Novel Antibodies

The present invention relates to antibodies and antigen binding fragments thereof that specifically bind human Insulin–like Growth Factor Receptor (hIGF-1R). The present invention also concerns methods of treating diseases or disorders with said antibodies and antigen binding fragments thereof, pharmaceutical compositions comprising said antibodies and antigen binding fragments thereof and methods of manufacture.

Background

The human insulin-like growth factor receptor (also known as IGF-1R, CD221 or EC 2.7.112) is a tyrosine kinase receptor with 70% homology to the insulin receptor. The receptor is activated by two ligands – IGF-I and IGF-II which bind the receptor with high affinity. The receptor is a disulphide linked αβ dimer, denoted $(\alpha\beta)_2$. The α-chain is entirely extracellular whilst the β-chain is membrane-spanning and has both an extracellular domain and an intracellular signalling domain. Ligand-mediated receptor activation triggers intracellular events including activation of MAPK and PI3K-protein kinase B pathways. Whilst IGF-1R is known to have an essential role in normal foetal and postnatal growth and development, it has also assumed an important role in cancer biology and has been implicated in a number of biological pathways such as mitogenesis, transformation and protection from apoptosis (reviewed extensively in Baserga et al. (1997) Endocrine, 7(1):99-102, Baserga (2003) Int J Cancer, 107(6):873-7, Larsson et al. (2005) Br J Cancer, 92(12):2097-101, Romano (2003) Drug News Perspect, 16(8):525-31). Furthermore the levels of receptor expression are known to be up-regulated on a variety of tumours types (reviewed by Khandwala et al. (2000) Endocr Rev., 21(3):215-44) and increased levels of the ligand IGF-I are associated with an increased risk of developing prostate cancer (Chan et al. (1998) Science, 279(5350):563-6).

Antagonists of the IGF-1R signalling pathway are known for their anti-tumour effects in vitro and in vivo (reviewed in Hofmann et al. (2005) Drug Discov Today, 10(15):1041-7 and Zhang et al. (2004) Expert Opin Investig Drugs, 13(12):1569-77). Approaches include neutralising antibodies (see Kull et al. (1983) J Biol Chem., 258(10):6561-6 and Li et al, (1993) Cancer Immunol Immunother., 49(4-5):243-52, Xiong et al. (1992) Proc Natl Acad Sci U S A., 89(12):5356-60, Burtrum et al. (2003) Cancer Res., 63(24):8912-21, Cohen et al. (2005) Clin Cancer Res., 11(5):2063-73,

Maloney et al. (2003) Cancer Res., 63(16):5073-83, Jackson-Booth et al. (2003) Horm Metab Res., 35(11-12):850-6), anti-sense (see Resnicoff et al. (1994) Cancer Res., 54(18):4848-50, Lee et al. (1996) Cancer Res., 56(13):3038-41, Muller et al. (1998) Int J Cancer., 77(4):567-71, Trojan et al. (1993) Science, 259(5091):94-7, Shapiro et al. (1994) J Clin Invest., 94(3):1235-42), dominant negative mutants (Prager et al. (1994) Proc Natl Acad Sci U S A., 91(6):2181-5) and small molecule tyrosine kinase inhibitors (see Hopfner et al. (2006) Biochem Pharmacol. 2006, 71(10):1435-48 and IGF-binding proteins (IGFBPs - see Nickerson et al. (1997) Biochem Biophys Res Commun., 237(3):690-3). Known monoclonal antibodies include those described in: WO99/60023, WO03/100008, WO02/053596, WO04/071529, EP0629240B, WO03/059951, WO03/106621, WO04/083248, WO04/087756, US2006452167A. However, there is a need for antibodies with improved effector function, for example with improved ADCC and/or CDC function.

Antibody structures are well known in the art and in particular it is known that the heavy chain constant region has a glycosylated sugar chain, this may be an N-glycoside linked sugar chain for example N-acetlyglucosamine and it may or may not be fucosylated.

Methods for measuring levels of fucosylation are well known in the art for example, for a population of antibodies, acid hydrolysis can be used to remove the monosaccharides of the glycosylated sugar chain from the antibody and these can be labelled with a dye such as 2-aminobenzoic acid (2-AA). Reverse phase high performance liquid chromatography with fluorescence detection can then be carried out and a standard curve constructed for sample quantitation. The ratio of fucose to mannose per antibody population can then be calculated

Brief Description of the Figures

Figure 1: Binding of purified murine monoclonal antibodies to human IGF-1R as determined by ELISA.

Figure 2A-E: Binding of purified 6E11 chimeric and 6E11 humanised antibodies to human IGF-1R as determined by ELISA. In Figure 2A, the binding curve for H0L0 was shifted to the right due to the fact that the antibody was at very low concentration and could not be accurately quantified. In Figure 2D, whist the overall trend was similar, the signal was reduced compared to other assays.

Figure 3: Down-regulation of IGF-1R receptor following incubation of 3T3/LISN c4 cells for 24 hours with purified 6E11 murine monoclonal antibody to human IGF-1R

Figure 4: Down-regulation of IGF-1R receptor following incubation of NCI-H838 cells for up to 24 hours with humanised H0L0 antibody to human IGF-1R

Figure 5: Down-regulation of IGF-1R receptor and Insulin receptor following incubation of NCI-H838 cells for 24 hours with H0L0, H0L0 IgG1m(AA) or non-targeting human IgG.

Figure 6: Histogram of fluorescent intensity for IGF-1R levels on granulocyte and lymphocyte populations following incubation with H0L0 at 4°C and 37°C.

Figure 7: Histogram of fluorescent intensity for IGF-1R levels in granulocyte and lymphocyte populations following incubation with H0Lo at 4°C and 37°C compared to an isotype control.

Figure 8: Inhibition of receptor phosphorylation mediated by purified murine monoclonal antibodies 6E11, 5G4 and 15D9,

Figure 9: shows an example of the inhibition of receptor phosphorylation mediated by H1L0 in comparison to the 6E11c.

Figure 10: shows an example of the inhibition of receptor phosphorylation mediated by H0L0 and H0L0 IgG1m(AA) and H1L0 and H10L0 IgG1m(AA).

Figure 11A: shows an example of the activity of various purified murine monoclonal antibodies in the competition ELISA.

Figure 11B: shows an example of the activity of H1L0 in the competition ELISA in

comparison to 6E11c.

Figure 12A-C: Competition ELISA to demonstrate the ability of purified 6E11 murine monoclonal or 6E11 chimeric or 6E11 humanised antibodies to inhibit the binding of IGF-1R receptor to a second neutralising antibody.

Figure 13A: Binding of purified murine monoclonal antibodies to recombinant cynomolgus macaque IGF-1R as determined by ELISA.

Figure 13B: Binding of purified humanised monoclonal antibodies to recombinant cynomolgus macaque IGF-1R in comparison to the 6E11 chimera (6E11c).

Figure 14: Insulin receptor binding ELISA using purified murine monoclonal antibodies.

Figure 15: Insulin receptor binding ELISA using purified humanised antibodies

Figure 16: FACS assay to demonstrate that the antibodies recognize the Colo205 tumour cell line .

Figure 17: FACS assay to demonstrate that the antibodies recognize the NCI-H838 lung carcinoma tumour cell line.

Figure 18: FACS assay to demonstrate that the antibodies recognize the MCF7 carcinoma and A549 lunch carcinoma cell line.

Figure 19: Immunohistochemistry on frozen tissue samples of tumour and normal prostate samples using purified murine monoclonal antibody

Figure 20: Immunohistochemistry on frozen tissue samples of tumour breast samples using purified murine monoclonal antibody

Figure 21: Immunohistochemistry on frozen tissue samples of tumour breast samples using purified H1L0 humanised and 6E11 chimeric monoclonal antibodies.

Figure 22: Immunohistochemistry on frozen tumour tissue samples of tumour breast samples using biotinylated H0L0 antibody.

Figure 23: Inhibition of IGF-I mediated proliferation of 3T3/LISN c4 cells inhibited by purified murine monoclonal antibodies

Figure 24: Inhibition of IGF-I mediated proliferation of 3T3/LISN c4 cells inhibited by

purified H1L0 humanised or 6E11 chimeric monoclonal antibodies

Figure 25A-E: Inhibition of IGF-I mediated proliferation of 3T3/LISN c4 cells inhibited by purified humanised or purified murine 6E11 monoclonal antibodies

Figure 26: Inhibition of IGF-I mediated cell cycling by purified murine monoclonal antibodies as determined by propidium iodide staining and flow cytometry.

Figure 27: Reversal of IGF-1 mediated protection from camptothecin induced apoptosis in NCI-H838 cells by murine 6E11 monoclonal antibody.

Figure 28: : Reversal of IGF-1 mediated protection from camptothecin induced apoptosis in A549 cells by selected antibodies.

Figure 29: Absence of agonistic activity of purified murine monoclonal antibody in the presence of cross-linking antibody.

Figure 30: Activity of H0L0 antibody in differentiated pre-adipocytes on the basal levels of phospho-AKT compared to a negative control antibody.

Figure 31: Activity of H0L0 antibody on the basal levels of phosphor-AKT in A549 cells.

Figure:32 Activity of humanised antibodies on basal IGF-1R receptor phosphorylation levels in 3T3/LISNc4 cells.

Figure:33 Activity of cross-linked humanised antibodies on basal IGF-1R receptor phosphorylation levels in 3T3/LISn c4 cells.

Figure:34 Activity of humanised antibodies on the proliferation of NCI-H929 cells in the absence of ligand stimulation.

Figure 35: Inhibition of 3T3/LISN c4 tumour growth in nude mice following treatment with 6E11 monoclonal antibody

Figure 36: Inhibition of 3T3/LISN c4 tumour growth in nude mice following treatment with 6E11 monoclonal antibody

Figure 37: Inhibition of 3T3/LISN c4 tumour growth in nude mice following treatment with 6E11 and humanised monoclonal antibodies.

Figure 38: Inhibition of Colo205 tumour growth in nude mice following treatment with

6E11 monoclonal antibody

Figure 39: Kinetics of receptor binding using NCI – H838 cellls incubated with H0L0

Figure 40: IGF – 1R/IR heterodimer binding assay using Colo-205 cells and recombinant NIH-3T3 cells against 6E11, 6E11c and H0L0 mIgG(AA) antibody.

Figure 41: Proliferation of NCI – H929 cells in the presence of 6E11 and H0L0.

Figure 42: Determination of stability of H0L0 in serum.

Figure 43 : Downregulation of IGR-1R in vivo in LISN/3T3 c4 cells following treatment with H0L0 or 6E11..

Brief description of the Tables.

Table 1: SEQ ID NO's of the hybridoma variable heavy and light chains.

Table 2:IC50 values for selected antibodies in phosphorytlation assays.

Table 2a:IC50 values for selected antibodies in phosphorytlation assays.

Table 3: Kinetic data for murine monoclonal antibodies.

Table 4: Kinetic data for humanised monoclonal antibodies

Table 5: Kinetic data for humanised monoclonal antibodies both wildtype and disabled Fc.

Table 6: Kinetic data for humanised monoclonal antibodies.

Table 7: Kinetic data for humanised monoclonal antibodies.

Table 8: Kinetic data for anti-IGF-1R versus human and cyno IGF-1R.

Table 9: Inhibition values for the 200 RU's IGF-1 surface.

Table 10: Inhibition values for the 4000 RU's IGF-1 surface.

Tbale 11: Neutralisation of binding of receptor to ligand.

Table 12: Summary of immunohistochemistry analsysis of tumout tissue microarray.

Table 13: Activity of various antibodies in AKT phosphorylation assay.

Table 14: Activity of various antibodies in AKT phosphorylation assay.

Summary of Invention

In one embodiment the invention provides an antibody or antigen binding fragment thereof which specifically binds IGF-1R comprising CDR H3 of SEQ. ID. NO: 1 or a variant thereof which contains 1 or 2 amino acid substitutions in the CDRH3.

In one embodiment the invention provides an antibody or antigen binding fragment thereof which specifically binds IGF-1R, specifically hIGF-1R and neutralises the activity of hIGF-1R, which comprises a heavy chain variable domain comprising CDR H3 of SEQ. ID. NO: 1 or variants thereof in which one or two amino acid residues within CDR H3 differ from the amino acid residues in the corresponding position in SEQ. ID. NO: 1.

Also provided is a method of producing an antibody as described herein comprising expressing in a cell line an antibody or antigen binding fragment thereof.

In another embodiment is provided a kit-of-parts comprising the composition described herein together with instructions for use.

Also provided is a method of treating a human patient afflicted with cancer which method comprises the step of administering a therapeutically effective amount of the antibody preparation described herein.

Detailed Description of Invention

The present invention provides an antibody or antigen binding fragment thereof which specifically binds IGF-1R, for example which specifically binds hIGF-1R.

In one embodiment of the present invention there is provided an antibody or antigen binding fragment thereof which specifically binds hIGF-1R and neutralises the activity of hIGF-1R, which comprises a heavy chain variable domain which specifically binds IGF-1R comprising CDR H3 of SEQ. ID. NO: 1 or variants thereof in which one or two amino acid residues within CDR H3 differ from the amino acid residues in the corresponding position in SEQ. ID. NO: 1.

In one embodiment of the present invention these differences in amino acid residues

are conservative substitutions.

In another embodiment of the invention there is provided an antibody or antigen binding fragment thereof which specifically binds IGF-1R and comprises a CDRH3 which is a variant of the sequence set forth in SEQ ID NO:1 in which one or two residues within said CDRH3 of said variant differs from the residue in the corresponding position in SEQ ID NO:1 in position 7 and/or position 9 (where the first residue is position 1, W, and where the last residue, V, is in position 14).

In a further embodiment of the invention there is provided an antibody or antigen binding fragment thereof which specifically binds IGF-1R and comprises a CDRH3 which is a variant of the sequence set forth in SEQ ID NO:1 in which one or two residues within said CDRH3 of said variant differs from the residue in the corresponding position in SEQ ID NO:1 by a substitution of R to S at position 7, or by a substitution of K to R at position 9, or by a substitution of R to S at position 7 and K to R at position 9.

In another embodiment of the invention there is provided an antibody or antigen binding fragment thereof further comprising one or more of the following sequences CDRH2 as set out in SEQ. ID. NO: 2, CDRH1 as set out in SEQ. ID. NO: 3, CDRL1 as set out in SEQ. ID. NO: 4, CDRL2 as set out in SEQ. ID. NO: 5, and CDRL3 as set out in SEQ. ID. NO: 6.

In one embodiment of the invention there is provided an antibody or antigen binding fragment thereof wherein CDR H1, H2 and H3 and CDR L1 and L3 are from 6E11 and CDR L2 is from 9C7.

In one embodiment of the present invention one or more of the CDR's of the antibody or antigen binding fragment thereof may comprise variants of the CDR's set out in the sequences listed above. Each variant CDR will comprise one or two amino acid residues which differ from the amino acid residue in the corresponding position in the sequence listed. Such substitutions in amino acid residues may be conservative substitutions, for example, substituting one hydrophobic amino acid for an alternative hydrophobic amino acid, for example substituting Leucine with Valine, or Isoleucine.

In a further embodiment of the invention there is provided an antibody or antigen binding fragment thereof comprising CDRH3 and further comprises one or more of

the following sequences CDRH2: SEQ. ID. NO: 2, CDRH1: SEQ. ID. NO: 3, CDRL1: SEQ. ID. NO: 4, CDRL2: SEQ. ID. NO: 7, and CDRL3: SEQ. ID. NO: 6.

In yet a further embodiment of the invention there is provided an antibody or antigen binding fragment thereof comprising CDRH3 and further comprises one or more of the following sequences CDRH2: SEQ. ID. NO: 2, CDRH1: SEQ. ID. NO: 3, CDRL1: SEQ. ID. NO: 4, CDRL2: SEQ. ID. NO: 7, and CDRL3: SEQ. ID. NO: 6 wherein one or more of the CDR's may be replaced by a variant thereof, each variant CDR containing 1 or 2 amino acid substitutions.

In one embodiment the antibody or antigen binding fragment thereof of the present invention comprises CDR H3 of SEQ. ID. NO: 1 and CDR H1 of SEQ. ID. NO: 3. In a further embodiment the antibody or antigen binding fragment thereof comprises CDRH3 of SEQ ID NO: 1 and CDR L2 of SEQ. ID. NO: 7. In yet a further embodiment the antibody or antigen binding fragment thereof of the present invention comprises CDR H3 of SEQ. ID. NO: 1 and CDR H1 of SEQ. ID. NO: 3. and CDR L2 of SEQ. ID. NO: 7.

In another embodiment of the present invention there is provided an antibody or antigen binding fragment thereof according to the invention described herein and further comprising the following CDR's:

CDRH1: SEQ. ID. NO: 3
CDRH2: SEQ. ID. NO: 2
CDRH3: SEQ. ID. NO: 1
CDRL1: SEQ. ID. NO: 4
CDRL2: SEQ. ID. NO: 7
CDRL3: SEQ. ID. NO: 6

In another embodiment of the invention there is provided an antibody or antigen binding fragment thereof which specifically binds IGF-1R and comprises CDR's which are variants of the sequences set forth above.

In another embodiment of the present invention there is provided an antibody or antigen binding fragment thereof which specifically binds IGF-1R and comprises a heavy chain variable domain of SEQ. ID. NO: 8 and a light chain variable domain of SEQ. ID. NO: 9, or a heavy chain variable domain of SEQ. ID. NO: 10 and a light

chain variable domain of SEQ. ID. NO: 11, or a heavy chain variable domain of SEQ. ID. NO: 12 and a light chain variable domain of SEQ. ID. NO: 13, or a heavy chain variable domain of SEQ. ID. NO: 14 and a light chain variable domain of SEQ. ID. NO: 16, or a heavy chain variable domain of SEQ. ID. NO: 15 and a light chain variable domain of SEQ. ID. NO: 16.

In another embodiment of the invention there is provided an isolated heavy chain variable domain of an antibody comprising SEQ ID NO: 12, SEQ ID NO: 14 or SEQ ID NO: 15, for example it comprises SEQ ID NO: 12.

In another embodiment of the present invention there is provided an antibody or antigen binding fragment thereof comprising CDR's according to the invention described herein, or heavy or light chain variable domains according to the invention described herein, wherein the antibody or antigen binding fragment thereof is rat, mouse, primate (e.g. cynomolgus, Old World monkey or Great Ape) or human.

In another embodiment of the present invention the antibody or antigen binding fragment thereof described herein additionally binds primate IGF-1R, for example cynomolgus macaque monkey IGF-1R.

In another embodiment of the present invention there is provided an antibody or antigen binding fragment thereof comprising one or more of the following CDR's: CDRH3 as set out in as set out in SEQ. ID. NO: 1, CDRH2 as set out in SEQ. ID. NO: 2, CDRH1 as set out in SEQ. ID. NO: 3, CDRL1 as set out in SEQ. ID. NO: 4, CDRL2 as set out in SEQ. ID. NO: 5 and CDRL3 as set out in SEQ. ID. NO: 6 in the context of a human framework, for example as a humanised or chimaeric antibody.

In one embodiment of the present invention the humanised heavy chain variable domain comprises the CDR's listed in SEQ ID NO: 1-3 within an acceptor antibody framework having greater than 80% identity in the framework regions, or greater than 85%, or greater than 90%, or greater than 95%, or greater than 98%, or greater than 99% identity in the framework regions to the human acceptor sequence in SEQ ID NO: 59

In one embodiment of the present invention the humanised light chain variable domain comprises the CDR's listed in SEQ ID NO: 4-6 within an acceptor antibody framework having greater than 80% identity in the framework regions, or greater than

85%, or greater than 90%, or greater than 95%, or greater than 98%, or greater than 99% identity in the framework regions to the human acceptor sequence in SEQ ID NO: 60

In SEQ ID NO: 59 and SEQ ID NO: 60 the position of the CDR sequences have been denoted by Xaa's.

In one embodiment of the invention there is provided an antibody or antigen binding fragment thereof comprising CDR's according to the invention described herein, or heavy chain or light chain variable domains according to the invention described herein wherein the antibody has a half life of at least 5 days, or at least 7 days or at least 9 days in a murine animal model.

In another embodiment of the present invention there is provided an antibody or antigen binding fragment thereof comprising CDR's according to the invention described herein, or heavy or light chain variable domains according to the invention described wherein the antibody further comprises a constant region, which may be of any isotype or subclass. In one embodiment the heavy chain constant region is of the IgG isotype, for example IgG1, IgG2, IgG3, IgG4 or variants thereof. In one embodiment the antibody is IgG1.

In one embodiment of the present invention there is provided an antibody according to the invention described herein and comprising a constant region such that the antibody has reduced ADCC and/or complement activation or effector functionality. In one such embodiment the heavy chain constant region may comprise a naturally disabled constant region of IgG2 or IgG4 isotype or a mutated IgG1 constant region. Examples of suitable modifications are described in EP0307434. One example comprises the substitutions of alanine residues at positions 235 and 237 (EU index numbering).

In another embodiment of the present invention there is provided an antibody according to the invention described herein wherein the antibody is capable of at least some effector function for example wherein it is capable of some ADCC and/or CDC function. In one embodiment of the present invention there is provided an antibody comprising a constant region or antigen binding fragment thereof which is linked to a constant region which specifically binds IGF-1R, for example human IGF-1R comprising CDR H3 of SEQ. ID. NO: 1 or variant thereof which contains 1 or 2

amino acid substitutions in the CDRH3, for example an antibody comprising a constant region or antigen binding fragment thereof which is linked to a constant region comprising CDR's selected from CDRH1: SEQ. ID. NO: 3, CDRH2: SEQ. ID. NO: 2, CDRH3: SEQ. ID. NO: 1, CDRL1: SEQ. ID. NO: 4, CDRL2: SEQ. ID. NO: 7 and CDRL3: SEQ. ID. NO: 6, and which further comprises a constant region of IgG1 wild type, IgG2 wild type, IgG3 wild type, IgG4 wild type or enhanced versions thereof.

In one embodiment of the present invention the antibody described herein comprising a constant region or antigen binding fragment thereof which is linked to a constant region, specifically binds to a growth factor receptor selected from IGF-1R, EGFR, HER-2 or HER-3.For example which specifically binds to HER-2 or HER-3 or for example which specifically binds to IGF-1R or EGRF, for example human IGF-1R.

In one embodiment the antibody of the present invention is linked to one or more domain antibodies with specificity for VEGF or EGFR.

In one embodiment of the present invention there is provided an antibody or antigen binding fragment thereof according to the invention described herein which comprises one or more mutations in its heavy chain constant region such that the antibody or antigen binding fragment has enhanced effector function. For example, wherein it has enhanced ADCC or enhanced CDC or wherein it has both enhanced ADCC and CDC effector function. Examples of suitable modifications are described in Shields et al. J. Biol. Chem (2001) 276:6591-6604, Lazar et al. PNAS (2006) 103:4005-4010 and US6737056, WO2004063351 and WO2004029207.
In one embodiment of the present invention there is provided an antibody comprising a heavy chain constant region or antigen binding fragment thereof which is linked to a heavy chain constant region which specifically binds IGF-1R, for example human IGF-1R. The antibody or antigen binding fragment thereof may comprise CDR H3 of SEQ. ID. NO: 1 or variants thereof in which one or two amino acid residues within CDR H3 differ from the amino acid residues in the corresponding position in SEQ. ID. NO: 1 and comprising a mutated heavy chain constant region such that the antibody or antigen binding fragment thereof has enhanced effector function compared to wild type. For example, an antibody or antigen binding fragment thereof which specifically binds IGF-1R comprising CDR H3 of SEQ. ID. NO: 1, for example an antibody or antigen binding fragment thereof comprising CDR's selected from CDRH1: SEQ. ID. NO: 3, CDRH2: SEQ. ID. NO: 2, CDRH3: SEQ. ID. NO: 1, CDRL1: SEQ. ID. NO: 4,

CDRL2: SEQ. ID. NO: 7 and CDRL3: SEQ. ID. NO: 6 and comprising a mutated heavy chain constant region such that the antibody or antigen binding fragment thereof has enhanced effector function compared to wild type.

In one embodiment of the present invention, such mutations are in one or more of positions selected from 239, 332 and 330 (IgG1), or the equivalent positions in other IgG isotypes. Examples of suitable mutations are S239D and I332E and A330L. In one embodiment the antibody or antigen binding fragment is mutated at positions 239 and 332, for example S239D and I332E, for example it is mutated at three or more positions selected from 239 and 332 and 330, for example S239D and I332E and A330L.

In another embodiment of the present invention there is provided an antibody comprising a heavy chain constant region or antigen binding fragment thereof which is linked to a heavy chain constant region according to the invention described herein and comprising a constant region selected from those set out in SEQ ID NO: 64 and SEQ ID. NO: 66, for example an antibody or antigen binding fragment comprising the variable domains of SEQ ID NO: 14 and SEQ ID NO: 15 together with the heavy chain constant region as set out in SEQ ID NO: 64 or SEQ ID NO: 66, for example an antibody comprising a heavy chain constant region or antigen binding fragment thereof which is linked to a heavy chain constant region comprising SEQ ID NO: 14, SEQ ID NO: 15 and SEQ ID NO: 64. In a further embodiment of the present invention there is provided an antibody comprising a heavy chain constant region or antigen binding fragment thereof which is linked to a heavy chain constant region according to the invention described herein and comprising a heavy chain constant region selected from those set out in SEQ ID NO: 64 and SEQ ID. NO: 66, for example antibody or antigen binding fragment thereof comprising the variable domains of SEQ ID NO: 14 and SEQ ID NO: 16 together with the heavy chain constant region as set out in SEQ ID NO: 64 or SEQ ID NO: 66, for example an antibody or antigen binding fragment thereof comprising SEQ ID NO: 14, SEQ ID NO: 16 and SEQ ID NO: 64.

In one embodiment of the present invention there is provided an antibody comprising a heavy chain constant region or antigen binding fragment thereof which is linked to a heavy chain constant region according to the invention described herein which comprises a heavy chain constant region with an altered glycosylation profile such that the antibody or antigen binding fragment thereof has enhanced effector function. For example, wherein it has enhanced ADCC or enhanced CDC or wherein it has

both enhanced ADCC and CDC effector function. Examples of suitable methodologies to produce antibodies with an altered glycosylation profile are described in WO2003011878, WO2006014679 and EP1229125.

In one embodiment of the present invention there is provided an antibody comprising a heavy chain constant region or antigen binding fragment thereof which is linked to a heavy chain constant region according to the invention described herein which specifically binds IGF-1R, for example human IGF-1R. The antibody or antigen binding fragment thereof may comprise CDR H3 of SEQ. ID. NO: 1 or variants thereof in which one or two amino acid residues within CDR H3 differ from the amino acid residues in the corresponding position in SEQ. ID. NO: 1 and comprising a heavy chain constant region with an altered glycosylation profile such that the antibody or antigen binding fragment has enhanced effector function when compared to wild type.

For example, an antibody or antigen binding fragment thereof which specifically binds IGF-1R, for example human IGF-1R comprising CDR H3 of SEQ. ID. NO: 1, for example an antibody or antigen binding fragment thereof comprising CDR's selected from CDRH1: SEQ. ID. NO: 3, CDRH2: SEQ. ID. NO: 2, CDRH3: SEQ. ID. NO: 1, CDRL1: SEQ. ID. NO: 4, CDRL2: SEQ. ID. NO: 7 and CDRL3: SEQ. ID. NO: 6 and comprising a heavy chain constant region with an altered glycosylation profile such that the antibody or antigen binding fragment has enhanced effector function when compared to wild type.

In one embodiment the invention provides an antibody preparation wherein the ratio of fucose to mannose in said antibody preparation is 0.8:3 or less, for example is 0.7:3 or less, or is 0.6:3 or less or is 0.5:3 or less or is 0.4:3 or less or is 0.3:3 or less, or is 0.2:3 or less or is 0.1:3 or less. In one embodiment the antibody preparation contains negligible or no bound fucose.

In another embodiment of the present invention there is provided an antibody preparation comprising an antibody or antigen binding fragment thereof comprising the variable domains of SEQ ID NO: 14 and SEQ ID NO: 15 or SEQ ID NO: 14 and SEQ ID NO: 16 and wherein the ratio of fucose to mannose in said antibody preparation is 0.8:3 or less, for example is 0.7:3 or less, or is 0.6:3 or less or is 0.5:3 or less or is 0.4:3 or less or is 0.3:3 or less, or is 0.2:3 or less or is 0.1:3 or less. In one embodiment the antibody preparation contains negligible or no bound fucose.

In one embodiment of the present invention there is provided an antibody comprising a heavy chain constant region or antigen binding fragment thereof which is linked to a heavy chain constant region according to the invention described herein which comprises a mutated heavy chain constant region and an altered glycosylation profile such that the antibody or antigen binding fragment has enhanced effector function, for example wherein it has one or more of the following functions, enhanced ADCC or enhanced CDC, for example wherein it has enhanced ADCC function.

In one embodiment of the invention there is provided an antibody preparation comprising antibodies as described herein which comprise an immunoglobulin heavy chain constant region, or antigen binding fragments thereof which are linked to an immunoglobulin heavy chain constant region wherein said immunoglobulin heavy chain constant region confers an effector function to the antibody or antigen binding fragment, and wherein said antibody or antigen binding fragment specifically binds to a growth factor receptor and wherein said immunoglobulin heavy chain constant region is mutated in at least 2 positions and has an altered glycosylation profile such that the ratio of fucose to mannose is 0.8:3 or less so that said antibody or antigen binding fragment has an enhanced effector function in comparison with an equivalent antibody or antigen-binding fragment with an immunoglobulin heavy chain constant region lacking said mutations and altered glycosylation profile. The altered glycosylation profile of said antibody preparation is not a consequence of said immunoglobulin heavy chain mutations.

For example, such antibodies or antigen binding fragments specifically bind IGF-1R, for example human IGF-1R and comprise CDR H3 of SEQ. ID. NO: 1, for example an antibody or antigen binding fragment comprising CDR's selected from CDRH1: SEQ. ID. NO: 3, CDRH2: SEQ. ID. NO: 2, CDRH3: SEQ. ID. NO: 1, CDRL1: SEQ. ID. NO: 4, CDRL2: SEQ. ID. NO: 7 and CDRL3: SEQ. ID. NO: 6 and comprise a mutated heavy chain constant region and have an altered glycosylation profile such that the antibody or antigen binding fragment has enhanced effector function. For example such antibodies or antigen binding fragments may comprise the variable domains of SEQ ID NO: 14 and SEQ ID NO: 15 or SEQ ID NO: 14 and SEQ ID NO: 16.

In one such embodiment, the mutations are in one or more of positions selected from 239, 332 and 330 (IgG1), or the equivalent positions in other IgG isotypes. Examples of suitable mutations are S239D and I332E and A330L. In one embodiment the

antibody comprising a constant region or antigen binding fragment thereof which is linked to a constant region has a mutation at 239 and 332, for example S239D and I332E or further may comprise mutations at three or more positions selected from 239 and 332 and 330, for example S239D and I332E and A330L.

In one embodiment the ratio of fucose to mannose in said antibody preparation is 0.8:3 or less, for example is 0.7:3 or less, or is 0.6:3 or less or is 0.5:3 or less or is 0.4:3 or less or is 0.3:3 or less, or is 0.2:3 or less or is 0.1:3 or less. In one embodiment the antibody preparation contains negligible or no bound fucose.

In one embodiment of the present invention there is provided an antibody comprising a heavy chain constant region or antigen binding fragment thereof which is linked to a heavy chain constant region according to the invention described herein which comprises a chimaeric heavy chain constant region for example wherein it comprises at least one CH2 domain from IgG3 such that the antibody or antigen binding fragment has enhanced effector function, for example wherein it has one or more of the following functions, enhanced ADCC or enhanced CDC, for example wherein it has enhanced CDCC. For example the antibody or antigen binding fragment may comprise one CH2 domain from IgG3 or both CH2 domains may be from IgG3.

In a further embodiment of the present invention there is provided an antibody comprising a heavy chain constant region or antigen binding fragment thereof which is linked to a heavy chain constant region according to the invention described herein which comprises a mutated and chimaeric heavy chain constant region for example wherein it comprises at least one CH2 domain from IgG3 and one CH2 domain from IgG1 wherein the IgG1 CH2 domain has one or more mutations at positions selected from 239 and 332 and 330, for example the mutations are selected from S239D and I332E and A330L such that the antibody has enhanced effector function, for example wherein it has one or more of the following functions, enhanced ADCC or enhanced CDC, for example wherein it has enhanced ADCC and enhanced CDCC. In one embodiment the IgG1 CH2 domain has the mutations S239D and I332E.

In one embodiment of the present invention there is provided an antibody comprising a heavy chain constant region or antigen binding fragment thereof which is linked to a heavy chain constant region according to the invention described herein which

comprises a chimaeric heavy chain constant region and an altered glycosylation profile such that the heavy chain constant region comprises at least one CH2 domain from IgG3 and one CH2 domain from IgG1and and which has an altered glycosylation profile such that the ratio of fucose to mannose is 0.8:3 or less so that said antibody or antigen binding fragment has an enhanced effector function in comparison with an equivalent antibody or antigen-binding fragment with an immunoglobulin heavy chain constant region lacking said mutations and altered glycosylation profile, such that the antibody or antigen binding fragment has enhanced effector function, for example wherein it has one or more of the following functions, enhanced ADCC or enhanced CDC, for example wherein it has enhanced ADCC and enhanced CDCC.

In an alternative embodiment the antibody or antigen binding fragment has at least one IgG3 CH2 domain and at least one heavy chain constant domain from IgG1 wherein both IgG CH2 domains are mutated in accordance with the limitations described herein.

In one embodiment of the present invention there is provided an antibody preparation comprising an antibody comprising a heavy chain constant region or antigen binding fragment thereof which is linked to a heavy chain constant region which comprises a mutated and chimaeric heavy chain constant region wherein said antibody preparation has an altered glycosylation profile such that the antibody or antigen binding fragment has enhanced effector function, for example wherein it has one or more of the following functions, enhanced ADCC or enhanced CDC. In one embodiment the mutations are selected from positions 239 and 332 and 330, for example the mutations are selected from S239D and I332E and A330L. In a further embodiment the heavy chain constant region comprises at least one CH2 domain from IgG3 and one Ch2 domain from IgG1. In one embodiment the heavy chain constant region has an altered glycosylation profile such that the ratio of fucose to mannose is 0.8:3 or less so that said antibody or antigen binding fragment has an enhanced effector function in comparison with an equivalent non-chimaeric antibody or antigen-binding fragment thereof with an immunoglobulin heavy chain constant region lacking said mutations and altered glycosylation profile.

In one embodiment of the present invention there is provided a recombinant transformed, transfected or transduced host cell comprising at least one expression cassette, for example where the expression cassette comprises a polynucleotide encoding a heavy chain of an antibody or antigen binding fragment thereof according

to the invention described herein and further comprises a polynucleotide encoding a light chain of a antibody or antigen binding fragment thereof according to the invention described herein or where there are two expression cassettes and the 1st encodes the light chain and the second encodes the heavy chain. For example in one embodiment the first expression cassette comprises a polynucleotide encoding a heavy chain of an antibody comprising a constant region or antigen binding fragment thereof which is linked to a constant region according to the invention described herein and further comprises a second cassette comprising a polynucleotide encoding a light chain of an antibody comprising a constant region or antigen binding fragment thereof which is linked to a constant region according to the invention described herein for example the first expression cassette comprises a polynucleotide encoding a heavy chain selected from SEQ. ID. NO: 40, SEQ. ID. NO: 41 or SEQ. ID. NO: 67 or SEQ. ID. NO: 70 and a second expression cassette comprising a polynucleotide encoding a light chain selected from SEQ. ID. NO: 42 or SEQ. ID. NO: 69.

In another embodiment of the invention there is provided a stably transformed host cell comprising a vector comprising one or more expression cassettes encoding a heavy chain and/or a light chain of the antibody comprising a constant region or antigen binding fragment thereof which is linked to a constant region as described herein. For example such host cells may comprise a first vector encoding the light chain and a second vector encoding the heavy chain, for example the first vector encodes a heavy chain selected from SEQ. ID. NO: 37, SEQ. ID. NO: 38 or SEQ. ID. NO: 68 and a second vector encoding a light chain for example the light chain of SEQ ID NO: 39.

In another embodiment of the present invention there is provided a host cell according to the invention described herein wherein the cell is eukaryotic, for example where the cell is mammalian. Examples of such cell lines include CHO or NS0.

In another embodiment of the present invention there is provided a method for the production of an antibody comprising a constant region or antigen binding fragment thereof which is linked to a constant region according to the invention described herein which method comprises the step of culturing a host cell in a culture media, for example serum- free culture media.

Also provided is a method of producing an antibody as described herein comprising

expressing in a cell line an antibody or antigen binding fragment thereof which has been adapted to regulate the presence or absence of binding of fucose to an N-glycoside linked sugar chain which binds to the immunologically functional molecule.

In another embodiment of the present invention there is provided a method according to the invention described herein wherein said antibody is further purified to at least 95% or greater (e.g. 98% or greater) with respect to said antibody containing serum-free culture media.

In another embodiment of the present invention there is provided a pharmaceutical composition comprising an antibody comprising a constant region or antigen binding fragment thereof which is linked to a constant region according to the invention described herein and a pharmaceutically acceptable carrier.

In another embodiment of the present invention there is provided a kit-of-parts comprising the composition according to the invention described herein described together with instructions for use.

In another embodiment of the present invention there is provided a method of treating a human patient afflicted with rheumatoid arthritis which method comprises the step of administering a therapeutically effective amount of the antibody comprising a constant region or antigen binding fragment thereof which is linked to a constant region according to the invention described herein. The antibody comprising a constant region or antigen binding fragment thereof which is linked to a constant region may be in combination with a pharmaceutically acceptable carrier.

In another embodiment of the present invention there is provided a method of treating a human patient afflicted with cancer which method comprises the step of administering a therapeutically effective amount of antibody comprising a constant region or antigen binding fragment thereof which is linked to a constant region according to the invention described herein. The antibody comprising a constant region or antigen binding fragment thereof which is linked to a constant region may be in combination with a pharmaceutically acceptable carrier.

In another embodiment of the present invention there is provided a method of treating a human patient afflicted with diabetic retinopathy which method comprises the step of administering a therapeutically effective amount of antibody comprising a

constant region or antigen binding fragment thereof which is linked to a constant region according to the invention described herein. The antibody comprising a constant region or antigen binding fragment thereof which is linked to a constant region may be in combination with a pharmaceutically acceptable carrier.

In another embodiment of the present invention there is provided a method of treating a human patient afflicted with macular degeneration which method comprises the step of administering a therapeutically effective amount of antibody comprising a constant region or antigen binding fragment thereof which is linked to a constant region according to the invention described herein. The antibody comprising a constant region or antigen binding fragment thereof which is linked to a constant region may be in combination with a pharmaceutically acceptable carrier.

In a further embodiment of the present invention there is provided a method of treating a human patient afflicted with cancer which method comprises the step of administering a therapeutically effective amount of the pharmaceutical composition comprising an antibody comprising a constant region or antigen binding fragment thereof which is linked to a constant region according to the invention described herein and a pharmaceutically acceptable carrier.

In another embodiment of the present invention there is provided use of an antibody comprising a constant region or antigen binding fragment thereof which is linked to a constant region according to the invention described herein in the manufacture of a medicament for the treatment of a disease or disorder selected from the group consisting of neovascularisation diseases such as proliferative Diabetic retinopathy, neovascular glaucoma and Age related Macular degeneration (AMD) also diseases or disorders selected from the group consisting of; Rheumatoid arthritis, Psoriasis or Cancers for example: Acute Lymphoblastic Leukemia, Adrenocortical Carcinoma, AIDS-Related Cancers, AIDS Related Lymphoma, Anal Cancer, Childhood Cerebellar Astrocytoma, Childhood Cerebral Astrocytoma, Colorectal Cancer, Basal Cell Carcinoma, Extrahepatic Bile Duct Cancer, Bladder Cancer, Osteosarcorna/Malignant Fibrous Histiocytoma Bone Cancer, Brain Tumors (e.g., Brain Stem Glioma, Cerebellar Astrocytoma, Cerebral Astrocytoma/Malignant Glioma, Ependymoma, Medulloblastoma, Supratentorial Primitive Neuroectodermal Tumors, Visual Pathway and Hypothalamic Glioma), Breast Cancer, Bronchial Adenomas/Carcinoids, Burkitt's Lymphoma, Carcinoid Tumor, Gastrointestinal

Carcinoid Tumor, Carcinoma of Unknown Primary, Primary Central Nervous System, Cerebellar Astrocytoma, Cerebral Astrocytoma/Malignant Glioma, Cervical Cancer, Childhood Cancers, Chronic Lymphocytic Leukemia, Chronic Myelogenous Leukemia, Chronic Myeloproliferative Disorders, Colon Cancer, Colorectal Cancer, Cutaneous T-Cell Lymphoma, Endometrial Cancer, Ependymoma, Esophageal Cancer, Ewing's Family of Tumors, Extracranial Germ Cell Tumor, Extragonadal Germ Cell Tumor, Extrahepatic Bile Duct Cancer, Intraocular Melanoma Eye Cancer, Retinoblastoma Eye Cancer, Gallbladder Cancer, Gastric (Stomach) Cancer, Gastrointestinal Carcinoid Tumor, Germ Cell Tumors (e.g., Extracranial, Extragonadal, and Ovarian), Gestational Trophoblastic Tumor, Glioma (e.g., Adult, Childhood Brain Stem, Childhood Cerebral Astrocytoma, Childhood Visual Pathway and Hypothalamic), Hairy Cell Leukemia, Head and Neck Cancer, Hepatocellular (Liver) Cancer, Hodgkin's Lymphoma, Hypopharyngeal Cancer, Hypothalamic and Visual Pathway Glioma, Intraocular Melanoma, Islet Cell Carcinoma (Endocrine Pancreas), Kaposi's Sarcoma, Kidney (Renal Cell) Cancer, Laryngeal Cancer, Leukemia (e.g., Acute Lymphoblastic, Acute Myeloid, Chronic Lymphocyhc, Chronic Myelogenous, and Hairy Cell), Lip and Oral Cavity Cancer, Liver Cancer, Non-Small Cell Lung Cancer, Small Cell Lung Cancer, Lymphoma (e.g., AIDS-Related, Burkitt's, Cutaneous T-cell, Hodgkin's, Non-Hodgkin's, and Primary Central Nervous System), Waldenstrom's Macroglobulinemia, Malignant Fibrous Histiocytoma of Bone/Osteosarcoma, Medulloblastoma, Melanoma, Intraocular (Eye) Melanoma, Merkel Cell Carcinoma, Mesothelioma, Metastatic Squamous Neck Cancer with Occult Primary, Multiple Endocrine Neoplasia Syndrome, Multiple Myeloma/Plasma Cell Neoplasm, Mycosis Fungoides, Myelodysplastic Syndromes, Myelodysplastic/Myeloproliferative Diseases, Myelogenous Leukemia, Chronic Myeloid Leukemia, Multiple Myeloma, Chronic Myeloproliferative Disorders, Nasal Cavity and Paranasal Sinus Cancer, Nasopharyngeal Cancer, Neuroblastoma, Oral Cancer, Oropharyngeal Cancer, Osteosarcoma/Malignant Fibrous Histiocytoma of Bone, Ovarian Cancer, Ovarian Epithelial Cancer, Ovarian Germ Cell Tumor, Ovarian Low Malignant Potential Tumor, Pancreatic Cancer, Islet Cell Pancreatic Cancer, Paranasal Sinus and Nasal Cavity Cancer, Parathyroid Cancer, Penile Cancer, Pheochromocytoma, Pineoblastoma, Pituitary Tumor, Plasma Cell Neoplasm/Multiple Myeloma, Pleuropulmonary Blastoma, Primary Central Nervous System Lymphoma, Prostate Cancer, Rectal Cancer, Renal Cell (Kidney) Cancer, Renal Pelvis and Ureter Transitional Cell Cancer, Retinoblastoma, Rhabdomyosarcoma, Salivary Gland Cancer, Soft Tissue Sarcoma, Uterine Sarcoma, Sezary Syndrome, non-Melanoma Skin Cancer, Merkel Cell Skin

Carcinoma, Small Intestine Cancer, Soft Tissue Sarcoma, Squamous Cell Carcinoma, Cutaneous T-cell Lymphoma, Testicular Cancer, Thyrnoma, Thymic Carcinoma, Thyroid Cancer, Gestational Trophoblastic Tumor, Carcinoma of Unknown Primary Site, Cancer of Unknown Primary Site, Urethral Cancer, Endometrial Uterine Cancer, Uterine Sarcoma, Vaginal Cancer, Visual Pathway and Hypothalamic Glioma, Vulvar Cancer, Waldenstrom's Macroglobulinemia, and Wilms' Tumor.

In another embodiment of the present invention there is provided a method according to the invention described herein wherein the patient is afflicted with one or more of: proliferative Diabetic retinopathy, Age-related Macular degeneration (AMD), neovascular glaucoma, Rheumatoid Arthritis, Psoriasis, Colorectal Cancer, Breast Cancer, Prostate Cancer, Lung Cancer or Myeloma

Definitions

The term "antibody" is used herein in the broadest sense and specifically covers monoclonal antibodies (including full length monoclonal antibodies), polyclonal antibodies, multispecific antibodies (e.g. bispecific antibodies), and antibody fragments so long as they exhibit the desired biological activity. These are explained later in further detail.

The term "monoclonal antibody" as used herein refers to an antibody obtained from a population of substantially homogenous antibodies i.e. the individual antibodies comprising the population are identical except for possible naturally occurring mutations that may be present in minor amounts. Monoclonal antibodies are highly specific being directed against a single antigenic binding site. Furthermore, in contrast to polyclonal antibody preparations which typically include different antibodies directed against different determinants (epitopes), each monoclonal antibody is directed against a single determinant on the antigen.

"Identity," means, for polynucleotides and polypeptides, as the case may be, the comparison calculated using an algorithm provided in (1) and (2) below:

(1) Identity for polynucleotides is calculated by multiplying the total number of nucleotides in a given sequence by the integer defining the percent identity divided by 100 and then subtracting that product from said total number of nucleotides in said sequence, or:

$$nn \leq xn - (xn \bullet y),$$

wherein nn is the number of nucleotide alterations, xn is the total number of nucleotides in a given sequence, y is 0.95 for 95%, 0.97 for 97% or 1.00 for 100%, and • is the symbol for the multiplication operator, and wherein any non-integer product of xn and y is rounded down to the nearest integer prior to subtracting it from xn. Alterations of a polynucleotide sequence encoding a polypeptide may create nonsense, missense or frameshift mutations in this coding sequence and thereby alter the polypeptide encoded by the polynucleotide following such alterations.

(2) Identity for polypeptides is calculated by multiplying the total number of amino acids by the integer defining the percent identity divided by 100 and then subtracting that product from said total number of amino acids, or:

$$na \leq xa - (xa \bullet y),$$

wherein na is the number of amino acid alterations, xa is the total number of amino acids in the sequence, y is 0.95 for 95%, 0.97 for 97% or 1.00 for 100%, and • is the symbol for the multiplication operator, and wherein any non-integer product of xa and y is rounded down to the nearest integer prior to subtracting it from xa.

The term "Variant(s)" as used herein, refers to a polynucleotide or polypeptide that differs from a reference polynucleotide or polypeptide respectively, but retains essential properties. A typical variant of a polynucleotide differs in nucleotide sequence from another, reference polynucleotide. Changes in the nucleotide sequence of the variant may or may not alter the amino acid sequence of a polypeptide encoded by the reference polynucleotide. Nucleotide changes may result in amino acid substitutions, additions, deletions, fusion proteins and truncations in the polypeptide encoded by the reference sequence, as discussed below. A typical variant of a polypeptide differs in amino acid sequence from another, reference polypeptide. Generally, differences are limited so that the sequences of the reference polypeptide and the variant are closely similar overall and, in many regions, identical. A variant and reference polypeptide may differ in amino acid sequence by one or more substitutions, additions, deletions in any combination. A substituted or inserted amino acid residue may or may not be one encoded by the genetic code. It is well recognised in the art that certain amino acid substitutions are regarded as being "conservative". Amino acids are divided into groups based on common side-chain properties and substitutions within groups that maintain all or substantially all of the binding affinity of the antibody of the invention or antigen binding fragment thereof are regarded as conservative substitutions, see table below:

Side chain	Members
Hydrophobic	Met, Ala, Val, Leu, Ile
neutral hydrophilic	Cys, Ser, Thr
Acidic	Asp, Glu
Basic	Asn, Gln, His, Lys, Arg
residues that influence chain orientation	Gly, Pro
Aromatic	Trp, Tyr, Phe

In some aspects of the invention variants in which several, for example 5-10, 1-5, 1-3, 1-2 amino acid residues or 1 amino acid residue are substituted, deleted, or added in any combination may be included. A variant of a polynucleotide or polypeptide may be a naturally occurring such as an allelic variant, or it may be a variant that is

not known to occur naturally. Non-naturally occurring variants of polynucleotides and polypeptides may be made by mutagenesis techniques, by direct synthesis, and by other recombinant methods known to skilled artisans.

"Isolated" means altered "by the hand of man" from its natural state, has been changed or removed from its original environment, or both. For example, a polynucleotide or a polypeptide naturally present in a living organism is not "isolated," but the same polynucleotide or polypeptide separated from the coexisting materials of its natural state is "isolated", including but not limited to when such polynucleotide or polypeptide is introduced back into a cell, even if the cell is of the same species or type as that from which the polynucleotide or polypeptide was separated.

Throughout the present specification and the accompanying claims the term "comprising" and "comprises" incorporates "consisting of" and "consists of". That is, these words are intended to convey the possible inclusion of other elements or integers not specifically recited, where the context allows.

The term "glycosylation profile" as used herein refers to the levels of glycosylation in an antibody population.

The term "specifically binds" as used throughout the present specification in relation to antibodies and antigen binding fragments thereof of the invention means that the antibody binds human IGF-1R (hIGF-1R) with no or insignificant binding to other human proteins. The term however does not exclude the fact that antibodies of the invention may also be cross-reactive with other forms of IGF-1R, for example primate IGF-1R.

The term "neutralises" as used throughout the present specification in relation to antibodies and antigen binding fragments thereof of the invention means that the biological activity of IGF-1R is reduced in the presence of the antibodies and antigen binding fragments thereof of the present invention in comparison to the activity of IGF-1R in the absence of such antibodies and antigen binding fragments thereof. Neutralisation may be due to but not limited to one or more of blocking ligand binding, preventing the ligand activating the receptor, down regulating the IGF-1R or affecting effector functionality. Levels of neutralisation can be measured in several ways, for example by use of the assays as set out in the examples below, for example in a LISN cell proliferation assay which may be carried out for example as

described in Example 23. The neutralisation of IGF-1R in this assay is measured by assessing the decreased tumour cell proliferation in the presence of neutralising antibody.

Levels of neutralisation can also be measured, for example in a receptor phosphorylation assay which may be carried out for example as described in Example 13. The neutralisation of IGF-1R in this assay is measured by assessing the inhibition of receptor phosphorylation in the presence of neutralising antibody.

If an antibody or antigen binding fragment thereof is capable of neutralisation then this is indicative of inhibition of the interaction between human IGF-1R binding proteins for example hIGF-I or hIGF-II and its receptor. Antibodies which are considered to have neutralising activity against human IGF-1R would have an IC_{50} of less than 10 micrograms/ml, or less than 5 micrograms/ml, or less than 2 micrograms/ml, or less than 1 microgram/ml in the LISN cell proliferation assay or receptor phosphorylation assay as set out in Examples 23 and Example 13 respectively.

In an alternative aspect of the present invention there is provided antibodies or antigen binding fragments thereof which have equivalent neutralising activity to the antibodies exemplified herein, for example antibodies which retain the neutralising activity of H0L0 and H0L0 IgG1m(AA) and H1L0 and H10L0 IgG1m(AA) in the LISN cell proliferation assay or receptor phosphorylation assay as set out in Examples 23 and 13 respectively. 5

Throughout this specification, amino acid residues in antibody sequences are numbered according to the Kabat scheme. Similarly, the terms "CDR", "CDRL1", "CDRL2", "CDRL3", "CDRH1", "CDRH2", "CDRH3" follow the Kabat numbering system as set forth in Kabat et al; Sequences of proteins of Immunological Interest NIH, 1987. It will be apparent to those skilled in the art that there are alternative definitions of CDR sequences such as for example those set out in Chothia et al. (1989).

It will be apparent to those skilled in the art that the term "derived" is intended to define not only the source in the sense of it being the physical origin for the material but also to define material which is structurally identical (in terms of primary amino acid sequence) to the material but which does not originate from the reference source. Thus "residues found in the donor antibody from which CDRH3 is derived"

need not necessarily have been purified from the donor antibody.

The term "stability" as used throughout the present specification in relation to antibodies and antigen binding fragments thereof of the invention means that the activity of the antibody or antigen binding fragment when determined by direct binding ELISA is comparable 12 days after incubation in serum to the EC-50 starting values at -20°C, 4°C or 37°C.

A "chimeric antibody" refers to a type of engineered antibody which contains a naturally-occurring variable domain (light chain and heavy chains) derived from a donor antibody in association with light and heavy chain constant regions derived from an acceptor antibody.

A "humanised antibody" refers to a type of engineered antibody having its CDRs derived from a non-human donor immunoglobulin, the remaining immunoglobulin-derived parts of the molecule being derived from one (or more) human immunoglobulin(s). In addition, framework support residues may be altered to preserve binding affinity (see, e.g., Queen et al., Proc. Natl Acad Sci USA, 86:10029-10032 (1989), Hodgson et al., Bio/Technology, 9:421 (1991)). A suitable human acceptor antibody may be one selected from a conventional database, e.g., the KABAT® database, Los Alamos database, and Swiss Protein database, by homology to the nucleotide and amino acid sequences of the donor antibody. A human antibody characterized by a homology to the framework regions of the donor antibody (on an amino acid basis) may be suitable to provide a heavy chain constant region and/or a heavy chain variable framework region for insertion of the donor CDRs. A suitable acceptor antibody capable of donating light chain constant or variable framework regions may be selected in a similar manner. It should be noted that the acceptor antibody heavy and light chains are not required to originate from the same acceptor antibody. The prior art describes several ways of producing such humanised antibodies – see for example EP-A-0239400 and EP-A-054951.

The term "donor antibody" refers to an antibody (monoclonal, and/or recombinant) which contributes the amino acid sequences of its variable domains, CDRs, or other functional fragments or analogs thereof to a first immunoglobulin partner, so as to provide the altered immunoglobulin coding region and resulting expressed altered antibody with the antigenic specificity and neutralizing activity characteristic of the donor antibody.

The term "acceptor antibody" refers to an antibody (monoclonal and/or recombinant) heterologous to the donor antibody, which contributes all (or any portion, but preferably all) of the amino acid sequences encoding its heavy and/or light chain framework regions and/or its heavy and/or light chain constant regions to the first immunoglobulin partner. The human antibody is the acceptor antibody.

"CDRs" are defined as the complementarity determining region amino acid sequences of an antibody which are the hypervariable domains of immunoglobulin heavy and light chains. See, e.g., Kabat et al., Sequences of Proteins of Immunological Interest, 4th Ed., U.S. Department of Health and Human Services, National Institutes of Health (1987). There are three heavy chain and three light chain CDRs (or CDR regions) in the variable portion of an immunoglobulin. Thus, "CDRs" as used herein refers to all three heavy chain CDRs, or all three light chain CDRs (or both all heavy and all light chain CDRs, if appropriate). The structure and protein folding of the antibody may mean that other residues are considered part of the antigen binding region and would be understood to be so by a skilled person. See for example Chothia et al., (1989) Conformations of immunoglobulin hypervariable domains; Nature 342, p877-883.

CDRs provide the majority of contact residues for the binding of the antibody to the antigen or epitope. CDRs of interest in this invention are derived from donor antibody variable heavy and light chain sequences, and include analogs of the naturally occurring CDRs, which analogs also share or retain the same antigen binding specificity and/or neutralizing ability as the donor antibody from which they were derived.

The terms "V_H" and "V_L" are used herein to refer to the heavy chain variable domain and light chain variable domain respectively of an antibody.

The term "Effector Function" as used herein is meant to refer to one or more of Antibody dependant cell mediated cytotoxic activity (ADCC) and complement–dependant cytotoxic activity (CDC) mediated responses, Fc-mediated phagocytosis and antibody recycling via the FcRn receptor. The interaction between the constant region of an antibody and various Fc receptors (FcR) is believed to mediate the effector functions of the antibody. Significant biological effects can be a consequence of effector functionality, in particular, antibody-dependent cellular cytotoxicity (ADCC), fixation of complement (complement dependent cytotoxicity or CDC),

phagocytosis (antibody-dependent cell-mediated phagocytosis or ADCP) and half-life/clearance of the antibody. Usually, the ability to mediate effector function requires binding of the antibody to an antigen and not all antibodies will mediate every effector function.

Effector function can be measured in a number of ways including for example via binding of the FcγRIII to Natural Killer cells or via FcγRI to monocytes/macrophages to measure for ADCC effector function. For example the antibody or antigen binding fragment of the present invention has an increased ADCC effector function when measured against the equivalent wild type antibody or antigen binding fragment thereof in a Natural Killer cell assay. Examples of such assays can be found in Shields *et al*, 2001 The Journal of Biological Chemistry, Vol. 276, p6591-6604; Chappel *et al*, 1993 The Journal of Biological Chemistry, Vol 268, p25124-25131; Lazar et al, 2006 PNAS, 103; 4005-4010.

Examples of assays to determine CDC function include that described in 1995 J Imm Meth 184:29-38.

Various modifications to the heavy chain constant region of antibodies may be carried out depending on the desired effector property. Human constant regions which essentially lack the functions of a) activation of complement by the classical pathway; and b) mediating antibody-dependent cellular cytotoxicity include the IgG4 constant region and the IgG2 constant region. IgG1 constant regions containing specific mutations have separately been described to reduce binding to Fc receptors and therefore reduce ADCC and CDC (Duncan et al. Nature 1988, 332; 563-564; Lund et al. J. Immunol. 1991, 147; 2657-2662; Chappel et al. PNAS 1991, 88; 9036-9040; Burton and Woof, Adv. Immunol. 1992, 51;1-84; Morgan et al., Immunology 1995, 86; 319-324; Hezareh et al., J. Virol. 2001, 75 (24); 12161-12168). Human IgG1 constant regions containing specific mutations or altered glycosylation on residue Asn297 have also been described to enhance binding to Fc receptors. These have also been shown to enhance ADCC and CDC, in some cases (Lazar et al. PNAS 2006, 103; 4005-4010; Shields et al. J Biol Chem 2001, 276; 6591-6604; Nechansky et al. Mol Immunol, 2007, 44; 1815-1817).

For IgG antibodies, effector functionalities including ADCC and ADCP are mediated by the interaction of the heavy chain constant region with a family of Fcγ receptors present on the surface of immune cells. In humans these include FcγRI (CD64), FcγRII (CD32) and FcγRIII (CD16). Interaction between the antibody bound to antigen and the formation of the Fc/ Fcγ complex induces a range of effects including

cytotoxicity, immune cell activation, phagocytosis and release of inflammatory cytokines. Specific substitutions in the constant region (including S239D/I332E) are know to increase the affinity of the heavy chain constant region for certain Fc receptors, thus enhancing the effector functionality of the antibody (Lazar et al. PNAS 2006).

1. Antibody Structures

1.1 Intact Antibodies

Intact antibodies include heteromultimeric glycoproteins comprising at least two heavy and two light chains. Aside from IgM, intact antibodies are usually heterotetrameric glycoproteins of approximately 150Kda, composed of two identical light (L) chains and two identical heavy (H) chains. Typically, each light chain is linked to a heavy chain by one covalent disulfide bond while the number of disulfide linkages between the heavy chains of different immunoglobulin isotypes varies. Each heavy and light chain also has intrachain disulfide bridges. Each heavy chain has at one end a variable domain (V_H) followed by a number of constant regions (CH_1, CH_2, CH_3). Each light chain has a variable domain (V_L) and a constant region at its other end; the heavy chain constant region of the light chain is aligned with the first constant region of the heavy chain and the light chain variable domain is aligned with the variable domain of the heavy chain. The light chains of antibodies from most vertebrate species can be assigned to one of two types called Kappa and Lambda based on the amino acid sequence of the constant region. Depending on the amino acid sequence of the heavy chain constant region of their heavy chains, human antibodies can be assigned to five different classes, IgA, IgD, IgE, IgG and IgM. IgG and IgA can be further subdivided into subclasses, IgG1, IgG2, IgG3 and IgG4; and IgA1 and IgA2. Species variants exist with mouse and rat having at least IgG2a, IgG2b. The variable domain of the antibody confers binding specificity upon the antibody with certain regions displaying particular variability called complementarity determining regions (CDRs). The more conserved portions of the variable domain are called Framework regions (FR). The variable domains of intact heavy and light chains each comprise four FR connected by three CDRs. The CDRs in each chain are held together in close proximity by the FR regions and with the CDRs from the other chain contribute to the formation of the antigen binding site of antibodies. The constant regions are not directly involved in the binding of the antibody to the antigen but exhibit various effector functions such as participation in antibody dependent cell-

mediated cytotoxicity (ADCC), phagocytosis via binding to Fcγ receptor, half-life/clearance rate via neonatal Fc receptor (FcRn) and complement dependent cytotoxicity via the C1q component of the complement cascade.

1.1.2 Human antibodies

Human antibodies may be produced by a number of methods known to those of skill in the art. Human antibodies can be made by the hybridoma method using human myeloma or mouse-human heteromyeloma cells lines see Kozbor J.Immunol 133, 3001, (1984) and Brodeur, Monoclonal Antibody Production Techniques and Applications, pp51-63 (Marcel Dekker Inc, 1987). Alternative methods include the use of phage libraries or transgenic mice both of which utilize human variable domain repertories (see Winter G, (1994), Annu.Rev.Immunol 12,433-455, Green LL (1999), J.Immunol.methods 231, 11-23).

Several strains of transgenic mice are now available wherein their mouse immunoglobulin loci has been replaced with human immunoglobulin gene segments (see Tomizuka K, (2000) PNAS 97,722-727; Fishwild D.M (1996) Nature Biotechnol. 14,845-851, Mendez MJ, 1997, Nature Genetics, 15,146-156). Upon antigen challenge such mice are capable of producing a repertoire of human antibodies from which antibodies of interest can be selected. Of particular note is the Trimera™ system (see Eren R et al, (1998) Immunology 93:154-161) where human lymphocytes are transplanted into irradiated mice, the Selected Lymphocyte Antibody System (SLAM, see Babcook et al, PNAS (1996) 93:7843-7848) where human (or other species) lymphocytes are effectively put through a massive pooled in vitro antibody generation procedure followed by deconvulated, limiting dilution and selection procedure and the Xenomouse II™ (Abgenix Inc). An alternative approach is available from Morphotek Inc using the Morphodoma™ technology.

Phage display technology can be used to produce human antibodies (and fragments thereof), see McCafferty; Nature, 348, 552-553 (1990) and Griffiths AD et al (1994) EMBO 13:3245-3260. According to this technique antibody variable domain genes are cloned in frame into either a major or minor coat of protein gene of a filamentous bacteriophage such as M13 or fd and displayed (usually with the aid of a helper phage) as functional antigen binding fragments thereof on the surface of the phage particle. Selections based on the functional properties of the antibody result in selection of the gene encoding the antibody exhibiting those properties. The phage

display technique can be used to select antigen specific antibodies from libraries made from human B cells taken from individuals afflicted with a disease or disorder described above or alternatively from unimmunized human donors (see Marks; J.Mol.Bio. 222,581-597, 1991). Where an intact human antibody is desired comprising a constant domain it is necessary to reclone the phage displayed derived fragment into a mammalian expression vectors comprising the desired constant regions and establishing stable expressing cell lines.

The technique of affinity maturation (Marks; Bio/technol 10,779-783 (1992)) may be used to improve binding affinity wherein the affinity of the primary human antibody is improved by sequentially replacing the H and L chain variable domains with naturally occurring variants and selecting on the basis of improved binding affinities. Variants of this technique such as "epitope imprinting" are now also available see WO 93/06213. See also Waterhouse; Nucl.Acids Res 21, 2265-2266 (1993).

1.2 Chimaeric and Humanised Antibodies

The use of intact non-human antibodies in the treatment of human diseases or disorders carries with it the potential for the now well established problems of immunogenicity, that is the immune system of the patient may recognise the non-human intact antibody as non-self and mount a neutralising response. This is particularly evident upon multiple administration of the non-human antibody to a human patient. Various techniques have been developed over the years to overcome these problems and generally involve reducing the composition of non-human amino acid sequences in the intact antibody whilst retaining the relative ease in obtaining non-human antibodies from an immunised animal e.g. mouse, rat or rabbit. Broadly two approaches have been used to achieve this. The first are chimaeric antibodies, which generally comprise a non-human (e.g. rodent such as mouse) variable domain fused to a human constant region. Because the antigen-binding site of an antibody is localised within the variable domains the chimaeric antibody retains its binding affinity for the antigen but acquires the effector functions of the human constant region and are therefore able to perform effector functions such as described supra. Chimaeric antibodies are typically produced using recombinant DNA methods. DNA encoding the antibodies (e.g. cDNA) is isolated and sequenced using conventional procedures (e.g. by using oligonucleotide probes that are capable of binding specifically to genes encoding the H and L chains of the antibody of the invention. Hybridoma cells serve as a typical source of such DNA. Once isolated, the DNA is placed into expression vectors which are then transfected into host cells such as *E.Coli*, COS cells, CHO

cells or myeloma cells that do not otherwise produce immunoglobulin protein to obtain synthesis of the antibody. The DNA may be modified by substituting the coding sequence for human L and H chains for the corresponding non-human (e.g. murine) H and L constant regions see e.g. Morrison; PNAS 81, 6851 (1984).

The second approach involves the generation of humanised antibodies wherein the non-human content of the antibody is reduced by humanizing the variable domains. Two techniques for humanisation have gained popularity. The first is humanisation by CDR grafting. CDRs build loops close to the antibody's N-terminus where they form a surface mounted in a scaffold provided by the framework regions. Antigen-binding specificity of the antibody is mainly defined by the topography and by the chemical characteristics of its CDR surface. These features are in turn determined by the conformation of the individual CDRs, by the relative disposition of the CDRs, and by the nature and disposition of the side chains of the residues comprising the CDRs. A large decrease in immunogenicity can be achieved by grafting only the CDRs of a non-human (e.g. murine) antibodies ("donor" antibodies) onto human framework ("acceptor framework") and constant regions (see Jones et al (1986) Nature 321,522-525 and Verhoeyen M et al (1988) Science 239, 1534-1536). However, CDR grafting per se may not result in the complete retention of antigen-binding properties and it is frequently found that some framework residues (sometimes referred to as “back mutations”) of the donor antibody need to be preserved in the humanised molecule if significant antigen-binding affinity is to be recovered (see Queen C et al (1989) PNAS 86, 10,029-10,033, Co, M et al (1991) Nature 351, 501-502). In this case, human variable domains showing the greatest sequence homology to the non-human donor antibody are chosen from a database in order to provide the human framework (FR). The selection of human FRs can be made either from human consensus or individual human antibodies. Where necessary key residues from the donor antibody are substituted into the human acceptor framework to preserve CDR conformations. Computer modelling of the antibody maybe used to help identify such structurally important residues, see WO99/48523.

Alternatively, humanisation maybe achieved by a process of "veneering". A statistical analysis of unique human and murine immunoglobulin heavy and light chain variable domains revealed that the precise patterns of exposed residues are different in human and murine antibodies, and most individual surface positions have a strong preference for a small number of different residues (see Padlan E.A. et al; (1991) Mol.Immunol.28, 489-498 and Pedersen J.T. et al (1994) J.Mol.Biol. 235; 959-973).

Therefore it is possible to reduce the immunogenicity of a non-human Fv by replacing exposed residues in its framework regions that differ from those usually found in human antibodies. Because protein antigenicity may be correlated with surface accessibility, replacement of the surface residues may be sufficient to render the mouse variable domain "invisible" to the human immune system (see also Mark G.E. et al (1994) in Handbook of Experimental Pharmacology vol.113: The pharmacology of monoclonal Antibodies, Springer-Verlag, pp105-134). This procedure of humanisation is referred to as "veneering" because only the surface of the antibody is altered, the supporting residues remain undisturbed.

1.3 Bispecific antibodies

A bispecific antibody is an antibody having binding specificities for at least two different epitopes. Methods of making such antibodies are known in the art. Traditionally, the recombinant production of bispecific antibodies is based on the co-expression of two immunoglobulin H chain-L chain pairs, where the two H chains have different binding specificities see Millstein et al, Nature 305 537-539 (1983), WO93/08829 and Traunecker et al EMBO, 10, 1991, 3655-3659. Because of the random assortment of H and L chains, a potential mixture of ten different antibody structures are produced of which only one has the desired binding specificity. An alternative approach involves fusing the variable domains with the desired binding specificities to heavy chain constant region comprising at least part of the hinge region, CH2 and CH3 regions. In one embodiment the CH1 region containing the site necessary for light chain binding is present in at least one of the fusions. DNA encoding these fusions, and if desired the L chain are inserted into separate expression vectors and are then co-transfected into a suitable host organism. It is possible though to insert the coding sequences for two or all three chains into one expression vector. In one approach, the bispecific antibody is composed of a H chain with a first binding specificity in one arm and a H-L chain pair, providing a second binding specificity in the other arm, see WO94/04690. Also see Suresh et al Methods in Enzymology 121, 210, 1986.

In one embodiment of the invention there is provided a bispecific antibody wherein at least one binding specificity of said antibody is for hIGF-1R, and said antibody neutralises the activity of hIGF-1R. Such antibodies may further comprise a human constant region of the IgG isotype, e.g. IgG1, IgG2, IgG3 or IgG4. Antibodies of the present invention may also be multispecific, for example multispecific antibodies

formed by assembly of a number of antigen-binding fragments.

1.4 Antigen Binding Fragments

Such antigen binding fragments comprise a partial heavy or light chain variable sequence (e.g., minor deletions at the amino or carboxy terminus of the immunoglobulin variable domain) which retains the same antigen binding specificity and the same or similar neutralizing ability as the antibody from which the fragment was derived.

In certain embodiments of the invention there is provided antigen binding fragments which neutralise the activity of hIGF-1R. Such fragments may be functional antigen binding fragments of intact and/or humanised and/or chimaeric antibodies such as Fab, Fab', $F(ab')_2$, Fv, ScFv fragments of the antibodies described supra. Traditionally such fragments are produced by the proteolytic digestion of intact antibodies by e.g. papain digestion (see for example, WO 94/29348) but may be produced directly from recombinantly transformed host cells. For the production of ScFv, see Bird et al ;(1988) Science, 242, 423-426. In addition, antigen binding fragments may be produced using a variety of engineering techniques as described below.

Fv fragments appear to have lower interaction energy of their two chains than Fab fragments. To stabilise the association of the V_H and V_L domains, they have been linked with peptides (Bird et al, (1988) Science 242, 423-426, Huston et al, PNAS, 85, 5879-5883), disulphide bridges (Glockshuber et al, (1990) Biochemistry, 29, 1362-1367) and "knob in hole" mutations (Zhu et al (1997), Protein Sci., 6, 781-788). ScFv fragments can be produced by methods well known to those skilled in the art see Whitlow et al (1991) Methods companion Methods Enzymol, 2, 97-105 and Huston et al (1993) Int.Rev.Immunol 10, 195-217. ScFv may be produced in bacterial cells such as E.Coli but are more preferably produced in eukaryotic cells. One disadvantage of ScFv is the monovalency of the product, which precludes an increased avidity due to polyvalent binding, and their short half-life. Attempts to overcome these problems include bivalent $(ScFv')_2$ produced from ScFV containing an additional C terminal cysteine by chemical coupling (Adams et al (1993) Can.Res 53, 4026-4034 and McCartney et al (1995) Protein Eng. 8, 301-314) or by spontaneous site-specific dimerization of ScFv containing an unpaired C terminal cysteine residue (see Kipriyanov et al (1995) Cell. Biophys 26, 187-204).

Alternatively, ScFv can be forced to form multimers by shortening the peptide linker to 3 to 12 residues to form "diabodies", see Holliger et al PNAS (1993), 90, 6444-6448. Reducing the linker still further can result in ScFV trimers ("triabodies", see Kortt et al (1997) Protein Eng, 10, 423-433) and tetramers ("tetrabodies", see Le Gall et al (1999) FEBS Lett, 453, 164-168). Construction of bivalent ScFV molecules can also be achieved by genetic fusion with protein dimerizing motifs to form "miniantibodies" (see Pack et al (1992) Biochemistry 31, 1579-1584) and "minibodies" (see Hu et al (1996), Cancer Res. 56, 3055-3061). ScFv-Sc-Fv tandems ((ScFV)2) may also be produced by linking two ScFv units by a third peptide linker, see Kurucz et al (1995) J.Immol.154, 4576-4582. Bispecific diabodies can be produced through the noncovalent association of two single chain fusion products consisting of V_H domain from one antibody connected by a short linker to the V_L domain of another antibody, see Kipriyanov et al (1998), Int.J.Can 77,763-772. The stability of such bispecific diabodies can be enhanced by the introduction of disulphide bridges or "knob in hole" mutations as described supra or by the formation of single chain diabodies (ScDb) wherein two hybrid ScFv fragments are connected through a peptide linker see Kontermann et al (1999) J.Immunol.Methods 226 179-188. Tetravalent bispecific molecules are available by e.g. fusing a ScFv fragment to the CH3 domain of an IgG molecule or to a Fab fragment through the hinge region see Coloma et al (1997) Nature Biotechnol. 15, 159-163. Alternatively, tetravalent bispecific molecules have been created by the fusion of bispecific single chain diabodies (see Alt et al, (1999) FEBS Lett 454, 90-94. Smaller tetravalent bispecific molecules can also be formed by the dimerization of either ScFv-ScFv tandems with a linker containing a helix-loop-helix motif (DiBi miniantibodies, see Muller et al (1998) FEBS Lett 432, 45-49) or a single chain molecule comprising four antibody variable domains (V_H and V_L) in an orientation preventing intramolecular pairing (tandem diabody, see Kipriyanov et al, (1999) J.Mol.Biol. 293, 41-56). Bispecific $F(ab')_2$ fragments can be created by chemical coupling of Fab' fragments or by heterodimerization through leucine zippers (see Shalaby et al, (1992) J.Exp.Med. 175, 217-225 and Kostelny et al (1992), J.Immunol. 148, 1547-1553).

The phrase an “immunoglobulin single variable domain” refers to an antibody variable domain (V_H, V_{HH}, V_L) that specifically binds an antigen or epitope independently of a different V region or domain. An immunoglobulin single variable domain can be present in a format (*e.g.,* homo- or hetero-multimer) with other, different variable regions or variable domains where the other regions or domains are not required for antigen binding by the single immunoglobulin variable domain (*i.e.*, where the immunoglobulin single variable domain binds antigen independently of the

additional variable domains).

Also available are isolated V_H and V_L domains (Domantis plc), see US 6, 248,516; US 6,291,158; US 6, 172,197 these are known as domain antibodies. A "domain antibody" or "dAb" is the same as an "immunoglobulin single variable domain" which is capable of binding to an antigen as the term is used herein. An immunoglobulin single variable domain may be a human antibody variable domain, but also includes single antibody variable domains from other species such as rodent (for example, as disclosed in WO 00/29004, nurse shark and *Camelid* V_{HH} dAbs. Camelid V_{HH} are immunoglobulin single variable domain polypeptides that are derived from species including camel, llama, alpaca, dromedary, and guanaco, which produce heavy chain antibodies naturally devoid of light chains. Such V_{HH} domains may be humanised according to standard techniques available in the art, and such domains are still considered to be "domain antibodies" according to the invention. As used herein "V_H includes camelid V_{HH} domains.

In one embodiment there is provided an antigen binding fragment (e.g. ScFv, Fab, Fab', $F(ab')_2$) or an engineered antigen binding fragment as described supra that specifically binds hIGF-1R neutralises the activity of hIGF-1R. The antigen binding fragment may comprise one or more of the following sequences CDRH3 as set out in SEQ. ID. NO: 1, CDRH2 as set out in SEQ. ID. NO: 2, CDRH1 as set out in SEQ. ID. NO: 3, CDRL1 as set out in SEQ. ID. NO: 4, CDRL2 as set out in SEQ. ID. NO: 5, and CDRL3 as set out in SEQ. ID. NO: 6.

1.5 Heteroconjugate antibodies

Heteroconjugate antibodies also form an embodiment of the present invention. Heteroconjugate antibodies are composed of two covalently joined antibodies formed using any convenient cross-linking methods. See, for example, US 4,676,980. Additionally, combinations of antibodies and antigen binding fragments are included within the present invention for example, one or more domain antibodies and or ScFv bound to a monoclonal antibody.

1.6 Other Modifications.

The interaction between the constant region of an antibody and various Fc receptors (FcγR) is believed to mediate the effector functions of the antibody which include

antibody-dependent cellular cytotoxicity (ADCC), fixation of complement, phagocytosis and half-life/clearance of the antibody. Various modifications to the constant region of antibodies of the invention may be carried out depending on the desired property. For example, specific mutations in the constant region to render an otherwise lytic antibody, non-lytic is detailed in EP 0629 240B1 and EP 0307 434B2 or one may incorporate a salvage receptor binding epitope into the antibody to increase serum half life see US 5,739,277. There are five currently recognised human Fcγ receptors, FcγR (I), FcγRIIa, FcγRIIb, FcγRIIIa and neonatal FcRn. Shields et al, (2001) J.Biol.Chem 276, 6591-6604 demonstrated that a common set of IgG1 residues is involved in binding all FcγRs, while FcγRII and FcγRIII utilize distinct sites outside of this common set. One group of IgG1 residues reduced binding to all FcγRs when altered to alanine: Pro-238, Asp-265, Asp-270, Asn-297 and Pro-239. All are in the IgG CH2 domain and clustered near the hinge joining CH1 and CH2. While FcγRI utilizes only the common set of IgG1 residues for binding, FcγRII and FcγRIII interact with distinct residues in addition to the common set. Alteration of some residues reduced binding only to FcγRII (e.g. Arg-292) or FcγRIII (e.g. Glu-293). Some variants showed improved binding to FcγRII or FcγRIII but did not affect binding to the other receptor (e.g. Ser-267Ala improved binding to FcγRII but binding to FcγRIII was unaffected). Other variants exhibited improved binding to FcγRII or FcγRIII with reduction in binding to the other receptor (e.g. Ser-298Ala improved binding to FcγRIII and reduced binding to FcγRII). For FcγRIIIa, the best binding IgG1 variants had combined alanine substitutions at Ser-298, Glu-333 and Lys-334. The neonatal FcRn receptor is believed to be involved in both antibody clearance and the transcytosis across tissues (see Junghans R.P (1997) Immunol.Res 16. 29-57 and Ghetie et al (2000) Annu.Rev.Immunol. 18, 739-766). Human IgG1 residues determined to interact directly with human FcRn includes Ile253, Ser254, Lys288, Thr307, Gln311, Asn434 and His435. Switches at any of these positions described in this section may enable increased serum half-life and/or altered effector properties of antibodies of the invention.

Other modifications include glycosylation variants of the antibodies of the invention. Glycosylation of antibodies at conserved positions in their constant regions is known to have a profound effect on antibody function, particularly effector functioning such as those described above, see for example, Boyd et al (1996), Mol.Immunol. 32, 1311-1318. Glycosylation variants of the antibodies or antigen binding fragments thereof of the present invention wherein one or more carbohydrate moiety is added,

substituted, deleted or modified are contemplated. Introduction of an asparagine-X-serine or asparagine-X-threonine motif creates a potential site for enzymatic attachment of carbohydrate moieties and may therefore be used to manipulate the glycosylation of an antibody. In Raju et al (2001) Biochemistry 40, 8868-8876 the terminal sialyation of a TNFR-IgG immunoadhesin was increased through a process of regalactosylation and/or resialylation using beta-1, 4-galactosyltransferace and/or alpha, 2,3 sialyltransferase. Increasing the terminal sialylation is believed to increase the half-life of the immunoglobulin. Antibodies, in common with most glycoproteins, are typically produced as a mixture of glycoforms. This mixture is particularly apparent when antibodies are produced in eukaryotic, particularly mammalian cells. A variety of methods have been developed to manufacture defined glycoforms, see Zhang et al Science (2004), 303, 371, Sears et al, Science, (2001) 291, 2344, Wacker et al (2002) Science, 298 1790, Davis et al (2002) Chem.Rev. 102, 579, Hang et al (2001) Acc.Chem.Res 34, 727. Thus the invention contemplates a plurality of (monoclonal) antibodies (which maybe of the IgG isotype, e.g. IgG1) as herein described comprising a defined number (e.g. 7 or less, for example 5 or less such as two or a single) glycoform(s) of said antibodies or antigen binding fragments thereof.

Further embodiments of the invention include antibodies of the invention or antigen binding fragments thereof coupled to a non-proteinaeous polymer such as polyethylene glycol (PEG), polypropylene glycol or polyoxyalkylene. Conjugation of proteins to PEG is an established technique for increasing half-life of proteins, as well as reducing antigenicity and immunogenicity of proteins. The use of PEGylation with different molecular weights and styles (linear or branched) has been investigated with intact antibodies as well as Fab' fragments, see Koumenis I.L. et al (2000) Int.J.Pharmaceut. 198:83-95.

2. Production Methods

Antibodies of the invention may be produced as a polyclonal population but are more preferably produced as a monoclonal population (that is as a substantially homogenous population of identical antibodies directed against a specific antigenic binding site). It will of course be apparent to those skilled in the art that a population implies more than one antibody entity. Antibodies of the present invention may be produced in transgenic organisms such as goats (see Pollock et al (1999),

J.Immunol.Methods 231:147-157), chickens (see Morrow KJJ (2000) Genet.Eng.News 20:1-55, mice (see Pollock et al) or plants (see Doran PM, (2000) Curr.Opinion Biotechnol. 11, 199-204, Ma JK-C (1998), Nat.Med. 4; 601-606, Baez J et al, BioPharm (2000) 13: 50-54, Stoger E et al; (2000) Plant Mol.Biol. 42:583-590).

Antibodies may also be produced by chemical synthesis. However, antibodies of the invention are typically produced using recombinant cell culturing technology well known to those skilled in the art. A polynucleotide encoding the antibody is isolated and inserted into a replicable vector such as a plasmid for further cloning (amplification) or expression. One useful expression system is a glutamate synthetase system (such as sold by Lonza Biologics), particularly where the host cell is CHO or NS0 (see below). Polynucleotide encoding the antibody is readily isolated and sequenced using conventional procedures (e.g. oligonucleotide probes). Vectors that may be used include plasmid, virus, phage, transposons, minichromsomes of which plasmids are a typical embodiment. Generally such vectors further include a signal sequence, origin of replication, one or more marker genes, an enhancer element, a promoter and transcription termination sequences operably linked to the light and/or heavy chain polynucleotide so as to facilitate expression. Polynucleotide encoding the light and heavy chains may be inserted into separate vectors and transfected into the same host cell or, if desired both the heavy chain and light chain can be inserted into the same vector for transfection into the host cell. Thus according to one aspect of the present invention there is provided a process of constructing a vector encoding the light and/or heavy chains of an antibody or antigen binding fragment thereof of the invention, which method comprises inserting into a vector, a polynucleotide encoding either a light chain and/or heavy chain of an antibody of the invention.

It is known to those skilled in the art that synthetic genes, which encode the same protein as a naturally occurring or wild type gene, may be designed by changing the codons that are used in the gene.

These design techniques involve replacing those codons in a gene that are rarely used in mammalian genes with codons that are more frequently used for that amino acid in mammalian gene. This process, called codon optimisation, is used with the intent that the total level of protein produced by the host cell is greater when transfected with the codon-optimised gene in comparison with the level when transfected with the wild-type sequence. Several methods have been published (Nakamura et. al., Nucleic Acids Research 1996,24 : 214-215; W098/34640; W097/11086).

Codon frequencies can be derived from literature sources for the highly expressed genes of many species (see e. g. Nakamura et al. Nucleic Acids Research 1996,24 : 214-215). Codon usage tables for humans (have also been published (WO2005025614).

It will be immediately apparent to those skilled in the art that due to the redundancy of the genetic code, alternative polynucleotides to those disclosed herein (particularly those codon optimised for expression in a given host cell) are also available that will encode the polypeptides of the invention.

3.1 Signal sequences

Antibodies of the present invention may be produced as a fusion protein with a heterologous signal sequence having a specific cleavage site at the N terminus of the mature protein. The signal sequence should be recognised and processed by the host cell. For prokaryotic host cells, the signal sequence may be for example an alkaline phosphatase, penicillinase, or heat stable enterotoxin II leaders. For yeast secretion the signal sequences may be for example a yeast invertase leader, α factor leader or acid phosphatase leaders see e.g. WO90/13646. In mammalian cell systems, viral secretory leaders such as herpes simplex gD signal and a native immunoglobulin signal sequence may be suitable. Typically the signal sequence is ligated in reading frame to DNA encoding the antibody of the invention.

3.2 Origin of replication

Origin of replications are well known in the art with pBR322 suitable for most gram-negative bacteria, 2μ plasmid for most yeast and various viral origins such as SV40, polyoma, adenovirus, VSV or BPV for most mammalian cells. Generally the origin of replication component is not needed for mammalian expression vectors but the SV40 may be used since it contains the early promoter.

3.3 Selection marker

Typical selection genes encode proteins that (a) confer resistance to antibiotics or other toxins e.g. ampicillin, neomycin, methotrexate or tetracycline or (b) complement auxiotrophic deficiencies or supply nutrients not available in the complex media. The selection scheme may involve arresting growth of the host cell. Cells, which have

been successfully transformed with the genes encoding the antibody of the present invention, survive due to e.g. drug resistance conferred by the selection marker. Another example is the so-called DHFR selection marker wherein transformants are cultured in the presence of methotrexate. In typical embodiments, cells are cultured in the presence of increasing amounts of methotrexate to amplify the copy number of the exogenous gene of interest. CHO cells are a particularly useful cell line for the DHFR selection. A further example is the glutamate synthetase expression system (Lonza Biologics). A suitable selection gene for use in yeast is the trp1 gene, see Stinchcomb et al Nature 282, 38, 1979.

3.4 Promoters

Suitable promoters for expressing antibodies of the invention are operably linked to DNA/polynucleotide encoding the antibody. Promoters for prokaryotic hosts include phoA promoter, Beta-lactamase and lactose promoter systems, alkaline phosphatase, tryptophan and hybrid promoters such as Tac. Promoters suitable for expression in yeast cells include 3-phosphoglycerate kinase or other glycolytic enzymes e.g. enolase, glyceralderhyde 3 phosphate dehydrogenase, hexokinase, pyruvate decarboxylase, phosphofructokinase, glucose 6 phosphate isomerase, 3-phosphoglycerate mutase and glucokinase. Inducible yeast promoters include alcohol dehydrogenase 2, isocytochrome C, acid phosphatase, metallothionein and enzymes responsible for nitrogen metabolism or maltose/galactose utilization. Promoters for expression in mammalian cell systems include viral promoters such as polyoma, fowlpox and adenoviruses (e.g. adenovirus 2), bovine papilloma virus, avian sarcoma virus, cytomegalovirus (in particular the immediate early gene promoter), retrovirus, hepatitis B virus, actin, rous sarcoma virus (RSV) promoter and the early or late Simian virus 40. Of course the choice of promoter is based upon suitable compatibility with the host cell used for expression. In one embodiment therefore there is provided a first plasmid comprising a RSV and/or SV40 and/or CMV promoter, DNA encoding light chain variable domain (V_L) of the invention, κC region together with neomycin and ampicillin resistance selection markers and a second plasmid comprising a RSV or SV40 promoter, DNA encoding the heavy chain variable domain (V_H) of the invention, DNA encoding the γ1 constant region, DHFR and ampicillin resistance markers

3.5 Enhancer element

Where appropriate, e.g. for expression in higher eukaroytics, an enhancer element operably linked to the promoter element in a vector may be used. Suitable mammalian enhancer sequences include enhancer elements from globin, elastase, albumin, fetoprotein and insulin. Alternatively, one may use an enhancer element from a eukaroytic cell virus such as SV40 enhancer (at bp100-270), cytomegalovirus early promoter enhancer, polyma enhancer, baculoviral enhancer or murine IgG2a locus (see WO04/009823). The enhancer may be located on the vector at a site upstream to the promoter.

3.5.5 - Polyadenylation signals

In eukaryotic systems, polyadenylation signals are operably linked to DNA/polynucleotide encoding the antibody of this invention. Such signals are typically placed 3' of the open reading frame. In mammalian systems, non-limiting example include signals derived from growth hormones, elongation factor-1 alpha and viral (eg SV40) genes or retroviral long terminal repeats. In yeast systems non-limiting examples of polydenylation/termination signals include those derived from the phosphoglycerate kinase (PGK) and the alcohol dehydrogenase 1 (ADH) genes. In prokaryotic system polyadenylation signals are typically not required and it is instead usual to employ shorter and more defined terminator sequences. Of course the choice of polyadenylation/termination sequences is based upon suitable compatibility with the host cell used for expression.

3.6 Host cells

Suitable host cells for cloning or expressing vectors encoding antibodies of the invention are prokaryotic, yeast or higher eukaryotic cells. Suitable prokaryotic cells include eubacteria e.g. enterobacteriaceae such as Escherichia e.g. E.Coli (for example ATCC 31,446; 31,537; 27,325), Enterobacter, Erwinia, Klebsiella Proteus, Salmonella e.g. Salmonella typhimurium, Serratia e.g. Serratia marcescans and Shigella as well as Bacilli such as B.subtilis and B.licheniformis (see DD 266 710), Pseudomonas such as P.aeruginosa and Streptomyces. Of the yeast host cells, Saccharomyces cerevisiae, schizosaccharomyces pombe, Kluyveromyces (e.g. ATCC 16,045; 12,424; 24178; 56,500), yarrowia (EP402, 226), Pichia Pastoris (EP183, 070, see also Peng et al J.Biotechnol. 108 (2004) 185-192), Candida,

Trichoderma reesia (EP244, 234), Penicillin, Tolypocladium and Aspergillus hosts such as A.nidulans and A.niger are also contemplated.

Although Prokaryotic and yeast host cells are specifically contemplated by the invention, host cells of the present invention are higher eukaryotic cells. Suitable higher eukaryotic host cells include mammalian cells such as COS-1 (ATCC NO:CRL 1650) COS-7 (ATCC CRL 1651), human embryonic kidney line 293, baby hamster kidney cells (BHK) (ATCC CRL.1632), BHK570 (ATCC NO: CRL 10314), 293 (ATCC NO:CRL 1573), Chinese hamster ovary cells CHO (e.g. CHO-K1, ATCC NO: CCL 61, DHFR-CHO cell line such as DG44 (see Urlaub et al, (1986) Somatic Cell Mol.Genet.12, 555-556)), particularly those CHO cell lines adapted for suspension culture, mouse sertoli cells, monkey kidney cells, African green monkey kidney cells (ATCC CRL-1587), HELA cells, canine kidney cells (ATCC CCL 34), human lung cells (ATCC CCL 75), Hep G2 and myeloma or lymphoma cells e.g. NS0 (see US 5,807,715), Sp2/0, Y0.

Thus in one embodiment of the invention there is provided a stably transformed host cell comprising a vector encoding a heavy chain and/or light chain of the antibody or antigen binding fragment thereof as herein described. Such host cells comprise a first vector encoding the light chain and a second vector encoding said heavy chain.

Bacterial fermentation

Bacterial systems are particularly suited for the expression of antigen binding fragments. Such fragments are localised intracellularly or within the periplasma. INS0luble periplasmic proteins can be extracted and refolded to form active proteins according to methods known to those skilled in the art, see Sanchez et al (1999) J.Biotechnol. 72, 13-20 and Cupit PM et al (1999) Lett Appl Microbiol, 29, 273-277.

3.7 Cell Culturing Methods.

Host cells transformed with vectors encoding the antibodies of the invention or antigen binding fragments thereof may be cultured by any method known to those skilled in the art. Host cells may be cultured in spinner flasks, roller bottles or hollow fibre systems but for large scale production that stirred tank reactors are used particularly for suspension cultures. Preferably the stirred tankers are adapted for aeration using e.g. spargers, baffles or low shear impellers. For bubble columns and

airlift reactors direct aeration with air or oxygen bubbles maybe used. Where the host cells are cultured in a serum free culture media, the media is supplemented with a cell protective agent such as pluronic F-68 to help prevent cell damage as a result of the aeration process. Depending on the host cell characteristics, either microcarriers maybe used as growth substrates for anchorage dependent cell lines or the cells maybe adapted to suspension culture (which is typical). The culturing of host cells, particularly invertebrate host cells may utilise a variety of operational modes such as fed-batch, repeated batch processing (see Drapeau et al (1994) cytotechnology 15: 103-109), extended batch process or perfusion culture. Although recombinantly transformed mammalian host cells may be cultured in serum-containing media such as fetal calf serum (FCS), for example such host cells are cultured in synthetic serum –free media such as disclosed in Keen et al (1995) Cytotechnology 17:153-163, or commercially available media such as ProCHO-CDM or UltraCHO™ (Cambrex NJ, USA), supplemented where necessary with an energy source such as glucose and synthetic growth factors such as recombinant insulin. The serum-free culturing of host cells may require that those cells are adapted to grow in serum free conditions. One adaptation approach is to culture such host cells in serum containing media and repeatedly exchange 80% of the culture medium for the serum-free media so that the host cells learn to adapt in serum free conditions (see e.g. Scharfenberg K et al (1995) in Animal Cell technology: Developments towards the 21st century (Beuvery E.C. et al eds), pp619-623, Kluwer Academic publishers).

Antibodies of the invention secreted into the media may be recovered and purified using a variety of techniques to provide a degree of purification suitable for the intended use. For example the use of antibodies of the invention for the treatment of human patients typically mandates at least 95% purity, more typically 98% or 99% or greater purity (compared to the crude culture medium). In the first instance, cell debris from the culture media is typically removed using centrifugation followed by a clarification step of the supernatant using e.g. microfiltration, ultrafiltration and/or depth filtration. A variety of other techniques such as dialysis and gel electrophoresis and chromatographic techniques such as hydroxyapatite (HA), affinity chromatography (optionally involving an affinity tagging system such as polyhistidine) and/or hydrophobic interaction chromatography (HIC, see US 5, 429,746) are available. In one embodiment, the antibodies of the invention, following various clarification steps, are captured using Protein A or G affinity chromatography followed by further chromatography steps such as ion exchange and/or HA chromatography, anion or cation exchange, size exclusion chromatography and ammonium sulphate

precipitation. Typically, various virus removal steps are also employed (e.g. nanofiltration using e.g. a DV-20 filter). Following these various steps, a purified (preferably monoclonal) preparation comprising at least 75mg/ml or greater e.g. 100mg/ml or greater of the antibody of the invention or antigen binding fragment thereof is provided and therefore forms an embodiment of the invention. Suitably such preparations are substantially free of aggregated forms of antibodies of the invention.

4. Pharmaceutical Compositions

Purified preparations of antibodies of the invention (particularly monoclonal preparations) as described supra, may be incorporated into pharmaceutical compositions for use in the treatment of human diseases and disorders such as Rheumatoid Arthritis, Psoriasis or Cancers e.g; Acute Lymphoblastic Leukemia, Adrenocortical Carcinoma, AIDS-Related Cancers, AIDS Related Lymphoma, Anal Cancer, Childhood Cerebellar Astrocytoma, Childhood Cerebral Astrocytoma, Colorectal Cancer, Basal Cell Carcinoma, Extrahepatic Bile Duct Cancer, Bladder Cancer, Osteosarcorna/Malignant Fibrous Histiocytoma Bone Cancer, Brain Tumors (e.g., Brain Stem Glioma, Cerebellar Astrocytoma, Cerebral Astrocytoma/Malignant Glioma, Ependymoma, Medulloblastoma, Supratentorial Primitive Neuroectodermal Tumors, Visual Pathway and Hypothalamic Glioma), Breast Cancer, Bronchial Adenomas/Carcinoids, Burkitt's Lymphoma, Carcinoid Tumor, Gastrointestinal Carcinoid Tumor, Carcinoma of Unknown Primary, Primary Central Nervous System, Cerebellar Astrocytoma, Cerebral Astrocytoma/Malignant Glioma, Cervical Cancer, Childhood Cancers, Chronic Lymphocytic Leukemia, Chronic Myelogenous Leukemia, Chronic Myeloproliferative Disorders, Colon Cancer, Colorectal Cancer, Cutaneous T-Cell Lymphoma, Endometrial Cancer, Ependymoma, Esophageal Cancer, Ewing's Family of Tumors, Extracranial Germ Cell Tumor, Extragonadal Germ Cell Tumor, Extrahepatic Bile Duct Cancer, Intraocular Melanoma Eye Cancer, Retinoblastoma Eye Cancer, Gallbladder Cancer, Gastric (Stomach) Cancer, Gastrointestinal Carcinoid Tumor, Germ Cell Tumors (e.g., Extracranial, Extragonadal, and Ovarian), Gestational Trophoblastic Tumor, Glioma (e.g., Adult, Childhood Brain Stem, Childhood Cerebral Astrocytoma, Childhood Visual Pathway and Hypothalamic), Hairy Cell Leukemia, Head and Neck Cancer, Hepatocellular (Liver) Cancer, Hodgkin's Lymphoma, Hypopharyngeal Cancer, Hypothalamic and Visual Pathway Glioma, Intraocular Melanoma, Islet Cell Carcinoma (Endocrine Pancreas), Kaposi's Sarcoma, Kidney (Renal Cell) Cancer, Laryngeal Cancer,

Leukemia (e.g., Acute Lymphoblastic, Acute Myeloid, Chronic Lymphocyhc, Chronic Myelogenous, and Hairy Cell), Lip and Oral Cavity Cancer, Liver Cancer, Non-Small Cell Lung Cancer, Small Cell Lung Cancer, Lymphoma (e.g., AIDS-Related, Burkitt's, Cutaneous T-cell, Hodgkin's, Non-Hodgkin's, and Primary Central Nervous System), Waldenstrom's Macroglobulinemia, Malignant Fibrous Histiocytoma of Bone/Osteosarcoma, Medulloblastoma, Melanoma, Intraocular (Eye) Melanoma, Merkel Cell Carcinoma, Mesothelioma, Metastatic Squamous Neck Cancer with Occult Primary, Multiple Endocrine Neoplasia Syndrome, Multiple Myeloma/Plasma Cell Neoplasm, Mycosis Fungoides, Myelodysplastic Syndromes, Myelodysplastic/Myeloproliferative Diseases, Myelogenous Leukemia, Chronic Myeloid Leukemia, Multiple Myeloma, Chronic Myeloproliferative Disorders, Nasal Cavity and Paranasal Sinus Cancer, Nasopharyngeal Cancer, Neuroblastoma, Oral Cancer, Oropharyngeal Cancer, Osteosarcoma/Malignant Fibrous Histiocytoma of Bone, Ovarian Cancer, Ovarian Epithelial Cancer, Ovarian Germ Cell Tumor, Ovarian Low Malignant Potential Tumor, Pancreatic Cancer, Islet Cell Pancreatic Cancer, Paranasal Sinus and Nasal Cavity Cancer, Parathyroid Cancer, Penile Cancer, Pheochromocytoma, Pineoblastoma, Pituitary Tumor, Plasma Cell Neoplasm/Multiple Myeloma, Pleuropulmonary Blastoma, Primary Central Nervous System Lymphoma, Prostate Cancer, Rectal Cancer, Renal Cell (Kidney) Cancer, Renal Pelvis and Ureter Transitional Cell Cancer, Retinoblastoma, Rhabdomyosarcoma, Salivary Gland Cancer, Soft Tissue Sarcoma, Uterine Sarcoma, Sezary Syndrome, non-Melanoma Skin Cancer, Merkel Cell Skin Carcinoma, Small Intestine Cancer, Soft Tissue Sarcoma, Squamous Cell Carcinoma, Cutaneous T-cell Lymphoma, Testicular Cancer, Thyrnoma, Thymic Carcinoma, Thyroid Cancer, Gestational Trophoblastic Tumor, Carcinoma of Unknown Primary Site, Cancer of Unknown Primary Site, Urethral Cancer, Endometrial Uterine Cancer, Uterine Sarcoma, Vaginal Cancer, Visual Pathway and Hypothalamic Glioma, Vulvar Cancer, Waldenstrom's Macroglobulinemia, and Wilms' Tumor.

Typically such compositions comprise a pharmaceutically acceptable carrier as known and called for by acceptable pharmaceutical practice, see e.g. Remingtons Pharmaceutical Sciences, 16th edition, (1980), Mack Publishing Co. Examples of such carriers include sterilised carrier such as saline, Ringers solution or dextrose solution, buffered with suitable buffers to a pH within a range of 5 to 8. Pharmaceutical compositions for injection (e.g. by intravenous, intraperitoneal, intradermal, subcutaneous, intramuscular or intraportal) or continuous infusion are

suitably free of visible particulate matter and may comprise between 0.1ng to 100mg of antibody, for example between 5mg and 25mg of antibody. Methods for the preparation of such pharmaceutical compositions are well known to those skilled in the art. In one embodiment, pharmaceutical compositions comprise between 0.1ng to 100mg of antibodies of the invention in unit dosage form, optionally together with instructions for use. Pharmaceutical compositions of the invention may be lyophilised (freeze dried) for reconstitution prior to administration according to methods well known or apparent to those skilled in the art. Where embodiments of the invention comprise antibodies of the invention with an IgG1 isotype, a chelator of copper such as citrate (e.g. sodium citrate) or EDTA or histidine may be added to pharmaceutical composition to reduce the degree of copper-mediated degradation of antibodies of this isotype, see EP0612251.

Effective doses and treatment regimes for administering the antibody or antigen binding fragment thereof of the invention are generally determined empirically and are dependent on factors such as the age, weight and health status of the patient and disease or disorder to be treated. Such factors are within the purview of the attending physician. Guidance in selecting appropriate doses may be found in e.g. Smith et al (1977) Antibodies in human diagnosis and therapy, Raven Press, New York but will in general be between 1mg and 1000mg.

Conveniently, a pharmaceutical composition comprising a kit of parts of the antibody of the invention or antigen binding fragment thereof together with other medicaments with instructions for use is also contemplated by the present invention.

The invention furthermore contemplates a pharmaceutical composition comprising a therapeutically effective amount of an antibody or antigen binding fragment thereof as herein described for use in the treatment of diseases responsive to neutralisation of the interaction between IGF-I and IGF-1R or IGF-II and IGF-IR.

In accordance with the present invention there is provided a pharmaceutical composition comprising a therapeutically effective amount of a monoclonal humanised antibody which antibody comprises a V_H domain selected from the group consisting of: SEQ ID NO:14 and a V_L domain selected from the group consisting of: SEQ ID NO:16

In accordance with the present invention there is provided a pharmaceutical

composition comprising a therapeutically effective amount of a monoclonal humanised antibody which antibody comprises a V_H domain selected from the group consisting of: SEQ ID NO:15 and a V_L domain selected from the group consisting of: SEQ ID NO:16

Conveniently, a pharmaceutical composition comprising a kit of parts of the antibody of the invention or antigen binding fragment thereof together with such another medicaments optionally together with instructions for use is also contemplated by the present invention.

The invention furthermore contemplates a pharmaceutical composition comprising a therapeutically effective amount of monoclonal antibody or antigen binding fragment thereof as herein described for use in the treatment of diseases responsive to neutralisation of the activity of IGF-1R.

In another embodiment of the invention a pharmaceutical composition comprising the antibody in combination with other therapeutic agents or radiation therapy, for example in combination with other classes of drug including mitotic inhibitors, alkylating agents, anti-metabolites, intercalating antibiotics, growth factor inhibitors, cell cycle inhibitors, enzymes, topoisomerase inhibitors, anti-survival agents, biological response modifiers, anti-hormones and anti-angiogenesis agents, including anti-growth factor receptor antagonists including trastuzumab (Herceptin), Erbitux (cetuximab), anti-growth factor antibodies such as bevacizumab (Avastin), antagonists of platelet-derived growth factor receptor (PDGFR), nerve growth factor (NGFR), fibroblast growth factor receptor (FGFR), , small molecular tyrosine kinase inhibitors for example lapatinib, gefitinib, etc, chemotherapeutic agents including gemcitabine, irinotecan, paclitaxel, cisplatin, doxorubicin, topotecan, cyclophosphamide, melphalan, dacarbazine, daunorubicin, aminocamptothecin, etoposide, teniposide, adriamycin, 5-Fluorouracil, cytosine arabinoside (Ara-C), Thiotepa, Taxotere, Buslfan, Cytoxin, Txaol, Methotrexate, Vinblastine, Bleomycin, Ifosfamide, Mitomycin C, Mitoxantrone, Vincreistine, Vinorelbine, Carboplatin, Carminomycin, Aminopterin, Dactinomycin, used in the treatment of human diseases and disorders such as Rheumatoid Arthritis, Psoriasis or Cancers such as: Acute Lymphoblastic Leukemia, Adrenocortical Carcinoma, AIDS-Related Cancers, AIDS Related Lymphoma, Anal Cancer, Childhood Cerebellar Astrocytoma, Childhood Cerebral Astrocytoma, Colorectal Cancer, Basal Cell Carcinoma, Extrahepatic Bile

Duct Cancer, Bladder Cancer, Osteosarcorna/Malignant Fibrous Histiocytoma Bone Cancer, Brain Tumors (e.g., Brain Stem Glioma, Cerebellar Astrocytoma, Cerebral Astrocytoma/Malignant Glioma, Ependymoma, Medulloblastoma, Supratentorial Primitive Neuroectodermal Tumors, Visual Pathway and Hypothalamic Glioma), Breast Cancer, Bronchial Adenomas/Carcinoids, Burkitt's Lymphoma, Carcinoid Tumor, Gastrointestinal Carcinoid Tumor, Carcinoma of Unknown Primary, Primary Central Nervous System, Cerebellar Astrocytoma, Cerebral Astrocytoma/Malignant Glioma, Cervical Cancer, Childhood Cancers, Chronic Lymphocytic Leukemia, Chronic Myelogenous Leukemia, Chronic Myeloproliferative Disorders, Colon Cancer, Colorectal Cancer, Cutaneous T-Cell Lymphoma, Endometrial Cancer, Ependymoma, Esophageal Cancer, Ewing's Family of Tumors, Extracranial Germ Cell Tumor, Extragonadal Germ Cell Tumor, Extrahepatic Bile Duct Cancer, Intraocular Melanoma Eye Cancer, Retinoblastoma Eye Cancer, Gallbladder Cancer, Gastric (Stomach) Cancer, Gastrointestinal Carcinoid Tumor, Germ Cell Tumors (e.g., Extracranial, Extragonadal, and Ovarian), Gestational Trophoblastic Tumor, Glioma (e.g., Adult, Childhood Brain Stem, Childhood Cerebral Astrocytoma, Childhood Visual Pathway and Hypothalamic), Hairy Cell Leukemia, Head and Neck Cancer, Hepatocellular (Liver) Cancer, Hodgkin's Lymphoma, Hypopharyngeal Cancer, Hypothalamic and Visual Pathway Glioma, Intraocular Melanoma, Islet Cell Carcinoma (Endocrine Pancreas), Kaposi's Sarcoma, Kidney (Renal Cell) Cancer, Laryngeal Cancer, Leukemia (e.g., Acute Lymphoblastic, Acute Myeloid, Chronic Lymphocyhc, Chronic Myelogenous, and Hairy Cell), Lip and Oral Cavity Cancer, Liver Cancer, Non-Small Cell Lung Cancer, Small Cell Lung Cancer, Lymphoma (e.g., AIDS-Related, Burkitt's, Cutaneous T-cell, Hodgkin's, Non-Hodgkin's, and Primary Central Nervous System), Waldenstrom's Macroglobulinemia, Malignant Fibrous Histiocytoma of Bone/Osteosarcoma, Medulloblastoma, Melanoma, Intraocular (Eye) Melanoma, Merkel Cell Carcinoma, Mesothelioma, Metastatic Squamous Neck Cancer with Occult Primary, Multiple Endocrine Neoplasia Syndrome, Multiple Myeloma/Plasma Cell Neoplasm, Mycosis Fungoides, Myelodysplastic Syndromes, Myelodysplastic/Myeloproliferative Diseases, Myelogenous Leukemia, Chronic Myeloid Leukemia, Multiple Myeloma, Chronic Myeloproliferative Disorders, Nasal Cavity and Paranasal Sinus Cancer, Nasopharyngeal Cancer, Neuroblastoma, Oral Cancer, Oropharyngeal Cancer, Osteosarcoma/Malignant Fibrous Histiocytoma of Bone, Ovarian Cancer, Ovarian Epithelial Cancer, Ovarian Germ Cell Tumor, Ovarian Low Malignant Potential Tumor, Pancreatic Cancer, Islet Cell Pancreatic Cancer, Paranasal Sinus and Nasal Cavity Cancer, Parathyroid Cancer, Penile Cancer, Pheochromocytoma,

Pineoblastoma, Pituitary Tumor, Plasma Cell Neoplasm/Multiple Myeloma, Pleuropulmonary Blastoma, Primary Central Nervous System Lymphoma, Prostate Cancer, Rectal Cancer, Renal Cell (Kidney) Cancer, Renal Pelvis and Ureter Transitional Cell Cancer, Retinoblastoma, Rhabdomyosarcoma, Salivary Gland Cancer, Soft Tissue Sarcoma, Uterine Sarcoma, Sezary Syndrome, non-Melanoma Skin Cancer, Merkel Cell Skin Carcinoma, Small Intestine Cancer, Soft Tissue Sarcoma, Squamous Cell Carcinoma, Cutaneous T-cell Lymphoma, Testicular Cancer, Thyrnoma, Thymic Carcinoma, Thyroid Cancer, Gestational Trophoblastic Tumor, Carcinoma of Unknown Primary Site, Cancer of Unknown Primary Site, Urethral Cancer, Endometrial Uterine Cancer, Uterine Sarcoma, Vaginal Cancer, Visual Pathway and Hypothalamic Glioma, Vulvar Cancer, Waldenstrom's Macroglobulinemia, and Wilms' Tumor.

The antibody or antigen binding fragments thereof of the present invention may be used in combination with one or more other therapeutically active agents or radiation for example in combination with other classes of drug including mitotic inhibitors, alkylating agents, anti-metabolites, intercalating antibiotics, growth factor inhibitors, cell cycle inhibitors, enzymes, topoisomerase inhibitors, anti-survival agents, biological response modifiers, anti-hormones and anti-angiogenesis agents, including anti-growth factor receptor antagonists including trastuzumab (Herceptin), Erbitux (cetuximab), anti-growth factor antibodies such as bevacizumab (Avastin), antagonists of platelet-derived growth factor receptor (PDGFR), nerve growth factor (NGFR), fibroblast growth factor receptor (FGFR), small molecule anti-IGF-1R agents, small molecular tyrosine kinase inhibitors including lapatinib, gefitinib, etc, chemotherapeutic agents including gemcitabine, irinotecan, paclitaxel, cisplatin, doxorubicin, topotecan, cyclophosphamide, melphalan, dacarbazine, daunorubicin, aminocamptothecin, etoposide, teniposide, adriamycin, 5-Fluorouracil, cytosine arabinoside (Ara-C), Thiotepa, Taxotere, Buslfan, Cytoxin, Txaol, Methotrexate, Vinblastine, Bleomycin, Ifosfamide, Mitomycin C, Mitoxantrone, Vincreistine, Vinorelbine, Carboplatin, Carminomycin, Aminopterin, Dactinomycin

The invention thus provides, in a further embodiment, the use of such a combination in the treatment of diseases where IGF-1 receptor signalling contributes to the disease or where neutralising the activity of the receptor will be beneficial and the use of the antibody or antigen binding fragment thereof in the manufacture of a medicament for the combination therapy of disorders such as Rheumatoid Arthritis, Psoriasis or Cancers such as: Acute Lymphoblastic Leukemia, Adrenocortical

Carcinoma, AIDS-Related Cancers, AIDS Related Lymphoma, Anal Cancer, Childhood Cerebellar Astrocytoma, Childhood Cerebral Astrocytoma, Colorectal Cancer, Basal Cell Carcinoma, Extrahepatic Bile Duct Cancer, Bladder Cancer, Osteosarcorna/Malignant Fibrous Histiocytoma Bone Cancer, Brain Tumors (e.g., Brain Stem Glioma, Cerebellar Astrocytoma, Cerebral Astrocytoma/Malignant Glioma, Ependymoma, Medulloblastoma, Supratentorial Primitive Neuroectodermal Tumors, Visual Pathway and Hypothalamic Glioma), Breast Cancer, Bronchial Adenomas/Carcinoids, Burkitt's Lymphoma, Carcinoid Tumor, Gastrointestinal Carcinoid Tumor, Carcinoma of Unknown Primary, Primary Central Nervous System, Cerebellar Astrocytoma, Cerebral Astrocytoma/Malignant Glioma, Cervical Cancer, Childhood Cancers, Chronic Lymphocytic Leukemia, Chronic Myelogenous Leukemia, Chronic Myeloproliferative Disorders, Colon Cancer, Colorectal Cancer, Cutaneous T-Cell Lymphoma, Endometrial Cancer, Ependymoma, Esophageal Cancer, Ewing's Family of Tumors, Extracranial Germ Cell Tumor, Extragonadal Germ Cell Tumor, Extrahepatic Bile Duct Cancer, Intraocular Melanoma Eye Cancer, Retinoblastoma Eye Cancer, Gallbladder Cancer, Gastric (Stomach) Cancer, Gastrointestinal Carcinoid Tumor, Germ Cell Tumors (e.g., Extracranial, Extragonadal, and Ovarian), Gestational Trophoblastic Tumor, Glioma (e.g., Adult, Childhood Brain Stem, Childhood Cerebral Astrocytoma, Childhood Visual Pathway and Hypothalamic), Hairy Cell Leukemia, Head and Neck Cancer, Hepatocellular (Liver) Cancer, Hodgkin's Lymphoma, Hypopharyngeal Cancer, Hypothalamic and Visual Pathway Glioma, Intraocular Melanoma, Islet Cell Carcinoma (Endocrine Pancreas), Kaposi's Sarcoma, Kidney (Renal Cell) Cancer, Laryngeal Cancer, Leukemia (e.g., Acute Lymphoblastic, Acute Myeloid, Chronic Lymphocyhc, Chronic Myelogenous, and Hairy Cell), Lip and Oral Cavity Cancer, Liver Cancer, Non-Small Cell Lung Cancer, Small Cell Lung Cancer, Lymphoma (e.g., AIDS-Related, Burkitt's, Cutaneous T-cell, Hodgkin's, Non-Hodgkin's, and Primary Central Nervous System), Waldenstrom's Macroglobulinemia, Malignant Fibrous Histiocytoma of Bone/Osteosarcoma, Medulloblastoma, Melanoma, Intraocular (Eye) Melanoma, Merkel Cell Carcinoma, Mesothelioma, Metastatic Squamous Neck Cancer with Occult Primary, Multiple Endocrine Neoplasia Syndrome, Multiple Myeloma/Plasma Cell Neoplasm, Mycosis Fungoides, Myelodysplastic Syndromes, Myelodysplastic/Myeloproliferative Diseases, Myelogenous Leukemia, Chronic Myeloid Leukemia, Multiple Myeloma, Chronic Myeloproliferative Disorders, Nasal Cavity and Paranasal Sinus Cancer, Nasopharyngeal Cancer, Neuroblastoma, Oral Cancer, Oropharyngeal Cancer, Osteosarcoma/Malignant Fibrous Histiocytoma of Bone, Ovarian Cancer, Ovarian Epithelial Cancer, Ovarian Germ Cell Tumor,

Ovarian Low Malignant Potential Tumor, Pancreatic Cancer, Islet Cell Pancreatic Cancer, Paranasal Sinus and Nasal Cavity Cancer, Parathyroid Cancer, Penile Cancer, Pheochromocytoma, Pineoblastoma, Pituitary Tumor, Plasma Cell Neoplasm/Multiple Myeloma, Pleuropulmonary Blastoma, Primary Central Nervous System Lymphoma, Prostate Cancer, Rectal Cancer, Renal Cell (Kidney) Cancer, Renal Pelvis and Ureter Transitional Cell Cancer, Retinoblastoma, Rhabdomyosarcoma, Salivary Gland Cancer, Soft Tissue Sarcoma, Uterine Sarcoma, Sezary Syndrome, non-Melanoma Skin Cancer, Merkel Cell Skin Carcinoma, Small Intestine Cancer, Soft Tissue Sarcoma, Squamous Cell Carcinoma, Cutaneous T-cell Lymphoma, Testicular Cancer, Thyrnoma, Thymic Carcinoma, Thyroid Cancer, Gestational Trophoblastic Tumor, Carcinoma of Unknown Primary Site, Cancer of Unknown Primary Site, Urethral Cancer, Endometrial Uterine Cancer, Uterine Sarcoma, Vaginal Cancer, Visual Pathway and Hypothalamic Glioma, Vulvar Cancer, Waldenstrom's Macroglobulinemia, and Wilms' Tumor.

When the antibody or antigen binding fragments thereof of the present invention are used in combination with other therapeutically active agents, the components may be administered either together or separately, sequentially or simultaneously by any convenient route.

The combinations referred to above may conveniently be presented for use in the form of a pharmaceutical formulation and thus pharmaceutical formulations comprising a combination as defined above optimally together with a pharmaceutically acceptable carrier or excipient comprise a further embodiment of the invention. The individual components of such combinations may be administered either together or separately, sequentially or simultaneously in separate or combined pharmaceutical formulations.

When combined in the same formulation it will be appreciated that the two components must be stable and compatible with each other and the other components of the formulation and may be formulated for administration. When formulated separately they may be provided in any convenient formulation, conveniently in such a manner as are known for antibodies and antigen binding fragments thereof in the art.

When in combination with a second therapeutic agent active against the same

disease, the dose of each component may differ from that when the antibody or antigen binding fragment thereof is used alone. Appropriate doses will be readily appreciated by those skilled in the art.

The invention thus provides, in a further embodiment, a combination comprising an antibody or antigen binding fragment thereof of the present invention together with another therapeutically active agent.

The combination referred to above may conveniently be presented for use in the form of a pharmaceutical formulation and thus pharmaceutical formulations comprising a combination as defined above together with a pharmaceutically acceptable carrier thereof represent a further embodiment of the invention.

The following examples illustrate various aspects of the invention. These examples do not limit the scope of this invention which is defined by the appended claims.

Examples

Example 1 – Generation of monoclonal antibodies

Monoclonal antibodies (mAbs) were produced by hybridoma cells generally in accordance with the method set forth in E Harlow and D Lane, Antibodies a Laboratory Manual, Cold Spring Harbor Laboratory, 1988. SJL mice were primed and boosted by intraperitoneal injection with recombinant human IGF-1R (R&D Systems, #305-GR) in RIBI adjuvant. Spleens from responder animals were harvested and fused to X63Ag8653GFP1L5 myeloma cells to generate hybridomas. The hybridoma supernatant material was screened for binding to IGF-1R using the FMAT (ABI8200) and BIAcore A100. The ABI8200 was used to confirm binding to recombinant IGF-1R (R&D Systems - 305-GR-050 and 391-GR-050) and HEK293T-expressed human IGF-1R, HEK293T expressed cynomolgus macaque IGF-1R and absence of binding to HEK293T-expressed human insulin receptor. The BIAcore A100 was used to estimate the kinetics of binding of hybridoma produced antibodies to recombinant IGF-1R (R&D Systems, #305-GR). Antibodies were captured onto the chip using a rabbit anti-mouse IgG (BR-1005-14, Biacore AB). Hybridomas of interest were monocloned using semi-solid media (methyl cellulose solution), Omnitrays and the ClonePix FL system.

Example 2 - Scale-up and purification of hybridoma material and monoclonal antibodies.

Hybridomas to be scaled up were grown in tissue culture to the scale of 4 confluent 225cm^2 flasks. At this point the cells were harvested by centrifugation at 400g for 5 minutes. The pellet was resuspended with 100ml serum free media (JRH610) to wash the cells. The cells were then centrifuged at 400g for 5 minutes. The supernatant was aspirated and discarded. 150ml of fresh serum free media was used to resuspend the cell pellet. The cell suspension was then transferred into a fresh 225cm2 flask and placed in an incubator for a period of 5 days. The supernatant was then harvested and centrifuged at 400g for 20 minutes. The supernatant was harvested and sterile filtered with a 0.2µM filter in preparation for purification. The antibody was purified using protein A resin columns. The purified antibody was dialysed against PBS pH7.4.

Example 3– Construction of IGF-1R expression vectors

Generation of expression cassette for full length human IGF-1R

The human IGF-1R cDNA expression cassette was identical to Genbank X04434 except for a change at nucleotide 3510. This results in the silent change of the codon for glycine 1170 from "GGC" to "GGG". Human IGF-1R cDNA was expressed from the pcDNA3.1(-) vector (Invitrogen). The sequence of human IGF-1R is set out in SEQ ID NO 44.

Generation of expression cassette for full length murine IGF-1R

The murine IGF-1R cDNA expression cassette was identical to Genbank AF056187 except for a change at nucleotide 3522. This results in the silent change of the codon for glycine 1174 from "GGT" to "GGG". The murine IGF-1R cDNAs was expressed from pcDNA3.1D-V5-His TOPO vectors (Invitrogen). The sequence of murine IGF-1R is set out in SEQ ID NO 46.

Generation of expression cassette for full length Cynomolgus macaque monkey (Macaca fascicularis) IGF-1R

The novel sequence for cynomolgus macaque monkey IGF-1R was cloned by PCR from a cynomolgus macaque monkey kidney cDNA library. Primers were based on the human IGF-1R database entry, NM_000875. PCR primers were designed with a Kozak motif at the 5' end and with flanking BamHI and NotI restriction sites. The BamHI-NotI PCR product was cloned into pCDNA3.1D with the vector T7 sequences proximal to the 5' end of the IGF-1R coding sequence. The cDNA obtained is 99.6% identical to the human sequence at the protein level (4aa differences from human). The sequence of cynomolgus macaque IGF-1R is set out in SEQ ID NO 45.

Generation of expression cassette for full length Human Insulin receptor (Type B)

A DNA cassette encoding human insulin receptor type B (SEQ ID NO 53) was cloned into pcDNA3.1 (Invitrogen). For comparison, the coding sequence of SEQ ID NO 53 is identical to the sequence given in Genbank entry:M10051, with the exception of the following changes:
The nucleotide numbering is based on the "A" of the initiation methionine being

nucleotide 1 (which corresponds to position 139 of the nucleotide sequence in M10051).

Nucleotide	Amino Acid	SEQ ID No 53	M10051
511	171	TAC (Tyr)	CAC (His)
783	261	GAT (Asp)	GAC (Asp)
909	303	CAG (Gln)	CAA (Gln)
1343	448	ATC (Ile)	ACC (Thr)
1474	492	CAG (Gln)	AAG (Lys)
1638	546	GAC (Asp)	GAT (Asp)
1650	550	GCA (Ala)	GCG (Ala)
3834	1278	AAC (Asn)	AAG (Lys)

Vectors for human, murine and cynomolgus macaque monkey IGF-1Rs and human insulin receptor type B were expressed transiently in 293 HEK-T cells using standard protocols and Lipofectamine reagent (Invitrogen).

Example 4 - Generation of and expression of recombinant proteins using BacMam

Construction of pFastBacMam vector backbone

pFastBac 1 (Invitrogen) was digested with SnaBI and Hpa1 to remove the polyhedrin promoter. This was ligated with a 3.1kb NruI-Bst1107I fragment from pcDNA3 (Invitrogen) which contains the cytomegalovirus immediate early (CMV IE) promoter with a polylinker and BGH poly A site and the SV40 promoter driving expression of the G418 resistance gene. This vector will allow production of a baculovirus which expresses a gene under the control of the CMV promoter in mammalian cells. It is also possible to select for stable derivatives by putting cells under G418 selection.

Human IGF-1R –Fc fusion protein

A plasmid designed to express human IGF-1R extracellular domain sequences fused to a factor Xa cleavage site and human Fc sequences from IgG1 was constructed. Sequences encoding the extracellular domain (amino acids 1-935) of the human IGF-1R cDNA were amplified by PCR and fused to a Factor Xa cleavage site and Fc sequences from human IgG1. The entire insert was then sub-cloned as a HindIII-

BamHI fragment into the pFastBacMam expression vector. The sequence of human IGF-1R-Fc fusion protein is set out in SEQ ID NO 47.

Cynomolgus macaque monkey (Macaca fascicularis) IGF-1R –Fc fusion protein

A plasmid designed to express cynomolgus macaque monkey IGF-1R extracellular domain sequences fused to a factor Xa cleavage site and human Fc sequences from IgG1 was constructed. The human IGF-1R expression plasmid was modified by the removal of a 82bp XbaI fragment of vector backbone by cutting with XbaI and re-ligating. This removes a second NotI site. The coding sequence for the extracellular domain of cynomolgus macaque monkey IGF-1R (amino acids 1-935) was amplified by PCR as a HindIII-NotI fragment and ligated into the modified human IGF-1R expression plasmid which had been cut with HindIII and Not I to remove the human sequences. The sequence of cynomolgus macaque IGF-1R-Fc fusion protein is set out in SEQ ID NO 48.

Expression of recombinant proteins using BacMam

Plasmid vectors encoding human and cynomolgus macaque monkey IGF-1R extracellular domain sequences fused to a Factor Xa cleavage site and Fc sequences from human IgG1 were used to direct protein expression using the BacMam system. Baculoviruses were generated using the Invitrogen Bac-to-Bac system. The initial P0 stock was scaled to a one litre P1 stock using standard procedures. Protein production was initiated by the infection of 1-5 litres of HEK293-F cells in suspension culture with the required BacMam virus (typically at a multiplicity of infection (MOI)) of 10 to 100 to 1 although this was usually optimized to maximize protein production). After 2-3 days culture the cell culture supernatant was harvested, cells were removed by centrifugation and the expressed protein was then purified from the cleared supernatant.

Example 5 - Construction of IGF-1R ligand expression plasmids

Gene sequences for the processed forms of IGF-I (amino acids 49-118, Swiss-prot P01343) and IGF-II (amino acids 25-91, Swiss-prot P01344) were codon optimised for *E.coli* expression. The genes were prepared de novo by build up of overlapping oligonucleotides and cloned into the NdeI-BamHI sites of pET-21b (Novagen). For

the production of biotinylated IGF-1R ligands, a C-terminal 15 amino acid biotinylation tag sequence (GLNDIFEAQKIEWHE, ref: Schatz (1993) Biotechnology (N Y), 11(10):1138-43) SEQ ID NO:17 was included in the gene build up.

The sequences of human IGF-I ligand and IGF-II ligand are set out in SEQ ID NO 49. and SEQ ID NO 51 respectively.

Example 6 - Expression and purification of IGF-1R ligands

Plasmids were transformed in E.coli BL21(DE3) cells then expression carried out using LB medium with 100µg/ml ampicillin following induction with 1mM IPTG at 37°C for 16 hours, The cell pellets were harvested by centrifugation. IGF-1R ligands were isolated as insoluble inclusion bodies by resuspending cell pellets in 50mM Tris pH8.0, 200mM NaCl, 1mM EDTA, 5mM DTT, lysed by sonication and recovered in the inclusion body fraction by centrifugation. Soluble IGF-1R ligands were produced by solubilising the inclusion bodies in 50mM Tris pH8.0, 6M Guanidine Hydrochloride, then rapidly diluting into a 100 fold excess volume of 50mM Tris pH8.0, 1mM oxidised glutathione, 1mM reduced glutathione followed by mixing for 16 hours at 4°C. Soluble protein was concentrated and centrifuged to remove insoluble material then biologically active IGF-1R ligands purified by reverse-phase HPLC using a Spherisorb C6 column (Waters) with an acetonitrile gradient.

For IGF-1R ligands with biotinylation tags, biotinylation was carried out by adding 5mM ATP, 5mM $MgCl_2$, 1mM d-biotin and 1µM biotin ligase to the purified proteins. The mixture was incubated at room temperature for 3 hours. The biotinylated IGF-1R ligands were purified by size exclusion chromatography using a Superdex 75 column (GE Healthcare). Purified IGF-1R ligands were dialysed against PBS, quantified using BSA standards and a BioRad coomassie based protein assay then stored in aliquots at -80C. Molecular weights of purified proteins were verified by mass spectroscopy. The sequences of human tagged IGF-I ligand and tagged IGF-II ligand are set out in SEQ ID NO 50. and SEQ ID NO 52 respectively.

Example 7. - Sequencing of Variable domains of Hybridomas

Total RNA was extracted from pellets of approximately 10^6 cells for each hybridoma clone using the RNeasy kit from Qiagen (#74106). Promega AccessQuick RT-PCR System (A1702) was used to produce cDNA of the variable heavy and light regions

using degenerate primers specific for the murine leader sequences and murine IgG1/$_K$ or IgG2b/$_K$ constant regions. The purified RT-PCR fragments were cloned using TA cloning kit (Invitrogen (K2030-40)) and a consensus sequence was obtained for each hybridoma by sequence alignment, database searching and alignment with known immunoglobulin variable sequences listed in KABAT (Sequences of Proteins of Immunological Interest, 4th Ed., U.S. Department of Health and Human Services, National Institutes of Health (1987).
The sequences listing numbers of the variable domains of hybridomas 6E11, 9C7, 2B9, 15D9 and 5G4 and shown in the Table 1 below:

Hybridoma	SEQ ID. NO: of variable heavy region	SEQ ID. NO: of variable light region
6E11	8	9
9C7	18	19
2B9	10	11
5G4	20	21
15D9	22	23

Table 1 - SEQ I.D. NO: of the variable heavy and light regions of the hybridomas. Note sequences shown in table 1 do not include signal sequences.

Example 8: Construction of chimaeric antibodies

Chimaeric antibodies, comprising parent murine variable domains grafted onto human IgG1/κ wild type constant regions were constructed by PCR cloning. Based on the consensus sequence, primers to amplify the murine variable domains were designed, incorporating restriction sites required to facilitate cloning into mammalian expression vectors. The full length heavy and light chains of the 6E11 chimeric antibody (6E11c) are given in SEQ I.D. NO: 24 and SEQ I.D. NO: 25.

Example 9: Humanisation strategy

Humanised antibodies were generated by a process of grafting CDRH1, CDRH2, CDRH3, CDRL1 and CDRL3 from the murine 6E11 antibody and CDRL2 from the murine 9C7 antibody onto a suitable human framework sequence.

The sequence of the humanised variable light domain of L0 is given in (SEQ I.D. NO: 16)

The sequences of the humanised variable heavy domains of H0 and H1 are given in (SEQ I.D. NO: 14 and SEQ I.D. NO: 15 respectively). Optimised nucleotide sequences encoding these sequences are shown in SEQ ID NO:34 (H0), 35 (H1) and 36 (L0). Alternative L0 and and H0 variable domain sequences are given in SEQ ID NO: 61 and 62 respectively, alternative L0 light chain and H0 heavy chain sequence are given in SEQ ID NO: 69 and 70.

Construction of humanised antibody vectors

DNA fragments encoding the humanised variable heavy and variable light regions were constructed de novo using a PCR-based strategy and overlapping oligonucleotides. The PCR product was cloned into mammalian expression vectors containing the human gamma 1 constant region and the human kappa constant region respectively. This is the wild type Fc region.

Using a similar strategy, the variable heavy regions were also cloned onto a variant of the human gamma 1 constant region which contained two alanine substitutions L235A and G237A (EU index numbering). These constructs are referred to herein as IgG1m(AA). The two humanised constructs which comprised the IgG1m(AA) variant are set out as H0L0 IgG1m(AA) (SEQ ID NO 54 and SEQ ID NO 39) and H1L0 IgG1m(AA) (SEQ ID NO 56 and SEQ ID NO 39).

Unless otherwise stated all humanised constructs used in the examples herein comprise wild type human gamma 1 constant regions.

Example 10 - Recombinant antibody expression in CHO cells

Expression plasmids encoding the heavy and light chains respectively of chimeric or humanised antibodies were transiently co-transfected into CHO-K1 cells. In some instances the supernatant material was used as the test article in binding and activity assays. In other instances, the supernatant material was filter sterilised and the antibody recovered by affinity chromatography using a Protein A. Antibodies were also expressed in a stable polyclonal CHO cell system. DNA vectors encoding the

heavy and light chains were co-electroporated into suspension CHO cells. Cells were passaged in shake flasks in MR1 basal selective medium at 37°C, 5%CO_2, 130-150rpm until cell viability and cell counts improved. CHO cells were then inoculated into MR1 basal x2 selective medium and incubated for 10 to 14 days at 34°C, 5%CO_2, 130-150rpm. The cells were pelleted by centrifugation and the supernatant sterile filtered. Antibody was recovered by Protein A purification.

Comparative data between hybridomas and/or chimaeric mAbs and/or humanised Mabs

Example 11 - Receptor Binding ELISA

0.4µg/mL Histidine tagged recombinant human IGF-1R (R&D Systems, #305-GR-050) was captured onto an ELISA plate coated with 0.5-1µg/mL of rabbit polyclonal antibody to 6xHis (Abcam, #ab9108). Anti-IGF-1R antibodies from the test supernatants or purified material were titrated across the plate. The levels of receptor-bound was detected by treatment with a horse-radish peroxidase (HRP) - conjugated goat-anti-mouse IgG antibody (Dako, P0260) or goat anti-human Kappa Light Chains peroxidase conjugate (Sigma, A7164). The ELISA was developed using O-phenylenediamine dihydrochloride (OPD) peroxidase substrate (Sigma, P9187).

Figure 1. shows the binding curves for murine antibodies 6E11, 5G4 and 15D9. Figure 2. shows the binding curves for H0L0 and H1L0 and H0L0 IgG1m(AA) and H1L0 IgG1m(AA) confirming they have similar binding activity when compared to the 6E11 chimaera. Other monoclonal antibodies were tested (data not shown).

Example 12 - Receptor Down Regulation

3T3/LISN c4 cells (murine NIH 3T3 cell line expressing human IGF-1R, see Kaleko et al. (1990) Molecular and Cellular Biology, 10 (2): 464-473) were incubated with 5µg/mL antibody at 37°C for 24 hours before the cells were harvested. Lysates of these cell pellets were run on an SDS PAGE gel and transferred to PVDF membrane (Western blot). IGF-1R was detected by treatment with a rabbit anti IGF-1R_β C-20 antibody (Santa Cruz Biotechnology, sc-713) followed by treatment with anti rabbit HRP-conjugated secondary antibody (P0217) and detected using enhanced chemiluminesence (ECL) reagent (GE Healthcare).

Figure 3 shows that incubation of 3T3/LISN c4 cells with monoclonal antibody 6E11 results in down-regulation of the IGF-1R$_\beta$ chain.

In a similar experiment, levels of receptor were assayed in NCI-H838 cells following treatment with H0L0. NCI-H838 cells ($1x10^6$/well) expressing human IGF-1R were incubated with 5µg/well H0L0 for various times up to 24 hours. Cells were then harvested and the cell pellets were lysed and run on an SDS PAGE gel, transferred to PVDF membrane and blotted for IGF-1R using a rabbit anti IGF-1Rβ c-20 antibody (Santa Cruz, sc713). Binding was detected with an anti-rabbit HRP antibody (Dako, PO217).The lanes on the Western blot from left to right are: No antibody control (harvested at 24 hours), then harvests at 0, 0.2, 0.5, 1, 1.5, 3, 6 and 24 hours. Figure 4 shows that H0L0 induces degradation as early as 30 minutes (0.5h) after exposure, although 3 hours continued exposure is required for IGF-1R levels to reach a basal level. In another experiment, incubation of Colo205 cells with various antibodies including 6E11 parental, 6E11 chimera, H0L0 for 24 hours caused a substantial reduction in receptor levels as determined by Western blot for the IGF-1R β-chain (data not shown)

In a follow-up experiment, NCI-H838 lung carcinoma cells were treated with the humanised therapeutic anti-IGF-1R antibodies. NCI-H838 cells were serum-starved for 24 hours and either untreated (control), or treated for 24 hours with 120nM H0L0, H0L0 IgG1m(AA) or non-targeting human IgG (control). Cells were washed twice with ice-cold PBS and lysed on ice for 10 minutes with 1% NP40 lysis buffer, clarified by centrifugation at 16400rpm for 20 minutes at 4°C and 50µg of soluble cellular proteins subjected to reducing SDS PAGE on 4 to12% polyacrylamide gels. After electrophoresis, proteins were transferred to PVDF membranes and immunoblotted for either IGF-1R or Insulin receptor. Membranes were also immunoblotted for α-tubulin to assess loading across each lane. Fluorescently-tagged secondary antibodies were used and the amounts of IGF-1R, Insulin receptor and α-tubulin quantified using a LI COR Odyssey imaging system. The primary and secondary antibodies used were selected from Alexa680, goat anti-rabbit, Molecular Probes #A-21076. Used 1/20000 = 0.5ml in 10ml (45min at 25°C), IRDye800, donkey anti-mouse, Rockland #610-732-124. Used 1/10000 = 1ml in 10ml (45min at 25°C) and IRDye800, donkey anti-rabbit, Rockland #611-732-127. Used 1/10000 = 0.5ml in 5ml (45min at 25°C).

The 97kDa β subunit of IGF-1R and the 200kDa full- length IGF-1R and the 95kDa β subunit of Insulin receptor and the 200kDa full length Insulin receptor and 50kDa α tubulin are shown in figure 5. Incubation with the humanised antibodies resulted in significantly decreased levels of IGF-1R – approximately 80% reduction relative to control antibody treated samples (Figure 5). This was accompanied by a decrease in Insulin receptor levels, most likely due to degradation of IGF-1R/Insulin receptor heterodimers.

Receptor down-regulation on LISN/3T3 c4 cells was also demonstrated using a FACS-based assay on whole blood (depleted for red blood cells). H0L0 was added for 24 hours at 4°C or 37°C to a whole blood sample from one donor (Donor 90263). Figure 6 shows an overlay histogram of fluorescent intensity for the granulocyte and lymphocyte population (as gated using forward and side-scatter profiles) at 4°C (solid line) and 37°C (dotted line). Following incubation, receptor levels were assessed using the PE labelled anti-IGF-1R antibody 1H7 (BD Pharmingen, #555999). In a different experiment H0L0 or Control IgG were added for 24 hours at 4°C or 37°C to a whole blood sample from a different donor (Donor 90691). Figure 7 shows an overlay histogram of fluorescent intensity or the granulocyte population at 4°C and 37°C compared to an isotype control. In both donors, incubation at 37°C for 24 hours caused a marked reduction in the mean fluorescence intensity compared to incubation of the same cell line at 4°C. The assay was repeated with whole blood samples from several other donors with a similar overall effect, although the magnitude of receptor expression and down-regulation varied from donor-to-donor. A small number of donors show very low receptor expression which was unaffected by incubation with antibody.

Example 13 - Inhibition of IGF-1 or IGF-II stimulated receptor phosphorylation

3T3/LISN c4 cells were plated at a density of 10 000 cells/well into 96 well plates and allowed to grow for 1-2 days in complete DMEM (DMEM-Hepes modification +10%FCS). Anti hIGF-1R antibodies (hybridoma supernatants or purified antibodies) were added to the cells and incubated for 1 hour. Either 30-50ng rhIGF-1(R&D Systems 291-G1 or 50ng/ml rhIGF-I (see Example 5 and 6) or 100ng/ml rhIGF-2 (R&D Systems 292-G2) (see Example 5 and 6) was added to the treated cells and incubated for a further 20-30 mins to stimulate receptor phosphorylation. Cells were washed once in PBS and then lysed by the addition of RIPA lysis buffer (150mM

NaCl, 50mM TrisHCl, 6mM Na Deoxycholate, 1% Tween 20) plus protease inhibitor cocktail (Roche 11 697 498 001). The plate was frozen for 30 minutes or overnight. After thawing, lysate from each well was transferred to a 96 well ELISA plate pre-coated with an anti IGF-1R capture antibody (R&D Systems MAB391) at 2µg/ml and blocked with 4%BSA/TBS. In some experiments an alternative capture antibody was used (2B9 SEQ ID NO: 10 and 11 coated at 1µg/ml). The plate was incubated overnight at 4°C. The plate was washed with TBST (TBS+0.1%Tween 20) and a Europium labelled anti Phosphotyrosine antibody (PerkinElmer DELFIA Eu-N1 PT66) diluted 1/2500 in 4%BSA/TBS was added to each well. After 1 hour incubation the plate was washed and DELFIA Enhancement (PerkinElmer 1244-105) solution added. After 10 min incubation the level of receptor phosphorylation was determined using a plate reader set up to measure Europium time resolved fluorescence (TRF).

Figure 8 shows an example of the inhibition of receptor phosphorylation mediated by purified murine monoclonal antibodies 6E11, 5G4 and 15D9, data collocated for experiments done at the same time for different plates with experiments done at the same time.

Figure 9 shows an example of the inhibition of receptor phosphorylation mediated by H1L0 in comparison to the chimeric 6E11 antibody (6E11c).

Figure 10 shows an example of the inhibition of receptor phosphorylation mediated by H0L0 and H1L0 in the context of a wild-type IgG1 Fc region and a substituted IgG1 Fc region (IgG1m(AA)).

In a different sets of experiments using a similar methodology, the IC50 values for inhibition of receptor phosphorylation were obtained and confirm that the humanised and 6E11 murine parental antibody show comparable profiles for inhibition of IGF-I and IGF-II mediated receptor phosphorylation (Table 2).

Table 2 - IC50 values for selected antibodies in IGF-I and IGF-II stimulated phosphorylation assays with 95% upper and lower confidence intervals

Antibody	IGF-1 Stimulation IC50 (ug/ml)	IGF-II Stimulation IC50 (ug/ml)
H0L0	0.06069	0.08145
H1L0	0.08359	0.10546
H0L0 IgG1m(AA)	0.08485	0.09968
H1L0 IgG1m(AA)	0.08263	0.10546
6E11 Parental	0.02445	n/a

In a different set of experiments using a similar methodology, the IC50 values for inhibition of receptor phosphorylation were obtained and confirm that the humanised antibody H0L0 and the murine parental antibody 6E11 show comparable activity. In parallel, the activity of the antibodies against the insulin receptor was tested using a 3T3 cell line engineered to express the human insulin receptor. In this experiment, none of the antibodies showed inhibition of insulin-induced receptor phosphorylation.

Table 2a IC50 values for selected antibodies in IGF-IR and IR DELFIA assays. Each value represents the mean of two data points. The negative control antibody is a hybrid IgG1.

	IGF-1R DELFIA - IC50 (nM)		IR DELFIA - IC50 (nM)	
Antibody	**Plate A**	**Plate B**	**Plate A**	**Plate B**
H0L0	0.25	0.19	>133nM	>133nM
6E11	0.18	0.16	>133nM	>133nM
Negative control	>133nM	>133nM	>133nM	>133nM

Example 14 - Competition ELISA

ELISA plates were coated with an anti human IGF-1R antagonistic antibody (MAB391, R&D Systems) at 2µg/ml and blocked with 4%BSA/PBS. Poly-His tagged recombinant human IGF-1R (R&D Systems #305-GR) was added at 400ng/ml in the presence of purified monoclonal antibodies and incubated for 1 hour at room temperature. The plate was washed in TBST (TBS+0.1% Tween 20) before the

addition of HRP labelled anti poly-his antibody (Sigma A7058-1VC) at 12-30µg/ml. The plate was incubated for 1 hour before further washing and development with OPD substrate (Sigma P9187). The reaction was stopped by the addition of 2M Sulphuric acid and absorbance was measured at 490nm.

Figure 11A shows an example of the activity of various purified murine monoclonal antibodies in the competition ELISA. Data collocated for experiments done at the same time.

Figure 11B shows an example of the activity of H1L0 in the competition ELISA in comparison to the 6E11 chimera (6E11c). Data collocated for experiments done at the same time.

Figure 12A shows an example of the activity of various purified humanised antibodies in the competition ELISA in comparison to the murine parental antibody (6E11) and chimera (6E11c). In figure 12A, H0L0 and H0L0 IgG1m(AA) showed an increased signal compared to the repeat assays shown in Figures 12B and 12C.

Figure 12B-C show examples of the activity of various purified humanised antibodies in the competition ELISA.

Example 15 - Cynomolgus macaque IGF-1R Binding ELISA

96 well ELISA plates were coated overnight with recombinant Cynomolgus macaque IGF-1R (see Example 4) at 1-2µg/ml and blocked with 4%BSA/PBS. Purified anti-hIGF-1R antibodies were added and incubated for 1 hour at room temperature. The plates were washed in TBST and HRP conjugated anti mouse Ig (DAKO #P0260) was added to each well at 0.6-1.0 µg/ml. Plates were incubated for 1 hour at room temperature, washed with TBST and developed with OPD substrate (Sigma P9187) or TMB substrate (Sigma T8665). The reaction was stopped with 2M Sulphuric acid and the level of binding determined by measuring the absorbance at 490nm (for OPD) and 450nM (for TMB). For antibodies containing a human IgG1/Cκ constant region, the HRP conjugated anti mouse Ig detection antibody was substituted with a goat anti-human Kappa Light Chains peroxidase conjugate (Sigma, A7164)

Figure 13A shows an example of purified murine monoclonal antibodies binding to recombinant cynomolgus macaque IGF-1R. Data collocated for experiments done at the same time.

Figure 13B shows an example of purified humanised monoclonal antibodies binding to recombinant cynomolgus macaque IGF-1R in comparison to the 6E11 chimera (6E11c).

Example 16 - Insulin Receptor binding ELISA

96 well ELISA plates were coated overnight with recombinant human Insulin Receptor (R&D Systems 1544-IR) at 0.5µg/ml and blocked with 4%BSA/PBS. Purified anti-hIGF-1R antibodies or mouse anti-human Insulin Receptor antibody (R&D Systems MAB15441) were added to the plates and incubated for 1 hour at room temperature before washing with TBST. HRP conjugated anti mouse Ig (DAKO #P0260) was added to each well at 1/500 or 1/2000 in 4%BSA/PBS and the plates incubated for 1 hour. Plates were washed and developed by the addition of TMB substrate (Sigma T8665) or OPD (Sigma P9187). The reaction was stopped with 2M Sulphuric acid and binding detected by measuring absorbance at 450nm or 490nm. For the detection of antibodies containing a human IgG1/Cκ constant region, the detection antibody listed above (HRP conjugated anti mouse Ig) was substituted with a goat anti-human Kappa Light Chains peroxidase conjugate (Sigma, A7164).

Figure 14 shows an example of the insulin receptor binding ELISA using purified murine monoclonal antibodies. In contrast to the positive control antibody (R&D Systems MAB15441), purified antibodies 6E11, 5G4 and 15D9 showed no binding to the insulin receptor at concentrations up to 10µg/ml. Data collocated for experiments.

In a different experiment, the insulin receptor binding profile of various humanised antibodies was tested using a similar methodology. Whilst a positive control antibody (R&D systems AF1544) showed good binding, there was no detectable binding of the humanised antibodies to recombinant insulin receptor at concentrations of up to 50µg/ml (Figure 15).

Example 17 - Determination of kinetics of binding

The binding kinetics of anti-IGF-1R antibodies for human IGF-1R were assessed using the Biacore™ system. The kinetic analysis was carried out using an antibody capture method. Briefly, an anti-mouse IgG antibody (Biacore, catalogue number BR-

1005-14) was used for analysis of mouse parental antibodies and Protein A, for humanised antibodies. Either the anti mouse antibody or the Protein A was immobilised on a CM5 Biosensor chip by primary amine coupling in accordance with Biacore™ standard protocols, utilising the immobilisation Wizard facility, inherent in the machines software, (levels of 3000-4000 resonance units (RU's) where typically immobilised). Anti-IGF-1R antibodies were then captured either directly from hybridoma supernatants or from purified material. The capture levels for supernatants depended upon the starting concentration of the hybridoma and these varied between around 20RU's to 650RU's. For the purified material, the level captured for the antibodies tested were generally between 20 and 600RU's. After capture, the baseline was allowed to stabilise before recombinant IGF-1R, histidine tagged material from R&D Systems (catalogue number 305-GR) was then passed over the surface at defined concentrations (usually in the range of 0-256nM). Due to the high affinity of the interaction, dissociation times of up to one hour were used. Regeneration was by acid elution using either 100mM phosphoric acid or 10mM Glycine, pH 1.5, the regeneration did not significantly affect the surfaces ability to capture antibody for another analysis step.. The runs were carried out at both 25 °C and 37°C. The experiments were carried out on the T100 Biacore™ system, using the T100 control and analysis software. The experimental data was fitted to the 1:1 model of binding inherent in the machines analysis software.

Tables 3 - 7 show a series of experiments conducted with supernatant and purified material.

Table 3 - Kinetic data for a selection of the purified murine IGF-1R monoclonals at 25 °C and 37 °C

Antibody	**Affinity (nM) 25°C (Run 1-T0011 R6)**	**Affinity (nM) 37°C (Run 2-T0011 R4)**	**Affinity (nM) 25°C (Run 3-T0022 R5)**
6E11	0.09	0.164	0.14
5G4	3.0	5.9	Not tested
15D9	0.233	0.558	Not tested

Table 4 – Kinetic data for supernatant material of a H1L0 and H0L0 in comparison with 6E11c. The run (T0037 R3) was carried out at 37°C.

Antibodies	***Ka***	***Kd***	**KD (nM)**
H1L0	7.56e4	3.52e-5	0.47
6E11c Supernatant	8.14e4	3.13e-5	0.38
6E11c Purified	8.52e4	3.32e-5	0.39

Table 5 – Kinetic data for supernatant material H0L0 and H0L0 IgG1m(AA) and H1L0 and H10L0 IgG1m(AA) in comparison with 6E11c. The run (T0040 R2) was carried out at 37°C.

Run 1

Antibodies	**Ka**	**Kd**	**KD (nM)**

H1L0 ((supernatant)	7.56e4	3.52e-5	0.47
6E11c (supernatant)	8.14e4	3.13e-5	0.38
6E11c (purified)	8.52e4	3.32e-5	0.39

Run 2

Antibodies	**Ka**	**Kd**	**KD (nM)**
H1L0 (supernatant)	6.82e4	4.28e-5	0.63
6E11c (purified)	7.59e4	3.25e-5	0.43

(H1L0 supernatants are the same for runs 1 and 2, however the 6E11c purified are different batches.)

Table 6 – Kinetic data for purified H0L0 and H1L0 in comparison with the 6E11 chimera (6E11c). The run (T0041 R1) was carried out at 37°C.

Antibodies	**Ka**	**Kd**	**KD (nM)**
H0L0	6.24e4	3.93e-5	0.63
H1L0	6.54e4	2.95e-5	0.45
6E11c	6.60e4	2.45e-5	0.37

Table 7 – Kinetic data for purified H0L0 and H0L0 IgG1m(AA) and H1L0 and H10L0 IgG1m(AA) in comparison with the 6E11 chimera (6E11c). Three independent runs were carried out at 37°C.

	Run 1 – T0044 R3			Run 2 – T0044 R4			Run 3 – T0044 R6		
Antibody	***Ka***	***Kd***	**KD (nM)**	***Ka***	***Kd***	**KD (nM)**	***Ka***	***Kd***	**KD (nM)**
H0L0	5.13e4	2.68e-5	0.52	6.62e4	3.97e-5	0.59	6.17e4	5.56e-5	0.90
H0L0 IgG1m(AA)	5.40e4	2.67e-5	0.49	7.68e4	4.00e-5	0.52	7.38e4	5.71e-5	0.77
H1L0	4.97e4	2.09e-5	0.42	6.67e4	3.47e-5	0.52	7.04e4	4.18e-5	0.59
H1L0 IgG1m(AA)	5.07e4	2.17e-5	0.43	6.61e4	3.22e-5	0.49	6.48e4	4.44e-5	0.69
6E11c	3.99e4	8.71e-6	0.22	6.78e4	2.29e-5	0.34	6.75e4	4.02e-5	0.60

In a different experiment the following approach was used. Protein A was immobilised on a CM5 surface by primary amine coupling in accordance with the manufacturers standard protocols. Antibodies against IGF-1R were captured on this surface and after a period of stabilisation recombinant human or cynomolgus IGF-1R was injected over this captured surface. Generally, the concentrations of IGF-1R used were 256-16nM, with a 0nM injection (buffer only) also used for double referencing in accordance with best practice for Biacore kinetic analysis. Data was analysed using the 1:1 model inherent to the Biacore machine, the work was carried out on the T100 using HBS-EP running buffer at 37°C. The results confirm that the humanised variants H0L0 and H0L0 IgG1m(AA) show high affinity binding (~300-600pM) to recombinant human and cynomolgus IGF-1R and comparable kinetics to the 6E11 chimera.

Table 8 Kinetics of anti-IGF-1R antibodies versus recombinant human IGF-1R and cyno IGF-1R at 37°C. Data shown is from a single experiment.

	Human IGF-1R			Cynomolgus IGF-1R		
	ka	Kd	KD(nM)	ka	Kd	KD (nM)
6E11 chimera	1.22e5	2.61e-5	0.21	1.10e5	2.14e-5	0.19
H0L0	1.07e5	2.73e-5	0.25	9.18e4	2.56e-5	0.28
H0L0 IgG1m(AA)	1.14e5	3.20e-5	0.28	1.07e5	2.73e-5	0.25

A similar experiment has been performed which confirms that H0L0, H0L0 IgG1m(AA) and 6E11 chimera show comparable kinetics of binding. However the overall affinity was lower than seen in all previous experiments (at 1.88, 1.84 and 1.52nM respectively). The reasons for the apparent difference are unknown.

Example 18 - Inhibition of ligand binding determined using Biacore

The experiment was carried out using two different densities of captured biotinylated IGF-I. Briefly either 200 or 4000 RU's was stably captured on a streptavidin sensor chip. To test the neutralisation capacity of anti-IGF-1R antibodies, different concentrations of antibodies were pre-mixed with a fixed concentration of recombinant IGF-1R. As a control non biotinylated IGF-1 was also mixed with the same concentration of IGF-1R. This mixture was then passed over the IGF-I surface and the point of maximal association measured. This reading was then compared to a sample with the same concentration of his-tagged IGF-1R in the absence of anti-IGF-1R antibodies. The presence of a neutralising antibody blocked binding of IGF-1R to IGF-I and reduced the maximal observed association. Percentage inhibition was calculated by comparing the values. Regeneration was carried out using two pulses of 4M magnesium chloride. The experiments were carried out on a Biacore 3000 system.

Tables 9 and 10 below show the percentage inhibition obtained and also detail the concentrations of antibodies, IGF-1 and IGF-1R used to obtain these results.

Table 9 - Inhibition Values for the 200 RU's IGF-1 Surface

Antibody + IGF-1R complex	% Inhibition
IGF1 (125nM) + His IGF-1R (25nM)	69
IGF1 (500nM) + His IGF-1R (25nM)	89
6E11 (125nM) + His IGF-1R (25nM)	48
6E11 (500nM) + His IGF-1R (25nM)	50

Table 10 - Inhibition Values for the 4000 RU's IGF-1 Surface

Antibody + IGF-1R complex	% Inhibition
IGF1 (5μM) + His IGF-1R (50nM)	93
IGF1 (500nM) + His IGF-1R (50nM)	86
6E11 (500nM) + His IGF-1R (50nM)	48

In a different experiment, the ability of the humanised antibodies to directly block ligand binding was assessed using a Biacore-based methodology using captured biotinylated ligand (IGF-I, IGF-2). Biotinylated IGF-1 or IGF-2 was immobilised on a strepavidin biosensor chip to around 300RUs and 350RUs respectively. IGF-1R was passed over the surface at 50nM alone or at 50nM in a premixed solution containing either 250nM of anti-IGF-1R antibodies (for the IGF-I experiments) or 500nM of anti-IGF-1R antibodies (for the IGF-2 experiment). IGF-1R binding was also carried out in the presence of the natural ligands IGF-1 and IGF-2 (both unbiotinylated). The surface was regenerated using 100mM phosphoric acid. The experiments were carried out using HBS-EP buffer at 25oC on the Biacore 3000. Note: The analysis of the IGF-2 assay data was complicated by the fact that some of the antibodies showed non-specific binding to the IGF-2 surface alone. For immobolised IGF-I, the most efficient inhibitor of receptor binding was unlabelled IGF-I, with IGF-2 and H0L0 showing about 60% inhibition (Table 11). For immobilised IGF-2, the most efficient inhibitor of receptor binding were unlabelled IGF-I or IGF-2. In this assay, the neutralising antibodies, including H0L0 showed partial inhibition of IGF-2 binding (Table 11).

Table 11 – Neutralisation of binding of receptor to ligand

Sample	% Neutralisation binding of receptor to IGF-I	% Neutralisation of receptor binding to IGF-2
H0L0	60	37
H0L0 IgG1m(AA)	58	17
6E11 Chimera	44	20
IGF-1	87	98
IGF-2	68	92

Example 19 - Fluorescence Activated Cell Sorting (FACS) analysis

Colo205 cells ($2x10^7$ cells/ml) were stained with anti hIGF-1R purified antibodies at 10µg/ml for 1 hour in FACs buffer (4% FCS in PBS). Cells were also stained in a suitable negative control mouse antibody (Sigma #I5154). Cells were washed in FACS buffer and then stained with an anti-mouse IgG PE secondary antibody 1:100(Sigma P8547). After washing in FACS buffer and fixing in Cell Fix (Becton Dickinson) cells were analysed by flow cytometry..

Figure 16 demonstrates that antibody 6E11 is capable of recognising natively expressed IGF-1R on the surface of a human tumour cell line.

In a different experiment, the humanised antibodies were tested for their ability to stain various human tumour cell lines known to over-express IGF-1R. NCI-H838 lung carcinoma cells were stained with the selected antibodies at 100µg/ml for 45mins at 4°C. Binding was detected with a PE conjugated anti human IgG antibody (Sigma P8047). Samples were analysed by flow cytometry using a Becton Dickinson FACscan cytometer. The results shown in Figure 17 confirm that the humanised variants can bind to NCI-H838. A similar result has been obtained for MCF7 breast carcinoma cells and A549 lung carcinoma cells (Figure 18).

Example 20 - Immunohistochemistry on frozen tissue sections

Tissues were sectioned onto glass slides, fixed with acetone for 2 minutes and then loaded into an automated slide stainer (DakoCytomation S3400). Slides were then

blocked and stained with murine antibodies (primary antibody) and an anti-mouse Ig-HRP secondary antibody (DakoCytomation Envision Kit) using standard immunochemical staining methods. Following this secondary incubation, the slides were washed and developed using the DakoCytomation Envision DAB solution, rinsed, dehydrated and cover-slipped for viewing. An irrelevant control antibody (mouse IgG1 purified from a MOPC21 hybridoma) was used as a negative control.

The humanised and chimeric antibodies were analysed in a similar manner except that these antibodies were biotinylated to facilitate detection. However, the presence of the biotin-tag was found to decrease the activity of these` antibodies as determined by ELISA (data not shown), therefore the concentration of primary antibody used was increased to up to 100ug/ml. The secondary antibody listed above (DakoCytomation Envision Kit – Anti-mouse Ig-HRP conjugate) was substituted with streptavidin-HRP, (DakoCytomation Cat# 1016). An alternative irrelevant antibody was also biotinylated and used as a negative control (Sigma #I5154).

The samples were analysed as follows. After calibrating the instrument using the calibration carrier (#69935000, 05041103097), the slides were loaded into the ChromaVison automated cellular imaging system and scanned at 10X. Data analysis was performed to calculate the % tissue staining (defined as brown/brown+blue*100).

Figure 19 and 20 show that 6E11 stains human tumour tissue samples. A positive control antibody was included as a reference (Abcam, #4065).

Figure 21 shows that 6E11 chimera (6E11c) and H1L0 stain human tumour tissue samples.

In a separate experiment, humanised H0L0 was tested for it's ability to recognise human IGF-1R on human tumour tissue samples using a frozen tissue microarray obtained from Cytomyxx (array ID: MB-1002). This array contains 10 lung, 10 breast, 10 colon and 10 prostate tumour cores. H0L0 was biotinylated to facilitate detection. A frozen microarray slide (Cytomyxx MB-1002) was fixed with a 4°C solution of acetone/ethanol (50:50), for 5 minutes, washed and then treated with 3% hydrogen peroxide for 5 minutes to remove any endogenous peroxidase, The slides were stained with biotinylated anti-IGF-1R H0L0 at 7.0µg/ml for 1 hour at room temperature. After washing, a streptavidin peroxidase was applied for 20 minutes

and visualised with DAB (diaminobenzidine) for 2 minutes. Finally, the sections were washed and counterstained in Mayer haematoxylin, rinsed in tap water, dehydrated, cleared and mounted. The tumour samples are as follows: Lung (Right panel: Squamous cell tumour, Left panel: Adenocarcinoma; Breast (Right panel: Adenocarcinoma, ductal epithelium, Left panel: Adenocarcinoma, invasive); Colon (Right panel: Adenocarcinoma well differentiated, Left panel: Adenocarcinoma well differentiated; Prostate (Right panel: Adenocarcinoma, Left panel: Adenocarcinoma). The results confirm that H0L0 showed moderate/strong staining of the viable sections as summarised in Table 12 below. Representative high powered images (200x) of the lung, breast, colon and prostate tumours are shown in Figure 22, confirming that the biotinylated H0L0 antibody predominantly stains epithelial cells.

Table 12- *Summary of immunohistochemistry analysis of tumour tissue microarray*

Tumour type	**Positive staining (moderate/strong)**	**No staining**	**Missing sample**
Lung	4	3	3
Breast	7	3	0
Colon	6	3	1
Prostate	6	0	4

Example 21 - Inhibition of AKT signalling

Costar 96-well plates (#3598) were coated with 50µl of 2% Gelatin in PBS and incubated in a 37°C incubator for at least one hour. Prior to use, the plates were rinsed once with PBS. Primary human pre-adipocytes were trypsinized, centrifuge and the medium siphoned off. The cells were resuspended with 10 mL of warmed PreAdipocyte growth medium (ZenBio, #PM-1). Cell density was adjusted to 150,000 cells per mL in PreAdipocyte growth media (ZenBio). Two T225 Costar Flasks containing 50ml of media were each seeded with 1 million cells. The remaining cells were used to seed the Gelatin-coated 96-well plates (100µl = 15,000 cells per well) using a Multidrop384 or similar instrument. The cells were incubated overnight at 37°C in a 5% CO2 atmosphere, 90% humidity. The following day, the medium was removed, 200µl of Induction Medium) added and the plates covered with Breath-Easy gas-permeable film (Sigma#Z380059). The plates were incubated for 6 days at

37°C, in a 5% CO2 atmosphere, 90% humidity. After 6 days, the medium was aspirated and 200µl of Differentiation Medium added. The plates were covered with Breath-Easy gas-permeable film and incubated for 7 days at 37°C in a 5% CO2 atmosphere, 90% humidity. Following differentiation of the cells, the medium was aspirated and the cells rinsed once with 200µL of PBS. 75µl of Adipocyte Starve Medium was added and the plates covered and incubated overnight at 37°C in a 5% CO2 atmosphere, 90% humidity. Test samples were diluted in Adipocyte Starve media at 4X the final concentration. 25µL of diluted test compound was added to each well and incubated at 37°C for 1 hour. IGF-I ligand (R&D Systems, #291-G1) was diluted to 30 nM in Adipocyte Starve Medium and 20µL of 30nM IGF-I was added to each well (final conc. 5 nM). The plates were incubated at 37°C for precisely 5 min after which time the supernatant was removed by flicking the media into a sink. The plates were dried on paper towels.

65µl of Complete Lysis buffer (MSD Lysis buffer containing phosphatase and protease inhibitors) was added to each well and the plate sealed with heated plate sealer. The plates were either stored at -80°C (for later analysis) or placed on a shaker (approx. 500 rpm) for 15 mins at room temperature before performing the MSD Assay.

Levels of phosphorylated AKT (pSer473) were assessed using the MSD phosphorylation assay kit (#K111CAD). Briefly, 150µL per well of Blocking solution (MSD Blocker A dissolved in MSD Tris Wash buffer) was added to each well of an MSD Assay plate. The plate was sealed and placed on a shaker at 300 rpm using a bench top plate shaker for 1 hour at room temperature. The Blocking solution was removed from the MSD plate(s) and the plates washed four times with 200 µL/well of 1x MSD Tris wash buffer. 50µL/well of cell lysate from the cell plate(s) was transferred to the corresponding well of the MSD plate(s) and sealed. The plates were shaken at 300 rpm using a benchtop plate shaker for 1 hour at room temperature. The MSD plates were washed four times with 200µL per well using 1x MSD Tris wash buffer (ELx405).

25µL of diluted detection antibody mixture (10 nM final concentration) was added to each well of the MSD plate(s). The plates were shaken at 300 rpm using a bench top plate shaker for 1 hour at room temperature and then washed four times with 200µL per well using 1x MSD Tris wash buffer (ELx405). 150µL of Read Buffer T with surfactant was added to each well and the plates read with MSD 6000 SECTOR reader. Although signal intensity decreased with time in Read Buffer, the signal

window typically remained steady for approximately 20-30 minutes.

Table 13 below shows a summary of the data from three independent plates and indicates that purified murine parental, chimeric and humanised Mabs inhibit IGF-I mediated induction of AKT phosphorylation. . Plates 1 and 2 were run in parallel. Plate 3 was run on a separate day. The values are represented as pIC50 (= -log10(IC50) in g/ml)

Table 13 – Activity of various purified antibodies in the AKT phosphorylation assay

Antibody	Plate 1	Plate 2	Plate 3
6E11 parental	7.75	7.79	7.67
H0L0	7.65	7.76	7.34
H1L0	7.62	7.68	7.30
6E11c	7.59	7.32	7.34
Negative control	6.05	6.25	<5.82

In two different experiments, various humanised antibodies were tested for inhibition of AKT phosphorylation in response to IGF-I or insulin stimulation. For the IGF-I results shown in Table 14, the values are represented as mean pIC50 where pIC50 = -log10(IC50) in g/ml. Vmax = is the maxmimum inhibition expressed as a percentage of the signal in the absence of ligand. Experiment 1 is the mean of 4 runs. Experiment 2 is the mean of 3 runs. Experiments 1 and 2 were run approximately 6 months apart. For some antibodies (**H0L0), different batches of material were tested in parallel.In comparison to the negative control antibody. All the anti-IGF-1R antibodies showed a dose dependent inhibition of phosphorylation of AKT with the humanised antibody H0L0 showing comparable activity to the 6E11 mouse parental antibody or 6E11 chimeric antibody (Table 14).

Table 14 - Activity of various purified antibodies in the AKT phosphorylation assay.

	Experiment 1		Experiment 2	
Antibody	**Mean pIC50 (g/ml) +/- SD**	**Mean Vmax (g/ml) +/- SD**	**Mean pIC50 (g/ml) +/- SD**	**Mean Vmax (g/ml) +/- SD**
6E11 parental	ND	ND	7.58 (0.17)	113 (2.6)
H0L0**	7.33 (0.11)	107 (9.5)	7.42 (0.14)**	106 (3.6)
			7.53 (0.15)**	100 (7.9)
H1L0 IgG1m(AA)	7.24 (0.14)	116 (11.0)	ND	ND
6E11 chimera	7.34 (0.08)	114 (14.1)	ND	ND
IR3	7.17 (0.15)	105 (7.0)	7.45 (0.07)	108 (10.6)
Negative control	7.17 (0.30)	57.2 (9.4)	7.30 (0.89)	52 (1.2)

Since the assay system was also sensitive to insulin mediated phosphorylation of AKT (via the insulin receptor), the effect of the humanised antibodies on insulin signalling was assessed in parallel to the IGF-I experiments described above. In contrast to the results with IGF-I stimulation, the humanised antibodies showed no inhibition of insulin receptor signalling over and above the non-specific effects observed with the negative control antibodies (data not shown). These results were observed in two independent experiments (seven runs in total).

Example 22 - Proliferation assay with MCF7 cells

MCF-7 cells (ATCC HBT-22) were seeded into 96 well plates at a density of 10000 cells/well and grown for 2 days in complete media (MEM+Earles salts + 10% FCS + 0.1mg/ml bovine insulin (Sigma I0516)). Cells were washed and incubated in serum free MEM (no serum, no insulin) for 4 hours. Media was removed and replaced with a range of concentrations of purified antibodies (0.014 - 10µg/ml) diluted in serum free media (100µl/well). Cells were incubated for 1 hour before the further addition of IGF-I (R&D Systems #291-G1) to a final concentration of 50ng/ml. All treatments were carried out in triplicate. Cells were incubated for 5 days at 37degC, 5% CO2. After incubation, 15µl of MTT dye solution (Promega #G402A) was added to each well and

the plates incubated for a further 4 hours. 100µl of Stop/Solublisation solution (Promega #G401A) was added to each well and the plate shaken gently overnight at room temperature. The following day the level of proliferation was determined by measuring the absorbance at 570nm using a plate reader.

Figure 23 shows the activity of various purified mouse monoclonal antibodies to inhibit the proliferation of tumour cells.Data collocated for experiments.

Example 23 - Proliferation assay – LISN cells

LISN cells (3T3 hIGF-1R) were seeded into white walled 96 well plates (Corning 3610) at a density of 10 000 cells/well and grown for 1 day in complete media (DMEM-Hepes modification + 10% FCS). The media was removed and cells incubated in serum free DMEM for 4 hours. Media was removed and replaced with a range of concentrations (0.0041-3µg/ml final concentration) of purified antibodies diluted in serum free media (50µl/well). Cells were incubated for 1 hour before the further addition of 50µl/well IGF-1 (R&D Systems 291-G1 or IGF-I - see Examples 5 and 6) to a final concentration of 50-60ng/ml. All treatments were carried out in triplicate. Cells were incubated for 0-3 days at 37degC, 5% CO2. After incubation, 100µl of freshly prepared Promega CellTitre-Glo reagent (Promega G7571) was added to each well and the plates shaken for 2 mins. The plate was further incubated at room temperature for 10mins to allow the signal to stabilise before measuring the luminescence signal with a Wallac Victor plate reader.

Figure 24 and 25 A-E show the activity of purified 6E11 murine monoclonal antibody, 6E11c H0L0 and H0L0 IgG1m(AA) and H1L0 and H1L0 IgG1m(AA). The data confirms that the H0L0 and H1L0 can inhibit tumour cell proliferation in vitro.Data collocated for experiments.

Example 24 - Inhibition of cell cycling

NCI-H838 (ATCC CRL-5844) cells were seeded into 24 well microplates at a density of $2X10^5$ cells/well and grown overnight in 1ml complete RPMI (RPMI+10%FCS). The following day the cells were washed with SFM (serum free RPMI media) and incubated in 1ml of the same media for 4 hours. The media was aspirated from the cells and 500µl of SFM containing 20µg/ml of purified antibodies was added (10µg/ml final concentration). Cells were incubated for 1 hour. In some wells, IGF-I (R&D Systems 291-G1) in SFM was added to a final concentration of 50ng/ml. The treated cells were incubated overnight. The following day the cells were washed gently in

PBS and then harvested by adding 200µl of Versene solution (Invitrogen #15040). The cell suspensions were transferred to a 96 well V-bottomed plate. After pelleting the cells by centrifugation they were fixed by the addition of chilled 80% Ethanol and incubation on ice for 30min. Cells were pelleted and re-suspended in 200µl of 50µg/ml Propidium Iodide, 0.1mM EDTA, 0.1% Triton X-100, 0.05mg/ml RNAse A. Cells were incubated on ice in the dark until being analysed by flow cytometry.

Figure 26. shows the cell cycle status of the various treatment groups in the presence of IGF-I, the cells are induced to cycle. In the presence of 6E11 antibody, cell cycling was inhibited at levels comparable to that of cells incubated in the absence of IGF-I.

Example 25 - Protection from apoptosis

A 96 well microplate was seeded with NCI-H838 cells (ATCC CRL-5844) at a density of 10000 cells/well in 100µl complete RPMI media and grown for 2 days. Cells were then washed in SFM (RPMI no serum) and incubated in 100µl SFM for 4-5 hours. The media was removed prior to treatment with either no antibody, a negative control antibody or a purified anti hIGF-1R antibody (6E11) at 20µg/ml. Cells were additionally treated with either SFM alone, SFM + IGF-1at 20ng/ml, SFM + Camptothecin at 5µM or SFM + Camptothecin at 5µM + IGF-1 at 20ng/ml. All treatments were tested in triplicate in a final volume of 100µl. The plate was then incubated for 20 hours. The media was aspirated from the wells and the cells lysed by the addition of 200µl of 0.5% NP-40 in PBS followed by 5 min incubation with shaking at room temperature. 20µl of lysate was transferred to a prepared microplate from the Roche Cell Death ELISA Kit and 80µl of incubation buffer added. The protocol described in the kit insert (Roche Cat.NO: 1 544 675) was followed and the absorbance at 405nm measured using a microplate reader.

Figure 27 shows that the presence of IGF-I affords NCI-H838 cells some protection from camptothecin induced apoptosis. The addition of 6E11 reversed the IGF-1 mediated protection from apoptosis

In a different experiment, selected antibodies were tested for their ability to prevent IGF-1 rescue from camptothecin induced apoptosis in A549 cells. A549 cells were plated at $1x10^4$, grown in 96 well plates and treated with 20µg/ml of the selected antibodies in serum free conditions for 1 hour. IGF-1 at 15ng/ml and Camptothecin at 5µg/ml were added together and cells incubated overnight. The level of apoptosis

was measured using a Roche Cell Death Detection ELISA kit (Roche 11774425001) which estimates the relative level of DNA fragmentation. As shown in Figure 28, all the humanised antibodies prevented IGF-1 induced rescue from apoptosis.

Example 26 - Absence of agonism in the presence or absence of cross-linking antibodies

96 well microplates were seeded with 3T3/LISN c4 cells at a density of 10,000 cells/well in complete DMEM (DMEM Hepes modification + 10%FCS) and grown for 2 days. Purified anti IGF-1R antibodies were titrated onto the cells in complete DMEM, each dilution being tested in triplicate. An antibody reported to have agonistic activity (#556000, BD Biosciences) and/or 50ng/ml of IGF-I (incubate for 20-30 mins) were included in some experiments as a positive control. Negative controls of irrelevant antibody and media alone were included. In other experiments, an anti-mouse cross-linking antibody (Sigma M8144) or an anti human cross linking antibody (Sigma I3382) were included in the antibody titration at a ratio of 2:1 [anti IGF-I Ab]:[cross linking Ab]. Plates were incubated for 30mins. Media was aspirated and cells were washed gently with PBS once before being lysed with RIPA lysis buffer(150mM NaCl, 50mM TrisHCl, 6mM Na Deoxycholate, 1% Tween 20) plus protease inhibitor cocktail (Roche 11 697 498 001). The plate was placed at -20°C overnight. After thawing, 100µl samples of lysate were transferred to a 96 well ELISA plate pre-coated with an anti IGF-1R capture antibody (2B9) at 2µg/ml and blocked with 4%BSA/TBS. The plate was incubated overnight at 4°C. The plate was washed 4 times with TBST (TBS+0.1%Tween 20) and a Europium labelled anti Phosphotyrosine antibody (DELFIA Eu-N1 PT66, PerkinElmer) diluted 1/2500 in 4%BSA/TBS was added to each well. After 1 hour incubation the plate was washed as before and 100µl DELFIA Enhancement solution (PerkinElmer 1244-105) added. After 10 min incubation the level of receptor phosphorylation was determined using a plate reader set up to measure Europium time resolved fluorescence (TRF).

Figure 29. shows that 6E11 had no agonistic activity at concentrations up 10µg/ml in the presence of cross-linking antibodies.Data collocated for experiments.

In a different experiment, the pre-adipocyte assay system was used to determine if the 6E11-series of humanised anti-IGF-1R antibodies modulate the basal levels of phospho-AKT which might indicate agonistic properties. In this experiment, the pre-adipocytes were differentiated and treated as described in example 21. However, the stimulation step was removed in order to assess the basal level of AKT phosphorylation in the presence of humanised antibody. The results show that the

humanised antibodies at concentrations up to 20μg/ml, there was no increase in basal AKT phosphorylation (Figure 30).

In a parallel set of experiments using the lung carcinoma cell line A549, the basal AKT phosphorylation levels were assessed in the presence of 0-20μg/ml antibody and absence of ligand. Two different batches of H0L0, a negative control sample and 11C11 (an antibody which shows moderate activation of receptor phosphorylation, see Figure 31). The y-axis shows AKT phosphorylation levels in arbitrary units. Consistent with the data presented in example 21, H0L0 inhibited IGF-I mediated Akt phosphorylation in a dose dependent manner (with IC50s in the 200-500ng/ml range, data not shown). However, increasing concentrations of antibody in the absence of ligand appear to cause a small increase the basal levels of phospho-Akt. However the signal appears to plateau and reaches no greater than 3-fold the resting levels with two different batches of material. The reason for the small increase in signal is unknown and this data is not supported by any other experimental data.

In a parallel set of experiments using the LISN-c4 3T3 cells, the effects of the humanised antibodies on basal levels of receptor phosphorylation was assessed in the absence of ligand stimulation. LISN cells were incubated in the presence of selected antibodies at a range of concentrations (27ng/ml -20μg/ml) for 30mins. A titration of the known agonistic antibodies 11C11 and BD556000 (Becton Dickinson) were also included. Control wells stimulated with IGF-2 at 100ng/ml can be included. Phosphorylation of IGF-1R was measured using the DELFIA assay described previously. In contrast to two positive control antibodies, 11C11 and BD556000, which induced a dose dependent increase in receptor phosphorylation, the humanised antibodies showed no increase in the basal levels of phosphorylated receptor (Figure 32). Since cross-linking of surface bound antibody has the potential to induce receptor signalling, the experiment was repeated in the presence of cross-linking antibodies. LISN cells were incubated with a range of concentrations of selected antibodies mixed with appropriate cross linking antibodies at a ratio of 2:1 for 30min (Sigma I3382 for the humanised antibodies and Sigma M8144 for the mouse antibodies). Phosphorylation of IGF-1R was measured using the DELFIA assay described previously. The results show that in contrast to 11C11 which increased the levels of phosphorylated receptor, the humanised antibodies showed no increase in basal levels of phosphorylated receptor (Figure 33).

The effects of addition of humanised antibodies on the proliferation of both LISN-c4 and NCI-H929 (human multiple myeloma cell line) in the absence of ligand were also

tested. NCI-H929 cells were plated into a 96well plate at $4x10^4$ cells/ well in serum free media. Dilutions of the selected antibodies were added in the range 20-0.019µg/ml. Cells were incubated for 4 days at 37°C before the proliferation was measured using a Promega Cell Titre Blue assay kit (Promega G8081). Figure 34 shows that H0L0 does not stimulate the proliferation of NCI-H929 cells in serum free media. Similar results were observed with LISN-c4 cells (data not shown).

Example 27 - Allograft model – 3T3/LISN c4

An in vivo tumour model using 3T3/LISN c4 cells was used to establish the ability of 6E11 murine monoclonal antibody to inhibit the growth of pre-established tumours in athymic nude mice. Tumours were induced by similar methods to those published in Cohen et al, Clinical Cancer Research 11:2063-2073 ((2005). In summary, 2.5 $x10^6$ LISN cells suspended in 0.1ml of Matrigel™ were subcutaneously inoculated into 4-6 week old athymic CD1 nu/nu mice. Once tumours had reached approximately $150mm^3$ in size, mice were treated twice weekly for three weeks with 250µg of antibody in 0.2ml of PBS by intraperitoneal injection. Tumours were measured by Vernier callipers across two diameters three times per week and the volume calculated using the formula (length x $[width]^2$)/2. Data were analysed as follows: Log_{10} transformed tumour volumes were analysed using a random coefficient regression analysis. This estimates the intercept (baseline) and slope (rate of tumour growth) for each group. Compared with the PBS treated group, there was a reduction of 31% in the growth rate in the 6E11 group (Figure 35, p=0.0007).

In a similar experiment, nude mice were implanted sub-cutaneously with $2.5x10^6$ cells in Matrigel. Eighteen days after implantation, mice with tumor volumes of 100-200 mm^3 were randomized into groups of 8 animals/treatment group. Anti-IGF-1R antibody 6E11 was administered by intraperitoneal injection at 250µg/mouse and 100µg/mouse dose, twice weekly for 3 weeks. Control animals received saline at the same schedule. Tumor size and mouse body weight were measured twice weekly. Compared with the saline treated group, there was a reduction of 56% and 70% in the tumour volume measured at day 35 for the 100µg/mouse and 250µg/mouse groups respectively (Figure 36).

In a different study, the activity of various antibodies was tested compared to the PBS negative control group. CD1 nu/nu athymic mice were each implanted with $2.5x10^6$ LISN/3T3-c4 cells suspended in Matrigel subcutaneously and tumour size measured by callipers and volume calculated by the following equation: Tumour

volume = length x (width)2 x 0.5 . Once tumours had reached a mean volume of ~150mm3, animals were randomised into groups with comparable tumour sizes and each animal received 250µg of the appropriate monoclonal antibody in PBS intraperitoneally twice weekly for three weeks. The treatment groups were 6E11 (mouse parental mAb), two different batches of H0L0 IgGm(AA) and H0L0. PBS only treatment was used as a vehicle control for these studies. Group size for this study was a minimum of fourteen animals. Data is presented as of mean tumour volume ± standard error versus days post antibody treatment. Both 6E11 and H0L0 showed statistically significant reductions in the tumour growth rate of 52% ($P<0.0001$) and 60% ($P<0.0001$) respectively (Figure 37). Consistent with an earlier study (data not shown), neither batch of H0L0 IgG1m(AA) antibody significantly altered the tumour growth rate compared with the PBS control group.

In addition to a reduction in tumour growth rate, there was an improvement in survival in time-to-cull of mice treated with 6E11 and H0L0 groups compared with the PBS control (data not shown). Consistent with the tumour growth data, the H0L0 IgGm(AA) showed no benefit in delaying the time-to-cull (data not shown).

A repeat of this study was carried out using 6E11 and H0L0 antibodies with the exception that Matrigel was not used. The results from this study mirrored those of the previous study, with 6E11 and H0L0 showing a significant reduction in tumour growth rate of 20% ($p=0.0464$) and 29.7% ($p=0.0037$) respectively compared to the PBS control. There was also an improvement in survival in time-to-cull of mice treated with 6E11 and H0L0 groups compared with the PBS control. An irrelevant antibody was used as a control in this experiment and exhibited a similar profile to the PBS group (data not shown).

Example 28 - Growth inhibition of Colo205 cell tumours by 6E11 mouse parental antibody

An in vivo tumour model using Colo205 cells was used to establish the ability of 6E11 murine monoclonal antibody to inhibit the growth of pre-established tumours in HRLN female nu/nu mice. $1x10^6$ Colo205 cells were suspended in 50% Matrigel and subcutaneously implanted into the flank of the nude mice. Once tumours had reached approximately 80-120mm^3 in size (equivalent to day 1 in Figure 38), mice

were treated every 3 days with 10mg/kg of antibody by intraperitoneal injection, for a total of 10 injections. Tumours were measured by callipers and the volume calculated using the formula (length x [width] ^{2})/2. Data were analysed as follows: Log_{10} transformed tumour volumes were analysed using a random coefficient regression analysis. This estimates the intercept (baseline) and slope (rate of tumour growth) for each group. Compared with the vehicle control (PBS), there was a 58% reduction in the growth rate in the 6E11 antibody (Figure 38, p=0.0019).

A similar experiment to that described above was performed on a separate occasion. However, in this second experiment using Colo205 cells no inhibition of tumour growth was observed for the 6E11 treated animals. The reasons for the absence of inhibition with 6E11 are unknown (data not shown).

A similar experiment to that described above was also performed using mice implanted with $1x10^7$ A549 cells. However, in this experiment no inhibition of tumour growth was observed for the 6E11 treated animals. The reasons for the absence of inhibition with 6E11 are unknown (data not shown).

Whilst the data from these last two experiments appears to show that the antibodies of the invention do not inhibit tumour growth in these models, it is believed that the first two tumour models (the allograft model and the first colo205 model) are more robust. A control antibody that gave a positive signal (i.e. showed inhibition of tumour growth) in these first two models showed no inhibition in the second Colo205 tumour model study or the A549 tumour model study, hence we have more confidence that the data from the first two tumour models is more indicative of activity of the test antibody than that of the second two models.

Three additional xenograft studies have been carried out with 6E11 or humanised variants thereof. In the first study, the activity of 6E11 was compared to H0L0 IgGm(AA) in the Colo205 model. In contrast to the results presented in Figure 38, there was no evidence for inhibition of tumour growth following treatment with 6E11 or H0L0 IgGm(AA) compared to the PBS control group. In a repeat study where the treatment groups were PBS, 6E11, H0L0, a positive control antibody and an irrelevant antibody control, no groups showed any evidence of inhibiting tumour growth with the exception of positive control which showed a 23% reduction in the rate of tumour growth relative to PBS (p=0.003). However, the irrelevant antibody control showed a 15% inhibition of tumour growth in comparison to PBS (p=0.053). No antibodies showed inhibition of tumour growth relative to the irrelevant antibody

control.

In a different xenograft model (MCF-7 breast tumour model), animals were treated with PBS, 6E11, H0L0, a positive control antibody and an irrelevant antibody control. In this study, no treatment groups separated from either PBS or the irrelevant antibody control. In both studies, treatment with paclitaxel significantly inhibited tumour growth (data not shown).

In a post-study analysis, tumour samples harvested at the end of all three studies were assessed for receptor expression by immunohistochemistry. No evidence for IGF-1R receptor expression was observed using a biotinylated H0L0 probe whilst the same labelled antibody positively stained a LISN/3T3 c4 tumour sample and patient tumour samples.

The reasons for the apparent difference in activity of the antibodies between the initial Colo205 study presented in Figure 38, the in-house results from the LISN model (studies IGF-1R-11 and IGF-1R-12) and the results from the three additional studies which did not show antibody mediated inhibition of tumour growth are unknown. However, the fact that the tumours derived from at least some of the Colo205 and MCF-7 cells lines are receptor negative (including the PBS control group) at the end of the study raises concerns about the validity of these particular experiments for assessing the in vivo efficacy of anti-IGF-1R antibodies.

Example 29 - Kinetics of receptor recycling

To investigate the re-appearance of receptor upon withdrawal of antibody, NCI-H838 cells were incubated in growth media in the presence of H0L0 (H0L0) antibody at 1.67μg/ml presented in Figure 39 The first sample of cells was harvested at 0.5 hours post antibody addition. All other samples were washed thoroughly with PBS at 3 hours post antibody addition before returning to growth media. Cells were then harvested at 3, 4, 5, 6, 7 and 24 hours post antibody addition and assessed for IGF-1R expression by Western blot using a rabbit anti IGF-1Rβ c20 antibody (Santa Cruz, sc713). Binding was detected using HRP antibody and IgG1 kappa was used as a negative control antibody. Lanes 1 to 7 are harvests at: 0.5, 3, 4, 5, 6, 7, 24 hours. Lane 8 & 9 are the no antibody control and negative control antibody (Sigma I5154) respectively and were harvested at 3 hours. Magic Mark (Sigma, LC5602) is shown

in Lane 10. Following removal of the antibody (at t=3 hours), the western blot shows that at t=7 hours (lane 6) there is no evidence for IGF-1R expression. In contrast by t=24 hours (lane 7, 21 hours after antibody removal), there is a strong band consistent with the IGF-1R$_\beta$ chain.

A second experiment (data not shown) confirmed that the re-appearance of receptor was maintained at 48, 72 and 96 hours. These data suggest that re-appearance of the majority of the receptor occurs within the first 4-21 hours following removal of the antibody.

Example 30 - IGF-1R/IR Heterodimer binding assay

In cells which express both the IGF-1R and insulin receptors (Insulin receptor), a hybrid receptor IGF-1R:Insulin receptor (InsR) has been shown to exist (Pandini et al. (1999) Clin Cancer Res., 5(7):1935-44). It has recently been shown that an anti-IGF-1R antibody which did not cross-react against the Insulin receptor, reduced Insulin receptor levels, most likely by internalising and degrading the hybrid receptor (Sachdev et al. (2006) Cancer Res., 66(4):2391-402).

Humanised antibodies were assessed for their activity against the hybrid receptor using co-immunoprecipitation assays. In the first experiment shown in Figure 40A, COLO-205 colon carcinoma cells and recombinant NIH-3T3 cells expressing human Insulin receptor were lysed on ice for 10 minutes with 1% NP40 lysis buffer, clarified by centrifugation at 16400rpm for 20 minutes at 4°C. Soluble cellular proteins (500µg) were immunoprecipitated with 5µg antibody against either IGF-1R (6E11, 6E11 chimera, H0L0 IgG1m(AA), IGF-1R beta, Insulin receptor (beta), or non-targeting human IgG (control). Immunoprecipitated proteins were subjected to reducing SDS PAGE on 4-12% polyacrylamide gels, transferred to PVDF membranes and immunoblotted for either IGF-1R or Insulin receptor. Fluorescently-tagged secondary antibodies were used and the IGF-1R/Insulin receptor immunoblots imaged using a LI COR Odyssey system. In a different experiment (Figure 40B) using a similar methodology as described above H0L0 and H0L0 IgG1m(AA) were compared. Note the unbound fraction relates to the material in the supernatant post-immunoprecipitation.. The 97kDa β subunit of IGF-1R and 200kDa full length IGF-1R and 95kDa β subunit of Insulin receptor and 200kDa full length Insulin receptor are shown in the figures. As shown in Figure 40A and 40B, the humanised antibodies were able to co-immunoprecipitate the insulin receptor and

IGF-1R from the Colo205 human carcinoma cell-line at levels comparable to the 6E11 chimera. The same antibodies did not immunoprecipitate the Insulin receptor in the absence of IGF-1R (see Figure 40A, fourth panel, lane 1-5), indicating that the antibodies were not reactive against Insulin receptor. Although the vast majority of IGF-1R was removed from the cell lysate after immunoprecipitating with the anti-IGF-1R antibodies further indicating the humanised antibodies can bind both heterodimeric IGF-1R:Insulin receptor and homodimeric IGF-1R receptors (Figure 40B, upper panel lanes 3-5), a good proportion of Insulin receptor (most likely the homodimeric portion) remained in the cell-lysates post-IP (Figure 40B, lower panel lanes 3-5)

Example 31 – Proliferation of NCI-H929 cells

The human myeloma cell line NCI-H929 was stimulated with IGF-1 in the presence of various concentrations of the selected antibodies in serum free media. NCI-H929 cells were washed in serum-free medium and plated into a 96well plate at $4x10^4$ cells/well. Dilutions of the selected antibodies were added in the range 20-0.019µg/ml for 1 hr at 37°C before addition of a fixed concentration of IGF-1 (25ng/ml). Cells were incubated for 4 days at 37°C before the proliferation was measured using a Promega Cell Titre Blue assay kit (Promega G8081). Figure 41 shows the dose dependent inhibition of IGF-1 driven proliferation of NCI-H929 cells in vitro by H0L0 and parental 6E11.

Example 32 - Stability in Serum

To investigate the stability of H0L0 in human serum, a 500ml aliquot of antibody at 100µg/mL was incubated for periods up to 12 days in human 100% serum at -20°C, 4°C and 37°C. The activity after 12 days was determined by direct binding ELISA. EC-50 values were calculated and compared to historical EC-50 values from a number of previous binding ELISAs. The results displayed below in Figure 42 confirm that H0L0 shows no drop off in activity when incubated in serum for a period of up to 12 days at -20°C, 4°C or 37°C.

Example 33 - Pharmacodynamics of receptor down-regulation in xenograft model

In order to confirm that H0L0 can down-regulate IGF-1R receptor in vivo, an in vivo assay based on the LISN/3T3 c4 cell line is currently being developed based on the findings of others (Cohen et al. (2005) Clin Cancer Res., 11(5):2063-73). In the first study, athymic CD1 nu/nu mice mice were implanted with $2.5x10^6$ 3T3/LISN cells in matrigel (Becton Dickinson). When tumours reached a size of 400-500mm^3, mice were dosed with 125ug of either control antibody or H0L0. Mice were culled at T= 16, 24, 48, 72 or 120 hours after dosing. Tumours were excised and immediately frozen in liquid nitrogen. Weighed tumour samples were homogenised in RIPA buffer plus protease and phosphatase inhibitors and a protein assay performed on the lysate after centrifugation. A DELFIA assay (see Example 34) was performed to assess the relative levels of total IGF-1R in the tumour samples. As shown in Figure 43A, treatment with H0L0 appears to have some impact on total receptor in the 24-72 hour timeframe although the magnitude of the effect is substantially less than has been previously reported (Cohen, 2005)

In a second study, groups of mice (n=6) were implanted as above. When tumours reached a size of 400-500mm^3 mice were dosed twice 72 hours apart with 250ug of either control antibody or H0L0. 24 hours after the final dose, 10µg human recombinant IGF-1 was administered intravenously. After 10 mins, tumours were excised and frozen in liquid nitrogen. Total IGF-1R DELFIA assay was performed as described in Example 34. Although the animals were treated with IGF-I, no consistent changes in the phosphorylated receptor levels were observed compared to the untreated control group. However, groups treated with anti-IGF-1R antibodies (6E11, H0L0) showed a reduction in total receptor levels (Figure 43B). A similar effect was seen in the terminal tumour samples from the efficacy study (data not shown). A combined study which investigated the effects on receptor phosphorylation and total receptor levels over time proved inconclusive (data not shown,).

Example 34 - Total IGF-1R DELFIA assay

100 µl of tumour lysate containing 10 or 25µg of protein was loaded onto ELISA plates previously coated with an anti IGF-1R capture antibody 2B9 (see example 13). Plates were incubated overnight at 4°C and then washed in TBST. 100µl of polyclonal anti IGF-1R biotinylated antibody (R&D Systems BAF391) at 400ng/ml in

4%BSA/TBS was added to each well and incubated for 1 hour at room temperature. Plates were washed in TBST and 100µl of Eu labeled Streptavidin (Perkin Elmer 1244-360) at 1/1000 dilution was added to each well and incubated for 1 hour at room temperature. Plates were washed in TBST and 100µl of DELFIA Enhancement solution (Perkin Elmer 1244-105) added to each well and incubated for 10mins. A time resolved fluorescence signal was measured using a Wallac Victor multilabel plate reader.

SEQUENCE LISTING

Polynucleotide or amino acid sequence:	Sequence identifier (SEQ.I.D.NO)
6E11 VH CDR3	1
6E11 VH CDR2	2
6E11 VH CDR1	3
6E11 VL CDR1	4
6E11 VL CDR2	5
6E11 VL CDR3	6
9C7 VL CDR2	7
6E11 VH variable domain	8
6E11 VL variable domain	9
2B9 VH variable domain	10
2B9 VL variable domain	11
6E11 chimera VH variable domain	12
6E11 chimera VL variable domain	13
H0 variable domain	14
H1 variable domain	15
L0 variable domain	16
Biotinylated Tag sequence	17
9C7 VH variable domain	18
9C7 VL variable domain	19
5G4 VH variable domain	20
5G4 VL variable domain	21
15D9 VH variable domain	22
15D9 VL variable domain	23
6E11 chimera Heavy chain	24
6E11 chimera Light chain	25
6E11 VH variable domain (polynucleotide sequence)	26
6E11 VL variable domain (polynucleotide sequence)	27
9C7 VH variable domain (polynucleotide sequence)	28

9C7 VL variable domain (polynucleotide sequence)	29
6E11 chimera VH variable domain (polynucleotide sequence)	30
6E11 chimera VL variable domain (polynucleotide sequence)	31
6E11 chimera Heavy chain (polynucleotide sequence)	32
6E11 chimera Light chain (polynucleotide sequence)	33
H0 variable domain (polynucleotide sequence)	34
H1 variable domain (polynucleotide sequence)	35
L0 variable domain (polynucleotide sequence)	36
H0 Heavy chain	37
H1 Heavy chain	38
L0 light chain	39
H0 Heavy chain (polynucleotide sequence)	40
H1 Heavy chain (polynucleotide sequence)	41
L0 Light chain (polynucleotide sequence)	42
Campath leader	43
Human IGF-1R	44
Cynomolgus macaque IGF-1R	45
Mouse IGF-1R	46
Human IGF-1R-Fc fusion	47
Cynomolgus macaque IGF-1R-Fc fusion	48
IGF-1	49
Tagged IGF-1	50
IGF-2	51
Tagged IGF-2	52
Human Insulin receptor type B	53
H0 IgG1m(AA) Heavy chain	54
H0 IgG1m(AA) Heavy chain (polynucleotide sequence)	55
H1 IgG1m(AA) Heavy chain	56
H1 IgG1m(AA) Heavy chain (polynucleotide	57

sequence) Alternative L0 light chain (polynucleotide sequence)	58
Human Acceptor Framework Sequence-VH region	59
Human Acceptor Framework Sequence-VL region	60
H0 humanised variable domain (polynucleotide sequence)	61
L0 humanised variable domain (polynucleotide sequence)	62
Heavy chain constant region (S239D, I332E) (polynucleotide sequence)	63
Heavy chain constant region (S239D, I332E)	64
Heavy chain constant region (S239D, I332E, A330L) (polynucleotide sequence)	65
Heavy chain constant region (S239D, I332E, A330L) enhanced region.	66
H0 heavy chain (S239D, I332E) (polynucleotide sequence)	67
H0 heavy chain (S239D, I332E)	68
Alternative L0 light chain (polynucleotide sequence)	69
Alternative H0 heavy chain (polynucleotide sequence)	70

Sequence listing

SEQ ID 1:

WILYYGRSKWYFDV

SEQ ID 2:

NINPNNGGTNYNQKFKD

SEQ ID 3:

DYYMN

SEQ ID 4:

RSSQSIVQSNGDTYLE

SEQ ID 5:

RISNRFS

SEQ ID 6:

FQGSHVPYT

SEQ ID 7:

RVSNRFS

SEQ ID 8:

EVQLQQSGPELVKPGASVRISCKASGYAFTDYYMNWVKQSHGKSLEWVANINPNN
GGTNYNQKFKDKATLTVDKSSNTAYMELRSLTSEDTAVYYCARWILYYGRSKWYF
DVWGTGTTVTVSS

SEQ ID 9:

DVLMTQTPLSLPVSLGDHASISCRSSQSIVQSNGDTYLEWYLQKPGQSPKLLIYRIS
NRFSGVPDRFSGSGSGTDFTLKISRVEAEDLGVYYCFQGSHVPYTFGGGTKLEIKR
A

SEQ ID 10:

QVQLKQSGPGLVQSSQSLSITCTISGFSLTSHGIYWLRQSPGKGLEWLGVIWSGGS
ADYNAAFISRLSISKDNSKSQVFFKMNSLQADDTAIYYCARSPYYYRSSLYAMDYW
GQGTSVTVSS

SEQ ID 11:

NIVLTQSPKSMSMSIGERVTLSCKASENVGTYVSWYQQKAEQSPKLLIYGASNRHT
GVPDRFTGSGSSTDFTLTISSVQAEDLADYHCGQSYSDPLTFGAGTKLELKRA

SEQ ID 12:

EVQLQQSGPELVKPGASVRISCKASGYAFTDYYMNWVKQSHGKSLEWVANINPNN
GGTNYNQKFKDKATLTVDKSSNTAYMELRSLTSEDTAVYYCARWILYYGRSKWYF
DVWGTGTLVTVSS

SEQ ID 13:

DVLMTQTPLSLPVSLGDHASISCRSSQSIVQSNGDTYLEWYLQKPGQSPKLLIYRIS
NRFSGVPDRFSGSGSGTDFTLKISRVEAEDLGVYYCFQGSHVPYTFGGGTKLEIKR
T

SEQ ID 14:

QVQLVQSGAEVKKPGASVKVSCKASGYTFTDYYMNWVRQAPGQGLEWMGNINPN
NGGTNYNQKFKDRVTMTTDTSTSTAYMELRSLRSDDTAVYYCARWILYYGRSKWY
FDVWGRGTLVTVSS

SEQ ID 15:

QVQLVQSGAEVKKPGASVKVSCKASGYAFTDYYMNWVRQAPGQGLEWMGNINP
NNGGTNYNQKFKDRVTMTTDTSTSTAYMELRSLRSDDTAVYYCARWILYYGRSKW
YFDVWGRGTLVTVSS

SEQ ID 16:

DIVMTQSPLSLPVTPGEPASISCRSSQSIVQSNGDTYLEWYLQKPGQSPQLLIYRVS
NRFSGVPDRFSGSGSGTDFTLKISRVEAEDVGVYYCFQGSHVPYTFGQGTKLEIKR
T

SEQ ID 17:

GLNDIFEAQKIEWHE

SEQ ID 18:

EVQLQQSGPELVKPGASVRISCKASGYAFTDYYMNWVKQSHGKSLEWMANINPNN
GGTNYNQKFKDKATLTVDKSSNTAYMELRSLTSEDSAVYYCARWILYYGRSKWYF
DVWGPGTTVTVSS

SEQ ID 19:

DVLMTQSPLSLPVSLGDHASISCRSSQSIVQSNGDTYLEWYLQKPGQSPKLLIYRVS
NRFSGVPDRFSGSGSGTDFTLKISRVEAEDLGVYYCFQGSHVPYTFGGGTKLEIKR
A

SEQ ID 20:

EVQLQQSGPELVKPGASVKISCKASGYAFTDYYMNWVKQTHGRSLEWMANINPNT
GGTNYNQKFRGKATLTVDKSSTTAYMELRSLTSEDSAVYYCARWILYYGSSRWYF
DVWGTGTTVTVSS

SEQ ID 21:

DVLMTQTPLSLPVSLGDQASISCRSSQTIVHSNGNTYLEWYLQKPGQSPKLLIYKVS

NRFSGVPDRFSGSGSGTDFTLKISRVEAEDLGIYYCFQGSHVPYTFGGGTKLEIKRA

SEQ ID 22:

EVQLQQSGPELVKPGASVKISCKASGYAFTDYYMNWVKQSHGKSLEWMANINPNT
GGTNYNQKFTGKATLTVDKSSTTAYMELRSLTSEDSAVYYCTRWILYYGSSKWYFD
VWGTGTTVTVSS

SEQ ID 23:

DVLMTQTPLSLPVSLGDQASISCRSSQTIVHSNGNTYLEWYLQKPGQSPKLLIYRVS
YRFSGVPDRFSGSGSGTDFTLKISRLEAEDLGIYYCFQGSHVPYTFGGGTKLEIKRA

SEQ ID 24:

MGWSWIFFFLLSETAGVLSEVQLQQSGPELVKPGASVRISCKASGYAFTDYYMNW
VKQSHGKSLEWVANINPNNGGTNYNQKFKDKATLTVDKSSNTAYMELRSLTSEDT
AVYYCARWILYYGRSKWYFDVWGTGTLVTVSSASTKGPSVFPLAPSSKSTSGGTA
ALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGTQ
TYICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMI
SRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTV
LHQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQPREPQVYTLPPSRDELTKNQVS
LTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQQ
GNVFSCSVMHEALHNHYTQKSLSLSPGK

SEQ ID 25:

MKLPVRLVVLMFWIPASSSDVLMTQTPLSLPVSLGDHASISCRSSQSIVQSNGDTYL
EWYLQKPGQSPKLLIYRISNRFSGVPDRFSGSGSGTDFTLKISRVEAEDLGVYYCFQ
GSHVPYTFGGGTKLEIKRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAKVQ
WKVDNALQSGNSQESVTEQDSKDSTYSLSSTLTLSKADYEKHKVYACEVTHQGLS
SPVTKSFNRGEC

SEQ ID 26:

GAGGTCCAGCTGCAACAATCTGGACCTGAGCTGGTGAAGCCTGGGGCTTCAGT

GAGGATATCCTGTAAGGCTTCTGGATACGCGTTCACTGACTACTACATGAACTG
GGTGAAGCAGAGCCATGGAAAGAGCCTTGAGTGGGTGGCAAATATTAATCCCA
ACAATGGTGGTACTAACTACAACCAGAAGTTCAAGGACAAGGCCACATTGACTG
TAGACAAGTCCTCCAACACAGCCTACATGGAGCTCCGCAGTCTGACATCTGAGG
ACACTGCAGTCTATTACTGTGCAAGATGGATTCTTTACTACGGTCGTAGCAAATG
GTACTTCGATGTCTGGGGCACAGGGACCACGGTCACCGTCTCCTCG

SEQ ID 27:

GATGTTTTGATGACCCAAACTCCACTCTCCCTGCCTGTCAGTCTTGGAGATCAC
GCCTCCATCTCTTGCAGATCTAGTCAGAGTATTGTTCAAAGTAATGGAGACACCT
ATTTAGAATGGTACCTGCAGAAACCAGGCCAGTCTCCAAAGCTCCTGATCTACA
GAATTTCCAACCGATTTTCTGGGGTCCCAGACAGGTTCAGTGGCAGTGGATCAG
GGACAGATTTCACACTCAAGATCAGTAGAGTGGAGGCTGAGGATCTGGGAGTTT
ATTACTGCTTTCAGGGTTCACATGTTCCGTACACGTTCGGAGGGGGGACCAAGC
TGGAAATAAAACGGGCT

SEQ ID 28:

GAGGTCCAGCTGCAACAATCTGGACCTGAGCTGGTGAAGCCTGGGGCTTCAGT
GAGGATATCCTGTAAGGCTTCTGGATACGCGTTCACTGACTACTACATGAACTG
GGTGAAACAGAGCCATGGAAAGAGCCTTGAGTGGATGGCAAATATTAATCCCAA
CAATGGTGGTACTAACTACAACCAGAAGTTCAAGGACAAGGCCACATTGACTGT
AGACAAGTCCTCCAACACAGCCTACATGGAGCTCCGCAGTCTGACATCTGAGGA
CTCTGCAGTCTATTACTGTGCAAGATGGATTCTTTACTACGGTCGTAGCAAGTG
GTACTTCGATGTCTGGGGCCCAGGGACCACGGTCACCGTCTCCTCG

SEQ ID 29:

GATGTTTTGATGACCCAAAGTCCACTCTCCCTGCCTGTCAGTCTTGGAGATCAC
GCCTCCATCTCTTGCAGATCTAGTCAGAGCATTGTTCAAAGTAATGGAGACACC
TATTTAGAATGGTACCTGCAGAAACCAGGCCAGTCTCCAAAGCTCCTGATCTATA
GAGTTTCCAACCGATTTTCTGGGGTCCCAGACAGGTTCAGTGGCAGTGGATCAG
GGACAGATTTCACACTCAAGATCAGTAGAGTGGAGGCTGAGGATCTGGGAGTTT
ATTACTGCTTTCAGGGTTCACATGTTCCGTACACGTTCGGAGGGGGGACCAAGC
TGGAAATAAAACGGGCT

SEQ ID 30:

GAGGTCCAGCTGCAACAATCTGGACCTGAGCTGGTGAAGCCTGGGGCTTCAGT
GAGGATATCCTGTAAGGCTTCTGGATACGCGTTCACTGACTACTACATGAACTG
GGTGAAGCAGAGCCATGGAAAGAGCCTTGAGTGGGTGGCAAATATTAATCCCA
ACAATGGTGGTACTAACTACAACCAGAAGTTCAAGGACAAGGCCACATTGACTG
TAGACAAGTCCTCCAACACAGCCTACATGGAGCTCCGCAGTCTGACATCTGAGG
ACACTGCAGTCTATTACTGTGCAAGATGGATTCTTTACTACGGTCGTAGCAAATG
GTACTTCGATGTCTGGGGCACAGGGACACTAGTCACAGTCTCCTCA

SEQ ID 31:

GATGTTTTGATGACCCAAACTCCACTCTCCCTGCCTGTCAGTCTTGGAGATCAC
GCCTCCATCTCTTGCAGATCTAGTCAGAGTATTGTTCAAAGTAATGGAGACACCT
ATTTAGAATGGTACCTGCAGAAACCAGGCCAGTCTCCAAAGCTCCTGATCTACA
GAATTTCCAACCGATTTTCTGGGGTCCCAGACAGGTTCAGTGGCAGTGGATCAG
GGACAGATTTCACACTCAAGATCAGTAGAGTGGAGGCTGAGGATCTGGGAGTTT
ATTACTGCTTTCAGGGTTCACATGTTCCGTACACGTTCGGAGGGGGGACCAAGC
TGGAAATAAAACGTACG

SEQ ID 32:

ATGGGATGGAGCTGGATCTTTTTCTTCCTCCTGTCAGAAACTGCAGGTGTCCTC
TCTGAGGTCCAGCTGCAACAATCTGGACCTGAGCTGGTGAAGCCTGGGGCTTC
AGTGAGGATATCCTGTAAGGCTTCTGGATACGCGTTCACTGACTACTACATGAA
CTGGGTGAAGCAGAGCCATGGAAAGAGCCTTGAGTGGGTGGCAAATATTAATC
CCAACAATGGTGGTACTAACTACAACCAGAAGTTCAAGGACAAGGCCACATTGA
CTGTAGACAAGTCCTCCAACACAGCCTACATGGAGCTCCGCAGTCTGACATCTG
AGGACACTGCAGTCTATTACTGTGCAAGATGGATTCTTTACTACGGTCGTAGCA
AATGGTACTTCGATGTCTGGGGCACAGGGACACTAGTCACAGTCTCCTCAGCCT
CCACCAAGGGCCCATCGGTCTTCCCCCTGGCACCCTCCTCCAAGAGCACCTCT
GGGGGCACAGCGGCCCTGGGCTGCCTGGTCAAGGACTACTTCCCCGAACCGG
TGACGGTGTCGTGGAACTCAGGCGCCCTGACCAGCGGCGTGCACACCTTCCCG
GCTGTCCTACAGTCCTCAGGACTCTACTCCCTCAGCAGCGTGGTGACCGTGCC
CTCCAGCAGCTTGGGCACCCAGACCTACATCTGCAACGTGAATCACAAGCCCA

GCAACACCAAGGTGGACAAGAAAGTTGAGCCCAAATCTTGTGACAAAACTCACA
CATGCCCACCGTGCCCAGCACCTGAACTCCTGGGGGGACCGTCAGTCTTCCTC
TTCCCCCCAAAACCCAAGGACACCCTCATGATCTCCCGGACCCCTGAGGTCACA
TGCGTGGTGGTGGACGTGAGCCACGAAGACCCTGAGGTCAAGTTCAACTGGTA
CGTGGACGGCGTGGAGGTGCATAATGCCAAGACAAAGCCGCGGGAGGAGCAG
TACAACAGCACGTACCGTGTGGTCAGCGTCCTCACCGTCCTGCACCAGGACTG
GCTGAATGGCAAGGAGTACAAGTGCAAGGTCTCCAACAAAGCCCTCCCAGCCC
CCATCGAGAAAACCATCTCCAAAGCCAAAGGGCAGCCCCGAGAACCACAGGTG
TACACCCTGCCCCCATCCCGGGATGAGCTGACCAAGAACCAGGTCAGCCTGAC
CTGCCTGGTCAAAGGCTTCTATCCCAGCGACATCGCCGTGGAGTGGGAGAGCA
ATGGGCAGCCGGAGAACAACTACAAGACCACGCCTCCCGTGCTGGACTCCGAC
GGCTCCTTCTTCCTCTACAGCAAGCTCACCGTGGACAAGAGCAGGTGGCAGCA
GGGGAACGTCTTCTCATGCTCCGTGATGCATGAGGCTCTGCACAACCACTACAC
GCAGAAGAGCCTCTCCCTGTCTCCGGGTAAATGA

SEQ ID 33:

ATGAAGTTGCCTGTTCGGCTCGTGGTGCTGATGTTCTGGATTCCTGCTTCCAGC
AGTGATGTTTTGATGACCCAAACTCCACTCTCCCTGCCTGTCAGTCTTGGAGAT
CACGCCTCCATCTCTTGCAGATCTAGTCAGAGTATTGTTCAAAGTAATGGAGACA
CCTATTTAGAATGGTACCTGCAGAAACCAGGCCAGTCTCCAAAGCTCCTGATCT
ACAGAATTTCCAACCGATTTTCTGGGGTCCCAGACAGGTTCAGTGGCAGTGGAT
CAGGGACAGATTTCACACTCAAGATCAGTAGAGTGGAGGCTGAGGATCTGGGA
GTTTATTACTGCTTTCAGGGTTCACATGTTCCGTACACGTTCGGAGGGGGGACC
AAGCTGGAAATAAAACGTACGGTGGCTGCACCATCTGTCTTCATCTTCCCGCCA
TCTGATGAGCAGTTGAAATCTGGAACTGCCTCTGTTGTGTGCCTGCTGAATAAC
TTCTATCCCAGAGAGGCCAAAGTACAGTGGAAGGTGGACAACGCCCTCCAATC
GGGTAACTCCCAGGAGAGTGTCACAGAGCAGGACAGCAAGGACAGCACCTACA
GCCTCAGCAGCACCCTGACGCTGAGCAAAGCAGACTACGAGAAACACAAAGTC
TACGCCTGCGAAGTCACCCATCAGGGCCTGAGCTCGCCCGTCACAAAGAGCTT
CAACAGGGGAGAGTGTTAG

SEQ ID 34:

CAGGTCCAGCTGGTGCAGAGCGGCGCAGAGGTGAAGAAGCCCGGAGCTAGCG
TCAAGGTCTCCTGCAAGGCTTCAGGCTACACATTCACCGACTACTACATGAACT

GGGTGAGACAGGCTCCAGGACAGGGCCTCGAGTGGATGGGCAACATCAACCC
CAACAATGGCGGGACAAACTACAACCAGAAGTTCAAGGATCGCGTGACCATGA
CCACCGACACTAGCACCTCAACAGCCTACATGGAGCTGAGGTCTCTGCGGAGC
GATGACACTGCCGTGTACTACTGTGCCAGGTGGATTCTGTACTACGGGAGGAG
CAAGTGGTACTTCGACGTCTGGGGAAGAGGGACACTAGTGACCGTGAGCAGC

SEQ ID 35:

CAGGTCCAGCTGGTGCAGAGCGGCGCAGAGGTGAAGAAGCCCGGAGCTAGCG
TCAAGGTCTCCTGCAAGGCTTCAGGCTACGCCTTCACCGACTACTACATGAACT
GGGTGAGACAGGCTCCAGGACAGGGCCTCGAGTGGATGGGCAACATCAACCC
CAACAATGGCGGGACAAACTACAACCAGAAGTTCAAGGATCGCGTGACCATGA
CCACCGACACTAGCACCTCAACAGCCTACATGGAGCTGAGGTCTCTGCGGAGC
GATGACACTGCCGTGTACTACTGTGCCAGGTGGATTCTGTACTACGGGAGGAG
CAAGTGGTACTTCGACGTCTGGGGAAGAGGGACACTAGTGACCGTGAGCAGC

SEQ ID 36:

GACATCGTCATGACCCAGAGCCCACTGTCACTCCCCGTGACACCCGGAGAGCC
CGCTAGCATCAGCTGTAGAAGCTCCCAGAGCATCGTGCAGTCTAACGGCGATA
CCTACCTCGAGTGGTACCTGCAGAAGCCCGGACAGTCTCCTCAGCTCCTGATTT
ACCGCGTCAGCAATCGCTTTTCCGGGGTGCCTGATCGGTTTAGCGGCTCAGGA
AGCGGAACCGACTTCACCCTGAAGATCTCAAGGGTGGAGGCTGAGGATGTGGG
CGTGTACTACTGCTTCCAGGGATCTCACGTGCCTTACACCTTCGGACAGGGCAC
AAAGCTCGAGATTAAGCGTACG

SEQ ID 37:

MGWSCIILFLVATATGVHSQVQLVQSGAEVKKPGASVKVSCKASGYTFTDYYMNW
VRQAPGQGLEWMGNINPNNGGTNYNQKFKDRVTMTTDTSTSTAYMELRSLRSDD
TAVYYCARWILYYGRSKWYFDVWGRGTLVTVSSASTKGPSVFPLAPSSKSTSGGT
AALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGT
QTYICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPELLGGPSVFLFPPKPKDTL
MISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVL
TVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQPREPQVYTLPPSRDELTKNQ
VSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQ

QGNVFSCSVMHEALHNHYTQKSLSLSPGK

SEQ ID 38:

MGWSCIILFLVATATGVHSQVQLVQSGAEVKKPGASVKVSCKASGYAFTDYYMNW
VRQAPGQGLEWMGNINPNNGGTNYNQKFKDRVTMTTDTSTSTAYMELRSLRSDD
TAVYYCARWILYYGRSKWYFDVWGRGTLVTVSSASTKGPSVFPLAPSSKSTSGGT
AALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGT
QTYICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPELLGGPSVFLFPPKPKDTL
MISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVL
TVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQPREPQVYTLPPSRDELTKNQ
VSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQ
QGNVFSCSVMHEALHNHYTQKSLSLSPGK

SEQ ID 39:

MGWSCIILFLVATATGVHSDIVMTQSPLSLPVTPGEPASISCRSSQSIVQSNGDTYLE
WYLQKPGQSPQLLIYRVSNRFSGVPDRFSGSGSGTDFTLKISRVEAEDVGVYYCFQ
GSHVPYTFGQGTKLEIKRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAKVQ
WKVDNALQSGNSQESVTEQDSKDSTYSLSSTLTLSKADYEKHKVYACEVTHQGLS
SPVTKSFNRGEC

SEQ ID 40:

ATGGGATGGTCCTGTATCATCCTGTTTCTGGTGGCCACAGCAACTGGCGTGCAC
TCTCAGGTCCAGCTGGTGCAGAGCGGCGCAGAGGTGAAGAAGCCCGGAGCTA
GCGTCAAGGTCTCCTGCAAGGCTTCAGGCTACACATTCACCGACTACTACATGA
ACTGGGTGAGACAGGCTCCAGGACAGGGCCTCGAGTGGATGGGCAACATCAAC
CCCAACAATGGCGGGACAAACTACAACCAGAAGTTCAAGGATCGCGTGACCAT
GACCACCGACACTAGCACCTCAACAGCCTACATGGAGCTGAGGTCTCTGCGGA
GCGATGACACTGCCGTGTACTACTGTGCCAGGTGGATTCTGTACTACGGGAGG
AGCAAGTGGTACTTCGACGTCTGGGGAAGAGGGACACTAGTGACCGTGTCCAG
CGCCAGCACCAAGGGCCCCAGCGTGTTCCCCCTGGCCCCCAGCAGCAAGAGC
ACCAGCGGCGGCACAGCCGCCCTGGGCTGCCTGGTGAAGGACTACTTCCCCG
AACCGGTGACCGTGTCCTGGAACAGCGGAGCCCTGACCAGCGGCGTGCACAC
CTTCCCCGCCGTGCTGCAGAGCAGCGGCCTGTACAGCCTGAGCAGCGTGGTG
ACCGTGCCCAGCAGCAGCCTGGGCACCCAGACCTACATCTGTAACGTGAACCA

CAAGCCCAGCAACACCAAGGTGGACAAGAAGGTGGAGCCCAAGAGCTGTGACA
AGACCCACACCTGCCCCCCCTGCCCTGCCCCCGAGCTGCTGGGAGGCCCCAG
CGTGTTCCTGTTCCCCCCCAAGCCTAAGGACACCCTGATGATCAGCAGAACCCC
CGAGGTGACCTGTGTGGTGGTGGATGTGAGCCACGAGGACCCTGAGGTGAAGT
TCAACTGGTACGTGGACGGCGTGGAGGTGCACAATGCCAAGACCAAGCCCAGG
GAGGAGCAGTACAACAGCACCTACCGGGTGGTGTCCGTGCTGACCGTGCTGCA
CCAGGATTGGCTGAACGGCAAGGAGTACAAGTGTAAGGTGTCCAACAAGGCCC
TGCCTGCCCCTATCGAGAAAACCATCAGCAAGGCCAAGGGCCAGCCCAGAGAG
CCCCAGGTGTACACCCTGCCCCCTAGCAGAGATGAGCTGACCAAGAACCAGGT
GTCCCTGACCTGCCTGGTGAAGGGCTTCTACCCCAGCGACATCGCCGTGGAGT
GGGAGAGCAACGGCCAGCCCGAGAACAACTACAAGACCACCCCCCCTGTGCTG
GACAGCGATGGCAGCTTCTTCCTGTACAGCAAGCTGACCGTGGACAAGAGCAG
ATGGCAGCAGGGCAACGTGTTCAGCTGCTCCGTGATGCACGAGGCCCTGCACA
ATCACTACACCCAGAAGAGCCTGAGCCTGTCCCCTGGCAAGTGA

SEQ ID 41:

ATGGGATGGTCCTGTATCATCCTGTTTCTGGTGGCCACAGCAACTGGCGTGCAC
TCTCAGGTCCAGCTGGTGCAGAGCGGCGCAGAGGTGAAGAAGCCCGGAGCTA
GCGTCAAGGTCTCCTGCAAGGCTTCAGGCTACGCCTTCACCGACTACTACATGA
ACTGGGTGAGACAGGCTCCAGGACAGGGCCTCGAGTGGATGGGCAACATCAAC
CCCAACAATGGCGGGACAAACTACAACCAGAAGTTCAAGGATCGCGTGACCAT
GACCACCGACACTAGCACCTCAACAGCCTACATGGAGCTGAGGTCTCTGCGGA
GCGATGACACTGCCGTGTACTACTGTGCCAGGTGGATTCTGTACTACGGGAGG
AGCAAGTGGTACTTCGACGTCTGGGGAAGAGGGACACTAGTGACCGTGTCCAG
CGCCAGCACCAAGGGCCCCAGCGTGTTCCCCCTGGCCCCCAGCAGCAAGAGC
ACCAGCGGCGGCACAGCCGCCCTGGGCTGCCTGGTGAAGGACTACTTCCCCG
AACCGGTGACCGTGTCCTGGAACAGCGGAGCCCTGACCAGCGGCGTGCACAC
CTTCCCCGCCGTGCTGCAGAGCAGCGGCCTGTACAGCCTGAGCAGCGTGGTG
ACCGTGCCCAGCAGCAGCCTGGGCACCCAGACCTACATCTGTAACGTGAACCA
CAAGCCCAGCAACACCAAGGTGGACAAGAAGGTGGAGCCCAAGAGCTGTGACA
AGACCCACACCTGCCCCCCCTGCCCTGCCCCCGAGCTGCTGGGAGGCCCCAG
CGTGTTCCTGTTCCCCCCCAAGCCTAAGGACACCCTGATGATCAGCAGAACCCC
CGAGGTGACCTGTGTGGTGGTGGATGTGAGCCACGAGGACCCTGAGGTGAAGT
TCAACTGGTACGTGGACGGCGTGGAGGTGCACAATGCCAAGACCAAGCCCAGG
GAGGAGCAGTACAACAGCACCTACCGGGTGGTGTCCGTGCTGACCGTGCTGCA

CCAGGATTGGCTGAACGGCAAGGAGTACAAGTGTAAGGTGTCCAACAAGGCCC
TGCCTGCCCCTATCGAGAAAACCATCAGCAAGGCCAAGGGCCAGCCCAGAGAG
CCCCAGGTGTACACCCTGCCCCCTAGCAGAGATGAGCTGACCAAGAACCAGGT
GTCCCTGACCTGCCTGGTGAAGGGCTTCTACCCCAGCGACATCGCCGTGGAGT
GGGAGAGCAACGGCCAGCCCGAGAACAACTACAAGACCACCCCCCCTGTGCTG
GACAGCGATGGCAGCTTCTTCCTGTACAGCAAGCTGACCGTGGACAAGAGCAG
ATGGCAGCAGGGCAACGTGTTCAGCTGCTCCGTGATGCACGAGGCCCTGCACA
ATCACTACACCCAGAAGAGCCTGAGCCTGTCCCCTGGCAAGTGA

SEQ ID 42:

ATGGGATGGTCCTGCATCATCCTGTTCCTGGTGGCAACTGCCACTGGAGTCCAC
TCCGACATCGTCATGACCCAGAGCCCACTGTCACTCCCCGTGACACCCGGAGA
GCCCGCTAGCATCAGCTGTAGAAGCTCCCAGAGCATCGTGCAGTCTAACGGCG
ATACCTACCTCGAGTGGTACCTGCAGAAGCCCGGACAGTCTCCTCAGCTCCTGA
TTTACCGCGTCAGCAATCGCTTTTCCGGGGTGCCTGATCGGTTTAGCGGCTCAG
GAAGCGGAACCGACTTCACCCTGAAGATCTCAAGGGTGGAGGCTGAGGATGTG
GGCGTGTACTACTGCTTCCAGGGATCTCACGTGCCTTACACCTTCGGACAGGG
CACAAAGCTCGAGATTAAGCGTACGGTGGCCGCCCCCAGCGTGTTCATCTTCC
CCCCCAGCGATGAGCAGCTGAAGAGCGGCACCGCCAGCGTGGTGTGTCTGCT
GAACAACTTCTACCCCCGGGAGGCCAAGGTGCAGTGGAAGGTGGACAATGCCC
TGCAGAGCGGCAACAGCCAGGAGAGCGTGACCGAGCAGGACAGCAAGGACTC
CACCTACAGCCTGAGCAGCACCCTGACCCTGAGCAAGGCCGACTACGAGAAGC
ACAAGGTGTACGCCTGTGAGGTGACCCACCAGGGCCTGTCCAGCCCCGTGACC
AAGAGCTTCAACCGGGGCGAGTGCTGA

SEQ ID 43:

MGWSCIILFLVATATGVHS

SEQ ID 44:

MKSGSGGGSPTSLWGLLFLSAALSLWPTSGEICGPGIDIRNDYQQLKRLENCTVIEG
YLHILLISKAEDYRSYRFPKLTVITEYLLLFRVAGLESLGDLFPNLTVIRGWKLFYNYAL
VIFEMTNLKDIGLYNLRNITRGAIRIEKNADLCYLSTVDWSLILDAVSNNYIVGNKPPK
ECGDLCPGTMEEKPMCEKTTINNEYNYRCWTTNRCQKMCPSTCGKRACTENNEC
CHPECLGSCSAPDNDTACVACRHYYYAGVCVPACPPNTYRFEGWRCVDRDFCAN

ILSAESSDSEGFVIHDGECMQECPSGFIRNGSQSMYCIPCEGPCPKVCEEEKKTKTI
DSVTSAQMLQGCTIFKGNLLINIRRGNNIASELENFMGLIEVVTGYVKIRHSHALVSLS
FLKNLRLILGEEQLEGNYSFYVLDNQNLQQLWDWDHRNLTIKAGKMYFAFNPKLCV
SEIYRMEEVTGTKGRQSKGDINTRNNGERASCESDVLHFTSTTTSKNRIIITWHRYR
PPDYRDLISFTVYYKEAPFKNVTEYDGQDACGSNSWNMVDVDLPPNKDVEPGILLH
GLKPWTQYAVYVKAVTLTMVENDHIRGAKSEILYIRTNASVPSIPLDVLSASNSSSQL
IVKWNPPSLPNGNLSYYIVRWQRQPQDGYLYRHNYCSKDKIPIRKYADGTIDIEEVT
ENPKTEVCGGEKGPCCACPKTEAEKQAEKEEAEYRKVFENFLHNSIFVPRPERKRR
DVMQVANTTMSSRSRNTTAADTYNITDPEELETEYPFFESRVDNKERTVISNLRPFT
LYRIDIHSCNHEAEKLGCSASNFVFARTMPAEGADDIPGPVTWEPRPENSIFLKWPE
PENPNGLILMYEIKYGSQVEDQRECVSRQEYRKYGGAKLNRLNPGNYTARIQATSL
SGNGSWTDPVFFYVQAKTGYENFIHLIIALPVAVLLIVGGLVIMLYVFHRKRNNSRLG
NGVLYASVNPEYFSAADVYVPDEWEVAREKITMSRELGQGSFGMVYEGVAKGVVK
DEPETRVAIKTVNEAASMRERIEFLNEASVMKEFNCHHVVRLLGVVSQGQPTLVIME
LMTRGDLKSYLRSLRPEMENNPVLAPPSLSKMIQMAGEIADGMAYLNANKFVHRDL
AARNCMVAEDFTVKIGDFGMTRDIYETDYYRKGGKGLLPVRWMSPESLKDGVFTT
YSDVWSFGVVLWEIATLAEQPYQGLSNEQVLRFVMEGGLLDKPDNCPDMLFELMR
MCWQYNPKMRPSFLEIISSIKEEMEPGFREVSFYYSEENKLPEPEELDLEPENMES
VPLDPSASSSSLPLPDRHSGHKAENGPGPGVLVLRASFDERQPYAHMNGGRKNE
RALPLPQSSTC

SEQ ID 45:

MKSGSGGGSPTSLWGLLFLSAALSLWPTSGEICGPGIDIRNDYQQLKRLENCTVIEG
YLHILLISKAEDYRSYRFPKLTVITEYLLLFRVAGLESLGDLFPNLTVIRGWKLFYNYAL
VIFEMTNLKDIGLYNLRNITRGAIRIEKNADLCYLSTVDWSLILDAVSNNYIVGNKPPK
ECGDLCPGTMEEKPMCEKTTINNEYNYRCWTTNRCQKMCPSACGKRACTENNEC
CHPECLGSCSAPDNDTACVACRHYYYAGVCVPACPPNTYRFEGWRCVDRDFCAN
ILSAESSDSEGFVIHDGECMQECPSGFIRNGSQSMYCIPCEGPCPKVCEEEKKTKTI
DSVTSAQMLQGCTIFKGNLLINIRRGNNIASELENFMGLIEVVTGYVKIRHSHALVSLS
FLKNLRLILGEEQLEGNYSFYVLDNQNLQQLWDWDHRNLTIKAGKMYFAFNPKLCV
SEIYRMEEVTGTKGRQSKGDINTRNNGERASCESDVLHFTSTTTWKNRIIITWHRYR
PPDYRDLISFTVYYKEAPFKNVTEYDGQDACGSNSWNMVDVDLPPNKDVEPGILLH
GLKPWTQYAVYVKAVTLTMVENDHIRGAKSEILYIRTNASVPSIPLDVLSASNSSSQL
IVKWNPPSLPNGNLSYYIVRWQRQPQDGYLYRHNYCSKDKIPIRKYADGTIDIEEVT
ENPKTEVCGGEKGPCCACPKTEAEKQAEKEEAEYRKVFENFLHNSIFVPRPERKRR
DVMQVANTTMSSRSRNTTAADTYNITDLEELETEYPFFESRVDNKERTVISNLRPFT

LYRIDIHSCNHEAEKLGCSASNFVFARTMPAEGADDIPGPVTWEPRPENSIFLKWPE
PENPNGLILMYEIKYGSQVEDQRECVSRQEYRKYGGAKLNRLNPGNYTARIQATSL
SGNGSWTDPVFFYVQAKTGYENFIHLIIALPVAVLLIVGGLVIMLYVFHRKRNNSRLG
NGVLYASVNPEYFSAADVYVPDEWEVAREKITMSRELGQGSFGMVYEGVAKGVVK
DEPETRVAIKTVNEAASMRERIEFLNEASVMKEFNCHHVVRLLGVVSQGQPTLVIME
LMTRGDLKSYLRSLRPEMENNPVLAPPSLSKMIQMAGEIADGMAYLNANKFVHRDL
AARNCMVAEDFTVKIGDFGMTRDIYETDYYRKGGKGLLPVRWMSPESLKDGVFTT
YSDVWSFGVVLWEIATLAEQPYQGLSNEQVLRFVMEGGLLDKPDNCPDMLFELMR
MCWQYNPKMRPSFLEIISSIKDEMEPGFREVSFYYSEENKLPEPEELDLEPENMES
VPLDPSASSSSLPLPDRHSGHKAENGPGPGVLVLRASFDERQPYAHMNGGRKNE
RALPLPQSSTC

SEQ ID 46:

MKSGSGGGSPTSLWGLVFLSAALSLWPTSGEICGPGIDIRNDYQQLKRLENCTVIE
GFLHILLISKAEDYRSYRFPKLTVITEYLLLFRVAGLESLGDLFPNLTVIRGWKLFYNY
ALVIFEMTNLKDIGLYNLRNITRGAIRIEKNADLCYLSTIDWSLILDAVSNNYIVGNKPP
KECGDLCPGTLEEKPMCEKTTINNEYNYRCWTTNRCQKMCPSVCGKRACTENNE
CCHPECLGSCHTPDDNTTCVACRHYYYKGVCVPACPPGTYRFEGWRCVDRDFCA
NIPNAESSDSDGFVIHDDECMQECPSGFIRNSTQSMYCIPCEGPCPKVCGDEEKKT
KTIDSVTSAQMLQGCTILKGNLLINIRRGNNIASELENFMGLIEVVTGYVKIRHSHALV
SLSFLKNLRLILGEEQLEGNYSFYVLDNQNLQQLWDWNHRNLTVRSGKMYFAFNP
KLCVSEIYRMEEVTGTKGRQSKGDINTRNNGERASCESDVLRFTSTTTWKNRIIITW
HRYRPPDYRDLISFTVYYKEAPFKNVTEYDGQDACGSNSWNMVDVDLPPNKEGEP
GILLHGLKPWTQYAVYVKAVTLTMVENDHIRGAKSEILYIRTNASVPSIPLDVLSASN
SSSQLIVKWNPPTLPNGNLSYYIVRWQRQPQDGYLYRHNYCSKDKIPIRKYADGTID
VEEVTENPKTEVCGGDKGPCCACPKTEAEKQAEKEEAEYRKVFENFLHNSIFVPRP
ERRRRDVMQVANTTMSSRSRNTTVADTYNITDPEEFETEYPFFESRVDNKERTVIS
NLRPFTLYRIDIHSCNHEAEKLGCSASNFVFARTMPAEGADDIPGPVTWEPRPENSI
FLKWPEPENPNGLILMYEIKYGSQVEDQRECVSRQEYRKYGGAKLNRLNPGNYTA
RIQATSLSGNGSWTDPVFFYVPAKTTYENFMHLIIALPVAILLIVGGLVIMLYVFHRKR
NNSRLGNGVLYASVNPEYFSAADVYVPDEWEVAREKITMNRELGQGSFGMVYEGV
AKGVVKDEPETRVAIKTVNEAASMRERIEFLNEASVMKEFNCHHVVRLLGVVSQGQ
PTLVIMELMTRGDLKSYLRSLRPEVEQNNLVLIPPSLSKMIQMAGEIADGMAYLNAN
KFVHRDLAARNCMVAEDFTVKIGDFGMTRDIYETDYYRKGGKGLLPVRWMSPESL

KDGVFTTHSDVWSFGVVLWEIATLAEQPYQGLSNEQVLRFVMEGGLLDKPDNCPD
MLFELMRMCWQYNPKMRPSFLEIIGSIKDEMEPSFQEVSFYYSEENKPPEPEELEM
ELEMEPENMESVPLDPSASSASLPLPERHSGHKAENGPGPGVLVLRASFDERQPY
AHMNGGRANERALPLPQSSTC

SEQ ID 47:

MKSGSGGGSPTSLWGLLFLSAALSLWPTSGEICGPGIDIRNDYQQLKRLENCTVIEG
YLHILLISKAEDYRSYRFPKLTVITEYLLLFRVAGLESLGDLFPNLTVIRGWKLFYNYAL
VIFEMTNLKDIGLYNLRNITRGAIRIEKNADLCYLSTVDWSLILDAVSNNYIVGNKPPK
ECGDLCPGTMEEKPMCEKTTINNEYNYRCWTTNRCQKMCPSTCGKRACTENNEC
CHPECLGSCSAPDNDTACVACRHYYYAGVCVPACPPNTYRFEGWRCVDRDFCAN
ILSAESSDSEGFVIHDGECMQECPSGFIRNGSQSMYCIPCEGPCPKVCEEEKKTKTI
DSVTSAQMLQGCTIFKGNLLINIRRGNNIASELENFMGLIEVVTGYVKIRHSHALVSLS
FLKNLRLILGEEQLEGNYSFYVLDNQNLQQLWDWDHRNLTIKAGKMYFAFNPKLCV
SEIYRMEEVTGTKGRQSKGDINTRNNGERASCESDVLHFTSTTTSKNRIIITWHRYR
PPDYRDLISFTVYYKEAPFKNVTEYDGQDACGSNSWNMVDVDLPPNKDVEPGILLH
GLKPWTQYAVYVKAVTLTMVENDHIRGAKSEILYIRTNASVPSIPLDVLSASNSSSQL
IVKWNPPSLPNGNLSYYIVRWQRQPQDGYLYRHNYCSKDKIPIRKYADGTIDIEEVT
ENPKTEVCGGEKGPCCACPKTEAEKQAEKEEAEYRKVFENFLHNSIFVPRPERKRR
DVMQVANTTMSSRSRNTTAADTYNITDPEELETEYPFFESRVDNKERTVISNLRPFT
LYRIDIHSCNHEAEKLGCSASNFVFARTMPAEGADDIPGPVTWEPRPENSIFLKWPE
PENPNGLILMYEIKYGSQVEDQRECVSRQEYRKYGGAKLNRLNPGNYTARIQATSL
SGNGSWTDPVFFYVQAKTGYENFIHAAAIEGRSGSCDKTHTCPPCPAPELLGGPSV
FLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKTKPREEQY
NSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQPREPQVYTLPP
SRDELTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLYSK
LTVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGKLRRASLG

SEQ ID 48:

MKSGSGGGSPTSLWGLLFLSAALSLWPTSGEICGPGIDIRNDYQQLKRLENCTVIEG
YLHILLISKAEDYRSYRFPKLTVITEYLLLFRVAGLESLGDLFPNLTVIRGWKLFYNYAL
VIFEMTNLKDIGLYNLRNITRGAIRIEKNADLCYLSTVDWSLILDAVSNNYIVGNKPPK
ECGDLCPGTMEEKPMCEKTTINNEYNYRCWTTNRCQKMCPSACGKRACTENNEC

CHPECLGSCSAPDNDTACVACRHYYYAGVCVPACPPNTYRFEGWRCVDRDFCAN
ILSAESSDSEGFVIHDGECMQECPSGFIRNGSQSMYCIPCEGPCPKVCEEEKKTKTI
DSVTSAQMLQGCTIFKGNLLINIRRGNNIASELENFMGLIEVVTGYVKIRHSHALVSLS
FLKNLRLILGEEQLEGNYSFYVLDNQNLQQLWDWDHRNLTIKAGKMYFAFNPKLCV
SEIYRMEEVTGTKGRQSKGDINTRNNGERASCESDVLHFTSTTTWKNRIIITWHRYR
PPDYRDLISFTVYYKEAPFKNVTEYDGQDACGSNSWNMVDVDLPPNKDVEPGILLH
GLKPWTQYAVYVKAVTLTMVENDHIRGAKSEILYIRTNASVPSIPLDVLSASNSSSQL
IVKWNPPSLPNGNLSYYIVRWQRQPQDGYLYRHNYCSKDKIPIRKYADGTIDIEEVT
ENPKTEVCGGEKGPCCACPKTEAEKQAEKEEAEYRKVFENFLHNSIFVPRPERKRR
DVMQVANTTMSSRSRNTTAADTYNITDLEELETEYPFFESRVDNKERTVISNLRPFT
LYRIDIHSCNHEAEKLGCSASNFVFARTMPAEGADDIPGPVTWEPRPENSIFLKWPE
PENPNGLILMYEIKYGSQVEDQRECVSRQEYRKYGGAKLNRLNPGNYTARIQATSL
SGNGSWTDPVFFYVQAKTGYENFIHAAAIEGRSGSCDKTHTCPPCPAPELLGGPSV
FLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKTKPREEQY
NSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQPREPQVYTLPP
SRDELTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLYSK
LTVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGKLRRASLG

SEQ ID 49:

MGPETLCGAELVDALQFVCGDRGFYFNKPTGYGSSSRRAPQTGIVDECCFRSCDL
RRLEMYCAPLKPAKSA

SEQ ID 50:

MGPETLCGAELVDALQFVCGDRGFYFNKPTGYGSSSRRAPQTGIVDECCFRSCDL
RRLEMYCAPLKPAKSAGLNDIFEAQKIEWHE

SEQ ID 51:

MAYRPSETLCGGELVDTLQFVCGDRGFYFSRPASRVSRRSRGIVEECCFRSCDLA
LLETYCATPAKSE

SEQ ID 52:

MAYRPSETLCGGELVDTLQFVCGDRGFYFSRPASRVSRRSRGIVEECCFRSCDLA

LLETYCATPAKSEGLNDIFEAQKIEWHE

SEQ ID 53:

MGTGGRRGAAAAPLLVAVAALLLGAAGHLYPGEVCPGMDIRNNLTRLHELENCSVI
EGHLQILLMFKTRPEDFRDLSFPKLIMITDYLLLFRVYGLESLKDLFPNLTVIRGSRLF
FNYALVIFEMVHLKELGLYNLMNITRGSVRIEKNNELCYLATIDWSRILDSVEDNYIVL
NKDDNEECGDICPGTAKGKTNCPATVINGQFVERCWTHSHCQKVCPTICKSHGCT
AEGLCCHSECLGNCSQPDDPTKCVACRNFYLDGRCVETCPPPYYHFQDWRCVNF
SFCQDLHHKCKNSRRQGCHQYVIHNNKCIPECPSGYTMNSSNLLCTPCLGPCPKV
CHLLEGEKTIDSVTSAQELRGCTVINGSLIINIRGGNNLAAELEANLGLIEEISGYLKIR
RSYALVSLSFFRKLRLIRGETLEIGNYSFYALDNQNLRQLWDWSKHNLTITQGKLFF
HYNPKLCLSEIHKMEEVSGTKGRQERNDIALKTNGDQASCENELLKFSYIRTSFDKIL
LRWEPYWPPDFRDLLGFMLFYKEAPYQNVTEFDGQDACGSNSWTVVDIDPPLRSN
DPKSQNHPGWLMRGLKPWTQYAIFVKTLVTFSDERRTYGAKSDIIYVQTDATNPSV
PLDPISVSNSSSQIILKWKPPSDPNGNITHYLVFWERQAEDSELFELDYCLKGLKLPS
RTWSPPFESEDSQKHNQSEYEDSAGECCSCPKTDSQILKELEESSFRKTFEDYLHN
VVFVPRKTSSGTGAEDPRPSRKRRSLGDVGNVTVAVPTVAAFPNTSSTSVPTSPEE
HRPFEKVVNKESLVISGLRHFTGYRIELQACNQDTPEERCSVAAYVSARTMPEAKA
DDIVGPVTHEIFENNVVHLMWQEPKEPNGLIVLYEVSYRRYGDEELHLCVSRKHFAL
ERGCRLRGLSPGNYSVRIRATSLAGNGSWTEPTYFYVTDYLDVPSNIAKIIIGPLIFVF
LFSVVIGSIYLFLRKRQPDGPLGPLYASSNPEYLSASDVFPCSVYVPDEWEVSREKI
TLLRELGQGSFGMVYEGNARDIIKGEAETRVAVKTVNESASLRERIEFLNEASVMKG
FTCHHVVRLLGVVSKGQPTLVVMELMAHGDLKSYLRSLRPEAENNPGRPPPTLQE
MIQMAAEIADGMAYLNAKKFVHRDLAARNCMVAHDFTVKIGDFGMTRDIYETDYYR
KGGKGLLPVRWMAPESLKDGVFTTSSDMWSFGVVLWEITSLAEQPYQGLSNEQVL
KFVMDGGYLDQPDNCPERVTDLMRMCWQFNPNMRPTFLEIVNLLKDDLHPSFPEV
SFFHSEENKAPESEELEMEFEDMENVPLDRSSHCQREEAGGRDGGSSLGFKRSY
EEHIPYTHMNGGKKNGRILTLPRSNPS

SEQ ID 54:

MGWSCIILFLVATATGVHSQVQLVQSGAEVKKPGASVKVSCKASGYTFTDYYMNW
VRQAPGQGLEWMGNINPNNGGTNYNQKFKDRVTMTTDTSTSTAYMELRSLRSDD
TAVYYCARWILYYGRSKWYFDVWGRGTLVTVSSASTKGPSVFPLAPSSKSTSGGT

AALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGT
QTYICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPELAGAPSVFLFPPKPKDTL
MISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVL
TVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQPREPQVYTLPPSRDELTKNQ
VSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQ
QGNVFSCSVMHEALHNHYTQKSLSLSPGK

SEQ ID 55:

ATGGGATGGTCCTGTATCATCCTGTTTCTGGTGGCCACAGCAACTGGCGTGCAC
TCTCAGGTCCAGCTGGTGCAGAGCGGCGCAGAGGTGAAGAAGCCCGGAGCTA
GCGTCAAGGTCTCCTGCAAGGCTTCAGGCTACACATTCACCGACTACTACATGA
ACTGGGTGAGACAGGCTCCAGGACAGGGCCTCGAGTGGATGGGCAACATCAAC
CCCAACAATGGCGGGACAAACTACAACCAGAAGTTCAAGGATCGCGTGACCAT
GACCACCGACACTAGCACCTCAACAGCCTACATGGAGCTGAGGTCTCTGCGGA
GCGATGACACTGCCGTGTACTACTGTGCCAGGTGGATTCTGTACTACGGGAGG
AGCAAGTGGTACTTCGACGTCTGGGGAAGAGGGACACTAGTGACCGTGAGCAG
CGCCAGCACCAAGGGCCCCAGCGTGTTCCCCCTGGCCCCCAGCAGCAAGAGC
ACCAGCGGCGGCACAGCCGCCCTGGGCTGCCTGGTGAAGGACTACTTCCCCG
AGCCCGTGACCGTGTCCTGGAACAGCGGAGCCCTGACAAGCGGGGTGCACAC
CTTCCCCGCCGTGCTGCAGAGCAGCGGCCTGTACAGCCTGAGCAGCGTGGTG
ACAGTGCCCAGCAGCAGCCTGGGCACCCAGACCTACATCTGCAACGTGAACCA
CAAGCCCAGCAACACCAAGGTGGACAAGAAGGTGGAGCCCAAGAGCTGCGAC
AAGACCCACACCTGCCCCCCCTGCCCTGCCCCTGAACTGGCCGGAGCCCCCTC
CGTGTTCCTGTTCCCCCCCAAGCCCAAGGACACCCTGATGATCAGCCGGACCC
CCGAGGTGACCTGCGTGGTGGTGGACGTGAGCCACGAGGACCCTGAGGTGAA
GTTCAATTGGTACGTGGACGGCGTGGAGGTGCACAACGCCAAGACCAAGCCCC
GGGAGGAACAGTACAACAGCACCTACCGGGTGGTGTCCGTGCTGACCGTGCTG
CACCAGGACTGGCTGAACGGCAAAGAATACAAGTGCAAGGTGTCCAACAAGGC
CCTGCCTGCCCCCATCGAGAAAACCATCAGCAAGGCCAAGGGCCAGCCCAGG
GAACCCCAGGTGTACACCCTGCCCCCCTCCCGGGACGAGCTGACCAAGAACCA
GGTGTCCCTGACCTGTCTGGTGAAGGGCTTCTACCCCAGCGACATCGCCGTGG
AGTGGGAGAGCAACGGCCAGCCCGAGAACAACTACAAGACCACCCCCCCTGTG
CTGGACAGCGACGGCAGCTTCTTCCTGTACAGCAAGCTGACCGTGGACAAGAG
CCGGTGGCAGCAGGGCAACGTGTTCAGCTGCAGCGTGATGCACGAGGCCCTG

CACAACCACTACACCCAGAAGAGCCTGAGCCTGTCCCCCGGCAAGTGA

SEQ ID 56:

MGWSCIILFLVATATGVHSQVQLVQSGAEVKKPGASVKVSCKASGYAFTDYYMNW
VRQAPGQGLEWMGNINPNNGGTNYNQKFKDRVTMTTDTSTSTAYMELRSLRSDD
TAVYYCARWILYYGRSKWYFDVWGRGTLVTVSSASTKGPSVFPLAPSSKSTSGGT
AALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGT
QTYICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPELAGAPSVFLFPPKPKDTL
MISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVL
TVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQPREPQVYTLPPSRDELTKNQ
VSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQ
QGNVFSCSVMHEALHNHYTQKSLSLSPGK.

SEQ ID 57:

ATGGGATGGTCCTGTATCATCCTGTTTCTGGTGGCCACAGCAACTGGCGTGCAC
TCTCAGGTCCAGCTGGTGCAGAGCGGCGCAGAGGTGAAGAAGCCCGGAGCTA
GCGTCAAGGTCTCCTGCAAGGCTTCAGGCTACGCCTTCACCGACTACTACATGA
ACTGGGTGAGACAGGCTCCAGGACAGGGCCTCGAGTGGATGGGCAACATCAAC
CCCAACAATGGCGGGACAAACTACAACCAGAAGTTCAAGGATCGCGTGACCAT
GACCACCGACACTAGCACCTCAACAGCCTACATGGAGCTGAGGTCTCTGCGGA
GCGATGACACTGCCGTGTACTACTGTGCCAGGTGGATTCTGTACTACGGGAGG
AGCAAGTGGTACTTCGACGTCTGGGGAAGAGGGACACTAGTGACCGTGAGCAG
CGCCAGCACCAAGGGCCCCAGCGTGTTCCCCCTGGCCCCCAGCAGCAAGAGC
ACCAGCGGCGGCACAGCCGCCCTGGGCTGCCTGGTGAAGGACTACTTCCCCG
AGCCCGTGACCGTGTCCTGGAACAGCGGAGCCCTGACAAGCGGGGTGCACAC
CTTCCCCGCCGTGCTGCAGAGCAGCGGCCTGTACAGCCTGAGCAGCGTGGTG
ACAGTGCCCAGCAGCAGCCTGGGCACCCAGACCTACATCTGCAACGTGAACCA
CAAGCCCAGCAACACCAAGGTGGACAAGAAGGTGGAGCCCAAGAGCTGCGAC
AAGACCCACACCTGCCCCCCCTGCCCTGCCCCTGAACTGGCCGGAGCCCCCTC
CGTGTTCCTGTTCCCCCCCAAGCCCAAGGACACCCTGATGATCAGCCGGACCC
CCGAGGTGACCTGCGTGGTGGTGGACGTGAGCCACGAGGACCCTGAGGTGAA
GTTCAATTGGTACGTGGACGGCGTGGAGGTGCACAACGCCAAGACCAAGCCCC
GGGAGGAACAGTACAACAGCACCTACCGGGTGGTGTCCGTGCTGACCGTGCTG

CACCAGGACTGGCTGAACGGCAAAGAATACAAGTGCAAGGTGTCCAACAAGGC
CCTGCCTGCCCCCATCGAGAAAACCATCAGCAAGGCCAAGGGCCAGCCCAGG
GAACCCCAGGTGTACACCCTGCCCCCCTCCCGGGACGAGCTGACCAAGAACCA
GGTGTCCCTGACCTGTCTGGTGAAGGGCTTCTACCCCAGCGACATCGCCGTGG
AGTGGGAGAGCAACGGCCAGCCCGAGAACAACTACAAGACCACCCCCCCTGTG
CTGGACAGCGACGGCAGCTTCTTCCTGTACAGCAAGCTGACCGTGGACAAGAG
CCGGTGGCAGCAGGGCAACGTGTTCAGCTGCAGCGTGATGCACGAGGCCCTG
CACAACCACTACACCCAGAAGAGCCTGAGCCTGTCCCCCGGCAAGTGA

SEQ ID 58:

ATGGGATGGTCCTGCATCATCCTGTTCCTGGTGGCAACTGCCACTGGAGTCCAC
TCCGACATCGTCATGACCCAGAGCCCACTGTCACTCCCCGTGACACCCGGAGA
GCCCGCTAGCATCAGCTGTAGAAGCTCCCAGAGCATCGTGCAGTCTAACGGCG
ATACCTACCTCGAGTGGTACCTGCAGAAGCCCGGACAGTCTCCTCAGCTCCTGA
TTTACCGCGTCAGCAATCGCTTTTCCGGGGTGCCTGATCGGTTTAGCGGCTCAG
GAAGCGGAACCGACTTCACCCTGAAGATCTCAAGGGTGGAGGCTGAGGATGTG
GGCGTGTACTACTGCTTCCAGGGATCTCACGTGCCTTACACCTTCGGACAGGG
CACAAAGCTCGAGATTAAGCGTACGGTGGCCGCTCCCAGCGTGTTCATCTTCCC
CCCCAGCGACGAGCAGCTGAAGAGCGGCACCGCCAGCGTGGTGTGCCTGCTG
AACAACTTCTACCCCCGGGAGGCCAAGGTGCAGTGGAAGGTGGACAACGCCCT
GCAGAGCGGCAACAGCCAGGAAAGCGTCACCGAGCAGGACAGCAAGGACTCC
ACCTACAGCCTGAGCAGCACCCTGACACTGAGCAAGGCCGACTACGAGAAGCA
CAAGGTGTACGCCTGCGAGGTGACCCACCAGGGCCTGTCCAGCCCCGTGACC
AAGAGCTTCAACCGGGGCGAGTGCTAG

SEQ ID 59:

QVQLVQSGAEVKKPGASVKVSCKASGYTFTXaaXaaXaaXaaXaaWVRQAPGQGLE
WMGXaaXaaXaaXaaXaaXaaXaaXaaXaaXaaXaaXaaXaaXaaXaaXaaRVTMTTD
TSTSTAYMELRSLRSDDTAVYYCARXaaXaaXaaXaaXaaXaaXaaXaaXaaXaaXaaXa
aXaaXaaWGRGTLVTVSS

SEQ ID 60:

DIVMTQSPLSLPVTPGEPASISCXaaXaaXaaXaaXaaXaaXaaXaaXaaXaaXaaXaaXa
aXaaXaaXaaWYLQKPGQSPQLLIYXaaXaaXaaXaaXaaXaaXaaGVPDRFSGSGSGT
DFTLKISRVEAEDVGVYYCXaaXaaXaaXaaXaaXaaXaaXaaXaaFGQGTKLEIKRT

SEQ ID 61:

CAGGTGCAGCTGGTGCAGAGCGGAGCCGAGGTGAAGAAGCCTGGCGCCAGCG
TCAAGGTGTCCTGCAAGGCCAGCGGCTACACCTTCACCGACTACTACATGAACT
GGGTGCGGCAGGCCCCAGGCCAGGGACTGGAATGGATGGGCAACATCAACCC
CAACAACGGCGGCACCAACTACAACCAGAAGTTCAAGGACCGGGTCACCATGA
CCACCGACACCAGCACCAGCACCGCCTACATGGAACTGCGGAGCCTGAGAAGC
GACGACACCGCCGTGTACTACTGCGCCCGGTGGATCCTGTACTACGGCCGGTC
CAAGTGGTACTTCGACGTGTGGGGCAGGGGCACACTAGT

SEQ ID NO 62:

GACATCGTGATGACCCAGAGCCCCCTGAGCCTGCCCGTGACCCCTGGCGAGC
CCGCCAGCATCAGCTGCAGAAGCAGCCAGAGCATCGTCCAGAGCAACGGCGA
CACCTACCTGGAATGGTATCTGCAGAAGCCCGGCCAGTCCCCCCAGCTGCTGA
TCTACAGAGTGAGCAACCGGTTCAGCGGCGTGCCCGACAGATTCAGCGGCAGC
GGCTCCGGCACCGACTTCACCCTGAAGATCAGCCGGGTGGAGGCCGAGGACG
TGGGCGTGTACTACTGCTTTCAAGGCAGCCACGTGCCCTACACCTTCGGCCAG
GGCACCAAGCTGGAAATCAAGCGTACG

SEQ ID NO: 63

ACTAGTCACCGTGAGCAGCGCCAGCACCAAGGGCCCCAGCGTGTTCCCCCTGG
CCCCCAGCAGCAAGAGCACCAGCGGCGGCACAGCCGCCCTGGGCTGCCTGGT
GAAGGACTACTTCCCCGAGCCCGTGACCGTGAGCTGGAACAGCGGAGCCCTGA
CCTCCGGCGTGCACACCTTCCCCGCCGTGCTGCAGAGCAGCGGCCTGTACAG
CCTGAGCAGCGTGGTGACCGTGCCCAGCAGCAGCCTGGGCACCCAGACCTAC
ATCTGCAACGTGAACCACAAGCCCAGCAACACCAAGGTGGACAAGAAGGTGGA
GCCCAAGAGCTGCGACAAGACCCACACCTGCCCCCCCTGCCCTGCCCCTGAGC
TGCTGGGCGGACCCGACGTGTTCCTGTTCCCCCCCAAGCCCAAGGACACCCTG
ATGATCAGCCGGACCCCCGAGGTGACCTGCGTGGTGGTGGACGTGAGCCACG
AGGACCCTGAGGTGAAGTTCAATTGGTACGTGGACGGCGTGGAGGTGCACAAC

GCCAAGACCAAGCCCCGGGAGGAACAGTACAACAGCACCTACCGGGTGGTGTC
CGTGCTGACCGTGCTGCACCAGGACTGGCTGAACGGCAAAGAATACAAGTGCA
AGGTGTCCAACAAGGCCCTGCCTGCCCCCGAGGAAAAGACCATCAGCAAGGCC
AAGGGCCAGCCCAGGGAACCCCAGGTGTACACCCTGCCCCCCTCCCGGGACG
AGCTGACCAAGAACCAGGTGTCCCTGACCTGTCTGGTGAAGGGCTTCTACCCC
AGCGACATCGCCGTGGAGTGGGAGAGCAACGGCCAGCCCGAGAACAACTACA
AGACCACCCCCCCTGTGCTGGACAGCGACGGCAGCTTCTTCCTGTACAGCAAG
CTGACCGTGGACAAGAGCCGGTGGCAGCAGGGCAACGTGTTCAGCTGCAGCG
TGATGCACGAGGCCCTGCACAACCACTACACCCAGAAGAGCCTGAGCCTGTCC
CCCGGCAAGTGA

SEQ ID NO: 64

LVTVSSASTKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSGV
HTFPAVLQSSGLYSLSSVVTVPSSSLGTQTYICNVNHKPSNTKVDKKVEPKSCDKT
HTCPPCPAPELLGGPDVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYV
DGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPEEK
TISKAKGQPREPQVYTLPPSRDELTKNQVSLTCLVKGFYPSDIAVEWESNGQPENN
YKTTPPVLDSDGSFFLYSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSP
GK

SEQ ID NO: 65

ACTAGTCACCGTGAGCAGCGCCAGCACCAAGGGCCCCAGCGTGTTCCCCCTGG
CCCCCAGCAGCAAGAGCACCAGCGGCGGCACAGCCGCCCTGGGCTGCCTGGT
GAAGGACTACTTCCCCGAGCCCGTGACCGTGAGCTGGAACAGCGGAGCCCTGA
CCTCCGGCGTGCACACCTTCCCCGCCGTGCTGCAGAGCAGCGGCCTGTACAG
CCTGAGCAGCGTGGTGACCGTGCCCAGCAGCAGCCTGGGCACCCAGACCTAC
ATCTGCAACGTGAACCACAAGCCCAGCAACACCAAGGTGGACAAGAAGGTGGA
GCCCAAGAGCTGCGACAAGACCCACACCTGCCCCCCCTGCCCTGCCCCTGAGC
TGCTGGGCGGACCCGACGTGTTCCTGTTCCCCCCCAAGCCCAAGGACACCCTG
ATGATCAGCCGGACCCCCGAGGTGACCTGCGTGGTGGTGGACGTGAGCCACG
AGGACCCTGAGGTGAAGTTCAATTGGTACGTGGACGGCGTGGAGGTGCACAAC
GCCAAGACCAAGCCCCGGGAGGAACAGTACAACAGCACCTACCGGGTGGTGTC

CGTGCTGACCGTGCTGCACCAGGACTGGCTGAACGGCAAAGAATACAAGTGCA
AGGTGTCCAACAAGGCCCTGCCTCTGCCCGAGGAAAAGACCATCAGCAAGGCC
AAGGGCCAGCCCAGGGAACCCCAGGTGTACACCCTGCCCCCCTCCCGGGACG
AGCTGACCAAGAACCAGGTGTCCCTGACCTGTCTGGTGAAGGGCTTCTACCCC
AGCGACATCGCCGTGGAGTGGGAGAGCAACGGCCAGCCCGAGAACAACTACA
AGACCACCCCCCCTGTGCTGGACAGCGACGGCAGCTTCTTCCTGTACAGCAAG
CTGACCGTGGACAAGAGCCGGTGGCAGCAGGGCAACGTGTTCAGCTGCAGCG
TGATGCACGAGGCCCTGCACAACCACTACACCCAGAAGAGCCTGAGCCTGTCC
CCCGGCAAGTGA

SEQ ID NO: 66

LVTVSSASTKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSGV
HTFPAVLQSSGLYSLSSVVTVPSSSLGTQTYICNVNHKPSNTKVDKKVEPKSCDKT
HTCPPCPAPELLGGPDVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYV
DGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPLPEEK
TISKAKGQPREPQVYTLPPSRDELTKNQVSLTCLVKGFYPSDIAVEWESNGQPENN
YKTTPPVLDSDGSFFLYSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSP
GK

SEQ ID NO: 67

ATGGGCTGGTCCTGCATCATCCTGTTTCTGGTGGCCACCGCCACCGGCGTGCA
CAGCCAGGTGCAGCTGGTGCAGAGCGGAGCCGAGGTGAAGAAGCCTGGCGCC
AGCGTCAAGGTGTCCTGCAAGGCCAGCGGCTACACCTTCACCGACTACTACAT
GAACTGGGTGCGGCAGGCCCCAGGCCAGGGACTGGAATGGATGGGCAACATC
AACCCCAACAACGGCGGCACCAACTACAACCAGAAGTTCAAGGACCGGGTCAC
CATGACCACCGACACCAGCACCAGCACCGCCTACATGGAACTGCGGAGCCTGA
GAAGCGACGACACCGCCGTGTACTACTGCGCCCGGTGGATCCTGTACTACGGC
CGGTCCAAGTGGTACTTCGACGTGTGGGGCAGGGGCACACTAGTCACCGTGAG
CAGCGCCAGCACCAAGGGCCCCAGCGTGTTCCCCCTGGCCCCCAGCAGCAAG
AGCACCAGCGGCGGCACAGCCGCCCTGGGCTGCCTGGTGAAGGACTACTTCC
CCGAGCCCGTGACCGTGAGCTGGAACAGCGGAGCCCTGACCTCCGGCGTGCA
CACCTTCCCCGCCGTGCTGCAGAGCAGCGGCCTGTACAGCCTGAGCAGCGTG
GTGACCGTGCCCAGCAGCAGCCTGGGCACCCAGACCTACATCTGCAACGTGAA

CCACAAGCCCAGCAACACCAAGGTGGACAAGAAGGTGGAGCCCAAGAGCTGC
GACAAGACCCACACCTGCCCCCCCTGCCCTGCCCCTGAGCTGCTGGGCGGAC
CCGACGTGTTCCTGTTCCCCCCCAAGCCCAAGGACACCCTGATGATCAGCCGG
ACCCCCGAGGTGACCTGCGTGGTGGTGGACGTGAGCCACGAGGACCCTGAGG
TGAAGTTCAATTGGTACGTGGACGGCGTGGAGGTGCACAACGCCAAGACCAAG
CCCCGGGAGGAACAGTACAACAGCACCTACCGGGTGGTGTCCGTGCTGACCGT
GCTGCACCAGGACTGGCTGAACGGCAAAGAATACAAGTGCAAGGTGTCCAACA
AGGCCCTGCCTGCCCCCGAGGAAAAGACCATCAGCAAGGCCAAGGGCCAGCC
CAGGGAACCCCAGGTGTACACCCTGCCCCCCTCCCGGGACGAGCTGACCAAG
AACCAGGTGTCCCTGACCTGTCTGGTGAAGGGCTTCTACCCCAGCGACATCGC
CGTGGAGTGGGAGAGCAACGGCCAGCCCGAGAACAACTACAAGACCACCCCC
CCTGTGCTGGACAGCGACGGCAGCTTCTTCCTGTACAGCAAGCTGACCGTGGA
CAAGAGCCGGTGGCAGCAGGGCAACGTGTTCAGCTGCAGCGTGATGCACGAG
GCCCTGCACAACCACTACACCCAGAAGAGCCTGAGCCTGTCCCCCGGCAAGTG
A

SEQ ID NO: 68

MGWSCIILFLVATATGVHSQVQLVQSGAEVKKPGASVKVSCKASGYTFTDYYMNW
VRQAPGQGLEWMGNINPNNGGTNYNQKFKDRVTMTTDTSTSTAYMELRSLRSDD
TAVYYCARWILYYGRSKWYFDVWGRGTLVTVSSASTKGPSVFPLAPSSKSTSGGT
AALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGT
QTYICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPELLGGPDVFLFPPKPKDTL
MISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVL
TVLHQDWLNGKEYKCKVSNKALPAPEEKTISKAKGQPREPQVYTLPPSRDELTKNQ
VSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQ
QGNVFSCSVMHEALHNHYTQKSLSLSPGK

SEQ ID NO: 69

ATGGGCTGGTCCTGCATCATCCTGTTTCTGGTGGCCACCGCCACCGGCGTGCA
CAGCGACATCGTGATGACCCAGAGCCCCCTGAGCCTGCCCGTGACCCCTGGC
GAGCCCGCCAGCATCAGCTGCAGAAGCAGCCAGAGCATCGTCCAGAGCAACG
GCGACACCTACCTGGAATGGTATCTGCAGAAGCCCGGCCAGTCCCCCCAGCTG
CTGATCTACAGAGTGAGCAACCGGTTCAGCGGCGTGCCCGACAGATTCAGCGG

CAGCGGCTCCGGCACCGACTTCACCCTGAAGATCAGCCGGGTGGAGGCCGAG
GACGTGGGCGTGTACTACTGCTTTCAAGGCAGCCACGTGCCCTACACCTTCGG
CCAGGGCACCAAGCTGGAAATCAAGCGTACGGTGGCCGCCCCCAGCGTGTTCA
TCTTCCCCCCCAGCGATGAGCAGCTGAAGAGCGGCACCGCCAGCGTGGTGTGT
CTGCTGAACAACTTCTACCCCCGGGAGGCCAAGGTGCAGTGGAAGGTGGACAA
TGCCCTGCAGAGCGGCAACAGCCAGGAGAGCGTGACCGAGCAGGACAGCAAG
GACTCCACCTACAGCCTGAGCAGCACCCTGACCCTGAGCAAGGCCGACTACGA
GAAGCACAAGGTGTACGCCTGTGAGGTGACCCACCAGGGCCTGTCCAGCCCC
GTGACCAAGAGCTTCAACCGGGGCGAGTGCTGA

Seq ID NO: 70

ATGGGCTGGTCCTGCATCATCCTGTTTCTGGTGGCCACCGCCACCGGCGTGCA
CAGCCAGGTGCAGCTGGTGCAGAGCGGAGCCGAGGTGAAGAAGCCTGGCGCC
AGCGTCAAGGTGTCCTGCAAGGCCAGCGGCTACACCTTCACCGACTACTACAT
GAACTGGGTGCGGCAGGCCCCAGGCCAGGGACTGGAATGGATGGGCAACATC
AACCCCAACAACGGCGGCACCAACTACAACCAGAAGTTCAAGGACCGGGTCAC
CATGACCACCGACACCAGCACCAGCACCGCCTACATGGAACTGCGGAGCCTGA
GAAGCGACGACACCGCCGTGTACTACTGCGCCCGGTGGATCCTGTACTACGGC
CGGTCCAAGTGGTACTTCGACGTGTGGGGCAGGGGCACACTAGTGACCGTGTC
CAGCGCCAGCACCAAGGGCCCCAGCGTGTTCCCCCTGGCCCCCAGCAGCAAG
AGCACCAGCGGCGGCACAGCCGCCCTGGGCTGCCTGGTGAAGGACTACTTCC
CCGAACCGGTGACCGTGTCCTGGAACAGCGGAGCCCTGACCAGCGGCGTGCA
CACCTTCCCCGCCGTGCTGCAGAGCAGCGGCCTGTACAGCCTGAGCAGCGTG
GTGACCGTGCCCAGCAGCAGCCTGGGCACCCAGACCTACATCTGTAACGTGAA
CCACAAGCCCAGCAACACCAAGGTGGACAAGAAGGTGGAGCCCAAGAGCTGTG
ACAAGACCCACACCTGCCCCCCCTGCCCTGCCCCCGAGCTGCTGGGAGGCCC
CAGCGTGTTCCTGTTCCCCCCCAAGCCTAAGGACACCCTGATGATCAGCAGAAC
CCCCGAGGTGACCTGTGTGGTGGTGGATGTGAGCCACGAGGACCCTGAGGTG
AAGTTCAACTGGTACGTGGACGGCGTGGAGGTGCACAATGCCAAGACCAAGCC
CAGGGAGGAGCAGTACAACAGCACCTACCGGGTGGTGTCCGTGCTGACCGTG
CTGCACCAGGATTGGCTGAACGGCAAGGAGTACAAGTGTAAGGTGTCCAACAA
GGCCCTGCCTGCCCCTATCGAGAAAACCATCAGCAAGGCCAAGGGCCAGCCCA
GAGAGCCCCAGGTGTACACCCTGCCCCCTAGCAGAGATGAGCTGACCAAGAAC
CAGGTGTCCCTGACCTGCCTGGTGAAGGGCTTCTACCCCAGCGACATCGCCGT
GGAGTGGGAGAGCAACGGCCAGCCCGAGAACAACTACAAGACCACCCCCCCT
GTGCTGGACAGCGATGGCAGCTTCTTCCTGTACAGCAAGCTGACCGTGGACAA

GAGCAGATGGCAGCAGGGCAACGTGTTCAGCTGCTCCGTGATGCACGAGGCC
CTGCACAATCACTACACCCAGAAGAGCCTGAGCCTGTCCCCTGGCAAGTGATG
A

CLAIMS

1) An antibody or antigen binding fragment thereof which specifically binds IGF-1R comprising CDR H3 of SEQ. ID. NO: 1 or variant thereof which contains 1 or 2 amino acid substitutions in the CDRH3.

2) An antibody or antigen binding fragment according to claim 1 wherein the amino acid residues of SEQ. ID. NO: 1 differ by a substitution at one or more positions selected from 7 and 9.

3) An antibody or antigen binding fragment according to claim 2 wherein the amino acid residues of SEQ. ID. NO: 1 differ by one or more substitutions selected from R to S at position 7 and K to R at position 9.

4) An antibody or antigen binding fragment thereof according to any one of claims 1 to 3 wherein the antibody or antigen binding fragment further comprises one or more of the following sequences CDRH2: SEQ. ID. NO: 2 or CDRH1: SEQ. ID. NO: 3, CDRL1: SEQ. ID. NO: 4, CDRL2: SEQ. ID. NO: 7 and CDRL3: SEQ. ID. NO: 6.

5) An antibody or antigen binding fragment according to claim 4 wherein one or more of the CDR's may be replaced by a variant thereof, each variant CDR containing 1 or 2 amino acid substitutions.

6) An antibody or antigen binding fragment according to claim 4 or claim 5 and further comprising CDR H1 of SEQ. ID. NO: 3.

7) An antibody or antigen binding fragment according to claim 4 or claim 5 further comprising CDR L2 of SEQ. ID. NO: 7.

8) An antibody or antigen binding fragment according to any one of claims 1 to 7 further comprising the following CDRs:

CDRH1: SEQ. ID. NO: 3
CDRH2: SEQ. ID. NO: 2
CDRH3: SEQ. ID. NO: 1
CDRL1: SEQ. ID. NO: 4
CDRL2: SEQ. ID. NO: 7
CDRL3: SEQ. ID. NO: 6

9) An antibody or antigen binding fragment thereof which specifically binds IGF-1R and

comprises a heavy chain variable region of SEQ. ID. NO: 8 and a light chain variable region of SEQ. ID. NO: 9.

10) An antibody or antigen binding fragment thereof which specifically binds IGF-1R and comprises a heavy chain variable region of SEQ. ID. NO: 10 and a light chain variable region of SEQ. ID. NO: 11.

11) An antibody or antigen binding fragment thereof which specifically binds IGF-1R and comprises a heavy chain variable region of SEQ. ID. NO: 12 and a light chain variable region of SEQ. ID. NO: 13.

12) An antibody or antigen binding fragment thereof which specifically binds IGF-1R and comprises a heavy chain variable region domain of SEQ. ID. NO: 14 and a light chain variable region domain of SEQ. ID. NO: 16.

13) An antibody or antigen binding fragment thereof which specifically binds IGF-1R and comprises a heavy chain variable region of SEQ. ID. NO:15 and a light chain variable region of SEQ. ID. NO:16.

14) An antibody or antigen binding fragment comprising CDR's according to any one of claims 1 to 8 or heavy or light chain variable regions according to any one of claims 9 to 13 wherein the antibody or antigen binding fragment is rat, mouse, primate (eg cynomolgus, Old World monkey or Great Ape) or human.

15) An antibody or antigen binding fragment according to any one of claims 1 to 14 wherein the antibody is a humanised or chimaeric antibody.

16) An antibody or antigen binding fragment according to any one of claims 1 to 15 wherein the antibody or antigen binding fragment additionally binds primate IGF-1R.

17) An antibody or antigen binding fragment according to any one of claims 1 to 16 wherein the antibody comprises a constant region.

18) An antibody according to claim 17 wherein the antibody comprises a constant region of IgG isotype.

19) An antibody according to claim 18 wherein the antibody is IgG1.

20) An antibody according to any one of claims 1 to 19 comprising a constant domain region such that the antibody has a reduced ADCC and/or complement activation or effector

functionality.

21) An antibody according to any one of claims 1 to 19 comprising a mutated constant domain or constant domain with an altered glycosylation profile such that the antibody has enhanced effector functions/ ADCC and/or complement activation.

22) An antigen binding fragment according to any one of claims 1 to 17 wherein the fragment is a Fab, Fab', $F(ab')_2$, Fv, diabody, triabody, tetrabody, miniantibody, minibody, isolated VH or isolated VL.

23) An antibody according to any one of claims 1 to 21 or antigen binding fragment according to any one of claims 1 to 22 wherein the antibody or antigen binding fragment thereof is capable of at least some effector function for example where it is capable of some ADCC or CDC function.

24) A recombinant transformed, transfected or transduced host cell comprising at least one expression cassette, whereby said expression cassette comprises a polynucleotide encoding a heavy chain of an antibody or antigen binding fragment according to the invention described herein and further comprises a polynucleotide encoding a light chain of an antibody or antigen binding fragment according to the invention described herein.

25) A recombinant transformed, transfected or transduced host cell comprising at least one expression cassette, whereby a first expression cassette comprises a polynucleotide encoding a heavy chain of an antibody or antigen binding fragment according to the invention described herein and further comprises a second expression cassette comprises a polynucleotide encoding a light chain of an antibody or antigen binding fragment according to the invention described herein.

26) A host cell according to claim 24 or claim 25 wherein the cell is eukaryotic.

27) A host cell according to claim 26 wherein the cell is mammalian.

28) A host cell according to claim 27 wherein the cell is CHO or NSO.

29) A method for the production of an antibody according to claims 1 to 21 or antigen binding fragment thereof according to any one of claims 1-22 which method comprises the step of culturing a host cell of any one of claims 24-28 in a serum- free culture media.

30) A method according to claim 29 wherein said antibody is secreted by said host cell into a culture media.

31) A method according to claim 30 wherein said antibody is further purified to at least 95% or greater (ege.g. 98% or greater) with respect to said antibody containing culture media.

32) A pharmaceutical composition comprising an antibody or antigen binding fragment thereof according to any preceding claim and a pharmaceutically acceptable carrier.

33) A kit-of-parts comprising the composition according to claim 32 together with instructions for use.

34) A method of treating a human patient afflicted with cancer which method comprises the step of administering a therapeutically effective amount of an antibody or antigen binding fragment thereof according to any preceding claim or composition of claim 32.

35) A method according to claim 34 wherein the patient is afflicted with breast cancer.

36) A method according to claim 34 wherein the patient is afflicted with prostate cancer.

37) Use of an antibody or antigen binding fragment thereof according to any preceding claim in the manufacture of a medicament for the treatment of a disease or disorder selected from the group consisting of; rheumatoid arthritis, breast cancer, prostrate cancer, lung cancer or myeloma.

38) An antibody or antigen binding fragment thereof according to any preceding claim wherein the antibody neutralises the activity of IGF-1R.