

REPLIKIN PEPTIDES AND USES THEREOF

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CROSS REFERENCE TO OTHER APPLICATIONS

[0001] This application is a Continuation-In-Part of PCT Application
No.PCT/US02/09240, filed March 26, 2002, which is a Continuation-In-Part of
Applications Nos. US 09/984,057 and US 09/984,056, both filed October 26,
10 2001, which claims priority from Provisional Applications 60/303,396, filed July
9, 2001, the subject matter of which is incorporated herein by reference.

FIELD OF THE INVENTION

[0002] This invention relates to the identification and use of Replikins, a newly
15 discovered class of peptides that share structural characteristics. In particular, this
invention relates to Replikins which have been found in viruses, bacteria, fungus,
cancer associated proteins, plants and unicellular parasites and their use as targets
in the development of methods of treating or preventing diseases. Further, this
invention relates to the use of Replikins in the detection of these diseases. Also
20 this invention relates to the use of Replikins to stimulate growth of plants used for
food.

INTRODUCTION AND BACKGROUND OF THE INVENTION

[0003] Rapid replication is characteristic of virulence in certain bacteria, viruses
25 and malignancies, but no chemistry common to rapid replication in different
organisms has been described previously. This patent application discloses a new
class of protein structures related to rapid replication. A new family of conserved
small proteins related to rapid replication, named Replikins, which are used to

predict and control rapid replication in multiple organisms and diseases and to induce rapid replication in plant and animal life.

[0004] We constructed an algorithm search for Replikins. In applying the

5 algorithm invented herein not only was the function of the epitope revealed - rapid replication, but an entire family of homologues whose function is related to rapid replication was discovered, which we named Replikins.

[0005] The algorithm is based on the following: 1) Evidence that the immune

10 system looks to parts rather than a whole protein in recognition. Protein chains are first hydrolyzed by the immune system into smaller pieces, frequently six (6) to ten (10) amino acids long, as part of the immune systems' process of recognition of foreign structures against which it may mount an immune defense. By way of example, the immune system recognizes the presence of disease by chopping up
15 proteins of the disease agent into smaller peptide sequences and reading them.

This principle is used as a basis for the algorithm with which to search for homologues of the malignin cancer epitope, once the structure of the epitope was known; 2) The specific structure of the malignin epitope, in which two of the three lysines (K's) are eight residues apart is in accordance with the apparent 'rules'

20 used by the immune system for recognition referred to above (6-10 amino acids long); 3) The fact that the malignin cancer epitope was shown to be a very strong antigen, that is -- a generator of a strong immune response; that there are three lysines (K's) in the 10-mer peptide glioma Replikin and that K's are known to bind frequently to DNA and RNA as potential anchors for the entry of viruses; and

25 4) One histidine (H) is included in the sequence of the malignin epitope, between

the two K's which are eight (8) residues apart, suggesting a connection to the metals of redox systems which are required to provide the energy for replication.

[0006] Engineered enzymes and catalytic antibodies, possessing tailored binding pockets with appropriately positioned functional groups have been successful in catalyzing a number of chemical transformations, sometimes with impressive efficiencies. Just as two or more separate proteins with specific and quite different functions are now often recognized to be synthesized together by organisms, and then separately cleaved to 'go about their separate functions', so the Replikin structure is a unique protein with a unique function that appears to be recognized separately by the immune system and may be now rationally engineered- e.g. synthesized to produce a functional unit.

[0007] From a proteomic point of view, this template based on the newly determined glioma peptide sequence has led to the discovery of a wide class of proteins with related conserved structures and a particular function, in this case replication. Examples of the increase in Replikin concentration with virulence of a disease appear in diseases including, influenza, HIV, cancer and tomato leaf curl virus. This class of structures is related to the phenomenon of rapid replication in organisms as diverse as yeast, algae, plants, the gemini curl leaf tomato virus, HIV and cancer.

[0008] In addition to detecting the presence of Replikins in rapidly replicating organisms, we found that 1) Replikin concentration (number of Replikins per 100 amino acids) and 2) Replikin compositions in specific functional states dependant

on rapid replication, provide the basis for the finding that Replikins are related quantitatively as well as qualitatively to the rate of replication of the organism in which they reside. Examples of these functional proofs include the relationship found between rapid replication and virulence in glioblastoma cells, between
5 Replikins in influenza virus and the prediction of influenza pandemics and epidemics, and the relationship between Replikin concentration and rapid replication in HIV.

[0009] The first functional basis for Replikins' role in rapid replication was found
10 in the properties of the glioma Replikin, a 10KD peptide called Malignin in brain glioblastoma multiforme (glioma) - a 250KD cell protein. Antimalignin antibody increased in concentration in serum (AMAS), measured by an early stage diagnostic test for cancer now used for most or all cell types. Malignin was so named because in tissue culture the expression of this peptide and its concentration
15 per milligram membrane protein extractable increased with increased rate of cell division per unit time. Not only is there an increase in the amount of malignin in proportion to the cell number increase but the amount of malignin is enriched, that is - increased ten fold whereas the cell number increased only five fold.

20 [0010] The structure of malignin protein was determined through hydrolysis and mass spectrometry which revealed what proved to be a novel 16 mer peptide sequence. We searched for the 16 mer peptide sequence which we have named a Glioma Replikin protein in databases for the healthy human genome and found that it was not present in these databases.

[0011] As such, the fixed requirement algorithm was used to search in other organisms for the Glioma Replikin protein or homologues thereof. Over 4,000 protein sequences in the "Pub Med" database were searched and homologues were found in viruses and plant forms specifically associated with rapid replication.

5 Homologues of such Replikin proteins occurred frequently in proteins called 'replicating proteins' by their investigators.

[0012] Homologues of the Replikin sequence were found in all tumor viruses (that is viruses that cause cancer), and in 'replicating proteins' of algae, plants, fungi,
10 viruses and bacteria.

[0013] That malignin is enriched ten-fold compared to the five-fold increase in cell number and membrane protein concentration in rapid replication of glioma cells suggests an integral relationship of the Replikins to replication. When the glioma
15 replikin was synthesized in vitro and administered as a synthetic vaccine to rabbits, abundant antimalignin antibody was produced - establishing rigourously the antigenic basis of the antimalignin antibody in serum (AMAS) test, and providing the first potential synthetic cancer vaccine and the prototype for Replikin vaccines in other organisms.

20 [0014] The demonstration of the relationship of the Replikins to replication and the natural immune response to cancer Replikins (overriding cell type) based upon the shared specificity of cancer Replikins, permits passive augmentation of immunity with antimalignin antibody and active augmentation with synthetic
25 Replikin vaccines.

[0015] A study of 8,090 serum specimens from cancer patients and controls has demonstrated that the concentration of antimalignin antibody increases with age in healthy individuals, as the incidence of cancer in the population increases, and increases further two to three-fold in early malignancy, regardless of cell type. In vitro this antibody is cytotoxic to cancer cells at picograms (femtomoles) per cancer cell, and in vivo the concentration of antimalignin antibody relates quantitatively to the survival of cancer patients. As shown in glioma cells, the stage in cancer at which cells only have been transformed to the immortal malignant state but remain quiescent or dormant, now can be distinguished from the more active life-threatening replicating state which is characterized by the increased concentration of Replikins. In addition, clues to the viral pathogenesis of cancer may be found in the fact that glioma glycoprotein 10B has a 50% reduction in carbohydrate residues when compared to the normal 10B. This reduction is associated with virus entry in other instances and so may be evidence of the attachment of virus for the delivery of virus Replikins to the 10B of glial cells as a step in the transformation to the malignant state.

[0016] The sharing of immunological specificity by diverse members of the class, as demonstrated with antimalignin antibody for the glioma and related cancer Replikins, suggests that B cells and their product antibodies may recognize Replikins by means of a similar recognition 'language'. With the discovery of the Replikins, this shared immunological specificity may explain what was previously difficult to understand: why the antimalignin antibody is elevated in all cancers, and is cytotoxic to cancer cells and related to survival of cancer patients in most or

all cell types. Thus antimalignin antibody is produced against cancer Replikins, which share immunological specificity and which are related to the phenomenon of rapid replication, not to cell type.

5 [0017] A second functional basis for the Replikins' role in rapid replication is the study of data from the past 100 years on influenza virus hemagglutinin protein sequences and epidemiology of influenza epidemics and pandemics. To date, only serological hemagglutinin and antibody classification, but no strain-specific conserved peptide sequences have previously been described in influenza, and no
10 changes in concentration and composition of any strain-specific peptide sequences have been described previously which correlate with epidemiologically documented epidemics or rapid replication.

[0018] A four to ten-fold increase in the concentration of strain-specific influenza
15 Replikins in one of each of the four major strains, influenza B, (A)H1N1, (A)H2N2 and (A)H3N2 was found, and that such increase of Replikin concentration was related to influenza epidemics caused specifically by each strain from 1902 to 2001. These increases in concentration were then shown to be due to the reappearance of at least one specific Replikin composition from 1 to up to 64
20 years after its disappearance, plus the emergence of new strain-specific Replikin compositions. Previously, no strain-specific chemical structures were known with which to predict which strains would predominate in coming influenza seasons, nor to devise annual mixtures of whole-virus strains for vaccines. The recent sharp increase in H3N2 Replikin concentration (1997 to 2000), the largest in
25 H3N2's history, and the reappearance of specific Replikin compositions which

were last seen in the high mortality H3N2 pandemic of 1968 and in the two high mortality epidemics of 1975 and 1977, but were absent for 20-25 years, together may be a warning of coming epidemics.

5 [0019] Synthetic Replikins are new vaccines. This high degree of conservation of Replikin structures observed whereby the identical structure can persist for 100 years, or reappear after an absence of from one to 64 years reappears indicates that what was previously thought to be change in virulence due to random substitution of amino acids in influenza proteins is more likely to be change due to an
10 organized process of conservation of Replikins. In fact, if random substitutions of each amino acid occurred, the chance against an average length influenza Replikin sequence being conserved for one year (let alone 100) is calculated to be in the order of 2 to the 27th power to 1.

15 [0020] The significant conservation of Replikins is not unique to influenza virus is also present in foot and mouth disease virus type O and in HIV, as well as in wheat.

[0021] A third functional basis for Replikins' role in rapid replication is the
20 increase in Replikin concentration shown to be related to rapid replication in HIV. The Replikin concentration in the slow-growing low-titre strain of HIV (NS1, "Bru"), prevalent in early stage infection, was found to be one-sixth of the Replikin concentration in the rapidly-growing high-titre strain of HIV (SI, "Lai"), prevalent in late stage HIV infection.

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[0022] Other examples are given of the relation of Replikins to rapid replication.

For example, in tomato curl leaf gemini virus, which devastates tomato crops, the first 161 amino acids of the 'replicating protein', which have been shown to bind to DNA, contain five Replikins.

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[0023] In malaria, legendary for rapid replication, trypanosomes are released from the liver in tens of thousands from one trypanosome. Multiple, novel, almost 'flamboyant' Replikin structures with concentrations of up to 36 overlapping Replikins per 100 amino acids are found therein.

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[0024] The increase in Replikin concentration in influenza epidemics is functionally comparable to the glioma Replikin's increase in concentration during rapid replication of malignant glioma cells and comparable to rapid replication in HIV and in a diverse range of other organisms. Replikins thus are associated with and appear to be part of the structural bases of rapid replication in different organisms.

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[0025] Replikin concentration and composition therefore provide new methods to detect and to control the process of replication, which is central to the survival and dominance of each biological population. The discovery of these new proteins related to rapid replication provides new opportunities 1) for detection of pathogens by qualitative and quantitative determinations of Replikins, 2) for the control of a broad range of diseases in which rapid replication is a key factor by targeting native Replikins and by using synthetic Replikins as vaccines, and 3) for the use of Replikins to foster growth of algal and plant foods.

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[0026] There is a significant number of diseases and pathogens which have proved difficult to detect and treat and for which there is no effective vaccine. Thus, for each disorder there is a need for developing a target that will provide effective methods of detecting, treating or preventing these diseases and pathogens.

SUMMARY OF THE INVENTION

[0027] The present invention provides a method for identifying nucleotide or amino acid sequences that include a Replikin sequence. The method is referred to herein as a 3-point-recognition method. By use of the "3-point recognition" method, namely, peptides comprising from 7 to about 50 amino acids including (1) at least one lysine residue located six to ten amino acid residues from a second lysine residue; (2) at least one histidine residue; and (3) at least 6% lysine residues (Replikin) – constituting a new class of peptides was revealed in algae, yeast, fungi, amoebae, bacteria, plant and virus proteins having replication, transformation, or redox functions.

[0028] In one aspect of the invention there are provided isolated or synthesized peptides containing a Replikin sequence. The peptides comprise from 7 to about 50 amino acids including (1) at least one lysine residue located six to ten amino acid residues from a second lysine residues; (2) at least one histidine residue; and (3) at least 6% lysine residues.

[0029] The present invention also provides methods for detecting the presence

of a contaminating organism in a body sample or environmental sample
comprising:

- (1) isolating nucleic acids from the body sample or environmental sample;
- (2) screening the nucleic acids for the presence of a Replikin structure; and
- 5 (3) correlating the presence of a Replikin structure with the presence of the
contaminating organism.

[0030] In another aspect of the invention there is provided a process for
stimulating the immune system of a subject to produce antibodies that bind
10 specifically to a Replikin sequence, said process comprising administering to the
subject an effective amount of a dosage of a composition comprising at least one
Replikin peptide. One embodiment comprises at least one peptide that is present
in an emerging strain of the organism if such new strain emerges.

15 [0031] The present invention also provides antibodies that bind specifically to a
Replikin, as defined herein, as well as antibody cocktails containing a plurality of
antibodies that specifically bind to Replikins. In one embodiment of the
invention, there are provided compositions comprising an antibody or antibodies
that specifically bind to a Replikin and a pharmaceutically acceptable carrier.

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[0032] In one aspect of the invention there are provided isolated, or separated from
other proteins, recombinant, or synthesized peptides or other methods containing a
viral Replikin sequence. The viral Replikin peptides comprise from 7 to about 50
amino acids including (1) at least one lysine residue located six to ten amino acid

residues from a second lysine residue; (2) at least one histidine residue; and (3) at least 6% lysine residues. (viral Replikin).

[0033] The present invention also provides methods for detecting the presence of a contaminating virus in a body sample or environmental sample comprising:

- (1) isolating nucleic acids from the body sample or environmental sample;
- (2) screening the nucleic acids for the presence of a viral Replikin structure; and
- (3) correlating the presence of viral Replikin structures, their concentration and composition, with the presence of the contaminating virus.

[0034] In another aspect of the invention there is provided a process for stimulating the immune system of a subject to produce antibodies that bind specifically to a viral Replikin sequence, said process comprising administering to the subject an effective amount of a dosage of a composition comprising at least one Replikin peptide. One embodiment comprises at least one peptide that is present in an emerging strain of the virus if such new strain emerges.

[0035] The present invention also provides antibodies that bind specifically to a viral Replikin, as defined herein, as well as antibody cocktails containing a plurality of antibodies that specifically bind to viral Replikins. In one embodiment of the invention, there are provided compositions comprising an antibody or antibodies that specifically bind to a viral Replikin and a pharmaceutically acceptable carrier.

[0036] The present invention also provides therapeutic compositions comprising one or more of isolated virus peptides having from 7 to about 50 amino acids comprising: (1) at least one lysine residue located six to ten residues from a second lysine residue; (2) at least one histidine residue; and (3) at least 6% lysine residues, and a pharmaceutically acceptable carrier.

[0037] In another aspect of the invention there is provided an antisense nucleic acid molecule complementary to a virus Replikin mRNA sequence, said Replikin mRNA sequence denoting from 7 to about 50 amino acids comprising:

(1) at least one lysine residue located six to ten residues from a second lysine residue; (2) at least one histidine residue; and (3) at least 6% lysine residues.

[0038] In yet another aspect of the invention there is provided a method of simulating the immune system of a subject to produce antibodies to viruses, said method comprising: administering an effective amount of at least one virus Replikin peptide having from 7 to about 50 amino acids comprising (1) at least one lysine residue located six to ten amino acid residues from a second lysine residue; (2) at least one histidine residue; (3) and at least 6% lysine residues.

[0039] In another aspect, there is provided a method of selecting a virus peptide for inclusion in a preventive or therapeutic virus vaccine comprising:

- (1) obtaining at least one isolate of each strain of a plurality of strains of said virus;

- (2) analyzing the amino acid sequence of the at least one isolate of each strain of the plurality of strains of the virus for the presence and concentration of Replikin sequences;
- (3) comparing the concentration of Replikin sequences in the amino acid sequence of the at least one isolate of each strain of the plurality of strains of the virus to the concentration of Replikin sequences observed in the amino acid sequence of each of the strains at least one earlier time period to provide the concentration of Replikins for at least two time periods, said at least one earlier time period being within about six months to about three years prior to step (1);
- (4) indentifying the strain of the virus having the highest increase in concentration of Replikin sequences during the at least two time periods; and
- (5) selecting at least one Replikin sequence present in the strain of the virus peptide identified in step (4) as a peptide for inclusion in the virus vaccine.

[0040] The present invention also provides a method of making a preventive or therapeutic virus vaccine comprising:

- (1) identifying a strain of a virus as an emerging strain,
- (2) selecting at least one Replikin sequence present in the emerging strain as a peptide template for the virus vaccine manufacture,
- (3) synthesizing peptides having the amino acid sequence of the at least one Replikin sequence selected in step (2), and

- (4) combining a therapeutically effective amount of the peptides of step (3) with a pharmaceutically acceptable carrier and/or adjuvant.

[0041] In another aspect, the invention is directed to a method of identifying an emerging strain of a virus for diagnostic, preventive or therapeutic purposes comprising:

- (1) obtaining at least one isolate of each strain of a plurality of strains of the virus;
- (2) analyzing the amino acid sequence of the at least one isolate of each strain of the plurality of strains of the virus for the presence and concentration of Replikin sequences;
- (3) comparing the concentration of Replikin sequences in the amino acid sequence of the at least one isolate of each strain of the plurality of strains of the virus to the concentration of Replikin sequences observed in the amino acid sequence of each of the strains at at least one earlier time period to provide the concentration of Replikins for at least two time periods, said at least one earlier time period being within about six months to about three years prior to step (1); and
- (4) identifying the strain of the virus having the highest increase in concentration of Replikin sequences during the at least two time periods.

[0042] In yet another aspect of the invention, there is provided a preventive or therapeutic virus vaccine comprising at least one isolated Replikin present in a

protein of an emerging strain of the virus and a pharmaceutically acceptable carrier and/or adjuvant.

[0043] Also provided by the present invention is a method of preventing or
5 treating a virus infection comprising administering to a patient in need thereof a preventive or therapeutic virus vaccine comprising at least one isolated Replikin present in a protein of an emerging strain of the virus and a pharmaceutically acceptable carrier and/or adjuvant.

10 INFLUENZA

[0044] Influenza is an acute respiratory illness of global importance. Despite international attempts to control influenza virus outbreaks through vaccination, influenza infections remain an important cause of morbidity and mortality.

Worldwide influenza epidemics and pandemics have occurred at irregular and
15 previously unpredictable intervals throughout history and it is expected that they will continue to occur in the future. The impact of both pandemic and epidemic influenza is substantial in terms of morbidity, mortality and economic cost.

[0045] Influenza vaccines remain the most effective defense against influenza
20 virus, but because of the ability of the virus to mutate and the availability of non-human host reservoirs, it is expected that influenza will remain an emergent or re-emergent infection. Global influenza surveillance indicates that influenza viruses may vary within a country and between countries and continents during an influenza season. Virological surveillance is of importance in monitoring
25 antigenic shift and drift. Disease surveillance is also important in assessing the

impact of epidemics. Both types of information have provided the basis of the vaccine composition and the correct use of antivirals. However, to date there has been only annual post hoc hematological classification of the increasing number of emerging influenza virus strains, and no specific chemical structure of the viruses
5 has been identified as an indicator of approaching influenza epidemics or pandemics. Currently, the only basis for annual classification of influenza virus as active, inactive or prevalent in a given year is the activities of the virus hemagglutinin and neuraminidase proteins. No influenza viral chemical structure has been identified prior to this application that can be used for quantitative
10 warning of epidemics or pandemics or to design more effective and safer vaccines.

[0046] Because of the annual administration of influenza vaccines and the short period of time when a vaccine can be administered, strategies directed at improving vaccine coverage are of critical importance.

15 [0047] In one aspect of the invention there are provided isolated or synthesized influenza virus peptides containing a Replikin sequence. The influenza Replikin virus peptides comprise from 7 to about 50 amino acids including (1) at least one lysine residue located six to ten amino acid residues from a second lysine residue;
20 (2) at least one histidine residue; and (3) at least 6% lysine residues. (Influenza Replikin).

[0048] In another aspect of the invention, there is provided a process for stimulating the immune system of a subject to produce antibodies that bind
25 specifically to an influenza virus Replikin sequence, said process comprising

administering to the subject an effective amount of dosage of a composition comprising at least one influenza virus Replikin peptide. In a preferred embodiment the composition comprises at least one peptide that is present in an emerging strain of influenza virus.

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[0049] The present invention also provides antibodies that bind specifically to an influenza virus Replikin, as defined herein, as well as antibody cocktails containing a plurality of antibodies that specifically bind to influenza virus Replikins. In one embodiment of the invention, there are provided compositions comprising an antibody or antibodies that specifically bind to an influenza Replikin and a pharmaceutically acceptable carrier.

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[0050] The present invention also provides therapeutic compositions comprising one or more of isolated influenza virus peptides having from 7 to about 50 amino acids comprising:

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- (1) at least one lysine residue located six to ten residues from a second lysine residue;
- (2) at least one histidine residue; and
- (3) at least 6% lysine residues, and a pharmaceutical acceptable carrier.

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[0051] In another aspect of the invention there is provided an antisense nucleic acid molecule complementary to an influenza virus hemagglutinin Replikin mRNA sequence, said Replikin mRNA sequence denoting from 7 to about 50 amino acids comprising:

- (1) at least one lysine residue located six to ten residues from a second lysine residue;
- (2) at least one histidine residue; and
- (3) at least 6% lysine residues.

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[0052] In yet another aspect of the invention there is provided a method of simulating the immune system of a subject to produce antibodies to influenza virus comprising administering an effective amount of at least one influenza virus Replikin peptide having from 7 to about 50 amino acids comprising:

- 10 (1) at least one lysine residue located six to ten amino acid residues from a second lysine residue;
- (2) at least one histidine residue; and
- (3) at least 6% lysine residues.

15 [0053] In another aspect, there is provided a method of selecting an influenza virus peptide for inclusion in a preventive or therapeutic influenza virus vaccine comprising:

- (1) obtaining at least one isolate of each strain of a plurality of strains of influenza virus;
- 20 (2) analyzing the hemagglutinin amino acid sequence of the at least one isolate of each strain of the plurality of strains of influenza virus for the presence and concentration of Replikin sequences;
- (3) comparing the concentration of Replikin sequences in the hemagglutinin amino acid sequence of the at least one isolate of each strain of the plurality of strains of influenza virus to the concentration
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of Replikin sequences observed in the hemagglutinin amino acid sequence of each of the strains at least one earlier time period to provide the concentration of Replikins for at least two time periods, said at least one earlier time period being within about six months to about three years prior to step (1);

- (4) identifying the strain of influenza virus having the highest increase in concentration of Replikin sequences during the at least two time periods;
- (5) selecting at least one Replikin sequence present in the strain of influenza virus peptide identified in step (4) as a peptide for inclusion in an influenza virus vaccine.

[0054] The present invention also provides a method of making a preventive or therapeutic influenza virus vaccine comprising:

- (1) identifying a strain of influenza virus as an emerging strain;
- (2) selecting at least one Replikin sequence present in the emerging strain as a peptide template for influenza virus vaccine manufacture,
- (3) synthesizing peptides having the amino acid sequence of the at least one Replikin sequence selected in step (2), and
- (4) combining a therapeutically effective amount of the peptides of step (3) with a pharmaceutically acceptable carrier and/or adjuvant.

[0055] In another aspect, the invention is directed to a method of identifying an emerging strain of influenza virus for diagnostic, preventive or therapeutic purposes comprising:

- (1) obtaining at least one isolate of each strain of a plurality of strains of influenza virus;
- (2) analyzing the hemagglutinin amino acid sequence of the at least one isolate of each strain of the plurality of strains of influenza virus for the presence and concentration of Replikin sequences;
- (3) comparing the concentration of Replikin sequences in the hemagglutinin amino acid sequence of the at least one isolate of each strain of the plurality of strains of influenza virus to the concentration of Replikin sequences observed in the hemagglutinin amino acid sequence of each of the strains at at least one earlier time period to provide the concentration of Replikins for at least two time periods, said at least one earlier time period being within about six months to about three years prior to step (1); and
- (4) identifying the strain of influenza virus having the highest increase in concentration of Replikin sequences during the at least two time periods.

[0056] In yet another aspect of the invention, there is provided a preventive or therapeutic influenza virus vaccine comprising at least one isolated Replikin present in the hemagglutinin protein of an emerging strain of influenza virus and a pharmaceutically acceptable carrier and/or adjuvant.

[0057] Also provided by the present invention is a method of preventing or treating influenza virus infection comprising administering to a patient in need thereof a preventive or therapeutic vaccine comprising at least one isolated

Replikin present in the hemagglutinin protein of an emerging strain of influenza virus and a pharmaceutically acceptable carrier and/or adjuvant.

TRYPANOSOMES

5 [0058] In one aspect of the invention there are provided isolated or synthesized trypanosome peptides containing a Replikin sequence. The trypanosome Replikin peptides comprise from 7 to about 50 amino acids including (1) at least one lysine residue located six to ten amino acid residues from a second lysine residue; (2) at least one histidine residue; and (3) at least 6% lysine residues. (Trypanosome
10 Replikins).

MALARIA

[0059] One trypanosome disorder which has proved difficult to treat and for which there is no effective vaccine is malaria. Malaria causes much death, and physical
15 and economic hardship in tropical regions. Malaria is caused mainly by *Plasmodium falciparum*, which has proved to be extremely resistant to treatment and to date, a vaccine for malaria has remained elusive. Thus there is a need for effective malaria vaccines and methods of treating or preventing the disease. This application provides the basis for such vaccines and methods of treatment and
20 prevention. All of the methods described above for production of and treatment with Replikin virus vaccines and Replikin influenza virus vaccines are applicable to the production of and treatment with Replikin malaria vaccines.

[0060] In the present invention, there are provided vaccines and methods for
25 preventing or treating malaria. The malaria vaccines comprise at least one isolated

Plasmodium falciparum Replikin. The present invention also provides methods for treating or preventing malaria comprising administering to a patient an effective amount of preventive or therapeutic vaccine comprising at least one isolated Plasmodium falciparum Replikin.

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[0061] Also provided by the present invention are antibodies, antibody cocktails and compositions that comprise antibodies that specifically bind to a Replikin or Replikins present in a malaria antigen of Plasmodium falciparum.

10 [0062] Another example of a trypanosome which may be treated under the present invention as is the case for malaria, the Replikins of Treponema Pallidum (syphilis), can be used for detection, prevention, treatment of syphilis.

BACTERIA

15 [0063] In one aspect of the invention there are provided isolated or synthesized bacterial peptides containing a Replikin sequence (bacterial Replikins). The bacterial peptides comprise from 7 to about 50 amino acids including (1) at least one lysine residue located six to ten amino acid residues from a second lysine residue; (2) at least one histidine residue; and (3) at least 6% lysine residues.

20 (bacterial Replikins). US Application Serial No. 10/105,232 filed March 26, 2002 is incorporated by reference in its entirety, including but not limited to the bacterial sequence listing and information.

[0064] The present invention also provides methods for detecting the presence

of a contaminating bacterial organism in a body sample or environmental sample comprising:

- (1) isolating nucleic acids from the body sample or environmental sample;
- (2) screening the nucleic acids for the presence of a Replikin structure; and
- 5 (3) correlating the presence of a Replikin structure with the presence of the contaminating organism.

[0065] In another aspect of the invention there is provided a process for stimulating the immune system of a subject to produce antibodies that bind
10 specifically to a bacterial Replikin sequence, said process comprising administering to the subject an effective amount of a dosage of a composition comprising at least one bacterial Replikin peptide. One embodiment comprises at least one bacterial peptide that is present in an emerging strain of the bacterial organism if such new strain emerges.

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[0066] The present invention also provides antibodies that bind specifically to a bacterial Replikin, as defined herein, as well as antibody cocktails containing a plurality of antibodies that specifically bind to bacterial Replikins. In one embodiment of the invention, there are provided compositions comprising an
20 antibody or antibodies that specifically bind to a bacterial Replikin and a pharmaceutically acceptable carrier.

[0067] The present invention also provides therapeutic compositions comprising one or more of isolated bacterial peptides having from 7 to about 50 amino acids
25 comprising:

- (1) at least one lysine residue located six to ten residues from a second lysine residue;
- (2) at least one histidine residue;
- (3) at least 6% lysine residues; and
- 5 (4) a pharmaceutically acceptable carrier.

[0068] In another aspect of the invention there is provided an antisense nucleic acid molecule complementary to a bacterial Replikin mRNA sequence, said Replikin mRNA sequence denoting from 7 to about 50 amino acids comprising:

- 10 (1) at least one lysine residue located six to ten residues from a second lysine residue;
- (2) at least one histidine residue; and
- (3) at least 6% lysine residues.

15 [0069] In yet another aspect of the invention there is provided a method of simulating the immune system of a subject to produce antibodies to bacteria comprising administering an effective amount of at least one bacterial Replikin peptide having from 7 to about 50 amino acids comprising:

- 20 (1) at least one lysine residue located six to ten amino acid residues from a second lysine residue;
- (2) at least one histidine residue; and
- (3) at least 6% lysine residues.

[0070] In another aspect, there is provided a method of selecting a bacterial Replikin peptide for inclusion in a preventive or therapeutic bacterial vaccine comprising:

- 5 (1) obtaining at least one isolate of each strain of a plurality of strains of the bacteria;
- (2) analyzing the amino acid sequence of the at least one isolate of each strain of the plurality of strains of the bacteria for the presence and concentration of bacterial Replikin sequences;
- 10 (3) comparing the concentration of bacterial Replikin sequences in the amino acid sequence of the at least one isolate of each strain of the plurality of strains of the bacteria to the concentration of bacterial Replikin sequences observed in the amino acid sequence of each of the strains at least one earlier time period to provide the concentration of
15 bacterial Replikins for at least two time periods, said at least one earlier time period being within about six months to about three years prior to step (1), or earlier in rapidly mutating bacteria;
- (4) indentifying the strain of the bacteria having the highest increase in concentration of bacterial Replikin sequences during the at least two
20 time periods; and
- (5) selecting at least one bacterial Replikin sequence present in the strain of the bacterial peptide identified in step (4) as a peptide for inclusion in the bacterial vaccine.

[0071] The present invention also provides a method of making a preventive or therapeutic bacterial vaccine comprising:

- (1) identifying a strain of a bacteria as an emerging strain;
- (2) selecting at least one bacterial Replikin sequence present in the emerging strain as a peptide template for the bacterial vaccine manufacture;
- (3) synthesizing peptides having the amino acid sequence of the at least one bacterial Replikin sequence selected in step (2); and
- (4) combining a therapeutically effective amount of the peptides of step (3) with a pharmaceutically acceptable carrier and/or adjuvant.

[0072] In another aspect, the invention is directed to a method of identifying an emerging strain of bacteria for diagnostic, preventive or therapeutic purposes comprising:

- (1) obtaining at least one isolate of each strain of a plurality of strains of the bacteria;
- (2) analyzing the amino acid sequence of the at least one isolate of each strain of the plurality of strains of the bacteria for the presence and concentration of bacterial Replikin sequences;
- (3) comparing the concentration of bacterial Replikin sequences in the amino acid sequence of the at least one isolate of each strain of the plurality of strains of the bacteria to the concentration of bacterial Replikin sequences observed in the amino acid sequence of each of the strains at at least one earlier time period to provide the concentration of bacterial Replikins for at least two time periods, said at least one earlier

time period being within about six months to about three years prior to
step (1); and

- (4) identifying the strain of the bacteria having the highest increase in
concentration of bacterial Replikin sequences during the at least two
time periods.

5

[0073] In yet another aspect of the invention, there is provided a preventive or
therapeutic bacterial vaccine comprising at least one isolated bacterial Replikin
present in a protein of an emerging strain of the bacteria and a pharmaceutically
acceptable carrier and/or adjuvant.

10

[0074] Two important sub-species of bacteria, classified under mycobacteria, are
Mycobacterium leprae (leprosy) whose 30-s ribosomal protein has a C-terminal
Replikin and Mycobacterium tuberculosis (tuberculosis) whose ATPase has three
Replikins.

15

Replikin in 30s ribosomal protein s6 of Mycobacterium leprae (leprosy) is:
kvimrtdkh

Replikins in the ATPase of Mycobacterium tuberculosis are:

hprpkvaaalkdsyrk

hprpkvaaalk

ksaqkwpdkflagaaqvah

Replikins in the B-D-galactosidase of E. coli:

25

hawqhkgktlfisrk

hqgktlfisrk

Replikins in *Agrobacterium tumefaciens*:

5 hsdqqlavmiaakrlddyk

hlldhпасvgqldlramlaveevkidnpvymek

hпасvgqldlramlaveevkidnpvymek

kcvmakncnikcpaglttnqeafngdpralaqylmniah

kncnikcpaglttnqeafngdpralaqylmniah

10 hhdtysiedlaqlihdakaarvrivk

hdtysiedlaqlihdakaarvrivk

hdakaarvrivk

kigqgakpgeggqlpspkvtveiaaarggtpgvelvsppph

kigqgakpgeggqlpspkvtveiaaarggtpgvelvsppph

15 kaseitktlasgamshgalvaaheavahgtmvggmsnsgeggeh

kaseitktlasgamshgalvaaheavah

kaseitktlasgamshgalvaaah

kaseitktlasgamsh

kryfpnvktpvggvtfaviaqavadwh

20 hhiaaglgfgasavypigvqfraeckfgadadkafkrfakaaekslmk

hhiaaglgfgasavypigvqfraeckfgadadkafkrfakaaekslmk

hhiaaglgfgasavypigvqfraeckfgadadkafkrfakaaek

hhiaaglgfgasavypigvqfraeckfgadadkafkrfak

hhiaaglgfgasavypigvqfraeckfgadadk

25 hiaaglgfgasavypigvqfraeckfgadadkafkrfakaaekslmk

hiaaglgfgasavypigvqfraeckfgadadkafkrfakaaek

hiaaglgfgasavypigvqfraeckfgadadkafkrfak

hiaaglgfgasavypigvqfraeckfgadadk

kfglydaafeksscgvftrkdgvqth

5

[0075] Also provided by the present invention is a method of preventing or treating a bacterial infection comprising administering to a patient in need thereof a preventive or therapeutic vaccine comprising at least one isolated bacterial Replikin present in a protein of an emerging strain of the bacteria and a

10 pharmaceutically acceptable carrier and/or adjuvant.

FUNGUS

[0076] In one aspect of the invention there are provided isolated or synthesized fungal peptides containing a Replikin sequence. The fungal Replikin peptides

15 comprise from 7 to about 50 amino acids including (1) at least one lysine residue located six to ten amino acid residues from a second lysine residue; (2) at least one histidine residue; and (3) at least 6% lysine residues (fungal Replikins).

[0077] All of the methods described above for production of and treatment with

20 bacterial Replikin vaccines are applicable to the production of and treatment with fungal Replikin vaccines.

[0078] In another aspect of the invention there is provided a process for stimulating the immune system of a subject to produce antibodies that bind

specifically to a fungal Replikin sequence, said process comprising administering to the subject an effective amount of a dosage of a composition comprising at least one fungal Replikin peptide.

5 [0079] The present invention also provides antibodies that bind specifically to a fungal Replikin, as defined herein, as well as antibody cocktails containing a plurality of antibodies that specifically bind to viral Replikins. In one embodiment of the invention, there are provided compositions comprising an antibody or antibodies that specifically bind to a fungal Replikin and a
10 pharmaceutically acceptable carrier.

[0080] The present invention also provides therapeutic compositions comprising one or more of isolated fungal peptides having from 7 to about 50 amino acids comprising:

- 15 (1) at least one lysine residue located six to ten residues from a second lysine residue;
- (2) at least one histidine residue;
- (3) at least 6% lysine residues; and
- (4) a pharmaceutically acceptable carrier.

20

[0081] In another aspect of the invention there is provided an antisense nucleic acid molecule complementary to an fungal Replikin mRNA sequence, said Replikin mRNA sequence having from 7 to about 50 amino acids comprising:

- (1) at least one lysine residue located six to ten residues from a second
25 lysine residue;

- (2) at least one histidine residue; and
- (3) at least 6% lysine residues.

[0082] In another aspect of the invention there is provided a process for
stimulating the immune system of a subject to produce antibodies that bind
specifically to a fungal Replikin sequence, said process comprising administering
to the subject an effective amount of a dosage of a composition comprising at least
one Replikin peptide.

10 INCREASING REPLICATION

[0083] In yet another aspect of the invention there is provided a method for
increasing the replication rate of an organism comprising transforming a gene
encoding an enzyme or other protein having a replication function in the organism
with at least one Replikin structure.

15

DEFINITIONS

[0084] As used herein, the term "peptide" or "protein" refers to a compound of
two or more amino acids in which the carboxyl group of one is united with an
amino group of another, forming a peptide bond. The term peptide is also used to
denote the amino acid sequence encoding such a compound. As used herein,
"isolated" or "synthesized" peptide or biologically active portion thereof refers to a
peptide that is substantially free of cellular material or other contaminating
peptides from the cell or tissue source from which the peptide is derived, or
substantially free from chemical precursors or other chemicals when chemically

synthesized by any method, or substantially free from contaminating peptides when synthesized by recombinant gene techniques.

[0085] As used herein, a Replikin peptide or Replikin protein is an amino acid sequence having 7 to about 50 amino acids comprising:

- (1) at least one lysine residue located six to ten amino acid residues from a second lysine residue;
- (2) at least one histidine residue;
- (3) at least 6% lysine residues.

Similarly, a Replikin sequence is the amino acid sequence encoding such a peptide or protein.

[0086] As used herein, "emerging strain" as used herein refers to a strain of a virus, bacterium, fungus, or other organisms identified as having an increased increasing concentration of Replikin sequences in one or more of its protein sequences relative to the concentration of Replikins in other strains of such organism. The increase or increasing concentration of Replikins occurs over a period of at least about six months, and preferably over a period of at least about one year, most preferably over a period of at least about three years or more, for example, in influenza virus, but may be a much shorter period of time for bacteria and other organisms.

[0087] As used herein, "mutation" refers to change in this structure and properties of an organism caused by substitution of amino acids. In contrast, the term

“conservation” as used herein, refers to conservation of particular amino acids due to lack of substitution.

BRIEF DESCRIPTION OF THE DRAWINGS

5 [0088] Figure 1 is a bar graph depicting the frequency of occurrence of Replikins in various organisms.

[0089] Figure 2 is a graph depicting the percentage of malignin per milligram total membrane protein during anaerobic replication of glioblastoma cells.

10

[0090] Figure 3 is a bar graph showing amount of antimalignin antibody produced in response to exposure to the recognin 16-mer.

15

[0091] Figure 4A is a photograph of a blood smear taken with ordinary and fluorescent light. Figure 4B is a photograph of a blood smear taken with ordinary and fluorescent light illustrating the presence of two leukemic cells. Figure 4C is a photograph of a dense layer of glioma cells in the presence of antimalignin antibody. Figure 4D and Figure 4E are photographs of the layer of cells in Figure 4C taken at 30 and 45 minutes following addition of antimalignin antibody

20

[0092] Figure 4F is a bar graph showing the inhibition of growth of small cell lung carcinoma cells in vitro by antimalignin antibody.

25

[0093] Figure 5 is a plot of the amount of antimalignin antibody present in the serum of patients with benign or malignant breast disease pre-and post surgery.

[0094] Figure 6 is a box diagram depicting an embodiment of the invention wherein a computer is used to carry out the 3-point-recognition method of identifying Replikin sequences.

5

[0095] Figure 7 is a graph showing the concentration of Replikins observed in hemagglutinin of influenza B and influenza A strain, H1N1, on a year by year basis from 1918 through 2001.

10

[0096] Figure 8 is a graph of the Replikin concentration observed in hemagglutinin of influenza A strains, H2N2 and H3N2, as well as an emerging strain defined by its constituent Replikins, designated H3N2(R), on a year by year basis from 1950 to 2001.

15

DETAILED DESCRIPTION OF THE INVENTION

[0097] The identification of a new family of small peptides related to the phenomenon of rapid replication, referred to herein as Replikins, provides targets for detection of pathogens in a sample and developing therapies, including vaccine development. In general, knowledge of and identification of this family of peptides enables development of effective therapies and vaccines for any organism that harbors Replikins. Identification of this family of peptides also provides for the detection of viruses and virus vaccine development.

20

[0098] For example, identification of this family of peptides provides for the detection of influenza virus and provides new targets for influenza treatment.

25

Identification of this family of peptides also provides for example, for the detection of malaria and provides new targets for malaria vaccine development. Further examples provided by the identification of this family of peptides include the detection of infectious disease Replikins, cancer immune Replikins and structural protein Replikins.

[0099] Rapid replication is characteristic of virulence in certain bacteria, viruses and malignancies, but no chemistry common to rapid replication in different organisms has been described. We have found a family of conserved small protein sequences related to rapid replication, which we have named Replikins. Such Replikins offer new targets for developing effective detection methods and therapies. The first Replikin found was the glioma Replikin, which was identified in brain glioblastoma multiforme (glioma) cell protein called malignin.

[00100] Hydrolysis and mass spectrometry of malignin revealed the novel 16 mer peptide sequence which contains the glioma Replikin. This Replikin was not found in databases for the normal healthy human genome and therefore appeared to be derived from some source outside the body.

[0101] We have devised an algorithm to search for the glioma Replikin or homologue thereof. Homologues were not common in over 4,000 protein sequences, but were found, surprisingly, in all tumor viruses, and in the replicating proteins of algae, plants, fungi, viruses and bacteria.

[0102] We have identified that both 1) Replikin concentration (number of Replikins per 100 amino acids) and 2) Replikin composition correlate with the functional phenomenon of rapid replication. These relationships provide functional basis for the determination that Replikins are related quantitatively as well as qualitatively to the rate of replication.

[0103] The first functional basis for Replikins role to rapid replication is seen in glioma replication. The fact that glioma malignin is enriched ten-fold compared to the five-fold increase in cell number and membrane protein concentration in rapid replication of glioma cells suggests an integral relationship of the Replikins to replication. When the glioma Replikin was synthesized in vitro and administered as a synthetic vaccine to rabbits, abundant antimalignin antibody was produced. This establishes the antigenic basis of the antimalignin antibody in serum (AMAS) test, and provides the first potential synthetic cancer vaccine and the prototype for Replikin vaccines in other organisms. With the demonstration of this natural immune relationship of the Replikins to replication and this natural immune response to cancer Replikins, which overrides cell type, based upon the shared specificity of cancer Replikins and rapid replication, both passive augmentation of this immunity with antimalignin antibody and active augmentation with synthetic Replikin vaccines now is possible.

[0104] The relationship between the presence of antimalignin antibody and survival in patients was shown in a study of 8,090 serum specimens from cancer patients. The study showed that the concentration of antimalignin antibody increases with age, as the incidence of cancer in the population increases, and

increases further two to three-fold in early malignancy, regardless of cell type. In vitro, the antimalignin antibody is cytotoxic to cancer cells at picograms (femtomoles) per cancer cell, and in vivo the concentration of antimalignin antibody relates quantitatively to the survival of cancer patients. As shown in glioma cells, the stage in cancer at which cells have only been transformed to the immortal malignant state but remain quiescent or dormant, now can be distinguished from the more active life-threatening replicating state, which is characterized by the increased concentration of Replikins. In addition, clues to the viral pathogenesis of cancer may be found in the fact that glioma glycoprotein 10B has a 50% reduction in carbohydrate residues when compared to the normal 10B. This reduction is associated with virus entry in other instances, and so may be evidence of the attachment of virus for the delivery of virus Replikins to the 10B of glial cells as a step in the transformation to the malignant state.

[0105] Our study concerning influenza virus hemagglutinin protein sequences and influenza epidemiology over the past 100 years, has provided a second functional basis for the relations of Replikins to rapid replication. Only serological hemagglutinin and antibody classification, but no strain-specific conserved peptide sequences have previously been described in influenza. Further, no changes in concentration and composition of any strain-specific peptide sequences have been described previously that correlate with epidemiologically documented epidemics or rapid replication. In this study, a four to ten-fold increase in the concentration of strain-specific influenza Replikins in one of each of the four major strains, influenza B, (A)H1N1, (A)H2N2 and (A)143N2, is shown to relate to influenza epidemics caused by each strain from 1902 to 2001.

[0106] We then showed that these increases in concentration are due to the reappearance of at least one specific Replikin composition from 1 to up to 64 years after its disappearance, plus the emergence of new strain-specific Replikin compositions. Previously, no strain-specific chemical structures were known with which to predict the strains that would predominate in coming influenza seasons, nor to devise annual mixtures of whole-virus strains for vaccines. The recent sharp increase in H3N2 Replikin concentration (1997 to 2000), the largest in H3N2's history, and the reappearance of specific Replikin compositions that were last seen in the high mortality H3N2 pandemic of 1968, and in the two high mortality epidemics of 1975 and 1977, but were absent for 20-25 years, together may be a warning of coming epidemics. This high degree of conservation of Replikin structures observed, whereby the identical structure can persist for 100 years, or reappear after an absence of from one to 64 years, indicate that what was previously thought to be change due to random substitution of amino acids in influenza proteins is more likely to be change due to an organized process of conservation of Replikins.

[0107] The conservation of Replikins is not unique to influenza virus but was also observed in other sources, for example in foot and mouth disease virus, type 0, HIV tat, and wheat.

[0108] A third functional basis for Replikins' role in rapid replication is seen in the increase in rapid replication in HIV. Replikin concentration was shown to be related to rapid replication in HIV. We found the Replikin concentration in the

slow growing low-titre strain of HIV (NS1, "Bru"), which is prevalent in early stage infection, to be one-sixth of the Replikin concentration in the rapidly-growing high-titre strain of HIV (SI, "Lai")(prevalent in late stage HIV infection).

5 [0109] Further examples demonstrate the relationship of Replikins to rapid replication. In the "replicating protein," of tomato curl leaf gemini virus which devastates tomato crops, the first 161 amino acids, the sequence which has been shown to bind to DNA, was shown to contain five Replikins. In malaria, legendary for rapid replication when trypanosomes are released from the liver in
10 the tens of thousands from one trypanosome, multiple, novel, almost 'flamboyant' Replikin structures have been found with concentrations of up to 36 overlapping Replikins per 100 amino acids.

[0110] The conservation of any structure is critical to whether that
15 structure provides a stable invariant target to attack and destroy or to stimulate. When a structure is tied in some way to a basic survival mechanism of the organism, the structures tend to be conserved. A varying structure provides an inconstant target, which is a good strategy for avoiding attackers, such as antibodies that have been generated specifically against the prior structure and thus
20 are ineffective against the modified form. This strategy is used by influenza virus, for example, so that a previous vaccine may be quite ineffective against the current virulent virus.

REPLIKINS AS STABLE TARGETS FOR TREATMENT

[0111] Both bacteria and HIV have both Replikin and non-Replikin amino acids.

In HIV, for example, there has been a recent increase in drug-resistance from 9% to 13% due to mutation, that is substitution of non-Replikin amino acids. (See

5 detailed analysis of TAT protein of HIV discussed herein). In bacteria, the development of 'resistant strains' is due to a similar mechanism. However, we have found that Replikin structures do not mutate or change to the same degree as non Replikin amino acids (see also discussion of foot and mouth disease virus conservation of Replikins discussed herein). The Replikin structures, as opposed
10 to the non-Replikin structures are conserved and thus provide new constant targets for treatment.

[0112] Certain structures too closely related to survival functions apparently cannot change constantly. Because an essential component of the Replikin

15 structure is histidine (h), which is know for its frequent binding to metal groups in redox enzymes and probable source of energy needed for replication, and since this histidine structure remains constant, this structure remains all the more attractive a target for destruction or stimulation.

20 [0113] From a proteomic point of view, inventors construction of a template based on the newly determined glioma peptide sequence led them to the discovery of a wide class of proteins with related conserved structures and a particular function, in this case replication. Examples of the increase in Replikin concentration with virulence of a disease include, influenza, HIV, cancer and tomato leaf curl virus.

25 This newly recognized class of structures is related to the phenomenon of rapid

replication in organisms as diverse as yeast, algae, plants, the gemini curl leaf tomato virus, HIV and cancer.

[0114] Replikin concentration and composition provide new quantitative
5 methods to detect and control the process of replication, which is central to the survival and dominance of each biological population. The sharing of immunological specificity by diverse members of the class, as demonstrated with antimalignin antibody for the glioma and related cancer Replikins, suggests that B cells and their product antibodies may recognize Replikins by means of a similar
10 recognition language.

[0115] Examples of peptide sequences of cancer Replikins or as containing a Replikin, i.e., a homologue of the glioma peptide, kagvafihkk, may be found in such cancers of, but not limited to, the lung, brain, liver, soft-tissue, salivary gland,
15 nasopharynx, esophagus, stomach, colon, rectum, gallbladder, breast, prostate, uterus, cervix, bladder, eye, forms of melanoma, lymphoma, leukemia, and kidney.

[0116] Replikins provide for: 1) detection of pathogens by qualitative and quantitative determinations of Replikins; 2) treatment and control of a broad range
20 of diseases in which rapid replication is a key factor by targeting native Replikins and by using synthetic Replikins as vaccines; and 3) fostering increased growth rates of algal and plant foods.

[0117] The first Replikin sequence to be identified was the cancer cell Replikin
25 found in a brain cancer protein, malignin, which was demonstrated to be enriched

ten-fold during rapid anaerobic replication of glioblastoma multiforme (glioma) cells. (Figure 2) Malignin is a 10KDa portion of the 250 KDa glycoprotein 10B, which was isolated in vivo and in vitro from membranes of glioblastoma multiforme (glioma) cells. Hydrolysis and mass spectroscopy of malignin

5 revealed a 16-mer peptide sequence, ykagvafllhkkndide (SEQ ID NO.:4), which is referred to herein as the glioma Replikin and which includes the shorter peptide, kagvafllhkk (SEQ ID NO.: 1), both of which apparently are absent in the normal human genome.

10

Table 1

16-mer peptide sequence ykagvafllhkkndide obtained from malignin by hydrolysis and mass spectrometry

Seq ID NO.	Fragment Identified	MH ⁺ (mass)	Sequence	Method By Which Fragment Obtained			
				Auto-hydrolysis of malignin free in solution	Auto-hydrolysis of malignin immobilized on bromoacetyl cellulose	Microwaved 5 seconds	Microwaved 30 seconds
19	1-3	331.21	(y)ka(g)				+
20	1-5	537.40	(y)kagv(a)		+		
21	2-6	445.28	(y)kagva(f)		+		
22	2-7	592.45	(Y)kagvaf(l)				
23	4-11	899.55	(a)gvafllhkk(n)				+
24	5-7	336.19	(g)vaf(l)				+
25	6-7	230.12	(v)af(l)				
26	6-10	615.36	(v)afllhk(k)				+
27	6-10	615.36	(v)afllhk(k)				
28	6-12	859.50	(v)afllhkkn(d)		+		
29	6-12	859.50	(v)afhkkn(d)				
30	7-8	279.17	(a)fl(h)				
31	10-16	861.43	(h)kkndide(f)		+		
32	11-14	489.27	(k)kndi(d)		+		
33	12-15	476.2	(k)ndid(e)				

[0118] When the 16-mer glioma Replikin was synthesized and injected as a synthetic vaccine into rabbits, abundant antimalignin antibody was produced.

5 (Bogoch et al., Cancer Detection and Prevention, 26 (Suppl. 1): 402 (2002)). The concentration of antimalignin antibody in serum in vivo has been shown to relate quantitatively to the survival of cancer patients. (Bogoch et al., Protides of Biological Fluids, 31:739-747 (1984). In vitro antimalignin antibodies have been shown to be cytotoxic to cancer cells at a concentration of picograms (femtomolar) per cancer cell. (Bogoch et al., Cancer Detection and Prevention, 26 (Suppl. 1):
10 402 (2002).

[0119] Studies carried out by the inventors showed that the glioma Replikin is not represented in the normal healthy human genome. Consequently, a search for the
15 origin and possible homologues of the Replikin sequence was undertaken by analysis of published sequences of various organisms.

[0120] By using the 16-mer glioma Replikin sequence as a template and constructing a recognition proteomic system to visually scan the amino acid
20 sequences of proteins of several different organisms, a new class of peptides, the Replikins, was identified. The present invention provides a method for identifying nucleotide or amino acid sequences that include a Replikin sequence. The method is referred to herein as a 3-point-recognition method. The three point recognition method comprises: a peptide from 7 to about 50 amino acids including
25 (1) at least one lysine residue located six to ten amino acid residues from a second lysine residue; (2) at least one histidine residue; and (3) at least 6% lysine residues. (Replikin). These peptides or proteins constitute a new class of peptides

in species including algae, yeast, fungi, amoebae, bacteria, plant, virus and cancer proteins having replication, transformation, or redox functions. Replikin peptides have been found to be concentrated in larger 'replicating' and 'transforming' proteins (so designated by their investigators, See Table 2) and cancer cell

5 proteins. No sequences were found to be identical to the malignin 16-mer peptide.

Table 2:
Examples of Replikins in various organisms - prototype: Glioma Replikin* kagvafllhkk
 (SEQ ID No.:1)

Algae:	SEQ ID NO.		
	34	Caldophera prolifera	kaskfllh
	35	Isolepis prolifera	kaqaetgeikgh
Yeast:	36	Schizosaccharomyces pombe	ksfkypkhhk
	37	Oryza sativa	kkaygnelhk
	2	Sacch. cerevisiae replication binding protein	hsikrelgiifdk
Fungi:		Isocitrate lyase ICI 1, Penicillium marneffeii	kvdjvthqk
	38	DNA-dependent RNA polymerase II, Discula destructiva	kleedaayhrkk
	39	Ophiostoma novo-ulm I, RNA in Dutch elm disease	kvilplrgnikgiffkh
	40	fungus	
Amoeba:	41	Entamoeba invadens, histone H2B	klilkgdlnkh
Bacteria:	42	Pribosomal protein replication factor, Helicobacter pylori	kshvafik
		Replication-associated protein Staph. aureus	
	10	Mycoplasma pulmonis, chromosome replication	kkektthnk
	43	Macrophage infectivity potentiator, L. legionella	kvaffqlkk
	90	Bacillus anthracis	khlisvkk
	91	Bacillus anthracis	hvkkekekknk
	92	Bacillus anthracis	khivkievk
	93	Bacillus anthracis	kkkkikdiygkdallh
	94	Bacillus anthracis	kwekikqh
	95	Bacillus anthracis	kkqippiepkddiik
	96	Bacillus anthracis	huryasnivesayllilnew-knniqsdlikk
	97	Bacillus anthracis	havddyagytlldknqsdv-tnskk
	98	Bacillus anthracis	haerlkvqknapk
Plants:	44	Arabidopsis thaliana, prolifera	kdhdffgdk
	45	Arabidopsis thaliana, cytoplasmic ribosomal	kmkgllqkkah
	46	Arabidopsis thaliana, DNA binding protein	kelssttqeksh
Viruses:	9	Replication associated protein A [Maize streak virus]	Kekkpskdeimrdiish
	11	Bovine herpes virus 4, DNA replication protein	hkinitngqk
	12	Meleagrid herpesvirus 1, replication binding protein	hkdllyrlmk
	47	Feline immunodeficiency	hlkdyklvk
	3	Foot and Mouth Disease (O)	hkqkivapvk
	5	HIV Type 1	kcfcgckegh
	7	HIV Type 2	kcnwckegh
	99	Small Pox Virus (Variola)	khymitwyk
	100	Small Pox Virus (Variola)	kysqtgkeliik
	101	Small Pox Virus (Variola)	hyddvrikndivvsrck
	102	Small Pox Virus (Variola)	hrfkildski
	103	Small Pox Virus (Variola)	kerghnyyfe

Tumor Viruses:	48	Rous sarcoma virus tyrosine-protein kinase	kklrhek
	49	v-yes, avian sarcoma	kklrhdk
	50	c-yes, colon cancer, malignant melanoma	kklrhdk
	51	v-srcC, avian sarcoma	kklrhek
	52	c-src, colon, mammary, pancreatic cancer	kklrhek
	53	Neuroblastoma RAS viral (v-ras) oncogene	kqahelak
	54	VPI (major capsid protein) [Polyomavirus sp.]	kthrfslk
	55	Sindbis	knlhkek
	56	E1 [Human papillomavirus type 71]	khrpilqlk
	57	v-erbB from AEV and c-erb	kspnhvk
	58	v-fms (feline sarcoma)	knihlek
	59	c-fms (acute and chronic myelomonocytic tumors)	knihlek
	60	large t-antigen 1 [Polyomavirus sp.]	kphlaqslek
	61	middle t-antigen [Polyomavirus sp.]	kqhrelkdk
	62	small t-antigen [Polyomavirus sp.]	kqhrelkdk
	63	v-abl, murine acute leukemia	kvpvlisptlkh
	64	Human T-cell lymphotropic virus type 2	kslllevdkdish
	65	c-kit, GI tumors, small cell lung carcinoma	kagitimvkreyh
	18	Hepatitis C	hyppkpgcivpak
Trans-Forming Proteins:	66	Transforming protein myb	Ksgkhlglk
	67	Transforming protein myc, Burkitt lymphoma	krreqlklhk
	68	Ras-related GTP-binding protein	ksfevikvih
	69	Transforming protein ras (teratocarcinoma)	kkkhtvkk
	70	TRAF-associated NF-kB activator TANK	kaqkdhlslk
	71	RFP transforming protein	hllcrvkdllk
	72	Transforming protein D (S.C.)	kygspkhrlik
	73	Papilloma virus type 11, transforming protein	klkhilgkarfik
	74	Protein tyrosine kinase (EC 2.7.1.112slk	kgdhvdkhykirk
	75	Transforming protein (axl(-))	kekldvmvdrhk
Cancer Cell Proteins:	76	Transforming protein (N-myc)	klqarqqqlkkieh
	77	Fibroblast growth factor 4 (Kaposi sarcoma)	kkgnrvsptmkvth
	78	Matrix metalloproteinase 7 (uterine)	keiplhfrk
	79	Transcription factor 7-like	kkkphilk
	80	Breast cancer antigen NY-BR-87	ktrhdplak
	81	BRCA-1-Associated Ring Domain Protein (breast)	khhpkdnlk
	82	'Autoantigen from a breast tumor'	khkrkkfrqk
	83	Glioma Replikin (this study)	kagvafllhkk
	84	Ovarian cancer antigen	khkrkkfrqk
	85	EE L leukemia	kkkskkhdk
	86	Proto-oncogene tyrosine-protein kinase C-ABLE	hksekalprk
	87	Adenomatosis polyposis coli	kkkkpsrlkgdnek
	88	Gastric cancer transforming protein	kkkgnrvsptmkvth
	89	Transforming protein (K-RAS 2B),lung	khkekmskdgkkkkkksk

[0121] Identification of an amino acid sequence as a Replikin or as containing a Replikin, i.e., a homologue of the glioma peptide, kagvafllhkk, requires that the three following requirements be met. According to the three point recognition system the sequences have three elements: (1) at least one lysine residue located six to ten residues from another lysine residue; (2) at least one histidine residue; and (3) a composition of at least 6% lysine within an amino acid sequence of 7 to about 50 residues.

[0122] Databases were searched using the National Library of Medicine keyword "PubMed" descriptor for protein sequences containing Replikin sequences. Over 4,000 protein sequences were visually examined for homologues. Sequences of all individual proteins within each group of PubMed-classified proteins were visually scanned for peptides meeting the three above-listed requirements. An infrequent occurrence of homologues was observed in "virus peptides" as a whole (1.5%) (N=953), and in other peptides not designated as associated with malignant transformation or replication such as "brain peptides" and "neuropeptides" (together 8.5%) (N=845). However, surprisingly, homologues were significantly more frequently identified in large "replicating proteins," which were identified as having an established function in replication in bacteria, algae, and viruses. Even more surprising was the finding that Replikin homologues occurred in 100% of "tumor viruses" (N=250), in 97% of "cancer proteins" (N=401), and in 85% of "transforming viruses" (N=248). These results suggest that there are shared properties of cancer pathogenesis regardless of cell type and suggest a role of viruses in carcinogenesis, i.e., conversion of cells from a transformed albeit dormant state to a more virulent actively replicating state.

[0123] Homologues of the following amino acid sequence, kagvafllhkk, as defined by the three point recognition method, were found in such viruses, or viral peptides, as, but not limited to, adenovirus, lentivirus, a-virus, retrovirus, andeno-associated virus, human immunodeficiency virus, hepatitis virus, influenza virus, maize streak virus, herpes virus, bovine herpes virus, feline immunodeficiency virus, foot and mouth disease virus, small pox virus, rous sarcoma virus, neuroblastoma RAS viral oncogene, polyomavirus, sindbis, human papilloma

virus, myelomonocytic tumor virus, murine acute leukemia, T-cell lymphotropic virus, and tomato leaf curl virus.

[0124] Replikins are present in such bacteria as, but not limited to, *Acetobacter*,
5 *Achromobacter*, *Actinomyces*, *Aerobacter*, *Alcaligenes*, *Arthrobacter*,
Azotobacter, *Bacillus*, *Brevibacterium*, *Chainia*, *Clostridium*, *Corynebacterium*,
Erwinia, *Escheria*, *Lebsiella*, *Lactobacillus*, *Haemophilus*, *Flavobacterium*,
Methylomonas, *Micrococcus*, *Mycobacterium*, *Micronomspora*, *Mycoplasma*,
Neisseria, *Nocardia*, *Proteus*, *Pseudomonas*, *Rhizobium*, *Salmonella*, *Serratia*,
10 *Staphylococcus*, *Streptococcus*, *Streptomyces*, *Streptosporangium*,
Streptovorticillium, *Vibrio*, peptide, and *Xanthomas*.

[0125] Replikins are present in such fungi as, but not limited to, *Penicillium*,
Diseula, *Ophiostoma novo-ulim*, *Mycophycophta*, *Phytophthora infestans*,
15 *Absidia*, *Aspergillus*, *Candida*, *Cephalosporium*, *Fusarium*, *Hansenula*, *Mucor*,
Paecilomyces, *Pichia*, *Rhizopus*, *Torulopsis*, *Trichoderma*, and *Erysiphe*.

[0126] Replikins are present in such yeast as, but not limited to, *Saccharomyces*,
Cryptococcus, including *Cryptococcus neoformans*, *Schizosaccharomyces*, and
20 *Oryza*.

[0127] Replikins are present in algae such as, but not limited to, *Caldophera*,
Isolepisprolifera, *Chondrus*, *Gracilaria*, *Gelidium*, *Caulerpa*, *Laurencia*,
Cladophexa, *Sargassum*, *Penicillos*, *Halimeda*, *Laminaria*, *Fucus*, *Ascophyllum*,
25 *Undari*, *Rhodymenia*, *Macrocystis*, *Eucheuma*, *Ahnfeltia*, and *Pteroclasia*.

[0128] Replikins are present in amoeba such as, but not limited to, Entamoeba (including Entamoeba invadens), Amoebidae, Acanthamoeba and Naegleria.

5 [0129] Replikins are present in plants such as, but not limited to, Arabidopsis, wheat, rice, and maize.

AUXILIARY SPECIFICATIONS

10 [0130] To permit classification of subtypes of Replikins, additional or "auxiliary specifications" to the basic "3-point-recognition" requirements may be added: (a) on a structural basis, such as the common occurrence of adjacent di- and polylysines in cancer cell proteins (e.g., transforming protein P21B(K-RAS 2B), lung, Table 2, SEQ ID NO.: 89), and other adjacent di-amino acids in TOLL-like
15 receptors, or b) on a functional basis, such as exhibiting ATPase, tyrosine kinase or redox activity as seen in Table 2.

FUNCTIONAL DERIVATIVES

[0131] "Functional derivatives" of the Replikins as described herein are
20 fragments, variants, analogs, or chemical derivatives of the Replikins, which retain at least a portion of the immunological cross reactivity with an antibody specific for the Replikin. A fragment of the Replikin peptide refers to any subset of the molecule. Variant peptides may be made by direct chemical synthesis, for example, using methods well known in the art. An analog of a Replikin to a non-
25 natural protein substantially similar to either the entire protein or a fragment

thereof. Chemical derivatives of a Replikin contain additional chemical moieties not normally a part of the peptide or peptide fragment.

[0132] As seen in Figure 2, during anaerobic respiration when the rate of cell replication is increased, malignin is enriched. That is, malignin is found to increase not simply in proportion to the increase in cell number and total membrane proteins, but is enriched as much as ten-fold in concentration, starting with 3% at rest and reaching 30% of total membrane protein. This clear demonstration of a marked increase in Replikin concentration with glioma cell replication points to, and is consistent with, the presence of Replikins identified with the 3-point recognition method in various organisms. For example, Replikins were identified in such proteins as "Saccharomyces cerevisiae replication binding protein" (SEQ ID NO.: 2) (hsikrelgiifdk); the "replication associated protein A of maize streak virus" (SEQ ID NO.: 8) (kyivcareahk) and (SEQ ID NO.: 9) (kekkpskdeimrdiish); the "replication-associated protein of Staphylococcus aureus" (SEQ ID NO.: 10) (kkektthink); the "DNA replication protein of bovine herpes virus 4" (SEQ ID NO.: 11) (hkinitngqk); and the "Mealigrad herpes virus 1 replication binding protein" (SEQ ID NO.: 12) (hkdlyrllmk). Previous studies of tomato leaf curl gemini virus show that the regulation of virus accumulation appears to involve binding of amino acids 1-160 of the "replicating protein" of that virus to leaf DNA and to other replication protein molecules during virus replication. Analysis of this sequence showed that amino acids 1-163 of this "replicating protein" contain five Replikins, namely: (SEQ ID NO.: 13) kfrinaknyfltyph, (SEQ ID NO.: 14) knletpvnlfiricrefh, (SEQ ID NO.: 15)

hpniqaaksstdvk, (SEQ ID NO.: 16) ksstdvkaymdkdgdvldh, and (SEQ ID NO.:
17) kasalnilrekapkdfvlqfh.

[0133] Table 2 shows that Replikin-containing proteins also are associated

5 frequently with redox functions, and protein synthesis or elongation, as well as
with cell replication. The association with metal-based redox functions, the
enrichment of the Replikin-containing glioma malignin concentration during
anaerobic replication, and the cytotoxicity of antimalignin at low concentrations
(picograms/cell) (Figure 4c-f), all suggest that the Replikins are related to central
10 respiratory survival functions, have been found less often subjected to the
mutations characteristic of non-Replikin amino acids.

[0134] Of particular interest, it was observed that at least one Replikin per

100 amino acids was found to be present in the hemagglutinin proteins of almost
15 all of the individual strains of influenza viruses examined. The Replikin
sequences that were observed to occur in the hemagglutinin proteins of isolates of
each of the four prevalent strains of influenza virus, influenza B, H1N1, H2N2,
and H3N2, for each year that amino acid sequence data are available (1902-2001),
are shown in Tables 3, 4, 5 and 6, below.

20

Table 3

Replikin Sequences present in hemagglutinins of Influenza B viruses in each year for which amino acid sequences were available (1902-2001).

<i>Influenza B Replikins</i>		<i>Year Detected in Influenza B strain</i>	
(Peak in Figure 7:		<u>EB1</u>	<u>EB2</u>
kshfanlk (SEQ ID NO. 104)		1902,19,24,38,40,43, <u>51</u> ,59,75,76, <u>77</u> ,89,90,93,97,98,99,00,01	
kshfanlkgtk (SEQ ID NO. 105)		1902,19,24,38,40,43, <u>51</u> ,59,75,76, <u>77</u> ,89,90,93,97,98,99,00,01	
kshfanlkgtktrgkclcpk (SEQ ID NO. 106)		1902,19,24,38,40,43, <u>51</u> ,59,75,76, <u>77</u> ,89,90,93,97,98,99,00,01	
hekygglnk (SEQ ID NO. 107)		1902,19,24,38,40,43, <u>51</u> ,59,75,76, <u>77</u> ,89,90,93,97,98,99,00,01	
hekygglnsk (SEQ ID NO. 108)		1902,19,24,38,40,43, <u>51</u> ,59,75,76, <u>77</u> ,89,90,93,97,98,99,00,01	
hekygglnskpyytgehak (SEQ ID NO. 109)		1902,19,24,38,40,43, <u>51</u> ,59,75,76, <u>77</u> ,89,90,93,97,98,99,00,01	
hakaigncpiwvk (SEQ ID NO. 110)		1902,19,24,38,40,43, <u>51</u> ,59,75,76, <u>77</u> ,89,90,93,97,98,99,00,01	
hakaigncpiwvktplklangtk (SEQ ID NO. 111)		1902,19,24,38,40,43, <u>51</u> ,59,75,76, <u>77</u> ,89,90,93,97,98,99,00,01	
hakaigncpiwvktplklangtkyrppak (SEQ ID NO. 112)		1902,19,24,38,40,43, <u>51</u> ,59,75,76, <u>77</u> ,89,90,93,97,98,99,00,01	
hakaigncpiwvktplklangtkyrppakllk (SEQ ID NO. 113)		1902,19,24,38,40,43, <u>51</u> ,59,75,76, <u>77</u> ,89,90,93,97,98,99,00,01	
hfanlkgtktrgk (SEQ ID NO. 114)		1919,	76, 89,90, 99,00,01
hfanlkgtktrgkclcpk (SEQ ID NO. 115)		1919,	76, 90 00,01
hsdneiqmvklygdsk (SEQ ID NO. 116)		1919	
hsdneiqdkmvklygdskpqr (SEQ ID NO. 117)		1919	
hsdneiqmvklygdskpqr (SEQ ID NO. 118)		1919, 24,	97,98, 00
k(a/v)silhevk (SEQ ID NO. 119)		1919, 40, 59,	90,93
kctgtipsakasilh (SEQ ID NO. 120)		1919,	00
kctgtipsakasilhevk (SEQ ID NO. 121)		1919,	93

kygginkskpyytgeh (SEQ ID NO. 122)	1919								
kvwcasgrskvikgsipligeadclh (SEQ ID NO. 123)	1919,	38,40,43,	59,75,76, <u>77</u> ,89,90,	98,99,00					
kpyytgehak (SEQ ID NO. 124)	1919,	38,40,	59,	89,90,93,97,98,	01				
kcmgtipsakasilhevk (SEQ ID NO. 125)	1924,	43,	75,76, <u>77</u> ,	93					
hnvinaekapggpyk (SEQ ID NO. 126)	1938,			93,97,	00				
hsdnetqmaklygdsk (SEQ ID NO. 127)	1938,			93,97,	00				
hgvavaadlkstqeaink (SEQ ID NO. 128)	1940,	59,	00						
hgvavaadlkstqeainkdtistqeaink (SEQ ID NO. 129)	1940								
klygdskpqkftssangvith (SEQ ID NO. 130)	1943,	75,76, <u>77</u> ,	93,97,	00					
hsdnetqmaklygdskpqk (SEQ ID NO. 131)	1943,	75,76, <u>77</u> ,	93						
hfankgtqtrgk (SEQ ID NO. 132)	1959								
kprsalkckgfh (SEQ ID NO. 133)	1988								
kskpyytgehakai(g/a)ncpiwvk (SEQ ID NO. 134)	2000								

1. Influenza B has not been responsible for any human pandemic (global distribution).
2. Abbreviation for years: eg. "19" = 1919, "01" = 2001.
3. The first year that a given Replikin appears is indicated at the beginning of the series of years in which that Replikin has been found.
4. Overlapping Replikin sequences are listed separately.
5. Increase in number of new Replikin structures occurs in years of epidemics (underlined): eg. 1951 and 1977 and correlates with increased total Replikin concentration (number of Replikins per 100 amino acid residues). See Figure 7.

Table 4

H1N1 Replikin Sequences present in H1N1 hemagglutinins of Influenza viruses in each year for which amino acid sequences were available (1918-2000)

H1N1 Replikin Year Detected in Influenza

H1N1 Strain

Peak in Figure7 : P1 E1 E1.1, 1.2, 1.3 E1.4)

hp(v/i)tigecpkyv(r/k)(s/t)(t/a)k (SEQ ID NO. 135)	1918,25,28,30,31,35,47,48,51,52,55,56,57,59,63,77,79,80,81,85,87,88,89,91,92,95,96,97,98,99,00				
hdsnvknly(e/g)kv(k/r)(n/s)ql(k/r)nnak (SEQ ID NO. 136)	1918, 28,30,31,	77,79,80,	88, 91, 95, 98		
hdsnvknly(e/g)kv(k/r)(n/s)qlk (SEQ ID NO. 137)	1918, 28,30,31,	77,79,80,	88, 91, 95, 98		
hkc(nn/dd)(a/t/e)cmesv(r/k)ngtydypkyseesklre(e/k)idgvk (SEQ ID NO. 138)	1918, 30, 35,	77, 80,			98
hkc(nn/dd)(a/t/e)cmesv(r/k)ngtydypkyseesk (SEQ ID NO. 139)	1918, 30, 35,	77, 80,			98
hqn(e/g)qgsqyaadqkstqnai(d/n)gitmkvnsviekmntqftavgkefnklek (SEQ ID NO. 140)	1918, 28,30,31,35,	59, 79,		95	
hqn(e/g)qgsqyaadqkstqnai(d/n)gitmkvnsviek (SEQ ID NO. 141)	1918, 28,30,31,35,	59, 79,		95	
hqn(e/g)qgsqyaadqkstqnai(d/n)gitmk (SEQ ID NO. 142)	1918, 28,30,31,35,	59, 79,		95	
kfeifpktsswpnh (SEQ ID NO. 143)	1918,	77			
kg(n/s/t)sypkl(n/s)ksy(v/t)nnkgkevlvlgvvh (SEQ ID NO. 144)	1918, 35,	77		96	
ksy(v/t)nnkgkevlvlgvvh (SEQ ID NO. 145)	1918, 35,	77		96	
hkcncmesvknngtydypkyseesklrekidgvk (SEQ ID NO. 146)	1928, 31,			95	
hkcncmesvknngtydypkyseesk (SEQ ID NO. 147)	1928, 31,			95	
hkcncmesvknngtydypk (SEQ ID NO. 148)	1928, 31,			95	
hkcncmesvk (SEQ ID NO. 149)	1928, 31,			95	
hngkssfy(k/r)nllwlt(e/g)knglypnlsksyvnnkek (SEQ ID NO. 150)	1928,			95,	00
hngkssfy(k/r)nllwlt(e/g)knglypnlsksyvnnk (SEQ ID NO. 151)	1928, 31,			95,	00
hngkssfy(k/r)nllwlt(e/g)knglypnlsk (SEQ ID NO. 152)	1928, 31,			95,	00

hngkssfy(k/r)nllwlt(e/g)k (SEQ ID NO. 153)	1928, <u>31</u> ,		95,	00
kssfyknllwiteknglypnlsksyvnnkekevlwgvh (SEQ ID NO. 154)	1928, <u>31</u> ,		95	
knllwiteknglypnlsksyvnnkekevlwgvh (SEQ ID NO. 155)	1928, <u>31</u> ,		95	
knglypnlsksyvnnkekevlwgvh (SEQ ID NO. 156)	1928, <u>31</u> ,		95,96,	00
ksy(v/a)nnkekev(l/-)(v/-)lwgvh (SEQ ID NO. 157)	1928, <u>31</u> ,	51,	95,96,	98, 00
kesswpnhtvtk (SEQ ID NO. 158)	1928, <u>31</u> ,		95	
het(t/n)kgvtaacpyagassfyrnllwlvkkensypklksysvnnk (SEQ ID NO. 159)	1930, 35			
het(t/n)kgvtaacpyagassfyrnllwlvkkensypklsk (SEQ ID NO. 160)	1930, 35			
kfeifpktsswpnevvlwgvh (SEQ ID NO. 161)	1930			
karswpkh (SEQ ID NO. 162)	1947, 51,52,55,56,	<u>79</u> ,	82	
kisksyvnnkekevlwqvvh (SEQ ID NO. 163)	1947, 51			
kankekevlwqvvh (SEQ ID NO. 164)	1947			
h(k/n)(g/q)kssfy(r/k)nllwitekng(l/s)yp(n/t)lsksyannkek (SEQ ID NO. 165)	1948	<u>79</u> ,	89,	<u>96</u>
h(k/n)(g/q)kssfy(r/k)nllwitek (SEQ ID NO. 166)	1948	<u>79</u> ,	89,	<u>96</u>
hakksfyk (SEQ ID NO. 167)	<u>1951</u> ,	57,59		
hngkclerkkgk (SEQ ID NO. 168)	<u>1951</u> ,52,55,56,57,59,	<u>79</u> ,		
hyklnn(q/g)kk (SEQ ID NO. 169)		1956,		00
hdiyrdeainnrfqigvkltagyk (SEQ ID NO. 170)		1956		
kgngcfeifhk (SEQ ID NO. 171)		1956		
klrlietmdkyhqiek (SEQ ID NO. 172)		1956		
klrlietmdkyh (SEQ ID NO. 173)		1956		
kchtdkgsisttk (SEQ ID NO. 174)		1956		
kiningdyakdyiwgvh (SEQ ID NO. 175)		1956		
hngkclerkgiaplqlgk (SEQ ID NO. 176)		1959,	82	

hetnrqvtaacpyagansffmliwlvkkessypklsk (SEQ ID NO. 177)	<u>1963</u>	81
hetnrqvtaacpyagansffmliwlvkkessypk (SEQ ID NO. 178)	<u>1963</u>	81
hpptstdqqslyqnadayifvgsskynrkfk (SEQ ID NO. 179)	<u>1963</u>	81
hpptstdqqslyqnadayifvgsskynrkfkpeia (SEQ ID NO. 180)	<u>1963</u>	81
hdiyrdeainnrfiqigvkitqgyk (SEQ ID NO. 181)	<u>1977,79</u>	91
hqneqsgsgyaadqkstknaidgitnkvnsviekmntqftavgk (SEQ ID NO. 182)	<u>1977</u>	
hqneqsgsgyaadqkstknaidgitnkvnsviek (SEQ ID NO. 183)	<u>1977</u>	
hqneqsgsgyaadqkstknaingitnkvnsviekmntqftavgkefnklek (SEQ ID NO. 184)	<u>1979</u>	91
hngklcrkigiaplqlgk (SEQ ID NO. 185)	<u>1979</u>	
hkcncnecmesvk (SEQ ID NO. 186)	<u>1979</u>	
kfeifpkasswpnh (SEQ ID NO. 187)	1981	
hdsnvknlyekvrsqimnak (SEQ ID NO. 188)	1981	
kvnsvikkmntqfaavgkefnh (SEQ ID NO. 189)	1981	
khngklck (SEQ ID NO. 190)	1981	
kkgtsypklksytnkgkevlvlgvh (SEQ ID NO. 191)	1981	
kgtsypklksytnkgkevlvlgvh (SEQ ID NO. 192)	1981	
kisksytnkgkevlvlgvh (SEQ ID NO. 193)	1981	
ksytnkgkevlvlgvh (SEQ ID NO. 194)	1981	
kgvtascshk (SEQ ID NO. 195)	1985,87	
kgvtascshkgrssfymllwiteknglypnlsk (SEQ ID NO. 196)	1985,87	
kgnsypklksyvnnkekevlvlgih (SEQ ID NO. 197)	1988	
kefnhle (SEQ ID NO. 198)	1988	
hpptstdqqslyqnadayvfgsskynkkfkpeiatrp (SEQ ID NO. 199)	1988	
hpptstdqqslyqnadayvfgsskynkkfk (SEQ ID NO. 200)	1988	

hegkssfymlwitekegsypklksyvnk (SEQ ID NO. 201)	1991	
hegkssfymlwitekegsypk (SEQ ID NO. 202)	1991	
hkcdnecmesvrngtydypkyseesk (SEQ ID NO. 203)	1991	
kesswpnhvtk (SEQ ID NO. 204)	1991,92	
knllwiteknglypnlsksyvnkekeilvlgvvh (SEQ ID NO. 205)	1991,92, <u>96</u>	
hngkssfy(k/m)(n/-)llwlt(e/g)(-/k)knglypnlsk (SEQ ID NO. 206)	1991,92, <u>96</u>	00
hngkssfyknllwitek (SEQ ID NO. 207)	1991,92, <u>96</u>	
htvtkgvtascshngkssfyknllwiteknglypnlsksyvnkekevlvlgvvh (SEQ ID NO. 208)	1995	
htvt(k/g)gv(t/s)ascshngkssfy(k/m)(n/-)llwlt(e/g)k(-n/k)glypnlsk (SEQ ID NO. 209)	1995,	00
htvtkgvtascshngkssfyknllwitek (SEQ ID NO. 210)	1995	
kyvrstklrmvtglmpisqsrqifgaigfeggtgmtdgwygyh (SEQ ID NO. 211)	1995	
hqneqsgyaadqkstknaingitnkvnsiiekmatqftavgk (SEQ ID NO. 212)	1995	
hqneqsgyaadqkstknaingitnkvnsiiek (SEQ ID NO. 213)	1995	
hqneqsgyaadqkstknaingitnk (SEQ ID NO. 214)	1995	
hsgarsfyrnllwivkkgsnsypk (SEQ ID NO. 215)	<u>1996</u>	
hsgarsfyrnllwivkkgsnsypklnk (SEQ ID NO. 216)	<u>1996</u>	
hsgarsfyrnllwivkkgsnsypklksytnck (SEQ ID NO. 217)	<u>1996</u>	
hsgarsfyrnllwivkkgsnsypklksytnckgk (SEQ ID NO. 218)	<u>1996</u>	
htvskgvtscshngk (SEQ ID NO. 219)	<u>1996</u>	
katswpnhettk (SEQ ID NO. 220)	<u>1996</u>	
kqvtscshnqk (SEQ ID NO. 221)	<u>1996</u>	
kgnsypklksytnckgkevlvlgvvh (SEQ ID NO. 222)	<u>1996</u>	
klksytnckgkevlvlgvvh (SEQ ID NO. 223)	<u>1996</u>	
ksytnckgkevlvlgvvh (SEQ ID NO. 224)	<u>1996</u>	

hnqkssfyrnlwlt(e/q)knglypnlsksy(v/a)annkek (SEQ ID NO. 225)	1997,98,99
hptigecpkyvrsak (SEQ ID NO. 226)	1997
hqneqsgyaadqkstqnaingitknvsviekmntqftavgk (SEQ ID NO. 227)	1998
hqneqsgyaadqkstqnaingitknvsviek (SEQ ID NO. 228)	1998
hngkssfyrnlwltelnglypnlsksyvnnkek (SEQ ID NO. 229)	1998

-
1. Influenza H1N1 was responsible for the human pandemic (global distribution) of 1918.
 2. Abbreviation for years: eg. "96" = 1996.
 3. The first year that a given Replikin appears is indicated at the beginning of the series of years in which that Replikin has been found in this work.
 4. Overlapping Replikin sequences are listed separately.
 5. *Increase in number of new Replikin structures occurs in years of epidemics (underlined): eg. 1918 and 1977 and correlates with increased total Replikin concentration (number of Replikins per 100 amino acid residues). See Figure 7.*

Table 5

Replikin Sequences present in hemagglutinins of Influenza H2N2 viruses in years 1957-2000

Influenza H2N2 Replikins Year Detected in Influenza H2N2 strain

(Peak in Figure 8: P2 E2)

khfekvkilpk (SEQ ID NO. 230)	<u>1957</u> , 58, 59, 60, 61, 64, <u>65</u> , 68,	78, 83, 84, 91
khllssvkhfekvk (SEQ ID NO. 231)	<u>1957</u> , 58, 59, 60, 61,	83, 84, 91
ha(k/q/m)(d/n)ilekthngk (SEQ ID NO. 232)	<u>1957</u> , 58, 59, 60, 61, 64, <u>65</u> , 68,	78, 83, 84, 91, 95
ha(k/q/m)(d/n)ilekthngkic(k/r) (SEQ ID NO. 233)	<u>1957</u> , 58, 59, 60, 61, 64, <u>65</u> , 68,	78, 83, 84, 91, 95
lnvhlptigecpkvksek (SEQ ID NO. 234)	<u>1957</u> , 58, 59, <u>65</u> , 68	
hpltigecpkvksek (SEQ ID NO. 235)	<u>1957</u> , 58, 59, <u>65</u> , 68, 64, 65, 68, 78, 83, 84, 91	
khllssvkhfekvkilpk (SEQ ID NO. 236)	<u>1957</u> , 58, 59, 60, 61, 64, <u>65</u> , 68,	78
krqssgimktegtlencetkqtpigainttlfhnh (SEQ ID NO. 237)	<u>1957</u> , 59,	83
kgsnyp(v/i)ak(g/r)syntsgseqmliiwq(v/i)h (SEQ ID NO. 238)	<u>1957</u> , 58, 59, 61,	83, 91, 95
htlgqsracavsgnpsffmmvwltekgsnypvak (SEQ ID NO. 239)	<u>1957</u>	
khfekvk (SEQ ID NO. 240)	<u>1957</u> , 59, <u>65</u>	
kiskrgssgimktegtlencetkqtpigainttlfh (SEQ ID NO. 241)	<u>1957</u> , 59, <u>65</u> ,	91
krqssgimktegtlencetkqtpigainttlfh (SEQ ID NO. 242)	<u>1957</u> , 59, <u>65</u> ,	91
ktegtlencetkqtpigainttlfh (SEQ ID NO. 243)	<u>1957</u> , 59, <u>65</u> ,	91
kiskrgssgimktegtlencetkqtpigainttlfh (SEQ ID NO. 244)	<u>1957</u> , 59, <u>65</u> ,	91
ktegtlencetkqtpigainttlfh(v/i)h (SEQ ID NO. 245)	<u>1957</u> , 59, <u>65</u> ,	91
kiskrgssgimktegtlencetkqtpigainttlfh (SEQ ID NO. 246)	<u>1957</u> , 59, <u>65</u> ,	91

19	k(e/g)snypvakgsynntsgeqmliiwgvh (SEQ ID NO. 247)	1957,	60,	65	
	hpltigecpkyvksek (SEQ ID NO. 248)	1957,	60,	65	
	kcqtplgaiktllpfh (SEQ ID NO. 249)	1957,		65	
	hsndqsgsgyaadkestqka(f/i)dgimkvnsviek-				
	-mntqfeavgklf(n/s)nleklelnkk (SEQ ID NO. 250)		1961,	65,68,	83,84
	hsndqsgsgyaadkestqka(f/i)dgimkvnsviek-				
	-mntqfeavgklf(n/s)nleklelnkk (SEQ ID NO. 251)		1961,	65, 68,	83,84
	hsndqsgsgyaadkestqka(f/i)dgimk (SEQ ID NO. 252)		1961,	65, 68,	83,84
	hdsnvrnlydkvrnqlrdnak (SEQ ID NO. 253)		1964,	68,76,	84,91
	hkcddecrnsvknngtydypklnrneikgvk (SEQ ID NO. 254)		1964,65,68,76,		83,84,91
	hkcddecrnsvknngtydypklnrneik (SEQ ID NO. 255)		1964,65,68,76,		83,84,91
	hkcddecrnsvknngtydypk (SEQ ID NO. 256)		1964,65,68,76,		83,84,91
	hkcddecrnsvk (SEQ ID NO. 257)		1964,65,68,76,		83,84,91
	kgsnypvakgsynntngeqiliiwgvh (SEQ ID NO. 258)			1976,78	
	hsndqsgsgyaadkestqkavdgimkvnsviekmntqfeavgk (SEQ ID NO. 259)			1976,	91
	krsgsgimktegtlencetkcqtplgaintllpfh (SEQ ID NO. 260)			1976,78,	83,84
	hpltigecpkyvksek (SEQ ID NO. 261)			1976	
	hakdilekthngklck (SEQ ID NO. 262)			1976	

1. Influenza H2N2 was responsible for the human pandemic (global distribution) of 1957.
2. Abbreviation for years: eg. "58" =1958.

3. The first year that a given Replikin appears is indicated at the beginning of the series of years in which that Replikin has been found in this work.
4. Overlapping Replikin sequences are listed separately.
5. Increase in number of new Replikin structures occurs in years of epidemics (underlined): eg. 1957 and 1965 and correlates with increased total Replikin concentration (number of Replikins per 100 amino acid residues). See Figure 8.

Table 6

H3N2 Replikin Sequences present in H3N2 hemagglutinins of Influenza viruses in each year for which amino acid sequences were available (1968-2000)

Influenza H3N2 ReplikinsYear Detected in Influenza H3N2 strain Influenza Replikins

(Peak in Figure 8: P3

E3

E4)

hdvyrdealnrfqikgvelksy (SEQ ID NO. 263)	<u>1968,72,</u>	<u>75</u>		96,97,98
htidltldsemnklfertrk (SEQ ID NO. 264)	<u>1968</u>			
kfhqiek (SEQ ID NO. 265)	<u>1968,72,</u>	<u>75,</u>	<u>77</u>	96,97,98
ktnekfh(g/q)iek (SEQ ID NO. 266)	<u>1968</u>		86	98
klmr(v/l)iektnekfh (SEQ ID NO. 267)	<u>1968,72,</u>	<u>75,</u>	<u>77</u>	97,98
hgiekefsevegriqdlekyvedtk (SEQ ID NO. 268)	<u>1968,72,</u>			98
kicnnphk (SEQ ID NO. 269)		<u>1975</u>		
klmrviikktnekfh (SEQ ID NO. 270)		<u>1975</u>		
hd(I,v)yrdealnrfqik(g/q)ve(r/k)s(q/g)yk (SEQ ID NO. 271)		<u>1975,76,77,</u>	86	
hgiekefsevegriqdlekyvedtk (SEQ ID NO. 272)		<u>1975</u>		
kyvedtkidlwsynaellvalenqh (SEQ ID NO. 273)		<u>1975</u>		
kyvkqnsiklatgmrvpektqtrglfgaiagfiengwegmidgwygfrh (SEQ ID NO. 274)		<u>1975</u>		
kefsevegriqdlekyvedtkidlwsynaellvalenqh (SEQ ID NO. 275)		<u>1975</u>		<u>2000</u>
hqn(s/e)(e/q)g(t/s)g(q/y)aad(l/q)k-				
-stq(a/n)a(i/l)d(q/g)l(n/t)(g/n)k(l/v)n(r/s)vi(e/c)k (SEQ ID NO. 276)		<u>1975</u>		<u>2000</u>
hcd(g/q)f(q,r)nekwdlf(v,i)er(s/t)k (SEQ ID NO. 277)		<u>1975,76,77,78,80,81,82,83,84,85,86,88,89,90,91,92,93,94,95,96,97,98</u>		

htidldsemnkkkfetrk (SEQ ID NO. 278)	<u>1977</u>		
ksgstypvlkvtmpnndnfdklyiwgvh (SEQ ID NO. 279)	<u>1977</u>		
klnwltsksgntypvlnvtmpnndnfdklyiwgvh (SEQ ID NO. 280)		1982	
htidldsemnkkkfetrk (SEQ ID NO. 281)			1986
klnrlietmekfhqtek (SEQ ID NO. 282)			<u>1987</u>
htgkssvmrsdapidfcnsecitpnqsiipndkpfqnvnkitygacpk (SEQ ID NO. 283)			1994
htgkssvmrsdapidfcnsecitpnqsiipndkpfqnvnk (SEQ ID NO. 284)			1994
hpstdsdqtslyvrasgrvtvstkrsgqtvipk (SEQ ID NO. 285)			1994
kyvedtkidlsynaellvalenqh (SEQ ID NO. 286)			1997,98
klfetrkqlrenaedmgngcfkiyh (SEQ ID NO. 287)			1998
krrsikssfrlnwlh (SEQ ID NO. 288)			1998
hptigecpky(v/r)kstk (SEQ ID NO. 289)			<u>2000</u>
kgnsypklsklsksyiinkkkevliwgih (SEQ ID NO. 290)			<u>2000</u>
klsklsks(v/y)iinkkkevliwgih (SEQ ID NO. 291)			<u>2000</u>
klks(v/y)iinkkkevliwgih (SEQ ID NO. 292)			<u>2000</u>

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1. Influenza H3N2 was responsible for the human pandemic (global distribution) of 1968.
 2. Abbreviation for years: eg. "77" =1977.
 3. The first year that a given Replikin appears is indicated at the beginning of the series of years in which that Replikin has been found.
 4. Overlapping Replikin sequences are listed separately.
 5. Increase in number of new Replikin structures occurs in years of epidemics (underlined) : eg. 1975 and correlates with increased total Replikin concentration (number of Replikins per 100 amino acid residues). See Figure 8.

[0135] Both the concentration and type, i.e., composition of Replikins observed, were found to relate to the occurrence of influenza pandemics and epidemics. The concentration of Replikins in influenza viruses was examined by visually scanning the hemagglutinin amino acid sequences published in the National Library of Medicine "PubMed" data base for influenza strains isolated world wide from human and animal reservoirs year by year over the past century, i.e., 1900 to 2001. These Replikin concentrations (number of Replikins per 100 amino acids, mean +/- SD) were then plotted for each strain.

[0136] The concentration of Replikins was found to directly relate to the occurrence of influenza pandemics and epidemics. The concentration of Replikins found in influenza B hemagglutinin and influenza A strain, H1N1, is shown in Figure 7, and the concentration of Replikins found in the two other common influenza virus A strains, H2N2 and H3N2 is shown in Figure 8 (H2N2, H3N2). The data in Figure 8 also demonstrate an emerging new strain of influenza virus as defined by its constituent Replikins (H3N2(R)).

[0137] Each influenza A strain has been responsible for one pandemic: in 1918, 1957, and 1968, respectively. The data in Figures 7 and 8 show that at least one Replikin per 100 amino acids is present in each of the influenza hemagglutinin proteins of all isolates of the four common influenza viruses examined, suggesting a function for Replikins in the maintenance of survival levels of replication. In the 1990s, during the decline of the H3N2 strain, there were no Replikins in many isolates of H3N2, but a high concentration of new Replikins appeared in H3N2 isolates, which define the emergence of the H3N2(R) strain.

[0138] Several properties of Replikin concentration are seen in Figure 7 and Figure 8 to be common to all four influenza virus strains. First, the concentration is cyclic over the years, with a single cycle of rise and fall occurring over a period of two to thirty years. This rise and fall is consistent with the known waxing and waning of individual influenza virus strain predominance by hemagglutinin and neuraminidase classification. Second, peak Replikin concentrations of each influenza virus strain previously shown to be responsible for a pandemic were observed to relate specifically and individually to each of the three years of the pandemics. For example, for the pandemic of 1918, where the influenza virus strain, H1N1, was shown to be responsible, a peak concentration of the Replikins in H1N1 independently occurred (P1); for the pandemic of 1957, where H2N2 emerged and was shown to be responsible, a peak concentration of the Replikins in H2N2 occurred (P2); and for the pandemic of 1968, where H3N2 emerged and was shown to be the cause of the pandemic, a peak concentration of the Replikins in H3N2 occurred (P3). Third, in the years immediately following each of the above three pandemics, the specific Replikin concentration decreased markedly, perhaps reflecting the broadly distributed immunity generated in each case. Thus, this post-pandemic decline is specific for H1N1 immediately following the pandemic (P1) for which it was responsible, and is not a general property of all strains at the time. An increase of Replikin concentration in influenza B repeatedly occurred simultaneously with the decrease in Replikin concentration in H1N1, e.g., EB1 in 1951 and EB2 in 1976, both associated with influenza B epidemics having the highest mortality. (Stuart-Harris, et al., Edward Arnold Ltd. (1985). Fourth, a secondary peak concentration, which exceeded the primary peak

increase in concentration, occurred 15 years after each of the three pandemics, and this secondary peak was accompanied by an epidemic: 15 years after the 1918 pandemic in an H1N1 'epidemic' year (E1); eight years after the 1957 pandemic in an H2N2 'epidemic' year (E2); and occurred seven years after the 1968 pandemic in an H3N2 'epidemic' year (E3). These secondary peak concentrations of specific Replikins may reflect recovery of the strain. Fifth, peaks of each strain's specific Replikin concentration frequently appear to be associated with declines in Replikin concentration of one or both other strains, suggesting competition between strains for host sites. Sixth, there is an apparent overall tendency for the Replikin concentration of each strain to decline over a period of 35 years (H2N2) to 60 years (influenza B). This decline cannot be ascribed to the influence of vaccines because it was evident in the case of influenza B from 1901 to 1964, prior to common use of influenza vaccines. In the case of influenza B, Replikin recovery from the decline is seen to occur after 1965, but Replikin concentration declined again between 1997 and 2000 (Figure 7). This correlates with the low occurrence of influenza B in recent case isolates. H1N1 Replikin concentration peaked in 1978-1979 (Figure 7) together with the reappearance and prevalence of the H1N1 strain, and then peaked in 1996 coincident with an H1N1 epidemic. (Figure 7). H1N1 Replikin concentration also declined between 1997 and 2000, and the presence of H1N1 strains decreased in isolates obtained during these years. For H2N2 Replikins, recovery from a 35 year decline has not occurred (Figure 8), and this correlates with the absence of H2N2 from recent isolates. For H3N2, the Replikin concentration of many isolates fell to zero during the period from 1996 to 2000, but other H3N2 isolates showed a significant, sharp increase in

Replikin concentration. This indicates the emergence of a substrain of H3N2, which is designated herein as H3N2(R).

[0139] Figures 7 and 8 demonstrate that frequently, a one to three year
5 stepwise increase is observed before Replikin concentration reaches a peak. This
stepwise increase proceeds the occurrence of an epidemic, which occurs
concurrently with the Replikin peak. Thus, the stepwise increase in concentration
of a particular strain is a signal that particular strain is the most likely candidate to
cause an epidemic or pandemic.

10 [0140] Currently, Replikin concentration in the H3N2(R) strain of influenza
virus is increasing (Figure 8, 1997 to 2000). Three similar previous peak increases
in H3N2 Replikin concentration are seen to have occurred in the H3N2-based
pandemic of 1968 (Figure 8), when the strain first emerged; and in the H3N2-
15 based epidemics of 1972 and 1975 (Figure 8). Each of these pandemic and
epidemics was associated with excess mortality. (Ailing, et al., Am J.
Epidemiol., 113(1):30-43 (1981). The rapid ascent in concentration of the
H3N2(R) subspecies of the H3N2 Replikins in 1997-2000, therefore, statistically
represents an early warning of an approaching severe epidemic or pandemic. An
20 H3N2 epidemic occurred in Russia in 2000 (Figure 8, E4); and the CDC report of
December 2001 states that currently, H3N2 is the most frequently isolated strain of
influenza virus world wide. (Morbidity and Mortality Weekly Reports (MMWR),
Center for Disease Control; 50(48):1084-68 (Dec.7, 2001).

25 [0141] In each case of influenza virus pandemic or epidemic new Replikins

emerge. There has been no observation of two of the same Replikins in a given hemagglutinin in a given isolate. To what degree the emergence of a new Replikin represents mutations versus transfer from another animal or avian pool is unknown. In some cases, each year one or more of the original Replikin structures is conserved, while at the same time, new Replikins emerge. For example, in influenza virus B hemagglutinin, five Replikins were constantly conserved between 1919 and 2001, whereas 26 Replikins came and went during the same period (some recurred after several years absence). The disappearance and re-emergence years later of a particular Replikin structure suggests that the Replikins return from another virus host pool rather than through de novo mutation.

[0142] In the case of H1N1 Replikins, the two Replikins present in the P1 peak associated with the 1918 pandemic were not present in the recovery B1 peak of 1933, which contains 12 new Replikins. Constantly conserved Replikins, therefore, are the best choice for vaccines, either alone or in combination. However, even recently appearing Replikins accompanying one year's increase in concentration frequently persist and increase further for an additional one or more years, culminating in a concentration peak and an epidemic, thus providing both an early warning and time to vaccinate with synthetic Replikins (see for example, H1N1 in the early 1990's, Figure 7).

[0143] The data in FIGURES 7 and 8 demonstrate a direct relationship between the presence and concentration of a particular Replikin in influenza protein sequences and the occurrence of pandemics and epidemics of influenza. Thus, analysis of the influenza virus hemagglutinin protein sequence for the

presence and concentration of Replikins provides a predictor of influenza pandemics and/or epidemics, as well as a target for influenza vaccine formulation.

[0144] Composition of Replikins in Strains of Influenza Virus B: Of a total of 26 Replikins identified in this strain (Table 3), the following ten Replikins are present in every influenza B isolate examined from 1902-2001. Overlapping Replikin sequences are listed separately. Lysines and histidines are in bold type to demonstrate homology consistent with the "3-point recognition."

kshfanlk (SEQ ID NO. 104)

10 kshfanlkgtk (SEQ ID NO. 105)

kshfanlkgtktrgklcpk (SEQ ID NO. 106)

hekygglnk (SEQ ID NO. 107)

hekygglnksk (SEQ ID NO. 108)

hekygglnkskpyytgehak (SEQ ID NO. 109)

15 hakaigncpiwvk (SEQ ID NO. 110)

hakaigncpiwvkvktpklkangtk (SEQ ID NO. 111)

hakaigncpiwkvktpklkangtkyrppak (SEQ ID NO. 112)

hakaigncpiwkvktpklkangtkyrppakllk (SEQ ID NO. 113)

20 [0145] Tables 3 and 4 indicate that there appears to be much greater stability of the Replikin structures in influenza B hemagglutinins compared with H1N1 Replikins. Influenza B has not been responsible for any pandemic, and it appears not to have an animal or avian reservoirs. (Stuart-Harris et al., Edward Arnold Ltd., London (1985)).

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[0146] Influenza H1N1 Replikins: Only one Replikin

"hp(v/i)tigecpkyv(r/k)(s/t)(t/a)k" is present in every H1N1 isolate for which sequences are available from 1918, when the strain first appeared and caused the pandemic of that year, through 2000. (Table 4). ("v/i" indicates that the amino acid v or i is present in the same position in different years.) Although H1N1 contains only one persistent Replikin, H1N1 appears to be more prolific than influenza B. There are 95 different Replikin structures in 82 years on H1N1 versus only 31 different Replikins in 100 years of influenza B isolates (Table 4). An increase in the number of new Replikin structures occurs in years of epidemics (Tables 3, 4, 5 and 6) and correlates with increased total Replikin concentration (Figures 7 and 8).

[0147] Influenza H2N2 Replikins: Influenza H2N2 was responsible for the human pandemic of 1957. Three of the 20 Replikins identified in that strain for 1957 were conserved in each of the H2N2 isolates available for examination on PubMed until 1995 (Table 5).

ha(k/q/m)(d/n)ilekthngk (SEQ ID NO. 232)

ha(k/q/m)(d/n)ilekthngklc(k/r) (SEQ ID NO. 233)

kgsnyp(v/i)ak(g/r)synntsgeqmliiwq(v/i)h (SEQ ID No. 238)

[0148] However, in contrast to H1N1, only 13 additional Replikins have been found in H2N2 beginning in 1961. This paucity of appearance of new Replikins correlates with the decline in the concentration of the H2N2 Replikins and the appearance of H2N2 in isolates over the years. (Figure 8).

[0149] Influenza H3N2 Replikins: Influenza H3N2 was responsible for the human pandemic of 1968. Five Replikins which appeared in 1968 disappeared after 1977, but reappeared in the 1990s (Table 6). The only Replikin structure which persisted for 22 years was hcd(g/q)f(q/r)nekwdlf(v/i)er(s/t)k, which
5 appeared first in 1977 and persisted through 1998. The emergence of twelve new H3N2 Replikins in the mid 1990s (Table 6) correlates with the increase in Replikin concentration at the same time (Figure 8), and with the prevalence of the H3N2 strain in recent isolates together with the concurrent disappearance of all Replikins from some of these isolates (Figure 8), this suggests the emergence of
10 the new substrain H3N2(R).

[0150] Figures 1 and 2 show that influenza epidemics and pandemics correlate with the increased concentration of Replikins in influenza virus, which is due to the reappearance of at least one Replikin from one to 59 years after its
15 disappearance. Also, in the A strain only, there is an emergence of new strain-specific Replikin compositions (Tables 4-6). Increase in Replikin concentration by repetition of individual Replikins within a single protein appears not to occur in influenza virus, but is seen in other organisms.

[0151] It has been believed that changes in the activity of different influenza strains are related to sequence changes in influenza hemagglutinins, which in turn are the products of substitutions effected by one of two poorly understood processes: i) antigenic drift, thought to be due to the accumulation of a series of point mutations in the hemagglutinin molecule, or ii) antigenic shift, in which the
20 changes are so great that genetic reassortment is postulated to occur between the
25

viruses of human and non-human hosts. First, the present data suggests that the change in activity of different influenza strains, rather than being related to non-specific sequence changes, are based upon, or relate to the increased concentration of strain-specific Replikins and strain-specific increases in the replication associated with epidemics. In addition, the data were examined for a possible insight into which sequence changes are due to "drift" or "shift", and which are due to conservation, storage in reservoirs, and reappearance. The data show that the epidemic-related increase in Replikin concentration is not due to the duplication of existing Replikins per hemagglutinin, but is due to the reappearance of at least one Replikin composition from 1 to up to 59 years after its disappearance, plus in the A strains only, the emergence of new strain-specific Replikin compositions (Tables 3-6). Thus the increase in Replikin concentration in the influenza B epidemics of 1951 and 1977 are not associated with the emergence of new Replikin compositions in the year of the epidemic but only with the reappearance of Replikin compositions which had appeared in previous years then disappeared (Table 3). In contrast, for the A strains, in addition to the reappearance of previously disappeared virus Replikins, new compositions appear (e.g. in H1N1 in the year of the epidemic of 1996, in addition to the reappearance of 6 earlier Replikins, 10 new compositions emerged). Since the A strains only, not influenza B, have access to non-human animal and avian reservoirs, totally new compositions probably derive from non-human host reservoirs rather than from mutations of existing human Replikins which appear to bear no resemblance to the new compositions other than the basic requirements of "3-point recognition" (Tables 2-5). The more prolific nature of H1N1 compared with B, and the fact that pandemics have been produced by the three A strains only, but not by the B strain,

both may also be a function of the ability of the human A strains to receive new Replikin compositions from non-human viral reservoirs.

[0152] Some Replikins have appeared in only one year, disappeared, and not
5 reappeared to date (Tables 3-6). Other Replikins disappear from one to up to 81
years, when the identical Replikin sequence reappears. Key Replikin 'k' and 'h'
amino acids, and the spaces between them, are conserved during the constant
presence of particular Replikins over many years, as shown in Tables 23-6 for the
following strain-specific Replikins: ten of influenza B, the single Replikin of
10 H1N1, and the single Replikin of H2N3, as well as for the reappearance of
identical Replikins after an absence. Despite the marked replacement or
substitution activity of other amino acids both inside the Replikin structure and
outside it in the rest of the hemagglutinin sequences, influenza Replikin histidine
(h) appears never to be, and lysine (k) is rarely replaced. Examples of this
15 conservation are seen in the H1N1 Replikin "hp(v/i)tigecpkyv(r/k)(s/t)(t/a)k," (SEQ
ID NO. 135) constant between 1918 and 2000, in the H3N2 Replikin
"hcd(g/q)f(q,r)nekwdlf(v/i)er(s/t)k" (SEQ ID NO. 277) constant between 1975 and
1998 and in the H3N2
Replikin "hqn(s/e)(e/q)g(t/s)g(q/y)aad(l/q)kstq(a/n)a(i/l)d(q/g)l(n/t)(g/n)k,(l/v)n(r/s)
20 vi(e/c)k" (SEQ ID NO. 276) which first appeared in 1975, disappeared for 25
years, and then reappeared in 2000. While many amino acids were substituted, the
basic Replikin structure of 2 Lysines, 6 to 10 residues apart, one histidine, a
minimum of 6% lysine in not more than approximately 50 amino acids, was
conserved.

[0153] Totally random substitution would not permit the persistence of these H1N1 and H3N2 Replikins, nor from 1902 to 2001 in influenza B the persistence of 10 Replikin structures, nor the reappearance in 1993 of a 1919 18mer Replikin after an absence of 74 years. Rather than a random type of substitution, the constancy suggests an orderly controlled process, or in the least, protection of the key Replikin residues so that they are fixed or bound in some way: lysines, perhaps bound to nucleic acids, and histidines, perhaps bound to respiratory redox enzymes. The mechanisms which control this conservation are at present unknown.

10

CONSERVATION OF REPLIKIN STRUCTURES

[0154] Whether Replikin structures are conserved or are subject to extensive natural mutation was examined by scanning the protein sequences of various isolates of foot and mouth disease virus (FMDV), where mutations in proteins of these viruses have been well documented worldwide for decades. Protein sequences of FMDV isolates were visually examined for the presence of both the entire Replikin and each of the component Replikin amino acid residues observed in a particular Replikin.

20 [0155] Rather than being subject to extensive substitution over time as occurs in neighboring amino acids, the amino acids which comprise the Replikin structure are substituted little or not at all, that is the Replikin structure is conserved.

[0156] For example, in the protein VP1 of FMDV type O, the Replikin (SEQ ID NO.: 3) "hkqkivapvk" was found to be conserved in 78% of the 236 isolates reported in PubMed, and each amino acid was found to be conserved in individual isolates as follows: his, 95.6%; lys, 91.8%; gln 92.3%; lys, 84.1%; ile, 90.7%; val, 91.8%; ala, 97.3%; pro, 96.2%; ala, 75.4%; and lys, 88.4%. The high rate of conservation suggests structural and functional stability of the Replikin structure and provides constant targets for treatment.

[0157] Similarly, sequence conservation was found in different isolates of HIV for its Replikins, such as (SEQ ID NO.: 5) "kcfncgkegh" or (SEQ ID NO.: 6) "kvylawvpahk" in HIV Type 1 and (SEQ ID NO.: 7) "kowncgkegh" in HIV Type 2 (Table 2). Further examples of sequence conservation were found in the HIV tat proteins, such as (SEQ ID NO.: 613) "hclvckqkkglgisygrkk," wherein the key lysine and histidine amino acids are conserved. (See Table 7)

[0158] Similarly, sequence conservation was observed in plants, for example in wheat, such as in wheat ubiquitin activating enzyme E (SEQ ID NOs. 614-616). The Replikins in wheat even provided a reliable target for stimulation of plant growth as described within. Other examples of conservation are seen in the constant presence of malignin in successive generations, over ten years of tissue culture of glioma cells, and by the constancy of affinity of the glioma Replikin for antimalignin antibody isolated by immunoadsorption from 8,090 human sera from the U.S., U.K., Europe and Asia (e.g., Figure 5 and U.S. Patent 6,242,578 B1).

[0159] Similarly, conservation was observed in trans-activator (Tat) proteins

in isolates of HIV. Tat (trans-activator) proteins are early RNA binding proteins regulating lentiviral transcription. These proteins are necessary components in the life cycle of all known lentiviruses, such as the human immunodeficiency viruses (HIV). Tat is a transcriptional regulator protein that acts by binding to the trans-activating response sequence (TAR) RNA element and activates transcription Initiation and/or elongation from the LTR promoter. HIV cannot replicate without tat, but the chemical basis of this has been unknown. In the HIV tat protein sequence from 89 to 102 residues, we have found a Replikin that is associated with rapid replication in other organisms. The amino acid sequence of this Replikin is "hclvcfqkkglgisygrkk." In fact, we found that this Replikin is present in every HIV tat protein. Some tat amino acids are substituted frequently, as shown in Table 8, by alternate amino acids (in small size fonts lined up below the most frequent amino acid (Table 7), the percentage of conservation for the predominant Replikin "hclvcfqkkglgisygrkk"). These substitutions have appeared for most of the individual amino acids. However, the key lysine and histidine amino acids within the Replikin sequence, which define the Replikin structure, are conserved 100% in the sequence; while substitutions are common elsewhere in other amino acids, both within and outside the Replikin, none occurs on these key histidine amino acids.

20

[0160] As shown in Table 7, it is not the case that lysines are not substituted in the tat protein amino acid sequence. From the left side of the table, the very first lysine in the immediate neighboring sequence, but outside the Replikin sequence, and the second lysine (k) in the sequence inside the Replikin, but "extra" in that it is not essential for the Replikin formation, are both substituted

25

frequently. However, the 3rd, 4th and 5th lysines, and the one histidine, in parentheses, which together set up the Replikin structure, are never substituted. Thus, these key amino acid sequences are 100% conserved. As observed in the case of the influenza virus Replikins, random substitution would not permit this
5 selective substitution and selective non-substitution to occur due to chance.

Table 7

% Replikin CONSERVATION of each constituent amino acid in the first 117 different isolates of HIV tat protein as reported in PubMed:

38 100 57 86 100 100 66 76 100 99 57 49 100 94 100 97 98 85 97 99 100 100 100%
Neighboring

Amino acids [tat Replikin]

k (c) s y [(h) (c) l v (c) f q k (k) g (l) g i s y g (r) (k) (k)]

below are the amino acid substitutions observed for each amino acid above:

h	c f	q i	l h t	a	a l y h q
r	w p	l l	i h	q	v
y	s	s	l m	r	s
i		s	m s		
s		r	n		
v					
a					
f					
p					
q					

[0161] The conservation of the Replikin structure suggests that the Replikin structure has a specific survival function for the HIV virus which must be

30 preserved and conserved, and cannot be sacrificed to the virus 'defense' maneuver of amino acid substitution crested to avoid antibody and other 'attack.' These 'defense' functions, although also essential, cannot 'compete' with the virus survival function of HIV replication.

35 [0162] Further conservation was observed in different isolates of HIV for its Replikins such as "kcfncgkegh" (SEQ ID NO. 5) or "kvylawvpahk" (SEQ ID NO. 6) in HIV Type 1 and "kcwncgkegh" (SEQ ID NO. 7) in HIV Type 2.

[0163] The high rate of conservation observed in FMVD and HIV Replikins

suggests that conservation also observed in the Replikins of influenza Replikins is a general property of viral Replikins. This conservation makes them a constant and reliable target for either destruction, for example by using specific Replikins such as for influenza, FMDV or HIV vaccines as illustrated for the glioma Replikin, or stimulation.

[0164] Similarly, as provided in examples found in viruses including influenza viruses, FMDV, and HIV, where high rates of conservation in Replikins suggest that conservation is a general property of viral Replikins and thus making Replikins a constant and reliable target for destruction or stimulation, conservation of Replikin structures occurs in plants. For example, in wheat plants, Replikins are conserved and provide a reliable target for stimulation. Examples of conserved Replikins in wheat plants ubiquitin activating enzyme E include:

E3 hkdrltkkvvdiarevakvdpvpyrrh (SEQ ID NO. 614)
E2 hkerldrkvvdvarevakvevpsyrh (SEQ ID NO. 615)
E1 hkerldrkvvdvarevakmevpsyrh (SEQ ID NO. 616)

* * * ** *

[0165] Similarly to conservation found in the HIV tat protein, the Replikin in the wheat ubiquitin activating enzyme E is conserved. As with the HIV tat protein, substitutions of amino acids (designated with an '*') adjacent to the Replikin variant forms in wheat ubiquitin activating enzyme E are common. The key k and h amino acids that form the Replikin structure, however, do not vary whereas the 'unessential' k that is only 5 amino acids (from the first k on the left) is substituted.

ANTI-REPLIKIN ANTIBODIES

[0166] An anti-Replikin antibody is an antibody against a Replikin. Data on anti-Replikin antibodies also support Replikin class unity. An anti-Replikin antibody response has been quantified by immunoadsorption of serum antimalignin

5 antibody to immobilized malignin (see Methods in U.S. Patent #5,866,690). The abundant production of antimalignin antibody by administration to rabbits of the synthetic version of the 16-mer peptide whose sequence was derived from malignin, absent carbohydrate or other groups, has established rigorously that this peptide alone is an epitope, that is, provides a sufficient basis for this immune
10 response (Figure 3). The 16-mer peptide produced both IgM and IgG forms of the antibody. Antimalignin antibody was found to be increased in concentration in serum in 37% of 79 cases in the U.S. and Asia of hepatitis B and C, early, in the first five years of infection, long before the usual observance of liver cancer, which develops about fifteen to twenty-five years after infection. Relevant to both
15 infectious hepatitis and HIV infections, transformed cells may be one form of safe haven for the virus: prolonging cell life and avoiding virus eviction, so that the virus remains inaccessible to anti-viral treatment.

[0167] Because administration of Replikins stimulates the immune system to
20 produce antibodies having a cytotoxic effect, peptide vaccines based on the particular influenza virus Replikin or group of Replikins observed to be most concentrated over a given time period provide protection against the particular strain of influenza most likely to cause an outbreak in a given influenza season., e.g., an emerging strain or re-emerging strain For example, analysis of the
25 influenza virus hemagglutinin amino acid sequence on a yearly or bi-yearly basis,

provides data which are useful in formulating a specifically targeted influenza vaccine for that year. It is understood that such analysis may be conducted on a region-by-region basis or at any desired time period, so that strains emerging in different areas throughout the world can be detected and specifically targeted vaccines for each region can be formulated.

INFLUENZA

[0168] Currently, vaccine formulations are changed twice yearly at international WHO and CDC meetings. Vaccine formulations are based on serological evidence of the most current preponderance of influenza virus strain in a given region of the world. However, prior to the present invention there has been no correlation of influenza virus strain specific amino acid sequence changes with occurrence of influenza epidemics or pandemics.

[0169] The observations of specific Replikins and their concentration in influenza virus proteins provides the first specific quantitative early chemical correlates of influenza pandemics and epidemics and provides for production and timely administration of influenza vaccines tailored specifically to treat the prevalent emerging or re-emerging strain of influenza virus in a particular region of the world. By analyzing the protein sequences of isolates of strains of influenza virus, such as the hemagglutinin protein sequence, for the presence, concentration and/or conservation of Replikins, influenza virus pandemics and epidemics can be predicted. Furthermore, the severity of such outbreaks of influenza can be significantly lessened by administering an influenza peptide vaccine based on the Replikin sequences found to be most abundant or shown to be on the rise in virus

isolates over a given time period, such as about one to about three years.

[0170] An influenza peptide vaccine of the invention may include a single

Replikinpeptide sequence or may include a plurality of Replikin sequences

5 observed in influenza virus strains. Preferably, the peptide vaccine is based on

Replikin sequence(s) shown to be increasing in concentration over a given time

period and conserved for at least that period of time. However, a vaccine may

include a conserved Replikin peptide(s) in combination with a new Replikin(s)

peptide or may be based on new Replikin peptide sequences. The Replikin

10 peptides can be synthesized by any method, including chemical synthesis or

recombinant gene technology, and may include non-Replikin sequences, although

vaccines based on peptides containing only Replikin sequences are preferred.

Preferably, vaccine compositions of the invention also contain a pharmaceutically

acceptable carrier and/or adjuvant.

15

[0171] The influenza vaccines of the present invention can be administered alone

or in combination with antiviral drugs, such as gancyclovir; interferon; interleukin;

M2 inhibitors, such as, amantadine, rimantadine; neuraminidase inhibitors, such as

zanamivir and oseltamivir; and the like, as well as with combinations of antiviral

20 drugs.

REPLIKIN DECOYS IN MALARIA

[0172] Analysis of the primary structure of a Plasmodium falciparum malaria

antigen located at the merozoite surface and/or within the parasitophorous vacuole

revealed that this organism, like influenza virus, also contains numerous Replikins (Table 8). However, there are several differences between the observation of Replikins in *Plasmodium falciparum* and influenza virus isolates. For example, *Plasmodium falciparum* contains several partial Replikins, referred to herein as
5 "Replikin decoys." These decoy structures contain an abundance of lysine residues, but lack the histidine required of Replikin structures. Specifically, these decoys contain many lysines 6 to 10 residues apart in overlapping fashion, similar to the true malaria recognins but without histidine residues. It is believed that the decoy structure maximizes the chances that an anti-malarial antibody or other
10 agent will bind to the relatively less important structure containing the lysines, i.e., the Replikin decoys, rather than binding to histidine, which is present in Replikin structure, such as Replikins in respiratory enzymes, which could result in destruction of the trypanosome. For example, an incoming antibody, with specificity for Replikin structures, might attach to the Replikin decoy structure,
15 leaving the true Replikin structure remains untouched.

[0173] Therefore, anti-Replikin treatment of malaria requires two phases (dual treatment): i) preliminary treatment with proteolytic enzymes that cleave the Replikin decoys, permitting 'safe passage' of the specific anti-Replikin treatment;
20 and ii) attacking malaria Replikins either with specific antibodies or by cellular immunity engendered by synthetic malaria Replikin vaccines or by organic means targeting the malaria Replikins.

REPETITION AND OVERLAPPING OF REPLIKIN STRUCTURES

[0174] Another difference seen in *Plasmodium falciparum* is a frequent repetition of individual Replikin structures within a single protein, which was not observed with influenza virus. Repetition may occur by (a) sharing of lysine residues
5 between Replikins, and (b) by repetition of a portion of a Replikin sequence within another Replikin sequence.

[0175] A third significant difference between Replikin structures observed in influenza virus isolates and *Plasmodium falciparum* is a marked overlapping of
10 Replikin structures throughout malarial proteins, e.g., there are nine overlapping Replikins in the 39 amino acid sequence of SEQ ID NO. 393 (Replikin concentration = 23.1/100 amino acids); and 15 overlapping Replikins in the 41 amino acids of SEQ ID NO. 467 (Replikin concentration = 36.6/100 amino acids). Both of these overlapping Replikin structures occur in blood stage trophozoites
15 and schizonts. In contrast, influenza virus Replikins are more scattered throughout the protein and the maximum Replikin concentration is about 7.5/100 amino acids (Figure 7); and tomato leaf curl gemini virus, which was also observed to have overlapping Replikins has only about 3.1/100 amino acids.

[0176] This mechanism of lysine multiples is also seen in the Replikins of cancer
20 proteins such as in gastric cancer transforming protein, ktkkgnrvsptmkvth (SEQ ID NO. 88), and in transforming protein P21B (K-RAS 2B) of lung, khkekmskdgkkkkks (SEQ ID NO. 89).

[0177] The relationship of higher Replikin concentration to rapid replication is also confirmed by analysis of HIV isolates. It was found that the slow-growing low titer strain of HIV (NSI, "Bru," which is prevalent in early stage HIV infection) has a Replikin concentration of 1.1 (+/- 1.6) Replikins per 100 amino acids, whereas the rapidly-growing high titer strain of HIV (S1, "Lai", which is prevalent in late stage HIV infection) has a Replikin concentration of 6.8 (+/- 2.7) Replikins per 100 amino acid residues.

[0178] The high concentration of overlapping Replikins in malaria, influenza virus and cancer cells is consistent with the legendary high and rapid replicating ability of malaria organisms. The multitude of overlapping Replikins in malaria also provides an opportunity for the organism to flood and confuse the immune system of its host and thereby maximize the chance that the wrong antibody will be made and perpetuated, leaving key malaria antigens unharmed.

[0179] As in the case of influenza virus, for example, peptide vaccines based on the Replikin structure(s) found in the malaria organism can provide an effective means of preventing and/or treating malaria. Vaccination against malaria can be achieved by administering a composition containing one or a mixture of Replikin structures observed in Plasmodium falciparum. Furthermore, antibodies to malaria Replikins can be generated and administered for passive immunity or malaria detection

[0180] Table 8 provides a list of several *Plasmodium falciparum* Replikin sequences. It should be noted that this list is not meant to be complete. Different isolates of the organism may contain other Replikin structures.

Table 8

Malaria Replikins

a) Primary structure of a Plasmodium falciparum malaria antigen located at the merozoite surface and within the parasitophorous vacuole

a) i) DECOYS:

(C-Terminal)

keeekekekekekekekekekekekekekekekekekeeeek (SEQ ID NO. 293), or
keeekekekekekekekekekekekekekekekekekeeeek (SEQ ID NO. 294), or
keeekekekekekekekekekekekekekekekekekeeeek (SEQ ID NO. 295), or
keeekekek (SEQ ID NO. 296)

ii) ReplikinS:

Hkklikalkkniesiqnkk (SEQ ID NO. 297)
hkklikalkkniesiqnkm (SEQ ID NO. 298)
hkklikalkk (SEQ ID NO. 299)
hkklikalk (SEQ ID NO. 300)
katysfvntkkkiislsqghkk (SEQ ID NO. 301)
katysfvntkkkiislsqghk (SEQ ID NO. 302)
katysfvntkkkiislsqgh (SEQ ID NO. 303)
htyvkqkapsdpqca dikeekellkek (SEQ ID NO. 304)
kiislsqghk (SEQ ID NO. 305)
kkkkfepkngnvsetikli (SEQ ID NO. 306)
kkkfepkngnvsetikli (SEQ ID NO. 307)
kkfepkngnvsetikli (SEQ ID NO. 308)
kngnvsetikli (SEQ ID NO. 309)
klichgkdkk (SEQ ID NO. 310)
kvkkgvtlkkfepkngnvsetiklichgkdkkh (SEQ ID NO. 311)
hliynksynpillsckvkkmmmlkenvdyiqnqlfkelmnqkatysfvntkkkiislk (SEQ ID NO. 312)
hliynksynpillsckvkkmmmlkenvdyiqnqlfkelmnqkatysfvntk (SEQ ID NO. 313)
hliynksynpillsckvkkmmmlkenvdyiqnqlfkelmnq (SEQ ID NO. 314)
hliynksynpillsckvkkmmmlkenvdyiqnqlf (SEQ ID NO. 315)
hliynksynpillsckvkkmmmlk (SEQ ID NO. 316)
ksannsanngkknnaeemknlvnflqshkklikalkkniesiqnkkh (SEQ ID NO. 317)
kknnaeemknlvnflqshkklikalkkniesiqnkkh (SEQ ID NO. 318)
knlvnflqshkklikalkkniesiqnkkh (SEQ ID NO. 319)
kklikalkkniesiqnkkh (SEQ ID NO. 320)
klikalkkniesiqnkkh (SEQ ID NO. 321)
kniesiqnkkh (SEQ ID NO. 322)
kniesiqnkkh (SEQ ID NO. 323)
knnaeemknlvnflqsh (SEQ ID NO. 324)
kklikalkkniesiqnkkqghkk (SEQ ID NO. 325)
kknnaeemknlvnflqshk (SEQ ID NO. 326)
knnaeemknlvnflqsh (SEQ ID NO. 327)
klikalkkniesiqnkkqghkk (SEQ ID NO. 328)
kvkkgvtlkkfepkngnvsetikli (SEQ ID NO. 329)
kngnvsetikli (SEQ ID NO. 330)
klichgkdkk (SEQ ID NO. 331)

ksannsanngkknnaeemknlvnflqsh (SEQ ID NO. 332)
 kknnaeemknlvnflqsh (SEQ ID NO. 333)
 kklikalkkniesiqnkkh (SEQ ID NO. 334)
 kalkkniesiqnkkh (SEQ ID NO. 335)
 kkniesiqnkkh (SEQ ID NO. 336)
 kelmnqkatysfvntkkkiislsqgh (SEQ ID NO. 337)
 ksqghkk (SEQ ID NO. 338)
 kkkiiislsqgh (SEQ ID NO. 339)
 kkiislsqgh (SEQ ID NO. 340)
 kkniesiqnkkh (SEQ ID NO. 341)
 kniesiqnkkh (SEQ ID NO. 342)
 htyvkgkkapsdpqcadikeeeckellk (SEQ ID NO. 343)
 htyvkgkkapsdpqcadikeeeckellk (SEQ ID NO. 344)

b)"liver stage antigen-3" gene="LSA-3" Replikins
 henvlsaalentqseeekkevidvieevk (SEQ ID NO. 345)
 kenvvttilekveettaesvttfsnileeigentitndtieekleelh (SEQ ID NO. 346)
 hylqqmkekfskek (SEQ ID NO. 347)
 hylqqmkekfskeknnnvievtlnkaekkgnvqvtnktekttk (SEQ ID NO. 348)
 hylqqmkekfskeknnnvievtlnkaekkgnvqvtnktekttkvdknnk (SEQ ID NO. 349)
 hylqqmkekfskeknnnvievtlnkaekkgnvqvtnktekttkvdknnkvpkkrqtqk (SEQ ID NO. 350)
 hylqqmkekfskeknnnvievtlnkaekkgnvqvtnktekttkvdknnkvpkkrqtqsk (SEQ ID NO. 351)
 hvdevmkyvqkidkevdkvskaleskndvtnlvkqnqdfvskvknfvkkyk (SEQ ID NO. 352)
 hvdevmkyvqkidkevdkvskaleskndvtnlvkqnqdfvskvknfvk (SEQ ID NO. 353)
 hvdevmkyvqkidkevdkvskaleskndvtnlvkqnqdfvsk (SEQ ID NO. 354)
 hvdevmkyvqkidkevdkvskaleskndvtnlvk (SEQ ID NO. 355)
 hvdevmkyvqkidkevdkvskalesk (SEQ ID NO. 356)
 hvdevmkyvqkidkevdkvsk (SEQ ID NO. 357)
 hvdevmkyvqkidkevdk (SEQ ID NO. 358)
 hvdevmkyvqkidk (SEQ ID NO. 359)
 kdevidlivqkekriekvkakkkklekkveegvsglkkh (SEQ ID NO. 360)
 kvkakkkklekkveegvsglkkh (SEQ ID NO. 361)
 kakkkkklekkveegvsglkkh (SEQ ID NO. 362)
 kkkklekkveegvsglkkh (SEQ ID NO. 363)
 kkkklekkveegvsglkkh (SEQ ID NO. 364)
 kklekkveegvsglkkh (SEQ ID NO. 365)
 klekkveegvsglkkh (SEQ ID NO. 366)
 kkveegvsglkkh (SEQ ID NO. 367)
 kveegvsglkkh (SEQ ID NO. 368)
 hveqnvyvdvdpamkdqflgilneagglkemffnledvfksesdvitveeikdepvqk (SEQ ID NO. 369)
 hikgleeddleevddllkgsildmlkgdmelgdmkesledvttklgerveslk (SEQ ID NO. 370)
 hikgleeddleevddllkgsildmlkgdmelgdmkesledvttk (SEQ ID NO. 371)
 hikgleeddleevddllkgsildmlkgdmelgdmk (SEQ ID NO. 372)
 hikgleeddleevddllkgsildmlk (SEQ ID NO. 373)
 hiisgdadvlssalgmdeeqmtrkkaqrpk (SEQ ID NO. 374)
 hdittldevvelkdveedkiek (SEQ ID NO. 375)
 kkleevhelk (SEQ ID NO. 376)
 kleevhelk (SEQ ID NO. 377)
 ktietdileekkkeiekdh (SEQ ID NO. 378)
 kkeiekdhfek (SEQ ID NO. 379)

kdhfek (SEQ ID NO. 380)
kfeeeaeikh (SEQ ID NO. 381)

c) 28 KDA ookinete surface antigen precursor Replikins:

kdgdtkctlecaqgkckcihksdnhhksdnhhksdpnhkkknnnnk (SEQ ID NO. 382)
kdgdtkctlecaqgkckcihksdnhhksdnhhksdpnhkk (SEQ ID NO. 383)
kdgdtkctlecaqgkckcihksdnhhksdnhhksdpnhk (SEQ ID NO. 384)
kdgdtkctlecaqgkckcihksdnhhksdnhhk (SEQ ID NO. 385)
kdgdtkctlecaqgkckcihksdnhhk (SEQ ID NO. 386)
kdgdtkctlecaqgkckcihk (SEQ ID NO. 387)
kdgdtkctlecaqgk (SEQ ID NO. 388)
kdgdtkctlecaqgk (SEQ ID NO. 389)
kciqaecnykecgeqkcvwdgih (SEQ ID NO. 390)
kecgeqkcvwdgih (SEQ ID NO. 391)
hieckcnndyvltnryecepknktsledtnk (SEQ ID NO. 392)

d) Blood stage trophozoites and schizonts Replikins:

ksdhnhsdsdhnhksdhnhsksdhnhsksdpnhkkknmmnnk (SEQ ID NO. 393)
ksdhnhsdsdhnhksdhnhsksdpnhkkknmmnnk (SEQ ID NO. 394)
ksdhnhsdsdhnhksdpnhkkknmmnnk (SEQ ID NO. 395)
ksdhnhsksdpnhkkknmmnnk (SEQ ID NO. 396)
kkknmmnnkdnksdpnhk (SEQ ID NO. 397)
kkmmnnkdnksdpnhk (SEQ ID NO. 398)
knmmnnkdnksdpnhk (SEQ ID NO. 399)
kdnksdpnhk (SEQ ID NO. 400)
ksdpnhk (SEQ ID NO. 401)
hslyalqqneeyqkvknekdkneikkikqlieknk (SEQ ID NO. 402)
hslyalqqneeyqkvknekdkneikkik (SEQ ID NO. 403)
hslyalqqneeyqkvknekdkneikk (SEQ ID NO. 404)
hslyalqqneeyqkvknekdkneik (SEQ ID NO. 405)
hklenleemdk (SEQ ID NO. 406)
khfddntneqk (SEQ ID NO. 407)
kkeddek (SEQ ID NO. 408)
keennkkeddek (SEQ ID NO. 409)
ktssgilnkeennkkeddek (SEQ ID NO. 410)
knihkk (SEQ ID NO. 411)
hikkkegidigyk (SEQ ID NO. 412)
kkmwtklwdnkgneitknih (SEQ ID NO. 413)
kkgiqwnllkkmwtklwdnkgneitknih (SEQ ID NO. 414)
kekksdenrkkkqkedkknpnklkkieytnkithffkaknnkqqnnvth (SEQ ID NO. 415)
kkksdenrkkkqkedkknpnklkkieytnkithffkaknnkqqnnvth (SEQ ID NO. 416)
kksdenrkkkqkedkknpnklkkieytnkithffkaknnkqqnnvth (SEQ ID NO. 417)
kkqkedkknpnklkkieytnkithffkaknnkqqnnvth (SEQ ID NO. 418)
kqkedkknpnklkkieytnkithffkaknnkqqnnvth (SEQ ID NO. 419)
kedkknpnklkkieytnkithffkaknnkqqnnvth (SEQ ID NO. 420)
knpnklkkieytnkithffkaknnkqqnnvth (SEQ ID NO. 421)
kkieytnkithffkaknnkqqnnvth (SEQ ID NO. 422)
kieytnkithffkaknnkqqnnvth (SEQ ID NO. 423)
kithffkaknnkqqnnvth (SEQ ID NO. 424)
hknnedikndnsdikndnsdikndnsdikndnsdikndnsdikndnsdik (SEQ ID NO. 425)

hknmedikndnskdikndnskdikndnskdikndnnedikndnsk (SEQ ID NO. 426)
hknmedikndnskdikndnskdikndnskdikndnnedik (SEQ ID NO. 427)
hknmedikndnskdikndnskdikndnskdik (SEQ ID NO. 428)
hknmedikndnskdikndnskdikndnsk (SEQ ID NO. 429)
hknmedikndnskdikndnskdik (SEQ ID NO. 430)
hknmedikndnskdikndnsk (SEQ ID NO. 431)
hknmedikndnskdik (SEQ ID NO. 432)
hknmedik (SEQ ID NO. 433)
kkyddlqnkynilnlklsleekneelkkyh (SEQ ID NO. 434)
kyddlqnkynilnlklsleekneelkkyh (SEQ ID NO. 435)
kynilnlklsleekneelkkyh (SEQ ID NO. 436)
klksleekneelkkyh (SEQ ID NO. 437)
knsleekneelkkyh (SEQ ID NO. 438)
kneelkkyh (SEQ ID NO. 439)
hmggnqdinenvynikpqefkeeeedismvntkk (SEQ ID NO. 440)
knsnelkrindnffklh (SEQ ID NO. 441)
kpolykkckisqlykkckisqvwwcmpvkdftfyernnvlnskienniekiph (SEQ ID NO. 442)
hinneytnknpkncillykneernyndnnikdyinsmnfk (SEQ ID NO. 443)
hinneytnknpkncillykneernyndnnikdyinsmnfk (SEQ ID NO. 444)
hinneytnknpkncillyk (SEQ ID NO. 445)
knktnqskgvkgeyekkketngh (SEQ ID NO. 446)
ktnqskgvkgeyekkketngh (SEQ ID NO. 447)
kgvkgeyekkketngh (SEQ ID NO. 448)
kgeyekkketngh (SEQ ID NO. 449)
ksgmytnegnkscecsykkkssssnkvh (SEQ ID NO. 450)
kscecsykkkssssnkvh (SEQ ID NO. 451)
kkkssssnkvh (SEQ ID NO. 452)
kkssssnkvh (SEQ ID NO. 453)
kssssnkvh (SEQ ID NO. 454)
himlksmytnegnkscecsykkkssssnk (SEQ ID NO. 455)
himlksmytnegnkscecsykkk (SEQ ID NO. 456)
himlksmytnegnkscecsykk (SEQ ID NO. 457)
himlksmytnegnkscecsyk (SEQ ID NO. 458)
kplaklrrektqinktkyergdviidnteiqkiiirdyhetlnvhkldh (SEQ ID NO. 459)
krektqinktkyergdviidnteiqkiiirdyhetlnvhkldh (SEQ ID NO. 460)
ktqinktkyergdviidnteiqkiiirdyhetlnvhkldh (SEQ ID NO. 461)
kplaklrrektqinktkyergdviidnteiqkiiirdyhetlnvh (SEQ ID NO. 462)
kplaklrrektqinktkyergdviidnteiqkiiirdyh (SEQ ID NO. 463)
klrrektqinktkyergdviidnteiqkiiirdyh (SEQ ID NO. 464)
krektqinktkyergdviidnteiqkiiirdyh (SEQ ID NO. 465)
ktqinktkyergdviidnteiqkiiirdyh (SEQ ID NO. 466)
kdkkekksdsnenrkkkqkedkknnpndnklkkieytnkith (SEQ ID NO. 467)
kdkekksdsnenrkkkqkedkknnpndnklkkieytnkith (SEQ ID NO. 468)
kekksdsnenrkkkqkedkknnpndnklkkieytnkith (SEQ ID NO. 469)
kkksdsnenrkkkqkedkknnpndnklkkieytnkith (SEQ ID NO. 470)
kkdsnenrkkkqkedkknnpndnklkkieytnkith (SEQ ID NO. 471)
kdsnenrkkkqkedkknnpndnklkkieytnkith (SEQ ID NO. 472)
kkkqkedkknnpndnklkkieytnkith (SEQ ID NO. 473)
kkqkedkknnpndnklkkieytnkith (SEQ ID NO. 474)
kqkedkknnpndnklkkieytnkith (SEQ ID NO. 475)

kedkknpnndnklkkieytnkith (SEQ ID NO. 476)
 knpnndnklkkieytnkith (SEQ ID NO. 477)
 knpnndnklkkieytnkith (SEQ ID NO. 478)
 klkkieytnkith (SEQ ID NO. 479)
 kkieytnkith (SEQ ID NO. 480)
 kieytnkith (SEQ ID NO. 481)
 hgqikiedvnnenfneeqmknkyndeekmdiskskslsdflek (SEQ ID NO. 482)
 hgqikiedvnnenfneeqmknkyndeekmdiskskslk (SEQ ID NO. 483)
 hgqikiedvnnenfneeqmknkyndeekmdisksk (SEQ ID NO. 484)
 hgqikiedvnnenfneeqmknkyndeekmdisk (SEQ ID NO. 485)
 kkyddlqnkynilnklksleekneelkkyh (SEQ ID NO. 486)
 kyddlqnkynilnklksleekneelkkyh (SEQ ID NO. 487)
 kynilnklksleekneelkkyh (SEQ ID NO. 488)
 klksleekneelkkyh (SEQ ID NO. 489)
 knsleekneelkkyh (SEQ ID NO. 490)
 kneelkkyh (SEQ ID NO. 491)
 hmggnqdinenynikpqefkeeeedismvntkkoddiqenik (SEQ ID NO. 492)
 ktnlyniynnknnddkdnildnenreglylcvmknsnelkrindnffklh (SEQ ID NO. 493)
 knsnelkrindnffklh (SEQ ID NO. 494)
 krindnffklh (SEQ ID NO. 495)
 hinneytnknpkncellykneernyndnnikdyinsmnfkk (SEQ ID NO. 496)
 hinneytnknpkncellykneernyndnnikdyinsmnfk (SEQ ID NO. 497)
 hinneytnknpkncellyk (SEQ ID NO. 498)
 kpclykkckisqvwvwcmpvkdftfityernvlnskienniekiph (SEQ ID NO. 499)
 kckisqvwvwcmpvkdftfityernvlnskienniekiph (SEQ ID NO. 500)
 kienniekiph (SEQ ID NO. 501)
 knktngskgvkgeyekkketngh (SEQ ID NO. 502)
 ktngskgvkgeyekkketngh (SEQ ID NO. 503)
 kgvkgeyekkketngh (SEQ ID NO. 504)
 kgeyekkketngh (SEQ ID NO. 505)
 ktiekinkskswffeeldeidkplaklrrektqinktkyergdviidnteiqkiirdyh (SEQ ID NO. 506)
 kinkskswffeeldeidkplaklrrektqinktkyergdviidnteiqkiirdyh (SEQ ID NO. 507)
 kplaklrrektqinktkyergdviidnteiqkiirdyh (SEQ ID NO. 508)
 himlksqmytnegnkscecsykkkssssnkvh (SEQ ID NO. 509)
 klrrektqinktkyergdviidnteiqkiirdyh (SEQ ID NO. 510)
 krektqinktkyergdviidnteiqkiirdyh (SEQ ID NO. 511)
 ktqinktkyergdviidnteiqkiirdyh (SEQ ID NO. 512)
 kplaklrrektqinktkyergdviidnteiqkiirdyhtlnvhkldh (SEQ ID NO. 513)
 klrrektqinktkyergdviidnteiqkiirdyhtlnvhkldh (SEQ ID NO. 514)
 krektqinktkyergdviidnteiqkiirdyhtlnvhkldh (SEQ ID NO. 515)
 ktqinktkyergdviidnteiqkiirdyhtlnvhkldh (SEQ ID NO. 516)
 kplaklrrektqinktkyergdviidnteiqkiirdyhtlnvh (SEQ ID NO. 517)
 klrrektqinktkyergdviidnteiqkiirdyhtlnvh (SEQ ID NO. 518)
 krektqinktkyergdviidnteiqkiirdyhtlnvh (SEQ ID NO. 519)
 ktqinktkyergdviidnteiqkiirdyhtlnvh (SEQ ID NO. 520)
 himlksqmytnegnkscecsykkkssssnkvh (SEQ ID NO. 521)
 ksqmytnegnkscecsykkkssssnkvh (SEQ ID NO. 522)
 kscecsykkkssssnkvh (SEQ ID NO. 523)
 kkkssssnkvh (SEQ ID NO. 524)
 kkssssnkvh (SEQ ID NO. 525)

kssssnkvh (SEQ ID NO. 526)
 himlksqmytnegnkscecsykkssssnk (SEQ ID NO. 527)
 himlksqmytnegnkscecsykk (SEQ ID NO. 528)
 himlksqmytnegnkscecsykk (SEQ ID NO. 529)
 himlksqmytnegnkscecsyk (SEQ ID NO. 530)
 hnnhniqiydkrinfnnphkvmyhdnmsknertek (SEQ ID NO. 531)
 hnnhniqiydkrinfnnphkvmyhdnmsk (SEQ ID NO. 532)
 hnnhniqiydkrinfnnphk (SEQ ID NO. 533)
 hkvmyhdnmsknertek (SEQ ID NO. 534)
 hkvmyhdnmsk (SEQ ID NO. 535)

REPLIKINS IN STRUCTURAL PROTEINS

[0181] It has also been determined that some structural proteins include Replikin structures. Structural proteins are molecules involved in tissue and organ support, such as collagen in skin and connective tissue and in membrane structures, for example amyloid A4 precursor protein (APP) in brain. Overproduction of these proteins is associated with disease; specifically, scleroderma in the case of overproduction of collagen in skin (Table 9) and Alzheimer's Disease in the case of overproduction of APP in the brain (Table 10).

[0182] The association of scleroderma and malignancy has been a source of controversy during recent years. Several mechanisms of interrelationship have been suggested in earlier reports. Recent long-term studies suggest an increased association-ratio of scleroderma and malignancy. However, the underlying mechanisms remain elusive. (Wenzel, J. Eur. J. Dermatol. 20002 May-Jun; 12(3): 296-300).

[0183] Several proteins concerned with the excessive production of proteins in scleroderma have been found to contain Replikin structures. Thus, these provide further examples of unrecognized targets for inhibition or cessation of excessive

collagen production. Table 9 provides a list of proteins in scleroderma and the associated Replikins.

[0184] The APP protein is the source of the amyloid beta A4 protein, which in
5 excessive amounts forms plaques in the extracellular spaces in the brain,
producing toxic effects associated with nerve cell loss in Alzheimer's Disease.
Most studies to date have focused on the inability to clear the excessive deposits of
A4, but have not considered that, rather than a waste clearance problem, this may
actually be a problem of overproduction of the precursor protein APP. The high
10 concentration of the Replikins in APP (3.3 Replikins per 100 amino acids)
strongly suggest that overproduction may well be the cause of Alzheimer's
Disease (Table 10). Therefore, the Replikins contained in Table 10 can be blocked
or inhibited by the same methods as illustrated in detail for the glioma Replikin.

Table 9

Proteins overproduced in scleroderma and associated Replikins:

PMCI HUMAN:

hreictiqssggimllkdqvlrcskiagvkvaetelilk (SEQ ID NO.536)

hreictiqssggimllkdqviresk (SEQ ID NO.537)

34KD nucleolar scleroderma antigen:

hreictiqssggimllkdqvlrcskiagvkvaetelikkalendqk (SEQ ID NO.538)

hreictiqssggimllkdqvlrcskiagvkvaetelilk (SEQ ID NO.539)

Fibrillarin:

kkmqenmkpqqtlepyerdh (SEQ ID NO.540)

kmqenmkpqqtlepyerdh (SEQ ID NO. 541)

SPOP HUMAN:

hemeeskknrveindvepevfkemmcfiytkapnldk (SEQ ID NO.542)

hemeeskknrveindvepevfkemmcfiytk (SEQ ID NO.543)

Centromere protein C:

khgelkvyk (SEQ ID NO.544)

klilgpqeekgkqh (SEQ ID NO. 545)

hnrihhk (SEQ ID NO. 546)

hnnssikstkktnqssk (SEQ ID NO. 547)

hnnsrkstkktnqssk (SEQ ID NO. 548)

khnilpktlandkhshkph (SEQ ID NO.549)

hnilpktlandkhshk (SEQ ID NO. 550)

hnilpktlandkhshk (SEQ ID NO. 551)

hnilpktlandk (SEQ ID NO. 552)

kntpdskkissmindhh (SEQ ID NO.553)

kntpdskkissmindh (SEQ ID NO. 554)

kdtciqspskceqkshpksvpvsskkk (SEQ ID NO. 555)

kdtciqspskceqkshpksvpvsskk (SEQ ID NO. 556)

hpksvpvsskkk (SEQ ID NO. 557)

hpksvpvsskk (SEQ ID NO. 558)

hpksvpvssk (SEQ ID NO. 559)

Factor CTCBF, KU antigen:

kalqekveikqlnh (SEQ ID NO. 560)

ktlfplicakkdqvtageifgdnhedgptakklkteggah (SEQ ID NO. 561)

ktlfplicakkdqvtageifqdnb (SEQ ID NO. 562)

klcvfkierhsih (SEQ ID NO. 563)

klcvfkierh (SEQ ID NO. 564)

kgpsfplkgiteqqkegleivk (SEQ ID NO. 565)

hgpsfplkgiteqqk (SEQ ID NO. 566)

ATP synthase subunit 6:

htllkilstflfk (SEQ ID NO. 567)

hllgnndknllpsk (SEQ ID NO. 568)

FBRL nuclear protein:

hrhegvficrgkedalvtk (SEQ ID NO. 569)

hegvficrgkedalvtk (SEQ ID NO. 570)

hsggnrgrgrggkrghqsgk (SEQ ID NO. 571)

krqnqsgknvmveph (SEQ ID NO. 572)

krqnqsgknvmvephrh (SEQ ID NO. 573)

kkmqgenmkpgeqltlepyerdh (SEQ ID NO.574)
 kmqgenmkpgeqltlepyerdh (SEQ ID NO. 575)
HP1Hs-alpha protein:
 haypedaenkeketak (SEQ ID NO. 576)
 keanvkcpqiviafyeerltwh (SEQ ID NO. 577)
 kvldrrvvkqgveyllkwkgfseeh (SEQ ID NO. 578)
 kgqveyllkwkgfseeh (SEQ ID NO. 579)
FM/Scl nucleolar protein:
 ksevaagvkksqplsaerlenvlfghdesh (SEQ ID NO.580)
 ksevaagvkksqplsaerlenvlfgh (SEQ ID NO. 581)
 kaaeygkkaaksetfrillhakniirpqlk (SEQ ID NO. 582)
 kaaeygkkaaksetfrillhak (SEQ ID NO. 583)
 ksetfrillhak (SEQ ID NO. 584)
 hakniirpqlk (SEQ ID NO. 585)
 hmnlkiaelpk (SEQ ID NO. 586)
 hslldhllklycnvdsnk (SEQ ID NO. 587)
 hllklycnvdsnk (SEQ ID NO. 588)

Table 10

Amyloid beta A4 precursor protein (APP) Replikins:

kakerleakh (SEQ ID NO. 589)
 kdrqhtlk (SEQ ID NO. 590)
 kdrqhtlkh (SEQ ID NO. 591)
 ketcsekstnlh (SEQ ID NO. 592)
 kteisevkmadaefgh (SEQ ID NO. 593)
 kteisevkmadaefghdsgfevrh (SEQ ID NO. 594)
 kkyvraeqkdrqhtlkh (SEQ ID NO. 595)
 kyvraeqkdrqhtlkh (SEQ ID NO. 596)
 kkyvraeqkdrqh (SEQ ID NO. 597)
 kyvraeqkdrqht (SEQ ID NO. 598)
 hhvfmmlkkyvraeqk (SEQ ID NO. 599)
 hvfmmlkkyvraeqk (SEQ ID NO. 600)
 hhvfmmlkkyvraeqkdrqhtlkh (SEQ ID NO. 601)
 hvfmmlkkyvraeqkdrqhtlkh (SEQ ID NO. 602)
 hahfqkakerleakh (SEQ ID NO. 603)
 hahfqkakerleak (SEQ ID NO. 604)
 hfqkakerleak (SEQ ID NO. 605)
 hqermdivcethllhwhtvaketcsekstnlh (SEQ ID NO. 606)
 hqermdivcethllhwhtvaketcsek (SEQ ID NO. 607)
 hwhtvaketcsek (SEQ ID NO. 608)
 htvaketcsek (SEQ ID NO. 609)
 hllhwhtvaketcsek (SEQ ID NO. 610)
 hmnvqngkwesdpsgtkctgk (SEQ ID NO. 611)
 hmnvqngkwesdpsgtk (SEQ ID NO. 612)

PASSIVE IMMUNITY

[0185] In another embodiment of the invention, isolated Replikin peptides may be used to generate antibodies, which may be used, for example to provide passive immunity in an individual. Passive immunity to the strain of influenza identified by the method of the invention to be the most likely cause of future influenza infections may be obtained by administering antibodies to Replikin sequences of the identified strain of influenza virus to patients in need. Similarly, passive immunity to malaria may be obtained by administering antibodies to Plasmodium falciparum Replikin(s).

[0186] Various procedures known in the art may be used for the production of antibodies to Replikin sequences. Such antibodies include but are not limited to polyclonal, monoclonal, chimeric, humanized, single chain, Fab fragments and fragments produced by an Fab expression library. Antibodies that are linked to a cytotoxic agent may also be generated. Antibodies may also be administered in combination with an antiviral agent. Furthermore, combinations of antibodies to different Replikins may be administered as an antibody cocktail.

[0187] For the production of antibodies, various host animals or plants may be immunized by injection with a Replikin peptide or a combination of Replikin peptides, including but not limited to rabbits, mice, rats, and larger mammals.

[0188] Monoclonal antibodies to Replikins may be prepared by using any technique that provides for the production of antibody molecules. These include but are not limited to the hybridoma technique originally described by Kohler and

Milstein, (Nature, 1975, 256:495-497), the human B-cell hybridoma technique (Kosbor et al., 1983, Immunology Today, 4:72), and the EBV hybridoma technique (Cole et al., Monoclonal Antibodies and Cancer Therapy, Alan R. Liss, Inc., pp. 77-96). In addition, techniques developed for the production of chimeric antibodies (Morrison et al., 1984, Proc. Nat. Acad. Sci USA, 81:6851-6855) or other techniques may be used. Alternatively, techniques described for the production of single chain antibodies (US 4,946,778) can be adapted to produce Replikin-specific single chain antibodies.

10 [0189] Particularly useful antibodies of the invention are those that specifically bind to Replikin sequences contained in peptides and/or polypeptides of influenza virus. For example, antibodies to any of peptides observed to be present in an emerging or re-emerging strain of influenza virus and combinations of such antibodies are useful in the treatment and/or prevention of influenza. Similarly, 15 antibodies to any Replikins present on malaria antigens and combinations of such antibodies are useful in the prevention and treatment of malaria.

[0190] Antibody fragments which contain binding sites for a Replikin may be generated by known techniques. For example, such fragments include but are not 20 limited to F(ab')₂ fragments which can be produced by pepsin digestion of the antibody molecules and the Fab fragments that can be generated by reducing the disulfide bridges of the F(ab')₂ fragments. Alternatively, Fab expression libraries can be generated (Huse et al., 1989, Science, 246:1275-1281) to allow rapid and easy identification of monoclonal Fab fragments with the desired specificity.

25

[0191] The fact that antimalignin antibody is increased in concentration in human malignancy regardless of cancer cell type (Figure 5), and that this antibody binds to malignant cells regardless of cell type now may be explained by the presence of the Replikin structures herein found to be present in most malignancies (Figure 1 and Table 2). Population studies have shown that antimalignin antibody increases in concentration in healthy adults with age, and more so in high-risk families, as the frequency of cancer increases. An additional two-fold or greater antibody increase which occurs in early malignancy has been independently confirmed with a sensitivity of 97% in breast cancers 1-10 mm in size. Shown to localize preferentially in malignant cells in vivo, histochemically the antibody does not bind to normal cells but selectively binds to (Figure 4a,b) and is highly cytotoxic to transformed cells in vitro (Figure 4c-f). Since in these examples the same antibody is bound by several cell types, that is, brain glioma, hematopoietic cells (leukemia), and small cell carcinoma of lung, malignant Replikin class unity is again demonstrated.

[0192] Antimalignin does not increase with benign proliferation, but specifically increases only with malignant transformation and replication in breast in vivo and returns from elevated to normal values upon elimination of malignant cells (Figure 5). Antimalignin antibody concentration has been shown to relate quantitatively to the survival of cancer patients, that is, the more antibody, the longer the survival. Taken together, these results suggest that anti-Replikin antibodies may be a part of a mechanism of control of cell transformation and replication. Augmentation of this immune response may be useful in the control of replication, either actively with synthetic Replikins as vaccines, or passively by the administration of anti-

Replikin antibodies, or by the introduction of non-immune based organic agents, such as for example, carbohydrates, lipids and the like, which are similarly designed to target the Replikin specifically.

5 [0193] In another embodiment of the invention, immune serum containing antibodies to one or more Replikins obtained from an individual exposed to one or more Replikins may be used to induce passive immunity in another individual or animal. Immune serum may be administered via i.v. to a subject in need of treatment. Passive immunity also can be achieved by injecting a recipient with
10 preformed antibodies to one or more Replikins. Passive immunization may be used to provide immediate protection to individuals who have been exposed to an infectious organism. Administration of immune serum or preformed antibodies is routine and the skilled practitioner can readily ascertain the amount of serum or antibodies needed to achieve the desired effect.

15

SYNTHETIC REPLIKIN VACCINE (ACTIVE IMMUNITY)

[0194] Synthetic Replikin vaccines, based on Replikins such as the glioma Replikin (SEQ ID NO.: 1) "kagvafihkk" or the hepatitis C Replikin (SEQ ID NO.:
18) "hyppkpgcivpak", or HIV Replikins such as (SEQ ID NO.: 5) "kcfncgkegh"
20 or (SEQ ID NO.: 6) "kvylawvpahk" or preferably, an influenza vaccine based on conserved and/or emerging or re-emerging Replikin(s) over a given time period may be used to augment antibody concentration in order to lyse the respective virus infected cells and release virus extracellularly where chemical treatment can then be effective. Similarly, a malaria vaccine, based on Replikins observed in
25 Plasmodium falciparum malaria antigens on the merozoite surface or within the

parasitophorous vacuole, for example, can be used to generate cytotoxic antibodies to malaria.

[0195] Recognin and/or Replikin peptides may be administered to a subject to induce the immune system of the subject to produce anti-Replikin antibodies.

Generally, a 0.5 to about 2 mg dosage, preferably a 1 mg dosage of each peptide is administered to the subject to induce an immune response. Subsequent dosages may be administered if desired.

[0196] The Replikin sequence structure is associated with the function of replication. Thus, whether the Replikins of this invention are used for targeting sequences that contain Replikins for the purpose of diagnostic identification, promoting replication, or inhibiting or attacking replication, for example, the structure-function relationship of the Replikin is fundamental.

[0197] It is preferable to utilize only the specific Replikin structure when seeking to induce antibodies that will recognize and attach to the Replikin fragment and thereby cause destruction of the cell. Even though the larger protein sequence may be known in the art as having a "replication associated function," vaccines using the larger protein often have failed or proven ineffective.

[0198] Although the present inventors do not wish to be held to a single theory, the studies herein suggest that the prior art vaccines are ineffective because they are based on the use of the larger protein sequence. The larger protein sequence invariably has one or more epitopes (independent antigenic sequences that can

induce specific antibody formation); Replikin structures usually comprise one of these potential epitopes. The presence of other epitopes within the larger protein may interfere with adequate formation of antibodies to the Replikin, by "flooding" the immune system with irrelevant antigenic stimuli that may preempt the Replikin antigens, See, e.g., Webster, R.G., *J. Immunol.*, 97(2):177-183 (1966); and Webster et al., *J. Infect. Dis.*, 134:48-58, 1976; Klenerman et al, *Nature* 394:421-422 (1998) for a discussion of this well-known phenomenon of antigenic primacy whereby the first peptide epitope presented and recognized by the immune system subsequently prevails and antibodies are made to it even though other peptide epitopes are presented at the same time. This is another reason that, in a vaccine formulation, it is important to present the constant Replikin peptide to the immune system first, before presenting other epitopes from the organism so that the Replikin is not pre-empted but lodged in immunological memory.

[0199] The formation of an antibody to a non-Replikin epitope may allow binding to the cell, but not necessarily lead to cell destruction. The presence of structural "decoys" on the C-termini of malaria proteins is another aspect of this ability of other epitopes to interfere with binding of effective anti-Replikin antibodies, since the decoy epitopes have many lysine residues, but no histidine residues. Thus, decoy epitopes may bind anti-Replikin antibodies, but may keep the antibodies away from histidine-bound respiratory enzymes. Treatment may therefore be most efficacious in two stages: 1) proteases to hydrolyze decoys, then; 2) anti-Replikin antibodies or other anti-Replikin agents.

[0200] It is well known in the art that in the course of antibody production against a "foreign" protein, the protein is first hydrolyzed into smaller fragments. Usually fragments containing from about six to ten amino acids are selected for antibody formation. Thus, if hydrolysis of a protein does not result in Replikin-containing
5 fragments, anti-Replikin antibodies will not be produced. In this regard, it is interesting that Replikins contain lysine residues located six to ten amino acids apart, since lysine residues are known to bind to membranes.

[0201] Furthermore, Replikin sequences contain at least one histidine residue.
10 Histidine is frequently involved in binding to redox centers. Thus, an antibody that specifically recognizes a Replikin sequence has a better chance of inactivating or destroying the cell in which the Replikin is located, as seen with anti-malignin antibody, which is perhaps the most cytotoxic anti-cancer antibody yet described, being active at picograms per cell.

15

[0202] One of the reasons that vaccines directed towards a particular protein antigen of a disease causing agent have not been fully effective in providing protection against the disease (such as foot and mouth vaccine which has been developed against the VP1 protein or large segments of the VP1 protein) is that the
20 best antibodies have not been produced, that is – it is likely that the antibodies to the Replikins have not been produced. Replikins have not been produced. That is, either epitopes other than Replikins present in the larger protein fragments may interfere according to the phenomenon of antigenic primacy referred to above, and/or because the hydrolysis of larger protein sequences into smaller sequences
25 for processing to produce antibodies results in loss of integrity of any Replikin

structure that is present, e.g., the Replikin is cut in two and/or the histidine residue is lost in the hydrolytic processing. The present studies suggest that for an effective vaccine to be produced, the Replikin sequences, and no other epitope, should be used as the vaccine. For example, a vaccine of the invention can be
5 generated using any one of the Replikin peptides identified by the three point recognition system.

[0203] Particularly preferred peptides - for example - an influenza vaccine include peptides that have been demonstrated to be conserved over a period of one or more
10 years, preferably about three years or more, and/or which are present in a strain of influenza virus shown to have the highest increase in concentration of Replikins relative to Replikin concentration in other influenza virus strains, e.g., an emerging strain. The increase in Replikin concentration preferably occurs over a period of at least about six months to one year, preferably at least about two years or more, and
15 most preferably about three years or more. Among the preferred Replikin peptides for use in an influenza virus vaccine are those Replikins observed to "re- emerge" after an absence from the hemagglutinin amino acid sequence for one or more years.

20 [0204] The Replikin peptides of the invention, alone or in various combinations are administered to a subject, preferably by i.v. or intramuscular injection, in order to stimulate the immune system of the subject to produce antibodies to the peptide. Generally the dosage of peptides is in the range of from about 0.1 µg to about 10 mg, preferably about 10 µg to about 1 mg, and most preferably about 50 µg to

about 500 ug. The skilled practitioner can readily determine the dosage and number of dosages needed to produce an effective immune response.

QUANTITATIVE MEASUREMENT EARLY RESPONSE(S) TO REPLIKIN VACCINES

[0205] The ability to measure quantitatively the early specific antibody response in days or a few weeks to a Replikin vaccine is a major practical advantage over other vaccines for which only a clinical response months or years later can be measured.

ADJUVANTS

[0206] Various adjuvants may be used to enhance the immunological response, depending on the host species, including but not limited to Freund's (complete and incomplete), mineral gels, such as aluminum hydroxide, surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, key limpet hemocyanin, dinitrophenol, and potentially useful human adjuvants such as BCG and *Corynebacterium parvum*.

REPLIKIN NUCLEOTIDE SEQUENCES

[0207] Replikin DNA or RNA may have a number of uses for the diagnosis of diseases resulting from infection with a virus, bacterium or other Replikin encoding agent. For example, Replikin nucleotide sequences may be used in hybridization assays of biopsied tissue or blood, e.g., Southern or Northern analysis, including in situ hybridization assays, to diagnose the presence of a particular organism in a tissue sample or an environmental sample, for example. The present invention also contemplates kits containing antibodies specific for particular Replikins that are present in a particular pathogen of interest, or

containing nucleic acid molecules (sense or antisense) that hybridize specifically to a particular Replikin, and optionally, various buffers and/or reagents needed for diagnosis.

5 [0208] Also within the scope of the invention are oligoribonucleotide sequences, that include antisense RNA and DNA molecules and ribozymes that function to inhibit the translation of Replikin- or recognin-containing mRNA. Both antisense RNA and DNA molecules and ribozymes may be prepared by any method known in the art. The antisense molecules can be incorporated into a wide variety of
10 vectors for delivery to a subject. The skilled practitioner can readily determine the best route of delivery, although generally i.v. or i.m. delivery is routine. The dosage amount is also readily ascertainable.

[0209] Particularly preferred antisense nucleic acid molecules are those that are
15 complementary to a Replikin sequence contained in a mRNA encoding, for example, an influenza virus polypeptide, wherein the Replikin sequence comprises from 7 to about 50 amino acids including (1) at least one lysine residue located six to ten residues from a second lysine residue; (2) at least one histidine residue; and (3) at least 6% lysine residues. More preferred are antisense nucleic acid
20 molecules that are complementary to a Replikin present in the coding strand of the gene or to the mRNA encoding the influenza virus hemagglutinin protein, wherein the antisense nucleic acid molecule is complementary to a nucleotide sequence encoding a Replikin that has been demonstrated to be conserved over a period of six months to one or more years and/or which are present in a strain of influenza
25 virus shown to have an increase in concentration of Replikins relative to Replikin

concentration in other influenza virus strains. The increase in Replikin concentration preferably occurs over a period of at least six months, preferably about one year, most preferably about two or three years or more.

5 [0210] Similarly, antisense nucleic acid molecules that are complementary to mRNA those that are complementary to a mRNA encoding bacterial Replikins comprising a Replikin sequence of from 7 to about 50 amino acids including (1) at least one lysine residue located six to ten residues from a second lysine residue; (2) at least one histidine residue; and (3) at least 6% lysine residues. More
10 preferred are antisense nucleic acid molecules that are complementary to the coding strand of the gene or to the mRNA encoding a protein of the bacteria.

DIAGNOSTIC APPLICATIONS

[0211] For organisms such as diatom plankton, foot and mouth disease virus,
15 tomato leaf curl gemini virus, hepatitis B and C, HIV, influenza virus and malignant cells, identified constituent Replikins are useful as vaccines, and also may be usefully targeted for diagnostic purposes. For example, blood collected for transfusions may be screened for contamination of organisms, such as HIV, by screening for the presence of Replikins shown to be specific for the contamination
20 organism. Also, screening for Replikin structures specific for a particular pathological organism leads to diagnostic detection of the organism in body tissue or in the environment.

REPLIKIN STIMULATION OF GROWTH

25 [0212] In another embodiment of the invention, Replikin structures are used to

increase the replication rate of cells, tissues or organs. A method is available to increase replication rates by the addition of specific Replikin structures for other cells, tissues or organs that it is desired to replicate more rapidly, together with or without appropriate stimulæ to cell division know in the art for said cells, tissues
5 or organs to increase the rate of replication and yield. This may be accomplished, for example, by methods known in the art, by modifying or transforming a gene encoding for or associated with a protein or enzyme having a replication function in the organism with at least one Replikin structure.

10 [0213] In another aspect of the invention, Replikin structures are used to increase the replication of organisms. The present invention demonstrates that in influenza virus, for example, increased replication associated with epidemics is associated with increased concentration of Replikins. The increase is due to 1) the reappearance of particular Replikin structures, which were present in previous
15 years, but which then disappeared for one or more years; and/or 2) by the appearance of new Replikin compositions. In addition, in malaria Replikins, repetition of the same Replikin in a single protein occurs.

[0214] Thus, the present invention provides methods and compositions for
20 increasing the replication of organisms. Similarly, in the manner that Replikins of different organisms can be targeted to inhibit replication of any organism, Replikins can be used to increase the replication of any organism. For example, production of rice, maize, and wheat crops, which are critical to feeding large populations in the world, can be improved, for example, by increasing the

concentration (number of Replikins/100 amino acid residues) of any particular strain of rice.

[0215] As an example, in the *Oryza sativa* strain of rice, catalase isolated from immature seeds was observed to contain the following different Replikins within the 491 amino acid sequence of the protein:

kfpdvihafkpnprsh (SEQ ID NO. 638)

kfpdvihafk (SEQ ID NO. 639)

karyvkfhwk (SEQ ID NO. 640)

hpkvspelraiwnylsqedeslgvkianlnvk (SEQ ID NO. 641)

katihkqndfk (SEQ ID NO. 642)

haptpitprpvvgrrqkatihkqndfk (SEQ ID NO. 643)

kfrpsssfdtkttttnagapvwndnealtvgprgpilledyhliekvah (SEQ ID NO. 644)

kfrpsssfdtkttttnagapvwndnealtvgprgpilledyn (SEQ ID NO. 645)

[0216] Thus, by using recombinant gene cloning techniques well known in the art, the concentration of Replikin structures in an organism, such as a food crop plant, can be increased, which will promote increased replication of the organism. For example, inserting additional Replikin sequences like the Replikins identified above into the *Oryza sativa* catalase gene by methods well known in the art will promote this organism's replication.

[0217] Similarly, in the NBS-LRR protein of *Oryza sativa* (japonica cultivar group), the following Replikins were found:

kvkahfqkh (SEQ ID NO. 647)

kvkahfqk (SEQ ID NO. 648)

kdyeidkddlih (SEQ ID NO. 648)

hmkqcfafcavfpk (SEQ ID NO. 650)

hvfweivwrsffqnvkqigsifqrkvryrgqsdvttskihdlmhdlavh (SEQ ID NO. 651)

5 kqigsifqrkvryrgpsdvttskihdlmhdlavh (SEQ ID NO. 652)

kqigsifqrkvryrgpsdvttskihdlmh (SEQ ID NO. 653)

kqigsifqrkvryrgqsdvttskih (SEQ ID NO. 654)

[0218] Further, for aspartic proteinase oryzasin 1 precursor protein, the following

10 Replikins were found:

khgvsagik (SEQ ID NO.655)

htvfdygmrvgfak (SEQ ID NO. 657)

hsryksgqsstyqkngk (SEQ ID NO. 658)

15 [0219] Similarly, in the MADS-box protein FDRMADS3 transcription factor of
Oryza sativa (indica cultivar-group), the following Replikins were found:

kqeamvlkqeinllqkglryiygnraneh (SEQ ID NO. 659)

kqeinllqkglryiygnraneh (SEQ ID NO. 660)

kskegmkaaneilqekiveqnglidvgmmvadqqngh (SEQ ID NO.661)

20 kaaneilqekiveqnglidvgmmvadqqngh (SEQ ID NO. 662)

[0220] Similarly, in LONI MAIZE (ATP-binding redox associated Hydrolase;
Serine protease; Multigene family; Mitochondrion), the following Replikins were
found:

25 kvlaahrygik (SEQ ID NO. 663)

klkiamkhliprvleqh (SEQ ID NO.664)

klkiamkh (SEQ ID NO. 665)

ktslassiakalnrfirislggvkdeadirgh (SEQ ID NO.666)

kalnrkfirislggvkdeadirgh (SEQ ID NO. 667)

5 kfirislggvkdeadirgh (SEQ ID NO. 668)

kvrlskatelvdrlhqlsilvaekitqkvegglsksqk (SEQ ID NO. 669)

hlqsilvaekitqkvegglsksqk (SEQ ID NO. 670)

kvrlskatelvdrlh (SEQ ID NO. 671)

kvggsavesskqdtkngkepihwhskgvaaralh (SEQ ID NO. 672)

10 kvggsavesskqdtkngkepihwh (SEQ ID NO. 673)

kvggsavesskqdtkngkepih (SEQ ID NO. 674)

kqdtkngkepihwhskgvaaralh (SEQ ID NO. 675)

kqdtkngkepih (SEQ ID NO. 676)

15 [0221] Similarly, for Glyceraldehyde 3-phosphate dehydrogenase A, a chloroplast precursor, the following Replikins are found:

hndlrraraalnivptstgaakavslvpnlk (SEQ ID NO. 677)

kvlddqkfgiikgtmttth (SEQ ID NO. 678)

hiqagakkvlitapgk (SEQ ID NO. 679)

20 hgrgdaspldviaindtggvkqashllk (SEQ ID NO. 680)

kqashllk (SEQ ID NO. 710)

[0222] Further, examples of rust resistance-like protein RP1-4 (Zea mays) found include the following Replikins:

25 kvrrvlskdysslkqlmtlmdddiskhlqiiesgleeredkvwmkeniik (SEQ ID NO. 681)

- kvrrvlskdysslkqlmtlmmdddiskh (SEQ ID NO. 682)
- hlqiiesgleeredkvwmkeniik (SEQ ID NO.683)
- hdlreniimkaddlask (SEQ ID NO. 684)
- hvnqlenvigkdealask (SEQ ID NO. 685)
- 5 kkqgyelrqldlnelggslh (SEQ ID NO. 686)
- kqgyelrqldlnelggslh (SEQ ID NO. 687)
- klylksrlkelilewssengmdamnilh (SEQ ID NO. 688)
- hlqllqlngmverlpnkvcnlskrylrgykdqipnigk (SEQ ID NO. 689)
- hlqllqlngmverlpnkvcnlskrylrgyk (SEQ ID NO. 690)
- 10 hlqllqlngmverlpnkvcnlsk (SEQ ID NO. 691)
- hnsnklpksvgelk (SEQ ID NO. 692)
- klpkvgelkh (SEQ ID NO. 693)
- hlsvrvesmqkhkeiyyk (SEQ ID NO. 694)
- khkeiyyk (SEQ ID NO. 695)
- 15 klrdilqesqkflvldlalfkh (SEQ ID NO. 696)
- hafsgaeikdqlrmklqdtaceiakrlgqcplaakvlgsmcrrk (SEQ ID NO. 697)
- hafsgaeikdqlrmk (SEQ ID NO. 698)
- klqdtaceiakrlgqcplaakvlgsmcrrkdiaewkaadvwfeksh (SEQ ID NO. 699)
- kvlgsmcrrkdiaewkaadvwfeksh (SEQ ID NO. 700)
- 20 kdiaewkaadvwfeksh (SEQ ID NO. 701)
- kaadvwfeksh (SEQ ID NO. 702)
- hvppttslptskvfgrnsdrdrvkfllgktttaeastk (SEQ ID NO. 703)
- kailteakqlrdllglph (SEQ ID NO. 704)
- kakaksgkgpillredessstattvmkpfh (SEQ ID NO. 705)
- 25 ksphrgkleswlrrlkeafydaedlldeh (SEQ ID NO. 706)

ksphrgkleswlrrlk (SEQ ID NO. 707)

hrgkleswlrrlk (SEQ ID NO. 708)

ksphrgk (SEQ ID NO. 709)

5 [0223] As discussed previously, the Replikin in wheat ubiquitin activating enzyme E (SEQ ID Nos. 614-616) is conserved. This conservation of Replikin structure provides reliable targets for stimulation of plant growth.

[0224] The close relationship of Replikins to redox enzymes is also clearly
10 indicated in this structure in wheat. Thus, this wheat ubiquitin activating enzyme E activates ubiquitin by first adenylating with ATP its carboxy-terminal glycine residue and, thereafter, linking this residue to the side chain of a cysteine residue in E1 (SEQ ID NO. 614), yielding an ubiquitin-E1 thiolester and free AMP.

15 [0225] A further example of the relationship of wheat Replikins to redox enzymes was also found in the PSABWheat Protein, Photosystem I P700 chlorophyll A apoprotein A2 (PsaB) (PSI-B) isolated from bread Chinese spring wheat Chloroplast *Triticum aestivum*. This protein functions as follows: PsaA and PsaB bind P700, the primary electron donor of photosystem I (PSI), as well as the
20 electron acceptors A0, A1, and FX. PSI functions as a plastocyanin/cytochrome c6-ferredoxin oxidoreductase. Cofactor P700 is a chlorophyll A dimer, A0 is chlorophyll A, A1 is a phylloquinone and FX is a 4Fe- 4S iron-sulfur center. The subunit A psaA/S heterodimer binds the P700 chlorophyll special pair and
25 subsequent electron acceptors. The PSI reaction center of higher plants and algae is composed of one at least 11 subunits. This is an integral membrane protein of

the Chloroplast thylakoid membrane. The 4Fe-4S iron-sulfur"center" to which 'h' bind is critical; hence the significance of 'h' in Replikin structure. Next to bacterial Replikins, these wheat Replikins and plant Replikins are the most primitive evolutionary illustrations of the importance of the Replikin structure to replication and the energy source needed for replication. This basic relationship carries through algae, virus Replikins, bacteria, cancer cells, and apparently all organisms with regard to replication.

[0226] Further examples of Replikins were found in the PSAB Wheat protein, which is critical for wheat growth. These include:

hlqpkwkpslswfknaesrlnhh (SEQ ID NO.617)
hlqpkwkpslswfk (SEQ ID NO. 618)
kwkpslswfknaesrlnhh (SEQ ID NO. 619)
kwkpslswfknaesrlnh (SEQ ID NO. 620)
kpslswfknaesrlnhh (SEQ ID NO. 621)
kpslswfknaesrlnh (SEQ ID NO. 622)
hhaialglhttlilvkgaldargsklmpdkk (SEQ ID NO. 623)
haialglhttlilvkgaldargsklmpdkk (SEQ ID NO. 624)
hhaialglhttlilvkgaldargsk (SEQ ID NO. 625)
haialglhttlilvkgaldargsk (SEQ ID NO. 626)
httlilvkgaldargsklmpdkk (SEQ ID NO.627)
httlilvkgaldargsklmpdk (SEQ ID NO. 628)
httlilvkgaldargsk (SEQ ID NO. 629)

[0227] A further example of the relationship of wheat Replikins to redox is provide in the PSAA_WHEAT Photosystem I 9700 chlorophyll A apoprotein A1, that include:

hhhlaiailfliaghmyrtnwghghgldileahkgpftgqghk (SEQ ID NO. 630)

5 hhlaiailfliaghmyrtnwghghgldileahkgpftgqghk (SEQ ID NO. 631)

hlaiaailfliaghmyrtnwghghgldileahkgpftgqghk (SEQ ID NO. 632)

hmyrtnwghghgldileahkgpftgqghk (SEQ ID NO. 633)

hglkdileahkgpftgqghk (SEQ ID NO. 634)

hdileahkgpftgqghk (SEQ ID NO. 635)

10 hkgpftgqghk (SEQ ID NO. 636)

kgpftgqghk (SEQ ID NO. 637)

COMPUTER SOFTWARE FOR IDENTIFYING REPLIKINS

[0228] The present invention also provides methods for identifying Replikin
15 sequences in an amino acid or nucleic acid sequence. Visual scanning of over four thousand sequences was performed in developing the present 3-point-recognition methods. However, data banks comprising nucleotide and/or amino acid sequences can also be scanned by computer for the presence of sequences meeting the 3 point recognition requirements.

20

[0229] According to another embodiment of the invention, three-point recognition methods described herein may be performed by a computer. Figure 6 is a block diagram of a computer available for use with the foregoing embodiments of the present invention. The computer may include a processor, an input/output device
25 and a memory storing executable program instructions representing the 3-point-

recognition methods of the foregoing embodiments. The memory may include a static memory, volatile memory and/or a nonvolatile memory. The static memory conventionally may be a read only memory ("ROM") provided on a magnetic, or an electrical or optical storage medium. The volatile memory conventionally may be a random access memory ("RAM") and may be integrated as a cache within the processor or provided externally from the processor as a separate integrated circuit. The non-volatile memory may be an electrical, magnetic or optical storage medium.

[0230] From a proteomic point of view the construction of a "3-point recognition" template based on the new glioma peptide sequence led directly to identification of a biology-wide class of proteins having related structures and functions. The operation of the 3-point-recognition method resembles identification by the use of a "keyword" search; but instead of using the exact spelling of the keyword "kagvafihkk" (SEQ ID NO.: 1) as in a typical sequence homology search, or in the nucleotide specification of an amino acid, an abstraction of the keyword delimited by the "3-point-recognition" parameters is used. This delimited abstraction, although derived from a single relatively short amino acid sequence leads to identification of a class of proteins with structures that are defined by the same specifications. That particular functions, in this case transformation and replication, in addition to structures, turn out also to be shared by members of the exposed class suggests that these structures and functions are related. Thus, from this newly identified short peptide sequence, a molecular recognition 'language' has been formulated, which previously has not been described. Further, the sharing of immunological specificity by diverse members of the class, as here

demonstrated for the cancer Replikins, suggests that B cells and their product antibodies recognize Replikins by means of a similar recognition language.

OTHER USES OF THE THREE POINT RECOGNITION METHOD

5 [0231] Since "3-point-recognition" is a proteomic method that specifies a particular class of proteins, using three or more different recognition points for other peptides similarly should provide useful information concerning other proteins classes. Further, the "3-point- recognition" method is applicable to other recognins, for example to the TOLL 'innate' recognition of lipopolyssacharides of
10 organisms. The three point recognition method may also be modified to identify other useful compounds of covalently linked organic molecules, including other covalently linked amino acids, nucleotides, carbohydrates, lipids or combinations thereof. In this embodiment of the invention a sequence is screened for subsequences containing three or more desired structural characteristics. In the
15 case of screening compounds composed of covalently linked amino acids, lipids or carbohydrates the subsequence of 7 to about 50 covalently linked units should contain (1) at least one first amino acid, carbohydrate or lipid residue located seven to ten residues from a second of the first amino acid, carbohydrate or lipid residue; (2) encoding at least one second amino acid, lipid or carbohydrate residue;
20 and (3) at least 6% of the first amino acid, carbohydrate or lipid residue. In the case of screening nucleotide sequences, the subsequence of about 21 to about 150 nucleotides should contain (1) at least one codon encoding a first amino acid located within eighteen to thirty nucleotides from a second codon encoding the first amino acid residue; (2) at least one second amino acid residue; and (3)
25 encodes at least 6% of said first amino acid residue.

[0232] Several embodiments of the present invention are specifically illustrated and described herein. However, it will be appreciated that modifications and variations of the present invention are encompassed by the above teachings and within the purview of the appended claims without departing from the spirit and intended scope of the invention.

EXAMPLE 1

PROCESS FOR EXTRACTION, ISOLATION AND

IDENTIFICATION OF ReplikinS AND THE USE OF *REPLIKINS* TO
TARGET, LABEL OR DESTROY *REPLIKIN*-CONTAINING ORGANISMS

a) Algae

[0233] The following algae were collected from Bermuda water sites and either extracted on the same day or frozen at -20 degrees C and extracted the next day. The algae were homogenized in a cold room (at 0 to 5 degrees C) in 1 gram aliquots in neutral buffer, for example 100 cc. of 0.005M phosphate buffer solution, pH 7 ("phosphate buffer") for 15 minutes in a Waring blender, centrifuged at 3000 rpm, and the supernatant concentrated by preevaporation and dialyzed against phosphate buffer in the cold to produce a volume of approximately 15 ml. The volume of this extract solution was noted and an aliquot taken for protein analysis, and the remainder was fractionated to obtain the protein fraction having a pK range between 1 and 4.

[0234] The preferred method of fractionation is chromatography as follows:

The extract solution is fractionated in the cold room (4 degrees C) on a DEAE cellulose (Cellex-D) column 2.5x11.0 cm, which has been equilibrated with 0.005M phosphate buffer. Stepwise eluting solvent changes are made with the following solutions:

Solution 1 - 4.04 g. NaH_2P_04 and 0.5g NaH_2P_04 are dissolved in 15 litres of distilled water (0.005 molar, pH 7);

Solution 2 - 8.57 g. NaH_2P_04 is dissolved in 2,480 ml. of distilled water;

Solution 3 - 17.1 g. of NaH_2P_04 is dissolved in 2480 ml of distilled water (0.05 molar, pH 4.7);

Solution 4 - 59.65 g. of NaH_2P_04 is dissolved in 2470 ml distilled water (0.175 molar);

Solution 5 - 101.6 g. of NaH_2P_04 is dissolved in 2455 ml distilled water (pH 4.3);

Solution 6 - 340.2 g. of NaH_2P_04 is dissolved in 2465 of distilled water (1.0 molar, pX-i 4.1);

Solution 7 - 283.63 g. of 80% phosphoric acid (H_3P_04) is made up in 2460 ml of distilled water (1.0 molar, pH 1.0).

[0235] The extract solution, in 6 to 10 ml volume, is passed onto the column and overlayed with Solution 1, and a reservoir of 300 ml of Solution 1 is attached and allowed to drip by gravity onto the column. Three ml aliquots of eluant are collected and analyzed for protein content at OD 280 until all of the protein to be removed with Solution 1 has been removed from the column. Solution 2 is then applied to the column, followed in succession by Solutions 3, 4, 5, 6 aid 7 until all

of the protein which can, be removed with each Solution is removed from the column. The eluates from Solution 7 are combined, dialyzed against phosphate buffer, the protein content determined of both dialysand and dialyzate, and both analyzed by gel electrophoresis. One or two bands of peptide or protein of molecular weight between 3,000 and 25,000 Daltons are obtained in Solution 7. For example the algae *Caulerpa mexicana*, *Laurencia obtusa*, *Cladophora prolifera*, *Sargassum natans*, *Caulerpa verticillata*, *Halimeda tuna*, and *Penicillus capitatus*, after extraction and treatment as above, all demonstrated in Solution 7 eluates sharp peptide bands in this molecular weight region with no contaminants. These Solution 7 proteins or their eluted bands are hydrolyzed, and the amino acid composition determined. The peptides so obtained, which have a lysine composition of 6% or greater are Replikin precursors. These Replikin peptide precursors are then determined for amino acid sequence and the Replikins are determined by hydrolysis and mass spectrometry as detailed in U.S. patent 6,242,578 B1. Those which fulfill the criteria defined by the "3-point-recognition" method are identified as Replikins. This procedure can also be applied to obtain yeast, bacterial and any plant Replikins.

b) Virus

[0236] Using the same extraction and column chromatography separation methods as above in a) for algae, Replikins in virus-infected cells are isolated and identified.

c) Tumor cells in vivo and in vitro tissue culture

[0237] Using the same extraction and column chromatography separation methods as above in a) for algae, Replikins in tumor cells are isolated and identified. For example, Replikin precursors of Astrocytin isolated from malignant brain tumors, 5 Malignin (Aglyco IOB) isolated from glioblastoma tumor cells in tissue culture, MCF7 mammary carcinoma cells in tissue culture, and P3J Lymphoma cells in tissue culture each treated as above in a) yielded Replikin precursors with lysine content of 9.1%, 6.7%, 6.7%, and 6.5% respectively. Hydrolysis and mass spectrometry of Aglyco IOB as described in Example 10 U.S. 6,242,578 B1 10 produced the amino acid sequence, ykagvafllhkkndiide the 16-mer Replikin.

EXAMPLE 2:

[0238] As an example of diagnostic use of Replikins: Aglyco IOB or the 16-mer 15 Replikin may be used as antigen to capture and quantify the amount of its corresponding antibody present in serum for diagnostic purposes are as shown in Figures 2,3,4 and 7 of U.S. 6,242,578 B1.

[0239] As an example of the production of agents to attach to Replikins for 20 labeling, nutritional or destructive purposes: Injection of the 16-mer Replikin into rabbits to produce the specific antibody to the 16-mer Replikin is shown in Example 6 and Figures 9A and 9B of U.S. 6,242,578 B1.

[0240] As an example of the use of agents to label Replikins: The use of antibodies to the 16-mer Replikin to label specific cells which contain this Replikin is shown in Figure 5 and Example 6 of U.S. 6,242,578 B1.

5 [0241] As an example of the use of agents to destroy Replikins: The use of antibodies to the 16-mer Replikin to inhibit or destroy specific cells which contain this Replikin is shown in Figure 6 of U.S. 6,242,578 B1.

EXAMPLE 3

10

[0242] Analysis of sequence data of isolates of influenza virus hemagglutinin protein or neuraminidase protein for the presence and concentration of Replikins is carried out by visual scanning of sequences or through use of a computer program based on the 3-point recognition system described herein. Isolates of influenza virus are obtained and the amino acid sequence of the influenza hemagglutinin and/or neuraminidase protein is obtained by any art known method, such as by sequencing the hemagglutinin or neuraminidase gene and deriving the protein sequence therefrom. Sequences are scanned for the presence of new Replikins, conservation of Replikins over time and concentration of Replikins in each isolate.

15

20 Comparison of the Replikin sequences and concentrations to the amino acid sequences obtained from isolates at an earlier time, such as about six months to about three years earlier, provides data that are used to predict the emergence of strains that are most likely to be the cause of influenza in upcoming flu seasons, and that form the basis for seasonal influenza peptide vaccines or nucleic acid

25 based vaccines. Observation of an increase in concentration, particularly a

stepwise increase in concentration of Replikins in a given strain of influenza virus for a period of about six months to about three years or more is a predictor of emergence of the strain as a likely cause of influenza epidemic or pandemic in the future.

5

[0243] Peptide vaccines or nucleic acid-based vaccines based on the Replikins observed in the emerging strain are generated. An emerging strain is identified as the strain of influenza virus having the highest increase in concentration of Replikin sequences within the hemagglutinin and/or neuraminidase sequence during the time period. Preferably, the peptide or nucleic acid vaccine is based on or includes any Replikin sequences that are observed to be conserved in the emerging strain. Conserved Replikins are preferably those Replikin sequences which are present in the hemagglutinin or neuraminidase protein sequence for about two years and preferably longer. The vaccines may include any combination of Replikin sequences identified in the emerging strain.

15

[0244] For vaccine production, the Replikin peptide or peptides identified as useful for an effective vaccine are synthesized by any method, including chemical synthesis and molecular biology techniques, including cloning, expression in a host cell and purification therefrom. The peptides are preferably admixed with a pharmaceutically acceptable carrier in an amount determined to induce a therapeutic antibody reaction thereto. Generally, the dosage is about 0.1 μg to about 10 mg.

20

[0245] The influenza vaccine is preferably administered to a patient in need thereof prior to the onset of "flu season." Influenza flu season generally occurs in late October and lasts through late April. However, the vaccine may be administered at any time during the year. Preferably, the influenza vaccine is administered once yearly, and is based on Replikin sequences observed to be present, and preferably conserved in the emerging strain of influenza virus. Another preferred Replikin for inclusion in an influenza vaccine is a Replikin demonstrated to have re-emerged in a strain of influenza after an absence of one or more years.

10

EXAMPLE 4

[0246] Analysis of sequence data of isolates of *Plasmodium falciparum* antigens for the presence and concentration of Replikins is carried out by visual scanning of sequences or through use of a computer program based on the 3-point recognition method described herein. Isolates of *Plasmodium falciparum* are obtained and the amino acid sequence of the protein is obtained by any art known method, such as by sequencing the gene and deriving the protein sequence therefrom. Sequences are scanned for the presence of Replikins, conservation of Replikins over time and concentration of Replikins in each isolate. This information provides data that are used to form the basis for anti-malarial peptide vaccines or nucleic acid based vaccines.

15

20

[0247] Peptide vaccines or nucleic acid-based vaccines based on the Replikins

observed in the malaria causing organism are generated. Preferably, the peptide or nucleic acid vaccine is based on or includes any Replikin sequences that are observed to be present on a surface antigen of the organism. The vaccines may include any combination of Replikin sequences identified in the malaria causing strain.

[0248] For vaccine production, the Replikin peptide or peptides identified as useful for an effective vaccine are synthesized by any method, including chemical synthesis and molecular biology techniques, including cloning, expression in a host cell and purification therefrom. The peptides are preferably admixed with a pharmaceutically acceptable carrier in an amount determined to induce a therapeutic antibody reaction thereto. Generally, the dosage is about 0.1 μ g to about 10 mg.

[0249] Then malaria vaccine is preferably administered to a patient in need thereof at any time during the year, and particularly prior to travel to a tropical environment.

[250] Another embodiment includes an antisense nucleic acid molecule complementary to the coding strand of the gene or the mRNA encoding organism for the replikins in organisms including, but not limited to, viruses, trypanosomes, bacteria, fungi, algae, amoeba, and plants, wherein said antisense nucleic acid molecules is complementary to a nucleotide sequence of a replikin containing organism.

CLAIMS

1. An isolated or synthesized Replikin peptide consisting of 7 to about 50 amino acids wherein said Replikin peptide consists of a motif comprising:

- (1) at least one lysine residue located at a first terminus of said motif and at least one lysine residue or at least one histidine residue located at a second terminus of said motif;
- (2) a first lysine residue located six to ten residues from a second lysine residue;
- (3) at least one histidine residue; and
- (4) at least 6% lysine residues,

and wherein said Replikin peptide is not isolated from a human, a human immunodeficiency virus, or an arenavirus.

2. The isolated or synthesized Replikin peptide of claim 1 wherein said peptide amino acid sequence consists of 7 to 25 amino acid residues.

3. The isolated or synthesized Replikin peptide of claim 1 wherein said peptide amino acid sequence consists of 7 to 16 amino acid residues.

4. An isolated or synthesized viral Replikin peptide consisting of 7 to about 50 amino acids wherein said Replikin peptide consists of a motif comprising:

- (1) at least one lysine residue located at a first terminus of said motif and at least one lysine residue or at least one histidine residue located at a second terminus of said motif;
- (2) a first lysine residue located six to ten residues from a second lysine residue;
- (3) at least one histidine residue; and
- (4) at least 6% lysine residues,

and wherein said Replikin peptide is not isolated from a human immunodeficiency virus or an arenavirus.

5. An isolated or synthesized viral Replikin peptide consisting of 7 to about 50 amino acids wherein said viral Replikin peptide consists of a motif comprising:

- (1) at least one lysine residue located at a first terminus of said motif and at least one lysine residue or at least one histidine residue located at a second terminus of said motif;
- (2) a first lysine residue located six to ten residues from a second lysine residue;
- (3) at least one histidine residue; and
- (4) at least 6% lysine residues,

and wherein said peptide is a human immunodeficiency virus (HIV) peptide of SEQ ID NO:613, a hepatitis virus peptide, a maize streak virus peptide, a herpes virus peptide, a bovine herpes virus peptide, a meleagrid herpes virus peptide, a feline immunodeficiency virus peptide, a foot and mouth disease virus peptide, a rous sarcoma virus peptide, an avian sarcoma virus peptide, a neuroblastoma RAS viral oncogene peptide, a polyoma virus peptide, a sindbis peptide, a human papilloma virus peptide, a myelomonocytic tumor virus peptide, a murine acute leukaemia peptide, a human T-cell lymphotropic virus peptide, or a tomato leaf curl virus peptide.

6. The peptide according to claim 5, wherein said viral peptide has the sequence as set forth in SEQ ID NO:613.

7. An isolated or synthesized bacterial Replikin peptide consisting of 7 to about 50 amino acids wherein said Replikin peptide consists of a motif comprising:

- (1) at least one lysine residue located at a first terminus of said motif and at least one lysine residue or at least one histidine residue located at a second terminus of said motif;
- (2) a first lysine residue located six to ten residues from a second lysine residue;
- (3) at least one histidine residue; and
- (4) at least 6% lysine residues.

8. The peptide of claim 7, wherein said bacterial peptide is a *Helicobacter* peptide.

9. The peptide of claim 8, wherein said *Helicobacter* peptide is a *Helicobacter pylori* peptide.

10. The peptide of claim 7, wherein said bacterial peptide is a *Staphylococcus* peptide.

11. The peptide of claim 10, wherein said *Staphylococcus* peptide is a *Staphylococcus aureus* peptide.

12. The peptide of claim 7, wherein said bacterial peptide is a *Legionella* peptide.

13. The peptide of claim 12, wherein said *Legionella* peptide is a *Legionella pneumophila* peptide.

14. The peptide of claim 7, wherein said bacterial peptide is an *Acetobacter* peptide, an *Actinomyces* peptide, an *Aerobacter* peptide, an *Arthrobacter* peptide, an *Azotobacter* peptide, a *Bacillus* peptide, a *Brevibacterium* peptide, a *Clostridium* peptide, a *Corynebacterium* peptide, an *Erwinia* peptide, an *Escheria* peptide, a *Klebsiella* peptide, a *Lactobacillus* peptide, a *Nocardia* peptide, a *Proteus* peptide, a *Pseudomonas* peptide, a *Rhizobium* peptide, a *Salmonella* peptide, a *Serratia* peptide, a *Staphylococcus* peptide, a *Streptococcus* peptide, a *Haemophilus* peptide, a *Flavobacterium* peptide, a *Methylomonas* peptide, a *Micrococcus* peptide, a *Mycobacterium* peptide, a *Micronomospora* peptide, a *Mycoplasma* peptide, a *Neisseria* peptide, a *Streptomyces* peptide, a *Streptosporangium* peptide, a *Streptovorticillium* peptide, a *Vibrio* peptide or a *Xanthomas* peptide.

15. An isolated or synthesized fungal Replikin peptide consisting of 7 to about 50 amino acids wherein said Replikin peptide comprises a motif comprising:

- (1) at least one lysine residue located at a first terminus of said motif and at least one lysine residue or at least one histidine residue located at a second terminus of said motif;
- (2) a first lysine residue located six to ten residues from a second lysine residue;
- (3) at least one histidine residue; and
- (4) at least 6% lysine residues

16. The peptide of claim 15, wherein said fungal peptide is a *Penicillium* peptide.

17. The peptide of claim 16, wherein said *Penicillium* peptide is a *Penicillium marneffei* peptide.

18. The peptide of claim 15, wherein said fungal peptide is an *Ophiostoma* peptide.

19. The peptide of claim 18, wherein said *Ophiostoma* peptide is an *Ophiostoma novo-ulmi* peptide.

20. The peptide of claim 15, wherein said fungal peptide is a *Diseula* peptide, a *Candida* peptide, an *Absidia* peptide, an *Aspergillus* peptide, a *Cephalosporium* peptide, a *Fusarium* peptide, a *Hansenula* peptide, a *Mucor* peptide, a *Paecilomyces* peptide, a *Pichia* peptide, a *Rhizopus* peptide, a *Saccaromyces* peptide, a *Torulopsis* peptide, a *Trichoderma* peptide, an *Erysiphe* peptide or a *Phytophthora infenstans* peptide.

21. An isolated or synthesized algal Replikin peptide consisting of 7 to about 50 amino acids wherein said Replikin peptide comprises a motif comprising:

- (1) at least one lysine residue located at a first terminus of said motif and at least one lysine residue or at least one histidine residue located at a second terminus of said motif;
- (2) a first lysine residue located six to ten residues from a second lysine residue;
- (3) at least one histidine residue; and
- (4) at least 6% lysine residues.

22. The peptide of claim 21, wherein said algal peptide is a *Caldophera* peptide, an *Isolepisprolifera* peptide, a *Porphyra* peptide, a *Chondrus* peptide, a *Gracilaria* peptide, a *Gelidium* peptide, a *Caulerpa* peptide, a *Laurencia* peptide, a *Cladophexa* peptide, a *Sargassum* peptide, a *Penicillos* peptide, a *Halimeda* peptide, a *Laminaria* peptide, a *Fucus* peptide, an *Ascophyllum* peptide, an *Undari* peptide, a *Rhodymenia* peptide, a *Macrocystic* peptide, an *Eucheuma* peptide, an *Ahnfeltia* peptide or a

Pteroclaisa peptide.

23. An isolated or synthesized yeast Replikin peptide consisting of 7 to about 50 amino acids wherein said Replikin peptide comprises a motif comprising:

- (1) at least one lysine residue located at a first terminus of said motif and at least one lysine residue or at least one histidine residue located at a second terminus of said motif;
- (2) a first lysine residue located six to ten residues from a second lysine residue;
- (3) at least one histidine residue; and
- (4) at least 6% lysine residues.

24. The peptide of claim 23, wherein said yeast peptide is a *Saccharomyces* peptide, a *Cryptococcus* peptide, a *Cryptococcus neoformans* peptide, or a *Schizosaccharomyces* peptide.

25. An isolated or synthesized amoeba Replikin peptide consisting of 7 to about 50 amino acids wherein said Replikin peptide comprises a motif comprising:

- (1) at least one lysine residue located at a first terminus of said motif and at least one lysine residue or at least one histidine residue located at a second terminus of said motif;
- (2) a first lysine residue located six to ten residues from a second lysine residue;
- (3) at least one histidine residue; and
- (4) at least 6% lysine residues

26. The peptide of claim 25, wherein said amoeba peptide is an *Entamoeba invadens* peptide, an *Amoebidae* peptide, an *Acanthamoeba* peptide, or a *Naegleria* peptide.

27. An isolated or synthesized plant Replikin peptide consisting of 7 to about

50 amino acids wherein said Replikin peptide consists of a motif comprising:

- (1) at least one lysine residue located at a first terminus of said motif and at least one lysine residue or at least one histidine residue located at a second terminus of said motif;
- (2) a first lysine residue located six to ten residues from a second lysine residue;
- (3) at least one histidine residue; and
- (4) at least 6% lysine residues.

28. The peptide of claim 27, wherein said plant peptide is a wheat peptide, a rice peptide, or a maize peptide.

29. The peptide of claim 28, wherein said peptide is a wheat peptide, and said wheat peptide is an Ubiquitin activating enzyme peptide.

30. The peptide of claim 27, wherein said peptide is a PsaB wheat peptide, or a PsaA wheat peptide.

31. An isolated or synthesized structural replication-associated Replikin peptide consisting of 7 to about 50 amino acids wherein said Replikin peptide comprises a motif comprising:

- (1) at least one lysine residue located at a first terminus of said motif and at least one lysine residue or at least one histidine residue located at a second terminus of said motif;
- (2) a first lysine residue located six to ten residues from a second lysine residue;
- (3) at least one histidine residue; and
- (4) at least 6% lysine residues.

32. The peptide of claim 31, wherein said structural replication-associated peptide is a PMCl peptide, a nucleolar scleroderma antigen, a fibrillarin peptide, a SPOP

peptide, a Centromere protein C peptide, a CTCBF, KU antigen peptide, an ATP synthase peptide, a FBRL nuclear peptide, a HP1Hs-alpha peptide, a PM/Sc1 nucleolar peptide, or an amyloid beta A4 precursor peptide.

33. An isolated or synthesized tumor virus associated Replikin peptide consisting of 7 to about 50 amino acids wherein said Replikin peptide comprises a motif comprising:

- (1) at least one lysine residue located at a first terminus of said motif and at least one lysine residue or at least one histidine residue located at a second terminus of said motif;
- (2) a first lysine residue located six to ten residues from a second lysine residue;
- (3) at least one histidine residue; and
- (4) at least 6% lysine residues.

34. An isolated or synthesized transforming-associated Replikin peptide consisting of 7 to about 50 amino acids wherein said Replikin peptide comprises a motif comprising:

- (1) at least one lysine residue located at a first terminus of said motif and at least one lysine residue or at least one histidine residue located at a second terminus of said motif;
- (2) a first lysine residue located six to ten residues from a second lysine residue;
- (3) at least one histidine residue; and
- (4) at least 6% lysine residues.

35. An isolated or synthesized cancer-cell associated Replikin peptide consisting of 7 to about 50 amino acids wherein said Replikin peptide consists of a motif comprising:

- (1) at least one lysine residue located at a first terminus of said motif and at least one lysine residue or at least one histidine residue located at a second

- terminus of said motif;
 - (2) a first lysine residue located six to ten residues from a second lysine residue;
 - (2) at least one histidine residue; and
 - (3) at least 6% lysine residues.
36. An antibody that specifically binds to a viral peptide of claim 4.
37. An antibody that specifically binds to a bacterial peptide of claim 7.
38. An antibody that specifically binds to a fungal peptide of claim 15.
39. An antibody that specifically binds to a yeast peptide or polypeptide of claim 23.
40. An antibody that specifically binds to an amoeba peptide or polypeptide of claim 25.
41. A composition comprising the antibody of any one of claims 36, 37, 38, 39 or 40 and a pharmaceutically acceptable carrier or adjuvant.
42. An antibody cocktail comprising a plurality of antibodies, wherein each of the antibodies specifically binds to a viral peptide of claim 4.
43. An antibody cocktail comprising a plurality of antibodies, wherein each of the antibodies specifically binds to a bacterial peptide of claim 7.
44. An antibody cocktail comprising a plurality of antibodies, wherein each of the antibodies specifically binds to a fungal peptide of claim 15.
45. An antibody cocktail comprising a plurality of antibodies, wherein each of the antibodies specifically binds to a yeast peptide of claim 23.
46. An antibody cocktail comprising a plurality of antibodies, wherein each of the antibodies specifically binds to an amoeba peptide of claim 25.

47. Use of a composition comprising at least one viral Replikin peptide of claim 4 for the preparation of a pharmaceutical composition for stimulating the immune system of a subject to produce antibodies that bind specifically to viral Replikins.

48. Use of a composition comprising at least one non-immune-based organic agent that specifically targets viral Replikin sequences of claim 4.

49. The use of claim 48, wherein the agent is a nucleic acid.

50. The use of claim 49, wherein the nucleic acid is in antisense configuration.

51. The use of claim 48, wherein the composition is administered as an approximately 1 mg dosage form.

52. Use of at least one proteolytic enzyme; and at least one antibody specific for at least one isolated Plasmodium falciparum Replikin peptide wherein said Replikin peptide consists of 7 to about 50 amino acids comprising a motif comprising:

- (1) at least one lysine residue located at a first terminus of said motif and at least one lysine residue or at least one histidine residue located at a second terminus of said motif;
- (2) a first lysine residue located six to ten residues from a second lysine residue;
- (3) at least one histidine residue; and
- (4) at least 6% lysine residues,

for the preparation of a pharmaceutical composition for a dual treatment method of treating malaria.

53. The use of claim 52, wherein a plurality of antibodies specific for Plasmodium falciparum Replikins are administered.

54. Use of at least one proteolytic enzyme; and a preventive or therapeutic vaccine comprising at least one isolated Plasmodium falciparum Replikin peptide wherein said Replikin peptide consists of 7 to about 50 amino acids and comprises a Replikin motif comprising:

- (1) at least one lysine residue located at a first terminus of said motif and at least one lysine residue or at least one histidine residue located at a second terminus of said motif;
 - (2) a first lysine residue located six to ten residues from a second lysine residue;
 - (3) at least one histidine residue; and
 - (4) at least 6% lysine residues,
- for the preparation of a vaccine for treating malaria.

55. The use of claim 54, wherein the vaccine comprises a plurality of Replikin peptides specific for *Plasmodium falciparum*.

56. The use of claim 54, wherein the vaccine comprises overlapping Replikin motifs.

57. A peptide of any one of claims 15 to 26 or 31 to 34 which consists of said motif.