

T CELL RECEPTORS RECOGNIZING MUTATED P53

CROSS-REFERENCE TO RELATED APPLICATION

[0001] This patent application claims the benefit of U.S. Provisional Patent Application No. 62/565,383, filed September 29, 2017, which is incorporated by reference in its entirety herein.

STATEMENT REGARDING FEDERALLY SPONSORED RESEARCH OR
DEVELOPMENT

[0002] This invention was made with Government support under project number BC010985 by the National Institutes of Health, National Cancer Institute. The Government has certain rights in the invention.

INCORPORATION-BY-REFERENCE OF MATERIAL SUBMITTED
ELECTRONICALLY

[0003] Incorporated by reference in its entirety herein is a computer-readable nucleotide/amino acid sequence listing submitted concurrently herewith and identified as follows: One 687,555 Byte ASCII (Text) file named "739980_ST25.txt," dated September 7, 2018.

BACKGROUND OF THE INVENTION

[0004] Some cancers may have very limited treatment options, particularly when the cancer becomes metastatic and unresectable. Despite advances in treatments such as, for example, surgery, chemotherapy, and radiation therapy, the prognosis for many cancers, such as, for example, pancreatic, colorectal, lung, endometrial, ovarian, and prostate cancers, may be poor. Accordingly, there exists an unmet need for additional treatments for cancer.

BRIEF SUMMARY OF THE INVENTION

[0005] An embodiment of the invention provides an isolated or purified T cell receptor (TCR) having antigenic specificity for mutated human p53.

[0006] Further embodiments of the invention provide related polypeptides and proteins, as well as related nucleic acids, recombinant expression vectors, host cells, populations of cells, and pharmaceutical compositions relating to the TCRs of the invention.

[0007] Still further embodiments of the invention provide methods of detecting the presence of cancer in a mammal and methods of treating or preventing cancer in a mammal.

BRIEF DESCRIPTION OF THE SEVERAL VIEWS OF THE DRAWINGS

[0008] Figure 1 is a graph showing the concentration of IFN- γ (pg/mL) measured following co-culture of autologous PBL transduced with one of the indicated TCRs from patient 4127 following co-culture with autologous dendritic cells (DCs) pulsed with either WT p53-G245 peptide (wt) (unshaded bars) or mutated p53-G245S peptide (mut) (shaded bars) at the indicated decreasing peptide concentrations (μ g/mL).

[0009] Figure 2 is a graph showing the percentage of 4-1BB positive cells (% of CD4+ mTCR β +) detected following co-culture of autologous PBL transduced with one of the indicated TCRs from patient 4127 with autologous DCs pulsed with either WT p53-G245 peptide (wt) (unshaded bars) or mutated p53-G245S peptide (mut) (shaded bars) at the indicated decreasing peptide concentrations (μ g/mL).

[0010] Figure 3 is a graph showing the percentage of 4-1BB positive cells (% of CD4+ mTCR β +) detected following co-culture of autologous PBL transduced with one of the indicated TCRs from patient 4127 with autologous DCs pulsed with one of the peptides shown in Table A.

[0011] Figure 4 is a graph showing the concentration of IFN- γ (pg/mL) measured following co-culture of TIL from patient 4127 or T cells transduced with one of the indicated TCRs with COS7 cells which were transduced with one of the indicated HLA alleles and pulsed with DMSO or one of the indicated peptides.

[0012] Figure 5 is a graph showing the percentage of 4-1BB positive cells (% of CD4+ mTCR β +) detected following co-culture of TIL from patient 4127 or T cells which were transduced with one of the indicated TCRs with COS7 cells which were transduced with one of the indicated HLA alleles and pulsed with DMSO or one of the indicated peptides.

[0013] Figure 6 is a graph showing the number of spots per 2×10^4 effector cells measured following co-culture of TIL from patient 4127 with allogeneic (DRB3*01:01:01 or DRB3*02:02:01) APCs which were (1) electroporated with TMG composed of irrelevant (IRR; left hatched open bars), WT p53 (p53-WT; left hatched gray bars) or mutated p53 (p53-MUT; gray bars) sequence or (2) pulsed with peptide vehicle (DMSO; right hatched open bars) or purified (>95% by HPLC) 25-amino acid peptides composed of WT p53-G245

sequence (LP-p53-wt-G245; right hatched gray bars) or mutated p53-G245S (LP-p53-mut-G245S; black bars) sequence.

[0014] Figure 7 is a graph showing the number of spots per 2×10^4 effector cells measured following co-culture of cells expressing the 4127-TCR1 with allogeneic (DRB3*01:01:01 or DRB3*02:02:01) APCs which were (1) electroporated with TMG composed of irrelevant (IRR; left hatched open bars), WT p53 (p53-WT; left hatched gray bars) or mutated p53 (p53-MUT; gray bars) sequence or (2) pulsed with peptide vehicle (DMSO; right hatched open bars) or purified (>95% by HPLC) 25-amino acid peptides composed of WT p53-G245 sequence (LP-p53-wt-G245; right hatched gray bars) or mutated p53-G245S (LP-p53-mut-G245S; black bars) sequence.

[0015] Figure 8A is a graph showing IFN- γ secretion (spots/ 2×10^4 cells) measured following co-culture of TIL from the infusion bag used to treat patient 4196 with Cos7 cells transfected with plasmids for TMG1 and the indicated HLA allele.

[0016] Figure 8B is a graph showing IFN- γ secretion (spots/ 2×10^4 cells) measured following co-culture of TIL from the infusion bag used to treat patient 4196 with autologous DCs pulsed with candidate minimal epitope (HMTEVVRHC (SEQ ID NO: 530) or (SQHMTEVVRH (SEQ ID NO: 531)). T cells cultured alone served as a control.

[0017] Figures 9A-9D are graphs showing the percentages of mTCR β +CD8+41BB+ cells detected following co-culture of DCs pulsed with minimal WT (circles) or mutated (mut) (squares) epitope at the indicated concentrations (μ g/mL) with cells transduced with one of the following TCRs: AV12/BV6-1 (A), AV2/BV6-1 (B), AV6/BV11 (C), or AV38/BV10-3 (D). Co-culture of pulsed DCs with untransduced (UT) cells (triangles) served as a control. Results shown are representative of two experiments.

[0018] Figures 10A-10B are graphs showing IFN- γ secretion (pg/mL) (A) and percentage of 4-1BB positive cells (% mTCR β +) (B) measured following co-culture of T2 cells pulsed with the minimal mutated epitope at the indicated concentrations with cells transduced with one of the following TCRs: AV12/BV6-1 (closed triangles), AV6/BV11 (circles), or AV38/BV10-3 (squares). Co-culture of pulsed T2 cells with untransduced (UT) cells (open triangles) served as a control.

[0019] Figure 11 is a graph showing the number of IFN- γ -positive spots per 2×10^4 effector cells detected following co-culture of TCR-transduced cells with the indicated target cell lines. Effector cells were transduced with AV12/BV6-1 (black bars), AV6/BV11 (striped bars), or AV38/BV10-3 (grey bars). HLA-A*02:01-transduced target cells are indicated by

LS123-A2, CCRF-CEM-A2 and AU-565-A2. Peptide-pulsed target cells are indicated by (+). Target cells not pulsed with peptide are indicated by (-). Untransduced effector cells (unshaded bars) co-cultured with target cell lines were used as a control.

[0020] Figure 12 is a graph showing the number of IFN- γ -positive spots per 2×10^4 effector cells measured following co-culture of Patient 4238 TIL fragments F1-F24 (n=24) with autologous APCs electroporated with TMG composed of irrelevant (IRR; open bars) or mutated p53 (p53-MUT; black bars) sequence.

[0021] Figure 13 is a graph showing the number of IFN- γ -positive spots per 2×10^4 effector cells measured following co-culture of Patient 4238 TIL fragments F1-F24 (n=24) with autologous APCs pulsed with peptide vehicle (DMSO; open bars) or purified (>95% by HPLC) 25-amino acid peptide composed of mutated p53-R248Q (LP-p53-R248Q-MUT; black bars) sequence.

[0022] Figure 14 is a graph showing the percentage of CD8+4-1BB+ cells detected following co-culture of Patient 4238 TIL fragments F1-F24 (n=24) with autologous APCs electroporated with TMG composed of irrelevant (IRR; open bars) or mutated p53 (p53-MUT; black bars) sequence.

[0023] Figure 15 is a graph showing the percentage of CD8+4-1BB+ cells detected following co-culture of Patient 4238 TIL fragments F1-F24 (n=24) with autologous APCs pulsed with peptide vehicle (DMSO; open bars) or purified (>95% by HPLC) 25-amino acid peptide composed of mutated p53-R248Q (LP-p53-R248Q-MUT; black bars) sequence.

[0024] Figure 16 is a graph showing the number of IFN- γ -positive spots per 2×10^4 effector cells measured following co-culture of TIL fragments (F1-F24; n=24) from patient 4253 with autologous APCs (1) pulsed with peptide vehicle (DMSO; open bars), (2) pulsed with purified (>95% by HPLC) 25-amino acid peptides composed of mutated p53-R248W (LP-p53-R248W-MUT; black bars) sequence, (3) electroporated with an irrelevant TMG (TMG-IRR; gray hatched bars), or (4) electroporated with p53-mut-TMG containing mutated p53-R248W sequence.

[0025] Figure 17 is a graph showing the number of IFN- γ -positive spots per 2×10^4 effector cells measured following co-culture of TIL fragments (F1-F24, n=24) from patient 4273 with autologous APCs electroporated with TMG (TMG) composed of irrelevant (IRR; open bars) WT p53 (p53-WT; gray bars) or mutated p53 (p53-MUT; black bars) sequence.

[0026] Figure 18 is a graph showing the percentage of CD4+4-1BB+ cells detected following co-culture of patient 4273 TIL fragments F1-F24 (n=24) with autologous APCs

electroporated with TMG composed of irrelevant (IRR; open bars) WT p53 (p53-WT; gray bars) or mutated p53 (p53-MUT; black bars) sequence.

[0027] Figure 19 is a graph showing the number of IFN- γ -positive spots per 2×10^4 effector cells measured following co-culture of TIL fragments (F1-F24, n=24) from patient 4273 with autologous APCs pulsed with peptide vehicle (DMSO; open bars) or purified (>95% by HPLC) 25-amino acid peptides composed of WT p53-R248 sequence (LP-p53-R248-WT; gray bars) or mutated p53-R248W (LP-p53-R248W-MUT; black bars) sequence.

[0028] Figure 20 is a graph showing the percentage of CD4+4-1BB+ cells detected following co-culture of TIL fragments (F1-F24, n=24) from patient 4273 with autologous APCs pulsed with peptide vehicle (DMSO; open bars) or purified (>95% by HPLC) 25-amino acid peptides composed of WT p53-R248 sequence (LP-p53-R248-WT; gray bars) or mutated p53-R248W (LP-p53-R248W-MUT; black bars) sequence.

[0029] Figure 21 is a graph showing the number of IFN- γ -positive spots per 2×10^4 effector cells measured following co-culture of autologous PBL from patient 4149 (transposed with 4149-TCRa2b1 or 4149-TCRa2b2) with autologous APCs which were (1) electroporated with TMG composed of irrelevant (IRR; right hatched black bars), WT p53 (p53-wt-TMG; left hatched black bars) or mutated p53 (p53-mut-TMG; gray bars) sequence or (2) pulsed with peptide vehicle (DMSO; open bars) or purified (>95% by HPLC) 25-amino acid peptides composed of WT p53-Y220 sequence (LP-p53-Y220-WT; horizontal hatched black bars) or mutated p53-Y220C (LP-p53-Y220C-MUT; black bars) sequence. Phorbol 12-myristate 13-acetate (PMA) and Ionomycin (Iono) was positive control (gray bars).

[0030] Figure 22 is a graph showing the percentage of CD4+4-1BB+ cells detected following co-culture of autologous PBL from patient 4149 that were transposed with a TCR (4149-TCRa2b1 or 4149-TCRa2b2) with autologous APCs which were (1) electroporated with TMG composed of irrelevant (IRR; right hatched black bars), WT p53 (p53-wt-TMG; left hatched black bars), or mutated p53 (p53-mut-TMG; gray bars) sequence or (2) pulsed with peptide vehicle (DMSO; open bars) or purified (>95% by HPLC) 25-amino acid peptides composed of WT p53-Y220 sequence (LP-p53-Y220-WT; horizontal hatched black bars) or mutated p53-Y220C (LP-p53-Y220C-MUT; black bars) sequence. PMA and Iono was positive control (gray bars).

[0031] Figure 23 is a graph showing the percentage of 4-1BB+ cells (% of CD4+ T cells) detected following co-culture of TIL from patient 4149 with autologous DCs pulsed with one of the peptides of Table F.

[0032] Figure 24 is a graph showing IFN- γ secretion (pg/mL) following co-culture of 4149-TCRa2b2 transposed T cells with Cos7 cells co-transfected with individual HLA alleles +/- TMGs. Cells not transfected with TMG were pulsed with p53Y220C 15-mer peptide. Pulsed target cells are indicated by shaded bars. Target cells transfected with TMG are indicated by unshaded bars.

[0033] Figure 25 is a graph showing the percentage of CD8+4-1BB+ cells detected following co-culture of TIL fragments (F2 and F24) from patient 4213 with autologous APCs pulsed with peptide vehicle (DMSO; open bars) or purified (>95% by HPLC) 25-amino acid peptides composed of the mutated p53-R248Q (LP-p53-R248Q-MUT; black bars) sequence.

[0034] Figure 26 is a graph showing the number of IFN- γ -positive spots per 2×10^4 effector cells measured following co-culture of CD4+ T cells from patient 4213 with autologous APCs pulsed with peptide vehicle (DMSO; open bars) or purified (>95% by HPLC) 25-amino acid peptides composed of the mutated p53-R248Q (LP-p53-R248Q-MUT; black bars) sequence. Secretion of IFN- γ is shown by open bars. Expression of 4-1BB is shown by closed bars.

[0035] Figure 27 is a graph showing the number of IFN- γ -positive spots per 2×10^4 effector cells measured following co-culture of TIL fragments (F1-F24, n=24) from patient 4268 with autologous APCs electroporated with TMG composed of irrelevant (IRR; open bars), WT p53 (p53-WT; gray bars), or mutated p53 (p53-MUT; black bars) sequence.

[0036] Figure 28 is a graph showing the number of IFN- γ -positive spots per 2×10^4 effector cells measured following co-culture of TIL fragments (F1-F24, n=24) from patient 4268 with autologous APCs pulsed with peptide vehicle (DMSO; open bars) or purified (>95% by HPLC) 25-amino acid peptides composed of WT p53-R248 sequence (LP-p53-R248-WT; gray bars) or mutated p53-R248Q (LP-p53-R248Q-MUT; black bars) sequence.

[0037] Figure 29 is a graph showing the percentage of CD4+4-1BB+ cells detected following co-culture of TIL fragments (F1-F24, n=24) from patient 4268 with autologous APCs pulsed with peptide vehicle (DMSO; open bars) or purified (>95% by HPLC) 25-amino acid peptides composed of WT p53-R248 sequence (LP-p53-R248-WT; gray bars) or mutated p53-R248Q (LP-p53-R248Q-MUT; black bars) sequence.

[0038] Figure 30 is a graph showing the percentage of CD8+4-1BB+ cells detected following co-culture of TIL fragments (F1-F24, n=24) from patient 4268 with autologous APCs pulsed with peptide vehicle (DMSO; open bars) or purified (>95% by HPLC) 25-

amino acid peptides composed of WT p53-R248 sequence (LP-p53-R248-WT; gray bars) or mutated p53-R248Q (LP-p53-R248Q-MUT; black bars) sequence.

[0039] Figure 31 is a graph showing the number of IFN- γ -positive spots per 2×10^4 effector cells measured following co-culture of TIL fragments (F1-F24, n=24) from patient 4266 with autologous APCs electroporated with TMG composed of irrelevant (IRR; open bars), WT p53 (p53-WT; gray bars), or mutated p53 (p53-MUT; black bars) sequence.

[0040] Figure 32 is a graph showing the percentage of CD8+4-1BB+ cells detected following co-culture of TIL fragments (F1-F24, n=24) from patient 4266 with autologous APCs electroporated with TMG composed of irrelevant (IRR; open bars), WT p53 (p53-WT; gray bars), or mutated p53 (p53-MUT; black bars) sequence.

[0041] Figure 33 is a graph showing the number of IFN- γ -positive spots per 2×10^4 effector cells measured following co-culture of TIL fragments (F1-F24, n=24) from patient 4266 with autologous APCs pulsed with peptide vehicle (DMSO; open bars) or purified (>95% by HPLC) 25-amino acid peptides composed of WT p53-R248 sequence (LP-p53-R248-WT; gray bars) or mutated p53-R248W (LP-p53-R248W-MUT; black bars) sequence.

[0042] Figure 34 is a graph showing the percentage of CD8+4-1BB+ cells detected following co-culture of TIL fragments (F1-F24, n=24) from patient 4266 with autologous APCs pulsed with peptide vehicle (DMSO; open bars) or purified (>95% by HPLC) 25-amino acid peptides composed of WT p53-R248 sequence (LP-p53-R248-WT; gray bars) or mutated p53-R248W (LP-p53-R248W-MUT; black bars) sequence.

[0043] Figure 35 shows an alignment of the amino acid sequences of the nine p53 splice variants. SP|P04637|P53_HUMAN (SEQ ID NO: 1); SP|P04637-2|P53_HUMAN (SEQ ID NO: 535); SP|P04637-3|P53_HUMAN (SEQ ID NO: 536); SP|P04637-4|P53_HUMAN (SEQ ID NO: 537); SP|P04637-5|P53_HUMAN (SEQ ID NO: 538); SP|P04637-6|P53_HUMAN (SEQ ID NO: 539); SP|P04637-7|P53_HUMAN (SEQ ID NO: 540); SP|P04637-8|P53_HUMAN (SEQ ID NO: 541); and SP|P04637-9|P53_HUMAN (SEQ ID NO: 542).

[0044] Figure 36 is a graph showing the percentage of 4-1BB positive cells (% of mTCR β +) detected following co-culture of T cells transduced with the 4127-O37-TCR with autologous APCs pulsed with decreasing concentrations of 25-amino acid peptides corresponding to the WT (open circles) or mutated (closed squares) p53-G245S sequence.

[0045] Figures 37A-37C are graphs showing percentage of 4-1BB positive cells (% of mTCR β +) (left y-axis; hatched bars) and IFN- γ (pg/mL) (right y-axis; black bars) measured

following co-culture of T cells transduced with the 4127-O37-TCR (A), 4127-TP53-G245S-TCR1 (B), or 4127-O102-TCR (C) with COS7 cells co-transfected with one of the indicated HLA alleles and pulsed with the 25-amino acid p53-G245S peptide.

[0046] Figure 38 is a graph showing the percentage of 4-1BB positive cells (% of CD8+) (right y-axis; black bars) and IFN- γ (spots per 2×10^4 cells) (left y-axis; hatched bars) measured following co-culture of TIL from Patient 4141 (fragment culture 12) with autologous APCs transfected with TMG encoding irrelevant mutations (TMG-IRR), WT p53 sequence (TP53-wt-TMG) or mutated p53 sequence including R175H (TP53-mut-TMG). Media alone and PMA and ionomycin were negative and positive controls, respectively.

[0047] Figure 39 is a graph showing the number of IFN- γ -positive spots per 2×10^4 effector cells measured following co-culture of TIL from Patient 4141 (fragment culture 12) with Cos7 cells co-transfected with the indicated HLA alleles and either no extra gene (HLA only; open bars), WT TP53 TMG (gray hatched bars), or mutated (black bars) TP53 TMG containing the p53-R175H sequence.

[0048] Figure 40 is a graph showing the concentration of IFN- γ (pg/mL) measured following co-culture of T cells expressing mock (no TCR) or 4141-TCR1a2 with T2 tumor cells (expressing HLA-A*02:01). T2 cells were pulsed with peptide vehicle (DMSO; gray bars) or purified (>95% by HPLC) peptides composed of WT p53-R175 peptide (hatched gray bars) or mutated p53-R175H peptide (black bars). Media alone (open bars) and PMA and Ionomycin (lattice bars) were negative and positive controls, respectively. Data are mean $\pm$ SEM (n=3).

[0049] Figure 41 is a graph showing the percentage of cells positive for expression of one of the indicated markers following co-culture of T cells expressing 4141-TCR1a2 with Saos2 cells (p53-NUL and HLA-A*02:01+), which were either unmanipulated (unshaded bars) or made to overexpress full length p53-R175H protein (shaded bars). Data are mean $\pm$ SEM (n=3). Student's two-tailed t-tests were performed for each cytokine between the two cell lines for statistical analyses (**p<0.001).

[0050] Figure 42 is a graph showing the percentage of CD4+4-1BB positive cells detected following co-culture of TIL (fragment culture 6 from patient 4259) with autologous APCs either (1) electroporated with TMG composed of irrelevant (TMG-IRR), WT p53 (TMG-p53-WT) or mutated p53 (TMG-p53-MUT) sequence or (2) pulsed with peptide vehicle (DMSO) or purified (>95% by HPLC) 25-amino acid peptides composed of WT p53-

Y220 sequence (LP-p53-Y220-WT) or mutated p53-Y220C (LP-p53-Y220C-MUT) sequence.

[0051] Figure 43 is a graph showing the percentage of CD4+4-1BB positive cells detected following co-culture of TIL fragment culture (no. 6) from patient 4259 with autologous APCs pulsed with decreasing concentrations of 25-amino acid peptides corresponding to the WT p53 sequence (open circles) or mutated p53-Y220C (closed squares) for 2 hours at 37 °C.

[0052] Figure 44 is a graph showing the percentage of CD4+4-1BB positive cells detected following co-culture of TIL from patient 4259 with autologous APCs pulsed with DMSO, WT p53-Y220 peptide, or mutated p53-Y220C peptide.

[0053] Figure 45 is a graph showing the percentage of 4-1BB positive cells (% of CD4+) detected following co-culture of TIL fragment culture no. 6 from Patient 4259 with Cos7 cells co-transfected with the indicated HLA alleles from patient 4259 and pulsed with DMSO (open bars) or the p53-Y220C peptide (closed bars).

[0054] Figure 46 is a graph showing the percentage of 4-1BB+ cells detected following co-culture of TC#4259 target cells (endogenously expressing p53-Y220C and HLA-DRB1*04:01:01) with effector T cells (10^5) expressing mock (no TCR; open bars) or p53-Y220C-specific TCR (4259-F6-TCR; black bars). The TC#4259 cells were either incubated with nothing, W6/32 pan-HLA Class-I specific blocking antibody, IVA12 pan-HLA Class-II specific blocking antibody or mutated p53-Y220C peptide. Media alone (no TC) and PMA and Ionomycin were negative and positive controls, respectively. Data are mean $\pm$ SEM (n=3). Student's two-tailed t-tests were performed between groups as indicated for statistical analyses (**p<0.01 and ***p<0.001).

[0055] Figure 47 is a graph showing the number of IFN- γ -positive spots per 2×10^4 effector cells measured following co-culture of TIL fragments (F1-F22 and F24, n=23) from patient 4285 with autologous APCs electroporated with TMG composed of irrelevant (IRR; open bars), WT p53 (p53-WT; gray bars) or mutated p53 (p53-MUT; black bars) sequence.

[0056] Figure 48 is a graph showing the number of IFN- γ -positive spots per 2×10^4 effector cells measured following co-culture of TIL fragments (F1-F22 and F24, n=23) from patient 4285 with autologous APCs pulsed with peptide vehicle (DMSO; open bars) or purified (>95% by HPLC) 25-amino acid peptides composed of WT p53-R175 sequence (LP-p53-R175-WT; gray bars) or mutated p53-R175H (LP-p53-R175H-MUT; black bars) sequence.

[0057] Figure 49 is a graph showing the percentage of CD4⁺4-1BB⁺ cells detected following co-culture of TIL fragments (F1-F22 and F24, n=23) from patient 4285 with autologous APCs electroporated with TMG composed of irrelevant (IRR; open bars), WT type p53 (p53-WT; gray bars) or mutated p53 (p53-MUT; black bars) sequence.

[0058] Figure 50 is a graph showing the percentage of CD4⁺4-1BB⁺ cells detected following co-culture of TIL fragments (F1-F22 and F24, n=23) from patient 4285 with autologous APCs pulsed with peptide vehicle (DMSO; open bars) or purified (>95% by HPLC) 25-amino acid peptides composed of WT p53-R175 sequence (LP-p53-R175-WT; gray bars) or mutated p53-R175H (LP-p53-R175H-MUT; black bars) sequences.

[0059] Figure 51 is a graph showing the percentage of 4-1BB⁺ cells (% of CD4⁺) detected following co-culture of TIL fragment cultures 4285-F6, 4285-F9 and 4285-F10 with Cos7 cells transfected with the indicated HLA alleles and pulsed with DMSO (peptide vehicle; gray and black hatched bars) or mutated p53-R175H peptide (gray, lattice and black bars).

[0060] Figure 52 is a graph showing the percentage of 4-1BB⁺ cells (% of CD4⁺) detected following co-culture of T cells transposed with 4285-TCR1 with autologous APCs pulsed with decreasing concentrations of 25- or 15-amino acid peptides corresponding to the WT (open circles and squares) or mutated (closed circles and squares) p53-R175H sequence.

[0061] Figure 53 is a graph showing the percentage of 4-1BB⁺ cells (% of CD8⁺) detected following co-culture of TIL from patient 4266 with Cos7 cells which were co-transfected with individual HLA alleles from patient 4266 and pulsed with no peptide (open bars), DMSO (peptide vehicle; gray bars), wild type p53-R248 peptide (gray hatched bars) or mutated p53-R248W peptide (black bars). Data are mean $\pm$ SEM (n=3).

[0062] Figure 54 is a graph showing the percentage of 4-1BB⁺ cells (% of CD8⁺) detected following co-culture of T cells expressing mock (no TCR), 4266-TCR1, 4266-TCR2, 4266-TCR3 or 4266-TCR4 with autologous APCs which were pulsed with peptide vehicle (DMSO; gray bars) or purified (>95% by HPLC) 25 amino acid peptides composed of WT p53-R248 sequence (hatched gray bars) or mutated p53-R248W (black bars) sequences. Media alone (open bars) and PMA and Ionomycin (lattice bars) were negative and positive controls, respectively.

[0063] Figure 55 is a graph showing the percentage of 4-1BB⁺ cells (% of CD8⁺) detected following co-culture of T cells expressing mock (no TCR) or p53-R248W-specific TCRs (4266-TCR2, 4266-TCR3 or 4266-TCR4) with tumor cell (TC) line established from

xenografted tumor fragment resected from Patient 4266 then serially passaged through immunocompromised mice (TC#4266). The TC#4266 cells were either incubated with nothing (black bars), W6/32 pan-HLA Class-I specific blocking antibody (right gray hatched bars), IVA12 pan-HLA Class-II specific blocking antibody (gray bars) or mutated p53-R248W peptide (left gray hatched bars). Media alone (no TC; open bars) and PMA and Ionomycin (gray lattice bars) were negative and positive controls, respectively.

[0064] Figure 56 is a graph showing the percentage of CD4⁺ 4-1BB⁺ cells detected following co-culture of TIL from patient 4273 with autologous APCs which were transfected with TMG encoding irrelevant mutations (gray bars), wild type p53 sequences (gray hatched bars) or mutated p53 sequences including p53-R248W (black bars). Media alone (open bars) and PMA and Ionomycin (lattice bars) were negative and positive controls, respectively.

[0065] Figure 57 is a graph showing the percentage of CD4⁺ 4-1BB⁺ cells detected following co-culture of TIL from patient 4273 with autologous APCs which were pulsed with 25 amino acid peptides corresponding to the wild type (open circles) or mutated (closed squares) from the p53-R248W neoepitope. DMSO was peptide vehicle.

[0066] Figure 58 is a graph showing the percentage of CD4⁺ 4-1BB⁺ cells detected following co-culture of TIL from patient 4273 with autologous APCs pulsed with 15 amino acid peptides from the p53-R248W neoepitope (amino acid substitution in bold) overlapping 14 amino acids. DMSO was peptide vehicle, media alone (T cells only) and PMA and ionomycin were controls. The 25 amino acid peptides (wt p53-R248 and mutated p53-R248W) were additional controls for the 15 amino acid peptides. The peptides are: YMCNSSCMGGMN**W**RP (SEQ ID NO: 592); MCNSSCMGGMN**W**RPI (SEQ ID NO: 593); CNSSCMGGMN**W**RPIL (SEQ ID NO: 594); NSSCMGGMN**W**RPILT (SEQ ID NO: 595); SSCMGGMN**W**RPILTI (SEQ ID NO: 596); SCMGGMN**W**RPILTII (SEQ ID NO: 597); CMGGMN**W**RPILTIIT (SEQ ID NO: 598); MGGMN**W**RPILTIITL (SEQ ID NO: 599); GGMN**W**RPILTIITL (SEQ ID NO: 600); GMN**W**RPILTIITLED (SEQ ID NO: 601); and MNWRPILTIITLED (SEQ ID NO: 602).

[0067] Figure 59 is a graph showing the concentration of IFN- γ (pg/mL) secreted following co-culture of TIL from Patient 4273 with Cos7 cells co-transfected with individual HLA alleles from patient 4273 and either WT (open bars) or mutated (black bars) TP53 TMG with or without the p53-R248W neoantigen, respectively.

[0068] Figure 60 is a graph showing the number of IFN- γ spots per 2×10^4 cells measured following co-culture of T cells expressing mock (no TCR) or 4273-TCR1a2 with autologous

APCs which were pulsed with peptide vehicle (DMSO; gray bars) or purified (>95% by HPLC) 25 amino acid peptides composed of WT p53-R248 sequence (hatched gray bars) or mutated p53-R248W (black bars) sequences. Media alone (open bars) and PMA and Ionomycin (lattice bars) were negative and positive controls, respectively.

DETAILED DESCRIPTION OF THE INVENTION

[0069] Tumor Protein P53 (also referred to as “TP53” or “p53”) acts as a tumor suppressor by, for example, regulating cell division. The p53 protein is located in the nucleus of the cell, where it binds directly to DNA. When DNA becomes damaged, the p53 protein is involved in determining whether the DNA will be repaired or the damaged cell will undergo apoptosis. If the DNA can be repaired, p53 activates other genes to fix the damage. If the DNA cannot be repaired, the p53 protein prevents the cell from dividing and signals it to undergo apoptosis. By stopping cells with mutated or damaged DNA from dividing, p53 helps prevent the development of tumors. Wild-type (WT) (normal) full-length p53 comprises the amino acid sequence of SEQ ID NO: 1.

[0070] Mutations in the p53 protein may reduce or eliminate the p53 protein’s tumor suppressor function. Alternatively or additionally, a p53 mutation may be a gain-of-function mutation by interfering with WT p53 in a dominant negative fashion. Mutated p53 protein may be expressed in any of a variety of human cancers such as, for example, cholangiocarcinoma, melanoma, colon cancer, rectal cancer, ovarian cancer, endometrial cancer, non-small cell lung cancer (NSCLC), glioblastoma, uterine cervical cancer, head and neck cancer, breast cancer, pancreatic cancer, or bladder cancer.

[0071] An embodiment of the invention provides an isolated or purified T cell receptor (TCR) having antigenic specificity for mutated human p53 (hereinafter, “mutated p53”). Hereinafter, references to a “TCR” also refer to functional portions and functional variants of the TCR, unless specified otherwise. Mutations of p53 are defined herein by reference to the amino acid sequence of full-length, WT p53 (SEQ ID NO: 1). Thus, mutations of p53 are described herein by reference to the amino acid residue present at a particular position, followed by the position number, followed by the amino acid with which that residue has been replaced in the particular mutation under discussion. For example, when the positions are as defined by SEQ ID NO: 1, the term “R175” refers to the arginine present at position 175 of SEQ ID NO: 1, “R175H” indicates that the arginine present at position 175 of SEQ ID NO: 1 is replaced by histidine, while “G245S” indicates that the glycine present at position

245 of SEQ ID NO: 1 has been replaced with serine. P53 has nine known splice variants. The p53 mutations described herein are conserved over all nine p53 splice variants. An alignment of the nine p53 splice variants is shown in Figure 35. Accordingly, T cells isolated by the inventive methods may have antigenic specificity for any mutated p53 amino acid sequence described herein encoded by any of the nine p53 splice variants. When the positions are as defined by SEQ ID NO: 1, then the actual positions of the amino acid sequence of a particular splice variant of p53 are defined relative to the corresponding positions of SEQ ID NO: 1, and the positions as defined by SEQ ID NO: 1 may be different than the actual positions in a particular splice variant. Thus, for example, mutations refer to a replacement of an amino acid residue in the amino acid sequence of a particular splice variant of p53 corresponding to the indicated position of the 393-amino acid sequence of SEQ ID NO: 1 with the understanding that the actual positions in the splice variant may be different.

[0072] In an embodiment of the invention, the TCR has antigenic specificity for human p53 with a mutation at position 175, 220, 245, 248, 249, 273, or 282 of SEQ ID NO: 1. The TCR may have antigenic specificity for human p53 with a mutation at position 175, 220, 245, or 248 of SEQ ID NO: 1. The p53 mutation may be any missense mutation. Accordingly, the mutation at position 175, 220, 245, 248, 249, 273, or 282 of SEQ ID NO: 1 may be a substitution of the native (WT) amino acid residue present at position 175, 220, 245, 248, 249, 273, or 282 of SEQ ID NO: 1 with any amino acid residue other than the native (WT) amino acid residue present at the particular position under discussion. In an embodiment of the invention, the TCR has antigenic specificity for human p53 with one of the following human p53 mutations: R175H, Y220C, G245D, G245S, R248L, R248Q, R248W, R249S, R273H, R273C, R273L, or R282W. The TCR may have antigenic specificity for human p53 with one of the following human p53 mutations: R175H, Y220C, G245S, R248Q, or R248W. For example, the inventive TCR may have antigenic specificity for a mutated p53 amino acid sequence selected from the group consisting of SEQ ID NOs: 2-13.

[0073] In an embodiment of the invention, the inventive TCRs may be able to recognize mutated p53 in an HLA (human leukocyte antigen)-molecule-dependent manner. "HLA-molecule-dependent manner," as used herein, means that the TCR elicits an immune response upon binding to mutated p53 within the context of an HLA molecule, which HLA molecule is expressed by the patient from which the TCR was isolated. The inventive TCRs may be able to recognize mutated p53 that is presented by the applicable HLA molecule and may bind to the HLA molecule in addition to mutated p53.

[0074] The TCRs of the invention provide many advantages, including when expressed by cells used for adoptive cell transfer. Mutated p53 is expressed by cancer cells and is not expressed by normal, noncancerous cells. Without being bound to a particular theory or mechanism, it is believed that the inventive TCRs advantageously target the destruction of cancer cells while minimizing or eliminating the destruction of normal, non-cancerous cells, thereby reducing, for example, by minimizing or eliminating, toxicity. Moreover, the inventive TCRs may, advantageously, successfully treat or prevent mutated p53-positive cancers that do not respond to other types of treatment such as, for example, chemotherapy, surgery, or radiation. Additionally, the inventive TCRs may provide highly avid recognition of mutated p53, which may provide the ability to recognize unmanipulated tumor cells (e.g., tumor cells that have not been treated with interferon (IFN)- γ , transfected with a vector encoding one or both of mutated p53 and the applicable HLA molecule, pulsed with a p53 peptide with the p53 mutation, or a combination thereof). Roughly half of all tumors harbor a mutation in p53, about half of which will be a missense mutation and about 30% of the missense mutations occur at the following “hotspot” residues: R175H, Y220C, G245D, G245S, R248L, R248Q, R248W, R249S, R273C, R273L, R273H and R282W. Moreover, the same “hotspot” mutations in p53 (e.g., R175H, Y220C, G245D, G245S, R248L, R248Q, R248W, R249S, R273H, R273C, R273L, or R282W) occur frequently (cumulatively about 30% of the p53 missense mutations) in tumors of unrelated people. Accordingly, the inventive TCRs may increase the number of patients who may be eligible for treatment with immunotherapy.

[0075] The phrase “antigenic specificity,” as used herein, means that the TCR can specifically bind to and immunologically recognize mutated p53 with high avidity. For example, a TCR may be considered to have “antigenic specificity” for mutated p53 if about 1×10^4 to about 1×10^5 T cells expressing the TCR secrete at least about 200 pg/mL or more (e.g., 200 pg/mL or more, 300 pg/mL or more, 400 pg/mL or more, 500 pg/mL or more, 600 pg/mL or more, 700 pg/mL or more, 1000 pg/mL or more, 5,000 pg/mL or more, 7,000 pg/mL or more, 10,000 pg/mL or more, 20,000 pg/mL or more, or a range defined by any two of the foregoing values) of IFN- γ upon co-culture with (a) antigen-negative, applicable HLA molecule positive target cells pulsed with a low concentration of mutated p53 peptide (e.g., about 0.05 ng/mL to about 5 ng/mL, 0.05 ng/mL, 0.1 ng/mL, 0.5 ng/mL, 1 ng/mL, 5 ng/mL, or a range defined by any two of the foregoing values) or (b) antigen-negative, applicable HLA molecule positive target cells into which a nucleotide sequence encoding mutated p53

has been introduced such that the target cell expresses mutated p53. Cells expressing the inventive TCRs may also secrete IFN- γ upon co-culture with antigen-negative, applicable HLA molecule positive target cells pulsed with higher concentrations of mutated p53 peptide.

[0076] Alternatively or additionally, a TCR may be considered to have “antigenic specificity” for mutated p53 if T cells expressing the TCR secrete at least twice as much IFN- γ upon co-culture with (a) antigen-negative, applicable HLA molecule positive target cells pulsed with a low concentration of mutated p53 peptide or (b) antigen-negative, applicable HLA molecule positive target cells into which a nucleotide sequence encoding mutated p53 has been introduced such that the target cell expresses mutated p53 as compared to the amount of IFN- γ expressed by a negative control. The negative control may be, for example, (i) T cells expressing the TCR, co-cultured with (a) antigen-negative, applicable HLA molecule positive target cells pulsed with the same concentration of an irrelevant peptide (e.g., some other peptide with a different sequence from the mutated p53 peptide) or (b) antigen-negative, applicable HLA molecule positive target cells into which a nucleotide sequence encoding an irrelevant peptide has been introduced such that the target cell expresses the irrelevant peptide, or (ii) untransduced T cells (e.g., derived from PBMC, which do not express the TCR) co-cultured with (a) antigen-negative, applicable HLA molecule positive target cells pulsed with the same concentration of mutated p53 peptide or (b) antigen-negative, applicable HLA molecule positive target cells into which a nucleotide sequence encoding mutated p53 has been introduced such that the target cell expresses mutated p53. IFN- γ secretion may be measured by methods known in the art such as, for example, enzyme-linked immunosorbent assay (ELISA).

[0077] Alternatively or additionally, a TCR may be considered to have “antigenic specificity” for mutated p53 if at least twice as many of the numbers of T cells expressing the TCR secrete IFN- γ upon co-culture with (a) antigen-negative, applicable HLA molecule positive target cells pulsed with a low concentration of mutated p53 peptide or (b) antigen-negative, applicable HLA molecule positive target cells into which a nucleotide sequence encoding mutated p53 has been introduced such that the target cell expresses mutated p53 as compared to the numbers of negative control T cells that secrete IFN- γ . The concentration of peptide and the negative control may be as described herein with respect to other aspects of the invention. The numbers of cells secreting IFN- γ may be measured by methods known in the art such as, for example, enzyme-linked immunospot (ELISOT) assay.

[0078] Alternatively or additionally, a TCR may be considered to have “antigenic specificity” for mutated p53 if at least twice as many spots are detected by ELISPOT for the T cells expressing the TCR upon co-culture with (a) antigen-negative, applicable HLA molecule positive target cells pulsed with a low concentration of mutated p53 peptide or (b) antigen-negative, applicable HLA molecule positive target cells into which a nucleotide sequence encoding mutated p53 has been introduced such that the target cell expresses mutated p53 as compared to the number of spots detected by ELISPOT for negative control T cells co-cultured with the same target cells. The concentration of peptide and the negative control may be as described herein with respect to other aspects of the invention.

[0079] Alternatively or additionally, a TCR may be considered to have “antigenic specificity” for mutated p53 if greater than about 50 spots are detected by ELISPOT for the T cells expressing the TCR upon co-culture with (a) antigen-negative, applicable HLA molecule positive target cells pulsed with a low concentration of mutated p53 peptide or (b) antigen-negative, applicable HLA molecule positive target cells into which a nucleotide sequence encoding mutated p53 has been introduced such that the target cell expresses mutated p53. The concentration of peptide may be as described herein with respect to other aspects of the invention.

[0080] Alternatively or additionally, a TCR may be considered to have “antigenic specificity” for mutated p53 if T cells expressing the TCR upregulate expression of one or both of 4-1BB and OX40 as measured by, for example, flow cytometry after stimulation with target cells expressing mutated p53.

[0081] The invention provides a TCR comprising two polypeptides (i.e., polypeptide chains), such as an alpha (α) chain of a TCR, a beta (β) chain of a TCR, a gamma (γ) chain of a TCR, a delta (δ) chain of a TCR, or a combination thereof. The polypeptides of the inventive TCR can comprise any amino acid sequence, provided that the TCR has antigenic specificity for mutated p53.

[0082] In an embodiment of the invention, the TCR comprises two polypeptide chains, each of which comprises a variable region comprising a complementarity determining region (CDR)1, a CDR2, and a CDR3 of a TCR. In an embodiment of the invention, the TCR comprises a first polypeptide chain comprising an α chain CDR1 (CDR1 α), an α chain CDR2 (CDR2 α), and an α chain CDR3 (CDR3 α), and a second polypeptide chain comprising a β chain CDR1 (CDR1 β), a β chain CDR2 (CDR2 β), and a β chain CDR3 (CDR3 β). In an embodiment of the invention, the TCR comprises the amino acid sequences of: (1) all of

SEQ ID NOs: 27-32; (2) all of SEQ ID NOs: 37-42; (3) all of SEQ ID NOs: 47-52; (4) all of SEQ ID NOs: 57-62; (5) all of SEQ ID NOs: 67-72; (6) all of SEQ ID NOs: 77-82; (7) all of SEQ ID NOs: 87-92; (8) all of SEQ ID NOs: 97-102; (9) all of SEQ ID NOs: 107-112; (10) all of SEQ ID NOs: 117-122; (11) all of SEQ ID NOs: 127-132; (12) all of SEQ ID NOs: 137-142; (13) all of SEQ ID NOs: 147-152; (14) all of SEQ ID NOs: 157-162; (15) all of SEQ ID NOs: 167-172; (16) all of SEQ ID NOs: 177-182; (17) all of SEQ ID NOs: 187-192; (18) all of SEQ ID NOs: 197-202; (19) all of SEQ ID NOs: 207-212; (20) all of SEQ ID NOs: 217-222; (21) all of SEQ ID NOs: 227-232; (22) all of SEQ ID NOs: 237-242; (23) all of SEQ ID NOs: 247-252; (24) all of SEQ ID NOs: 257-262; (25) all of SEQ ID NOs: 267-272; (26) all of SEQ ID NOs: 277-282; (27) all of SEQ ID NOs: 287-292; (28) all of SEQ ID NOs: 297-302; (29) all of SEQ ID NOs: 307-312; (30) all of SEQ ID NOs: 317-322; (31) all of SEQ ID NOs: 327-332; (32) all of SEQ ID NOs: 337-342; (33) all of SEQ ID NOs: 347-352; (34) all of SEQ ID NOs: 357-362; (35) all of SEQ ID NOs: 367-372; (36) all of SEQ ID NOs: 377-382; (37) all of SEQ ID NOs: 387-392; (38) all of SEQ ID NOs: 397-402; (39) all of SEQ ID NOs: 407-412; (40) all of SEQ ID NOs: 417-422; (41) all of SEQ ID NOs: 427-432; (42) all of SEQ ID NOs: 437-442; (43) all of SEQ ID NOs: 447-452; (44) all of SEQ ID NOs: 457-462; (45) all of SEQ ID NOs: 467-472; (46) all of SEQ ID NOs: 477-482; or (47) all of SEQ ID NOs: 487-492. Each one of the foregoing 47 collections of amino acid sequences in this paragraph sets forth the six CDR regions of each of 47 different TCRs having antigenic specificity for mutated human p53. The six amino acid sequences in each collection correspond to the CDR1 α , CDR2 α , CDR3 α , CDR1 β , CDR2 β , and CDR3 β of a TCR, respectively.

[0083] In an embodiment of the invention, the TCR comprises an α chain variable region amino acid sequence and a β chain variable region amino acid sequence which together comprise one of the collections of CDRs set forth above. In this regard, the TCR can comprise the amino acid sequences of: (1) both of SEQ ID NOs: 33-34; (2) both of SEQ ID NOs: 43-44; (3) both of SEQ ID NOs: 53-54; (4) both of SEQ ID NOs: 63-64; (5) both of SEQ ID NOs: 73-74; (6) both of SEQ ID NOs: 83-84; (7) both of SEQ ID NOs: 93-94; (8) both of SEQ ID NOs: 103-104; (9) both of SEQ ID NOs: 113-114; (10) both of SEQ ID NOs: 123-124; (11) both of SEQ ID NOs: 133-134; (12) both of SEQ ID NOs: 143-144; (13) both of SEQ ID NOs: 153-154; (14) both of SEQ ID NOs: 163-164; (15) both of SEQ ID NOs: 173-174; (16) both of SEQ ID NOs: 183-184; (17) both of SEQ ID NOs: 193-194; (18) both of SEQ ID NOs: 203-204; (19) both of SEQ ID NOs: 213-214; (20) both of SEQ ID NOs:

223-224; (21) both of SEQ ID NOs: 233-234; (22) both of SEQ ID NOs: 243-244; (23) both of SEQ ID NOs: 253-254; (24) both of SEQ ID NOs: 263-264; (25) both of SEQ ID NOs: 273-274; (26) both of SEQ ID NOs: 283-284; (27) both of SEQ ID NOs: 293-294; (28) both of SEQ ID NOs: 303-304; (29) both of SEQ ID NOs: 313-314; (30) both of SEQ ID NOs: 323-324; (31) both of SEQ ID NOs: 333-334; (32) both of SEQ ID NOs: 343-344; (33) both of SEQ ID NOs: 353-354; (34) both of SEQ ID NOs: 363-364; (35) both of SEQ ID NOs: 373-374; (36) both of SEQ ID NOs: 383-384; (37) both of SEQ ID NOs: 393-394; (38) both of SEQ ID NOs: 403-404; (39) both of SEQ ID NOs: 413-414; (40) both of SEQ ID NOs: 423-424; (41) both of SEQ ID NOs: 433-434; (42) both of SEQ ID NOs: 443-444; (43) both of SEQ ID NOs: 453-454; (44) both of SEQ ID NOs: 463-464; (45) both of SEQ ID NOs: 473-474; (46) both of SEQ ID NOs: 483-484; or (47) both of SEQ ID NOs: 493-494. Each one of the foregoing 47 collections of amino acid sequences in this paragraph sets forth the two variable regions of each of 47 different TCRs having antigenic specificity for mutated human p53. The two amino acid sequences in each collection correspond to the variable region of the α chain and the variable region of the β chain of a TCR, respectively.

[0084] The inventive TCRs may further comprise a constant region. The constant region may be derived from any suitable species such as, e.g., human or mouse. In an embodiment of the invention, the TCRs further comprise a murine constant region. As used herein, the term “murine” or “human,” when referring to a TCR or any component of a TCR described herein (e.g., complementarity determining region (CDR), variable region, constant region, alpha chain, and/or beta chain), means a TCR (or component thereof) which is derived from a mouse or a human, respectively, i.e., a TCR (or component thereof) that originated from or was, at one time, expressed by a mouse T cell or a human T cell, respectively. In an embodiment of the invention, the TCR may comprise a murine α chain constant region and a murine β chain constant region. The murine α chain constant region may be modified or unmodified. A modified murine α chain constant region may be, e.g., cysteine-substituted, LVL-modified, or both cysteine-substituted and LVL-modified, as described, for example, in US 2017/0145070. The murine β chain constant region may be modified or unmodified. A modified murine β chain constant region may be, e.g., cysteine-substituted, as described, for example, in US 2017/0145070. In an embodiment of the invention, the TCR comprises a cysteine-substituted, LVL-modified murine α chain constant region comprising the amino acid sequence of SEQ ID NO: 23 or 24. In an embodiment of the invention, the TCR

comprises a cysteine-substituted murine β chain constant region comprising the amino acid sequence of SEQ ID NO: 25.

[0085] In an embodiment of the invention, the inventive TCR can comprise an α chain of a TCR and a β chain of a TCR. The α chain of the TCR may comprise a variable region of an α chain and a constant region of an α chain. An α chain of this type can be paired with any β chain of a TCR. The β chain may comprise a variable region of a β chain and a constant region of a β chain. In an embodiment of the invention, the TCR comprises the amino acid sequences of: (1) both of SEQ ID NOs: 35-36; (2) both of SEQ ID NOs: 45-46; (3) both of SEQ ID NOs: 55-56; (4) both of SEQ ID NOs: 65-66; (5) both of SEQ ID NOs: 75-76; (6) both of SEQ ID NOs: 85-86; (7) both of SEQ ID NOs: 95-96; (8) both of SEQ ID NOs: 105-106; (9) both of SEQ ID NOs: 115-116; (10) both of SEQ ID NOs: 125-126; (11) both of SEQ ID NOs: 135-136; (12) both of SEQ ID NOs: 145-146; (13) both of SEQ ID NOs: 155-156; (14) both of SEQ ID NOs: 165-166; (15) both of SEQ ID NOs: 175-176; (16) both of SEQ ID NOs: 185-186; (17) both of SEQ ID NOs: 195-196; (18) both of SEQ ID NOs: 205-206; (19) both of SEQ ID NOs: 215-216; (20) both of SEQ ID NOs: 225-226; (21) both of SEQ ID NOs: 235-236; (22) both of SEQ ID NOs: 245-246; (23) both of SEQ ID NOs: 255-256; (24) both of SEQ ID NOs: 265-266; (25) both of SEQ ID NOs: 275-276; (26) both of SEQ ID NOs: 285-286; (27) both of SEQ ID NOs: 295-296; (28) both of SEQ ID NOs: 305-306; (29) both of SEQ ID NOs: 315-316; (30) both of SEQ ID NOs: 325-326; (31) both of SEQ ID NOs: 335-336; (32) both of SEQ ID NOs: 345-346; (33) both of SEQ ID NOs: 355-356; (34) both of SEQ ID NOs: 365-366; (35) both of SEQ ID NOs: 375-376; (36) both of SEQ ID NOs: 385-386; (37) both of SEQ ID NOs: 395-396; (38) both of SEQ ID NOs: 405-406; (39) both of SEQ ID NOs: 415-416; (40) both of SEQ ID NOs: 425-426; (41) both of SEQ ID NOs: 435-436; (42) both of SEQ ID NOs: 445-446; (43) both of SEQ ID NOs: 455-456; (44) both of SEQ ID NOs: 465-466; (45) both of SEQ ID NOs: 475-476; (46) both of SEQ ID NOs: 485-486; or (47) both of SEQ ID NOs: 495-496. Each one of the foregoing 47 collections of amino acid sequences in this paragraph sets forth the α chain and β chain of each of 47 different TCRs having antigenic specificity for mutated human p53. The two amino acid sequences in each collection correspond to the α chain and the β chain of a TCR, respectively.

[0086] Included in the scope of the invention are functional variants of the inventive TCRs described herein. The term “functional variant,” as used herein, refers to a TCR, polypeptide, or protein having substantial or significant sequence identity or similarity to a

parent TCR, polypeptide, or protein, which functional variant retains the biological activity of the TCR, polypeptide, or protein of which it is a variant. Functional variants encompass, for example, those variants of the TCR, polypeptide, or protein described herein (the parent TCR, polypeptide, or protein) that retain the ability to specifically bind to mutated p53 for which the parent TCR has antigenic specificity or to which the parent polypeptide or protein specifically binds, to a similar extent, the same extent, or to a higher extent, as the parent TCR, polypeptide, or protein. In reference to the parent TCR, polypeptide, or protein, the functional variant can, for instance, be at least about 30%, at least about 50%, at least about 75%, at least about 80%, at least about 90%, at least about 95%, at least about 96%, at least about 97%, at least about 98%, at least about 99% or more identical in amino acid sequence to the parent TCR, polypeptide, or protein, respectively.

[0087] The functional variant can, for example, comprise the amino acid sequence of the parent TCR, polypeptide, or protein with at least one conservative amino acid substitution. Conservative amino acid substitutions are known in the art, and include amino acid substitutions in which one amino acid having certain physical and/or chemical properties is exchanged for another amino acid that has the same chemical or physical properties. For instance, the conservative amino acid substitution can be an acidic amino acid substituted for another acidic amino acid (e.g., Asp or Glu), an amino acid with a nonpolar side chain substituted for another amino acid with a nonpolar side chain (e.g., Ala, Gly, Val, Ile, Leu, Met, Phe, Pro, Trp, Val, etc.), a basic amino acid substituted for another basic amino acid (Lys, Arg, etc.), an amino acid with a polar side chain substituted for another amino acid with a polar side chain (Asn, Cys, Gln, Ser, Thr, Tyr, etc.), etc.

[0088] Alternatively or additionally, the functional variants can comprise the amino acid sequence of the parent TCR, polypeptide, or protein with at least one non-conservative amino acid substitution. In this case, it is preferable for the non-conservative amino acid substitution to not interfere with or inhibit the biological activity of the functional variant. Preferably, the non-conservative amino acid substitution enhances the biological activity of the functional variant, such that the biological activity of the functional variant is increased as compared to the parent TCR, polypeptide, or protein.

[0089] The TCR, polypeptide, or protein can consist essentially of the specified amino acid sequence or sequences described herein, such that other components of the TCR, polypeptide, or protein, e.g., other amino acids, do not materially change the biological activity of the TCR, polypeptide, or protein.

[0090] Also provided by the invention is a polypeptide comprising a functional portion of any of the TCRs described herein. The term "polypeptide," as used herein, includes oligopeptides and refers to a single chain of amino acids connected by one or more peptide bonds.

[0091] With respect to the inventive polypeptides, the functional portion can be any portion comprising contiguous amino acids of the TCR of which it is a part, provided that the functional portion specifically binds to mutated p53. The term "functional portion," when used in reference to a TCR, refers to any part or fragment of the TCR of the invention, which part or fragment retains the biological activity of the TCR of which it is a part (the parent TCR). Functional portions encompass, for example, those parts of a TCR that retain the ability to specifically bind to mutated p53 (e.g., in an applicable HLA molecule-dependent manner), or detect, treat, or prevent cancer, to a similar extent, the same extent, or to a higher extent, as the parent TCR. In reference to the parent TCR, the functional portion can comprise, for instance, about 10%, about 25%, about 30%, about 50%, about 68%, about 80%, about 90%, about 95%, or more, of the parent TCR.

[0092] The functional portion can comprise additional amino acids at the amino or carboxy terminus of the portion, or at both termini, which additional amino acids are not found in the amino acid sequence of the parent TCR. Desirably, the additional amino acids do not interfere with the biological function of the functional portion, e.g., specifically binding to mutated p53; and/or having the ability to detect cancer, treat or prevent cancer, etc. More desirably, the additional amino acids enhance the biological activity, as compared to the biological activity of the parent TCR.

[0093] The polypeptide can comprise a functional portion of either or both of the α and β chains of the TCRs of the invention, such as a functional portion comprising one of more of CDR1, CDR2, and CDR3 of the variable region(s) of the α chain and/or β chain of a TCR of the invention. In an embodiment of the invention, the polypeptide can comprise a functional portion comprising the amino acid sequences of: (1) all of SEQ ID NOs: 27-32; (2) all of SEQ ID NOs: 37-42; (3) all of SEQ ID NOs: 47-52; (4) all of SEQ ID NOs: 57-62; (5) all of SEQ ID NOs: 67-72; (6) all of SEQ ID NOs: 77-82; (7) all of SEQ ID NOs: 87-92; (8) all of SEQ ID NOs: 97-102; (9) all of SEQ ID NOs: 107-112; (10) all of SEQ ID NOs: 117-122; (11) all of SEQ ID NOs: 127-132; (12) all of SEQ ID NOs: 137-142; (13) all of SEQ ID NOs: 147-152; (14) all of SEQ ID NOs: 157-162; (15) all of SEQ ID NOs: 167-172; (16) all of SEQ ID NOs: 177-182; (17) all of SEQ ID NOs: 187-192; (18) all of SEQ ID NOs: 197-

202; (19) all of SEQ ID NOs: 207-212; (20) all of SEQ ID NOs: 217-222; (21) all of SEQ ID NOs: 227-232; (22) all of SEQ ID NOs: 237-242; (23) all of SEQ ID NOs: 247-252; (24) all of SEQ ID NOs: 257-262; (25) all of SEQ ID NOs: 267-272; (26) all of SEQ ID NOs: 277-282; (27) all of SEQ ID NOs: 287-292; (28) all of SEQ ID NOs: 297-302; (29) all of SEQ ID NOs: 307-312; (30) all of SEQ ID NOs: 317-322; (31) all of SEQ ID NOs: 327-332; (32) all of SEQ ID NOs: 337-342; (33) all of SEQ ID NOs: 347-352; (34) all of SEQ ID NOs: 357-362; (35) all of SEQ ID NOs: 367-372; (36) all of SEQ ID NOs: 377-382; (37) all of SEQ ID NOs: 387-392; (38) all of SEQ ID NOs: 397-402; (39) all of SEQ ID NOs: 407-412; (40) all of SEQ ID NOs: 417-422; (41) all of SEQ ID NOs: 427-432; (42) all of SEQ ID NOs: 437-442; (43) all of SEQ ID NOs: 447-452; (44) all of SEQ ID NOs: 457-462; (45) all of SEQ ID NOs: 467-472; (46) all of SEQ ID NOs: 477-482; or (47) all of SEQ ID NOs: 487-492.

[0094] In an embodiment of the invention, the inventive polypeptide can comprise, for instance, the variable region of the inventive TCR comprising a combination of the CDR regions set forth above. In this regard, the polypeptide can comprise the amino acid sequences of: (1) both of SEQ ID NOs: 33-34; (2) both of SEQ ID NOs: 43-44; (3) both of SEQ ID NOs: 53-54; (4) both of SEQ ID NOs: 63-64; (5) both of SEQ ID NOs: 73-74; (6) both of SEQ ID NOs: 83-84; (7) both of SEQ ID NOs: 93-94; (8) both of SEQ ID NOs: 103-104; (9) both of SEQ ID NOs: 113-114; (10) both of SEQ ID NOs: 123-124; (11) both of SEQ ID NOs: 133-134; (12) both of SEQ ID NOs: 143-144; (13) both of SEQ ID NOs: 153-154; (14) both of SEQ ID NOs: 163-164; (15) both of SEQ ID NOs: 173-174; (16) both of SEQ ID NOs: 183-184; (17) both of SEQ ID NOs: 193-194; (18) both of SEQ ID NOs: 203-204; (19) both of SEQ ID NOs: 213-214; (20) both of SEQ ID NOs: 223-224; (21) both of SEQ ID NOs: 233-234; (22) both of SEQ ID NOs: 243-244; (23) both of SEQ ID NOs: 253-254; (24) both of SEQ ID NOs: 263-264; (25) both of SEQ ID NOs: 273-274; (26) both of SEQ ID NOs: 283-284; (27) both of SEQ ID NOs: 293-294; (28) both of SEQ ID NOs: 303-304; (29) both of SEQ ID NOs: 313-314; (30) both of SEQ ID NOs: 323-324; (31) both of SEQ ID NOs: 333-334; (32) both of SEQ ID NOs: 343-344; (33) both of SEQ ID NOs: 353-354; (34) both of SEQ ID NOs: 363-364; (35) both of SEQ ID NOs: 373-374; (36) both of SEQ ID NOs: 383-384; (37) both of SEQ ID NOs: 393-394; (38) both of SEQ ID NOs: 403-404; (39) both of SEQ ID NOs: 413-414; (40) both of SEQ ID NOs: 423-424; (41) both of SEQ ID NOs: 433-434; (42) both of SEQ ID NOs: 443-444; (43) both of SEQ ID NOs: 453-454; (44) both of SEQ ID NOs: 463-464; (45) both of SEQ ID NOs: 473-474; (46) both of SEQ ID NOs: 483-484; or (47) both of SEQ ID NOs: 493-494.

[0095] In an embodiment of the invention, the inventive polypeptide can further comprise the constant region of the inventive TCR set forth above. In this regard, the polypeptide can comprise the amino acid sequence of (i) one of SEQ ID NOs 23-25 or (ii) SEQ ID NO: 25 and one of SEQ ID NOs: 23-24.

[0096] In an embodiment of the invention, the inventive polypeptide may comprise an α chain and a β chain of the inventive TCR. In this regard, the polypeptide can comprise the amino acid sequences of: (1) both of SEQ ID NOs: 35-36; (2) both of SEQ ID NOs: 45-46; (3) both of SEQ ID NOs: 55-56; (4) both of SEQ ID NOs: 65-66; (5) both of SEQ ID NOs: 75-76; (6) both of SEQ ID NOs: 85-86; (7) both of SEQ ID NOs: 95-96; (8) both of SEQ ID NOs: 105-106; (9) both of SEQ ID NOs: 115-116; (10) both of SEQ ID NOs: 125-126; (11) both of SEQ ID NOs: 135-136; (12) both of SEQ ID NOs: 145-146; (13) both of SEQ ID NOs: 155-156; (14) both of SEQ ID NOs: 165-166; (15) both of SEQ ID NOs: 175-176; (16) both of SEQ ID NOs: 185-186; (17) both of SEQ ID NOs: 195-196; (18) both of SEQ ID NOs: 205-206; (19) both of SEQ ID NOs: 215-216; (20) both of SEQ ID NOs: 225-226; (21) both of SEQ ID NOs: 235-236; (22) both of SEQ ID NOs: 245-246; (23) both of SEQ ID NOs: 255-256; (24) both of SEQ ID NOs: 265-266; (25) both of SEQ ID NOs: 275-276; (26) both of SEQ ID NOs: 285-286; (27) both of SEQ ID NOs: 295-296; (28) both of SEQ ID NOs: 305-306; (29) both of SEQ ID NOs: 315-316; (30) both of SEQ ID NOs: 325-326; (31) both of SEQ ID NOs: 335-336; (32) both of SEQ ID NOs: 345-346; (33) both of SEQ ID NOs: 355-356; (34) both of SEQ ID NOs: 365-366; (35) both of SEQ ID NOs: 375-376; (36) both of SEQ ID NOs: 385-386; (37) both of SEQ ID NOs: 395-396; (38) both of SEQ ID NOs: 405-406; (39) both of SEQ ID NOs: 415-416; (40) both of SEQ ID NOs: 425-426; (41) both of SEQ ID NOs: 435-436; (42) both of SEQ ID NOs: 445-446; (43) both of SEQ ID NOs: 455-456; (44) both of SEQ ID NOs: 465-466; (45) both of SEQ ID NOs: 475-476; (46) both of SEQ ID NOs: 485-486; or (47) both of SEQ ID NOs: 495-496.

[0097] The invention further provides a protein comprising at least one of the polypeptides described herein. By "protein" is meant a molecule comprising one or more polypeptide chains. In an embodiment, the protein of the invention can comprise: (1) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 27-29 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 30-32; (2) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 37-39 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 40-42; (3) a first polypeptide chain comprising the amino acid sequences of all of SEQ

ID NOs: 47-49 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 50-52; (4) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 57-59 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 60-62; (5) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 67-69 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 70-72; (6) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 77-79 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 80-82; (7) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 87-89 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 90-92; (8) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 97-99 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 100-102; (9) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 107-109 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 110-112; (10) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 117-119 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 120-122; (11) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 127-129 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 130-132; (12) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 137-139 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 140-142; (13) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 147-149 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 150-152; (14) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 157-159 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 160-162; (15) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 167-169 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 170-172; (16) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 177-179 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 180-182; (17) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 187-189 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 190-192; (18) a first polypeptide chain comprising the amino acid sequences of all of

SEQ ID NOs: 197-199 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 200-202; (19) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 207-209 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 210-212; (20) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 217-219 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 220-222; (21) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 227-229 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 230-232; (22) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 237-239 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 240-242; (23) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 247-249 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 250-252; (24) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 257-259 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 260-262; (25) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 267-269 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 270-272; (26) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 277-279 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 280-282; (27) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 287-289 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 290-292; (28) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 297-299 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 300-302; (29) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 307-309 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 310-312; (30) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 317-319 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 320-322; (31) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 327-329 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 330-332; (32) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 337-339 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 340-342; (33)

a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 347-349 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 350-352; (34) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 357-359 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 360-362; (35) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 367-369 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 370-372; (36) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 377-379 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 380-382; (37) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 387-389 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 390-392; (38) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 397-399 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 400-402; (39) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 407-409 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 410-412; (40) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 417-419 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 420-422; (41) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 427-429 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 430-432; (42) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 437-439 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 440-442; (43) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 447-449 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 450-452; (44) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 457-459 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 460-462; (45) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 467-469 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 470-472; (46) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 477-479 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 480-482; or (47) a first polypeptide chain comprising the amino acid

sequences of all of SEQ ID NOs: 487-489 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 490-492.

[0098] In an embodiment of the invention, the protein comprises: (1) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 33 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 34; (2) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 43 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 44; (3) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 53 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 54; (4) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 63 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 64; (5) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 73 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 74; (6) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 83 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 84; (7) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 93 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 94; (8) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 103 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 104; (9) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 113 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 114; (10) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 123 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 124; (11) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 133 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 134; (12) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 143 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 144; (13) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 153 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 154; (14) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 163 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 164; (15) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 173 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 174; (16) a first polypeptide chain

comprising the amino acid sequence of SEQ ID NO: 183 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 184; (17) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 193 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 194; (18) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 203 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 204; (19) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 213 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 214; (20) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 223 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 224; (21) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 233 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 234; (22) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 243 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 244; (23) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 253 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 254; (24) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 263 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 264; (25) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 273 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 274; (26) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 283 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 284; (27) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 293 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 294; (28) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 303 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 304; (29) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 313 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 314; (30) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 323 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 324; (31) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 333 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 334; (32) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 343 and a second polypeptide chain

comprising the amino acid sequence of SEQ ID NO: 344; (33) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 353 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 354; (34) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 363 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 364; (35) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 373 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 374; (36) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 383 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 384; (37) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 393 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 394; (38) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 403 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 404; (39) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 413 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 414; (40) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 423 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 424; (41) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 433 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 434; (42) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 443 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 444; (43) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 453 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 454; (44) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 463 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 464; (45) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 473 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 474; (46) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 483 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 484; or (47) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 493 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 494.

[0099] In an embodiment of the invention, the protein comprises: (1) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 35 and a second polypeptide chain

comprising the amino acid sequence of SEQ ID NO: 36; (2) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 45 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 46; (3) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 55 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 56; (4) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 65 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 66; (5) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 75 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 76; (6) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 85 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 86; (7) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 95 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 96; (8) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 105 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 106; (9) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 115 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 116; (10) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 125 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 126; (11) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 135 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 136; (12) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 145 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 146; (13) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 155 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 156; (14) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 165 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 166; (15) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 175 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 176; (16) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 185 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 186; (17) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 195 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 196; (18) a first polypeptide chain

comprising the amino acid sequence of SEQ ID NO: 205 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 206; (19) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 215 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 216; (20) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 225 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 226; (21) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 235 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 236; (22) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 245 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 246; (23) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 255 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 256; (24) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 265 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 266; (25) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 275 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 276; (26) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 285 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 286; (27) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 295 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 296; (28) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 305 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 306; (29) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 315 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 316; (30) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 325 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 326; (31) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 335 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 336; (32) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 345 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 346; (33) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 355 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 356; (34) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 365 and a second polypeptide chain

comprising the amino acid sequence of SEQ ID NO: 366; (35) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 375 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 376; (36) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 385 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 386; (37) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 395 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 396; (38) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 405 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 406; (39) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 415 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 416; (40) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 425 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 426; (41) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 435 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 436; (42) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 445 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 446; (43) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 455 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 456; (44) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 465 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 466; (45) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 475 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 476; (46) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 485 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 486; or (47) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 495 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 496.

[0100] The protein of the invention may be a TCR. Alternatively, if the first and/or second polypeptide chain(s) of the protein further comprise(s) other amino acid sequences, e.g., an amino acid sequence encoding an immunoglobulin or a portion thereof, then the inventive protein can be a fusion protein. In this regard, the invention also provides a fusion protein comprising at least one of the inventive polypeptides described herein along with at least one other polypeptide. The other polypeptide can exist as a separate polypeptide of the

fusion protein, or can exist as a polypeptide, which is expressed in frame (in tandem) with one of the inventive polypeptides described herein. The other polypeptide can encode any peptidic or proteinaceous molecule, or a portion thereof, including, but not limited to an immunoglobulin, CD3, CD4, CD8, an MHC molecule, a CD1 molecule, e.g., CD1a, CD1b, CD1c, CD1d, etc.

[0101] The fusion protein can comprise one or more copies of the inventive polypeptide and/or one or more copies of the other polypeptide. For instance, the fusion protein can comprise 1, 2, 3, 4, 5, or more, copies of the inventive polypeptide and/or of the other polypeptide. Suitable methods of making fusion proteins are known in the art, and include, for example, recombinant methods.

[0102] In some embodiments of the invention, the TCRs, polypeptides, and proteins of the invention may be expressed as a single protein comprising a linker peptide linking the α chain and the β chain. In this regard, the TCRs, polypeptides, and proteins of the invention may further comprise a linker peptide. The linker peptide may advantageously facilitate the expression of a recombinant TCR, polypeptide, and/or protein in a host cell. The linker peptide may comprise any suitable amino acid sequence. For example, the linker peptide may comprise the amino acid sequence of SEQ ID NO: 26. Upon expression of the construct including the linker peptide by a host cell, the linker peptide may be cleaved, resulting in separated α and β chains. In an embodiment of the invention, the TCR, polypeptide, or protein may comprise an amino acid sequence comprising a full-length α chain, a full-length β chain, and a linker peptide positioned between the α and β chains.

[0103] The protein of the invention can be a recombinant antibody, or an antigen binding portion thereof, comprising at least one of the inventive polypeptides described herein. As used herein, "recombinant antibody" refers to a recombinant (e.g., genetically engineered) protein comprising at least one of the polypeptides of the invention and a polypeptide chain of an antibody, or an antigen binding portion thereof. The polypeptide of an antibody, or antigen binding portion thereof, can be a heavy chain, a light chain, a variable or constant region of a heavy or light chain, a single chain variable fragment (scFv), or an Fc, Fab, or F(ab)₂' fragment of an antibody, etc. The polypeptide chain of an antibody, or an antigen binding portion thereof, can exist as a separate polypeptide of the recombinant antibody. Alternatively, the polypeptide chain of an antibody, or an antigen binding portion thereof, can exist as a polypeptide, which is expressed in frame (in tandem) with the polypeptide of the invention. The polypeptide of an antibody, or an antigen binding portion thereof, can be a

polypeptide of any antibody or any antibody fragment, including any of the antibodies and antibody fragments described herein.

[0104] The TCRs, polypeptides, and proteins of the invention can be of any length, i.e., can comprise any number of amino acids, provided that the TCRs, polypeptides, or proteins retain their biological activity, e.g., the ability to specifically bind to mutated p53; detect cancer in a mammal; or treat or prevent cancer in a mammal, etc. For example, the polypeptide can be in the range of from about 50 to about 5000 amino acids long, such as 50, 70, 75, 100, 125, 150, 175, 200, 300, 400, 500, 600, 700, 800, 900, 1000 or more amino acids in length. In this regard, the polypeptides of the invention also include oligopeptides.

[0105] The TCRs, polypeptides, and proteins of the invention of the invention can comprise synthetic amino acids in place of one or more naturally-occurring amino acids. Such synthetic amino acids are known in the art, and include, for example, aminocyclohexane carboxylic acid, norleucine, α -amino n-decanoic acid, homoserine, S-acetylaminomethyl-cysteine, trans-3- and trans-4-hydroxyproline, 4-aminophenylalanine, 4-nitrophenylalanine, 4-chlorophenylalanine, 4-carboxyphenylalanine, β -phenylserine β -hydroxyphenylalanine, phenylglycine, α -naphthylalanine, cyclohexylalanine, cyclohexylglycine, indoline-2-carboxylic acid, 1,2,3,4-tetrahydroisoquinoline-3-carboxylic acid, aminomalonic acid, aminomalonic acid monoamide, N'-benzyl-N'-methyl-lysine, N',N'-dibenzyl-lysine, 6-hydroxylysine, ornithine, α -aminocyclopentane carboxylic acid, α -aminocyclohexane carboxylic acid, α -aminocycloheptane carboxylic acid, α -(2-amino-2-norbornane)-carboxylic acid, α,γ -diaminobutyric acid, α,β -diaminopropionic acid, homophenylalanine, and α -tert-butylglycine.

[0106] The TCRs, polypeptides, and proteins of the invention can be glycosylated, amidated, carboxylated, phosphorylated, esterified, N-acylated, cyclized via, e.g., a disulfide bridge, or converted into an acid addition salt and/or optionally dimerized or polymerized, or conjugated.

[0107] The TCR, polypeptide, and/or protein of the invention can be obtained by methods known in the art such as, for example, *de novo* synthesis. Also, polypeptides and proteins can be recombinantly produced using the nucleic acids described herein using standard recombinant methods. See, for instance, Green and Sambrook, Molecular Cloning: A Laboratory Manual, 4th ed., Cold Spring Harbor Press, Cold Spring Harbor, NY (2012). Alternatively, the TCRs, polypeptides, and/or proteins described herein can be commercially

synthesized by companies, such as Synpep (Dublin, CA), Peptide Technologies Corp. (Gaithersburg, MD), and Multiple Peptide Systems (San Diego, CA). In this respect, the inventive TCRs, polypeptides, and proteins can be synthetic, recombinant, isolated, and/or purified.

[0108] An embodiment of the invention provides a nucleic acid comprising a nucleotide sequence encoding any of the TCRs, polypeptides, or proteins described herein. "Nucleic acid," as used herein, includes "polynucleotide," "oligonucleotide," and "nucleic acid molecule," and generally means a polymer of DNA or RNA, which can be single-stranded or double-stranded, which can contain natural, non-natural or altered nucleotides, and which can contain a natural, non-natural or altered internucleotide linkage, such as a phosphoroamidate linkage or a phosphorothioate linkage, instead of the phosphodiester found between the nucleotides of an unmodified oligonucleotide. In an embodiment, the nucleic acid comprises complementary DNA (cDNA). It is generally preferred that the nucleic acid does not comprise any insertions, deletions, inversions, and/or substitutions. However, it may be suitable in some instances, as discussed herein, for the nucleic acid to comprise one or more insertions, deletions, inversions, and/or substitutions.

[0109] Preferably, the nucleic acids of the invention are recombinant. As used herein, the term "recombinant" refers to (i) molecules that are constructed outside living cells by joining natural or synthetic nucleic acid segments to nucleic acid molecules that can replicate in a living cell, or (ii) molecules that result from the replication of those described in (i) above. For purposes herein, the replication can be in vitro replication or in vivo replication.

[0110] The nucleic acids can be constructed based on chemical synthesis and/or enzymatic ligation reactions using procedures known in the art. See, for example, Green and Sambrook et al., *supra*. For example, a nucleic acid can be chemically synthesized using naturally occurring nucleotides or variously modified nucleotides designed to increase the biological stability of the molecules or to increase the physical stability of the duplex formed upon hybridization (e.g., phosphorothioate derivatives and acridine substituted nucleotides). Examples of modified nucleotides that can be used to generate the nucleic acids include, but are not limited to, 5-fluorouracil, 5-bromouracil, 5-chlorouracil, 5-iodouracil, hypoxanthine, xanthine, 4-acetylcytosine, 5-(carboxyhydroxymethyl) uracil, 5-carboxymethylaminomethyl-2-thiouridine, 5-carboxymethylaminomethyluracil, dihydrouracil, beta-D-galactosylqueosine, inosine, N⁶-isopentenyladenine, 1-methylguanine, 1-methylinosine, 2,2-dimethylguanine, 2-methyladenine, 2-methylguanine, 3-methylcytosine, 5-methylcytosine, N⁶-substituted

adenine, 7-methylguanine, 5-methylaminomethyluracil, 5-methoxyaminomethyl-2-thiouracil, beta-D-mannosylqueosine, 5'-methoxycarboxymethyluracil, 5-methoxyuracil, 2-methylthio-N⁶-isopentenyladenine, uracil-5-oxyacetic acid (v), wybutoxosine, pseudouracil, queosine, 2-thiocytosine, 5-methyl-2-thiouracil, 2-thiouracil, 4-thiouracil, 5-methyluracil, uracil-5-oxyacetic acid methylester, 3-(3-amino-3-N-2-carboxypropyl) uracil, and 2,6-diaminopurine. Alternatively, one or more of the nucleic acids of the invention can be purchased from companies, such as Macromolecular Resources (Fort Collins, CO) and Synthege (Houston, TX).

[0111] In an embodiment of the invention, the nucleic acid comprises a codon-optimized nucleotide sequence encoding any of the TCRs, polypeptides, or proteins described herein. Without being bound to any particular theory or mechanism, it is believed that codon optimization of the nucleotide sequence increases the translation efficiency of the mRNA transcripts. Codon optimization of the nucleotide sequence may involve substituting a native codon for another codon that encodes the same amino acid, but can be translated by tRNA that is more readily available within a cell, thus increasing translation efficiency. Optimization of the nucleotide sequence may also reduce secondary mRNA structures that would interfere with translation, thus increasing translation efficiency.

[0112] The invention also provides a nucleic acid comprising a nucleotide sequence which is complementary to the nucleotide sequence of any of the nucleic acids described herein or a nucleotide sequence which hybridizes under stringent conditions to the nucleotide sequence of any of the nucleic acids described herein.

[0113] The nucleotide sequence which hybridizes under stringent conditions preferably hybridizes under high stringency conditions. By "high stringency conditions" is meant that the nucleotide sequence specifically hybridizes to a target sequence (the nucleotide sequence of any of the nucleic acids described herein) in an amount that is detectably stronger than non-specific hybridization. High stringency conditions include conditions which would distinguish a polynucleotide with an exact complementary sequence, or one containing only a few scattered mismatches from a random sequence that happened to have a few small regions (e.g., 3-10 bases) that matched the nucleotide sequence. Such small regions of complementarity are more easily melted than a full-length complement of 14-17 or more bases, and high stringency hybridization makes them easily distinguishable. Relatively high stringency conditions would include, for example, low salt and/or high temperature conditions, such as provided by about 0.02-0.1 M NaCl or the equivalent, at temperatures of

about 50-70 °C. Such high stringency conditions tolerate little, if any, mismatch between the nucleotide sequence and the template or target strand, and are particularly suitable for detecting expression of any of the inventive TCRs. It is generally appreciated that conditions can be rendered more stringent by the addition of increasing amounts of formamide.

[0114] The invention also provides a nucleic acid comprising a nucleotide sequence that is at least about 70% or more, e.g., about 80%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, or about 99% identical to any of the nucleic acids described herein. In this regard, the nucleic acid may consist essentially of any of the nucleotide sequences described herein.

[0115] The nucleic acids of the invention can be incorporated into a recombinant expression vector. In this regard, the invention provides a recombinant expression vector comprising any of the nucleic acids of the invention. In an embodiment of the invention, the recombinant expression vector comprises a nucleotide sequence encoding the α chain, the β chain, and linker peptide.

[0116] For purposes herein, the term "recombinant expression vector" means a genetically-modified oligonucleotide or polynucleotide construct that permits the expression of an mRNA, protein, polypeptide, or peptide by a host cell, when the construct comprises a nucleotide sequence encoding the mRNA, protein, polypeptide, or peptide, and the vector is contacted with the cell under conditions sufficient to have the mRNA, protein, polypeptide, or peptide expressed within the cell. The vectors of the invention are not naturally-occurring as a whole. However, parts of the vectors can be naturally-occurring. The inventive recombinant expression vectors can comprise any type of nucleotide, including, but not limited to DNA and RNA, which can be single-stranded or double-stranded, synthesized or obtained in part from natural sources, and which can contain natural, non-natural or altered nucleotides. The recombinant expression vectors can comprise naturally-occurring, non-naturally-occurring internucleotide linkages, or both types of linkages. Preferably, the non-naturally occurring or altered nucleotides or internucleotide linkages do not hinder the transcription or replication of the vector.

[0117] The recombinant expression vector of the invention can be any suitable recombinant expression vector, and can be used to transform or transfect any suitable host cell. Suitable vectors include those designed for propagation and expansion or for expression or both, such as plasmids and viruses. The vector can be selected from the group consisting of the transposon/transposase series, pUC series (Fermentas Life Sciences), the pBluescript

series (Stratagene, LaJolla, CA), the pET series (Novagen, Madison, WI), the pGEX series (Pharmacia Biotech, Uppsala, Sweden), and the pEX series (Clontech, Palo Alto, CA). Bacteriophage vectors, such as λ GT10, λ GT11, λ ZapII (Stratagene), λ EMBL4, and λ NM1149, also can be used. Examples of plant expression vectors include pBI01, pBI101.2, pBI101.3, pBI121 and pBIN19 (Clontech). Examples of animal expression vectors include pEUK-Cl, pMAM and pMAMneo (Clontech). Preferably, the recombinant expression vector is a transposon vector or a viral vector, e.g., a retroviral vector.

[0118] The recombinant expression vectors of the invention can be prepared using standard recombinant DNA techniques described in, for example, Green and Sambrook et al., *supra*. Constructs of expression vectors, which are circular or linear, can be prepared to contain a replication system functional in a prokaryotic or eukaryotic host cell. Replication systems can be derived, e.g., from ColEI, 2 μ plasmid, λ , SV40, bovine papillomavirus, and the like.

[0119] Desirably, the recombinant expression vector comprises regulatory sequences, such as transcription and translation initiation and termination codons, which are specific to the type of host cell (e.g., bacterium, fungus, plant, or animal) into which the vector is to be introduced, as appropriate and taking into consideration whether the vector is DNA- or RNA-based.

[0120] The recombinant expression vector can include one or more marker genes, which allow for selection of transformed or transfected host cells. Marker genes include biocide resistance, e.g., resistance to antibiotics, heavy metals, etc., complementation in an auxotrophic host cell to provide prototrophy, and the like. Suitable marker genes for the inventive expression vectors include, for instance, neomycin/G418 resistance genes, hygromycin resistance genes, histidinol resistance genes, tetracycline resistance genes, and ampicillin resistance genes.

[0121] The recombinant expression vector can comprise a native or nonnative promoter operably linked to the nucleotide sequence encoding the TCR, polypeptide, or protein, or to the nucleotide sequence which is complementary to or which hybridizes to the nucleotide sequence encoding the TCR, polypeptide, or protein. The selection of promoters, e.g., strong, weak, inducible, tissue-specific and developmental-specific, is within the ordinary skill of the artisan. Similarly, the combining of a nucleotide sequence with a promoter is also within the skill of the artisan. The promoter can be a non-viral promoter, e.g., a human elongation factor-1 α promoter, or a viral promoter, e.g., a cytomegalovirus (CMV) promoter, an SV40

promoter, an RSV promoter, and a promoter found in the long-terminal repeat of the murine stem cell virus.

[0122] The inventive recombinant expression vectors can be designed for either transient expression, for stable expression, or for both. Also, the recombinant expression vectors can be made for constitutive expression or for inducible expression.

[0123] Further, the recombinant expression vectors can be made to include a suicide gene. As used herein, the term "suicide gene" refers to a gene that causes the cell expressing the suicide gene to die. The suicide gene can be a gene that confers sensitivity to an agent, e.g., a drug, upon the cell in which the gene is expressed, and causes the cell to die when the cell is contacted with or exposed to the agent. Suicide genes are known in the art and include, for example, the Herpes Simplex Virus (HSV) thymidine kinase (TK) gene, cytosine deaminase, purine nucleoside phosphorylase, and nitroreductase.

[0124] Another embodiment of the invention further provides a host cell comprising any of the recombinant expression vectors described herein. As used herein, the term "host cell" refers to any type of cell that can contain the inventive recombinant expression vector. The host cell can be a eukaryotic cell, e.g., plant, animal, fungi, or algae, or can be a prokaryotic cell, e.g., bacteria or protozoa. The host cell can be a cultured cell or a primary cell, i.e., isolated directly from an organism, e.g., a human. The host cell can be an adherent cell or a suspended cell, i.e., a cell that grows in suspension. Suitable host cells are known in the art and include, for instance, DH5 α *E. coli* cells, Chinese hamster ovarian cells, monkey VERO cells, COS cells, HEK293 cells, and the like. For purposes of amplifying or replicating the recombinant expression vector, the host cell is preferably a prokaryotic cell, e.g., a DH5 α cell. For purposes of producing a recombinant TCR, polypeptide, or protein, the host cell is preferably a mammalian cell. Most preferably, the host cell is a human cell. While the host cell can be of any cell type, can originate from any type of tissue, and can be of any developmental stage, the host cell preferably is a peripheral blood lymphocyte (PBL) or a peripheral blood mononuclear cell (PBMC). More preferably, the host cell is a T cell.

[0125] For purposes herein, the T cell can be any T cell, such as a cultured T cell, e.g., a primary T cell, or a T cell from a cultured T cell line, e.g., Jurkat, SupT1, etc., or a T cell obtained from a mammal. If obtained from a mammal, the T cell can be obtained from numerous sources, including but not limited to blood, bone marrow, lymph node, the thymus, or other tissues or fluids. T cells can also be enriched for or purified. Preferably, the T cell is a human T cell. The T cell can be any type of T cell and can be of any developmental stage,

including but not limited to, CD4⁺/CD8⁺ double positive T cells, CD4⁺ helper T cells, e.g., Th₁ and Th₂ cells, CD4⁺ T cells, CD8⁺ T cells (e.g., cytotoxic T cells), tumor infiltrating lymphocytes (TILs), memory T cells (e.g., central memory T cells and effector memory T cells), naïve T cells, and the like.

[0126] Also provided by the invention is a population of cells comprising at least one host cell described herein. The population of cells can be a heterogeneous population comprising the host cell comprising any of the recombinant expression vectors described, in addition to at least one other cell, e.g., a host cell (e.g., a T cell), which does not comprise any of the recombinant expression vectors, or a cell other than a T cell, e.g., a B cell, a macrophage, a neutrophil, an erythrocyte, a hepatocyte, an endothelial cell, an epithelial cells, a muscle cell, a brain cell, etc. Alternatively, the population of cells can be a substantially homogeneous population, in which the population comprises mainly of host cells (e.g., consisting essentially of) comprising the recombinant expression vector. The population also can be a clonal population of cells, in which all cells of the population are clones of a single host cell comprising a recombinant expression vector, such that all cells of the population comprise the recombinant expression vector. In one embodiment of the invention, the population of cells is a clonal population comprising host cells comprising a recombinant expression vector as described herein.

[0127] In an embodiment of the invention, the numbers of cells in the population may be rapidly expanded. Expansion of the numbers of T cells can be accomplished by any of a number of methods as are known in the art as described in, for example, U.S. Patent 8,034,334; U.S. Patent 8,383,099; U.S. Patent Application Publication No. 2012/0244133; Dudley et al., *J. Immunother.*, 26:332-42 (2003); and Riddell et al., *J. Immunol. Methods*, 128:189-201 (1990). In an embodiment, expansion of the numbers of T cells is carried out by culturing the T cells with OKT3 antibody, IL-2, and feeder PBMC (e.g., irradiated allogeneic PBMC).

[0128] The inventive TCRs, polypeptides, proteins, nucleic acids, recombinant expression vectors, and host cells (including populations thereof), can be isolated and/or purified. The term "isolated" as used herein means having been removed from its natural environment. The term "purified" as used herein means having been increased in purity, wherein "purity" is a relative term, and not to be necessarily construed as absolute purity. For example, the purity can be at least about 50%, at least about 60%, at least about 70%, at least about 80%, at least about 90%, at least about 95%, or can be about 100%.

[0129] The inventive TCRs, polypeptides, proteins, nucleic acids, recombinant expression vectors, and host cells (including populations thereof), all of which are collectively referred to as "inventive TCR materials" hereinafter, can be formulated into a composition, such as a pharmaceutical composition. In this regard, the invention provides a pharmaceutical composition comprising any of the TCRs, polypeptides, proteins, nucleic acids, expression vectors, and host cells (including populations thereof), described herein, and a pharmaceutically acceptable carrier. The inventive pharmaceutical compositions containing any of the inventive TCR materials can comprise more than one inventive TCR material, e.g., a polypeptide and a nucleic acid, or two or more different TCRs. Alternatively, the pharmaceutical composition can comprise an inventive TCR material in combination with another pharmaceutically active agent(s) or drug(s), such as a chemotherapeutic agents, e.g., asparaginase, busulfan, carboplatin, cisplatin, daunorubicin, doxorubicin, fluorouracil, gemcitabine, hydroxyurea, methotrexate, paclitaxel, rituximab, vinblastine, vincristine, etc.

[0130] Preferably, the carrier is a pharmaceutically acceptable carrier. With respect to pharmaceutical compositions, the carrier can be any of those conventionally used for the particular inventive TCR material under consideration. Methods for preparing administrable compositions are known or apparent to those skilled in the art and are described in more detail in, for example, *Remington: The Science and Practice of Pharmacy*, 22nd Ed., Pharmaceutical Press (2012). It is preferred that the pharmaceutically acceptable carrier be one which has no detrimental side effects or toxicity under the conditions of use.

[0131] The choice of carrier will be determined in part by the particular inventive TCR material, as well as by the particular method used to administer the inventive TCR material. Accordingly, there are a variety of suitable formulations of the pharmaceutical composition of the invention. Suitable formulations may include any of those for parenteral, subcutaneous, intravenous, intramuscular, intraarterial, intrathecal, intratumoral, or interperitoneal administration. More than one route can be used to administer the inventive TCR materials, and in certain instances, a particular route can provide a more immediate and more effective response than another route.

[0132] Preferably, the inventive TCR material is administered by injection, e.g., intravenously. When the inventive TCR material is a host cell expressing the inventive TCR, the pharmaceutically acceptable carrier for the cells for injection may include any isotonic carrier such as, for example, normal saline (about 0.90% w/v of NaCl in water, about 300 mOsm/L NaCl in water, or about 9.0 g NaCl per liter of water), NORMOSOL R electrolyte

solution (Abbott, Chicago, IL), PLASMA-LYTE A (Baxter, Deerfield, IL), about 5% dextrose in water, or Ringer's lactate. In an embodiment, the pharmaceutically acceptable carrier is supplemented with human serum albumen.

[0133] For purposes of the invention, the amount or dose (e.g., numbers of cells when the inventive TCR material is one or more cells) of the inventive TCR material administered should be sufficient to effect, e.g., a therapeutic or prophylactic response, in the subject or animal over a reasonable time frame. For example, the dose of the inventive TCR material should be sufficient to bind to a cancer antigen (e.g., mutated p53), or detect, treat or prevent cancer in a period of from about 2 hours or longer, e.g., 12 to 24 or more hours, from the time of administration. In certain embodiments, the time period could be even longer. The dose will be determined by the efficacy of the particular inventive TCR material and the condition of the animal (e.g., human), as well as the body weight of the animal (e.g., human) to be treated.

[0134] Many assays for determining an administered dose are known in the art. For purposes of the invention, an assay, which comprises comparing the extent to which target cells are lysed or IFN- γ is secreted by T cells expressing the inventive TCR, polypeptide, or protein upon administration of a given dose of such T cells to a mammal among a set of mammals of which each is given a different dose of the T cells, could be used to determine a starting dose to be administered to a mammal. The extent to which target cells are lysed or IFN- γ is secreted upon administration of a certain dose can be assayed by methods known in the art.

[0135] The dose of the inventive TCR material also will be determined by the existence, nature and extent of any adverse side effects that might accompany the administration of a particular inventive TCR material. Typically, the attending physician will decide the dosage of the inventive TCR material with which to treat each individual patient, taking into consideration a variety of factors, such as age, body weight, general health, diet, sex, inventive TCR material to be administered, route of administration, and the severity of the cancer being treated. In an embodiment in which the inventive TCR material is a population of cells, the number of cells administered per infusion may vary, e.g., from about 1×10^6 to about 1×10^{12} cells or more. In certain embodiments, fewer than 1×10^6 cells may be administered.

[0136] One of ordinary skill in the art will readily appreciate that the inventive TCR materials of the invention can be modified in any number of ways, such that the therapeutic

or prophylactic efficacy of the inventive TCR materials is increased through the modification. For instance, the inventive TCR materials can be conjugated either directly or indirectly through a bridge to a chemotherapeutic agent. The practice of conjugating compounds to a chemotherapeutic agent is known in the art. One of ordinary skill in the art recognizes that sites on the inventive TCR materials, which are not necessary for the function of the inventive TCR materials, are ideal sites for attaching a bridge and/or a chemotherapeutic agent, provided that the bridge and/or chemotherapeutic agent, once attached to the inventive TCR materials, do(es) not interfere with the function of the inventive TCR materials, i.e., the ability to bind to mutated p53 or to detect, treat, or prevent cancer.

[0137] It is contemplated that the inventive pharmaceutical compositions, TCRs, polypeptides, proteins, nucleic acids, recombinant expression vectors, host cells, or populations of cells can be used in methods of treating or preventing cancer. Without being bound to a particular theory, the inventive TCRs are believed to bind specifically to mutated p53, such that the TCR (or related inventive polypeptide or protein), when expressed by a cell, is able to mediate an immune response against a target cell expressing mutated p53. In this regard, an embodiment of the invention provides a method of treating or preventing cancer in a mammal, comprising administering to the mammal any of the pharmaceutical compositions, TCRs, polypeptides, or proteins described herein, any nucleic acid or recombinant expression vector comprising a nucleotide sequence encoding any of the TCRs, polypeptides, proteins described herein, or any host cell or population of cells comprising a recombinant vector which encodes any of the TCRs, polypeptides, or proteins described herein, in an amount effective to treat or prevent cancer in the mammal.

[0138] An embodiment of the invention provides any of the pharmaceutical compositions, TCRs, polypeptides, or proteins described herein, any nucleic acid or recombinant expression vector comprising a nucleotide sequence encoding any of the TCRs, polypeptides, proteins described herein, or any host cell or population of cells comprising a recombinant vector which encodes any of the TCRs, polypeptides, or proteins described herein, for use in the treatment or prevention of cancer in a mammal.

[0139] The terms "treat," and "prevent" as well as words stemming therefrom, as used herein, do not necessarily imply 100% or complete treatment or prevention. Rather, there are varying degrees of treatment or prevention of which one of ordinary skill in the art recognizes as having a potential benefit or therapeutic effect. In this respect, the inventive methods can provide any amount of any level of treatment or prevention of cancer in a mammal.

Furthermore, the treatment or prevention provided by the inventive method can include treatment or prevention of one or more conditions or symptoms of the cancer being treated or prevented. For example, treatment or prevention can include promoting the regression of a tumor. Also, for purposes herein, "prevention" can encompass delaying the onset of the cancer, or a symptom or condition thereof. Alternatively or additionally, "prevention" may encompass preventing or delaying the recurrence of cancer, or a symptom or condition thereof.

[0140] Also provided by an embodiment of the invention is a method of detecting the presence of cancer in a mammal. The method comprises (i) contacting a sample comprising one or more cells from the mammal with any of the inventive TCRs, polypeptides, proteins, nucleic acids, recombinant expression vectors, host cells, populations of cells, or pharmaceutical compositions described herein, thereby forming a complex, and detecting the complex, wherein detection of the complex is indicative of the presence of cancer in the mammal.

[0141] With respect to the inventive method of detecting cancer in a mammal, the sample of cells can be a sample comprising whole cells, lysates thereof, or a fraction of the whole cell lysates, e.g., a nuclear or cytoplasmic fraction, a whole protein fraction, or a nucleic acid fraction.

[0142] For purposes of the inventive detecting method, the contacting can take place *in vitro* or *in vivo* with respect to the mammal. Preferably, the contacting is *in vitro*.

[0143] Also, detection of the complex can occur through any number of ways known in the art. For instance, the inventive TCRs, polypeptides, proteins, nucleic acids, recombinant expression vectors, host cells, or populations of cells, described herein, can be labeled with a detectable label such as, for instance, a radioisotope, a fluorophore (e.g., fluorescein isothiocyanate (FITC), phycoerythrin (PE)), an enzyme (e.g., alkaline phosphatase, horseradish peroxidase), and element particles (e.g., gold particles).

[0144] For purposes of the inventive methods, wherein host cells or populations of cells are administered, the cells can be cells that are allogeneic or autologous to the mammal. Preferably, the cells are autologous to the mammal.

[0145] With respect to the inventive methods, the cancer can be any cancer, including any of acute lymphocytic cancer, acute myeloid leukemia, alveolar rhabdomyosarcoma, bone cancer, brain cancer, breast cancer, cancer of the anus, anal canal, or anorectum, cancer of the eye, cancer of the intrahepatic bile duct, cancer of the joints, cancer of the neck, gallbladder,

or pleura, cancer of the nose, nasal cavity, or middle ear, cancer of the oral cavity, cancer of the vagina, cancer of the vulva, chronic lymphocytic leukemia, chronic myeloid cancer, colon cancer, colorectal cancer, endometrial cancer, esophageal cancer, uterine cervical cancer, gastrointestinal carcinoid tumor, glioma, Hodgkin lymphoma, hypopharynx cancer, kidney cancer, larynx cancer, liver cancer, lung cancer, malignant mesothelioma, melanoma, multiple myeloma, nasopharynx cancer, non-Hodgkin lymphoma, cancer of the oropharynx, ovarian cancer, cancer of the penis, pancreatic cancer, peritoneum, omentum, and mesentery cancer, pharynx cancer, prostate cancer, rectal cancer, renal cancer, skin cancer, small intestine cancer, soft tissue cancer, stomach cancer, testicular cancer, thyroid cancer, cancer of the uterus, ureter cancer, and urinary bladder cancer. In a preferred embodiment, the cancer is a cancer which expresses mutated p53. The cancer may express p53 with a mutation at any one or more of positions 175, 220, 245, 248, 249, 273, and 282 of SEQ ID NO: 1. The cancer may express p53 with any one or more of the following human p53 mutations: R175H, Y220C, G245D, G245S, R248L, R248Q, R248W, R249S, R273H, R273C, R273L, and R282W. Preferably, the cancer is an epithelial cancer or cholangiocarcinoma, melanoma, colon cancer, rectal cancer, ovarian cancer, endometrial cancer, non-small cell lung cancer (NSCLC), glioblastoma, uterine cervical cancer, head and neck cancer, breast cancer, pancreatic cancer, or bladder cancer.

[0146] The mammal referred to in the inventive methods can be any mammal. As used herein, the term "mammal" refers to any mammal, including, but not limited to, mammals of the order Rodentia, such as mice and hamsters, and mammals of the order Logomorpha, such as rabbits. It is preferred that the mammals are from the order Carnivora, including Felines (cats) and Canines (dogs). It is more preferred that the mammals are from the order Artiodactyla, including Bovines (cows) and Swines (pigs) or of the order Perssodactyla, including Equines (horses). It is most preferred that the mammals are of the order Primates, Ceboids, or Simoids (monkeys) or of the order Anthropoids (humans and apes). An especially preferred mammal is the human.

[0147] The following examples further illustrate the invention but, of course, should not be construed as in any way limiting its scope.

EXAMPLES

[0148] The amino acid sequences set forth in Tables 1-3 were employed in the experiments described in the following Examples. In Tables 1-2, “LP” stands for “long peptide.” In Table 3, “TMG” stands for “tandem minigene.”

TABLE 1

SEQ ID NO:	Name	Sequence
2	LP-p53-R175H-MUT	YKQSQHMTEVVR <u>H</u> CPHHERCSDSDG
3	LP-p53-R273H-MUT	SGNLLGRNSFEV <u>H</u> VCACPGRRDRTE
4	LP-p53-R248L-MUT	YMCNSSCMGGMN <u>L</u> RPILTIITLEDSS
5	LP-p53-R282W-MUT	FEVRVCACPGRD <u>W</u> RTEEEENLRKKGE
6	LP-p53-R273C-MUT	SGNLLGRNSFEV <u>C</u> VCACPGRRDRTE
7	LP-p53-G245S-MUT	HNYMCNSSCMG <u>S</u> MNRRPILTIITL
8	LP-p53-R248Q-MUT	YMCNSSCMGGMN <u>Q</u> RPILTIITLEDSS
9	LP-p53-G245D-MUT	HNYMCNSSCMG <u>D</u> MNRRPILTIITL
10	LP-p53-R273L-MUT	SGNLLGRNSFEV <u>L</u> VCACPGRRDRTE
11	LP-p53-R248W-MUT	YMCNSSCMGGMN <u>W</u> RPILTIITLEDSS
12	LP-p53-Y220C-MUT	DRNTFRHSVVVP <u>C</u> EPPEVGSDCTTI
13	LP-p53-R249S-MUT	MCNSSCMGGMN <u>R</u> SPIITILEDSS

[0149] The WT versions of the peptides of Table 1 are set forth in Table 2.

TABLE 2

SEQ ID NO:	Name	Sequence
15	LP-p53-R175-WT	YKQSQHMTEVVR <u>R</u> CPHHERCSDSDG
16	LP-p53-R273-WT	SGNLLGRNSFEV <u>R</u> VCACPGRRDRTE
17	LP-p53-R248-WT	YMCNSSCMGGMN <u>R</u> RPILTIITLEDSS
18	LP-p53-R282-WT	FEVRVCACPGRD <u>R</u> RTEEEENLRKKGE
19	LP-p53-G245-WT	HNYMCNSSCMG <u>G</u> MNRRPILTIITL
20	LP-p53-Y220-WT	DRNTFRHSVVVP <u>Y</u> EPPEVGSDCTTI
21	LP-p53-R249-WT	MCNSSCMGGMN <u>R</u> RPIITILEDSS

TABLE 3

SEQ ID NO:	Name	Sequence
14	TMG-p53-MUT	YKQSQHMTVEVVRHCPHHERCSDSDGSGNLLGRNSFEVHVACAPGRDRRTEYMCNSSCMGGMNLRPILTIITLEDSEFEVRVCACPGRDWRTEEEENLRKKGESGNLLGRNSFEVVCACAPGRDRRTEHYNYMCNSSCMGSMNRRPILTIITLYMCNSSCMGGMNQRPIITILEDSDSHYNYMCNSSCMGDMNRRPILTIITLSGNLLGRNSFEVLVCACAPGRDRRTEYMCNSSCMGGMNWRPILTIITLEDSDRNTFRHSVVVPCEPPEVGSDCTTIMCNSSCMGGMNRSPILTIITLEDSS
22	TMG-p53-WT	YKQSQHMTVEVVRRCPPHHERCSDSDGSGNLLGRNSFEVRVCACAPGRDRRTEYMCNSSCMGGMNRPRILTIITLEDSEFEVRVCACPGRDWRTEEEENLRKKGESGNLLGRNSFEVRVCACAPGRDRRTEHYNYMCNSSCMGGMNRRPILTIITLYMCNSSCMGGMNRPRILTIITLEDSDSHYNYMCNSSCMGGMNRRPILTIITLSGNLLGRNSFEVRVCACAPGRDRRTEYMCNSSCMGGMNRPRILTIITLEDSDRNTFRHSVVVPYEPPEVGSDCTTIMCNSSCMGGMNRPRILTIITLEDSS

EXAMPLE 1

[0150] This example demonstrates the isolation and specific reactivity of four anti-mutated p53 TCRs from patient 4127.

[0151] Experiments were carried out as described for Figures 1-7, 36, and 37A-37C for Patient 4127. The mutated p53 reactive T cells for this patient were identified by the method described in U.S. Patent Application Number 2017/0224800 ("Tran method"). The methods for isolating the individual TCRs are set forth below.

[0152] Autologous PBL were transduced with one of the TCRs shown in Figure 1 and were tested 2 weeks later. Autologous DC cells were plated at 3×10^4 cells/well and pulsed overnight with either WT p53-G245 peptide (wt) or mutated p53-G245S peptide (mut) at decreasing peptide concentrations. 3×10^4 transduced T cells were added per well and co-cultured overnight at 37 °C. Supernatants were harvested for IFN- γ ELISA. The IFN- γ ELISA results are shown in Figure 1. 4-1BB expression was measured by FACS gate: lymphocytes\PI(neg)CD3⁺\CD3⁺mTCR⁺\CD8(neg)CD4⁺. The FACS results are shown in Figure 2.

[0153] Autologous PBL were transduced with one of the TCRs shown in Figure 3 and co-cultured as described for the experiment of Figure 2 except that the DCs were pulsed with DMSO (peptide vehicle) or one of the 15-mer p53-G245S peptides with overlapping 14 amino acids shown in Table A. 4-1BB expression was carried out as described for Figure 2. The results are shown in Figure 3.

TABLE A

Peptide no. in X-axis of Fig. 3	Peptide	SEQ ID NO:
1	HNYMCNSSCMG <u>S</u> MN	497
2	YNYMCNSSCMG <u>S</u> MNR	498
3	NYMCNSSCMG <u>S</u> MNRR	499
4	YMCNSSCMG <u>S</u> MNRRP	500
5	MCNSSCMG <u>S</u> MNRRPI	501
6	CNSSCMG <u>S</u> MNRRPIL	502
7	NSSCMG <u>S</u> MNRRPILT	503
8	SSCMG <u>S</u> MNRRPILTI	504
9	SCMG <u>S</u> MNRRPILTII	505
10	CMG <u>S</u> MNRRPILTIIT	506
11	MG <u>S</u> MNRRPILTIITL	507

[0154] Cos7 cells (2.5×10^4 per well) were plated on wells of flat-bottom 96 well plates. After 20 hours, cells were co-transfected with individual HLA alleles. After 20 hours, cells were pulsed with p53G245S-15-mer peptide for 3 hours at 37 °C at 10 µg/mL. After washing, T cells (10^5) were added to wells and co-cultured overnight (for 20 hours) at 37 °C. IFN-γ secretion was measured by ELISA; prediction by NetMHCIIpan: cbs.dtu.dk/services/NetMHCIIpan/. 4-1BB expression was measured by FACS. FACS gate: lymphocytes → live (PI negative) → CD3+ (T cells) → CD4+ (4127-TIL) or CD4+mTCR+ (TCR transduced T cells). The results are shown in Figures 4-5 and Table B.

TABLE B

HLA	Peptide	Affinity, nM	Rank
DRB3*02:02	HNYMCNSSCMG <u>S</u> MN SEQ ID NO: 497	533.4	21

[0155] DRB3*02 is expressed by 1367 of 3719 (37%) of DRB_ typed patients in the NCI HLA database and 5 of 9 (56%) endometrial and ovarian cancer patients at NCI-SB (National Cancer Institute Surgery Branch). The reported frequency of the DRB3*02 allele is very high according to the allelefrequencies.net website. For example, this website reports that the frequency of the DRB3*02 allele is 0.3447 of the USA NMDP Middle Eastern or North Coast of Africa population.

[0156] TIL from patient 4127 were co-cultured with allogeneic (DRB3*01:01:01 or DRB3*02:02:01) antigen presenting cells (APCs) which were (1) electroporated with tandem minigenes (TMG) composed of irrelevant, WT p53, or mutated p53 sequence or (2) pulsed with peptide vehicle (DMSO) or purified (>95% by high-performance liquid chromatography (HPLC)) 25-amino acid peptides composed of WT p53-G245 sequence or mutated p53-G245S sequence. Co-cultures were performed overnight at 37 °C. Secretion of IFN- γ was evaluated using ELISPOT assay. The results are shown in Figure 6.

[0157] T cells expressing the 4127-TCR1 specific to p53-G245S were co-cultured with allogeneic APCs as described for the experiment of Figure 6. The results are shown in Figure 7.

[0158] Autologous APCs were pulsed with decreasing concentrations of 25-amino acid peptides corresponding to the WT p53-G245 or mutated p53-G245S sequence. T cells transduced with the 4127-O37-TCR from patient 4127 were co-cultured overnight at 37 °C with peptide-pulsed APCs. Expression of 4-1BB was assayed by flow cytometry after gating lymphocytes $\rightarrow$ single cells $\rightarrow$ live $\rightarrow$ CD3+mTCR+. The results are shown in Figure 36.

[0159] Cos7 cells (2.5×10^4 per well) were plated on wells of flat-bottom 96 well plates. The following day, cells were co-transfected with individual HLA alleles from patient 4127. The next day, the 25-amino acid p53-G245S peptide was pulsed on transfected Cos7 cells. Excess peptide was washed away. T cells were transduced with one of the TCRs from Patient 4127. The transduced T cells were added (2×10^4 cells/well) to the co-culture with the Cos7 cells. Co-cultures were incubated overnight at 37 °C. Secretion of IFN- γ was evaluated by ELISA. Expression of 4-1BB was assayed by flow cytometry after gating lymphocytes $\rightarrow$ single cells $\rightarrow$ live $\rightarrow$ CD3+mTCR+. The results are shown in Figures 37A-37C.

[0160] The sequence of TCR **4127-TP53-G245S-TCR1**, which was isolated from Patient 4127, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1beta (SEQ ID NO: 30), the second underlined region is the CDR2beta (SEQ ID NO: 31), the third underlined region is the CDR3beta (SEQ ID NO: 32), the fourth underlined region is the CDR1alpha (SEQ ID NO: 27), the fifth underlined region is the CDR2alpha (SEQ ID NO: 28), and the sixth underlined region is the CDR3alpha (SEQ ID NO: 29). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the beta chain constant region (SEQ ID NO: 25) and the second italicized region is the alpha chain constant region (SEQ ID NO: 23). The beta chain variable region (SEQ ID NO: 34) includes the sequence starting from the amino terminus and ending

immediately prior to the start of the beta chain constant region. The alpha chain variable region (SEQ ID NO: 33) includes the sequence starting immediately after the linker and ending immediately prior to the start of the alpha chain constant region. The full-length beta chain (SEQ ID NO: 36) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length alpha chain (SEQ ID NO: 35) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0161] Cancer reactive T cells were identified as described below. The TCR was isolated as described below.

TCR name: 4127-TP53-G245S-TCR1

Recognition of p53 mutation: G245S

Method: Tran method

Co-culture to identify TCR: Co-culture 4127-F10 TIL fragment with G245S long peptide, sorted CD4+41BB+ T cells (single cell RT-PCR only)

Method to identify TCR: single-cell RT-PCR (below), Adaptive Pairseq (4127-F10 fragment), 5'RACE from TIL clone O71

Abundance of TCR amongst all paired TCRs: 31.3% (observed 5 times of 16 pairs)

TCR orientation: beta-alpha

Expression vector: gamma-retrovirus

MATRLLCWAALCLLGAELTEAGVAQSPRYKIIKRQSVAFWCNPISGHATLYWYQQ
ILGQGPKLLIQFQNNGVVDDSQLPKDRFSAERLKGVDSTLKIQPAKLEDSAVVYLCASS
LVNTEAFFGQGTRLTVVEDLRNVTTPPKVSLFEPKAEIANKQKATLVCLARGFFPDHVEL
SWWVNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNPRNHFRCQVQFHGLSEE
DKWPEGSPKPVTONISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLYAVLVSTLVV
MAMVKRKNSRAKRSGSGATNFSLLKQAGDVEENPGPMLLLLIPVLGMIFALRDAR
AQSVSQHNHHVILSEAASLELGCNYSYGGTVNLFWYVQYPGQHLQLLLKYFSGDPL
VKGIGKFEEAFIKSKFSFNLKPSVQWSDTAEYFCAVKGDYKLSFGAGTTVTVRANI
QNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTMESGTFITDKCVLDMKAMDSKNGA
IAWSNQTSFTCQDIFKETNATYPSSDVPCDATLTEKSFETDMNLNFQNLVIVLRILLKVA
GFNLLMTLRLWSS (SEQ ID NO: 543)

[0162] The statistics for TCR **4127-TP53-G245S-TCR1** for Patient 4127 are set forth in Table 4 below.

Table 4

Parameter	#	Frequency
Total wells	96	100%
CDR3alpha	6	6.3%
CDR3beta	13	13.5%
4127-TP53-G245S-TCR1 pairs	5	5.2%

Parameter	#	Frequency
Total paired TCRs	16	16.7%

[0163] The sequence of TCR **4127-TP53-G245S-TCR4**, which was isolated from Patient 4127, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1beta (SEQ ID NO: 40), the second underlined region is the CDR2beta (SEQ ID NO: 41), the third underlined region is the CDR3beta (SEQ ID NO: 42), the fourth underlined region is the CDR1alpha (SEQ ID NO: 37), the fifth underlined region is the CDR2alpha (SEQ ID NO: 38), and the sixth underlined region is the CDR3alpha (SEQ ID NO: 39). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the beta chain constant region (SEQ ID NO: 25) and the second italicized region is the alpha chain constant region (SEQ ID NO: 23). The beta chain variable region (SEQ ID NO: 44) includes the sequence starting from the amino terminus and ending immediately prior to the start of the beta chain constant region. The alpha chain variable region (SEQ ID NO: 43) includes the sequence starting immediately after the linker and ending immediately prior to the start of the alpha chain constant region. The full-length beta chain (SEQ ID NO: 46) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length alpha chain (SEQ ID NO: 45) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0164] Cancer reactive T cells were identified as described below. The TCR was isolated as described below.

TCR name: 4127-TP53-G245S-TCR4

Recognition of p53 mutation: G245S

Method: Tran method

Co-culture to identify TCR: Co-culture 4127-F10 TIL fragment with G245S long peptide, sorted CD4+41BB+ T cells (single cell RT-PCR only)

Method to identify TCR: single-cell RT-PCR (below) and Adaptive Pairseq (4127-F10 fragment)

Abundance of TCR amongst all paired TCRs: 12.5% (observed 2 times of 16 pairs)

TCR orientation: beta-alpha

Expression vector: gamma-retrovirus

MAPGLLHWMALCCLLTGHGDAMVIQNPRYQVTQFGKPVTLSQSLNHNVMYWY
 QKSSQAPKLLFHY~~YDKDEN~~NEADTPDNFQSRPNTSFCFLDIRSPGLGDAAMYLC~~A~~
~~TSRELRGNEQFF~~GPGR~~LT~~VL~~ED~~LRNVTPPKVSLFEPKAEIANKQKATLVCLARGFFPD
 HVELSWWVNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNPRNHFRCQVQFHG
 LSEEDKWPEGSPKPVTQNISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLYAVLVST

*L*VVMAMVKKNS**RAK**RS**G**S**G**ATN**F**S**L**L**K**Q**A**G**D**VEEN**P**G**P**MLTASLLRAVIASICVV
SSMAQKV**T**QAQTEISVVEKEDVTLDCVYET**R**DTTYYLFWYKQPPSGELVFLIR**R**NS**F**
DEQNEISGRYSWN**F**QKSTSSFNFTITASQVVDSAVY**F**CAL**S**EGGSNYKL**T**FGKG**T**LLT
VNPNIQ**N**PEPAVYQLKDPRS**Q**DSLCLFTDFDSQINVPK**T**MESGTFITDKCVLDMKAMDS
K**S**NGAIAWSN**Q**TSFT**C**QDIFKETNATYPSSDVPCD**A**T**L**TEKS**F**ETDMNLN**F**Q**N**LLVIVLRIL
LLKVAGFNLLMTLRLWSS (SEQ ID NO: 544)

[0165] The statistics for TCR **4127-TP53-G245S-TCR4** for Patient 4127 are set forth in Table 5 below.

Table 5

Parameter	#	Frequency
Total wells	96	100%
CDR3alpha	2	2.1%
CDR3beta	39	40.6%
4127-TP53-G245S-TCR4 pairs	2	2.1%
Total paired TCRs	16	16.7%

[0166] The sequence of TCR **4127_O102_TCR**, which was isolated from Patient 4127, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1beta (SEQ ID NO: 50), the second underlined region is the CDR2beta (SEQ ID NO: 51), the third underlined region is the CDR3beta (SEQ ID NO: 52), the fourth underlined region is the CDR1alpha (SEQ ID NO: 47), the fifth underlined region is the CDR2alpha (SEQ ID NO: 48), and the sixth underlined region is the CDR3alpha (SEQ ID NO: 49). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the beta chain constant region (SEQ ID NO: 25) and the second italicized region is the alpha chain constant region (SEQ ID NO: 23). The beta chain variable region (SEQ ID NO: 54) includes the sequence starting from the amino terminus and ending immediately prior to the start of the beta chain constant region. The alpha chain variable region (SEQ ID NO: 53) includes the sequence starting immediately after the linker and ending immediately prior to the start of the alpha chain constant region. The full-length beta chain (SEQ ID NO: 56) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length alpha chain (SEQ ID NO: 55) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0167] Cancer reactive T cells were identified as described below. The TCR was isolated as described below.

TCR name: 4127_O102_TCR

Recognition of p53 mutation: G245S

Method: Tran method

Co-culture to identify TCR: Not performed. TCR was directly identified from RNA of T cell clone (O102) using 5'RACE. Clone was developed by sorting for OX40+ T cells and establishing T cell clones by limiting dilution. T cell clones were screened using the Tran method.

Method to identify TCR: 5'RACE from TIL clone O102

Abundance of TCR amongst all paired TCRs: 100% of T cell clone (not applicable)

TCR orientation: beta-alpha

Expression vector: gamma-retrovirus

MAMSIGLLCCAALSLLWAGPVNAGVTQTPKFQVLKTGQSMTLQCAQDMNHEYMS
WYRQDPGMGLRLIHYSVGAGITDQGEVPGYNVSRSTTEDFPLRLLSAAPSQTSVYF
CASSYRESHYGYTFGSGTRLTVVEDLRNVTPPKVSLFEPSCAEIANKQKATLVCLARGFF
PDHVELSWVNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNP RNHFRCQVQF
HGLSEEDKWPEGSPKPVTONISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLYAVL
VSTLVVMAMVKRKNSRAKRSGSGATNFSLLKQAGDVEENPGPMWGVFLLYVSMK
MGGTTGQNIDQPTMTATEGAIVQINCTYQTSGFNGLFWYQQHAGEAPTFLSYNVL
DGLEEKGRFSSFLSRSGYSYLLKELQMKDSASYLCAVKWTGGFKTIFGAGTRLFV
KANIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTMESGTFITDKCVLDMKAMDSK
SNGALAWSNQTSFTCQDIFKETNATYPSSDVPCDATLTEKSFETDMNLFQNLVIVLRILL
LKVAGFNLLMTLRLWSS (SEQ ID NO: 545)

[0168] The sequence of TCR **4127-O37-TCR**, which was isolated from Patient 4127, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1beta (SEQ ID NO: 460), the second underlined region is the CDR2beta (SEQ ID NO: 461), the third underlined region is the CDR3beta (SEQ ID NO: 462), the fourth underlined region is the CDR1alpha (SEQ ID NO: 457), the fifth underlined region is the CDR2alpha (SEQ ID NO: 458), and the sixth underlined region is the CDR3alpha (SEQ ID NO: 459).

[0169] The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the beta chain constant region (SEQ ID NO: 25) and the second italicized region is the alpha chain constant region (SEQ ID NO: 23). The beta chain variable region (SEQ ID NO: 464) includes the sequence starting from the amino terminus and ending immediately prior to the start of the beta chain constant region. The alpha chain variable region (SEQ ID NO: 463) includes the sequence starting immediately after the linker and ending immediately prior to the start of the alpha chain constant region. The full-length beta chain (SEQ ID NO: 466) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length alpha chain (SEQ ID NO: 465)

includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0170] Cancer reactive T cells were identified as described below. The TCR was isolated as described below.

TCR name: 4127-O37-TCR

Recognition of p53 mutation: G245S

Method: Tran method

Co-culture to identify TCR: Not performed. TCR was directly identified from genomic DNA of T cell clone (O37) using Adaptive TCRAD and TCRB surveys (next-generation sequencing (NGS) platform). The O37 clone was developed by sorting for OX40+ T cells and establishing T cell clones by limiting dilution. T cell clones were screened using the Tran method.

Method to identify TCR: Adaptive TCR sequencing (NGS) of TIL clone O37

Abundance of TCR amongst all paired TCRs: 100% of T cell clone (n/a)

TCR orientation: beta-alpha

Expression vector: gamma-retrovirus

MALLLLLLGPGISLLLPGLAGSGLGAVVSQHPSWVICKSGTSVKIECRSLDFQATTM
FWYRQFPKQSLMLMATSNESKATYEQGVEKDKFLINHASLTSTLTVTSAHPEDSS
FYICSAAGQANTEAFFGQGTRLTVVEDLRNVTPPKVSLFEPKAEIANKQKATLVCLARG
FFPDHVELSWVNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNPRNHFRCQV
QFHGLSEEDKWPEGSPKPVTQNISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLYA
VLVSTLVMMAMVKKRNSRAKRS~~SG~~GATNFSLLKQAGDVEENPGPMHSLRVLLVILW
LQLSWVWSQQKEVEQNSGPLSVPEGAIASLNCTYS~~DR~~GSQSFFWYRQYSGKSPELIM
FIYSNGDKEDGRFTAQLNKASQYVSLIRDSQPSDSATYLC~~AV~~NDAGNMLTFGGGTR
LMVKPNIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTMESGTFITDKCVLDMKA
MDSKSNGAIAWSNQTSFTCQDIFKETNATYPSSDVPCDATLTEKSFETDMNLNFQNLLVIV
LRLLLLKVAGFNLLMTLRLWSS (SEQ ID NO: 546)

EXAMPLE 2

[0171] This example demonstrates the isolation and specific reactivity of three anti-mutated p53 TCRs from patient 4196.

[0172] Experiments were carried out as described for Figures 8-11 for Patient 4196. The p53 reactive T cells for this patient were identified by the Tran method, as described in U.S. Patent Application Number 2017/0224800. The Fluidigm method was used for isolating the individual TCRs. The statistics for Patient 4196 are shown in Table D.

[0173] Characterization of p53-reactive cells: challenges with identifying the minimal epitope: The first predicted peptide with mutated amino acid p53 R175H is shown in Table C.

TABLE C

HLA	HLA-A0201
Peptide	HMTEVVRHC (SEQ ID NO: 530)
Core	HMTEVVRHC (SEQ ID NO: 530)
Affinity (nM)	7826.06
% rank	13.00

[0174] TMG1 versus 4196-Rx1 TIL: Cos7 cells were transfected with plasmids for TMG1 and HLA allele. Cells were co-cultured with 4196 Rx1 TIL infusion bag for 20 hours (h). The results are shown in Figure 8A.

[0175] P53-R175H minimal peptide versus 4196-Rx1 TIL: Day 4 autologous DCs were pulsed with either candidate minimal epitope (HMTEVVRHC (SEQ ID NO: 530) or (SQHMTEVVRH (SEQ ID NO: 531)) for 2h. Cells were washed and co-cultured with 4196 Rx1 TIL infusion bag for 20 h. The results are shown in Figure 8B.

[0176] A*02:01-restricted TP53 tetramer can be used to isolate TP53-reactive cells from the TIL infusion bag: Isolation of TP53-reactive cells using 4-1BB+ sort following co-culture with TP53 peptide was technically limited. To isolate TP53-reactive cells, an A*02:01-TP53 tetramer was generated. An irrelevant tetramer (A*02:01-gp100) was generated as a control. A FACS analysis was carried out gated on live, CD3+, CD8+ cells. For the A*02:01-TP53 tetramer, 14.9% p53 tetramer+ cells were detected. For the A*02:01-gp100 tetramer, 0.7% p53 tetramer+ cells were detected.

[0177] Isolation and characterization of TP53-reactive cells: The strategy for the isolation and characterization of TP53-reactive cells was as follows: (1) sort for tetramer-positive cells using FACS as described above. (2) Carry out single cell PCR. (3) Carry out V β sequencing.

[0178] Several candidate clones were identified for the TP53 TCR using single cell PCR, as shown in Table D. TCR-1a and TCR-1b were alternative candidate clones.

TABLE D

	Candidate TCR	CDR3 (amino acids)	
TCR-1a	TRBV6-1	CASSEGLWQVGDEQYF (SEQ ID NO: 72)	63.3% of 120 pairs
	TRAV12-1	CVVQPGGYQKVTF	

	Candidate TCR	CDR3 (amino acids)	
		(SEQ ID NO: 69)	
TCR-1b	TRBV6-1	Not functional	
	TRAV2		
TCR-2	TRBV11-2	CASSLDPGDTGELFF (SEQ ID NO: 92)	25.8% of 120 pairs
	TRAV6	CALDIYPHDMRF (SEQ ID NO: 89)	
TCR-3	TRBV10-3	CAISELVTGDSPLHF (SEQ ID NO: 82)	9.2% of 120 pairs
	TRAV38-1	CAFMGYSGAGSYQLTF (SEQ ID NO: 79)	

[0179] Plasmids encoding candidate TCRs were cloned with MSGV1 vectors. Retroviral transduction was used to introduce TCR into donor lymphocytes.

[0180] HPLC grade minimal epitope was pulsed on day 4 on A*02:01 DCs for 2 h. DCs were co-cultured with TCR-transduced cells for 20 h. The results are shown in Figures 9A-9D. Three different A*02:01 restricted TP53 receptors were identified.

[0181] T2 cells were thawed and rested for 24 hours, then pulsed with decreasing concentrations of HPLC minimal epitope for 2h. Cells were then washed and co-cultured with TP53 TCR-transduced cells for 17h. The results are shown in Figures 10A-10B.

[0182] The following parental tumor cell lines were transduced with HLA-A*02:01: Colon line – LS123 (ATCC CCL-255) with R175H but unknown HLA-A2 status. Leukemia line: CCRF-CEM (ATCC CRM-CCL-119, leukemia) with R175H but unknown HLA-A2 status. Breast line: AU-565 (ATCC CRL-2351, breast adenocarcinoma) with R175H but unknown HLA-A2 status. Melanoma cell line: MEL624; endogenous expression of HLA-A2 and wt R175.

[0183] Target transduced tumor cells were harvested and plated on the morning of co-culture (1×10^5 cells/well). R175H minimal epitope was pulsed onto target cells (HPLC, 5 μ g/mL) for 2h. Cells were washed twice and co-cultured with 2×10^4 TP53 TCR-transduced cells for 20h (patient 4196). IFN- γ ELISPOT was used for the readout of response. The results are shown in Figure 11. Additional experimental data regarding target cell HLA-A2 and TP53 expression and the reactivity of the TCRs of this Example is provided in Example 15.

[0184] The sequence of TCR **4196_AV12-1_with_BV6-1**, which was isolated from Patient 4196, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1beta (SEQ ID NO: 70), the second underlined region is the CDR2beta (SEQ ID NO: 71), the third underlined region is the CDR3beta (SEQ ID NO: 72), the fourth underlined region is the CDR1alpha (SEQ ID NO: 67), the fifth underlined region is the CDR2alpha (SEQ ID NO: 68), and the sixth underlined region is the CDR3alpha (SEQ ID NO: 69). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the beta chain constant region (SEQ ID NO: 25) and the second italicized region is the alpha chain constant region (SEQ ID NO: 24). The beta chain variable region (SEQ ID NO: 74) includes the sequence starting from the amino terminus and ending immediately prior to the start of the beta chain constant region. The alpha chain variable region (SEQ ID NO: 73) includes the sequence starting immediately after the linker and ending immediately prior to the start of the alpha chain constant region. The full-length beta chain (SEQ ID NO: 76) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length alpha chain (SEQ ID NO: 75) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0185] Cancer reactive T cells were identified as described below.

TCR name: 4196_AV12-1_with_BV6-1

Recognition of p53 mutation: R175H

Method: Tran method

MAIGLLCCVAFSLLWASPVNAGVTQTPKFQVLKTGQSMTLQCAQDMNHNSMYWY
 RQDPGMGLRLIYYSASEGTTDKGEVPNGYNVSRLNKREFSLRLESAAPSQTSVYFCA
SSEGLWQVGDEQYFGPGTRLTVTEDLRNVTPPKVSLFEPSKAEIANKQKATLVCLARGF
FPDHVELSWWWNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNPRNHFRCQVQ
FHGLSEEDKWPEGSPKPVTQNISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLYAV
LVSTLVVMAMVKRKNSRAKRSSGGATNFSLLKQAGDVEENPGPMISLRVLLVILWL
QLSWVWSQRKEVEQDPGPFNVPEGATVAFNCTYSNSASQSFFWYRQDCRKEPKLL
MSVYSSGNEDGRFTAQLNRASQYISLLIRDSKLSDSATYLCVVQPGGYQKVTFGTGT
KLQVIPDIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTMESGTFITDKCVLDMKA
MDSKSNGAIAWSNQTSTFCQDIFKETNATYPSSDVPCDATLTEKSFETDMNLNFQNLLVIV
LRILLKVAGFNLLMTLRLWSS (SEQ ID NO: 547)

[0186] The sequence of TCR **4196_AV38-1_with_BV10-3**, which was isolated from Patient 4196, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1beta (SEQ ID NO: 80), the second underlined region is the CDR2beta (SEQ ID

NO: 81), the third underlined region is the CDR3beta (SEQ ID NO: 82), the fourth underlined region is the CDR1alpha (SEQ ID NO: 77), the fifth underlined region is the CDR2alpha (SEQ ID NO: 78), and the sixth underlined region is the CDR3alpha (SEQ ID NO: 79). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the beta chain constant region (SEQ ID NO: 25) and the second italicized region is the alpha chain constant region (SEQ ID NO: 24). The beta chain variable region (SEQ ID NO: 84) includes the sequence starting from the amino terminus and ending immediately prior to the start of the beta chain constant region. The alpha chain variable region (SEQ ID NO: 83) includes the sequence starting immediately after the linker and ending immediately prior to the start of the alpha chain constant region. The full-length beta chain (SEQ ID NO: 86) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length alpha chain (SEQ ID NO: 85) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0187] Cancer reactive T cells were identified as described below.

TCR name: 4196_AV38-1_with_BV10-3

Recognition of p53 mutation: R175H

Method: Tran method

MATRLFFYVALCLLWTGHMDAGITQSPRHKVTETGTPVTLRCHQTENHRYMYWYR
QDPGHGLRLIHYSYGVDKTDKGEVSDGYSVSRSKTEDFLLTLESATSSQTSVYFCAIS
ELVTGDSPLHFGNGTRLTVTEDLRNVTPPKVSLFEPKAEIANKQKATLVCLARGFFPDH
VELSWVNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNP RNHFRCQVQFHGL
SEEDKWPEGSPKPVTONISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLYAVLVSTL
VVMAMVKRKNSRAKRSGSGATNFSLLKQAGDVEENPGPMTRVSLWAVVSTCLE
SGMAQTVTQSQPEMSVQEAETVTLSCITYDTSENYYLFWYKQPPSRQMILVIRQEA
YKQQNATENRFSVNFQKAASFSLKISDSQLGDTAMYFCAFMGYSGAGSYQLTFGK
GTKLSVIPDIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTMESGTFITDKCVLDMK
AMDSKSNAGIAWSNQTSTFCQDIFKETNATYPSSDVPCDATLTEKSFETDMNLFQNLVI
VLRILLKLVAGFNLLMTLRLWSS (SEQ ID NO: 548)

[0188] The sequence of TCR 4196_AV6_with_BV11-2, which was isolated from Patient 4196, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1beta (SEQ ID NO: 90), the second underlined region is the CDR2beta (SEQ ID NO: 91), the third underlined region is the CDR3beta (SEQ ID NO: 92), the fourth underlined region is the CDR1alpha (SEQ ID NO: 87), the fifth underlined region is the CDR2alpha (SEQ ID NO: 88), and the sixth underlined region is the CDR3alpha (SEQ ID NO: 89). The

bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the beta chain constant region (SEQ ID NO: 25) and the second italicized region is the alpha chain constant region (SEQ ID NO: 24). The beta chain variable region (SEQ ID NO: 94) includes the sequence starting from the amino terminus and ending immediately prior to the start of the beta chain constant region. The alpha chain variable region (SEQ ID NO: 93) includes the sequence starting immediately after the linker and ending immediately prior to the start of the alpha chain constant region. The full-length beta chain (SEQ ID NO: 96) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length alpha chain (SEQ ID NO: 95) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0189] Cancer reactive T cells were identified as described below.

TCR name: 4196_AV6_with_BV11-2

Recognition of p53 mutation: R175H

Method: Tran method

MATRLLCWAALCLLGAELTEAGVAQSPRYKIIKRQSVAFWCNPISGHATLYWYQQ
 ILGQGPKLLIQFQNGVVDSDQLPKDRFSAERLKGVDSTLKIQPAKLEDSAVYLCASS
 LDPGDTGELFFGEGSRLTVLEDLRNVTPPKVSLFEPKAEIANKQKATLVCLARGFFPDH
 VELSWWVNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSAFWHNP RNHFRCQVQFHGL
 SEEDKWPEGSPKPVTONISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLYAVLVSTL
 VVMAMVKKRNSRAKRSGSGATNFSLLKQAGDVEENPGPMESFLGGVLLILWLQVD
 WVKSQKIEQNSEALNIQEGKTATLTCNYTNYSPAYLQWYRQDPGRGPVFLLLIRENE
 KEKRKERLKVTFDTTLKQSLFHITASQPADSATYLCALDIYPHDMRFGAGTRLTVPK
 DIQNPEPAVYQLKDPQRSQDSTLCLFTDFDSQINVPKTMESGTFITDKCVLDMKAMDSKSN
 GAIAWSNQTSTFCQDIFKETNATYPSSDVPCDATLTEKSFETDMNLFQNLVIVLRILLK
 VAGFNLLMTLRLWSS (SEQ ID NO: 549)

EXAMPLE 3

[0190] This example demonstrates the isolation of eleven anti-mutated p53 TCRs from patient 4238.

[0191] Experiments were carried out as described for Figures 12-15 for Patient 4238.

[0192] TIL fragments (F1-F24, n=24) from patient 4238 were co-cultured with autologous APCs electroporated with TMG composed of the irrelevant or mutated p53 sequence. Co-cultures were performed overnight at 37 °C. Secretion of IFN- γ was evaluated using ELISPOT assay. The results are shown in Figure 12. Expression of 4-1BB was

evaluated by flow cytometry after gating for lymphocytes → living cells (PI negative) → CD3+ (T cells). The results are shown in Figure 14.

[0193] TIL fragments (F1-F24, n=24) from patient 4238 were co-cultured with autologous APCs pulsed with peptide vehicle (DMSO) or purified (>95% by HPLC) 25-amino acid peptide composed of the mutated p53-R248Q sequence. Co-cultures were performed overnight at 37 °C. Secretion of IFN-γ was evaluated using ELISPOT assay. The results are shown in Figure 13. Expression of 4-1BB was evaluated by flow cytometry after gating for lymphocytes → living cells (PI negative) → CD3+ (T cells). The results are shown in Figure 15.

[0194] TIL fragments F10, F11 and F17 were sources of TCRs after sorting CD8+41BB+ T cells.

[0195] The sequence of TCR **4238-F10-TCR1**, which was isolated from Patient 4238, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 207), the second underlined region is the CDR2alpha (SEQ ID NO: 208), the third underlined region is the CDR3alpha (SEQ ID NO: 209), the fourth underlined region is the CDR1beta (SEQ ID NO: 210), the fifth underlined region is the CDR2beta (SEQ ID NO: 211), and the sixth underlined region is the CDR3beta (SEQ ID NO: 212). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 213) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 214) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 215) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 216) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0196] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4238-F10-TCR1

Recognition of p53 mutation: R248Q

Screening method: p53 “hotspot” mutation universal screening

Co-culture to identify TCR: Co-culture 4238-F10 with R248Q long peptide, sorted CD8+41BB+ T cells

Method to identify TCR: single-cell reverse transcriptase polymerase chain reaction (RT-PCR)

Abundance of TCR amongst all paired TCRs: 25.0% (observed 3 times of 12 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon

MATASLLRAVIASICVVSSMAQKVTQAQTEISVVEKEDVTLDVCVYETRDTTTYLFW
YKQPPSGELVFLIRNSFDEQNEISGRYSWNFQKSTSSFNFTITASQVVDSAVYFCALS
EVDSGNTPLVFGKGTRLSVIANIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTME
SGTFITDKCVLDMKAMDSKSNAGIAWSNQTSFTCQDIFKETNATYPSSDVPCDATLTEKSF
ETDMNLFQNLVIVLRILLKLVAGFNLLMTLRLWSSRAKRSRSGSGATNFSLLKQAGDV
EENPGPMHFRLLCCVAFCLLGAGPVD SGVTQTPKHLITATGQRVTLRCSPRSGDLSV
YWYQQSLDQGLQFLIQYYNGEERAKGNILERFSAQQFPDLHSELNLSLELGDSALY
FCASSVGSSSSTDQYFGPGTRLTVLEDLRNVTPPKVSLFEPSKAEIANKQKATLVCLAR
GFFPDHVELSWWWNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNP RNHFRCQ
VQFHGLSEEDKWPEGSPKPVTQNISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLY
AVLVSTLVVMAMVKKRNS (SEQ ID NO: 550)

[0197] The statistics for TCR **4238-F10-TCR1** for Patient 4238 are set forth in Table 6 below.

Table 6

Parameter	#	Frequency
Total wells	96	100%
CDR3alpha	4	4.2%
CDR3beta	4	4.2%
4238-F10-TCR1 pairs	3	3.1%
Total paired TCRs	12	12.5%

[0198] The sequence of TCR **4238-F10-TCR2**, which was isolated from Patient 4238, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 217), the second underlined region is the CDR2alpha (SEQ ID NO: 218), the third underlined region is the CDR3alpha (SEQ ID NO: 219), the fourth underlined region is the CDR1beta (SEQ ID NO: 220), the fifth underlined region is the CDR2beta (SEQ ID NO: 221), and the sixth underlined region is the CDR3beta (SEQ ID NO: 222). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable

region (SEQ ID NO: 223) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 224) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 225) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 226) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0199] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolated the TCR is set forth below.

TCR name: 4238-F10-TCR2

Recognition of p53 mutation: R248Q

Screening method: p53 “hotspot” mutation universal screening

Co-culture to identify TCR: Co-culture 4238-F10 with R248Q long peptide, sorted CD8+41BB+ T cells

Method to identify TCR: single-cell RT-PCR

Abundance of TCR amongst all paired TCRs: 25.0% (observed 3 times of 12 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon

MATASLLRAVIASICVVSSMAQKVTQAQTEISVVEKEDVTLDVCVYETRDTTYLFW
YKQPPSGELVFLIRNSFDEQNEISGRYSWNFQKSTSSFNFTITASQVVDSAVYFCALS
EVDSGNTPLVFGKGTRLSVIANIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTM
SGTFITDKCVLDMKAMDSKSNGAIAWSNQTSFTCQDIFKETNATYPSSDVPCDATLTEKSF
ETDMNLNFQNLVIVLRILLKLVAGFNLLMTLRLWSSRAKRSRSGGATNFSLLKQAGDV
EENPGPMHFRLLCCVAFCLLGAGPVDSGVTQTPKHLITATGQRVTLRCSRSGDLSV
YWYQQSLDQGLQFLIHYYNGEERAKGNILERFSAQQFPDLHSELNLSSLELGDSALY
FCASSVGSSSSTDTQYFGPGTRTLVLEDLRNVTPPKVSLFEPSKAELANKQKATLVCLAR
GFFPDHVELSWVWNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNP RNHFRCQ
VQFHGLSEEDKWPEGSPKPTQNISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLY
AVLVSTLVMMAMVKRKNS (SEQ ID NO: 551)

[0200] The statistics for TCR **4238-F10-TCR2** for Patient 4238 are set forth in Table 7 below.

Table 7

Parameter	#	Frequency
Total wells	96	100%
CDR3alpha	4	4.2%
CDR3beta	4	4.2%
4238-F10-TCR2 pairs	3	3.1%

Parameter	#	Frequency
Total paired TCRs	12	12.5%

[0201] The sequence of TCR **4238-F10-TCR3**, which was isolated from Patient 4238, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 227), the second underlined region is the CDR2alpha (SEQ ID NO: 228), the third underlined region is the CDR3alpha (SEQ ID NO: 229), the fourth underlined region is the CDR1beta (SEQ ID NO: 230), the fifth underlined region is the CDR2beta (SEQ ID NO: 231), and the sixth underlined region is the CDR3beta (SEQ ID NO: 232). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 233) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 234) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 235) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 236) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0202] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4238-F10-TCR3

Recognition of p53 mutation: R248Q

Screening method: p53 “hotspot” mutation universal screening

Co-culture to identify TCR: Co-culture 4238-F10 with R248Q long peptide, sorted CD8+41BB+ T cells

Method to identify TCR: single-cell RT-PCR

Abundance of TCR amongst all paired TCRs: 58.3% (observed 7 times of 12 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon

MAKSLRVLLVILWLQLSWVWSQQKEVEQDPGPLSVPEGAIIVSLNCTYSNSAFQYFM
WYRQYSRKGPPELLMYTYSSGNKEDGRFTAQVDKSSKYISLFIRDSQPSDSATYLCAM
TSPYNNNDMRFGAGTRLTVKPNIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTM
ESGTFITDKCVLDMKAMDSKSNGAIAWSNQTSFTCQDIFKETNATYPSSDVPCDATLTEKS
FETDMNLNFQNLVIVLRILLKLVAGFNLLMTLRLWSSRAKRSGSGATNFSLLKQAGD
VEENPGPMHLLLLLLGPGISLLLPGLAGSGLGAVVSQHPSWVICKSGTSVKIECRSL

DFQATTMFWYRQFPKQSLMLMATSNEGSKATYEQGVEKDKFLINHASLTLSTLTVT
SAHPEDSSFYICSGGLEEAARQFIGPGTRLTVLEDLRNVTPPKVSLFEPSKAEIANKQKAT
LVCLARGFFPDHVELSWVWNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNPRN
HFRCQVQFHGLSEEDKWPEGSPKPVTONISAEAWGRADCGITSASYQQGVLSATILYEILL
 GKATLYAVLVSTLVVMAMVKRKNS (SEQ ID NO: 552)

[0203] The statistics for TCR **4238-F10-TCR3** for Patient 4238 are set forth in Table 8 below.

Table 8

Parameter	#	Frequency
Total wells	96	100%
CDR3alpha	16	16.7%
CDR3beta	26	27.1%
4238-F10-TCR3 pairs	7	7.3%
Total paired TCRs	12	12.5%

[0204] The sequence of TCR **4238-F10-TCR4**, which was isolated from Patient 4238, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 237), the second underlined region is the CDR2alpha (SEQ ID NO: 238), the third underlined region is the CDR3alpha (SEQ ID NO: 239), the fourth underlined region is the CDR1beta (SEQ ID NO: 240), the fifth underlined region is the CDR2beta (SEQ ID NO: 241), and the sixth underlined region is the CDR3beta (SEQ ID NO: 242). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 243) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 244) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 245) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 246) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0205] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4238-F10-TCR4

Recognition of p53 mutation: R248Q

Screening method: p53 “hotspot” mutation universal screening

Co-culture to identify TCR: Co-culture 4238-F10 with R248Q long peptide, sorted CD8+41BB+ T cells

Method to identify TCR: single-cell RT-PCR

Abundance of TCR amongst all paired TCRs: 58.3% (observed 7 times of 12 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon

MAKSLRVLLVILWLQLSWVWSQQKEVEQDPGPLSVPEGAIIVSLNCTYSNSAFQYFM
WYRQYSRKGPPELLMYTYSSGNKEDGRFTAQVDKSSKYISLFIRDSQPSDSATYLCAM
TSPYNNNDMRFGAGTRLTVKPNIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTM
ESGFTITDKCVLDMKAMDSSNGAIAWSNQTSFTCQDIFKETNATYPSSDVPCDATLTEKS
FETDMNLNFQNLIVIVLRILLKLVAGFNLLMTLRLWSSRAKRSGSGATNFSLLKQAGD
VEENPGPMHLLLLLLGPAGSGLGAVVSQHPSRVICKSGTSVKIECRSLDFQATTMFW
YRQFPKQSLMLMATSNEGSKATYEQGVEKDKFLINHASLTLSTLTVTSAHPEDSSFYI
CSGGLLEEAAARQFIGPGTRLTVLEDLRNVTTPPKVSLFEPSKAEIANKQKATLVCLARGFFP
DHVELSWVNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNP RNHFRCQVQFH
GLSEEDKWPEGSPKPVTONISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLYAVLV
STLVVMAMVKRKNS (SEQ ID NO: 553)

[0206] The statistics for TCR **4238-F10-TCR4** for Patient 4238 are set forth in Table 9 below.

Table 9

Parameter	#	Frequency
Total wells	96	100%
CDR3alpha	16	16.7%
CDR3beta	26	27.1%
4238-F10-TCR4 pairs	7	7.3%
Total paired TCRs	12	12.5%

[0207] The sequence of TCR **4238-F11-TCR1**, which was isolated from Patient 4238, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 247), the second underlined region is the CDR2alpha (SEQ ID NO: 248), the third underlined region is the CDR3alpha (SEQ ID NO: 249), the fourth underlined region is the CDR1beta (SEQ ID NO: 250), the fifth underlined region is the CDR2beta (SEQ ID NO: 251), and the sixth underlined region is the CDR3beta (SEQ ID NO: 252). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second

italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 253) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 254) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 255) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 256) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0208] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4238-F11-TCR1

Recognition of p53 mutation: R248Q

Screening method: p53 “hotspot” mutation universal screening

Co-culture to identify TCR: Co-culture 4238-F11 with R248Q long peptide, sorted CD8+41BB+ T cells

Method to identify TCR: single-cell RT-PCR

Abundance of TCR amongst all paired TCRs: 25.0% (observed 3 times of 12 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon

MALVARVTVFLTFGTIIDAKTTQPPSMDCAEGRAANLPCNHSTISGNEYVYWYRQIH
SQGPQYIIHGLKNNETNEMASLIITEDRKSSTLILPHATLRDTAVYYCIVPNDYKLSFG
AGTTVTVRANIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTMESGTFITDKCVLD
MKAMDSKSNGAIAWSNQTSFTCQDIFKETNATYPSSDVPCDATLTEKSFETDMNLFQNL
LVIVLRILLKLVAGFNLLMTLRLWSSRAKRSGSGATNFSLLKQAGDVEENPGPMHIRL
LCRVAFCFLAVGLVDVKVTQSSRYLVKRTGEKVFLECVQDMDHENMFWYRQDPGL
GLRLIYFSYDVKMKEKGDIPEGYSVSREKKERFSLILESASTNQTSMYLCASSFGTGS
QETQYFGPGTRLLVLEDLRNVTPPKVSLFEPSKAEIANKQKATLVCLARGFFPDHVELSW
WVNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNP RNHFRCQVQFHGLSEEDK
WPEGSPKPVTONISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLYAVLVSTLVVMA
MVKRKNS (SEQ ID NO: 554)

[0209] The statistics for TCR **4238-F11-TCR1** for Patient 4238 are set forth in Table 10 below.

Table 10

Parameter	#	Frequency
Total wells	96	100%
CDR3alpha	5	5.2%

Parameter	#	Frequency
CDR3beta	3	3.1%
4238-F11-TCR1 pairs	3	3.1%
Total paired TCRs	12	12.5%

[0210] The sequence of TCR **4238-F11-TCR2**, which was isolated from Patient 4238, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 257), the second underlined region is the CDR2alpha (SEQ ID NO: 258), the third underlined region is the CDR3alpha (SEQ ID NO: 259), the fourth underlined region is the CDR1beta (SEQ ID NO: 260), the fifth underlined region is the CDR2beta (SEQ ID NO: 261), and the sixth underlined region is the CDR3beta (SEQ ID NO: 262). The bold region is the linker (SEQ ID NO: 262). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 263) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 264) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 265) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 266) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0211] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4238-F11-TCR2

Recognition of p53 mutation: R248Q

Screening method: p53 “hotspot” mutation universal screening

Co-culture to identify TCR: Co-culture 4238-F11 with R248Q long peptide, sorted CD8+41BB+ T cells

Method to identify TCR: single-cell RT-PCR

Abundance of TCR amongst all paired TCRs: 25.0% (observed 3 times of 12 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon

MAYSPGLVSLILLLLGRTRGNSVTQMEGPVTLSEEAFLTINCTYTATGYPSLFWYVQ
YPGEGLQLLLKATKADDKGSNKGFEATYRKETTFSHLEKGSVQVSDSAVYFCALNP
NAGGTSYGKLTFGQGTILTVHPNIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKT
MESGTFITDKCVLDMKAMDSKSNGAIAWSNQTSFTCQDIFKETNATYPSSDVPCDATLTE

*KSFETDMNLFQNLIVIVLRILLKLVAGFNLLMTLRLWSSRAKRSGSGATNFSLLKQAG
DVEENPGPMHPRLFWALLCLLGTGPVEAGVTQSPTHLIKTRGQQATLRCSPISGHT
SVYWYQQALGLGLQFLLWYDEGEERNRGNFPPRFSGRQFPNYSSSELNVNALELED
 ALYLCASSSPGATSGGANTGELFFGEGSRLTVLEDLRNVTPPKVSLFEPKAEIANKQK
 ATLVCLARGFFPDHVELSWVNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNP
 RNFHRCQVQFHGLSEEDKWPEGSPKPTQNISAEAWGRADCGITSASYQQGVLSATILYEI
 LLGKATLYAVLVSTLVVMAMVKRKNS (SEQ ID NO: 555)*

[0212] The statistics for TCR **4238-F11-TCR2** for Patient 4238 are set forth in Table 11 below.

Table 11

Parameter	#	Frequency
Total wells	96	100%
CDR3alpha	5	5.2%
CDR3beta	8	8.3%
4238-F11-TCR2 pairs	3	3.1%
Total paired TCRs	12	12.5%

[0213] The sequence of TCR **4238-F11-TCR3**, which was isolated from Patient 4238, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 267), the second underlined region is the CDR2alpha (SEQ ID NO: 268), the third underlined region is the CDR3alpha (SEQ ID NO: 269), the fourth underlined region is the CDR1beta (SEQ ID NO: 270), the fifth underlined region is the CDR2beta (SEQ ID NO: 271), and the sixth underlined region is the CDR3beta (SEQ ID NO: 272). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 273) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 274) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 275) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 276) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0214] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4238-F11-TCR3

Recognition of p53 mutation: R248Q

Screening method: p53 “hotspot” mutation universal screening

Co-culture to identify TCR: Co-culture 4238-F11 with R248Q long peptide, sorted CD8+41BB+ T cells

Method to identify TCR: single-cell RT-PCR

Abundance of TCR amongst all paired TCRs: 16.7% (observed 2 times of 12 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon

MATFAGFSFLFLWLQLDCMSRGEDVEQSLFLSVREGDSSVINCTYTDSSSTYLYWYK
QEPGAGLQLLTYIFSNMDMKQDQRLTVLLNKKDKHLSLRIADTQTGDSAIYFCAEIP
RDSGGGADGLTFGKGTHLIQPNIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTM
ESGFTITDKCVLDMKAMDSKSNAGIAWSNQTSTFCQDIFKETNATYPSSDVPCDATLTEKS
FETDMNLNFQNLVIVLRILLKLVAGFNLLMTLRLWSSRAKRS~~SG~~GATNFSLLKQAGD
VEENPGPMHLLLLLLGPGISLLLPGLAGSGLGAVVSQHPSWVICKSGTSVKIECRSL
DFQATTMFWRQFPKQSLMLMATSNEGSKATYEQGVEKDKFLINHASLTLSTLTVT
SAHPEDSSFYICSARDLQRSYEQYFGPGTRTLVTEDLRNVTPPKVSLFEPKAEIANKQK
ATLVCLARGFFPDHVELSWVWNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNP
RNHFRCQVQFHGLSEEDKWPEGSPKPVTONISAEAWGRADCGITSASYQQGVLSATILYEI
LLGKATLYAVLVSTLVVMAMVKRKNS (SEQ ID NO: 556)

[0215] The statistics for TCR **4238-F11-TCR3** for Patient 4238 are set forth in Table 12 below.

Table 12

Parameter	#	Frequency
Total wells	96	100%
CDR3alpha	4	4.2%
CDR3beta	2	2.1%
4238-F11-TCR3 pairs	2	2.1%
Total paired TCRs	12	12.5%

[0216] The sequence of TCR **4238-F11-TCR4**, which was isolated from Patient 4238, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 277), the second underlined region is the CDR2alpha (SEQ ID NO: 278), the third underlined region is the CDR3alpha (SEQ ID NO: 279), the fourth underlined region is the CDR1beta (SEQ ID NO: 280), the fifth underlined region is the CDR2beta (SEQ ID NO: 281), and the sixth underlined region is the CDR3beta (SEQ ID NO:

282). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 283) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 284) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 285) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 286) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0217] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4238-F11-TCR4

Recognition of p53 mutation: R248Q

Screening method: p53 "hotspot" mutation universal screening

Co-culture to identify TCR: Co-culture 4238-F11 with R248Q long peptide, sorted CD8+41BB+ T cells

Method to identify TCR: single-cell RT-PCR

Abundance of TCR amongst all paired TCRs: 16.7% (observed 2 times of 12 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon

MATFAGFSFLFLWLQLDCMSRGEDVEQSLFLSVREGDSSVINCTYTDSSSTYLYWYK
QEPGAGLQLLTYIFSNMDMKQDQRLTVLLNKKDKHLSLRIADTQTGDSAIYFCAEIP
RDSGGGADGLTFGKGTHLIIQPNIQNPEPAVYQLKDPRSQDSTLCFLTDFDSQINVPKTM
ESGTFITDKCVLDMKAMDSKSNGAIAWSNQTSTFCQDIFKETNATYPSSDVPCDATLTEKS
FETDMNLNFQNLLVIVLRILLKLVAGFNLLMTLRLWSSRAKRSGSGATNFSLLKQAGD
VEENPGPMHLLLLLLGPAGSGLGAVVSQHPSRVICKSGTSVKIECRSLDFQATTMFW
YRQFPKQSLMLMATSNEGSKATYEQGVEKDKFLINHASLTLSTLTVTSAHPEDSSFYI
CSARDLQRSYEQYFGPGTRLTVTEDLRNVTPPKVSLFEPKAEIANKQKATLVCLARGFF
PDHVELSWVWNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNPNNHFRQVQF
HGLSEEDKWPEGSPKPVTONISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLYAVL
VSTLVVMAMVKRKNS (SEQ ID NO: 557)

[0218] The statistics for TCR **4238-F11-TCR4** for Patient 4238 are set forth in Table 13 below.

Table 13

Parameter	#	Frequency
Total wells	96	100%
CDR3alpha	4	4.2%
CDR3beta	2	2.1%
4238-F11-TCR4 pairs	2	2.1%
Total paired TCRs	12	12.5%

[0219] The sequence of TCR **4238-F17-TCR1**, which was isolated from Patient 4238, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 287), the second underlined region is the CDR2alpha (SEQ ID NO: 288), the third underlined region is the CDR3alpha (SEQ ID NO: 289), the fourth underlined region is the CDR1beta (SEQ ID NO: 290), the fifth underlined region is the CDR2beta (SEQ ID NO: 291), and the sixth underlined region is the CDR3beta (SEQ ID NO: 292). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 293) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 294) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 295) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 296) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0220] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4238-F17-TCR1

Recognition of p53 mutation: R248Q

Screening method: p53 “hotspot” mutation universal screening

Co-culture to identify TCR: Co-culture 4238-F17 with R248Q long peptide, sorted CD8+41BB+ T cells

Method to identify TCR: single-cell RT-PCR

Abundance of TCR amongst all paired TCRs: 33.3% (observed 2 times of 6 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon

MAGIRALFMYLWLQLDWVSRGESVGLHLPTLSVQEGDNSIINCAYSNSASDYFIWY
 KQESGKGPFIIIDIRSNMDKRQGGQRTVLLNKTVKHLSLQIAATQPGDSAVYFCAEP
 VGGLNSGYALNFGKGTSLLVTPNIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKT
 MESGTFITDKCVLDMKAMDSKSNGAIAWSNQTSFTCQDIFKETNATYPSSDVPCDATLTE
 KSFETDMNLNFQNLVIVLRILLKLVAGFNLLMTLRLWSSRAKRSGSGATNFSLLKQAG
DVEENPGPMHPGLLCWVLLCLLGAGPVDAGVTQSPTHLIKTRGQQVTLRCSPISGH
KSVSWYQQVLGQGPQFIFQYYEKEERGRGNFPDRFSARQFPNYSSELNVNALLGDS
 ALYLCASSGGRTSGAYEQFFGPGTRLTVLEDLRNVTPPKVSLFEPSKAEIANKQKATLV
 CLARGFFPDHVELSWVNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNPRNH
 FRCQVQFHGLSEEDKWPEGSPKPVTONISAEAWGRADCGITSASYQQGVLSATILYEILLG
 KATLYAVLVSTLVVMAMVKRKNS (SEQ ID NO: 558)

[0221] The statistics for TCR **4238-F17-TCR1** for Patient 4238 are set forth in Table 14 below.

Table 14

Parameter	#	Frequency
Total wells	96	100%
CDR3alpha	3	3.1%
CDR3beta	2	2.1%
4238-F17-TCR1 pairs	2	2.1%
Total paired TCRs	6	6.3%

[0222] The sequence of TCR **4238-F17-TCR2**, which was isolated from Patient 4238, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 297), the second underlined region is the CDR2alpha (SEQ ID NO: 298), the third underlined region is the CDR3alpha (SEQ ID NO: 299), the fourth underlined region is the CDR1beta (SEQ ID NO: 300), the fifth underlined region is the CDR2beta (SEQ ID NO: 301), and the sixth underlined region is the CDR3beta (SEQ ID NO: 302). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 303) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 304) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 305) includes the sequence starting from the amino terminus and ending

immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 306) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0223] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4238-F17-TCR2

Recognition of p53 mutation: R248Q

Screening method: p53 “hotspot” mutation universal screening

Co-culture to identify TCR: Co-culture 4238-F17 with R248Q long peptide, sorted CD8+41BB+ T cells

Method to identify TCR: single-cell RT-PCR

Abundance of TCR amongst all paired TCRs: 33.3% (observed 2 times of 6 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon

MAKMLECAFIVLWLQLGWLSGEDQVTQSPEALRLQEGESSLNCSYTVSGLRGLFW
YRQDPGKGPEFLFTLYSAGEEKEKERLKATLTKKESFLHITAPKPEDSATYLCAVTAH
RGSTLGRLYFGRGTQLTVWPNIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTME
SGTFITDKCVLDMKAMDSKSNAGIAWSNQTSTFCQDIFKETNATYPSSDVPCDATLTEKSF
ETDMNLNFQNLLVIVLRILLKLVAGFNLLMTLRLWSSRAKRSGSGATNFSLLKQAGDV
EENPGPMHPGLLCWALLCLLGAGLVDAGVTQSPTHLIKTRGQQVTLRCSPKSGHDT
VSWYQQALGQGPQFIFQYEEEEERQRGNFPDRFSGHQFPNYSSELNVNALLLGDSAL
YLCASSRRGGAYNEQFFGPGTRLTVLEDLRNVTTPPKVSLFEPKAEIANKQKATLVCLAR
GFFPDHVELSWWWNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNPNNHFRQC
VQFHGLSEEDKWPEGSPKPVTONISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLY
AVLVSTLVVMAMVKKNS (SEQ ID NO: 559)

[0224] The statistics for TCR **4238-F17-TCR2** for Patient 4238 are set forth in Table 15 below.

Table 15

Parameter	#	Frequency
Total wells	96	100%
CDR3alpha	6	6.3%
CDR3beta	2	2.1%
4238-F17-TCR2 pairs	2	2.1%
Total paired TCRs	6	6.3%

[0225] The sequence of TCR **4238-F17-TCR3**, which was isolated from Patient 4238, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 307), the second underlined region is the CDR2alpha (SEQ ID NO: 308), the third underlined region is the CDR3alpha (SEQ ID NO: 309), the fourth

underlined region is the CDR1beta (SEQ ID NO: 310), the fifth underlined region is the CDR2beta (SEQ ID NO: 311), and the sixth underlined region is the CDR3beta (SEQ ID NO: 312). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 313) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 314) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 315) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 316) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0226] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4238-F17-TCR3

Recognition of p53 mutation: R248Q

Screening method: p53 "hotspot" mutation universal screening

Co-culture to identify TCR: Co-culture 4238-F17 with R248Q long peptide, sorted CD8+41BB+ T cells

Method to identify TCR: single-cell RT-PCR

Abundance of TCR amongst all paired TCRs: 33.3% (observed 2 times of 6 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon

MAKNPLAAPLLILWFHLDCVSSILNVEQSPQSLHVQEGDSTNFTCSFPSSNFYALHWY
RWETAKSPEALFVMTLNGDEKKKGRISATLNTKEGYSYLYIKGSQPEDSATYLCASV
GGGADGLTFGKGTHLIIQPNIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTMESG
TFITDKCVLDMKAMDSDSKSNGALAWSNQTSFTCQDIFKETNATYPSSDVPCDATLTEKSFET
DMNLNFQNLVIVLRILLKLVAGFNLLMTLRLWSS**RAKRSGSGATNFSLLKQAGDVEE**
NPGPMHTRLLFWVAFCLLGADHTGAGVSQSPSNKVTEKGKDVELRCDPISGHTALY
WYRQSLGQGLEFLIYFQGN**SAPDKSGLPSDRFSAERTGGSVSTLTIQRTQQEDSAVYL**
CASTWDRGSYNEQFFGPGTRLTVLEDLRNVTPPKVSLFEP**SKAEIANKQKATLVCLARG**
FFPDHVELSWVNGKEVHSGVCTDPQAYKESNYSYCLSSRLRV**SATFWHNPRNHFRCQV**
QFHGLSEEDKWPEGSPKPVTQNISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLYA
VLVSTLVVMAMVKRKNS (SEQ ID NO: 560)

[0227] The statistics for TCR **4238-F17-TCR3** for Patient 4238 are set forth in Table 16 below.

Table 16

Parameter	#	Frequency
Total wells	96	100%
CDR3alpha	2	2.1%
CDR3beta	2	2.1%
4238-F17-TCR3 pairs	2	2.1%
Total paired TCRs	6	6.3%

EXAMPLE 4

[0228] This example demonstrates the isolation of two anti-mutated p53 TCRs from patient 4253.

[0229] An experiment was carried out as described for Figure 16 for Patient 4253.

[0230] TIL fragments (F1-F24; n=24) from patient 4253 were co-cultured with autologous APCs (1) pulsed with peptide vehicle (DMSO), (2) pulsed with purified (>95% by HPLC) 25-amino acid peptides composed of mutated p53-R248W sequence, (3) electroporated with an irrelevant TMG or (4) electroporated with p53-mut-TMG containing the mutated p53-R248W sequence. Co-cultures were performed overnight at 37 °C. Secretion of IFN- γ was evaluated by ELISPOT. The results are shown in Figure 16. F15 was selected for sorting for TCRs because it was reactive to both p53-mut-TMG and p53-R248W long peptide.

[0231] The sequence of TCR **4253-TIL-TCR1**, which was isolated from Patient 4253, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 187), the second underlined region is the CDR2alpha (SEQ ID NO: 188), the third underlined region is the CDR3alpha (SEQ ID NO: 189), the fourth underlined region is the CDR1beta (SEQ ID NO: 190), the fifth underlined region is the CDR2beta (SEQ ID NO: 191), and the sixth underlined region is the CDR3beta (SEQ ID NO: 192). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 193) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 194) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha

chain (SEQ ID NO: 195) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 196) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0232] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4253-TIL-TCR1

Recognition of p53 mutation: R248W

Method: Abbreviated p53 “hotspot” mutation universal screening (only mutated TMG and R248W long peptide evaluated)

Co-culture to identify TCR: Co-culture 4253-F15 with p53-R248W long peptide, sorted CD3+41BB+ T cells

Method to identify TCR: single-cell RT-PCR

Abundance of TCR amongst all paired TCRs: 5.6% (observed 2 times of 36 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon

MAKCPQALLAIFWLLLSWVSSDKVVQSPLSLVVHEGDTVTLNCSYEVTNFRSLLW
YKQEKKAPTFLFMLTSSGIEKKSGRLSSILDKKELSSILNITATQTGDSAIIYLCAGQNY
GGSQGNLIFGKGTKLSVKPNIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTMESG
TFITDKCVLDMKAMDSKSNGLAWSNQTSFTCQDIFKETNATYPSSDVPCDNLTEKSFET
DMNLNFQNLVIVLRILLKLVAGFNLLMTLRLWSSRAKRSGSGATNFSLLKQAGDVEE
NPGPMHTWLVCWAIFSLKAGLTEPEVTQTPSHQVTQMGEVILRCVPISNHLIFY
WYRQILGQKVEFLVSFYNNSEISEKSEIFDDQFSVERPDGSNFTLKIRSTKLEDSAMYFC
ASRDPAYEQYFGPGTRLTVTEDLRNVTPPKVSLFEPKAEIANKQKATLVCLARGFFPD
HVELSWVWNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNPVNHFRQVQVQFHG
LSEEDKWPEGSPKPVTONISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLYAVLVST
LVVMAMVKKRNS (SEQ ID NO: 561)

[0233] The statistics for TCR **4253-TIL-TCR1** for Patient 4253 are set forth in Table 17 below.

Table 17

Parameter	#	Frequency
Total wells	55	100%
CDR3alpha	3	5.5%
CDR3beta	2	3.6%
4253-TIL-TCR1 pairs	2	3.6%
Total paired TCRs	36	65.5%

[0234] The sequence of TCR **4253-TIL-TCR2**, which was isolated from Patient 4253, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 197), the second underlined region is the CDR2alpha (SEQ ID NO: 198), the third underlined region is the CDR3alpha (SEQ ID NO: 199), the fourth underlined region is the CDR1beta (SEQ ID NO: 200), the fifth underlined region is the CDR2beta (SEQ ID NO: 201), and the sixth underlined region is the CDR3beta (SEQ ID NO: 202). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 203) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 204) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 205) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 206) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0235] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4253-TIL-TCR2

Recognition of p53 mutation: R248W

Method: Abbreviated p53 "hotspot" mutation universal screening (only mutated TMG and R248W long peptide evaluated)

Co-culture to identify TCR: Co-culture 4253-F15 with p53-R248W long peptide, sorted CD3+41BB+ T cells

Method to identify TCR: single-cell RT-PCR

Abundance of TCR amongst all paired TCRs: 91.7% (observed 33 times of 36 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon

MASLRVLLVILWLQLSWVWSQQKEVEQNSGPLSVPEGAIASLNCTYSDRGSQSFFW
YRQYSGKSPELIMFIYSNGDKEDGRFTAQLNKASQYVSLIRDSQPSDSATYLCVNP
PVKTSYDKVIFGPGTSLSVIPNIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTMES
GTFITDKCVLDMKAMDSSNGAIAWSNQTSFTCQDIFKETNATYPSSDVPCDATLTEKSFE
TDMNLNFQNLVIVLRILLKLVAGFNLLMTLRWSSRAKRSRSGATNFSLLKQAGDVE
ENPGPMHTSLLCWVVLGFLGTDHTGAGVSQSPRYKVTKRGQDVALRCDPISGHVSL
YWYRQALGQGPEFLTYFNIEAQQDKSGLPNDRFSAERPEGSISTLTIQRTQEQRDSAM
YRCASSHREPHGTGELFFGEGSRLTVLEDLRNVTPPKVSLFEPKAEIANKQKATLVCLAR
GFFPDHVELSWVWNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNPNNHFRQ

*VQFHGLSEEDKWPEGSPKPVTQNISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLY
AVLVSTLVVMAMVKRKNS* (SEQ ID NO: 562)

[0236] The statistics for TCR **4253-TIL-TCR2** for Patient 4253 are set forth in Table 18 below.

Table 18

Parameter	#	Frequency
Total wells	55	100%
CDR3alpha	35	63.6%
CDR3beta	33	60.0%
4253-TIL-TCR2 pairs	33	60.0%
Total paired TCRs	36	65.5%

EXAMPLE 5

[0237] This example demonstrates the identification of anti-mutated p53 T cells in Patient 4273 by co-culturing autologous APCs induced to express mutated p53 within autologous T cells (“p53 hotspot mutation universal screening”). This example also demonstrates the isolation of two anti-mutated p53 TCRs from patient 4273.

[0238] Experiments were carried out as described for Figures 17-20 and 56-60 for Patient 4273.

[0239] TIL fragments (F1-F24, n=24) from patient 4273 were co-cultured with autologous APCs electroporated with TMG composed of irrelevant, WT p53 or mutated p53 sequence. Co-cultures were performed overnight at 37 °C. Secretion of IFN-γ was evaluated using ELISPOT assay. The results are shown in Figure 17. Expression of 4-1BB was evaluated by flow cytometry after gating for lymphocytes → living cells (PI negative) → CD3+ (T cells). The results are shown in Figure 18.

[0240] TIL fragments (F1-F24, n=24) from patient 4273 were co-cultured with autologous APCs pulsed with peptide vehicle (DMSO) or purified (>95% by HPLC) 25-amino acid peptides composed of WT p53-R248 sequence or mutated p53-R248W sequence. Co-cultures were performed overnight at 37 °C. Secretion of IFN-γ was evaluated using ELISPOT assay. The results are shown in Figure 19. Expression of 4-1BB was evaluated by flow cytometry after gating for lymphocytes → living cells (PI negative) → CD3+ (T cells). The results are shown in Figure 20.

[0241] For patient 4273, F15 was the most reactive fragment and the responses by F15 to the LP and TMG were comparable and primarily by CD4 T cells. So 4273-F15 was co-cultured with APCs pulsed with the R248W LP, and the following day CD4+41BB⁺ T cells were sorted as single cells into the wells of a 96 well PCR plate at one cell per well. The PCR plate has an RT-PCR solution in each well, which amplifies the TCR alpha and beta CDR3 regions in the same solution. The wells of the plate are then split into two 96 well PCR plates and a second PCR round is performed to amplify either the CDR3 alpha or the CDR3 beta as separate reactions. The PCR products from each well (total of 192 PCR products mapped to each well – alpha or beta) are sequenced by Sanger sequencing. The nucleotide sequence is inputted into IMGT/V-QUEST (imgt.org/IMGT_vquest/vquest?livret=0&Option=humanTcR), IgBlast (ncbi.nlm.nih.gov/igblast/igblast.cgi?CMD=Web&SEARCH_TYPE=TCR&LINK_LOC=igtab) and translated by Expasy (web.expasy.org/translate/). The variable family is determined and fused to the CDR3 and junction (J or DJ) from the translated sequence. The variable sequence is fused to the murine constant sequence and the reconstructed TCRalpha and TCRbeta are linked by furin-felxible-P2A (RAKR-SGSG-ATNFSLLKQAGDVEENPGP) (SEQ ID NO: 26). Then the sequence is synthesized into DNA de novo and cloned into an expression vector (gamma-retrovirus or SLEEPING BEAUTY (SB) transposon (University of Minnesota, Minneapolis, MN)). T cells are then made to express the TCR using the standard viral transduction or non-viral transposition protocols and T cells expressing murinized TCRs (as detected by mouse TCR beta constant chain) are tested against the putative peptide.

[0242] Autologous APCs were transfected with TMG encoding irrelevant mutations, WT p53 sequences or mutated p53 sequences including p53-R248W. Media alone and PMA and Ionomycin were negative and positive controls, respectively. TIL from patient 4273 (fragment cultures 8 and 15) were co-cultured overnight at 37 °C with TMG transfected APCs. Expression of 4-1BB was evaluated by flow cytometry after gating for lymphocytes → living cells (PI negative) → CD3⁺ (T cells). The results are shown in Figure 56.

[0243] Autologous APCs were pulsed with 25 amino acid peptides corresponding to the WT or mutated p53-R248W neoepitope for 2 hours at 37 °C. TIL from patient 4273 (fragment culture 15) with specificity to p53-R248W were co-cultured overnight at 37 °C with peptide-pulsed APCs. DMSO was peptide vehicle. Expression of 4-1BB was evaluated

by flow cytometry after gating for lymphocytes → living cells (PI negative) → CD3+ (T cells). The results are shown in Figure 57.

[0244] Autologous APCs were pulsed with 15 amino acid peptides from the p53-R248W neoepitope overlapping 14 amino acids. TIL from patient 4273 (fragment culture 15) with specificity to p53-R248W were co-cultured overnight at 37 °C with peptide-pulsed APCs. DMSO was peptide vehicle, media alone (T cells only) and PMA and ionomycin were controls. The 25 amino acid peptides (wt p53-R248 and mutated p53-R248W) were additional controls for the 15 amino acid peptides. Expression of 4-1BB was evaluated by flow cytometry after gating for lymphocytes → living cells (PI negative) → CD3+ (T cells) → CD4+CD8-. The results are shown in Figure 58.

[0245] Cos7 cells (2.5×10^4 per well) were plated on wells of flat-bottom 96 well plates. The following day, cells were co-transfected with individual HLA alleles from patient 4273 and either wild type or mutated TP53 TMG with or without the p53-R248W neoantigen, respectively. The following day, TIL with specificity to p53-R248W from Patient 4273 (fragment culture 15) were co-cultured with transfected Cos7 cells overnight at 37 °C. Secretion of IFN- γ was evaluated by ELISA. The results are shown in Figure 59.

[0246] T cells expressing mock (no TCR) or 4273-TCR1a2 were co-cultured with autologous APCs which were pulsed with peptide vehicle (DMSO) or purified (>95% by HPLC) 25 amino acid peptides composed of wild type p53-R248 sequence or mutated p53-R248W sequences. Media alone and PMA and Ionomycin were negative and positive controls, respectively. Co-cultures were performed overnight at 37 °C. Secretion of IFN- γ was evaluated by ELISPOT. The results are shown in Figure 60.

[0247] The sequence of TCR 4273-TP53-R248W-TCR1a1, which was isolated from Patient 4273, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 437), the second underlined region is the CDR2alpha (SEQ ID NO: 438), the third underlined region is the CDR3alpha (SEQ ID NO: 439), the fourth underlined region is the CDR1beta (SEQ ID NO: 440), the fifth underlined region is the CDR2beta (SEQ ID NO: 441), and the sixth underlined region is the CDR3beta (SEQ ID NO: 442). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 443) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable

region (SEQ ID NO: 444) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 445) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 446) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

TCR name: 4273-TP53-R248W-TCR1a1

Recognition of p53 mutation: R248W

Method: p53 “hotspot” mutation universal screening

MASLRVLLVILWLQLSWVWSQQKEVEQNSGPLSVPEGAIASLNCTYSDRGSQSFFW
 YRQYSGKSPELIMFIYSNGDKEDGRFTAQLNKASQYVSLIRDSQPSDSATYLCAVTL
CGGYNKLIFGAGTRLAVHPNIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTMES
GTFITDKCVLDMKAMDSKSNGAIAWSNQTSFTCQDIFKETNATYPSSDVPCDATLTEKSFE
TDMNLNFQNLLIVLRILLKVAGFNLLMTLRLWSSRAKRSGSGATNFSLLKQAGDVE
ENPGPMHNQVLCCVL**CFLGANTVDGGITQSPKYLFRKEGQNV**TL**SCEQNLNHDA**
 MYWYRQDPGQGLRLIYYSQIVNDFQKGDIVEGYVSREKKESFPLTVTSAQKNPTAF
 YLCASSSRDYEQYFGPGTRLTVTEDLRNVTPPKVSLFEPSKAEIANKQKATLVCLARGFF
 PDHVELSWVWNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSAFWHNPRNHFRCQVQF
 HGLSEEDKWPEGSPKPVTONISAEAWGRADCGITSASYQOGVLSATILYEILLGKATLYAVL
 VSTLVVMAMVKKNS (SEQ ID NO: 563)

[0248] The sequence of TCR 4273-TP53-R248W-TCR1a2, which was isolated from Patient 4273, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 447), the second underlined region is the CDR2alpha (SEQ ID NO: 448), the third underlined region is the CDR3alpha (SEQ ID NO: 449), the fourth underlined region is the CDR1beta (SEQ ID NO: 450), the fifth underlined region is the CDR2beta (SEQ ID NO: 451), and the sixth underlined region is the CDR3beta (SEQ ID NO: 452). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 453) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 454) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 455) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 456)

includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

TCR name: 4273-TP53-R248W-TCR1a2

Recognition of p53 mutation: R248W

Method: p53 “hotspot” mutation universal screening

MASLRVLLVILWLQLSWVWSQQKEVEQNSGPLSVPEGAIASLNCTYSDRGSSQFFW
 YRQYSGKSPELIMFIYSNGDKEDGRFTAQLNKASQYVSLIRDSQPSDSATYLCVTL
SGGYNKLIFGAGTRLAVHPNIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTMES
GTFTDKCVLDMKAMDSKNGAIAWSNQTSTFCQDIFKETNATYPSSDVPCDATLTEKSFE
TDMNLNFQNLLVIVLRILLKLVAGFNLLMTLRLWSSRAKRSGSGATNFSLLKQAGDVE
ENPGPMHNQVLCVFLGANTVDGGITQSPKYLFRKEGQNVTLSCQNINHDA
 MYWYRQDPGQGLRLIYYSQIVNDFQKGDIVEGYSVSREKKESFPLTVTSAQKNPTAF
YLCASSRDYEQYFGPGTRLTVTEDLRNVTPPKVSLFEPKAEIANKQKATLVCLARGFF
PDHVELSWVNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNPRNHFRCQVQF
HGLSEEDKWPEGSPKPVTONISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLYAVL
VSTLVVMAMVKRKNS (SEQ ID NO: 564)

[0249] The statistics for the TCRs of Patient 4273 are set forth in Table 19 below. In Table 19, 96 total wells sorted with 41BB+ T cells after co-culture with mutated p53 protein (TMG or peptide). 77 wells had productive pairs (meaning had (1) a sequence and (2) no stop codons in the sequence) for a pairing frequency of 80.2%. 43 of those pairs were the CDR3A/CDR3B combination to make 4273-TP53-R248W-TCR1a1 (55.8% of the productive pairs). 30 of those pairs were the CDR3A/CDR3B combination to make 4273-TP53-R248W-TCR1a2 (39% of the productive pairs). Overall, the CDR3A and CDR3B for 4273-TP53-R248W-TCR1a1 were found 50 and 83 times, respectively, out of 96 wells. Overall, the CDR3A and CDR3B for 4273-TP53-R248W-TCR1a2 were found 33 and 83 times, respectively, out of 96 wells.

TABLE 19

TCR name	4273-TP53-R248W-TCR1a1	4273-TP53-R248W-TCR1a2
CDR3a	CAVTLGGYNKLIF (SEQ ID NO: 439)	CAVTLSSGGYNKLIF (SEQ ID NO: 449)
CDR3b	CASSSRDYEYF (SEQ ID NO: 442)	CASSSRDYEYF (SEQ ID NO: 452)
total wells	96	
total CDR3a/CDR3b pairs	77	
% paired	80.21%	
# times CDR3a	50	33

TCR name	4273-TP53-R248W-TCR1a1	4273-TP53-R248W-TCR1a2
# times CDR3b	83	83
paired CDR3a/CDR3b	43	30
% of paired CDR3a/CDR3b pairs	55.84%	38.96%

EXAMPLE 6

[0250] This example demonstrates the identification of anti-mutated p53 T cells in Patient 4149. This example also demonstrates the isolation and specific reactivity of one anti-mutated p53 TCR from patient 4149.

[0251] Experiments were carried out as described for Figures 21-24 for Patient 4149. The TCR was found using the Tran method. The TCR was then used to validate the “p53 hotspot mutation universal screening” method.

[0252] A TCR (4149-TCRa2b1 or 4149-TCRa2b2) was transposed into autologous PBL from patient 4149 and co-cultured with autologous APCs which were (1) electroporated with TMG composed of irrelevant, WT p53, or mutated p53 sequence or (2) pulsed with peptide vehicle (DMSO) or purified (>95% by HPLC) 25-amino acid peptides composed of WT p53-Y220 sequence or mutated p53-Y220C sequence. Combination of PMA and ionomycin was positive control. Co-cultures were performed overnight at 37 °C. Secretion of IFN- γ was evaluated using ELISPOT assay. The results are shown in Figure 21. Expression of 4-1BB was evaluated by flow cytometry after gating for lymphocytes $\rightarrow$ living cells (PI negative) $\rightarrow$ CD3+ (T cells) $\rightarrow$ CD4+ mTCR+ (TCR transposed T cells). The results are shown in Figure 22.

[0253] The percentage of CD4+4-1BB+ cells by TCRAD deep sequencing and TCRB deep sequencing was also performed. The results are shown in Table E.

TABLE E

TCR name	% of CD4+4-1BB+ by TCRAD deep sequencing	% of CD4+4-1BB+ by TCRB deep sequencing
4149-TCR-a2b1	20%	66%
4149-TCR-a2b2	20%	18%

[0254] Mapping of putative p53^{Y220C} minimal epitope recognized by 4149-F11: Autologous DC cells were peptide pulsed (10 µg/mL) and rested overnight in granulocyte-macrophage colony-stimulating factor (GM-CSF) and IL-4. TIL were rested for 2-3 days in 500 CU/mL IL-2. 2×10^4 TIL and 10^5 target cells were co-cultured overnight at 37 °C. IFN-γ was measured by ELISPOT. The results are shown in Table F. 4-1BB expression was measured by FACS with the gate lymphocytes\PI(neg)CD3+\CD3+CD4+. The results are shown in Figure 23.

TABLE F

Peptide No.	ELISPOT Result Positive (+) or negative (-) for IFN-γ production	Peptide	SEQ ID NO:
0	-	Vehicle	Not applicable
1	+	DRNTFRHSVVVP <u>C</u> EP	508
2	+	RNTFRHSVVVP <u>C</u> EPP	509
3	+	NTRHSVVVP <u>C</u> EPPE	510
4	+	TRHSVVVP <u>C</u> EPPEV	511
5	+ (weak)	FRHSVVVP <u>C</u> EPPEVG	512
6	-	RHSVVVP <u>C</u> EPPEVGS	513
7	-	HSVVVP <u>C</u> EPPEVGSD	514
8	-	SVVVVP <u>C</u> EPPEVGSDC	515
9	-	VVVVP <u>C</u> EPPEVGSDCT	516
10	-	VVP <u>C</u> EPPEVGSDCTT	517
11	-	VP <u>C</u> EPPEVGSDCTTI	518

[0255] Cos7 cells (2.5×10^4 per well) were plated on wells of flat-bottom 96 well plates. After 20 hours, cells were co-transfected with individual HLA alleles with or without TMGs. After 20 hours, autologous DC cells were transfected with TMG in parallel. All HLA Class-II alleles were co-transfected into one set of wells with or without TMG. Cells not transfected with TMG were pulsed with p53-Y220C 15-mer peptide for 2-3 hours at 37 °C at 10 µg/mL. After washing, 4149-TCRa2b2-transposed T cells (10^5) at day+14 of second REP were added to wells and co-cultured overnight at 37 °C. IFN-γ secretion was measured by

ELISA. The results are shown in Figure 24. Prediction (Table G) by NetMHCIIpan:
 cbs.dtu.dk/services/NetMHCIIpan/.

TABLE G

HLA	Peptide	Affinity, nM	Rank
DRB3*02:02	NTFRHSVVVP <u>C</u> EPPE (SEQ ID NO: 510)	433.8	17

[0256] DRB3*02 expression was detected in 1367 of 3719 (37%) of DRB_ typed patients in the NCI HLA database and in 5 of 9 (56%) endometrial and ovarian cancer patients at NCI-SB. The reported frequency of the DRB3*02 allele is very high, as described in Example 1.

[0257] The sequence of TCR **4149TCRa2b2**, which was isolated from Patient 4149, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 57), the second underlined region is the CDR2alpha (SEQ ID NO: 58), the third underlined region is the CDR3alpha (SEQ ID NO: 59), the fourth underlined region is the CDR1beta (SEQ ID NO: 60), the fifth underlined region is the CDR2beta (SEQ ID NO: 61), and the sixth underlined region is the CDR3beta (SEQ ID NO: 62). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 63) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 64) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 65) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 66) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0258] Cancer reactive T cells were identified using the screening method set forth below. The p53 reactive cells for this patient were identified by the Transcreening method as described in U.S. Application No. 2017/0224800.

TCR name: 4149TCRa2b2

Recognition of p53 mutation: Y220C

Screening method: Used to validate the p53 “hotspot” mutation universal screening

Co-culture to identify TCR: Co-culture 4149-F11 TIL fragment with p53-Y220C long peptide, sorted CD4+41BB+ T cells

Method to identify TCR: Frequency pairing. CD4+41BB+ sorted T cells were expanded in REP (rapid expansion protocol) and the resulting T cell culture was subjected to TCRAD (alpha) and TCRB (beta) deep sequencing by Adaptive Biotechnologies. The top two TCR alphas were paired with the top two TCR betas in a matrix of 4 total TCRs. The second TCR alpha and second TCR beta was the reactive TCR hence TCRA2b2 nomenclature.

Abundance of TCR amongst all TCRs: as below

TCR orientation: alpha-beta

Expression vector: SB transposon

MASAPISMLAMLFTLSGLRAQSVAQPEDQVNVAEGNPLTVKCTYSVSGNPYLFWYV
QYPNRGLQFLLKYITGDNLVKGSYGFEAEFNKSQTSFHLKKPSALVSDSALYFC AVR
VWDYKLSFGAGTTVTVRANIQNPEPAVYQLKDPRSQDSTLCCLFTDFDSQINVPKTMESG
TFITDKCVLDMKAMDSKSNGAIAWSNQTSTFCQDIFKETNATYPSSDVPCDATLTEKSFET
DMNLNFQNLVIVLRILLKLVAGFNLLMTLRLWSSRAKRSGSGATNFSLLKQAGDVEE
NPGPMHNQVLCVVLCCFLGANTVDGGITQSPKYLFRKEGQNVTLSCQNLNHDAM
YWYRQDPGQGLRLIYYSQIVNDFQKGDIAEGYSVSREKKESFPLTVTSAQKNPTAFY
LCASSISAGGDGYTFGSGTRLT VVEDLRNVTPPKVSLFEPKAEIANKQKATLVCLARGF
FPDHVELSWVWNGKEVHSGVCTDPQAYKESNYSYCLSSRLRV SATFWHNP RNHFRCQVQ
FHGLSEEDKWPEGSPKPVTONISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLYAV
LVSTLVMMAMVKKRNS (SEQ ID NO: 565)

[0259] The statistics for TCR **4149TCRa2b2** of Patient 4149 are set forth in Table 20 below.

Table 20

CDR3	Rank of unique, productive CDR3s	CDR3 Counts	CDR3 Frequency
Alpha: CAVRVWDYKLSF (SEQ ID NO: 59)	2 of 4,308	648,707 of 3,309,400	19.6 %
Beta: CASSISAGGDGYTF (SEQ ID NO: 62)	2 of 3,176	104,325 of 578,948	18.0 %

EXAMPLE 7

[0260] This example demonstrates the identification of anti-mutated p53 T cells in Patient 4213 by co-culturing autologous APCs induced to express mutated p53 within autologous T cells ("p53 hotspot mutation universal screening"). This example also demonstrates the isolation of twelve anti-mutated p53 TCRs from patient 4213.

[0261] Experiments were carried out as described for Figures 25-26 for Patient 4213.

[0262] TIL fragments (F2 and F24) from patient 4213 were co-cultured with autologous APCs pulsed with peptide vehicle (DMSO) or purified (>95% by HPLC) 25-amino acid peptides composed of the mutated p53-R248Q sequence. Co-cultures were performed overnight at 37 °C. Expression of 4-1BB was evaluated by flow cytometry after gating for lymphocytes → living cells (PI negative) → CD3+ (T cells). The results are shown in Figure 25. CD8+4-1BB+ T cells were sorted into wells of 96 wells plates. TCRs were identified using single-cell RT-PCR.

[0263] CD4+ T cells came from patient 4213's peripheral blood lymphocytes. The CD4+ T cell culture was co-cultured with autologous APCs pulsed with peptide vehicle (DMSO) or purified (>95% by HPLC) 25-amino acid peptides composed of the mutated p53-R248Q sequence. Co-cultures were performed overnight at 37 °C. Secretion of IFN-γ was evaluated by ELISPOT. Expression of 4-1BB was evaluated by flow cytometry after gating for lymphocytes → living cells (PI negative) → CD3+ (T cells). The results are shown in Figure 26. CD4+4-1BB+ T cells were sorted into wells of 96 wells plates. TCRs were identified using single-cell RT-PCR.

[0264] The sequence of **4213-F2-TCR1**, which was isolated from Patient 4213, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 317), the second underlined region is the CDR2alpha (SEQ ID NO: 318), the third underlined region is the CDR3alpha (SEQ ID NO: 319), the fourth underlined region is the CDR1beta (SEQ ID NO: 320), the fifth underlined region is the CDR2beta (SEQ ID NO: 321), and the sixth underlined region is the CDR3beta (SEQ ID NO: 322). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 323) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 324) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 325) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 326) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0265] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4213-F2-TCR1

Recognition of p53 mutation: R248Q

Screening method: p53 "hotspot" mutation universal screening

Co-culture to identify TCR: Co-culture 4213-F2 with R248Q long peptide, sorted CD8+41BB+ T cells

Method to identify TCR: single-cell RT-PCR

Abundance of TCR amongst all paired TCRs: 8.3% (observed 2 times of 24 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon

MAMLLGASVLILWLQPDWVNSQQKNDDQQVKQNSPSLSVQEGRISILNCDYTNSMF
DYFLWYKKYPAEGPTFLISISSIKDKNEDGRFTVFLNKS AKHLSLHIVPSQPGDSAVYF
CAANTGNQFYFGTGTSLTVIPNIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTME
 SGTFTDKCVLDMKAMDSKSNGAIAWSNQTSFTCQDIFKETNATYPSSDVPCDATLTEKSF
 ETDMNLNFQNLVIVLRILLKLVAGFNLLMTLRLWSS**RAKRSGSGATNFSLLKQAGDV**
EENPGPMHTRLLCWAALCCLGAELTEAGVAQSPRYKIIKRQSVAFWCNPISGHATL
 YWYQQILGQGPKLLIQFNNGVVDDSQLPKDRFSAERLKGVDSTLKIQPAKLEDSAV
 YLCASSHLAGEFYNEQFFGPGTRTLTVLEDLRNVTPPKVSLFEPKAEIANKQKATLVCLA
 RGFFPDHVELSWVWNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSAFWHNP RNHFRC
 QVQFHGLSEEDKWPEGSPKPVTONISAEAWGRADCGITSASYQQGVLSATILYEILLGKAT
 LYAVLVSTLVVMAMVKRKNS (SEQ ID NO: 566)

[0266] The statistics for TCR **4213-F2-TCR1** of Patient 4213 are set forth in Table 21 below.

Table 21

Parameter	#	Frequency
Total wells	96	100%
CDR3alpha	3	3.1%
CDR3beta	8	8.3%
4213-F2-TCR1 pairs	2	2.1%
Total paired TCRs	24	25.0%

[0267] The sequence of **4213-F2-TCR2**, which was isolated from Patient 4213, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 327), the second underlined region is the CDR2alpha (SEQ ID NO: 328), the third underlined region is the CDR3alpha (SEQ ID NO: 329), the fourth underlined region is the CDR1beta (SEQ ID NO: 330), the fifth underlined region is the CDR2beta (SEQ ID NO: 331), and the sixth underlined region is the CDR3beta (SEQ ID NO: 332). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 333)

includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 334) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 335) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 336) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0268] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4213-F2-TCR2

Recognition of p53 mutation: R248Q

Screening method: p53 “hotspot” mutation universal screening

Co-culture to identify TCR: Co-culture 4213-F2 with R248Q long peptide, sorted CD8+41BB+ T cells

Method to identify TCR: single-cell RT-PCR

Abundance of TCR amongst all paired TCRs: 12.5% (observed 3 times of 24 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon

MAGAFLLYVSMKMGGTAGQSLEQPSEVTAVEGAIVQINCTYQTSGFYGLSWYQQH
DGGAPTFLSYNALDGLEETGRFSSFLSRSDSYGYLLLQELQMKDSASYFCAFAAYGQN
FVFGPGTRL SVLPNIQNPEPAVYQLKDP RSQDSTLCLFTDFDSQINVPKTMESGTFITDKC
VLDMKAMDSKSNGAIAWSNQTSFTCQDIFKETNATYPSSDVPCDATLTEKSFETDMNLF
QNLLVIVLRILLKLVAGFNLLMTLRLWSSRAKRS GSGATNFSLLKQAGDVEENPGPMH
FRLCCVAFCLLGAGPVD SGVTQTPKHLITATGQRVTLRCS PRSGDLSVYWYQQSLD
QGLQFLIQYYNGEERAKGNILERFSAQQFPDLHSELNLSSLELGDSALYFCASSPLGD
SGNTIYFGE GSWLTVVEDLRNVTPPKVSLFEPSKAEIANKQKATLVCLARGFFPDHVELS
WWVNGKEVHSGVCTDPQAYKESNYSYCLSSRLRV SATFWHNP RNHFRCQVQFHGLSEED
KWPEGSPK PVTQNISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLYAVLVSTLVVM
AMVKRKNS (SEQ ID NO: 567)

[0269] The statistics for TCR **4213-F2-TCR2** of Patient 4213 are set forth in Table 22 below.

Table 22

Parameter	#	Frequency
Total wells	96	100%
CDR3alpha	4	4.2%
CDR3beta	4	4.2%
4213-F2-TCR2 pairs	3	3.1%
Total paired TCRs	24	25.0%

[0270] The sequence of **4213-F2-TCR3**, which was isolated from Patient 4213, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 337), the second underlined region is the CDR2alpha (SEQ ID NO: 338), the third underlined region is the CDR3alpha (SEQ ID NO: 339), the fourth underlined region is the CDR1beta (SEQ ID NO: 340), the fifth underlined region is the CDR2beta (SEQ ID NO: 341), and the sixth underlined region is the CDR3beta (SEQ ID NO: 342). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 343) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 344) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 345) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 346) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0271] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4213-F2-TCR3

Recognition of p53 mutation: R248Q

Screening method: p53 "hotspot" mutation universal screening

Co-culture to identify TCR: Co-culture 4213-F2 with R248Q long peptide, sorted CD8+41BB+ T cells

Method to identify TCR: single-cell RT-PCR

Abundance of TCR amongst all paired TCRs: 70.8% (observed 17 times of 24 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon

MATLLGVSLVILWLQLARVNSQQGEEDPQALSIQEGENATMNC SYKTSINN LQWYR
 QNSGRGLVHLILIRSNEREKHSGRRLRVTLDTSKSSSLLITASRAADTASYFCATDAW
 NNDMRFGAGTRLTVKPNIQNPEPAVYQLKDP RSQDSTLCLFTDFDSQINVPKTMESGTF
 ITDKCVLDMKAMDSKSNGAIAWSNQTSFTCQDIFKETNATYPSSDVPCDATLTEKSFETD
 MNLNFQNL LVIVLRILLK VAGFNLLMTLRLWSSRAKRSGSGATNFSLLKQAGDVEEN
PGPMHIGLLCCVAFSLLWASPVNAGVTQTPKFQVLKTGQSMTLQCAQDMNHNSMY
 WYRQDPGMGLRLIYYSASEGTTDKGEVPNGYNVSRLNKREFSLRLESAAPSQTSVYF
 CASSESQGNTEAFFGQGTRLTVVEDLRNVTPPKVSLFEPSKAELANKQKATLVCLARGFF
 PDHVELSWVWNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNP RNHFRCQVQF
 HGLSEEDKWPEGSPKPVTONISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLYAVL
 VSTLVVMAMVKRKNS (SEQ ID NO: 568)

[0272] The statistics for TCR **4213-F2-TCR3** of Patient 4213 are set forth in Table 23 below.

Table 23

Parameter	#	Frequency
Total wells	96	100%
CDR3alpha	19	19.8%
CDR3beta	42	43.8%
4213-F2-TCR3 pairs	17	17.7%
Total paired TCRs	24	25.0%

[0273] The sequence of **4213-F24-TCRa1**, which was isolated from Patient 4213, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 347), the second underlined region is the CDR2alpha (SEQ ID NO: 348), the third underlined region is the CDR3alpha (SEQ ID NO: 349), the fourth underlined region is the CDR1beta (SEQ ID NO: 350), the fifth underlined region is the CDR2beta (SEQ ID NO: 351), and the sixth underlined region is the CDR3beta (SEQ ID NO: 352). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 353) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 354) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 355) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 356) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0274] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4213-F24-TCRa1

Recognition of p53 mutation: R248Q

Screening method: p53 “hotspot” mutation universal screening

Co-culture to identify TCR: Co-culture 4213-F24 with R248Q long peptide, sorted CD8+41BB+ T cells

Method to identify TCR: single-cell RT-PCR**Abundance of TCR amongst all paired TCRs: 15.9% (observed 7 times of 44 pairs)****TCR orientation: alpha-beta****Expression vector: SB transposon**

MAKHLTTFLVILWLYFYRGNQVEQSPQSLIILEGKNCTLQCNYTVSPFSNLRWY
 KQDTGRGPVSLTIMTFSENTKSNGRYTATLDADTKQSSLHITASQLSDSASYICVVSS
YKIIFGTGTRLHVFPNIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTMESGTFITD
 KCVLDMKAMDSKSNGAIAWSNQTSTFCQDIFKETNATYPSSDVPCDATLTEKSFETDMNL
 NFQNLVIVLRILLKLVAGFNLLMTLRLWSSRAKRSGSGATNFSLLKQAGDVEENPGP
 MHTRLLFVWVAFCLLGAYHTGAGVVSQSPSNKVTEKGKDVELRCDPISGHTALYWYR
 QRLGQGLEFLIYFQGNAPDKSGLPSDRFSAERTGESVSTLTIQRTQQEDSAVYLCAS
SPIQGENSPLHFGNGTRLTVTEDLRNVTPPKVSLFEPSKAEIANKQKATLVCLARGFFPD
 HVELSWVWNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSAFWHNPRNHFRCQVQFHG
 LSEEDKWPEGSPKPVTONISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLYAVLVST
 LVVMAMVKKRNS (SEQ ID NO: 569)

[0275] The statistics for TCR **4213-F24-TCRa1** of Patient 4213 are set forth in Table 24 below.

Table 24

Parameter	#	Frequency
Total wells	96	100%
CDR3alpha	7	7.3%
CDR3beta	80	83.3%
4213-F24-TCRa1 pairs	7	7.3%
Total paired TCRs	44	45.8%

[0276] The sequence of **4213-F24-TCRa2**, which was isolated from Patient 4213, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 357), the second underlined region is the CDR2alpha (SEQ ID NO: 358), the third underlined region is the CDR3alpha (SEQ ID NO: 359), the fourth underlined region is the CDR1beta (SEQ ID NO: 360), the fifth underlined region is the CDR2beta (SEQ ID NO: 361), and the sixth underlined region is the CDR3beta (SEQ ID NO: 362). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 363) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 364)

includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 365) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 366) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0277] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4213-F24-TCRa2

Recognition of p53 mutation: R248Q

Screening method: p53 “hotspot” mutation universal screening

Co-culture to identify TCR: Co-culture 4213-F24 with R248Q long peptide, sorted CD8+41BB+ T cells

Method to identify TCR: single-cell RT-PCR

Abundance of TCR amongst all paired TCRs: 84.1% (observed 37 times of 44 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon

MAKHLTTFLVILWLYFYRGNGKNQVEQSPQSLIILEGKNCTLQCNYTVSPFSNLRWY
KQDTGRGPVSLTIMTFSENTKSNGRYTATLDADTKQSSLHITASQLSDSASYICVVSS
YKLIFGTGTRLQVFPNIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTMESGTFITD
KCVLDMKAMDSKSNGAIAWSNQTSTFCQDIFKETNATYPSSDVPCDATLTEKSFETDMNL
NFQNLVIVLRILLKLVAGFNLLMTLRLWSSRAKRSGSGATNFSLLKQAGDVEENPGP
MHTRLLFWVAFCLLGAYHTGAGVSQSPSNKVTEKGDVELRCDPISGHTALYWYR
QRLGQGLEFLIYFQGNAPDKSGLPSDRFSAERTGESVSTLTQRTQQEDSAVYLCAS
SPIQGENSPLHFGNGTRLTVTEDLRNVTPPKVSLFEPSKAEIANKQKATLVCLARGFFPD
HVELSWVNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNPРНHFRСQVQFHG
LSEEDKWPEGSPKPVTONISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLYAVLVST
LVVMAMVKRKNS (SEQ ID NO: 570)

[0278] The statistics for TCR 4213-F24-TCRa2 of Patient 4213 are set forth in Table 25 below.

Table 25

Parameter	#	Frequency
Total wells	96	100%
CDR3alpha	39	40.6%
CDR3beta	80	83.3%
4213-F24-TCRa2 pairs	37	38.5%
Total paired TCRs	44	45.8%

[0279] The sequence of **4213-PBL-TCR1**, which was isolated from Patient 4213, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 367), the second underlined region is the CDR2alpha (SEQ ID NO: 368), the third underlined region is the CDR3alpha (SEQ ID NO: 369), the fourth underlined region is the CDR1beta (SEQ ID NO: 370), the fifth underlined region is the CDR2beta (SEQ ID NO: 371), and the sixth underlined region is the CDR3beta (SEQ ID NO: 372). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 373) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 374) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 375) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 376) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0280] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4213-PBL-TCR1

Recognition of p53 mutation: R248Q

Screening method: p53 "hotspot" mutation universal screening

Co-culture to identify TCR: CD4+ Memory T cells after *in vitro* sensitization with R248Q long peptide were co-cultured with R248Q long peptide and CD4+41BB+ T cells were sorted

Method to identify TCR: single-cell RT-PCR

Abundance of TCR amongst all paired TCRs: 9.5% (observed 6 times of 63 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon (University of Minnesota, Minneapolis, MN)

MALLLVPAFQVIFTLGGTRAQSVTQLDSQVPVFEEAPVELRCNYSSSVSVYLFWYVQ
 YPNQGLQLLLKYLSGSTLVESINGFEAEFNKSQTSFHLRKPSVHISDTAEYFCVSKG
TGAQKLVFGQGTRLTINPNIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTMESGT
FITDKCVLDMMKAMDSKSNAGIAWSNQTSFTCQDIFKETNATYPSSDVPCDATLTEKSFETD
 MNLNFQNLVIVLRILLKLVAGFNLLMTLRLWSSRAKRSGSGATNFSLLKQAGDVEEN
PGPMHNQVLCCVVLCFLGANTVDGGITQSPKYLFRKEGQNVTLSCQNLNHDAMY
 WYRQDPGQGLRLIYYSQIVNDFQKGDIAEGYSVSREKKESFPLIVTSAQKNPTASYLC
ASEAFFGQGTRLTVVEDLRNVTPPKVSLFEPKAEIANKQKATLVCLARGFFPDHVELSW
 WYNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSAFWHNP RNHFRCQVQFHGLSEEDK
 WPEGSPKPVTONISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLYAVLVSTLVVMA
 MVKRKNS (SEQ ID NO: 571)

[0281] The statistics for TCR **4213-PBL-TCR1** of Patient 4213 are set forth in Table 26 below.

Table 26

Parameter	#	Frequency
Total wells	192	100%
CDR3alpha	9	4.7%
CDR3beta	6	3.1%
4213-PBL-TCR1 pairs	6	3.1%
Total paired TCRs	63	32.8%

[0282] The sequence of **4213-PBL-TCR2**, which was isolated from Patient 4213, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 377), the second underlined region is the CDR2alpha (SEQ ID NO: 378), the third underlined region is the CDR3alpha (SEQ ID NO: 379), the fourth underlined region is the CDR1beta (SEQ ID NO: 380), the fifth underlined region is the CDR2beta (SEQ ID NO: 381), and the sixth underlined region is the CDR3beta (SEQ ID NO: 382). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 383) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 384) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 385) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 386) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0283] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4213-PBL-TCR2

Recognition of p53 mutation: R248Q

Screening method: p53 “hotspot” mutation universal screening

Co-culture to identify TCR: CD4+ Memory T cells after *in vitro* sensitization with R248Q long peptide were co-cultured with R248Q long peptide and CD4+41BB+ T cells were sorted

Method to identify TCR: single-cell RT-PCR**Abundance of TCR amongst all paired TCRs: 7.9% (observed 5 times of 63 pairs)****TCR orientation: alpha-beta****Expression vector: SB transposon**

MALVTSITVLLSLGIMGDAKTTQPNSMESNEEEPVHLPCNHSTISGTDYIHWYRQLPS
 QGPEYVIHGLTSNVNNRMA SLAIAEDRKSSTLILHRATLRDAAVYYC ILASGAGSYQ
 LTFGKGTKLSVIPNIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTMESGTFITDKC
 VLDMKAMDSKSNGAIAWSNQTSTFCQDIFKETNATYPSSDVPCDATLTEKSFETDMNLNF
 QNLLVIVLRILLKLVAGFNLLMTLRLWSSRAKRSGSGATNFSLLKQAGDVEENPGPMH
 CRLCCVVFCLLQAGPLDTAVSQTPKYLVTQMGNPKSIKCEQNLGHDTMYWYKQD
 SKKFLKIMFSYNNKELIINETVPNRFSPKSPDKAHLNLHINSLELGDSAVYFCASRTIG
 YNTEAFFGQGTRLTVVEDLRNVTPPKVSLFEPSKAEIANKQKATLVCLARGFFPDIIVELS
 WWWNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNP RNHFRCQVQFHGLSEED
 KWPEGSPKPVTQNISA EAWGRADCGITSASYQQGVLSATILYEILLGKATLYAVLVSTLVVM
 AMVKRKNS (SEQ ID NO: 572)

[0284] The statistics for TCR **4213-PBL-TCR2** of Patient 4213 are set forth in Table 27 below.

Table 27

Parameter	#	Frequency
Total wells	192	100%
CDR3alpha	8	4.2%
CDR3beta	8	4.2%
4213-PBL-TCR2 pairs	5	2.6%
Total paired TCRs	63	32.8%

[0285] The sequence of **4213-PBL-TCR3**, which was isolated from Patient 4213, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 387), the second underlined region is the CDR2alpha (SEQ ID NO: 388), the third underlined region is the CDR3alpha (SEQ ID NO: 389), the fourth underlined region is the CDR1beta (SEQ ID NO: 390), the fifth underlined region is the CDR2beta (SEQ ID NO: 391), and the sixth underlined region is the CDR3beta (SEQ ID NO: 392). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 393) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 394)

includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 395) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 396) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0286] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4213-PBL-TCR3

Recognition of p53 mutation: R248Q

Screening method: p53 “hotspot” mutation universal screening

Co-culture to identify TCR: CD4+ Memory T cells after *in vitro* sensitization with R248Q long peptide were co-cultured with R248Q long peptide and CD4+41BB+ T cells were sorted

Method to identify TCR: single-cell RT-PCR

Abundance of TCR amongst all paired TCRs: 6.3% (observed 4 times of 63 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon

MAKIRQFLAILWLQLSCVSAAKNEVEQSPQNLTAEQEGFITINCSYSVGISALHWLQ
QHPGGGIVSLFMLSSGKKKHGRLLIATINIQEKHSSLHITASHPRDSAVYICAALSYNTD
KLIFGTGTRLQVFPNIQNPEPAVYQLKDPQSQDSTLCLFTDFDSQINVPKTMESGTFITDK
CVLDMKAMDSKSNAGIAWSNQTSFTCQDIFKETNATYPSSDVPCDATLTEKSFETDMNLN
FQNLLVIVLRILLKLVAGFNLLMTLRLWSSRAKRSGSGATNFSLLKQAGDVEENPGPM
HTRLLCWAALCLLGADHTGAGVVSQTPSNKVTEKGKYVELRCDPISGHTALYWYRQS
LGQGPEFLIYFQGTGAADDSGLPNDRFFAVRPEGSVSTLKIQRTERGDSAVYLCASSL
SGLLQETQYFGPGTRLLVLEDLRNVTPPKVSLFEPSKAEIANKQKATLVCLARGFFPDHV
ELSWWVNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNPRNHFRCQVQFHGLS
EEDKWPEGSPKPVTONISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLYAVLVSTL
VVMAMVKRKNS (SEQ ID NO: 573)

[0287] The statistics for TCR **4213-PBL-TCR3** of Patient 4213 are set forth in Table 28 below.

Table 28

Parameter	#	Frequency
Total wells	192	100%
CDR3alpha	4	2.1%
CDR3beta	7	3.6%
4213-PBL-TCR3 pairs	4	2.1%
Total paired TCRs	63	32.8%

[0288] The sequence of **4213-PBL-TCR4a1**, which was isolated from Patient 4213, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 397), the second underlined region is the CDR2alpha (SEQ ID NO: 398), the third underlined region is the CDR3alpha (SEQ ID NO: 399), the fourth underlined region is the CDR1beta (SEQ ID NO: 400), the fifth underlined region is the CDR2beta (SEQ ID NO: 401), and the sixth underlined region is the CDR3beta (SEQ ID NO: 402). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 403) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 404) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 405) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 406) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0289] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4213-PBL-TCR4a1

Recognition of p53 mutation: R248Q

Screening method: p53 "hotspot" mutation universal screening

Co-culture to identify TCR: CD4+ Memory T cells after *in vitro* sensitization with R248Q long peptide were co-cultured with R248Q long peptide and CD4+41BB+ T cells were sorted

Method to identify TCR: single-cell RT-PCR

Abundance of TCR amongst all paired TCRs: 3.2% (observed 2 times of 63 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon

MAYSPGLVSLILLLLGRTRGNSVTQMEGPVTLSEEAFLTINCTYTATGYPSLFWYVQ
YPGEGLQLLLKATKADDDKGSNKGFEATYRKETTSFHLEKGSVQVSDSAVYFCALSH
TGSSNTGKLIFGQGTRLQVKPNIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTM
SGTFITDKCVLDMKAMDSSKNGAIAWSNQTSTFCQDIFKETNATYPSSDVPCDATLTEKSF
ETDMNLFQNLVIVLRILLKVGAFNLLMTLRWSS**RAKRSGSGATNFSLLKQAGDV**
EENPGPMHTRLLCWAALCLLGAELTEAGVAQSPRYKIIKRQSVAFWCNPISGHATL
YWYQQILGQGPKLLIQFQNGVVDDSQLPKDRFSAERLKGVDSTLKIQAILEDASV
YLCASSTGGGRHQPHQFGDGRLSILEDLRNVTPPKVSLFEPKAEIANKQKATLVCLAR
GFFPDHVELSWVNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNPРНHFRСQ

*VQFHGLSEEDKWPEGSPKPVTQNISAEAWGRADCGITSASYQOGVLSATILYEILLGKATLY
AVLVSTLVVMAMVKRKNS* (SEQ ID NO: 574)

[0290] The statistics for TCR **4213-PBL-TCR4a1** of Patient 4213 are set forth in Table 29 below.

Table 29

Parameter	#	Frequency
Total wells	192	100%
CDR3alpha	2	1.0%
CDR3beta	36	18.8%
4213-PBL-TCR4a1 pairs	2	1.0%
Total paired TCRs	63	32.8%

[0291] The sequence of **4213-PBL-TCR4a2**, which was isolated from Patient 4213, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 407), the second underlined region is the CDR2alpha (SEQ ID NO: 408), the third underlined region is the CDR3alpha (SEQ ID NO: 409), the fourth underlined region is the CDR1beta (SEQ ID NO: 410), the fifth underlined region is the CDR2beta (SEQ ID NO: 411), and the sixth underlined region is the CDR3beta (SEQ ID NO: 412). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 413) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 414) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 415) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 416) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0292] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4213-PBL-TCR4a2

Recognition of p53 mutation: R248Q

Screening method: p53 “hotspot” mutation universal screening

Co-culture to identify TCR: CD4+ Memory T cells after *in vitro* sensitization with R248Q long peptide were co-cultured with R248Q long peptide and CD4+41BB+ T cells were sorted

Method to identify TCR: single-cell RT-PCR

Abundance of TCR amongst all paired TCRs: 3.2% (observed 2 times of 63 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon

MAYSPGLVSLILLLLGRTRGNSVTQMEGPVTLSEEAFLTINCTYATATGYPSLFWYVQ
YPGEGLQLLLKATKADDKGSNKGFEATYRKETTSFHLEKGSVQVSDSAVYFCALSQ
TGSSKTGKLIFGQGTRLQVKPNIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTM
SGTFITDKCVLDMKAMDSKSNGAIAWSNQTSTFCQDIFKETNATYPSSDVPCDATLTEKSF
ETDMNLNFQNLVIVLRILLKLVAGFNLLMTLRLWSSRAKRSGSGATNFSLLKQAGDV
EENPGPMHTRLLCWAALCCLLGAELTEAGVAQSPRYKIIKRQSVAFWCNPISGHATL
YWYQQILGQGPKLLIQFNNGVVDDSQLPKDRFSAERLKGVDSTLKIQPAKLEDSAV
YLCASSTGGGRHQPHFGDGTRLSELEDLRNVTTPPKVSLFEPKAEIANKQKATLVCLAR
GFFPDHVELSWWVNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNPRNHFCQ
VQFHGLSEEDKWPEGSPKPTQNISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLY
AVLVSTLVMMAMVKRKNS (SEQ ID NO: 575)

[0293] The statistics for TCR **4213-PBL-TCR4a2** of Patient 4213 are set forth in Table 30 below.

Table 30

Parameter	#	Frequency
Total wells	192	100%
CDR3alpha	2	1.0%
CDR3beta	36	18.8%
4213-PBL-TCR4a2 pairs	2	1.0%
Total paired TCRs	63	32.8%

[0294] The sequence of **4213-PBL-TCR4a3**, which was isolated from Patient 4213, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 417), the second underlined region is the CDR2alpha (SEQ ID NO: 418), the third underlined region is the CDR3alpha (SEQ ID NO: 419), the fourth underlined region is the CDR1beta (SEQ ID NO: 420), the fifth underlined region is the CDR2beta (SEQ ID NO: 421), and the sixth underlined region is the CDR3beta (SEQ ID NO: 422). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable

region (SEQ ID NO: 423) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 424) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 425) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 426) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0295] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4213-PBL-TCR4a3

Recognition of p53 mutation: R248Q

Screening method: p53 “hotspot” mutation universal screening

Co-culture to identify TCR: CD4+ Memory T cells after *in vitro* sensitization with R248Q long peptide were co-cultured with R248Q long peptide and CD4+41BB+ T cells were sorted

Method to identify TCR: single-cell RT-PCR

Abundance of TCR amongst all paired TCRs: 4.8% (observed 3 times of 63 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon

MAYSPGLVSLILLLLGRTRGNSVTQMEGPVTLSEEAFLTINCTYTATGYPSLFWYVQ
YPGEGLQLLLKATKADDKGSNKGFEATYRKETTTFHLEKGSVQVSDSAVYFCALSQ
TGSSNTGKLIFGQGTRLQVKPNIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTME
SGTFTDKCVLDMKAMDSKSNAGIAWSNQTSFTCQDIFKETNATYPSSDVPCDATLTEKSF
ETDMNLNFNQLLVIVLRILLKLVAGFNLLMTLRLWSSRAKRSGSGATNFSLLKQAGDV
EENPGPMHTRLLCWAALCLLGAELTEAGVAQSPRYKIIKRQSVAFWCNPISGHATL
YWYQQILGQGPKLLIQFNNGVVDDSQLPKDRFSAERLKGVDSTLKIQPAKLEDSAV
YLCASSTGGGRHQPHFGDGTLSILEDLRNVTPPKVSLFEPKAEIANKQKATLVCLAR
GFFPDHVELSWVWNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNPNNHFRQ
VQFHGLSEEDKWPEGSPKPVTONISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLY
AVLVSTLVVMAMVKKRNS (SEQ ID NO: 576)

[0296] The statistics for TCR **4213-PBL-TCR4a3** of Patient 4213 are set forth in Table 31 below.

Table 31

Parameter	#	Frequency
Total wells	192	100%
CDR3alpha	4	2.1%

Parameter	#	Frequency
CDR3beta	36	18.8%
4213-PBL-TCR4a3 pairs	3	1.6%
Total paired TCRs	63	32.8%

[0297] The sequence of **4213-PBL-TCR4a4**, which was isolated from Patient 4213, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 427), the second underlined region is the CDR2alpha (SEQ ID NO: 428), the third underlined region is the CDR3alpha (SEQ ID NO: 429), the fourth underlined region is the CDR1beta (SEQ ID NO: 430), the fifth underlined region is the CDR2beta (SEQ ID NO: 431), and the sixth underlined region is the CDR3beta (SEQ ID NO: 432). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 433) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 434) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 435) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 436) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0298] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4213-PBL-TCR4a4

Recognition of p53 mutation: R248Q

Screening method: p53 "hotspot" mutation universal screening

Co-culture to identify TCR: CD4+ Memory T cells after *in vitro* sensitization with R248Q long peptide were co-cultured with R248Q long peptide and CD4+41BB+ T cells were sorted

Method to identify TCR: single-cell RT-PCR

Abundance of TCR amongst all paired TCRs: 3.2% (observed 2 times of 63 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon

MAYSPGLVSLILLLLGRTRGNSVTQMEGPVTLSEEAFLTINCTYTATGYPSLFWYVQ
YPGEGLQLLLKATKADDKGSNKGFEATYRKETTFSHLEKGSVQVSDSAVYFCALSTT
GSSNTGKLIFGQGTTLQVKPNIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTMES

GTFITDKCVLDMKAMDSKSNGAIAWSNQTSTFCQDIFKETNATYPSSDVPCDATLTEKSFE
 TDMNLNFQNLVIVLRILLKLVAGFNLLMTLRWSSRAKRSGSGATNFSLLKQAGDVE
 ENPGPMHTRLLCWAALCLLGAELTEAGVAQSPRYKIIKQSVAFWCNPISGHATLY
 WYQQILGQGPKLLIQFQNGVVDSDQLPKDRFSAERLKGVDSTLKIQPAKLEDSAVY
 LCASSTGGGRHQPHFQNGDTRLSELDLRNVTPPKVSLFEPKAEIANKQKATLVCLARG
 FFPDHVELSWWVNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNPVNHFRQV
 QFHGLSEEDKWPEGSPKPVTONISAEAWGRADCGITSASYQOGVLSATILYEILLGKATLYA
 VLVSTLVVMAMVKRKNS (SEQ ID NO: 577)

[0299] The statistics for TCR **4213-PBL-TCR4a4** of Patient 4213 are set forth in Table 32 below.

Table 32

Parameter	#	Frequency
Total wells	192	100%
CDR3alpha	2	1.0%
CDR3beta	36	18.8%
4213-PBL-TCR4a3 pairs	2	1.0%
Total paired TCRs	63	32.8%

EXAMPLE 8

[0300] This example demonstrates the identification of anti-mutated p53 T cells in Patient 4268 by co-culturing autologous APCs induced to express mutated p53 within autologous T cells (“p53 hotspot mutation universal screening”). This example also demonstrates the isolation of five anti-mutated p53 TCRs from patient 4268.

[0301] Experiments were carried out as described for Figures 27-30 for Patient 4268.

[0302] TIL fragments (F1-F24, n=24) from patient 4268 were co-cultured with autologous APCs electroporated with TMG composed of irrelevant, WT p53, or mutated p53 sequence. Co-cultures were performed overnight at 37 °C. Secretion of IFN- γ was evaluated using ELISPOT assay. The results are shown in Figure 27.

[0303] TIL fragments (F1-F24, n=24) from patient 4268 were co-cultured with autologous APCs pulsed with peptide vehicle (DMSO) or purified (>95% by HPLC) 25-amino acid peptides composed of WT p53-R248 sequence or mutated p53-R248Q sequence. Co-cultures were performed overnight at 37 °C. Secretion of IFN- γ was evaluated using ELISPOT assay. The results are shown in Figure 28.

[0304] TIL fragments (F1-F24, n=24) from patient 4268 were co-cultured with autologous APCs pulsed with peptide vehicle (DMSO) or purified (>95% by HPLC) 25-

amino acid peptides composed of wt p53-R248 sequence or mutated p53-R248Q sequence. Co-cultures were performed overnight at 37 °C. Expression of 4-1BB was evaluated by flow cytometry after gating for lymphocytes → living cells (PI negative) → CD3+ (T cells). The results are shown in Figure 29. TIL fragments F18 and F19 were sources of TCRs after sorting CD4+41BB+ T cells.

[0305] TIL fragments (F1-F24, n=24) from patient 4268 were co-cultured with autologous APCs pulsed with peptide vehicle (DMSO) or purified (>95% by HPLC) 25-amino acid peptides composed of WT p53-R248 sequence or mutated p53-R248Q sequence. Co-cultures were performed overnight at 37 °C. Expression of 4-1BB was evaluated by flow cytometry after gating for lymphocytes → living cells (PI negative) → CD3+ (T cells). The results are shown in Figure 30. TIL fragments F7, F8, and F15 were sources of TCRs after sorting CD8+4-1BB+ T cells.

[0306] The sequence of **4268-TCR1**, which was isolated from Patient 4268, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 137), the second underlined region is the CDR2alpha (SEQ ID NO: 138), the third underlined region is the CDR3alpha (SEQ ID NO: 139), the fourth underlined region is the CDR1beta (SEQ ID NO: 140), the fifth underlined region is the CDR2beta (SEQ ID NO: 141), and the sixth underlined region is the CDR3beta (SEQ ID NO: 142). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 143) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 144) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 145) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 146) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0307] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4268-TCR1

Recognition of p53 mutation: R248Q

Method: p53 “hotspot” mutation universal screening

Co-culture to identify TCR: Co-culture 4268-F7 and 4268-F8 with p53-R248Q long peptide, sorted CD8+41BB+ T cells

Method to identify TCR: single-cell RT-PCR

Abundance of TCR amongst all paired TCRs: 78.7% (observed 48 times of 61 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon

MASLRVLLVILWLQLSWVWSQQKEVEQNSGPLSVPEGAIASLNCTYSDRGSQSFFW
 YRQYSGKSPELIMFIYSNGDKEDGRFTAQLNKASQYVSLIRDSQPSDSATYLCAVS
WYSTLTFGKGTMLLVSPNIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTMESGT
 FITDKCVLDMKAMDSKSNGAIAWSNQTSFTCQDIFKETNATYPSSDVPCDATLTEKSFETD
 MNLNFQNLVIVLRILLKVAGFNLLMTLRLWSSRAKRSGSGATNFSLLKQAGDVEEN
 PGPMTLRLFFYVALCLLWAGHRDAGITQSPRYKITETGRQVTLMCHQTWSHSYMF
 WYRQDLGHGLRLIYYSAADITDKGEVPDGYVVSRSKTENFPLTLESATRSQTSVYF
 CASSGSRTDTQYFGPGTRLTVLEDLRNVTPPKVSLFEPKAEIANKQKATLVCLARGFFP
 DHVELSWWVNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNPРНHFRQVQFH
 GLSEEDKWPEGSPKPVTONISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLYAVLV
 STLVVMAMVKRKNS (SEQ ID NO: 578)

[0308] The statistics for TCR **4268-TCR1** of Patient 4268 are set forth in Table 33 below.

Table 33

Parameter	#	Frequency
Total wells	96	100%
CDR3alpha	54	56.3%
CDR3beta	71	74.0%
4268-TCR1 pairs	48	50.0%
Total paired TCRs	61	63.5%

[0309] The sequence of **4268-TCR2**, which was isolated from Patient 4268, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 147), the second underlined region is the CDR2alpha (SEQ ID NO: 148), the third underlined region is the CDR3alpha (SEQ ID NO: 149), the fourth underlined region is the CDR1beta (SEQ ID NO: 150), the fifth underlined region is the CDR2beta (SEQ ID NO: 151), and the sixth underlined region is the CDR3beta (SEQ ID NO: 152). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 153) includes the sequence starting from the amino terminus and ending immediately prior to the

start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 154) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 155) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 156) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0310] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4268-TCR2

Recognition of p53 mutation: R248Q

Method: p53 “hotspot” mutation universal screening

Co-culture to identify TCR: Co-culture 4268-F7 and 4268-F8 with p53-R248Q long peptide, sorted CD8+41BB+ T cells

Method to identify TCR: single-cell RT-PCR

Abundance of TCR amongst all paired TCRs: 6.6% (observed 4 times of 61 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon

MALKFSVSILWQLAWVSTQLLEQSPQFLSIQEGENLTVYCNSSSVFSSLQWYRQEPG
EGPVLLVTVTGGEVKKLRKLTQFGDARKDSSLHITAAQPGDTGLYLCAGEFAGN
QFYFGTGTSLTVIPNIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTMESGTFITDK
CVLDMKAMDSKSNGLAWSNQTSTFCQDIFKETNATYPSSDVPCDATLTEKSFETDMNLN
FQNLVIVLRILLKLVAGFNLLMTLRLWSSRAKRSKSGGATNFSLLKQAGDVEENPGPM
HCRLLCCAVLCLLGAVPIDTEVTQTPKHLVGMNTNKKSLKCEQHMGHRAMYWYK
QKAKKPELMFVYSYEKLSINESVPSRFSPECNSSLNLHLHALQPEDSALYLCASS
QVGLTYEQYFGPGTRLTVTEDLRNVTPPKVSLFEPSKAEIANKQKATLVCLARGFFPDH
VELSWWVNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNP RNHFRCQVQFHGL
SEEDKWPEGSPKPVTONISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLYAVLVSTL
VVMAMVKRKNS (SEQ ID NO: 579)

[0311] The statistics for TCR **4268-TCR2** of Patient 4268 are set forth in Table 34 below.

Table 34

Parameter	#	Frequency
Total wells	96	100%
CDR3alpha	6	6.3%
CDR3beta	4	4.2%
4268-TCR2 pairs	4	4.2%
Total paired TCRs	61	63.5%

[0312] The sequence of **4268-TCR3**, which was isolated from Patient 4268, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 157), the second underlined region is the CDR2alpha (SEQ ID NO: 158), the third underlined region is the CDR3alpha (SEQ ID NO: 159), the fourth underlined region is the CDR1beta (SEQ ID NO: 160), the fifth underlined region is the CDR2beta (SEQ ID NO: 161), and the sixth underlined region is the CDR3beta (SEQ ID NO: 162). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 163) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 164) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 165) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 166) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0313] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4268-TCR3

Recognition of p53 mutation: R248Q

Method: p53 "hotspot" mutation universal screening

Co-culture to identify TCR: Co-culture 4268-F15 with p53-R248Q long peptide, sorted CD8+41BB+ T cells

Method to identify TCR: single-cell RT-PCR

Abundance of TCR amongst all paired TCRs: 88.6% (observed 31 times of 35 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon

MASAPISMLAMLFTLSGLRAQSVAQPEDQVNVAEGNPLTVKCTYSVSGNPYLFWYV
 QYPNRGLQFLLKYITGDNLVKGSYGFEAEFNKSQTSFHLKKPSALVSDSALYFC AVR
 DNSGGSNYKLTFGKGTLLTVNPNIQNPEPAVYQLKDPRS QDSTLCLFTDFDSQINVPKT
 MESGTFITDKCVLDMKAMDSKSNGAIAWSNQTSTFCQDIFKETNATYPSSDVPCDATLTE
 KSFETDMNLFQNLVIVLRILLKLVAGFNLLMTLRLWSSRAKRSGSGATNFSLLKQAG
DVEENPGPMHTRLLCWAALCCLLGAELTEAGVAQSPRYKIIKRQSVAFWCNPISGH
 ATLYWYQQILGQGPKLLIQFQNNGVVDDSQLPKDRFSAERLKGVDSTLKIQPAKLED
 SAVYLCASSLGQGQTQYFGPGTRLLVLEDLRNVTTPKVSLFEPSKAEIANKQKATLVCL
 ARGFFPDHVELSWVNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNPRNHFR
 CQVQFHGLSEEDKWPEGSPKPTQNISAEAWGRADCGITSASYQQGVLSATILYEILLGKA
 TLYAVLVSTLVVMAMVKRKNS (SEQ ID NO: 580)

[0314] The statistics for TCR **4268-TCR3** of Patient 4268 are set forth in Table 35 below.

Table 35

Parameter	#	Frequency
Total wells	96	100%
CDR3alpha	42	43.8%
CDR3beta	37	38.5%
4268-TCR3 pairs	31	32.3%
Total paired TCRs	35	36.5%

[0315] The sequence of **4268-TCR4**, which was isolated from Patient 4268, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 167), the second underlined region is the CDR2alpha (SEQ ID NO: 168), the third underlined region is the CDR3alpha (SEQ ID NO: 169), the fourth underlined region is the CDR1beta (SEQ ID NO: 170), the fifth underlined region is the CDR2beta (SEQ ID NO: 171), and the sixth underlined region is the CDR3beta (SEQ ID NO: 172). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 173) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 174) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 175) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 176) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0316] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4268-TCR4

Recognition of p53 mutation: R248Q

Method: p53 “hotspot” mutation universal screening

Co-culture to identify TCR: Co-culture 4268-F18 with p53-R248Q long peptide, sorted CD4+41BB+ T cells

Method to identify TCR: single-cell RT-PCR**Abundance of TCR amongst all paired TCRs: 95.2% (observed 40 times of 42 pairs)****TCR orientation: alpha-beta****Expression vector: SB transposon**

MALLLVPAFQVIFTLGGTRAQSVTQLDSQVPVFEEAPVELRCNYSSSVSVYLFWYVQ
 YPNQGLQLLLKYLSGSTLVESINGFEAEFNKSQTSFHLRKPSVHISDTAEYFCAVRGSS
 GTYKYIFGTGTRLKVLANIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTMESGTF
 ITDKCVLDMKAMDSDKSNAGIAWSNQTSTFCQDIFKETNATYPSSDVPCDATLTEKSFETD
 MNLNFQNLVIVLRILLKLVAGFNLLMTLRLWSSRAKRSGSGATNFSLLKQAGDVEEN
 PGP MHIRLLCRVAFCLAVGLVDVKVTQSSRYLVKRTGEKVFLECVQDMDHENMF
 WYRQDPGLGLRLIYFSYDVKMKEKGDIPGYSVSREKKERFSLILESASTNQTSMYL
 CASKGDQNTAEFFGQGTRLTVVEDLRNVTPPKVSLFEPKAEIANKQKATLVCLARGFF
 PDHVELSWVNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNP RNHFRCQVQF
 HGLSEEDKWPEGSPKPVTONISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLYAVL
 VSTLVVMAMVKKRNS (SEQ ID NO: 581)

[0317] The statistics for TCR **4268-TCR4** of Patient 4268 are set forth in Table 36 below.

Table 36

Parameter	#	Frequency
Total wells	96	100%
CDR3alpha	43	44.8%
CDR3beta	53	55.2%
4268-TCR4 pairs	40	41.7%
Total paired TCRs	42	43.8%

[0318] The sequence of **4268-TCR5**, which was isolated from Patient 4268, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 177), the second underlined region is the CDR2alpha (SEQ ID NO: 178), the third underlined region is the CDR3alpha (SEQ ID NO: 179), the fourth underlined region is the CDR1beta (SEQ ID NO: 180), the fifth underlined region is the CDR2beta (SEQ ID NO: 181), and the sixth underlined region is the CDR3beta (SEQ ID NO: 182). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 183) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 184)

includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 185) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 186) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0319] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4268-TCR5

Recognition of p53 mutation: R248Q

Method: p53 “hotspot” mutation universal screening

Co-culture to identify TCR: Co-culture 4268-F19 with p53-mut-TMG, sorted CD4+41BB+ T cells

Method to identify TCR: single-cell RT-PCR

Abundance of TCR amongst all paired TCRs: 11.8% (observed 2 times of 17 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon

MAGVFLLYVSMKMGTTGQNIDQPTMTATEGAIVQINCTYQTSGFNGLFWYQQH
AGEAPTFLSYNVLDGLEEKGRFSSFLSRSGYSYLLKELQMKDSASYLCAVRDLQT
GANNLFFGTGTRLTVIPNIQNPEPAVYQLKDPQSQDSTLCLFTDFDSQINVPKTMESGTFI
TDKCVLDMKAMDSKSNAGIAWSNQTSFTCQDIFKETNATYPSSDVPCDATLTEKSFETDM
NLNFQNLVIVLRILLKLVAGFNLLMTLRLWSSRAKRSGSGATNFSLLKQAGDVEENP
GPMHIRLLCRVAFCLAVGLVDVKVTQSSRYLVKRTGKVFLECVQDMDHENMFW
YRQDPGLGLRLIYFSYDVKMKEKGDIEGYSVSREKKERFSLILESASTNQTSMYLCA
SSLTFGTTEAFFGQGTTRLTVVEDLRNVTTPPKVSLFEPSKAEIANKQKATLVCLARGFFPD
HVELSWVWNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSAFWHNPNNHFRQCQVQFHG
LSEEDKWPEGSPKPVTONISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLYAVLVST
LVVMAMVVRKNS (SEQ ID NO: 582)

[0320] The statistics for TCR 4268-TCR5 of Patient 4268 are set forth in Table 37 below.

Table 37

Parameter	#	Frequency
Total wells	96	100%
CDR3alpha	2	2.1%
CDR3beta	87	90.6%
4268-TCR5 pairs	2	2.1%
Total paired TCRs	17	17.7%

EXAMPLE 9

[0321] This example demonstrates the identification of anti-mutated p53 T cells in Patient 4266 by co-culturing autologous APCs induced to express mutated p53 within autologous T cells (“p53 hotspot mutation universal screening”). This example also demonstrates the isolation of four anti-mutated p53 TCRs from patient 4266.

[0322] Experiments were carried out as described for Figures 31-34 and 53-55 for Patient 4266.

[0323] TIL fragments (F1-F24, n=24) from patient 4266 were co-cultured with autologous APCs electroporated with TMG composed of irrelevant, WT p53, or mutated p53 sequence. Co-cultures were performed overnight at 37 °C. Secretion of IFN- γ was evaluated using ELISPOT assay. The results are shown in Figure 31. Expression of 4-1BB was evaluated by flow cytometry after gating for lymphocytes $\rightarrow$ living cells (PI negative) $\rightarrow$ CD3+ (T cells). The results are shown in Figure 32.

[0324] TIL fragments (F1-F24, n=24) from patient 4266 were co-cultured with autologous APCs pulsed with peptide vehicle (DMSO) or purified (>95% by HPLC) 25-amino acid peptides composed of WT p53-R248 sequence or mutated p53-R248W sequence. Co-cultures were performed overnight at 37 °C. Secretion of IFN- γ was evaluated using ELISPOT assay. The results are shown in Figure 33. Expression of 4-1BB was evaluated by flow cytometry after gating for lymphocytes $\rightarrow$ living cells (PI negative) $\rightarrow$ CD3+ (T cells). The results are shown in Figure 34.

[0325] Cos7 cells (2.5×10^4 per well) were plated on wells of flat-bottom 96 well plates. The following day, cells were co-transfected with individual HLA alleles from patient 4266. The next day cells were pulsed with no peptide, DMSO, wild type p53-R248 peptide SSCMGGMNRR (SEQ ID NO: 590) or mutated p53-R248W peptide SSCMGGMNWR (SEQ ID NO: 591) for 2 hours at 37 °C at 1 μ g/mL. TIL cultures from patient 4266 (10^5) were added to wells and co-cultured overnight at 37°C. Expression of 4-1BB was evaluated by flow cytometry after gating for lymphocytes $\rightarrow$ living cells (PI negative) $\rightarrow$ CD3+ (T cells) $\rightarrow$ CD4-CD8+. The results are shown in Fig. 53.

[0326] T cells expressing mock (no TCR), 4266-TCR1, 4266-TCR2, 4266-TCR3 or 4266-TCR4 with putative specificity to p53-R248W identified from 4266-TIL were co-cultured with autologous APCs which were pulsed with peptide vehicle (DMSO) or purified (>95% by HPLC) 25 amino acid peptides composed of WT p53-R248 sequence or mutated p53-R248W sequences. Media alone and PMA and Ionomycin were negative and positive

controls, respectively. Co-cultures were performed overnight at 37 °C. Expression of 4-1BB was evaluated by flow cytometry after gating for lymphocytes → living cells (PI negative) → CD3+ (T cells) → CD4-CD8+. The results are shown in Fig. 54.

[0327] A tumor cell (TC) line was established from a xenografted tumor fragment resected from Patient 4266 then serially passaged through immunocompromised mice (TC#4266). The TC#4266 was co-cultured with T cells (10^5) expressing mock (no TCR) or p53-R248W-specific TCRs (4266-TCR2, 4266-TCR3 or 4266-TCR4) overnight at 37°C. The TC#4266 cells were either incubated with nothing, W6/32 pan-HLA Class-I specific blocking antibody, IVA12 pan-HLA Class-II specific blocking antibody or mutated p53-R248W peptide SSCMGGMNWR (SEQ ID NO: 591) for 2 hours at 37 °C. The antibodies were kept in the co-culture at 5 µg/mL final concentration. The peptide was incubated at 1 µg/mL and excess peptide was washed after incubation. Media alone (no TC) and PMA and Ionomycin were negative and positive controls, respectively. Expression of 4-1BB was evaluated by flow cytometry after gating for lymphocytes → living cells (PI negative) → CD3+ (T cells) → CD4-CD8+. The results are shown in Fig. 55.

[0328] The sequence of **4266-TCR1**, which was isolated from Patient 4266, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 97), the second underlined region is the CDR2alpha (SEQ ID NO: 98), the third underlined region is the CDR3alpha (SEQ ID NO: 99), the fourth underlined region is the CDR1beta (SEQ ID NO: 100), the fifth underlined region is the CDR2beta (SEQ ID NO: 101), and the sixth underlined region is the CDR3beta (SEQ ID NO: 102). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 103) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 104) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 105) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 106) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0329] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4266-TCR1

Recognition of p53 mutation: R248W

Screening method: p53 “hotspot” mutation universal screening

Co-culture to identify TCR: Co-culture 4266-F1, 4266-F3, 4266-F5 and 4266-F6 with p53mutTMG or R248W long peptide (both co-cultures detected the same TCR), sorted CD8+41BB+ T cells

Method to identify TCR: single-cell RT-PCR

Abundance of TCR amongst all paired TCRs: 17.0% (observed 9 times of 53 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon

MALLLVPAFQVIFTLGGTRAQSVTQLDSQVPVFEEAPVELRCNYSSSVSVYLFWYVQ
YPNQGLQLLKYLSGSTLVESINGFEAEFNKSQTSFHLRKPSVHISDTAEYFCAVSDL
VRDDKIIFGKGTRLHILPNIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTMESGTF
ITDKCVLDMKAMDSKSNAGIAWSNQTSTFCQDIFKETNATYPSSDVPCDNLTEKSFETD
MNLNFQNLVIVLRILLKLVAGFNLLMTLRLWSSRAKRSGSGATNFSLLKQAGDVEEN
PGPMHIGLLCCVAFSLLWASPVNAGVTQTPKFQVLKTGQSMTLQCAQDMNHNSMY
WYRQDPGMGLRLIYYSASEGTTDKGEVPNGYNVSRNLNREFSLRLESAAPSQTSVYF
CASIGGFEEAFFGQGTRLTVVEDLRNVTPPKVSLFEPSKAEIANKQKATLVCLARGFFPDH
VELSWVWNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNPNNHFRQVQVQFHGL
SEEDKWPEGSPKPVTONISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLYAVLVSTL
VVMAMVKRKNS (SEQ ID NO: 583)

[0330] The statistics for TCR **4266-TCR1** of Patient 4266 are set forth in Table 38 below.

Table 38

Parameter	#	Frequency
Total wells	96	100%
CDR3alpha	9	9.4%
CDR3beta	10	10.4%
4266-TCR1 pairs	9	9.4%
Total paired TCRs	53	55.2%

[0331] The sequence of **4266-TCR2**, which was isolated from Patient 4266, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 107), the second underlined region is the CDR2alpha (SEQ ID NO: 108), the third underlined region is the CDR3alpha (SEQ ID NO: 109), the fourth underlined region is the CDR1beta (SEQ ID NO: 110), the fifth underlined region is the CDR2beta (SEQ ID NO: 111), and the sixth underlined region is the CDR3beta (SEQ ID NO: 112). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is

the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 113) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 114) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 115) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 116) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0332] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4266-TCR2

Recognition of p53 mutation: R248W

Screening method: p53 “hotspot” mutation universal screening

Co-culture to identify TCR: Co-culture 4266-F1, 4266-F3, 4266-F5 and 4266-F6 with p53mutTMG or R248W long peptide (both co-cultures detected the same TCR), sorted CD8+41BB+ T cells

Method to identify TCR: single-cell RT-PCR

Abundance of TCR amongst all paired TCRs: 24.5% (observed 13 times of 53 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon

MAGVFLLYVSMKMGGTTGQNIDQPTMTATEGAIVQINCTYQTSGFNGLFWYQQH
AGEAPTFLSYNVLDGLEEKGRFSSFLSRSGYSYLLKELQMKDSASYLCAVYTGGF
KTIFGAGTRLFVKANIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTMESGTFITD
KCVLDMKAMD SKSNGALAWSNQTSFTCQDIFKETNATYPSSDVPCD ATLTEKSFETDMNL
NFQNLVIVLRILLKLVAGFNLLMTLRLWSSRAKRSGSGATNFSLLKQAGDVEENPGP
MHTRLLCWAALCLLGAELTEAGVAQSPRYKIIKRQSVAFWCNPISGHATLYWYQQ
ILGQGPKLLIQFQNGGVVDDSQLPKDRFSAERLKGVDSTLKIQPAKLEDSAVYLCAS
NLGGGSTDTQYFGPGTRLTVLEDLRNVTPPKVSLFEP SKAEIANKQKATLVCLARGFFP
DHVELSWWWNGKEVHSGVCTDPQAYKESNYSYCLSSRLRV SATFWHNP RNHFRCQVQFH
GLSEEDKWPEGSPKPVTONISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLYAVLV
STLVVMAMVKRKNS (SEQ ID NO: 584)

[0333] The statistics for TCR **4266-TCR2** of Patient 4266 are set forth in Table 39 below.

Table 39

Parameter	#	Frequency
Total wells	96	100%

Parameter	#	Frequency
CDR3alpha	21	21.9%
CDR3beta	18	18.8%
4266-TCR2 pairs	13	13.5%
Total paired TCRs	53	55.2%

[0334] The sequence of **4266-TCR3**, which was isolated from Patient 4266, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 117), the second underlined region is the CDR2alpha (SEQ ID NO: 118), the third underlined region is the CDR3alpha (SEQ ID NO: 119), the fourth underlined region is the CDR1beta (SEQ ID NO: 120), the fifth underlined region is the CDR2beta (SEQ ID NO: 121), and the sixth underlined region is the CDR3beta (SEQ ID NO: 122). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 123) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 124) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 125) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 126) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0335] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4266-TCR3

Recognition of p53 mutation: R248W

Screening method: p53 “hotspot” mutation universal screening

Co-culture to identify TCR: Co-culture 4266-F1, 4266-F3, 4266-F5 and 4266-F6 with p53mutTMG or R248W long peptide (both co-cultures detected the same TCR), sorted CD8+41BB+ T cells

Method to identify TCR: single-cell RT-PCR

Abundance of TCR amongst all paired TCRs: 34.0% (observed 18 times of 53 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon

MAGVFLLYVSMKMGGTTGQNI^DQPT^EMTAT^EGAI^VQIN^CTYQTSGFNGLFWYQQH
AGEAPTFLSYNVLDGLEEKGRFSSFLSRSGYSYLLKELQMKDSASYLCAFYYGGS
QGNLIFGKGTKLSVKPNIQNPEPAVYQLK^DPRSQDSTLCLFTDFDSQINVPKTMESGTFIT

DKCVLDMKAMDSKSNGAIAWSNQTSFTCQDIFKETNATYPSSDVPCDATLTEKSFETDMN
*LNFQNLVIVLRILLKLVAGFNLLMTLRLWSS**RAKRSGSGATNFSLLKQAGDVEENPG***
PMHTRLLCWVVLGFLGTDHTGAGVSQSPRYKVAKRGQDVALRCDPISGHVSLFWY
QQALGQGPEFLTYFQNEAQLDKSGLPSDRFFAERPEGSVSTLKIQR**TQQEDSAVYLC**
ASSFGSGSTDTQYFGPGTRLTVLEDLRNVTPPKVSLFEP**SKAEIANKQKATLVCLARGFF**
PDHVELSWVNGKEVHSGVCTDPQAYKESNYSYCLSSRLRV**SATFWHNPRNHFRCQVQF**
HGLSEEDKWPEGSPKPVTONISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLYAVL
VSTLVVMAMVKKRNS (SEQ ID NO: 585)

[0336] The statistics for TCR **4266-TCR3** of Patient 4266 are set forth in Table 40 below.

Table 40

Parameter	#	Frequency
Total wells	96	100%
CDR3alpha	19	19.8%
CDR3beta	26	27.1
4266-TCR3 pairs	18	18.8%
Total paired TCRs	53	55.2%

[0337] The sequence of **4266-TCR4**, which was isolated from Patient 4266, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 127), the second underlined region is the CDR2alpha (SEQ ID NO: 128), the third underlined region is the CDR3alpha (SEQ ID NO: 129), the fourth underlined region is the CDR1beta (SEQ ID NO: 130), the fifth underlined region is the CDR2beta (SEQ ID NO: 131), and the sixth underlined region is the CDR3beta (SEQ ID NO: 132). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 133) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 134) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 135) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 136) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0338] Cancer reactive T cells were identified using the screening method set forth below. The method used to isolate the TCR is set forth below.

TCR name: 4266-TCR4

Recognition of p53 mutation: R248W

Screening method: p53 “hotspot” mutation universal screening

Co-culture to identify TCR: Co-culture 4266-F1, 4266-F3, 4266-F5 and 4266-F6 with p53mutTMG or R248W long peptide (both co-cultures detected the same TCR), sorted CD8+41BB+ T cells

Method to identify TCR: single-cell RT-PCR

Abundance of TCR amongst all paired TCRs: 9.4% (observed 5 times of 53 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon

MAGVFLLYVSMKMGTTGQNIDQPTMTATEGAIVQINCTYQTSGFNGLFWYQQH
AGEAPTFLSYNVLDGLEEKGRFSSFLSRSGYSYLLKELQMKDSASYLCAVYPGGS
QGNLIFGKGTKLSVKPNIQNPEPAVYQLKDPQSQDSTLCLFTDFDSQINVPKTMESGTFIT
DKCVLDMKAMDSKSNGAIAWSNQTSFTCQDIFKETNATYPSSDVPCDATLTEKSFETDMN
LNFQNLVIVLRILLKLVAGFNLLMTLRLWSSRAKRSGSGATNFSLLKQAGDVEENPG
PMHTRLLCWAALCLLGAELTEAGVAQSPRYKIIKRQSVAFWCNPISGHATLYWYQ
QILGQGPKLLIQFNNGVVDSDQLPKDRFSAERLKGVDSTLKIQPAKLEDSAVYLCA
SSLGTGSTDTQYFGPGTRLTVLEDLRNVTPPKVSLFEPKAEIANKQKATLVCLARGFFP
DHVELSWVNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNPVHFRQCQVQFH
GLSEEDKWPEGSPKPVTONISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLYAVLV
STLVVMAMVKRKNS (SEQ ID NO: 586)

[0339] The statistics for TCR **4266-TCR4** of Patient 4266 are set forth in Table 41 below.

Table 41

Parameter	#	Frequency
Total wells	96	100%
CDR3alpha	5	5.2%
CDR3beta	13	13.5
4266-TCR3 pairs	5	5.2%
Total paired TCRs	53	55.2%

EXAMPLE 10

[0340] This example demonstrates a summary of responses to p53 “hotspot” mutations by T cells.

[0341] A summary of responses to p53 “hotspot” mutations by T cells is provided in Table 42. Numbers 1-15 of Table 42 were a retrospective study. Numbers 16-33 of Table 42 were a prospective study.

TABLE 42

#	Tumor type	Patient	p53 mut	T cell screen	HLA
1	Gastric	3446	G245S	n/a	n/a
2	Gastroesophageal	3788	Y220C	N	n/a
3	Rectal	3942	R273C	n/a	n/a
4	Colon	4023	R282W	Y	Class-II
5	Colon	4095	R282W	n/a	n/a
6	Ovarian	4127	G245S	Y	DRB3*02:02
7	Breast	4130	R273H	Y	Class-II
8	Colon	4141	R175H	Y	A*02:01
9	Ovarian	4149	Y220C	Y	DRB3*02:02
10	Colon	4160	R273H	N	n/a
11	Melanoma	4165	G245D	N	n/a
12	Colon	4166	R248W	n/a	n/a
13	Rectal	4171	R248Q	N	n/a
14	Melanoma	4187	R273H	N	n/a
15	Colon	4196	R175H	Y	A*02:01
16	Colon	4213	R248Q	Y	both
17	Colon	4217	R175H	N	n/a
18	Cholangiocarcinoma	4220	R248Q	N	n/a
19	Rectal	4235	R273C	N	n/a
20	Colon	4238	R248Q	Y	Class-I
21	Colon	4244	R282W	N	n/a
22	Colon	4245	R248Q	N	n/a
23	Colon	4252	R175H	N	n/a
24	Melanoma	4253	R248W	Y	unknown
25	Colon	4254	R273H	N	n/a
26	Colon	4257	R248W	N	n/a
27	Endometrial	4258	R273H	N	n/a

#	Tumor type	Patient	p53 mut	T cell screen	HLA
28	Colon	4259	Y220C	Y	A*02:01, DRB1*04
29	Colon	4266	R248W	Y	A*68:01
30	Colon	4268	R248Q	Y	both
31	Pancreatic	4270	R282W	Y	unknown
32	Rectal	4273	R248W	Y	DPB1*02:01
33	Rectal	4274	R175H	n/a	
34	Colon	4283	R175H	N	n/a
35	Colon	4285	R175H	Y	DRB1*13:01
36	Colon	4287	R248W	N	n/a
37	Colon	4312	R175H	N	n/a

n/a=not applicable; N=negative; Y=confirmed reactive; TBS=to be screened.

EXAMPLE 11

[0342] This example demonstrates the treatment of patients with p53 mutation-reactive TIL.

[0343] A summary of the treatment of patients with p53 mutation-reactive TIL is provided in Table 43.

TABLE 43

#	Tumor Type	Patient	p53 mut	Infusion bag screening	Total # of Cells ($\times 10^9$)	% of p53 reactive TIL	# of p53 reactive TIL ($\times 10^9$)	Response	Duration (months)
1	Ovarian	4127	G245S	Y	143	2.8	4.0	P.R.	4
2	Colon	4141	R175H	Y	69	0.8	0.6	N.R.	-
3	Colon	4196	R175H	Y	92	3.3	3.0	N.R.	-
4	Ovarian	4149	Y220C	Y	37	11.1	4.1	N.R.	-
5	Colon	4213	R248Q	TBS	33			P.R.	4
6	Colon	4238	R248Q	TBS	57			N.R.	

#	Tumor Type	Patient	p53 mut	Infusion bag screening	Total # of Cells ($\times 10^9$)	% of p53 reactive TIL	# of p53 reactive TIL ($\times 10^9$)	Response	Duration (months)
7	Colon	4266	R248W	Y	104	50.8	52.8		
8	Colon	4268	R248Q	**TBS					
9	Rectal	4273	R248W	**Y	117	6.8	8.0	N.R.	-
10	Colon	4285	R175H	Y	69.6	2.4	1.7	N.R.	-

NT=not treated; TBS=to be screened; N.R.=no response; P.R.=partial response; **patient not yet treated.

EXAMPLE 12

[0344] This example demonstrates the isolation and specific reactivity of a TCR from patient 4141.

[0345] Autologous APCs were transfected with TMG encoding irrelevant mutations, WT p53 sequence, or mutated p53 sequence including R175H. Media alone and PMA and ionomycin were negative and positive controls, respectively. The following day, TIL from patient 4141 (fragment culture 12) were co-cultured overnight at 37 °C with TMG-transfected APCs. Secretion of IFN- γ was evaluated by ELISPOT. Expression of 4-1BB was evaluated by flow cytometry after gating for lymphocytes $\rightarrow$ living cells (PI negative) $\rightarrow$ CD3+ (T cells) $\rightarrow$ CD4-CD8+. The results are shown in Figure 38.

[0346] Cos7 cells (2.5×10^4 per well) were plated on wells of flat-bottom 96 well plates. The following day, cells were co-transfected with individual HLA alleles from patient 4141 and either no extra gene, WT TP53 TMG, or mutated TP53 TMG containing the p53-R175H sequence. TIL with specificity to p53-R175H from Patient 4141 (fragment culture 12) were co-cultured the following day with transfected Cos7 cells and were incubated overnight at 37 °C. Secretion of IFN- γ was evaluated by ELISPOT. The results are shown in Figure 39.

[0347] T cells expressing mock (no TCR) or 4141-TCR1a2 were co-cultured with T2 tumor cells (expressing HLA-A*02:01). T2 cells were pulsed for 2 hours at 37 °C with peptide vehicle (DMSO) or purified (>95% by HPLC) peptides composed of WT p53-R175 peptide HMTEVVRRC (SEQ ID NO: 532) or mutated p53-R175H peptide HMTEVVRHC

(SEQ ID NO: 530). Media alone and PMA and Ionomycin were negative and positive controls, respectively. Co-cultures were performed overnight at 37 °C. Secretion of IFN- γ was evaluated by ELISA. The results are shown in Figure 40.

[0348] T cells expressing 4141-TCR1a2 were co-cultured overnight at 37 °C with Saos2 cells (p53-NUL and HLA-A*02:01+), which were either unmanipulated or made to overexpress full length p53-R175H protein. Inhibitors of secretion (monensin and brefeldin A) were added to co-cultures to trap cytokines within T cells. After 6 hours of co-culture, cells were fixed and permeabilized then stained for IL-2, CD107a, IFN- γ and tumor necrosis factor-alpha (TNF α). Flow cytometry was used to analyze co-cultures based on a lymphocyte gate. The results are shown in Figure 41.

[0349] The sequence of TCR **4141-TCR1a2**, which was isolated from Patient 4141, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 467), the second underlined region is the CDR2alpha (SEQ ID NO: 468), the third underlined region is the CDR3alpha (SEQ ID NO: 469), the fourth underlined region is the CDR1beta (SEQ ID NO: 470), the fifth underlined region is the CDR2beta (SEQ ID NO: 471), and the sixth underlined region is the CDR3beta (SEQ ID NO: 472). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 473) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 474) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 475) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 476) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0350] Cancer reactive T cells were identified as described below. The TCR was isolated as described below.

TCR name: 4141-TCR1a2

Recognition of p53 mutation: R175H

Screening method: p53 “hotspot” mutation universal screening

Co-culture to identify TCR: Co-culture 4141 infusion bag TIL with p53mutTMG and sorted CD8+41BB+ T cells

Method to identify TCR: single-cell RT-PCR then TA TOPO cloning kit (Thermo Fisher Scientific, Waltham, MA) for alpha chain

TCR orientation: alpha-beta

Expression vector: SB transposon

MASIRAVFIFLWLQLDLVNGENVEQHPSTLSVQEGDSAVIKCTYSDSASNYFPWYKQ
ELGKGPQLIIDIRSNVGEKKDQRIAVTLNKTAKHFSLHITETQPEDSAVYFCAASKSAI
MVVLQTSSSLELALCLLSSQVNIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTM
SGTFITDKCVLDMKAMDSDSKSNGAIAWSNQTSTFCQDIFKETNATYPSSDVPCDATLTEKSF
ETDMNLFQNLVIVLRILLKLVAGFNLLMTLRLWSSRAKRS~~SG~~GATNFSLLKQAGDV
EENPGPMHPQLLGYVVLCLLGAGPLEAQVTQNPRYLITVTGKCLTVTCSQNMNHEY
MSWYRQDPGLGLRQIYYSMNVEVTDKGDVPEGYKVSKEKRNFLILESPSPNQTS
YFCASSIQQGADTQYFGPGTRLTVLEDLRNVTPPKVSLFEPKAEIANKQKATLVCLARG
FFPDHVELSWVNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNP RNHFRCQV
QFHGLSEEDKWPEGSPKPVTONISAEAWGRADCGITSASYQQGVLSATILYEILLGKATLYA
VLVSTLVVMAMVKRKNS (SEQ ID NO: 587)

[0351] The statistics for 4141-TCR1a2 from patient 4141 are set forth in Table 44 below.

TABLE 44

Parameter	#	Frequency
Total wells	96	100%
CDR3alpha	Unknown (TA TOPO cloning)	Not applicable
CDR3beta	58	60.4%

EXAMPLE 13

[0352] This example demonstrates the isolation and specific reactivity of a TCR isolated from patient 4259.

[0353] TIL fragment culture (no. 6) from patient 4259 was co-cultured with autologous APCs either (1) electroporated with TMG composed of irrelevant, WT p53, or mutated p53 sequence or (2) pulsed with peptide vehicle (DMSO) or purified (>95% by HPLC) 25-amino acid peptides composed of WT p53-Y220 sequence or mutated p53-Y220C sequence. Co-cultures were performed overnight at 37 °C. Expression of 4-1BB was evaluated by flow cytometry after gating for lymphocytes → living cells (PI negative) → CD3+ (T cells). The results are shown in Figure 42.

[0354] Autologous APCs were pulsed with decreasing concentrations of 25-amino acid peptides corresponding to the WT p53-Y220 or mutated p53-Y220C sequence for 2 hours at 37 °C. TIL fragment culture (no. 6) from patient 4259 was co-cultured with peptide-pulsed

APCs. Expression of 4-1BB was assayed by flow cytometry after gating lymphocytes → living cells (PI negative) → CD3+ (T cells). The results are shown in Figure 43.

[0355] Autologous antigen presenting cells were pulsed with DMSO, WT p53-Y220 peptide RNTFRHSVVPYE (SEQ ID NO: 533) or mutated p53-Y220C peptide RNTFRHSVVPCE (SEQ ID NO: 534) for 2 hours at 37 °C. Excess peptide was washed away. TIL from patient 4259 (fragment culture 6) with specificity to p53-Y220C was co-cultured overnight at 37 °C with peptide-pulsed APCs. Expression of 4-1BB was evaluated by flow cytometry after gating for lymphocytes → living cells (PI negative) → CD3+ (T cells). The results are shown in Figure 44.

[0356] Cos7 cells (2.5×10^4 per well) were plated on wells of flat-bottom 96 well plates. The following day, cells were co-transfected with individual HLA alleles from patient 4259. The next day, DMSO or the p53-Y220C peptide RNTFRHSVVPCE (SEQ ID NO: 534) were pulsed for 2 hours on transfected Cos7 cells. Excess peptide was washed away. TIL fragment culture no.6 from Patient 4259 was added (10^5 cells/well). Co-cultures were incubated overnight at 37 °C. Expression of 4-1BB was assayed by flow cytometry after gating lymphocytes → live → CD3+ (T cells) → CD8-CD4+. The results are shown in Figure 45.

[0357] APCs autologous to Patient 4259 were pulsed with 25-amino acid peptides corresponding to the WT p53-Y220 or mutated p53-Y220C sequence for 2 hours at 37 °C. Excess peptide was washed away. T cells expressing 4259-F6-TCR were co-cultured overnight at 37 °C with peptide-pulsed APCs. Expression of 4-1BB was evaluated by flow cytometry after gating for lymphocytes → living cells (PI negative) → CD3+ (T cells). The introduced TCR was measured by mouse TCRbeta (mTCR). The results are shown in Table 45.

TABLE 45

	WT p53-Y220	Mut p53-Y220C
4-1BB+/mTCR+	0.27	5.22
4-1BB-/mTCR-	68.5	73.7
4-1BB+/mTCR-	0.18	1.59
4-1BB-/mTCR+	31.0	19.5

[0358] A tumor cell (TC) line was established from a xenografted tumor fragment resected from Patient 4259 and then serially passaged through immunocompromised mice

(TC#4259). TC#4259 was co-cultured with T cells (10^5) expressing mock (no TCR) or p53-Y220C-specific TCR (4259-F6-TCR) overnight at 37 °C. The TC#4259 cells were either incubated with nothing, W6/32 pan-HLA Class-I specific blocking antibody, IVA12 pan-HLA Class-II specific blocking antibody or mutated p53-Y220C peptide RNTFRHSVVVPCE (SEQ ID NO: 534) for 2 hours at 37 °C. The antibodies were kept in the co-culture at 5 µg/mL final concentration. The peptide was incubated at 10 µg/mL and excess peptide was washed after incubation. Media alone (no TC) and PMA and Ionomycin were negative and positive controls, respectively. Expression of 4-1BB was evaluated by flow cytometry after gating for lymphocytes → living cells (PI negative) → CD3⁺ (T cells). The results are shown in Figure 46.

[0359] The sequence of TCR **4259-F6-TCR**, which was isolated from Patient 4259, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 477), the second underlined region is the CDR2alpha (SEQ ID NO: 478), the third underlined region is the CDR3alpha (SEQ ID NO: 479), the fourth underlined region is the CDR1beta (SEQ ID NO: 480), the fifth underlined region is the CDR2beta (SEQ ID NO: 481), and the sixth underlined region is the CDR3beta (SEQ ID NO: 482). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 483) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 484) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 485) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 486) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0360] Cancer reactive T cells were identified as described below. The TCR was isolated as described below.

TCR name: 4259-F6-TCR

Recognition of p53 mutation: Y220C

Screening method: p53 “hotspot” mutation universal screening

Co-culture to identify TCR: Co-culture 4259-F6 with p53-Y220C peptide and sorted CD4⁺41BB⁺ T cells

Method to identify TCR: single-cell RT-PCR

Abundance of TCR amongst all paired TCRs: 81.1% (observed 36 times of 44 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon

MASLRVLLVILWLQLSWVWSQQKEVEQNSGPLSVPEGAIASLNCTYSDRGSQSFFW
 YRQYSGKSPELIMFIYSNGDKEDGRFTAQLNKASQYVSLIRDSQPSDSATYLCAWN
 SGGSNYKLTFGKGTLT VNPNIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTMES
 GTFITDKCVLDMKAMDSKSNGAIAWSNQTSTFCQDIFKETNATYPSSDVPCDATLTEKSFE
 TDMNLNFQNLVIVLRILLKLVAGFNLLMTLRLWSSRAKRSGSGATNFSLLKQAGDVE
 ENPGPMHLGLLCCGAFSLLWAGPVNAGVTQTPKFRVLKTGQSM TLLCAQDMNHEY
 MYWYRQDPGMGLRLIHYSVGEGTTAKGEVPDGYNVSRLLKQNFLLGLESAAPSQTS
 VYFCASSYSQAWGQPQHFGDGT RL SILEDLRNVTPPKVSLFEP SKAEIANKQKATLVCL
 ARGFFPDHVELSWWWNGKEVHSGVCTDPQAYKESNYSYCLSSRLRV SATFWHNP RNHFR
 CQVQFHGLSEEDKWPEGSPKPVTONISAEAWGRADCGITSASYQQGVLSATILYEILLGKA
 TLYAVLVSTLVVMAMVKRKNS (SEQ ID NO: 588)

[0361] The statistics for **4259-F6-TCR** from patient 4259 are set forth in Table 46 below.

TABLE 46

Parameter	#	Frequency
Total wells	96	100%
CDR3alpha	41	42.7%
CDR3beta	47	49.0%
4259-F6-TCR pairs	36	37.5%
Total paired TCRs	44	45.8%

EXAMPLE 14

[0362] This example demonstrates the isolation and specific reactivity of a TCR from patient 4285.

[0363] TIL fragments (F1-F22 and F24, n=23) from patient 4285 were co-cultured with autologous APCs electroporated with TMG composed of irrelevant, WT p53, or mutated p53 sequence. Co-cultures were performed overnight at 37 °C. Secretion of IFN- γ was evaluated using ELISPOT assay. The results are shown in Figure 47. Expression of 4-1BB was evaluated by flow cytometry after gating for lymphocytes $\rightarrow$ living cells (PI negative) $\rightarrow$ CD3+ (T cells). The results are shown in Figure 49.

[0364] TIL fragments (F1-F22 and F24, n=23) from patient 4285 were co-cultured with autologous APCs pulsed with peptide vehicle (DMSO) or purified (>95% by HPLC) 25-amino acid peptides composed of WT p53-R175 sequence or mutated p53-R175H sequence. Co-cultures were performed overnight at 37 °C. Secretion of IFN- γ was evaluated using

ELISPOT assay. The results are shown in Figure 48. Expression of 4-1BB was evaluated by flow cytometry after gating for lymphocytes → living cells (PI negative) → CD3+ (T cells). The results are shown in Figure 50. TIL fragment F6 was the source of 4285-TCR1 after sorting CD4+41BB+ T cells.

[0365] Autologous APCs were pulsed with 15-amino acid peptides from the p53-R175H sequence (amino acid substitution underlined in Table 45) overlapping 14 amino acids. TIL from patient 4285 (fragment cultures 10, 6 and 9) with specificity to p53-R175H were co-cultured overnight at 37 °C with peptide-pulsed APCs. DMSO was peptide vehicle. Secretion of IFN-γ was evaluated using ELISPOT assay. The results are shown in Table 47.

TABLE 47

		ELISPOT Result Positive (+) or negative (-) for IFN-γ production		
Peptide	SEQ ID NO:	4285-F10	4285-F6	4285-F9
None (vehicle)	Not applicable	-	-	-
YKQSQHMTEVVR <u>H</u> CP	519	-	-	-
KQSQHMTEVVR <u>H</u> CPH	520	-	-	+
QSQHMTEVVR <u>H</u> CPHH	521	-	+	+
SQHMTEVVR <u>H</u> CPHHE	522	+	+	+
QHMTEVVR <u>H</u> CPHHER	523	+	+	+
HMTEVVR <u>H</u> CPHHERC	524	+	+	+
MTEVVR <u>H</u> CPHHERCS	525	+	+	+
TEVVR <u>H</u> CPHHERCSD	526	+	+	+
EVVR <u>H</u> CPHHERCSDS	527	+	+	+
VVR <u>H</u> CPHHERCSDSD	528	+	+	-
VR <u>H</u> CPHHERCSDSDG	529	+	+	-

[0366] Cos7 cells (2.5×10^4 per well) were plated on wells of flat-bottom 96 well plates. The following day, cells were co-transfected with individual HLA alleles from patient 4285. The next day cells were pulsed with DMSO or mutated p53-R175H peptide YKQSQHMTEVVRHCPHHERCSDSDG (SEQ ID NO: 2) for 2 hours at 37 °C at 10 µg/mL. Selected TIL fragment cultures with specificity to p53-R175H from Patient 4285 (4285-F6, 4285-F9 and 4285-F10) were co-cultured with transfected Cos7 cells overnight at 37 °C.

Expression of 4-1BB was assayed by flow cytometry after gating lymphocytes → live → CD3+ (T cells) → CD8-CD4+. The results are shown in Figure 51.

[0367] Autologous APCs were pulsed with decreasing concentrations of 25- or 15-amino acid peptides corresponding to the WT or mutated p53-R175H sequence for 2 hours at 37 °C. T cells transposed with 4285-TCR1 from patient 4285 were co-cultured overnight at 37 °C with peptide-pulsed APCs. Expression of 4-1BB was assayed by flow cytometry after gating lymphocytes → live → CD3+ (T cells) → CD8-CD4+. The results are shown in Figure 52.

[0368] The sequence of **4285-TCR1**, which was isolated from Patient 4285, is set forth below. Starting from the amino terminus, the first underlined region is the CDR1alpha (SEQ ID NO: 487), the second underlined region is the CDR2alpha (SEQ ID NO: 488), the third underlined region is the CDR3alpha (SEQ ID NO: 489), the fourth underlined region is the CDR1beta (SEQ ID NO: 490), the fifth underlined region is the CDR2beta (SEQ ID NO: 491), and the sixth underlined region is the CDR3beta (SEQ ID NO: 492). The bold region is the linker (SEQ ID NO: 26). Starting from the amino terminus, the first italicized region is the alpha chain constant region (SEQ ID NO: 23) and the second italicized region is the beta chain constant region (SEQ ID NO: 25). The alpha chain variable region (SEQ ID NO: 493) includes the sequence starting from the amino terminus and ending immediately prior to the start of the alpha chain constant region. The beta chain variable region (SEQ ID NO: 494) includes the sequence starting immediately after the linker and ending immediately prior to the start of the beta chain constant region. The full-length alpha chain (SEQ ID NO: 495) includes the sequence starting from the amino terminus and ending immediately prior to the start of the linker. The full-length beta chain (SEQ ID NO: 496) includes the sequence starting immediately after the linker and ending with the carboxyl terminus.

[0369] Cancer reactive T cells were identified as described below. The TCR was isolated as described below.

TCR name: 4285-TCR1

Recognition of p53 mutation: R175H

Screening method: p53 “hotspot” mutation universal screening

Co-culture to identify TCR: Co-culture 4285-F6 with p53-R175H peptide and sorted CD4+41BB+ T cells

Method to identify TCR: single-cell RT-PCR

Abundance of TCR amongst all paired TCRs: 81.8% (observed 36 times of 44 pairs)

TCR orientation: alpha-beta

Expression vector: SB transposon

MAKNPLAAPLLILWFHLDCVSSILNVEQSPQSLHVQEGDSTNFTCSFPSSNFYALHWY
 RWETAKSPEALFVMTLNGDEKKKGRISATLNTKEGYSYLYIKGSQPEDSATYLCALI
 TGGGNKLTFGTGTQLKVELNIQNPEPAVYQLKDPRSQDSTLCLFTDFDSQINVPKTMES
 GTFITDKCVLDMKAMDSKSNGAIAWSNQTSTFCQDIFKETNATYPSSDVPCDATLTEKSFE
 TDMNLNFQNLVIVLRILLKLVAGFNLLMTLRLWSSRAKRSGSGATNFSLLKQAGDVE
ENPGPMHLGLLCCGAFSLLWAGPVNAGVTQTPKFRVLKTGQSM TLLCAQDMNHEY
 MYWYRQDPGMGLRLIHYSVGEGTTAKGEVPDGYNVSRLLKKQNFLLGLESAAPSQTS
 VYFCASRLQGWNSPLHFGNGTRLTVTEDLRNVTPPKVSLFEPKAEIANKQKATLVCLA
 RGFFPDHVELSWVWNGKEVHSGVCTDPQAYKESNYSYCLSSRLRVSATFWHNP RNHFR
 QVQFHGLSEEDKWPEGSPKPVTONISAEAWGRADCGITSASYQQGVLSATILYEILLGKAT
 LYAVLVSTLVVMAMVKKRNS (SEQ ID NO: 589)

[0370] The statistics for 4285-TCR1 from patient 4285 are set forth in Table 48.

TABLE 48

Parameter	#	Frequency
Total wells	96	100%
CDR3alpha	37	39.8%
CDR3beta	39	40.6%
4285-TCR1 pairs	36	37.5%
Total paired TCRs	44	45.8%

EXAMPLE 15

[0371] This example demonstrates the specific reactivity of the three anti-mutated p53 TCRs from patient 4196 of Example 2.

[0372] The expression of HLA-A*0201 and p53 R175H by various target cell lines is presented in Table 49.

TABLE 49

Cell line	HLA-A*0201 reported	HLA-A2 by FACS MFI (MFI isotype)	P53 R175H reported	Estimated P53 R175H copy # per 1e5 GAPDH
KLE	+	+ 46733 (2038)	+	167
SKUT1	+	- 1009 (1009)	+	308
TYKNU	+	+ 6556 (1537)	+	175
TYKNU CpR	+	+ 8611 (1511)	+	179
VMRCL CD	+	+ 8340 (949)	+	43
SKBR3	+	weak 3414 (2546)	+	99
CEM/C1	-	- 387 (330)	+	138
HCC139 5	-	- 1743 (1537)	+	105

Cell line	HLA-A*0201 reported	HLA-A2 by FACS MFI (MFI isotype)	P53 R175H reported	Estimated P53 R175H copy # per 1e5 GAPDH
KMS26	+	+ 57284 (679)	+	87
AU565	+	weak 3839 (1870)	+	80
LS123	-	- 1916 (2009)	+	190
CCRF CEM	-	- 235 (177)	+	149
SAOS2 R175H	+	+ 51049 (1417)	+	N/A
SAOS Parental	+	+ 40003 (1272)	-	22 (detection limit)
media	Not applicable (n/a)	n/a	n/a	n/a
PMA/lo	n/a	n/a	n/a	n/a

[0373] The target cells of Table 49 were co-cultured with cells that were transduced with one of the TCRs of Example 2. Mock-transduced cells (no TCR) were used as control effector cells. IFN- γ secretion (pg/mL) (Table 50) and 4-1BB expression (% 4-1BB (of mTCR β +)) (Table 51) were measured.

TABLE 50

Cell line	TP53 TCR AV38/BV10 CD8+	TP53 TCR AV12/BV6 CD8+	TP53 TCR AV6/BV11 CD8+	mock TCR
KLE	<u>741</u>	110	<u>71</u>	51
SKUT1	28	24	26	38
TYKNU	196	41	27	33
TYKNU CpR	<u>298</u>	65	34	24
VMRCLCD	23	22	23	23
SKBR3	26	23	24	26
CEM/C1	26	23	22	28
HCC1395	25	23	26	24
KMS26	<u>472</u>	<u>168</u>	95	23
AU565	21	30	25	28
LS123	25	23	25	26
CCRF CEM	23	23	30	30
SAOS2 R175H	<u>>2000</u>	<u>1064</u>	<u>190</u>	24
SAOS Parental	29	26	26	25
media	25	26	26	25
PMA/lo	<u>1166</u>	<u>485</u>	<u>98</u>	<u>161</u>

TABLE 51

Cell line	TP53 TCR AV38/BV10 CD8+	TP53 TCR AV12/BV6 CD8+	TP53 TCR AV6/BV11 CD8+	mock TCR
KLE	<u>91.5</u>	<u>80.9</u>	<u>84.3</u>	2.2
SKUT1	2.6	2.5	2.5	8.3
TYKNU	<u>70.4</u>	<u>67.3</u>	<u>56.4</u>	2.7
TYKNU CpR	<u>87.7</u>	<u>78.5</u>	<u>74.7</u>	2.8
VMRCLC D	3.0	2.8	8.0	6.3
SKBR3	1.5	2.4	2.1	5.2
CEM/C1	2.6	3.5	5.0	4.1
HCC1395	1.5	2.3	3.4	2.6
KMS26	<u>65.7</u>	<u>58.7</u>	<u>61.5</u>	6.9
AU565	2.0	2.0	2.3	5.6
LS123	2.0	2.7	3.2	4.6
CCRF CEM	3.9	5.6	6.2	2.4
SAOS2 R175H	<u>89.3</u>	<u>88.4</u>	<u>88.5</u>	5.5
SAOS Parental	2.3	0.0	11.0	5.2

EXAMPLE 16

[0374] This example demonstrates a summary of the reactivity of TCRs of Examples 1-15.

[0375] The TCRs of Table 52 were isolated, expressed in T cells and tested against the relevant antigen. A summary of the results is shown in Table 52.

TABLE 52

TCR name	p53 a.a. substitution	TCR tested?	p53 mutation reactive?
4127-TP53-G245S-TCR1	G245S	yes	yes
4127-TP53-G245S-TCR4	G245S	yes	yes
4127_O102_TCR	G245S	yes	yes
4149TCRa2b2	Y220C	yes	yes
4196_AV12-1_with_BV6-1	R175H	yes	yes

TCR name	p53 a.a. substitution	TCR tested?	p53 mutation reactive?
4196_AV38-1_with_BV10-3	R175H	yes	yes
4196_AV6_with_BV11-2	R175H	yes	yes
4253-TIL-TCR1	R248W	yes	no
4253-TIL-TCR2	R248W	yes	no
4266-TCR1	R248W	yes	no
4266-TCR2	R248W	yes	yes
4266-TCR3	R248W	yes	yes
4266-TCR4	R248W	yes	yes
4268-TCR1	R248Q	yes	no
4268-TCR2	R248Q	yes	no
4268-TCR3	R248Q	yes	no
4268-TCR4	R248Q	yes	no
4268-TCR5	R248Q	yes	no
4273-TP53-R248W-TCR1a1	R248W	yes	no
4273-TP53-R248W-TCR1a2	R248W	yes	Yes
4141-TCR1a2	R175H	yes	yes
4285-TCR1	R175H	yes	yes
4259-F6-TCR	Y220C	yes	yes
4127-O37-TCR	G245S	yes	yes

[0376] All references, including publications, patent applications, and patents, cited herein are hereby incorporated by reference to the same extent as if each reference were

individually and specifically indicated to be incorporated by reference and were set forth in its entirety herein.

[0377] The use of the terms “a” and “an” and “the” and “at least one” and similar referents in the context of describing the invention (especially in the context of the following claims) are to be construed to cover both the singular and the plural, unless otherwise indicated herein or clearly contradicted by context. The use of the term “at least one” followed by a list of one or more items (for example, “at least one of A and B”) is to be construed to mean one item selected from the listed items (A or B) or any combination of two or more of the listed items (A and B), unless otherwise indicated herein or clearly contradicted by context. The terms “comprising,” “having,” “including,” and “containing” are to be construed as open-ended terms (i.e., meaning “including, but not limited to,”) unless otherwise noted. Recitation of ranges of values herein are merely intended to serve as a shorthand method of referring individually to each separate value falling within the range, unless otherwise indicated herein, and each separate value is incorporated into the specification as if it were individually recited herein. All methods described herein can be performed in any suitable order unless otherwise indicated herein or otherwise clearly contradicted by context. The use of any and all examples, or exemplary language (e.g., “such as”) provided herein, is intended merely to better illuminate the invention and does not pose a limitation on the scope of the invention unless otherwise claimed. No language in the specification should be construed as indicating any non-claimed element as essential to the practice of the invention.

[0378] Preferred embodiments of this invention are described herein, including the best mode known to the inventors for carrying out the invention. Variations of those preferred embodiments may become apparent to those of ordinary skill in the art upon reading the foregoing description. The inventors expect skilled artisans to employ such variations as appropriate, and the inventors intend for the invention to be practiced otherwise than as specifically described herein. Accordingly, this invention includes all modifications and equivalents of the subject matter recited in the claims appended hereto as permitted by applicable law. Moreover, any combination of the above-described elements in all possible variations thereof is encompassed by the invention unless otherwise indicated herein or otherwise clearly contradicted by context.

CLAIM(S):

1. An isolated or purified T cell receptor (TCR) having antigenic specificity for mutated human p53.
2. The TCR of claim 1, wherein the TCR has antigenic specificity for human p53 with a mutation at position 175, 220, 245, or 248 of SEQ ID NO: 1.
3. The TCR of claim 1, wherein the TCR has antigenic specificity for human p53 with one of the following human p53 mutations: R175H, Y220C, G245S, R248Q, or R248W.
4. The TCR of any one of claims 1-3, wherein the TCR comprises the amino acid sequences of:
 - (1) all of SEQ ID NOs: 27-32;
 - (2) all of SEQ ID NOs: 37-42;
 - (3) all of SEQ ID NOs: 47-52;
 - (4) all of SEQ ID NOs: 57-62;
 - (5) all of SEQ ID NOs: 67-72;
 - (6) all of SEQ ID NOs: 77-82;
 - (7) all of SEQ ID NOs: 87-92;
 - (8) all of SEQ ID NOs: 97-102;
 - (9) all of SEQ ID NOs: 107-112;
 - (10) all of SEQ ID NOs: 117-122;
 - (11) all of SEQ ID NOs: 127-132;
 - (12) all of SEQ ID NOs: 137-142;
 - (13) all of SEQ ID NOs: 147-152;
 - (14) all of SEQ ID NOs: 157-162;
 - (15) all of SEQ ID NOs: 167-172;
 - (16) all of SEQ ID NOs: 177-182;
 - (17) all of SEQ ID NOs: 187-192;
 - (18) all of SEQ ID NOs: 197-202;
 - (19) all of SEQ ID NOs: 207-212;

- (20) all of SEQ ID NOs: 217-222;
- (21) all of SEQ ID NOs: 227-232;
- (22) all of SEQ ID NOs: 237-242;
- (23) all of SEQ ID NOs: 247-252;
- (24) all of SEQ ID NOs: 257-262;
- (25) all of SEQ ID NOs: 267-272;
- (26) all of SEQ ID NOs: 277-282;
- (27) all of SEQ ID NOs: 287-292;
- (28) all of SEQ ID NOs: 297-302;
- (29) all of SEQ ID NOs: 307-312;
- (30) all of SEQ ID NOs: 317-322;
- (31) all of SEQ ID NOs: 327-332;
- (32) all of SEQ ID NOs: 337-342;
- (33) all of SEQ ID NOs: 347-352;
- (34) all of SEQ ID NOs: 357-362;
- (35) all of SEQ ID NOs: 367-372;
- (36) all of SEQ ID NOs: 377-382;
- (37) all of SEQ ID NOs: 387-392;
- (38) all of SEQ ID NOs: 397-402;
- (39) all of SEQ ID NOs: 407-412;
- (40) all of SEQ ID NOs: 417-422;
- (41) all of SEQ ID NOs: 427-432;
- (42) all of SEQ ID NOs: 437-442;
- (43) all of SEQ ID NOs: 447-452;
- (44) all of SEQ ID NOs: 457-462;
- (45) all of SEQ ID NOs: 467-472;
- (46) all of SEQ ID NOs: 477-482; or
- (47) all of SEQ ID NOs: 487-492.

5. The TCR of any one of claims 1-4, wherein the TCR comprises the amino acid sequences of:

- (1) both of SEQ ID NOs: 33-34;
- (2) both of SEQ ID NOs: 43-44;

- (3) both of SEQ ID NOs: 53-54;
- (4) both of SEQ ID NOs: 63-64;
- (5) both of SEQ ID NOs: 73-74;
- (6) both of SEQ ID NOs: 83-84;
- (7) both of SEQ ID NOs: 93-94;
- (8) both of SEQ ID NOs: 103-104;
- (9) both of SEQ ID NOs: 113-114;
- (10) both of SEQ ID NOs: 123-124;
- (11) both of SEQ ID NOs: 133-134;
- (12) both of SEQ ID NOs: 143-144;
- (13) both of SEQ ID NOs: 153-154;
- (14) both of SEQ ID NOs: 163-164;
- (15) both of SEQ ID NOs: 173-174;
- (16) both of SEQ ID NOs: 183-184;
- (17) both of SEQ ID NOs: 193-194;
- (18) both of SEQ ID NOs: 203-204;
- (19) both of SEQ ID NOs: 213-214;
- (20) both of SEQ ID NOs: 223-224;
- (21) both of SEQ ID NOs: 233-234;
- (22) both of SEQ ID NOs: 243-244;
- (23) both of SEQ ID NOs: 253-254;
- (24) both of SEQ ID NOs: 263-264;
- (25) both of SEQ ID NOs: 273-274;
- (26) both of SEQ ID NOs: 283-284;
- (27) both of SEQ ID NOs: 293-294;
- (28) both of SEQ ID NOs: 303-304;
- (29) both of SEQ ID NOs: 313-314;
- (30) both of SEQ ID NOs: 323-324;
- (31) both of SEQ ID NOs: 333-334;
- (32) both of SEQ ID NOs: 343-344;
- (33) both of SEQ ID NOs: 353-354;
- (34) both of SEQ ID NOs: 363-364;
- (35) both of SEQ ID NOs: 373-374;

- (36) both of SEQ ID NOs: 383-384;
- (37) both of SEQ ID NOs: 393-394;
- (38) both of SEQ ID NOs: 403-404;
- (39) both of SEQ ID NOs: 413-414;
- (40) both of SEQ ID NOs: 423-424;
- (41) both of SEQ ID NOs: 433-434;
- (42) both of SEQ ID NOs: 443-444;
- (43) both of SEQ ID NOs: 453-454;
- (44) both of SEQ ID NOs: 463-464;
- (45) both of SEQ ID NOs: 473-474;
- (46) both of SEQ ID NOs: 483-484; or
- (47) both of SEQ ID NOs: 493-494.

6. The TCR of any one of claims 1-5, wherein the TCR comprises the amino acid sequences of:

- (1) both of SEQ ID NOs: 35-36;
- (2) both of SEQ ID NOs: 45-46;
- (3) both of SEQ ID NOs: 55-56;
- (4) both of SEQ ID NOs: 65-66;
- (5) both of SEQ ID NOs: 75-76;
- (6) both of SEQ ID NOs: 85-86;
- (7) both of SEQ ID NOs: 95-96;
- (8) both of SEQ ID NOs: 105-106;
- (9) both of SEQ ID NOs: 115-116;
- (10) both of SEQ ID NOs: 125-126;
- (11) both of SEQ ID NOs: 135-136;
- (12) both of SEQ ID NOs: 145-146;
- (13) both of SEQ ID NOs: 155-156;
- (14) both of SEQ ID NOs: 165-166;
- (15) both of SEQ ID NOs: 175-176;
- (16) both of SEQ ID NOs: 185-186;
- (17) both of SEQ ID NOs: 195-196;
- (18) both of SEQ ID NOs: 205-206;

- (19) both of SEQ ID NOs: 215-216;
- (20) both of SEQ ID NOs: 225-226;
- (21) both of SEQ ID NOs: 235-236;
- (22) both of SEQ ID NOs: 245-246;
- (23) both of SEQ ID NOs: 255-256;
- (24) both of SEQ ID NOs: 265-266;
- (25) both of SEQ ID NOs: 275-276;
- (26) both of SEQ ID NOs: 285-286;
- (27) both of SEQ ID NOs: 295-296;
- (28) both of SEQ ID NOs: 305-306;
- (29) both of SEQ ID NOs: 315-316;
- (30) both of SEQ ID NOs: 325-326;
- (31) both of SEQ ID NOs: 335-336;
- (32) both of SEQ ID NOs: 345-346;
- (33) both of SEQ ID NOs: 355-356;
- (34) both of SEQ ID NOs: 365-366;
- (35) both of SEQ ID NOs: 375-376;
- (36) both of SEQ ID NOs: 385-386;
- (37) both of SEQ ID NOs: 395-396;
- (38) both of SEQ ID NOs: 405-406;
- (39) both of SEQ ID NOs: 415-416;
- (40) both of SEQ ID NOs: 425-426;
- (41) both of SEQ ID NOs: 435-436;
- (42) both of SEQ ID NOs: 445-446;
- (43) both of SEQ ID NOs: 455-456;
- (44) both of SEQ ID NOs: 465-466;
- (45) both of SEQ ID NOs: 475-476;
- (46) both of SEQ ID NOs: 485-486; or
- (47) both of SEQ ID NOs: 495-496.

7. An isolated or purified polypeptide comprising a functional portion of the TCR of any one of claims 1-6, wherein the polypeptide comprises the amino acid sequences of:

- (1) all of SEQ ID NOs: 27-32;

- (2) all of SEQ ID NOs: 37-42;
- (3) all of SEQ ID NOs: 47-52;
- (4) all of SEQ ID NOs: 57-62;
- (5) all of SEQ ID NOs: 67-72;
- (6) all of SEQ ID NOs: 77-82;
- (7) all of SEQ ID NOs: 87-92;
- (8) all of SEQ ID NOs: 97-102;
- (9) all of SEQ ID NOs: 107-112;
- (10) all of SEQ ID NOs: 117-122;
- (11) all of SEQ ID NOs: 127-132;
- (12) all of SEQ ID NOs: 137-142;
- (13) all of SEQ ID NOs: 147-152;
- (14) all of SEQ ID NOs: 157-162;
- (15) all of SEQ ID NOs: 167-172;
- (16) all of SEQ ID NOs: 177-182;
- (17) all of SEQ ID NOs: 187-192;
- (18) all of SEQ ID NOs: 197-202;
- (19) all of SEQ ID NOs: 207-212;
- (20) all of SEQ ID NOs: 217-222;
- (21) all of SEQ ID NOs: 227-232;
- (22) all of SEQ ID NOs: 237-242;
- (23) all of SEQ ID NOs: 247-252;
- (24) all of SEQ ID NOs: 257-262;
- (25) all of SEQ ID NOs: 267-272;
- (26) all of SEQ ID NOs: 277-282;
- (27) all of SEQ ID NOs: 287-292;
- (28) all of SEQ ID NOs: 297-302;
- (29) all of SEQ ID NOs: 307-312;
- (30) all of SEQ ID NOs: 317-322;
- (31) all of SEQ ID NOs: 327-332;
- (32) all of SEQ ID NOs: 337-342;
- (33) all of SEQ ID NOs: 347-352;
- (34) all of SEQ ID NOs: 357-362;

- (35) all of SEQ ID NOs: 367-372;
- (36) all of SEQ ID NOs: 377-382;
- (37) all of SEQ ID NOs: 387-392;
- (38) all of SEQ ID NOs: 397-402;
- (39) all of SEQ ID NOs: 407-412;
- (40) all of SEQ ID NOs: 417-422;
- (41) all of SEQ ID NOs: 427-432;
- (42) all of SEQ ID NOs: 437-442;
- (43) all of SEQ ID NOs: 447-452;
- (44) all of SEQ ID NOs: 457-462;
- (45) all of SEQ ID NOs: 467-472;
- (46) all of SEQ ID NOs: 477-482; or
- (47) all of SEQ ID NOs: 487-492.

8. The polypeptide of claim 7, wherein the polypeptide comprises the amino acid sequences of:

- (1) both of SEQ ID NOs: 33-34;
- (2) both of SEQ ID NOs: 43-44;
- (3) both of SEQ ID NOs: 53-54;
- (4) both of SEQ ID NOs: 63-64;
- (5) both of SEQ ID NOs: 73-74;
- (6) both of SEQ ID NOs: 83-84;
- (7) both of SEQ ID NOs: 93-94;
- (8) both of SEQ ID NOs: 103-104;
- (9) both of SEQ ID NOs: 113-114;
- (10) both of SEQ ID NOs: 123-124;
- (11) both of SEQ ID NOs: 133-134;
- (12) both of SEQ ID NOs: 143-144;
- (13) both of SEQ ID NOs: 153-154;
- (14) both of SEQ ID NOs: 163-164;
- (15) both of SEQ ID NOs: 173-174;
- (16) both of SEQ ID NOs: 183-184;
- (17) both of SEQ ID NOs: 193-194;

- (18) both of SEQ ID NOs: 203-204;
- (19) both of SEQ ID NOs: 213-214;
- (20) both of SEQ ID NOs: 223-224;
- (21) both of SEQ ID NOs: 233-234;
- (22) both of SEQ ID NOs: 243-244;
- (23) both of SEQ ID NOs: 253-254;
- (24) both of SEQ ID NOs: 263-264;
- (25) both of SEQ ID NOs: 273-274;
- (26) both of SEQ ID NOs: 283-284;
- (27) both of SEQ ID NOs: 293-294;
- (28) both of SEQ ID NOs: 303-304;
- (29) both of SEQ ID NOs: 313-314;
- (30) both of SEQ ID NOs: 323-324;
- (31) both of SEQ ID NOs: 333-334;
- (32) both of SEQ ID NOs: 343-344;
- (33) both of SEQ ID NOs: 353-354;
- (34) both of SEQ ID NOs: 363-364;
- (35) both of SEQ ID NOs: 373-374;
- (36) both of SEQ ID NOs: 383-384;
- (37) both of SEQ ID NOs: 393-394;
- (38) both of SEQ ID NOs: 403-404;
- (39) both of SEQ ID NOs: 413-414;
- (40) both of SEQ ID NOs: 423-424;
- (41) both of SEQ ID NOs: 433-434;
- (42) both of SEQ ID NOs: 443-444;
- (43) both of SEQ ID NOs: 453-454;
- (44) both of SEQ ID NOs: 463-464;
- (45) both of SEQ ID NOs: 473-474;
- (46) both of SEQ ID NOs: 483-484; or
- (47) both of SEQ ID NOs: 493-494.

9. The polypeptide of claim 7 or 8, wherein the polypeptide comprises the amino acid sequences of:

- (1) both of SEQ ID NOs: 35-36;
- (2) both of SEQ ID NOs: 45-46;
- (3) both of SEQ ID NOs: 55-56;
- (4) both of SEQ ID NOs: 65-66;
- (5) both of SEQ ID NOs: 75-76;
- (6) both of SEQ ID NOs: 85-86;
- (7) both of SEQ ID NOs: 95-96;
- (8) both of SEQ ID NOs: 105-106;
- (9) both of SEQ ID NOs: 115-116;
- (10) both of SEQ ID NOs: 125-126;
- (11) both of SEQ ID NOs: 135-136;
- (12) both of SEQ ID NOs: 145-146;
- (13) both of SEQ ID NOs: 155-156;
- (14) both of SEQ ID NOs: 165-166;
- (15) both of SEQ ID NOs: 175-176;
- (16) both of SEQ ID NOs: 185-186;
- (17) both of SEQ ID NOs: 195-196;
- (18) both of SEQ ID NOs: 205-206;
- (19) both of SEQ ID NOs: 215-216;
- (20) both of SEQ ID NOs: 225-226;
- (21) both of SEQ ID NOs: 235-236;
- (22) both of SEQ ID NOs: 245-246;
- (23) both of SEQ ID NOs: 255-256;
- (24) both of SEQ ID NOs: 265-266;
- (25) both of SEQ ID NOs: 275-276;
- (26) both of SEQ ID NOs: 285-286;
- (27) both of SEQ ID NOs: 295-296;
- (28) both of SEQ ID NOs: 305-306;
- (29) both of SEQ ID NOs: 315-316;
- (30) both of SEQ ID NOs: 325-326;
- (31) both of SEQ ID NOs: 335-336;
- (32) both of SEQ ID NOs: 345-346;
- (33) both of SEQ ID NOs: 355-356;

- (34) both of SEQ ID NOs: 365-366;
- (35) both of SEQ ID NOs: 375-376;
- (36) both of SEQ ID NOs: 385-386;
- (37) both of SEQ ID NOs: 395-396;
- (38) both of SEQ ID NOs: 405-406;
- (39) both of SEQ ID NOs: 415-416;
- (40) both of SEQ ID NOs: 425-426;
- (41) both of SEQ ID NOs: 435-436;
- (42) both of SEQ ID NOs: 445-446;
- (43) both of SEQ ID NOs: 455-456;
- (44) both of SEQ ID NOs: 465-466;
- (45) both of SEQ ID NOs: 475-476;
- (46) both of SEQ ID NOs: 485-486; or
- (47) both of SEQ ID NOs: 495-496;

10. An isolated or purified protein comprising:

- (1) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 27-29 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 30-32;
- (2) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 37-39 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 40-42;
- (3) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 47-49 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 50-52;
- (4) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 57-59 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 60-62;
- (5) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 67-69 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 70-72;

(6) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 77-79 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 80-82;

(7) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 87-89 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 90-92;

(8) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 97-99 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 100-102;

(9) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 107-109 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 110-112;

(10) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 117-119 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 120-122;

(11) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 127-129 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 130-132;

(12) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 137-139 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 140-142;

(13) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 147-149 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 150-152;

(14) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 157-159 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 160-162;

(15) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 167-169 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 170-172;

(16) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 177-179 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 180-182;

(17) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 187-189 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 190-192;

(18) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 197-199 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 200-202;

(19) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 207-209 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 210-212;

(20) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 217-219 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 220-222;

(21) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 227-229 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 230-232;

(22) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 237-239 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 240-242;

(23) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 247-249 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 250-252;

(24) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 257-259 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 260-262;

(25) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 267-269 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 270-272;

(26) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 277-279 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 280-282;

(27) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 287-289 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 290-292;

(28) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 297-299 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 300-302;

(29) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 307-309 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 310-312;

(30) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 317-319 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 320-322;

(31) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 327-329 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 330-332;

(32) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 337-339 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 340-342;

(33) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 347-349 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 350-352;

(34) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 357-359 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 360-362;

(35) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 367-369 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 370-372;

(36) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 377-379 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 380-382;

(37) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 387-389 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 390-392;

(38) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 397-399 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 400-402;

(39) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 407-409 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 410-412;

(40) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 417-419 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 420-422;

(41) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 427-429 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 430-432;

(42) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 437-439 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 440-442;

(43) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 447-449 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 450-452;

(44) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 457-459 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 460-462;

(45) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 467-469 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 470-472;

(46) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 477-479 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 480-482; or

(47) a first polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 487-489 and a second polypeptide chain comprising the amino acid sequences of all of SEQ ID NOs: 490-492.

11. The protein of claim 10, wherein the protein comprises:

(1) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 33 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 34;

(2) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 43 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 44;

(3) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 53 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 54;

(4) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 63 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 64;

(5) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 73 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 74;

(6) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 83 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 84;

(7) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 93 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 94;

(8) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 103 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 104;

(9) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 113 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 114;

(10) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 123 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 124;

(11) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 133 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 134;

(12) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 143 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 144;

(13) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 153 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 154;

(14) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 163 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 164;

(15) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 173 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 174;

(16) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 183 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 184;

(17) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 193 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 194;

(18) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 203 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 204;

(19) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 213 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 214;

(20) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 223 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 224;

(21) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 233 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 234;

(22) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 243 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 244;

(23) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 253 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 254;

(24) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 263 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 264;

(25) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 273 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 274;

(26) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 283 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 284;

(27) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 293 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 294;

(28) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 303 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 304;

(29) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 313 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 314;

(30) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 323 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 324;

(31) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 333 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 334;

(32) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 343 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 344;

(33) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 353 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 354;

(34) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 363 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 364;

(35) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 373 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 374;

(36) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 383 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 384;

(37) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 393 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 394;

(38) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 403 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 404;

(39) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 413 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 414;

(40) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 423 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 424;

(41) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 433 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 434;

(42) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 443 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 444;

(43) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 453 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 454;

(44) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 463 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 464;

(45) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 473 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 474;

(46) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 483 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 484; or

(47) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 493 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 494.

12. The protein of claim 10 or 11, wherein the protein comprises:

(1) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 35 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 36;

(2) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 45 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 46;

(3) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 55 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 56;

(4) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 65 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 66;

(5) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 75 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 76;

(6) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 85 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 86;

(7) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 95 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 96;

(8) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 105 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 106;

(9) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 115 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 116;

(10) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 125 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 126;

(11) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 135 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 136;

(12) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 145 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 146;

(13) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 155 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 156;

(14) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 165 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 166;

(15) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 175 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 176;

(16) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 185 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 186;

(17) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 195 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 196;

(18) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 205 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 206;

(19) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 215 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 216;

(20) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 225 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 226;

(21) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 235 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 236;

(22) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 245 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 246;

(23) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 255 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 256;

(24) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 265 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 266;

(25) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 275 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 276;

(26) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 285 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 286;

(27) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 295 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 296;

(28) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 305 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 306;

(29) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 315 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 316;

(30) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 325 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 326;

(31) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 335 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 336;

(32) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 345 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 346;

(33) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 355 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 356;

(34) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 365 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 366;

(35) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 375 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 376;

(36) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 385 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 386;

(37) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 395 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 396;

(38) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 405 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 406;

(39) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 415 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 416;

(40) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 425 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 426;

(41) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 435 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 436;

(42) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 445 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 446;

(43) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 455 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 456;

(44) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 465 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 466;

(45) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 475 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 476;

(46) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 485 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 486; or

(47) a first polypeptide chain comprising the amino acid sequence of SEQ ID NO: 495 and a second polypeptide chain comprising the amino acid sequence of SEQ ID NO: 496.

13. An isolated or purified nucleic acid comprising a nucleotide sequence encoding the TCR according to any one of claims 1-6, the polypeptide according to any one of claims 7-9, or the protein according to any one of claims 10-12.

14. A recombinant expression vector comprising the nucleic acid according to claim 13.

15. An isolated or purified host cell comprising the recombinant expression vector according to claim 14.

16. An isolated or purified population of cells comprising the host cell according to claim 15.

17. A pharmaceutical composition comprising (a) the TCR according to any one of claims 1-6, the polypeptide according to any one of claims 7-9, the protein according to any one of claims 10-12, the nucleic acid according to claim 13, the recombinant expression vector according to claim 14, the host cell according to claim 15, or the population of cells according to claim 16 and (b) a pharmaceutically acceptable carrier.

18. A method of detecting the presence of cancer in mammal, the method comprising:

(a) contacting a sample comprising cells of the cancer with the TCR according to any one of claims 1-6, the polypeptide according to any one of claims 7-9, the protein according to any one of claims 10-12, the nucleic acid according to claim 13, the recombinant expression vector according to claim 14, the host cell according to claim 15, the population of cells according to claim 16, or the pharmaceutical composition of claim 17, thereby forming a complex; and

(b) detecting the complex,

wherein detection of the complex is indicative of the presence of cancer in the mammal.

19. The TCR according to any one of claims 1-6, the polypeptide according to any one of claims 7-9, the protein according to any one of claims 10-12, the nucleic acid

according to claim 13, the recombinant expression vector according to claim 14, the host cell according to claim 15, the population of cells according to claim 16, or the pharmaceutical composition of claim 17, for use in the treatment or prevention of cancer in a mammal.

20. The population of cells for the use according to claim 19, wherein the population of cells is autologous to the mammal.

21. The population of cells for the use according to claim 19, wherein the population of cells is allogeneic to the mammal.

22. The method according to claim 18, or the TCR, polypeptide, protein, nucleic acid, recombinant expression vector, host cell, population of cells, or pharmaceutical composition for the use according to any one of claims 19-21, wherein the cancer is an epithelial cancer.

23. The method according to claim 18, or the TCR, polypeptide, protein, nucleic acid, recombinant expression vector, host cell, population of cells, or pharmaceutical composition for the use according to any one of claims 19-21, wherein the cancer is cholangiocarcinoma, melanoma, colon cancer, rectal cancer, ovarian cancer, endometrial cancer, non-small cell lung cancer (NSCLC), glioblastoma, uterine cervical cancer, head and neck cancer, breast cancer, pancreatic cancer, or bladder cancer.