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Case 100-7908

PEPTIDES

The present invention relates to variants of parathyroid hormone (PTH), a process for their production, pharmaceutical preparations comprising them and their use as a pharmaceutical.

The term "PTH" as used herein refers to any genetically encoded form of parathyroid hormone, including the mature form containing 84 amino acids of a given vertebrate PTH species, e.g human, porcine, rat, bovine, chicken, and fragments thereof as well as analogues and derivatives thereof having PTH-like activity. The position of each amino acid involved in the PTH sequence is numbered according to the internationally accepted procedure. For consistency and as is conventional, in the following description, the same numbering system will be applied to the amino acids of the PTH sequence independently of the substitution pattern in the molecule.

More particularly, the present invention provides a PTH compound having PTH-like activity and comprising at least one modification, said modification being either

1. at least one radical selected from a L- or D- α -amino acid, C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to the terminal amino group of the PTH compound, and/or at least one radical selected from C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl

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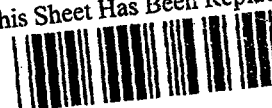


the alkyl, alkenyl, aralkyl, aralkenyl or cycloalkylalkyl group are hydroxy, amino and for the aryl moiety also halogen and/or C₁₋₄alkoxy. The alkyl group or alkyl moiety may be linear or branched and optionally interrupted by O, S or N. Preferably any C₁₋₈alkyl, C₂₋₈alkenyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl on the amino group of a PTH compound, is non-substituted. Examples for C₁₋₈alkyl are C₁₋₆alkyl, preferably methyl, ethyl, propyl, isopropyl, butyl, isobutyl, t.-butyl; for C₂₋₈alkenyl, C₂₋₄alkenyl, preferably allyl; for C₂₋₈alkynyl, C₂₋₄alkynyl, preferably prop-2-ynyl; for aralkyl, phenyl or benzyl; for aralkenyl, styryl; for C₃₋₆cycloalkyl-C₁₋₄alkyl, cyclohexyl-methyl. C₂₋₆alkoxycarbonyl is preferably formyl or acetyl. Suitable examples of D- or L- α -amino acids attached to the N-terminus include e.g. D- or L- Pro, Ala. Preferred substituents when present are alkyl, alkynyl, alkoxy-carbonyl or a D- or L- α -amino acid attached to the N-terminus of the PTH compound and/or at least one alkyl or alkoxy-carbonyl attached to one or more side chain amino groups.

Preferred PTH compounds of the invention are those comprising at least one amino acid unit replaced in one of the following positions of the PTH sequence: 1, 2, 3, 8-11, 13 to 19, 21, 22, 29 to 34, particularly 8-11, 16-19, 33 and/or 34. Further preferred PTH compounds of the invention are those comprising more than one amino acid unit replaced, particularly more than two amino acid units, more particularly more than three amino acid units, especially from 5 to 7 amino acid units replaced, preferably at any combination of the above mentioned positions of the PTH sequence.

In a series of specific or alternative embodiments, the present invention provides a PTH compound as disclosed above wherein in particular the α -amino acids in positions 1 and 2 at the amino terminus of the PTH sequence are replaced by a pseudo-dipeptide.

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The term "pseudo-dipeptide" as used herein refers to a dipeptide isostere in which the peptide bond between the 2 amino acid residues, whether natural or unnatural, is replaced with any isosteric group, e.g. $-\text{CH}_2-\text{NH}-$, $-\text{C}(\text{Halogen})=\text{CH}-$ or $-\text{C}(\text{alkyl})=\text{CH}-$.

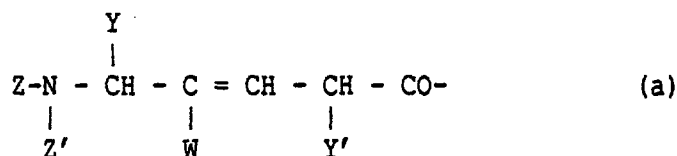
Examples of compounds of the invention wherein the α -amino acids in positions 1 and 2 are replaced by a pseudo-dipeptide include e.g compounds of formula I

X - P₁

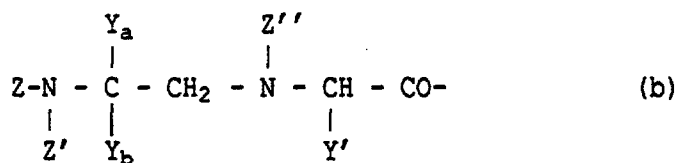
I

wherein

X is a residue of formula (a)



or (b)



wherein each of Z and Z', independently, is H; optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl; or a protecting group, at most one of Z and Z' being a protecting group, Z'' is H, C₁₋₈alkyl or a protecting group each of Y and Y', independently, is an optionally protected side chain of a natural or unnatural α -amino acid, one of Y_a and Y_b is hydrogen and the other is an optionally protected side chain of a natural or unnatural α -amino acid or Y_a and Y_b form together with the carbon atom to which they are attached a C₃₋₆cycloalkyl group,

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and

W is halogen or C₁₋₄alkyl,

or W and Y' together with the moiety ---C=CH-CH-

to which they are attached, form an optionally substituted aromatic cyclic or heterocyclic residue, and

P₁ is a PTH sequence, as defined above, in which amino acid units in positions 1 and 2 of the N-terminus are omitted, in free form or in salt or complex form.

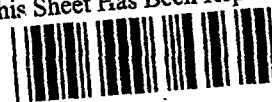
In compounds of formula I, P₁ may be a fragment (3-z) of PTH wherein z is 84 or an integer from 27 to 38, preferably from 34 to 38, particularly 34, 36, 37 or 38, especially 34 or 36. The PTH sequence represented by P₁ may correspond either to a naturally occurring PTH sequence, or to a naturally occurring PTH sequence wherein at least one α -amino acid unit has been replaced by a natural or unnatural amino acid optionally in protected form and/or one or more α -amino acid units are also omitted. P₁ may also comprise at least one side chain amino group substituted as disclosed above.

Halogen is preferably fluorine or chlorine.

In the residue of formula (a), the double bond has preferably the configuration trans (E).

As residue attaching to the α -carbon atom of a natural α -amino acid, Y, Y', Y_a or Y_b may be the side chain present in a natural α -amino acid as listed above, e.g. as present in Gly (i.e. H), Ala, Val, Ser, Leu, Ile, Phe, Trp. Y, Y', Y_a or Y_b may also be the residue attaching to the α -carbon atom of an unnatural α -amino acid as indicated above, e.g. as present in Nva, Orn, Abu, Aib, Nle. If the side chain of the natural or unnatural amino acids as Y, Y', Y_a or Y_b includes heteroatoms such as O, S or N, the heteroatoms on the side chain may optionally be pro-

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tected with an O-, S- or N-protecting group, e.g. as indicated above.

Protecting groups as Z or Z' or Z" may be as disclosed above.
C₁₋₈alkyl as Z or Z' or Z" may be as indicated above.

Preferably Z or Z' is hydrogen or C₁₋₄alkyl, particularly hydrogen or methyl.

Preferably Y is H or CH₃.

When W is C₁₋₄alkyl, it is preferably CH₃.

When W and Y' together with the moiety to which they are attached form an optionally substituted aromatic cyclic or heterocyclic residue, it may be an aromatic 5- or 6-membered cyclic residue comprising optionally 1 or 2 heteroatoms selected from N, S and O, e.g. phenyl, imidazolyl, pyridyl, oxazolyl or thiazolyl. A preferred substituent is hydroxy or methoxy, particularly for the phenyl ring.

Y' is preferably H, CH₃, isopropyl or benzyl.

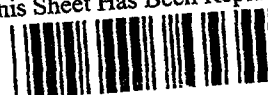
When Z" is C₁₋₈alkyl, it is preferably CH₃, C₂H₅ or isopropyl.

C₃₋₆cycloalkyl as Y_a and Y_b together is preferably cyclopropyl or cyclopentyl.

When Z" is a protecting group, it is preferably acyl, particularly acetyl.

In a series of specific and alternative embodiments, the present invention provides a PTH compound as disclosed above wherein the α -amino acid unit in position 1 and/or the α -amino acid unit in

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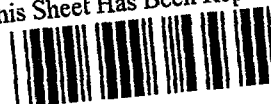
position 2 of the N-terminus of the PTH sequence is replaced by an optionally protected natural or unnatural amino acid residue, provided that

when the PTH compound is free from D- or L- α -amino acid attached to the N-terminus or from C₂₋₆alkoxycarbonyl or optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl, it is other than a PTH compound having a naturally occurring α -amino acid sequence;

or the PTH compound is other than PTH(1-34) wherein

- i. the α -amino acid in position 1 is Aib, Gly, D-Ser, D-Ala or Tyr; or
- ii. the α -amino acid in position 2 is Ala, D-Val, Lys, Cit or Arg and the α -amino acid in position 34 is Tyr; or the α -amino acid in position 2 is D-Val and the α -amino acid in position 34 is D-Tyr and optionally the α -amino acids in positions 8 and 18 are each Nle; or
- iii. the α -amino acid in position 1 is Aib and at least one further α -amino acid unit has been replaced as follows: the α -amino acid in position 8 and/or 18 is Leu, Ile, Val, Phe or Trp, and/or the α -amino acid in position 11 is Ser, Lys, Phe, β -Nal, Trp or Tyr, and/or the α -amino acid in position 12 is D-Leu, D-Ile, D-Nle, D-Val, D-Ser, D-Ser(Butyl), D-Abu, D-Thr, D-Nva, D-Met, D- β -Nal, D-Trp, D-Lys, D-Tyr, D-Lys(Fmoc), D-Phe or D-Asn, and/or the α -amino acid in position 13 is Leu, and/or the α -amino acid in position 19 and/or in position 21 is Arg, Lys, Asn or His, and/or the α -amino acid in position 23 is 2-(1,3-dithiolane-2-yl)Trp, and/or the α -amino acid in position 25 and/or in position 26 is His, and/or the α -amino acid in position 27 is Gln or Leu,

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or the PTH compound is other than PTH(1-84) wherein

- i. the α -amino acid in position 1 is Tyr, Val, Pro, Asp or Cys; or
- ii. the α -amino acid in position 2 is Ala, Glu, Leu, Ser or Arg.

Examples of compounds of the invention wherein the α -amino acids in position 1 and/or in position 2 are replaced as indicated above include e.g compounds of formula II



wherein

X_{1a} is a residue of an optionally protected natural α -amino acid or a residue of formula (IIa)

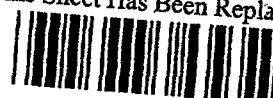


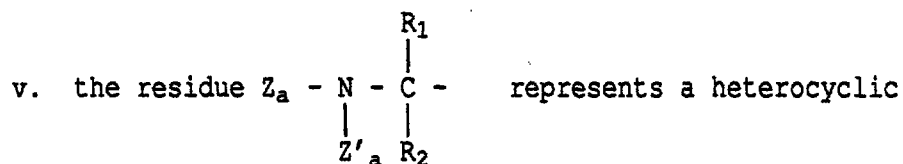
wherein

each of Z_a and Z'_a , independently, is H, C_{1-6} alkyl or a protecting group, at most one of Z_a and Z'_a being a protecting group, and either n is 1 and

- i. each of R_1 and R_2 , independently, is C_{1-6} alkyl, or
- ii. one of R_1 and R_2 is methyl or ethyl and the other is an optionally protected residue attaching to the α -carbon atom of a natural α -amino acid other than Ala, Leu, Ile or Val, or
- iii. one of R_1 and R_2 is H, methyl or ethyl and the other is an optionally protected residue attaching to the α -carbon atom of an unnatural α -amino acid, or
- iv. R_1 and R_2 form together with the carbon atom to which they are attached a C_{3-6} cycloalkyl group, or

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residue optionally condensed to a benzene ring,

or n is 2, 3, 4 or 5 and each of R_1 and R_2 is H or CH_3 ,

Y_{2a} is a residue of an optionally protected natural α -amino acid or a residue of formula (IIb)

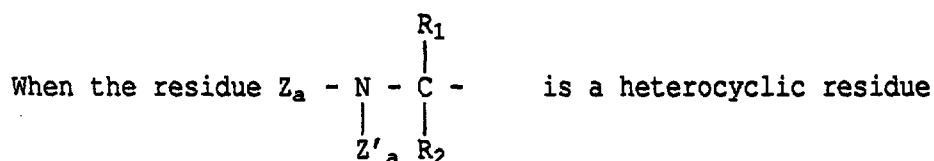


Z_b is H or C_{1-6} alkyl and either m is 1 and

- i. each of R_3 and R_4 , independently, is C_{1-6} alkyl, or
- ii. one of R_3 and R_4 is methyl or ethyl and the other is an optionally protected residue attaching to the α -carbon atom of a natural α -amino acid other than Ala, Leu, Ile or Val, or
- iii. one of R_3 and R_4 is H, methyl or ethyl and the other is an optionally protected residue attaching to the α -carbon atom of an unnatural α -amino acid,

or m is 2, 3, 4 or 5 and each of R_3 and R_4 is H or CH_3 ,
and

P_1 is as defined above.



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optionally condensed to a benzene ring, it may be a residue derived from a saturated or unsaturated 5- or 6-membered heterocycle comprising one or more heteroatoms selected from N, O and S. Examples of such heterocyclic residues include e.g. 2-pyridyl, 3-morpholinyl, hexahydropyridazinyl, 2- or 3-indolyl.

When n or m is > 1, the moiety CR_1R_2 or CR_3R_4 may be $-CH(CH_3)-CH_2-$, propyl, butyl or pentyl.

C_{3-6} cycloalkyl as R_1 and R_2 together is preferably cyclopropyl or cyclopentyl.

As residue attaching to the α -carbon atom of a natural α -amino acid, R_1 , R_2 , R_3 or R_4 may be the side chain present in a natural α -amino acid as listed above, e.g. as present in Gly, Ser or Thr. One of R_1 and R_2 or one of R_3 and R_4 may be methyl or ethyl and the other may be CH_3 , isopropyl, isobutyl or $CH_3-CH_2-CH(CH_3)-$. R_1 , R_2 , R_3 or R_4 may also be the residue attaching to the α -carbon atom of an unnatural α -amino acid as indicated above, e.g. as present in Abu, Gaba, Nva, Aib, TMSA or Nle. If the side chain of the natural or unnatural amino acids as R_1 , R_2 , R_3 or R_4 includes heteroatoms such as O, S or N, the heteroatoms on the side chain may optionally be protected with an O-, S- or N-protecting group, e.g. as indicated above. Protecting group as Z_a or Z'_a may be as disclosed above. C_{1-6} alkyl as Z_b is preferably methyl or ethyl. Preferably Z_b is H. C_{1-6} alkyl as Z_a or Z'_a is preferably straight chain or branched C_{1-4} alkyl. Preferably one of Z_a and Z'_a is H and the other is H, C_{1-4} alkyl or a protecting group, particularly H.

In a further or alternative embodiment, the present invention also provides a PTH compound as disclosed above wherein the α -amino acid unit in position 8 and/or the α -amino acid unit in

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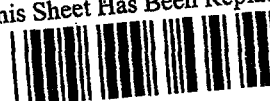
position 18 is replaced by an optionally protected natural or unnatural amino acid residue, provided that

when the PTH compound is free from D- or L- α -amino acid attached to the N-terminus or from C₂₋₆alkoxycarbonyl or optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl, it is other than a PTH compound having a naturally occurring α -amino acid sequence;

or the PTH compound is other than PTH(1-34) wherein

- i. the α -amino acids in positions 8 and 18 are each Nle or each Met(O) and optionally the α -amino acid in position 34 is Tyr; or the α -amino acids in positions 8 and 18 are each Nle and the α -amino acid in position 34 is Tyr and either the α -amino acid in position 12 is L- or D-Pro, L- or D-Ala, Aib or NMeGly or the α -amino acid in position 23 is Phe, Leu, Nle, Val, Tyr, α -Nal or β -Nal; or the α -amino acids in positions 8 and 18 are each Nle and the α -amino acid in position 2 is D-Val and the α -amino acid in position 34 is D-Tyr; or
- ii. the α -amino acid in position 8 and/or 18 is Leu, Ile, Val, Phe or Trp and optionally at least one further α -amino acid unit has been replaced as follows: the α -amino acid in position 1 is Aib and/or the α -amino acid in position 11 is Ser, Lys, Phe, β -Nal, Trp or Tyr, and/or the α -amino acid in position 12 is D-Leu, D-Ile, D-Nle, D-Val, D-Ser, D-Ser(Butyl), D-Abu, D-Thr, D-Nva, D-Met, D- β -Nal, D-Trp, D-Lys, D-Tyr, D-Lys(Fmoc), D-Phe or D-Asn, and/or the α -amino acid in position 13 is Leu, and/or the α -amino acid in position 19 and/or in position 21 is Arg, Lys, Asn or His, and/or the α -amino acid in position 23 is 2-(1,3-dithiolane-2-yl)Trp, and/or the α -amino acid in

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position 25 and/or in position 26 is His, and/or the α -amino acid in position 27 is Gln or Leu; or

iii. the α -amino acid in position 8 and/or 18 is Ala or Ser; or the α -amino acid in position 8 and/or 18 is Leu, and the α -amino acid in position 34 is Tyr; or

the PTH compound is other than PTH(1-84) wherein

the α -amino acid in position 8 is Met, Met(O), Ala, Val, Leu, Ile, Ser, Trp, Asn, Gln, Asp, Glu, Lys, Arg, Tyr or Gly and the α -amino acid in position 18 is Leu; or the α -amino acid in position 8 and in position 18 are each Met(O); or the α -amino acid in position 8 is Leu and the α -amino acid in position 18 is Met(O); or

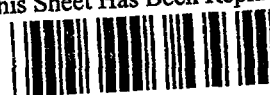
the PTH compound is other than [Leu¹⁸, Tyr³⁴]hPTH(1-84), [Leu⁸, Tyr³⁴]hPTH(1-84), [Ile⁸, Leu¹⁸, Tyr³⁴]hPTH(1-84) or [Leu⁸, Leu¹⁸, Tyr³⁴]hPTH(1-84).

When the α -amino acid unit in position 8 is replaced by an unnatural amino acid, it may be an unnatural amino acid as disclosed above e.g. Nva, Nle or Cha. Preferably it is an unnatural lipophilic amino acid preferably such containing a straight chain alkyl residue.

Nva (norvaline) and its homologues are particularly preferred. The unnatural amino acid in position 8 of the PTH sequence may also be a C α -methylated or C α -ethylated natural α -amino acid.

Examples of PTH compounds of the invention wherein the α -amino acid unit in position 18 is replaced, are e.g. PTH compounds wherein the α -amino acid residue in position 18 is replaced by a natural amino acid residue such as Gln, Tyr or Lys, for example [Gln¹⁸]-PTH, [Lys¹⁸]-PTH and [Tyr¹⁸]-PTH, particularly [Gln¹⁸ or Lys¹⁸ or Tyr¹⁸]-hPTH-(1-x) wherein x is 84 or an integer from 27 to 38, particularly from 34 to 38. Ala in position 18 is also

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preferred particularly when the PTH sequence is a (1-36) PTH fragment. In these compounds, the amino acid in position 8 may be Met or it may be replaced by an amino acid selected from Leu, Ile, Val, Phe, Gln, Trp, Ser and Ala or it may be an unnatural lipophilic amino acid residue, e.g. Nva or Nle.

Another group of such PTH compounds of the invention comprises e.g. PTH compounds wherein the amino acid residue in position 8 is replaced by an unnatural lipophilic amino acid residue, e.g. Nle or more preferably Nva. In these compounds the amino acid in position 18 may be Met or it may be replaced by an amino acid selected from Leu, Ile, Val, Phe, Trp, Ser, Ala, Gln, Lys or Tyr, preferably Leu, Gln, Ala or Tyr.

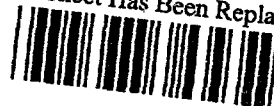
A further group of such PTH compounds of the invention are those wherein one or more amino acid units in the remaining positions, e.g. 1 to 7, 9 to 17 and/or 19 to 38 are either omitted or replaced in addition to the replacement of the α -amino acid unit in position 8 and/or 18. In such a case, the α -amino acid in position 8 is preferably replaced by Leu and the α -amino acid in position 18 by Gln, Leu, Ala or Tyr.

In a yet further or alternative embodiment, the present invention also provides a PTH compound as disclosed above wherein at least one of the α -amino acid unit of the PTH sequence is replaced by the α -amino acid unit which is present at the corresponding position in PTHrP,

provided that

when the PTH compound is free from D- or L- α -amino acid attached to the N-terminus or from C₂₋₆alkoxycarbonyl or optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl, it is other than a PTH compound having a naturally occurring α -amino acid sequence;

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or the PTH compound is other than PTH(1-34) wherein

- i. the α -amino acid in position 8 and/or 18 is Leu; and/or the α -amino acid in position 11 is Lys; and/or the α -amino acid in position 19 and/or 21 is Arg; and/or the α -amino acid in position 25 and/or 26 is His; and/or the α -amino acid in position 27 is Leu; or
- ii. the α -amino acid in position 25 is Gln and/or the α -amino acid in position 26 is Asn, the α -amino acid in position 27 being optionally Leu; or

the PTH compound is other than [Leu⁸,Leu¹⁸]PTH(1-84).

The term "PTHrP" refers to any genetically encoded form of PTHrP, e.g. human, chicken, rat or mouse PTHrP. For consistency and as is conventional, in the following description, the same numbering system will be applied to the amino acids of the PTHrP sequence starting with Ala in position 1 and comprising e.g. Ala in position 38.

This group of compounds according to the invention are hybrids between PTH and the homologous PTHrP sequences (referred to herein as PTH-rP hybrids). The amino acid sequence wherein the substitution according to the invention takes place is from position 1 up to position 38, particularly 1 up to 36, especially 1 up to 34.

Preferred PTH compounds of the invention are PTH-rP hybrids wherein more than one α -amino acid unit of the PTH sequence, e.g. at least 2, preferably from 3 to 5 α -amino acid units, are replaced according to the invention.

A preferred group of the PTH-rP hybrids according to the invention comprises PTH compounds wherein at least one α -amino acid unit of the PTH α -amino acid sequence in positions 8 to 11, i.e. Met⁸, His⁹, Asn¹⁰ or Leu¹¹ is replaced by the corresponding

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α -amino acid unit of the corresponding sequence of PTHrP, i.e. Leu⁸, His⁹, Asp¹⁰ or Lys¹¹. The replacement at positions 8 and/or 10 is preferred, particularly at positions 8 and 10. More preferably the α -amino acids at positions 8, 10 and 11 of said PTH sequence are replaced by Leu⁸, Asp¹⁰ and Lys¹¹, respectively.

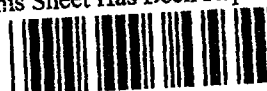
Another preferred group of the PTH-rP hybrids according to the invention comprises PTH compounds wherein at least one α -amino acid unit of the PTH α -amino acid sequence in positions 16 to 19 is replaced by the corresponding α -amino acid unit of the corresponding sequence of PTHrP, i.e. Gln¹⁶, Asp¹⁷, Leu¹⁸, Arg¹⁹. Preferably 3 or all 4 α -amino acids of said sequence are replaced. Particularly preferred are PTH compounds of the invention wherein the α -amino acid in position 16 is Gln, the α -amino acid in position 18 is Leu and the α -amino acid in position 19 is Arg.

A further preferred group of the PTH-rP hybrids according to the invention comprises PTH compounds wherein at least one α -amino acid unit of the PTH α -amino acid sequence in positions 33 and 34 is replaced by the corresponding α -amino acid unit of the corresponding sequence of PTHrP, i.e. Thr³³, Ala³⁴. Preferably both α -amino acids are replaced.

Yet further preferred groups of the PTH-rP hybrids according to the invention are PTH compounds comprising any combination of the above mentioned α -amino acid substitutions (8-11, 16-19, 33-34), more preferably a combination of substitutions selected from the above indicated 8-11 and 33-34 sequences.

As it will be appreciated, in the PTH-rP hybrids according to the invention one or more α -amino acids in positions 1 up to 38 may be further replaced by a natural or unnatural amino acid unit as indicated above or be omitted. In position 10 there may be a

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α -amino acid selected from Gly, Gln, Glu, His, Ser, Thr or Tyr. In position 13 there may be a D- or L- α -amino acid other than Arg, e.g. Ala, Cys, Gln, Ile, Asn, Trp, Asp, Val, Ser, Thr, Tyr, Met, Leu or Gly; in position 16 there may be a D- or L- α -amino acid, e.g. Lys, Ser, Leu, Ala, Gln or Gly; in position 17 Ala or Ser or preferably an amino acid having a bigger side chain than Ala or Ser, e.g. Glu, Gln, Phe, His, Ile or Lys; in position 18 Gln or Tyr; in position 19 Ala, Arg, Val, Tyr, Ser, Lys, Met, His, Gly, Pro, Asn or Ile; Gln or Arg in position 26; and/or in position 33 a D- or L- α -amino acid e.g. Ser, Thr, Leu, Gly, Gln, Arg, Pro, Asp, Ile, Lys, or Thr. They also may be substituted on the N-terminus or on a side chain amino group as indicated above. If desired, S- or O-containing side chains of the PTH-rP hybrids of the invention may also be protected as disclosed above.

Examples of preferred PTH-rP hybrids according to the invention are [Leu⁸, Gln¹⁸, Thr³³, Ala³⁴]PTH, [Leu⁸, Ala¹⁶, Gln¹⁸, Thr³³, Ala³⁴]PTH, [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸, Thr³³, Ala³⁴]PTH, [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸]PTH, [Leu⁸, Ala¹⁶, Gln¹⁸, Ala¹⁹]PTH, [Leu⁸, Asp¹⁰, Lys¹¹, Gln¹⁸]PTH, [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸, Ala¹⁹]PTH, [Leu⁸, Ala¹⁶, Gln¹⁸, Ala¹⁹, Thr³³, Ala³⁴]PTH and [Leu⁸, Asp¹⁰, Lys¹¹, Gln¹⁸, Thr³³, Ala³⁴]PTH.

Preferred PTH-rP hybrids according to the invention are PTH(1-x') compounds wherein x' is an integer from 34 to 38, particularly 34, 36, 37 or 38 especially hPTH(1-x'), wherein one or more α -amino acids are replaced by the α -amino acid(s) present at the corresponding position in PTHrP preferably as indicated above. When the positions 33 and 34 are replaced by the corresponding α -amino acid sequence of PTHrP, the PTH compound is preferably a hPTH fragment comprising 34 α -amino acid units. PTH-rP hybrids with carboxy terminus are preferred.

Preferably PTHrP is hPTHrP.

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The compounds of the invention may exist e.g. in free form, salt form or in the form of complexes thereof. Acid addition salts may be formed with e.g. organic acids, polymeric acids and inorganic acids. Such acid addition salt forms include e.g. the hydrochlorides and the acetates. Complexes are e.g. formed from the compound of the invention on addition of inorganic substances, e.g. inorganic salts or hydroxides such as Ca- and Zn-salts, and/or an addition of polymeric organic substances.

The present invention also provides a process for the production of the compounds of the invention. They may be prepared in a stepwise manner either in solution or using the solid phase synthesis process or genetic engineering.

The compounds of the invention may be produced for example as follows:

- a) removing at least one protecting group which is present in a compound of the invention in protected form; or
- b) linking together by an amide bond two peptide fragments, the peptide fragments being such that the desired amino acid sequence of the desired compound is obtained, and then effecting optionally stage a) of the process,
- c) for the production of a PTH compound wherein the α -amino acid units in positions 1 and 2 at the amino terminus are replaced by a pseudo-dipeptide, reacting a pseudo-dipeptide in protected or unprotected form with a PTH peptide in protected or unprotected form in which the amino acid residues in positions 1 and 2 are omitted and if necessary carrying out process step a); or

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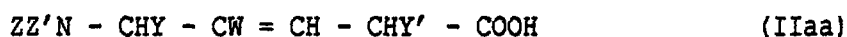
d) for the production of a PTH compound comprising at least one radical selected from D- or L- α -amino acid attached to the N-terminus and C₂₋₆alkoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl attached to the terminal amino group and/or to a side chain amino group, introducing such a radical in a PTH compound in protected or unprotected form and free from such a radical, and if necessary carrying out process step a);

and recovering the compounds thus obtained in free form, in salt form or in complex form.

Process steps a), b) and c) may be effected in analogy with known methods, e.g. as known in the art of peptide chemistry or as described in the following examples. Where desired, in these reactions, protecting groups which are suitable for use in peptides may be used for functional groups which do not participate in the reaction. The term protecting group may also include a polymer resin having functional groups.

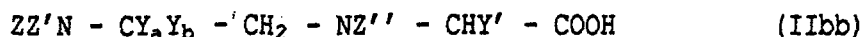
In process step b), for the production of a PTH compound wherein the α -amino acid units in positions 1 and 2 at the amino terminus are replaced by a pseudo-dipeptide, one peptide fragment used as starting material may comprise the pseudo-dipeptide at its N-terminus. This starting material may be prepared in accordance with process step c).

In process step c) the pseudo-dipeptide used as starting material may be a compound of formula IIaa or IIbb



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wherein Y, Y', Y_a, Y_b, Z, Z', Z'' and W are as defined above, or a functional derivative thereof, e.g. ester, acid halide or a symmetric or asymmetric anhydride. Compounds of formula IIaa wherein W and Y' together with the moiety $-C=CH-CH-$ to which they are attached, form an optionally substituted aromatic cyclic or heterocyclic residue, and compounds of formula IIbb wherein Z'' is alkyl or a protecting group, e.g. acetyl, or wherein $-CY_aY_b$ is C₃₋₆cycloalkyl and Z'' is H are novel and form part of the invention.

Process step d) may be carried out in analogy with known methods, e.g. alkylating methods. In a preferred embodiment of the invention, the alkylation is performed under reductive conditions, e.g. using a corresponding aldehyde or ketone. It may be performed for example in the presence of NaBH₃CN, preferably at an acidic pH, e.g. from 5 to 7. The temperature of the reaction may be e.g. from -20° C to 100° C. It may be advantageous to carry out the reaction in an inert solvent, e.g. water, an alcohol, dioxane or DMF or a mixture thereof. A D- or L-α-amino acid may also be attached to the N-terminus in accordance with known procedure.

The peptide fragment used as starting material in process step b) may comprise one or more unnatural amino acid units; it may also be substituted on the N-terminus and/or on a side chain amino group by a radical selected from C₂₋₆alkoxycarbonyl, optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl or it may bear a D- or L-α-amino acid on the N-terminus.

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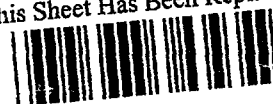
The compounds of the invention or fragments thereof comprising natural α -amino acid units may also be prepared using recombinant technology.

According to a preferred embodiment of the invention there is also provided a process for the production of a medium-sized polypeptide, in which a fusion protein comprising an N-terminal bacteriophage T4 gene 55 polypeptide having the desired polypeptide linked to the C-terminal thereof, is expressed in bacterial cells.

The gene 55 polypeptide may comprise the complete gp55 protein of bacteriophage T4 (188 amino acid residues) which is described in Gram, H. and Rüger, R; 1985; The EMBO Journal, 4, (1), pp 257-264. Alternatively a fragment, preferably an N-terminal fragment of the gp55 protein may be used. For example, a gp55 protein fragment comprising the first 112 N-terminal amino acid residues of the protein may be used. Typically, however, the gene 55 polypeptide comprises at least the first 25 N-terminal residues of the gp55 protein.

The medium-sized polypeptide may be from about 20 up to 100, e.g. 150, preferably from about 20 to about 50, amino acids in length. Examples of medium-sized polypeptides include calcitonins, endorphins, gastric inhibitory peptide(s), glucagon, neuropeptide Y, growth hormone releasing factor (GHRF), amyloid β protein fragments, hirudin, insulin, somatostatin, epidermal growth factor, nerve growth factor and PTH or fragments thereof. Particularly preferred medium-sized polypeptide which may be prepared according to the process of the present invention are the compounds of the invention or fragments thereof having natural amino acid substitutions.

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Conveniently the gene 55-PTH fusion protein also comprises a cleavable linker between the gene 55 polypeptide and the desired polypeptide. Preferably the cleavable linker is chemically cleavable and more preferably contains the amino acid sequences Asp-Pro-Pro or Asn-Gly-Pro.

In a further aspect the invention provides a nucleotide sequence coding for a fusion protein comprising an N-terminal bacteriophage T4 gene 55 polypeptide and a desired medium-sized polypeptide linked to the C-terminal thereof.

The nucleotide sequence is preferably a DNA sequence suitable for expression in bacteria. The sequence typically contains nucleotides coding for a cleavable linker between the gene 55 and desired polypeptide coding sequences.

The invention also provides a bacterial expression vector comprising a DNA sequence coding for the fusion protein.

The expression vector typically contains, in addition to the fusion protein coding sequence, appropriate expression control sequences including a suitable promoter, an operator and a ribosome binding site and other appropriate regulatory sequences. Usually, also the expression vector contains one or more selectable markers.

Any suitable promoter may be used, such as the TrpE, λ P1, λ Pr, lac or Tac promoter. Preferably the promoter is the bacteriophage T7 promoter and the expression vector is a plasmid. For E. coli expression an R1 or Col-E1 plasmid-derived vector may be used. For example, expression vector pET 17B, obtainable from Novagen, or similar expression vectors may be used.

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In another aspect the invention provides bacterial host cells transformed with a fusion protein coding sequence or an expression vector as described above. Any suitable bacterial host may be used, though preferably the host is E. coli. For example, E. coli strain BL21 (DE3) Lys E and similar strains may be used.

Advantageously the fusion protein accumulates within the host cells in the form of insoluble inclusion bodies, and thereby facilitates recovery of the fusion protein product from the cells. In order to recover the desired polypeptide product from the fusion protein, the protein of the inclusion bodies is solubilised, and the desired polypeptide product cleaved from the fusion protein. Any suitable solubilisation treatment may be used, including acid treatment, e.g. at about pH 2 to 4, preferably with acid at about pH 2.5, or treatment with a denaturing agent, e.g. a mild denaturing agent such as guanidine hydrochloride . In addition it may be necessary to remove one or more unwanted N-terminal amino acid residues from the product which remain after cleavage.

Thus in a further aspect the invention provides a process for the production of a desired medium-sized polypeptide, in a bacterial host, the process comprising:
causing transformed bacterial host cells as defined above to express the fusion protein in the form of inclusion bodies;
isolating the inclusion bodies;
solubilizing the inclusion bodies, and
cleaving the desired polypeptide from the fusion protein.

In the case of some medium-sized polypeptides, such as the PTH compounds, it is not normally necessary to carry out carefully-controlled denaturation and renaturation procedures to obtain PTH products in active form, e.g. having PTH-like activity. The medium-sized polypeptide products may be obtained in active form

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merely by neutralising the acid conditions used for solubilisation. Solubilisation and cleavage of the fusion protein may be carried out in a single step.

The invention also includes a fusion protein comprising an N-terminal bacteriophage T4 gene 55 polypeptide and, linked to the C-terminal thereof, a desired medium-sized polypeptide.

Using the processes and vectors of the invention it has been found that it is possible to produce high levels of medium-sized polypeptides, particularly PTH products. In E. coli, we have found that up to about 50% of the total protein expressed is the desired fusion protein. Also, the fusion protein forms inclusion bodies which are easy to separate from the other material of the cells. Moreover the inclusion bodies often consist of at least about 90% of the desired fusion protein which can significantly reduce the amount of purification that is required. Further, expressing the PTH product, in the form of the fusion protein can substantially decrease endogenous processing of the polypeptide in the bacterial cell, allowing homogenous products to be obtained.

Preferably the cleavable linker contains chemically cleavable amino acids adjacent the N-terminus of the desired polypeptide, to permit the desired polypeptide to be cleaved from the fusion protein by simple chemical means. Preferably the desired polypeptide does not contain the chemically cleavable groups.

Preferred chemically cleavable linkers contain the amino acids Asp-Pro-Pro or Asn-Gly-Pro. The Asp-Pro or Asn-Gly bonds may be chemically cleaved using acid conditions e.g. formic acid (normally about pH 2.5) or hydroxylamine respectively. The dipeptides Pro-Pro or Gly-Pro that remain on the N-terminus of the desired polypeptide may then be removed using a suitable

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dipeptidyl-peptidase. For example the L. lactis X-Pro dipeptidyl-peptidase EC 3.4.14.5 can be used. This peptidase is described in Nardi et al; 1991; Applied and Environmental Microbiology, 57, p45 to 50.

The desired polypeptide may be purified following cleavage from the fusion protein. HPLC purification techniques may be used. Purification may be carried out before removal of any unwanted N-terminal amino acid residues.

The process of the invention is further illustrated in the examples 308 to 314 Fig. 1 shows the amino acid sequence and corresponding DNA sequence of a truncated gp55-Asp-Pro-Pro-hPTH(1-38)OH fusion protein as cloned into the plasmid pET17B, and Fig. 2 shows the amino acid sequence and corresponding DNA sequence of a full length gp55-Asn-Gly-Pro-hPTH(1-38)OH fusion protein as cloned into the plasmid pET17B.

In the following Examples all temperatures are in °C. The following abbreviations are employed. In the present specification the term PTH without any further indication includes the carboxy terminus as well as a carboxamide terminus.

BOC	=	tert.Butyloxycarbonyl
DMF	=	dimethylformamide
DCM	=	dichloromethane
Fmoc	=	9-fluorenylmethoxycarbonyl
HOBt	=	1-hydroxybenzotriazole
TFA	=	trifluoroacetic acid
Trt	=	trityl = triphenylmethyl
RP-HPLC	=	reverse phase high performance liquid chromatography
r.t.	=	room temperature
DNP	=	Dinitrophenyl

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Tos	=	Tosyl
DIPC	=	Diisopropylcarbodiimide
Pmc	=	2,2,5,7,8-Pentamethylchroman-6-sulfonyl
Ip	=	Isopropyl
For	=	Formyl
Cpe	=	1-amino-cyclopentane-1-carboxylic acid
Cpp	=	1-amino-cyclopropane-1-carboxylic acid
Cha	=	cyclohexylalanine
tBu	=	t.butyl

EXAMPLE 1: N^ε-Isopropyl hPTH(1-34) NH₂

This peptide is assembled in a stepwise manner on a poly-styrene-based resin support. The Boc-group is used for protection of the alpha amino-group and side-chain functional groups are protected as follows: Lys(2-chlorobenzyloxycarbonyl), Ser(benzyl), Thr(benzyl), Glu(benzyl), Asp(benzyl), Met(O), His(DNP), Arg(Tos), and Trp(HCO). Other residues are left unprotected.

Amino-4-methylphenyl-methyl-co(polystyrene-divinylbenzene = MBHA-resin) (0,7 mmol/g) is subjected to the following cycle, steps (1) to (7), of treatments:

- (1) DCM
- (2) trifluoroacetic acid (50 %) in DCM
- (3) DCM
- (4) diisopropylethylamine (10 %) in DMF
- (5) DMF
- (6) preformed symmetrical anhydride (1.4 mmol per g starting resin) of Boc-amino acid in DMF
- (7) DMF

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Volumes of washes and reagents are from 5 to 20 ml per gram of starting resin. Each step is repeated as many times as necessary for either complete reaction of the resin (steps 2, 4, 6) or complete displacement of the previous reagent from the resin (steps 1, 3, 5, 7). Samples of resin are removed after each cycle and checked for completeness of the coupling reaction by a colorimetric test for residual amino groups using ninhydrin.

Symmetrical anhydrides of Boc-amino acids are prepared just prior to use by reacting Boc-amino acid (2,8 mmol per g resin) and DCCI (1,4 mmol per g resin) in DCM, containing DMF in amounts sufficient for complete dissolution of the Boc-amino acid. The mixture is filtered, more DMF is added to the filtrate which is concentrated by evaporation of the volatile components at a temperature not exceeding 15°C and the resulting solution is used in step (6).

The cycle of reactions, (1) to (7), is repeated for each amino acid residue such as to provide the sequence of the title compound except for Boc-Gln-OH and Boc-Arg(Tos)-OH, which are coupled in step (6) as their preformed 1-hydroxybenzo-triazole esters in DMF.

The fully assembled peptide resin is treated twice with 20 ml of thiophenol 5% in DMF at r.t. for 1 hour and washed with DMF, methanol and dichloromethane. The peptide resin is again subjected to steps (1) to (5) followed by the addition of 0.5 ml of acetone, 84 mg of sodium cyanoborohydride and 0.2 ml of acetic acid in 20 ml of DMF per gram of resin. After 16 hours at RT the resin is washed with DMF and dichloromethane and dried.

The peptide resin is treated with liquid HF, dimethyl sulfide, p-cresol and ethane-1,2-dithiol according to the 'low-high' procedure which is described, e.g., in International Journal of Peptide and Protein Research, vol.26, page 262-273 (1985). After removal of the volatile components and washing with ethylacetate

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the residue is extracted with several portions of 1%-acetic acid, filtered and the filtrate lyophilized. The lyophilized product is purified by repeated reversed-phase chromatography on a column of octadecyl-silica using a gradient of acetonitril in 2%-phosphoric acid or in 20 mM tetramethylammonium phosphate. Fractions containing the pure compound are combined, filtered through a weakly basic ion-exchange resin in the acetate form and the filtrate lyophilized to give the title compound.

MS (Ion-Spray): 4160.

EXAMPLE 2: [Lys (NE-Isopropyl)^{26,27}]hPTH(1-34) NH₂

The peptide is assembled as described for Example 1 but using Lys(Fmoc) for incorporation in position 13 and Fmoc-Ser(tBu) in terminal position 1.

The fully assembled peptide resin is treated twice with 20 ml of thiophenol 5% in DMF at r.t. for 1 hour and washed with DMF, methanol and dichloromethane. The dry resin is treated with liquid HF as described for Example 1. The cleavage product (350 mg) is suspended in a mixture of methanol (5 ml), phosphate buffer pH = 5.0 (5 ml), sodium cyanoborohydride (48 mg) and acetone (0.28 ml). After 16 hours at RT the reaction mixture is filtered and the filtrate evaporated. The residue is suspended in 20%-piperidine in DMF for 15 minutes, diluted with 20 volumes of ether and filtered. The residue is dissolved in 100 ml of water and acetic acid added to pH = 3, filtered and the filtrate lyophilized. The lyophilizate is purified by reversed-phase chromatography on a column of octadecyl-silica using a gradient of acetonitril in 2%-phosphoric acid. Fractions containing the pure compound are combined, filtered through a weakly basic ion-exchange resin in the acetate form and the filtrate lyophilized to give the title compound.

MS (Ion Spray): 4205.6.

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EXAMPLE 3: ~~NE~~-Isopropyl [Lys (~~NE~~-Isopropyl)]^{13, 26, 27}hPTH-(1-38)-OH**a) Solid phase peptide synthesis**

The peptide is synthesized in a stepwise manner on a polystyrene based resin support. The alpha-amino group is protected by Fmoc and the side-chain functional groups as followed: Asp(OtBu), Glu(OtBu), His(Trt), Lys(Boc), Asn(Trt), Gln(Trt), Arg(Pmc) and Ser(tBu). Other amino acids are left unprotected.

Fmoc-Gly esterified to 4-Hydroxymethyl-phenoxyethyl-co(polystyrene-1%-divinylbenzene), 0.5 mmol/g, is used as starting material for the stepwise solid phase synthesis, which consists of repetitive cycles of alpha-amino deprotection, washing, coupling (attachment of new amino acid) and washing. A three to five fold excess of Fmoc-amino acids is coupled as preformed HOBt-esters using DIPC.

Fmoc-Arg(Pmc), Fmoc-Asn(Trt) and Fmoc-Gln(Trt) are coupled as symmetrical anhydrides using DIPC. After complete assembly of the peptide chain the Fmoc-protecting group of Ser¹ is removed by 20% piperidine/DMF. Cleavage of the peptide from the polystyrene resin and removal of all side chain protecting groups is obtained by treatment of the peptide resin with a mixture of 10% ethanedithiol, thioanisole, thiocresole and water in 90% TFA for three hours at room temperature. The resin particles are filtered off and washed with some TFA. The product is precipitated from the combined filtrates by addition of ether, filtered and dried. The product is purified by chromatography on a C-18 silica column using a gradient of acetonitrile in 2% H₃PO₄/water. Fractions containing the pure compound are collected, filtered through an anion-exchange resin (Biorad, AG 4-X4, 100-200mesh acetate form) and lyophilized to give hPTH-(1-38)-OH.

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b) Alkylation

40 mg (9 μ mol) hPTH-(1-38)-OH (4458.2 g/mol) are dissolved in 1.2 ml methanol, 1.2 ml phosphate buffer (Merck no. 9887 adjusted to pH 5.0), 45 mg (716 μ mol) NaBH₃CN (62.84 g/mol) and 0.8 ml (11 mmol) acetone (73.52 ml/mol). The reaction is monitored by RP-HPLC on a C-18 silica column and finished after 15 hours at room temperature. The reaction mixture is diluted with water and chromatographed on a C-18 silica column using a gradient of acetonitrile in 2% H₃PO₄/water. Fractions containing the pure compound are collected, filtered through an anion-exchange resin (acetate form) and lyophilized to give the title compound as polyacetate, polyhydrate.

$$[\alpha]_D^{20} = -5.7^\circ \quad (c = 0.317 \text{ in } 95\% \text{ AcOH})$$

MS (Ion-Spray): 4626

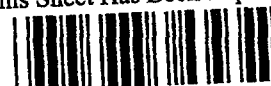
EXAMPLE 4: N^α-Methyl [Ala¹]PTH-(1-38)-OH

The peptide chain is assembled in the same manner as in example 3a. At position 1 instead of Fmoc-Ser(tBu)-OH the unnatural amino acid Fmoc-N^α-Methyl-Ala-OH is coupled to the peptide resin. Cleavage, deprotection and purification is performed as in example 3a.

MS (Ion Spray) = 4455, 91

EXAMPLE 5: [Ala¹,Ala³,Leu⁸,Gln¹³,Ala¹⁶,Gln¹⁸,Ala¹⁹,Phe²³,His²⁵,His²⁶,Leu²⁷,Thr³³,Ala³⁴]hPTH(1-34)OH

This peptide is prepared in analogy with the procedure of Example 3a. Instead of Fmoc-Gly esterified to 4-hydroxymethyl-phenoxy-methyl-co(polystyrene-divinylbenzene) the appropriate Fmoc-amino acid, e.g. Fmoc-Ala, Fmoc-Phe, Fmoc-D-Ala, Fmoc-D-Phe, etc. is used. Additionally the side-chain functional groups are protected as follows: Thr(t.-butyl), Trp(Boc) and Tyr(t.-butyl).



By repeating the procedure disclosed in Examples 3 and 5 but using the appropriate starting materials, the following compounds may be obtained:

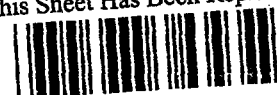
- EXAMPLE 6:** [Leu⁸, Asp¹⁰, Ala¹⁶, Gln¹⁸, Thr³³]hPTH(1-34)OH
- EXAMPLE 7:** [Ile¹]hPTH(1-38)OH (IS-MS: 4484)
- EXAMPLE 8:** [Ala¹, Abu²]hPTH(1-38)OH (IS-MS: 4428)
- EXAMPLE 9:** [Ala¹, Nva²]hPTH(1-38)OH (IS-MS: 4442)
- EXAMPLE 10:** [Ala¹, Ile²]hPTH(1-38)OH (IS-MS: 4456)
- EXAMPLE 11:** [Ala¹, Ala³, Leu⁸, Gln¹³, Ala¹⁶, Gln¹⁸, Ala¹⁹, His²⁶, Leu²⁷, Thr³³, Ala³⁴]hPTH(1-34)OH
- EXAMPLE 12:** [N-MeAla¹]hPTH(1-36)OH (IS-MS: 4286)
- EXAMPLE 13:** [Ala¹, Ala³, Leu⁸, Gln¹⁸]hPTH(1-36)OH (IS-MS: 4235)
- EXAMPLE 14:** [Thr¹]-hPTH-(1-38)OH (IS-MS: 4472)
- EXAMPLE 15:** [Leu¹]-hPTH-(1-38)OH (IS-MS: 4484)
- EXAMPLE 16:** [Abu¹]-hPTH-(1-38)OH (IS-MS: 4456)
- EXAMPLE 17:** [Gaba¹]-hPTH-(1-38)OH (IS-MS: 4456)
- EXAMPLE 18:** [Leu⁸, Lys¹¹, Gln¹⁸]hPTH(1-36)OH
- EXAMPLE 19:** [Leu⁸, Gln¹⁶, Asp¹⁷, Leu¹⁸, Arg¹⁹, Arg²²]hPTH(1-36)OH (IS-MS: 4347)
- EXAMPLE 20:** [Leu⁸, Gln¹⁸, Thr³³, Ala³⁴]-hPTH(1-34)OH (IS-MS: 4007)
- EXAMPLE 21:** [Leu⁸, Ala¹⁶, Gln¹⁸, Ala¹⁹, Thr³³, Ala³⁴]hPTH(1-34)OH (IS-MS: 3906)
- EXAMPLE 22:** [Leu⁸, Ala¹³, Gln¹⁸, Gln²⁶, Phe²⁷, Thr³³, Ala³⁴]hPTH(1-34)OH
- EXAMPLE 23:** Ip-[Leu⁸, Lys(Ip)¹³, Gln¹⁸, Lys(Ip)^{26, 27}, Thr³³, Ala³⁴]hPTH(1-34)OH (IS-MS: 4175)
- EXAMPLE 24:** Ip-[Leu⁸, Ala¹³, Gln¹⁸, Gln²⁶, Phe²⁷, Thr³³, Ala³⁴]hPTH(1-34)OH
- EXAMPLE 25:** [Gln¹⁶]hPTH(1-38)OH
- EXAMPLE 26:** [Ser¹⁴]hPTH(1-38)OH

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- EXAMPLE 27: [Ala¹,Ala³,Leu⁸,Gln¹³,Ala¹⁶,Gln¹⁸,Ala¹⁹,His²⁵,His²⁶,
Leu²⁷,Thr³³,Ala³⁴]hPTH(1-34)OH
- EXAMPLE 28: [Ala¹,Ala³,Leu⁸,Gln¹³,Ala¹⁶,Gln¹⁸,Ala¹⁹,Gln²⁶,Phe²⁷,
Thr³³,Ala³⁴]hPTH(1-34)OH
- EXAMPLE 29: [Leu⁸,Ala¹⁶,Gln¹⁸,Thr³³,Ala³⁴]hPTH(1-34)OH
(IS-MS: 3965)
- EXAMPLE 30: [Leu⁸,Gln¹⁸,Ala²⁹,Glu³⁰,Ile³¹]hPTH(1-34)OH
- EXAMPLE 31: [Leu⁸,Asp¹⁰,Lys¹¹,Gln¹⁸]hPTH(1-36)OH
- EXAMPLE 32: [Leu⁸,Asp¹⁰,Lys¹¹,Ser¹⁴,Ile¹⁵,Gln¹⁶,Asp¹⁷,Leu¹⁸,
Arg¹⁹]hPTH(1-36)OH
- EXAMPLE 33: [Leu⁸,Asp¹⁰,Lys¹¹,Leu¹⁸]hPTH(1-36)OH
- EXAMPLE 34: [Leu⁸,Gln¹⁶,Asp¹⁷,Leu¹⁸,Arg¹⁹]hPTH(1-36)OH
- EXAMPLE 35: [Leu⁸,Asp¹⁰,Lys¹¹,Ala¹⁷,Leu¹⁸]hPTH(1-36)OH (IS-MS:
4252)
- EXAMPLE 36: [Leu⁸,Gln¹⁶,Asp¹⁷,Leu¹⁸,Arg¹⁹,Thr³³,Ala³⁴]
hPTH(1-34)OH
- EXAMPLE 37: [Leu⁸,Asp¹⁰,Lys¹¹,Gln¹⁸,Thr³³,Ala³⁴]hPTH(1-34)OH
(IS-MS: 4022)
- EXAMPLE 38: [Leu⁸,Ala¹⁶,Asp¹⁷,Leu¹⁸,Ala¹⁹]hPTH(1-36)OH
- EXAMPLE 39: [Leu⁸,Asp¹⁰,Ala¹⁶,Asp¹⁷,Leu¹⁸,Ala¹⁹]hPTH(1-36)OH
- EXAMPLE 40: [Leu⁸,Gln¹⁸,Arg²²,Thr³³,Ala³⁴]hPTH(1-34)OH
- EXAMPLE 41: [Leu⁸,Asp¹⁰,Lys¹¹,Gln¹⁶,Asp¹⁷,Leu¹⁸,Arg¹⁹,Thr³³,
Ala³⁴]hPTH(1-34)OH (IS-MS: 4077)
- EXAMPLE 42: [Leu⁸,Asp¹⁰,Lys¹¹,Ala¹⁶,Gln¹⁸,Ala¹⁹]hPTH(1-36)OH
(IS-MS: 4181)
- EXAMPLE 43: [Leu⁸,Ala¹⁶,Asp¹⁷,Gln¹⁸,Ala¹⁹]hPTH(1-36)OH (IS-MS:
4193)
- EXAMPLE 44: [Leu⁸,Ala¹⁶,Ala¹⁷,Gln¹⁸,Ala¹⁹]hPTH(1-36)OH (IS-MS:
4149)
- EXAMPLE 45: [Leu⁸,Ala¹⁷,Gln¹⁸,Ala¹⁹]hPTH(1-36)OH
- EXAMPLE 46: [Leu⁸,Ala¹⁷,Gln¹⁸,Ala¹⁹,Arg²²]hPTH(1-36)OH (IS-MS:
4219)
- EXAMPLE 47: [Leu⁸,Ala¹⁷,Gln¹⁸,Ala¹⁹,Arg²²,Thr³³,Ala³⁴]
hPTH(1-34)OH (IS-MS: 3960)

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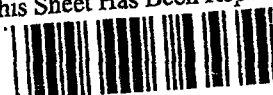
- EXAMPLE 48: [Leu⁸,Gln¹⁸]hPTH(1-36)OH
- EXAMPLE 49: [Leu⁸,Asp¹⁰,Lys¹¹,Ala¹⁶,Gln¹⁸,Thr³³,Ala³⁴]
hPTH(1-34)OH (IS-MS: 3980)
- EXAMPLE 50: [Leu⁸,Asp¹⁰,Lys¹¹,Ala¹⁶,Gln¹⁸]hPTH(1-36)OH
(IS-MS: 4240)
- EXAMPLE 51: [Leu⁸,Asp¹⁰,Lys¹¹,Ala¹⁶,Gln¹⁸,Ala¹⁹,Thr³³,Ala³⁴]
hPTH(1-34)OH (IS-MS: 3923)
- EXAMPLE 52: [Leu⁸,Asp¹⁰,Ala¹⁶,Gln¹⁸]hPTH(1-36)OH
(IS-MS: 4225)
- EXAMPLE 53: [Leu⁸,Asp¹⁰,Lys¹¹,Ala¹⁶,Gln¹⁸,Ala¹⁹,Thr³³]
hPTH(1-34)OH (IS-MS: 3999)
- EXAMPLE 54: [Leu⁸,Gln¹³,Ala¹⁶,Gln¹⁸,Ala¹⁹,His²⁶,Leu²⁷,Thr³³]
hPTH(1-34)OH
- EXAMPLE 55: [Leu⁸,Ala¹⁶,Gln¹⁸,Ala¹⁹,Arg²²]hPTH(1-36)OH
- EXAMPLE 56: [Ile¹⁵]hPTH(1-38)OH
- EXAMPLE 57: [Leu⁸,Gln¹³,Ala¹⁶,Gln¹⁸,Ala¹⁹,Arg²²,His²⁶,Leu²⁷,
Thr³³,Ala³⁴]hPTH(1-34)OH
- EXAMPLE 58: [Leu⁸,Gln¹³,Ala¹⁶,Gln¹⁸,Arg¹⁹,His²⁶,Leu²⁷,Thr³³,
Ala³⁴]hPTH(1-34)OH
- EXAMPLE 59: [Leu⁸,Ser¹³,Ala¹⁶,Gln¹⁸,Ala¹⁹,Arg²²]hPTH(1-36)OH
- EXAMPLE 60: [Leu⁸,Ala¹³,Ala¹⁶,Gln¹⁸,Ala¹⁹,Arg²⁶,Arg²⁷]
hPTH(1-36)OH
- EXAMPLE 61: [Leu⁸,Gln¹³,Ala¹⁶,Gln¹⁸,Ala¹⁹,His²⁶,Leu²⁷,Arg³³,
Ala³⁴]hPTH(1-34)OH
- EXAMPLE 62: [Leu⁸,Ala^{16,17,18,19}]hPTH(1-36)OH
- EXAMPLE 63: [Leu⁸,Ala¹³,Ala¹⁶,Gln¹⁸,Ala¹⁹,Gln²⁶,Phe²⁷]
hPTH(1-36)OH (IS-MS: 4127)
- EXAMPLE 64: Ip-[Leu⁸,Ala¹³,Ala¹⁶,Gln¹⁸,Ala¹⁹,Gln²⁶,Phe²⁷]
hPTH(1-36)OH
- EXAMPLE 65: Ip-[Leu⁸,Lys(Ip)¹³,Ala¹⁶,Gln¹⁸,Ala¹⁹,
Lys(Ip)^{26,27}]hPTH(1-36)OH
- EXAMPLE 66: [Leu⁸,Ala¹⁶,Gln¹⁸,Ala¹⁹]hPTH(1-36)OH (IS-MS: 4166)
- EXAMPLE 67: Ip-[Leu⁸,Lys(Ip)¹³,Ala¹⁶,Ala¹⁷,Ala¹⁸,Ala¹⁹,
Lys(Ip)^{26,27}]hPTH(1-36)OH

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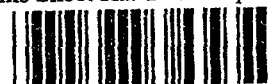
- EXAMPLE 68: [Aib³,Gln¹⁸]hPTH(1-36)OH (IS-MS: 4283)
- EXAMPLE 69: Ip-[Leu⁸,Asp¹⁰,Lys(Ip)^{11,13,26,27},Gln¹⁸]
hPTH(1-36)OH
- EXAMPLE 70: Ip-[Leu⁸,Ser¹³,Ala¹⁶,Gln¹⁸,Ala¹⁹,Arg²²,
Lys(Ip)^{26,27}]hPTH(1-36)OH
- EXAMPLE 71: [Leu⁸,Tyr¹⁸]hPTH(1-36)OH
- EXAMPLE 72: [Ser³³]-hPTH-(1-38)-OH
- EXAMPLE 73: [Thr³³]-hPTH-(1-38)-OH
- EXAMPLE 74: [Leu³³]-hPTH-(1-38)-OH
- EXAMPLE 75: [Gly³³]-hPTH-(1-38)-OH
- EXAMPLE 76: [Leu⁸,His¹⁰,Gln¹⁸,Arg²²,Thr³³,Ala³⁴]hPTH(1-34)OH
- EXAMPLE 77: [Leu⁸,Gly¹⁰,Gln¹⁸,Arg²²,Thr³³,Ala³⁴]hPTH(1-34)OH
- EXAMPLE 78: [Leu⁸,Glu¹⁰,Gln¹⁸,Arg²²,Thr³³,Ala³⁴]hPTH(1-34)OH
- EXAMPLE 79: [Leu⁸,Thr¹⁰,Gln¹⁸,Arg²²,Thr³³,Ala³⁴]hPTH(1-34)OH
- EXAMPLE 80: [Leu⁸,Gln¹⁰,Gln¹⁸,Arg²²,Thr³³,Ala³⁴]hPTH(1-34)OH
- EXAMPLE 81: [Gln³³]-hPTH-(1-38)-OH
- EXAMPLE 82: [Leu⁸,Tyr¹⁰,Gln¹⁸,Arg²²,Thr³³,Ala³⁴]hPTH(1-34)OH
- EXAMPLE 83: [1-amino-cyclopentane-1-carboxylic
acid¹,Leu⁸,Ala¹⁶,Gln¹⁸,Ala¹⁹]hPTH(1-36)OH
- EXAMPLE 84: [1-amino-cyclopentane-1-carboxylic acid¹,Leu⁸,
Ala^{13,16},Gln¹⁸,Ala¹⁹,Arg^{26,27}]hPTH(1-36)OH
- EXAMPLE 85: [1-amino-cyclopentane-1-carboxylic acid³,Gln¹⁸]
hPTH(1-36)OH (IS-MS: 4309)
- EXAMPLE 86: [Arg¹²]-hPTH-(1-38)-OH
- EXAMPLE 87: [Ser¹²]-hPTH-(1-38)-OH
- EXAMPLE 88: [Cys¹³]-hPTH-(1-38)-OH
- EXAMPLE 89: [Ile¹³]-hPTH-(1-38)-OH
- EXAMPLE 90: [Asn¹³]-hPTH-(1-38)-OH
- EXAMPLE 91: [Trp¹³]-hPTH-(1-38)-OH
- EXAMPLE 92: [Asp¹³]-hPTH-(1-38)-OH
- EXAMPLE 93: [Val¹³]-hPTH-(1-38)-OH
- EXAMPLE 94: [Thr¹³]-hPTH-(1-38)-OH
- EXAMPLE 95: [Ser¹³]-hPTH-(1-38)-OH

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EXAMPLE 96: [Tyr¹³]-hPTH-(1-38)-OH
EXAMPLE 97: [Met¹³]-hPTH-(1-38)-OH
EXAMPLE 98: [Gln¹³]-hPTH-(1-38)-OH
EXAMPLE 99: [Leu¹³]-hPTH-(1-38)-OH
EXAMPLE 100: [Ala¹³]-hPTH-(1-38)-OH
EXAMPLE 101: [Gly¹³]-hPTH-(1-38)-OH
EXAMPLE 102: [Val¹⁴]-hPTH-(1-38)-OH
EXAMPLE 103: [Ala¹⁴]-hPTH-(1-38)-OH
EXAMPLE 104: [Lys¹⁴]-hPTH-(1-38)-OH
EXAMPLE 105: [Arg¹⁴]-hPTH-(1-38)-OH
EXAMPLE 106: [Thr¹⁴]-hPTH-(1-38)-OH
EXAMPLE 107: [Ile¹⁴]-hPTH-(1-38)-OH
EXAMPLE 108: [Tyr¹⁴]-hPTH-(1-38)-OH
EXAMPLE 109: [Tyr¹⁵]-hPTH-(1-38)-OH
EXAMPLE 110: [Arg¹⁵]-hPTH-(1-38)-OH
EXAMPLE 111: [Val¹⁵]-hPTH-(1-38)-OH
EXAMPLE 112: [Lys¹⁶]-hPTH-(1-38)-OH
EXAMPLE 113: [Ser¹⁶]-hPTH-(1-38)-OH
EXAMPLE 114: [Leu¹⁶]-hPTH-(1-38)-OH
EXAMPLE 115: [Ala¹⁶]-hPTH-(1-38)-OH
EXAMPLE 116: [Gly¹⁶]-hPTH-(1-38)-OH
EXAMPLE 117: [Ala¹⁷]-hPTH-(1-38)-OH
EXAMPLE 118: [Met¹⁷]-hPTH-(1-38)-OH
EXAMPLE 119: [Ile¹⁷]-hPTH-(1-38)-OH
EXAMPLE 120: [Ser¹⁹]-hPTH-(1-38)-OH
EXAMPLE 121: [Lys¹⁹]-hPTH-(1-38)-OH
EXAMPLE 122: [Leu¹⁹]-hPTH-(1-38)-OH
EXAMPLE 123: [Ala¹⁹]-hPTH-(1-38)-OH
EXAMPLE 124: [Tyr¹⁹]-hPTH-(1-38)-OH
EXAMPLE 125: [Met¹⁹]-hPTH-(1-38)-OH
EXAMPLE 126: [His¹⁹]-hPTH-(1-38)-OH
EXAMPLE 127: [Val¹⁹]-hPTH-(1-38)-OH
EXAMPLE 128: [Gly¹⁹]-hPTH-(1-38)-OH
EXAMPLE 129: [Pro¹⁹]-hPTH-(1-38)-OH

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EXAMPLE 130: [Asp¹⁹]-hPTH-(1-38)-OH
EXAMPLE 131: [Ile¹⁹]-hPTH-(1-38)-OH
EXAMPLE 132: [Val¹⁹,Gln²⁴]-hPTH-(1-38)-OH
EXAMPLE 133: [Arg¹⁹]-hPTH-(1-38)-OH
EXAMPLE 134: [Phe²⁰]-hPTH-(1-38)-OH
EXAMPLE 135: [Ala²¹]-hPTH-(1-38)-OH
EXAMPLE 136: [Gly²¹]-hPTH-(1-38)-OH
EXAMPLE 137: [Phe²¹]-hPTH-(1-38)-OH
EXAMPLE 138: [Leu²¹]-hPTH-(1-38)-OH
EXAMPLE 139: [Asn²¹]-hPTH-(1-38)-OH
EXAMPLE 140: [Gln²¹]-hPTH-(1-38)-OH
EXAMPLE 141: [Ser²¹]-hPTH-(1-38)-OH
EXAMPLE 142: [Gly²²]-hPTH-(1-38)-OH
EXAMPLE 143: [Leu²²]-hPTH-(1-38)-OH
EXAMPLE 144: [His²²]-hPTH-(1-38)-OH
EXAMPLE 145: [Ala²²]-hPTH-(1-38)-OH
EXAMPLE 146: [Ile²²]-hPTH-(1-38)-OH
EXAMPLE 147: [Val²²]-hPTH-(1-38)-OH
EXAMPLE 148: [Ser²²]-hPTH-(1-38)-OH
EXAMPLE 149: [Arg²²]-hPTH-(1-38)-OH
EXAMPLE 150: [Arg²⁶]-hPTH-(1-38)-OH
EXAMPLE 151: [Val²⁷]-hPTH-(1-38)-OH
EXAMPLE 152: [Ile²⁷]-hPTH-(1-38)-OH
EXAMPLE 153: [Leu²⁷]-hPTH-(1-38)-OH
EXAMPLE 154: [Arg²⁷]-hPTH-(1-38)-OH
EXAMPLE 155: [Ala²⁷]-hPTH-(1-38)-OH
EXAMPLE 156: [Val²⁸]-hPTH-(1-38)-OH
EXAMPLE 157: [Ile²⁸]-hPTH-(1-38)-OH
EXAMPLE 158: [Pro³,Thr³³]-hPTH-(1-38)-OH
EXAMPLE 159: [Arg³³]-hPTH-(1-38)-OH
EXAMPLE 160: [Pro³³]-hPTH-(1-38)-OH
EXAMPLE 161: [Asp³³]-hPTH-(1-38)-OH
EXAMPLE 162: [Ile³³]-hPTH-(1-38)-OH
EXAMPLE 163: [Lys³³]-hPTH-(1-38)-OH

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EXAMPLE 164: [Ile³¹, Arg³³]-hPTH-(1-38)-OH

EXAMPLE 165: [1-aminocyclopentane-1-carboxylic acid¹]hPTH-
(1-36)NH₂

The peptide is assembled in a stepwise manner on a polystyrene based resin support. The Fmoc-group is used for protection of the alpha-amino groups.

Side-chain functional groups are protected as Arg(Pmc), Asn(Trt), Asp(OtBu), Gln(Trt), Glu(OtBu), His(Trt), Lys(Boc), Ser(tBu), Thr(tBu), Trp(Boc) and Tyr(tBu). Other amino acids are left unprotected.

4-(2',4'-Dimethoxyphenyl-Fmoc-amino-methyl)-phenoxy-co(polystyrene-divinylbenzene), 0.4 mmol/g, which may be prepared, e.g., as described in Tetrah. Letters, 28, 3787-3790 (1987) is subjected to the following cycle, steps (1) to (5), of treatments:

- (1) DMF
- (2) piperidine (20%) in DMF
- (3) DMF
- (4) mixture of HOBt, diisopropylcarbodiimide, and Fmoc-alanine (0.8 mmol per gram starting resin each)
- (5) DMF

Volumes of washes and reagents are from 5 to 20 mL per gram of starting resin.

In the next cycle of treatments (1) to (5), Fmoc-valine is substituted for Fmoc-alanine and so on for each cycle such as to assemble on the resin the correct amino acid sequence of the title compound.

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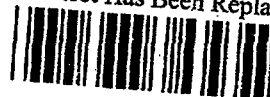
Each step is repeated as many times as necessary for either complete reaction of the resin (steps 2, 4) or complete displacement of the previous reagent(s) from the resin (steps 3, 5). Samples of the resin are removed after each cycle and checked for completeness of the coupling reaction by a colorimetric test for residual amino groups using ninhydrin.

At the end of the synthesis a final cycle comprising steps (1) to (3) only is performed and the peptide resin is washed with 2-propanol, then with a mixture of methanol and methylene chloride (1:1 v/v) and dried thoroughly in a vacuum desiccator.

The peptide resin (1 g) is suspended in a mixture (20 ml) of trifluoroacetic acid, water, and 1,2-ethanedithiol (90:5:5 v/v) for 2 hours at room temperature, the resin particles are filtered off and washed with a small volume of TFA. The product is precipitated from the combined filtrates by addition of ether (20 volumes), filtered, washed with more ether and dried. The residue is dissolved in 2 % acetic acid, the solution let to stand at r.t. for 8 hours prior to lyophilization. The lyophilisate is chromatographed on a C-18 silica column using a gradient of acetonitrile in 2% H_3PO_4 . Fractions are checked by analytical HPLC and those containing the pure compound are collected, filtered through an anion-exchange resin in the acetate form and lyophilised to give the title compound as a polyacetate, polyhydrate.

Fmoc-1-aminocyclopentane-1-carboxylic acid used in the preparation of the peptide resin intermediate may be prepared, e.g. as described by G. Valle et al., 1988, in Can.J.Chem. 66:2575-2582.
MS (Ion-Spray): 4312

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EXAMPLE 166: **[1-(1-aminocyclopropane-1-carboxylic acid)]hPTH-**
(1-36)NH₂

This peptide is prepared in analogy with the procedure of Example 165. Fmoc-1-aminocyclopropane-1-carboxylic acid used in the preparation of the peptide resin intermediate may be prepared, e.g., as described by M. Crisma et al. (1989) in Int.J.Biol.Macromol.11:345-352. MS(Ion-Spray): 4283

By repeating the procedure as disclosed in Example 165 but using the appropriate starting materials, following compounds may be obtained:

EXAMPLE 167: [D-Pro¹]hPTH(1-36)NH₂

EXAMPLE 168: [Nva¹]hPTH(1-36)NH₂

EXAMPLE 169: [N-Me-Ser¹]hPTH(1-36)NH₂

EXAMPLE 170: [Indole-2-carboxylic acid¹]hPTH(1-36)NH₂

EXAMPLE 171: [Indole-3-carboxylic acid¹]hPTH(1-36)NH₂

EXAMPLE 172: [Pyridine-3-carboxylic acid¹]hPTH(1-36)NH₂

EXAMPLE 173: [Pyridine-2-carboxylic acid¹]hPTH(1-36)NH₂

EXAMPLE 174: [Hexahydropyridazine-3-carboxylic
acid¹]hPTH(1-36)NH₂

EXAMPLE 175: [Morpholine-2-carboxylic acid¹]hPTH(1-36)NH₂

EXAMPLE 176: [Pro¹]hPTH(1-36)NH₂

EXAMPLE 177: [Leu¹]hPTH(1-36)NH₂

EXAMPLE 178: [Ile¹]hPTH(1-36)NH₂

EXAMPLE 179: [Thr³³,Ala³⁴]hPTH(1-34)NH₂

EXAMPLE 180: [Nva⁸]hPTH(1-36)NH₂

EXAMPLE 181: [Gln¹⁸]hPTH(1-36)NH₂

EXAMPLE 182: [Tyr¹⁸]hPTH(1-36)NH₂

EXAMPLE 183: [Lys¹⁸]hPTH(1-36)NH₂

EXAMPLE 184: [Ala¹⁸]hPTH(1-36)NH₂

EXAMPLE 185: [Phe²³,His²⁵,His²⁶,Leu²⁷]-hPTH(1-34)NH₂

EXAMPLE 186: [Phe²³]-hPTH(1-36)NH₂ (IS-MS: 4247)

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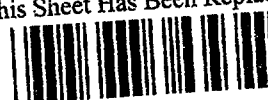
- EXAMPLE 187: [Phe²³, His²⁵, His²⁶, Leu²⁷, Ile²⁸, Ala²⁹, Glu³⁰, Ile³¹, Thr³³, Ala³⁴]-hPTH(1-34)NH₂ (IS-MS:3934)
- EXAMPLE 188: [Ala²⁹]-hPTH(1-36)NH₂ (IS-MS: 4248)
- EXAMPLE 189: [Ala³⁴]-hPTH(1-36)NH₂ (IS-MS: 4210)
- EXAMPLE 190: [Gln²⁵]hPTH(1-36)NH₂
- EXAMPLE 191: [Leu⁸, Asp¹⁰, Lys¹¹, Thr³³, Ala³⁴]hPTH(1-34)NH₂
- EXAMPLE 192: [Ala¹, His⁵, Leu⁸, Asp¹⁰, Lys¹¹, Thr³³, Ala³⁴]hPTH(1-34)NH₂
- EXAMPLE 193: [Ser¹⁴, Ile¹⁵, Gln¹⁶, Asp¹⁷, Leu¹⁸, Arg¹⁹, Thr³³, Ala³⁴]hPTH(1-34)NH₂
- EXAMPLE 194: [D-Phe³⁴, D-Ala³⁶]hPTH(1-36)-NH₂ [MS(IS): 4290]
- EXAMPLE 195: [Ala³]hPTH(1-36)NH₂ [MS(IS):4271]
- EXAMPLE 196: [D-Ser³]hPTH(1-36)NH₂ [MS(IS):4290]
- EXAMPLE 197: [D-Glu⁴]hPTH(1-36)NH₂ [MS(IS):4290]
- EXAMPLE 198: [D-His⁹]hPTH(1-36)NH₂ [MS(IS):4286]
- EXAMPLE 199: [Ala¹⁰]hPTH(1-36)NH₂ [MS(IS):4243]
- EXAMPLE 200: [D-Asn¹⁰]hPTH(1-36)NH₂ [MS(IS): 4288]
- EXAMPLE 201: [Ala¹²]hPTH(1-36)NH₂ [MS(IS): 4299]
- EXAMPLE 202: [Gln¹³]hPTH(1-36)NH₂ [MS(IS): 4287]
- EXAMPLE 203: [His¹³]hPTH(1-36)NH₂ [MS(IS): 4296]
- EXAMPLE 204: [Leu¹³]hPTH(1-36)NH₂ [MS(IS): 4371]
- EXAMPLE 205: [Ala¹³]hPTH(1-36)NH₂ [MS(IS): 4228]
- EXAMPLE 206: [Ala¹³, Gln²⁶, Phe²⁷, D-Phe³⁴, D-Ala³⁶]hPTH(1-36)NH₂
[MS(IS): 4249]
- EXAMPLE 207: [Ala¹⁴]hPTH(1-36)NH₂ [MS(IS): 4219]
- EXAMPLE 208: [D-His¹⁴]hPTH(1-36)NH₂ [MS(IS): 4288]
- EXAMPLE 209: [D-Asn¹⁶]hPTH(1-36)NH₂ [MS(IS): 4284]
- EXAMPLE 210: [Ala¹⁷]hPTH(1-36)NH₂ [MS(IS): 4270]
- EXAMPLE 211: [D-Ser¹⁷]hPTH(1-36)NH₂ [MS(IS): 4288]
- EXAMPLE 212: [Ala¹⁹]hPTH(1-36)NH₂ [MS(IS): 4228]
- EXAMPLE 213: [D-Glu¹⁹]hPTH(1-36)NH₂ [MS(IS): 4288]
- EXAMPLE 214: [Ala²¹]hPTH(1-36)NH₂ [MS(IS): 4257]
- EXAMPLE 215: [Ala²²]hPTH(1-36)NH₂ [MS(IS): 4227]
- EXAMPLE 216: [Gln²⁶]hPTH(1-36)NH₂ [MS(IS): 4287]

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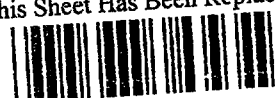
- EXAMPLE 217: [Nle²⁶]hPTH(1-36)NH₂ [MS(IS): 4271]
- EXAMPLE 218: [D-Lys²⁶]hPTH(1-36)NH₂ [MS(IS): 4288]
- EXAMPLE 219: [Nle^{8, 18, 27}]hPTH(1-36)NH₂
- EXAMPLE 220: [His²⁷]hPTH(1-36)NH₂ [MS(IS): 4294]
- EXAMPLE 221: [Phe²⁷]hPTH(1-36)NH₂ [MS(IS): 4304]
- EXAMPLE 222: [Nle²⁷]hPTH(1-36)NH₂ [MS(IS): 4271]
- EXAMPLE 223: [Asn²⁷]hPTH(1-36)NH₂ [MS(IS): 4271]
- EXAMPLE 224: [Ala²⁷]hPTH(1-36)NH₂ [MS(IS): 4228]
- EXAMPLE 225: [D-Gln²⁹]hPTH(1-36)NH₂ [MS(IS): 4289]
- EXAMPLE 226: [D-Asp³⁰]hPTH(1-34)NH₂ [MS(IS): 4118]
- EXAMPLE 227: [Ala³⁰]hPTH(1-36)NH₂ [MS(IS): 4241]
- EXAMPLE 228: [D-Val³¹]hPTH(1-36)NH₂ [MS(IS): 4290]
- EXAMPLE 229: [Ala³¹]hPTH(1-36)NH₂ [MS(IS): 4258]
- EXAMPLE 230: [Lys³²]hPTH(1-34)NH₂ [MS(IS): 3832]
- EXAMPLE 231: [D-His³²]hPTH(1-36)NH₂ [MS(IS): 4288]
- EXAMPLE 232: [Ala³²]hPTH(1-36)NH₂ [MS(IS): 4219]
- EXAMPLE 233: [D-Asn³³]hPTH(1-36)NH₂ [MS(IS): 4288]
- EXAMPLE 234: [Ala³³]hPTH(1-36)NH₂ [MS(IS): 4210]
- EXAMPLE 235: [NMePhe³⁴]hPTH(1-36)NH₂ [MS(IS): 4301]
- EXAMPLE 236: [D-Asp³⁰]hPTH(1-36)NH₂ [MS(IS): 4290]
- EXAMPLE 237: Ip-[Nle^{8, 18}, Lys(Ip)^{13, 26, 27}, D-Asn³³, D-Phe³⁴]
hPTH(1-34)NH₂
- EXAMPLE 238: Ip-[Nle^{8, 18, 27}, Lys(Ip)^{13, 26}]hPTH(1-36)NH₂
- EXAMPLE 239: [Nle^{8, 18}, D-Asn³³, D-Phe³⁴]hPTH(1-34)NH₂
[MS(IS): 4080]
- EXAMPLE 240: [Lys(Ip)¹³]hPTH(1-36)NH₂ [MS(IS): 4331]
- EXAMPLE 241: Propargyl-[A¹]-hPTH-(1-36)-NH₂
- EXAMPLE 242: [Ala⁰]-hPTH-(1-36)-NH₂
- EXAMPLE 243: [Pro⁰]-hPTH-(1-36)-NH₂
- EXAMPLE 244: Acetyl-hPTH-(1-36)-NH₂
- EXAMPLE 245: [HyPro¹]-hPTH-(1-36)-NH₂
- EXAMPLE 246: N-Dimethyl-[Ala¹]-hPTH-(1-36)-NH₂
- EXAMPLE 247: [D-Ser¹]-hPTH-(1-36)-NH₂ [MS(IS): 4288]
- EXAMPLE 248: [Lys(For)¹]-hPTH-(1-36)-NH₂ [MS(IS): 4356]

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- EXAMPLE 249: [D-Glyceric acid¹]-hPTH-(1-36)-NH₂
- EXAMPLE 250: [Asn¹]-hPTH-(1-36)-NH₂
- EXAMPLE 251: [4-Aminobenzoic acid¹]-hPTH-(1-36)-NH₂
- EXAMPLE 252: [4-Aminosalicylic acid¹]-hPTH-(1-36)-NH₂
- EXAMPLE 253: [TMSA¹]-hPTH-(1-36)-NH₂ [MS (IS): 4343]
- EXAMPLE 254: [Phe¹]-hPTH-(1-36)-NH₂
- EXAMPLE 255: [Propargylglycin¹]-hPTH-(1-36)-NH₂
- EXAMPLE 256: [Ala¹, His⁵, Leu⁸, Asp¹⁰, Lys¹¹, Gln¹⁸, Phe²², Phe²³, His²⁵, His²⁶,
Leu²⁷, Thr³³, Ala³⁴]-hPTH-(1-34)-NH₂
- EXAMPLE 257: [Abu²]-hPTH-(1-36)-NH₂
- EXAMPLE 258: [D-Val²]-hPTH-(1-36)-NH₂ [MS (IS): 4288]
- EXAMPLE 259: [tert. Leu²]-hPTH-(1-36)-NH₂
- EXAMPLE 260: [Ala¹]-hPTH-(1-36)-NH₂
- EXAMPLE 261: [D-Ile⁵]-hPTH-(1-36)-NH₂ [MS (IS): 4288]
- EXAMPLE 262: [D-Gln⁶]-hPTH-(1-36)-NH₂ [MS (IS): 4288]
- EXAMPLE 263: [D-Leu⁷]-hPTH-(1-36)-NH₂ [MS (IS): 4288]
- EXAMPLE 264: [Nle⁸]-hPTH-(1-36)-NH₂ [MS (IS): 4268]
- EXAMPLE 265: [Phe⁸]-hPTH-(1-36)-NH₂ [MS (IS): 4303]
- EXAMPLE 266: [Cha⁸]-hPTH-(1-36)-NH₂ [MS (IS): 4309]
- EXAMPLE 267: [Leu⁸]-hPTH-(1-38)-NH₂
- EXAMPLE 268: [D-Leu¹¹]-hPTH-(1-36)-NH₂ [MS (IS): 4287]
- EXAMPLE 269: [Ala¹¹]-hPTH-(1-36)-NH₂ [MS (IS): 4244]
- EXAMPLE 270: [D-Lys¹³]-hPTH-(1-36)-NH₂ [MS (IS): 4288]
- EXAMPLE 271: [D-Leu¹⁵]-hPTH-(1-36)-NH₂ [MS (IS): 4288]
- EXAMPLE 272: [Ala¹⁵]-hPTH-(1-36)-NH₂ [MS (IS): 4243]
- EXAMPLE 273: [Ala¹⁶]-hPTH-(1-36)-NH₂ [MS (IS): 4243]
- EXAMPLE 274: [Met (O₂)¹⁸]-hPTH-(1-36)-NH₂ [MS (IS): 4320]
- EXAMPLE 275: [Nle¹⁸]-hPTH-(1-36)-NH₂ [MS (IS): 4268]
- EXAMPLE 276: [D-Met¹⁸]-hPTH-(1-36)-NH₂ [MS (IS): 4288]
- EXAMPLE 277: [Lys²⁰]-hPTH-(1-36)-NH₂ [MS (IS): 4259]
- EXAMPLE 278: [D-Arg²⁰]-hPTH-(1-36)-NH₂ [MS (IS): 4288]
- EXAMPLE 279: [D-Val²¹]-hPTH-(1-36)-NH₂ [MS (IS): 4288]
- EXAMPLE 280: [Trp (SO₂Pmc)²³]-hPTH-(1-38)-NH₂ [MS (IS): 4723]
- EXAMPLE 281: [Trp (Pmc)²³]-hPTH-(1-38)-NH₂ [MS (IS): 4660]

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- EXAMPLE 282: [D-Trp²³]-hPTH-(1-36)-NH₂ [MS (IS): 4288]
- EXAMPLE 283: [Ala²³]-hPTH-(1-36)-NH₂ [MS (IS): 4171]
- EXAMPLE 284: [D-Leu²⁴]-hPTH-(1-36)-NH₂ [MS (IS): 4288]
- EXAMPLE 285: [Phe²⁵]-hPTH-(1-36)-NH₂ [MS (IS): 4277]
- EXAMPLE 286: [Lys²⁵]-hPTH-(1-36)-NH₂ [MS (IS): 4258]
- EXAMPLE 287: [Ala²⁵]-hPTH-(1-36)-NH₂ [MS (IS): 4201]
- EXAMPLE 288: [Ala²⁶]-hPTH-(1-36)-NH₂ [MS (IS): 4229]
- EXAMPLE 289: [Lys^{26/27} (For)]-hPTH-(1-34)-NH₂
- EXAMPLE 290: [D-Lys²⁷]-hPTH-(1-36)-NH₂ [MS (IS): 4288]
- EXAMPLE 291: [D-Leu²⁸]-hPTH-(1-36)-NH₂ [MS (IS): 4288]
- EXAMPLE 292: [D-Phe³⁴]-hPTH-(1-36)-NH₂ [MS (IS): 4288]
- EXAMPLE 293: [D-Val³⁵]-hPTH-(1-36)-NH₂
- EXAMPLE 294: [Ala³⁵]-hPTH-(1-36)-NH₂
- EXAMPLE 295: [Pro³⁵]-hPTH-(1-36)-NH₂
- EXAMPLE 296: [NMeVal³⁵]-hPTH-(1-36)-NH₂
- EXAMPLE 297: [Thr³⁵, Ala³⁶]-hPTH-(1-36)-NH₂
- EXAMPLE 298: [D-Ala³⁶]-hPTH-(1-36)-NH₂
- EXAMPLE 299: [NMeAla³⁶]-hPTH-(1-36)-NH₂
- EXAMPLE 300: (5-Amino-4-fluoro-2-isopropyl)-hex-3-enoyl-hPTH(3-36)NH₂

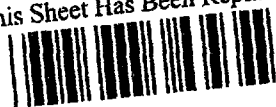
a) Preparation of hPTH-(3-36)-NH-resin

This intermediate is prepared in analogy with the procedure of Example 165.

b) [(5-Amino-4-fluoro-2-isopropyl)-hex-3-enoyl]hPTH(3-36)-amide

204.6 mg Fmoc-hPTH(3-36)-NH-resin obtained in a) above are treated with DMF for 5 min. and then washed several times with isopropanol and DMF. The Fmoc-protecting group is removed by treatment with DMF/piperidin 8:2 for 10 min.. The deprotected

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PTH-fragment, 38.1 mg of 4-fluoro-2-isopropyl-5-tert-butoxycarbonylamino-hex-3-enoic acid, 68.7 mg benzotriazol-1-yloxytris(pyrrolidino)phosphonium hexafluorophosphate (PyBOP) and 26.6 mg 4-methylmorpholin are shaken for 80 min. in 0.7 ml DMF. After several washings with isopropanol and DMF, the solid residue is treated with 4.5 ml TFA/0.25 ml water/0.25 ml ethanedithiol for 55 min. This mixture is then filtered and by addition of diethyl ether the title compound precipitated from the remaining solution. The title compound is purified by prep. HPLC (Vydac column, gradient A = water, B = 7 acetonitril/3 water/0.2 phosphoric acid).

MS (Ion-spray): mass 4272

$[\alpha]_{D}^{25} = -66,8^{\circ}$ (c=0.25 in 95% AcOH)
365 nm

EXAMPLE 301: 4-Fluoro-2-isopropyl-5-tert.butoxycarbonylamino-hex-3-enoate

a) (S)-3-(1-Oxo(4-fluoro-3-hydroxy-2-isopropyl)hex-4-enyl)-4-phenylmethyl-2-oxazolidinone

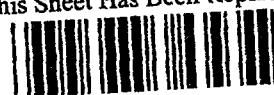
The title compound is prepared analogous to the procedure described by D.A. Evans et al., Org. Synth. 68, 1990; starting from (S)-3-(1-oxoisobutyl)-4-phenylmethyl-2-oxazolidinone and 2-fluoro-but-2-enal.

MS (FAB): MH⁺ 350

b) (S)-3-(1-Oxo(4-fluoro-3-O-(2,2,2trichlorethanimino)-2-isopropyl)hex-4-enyl)-4-phenylmethyl-2-oxazolidinone

1.5 g of the compound of Example 301a) are dissolved in 6 ml DCM. At 0° 0.0958 ml 1,8-diazabicyclo(5,4,0)undec-7-en (DBU) are added. To this solution 0.475 ml trichloroacetonitril in 2 ml DCM is added dropwise at 0°. After 1 hour the reaction mixture is

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evaporated and the residue is purified by column chromatography on silica gel (hexan/diethyl ether 2:1).

MS (FAB): MH⁺ 493

c) (S)-3-(1-Oxo(4-fluoro-5-N-2,2,2-trichloroacetyl-amino-2-isopropyl)hex-3-enyl)-4-phenylmethyl-2-oxazolidinone

2 g of the compound of 301b) is dissolved in 150 ml o-xylene and stirred under reflux at 160° for 3 hours. Then o-xylene is removed and the residue is purified by column chromatography on silica gel (toluol/ethyl acetate 98:2).

MS (FAB): MH⁺ 493

d) 4-Fluoro-5-N-2,2,2-trichloroacetyl-amino-2-isopropyl-hex-3-enoic acid

The title compound is prepared analogous to the procedure described by D.A. Evans et al. Org. Synth. 68 1990, but using as starting material the compound of 301c).

MS (FAB): MH⁺ 335

e) 5-Amino-4-fluoro-2-isopropyl-hex-3-enoic acid

899 mg of the compound of 301d) are dissolved in 20 ml ethanol and 13.5 ml 6N sodium hydroxide solution are added. This mixture is stirred for 20 hours at r.t. Then ethanol is removed and the remaining aqueous fraction is acidified with 2N HCl to pH=2 and extracted with n-butanol. The extract is evaporated and the residue is filtered through silica gel (DCM/methanol 1:1), thus yielding pure title compound.

MS (FAB): MH⁺ 190

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f) 4-Fluoro-2-isopropyl-5-tert.butoxycarbonylamino-hex-3-enoic acid

0.56 g of the compound of 301e) are dissolved in 12 ml water and 1.5 g sodium carbonate are added. To this solution 0.4 g di-tert.butylcarbonate dissolved in 12 ml tetrahydrofuran are added. This mixture is stirred at room temperature for 18 hours. Then 50 ml n-butanol and 50 ml water are added and under vigorous stirring 1N HCl is added until pH=2. The organic fraction is evaporated and the residue purified by column chromatography on silica gel (hexan/acetone 1:2).

MS (FAB): MH^+ 290

$[\alpha]_D = -61,3^\circ$ (1% in methanol)

By repeating the above procedures but using the appropriate starting materials the following compounds may be prepared:

- 4-Fluoro-2-methyl-5-tert.butoxycarbonylamino-hex-3-enoic acid
- 4-Chloro-2-methyl-5-tert.butoxycarbonylamino-hex-3-enoic acid
- 4-Fluoro-2-benzyl-5-tert.butoxycarbonylamino-hex-3-enoic acid
- 4-Chloro-2-isopropyl-5-tert.butoxycarbonylamino-hex-3-enoic acid
- 4-Methyl-2-isopropyl-5-tert.butoxycarbonylamino-hex-3-enoic acid

EXAMPLE 302:

By following the procedure of Example 300 above but using the appropriate starting materials, the following compounds may be prepared:

- (5-Amino-4-fluoro-2-methyl)-hex-3-enoyl-hPTH(3-36)-amide
MS (Ion-Spray): Mass 4245

- (5-Amino-4-chloro-2-methyl)-hex-3-enoyl-hPTH(3-36)-amide
MS (Ion-Spray): Mass 4260

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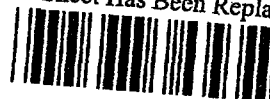
- (5-Amino-4-fluoro-2-benzyl)-hex-3-enoyl-hPTH(3-36)-amide
MS (Ion-Spray): Mass 4320
- (5-Amino-4-chloro-2-isopropyl)-hex-3-enoyl-hPTH(3-36)-amide
MS (Ion-Spray): Mass 4289
- (5-Amino-4-methyl-2-isopropyl)-hex-3-enoyl-hPTH(3-36)-amide
MS (Ion-Spray): Mass 4269

EXAMPLE 303:

By following the procedure of Example 300 above but using the appropriate starting materials, the following compounds may be prepared:

- a) (5-amino-3-aza-2-isopropyl)-hexanoyl-hPTH(3-36)-amide [MS (Ion-Spray): 4257] using as starting material:
(4-aza-2-t.-butoxycarbonylamino-5-isopropyl)-hexanoic acid
- b) (5-amino-3-aza-3-N-acetyl-2-isopropyl)-hexanoyl-hPTH(3-36)-amide [MS (Ion-Spray): 4299] using as starting material:
(4-aza-4-N-acetyl-2-t.-butoxycarbonylamino-5-isopropyl)-hexanoic acid
- c) (5-amino-3-aza-3-N-acetyl)-hexanoyl-hPTH(3-36)-amide [MS (Ion-Spray): 4257] using as starting material: (4-aza-4-N-acetyl-2-t.-butoxycarbonylamino)-hexanoic acid
- d) (5-methylamino-3-aza-2-isopropyl)-hexanoyl-hPTH(3-36)-amide [MS (Ion-Spray): 4271] using as starting material:
(4-aza-2-(N-t.-butoxycarbonyl,N-methylamino)-5-isopropyl)-hexanoic acid

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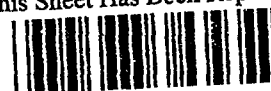
- e) (5-methylamino-3-aza-3-N-methyl-2-isopropyl)-hexanoyl-hPTH(3-36)-amide [MS (Ion-Spray): 4285] using as starting material: [4-aza-4-N-methyl-2-(N-t.-butoxycarbonylamino,N-methyl-amino)-5-isopropyl]-hexanoic acid
- f) (5-amino-3-aza-3-N-methyl-2-isopropyl)-hexanoyl-hPTH(3-36)-amide [MS (Ion-Spray): 4271] using as starting material: (4-aza-4-N-methyl-2-t.-butoxycarbonylamino-5-isopropyl)-hexanoic acid
- g) (5-amino-3-aza-3-N-isopropyl)-hexanoyl-hPTH(3-36)-amide [MS (Ion-Spray): 4257] using as starting material: (4-aza-4-N-isopropyl-2-t.-butoxycarbonylamino-5-isopropyl)-hexanoic acid
- h) [1-cyclopentane-1-amino-1-carboxylic acid(psi CH₂-NH)-Val²]-hPTH(1-36)-amide using 1-cyclopentane-1-t.-butoxycarbonylamino-1-carboxylic acid (psi CH₂-NH)-Val-OH as starting material.

EXAMPLE 304:

By following the procedure of Example 3a) above but using the appropriate starting material, the following compound may be prepared:

(5-amino-3-aza-2-isopropyl)-hexanoyl-[Leu⁸,Ala¹⁶,Gln¹⁸,Ala¹⁹]hPTH(3-36)OH [MS (Ion-Spray): 4135] using as starting material: (4-aza-2-t.-butoxycarbonylamino)-hexanoic acid

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EXAMPLE 305: (4-Aza-4-N-acetyl-2-tert.butoxycarbonylamino-5-isopropyl)-hexanoic acid

a) Methyl(4-aza-2-tert.butoxycarbonylamino-5-isopropyl)-hexanoic acid

Boc-Ala-Val-OMe is treated with the Lawesson-reagent (S.-O. Lawesson, et al. Tetrahedron 1981, 37, 3635). The resulting endothiopeptide is then reduced in accordance with the procedure described by F.S. Guziec et. al. THL, 1990, 23-26.

MS (EI): MH⁺ 289

b) Methyl (4-aza-4-N-acetyl-2-tert.butoxycarbonylamino-5-isopropyl) hexanoate

264 mg of the compound of 305a) is dissolved in 5 ml DCM and then 0.14 ml triethylamin and 0.072 ml acetyl chloride are added. This mixture is stirred for 24 hours at 40°. After addition of 50 ml DCM, the solution is washed twice with water. The organic fraction is dried and evaporated. The title compound thus obtained is purified by column chromatography (hexan/ethyl acetate 4:1)

MS (EI): M⁺ 330

c) (4-Aza-4-N-acetyl-2-tert.butoxycarbonylamino-5-isopropyl)-hexanoic acid

356 mg of the compound of 305b) are dissolved in 4 ml methanol/-water 3:1 and 53 mg lithium hydroxide are added. This mixture is stirred at 60° for two hours. Then 25 ml DCM are added and 1N HCl is added under vigorous stirring until pH=2. The organic fraction is separated, dried and evaporated, thus yielding the title compound in pure form.

MS (FAB): MH⁺ 317

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By repeating the procedure but using the appropriate starting materials, following compounds may be obtained:

(4-Aza-4-N-acetyl-2-tert.butoxycarbonylamino)-hexanoic acid
(4-Aza-2-tert.butoxycarbonylamino-5-isopropyl)-hexanoic acid
[4-aza-2-(N-t.-butoxycarbonyl,N-methylamino)-5-isopropyl]-
hexanoic acid
(4-aza-4-N-methyl-2-t.-butoxycarbonylamino-5-isopropyl)-hexanoic
acid
(4-aza-4-N-isopropyl-2-t.-butoxycarbonylamino-5-isopropyl)-
hexanoic acid
(4-aza-2-t.-butoxycarbonylamino)-hexanoic acid

The compounds of Examples 1 to 300 and 302 to 304 show correct amino acid ratios in quantitative amino acid analysis.

In the following examples illustrating the recombinant process of the invention, if not otherwise specified, the standard procedures described in Ausubel et al; 1991; Current Protocols in Molecular Biology, John Wiley & Sons, New York are used.

EXAMPLE 306: Cloning bacteriophage T4 gene 55

Two DNA fragments, one containing the entire coding sequence of bacteriophage T4 gene 55, and the other fragment encoding part of it are amplified by polymerase chain reaction (PCR) using purified T4 DNA as the template. Oligonucleotides which contain NdeI or BamHI endonuclease recognition sequences are used as primers. The PCR reactions (25 cycles, each 30" 94 °C, 30" 50 °C, 30" 72 °C) are performed with 1 ng of template and 100 pM of the upstream primer (GAGGTGCATA TGTCAGAAAC TAAGCCT 27), the NdeI recognition site being provided by nucleotides 7-12 thereof) and

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100 pM of the downstream primer

(GGTCGGATCC ATCGTTAGCG TTAGCCTCAT ATAAAAATC 40 the BamHI, recognition site being provided by nucleotides 5-10 thereof) in 100 µl reaction buffer containing 10 mM Tris-HCl, 50 mM KCl, 1.5 mM MgCl₂, 0.2 mM of each of dATP, dCTP, dGTP, dTTP, and 0.1% Triton X 100, pH 9.0. A 0.6kb fragment is obtained.

The truncated form of gene 55 is prepared by PCR as described for the entire gene, except that a different oligonucleotide is used as downstream primer (GGTCGGATCC TACGTTGGAC GAATGCAT 28, the BamHI recognition site being provided by nucleotides 5-10 thereof). A 0.35 kb fragment is obtained.

The 0.6 kb and 0.35 kb DNA fragments are digested with restriction endonucleases NdeI/BamHI and are cloned into a NdeI/BamHI digest of expression vector pET17B (obtained from Novagen). The 0.6 kb DNA fragment encodes the entire gp55 protein (188 amino acid residues - see Fig. 1), whereas the 0.35 kb fragment encodes a 12 kD amino-terminal fragment of gp55 (112 amino acid residues - see Fig. 2).

Example 307: Preparation and cloning of the DNA encoding PTH (1-38)

DNA fragments encoding a) the linker peptide corresponding to amino acid residues 110-115 of Fig. 1 and PTH (1-38) or b) the linker peptide corresponding to amino acid residues 186-191 of Fig. 2 and PTH (1-38) are generated by PCR using a cloned synthetic PTH (1-38) coding sequence as a template. The oligonucleotides encoding the linker a)

(GAGTGGATCC ATCGATCCAC CATCCGTATC AGAAATACAA CT 42), or encoding linker b)

GAGTGGATCC GTTAACGGTC CATCCGTATC AGAAATACAA CT 42 are used as upstream primers. The downstream primer TCTACTCGAG TTAACCCAGA GCTACAAAAT TATG 34

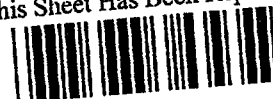
The upstream primers each incorporate a BamHI cloning site and the downstream primer contains a XhoI recognition site (nucleotides 5-10 of each sequence). The PCR reaction is performed under the same conditions as described in Example 306. After cleavage with BamHI and XhoI, the 0.14 kb PCR products are inserted into BamHI-XhoI digests of the gp55-pET17B plasmids, obtained in Example 306.

For expression of the fusion proteins the resultant plasmid constructs are transformed into E.coli BL21 (DE3) LysE and single bacterial colonies are further propagated. The expression of the fusion proteins is obtained by incubating a preculture at 37°C on a rotary shaker overnight in circle growth medium (Bio 101). A main culture is inoculated with the preculture making up 1 to 5 % of the volume of the main culture. The main culture is then fermented at a temperature of 37°C and at a pH of 6.9 to 7.1 whilst being aerated at 10 l/min and agitated at 200-400 rpm. Expression is induced by the addition of 1 mM IPTG as soon as the culture reaches an OD₅₅₀ of about 1.0 to 5.0. After induction the fermentation is continued for five hours and then the fermentation broth is harvested without killing the E. coli cells. The harvested cells are spun down with a tubular centrifuge and resuspended in sample buffer and assayed for expression by SDS PAGE. The cell pellet is then frozen until purification. The procedures and host cells used are fully described in Studier et al; 1990; "Use of T7 RNA Polymerase to direct expression of Cloned Genes", Methods in Enzymology, Academic Press, pages 60 to 89.

EXAMPLE 308: Preparation of inclusion bodies

Frozen E. coli pellets obtained from Example 307 are resuspended to 25% w/v in Buffer A (50mM Tris pH 8.0; containing 2mM DTT, 5mM Benzamidinium-HCl, 1.5mM MgCl₂, 1.0mM MnCl₂, 10µg/ml DNase I and

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2mg/ml Lysozyme) and mixed by stirring for 1 hour at room temperature. The resuspended cells are lysed by passage through a Manton-Gaulin homogeniser (2 passes at 1200 bar) and the resultant lysate kept on ice. The lysate is diluted 2 fold with ice-cold buffer B (50mM Tris pH 8.0; containing 2mM DTT, 5mM Benzamidine-HCl and 4mM EDTA) and centrifuged for 30min at 27,500g.

The supernatant from the centrifuge is carefully removed and discarded. The pellet is resuspended in buffer B to 25% w/v, passed once more through the Manton-Gaulin and centrifuged as before. This process is repeated once using buffer B and once using water. The resultant inclusion body pellets are weighed and frozen at -20°C until required.

EXAMPLE 309: Solubilisation and cleavage of gp55-Asp-Pro-Pro-(1-38)hPTH inclusion bodies

Frozen inclusion bodies containing gp55-Asp-Pro-Pro-(1-38)hPTH as obtained in Example 308 are suspended in 10 mM HCl acid solution (analytical grade) to a final concentration of 6%w/v. The solution is incubated for 24 hours at 50°C and the reaction terminated by addition of 1 volume of aqueous 100 mM NaOAc solution. The diluted supernatant is clarified by centrifugation at 27,500g for 30 minutes and filtered through a glass fibre filter.

EXAMPLE 310: Solubilisation and cleavage of gp55-Asn-Gly-Pro-(1-38)hPTH inclusion bodies

Frozen inclusion bodies containing gp55-Asn-Gly-Pro-(1-38)hPTH and obtained from example 308 are dissolved in 6M Guanidine-HCl (containing 2M Hydroxylamine-HCl and brought to pH 9.0 by addition of 4.5M LiOH) to give an estimated 5mg/ml protein. The

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pH is readjusted to pH 9 by further addition of LiOH and the solution incubated for 4 hours at 45°C. The reaction is terminated by addition of formic acid to pH 4 and the solution centrifuged and filtered as described in example 309.

EXAMPLE 311: Preparative HPLC of Gly-Pro and Pro-Pro (1-38)hPTH

Filtered cleavage supernatants are analysed on analytical reversed phase HPLC (Orpogen HD-Gel-RP-7s-300; 4 x 150mm column) and compared with a known (1-38)hPTH standard (Bachem) to determine the PTH concentration. The column is equilibrated with 85% HPLC buffer A (90% water, 10% acetonitrile; containing 0.1%v/v TFA) and 15% HPLC buffer B (10% water, 90% acetonitrile; containing 0.1% TFA) and is eluted with a gradient of 15% HPLC buffer B to 100% HPLC buffer B over 15 minutes at a flow rate of 0.75ml/min.

Depending upon the scale of the preparation, supernatants are either loaded on a 1.0 x 25cm C4 Vydac or a 2.2 x 25cm C4 Vydac. The columns are equilibrated with 85% HPLC buffer A and 15% HPLC buffer B at flow rates of 4ml/min and 10ml/min respectively. The columns are eluted with gradients of 15%B to 85%B over 80ml and 15%B to 55%B over 300ml respectively.

Gly-Pro-hPTH(1-38) or Pro-Pro-hPTH(1-38) peaks are collected manually and the acetonitrile is later removed under vacuum. Analytical HPLC is used to determine the peptide concentration both after cleavage and in the collected fractions.

The solvent free peptide solutions are diluted with an equal volume of 100mM sodium acetate and the pH adjusted, if necessary, to pH 5.4. Following filtration (0.2µm), the solution is loaded on a cation exchange column (either Mono S or SP-Sepharose High Performance, packed into HR10/10 or XK16/10 columns respectively) equilibrated with 50mM sodium acetate pH 5.4 (buffer C). The column is eluted with a 0 to 500mM NaCl gradient in buffer C over 7.5 column volumes. 10ml fractions are collected and the

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X-X-(1-38)hPTH peak is pooled. The peptide concentration is determined as before using HPLC.

This column serves both to exchange the peptides into a more physiological buffer and to remove additional peptides produced by chemical cleavage of the fusion protein itself.

EXAMPLE 312: Enzymatic removal of X-X from X-X-(1-38)hPTH

Pooled peptide fractions, as obtained in Example 311, (concentration range 1.5 - 2.0 mg/ml) are brought to pH 8.0 by addition of solid Tris. Purified X-Prolyl dipeptidase (0.5mg/ml in PBS pH 7.2) (Nardi et al. (1991) App. Env. Microbiol. 57, No.1, 45-50) is added to 1:1000 and the solution is incubated for 24h at 37 C. The reaction is stopped by addition of TFA to 0.2% (pH = 5.0) and the resultant (1-38)hPTH is isolated using preparative HPLC (using the same conditions as described in Example 311). The solvent is removed and the product is lyophilised after removal of aliquots for N-terminal sequence and mass spectroscopic analyses.

Examples of the yields which are obtained using the two fusion protein approaches described above are as follows:

Gen55-Asp-Pro-Pro-(1-38)PTH (using the "truncated" form of Gen55)

Under partially optimized fermentation conditions, a 215 g wet cell pellet is obtained from a 10 litre fermentation. The percentage level of expression of the fusion protein is approximately 37%. From this pellet, 31 g of inclusion bodies are isolated. The protein purity (with respect to the fusion protein) of these bodies is 61%. After solubilisation, cleavage and centrifugation/filtration approximately 620mg Pro-Pro-PTH are detected. This material yields 580mg Pro-Pro-PTH after subsequent C4-Vydac HPLC. After dilution, cation-exchange on Mono S,

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digestion with X-Prolydipeptidase (at a Pro-Pro-PTH concentration of 2mg/ml), HPLC and lyophilisation, 440mg of lyophilized pure product with full biological activity is obtained. This represents a yield of 71% between the stopped cleavage step and the final product.

Gen55-Asn-Gly-Pro-(1-38)PTH (using the "long" form of Gen55)

From a non-optimized 20 litre fermentation a 78g wet cell pellet with 50% expression of the fusion protein is harvested. Following lysis and centrifugation, 18g of "90% pure" inclusion bodies are obtained. Following the acid stop step and centrifugation/filtration about 470mg Gly-Pro-PTH are detected. After HPLC on C4-Vydac, 355mg of peptide are detected, decreasing to 321mg after SP-Sepharose cation-exchange. Enzymatic removal of the Gly and Pro residues is carried out at 2mg/ml. After the final HPLC step and lyophilization, 244mg of pure, fully biologically active (1-38)PTH is recovered representing a yield of 52% from the stopped cleavage reaction stage.

Compounds of Examples 25, 26, 56 and 86 to 164 are also prepared using the process of Examples 306 to 312.

The compounds of the invention in free form or in the form of pharmaceutically acceptable salts and complexes exhibit valuable pharmacological properties as indicated in animal tests and are therefore indicated for therapy.

The biological activity of the compounds of the invention is assessed in vitro by measuring their ability of stimulating the synthesis of cyclic AMP in UMR 106-06 rat and SaOS-2 human osteosarcoma cells according to the method of Marcus and Aurbach in Endocrinology, 85, 801-810 (1969). Rat osteosarcoma UMR 106 cells are grown to confluence in Eagle's Minimum Essential Medium - 10% FCS in 12 well plates, human SaOS-2 osteosarcoma cells are

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grown in McCoy's 5A medium - 10% FCS. The medium is then changed to medium with 2% FCS and 1-5 $\mu\text{Ci}/\text{well}$ $[^3\text{H}]$ -adenine is added. Two hours later, cells are washed twice with serum-free medium and incubated in DMEM - 1% BSA containing 1 mM 3-isobutyl-1-methyl-xanthine to exclude actions on phosphodiesterases. Test substances are added 20 min later. The reaction is stopped and cAMP extracted after 15 min by adding ice cold 5% trichloroacetic acid. A carrier solution (0.5 ml/well) containing 5 mM of unlabelled adenine, adenosine, AMP, ADP, ATP, and cAMP as well as 0.4 μCi $[^{14}\text{C}]$ -adenosine for determination of recovery is added. $[^3\text{H}]$ -cAMP is separated using serial Dowex 50W-X4 (200-400 mesh) and alumina chromatography and counted. Results are calculated in % of solvent control and EC_{50} values determined from DRC curves. Compounds of the invention stimulate cAMP production in UMR 106-06 rat and SaOS-2 human osteosarcoma cells at a concentration of 10^{-11} to 10^{-6} M.

The compounds of the invention also have binding affinity to PTH receptors, e.g. as follows:

Chicken $[\text{Tyr}^{36}]$ PTHrP(1-36)amide is iodinated to a specific activity of 2,200 Ci/mmol using the lactoperoxidase method (Anawa Lab. AG, Wangen). Monolayers of opossum kidney cells are washed with 200 μl DMEM and HAM's F12 (1:1) containing 1% BSA and incubated at 16°C with 50,000cpm of $[^{125}\text{I}-\text{Tyr}^{36}]$ chPTHrP(1-36)-amide per well in the presence or absence of 1 μM $[\text{Tyr}^{36}]$ chPTHrP(1-36)amide. After incubation, cells are washed with 0.5 ml medium (4°C), lysed with 0.5 ml 1N NaOH and radioactivity is determined. Specific binding is defined as total binding minus nonspecific binding. Competition curves are analyzed using SCTFIT, a non-linear regression computer program (Feyen et al, 1992, Biochem. Biophys. Res. Commun. 187:8-13) and data presented as mean pK_D values ($n=2$ to 3). Compounds of the invention show in

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this test a binding affinity expressed as a mean pK_D value of from 7 to 10.

Furthermore, the compounds of the invention increase plasma calcium level after continuous s.c. infusion, e.g. as determined in male thyroparathyroidectomized Wistar rats weighing 140 to 170 g. 5 days after thyroparathyroidectomy, rats are implanted in the neck with Alzet minipumps and test compounds infused at rates of 0.3 to 30 $\mu\text{g/day/rat}$. Blood is drawn by retro-orbital puncture in the morning of days 1, 2, 3, and 6 thereafter and plasma calcium concentrations are determined photometrically. In this test compounds of the invention increase plasma calcium.

The compounds of the invention are accordingly indicated for preventing or treating all bone conditions which are associated with increased calcium depletion or resorption or in which calcium fixation in the bone is desirable, e.g. osteoporosis of various genesis (e.g. juvenile, menopausal, post-menopausal, post-traumatic, caused by old age or by cortico-steroid therapy or inactivity), fractures, osteopathy, including acute and chronic states associated with skeletal demineralisation, osteo-malacia, periodontal bone loss or bone loss due to arthritis or osteoarthritis or for treating hypoparathyroidism.

The compounds of the invention are particularly indicated for preventing or treating osteoporosis of various genesis.

For all the above uses, an indicated daily dosage is in the range from about 0.003 to about 10 mg preferably 0.003 to 3, more preferably 0.01 to 1 mg of a compound of the invention. Compounds of the invention may be administered once a day or up to twice a week.

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The compounds of the invention may be administered in free form or in pharmaceutically acceptable salt form or complexes. Such salts and complexes may be prepared in conventional manner and exhibit the same order of activity as the free compounds. The present invention also provides a pharmaceutical composition comprising a compound of the invention in free base form or in pharmaceutically acceptable salt form or complex form in association with a pharmaceutically acceptable diluent or carrier. Such compositions may be formulated in conventional manner. The compounds of the invention may be administered by any conventional route, for example parenterally e.g. in form of injectable solutions or suspensions, enterally, e.g. orally, for example in the form of tablets or capsules or in a transdermal, nasal or a suppository form.

In accordance with the foregoing the present invention further provides:

- a) a compound of the invention or a pharmaceutically acceptable salt or complex thereof for use as a pharmaceutical;
- b) a method for improving bone formation, e.g. preventing or treating all bone conditions which are associated with increased calcium depletion or resorption or in which calcium fixation in the bone is desirable, e.g. osteoporosis of various genesis (e.g. juvenile, menopausal, post-menopausal, post-traumatic, caused by old age or by cortico-steroid therapy or inactivity), fractures, osteopathy, including acute and chronic states associated with skeletal demineralisation, osteo-malacia, periodontal bone loss or bone loss due to arthritis or osteoarthritis, or for treating hypoparathyroidism in a subject in need of such treatment, which method comprises administering to said subject an effective amount

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of a compound of the invention or a pharmaceutically acceptable salt or complex thereof;

- c) a compound of the invention or a pharmaceutically acceptable salt or complex thereof for use in the preparation of a pharmaceutical composition for use in the method as in b) above.

According to a further embodiment of the invention, the compounds of the invention may be employed as adjunct or adjuvant to other therapy, e.g. a therapy using a bone resorption inhibitor, for example as in osteoporosis therapy, in particular a therapy employing calcium, a calcitonin or an analogue or derivative thereof, e.g. salmon, eel or human calcitonin, a steroid hormone, e.g. an estrogen, a partial estrogen agonist or estrogen-gestagen combination, a biphosphonate, vitamin D or an analog thereof, or any combination thereof.

When the compounds of the invention are administered in conjunction with, e.g. as an adjuvant to bone resorption inhibition therapy, dosages for the co-administered inhibitor will of course vary depending on the type of inhibitor drug employed, e.g. whether it is a steroid or a calcitonin, on the condition to be treated, whether it is a curative or preventive therapy, on the regimen and so forth.

In accordance with the foregoing the present invention provides in a yet further aspect:

- d) a method for improving bone formation, e.g. preventing or treating calcium depletion, for example for preventing or treating any of the specific conditions or diseases hereinbefore set forth, in a subject in need of such a treatment which method comprises administering to said subject an effective amount of a) a compound of the invention and b) a

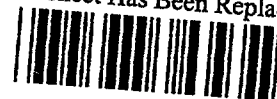
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second drug substance, said second drug substance being a bone resorption inhibitor, for example as indicated above.

Compounds of Examples 20, 29, 37, 49 and 50 are preferred.

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CLAIMS

1. A PTH compound having PTH-like activity and comprising at least one modification, said modification being either
 1. at least one radical selected from a L- or D- α -amino acid, C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to the terminal amino group of the PTH compound, and/or at least one radical selected from C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to one or more side chain amino groups of the PTH compound,

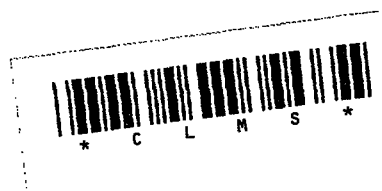
or

 - 2. at least one α -amino acid unit in the positions 1 to 38 of a naturally occurring PTH sequence being replaced by a natural or unnatural amino acid unit optionally in protected form, whereby the α -amino acid units present in positions 1 and 2 at the amino terminus of the PTH sequence together may be replaced by a pseudo-peptide,
- or a combination of such modifications,

provided that

when the PTH compound is free from D- or L- α -amino acid attached to the N-terminus or from C₂₋₆alcoxycarbonyl or optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, C₂₋₈alkynyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl, it is other than a PTH compound having a naturally occurring α -amino acid sequence;

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or the PTH compound is other than PTH(1-34) wherein

- i. the α -amino acid in position 1 is Gly, D-Ser, D-Ala or Tyr;
or
- ii. the α -amino acid in position 2 is Ala, D-Val, Lys, Arg or Cit and the α -amino acid in position 34 is Tyr; or the α -amino acid in position 2 is D-Val and the α -amino acid in position 34 is D-Tyr and optionally the α -amino acids in positions 8 and 18 are each Nle; or
- iii. the α -amino acid in positions 3 and/or 6 and/or 9 are replaced by a natural or unnatural amino acid; or
- iv. the α -amino acid in position 23 is replaced by Ala, Arg, Asn, Asp, Cys, Gln, Glu, Gly, His, Ile, Lys, Met, Pro, Ser or Thr; or
- v. the α -amino acid in position 25 and/or 26 and/or 27 is replaced by Ala, Asn, Asp, Cys, Gln, Glu, Gly, His, Ile, Leu, Met, Phe, Pro, Ser, Thr, Trp, Tyr or Val; or
- vi. the α -amino acids in positions 8 and 18 are each Nle or each Met(O) and optionally the α -amino acid in position 34 is Tyr; or the α -amino acids in positions 8 and 18 are each Nle and the α -amino acid in position 34 is Tyr and either the α -amino acid in position 12 is L- or D-Pro, L- or D-Ala, Aib or NMeGly or the α -amino acid in position 23 is Phe, Leu, Nle, Val, Tyr, α -Nal or β -Nal; or
- vii. the α -amino acid in position 28 is Lys and the α -amino acid in position 30 is Leu; or
- viii. the α -amino acid in position 1 is Aib; and/or the α -amino acid in position 8 and/or 18 is Leu, Ile, Val, Phe or Trp; and/or the α -amino acid in position 11 is Ser, Lys, Phe, β -Nal, Trp or Tyr; and/or the α -amino acid in position 12 is D-Leu, D-Ile, D-Nle,

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D-Val, D-Ser, D-Ser(Butyl), D-Abu, D-Thr, D-Nva, D-Met, D- β -Nal, D-Trp, D-Lys, D-Tyr, D-Lys(Fmoc), D-Phe or D-Asn; and/or the α -amino acid in position 13 is Leu; and/or the α -amino acid in position 19 and/or in position 21 is Arg, Lys, Asn or His; and/or the α -amino acid in position 23 is 2-(1,3-dithiolane-2-yl)Trp; and/or the α -amino acid in position 25 and/or in position 26 is His; and/or the α -amino acid in position 27 is Gln or Leu; or

- ix. the α -amino acid in position 8 and/or 18 is Ala or Ser; or the α -amino acid in position 8 and/or 18 is Ala, Val, Leu, Ile, Ser or Trp and the α -amino acid in position 34 is Tyr; or

the PTH compound is other than PTH(1-84) wherein

- i. the α -amino acid in position 1 is Tyr, Val, Pro, Asp or Cys; or
- ii. the α -amino acid in position 2 is Ala, Glu, Leu, Ser or Arg; or
- iii. the α -amino acid in positions 3 and/or 6 and/or 9 are replaced by a natural or unnatural amino acid; or
- iv. the α -amino acid in position 23 is replaced by Ala, Arg, Asn, Asp, Cys, Gln, Glu, Gly, His, Ile, Lys, Met, Pro, Ser or Thr; or
- v. the α -amino acid in position 25 and/or 26 and/or 27 is replaced by Ala, Asn, Asp, Cys, Gln, Glu, Gly, His, Ile, Leu, Met, Phe, Pro, Ser, Thr, Trp, Tyr or Val; or
- vi. the α -amino acid in position 8 is Met, Met(O), Ala, Val, Leu, Ile, Ser, Trp, Asn, Gln, Asp, Glu, Lys, Arg, Tyr or Gly and the α -amino acid in position 18 is Leu; or the α -amino acid in position 8 and/or 18 is Ala, Val, Leu, Ile, Ser or Trp, and optionally the α -amino acid

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in position 34 is Tyr; or the α -amino acid in position 8 and in position 18 are each Met(O); or the α -amino acid in position 8 is Leu and the α -amino acid in position 18 is Met(O); or
vii. the α -amino acid in position 26 is Gln;

the compound is other than Pro⁰PTH(1-84) or [Met⁰,Leu⁸,Leu¹⁸]PTH(1-84); or

the PTH compound is other than hPTH(1-36) wherein the α -amino acid in position 36 is Leu;

in free form or in salt form or complex form.

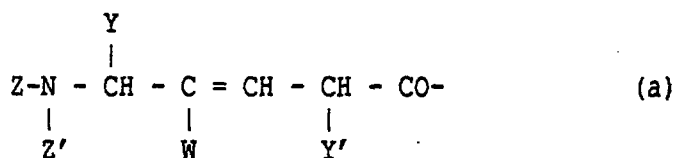
2. A PTH compound according to claim 1 comprising at least one amino acid unit replaced in one of the following positions of the PTH sequence: 1, 2, 3, 8-11, 13 to 19, 21, 22, 29 to 34.
3. A PTH compound according to claim 1 or 2 wherein the α -amino acid units in positions 1 and 2 at the amino terminus of the PTH sequence is replaced by a pseudo-di-peptide.
4. A PTH compound according to claim 3 of formula I

X - P₁

I

wherein

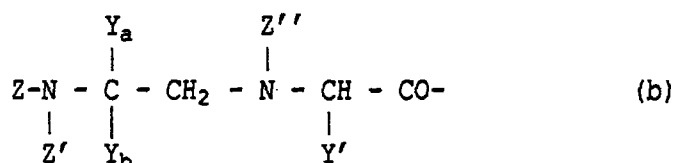
X is a residue of formula (a)



or (b)

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wherein each of Z and Z', independently, is H; optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl; or a protecting group, at most one of Z and Z' being a protecting group,

Z'' is H, C₁₋₈alkyl or a protecting group

each of Y and Y', independently, is an optionally protected side chain of a natural or unnatural α-amino acid,

one of Y_a and Y_b is hydrogen and the other is an

optionally protected side chain of a natural or unnatural

α-amino acid or Y_a and Y_b form together with the carbon

atom to which they are attached a C₃₋₆cycloalkyl group,

and

W is halogen or C₁₋₄alkyl,

or W and Y' together with the moiety $\begin{array}{c} \text{---C=CH-CH-} \\ | \quad | \end{array}$

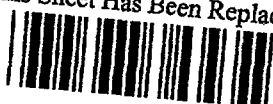
to which they are attached, form an optionally substituted aromatic cyclic or heterocyclic residue, and

P₁ is a PTH sequence in which amino acid units in positions 1 and 2 of the N-terminus are omitted.

5. A PTH compound according to claim 1 or 2, wherein the α-amino acid unit in position 1 and/or the α-amino acid unit in position 2 of the N-terminus of the PTH sequence is replaced by an optionally protected natural or unnatural amino acid residue, provided that

when the PTH compound is free from D- or L-α-amino acid attached to the N-terminus or from C₂₋₆alkoxycarbonyl or

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optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl, it is other than a PTH compound having a naturally occurring α -amino acid sequence;

or the PTH compound is other than PTH(1-34) wherein

- i. the α -amino acid in position 1 is Aib, Gly, D-Ser, D-Ala or Tyr; or
- ii. the α -amino acid in position 2 is Ala, D-Val, Lys, Cit or Arg and the α -amino acid in position 34 is Tyr; or the α -amino acid in position 2 is D-Val and the α -amino acid in position 34 is D-Tyr and optionally the α -amino acids in positions 8 and 18 are each Nle; or
- iii. the α -amino acid in position 1 is Aib and at least one further α -amino acid unit has been replaced as follows: the α -amino acid in position 8 and/or 18 is Leu, Ile, Val, Phe or Trp, and/or the α -amino acid in position 11 is Ser, Lys, Phe, β -Nal, Trp or Tyr, and/or the α -amino acid in position 12 is D-Leu, D-Ile, D-Nle, D-Val, D-Ser, D-Ser(Butyl), D-Abu, D-Thr, D-Nva, D-Met, D- β -Nal, D-Trp, D-Lys, D-Tyr, D-Lys(Fmoc), D-Phe or D-Asn, and/or the α -amino acid in position 13 is Leu, and/or the α -amino acid in position 19 and/or in position 21 is Arg, Lys, Asn or His, and/or the α -amino acid in position 23 is 2-(1,3-dithiolane-2-yl)Trp, and/or the α -amino acid in position 25 and/or in position 26 is His, and/or the α -amino acid in position 27 is Gln or Leu,

or the PTH compound is other than PTH(1-84) wherein

- i. the α -amino acid in position 1 is Tyr, Val, Pro, Asp or Cys; or



- ii. the α -amino acid in position 2 is Ala, Glu, Leu, Ser or Arg.

6. A PTH compound according to claim 5 of formula II



wherein

X_{1a} is a residue of an optionally protected natural α -amino acid or a residue of formula (IIa)



wherein

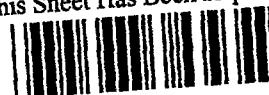
each of Z_a and Z'_a , independently, is H, C_{1-6} alkyl or a protecting group, at most one of Z_a and Z'_a being a protecting group, and either n is 1 and

- i. each of R_1 and R_2 , independently, is C_{1-6} alkyl, or
- ii. one of R_1 and R_2 is methyl or ethyl and the other is an optionally protected residue attaching to the α -carbon atom of a natural α -amino acid other than Ala, Leu, Ile or Val, or
- iii. one of R_1 and R_2 is H, methyl or ethyl and the other is an optionally protected residue attaching to the α -carbon atom of an unnatural α -amino acid, or
- iv. R_1 and R_2 form together with the carbon atom to which they are attached a C_{3-6} cycloalkyl group, or

- v. the residue $Z_a - N - C -$ represents a heterocyclic
- $$\begin{array}{c} R_1 \\ | \\ Z_a - N - C - \\ | \quad | \\ Z'_a \quad R_2 \end{array}$$



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residue optionally condensed to a benzene ring,

or n is 2, 3, 4 or 5 and each of R₁ and R₂ is H or CH₃,

Y_{2a} is a residue of an optionally protected natural α-amino acid
or a residue of formula (IIb)



Z_b is H or C₁₋₆alkyl and either m is 1 and

- i. each of R₃ and R₄, independently, is C₁₋₆alkyl, or
- ii. one of R₃ and R₄ is methyl or ethyl and the other is an optionally protected residue attaching to the α-carbon atom of a natural α-amino acid other than Ala, Leu, Ile or Val, or
- iii. one of R₃ and R₄ is H, methyl or ethyl and the other is an optionally protected residue attaching to the α-carbon atom of an unnatural α-amino acid,

or m is 2, 3, 4 or 5 and each of R₃ and R₄ is H or CH₃,
and

P₁ is as defined in claim 4.

7. A PTH compound according to any one of the preceding claims wherein the α-amino acid unit in position 8 and/or the α-amino acid unit in position 18 is replaced by an optionally protected natural or unnatural amino acid residue, provided that

when the PTH compound is free from D- or L-α-amino acid attached to the N-terminus or from C₂₋₆alkoxycarbonyl or



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optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl, it is other than a PTH compound having a naturally occurring α -amino acid sequence;

or the PTH compound is other than PTH(1-34) wherein

- i. the α -amino acids in positions 8 and 18 are each Nle or each Met(O) and optionally the α -amino acid in position 34 is Tyr; or the α -amino acids in positions 8 and 18 are each Nle and the α -amino acid in position 34 is Tyr and either the α -amino acid in position 12 is L- or D-Pro, L- or D-Ala, Aib or NMeGly or the α -amino acid in position 23 is Phe, Leu, Nle, Val, Tyr, α -Nal or β -Nal; or the α -amino acids in positions 8 and 18 are each Nle and the α -amino acid in position 2 is D-Val and the α -amino acid in position 34 is D-Tyr; or
- ii. the α -amino acid in position 8 and/or 18 is Leu, Ile, Val, Phe or Trp and optionally at least one further α -amino acid unit has been replaced as follows: the α -amino acid in position 1 is Aib and/or the α -amino acid in position 11 is Ser, Lys, Phe, β -Nal, Trp or Tyr, and/or the α -amino acid in position 12 is D-Leu, D-Ile, D-Nle, D-Val, D-Ser, D-Ser(Butyl), D-Abu, D-Thr, D-Nva, D-Met, D- β -Nal, D-Trp, D-Lys, D-Tyr, D-Lys(Fmoc), D-Phe or D-Asn, and/or the α -amino acid in position 13 is Leu, and/or the α -amino acid in position 19 and/or in position 21 is Arg, Lys, Asn or His, and/or the α -amino acid in position 23 is 2-(1,3-dithiolane-2-yl)Trp, and/or the α -amino acid in position 25 and/or in position 26 is His, and/or the α -amino acid in position 27 is Gln or Leu; or



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iii. the α -amino acid in position 8 and/or 18 is Ala or Ser; or the α -amino acid in position 8 and/or 18 is Leu, and the α -amino acid in position 34 is Tyr; or

the PTH compound is other than PTH(1-84) wherein

the α -amino acid in position 8 is Met, Met(O), Ala, Val, Leu, Ile, Ser, Trp, Asn, Gln, Asp, Glu, Lys, Arg, Tyr or Gly and the α -amino acid in position 18 is Leu; or the α -amino acid in position 8 and in position 18 are each Met(O); or the α -amino acid in position 8 is Leu and the α -amino acid in position 18 is Met(O); or the PTH compound is other than [Leu¹⁸,Tyr³⁴]hPTH(1-84), [Leu⁸,Tyr³⁴]hPTH(1-84), [Ile⁸,Leu¹⁸,Tyr³⁴]hPTH(1-84) or [Leu⁸,Leu¹⁸,Tyr³⁴]hPTH(1-84).

8. A PTH compound having PTH-like activity wherein at least one of the α -amino acid units in the positions 1 to 38 of a PTH sequence is replaced by the α -amino acid unit which is present at the corresponding position in PTHrP, the PTH compound optionally comprising at least one radical selected from a L- or D- α -amino acid, C₂₋₆alkoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to the terminal amino group of the PTH compound, and/or at least one radical selected from C₂₋₆alkoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to one or more side chain amino groups of the PTH compound, provided that

when the PTH compound is free from D- or L- α -amino acid attached to the N-terminus or from C₂₋₆alkoxycarbonyl or optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl,

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aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl, it is other than a PTH compound having a naturally occurring α -amino acid sequence;

or the PTH compound is other than PTH(1-34) wherein

- i. the α -amino acid in position 8 and/or 18 is Leu; and/or the α -amino acid in position 11 is Lys; and/or the α -amino acid in position 19 and/or 21 is Arg; and/or the α -amino acid in position 25 and/or 26 is His; and/or the α -amino acid in position 27 is Leu; and optionally at least one further α -amino acid unit has been replaced as follows: the α -amino acid in position 1 is Aib and/or the α -amino acid in position 11 is Ser, Phe, β -Nal, Trp or Tyr, and/or the α -amino acid in position 12 is D-Leu, D-Ile, D-Nle, D-Val, D-Ser, D-Ser(Butyl), D-Abu, D-Thr, D-Nva, D-Met, D- β -Nal, D-Trp, D-Lys, D-Tyr, D-Lys(Fmoc), D-Phe or D-Asn, and/or the α -amino acid in position 13 is Leu, and/or the α -amino acid in position 19 and/or in position 21 is Lys, Asn or His, and/or the α -amino acid in position 23 is 2-(1,3-dithiolane-2-yl)Trp, and/or the α -amino acid in position 27 is Gln; or
 - ii. the α -amino acid in position 25 is Gln and/or the α -amino acid in position 26 is Asn, the α -amino acid in position 27 being optionally Leu; or
- the PTH compound is other than [Leu⁸,Leu¹⁸]PTH(1-84), in free form or in salt or complex form.

9. A PTH compound according to claim 8 wherein at least 2 α -amino acid units of the PTH sequence is replaced by the α -amino acid unit which is present at the corresponding position in PTHrP.

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10. A PTH compound having PTH-like activity wherein at least one of the α -amino acid units in the positions 8 to 11 of a PTH sequence is replaced by the α -amino acid unit which is present at the corresponding position in PTHrP, the PTH compound optionally comprising at least one radical selected from a L- or D- α -amino acid, C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to the terminal amino group of the PTH compound, and/or at least one radical selected from C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to one or more side chain amino groups of the PTH compound, provided that the PTH compound is other than [Leu⁸,Leu¹⁸]-PTH(1-84) or PTH(1-34) wherein the α -amino acid in position 8 is Leu and/or the α -amino acid in position 11 is Lys and optionally at least one further α -amino acid unit is replaced as follows: the α -amino acid unit in position 1 is Aib, and/or the α -amino acid in position 18 is Leu, Ile, Val, Phe or Trp, and/or the α -amino acid in position 11 is Ser, Phe, β -Nal, Trp or Tyr, and/or the α -amino acid in position 12 is D-Leu, D-Ile, D-Nle, D-Val, D-Ser, D-Ser(Butyl), D-Abu, D-Thr, D-Nva, D-Met, D- β -Nal, D-Trp, D-Lys, D-Tyr, D-Lys(Fmoc), D-Phe or D-Asn, and/or the α -amino acid in position 13 is Leu, and/or the α -amino acid in position 19 and/or in position 21 is Arg, Lys, Asn or His, and/or the α -amino acid in position 23 is 2-(1,3-dithiolane-2-yl)Trp, and/or the α -amino acid in position 25 and/or in position 26 is His, and/or the α -amino acid in position 27 is Gln or Leu, in free form or in salt or complex form.
11. A PTH compound which is [Leu⁸,Asp¹⁰]PTH or [Leu⁸,Asp¹⁰,Lys¹¹]PTH, the PTH compound optionally comprising at least one radical selected from a L- or D- α -amino acid,

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C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to the terminal amino group of the PTH compound, and/or at least one radical selected from C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to one or more side chain amino groups of the PTH compound, in free form or in salt or complex form.

12. A PTH compound having PTH-like activity wherein at least one of the α -amino acid units in the positions 16 to 19 of a PTH sequence is replaced by the α -amino acid unit which is present at the corresponding position in PTHrP, the PTH compound optionally comprising at least one radical selected from a L- or D- α -amino acid, C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to the terminal amino group of the PTH compound, and/or at least one radical selected from C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to one or more side chain amino groups of the PTH compound, provided that the PTH compound is other than [Leu⁸, Leu¹⁸]-PTH(1-84) or PTH(1-34) wherein the α -amino acid in position 18 is Leu and/or the α -amino acid in position 19 is Arg and optionally at least one further α -amino acid unit is replaced as follows: the α -amino acid unit in position 1 is Aib, and/or the α -amino acid in position 8 is Leu, Ile, Val, Phe or Trp, and/or the α -amino acid in position 11 is Ser, Lys, Phe, β -Nal, Trp or Tyr, and/or the α -amino acid in position 12 is D-Leu, D-Ile, D-Nle, D-Val, D-Ser, D-Ser(Butyl), D-Abu, D-Thr, D-Nva, D-Met, D- β -Nal, D-Trp, D-Lys, D-Tyr, D-Lys(Fmoc), D-Phe or D-Asn, and/or the α -amino acid in

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position 13 is Leu, and/or the α -amino acid in position 19 is Lys, Asn or His and/or the α -amino acid in position 21 is Arg, Lys, Asn or His, and/or the α -amino acid in position 23 is 2-(1,3-dithiolane-2-yl)Trp, and/or the α -amino acid in position 25 and/or in position 26 is His, and/or the α -amino acid in position 27 is Gln or Leu, in free form or in salt or complex form.

13. A PTH compound having PTH-like activity wherein at least one of the α -amino acid units in the positions 33 and 34 of a PTH sequence is replaced by the α -amino acid unit which is present at the corresponding position in PTHrP, the PTH compound optionally comprising at least one radical selected from a L- or D- α -amino acid, C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to the terminal amino group of the PTH compound, and/or at least one radical selected from C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to one or more side chain amino groups of the PTH compound, in free form or in salt or complex form.
14. A PTH compound according to claims 10 and 13 wherein at least 2 α -amino acid units in the positions 8-11 and 33-34 of a PTH sequence are replaced.
15. A PTH compound selected from [Thr³³,Ala³⁴]PTH, [Leu⁸,Thr³³,Ala³⁴]PTH and [Leu⁸,Asp¹⁰,Lys¹¹,Thr³³,Ala³⁴]PTH, the PTH compound optionally comprising at least one radical selected from a L- or D- α -amino acid, C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to the terminal amino group of the PTH compound, and/or at

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least one radical selected from C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to one or more side chain amino groups of the PTH compound, in free form or in salt or complex form.

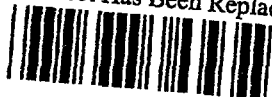
16. A PTH compound according to any one of claims 8 to 15, wherein one or more α -amino acid units in the remaining positions 1 to 38 is further replaced by a natural or unnatural amino acid unit.
17. A PTH compound according to claim 16 wherein the α -amino acid in position 10 is selected from Gly, Gln, Glu, His, Ser, Thr, Lys and Tyr.
18. A PTH compound according to claim 16 or 17, wherein the α -amino acid in position 13 is a D- or L- α -amino acid other than Arg.
19. A PTH compound according to claim 16, 17 or 18, wherein the α -amino acid in position 16 is a D- or L- α -amino acid.
20. A PTH compound according to any one of claims 16 to 19 wherein the α -amino acid in position 18 is Gln or Tyr.
21. A PTH compound according to any one of claims 16 to 20, wherein the α -amino acid in position 19 is selected from Ala, Arg, Val, Tyr, Ser, Lys, Met, His, Gly, Pro, Asn and Ile.
22. A PTH compound according to any one of claims 16 to 21, wherein the α -amino acid in position 33 is a D- or L- α -amino acid other than Thr, Phe or Ala.

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23. A PTH compound which is selected from [Leu⁸, Gln¹⁸, Thr³³, Ala³⁴]PTH, [Leu⁸, Ala¹⁶, Gln¹⁸, Thr³³, Ala³⁴]PTH, [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸, Thr³³, Ala³⁴]PTH, [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸]PTH, [Leu⁸, Ala¹⁶, Gln¹⁸, Ala¹⁹]PTH, [Leu⁸, Asp¹⁰, Lys¹¹, Gln¹⁸]PTH, [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸, Ala¹⁹]PTH, [Leu⁸, Ala¹⁶, Gln¹⁸, Ala¹⁹, Thr³³, Ala³⁴]PTH and [Leu⁸, Asp¹⁰, Lys¹¹, Gln¹⁸, Thr³³, Ala³⁴]PTH, the PTH compound optionally comprising at least one radical selected from a L- or D- α -amino acid, C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to the terminal amino group of the PTH compound, and/or at least one radical selected from C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to one or more side chain amino groups of the PTH compound, in free form or in salt or complex form.
24. A PTH compound according to any one of preceding claims which is a N-terminal PTH fragment comprising 1-34 to 1-38 amino acid units.
25. A PTH compound according to claim 24 which is a N-terminal hPTH fragment.
26. A PTH compound according to any one of claims 1 to 24 which is hPTH(1-34) or hPTH(1-36).
27. A PTH compound according to any one of the preceding claims wherein the C-terminus is -COOH, esterified -COOH, -CONH₂, mono- or disubstituted -CONH₂.

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28. A PTH compound selected from
[Leu⁸,Gln¹⁸,Thr³³,Ala³⁴]-hPTH(1-34)OH
[Leu⁸,Ala¹⁶,Gln¹⁸,Ala¹⁹,Thr³³,Ala³⁴]hPTH(1-34)OH
[Leu⁸,Ala¹⁶,Gln¹⁸,Thr³³,Ala³⁴]hPTH(1-34)OH
[Leu⁸,Asp¹⁰,Lys¹¹,Gln¹⁸]hPTH(1-36)OH
[Leu⁸,Asp¹⁰,Lys¹¹,Gln¹⁸,Thr³³,Ala³⁴]hPTH(1-34)OH
[Leu⁸,Asp¹⁰,Lys¹¹,Ala¹⁶,Gln¹⁸,Ala¹⁹]hPTH(1-36)OH
[Leu⁸,Asp¹⁰,Lys¹¹,Ala¹⁶,Gln¹⁸,Thr³³,Ala³⁴]hPTH(1-34)OH
[Leu⁸,Asp¹⁰,Lys¹¹,Ala¹⁶,Gln¹⁸]hPTH(1-36)OH, and
[Leu⁸,Ala¹⁶,Gln¹⁸,Ala¹⁹]hPTH(1-36)OH

in free form or in salt or complex form.

29. A process for the production of a desired medium-sized polypeptide in which a fusion protein comprising an N-terminal bacteriophage T4 gene 55 polypeptide having the desired polypeptide linked to the C-terminal thereof, is expressed in bacterial cells.
30. A process according to claim 29 in which the fusion protein comprises a chemically cleavable linker between the gene 55 and the desired polypeptides.
31. A process according to claim 30 in which the cleavable linker comprises the amino acid sequences Asp-Pro-Pro or Asn-Gly-Pro.
32. A process according to any one of the claims 29 to 31 in which the desired polypeptide is selected from the group consisting of calcitonins, endorphins, gastric inhibitory peptide(s), glucagon, neuropeptide Y, growth hormone releasing factor (GHRF), amyloid β protein fragments, hirudin, insulin, somatostatin, epidermal growth factor,

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nerve growth factor, parathyroid hormone (PTH), and fragments thereof.

33. A nucleotide sequence which codes for a fusion protein comprising an N-terminal bacteriophage T4 gene 55 polypeptide and a desired medium-sized polypeptide linked to the C-terminal thereof.
34. A bacterial expression vector comprising a DNA sequence according to claim 33.
35. Bacterial host cells transformed with a DNA sequence according to claim 33, or a bacterial vector according to claim 34.
36. A process for the production of a desired medium-sized polypeptide comprising causing bacterial host cells according to claim 35 to express the fusion protein in the form of inclusion bodies; isolating the inclusion bodies; solubilising the inclusion bodies, and cleaving the desired polypeptide from the fusion protein.
37. A fusion protein comprising an N-terminal bacteriophage T4 gene 55 polypeptide and a desired medium-sized polypeptide linked to the C-terminal thereof.
38. A process for the production of a desired medium-sized polypeptide comprising cleaving the desired polypeptide from a fusion protein according to claim 37, and optionally removing unwanted amino acid residues from the N-terminus thereof.

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39. A process according to any one of claims 29-32, 36 or 38 in which the desired medium-sized polypeptide comprises a PTH polypeptide.
40. A nucleotide sequence according to claim 33, in which the desired medium-sized polypeptide comprises a PTH polypeptide.
41. A bacterial expression vector comprising a DNA sequence according to claim 40.
42. Bacterial host cells transformed with a DNA sequence according to claim 31 or a bacterial expression vector according to claim 41.
43. A process for the production of a medium-sized polypeptide comprising:
- culturing bacterial host cells transformed with a vector containing a DNA sequence coding for expression of a fusion protein comprising an N-terminal bacteriophage T4 polypeptide having the desired polypeptide linked to the C-terminal thereof by a chemically cleavable linker selected from the group consisting of peptides containing the amino acid sequence Asp-Pro-Pro or Asn-Gly-Pro and expressing the fusion protein in the form of inclusion bodies;
 - isolating the inclusion bodies;
 - solubilising the inclusion bodies under acid conditions;
 - cleaving the desired polypeptide from the fusion protein, and
 - removing unwanted N-terminal amino acid residues from the desired polypeptide product.

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44. A PTH compound according to any one of claims 1 to 28 for use as a pharmaceutical.
45. A pharmaceutical composition comprising a PTH compound as defined in any one of claims 1 to 28, in free form or in physiologically acceptable salt form, together with a pharmaceutically acceptable diluent or carrier therefor.
46. A PTH compound substantially as herein before described with reference to any one of Examples 1 to 300, 302-304 and 306 to 312.

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FIGURE 1

AGATCTCGAT CCCGCGAAAT TAATACGACT CACTATAGGG AGACCACAAC GGTTTCCCTC	60
TAGAAATAAT TTTGTTTAAC TTTAAGAAGG AGATATACAT ATG TCA GAA ACT AAG	115
Met Ser Glu Thr Lys	
1 5	
CCT AAA TAT AAT TAC GTA AAC AAT AAA GAG CTT TTA CAA GCT ATT ATT	163
Pro Lys Tyr Asn Tyr Val Asn Asn Lys Glu Leu Leu Gln Ala Ile Ile	
10 15 20	
GAT TGG AAA ACA GAA TTA GCA AAT AAT AAA GAC CCA AAT AAA GTA GTT	211
Asp Trp Lys Thr Glu Leu Ala Asn Asn Lys Asp Pro Asn Lys Val Val	
25 30 35	
CGT CAG AAT GAT ACT ATC GGA TTA GCC ATT ATG CTT ATT GCA GAA GGC	259
Arg Gln Asn Asp Thr Ile Gly Leu Ala Ile Met Leu Ile Ala Glu Gly	
40 45 50	
TTA TCT AAA CGT TTC AAC TTT TCA GGA TAC ACC CAG TCT TGG AAA CAA	307
Leu Ser Lys Arg Phe Asn Phe Ser Gly Tyr Thr Gln Ser Trp Lys Gln	
55 60 65	
GAA ATG ATT GCA GAT GGT ATA GAA GCT TCT ATT AAG GGG CTT CAC AAT	355
Glu Met Ile Ala Asp Gly Ile Glu Ala Ser Ile Lys Gly Leu His Asn	
70 75 80 85	
TTT GAT GAA ACG AAA TAT AAA AAC CCA CAT GCG TAT ATA ACT CAA GCT	403
Phe Asp Glu Thr Lys Tyr Lys Asn Pro His Ala Tyr Ile Thr Gln Ala	
90 95 100	

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TGT TTT AAT GCA TTC GTC CAA CGT GGA TCC ATC GAT CCA CCA TCC GTA	451
Cys Phe Asn Ala Phe Val Gln Arg Gly Ser Ile Asp Pro Pro Ser Val	
105 110 115	
TCA GAA ATA CAA CTA ATG CAT AAT CTG GGT AAA CAT CTG AAT TCA ATG	499
Ser Glu Ile Gln Leu Met His Asn Leu Gly Lys His Leu Asn Ser Met	
120 125 130	
GAA CGT GTA GAA TGG CTG CGT AAA AAA CTG CAG GAT GTA CAT AAT TTT	547
Glu Arg Val Glu Trp Leu Arg Lys Lys Leu Gln Asp Val His Asn Phe	
135 140 145	
GTA GCT CTG GGT TAACTCGAGC AGATCCGGCT GCTAACAAAG CCCGAAAGGA	599
Val Ala Leu Gly	
150	
AGCTGAGTTG GCTGCTGCCA CCGCTGAGCA ATAAGTAGCA TAACCCCTTG GGGCCCTTGG	659
GGCCTCTAAA CGGGTCTTGA GGGGTTTTTT GCTGAAAGGA GGAAGTATAT CCGGATAA	717

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FIGURE 2

AGATCTCGAT CCCGCGAAAT TAATACGACT CACTATAGGG AGACCACAAC GGTTTCCCTC	60
TAGAAATAAT TTTGTTTAAC TTTAAGAAGG AGATATACAT ATG TCA GAA ACT AAG	115
Met Ser Glu Thr Lys	
1 5	
CCT AAA TAT AAT TAC GTA AAC AAT AAA GAG CTT TTA CAA GCT ATT ATT	163
Pro Lys Tyr Asn Tyr Val Asn Asn Lys Glu Leu Leu Gln Ala Ile Ile	
10 15 20	
GAT TGG AAA ACA GAA TTA GCA AAT AAT AAA GAC CCA AAT AAA GTA GTT	211
Asp Trp Lys Thr Glu Leu Ala Asn Asn Lys Asp Pro Asn Lys Val Val	
25 30 35	
CGT CAG AAT GAT ACT ATC GGA TTA GCC ATT ATG CTT ATT GCA GAA GGC	259
Arg Gln Asn Asp Thr Ile Gly Leu Ala Ile Met Leu Ile Ala Glu Gly	
40 45 50	
TTA TCT AAA CGT TTC AAC TTT TCA GGA TAC ACC CAG TCT TGG AAA CAA	307
Leu Ser Lys Arg Phe Asn Phe Ser Gly Tyr Thr Gln Ser Trp Lys Gln	
55 60 65	
GAA ATG ATT GCA GAT GGT ATA GAA GCT TCT ATT AAG GGG CTT CAC AAT	355
Glu Met Ile Ala Asp Gly Ile Glu Ala Ser Ile Lys Gly Leu His Asn	
70 75 80 85	
TTT GAT GAA ACG AAA TAT AAA AAC CCA CAT GCG TAT ATA ACT CAA GCT	403
Phe Asp Glu Thr Lys Tyr Lys Asn Pro His Ala Tyr Ile Thr Gln Ala	
90 95 100	

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TGT TTT AAT GCA TTC GTC CAA CGT ATT AAA AAA GAA CGT AAG GAA GTT	451
Cys Phe Asn Ala Phe Val Gln Arg Ile Lys Lys Glu Arg Lys Glu Val	
105 110 115	
GCA AAG AAA TAT AGT TAC TTC GTT CAC AAT GTC TAT GAC AGC CGT GAC	499
Ala Lys Lys Tyr Ser Tyr Phe Val His Asn Val Tyr Asp Ser Arg Asp	
120 125 130	
GAC GAT ATG GTT GCG TTA GTA GAT GAA ACT TTT ATT CAA GAC ATC TAT	547
Asp Asp Met Val Ala Leu Val Asp Glu Thr Phe Ile Gln Asp Ile Tyr	
135 140 145	
GAT AAA ATG ACG CAT TAC GAA GAA TCA ACC TAT AGA ACA CCG GGG GCT	595
Asp Lys Met Thr His Tyr Glu Glu Ser Thr Tyr Arg Thr Pro Gly Ala	
150 155 160 165	
GAA AAG AAA AGT GTT GTA GAT GAT TCT CCT AGT TTG GAT TTT TTA TAT	643
Glu Lys Lys Ser Val Val Asp Asp Ser Pro Ser Leu Asp Phe Leu Tyr	
170 175 180	
GAG GCT AAC GAT GGA TCC GTT AAC GGT CCA TCC GTA TCA GAA ATA CAA	691
Glu Ala Asn Asp Gly Ser Val Asn Gly Pro Ser Val Ser Glu Ile Gln	
185 190 195	
CTA ATG CAT AAT CTG GGT AAA CAT CTG AAT TCA ATG GAA CGT GTA GAA	739
Leu Met His Asn Leu Gly Lys His Leu Asn Ser Met Glu Arg Val Glu	
200 205 210	
TGG CTG CGT AAA AAA CTG CAG GAT GTA CAT AAT TTT GTA GCT CTG GGT	787
Trp Leu Arg Lys Lys Leu Gln Asp Val His Asn Phe Val Ala Leu Gly	
215 220 225	

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TAAC	TCGAGC	AGAT	CCGGCT	GCTA	ACAAAG	CCCG	AAAGGA	AGCT	GAGTTG	GCTG	CTGCCA	847
CCGCT	GAGCA	ATAA	CTAGCA	TAAC	CCCTTG	GGGC	CCCTTG	GGCT	CTAAA	CGGT	CTTGA	907
GGGT	TTTTTT	GCTG	AAAGGA	GGA	ACTATAT	CCG	GATAA					945

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Abstract of the invention

PTH compounds having PTH-like activity and comprising at least one modification, said modification being either

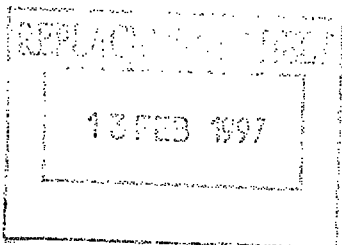
1. at least one radical selected from a L- or D- α -amino acid, C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to the terminal amino group of the PTH compound, and/or at least one radical selected from C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to one or more side chain amino groups of the PTH compound,

or

2. at least one α -amino acid unit in the positions 1 to 38 of a naturally occurring PTH sequence being replaced by a natural or unnatural amino acid unit optionally in protected form, whereby the α -amino acid units present in positions 1 and 2 at the amino terminus of the PTH sequence may be replaced by a pseudo-peptide,

or a combination of such modifications,

in free form or in salt form, have pharmacological activity, e.g. for preventing or treating all bone conditions which are associated with increased calcium depletion or resorption or in which calcium fixation in the bone is desirable.



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or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to one or more side chain amino groups of the PTH compound,

or

2. at least one α -amino acid unit in the positions 1 to 38 of a naturally occurring PTH sequence being replaced by a natural or unnatural amino acid unit optionally in protected form, whereby the α -amino acid units present in positions 1 and 2 at the amino terminus of the PTH sequence together may be replaced by a pseudo-peptide, or a combination of such modifications,

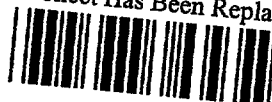
provided that

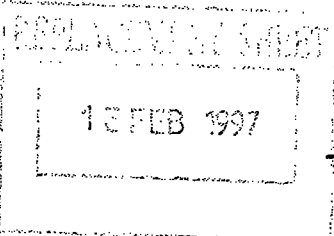
when the PTH compound is free from D- or L- α -amino acid attached to the N-terminus or from C₂₋₆alcoxycarbonyl or optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, C₂₋₈alkynyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl, it is other than a PTH compound having a naturally occurring α -amino acid sequence;

or the PTH compound is other than PTH(1-34) wherein

- i. the α -amino acid in position 1 is Gly, D-Ser, D-Ala or Tyr; or
- ii. the α -amino acid in position 2 is Ala, D-Val, Lys, Arg or Cit and the α -amino acid in position 34 is Tyr; or the α -amino acid in position 2 is D-Val and the α -amino acid in position 34 is D-Tyr and optionally the α -amino acids in positions 8 and 18 are each Nle; or
- iii. the α -amino acid in positions 3 and/or 6 and/or 9 are replaced by a natural or unnatural amino acid; or
- iv. the α -amino acid in position 23 is replaced by Ala, Arg, Asn, Asp, Cys, Gln, Glu, Gly, His, Ile, Lys, Met, Pro, Ser or Thr; or
- v. the α -amino acid in position 25 and/or 26 and/or 27 is replaced by Ala, Asn, Asp, Cys, Gln, Glu, Gly, His, Ile, Leu, Met, Phe, Pro, Ser, Thr, Trp, Tyr or Val; or
- vi. the α -amino acids in positions 8 and 18 are each Nle or each Met(O) and optionally the α -amino acid in position 34 is Tyr; or the α -amino acids in positions 8 and 18 are each Nle and the

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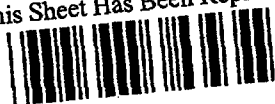




3,4-dimethyl; arylmethyl such as benzyl optionally ring substituted with p-methoxy, p-nitro or p-chloro; or arylsulfonyl such as phenylsulfonyl optionally ring substituted with p-methyl or p-methoxy, or naphthylsulfonyl optionally ring substituted with e.g. amino or di(C₁₋₄alkyl)amino.

O-protecting groups of O-containing side chains are e.g. as described in "The Peptides", 3, (1981), E. Gross and J. Meienhofer (Ed.). For aliphatic hydroxy functionalities, suitable O-protecting groups are e.g. as disclosed in the above reference in "Protection of the Hydroxy Group", J.M. Stewart, 169-201 and include the benzyl, t.-butyl and methyl groups. For aromatic hydroxy functionalities, suitable O-protecting groups include the benzyl, t.-butyl, methyl, tosyl and benzyloxycarbonyl groups. O-protecting groups for carboxy functionalities on amino acid side chains are well known ester groups and described e.g. in "The Peptides", 3, 101-135 ("Carboxyl Protecting Groups" by R.W. Roeske) and include the methyl, ethyl, t.-butyl and benzyl groups. S-protecting groups for thiol functionalities on amino acid side chains are known and described e.g. in "The Peptides", 3, 137-167 ("Sulfhydryl Group Protection" by R.G. Hiskey). Examples include the methyl, t.-butyl, benzyl, p-methoxyphenyl-methyl, ethylamino-carbonyl and benzyloxycarbonyl groups.

According to the invention the PTH compounds may bear on their terminal amino group at least one radical selected from a L- or D- α -amino acid, C₂₋₆alkoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl, and/or on one or more side chain amino group(s) at least one radical selected from C₂₋₆alkoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl. When such a group is attached to a side chain amino group, it is preferably on the ϵ -amino group of a Lys unit. Preferred substituents for



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REPLACED SHEETS		8G
✓ Description	: paged 1 to paged 65	
✓ Claim(s)	: paged 66 to paged 85	
✓ Drawing(s)	: paged 15 to paged 56	
✓ Abstract	: paged 85 to paged 86	
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✓ Claim(s)	: paged 66 to paged 85	
✓ Drawing(s)	: paged 15 to paged 56	
✓ Abstract	: paged 85 to paged 86	
DATE OF RECEIPT 22/5/97		

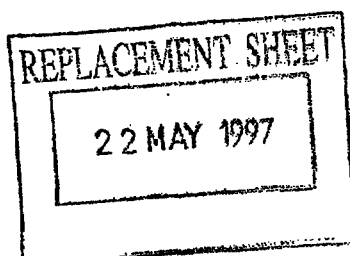
PEPTIDES

The present invention relates to variants of parathyroid hormone (PTH), a process for their production, pharmaceutical preparations comprising them and their use as a pharmaceutical.

The term "PTH" as used herein refers to any genetically encoded form of parathyroid hormone, including the mature form containing 84 amino acids of a given vertebrate PTH species, e.g human, porcine, rat, bovine, chicken, and fragments thereof as well as analogues and derivatives thereof having PTH-like activity. The position of each amino acid involved in the PTH sequence is numbered according to the internationally accepted procedure. For consistency and as is conventional, in the following description, the same numbering system will be applied to the amino acids of the PTH sequence independently of the substitution pattern in the molecule.

More particularly, the present invention provides a PTH compound having PTH-like activity and comprising at least one modification, said modification being either

1. at least one radical selected from a L- or D- α -amino acid, C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to the terminal amino group of the PTH compound, and/or at least one radical selected from C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl



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or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to one or more side chain amino groups of the PTH compound,

or

2. at least one α -amino acid unit in the positions 1 to 38 of a naturally occurring PTH sequence being replaced by a natural or unnatural amino acid unit optionally in protected form, whereby the α -amino acid units present in positions 1 and 2 at the amino terminus of the PTH sequence together may be replaced by a pseudo-peptide, or a combination of such modifications,

provided that

when the PTH compound is free from D- or L- α -amino acid attached to the N-terminus or from C₂₋₆alkoxycarbonyl or optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, C₂₋₈alkynyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl, it is other than a PTH compound having a naturally occurring α -amino acid sequence;

or the PTH compound is other than PTH(1-34) wherein

- i. the α -amino acid in position 1 is Gly, D-Ser, D-Ala or Tyr; or
- ii. the α -amino acid in position 2 is Ala, D-Val, Lys, Arg or Cit and the α -amino acid in position 34 is Tyr; or the α -amino acid in position 2 is D-Val and the α -amino acid in position 34 is D-Tyr and optionally the α -amino acids in positions 8 and 18 are each Nle; or
- iii. the α -amino acid in positions 3 and/or 6 and/or 9 are replaced by a natural or unnatural amino acid; or
- iv. the α -amino acid in position 23 is replaced by Ala, Arg, Asn, Asp, Cys, Gln, Glu, Gly, His, Ile, Lys, Met, Pro, Ser or Thr; or
- v. the α -amino acid in position 25 and/or 26 and/or 27 is replaced by Ala, Asn, Asp, Cys, Gln, Glu, Gly, His, Ile, Leu, Met, Phe, Pro, Ser, Thr, Trp, Tyr or Val; or
- vi. the α -amino acids in positions 8 and 18 are each Nle or each Met(O) and optionally the α -amino acid in position 34 is Tyr; or the α -amino acids in positions 8 and 18 are each Nle and the

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α -amino acid in position 34 is Tyr and either the α -amino acid in position 12 is L- or D-Pro, L- or D-Ala, Aib or NMeGly or the α -amino acid in position 23 is Phe, Leu, Nle, Val, Tyr, α -Nal or β -Nal; or

- vii. the α -amino acid in position 28 is Lys and the α -amino acid in position 30 is Leu; or
- viii. the α -amino acid in position 1 is Aib; and/or the α -amino acid in position 8 and/or 18 is Leu, Ile, Val, Phe or Trp; and/or the α -amino acid in position 11 is Ser, Lys, Phe, β -Nal, Trp or Tyr; and/or the α -amino acid in position 12 is D-Leu, D-Ile, D-Nle, D-Val, D-Ser, D-Ser(Butyl), D-Abu, D-Thr, D-Nva, D-Met, D- β -Nal, D-Trp, D-Lys, D-Tyr, D-Lys(Fmoc), D-Phe or D-Asn; and/or the α -amino acid in position 13 is Leu; and/or the α -amino acid in position 19 and/or in position 21 is Arg, Lys, Asn or His; and/or the α -amino acid in position 23 is 2-(1,3-dithiolane-2-yl)Trp; and/or the α -amino acid in position 25 and/or in position 26 is His; and/or the α -amino acid in position 27 is Gln or Leu; or
- ix. the α -amino acid in position 8 and/or 18 is Ala or Ser; or the α -amino acid in position 8 and/or 18 is Ala, Val, Leu, Ile, Ser or Trp and the α -amino acid in position 34 is Tyr; or

the PTH compound is other than PTH(1-84) wherein

- i. the α -amino acid in position 1 is Tyr, Val, Pro, Asp or Cys; or
- ii. the α -amino acid in position 2 is Ala, Glu, Leu, Ser or Arg; or
- iii. the α -amino acid in positions 3 and/or 6 and/or 9 are replaced by a natural or unnatural amino acid; or
- iv. the α -amino acid in position 23 is replaced by Ala, Arg, Asn, Asp, Cys, Gln, Glu, Gly, His, Ile, Lys, Met, Pro, Ser or Thr; or
- v. the α -amino acid in position 25 and/or 26 and/or 27 is replaced by Ala, Asn, Asp, Cys, Gln, Glu, Gly, His, Ile, Leu, Met, Phe, Pro, Ser, Thr, Trp, Tyr or Val; or

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- vi. the α -amino acid in position 8 is Met, Met(O), Ala, Val, Leu, Ile, Ser, Trp, Asn, Gln, Asp, Glu, Lys, Arg, Tyr or Gly and the α -amino acid in position 18 is Leu; or the α -amino acid in position 8 and/or 18 is Ala, Val, Leu, Ile, Ser or Trp, and optionally the α -amino acid in position 34 is Tyr; or the α -amino acid in position 8 and in position 18 are each Met(O); or the α -amino acid in position 8 is Leu and the α -amino acid in position 18 is Met(O); or
- vii. the α -amino acid in position 26 is Gln;

the compound is other than $\text{Pro}^0\text{PTH}(1-84)$ or $[\text{Met}^0, \text{Leu}^8, \text{Leu}^{18}]\text{PTH}(1-84)$; or

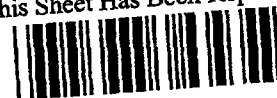
the PTH compound is other than $\text{hPTH}(1-36)$ wherein the α -amino acid in position 36 is Leu;

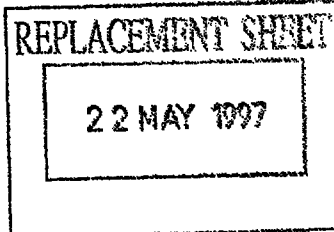
in free form or in salt form or complex form.

When the PTH sequence is derived from a PTH fragment, it is a PTH fragment having PTH-like activity and comprising at least the first 27 N-terminal amino acid units of PTH, preferably up to the first 38 N-terminal amino acid units of PTH, e.g. 1-34 to 1-38, e.g. 1-34, 1-36, 1-37 or 1-38 PTH, at least one of the α -amino acids being replaced according to the invention. One or more α -amino acid units normally present in the naturally occurring PTH sequence may also be omitted. hPTH fragments, particularly $\text{hPTH}(1-34)$ and $\text{hPTH}(1-36)$, are preferred.

The C-terminus of the PTH compounds may be $-\text{COOH}$, esterified $-\text{COOH}$, e.g. $-\text{COOR}_a$ wherein R_a is lower alkyl, for example C_{1-4} alkyl, $-\text{CONH}_2$ or a mono- or disubstituted amide, e.g. $-\text{CONR}_b\text{R}_c$ wherein one of R_b and R_c is H and the other is an aliphatic residue, e.g. C_{1-6} alkyl, or both are an aliphatic residue, or R_b and R_c together with the nitrogen atom to which

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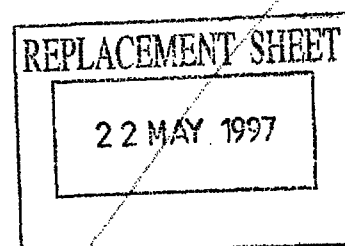
they are attached form a heterocyclic residue, e.g. pyrrolidinyl or piperidinyl.

Hereinafter, these compounds will be referred to as compounds of the invention.

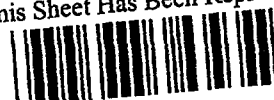
"Natural amino acids" refer to those well known in the art. They are listed and standard abbreviations are provided in the U.S.P.T.O. publication, Trademark Official Gazette, published May 15, 1990, p. 33 at 46. These amino acids and abbreviations are specifically incorporated herein by reference.

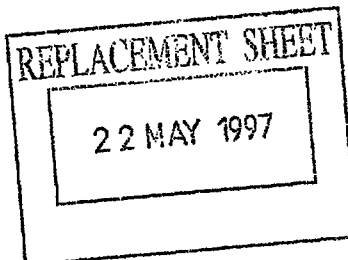
The natural amino acids are shown below:

A	Ala	alanine
D	Asp	aspartic acid
E	Glu	glutamic acid
F	Phe	phenylalanine
G	Gly	glycine
H	His	histidine
I	Ile	isoleucine
K	Lys	lysine
L	Leu	leucine
M	Met	methionine
N	Asn	asparagine
Q	Gln	glutamine
R	Arg	arginine
S	Ser	serine
T	Thr	threonine
V	Val	valine
W	Trp	tryptophane
Y	Tyr	tyrosine



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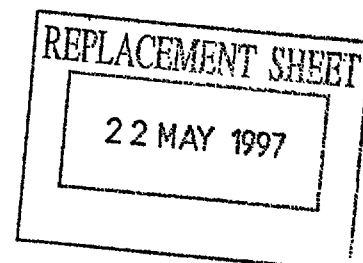


The term "unnatural amino acid" unit means an amino acid unit which is not genetically encoded. Examples of unnatural amino acid units include e.g. the D isomers of the natural α -amino acids as indicated above, Aib (amino-isobutyric acid), bAib (3-aminoisobutyric acid), Nva (norvaline), β -Ala, Aad (2-amino-adipic acid), bAad (3-aminoadipic acid), Abu (2-aminobutyric acid), Gaba (γ -aminobutyric acid), Acp (6-aminocaproic acid), Dbu (2,4-diaminobutyric acid), TMSA (trimethylsilyl-Ala), alle (allo-Isoleucine), Nle (Norleucine), tert.Leu, Cit (Citrulline), Orn, Dpm (2,2'-diaminopimelic acid), Dpr (2,3-diaminopropionic acid), α - or β -Nal, Cha (cyclohexyl-Ala), hydroxy-proline, Sar (Sarcosine) etc., cyclic amino acid units and N^{α} -alkylated amino acid units, e.g. MeGly (N^{α} -Methyl-glycine), EtGly (N^{α} -ethylglycine), EtAsn (N^{α} -ethyl-asparagine).

By amino acid in protected form is meant a natural or unnatural amino acid having e.g. a side chain including an heteroatom such as O, S or N which can be protected with an O-, S- or N-protecting group. The N-terminus of the PTH compounds of the invention may also be in protected form.

N-protecting groups as may be present on the N-terminus or side chain amino groups of amino acid units include such groups as e.g. disclosed in "Protective Groups in Organic Synthesis", T. W. Greene, J. Wiley & Sons NY (1981), 219-287, for example acyl such as formyl, acetyl, trifluoroacetyl, methoxysuccinyl, hydroxysuccinyl or benzoyl optionally substituted on the phenyl ring with e.g. p-methoxycarbonyl, p-methoxy, p-nitro or p-phenylsulfonamidocarbonyl; alkoxycarbonyl such as t-butyloxycarbonyl, isobutyloxycarbonyl or methoxycarbonyl; allyloxycarbonyl; trityl; 2,2,5,7,8-pentamethyl-chroman-6-sulfonyl; arylmethoxycarbonyl such as 9-fluorenylmethoxycarbonyl or benzyloxy carbonyl optionally substituted on the phenyl ring with p-methoxy, p-nitro, o- or p-chloro, m-phenyl or



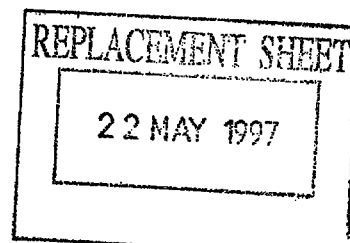


3,4-dimethyl; arylmethyl such as benzyl optionally ring substituted with p-methoxy, p-nitro or p-chloro; or arylsulfonyl such as phenylsulfonyl optionally ring substituted with p-methyl or p-methoxy, or naphthylsulfonyl optionally ring substituted with e.g. amino or di(C₁₋₄alkyl)amino.

O-protecting groups of O-containing side chains are e.g. as described in "The Peptides", 3, (1981), E. Gross and J. Meienhofer (Ed.). For aliphatic hydroxy functionalities, suitable O-protecting groups are e.g. as disclosed in the above reference in "Protection of the Hydroxy Group", J.M. Stewart, 169-201 and include the benzyl, t.-butyl and methyl groups. For aromatic hydroxy functionalities, suitable O-protecting groups include the benzyl, t.-butyl, methyl, tosyl and benzyloxycarbonyl groups. O-protecting groups for carboxy functionalities on amino acid side chains are well known ester groups and described e.g. in "The Peptides", 3, 101-135 ("Carboxyl Protecting Groups" by R.W. Roeske) and include the methyl, ethyl, t.-butyl and benzyl groups. S-protecting groups for thiol functionalities on amino acid side chains are known and described e.g. in "The Peptides", 3, 137-167 ("Sulfhydryl Group Protection" by R.G. Hiskey). Examples include the methyl, t.-butyl, benzyl, p-methoxyphenyl-methyl, ethylamino-carbonyl and benzyloxycarbonyl groups.

According to the invention the PTH compounds may bear on their terminal amino group at least one radical selected from a L- or D- α -amino acid, C₂₋₆alkoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl, and/or on one or more side chain amino group(s) at least one radical selected from C₂₋₆alkoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl. When such a group is attached to a side chain amino group, it is preferably on the ϵ -amino group of a Lys unit. Preferred substituents for





the alkyl, alkenyl, aralkyl, aralkenyl or cycloalkylalkyl group are hydroxy, amino and for the aryl moiety also halogen and/or C₁₋₄alkoxy. The alkyl group or alkyl moiety may be linear or branched and optionally interrupted by O, S or N. Preferably any C₁₋₈alkyl, C₂₋₈alkenyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl on the amino group of a PTH compound, is non-substituted. Examples for C₁₋₈alkyl are C₁₋₆alkyl, preferably methyl, ethyl, propyl, isopropyl, butyl, isobutyl, t.-butyl; for C₂₋₈alkenyl, C₂₋₄alkenyl, preferably allyl; for C₂₋₈alkynyl, C₂₋₄alkynyl, preferably prop-2-ynyl; for aralkyl, phenyl or benzyl; for aralkenyl, styryl; for C₃₋₆cycloalkyl-C₁₋₄alkyl, cyclohexyl-methyl. C₂₋₆alkoxycarbonyl is preferably formyl or acetyl. Suitable examples of D- or L- α -amino acids attached to the N-terminus include e.g. D- or L- Pro, Ala. Preferred substituents when present are alkyl, alkynyl, alkoxy-carbonyl or a D- or L- α -amino acid attached to the N-terminus of the PTH compound and/or at least one alkyl or alkoxy-carbonyl attached to one or more side chain amino groups.

Preferred PTH compounds of the invention are those comprising at least one amino acid unit replaced in one of the following positions of the PTH sequence: 1, 2, 3, 8-11, 13 to 19, 21, 22, 29 to 34, particularly 8-11, 16-19, 33 and/or 34. Further preferred PTH compounds of the invention are those comprising more than one amino acid unit replaced, particularly more than two amino acid units, more particularly more than three amino acid units, especially from 5 to 7 amino acid units replaced, preferably at any combination of the above mentioned positions of the PTH sequence.

In a series of specific or alternative embodiments, the present invention provides a PTH compound as disclosed above wherein in particular the α -amino acids in positions 1 and 2 at the amino terminus of the PTH sequence are replaced by a pseudo-dipeptide.

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The term "pseudo-dipeptide" as used herein refers to a dipeptide isostere in which the peptide bond between the 2 amino acid residues, whether natural or unnatural, is replaced with any isosteric group, e.g. $-\text{CH}_2-\text{NH}-$, $-\text{C}(\text{Halogen})=\text{CH}-$ or $-\text{C}(\text{alkyl})=\text{CH}-$.

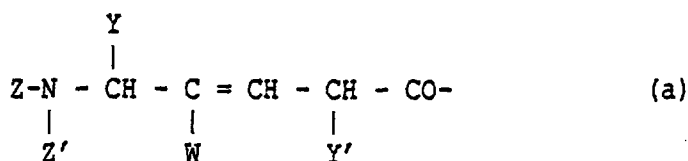
Examples of compounds of the invention wherein the α -amino acids in positions 1 and 2 are replaced by a pseudo-dipeptide include e.g compounds of formula I

X - P₁

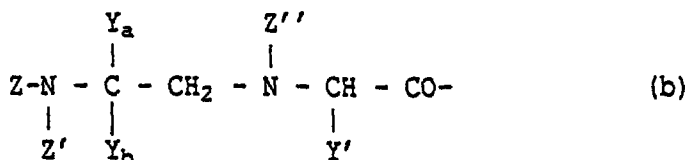
I

wherein

X is a residue of formula (a)



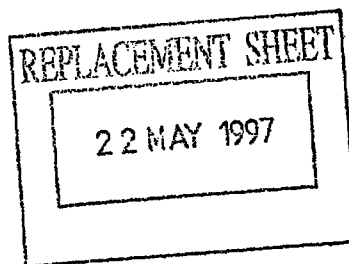
or (b)



wherein each of Z and Z', independently, is H; optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl; or a protecting group, at most one of Z and Z' being a protecting group, Z'' is H, C₁₋₈alkyl or a protecting group each of Y and Y', independently, is an optionally protected side chain of a natural or unnatural α -amino acid, one of Y_a and Y_b is hydrogen and the other is an optionally protected side chain of a natural or unnatural α -amino acid or Y_a and Y_b form together with the carbon atom to which they are attached a C₃₋₆cycloalkyl group,

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and

W is halogen or C₁₋₄alkyl,

or W and Y' together with the moiety —C=CH—CH—

to which they are attached, form an optionally substituted aromatic cyclic or heterocyclic residue, and

P₁ is a PTH sequence, as defined above, in which amino acid units in positions 1 and 2 of the N-terminus are omitted, in free form or in salt or complex form.

In compounds of formula I, P₁ may be a fragment (3-z) of PTH wherein z is 84 or an integer from 27 to 38, preferably from 34 to 38, particularly 34, 36, 37 or 38, especially 34 or 36. The PTH sequence represented by P₁ may correspond either to a naturally occurring PTH sequence, or to a naturally occurring PTH sequence wherein at least one α -amino acid unit has been replaced by a natural or unnatural amino acid optionally in protected form and/or one or more α -amino acid units are also omitted. P₁ may also comprise at least one side chain amino group substituted as disclosed above.

Halogen is preferably fluorine or chlorine.

In the residue of formula (a), the double bond has preferably the configuration trans (E).

As residue attaching to the α -carbon atom of a natural α -amino acid, Y, Y', Y_a or Y_b may be the side chain present in a natural α -amino acid as listed above, e.g. as present in Gly (i.e. H), Ala, Val, Ser, Leu, Ile, Phe, Trp. Y, Y', Y_a or Y_b may also be the residue attaching to the α -carbon atom of an unnatural α -amino acid as indicated above, e.g. as present in Nva, Orn, Abu, Aib, Nle. If the side chain of the natural or unnatural amino acids as Y, Y', Y_a or Y_b includes heteroatoms such as O, S or N, the heteroatoms on the side chain may optionally be pro-

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tected with an O-, S- or N-protecting group, e.g. as indicated above.

Protecting groups as Z or Z' or Z'' may be as disclosed above.

C₁₋₈alkyl as Z or Z' or Z'' may be as indicated above.

Preferably Z or Z' is hydrogen or C₁₋₄alkyl, particularly hydrogen or methyl.

Preferably Y is H or CH₃.

When W is C₁₋₄alkyl, it is preferably CH₃.

When W and Y' together with the moiety to which they are attached form an optionally substituted aromatic cyclic or heterocyclic residue, it may be an aromatic 5- or 6-membered cyclic residue comprising optionally 1 or 2 heteroatoms selected from N, S and O, e.g. phenyl, imidazolyl, pyridyl, oxazolyl or thiazolyl. A preferred substituent is hydroxy or methoxy, particularly for the phenyl ring.

Y' is preferably H, CH₃, isopropyl or benzyl.

When Z'' is C₁₋₈alkyl, it is preferably CH₃, C₂H₅ or isopropyl.

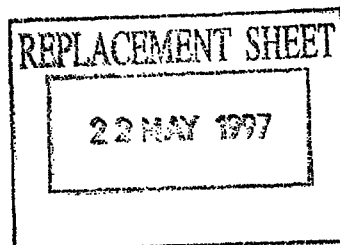
C₃₋₆cycloalkyl as Y_a and Y_b together is preferably cyclopropyl or cyclopentyl.

When Z'' is a protecting group, it is preferably acyl, particularly acetyl.

In a series of specific and alternative embodiments, the present invention provides a PTH compound as disclosed above wherein the α -amino acid unit in position 1 and/or the α -amino acid unit in

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position 2 of the N-terminus of the PTH sequence is replaced by an optionally protected natural or unnatural amino acid residue, provided that

when the PTH compound is free from D- or L- α -amino acid attached to the N-terminus or from C₂₋₆alkoxycarbonyl or optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl, it is other than a PTH compound having a naturally occurring α -amino acid sequence;

or the PTH compound is other than PTH(1-34) wherein

- i. the α -amino acid in position 1 is Aib, Gly, D-Ser, D-Ala or Tyr; or
- ii. the α -amino acid in position 2 is Ala, D-Val, Lys, Cit or Arg and the α -amino acid in position 34 is Tyr; or the α -amino acid in position 2 is D-Val and the α -amino acid in position 34 is D-Tyr and optionally the α -amino acids in positions 8 and 18 are each Nle; or
- iii. the α -amino acid in position 1 is Aib and at least one further α -amino acid unit has been replaced as follows: the α -amino acid in position 8 and/or 18 is Leu, Ile, Val, Phe or Trp, and/or the α -amino acid in position 11 is Ser, Lys, Phe, β -Nal, Trp or Tyr, and/or the α -amino acid in position 12 is D-Leu, D-Ile, D-Nle, D-Val, D-Ser, D-Ser(Butyl), D-Abu, D-Thr, D-Nva, D-Met, D- β -Nal, D-Trp, D-Lys, D-Tyr, D-Lys(Fmoc), D-Phe or D-Asn, and/or the α -amino acid in position 13 is Leu, and/or the α -amino acid in position 19 and/or in position 21 is Arg, Lys, Asn or His, and/or the α -amino acid in position 23 is 2-(1,3-dithiolane-2-yl)Trp, and/or the α -amino acid in position 25 and/or in position 26 is His, and/or the α -amino acid in position 27 is Gln or Leu,

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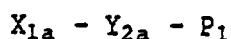
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or the PTH compound is other than PTH(1-84) wherein

- i. the α -amino acid in position 1 is Tyr, Val, Pro, Asp or Cys; or
- ii. the α -amino acid in position 2 is Ala, Glu, Leu, Ser or Arg.

Examples of compounds of the invention wherein the α -amino acids in position 1 and/or in position 2 are replaced as indicated above include e.g compounds of formula II



II

wherein

X_{1a} is a residue of an optionally protected natural α -amino acid or a residue of formula (IIa)

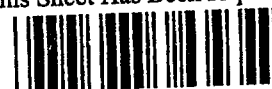


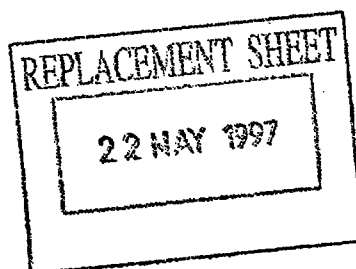
wherein

each of Z_a and Z'_a , independently, is H, C_{1-6} alkyl or a protecting group, at most one of Z_a and Z'_a being a protecting group, and either n is 1 and

- i. each of R_1 and R_2 , independently, is C_{1-6} alkyl, or
- ii. one of R_1 and R_2 is methyl or ethyl and the other is an optionally protected residue attaching to the α -carbon atom of a natural α -amino acid other than Ala, Leu, Ile or Val, or
- iii. one of R_1 and R_2 is H, methyl or ethyl and the other is an optionally protected residue attaching to the α -carbon atom of an unnatural α -amino acid, or
- iv. R_1 and R_2 form together with the carbon atom to which they are attached a C_{3-6} cycloalkyl group, or

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v. the residue $Z_a - \text{N} - \overset{\overset{R_1}{|}}{\underset{\underset{Z'_a}{|}}{\text{C}}} - R_2$ represents a heterocyclic

residue optionally condensed to a benzene ring,

or n is 2, 3, 4 or 5 and each of R_1 and R_2 is H or CH_3 ,

Y_{2a} is a residue of an optionally protected natural α -amino acid or a residue of formula (IIb)



Z_b is H or C_{1-6} alkyl and either m is 1 and

- i. each of R_3 and R_4 , independently, is C_{1-6} alkyl, or
- ii. one of R_3 and R_4 is methyl or ethyl and the other is an optionally protected residue attaching to the α -carbon atom of a natural α -amino acid other than Ala, Leu, Ile or Val, or

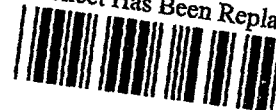
- iii. one of R_3 and R_4 is H, methyl or ethyl and the other is an optionally protected residue attaching to the α -carbon atom of an unnatural α -amino acid,

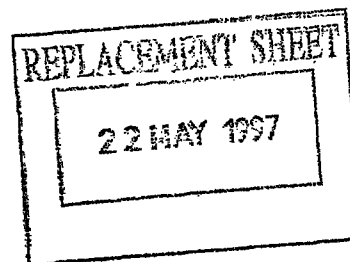
or m is 2, 3, 4 or 5 and each of R_3 and R_4 is H or CH_3 , and

P_1 is as defined above.

When the residue $Z_a - \text{N} - \overset{\overset{R_1}{|}}{\underset{\underset{Z'_a}{|}}{\text{C}}} - R_2$ is a heterocyclic residue

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optionally condensed to a benzene ring, it may be a residue derived from a saturated or unsaturated 5- or 6-membered heterocycle comprising one or more heteroatoms selected from N, O and S. Examples of such heterocyclic residues include e.g. 2-pyridyl, 3-morpholinyl, hexahydropyridazinyl, 2- or 3-indolyl.

When n or m is > 1 , the moiety CR_1R_2 or CR_3R_4 may be $-CH(CH_3)-CH_2-$, propyl, butyl or pentyl.

C_{3-6} cycloalkyl as R_1 and R_2 together is preferably cyclopropyl or cyclopentyl.

As residue attaching to the α -carbon atom of a natural α -amino acid, R_1 , R_2 , R_3 or R_4 may be the side chain present in a natural α -amino acid as listed above, e.g. as present in Gly, Ser or Thr. One of R_1 and R_2 or one of R_3 and R_4 may be methyl or ethyl and the other may be CH_3 , isopropyl, isobutyl or $CH_3-CH_2-CH(CH_3)-$. R_1 , R_2 , R_3 or R_4 may also be the residue attaching to the α -carbon atom of an unnatural α -amino acid as indicated above, e.g. as present in Abu, Gaba, Nva, Aib, TMSA or Nle. If the side chain of the natural or unnatural amino acids as R_1 , R_2 , R_3 or R_4 includes heteroatoms such as O, S or N, the heteroatoms on the side chain may optionally be protected with an O-, S- or N-protecting group, e.g. as indicated above. Protecting group as Z_a or Z'_a may be as disclosed above. C_{1-6} alkyl as Z_b is preferably methyl or ethyl. Preferably Z_b is H. C_{1-6} alkyl as Z_a or Z'_a is preferably straight chain or branched C_{1-4} alkyl. Preferably one of Z_a and Z'_a is H and the other is H, C_{1-4} alkyl or a protecting group, particularly H.

In a further or alternative embodiment, the present invention also provides a PTH compound as disclosed above wherein the α -amino acid unit in position 8 and/or the α -amino acid unit in



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position 18 is replaced by an optionally protected natural or unnatural amino acid residue, provided that

when the PTH compound is free from D- or L- α -amino acid attached to the N-terminus or from C₂₋₆alkoxycarbonyl or optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl, it is other than a PTH compound having a naturally occurring α -amino acid sequence;

or the PTH compound is other than PTH(1-34) wherein

- i. the α -amino acids in positions 8 and 18 are each Nle or each Met(O) and optionally the α -amino acid in position 34 is Tyr; or the α -amino acids in positions 8 and 18 are each Nle and the α -amino acid in position 34 is Tyr and either the α -amino acid in position 12 is L- or D-Pro, L- or D-Ala, Aib or NMeGly or the α -amino acid in position 23 is Phe, Leu, Nle, Val, Tyr, α -Nal or β -Nal; or the α -amino acids in positions 8 and 18 are each Nle and the α -amino acid in position 2 is D-Val and the α -amino acid in position 34 is D-Tyr; or
- ii. the α -amino acid in position 8 and/or 18 is Leu, Ile, Val, Phe or Trp and optionally at least one further α -amino acid unit has been replaced as follows: the α -amino acid in position 1 is Aib and/or the α -amino acid in position 11 is Ser, Lys, Phe, β -Nal, Trp or Tyr, and/or the α -amino acid in position 12 is D-Leu, D-Ile, D-Nle, D-Val, D-Ser, D-Ser(Butyl), D-Abu, D-Thr, D-Nva, D-Met, D- β -Nal, D-Trp, D-Lys, D-Tyr, D-Lys(Fmoc), D-Phe or D-Asn, and/or the α -amino acid in position 13 is Leu, and/or the α -amino acid in position 19 and/or in position 21 is Arg, Lys, Asn or His, and/or the α -amino acid in position 23 is 2-(1,3-dithiolane-2-yl)Trp, and/or the α -amino acid in

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position 25 and/or in position 26 is His, and/or the α -amino acid in position 27 is Gln or Leu; or

- iii. the α -amino acid in position 8 and/or 18 is Ala or Ser; or the α -amino acid in position 8 and/or 18 is Leu, and the α -amino acid in position 34 is Tyr; or

the PTH compound is other than PTH(1-84) wherein

the α -amino acid in position 8 is Met, Met(O), Ala, Val, Leu, Ile, Ser, Trp, Asn, Gln, Asp, Glu, Lys, Arg, Tyr or Gly and the α -amino acid in position 18 is Leu; or the α -amino acid in position 8 and in position 18 are each Met(O); or the α -amino acid in position 8 is Leu and the α -amino acid in position 18 is Met(O); or

the PTH compound is other than [Leu¹⁸, Tyr³⁴]hPTH(1-84), [Leu⁸, Tyr³⁴]hPTH(1-84), [Ile⁸, Leu¹⁸, Tyr³⁴]hPTH(1-84) or [Leu⁸, Leu¹⁸, Tyr³⁴]hPTH(1-84).

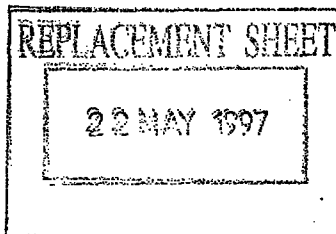
When the α -amino acid unit in position 8 is replaced by an unnatural amino acid, it may be an unnatural amino acid as disclosed above e.g. Nva, Nle or Cha. Preferably it is an unnatural lipophilic amino acid preferably such containing a straight chain alkyl residue.

Nva (norvaline) and its homologues are particularly preferred. The unnatural amino acid in position 8 of the PTH sequence may also be a α -methylated or α -ethylated natural α -amino acid.

Examples of PTH compounds of the invention wherein the α -amino acid unit in position 18 is replaced, are e.g. PTH compounds wherein the α -amino acid residue in position 18 is replaced by a natural amino acid residue such as Gln, Tyr or Lys, for example [Gln¹⁸]-PTH, [Lys¹⁸]-PTH and [Tyr¹⁸]-PTH, particularly [Gln¹⁸ or Lys¹⁸ or Tyr¹⁸]-hPTH-(1-x) wherein x is 84 or an integer from 27 to 38, particularly from 34 to 38. Ala in position 18 is also

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preferred particularly when the PTH sequence is a (1-36) PTH fragment. In these compounds, the amino acid in position 8 may be Met or it may be replaced by an amino acid selected from Leu, Ile, Val, Phe, Gln, Trp, Ser and Ala or it may be an unnatural lipophilic amino acid residue, e.g. Nva or Nle.

Another group of such PTH compounds of the invention comprises e.g. PTH compounds wherein the amino acid residue in position 8 is replaced by an unnatural lipophilic amino acid residue, e.g. Nle or more preferably Nva. In these compounds the amino acid in position 18 may be Met or it may be replaced by an amino acid selected from Leu, Ile, Val, Phe, Trp, Ser, Ala, Gln, Lys or Tyr, preferably Leu, Gln, Ala or Tyr.

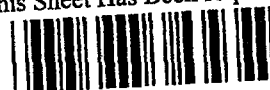
A further group of such PTH compounds of the invention are those wherein one or more amino acid units in the remaining positions, e.g. 1 to 7, 9 to 17 and/or 19 to 38 are either omitted or replaced in addition to the replacement of the α -amino acid unit in position 8 and/or 18. In such a case, the α -amino acid in position 8 is preferably replaced by Leu and the α -amino acid in position 18 by Gln, Leu, Ala or Tyr.

In a yet further or alternative embodiment, the present invention also provides a PTH compound as disclosed above wherein at least one of the α -amino acid unit of the PTH sequence is replaced by the α -amino acid unit which is present at the corresponding position in PTHrP,

provided that

when the PTH compound is free from D- or L- α -amino acid attached to the N-terminus or from C₂₋₆alkoxycarbonyl or optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl, it is other than a PTH compound having a naturally occurring α -amino acid sequence;

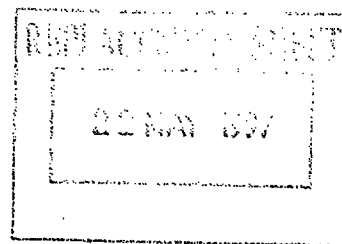
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or the PTH compound is other than PTH(1-34) wherein

- i. the α -amino acid in position 8 and/or 18 is Leu; and/or the α -amino acid in position 11 is Lys; and/or the α -amino acid in position 19 and/or 21 is Arg; and/or the α -amino acid in position 25 and/or 26 is His; and/or the α -amino acid in position 27 is Leu; or
- ii. the α -amino acid in position 25 is Gln and/or the α -amino acid in position 26 is Asn, the α -amino acid in position 27 being optionally Leu; or

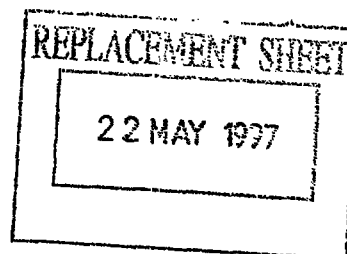
the PTH compound is other than [Leu⁸,Leu¹⁸]PTH(1-84).

The term "PTHrP" refers to any genetically encoded form of PTHrP, e.g. human, chicken, rat or mouse PTHrP. For consistency and as is conventional, in the following description, the same numbering system will be applied to the amino acids of the PTHrP sequence starting with Ala in position 1 and comprising e.g. Ala in position 38.

This group of compounds according to the invention are hybrids between PTH and the homologous PTHrP sequences (referred to herein as PTH-rP hybrids). The amino acid sequence wherein the substitution according to the invention takes place is from position 1 up to position 38, particularly 1 up to 36, especially 1 up to 34.

Preferred PTH compounds of the invention are PTH-rP hybrids wherein more than one α -amino acid unit of the PTH sequence, e.g. at least 2, preferably from 3 to 5 α -amino acid units, are replaced according to the invention.

A preferred group of the PTH-rP hybrids according to the invention comprises PTH compounds wherein at least one α -amino acid unit of the PTH α -amino acid sequence in positions 8 to 11, i.e. Met⁸, His⁹, Asn¹⁰ or Leu¹¹ is replaced by the corresponding



α -amino acid unit of the corresponding sequence of PTHrP, i.e. Leu⁸, His⁹, Asp¹⁰ or Lys¹¹. The replacement at positions 8 and/or 10 is preferred, particularly at positions 8 and 10. More preferably the α -amino acids at positions 8, 10 and 11 of said PTH sequence are replaced by Leu⁸, Asp¹⁰ and Lys¹¹, respectively.

Another preferred group of the PTH-rP hybrids according to the invention comprises PTH compounds wherein at least one α -amino acid unit of the PTH α -amino acid sequence in positions 16 to 19 is replaced by the corresponding α -amino acid unit of the corresponding sequence of PTHrP, i.e. Gln¹⁶, Asp¹⁷, Leu¹⁸, Arg¹⁹. Preferably 3 or all 4 α -amino acids of said sequence are replaced. Particularly preferred are PTH compounds of the invention wherein the α -amino acid in position 16 is Gln, the α -amino acid in position 18 is Leu and the α -amino acid in position 19 is Arg.

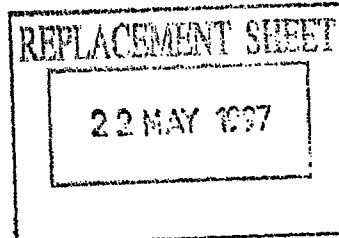
A further preferred group of the PTH-rP hybrids according to the invention comprises PTH compounds wherein at least one α -amino acid unit of the PTH α -amino acid sequence in positions 33 and 34 is replaced by the corresponding α -amino acid unit of the corresponding sequence of PTHrP, i.e. Thr³³, Ala³⁴. Preferably both α -amino acids are replaced.

Yet further preferred groups of the PTH-rP hybrids according to the invention are PTH compounds comprising any combination of the above mentioned α -amino acid substitutions (8-11, 16-19, 33-34), more preferably a combination of substitutions selected from the above indicated 8-11 and 33-34 sequences.

As it will be appreciated, in the PTH-rP hybrids according to the invention one or more α -amino acids in positions 1 up to 38 may be further replaced by a natural or unnatural amino acid unit as indicated above or be omitted. In position 10 there may be a

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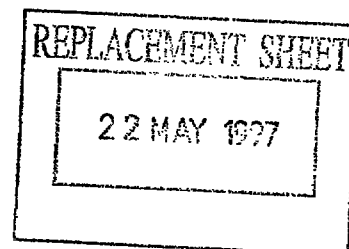
α -amino acid selected from Gly, Gln, Glu, His, Ser, Thr or Tyr. In position 13 there may be a D-or L- α -amino acid other than Arg, e.g. Ala, Cys, Gln, Ile, Asn, Trp, Asp, Val, Ser, Thr, Tyr, Met, Leu or Gly; in position 16 there may be a D- or L- α -amino acid, e.g. Lys, Ser, Leu, Ala, Gln or Gly; in position 17 Ala or Ser or preferably an amino acid having a bigger side chain than Ala or Ser, e.g. Glu, Gln, Phe, His, Ile or Lys; in position 18 Gln or Tyr; in position 19 Ala, Arg, Val, Tyr, Ser, Lys, Met, His, Gly, Pro, Asn or Ile; Gln or Arg in position 26; and/or in position 33 a D- or L- α -amino acid e.g. Ser, Thr, Leu, Gly, Gln, Arg, Pro, Asp, Ile, Lys, or Thr. They also may be substituted on the N-terminus or on a side chain amino group as indicated above. If desired, S- or O-containing side chains of the PTH-rP hybrids of the invention may also be protected as disclosed above.

Examples of preferred PTH-rP hybrids according to the invention are [Leu⁸, Gln¹⁸, Thr³³, Ala³⁴]PTH, [Leu⁸, Ala¹⁶, Gln¹⁸, Thr³³, Ala³⁴]PTH, [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸, Thr³³, Ala³⁴]PTH, [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸]PTH, [Leu⁸, Ala¹⁶, Gln¹⁸, Ala¹⁹]PTH, [Leu⁸, Asp¹⁰, Lys¹¹, Gln¹⁸]PTH, [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸, Ala¹⁹]PTH, [Leu⁸, Ala¹⁶, Gln¹⁸, Ala¹⁹, Thr³³, Ala³⁴]PTH and [Leu⁸, Asp¹⁰, Lys¹¹, Gln¹⁸, Thr³³, Ala³⁴]PTH.

Preferred PTH-rP hybrids according to the invention are PTH(1-x') compounds wherein x' is an integer from 34 to 38, particularly 34, 36, 37 or 38 especially hPTH(1-x'), wherein one or more α -amino acids are replaced by the α -amino acid(s) present at the corresponding position in PTHrP preferably as indicated above. When the positions 33 and 34 are replaced by the corresponding α -amino acid sequence of PTHrP, the PTH compound is preferably a hPTH fragment comprising 34 α -amino acid units. PTH-rP hybrids with carboxy terminus are preferred.

Preferably PTHrP is hPTHrP.

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The compounds of the invention may exist e.g. in free form, salt form or in the form of complexes thereof. Acid addition salts may be formed with e.g. organic acids, polymeric acids and inorganic acids. Such acid addition salt forms include e.g. the hydrochlorides and the acetates. Complexes are e.g. formed from the compound of the invention on addition of inorganic substances, e.g. inorganic salts or hydroxides such as Ca- and Zn-salts, and/or an addition of polymeric organic substances.

The present invention also provides a process for the production of the compounds of the invention. They may be prepared in a stepwise manner either in solution or using the solid phase synthesis process or genetic engineering.

The compounds of the invention may be produced for example as follows:

- a) removing at least one protecting group which is present in a compound of the invention in protected form; or
- b) linking together by an amide bond two peptide fragments, the peptide fragments being such that the desired amino acid sequence of the desired compound is obtained, and then effecting optionally stage a) of the process,
- c) for the production of a PTH compound wherein the α -amino acid units in positions 1 and 2 at the amino terminus are replaced by a pseudo-dipeptide, reacting a pseudo-dipeptide in protected or unprotected form with a PTH peptide in protected or unprotected form in which the amino acid residues in positions 1 and 2 are omitted and if necessary carrying out process step a); or

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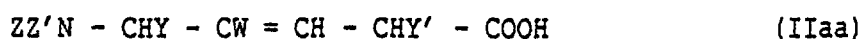
d) for the production of a PTH compound comprising at least one radical selected from D- or L- α -amino acid attached to the N-terminus and C₂₋₆alkoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl attached to the terminal amino group and/or to a side chain amino group, introducing such a radical in a PTH compound in protected or unprotected form and free from such a radical, and if necessary carrying out process step a);

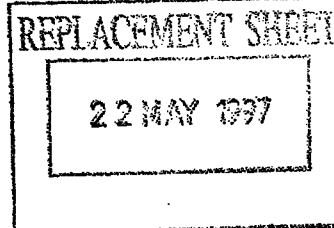
and recovering the compounds thus obtained in free form, in salt form or in complex form.

Process steps a), b) and c) may be effected in analogy with known methods, e.g. as known in the art of peptide chemistry or as described in the following examples. Where desired, in these reactions, protecting groups which are suitable for use in peptides may be used for functional groups which do not participate in the reaction. The term protecting group may also include a polymer resin having functional groups.

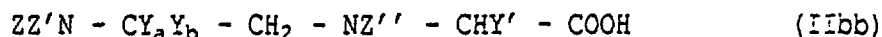
In process step b), for the production of a PTH compound wherein the α -amino acid units in positions 1 and 2 at the amino terminus are replaced by a pseudo-dipeptide, one peptide fragment used as starting material may comprise the pseudo-dipeptide at its N-terminus. This starting material may be prepared in accordance with process step c).

In process step c) the pseudo-dipeptide used as starting material may be a compound of formula IIaa or IIbb





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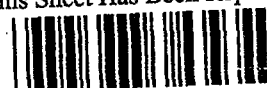


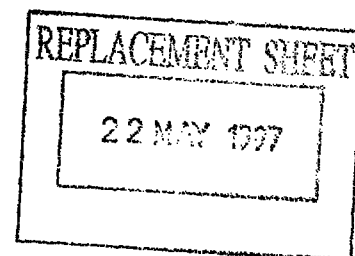
wherein Y, Y', Y_a, Y_b, Z, Z', Z'' and W are as defined above, or a functional derivative thereof, e.g. ester, acid halide or a symmetric or asymmetric anhydride. Compounds of formula IIaa wherein W and Y' together with the moiety $-C=CH-CH-$ to which they are attached, form an optionally substituted aromatic cyclic or heterocyclic residue, and compounds of formula IIbb wherein Z'' is alkyl or a protecting group, e.g. acetyl, or wherein $-CY_aY_b$ is C₃₋₆cycloalkyl and Z'' is H are novel and form part of the invention.

Process step d) may be carried out in analogy with known methods, e.g. alkylating methods. In a preferred embodiment of the invention, the alkylation is performed under reductive conditions, e.g. using a corresponding aldehyde or ketone. It may be performed for example in the presence of NaBH₃CN, preferably at an acidic pH, e.g. from 5 to 7. The temperature of the reaction may be e.g. from -20° C to 100° C. It may be advantageous to carry out the reaction in an inert solvent, e.g. water, an alcohol, dioxane or DMF or a mixture thereof. A D- or L-α-amino acid may also be attached to the N-terminus in accordance with known procedure.

The peptide fragment used as starting material in process step b) may comprise one or more unnatural amino acid units; it may also be substituted on the N-terminus and/or on a side chain amino group by a radical selected from C₂₋₆alkoxycarbonyl, optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl or it may bear a D- or L-α-amino acid on the N-terminus.

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The compounds of the invention or fragments thereof comprising natural α -amino acid units may also be prepared using recombinant technology.

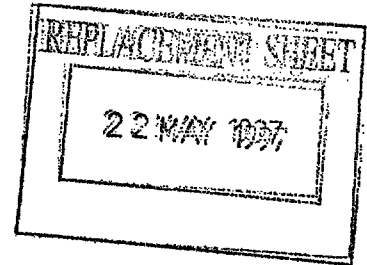
According to a preferred embodiment of the invention there is also provided a process for the production of a medium-sized polypeptide, in which a fusion protein comprising an N-terminal bacteriophage T4 gene 55 polypeptide having the desired polypeptide linked to the C-terminal thereof, is expressed in bacterial cells.

The gene 55 polypeptide may comprise the complete gp55 protein of bacteriophage T4 (188 amino acid residues) which is described in Gram, H. and Rüger, R; 1985; The EMBO Journal, 4, (1), pp 257-264. Alternatively a fragment, preferably an N-terminal fragment of the gp55 protein may be used. For example, a gp55 protein fragment comprising the first 112 N-terminal amino acid residues of the protein may be used. Typically, however, the gene 55 polypeptide comprises at least the first 25 N-terminal residues of the gp55 protein.

The medium-sized polypeptide may be from about 20 up to 100, e.g. 150, preferably from about 20 to about 50, amino acids in length. Examples of medium-sized polypeptides include calcitonins, endorphins, gastric inhibitory peptide(s), glucagon, neuropeptide Y, growth hormone releasing factor (GHRF), amyloid β protein fragments, hirudin, insulin, somatostatin, epidermal growth factor, nerve growth factor and PTH or fragments thereof. Particularly preferred medium-sized polypeptide which may be prepared according to the process of the present invention are the compounds of the invention or fragments thereof having natural amino acid substitutions.

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Conveniently the gene 55-PTH fusion protein also comprises a cleavable linker between the gene 55 polypeptide and the desired polypeptide. Preferably the cleavable linker is chemically cleavable and more preferably contains the amino acid sequences Asp-Pro-Pro or Asn-Gly-Pro.

In a further aspect the invention provides a nucleotide sequence coding for a fusion protein comprising an N-terminal bacteriophage T4 gene 55 polypeptide and a desired medium-sized polypeptide linked to the C-terminal thereof.

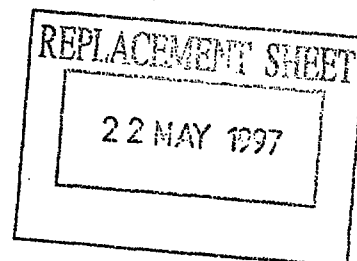
The nucleotide sequence is preferably a DNA sequence suitable for expression in bacteria. The sequence typically contains nucleotides coding for a cleavable linker between the gene 55 and desired polypeptide coding sequences.

The invention also provides a bacterial expression vector comprising a DNA sequence coding for the fusion protein.

The expression vector typically contains, in addition to the fusion protein coding sequence, appropriate expression control sequences including a suitable promoter, an operator and a ribosome binding site and other appropriate regulatory sequences. Usually, also the expression vector contains one or more selectable markers.

Any suitable promoter may be used, such as the TrpE, λ P1, λ Pr, lac or Tac promoter. Preferably the promoter is the bacteriophage T7 promoter and the expression vector is a plasmid. For E. coli expression an R1 or Col-E1 plasmid-derived vector may be used. For example, expression vector pET 17B, obtainable from Novagen, or similar expression vectors may be used.





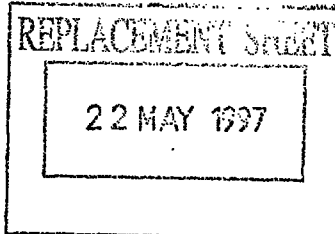
In another aspect the invention provides bacterial host cells transformed with a fusion protein coding sequence or an expression vector as described above. Any suitable bacterial host may be used, though preferably the host is E. coli. For example, E. coli strain BL21 (DE3) Lys E and similar strains may be used.

Advantageously the fusion protein accumulates within the host cells in the form of insoluble inclusion bodies, and thereby facilitates recovery of the fusion protein product from the cells. In order to recover the desired polypeptide product from the fusion protein, the protein of the inclusion bodies is solubilised, and the desired polypeptide product cleaved from the fusion protein. Any suitable solubilisation treatment may be used, including acid treatment, e.g. at about pH 2 to 4, preferably with acid at about pH 2.5, or treatment with a denaturing agent, e.g. a mild denaturing agent such as guanidine hydrochloride. In addition it may be necessary to remove one or more unwanted N-terminal amino acid residues from the product which remain after cleavage.

Thus in a further aspect the invention provides a process for the production of a desired medium-sized polypeptide, in a bacterial host, the process comprising:
causing transformed bacterial host cells as defined above to express the fusion protein in the form of inclusion bodies;
isolating the inclusion bodies;
solubilizing the inclusion bodies, and
cleaving the desired polypeptide from the fusion protein.

In the case of some medium-sized polypeptides, such as the PTH compounds, it is not normally necessary to carry out carefully-controlled denaturation and renaturation procedures to obtain PTH products in active form, e.g. having PTH-like activity. The medium-sized polypeptide products may be obtained in active form





merely by neutralising the acid conditions used for solubilisation. Solubilisation and cleavage of the fusion protein may be carried out in a single step.

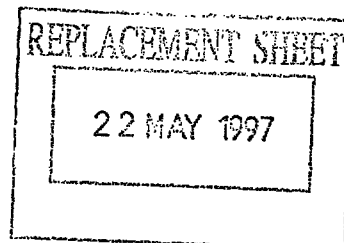
The invention also includes a fusion protein comprising an N-terminal bacteriophage T4 gene 55 polypeptide and, linked to the C-terminal thereof, a desired medium-sized polypeptide.

Using the processes and vectors of the invention it has been found that it is possible to produce high levels of medium-sized polypeptides, particularly PTH products. In E. coli, we have found that up to about 50% of the total protein expressed is the desired fusion protein. Also, the fusion protein forms inclusion bodies which are easy to separate from the other material of the cells. Moreover the inclusion bodies often consist of at least about 90% of the desired fusion protein which can significantly reduce the amount of purification that is required. Further, expressing the PTH product, in the form of the fusion protein can substantially decrease endogenous processing of the polypeptide in the bacterial cell, allowing homogenous products to be obtained.

Preferably the cleavable linker contains chemically cleavable amino acids adjacent the N-terminus of the desired polypeptide, to permit the desired polypeptide to be cleaved from the fusion protein by simple chemical means. Preferably the desired polypeptide does not contain the chemically cleavable groups.

Preferred chemically cleavable linkers contain the amino acids Asp-Pro-Pro or Asn-Gly-Pro. The Asp-Pro or Asn-Gly bonds may be chemically cleaved using acid conditions e.g. formic acid (normally about pH 2.5) or hydroxylamine respectively. The di-peptides Pro-Pro or Gly-Pro that remain on the N-terminus of the desired polypeptide may then be removed using a suitable





dipeptidyl-peptidase. For example the L. lactis X-Pro dipeptidyl-peptidase EC 3.4.14.5 can be used. This peptidase is described in Nardi et al; 1991; Applied and Environmental Microbiology, 57, p45 to 50.

The desired polypeptide may be purified following cleavage from the fusion protein. HPLC purification techniques may be used. Purification may be carried out before removal of any unwanted N-terminal amino acid residues.

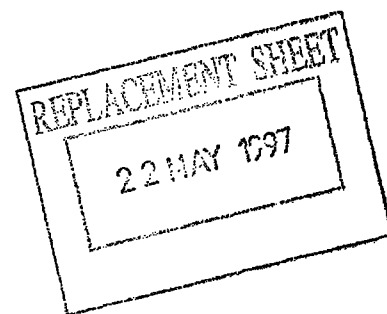
The process of the invention is further illustrated in the examples 308 to 314 Fig. 1 shows the amino acid sequence and corresponding DNA sequence of a truncated gp55-Asp-Pro-Pro-hPTH(1-38)OH fusion protein as cloned into the plasmid pET17B, and Fig. 2 shows the amino acid sequence and corresponding DNA sequence of a full length gp55-Asn-Gly-Pro-hPTH(1-38)OH fusion protein as cloned into the plasmid pET17B.

In the following Examples all temperatures are in °C. The following abbreviations are employed. In the present specification the term PTH without any further indication includes the carboxy terminus as well as a carboxamide terminus.

BOC	=	tert. Butyloxycarbonyl
DMF	=	dimethylformamide
DCM	=	dichloromethane
Fmoc	=	9-fluorenylmethoxycarbonyl
HOBt	=	1-hydroxybenzotriazole
TFA	=	trifluoroacetic acid
Trt	=	trityl = triphenylmethyl
RP-HPLC	=	reverse phase high performance liquid chromatography
r.t.	=	room temperature
DNP	=	Dinitrophenyl

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Tos = Tosyl
DIPC = Diisopropylcarbodiimide
Pmc = 2,2,5,7,8-Pentamethylchroman-6-sulfonyl
Ip = Isopropyl
For = Formyl
Cpe = 1-amino-cyclopentane-1-carboxylic acid
Cpp = 1-amino-cyclopropane-1-carboxylic acid
Cha = cyclohexylalanine
tBu = t.butyl

EXAMPLE 1: ~~NE~~-Isopropyl hPTH(1-34) NH₂

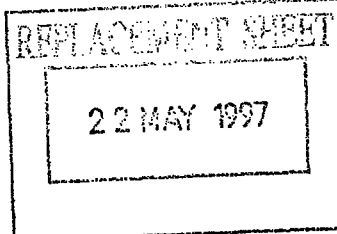
This peptide is assembled in a stepwise manner on a poly-styrene-based resin support. The Boc-group is used for protection of the alpha amino-group and side-chain functional groups are protected as follows: Lys(2-chlorobenzyloxycarbonyl), Ser(benzyl), Thr(benzyl), Glu(benzyl), Asp(benzyl), Met(O), His(DNP), Arg(Tos), and Trp(HCO). Other residues are left unprotected.

Amino-4-methylphenyl-methyl-co(polystyrene-divinylbenzene = MBHA-resin) (0,7 mmol/g) is subjected to the following cycle, steps (1) to (7), of treatments:

- (1) DCM
- (2) trifluoroacetic acid (50 %) in DCM
- (3) DCM
- (4) diisopropylethylamine (10 %) in DMF
- (5) DMF
- (6) preformed symmetrical anhydride (1.4 mmol per g starting resin) of Boc-amino acid in DMF
- (7) DMF

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Volumes of washes and reagents are from 5 to 20 ml per gram of starting resin. Each step is repeated as many times as necessary for either complete reaction of the resin (steps 2, 4, 6) or complete displacement of the previous reagent from the resin (steps 1, 3, 5, 7). Samples of resin are removed after each cycle and checked for completeness of the coupling reaction by a colorimetric test for residual amino groups using ninhydrin.

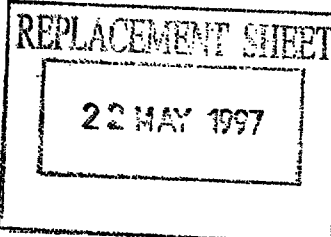
Symmetrical anhydrides of Boc-amino acids are prepared just prior to use by reacting Boc-amino acid (2,8 mmol per g resin) and DCCI (1,4 mmol per g resin) in DCM, containing DMF in amounts sufficient for complete dissolution of the Boc-amino acid. The mixture is filtered, more DMF is added to the filtrate which is concentrated by evaporation of the volatile components at a temperature not exceeding 15°C and the resulting solution is used in step (6).

The cycle of reactions, (1) to (7), is repeated for each amino acid residue such as to provide the sequence of the title compound except for Boc-Gln-OH and Boc-Arg(Tos)-OH, which are coupled in step (6) as their preformed 1-hydroxybenzo-triazole esters in DMF.

The fully assembled peptide resin is treated twice with 20 ml of thiophenol 5% in DMF at r.t. for 1 hour and washed with DMF, methanol and dichloromethane. The peptide resin is again subjected to steps (1) to (5) followed by the addition of 0.5 ml of acetone, 84 mg of sodium cyanoborohydride and 0.2 ml of acetic acid in 20 ml of DMF per gram of resin. After 16 hours at RT the resin is washed with DMF and dichloromethane and dried.

The peptide resin is treated with liquid HF, dimethyl sulfide, p-cresol and ethane-1,2-dithiol according to the 'low-high' procedure which is described, e.g., in International Journal of Peptide and Protein Research, vol.26, page 262-273 (1985). After removal of the volatile components and washing with ethylacetate





the residue is extracted with several portions of 1%-acetic acid, filtered and the filtrate lyophilized. The lyophilized product is purified by repeated reversed-phase chromatography on a column of octadecyl-silica using a gradient of acetonitril in 2%-phosphoric acid or in 20 mM tetramethylammonium phosphate. Fractions containing the pure compound are combined, filtered through a weakly basic ion-exchange resin in the acetate form and the filtrate lyophilized to give the title compound.

MS (Ion-Spray): 4160.

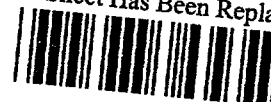
EXAMPLE 2: [Lys (~~NE~~-Isopropyl)^{26,27}]hPTH(1-34) NH₂

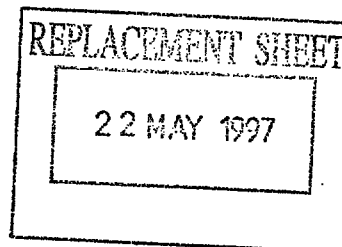
The peptide is assembled as described for Example 1 but using Lys(Fmoc) for incorporation in position 13 and Fmoc-Ser(tBu) in terminal position 1.

The fully assembled peptide resin is treated twice with 20 ml of thiophenol 5% in DMF at r.t. for 1 hour and washed with DMF, methanol and dichloromethane. The dry resin is treated with liquid HF as described for Example 1. The cleavage product (350 mg) is suspended in a mixture of methanol (5 ml), phosphate buffer pH = 5.0 (5 ml), sodium cyanoborohydride (48 mg) and acetone (0.28 ml). After 16 hours at RT the reaction mixture is filtered and the filtrate evaporated. The residue is suspended in 20%-piperidine in DMF for 15 minutes, diluted with 20 volumes of ether and filtered. The residue is dissolved in 100 ml of water and acetic acid added to pH = 3, filtered and the filtrate lyophilized. The lyophilizate is purified by reversed-phase chromatography on a column of octadecyl-silica using a gradient of acetonitril in 2%-phosphoric acid. Fractions containing the pure compound are combined, filtered through a weakly basic ion-exchange resin in the acetate form and the filtrate lyophilized to give the title compound.

MS (Ion Spray): 4205.6.

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EXAMPLE 3: N^ε-Isopropyl [Lys (N^ε-Isopropyl)^{13,26,27}]hPTH-(1-38)-OH

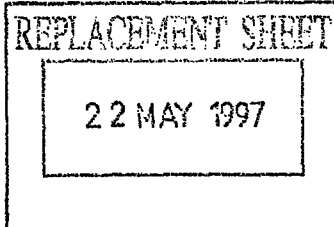
a) Solid phase peptide synthesis

The peptide is synthesized in a stepwise manner on a polystyrene based resin support. The alpha-amino group is protected by Fmoc and the side-chain functional groups as followed: Asp(OtBu), Glu(OtBu), His(Trt), Lys(Boc), Asn(Trt), Gln(Trt), Arg(Pmc) and Ser(tBu). Other amino acids are left unprotected.

Fmoc-Gly esterified to 4-Hydroxymethyl-phenoxyethyl-co(polystyrene-1%-divinylbenzene), 0.5 mmol/g, is used as starting material for the stepwise solid phase synthesis, which consists of repetitive cycles of alpha-amino deprotection, washing, coupling (attachment of new amino acid) and washing. A three to five fold excess of Fmoc-amino acids is coupled as preformed HOBt-esters using DIPC.

Fmoc-Arg(Pmc), Fmoc-Asn(Trt) and Fmoc-Gln(Trt) are coupled as symmetrical anhydrides using DIPC. After complete assembly of the peptide chain the Fmoc-protecting group of Ser¹ is removed by 20% piperidine/DMF. Cleavage of the peptide from the polystyrene resin and removal of all side chain protecting groups is obtained by treatment of the peptide resin with a mixture of 10% ethanedithiol, thioanisole, thiocresole and water in 90% TFA for three hours at room temperature. The resin particles are filtered off and washed with some TFA. The product is precipitated from the combined filtrates by addition of ether, filtered and dried. The product is purified by chromatography on a C-18 silica column using a gradient of acetonitrile in 2% H₃PO₄/water. Fractions containing the pure compound are collected, filtered through an anion-exchange resin (Biorad, AG 4-X4, 100-200mesh acetate form) and lyophilized to give hPTH-(1-38)-OH.





b) Alkylation

40 mg (9 μ mol) hPTH-(1-38)-OH(4458.2 g/mol) are dissolved in 1.2 ml methanol, 1.2 ml phosphate buffer (Merck no. 9887 adjusted to pH 5.0), 45 mg (716 μ mol) NaBH₃CN (62.84 g/mol) and 0.8 ml (11 μ mol) acetone (73.52 ml/mol). The reaction is monitored by RP-HPLC on a C-18 silica column and finished after 15 hours at room temperature. The reaction mixture is diluted with water and chromatographed on a C-18 silica column using a gradient of acetonitrile in 2% H₃PO₄/water. Fractions containing the pure compound are collected, filtered through an anion-exchange resin (acetate form) and lyophilized to give the title compound as polyacetate, polyhydrate.

$$[\alpha]_D^{20} = -5.7^{\circ} \quad (c = 0.317 \text{ in } 95\% \text{ AcOH})$$

MS (Ion-Spray): 4626

EXAMPLE 4: N^ε-Methyl [Ala¹]PTH-(1-38)-OH

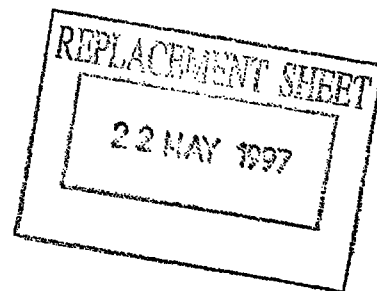
The peptide chain is assembled in the same manner as in example 3a. At position 1 instead of Fmoc-Ser(tBu)-OH the unnatural amino acid Fmoc-N^ε-Methyl-Ala-OH is coupled to the peptide resin. Cleavage, deprotection and purification is performed as in example 3a.

MS (Ion Spray) = 4455, 91

EXAMPLE 5: [Ala¹,Ala³,Leu⁸,Gln¹³,Ala¹⁶,Gln¹⁸,Ala¹⁹,Phe²³,His²⁵,His²⁶,Leu²⁷,Thr³³,Ala³⁴]hPTH(1-34)OH

This peptide is prepared in analogy with the procedure of Example 3a. Instead of Fmoc-Gly esterified to 4-hydroxymethyl-phenoxy-methyl-co(polystyrene-divinylbenzene) the appropriate Fmoc-amino acid, e.g. Fmoc-Ala, Fmoc-Phe, Fmoc-D-Ala, Fmoc-D-Phe, etc. is used. Additionally the side-chain functional groups are protected as follows: Thr(t.-butyl), Trp(Boc) and Tyr(t.-butyl).



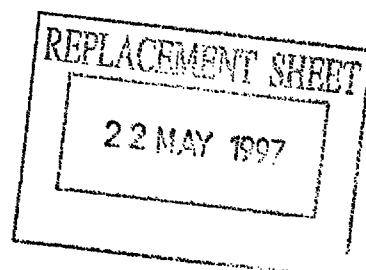


By repeating the procedure disclosed in Examples 3 and 5 but using the appropriate starting materials, the following compounds may be obtained:

- EXAMPLE 6: [Leu⁸, Asp¹⁰, Ala¹⁶, Gln¹⁸, Thr³³]hPTH(1-34)OH
- EXAMPLE 7: [Ile¹]hPTH(1-38)OH (IS-MS: 4484)
- EXAMPLE 8: [Ala¹, Abu²]hPTH(1-38)OH (IS-MS: 4428)
- EXAMPLE 9: [Ala¹, Nva²]hPTH(1-38)OH (IS-MS: 4442)
- EXAMPLE 10: [Ala¹, Ile²]hPTH(1-38)OH (IS-MS: 4456)
- EXAMPLE 11: [Ala¹, Ala³, Leu⁸, Gln¹³, Ala¹⁶, Gln¹⁸, Ala¹⁹, His²⁶, Leu²⁷, Thr³³, Ala³⁴]hPTH(1-34)OH
- EXAMPLE 12: [N-MeAla¹]hPTH(1-36)OH (IS-MS: 4286)
- EXAMPLE 13: [Ala¹, Ala³, Leu⁸, Gln¹⁸]hPTH(1-36)OH (IS-MS: 4235)
- EXAMPLE 14: [Thr¹]-hPTH-(1-38)OH (IS-MS: 4472)
- EXAMPLE 15: [Leu¹]-hPTH-(1-38)OH (IS-MS: 4484)
- EXAMPLE 16: [Abu¹]-hPTH-(1-38)OH (IS-MS: 4456)
- EXAMPLE 17: [Gaba¹]-hPTH-(1-38)OH (IS-MS: 4456)
- EXAMPLE 18: [Leu⁸, Lys¹¹, Gln¹⁸]hPTH(1-36)OH
- EXAMPLE 19: [Leu⁸, Gln¹⁶, Asp¹⁷, Leu¹⁸, Arg¹⁹, Arg²²]hPTH(1-36)OH (IS-MS: 4347)
- EXAMPLE 20: [Leu⁸, Gln¹⁸, Thr³³, Ala³⁴]-hPTH(1-34)OH (IS-MS: 4007)
- EXAMPLE 21: [Leu⁸, Ala¹⁶, Gln¹⁸, Ala¹⁹, Thr³³, Ala³⁴]hPTH(1-34)OH (IS-MS: 3906)
- EXAMPLE 22: [Leu⁸, Ala¹³, Gln¹⁸, Gln²⁶, Phe²⁷, Thr³³, Ala³⁴]hPTH(1-34)OH
- EXAMPLE 23: Ip-[Leu⁸, Lys(Ip)¹³, Gln¹⁸, Lys(Ip)^{26, 27}, Thr³³, Ala³⁴]hPTH(1-34)OH (IS-MS: 4175)
- EXAMPLE 24: Ip-[Leu⁸, Ala¹³, Gln¹⁸, Gln²⁶, Phe²⁷, Thr³³, Ala³⁴]hPTH(1-34)OH
- EXAMPLE 25: [Gln¹⁶]hPTH(1-38)OH
- EXAMPLE 26: [Ser¹⁴]hPTH(1-38)OH

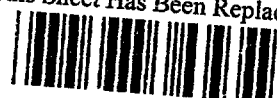
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- EXAMPLE 27: [Ala¹,Ala³,Leu⁸,Gln¹³,Ala¹⁶,Gln¹⁸,Ala¹⁹,His²⁵,His²⁶,
Leu²⁷,Thr³³,Ala³⁴]hPTH(1-34)OH
- EXAMPLE 28: [Ala¹,Ala³,Leu⁸,Gln¹³,Ala¹⁶,Gln¹⁸,Ala¹⁹,Gln²⁶,Phe²⁷,
Thr³³,Ala³⁴]hPTH(1-34)OH
- EXAMPLE 29: [Leu⁸,Ala¹⁶,Gln¹⁸,Thr³³,Ala³⁴]hPTH(1-34)OH
(IS-MS: 3965)
- EXAMPLE 30: [Leu⁸,Gln¹⁸,Ala²⁹,Glu³⁰,Ile³¹]hPTH(1-34)OH
- EXAMPLE 31: [Leu⁸,Asp¹⁰,Lys¹¹,Gln¹⁸]hPTH(1-36)OH
- EXAMPLE 32: [Leu⁸,Asp¹⁰,Lys¹¹,Ser¹⁴,Ile¹⁵,Gln¹⁶,Asp¹⁷,Leu¹⁸,
Arg¹⁹]hPTH(1-36)OH
- EXAMPLE 33: [Leu⁸,Asp¹⁰,Lys¹¹,Leu¹⁸]hPTH(1-36)OH
- EXAMPLE 34: [Leu⁸,Gln¹⁶,Asp¹⁷,Leu¹⁸,Arg¹⁹]hPTH(1-36)OH
- EXAMPLE 35: [Leu⁸,Asp¹⁰,Lys¹¹,Ala¹⁷,Leu¹⁸]hPTH(1-36)OH (IS-MS:
4252)
- EXAMPLE 36: [Leu⁸,Gln¹⁶,Asp¹⁷,Leu¹⁸,Arg¹⁹,Thr³³,Ala³⁴]
hPTH(1-34)OH
- EXAMPLE 37: [Leu⁸,Asp¹⁰,Lys¹¹,Gln¹⁸,Thr³³,Ala³⁴]hPTH(1-34)OH
(IS-MS: 4022)
- EXAMPLE 38: [Leu⁸,Ala¹⁶,Asp¹⁷,Leu¹⁸,Ala¹⁹]hPTH(1-36)OH
- EXAMPLE 39: [Leu⁸,Asp¹⁰,Ala¹⁶,Asp¹⁷,Leu¹⁸,Ala¹⁹]hPTH(1-36)OH
- EXAMPLE 40: [Leu⁸,Gln¹⁸,Arg²²,Thr³³,Ala³⁴]hPTH(1-34)OH
- EXAMPLE 41: [Leu⁸,Asp¹⁰,Lys¹¹,Gln¹⁶,Asp¹⁷,Leu¹⁸,Arg¹⁹,Thr³³,
Ala³⁴]hPTH(1-34)OH (IS-MS: 4077)
- EXAMPLE 42: [Leu⁸,Asp¹⁰,Lys¹¹,Ala¹⁶,Gln¹⁸,Ala¹⁹]hPTH(1-36)OH
(IS-MS: 4181)
- EXAMPLE 43: [Leu⁸,Ala¹⁶,Asp¹⁷,Gln¹⁸,Ala¹⁹]hPTH(1-36)OH (IS-MS:
4193)
- EXAMPLE 44: [Leu⁸,Ala¹⁶,Ala¹⁷,Gln¹⁸,Ala¹⁹]hPTH(1-36)OH (IS-MS:
4149)
- EXAMPLE 45: [Leu⁸,Ala¹⁷,Gln¹⁸,Ala¹⁹]hPTH(1-36)OH
- EXAMPLE 46: [Leu⁸,Ala¹⁷,Gln¹⁸,Ala¹⁹,Arg²²]hPTH(1-36)OH (IS-MS:
4219)
- EXAMPLE 47: [Leu⁸,Ala¹⁷,Gln¹⁸,Ala¹⁹,Arg²²,Thr³³,Ala³⁴]
hPTH(1-34)OH (IS-MS: 3960)

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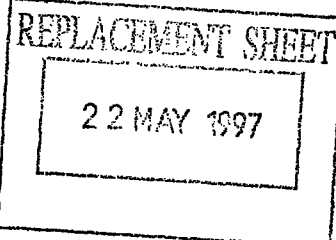


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- EXAMPLE 48: [Leu⁸,Gln¹⁸]hPTH(1-36)OH
- EXAMPLE 49: [Leu⁸,Asp¹⁰,Lys¹¹,Ala¹⁶,Gln¹⁸,Thr³³,Ala³⁴]
hPTH(1-34)OH (IS-MS: 3980)
- EXAMPLE 50: [Leu⁸,Asp¹⁰,Lys¹¹,Ala¹⁶,Gln¹⁸]hPTH(1-36)OH
(IS-MS: 4240)
- EXAMPLE 51: [Leu⁸,Asp¹⁰,Lys¹¹,Ala¹⁶,Gln¹⁸,Ala¹⁹,Thr³³,Ala³⁴]
hPTH(1-34)OH (IS-MS: 3923)
- EXAMPLE 52: [Leu⁸,Asp¹⁰,Ala¹⁶,Gln¹⁸]hPTH(1-36)OH
(IS-MS: 4225)
- EXAMPLE 53: [Leu⁸,Asp¹⁰,Lys¹¹,Ala¹⁶,Gln¹⁸,Ala¹⁹,Thr³³]
hPTH(1-34)OH (IS-MS: 3999)
- EXAMPLE 54: [Leu⁸,Gln¹³,Ala¹⁶,Gln¹⁸,Ala¹⁹,His²⁶,Leu²⁷,Thr³³]
hPTH(1-34)OH
- EXAMPLE 55: [Leu⁸,Ala¹⁶,Gln¹⁸,Ala¹⁹,Arg²²]hPTH(1-36)OH
- EXAMPLE 56: [Ile¹⁵]hPTH(1-38)OH
- EXAMPLE 57: [Leu⁸,Gln¹³,Ala¹⁶,Gln¹⁸,Ala¹⁹,Arg²²,His²⁶,Leu²⁷,
Thr³³,Ala³⁴]hPTH(1-34)OH
- EXAMPLE 58: [Leu⁸,Gln¹³,Ala¹⁶,Gln¹⁸,Arg¹⁹,His²⁶,Leu²⁷,Thr³³,
Ala³⁴]hPTH(1-34)OH
- EXAMPLE 59: [Leu⁸,Ser¹³,Ala¹⁶,Gln¹⁸,Ala¹⁹,Arg²²]hPTH(1-36)OH
- EXAMPLE 60: [Leu⁸,Ala¹³,Ala¹⁶,Gln¹⁸,Ala¹⁹,Arg²⁶,Arg²⁷]
hPTH(1-36)OH
- EXAMPLE 61: [Leu⁸,Gln¹³,Ala¹⁶,Gln¹⁸,Ala¹⁹,His²⁶,Leu²⁷,Arg³³,
Ala³⁴]hPTH(1-34)OH
- EXAMPLE 62: [Leu⁸,Ala^{16,17,18,19}]hPTH(1-36)OH
- EXAMPLE 63: [Leu⁸,Ala¹³,Ala¹⁶,Gln¹⁸,Ala¹⁹,Gln²⁶,Phe²⁷]
hPTH(1-36)OH (IS-MS: 4127)
- EXAMPLE 64: Ip-[Leu⁸,Ala¹³,Ala¹⁶,Gln¹⁸,Ala¹⁹,Gln²⁶,Phe²⁷]
hPTH(1-36)OH
- EXAMPLE 65: Ip-[Leu⁸,Lys(Ip)¹³,Ala¹⁶,Gln¹⁸,Ala¹⁹,
Lys(Ip)^{26,27}]hPTH(1-36)OH
- EXAMPLE 66: [Leu⁸,Ala¹⁶,Gln¹⁸,Ala¹⁹]hPTH(1-36)OH (IS-MS: 4166)
- EXAMPLE 67: Ip-[Leu⁸,Lys(Ip)¹³,Ala¹⁶,Ala¹⁷,Ala¹⁸,Ala¹⁹,
Lys(Ip)^{26,27}]hPTH(1-36)OH

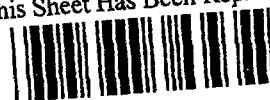
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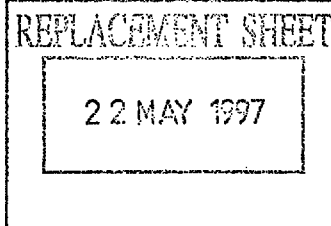




- EXAMPLE 68: [Aib³,Gln¹⁸]hPTH(1-36)OH (IS-MS: 4283)
- EXAMPLE 69: Ip-[Leu⁸,Asp¹⁰,Lys(Ip)^{11,13,26,27},Gln¹⁸]
hPTH(1-36)OH
- EXAMPLE 70: Ip-[Leu⁸,Ser¹³,Ala¹⁶,Gln¹⁸,Ala¹⁹,Arg²²,
Lys(Ip)^{26,27}]hPTH(1-36)OH
- EXAMPLE 71: [Leu⁸,Tyr¹⁸]hPTH(1-36)OH
- EXAMPLE 72: [Ser³³]-hPTH-(1-38)-OH
- EXAMPLE 73: [Thr³³]-hPTH-(1-38)-OH
- EXAMPLE 74: [Leu³³]-hPTH-(1-38)-OH
- EXAMPLE 75: [Gly³³]-hPTH-(1-38)-OH
- EXAMPLE 76: [Leu⁸,His¹⁰,Gln¹⁸,Arg²²,Thr³³,Ala³⁴]hPTH(1-34)OH
- EXAMPLE 77: [Leu⁸,Gly¹⁰,Gln¹⁸,Arg²²,Thr³³,Ala³⁴]hPTH(1-34)OH
- EXAMPLE 78: [Leu⁸,Glu¹⁰,Gln¹⁸,Arg²²,Thr³³,Ala³⁴]hPTH(1-34)OH
- EXAMPLE 79: [Leu⁸,Thr¹⁰,Gln¹⁸,Arg²²,Thr³³,Ala³⁴]hPTH(1-34)OH
- EXAMPLE 80: [Leu⁸,Gln¹⁰,Gln¹⁸,Arg²²,Thr³³,Ala³⁴]hPTH(1-34)OH
- EXAMPLE 81: [Gln³³]-hPTH-(1-38)-OH
- EXAMPLE 82: [Leu⁸,Tyr¹⁰,Gln¹⁸,Arg²²,Thr³³,Ala³⁴]hPTH(1-34)OH
- EXAMPLE 83: [1-amino-cyclopentane-1-carboxylic
acid¹,Leu⁸,Ala¹⁶,Gln¹⁸,Ala¹⁹]hPTH(1-36)OH
- EXAMPLE 84: [1-amino-cyclopentane-1-carboxylic acid¹,Leu⁸,
Ala^{13,16},Gln¹⁸,Ala¹⁹,Arg^{26,27}]hPTH(1-36)OH
- EXAMPLE 85: [1-amino-cyclopentane-1-carboxylic acid³,Gln¹⁸]
hPTH(1-36)OH (IS-MS: 4309)
- EXAMPLE 86: [Arg¹²]-hPTH-(1-38)-OH
- EXAMPLE 87: [Ser¹²]-hPTH-(1-38)-OH
- EXAMPLE 88: [Cys¹³]-hPTH-(1-38)-OH
- EXAMPLE 89: [Ile¹³]-hPTH-(1-38)-OH
- EXAMPLE 90: [Asn¹³]-hPTH-(1-38)-OH
- EXAMPLE 91: [Trp¹³]-hPTH-(1-38)-OH
- EXAMPLE 92: [Asp¹³]-hPTH-(1-38)-OH
- EXAMPLE 93: [Val¹³]-hPTH-(1-38)-OH
- EXAMPLE 94: [Thr¹³]-hPTH-(1-38)-OH
- EXAMPLE 95: [Ser¹³]-hPTH-(1-38)-OH

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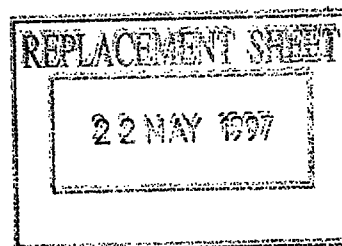




EXAMPLE 96: [Tyr¹³]-hPTH-(1-38)-OH
EXAMPLE 97: [Met¹³]-hPTH-(1-38)-OH
EXAMPLE 98: [Gln¹³]-hPTH-(1-38)-OH
EXAMPLE 99: [Leu¹³]-hPTH-(1-38)-OH
EXAMPLE 100: [Ala¹³]-hPTH-(1-38)-OH
EXAMPLE 101: [Gly¹³]-hPTH-(1-38)-OH
EXAMPLE 102: [Val¹⁴]-hPTH-(1-38)-OH
EXAMPLE 103: [Ala¹⁴]-hPTH-(1-38)-OH
EXAMPLE 104: [Lys¹⁴]-hPTH-(1-38)-OH
EXAMPLE 105: [Arg¹⁴]-hPTH-(1-38)-OH
EXAMPLE 106: [Thr¹⁴]-hPTH-(1-38)-OH
EXAMPLE 107: [Ile¹⁴]-hPTH-(1-38)-OH
EXAMPLE 108: [Tyr¹⁴]-hPTH-(1-38)-OH
EXAMPLE 109: [Tyr¹⁵]-hPTH-(1-38)-OH
EXAMPLE 110: [Arg¹⁵]-hPTH-(1-38)-OH
EXAMPLE 111: [Val¹⁵]-hPTH-(1-38)-OH
EXAMPLE 112: [Lys¹⁶]-hPTH-(1-38)-OH
EXAMPLE 113: [Ser¹⁶]-hPTH-(1-38)-OH
EXAMPLE 114: [Leu¹⁶]-hPTH-(1-38)-OH
EXAMPLE 115: [Ala¹⁶]-hPTH-(1-38)-OH
EXAMPLE 116: [Gly¹⁶]-hPTH-(1-38)-OH
EXAMPLE 117: [Ala¹⁷]-hPTH-(1-38)-OH
EXAMPLE 118: [Met¹⁷]-hPTH-(1-38)-OH
EXAMPLE 119: [Ile¹⁷]-hPTH-(1-38)-OH
EXAMPLE 120: [Ser¹⁹]-hPTH-(1-38)-OH
EXAMPLE 121: [Lys¹⁹]-hPTH-(1-38)-OH
EXAMPLE 122: [Leu¹⁹]-hPTH-(1-38)-OH
EXAMPLE 123: [Ala¹⁹]-hPTH-(1-38)-OH
EXAMPLE 124: [Tyr¹⁹]-hPTH-(1-38)-OH
EXAMPLE 125: [Met¹⁹]-hPTH-(1-38)-OH
EXAMPLE 126: [His¹⁹]-hPTH-(1-38)-OH
EXAMPLE 127: [Val¹⁹]-hPTH-(1-38)-OH
EXAMPLE 128: [Gly¹⁹]-hPTH-(1-38)-OH
EXAMPLE 129: [Pro¹⁹]-hPTH-(1-38)-OH

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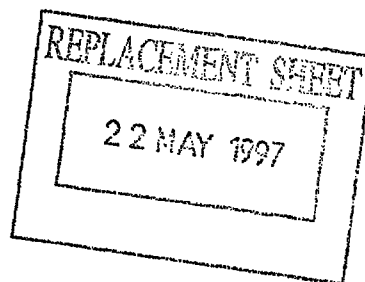




EXAMPLE 130: [Asp¹⁹]-hPTH-(1-38)-OH
EXAMPLE 131: [Ile¹⁹]-hPTH-(1-38)-OH
EXAMPLE 132: [Val¹⁹,Gln²⁴]-hPTH-(1-38)-OH
EXAMPLE 133: [Arg¹⁹]-hPTH-(1-38)-OH
EXAMPLE 134: [Phe²⁰]-hPTH-(1-38)-OH
EXAMPLE 135: [Ala²¹]-hPTH-(1-38)-OH
EXAMPLE 136: [Gly²¹]-hPTH-(1-38)-OH
EXAMPLE 137: [Phe²¹]-hPTH-(1-38)-OH
EXAMPLE 138: [Leu²¹]-hPTH-(1-38)-OH
EXAMPLE 139: [Asn²¹]-hPTH-(1-38)-OH
EXAMPLE 140: [Gln²¹]-hPTH-(1-38)-OH
EXAMPLE 141: [Ser²¹]-hPTH-(1-38)-OH
EXAMPLE 142: [Gly²²]-hPTH-(1-38)-OH
EXAMPLE 143: [Leu²²]-hPTH-(1-38)-OH
EXAMPLE 144: [His²²]-hPTH-(1-38)-OH
EXAMPLE 145: [Ala²²]-hPTH-(1-38)-OH
EXAMPLE 146: [Ile²²]-hPTH-(1-38)-OH
EXAMPLE 147: [Val²²]-hPTH-(1-38)-OH
EXAMPLE 148: [Ser²²]-hPTH-(1-38)-OH
EXAMPLE 149: [Arg²²]-hPTH-(1-38)-OH
EXAMPLE 150: [Arg²⁶]-hPTH-(1-38)-OH
EXAMPLE 151: [Val²⁷]-hPTH-(1-38)-OH
EXAMPLE 152: [Ile²⁷]-hPTH-(1-38)-OH
EXAMPLE 153: [Leu²⁷]-hPTH-(1-38)-OH
EXAMPLE 154: [Arg²⁷]-hPTH-(1-38)-OH
EXAMPLE 155: [Ala²⁷]-hPTH-(1-38)-OH
EXAMPLE 156: [Val²⁸]-hPTH-(1-38)-OH
EXAMPLE 157: [Ile²⁸]-hPTH-(1-38)-OH
EXAMPLE 158: [Pro³,Thr³³]-hPTH-(1-38)-OH
EXAMPLE 159: [Arg³³]-hPTH-(1-38)-OH
EXAMPLE 160: [Pro³³]-hPTH-(1-38)-OH
EXAMPLE 161: [Asp³³]-hPTH-(1-38)-OH
EXAMPLE 162: [Ile³³]-hPTH-(1-38)-OH
EXAMPLE 163: [Lys³³]-hPTH-(1-38)-OH

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EXAMPLE 164: [Ile³¹, Arg³³]-hPTH-(1-38)-OH

EXAMPLE 165: [1-aminocyclopentane-1-carboxylic acid¹]hPTH-
(1-36)NH₂

The peptide is assembled in a stepwise manner on a polystyrene based resin support. The Fmoc-group is used for protection of the alpha-amino groups.

Side-chain functional groups are protected as Arg(Pmc), Asn(Trt), Asp(OtBu), Gln(Trt), Glu(OtBu), His(Trt), Lys(Boc), Ser(tBu), Thr(tBu), Trp(Boc) and Tyr(tBu). Other amino acids are left unprotected.

4-(2',4'-Dimethoxyphenyl-Fmoc-amino-methyl)-phenoxy-co(polystyrene-divinylbenzene), 0.4 mmol/g, which may be prepared, e.g., as described in Tetrah. Letters, 28, 3787-3790 (1987) is subjected to the following cycle, steps (1) to (5), of treatments:

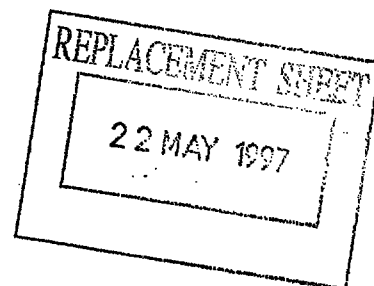
- (1) DMF
- (2) piperidine (20%) in DMF
- (3) DMF
- (4) mixture of HOBT, diisopropylcarbodiimide, and Fmoc-alanine (0.8 mmol per gram starting resin each)
- (5) DMF

Volumes of washes and reagents are from 5 to 20 mL per gram of starting resin.

In the next cycle of treatments (1) to (5), Fmoc-valine is substituted for Fmoc-alanine and so on for each cycle such as to assemble on the resin the correct amino acid sequence of the title compound.

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Each step is repeated as many times as necessary for either complete reaction of the resin (steps 2, 4) or complete displacement of the previous reagent(s) from the resin (steps 3, 5). Samples of the resin are removed after each cycle and checked for completeness of the coupling reaction by a colorimetric test for residual amino groups using ninhydrin.

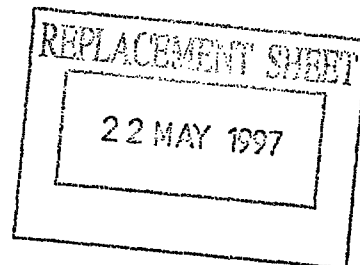
At the end of the synthesis a final cycle comprising steps (1) to (3) only is performed and the peptide resin is washed with 2-propanol, then with a mixture of methanol and methylene chloride (1:1 v/v) and dried thoroughly in a vacuum desiccator.

The peptide resin (1 g) is suspended in a mixture (20 ml) of trifluoroacetic acid, water, and 1,2-ethanedithiol (90:5:5 v/v) for 2 hours at room temperature, the resin particles are filtered off and washed with a small volume of TFA. The product is precipitated from the combined filtrates by addition of ether (20 volumes), filtered, washed with more ether and dried. The residue is dissolved in 2 % acetic acid, the solution let to stand at r.t. for 8 hours prior to lyophilization. The lyophilisate is chromatographed on a C-18 silica column using a gradient of acetonitrile in 2% H_3PO_4 . Fractions are checked by analytical HPLC and those containing the pure compound are collected, filtered through an anion-exchange resin in the acetate form and lyophilised to give the title compound as a polyacetate, polyhydrate.

Fmoc-1-aminocyclopentane-1-carboxylic acid used in the preparation of the peptide resin intermediate may be prepared, e.g. as described by G. Valle et al., 1988, in Can.J.Chem. 66:2575-2582.
MS (Ion-Spray): 4312

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EXAMPLE 166: [1-(1-aminocyclopropane-1-carboxylic acid)]hPTH-
(1-36)NH₂

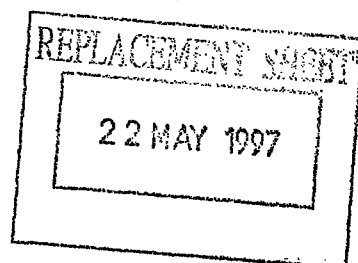
This peptide is prepared in analogy with the procedure of Example 165. Fmoc-1-aminocyclopropane-1-carboxylic acid used in the preparation of the peptide resin intermediate may be prepared, e.g., as described by M. Crisma et al. (1989) in Int.J.Biol.Macromol.11:345-352. MS(Ion-Spray): 4283

By repeating the procedure as disclosed in Example 165 but using the appropriate starting materials, following compounds may be obtained:

- EXAMPLE 167: [D-Pro¹]hPTH(1-36)NH₂
EXAMPLE 168: [Nva¹]hPTH(1-36)NH₂
EXAMPLE 169: [N-Me-Ser¹]hPTH(1-36)NH₂
EXAMPLE 170: [Indole-2-carboxylic acid¹]hPTH(1-36)NH₂
EXAMPLE 171: [Indole-3-carboxylic acid¹]hPTH(1-36)NH₂
EXAMPLE 172: [Pyridine-3-carboxylic acid¹]hPTH(1-36)NH₂
EXAMPLE 173: [Pyridine-2-carboxylic acid¹]hPTH(1-36)NH₂
EXAMPLE 174: [Hexahydropyridazine-3-carboxylic acid¹]hPTH(1-36)NH₂
EXAMPLE 175: [Morpholine-2-carboxylic acid¹]hPTH(1-36)NH₂
EXAMPLE 176: [Pro¹]hPTH(1-36)NH₂
EXAMPLE 177: [Leu¹]hPTH(1-36)NH₂
EXAMPLE 178: [Ile¹]hPTH(1-36)NH₂
EXAMPLE 179: [Thr³³,Ala³⁴]hPTH(1-34)NH₂
EXAMPLE 180: [Nva⁸]hPTH(1-36)NH₂
EXAMPLE 181: [Gln¹⁸]hPTH(1-36)NH₂
EXAMPLE 182: [Tyr¹⁸]hPTH(1-36)NH₂
EXAMPLE 183: [Lys¹⁸]hPTH(1-36)NH₂
EXAMPLE 184: [Ala¹⁸]hPTH(1-36)NH₂
EXAMPLE 185: [Phe²³,His²⁵,His²⁶,Leu²⁷]-hPTH(1-34)NH₂
EXAMPLE 186: [Phe²³]-hPTH(1-36)NH₂ (IS-MS: 4247)

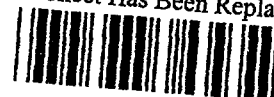
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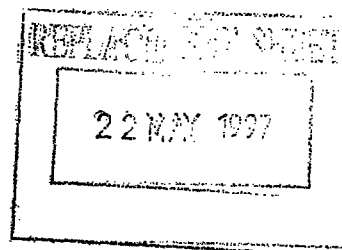




- EXAMPLE 187: [Phe²³, His²⁵, His²⁶, Leu²⁷, Ile²⁸, Ala²⁹, Glu³⁰, Ile³¹, Thr³³, Ala³⁴]-hPTH(1-34)NH₂ (IS-MS:3934)
- EXAMPLE 188: [Ala²⁹]-hPTH(1-36)NH₂ (IS-MS: 4248)
- EXAMPLE 189: [Ala³⁴]-hPTH(1-36)NH₂ (IS-MS: 4210)
- EXAMPLE 190: [Gln²⁵]hPTH(1-36)NH₂
- EXAMPLE 191: [Leu⁸, Asp¹⁰, Lys¹¹, Thr³³, Ala³⁴]hPTH(1-34)NH₂
- EXAMPLE 192: [Ala¹, His⁵, Leu⁸, Asp¹⁰, Lys¹¹, Thr³³, Ala³⁴]
hPTH(1-34)NH₂
- EXAMPLE 193: [Ser¹⁴, Ile¹⁵, Gln¹⁶, Asp¹⁷, Leu¹⁸, Arg¹⁹, Thr³³, Ala³⁴]
hPTH(1-34)NH₂
- EXAMPLE 194: [D-Phe³⁴, D-Ala³⁶]hPTH(1-36)-NH₂ [MS(IS): 4290]
- EXAMPLE 195: [Ala³]hPTH(1-36)NH₂ [MS(IS):4271]
- EXAMPLE 196: [D-Ser³]hPTH(1-36)NH₂ [MS(IS):4290]
- EXAMPLE 197: [D-Glu⁴]hPTH(1-36)NH₂ [MS(IS):4290]
- EXAMPLE 198: [D-His⁹]hPTH(1-36)NH₂ [MS(IS):4286]
- EXAMPLE 199: [Ala¹⁰]hPTH(1-36)NH₂ [MS(IS):4243]
- EXAMPLE 200: [D-Asn¹⁰]hPTH(1-36)NH₂ [MS(IS): 4288]
- EXAMPLE 201: [Ala¹²]hPTH(1-36)NH₂ [MS(IS): 4299]
- EXAMPLE 202: [Gln¹³]hPTH(1-36)NH₂ [MS(IS): 4287]
- EXAMPLE 203: [His¹³]hPTH(1-36)NH₂ [MS(IS): 4296]
- EXAMPLE 204: [Leu¹³]hPTH(1-36)NH₂ [MS(IS): 4371]
- EXAMPLE 205: [Ala¹³]hPTH(1-36)NH₂ [MS(IS): 4228]
- EXAMPLE 206: [Ala¹³, Gln²⁶, Phe²⁷, D-Phe³⁴, D-Ala³⁶]hPTH(1-36)NH₂
[MS(IS): 4249]
- EXAMPLE 207: [Ala¹⁴]hPTH(1-36)NH₂ [MS(IS): 4219]
- EXAMPLE 208: [D-His¹⁴]hPTH(1-36)NH₂ [MS(IS): 4288]
- EXAMPLE 209: [D-Asn¹⁶]hPTH(1-36)NH₂ [MS(IS): 4284]
- EXAMPLE 210: [Ala¹⁷]hPTH(1-36)NH₂ [MS(IS): 4270]
- EXAMPLE 211: [D-Ser¹⁷]hPTH(1-36)NH₂ [MS(IS): 4288]
- EXAMPLE 212: [Ala¹⁹]hPTH(1-36)NH₂ [MS(IS): 4228]
- EXAMPLE 213: [D-Glu¹⁹]hPTH(1-36)NH₂ [MS(IS): 4288]
- EXAMPLE 214: [Ala²¹]hPTH(1-36)NH₂ [MS(IS): 4257]
- EXAMPLE 215: [Ala²²]hPTH(1-36)NH₂ [MS(IS): 4227]
- EXAMPLE 216: [Gln²⁶]hPTH(1-36)NH₂ [MS(IS): 4287]

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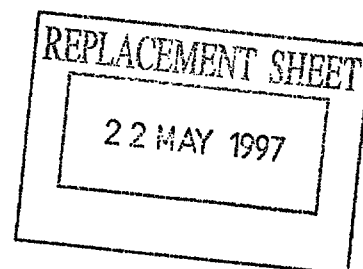




- EXAMPLE 217: [Nle²⁶]hPTH(1-36)NH₂ [MS(IS): 4271]
- EXAMPLE 218: [D-Lys²⁶]hPTH(1-36)NH₂ [MS(IS): 4288]
- EXAMPLE 219: [Nle^{8, 18, 27}]hPTH(1-36)NH₂
- EXAMPLE 220: [His²⁷]hPTH(1-36)NH₂ [MS(IS): 4294]
- EXAMPLE 221: [Phe²⁷]hPTH(1-36)NH₂ [MS(IS): 4304]
- EXAMPLE 222: [Nle²⁷]hPTH(1-36)NH₂ [MS(IS): 4271]
- EXAMPLE 223: [Asn²⁷]hPTH(1-36)NH₂ [MS(IS): 4271]
- EXAMPLE 224: [Ala²⁷]hPTH(1-36)NH₂ [MS(IS): 4228]
- EXAMPLE 225: [D-Gln²⁹]hPTH(1-36)NH₂ [MS(IS): 4289]
- EXAMPLE 226: [D-Asp³⁰]hPTH(1-34)NH₂ [MS(IS): 4118]
- EXAMPLE 227: [Ala³⁰]hPTH(1-36)NH₂ [MS(IS): 4241]
- EXAMPLE 228: [D-Val³¹]hPTH(1-36)NH₂ [MS(IS): 4290]
- EXAMPLE 229: [Ala³¹]hPTH(1-36)NH₂ [MS(IS): 4258]
- EXAMPLE 230: [Lys³²]hPTH(1-34)NH₂ [MS(IS): 3832]
- EXAMPLE 231: [D-His³²]hPTH(1-36)NH₂ [MS(IS): 4288]
- EXAMPLE 232: [Ala³²]hPTH(1-36)NH₂ [MS(IS): 4219]
- EXAMPLE 233: [D-Asn³³]hPTH(1-36)NH₂ [MS(IS): 4288]
- EXAMPLE 234: [Ala³³]hPTH(1-36)NH₂ [MS(IS): 4210]
- EXAMPLE 235: [NMePhe³⁴]hPTH(1-36)NH₂ [MS(IS): 4301]
- EXAMPLE 236: [D-Asp³⁰]hPTH(1-36)NH₂ [MS(IS): 4290]
- EXAMPLE 237: Ip-[Nle^{8, 18}, Lys(Ip)^{13, 26, 27}, D-Asn³³, D-Phe³⁴]
hPTH(1-34)NH₂
- EXAMPLE 238: Ip-[Nle^{8, 18, 27}, Lys(Ip)^{13, 26}]hPTH(1-36)NH₂
- EXAMPLE 239: [Nle^{8, 18}, D-Asn³³, D-Phe³⁴]hPTH(1-34)NH₂
[MS(IS): 4080]
- EXAMPLE 240: [Lys(Ip)¹³]hPTH(1-36)NH₂ [MS(IS): 4331]
- EXAMPLE 241: Propargyl-[A¹]-hPTH-(1-36)-NH₂
- EXAMPLE 242: [Ala⁰]-hPTH-(1-36)-NH₂
- EXAMPLE 243: [Pro⁰]-hPTH-(1-36)-NH₂
- EXAMPLE 244: Acetyl-hPTH-(1-36)-NH₂
- EXAMPLE 245: [HyPro¹]-hPTH-(1-36)-NH₂
- EXAMPLE 246: N-Dimethyl-[Ala¹]-hPTH-(1-36)-NH₂
- EXAMPLE 247: [D-Ser¹]-hPTH-(1-36)-NH₂ [MS(IS): 4288]
- EXAMPLE 248: [Lys(For)¹]-hPTH-(1-36)-NH₂ [MS(IS): 4356]

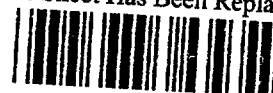
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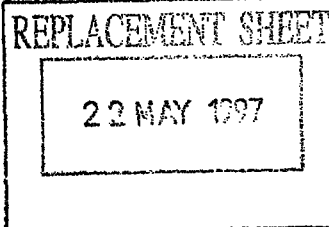




- EXAMPLE 249: [D-Glyceric acid¹]-hPTH-(1-36)-NH₂
- EXAMPLE 250: [Asn¹]-hPTH-(1-36)-NH₂
- EXAMPLE 251: [4-Aminobenzoic acid¹]-hPTH-(1-36)-NH₂
- EXAMPLE 252: [4-Aminosalicylic acid¹]-hPTH-(1-36)-NH₂
- EXAMPLE 253: [TMSA¹]-hPTH-(1-36)-NH₂ [MS (IS): 4343]
- EXAMPLE 254: [Phe¹]-hPTH-(1-36)-NH₂
- EXAMPLE 255: [Propargylglycin¹]-hPTH-(1-36)-NH₂
- EXAMPLE 256: [Ala¹, His⁵, Leu⁸, Asp¹⁰, Lys¹¹, Gln¹⁸, Phe²², Phe²³, His²⁵, His²⁶,
Leu²⁷, Thr³³, Ala³⁴]-hPTH-(1-34)-NH₂
- EXAMPLE 257: [Abu²]-hPTH-(1-36)-NH₂
- EXAMPLE 258: [D-Val²]-hPTH-(1-36)-NH₂ [MS (IS): 4288]
- EXAMPLE 259: [tert. Leu²]-hPTH-(1-36)-NH₂
- EXAMPLE 260: [Ala¹]-hPTH-(1-36)-NH₂
- EXAMPLE 261: [D-Ile⁵]-hPTH-(1-36)-NH₂ [MS (IS): 4288]
- EXAMPLE 262: [D-Gln⁶]-hPTH-(1-36)-NH₂ [MS (IS): 4288]
- EXAMPLE 263: [D-Leu⁷]-hPTH-(1-36)-NH₂ [MS (IS): 4288]
- EXAMPLE 264: [Nle⁸]-hPTH-(1-36)-NH₂ [MS (IS): 4268]
- EXAMPLE 265: [Phe⁸]-hPTH-(1-36)-NH₂ [MS (IS): 4303]
- EXAMPLE 266: [Cha⁸]-hPTH-(1-36)-NH₂ [MS (IS): 4309]
- EXAMPLE 267: [Leu⁸]-hPTH-(1-38)-NH₂
- EXAMPLE 268: [D-Leu¹¹]-hPTH-(1-36)-NH₂ [MS (IS): 4287]
- EXAMPLE 269: [Ala¹¹]-hPTH-(1-36)-NH₂ [MS (IS): 4244]
- EXAMPLE 270: [D-Lys¹³]-hPTH-(1-36)-NH₂ [MS (IS): 4288]
- EXAMPLE 271: [D-Leu¹⁵]-hPTH-(1-36)-NH₂ [MS (IS): 4288]
- EXAMPLE 272: [Ala¹⁵]-hPTH-(1-36)-NH₂ [MS (IS): 4243]
- EXAMPLE 273: [Ala¹⁶]-hPTH-(1-36)-NH₂ [MS (IS): 4243]
- EXAMPLE 274: [Met (O₂)¹⁸]-hPTH-(1-36)-NH₂ [MS (IS): 4320]
- EXAMPLE 275: [Nle¹⁸]-hPTH-(1-36)-NH₂ [MS (IS): 4268]
- EXAMPLE 276: [D-Met¹⁸]-hPTH-(1-36)-NH₂ [MS (IS): 4288]
- EXAMPLE 277: [Lys²⁰]-hPTH-(1-36)-NH₂ [MS (IS): 4259]
- EXAMPLE 278: [D-Arg²⁰]-hPTH-(1-36)-NH₂ [MS (IS): 4288]
- EXAMPLE 279: [D-Val²¹]-hPTH-(1-36)-NH₂ [MS (IS): 4288]
- EXAMPLE 280: [Trp (SO₂Pmc)²³]-hPTH-(1-38)-NH₂ [MS (IS): 4723]
- EXAMPLE 281: [Trp (Pmc)²³]-hPTH-(1-38)-NH₂ [MS (IS): 4660]

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EXAMPLE 282: [D-Trp²³]-hPTH-(1-36)-NH₂ [MS(IS): 4288]
EXAMPLE 283: [Ala²³]-hPTH-(1-36)-NH₂ [MS(IS): 4171]
EXAMPLE 284: [D-Leu²⁴]-hPTH-(1-36)-NH₂ [MS(IS): 4288]
EXAMPLE 285: [Phe²⁵]-hPTH-(1-36)-NH₂ [MS(IS): 4277]
EXAMPLE 286: [Lys²⁵]-hPTH-(1-36)-NH₂ [MS(IS): 4258]
EXAMPLE 287: [Ala²⁵]-hPTH-(1-36)-NH₂ [MS(IS): 4201]
EXAMPLE 288: [Ala²⁶]-hPTH-(1-36)-NH₂ [MS(IS): 4229]
EXAMPLE 289: [Lys^{26/27}(For)]-hPTH-(1-34)-NH₂
EXAMPLE 290: [D-Lys²⁷]-hPTH-(1-36)-NH₂ [MS(IS): 4288]
EXAMPLE 291: [D-Leu²⁸]-hPTH-(1-36)-NH₂ [MS(IS): 4288]
EXAMPLE 292: [D-Phe³⁴]-hPTH-(1-36)-NH₂ [MS(IS): 4288]
EXAMPLE 293: [D-Val³⁵]-hPTH-(1-36)-NH₂
EXAMPLE 294: [Ala³⁵]-hPTH-(1-36)-NH₂
EXAMPLE 295: [Pro³⁵]-hPTH-(1-36)-NH₂
EXAMPLE 296: [NMeVal³⁵]-hPTH-(1-36)-NH₂
EXAMPLE 297: [Thr³⁵,Ala³⁶]-hPTH-(1-36)-NH₂
EXAMPLE 298: [D-Ala³⁶]-hPTH-(1-36)-NH₂
EXAMPLE 299: [NMeAla³⁶]-hPTH-(1-36)-NH₂

EXAMPLE 300: (5-Amino-4-fluoro-2-isopropyl)-hex-3-enoyl-hPTH(3-
36)NH₂

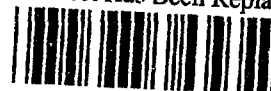
a) Preparation of hPTH-(3-36)-NH-resin

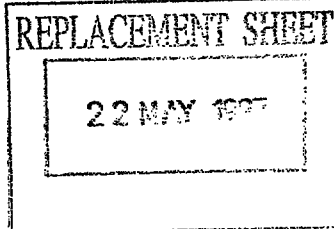
This intermediate is prepared in analogy with the procedure of Example 165.

b) [(5-Amino-4-fluoro-2-isopropyl)-hex-3-enoyl]hPTH(3-36)-amide

204.6 mg Fmoc-hPTH(3-36)-NH-resin obtained in a) above are treated with DMF for 5 min. and then washed several times with isopropanol and DMF. The Fmoc-protecting group is removed by treatment with DMF/piperidin 8:2 for 10 min.. The deprotected

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PTH-fragment, 38.1 mg of 4-fluoro-2-isopropyl-5-tert-butoxycarbonylamino-hex-3-enoic acid, 68.7 mg benzotriazol-1-yloxytris(pyrrolidino)phosphonium hexafluorophosphate (PyBOP) and 26.6 mg 4-methylmorpholin are shaken for 80 min. in 0.7 ml DMF. After several washings with isopropanol and DMF, the solid residue is treated with 4.5 ml TFA/0.25 ml water/0.25 ml ethanedithiol for 55 min. This mixture is then filtered and by addition of diethyl ether the title compound precipitated from the remaining solution. The title compound is purified by prep. HPLC (Vydac column, gradient A = water, B = 7 acetonitril/3 water/0.2 phosphoric acid).

MS (Ion-spray): mass 4272

$[\alpha]_{D}^{25} = -66,8^{\circ}$ (c=0.25 in 95% AcOH)

EXAMPLE 301: 4-Fluoro-2-isopropyl-5-tert.butoxycarbonylamino-hex-3-enoate

a) (S)-3-(1-Oxo(4-fluoro-3-hydroxy-2-isopropyl)hex-4-enyl)-4-phenylmethyl-2-oxazolidinone

The title compound is prepared analogous to the procedure described by D.A. Evans et al., Org. Synth. 68, 1990; starting from (S)-3-(1-oxoisobutyl)-4-phenylmethyl-2-oxazolidinone and 2-fluoro-but-2-enal.

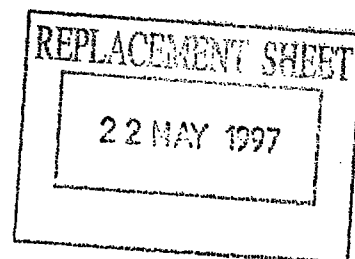
MS (FAB): MH⁺ 350

b) (S)-3-(1-Oxo(4-fluoro-3-O-(2,2,2trichlorethanimino)-2-isopropyl)hex-4-enyl)-4-phenylmethyl-2-oxazolidinone

1.5 g of the compound of Example 301a) are dissolved in 6 ml DCM. At 0° 0.0958 ml 1,8-diazabicyclo(5,4,0)undec-7-en (DBU) are added. To this solution 0.475 ml trichloroacetonitril in 2 ml DCM is added dropwise at 0°. After 1 hour the reaction mixture is

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evaporated and the residue is purified by column chromatography on silica gel (hexan/diethyl ether 2:1).

MS (FAB): MH⁺ 493

c) (S)-3-(1-Oxo(4-fluoro-5-N-2,2,2-trichloroacetyl-amino-2-isopropyl)hex-3-enyl)-4-phenylmethyl-2-oxazolidinone

2 g of the compound of 301b) is dissolved in 150 ml o-xylene and stirred under reflux at 160° for 3 hours. Then o-xylene is removed and the residue is purified by column chromatography on silica gel (toluol/ethyl acetate 98:2).

MS (FAB): MH⁺ 493

d) 4-Fluoro-5-N-2,2,2-trichloroacetyl-amino-2-isopropyl-hex-3-enoic acid

The title compound is prepared analogous to the procedure described by D.A. Evans et al. Org. Synth. 68 1990, but using as starting material the compound of 301c).

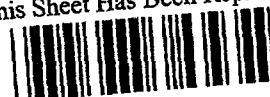
MS (FAB): MH⁺ 335

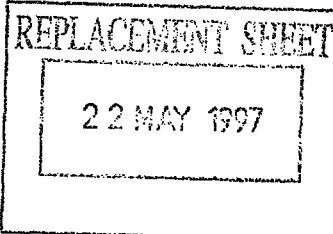
e) 5-Amino-4-fluoro-2-isopropyl-hex-3-enoic acid

899 mg of the compound of 301d) are dissolved in 20 ml ethanol and 13.5 ml 6N sodium hydroxide solution are added. This mixture is stirred for 20 hours at r.t. Then ethanol is removed and the remaining aqueous fraction is acidified with 2N HCl to pH=2 and extracted with n-butanol. The extract is evaporated and the residue is filtered through silica gel (DCM/methanol 1:1), thus yielding pure title compound.

MS (FAB): MH⁺ 190

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f) 4-Fluoro-2-isopropyl-5-tert.butoxycarbonylamino-hex-3-enoic acid

0.56 g of the compound of 301e) are dissolved in 12 ml water and 1.5 g sodium carbonate are added. To this solution 0.4 g di-tert.butylcarbonate dissolved in 12 ml tetrahydrofuran are added. This mixture is stirred at room temperature for 18 hours. Then 50 ml n-butanol and 50 ml water are added and under vigorous stirring 1N HCl is added until pH=2. The organic fraction is evaporated and the residue purified by column chromatography on silica gel (hexan/acetone 1:2).

MS (FAB): MH^+ 290

$[\alpha]_D = -61,3^\circ$ (1% in methanol)

By repeating the above procedures but using the appropriate starting materials the following compounds may be prepared:

4-Fluoro-2-methyl-5-tert.butoxycarbonylamino-hex-3-enoic acid
4-Chloro-2-methyl-5-tert.butoxycarbonylamino-hex-3-enoic acid
4-Fluoro-2-benzyl-5-tert.butoxycarbonylamino-hex-3-enoic acid
4-Chloro-2-isopropyl-5-tert.butoxycarbonylamino-hex-3-enoic acid
4-Methyl-2-isopropyl-5-tert.butoxycarbonylamino-hex-3-enoic acid

EXAMPLE 302:

By following the procedure of Example 300 above but using the appropriate starting materials, the following compounds may be prepared:

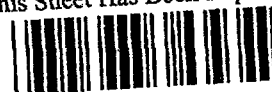
- (5-Amino-4-fluoro-2-methyl)-hex-3-enoyl-hPTH(3-36)-amide

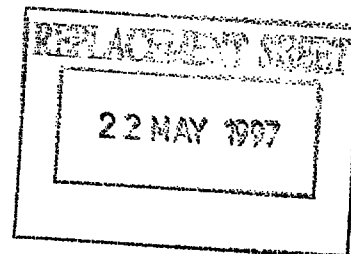
MS (Ion-Spray): Mass 4245

- (5-Amino-4-chloro-2-methyl)-hex-3-enoyl-hPTH(3-36)-amide

MS (Ion-Spray): Mass 4260

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- (5-Amino-4-fluoro-2-benzyl)-hex-3-enoyl-hPTH(3-36)-amide
MS (Ion-Spray): Mass 4320
- (5-Amino-4-chloro-2-isopropyl)-hex-3-enoyl-hPTH(3-36)-amide
MS (Ion-Spray): Mass 4289
- (5-Amino-4-methyl-2-isopropyl)-hex-3-enoyl-hPTH(3-36)-amide
MS (Ion-Spray): Mass 4269

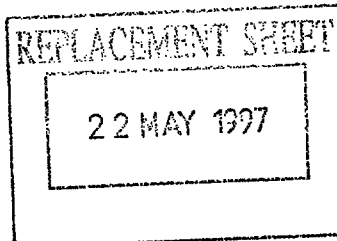
EXAMPLE 303:

By following the procedure of Example 300 above but using the appropriate starting materials, the following compounds may be prepared:

- a) (5-amino-3-aza-2-isopropyl)-hexanoyl-hPTH(3-36)-amide [MS (Ion-Spray): 4257] using as starting material:
(4-aza-2-t.-butoxycarbonylamino-5-isopropyl)-hexanoic acid
- b) (5-amino-3-aza-3-N-acetyl-2-isopropyl)-hexanoyl-hPTH(3-36)-amide [MS (Ion-Spray): 4299] using as starting material:
(4-aza-4-N-acetyl-2-t.-butoxycarbonylamino-5-isopropyl)-hexanoic acid
- c) (5-amino-3-aza-3-N-acetyl)-hexanoyl-hPTH(3-36)-amide [MS (Ion-Spray): 4257] using as starting material: (4-aza-4-N-acetyl-2-t.-butoxycarbonylamino)-hexanoic acid
- d) (5-methylamino-3-aza-2-isopropyl)-hexanoyl-hPTH(3-36)-amide [MS (Ion-Spray): 4271] using as starting material:
[4-aza-2-(N-t.-butoxycarbonyl,N-methylamino)-5-isopropyl]-hexanoic acid

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- e) (5-methylamino-3-aza-3-N-methyl-2-isopropyl)-hexanoyl-hPTH(3-36)-amide [MS (Ion-Spray): 4285] using as starting material: [4-aza-4-N-methyl-2-(N-t.-butoxycarbonylamino,N-methyl-amino)-5-isopropyl]-hexanoic acid
- f) (5-amino-3-aza-3-N-methyl-2-isopropyl)-hexanoyl-hPTH(3-36)-amide [MS (Ion-Spray): 4271] using as starting material: (4-aza-4-N-methyl-2-t.-butoxycarbonylamino-5-isopropyl)-hexanoic acid
- g) (5-amino-3-aza-3-N-isopropyl)-hexanoyl-hPTH(3-36)-amide [MS (Ion-Spray): 4257] using as starting material: (4-aza-4-N-isopropyl-2-t.-butoxycarbonylamino-5-isopropyl)-hexanoic acid
- h) [1-cyclopentane-1-amino-1-carboxylic acid(psi CH₂-NH)-Val²]-hPTH(1-36)-amide using 1-cyclopentane-1-t.-butoxycarbonyl-amino-1-carboxylic acid (psi CH₂-NH)-Val-OH as starting material.

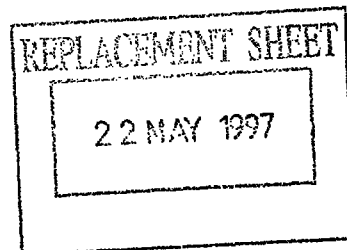
EXAMPLE 304:

By following the procedure of Example 3a) above but using the appropriate starting material, the following compound may be prepared:

(5-amino-3-aza-2-isopropyl)-hexanoyl-[Leu⁸,Ala¹⁶,Gln¹⁸,Ala¹⁹]hPTH(3-36)OH [MS (Ion-Spray): 4135] using as starting material: (4-aza-2-t.-butoxycarbonylamino)-hexanoic acid

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EXAMPLE 305: (4-Aza-4-N-acetyl-2-tert.butoxycarbonylamino-5-isopropyl)-hexanoic acid

a) Methyl (4-aza-2-tert.butoxycarbonylamino-5-isopropyl)-hexanoic acid

Boc-Ala-Val-OMe is treated with the Lawesson-reagent (S.-O. Lawesson, et al. Tetrahedron 1981, 37, 3635). The resulting endothiopeptide is then reduced in accordance with the procedure described by F.S. Guziec et. al. THL, 1990, 23-26.

MS (EI): MH^+ 289

b) Methyl (4-aza-4-N-acetyl-2-tert.butoxycarbonylamino-5-isopropyl) hexanoate

264 mg of the compound of 305a) is dissolved in 5 ml DCM and then 0.14 ml triethylamin and 0.072 ml acetyl chloride are added. This mixture is stirred for 24 hours at 40°. After addition of 50 ml DCM, the solution is washed twice with water. The organic fraction is dried and evaporated. The title compound thus obtained is purified by column chromatography (hexan/ethyl acetate 4:1)

MS (EI): M^+ 330

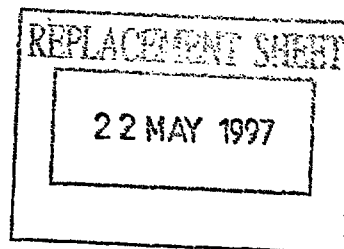
c) (4-Aza-4-N-acetyl-2-tert.butoxycarbonylamino-5-isopropyl)-hexanoic acid

356 mg of the compound of 305b) are dissolved in 4 ml methanol/-water 3:1 and 53 mg lithium hydroxide are added. This mixture is stirred at 60° for two hours. Then 25 ml DCM are added and 1N HCl is added under vigorous stirring until pH=2. The organic fraction is separated, dried and evaporated, thus yielding the title compound in pure form.

MS (FAB): MH^+ 317

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By repeating the procedure but using the appropriate starting materials, following compounds may be obtained:

(4-Aza-4-N-acetyl-2-tert.butoxycarbonylamino)-hexanoic acid
(4-Aza-2-tert.butoxycarbonylamino-5-isopropyl)-hexanoic acid
[4-aza-2-(N-t.-butoxycarbonyl,N-methylamino)-5-isopropyl]-
hexanoic acid
(4-aza-4-N-methyl-2-t.-butoxycarbonylamino-5-isopropyl)-hexanoic
acid
(4-aza-4-N-isopropyl-2-t.-butoxycarbonylamino-5-isopropyl)-
hexanoic acid
(4-aza-2-t.-butoxycarbonylamino)-hexanoic acid

The compounds of Examples 1 to 300 and 302 to 304 show correct amino acid ratios in quantitative amino acid analysis.

In the following examples illustrating the recombinant process of the invention, if not otherwise specified, the standard procedures described in Ausubel et al; 1991; Current Protocols in Molecular Biology, John Wiley & Sons, New York are used.

EXAMPLE 306: Cloning bacteriophage T4 gene 55

Two DNA fragments, one containing the entire coding sequence of bacteriophage T4 gene 55, and the other fragment encoding part of it are amplified by polymerase chain reaction (PCR) using purified T4 DNA as the template. Oligonucleotides which contain NdeI or BamHI endonuclease recognition sequences are used as primers. The PCR reactions (25 cycles, each 30" 94 °C, 30" 50 °C, 30" 72 °C) are performed with 1 ng of template and 100 pM of the upstream primer (GAGGTGCATA TGTCAGAAAC TAAGCCT 27), the NdeI recognition site being provided by nucleotides 7-12 thereof) and

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100 pM of the downstream primer

(GGTCGGATCC ATCGTTAGCG TTAGCCTCAT ATAAAAAATC 40 the BamHI, recognition site being provided by nucleotides 5-10 thereof) in 100 µl reaction buffer containing 10 mM Tris-HCl, 50 mM KCl, 1.5 mM MgCl₂, 0.2 mM of each of dATP, dCTP, dGTP, dTTP, and 0.1% Triton X 100, pH 9.0. A 0.6kb fragment is obtained.

The truncated form of gene 55 is prepared by PCR as described for the entire gene, except that a different oligonucleotide is used as downstream primer (GGTCGGATCC TACGTTGGAC GAATGCAT 28, the BamHI recognition site being provided by nucleotides 5-10 thereof). A 0.35 kb fragment is obtained.

The 0.6 kb and 0.35 kb DNA fragments are digested with restriction endonucleases NdeI/BamHI and are cloned into a NdeI/BamHI digest of expression vector pET17B (obtained from Novagen). The 0.6 kb DNA fragment encodes the entire gp55 protein (188 amino acid residues - see Fig. 1), whereas the 0.35 kb fragment encodes a 12 kD amino-terminal fragment of gp55 (112 amino acid residues - see Fig. 2).

Example 307: Preparation and cloning of the DNA encoding PTH (1-38)

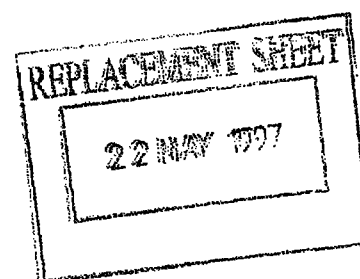
DNA fragments encoding a) the linker peptide corresponding to amino acid residues 110-115 of Fig. 1 and PTH (1-38) or b) the linker peptide corresponding to amino acid residues 186-191 of Fig. 2 and PTH (1-38) are generated by PCR using a cloned synthetic PTH (1-38) coding sequence as a template. The oligonucleotides encoding the linker a)

(GAGTGGATCC ATCGATCCAC CATCCGTATC AGAAATACAA CT 42), or encoding linker b

GAGTGGATCC GTTAACGGTC CATCCGTATC AGAAATACAA CT 42 are used as upstream primers. The downstream primer TCTACTCGAG TTAACCCAGA GCTACAAAAT TATG 34

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The upstream primers each incorporate a BamHI cloning site and the downstream primer contains a XhoI recognition site (nucleotides 5-10 of each sequence). The PCR reaction is performed under the same conditions as described in Example 306. After cleavage with BamHI and XhoI, the 0.14 kb PCR products are inserted into BamHI-XhoI digests of the gp55-pET17B plasmids, obtained in Example 306.

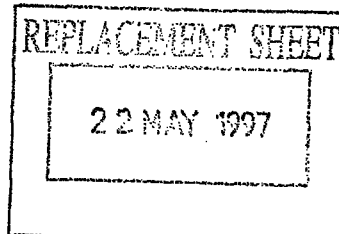
For expression of the fusion proteins the resultant plasmid constructs are transformed into E.coli BL21 (DE3) Lyse and single bacterial colonies are further propagated. The expression of the fusion proteins is obtained by incubating a preculture at 37°C on a rotary shaker overnight in circle growth medium (Bio 101). A main culture is inoculated with the preculture making up 1 to 5 % of the volume of the main culture. The main culture is then fermented at a temperature of 37°C and at a pH of 6.9 to 7.1 whilst being aerated at 10 l/min and agitated at 200-400 rpm. Expression is induced by the addition of 1 mM IPTG as soon as the culture reaches an OD₅₅₀ of about 1.0 to 5.0. After induction the fermentation is continued for five hours and then the fermentation broth is harvested without killing the E. coli cells. The harvested cells are spun down with a tubular centrifuge and resuspended in sample buffer and assayed for expression by SDS PAGE. The cell pellet is then frozen until purification. The procedures and host cells used are fully described in Studier et al; 1990; "Use of T7 RNA Polymerase to direct expression of Cloned Genes", Methods in Enzymology, Academic Press, pages 60 to 89.

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EXAMPLE 308: Preparation of inclusion bodies

Frozen E. coli pellets obtained from Example 307 are resuspended to 25% w/v in Buffer A (50mM Tris pH 8.0; containing 2mM DTT, 5mM Benzamidinium-HCl, 1.5mM MgCl₂, 1.0mM MnCl₂, 10µg/ml DNase 1 and



2mg/ml Lysozyme) and mixed by stirring for 1 hour at room temperature. The resuspended cells are lysed by passage through a Manton-Gaulin homogeniser (2 passes at 1200 bar) and the resultant lysate kept on ice. The lysate is diluted 2 fold with ice-cold buffer B (50mM Tris pH 8.0; containing 2mM DTT, 5mM Benzamidine-HCl and 4mM EDTA) and centrifuged for 30min at 27,500g.

The supernatant from the centrifuge is carefully removed and discarded. The pellet is resuspended in buffer B to 25% w/v, passed once more through the Manton-Gaulin and centrifuged as before. This process is repeated once using buffer B and once using water. The resultant inclusion body pellets are weighed and frozen at -20°C until required.

EXAMPLE 309: Solubilisation and cleavage of gp55-Asp-Pro-Pro-(1-38)hPTH inclusion bodies

Frozen inclusion bodies containing gp55-Asp-Pro-Pro-(1-38)hPTH as obtained in Example 308 are suspended in 10 mM HCl acid solution (analytical grade) to a final concentration of 6%w/v. The solution is incubated for 24 hours at 50°C and the reaction terminated by addition of 1 volume of aqueous 100 mM NaOAc solution. The diluted supernatant is clarified by centrifugation at 27,500g for 30 minutes and filtered through a glass fibre filter.

EXAMPLE 310: Solubilisation and cleavage of gp55-Asn-Gly-Pro-(1-38)hPTH inclusion bodies

Frozen inclusion bodies containing gp55-Asn-Gly-Pro-(1-38)hPTH and obtained from example 308 are dissolved in 6M Guanidine-HCl (containing 2M Hydroxylamine-HCl and brought to pH 9.0 by addition of 4.5M LiOH) to give an estimated 5mg/ml protein. The

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pH is readjusted to pH 9 by further addition of LiOH and the solution incubated for 4 hours at 45°C. The reaction is terminated by addition of formic acid to pH 4 and the solution centrifuged and filtered as described in example 309.

EXAMPLE 311: Preparative HPLC of Gly-Pro and Pro-Pro (1-38)hPTH

Filtered cleavage supernatants are analysed on analytical reversed phase HPLC (Orpogen HD-Gel-RP-7s-300; 4 x 150mm column) and compared with a known (1-38)hPTH standard (Bachem) to determine the PTH concentration. The column is equilibrated with 85% HPLC buffer A (90% water, 10% acetonitrile; containing 0.1%v/v TFA) and 15% HPLC buffer B (10% water, 90% acetonitrile; containing 0.1% TFA) and is eluted with a gradient of 15% HPLC buffer B to 100% HPLC buffer B over 15 minutes at a flow rate of 0.75ml/min.

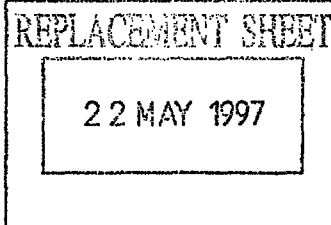
Depending upon the scale of the preparation, supernatants are either loaded on a 1.0 x 25cm C4 Vydac or a 2.2 x 25cm C4 Vydac. The columns are equilibrated with 85% HPLC buffer A and 15% HPLC buffer B at flow rates of 4ml/min and 10ml/min respectively. The columns are eluted with gradients of 15%B to 85%B over 80ml and 15%B to 55%B over 300ml respectively.

Gly-Pro-hPTH(1-38) or Pro-Pro-hPTH(1-38) peaks are collected manually and the acetonitrile is later removed under vacuum.

Analytical HPLC is used to determine the peptide concentration both after cleavage and in the collected fractions.

The solvent free peptide solutions are diluted with an equal volume of 100mM sodium acetate and the pH adjusted, if necessary, to pH 5.4. Following filtration (0.2µm), the solution is loaded on a cation exchange column (either Mono S or SP-Sepharose High Performance, packed into HR10/10 or XK16/10 columns respectively) equilibrated with 50mM sodium acetate pH 5.4 (buffer C). The column is eluted with a 0 to 500mM NaCl gradient in buffer C over 7.5 column volumes. 10ml fractions are collected and the





X-X-(1-38)hPTH peak is pooled. The peptide concentration is determined as before using HPLC.

This column serves both to exchange the peptides into a more physiological buffer and to remove additional peptides produced by chemical cleavage of the fusion protein itself.

EXAMPLE 312: Enzymatic removal of X-X from X-X-(1-38)hPTH

Pooled peptide fractions, as obtained in Example 311, (concentration range 1.5 - 2.0 mg/ml) are brought to pH 8.0 by addition of solid Tris. Purified X-Prolyl dipeptidase (0.5mg/ml in PBS pH 7.2) (Nardi et al. (1991) App. Env. Microbiol. 57, No.1, 45-50) is added to 1:1000 and the solution is incubated for 24h at 37 C. The reaction is stopped by addition of TFA to 0.2% (pH = 5.0) and the resultant (1-38)hPTH is isolated using preparative HPLC (using the same conditions as described in Example 311). The solvent is removed and the product is lyophilised after removal of aliquots for N-terminal sequence and mass spectroscopic analyses.

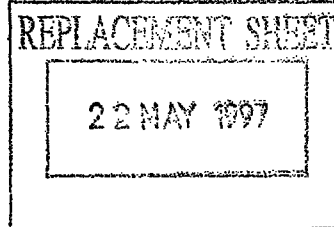
Examples of the yields which are obtained using the two fusion protein approaches described above are as follows:

Gen55-Asp-Pro-Pro-(1-38)PTH (using the "truncated" form of Gen55)

Under partially optimized fermentation conditions, a 215 g wet cell pellet is obtained from a 10 litre fermentation. The percentage level of expression of the fusion protein is approximately 37%. From this pellet, 31 g of inclusion bodies are isolated. The protein purity (with respect to the fusion protein) of these bodies is 61%. After solubilisation, cleavage and centrifugation/filtration approximately 620mg Pro-Pro-PTH are detected. This material yields 580mg Pro-Pro-PTH after subsequent C4-Vydac HPLC. After dilution, cation-exchange on Mono S,

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digestion with X-Prolydipeptidase (at a Pro-Pro-PTH concentration of 2mg/ml), HPLC and lyophilisation, 440mg of lyophilized pure product with full biological activity is obtained. This represents a yield of 71% between the stopped cleavage step and the final product.

Gen55-Asn-Gly-Pro-(1-38)PTH (using the "long" form of Gen55)

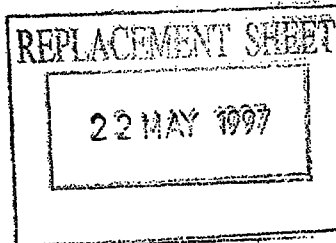
From a non-optimized 20 litre fermentation a 78g wet cell pellet with 50% expression of the fusion protein is harvested. Following lysis and centrifugation, 18g of "90% pure" inclusion bodies are obtained. Following the acid stop step and centrifugation/filtration about 470mg Gly-Pro-PTH are detected. After HPLC on C4-Vydac, 355mg of peptide are detected, decreasing to 321mg after SP-Sepharose cation-exchange. Enzymatic removal of the Gly and Pro residues is carried out at 2mg/ml. After the final HPLC step and lyophilization, 244mg of pure, fully biologically active (1-38)PTH is recovered representing a yield of 52% from the stopped cleavage reaction stage.

Compounds of Examples 25, 26, 56 and 86 to 164 are also prepared using the process of Examples 306 to 312.

The compounds of the invention in free form or in the form of pharmaceutically acceptable salts and complexes exhibit valuable pharmacological properties as indicated in animal tests and are therefore indicated for therapy.

The biological activity of the compounds of the invention is assessed in vitro by measuring their ability of stimulating the synthesis of cyclic AMP in UMR 106-06 rat and SaOS-2 human osteosarcoma cells according to the method of Marcus and Aurbach in Endocrinology, 85, 801-810 (1969). Rat osteosarcoma UMR 106 cells are grown to confluence in Eagle's Minimum Essential Medium - 10% FCS in 12 well plates, human SaOS-2 osteosarcoma cells are





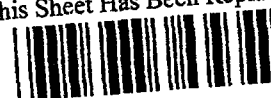
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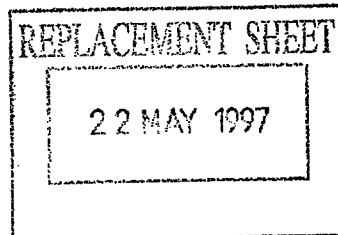
grown in McCoy's 5A medium - 10% FCS. The medium is then changed to medium with 2% FCS and 1-5 $\mu\text{Ci}/\text{well}$ $[3\text{H}]\text{-adenine}$ is added. Two hours later, cells are washed twice with serum-free medium and incubated in DMEM - 1% BSA containing 1 mM 3-isobutyl-1-methyl-xanthine to exclude actions on phosphodiesterases. Test substances are added 20 min later. The reaction is stopped and cAMP extracted after 15 min by adding ice cold 5% trichloroacetic acid. A carrier solution (0.5 ml/well) containing 5 mM of unlabelled adenine, adenosine, AMP, ADP, ATP, and cAMP as well as 0.4 μCi $[14\text{C}]\text{-adenosine}$ for determination of recovery is added. $[3\text{H}]\text{-cAMP}$ is separated using serial Dowex 50W-X4 (200-400 mesh) and alumina chromatography and counted. Results are calculated in % of solvent control and EC_{50} values determined from DRC curves. Compounds of the invention stimulate cAMP production in UMR 106-06 rat and SaOS-2 human osteosarcoma cells at a concentration of 10^{-11} to 10^{-6} M.

The compounds of the invention also have binding affinity to PTH receptors, e.g. as follows:

Chicken $[\text{Tyr}^{36}]\text{PTHrP}(1-36)\text{amide}$ is iodinated to a specific activity of 2,200 Ci/mmol using the lactoperoxidase method (Anawa Lab. AG, Wangen). Monolayers of opossum kidney cells are washed with 200 μl DMEM and HAM's F12 (1:1) containing 1% BSA and incubated at 16°C with 50,000cpm of $[^{125}\text{I-Tyr}^{36}]\text{chPTHrP}(1-36)\text{-amide}$ per well in the presence or absence of 1 μM $[\text{Tyr}^{36}]\text{chPTHrP}(1-36)\text{amide}$. After incubation, cells are washed with 0.5 ml medium (4°C), lysed with 0.5 ml 1N NaOH and radioactivity is determined. Specific binding is defined as total binding minus nonspecific binding. Competition curves are analyzed using SCTFIT, a non-linear regression computer program (Feyen et al, 1992, Biochem. Biophys. Res. Commun. 187:8-13) and data presented as mean pK_D values ($n=2$ to 3). Compounds of the invention show in

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this test a binding affinity expressed as a mean pK_D value of from 7 to 10.

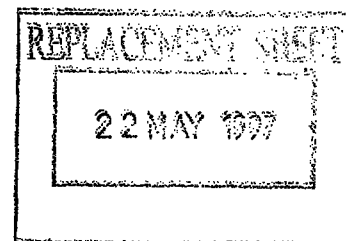
Furthermore, the compounds of the invention increase plasma calcium level after continuous s.c. infusion, e.g. as determined in male thyroparathyroidectomized Wistar rats weighing 140 to 170 g. 5 days after thyroparathyroidectomy, rats are implanted in the neck with Alzet minipumps and test compounds infused at rates of 0.3 to 30 $\mu\text{g/day/rat}$. Blood is drawn by retro-orbital puncture in the morning of days 1, 2, 3, and 6 thereafter and plasma calcium concentrations are determined photometrically. In this test compounds of the invention increase plasma calcium.

The compounds of the invention are accordingly indicated for preventing or treating all bone conditions which are associated with increased calcium depletion or resorption or in which calcium fixation in the bone is desirable, e.g. osteoporosis of various genesis (e.g. juvenile, menopausal, post-menopausal, post-traumatic, caused by old age or by cortico-steroid therapy or inactivity), fractures, osteopathy, including acute and chronic states associated with skeletal demineralisation, osteomalacia, periodontal bone loss or bone loss due to arthritis or osteoarthritis or for treating hypoparathyroidism.

The compounds of the invention are particularly indicated for preventing or treating osteoporosis of various genesis.

For all the above uses, an indicated daily dosage is in the range from about 0.003 to about 10 mg preferably 0.003 to 3, more preferably 0.01 to 1 mg of a compound of the invention. Compounds of the invention may be administered once a day or up to twice a week.





The compounds of the invention may be administered in free form or in pharmaceutically acceptable salt form or complexes. Such salts and complexes may be prepared in conventional manner and exhibit the same order of activity as the free compounds. The present invention also provides a pharmaceutical composition comprising a compound of the invention in free base form or in pharmaceutically acceptable salt form or complex form in association with a pharmaceutically acceptable diluent or carrier. Such compositions may be formulated in conventional manner. The compounds of the invention may be administered by any conventional route, for example parenterally e.g. in form of injectable solutions or suspensions, enterally, e.g. orally, for example in the form of tablets or capsules or in a transdermal, nasal or a suppository form.

In accordance with the foregoing the present invention further provides:

- a) a compound of the invention or a pharmaceutically acceptable salt or complex thereof for use as a pharmaceutical;
- b) a method for improving bone formation, e.g. preventing or treating all bone conditions which are associated with increased calcium depletion or resorption or in which calcium fixation in the bone is desirable, e.g. osteoporosis of various genesis (e.g. juvenile, menopausal, post-menopausal, post-traumatic, caused by old age or by cortico-steroid therapy or inactivity), fractures, osteopathy, including acute and chronic states associated with skeletal demineralisation, osteo-malacia, periodontal bone loss or bone loss due to arthritis or osteoarthritis, or for treating hypoparathyroidism in a subject in need of such treatment, which method comprises administering to said subject an effective amount

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of a compound of the invention or a pharmaceutically acceptable salt or complex thereof;

- c) a compound of the invention or a pharmaceutically acceptable salt or complex thereof for use in the preparation of a pharmaceutical composition for use in the method as in b) above.

According to a further embodiment of the invention, the compounds of the invention may be employed as adjunct or adjuvant to other therapy, e.g. a therapy using a bone resorption inhibitor, for example as in osteoporosis therapy, in particular a therapy employing calcium, a calcitonin or an analogue or derivative thereof, e.g. salmon, eel or human calcitonin, a steroid hormone, e.g. an estrogen, a partial estrogen agonist or estrogen-gestagen combination, a biphosphonate, vitamin D or an analog thereof, or any combination thereof.

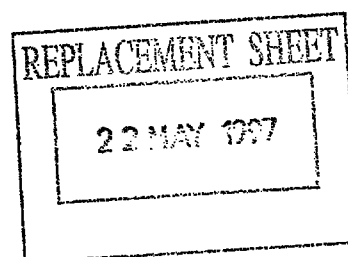
When the compounds of the invention are administered in conjunction with, e.g. as an adjuvant to bone resorption inhibition therapy, dosages for the co-administered inhibitor will of course vary depending on the type of inhibitor drug employed, e.g. whether it is a steroid or a calcitonin, on the condition to be treated, whether it is a curative or preventive therapy, on the regimen and so forth.

In accordance with the foregoing the present invention provides in a yet further aspect:

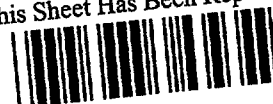
- d) a method for improving bone formation, e.g. preventing or treating calcium depletion, for example for preventing or treating any of the specific conditions or diseases hereinbefore set forth, in a subject in need of such a treatment which method comprises administering to said subject an effective amount of a) a compound of the invention and b) a

second drug substance, said second drug substance being a bone resorption inhibitor, for example as indicated above.

Compounds of Examples 20, 29, 37, 49 and 50 are preferred.



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CLAIMS

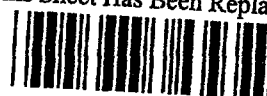
1. A PTH compound having PTH-like activity and comprising at least one modification, said modification being either
 1. at least one radical selected from a L- or D- α -amino acid, C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to the terminal amino group of the PTH compound, and/or at least one radical selected from C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to one or more side chain amino groups of the PTH compound,
 - or
 2. at least one α -amino acid unit in the positions 1 to 38 of a naturally occurring PTH sequence being replaced by a natural or unnatural amino acid unit optionally in protected form, whereby the α -amino acid units present in positions 1 and 2 at the amino terminus of the PTH sequence together may be replaced by a pseudo-peptide,
 - or a combination of such modifications,

provided that

when the PTH compound is free from D- or L- α -amino acid attached to the N-terminus or from C₂₋₆alcoxycarbonyl or optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, C₂₋₈alkynyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl, it is other than a PTH compound having a naturally occurring α -amino acid sequence;



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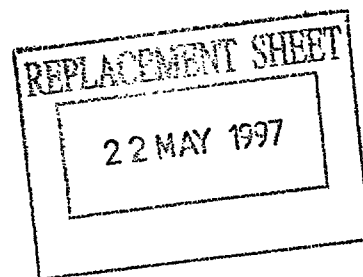
or the PTH compound is other than PTH(1-34) wherein

- i. the α -amino acid in position 1 is Gly, D-Ser, D-Ala or Tyr;
or
- ii. the α -amino acid in position 2 is Ala, D-Val, Lys, Arg or Cit and the α -amino acid in position 34 is Tyr; or the α -amino acid in position 2 is D-Val and the α -amino acid in position 34 is D-Tyr and optionally the α -amino acids in positions 8 and 18 are each Nle; or
- iii. the α -amino acid in positions 3 and/or 6 and/or 9 are replaced by a natural or unnatural amino acid; or
- iv. the α -amino acid in position 23 is replaced by Ala, Arg, Asn, Asp, Cys, Gln, Glu, Gly, His, Ile, Lys, Met, Pro, Ser or Thr; or
- v. the α -amino acid in position 25 and/or 26 and/or 27 is replaced by Ala, Asn, Asp, Cys, Gln, Glu, Gly, His, Ile, Leu, Met, Phe, Pro, Ser, Thr, Trp, Tyr or Val; or
- vi. the α -amino acids in positions 8 and 18 are each Nle or each Met(O) and optionally the α -amino acid in position 34 is Tyr; or the α -amino acids in positions 8 and 18 are each Nle and the α -amino acid in position 34 is Tyr and either the α -amino acid in position 12 is L- or D-Pro, L- or D-Ala, Aib or NMeGly or the α -amino acid in position 23 is Phe, Leu, Nle, Val, Tyr, α -Nal or β -Nal; or
- vii. the α -amino acid in position 28 is Lys and the α -amino acid in position 30 is Leu; or
- viii. the α -amino acid in position 1 is Aib; and/or the α -amino acid in position 8 and/or 18 is Leu, Ile, Val, Phe or Trp; and/or the α -amino acid in position 11 is Ser, Lys, Phe, β -Nal, Trp or Tyr; and/or the α -amino acid in position 12 is D-Leu, D-Ile, D-Nle,



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D-Val, D-Ser, D-Ser(Butyl), D-Abu, D-Thr, D-Nva, D-Met, D- β -Nal, D-Trp, D-Lys, D-Tyr, D-Lys(Fmoc), D-Phe or D-Asn; and/or the α -amino acid in position 13 is Leu; and/or the α -amino acid in position 19 and/or in position 21 is Arg, Lys, Asn or His; and/or the α -amino acid in position 23 is 2-(1,3-dithiolane-2-yl)Trp; and/or the α -amino acid in position 25 and/or in position 26 is His; and/or the α -amino acid in position 27 is Gln or Leu; or

- ix. the α -amino acid in position 8 and/or 18 is Ala or Ser; or the α -amino acid in position 8 and/or 18 is Ala, Val, Leu, Ile, Ser or Trp and the α -amino acid in position 34 is Tyr; or

the PTH compound is other than PTH(1-84) wherein

- i. the α -amino acid in position 1 is Tyr, Val, Pro, Asp or Cys; or
- ii. the α -amino acid in position 2 is Ala, Glu, Leu, Ser or Arg; or
- iii. the α -amino acid in positions 3 and/or 6 and/or 9 are replaced by a natural or unnatural amino acid; or
- iv. the α -amino acid in position 23 is replaced by Ala, Arg, Asn, Asp, Cys, Gln, Glu, Gly, His, Ile, Lys, Met, Pro, Ser or Thr; or
- v. the α -amino acid in position 25 and/or 26 and/or 27 is replaced by Ala, Asn, Asp, Cys, Gln, Glu, Gly, His, Ile, Leu, Met, Phe, Pro, Ser, Thr, Trp, Tyr or Val; or
- vi. the α -amino acid in position 8 is Met, Met(O), Ala, Val, Leu, Ile, Ser, Trp, Asn, Gln, Asp, Glu, Lys, Arg, Tyr or Gly and the α -amino acid in position 18 is Leu; or the α -amino acid in position 8 and/or 18 is Ala, Val, Leu, Ile, Ser or Trp, and optionally the α -amino acid



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in position 34 is Tyr; or the α -amino acid in position 8 and in position 18 are each Met(O); or the α -amino acid in position 8 is Leu and the α -amino acid in position 18 is Met(O); or
vii. the α -amino acid in position 26 is Gln;

the compound is other than Pro⁰PTH(1-84) or [Met⁰, Leu⁸, Leu¹⁸]PTH(1-84); or

the PTH compound is other than hPTH(1-36) wherein the α -amino acid in position 36 is Leu;

in free form or in salt form or complex form.

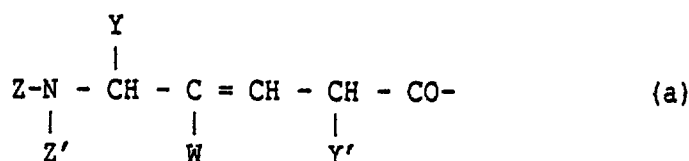
2. A PTH compound according to claim 1 comprising at least one amino acid unit replaced in one of the following positions of the PTH sequence: 1, 2, 3, 8-11, 13 to 19, 21, 22, 29 to 34.
3. A PTH compound according to claim 1 or 2 wherein the α -amino acid units in positions 1 and 2 at the amino terminus of the PTH sequence is replaced by a pseudo-di-peptide.
4. A PTH compound according to claim 3 of formula I

X - P₁

I

wherein

X is a residue of formula (a)

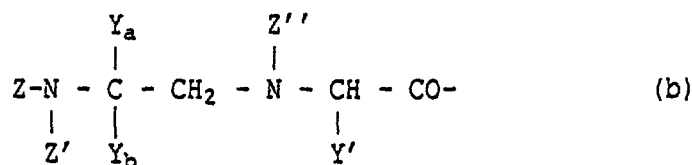
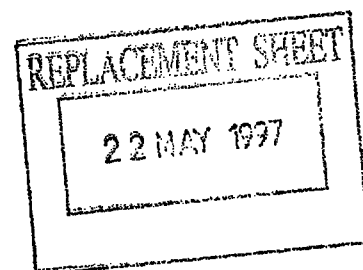


or (b)



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wherein each of Z and Z', independently, is H; optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl; or a protecting group, at most one of Z and Z' being a protecting group,

Z'' is H, C₁₋₈alkyl or a protecting group

each of Y and Y', independently, is an optionally protected side chain of a natural or unnatural α-amino acid, one of Y_a and Y_b is hydrogen and the other is an optionally protected side chain of a natural or unnatural α-amino acid or Y_a and Y_b form together with the carbon atom to which they are attached a C₃₋₆cycloalkyl group, and

W is halogen or C₁₋₄alkyl,

or W and Y' together with the moiety ---C=CH-CH-

to which they are attached, form an optionally substituted aromatic cyclic or heterocyclic residue, and

P₁ is a PTH sequence in which amino acid units in positions 1 and 2 of the N-terminus are omitted.

5. A PTH compound according to claim 1 or 2, wherein the α-amino acid unit in position 1 and/or the α-amino acid unit in position 2 of the N-terminus of the PTH sequence is replaced by an optionally protected natural or unnatural amino acid residue, provided that

when the PTH compound is free from D- or L-α-amino acid attached to the N-terminus or from C₂₋₆alkoxycarbonyl or

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optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl, it is other than a PTH compound having a naturally occurring α -amino acid sequence;

or the PTH compound is other than PTH(1-34) wherein

- i. the α -amino acid in position 1 is Aib, Gly, D-Ser, D-Ala or Tyr; or
- ii. the α -amino acid in position 2 is Ala, D-Val, Lys, Cit or Arg and the α -amino acid in position 34 is Tyr; or the α -amino acid in position 2 is D-Val and the α -amino acid in position 34 is D-Tyr and optionally the α -amino acids in positions 8 and 18 are each Nle; or
- iii. the α -amino acid in position 1 is Aib and at least one further α -amino acid unit has been replaced as follows: the α -amino acid in position 8 and/or 18 is Leu, Ile, Val, Phe or Trp, and/or the α -amino acid in position 11 is Ser, Lys, Phe, β -Nal, Trp or Tyr, and/or the α -amino acid in position 12 is D-Leu, D-Ile, D-Nle, D-Val, D-Ser, D-Ser(Butyl), D-Abu, D-Thr, D-Nva, D-Met, D- β -Nal, D-Trp, D-Lys, D-Tyr, D-Lys(Fmoc), D-Phe or D-Asn, and/or the α -amino acid in position 13 is Leu, and/or the α -amino acid in position 19 and/or in position 21 is Arg, Lys, Asn or His, and/or the α -amino acid in position 23 is 2-(1,3-dithiolane-2-yl)Trp, and/or the α -amino acid in position 25 and/or in position 26 is His, and/or the α -amino acid in position 27 is Gln or Leu,

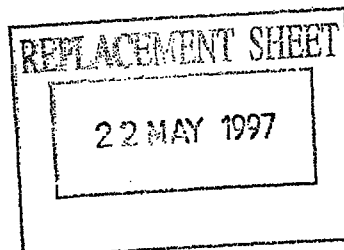
or the PTH compound is other than PTH(1-84) wherein

- i. the α -amino acid in position 1 is Tyr, Val, Pro, Asp or Cys; or

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ii. the α -amino acid in position 2 is Ala, Glu, Leu, Ser or Arg.

6. A PTH compound according to claim 5 of formula II



wherein

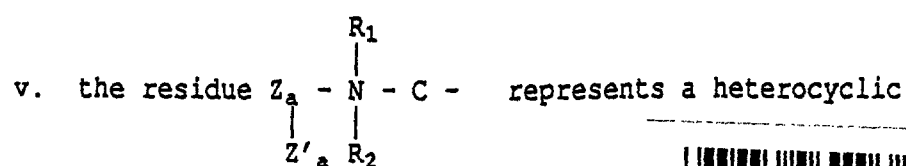
X_{1a} is a residue of an optionally protected natural α -amino acid or a residue of formula (IIa)



wherein

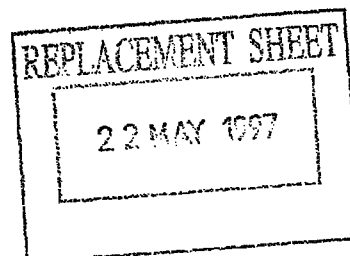
each of Z_a and Z'_a , independently, is H, C_{1-6} alkyl or a protecting group, at most one of Z_a and Z'_a being a protecting group, and either n is 1 and

- i. each of R_1 and R_2 , independently, is C_{1-6} alkyl, or
- ii. one of R_1 and R_2 is methyl or ethyl and the other is an optionally protected residue attaching to the α -carbon atom of a natural α -amino acid other than Ala, Leu, Ile or Val, or
- iii. one of R_1 and R_2 is H, methyl or ethyl and the other is an optionally protected residue attaching to the α -carbon atom of an unnatural α -amino acid, or
- iv. R_1 and R_2 form together with the carbon atom to which they are attached a C_{3-6} cycloalkyl group, or



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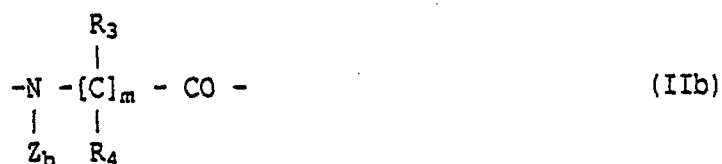


residue optionally condensed to a benzene ring,

or n is 2, 3, 4 or 5 and each of R₁ and R₂ is H or CH₃,

Y_{2a} is a residue of an optionally protected natural α-amino acid

or a residue of formula (IIb)



Z_b is H or C₁₋₆alkyl and either m is 1 and

- i. each of R₃ and R₄, independently, is C₁₋₆alkyl, or
- ii. one of R₃ and R₄ is methyl or ethyl and the other is an optionally protected residue attaching to the α-carbon atom of a natural α-amino acid other than Ala, Leu, Ile or Val, or
- iii. one of R₃ and R₄ is H, methyl or ethyl and the other is an optionally protected residue attaching to the α-carbon atom of an unnatural α-amino acid,

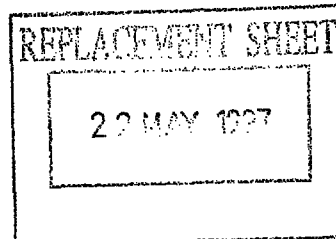
or m is 2, 3, 4 or 5 and each of R₃ and R₄ is H or CH₃, and

P₁ is as defined in claim 4.

7. A PTH compound according to any one of the preceding claims wherein the α-amino acid unit in position 8 and/or the α-amino acid unit in position 18 is replaced by an optionally protected natural or unnatural amino acid residue, provided that

when the PTH compound is free from D- or L-α-amino acid attached to the N-terminus or from C₂₋₆alkoxycarbonyl or





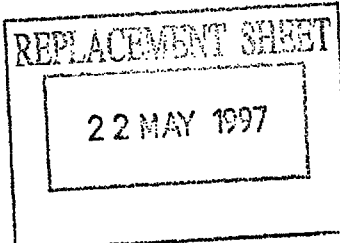
optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl, it is other than a PTH compound having a naturally occurring α -amino acid sequence;

or the PTH compound is other than PTH(1-34) wherein

- i. the α -amino acids in positions 8 and 18 are each Nle or each Met(O) and optionally the α -amino acid in position 34 is Tyr; or the α -amino acids in positions 8 and 18 are each Nle and the α -amino acid in position 34 is Tyr and either the α -amino acid in position 12 is L- or D-Pro, L- or D-Ala, Aib or NMeGly or the α -amino acid in position 23 is Phe, Leu, Nle, Val, Tyr, α -Nal or β -Nal; or the α -amino acids in positions 8 and 18 are each Nle and the α -amino acid in position 2 is D-Val and the α -amino acid in position 34 is D-Tyr; or
- ii. the α -amino acid in position 8 and/or 18 is Leu, Ile, Val, Phe or Trp and optionally at least one further α -amino acid unit has been replaced as follows: the α -amino acid in position 1 is Aib and/or the α -amino acid in position 11 is Ser, Lys, Phe, β -Nal, Trp or Tyr, and/or the α -amino acid in position 12 is D-Leu, D-Ile, D-Nle, D-Val, D-Ser, D-Ser(Butyl), D-Abu, D-Thr, D-Nva, D-Met, D- β -Nal, D-Trp, D-Lys, D-Tyr, D-Lys(Fmoc), D-Phe or D-Asn, and/or the α -amino acid in position 13 is Leu, and/or the α -amino acid in position 19 and/or in position 21 is Arg, Lys, Asn or His, and/or the α -amino acid in position 23 is 2-(1,3-dithiolane-2-yl)Trp, and/or the α -amino acid in position 25 and/or in position 26 is His, and/or the α -amino acid in position 27 is Gln or Leu; or

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iii. the α -amino acid in position 8 and/or 18 is Ala or Ser; or the α -amino acid in position 8 and/or 18 is Leu, and the α -amino acid in position 34 is Tyr; or

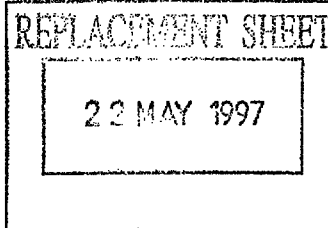
the PTH compound is other than PTH(1-84) wherein the α -amino acid in position 8 is Met, Met(O), Ala, Val, Leu, Ile, Ser, Trp, Asn, Gln, Asp, Glu, Lys, Arg, Tyr or Gly and the α -amino acid in position 18 is Leu; or the α -amino acid in position 8 and in position 18 are each Met(O); or the α -amino acid in position 8 is Leu and the α -amino acid in position 18 is Met(O); or the PTH compound is other than [Leu¹⁸, Tyr³⁴]hPTH(1-84), [Leu⁸, Tyr³⁴]hPTH(1-84), [Ile⁸, Leu¹⁸, Tyr³⁴]hPTH(1-84) or [Leu⁸, Leu¹⁸, Tyr³⁴]hPTH(1-84).

8. A PTH compound having PTH-like activity wherein at least one of the α -amino acid units in the positions 1 to 38 of a PTH sequence is replaced by the α -amino acid unit which is present at the corresponding position in PTHrP, the PTH compound optionally comprising at least one radical selected from a L- or D- α -amino acid, C₂₋₆alkoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to the terminal amino group of the PTH compound, and/or at least one radical selected from C₂₋₆alkoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to one or more side chain amino groups of the PTH compound, provided that

when the PTH compound is free from D- or L- α -amino acid attached to the N-terminus or from C₂₋₆alkoxycarbonyl or optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl,

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aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl, it is other than a PTH compound having a naturally occurring α -amino acid sequence;

or the PTH compound is other than PTH(1-34) wherein

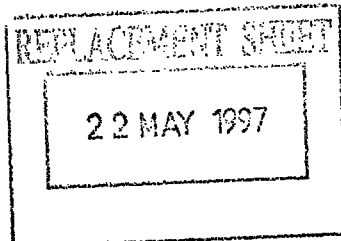
- i. the α -amino acid in position 8 and/or 18 is Leu; and/or the α -amino acid in position 11 is Lys; and/or the α -amino acid in position 19 and/or 21 is Arg; and/or the α -amino acid in position 25 and/or 26 is His; and/or the α -amino acid in position 27 is Leu; and optionally at least one further α -amino acid unit has been replaced as follows: the α -amino acid in position 1 is Aib and/or the α -amino acid in position 11 is Ser, Phe, β -Nal, Trp or Tyr, and/or the α -amino acid in position 12 is D-Leu, D-Ile, D-Nle, D-Val, D-Ser, D-Ser(Butyl), D-Abu, D-Thr, D-Nva, D-Met, D- β -Nal, D-Trp, D-Lys, D-Tyr, D-Lys(Fmoc), D-Phe or D-Asn, and/or the α -amino acid in position 13 is Leu, and/or the α -amino acid in position 19 and/or in position 21 is Lys, Asn or His, and/or the α -amino acid in position 23 is 2-(1,3-dithiolane-2-yl)Trp, and/or the α -amino acid in position 27 is Gln; or
 - ii. the α -amino acid in position 25 is Gln and/or the α -amino acid in position 26 is Asn, the α -amino acid in position 27 being optionally Leu; or
- the PTH compound is other than [Leu⁸,Leu¹⁸]PTH(1-84), in free form or in salt or complex form.

9. A PTH compound according to claim 8 wherein at least 2 α -amino acid units of the PTH sequence is replaced by the α -amino acid unit which is present at the corresponding position in PTHrP.



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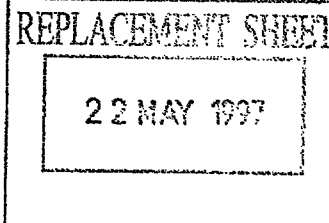


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10. A PTH compound having PTH-like activity wherein at least one of the α -amino acid units in the positions 8 to 11 of a PTH sequence is replaced by the α -amino acid unit which is present at the corresponding position in PTHrP, the PTH compound optionally comprising at least one radical selected from a L- or D- α -amino acid, C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to the terminal amino group of the PTH compound, and/or at least one radical selected from C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to one or more side chain amino groups of the PTH compound, provided that the PTH compound is other than [Leu⁸,Leu¹⁸]-PTH(1-84) or PTH(1-34) wherein the α -amino acid in position 8 is Leu and/or the α -amino acid in position 11 is Lys and optionally at least one further α -amino acid unit is replaced as follows: the α -amino acid unit in position 1 is Aib, and/or the α -amino acid in position 18 is Leu, Ile, Val, Phe or Trp, and/or the α -amino acid in position 11 is Ser, Phe, β -Nal, Trp or Tyr, and/or the α -amino acid in position 12 is D-Leu, D-Ile, D-Nle, D-Val, D-Ser, D-Ser(Butyl), D-Abu, D-Thr, D-Nva, D-Met, D- β -Nal, D-Trp, D-Lys, D-Tyr, D-Lys(Fmoc), D-Phe or D-Asn, and/or the α -amino acid in position 13 is Leu, and/or the α -amino acid in position 19 and/or in position 21 is Arg, Lys, Asn or His, and/or the α -amino acid in position 23 is 2-(1,3-dithiolane-2-yl)Trp, and/or the α -amino acid in position 25 and/or in position 26 is His, and/or the α -amino acid in position 27 is Gln or Leu, in free form or in salt or complex form.
11. A PTH compound which is [Leu⁸,Asp¹⁰]PTH or [Leu⁸,Asp¹⁰,Lys¹¹]PTH, the PTH compound optionally comprising at least one radical selected from a L- or D- α -amino acid,

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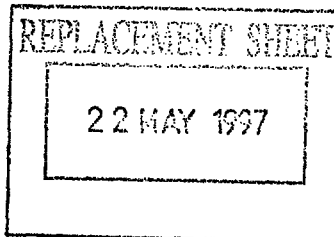
C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to the terminal amino group of the PTH compound, and/or at least one radical selected from C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to one or more side chain amino groups of the PTH compound, in free form or in salt or complex form.

12. A PTH compound having PTH-like activity wherein at least one of the α -amino acid units in the positions 16 to 19 of a PTH sequence is replaced by the α -amino acid unit which is present at the corresponding position in PTHrP, the PTH compound optionally comprising at least one radical selected from a L- or D- α -amino acid, C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to the terminal amino group of the PTH compound, and/or at least one radical selected from C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to one or more side chain amino groups of the PTH compound, provided that the PTH compound is other than [Leu⁸, Leu¹⁸]-PTH(1-84) or PTH(1-34) wherein the α -amino acid in position 18 is Leu and/or the α -amino acid in position 19 is Arg and optionally at least one further α -amino acid unit is replaced as follows: the α -amino acid unit in position 1 is Aib, and/or the α -amino acid in position 8 is Leu, Ile, Val, Phe or Trp, and/or the α -amino acid in position 11 is Ser, Lys, Phe, β -Nal, Trp or Tyr, and/or the α -amino acid in position 12 is D-Leu, D-Ile, D-Nle, D-Val, D-Ser, D-Ser(Butyl), D-Abu, D-Thr, D-Nva, D-Met, D- β -Nal, D-Trp, D-Lys, D-Tyr, D-Lys(Fmoc), D-Phe or D-Asn, and/or the α -amino acid in _____



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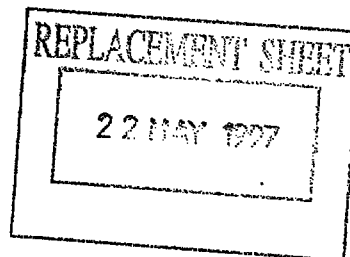
position 13 is Leu, and/or the α -amino acid in position 19 is Lys, Asn or His and/or the α -amino acid in position 21 is Arg, Lys, Asn or His, and/or the α -amino acid in position 23 is 2-(1,3-dithiolane-2-yl)Trp, and/or the α -amino acid in position 25 and/or in position 26 is His, and/or the α -amino acid in position 27 is Gln or Leu, in free form or in salt or complex form.

13. A PTH compound having PTH-like activity wherein at least one of the α -amino acid units in the positions 33 and 34 of a PTH sequence is replaced by the α -amino acid unit which is present at the corresponding position in PTHrP, the PTH compound optionally comprising at least one radical selected from a L- or D- α -amino acid, C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to the terminal amino group of the PTH compound, and/or at least one radical selected from C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to one or more side chain amino groups of the PTH compound, in free form or in salt or complex form.
14. A PTH compound according to claims 10 and 13 wherein at least 2 α -amino acid units in the positions 8-11 and 33-34 of a PTH sequence are replaced.
15. A PTH compound selected from [Thr³³,Ala³⁴]PTH, [Leu⁸,Thr³³,Ala³⁴]PTH and [Leu⁸,Asp¹⁰,Lys¹¹,Thr³³,Ala³⁴]PTH, the PTH compound optionally comprising at least one radical selected from a L- or D- α -amino acid, C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to the terminal amino group of the PTH compound, and/or at



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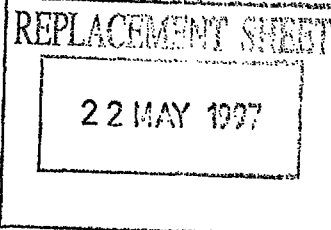
least one radical selected from C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to one or more side chain amino groups of the PTH compound, in free form or in salt or complex form.

16. A PTH compound according to any one of claims 8 to 15, wherein one or more α -amino acid units in the remaining positions 1 to 38 is further replaced by a natural or unnatural amino acid unit.
17. A PTH compound according to claim 16 wherein the α -amino acid in position 10 is selected from Gly, Gln, Glu, His, Ser, Thr, Lys and Tyr.
18. A PTH compound according to claim 16 or 17, wherein the α -amino acid in position 13 is a D- or L- α -amino acid other than Arg.
19. A PTH compound according to claim 16, 17 or 18, wherein the α -amino acid in position 16 is a D- or L- α -amino acid.
20. A PTH compound according to any one of claims 16 to 19 wherein the α -amino acid in position 18 is Gln or Tyr.
21. A PTH compound according to any one of claims 16 to 20, wherein the α -amino acid in position 19 is selected from Ala, Arg, Val, Tyr, Ser, Lys, Met, His, Gly, Pro, Asn and Ile.
22. A PTH compound according to any one of claims 16 to 21, wherein the α -amino acid in position 33 is a D- or L- α -amino acid other than Thr, Phe or Ala.



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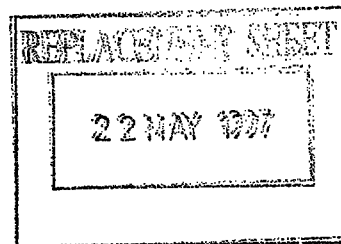


23. A PTH compound which is selected from [Leu⁸, Gln¹⁸, Thr³³, Ala³⁴]PTH, [Leu⁸, Ala¹⁶, Gln¹⁸, Thr³³, Ala³⁴]PTH, [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸, Thr³³, Ala³⁴]PTH, [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸]PTH, [Leu⁸, Ala¹⁶, Gln¹⁸, Ala¹⁹]PTH, [Leu⁸, Asp¹⁰, Lys¹¹, Gln¹⁸]PTH, [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸, Ala¹⁹]PTH, [Leu⁸, Ala¹⁶, Gln¹⁸, Ala¹⁹, Thr³³, Ala³⁴]PTH and [Leu⁸, Asp¹⁰, Lys¹¹, Gln¹⁸, Thr³³, Ala³⁴]PTH, the PTH compound optionally comprising at least one radical selected from a L- or D- α -amino acid, C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to the terminal amino group of the PTH compound, and/or at least one radical selected from C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to one or more side chain amino groups of the PTH compound, in free form or in salt or complex form.
24. A PTH compound according to any one of preceding claims which is a N-terminal PTH fragment comprising 1-34 to 1-38 amino acid units.
25. A PTH compound according to claim 24 which is a N-terminal hPTH fragment.
26. A PTH compound according to any one of claims 1 to 24 which is hPTH(1-34) or hPTH(1-36).
27. A PTH compound according to any one of the preceding claims wherein the C-terminus is -COOH, esterified -COOH, -CONH₂, mono- or disubstituted -CONH₂.



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28. A PTH compound selected from
[Leu⁸,Gln¹⁸,Thr³³,Ala³⁴]-hPTH(1-34)OH
[Leu⁸,Ala¹⁶,Gln¹⁸,Ala¹⁹,Thr³³,Ala³⁴]hPTH(1-34)OH
[Leu⁸,Ala¹⁶,Gln¹⁸,Thr³³,Ala³⁴]hPTH(1-34)OH
[Leu⁸,Asp¹⁰,Lys¹¹,Gln¹⁸]hPTH(1-36)OH
[Leu⁸,Asp¹⁰,Lys¹¹,Gln¹⁸,Thr³³,Ala³⁴]hPTH(1-34)OH
[Leu⁸,Asp¹⁰,Lys¹¹,Ala¹⁶,Gln¹⁸,Ala¹⁹]hPTH(1-36)OH
[Leu⁸,Asp¹⁰,Lys¹¹,Ala¹⁶,Gln¹⁸,Thr³³,Ala³⁴]hPTH(1-34)OH
[Leu⁸,Asp¹⁰,Lys¹¹,Ala¹⁶,Gln¹⁸]hPTH(1-36)OH, and
[Leu⁸,Ala¹⁶,Gln¹⁸,Ala¹⁹]hPTH(1-36)OH

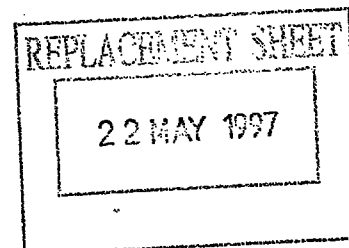
in free form or in salt or complex form.

29. A process for the production of a desired medium-sized polypeptide in which a fusion protein comprising an N-terminal bacteriophage T4 gene 55 polypeptide having the desired polypeptide linked to the C-terminal thereof, is expressed in bacterial cells.
30. A process according to claim 29 in which the fusion protein comprises a chemically cleavable linker between the gene 55 and the desired polypeptides.
31. A process according to claim 30 in which the cleavable linker comprises the amino acid sequences Asp-Pro-Pro or Asn-Gly-Pro.
32. A process according to any one of the claims 29 to 31 in which the desired polypeptide is selected from the group consisting of calcitonins, endorphins, gastric inhibitory peptide(s), glucagon, neuropeptide Y, growth hormone releasing factor (GHRF), amyloid β protein fragments, hirudin, insulin, somatostatin, epidermal growth factor,



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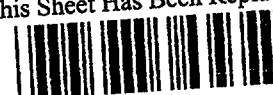
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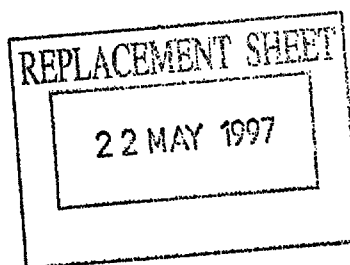
nerve growth factor, parathyroid hormone (PTH), and fragments thereof.

33. A nucleotide sequence which codes for a fusion protein comprising an N-terminal bacteriophage T4 gene 55 polypeptide and a desired medium-sized polypeptide linked to the C-terminal thereof.
34. A bacterial expression vector comprising a DNA sequence according to claim 33.
35. Bacterial host cells transformed with a DNA sequence according to claim 33, or a bacterial vector according to claim 34.
36. A process for the production of a desired medium-sized polypeptide comprising causing bacterial host cells according to claim 35 to express the fusion protein in the form of inclusion bodies; isolating the inclusion bodies; solubilising the inclusion bodies, and cleaving the desired polypeptide from the fusion protein.
37. A fusion protein comprising an N-terminal bacteriophage T4 gene 55 polypeptide and a desired medium-sized polypeptide linked to the C-terminal thereof.
38. A process for the production of a desired medium-sized polypeptide comprising cleaving the desired polypeptide from a fusion protein according to claim 37, and optionally removing unwanted amino acid residues from the N-terminus thereof.



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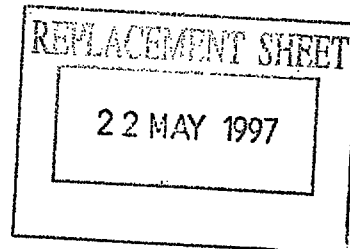


39. A process according to any one of claims 29-32, 36 or 38 in which the desired medium-sized polypeptide comprises a PTH polypeptide.
40. A nucleotide sequence according to claim 33, in which the desired medium-sized polypeptide comprises a PTH polypeptide.
41. A bacterial expression vector comprising a DNA sequence according to claim 40.
42. Bacterial host cells transformed with a DNA sequence according to claim 31 or a bacterial expression vector according to claim 41.
43. A process for the production of a medium-sized polypeptide comprising:
- culturing bacterial host cells transformed with a vector containing a DNA sequence coding for expression of a fusion protein comprising an N-terminal bacteriophage T4 polypeptide having the desired polypeptide linked to the C-terminal thereof by a chemically cleavable linker selected from the group consisting of peptides containing the amino acid sequence Asp-Pro-Pro or Asn-Gly-Pro and expressing the fusion protein in the form of inclusion bodies;
 - isolating the inclusion bodies;
 - solubilising the inclusion bodies under acid conditions;
 - cleaving the desired polypeptide from the fusion protein, and
 - removing unwanted N-terminal amino acid residues from the desired polypeptide product.

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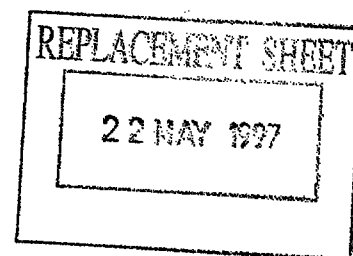
- 85 -

44. A PTH compound according to any one of claims 1 to 28 for use as a pharmaceutical.
45. A pharmaceutical composition comprising a PTH compound as defined in any one of claims 1 to 28, in free form or in physiologically acceptable salt form, together with a pharmaceutically acceptable diluent or carrier therefor.
46. A PTH compound substantially as herein before described with reference to any one of Examples 1 to 300, 302-304 and 306 to 312.

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PEPTIDES



Abstract of the invention

PTH compounds having PTH-like activity and comprising at least one modification, said modification being either

1. at least one radical selected from a L- or D- α -amino acid, C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to the terminal amino group of the PTH compound, and/or at least one radical selected from C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to one or more side chain amino groups of the PTH compound,

or

2. at least one α -amino acid unit in the positions 1 to 38 of a naturally occurring PTH sequence being replaced by a natural or unnatural amino acid unit optionally in protected form, whereby the α -amino acid units present in positions 1 and 2 at the amino terminus of the PTH sequence may be replaced by a pseudo-peptide,

or a combination of such modifications,

in free form or in salt form, have pharmacological activity, e.g. for preventing or treating all bone conditions which are associated with increased calcium depletion or resorption or in which calcium fixation in the bone is desirable.

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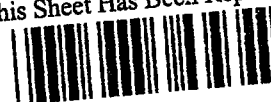
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A representative compound is e.g. of formula 1

Scr-Val-Ser-Glu-Ile-Gln-Leu-R₁-His-R₂-R₃-Gly-R₄-His-Leu-
R₅-R₆-R₇-R₈-Arg-Val-Glu-Trp-Leu-Arg-R₉-Lys-Leu-Gln-Asp-
Val-His-R₁₀-R₁₁-(R₁₂)_n-X

wherein each of R₁ to R₁₁ has a meaning within the invention, R₁₂ is Val, Val-Ala, Val-Ala-Leu or Val-Ala-Leu-Gly; n is 0 or 1; and X is OH, NH₂ or substituted OH or NH₂, provided that R₇ is Gln or Tyr, and/or R₁₀ is Thr and/or R₁₁ is Ala.

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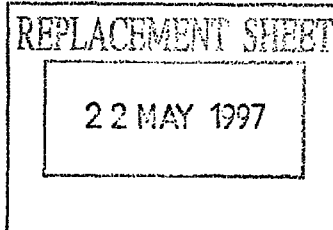
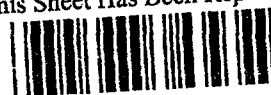
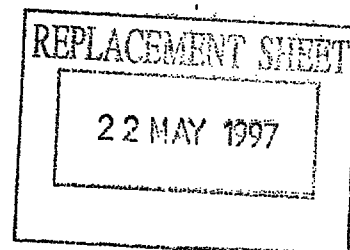


FIGURE 1

AGATCTCGAT CCCGCGAAAT TAATACGACT CACTATAGGG AGACCACAAC GGTTTCCCTC	60
TAGAAATAAT TTTGTTTAAC TTTAAGAAGG AGATATACAT ATG TCA GAA ACT AAG	115
Met Ser Glu Thr Lys	
1 5	
CCT AAA TAT AAT TAC GTA AAC AAT AAA GAG CTT TTA CAA GCT ATT ATT	163
Pro Lys Tyr Asn Tyr Val Asn Asn Lys Glu Leu Leu Gln Ala Ile Ile	
10 15 20	
GAT TGG AAA ACA GAA TTA GCA AAT AAT AAA GAC CCA AAT AAA GTA GTT	211
Asp Trp Lys Thr Glu Leu Ala Asn Asn Lys Asp Pro Asn Lys Val Val	
25 30 35	
CGT CAG AAT GAT ACT ATC GGA TTA GCC ATT ATG CTT ATT GCA GAA GGC	259
Arg Gln Asn Asp Thr Ile Gly Leu Ala Ile Met Leu Ile Ala Glu Gly	
40 45 50	
TTA TCT AAA CGT TTC AAC TTT TCA GGA TAC ACC CAG TCT TGG AAA CAA	307
Leu Ser Lys Arg Phe Asn Phe Ser Gly Tyr Thr Gln Ser Trp Lys Gln	
55 60 65	
GAA ATG ATT GCA GAT GGT ATA GAA GCT TCT ATT AAG GGG CTT CAC AAT	355
Glu Met Ile Ala Asp Gly Ile Glu Ala Ser Ile Lys Gly Leu His Asn	
70 75 80 85	
TTT GAT GAA ACG AAA TAT AAA AAC CCA CAT GCG TAT ATA ACT CAA GCT	403
Phe Asp Glu Thr Lys Tyr Lys Asn Pro His Ala Tyr Ile Thr Gln Ala	
90 95 100	

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 105 110 115

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 120 125 130

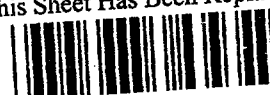
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 Val Ala Leu Gly
 150

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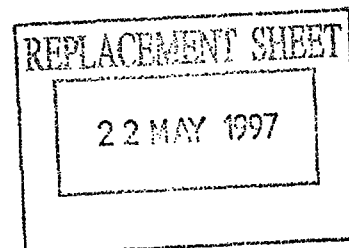
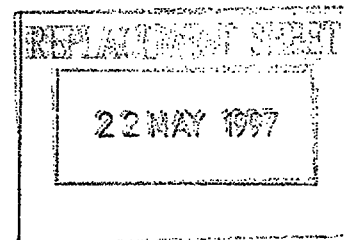


FIGURE 2

AGATCTCGAT CCCGCGAAAT TAATACGACT CACTATAGGG AGACCACAAC GGTTCCTC	60
TAGAAATAAT TTTGTTTAAC TTAAAGAAGG AGATATACAT ATG TCA GAA ACT AAG	115
Met Ser Glu Thr Lys	
1 5	
CCT AAA TAT AAT TAC GTA AAC AAT AAA GAG CTT TTA CAA GCT ATT ATT	163
Pro Lys Tyr Asn Tyr Val Asn Asn Lys Glu Leu Leu Gln Ala Ile Ile	
10 15 20	
GAT TGG AAA ACA GAA TTA GCA AAT AAT AAA GAC CCA AAT AAA GTA GTT	211
Asp Trp Lys Thr Glu Leu Ala Asn Asn Lys Asp Pro Asn Lys Val Val	
25 30 35	
CGT CAG AAT GAT ACT ATC GGA TTA GCC ATT ATG CTT ATT GCA GAA GGC	259
Arg Gln Asn Asp Thr Ile Gly Leu Ala Ile Met Leu Ile Ala Glu Gly	
40 45 50	
TTA TCT AAA CGT TTC AAC TTT TCA GGA TAC ACC CAG TCT TGG AAA CAA	307
Leu Ser Lys Arg Phe Asn Phe Ser Gly Tyr Thr Gln Ser Trp Lys Gln	
55 60 65	
GAA ATG ATT GCA GAT GGT ATA GAA GCT TCT ATT AAG GGG CTT CAC AAT	355
Glu Met Ile Ala Asp Gly Ile Glu Ala Ser Ile Lys Gly Leu His Asn	
70 75 80 85	
TTT GAT GAA ACG AAA TAT AAA AAC CCA CAT GCG TAT ATA ACT CAA GCT	403
Phe Asp Glu Thr Lys Tyr Lys Asn Pro His Ala Tyr Ile Thr Gln Ala	
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 105 110 115

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 120 125 130

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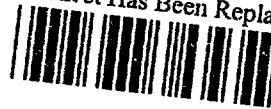
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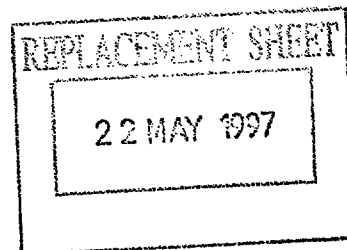
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 185 190 195

CTA ATG CAT AAT CTG GGT AAA CAT CTG AAT TCA ATG GAA CGT GTA GAA 739
 Leu Met His Asn Leu Gly Lys His Leu Asn Ser Met Glu Arg Val Glu
 200 205 210

TGG CTG CGT AAA AAA CTG CAG GAT GTA CAT AAT TTT GTA GCT CTG GGT 787
 Trp Leu Arg Lys Lys Leu Gln Asp Val His Asn Phe Val Ala Leu Gly
 215 220 225

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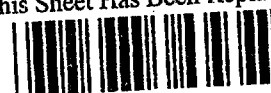
- 5 -

TAACTCGAGC AGATCCGGCT GCTAACAAAG CCCGAAAGGA AGCTGAGTTG GCTGCTGCCA 847

CCGCTGAGCA ATAAC TAGCA TAACCCCTTG GGGCCCTTGG GGCTCTAAA CGGGTCTTGA 907

GGGGTTTTTT GCTGAAAGGA GGAAC TATAT CCGGATAA 945

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REPLACEMENT SHEET

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ANALOGS OF PTH

REPLACED SHEETS		50
☒	Description : paged 1 to paged 21	
☒	Claim(s) : paged 23 to paged 25	
☐	Drawing(s) : paged to paged	
☒	Abstract : paged 4 to paged 7	
DATE OF RECEIPT		

The present invention relates to variants of parathyroid hormone (PTH), a process for their production, pharmaceutical preparations comprising them and their use as a pharmaceutical.

The term "PTH" as used herein refers to any genetically encoded form of parathyroid hormone, including the mature form containing 84 amino acids of a given vertebrate PTH species, e.g human, porcine, rat, bovine, chicken, and fragments thereof as well as analogues and derivatives thereof having PTH-like activity. The position of each amino acid involved in the PTH sequence is numbered according to the internationally accepted procedure: For consistency and as is conventional, in the following description, the same numbering system will be applied to the amino acids of the PTH sequence independently of the substitution pattern in the molecule.

More particularly, the present invention provides a PTH compound having PTH-like activity of formula 1

Ser-Val-Ser-Glu-Ile-Gln-Leu-R₁-His-R₂-R₃-Gly-R₄-His-Leu-R₅-R₆-R₇-R₈-Arg-Val-Glu-Trp-Leu-Arg-R₉-Lys-Leu-Gln-Asp-Val-His-R₁₀-R₁₁-(R₁₂)_n-X

wherein

R₁ is Met or Leu;

R₂ is Asn, Asp, Gly, Gln, Glu, His, Ser, Thr or Tyr;

R₃ is Leu or Lys;

R₄ is Lys or D- or L-Ala, -Cys, -Gln, -Ile, -Asn, -Trp, -Asp, -Val, -Thr, -Ser, -Tyr, -Met, -Leu or -Gly;

R₅ is Asn, Gln, D-Gln or D- or L-Lys, -Ser, -Leu, -Ala or -Gly;

R₆ is Ser, Asp, Ala, Glu, Gln, Phe, His, Ile or Lys;

R₇ is Gln

R₈ is Glu, Arg, Ala, Val, Tyr, Ser, Lys, Met, His, Gly, Pro, Asn or Ile;

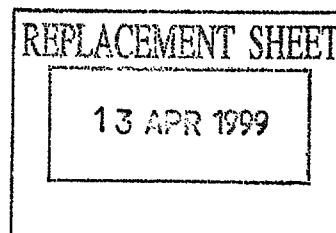
R₉ is Lys, Gln or Arg;

R₁₀ is Asn, Thr, D-Thr or D- or L-Ser, -Leu, -Gly, -Gln, -Arg, -Pro, -Asp, -Ile or -Lys;

R₁₁ is Phe or Ala;

R₁₂ is Val, Val-Ala, Val-Ala-Leu or Val-Ala-Leu-Gly;

n is 0 or 1; and



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☒	Claim(s) : paged 22 to paged 25	
☐	Drawing(s) : paged to paged	
☒	Abstract : paged 26 to paged 27	
DATE OF RECEIPT		13/4/99

X is OH, OR_a, NH₂ or NR_bR_c wherein R_a is C₁₋₄alkyl, one of R_b and R_c is H and the other is C₁₋₆alkyl, the compound optionally comprising at least one radical selected from a L- or D- α -amino acid, C₂₋₆alkoxy-carbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, arakyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to its terminal amino group, and/or

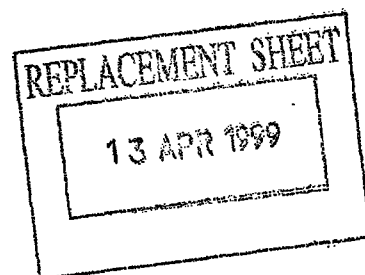
the compound optionally comprising at least one radical selected from C₂₋₆alkoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, arakyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl attached to one or more side chain amino groups of the compound, in free form or in salt or complex form.

Hereinafter, these compounds will be referred to as compounds of the invention.

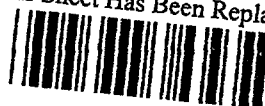
"Natural amino acids" refer to those well known in the art. They are listed and standard abbreviations are provided in the U.S.P.T.O. publication, Trademark Official Gazette, published May 15, 1990, P. 33 at 46. These amino acids and abbreviations are specifically incorporated herein by reference.

The natural amino acids are shown below:

A	Ala	alanine
D	Asp	aspartic acid
E	Glu	glutamic acid
F	Phe	phenylalanine
G	Gly	glycine
H	His	histidine
I	Ile	isoleucine
K	Lys	lysine
L	Leu	leucine
M	Met	methionine
N	Asn	asparagine
Q	Gln	glutamine
R	Arg	arginine
S	Ser	serine
T	Thr	threonine
V	Val	valine
W	Trp	tryptophane



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Y Tyr tyrosine

By amino acid in protected form is meant a natural amino acid having e.g. a side chain including an heteroatom such as O, S or N which can be protected with an O-, S- or N-protecting group. The N-terminus of the PTH compounds of the invention may also be in protected form.

N-protecting groups as may be present on the N-terminus or side chain amino groups of amino acid units include such groups as e.g. disclosed in "Protective Groups in Organic Synthesis", T. W. Greene, J. Wiley & Sons NY (1981), 219-287, for example acyl such as formyl, acetyl, trifluoroacetyl, methoxysuccinyl, hydroxysuccinyl or benzoyl optionally substituted on the phenyl ring with e.g. p-methoxycarbonyl, p-methoxy, p-nitro or p-phenylsulfonamidocarbonyl; alkoxycarbonyl such as t-butyloxycarbonyl, isobutyloxycarbonyl or methoxycarbonyl; allyloxycarbonyl; trityl; 2,2,5,7,8-pentamethyl-chroman-6-sulfonyl; arylmethoxycarbonyl such as 9-fluorenylmethoxycarbonyl or benzyloxy carbonyl optionally substituted on the phenyl ring with p-methoxy, p-nitro, o- or p-chloro, m-phenyl or 3,4-dimethyl; arylmethyl such as benzyl optionally ring substituted with p-methoxy, p-nitro or p-chloro; or arylsulfonyl such as phenylsulfonyl optionally ring substituted with p-methyl or p-methoxy, or naphthylsulfonyl optionally ring substituted with e.g. amino or di(C₁₋₄alkyl)amino.

O-protecting groups of O-containing side chains are e.g. as described in "The Peptides", 3, (1981), E. Gross and J. Meienhofer (Ed.). For aliphatic hydroxy functionalities, suitable O-protecting groups are e.g. as disclosed in the above reference in "Protection of the Hydroxy Group", J.M. Stewart, 169-201 and include the benzyl, t.-butyl and methyl groups. For aromatic hydroxy functionalities, suitable O-protecting groups include the benzyl, t.-butyl, methyl, tosyl and benzyloxycarbonyl groups.

O-protecting groups for carboxy functionalities on amino acid side chains are well known ester groups and described e.g. in "The Peptides", 3, 101-135 (11 Carboxyl Protecting Groups" by R.W. Roeske) and include the methyl, ethyl, t.-butyl and benzyl groups. S-protecting groups for thiol functionalities on amino acid side chains are known and described e.g. in "The Peptides", 3, 137-167 ("Sulfhydryl Group Protection" by R.G. Hiskey). Examples include the methyl, t.-butyl, benzyl, p-methoxyphenylmethyl, ethylamino-carbonyl and benzyloxycarbonyl groups.

According to the invention the PTH compounds may bear on their terminal amino group at least one radical selected from a L- or D- α -amino acid, C₂₋₆alkoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl, and/or on one or more side chain amino group(s) at least one radical selected from C₂₋₆alkoxycarbonyl and optionally substituted

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REPLACEMENT SHEET

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C_{1-8} alkyl, C_{2-8} alkenyl, C_{2-8} alkynyl, aralkyl, aralkenyl or C_{3-6} cycloalkyl- C_{1-4} alkyl. When such a group is attached to a side chain amino group, it is preferably on the ϵ -amino group of a Lys unit. Preferred substituents for the alkyl, alkenyl, aralkyl, aralkenyl or cycloalkylalkyl group are hydroxy, amino and for the aryl moiety also halogen and/or C_{1-4} alkoxy. The alkyl group or alkyl moiety may be linear or branched and optionally interrupted by O, S or N. Preferably any C_{1-8} alkyl, C_{2-8} alkenyl, aralkyl, aralkenyl or C_{3-6} cycloalkyl- C_{1-4} alkyl on the amino group of a PTH compound, is non-substituted. Examples for C_{1-8} alkyl are C_{1-6} alkyl, preferably methyl, ethyl, propyl, isopropyl, butyl, isobutyl, t.-butyl; for C_{2-8} alkenyl, C_{2-4} alkenyl, preferably allyl; for C_{2-8} alkynyl, C_{2-4} alkynyl, preferably prop-2-ynyl; for aralkyl, phenyl or benzyl; for aralkenyl, styryl; for C_{3-6} cycloalkyl- C_{1-4} alkyl, cyclohexyl-methyl. C_{2-6} alkoxycarbonyl is preferably formyl or acetyl. Suitable examples of D- or L- α -amino acids attached to the N-terminus include e.g. D- or L- Pro, Ala. Preferred substituents when present are alkyl, alkynyl, alcoxycarbonyl or a D- or L- α -amino acid attached to the N-terminus of the PTH compound and/or at least one alkyl or alcoxycarbonyl attached to one or more side chain amino groups.

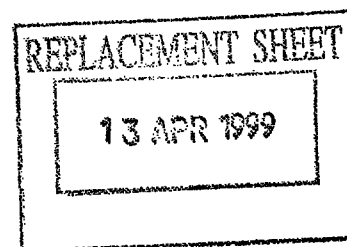
Examples of preferred compounds according to the invention are [Leu⁸, Gln¹⁸, Thr³³, Ala³⁴]PTH(1-x'), [Leu⁸, Ala¹⁶, Gln¹⁸, Thr³³, Ala³⁴]PTH(1-x'), [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸, Thr³³, Ala³⁴]PTH(1-x'), [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸]PTH(1-x'), [Leu⁸, Ala¹⁶, Gln¹⁸, Ala¹⁹]PTH(1-x'), [Leu⁸, Asp¹⁰, Lys¹¹, Gln¹⁸]PTH(1-x'), [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸, Ala¹⁹]PTH(1-x'), [Leu⁸, Ala¹⁶, Gln¹⁸, Ala¹⁹, Thr³³, Ala³⁴]PTH(1-x'), and [Leu⁸, Asp¹⁰, Lys¹¹, Gln¹⁸, Thr³³, Ala³⁴]PTH, wherein x' is an integer from 34 to 38, particularly 34, 36, 37 or 38. Especially PTH(1-x') is hPTH(1-x').

The compounds of the invention may exist e.g. in free form, salt form or in the form of complexes thereof. Acid addition salts may be formed with e.g. organic acids, polymeric acids and inorganic acids. Such acid addition salt forms include e.g. the hydrochlorides and the acetates. Complexes are e.g. formed from the compound of the invention on addition of inorganic substances, e.g. inorganic salts or hydroxides such as Ca- and Zn-salts, and/or an addition of polymeric organic substances.

The present invention also provides a process for the production of the compounds of the invention. They may be prepared in a stepwise manner either in solution or using the solid phase synthesis process or genetic engineering.

The compounds of the invention may be produced for example as follows:

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- a) removing at least one protecting group which is present in a compound of the invention in protected form; or
- b) linking together by an amide bond two peptide fragments, the peptide fragments being such that the desired amino acid sequence of the desired compound is obtained, and then effecting optionally stage a) of the process,
- c) for the production of a PTH compound comprising at least one radical selected from D- or L- α -amino acid attached to the N-terminus and C₂₋₆alkoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl attached to the terminal amino group and/or to a side chain amino group, introducing such a radical in a PTH compound in protected or unprotected form and free from such a radical, and if necessary carrying out process step a);

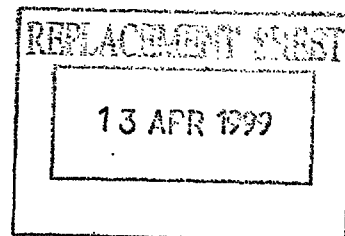
and recovering the compounds thus obtained in free form, in salt form or in complex form.

Process steps a), b) and c) may be effected in analogy with known methods, e.g. as known in the art of peptide chemistry or as described in the following examples. Where desired, in these reactions, protecting groups which are suitable for use in peptides may be used for functional groups which do not participate in the reaction. The term protecting group may also include a polymer resin having functional groups.

Process step c) may be carried out in analogy with known methods, e.g. alkylating methods. In a preferred embodiment of the invention, the alkylation is performed under reductive conditions, e.g. using a corresponding aldehyde or ketone. It may be performed for example in the presence of NaBH₃CN, preferably at an acidic pH, e.g. from 5 to 7. The temperature of the reaction may be e.g. from -20° C to 100° C. It may be advantageous to carry out the reaction in an inert solvent, e.g. water, an alcohol, dioxane or DMF or a mixture thereof. A D- or L- α -amino acid may also be attached to the N-terminus in accordance with known procedure.

The peptide fragment used as starting material in process step b) may be substituted on the N-terminus and/or on a side chain amino group by a radical selected from C₂₋₆alkoxycarbonyl, optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl or it may bear a D- or L- α -amino acid on the N-terminus.

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The compounds of the invention or fragments thereof comprise natural α -amino acid units and may also be prepared using recombinant technology.

According to a preferred embodiment of the invention there is also provided a process for the production of a compound of the invention, in which a fusion protein comprising an N-terminal bacteriophage T4 gene 55 polypeptide having the desired compound linked to the C-terminal thereof, is expressed in bacterial cells and where required, introducing a C_{2-6} alkoxy carbonyl or optionally substituted C_{1-8} alkyl, C_{2-8} alkenyl, C_{2-8} alkynyl, aralkyl, aralkenyl or C_{3-6} cycloalkyl- C_{1-4} alkyl on the terminal and/or side chain amino group of the compound in protected or unprotected form and, if required, removing the protecting group present in the protected form.

The gene 55 polypeptide may comprise the complete gp55 protein of bacteriophage T4 (188 amino acid residues) which is described in Gram, H. and Rüger, R; 1985; The EMBO Journal, 4, (1), pp 257-264. Alternatively a fragment, preferably an N-terminal fragment of the gp55 protein may be used. For example, a gp55 protein fragment comprising the first 112 N-terminal amino acid residues of the protein may be used. Typically, however, the gene 55 polypeptide comprises at least the first 25 N-terminal residues of the gp⁵⁵ protein.

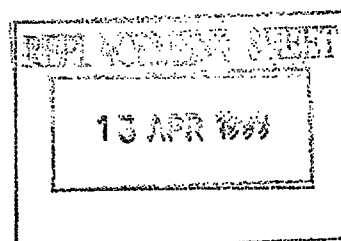
Conveniently the gene 55-PTH fusion protein also comprises a cleavable linker between the gene 55 polypeptide and the desired compound. Preferably the cleavable linker is chemically cleavable and more preferably contains the amino acid sequences Asp-Pro-Pro or Asn-Gly-Pro.

In a further aspect the invention provides a nucleotide sequence coding for a fusion protein comprising an N-terminal bacteriophage T4 gene 55 polypeptide and a desired compound of the invention linked to the C-terminal thereof.

The nucleotide sequence is preferably a DNA sequence suitable for expression in bacteria. The sequence typically contains nucleotides coding for a cleavable linker between the gene 55 and desired compound coding sequences.

The invention also provides a bacterial expression vector comprising a DNA sequence coding for the fusion protein.

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The expression vector typically contains, in addition to the fusion protein coding sequence, appropriate expression control sequences including a suitable promoter, an operator and a ribosome binding site and other appropriate regulatory sequences. Usually, also the expression vector contains one or more selectable markers.

Any suitable promoter may be used, such as the TrpE, λ P1, λ Pr, lac or Tac promoter. Preferably the promoter is the bacteriophage T7 promoter and the expression vector is a plasmid. For *E. coli* expression an RI or Col-E1 plasmid-derived vector may be used. For example, expression vector pET 17B, obtainable from Novagen, or similar expression vectors may be used.

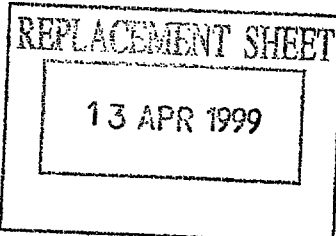
In another aspect the invention provides bacterial host cells transformed with a fusion protein coding sequence or an expression vector as described above. Any suitable bacterial host may be used, though preferably the host is *E. coli*. For example, *E. coli* strain BL21 (DE3) Lys E and similar strains may be used.

Advantageously the fusion protein accumulates within the host cells in the form of insoluble inclusion bodies, and thereby facilitates recovery of the fusion protein product from the cells. In order to recover the desired polypeptide product from the fusion protein, the protein of the inclusion bodies is solubilised, and the desired polypeptide product cleaved from the fusion protein. Any suitable solubilisation treatment may be used, including acid treatment, e.g. at about pH 2 to 4, preferably with acid at about pH 2.5, or treatment with a denaturing agent, e.g. a mild denaturing agent such as guanidine hydrochloride. In addition it may be necessary to remove one or more unwanted N-terminal amino acid residues from the product which remain after cleavage.

Thus in a further aspect the invention provides a process for the production of a desired compound of the invention, in a bacterial host, the process comprising: causing transformed bacterial host cells as defined above to express the fusion protein in the form of inclusion bodies; isolating the inclusion bodies; solubilizing the inclusion bodies, and cleaving the desired polypeptide from the fusion protein.

In the case of PTH compounds, it is not normally necessary to carry out carefully controlled denaturation and renaturation procedures to obtain PTH products in active form, e.g. having PTH-like activity. The medium-sized polypeptide products may be obtained in active form merely by neutralising the acid conditions used for solubilisation. Solubilisation and cleavage of the fusion protein may be carried out in a single step.

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The invention also includes a fusion protein comprising an N-terminal bacteriophage T4 gene 55 polypeptide and, linked to the C-terminal thereof, a desired compound of the invention.

Using the processes and vectors of the invention it has been found that it is possible to produce high levels of PTH products. In *E. coli*, we have found that up to about 50% of the total protein expressed is the desired fusion protein. Also, the fusion protein forms inclusion bodies which are easy to separate from the other material of the cells. Moreover the inclusion bodies often consist of at least about 90% of the desired fusion protein which can significantly reduce the amount of purification that is required. Further, expressing the PTH product, in the form of the fusion protein can substantially decrease endogenous processing of the polypeptide in the bacterial cell, allowing homogenous products to be obtained.

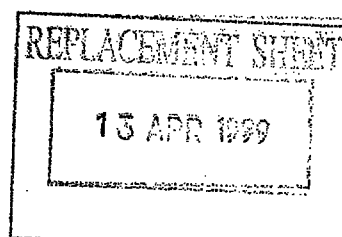
Preferably the cleavable linker contains chemically cleavable amino acids adjacent the N-terminus of the desired polypeptide, to permit the desired polypeptide to be cleaved from the fusion protein by simple chemical means. Preferably the desired compound does not contain the chemically cleavable groups.

Preferred chemically cleavable linkers contain the amino acids Asp-Pro-Pro or Asn-Gly-Pro. The Asp-Pro or Asn-Gly bonds may be chemically cleaved using acid conditions e.g. formic acid (normally about pH 2.5) or hydroxylamine respectively. The dipeptides Pro-Pro or Gly-Pro that remain on the N-terminus of the desired polypeptide may then be removed using a suitable dipeptidyl-peptidase. For example the *L. lactis* X-Pro dipeptidyl-peptidase EC 3.4.14.5 can be used. This peptidase is described in Nardi et al; 1991; Applied and Environmental Microbiology, 57, p45 to 50.

The compound of the invention may be purified following cleavage from the fusion protein. HPLC purification techniques may be used. Purification may be carried out before removal of any unwanted N-terminal amino acid residues.

The process of the invention is further illustrated in the examples 25 to 31 Fig. 1 shows the amino acid sequence and corresponding DNA sequence of a truncated gp55-Asp-Pro-ProhPTH(1-38)OH fusion protein as cloned into the plasmid pET17B, and Fig. 2 shows the amino acid sequence and corresponding DNA sequence of a full length gp55-Asn-Gly-Pro-hPTH(1-38)OH fusion protein as cloned into the plasmid pET17B.

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In the following Examples all temperatures are in 0°C. The following abbreviations are employed. In the present specification the term PTH without any further indication includes the carboxy terminus as well as a carboxamide terminus.

BOC	tert. Butyloxycarbonyl
DMF	dimethylformamide
DCM	dichloromethane
Fmoc	9-fluorenylmethoxycarbonyl
HOBt	1-hydroxybenzotriazole
TFA	trifluoroacetic acid
Trt	trityl = triphenylmethyl
RP-HPLC	reverse phase high performance liquid chromatography
r.t.	room temperature
DNP	Dinitrophenyl
Tos	Tosyl
DIPC	Diisopropylcarbodiimide
Pmc	2,2,5,7,8-Pentamethylchroman-6-sulfonyl
IP	Isopropyl
For	Formyl
Cpe	1-amino-cyclopentane-1-carboxylic acid
Cpp	1-amino-cyclopropane-1-carboxylic acid
Cha	cyclohexylalanine
tBu	t. butyl

Comparative EXAMPLE 1: N^α--Isopropyl hPTH(1-34) NH₂

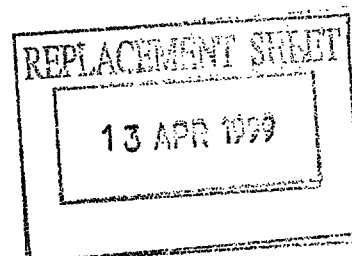
This peptide is assembled in a stepwise manner on a polystyrene based resin support. The Boc-group is used for protection of the alpha amino-group and side-chain functional groups are protected as follows: Lys(2-chlorobenzoyloxycarbonyl), Ser(benzyl), Thr(benzyl), Glu(benzyl), Asp(benzyl), Met(O), His(DNP), Arg(Tos), and Trp(HCO). Other residues are left unprotected.

Amino-4-methylphenyl-methyl-co(polystyrene-divinylbenzene = MBHA resin) (0,7 mmol/g) is subjected to the following cycle, steps (1) to (7), of treatments:

(1) DCM

(2) trifluoroacetic acid (50 %) in DCM

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- (3) DCM
- (4) diisopropylethylamine (10 %) in DMF
- (5) DMF
- (6) preformed symmetrical anhydride (1.4 mmol per g starting resin) of Boc-amino acid in DMF
- (7) DMF

Volumes of washes and reagents are from 5 to 20 ml per gram of starting resin. Each step is repeated as many times as necessary for either complete reaction of the resin (steps 2, 4, 6) or complete displacement of the previous reagent from the resin (steps 1, 3, 5, 7). Samples of resin are removed after each cycle and checked for completeness of the coupling reaction by a colorimetric test for residual amino groups using ninhydrin.

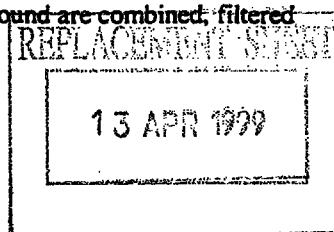
Symmetrical anhydrides of Boc-amino acids are prepared just prior to use by reacting Boc-amino acid (2.8 mmol per g resin) and DCCI (1.4 mmol per g resin) in DCM, containing DMF in amounts sufficient for complete dissolution of the Boc-amino acid. The mixture is filtered, more DMF is added to the filtrate which is concentrated by evaporation of the volatile components at a temperature not exceeding 15°C and the resulting solution is used in step (6).

The cycle of reactions, (1) to (7), is repeated for each amino acid residue such as to provide the sequence of the title compound except for Boc-Gln-OH and Boc-Arg(Tos)-OH, which are coupled in step (6) as their preformed 1-hydroxybenzo-triazole esters in DMF.

The fully assembled peptide resin is treated twice with 20 ml of thiophenol A in DMF at r.t. for 1 hour and washed with DMF, methanol and dichloromethane. The peptide resin is again subjected to steps (1) to (5) followed by the addition of 0.5 ml of acetone, 84 mg of sodium cyanoborohydride and 0.2 ml of acetic acid in 20 ml of DMF per gram of resin. After 16 hours at RT the resin is washed with DMF and dichloromethane and dried.

The peptide resin is treated with liquid HF, dimethyl sulfide, p-cresol and ethane-1,2-dithiol according to the 'low-high' procedure which is described, e.g., in International Journal of Peptide and Protein Research, vol.26, page 262-273 (1985). After removal of the volatile components and washing with ethylacetate the residue is extracted with several portions of 1%-acetic acid, filtered and the filtrate lyophilized. The lyophilized product is purified by repeated reversed-phase chromatography on a column of octadecyl-silica using a gradient of acetonitrile in 2%-phosphoric acid or in 20 mM tetramethylammonium phosphate. Fractions containing the pure compound are combined, filtered

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through a weakly basic ion-exchange resin in the acetate form and the filtrate lyophilized to give the title compound. MS (Ion-Spray): 4160.

Comparative EXAMPLE 2: [Lys (N^ε-Isopropyl)]^{26, 27}hPTH(1-34) NH₂

The peptide is assembled as described for Example 1 but using Lys(Fmoc) for incorporation in position 13 and Fmoc-Ser(tBu) in terminal position 1.

The fully assembled peptide resin is treated twice with 20 ml of thiophenol 5% in DMF at r.t. for 1 hour and washed with DMF, methanol and dichloromethane. The dry resin is treated with liquid HF as described for Example 1. The cleavage product (350 mg) is suspended in a mixture of methanol (5 ml), phosphate buffer pH 5.0 (5 ml), sodium cyanoborohydride (48 mg) and acetone (0.28 ml). After 16 hours at RT the reaction mixture is filtered and the filtrate evaporated. The residue is suspended in 20%-piperidine in DMF for 15 minutes, diluted with 20 volumes of ether and filtered. The residue is dissolved in 100 ml of water and acetic acid added to pH = 3, filtered and the filtrate lyophilized. The lyophilizate is purified by reversed-phase chromatography on a column of octadecyl-silica using a gradient of acetonitrile in 2%-phosphoric acid. Fractions containing the pure compound are combined, filtered through a weakly basic ion-exchange resin in the acetate form and the filtrate lyophilized to give the title compound. MS (Ion Spray): 4205.6.

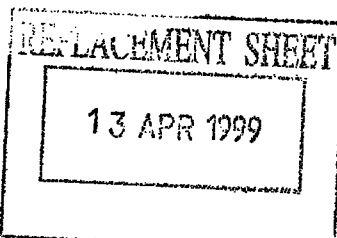
Comparative EXAMPLE 3: N^α-Isopropyl [Lys (N^ε-Isopropyl)]^{13,26, 27}hPTH-(1-38)-OH

a) Solid phase peptide synthesis

The peptide is synthesized in a stepwise manner on a polystyrene based resin support. The α-amino group is protected by Fmoc and the side-chain functional groups as followed: Asp(OtBu), Glu(OtBu), His(Trt), Lys(Boc), Asn(Trt), Gln(Trt), Arg(Pmc) and Ser(tBu). Other amino acids are left unprotected.

Fmoc-Gly esterified to 4-Hydroxymethyl-phenoxyethylco(polystyrene-1%-divinylbenzene), 0.5 mmol/g, is used as starting material for the stepwise solid phase synthesis, which consists of repetitive cycles of α-amino deprotection, washing, coupling (attachment of new amino acid) and washing. A three to five fold excess of Fmoc-amino acids is coupled as preformed HOBt-esters using DIPC. Fmoc-Arg(Pmc), Fmoc-Asn(Trt) and Fmoc-Gln(Trt) are coupled as symmetrical anhydrides using DIPC. After complete assembly of the peptide chain the Fmoc-protecting group of Ser¹ is removed by 20% piperidine/DMF. Cleavage of the peptide from the polystyrene resin and removal of

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all side chain protecting groups is obtained by treatment of the peptide resin with a mixture of 10% ethanedithiol, thioanisole, thiocresole and water in 90% TFA for three hours at room temperature. The resin particles are filtered off and washed with some TFA. The product is precipitated from the combined filtrates by addition of ether, filtered and dried. The product is purified by chromatography on a C-18 silica column using a gradient of acetonitrile in 2% H_3PO_4 /water. Fractions containing the pure compound are collected, filtered through an anion-exchange resin (Biorad, AG 4-X4, 100-200mesh acetate form) and lyophilized to give hPTH-(1-38)-OH.

b) Alkylation

40 mg (9 μmol) hPTH-(1-38)-OH (4458.2 g/mol) are dissolved in 1.2 ml methanol, 1.2 ml phosphate buffer (Merck no. 9887 adjusted to pH 5.0), 45 mg (716 nmol) NaBH_3CN (62.84 g/mol) and 0.8 ml (11 mmol) acetone (73.52 ml/mol). The reaction is monitored by RP-HPLC on a C-18 silica column and finished after 15 hours at room temperature. The reaction mixture is diluted with water and chromatographed on a C-18 silica column using a gradient of acetonitrile in 2% H_3PO_4 /water. Fractions containing the pure compound are collected, filtered through an anion-exchange resin (acetate form) and lyophilized to give the title compound as polyacetate, polyhydrate.

$$[\alpha]_D^{20} = 5.7^\circ \quad (c = 0.317 \text{ in } 95\% \text{ AcOH}).$$

MS(Ion-Spray): 4626

Comparative EXAMPLE 4: N α -Methyl [Ala¹-]PTH-(1-38)-OH

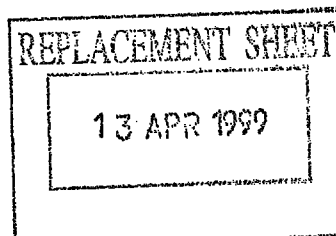
The peptide chain is assembled in the same manner as in Comparative example 3a. At position 1 instead of Fmoc-Ser(tBu)-OH the unnatural amino acid Fmoc-N α -Methyl-Ala-OH is coupled to the peptide resin. Cleavage, deprotection and purification is performed as in example 3a.

MS (Ion Spray) = 4455,91

Comparative EXAMPLE 5: [Ala¹,Ala³,Leu⁸,Gln¹³,Ala¹⁶,Gln¹⁸,Ala¹⁹,Phe²³,His²⁵,His²⁶,Leu²⁷,Thr³³,Ala³⁴]hPTH (1-34)OH

This peptide is prepared in analogy with the procedure of comparative Example 3a. Instead of Fmoc-Gly esterified to 4-hydroxymethyl-phenoxymethyl-co(polystyrene-divinylbenzene) the appropriate Fmoc-amino acid, e.g. Fmoc-Ala, Fmoc-Phe, Fmoc-D-Ala, Fmoc-D-Phe, etc. is used. Additionally the side-chain functional groups are protected as follows: Thr(t.-butyl), Trp(Boc) and Tyr(t.-butyl).

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By repeating the procedure disclosed in Examples 3 and 5 but using the appropriate starting materials, the following compounds may be obtained:

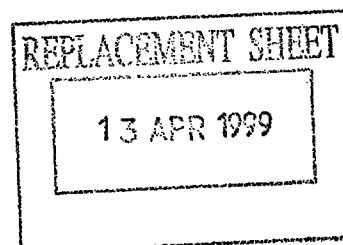
- Example 1: [Leu⁸, Asp¹⁰, Ala¹⁶, Gln¹⁸, Thr³³]hPTH (1-34)OH
 Example 2: [Leu⁸, Lys¹¹, Gln¹⁸]hPTH (1-36)OH
 Example 3: [Leu⁸, Gln¹⁸, Thr³³, Ala³⁴]-hPTH (1-34)OH (IS-MS: 4007)
 Example 4: [Leu⁸, Ala¹⁶, Gln¹⁸, Ala¹⁹, Thr³³, Ala³⁴]hPTH (1-34)OH
 Example 5: [Leu⁸, Ala¹⁶, Gln¹⁸, Thr³³, Ala³⁴]hPTH (1-34)OH (IS-MS: 3965)
 Example 6: [Leu⁸, Asp¹⁰, Lys¹¹, Gln¹⁸]hPTH (1-36)OH
 Example 7: [Leu⁸, Gln¹⁶, Asp¹⁷, Leu¹⁸, Arg¹⁹, Thr³³, Ala³⁴]-hPTH (1-34)OH
 Example 8: [Leu⁸, Asp¹⁰, Lys¹¹, Gln¹⁸, Thr³³, Ala³⁴]hPTH (1-34)OH (IS-MS: 4022)
 Example 9: [Leu⁸, Asp¹⁰, Lys¹¹, Gln¹⁶, Asp¹⁷, Leu¹⁸, Arg¹⁹, Thr³³, Ala³⁴]hPTH (1-34)OH (IS-MS: 4077)
 Example 10: [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸, Ala¹⁹]hPTH (1-36)OH (IS-MS: 4181)
 Example 11: [Leu⁸, Ala¹⁶, Asp¹⁷, Gln¹⁸, Ala¹⁹]hPTH (1-36)OH (IS-MS: 4193)
 Example 12: [Leu⁸, Ala¹⁶, Ala¹⁷, Gln¹⁸, Ala¹⁹]hPTH (1-36)OH (IS-MS: 4149)
 Example 13: [Leu⁸, Ala¹⁷, Gln¹⁸, Ala¹⁹]hPTH (1-36)OH
 Example 14: [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸, Thr³³, Ala³⁴]hPTH (1-34)OH (IS-MS: 3980)
 Example 15: [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸]hPTH (1-36)OH (IS-MS: 4240)
 Example 16: [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸, Ala¹⁹, Thr³³, Ala³⁴]hPTH (1-34)OH (IS-MS: 3923)
 Example 17: [Leu⁸, Asp¹⁰, Ala¹⁶, Gln¹⁸]hPTH (1-36)OH (IS-MS: 4225)
 Example 18: [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸, Ala¹⁹, Thr³³]hPTH (1-34)OH (IS-MS: 3999)
 Example 19: [Leu⁸, Ala¹⁶, Gln¹⁸, Ala¹⁹]hPTH (1-36)OH (IS-MS: 4166)
 Example 20: [Thr³³]-hPTH-(1-38)-OH

Comparative EXAMPLE 6: [1-aminocyclopentane-1-carboxylic acid¹-hPTH-(1-36)NH₂

The peptide is assembled in a stepwise manner on a polystyrene based resin support. The Fmoc-group is used for protection of the alpha-amino groups.

Side-chain functional groups are protected as Arg(Pmc), Asn(Trt), Asp(OtBu), Gln(Trt), Glu(OtBu), His(Trt), Lys(Boc), Ser(tBu), Thr(tBu), Trp(Boc) and Tyr(tBu). Other amino acids are left unprotected.

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4-(2',4'-Dimethoxyphenyl-Fmoc-amino-methyl)-phenoxy-co(polystyrene-divinylbenzene), 0.4 mmol/g, which may be prepared, e.g., as described in Tetrah. Letters, 28, 3787-3790 (1987) is subjected to the following cycle, steps (1) to (5), of treatments:

- (1) DMF
- (2) piperidine (20%) in DMF
- (3) DMF
- (4) mixture of HOBt, diisopropylcarbodiimide, and Fmoc-alanine (0.8 mmol per gram starting resin each)
- (5) DMF

Volumes of washes and reagents are from 5 to 20 mL per gram of starting resin.

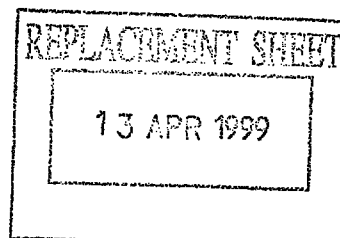
In the next cycle of treatments (1) to (5), Fmoc-valine is substituted for Fmoc-alanine and so on for each cycle such as to assemble on the resin the correct amino acid sequence of the title compound.

Each step is repeated as many times as necessary for either complete reaction of the resin (steps 2, 4) or complete displacement of the previous reagent(s) from the resin (steps 3, 5). Samples of the resin are removed after each cycle and checked for completeness of the coupling reaction by a colorimetric test for residual amino groups using ninhydrin.

At the end of the synthesis a final cycle comprising steps (1) to (3) only is performed and the peptide resin is washed with 2-propanol, then with a mixture of methanol and methylene chloride (1:1 v/v) and dried thoroughly in a vacuum desiccator.

The peptide resin (1 g) is suspended in a mixture (20 ml) of trifluoroacetic acid, water, and 1,2-ethanedithiol (90:5:5 v/v) for 2 hours at room temperature, the resin particles are filtered off and washed with a small volume of TFA. The product is precipitated from the combined filtrates by addition of ether (20 volumes), filtered, washed with more ether and dried. The residue is dissolved in 2 % acetic acid, the solution let to stand at r.t. for 8 hours prior to lyophilization. The lyophilisate is chromatographed on a C-18 silica column using a gradient of acetonitrile in 2% H₃PO₄. Fractions are checked by analytical HPLC and those containing the pure compound are collected, filtered through an anion-exchange resin in the acetate form and lyophilised to give the title compound as a polyacetate, polyhydrate.

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Fmoc-L-aminocyclopentane-L-carboxylic acid used in the preparation of the peptide resin intermediate may be prepared, e.g. as described by G. Valle et al., 1988, in Can.J.Chem.66:2575-2582.

MS(Ion-Spray): 4312

By repeating the procedure as disclosed in Comparative Example 6 but using the appropriate starting materials, following compounds may be obtained:

Example 21: [Thr³³, Ala³⁴]hPTH (1-34)NH₂

Example 22: [Gln¹⁸]hPTH (1-36)NH₂

Example 23: [Ala³⁴]hPTH (1-36)NH₂ (IS-MS: 4210)

Example 24: [Leu⁸, Asp¹⁰, Lys¹¹, Thr³³, Ala³⁴]hPTH (1-34) NH₂

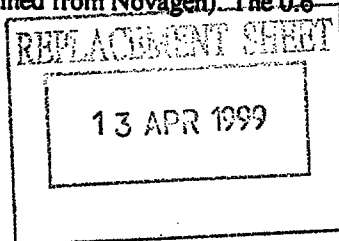
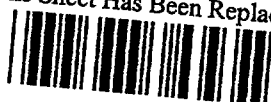
The compounds of Examples 1 to 24 show correct amino acid ratios in quantitative amino acid analysis.

In the following examples illustrating the recombinant process of the invention, if not otherwise specified, the standard procedures described in Ausubel et al; 1991; Current Protocols in Molecular Biology, John Wiley & Sons, New York are used.

EXAMPLE 25: Cloning bacteriophage T4 gene 55

Two DNA fragments, one containing the entire coding sequence of bacteriophage T4 gene 55, and the other fragment encoding part of it are amplified by polymerase chain reaction (PCR) using purified T4 DNA as the template. Oligonucleotides which contain NdeI or BamHI endonuclease recognition sequences are used as primers. The PCR reactions (25 cycles, each 30" 94°C, 30" 50°C, 30" 72°C) are performed with 1 ng of template and 100 pM of the upstream primer (GAGGTGCATA TGTCAGAAAC TAAGCCT 27), the NdeI recognition site being provided by nucleotides 7-12 thereof and 100 pM of the downstream primer (GGTCGGATCC ATCGTTAGCG TTAGCCTCAT ATAAAAAATC 40 the BamHI, recognition site being provided by nucleotides 5-10 thereof) in 100 µl reaction buffer containing 10 mM Tris-HCl, 50 mM KCl, 1.5 mM MgCl₂, 0.2 mM of each of dATP, dCTP, dGTP, dTTP, and 0.1% Triton X 100, pH 9.0. A 0.6kb fragment is obtained. The truncated form of gene 55 is prepared by PCR as described for the entire gene, except that a different oligonucleotide is used as downstream primer (GGTCGGATCC TACGTTGGAC GAATGCAT 28, the BamHI recognition site being provided by nucleotides 5-10 thereof). A 0.35 kb fragment is obtained. The 0.6 kb and 0.35 kb DNA fragments are digested with restriction endonucleases NdeI/BamHI and are cloned into a NdeI/BamHI digest of expression vector pET17B (obtained from Novagen). The 0.6-

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kb DNA fragment encodes the entire gp55 protein (188 amino acid residues - see Fig. 1), whereas the 0.35 kb fragment encodes a 12 kD amino-terminal fragment of gp55 (112 amino acid residues - see Fig. 2).

Example 26: Preparation and cloning of the DNA encoding PTH (1-38)

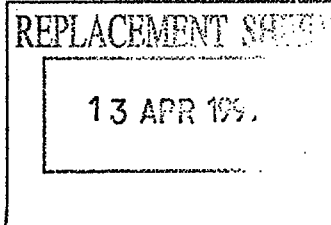
DNA fragments encoding a) the linker peptide corresponding to amino acid residues 110-115 of Fig. 1 and PTH (1-38) or b) the linker peptide corresponding to amino acid residues 186-191 of Fig. 2 and PTH (1-38) are generated by PCR using a cloned synthetic PTH (1-38) coding sequence as a template. The oligonucleotides encoding the linker a) (GAGTGATCC ATCGATCCAC CATCCGTATC AGAAATACAA CT 42), or encoding linker b) GAGTGGATCC GTTAACGGTC CATCCGTATC AGAAATACAA CT 42 are used as upstream primers. The downstream primer TCTACTCGAG TTAACCCAGA GCTACAAAAT TATG 34

The upstream primers each incorporate a BamHI cloning site and the downstream primer contains a XhoI recognition site (nucleotides 5-10 of each sequence). The PCR reaction is performed under the same conditions as described in Example 25. After cleavage with BamHI and XhoI, the 0.14 kb PCR products are inserted into BamHI-XhoI digests of the gp55-pET17B plasmids, obtained in Example 25. For expression of the fusion proteins the resultant plasmid constructs are transformed into E.coli BL21 (DE3) LysE and single bacterial colonies are further propagated. The expression of the fusion proteins is obtained by incubating a preculture at 37°C on a rotary shaker overnight in circle growth medium (Bio 101). A main culture is inoculated with the preculture making up 1 to 5 % of the volume of the main culture. The main culture is then fermented at a temperature of 37°C and at a pH of 6.9 to 7.1 whilst being aerated at 10 l/min and agitated at 200-400 rpm. Expression is induced by the addition of 1 mM IPTG as soon as the culture reaches an OD₅₅₀ of about 1.0 to 5.0. After induction the fermentation is continued for five hours and then the fermentation broth is harvested without killing the *E. coli* cells. The harvested cells are spun down with a tubular centrifuge and resuspended in sample buffer and assayed for expression by SDS PAGE. The cell pellet is then frozen until purification. The procedures and host cells used are fully described in Studier et al; 1990; "Use of T7 RNA Polymerase to direct expression of Cloned Genes", Methods in Enzymology Academic Press. pages 60 to 89.

EXAMPLE 27: Preparation of inclusion bodies

Frozen E. coli pellets obtained from Example 26 are resuspended to 25% w/v in Buffer A (50mM Tris PH 8.0; containing 2mM DTT, 5mM Benzamidine-HCl, 1.5mM MgCl₂, 1.0mM MnCl₂, 10pg/ml. DNase I and

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2mg/ml Lysozyme) and mixed by stirring for 1 hour at room temperature. The resuspended cells are lysed by passage through a Manton-Gaulin homogeniser (2 passes at 1200 bar) and the resultant lysate kept on ice. The lysate is diluted 2 fold with ice-cold buffer B (50mM Tris pH 8.0; containing 2mM DTT, 5mM Benzamidine-HCl and 4mM EDTA) and centrifuged for 30min at 27,500g. The supernatant from the centrifuge is carefully removed and discarded. The pellet is resuspended in buffer B to 25% w/v, passed once more through the Manton-Gaulin and centrifuged as before. This process is repeated once using buffer B and once using water. The resultant inclusion body pellets are weighed and frozen at -20°C until required.

EXAMPLE 28: Solubilisation and cleavage of gp55-Asp-Pro-Pro-(1-38)hPTH inclusion bodies

Frozen inclusion bodies containing gp55-Asp-Pro-Pro-(1-38)hPTH as obtained in Example 27 are suspended in 10 mM HCl acid solution (analytical grade) to a final concentration of 6%w/v. The solution is incubated for 24 hours at 50°C and the reaction terminated by addition of 1 volume of aqueous 100 mM NaOAc solution. The diluted supernatant is clarified by centrifugation at 27,500g for 30 minutes and filtered through a glass fibre filter.

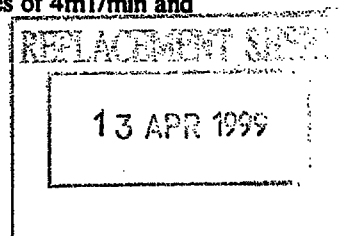
EXAMPLE 29: Solubilisation and cleavage of gp55-Asn-Gly-Pro-(1-38)hPTH inclusion bodies

Frozen inclusion bodies containing gp55-Asn-Gly-Pro-(1-38)hPTH and obtained from example 27 are dissolved in 6M Guanidine-HCl (containing 2M Hydroxylamine-HCl and brought to pH 9.0 by addition of 4.5M LiOH) to give an estimated 5mg/ml protein. The pH is readjusted to pH 9 by further addition of LiOH and the solution incubated for 4 hours at 45°C. The reaction is terminated by addition of formic acid to pH 4 and the solution centrifuged and filtered as described in example 28.

EXAMPLE 30: Preparative HPLC of Gly-Pro and Pro-Pro (1-38)hPTH

Filtered cleavage supernatants are analysed on analytical reversed phase HPLC (Orpogen HD-Gel-RP-7s-300; 4 x 150mm column) and compared with a known (1-38)hPTH standard (Bachem) to determine the PTH concentration. The column is equilibrated with 85% HPLC buffer A (90% water, 10% acetonitrile; containing 0.1%v/v TFA) and 15% HPLC buffer B (10% water, 90% acetonitrile; containing 0.1% TFA) and is eluted with a gradient of 15% HPLC buffer B to 100% HPLC buffer B over 15 minutes at a flow rate of 0.75ml/min. Depending upon the scale of the preparation, supernatants are either loaded on a 1.0 x 25cm C4 Vydac or a 2.2 x 25cm C4 Vydac. The columns are equilibrated with 85% HPLC buffer A and 15% HPLC buffer B at flow rates of 4ml/min and

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10ml/min respectively. The columns are eluted with gradients of 15%B to 85%B over 80ml and 15%B to 55%B over 300ml respectively. Gly-Pro-hPTH(1-38) or Pro-Pro-hPTH(1-38) peaks are collected manually and the acetonitrile is later removed under vacuum. Analytical HPLC is used to determine the peptide concentration both after cleavage and in the collected fractions. The solvent free peptide solutions are diluted with an equal volume of 100mM sodium acetate and the pH adjusted, if necessary, to pH 5.4. Following filtration (0.2µm), the solution is loaded on a cation exchange column (either Mono S or SP-Sepharose High Performance, packed into HR10/10 or XK16/10 columns respectively) equilibrated with 50mM sodium acetate pH 5.4 (buffer C). The column is eluted with a 0 to 500mM NaCl gradient in buffer C over 7.5 column volumes. 10ml fractions are collected and the X-X-(1-38)hPTH peak is pooled. The peptide concentration is determined as before using HPLC. This column serves both to exchange the peptides into a more physiological buffer and to remove additional peptides produced by chemical cleavage of the fusion protein itself.

EXAMPLE 31: Enzymatic removal of X-X from X-X-(1-38)hPTH

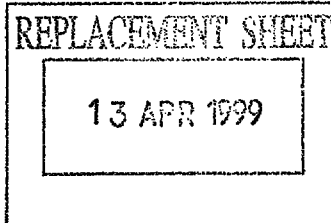
Pooled peptide fractions, as obtained in Example 30, (concentration range 1.5 - 2.0 mg/ml) are brought to pH 8.0 by addition of solid Tris. Purified X-Prolyl dipeptidase (0.5mg/ml in PBS pH 7.2) (Nardi et al. (1991) App. Env. Microbiol. 57, No.1, 45-50) is added to 1:1000 and the solution is incubated for 24h at 37 C. The reaction is stopped by addition of TFA to 0.2% (pH = 5.0) and the resultant (1-38)hPTH is isolated using preparative HPLC (using the same conditions as described in Example 30). The solvent is removed and the product is lyophilised after removal of aliquots for N-terminal sequence and mass spectroscopic analyses.

Examples of the yields which are obtained using the two fusion protein approaches described above are as follows:

Gen55-Asp-Pro-Pro-(1-38)PTH (using the "truncated" form of Gen55)

Under partially optimized fermentation conditions, a 215 g wet cell pellet is obtained from a 10 litre fermentation. The percentage level of expression of the fusion protein is approximately 37%. From this pellet, 31 g of inclusion bodies are isolated. The protein purity (with respect to the fusion protein) of these bodies is 61%. After solubilisation, cleavage and centrifugation/filtration approximately 620mg Pro-Pro-PTH are detected. This material yields 580mg Pro-Pro-PTH after subsequent C4-Vydac HPLC. After dilution, cation-exchange on Mono S, digestion with X-Prolyldipeptidase (at a Pro-Pro-PTH concentration of 2mg/ml), HPLC and lyophilisation, 440mg of lyophilized pure product

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with full biological activity is obtained. This represents a yield of 71% between the stopped cleavage step and the final product.

Gen55-Asn-Gly-Pro-(1-38)PTH (using the "long" form of Gen55)

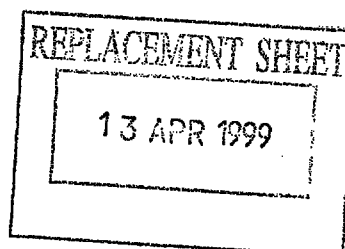
From a non-optimized 20 litre fermentation a 78g wet cell pellet with 50% expression of the fusion protein is harvested. Following lysis and centrifugation, 18g of "90% pure" inclusion bodies are obtained. Following the acid stop step and centrifugation/filtration about 470mg Gly-Pro-PTH are detected. After HPLC on C4-Vydac, 355mg of peptide are detected, decreasing to 321mg after SP-Sepharose cation-exchange. Enzymatic removal of the Gly and Pro residues is carried out at 2mg/ml. After the final HPLC step and lyophilization, 244mg of pure, fully biologically active (1-38)PTH is recovered representing a yield of 52% from the stopped cleavage reaction stage.

The compounds of the invention in free form or in the form of pharmaceutically acceptable salts and complexes exhibit valuable pharmacological properties as indicated in animal tests and are therefore indicated for therapy.

The biological activity of the compounds of the invention is assessed in vitro by measuring their ability of stimulating the synthesis of cyclic AMP in UMR 106-06 rat and SaOS-2 human osteosarcoma cells according to the method of Marcus and Aurbach in *Endocrinology*, 85, 801-810 (1969). Rat osteosarcoma UMR 106 cells are grown to confluence in Eagle's Minimum Essential Medium - 10% FCS in 12 well plates, human SaOS-2 osteosarcoma cells are grown in McCoy's 5A medium - 10% FCS. The medium is then changed to medium with 2% FCS and 1-5 pCi/well [3H]-adenine is added. Two hours later, cells are washed twice with serum-free medium and incubated in DMEM - 1% BSA containing 1 mM 3-isobutyl-1-methylxanthine to exclude actions on phosphodiesterases. Test substances are added 20 min later. The reaction is stopped and cAMP extracted after 15 min by adding ice cold 5% trichloroacetic acid. A carrier solution (0.5 ml/well) containing 5 mM of unlabelled adenine, adenosine, AMP, ADP, ATP, and cAMP as well as 0.4 μ Ci [14C]-adenosine for determination of recovery is added. [M]-cAMP is separated using serial Dowex 50W-X4 (200-400 mesh) and alumina chromatography and counted. Results are calculated in % of solvent control and EC_{50} values determined from DRC curves. Compounds of the invention stimulate cAMP production in UMR 106-06 rat and SaOS-2 human osteosarcoma cells at a concentration of 10^{-11} to 10^{-6} M.

The compounds of the invention also have binding affinity to PTH receptors, e.g. as follows:

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Chicken [Tyr³⁶]PTHrP(1-36)amide is iodinated to a specific activity of 2.200 Ci/mmol using the lactoperoxidase method (Anawa Lab. AG, Wangen). Monolayers of opossum kidney cells are washed with 200 μ l DMEM and HAM's F12 (1:1) containing 1% BSA and incubated at 16°C with 50.000cpm of [¹²⁵I-Tyr³⁶]chPTHrP(1-36)amide per well in the presence or absence of 1 μ M [Tyr³⁶]chPTHrP(1-36)amide. After incubation, cells are washed with 0.5 ml medium (4°C), lysed with 0.5 ml 1N NaOH and radioactivity is determined. Specific binding is defined as total binding minus nonspecific binding. Competition curves are analyzed using SCTFIT, a non-linear regression computer program (Feyen et al. 1992, Biochem. Biophys. Res. Commun. 187:8-13) and data presented as mean pK_D values (n=2 to 3). Compounds of the invention show in this test a binding affinity expressed as a mean pK_D value of from 7 to 10.

Furthermore, the compounds of the invention increase plasma calcium level after continuous s.c. infusion, e.g. as determined in male thyroparathyroidectomized Wistar rats weighing 140 to 170 g. 5 days after thyroparathyroidectomy, rats are implanted in the neck with Alzet minipumps and test compounds infused at rates of 0.3 to 30 μ g/day/rat. Blood is drawn by retro-orbital puncture in the morning of days 1, 2, 3, and 6 thereafter and plasma calcium concentrations are determined photometrically. In this test compounds of the invention increase plasma calcium.

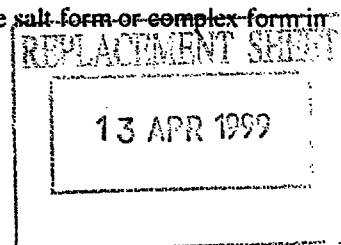
The compounds of the invention are accordingly indicated for preventing or treating all bone conditions which are associated with increased calcium depletion or resorption or in which calcium fixation in the bone is desirable, e.g. osteoporosis of various genesis (e.g. juvenile, menopausal, post-menopausal, post-traumatic, caused by old age or by cortico-steroid therapy or inactivity), fractures, osteopathy, including acute and chronic states associated with skeletal demineralisation, osteo-malacia, periodontal bone loss or bone loss due to arthritis or osteoarthritis or for treating hypoparathyroidism.

The compounds of the invention are particularly indicated for preventing or treating osteoporosis of various genesis.

For all the above uses, an indicated daily dosage is in the range from about 0.003 to about 10 mg preferably 0.003 to 3, more preferably 0.01 to 1 mg of a compound of the invention. Compounds of the invention may be administered once a day or up to twice a week.

The compounds of the invention may be administered in free form or in pharmaceutically acceptable salt form or complexes. Such salts and complexes may be prepared in conventional manner and exhibit the same order of activity as the free compounds. The present invention also provides a pharmaceutical composition comprising a compound of the invention in free base form or in pharmaceutically acceptable salt form or complex form in

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association with a pharmaceutically acceptable diluent or carrier. Such compositions may be formulated in conventional manner. The compounds of the invention may be administered by any conventional route, for example parenterally e.g. in form of injectable solutions or suspensions, enterally, e.g. orally, for example in the form of tablets or capsules or in a transdermal, nasal or a suppository form.

In accordance with the foregoing the present invention further provides:

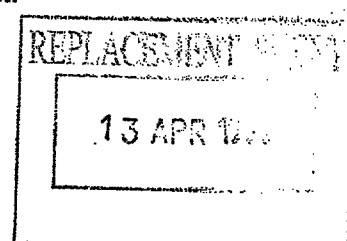
- a) a compound of the invention or a pharmaceutically acceptable salt or complex thereof for use as a pharmaceutical;
- b) a method for improving bone formation, e.g. preventing or treating all bone conditions which are associated with increased calcium depletion or resorption or in which calcium fixation in the bone is desirable, e.g. osteoporosis of various genesis (e.g. juvenile, menopausal, post-menopausal, post-traumatic, caused by old age or by cortico-steroid therapy or inactivity), fractures, osteopathy, including acute and chronic states associated with skeletal demineralisation, osteomalacia, periodontal bone loss or bone loss due to arthritis or osteoarthritis, or for treating hypoparathyroidism in a subject in need of such treatment, which method comprises administering to said subject an effective amount of a compound of the invention or a pharmaceutically acceptable salt or complex thereof; or
- c) a compound of the invention or a pharmaceutically acceptable salt or complex thereof for use in the preparation of a pharmaceutical composition for use in the method as in b) above.

According to a further embodiment of the invention, the compounds of the invention may be employed as adjunct or adjuvant to other therapy, e.g. a therapy using a bone resorption inhibitor, for example as in osteoporosis therapy, in particular a therapy employing calcium, a calcitonin or an analogue or derivative thereof, e.g. salmon, eel or human calcitonin, a steroid hormone, e.g. an estrogen, a partial estrogen agonist or estrogen-gestagen combination, a biphosphonate, vitamin D or an analog thereof, or any combination thereof.

When the compounds of the invention are administered in conjunction with, e.g. as an adjuvant to bone resorption inhibition therapy, dosages for the co-administered inhibitor will of course vary depending on the type of inhibitor drug employed, e.g. whether it is a steroid or a calcitonin, on the condition to be treated, whether it is a curative or preventive therapy, on the regimen and so forth.

In accordance with the foregoing the present invention provides in a yet further aspect:

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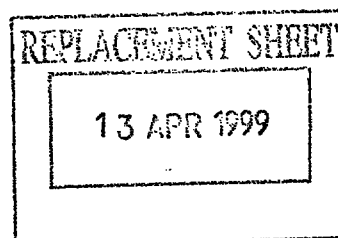


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d) a method for improving bone formation, e.g. preventing or treating calcium depletion, for example for preventing or treating any of the specific conditions or diseases hereinbefore set forth, in a subject in need of such a treatment which method comprises administering to said subject an effective amount of a) a compound of the invention and b) a second drug substance, said second drug substance being a bone resorption inhibitor, for example as indicated above.

Compounds of Examples 3,5,8,14 and 15 are preferred.

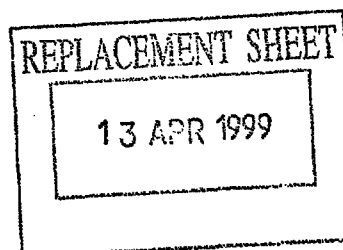
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CLAIMS

I. A Compound of formula I

Ser-Val-Ser-Glu-Ile-Gln-Leu-R₁-His-R₂-Ile-Gly-R₃-His-Leu
 R₅-R₆-R₇-R₈-Arg-Val-Glu-Trp-Leu-Arg-R₉-Lys-Leu-Gln-Asp-
 Val-His-R₁₀-R₁₁-(R₁₂)_n-X



wherein

R₁ is Met or Leu;

R₂ is Asn, Asp, Gly, Gln, Glu, His, Ser, Thr or Tyr;

R₃ is Leu or Lys;

R₄ is Lys or D- or L-Ala, -Cys, -Gln, -Ile, -Asn, -Trp, -Asp, -Val, -Thr, -Ser, -Tyr, -Met, -Leu or -Gly;

R₅ is Asn, Gln, D-Gln or D- or L-Lys, -Ser, -Leu, -Ala or -Gly;

R₆ is Ser, Asp, Ala, Glu, Gln, Phe, His, Ile or Lys;

R₇ is Gln

R₈ is Glu, Arg, Ala, Val, Tyr, Ser, Lys, Met, His, Gly, Pro, Asn or Ile;

R₉ is Lys, Gln or Arg;

R₁₀ is Asn, Thr, D-Thr or D- or L-Ser, -Leu, -Gly, -Gln, -Arg, -Pro, -Asp, -Ile or -Lys;

R₁₁ is Phe or Ala;

R₁₂ is Val, Val-Ala, Val-Ala-Leu or Val-Ala-Leu-Gly;

n is 0 or 1; and

X is OH, OR_a, NH₂ or NR_bR_c wherein R_a is C₁₋₄alkyl, one of

R_b and R_c is H and the other is C₁₋₆alkyl,

the compound optionally comprising at least one radical selected from a L- or D-α-amino acid,

C₂₋₆alkoxy-carbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, arakyl, aralkenyl or

C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to its terminal amino group, and/or

the compound optionally comprising at least one radical selected from C₂₋₆alkoxycarbonyl and

optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, arakyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl

attached to one or more side chain amino groups of the compound, in free form or in salt or complex form.

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* C L M S *

2. A compound according to claim 1 wherein R_{10} is Thr.
3. A compound according to claim 1 or 2, wherein R_{11} is Ala.

4. A compound selected from [Leu⁸, Gln¹⁸, Thr³³, Ala³⁴]hPTH, [Leu⁸, Ala¹⁶, Gln¹⁸, Thr³³, Ala³⁴]hPTH, [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸, Thr³³, Ala³⁴]hPTH, [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸]hPTH, [Leu⁸, Ala¹⁶, Gln¹⁸, Ala¹⁹]hPTM, [Leu⁸, Asp¹⁰, Lys¹¹, Gln¹⁸]hPTH [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸, Ala¹⁹, Thr³³, Ala³⁴]hPTH and [Leu⁸, Asp¹⁰, Lys¹¹, Gln¹⁸, Thr³³, Ala³⁴]hPTH

the compound optionally comprising at least one radical selected from a L- or D-a-amino acid, C₂₋₆alkoxy-carbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to its terminal amino group,

and/or at least one radical selected from C₂₋₆alkoxycarbonyl and optionally substituted C₁₋₈alkyl,

C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to one or more side chain amino groups of the compound,

in free form or in salt or complex form.

5A compound selected from

[Leu⁸, Gln¹⁸, Thr³³, Ala³⁴] 1-hPTH(1-34)OH

[Leu⁸, Ala¹⁶, Gln¹⁸, Ala¹⁹, Thr³³, Ala³⁴]hPTH(1-34)OH

[Leu⁸, Ala¹⁶, Gln¹⁸, Thr³³, Ala³⁴]hPTH(1-34)OH

[Leu⁸, Asp¹⁰, Lys¹¹, Gln¹⁸]hPTH(1-36)OH

[Leu⁸, Asp¹⁰, Lys¹¹, Gln¹⁸, Thr³³, Ala³⁴]hPTH(1-34)OH

[Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸, Ala¹⁹]hPTH(1-36)OH

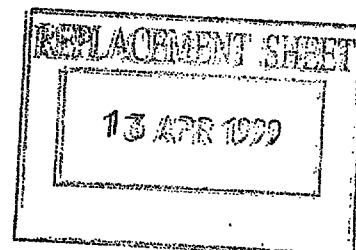
[Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸, Thr³³, Ala³⁴]hPTH(1-34)OH

[Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸]hPTH(1-36)OH, and

[Leu⁸, Ala¹⁶, Gln¹⁸, Ala¹⁹]hPTH(1-36)OH

in free form or in salt or complex form.

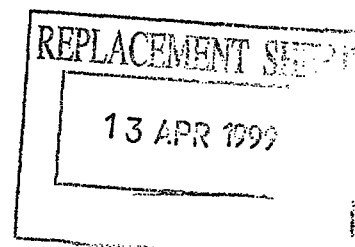
6. A process for the production of a compound according to any one of the preceding claims in which a fusion protein comprising a N-terminal bacteriophage T4 gene 55 polypeptide having the desired compound linked to the C-terminal thereof, is expressed in bacterial cells, and, where required, introducing a C₂₋₆alkoxy-carbonyl or optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl on the terminal and/or side chain amino group of the compound in protected or unprotected form and, if required, removing the protecting group present in the protected form.



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7. A process according to claim 6 in which the fusion protein comprises a chemically cleavable linker between the gene 55 and the desired compound.
8. A process according to claim 7 in which the cleavable linker comprises the amino acid sequences Asp-Pro-Pro or Asn-Gly-Pro.
9. A nucleotide sequence which codes for a fusion protein comprising an N-terminal bacteriophage T4 gene 55 polypeptide and a desired compound according to any one of the claims 1 to 5, linked to the C-terminal thereof.
10. A bacterial expression vector comprising a DNA sequence according to claim 9.
11. Bacterial host cells transformed with a DNA sequence according to claim 9, or a bacterial vector according to claim 10.
12. A fusion protein comprising an N-terminal bacteriophage T4 gene 55 polypeptide and a desired compound according to any of the claims 1 to 5, linked to the C-terminal thereof.
13. A compound according to any one of claims 1 to 5 in free form or in pharmaceutically acceptable salt form for use as a pharmaceutical.
14. A pharmaceutical composition comprising a compound as defined in any one of claims 1 to 5 in free form or in pharmaceutically acceptable salt form, together with a pharmaceutically acceptable diluent or carrier therefor.
15. A compound according to any one of claims 1 to 5, in free form or in pharmaceutically acceptable salt or complex form, for use in the preparation of a pharmaceutical composition for use in improving bone formation.
16. A compound according to any one of claims 1 to 5, in free form or in pharmaceutically acceptable salt or complex form, for use in association with a bone resorption inhibitor to improve bone formation.



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ANALOGS OF PTH

Abstract of the invention

PTH compounds having PTH-like activity and comprising at least one modification, said modification being either

1. at least one radical selected from a L- or D- α -amino acid, C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to the terminal amino group of the PTH compound, and/or at least one radical selected from C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to one or more side chain amino groups of the PTH compound,

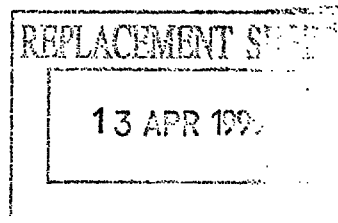
or

2. at least one α -amino acid unit in the positions 1 to 38 of a naturally occurring PTH sequence being replaced by a natural or unnatural amino acid unit optionally in protected form, whereby the α -amino acid units present in positions 1 and 2 at the amino terminus of the PTH sequence may be replaced by a pseudo-peptide,

or a combination of such modifications,

in free form or in salt form, have pharmacological activity, e.g. for preventing or treating all bone conditions which are associated with increased calcium depletion or resorption or in which calcium fixation in the bone is desirable.

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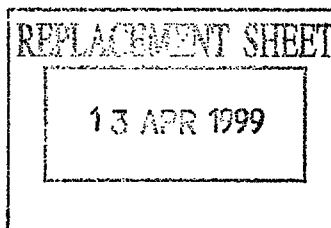


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A representative compound is e.g. of formula 1

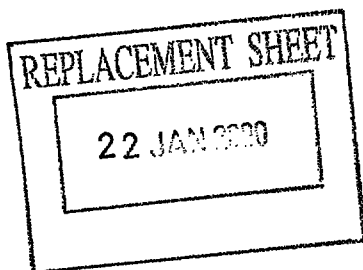
Ser-Val-Ser-Glu-Ile-Gln-Leu- R_1 -His- R_2 - R_3 -Gly- R_4 -His-Leu-
 R_5 - R_6 - R_7 - R_8 -Arg-Val-Glu-Trp-Leu-Arg- R_9 -Lys-Leu-Gln-Asp-
Val-His- R_{10} - R_{11} -(R_{12}) $_n$ -X

wherein each of R_1 to R_{11} has a meaning within the invention, R_{12} is Val, Val-Ala, Val-Ala-Leu or Val-Ala-Leu-Gly; n is 0 or 1; and X is OH, NH_2 or substituted OH or NH_2 , provided that R_7 is Gln or Tyr, and/or R_{10} is Thr and/or R_{11} is Ala.



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ANALOGS OF PTH

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REPLACEMENT SHEET

✓	Abstract		to	
✓	Summary		to	
✓	Claims		to	
✓	Drawings		to	
✓	References		to	
✓	Index		to	
✓	Abstract		to	
DATE OF RECEIPT 12/1/25				

The present invention relates to variants of parathyroid hormone (PTH), a process for their production, pharmaceutical preparations comprising them and their use as a pharmaceutical.

Compounds of this type are well-known, e.g., see WO 9211286.

The term "PTH" as used herein refers to any genetically encoded form of parathyroid hormone, including the mature form containing 84 amino acids of a given vertebrate PTH species, e.g. human, porcine, rat, bovine, chicken, and fragments thereof as well as analogues and derivatives thereof having PTH-like activity. The position of each amino acid involved in the PTH sequence is numbered according to the internationally accepted procedure: For consistency and as is conventional, in the following description, the same numbering system will be applied to the amino acids of the PTH sequence independently of the substitution pattern in the molecule.

More particularly, the present invention provides a PTH compound having PTH-like activity of formula 1

Ser-Val-Ser-Glu-Ile-Gln-Leu-R₁-His-R₂-R₃-Gly-R₄-His-Leu-R₅-R₆-R₇-R₈-Arg-Val-Glu-Trp-Leu-Arg-R₉-Lys-Leu-Gln-Asp-Val-His-R₁₀-R₁₁-(R₁₂)_n-X

wherein

R₁ is Met or Leu;

R₂ is Asn, Asp, Gly, Gln, Glu, His, Ser, Thr or Tyr;

R₃ is Leu or Lys;

R₄ is Lys or D- or L-Ala, -Cys, -Gln, -Ile, -Asn, -Trp, -Asp, -Val, -Thr, -Ser, -Tyr, -Met, -Leu or -Gly;

R₅ is Asn, Gln, D-Gln or D- or L-Lys, -Ser, -Leu, -Ala or -Gly;

R₆ is Ser, Asp, Ala, Glu, Gln, Phe, His, Ile or Lys;

R₇ is Gln

R₈ is Glu, Arg, Ala, Val, Tyr, Ser, Lys, Met, His, Gly, Pro, Asn or Ile;

R₉ is Lys, Gln or Arg;

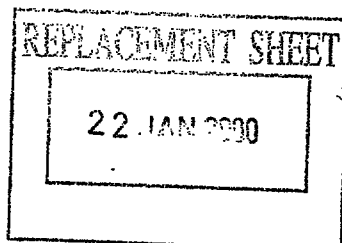
R₁₀ is Asn, Thr, D-Thr or D- or L-Ser, -Leu, -Gly, -Gln, -Arg, -Pro, -Asp, -Ile or -Lys;

R₁₁ is Phe or Ala;

R₁₂ is Val, Val-Ala, Val-Ala-Leu or Val-Ala-Leu-Gly;

n is 0 or 1; and

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REPLACEMENT SHEET

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X is OH, OR_a, NH₂ or NR_bR_c wherein R_a is C₁₋₄alkyl, one of R_b and R_c is H and the other is C₁₋₆alkyl, the compound optionally comprising at least one radical selected from a L- or D-α-amino acid, C₂alkoxy-carbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to its terminal amino group, and/or

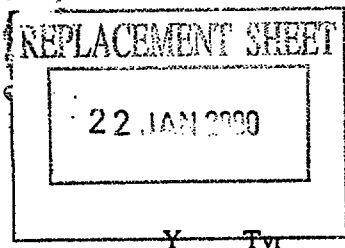
the compound optionally comprising at least one radical selected from C₂₋₆alkoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl attached to one or more side chain amino groups of the compound, in free form or in salt or complex form.

Hereinafter, these compounds will be referred to as compounds of the invention.

"Natural amino acids" refer to those well known in the art. They are listed and standard abbreviations are provided in the U.S.P.T.O. publication, Trademark Official Gazette, published May 15, 1990, P. 33 at 46. These amino acids and abbreviations are specifically incorporated herein by reference.

The natural amino acids are shown below:

A	Ala	alanine
D	Asp	aspartic acid
E	Glu	glutamic acid
F	Phe	phenylalanine
G	Gly	glycine
H	His	histidine
I	Ile	isoleucine
K	Lys	lysine
L	Leu	leucine
M	Met	methionine
N	Asn	asparagine
Q	Gln	glutamine
R	Arg	arginine
S	Ser	serine
T	Thr	threonine
V	Val	valine
W	Trp	tryptophane



REPLACEMENT SHEET

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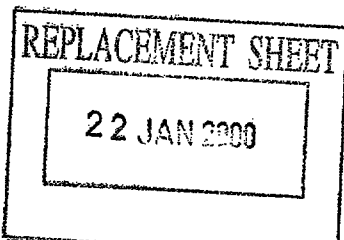
tyrosine

By amino acid in protected form is meant a natural amino acid having e.g. a side chain including an heteroatom such as O, S or N which can be protected with an O-, S- or N-protecting group. The N-terminus of the PTH compounds of the invention may also be in protected form.

N-protecting groups as may be present on the N-terminus or side chain amino groups of amino acid units include such groups as e.g. disclosed in "Protective Groups in Organic Synthesis", T. W. Greene, J. Wiley & Sons NY (1981), 219-287, for example acyl such as formyl, acetyl, trifluoroacetyl, methoxysuccinyl, hydroxysuccinyl or benzoyl optionally substituted on the phenyl ring with e.g. p-methoxycarbonyl, p-methoxy, p-nitro or p-phenylsulfonamidocarbonyl; alkoxycarbonyl such as t-butyloxycarbonyl, isobutyloxycarbonyl or methoxycarbonyl; allyloxycarbonyl; trityl; 2,2,5,7,8-pentamethyl-chroman-6-sulfonyl; arylmethoxycarbonyl such as 9-fluorenylmethoxycarbonyl or benzyloxy carbonyl optionally substituted on the phenyl ring with p-methoxy, p-nitro, o- or p-chloro, m-phenyl or 3,4-dimethyl; arylmethyl such as benzyl optionally ring substituted with p-methoxy, p-nitro or p-chloro; or arylsulfonyl such as phenylsulfonyl optionally ring substituted with p-methyl or p-methoxy, or naphthylsulfonyl optionally ring substituted with e.g. amino or di(C₁₋₄alkyl)amino.

O-protecting groups of O-containing side chains are e.g. as described in "The Peptides", 3, (1981), E. Gross and J. Meienhofer (Ed.). For aliphatic hydroxy functionalities, suitable O-protecting groups are e.g. as disclosed in the above reference in "Protection of the Hydroxy Group", J.M. Stewart, 169-201 and include the benzyl, t-butyl and methyl groups. For aromatic hydroxy functionalities, suitable O-protecting groups include the benzyl, t-butyl, methyl, tosyl and benzyloxycarbonyl groups. O-protecting groups for carboxy functionalities on amino acid side chains are well known ester groups and described e.g. in "The Peptides", 3, 101-135 ("Carboxyl Protecting Groups" by R.W. Roeske) and include the methyl, ethyl, t-butyl and benzyl groups. S-protecting groups for thiol functionalities on amino acid side chains are known and described e.g. in "The Peptides", 3, 137-167 ("Sulfhydryl Group Protection" by R.G. Hiskey). Examples include the methyl, t-butyl, benzyl, p-methoxyphenylmethyl, ethylamino-carbonyl and benzyloxycarbonyl groups.

According to the invention the PTH compounds may bear on their terminal amino group at least one radical selected from a L- or D- α -amino acid, C₂₋₆alkoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl, and/or on one or more side chain amino group(s) at least one radical selected from C₂₋₆alkoxycarbonyl and optionally substituted



REPLACEMENT SHEET

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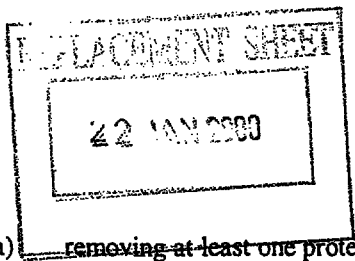
C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl. When such a group is attached to a side chain amino group, it is preferably on the ε-amino group of a Lys unit. Preferred substituents for the alkyl, alkenyl, aralkyl, aralkenyl or cycloalkylalkyl group are hydroxy, amino and for the aryl moiety also halogen and/or C₁₋₄alkoxy. The alkyl group or alkyl moiety may be linear or branched and optionally interrupted by O, S or N. Preferably any C₁₋₈alkyl, C₂₋₈alkenyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl on the amino group of a PTH compound, is non-substituted. Examples for C₁₋₈alkyl are C₁₋₆alkyl, preferably methyl, ethyl, propyl, isopropyl, butyl, isobutyl, t.-butyl; for C₂₋₈alkenyl, C₂₋₄alkenyl, preferably allyl; for C₂₋₈alkynyl, C₂₋₄alkynyl, preferably prop-2-ynyl; for aralkyl, phenyl or benzyl; for aralkenyl, styryl; for C₃₋₆cycloalkyl-C₁₋₄alkyl, cyclohexyl-methyl. C₂₋₆alkoxycarbonyl is preferably formyl or acetyl. Suitable examples of D- or L-α-amino acids attached to the N-terminus include e.g. D- or L- Pro, Ala. Preferred substituents when present are alkyl, alkynyl, alcoxycarbonyl or a D- or L-α-amino acid attached to the N-terminus of the PTH compound and/or at least one alkyl or alcoxycarbonyl attached to one or more side chain amino groups.

Examples of preferred compounds according to the invention are [Leu⁸, Gln¹⁸, Thr³³, Ala³⁴]PTH(1-x'), [Leu⁸, Ala¹⁶, Gln¹⁸, Thr³³, Ala³⁴]PTH (1- x'), [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸, Thr³³, Ala³⁴]PTH (1-x'), [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸]PTH(1- x'), [Leu⁸, Ala¹⁶, Gln¹⁸, Ala¹⁹]PTH(1- x'), [Leu⁸, Asp¹⁰, Lys¹¹, Gln¹⁸]PTH(1- x'), [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸, Ala¹⁹]PTH(1- x'), [Leu⁸, Ala¹⁶, Gln¹⁸, Ala¹⁹, Thr³³, Ala³⁴]PTH(1- x'), and [Leu⁸, Asp¹⁰, Lys¹¹, Gln¹⁸, Thr³³, Ala³⁴]PTH, wherein x' is an integer from 34 to 38, particularly 34, 36, 37 or 38. Especially PTH (1- x') is hPTH (1- x').

The compounds of the invention may exist e.g. in free form, salt form or in the form of complexes thereof. Acid addition salts may be formed with e.g. organic acids, polymeric acids and inorganic acids. Such acid addition salt forms include e.g. the hydrochlorides and the acetates. Complexes are e.g. formed from the compound of the invention on addition of inorganic substances, e.g. inorganic salts or hydroxides such as Ca- and Zn-salts, and/or an addition of polymeric organic substances.

The present invention also provides a process for the production of the compounds of the invention. They may be prepared in a stepwise manner either in solution or using the solid phase synthesis process or genetic engineering.

The compounds of the invention may be produced for example as follows:



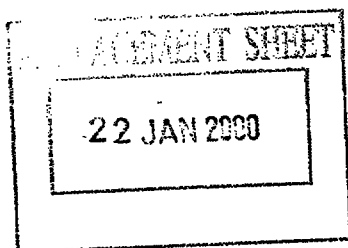
- a) ~~removing at least one protecting group which is present in a compound of the invention in~~
protected form; or
- b) linking together by an amide bond two peptide fragments, the peptide fragments being such that the desired amino acid sequence of the desired compound is obtained, and then effecting optionally stage a) of the process,
- c) for the production of a PTH compound comprising at least one radical selected from D- or L- α -amino acid attached to the N-terminus and C₂₋₆alkoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl attached to the terminal amino group and/or to a side chain amino group, introducing such a radical in a PTH compound in protected or unprotected form and free from such a radical, and if necessary carrying out process step a);

and recovering the compounds thus obtained in free form, in salt form or in complex form.

Process steps a), b) and c) may be effected in analogy with known methods, e.g. as known in the art of peptide chemistry or as described in the following examples. Where desired, in these reactions, protecting groups which are suitable for use in peptides may be used for functional groups which do not participate in the reaction. The term protecting group may also include a polymer resin having functional groups.

Process step c) may be carried out in analogy with known methods, e.g. alkylating methods. In a preferred embodiment of the invention, the alkylation is performed under reductive conditions, e.g. using a corresponding aldehyde or ketone. It may be performed for example in the presence of NaBH₃CN, preferably at an acidic pH, e.g. from 5 to 7. The temperature of the reaction may be e.g. from -20° C to 100°C. It may be advantageous to carry out the reaction in an inert solvent, e.g. water, an alcohol, dioxane or DMF or a mixture thereof. A D- or L- α -amino acid may also be attached to the N-terminus in accordance with known procedure.

The peptide fragment used as starting material in process step b) may be substituted on the N-terminus and/or on a side chain amino group by a radical selected from C₂₋₆alkoxycarbonyl, optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl or it may bear a D- or L- α -amino acid on the N-terminus.



The compounds of the invention or fragments thereof comprise natural α -amino acid units and may also be prepared using recombinant technology.

According to a preferred embodiment of the invention there is also provided a process for the production of a compound of the invention, in which a fusion protein comprising an N-terminal bacteriophage T4 gene 55 polypeptide having the desired compound linked to the C-terminal thereof, is expressed in bacterial cells and where required, introducing a C₂₋₆alkoxy carbonyl or optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl on the terminal and/or side chain amino group of the compound in protected or unprotected form and, if required, removing the protecting group present in the protected form.

The gene 55 polypeptide may comprise the complete gp55 protein of bacteriophage T4 (188 amino acid residues) which is described in Gram, H. and Rüger, R; 1985; The EMBO Journal, 4, (1), pp 257-264. Alternatively a fragment, preferably an N-terminal fragment of the gp55 protein may be used. For example, a gp55 protein fragment comprising the first 112 N-terminal amino acid residues of the protein may be used. Typically, however, the gene 55 polypeptide comprises at least the first 25 N-terminal residues of the gp⁵⁵ protein.

Conveniently the gene 55-PTH fusion protein also comprises a cleavable linker between the gene 55 polypeptide and the desired compound. Preferably the cleavable linker is chemically cleavable and more preferably contains the amino acid sequences Asp-Pro-Pro or Asn-Gly-Pro.

In a further aspect the invention provides a nucleotide sequence coding for a fusion protein comprising an N-terminal bacteriophage T4 gene 55 polypeptide and a desired compound of the invention linked to the C-terminal thereof.

The nucleotide sequence is preferably a DNA sequence suitable for expression in bacteria. The sequence typically contains nucleotides coding for a cleavable linker between the gene 55 and desired compound coding sequences.

The invention also provides a bacterial expression vector comprising a DNA sequence coding for the fusion protein.

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The expression vector typically contains, in addition to the fusion protein coding sequence, appropriate expression control sequences including a suitable promoter, an operator and a ribosome binding site and other appropriate regulatory sequences. Usually, also the expression vector contains one or more selectable markers.

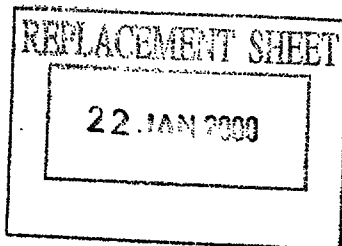
Any suitable promoter may be used, such as the TrpE, λ P1, λ Pr, lac or Tac promoter. Preferably the promoter is the bacteriophage T7 promoter and the expression vector is a plasmid. For E. coli expression an RI or Col-E1 plasmid-derived vector may be used. For example, expression vector pET 17B, obtainable from Novagen, or similar expression vectors may be used.

In another aspect the invention provides bacterial host cells transformed with a fusion protein coding sequence or an expression vector as described above. Any suitable bacterial host may be used, though preferably the host is E. coli. For example, E. coli strain BL21 (DE3) Lys E and similar strains may be used.

Advantageously the fusion protein accumulates within the host cells in the form of insoluble inclusion bodies, and thereby facilitates recovery of the fusion protein product from the cells. In order to recover the desired polypeptide product from the fusion protein, the protein of the inclusion bodies is solubilised, and the desired polypeptide product cleaved from the fusion protein. Any suitable solubilisation treatment may be used, including acid treatment, e.g. at about pH 2 to 4, preferably with acid at about pH 2.5, or treatment with a denaturing agent, e.g. a mild denaturing agent such as guanidine hydrochloride. In addition it may be necessary to remove one or more unwanted N-terminal amino acid residues from the product which remain after cleavage.

Thus in a further aspect the invention provides a process for the production of a desired compound of the invention, in a bacterial host, the process comprising: causing transformed bacterial host cells as defined above to express the fusion protein in the form of inclusion bodies; isolating the inclusion bodies; solubilizing the inclusion bodies, and cleaving the desired polypeptide from the fusion protein.

In the case of PTH compounds, it is not normally necessary to carry out carefully controlled denaturation and renaturation procedures to obtain PTH products in active form, e.g. having PTH-like activity. The medium-sized polypeptide products may be obtained in active form merely by neutralising the acid conditions used for solubilisation. Solubilisation and cleavage of the fusion protein may be carried out in a single step.



The invention also includes a fusion protein comprising an N-terminal bacteriophage T4 gene 55 polypeptide and, linked to the C-terminal thereof, a desired compound of the invention.

Using the processes and vectors of the invention it has been found that it is possible to produce high levels of PTH products. In *E. coli*, we have found that up to about 50% of the total protein expressed is the desired fusion protein. Also, the fusion protein forms inclusion bodies which are easy to separate from the other material of the cells. Moreover the inclusion bodies often consist of at least about 90% of the desired fusion protein which can significantly reduce the amount of purification that is required. Further, expressing the PTH product, in the form of the fusion protein can substantially decrease endogenous processing of the polypeptide in the bacterial cell, allowing homogenous products to be obtained.

Preferably the cleavable linker contains chemically cleavable amino acids adjacent the N-terminus of the desired polypeptide, to permit the desired polypeptide to be cleaved from the fusion protein by simple chemical means. Preferably the desired compound does not contain the chemically cleavable groups.

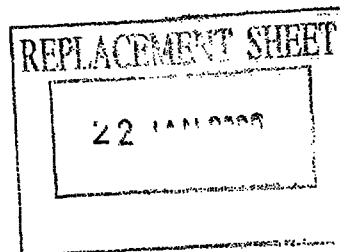
Preferred chemically cleavable linkers contain the amino acids Asp-Pro-Pro or Asn-Gly-Pro. The Asp-Pro or Asn-Gly bonds may be chemically cleaved using acid conditions e.g. formic acid (normally about pH 2.5) or hydroxylamine respectively. The dipeptides Pro-Pro or Gly-Pro that remain on the N-terminus of the desired polypeptide may then be removed using a suitable dipeptidyl-peptidase. For example the *L. lactis* X-Pro dipeptidyl-peptidase EC 3.4.14.5 can be used. This peptidase is described in Nardi et al; 1991; Applied and Environmental Microbiology, 57, p45 to 50.

The compound of the invention may be purified following cleavage from the fusion protein. HPLC purification techniques may be used. Purification may be carried out before removal of any unwanted N-terminal amino acid residues.

The process of the invention is further illustrated in the examples 25 to 31 which however only relate to fusion proteins comprising (1-38)PTH which are not comprised in the invention. SEQ 10 no. 1 shows the amino acid sequence and corresponding DNA sequence of a truncated gp55-Asp-Pro-ProhPTH(1-38)OH fusion protein as cloned into the plasmid pET17B, and SEQ 10 no.3 shows the amino acid sequence and corresponding DNA sequence of a full length gp55-Asn-Gly-Pro-hPTH(1-38)OH fusion protein as cloned into the plasmid pET17B.

In the following Examples all temperatures are in 0°C. The following abbreviations are employed. In the present specification the term PTH without any further indication includes the carboxy terminus as well as a carboxamide terminus.

BOC	tert. Butyloxycarbonyl
DMF	dimethylformamide
DCM	dichloromethane
Fmoc	9-fluorenylmethoxycarbonyl
HOBt	1-hydroxybenzotriazole
TFA	trifluoroacetic acid
Trt	trityl = triphenylmethyl
RP-HPLC	reverse phase high performance liquid chromatography
r.t.	room temperature
DNP	Dinitrophenyl
Tos	Tosyl
DIPC	Diisopropylcarbodiimide
Pmc	2,2,5,7,8-Pentamethylchroman-6-sulfonyl
IP	Isopropyl
For	Formyl
Cpe	1-amino-cyclopentane-1-carboxylic acid
Cpp	1-amino-cyclopropane-1-carboxylic acid
Cha	cyclohexylalanine
tBu	t.butyl

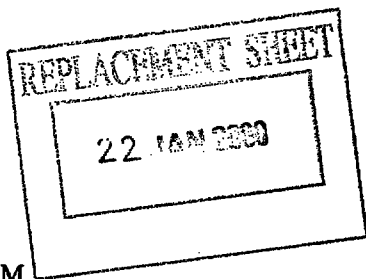


Comparative EXAMPLE 1: N^α--Isopropyl hPTH(1-34) NH₂

This peptide is assembled in a stepwise manner on a polystyrene based resin support. The Boc-group is used for protection of the alpha amino-group and side-chain functional groups are protected as follows: Lys(2-chlorobenzyloxycarbonyl), Ser(benzyl), Thr(benzyl), Glu(benzyl), Asp(benzyl), Met(O), His(DNP), Arg(Tos), and Trp(HCO). Other residues are left unprotected.

Amino-4-methylphenyl-methyl-co(polystyrene-divinylbenzene = MBHA resin) (0,7 mmol/g) is subjected to the following cycle, steps (1) to (7), of treatments:

- (1) DCM
- (2) trifluoroacetic acid (50 %) in DCM



- (3) DCM
- (4) diisopropylethylamine (10 %) in DMF
- (5) DMF
- (6) preformed symmetrical anhydride (1.4 mmol per g starting resin) of Boc-amino acid in DMF
- (7) DMF

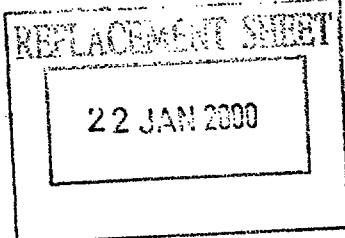
Volumes of washes and reagents are from 5 to 20 ml per gram of starting resin. Each step is repeated as many times as necessary for either complete reaction of the resin (steps 2, 4, 6) or complete displacement of the previous reagent from the resin (steps 1, 3, 5, 7). Samples of resin are removed after each cycle and checked for completeness of the coupling reaction by a colorimetric test for residual amino groups using ninhydrin.

Symmetrical anhydrides of Boc-amino acids are prepared just prior to use by reacting Boc-amino acid (2,8 mmol per g resin) and DCCI (1,4 mmol per g resin) in DCM, containing DMF in amounts sufficient for complete dissolution of the Boc-amino acid. The mixture is filtered, more DMF is added to the filtrate which is concentrated by evaporation of the volatile components at a temperature not exceeding 15°C and the resulting solution is used in step (6).

The cycle of reactions, (1) to (7), is repeated for each amino acid residue such as to provide the sequence of the title compound except for Boc-Gln-OH and Boc-Arg(Tos)-OH, which are coupled in step (6) as their preformed 1-hydroxybenzo-triazole esters in DMF.

The fully assembled peptide resin is treated twice with 20 ml of thiophenol A in DMF at r.t. for 1 hour and washed with DMF, methanol and dichloromethane. The peptide resin is again subjected to steps (1) to (5) followed by the addition of 0.5 ml of acetone, 84 mg of sodium cyanoborohydride and 0.2 ml of acetic acid in 20 ml of DMF per gram of resin. After 16 hours at RT the resin is washed with DMF and dichloromethane and dried.

The peptide resin is treated with liquid HF, dimethyl sulfide, p-cresol and ethane-1,2-dithiol according to the 'low-high' procedure which is described, e.g., in International Journal of Peptide and Protein Research, vol.26, page 262-273 (1985). After removal of the volatile components and washing with ethylacetate the residue is extracted with several portions of 1%-acetic acid, filtered and the filtrate lyophilized. The lyophilized product is purified by repeated reversed-phase chromatography on a column of octadecyl-silica using a gradient of acetonitrile in 2%-phosphoric acid or in 20 mM tetramethylammonium phosphate. Fractions containing the pure compound are combined, filtered



through a weakly basic ion-exchange resin in the acetate form and the filtrate lyophilized to give the title compound. MS (Ion-Spray): 4160.

Comparative EXAMPLE 2: [Lys (N^ε-Isopropyl)]^{26,27}hPTH(1-34) NH₂

The peptide is assembled as described for Example 1 but using Lys(Fmoc) for incorporation in position 13 and Fmoc-Ser(tBu) in terminal position 1.

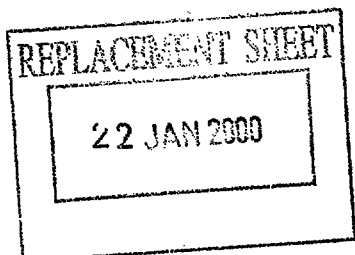
The fully assembled peptide resin is treated twice with 20 ml of thiophenol 5% in DMF at r.t. for 1 hour and washed with DMF, methanol and dichloromethane. The dry resin is treated with liquid HF as described for Example 1. The cleavage product (350 mg) is suspended in a mixture of methanol (5 ml), phosphate buffer pH 5.0 (5 ml), sodium cyanoborohydride (48 mg) and acetone (0.28 ml). After 16 hours at RT the reaction mixture is filtered and the filtrate evaporated. The residue is suspended in 20%-piperidine in DMF for 15 minutes, diluted with 20 volumes of ether and filtered. The residue is dissolved in 100 ml of water and acetic acid added to pH = 3, filtered and the filtrate lyophilized. The lyophilizate is purified by reversed-phase chromatography on a column of octadecyl-silica using a gradient of acetonitrile in 2%-phosphoric acid. Fractions containing the pure compound are combined, filtered through a weakly basic ion-exchange resin in the acetate form and the filtrate lyophilized to give the title compound. MS (Ion Spray): 4205.6.

Comparative EXAMPLE 3: N^α-Isopropyl [Lys (N^ε-Isopropyl)]^{13,26,27}hPTH-(1-38)-OH

a) Solid phase peptide synthesis

The peptide is synthesized in a stepwise manner on a polystyrene based resin support. The alpha-amino group is protected by Fmoc and the side-chain functional groups as followed: Asp(OtBu), Glu(OtBu), His(Trt), Lys(Boc), Asn(Trt), Gln(Trt), Arg(Pmc) and Ser(tBu). Other amino acids are left unprotected.

Fmoc-Gly esterified to 4-Hydroxymethyl-phenoxyethylco(polystyrene-1%-divinylbenzene), 0.5 mmol/g, is used as starting material for the stepwise solid phase synthesis, which consists of repetitive cycles of alpha-amino deprotection, washing, coupling (attachment of new amino acid) and washing. A three to five fold excess of Fmoc-amino acids is coupled as preformed HOBt-esters using DIPC. Fmoc-Arg(Pmc), Fmoc-Asn(Trt) and Fmoc-Gln(Trt) are coupled as symmetrical anhydrides using DIPC. After complete assembly of the peptide chain the Fmoc-protecting group of Ser¹ is removed by 20% piperidine/DMF. Cleavage of the peptide from the polystyrene resin and removal of



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all side chain protecting groups is obtained by treatment of the peptide resin with a mixture of 10% ethanedithiol, thioanisole, thiocresole and water in 90% TFA for three hours at room temperature. The resin particles are filtered off and washed with some TFA. The product is precipitated from the combined filtrates by addition of ether, filtered and dried. The product is purified by chromatography on a C-18 silica column using a gradient of acetonitrile in 2% H_3PO_4 /water. Fractions containing the pure compound are collected, filtered through an anion-exchange resin (Biorad, AG 4-X4, 100-200mesh acetate form) and lyophilized to give hPTH-(1-38)-OH.

b) Alkylation

40 mg (9 μ mol) hPTH-(1-38)-OH(4458.2 g/mol) are dissolved in 1.2 ml methanol, 1.2 ml phosphate buffer (Merck no. 9887 adjusted to pH 5.0), 45 mg (716 nmol) $NaBH_3CN$ (62.84 g/mol) and 0.8 ml (11 mmol) acetone (73.52 ml/mol). The reaction is monitored by RP-HPLC on a C-18 silica column and finished after 15 hours at room temperature. The reaction mixture is diluted with water and chromatographed on a C-18 silica column using a gradient of acetonitrile in 2% H_3PO_4 /water. Fractions containing the pure compound are collected, filtered through an anion-exchange resin (acetate form) and lyophilized to give the title compound as polyacetate, polyhydrate.

$[a]_D^{20} = 5.7^\circ$ (c= 0.317 in 95% AcOH).

MS(Ion-Spray): 4626

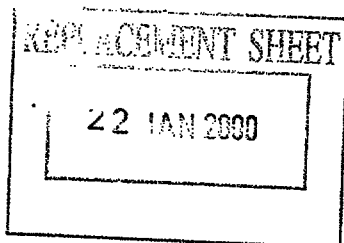
Comparative EXAMPLE 4: N α -Methyl [Ala¹]-PTH-(1-38)-OH

The peptide chain is assembled in the same manner as in Comparative example 3a. At position 1 instead of Fmoc-Ser(tBu)-OH the unnatural amino acid Fmoc-N $^{\alpha}$ -Methyl-Ala-OH is coupled to the peptide resin. Cleavage, deprotection and purification is performed as in example 3a.

MS (Ion Spray) = 4455,91

**Comparative EXAMPLE 5: [Ala¹,Ala³,Leu⁸,Gln¹³,Ala¹⁶,Gln¹⁸,Ala¹⁹,Phe²³,His²⁵
His²⁶,Leu²⁷,Thr³³,Ala³⁴]hPTH (1-34)OH**

This peptide is prepared in analogy with the procedure of comparative Example 3a. Instead of Fmoc-Gly esterified to 4-hydroxymethyl-phenoxymethyl-co(polystyrene-divinylbenzene) the appropriate Fmoc-amino acid, e.g. Fmoc-Ala, Fmoc-Phe, Fmoc-D-Ala, Fmoc-D-Phe, etc. is used. Additionally the side-chain functional groups are protected as follows: Thr(t.-butyl), Trp(Boc) and Tyr(t.-butyl).



By repeating the procedure disclosed in Examples 3 and 5 but using the appropriate starting materials, the following compounds may be obtained:

Example 1: [Leu⁸, Asp¹⁰, Ala¹⁶, Gln¹⁸, Thr³³]hPTH (1-34)OH

Example 2: [Leu⁸, Lys¹¹, Gln¹⁸]hPTH (1-36)OH

Example 3: [Leu⁸, Gln¹⁸, Thr³³, Ala³⁴]hPTH (1-34)OH (IS-MS: 4007)

Example 4: [Leu⁸, Ala¹⁶, Gln¹⁸, Ala¹⁹, Thr³³, Ala³⁴]hPTH (1-34)OH

Example 5: [Leu⁸, Ala¹⁶, Gln¹⁸, Thr³³, Ala³⁴]hPTH (1-34)OH (IS-MS: 3965)

Example 6: [Leu⁸, Asp¹⁰, Lys¹¹, Gln¹⁸]hPTH (1-36)OH

Comparative Example 7: [Leu⁸, Gln¹⁶, Asp¹⁷, Leu¹⁸, Arg¹⁹, Thr³³, Ala³⁴]hPTH (1-34)OH

Example 8: [Leu⁸, Asp¹⁰, Lys¹¹, Gln¹⁸, Thr³³, Ala³⁴]hPTH (1-34)OH (IS-MS: 4022)

Comparative Example 9: [Leu⁸, Asp¹⁰, Lys¹¹, Gln¹⁶, Asp¹⁷, Leu¹⁸, Arg¹⁹, Thr³³, Ala³⁴]hPTH (1-34)OH (IS-MS: 4077)

Example 10: [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸, Ala¹⁹]hPTH (1-36)OH (IS-MS: 4181)

Example 11: [Leu⁸, Ala¹⁶, Asp¹⁷, Gln¹⁸, Ala¹⁹]hPTH (1-36)OH (IS-MS: 4193)

Example 12: [Leu⁸, Ala¹⁶, Ala¹⁷, Gln¹⁸, Ala¹⁹]hPTH (1-36)OH (IS-MS: 4149)

Example 13: [Leu⁸, Ala¹⁷, Gln¹⁸, Ala¹⁹]hPTH (1-36)OH

Example 14: [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸, Thr³³, Ala³⁴]hPTH (1-34)OH (IS-MS: 3980)

Example 15: [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸]hPTH (1-36)OH (IS-MS: 4240)

Example 16: [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸, Ala¹⁹, Thr³³, Ala³⁴]hPTH (1-34)OH (IS-MS: 3923)

Example 17: [Leu⁸, Asp¹⁰, Ala¹⁶, Gln¹⁸]hPTH (1-36)OH (IS-MS: 4225)

Example 18: [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸, Ala¹⁹, Thr³³]hPTH (1-34)OH (IS-MS: 3999)

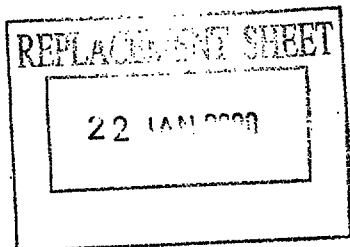
Example 19: [Leu⁸, Ala¹⁶, Gln¹⁸, Ala¹⁹]hPTH (1-36)OH (IS-MS: 4166)

Comparative Example 20: [Thr³³]hPTH-(1-38)-OH

Comparative EXAMPLE 6: [1-aminocyclopentane-1-carboxylic acid¹-hPTH-(1-36)NH₂

The peptide is assembled in a stepwise manner on a polystyrene based resin support. The Fmoc-group is used for protection of the alpha-amino groups.

Side-chain functional groups are protected as Arg(Pmc), Asn(Trt), Asp(OtBu), Gln(Trt), Glu(OtBu), His(Trt), Lys(Boc), Ser(tBu), Thr(tBu), Trp(Boc) and Tyr(tBu). Other amino acids are left unprotected.



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4-(2',4'-Dimethoxyphenyl-Fmoc-amino-methyl)-phenoxy-co(polystyrene-divinylbenzene), 0.4 mmol/g, which may be prepared, e.g., as described in Tetrah. Letters, 28, 3787-3790 (1987) is subjected to the following cycle, steps (1) to (5), of treatments:

- (1) DMF
- (2) piperidine (20%) in DMF
- (3) DMF
- (4) mixture of HOBt, diisopropylcarbodiimide, and Fmoc-alanine (0.8 mmol per gram starting resin each)
- (5) DMF

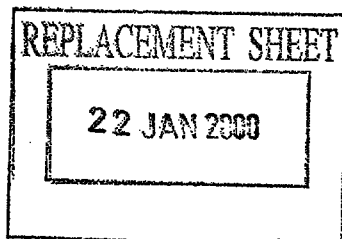
Volumes of washes and reagents are from 5 to 20 mL per gram of starting resin.

In the next cycle of treatments (1) to (5), Fmoc-valine is substituted for Fmoc-alanine and so on for each cycle such as to assemble on the resin the correct amino acid sequence of the title compound.

Each step is repeated as many times as necessary for either complete reaction of the resin (steps 2, 4) or complete displacement of the previous reagent(s) from the resin (steps 3, 5). Samples of the resin are removed after each cycle and checked for completeness of the coupling reaction by a colorimetric test for residual amino groups using ninhydrin.

At the end of the synthesis a final cycle comprising steps (1) to (3) only is performed and the peptide resin is washed with 2-propanol, then with a mixture of methanol and methylene chloride (1:1 v/v) and dried thoroughly in a vacuum desiccator.

The peptide resin (1 g) is suspended in a mixture (20 ml) of trifluoroacetic acid, water, and 1,2-ethanedithiol (90:5:5 v/v) for 2 hours at room temperature, the resin particles are filtered off and washed with a small volume of TFA. The product is precipitated from the combined filtrates by addition of ether (20 volumes), filtered, washed with more ether and dried. The residue is dissolved in 2 % acetic acid, the solution let to stand at r.t. for 8 hours prior to lyophilization. The lyophilisate is chromatographed on a C-18 silica column using a gradient of acetonitrile in 2% H₃PO₄. Fractions are checked by analytical HPLC and those containing the pure compound are collected, filtered through an anion-exchange resin in the acetate form and lyophilised to give the title compound as a polyacetate, polyhydrate.



Fmoc-1-aminocyclopentane-1-carboxylic acid used in the preparation of the peptide resin intermediate may be prepared, e.g. as described by G. Valle et al., 1988, in Can.J.Chem.66:2575-2582.

MS(Ion-Spray): 4312

By repeating the procedure as disclosed in Comparative Example 6 but using the appropriate starting materials, following compounds may be obtained:

Comparative Example 21: [Thr³³, Ala³⁴]hPTH (1-34)NH₂

Example 22: [Gln¹⁸]hPTH (1-36)NH₂

Comparative Example 23: [Ala³⁴]hPTH (1-36)NH₂ (IS-MS: 4210)

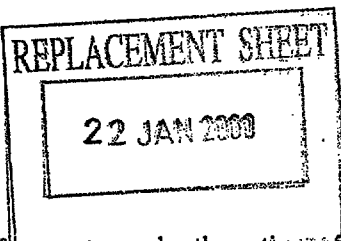
Comparative Example 24: [Leu⁸, Asp¹⁰, Lys¹¹, Thr³³, Ala³⁴]hPTH (1-34) NH₂

The compounds of Examples 1 to 24 show correct amino acid ratios in quantitative amino acid analysis.

In the following examples illustrating the recombinant process of the invention, if not otherwise specified, the standard procedures described in Ausubel et al; 1991; Current Protocols in Molecular Biology, John Wiley & Sons, New York are used. The fusion proteins obtained in this process comprising (1-28) PTH are not comprised in the invention.

Comparative EXAMPLE 25: Cloning bacteriophage T4 gene 55

Two DNA fragments, one containing the entire coding sequence of bacteriophage T4 gene 55, and the other fragment encoding part of it are amplified by polymerase chain reaction (PCR) using purified T4 DNA as the template. Oligonucleotides which contain NdeI or BamHI endonuclease recognition sequences are used as primers. The PCR reactions (25 cycles, each 30" 94°C, 30" 50°C, 30" 72°C) are performed with 1 ng of template and 100 pM of the upstream primer (GAGGTGCATA TGTCAGAAAC TAAGCCT 27), the NdeI recognition site being provided by nucleotides 7-12 thereof) and 100 pM of the downstream primer (GGTCGGATCC ATCGTTAGCG TTAGCCTCAT ATAAAAAATC 40 the BamHI, recognition site being provided by nucleotides 5-10 thereof) in 100 µl reaction buffer containing 10 mM Tris-HCl, 50 mM KCl, 1.5 mM MgCl₂, 0.2 mM of each of dATP, dCTP, dGTP, dTTP, and 0.1% Triton X 100, pH 9.0. A 0.6kb fragment is obtained. The truncated form of gene 55 is prepared by PCR as described for the entire gene, except that a different oligonucleotide is used as downstream primer (GGTCGGATCC TACGTTGGAC GAATGCAT 28, the BamHI recognition site being provided by nucleotides 5-10 thereof). A 0.35 kb fragment is obtained. The 0.6 kb and 0.35 kb DNA fragments are digested with restriction endonucleases NdeI/BamHI and are cloned into a NdeI/BamHI digest of expression vector pET17B (obtained from Novagen). The 0.6



Kb DNA fragment encodes the entire gp55 protein (188 amino acid residues – see SEQ 10 no. 1), whereas the 0.35 kb fragment encodes a 12 kD amino-terminal fragment of gp55 (112 amino acid residues – see Fig. 2).

Comparative Example 26: Preparation and cloning of the DNA encoding PTH (1-38)

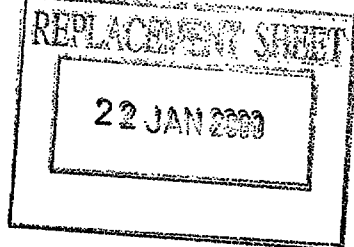
DNA fragments encoding a) the linker peptide corresponding to amino acid residues 110-115 of SEQ 10 no. 1 and PTH (1-38) or b) the linker peptide corresponding to amino acid residues 186-191 of SEQ 10 no. 3 and PTH (1-38) are generated by PCR using a cloned synthetic PTH (1-38) coding sequence as a template.

The oligonucleotides encoding the linker a) (GAGTGATCC ATCGATCCAC CATCCGTATC AGAAATACAA CT 42), or encoding linker b GAGTGGATCC GTTAACGGTC CATCCGTATC AGAAATACAA CT 42 are used as upstream primers. The downstream primer TCTACTCGAG TTAACCCAGA GCTACAAAAT TATG 34

The upstream primers each incorporate a BamHI cloning site and the downstream primer contains a XhoI recognition site (nucleotides 5-10 of each sequence). The PCR reaction is performed under the same conditions as described in Example 25. After cleavage with BamHI and XhoI, the 0.14 kb PCR products are inserted into BamHI-XhoI digests of the gp55-pET17B plasmids, obtained in Example 25. For expression of the fusion proteins the resultant plasmid constructs are transformed into E.coli BL21 (DE3) LysE and single bacterial colonies are further propagated. The expression of the fusion proteins is obtained by incubating a preculture at 37°C on a rotary shaker overnight in circle growth medium (Bio 101). A main culture is inoculated with the preculture making up 1 to 5 % of the volume of the main culture. The main culture is then fermented at a temperature of 37°C and at a pH of 6.9 to 7.1 whilst being aerated at 10 l/min and agitated at 200-400 rpm. Expression is induced by the addition of 1 mM IPTG as soon as the culture reaches an OD₅₅₀ of about 1.0 to 5.0. After induction the fermentation is continued for five hours and then the fermentation broth is harvested without killing the *E. coli* cells. The harvested cells are spun down with a tubular centrifuge and resuspended in sample buffer and assayed for expression by SDS PAGE. The cell pellet is then frozen until purification. The procedures and host cells used are fully described in Studier et al; 1990; "Use of T7 RNA Polymerase to direct expression of Cloned Genes", Methods in Enzymology Academic Press. pages 60 to 89.

Comparative EXAMPLE 27: Preparation of inclusion bodies

Frozen E. coli pellets obtained from Example 26 are resuspended to 25% w/v in Buffer A (50mM Tris PH 8.0; containing 2mM DTT, 5mM Benzamidine-HCl, 1.5mM MgCl₂, 1.0mM MnCl₂, 10pg/ml. DNase I and



2mg/ml Lysozyme) and mixed by stirring for 1 hour at room temperature. The resuspended cells are lysed by passage through a Manton-Gaulin homogeniser (2 passes at 1200 bar) and the resultant lysate kept on ice. The lysate is diluted 2 fold with ice-cold buffer B (50mM Tris pH 8.0; containing 2mM DTT, 5mM Benzamidine-HCl and 4mM EDTA) and centrifuged for 30min at 27,500g. The supernatant from the centrifuge is carefully removed and discarded. The pellet is resuspended in buffer B to 25% w/v, passed once more through the Manton-Gaulin and centrifuged as before. This process is repeated once using buffer B and once using water. The resultant inclusion body pellets are weighed and frozen at -20°C until required.

Comparative EXAMPLE 28: Solubilisation and cleavage of gp55-Asp-Pro-Pro-(1-38)hPTH inclusion bodies

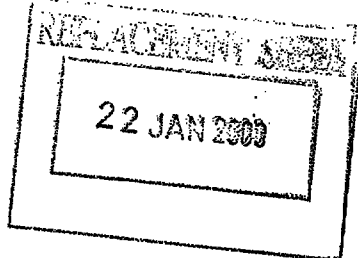
Frozen inclusion bodies containing gp55-Asp-Pro-Pro-(1-38)hPTH as obtained in Example 27 are suspended in 10 mM HCl acid solution (analytical grade) to a final concentration of 6%w/v. The solution is incubated for 24 hours at 50°C and the reaction terminated by addition of 1 volume of aqueous 100 mM NaOAc solution. The diluted supernatant is clarified by centrifugation at 27,500g for 30 minutes and filtered through a glass fibre filter.

Comparative EXAMPLE 29: Solubilisation and cleavage of gp55-Asn-Gly-Pro-(1-38)hPTH inclusion bodies

Frozen inclusion bodies containing gp55-Asn-Gly-Pro-(1-38)hPTH and obtained from example 27 are dissolved in 6M Guanidine-HCl (containing 2M Hydroxylamine-HCl and brought to pH 9.0 by addition of 4.5M LiOH) to give an estimated 5mg/ml protein. The pH is readjusted to pH 9 by further addition of LiOH and the solution incubated for 4 hours at 45°C. The reaction is terminated by addition of formic acid to pH 4 and the solution centrifuged and filtered as described in example 28.

Comparative EXAMPLE 30: Preparative HPLC of Gly-Pro and Pro-Pro (1-38)hPTH

Filtered cleavage supernatants are analysed on analytical reversed phase HPLC (Orpogen HD-Gel-RP-7s-300; 4 x 150mm column) and compared with a known (1-38)hPTH standard (Bachem) to determine the PTH concentration. The column is equilibrated with 85% HPLC buffer A (90% water, 10% acetonitrile; containing 0.1%v/v TFA) and 15% HPLC buffer B (10% water, 90% acetonitrile; containing 0.1% TFA) and is eluted with a gradient of 15% HPLC buffer B to 100% HPLC buffer B over 15 minutes at a flow rate of 0.75ml/min. Depending upon the scale of the preparation, supernatants are either loaded on a 1.0 x 25cm C4 Vydac or a 2.2 x 25cm C4 Vydac. The columns are equilibrated with 85% HPLC buffer A and 15% HPLC buffer B at flow rates of 4ml/min and



10ml/min respectively. The columns are eluted with gradients of 15%B to 85%B over 80ml and 15%B to 55%B over 300ml respectively. Gly-Pro-hPTH(1-38) or Pro-Pro-hPTH(1-38) peaks are collected manually and the acetonitrile is later removed under vacuum. Analytical HPLC is used to determine the peptide concentration both after cleavage and in the collected fractions. The solvent free peptide solutions are diluted with an equal volume of 100mM sodium acetate and the pH adjusted, if necessary, to pH 5.4. Following filtration (0.2µm), the solution is loaded on a cation exchange column (either Mono S or SP-Sepharose High Performance, packed into HR10/10 or XK16/10 columns respectively) equilibrated with 50mM sodium acetate pH 5.4 (buffer C). The column is eluted with a 0 to 500mM NaCl gradient in buffer C over 7.5 column volumes. 10ml fractions are collected and the X-X-(1-38)hPTH peak is pooled. The peptide concentration is determined as before using HPLC. This column serves both to exchange the peptides into a more physiological buffer and to remove additional peptides produced by chemical cleavage of the fusion protein itself.

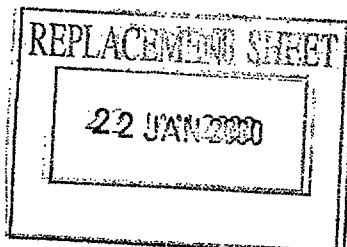
Comparative EXAMPLE 31: Enzymatic removal of X-X from X-X-(1-38)hPTH

Pooled peptide fractions, as obtained in Example 30, (concentration range 1.5 - 2.0 mg/ml) are brought to pH 8.0 by addition of solid Tris. Purified X-Prolyl dipeptidase (0.5mg/ml in PBS pH 7.2) (Nardi et al. (1991) App. Env. Microbiol. 57, No.1, 45-50) is added to 1:1000 and the solution is incubated for 24h at 37 C. The reaction is stopped by addition of TFA to 0.2% (pH = 5.0) and the resultant (1-38)hPTH is isolated using preparative HPLC (using the same conditions as described in Example 30). The solvent is removed and the product is lyophilised after removal of aliquots for N-terminal sequence and mass spectroscopic analyses.

Examples of the yields which are obtained using the two fusion protein approaches described above are as follows:

Gen55-Asp-Pro-Pro-(1-38)PTH (using the "truncated" form of Gen55)

Under partially optimized fermentation conditions, a 215 g wet cell pellet is obtained from a 10 litre fermentation. The percentage level of expression of the fusion protein is approximately 37%. From this pellet, 31 g of inclusion bodies are isolated. The protein purity (with respect to the fusion protein) of these bodies is 61%. After solubilisation, cleavage and centrifugation/filtration approximately 620mg Pro-Pro-PTH are detected. This material yields 580mg Pro-Pro-PTH after subsequent C4-Vydac HPLC. After dilution, cation-exchange on Mono S, digestion with X-Prolyldipeptidase (at a Pro-Pro-PTH concentration of 2mg/ml), HPLC and lyophilisation, 440mg of lyophilized pure product



with full biological activity is obtained. This represents a yield of 71% between the stopped cleavage step and the final product.

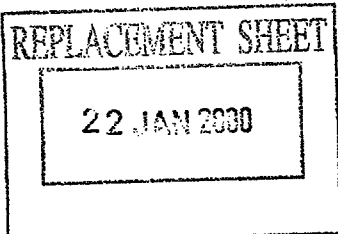
Gen55-Asn-Gly-Pro-(1-38)PTH (using the "long" form of Gen55)

From a non-optimized 20 litre fermentation a 78g wet cell pellet with 50% expression of the fusion protein is harvested. Following lysis and centrifugation, 18g of "90% pure" inclusion bodies are obtained. Following the acid stop step and centrifugation/filtration about 470mg Gly-Pro-PTH are detected. After HPLC on C4-Vydac, 355mg of peptide are detected, decreasing to 321mg after SP-Sepharose cation-exchange. Enzymatic removal of the Gly and Pro residues is carried out at 2mg/ml. After the final HPLC step and lyophilization, 244mg of pure, fully biologically active (1-38)PTH is recovered representing a yield of 52% from the stopped cleavage reaction stage.

The compounds of the invention in free form or in the form of pharmaceutically acceptable salts and complexes exhibit valuable pharmacological properties as indicated in animal tests and are therefore indicated for therapy.

The biological activity of the compounds of the invention is assessed in vitro by measuring their ability of stimulating the synthesis of cyclic AMP in UMR 106-06 rat and SaOS-2 human osteosarcoma cells according to the method of Marcus and Aurbach in *Endocrinology*, 85, 801-810 (1969). Rat osteosarcoma UMR 106 cells are grown to confluence in Eagle's Minimum Essential Medium - 10% FCS in 12 well plates, human SaOS-2 osteosarcoma cells are grown in McCoy's 5A medium - 10% FCS. The medium is then changed to medium with 2% FCS and 1-5 pCi/well [3H]-adenine is added. Two hours later, cells are washed twice with serum-free medium and incubated in DMEM - 1% BSA containing 1 mM 3-isobutyl-1-methylxanthine to exclude actions on phosphodiesterases. Test substances are added 20 min later. The reaction is stopped and cAMP extracted after 15 min by adding ice cold 5% trichloroacetic acid. A carrier solution (0.5 ml/well) containing 5 mM of unlabelled adenine, adenosine, AMP, ADP, ATP, and cAMP as well as 0.4 μ Ci [14C]-adenosine for determination of recovery is added. [M]-cAMP is separated using serial Dowex 50W-X4 (200-400 mesh) and alumina chromatography and counted. Results are calculated in % of solvent control and EC_{50} values determined from DRC curves. Compounds of the invention stimulate cAMP production in UMR 106-06 rat and SaOS-2 human osteosarcoma cells at a concentration of 10^{-11} to 10^{-6} M.

The compounds of the invention also have binding affinity to PTH receptors, e.g. as follows:



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Chicken [Tyr³⁶]PTHrP(1-36)amide is iodinated to a specific activity of 2,200 Ci/mmol using the lactoperoxidase method (Anawa Lab. AG, Wangen). Monolayers of opossum kidney cells are washed with 200 μ l DMEM and HAM's F12 (1:1) containing 1% BSA and incubated at 16°C with 50,000cpm of [I¹²⁵-Tyr³⁶]chPTHrP(1-36)amide per well in the presence or absence of 1 μ M [Tyr³⁶]chPTHrP(1-36)amide. After incubation, cells are washed with 0.5 ml medium (4°C), lysed with 0.5 ml 1N NaOH and radioactivity is determined. Specific binding is defined as total binding minus nonspecific binding. Competition curves are analyzed using SCTFIT, a non-linear regression computer program (Feyen et al, 1992, Biochem. Biophys. Res. Commun. 187:8-13) and data presented as mean pK_D values (n=2 to 3). Compounds of the invention show in this test a binding affinity expressed as a mean pK_D value of from 7 to 10.

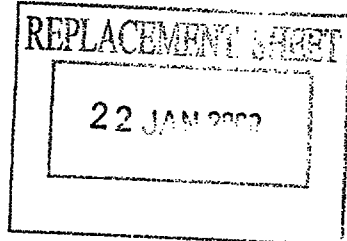
Furthermore, the compounds of the invention increase plasma calcium level after continuous s.c. infusion, e.g. as determined in male thyroparathyroidectomized Wistar rats weighing 140 to 170 g. 5 days after thyroparathyroidectomy, rats are implanted in the neck with Alzet minipumps and test compounds infused at rates of 0.3 to 30 μ g/day/rat. Blood is drawn by retro-orbital puncture in the morning of days 1, 2, 3, and 6 thereafter and plasma calcium concentrations are determined photometrically. In this test compounds of the invention increase plasma calcium.

The compounds of the invention are accordingly indicated for preventing or treating all bone conditions which are associated with increased calcium depletion or resorption or in which calcium fixation in the bone is desirable, e.g. osteoporosis of various genesis (e.g. juvenile, menopausal, post-menopausal, post-traumatic, caused by old age or by cortico-steroid therapy or inactivity), fractures, osteopathy, including acute and chronic states associated with skeletal demineralisation, osteo-malacia, periodontal bone loss or bone loss due to arthritis or osteoarthritis or for treating hypoparathyroidism.

The compounds of the invention are particularly indicated for preventing or treating osteoporosis of various genesis.

For all the above uses, an indicated daily dosage is in the range from about 0.003 to about 10 mg preferably 0.003 to 3, more preferably 0.01 to 1 mg of a compound of the invention. Compounds of the invention may be administered once a day or up to twice a week.

The compounds of the invention may be administered in free form or in pharmaceutically acceptable salt form or complexes. Such salts and complexes may be prepared in conventional manner and exhibit the same order of activity as the free compounds. The present invention also provides a pharmaceutical composition comprising a compound of the invention in free base form or in pharmaceutically acceptable salt form or complex form in



association with a pharmaceutically acceptable diluent or carrier. Such compositions may be formulated in conventional manner. The compounds of the invention may be administered by any conventional route, for example parenterally e.g. in form of injectable solutions or suspensions, enterally, e.g. orally, for example in the form of tablets or capsules or in a transdermal, nasal or a suppository form.

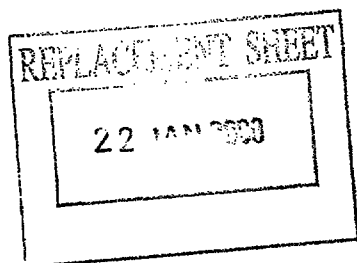
In accordance with the foregoing the present invention further provides:

- a) a compound of the invention or a pharmaceutically acceptable salt or complex thereof for use as a pharmaceutical;
- b) a pharmaceutical composition for improving bone formation, e.g. preventing or treating all bone conditions which are associated with increased calcium depletion or resorption or in which calcium fixation in the bone is desirable, e.g. osteoporosis of various genesis (e.g. juvenile, menopausal, post-menopausal, post-traumatic, caused by old age or by cortico-steroid therapy or inactivity), fractures, osteopathy, including acute and chronic states associated with skeletal demineralisation, osteomalacia, periodontal bone loss or bone loss due to arthritis or osteoarthritis, or for treating hypoparathyroidism in a subject in need of such treatment; or
- c) a compound of the invention or a pharmaceutically acceptable salt or complex thereof for use in the preparation of a pharmaceutical composition for use in the method as in b) above.

According to a further embodiment of the invention, the compounds of the invention may be employed as adjunct or adjuvant to other therapy, e.g. a therapy using a bone resorption inhibitor, for example as in osteoporosis therapy, in particular a therapy employing calcium, a calcitonin or an analogue or derivative thereof, e.g. salmon, eel or human calcitonin, a steroid hormone, e.g. an estrogen, a partial estrogen agonist or estrogen-gestagen combination, a biphosphonate, vitamin D or an analog thereof, or any combination thereof.

When the compounds of the invention are administered in conjunction with, e.g. as an adjuvant to bone resorption inhibition therapy, dosages for the co-administered inhibitor will of course vary depending on the type of inhibitor drug employed, e.g. whether it is a steroid or a calcitonin, on the condition to be treated, whether it is a curative or preventive therapy, on the regimen and so forth.

In accordance with the foregoing the present invention provides in a yet further aspect:



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d) a pharmaceutical composition for improving bone formation, e.g. preventing or treating calcium depletion, for example for preventing or treating any specific conditions or diseases hereinbefore set forth, in a subject in need of such treatment comprising a) a compound of the invention and b) a second drug substance, said second drug substance being a bone resorption inhibitor, for example as indicated above.

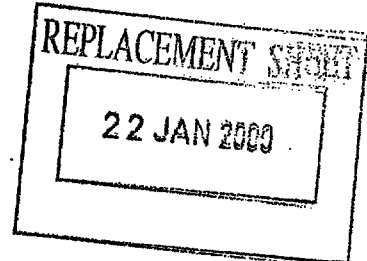
Compounds of Examples 3,5,8,14 and 15 are preferred.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT:

(A) NAME: SANDOZ LTD
 (B) STREET: PO BOX
 (C) CITY: BASEL
 (D) STATE: BS
 (E) COUNTRY: SWITZERLAND
 (F) POSTAL CODE (ZIP): CH 4002
 (G) TELEPHONE: 061 324 1111
 (H) TELEFAX: 061 322 7572
 (I) TELEX: 965 050 55



(ii) TITLE OF INVENTION: Organic Compounds

(iii) NUMBER OF SEQUENCES: 12

(iv) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: Floppy disk
 (B) COMPUTER: IBM PC compatible
 (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 (D) SOFTWARE: PatentIn Release #1.0, Version #1.25 (EPO)

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 717 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

(v) FRAGMENT TYPE: internal

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Bacteriophage T4/Homo sapiens

(ix) FEATURE:

(A) NAME/KEY: CDS
 (B) LOCATION: 101..559

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Asp Trp Lys Thr Glu Leu Ala Asn Asn Lys Asp Pro Asn Lys Val Val	
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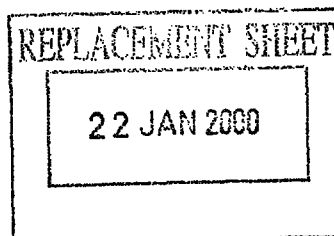
(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 153 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

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Lys	Gly	Leu	His	Asn	Phe	Asp	Glu	Thr	Lys	Tyr	Lys	Asn	Pro	His	Ala		
			85					90						95			
Tyr	Ile	Thr	Gln	Ala	Cys	Phe	Asn	Ala	Phe	Val	Gln	Arg	Gly	Ser	Ile		
			100					105					110				



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(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 945 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

(v) FRAGMENT TYPE: internal

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Bacteriophage T4/Homo sapiens

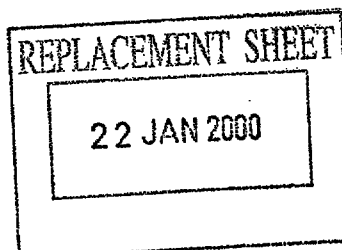
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(A) NAME/KEY: CDS

(B) LOCATION: 101..787

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Met Ser Glu Thr Lys	5
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CCT AAA TAT AAT TAC GTA AAC AAT AAA GAG CTT TTA CAA GCT ATT ATT	163
Pro Lys Tyr Asn Tyr Val Asn Asn Lys Glu Leu Leu Gln Ala Ile Ile	20
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Asp Trp Lys Thr Glu Leu Ala Asn Asn Lys Asp Pro Asn Lys Val Val	35
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Glu Met Ile Ala Asp Gly Ile Glu Ala Ser Ile Lys Gly Leu His Asn	85
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105	
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(2) INFORMATION FOR SEQ ID NO:4:

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 (A) LENGTH: 229 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

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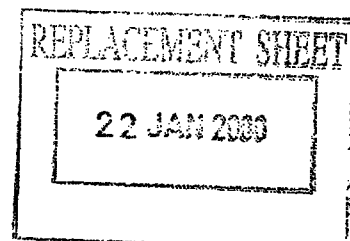
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 85 90 95

Tyr Ile Thr Gln Ala Cys Phe Asn Ala Phe Val Gln Arg Ile Lys Lys
 100 105 110



Glu Arg Lys Glu Val Ala Lys Lys Tyr Ser Tyr Phe Val His Asn Val
 115 120 125
 Tyr Asp Ser Arg Asp Asp Asp Met Val Ala Leu Val Asp Glu Thr Phe
 130 135 140
 Ile Gln Asp Ile Tyr Asp Lys Met Thr His Tyr Glu Glu Ser Thr Tyr
 145 150 155 160
 Arg Thr Pro Gly Ala Glu Lys Lys Ser Val Val Asp Asp Ser Pro Ser
 165 170 175
 Leu Asp Phe Leu Tyr Glu Ala Asn Asp Gly Ser Val Asn Gly Pro Ser
 180 185 190
 Val Ser Glu Ile Gln Leu Met His Asn Leu Gly Lys His Leu Asn Ser
 195 200 205
 Met Glu Arg Val Glu Trp Leu Arg Lys Lys Leu Gln Asp Val His Asn
 210 215 220
 Phe Val Ala Leu Gly
 225

(2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 27 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

(v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

GAGGTGCATA TGTCAGAAAC TAAGCCT

27

(2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 40 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

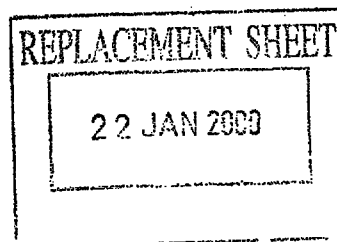
(iii) ANTI-SENSE: NO

(v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

GGTCGGATCC ATCGTTAGCG TAGCCTCAT ATAAAAATC

40



(2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 28 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA

- (iii) HYPOTHETICAL: NO

- (iii) ANTI-SENSE: NO

- (v) FRAGMENT TYPE: internal

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

GGTCGGATCC TACGTTGGAC GAATGCAT

28

(2) INFORMATION FOR SEQ ID NO:8:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 42 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA

- (iii) HYPOTHETICAL: NO

- (iii) ANTI-SENSE: NO

- (v) FRAGMENT TYPE: internal

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

GAGTGGATCC ATCGATCCAC CATCCGTATC AGAAATACAA CT

42

(2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 42 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA

- (iii) HYPOTHETICAL: NO

- (iii) ANTI-SENSE: NO

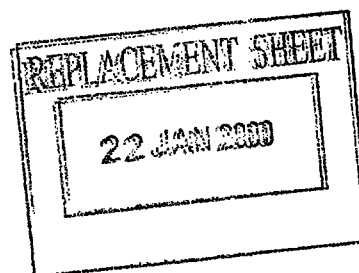
- (v) FRAGMENT TYPE: internal

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

GAGTGGATCC GTTAACGGTC CATCCGTATC AGAAATACAA CT

42

(2) INFORMATION FOR SEQ ID NO:10:



- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 34 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

(v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

TCTACTCGAG TTAACCCAGA GCTACAAAAT TATG

34

(2) INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 34 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Homo sapiens

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

Ser Val Ser Glu Ile Gln Leu Met His Asn Leu Gly Lys His Leu Asn
 1 5 10 15

Ser Met Glu Arg Val Glu Trp Leu Arg Lys Lys Leu Gln Asp Val His
 20 25 30

Asn Phe

(2) INFORMATION FOR SEQ ID NO:12:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 36 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Homo sapiens

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Ser Val Ser Glu Ile Gln Leu Met His Asn Leu Gly Lys His Leu Asn
 1 5 10 15
 Ser Met Glu Arg Val Glu Trp Leu Arg Lys Lys Leu Gln Asp Val His
 20 25 30
 Asn Phe Val Ala
 35

(2) INFORMATION FOR SEQ ID NO:13:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 38 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

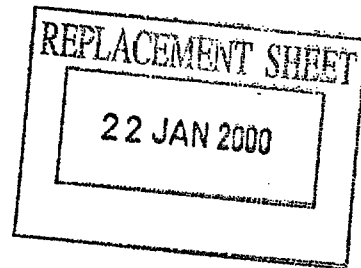
(iii) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Homo sapiens

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

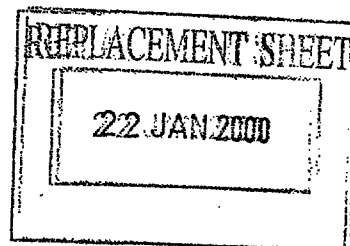
Ser Val Ser Glu Ile Gln Leu Met His Asn Leu Gly Lys His Leu Asn
 1 5 10 15
 Ser Met Glu Arg Val Glu Trp Leu Arg Lys Lys Leu Gln Asp Val His
 20 25 30
 Asn Phe Val Ala Leu Gly
 35



CLAIMS

1. A Compound of formula I

Ser-Val-Ser-Glu-Ile-Gln-Leu-R₁-His-R₂-R₃-Gly-R₄-His-Leu
 R₅-R₆-R₇-R₈-Arg-Val-Glu-Trp-Leu-Arg-R₉-Lys-Leu-Gln-Asp-
 Val-His-R₁₀-R₁₁-(R₁₂)_n-X



wherein

R₁ is Met or Leu;

R₂ is Asn, Asp, Gly, Gln, Glu, His, Ser, Thr or Tyr;

R₃ is Leu or Lys;

R₄ is Lys or D- or L-Ala, -Cys, -Gln, -Ile, -Asn, -Trp, -Asp, -Val, -Thr, -Ser, -Tyr, -Met, -Leu or -Gly;

R₅ is Asn, Gln, D-Gln or D- or L-Lys, -Ser, -Leu, -Ala or -Gly;

R₆ is Ser, Asp, Ala, Glu, Gln, Phe, His, Ile or Lys;

R₇ is Gln

R₈ is Glu, Arg, Ala, Val, Tyr, Ser, Lys, Met, His, Gly, Pro, Asn or Ile;

R₉ is Lys, Gln or Arg;

R₁₀ is Asn, Thr, D-Thr or D- or L-Ser, -Leu, -Gly, -Gln, -Arg, -Pro, -Asp, -Ile or -Lys;

R₁₁ is Phe or Ala;

R₁₂ is Val, Val-Ala, Val-Ala-Leu or Val-Ala-Leu-Gly;

n is 0 or 1; and

X is OH, OR_a, NH₂ or NR_bR_c wherein R_a is C₁₋₄alkyl, one of

R_b and R_c is H and the other is C₁₋₆alkyl,

the compound optionally comprising at least one radical selected from a L- or D-α-amino acid, C₂₋₆alkoxy-carbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to its terminal amino group, and/or

the compound optionally comprising at least one radical selected from C₂₋₆alkoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl attached to one or more side chain amino groups of the compound, in free form or in salt or complex form.



2. A compound according to claim 1 wherein R₁₀ is Thr.

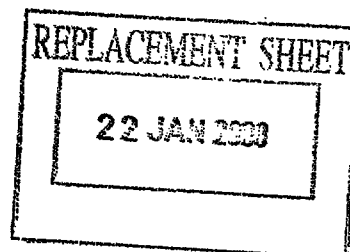
3. A compound according to claim 1 or 2, wherein R₁₁ is Ala.

4. A compound selected from [Leu⁸, Gln¹⁸, Thr³³, Ala³⁴]hPTH, [Leu⁸, Ala¹⁶, Gln¹⁸, Thr³³, Ala³⁴]hPTH, [Leu⁸ Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸, Thr³³, Ala³⁴]hPTH, [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸]hPTH, [Leu⁸, Ala¹⁶, Gln¹⁸, Ala¹⁹]hPTH, [Leu⁸, Asp¹⁰, Lys¹¹, Gln¹⁸]hPTH [Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸, Ala¹⁹]hPTH [Leu⁸, Ala¹⁶, Gln¹⁸, Ala¹⁹, Thr³³, Ala³⁴]hPTH and [Leu⁸, Asp¹⁰, Lys¹¹, Gln¹⁸, Thr³³, Ala³⁴]hPTH the compound optionally comprising at least one radical selected from a L- or D-α-amino acid, C₂₋₆alkoxy-carbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to its terminal amino group, and/or at least one radical selected from C₂₋₆alkoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to one or more side chain amino groups of the compound, in free form or in salt or complex form.

A compound selected from

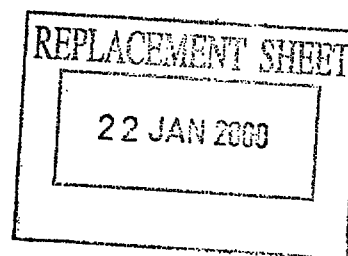
[Leu⁸, Gln¹⁸, Thr³³, Ala³⁴] 1-hPTH(1-34)OH
[Leu⁸, Ala¹⁶, Gln¹⁸, Ala¹⁹, Thr³³, Ala³⁴]hPTH(1-34)OH
[Leu⁸, Ala¹⁶, Gln¹⁸, Thr³³, Ala³⁴]hPTH(1-34)OH
[Leu⁸, Asp¹⁰, Lys¹¹, Gln¹⁸]hPTH(1-36)OH
[Leu⁸, Asp¹⁰, Lys¹¹, Gln¹⁸, Thr³³, Ala³⁴]hPTH(1-34)OH
[Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸, Ala¹⁹]hPTH(1-36)OH
[Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸, Thr³³, Ala³⁴]hPTH(1-34)OH
[Leu⁸, Asp¹⁰, Lys¹¹, Ala¹⁶, Gln¹⁸]hPTH(1-36)OH, and
[Leu⁸, Ala¹⁶, Gln¹⁸, Ala¹⁹]hPTH(1-36)OH

in free form or in salt or complex form.



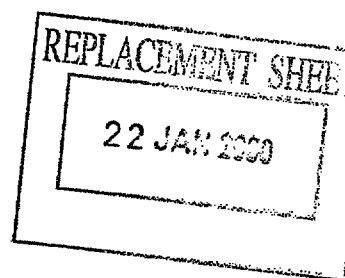
6. A process for the production of a compound according to any one of the preceding claims in which a fusion protein comprising a N-terminal bacteriophage T4 gene 55 polypeptide having the desired compound linked to the C-terminal thereof, is expressed in bacterial cells, and, where required, introducing a C₂₋₆alkoxy-carbonyl or optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl on the terminal and/or side chain amino group of the compound in protected or unprotected form and, if required, removing the protecting group present in the protected form.

7. A process according to claim 6 in which the fusion protein comprises a chemically cleavable linker between the gene 55 and the desired compound.
8. A process according to claim 7 in which the cleavable linker comprises the amino acid sequences Asp-Pro-Pro or Asn-Gly-Pro.
9. A nucleotide sequence which codes for a fusion protein comprising an N-terminal bacteriophage T4 gene 55 polypeptide and a desired compound according to any one of the claims 1 to 5, linked to the C-terminal thereof.
10. A bacterial expression vector comprising a DNA sequence according to claim 9.
11. Bacterial host cells transformed with a DNA sequence according to claim 9, or a bacterial vector according to claim 10.
12. A fusion protein comprising an N-terminal bacteriophage T4 gene 55 polypeptide and a desired compound according to any of the claims 1 to 5, linked to the C-terminal thereof.
13. A compound according to any one of claims 1 to 5 in free form or in pharmaceutically acceptable salt form for use as a pharmaceutical.
14. A pharmaceutical composition comprising a compound as defined in any one of claims 1 to 5 in free form or in pharmaceutically acceptable salt form, together with a pharmaceutically acceptable diluent or carrier therefor.
15. A compound according to any one of claims 1 to 5, in free form or in pharmaceutically acceptable salt or complex form, for use in the preparation of a pharmaceutical composition for use in improving bone formation.
16. A compound according to any one of claims 1 to 5, in free form or in pharmaceutically acceptable salt or complex form, for use in association with a bone resorption inhibitor to improve bone formation.



ANALOGS OF PTH

Abstract of the invention



PTH compounds having PTH-like activity and comprising at least one modification, said modification being either

1. at least one radical selected from a L- or D- α -amino acid, C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to the terminal amino group of the PTH compound, and/or at least one radical selected from C₂₋₆alcoxycarbonyl and optionally substituted C₁₋₈alkyl, C₂₋₈alkenyl, C₂₋₈alkynyl, aralkyl, aralkenyl or C₃₋₆cycloalkyl-C₁₋₄alkyl and attached to one or more side chain amino groups of the PTH compound,

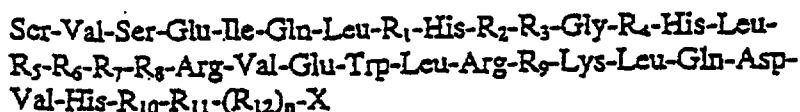
or

2. at least one α -amino acid unit in the positions 1 to 38 of a naturally occurring PTH sequence being replaced by a natural or unnatural amino acid unit optionally in protected form, whereby the α -amino acid units present in positions 1 and 2 at the amino terminus of the PTH sequence may be replaced by a pseudo-peptide,

or a combination of such modifications,

in free form or in salt form, have pharmacological activity, e.g. for preventing or treating all bone conditions which are associated with increased calcium depletion or resorption or in which calcium fixation in the bone is desirable.

A representative compound is e.g. of formula 1



wherein each of R₁ to R₁₁ has a meaning within the invention, R₁₂ is Val, Val-Ala, Val-Ala-Leu or Val-Ala-Leu-Gly; n is 0 or 1; and X is OH, NH₂ or substituted OH or NH₂, provided that R₇ is Gln or Tyr, and/or R₁₀ is Thr and/or R₁₁ is Ala.

- 1 -

REPLACEMENT SHEET

FIGURE 1

AGATCTCGAT CCCGCGAAAT TAATACGACT CACTATAGGG AGACCACAAC GGTTCCTC 60

TAGAAATAAT TTTGTTTAAC TTTAAGAAGG AGATATACAT ATG TCA GAA ACT AAG 115
Met Ser Glu Thr Lys
1 5

CCT AAA TAT AAT TAC GTA AAC AAT AAA GAG CTT TTA CAA GCT ATT ATT 163
Pro Lys Tyr Asn Tyr Val Asn Asn Lys Glu Leu Leu Gln Ala Ile Ile
10 15 20

GAT TGG AAA ACA GAA TTA GCA AAT AAT AAA GAC CCA AAT AAA GTA GTT 211
Asp Trp Lys Thr Glu Leu Ala Asn Asn Lys Asp Pro Asn Lys Val Val
25 30 35

CGT CAG AAT GAT ACT ATC GGA TTA GCC ATT ATG CTT ATT GCA GAA GGC 259
Arg Gln Asn Asp Thr Ile Gly Leu Ala Ile Met Leu Ile Ala Glu Gly
40 45 50

TTA TCT AAA CGT TTC AAC TTT TCA GGA TAC ACC CAG TCT TGG AAA CAA 307
Leu Ser Lys Arg Phe Asn Phe Ser Gly Tyr Thr Gln Ser Trp Lys Gln
55 60 65

GAA ATG ATT GCA GAT GGT ATA GAA GCT TCT ATT AAG GGG CTT CAC AAT 355
Glu Met Ile Ala Asp Gly Ile Glu Ala Ser Ile Lys Gly Leu His Asn
70 75 80 85

TTT GAT GAA ACG AAA TAT AAA AAC CCA CAT GCG TAT ATA ACT CAA GCT 403
Phe Asp Glu Thr Lys Tyr Lys Asn Pro His Ala Tyr Ile Thr Gln Ala
90 95 100

REPLACEMENT SHEET

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TGT TTT AAT GCA TTC GTC CAA CGT GGA TCC ATC GAT CCA CCA TCC GTA 451
Cys Phe Asn Ala Phe Val Gln Arg Gly Ser Ile Asp Pro Pro Ser Val
105 110 115

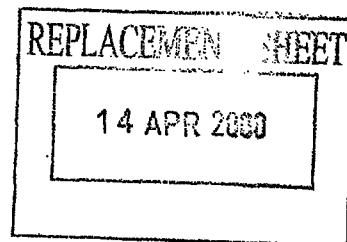
TCA GAA ATA CAA CTA ATG CAT AAT CTG GGT AAA CAT CTG AAT TCA ATG 499
Ser Glu Ile Gln Leu Met His Asn Leu Gly Lys His Leu Asn Ser Met
120 125 130

GAA CGT GTA GAA TGG CTG CGT AAA ~~AAA~~ CTG CAG GAT GTA CAT AAT TTT 547
Glu Arg Val Glu Trp Leu Arg Lys Lys Leu Gln Asp Val His Asn Phe
135 140 145

GTA GCT CTG GGT TAACTCGAGC AGATCCGGCT GCTAACAAAG CCCGAAAGGA 599
Val Ala Leu Gly
150

AGCTGAGTTG GCTGCTGCCA CCGCTGAGCA ATAACTAGCA TAACCCCTTG GGGCCCTTGG 659

GGCCTCTAAA CGGGTCTTGA GGGGTTTTTT GCTGAAAGGA GGAACCTATAT CCGGATAA 717



REPLACEMENT SHEET

FIGURE 2

AGATCTCGAT CCCGCGAAAT TAATACGACT CACTATAGGG AGACCACAAC GGTTTCCCTC 60

TAGAAATAAT TTTGTTTAAC TTTAAGAAGG AGATATACAT ATG TCA GAA ACT AAG 115
Met Ser Glu Thr Lys
1 5

CCT AAA TAT AAT TAC GTA AAC AAT AAA GAG CTT TTA CAA GCT ATT ATT 163
Pro Lys Tyr Asn Tyr Val Asn Asn Lys Glu Leu Leu Gln Ala Ile Ile
10 15 20

GAT TGG AAA ACA GAA TTA GCA AAT AAT AAA GAC CCA AAT AAA GTA GTT 211
Asp Trp Lys Thr Glu Leu Ala Asn Asn Lys Asp Pro Asn Lys Val Val
25 30 35

CGT CAG AAT GAT ACT ATC GGA TTA GCC ATT ATG CTT ATT GCA GAA GGC 259
Arg Gln Asn Asp Thr Ile Gly Leu Ala Ile Met Leu Ile Ala Glu Gly
40 45 50

TTA TCT AAA CGT TTC AAC TTT TCA GGA TAC ACC CAG TCT TGG AAA CAA 307
Leu Ser Lys Arg Phe Asn Phe Ser Gly Tyr Thr Gln Ser Trp Lys Gln
55 60 65

GAA ATG ATT GCA GAT GGT ATA GAA GCT TCT ATT AAG GGG CTT CAC AAT 355
Glu Met Ile Ala Asp Gly Ile Glu Ala Ser Ile Lys Gly Leu His Asn
70 75 80 85

TTT GAT GAA ACG AAA TAT AAA AAC CCA CAT GCG TAT ATA ACT CAA GCT 403
Phe Asp Glu Thr Lys Tyr Lys Asn Pro His Ala Tyr Ile Thr Gln Ala
90 95 100

REPLACEMENT SHEET

14 APR 2000

REPLACEMENT SHEET

TGT TTT AAT GCA TTC GTC CAA CGT ATT AAA AAA GAA CGT AAG GAA GTT 451
Cys Phe Asn Ala Phe Val Gln Arg Ile Lys Lys Glu Arg Lys Glu Val
105 110 115

GCA AAG AAA TAT AGT TAC TTC GTT CAC AAT GTC TAT GAC AGC CGT GAC 499
Ala Lys Lys Tyr Ser Tyr Phe Val His Asn Val Tyr Asp Ser Arg Asp
120 125 130

GAC GAT ATG GTT GCG TTA GTA GAT GAA ACT TTT ATT CAA GAC ATC TAT 547
Asp Asp Met Val Ala Leu Val Asp Glu Thr Phe Ile Gln Asp Ile Tyr
135 140 145

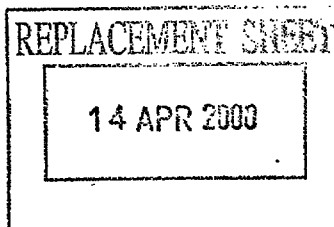
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Asp Lys Met Thr His Tyr Glu Glu Ser Thr Tyr Arg Thr Pro Gly Ala
150 155 160 165

GAA AAG AAA AGT GTT GTA GAT GAT TCT CCT AGT TTG GAT TTT TTA TAT 643
Glu Lys Lys Ser Val Val Asp Asp Ser Pro Ser Leu Asp Phe Leu Tyr
170 175 180

GAG GCT AAC GAT GGA TCC GTT AAC GGT CCA TCC GTA TCA GAA ATA CAA 691
Glu Ala Asn Asp Gly Ser Val Asn Gly Pro Ser Val Ser Glu Ile Gln
185 190 195

CTA ATG CAT AAT CTG GGT AAA CAT CTG AAT TCA ATG GAA CGT GTA GAA 739
Leu Met His Asn Leu Gly Lys His Leu Asn Ser Met Glu Arg Val Glu
200 205 210

TGG CTG CGT AAA AAA CTG CAG GAT GTA CAT AAT TTT GTA GCT CTG GGT 787
Trp Leu Arg Lys Lys Leu Gln Asp Val His Asn Phe Val Ala Leu Gly
215 220 225



REPLACEMENT SHEET

TAAGTCGAGC AGATCCGGCT GCTAACAAAG CCCGAAAGGA AGCTGAGTTG GCTGCTGCCA	847
CCGCTGAGCA ATAACTAGCA TAACCCCTTG GGGCCCTTGG GGCCTCTAAA CGGGTCTTGA	907
GGGGTTTTTT GCTGAAAGGA GGAACATAT CCGGATAA	945

