated as 1BJ1, and that formed between VEGF and matured anti-VEGF antibody 1CZ8. The results from 1CZ8 structural template were similar to those from 1BJ1 in the relative

ranking order of the scanned sequences.

1) Lead sequence

The lead sequence for V_H CDR3 is taken from the parental anti-VEGF antibody according to Kabat classification with amino acid residues CAK and WG from the adjacent framework regions flanking the V_H CDR3 sequence at N- and C- terminus, respectively (**Figure 9B**). As shown in **Figure 9B**, V_H CDR3 of the parental and matured antibodies differ only at two amino acid positions. Only V_H CDR3 sequence of the parental antibody was used to build the HMM for searching the protein databases.

2) Hit Library and Variant Profile

The HMM built using the single lead sequence, SEQ ID NO: 5 (**Figure 9B**), was calibrated and used to search the Kabat database (Johnson, G and Wu, TT (2001) Nucleic Acids Research, 29, 205-206). All sequence hits that are above expectation value or E-value are listed and aligned using HAMMER 2.1.1 package. After removing the redundant and the matured sequence (i.e., SEQ ID NO: 6 by assuming that no matured sequence is available) from the hit list, the remaining 107 hit sequences for the lead HMM form the hit library.

As shown in **Figure 10A**, the 107 hits have sequence identities ranging from 35 to 95% of the lead sequence from the Kabat database. The evolutionary distances between the hits are displayed in a phylogram in **Figure 10B** by using the program TreeView1.6.5 (http://taxonomy.zoology.gla.ac.uk/rod/rod.html). The phylogenetic tree was analyzed using the neighbor-joining method (Saitou N, Nei M (1987) Mol Biol Evol 4, 406-425) in ClustalW 1.81 (Thompson JD, Higgins DG, Gibson TJ (1994) Nucleic Acids Research 22, 4673-4680).

The variant profile at each position is shown in **Figure 11**. The AA-PVP table in **Figure 11** gives the number of occurrence of each amino acid residue at each position. The variant profile below the table lists, in the order of decreasing occurrence at each position, all variants found from the database with the lead sequence as the reference sequence. The dot indicates that the same amino acid as in the reference is found at that position.

The diversity of the 107 hit sequences from the hit library can be seen in the AA-PVP table that shows both the frequency and variability of amino acids at each position. Comparing the difference between sequences of the parental and matured anti-VEGF antibody in V_H CDR3, two different amino acids (H97Y and S100aT using the numbering in the Kabat system) are included in variants listed at each position. The H97Y which was reported to be the most important mutant to increase the binding affinity of the matured sequence (Chen Y, Wiesmann C, Fuh G, Li B, Christinger HW, McKay P, de Vos AM, Lowman HB (1999) J. Mol Biol 293, 865-881) is readily identified as the most frequent residue (~27%) in that position. The S100aT accounts for ~5% of the variants identified in that position. The right lower portion of Figure 11 shows the variant profile after filtering variants that occur at or less than the cutoff frequency of 10. After the filtering, it becomes clear that only a limited numbers of variants are allowed at each position of the sequence; however some important mutants such as S100aT in the matured sequence might be missed at such a cutoff although energy scoring would keep it.

The variant profile from the evolutionary sequence pool provides informative data to identify the positions in the lead sequence that can be either varied or fixed. The sites can be divided into three categories: i) Structurally conserved sites remain conserved over evolution. The high frequency residues can be used to maintain the scaffold of the target motif at these positions; ii) variable functional hot spots should be targeted with focused mutagenesis; iii) combination of both i) and ii) to stabilize the target scaffold while simultaneously providing variability in the functional hot spots.

A set of the amino acids from the functional variants should be included at the functional hot spots according to their frequencies in the variant profile because they are evolutionarily selected or optimized. Furthermore, the variants at each position can be filtered or prioritized to include other potentially beneficial mutants or exclude potentially undesirable mutants to meet the computational and experimental constraints.

3) Structure-based evaluation of combinatorial sequences of the hit library

Although the variant profile is informative on the preferred amino acid residues at each position and specific mutants in a preferred order, unmodified, it embodies an enormous number of recombinants. Some filtering using frequency cutoff can reduce the combinatorial sequences that need to be evaluated by computational screening or targeted directly by experimental libraries. Even with the cutoff applied to the variant profile, there is still a large number of combinatorial sequences that needs to be scored and evaluated in the final sequences for experimental screening (as shown in **Figure 13A-C and 28A-D**).

A structure-based scoring is applied to screen the hit library and its combinatorial sequences that form a hit variant library. Side chains of V_H CDR3 of the parental anti-VEGF antibody were substituted by rotamers of corresponding amino acid variants from the hit variant library at each residue position. The conformations of rotamers were built and optimized by using the program SCWRL® (version 2.1) using backbone-dependent rotamer library (Bower MJ, Cohen FE, Dunbrack RL (1997) JMB 267, 1268-82).

The scoring was done by searching the optimal rotamers and minimizing the energy by 100 steps using the Amber94 force field in CONGEN [Bruccoleri and Karplus (1987) Biopolymers 26:137-168] in the presence and absence of the structure of the antigen VEGF. Figure 12A & B shows the energy scores of an anti-VEGF variant library based on the total energy calculated with CONGEN with and/without VEGF antigen, using the structures of the parental (1bj1) and matured (1cz8) anti-VEGF antibodies, respectively. The scores of the parental and matured sequences are marked in **Figures 12A and B**. The matured sequence scores better than the parental sequence in both structures with/without antigen, suggesting that the mutants of the matured sequence stabilize both the antibody structure as well as its complex with VEGF antigen. **Figure 12C** shows that the scoring of the sequences in the presence and absence of antigen is in general correlated, which suggests that screening sequences based on a antibody structure alone would also provide good candidate sequences with good binding affinity with its antigen.

As shown in **Figures 12A and 12B**, there are a large number of sequences for various variant libraries with higher scores than the

parental and matured sequences. The distribution of the energy scoring in the energy diagram is shown in **Figure 13A** for 10 selected sequences from the hit variant library of V_H CDR3, its combinatorial peptides, combinatorial library of the degenerate nucleic acid library, and the experimentally selected sequnences. The scoring shows that Y97 in the matured sequence always scores better than H97, consistent with the experimental observation (Chen Y, Wiesmann C, Fuh G, Li B, Christinger HW, McKay P, de Vos AM, Lowman HB (1999) J. Mol Biol 293, 865-881). T100a is preferred over S100a as found in the matured sequences, whereas both T and S are equally preferred in 100b position. Thus, the structure-based energy scoring provides another independent way to reprofile the occurrence of variants at each position for the hit variant library which was originally built based on profiling of evolutionary sequences selected from protein databases.

In order to gauge the accuracy of the scoring function using the simple energy function implemented in CONGEN, the energies of a randomly selected set of sequences were calculated using a refined custom scoring function that includes sidechain entropy, nonpolar solvation energy and electrostatic solvation energy. Three energy terms were calculated: sidechain entropy, nonpolar solvation energy and electrostatic solvation energy. There was an additional option to calculate the backbone entropy for loops. The sidechain entropy was calculated using the conformational search command CGEN in CONGEN. Options under CGEN were defined to perform individual sidechain conformational tree search using the torsion space at each bond (node) to expand the tree. These included the SEARCH DEPTH and SIDE option for each sidechain with the SGRID parameter set to AUTO so that each torsion angle was rotated at discrete intervals. Specifically, the AUTO setting used torsion grid angle of 30 degrees for bonds with rotational symmetry such as in the phenyl, tyrosyl, carboxyl, and amino groups, and 10 degrees for all others. MIN option set rotational sampling to start at a local energy minimum for each specified torsion. Also VAVOID option was included to turn on van der Waals repulsion avoidance. MAXEVDW parameter was set to a relatively high 100 kcal/mol so as to relax the van der Waals repulsion, leading to a higher number of conformers in the enumeration.

This sidechain conformational search was repeated for each mutant residue sidechain. The code outputs the "number of bottom leaves" reached by the tree search in conformational space which is the number of completed tree search. As an approximation, the sidechain conformational search treats each residue independently, so that computational time can be minimized. For residues that do not contact one another, this is a good approximation. For residues that can potentially contact one another, the conformational enumeration will tend to overestimate the number of conformations. Since we use a relatively high van der Waals repulsion in order to obtain a larger sampling, the error due to residue contacts should be reduced in the context of this artificial gauge of the conformational space. Furthermore, the significance of the error due to residue contacts will tend to diminish with greater number of conformations since the relative change in entropy is a difference of the logarithms of the number of conformations in the mutant and the reference structures.

The nonelectrostatic solvation energy is made proportional to the molecular surface, as calculated by the GEPOL93 algorithm, with the scaling constant of 70 cal/mol/A^2 (Tunon I, Silla E, Pascual-Ahuir JL (1992) Prot Eng 5, 715-716) using GEPOL (Pascual-Ahuir JL, Silla E (1993) J Comput Chem 11, 1047-1060) command as implemented in CONGEN. NDIV which specifies the division level for the triangles on the surface is set to 3. Values range from 1 to 5 with 5 giving the highest accuracy but with significant increase in CPU time requirement. RGRID is set to 2.5A and describes the space grid used to find neighbor.

The electrostatic solvation energy is calculated using the finite-difference PB (FDPB) method as implemented in UHBD program (Davis ME, Madura JD, Luty BA, McCammon JA (1991) Comput Phys Commun 62, 187-197). The focusing method is used for the region surrounding the mutation. An automated protocol generates three grids: coarse, fine, and focus grids. The grid units are 1.5, 0.5, and 0.25 angstroms, respectively. The focusing grid is a cubic grid that spans the Cartesian volume occupied by the mutated residues. The fine grid is a cubic grid that spans the entire volume of the protein or the complex. The coarse grid is a cubic grid that is set to approximately twice the size of the fine grid in each axis and covers approximately 8 times the

volume of the fine grid. The coarse grid serves to account for the long-range solvent effects and sets the boundary conditions for the fine grid. Similarly, the fine grid accounts for the electrostatic contributions of the protein interior and sets the boundary condition for the focus grid. The focus grid accounts for finer details of the localized effects due to the mutation. The dielectric constants for the protein interior and exterior are set to 4 and 78, respectively. Temperature is set to 300 Kelvin and ionic strength is set to 150 mM. Maximum iteration is set to 200. The calculations are repeated with a uniform dielectric so that both the interior and exterior dielectrics are set to 4 and the difference between the two energies is computed. The latter calculations represent the energies due to bringing the charges onto the grids.

It was shown that the custom scoring function or the molecular mechanics energy using Amber94 forcefield in CONGEN plus the solvation terms from PB in UHBD used here is similar to MM-PBSA or MM-GBSA. The energy function shows better agreement with experimental data (Sharp KA. (1998) Proteins 33, 39-48; Novotny J, Bruccoleri RE, Davis M, Sharp KA (1997) J Mol Biol 268, 401-411), especially when structure ensembles by molecular dynamics calculations are used to provide more accurate methods to score sequence and its variants based on the ensemble averages of the energy functions (Kollman PA, Massova I, Reyes C, Kuhn B, Huo SH, Chong LT, Lee M, Lee TS, Duan Y, Wang W, Donini O, Cieplak P, Srinivasan P, Case DA, and Cheatham TE (2000) Acc. Chem Res. 33, 889-897).

4) Reduction of the variant profile of the hit variant library

The variant profile from the hit variant library as described above was filtered in order to reduce the potential library size while maintaining most of the preferred residues. The upper portion of **Figure 13A** shows the reduced variant profile of 10 selected sequences with top ranking from a hit variant library after eliminating amino acids with occurrences lower than the cutoff value and structure-based evaluation. The list was chosen as a blind test on the validation of the current method in selecting for diverse sequences that can bind with a target antigen. There are some common features shared among 10 selected sequences from one computationally screening variant library:

R94, Y97 and R100a are found always better than the corresponding residues at K94, H97 and S100a, for example for the top ranked 200 sequences using either 1bj1 or 1cz8 as the template structure in the presence or absence of VEGF antigen. As shown in the experimental selection later that H97Y is indeed a good mutant for affinity maturation. However, mutation such as K94R and S100aR into arginine is an interesting case: on the one hand, K94R is not a good mutant for affinity maturation although K94R lies in the boundary between CDR and framework according to Kabat classification and is preferred evolutionally for human framework sequence. K94 is favored over R94 as shown in experimental selection of the current invention (**Figures 30 & 36**), consistent with the observation in literature that R94K mutation increases the binding affinity of anti-VEGF antibody (Baca M, Presta LG, O'Connor SJ, Wells JA (1997) J Biol Chem 272, 10678-10684). One the other hand, S100aR turns out to be one of the most important single mutation for VH CDR3 maturation, it is favored over S100aT as reported in the literature and persist many rounds of panning under harsh washing conditions in phage display (see **Figures 30 & 36**).

In order to avoid missing some important mutants in a variant profile, some residues such as lysine (such as K94R) from the wild type might be included even though they are below the cutoff value used in filtering hit library or they somehow score less well as arginine because of the problems associated with assumption in computation involving charged residues with long side chains or conformational change etc. Therefore, for charged residues with long side chains, such as arginine and lysine, the predicted residues as well as the wild type residue at the same position might be included in the design libraries. The reduced variant profile was used to enumerate hit variant library II as the blind test on the inventive method used here for designing functional library with diverse sequences from the lead sequence.

5) Hit variant library II–an amino acid library designed from scoring selection and optimization

A strategy that selects top sequences based on favorable score and/or the presence of residues likely to participate in favorable

interactions was employed to identify a cluster or clusters of amino acid sequences for the nucleic acid library design (**Figure 7**). As described above, a cluster of sequences (e.g., 10 sequences) in **Figures 13A-C** for V_H CDR3, CDR1 and CDR2, respectively, from computational evaluation was chosen for further experimental test in vitro. The peptide sequence and variants at each position are listed in upper left portion of **Figure 13A**. A combinatorial library was generated based on the filtered variant profile, forming hit variant library II. For V_H CDR3 of anti-VEGF (**Figure 13A**), the size of hit variant library II is 72 based on the variant profile of the selected top 10 sequences with scores better than the lead sequence (the top 10 ranked sequences among the variant library used). See Figures **13B and C** for V_H CDR1 and CDR2.

6) Construction of degenerate nucleic acid library based on hit variant library II

The hit variant library constructed above was targeted with a single degenerate nucleic acid library. The lower portion of **Figure 13A** shows a nucleic acid sequence profile resulting from back-translation using the optimal E. coli codons for V_H CDR3. Based on this profile, a degenerate nucleic acid library was synthesized by incorporating a mixture of bases into each degenerate position. As a result of the combinatorial effect of the synthesis, this degenerate nucleic acid library encodes an expanded amino acid library (designated "hit variant library III") with a size of 4608. See **Figures 13B and C** for V_H CDR1 and CDR2.

The degenerate nucleic acid library constructed above was cloned into a phage display system and the phage-displayed antibodies (ccFv) were selected based on their binding to immobilized VEGF coated onto 96-well plates. As will be described in more detail in section 2 below, with a small nucleic acid library size, one to three round of washing and selection (i.e., panning) were performed and clones showing positive ELISA reaction were selected and sequenced as shown in **Figure 14B** for V_H CDR3. The positive clones show a diverse variant profile at targeted position with the incorporation of degenerate codons into the nucleic acid library.

The results of the designed versus the experimentally screened

antibody sequences are analyzed in **Figures14-18**. In brief, the sequences for V_H CDR1,2,3 have been designed based on the inventive method described above in detail for V_H CDR3. Top 10 sequences and their variant profiles selected from the computationally screened libraries for V_H CRD3, CDR2 and CDR2, respectively, are shown **Figures 13A-C**. **Figure 16A** is a table that lists the experimentally selected amino acids sequences from V_H CDR1, CDR2 and CDR3 libraries of degenerate nucleic acids shown in **Figures 13A-C**. **Figure 16B** shows the distribution of the sequence identities of selected sequences from V_H CDR1, CDR2 and CDR3 libraries relative to the corresponding parental sequence of anti-VEGF V_H CDR1, 2,3 respectively. **Figure 17A** shows the relationship among 4 different libraries (designed amino acid sequences, the combinatorial library of amino acid variant of the designed sequences, and combinatorial degenerate nucleic acid libraries encoding the unique amino acid sequences and the entire degenerate nucleic acid library) and the distribution of the experimentally selected positive clones shown in X, using anti-VEGF V_H CDR3 library from round 3 as an example (see table in **Figure 17B**). The distribution among different libraries depends on selection conditions, the effectiveness of library design, the relative size of the selected colons versus library or number of sequenced clones etc. **Figure 17B** shows a table delineating the relationships among the four libraries (**Figure 17A**) and the distribution of the experimentally selected sequences of the positive clones for anti-VEGF V_H CDR1, 2, 3 library.

Detailed analysis for V_H CDR3 is discussed below. **Figure 14A** shows UV reading of the ELISA positive clones identified in round 1 and round 3 selections of functional anti-VEGF ccFv antibodies with V_H CDR3 encoded by the designed nucleic acid library (**Figure 13A**). **Figure 14B** shows V_H CDR3 sequences of the positive clones from round 1 and 3 selection via phage display of the nucleic acid library shown in **Figure 13A**. It is clear that many diverse sequences are selected with large variations at several positions that are different from V_H CDR3 of parental and matured anti-VEGF antibody (**Figure 9B & C**). **Figure 14C** illustrates a phylogenic tree of the positive clones showing the diversity of the screened sequences. The sequence identities of the

selected positive clones from V_H CDR3 shown in **Figures 14B** ranged from 57 to 73 percent relative to the parental V_H CDR3 sequence. **Figures 15A-B** are pie charts showing the breakdown of the origins of the screened sequences in the first and third rounds into three groups: designed amino acid sequences, combinatorial amino acid sequences from the designed sequences, and the unique combinatorial amino acid sequences encoded by the synthesized degenerate nucleic acid library. Because only limited number of positive clones from each round are selected for sequence analysis, the figures are only used to illustrate percentage of the selected sequences from designed, its combinatorial amino acid and nucleic acid libraries.

These experiments demonstrated that by using the methods of the present invention, antibodies could be selected, not only with diverse sequences and phylogenic distances, but also with relevant biological function, e.g., ability to bind to the target antigen such as VEGF.

Figure 18 summarizes the progressive evolution of the sequence design using the scoring results for amino acid sequences at each stage for V_H CDR3 as an example. From left to right, the diagram shows the energy spectra for the lead sequence, the hit library generated from the database search, computationally screened combinatorial sequences in the hit variant library I, a selected group of designed amino acid sequences (hit variant library II), a degenerate nucleic acid library derived from library II profile, and experimentally screened positive clones and sequences. The process can be iterated with feedback from experiments until the sequences with enhanced or desired properties are selected experimentally.

Figure 19A-D show the comparison of the sequence homology distribution based on a lead sequence or a lead sequence derived from a multiple structure-based alignment. **Figure 19A** shows the lead profile generated from structure-based mutiple seqeuce alignment. The structural motif of the lead sequence is used to search protein structure database (PDB databank) for similar structures within certain distance cutoff. The five structures are superimposed using Ca atoms of the V_H CDR3. The average root mean squire difference (RMSD) between each structure and V_H CDR3 structure motif (colored in magenta) is within 2

Å. The corresponding mutiple sequence alignment is shown in the right of **Figure 19A**, together with their PDB IDs and color of the corresponding structure.

Figure 19B shows a variant profile for the 251 unique sequences of the hit library generated based on the lead sequence profile of V_H CDR3 of parental anti-VEGF antibody. The lower portion of the figure shows a filtered variant profile obtained by using a 5% cutoff of the frequency or 12 in this case. Interestingly, important mutants (H97Y and S100aR or S100aT, see **Figures 30 & 36**) are also observed in the variant profile generated from the lead sequence profile.

Figure 19C shows the distribution of the sequences from the hit library relative to the parental V_H CDR3 sequence. The circles indicate the sequence identity up to 36% can be identified using the single parental sequence for HMM search. The triangles indicate that even lower sequence identity up to ~20% can be found using the lead sequence profile from a structure-based multiple sequence alignment. The sequence searching strategy used here can find diverse hits with remote homology (as low as 20%) to the lead sequence.

Figure 19D shows the conceptual evolution of the inventive methods used here to search for promising candidates in sequence, structure and function spaces. The basic idea here is to expand the diversity of hits and variant libraries in sequence and structure space in order to find the candidates with improved function in function space. While the diversity and/or the size of the hit and variant library is increased by, for example, finding remote homologues of the lead sequence or sequence profile (as shown in **Figure 19A**), the intersection among the sequence, structure and function spaces can be focused into a smaller region with increased probability of finding sequences with enhanced function.

It is clear that using structurally-based multiple sequence alignment as the profile to build the HMM model makes it possible to find remote homologues (to 20% sequence identity of the query sequence) of a lead sequence. The inventive method described here will become more powerful for designing antibody CDR libraries with the increase in available sequence and structure information and improvement in the accuracy of the scoring functions.

2. Functional Screening of Designed Antibody Libraries in Vitro

The antibody libraries that were designed in silico, based on a lead sequence of the parental anti-VEGF antibody by using the methods described above were tested for their ability to bind to the antigen, VEGF, by using a novel phage display system. The structure of either the parental antibody or matured antibody would be used for structure-based computational screening. In contrast to the popular approach of screening antibodies adopting a form of single chain antibody (scFv) (see another novel method shown in Figure 20 & 32), a two-chain antibody library was expressed and displayed on the surface of bacteriophage. The two-chain antibody is formed by heterodimerization of V_H and V_L to functionally mimic the Fab of antibody. This two-chain antibody is designated as "ccFv". The ccFv library was constructed based on the degenerate nucleic acid library encoding sequences of the antibodies designed in silico as described above.

Described in detail below are the rationale for designing the ccFv, construction and expression of the ccFv library, and functional screening of the ccFv library.

1) ccFv—a heterodimeric coiled-coil stabilized antibody

The antibody Fv fragment is the smallest antibody fragment containing the whole antigen-binding site. The Fv fragments have very low interaction energy between their two V_H and V_L fragments, and are often too unstable for many applications at physiological condition. Naturally, V_H and V_L domain are linked by an interchain disulfide bond located in the constant domains, C_H1 and C_L, to form a Fab fragment. It has also been shown that the V_H and V_L fragments can also be artificially held together by a short peptide linker between the carboxy-terminus of one fragment and amino-terminus of another to form a single-chain Fv antibody fragment (scFv).

The present invention provides a new strategy to stabilize V_H and V_L heterodimer. A unique heterodimerization sequence pair was designed and used to create a Fab-like, functional artificial Fv fragment ccFv (**Figure 20**). Each of the heterodimeric sequence pair was derived

from heterodimeric receptors $GABA_B$ R1 and R2, respectively. This sequence pair specifically forms a coiled-coil structure and mediates the functional heterodimerization of $GABA_B$-R1 and $GABA_B$-R2 receptors. For the purpose of engineering a heterodimer of V_H and V_L of an antibody, $GABA_B$-R1 and $GABA_B$-R2 coiled coil domains (GR1 and GR2, respectively) are fused to the carboxy-terminus of V_H and V_L fragment, respectively. Thus, the functional pairing of V_H and V_L, ccFv (coiled coil Fv), is mediated by specific heterodimerization of GR1 and GR2. Furthermore, the carboxy-termini of GR1 and GR2 domains were modified by adding a flexible spacer or flexon "SerArgGlyGlyGlyGly" [SEQ ID NO: 7] (or "GlyGlyGlyGlySer" " [SEQ ID NO: 18]) To further stabilize the heterodimeric ccFv, a pair of cysteine residues were introduced by adding "ValGlyGlyCys" [SEQ ID NO: 8] spacer at the C-termini of the GR1 and GR2 coiled coils so that the coiled-coil GR1 & GR2 mediated heterodimer can be linked covalently by disulfide bond (**Figure 20-21**). ccFv were expressed in E.coli with a molecular weight 35 kDa.

2) Anti-VEGF (AM2-ccFv) and its display on phage surface

V_H and V_L sequences of an anti-VEGF antibody AM2 are shown in Fig. 22A-B. This is an antibody designed by modifying the parental anti-VEGF antibody. Unique restriction sites were introduced in both V_H and V_L genes of the parental anti-VEGF antibody to facilitate an efficient cloning of designed CDR sequence libraries. Both AM2 V_H and V_L genes were cloned into a phagemid vector to construct the phage display vector pABMD12. Figures 23A and 23B show the vector map and sequence [SEQ ID NO: 17], respectively. This vector will express two fusion proteins: V_H-GR1 and V_L-GR2-pIII fusions. The expressed V_H-GR1 and V_L-GR2-pIII fusions are secreted into periplasmic space, where they heterodimerize to form a stable ccFv antibody (designated as "AM2-ccFv") via the coiled-coil domain.

To display AM2-ccFv on phage, pABMD12 vector was transformed into bacterial TG1 cells. The TG1 cells carrying the pABMD12 vector were further superinfected with KO7 helper phage. The infected TG1 cells were grown in 2xYT/Amp/Kan at 30°C overnight. The phagemid particles were precipitated twice by PEG/NaCl from

culture supernatants, and resuspended in PBS for library selection against immobilized VEGF. After 2 hours of binding, unbound phages were washed away and bound phages were eluted and amplified for the next round of panning.

Binding of the ccFv displayed on phage particles was detected by antigen binding activity via phage ELISA. Briefly, the antigen (e.g., VEGF) was first coated onto the ELISA plates. After blocking with 5% milk/PBS, the phage solution was added to the ELISA plates. The phages bound to the immobilized antigen were detected by incubation with HRP-conjugated anti-M13 antibody against phage coat protein pVIII. The substrate ABTS [2,2'Azino-bis(3-ethylbenzthiazoline-6-sulfonic acid)] was used for measurement of HRP activity. The assay was shown to be highly specific for AM2.

The single-chain AM2 antibody (AM2-scFv) phage was also prepared for comparison with the AM2-ccFv in phage ELISA described above. As indicated in Figure 24, the apparent binding affinity of AM2-ccFv phage to immobilized VEGF is almost one order of magnitude higher than AM2-scFv phage. Thus, it is concluded that both AM2-ccFv and AM2-scFv are functional when displayed on a phage particle.

3) Enrichment of ccFv phages from a model antibody library

To prove that AM2-ccFv displayed phages can be enriched from background phages, we performed panning experiments to select for AM2-ccFv phage from "model libraries". The model libraries were prepared by mixing of AM2-ccFv phages with an unrelated AM1-ccFv displayed phage at a ratio of $1:10^6$ or $1: 10^7$. Two round of panning on immobilized VEGF antigen were carried out. 100 ul of 2ug/ml VEGF was coated on each well in a 96-wells plate. After blocking with 5% milk in PBS, $1X10^{12}$ library phages in 2% milk/PBS were added to the well, and incubated for 2 hours at room temperature. Phage solution was discarded and wells were washed 5 times with PBST (0.05% Tween-20 in PBS) and 5 times with PBS. Bound phages were eluted with 100 mM triethylamine, and were added to TG1 culture for infection. The phages prepared from infected TG1 cells were used for the next round panning and phage ELISA described above. After each round of panning, the ratio of AM2-ccFv phage to AM1-ccFv phage recovered was

also determined by analysis of infected TG1 colonies via PCR. Due to the difference in the sequences of AM2-ccFv gene and AM1-ccFv gene, a pair of primers was designed to specifically amplify only AM2-ccFv gene, but not AM1-ccFv. As shown in **Figure 25A**, phages from the second round panning yielded very high ELISA reading, suggesting that a high enrichment of AM2-ccFv phages was achieved from both the $1:10^6$ and the $1: 10^7$ libraries after 2 rounds of panning. PCR analysis confirmed that the occurrence rate of AM2-ccFv phage was 4.4% from $1: 10^7$ library after the first round panning, and 100% after the second round of panning (**Figure 25B**).

4) Construction and panning of phage library of designed ccFv antibodies

As diagramed in **Figure 8**, a modular, evolutionary approach was employed to construct an antibody library for computational and experimental screening. The oligos encoding a library of designed CDR sequences were synthesized and amplified by PCR. The primers for amplification contain the restriction sites to clone the synthetic CDR sequences into the pABMD12 vector. Three V_H libraries were prepared for AM2-ccFv, using restriction sites of NheI and XmaI, XmaI and SpeII, and PstI and StyI for the insertion of CDR1, CDR2 and CDR3, respectively. After ligation, DNA was transformed into TG1 cells. Phages were prepared from TG1 cells by KO7 helper phage infection. Three rounds of panning against immobilized VEGF were carried out as described below. 100 ul of 2ug/ml VEGF was first coated onto each well of a 96-well plate. After blocking with 5% milk in PBS, $1X10^{12}$ library phages in 2% milk/PBS were added to the well and incubated for 2 hours at room temperature. The phage containing solution was then discarded, and the wells were washed 5 times with PBST (0.05% Tween-20 in PBS) and 5 times with PBS. Bound phages were finally eluted with 100 mM triethylamine, and were added to TG1 culture for infection. The phages prepared from infected TG1 cells were consequently used for the next round of panning. For each round of panning, 94 to 376 clones were picked for phage ELISA (**Figures 26A and B**). Positive clones from the phage ELISA were amplified by PCR and sequenced. DNA sequences were then translated to amino acid sequences. The coding amino acid sequences from the three libraries

ware listed in a table in Figure 27.

5) Library design based on the sequence with and without constraints from tertiary structure or structural model

Another strategy for designing CDR libraries is to partition the CDR sequences into uncorrelated and correlated segments in structure space in order to detect the covariant mutants at structurally coupled positions such as the N- and C-termini regions of the CDR loops (low resolution structure should be enough in most cases). For example, **Figure 28A** shows a composite variant profile for V_H CDR3 of anti-VEGF antibody obtained by combining a filtered hit variant profile for V_H CDR3 with other variants from experimental selection. We would like to demonstrate that variants from diverse sources can be combined to generate a composite variant profile for library construction. This variant profile is parsed into several segments of smaller variant profile in order to make sure that each smaller variant profile can be covered by a nucleic acid library with a diversity around 10^6-10^7. Note, the combination of the V_H CDR3 mature sequence with H97Y and S101T (S100aT in Kabat) is deliberately avoided in the parsed segment libraries (see **Figures 28A-D**).

Figure 28A-D show the sequence library of anti-VEGF V_H CDR3. The library is parsed into 3 segments: **Figure 28B** covers the N- and C-termini that might contain coupled variants (1-3), **Figure 28C** contains segment (4) and **Figure 28D** contains another segment (5). All three segments are covered by nucleic acid libraries with a size around 10^6: (1-3) in **Figure 28B** are targeted by 3 degenerate nucleic acid libraries, whereas (4) and (5) in **Figures 28 C-D** are targeted by a separate degenerate nucleic acid library.

The rationale for designing these segment libraries is as follows. Structurally distant segments are often uncorrelated so that mutations widely separated in space can be treated independently. For the CDR3 loop, the sequence is partitioned into three segments: the first and third segments (base of the loop) form one profile for library design, whereas apex of the loop is parsed into two profiles for library design with a size of 10^6 in the degenerate nucleic acid libraries. As shown in **Figure 28B**,

fragments at N- and C-termini that couple with each other in space (the sequences forming the base of the loop are generally correlated due to loop closure) should be targeted simultaneously by the combinatorial nucleic acid libraries with only three degenerate oligonucleotides (1-3). Simple criteria such as the C_α or C_β distance matrix can be examined to identify correlated segments (see **Figure 28A** for the structure and distance contact matrix among C_α atoms within 8A). Optionally, a more detailed interaction matrix can be mapped out to explore number and types of interactions, but the underlying principle is the same for identifying correlated segments.

Libraries for the apex, such as (4) and (5) in **Figures 28C and 28D**, are often uncorrelated. They are targeted by degenerate oligonucleotide libraries along the primary sequence in a consecutive fashion as long as each library is limited to the size range that can be managed easily by experiment (< 10^6 in **Figures 28C-D**). There should be positional overlaps between the fragments to maintain a small level of local correlation among the resulting libraries. In a similar fashion, longer segments can be partitioned into overlapping segments to span the length of the sequence and the corresponding libraries can be generated.

The resulting re-profiling can be further modified and enhanced based on observed experimental or structural or computational criteria. These can include varying positions with known hydrogen bonds with additional polar amino acids, region of high van der Waals contacts with bulky aliphatic or aromatic groups, or region which might benefit from increased flexibility with glycine. In an experimental feedback, variants may be added based on assay results from earlier screening as a basis for subsequent design improvement as shown the variant profile in **Figure 28A**. A more sophisticated analysis might take into account the coupling of amino acid groups such as salt bridges or hydrogen bonds within the sequence.

6) Off-rate panning for ccFv library L14

In order to select high affinity antibodies, off-rate panning process was carried out for selection in library L14 (see **Figure 28A-D**).

The strength of the interaction between an antibody fragment on phage surface and an immobilized antigen is measured by their interacting affinity, which is determined by its on-rate (the rate of association) and off-rate (the rate of dissociation). According to previous studies, antibody of high affinity usually bears slow off-rate whereas antibody of low affinity usually bears fast off-rate, whereas their on-rates are similar. The off-rate panning was designed to facilitate the dissociation of those antibodies with lower affinities from immobilized antigen with gradual increase in harshness (stringency) of wash conditions. By applying washes of increasing stringency, phages with lower affinities will be washed away, leaving behind phages with increasingly higher affinities (i.e., the slower off-rates). Therefore, those phages that survive increasingly harsh washing conditions should have higher affinities and those whose occurrence becomes dominant must have higher affinities than those of low occurrence rate. We also demonstrate comparable off-rate panning at the phage level using two independent display platforms (**Figures 20 and 32**) under various panning conditions (**Figures 29 and 35A-B**). The resulting positive clones or consensus of clones from phage panning should suggest strongly that some sequences or variants should possess enhanced affinity with antigen relative to the parental sequences.

L14 was prepared as anti-VEGF V_H CDR3 library by parsing the V_H CDR3 sequence into short overlapping segments (see **Figure 28A-D**). In order to discriminate slow off-rates, a number of panning conditions were manipulated. During the first two rounds of panning, wells were briefly washed 6 times with PBST and PBS to remove phages with lower affinities. Starting from panning 3, the bound phages were further washed with additional hours to remove those with faster off-rates (dissociation). The duration and stringency of such a dissociation period were increased with the number of panning (**Figure 29**) so that more and more phages were allowed to dissociate and to be removed; in contrast, those with slow off-rate (higher affinity) would remain bound and are eventually enriched. As listed in **Figure 29**, panning 3 was performed in PBS for 1 hour at 37°C (PBS was refreshed every 10 min. and a brief wash was applied in between to remove the dissociated phages); panning 4 was performed in PBS for 2 hours at 37°C; panning

5 was performed in PBST for 1 hour at room temperature followed by PBS for 2 hours at 37°C; panning 6 applied an overnight wash in a large volume (20 ml) of PBS at room temperature; panning 7 further increased the temperature (30°C), volume (50 ml), and duration (24 hrs) of the wash. As indicated in **Figure 29**, in addition to changing the wash stringency described above, by lowering the concentration of antigen, the concentration of the phage input, and increasing the temperature of the binding period, dissociation is further enhanced. The surviving clones from the panning were randomly picked and assayed in phage ELISA to confirm their abilities to bind to VEGF. 100% ELISA positive rate was obtained from clones in both panning 5 and 7, suggesting that after panning 5, all survival phages were able to bind to VEGF, and therefore, phages being washed away had faster off-rates. Among the clones that were positive in phage ELISA, 20 clones from panning 5 and 10 clones from panning 7 were randomly picked for DNA sequencing. The coding amino acid sequences for VH CDR3 are summarized in **Figure 30**. The frequency of the wild-type anti-VEGF antibody was 20% in panning 5. After two additional rounds of off-rate panning with high stringency, the frequency of the wild-type sequence dropped to zero in panning 7. In contrast, the HR (H97, R101 or R100a in Kabat) mutant was continuously enriched from 35% in panning 5 to 70% in panning 7 (**Figure 30**), which became the sole dominant clone in the end. The presence of the HT (H97, T101 or T100a in Kabat) mutant (30%) remained unchanged in panning 5 and 7. The enrichment of HR mutant from P0 to P7 is shown in **Figure 31**. These data suggest that both HR and HT mutant have higher affinity than that of wild-type antibody. The affinity of HR mutant should be higher than that of HT mutant, which has a threonine, rather than arginine, at position 101 (or 100a in Kabat), as reported for the matured sequence (Chen Y, Wiesmann C, Fuh G, Li B, Christinger HW, McKay P, de Vos AM, Lowman HB (1999) J. Mol Biol 293, 865-881).

8) Panning of single-chain (scFv) anti-VEGF antibody library by adapter-mediated phage display system.

The off-rate panning strategy was further tested using an

independent system as described below.

In the conventional phage display system, a protein of interest is fused to a phage capsid protein such as pIII in order to be displayed on the surface of phage. This fusion protein will be assembled into phage particles with the wild-type phage proteins provided by a helper phage such as KO7. We have developed a new phage display system named 'adaptor-directed display system'. In general, a protein of interest is carried to the surface of the phage particle by a pair of adaptors that specifically form a heterodimer, one being fused with the displayed protein in an expression vector and the other being fused with a phage capsid protein in a helper vector. The present example for the pair of adaptors is GR1 and GR2, as described above. As illustrated in Figure 32, the protein of interest (scFv anti-VEGF) is expressed as a fusion with an adaptor (GR1) to form a construct of scFv-GR1 in an expression vector (Figure 33A and B). GR2 was inserted in the genome of a helper phage to form a fusion with pIII capsid protein (GR2-CT of pIII, **Figure 33A and B**). As a result, the helper phage with the modified genome is then designated the GMCT Ultra-Helper phage (**Figure 34A and B**). In TG1 cells, the expression vector expresses scFv-GR1, which is then secreted into bacterial periplasmid space. The cells are further infected with GMCT Ultra-Helper phage, which expresses GR2-CT of pIII, also secreted into the bacterial periplasmic space. Therefore, scFv-GR1 and GR2-CT of pIII specifically form a heterodimer through a coiled-coil interaction between GR1 and GR2, which ultimately assembles the scFv onto the surface of the phage.

Using this system, we constructed an anti-VEGF scFv library L17, equivalent to ccFv library L14 described above (anti-VEGF CDR3 V_H synthetic library). Similar to the selection of library L14, off-rate panning was applied. Library DNA was transformed into TG1 cells and then rescued with GMCT Ultra-Helper phage. Phages were prepared following standard protocol and tested for binding against immobilized VEGF in 96-well plate. As indicated in Figure 35A, wells from panning 1 and 2 were first washed 10 times with PBST and then 10 times with PBS at room temperature, followed by a dissociation period in PBST for 1 hour at room temperature (PBST was refreshed every 10 min. and a brief wash was applied in between to remove the dissociated phages);

the dissociation period was increased to 2 hours in panning 3. Using phages recovered from panning 3, two parallel pannings (**Figure 35B**), panning 4 and panning 5, were carried out in order to further enhance the dissociation of phages with lower affinities: 150 ml PBST for 18 hrs at 25°C for panning 4, and at 37°C for panning 5. Ten clones of ELISA positives from panning 4 and 8 clones from panning 5 were picked randomly for sequencing. The data are shown in Figure 36. In panning 4, the presence of WT sequence was 10%. The frequencies of both HT mutant (30%) and HA mutant (30%) were equal. Note that there is no arginine residue shown in position 101 (100a Kabat) among the 10 clones analyzed (**Figure 36**), suggesting its low occurrence at this stage. In contrast, by increasing the dissociation stringency at panning 5, the occurrence of arginine in position 101 (100a Kabat) increases to 50% (4 out of 8 clones) and becomes dominant in panning 5. In comparison, the HT mutant drops from 30% to 12.5% and the WT drops from 10% to 0, consistent with the observation in Figure 30. This result suggests strongly that the HR mutant has a higher affinity than either the HT mutant or the WT.

9) Summary of the library design, diversity and affinity maturation

Results shown in both **Figure 30 and 36** suggest that the off-rate panning of two independent novel phage display systems used here are able to select out a novel mutant, HR (H97, R101 or R100a Kabat). The HR mutant has a higher binding affinity than the corresponding HT (H97, T101 or T100a Kabat) mutant in the reported matured sequence (Figure 9B). Moreover, HR mutant binds the antigen better than the YS (Y97, S101 or S101a Kabat) mutant (see Panning 4 of Figure 36). The YS mutant was reported previously to improve the binding affinity 14-fold relative to the WT and was believed to be the single most important mutant in V_H CDR3 of the matured anti-VEGF antibody (Figure 9B and see Chen Y, Wiesmann C, Fuh G, Li B, Christinger HW, McKay P, de Vos AM (1999) J Mol Biol 293, 865-881). This mutant H97Y is also found to be important in the designed library both by database searching (Figure 11) and computational screening (Figure 13A).

K94 is an interesting case and deserves some discussion.

Strictly speaking, K94 does not belong to V_H CDR3 according to the Kabat nomenclature. However, the sequence CAK at the N-terminal of V_H CDR3 are included in building the HMM motif because this sequence puts a strong constraint on the boundary of the sequence motif. Because CAK is the boundary region between framework and V_H CDR3, we consider it here to test the impact of the mutation in this region on the binding affinity. Although R94 is found to be favorable in both the database search and computational screening (Figure 11 and 13A), K94 binds tighter than R94 in experimental screening (Figures 30 and 36). Only K94 was selected when both K94 and R94 were included in the libraries (**Figures 28B, 30 and 36**), although the R94 is still active in binding to VEGF (see **Figures 13A and 14B**). The reason for this might be that R94 in the joint region would change the orientation of the V_H CDR3 in binding to the antigen by interacting with other regions of the antibody, thereby invalidating the original K94 x-ray structure (matured antibody) used for computational screening. It was reported that R94 would reduce the binding affinity of the anti-VEGF antibody by ~ 5-fold during humanization (Presta LG, O'Connor SJ, Wells JA (1997) J Biol Chem 272, 10678-10684). Several approaches can be used to avoid this problem: (1) avoid designing the boundary residues if only CDRs should be designed; (2) combine both parental and preferred residues (e.g. both K and R at 94) in the experimental library. These should be reasonable and straightforward in this case because R and K are the two major residues (~90% for R94 and ~10% for K94) preferred at this position from the database search (see Figure 11); (3) sampling the conformations at this position for R94 computationally by molecular dynamics simulation and see if altered structure or structure ensemble should be used with R94.

To summarize, the three important sites around V_H CDR3 region of the anti-VEGF antibody have been found to have a direct impact on the binding affinity of the antibody for VEGF. Two of the mutations (Y97 and R101 or R100a Kabat) in the three positions (K94, H97 and S101) were found to be important for improved binding with antigen using either the parental or matured antibody structure in the presence and/or absence of antigen, whereas R94 was not predicted correctly because of the potential structural changes induced by the mutation at

the joint region. Y97 is known to be an important mutation for affinity improvement as shown in our own experimental screening. R101 (R100a Kabat) is a novel mutant confirmed by two independent phage display systems and may confer potentially higher affinity than that by Y97.

Most of these mutants including R94, Y97 and R101 are among the dominant variants in the hit variant profile (see Figure 11) (>5%). So a simple sequence search would have found them from the hit variant library. In structure-based screening of the variant library, these mutants are also ranked higher in the selected sequence profile as shown in Figure 13A. From an ensemble sequence scoring point of view, the pooling and reprofiling of the sequences scoring higher than the parental sequence, also ranks the observed variants at 94 (88%R, 12%K), 97 (60%Y, 17%H), and 101 (60%R, 17%T, 13%S) highly. Except for the problem associated with R94, the statistical preference for Y97 and R101 or T101 is apparent in our design. We have demonstrated our library design, using sequence searching and/or structure-based scoring to generate variant profiles. The experimental screening or selection using the two independent novel phage display systems have shown the utility of the inventive methods described here in designing sequences different from the parental sequence in V_H . Some of the mutants found here, such as Y97 and/or R101 or T101, have affinity higher than that of the parental sequence by at least 10-fold (Y97 is reported to account for a 14-fold improvement in affinity while R101 is shown in our experiments to have a higher affinity (see Figure 36). By extraploatoin, a combination of the mutants, such as Y97 and R101, is likely to have a higher affinity than that reported for the matured sequence.

The binding affinities of the affinity matured VH CDR3 were determined using SPR (surface plasma resonance) instrument (BIAcore) with VEGF immobilized on a biosensor chip as shown in Figure 37. The proteins were expressed and purified. The X50 is in ccFv format and contain the reference sequences for VH and VL shown in Figure 22A and 22B. X63 contains H97Y and S101T in VH CDR3 with 6.3-fold improvement in Kd vs 14-fold improvement in Fab format reported in literature (see Table 6 of Chen Y, Wiesmann C, Fuh G, Li B, Christinger

HW, McKay P, de Vos AM, Lowman HB (1999) J. Mol Biol 293, 865-881). X64 contains the S101R mutant in VH CDR3 with 2.5-fold improvement relative the reference; the improvement comes almost exclusively from the on-rate increase. The importance of this novel mutant for on-rate improvement is not reported, although exhaustive mutagenesis at this position has been done. Also, its frequency in database at this position is low. This demostrates the approach taken here is able to discover important mutants for affinity improvement. The X65 contains H97Y and S101R, showing 10-fold improvement using the ccFv format under the same condition, which is stronger in binding affinity than the best mutant combination (H97Y and S101T) for X63 of the affinity-matured VH CDR3 sequence (Chen Y, Wiesmann C, Fuh G, Li B, Christinger HW, McKay P, de Vos AM, Lowman HB (1999) J. Mol Biol 293, 865-881).

Example 2

Generation of anti-VEGF antibody libraries for framework optimization

VEGF is a key angiogenic factor in development and is involved in the growth of solid tumor by stimulating endothelial cells. A murine monoclonal antibody was found to block VEGF-dependent cell proliferation and slow the tumor growth in vivo (Kim KJ, Li B, Winer J, Armanini M, Gillett N, Phillips HS, Ferrara N (1993) Nature 362, 841-844). This murine antibody was humanized (Presta LG, Chen H, O'Connor SJ, Chisholm V, Meng YG, Krummen L, Winkler M, Ferrara N (1997) Cancer Res. 57, 4593-4599; Baca M, Presta LG, O'Connor SJ, Wells JA (1997) J Biol Chem 272, 10678-10684) using random mutagenesis at some key framework positions following grafting of antigen-binding loops. Typically, after rounds of site-directed mutagenesis and selection, humanized antibodies are generated by replacing a human or concensus human framework with non-human amino acids from the parental non-human antibody at certain pre-determined key positions. These humanized antibodies will usually bind to its cognate antigen of its parental antibody with the reduced affinity relative its parental antibody (about 6-fold weaker for

humanized anti-VEGF relative its parental murine antibody, see Baca M, Presta LG, O'Connor SJ, Wells JA (1997) J Biol Chem 272, 10678-10684, and 2-fold weaker for another version of the humanized anti-VEGF, see Presta LG, Chen H, O'Connor SJ, Chisholm V, Meng YG, Krummen L, Winkler M, Ferrara N (1997) Cancer Res. 57, 4593-4599; Baca M, Presta LG, O'Connor SJ, Wells JA (1997) J Biol Chem 272, 10678-10684). This loss of binding affinity would be recovered by using affinity maturation in CDRs (Chen Y, Wiesmann C, Fuh G, Li B, Christinger HW, McKay P, de Vos AM, Lowman HB (1999) J. Mol Biol 293, 865-881).

Using present inventive methods described, we have discovered 2 humanized frameworks that are 4-folder higher in binding affinity (in ccFv format) upon framework optimization than the parental/reference anti-VEGF antibody sequence (see **Figure 22A & B** for the humanized anti-VEGF antibody framework reported in the literature (Presta LG, Chen H, O'Connor SJ, Chisholm V, Meng YG, Krummen L, Winkler M, Ferrara N (1997) Cancer Res. 57, 4593-4599). Because the reported humanized anti-VEGF antibody (**Figures 22 A & B**) is ~ 2 times weaker than its corresponding murine antibody, these two humanized antibodies should have ~2-fold higher binding affinity upon humanization than the corresponding murine antibody.

1. In silico Design of Anti-VEGF Antibody Framework Libraries

Figure 38A upper panel shows the amino acid sequences of the framework fr123 regions of the murine anti-VEGF antibody (therein after referred to as "murine anti-VEGF antibody or A4.6.1"), the humanized antibodies (HU2.0 and HU2.10 sellected from the libraries and amino acids used for humanization at key positions for both VH and VL (see Baca M, Presta LG, O'Connor SJ, Wells JA (1997) J Biol Chem 272, 10678-10684). The framework and CDRs are designated according to the Kabat criteria (Kabat EA, Redi-Miller M, Perry HM, Gottesman KS (1987) Sequences of Proteins of Immunological Interest 4th edit, National Institutes of Health, Bethesda, MD), although other classification can be used also. **Figure 38A** lower panel shows the amino acid sequences of the framework fr123 regions of the murine

anti-VEGF antibody (therein after referred to as "murine anti-VEGF antibody") and the humanized antibody used as the parental and reference framework here (therein after referred to as "humanized anti-VEGF antibody") reported in the literature (see Presta LG, Chen H, O'Connor SJ, Chisholm V, Meng YG, Krummen L, Winkler M, Ferrara N (1997) Cancer Res. 57, 4593-4599). The framework 4 is not designed because it is relatedly constant. But it can be designed if desired using the same approach. Also, separate segment of framework FR1 or FR2 or FR3 and FR4 can be designed individually and pasted together if desired. The combination of CDRs and FRs can be designed simultaneously by designing each segments or combinations of segments used the approach described here. The positions of CDR1 and CDR2 are indicated using arrows but not listed in the figure. The CDRs are the same as in Figure 9B from the murine anti-VEGF. **Figure 38B** shows the variant profiles for the hit library generated using the human VH germline sequences based on the lead sequence of V_H FR123 of the murine anti-VEGF antibody. The variant profile at the bottom shows the amino acid positional diversity. The lower portion of the figure shows the filtered variant profiles obtained by using a cutoff frequency of 5 and 13, repsectively. All positional amino acids occurring 5 or less times or (13 or less) among the members of the hit list are filtered. **Figure 38B**-continuous shows that the reprofiled variant profile for the hit library generated using the human VH germline sequences based on the lead sequence of V_H FR123 of the murine anti-VEGF antibody without cutoff but the variant at each position is ranked based on its structural compatibility with the antibody structure using total energy or van der waals energy. Some reference amino acids are found to be favorable at certain positions based on their total energy or specific packing, although their occurrence frequency is very low (see for instance, 4 positions (F68(F67), L72(L71), S77(S76) and K98(K94) annotated using arrows). F68 and L72, for example are included in library for selection. **Figure 38C** shows the variant profiles for the hit library generated using the Kabat-derived human VH sequences based on the lead sequence of V_H FR123 of the murine anti-VEGF antibody with a filtered variant profile at a cutoff of 19. The profile underscores

the importance of certain amino acids occurring at low frequency but important in scaffolding. The murine VH FR123 sequence is listed as the reference above the dotted line with position annotated using consecutive number. All the variants of amino acids are listed below the dotted line. The dot in the variant represent the same amino acid as in the reference. Figure 38D shows the designer libraries using the filtered variant profile from human VH germline sequences at cutoff 5 (see Figure 38B). The sequence number annotated above the FR123 sequence is based on the kabat nomenclature (kabataa) and its consecutive order including and amino acids in its CDRs. This filtered variant profile can be further screened computationally to reflect the ranking order of the structural compatibility if only the antibody structure is used. Two amino acids, F70(F69) and L72(L71), missing from the filtered variant profile at cutoff 5 were also included because they are among the best preferred amino acids at these positons based on structure-based scoring. The final submitted library for top 100 ranked sequences from structure-based screening also include F70(F69), L72(L71), S77(S76) and K98(K94) (the number in the bracket representing sequence number based on kabat nomenclature), because some amino acids such as R is over predicted in the computation for both L72(L71) and K98(K94) as discussed previously for K94R in the VH CDR3 affinity maturation.

Figure 38D lower panel shows the designer libraries with amino acids used for humanization for VH fr123. As shown in **Figure 38D**, although the human vs non-human sequences differ in many positions across the entire chain for VH, the amino acid libraries used in other approaches are concentrated at a few key positions, whereas the present invention targets various positions across both VH and VL chains with a few mutants at those positions based on the designer libraries for the starting antibody.

According to the present invention, each motif such as frameworks FR1, FR2, FR3 and FR4 depicted in **Figure 8**, each framework motif or its combination such as FR123 of the antibody can be targeted using a modular in silico evolutionary design approach. It has been understood that there are only a limited number of conformations (called canonical structures) for each motif or its

combination. These structural features of an antibody provide an excellent system for testing the evolutionary sequence design by using structured motifs at various regions of an antibody based on the extensive analysis of antibody structures. These structure and sequence conservation are observed across different species. In fact, the scaffolding of antibodies, or the immunoglobin fold, is one of the most abundant structure observed in nature and is highly conserved among various antibodies and related molecules.

The inventors believe that parental anti-VEGF antibody described above can serve as a lead protein in a model system for directed antibody optimization for therapeutic and other applications using the methods of the present invention. The humanized anti-VEGF antibodies (Baca et al., *supra*; Presta et al., *supra*) can serve as a reference or positive control to validate the results obtained by using the inventive methods.

In addition, structural superposition revealed that the structure of the complex formed between VEGF and the parental antibody almost overlaps with that formed between VEGF and the matured antibody. Since the antibody structures, especially framework regions remain substantially the same, structures of both parental and matured antibodies were used in the design of digital libraries of anti-VEGF antibodies using the inventive methods. The inventive method can be also used to design antibody framework using sequence-based approach or structure ensembles that contain the induced structure changes in CDRs.

Using murine anti-VEGF antibody framework as the lead protein and its V_H FR123 as the lead sequence, digital libraries of V_H FR123 were constructed by following the procedure outlined as Route IV in **Figure 1D** and the diagram in **Figure 2**.

As an overview, a hit library was constructed by searching and selecting hit amino acid sequences with remote homology to V_H FR123. Variant profile was built to list all variants at each position based on the hit library and filtered with certain cutoff value to reduce of the size of the resulting hit variant library within computational or experimental limit. Variant profiles were also built in order to facilitate i) the sampling of the sequence space that covers the preferred region in the

fitness landscape; ii) the partitioning and synthesis of degenerate nucleic acid libraries that target the preferred peptide ensemble sequences; iii) the experimental screening of the antibody libraries for the desired function; and iv) the analysis of experimental results with feedback for further design and optimization.

The lead structural templates were obtained from the available X-ray structures of the complexes formed between VEGF and anti-VEGF antibodies. The complex structure of VEGF and parental anti-VEGF antibody is designated as 1BJ1, and that formed between VEGF and matured anti-VEGF antibody 1CZ8. The results from 1CZ8 structural template were similar to those from 1BJ1 in the relative ranking order of the scanned sequences. The modeled structure or structure ensemble or ensemble avaerage can be also used for screening sequences.

1) Lead sequence

The lead sequence for V_H FR123 is taken from the murine anti-VEGF antibody according to Kabat classification (**Figure 38B**).

2) Hit Library and Variant Profile

The HMM built using the single lead sequence, A4.6.1 (**Figure 38A**), was calibrated and used to search human heavy chain germline sequence database and/or human sequence database (including human germlines and humanized sequences) derived from Kabat database (Johnson, G and Wu, TT (2001) Nucleic Acids Research, 29, 205-206). All sequence hits that are above expectation value or E-value are listed and aligned using HAMMER 2.1.1 package. After removing the redundant sequences from the hit list, the remaining hit sequences for the lead HMM form the hit library.

The sequence identities of the hit sequences from the human VH germline ranges from 40 to 68% of the lead sequence, whereas the corresponding sequence identities of the hit sequences from human immunoglobin sequences derived from Kabat database (the database are parsed to fr123 fragment in order to increase the sensitivity of the search and their relative ranking) (other database would be used if the contain the immunoglobin sequences of human origins) ranging from

~30 to 75%. The evolutionary distances between the hits can be analysed by using the program TreeView1.6.5 (http://taxonomy.zoology.gla.ac.uk/rod/rod.html). The phylogenetic tree was analyzed using the neighbor-joining method (Saitou N, Nei M (1987) Mol Biol Evol 4, 406-425) in ClustalW 1.81 (Thompson JD, Higgins DG, Gibson TJ (1994) Nucleic Acids Research 22, 4673-4680).

The AA-PVP tables in **Figure 38B & D** give the number of occurrence of each amino acid residue at each position. The variant profile below the table lists, in the order of decreasing occurrence at each position, all variants found from the database with the lead sequence as the reference sequence. The dot indicates that the same amino acid as in the reference is found at that position. Comparing the difference in the identity of hit sequences between the human VH germlines and Kabat-derived human VH sequences, the difference in the AA-PVP is apparent: whereas all mutants at each position is of human original for the AA-PVP from human germline sequences, the AA-PVP also contains amino acids of non-human origins or of low occurrence frequency that might come from the starting non-human antibody sequence or amino acids that are structurally important to stabilize the scaffold of the target antibodies etc over the course of evolution. For instance, F70 and L72 in Figure 42B are not identified at AA-PVP from the VH3 germline family (see **Figure 42**, only I and R are allowed at these two positions in human VH3 germlines). But on the other hand, F75 and L77 are allowed in the human VH germline sequences with very low frequency of occurrence. These amino acids F70 and L72 occur at relatively higher frequency in AA-PVP from the Kabat-derived human sequences. All the variants of amino acids are listed below the dotted line. The dot in the variant represent the same amino acid as in the reference. Figure 38D shows the designer libraries using the filtered variant profile from human VH germline sequences at cutoff 5 (see Figure 38B). The sequence number annotated above the FR123 sequence is based on the kabat nomenclature (kabataa) and its consecutive order including and amino acids in its CDRs. This filtered variant profile can be further screened computationally to reflect the ranking order of the structural compatibility if only the antibody structure is used. Two amino acids, F70(F69) and L72(L71), missing

from the filtered variant profile at cutoff 5 were also included because they are among the best preferred amino acids at these positons based on structure-based scoring. The final submitted library for top 100 ranked sequences from structure-based screening also include F70(F69), L72(L71), S77(S76) and K98(K94) (the number in the bracket representing sequence number based on kabat nomenclature), because some amino acids such as R is over predicted in the computation for both L72(L71) and K98(K94) as discussed previously for K94R in the VH CDR3 affinity maturation.

Figure 42 also shows that both F and I can be identitied from this position from the panning while only dominant L72 can be identified at this position. In short, using different database of human origin for framework optimization would provide diverse but powerful choices of amino acids for framework optimization including humanization with improved binding affinity and stability. With the increase in our knowledges in developing therapeutic antibodies, more and more antibody sequence data will be accumulated and guide our design using present invention. No prior assumption is needed to assume the key positions and amino acids associated with those positions. Because this information is revealed automatically using present inventive method, it will become better defined with increase in their occurrence in database as more data are accumulated. Variants can be re-profiled or prioritized to include other potentially beneficial mutants using structurally-based criteria (see **Figure 38B-continuous**).

3) Structure-based evaluation of combinatorial sequences of the hit library

Although the variant profile is informative on the preferred amino acid residues at each position and specific mutants in a preferred order, unmodified, it embodies an enormous number of recombinants. The scoring shows that F70 and L72 should be kept in the profile because they are favored in the structure-based scoring, although their frequency of occurrence is lower than the cutoff used for profile derived from database search **(Figure 38B-continuous)**. Thus, the structure-based energy scoring provides another way to reprofile the occurrence of variants at each position for the hit variant library which was originally built based on profiling of evolutionary sequences selected from protein

databases. Some filtering using frequency cutoff can reduce the combinatorial sequences that need to be evaluated by computational screening or targeted directly by experimental libraries. Even with the cutoff applied to the variant profile, there is still a large number of combinatorial sequences that needs to be scored and evaluated in the final sequences for experimental screening (as shown in **Figure 38D lower panel**).

A structure-based scoring is applied to screen the hit library and its combinatorial sequences that form a hit variant library. Side chains of V_H FR123 of the anti-VEGF antibody in 1CZ8 or 1BJ1 were substituted by rotamers of corresponding amino acid variants from the hit variant library at each residue position. The conformations of rotamers were built and optimized by using the program SCWRL® (version 2.1) using backbone-dependent rotamer library (Bower MJ, Cohen FE, Dunbrack RL (1997) JMB 267, 1268-82).

The scoring was done by searching the optimal rotamers and minimizing the energy by 100 steps using the Amber94 force field in CONGEN [Bruccoleri and Karplus (1987) Biopolymers 26:137-168] in the presence and absence of the structure of the antigen VEGF.

Figure 39A depicts the distribution of scoring diagram for VH framework fr123 hit sequences of murine anti-VEGF using the human VH germline sequences in relatively densely populated blue strips in column 1 in x-axis, together with the murine and humanized framework fr123 (see Presta et al. supra) sequence and a widely used human VH germline DP47 in the relatively sparsely populated blue strips in column 0 in x-axis, using 1bj1 (upper panel) and 1cz8 (lower panel) as the template structures in the absence (leftmost column) and presence(middle column) of the VEGF antigen. The scores of sequences in the presence and absence of antigen is correlated (in the rightmost column), indicating the antibody structure for framework optimization is sufficient for most of the framework optimization because they have minimal contact with antigen. The scoring digrams for the combinatorial sequence libraries are not shown here.

Figure 39B depicts the ranking scoring in the left panel based on the difference between sequences in the library and the reference murine VH FR123 sequence and the phylogenetic distances in x-axis

(distance connecting them (see Figure 14C also) for the reference, murine VH FR123, humanized VH FR123 reported (Presta et al., supra 1997 and Chen et al. supra 1999) and the top ranked 200 designer sequences and human VH3 germlines including a widely used VH human germline called DP47. The top 200 ranking sequences from structure-based screening of one variant profile (AA-PVP) of human germlines are clustered with the human VH3 germline family in phylogenentic analysis (red cycle), whereas the lead murine antibody framework is genetically distant in its phylogentic distance from the designed (when only human germline VH sequences at high occurrence frequency are included and the humanized sequence from 1bj1 (see Presta et al., supra), although the phylogenetic distance would change slightly by including amino acids with relatively low occurrence frequency such as F70(F69) and K98(K94) (see Figure 42C and D). The y-axis shows most of the designed framework VH fr123 have good structural compatibility with the structure relative to the murine reference and humanized framework VH fr123, close to DP47. These support the human-like features of the framework optimization for the inventive method described here as defined partly by its database used.

4) Reduction of the variant profile of the hit variant library

The variant profile from the hit variant library as described above was filtered in order to reduce the potential library size while maintaining most of the preferred residues as shown in Figure 38B, obtained from a hit variant library after eliminating amino acids with occurrences lower than the cutoff value and/or by screening sequences based on their compatibility with the structural scaffolding. For example, some important mutants in a variant profile, such as F70 and L72 from the wild type might be included even though they are below the cutoff value used in filtering hit library. They are evaluated using structure-based profiling and persist many rounds of panning under harsh washing conditions in phage display (see **Figures 42**). The top 100 sequences from structure-based scoring were used, together with F70 and L72 from structure-based profiling of original profile.

5) Construction of degenerate nucleic acid library based on hit variant

library II

The hit variant library constructed above was targeted with a degenerate oligonucleotides shown in **Figure 40A**. The degenerate nucleic acid library constructed above was cloned into a phage display system and the phage-displayed antibodies (ccFv) were selected based on their binding to immobilized VEGF coated onto 96-well plates. The final designed humanized sequence of VH anti-VEGF is shown in **Figure 40A**. For some 120 amino acid residues of the VH of anti-VEGF, 34 amino acids were changed as the result of the computational design: 18 of them were fixed (in bold and underlined) and 16 were placed as a result of determination by phage display library screening (labeled by "X") using the ccFv system described. Accordingly, degeneracy of the DNA sequence corresponding to the 16 positions was created in order to generate multiple options of preferred amino acid residues during screening. The theoretical diversity of the library is approximately $2.6x10^5$. The library was installed into a phage display vector pABMD12 in which the VH of anti-VEGF was replaced by the library. As a result, VL and a variety of VH generated from the library would pair to form a functional ccFv of anti-VEGF. The phage display library was then used for further panning against immobilized VEGF protein antigen.

In order to generate a library that can cover such a wide range of scattered distribution of degenerative positions, multiple overlapping degenerative DNA oligos were synthesized with degenerative positions at the sites where the library was designed. The assembly process consisted of two PCR reactions, assembly PCR, and amplification PCR. The assembly oligos were designed with 35-40mers and overlapped by 15-20 bases with melting temperature of about 60°C by average. One additional pair of amplification oligo primers (Amp93 and Amp94) was created for final amplification of the designed products. Accordingly, the assembly PCR includes: equal amount of the assembly oligo primers in a final total concentration of 8 uM, dNTP of 0.8 uM, 1x pfu buffer (Strategene), and 2.5 units of pfu turbo (Strategene). The thermal cycle was performed as follows: 94°C x 45", 58°C x 45", 72°C x 45" for 30 cycles and a final extension of 10 minutes at 72°C. The PCR product mix was diluted 10 folds and used as the template for the amplification

PCR in which all reagents were remained same except for addition of the amplification primers at the final concentration of 1 uM. The thermal cycle was performed as follows: 94°C x 45", 60°C x 45", 72°C x 45" for 30 cycles and a final extension of 20 minutes at 72°C. The final product (the VH library) was purified, digested with HindIII and StyI (Fig. 26), and finally subcloned into vector pABMD12 to replace the original murine VH. The library was used to electrically transform (electroporate?) TG1 competent cells, which were in turn amplified and rescued by helper phage KO7 (Amersham) before production of phages of the library at 30°C overnight according to standard procedure.

6) Panning of phage display library of humanized VH of anti-VEGF

To screen the library constructed described in the above example, purified homodimeric VEGF protein (Calbiochem) was diluted in designated concentration in coating buffer (0.05 M $NaHCO_3$, pH 9.6) and immobilized on Maxisorb wells (Nunc) at 4°C overnight. The coated wells were then blocked in 5% milk at 37°C for 1 hr before phage library diluted in PBS was applied in the wells for incubation at 37°C for 2 hrs. The incubation mix also routinely contained 2% milk to minimize nonspecific binding. At the end of the incubation, the wells were washed and the phages bound were subsequently eluted by 1.4% triethylamine before infecting TG1 cells followed by rescue by KO7 helper phage for amplification. To amplify the phages, infected and rescued TG1 cells were then grown at 30°C overnight in the presence of carbenicilline and kanamycin before phage library was harvested. The phages amplified were used as the input library for the next round of panning. The panning procedure was summarized in **Figure. 41**. Meanwhile, individual clones from 5th panning and on were randomly sampled for phage ELISA, in which specific binding to immobilized VEGF would be confirmed, and demonstrated 100% positives from the 5th to 7th pannings. Finally, isolated clones grown on plates of 2xYT/carbenicilline (100 ug/ml)/kanamycin (70 ug/ml) were sampled for sequencing beginning from the 5th panning (P5) to define the hit positions and hit sequences against the design.

Summary of the sequence analysis of the hits from the above library panning is illustrated in **Figure 42A**, where comparison of

amino acid residues is made at the positions where the library was designed, together with dominant residues of VH of family III of human germlines, and the hits from the library panning. As indicated, among the sixteen positions that were designed for determination by the phage display library screening, a particular amino acid residue at positions 1, 11, 17, 24, 70, 72, 74, 77, 78, 79, 98 in consecutive numbering (Figure 42B) remained or became dominant from P5 (the 5th panning) to the last (the 8th) panning, whereas the rest of the positions demonstrated some fluctuations of the dominant residues. The final selection of residues at nine of the sixteen positions (shaded in Figure 42B) at the end are predominantly consistent with the residues at equivalent positions at family III of human immunoglobin VH, which makes the selected species most likely fall into family III.

Figure 42C shows the phylogenetic analysis of top hit VH sequences from panning of the phage display libraries of the anti-VEGF, together with human germline VH3 families, murine anti-VEGF VH framework FR123 and humanized VH framework fr123 as annotated. As shown in Figure 42C, the human germline VH3 family is clustered together in phylogenetic distance as expected. The selected optimized VH frameworks also cluster together with the humanized VH sequence (see annotation), very close in phylogenetic distance to the human germline VH3 family, while the murine VH framework is very distant from the optimized VH frameworks and human germlines. The phylogenic analysis of the hit sequences against the entire human immunoglubin repertoire of VH suggests that they are indeed most closely related to family III. The phylogenic analysis also demonstrates that the final hit sequences are much closely related to family III of human immunoglobin as compared to the murine-origin sequence of anti-VEGF (Y. Chen et al., 1999). In summary, the result showed that amino acid residues of human origin for majority of the 34 positions were successfully determined.

Furthermore, five positions, namely, positions 6, 72, 77, 79, 98 in consecutive numbering (Figure 42B), did not end up with preferred human residues after the selection, whereas positions 70 and 74 in consecutive numbering (Figure 42B) managed to pick up a minority population that are residues of human origin. Although remaining

minority, these populations consistently survived from the continuous harsh wash and multiple pannings, demonstrating that they indeed possess high affinity toward the antigen. Those positions did not choose a dominant residue of human origin. On the other hand, the existence of a minority population of human-origin residues (position 70 and 74 in consecutive numbering (Figure 42B)) suggests that it is probably feasible to humanize these positions.

This supports the conclusion that the present inventive method in designing optimized frameworks with fully human or human-like sequences of the optimized antibodies, depending on the fine balance between human-like and compatibility with structure template or templates from ensemble structure or structure average. Figure 42B shows the phylogenetic distances of these sequences in another tree view with annotation for a few well characterized sequences D36, D40 and D42 and related sequences. The D36 is as human as or a little better than the humanized sequence reported in its phylogenetic distance.

The full-length sequences of top hits (top hits from the final two pannings, the 7th and 8th pannings) from the anti-VEGF VH library panning are listed in **Figure 42A**, together with the murine anti-VEGF VH (Y. Chen et al., 1999) and dominant sequence of family III of human immunoglobin VH.

7) Selection of humanized VH of anti-VEGF with high affinity

In order to increase stringency of the wash to select high affinity binders, as summarized in **Figure 41**, procedures of applying prolonged wash, increasing the volume of wash, reducing coating VEGF concentration, reducing the input library phages, etc. were implemented. All these measures would tend to facilitate dissociation of those interactions of the relatively lower affinities, and selectively favor survival of those of higher affinities. The clones of surviving phages from this panning were then sampled for sequencing. The full-length anti-VEGF VH sequences of top hits from this panning are listed in **Figure 42A**. Using our inventive methods described, we have discovered 3 (D36, D40 and D42) humanized frameworks with higher binding affinity in ccFv format upon framework optimization (see

Figures 43A & B) than the parental or reference anti-VEGF antibody sequence (see **Figure 22A & B** for the humanized anti-VEGF antibody framework (Presta LG, Chen H, O'Connor SJ, Chisholm V, Meng YG, Krummen L, Winkler M, Ferrara N (1997) Cancer Res. 57, 4593-4599). These improvement comes mainly from a larger increase in the on-rate and small decrease in the off-rate by framework humanization alone. **Figure 43A** shows the sequences of the optimized VH frameworks (FR123) of anti-VEGF antibodies selected from the designer VH optimization libraries using ccFv phage display system (see description in Figures 23-25 above). The VH fr123 of D36, D40 and D42, together with the original murine antibody VH FR123 and humanized sequence (Presta et al supra) with the same CDRs from murine antibody. The dots the lower panel indicate the amino acids are the same as the reference (murine VH framework fr123).

Figure 43B shows the affinity data of 5 antibodies, parental antibody (X50) and the optimized frameworks (D36, D40, D41 and D42) of anti-VEGF antibody selected from designer libraries using BIAcore biosensor (see Figure 43A and notes in Figure 43B for their sequences). The measurement is done by measuring the change of SPR units (y-axis) vs time (x-axis) when a purified antibody binds its antigen (VEGF) immobilized on the CM5 biochip at 25°C. Both the on-rate and off-rate changes were determined from the data fitting using 1:1 Langmuir binding model. 2 humanized frameworks D36 and D40 are ~4-folder higher in binding affinity (in ccFv format) upon framework optimization than the parental/reference anti-VEGF antibody sequence (see **Figure 22A & B** for the humanized anti-VEGF antibody framework reported in the literature (Presta LG, Chen H, O'Connor SJ, Chisholm V, Meng YG, Krummen L, Winkler M, Ferrara N (1997) Cancer Res. 57, 4593-4599), while D42 is sabout the same as the reference antibody. Because the reported humanized anti-VEGF antibody (**Figures 22 A & B**) is ~ 2 times weaker than its corresponding murine antibody, these two humanized antibodies should have ~2-fold higher binding affinity upon humanization than the corresponding murine antibody.

Figure 44 shows the increased stability of the optimized VH frameworks (D36 and D40). The y-axis shows the percentage of the antibody remain active in binding to the immobilized VEGF antigen

using BIAcore at 25C after the purified antibody is incubated at 4, 37 and 42C for 17 hours for the parental X50 and optimized frameworks (D36 and D40). It shows that the optimized frameworks have higher stability than the humanized VH framework reported (Presta et al. supra, 1997).

Figure 45 shows the improved expression of the optimized VH frameworks. The optimized frameworks (D36, D40 and D42) also show the improved expression relative to the parental/wild type antibody (X50) as shown in the yield expression detected by SDS-PAGE/coomassie blue staining.

It should be noted that the antibody libraries designed by using the methods of the present invention can not only be expressed and screened in a bacteriophage system, but also in cells of other organisms, including but not limited to yeast, insect, plant, and mammalian cells. A designed antibody, including the antigen binding fragments and other antibody forms, may be produced by a variety of recombinant DNA or other techniques. For example, the DNA segment(s) encoding the designed antibody may be cloned into an expression vector and transferred into the host cells by well-known methods, which varies depending on the type of the cellular host, including but not limited to calcium chloride transfection, electroporation, lipofection, and viral transfection. The antibody may be purified according to standard procedures of the art, including but not limited to ammonium sulfate precipitation, affinity columns, column chromatography, gel electrophoresis, and the like. Various modifications may occur to those skilled in the art without departing from the spirit and scope of the invention as defined by the appended claims.

The antibodies designed by using the methods of present invention may be used for diagnosing or therapeutic treatment of various diseases, including but not limited to, cancer, autoimmune diseases such as multiple sclerosis, rheumatoid arthritis, systemic lupus erythematosus, Type I diabetes, and myasthenia gravis, graft-versus-host disease, cardiovascular diseases, viral infection such as HIV, hepatitis viruses, and herpes simplex virus, bacterial infection, allergy, Type II diabetes, hematological disorders such as anemia.

The antibodies can also be used as conjugates that are linked with diagnostic or therapeutic moieties, or in combination with chemotherapeutic or biological agents. The antibodies can also be formulated for delivery via a wide variety of routes of administration. For example, the antibodies may be administered or coadministered orally, topically, parenterally, intraperitoneally, intravenously, intraarterially, transdermally, sublingually, intramuscularly, rectally, transbuccally, intranasally, via inhalation, vaginally, intraoccularly, via local delivery (for example by a catheter or a stent), subcutaneously, intraadiposally, intraarticularly, or intrathecally.

The methods of present invention for designing protein libraries in silico can be implemented in various configurations in any computing systems, including but not limited to supercomputers, personal computers, personal digital assistants (PDAs), networked computers, distributed computers on the internet or other microprocessor systems. The methods and systems described herein above is amenable to execution on various types of executable mediums other than a memory device such as a random access memory (RAM). Other types of executable mediums can used, including but not limited to, a computer readable storage medium which can be any memory device, compact disc, zip disk or floppy disk.

The patents, patent applications and publications cited above are hereby incorporated by reference in their entirety.

CLAIMS

What is claimed is:

1. A method for constructing a library of antibody sequences, the method comprising the steps of:

providing an amino acid sequence of the variable region of the heavy chain (V_H) or light chain (V_L) of a lead antibody;

identifying the amino acid sequences in the CDRs of the lead antibody;

selecting one of the CDRs in the V_H or V_L region of the lead antibody;

providing an amino acid sequence that comprises at least 3 consecutive amino acid residues in the selected CDR, the selected amino acid sequence being a lead sequence;

comparing the lead sequence with a plurality of tester protein sequences; and

selecting from the plurality of tester protein sequences at least two peptide segments that have at least 15% sequence identity with the lead sequence, the selected peptide segments forming a hit library.

2. The method of claim 1, wherein the length of the lead sequence is between 5-100 aa.

3. The method of claim 1, wherein the length of the lead sequence is between 6-80 aa.

4. The method of claim 1, wherein the length of the lead sequence is between 8-50 aa.

5. The method of claim 1, wherein the step of identifying the amino sequences in the CDRs is carried out by using Kabat criteria or Chothia criteria.

6. The method of claim 1, wherein the lead sequence comprises an amino acid sequence from a region within the V_H or V_L of the lead antibody selected from the group consisting of CDR1, CDR2, CDR3, FR1-CDR1, CDR1-FR2, FR2-CDR2, CDR2-FR3, FR3-CDR3, CDR3-FR4, FR1-CDR1-FR2, FR2-CDR2-FR3, and FR3-CDR3-FR4.

7. The method of claim 1, wherein the lead sequence comprises at least 6 consecutive amino acid residues in the selected CDR.

8. The method of claim 1, wherein the lead sequence comprises at least 7 consecutive amino acid residues in the selected CDR.

9. The method of claim 1, wherein the lead sequence comprises all of the amino acid residues in the selected CDR.

10. The method of claim 1, wherein the lead sequence further comprises at least one of the amino acid residues immediately adjacent to the selected CDR.

11. The method of claim 1, wherein the lead sequence further comprises at least one of the amino acid residues in the FRs flanking the selected CDR.

12. The method of claim 1, wherein the lead sequence further comprises
one or more CDRs or FRs adjacent the C-terminus or N-terminus of the selected CDR.

13. The method of claim 1, wherein the plurality of tester protein sequences comprises antibody sequences.

14. The method of claim 1, wherein the plurality of tester protein sequences comprises human antibody sequences.

15. The method of claim 1, wherein the plurality of tester protein sequences comprises humanized antibody sequences each having at least 70% human sequence in V_H or V_L.

16. The method of claim 1, wherein the plurality of tester protein sequences comprises human germline antibody sequences.

17. The method of claim 1, wherein the plurality of tester protein sequences is retrieved from a database consisting of genbank of the NIH, Swiss-Prot database, and the Kabat database for CDRs of antibodies.

18. The method of claim 1, wherein the step of comparing the lead sequence with the plurality of tester protein sequences is implemented by an algorithm selected from the group consisting of BLAST, PSI-BLAST, profile HMM, and COBLATH.

19. The method of claim 1, wherein the sequence identity of the selected peptide segments in the hit library with the lead sequence is at least 25%.

20. The method of claim 1, wherein the sequence identity of the selected peptide segments in the hit library with the lead sequence is at least 35%.

21. The method of claim 1, wherein the sequence identity of the selected peptide segments in the hit library with the lead sequence is at least 45%.

22. The method of claim 1, further comprising the step of:

constructing a nucleic acid library comprising DNA segments encoding the amino acid sequences of the hit library.

23. The method of claim 1, further comprising the steps of:

building an amino acid positional variant profile of the hit library;

converting amino acid positional variant profile of the hit library into a nucleic acid positional variant profile by back-translating the amino acid positional variants into their corresponding genetic codons; and

constructing a degenerate nucleic acid library of DNA segments by combinatorially combining the nucleic acid positional variants.

24. The method of claim 23, wherein the genetic codons are the ones that are preferred for expression in bacteria.

25. The method of claim 23, wherein the genetic codons are chosen such that the diversity of the degenerate nucleic acid library of DNA segments is below $1x10^7$.

26. The method of claim 23, wherein the genetic codons are chosen such that the diversity of the degenerate nucleic acid library of DNA segments is below $1x10^6$.

27. The method of claim 23, further comprising the steps of:

introducing the DNA segments in the degenerate nucleic acid library into cells of a host organism;

expressing the DNA segments in the host cells such that recombinant antibodies containing the amino acid sequences of the hit library encoded by the degenerate nucleic acid library are produced in the cells of the host organism; and

selecting the recombinant antibody that binds to a target antigen with affinity higher than 10^6 M^{-1}.

28. The method of claim 27, wherein the affinity of the selected recombinant antibody is higher than 10^8 M^{-1}.

29. The method of claim 27, wherein the affinity of the selected recombinant antibody is higher than 10^9 M^{-1}.

30. The method of claim 27, wherein the host organism is selected from the group consisting of bacteria, yeast, plants, insects, and mammals.

31. The method of claim 27, wherein the recombinant antibodies are selected from the group consisting of fully assembled antibodies, Fab fragments, Fv fragments, and single chain antibodies.

32. The method of claim 27, wherein the recombinant antibodies are displayed on the surface of phage particles.

33. The method of claim 32, wherein the recombinant antibodies displayed on the surface of phage particles are double-chain heterodimers formed between V_H and V_L.

34. The method of claim 33, wherein heterodimerization of V_H and V_L chains is facilitated by a heterodimer formed between two non-antibody polypeptide chains fused to the V_H and V_L chains, respectively.

35. The method of claim 34, wherein the non-antibody polypeptide chains are derived from heterodimeric receptors $GABA_B$ R1 (GR1) and R2 (GR2), respectively.

36. The method of claim 32, wherein the recombinant antibodies displayed on the surface of phage particles are single-chain antibodies containing V_H and V_L linked by a peptide linker.

37. The method of claim 36, wherein display of the single chain antibody on the surface of phage particles is facilitated by a heterodimer formed between a fusion of the single chain antibody with GR1 and a fusion of phage pIII capsid protein with GR2.

38. The method of claim 27, wherein the target antigen is selected from the group consisting of small organic molecules, proteins, peptides, nucleic acids and polycarbohydrates.

39. A computer-implemented method for constructing a library of mutant antibodies based on a lead antibody, the method comprising the steps of:

taking as an input an amino acid sequence that comprises at least 3 consecutive amino acid residues in a CDR region of the lead antibody, the amino acid sequence being a lead sequence;

employing a computer executable logic to compare the lead sequence with a plurality of tester protein sequences;

selecting from the plurality of tester protein sequences at least two peptide segments that have at least 15% sequence identity with lead sequence; and

generating as an output the selected peptide segments which form a hit library.

40. A computer-readable medium, comprising: logic for constructing a library of mutant antibodies based on a lead antibody, the logic comprising

logic which

takes as an input an amino acid sequence that comprises at least 3 consecutive amino acid residues in a CDR of the lead antibody, the amino acid sequence being a lead sequence;

compares the lead sequence with a plurality of tester protein sequences;

selects from the plurality of tester protein sequences at least two peptide segments that have at least 15% sequence identity with lead sequence; and

generates as an output the selected peptide segments which form

a hit library.

41. A method for constructing a library of antibody based on a structure of a lead antibody, the method comprising:

providing an amino acid sequence of the variable region of the heavy chain (V_H) or light chain (V_L) of a lead antibody, the lead antibody

having a known three dimensional structure which is defined as a lead structural template;

identifying the amino acid sequences in the CDRs of the lead antibody;

selecting one of the CDRs in the V_H or V_L region of the lead antibody;

providing an amino acid sequence that comprises at least 3 consecutive amino acid residues in the selected CDR, the selected amino acid sequence being a lead sequence;

comparing the lead sequence with a plurality of tester protein sequences;

selecting from the plurality of tester protein sequences at least two peptide segments that have at least 10% sequence identity with lead sequence, the selected peptide segments forming a hit library;

determining if a member of the hit library is structurally compatible with the lead structural template using a scoring function; and

selecting the members of the hit library that score equal to or better than the lead sequence.

42. The method of claim 41, wherein the length of the lead sequence is between 5-100 aa.

43. The method of claim 41, wherein the length of the lead sequence is between 6-80 aa.

44. The method of claim 41, wherein the length of the lead sequence is between 8-50 aa.

45. The method of claim 41, wherein the step of identifying the amino sequences in the CDRs is carried out by using Kabat criteria or Chothia criteria.

46. The method of claim 41, wherein the lead sequence comprises an amino acid sequence from a region within the V_H or V_L of the lead antibody selected from the group consisting of CDR1, CDR2, CDR3, FR1-CDR1, CDR1-FR2, FR2-CDR2, CDR2-FR3, FR3-CDR3, CDR3-FR4, FR1-CDR1-FR2, FR2-CDR2-FR3, and FR3-CDR3-FR4.

47. The method of claim 41, wherein the lead sequence comprises at least 6 consecutive amino acid residues in the selected CDR.

48. The method of claim 41, wherein the lead sequence comprises at least 7 consecutive amino acid residues in the selected CDR.

49. The method of claim 41, wherein the lead sequence comprises all of the amino acid residues in the selected CDR.

50. The method of claim 41, wherein the lead sequence further comprises at least one of the amino acid residues immediately adjacent to the selected CDR.

51. The method of claim 41, wherein the lead sequence further comprises at least one of the amino acid residues in the FRs flanking the selected CDR.

52. The method of claim 41, wherein the lead sequence further comprises
one or more CDRs or FRs adjacent the C-terminus or N-terminus of the selected CDR.

53. The method of claim 41, wherein the plurality of tester protein sequences comprises antibody sequences.

54. The method of claim 41, wherein the plurality of tester protein sequences comprises human antibody sequences.

55. The method of claim 41, wherein the plurality of tester protein sequences comprises humanized antibody sequences having at least 70% human sequences in V_H or V_L.

56. The method of claim 41, wherein the plurality of tester protein sequences comprises human germline antibody sequences.

57. The method of claim 41, wherein the plurality of tester protein sequences is retrieved from a database consisting of genbank of the NIH, Swiss-Prot database, and the Kabat database for CDRs of antibodies.

58. The method of claim 41, wherein the step of comparing the lead sequence with the plurality of tester protein sequences is implemented by an algorithm selected from the group consisting of BLAST, PSI-BLAST, profile HMM, and COBLATH.

59. The method of claim 41, wherein the sequence identity of the selected peptide segments in the hit library with the lead sequence is at least 25%.

60. The method of claim 41, wherein the sequence identity of the selected peptide segments in the hit library with the lead sequence is at least 35%.

61. The method of claim 41, wherein the sequence identity of the selected peptide segments in the hit library with the lead sequence is at least 45%.

62. The method of claim 41, wherein the scoring function is an energy scoring function selected from the group consisting of electrostatic interactions, van der Waals interactions, electrostatic solvation energy, solvent-accessible surface solvation energy, and conformational entropy.

63. The method of claim 41, wherein the scoring function is a scoring function that incorporates a forcefield selected from the group consisting of the Amber forcefield, Charmm forcefield, the Discover cvff forcefields, the ECEPP forcefields, the GROMOS forcefields, the OPLS forcefields, the MMFF94 forcefield, the Tripos forcefield, the MM3 forcefield, the Dreiding forcefield, and UNRES forcefield.

64. The method of claim 41, wherein the step of selecting the members of the hit library includes selecting the members of the hit library that have a lower or equal total energy than that of the lead sequence calculated based on a formula of

$$\Delta E_{total} = E_{vdw} + E_{bond} + E_{angel} + E_{electrostatics} + E_{solvation.}$$

65. The method of claim 41, wherein the step of selecting the members of the hit library includes selecting the members of the hit library that have a lower binding free energy than that of the lead sequence calculated as the difference between the bound and unbound states using a refined scoring function

$$\Delta G_b = \Delta G_{MM} + \Delta G_{sol} - T\Delta S_{ss}$$

where

$$\Delta G_{MM} = \Delta G_{ele} + \Delta G_{vdw} \quad (1)$$

$$\Delta G_{sol} = \Delta G_{ele\text{-}sol} + \Delta G_{ASA} \quad (2)$$

66. The method of claim 41, wherein the lead structural template is a 3D structure of a fully assembled lead antibody.

67. The method of claim 41, wherein the lead structural template is a 3D structure of V_H or V_L of the lead antibody.

68. The method of claim 41, wherein the lead structural template is a 3D structure of a CDR or FR of the lead antibody, or combination thereof.

69. The method of claim 41, wherein the lead structural template is a structure derived from X-crystallography, nuclear magnetic resonance (NMR) spectroscopy or theoretical structural modeling.

70. The method of claim 41, further comprising the step of:

constructing a nucleic acid library comprising DNA segments encoding the amino acid sequences of the hit library.

71. The method of claim 41, further comprising the steps of:

building an amino acid positional variant profile of the hit library;

converting amino acid positional variant profile of the hit library into a nucleic acid positional variant profile by back-translating the amino acid positional variants into their corresponding genetic codons; and

constructing a degenerate nucleic acid library of DNA segments by combinatorially combining the nucleic acid positional variants.

72. The method of claim 71, wherein the genetic codons are the ones that are preferred for expression in bacteria.

73. The method of claim 71, wherein the genetic codons are chosen such that the diversity of the degenerate nucleic acid library of DNA segments is below $1x10^7$.

74. The method of claim 71, wherein the genetic codons are chosen such that the diversity of the degenerate nucleic acid library of DNA segments is below $1x10^6$.

75. The method of claim 71, further comprising the steps of:

introducing the DNA segments in the degenerate nucleic acid library into cells of a host organism;

expressing the DNA segments in the host cells such that recombinant antibodies containing the amino acid sequences of the hit

library encoded by the degenerate nucleic acid library are produced in the cells of the host organism; and

selecting the recombinant antibody that binds to a target antigen with affinity higher than 10^6 M^{-1}.

76. The method of claim 75, wherein the affinity of the selected recombinant antibody is higher than 10^8 M^{-1}.

77. The method of claim 75, wherein the affinity of the selected recombinant antibody is higher than 10^9 M^{-1}.

78. The method of claim 75, wherein the host organism is selected from the group consisting of bacteria, yeast, plants, insects, and mammals.

79. The method of claim 75, wherein the recombinant antibodies are selected from the group consisting of fully assembled antibodies, Fab fragments, Fv fragments, and single chain antibodies.

80. The method of claim 75, wherein the recombinant antibodies are displayed on the surface of phage particles.

81. The method of claim 80, wherein the recombinant antibodies displayed on the surface of phage particles are double-chain heterodimers formed between V_H and V_L.

82. The method of claim 81, wherein heterodimerization of V_H and V_L chains is facilitated by a heterodimer formed between two non-antibody polypeptide chains fused to the V_H and V_L chains, respectively.

83. The method of claim 82, wherein the non-antibody polypeptide chains are derived from heterodimeric receptors $GABA_B$ R1 (GR1) and R2 (GR2), respectively.

84. The method of claim 80, wherein the recombinant antibodies displayed on the surface of phage particles are single-chain antibodies containing V_H and V_L linked by a peptide linker.

85. The method of claim 84, wherein display of the single chain antibody on the surface of phage particles is facilitated by a heterodimer formed between a fusion of the single chain antibody with GR1 and a fusion of phage pIII capsid protein with GR2.

86. The method of claim 75, wherein the target antigen is selected from the group consisting of small organic molecules, proteins, peptides, nucleic acids and polycarbohydrates.

SEQUENCE LISTING

<110> Luo, Peizhi
Hsieh, Mark
Zhong, Pingyu
Wang, Caili
Cao, Yicheng
Liu, Shengjiang

<120> GENERATION AND SELECTION OF PROTEIN LIBRARY IN SILICO

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<150> US 10/153,176
<151> 2002-05-20

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Glu Val Gln Leu Val Glu Ser Gly Gly Gly Leu Val Gln Pro Gly Gly
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Ser Leu Arg Leu Ser Cys Ala Ala Ser Gly Tyr Thr Phe Thr Asn Tyr

20 25 30

Gly Met Asn Trp Val Arg Gln Ala Pro Gly Lys Gly Leu Glu Trp Val
35 40 45

Gly Trp Ile Asn Thr Tyr Thr Gly Glu Pro Thr Tyr Ala Ala Asp Phe
50 55 60

Lys Arg Arg Phe Thr Phe Ser Leu Asp Thr Ser Lys Ser Thr Ala Tyr
65 70 75 80

Leu Gln Met Asn Ser Leu Arg Ala Glu Asp Thr Ala Val Tyr Tyr Cys
85 90 95

Ala Lys Tyr Pro His Tyr Tyr Gly Ser Ser His Trp Tyr Phe Asp Val
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Asp Ile Gln Met Thr Gln Ser Pro Ser Ser Leu Ser Ala Ser Val Gly
1 5 10 15

Asp Arg Val Thr Ile Thr Cys Ser Ala Ser Gln Asp Ile Ser Asn Tyr
20 25 30

Leu Asn Trp Tyr Gln Gln Lys Pro Gly Lys Ala Pro Lys Val Leu Ile
35 40 45

Tyr Phe Thr Ser Ser Leu His Ser Gly Val Pro Ser Arg Phe Ser Gly
50 55 60

Ser Gly Ser Gly Thr Asp Phe Thr Leu Thr Ile Ser Ser Leu Gln Pro
65 70 75 80

Glu Asp Phe Ala Thr Tyr Tyr Cys Gln Gln Tyr Ser Thr Val Pro Trp
85 90 95

Thr Phe Gly Gln Gly Thr Lys Val Glu Ile Lys Arg
100 105

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Glu Val Gln Leu Val Glu Ser Gly Gly Gly Leu Val Gln Pro Gly Gly
1 5 10 15

Ser Leu Arg Leu Ser Cys Ala Ala Ser Gly Tyr Asp Phe Thr His Tyr
20 25 30

Gly Met Asn Trp Val Arg Gln Ala Pro Gly Lys Gly Leu Glu Trp Val
35 40 45

Gly Trp Ile Asn Thr Tyr Thr Gly Glu Pro Thr Tyr Ala Ala Asp Phe
50 55 60

Lys Arg Arg Phe Thr Phe Ser Leu Asp Thr Ser Lys Ser Thr Ala Tyr
65 70 75 80

Leu Gln Met Asn Ser Leu Arg Ala Glu Asp Thr Ala Val Tyr Tyr Cys
85 90 95

Ala Lys Tyr Pro Tyr Tyr Tyr Gly Thr Ser His Trp Tyr Phe Asp Val
100 105 110

Trp Gly Gln Gly Thr Leu Val Thr Val Ser Ser
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1 5 10 15

Asp Arg Val Thr Ile Thr Cys Ser Ala Ser Gln Asp Ile Ser Asn Tyr
20 25 30

Leu Asn Trp Tyr Gln Gln Lys Pro Gly Lys Ala Pro Lys Val Leu Ile
35 40 45

Tyr Phe Thr Ser Ser Leu His Ser Gly Val Pro Ser Arg Phe Ser Gly
50 55 60

Ser Gly Ser Gly Thr Asp Phe Thr Leu Thr Ile Ser Ser Leu Gln Pro
65 70 75 80

Glu Asp Phe Ala Thr Tyr Tyr Cys Gln Gln Tyr Ser Thr Val Pro Trp
85 90 95

Thr Phe Gly Gln Gly Thr Lys Val Glu Ile Lys Arg
100 105

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Val Trp Gly

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Tyr Phe Thr Ser Ser Leu His Ser Gly Val Pro Ser Arg Phe Ser Gly
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Ser Gly Ser Gly Thr Asp Phe Thr Leu Thr Ile Ser Ser Leu Gln Pro
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Arg

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<400> 59

Cys Ala Arg Asp Pro Tyr Tyr Tyr Gly Arg Ser Tyr Trp Tyr Phe Asp
1 5 10 15

Leu Trp Gly

<210> 60

<211> 57

<212> DNA

<213> Artificial Sequence

<220>

<223> DNA encoding variant profile lead sequence

<400> 60

tgcgcgcgta acccgtacta caacggtcgt agctactggt acttcgatct gtggggt 57

<210> 61

<211> 57

<212> DNA

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<400> 61
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<210> 62

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> Matured VH CDR3

<400> 62

Thr Ala Val Tyr Tyr Cys Ala Lys Tyr Pro Tyr Tyr Tyr Gly Thr Ser
1 5 10 15

His Trp Tyr Phe Asp Val Trp Gly Gln Gly
20 25

<210> 63

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> pVGD9-1-1B8

<400> 63

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Cys Gly Arg Thr
1 5 10 15

Cys Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 64

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> pVGD9-1-1D7

<400> 64

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Cys Gly Arg Thr
1 5 10 15

Cys Trp Tyr Phe Asp Leu Trp Gly Gln Gly
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<210> 65

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> pVGD9-1-2A6

<400> 65

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Cys Gly Arg Thr
1 5 10 15

Cys Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 66

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> pVGD9-1-3D10

<400> 66

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Cys Gly Arg Thr

1 5 10 15

Cys Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 67

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> pVGD9-1-2A12

<400> 67

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Cys Gly Arg Thr
1 5 10 15

Cys Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 68

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> pVGD9-1-1F8

<400> 68

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Cys Gly Arg Ser
1 5 10 15

Cys Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 69

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> pVGD9-1-3B5

<400> 69

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Cys Gly Arg Ser
1 5 10 15

Cys Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 70

<211> 26

<212> PRT

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<220>

<223> pVGD9-1-1C4

<400> 70

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Cys Ser Arg Ser
1 5 10 15

Cys Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 71

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> pVGD9-1-3F1

<400> 71

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Cys Ser Arg Ser
1 5 10 15

Cys Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 72

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> pVGD9-1-2F7

<400> 72

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Cys Gly Arg Ser
1 5 10 15

Asn Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 73

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> pVGD9-1-3B11

<400> 73

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Gly Ser Arg Thr
1 5 10 15

Cys Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 74

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> pVGD9-1-3B10

<400> 74

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Ser Gly Arg Ser
1 5 10 15

Tyr Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 75

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> pVGD9-1-1A11

<400> 75

Thr Ala Val Tyr Tyr Cys Ala Arg Asp Pro Tyr Tyr Asp Ser Arg Ser
1 5 10 15

Ser Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 76

<211> 26

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<213> Artificial Sequence

<220>

<223> pVGD9-1-2A1

<400> 76

Thr Ala Val Tyr Tyr Cys Ala Arg Asp Pro Tyr Tyr Asp Ser Arg Ser
1 5 10 15

Tyr Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 77

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> pVGD9-1-3A6

<400> 77

Thr Ala Val Tyr Tyr Cys Ala Arg Asp Pro Tyr Tyr Gly Gly Arg Ser
1 5 10 15

Tyr Trp Tyr Phe Asp Leu Trp Gly Gln Gly

20 25

<210> 78

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> pVGD9-1-2G8

<400> 78

Thr Ala Val Tyr Tyr Cys Ala Arg Asp Pro Tyr Tyr Gly Gly Arg Thr
1 5 10 15

Ser Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 79

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> pVGD9-1-3B8

<400> 79

Thr Ala Val Tyr Tyr Cys Ala Arg Asp Pro Tyr Tyr Cys Gly Arg Ser
1 5 10 15

Asn Trp Tyr Phe Asp Leu Trp Gly Gln Gly
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<210> 80

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> pVGD9-1-1B7

<400> 80

Thr Ala Val Ser Tyr Cys Ala Arg Asp Pro Tyr Tyr Tyr Gly Arg Ser
1 5 10 15

Asn Trp Tyr Phe Asp Leu Trp Gly Gln Gly
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<210> 81

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> pVGD9-1-3H3

<400> 81

Thr Ala Val Tyr Tyr Cys Ala Arg Asp Pro Tyr Tyr Tyr Gly Arg Thr
1 5 10 15

Ser Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 82

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> pVGD9-1-1D2

<400> 82

Thr Ala Val Tyr Tyr Cys Ala Arg Ala Pro Tyr Tyr Cys Ser Arg Thr
1 5 10 15

Cys Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 83

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> pVGD9-1-2A2

<400> 83

Thr Ala Val Tyr Tyr Cys Ala Arg Ala Pro Tyr Tyr Cys Ser Arg Thr
1 5 10 15

Cys Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 84

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> pVGD9-1-2A11

<400> 84

Thr Ala Val Tyr Tyr Cys Ala Arg Ala Pro Tyr Tyr Cys Gly Arg Ser
1 5 10 15

Cys Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 85

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> pVGD9-1-3A12

<400> 85

Thr Ala Val Tyr Tyr Cys Ala Arg Ala Pro Tyr Tyr Cys Gly Arg Ser
1 5 10 15

Cys Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 86

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> pVGD9-1-1C2

<400> 86

Thr Ala Val Tyr Tyr Tyr Ala Arg Ala Pro Tyr Tyr Cys Gly Arg Ser
1 5 10 15

Tyr Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 87

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> pVGD9-1-1B12

<400> 87

Thr Ala Val Tyr Tyr Tyr Ala Arg Ala Pro Tyr Tyr Cys Gly Arg Ser
1 5 10 15

Tyr Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 88

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> pVGD9-1-2A3

<400> 88

Thr Ala Val Tyr Tyr Cys Ala Arg Ser Pro Tyr Tyr Cys Gly Arg Ser
1 5 10 15

Cys Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 89

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> pVGD9-1-2G5

<400> 89

Thr Ala Val Tyr Tyr Cys Ala Arg Ser Pro Tyr Tyr Cys Gly Arg Ser
1 5 10 15

Cys Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 90

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> pVGD9-1-2A8

<400> 90

Thr Ala Val Tyr Tyr Cys Ala Arg Ser Pro Tyr Tyr Cys Ser Arg Thr
1 5 10 15

Cys Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 91

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> pVGD9-1-3A7

<400> 91

Thr Ala Val Tyr Tyr Cys Ala Arg Ser Pro Tyr Tyr Gly Gly Arg Ser
1 5 10 15

Tyr Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 92

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> pVGD9-1-2B7

<400> 92

Thr Ala Val Tyr Tyr Cys Ala Arg Asn Pro Tyr Tyr Cys Gly Arg Thr
1 5 10 15

Cys Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 93

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> pVGD9-1-2G6

<400> 93

Thr Ala Val Tyr Tyr Cys Ala Arg Asn Pro Tyr Tyr Cys Gly Arg Thr
1 5 10 15

Cys Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 94

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> pVGD9-1-3A11

<400> 94

Thr Ala Val Tyr Tyr Cys Ala Arg Thr Pro Tyr Tyr Gly Gly Arg Thr
1 5 10 15

Asn Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 95

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> pVGD9-1-3H2

<400> 95

Thr Ala Val Tyr Tyr Cys Ala Arg Thr Pro Tyr Tyr Tyr Gly Arg Thr
1 5 10 15

Ser Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 96

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> Library9

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<222> (9)..(9)

<223> X = Y, D, A, S, N, or T

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<223> X = Y, C, G, S, or D.

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<222> (14)..(14)

<223> X = G or S.

<220>

<221> MISC_FEATURE

<222> (16)..(16)

<223> X = S or T.

<220>

<221> MISC_FEATURE

<222> (17)..(17)

<223> X = H, C, N, Y, or S.

<400> 96

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Thr Ala Val Tyr Tyr Cys Ala Arg Xaa Pro Tyr Tyr Xaa Xaa Arg Xaa
1               5                  10                  15

Xaa Trp Tyr Phe Asp Leu Trp Gly Gln Gly
            20                  25
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<210> 97

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> PVG9-3-1G6

<400> 97

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Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Tyr Ser Arg Ser
1               5                  10                  15

Ser Trp Tyr Phe Asp Leu Trp Gly Gln Gly
            20                  25
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<210> 98

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> PVG9-3-2A1

<400> 98

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Tyr Ser Arg Ser
1 5 10 15

Ser Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 99

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> PVG9-3-1D3

<400> 99

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Tyr Ser Arg Ser
1 5 10 15

Ser Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 100

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> PVG9-3-1D5

<400> 100

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Tyr Ser Arg Ser
1 5 10 15

Ser Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 101

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> PVG9-3-1G4

<400> 101

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Tyr Ser Arg Ser
1 5 10 15

Ser Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 102

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> PVG9-3-2E1

<400> 102

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Tyr Ser Arg Ser
1 5 10 15

Ser Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 103

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> PVG9-3-1E5

<400> 103

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Tyr Ser Arg Ser

1 5 10 15

Ser Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 104

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> PVG9-3-1A5

<400> 104

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Tyr Ser Arg Ser
1 5 10 15

Ser Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 105

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> PVG9-3-1A10

<400> 105

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Tyr Ser Arg Ser
1 5 10 15

Ser Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 106

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> PVG9-3-2E6

<400> 106

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Tyr Ser Arg Ser
1 5 10 15

Asn Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 107

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> PVG9-3-1B10

<400> 107

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Tyr Ser Arg Ser
1 5 10 15

Asn Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 108

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> PVG9-3-1D6

<400> 108

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Tyr Ser Arg Ser
1 5 10 15

Asn Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 109

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> PVG9-3-2F4

<400> 109

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Tyr Gly Arg Ser
1 5 10 15

Asn Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 110

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> PVG9-3-1E3

<400> 110

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Tyr Gly Arg Ser
1 5 10 15

Asn Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 111

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> PVG9-3-2A2

<400> 111

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Tyr Gly Arg Ser
1 5 10 15

Asn Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 112

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> PVG9-3-1G7

<400> 112

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Tyr Gly Arg Ser
1 5 10 15

Ser Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 113

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> PVG9-3-1E2

<400> 113

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Tyr Gly Arg Ser
1 5 10 15

Ser Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 114

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> PVG9-3-2B12

<400> 114

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Tyr Gly Arg Thr
1 5 10 15

Ser Trp Tyr Phe Asp Leu Trp Gly Gln Gly

20 25

<210> 115

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> PVG9-3-2D6

<400> 115

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Tyr Gly Arg Thr
1 5 10 15

Ser Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 116

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> PVG9-3-2F2

<400> 116

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Ser Gly Arg Ser
1 5 10 15

Ser Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 117

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> PVG9-3-1D1

<400> 117

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Ser Gly Arg Ser
1 5 10 15

Ser Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 118

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> PVG9-3-1A11

<400> 118

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Ser Gly Arg Ser
1 5 10 15

Ser Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 119

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> PVG9-3-1E7

<400> 119

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Tyr Ser Arg Thr
1 5 10 15

Ser Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 120

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> PVG9-3-1C10

<400> 120

Thr Ala Val Tyr Tyr Cys Ala Arg Ala Pro Tyr Tyr Cys Gly Arg Ser
1 5 10 15

Cys Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 121

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> PVG9-3-1F9

<400> 121

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Cys Gly Arg Ser
1 5 10 15

Cys Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 122

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> PVG9-3-1F8

<400> 122

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Asp Gly Arg Ser
1 5 10 15

Asn Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 123

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> PVG9-3-2G5

<400> 123

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Cys Gly Arg Ser
1 5 10 15

Asn Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 124

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> PVG9-3-2B11

<400> 124

Thr Ala Val Tyr Tyr Cys Ala Arg Asp Pro Tyr Tyr Cys Ser Arg Ser
1 5 10 15

Cys Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 125

<211> 26

<212> PRT

<213> Artificial Sequence

<220>

<223> PVG9-3-2G9

<400> 125

Thr Ala Val Tyr Tyr Cys Ala Arg Tyr Pro Tyr Tyr Ser Gly Arg Ser
1 5 10 15

Tyr Trp Tyr Phe Asp Leu Trp Gly Gln Gly
20 25

<210> 126

<211> 123

<212> PRT

<213> Artificial Sequence

<220>

<223> VH of antibody X64

<400> 126

Glu Val Gln Leu Val Glu Ser Gly Gly Gly Leu Val Gln Pro Gly Gly
1 5 10 15

Ser Leu Arg Leu Ser Cys Ala Ala Ser Gly Tyr Thr Phe Thr Asn Tyr
20 25 30

Gly Met Asn Trp Val Arg Gln Ala Pro Gly Lys Gly Leu Glu Trp Val
35 40 45

Gly Trp Ile Asn Thr Tyr Thr Gly Glu Pro Thr Tyr Ala Ala Asp Phe
50 55 60

Lys Arg Arg Phe Thr Phe Ser Leu Asp Thr Ser Lys Ser Thr Ala Tyr
65 70 75 80

Leu Gln Met Asn Ser Leu Arg Ala Glu Asp Thr Ala Val Tyr Tyr Cys
85 90 95

Ala Lys Tyr Pro His Tyr Tyr Gly Arg Ser His Trp Tyr Phe Asp Val
100 105 110

Trp Gly Gln Gly Thr Leu Val Thr Val Ser Ser
115 120

<210> 127

<211> 108

<212> PRT

<213> Artificial Sequence

<220>

<223> VL of antibody X64, X65, D36, D40 and D42

<400> 127

Asp Ile Gln Met Thr Gln Ser Pro Ser Ser Leu Ser Ala Ser Val Gly
1 5 10 15

Asp Arg Val Thr Ile Thr Cys Ser Ala Ser Gln Asp Ile Ser Asn Tyr
20 25 30

Leu Asn Trp Tyr Gln Gln Lys Pro Gly Lys Ala Pro Lys Val Leu Ile
35 40 45

Tyr Phe Thr Ser Ser Leu His Ser Gly Val Pro Ser Arg Phe Ser Gly
50 55 60

Ser Gly Ser Gly Thr Asp Phe Thr Leu Thr Ile Ser Ser Leu Gln Pro
65 70 75 80

Glu Asp Phe Ala Thr Tyr Tyr Cys Gln Gln Tyr Ser Thr Val Pro Trp
85 90 95

Thr Phe Gly Gln Gly Thr Lys Val Glu Ile Lys Arg
100 105

<210> 128

<211> 123

<212> PRT

<213> Artificial Sequence

<220>

<223> VH of antibody X65

<400> 128

Glu Val Gln Leu Val Glu Ser Gly Gly Gly Leu Val Gln Pro Gly Gly
1 5 10 15

Ser Leu Arg Leu Ser Cys Ala Ala Ser Gly Tyr Thr Phe Thr Asn Tyr
20 25 30

Gly Met Asn Trp Val Arg Gln Ala Pro Gly Lys Gly Leu Glu Trp Val
35 40 45

Gly Trp Ile Asn Thr Tyr Thr Gly Glu Pro Thr Tyr Ala Ala Asp Phe
50 55 60

Lys Arg Arg Phe Thr Phe Ser Leu Asp Thr Ser Lys Ser Thr Ala Tyr
65 70 75 80

Leu Gln Met Asn Ser Leu Arg Ala Glu Asp Thr Ala Val Tyr Tyr Cys
85 90 95

Ala Lys Tyr Pro Tyr Tyr Tyr Gly Arg Ser His Trp Tyr Phe Asp Val
100 105 110

Trp Gly Gln Gly Thr Leu Val Thr Val Ser Ser
115 120

<210> 129

<211> 123

<212> PRT

<213> Artificial Sequence

<220>

<223> VH of antibody D36

<400> 129

Glu Val Gln Leu Val Gln Ser Gly Gly Gly Val Val Gln Pro Gly Gly
1 5 10 15

Ser Leu Arg Leu Ser Cys Ala Ala Ser Gly Tyr Thr Phe Thr Asn Tyr
20 25 30

Gly Met Asn Trp Val Arg Gln Ala Pro Gly Lys Gly Leu Glu Trp Val
35 40 45

Gly Trp Ile Asn Thr Tyr Thr Gly Glu Pro Thr Tyr Ala Ala Asp Phe
50 55 60

Lys Arg Arg Phe Thr Phe Ser Leu Asp Thr Ser Lys Ser Thr Ala Tyr
65 70 75 80

Leu Gln Met Asn Ser Leu Arg Ala Glu Asp Thr Ala Val Tyr Tyr Cys
85 90 95

Ala Lys Tyr Pro His Tyr Tyr Gly Ser Ser His Trp Tyr Phe Asp Val
100 105 110

Trp Gly Gln Gly Thr Leu Val Thr Val Ser Ser
115 120

<210> 130

<211> 123

<212> PRT

<213> Artificial Sequence

<220>

<223> VH of antibody D40

<400> 130

Glu Val Gln Leu Val Gln Ser Gly Gly Gly Val Val Gln Pro Gly Gly
1 5 10 15

Ser Leu Arg Leu Ser Cys Ala Ala Ser Gly Tyr Thr Phe Thr Asn Tyr
20 25 30

Gly Met Asn Trp Ile Arg Gln Ala Pro Gly Lys Gly Leu Glu Trp Val
35 40 45

Gly Trp Ile Asn Thr Tyr Thr Gly Glu Pro Thr Tyr Ala Ala Asp Phe
50 55 60

Lys Arg Arg Val Thr Phe Ser Leu Asp Thr Ser Lys Ser Thr Ala Tyr
65 70 75 80

Leu Gln Leu Asn Ser Leu Arg Ala Glu Asp Thr Ala Val Tyr Tyr Cys
85 90 95

Ala Lys Tyr Pro His Tyr Tyr Gly Ser Ser His Trp Tyr Phe Asp Val
100 105 110

Trp Gly Gln Gly Thr Leu Val Thr Val Ser Ser
115 120

<210> 131

<211> 123

<212> PRT

<213> Artificial Sequence

<220>

<223> VH of antibody D42

<400> 131

Glu Val Gln Leu Val Gln Ser Gly Gly Gly Val Val Gln Pro Gly Gly
1 5 10 15

Thr Leu Arg Leu Thr Cys Ala Ala Ser Gly Tyr Thr Phe Thr Asn Tyr
20 25 30

Gly Met Asn Trp Val Arg Gln Ala Pro Gly Lys Gly Leu Glu Trp Val
35 40 45

Gly Trp Ile Asn Thr Tyr Thr Gly Glu Pro Thr Tyr Ala Ala Asp Phe
50 55 60

Lys Arg Arg Val Thr Phe Ser Leu Asp Thr Ser Lys Ser Thr Ala Tyr

65 70 75 80

Leu Gln Met Asn Ser Leu Arg Ala Glu Asp Thr Ala Val Tyr Tyr Cys
85 90 95

Ala Lys Tyr Pro His Tyr Tyr Gly Ser Ser His Trp Tyr Phe Asp Val
100 105 110

Trp Gly Gln Gly Thr Leu Val Thr Val Ser Ser
115 120