

SPECIFICATION

ANALYSIS OF PREDISPOSITION BASED ON HUMAN AIRWAY
TRIPSIN PROTEASE GENE POLYMORPHISM

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Technical Field

10 This invention relates to a method for predicting the constitution susceptible to the onset of specific diseases, effects on methods of treatment for patients suffering from said diseases or predicting the prognosis of the treatment by analysis of genetic polymorphisms of a human trypsin-like enzyme of a respiratory tract.

15 Background Art

Research on related genes has recently been promoted not only in genetic diseases due to deletion or mutation of single genes but also in multifactorial diseases caused by entanglement of several genetic predispositions and environmental factors. As a result, the deletion or point mutation and isoforms related to the multifactorial diseases and further mutation of genetic parts (introns or promoters) without affecting actually translated amino acid sequences have come to be considered as risk factors for the diseases.

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Effect of the Invention

It has been published that the correlation is recognized between bone density and genetic polymorphisms of an intron of the vitamin D receptor in the osteopathic field as the prior art (Morrison, N. A. et al., Nature, 367: 284-287, 1994). In the field of circulatory organs, it has been reported that the I type (insertion type) and D type (deletion type) genetic polymorphisms of an angiotensin-converting enzyme are associated with the onset of myocardial infarctions (Cambien, F. et al., Nature 359: 641-644,

1992) and the amino acid substitution of M235T of angiotensinogen and the polymorphisms of a promoter region of G-6A are associated with the onset of essential hypertension (Inoue, I. et al., J. Clin. Invest., 99: 1786-1797, 1997). Furthermore, in the field of the nervous system, it has been reported on the association between the onset of dementia and the isoforms of apoE protein. Much research has been carried out in the association between the genetic polymorphisms of glutathione S-transferase and the onset of cancers in the cancer-related field. As the field of respiratory diseases, it has been reported on the association between the onset or morbid state of asthma and the TNF (Moffatt, M. F. et al., Hum. Mol. Genet. 6 (4): 551-554, 1997) and the association between the onset or morbid state of the asthma and the genetic polymorphisms of an angiotensin converting enzyme (Benessiano, J. et al., J. Allergy Clin. Immunol. 99 (1): 53-57, 1997).

Furthermore, attention has been paid to the genetic background as one of causes of difference between patients in sensitivity to drugs used for treating diseases, and it is has been desired even by the medical site to provide the directionality such as selection of methods of treatment according to the drug sensitivity of individual humans by diagnosis of the genetic polymorphisms. It is thought that the drug development by selecting patient groups expectable of drug effects according to the genetic polymorphisms is effective in clinical trials (Kleyn K. W. et al., Science, 281: 1820-1821, 1998). A report on the genetic polymorphisms of an intron of an angiotensin converting enzyme [ACE (angiotensin converting enzyme)] and effects of an ACE inhibitor (Yoshida, H. et al., J. Clin. Invest. 96: 2162-2169, 1995), a report on the genetic polymorphisms of beta 2-adrenergic receptor and effects of the beta-agonist on asthma (Liggett, S. B., Am. J. Respir. Crit. Care Med. 156 (4 Pt 2): S156-162, 1997) and the like are cited as the conventional reports related to the drug sensitivity and the genetic polymorphisms.

On the other hand, the human trypsin-like enzyme of the respiratory tract related to the present invention has been purified from the sputum of patients suffering from chronic airway diseases (Japanese Unexamined Patent Publication No. 7-067640 and Yasuoka, S. et al., Am. J. Respir. Cell Mol. Biol., 16: 300-308, 1997) and the amino acid sequence and

cDNA sequence thereof have been already made clear (Japanese Unexamined Patent Publication No. 8-89246 and Yamaoka K. et. al., J. Biol. Chem., 273(19): 11895-11901, 1998). Several studies have been made of the activity possessed by the enzyme in vitro. Since the enzyme has
5 production enhancing actions on cytokines such as IL-8 or GM-CSF derived from a human bronchial epithelial cell line including the association with mucociliary movement, the possibility for association with the morbid state of airway inflammations is considered (Terao, Noriko et al., the Japanese Respiratory Society, 1998). Since the enzyme has enzyme
10 activities such as hydrolytic activity for fibrinogen and activating actions on plasminogen activators (pro-urokinase) (Yoshinaga, Junko et al., Conference on Proteases and Inhibitors in Pathophysiology and Therapeutics, 1998), the possibility is assumed for anti-inflammatory actions through the formation of fibrins on the airway mucosal surfaces or
15 modification of the morbid state thereof in chronic airway diseases and the possibility is also considered for the association with cancer metastasis or the like. The association of the enzyme with physiological functions or the morbid state in vivo is not yet sufficiently elucidated, and the genetic parts (introns or promoters) without corresponding to the actually
20 translated amino acid sequences has not yet known about genes at all. Further, no investigation has hitherto been carried out on the association of the presence or absence of the genetic polymorphisms for the human trypsin-like enzyme of the respiratory tract or the genetic polymorphisms with diseases.

25 By the way, much information can be provided about the prediction of the onset of specific diseases, prognosis of treatment, selection of appropriate methods of the treatment and administered drugs or the like by the prediction of diseases-associated constitution by genetic analysis. Accordingly, the prediction is desired by many physicians and patients and
30 further makes the prophylaxis of onset and early therapy possible. Therefore, it is thought that the prediction is related with a reduction in medical care expenditures to become indispensable for the future medical care.

It is, however, very difficult to find out a gene associated with

diseases and establish an analytical method therefor, and there are few examples of genetic analytical methods in which the association with diseases is recognized as described above. Therefore, the development of the genetic analytical technique for making various disease-associated constitutions predictable is strongly desired.

On the other hand, it is not yet elucidated with what diseases the human trypsin-like enzyme of the respiratory tract is associated at present.

Disclosure of the Invention

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As a result of intensive research made in consideration of the problems of the prior art, the present inventors et al. have designed a primer for genetically amplifying an intron part on the genome of the human trypsin-like enzyme of the respiratory tract specifically expressing in the human respiratory tract, novelly determined the DNA sequence of both the termini of the amplified genetic fragment and the novelly determined DNA sequence in an exon/intron border part, found out that there are genetic polymorphisms in the amplified genetic fragment and the novelly determined DNA sequence and further firstly found out the association of the genotypes of the human trypsin-like enzyme of the respiratory tract with diseases in individual humans by analyzing the genetic polymorphisms, thus attaining the present invention.

That is, an object of the present invention is to provide a method for predicting the constitution of individual humans susceptible to the onset of specific diseases or effects of treatment on patients or prognosis of the treatment by analyzing the genetic polymorphisms of the human trypsin-like enzyme of the respiratory tract.

Further, the present invention is a method for diagnosing an abnormality in the mucociliary biophylactic system by the analysis of the genetic polymorphisms of the human trypsin-like enzyme of the respiratory tract and a method for predicting the constitution of individual humans susceptible to the onset of diseases, effects of treatment on patients or prognosis of the treatment.

Furthermore, the present invention is a genetic fragment containing

a part or all of the base sequence of an intron in the human trypsin-like enzyme of the respiratory tract.

Brief Description of Drawings:

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Fig. 1 illustrates agarose gel electrophoretic patterns of DNA fragments containing an intron region of a gene encoding a mature protein of the human trypsin-like enzyme of the respiratory tract obtained by genetic amplification. It is observed that lanes 1 and 5 are markers (10 Kb, 7 Kb, 5 Kb, 4 Kb, 3 Kb, 2.5 Kb, 2 Kb, 1.5 Kb and 1 Kb from above), lane 2 is a DNA fragment of about 6 Kb amplified by primers A1 (sequence No. 7) and A2 (sequence No. 8), lane 3 is a DNA fragment of about 1.5 Kb amplified by primers B1 (sequence No. 9) and B2 (sequence No. 10) and lane 4 is a DNA fragment of about 3.4 Kb amplified by primers C1 (sequence No. 11) and C2 (sequence No. 12).

Fig. 2 illustrates agarose gel electrophoretic patterns of DNA fragments containing an intron region of a gene encoding a propeptide of the human trypsin-like enzyme of the respiratory tract obtained by the genetic amplification. It is observed that lanes 1 and 4 are markers (10 Kb, 7 Kb, 5 Kb, 4 Kb, 3 Kb, 2.5 Kb, 2 Kb, 1.5 Kb and 1 Kb from above), lane 2 is a DNA fragment of about 5.5 Kb amplified by primers D1 (sequence No. 13) and D2 (sequence No. 14) and lane 3 is a DNA fragment of about 5 Kb amplified by primers E1 (sequence No. 15) and E2 (sequence No. 16).

Fig. 3 illustrates relative positions of introns A, B and C on the human trypsin-like enzyme genome of the respiratory tract and agarose gel electrophoretic patterns of genetic polymorphisms (RFLP) of the intron A detected by TaqI and of the intron C detected by MboI and BstUI.

Best Mode for Carrying Out the Invention

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According to the present invention, there are provided a method for predicting the association of the constitution of individual humans with specific diseases and a genetic fragment or a DNA sequence used for the genetic analysis thereof.

Respiratory diseases, pulmonary cancer, especially pulmonary emphysema (PE), sinobronchial syndrome, diffuse panbronchiolitis (DPB) and bronchiectasis (BE) belonging to chronic obstructive pulmonary diseases (COPD) or abnormalities in the mucociliary biophylactic system
5 are exemplified as specific diseases for judging the constitution susceptible to the onset or specified diseases for predicting the judgment on effects of treatment thereof or prognosis of the treatment by the method of the present invention.

Analysis of the genotypes classified by detecting one or more base
10 mutations and analysis of the haplotypes classified by detecting one or more of the base mutations are exemplified as an analytical method for the genetic polymorphisms of the human trypsin-like enzyme of the respiratory tract in the present invention.

The analytical method for the genetic polymorphisms of the
15 trypsin-like enzyme of the respiratory tract is carried out by, for example, an analytical method by a restriction fragment length polymorphisms (RFLP) according to the cleavage with a restriction enzyme. The analytical method for the genetic polymorphisms in the present invention includes even an analytical method for the genetic polymorphisms
20 detectable by the Southern hybridization using a cDNA sequence of the human trypsin-like enzyme of the respiratory tract. That is, it is a method for cleaving the genomic DNA with a restriction enzyme capable of detecting the genetic polymorphisms disclosed in the present invention, then carrying out a gel electrophoresis, performing transcription to a
25 nitrocellulose membrane or the like and subsequently analyzing the cleaved patterns of the human trypsin-like enzyme genome of the respiratory tract using the human trypsin-like enzyme cDNA of the respiratory tract as a probe. The analysis can be made even by a method for amplifying a DNA fragment by PCR so as to include sites for the genetic polymorphisms,
30 then converting the amplified DNA fragment into a single strand and analyzing the resulting single strand by a difference in mobility of electrophoresis [PCR-single strand conformation polymorphism (SSCP)] method or the like. Furthermore, many methods such as a mismatch PCR method, a PCR-allele specific oligo (ASO) method using an allele

specific oligonucleotide, a method for judgment by carrying out annealing using an oligo probe or a pinpoint sequencing method for directly determining the base sequence of the genetic polymorphic sites are cited as the method for making the detection of the polymorphic sites possible, and
5 the analytical method for the genetic polymorphisms can be applied even to genetic diagnosis by using a DNA chip.

An intron may be used as sites for analysis of the genetic polymorphisms, and the following genetic fragments are exemplified as the site for analysis of the specific genetic polymorphisms:

- 10 (a) a genetic fragment containing an intron region amplifiable by using the primers represented by sequence No. 7 and sequence No. 8,
- (b) a genetic fragment containing an intron region amplifiable by using the primers represented by sequence No. 9 and sequence No. 10.
- (c) a genetic fragment containing an intron region amplifiable by using the
15 primers represented by sequence No. 11 and sequence No. 12
- (d) a genetic fragment containing an intron region amplifiable by using the primers represented by sequence No. 13 and sequence No. 14,
- (e) a genetic fragment containing an intron region amplifiable by using the primers represented by sequence No. 15 and sequence No. 16,
- 20 (f) a genetic fragment containing intron C amplifiable by using the primers represented by the sequence No. 11 and the sequence No. 12 and
- (g) a genetic fragment containing intron A amplifiable by using the primers represented by the sequence No. 7 and the sequence No. 8.

In the intron C, a genetic fragment containing a sequential part
25 recognized herein by a restriction enzyme BstUI, MboI, MseI or FbaI is exemplified as the sites for analysis of the genetic polymorphisms. In the intron A, a part containing a sequential part recognized by a restriction enzyme MboI, TaqI or AfaI, one of the genetic polymorphic sites represented by sequence No. 17 or a combination of one or more thereof is exemplified as
30 the sites for the analysis of the genetic polymorphisms. The genotypic or haplotypic classification is judged by the analysis of the sites.

Furthermore, the present invention is a genetic fragment containing a part or all of the base sequence of an intron in the human trypsin-like enzyme of the respiratory tract and the following genetic fragments are

especially exemplified:

- (a) a genetic fragment containing an intron region amplifiable by using the primers represented by the sequence No. 7 and the sequence No. 8,
- (b) a genetic fragment containing an intron region amplifiable by using the primers represented by the sequence No. 9 and the sequence No. 10,
- (c) a genetic fragment containing an intron region amplifiable by using the primers represented by the sequence No. 11 and the sequence No. 12,
- (d) a genetic fragment containing an intron region amplifiable by using the primers represented by the sequence No. 13 and the sequence No. 14
- (e) a genetic fragment containing an intron region amplifiable by using the primers represented by the sequence No. 15 and the sequence No. 16,
- (f) a genetic fragment containing an intron in the human trypsin-like enzyme of the respiratory tract comprising the base sequence represented by either of the sequence No. 1 to the sequence No. 6 or a genetic fragment sandwiched between the base sequences and
- (g) a genetic fragment of the intron C in the human trypsin-like enzyme of the respiratory tract comprising the base sequence represented by the sequence No. 17.

Examples

The present invention is explained in detail hereinafter by way of examples, provided that the examples are not intended as a definition of the method for predicting the disease-associated constitution by the analysis of the genetic polymorphisms of the human trypsin-like enzyme of the respiratory tract.

[Standard methods for operations]

Standard DNA extracting operations, genetic amplifying operations, restriction enzyme cleavage operations and electrophoretic operations usable in the present invention are explained hereinafter.

(a) Standard DNA extracting operations

Nothing is especially limited in biosamples used for the genetic

amplification of the present invention; however, blood corpuscle components are suitable because specimens are readily collected and DNA is easily extracted.

1. The whole blood in a volume of 0.5 ml (using a 2Na-EDTA
5 anticoagulant) is placed in a micro-centrifugal tube having a capacity of 1.5 ml.

2. A dissolvent in a volume of 0.5 ml is added to the tube, and the tube is lightly tapped several times. The tube is then turned upside down, and the liquids are mixed.

10 (The following mixing operations are performed according to the above procedures).

An example of the dissolvent: $1 \times \text{SSC}$

This is prepared by diluting $20 \times \text{SSC}$ regulated to pH 7.0 with 10N NaOH containing 175.3 grams of NaCl and 88.2 grams of sodium citrate
15 in 1 liter 10-fold.

3. The mixture is centrifuged (at 4°C and 10,000 g for 20 seconds), and the supernatant is then removed so as not to discharge dark pellets.

4. The dissolvent in a volume of 1 ml is added to stir the mixture.

5. The mixture is centrifuged (at 4°C and 10,000 g for 20 seconds),
20 and the supernatant is then removed.

6. Steps 4 and 5 are repeated once more.

7. An enzyme reaction solution in a volume of 200 μl and a proteolytic enzyme in a volume of 10 μl are added and mixed therewith.

An example of the enzyme reaction solution: A mixture liquid of
25 0.04 M DTT (dithiothreitol) with 0.2 M NaOAc (sodium acetate) and 0.4% SDS

An example of the proteolytic enzyme liquid:

A 10 mg/ml proteinase (Proteinase K)

8. The mixture is kept warm at 37°C for 1 hour (the mixture is mixed
30 by lightly shaking 2 to 3 times in the course thereof).

9. A solution of sodium iodide in a volume of 300 μl is added and mixed therewith.

10. Isopropyl alcohol in a volume of 0.5 ml is added and mixed until a white linear DNA is completely visible.

11. The resulting mixture is centrifuged (at room temperature and 10,000 g for 10 minutes) , and the supernatant is then slowly removed. The solution remaining in a tube wall is sufficiently removed by a method for placing the tube on a filter paper upside down or the like.

- 5 12. A wash liquid (A) in a volume of 1 ml is added and mixed therewith. The mixture is sufficiently mixed so as to peel a precipitate from the tube wall.

An example of the wash liquid (A): 70% EtOH

13. The resulting mixture is centrifuged (at room temperature and
10 10,000 g for 5 minutes), and the supernatant is then removed.

14. A wash liquid (B) in a volume of 1 ml is added, and the prepared mixture is sufficiently mixed so as to peel the precipitate from the tube wall.

An example of the wash liquid (B): 80% EtOH

- 15 15. The mixture is centrifuged (at room temperature and 10,000 g for 5 minutes), and the supernatant is then removed.

16 The DNA precipitate is lightly vacuum-dried (the drying time is within 3 minutes because the DNA is sparingly dissolved when drying the precipitate too much).

(a) Standard genetic amplifying operations

- 20 Although several principles are known about the genetic amplifying method, the polymerase chain reaction method (PCR method) is described as a standard one hereinafter.

Composition of the reaction solution: 50 mM of KCl, 10 mM of Tris-HCl (pH 9.0, 25°C), 0.1% of TritonX-100, 1.5 mM of MgCl₂, 2 mM of dNTPs,
25 15 μ M of Forward Primer, 15 μ M of Reverse Primer, 1 mg/l of a genomic DNA and 1 unit of TaqDNA polymerase, the total volume of 50 μ l

Reaction cycle: at 94°C for 1 minute, 64°C for 1 minute and 72°C for 1 minute as one cycle. The reaction is conducted for 40 cycles.

Primers used are the following C1 (35 bases) and C2 (35 bases):

- 30 C1: 5'-GGAGC CATCT TGTCT GGAAT GCTGT GTGCT GGAGT-3'
C2: 5'-CACAA TAAAC CAAAG CCGCC GTGAG TCTTC TTGTA-3'

(c) Standard restriction enzyme cleavage operations

Composition of the reaction solution for MboI: 10 mM of Tris-HCl (pH7.4), 10 mM of MgCl₂, 100 mM of NaCl, 10 mM of KCl, 1 mM of DTT and

100 µg/ml of BSA (bovine serum albumin)

MboI at a concentration of 10 units/20 µl of reaction solution is added to carry out incubation at 37°C for 3 hours.

(b) Standard electrophoretic operations

5 The buffer solution for electrophoresis is 0.5 × TBE, with the proviso that 5 × TBE contains 54 grams of Tris base, 27.5 grams of boric acid and 1 mM of EDTA in 1 liter and is regulated to pH 8.0. The above buffer solution is used to carry out electrophoresis under a voltage of 100 V for 30 minutes by using a 1% or a 3% agarose gel [using Seakem GTA Agarose
10 (FMC Bio Products)] (containing 0.5 µl/ml of ethidium bromide). The band of the DNA is then observed with a UV lamp.

[Example 1] Obtaining of DNA fragment containing an intron on human trypsin-like enzyme genome of the respiratory tract and analysis of base
15 sequence thereof

In order to search for an intron site on the genome of a mature human trypsin-like enzyme of the respiratory tract, primers were prepared by referring to the constitution of exons and introns of the genome of several tryptase analogous enzymes to amplify a human genomic DNA.
20 A DNA fragment (genetic fragment) of about 6 Kb was amplified by the primers A1 (sequence No. 7) and A2 (sequence No. 8), and a DNA fragment of about 1.5 Kb was amplified by the primers B1 (sequence No. 9) and B2 (sequence No. 10). A DNA fragment of about 3.4 Kb was amplified by the primers C1 (sequence No. 11) and C2 (sequence No. 12). (Fig.1)

25 The DNA fragments were excised from gels and purified, and the respective fragments were inserted into TA cloning vectors (manufactured by Invitrogen Corporation). The base sequences on both sides (5'-terminus and 3'-terminus) of the insert DNA were determined for several clones thereof to find that the cDNA sequence of the human trypsin-like
30 enzyme of the respiratory tract was present contiguous to the sequences of the primers, consensus sequences were recognized as contiguous thereto in border regions of the exon-intron on both sides of the 5'-terminus and the 3'-terminus and the amplified DNA fragment was a DNA fragment containing the intron region in the human trypsin-like

enzyme of the respiratory tract.

The sequences of the introns which have been made clear in the present invention in a genetic region encoding the mature protein of the human trypsin-like enzyme of the respiratory tract are respectively referred to as intron A, intron B and intron C from the 5'-side. (Fig. 3). The base sequences at the 5'- and the 3'-termini of the clarified respective introns are as represented by the sequence Nos. 1, 2, 3, 4, 5 and 6. As for the intron C, the whole base sequence is represented by the sequence No. 17.

On the other hand, several primers were prepared for the region encoding a propeptide of the human trypsin-like enzyme of the respiratory tract to carry out the genetic amplification. Thereby, it was clear that a DNA fragment of about 5.5 Kb was amplified by the primers D1 (sequence No. 13) and D2 (sequence No. 14) and a DNA fragment of about 5 Kb was amplified by the primers E1 (sequence No. 15) and E2 (sequence No. 16). (Fig. 2). The genetic fragments are regarded as containing the introns.

[Example 2] Analysis of genetic polymorphisms of introns in human trypsin-like enzyme of the respiratory tract

In order to investigate whether or not the genetic polymorphisms are present in the introns A, B and C, the genomic DNA of normal 23 humans was extracted from the whole blood (the standard method for operations) and respectively amplified by the primers A1 and A2 for amplifying the intron A, the primers B1 and B2 for amplifying the intron B and the primers C1 and C2 for amplifying the intron C according to the PCR to compare the cleavage patterns with various kinds of restriction enzymes. As a result, it was found that the genetic polymorphisms were observed with the restriction enzymes MboI, TaqI and AfaI in the intron A.

Conversely, the genetic polymorphisms were not observed by restriction enzymes Tsp509I, AluI, NlaIII, MspI, BstUI, BfaI, HinPI, HaeIII, HindIII, SspI, PstI, EcoRI, SalI and EcoRV.

In the intron B, the genetic polymorphisms were not observed by the restriction enzymes Tsp509I, AluI, NlaIII, MspI, BstUI, BfaI, HinPI, HaeIII, MboI, AfaI, TaqI, MseI, ClaI, NsiI, EcoT14I, NdeI, PmlI, ApaLI, AatII, ApaI,

KpnI, BsmI, HindIII, SspI and EcoRV.

In the intron C, it was found that the genetic polymorphisms were observed by the restriction enzymes MboI, BstUI, MseI and FbaI.

Conversely, the genetic polymorphisms were not observed by restriction
5 enzymes AluI, NlaIII, MspI, BfaI, HinfI, HaeIII, AfaI, TaqI, HindIII, SspI, BglII, EcoT14I, PvuII, PvuI, EcoRI, BamHI, EcoRV and KpnI.

When the DNA fragment containing the intron C amplified by using a combination of the primers C1 and C2 was cleaved with the MboI or BstUI, experiments on cleavage by the MboI revealed that the case where a band
10 appeared at 1.3 Kb could be judged as genotype MM, the case where a band appeared at 1.05 Kb could be judged as genotype mm and the case where two bands appeared together at 1.3 Kb and 1.05 Kb could be judged as genotype Mm according to the electrophoresis. On the other hand, in the case of the BstUI, the case where a band appeared at 3.4 Kb could be judged
15 as BB, the case where two bands appeared at 2.45 Kb and 0.95 Kb could be judged as bb and the case where three bands appeared together could be judged as Bb.

Fig. 3 shows electrophoretic patterns of the genetic polymorphisms of the intron A detected by the TaqI and of the intron C detected by the
20 MboI and BstUI. Furthermore, the whole base sequence of the bm type haplotype was determined for the intron C from the DNA fragment obtained from the genome of one example of a bbmm type normal human by the PCR. (sequence No. 17). The genetic polymorphic sites detected by the BstUI and MboI in the base sequence of the intron C are indicated by
25 arrows with white spaces.

The base sequence of one example of a patient suffering from the BBmm type BE was determined to find that the base sequence of the Bm haplotype BstUI genetic polymorphic sites were not cleaved with the BstUI because CGCG was converted into ACCG.

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[Example 3] Analysis of disease-associated constitution by the analysis of the intron genetic polymorphisms of human trypsin-like enzyme of the respiratory tract

As for the statistical analysis, the analysis was made according to

the chi-square test using a statistical analysis software Stat View4.02 (Abacus Concepts Co.).

Example 3-1 Analysis of disease-associated constitution by genotypic classification

Among the genotypes disclosed in Example 2, investigation was made whether or not the classification of diseases associated with the human trypsin-like enzyme of the respiratory tract can be made for the genetic polymorphisms detected with the MboI and BstUI in the intron C. The diseases selected as objects are diffuse panbronchiolitis (DPB), bronchiectasis (BE), pulmonary emphysema (PE) and bronchial asthma (BA) which are respiratory diseases.

The genotypes of each human were judged by selecting 106 normal humans, 29 patients suffering from the diffuse panbronchiolitis (DPB), 38 patients suffering from the bronchiectasis (BE), 22 patients suffering from pulmonary emphysema (PE) and 32 patients suffering from the bronchial asthma (BA) according to the standard method for operations. MboI and BstUI were used as the restriction enzymes.

The number of humans having the occurrence and the frequency of occurrence of each genotype and the number of occurrence and the frequency of occurrence of each allelic type were as shown in Tables 1 and 2.

Table 1 BstUI genotype

Number of humans having
the occurrence (humans)

Number of occurrence (allele)

	BB	Bb	bb	Total
Normal	16	55	35	106
DPB	8	12	9	29
BE	11	15	12	38
PE	5	12	5	22

	B	b	Total
Normal	87	125	212
DPB	28	30	58
BE	37	39	76
PE	22	22	44

BA 2 12 18 32 BA 16 48 64

Frequency of occurrence (%)

	BB	Bb	bb
Normal	15.1	51.9	33.0
DPB	27.6	41.4	31.0
BE	28.9	39.5	31.6
PE	22.7	54.5	22.7

BA 6.3 37.5 56.3

Table 2 MboI genotype

Number of humans having

5 the occurrence (humans)

Frequency of occurrence (%)

	B	b
Normal	41.0	59.0
DPB	48.3	51.7
BE	48.7	51.3
PE	50.0	50.0

BA 25.0 75.0

Number of occurrence (allele)

	MM	Mm	mm	Total
Normal	10	40	56	106
DPB	3	7	19	29
BE	3	15	20	38
PE	3	6	13	22

BA 1 12 19 32

	M	m	Total
Normal	60	152	212
DPB	13	45	58
BE	21	55	76
PE	12	32	44

BA 14 50 64

Frequency of occurrence (%)

	MM	Mm	mm
Normal	9.4	37.7	52.8
DPB	10.3	24.1	65.5
BE	7.9	39.5	52.6
PE	13.6	27.3	59.1

BA 3.1 37.5 59.4

Frequency of occurrence (%)

	M	m
Normal	28.3	71.7
DPB	22.4	77.6
BE	27.6	72.4
PE	27.3	72.7

BA 21.9 78.1

10

Tables 1 and 2 show that the frequency of occurrence of BB type manifests a higher tendency in the DPB, BE and PE by judging the above genotypes. That is, the judgment can be made that individual humans having the genotypes have constitutions susceptible to the DPB, BE and PE. Furthermore, when the DPB, BE and PE are collected into the patient groups suffering from the respiratory three diseases according to the classification of the chronic obstructive pulmonary diseases (COPD), results are obtained as follows: The frequency of occurrence of the BB type is statistically significantly higher than that of normal humans (chi-square p value = 0.04 and chi-square value = 4.2).

20

Frequency of observation

Three diseases, BB/NotBB

	BB	NotBB	Total
Normal	16	90	106
Respiratory	24	65	89
3 diseases			
Total	40	155	195

5

Percent (row):

Three diseases, BB/Not BB

	BB	Not BB	Total
Normal	15	85	100
Respiratory	27	73	100
3 diseases			
Total	21	79	100

10

Contingency table analytical statistics:

Three diseases, BB/NotBB

Number of missing values	63
Degree of freedom	1
Chi-square value	4.182
Chi-square p value	.0409
G-square value	4.177
G-square p value	.0410
Contingency table analytical coefficient	.145
Phi	.146
Chi-square value (Yates' continuity correction)	3.489
Chi-square p value (Yates' continuity correction)	.0618
Fisher's direct method p value	.0503

Example 3-2 Analysis of disease-related constitution by haplotypic classification

Frequency of occurrence of haplotypes

5 The number of humans having the occurrence and the frequency of occurrence of the haplotypes according to a combination of both genetic polymorphisms of BstUI and MboI in 106 normal humans, 29 patients suffering from diffuse panbronchiolitis (DPB), 38 patients suffering from bronchiectasis (BE), 22 patients suffering from pulmonary emphysema (PE) and 32 patients suffering from bronchial asthma (BA) are shown in the tables. As a result of investigation on 238 humans, the tables suggest that the bM haplotype is almost absent in Japanese due to the absence of anyone having the genotypes of Bb-MM, bb-MM and bb-Mm at all.

Number of humans having the occurrence (humans)

	BB-MM	BB-Mm	BB-mm	Bb-MM	Bb-Mm	Bb-mm	bb-MM	bb-Mm	bb-mm	Total
Normal	10	6	0	0	34	21	0	0	35	106
DPB	3	2	3	0	5	7	0	0	9	29
BE	3	7	1	0	8	7	0	0	12	38
PE	3	0	2	0	6	6	0	0	5	22
15 BA	1	1	0	0	11	1	0	0	18	32

Frequency of occurrence (%)

	BB-MM	BB-Mm	BB-mm	Bb-MM	Bb-Mm	Bb-mm	bb-MM	bb-Mm	bb-mm	Total
Normal	9.4	5.7	0.0	0.0	32.1	19.8	0.0	0.0	33.0	100%
DPB	10.3	6.9	10.3	0.0	17.2	24.1	0.0	0.0	31.0	100%
BE	7.9	18.4	2.6	0.0	21.1	18.4	0.0	0.0	31.6	100%
PE	13.6	0.0	9.1	0.0	27.3	27.3	0.0	0.0	22.7	100%
BA	3.1	3.1	0.0	0.0	34.4	3.1	0.0	0.0	56.3	100%

As for the following analysis, the statistical technique (chi-square method) was used to promote the analysis of the association with

respiratory diseases on the assumption that the human trypsin-like enzyme of the respiratory tract genetic haplotypes of Japanese were the three kinds of BM, Bm and bm (when the number of occurrence was 5 or more, the chi-square p values were used as the following p values and the
 5 Fisher's direct method p values were indicated as the following p values in the case of 2×2 table including a frame of the number of occurrence of 4 or below).

Allelic classification (1)

10 The investigation was made on the frequency of occurrence of each allele for three kinds of haplotypes of Japanese to find that there was a deviation in distribution of the frequency of occurrence between the respiratory three disease groups (DPB, BE and PE) belonging to the COPD and normal humans with a statistical significant difference ($p = 0.027$).
 15 Particularly, the frequency of occurrence of Bm allele was higher in the respiratory three disease groups (DPB, BE and PE) belonging to the COPD than in normal humans.

The frequency of occurrence of Bm allele in the BE, DPB and PE was high with regard to each disease.

20 Conversely, the frequency of occurrence of Bm allele was lower for the BA.

Frequency of observation:

Three diseases, allele

	Bm.	BM	b.m.	Total
Normal	27	60	125	212
Respiratory 3 diseases	41	46	91	178
Total	68	106	216	390

Percent (row): Three diseases, allele

	Bm.	BM	b.m.	Total
Normal	13	28	59	100
Respiratory 3 diseases	23	26	51	100
Total	17	27	55	100

Contingency table analytical statistics:

Three diseases, allele

Number of missing values	107
Degree of freedom	2
Chi-square value	7.174
Chi-square p value	.0277
G-square value	7.164
G-square p value	.0278
Contingency table analytical coefficient	.134
Cramer's V value	.136

5

Frequency of observation: DPB, allele

	Bm.	BM	b.m.	Total
Normal	27	60	125	212
DPB	15	13	30	58
Total	42	73	155	270

Percent (row): DPB, allele

	Bm.	BM	b.m.	Total
Normal	13	28	59	100
DPB	26	22	52	100
Total	16	27	57	100

10

Contingency table analytical statistics:

DPB, allele

Number of missing values	227
Degree of freedom	2
Chi-square value	6.044
Chi-square p value	.0487
G-square value	5.488
G-square p value	.0643
Contingency table analytical coefficient	.148
Cramer's V value	.150

15

Frequency of observation: BE, allele

	Bm.	BM	b.m.	Total
BE	16	21	39	76
Normal	27	60	125	212
Total	43	81	164	288

5

Percent (row): BE, allele

	Bm.	BM	b.m.	Total
BE	21	28	51	100
Normal	13	28	59	100
Total	15	28	57	100

10

Contingency table analytical statistics:

BE,allele

Number of missing values	209
Degree of freedom	2
Chi-square value	3.175
Chi-square p value	.2044
G-square value	3.007
G-square p value	.2224
Contingency table analytical coefficient	.104
Cramer's V value	.105

Frequency of observation: PE, allele

	Bm.	BM	b.m.	Total
PE	10	12	22	44
Normal	27	60	125	212
Total	37	72	147	256

15

Percent (row): PE, allele

	Bm.	BM	b.m.	Total
PE	23	27	50	100
Normal	13	28	59	100
Total	14	28	57	100

20

Contingency table analytical statistics:

PE, allele

Number of missing values	241
Degree of freedom	2
Chi-square value	3.040
Chi-square p value	.2187
G-square value	2.765
G-square p value	.2510
Contingency table analytical coefficient	.108
Cramer's V value	.109

5

Frequency of observation: BA, allele

	Bm.	BM	b.m.	Total
BA	2	14	48	64
Normal	27	60	125	212
Total	29	74	173	276

10

Percent (row): BA, allele

	Bm.	BM	b.m.	Total
BA	3	22	75	100
Normal	13	28	59	100
Total	11	27	63	100

Contingency table analytic statistics:

BA, allele

Number of missing values	221
Degree of freedom	2
Chi-square value	7.096
Chi-square p value	.0288
G-square value	8.264
G-square p value	.0160
Contingency table analytical coefficient	.158
Cramer's V value	.160

15

Allelic classification (2)

Since the association of the Bm alleles with diseases is considered as strong from the above description, the relation of the number of Bm type alleles with the number of alleles other than the Bm type was analyzed by paying special attention to the type.

In view of the respiratory three disease groups (DPB, BE and PE) belonging to the COPD, the frequency of occurrence of the Bm allele is definitely higher in the patient groups suffering from the respiratory three diseases than that in the normal humans when the patient groups suffering from the three respiratory disease groups are compared with normal humans, and a statistical significant difference was recognized in the deviation of the distribution ($p = 0.0002$). The association of the Bm type alleles with the onset of the respiratory three disease groups (DPB, BE and PE) belonging to the COPD was shown by the comparison described above.

Any of the respiratory three disease groups (DPB, BE and PE) belonging to the COPD had a higher frequency of occurrence than that in normal humans in view of each disease (BE; $p = 0.012$, DPB; $p = 0.0025$ and PE; $p = 0.0052$).

On the other hand, the frequency of occurrence of the Bm type was significantly lower than that in normal humans ($p = 0.049$).

Frequency of observation:

	Three diseases, Bm/NotBm		Total
	Bm	NotBm	
Normal	27	185	212
Respiratory 3 diseases	49	129	178
Total	76	314	390

25

Percent (row): Three diseases,

	Bm/NotBm		Total
	Bm	NotBm	
Normal	13	87	100
Respiratory 3 diseases	28	72	100
Total	19	81	100

Contingency table analytical statistics:

Three diseases, Bm/NotBm

Number of missing values	107
Degree of freedom	1
Chi-square value	13.494
Chi-square p value	.0002
G-square value	13.534
G-square p value	.0002
Contingency table analytical coefficient	.183
Phi	.186
Chi-square value (Yates' continuity correction)	12.568
Chi-square p value (Yates' continuity correction)	.0004
Fisher's direct method p value	.0003

5

Frequency of observation:

DPB, Bm/NotBm

	Bm	NotBm	Total
Normal	27	185	212
DPB	17	41	58
Total	44	226	270

10

Percent (row):

DPB, Bm/NotBm

	Bm	NotBm	Total
Normal	13	87	100
DPB	29	71	100
Total	16	84	100

15

20

Contingency table analytical statistics:

DPB, Bm/NotBm

Number of missing values	227
Degree of freedom	1
Chi-square value	9.172
Chi-square p value	.0025
G-square value	8.202
G-square p value	.0042
Contingency table analytical coefficient	.181
Phi	.184
Chi-square value (Yates' continuity correction)	7.997
Chi-square p value (Yates' continuity correction)	.0047
Fisher's direct method p value	.0045

Frequency of observation:

BE, Bm/NotBm

	Bm	NotBm	Total
BE	19	57	76
Normal	27	185	212
Total	46	242	288

Percent (row): BE, Bm/NotBm

	Bm	NotBm	Total
BE	25	75	100
Normal	13	87	100
Total	16	84	100

5

10

15

Contingency table analytical statistics:

BE, Bm/NotBm

Number of missing values	209
Degree of freedom	1
Chi-square value	6.270
Chi-square p value	.0123
G-square value	5.824
G-square p value	.0158
Contingency table analytical coefficient	.146
Phi	.148
Chi-square value (Yates' continuity correction)	5.389
Chi-square p value (Yates' continuity correction)	.0203
Fisher's direct method p value	.0172

Frequency of observation:

5

PE, Bm/NotBm

	Bm	NotBm	Total
PE	13	31	44
Normal	27	185	212
Total	40	216	256

Percent (row): PE, Bm/NotBm

10

	Bm	NotBm	Total
PE	30	70	100
Normal	13	87	100
Total	16	84	100

15

Contingency table analytical statistics:

PE, Bm/NotBm

Number of missing values	241
Degree of freedom	1
Chi-square value	7.810
Chi-square p value	.0052
G-square value	6.802
G-square p value	.0091
Contingency table analytical coefficient	.172
Phi	.175
Chi-square value (Yates' continuity correction)	6.587
Chi-square p value (Yates' continuity correction)	.0103
Fisher's direct method p value	.0104

Frequency of observation:

BA, Bm/NotBm

	Bm	NotBm	Total
BA	2	62	64
Normal	27	185	212
Total	29	247	276

Percent (row):

BA, Bm/NotBm

	Bm	NotBm	Total
BA	3	97	100
Normal	13	87	100
Total	11	89	100

5

10

15

20

Contingency table analytic statistics:

BA, Bm/NotBm	
Number of missing values	221
Degree of freedom	1
Chi-square value	4.829
Chi-square p value	.0280
G-square value	6.035
G-square p value	.0140
Contingency table analytical coefficient	.131
Phi	.132
Chi-square value (Yates' continuity correction)	3.861
Chi-square p value (Yates' continuity correction)	.0494
Fisher's direct method p value	.0340

Individual classification (1)

5 Since the association of the Bm type among the three haplotypes with the respiratory diseases is considered as especially deep from the analytical results of the allelic classification, the classification and analysis were made of individuals without the Bm type, individuals having one Bm type (hetero) and individuals having two Bm types (homo) by particularly
10 noticing the Bm type.

 As for the respiratory three disease groups (DPB, BE and PE) belonging to the COPD, a significant difference was observed in distribution in relation to the frequency of occurrence of the haplotypic classifications Bm-0.1 and 2 based on the Bm in comparison of the normal
15 humans with the patient groups suffering from the respiratory three diseases ($p = 0.0093$).

 A significant difference was observed in the deviation of the distribution of patients developing the DPB and PE with regard to each disease (DPB: $p = 0.0024$ and PE: $p = 0.0069$). There was the tendency
20 even in the BE ($p = 0.089$).

Frequency of observation:

Three diseases, Bm.

	Bm-0	Bm-1	Bm-2	Total
Normal	79	27	0	106
Respiratory 3 diseases	54	29	6	89
Total	133	56	6	195

5

Percent (row): Three diseases, Bm.

	Bm-0	Bm-1	Bm-2	Total
Normal	75	25	0	100
Respiratory 3 diseases	61	33	7	100
Total	68	29	3	100

10

Contingency table analytical statistics

Three diseases, Bm.

Number of missing values	63
Degree of freedom	2
Chi-square value	9.360
Chi-square p value	.0093
G-square value	.
G-square p value	.
Contingency table analytical coefficient	.214
Cramer's V value	.219

Frequency of observation: DPB, Bm.

	Bm-0	Bm-1	Bm-2	Total
Normal	79	27	0	106
DPB	17	9	3	29
Total	96	36	3	135

15

Percent (row): DPB, Bm.

	Bm-0	Bm-1	Bm-2	Total
Normal	75	25	0	100
DPB	59	31	10	100
Total	71	27	2	100

Contingency table analytical statistics:

DPB, Bm.

Number of missing values	123
Degree of freedom	2
Chi-square value	12.040
Chi-square p value	.0024
G-square value	.
G-square p value	.
Contingency table analytical coefficient	.286
Cramer's V value	.299

Frequency of observation: BE, Bm.

5

	Bm-0	Bm-1	Bm-2	Total
BE	23	14	1	38
Normal	79	27	0	106
Total	102	41	1	144

Percent (row): BE, Bm.

	Bm-0	Bm-1	Bm-2	Total
BE	61	37	3	100
Normal	75	25	0	100
Total	71	28	1	100

10

Contingency table analytical statistics:

BE, Bm.

Number of missing values	114
Degree of freedom	2
Chi-square value	4.834
Chi-square p value	.0892
G-square value	.
G-square p value	.
Contingency table analytical coefficient	.180
Cramer's V value	.183

15

Frequency of observation: PE, Bm.				
	Bm-0	Bm-1	Bm-2	Total
PE	14	6	2	22
Normal	79	27	0	106
Total	93	33	2	128

5

Percent (row): PE, Bm.				
	Bm-0	Bm-1	Bm-2	Total
PE	64	27	9	100
Normal	75	25	0	100
Total	73	26	2	100

Contingency table analytical statistics:

PE, Bm.	
Number of missing values	130
Degree of freedom	2
Chi-square value	9.957
Chi-square p value	.0069
G-square value	.
G-square p value	.
Contingency table analytical coefficient	.0269
Cramer's V value	.0279

10

Individual classification (2)

Analysis was made whether or not the Bm haplotype was possessed (Bm-0 vs. Bm-1.2).

15 As for the respiratory three disease groups (DPB, BE and PE) belonging to the COPD, the frequency of occurrence of individuals having the Bm in the respiratory three disease groups is higher than that in normal humans in comparison thereof with the normal humans, and a significant difference was observed in the deviation of distribution ($p = 0.039$). On the other hand, there were more individuals without the Bm
20 haplotype in BA vice versa, and a significant deviation was noted in the distribution as compared with that in the normal human group ($p = 0.037$).

Three diseases, Bm-0/Bm-1.2

	Bm-0	Bm-1.2	Total
Normal	79	27	106
Respiratory 3 diseases	54	35	89
Total	133	62	195

5

Percent (row): Three diseases,
Bm-0/Bm-1.2

	Bm-0	Bm-1.2	Total
Normal	75	25	100
Respiratory 3 diseases	61	39	100
Total	68	32	100

10

Contingency table analytical statistics:

Three diseases, Bm-0/Bm-1.2

Number of missing values	63
Degree of freedom	1
Chi-square value	4.282
Chi-square p value	.0385
G-square value	4.279
G-square p value	.0386
Contingency table analytical coefficient	.147
Phi	.148
Chi-square value (Yates' continuity correction)	3.670
Chi-square p value (Yates' continuity correction)	.0554
Fisher's direct method p value	.0453

15

Frequency of observation:

BA, Bm-0/Bm-1.2

	Bm-0	Bm-1.2	Total
BA	30	2	32
Normal	79	27	106
Total	109	29	138

5

Percent (row): BA, Bm-0/Bm-1.2

	Bm-0	Bm-1.2	Total
BA	94	6	100
Normal	75	25	100
Total	79	21	100

10

Contingency table analytical statistics:

BA, Bm-0/Bm-1.2

Number of missing values	120
Degree of freedom	1
Chi-square value	5.471
Chi-square p value	.0193
G-square value	6.641
G-square p value	.0100
Contingency table analytical coefficient	.195
Phi	.199
Chi-square value (Yates' continuity correction)	4.375
Chi-square p value (Yates' continuity correction)	.0365
Fisher's direct method p value	.0241

15

20

Frequency of observation:

DPB, Bm-0/Bm-1.2

	Bm-0	Bm-1.2	Total
Normal	79	27	106
DPB	17	12	29
Total	96	39	135

5

Percent (row): DPB, Bm-0/Bm-1.2

	Bm-0	Bm-1.2	Total
Normal	75	25	100
DPB	59	41	100
Total	71	29	100

10

Contingency table analytical statistics:

DPB, Bm-0/Bm-1.2

Number of missing values	123
Degree of freedom	1
Chi-square value	2.805
Chi-square p value	.0940
G-square value	2.674
G-square p value	.1020
Contingency table analytical coefficient	.143
Phi	.144
Chi-square value (Yates' continuity correction)	2.089
Chi-square p value (Yates' continuity correction)	.1484
Fisher's direct method p value	.1086

Frequency of observation:

BE, Bm-0/Bm-1.2

	Bm-0	Bm-1.2	Total
BE	23	15	38
Normal	79	27	106
Total	102	42	144

15

Percent (row): BE, Bm-0/Bm-1.2

	Bm-0	Bm-1.2	Total
BE	61	39	100
Normal	75	25	100
Total	71	29	100

5

Contingency table analytical statistics:

BE, Bm-0/Bm-1.2

Number of missing values	114
Degree of freedom	1
Chi-square value	2.654
Chi-square p value	.1033
G-square value	2.564
G-square p value	.1093
Contingency table analytical coefficient	.135
Phi	.136
Chi-square value (Yates' continuity correction)	2.020
Chi-square p value (Yates' continuity correction)	.1552
Fisher's direct method p value	.1444

Frequency of observation

PE, Bm-0/Bm-1.2

	Bm-0	Bm-1.2	Total
PE	14	8	22
Normal	79	27	106
Total	93	35	128

10

Percent (row): PE, Bm-0/Bm-1.2

	Bm-0	Bm-1.2	Total
PE	64	36	100
Normal	75	25	100
Total	73	27	100

15

Contingency table analytical statistics

PE, Bm-0/Bm-1. 2

Number of missing values	130
Degree of freedom	1
Chi-square value	1.088
Chi-square p value	.2969
G-square value	1.040
G-square p value	.3079
Contingency table analytical coefficient	.092
Phi	.092
Chi-square value (Yates' continuity correction)	.609
Chi-square p value (Yates' continuity correction)	4.353
Fisher's direct method p value	.3035

Individual classification (3)

5 Furthermore, a comparison of the frequency of occurrence (Bm-0.1 vs. Bm-2) was made between individuals having the Bm haplotype as the homo (BBmm; Bm-2) and individuals without the haplotype (Bm-0.1). As for the respiratory three disease groups (DPB, BE and PE) belonging to the COPD, a significant difference was observed in the frequency of occurrence
10 between the individuals having the Bm haplotype as the homo (BBmm) and individuals without the Bm haplotype in comparison of the respiratory three disease groups with the normal humans ($p = 0.021$), and all the six individuals having the Bm homo type (Bm-2) were affected by any of the respiratory three diseases (DPE, BE and PE) belonging to the COPD.
15 Three individuals were affected by the DPB and one thereof was affected by the BE. No human having the Bm homo type (Bm-2) was found in 106 normal humans and 32 patients suffering from the BA.

As for each disease, a statistical significant difference was observed in the deviation of the distribution of patients suffering from the DPB and
20 PE (DPB: $p = 0.0082$ and PE: $p = 0.029$).

Frequency of observation:

Three diseases, Bm-0.1/Bm-2

	Bm-0.1	Bm-2	Total
Normal	106	0	106
Respiratory	83	6	89
3 diseases			
Total	189	6	195

5

Percent (row):

Three diseases, Bm-0.1/Bm-2.

	Bm-0.1	Bm-2	Total
Normal	100	0	100
Respiratory	93	7	100
3 diseases			
Total	97	3	100

10

Contingency table analytical statistics:

Three diseases, Bm-0.1/Bm-2

Number of missing values	63
Degree of freedom	1
Chi-square value	7.373
Chi-square p value	.0066
G-square value	.
G-square p value	.
Contingency table	.191
analytical coefficient	.194
Phi	5.295
Chi-square value (Yates' continuity correction)	
Chi-square p value (Yates' continuity correction)	.0214
Fisher's direct method p value	.0082

15

Frequency of observation:

DPB, Bm-0.1/Bm-2

	Bm-0.1	Bm-2	Total
Normal	106	0	106
DPB	26	3	29
Total	132	3	135

5

Percent (row): DPB, Bm-0.1/Bm-2

	Bm-0.1	Bm-2	Total
Normal	100	0	100
DPB	90	10	100
Total	98	2	100

Contingency table analytical statistics:

10

DPB, Bm-0.1/Bm-2

Number of missing values	123
Degree of freedom	1
Chi-square value	11.215
Chi-square p value	.0008
G-square value	.
G-square p value	.
Contingency table analytical coefficient	.277
Phi	.288
Chi-square value (Yates' continuity correction)	6.987
Chi-square p value (Yates' continuity correction)	.0082
Fisher's direct method p value	.0091

Frequency of observation:

PE, Bm-0.1/Bm-2

15

	Bm-0.1	Bm-2	Total
PE	20	2	22
Normal	106	0	106
Total	126	2	128

Percent (row):

PE, Bm-0.1/Bm-2

	Bm-0.1	Bm-2	Total
PE	91	9	100
Normal	100	0	100
Total	98	2	100

5

Contingency table analytical statistics:

PE, Bm-0.1/Bm-2

Number of missing values	130
Degree of freedom	1
Chi-square value	9.789
Chi-square p value	.0018
G-square value	.
G-square p value	.
Contingency table analytical coefficient	.267
Phi	.277
Chi-square value (Yates' continuity correction)	4.771
Chi-square p value (Yates' continuity correction)	.0289
Fisher's direct method p value	.0284

Frequency of observation:

BE, Bm-0.1/Bm-2

10

	Bm-0.1	Bm-2	Total
BE	37	1	38
Normal	106	0	106
Total	143	1	144

Percent (row): BE, Bm-0.1/Bm-2

	Bm-0.1	Bm-2	Total
BE	97	3	100
Normal	100	0	100
Total	99	1	100

15

Contingency table analytical statistics:

BE, Bm-0.1/Bm-2

Number of missing values	114
Degree of freedom	1
Chi-square value	2.809
Chi-square p value	.0937
G-square value	.
G-square p value	.
Contingency table analytical coefficient	.138
Phi	.140
Chi-square value (Yates' continuity correction)	.289
Chi-square p value (Yates' continuity correction)	.5909
Fisher's direct method p value	.2639

Frequency of observation:

BA, Bm-0.1/Bm-2

	Bm-0.1	Bm-2	Total
BA	32	0	32
Normal	106	0	106
Total	138	0	138

Percent (row):

BA, Bm-0.1/Bm-2

	Bm-0.1	Bm-2	Total
BA	100	0	100
Normal	100	0	100
Total	100	0	100

The above results definitely show that the Bm type haplotypes are associated with chronic respiratory tract inflammations in respiratory diseases according to a certain mechanism by analyzing the intron genetic polymorphisms of the human trypsin-like enzyme of the respiratory tract. Furthermore, it is also shown that the association of the human trypsin-like enzyme of the respiratory tract with diseases is different between the three diseases of DPB, BE and PE belonging to the chronic obstructive

pulmonary diseases (COPD) and the BA.

Since all the individuals having a certain genetic polymorphism do not develop some diseases, the genetic polymorphisms of the human trypsin-like enzyme of the respiratory tract are not a decisive onset factor
5 such as the so-called genetic disease and may safely be said as a readily onsetting factor. That is, when an environmental factor or the like is added to individuals having the Bm haplotypes, the individuals are susceptible to the onset of the respiratory diseases such as the BE, PE and DPB.

10 It is shown from the above results that diseases associated with the human trypsin-like enzyme of the respiratory tract can be classified by using the genetic polymorphisms of the human trypsin-like enzyme of the respiratory tract. The analytical method for the genetic polymorphisms of the human trypsin-like enzyme of the respiratory tract is a means
15 applicable to the prediction of onset constitution of diseases associated with the human trypsin-like enzyme of the respiratory tract in individual humans, the prediction of effects on the treatment of the diseases, prediction of the possibility for relapse of the prognosis thereof or the like.

20 Possibility of Industrial Utilization

The present invention provides a method for determining the disease-associated constitution of individual humans by the analyzing the genetic polymorphisms of the human trypsin-like enzyme of the respiratory
25 tract. Accordingly, when the diseases associated with the human trypsin-like enzyme of the respiratory tract can be identified by the analysis, it can be assumed that individuals having the certain genotype of the human trypsin-like enzyme of the respiratory tract are susceptible to some diseases.

30 That is, information about the methods of treatment for the individual humans can be provided in an early stage by predicting the disease onset constitution (individuals having the constitution susceptible to certain diseases) and related with early diagnosis and early treatment. The possibility for the relapse of the diseases can be estimated even after

the treatment. That is, physicians can pay careful attention to patients and provide proper direction by predicting the prognosis of the treatment (the course of curing after the treatment of patients and risk of relapse).

5 Furthermore, the genetic polymorphic analysis of the human trypsin-like enzyme of the respiratory tract has a possibility for providing a means for determining effects of drugs to be administered and narrowing the patient groups in which the administered drugs are effective in the development of the drugs in the morbid state associated with the human trypsin-like enzyme of the respiratory tract.

10 As described above, the early diagnosis, early treatment and direction of proper prophylactic methods, appropriate medication and proper afterfollow after the treatment are related to a reduction in huge medical expenditures causing problems at present.

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CLAIMS

1. A method for predicting the constitution susceptible to the onset of specific diseases in individual humans or effects of treatment on patients or prognosis after the treatment by analyzing polymorphisms of a human trypsin-like enzyme of a respiratory tract.

2. The method as set forth in claim 1, wherein the method for analyzing the genetic polymorphisms comprises the analysis of genotypes classified by detecting one or more base mutations.

3. The method as set forth in claim 1, the method for analyzing the genetic polymorphisms comprises the analysis of haplotypes classified by detecting the one or more base mutations.

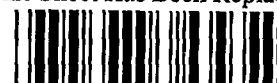
4. The method as set forth in claim 2 or 3, wherein sites for the analysis of genetic polymorphisms are introns.

5. The method as set forth in claim 2 or 3, wherein the sites for the analysis of the genetic polymorphisms are any of the following genetic fragments:

- (a) a genetic fragment containing an intron region amplifiable by using primers represented by sequence No. 7 and sequence No. 8.
- (b) a genetic fragment containing an intron region amplifiable by using primers represented by sequence No. 9 and sequence No. 10.
- (c) a genetic fragment containing an intron region amplifiable by using primers represented by sequence No. 11 and sequence No. 12.
- (d) a genetic fragment containing an intron region amplifiable by using primers represented by sequence No. 13 and sequence No. 14.
- (e) a genetic fragment containing an intron region amplifiable by using primers represented by sequence No. 15 and sequence No. 16.

6. The method as set forth in claim 2 or 3, wherein the sites for the analysis of the genetic polymorphisms are a genetic fragment containing intron C amplifiable by using the primers represented by the sequence No. 11 and the sequence No. 12.

7. The method as set forth in claim 2 or 3, wherein the sites for the analysis of the genetic polymorphisms are a genetic fragment containing intron A amplifiable by using the primers represented by the sequence No.





7 and the sequence No. 8.

8. The method as set forth in claim 6, wherein the sites for the analysis of the genetic polymorphisms in the intron C are the genetic fragment containing a sequential part recognizable by a restriction enzyme
5 BstUI, MboI, MseI or FbaI.

9. The method as set forth in claim 7, wherein the sites for the analysis of the genetic polymorphisms in the intron A are the genetic fragment containing a sequential part recognizable by a restriction enzyme MboI, TaqI or AfaI.

10 10. The method as set forth in claim 2 or 3, wherein the sites for the analysis of the genetic polymorphisms are a combination of the one or more sites for the genetic polymorphisms represented by sequence No. 17.

11. The method as set forth in any of claim 5 to claim 10, wherein the analytical method for genetic polymorphic sites are according to the
15 genotypic or haplotypic classification judged by cleavage with one or two or more restriction enzymes.

12. The method as set forth in claim 6, 8 or 10, wherein the analytical method for the genetic polymorphic sites in the intron C are according to the genotypic or haplotypic classification judged by cleavage
20 with the restriction enzyme BstUI, MboI, MseI or FbaI.

13. The method as set forth in claim 7 or 9, wherein the analytical method for the genetic polymorphic sites in the intron A are according to the genotypic or haplotypic classification judged by cleavage with the restriction enzyme MboI, TaqI or AfaI.

25 14. A method for predicting the constitution susceptible to the onset of specific diseases in individual humans or effects of treatment on patients or prognosis of the treatment which is a clinical diagnosis of the specific diseases by analyzing the genetic polymorphisms of the human trypsin-like enzyme of the respiratory tract.

30 15. The method as set forth in claim 1 or 14, wherein the specific diseases are respiratory diseases.

16. The method as set forth in claim 15, wherein the specific diseases are chronic obstructive pulmonary diseases.

17. The method as set forth in claim 15, wherein the specific

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diseases are sinobronchial syndrome.

18. The method as set forth in claim 15, wherein the specific diseases are pulmonary emphysema.

19. The method as set forth in claim 15, wherein the specific
5 diseases are diffuse panbronchiolitis.

20. The method as set forth in claim 15, wherein the specific diseases are bronchiectasis.

21. A method for diagnosing a mucociliary biophylactic system by analyzing the genetic polymorphisms of the human trypsin-like enzyme of
10 the respiratory tract.

22. A genetic fragment containing a part or all of the base sequence of the intron in the human trypsin-like enzyme of the respiratory tract.

23. Any of the following genetic fragments:

(a) a genetic fragment containing an intron region amplifiable by using
15 primers represented by the sequence No. 7 and the sequence No. 8.

(b) a genetic fragment containing an intron region amplifiable by using primers represented by the sequence No. 9 and the sequence No. 10.

(c) a genetic fragment containing an intron region amplifiable by using primers represented by the sequence No. 11 and the sequence No. 12.

20 (d) a genetic fragment containing an intron region amplifiable by using primers represented by the sequence No. 13 and the sequence No. 14.

(e) a genetic fragment containing an intron region amplifiable by using primers represented by the sequence No. 15 and the sequence No. 16.

24. A genetic fragment of an intron in a human trypsin-like enzyme
25 of the respiratory tract comprising a base sequence represented by any of the sequence No. 1 to the sequence No. 6 or a genetic fragment sandwiched between the base sequences.

25. A genetic fragment of an intron in a human trypsin-like enzyme of the respiratory tract comprising the base sequence represented by any of
30 the sequence No. 1 to the sequence No. 6.

26. A genetic fragment of intron C in a human trypsin-like enzyme of the respiratory tract comprising the base sequence represented by the sequence No. 17.

